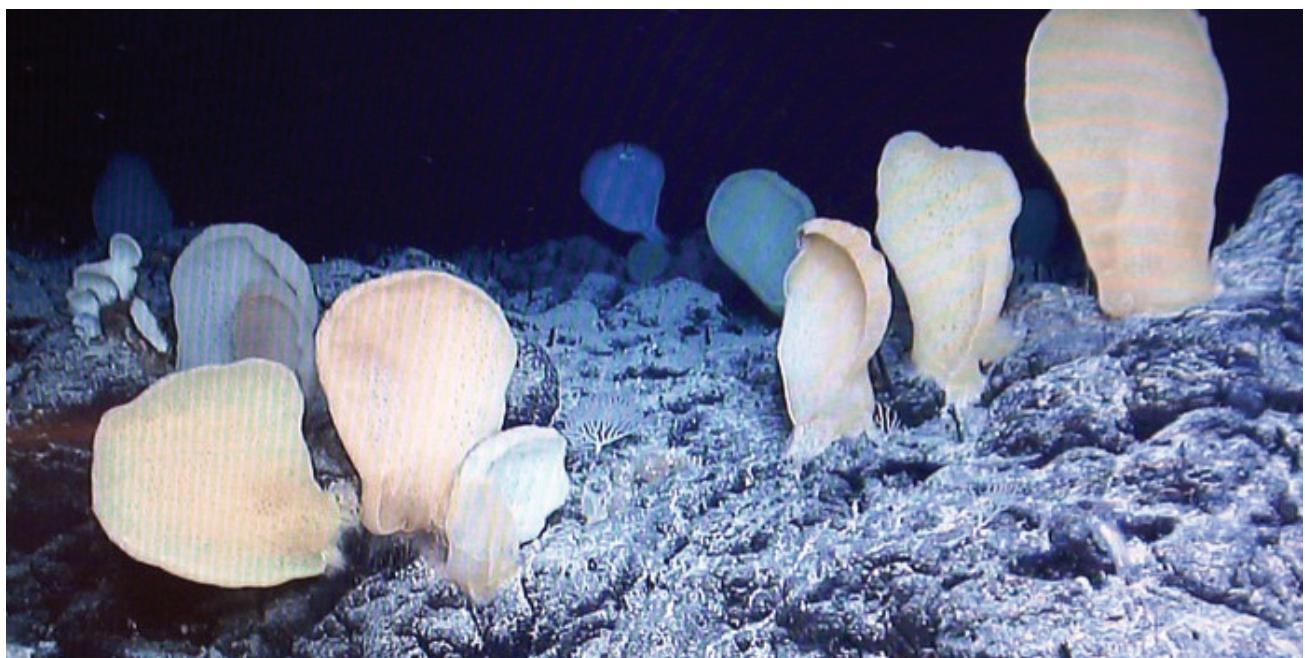


**Report of the**

**WORKSHOP ON DEEP-SEA SPECIES IDENTIFICATION**

**Rome, Italy, 2–4 December 2009**



Cover photo: An aggregation of the hexactinellid sponge *Poliopogon amadou* at the Great Meteor seamount, Northeast Atlantic.  
Courtesy of the Task Group for Maritime Affairs, Estrutura de Missão para os Assuntos do Mar – Portugal.

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## PREPARATION OF THIS DOCUMENT

This FAO Fisheries and Aquaculture Report is the report of the Workshop on Deep-sea Species Identification held in Rome, Italy, from 2 to 4 December 2009. The objective of the Workshop was to identify and review the key issues for vulnerable deep-sea species that should be addressed when developing user-friendly identification tools for corals, sponges and chondrichthyes and, thus, assist in the implementation of the FAO International Guidelines for the Management of Deep-sea Fisheries in the High Seas and enhance fisheries management tools in general. Part 1 of the report contains an overview of the presentations and discussions held during the Workshop and presents the conclusions and recommendations agreed upon by participants. Part 2 of the report contains the main elements of three background documents drafted for the Workshop. The background documents were harmonized for the purpose of this report and represent the consultants' knowledge on current information available on the different species groups.

FAO is grateful to the Workshop participants for their contributions to this report.

### **FAO.**

*Report of the Workshop on Deep-sea Species Identification, Rome, 2–4 December 2009.*

FAO Fisheries and Aquaculture Report No. 947. Rome, FAO. 2011. 209 pp.

### **ABSTRACT**

A Workshop on Deep-sea Species Identification was organized by FAO in Rome, Italy, from 2 to 4 December 2009. The meeting was organized in response to the need for a strategy for the development of appropriate deep-sea species identification tools for fishery purposes, in particular, to address the broadened requirements for reporting on not only target species, but also associated species following recent international developments with respect to fisheries management guidance and biodiversity conservation. The Workshop included an overview of relevant FAO programmes. The overview was followed by presentations and discussions on current knowledge and key issues to be addressed to improve knowledge of vulnerable deep-sea species groups such as chondrichthyes, corals, sponges and other selected deep-sea groups through the development of a database and appropriate identification tools to facilitate reporting on these species groups by fishery operators. Part 1 of the report includes these discussions as well as the conclusions and recommendations agreed upon by participants. Harmonized versions of the three background documents drafted for the Workshop are included in Part 2 of the report.

## ABBREVIATIONS AND ACRONYMS

ABNJ	area beyond national jurisdiction
CBD	Convention on Biological Diversity
CITES	Convention on International Trade in Endangered Species of Wild Fauna and Flora
CCAMLR	Commission for the Conservation of Antarctic Marine Living Resources
COFI	FAO Committee on Fisheries
EAF	ecosystem approach to fisheries
EBSA	ecologically or biologically significant area
EEZ	exclusive economic zone
GIS	geographic information system
GOBI	Global Ocean Biodiversity Initiative
IUCN	International Union for Conservation of Nature
IEO	Instituto Español de Oceanografía
NIWA	National Institute of Water and Atmospheric Research (New Zealand)
RFMA	regional fisheries management arrangement
RFMO	regional fisheries management organization
ROV	remotely operated vehicle
SEAFO	South East Atlantic Fisheries Organisation
SIODFA	Southern Indian Ocean Deepwater Fishers' Association
SPRFMO	South Pacific Regional Fisheries Management Organisation
UNGA	United Nations General Assembly
VME	vulnerable marine ecosystem

## CONTENTS

	<b>Page</b>
Preparation of this document	iii
Abstract	iii
Abbreviations and acronyms	iv
Contents	v
<b>PART 1: REPORT OF THE EXPERT WORKSHOP</b>	<b>1</b>
Background	1
Introduction	1
Relevant FAO Programmes	2
Discussion on chondrichthyes, corals and sponges	3
Discussion on vulnerable deep-sea or bycatch fish species	7
Deep-sea species identification guides for vulnerable species	8
Discussion on ways forward for the development of guides for vulnerable deep-sea species	9
Conclusions and recommendations	11
<b>APPENDIXES OF PART 1</b>	
Appendix 1: List of participants	12
Appendix 2: Agenda	13
<b>PART 2: EXPERT WORKSHOP BACKGROUND DOCUMENTS</b>	
Information to assist preparation of a deep-sea species identification guide – sharks, batoids and chimaeras (class Chondrichthyes)	
<i>by Peter M. Kyne and Colin A. Simpfendorfer</i>	15
Global list of cold-water corals (order Scleractinia; sub-order Filifera; subclass Octocorallia, order Antipatharia) from waters deeper than 200 m, vulnerable species, and draft recommendations for the production of identification guides	
<i>by Marcelo Visentini Kitahara</i>	97
Towards the development of an identification guide for vulnerable deep-sea sponges	
<i>by Joana R. Xavier and Rob W.M. van Soest</i>	149



## PART 1: REPORT OF THE EXPERT WORKSHOP

### **Background**

The international community has responded to increased concern regarding sustainable use of marine resources and marine conservation in areas beyond national jurisdiction (ABNJ) through a number of recently developed international instruments. The International Guidelines for the Management of Deep-sea Fisheries in the High Seas (FAO Deep-sea Guidelines [2009]) were one such instrument developed through FAO. The United Nations General Assembly (UNGA) has also addressed this issue in various resolutions (e.g. 61/105). In 2008, the Conference of Parties to the Convention on Biological Diversity (CBD), at its ninth meeting, adopted criteria for Ecologically or Biologically Significant Areas (EBSAs; Decision IX/20 Annex 1). Coordinated by the International Union for Conservation of Nature (IUCN), the Global Ocean Biodiversity Initiative (GOBI), an international partnership of marine institutions, has been working on developing both technical guidance and training materials concerning implementation of the CBD EBSA criteria.

The ecosystem approach to fisheries (EAF) is now widely advocated and applied in deep-sea fisheries. However, the inherent restrictions on obtaining sufficient information for stock assessment or benthic habitat data (compared with nearshore shelf/slope fisheries) mean that management regimes typically operate at a low level of knowledge, and management action must occur in a highly precautionary manner. In many deep-sea fisheries, there is a lack of the most basic underlying catch data to support adequate fisheries management and conservation efforts, but also, most of the stock are “poor data stock”, i.e. with sporadic or intermittent fishing activity, that do not allow traditional assessments.

The main commercial fish species are often well recorded in fishery logbooks, but many of the bycatch species are not because they are unable to be easily identified at sea. Detailed taxonomic literature is of little help at sea to fishers, observers or even trained scientists, because much of the information is provided as detailed keys to entire taxa, which are practically impossible to use on a ship without a lot of time and specialized equipment such as microscopes. However, many of the common species can be readily identified with the aid of pictorial guides that highlight the main physical characteristics, and are annotated with comments to avoid confusion with similar species. Hence, the development of identification guides specifically for use of non-specialist technical staff at sea will enable better information on bycatch composition and stock status, as well impacts on, and location of, vulnerable marine ecosystems (VMEs) that have been recognized as major gaps in knowledge.

The development of species identification guides is specifically recommended in the FAO Deep-sea Guidelines in paragraph 36 (below):

“36. National and international training programmes for fishers and scientific observers should be used to improve catch identification and biological data collection, including the use of existing FAO material for the identification of commercial species, and the development of field manuals for the identification of non-commercial species, particularly for benthic invertebrates. FAO should provide support to the development and coordination of such programmes.”

### **Introduction**

1. The Workshop on Deep-sea Species Identification was held in Rome, Italy, from 2 to 4 December 2009 and was attended by six participants from a wide range of experiences and geographic areas (see Appendix 1). The meeting was opened by Merete Tandstad, FAO Marine and Inland Fisheries Service, who welcomed the participants and asked participants to introduce themselves.

2. The agenda was presented and participants were given the opportunity to comment on the agenda, after which the agenda was adopted (see Appendix 2).

3. The Workshop was tasked with developing a strategy for the production of appropriate deep-sea species identification tools that can be used in fisheries operations. Johanne Fischer, FAO Marine and Inland Fisheries Service, introduced the workshop objectives and explained that the envisaged identification tools for vulnerable deep-sea species are meant to: (a) assist in the implementation of fisheries management measures (e.g. bycatch requirements, recording of catches, and inspection); (b) enhance scientific assessment; (c) be used by practitioners and by scientists; and (d) create public awareness. For the purpose of the Workshop, participants were requested to discuss a number of issues, such as the selection of species (e.g. pelagics yes or no; depth considerations, taxonomic levels for identification, and definition of vulnerability), as well as the types of products required (e.g. ID cards for on-the-spot identification, more comprehensive guides, and electronic databases). Other important topic consisted in the geographic resolution for different types of products, the information basis available so far for selected species (e.g. scientific drawings, and photographs), availability of cooperating experts for different species groups and the general logistic approach for each species group.

4. Malcolm Clark, National Institute of Water and Atmospheric Research (NIWA), New Zealand, was nominated chair of the meeting.

### **Relevant FAO Programmes**

5. Two FAO Programmes are of particular relevance to the work discussed at the meeting: the FAO Programme on Deep-sea Fisheries in the High Seas; and the FAO FishFinder Programme.

#### ***FAO Programme on Deep-sea Fisheries in the High Seas***

6. Jessica Sanders, FAO Policy, Economics and Institutions Service, presented the FAO Programme on Deep-sea Fisheries in the High Seas. Through the adoption of the FAO International Guidelines for the Management of Deep-sea Fisheries in the High Seas, FAO was requested by the Committee on Fisheries (COFI) to carry out a number of supporting activities to create awareness and facilitate the implementation of the FAO Deep-sea Guidelines. Building on these requests, FAO has initiated a programme with the aim of assisting States, institutions, the fishing industry and the regional fisheries management organization and arrangements (RFMO/As) in the implementation of the FAO Deep-sea Guidelines. The objective is to improve the current management systems through more and better information and tools, as well as to better engagement and communication among stakeholders, and capacity building. The four-year programme seeks to establish a knowledge baseline in relation to these fisheries and related ecosystems. It contains four major components: (i) support tools for the implementation of the FAO Deep-sea Guidelines; (ii) a VME information system; (iii) pilot implementation activities for enhanced management of deep-sea resources; and (iv) global coordination, monitoring and evaluation, and dissemination of information. The Programme is seen as a multidonor programme, where components or elements of components can be supported through a modular approach.

#### ***FAO FishFinder Programme***

7. Johanne Fischer, FAO Fisheries Management and Conservation Service, introduced the FAO FishFinder Programme. The Programme's objective is to improve the identification of marine organisms of actual and potential interest to fisheries by providing and disseminating tools to facilitate species identification in fisheries and by providing a global and coherent system of scientific and common nomenclature. Priority is assigned to resources of major commercial importance or threatened species and to developing countries/regions facing difficulties in species identification. The main activities of the Programme are: to secure the best up-to-date information (calling upon knowledgeable specialists in taxonomy); to compile information on species distribution in order to

produce distribution maps; to draw reliable and accurate illustrations of marine organisms and their anatomical details; and to produce and distribute, through different media, species identification information for fishery purposes. The outputs are publications on species identification such as regional and field guides, catalogues, CD-ROMs, synopses, fact sheets available on the Web, species distribution maps and scientific illustrations.

### **Discussion on chondrichthyes and corals and sponges**

8. As a basis for the technical discussions, the Workshop was presented with the following three comprehensive background documents:

- Information to assist preparation of a deep-sea species identification guide – sharks, batoids and chimaeras (class Chondrichthyes), by Peter M. Kyne and Colin A. Simpfendorfer;
- Global list of cold-water corals (order Scleractinia; sub-order Filifera; sub-class Octocorallia, order Antipatharia) from waters deeper than 200 m, vulnerable species, and draft recommendations for the production of identification guides, by Marcelo Visentini Kitahara;
- Towards the development of an identification guide of vulnerable deep-sea sponges, by Joana R. Xavier and Rob W.M. van Soest.

In addition, two presentations were made: one by José Luis López Abellán, Instituto Español de Oceanografía (IEO), Spain, on work on deep-sea species in the Southeast Atlantic; and one by Malcolm Clark on NIWA's experience on developing species identification guides for fish and invertebrates.

The Workshop participants commended the authors of the background documents on their work and provided guidance on improvements to be made to these documents. A summary of the background documents as well as the discussions resulting from them is provided below, whereas the comprehensive background documents can be found in Part 2 of this report.

#### ***Chondrichthyes***

9. Peter M. Kyne, Charles Darwin University, Research Institute for the Environment and Livelihoods, Australia, provided an extensive account on the immense information already available on these groups, noting that almost half of the total number of species occurs in deep-sea. An important note is that almost all of the deep-sea species have their bathymetric limit of distribution at around above 2 000 m and, therefore, cannot seek refuge (from fisheries) in deeper water. Mr Kyne further showed that there is a correlation between depth and species productivity (with deeper species generally producing fewer offspring or having later maturation age).

10. The class Chondrichthyes (cartilaginous fishes) comprises the sharks, batoids (together forming the subclass Elasmobranchii) and the chimaeras (subclass Holocephali). There has been considerable recent focus on the status of species and populations within this class given their inherent vulnerability resulting from life-history characteristics and well-documented and publicized cases of population declines and depletions. Particularly high levels of endemism are found in lanternsharks (Etmopteridae), catsharks (Scyliorhinidae) and batoids (Rajiformes), and this endemism is often associated with seamounts and mid-oceanic ridges.

11. Almost half of the known cartilaginous fishes (532 of 1 144 species) can be considered "deep-sea", although many other primarily shelf species have also been recorded at depths of more than 200 m. The deep-sea fauna comprises 254 sharks (~53 percent of known species; 23 families), 237 batoids (~35 percent of known species; 11 families) and 41 chimaeras (~91 percent of known species; 2 families). All 10 orders of chondrichthyans are represented in the deep-sea, however, the bulk of the fauna is attributable to four main groups: (i) squaloid dogfishes (order Squaliformes; 46.1 percent of the deep-sea shark fauna); (ii) scyliorhinid catsharks (order Carcharhiniformes, family

Scyliorhinidae; 40.2 percent of the deep-sea shark fauna); (iii) skates (order Rajiformes; families Arhynchobatidae, Rajidae and Anacanthobatidae; 89.7 percent of the deep-sea batoid fauna); and (iv) chimaeras (order Chimaeriformes, families Rhinochimaeridae and Chimaeridae).

12. A checklist of all known described extant deep-sea chondrichthyans (see Part 2, Appendix 1) was provided in phylogenetic order including species (with authority) and common names, a brief account of geographic distribution (including an overview of the respective FAO Areas for each species, allowing construction of species lists for any FAO Area), habitat preferences, depth range, and any relevant taxonomic issues. All deep-sea chondrichthyes are considered to be vulnerable to capture in fishing gear given their morphology, behaviour and habitat associations.

