



Contents lists available at ScienceDirect

Molecular Phylogenetics and Evolution

journal homepage: www.elsevier.com/locate/ympev

Phylogenetics of mud snakes (Squamata: Serpentes: Homalopsidae): A paradox of both undescribed diversity and taxonomic inflation

Justin M. Bernstein^{a,*}, John C. Murphy^b, Harold K. Voris^b, Rafe M. Brown^c, Sara Ruane^{a,d}^a Department of Biological Sciences, 206 Boyden Hall, Rutgers University-Newark, 195 University Ave, Newark, NJ 07102, USA^b Science and Education, Field Museum, 1400 S. Lake Shore Drive, Chicago, IL 60605, USA^c Biodiversity Institute and Department of Ecology and Evolutionary Biology, 1345 Jayhawk Boulevard, University of Kansas, Lawrence, KS 66045, USA^d Department of Earth and Environmental Science, Ecology & Evolution, Rutgers University-Newark, 195 University Ave, Newark, NJ 07102, USA

ARTICLE INFO

Keywords:

Biogeography
Diversification
Pleistocene
Sea level fluctuations
Southeast Asia
Quaternary

ABSTRACT

Mud snakes (Serpentes: Homalopsidae) are a family of 55 described, mainly aquatic, species primarily distributed throughout mainland Southeast Asia and the Indo-Australian Archipelago. Although they have been the focus of prior research, the basic relationships amongst genera and species remain poorly known. We used a combined mitochondrial and nuclear gene dataset to infer their phylogenetic relationships, using the highest levels of taxon and geographic sampling for any homalopsid phylogeny to date (62% generic and 62% species coverage; 140 individuals). Our results recover two reciprocally monophyletic groups: the fangless *Brachyorrhos* and its sister clade comprised of all rear-fanged homalopsids. Most genera and interspecific relationships were monophyletic and strongly supported, but intergeneric relationships and intraspecific population structure lack support. We find evidence of both undescribed diversity as well as cases of taxonomic inflation within several species. Tree-based species delimitation approaches (mPTP) support potential new candidate species as distinct from their conspecifics and also suggest that many named taxa may not be distinct species. Divergence date estimation and lineage-through-time analyses indicate lower levels of speciation in the Eocene, with a subsequent burst in diversification in the Miocene. Homalopsids may have diversified most rapidly during the Pliocene and Pleistocene, possibly in relation to tectonic shifts and sea-level fluctuations that took place in Sundaland and the Sahul Shelf. Our analyses provide new insights on homalopsid taxonomy, a baseline phylogeny for the family, and further biogeographic implications demonstrating how dynamic tectonics and Quaternary sea level changes may have shaped a widespread, diverse family of snakes.

1. Introduction

The field of systematics strives to discover and describe the evolutionary relationships of life and integrate this knowledge to understand how species and populations interact and change over time. Phylogenies are the baseline tool to discover biodiversity and provide the historical framework for analyses that investigate taxonomic and biogeographic hypotheses (Grismer et al., 2016; Li and Li, 2018; Miralles et al., 2018), hybridization (Burbrink and Gehara, 2018; Dufresnes et al., 2020), ancestral traits (Gamble et al., 2015; Takeuchi et al., 2018), biogeography and speciation (Hinckley et al., 2020; Tomasello et al., 2020), and phenotypic trait evolution (Mahler et al., 2010; Setiadi et al., 2011; Bergmann et al., 2020). Snakes have increasingly been used as a model

system to investigate evolutionary and ecological hypotheses (Shine and Bonnet, 2000; Lillywhite and Martins, 2019), in part due to their successful colonization of all continents except Antarctica, and having adapted to a wide variety of environments, including terrestrial, fossorial, arboreal, marine, and freshwater habitats (Greene, 1997). Even within families and genera, snakes are diverse in morphology, behavior, and habitat preference. Broadly construed, snakes include numerous lineages with both exceptionally wide and narrow, range-restricted geographic distributions, making them ideal for investigating biogeographic hypotheses, speciation, and adaptation, particularly in relation to body plan evolution, body size, and associated morphological traits. Although the monophyly of snakes is well-supported and among-family relationships have begun to stabilize (e.g., Pyron et al., 2013; Zheng and

* Corresponding author.

E-mail addresses: jmbernst223@gmail.com (J.M. Bernstein), hvoris@fieldmuseum.org (H.K. Voris), rafe@ku.edu (R.M. Brown), sara.ruane@rutgers.edu (S. Ruane).<https://doi.org/10.1016/j.ympev.2021.107109>

Received 18 September 2020; Received in revised form 20 November 2020; Accepted 5 February 2021

Available online 18 February 2021

1055-7903/© 2021 Elsevier Inc. All rights reserved.

Wiens, 2016; Burbrink et al., 2020), relationships within many families remain unresolved. Resolution of remaining intrafamilial systematic problems are critical challenges for the use of snakes as model study systems in evolutionary biology.

Mud snakes (Serpentes: Homalopsidae) are a group of 55 species in 29 genera, distributed as far west as the Indus River of Pakistan and ranging eastward throughout South and mainland Southeast Asia, the Philippines, the Indo-Australian archipelago, New Guinea, and northern Australia (Murphy and Voris, 2014). Homalopsids are primarily aquatic, with some species specializing in salt or brackish water systems (e.g., mangroves, coastlines), as well as freshwater systems (e.g., rivers, lakes; Murphy, 2007). Many homalopsids have morphological characters, such as semicircular valvular nostrils and dorsally located eyes, that are presumed adaptations for their largely aquatic lifestyles (Voris et al., 2002). The extraordinary morphological and ecological diversity within this group has been used to study evolutionary phenomena such as head shape and diversification (Fabre et al., 2016), feeding performance (Jayne et al., 2018), and for testing hypotheses regarding biogeography in relation to Southeast Asia's complex geologic history (Alfaro et al., 2008). Although these studies demonstrate the value of particular focal-clade homalopsid study systems for understanding broad evolutionary themes, they are dependent on incorporation of a well-resolved phylogenetic estimate, which currently does not exist for the Homalopsidae.

Homalopsids have been long recognized as a distinct group of snakes (Bonaparte, 1845; Jan, 1863) but various studies have led to disparate hypotheses regarding their phylogenetic position, resulting in a convoluted nomenclatural history (e.g., Gray, 1849; Boulenger 1890, 1896; Smith, 1943; Gyi, 1970; Knight and Mindell, 1994; Underwood, 1999). For example, species now recognized as homalopsids have previously been a part of/considered closely related to the families Colubridae, Natricidae, Viperidae, and Pareatidae (Murphy and Voris, 2014). Although the monophyly, position, and establishment of Homalopsidae as a family has consistently been supported in modern phylogenies (Voris et al., 2002; Lawson et al., 2005; Pyron et al., 2013; Burbrink et al., 2020), their intergeneric and interspecific relationships remain poorly resolved. Several fine-scale analyses have been conducted for the genera *Cerberus* (Alfaro et al., 2004) and *Enhydryis* (Karns et al., 2010), but taxon and geographic sampling remain low in familial-level trees, as many species are known from only a few specimens and/or collected before tissue sampling for molecular analysis was standard practice. The most taxonomically comprehensive homalopsid tree to date is based on a mitochondrial (mtDNA) gene and included 31 specimens (45% and 62% species and generic coverage, respectively; Quah et al., 2018). Prior work on the family (Alfaro et al., 2008) hypothesized divergence times for homalopsids, but with 38% of species, 48% of genera, and did not include the homalopsid genus *Brachyorrhos*; this genus, along with two other poorly known genera (*Calamophis*, *Karnsophis*), are ecologically disparate and hypothesized to be the fangless sister group to all other homalopsid taxa, which are rear-fanged. Thus, the inclusion of fangless homalopsids is essential for inferring accurate divergence dates and any inferences of evolutionary transitions in phenotypic or biogeographic character states.

Here, we infer a phylogeny of the Homalopsidae using a combined mitochondrial and nuclear dataset, with the highest species-, individual-, and geographic-level representation to date (62% and 62% species and generic coverage, respectively). We infer the first species tree and time-calibration analyses for the Homalopsidae to include the morphologically disparate, fangless genus *Brachyorrhos*. Here, we provide an initial phylogenetic framework to (1) reveal and resolve relationships within Homalopsidae with increased generic, species, individual, and geographic sampling (140 specimens across the geographic ranges of 34 species), (2) identify the position of species that have never been previously included in any phylogeny, (3) conduct time-calibration analyses to correlate homalopsid diversification with events in a geological record, and (4) provide insights into the biogeography of the family.

2. Methods and materials

2.1. Taxon sampling, DNA extraction, and sequencing

We sampled a total of 140 homalopsids, representing 34 (62%) species and 18 (62%) genera (see Table S1 in Supplemental Material) from the rear-fanged and fangless groups; all missing species are only known from holotypes and/or formalin-preserved museum specimens and were unavailable for this project. Most publicly available sequence data for homalopsids are from mtDNA cytochrome *b* (cyt-*b*) and nuclear prolactin receptor (PRLR) regions, so we sequenced these two loci for new samples to maximize compatibility of our data with previously sampled specimens. We obtained 98 homalopsid tissues from 16 museum collections and included 48 archived sequences from Genbank (NCBI; 34 cyt-*b*, 14 PRLR [Alfaro et al., 2008; Karns et al., 2010; Kumar et al., 2012; Murphy et al., 2012a; Wiens and Tiu, 2012; Alencar et al., 2016; Ukuwela et al., 2017]; see Table S1 in Supplemental Material). This sampling covers the majority of the geographic range of most species. For completeness, we also include a previously identified (*Hypsiscopus* from Lake Towuti, Sulawesi. Homalopsidae is consistently found as a distinct lineage from other caenophidian snakes (e.g., viperids, natricids, elapids, lamprophiids; Pyron et al., 2013; Figueroa et al., 2016; Burbrink et al., 2020), and thus we included outgroup sequences of the following from Genbank: *Crotalus horridus*, *Bitis nasicornis*, *Nerodia sipedon*, *Bungarus fasciatus*, *Laticauda laticaudata*, *Laticauda frontalis*, *Boaedon fuliginosus* (Alfaro and Arnold, 2001; Nagy et al., 2003; Sanders et al., 2008; Marges et al., 2015; see Table S1 in Supplemental Material, museum and institution codes follow Sabaj [2016]).

Genomic DNA was isolated from liver and muscle tissue using Qiagen® DNeasy blood and tissue kit protocols. In brief, we lysed tissues using proteinase K and ATL lysis buffer for 12 h at 56 °C, and subsequently followed the Qiagen® kit protocol. We amplified cyt-*b* and PRLR using the polymerase chain reaction (PCR) with primers L14910 and H16064 (cyt-*b*, ~1100 base pairs targeted; Burbrink et al., 2000) and PRLR_f1 and PRLR_r3 (PRLR, 532 base pairs targeted; Townsend et al., 2008). We ran 25 µl-PCR reactions with a 5-min 94 °C initial denaturation step, followed by 35 cycles of denaturing at 94 °C for 45 s, primer annealing at 52 | 57.2 °C (cyt-*b*|PRLR) for 45 s, and elongation at 72 °C for 60 s, followed by a final extension step of 5 min at 72 °C. We used ExoSAP-IT™ PCR Product cleanup reagent (Thermo Fisher Scientific) to clean PCR products prior to sequencing. Genomic DNA was quantified on a Qubit 3 fluorometer (Thermo Fisher Scientific: Invitrogen) and Sanger-sequenced on an ABI 3730xl DNA analyzer at Macrogen (New York, United States). Resulting sequences were aligned for each gene using the MUSCLE alignment option under default parameters in Geneious v7.1.9 (Geneious, 2020; <https://www.geneious.com>) and we checked alignments by eye to ensure these protein coding loci were within the correct reading frame and to remove ambiguous base pairs at the beginning or ends of sequences. We checked PRLR sequence chromatograms for heterozygous sites; no individuals were heterozygous at any sites, so PRLR data were not phased.

2.2. Phylogenetic analysis

To initially explore the genetic diversity and structure amongst taxonomically and geographically distinct populations, we used maximum likelihood (ML) and Bayesian inference (BI) to reconstruct phylogenetic relationships on a concatenated dataset. All gene trees and concatenated, dual-gene trees were generated using the CIPRES Science Gateway v3.3 (Miller et al., 2010). Evolutionary models and partitioning schemes for all DNA alignments and subsequent RAXML and MrBayes analyses were selected by partitioning the concatenated dataset by gene and codon position in PartitionFinder2; the best-fit nucleotide substitution models and partitioning schemes for ML and BI analyses were

selected under the Akaike information criterion (AICc; Lanfear et al. 2016). We ran RAxML v8.2.10 (Stamatakis, 2014) for 1000 bootstrap iterations under a general time-reversible (GTR; Rodríguez et al., 1990) evolutionary model with a gamma distribution (GTRGAMMA) to account for branch rate heterogeneity. For BI, we explored parameter space, using MCMC sampling procedures, for 100 million generations (25% burn-in) in MrBayes v3.2.6 under a GTR model, allowing for a proportion of invariant sites ('invgamma' option; Ronquist et al., 2012).

We calculated uncorrected pairwise cyt-b distances between genetically or geographically proximate specimens in Geneious v7.1.9 using the pairwise matrix function; PRLR distances were not calculated due to lower sampling and resolution for the PRLR gene tree (see Results: 3.1). Trees were visualized in FigTree v1.4 (Rambaut, 2014), and relationships were considered strongly supported when accompanied by nodal bootstrap (BS) values ≥ 70 (Hillis and Bull, 1993) and Bayesian posterior probabilities (PP) ≥ 0.95 . To make visual comparisons between ML and BI trees, we used the 'cophylo' function in the *phytools* package (Revell, 2012) in R v3.6.1 (R Core Team, 2019).