13. Lists were also provided of species known to be targeted (including historically) in fisheries (including those landed as byproduct or bycatch in multispecies fisheries) (17 sharks, 17 batoids, 3 chimaeras) and species known to be caught as bycatch (147 sharks, 146 batoids, 23 chimaeras; total of 316 species, although more have probably gone unrecorded). Approaches to assessing the “vulnerability” of species were discussed and the author suggested that these could include a combination of information on life history and productivity (the decreasing relationship between productivity and depth should be considered), extent of occurrence, rarity, target/bycatch species and conservation status (if any).

14. Reference lists were compiled of existing regional and global identification guides, general deep-sea chondrichthyan papers, general fisheries-related papers, and papers relating to species groups (the bulk of references): (i) squaloid dogfishes (divided by family); (ii) scyliorhinid catsharks; (iii) skates; (iv) chimaeras; and (v) other deep-sea groups. For these species groups, references related to taxonomy, distribution and occurrence, habitat, biology, ecology, fisheries and conservation were provided. While there are a considerable number of references available for some shark families (dogfishes and catsharks), the bulk in fact relate to a handful of species, mostly from the Northeast Atlantic. In addition, an overview of currents experts was provided.

15. The skate literature is diverse but much of it comes from shallower water species or from studies on the shelf for species with wide depth ranges, and chimaera literature is mostly limited to species descriptions.

16. As regards the development of new identification guides to deep-sea chondrichthyans, it was recommended that these should be undertaken at the species level, given the current status of knowledge on the group available information and the number of experts active in this field. Higher-level keys to families and genera should be provided. Keys to species are essential. Guides could include keys to all species (globally and regionally for regional guides), but then the species accounts could focus on the selected “vulnerable” species only. The selection of species should include those recorded as bycatch (including common species; not only “vulnerable” species). For individual accounts, the following components are considered critical: diagnostic characteristics, a list of similar species (referring the user to these species accounts), distribution maps and line drawings.

17. In the subsequent discussions, it was noted that a deep-sea chondrichthyan identification guide could be produced relatively quickly as there is considerable information available from existing FAO guides and publications. For the sharks, the bulk of species have detailed accounts prepared or in preparation from the Catalogue of Sharks of the World.<sup>1</sup> Material on batoids and chimaeras is more limited, but there is still a considerable amount available (e.g. 40 percent of deep-sea batoids have existing FAO line drawings).

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<sup>1</sup> Compagno, L.J.V. 2001. *Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Volume 2. Bullhead, mackerel and carpet sharks (Heterodontiformes, Lamniformes and Orectolobiformes)*. FAO Species Catalogue for Fishery Purposes No. 1, Vol. 2. Rome, FAO. 269 pp. Note that Vol. 1 and Vol. 3 are still in preparation.

18. For this group, it was recommended that a global guide be prepared first, followed by regional guides as appropriate or required (it would be possible to prepare species lists for each FAO Area, or larger ocean regions from checklists provided). This would imply preparation of all information in the first instance, from which information could then be extracted for regional guides. It was, however, stressed that the taxonomy remains unresolved for several groups (*Apristurus*, *Centrophorus*, *Squalus*, and some skate groups) and there are at least 65 undescribed chondrichthyan species presently known (with numbers continuing to increase). Development of a guide will need to monitor developments in taxonomy and systematics, but selected experts will have a good understanding of this.

19. The Workshop participants agreed that chondrichthyans constitute the group for which more existing information and illustrations for a guide would be available, recognizing that there would still be gaps to address.

### *Corals*

20. Marcelo Visentini Kitahara, James Cook University, Australia, introduced this topic, providing an overview of current knowledge and information available on this group, including a comprehensive list of all scleractinian (Anthozoa, Hexacorallia, Scleractinia) and calcified hydrozoans (Hydrozoa, Filifera, Stylsteridae) species, and all potential habitat-forming cold-water octocorallians (Anthozoa, Octocorallia) and antipatharians (Anthozoa, Hexacorallia, Antipatharia). An important note was that more than half of the total number of species occur in deep-sea, and provide habitat for a large variety of organisms. These organisms rely on corals as a source of food and shelter. In addition, it was noted that all species are vulnerable to many human activities, including deep-sea fisheries and that, to date, there are no data on the resilience of the habitats formed by deep-sea corals.

21. The term coral has been defined by Cairns<sup>2</sup> as: “animals in the cnidarian classes Anthozoa and Hydrozoa that produce either calcium carbonate (aragonitic or calcitic) secretions resulting in a continuous skeleton or as numerous microscopic, individualized sclerites, or that have a black, horn-like, proteinaceous axis”. From this definition, there are four orders belonging to two cnidarian classes: (i) class Anthozoa, subclass Hexacorallia – order Scleractinia, order Zoanthidea, order Antipatharia, subclass Octocorallia; and (ii) class Hydrozoa, subclass Hydroidolina – order Anthoathecata.

22. Known since the eighteenth century, cold-water corals are involved in the formation of large seabed structures such as reefs and giant carbonate mounds. These structures sustain some of the most species-rich marine ecosystems. Unlike tropical and subtropical coral reefs that grow in relatively shallow waters, cold-water corals do not rely on symbiosis with photosynthetic dinoflagellates (*Symbiodinium* species); instead, they capture microscopic animals and plant matter that drift past in the water column in waters deeper than 50 m.

23. A compilation was made for global distribution (in relation to the 19 FAO Major Fishing Areas) of all cold-water species of scleractinians (797 spp., including undescribed species), filiferians (246 spp.), all potential habitat-forming cold-water octocorallians (225 spp.), and antipatharians (33 spp.) known to occur in waters deeper than 200 m. Despite their important role in deep-sea environments, other cnidarians such as Corallimorpharia, Ceriantharia, Zoanthidea and Actiniaria are not part of the present report because they do not form large three-dimensional structures. All these groups, and many other taxa besides, can be important benthic invertebrate bycatch in commercial fisheries, yet, overall, can be considered less vulnerable to fishing gear. The species groups compiled in this report were proposed by the Workshop to be used as a starting point for any identification guides (but not excluding other, potentially interesting, groups).

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<sup>2</sup> Cairns, S.D. 2007. Deep-water corals: an overview with special reference to diversity and distribution of deep-water scleractinian corals. *Bulletin of Marine Science*, 81(3): 311–322.

24. From each of the four groups, there is a brief discussion of species known to be: (i) vulnerable to fishing gear or other factors; or/and (ii) of commercial importance; or/and (iii) bycatch species. In addition, draft recommendations for the production of identification guides are suggested.

25. For some species of corals, it is complicated to compile knowledge owing to the difficulty in obtaining samples. One recognized problem is the difficulties related to exchanging materials between countries for research owing to restrictions under the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES), especially in those countries that do not have institutions with permanent CITES permissions. With regard to illustrations, photographs of corals are available for approximately 70 percent of the known species, and it is possible to acquire photos for many species from coral collections in museums.

26. To date, the possible level of identification is as follows: (i) scleractinian species (all colonials and key solitary species); (ii) order Antipatharia; (iii) subclass Octocorallia and key genera; and (iv) suborder Filifera and key genera. It was recognized that identifying some of the Scleractinia genera down to species level is problematic.

27. It was noted that the ecological role of solitary species is unknown and, thus, it was recommended to include these species owing to their potential vulnerability. Moreover, reliable distribution maps for solitary species are not readily available, and an effort should be made to see if such maps could be developed.

28. Many coral species are widespread, leading to considerable overlaps between FAO statistical areas. Nevertheless, the suggestion was made to start developing guides for individual regions instead of a global guide, noting that information compiled for one region could be applicable also to other regions. The guides should contain information on multiple species and particular attention should be paid to differences between similar species to help the users avoid confusion.

29. Participants discussed the difficulties in assessing vulnerability of different species for inclusion in the guides. It was also mentioned that climate change impacts needed to be considered as part of longer-term projects.

### *Sponges*

30. Joana R. Xavier, Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, the Netherlands, introduced this topic, providing an overview of current knowledge and information available on this group. She noted that sponge taxonomy and systematics is mainly based on internal morphological features such as the type, shape, size and arrangement of the skeletal structures (spicules, collagen fibrils and spongin fibres) that require microscopic examination. Additional characteristics such as external features (colour, shape, consistency, and distribution of the aquiferous system's openings) can be informative but alone are unreliable for taxonomic assignment.

31. The phylum Porifera (from Latin *porus* + *ferō* = pore bearer) constitutes a group of aquatic animals that is widely distributed geographically and bathymetrically in both marine and freshwater ecosystems. Regarded among the most primitive extant animal groups (635 mya<sup>3</sup>), they are a dominant invertebrate group in hard-bottom benthic communities throughout the temperate, tropical, and polar zones from intertidal to abyssal depths. Sponges play important ecological roles in bioerosion, reef formation, substrate consolidation, benthoo-pelagic coupling, and habitat provision that have major implications for ecosystem functioning. In addition, as sessile organisms, sponges have developed a range of chemical defence strategies against predators, spatial competitors, or as antifouling. These secondary metabolites with antimicrobial, analgesic, antiviral and anticancer activities have placed sponges among the most prolific and promising producers of medical compounds and increasingly attract the interest of pharmaceutical companies.

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<sup>3</sup> Dating is from the International Commission on Stratigraphy ([www.stratigraphy.org](http://www.stratigraphy.org)); mya = million years ago.

32. The phylum comprises three extant (Demospongiae, Calcarea and Hexactinellida) and one extinct (Archaeocyatha) classes, 25 orders, 127 families, and 682 genera. Approximately 8 300 species are currently recognized (World Porifera Database<sup>4</sup>) but the true diversity of this group is estimated to comprise probably twice this number. It is estimated that there are approximately 4 000 sponge species occurring in deep-seas.

33. In some areas, deep-sea sponges form structurally complex habitats, e.g. sponge grounds and reefs, that support a diversity of associated fauna. There are three main types of sponge aggregations: (i) demosponge grounds, known as “ostur or cheese bottoms”, composed by multispecific assemblages of large-sized and very abundant astrophorid species of the genera *Geodia*, *Stelletta*, *Isops* and *Stryphnus*; (ii) “monospecific” aggregations of glass sponges such as *Pheronema carpenteri*, *Asconema setubalense*, *Rossella nodastrella* and *Poliopogon amadou*; and (iii) sponge reefs, thus far only known from the western Canadian continental shelf, formed by several hexactinellid species such as *Chonelasma* sp., *Heterochone calyx*, *Aphrocallistes vastus* and *Farrea occa*. In other deep-sea habitats, such as in cold-water coral reefs, sponges do not constitute the primary structural organisms although their diversity surpasses by far that of the structural taxa (e.g. up to 122 sponge species associated to bathyal *Madrepora oculata* and *Lophelia pertusa* reefs of the Rockall Bank).

34. The distribution of all deep-sea species within the class Hexactinellida (glass sponges – 5 orders; 20 families; 121 genera; 645 species); and the orders Lithistida (stony sponges – 14 families; 45 genera; 197 species), Astrophorida (5 families; 42 genera; 691 species) and family Cladorhizidae (carnivorous sponges – 6 genera; 105 species) within the class Demospongiae by FAO Major Fishing Areas was compiled.

35. A list of bibliographic references was compiled. This list provides the main references containing information on: general sponge taxonomy; historical campaigns that yielded extensive sponge reports; sponge aggregations; trawl-induced damage to sponge populations; general deep-sea sponge diversity; and recent group-specific taxonomy. In addition, an overview of current experts was provided.

36. The main recommendations in regard to strategy and limitations on the production of a deep-sea sponge guide were provided. The main limitations highlighted were: (i) the need for microscopic examination for accurate species and/or genera assignment; (ii) the lack of information on species distribution and abundance; and (iii) a general shortage of sponge taxonomists especially for some regions/FAO Areas.

37. To date, despite the scarce knowledge on deep-water sponges, their distribution at species level seems to be very restricted, and usually each species is reported for only one FAO area. Yet, largely in contrast to shallow-water species, most of the deep-sea records are associated with detailed collection data (geographic coordinates, depth ranges, substrate and habitat characteristics).

38. It was broadly accepted that, although difficult, the FAO deep-water vulnerable species guide must include a representative number of deep-sea sponges to increase awareness on the group and, in addition, it will probably increase knowledge once more specimens are collected. In this respect, it was suggested that the specimens, species pictures and/or data collected through the initiative could be forward to the group expert.

## **Discussion on vulnerable deep-sea or bycatch fish species**

39. Luis José López Abellán, IEO, presented a summary of the procedure adopted by the RFMO (South East Atlantic Fisheries Organisation [SEAFO]) in the region to address UNGA Resolution 61/105 and other impacts of bottom fishing on elasmobranchs and potentially other

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<sup>4</sup> World Porifera Database can be accessed at: [www.marinespecies.org/porifera](http://www.marinespecies.org/porifera)

deep-sea fishes. For the latter topic, different views in relation to the definition of vulnerability were introduced for discussion and some examples of the practical application of resilience/productivity and intrinsic vulnerability concepts were given.

40. The joint Namibian–Spanish multidisciplinary research activities conducted on the Walvis Ridge seamounts (Ewing and Valdivia Bank) in 2008 and 2009 were described. They focused on mapping and characterizing seamounts, and on identifying potential VME areas or locations. In addition, an analysis of specific composition of samplings was made.

41. A general overview of fisheries carried out in the area was provided and the species composition of catches, including bycatch, in commercial fishing was analysed by type of gear. Data from research cruises and commercial activity were compared, showing the loss of information in relation to species identification in the commercial fishing.

42. A general descriptive account of fisheries in the South Atlantic was also presented. The SEAFO region was described, which is divided into four reporting areas (A–D). This covers two oceanographically different sections, south of 40 °S with Antarctic waters and north of 40 °S, which is more temperate. It was noted that SEAFO has started the process of describing the intrinsic vulnerability or vulnerability to fishing gear of most commercially important species in relation to the levels of aggregations, life history and distributions.

43. Discussion focused on the work done assessing the relative vulnerability of fish species. It was suggested that doing this for a wider range of bycatch species could be useful. For example, elasmobranchs had not yet been analysed, but it would be of interest to see how their intrinsic vulnerability compared with some of the less-productive teleosts.

44. It was stated that Cheung's method,<sup>5</sup> a fuzzy expert system to estimate vulnerability to fishing, based on life-history and ecological characteristics of species is available through FishBase,<sup>6</sup> and has been included in the SEAFO species profiles developed by this organization, but this would have to be checked as the information might not be available for non-commercial species. The reference to the resilience (productivity) of species is also included in the species profiles using Musick's method.<sup>7</sup> This could feed into a general discussion of relative vulnerability if that aspect were included in the species identification guides.

45. The possibility of looking at the vulnerability of mixed species aggregations to fishing gear, the vulnerability of essential habitat and the productivity of species combined as an “ecosystem vulnerability” were also discussed.

### **Deep-sea species identification guides for vulnerable species**

46. An overview of work on deep-sea species guides in the Southern Pacific was presented as an example of what guides for vulnerable deep-sea species could contain, including suggestions of elements for the development of deep-sea species guides for vulnerable species under the FAO Deep-sea Programme. This was followed by discussions on next steps in the preparation process, including a discussion on possible elements for a future project proposal in support of the development of such guides and on general information for a database on vulnerable deep-sea species that would feed into these guides.

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<sup>5</sup> Cheung W.W.L., Pitcher, T.J. and Pauly, D. 2005. A fuzzy logic expert system to estimate intrinsic extinction vulnerability of marine fishes to fishing. *Biological Conservation* 124: 97–111.

Cheung W.W.L., Watson, R., Morato, T., Pitcher, T.J. and Pauly, D. 2007. Intrinsic vulnerability in the global fish catch. *Marine Ecology Progress Series*, 333: 1–12.

<sup>6</sup> FishBase can be accessed at: [www.fishbase.org](http://www.fishbase.org)

<sup>7</sup> Musick, J.A. 1999. Criteria to define extinction risk in marine fishes. *Fisheries*, 24(12): 6–14.

### *Overview of work on deep-sea species in the Southern Pacific*

47. Malcolm Clark gave a presentation entitled “Development of guides for VME species experience with identification of fish and invertebrate bycatch”. This described the context and progressive development of identification guides by NIWA since the 1990s, which include a wide range of fish and benthic invertebrate species from New Zealand waters. They have also been modified for use on fishing vessels that operate in the southern Indian ocean (through the Southern Indian Ocean Deepwater Fishers’ Association [SIODFA]). Separate guides have been developed for the Antarctic (Commission for the Conservation of Antarctic Marine Living Resources [CCAMLR]) and South Pacific (South Pacific Regional Fisheries Management Organisation [SPRFMO]).

48. Example pages were shown for both fish and benthic invertebrate guides. The structure and content were based on presenting information on several hundred species in a form appropriate for use by research scientists, as well as scientific observers on commercial vessels. The guides include sets of instructions for handling and preserving the samples. Photographic “keys” were provided for higher family or order levels, so that identification could occur at the appropriate taxonomic level depending on the condition of material and confidence of the user. Each species page included scientific and common names, recording codes (generally a three-letter code), a distribution map (for fishes), an annotated photograph and/or line drawing, and notes on distinguishing features, colour, size, distribution, depth, similar species, and references.

49. The guides prepared by NIWA were not initially “identification-at-a-glance”. However, this approach was applied for the first drafts of working observer identification sheets prepared for the SPRFMO and recently the CCAMLR in response to developing VME criteria.

### **Discussion on ways forward for the development of guides for vulnerable deep-sea species**

50. Following the different presentations, there was discussion focusing on how to ensure continued development of deep-sea species guides for fisheries purposes, including possible elements for a project proposal. Information on each species should be stored in a relational database format. The underlying concept of a database is that it would enable a variety of products to be output, and to evolve with new information and data. In addition, if species data or pages were available electronically, and searchable and selectable online, that would enable interested users to compile their own identification guide for a specific geographical area or group of species. General information and information requirements that should be included in that database can be found in Table 1.