Because concatenated analyses may not represent the true history of species due to gene-tree-species-tree discordance (Maddison, 1997), we estimated a species tree using the StarBEAST2 package in BEAST v2.6.2 (Bouckaert et al., 2014) and selected and averaged over all models of evolution using the 'bModelTest' package (Bouckaert and Drummond, 2017). We used the concatenated tree and mPTP analysis results (see Methods: 2.3 and Results: 3.2) to designate terminal taxa, including both described species and putatively-undescribed candidate species (>2 specimens). To identify the appropriate diversification model, both Yule and birth-death process analyses were run for 100 million generations, with an uncorrelated lognormal relaxed clock to allow for rate heterogeneity across lineages. The species-tree was rerun five times under the model with the highest likelihood (measured in Tracer v1.6 [Rambaut et al., 2018]) with default priors for 70 million generations and sampled and stored every 7000 generations each run. To reduce parameterization, we constrained the divergence-dated tree to recover Homalopsidae as monophyletic and the rear-fanged taxon (*Brachyorrhos*) as monophyletic (consistent with all previous homalopsid literature and our own analyses). We checked for run convergence (10% burn-in) and effective sample sizes (ESS) in Tracer v1.6. Effective sample sizes >200 (100 minimum) were considered efficiently sampled for the respective parameters (Bouckaert et al., 2014). All trees (14,001 trees) from the analysis were combined using TreeAnnotator and visualized in FigTree.

2.3. Tree-based assessment of taxonomic validity

The use of multispecies coalescent methods in BEAST involves setting *a priori* grouping of taxa for analysis, which, if based on current taxonomy, may not accurately represent true diversity of the group. Because this is the first time a species tree analysis has been performed on homalopsid snakes with this level of sampling, we also use Markov Chain Monte Carlo (MCMC) sampling with a multi-rate Poisson Tree Processes (mPTP; Kapli et al., 2017) for assessing and quantifying the level of statistical support for described species in our concatenated tree. The mPTP method delimits species, while taking phylogeny and number of substitutions into account; this method does not require any input similarity threshold (Kapli et al., 2017). More computationally intensive approaches, such as Bayesian MCMC programs in Bayesian Phylogenetics and Phylogeography (BPP; Yang and Rannala, 2010; Yang, 2015), are widely used for "discovery-stage" delimitation and proposition of cryptic or undescribed diversity (e.g., Brown et al., 2012; Ruane et al., 2014; Ramirez-Reyes et al., 2020; Burbrink and Ruane, 2021). However, in this instance, a lack of an *a priori* or hypothesized grouping of taxa, and our use of two, non-neutral loci, would likely provide poor estimates of population size (θ) and divergence time (τ) parameters, ultimately limiting the credibility of species delimitation results in BPP (McKay et al., 2013). Thus, we used a mPTP analysis, which has been found to produce similar results, or even outperform other single-locus methods,

such as the general mixed Yule coalescent (GMYC; Luo et al., 2018). We performed two MCMC runs using a single-threshold for 500 million generations and a 10% burn-in to quantitatively assess the taxonomic validity of named populations in our sampling, as well as to see if conspecifics separated by deep divergences are delimited as putative species. This analysis was only run on the concatenated dataset to maximize the delimitation analysis on the highest number of taxonomic and geographic samples. Whereas mPTP, similar to the general mixed Yule coalescent (GMYC) method, was originally intended as a single-locus delimitation technique, it is also used for concatenated datasets of nuclear genes (Parnmen et al., 2012), nuclear + mitochondrial loci (Arrigoni et al., 2016), and single nucleotide polymorphisms (SNPs; Nieto-Montes de Oca et al., 2017).

2.4. Divergence time estimation

No fossils of homalopsid snakes have been described; to calibrate our species tree, we relied on secondary calibrations from Burbrink et al. (2020), which contains the densest sampling of homalopsid snakes using genomic data (seven species) for any published dating analysis. Using the R packages *phytools* (Revell, 2012), *ape* (Paradis and Schliep, 2019), and *HDInterval* (Meredith and Kruschke, 2018), we extracted the distributional mean and 95% highest posterior density (HPD) of dates from two nodes in Burbrink et al. (2020): the most recent common ancestor (MRCA) of all homalopsids (95% HPD = 46.08–49.63 mya) and the MRCA of the three crustacean-eating homalopsids, *Fordonia leucobalia* + *Gerarda prevostiana* + *Cantorina violacea* ('FGC' clade; 95% HPD = 8.33–9.21 mya). We used the 95% HPD as log normal distributions for the upper and lower bounds on our calibrations. Although the reduced sampling of homalopsids in Burbrink et al. (2020) may raise concern as to the validity of the FGC clade date, the divergence analysis using only the Homalopsidae MRCA calibration consistently returned poor ESS values (<10) for almost all statistics, and several bimodal posterior distributions (i.e., lack of convergence; see File S1–S2 in Supplemental Material). Thus, we used both node calibrations in our analysis. The mean (M) date and standard deviation (S) set used in our analysis were: Homalopsidae: M = 47.723, S = 0.018; FGC clade: M = 8.783, S = 0.026.

To observe the rate of homalopsid diversification over time, we constructed lineage-through-time (LTT) plots and calculated Pybus and Harvey's gamma (γ ; Pybus and Harvey, 2000) statistic in R v1.1.4 using the 'litt' function in the package *phytools*. We compared the LTT plot of our chronogram to 100 simulations of the dataset under a pure birth model of diversification.

3. Results

3.1. Maximum likelihood, Bayesian inference, and pairwise distances

The final alignments for cyt-b, PRLR, and the concatenated dataset were 1053, 585, and 1638 base pairs, and the cyt-b and PRLR genealogies included 123 and 91 homalopsids respectively (See Figs. S1–S2 in Supplemental Material). Although our concatenated alignment (140 homalopsids) has 12.8% and 35.5% missing specimen data for cyt-b and PRLR, respectively, it has a higher species representation than the individual gene trees (eight species for cyt-b and one species for PRLR were only represented by that single gene in the concatenated dataset). Phylogenetic accuracy has been shown to increase if there is a higher coverage of taxon sampling compared to data coverage (Hedtke et al., 2006; Wiens and Tiu, 2012). Thus, we particularly focus on the results of the concatenated and species trees in our results and discussion, which include the highest sample representation; we include the single gene trees in the supplemental materials (See Figs. S1–S2 in Supplemental Material). The complete list of specimens used in this study and their associated Genbank accession numbers are provided in Supplemental Material Table S1.

For both loci, the ML and BI analyses indicated a strongly supported, monophyletic Homalopsidae (See Fig. S1 in [Supplementary Material](#)). Most genera in the ML and BI gene trees for PRLR were monophyletic; *Gyiophis* and *Myrrophis* are weakly supported within *Enhydris*. In contrast, all genera were monophyletic in the cyt-b gene trees. The BI cyt-b tree poorly supported *Hypsiscopus* as sister to all other homalopsids (including *Brachyorrhos*) and contained a polytomy for the placement of several genera. Bootstrap and posterior probabilities of both gene trees had comparable support values, and were similar to the concatenated tree, discussed below.

Concatenated ML and BI analyses strongly supported a monophyletic Homalopsidae (BS/PP = 1; [Fig. 1](#)), composed of two major clades: the fangless clade comprised solely of the genus *Brachyorrhos* and a rear-

fanged clade of all other homalopsids. Most relationships were congruent between ML and BI trees and had comparable support values, with well-supported branches at the genus, species, and sister species levels, and poorly supported relationships among most genera ([Fig. 1](#), [Supplemental Figs. S1–S2](#)). The major topological difference between the concatenated ML and BI analyses was the placement of *Raclitia indicia* ([Fig. 1](#)).

Uncorrected pairwise cyt-b distances were calculated for specimens that were either genetically related (based on results) or geographically proximate (or both), rather than pre-existing species taxonomy, due to several cases of possible undescribed diversity or taxonomic inflation ([Fig. 2](#)). Within *Brachyorrhos*, there was strong support for *B. albus* as part of *B. raffrayi*, with a minimal genetic distance of 0.48%. Low levels

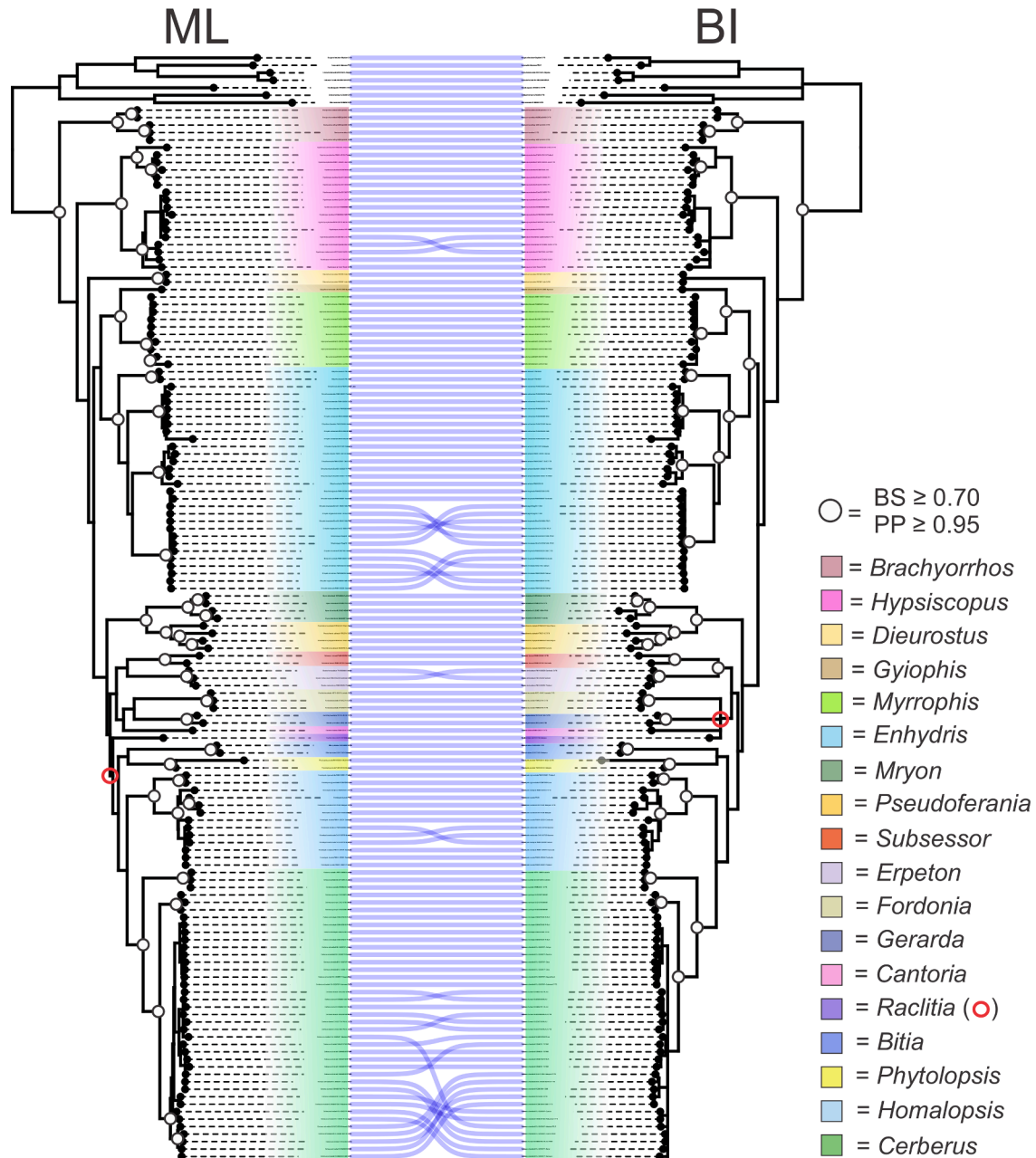


Fig. 1. Comparison of concatenated maximum likelihood (ML) and Bayesian inference (BI) homalopsid phylogenies. Genera are color-coded on each tree. Purple lines are attached to identical tips of both trees, emphasizing areas of congruence versus topological disparity among ML and BI estimates. Unhighlighted clade in both trees are the outgroups. The red circles denote the position of *Raclitia indicia*, the only genus recovered in different positions between both analyses. White circles at divergences indicate BS \geq 0.70 and PP \geq 0.95. Note: dashed lines from tips to taxon labels are for ease of visualization and are not representative of any analytical results. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

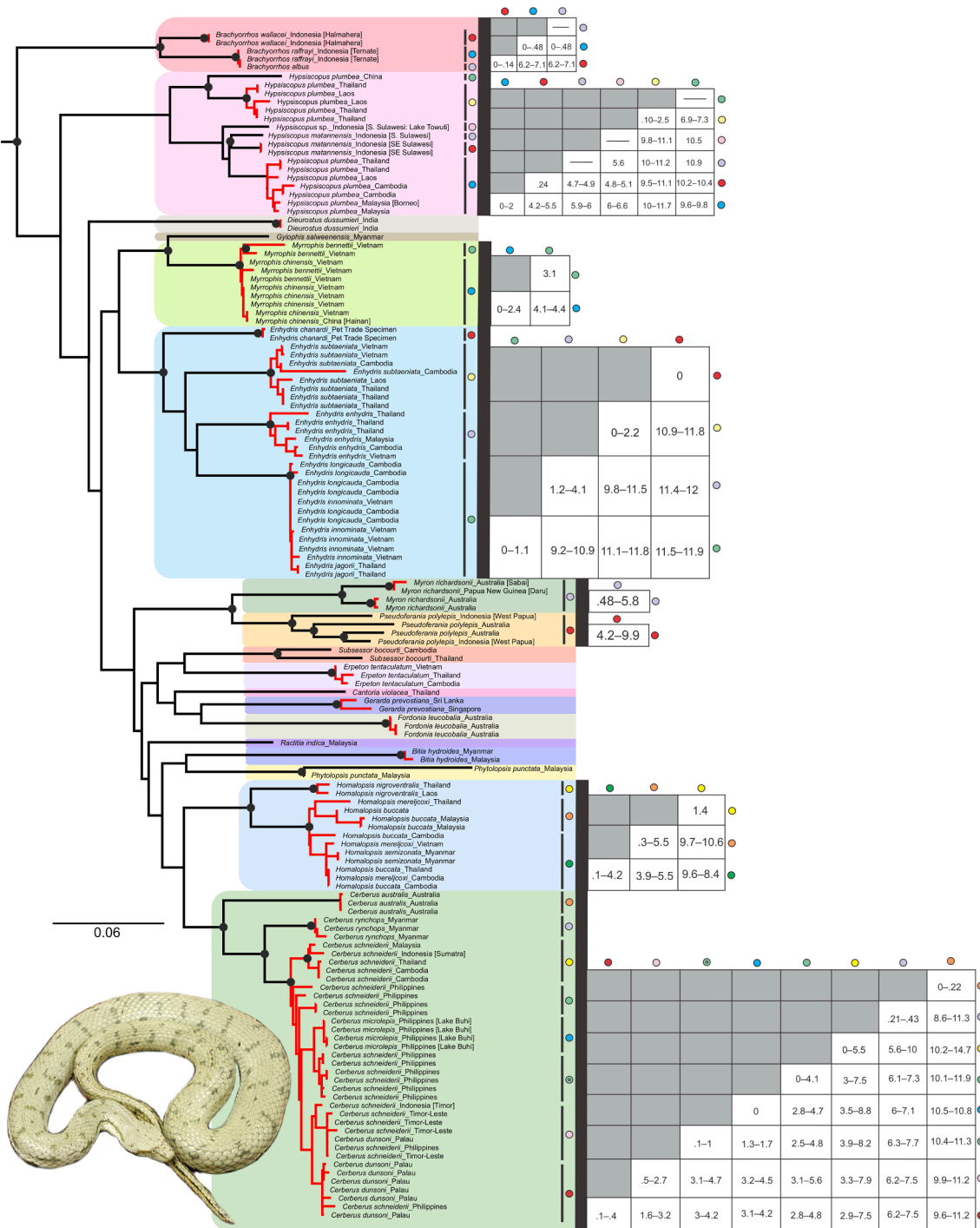


Fig. 2. Concatenated homalopsid ML tree with mPTP species delimitation results (outgroups not shown). Black nodes indicate bootstrap values ≥ 70 . Red clades and single black, terminal branches represent species from the mPTP analysis. Cytochrome *b* genetic distances are given for genera with high levels of diversity. Pairwise distance matrices are based by group, denoted by the adjacent colored circles. Photograph of *Cerberus australis* NTM R29853 courtesy of Museum and Art Gallery of the Northern Territory. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

of interspecific divergence were also seen in some specimens of *Myrophis bennettii* and *M. chinensis*. *Enhydris longicauda*, *E. innominata*, and *E. jagorii* are all recovered within a single clade with pairwise distances of 0–1.1%. Three of the seven *Homalopsis buccata* specimens, and all *H. mereljcoxi* and *H. semizonata*, were in a clade that is 0.5–1.27% in genetic distance from one another. Finally, the Palau-endemic *Cerberus dunsoni* and Philippine Lake Buih-endemic *C. microlepis* both formed clades within the geographically widespread *C. schneiderii*. The pairwise distances between *C. dunsoni*, *C. microlepis*, and closely-related

C. schneiderii were 1.6–3.2% and 1.3–1.7%, respectively.

Intraspecific and intragroup (for potentially synonymous taxa) distances for several species were much greater than intraspecific distances seen in other taxa. The two Australian *Pseudoferania polylepis* specimens are not sister to each other, and, with the New Guinea specimens, have an intraspecific variation of 4.2–9.9%. Two distinct clades of *Homalopsis buccata* + *Homalopsis mereljcoxi* were 3.9–5.5% divergent; one of these groups contains specimens from Malaysia and Thailand, whereas the other clade consists of Thailand, Cambodia, and Myanmar specimens

(Fig. 2). The latter clade also contains *H. semizonata*. Specimens of *Cerberus schneiderii* from Cambodia, Peninsular Malaysia, and Sumatra are sister to all other *C. schneiderii* (Philippines, Palau, Indonesia [Timor Island]); these two groups have an intergroup divergence of 3.0–8.8%. *Cerberus dunsoni*, *C. microlepis*, and the Timor population of *C. schneiderii* are all recovered as distinct geographic lineages within the Philippine *C. schneiderii* clade (Fig. 2). We note, however, that one specimen of Timor *C. schneiderii* and *C. dunsoni* are swapped from their respective clades; both specimens have missing cyt-b data and each group is monophyletic in the cyt-b gene tree (see Fig. S1–S2 in Supplemental Material) and thus this may be an artifact of missing data. The Philippines + Palau + Timor *C. schneiderii* has an intragroup cyt-b distance range of 0–5.6%. In *Hypsiscopus plumbea*, the specimen from China and those from Thailand and Laos are sister groups and are 6.9–7.3% divergent from one another; this group is sister to all other Southeast Asian *H. plumbea* and has an intraspecific cyt-b distance of 9.5–11.7% (Fig. 2). Finally, specimens of *Myron richardsonii* from Australia are ~6% divergent from the specimens with localities on or near Papua New Guinea (PNG).

3.2. Concatenated candidate species delimitation

The mPTP analysis proposed 36 species from the 34 formally described species comprising our dataset, and, while provided quantitative support of synonymy, also yielded support for (or proposed) undescribed species diversity. Non-monophyletic species in the concatenated tree were supported as one species in the mPTP analysis. The delimitation results supported the grouping of *Brachyorrhos raffrayi* and

Table 1

Results from the mPTP analysis that indicate taxonomic inflation or undescribed diversity for each genus. Each species recovered as one species (synonymous) or multiple species (undescribed) is given with the country-level localities of the specimens. International Organization of Standardization (ISO) codes for countries are as follows: CN = China; ID = Indonesia [Sw.: Sulawesi]; KH = Cambodia; LA = Laos; MM = Myanmar; MY = Malaysia; PH = Philippines; PW = Palau; TH = Thailand; VN = Vietnam. No proposed candidate or synonymized species are classified as not applicable (NA). Note: No species name is suggested as the preferred taxonomy. Morphological data from previous studies on these species are included in File S3 of the Supplemental Material.

Genus	Synonymous Species	Proposed Candidate Species
<i>Brachyorrhos</i>	<i>B. raffrayi</i> ID [Ternate] <i>B. albus</i>	NA
<i>Homalopsis</i>	<i>H. buccata</i> KH, MY, TH, <i>H. mereljcoxi</i> KH, TH, VN <i>H. semizonata</i> MM	NA
<i>Cerberus</i>	<i>C. schneiderii</i> ID, KH, PH, MY, TH <i>C. microlepis</i> PH <i>C. dunsoni</i> PW	NA
<i>Myrrophis</i>	<i>M. bennettii</i> VN <i>M. chinensis</i> CN, VN	NA
<i>Enhydris</i>	<i>E. longicauda</i> KH <i>E. innominata</i> VN <i>E. jagorii</i> TH	NA
<i>Hypsiscopus</i>	NA	<i>H. plumbea</i> CN <i>H. plumbea</i> TH, LA <i>H. plumbea</i> KH, TH, LA, MY <i>H. plumbea</i> ID [S. Sw.] <i>H. plumbea</i> ID [S. Sw.: Towuti] <i>H. plumbea</i> ID [SW. Sw.]
<i>Myron</i>	NA	<i>M. richardsonii</i> PG <i>M. richardsonii</i> AU
<i>Pseudoferania</i>	NA	<i>P. polylepis</i> ID [West Papua] <i>P. polylepis</i> ID [West Papua] <i>P. polylepis</i> AU <i>P. polylepis</i> AU
<i>Phytolopsis</i>	NA	<i>P. punctata</i> MY <i>P. punctata</i> MY
<i>Subsector</i>	NA	<i>S. bocourti</i> KH <i>S. bocourti</i> TH

B. albus as a single taxon (Table 1). The proposal to synonymize several species was also recovered in closely-related populations of *Homalopsis* (*H. buccata* + *H. mereljcoxi* + *H. semizonata*), *Cerberus* (*C. schneiderii* + *C. microlepis* + *C. dunsoni*), *Myrrophis* (*M. bennettii* + *M. chinensis*), and *Enhydris* (*E. longicauda* + *E. innominata* + *E. jagorii*; Table 1).

In contrast, undescribed species diversity was proposed in *Hypsiscopus*, *Myron*, *Pseudoferania*, and *Phytolopsis* (Table 1). Our analysis putatively splits *Hypsiscopus* into six potential species: *H. sp.* (Lake Towuti, Sulawesi), *H. plumbea* (China), *H. plumbea* (Thailand + Laos), *H. plumbea* (Thailand + Laos + Cambodia + Malaysia), *H. matannensis* (South Sulawesi), and *H. matannensis* (Southeast Sulawesi). The New Guinea and Australian specimens of *Myron richardsonii* were each recovered as potentially distinct species. This was also seen with *Subsector bocourti*, one specimen each from Cambodia and Thailand as potentially different species, and to a greater extent with *Pseudoferania polylepis*, with all four specimens identified as separate candidate species. Although from the same locality, the two *Phytolopsis punctata* were also preliminarily delimited; this is most likely an artifact due to missing PRLR data in one of the two specimens; mPTP analyses utilize branch lengths and phylogeny and, thus, the long branch from the sample with both gene regions was proposed as a distinct species. The same may also explain preliminary delimitation of the two specimens of *S. bocourti* as different species (but from two different localities).

3.3. Species-tree inference and divergence date estimation

For the species trees, the birth-death process of diversification had a higher likelihood score than the Yule process (-20052.08 for birth-death, -20056.17 for Yule), so the former model was used; ESS values were generally >200. In the species tree, Homalopsidae is well-supported as monophyletic, with a sister relationship between the fanged and fangless clades (Fig. 3A). The relationships in the species tree were similar to the concatenated tree, with most intergeneric relationships poorly supported. With the exception of *Enhydris*, all polytypic genera were recovered with strong support. The major differences between the species and concatenated trees were the placements of *Dieurostus*, *Racliitia*, *Erpeton* + *Subsector*, and *Bitia* + *Phytolopsis* (Figs. 2, 3A).

The *H. plumbea* from China was sister to the Thailand + Laos (northern) *H. plumbea* populations, and the Thailand + Laos + Cambodia + Malaysia (southern) populations were sister to *H. matannensis* + *H. sp.*-Lake Towuti (Fig. 3B). Similar to the concatenated analysis, the Australian and PNG *Myron richardsonii* were reciprocally monophyletic (Fig. 3C), and the West Papuan and Australian *Pseudoferania polylepis* were not sister taxa (Fig. 3D).

The LTT plot under the birth-death model and Pybus and Harvey's γ statistic indicate that homalopsids slowly diversified early in their evolution, and subsequently had a rapid increase in their diversification rate around 10 mya (HPD 8.75–11.66; $\gamma = 3.869$, p -value = 0.0001; Fig. 3A). Under a pure-birth model of diversification, LTT plot simulations ($n = 100$) showed a gradual increase in diversification through time (Fig. 3A). The dated analysis suggests a mid-Eocene (45.31 mya; 95% HPD 43.68–46.88 my) diversification of the crown homalopsid group (Fig. 3A). The fangless *Brachyorrhos* diversified during the Pleistocene (1.49 mya; 95% HPD 0.18–2.58 my), and the rear-fanged clade diversified in the upper-Miocene (10.14 mya; 95% HPD 8.75–11.66 my), with most subsequent divergences between rear-fanged genera occurring between 4.5 and 9.8 million years ago (Fig. 3A). The undescribed lineages of *H. plumbea*, *M. richardsonii*, and *P. polylepis* split from their sister lineages 2.47 ± 1.31 , 0.83 ± 0.75 , and 0.66 – 2.58 ± 0.60 – 0.91 mya respectively (Fig. 3B, C, D).