Furthermore, the following was noted with regard to what guides for vulnerable species should include and how these should be structured:

51. The structure of the guide should be hierarchical, so that identification of all specimens can occur, even if taxonomic resolution varies. A balance is needed between identifications being too general to be informative (e.g. simply coral), and too specific (or including too many species) to be usable.

52. Identification needs to be based on external and macro-size features (so species included do not require identification with microscopes or technical equipment) to facilitate onboard identification.

53. Rare and unusual species need to be retained, so the guide should include notes on preservation of specimens. Similarly, other species that cannot be easily identified using macro-size features might need to be retained.

54. Guides need to have general introductory notes on the species and groups, and identification drawings to show the anatomical features used or referred to in the guide.

**Table 1. General information for database on vulnerable deep-sea species**

This is a summary table of desirable information for a database (repository) on deep-sea species. Depending on the species, more or less information will be available to create either sheets or any type of documentation needed by the users.

Key pieces of information	Description (if available)
Nomenclature	<ul style="list-style-type: none"> <li>Species name, including species authority</li> <li>Common name</li> <li>Junior synonyms, if regularly used</li> </ul>
Codes	<p>Existing codes:</p> <ul style="list-style-type: none"> <li>FAO codes (chondrichthyes, bony fish)</li> <li>Other codes (e.g. RFMOs, etc.)</li> <li>New codes</li> <li>Create codes for sponges and corals (refer to CCAMLR, and others)<sup>1</sup></li> </ul>
Taxonomic systematics	<ul style="list-style-type: none"> <li>Phylum</li> <li>Class</li> <li>Order</li> <li>Family</li> </ul>
Line drawings and/or colour photography (both live and on deck)	<ul style="list-style-type: none"> <li>Annotated to highlight features</li> <li>Details in line drawing</li> </ul>
Distinguishing features	<ul style="list-style-type: none"> <li>Description of main characteristics, including diagnostic descriptions (key points to look for)</li> </ul>
Formatting	<ul style="list-style-type: none"> <li>Colour attribution</li> <li>Size (to be consistent)</li> </ul>
Distribution	<ul style="list-style-type: none"> <li>Description or map: it is better if it is an expert map of distribution, but record points (type of locality and others) should also be included</li> </ul>
Depth range	<ul style="list-style-type: none"> <li>If it is known to be more common at a certain depth range, e.g. 300–1 200 m, commonly 500–700 m</li> </ul>
Habitat and biology (optional, when the space is available)	<ul style="list-style-type: none"> <li>To be discussed according to the region</li> <li>Information that is useful or interesting to heighten interest/awareness, e.g. usually found on hydrothermal vents, seamounts, etc. / reproduction / life-history characteristics including age, growth, etc.</li> <li>Follow a consistent format and layout</li> <li>Habitat preferences, e.g. at broad scale — slope, seamounts</li> <li>Key biological features, e.g. size, age/growth, reproduction, maturity</li> <li>Elements to report, e.g. if elasmobranch pups expelled when on deck</li> <li>Assessing maturity</li> </ul>
Similar species	<p>Description of similar species to differentiate them from others:</p> <ul style="list-style-type: none"> <li>list names and page numbers if a key, or other species, are included; or</li> <li>list what to look at for distinguishing features</li> </ul>
Remarks	<p>Covers any other comments (e.g. general taxonomic confidence, distribution, poisonous):</p> <ul style="list-style-type: none"> <li>Symbol to signify category if it is listed on the IUCN Red List (if it is already described in the introduction)</li> <li>Regional guides to signify if it is already listed by an RFMO</li> </ul>

<sup>1</sup> To be evaluated based on different codes in use.

55. It is important to annotate with notes about similar species or taxa in order to minimize confusion with similar species and aid confidence in identifications.

56. Photographs showing fresh coloration have proved very useful, making a combination of annotated line drawing and good colour photographs the preferred option.

57. If the guide is in a book format, it will need multiple indexes (species, common name, family).

58. Observer guides are generally for higher taxa, for which sheet/poster-type formats can also work well.

59. There is a need for consistency in guide form, species covered, codes between FAO, RFMOs and national bodies.

60. Training with actual material is useful (essential for some groups such as stony corals).

### **Conclusions and recommendations**

61. The Workshop recognized the widespread distribution of deep-sea fisheries, the vulnerability of certain taxa taken as bycatch in these bottom contact fisheries and the lack of means to identify these species. The Workshop also recognized the immediate need for identification tools to inform fisheries management.

62. The Workshop recommended that a series of identification guides be developed for certain vulnerable groups of species affected by bottom gear, with an initial focus on three of the most affected groups: chondrichthyes, corals and sponges.

63. The Workshop agreed that the compilation of information should be on a global scale and be as comprehensive as possible, using existing information as well as encouraging the collection of new data.

64. The Workshop agreed that this information can be used as the basis for the development of a variety of guides that target different geographic areas and user groups at varying levels of detail.

65. The Workshop agreed on a number of details including the content and structure of such guides as outlined in Table 1, as well as an overall strategy for the development of a work plan. This included the development of a project proposal for the development of deep-sea species identification material that will be presented to potential donors for their consideration.

## APPENDIX 1

### List of participants

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## APPENDIX 2

### Agenda

#### WEDNESDAY 2 DECEMBER 2009

09:00	<b>Arrival in FAO</b>	
09:00–10:30	Introduction <ul style="list-style-type: none"> <li>• Opening and administrative arrangements</li> <li>• Election of chair and designation of note takers for the different sessions</li> <li>• Agenda</li> <li>• Objectives of the meeting</li> </ul>	Merete Tandstad/ Johanne Fischer
10:30–11:00	<b>Coffee break</b>	
11:00–13:00	Relevant FAO Programmes <ul style="list-style-type: none"> <li>• Introduction to the FAO Programme on Deep-sea Fisheries in the High Seas</li> <li>• Preparation of species identification material under the FAO FishFinder Programme</li> <li>• Discussion</li> </ul>	Merete Tandstad/ Jessica Sander/ Johanne Fischer
13:00–14:00	<b>Lunch</b>	
14:00–15:30	Corals <ul style="list-style-type: none"> <li>• Global list of known corals (by region), vulnerable species, relevant references and list of experts</li> <li>• Draft recommendations for the production of identification guides</li> <li>• Discussion</li> </ul>	Marcelo Visentini Kitahara
15:30–16:00	<b>Coffee break</b>	
16:00–17:00	Sponges <ul style="list-style-type: none"> <li>• Global list of known sponges (by region), vulnerable species, relevant references and list of experts</li> <li>• Draft recommendations for the production of identification guides</li> <li>• Discussion</li> </ul>	Joana R. Bogalho Teixeira Xavier

**THURSDAY 3 DECEMBER 2009**

09:00–10:30	<b>Elasmobranchs and chimaeras</b> <ul style="list-style-type: none"> <li>• Global list of known (a) sharks/rays and (b) chimeras (by region), vulnerable species, relevant references and list of experts</li> <li>• Draft recommendations for the production of identification guides</li> <li>• Discussion</li> </ul>	Peter Kyne
10:30–11:00	<b>Coffee break</b>	
11:00–13:00	<b>Vulnerable deep-sea or bycatch fish species</b> <ul style="list-style-type: none"> <li>• Overview of work on deep-sea species at IEO</li> <li>• Overview of work on deep-sea species in the Southern Pacific</li> <li>• Discussion</li> </ul>	Luis José López Abellán/ Malcolm Clark
13:00–14:00	<b>Lunch</b>	
14:00–15:30	<b>Next steps</b> <ul style="list-style-type: none"> <li>• Elements for project proposal</li> <li>• Way forward</li> </ul>	Johanne Fischer/ Merete Tandstad/ Jessica Sanders
15:30–16:00	<b>Coffee break</b>	
16:00–17:00	<ul style="list-style-type: none"> <li>• Discussion</li> </ul>	Malcolm Clark

**FRIDAY, 4 DECEMBER 2009**

09:00–10:30	<ul style="list-style-type: none"> <li>• Report writing</li> </ul>	All
10:30–11:00	<b>Coffee break</b>	
11:00–13:00	<ul style="list-style-type: none"> <li>• Report writing</li> </ul>	All
14:00–14:00	<b>Lunch</b>	
14:00–15:30	<ul style="list-style-type: none"> <li>• Discussion and adoption of report and main recommendations</li> <li>• Closure</li> </ul>	Malcolm Clark
15:30–16:00	<b>Coffee break</b>	

## PART 2: EXPERT WORKSHOP BACKGROUND DOCUMENTS

### Information to assist preparation of a deep-sea species identification guide – sharks, batoids and chimaeras (class Chondrichthyes)

by Peter M. Kyne and Colin A. Simpfendorfer

Charles Darwin University and James Cook University

#### **Overview**

Following the FAO terms of reference, a global list (in relation to the 19 FAO major fishing areas) of known chondrichthyans: sharks, batoids and chimaeras (by region and at species level) that occur in the deep seas in depths from about 200 down to about 2 000 m, was compiled. From this, extract lists of those species (or species groups) that are known to be (a) vulnerable to fishing gear or other factors or/and (b) of commercial importance or/and (c) by-catch species, were also compiled. A relevant bibliography for regional and global identification guides, general deep-sea chondrichthyans and for each species group was assembled. Finally, draft recommendations for the production of identification guides are suggested.

#### **1. List of relevant chondrichthyans**

The deep-sea chondrichthyans are those whose distribution is confined to (or predominantly at) depths below 200 m, or those that spend a considerable part of their lifecycle below this depth. The maximum depth considered in this list is 2 000 m. Excluded from the list are many chondrichthyans, which have been recorded at depths of more than 200 m but which are predominantly shelf species. These species are recorded far less commonly or irregularly in the deep-sea.

The total number of extant, formally described chondrichthyan species currently stands at 1 144; this comprises 482 sharks, 671 batoids and 45 holocephalans (W.T, *personal communications*). Of the global fauna, 532 chondrichthyans can be considered to be deep-sea species (according to our definition; 200–2 000 m), representing 46.5 percent of the global total. The deep-sea fauna is divided between 254 sharks (52.7 percent of global), 237 batoids (35.3 percent of global) and 41 holocephalans (91.1 percent of global). All nine orders of elasmobranchs and the single holocephalan order are represented in the deep-sea (Table 1).

The bulk of the deep-sea chondrichthyan fauna is attributable to four main groups: (i) squaloid dogfishes (order Squaliformes), which represent 46.1 percent of the deep-sea shark fauna; (ii) scyliorhinid catsharks (order Carcharhiniformes, family Scyliorhinidae) (40.2 percent of the deep-sea shark fauna); (iii) skates (order Rajiformes, families Arhynchobatidae, Rajidae and Anacanthobatidae) (89.7 percent of the deep-sea batoid fauna); and (iv) holocephalans (order Chimaeriformes, families Rhinochimaeridae and Chimaeridae).

#### **2. Annotated global checklist of extant deep-sea chondrichthyans**

Within the checklist, where a ? follows a generic name, the placement of this species in that particular genera is questionable and thus tentative. Further research may result in placement in another genus, evaluation of a subgenera to generic level or designation of an altogether new genus. Where a ? follows a specific name, the validity of this species is questionable or the use of that specific name may be invalid. Distribution, habitat and depth information was collated from Compagno and Duffy (2003), global and regional field guides, specifically Last and Stevens (2009), Carpenter and Niem (1998, 1999), Carpenter (2002), Ebert (2003), Compagno *et al.* (2005) and White *et al.* (2006), as well as the primary literature, grey literature sources and consultation with experts. Habitat zones broadly follow Compagno *et al.* (2005).

The checklist covers the depth range 200–2 000 m. Few chondrichthyans are restricted to waters more than 2 000 m. In fact, only three skates have a minimum depth of ~2 000 m. These are: abyssal skate *Bathyraja ishiharai* Stehmann, 2005 (abyssal plains at ~2 300 m), fine-spined skate *Bathyraja*

*microtrachys* (Osburn and Nichols, 1916) (deep slopes and abyssal plains at 1 995–2 900 m) and pallid skate *Bathyraja pallida* (Forster, 1967) (deep slopes and abyssal plains at 2 200–3 280 m). These species are not treated here. No chondrichthyans have ever been observed or recorded in the hadal or hadopelagic zones (Compagno *et al.*, 2005; Priede *et al.*, 2006). Priede *et al.* (2006) hypothesized that the high-energy demands of chondrichthyans exclude them from the deepest habitat zones.

**Table 1**  
**Diversity of deep-sea chondrichthyan fishes by order and family**

Order	Family	Common Name	Number of Species
<b>Sharks</b>			
Hexanchiformes	Chlamydoselachidae	Frilled Sharks	2
	Hexanchidae	Sixgill and Sevengill Sharks	3
Squaliformes	Echinorhinidae	Bramble Sharks	2
	Squalidae	Dogfish Sharks	25
	Centrophoridae	Gulper Sharks	17
	Etomopteridae	Lanternsharks	42
	Somniosidae	Sleeper Sharks	16
	Oxynotidae	Roughsharks	5
	Dalatiidae	Kitefin Sharks	9
Squatiniformes	Squatinaeidae	Angelsharks	7
Pristiophoriformes	Pristiophoridae	Sawsharks	3
Heterodontiformes	Heterodontidae	Bullhead Sharks	1
Orectolobiformes	Parascylliidae	Collared Carpetsharks	2
Lamniformes	Odontaspidae	Sand Tiger Sharks	2
	Pseudocarchariidae	Crocodile Sharks	1
	Mitsukurinidae	Goblin Sharks	1
	Alopiidae	Thresher Sharks	1
	Cetorhinidae	Basking Sharks	1
Carcharhiniformes	Scyliorhinidae	Catsharks	102
	Proscylliidae	Finback Catsharks	3
	Pseudotriakidae	False Catsharks	2
	Triakidae	Houndsharks	6
	Carcharhinidae	Requiem Sharks	1
		<b>Subtotal - sharks</b>	<b>254</b>
<b>Batoids</b>			
Rajiformes	Rhinobatidae	Guitarfishes	1
	Narcinidae	Numbfishes	7
	Narkidae	Sleeper Rays	4
	Torpedinidae	Torpedo Rays	8
	Arhynchobatidae	Softnose Skates	75
	Rajidae	Hardnose Skates	116
	Anacanthobatidae	Legskates	21
	Plesiobatidae	Giant Stingarees	1
	Urolophidae	Stingarees	2
	Hexatrygonidae	Sixgill Stingrays	1
	Dasyatidae	Whiptail Stingrays	1
		<b>Subtotal - batoids</b>	<b>237</b>
<b>Holocephalans</b>			
Chimaeriformes	Rhinochimaeridae	Longnose Chimaeras	8
	Chimaeridae	Shortnose Chimaeras	33
		<b>Subtotal - holocephalans</b>	<b>41</b>
		<b>Total</b>	<b>532</b>

A detailed list of species found for FAO major fishing areas is presented in Appendix 2.

### **3. Species vulnerable to fishing gear or other factors**

All deep-sea chondrichthyans are vulnerable to fishing gear to some extent. However, despite being susceptible to gear they may not currently encounter it. This may occur for species that only occur at great depths (more than 1 000 m) where fishing occurs infrequently in most regions, or that occur in habitats that are rarely fished. Although great depths may currently provide protection from fishing to some species, there is a general trend towards fishing deeper as resources in shallower seas become more difficult to catch (Morato *et al.*, 2006). Thus, over time it would be expected that many deep-sea species will encounter increasing fishing pressure. This trend may be of concern for fisheries managers as recent work has shown that there is an inverse relationship between depth and population productivity in chondrichthyans (Simpfendorfer and Kyne, 2009). In general, deep-sea species have intrinsic rebound potentials approximately half of those from continental slope and epipelagic habitats. As a result deep-sea species will have relatively limited scope to provide sustainable catches by targeted fisheries or be very slow to recover if overfished. In addition, unlike teleost fishes, few shark species occur below 3 000 m (Priede *et al.*, 2006) so they do not have a deepwater refuge where fishing does not currently occur.

### **4. Species of commercial importance and those taken as bycatch**

While all deep-sea chondrichthyans are vulnerable to fishing gear, few are targeted by fisheries (Appendix 3). There are two main groups of species that are targeted in deep-sea fishing operations:

- Squalene yielding deep-sea sharks. Many species of deep-sea sharks contain large quantities of squalene in their livers. In particular members of the families Centrophoridae (e.g. *Centrophorus granulosus* [Bloch and Schneider, 1801], *C. harrissoni* [McCulloch, 1915], *C. isodon* [Chu, Meng and Liu, 1981], *C. squamosus* [Bonnaterre, 1788], *C. zeehani* [White, Ebert and Compagno, 2008], *Deania calcea* [Lowe, 1839]), Etmopteridae (e.g. *Etmopterus virens* [Bigelow, Schroeder and Springer, 1953]), Dalatiidae (e.g. *Dalatias licha* [Bonnaterre, 1788]) and Somniosidae (e.g. *Centrosynmus coelolepis* [Bocage and Capello, 1864], *Centroscyllium crepidater* [Bocage and Capello, 1864], *Proscymnodon plunketi* [Waite, 1909]). Squalene is a high value product and demand has driven the retention of bycatch and in some instances targeting of these species where significant catches are made. Locations where this has occurred include the Northeast Atlantic, Southeast Australia, Southeast Asia, Maldives, and Southern Africa. In many locations where these species have been targeted or regularly taken as bycatch significant declines in abundance have been observed after relatively short periods of fishing (Graham *et al.*, 2001; Jones *et al.*, 2005).
- Skates. Skate flesh (often sold as “wings”) is considered of high quality and a range of species are taken as both target and bycatch in fisheries. Most are caught by deepwater trawl fisheries, but some are targeted by longlines. Significant catches are mostly taken in temperate areas, including in the Northeast, Northwest and Southwest Atlantic, Alaska, and Chile. The most commonly utilized groups include members of the families Rajidae (e.g. *Amblyraja radiata* [Donovan, 1808], *Dipturus* spp., *Raja* spp.) and Arhynchobatidae (e.g. *Bathyraja* spp.). In the Northeast Atlantic where skates have been a common target and bycatch species over many decades, populations of less productive species have declined significantly, and have been extirpated from substantial parts of their ranges (Brander, 1981; Dulvy and Reynolds, 2002), suggesting that intensive fisheries may not be sustainable for all species. Declines have been documented in other regions also, including Southeast Australia (Graham *et al.*, 2001) and the Northwest Atlantic (Devine *et al.*, 2006).