4. Discussion

Advances in phylogenomics and bioinformatics have resulted in a greater understanding of the evolution and diversity of numerous organismal groups, yet many families are still poorly known with respect

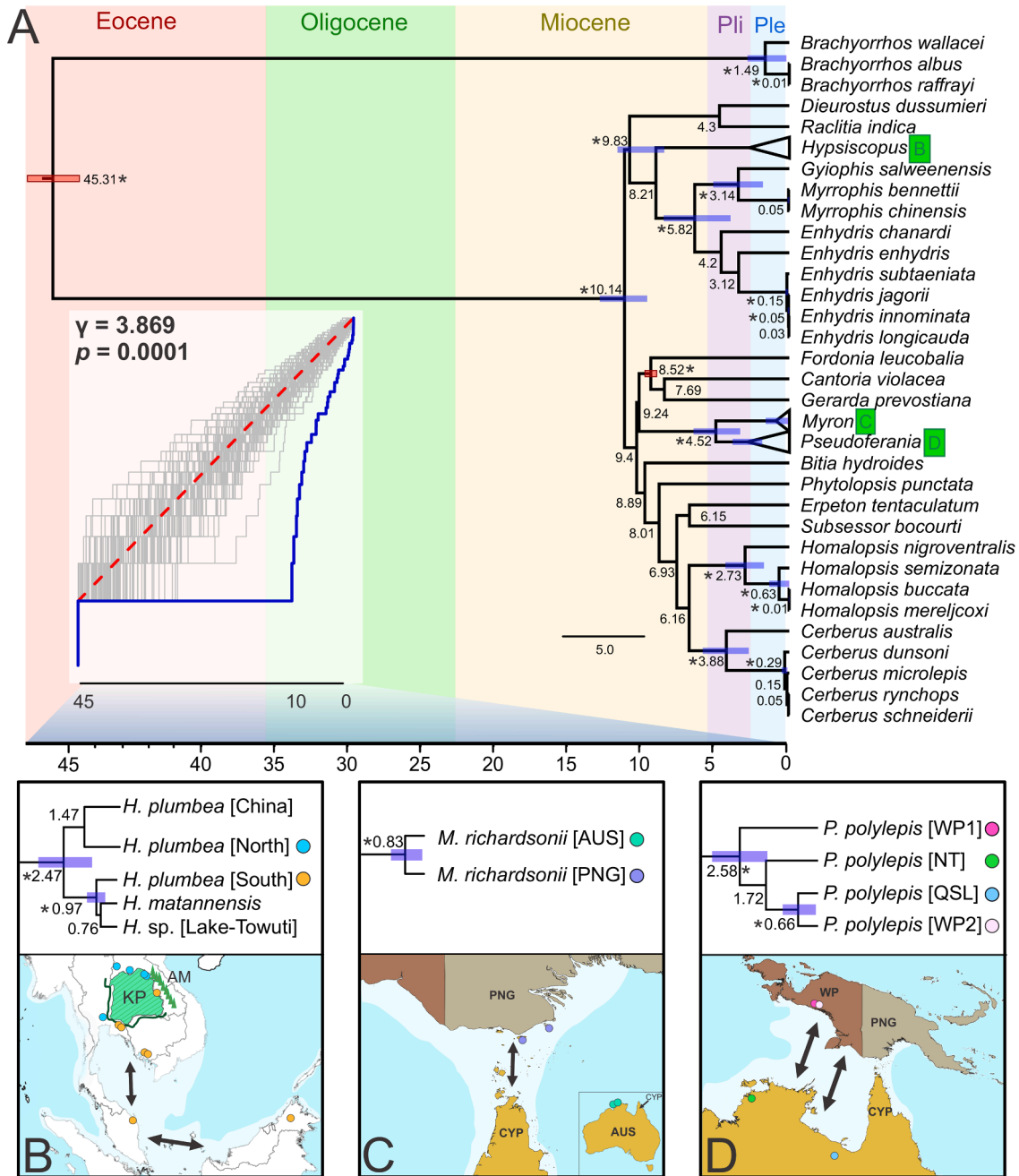


Fig. 3. (A) Chronogram of the Homalopsidae. Numbers and node bars represent mean ages of estimated divergence times and the respective 95% HPD. Asterisks (*) next to dates represent PP \geq 0.95 for the respective divergence. Bars for calibrated nodes using [Burbrink et al. \(2020\)](#) are colored red. Lineage-through time plots (grey = 100 pure birth simulations; red = log lineages under pure birth model; blue = birth-death model) inset with γ statistic and associated p -value. (B–D) Expanded clades from panel A for *Hypsiscopus*, *Myron*, and *Pseudoferania*. Colored dots indicate sample locations of respective taxa; black arrows show possible dispersal routes over areas of land bridges within the past 2.5 million to 17,000 years (light blue; [Voris, 2000](#)). KP = Khorat Plateau (green polygon, thick lines show raised elevation of western and southern ridges); AM = Annamite Mountain Range (green triangles); PNG = Papua New Guinea; AUS = Australia; CYP = Cape York Peninsula; WP = West Papua, Indonesia. Maps created using QGIS and downloaded shape files from [diva-gis.org](#). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

to intrageneric relationships and species limits. Our results indicate that snakes of the family Homalopsidae are in need of both broad- and fine-scale investigation with respect to species diversity, and their taxonomic placement within supra-specific higher taxa. The early branching order of our estimates and, thus, many of the intergeneric relationships of homalopsids, remain unresolved (Figs. 2, 3). However, we do find several strongly-supported sister-taxon relationships (Fig. 2, 3), which are consistent with previous studies (e.g., *Brachyorrhos*, [Murphy et al., 2011](#); *Cerberus* relationships, [Murphy et al., 2012a](#); a crustacean-eating

clade, [Alfaro et al., 2008](#)). Additionally, results indicate many currently recognized species are likely synonymous, whereas others may harbor undescribed taxa. Although our trees are based on just two loci, mtDNA-based trees have yielded significant evolutionary and natural history findings at the species and population levels ([Funk and Omland, 2003](#); [Rubinoff and Holland, 2005](#); [Burbrink and Ruane, 2021](#)); despite low coverage for both genes, our additional taxonomic sampling and advanced methods, compared to previous studies, provide evolutionary insights into homalopsid diversification and related taxonomy, as

discussed below (Wiens, 2003; Wiens and Morrill, 2011; Zheng and Wiens, 2015).

4.1. Systematic implications for Homalopsidae

4.1.1. Intergeneric relationships and the inclusion of novel taxa

Our resulting trees corroborate many of the lineages and relationships found in previous homalopsid phylogenies (e.g., Alfaro et al., 2004; Alfaro et al., 2008). Novel relationships among and within genera were also recovered, when compared to studies that only used mtDNA or with lower taxonomic and geographic sampling (Voris et al., 2002; Alfaro et al., 2008). The inclusion of *Cerberus dunsoni*, *Enhydris chanardi*, *Homalopsis semizonata*, *H. nigroventralis*, and *Myrrophis bennettii*, which have never been part of any phylogenetic analysis, support the monophyly of the respective genera (Figs. 2, 3). We also find alternative topologies to previous hypotheses of the placement of particular species. For example, the placement of *Racliitia indica* from Malaysia is hypothesized as sister to *Erpeton tentaculatum* from Thailand (Quah et al., 2018), but here we find *R. indica* as either sister to *Dieurostus dussumieri* or a clade containing *Cerberus*, *Homalopsis*, *Bitia*, and *Phytolopsis* (Figs. 2, 3A).

Studies on *Enhydris* (e.g., Karns et al., 2005, 2010; Lukoschek et al., 2011) have focused heavily on *E. jagorii*, *E. longicauda*, *E. innominata*, *E. enhydris*, and *E. subtaeniata*. The sixth species of the genus, *E. chanardi*, is a rare snake with an uncertain geographic range in Thailand (Murphy and Voris, 2014). We find support (concatenated tree BS = 98; species tree PP = 0.84) for *E. chanardi* as the sister to all other *Enhydris*. Although our specimens of *E. chanardi* (vouchers YPM 15033, 15037) are from the pet trade, and no other fresh tissues of this species were available for this study, an examination of the associated voucher skins of the tissues confirms their identity as *E. chanardi* based on scalation and color pattern (JMB, unpublished data). Additionally, our mPTP analysis supports the distinctiveness of *E. chanardi* from the other 29 *Enhydris* specimens included here. In contrast, our sampling of *H. mereljcoxi* and *H. nigroventralis* indicate that although *H. nigroventralis* is distinct, *H. mereljcoxi* is within *H. semizonata* (Fig. 2). Similarly, recognition of *Cerberus dunsoni* and *C. microlepis* renders *C. schneiderii* polyphyletic (Fig. 2).

4.1.2. Evidence of synonymy

The species richness of homalopsid snakes has increased since Gyi's (1970) morphology-based classification of the family (Murphy, 2007; Murphy and Voris, 2014), going from 34 species in 10 genera to the current 55 species in 29 genera. Although our study supports the recognition of many of these taxa, there are several questionably-distinct species (Table 1). We find some support from our distance calculations, concatenated tree, and mPTP species delimitation analysis, for the proposal that 13 species (among five genera) should be critically evaluated and possibly placed back in synonymy with their closely-related congeners. These results may indicate taxonomic inflation of species that have similar or overlapping diagnostic characters (See File S3 in Supplementary Material File for all comparisons, discussed below), many of which have geographically proximate, or even peripatric, distributions.

The low intraspecific genetic distances of *Brachyorrhos raffrayi* and *B. albus* indicates they may represent one species, and thorough investigation of the diagnostic characters of these understudied, semi-fossorial homalopsids is warranted. *Brachyorrhos raffrayi* and *B. albus* are both fangless homalopsids with allopatric distributions on the Indonesian islands of Ternate and Seram (and satellite islands), respectively. However, the known herpetofauna of island localities between *B. raffrayi* and *B. albus* are not as thoroughly investigated, and there are reports of a population of *B. albus* established on Pulau Bisa, off Obi Island (Murphy et al., 2012b; O'Shea, 2018), which would make the closest population *B. albus* 49 km away from the Bacan Islands, the northernmost which is Ternate and the known locality of *B. raffrayi*.

Although *B. raffrayi* has not been reported from the rest of the Bacan Islands, undiscovered populations may exist there. Additionally, these species have similar and overlapping scale morphologies (e.g., subcaudals [males], ventrals; Murphy et al., 2012b), which are commonly used as morphological diagnostic characters for snakes. Our results suggest that the phylogenetic relationships observed may be explained by a geographically expanded *B. albus*, and that putting *B. raffrayi* in synonymy with the former may be advisable.

Taxonomic inflation is also of concern in *Homalopsis*, *Cerberus*, *Myrrophis*, and *Enhydris*. The clade consisting of *Homalopsis buccata*, *H. mereljcoxi*, and *H. semizonata*, (Fig. 2) include Thailand *H. buccata* north of their known distribution (Murphy et al., 2012c; Murphy and Voris, 2014). These species may be difficult to accurately identify in the field and an examination of the *H. buccata* from northern Myanmar is needed to identify the species boundaries between this species and its congeners. *Homalopsis mereljcoxi* and *H. semizonata*, however, have proximate ranges in Indochina, and both are found along the Malay Peninsula, south of the Isthmus of Kra (Pauwels and Sumontha, 2016). Meristic characters (e.g., ventrals, subcaudals) overlap between these species, and some of the diagnostic characters, such as unique three-way fragmentation of the prefrontal head plates, have been shown to be variable and similar to other *Homalopsis* species (Pauwels and Sumontha, 2016).

Another set of species with problematic taxonomic and biogeographic implications is the placement of *Cerberus dunsoni* and *C. microlepis* within *C. schneiderii*. *Cerberus microlepis* and *C. dunsoni* differ from *C. schneiderii*, respectively, by a higher number of dorsal scale rows and plate-like parietal scale fragments on the head (Murphy et al., 2012a; Murphy and Voris, 2014). However, much of their ranges in scale counts and their color pattern overlap with *C. schneiderii*, which exhibits greater morphological variation than once thought (Barrera Jr. et al., 2017). The distribution of *C. microlepis* is restricted to Lake Bui in the Philippines, which is geographically located within the range of *C. schneiderii*; the endemic *C. dunsoni* is restricted to the Palau islands of Micronesia). It has been hypothesized that *C. microlepis* may be a freshwater-adapted form of *C. schneiderii* after a population became isolated to the lake when an earthquake caused the adjacent Mt. Iriga to form a natural dam a few hundred years ago (Alfaro et al., 2004). *Cerberus dunsoni*, which is 870 km away from its congeners, may represent a population of *C. schneiderii* that made successive colonizations across the Kyushu-Palau Ridge, a volcanic island chain that extended north of New Guinea (Allison, 1996). Such interpretation contrasts with current taxonomy and the hypothesis that *C. dunsoni* is independent from *C. schneiderii* (Murphy et al., 2012a). Nevertheless, lake and island populations are often distinctly grouped in our analyses (Table 1; Figs. 2, 3), suggesting some level of isolation. A detailed study focusing on these populations and including explicit gene-flow analysis is necessary to determine the degree of separation amongst these taxa.

The two known species of *Myrrophis*, both of which are found in southern China, are delimited by distinct numbers of several scale characters and color pattern, as well as habitat type; *M. chinensis* is known from freshwater habitats, whereas *M. bennettii* is known from brackish and marine waters (Murphy, 2007; Karns et al., 2010; Kumar et al., 2012). The grouping of these species in our analyses may represent greater variability in a single, wide-spread species and subsequent examination of the original voucher specimens would help to confirm these identifications. Similarly, the *Enhydris longicauda* + *E. innominata* + *E. jagorii* group is only known from Cambodia, Vietnam (and possibly Cambodia), and Thailand, respectively (Murphy and Voris, 2014). It is uncertain if these species are sympatric with one another, but their morphologies are similar in their number of dorsal scale rows and subcaudals. The relationship reported here has been recovered in other studies on *Enhydris* (Karns et al., 2010). We provide additional support for this potential synonymy in this genus with specific species delimitation analyses and increased sampling.

All of these cases support scenarios of synonymy for species in

Homalopsidae, which require greater sampling both for individual snakes as well as loci, ideally at the genomic level in conjunction with detailed morphological examination for the taxa in question. Thus, we remain conservative and do not suggest any taxonomic changes until future genomic analyses are conducted. The interspecific pairwise cyt-b genetic distances for most of these possibly synonymous species are much smaller (e.g., *Homalopsis*, *Enhydryis*; Fig. 2) than the interspecific distances for species considered distinct in other recent snake studies that calculated mtDNA distances (e.g., Ruane et al., 2018; Keates et al., 2019). Though changes in taxonomy ideally will include increased sample sizes and more loci than presented here, these results suggest potential cases of taxonomic inflation within several homalopsid genera.

4.1.3. Undescribed diversity and intraspecific biogeography

In contrast to evidence of synonymy in some taxa, we also find several instances of undescribed homalopsid lineages. There is strong support for a north-south split between populations of *Hypsiscopus plumbea* from northern Thailand + Laos and those from south-central Thailand + Cambodia + Malaysia. This divergence may be the result of two related geographic events in Thailand during the Quaternary. The Khorat Plateau (Khorat Basin; 180,000 km²), separating northern and southern clades of *H. plumbea*, in northeastern Thailand formed after Quaternary tectonic uplifting and tilting occurred alongside its western and southern edges (Hutchison, 1989; Rainboth, 1996). This uplift led to a river catchment event in which the Mekong River, originally flowing south towards the Gulf of Thailand (currently the Chao Phraya river plain; Carbonnel 1965; Workman, 1977), shifted to its current position running west to east, and then south through the now-Thailand-Laos border and Cambodia (Rainboth, 1966; Fontaine and Workman, 1978). During the mid-Quaternary, the Khorat Plateau had a heterogeneous landscape of river valleys and mountains that have shaped the local biodiversity (Fontaine and Workman, 1978; Hutchison, 1989). With respect to the mountains on the Khorat Plateau's western and southern margins, shifting of major aquatic riverways (e.g., the Mekong River), and the Annamite mountain range to the east of the Khorat Plateau, our divergence dating results indicate that tectonic uplift, the heterogeneous landscape, and secondary river catchment events may have ultimately acted as barriers to gene flow between northern and southern *H. plumbea* (Fig. 3B), as has been demonstrated in gastropods, fish, and other homalopsids (Glaubrecht and Köhler, 2004; Lukoschek et al., 2011; Adamson et al., 2012). Such divisions may also be supported by the phylogenetic placement of a *H. plumbea* specimen from Bangkok (FMNH 250124) with the northern specimens (Fig. 2), despite being geographically proximate to the samples in the southern clade; this likely reflects the river catchment, where some of the population remained in the Mekong (now in northern Thailand) while other parts of the population with a similar haplotype were isolated to the Chao Phraya near Bangkok. Although some of the mountains surrounding the Khorat Plateau are not particularly high in elevation, they may still be a sufficient barrier between homalopsid populations, possibly due to their aquatic nature, as evidenced by a significant decrease in homalopsid abundance at Khorat Plateau rim sites (Karns et al., 2005). The specimen of the southern clade from Malaysian Borneo also suggests dispersal via land bridges in the Pleistocene that connected peninsular Malaysia to Borneo (Woodruff, 2010). Additionally, a potentially new species of *H. plumbea* from Guangxi, China is supported as sister to its conspecifics in northern Thailand and Laos. This locality in China is over 500 km from the other *H. plumbea* in our sampling; Guangxi, being located within one of the most biodiverse regions on the planet, may harbor undescribed species, similar to many other vertebrate groups from that region (Luo et al., 2016).

Our species delimitation analysis provides support for previous studies (Alfaro et al., 2008; Murphy and Voris, 2014) that suggest that the Sulawesi Lake Towuti *Hypsiscopus* specimen is distinct from all other *Hypsiscopus* species. The specimens of *Hypsiscopus matannensis* between South and Southeast Sulawesi may also represent potentially new

species, a result recovered in unrelated vertebrates that have population structure between these and other regions of the mainland (e.g., Evans et al., 2003; McGuire et al., 2007; Burton and Nietsch, 2010). Sulawesi is a composite island, in which multiple landmasses collided together, and then subsequent Pleistocene sea-level fluctuations connected and disconnected different regions (Hamilton, 1979; Hall, 1996, 1998; Moss and Wilson, 1998; Nugraha and Hall, 2018). Faunal boundaries such as the Tempe Depression in South Sulawesi and the suture (tectonic) line of Southeast Sulawesi, the latter of which is near Lake Towuti, have been proposed as significant biogeographic transition zones, which may act as barriers to gene flow (Fooden, 1969; Hall, 1998; Evans et al., 2003; Burton and Nietsch, 2010).

The trees and mPTP analysis indicate undescribed diversity in the Australasian *Myron richardsonii* and *Pseudoferania polylepis* (Table 1; Figs. 2, 3). *Myron richardsonii* specimens from localities that are (geographically) from PNG are supported as the sister taxon to the specimens from Australia. Pleistocene sea-level fluctuations have repeatedly connected and disconnected major land masses and coastlines on the Sunda and Sahul Shelves (Voris, 2000; Woodruff, 2010), resulting in distinct genetic signatures in extant taxa (Hall, 1998; Hewitt, 2000). It is likely that *M. richardsonii* was a trans-Torresian species that dispersed between Australia and PNG during periods of inter-landmass activity in the Pleistocene (Fig. 3C), which is plausible given the shallow waters of the Torres Strait and these snakes' aquatic (estuarine/marine) nature. These patterns are not limited to our example here, but have also been shown in mammals (Aplin et al., 1993) and other snakes (e.g., elapids; Wüster et al., 2005). The phylogenetic and delimitation analyses of *Pseudoferania polylepis*, another homalopsid with an Australo-Papuan distribution, suggest four highly divergent specimens between West Papua, Indonesia, and Australia (Fig. 3D). Similar to biogeographic scenarios of the Torres Strait, the land bridge formed between Australia and PNG spanned the Arafura Sea, which is located between our *P. polylepis* specimens (Fig. 3D). Although more specimens and loci are required, multiple dispersal events may have produced the relationships observed (Fig. 3D).

These biogeographic hypotheses may explain the divergences in these taxa, though other genera included here that showed evidence of undescribed diversity (i.e., *Subsessor bocourti* and *Phytolopsis punctata*; Table 1) only included one or two specimens from each locality. More specimens and geographic sampling are needed to determine if these splits represent undescribed species or are artifacts in our analyses.

4.2. Evolution of the Homalopsidae

We present the first dating analysis specifically for the family Homalopsidae, with the inclusion of the fangless homalopsids and highest taxonomic coverage to date. These dates are likely to change with the addition of more taxa and loci, and so we suggest our biogeographic interpretations for Homalopsidae as a starting point for future studies. Our species tree indicates that the fangless *Brachyorrhos* inhabiting eastern Indonesia (Maluku Islands) and the rear-fanged group in Southeast Asia, Australia, and New Guinea diverged 45.31 ± 1.63 mya (Fig. 3). Homalopsids may have a mainland Southeast Asian origin, with subsequent expansion westward into South Asia and eastward towards the Greater and Lesser Sunda Islands, Wallacea, the Philippines, Palau, New Guinea, and Australia. An origin in mainland Southeast Asia is also supported by the observation that ~49% of homalopsid species are distributed throughout Indochina (and adjacent China), with the rest known from isolated regions or islands east or west of Indochina.

Dispersal events have likely been the result of the changing paleogeography that occurred through most of these regions, even across regions that are considered strong faunal barriers (e.g., *Hypsiscopus* crossing Wallace's Line; Alfaro et al., 2008; Esselstyn et al., 2010; Brown et al., 2013). We also find evidence for recent founder events, such as *Cerberus microlepis* of Lake Buhi, *C. dunsoni* of Palau, and the Timor population of *C. schneiderii*, all of which may actually be populations of

the phenotypically similar *C. schneiderii*. Our data suggests that dispersal into isolated habitats such as lakes and islands can be followed by inhibited gene flow from the surrounding populations.

Perhaps the most striking of our results is that of the timing of the initial split between the fangless and fanged homalopsids. The time period between the diversification of crown homalopsids ($\sim 45.31 \pm 1.63$ mya, Fig. 3) coincides with the timing of the Australian plate moving northward through the Pacific, away from Antarctica (Hall, 2009). The Australian plate contains the landmasses of Australia, New Zealand, and New Guinea, including the Bird's Head peninsula and eastern Indonesian Islands that harbor *Brachyorrhos*. The divergence date estimation (Fig. 3) indicates that the fangless clade was already present before any known connections to facilitate dispersal between Southeast Asia and Australia. Although these topologies are broadly consistent with previous studies inferring rear-fanged homalopsid biogeography (e.g., Alfaro et al., 2008), divergence date estimates of both fanged and fangless homalopsids greatly changes the inferred biogeographic implications on the evolution of this family.

The young ages between species of *Brachyorrhos wallacea* and *B. albus* + *B. raffrayi* (~ 10 kya–1.5 mya) may reflect the geologic history of some islands to which these species are seemingly restricted to; the Banda Arc, including Timor and Seram, are very young (~ 2 million years old; Hall, 2009; Lohman et al., 2011). Although some of the larger islands, such as Halmahera, are geologically dated no older than 11 my, volcanic activity of Halmahera ceased in the last 2 million years (Baker and Malaihollo, 1996; Hall, 2009). Indeed, our topology and divergence dates show a long period of ~ 35 million years in which there was no divergence amongst homalopsids, which may be due to missing nuclear data for *Brachyorrhos* vs. reflecting reality. The lack of a slowly-evolving nuclear gene and reliance on a fast-evolving mitochondrial gene may have resulted in the large temporal gap between the fangless and rear-fanged homalopsids in the species tree (Fig. 3). Alternatively, the temporal gap may represent missing data from real biological units (i.e., unsampled extant and/or extinct taxa). Halmahera formed due to ongoing arc collisions in the Molucca Sea, and in the next 5 million years will likely submerge below the ocean's surface with no subsurface traces of the arc itself (Hall, 2000). If other island arcs have also disappeared from the ocean's surface, as has been hypothesized (Hall, 2000), then the subsidence of island arcs may be correlated with the extinction of their flora and fauna. As such, if extinction has occurred in Wallacean island arcs, this can cause an artifactual gap (in our case, ~ 35 my long) in the chronogram and inflate the Pybus and Harvey's γ statistic that suggested early-slow and recent-rapid diversification (i.e., the Pull of the Present; Ricklefs, 2007).

5. Conclusion

Our study provides new evolutionary insights on homalopsid snakes using the largest and most comprehensively sampled phylogeny of the family to date. Although this dataset is limited to two loci and primarily informed by mtDNA, our primary conclusions, with regard to evolutionary relationships and biogeographic patterns, are observed using either locus. Furthermore, mtDNA is a heritable unit and provides evidence of evolutionary and biogeographic phenomena (Rubinoff and Holland, 2005; Burbrink and Ruane, 2021). Previous studies on snakes (Burbrink et al., 2000, 2021), lizards (Grismer et al., 2016), salamanders (Steinfartz et al., 2000; Vences et al., 2014; Rodríguez et al., 2017), birds (Lamichhaney et al., 2015), bats (Stadelmann et al., 2007; Morales et al., 2019), and eutherian mammals (Liu and Miyamoto, 1999; Song et al., 2012) have found that initial phylogenetic reconstructions using single or few mitochondrial and/or nuclear loci often find the same lineages, sister taxa, and/or species groups when reanalyzed using more comprehensive molecular sampling. Our increased taxonomic and biogeographic sampling will be critical for accurately revisiting the evolution of organismal, geographical, and ecological traits in this group (e.g., habitat preference, salt tolerance [Kumar et al., 2012], diet and

feeding behavior [Fabre et al., 2016; Jayne et al., 2018], biogeography [Alfaro et al., 2008]). Finally, increasing current knowledge of homalopsid diversity is crucial to conservation. Although mud snakes are a critical component to Southeast Asian ecosystems and contribute a large portion of resident vertebrate biomass (Murphy, 2007), populations are likely in danger of extirpation or extinction. For example, ~ 3.8 million homalopsid snakes harvested from the wild, yearly, in Tonle Sap, Cambodia (Brooks et al., 2007).

With the recent success of studies incorporating fluid-preserved specimens into systematic datasets (e.g., Ruane and Austin, 2017), our study and understanding of Homalopsidae can be greatly enhanced with the addition of species and populations only known from museum specimens and historical records in the literature, such as the other fangless homalopsids *Karnsophis* and *Calamophis*, as well as several rear-fanged genera. During the course of publishing this study, a new genus and species of homalopsid, endemic to Myanmar and sister to *Gyiophis*, was described (*Myanophis thanlyinensis*; Köhler et al., 2021), emphasizing that the diversity and evolution of this group is still far from being recognized. The use of genomic datasets (e.g. SNPs, target capture; Leaché and Oaks, 2017; Barrow et al., 2018) and the publication of draft genomes (Köhler et al., 2021) will likely provide a more accurate interpretation of the evolution, biogeography and historic demography of this group. Additionally, increasing our geographic sampling will be crucial to elucidating the polarity of gene flow and dispersal, and provide genome-scale insights, which will improve the accuracy of species delimitation analyses (Chan et al., 2020). Although increased loci and taxon sampling will be forthcoming from both our own and other studies in progress, the analyses presented here provide new data (taxon and gene sampling), a more comprehensive phylogenetic inference, and novel evolutionary hypotheses for the continued study of this highly unique, widespread—and yet poorly known—family of aquatic caenophidian snakes.

CRedit authorship contribution statement

Justin M. Bernstein: Conceptualization, Methodology, Software, Formal analysis, Investigation, Writing - original draft, Visualization. **John C. Murphy:** Conceptualization, Validation, Resources, Supervision. **Harold K. Voris:** Conceptualization, Validation, Resources, Supervision. **Rafe M. Brown:** Methodology, Validation, Resources, Supervision. **Sara Ruane:** Conceptualization, Methodology, Validation, Resources, Supervision, Project administration, Funding acquisition.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We acknowledge the following individuals and associated institutions for providing tissues to be used for DNA sequencing in this study: Frank Burbrink, David Kizirian, and Lauren Vonnahme (American Museum of Natural History); Molly Hagemann (Bernice P. Bishop Museum); Lauren Scheinberg and Erica Ely (California Academy of Sciences); Alan Resetar and Joshua Mata (Field Museum of Natural History); Lee Grismer (La Sierra University Herpetology Collection); Christopher Austin and Jackson Roberts (Louisiana State University Museum of Natural Science); Gavin Dally (Museum and Art Gallery of the Northern Territory); Jimmy McGuire and Carol Spencer (Museum of Vertebrate Zoology); Bryan Stuart (North Carolina Museum of Natural Sciences); Patrick Couper and Andrew Amey (Queensland Museum); Bob Murphy (Royal Ontario Museum); Sally South (South Australian Museum); Addison Wynn (National Museum of Natural History); Paul Doughty and Rebecca Bray (Western Australian Museum); Gregory

Watkins-Colwell (Yale Peabody Museum of Natural History). We also thank Hinrich Kaiser for providing tissues of *Cerberus schneiderii* from Timor through the National Museum of Natural History, and David Kizirian and Lauren Vonnahme for assisting in the importation of tissues from the South Australian Museum and Western Australian Museum. We also thank Frank Burbrink for assisting in providing the code and analysis for the divergence time estimation. We thank the Philippine Department of the Environment and Natural Resources, Biodiversity Management Bureau, for logistical support and for overseeing drafts of project Memoranda of Agreement, as well as issuing research, collecting, and export permits necessary for this and related studies. We also thank the KU Institutional Animal Care and Use Committee for approving animal handling protocols (KU IACUC AUS 185-05 to RMB). Finally, we thank Evan Quah for providing photographs of *Raclinia indica* and *Gyophis salweenensis*.