While these two groups represent the majority of catch of deep-sea chondrichthyans, many species from other groups are also captured as bycatch in fisheries (Appendix 3). For some species or groups the vast majority of individuals are discarded because of the lack of commercial value due to their small size, low quality products, lack of markets, difficulty in processing or high levels of mercury. This includes groups within the dogfish sharks (Squaliformes) and the catsharks (Scyliorhinidae) such

as some species of the genera *Etmopterus* and *Apristurus*, respectively. For many other species markets exist for flesh and livers and they are retained when they are taken as bycatch in other deep-sea fisheries. It should also be recognised that, although at present unrecorded, additional species of deep-sea chondrichthyan may also be taken as bycatch.

## 5. Recommendation for the production of the identification guide

The guide should be comprehensive in its coverage of species. For the chondrichthyans, this includes 532 deep-sea species across the three major groups (sharks, batoids and chimaeras).

There would need to be some consideration however, of dealing with other species not considered by definition to be deep-sea species, but which have been recorded in depths more than 200 m. This includes many chondrichthyans which have been recorded at depths of more than 200 m but which are predominantly species of the shelf. These species are recorded far less commonly or irregularly in the deep-sea and include such examples as spiny dogfish *Squalus acanthias* Linnaeus, 1758 (a shelf species which has been recorded exceptionally to 1 446 m) and many shelf skates whose bathymetrical distribution extends to the upper slope, including sandy skate *Leucoraja circularis* (Couch, 1838), blonde skate *Raja brachyura* (Lafont, 1873), thornback skate *R. clavata* Linnaeus, 1758, clearnose skate *R. eglanteria* Bosc, 1802, brown skate *R. miraleatus* Linnaeus, 1758 and rough skate *R. radula* Delaroche, 1809.

Some particular species which may require justification as to their absence from this report are the broadnose sevengill shark *Notorynchus cepedianus* (Péron, 1807), which although recorded to 570 m is primarily a coastal inshore shark of temperate waters; the sparseshark *Scymnodalatias oligodon* (Kukuyev and Konovalenko, 1988) and the largetooth cookiecutter shark *Isistius plutodus* (Garrick and Springer, 1964), although both occur over very deep water (to 4 000 m), they have both only ever been recorded in the epipelagic zone (-200 m depth in the water column) (it should be noted though that these species are potential vertical migrators and may also occur in the mesopelagic and bathypelagic zones); the megamouth shark *Megachasma pelagios* (Taylor, Compagno and Struhsaker, 1983), a coastal, shelf and epipelagic species recorded from the pelagic zone to depths of 166 m over water to 4 600 m depth, but never recorded from the deep-sea. One last notable species is the great white shark *Carcharodon carcharias* (Linnaeus, 1758), a coastal, inshore, shelf and epipelagic offshore species known to undertake considerable migrations. Great whites have rarely been recorded from the continental slope (Compagno, 2001), and thus in the deep-sea, and a reported capture at 1 280 m (Bigelow and Schroeder, 1948; Compagno, 2001) is doubtful (Bonfil *et al.*, 2005). However, more recent research of a shark tagged off South Africa has shown that during transoceanic migration great whites undertake periodic deep dives to depths of up to 980 m (Bonfil *et al.*, 2005). It should be noted that this depth is the sensor limit for the tag employed and so it is possible that the tagged shark dived to depths in excess of 980 m. While deep dives were regular in occurrence during its migration, with the shark spending 18 percent of the time at depths of 500–750 m, a far greater amount of time (61 percent) was spent at the surface in water – 0.5 m deep (Bonfil *et al.*, 2005). Thus, given the species' primarily coastal and epipelagic occurrence, it is not considered a deep-sea species.

- The guide should be strong on field identification. This should include accurate keys to species for each family (tested in the field and using museum collections). Keys are very valuable in the field identification of species.
- An introduction to each family should be included. These could follow the style of the FAO species identification guides of fishery purposes (e.g. the living marine resources of the Western Central Pacific: Carpenter and Niem, 1998; Carpenter and Niem, 1999).
- Each species account should include key field identification features. This should be different from any taxonomic description, and should highlight those characters which can be used to accurately identify a species, and separate it from all other species.
- The inclusion in each species account of a note on 'Similar Species' would prove valuable. This will allow the user to cross check their specimen with those species with which it is most likely to be misidentified.

- Each species account should include a distribution map, and a brief note on distribution, habitat and depth range. This is essential information to determine if a species' occurrence overlaps, or potentially overlaps with known fishing operations. Worthwhile would be an estimation (as accurate as possible using geographic information system [GIS]) of a species' extent of occurrence and/or area of occupancy. This information is vital in conservation planning.
- Each species account should include information on maximum size, size at maturity and size at birth, if known. A comment should be made if this, the most basic of information, is unknown for a species. This assists in highlighting knowledge gaps.
- Each species should include available biological information. However, there is no need to go into great detail, although summaries can be provided (fecundity, reproductive cycle, age and growth are particularly important). Information on diet and feeding ecology is not so vital for an identification guide. The above mentioned biological information is lacking for the vast majority of species (it has largely been summarised for all species in Kyne and Simpfendorfer (2007) and Kyne and Simpfendorfer (2010)).
- Each species account of targeted species (Appendix 3) should include information on fisheries they are captured in and any trend data, where available. For bycatch species, information on at least the type of fishery (trawl, longline etc.) should be provided. More general fishery information can be provided in the introduction to families.
- Each species account should include its conservation status according to the IUCN Red List of Threatened Species. The vast majority of chondrichthyans have now been assessed against the IUCN Red List Categories and Criteria and so this data is readily available (IUCN, 2009). It serves to highlight not only those threatened species, but also the large proportion of Data Deficient species, drawing attention to the poorly-known status of taxa, knowledge gaps, and information required to accurately assess conservation status.
- Each species account, or at least the family introduction should include information on taxonomic uncertainty. This can encourage the provision of material required for taxonomic resolution. This section should also include the results of molecular taxonomy studies that are helping to resolve the placement and relationships between many taxa.
- The guide should contain preliminary sections discussing issues relating to chondrichthyans and the deep-sea, management considerations, vulnerabilities, general fishery trends, habitat considerations, etc. (for example, the limited area of available habitat for species occurring only on the continental slope).
- The guide could be undertaken in two editions, a global guide which encompasses all regions and is comprehensive, and regional guides specific to ocean regions. These regional guides are more practical for users working in the field.

## **6. Potential difficulties/technical considerations**

There has been a recent resurgence in chondrichthyan taxonomy and more than a third of known species have been described in the last 30 years (Last, 2007), while in the last five years there has been considerable effort placed into describing new fauna and resolving taxonomic issues. However, there is still considerable taxonomic work required for the three major faunal groups. The total number of known species is ever increasing as exploratory and taxonomic work ensues and there is considerable material still awaiting description (at the time of writing, at least 37 sharks, 23 batoids and 5 chimaeras are known to represent new species). These numbers of species illustrate two important points. Firstly, the overall lack of knowledge of the deep-sea fauna at even the most basic (i.e. taxonomic) level. Secondly, that the deep-sea chondrichthyan fauna is far from fully documented. For example, a recent joint Australian/Indonesian project monitoring the landings of chondrichthyans at various fish markets in Indonesia has revealed several previously unknown deep-sea taxa amongst a larger number of new species (White *et al.*, 2006). The majority of fishing in Indonesia is still focused on coastal and pelagic resources, with relatively little deep-sea fishing activity. With continuing exploitation of traditional resources, fishing activities will likely move to deeper water around Indonesia, and the discovery of new fauna will likely continue. The checklist of known species will

also continue to grow as material is gathered and examined from exploratory research cruises to previously poorly surveyed ocean regions. The 2003 NORFANZ voyage<sup>1</sup>, surveying the seamounts and abyssal plains around the Lord Howe and Norfolk Ridges in the Western Pacific is one such example (Last, 2007). Priede *et al.* (2006) note, however, that the discovery of new species of chondrichthyans at considerable depth (more than 3 000 m) is unlikely given present bathymetrical distributions and the apparent inability of chondrichthyans to exploit the deepest habitat zones.

The systematics and inter-relationships of several groups of deep-sea chondrichthyans remains unresolved and as these groups are reviewed by taxonomists, the total number of species will ultimately change. One of the more taxonomically complex groups is the gulper sharks of the genus *Centrophorus*. A review of some of the Indo-West Pacific gulper shark fauna highlighted that many species traditionally considered to be wide-ranging with global distributions are likely species-complexes of regional endemics (White *et al.*, 2008; W.T. White, personal communications). Forms in the Atlantic likely represent distinct species to those in the Indo-West Pacific, for example, the lowfin gulper shark *Centrophorus lusitanicus* Bocage and Capello, 1864. At even a more localised scale, the Indo-West Pacific longnose gulper shark *C. harrissoni* McCulloch, 1915 was found to be distinct between the east and west coast of Australia, representing separate species (White *et al.*, 2008). Resolution of such issues will have a profound effect on conservation and management of gulper sharks, amongst the most exploited and vulnerable of the deep-sea sharks.

Another deep-sea group that has undergone recent taxonomic revision are the catsharks of the genus *Apristurus*. This large group has a complex and confused taxonomic history in part due to poor descriptions, lack of comparative material, poorly-defined morphometric characters and the large number of synonyms and undescribed species (Nakaya *et al.*, 2005; Sato, 2005). Resolution of the genus relied on an international collaboration of experts. These studies incorporated new fauna, some which proved to be limited range endemics (see papers in Last *et al.*, 2008).

Finally, the softnose and hardnose skates of the families Arhynchobatidae and Rajidae are considered to be a morphologically conservative, yet highly diverse group (McEachran and Dunn, 1998). Indeed, they are two of the largest families of chondrichthyans, yet the phylogeny and inter-relationships of the skates remain unresolved.

The resolution of taxonomic issues, and the description of known new species is a priority, and a push to complete this work is required. Keeping abreast of the description of new species and taxonomic changes is paramount to ensuring that a deep-sea species identification guide be comprehensive and up to date.

For the sharks, the Sharks of the World catalogues (FAO Species Catalogue for Fisheries Purposes), in particular Volume 1 (Hexanchiformes, Squaliformes, Squatiniformes and Pristiophoriformes) presently in preparation by David A. Ebert will serve as a valuable compilation of information on all deep-sea sharks. A new deep-sea identification guide should align closely to these catalogues. For the batoids and the holocephalans, such global catalogues do not exist and thus there is no baseline to measure against. The preparation of information on deep-sea batoids and holocephalans will likely prove more time consuming than the sharks.

The status of the FAO Batoid Catalogue needs to be considered. This guide is a priority and it may be difficult to convince some batoid taxonomists to work on a new guide for the deep-sea fauna, while the batoid catalogue remains unfinished.

Expertise is limited in some deep-sea groups, with a relatively small pool of taxonomist experts, many of which have been involved, or are being involved in the development of other guides and catalogues. This may limit their ability to be able to commit to a new guide, if other commitments are ongoing.

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<sup>1</sup> [www.environment.gov.au/coasts/discovery/voyages/norfanz/index.html](http://www.environment.gov.au/coasts/discovery/voyages/norfanz/index.html)

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## APPENDIX 1

### Annotated global checklist of extant deep-sea chondrichthyans

**Class Chondrichthyes.**

**Subclass Holocephali.**

**Order Chimaeriformes. Modern Chimaeras.**

#### Family Rhinochimaeridae. Longnose Chimaeras.

***Harriotta haeckeli* (Karrer, 1972).** Smallspine spookfish.

Patchy in the Southern Ocean and Atlantic. Abyssal plains and deep-sea troughs. 1 114-2 603 m.

***Harriotta raleighana* (Goode and Bean, 1895).** Narrownose or longnose chimaera, bentnose rabbitfish or bigspine spookfish.

Wide-ranging but patchy in the Indian, Pacific and Atlantic. Upper to deep slope, abyssal plains and seamounts. 380-2 600 m.

***Neoharriotta carri* (Bullis and Carpenter, 1966).** Dwarf sicklefin chimaera.

Western Central Pacific: southern Caribbean. Upper slope. 240-600 m.

***Neoharriotta pinnata* (Schnakenbeck, 1931).** Sickletail chimaera.

Eastern Central and Southeast Atlantic: West Africa. Outermost shelf and upper slope. 200-470 m.

***Neoharriotta pumila* (Didier and Stehmann, 1996).** Arabian sicklefin chimaera.

Western Indian: Arabian Sea and Gulf of Aden. Outer shelf and upper to mid slope. 100-1 120 m.

***Rhinochimaera africana* (Compagno, Stehmann and Ebert, 1990).** Paddlenose chimaera or spookfish.

Patchy in the Indo-West Pacific and Southeast Atlantic. Upper to mid slope and seamounts. 550-1 450 m.

***Rhinochimaera atlantica* (Holt and Byrne, 1909).** Spearnose chimaera or straightnose rabbitfish.

Wide-ranging in the Atlantic. Upper to mid slope. 500-1 500 m.

***Rhinochimaera pacifica* (Mitsukuri, 1895).** Pacific spookfish or knifenose chimaera.

Patchy in the Indo-West Pacific and the Southeast Pacific. Outermost shelf, upper to mid slope, deep-sea troughs, deep-sea plateaus and seamounts. 191-1 290 m (mostly >700 m).

#### Family Chimaeridae. Shortnose Chimaeras.

***Chimaera argiloba* (Last, White and Pogonoski, 2008).** Whitefin chimaera.

Eastern Indian: western Australia. Upper slope. 370-520 m.

***Chimaera cubana* (Howell-Rivero, 1936).** Cuban chimaera.

Western Central Atlantic: Caribbean. Upper slope. 234-450 m.

***Chimaera fulva* (Didier, Last and White, 2008).** Southern chimaera.

Eastern Indian and Southwest Pacific: southern Australia. Mid slope. 780-1 095 m.

***Chimaera jordani* (Tanaka, 1905).** Jordan's chimaera.

Confirmed from the Northwest Pacific (Japan) and Western Indian, but probably more wide-ranging. Upper to deep slope. 383-1 600 m.

***Chimaera lignaria* (Didier, 2002).** Giant, purple or carpenter's chimaera.

Primarily Southwest Pacific: southern Australia and New Zealand. Upper to deep slope, deep-sea plateaus and seamounts. 400-1 800 m (mostly >800 m).

***Chimaera macrospina* (Didier, Last and White, 2008).** Longspine chimaera.

Eastern Indian and Western Pacific: Australia. Upper to mid slope. 435-1 300 m (mostly >800 m).

*Chimaera monstrosa* (Linnaeus, 1758). Rabbitfish.

Wide-ranging in the Northeast Atlantic including the Mediterranean. Shelf and upper to mid slope. 50-1 000 m (mostly 300-500 m).

*Chimaera obscura* (Didier, Last and White, 2008). Shortspine chimaera.

Western Central and Southwest Pacific: eastern Australia. Upper to mid slope. 450-1 000 m.

*Chimaera owstoni* (Tanaka, 1905). Owston's chimaera.

Northwest Pacific: Japan. Upper to mid slope. 500-1 200 m.

*Chimaera panthera* (Didier, 1998). Leopard or roundfin chimaera.

Southwest Pacific: New Zealand. Upper to mid slope, deep-sea rises, submarine ridges and seamounts. 327-1 020 m.

*Chimaera phantasma* (Jordan and Snyder, 1900). Silver chimaera

Northwest Pacific: Japan and Taiwan. Shelf and upper to mid slope. 20-962 m. Includes the junior synonym *Chimaera pseudomonstrosa* Fang and Wang, 1932.

*Hydrolagus affinis* (Capello, 1867). Atlantic chimaera or smalleyed rabbitfish.

Wide-ranging in the North Atlantic. Upper to deep slope, abyssal plains and seamounts. 300-2 909 m (mostly >1 000 m).

*Hydrolagus africanus* (Gilchrist, 1922). African chimaera.

Western Indian and probably Southeast Atlantic: southern Africa. Upper to mid slope. 300-1 300 m (mostly 421-750 m).

*Hydrolagus alberti* (Bigelow and Schroeder, 1951). Gulf chimaera.

Western Central Atlantic: Caribbean and the Gulf of Mexico. Upper to mid slope. 348-1 100 m.

*Hydrolagus albus* (Quaranta, Didier, Long and Ebert, 2006). Whitespot ghostshark.

Southeast Pacific: Galapagos Islands. Upper to mid slope. 600-900 m.

*Hydrolagus barbouri* (Garman, 1908). Ninespot chimaera.

Northwest Pacific: Japan. Upper to mid slope. 250-1 100 m (most common 600-800 m).

*Hydrolagus bemisi* (Didier, 2002). Pale ghostshark.

Southwest Pacific: New Zealand. Upper to mid slope, deep-sea plateaus and rises. 400-1 100 m (mostly 500-700 m).

*Hydrolagus colliei* (Lay and Bennett, 1839). Spotted ratfish.

Northeast and Eastern Central Pacific: Alaska to Mexico. Shelf and upper to mid slope. 0-971 m.

*Hydrolagus homonycterus* (Didier, 2008). Black ghostshark.

Primarily Southwest Pacific: southern Australia and New Zealand. Mid slope and seamounts. 866-1 447 m.

*Hydrolagus lemures* (Whitley, 1939). Blackfin ghostshark.