Funding

Field work was supported by funds from NSF DEB 0743491 and 1654388 to RMB, and NSF DEB 1926772 to SR. This work was also supported by funds from Rutgers University-Newark granted to the Ruane Lab.

Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jmpev.2021.107109>.

References

- Adamson, E.A., Hurwood, D.A., Mather, P.B., 2012. Insights into historical drainage evolution based on the phylogeography of the chevron snakehead fish (*Channa striata*) in the Mekong Basin. *Freshw. Biol.* 57 (11), 2211–2229. <https://doi.org/10.1111/j.1365-2427.2012.02864.x>.
- Alencar, L.R., Quental, T.B., Graziotin, F.G., Alfaro, M.L., Martins, M., Venzon, M., Zaher, H., 2016. Diversification in vipers: phylogenetic relationships, time of divergence and shifts in speciation rates. *Mol. Phylogenet. Evol.* 105, 50–62. <https://doi.org/10.1016/j.jmpev.2016.07.029>.
- Alfaro, M.E., Arnold, S.J., 2001. Molecular systematics and evolution of *Regina* and the thamnophiine snakes. *Mol. Phylogenet. Evol.* 21 (3), 408–423. <https://doi.org/10.1006/mpev.2001.1024>.
- Alfaro, M.E., Karns, D.R., Voris, H.K., Abernathy, E., Sellins, S.L., 2004. Phylogeny of *Cerberus* (Serpentes: Homalopsinae) and phylogeography of *Cerberus rynchops*: diversification of a coastal marine snake in Southeast Asia. *J. Biogeogr.* 31 (8), 1277–1292. <https://doi.org/10.1111/j.1365-2699.2004.01114.x>.
- Alfaro, M.E., Karns, D.R., Voris, H.K., Brock, C.D., Stuart, B.L., 2008. Phylogeny, evolutionary history, and biogeography of Oriental-Australian rear-fanged water snakes (Colubroidea: Homalopsidae) inferred from mitochondrial and nuclear DNA sequences. *Mol. Phylogenet. Evol.* 46 (2), 576–593. <https://doi.org/10.1016/j.jmpev.2007.10.024>.
- Allison, A., 1996. Zoogeography of amphibians and reptiles of New Guinea and the Pacific region. In: Keast, E., Miller, S.E. (Eds.), *The Origin and Evolution of Pacific ISLAND BIOTAS, New Guinea to Eastern Polynesia: Patterns and Processes*. SPB Academic Publishing, Amsterdam, pp. 407–436.
- Aplin, K.P., Baverstock, P.R., Donnellan, S.C., 1993. Albumin immunological evidence for the time and mode of origin of the New Guinean terrestrial mammal fauna. *Sci. New Guinea* 19 (3), 131–145.
- Arrigoni, R., Berumen, M.L., Chen, C.A., Terraneo, T.I., Baird, A.H., Payri, C., Benzoni, F., 2016. Species delimitation in the reef coral genera *Echinophyllia* and *Oxypora* (Scleractinia, Lobophylliidae) with a description of two new species. *Mol. Phylogenet. Evol.* 105, 146–159. <https://doi.org/10.1016/j.jmpev.2016.08.023>.
- Baker, S., Malaihollo, J., 1996. Dating of Neogene igneous rocks in the Halmahera region: arc initiation and development. *Geol. Soc. Lond. Spec. Publ.* 106 (1), 499–509. <https://doi.org/10.1144/GSL.SP.1996.106.01.31>.
- Barrera, W.B., Jaluague, J.C.A., Albano, S.D.C., Jalbuena, G.L.L., 2017. New distributional record and intra-specific variation of *Cerberus schneiderii* in Iyam River, Lucena City, Quezon, Philippines. *Philippine J. Syst. Biol.* 11 (1), 29–36.
- Barrow, L.N., Lemmon, A.R., Lemmon, E.M., 2018. Targeted sampling and target capture: assessing phylogeographic concordance with genome-wide data. *Syst. Biol.* 67 (6), 979–996. <https://doi.org/10.1093/sysbio/syy021>.
- Bergmann, P.J., Morinaga, G., Freitas, E.S., Irschick, D.J., Wagner, G.P., Siler, C.D., 2020. Locomotion and palaeoclimate explain the re-evolution of quadrupedal body form in *Brachymeles* lizards. *Proc. Royal Soc. B* 287 (1938), 20201994. <https://doi.org/10.1098/rspb.2020.1994>.
- Bonaparte, C.L.P., 1845. Specchio generale dei sistemi erpetologico, anfibiologico ed ittiologico. *Atti Riunione degli Scienziati Italiani* 6, 376–378.
- Boulenger, G.A., 1890. *The Fauna of British India, Including Ceylon and Burma. Reptilia and Batrachia*. Taylor & Francis, London.
- Boulenger, G.A., 1896. *Catalogue of the Snakes in the British Museum (Natural History)*, vol. 3. London, Trustees of the British Museum, <https://doi.org/10.5962/bhl.title.8316>.
- Bouckaert, R.R., Drummond, A.J., 2017. bModelTest: Bayesian phylogenetic site model averaging and model comparison. *BMC Evol. Biol.* 17, 42. <https://doi.org/10.1186/s12862-017-0890-6>.
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M.A., Rambaut, A., Drummond, A.J., 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. *PLoS Comput. Biol.* 10 (4), e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>.
- Brooks S.E., Reynolds, J.D., Allison, E.A., Touch, B., 2007. The exploitation of homalopsid water snakes at Tonle Sap Lake, Cambodia. In: Murphy, J.C. (Ed.), *Homalopsid Snakes. Evolution in the Mud*. Kreiger Publishing Company, Florida, pp. 31–37.
- Brown, R.M., Siler, C.D., Grismer, L.L., Das, I., McGuire, J.A., 2012. Phylogeny and cryptic diversification in Southeast Asian flying geckos. *Mol. Phylogenet. Evol.* 65 (2), 351–361. <https://doi.org/10.1016/j.jmpev.2012.06.009>.
- Brown, R.M., Siler, C.D., Oliveros, C.H., Esselstyn, J.A., Diesmos, A.C., Hosner, P.A., Linkem, C.W., Barley, A.J., Oaks, J.R., Sanguilu, M.B., Welton, L.J., Blackburn, D.C., Moyle, R.G., Townsend Peterson, A., Alcalá, A.C., 2013. Evolutionary processes of diversification in a model island archipelago. *Annu. Rev. Ecol. Syst.* 44, 411–435. <https://doi.org/10.1146/annurev-ecolsys-110411-160323>.
- Burbrink, F.T., Gehara, M., 2018. The biogeography of deep time phylogenetic reticulation. *Syst. Biol.* 67 (5), 743–755. <https://doi.org/10.1093/sysbio/syy019>.
- Burbrink, F.T., Graziotin, F.G., Pyron, R.A., Cundall, D., Donnellan, S., Irish, F., Keogh, J.S., Kraus, F., Murphy, R.W., Raxworthy, C.J., Ruane, S., Lemmon, A.R., Lemmon, E.M., Zaher, H., 2020. Interrogating genomic-scale data for Squamata (lizards, snakes, and amphisbaenians) shows no support for key traditional morphological relationships. *Syst. Biol.* 69 (3), 502–520. <https://doi.org/10.1093/sysbio/syz062>.
- Burbrink, F., Gehara, M., Myers, E.A., 2021. Resolving spatial complexities of hybridization in the context of the gray zone of speciation in North American ratsnakes (*Pantherophis obsoletus* complex). *bioRxiv*, <https://doi.org/10.1101/2020.05.05.079467>.
- Burbrink, F.T., Lawson, R., Slowinski, J.B., 2000. Mitochondrial DNA phylogeography of the polytypic North American rat snake (*Elaphe obsoleta*): a critique of the subspecies concept. *Evolution* 54 (6), 2107–2118. [https://doi.org/10.1554/0014-3820\(2000\)054\[2107:mdpotp\]2.0.co;2](https://doi.org/10.1554/0014-3820(2000)054[2107:mdpotp]2.0.co;2).
- Burbrink, F.T., Ruane, S., 2021. Contemporary philosophy and methods for species delimitations. *Ichthyol. Herpetol.* In press.
- Burton, J.A., Nietsch, A., 2010. Geographical variation in duet songs of Sulawesi tarsiers: evidence for new cryptic species in south and southeast Sulawesi. *Int. J. Primatol.* 31 (6), 1123–1146. <https://doi.org/10.1007/s10764-010-9449-8>.
- Carbonnel, J.P., 1965. Essai d'interprétation morphotectonique de la cuvette Cambodgienne. *Revue de Géographie Physique et de Géologie Dynamique* 7, 277–281.
- Chan, K.O., Hutter, C.R., Wood Jr, P.L., Grismer, L.L., Das, I., Brown, R.M., 2020. Gene flow creates a mirage of cryptic species in a Southeast Asian spotted stream frog complex. *Mol. Ecol.* 29 (20), 3970–3987. <https://doi.org/10.1111/mec.15603>.
- Dufresnes, C., Pribille, M., Alard, B., Gonçalves, H., Amat, F., Crochet, P.A., Dubey, S., Perrin, N., Fumagalli, L., Vences, M., Martínez-Solano, I., 2020. Integrating hybrid zone analyses in species delimitation: lessons from two anuran radiations of the Western Mediterranean. *Heredity* 124 (3), 423–438. <https://doi.org/10.1038/s41437-020-0294-z>.
- Esselstyn, J.A., Oliveros, C.H., Moyle, R.G., Peterson, A.T., McGuire, J.A., Brown, R.M., 2010. Integrating phylogenetic and taxonomic evidence illuminates complex biogeographic patterns along Huxley's modification of Wallace's Line. *J. Biogeogr.* 37 (11), 2054–2066. <https://doi.org/10.1111/j.1365-2699.2010.02378.x>.
- Evans, B.J., Supriatna, J., Andayani, N., Setiadi, M.I., Cannatella, D.C., Melnick, D.J., 2003. Monkeys and toads define areas of endemism on Sulawesi. *Evolution* 57 (6), 1436–1443. <https://doi.org/10.1111/j.0014-3820.2003.tb00350.x>.
- Fabre, A.C., Bickford, D., Segall, M., Herrel, A., 2016. The impact of diet, habitat use, and behaviour on head shape evolution in homalopsid snakes. *Biol. J. Linn. Soc.* 118 (3), 634–647. <https://doi.org/10.1111/bj.12753>.
- Fooden, J., 1969. *Taxonomy and evolution of monkeys of Celebes (Primates: Cercopithecidae)*. Bibliotheca Primatologica, 10. Karger, New York.
- Fontaine, H., Workman, D.R., 1978. Review of the geology and mineral resources of Kampuchea, Laos and Vietnam. In: *Third Regional Conference on Geology and Mineral Resources of Southeast Asia, Bangkok, Thailand*, pp. 541–603.
- Figuerola, A., McKelvey, A.D., Grismer, L.L., Bell, C.D., Lailvaux, S.P., 2016. A species-level phylogeny of extant snakes with description of a new colubrid subfamily and genus. *PLoS One* 11 (9). <https://doi.org/10.1371/journal.pone.0161070>.
- Funk, D.J., Omland, K.E., 2003. Species-level paraphyly and polyphyly: frequency, causes, and consequences, with insights from animal mitochondrial DNA. *Annu. Rev. Ecol. Syst.* 34 (1), 397–423. <https://doi.org/10.1146/annurev.ecolsys.34.011802.132421>.
- Gamble, T., Greenbaum, E., Jackman, T.R., Bauer, A.M., 2015. Into the light: diurnality has evolved multiple times in geckos. *Biol. J. Linn. Soc.* 115 (4), 896–910. <https://doi.org/10.1111/bj.12536>.
- Geneious. Version 7.1.8. <https://www.geneious.com> (accessed 20 July 2020).
- Gray, J.E., 1849. *Catalogue of the Specimens of Snakes in the Collection of the British Museum*. London, Printed by Order of the Trustees, British Museum. <https://doi.org/10.5962/bhl.title.5503>.

- Glaubrecht, M., Köhler, F., 2004. Radiating in a river: systematics, molecular genetics and morphological differentiation of viviparous freshwater gastropods endemic to the Kaek River, central Thailand (Cerithioidea, Pachychilidae). *Biol. J. Linn. Soc.* 82 (3), 275–311. <https://doi.org/10.1111/j.1095-8312.2004.00361.x>.
- Greene, H.W., 1997. *Snakes: The Evolution of Mystery in Nature*. University of California Press, Berkeley.
- Grismer, J.L., Schulte, J.A., Alexander, A., Wagner, P., Travers, S.L., Buehler, M.D., Welton, L.J., Brown, R.M., 2016. The Eurasian invasion: phylogenomic data reveal multiple Southeast Asian origins for Indian Dragon Lizards. *BMC Evol. Biol.* 16 (1), 43. <https://doi.org/10.1186/s12862-016-0611-6>.
- Gyi, K.K., 1970. A revision of colubrid snakes of the sub-family Homalopsinae. University of Kansas Publications, Museum of Natural History, vol. 20, no. 2, pp. 44–223.
- Hamilton, W.B., 1979. Tectonics of the Indonesian Region, vol. 1078. Washington, D.C., US Government Printing Office, <https://doi.org/10.3133/pp1078>.
- Hall, R., 1996. Reconstructing Cenozoic SE Asia. In: Hall, R., Blundell, D. (Eds.), *Tectonic Evolution of Southeast Asia*. Geological Society of London, London, pp. 153–184.
- Hall, R., 1998. The plate tectonics of Cenozoic SE Asia and the distribution of land and sea. In: Hall, R., Holloway, J.D. (Eds.), *Biogeography and Geological Evolution of SE Asia*. Backhuys Publishers, Leiden, pp. 99–131.
- Hall, R., 2000. Neogene history of collision in the Halmahera region, Indonesia. In: *Proceedings, Indonesian Petroleum Association, 27th Annual Convention*, pp. 487–493.
- Hall, R., 2009. Southeast Asia's changing palaeogeography. *Blumea-Biodiversity, Evolut. Biogeogr. Plants* 54 (1–2), 148–161. <https://doi.org/10.3767/000651909X475941>.
- Hedtke, S.M., Townsend, T.M., Hillis, D.M., 2006. Resolution of phylogenetic conflict in large data sets by increased taxon sampling. *Syst. Biol.* 55 (3), 522–529. <https://doi.org/10.1080/10635150600697358>.
- Hewitt, G., 2000. The genetic legacy of the Quaternary ice ages. *Nature* 405 (6789), 907–913. <https://doi.org/10.1038/35016000>.
- Hillis, D.M., Bull, J.J., 1993. An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Syst. Biol.* 42 (2), 182–192. <https://doi.org/10.1093/sysbio/42.2.182>.
- Hinckley, A., Hawkins, M.T., Achmadi, A.S., Maldonado, J.E., Leonard, J.A., 2020. Ancient divergence driven by geographic isolation and ecological adaptation in forest dependent Sundaland tree squirrels. *Front. Ecol. Evolut.* 8, 208. <https://doi.org/10.3389/fevo.2020.00208>.
- Hutchison, C.S., 1989. *Geological Evolution of South-east Asia*. Clarendon Press, Oxford, UK.
- Jan, G., 1863. *Elenco sistematico degli ofidi descritti e disegnati per l'iconografia generale*. A. Lombardi, Milano, Italy. <https://doi.org/10.5962/bhl.title.106683>.
- Jayne, B.C., Voris, H.K., Ng, P.K., 2018. How big is too big? Using crustacean-eating snakes (Homalopsidae) to test how anatomy and behaviour affect prey size and feeding performance. *Biol. J. Linn. Soc.* 123 (3), 636–650. <https://doi.org/10.1093/biolinnean/bly007>.
- Kapli, P., Lutteropp, S., Zhang, J., Kobert, K., Pavlidis, P., Stamatakis, A., Flouri, T., 2017. Multi-rate Poisson tree processes for single-locus species delimitation under maximum likelihood and Markov chain Monte Carlo. *Bioinformatics* 33 (11), 1630–1638. <https://doi.org/10.1093/bioinformatics/btx025>.
- Karns, D.R., Lukoschek, V., Osterhage, J., Murphy, J.C., Voris, H.K., 2010. Phylogeny and biogeography of the *Enhydris* clade (Serpentes: Homalopsidae). *Zootaxa* 2452 (1), 18–30. <https://doi.org/10.11646/zootaxa.2452.1.2>.
- Karns, D.R., Murphy, J.C., Voris, H.K., Suddeth, J.S., 2005. Comparison of semi-aquatic snake communities associated with the Khorat Basin, Thailand. *Natl. History J. Chulalongkorn Univ.* 5, 73–90.
- Keates, C., Conradie, W., Greenbaum, E., Edwards, S., 2019. A snake in the grass: Genetic structuring of the widespread African grass snake (*Psemmophylax Fitzinger* 1843), with the description of a new genus and a new species. *J. Zool. Syst. Evol. Res.* 57 (4), 1039–1066. <https://doi.org/10.1111/jzs.12337>.
- Knight, A., Mindell, D.P., 1994. On the phylogenetic relationship of Colubrinae, Elapidae, and Viperidae and the evolution of front-fanged venom systems in snakes. *Copeia* 1994, 1–9. <https://doi.org/10.2307/1446664>.
- Köhler, G., Khaing, K.P.P., Than, N.L., Baranski, D., Schell, T., Greve, C., Janke, A., Pauls, S.U., 2021. A new genus and species of mud snake from Myanmar (Reptilia, Squamata, Homalopsidae). *Zootaxa* 4915 (3), 301–325. <https://doi.org/10.11646/zootaxa.4915.3.1>.
- Kumar, A.B., Sanders, K.L., George, S., Murphy, J.C., 2012. The status of *Eurostus dussumieri* and *Hypsirhina chinensis* (Reptilia, Squamata, Serpentes): with comments on the origin of salt tolerance in homalopsid snakes. *Syst. Biodivers* 10 (4), 479–489. <https://doi.org/10.1080/14772000.2012.751940>.
- Lamichhane, S., Berglund, J., Almén, M.S., Maqbool, K., Grabherr, M., Martinez-Barrio, A., Promerová, M., Rubin, C.J., Wang, C., Zamani, N., Grant, B.R., Grant, P.R., Webster, M.T., Andersson, L., 2015. Evolution of Darwin's finches and their beaks revealed by genome sequencing. *Nature* 518 (7539), 371–375. <https://doi.org/10.1038/nature14181>.
- Lanfear, R., Frandsen, P.B., Wright, A.M., Senfeld, T., Calcott, B., 2016. PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. *Mol. Biol. Evol.* 34 (3), 772–773. <https://doi.org/10.1093/molbev/msw260>.
- Lawson, R., Slowinski, J.B., Crother, B.I., Burbrink, F.T., 2005. Phylogeny of the Colubroidea (Serpentes): new evidence from mitochondrial and nuclear genes. *Mol. Phylogenet. Evol.* 37 (2), 581–601. <https://doi.org/10.1016/j.ympev.2005.07.016>.
- Leaché, A.D., Oaks, J.R., 2017. The utility of single nucleotide polymorphism (SNP) data in phylogenetics. *Annu. Rev. Ecol. Syst.* 48, 69–84. <https://doi.org/10.1146/annurev-ecolsys-110316-022645>.
- Li, F., Li, S., 2018. Paleocene-Eocene and Plio-Pleistocene sea-level changes as “species pumps” in Southeast Asia: evidence from *Altheus* spiders. *Mol. Phylogenet. Evol.* 127, 545–555. <https://doi.org/10.1016/j.ympev.2018.05.014>.
- Lillywhite, H., Martins, M., 2019. *Islands and Snakes: Isolation and Adaptive Evolution*. Oxford University Press, New York, New York.
- Liu, F.G.R., Miyamoto, M.M., 1999. Phylogenetic assessment of molecular and morphological data for eutherian mammals. *Syst. Biol.* 48 (1), 54–64. <https://doi.org/10.1080/106351599260436>.
- Lohman, D.J., de Bruyn, M., Page, T., von Rintelen, K., Hall, R., Ng, P.K.L., Shih, H.T., Carvalho, G.R., von Rintelen, T., 2011. Biogeography of the Indo-Australian archipelago. *Annu. Rev. Ecol. Evol. Syst.* 42, 205–226. <https://doi.org/10.1146/annurev-ecolsys-102710-145001>.
- Lukoschek, V., Osterhage, J.L., Karns, D.R., Murphy, J.C., Voris, H.K., 2011. Phylogeography of the Mekong mud snake (*Enhydris subtaeniata*): the biogeographic importance of dynamic river drainages and fluctuating sea levels for semiaquatic taxa in Indochina. *Ecol. Evol.* 1 (3), 330–342. <https://doi.org/10.1002/ece3.29>.
- Luo, Z., Tang, S., Jiang, Z., Chen, J., Fang, H., Li, C., 2016. Conservation of terrestrial vertebrates in a global hotspot of karst area in southwestern China. *Sci. Rep.* 6, 1–12. <https://doi.org/10.1038/srep25717>.
- Luo, A., Ling, C., Ho, S.Y., Zhu, C.D., 2018. Comparison of methods for molecular species delimitation across a range of speciation scenarios. *Syst. Biol.* 67 (5), 830–846. <https://doi.org/10.1093/sysbio/syy011>.
- Maddison, W.P., 1997. Gene trees in species trees. *Syst. Biol.* 46 (3), 523–536. <https://doi.org/10.1093/sysbio/46.3.523>.
- Mahler, D.L., Revell, L.J., Glor, R.E., Losos, J.B., 2010. Ecological opportunity and the rate of morphological evolution in the diversification of Greater Antillean anoles. *Evolution: International Journal of Organic. Evolution* 64 (9), 2731–2745. <https://doi.org/10.1111/j.1558-5646.2010.01026.x>.
- Margres, M.J., Wray, K.P., Seavy, M., McGivern, J.J., Sanader, D., Rokyta, D.R., 2015. Phenotypic integration in the feeding system of the eastern diamondback rattlesnake (*Crotalus adamanteus*). *Mol. Ecol.* 24 (13), 3405–3420. <https://doi.org/10.1111/mec.13240>.
- McKay, B.D., Mays Jr, H.L., Wu, Y., Li, H., Yao, C.T., Nishiumi, I., Zou, F., 2013. An empirical comparison of character-based and coalescent-based approaches to species delimitation in a young avian complex. *Mol. Ecol.* 22 (19), 4943–4957. <https://doi.org/10.1111/mec.12446>.
- McGuire, J.A., Brown, R.M., Riyanto, A., Andayani, N., 2007. The flying lizards of the *Draco lineatus* group (Squamata: Iguania: Agamidae): a taxonomic revision with descriptions of two new species. *Herpetol. Monograph* 21, 179–212. <https://doi.org/10.1655/07-012.1>.
- Meredith, M., Kruschke, J., 2018. HDInterval: Highest (Posterior) Density Intervals. R package version 0.2.0. <https://CRAN.R-project.org/package=HDInterval> (accessed 20 July 2020).
- Miller, M.A., Pfeiffer, W., Schwartz, T., 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: *Proceedings of the Gateway Computing Environments Workshop (GCE)*. IEEE, New Orleans, USA, pp. 1–8. <http://doi.org/10.1109/GCE.2010.5676129>.
- Miralles, A., Marin, J., Markus, D., Herrel, A., Hedges, S.B., Vidal, N., 2018. Molecular evidence for the paraphyly of Scoleophidia and its evolutionary implications. *J. Evol. Biol.* 31 (12), 1782–1793. <https://doi.org/10.1111/jeb.13373>.
- Morales, A.E., Ruedi, M., Field, K., Carstens, B.C., 2019. Diversification rates have no effect on the convergent evolution of foraging strategies in the most speciose genus of bats. *Myotis*. *Evolution* 73 (11), 2263–2280. <https://doi.org/10.1111/evo.13849>.
- Moss, S.J., Wilson, M.E., 1998. Biogeographic implications of the Tertiary palaeogeographic evolution of Sulawesi and Borneo. In: Hall, R., Holloway, J.D. (Eds.), *Biogeography and Geological Evolution of SE Asia*. Backhuys Publishers, The Netherlands, pp. 133–163.
- Murphy, J.C., 2007. *Homalopsid Snakes. Evolution in the Mud*. Kreiger, Melbourne, Florida.
- Murphy, J.C., Lang, R.D., Gower, D.J., Sanders, K.L., 2012a. The Moluccan short-tailed snakes of the genus *Brachyorrhos* Kuhl (Squamata: Serpentes: Homalopsidae), and the status of *Calamophis* Meyer. *Raffles Bull. Zool.* 60 (2), 501–514.
- Murphy, J.C., Mumpuni, Sanders, K.L., 2011. First molecular evidence for the phylogenetic placement of the enigmatic snake genus *Brachyorrhos* (Serpentes: Caenophidia). *Mole. Phylogenetics Evolut.* 61, 953–957. <https://doi.org/10.1016/j.ympev.2011.08.013>.
- Murphy, J.C., Voris, H.K., 2014. A checklist and key to the homalopsid snakes (Reptilia, Squamata, Serpentes), with the description of new genera. *Fieldiana Life Earth Sci.* 2014 (8), 1–43. <https://doi.org/10.3158/2158-5520-14.8.1>.
- Murphy, J.C., Voris, H.K., Karns, D.R., 2012b. The dog-faced water snakes, a revision of the genus *Cerberus* Cuvier, (Squamata, Serpentes, Homalopsidae), with the description of a new species. *Zootaxa* 3484 (1), 1–34. [10.11646/zootaxa.3484.1.1](https://doi.org/10.11646/zootaxa.3484.1.1).
- Murphy, J.C., Voris, H.K., Murthy, B.H.C.K., Traub, J., Cumberbatch, C., 2012c. The masked water snakes of the genus *Homalopsis* Kuhl & van Hasselt, 1822 (Squamata, Serpentes, Homalopsidae), with the description of a new species. *Zootaxa* 3208 (1), 1–26. <https://doi.org/10.11646/zootaxa.3208.1.1>.
- Nagy, Z.T., Joger, U., Wink, M., Glaw, F., Vences, M., 2003. Multiple colonization of Madagascar and Socotra by colubrid snakes: evidence from nuclear and mitochondrial gene phylogenies. *Proc. R. Soc. Lond. B Biol. Sci.* 270 (1533), 2613–2621. <https://doi.org/10.1098/rspb.2003.2547>.
- Nieto-Montes de Oca, A., Barley, A.J., Meza-Lázaro, R.N., García-Vázquez, U.O., Zamora-Abrego, J.G., Thomson, R.C., Leaché, A.D., 2017. Phylogenomics and species delimitation in the knob-scaled lizards of the genus *Xenosaurus* (Squamata: Xenosauridae) using ddRADseq data reveal a substantial underestimation of diversity. *Mol. Phylogenet. Evol.* 106, 241–253. <https://doi.org/10.1016/j.ympev.2016.09.001>.