Eastern Indian and Western Central and Southwest Pacific: Australia. Outer shelf and upper slope. 146-700 m.

*Hydrolagus lusitanicus* (Moura, Figueiredo, Machado, Almeida and Gordo, 2005).

Northeast Atlantic: Portugal. Deep slope. 1 600 m.

*Hydrolagus macrourus* (de Buen, 1959). Bigeye chimaera.

Southeast Pacific: Chile. Habitat data not available.

*Hydrolagus marmoratus* (Didier, 2008). Marbled ghostshark.

Western Central and Southwest Pacific: eastern Australia. Upper to mid slope. 548-995 m.

*Hydrolagus matallanasi* (Soto and Vooren, 2004). Striped Rabbitfish.

Southwest Atlantic: southern Brazil. Upper slope. 416-736 m.

*Hydrolagus mcoskeri* (Barnett, Didier, Long and Ebert, 2006). Galapagos ghostshark.

Southeast Pacific: Galapagos Islands. Upper slope. 396-506 m.

*Hydrolagus melanophasma* (James, Ebert, Long and Didier, 2009). Eastern Pacific black ghostshark

Eastern Central Pacific: California (United States) to Gulf of California (Mexico). Shelf and upper to deep slope. 31-1 667 m.

*Hydrolagus mirabilis* (Collett, 1904). Large-eyed rabbitfish or spectral chimaera.

Wide-ranging in the Eastern Atlantic and also in the Western Central Atlantic. Upper to deep slope. 450-1 933 m (mostly >800 m).

***Hydrolagus mitsukurii* (Dean, 1904).** Mitsukurii's chimaera.

Northwest and Western Central Pacific: Japan to the Philippines. Upper to mid slope. 325-770 m. Includes the junior synonym *Hydrolagus deani* (Smith and Radcliffe, 1912).

***Hydrolagus novaezealandiae* (Fowler, 1910).** Dark or New Zealand ghostshark.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 25-950 m (most common 150-500 m).

***Hydrolagus ogilbyi* (Waite, 1898).** Ogilby's ghostshark.

Eastern Indian and Southwest Pacific: southeastern Australia. Outer shelf and upper slope. 120-350 m.

***Hydrolagus pallidus* (Hardy and Stehmann, 1990).** Pale chimaera.

Scattered records in the Northeast Atlantic. Mid to deep slope and deep-sea troughs. 1 200-2 650 m.

***Hydrolagus purpureascens* (Gilbert, 1905).** Purple chimaera.

Northwest Pacific (Japan) and Eastern Central Pacific (Hawaii). Mid to deep slope, deep-sea troughs and seamounts. 920-1 130 m (Japan), 1 750-1 951 m (Hawaii). Includes the probable junior synonym *Hydrolagus eidolon* (Jordan and Hubbs, 1925).

***Hydrolagus trolli* (Didier and Séret, 2002).** Pointy-nosed blue chimaera.

Patchy in the Southwest and Western Central Pacific. Upper to mid slope and seamounts. 610-2 000 m (mostly >1 000 m).

## Subclass Elasmobranchii.

### Superorder Squalomorphii. Squalomorph Sharks.

#### Order Hexanchiformes. Cow and Frilled Sharks.

##### Family Chlamydoselachidae. Frilled Sharks.

***Chlamydoselachus africana* (Ebert and Compagno, 2009).** Southern African frilled shark.

Southeast Atlantic and Western Indian: southern Africa. Upper to mid slope. 300-1 400 m.

***Chlamydoselachus anguineus* (Garman, 1884).** Frilled shark.

Wide-ranging but patchy in temperate and tropical waters of the Atlantic and Indo-Pacific. Shelf and upper to mid slope. 50-1 500 m.

##### Family Hexanchidae. Sixgill and Sevengill Sharks.

***Heptranchias perlo* (Bonnaterre, 1788).** Sharpnose sevengill shark or perlon.

Wide-ranging but patchy in temperate and tropical waters of the Atlantic and Indo-Pacific. Shelf (occasional) and upper to mid slope. 27-1 000 m.

***Hexanchus griseus* (Bonnaterre, 1788).** Bluntnose sixgill shark.

Wide-ranging but patchy in temperate and tropical waters of the Atlantic and Indo-Pacific. Shelf, upper to deep slope, submarine ridges and seamounts. Mainly deepwater, young inshore in cold water. 0-2 490 m (500-1 100 m usual).

***Hexanchus nakamurai* (Teng, 1962).** Bigeye sixgill shark.

Wide-ranging but patchy in warm-temperate and tropical waters of the Atlantic and Indo-Pacific. Shelf (occassional) and upper slope. 90-621 m.

#### Order Squaliformes. Dogfish Sharks.

##### Family Echinorhinidae. Bramble Sharks.

***Echinorhinus brucus* (Bonnaterre, 1788).** Bramble shark.

Wide-ranging but patchy in the Atlantic and Indo-Pacific. Shelf (occassional) and upper to mid slope. 18-900 m.

*Echinorhinus cookei* (Pietschmann, 1928). Prickly shark.

Patchy in the Eastern, Central and Western Pacific. Shelf and upper to mid slope and seamounts. 11-1 100 m.

### Family Squalidae. Dogfish Sharks.

*Cirrhigaleus asper* (Merrett, 1973). Roughskin spurdog.

Wide-ranging but patchy in warm-temperate and tropical waters of the Atlantic and Indo-Pacific. Shelf and upper slope. 73-600 m.

*Cirrhigaleus australis* (White, Last and Stevens, 2007). Southern mandarin dogfish.

Eastern Indian and Southwest Pacific: southern Australia and New Zealand. Upper slope. 360-640 m.

*Cirrhigaleus barbifer* (Tanaka, 1912). Mandarin dogfish.

Patchy in the Western Pacific. Outer shelf and upper slope. 146-640 m. May represent a species complex.

*Squalus albifrons* (Last, White and Stevens, 2007). Eastern highfin spurdog.

Western Central and Southwest Pacific: eastern Australia. Outer shelf and upper slope. 131-450 m.

*Squalus altipinnis* (Last, White and Stevens, 2007). Western highfin spurdog.

Eastern Indian: western Australia. Upper slope. ~300 m.

*Squalus blainville* (Risso, 1826). Longnose spurdog.

Nominally from the Eastern Atlantic and the Mediterranean, but not well defined due to confusion with other species and taxonomic issues. Shelf and upper slope. 16-440 m. Considerable taxonomic issues (records from elsewhere in the Atlantic and the Indo-Pacific based in part on *S. mitsukurii* or close relatives).

*Squalus brevirostris* (Tanaka, 1917). Japanese shortnose spurdog.

Northwest Pacific: Japan and Taiwan. Habitat information not available. Tentatively placed on checklist as distinct from *S. megalops*.

*Squalus bucephalus* (Last, Séret and Pogonoski, 2007). Bighead spurdog.

Western Pacific: Norfolk Ridge and New Caledonia. Upper to mid slope and submarine ridges. 448-880 m.

*Squalus chloroculus* (Last, White and Motomura, 2007). Greeneye spurdog.

Southwest Pacific and Eastern Indian: southern Australia. Upper to mid slope. 216-1 360 m.

*Squalus crassispinus* (Last, Edmunds and Yearsley, 2007). Fatspine spurdog.

Eastern Indian: northwestern Australia. Outermost shelf and upper slope. 187-262 m.

*Squalus cubensis* (Howell-Rivero, 1936). Cuban dogfish.

Warm-temperate waters of the Western Atlantic. Shelf and upper slope. 60-380 m.

*Squalus edmundsi* (White, Last and Stevens, 2007). Edmund's spurdog.

Eastern Indian: western Australia and Indonesia. Upper to mid slope. 204-850 m (mostly 300-500 m).

*Squalus grahami* (White, Last and Stevens, 2007). Eastern longnose spurdog.

Western Central and Southwest Pacific: eastern Australia. Outer shelf and upper slope. 148-504 m (mainly 220-450 m).

*Squalus griffini* (Phillipps, 1931). Northern spiny dogfish.

Southwest Pacific: New Zealand. Shelf and upper slope. 37-616 m.

*Squalus hemipinnis* (White, Last and Yearsley, 2007). Indonesian shortsnot spurdog

Eastern Indian: Indonesia. Upper slope.

*Squalus japonicus* (Ishikawa, 1908). Japanese spurdog.

Northwest and Western Central Pacific: East Asia. Outer shelf and upper slope. 120-340 m.

*Squalus lalannei* (Baranes, 2003). Seychelles spurdog.

Western Indian: Seychelles. Mid slope. 1 000 m.

*Squalus megalops* (Macleay, 1881). Shortnose spurdog.

Widespread in temperate and warm-temperate waters of the Indo-West Pacific and Eastern Atlantic. Shelf and upper slope. 0-732 m. May represent a species complex.

*Squalus melanurus* (Fourmanoir, 1979). Blacktail spurdog.

Western Central Pacific: New Caledonia. Shelf and upper slope. 34-480 m.

*Squalus mitsukurii* (Jordan and Snyder, in Jordan and Fowler, 1903). Shortspine spurdog.

Wide-ranging, but patchy in temperate and tropical waters of the Atlantic and Indo-Pacific, but not well defined due to taxonomic issues. Shelf and upper to mid slope, submarine ridges and seamounts. 4-954 m (100-500 m usual). Resolution of considerable taxonomic issues is ongoing. Tentatively includes the junior synonym *Squalus probatovi* Myagkov and Kondyrin, 1986.

*Squalus montalbani* (Whitley, 1931). Philippine spurdog.

Patchy in the Eastern Indian and Western Pacific. Upper slope. 295-670 m.

*Squalus nasutus* (Last, Marshall and White, 2007). Western longnose spurdog.

Patchy in the Eastern Indian and Western Pacific. Upper to mid slope. 300-850 m (mainly 300-400 m).

*Squalus notocaudatus* (Last, White and Stevens, 2007). Bartail spurdog.

Western Central Pacific: northeastern Australia. Upper slope. 225-454 m.

*Squalus rancureli* (Fourmanoir, 1978). Cyrano spurdog.

Western Central Pacific: Vanuatu. Upper slope. 320-400 m.

*Squalus raoulensis* (Duffy and Last, 2007). Kermadec spiny dogfish.

Southwest Pacific: Kermadec Ridge. Submarine ridges. 320 m.

### Family Centrophoridae. Gulper Sharks.

*Centrophorus acus* (Garman, 1906). Needle dogfish.

Patchy in the Eastern Indian and Western Pacific. Outer shelf and upper to mid slope. 150-950 m (mostly >200 m, possibly to 1 786 m).

*Centrophorus atromarginatus* (Garman, 1913). Dwarf gulper shark.

Patchy in the Indo-West Pacific, but not well defined. Outer shelf and upper slope. 150-450 m.

*Centrophorus granulosus* (Bloch and Schneider, 1801). Gulper shark.

Wide-ranging but patchy in the Atlantic and Indo-Pacific, but some records may represent additional species. Shelf and upper to mid slope. 50-1 440 m (mostly 200-600 m). Resolution of taxonomic issues ongoing and as such occurrence and distribution not well defined.

*Centrophorus harrissoni* (McCulloch, 1915). Longnose gulper shark.

Patchy in the Western Central and Southwest Pacific. Upper to mid slope and submarine ridges. 220-1 050 m.

*Centrophorus isodon* (Chu, Meng and Liu, 1981). Blackfin gulper shark.

Patchy in the Indo-West Pacific. Mid slope. 760-770 m.

*Centrophorus lusitanicus* (Bocage and Capello, 1864). Lowfin gulper shark.

Patchy in the Eastern Atlantic and Indo-West Pacific. Upper to mid slope. 300-1 400 m (mostly 300-600 m). Indo-West Pacific form may represent a separate species (*Centrophorus* cf. *lusitanicus* in White et al. 2006).

*Centrophorus moluccensis* (Bleeker, 1860). Smallfin gulper shark.

Patchy in the Indo-West Pacific. Outer shelf and upper to mid slope. 125-820 m. Resolution of taxonomic issues ongoing. May represent a species complex.

*Centrophorus niaukang* (Teng, 1959.) Taiwan gulper shark.

Patchy in the Eastern Indian and Western Pacific. Outer shelf and upper to mid slope. 98-~1 000 m.

*Centrophorus seychellorum* (Baranes, 2003). Seychelles gulper shark.

Western Indian: Seychelles. Mid slope. 1 000 m.

*Centrophorus squamosus* (Bonnaterre, 1788). Leafscale gulper shark.

Wide-ranging, but patchy in the Eastern Atlantic and Indo-West Pacific. Upper to deep slope and abyssal plains. 230-3 280 m.

***Centrophorus tessellatus* (Garman, 1906).** Mosaic gulper shark.

Scattered locations in the Western Atlantic and Pacific, but some records provisional. Upper slope. 260–730 m. Taxonomic issues mean that occurrence and distribution are not well defined and as such many records are provisional.

***Centrophorus westraliensis* (White, Ebert and Compagno, 2008).** Western gulper shark.

Eastern Indian: western Australia. Upper to mid slope. 616–750 m.

***Centrophorus zeehaani* (White, Ebert and Compagno, 2008).** Southern dogfish.

Eastern Indian and Southwest Pacific: southern Australia. Upper slope. 208–701 m (usually >400 m).

***Deania calcea* (Lowe, 1839).** Birdbeak or shovelnose dogfish.

Wide-ranging in the Eastern Atlantic and Indo-West Pacific. Shelf and upper to mid slope. 70–1 470 m (usually 400–900 m). Indonesian form may represent a distinct species (*Deania* cf. *calcea* in White *et al.* 2006).

***Deania hystricosum* (Garman, 1906).** Rough longnose dogfish.

Scattered locations in the Eastern Atlantic and Western Pacific. Upper to mid slope. 470–1 300 m.

***Deania profundorum* (Smith and Radcliffe, 1912).** Arrowhead dogfish.

Scattered locations in the Atlantic and Indo-West Pacific. Upper to deep slope. 275–1 785 m.

***Deania quadrispinosum* (McCulloch, 1915).** Longsnout dogfish.

Southern regions of the Eastern Atlantic and Indo-West Pacific. Outer shelf and upper to mid slope. 150–1 360 m (usually <400 m).

### Family Etmopteridae. Lanternsharks.

***Aculeola nigra* (de Buen, 1959).** Hooktooth dogfish.

Southeast Pacific: Peru to Chile. Outer shelf and upper slope. 110–735 m.

***Centroscyllium excelsum* (Shirai and Nakaya, 1990).** Highfin dogfish.

Northwest Pacific: Emperor Seamount Chain. Seamounts. 800–1 000 m.

***Centroscyllium fabricii* (Reinhardt, 1825).** Black dogfish.

Wide-ranging in temperate waters of the Atlantic. Outer shelf and upper to deep slope. 180–1 600 m (usually >275 m, probably to 2 250 m).

***Centroscyllium granulatum* (Günther, 1887).** Granular dogfish.

Southeast Pacific: Chile. Upper slope. 300–500 m.

***Centroscyllium kamoharai* (Abe, 1966).** Bareskin dogfish.

Indo-West Pacific: Japan, Australia and possibly the Philippines. Upper to mid slope. 500–1 200 m (mostly >900 m).

***Centroscyllium nigrum* (Garman, 1899).** Combtooth dogfish.

Patchy in the Central and Eastern Pacific. Upper to mid slope. 400–1 143 m.

***Centroscyllium ornatum* (Alcock, 1889).** Ornate dogfish.

Northern Indian: Arabian Sea and the Bay of Bengal. Upper to mid slope. 521–1 262 m.

***Centroscyllium ritteri* (Jordan and Fowler, 1903).** Whitefin dogfish.

Northwest Pacific: Japan. Upper to mid slope and seamounts. 320–1 100 m.

***Etmopterus baxteri* (Garrick, 1957).** New Zealand lanternshark.

Southern regions of the Eastern Atlantic and Indo-West Pacific. Upper to mid slope. 250–1 500 m. Occurrence and distribution not well defined in the Southeast Atlantic and Western Indian.

***Etmopterus bigelowi* (Shirai and Tachikawa, 1993).** Blurred smooth lanternshark.

Wide-ranging but patchy in the Atlantic and Indo-Pacific. Outer shelf, upper to mid slope, submarine ridges and seamounts. 163–1 000 m.

***Etmopterus brachyurus* (Smith and Radcliffe, 1912).** Shorttail lanternshark.

Scattered in the Indo-West Pacific: Japan, the Philippines and Australia. Upper slope. 400–610 m. References to the species off southern Africa refer to the as yet undescribed sculpted lanternshark.

***Etmopterus bullisi* (Bigelow and Schroeder, 1957).** Lined lanternshark.

Western Central Atlantic: Caribbean. Upper to mid slope. 275–824 m (mostly >350 m).

*Etmopterus burgesii* (Schaaf-da Silva and Ebert, 2006). Broad-snout lanternshark.

Northwest Pacific: Taiwan. Slope. >300 m.

*Etmopterus carteri* (Springer and Burgess, 1985). Cylindrical lanternshark.

Western Central Atlantic: Caribbean coast of Colombia. Upper slope. 283–356 m.

*Etmopterus caudistigmus* (Last, Burgess and Séret, 2002). New Caledonia tailspot lanternshark

Western Central Pacific: New Caledonia. Upper to mid slope. 638–793 m.

*Etmopterus decacuspidatus* (Chan, 1966). Combtooth lanternshark.

Northwest Pacific: South China Sea. Upper slope. 512–692 m.

*Etmopterus dianthus* (Last, Burgess and Séret, 2002). Pink lanternshark.

Western Central Pacific: Australia and New Caledonia. Upper to mid slope. 700–880 m.

*Etmopterus dislineatus* (Last, Burgess and Séret, 2002). Lined lanternshark.

Western Central Pacific: Australia. Upper slope. 590–700 m.

*Etmopterus evansi* (Last, Burgess and Séret, 2002). Blackmouth lanternshark.

Eastern Indian: Australia and Arafura Sea. Shoals and reefs on the upper slope. 430–550 m.

*Etmopterus fusus* (Last, Burgess and Séret, 2002). Pygmy lanternshark.

Eastern Indian: Australia. Upper slope. 430–550 m.

*Etmopterus gracilispinis* (Krefft, 1968). Broadband lanternshark.

Wide-ranging but patchy in the Western Atlantic and off southern Africa. Outer shelf and upper to mid slope. 70–1 000 m.

*Etmopterus granulosus* (Günther, 1880). Southern lanternshark.

Southwest Atlantic and Southeast Pacific: southern South America. Upper slope. 220–637 m.

*Etmopterus hillianus* (Poey, 1861). Caribbean lanternshark.

Northwest and Western Central Atlantic including the Caribbean. Upper slope. 311–695 m.

*Etmopterus litvinovi* (Parin and Kotlyar, in Kotlyar, 1990). Smalleye lanternshark.

Southeast Pacific: Nazca and Sala y Gomez Submarine Ridges. Submarine ridges. 630–1 100 m.

*Etmopterus lucifer* (Jordan and Snyder, 1902). Blackbelly lanternshark.

Patchy in the Western Pacific, but provisional records from elsewhere require confirmation. Outer shelf, upper to mid slope. 158–1 357 m.

*Etmopterus molleri* (Whitley, 1939). Slendertail lanternshark.

Patchy in the Western Pacific and possibly also the Western Indian (Mozambique). Upper slope. 238–655 m. Includes the probable junior synonym *Etmopterus schmidti* Dolganov, 1986.

*Etmopterus perryi* (Springer and Burgess, 1985). Dwarf lanternshark.

Western Central Atlantic: Caribbean off Colombia. Upper slope. 283–375 m.

*Etmopterus polli* (Bigelow, Schroeder and Springer, 1953). African lanternshark.

Eastern Atlantic: West Africa. Possibly Caribbean off Venezuela. Upper to mid slope. 300–1 000 m.

*Etmopterus princeps* (Collett, 1904). Great lanternshark.

Wide-ranging in the North and Central Atlantic. Upper to deep slope, deep-sea rises, deep-sea plateaus and abyssal plains. 567–4 500 m.

*Etmopterus pseudosqualiolus* (Last, Burgess and Séret, 2002). False pygmy lanternshark.

Western Central Pacific: Norfolk and Lord Howe Ridges. Submarine ridges. 1 043–1 102 m.

*Etmopterus pusillus* (Lowe, 1839). Smooth lanternshark.

Wide-ranging in temperate and warm-temperate waters of the Atlantic and Indo-West and Central Pacific. Upper to mid (possibly to deep) slope. 274–1 000 m (possibly to 1 998 m).

*Etmopterus pycnolepis* (Kotlyar, 1990). Densescale lanternshark.

Southeast Pacific: Nazca and Sala y Gomez Submarine Ridges. Submarine ridges. 330–763 m.

*Etmopterus robinsi* (Schofield and Burgess, 1997). West Indian lanternshark.

Western Central Atlantic: Caribbean. Upper to mid slope. 412–787 m.

*Etmopterus schultzi* (Bigelow, Schroeder and Springer, 1953). Fringefin lanternshark.

Western Central Atlantic: Gulf of Mexico. Upper to mid slope. 220–915 m (mostly >350 m).

*Etmopterus sentosus* (Bass, D'Aubrey and Kistnasamy, 1976). Thorny lanternshark.

Western Indian: East Africa. Upper slope. 200–500 m.

***Etmopterus spinax* (Linnaeus, 1758).** Velvet belly.

Widespread in the Eastern Atlantic and Mediterranean. Shelf, upper to deep slope and deep-sea rises. 70–2 490 m (mostly 200–500 m).

***Etmopterus splendidus* (Yano, 1988).** Splendid lanternshark.

Northwest Pacific: Japan and Taiwan. Uppermost slope. 210 m.

***Etmopterus unicolor* (Engelhardt, 1912).** Brown lanternshark.

Patchy in the Eastern Atlantic and Indo-West Pacific. Upper to mid slope and seamounts. 750–1 500 m.

***Etmopterus villosus* (Gilbert, 1905).** Hawaiian lanternshark.

Eastern Central Pacific: Hawaii. Upper to mid slope. 406–911 m.

***Etmopterus virens* (Bigelow, Schroeder and Springer, 1953).** Green lanternshark.

Western Central Atlantic: Gulf of Mexico and Caribbean. Upper to mid slope. 196–915 m (mostly >350 m).

***Miroscyllium sheikoi* (Dolganov, 1986).** Rasptooth dogfish.

Northwest Pacific: off Japan. Upper slope of submarine ridges. 340–370 m.

***Trigonognathus kabeyai* (Mochizuki and Ohe, 1990).** Viper dogfish.

North and Central Pacific: Japan and Hawaii. Upper slope and seamounts. 270–360 m.

### Family Somniosidae. Sleeper Sharks.

***Centroscymnus coelolepis* (Bocage and Capello, 1864).** Portuguese dogfish.

Wide-ranging in the Atlantic and Indo-Pacific. Outer shelf, upper to deep slope and abyssal plains. 128–3 675 m (mostly >400 m).

***Centroscymnus owstoni* (Garman, 1906).** Roughskin dogfish.

Wide-ranging, but patchy in the Atlantic, Pacific and Eastern Indian. Upper to mid slope and submarine ridges. 426–1 459 m (mostly >600 m). Includes the junior synonym *Centroscymnus cryptacanthus* Regan, 1906.

***Centroselachus crepidater* (Bocage and Capello, 1864).** Longnose velvet dogfish.

Wide-ranging, but patchy in the Western Atlantic, Pacific and Western Indian. Upper to deep slope. 270–2 080 m (mostly >500 m).

***Proscymnodon macracanthus* (Regan, 1906).** Largespine velvet dogfish.

Southeast Pacific: Straits of Magellan (Chile). Habitat and depth unrecorded.

***Proscymnodon plunketi* (Waite, 1909).** Plunket shark.

Patchy in southern regions of the Indo-West Pacific. Upper to mid slope. 219–1 427 m (most common 550–732 m).

***Scymnodalatias albicauda* (Taniuchi and Garrick, 1986).** Whitetail dogfish.

Patchy in the Southern Ocean. Outer shelf, upper slope and submarine ridges. 150–500 m.

***Scymnodalatias garricki* (Kukuyev and Konovalenko, 1988).** Azores dogfish.

Northeast Atlantic: North Atlantic Ridge. Mesopelagic over seamounts. 300 m.

***Scymnodalatias sherwoodi* (Archey, 1921).** Sherwood's dogfish.

Patchy in the Southern Ocean. Upper slope. 400–500 m.

***Scymnodon ringens* (Bocage and Capello, 1864).** Knifetooth dogfish.

Northeast and Eastern Central Atlantic. Uncertain from New Zealand in the Southwest Pacific. Upper to deep slope. 200–1 600 m.

***Somniosus antarcticus* (Whitley, 1939).** Southern sleeper shark.

Patchy in the Southern Ocean. Outer shelf and upper to mid slope. 145–1 200 m.

***Somniosus longus* (Tanaka, 1912).** Frog shark.

Western Pacific: Japan and New Zealand. Upper to mid slope. 250–1 160 m.

***Somniosus microcephalus* (Bloch and Schneider, 1801).** Greenland shark.

Cool temperate and boreal waters of the North Atlantic. Shelf (inshore in Arctic winter) and upper to mid slope. 0–1 200 m.

***Somniosus pacificus* (Bigelow and Schroeder, 1944).** Pacific sleeper shark.

Wide-ranging in the North Pacific. Shelf and upper to deep slope (shallower in north, deeper in south of range). 0–2 000 m.

*Somniosus rostratus* (Risso, 1810). Little sleeper shark.

Northeast Atlantic and the Mediterranean. Uncertain from Cuba in the Western Central Atlantic. Outermost shelf and upper to mid slope. 200–2 200 m.

*Zameus ichiharai* (Yano and Tanaka, 1984). Japanese velvet dogfish.

Northwest Pacific: Japan. Upper to mid slope. 450–830 m.

*Zameus squamulosus* (Günther, 1877). Velvet dogfish.

Wide-ranging but patchy in the Atlantic, Indian and Pacific. Upper to mid slope. Also epipelagic and mesopelagic. 550–1 450 m (when benthic); 0–580 m in water up to 6 000 m deep (when pelagic).

### Family Oxynotidae. Roughsharks.

*Oxynotus bruniensis* (Ogilby, 1893). Prickly dogfish.

Indo-West Pacific: New Zealand and Australia. Shelf and upper to mid slope. 45–1 067 m (most common 350–650 m).

*Oxynotus caribbaeus* (Cervigon, 1961). Caribbean roughshark.

Western Central Atlantic: Gulf of Mexico and Caribbean (Venezuela). Upper slope. 402–457 m.

*Oxynotus centrina* (Linnaeus, 1758). Angular roughshark.

Wide-ranging in the Eastern Atlantic and the Mediterranean. Shelf and upper slope. 50–660 m (mostly >100 m).

*Oxynotus japonicus* (Yano and Murofushi, 1985). Japanese roughshark.

Northwest Pacific: Japan. Uppermost slope. 225–270 m.

*Oxynotus paradoxus* (Frade, 1929). Sailfin roughshark.

Northeast and Eastern Central Atlantic: Scotland to West Africa. Upper slope. 265–720 m.

### Family Dalatiidae. Kitefin Sharks.

*Dalatias licha* (Bonnaterre, 1788). Kitefin shark.

Wide-ranging but patchy in the Atlantic, Indo-West and Central Pacific. Shelf and upper to deep slope. 37–1 800 m (mainly >200 m).

*Euprotomicroides zantedeschia* (Hulley and Penrith, 1966). Taillight shark.

Patchy in the South Atlantic: off Brazil and southern Africa. Upper slope. Also epipelagic. 458–641 m (when benthic), 0–25 m (when pelagic).

*Euprotomicrus bispinatus* (Quoy and Gaimard, 1824). Pygmy shark.

Wide-ranging but scattered records in the Atlantic and Indo-Pacific. Epipelagic, mesopelagic and bathypelagic. 0–>1 500 m in water up to 9 938 m deep.

*Heteroscymnoides marleyi* (Fowler, 1934). Longnose pygmy shark.

Scattered records in the southern Atlantic, Southeast Pacific and Southwest Indian. Epipelagic and mesopelagic. 0–502 m in water 830–>4 000 m deep.

*Isistius brasiliensis* (Quoy and Gaimard, 1824). Cookiecutter or cigar shark.

Circumglobal in tropical and warm temperate waters. Epipelagic, mesopelagic and bathypelagic. 0–3 500 m.

*Isistius labialis* (Meng, Chu and Li, 1985). South China cookiecutter shark.

Northwest Pacific: South China Sea. Mesopelagic over slope. 520 m.

*Mollisquama parini* (Dolganov, 1984). Pocket shark.

Southeast Pacific: Nazca Submarine Ridge. Submarine ridges. 330 m.

*Squaliolus aliae* (Teng, 1959). Smalleye pygmy shark.

Patchy in the Western Pacific and Eastern Indian. Epipelagic, mesopelagic and bathypelagic near land. 200–2 000 m.

*Squaliolus laticaudus* (Smith and Radcliffe, 1912). Spined pygmy shark.

Wide-ranging but scattered records in the Atlantic, Western Pacific and Western Indian. Epipelagic and mesopelagic. 200–500 m.

## **Order Squatiniformes. Angelsharks.**

### **Family Squatinidae. Angelsharks.**

*Squatina aculeata* (Dumeril, in Cuvier, 1817). Sawback angelshark.

Eastern Atlantic off West Africa, and the Western Mediterranean. Shelf and upper slope. 30–500 m.

*Squatina africana* (Regan, 1908). African angelshark.

Western Indian: East Africa. Shelf and upper slope. 0–494 m (mainly 60–300 m).

*Squatina albipunctata* (Last and White, 2008). Eastern angelshark.

Western Pacific and (marginally) Eastern Indian: eastern Australia. Shelf and upper slope. 37–415 m.

*Squatina argentina* (Marini, 1930). Argentine angelshark.

Southwest Atlantic: Brazil to Argentina. Shelf and upper slope. 51–320 m (mostly 120–320 m).

*Squatina formosa* (Shen and Ting, 1972). Taiwan angelshark.

Western Pacific: Taiwan and the Philippines. Outer shelf and upper slope. 183–385 m.

*Squatina pseudocellata* (Last and White, 2008). Western angelshark.

Eastern Indian: western Australia. Outer shelf and upper slope. 150–312 m.

*Squatina tergocellata* (McCulloch, 1914). Ornate angelshark.

Eastern Indian: Australia. Outer shelf and upper slope. 130–400 m (most common ~300 m).

## **Order Pristiophoriformes. Sawsharks.**

### **Family Pristiophoridae. Sawsharks.**

*Pliotrema warreni* (Regan, 1906). Sixgill sawshark.

Southeast Atlantic and Western Indian: southern Africa. Shelf and upper slope. 37–500 m.

*Pristiophorus delicatus* (Yearsley, Last and White, 2008). Tropical sawshark.

Western Central Pacific: northeastern Australia. Upper slope. 246–405 m.

*Pristiophorus schroederi* (Springer and Bullis, 1960). Bahamas sawshark

Western Central Atlantic: between Cuba, Florida (United States) and Bahamas. Upper to mid slope. 438–952 m.

## **Order Rajiformes. Batoids.**

### **Family Rhinobatidae. Guitarfishes.**

*Rhinobatos variegatus* (Nair and Lal Mohan, 1973). Stripenose guitarfish

Eastern Indian: Gulf of Mannar, India. Upper slope. 366 m.

### **Family Narcinidae. Numbfishes.**

*Benthobatis krefftii* (Rincon, Stehmann and Vooren, 2001). Brazilian blind torpedo.

Southwest Atlantic: Brazil. Upper slope. 400–600 m.

*Benthobatis marcida* (Bean and Weed, 1909). Pale or blind torpedo.

Western Central Atlantic: South Carolina (United States) to northern Cuba. Upper to mid slope. 274–923 m.

*Benthobatis moresbyi* (Alcock, 1898). Dark blidray.

Western Indian: Arabian Sea. Mid slope. 787–1 071 m.

*Benthobatis yangi* (Carvalho, Compagno and Ebert, 2003). Narrow blidray.

Northwest and Western Central Pacific: Taiwan. Upper slope (possibly also outer shelf). <300 m.

*Narcine lasti* (Carvalho and Séret, 2002). Western numbfish.

Eastern Indian: western Australia. Outermost shelf and upper slope. 180–320 m.

***Narcine nelsoni* (Carvalho, 2008).** Eastern numbfish.

Western Central Pacific: northeastern Australia. Outer shelf and upper slope. 140–540 m.

***Narcine tasmaniensis* (Richardson, 1840).** Tasmanian numbfish.

Eastern Indian and Southwest Pacific: southeastern Australia. Shelf (south of range) and upper slope (north of range). 5–640 m (in north of range mainly 200–640 m).

### Family Narkidae. Sleeper Rays.

***Heteronarce garmani* (Regan, 1921).** Natal sleeper ray.

Western India: Mozambique and South Africa. Shelf and upper slope. 73–329 m.

***Heteronarce mollis* (Lloyd, 1907).** Soft sleeper ray.

Western Indian: Gulf of Aden. Shelf and upper slope. 73–346 m.

***Typhlonarke aysoni* (Hamilton, 1902).** Blind electric ray.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 46–900 m (most common 300–400 m).

***Typhlonarke tarakea* (Phillipps, 1929).** Oval electric ray.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 46–900 m (most common 300–400 m).

### Family Torpedinidae. Torpedo Rays.

***Torpedo fairchildi* (Hutton, 1872).** New Zealand torpedo ray.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 5–1 135 m (most common 100–300 m).

***Torpedo fuscomaculata* (Peters, 1855).** Blackspotted torpedo.

Western Indian: Mozambique and South Africa. Reports from Western Indian Ocean islands likely refer to similar undescribed species. Shelf and upper slope. 0–439 m.

***Torpedo macneilli* (Whitley, 1932).** Short-tail torpedo ray.

Eastern Indian and Western Pacific: Australia. Outer shelf and upper slope. 90–825 m.

***Torpedo microdiscus* (Parin and Kotlyar, 1985).** Smalldisk torpedo.

Southeast Pacific: Nazca and Sala y Gomez Submarine Ridges. Submarine ridges.

***Torpedo nobiliana* (Bonaparte, 1835).** Great, Atlantic, or black torpedo.

Wide-ranging in the Eastern and Western Atlantic. Shelf and upper slope. 2–530 m.

***Torpedo puelcha* (Lahille, 1928).** Argentine torpedo.

Southwest Atlantic: Brazil to Argentina. Shelf and upper slope. Inshore–600 m.

***Torpedo tokionis* (Tanaka, 1908).** Longtail torpedo ray.

Patchy in the Eastern Indian and Western Pacific. Upper slope. 410–735 m.

***Torpedo tremens* (de Buen, 1959).** Chilean torpedo.

Patchy in the Eastern Central and Southeast Pacific. Shelf and upper slope. Inshore 0–700 m.

### Family Arhynchobatidae. Softnose Skates.

***Arhynchobatis asperrimus* (Waite, 1909).** Longtailed skate.

Southwest Pacific: New Zealand. Outer shelf and upper to mid slope. 90–1 070 m.

***Bathyraja abyssicola* (Gilbert, 1896).** Deepsea skate.

Wide-ranging in the North Pacific. Upper to deep slope and abyssal plains. 362–2 906 m.