- Nugraha, A.M.S., Hall, R., 2018. Late Cenozoic palaeogeography of Sulawesi, Indonesia. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 490, 191–209. <https://doi.org/10.1016/j.palaeo.2017.10.033>.
- O'Shea, M., 2018. *The Book of Snakes: A Life-Size Guide to Six Hundred Species from Around the World*. University of Chicago Press, Chicago.
- Paradis, E., Schliep, K., 2019. ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* 35 (3), 526–528. <https://doi.org/10.1093/bioinformatics/bty633>.
- Parnmen, S., Rangsitruji, A., Mongkolsuk, P., Boonpragob, K., Nutakki, A., Lumbsch, H.T., 2012. Using phylogenetic and coalescent methods to understand the species diversity in the *Cladia aggregata* complex (Ascomycota, Lecanorales). *PLoS One* 7 (12). <https://doi.org/10.1371/journal.pone.0052245>.
- Pauwels, O.S., Sumontha, M., 2016. Taxonomic identity reveals great cryptic aquatic snake populations (Squamata: Homalopsidae: *Cerberus* and *Homalopsis*) from southern Thailand. *Zootaxa* 4107 (2), 293–300. <https://doi.org/10.11646/zootaxa.4107.2.11>.
- Pybus, O.G., Harvey, P.H., 2000. Testing macro-evolutionary models using incomplete molecular phylogenies. *Proc. R. Soc. Lond. B Biol. Sci.* 267, 2267–2272. <https://doi.org/10.1098/rspb.2000.1278>.
- Pyron, R.A., Burbrink, F.T., Wiens, J.J., 2013. A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol. Biol.* 13 (1), 93. <https://doi.org/10.1186/1471-2148-13-93>.
- Quah, E.S., Wood Jr., P.L., Grismer, L., Sah, S.A.M., 2018. On the taxonomy and phylogeny of the rare Selangor Mud Snake (*Racliia indica*) Gray (Serpentes, Homalopsidae) from Peninsular Malaysia. *Zootaxa* 4514 (1), 53–64. <https://doi.org/10.11646/zootaxa.4514.1.4>.
- R Core Team, 2019. *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria (accessed 20 July 2020).
- Rainboth, W.J., 1996. *Fishes of the Cambodian Mekong*. Food and Agriculture Organization of the United Nations, Rome.
- Rambaut, A., 2014. FigTree v1.4.2. <http://tree.bio.ed.ac.uk/software/figtree/> (accessed 20 July 2020).
- Rambaut, A., Drummond, A.J., Xie, D., Baele, G., Suchard, M.A., 2018. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Syst. Biol.* 67 (5), 901–904. <https://doi.org/10.1093/sysbio/syy032>.
- Ramírez-Reyes, T., Blair, C., Flores-Villela, O., Piñero, D., Lathrop, A., Murphy, R., 2020. Phylogenomics and molecular species delimitation reveals great cryptic diversity of leaf-toed geckos (Phyllodactylidae: *Phyllodactylus*), ancient origins, and diversification in Mexico. *Mol. Phylogenet. Evol.* 150, 106880. <https://doi.org/10.1016/j.ympev.2020.106880>.
- Ricklefs, R.E., 2007. Estimating diversification rates from phylogenetic information. *Trends Ecol. Evol.* 22 (11), 601–610. <https://doi.org/10.1016/j.tree.2007.06.013>.
- Revell, L.J., 2012. phytools: an R package for phylogenetic comparative biology (and other things). *Methods Ecol. Evol.* 3, 217–223. <https://doi.org/10.1111/j.2041-210X.2011.00169.x>.
- Rodríguez, A., Burgon, J.D., Lyra, M., Irisarri, I., Baurain, D., Blaustein, L., Göçmen, B., Künzel, S., Mable, B.K., Nolte, A.W., Vieth, M., Steinfartz, M., Elmer, K.R., Philippe, H., Vences, M., 2017. Inferring the shallow phylogeny of true salamanders (*Salamandra*) by multiple phylogenomic approaches. *Mol. Phylogenet. Evol.* 115, 16–26. <https://doi.org/10.1016/j.ympev.2017.07.009>.
- Rodríguez, F.J., Oliver, J.L., Marin, A., Medina, J.R., 1990. The general stochastic model of nucleotide substitution. *J. Theor. Biol.* 142 (4), 485–501. [https://doi.org/10.1016/S0022-5193\(05\)80104-3](https://doi.org/10.1016/S0022-5193(05)80104-3).
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Höhna, S., Larget, B., Liu, L., Suchard, M.A., Huelsenbeck, J.P., 2012. MrBayes 3.2: efficient bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* 61 (3), 539–542. <https://doi.org/10.1093/sysbio/sys029>.
- Ruane, S., Austin, C.C., 2017. Phylogenomics using formalin-fixed and 100+ year-old intractable natural history specimens. *Mol. Ecol. Resour.* 17 (5), 1003–1008. <https://doi.org/10.1111/1755-0998.12655>.
- Ruane, S., Bryson Jr., R.W., Pyron, R.A., Burbrink, F.T., 2014. Coalescent species delimitation in milksnakes (genus *Lampropeltis*) and impacts on phylogenetic comparative analyses. *Syst. Biol.* 63 (2), 231–250. <https://doi.org/10.1093/sysbio/syt099>.
- Ruane, S., Myers, E.A., Lo, K., Yuen, S., Welt, R.S., Juman, M., Futterman, I., Nussbaum, R.A., Schneider, G., Burbrink, T., Raxworthy, C.J., 2018. Unrecognized species diversity and new insights into colour pattern polymorphism within the widespread Malagasy snake *Mimophis* (Serpentes: Lamprophiidae). *Syst. Biodivers.* 16 (3), 229–244. <https://doi.org/10.1080/14772000.2017.1375046>.
- Rubinoff, D., Holland, B.S., 2005. Between two extremes: mitochondrial DNA is neither the panacea nor the nemesis of phylogenetic and taxonomic inference. *Syst. Biol.* 54 (6), 952–961. <https://doi.org/10.1080/10635150500234674>.
- Sabaj, M.H., 2016. Standard symbolic codes for institutional resource collections in herpetology and ichthyology: an Online Reference. Version 6.5 (16 August 2016). American Society of Ichthyologists and Herpetologists, Washington, D.C., USA. <http://www.asih.org> (accessed 18 July 2020).
- Sanders, K.L., Lee, M.S., Leys, R., Foster, R., Scott Keogh, J., 2008. Molecular phylogeny and divergence dates for Australasian elapids and sea snakes (Hydrophiinae): evidence from seven genes for rapid evolutionary radiations. *J. Evol. Biol.* 21 (3), 682–695. <https://doi.org/10.1111/j.1420-9101.2008.01525.x>.
- Setiadi, M.I., McGuire, J.A., Brown, R.M., Zubairi, M., Iskandar, D.T., Andayani, N., Supriatna, J., Evans, B.J., 2011. Adaptive radiation and ecological opportunity in Sulawesi and Philippine fanged frog (*Limnodynastes*) communities. *Am. Nat.* 178 (2), 221–240. <https://doi.org/10.1086/660830>.
- Shine, R., Bonnet, X., 2000. Snakes: a new 'model organism' in ecological research? *Trends Ecol. Evol.* 15 (6), 221–222. [https://doi.org/10.1016/S0169-5347\(00\)01853-X](https://doi.org/10.1016/S0169-5347(00)01853-X).
- Smith, M.A., 1943. The Fauna of British India, Ceylon and Burma, including the whole of the Indo-Chinese sub-region. In: *Reptilia and Amphibia*, vol. III. Serpentes. Taylor and Francis, London, pp. 1–583.
- Song, S., Liu, L., Edwards, S.V., Wu, S., 2012. Resolving conflict in eutherian mammal phylogeny using phylogenomics and the multispecies coalescent model. *Proc. Natl. Acad. Sci.* 109 (37), 14942–14947. <https://doi.org/10.1073/pnas.1211733109>.
- Stadelmann, B., Lin, L.K., Kunz, T.H., Ruedi, M., 2007. Molecular phylogeny of New World *Myotis* (Chiroptera, Vespertilionidae) inferred from mitochondrial and nuclear DNA genes. *Mol. Phylogenet. Evol.* 43 (1), 32–48. <https://doi.org/10.1016/j.ympev.2006.06.019>.
- Stamatakis, A., 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30 (9), 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>.
- Steinfartz, S., Veith, M., Tautz, D., 2000. Mitochondrial sequence analysis of *Salamandra* taxa suggests old splits of major lineages and postglacial recolonizations of Central Europe from distinct source populations of *Salamandra salamandra*. *Mol. Ecol.* 9 (4), 397–410. <https://doi.org/10.1046/j.1365-294x.2000.00870.x>.
- Takeuchi, H., Savitzky, A.H., Ding, L., de Silva, A., Das, I., Nguyen, T.T., Tsei, T.S., Jono, T., Zhu, G.X., Mahalpatha, D., Tang, Y., Mori, A., 2018. Evolution of nuchal glands, unusual defensive organs of Asian natrixine snakes (Serpentes: Colubridae), inferred from a molecular phylogeny. *Ecol. Evol.* 8 (20), 10219–10232. <https://doi.org/10.1002/ece3.4497>.
- Tomasello, S., Karstein, K., Hodač, L., Paetzold, C., Hörandl, E., 2020. Phylogenomics unravels Quaternary vicariance and allopatric speciation patterns in temperate-montane plant species: a case study on the *Ranunculus auricomus* species complex. *Mol. Ecol.* 29 (11), 2031–2049. <https://doi.org/10.1111/mec.15458>.
- Townsend, T.M., Alegre, R.E., Kelley, S.T., Wiens, J.J., Reeder, T.W., 2008. Rapid development of multiple nuclear loci for phylogenetic analysis using genomic resources: an example from squamate reptiles. *Mol. Phylogenet. Evol.* 47 (1), 129–142. <https://doi.org/10.1016/j.ympev.2008.01.008>.
- Ukuwela, K.D., De Silva, A., Sanders, K.L., 2017. Further specimens of the mud snake, *Gerarda prevostiana* (Homalopsidae) from Sri Lanka with insights from molecular phylogenetics. *Raffles Bull. Zool.* 65, 29–34.
- Underwood, G., 1999. Morphological evidence on the affinities of vipers. *Kaupia* 8, 3–8.
- Vences, M., Sanchez, E., Hauswaldt, J.S., Eikelmann, D., Rodríguez, A., Carranza, S., Donaire, D., Gehara, M., Helfer, V., Lötters, S., Werner, P., Schulz, S., Steinfartz, S., 2014. Nuclear and mitochondrial multilocus phylogeny and survey of alkaloid content in true salamanders of the genus *Salamandra* (Salamandridae). *Mol. Phylogenet. Evol.* 73, 208–216. <https://doi.org/10.1016/j.ympev.2013.12.009>.
- Voris, H.K., 2000. Maps of Pleistocene sea levels in Southeast Asia: shorelines, river systems and time durations. *J. Biogeogr.* 27 (5), 1153–1167. <https://doi.org/10.1046/j.1365-2699.2000.00489.x>.
- Voris, H.K., Alfaro, M.E., Karns, D.R., Starnes, G.L., Thompson, E., Murphy, J.C., 2002. Phylogenetic relationships of the Oriental-Australian rear-fanged water snakes (Colubridae: Homalopsinae) based on mitochondrial DNA sequences. *Copeia* 2002 (4), 906–915. [https://doi.org/10.1643/0045-8511\(2002\)002\[0906:protoa\]2.0.co;2](https://doi.org/10.1643/0045-8511(2002)002[0906:protoa]2.0.co;2).
- Yang, Z., 2015. The BPP program for species tree estimation and species delimitation. *Curr. Opin. Genet. Evol.* 10, 854–865. <https://doi.org/10.1093/czoolo/61.5.854>.
- Yang, Z., Rannala, B., 2010. Bayesian species delimitation using multilocus sequence data. *Proc. Natl. Acad. Sci.* 107 (20), 9264–9269. <https://doi.org/10.1073/pnas.0913022107>.
- Wiens, J.J., 2003. Missing data, incomplete taxa, and phylogenetic accuracy. *Syst. Biol.* 52 (4), 528–538. <https://doi.org/10.1080/10635150390218330>.
- Wiens, J.J., Morrill, M.C., 2011. Missing data in phylogenetic analysis: reconciling results from simulations and empirical data. *Syst. Biol.* 60 (5), 719–731. <https://doi.org/10.1093/sysbio/syr025>.
- Wiens, J.J., Tiu, J., 2012. Highly incomplete taxa can rescue phylogenetic analyses from the negative impacts of limited taxon sampling. *PLoS ONE* 7 (8). <https://doi.org/10.1371/journal.pone.0042925>.
- Woodruff, D.S., 2010. Biogeography and conservation in Southeast Asia: how 2.7 million years of repeated environmental fluctuations affect today's patterns and the future of the remaining refugial-phase biodiversity. *Biodivers. Conserv.* 19 (4), 919–941. <https://doi.org/10.1007/s10531-010-9783-3>.
- Wüster, W., Dumbrell, A.J., Hay, C., Pook, C.E., Williams, D.J., Fry, B.G., 2005. Snakes across the Strait: trans-Torresian phylogeographic relationships in three genera of Australasian snakes (Serpentes: Elapidae: *Acanthophis*, *Oxyuranus*, and *Pseudechis*). *Mol. Phylogenet. Evol.* 34 (1), 1–14. <https://doi.org/10.1007/s10531-010-9783-3>.
- Workman, D.R., 1977. *Geology of Laos, Cambodia, South Vietnam and the Eastern Part of Thailand*. Overseas Geology and Mineral Resources, vol. 50, Institute of Geological Sciences, Her Majesty's Stationary Office, London, UK.
- Zheng, Y., Wiens, J.J., 2015. Do missing data influence the accuracy of divergence-time estimation with BEAST? *Mol. Phylogenet. Evol.* 85, 41–49. <https://doi.org/10.1016/j.ympev.2015.02.002>.
- Zheng, Y., Wiens, J.J., 2016. Combining phylogenomic and supermatrix approaches, and a time-calibrated phylogeny for squamate reptiles (lizards and snakes) based on 52 genes and 4162 species. *Mol. Phylogenet. Evol.* 94, 537–547. <https://doi.org/10.1016/j.ympev.2015.10.009>.