***Bathyraja aguja* (Kendall and Radcliffe, 1912).** Aguja skate.

Southeast Pacific: Peru. Mid slope. 980 m.

***Bathyraja albomaculata* (Norman, 1937).** White-dotted skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper to mid slope. 55–861 m.

***Bathyraja aleutica* (Gilbert, 1895).** Aleutian skate.

Wide-ranging in the North Pacific. Outer shelf and upper slope. 91–700 m.

***Bathyraja andriashevi* (Dolganov, 1985).** Little-eyed skate.

Northwest Pacific: Japan and Russia. Mid to deep slope. 1 400–2 000 m.

*Bathyraja bergi* (Dolganov, 1985). Bottom skate.

Northwest Pacific: Japan and Russia. Outer shelf and upper to mid slope. 70–900 m. Includes the junior synonym *Bathyraja pseudoisotachys* Ishihara and Ishiyama, 1985.

*Bathyraja brachyurops* (Fowler, 1910). Broadnose skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 28–604 m.

*Bathyraja caeluronigricans* (Ishiyama and Ishihara, 1977). Purpleblack skate.

Northwest Pacific: Japan and Russia. Upper slope. 200–400 m. Possible synonym of *Bathyraja matsubarai* (Ishiyama, 1952).

*Bathyraja cousseauae* (Diaz de Astarloa and Mabragaña, 2004). Cousseau's skate.

Southwest Atlantic: Argentina and the Falkland/Malvinas Islands. Outer shelf and upper to mid slope. 119–1 011 m.

*Bathyraja diplotaenia* (Ishiyama, 1950). Dusky pink skate.

Northwest Pacific: Japan. Outer shelf and upper slope. 100–700 m.

*Bathyraja eatoni* (Günther, 1876). Eaton's skate.

Circum-Antarctic. Shelf, upper to mid slope, deep-sea plateaus and submarine ridges. 15–800 m.

*Bathyraja fedorovi* (Dolganov, 1985). Cinnamon skate.

Northwest Pacific: Japan and Russia. Upper to deep slope and abyssal plains. 447–2 025 m.

*Bathyraja griseocauda* (Norman, 1937). Graytail skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper to mid slope. 82–941 m.

*Bathyraja hesperafricana* (Stehmann, 1995). West African skate.

Eastern Central Atlantic: West Africa. Mid to deep slope. 750–2 000 m.

*Bathyraja interrupta* (Gill and Townsend, 1897). Bering skate.

Wide-ranging in the North and Eastern Central Pacific, although not well defined due to possible misidentifications. Shelf and upper to mid slope. 23–1 500 m.

*Bathyraja irrasa* (Hureau and Ozouf-Costaz, 1980). Kerguelen sandpaper skate.

Antarctic Indian: Kerguelen Islands. Upper to mid slope. 300–1 200 m.

*Bathyraja isotrachys* (Günther, 1877). Raspback skate.

Northwest Pacific: Russia, Japan and Korea. Upper to deep slope. 370–2 000 m.

*Bathyraja kincaidii* (Garman, 1908). Sandpaper skate.

Eastern Central and Northeast Pacific: Baja California north to Gulf of Alaska. Shelf and upper to mid slope. 55–1 372 m (most common 200–500 m).

*Bathyraja lindbergi* (Ishiyama and Ishihara, 1977). Commander skate.

Northwest Pacific: Japan, Russia and the Bering Sea. Outer shelf and upper to mid slope. 120–950 m.

*Bathyraja longicauda* (de Buen, 1959). Slimtail skate.

Southeast Pacific: Peru and Chile. Upper slope. 580–735 m.

*Bathyraja maccaini* (Springer, 1972). McCain's skate.

Southeast Pacific and Antarctic Atlantic: Chile and Antarctica (Orkney and South Shetland islands to the Antarctic Peninsula). Shelf and upper slope. Inshore–500 m.

*Bathyraja macloviana* (Norman, 1937). Patagonian skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 53–514 m.

*Bathyraja maculata* (Ishiyama and Ishihara, 1977). Whiteblotched skate.

Northern Pacific: Aleutian Islands and Bering Sea westwards to Japan. Shelf and upper to mid slope. 73–1 110 m.

*Bathyraja magellanica* (Philippi, 1902 or Steindachner, 1903). Magellan skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 51–600 m (mostly <70 m in Falklands).

*Bathyraja matsubarai* (Ishiyama, 1952). Dusky purple skate.

Northwest Pacific: Japan and Russia. Occurrence in the Northeast Pacific requires confirmation. Outer shelf and upper to deep slope. 120–2 000 m.

***Bathyraja meridionalis* (Stehmann, 1987).** Darkbelly skate.

Primarily Sub-Antarctic in the Atlantic, possibly circum-Antarctic. Upper to deep slope, abyssal plains and seamounts. 300–2 240 m.

***Bathyraja minispinosa* (Ishiyama and Ishihara, 1977).** Smallthorn skate.

Northern Pacific: Aleutian Islands and Bering Sea westwards to Japan. Outer shelf and upper to mid slope. 150–1 420 m.

***Bathyraja multispinis* (Norman, 1937).** Multispine skate.

Southwest Atlantic and Southeast Pacific: southern South America. Outer shelf and upper slope. 82–740 m. A record at 1 900 m off Uruguay requires confirmation.

***Bathyraja murrayi* (Günther, 1880).** Murray's skate.

Antarctic Indian: Kerguelen and Heard Islands, but possibly circum-Antarctic. Shelf and upper slope. 30–650 m.

***Bathyraja notoroensis* (Ishiyama and Ishihara, 1977).** Notoro skate.

Northwest Pacific: Japan. Upper slope. ~600 m. Possible synonym of *Bathyraja matsubarai* (Ishiyama, 1952).

***Bathyraja papilionifera* (Stehmann, 1985).** Butterfly skate.

Southwest Atlantic: Argentina and the Falkland/Malvinas Islands. Upper to deep slope and seamounts. 637–2 000 m.

***Bathyraja parmifera* (Bean, 1881).** Alaska or flathead skate.

Wide-ranging in the North Pacific: Gulf of Alaska to Japan. Shelf and upper to mid slope. 20–1 425 m. Includes the synonym *Bathyraja simoterus* (Ishiyama, 1967).

***Bathyraja peruana* (McEachran and Miyake, 1984).** Peruvian skate.

Southeast Pacific: Ecuador to Chile. Upper to mid slope. 600–1 060 m.

***Bathyraja richardsoni* (Garrick, 1961).** Richardson's skate.

Southwest Pacific: Australia and New Zealand (possibly also in the North Atlantic). Mid to deep slope, abyssal plains and seamounts. 1 370–2 909 m.

***Bathyraja scaphiops* (Norman, 1937).** Cuphead skate.

Southwest Atlantic: Argentina and the Falkland/Malvinas Islands. Outer shelf and upper slope. 104–509 m (most common 104–159 m).

***Bathyraja schroederi* (Krefft, 1968).** Whitemouth skate.

Southwest Atlantic and Southeast Pacific: southern South America. Mid to deep slope and abyssal plains. 800–2 380 m.

***Bathyraja shuntovi* (Dolganov, 1985).** Narrownose skate.

Southwest Pacific: New Zealand. Upper to mid slope. 300–1 470 m.

***Bathyraja smirnovi* (Soldatov and Lindberg, 1913).** Golden skate.

Northwest Pacific: Bering Sea to Japan. Outer shelf and upper to mid slope. 100–1 000 m.

***Bathyraja smithii* (Müller and Henle, 1841).** African softnose skate.

Southeast Atlantic: Namibia and South Africa. Upper to mid slope. 400–1 020 m.

***Bathyraja spinicauda* (Jensen, 1914).** Spinetail or spinytail skate.

Wide-ranging in the Northeast and Northwest Atlantic. Outer shelf and upper to mid slope. 140–1 209 m (mainly >400 m).

***Bathyraja spinosissima* (Beebe and Tee-Van, 1941).** White skate.

Eastern Central and Southeast Pacific. Records from the Northwest Pacific may be a separate species. Mid to deep slope and abyssal plains. 800–2 938 m.

***Bathyraja trachouros* (Ishiyama, 1958).** Eremo skate.

Northwest Pacific: Japan. Upper slope. 300–500 m.

***Bathyraja trachura* (Gilbert, 1892).** Roughtail skate.

Wide-ranging in the North Pacific. Upper to deep slope and abyssal plains. 213–2 550 m.

***Bathyraja tzinovskii* (Dolganov, 1985).** Creamback skate.

Northwest Pacific: Japan and Russia. Deep slope and abyssal plains. 1 776–2 500 m.

***Bathyraja violacea* (Suvorov, 1935).** Okhotsk skate.

North Pacific: Bering Sea to Japan. Shelf and upper to mid slope. 23–1 100 m.

***Brochiraja aenigma* (Last and McEachran, 2006).**

Southwest Pacific: New Zealand. Upper slope. 660–665 m.

*Brochiraja albilabiata* (Last and McEachran, 2006).

Southwest Pacific: New Zealand. Mid slope. 785–1 180 m.

*Brochiraja asperula* (Garrick and Paul, 1974). Prickly deepsea skate.

Southwest Pacific: New Zealand. Mid slope. 350–1 010 m.

*Brochiraja leviveneta* (Last and McEachran, 2006).

Southwest Pacific: New Zealand. Mid slope. 960–1 015 m.

*Brochiraja microspinifera* (Last and McEachran, 2006).

Southwest Pacific: New Zealand. Upper to mid slope. 510–900 m.

*Brochiraja spinifera* (Garrick and Paul, 1974). Spiny deepsea skate.

Southwest Pacific: New Zealand. Upper slope. 500–750 m.

*Insentiraja laxipella* (Yearsley and Last, 1992). Eastern looseskin skate.

Western Central Pacific: northeastern Australia. Mid slope. 800–880 m.

*Insentiraja subtilispinosa* (Stehmann, 1985). Velvet or western looseskin skate.

Patchy in the Eastern Indian and Western Central Pacific. Upper to mid slope. 320–1 460 m.

*Notoraja azurea* (McEachran and Last, 2008). Blue skate.

Eastern Indian: western and southern Australia. Mid slope. 765–1 440 m.

*Notoraja hirticauda* (Last and McEachran, 2006). Ghost skate.

Eastern Indian: western Australia. Upper to mid slope. 500–760 m.

*Notoraja lira* (McEachran and Last, 2008). Broken Ridge skate.

Eastern Indian: Broken Ridge. Submarine ridges. 1 050 m.

*Notoraja ochroderma* (McEachran and Last, 1994). Pale skate.

Western Central Pacific: northeastern Australia. Upper slope. 400–455 m.

*Notoraja sapphira* (Séret and Last, 2009). Sapphire skate.

Southwest and Western Central Pacific: Norfolk Ridge. Submarine Ridges. 1 195–1 313 m.

*Notoraja sticta* (McEachran and Last, 2008). Blotched skate.

Eastern Indian: southern Australia. Mid slope. 820–1 200 m.

*Notoraja tobitukai* (Hiyama, 1940). Leadhued skate.

Northwest Pacific: Japan and Taiwan. Shelf and upper to mid slope. 60–900 m.

*Pavoraja allenii* (McEachran and Fechhelm, 1982). Allen's skate.

Eastern Indian: western Australia. Upper slope. 304–458 m.

*Pavoraja arenaria* (Last, Mallick and Yearsley, 2008). Sandy peacock skate.

Eastern Indian: southern Australia. Outermost shelf and upper slope. 192–712 m (mainly 300–400 m).

*Pavoraja mosaica* (Last, Mallick and Yearsley, 2008). Mosaic skate.

Western Central Pacific: northeastern Australia. Upper slope. 300–405 m.

*Pavoraja nitida* (Günther, 1880). Peacock skate.

Eastern Indian and Southwest Pacific: southern Australia. Shelf and upper slope. 75–432 m.

*Pavoraja pseudonitida* (Last, Mallick and Yearsley, 2008). False peacock skate.

Western Central Pacific: northeastern Australia. Upper slope. 212–512 m.

*Pavoraja umbrosa* (Last, Mallick and Yearsley, 2008). Dusky skate.

Western Pacific: eastern Australia. Upper slope. 360–731 m.

*Psammobatis scobina* (Philippi, 1857). Rasphorn sandskate.

Southeast Pacific: Chile. Shelf and upper slope. 40–450 m.

*Pseudoraja fischeri* (Bigelow and Schroeder, 1954). Fanfin skate.

Scattered records in the Western Central Atlantic. Upper slope. 412–576 m.

*Rhinoraja kujiensis* (Tanaka, 1916). Dapple-bellied softnose skate.

Northwest Pacific: Japan. Upper to mid slope. 450–~1 000 m.

*Rhinoraja longi* (Raschi and McEachran, 1991). Aleutian dotted skate.

North Pacific: Aleutian Islands. Upper slope. 390–435 m.

*Rhinoraja longicauda* (Ishiyama, 1952). White-bellied softnose skate.

Northwest Pacific: Japan and Russia. Upper to mid slope. 300–1 000 m.

*Rhinoraja obtusa* (Gill and Townsend, 1897). Blunt skate.

North Pacific: Bering Sea. Habitat and depth information not available.

*Rhinoraja odai* (Ishiyama, 1952). Oda's skate.

Northwest Pacific: Japan. Upper to mid slope. 300–870 m.

***Rhinoraja taranetzi* (Dolganov, 1985).** Mudskate.

Reasonably wide-ranging in the North Pacific. Shelf and upper to mid slope. 81–1 000 m.  
Includes the junior synonym *Bathyraja hubbsi* Ishihara and Ishiyama, 1985.

**Family Rajidae. Hardnose Skates.*****Amblyraja badia* (Garman, 1899).** Broad skate.

Patchy in the North Pacific. Mid to deep slope and abyssal plains. 846–2 324 m.

***Amblyraja doellojuradoi* (Pozzi, 1935).** Southern thorny skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 51–642 m.

***Amblyraja frerichsi* (Krefft, 1968).** Thickbody skate.

Southwest Atlantic and Southeast Pacific: southern South America. Mid to deep slope and abyssal plains. 720–2 609 m.

***Amblyraja georgiana* (Norman, 1938).** Antarctic starry skate.

Circum-Antarctic, including off Chile and the Falkland/Malvinas Islands. Shelf and upper to mid slope. 20–800 m.

***Amblyraja hyperborea* (Collett, 1879).** Arctic or boreal skate.

Wide-ranging but patchy in deep temperate waters. Upper to deep slope and abyssal plains. 300–2 500 m. Taxonomic issues unresolved, may represent a species complex.

***Amblyraja jensenii* (Bigelow and Schroeder, 1950).** Jensen's skate.

North Atlantic: Nova Scotia (Canada) to southern New England (United States) and Iceland. Upper to deep slope. 366–2 196 m.

***Amblyraja radiata* (Donovan, 1808).** Thorny skate.

Wide-ranging in the North Atlantic and also off South Africa. Shelf and upper to mid slope. 18–1 400 m (most common 27–439 m).

***Amblyraja reversa* (Lloyd, 1906).** Reversed skate.

Western Indian: Arabian Sea. Deep slope. 1 500 m.

***Amblyraja robertsi* (Hulley, 1970).** Bigmouth skate.

Southeast Atlantic: South Africa. Mid slope. 1 350 m.

***Amblyraja taaf* (Meisner, 1987).** Whiteleg skate.

Antarctic Indian: Crozet Islands. Outer shelf and upper slope. 150–600 m.

***Breviraja claramaculata* (McEachran and Matheson, 1985).** Brightspot skate.

Western Central Atlantic: South Carolina to Florida (United States). Upper to mid slope. 293–896 m.

***Breviraja colesi* (Bigelow and Schroeder, 1948).** Lightnose skate.

Western Central Atlantic: Florida (United States), Bahamas and Cuba. Upper slope. 220–415 m.

***Breviraja marklei* (McEachran and Miyake, 1987).** Nova Scotia skate.

Northwest Atlantic: Canada. Upper to mid slope. 443–988 m.

***Breviraja mouldsi* (McEachran and Matheson, 1995).** Blacknose skate.

Western Central Atlantic: Honduras to Panama. Upper to mid slope. 353–776 m.

***Breviraja nigriventris* (McEachran and Matheson, 1985).** Blackbelly skate.

Western Central Atlantic: Panama and the northern coast of South America. Upper to mid slope. 549–776 m.

***Breviraja spinosa* (Bigelow and Schroeder, 1950).** Spinose skate.

Western Central Atlantic: North Carolina to Florida (United States). Upper slope. 366–671 m.

***Dactylobatus armatus* (Bean and Weed, 1909).** Skilletskate.

Patchy in the Western Central Atlantic. Upper slope. 338–685 m.

***Dactylobatus clarki* (Bigelow and Schroeder, 1958).** Hookskeate.

Patchy in the Western Central Atlantic. Upper to mid slope. 366–915 m.

***Dipturus acrobelus* (Last, White and Pogonoski, 2008).** Deepwater skate.

Eastern Indian and Southwest Pacific: southern Australia. Upper to mid slope. 446–1 328 m (mainly 800–1 000 m).

*Dipturus apricus* (Last, White and Pogonoski, 2008). Pale tropical skate.

Western Central Pacific: northeastern Australia. Outermost shelf and upper slope. 196–606 m (mainly 300–500 m).

*Dipturus batis* (Linnaeus, 1758). Gray or blue skate.

Formerly wide-ranging in the Northeast and Eastern Central Atlantic, including the Mediterranean, but now extirpated from or reduced in much of its historical range. Outer shelf, upper to deep slope and deep-sea rises. 100–2 619 m.

*Dipturus bullisi* (Bigelow and Schroeder, 1962). Tortugas skate.

Patchy in the Western Central Atlantic. Upper slope. 183–549 m.

*Dipturus campbelli* (Wallace, 1967). Blackspot skate.

Western Indian: patchy off South Africa and Mozambique. Outer shelf and upper slope. 137–403 m.

*Dipturus canutus* (Last, 2008). Grey skate.

Eastern Indian and Southwest Pacific: southeastern Australia. Upper slope. 330–730 m (mainly 400–600 m).

*Dipturus cerva* (Whitley, 1939). White-spotted skate.

Eastern Indian and Southwest Pacific: southern Australia. Shelf and upper slope. 20–470 m.

*Dipturus croisnieri* (Séret, 1989). Madagascar skate.

Western Indian: Madagascar. Upper to mid slope. 300–850 m.

*Dipturus doutei* (Cadenat, 1960). Violet or javelin skate.

Eastern Central and Southeast Atlantic: West Africa. Outer shelf and upper to mid slope. 163–800 m (mostly >400 m).

*Dipturus endeavouri* (Last, 2008). Endeavour skate.

Western Central and Southwest Pacific: eastern Australia. Outer shelf and upper slope. 125–500 m.

*Dipturus garricki* (Bigelow and Schroeder, 1958). San Blas skate.

Western Central Atlantic: northern Gulf of Mexico and Nicaragua. Upper slope. 275–476 m.

*Dipturus gigas* (Ishiyama, 1958). Giant skate.

Northwest and Western Central Pacific: Japan to the Philippines. Upper to mid slope. 300–1 000 m.

*Dipturus grahami* (Last, 2008). Graham's skate.

Western Central and Southwest Pacific: eastern Australia. Outer shelf and upper slope. 146–490 m (mainly 250–450 m).

*Dipturus gadgei* (Whitley, 1940). Bight skate.

Eastern Indian and Southwest Pacific: southern Australia. Outer shelf and upper slope. 160–765 m (most common 400–550 m).

*Dipturus healdi* (Last, White and Pogonoski, 2008). Heald's skate.

Eastern Indian: western Australia. Upper slope. 304–520 m.

*Dipturus innominatus* (Garrick and Paul, 1974). New Zealand smooth skate.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 15–1 310 m.

*Dipturus johannisdavesi* (Alcock, 1899). Travancore skate.

Western Indian: Gulf of Aden and India. Upper slope. 457–549 m.

*Dipturus laevis* (Mitchell, 1817). Barndoor skate.

Northwest Atlantic: Canada and United States. Shelf and upper slope. 0–430 m.

*Dipturus lanceorostratus* (Wallace, 1967). Rattail skate.

Western Indian: Mozambique. Upper slope. 430–439 m.

*Dipturus leptocaudus* (Krefft and Stehmann, 1975). Thintail skate.

Southwest Atlantic: Brazil. Upper slope. 400–550 m.

*Dipturus linteus* (Fries, 1838). Sailskate.

North Atlantic, primarily in the Northeast Atlantic. Upper to mid slope. 150–1 200+ m.

*Dipturus macrocaudus* (Ishiyama, 1955). Bigtail skate.

Northwest Pacific: Japan, Korea and Taiwan. Upper to mid slope. 300–800 m.

*Dipturus melanospilus* (Last, White and Pogonoski, 2008). Blacktip skate.

Western Central and Southwest Pacific: eastern Australia. Upper slope. 239–695 m.

*Dipturus mennii* (Gomes and Paragó, 2001). South Brazilian skate.

Southwest Atlantic: Brazil. Outer shelf and upper slope. 133–500 m. Includes the likely synonym *Dipturus diehli* Soto and Mincarone, 2001.

*Dipturus nidarosiensis* (Collett, 1880). Norwegian skate.

Northeast Atlantic: Norway, Iceland and the Rockall Trough. Upper to mid slope and deep-sea rises. 200–1 000 m.

*Dipturus oculus* (Last, 2008). Ocellate skate

Eastern Indian: western Australia. Upper slope. 200–389 m.

*Dipturus oregoni* (Bigelow and Schroeder, 1958). Hooktail skate.

Western Central Atlantic: northern Gulf of Mexico. Upper to mid slope. 475–1 079 m.

*Dipturus oxyrhynchus* (Linnaeus, 1758). Sharpnose skate.

Wide-ranging in the Northeast and Eastern Central Atlantic, including the Mediterranean.

Shelf and upper to mid slope. 15–900 m.

*Dipturus polyommata* (Ogilby, 1910). Argus skate.

Western Central Pacific: northeastern Australia. Outer shelf and upper slope. 135–320 m.

*Dipturus pullopunctata* (Smith, 1964). Slime skate.

Southeast Atlantic: Namibia and South Africa. Shelf and upper slope. 15–457 m (most common 100–300 m).

*Dipturus queenslandicus* (Last, White and Pogonoski, 2008). Queensland deepwater skate.

Western Central Pacific: northeastern Australia. Upper slope. 399–606 m.

*Dipturus springeri* (Wallace, 1967). Roughbelly skate.

Southeast Atlantic and Western Indian: southern Africa. Upper slope. 400–740 m (mostly 400–500 m).

*Dipturus stenorhynchus* (Wallace, 1967). Prownose skate.

Southeast Atlantic and Western Indian: South Africa and Mozambique. Upper to mid slope. 253–761 m.

*Dipturus teevani* (Bigelow and Schroeder, 1951). Caribbean skate.

Patchy throughout the Western Central Atlantic. Upper slope. 311–732 m.

*Dipturus tengu* (Jordan and Fowler, 1903). Acutenose or tengu skate.

Northwest and Western Central Pacific: Japan to the Philippines. Shelf and upper slope. 45–300 m.

*Dipturus trachydermus* (Krefft and Stehmann, 1974). Roughskin skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 20–450 m.

*Dipturus wengi* (Séret and Last, 2008). Weng's skate.

Eastern Indian and Western Pacific: Australia and Indonesia. Upper to mid slope. 486–1 165 m.

*Fenestraja atripinna* (Bigelow and Schroeder, 1950). Blackfin pygmy skate.

Western Central Atlantic: North Carolina (United States) to Cuba. Upper to mid slope. 366–951 m.

*Fenestraja cubensis* (Bigelow and Schroeder, 1950). Cuban pygmy skate.

Western Central Atlantic: Florida (United States), Bahamas and Cuba. Upper to mid slope. 440–869 m.

*Fenestraja ishiyamai* (Bigelow and Schroeder, 1962). Plain pygmy skate.

Scattered records in the Western Central Atlantic. Upper to mid slope. 503–950 m.

*Fenestraja maceachrani* (Séret, 1989). Madagascar pygmy skate.

Western Indian: Madagascar. Upper to mid slope. 600–765 m.

*Fenestraja mamillidens* (Alcock, 1889). Prickly skate.

Eastern Indian: Bay of Bengal. Mid slope. 1 093 m.

*Fenestraja plutonia* (Garman, 1881). Pluto skate.

Patchy in the Western Central Atlantic. Upper to mid slope. 293–1 024 m.

*Fenestraja sibogae* (Weber, 1913). Siboga pygmy skate.

Western Central Pacific: Bali Sea, Indonesia. Upper slope. 290 m.

*Fenestraja sinusmexicanus* (Bigelow and Schroeder, 1950). Gulf of Mexico pygmy skate.

Patchy in the Western Central Atlantic. Shelf and upper to mid slope. 56–1 096 m.

***Gurgesiella atlantica* (Bigelow and Schroeder, 1962).** Atlantic pygmy skate.

Western Atlantic: Central and South America. Upper to mid slope. 247–960 m.

***Gurgesiella dorsalifera* (McEachran and Compagno, 1980).** Onefin skate.

Southwest Atlantic: Brazil. Upper to mid slope. 400–800 m.

***Gurgesiella furvescens* (de Buen, 1959).** Dusky finless skate.

Southeast Pacific: Galapagos Islands and Peru to Chile. Upper to mid slope. 300–960 m.

***Leucoraja compagnoi* (Stehmann, 1995).** Tigertail skate.

Southeast Atlantic and Western Indian: South Africa. Upper slope. 497–625 m.

***Leucoraja fullonica* (Linnaeus, 1758).** Shagreen skate.

Northeast (and marginally into the Eastern Central) Atlantic, including the western Mediterranean. Shelf and upper to mid slope. 30–800 m.

***Leucoraja garmani* (Whitley, 1939).** Rosette skate.

Northwest and Western Central Atlantic: Cape Cod to Florida (United States). Shelf and upper slope. 37–530 m.

***Leucoraja lentiginosa* (Bigelow and Schroeder, 1951).** Freckled skate.

Western Central Atlantic: northern Gulf of Mexico. Shelf and upper slope. 53–588 m.

***Leucoraja leucosticta* (Stehmann, 1971).** Whitedappled skate.

Eastern Central and Southeast Atlantic: West Africa. Shelf and upper slope. 70–600 m.

***Leucoraja melitensis* (Clark, 1926).** Maltese skate.

Southwestern and south-central Mediterranean. Shelf and upper to mid slope. 60–800 m (more common 400–800 m).

***Leucoraja naevus* (Müller and Henle, 1841).** Cuckoo skate.

Reasonably wide-ranging in the Northeast and Eastern Central Atlantic, including the Mediterranean. Shelf and upper slope. 20–500 m.

***Leucoraja pristispina* (Last, Stehmann and Séret, 2008).** Sawback skate.

Eastern Indian: western Australia. Upper slope. 202–504 m.

***Leucoraja wallacei* (Hulley, 1970).** Yellowspot or blancmange skate.

Southeast Atlantic and Western Indian: southern Africa. Shelf and upper slope. 70–500 m (most common 150–300 m).

***Leucoraja yucatanensis* (Bigelow and Schroeder, 1950).** Yucatan skate.

Western Central Atlantic: Central America. Outermost shelf and upper slope. 192–457 m.

***Malacoraja kreffti* (Stehmann, 1978).** Krefft's skate.

Northeast Atlantic: Rockall Trough and Iceland. Mid slope and deep-sea rises. 1 200 m.

***Malacoraja obscura* (Carvalho, Gomes and Gadig, 2006).** Brazilian soft Skate.

Southwest Atlantic: Brazil. Mid slope. 808–1 105 m.

***Malacoraja senta* (Garman, 1885).** Smooth skate.

Northwest Atlantic: Canada and United States. Shelf and upper to mid slope. 46–914 m.

***Malacoraja spinacidermis* (Barnard, 1923).** Prickled or roughskin skate.

Wide-ranging but patchy in the Eastern Atlantic. Upper to deep slope. 450–1 568 m.

***Neoraja africana* (Stehmann and Séret, 1983).** West African pygmy skate.

Eastern Central Atlantic: Gabon and Western Sahara. Mid to deep slope. 900–1 550 m.

***Neoraja caerulea* (Stehmann, 1976).** Blue pygmy skate.

Northeast Atlantic: Rockall Trough. Upper to mid slope and deep-sea rises. 600–1 262 m.

***Neoraja carolinensis* (McEachran and Stehmann, 1984).** Carolina pygmy skate.

Western Central Atlantic: North Carolina to Florida (United States). Upper to mid slope. 695–1 010 m.

***Neoraja iberica* (Stehmann, Séret, Costa and Baro, 2008).** Iberian pygmy skate

Northeast Atlantic: Iberian Peninsula. Upper slope. 270–670 m.

***Neoraja stehmanni* (Hulley, 1972).** South African pygmy skate.

Southeast Atlantic: South Africa. Upper to mid slope. 292–1 025 m (most common > 600 m).

***Okamejei arafurensis* (Last and Gledhill, 2008).** Arafura skate.

Western Central Pacific and Eastern Indian: northern Australia. Outermost shelf and upper slope. 179–298 m.

***Okamejei heemstrai* (McEachran and Fechhelm, 1982).** Narrow or East African skate.

Western Indian: East Africa. Upper slope. 200–500 m.

*Okamejei leptoura* (Last and Gledhill, 2008). Thintail skate.

Eastern Indian: western Australia. Upper slope. 202–735 m.

*Raja africana?* (Capapé, 1977). African skate.

Disjunct range in the Mediterranean (Tunisia) and Eastern Central Atlantic (Mauritania).

Shelf and upper slope. 50–400 m. Homonym, requires a replacement name.

*Raja bahamensis* (Bigelow and Schroeder, 1965). Bahama skate.

Western Central Atlantic: Florida (United States) and Bahamas. Upper slope. 366–411 m.

*Raja binoculata* (Girard, 1854). Big skate

Wide-ranging in the Northeast and Eastern Central Pacific (and marginally into the Northwest Pacific). Shelf and upper to mid slope. 3–800 m.

*Raja inornata* (Jordan and Gilbert, 1880). California skate.

Eastern Central Pacific: western United States and Mexico. Shelf and upper to mid slope. 17–671 m.

*Raja maderensis* (Lowe, 1841). Madeira skate.

Eastern Central Atlantic: Madeira. Shelf and upper slope. ?–500 m.

*Raja polystigma* (Regan, 1923). Speckled skate.

Western Mediterranean. Outer shelf and upper slope. 100–400 m.

*Raja rhina* (Jordan and Gilbert, 1880). Longnose skate.

Wide-ranging in the Northeast and Eastern Central Pacific. Shelf and upper to mid slope. 9–1 069 m.

*Raja stellulata* (Jordan and Gilbert, 1880). Pacific starry skate.

Eastern Central Pacific: United States and Mexico. Shelf and upper slope. 18–732 m (usually <100 m). Records from Alaska and Bering Sea likely other species (i.e. *B. parmifera*).

*Raja straeleni* (Poll, 1951). Biscuit skate.

Wide-ranging in the Eastern Central and Southeast Atlantic: West Africa; Western Indian: southern Africa. Shelf and upper slope. 80–690 m (mostly 200–300 m).

*Rajella annandalei* (Weber, 1913). Indonesian round or Annandale's skate.

Western Central Pacific: eastern Indonesia. Upper to mid slope. 400–830 m.

*Rajella barnardi* (Norman, 1935). Bighorn skate.

Wide-ranging in the Eastern Atlantic: West Africa. Outer shelf and upper to deep slope. 102–1 700 m. Includes the junior synonym *Raja confundens* Hulley, 1970.

*Rajella bathyphila* (Holt and Byrne, 1908). Deepwater skate.

Wide-ranging in the North Atlantic. Upper to deep slope and abyssal plains. 600–2 050 m (mostly >1 000 m).

*Rajella bigelowi* (Stehmann, 1978). Bigelow's skate.

Wide-ranging but patchy in the North and Central Atlantic. Upper to deep slope, deep-sea rises and abyssal plains. 650–4 156 m (mostly >1 500 m).

*Rajella caudaspinosa* (von Bonde and Swart, 1923). Munchkin skate.

Southeast Atlantic and Western Indian: Namibia and South Africa. Upper slope. 310–520 m.

*Rajella challengereri* (Last and Stehmann, 2008). Challenger skate.

Eastern Indian and Southwest Pacific: southern Australia. Mid slope. 860–1 500 m.

*Rajella dissimilis* (Hulley, 1970). Ghost skate.

Patchy in the Eastern Atlantic. Upper to deep slope. 400–1 570 m.

*Rajella eisenhardti* (Long and McCosker, 1999). Galapagos gray skate.

Southeast Pacific: Galapagos Islands. Mid slope. 757–907 m.

*Rajella fuliginea* (Bigelow and Schroeder, 1954). Sooty skate.

Patchy in the Western Central Atlantic. Upper to mid slope. 731–1 280 m.

*Rajella fyllae* (Luetken, 1888). Round skate.

Wide-ranging in the Northeast and Northwest Atlantic. Outer shelf and upper to deep slope. 170–2 050 m (average depth range 400–800 m).

*Rajella kukujevi* (Dolganov, 1985). Mid-Atlantic skate.

Patchy in the Northeast Atlantic. Mid slope. 750–1 341 m.

*Rajella leopardus* (von Bonde and Swart, 1923). Leopard skate.

Patchy in the Eastern Central and Southeast Atlantic and possibly Western Indian (South Africa). Outer shelf and upper to deep slope. 130–1 920 m.

*Rajella nigerrima* (de Buen, 1960). Blackish skate.

Southeast Pacific: Ecuador to Chile. Upper to mid slope. 590–1 000 m.

*Rajella purpuriventris* (Bigelow and Schroeder, 1962). Purplebelly skate.

Patchy in the Western Central Atlantic. Upper to deep slope. 732–2 010 m.

*Rajella ravidula* (Hulley, 1970). Smoothback skate.

Eastern Central and Southeast Atlantic: Morocco and South Africa. Mid slope. 1 000–1 250 m.

*Rajella sadowskyii* (Krefft and Stehmann, 1974). Brazilian skate.

Southwest Atlantic and Southeast Pacific: southern South America. Mid slope. 1 200 m.

*Rostroraja alba* (Lacepede, 1803). White, bottlenose or spear-nose skate.

Wide-ranging in the Eastern Atlantic, including the western Mediterranean, and the Western Indian (to Mozambique). Shelf and upper slope. 30–600 m.

*Zearaja chilensis* (Guichenot, 1848). Yellownose skate.

Southwest Atlantic and Southeast Pacific: southern South America. Shelf and upper slope. 28–500 m. Includes the junior synonym *Dipturus flavirostris* (Philippi, 1892).

*Zearaja nasutus* (Banks, in Müller and Henle, 1841). Rough skate.

Southwest Pacific: New Zealand. Shelf and upper to mid slope. 10–1 500 m.

### Family Anacanthobatidae. Legskates.

*Anacanthobatis americanus* (Bigelow and Schroeder, 1962). American legskate.

Western Atlantic: Caribbean and the northern coast of South America. Outermost shelf and upper to mid slope. 183–915 m.

*Anacanthobatis donghaiensis* (Deng, Xiong and Zhan, 1983). East China legskate.

Northwest Pacific: East China Sea. Upper to mid slope. 200–1 000 m.

*Anacanthobatis folirostris* (Bigelow and Schroeder, 1951). Leafnose legskate.

Western Central Atlantic: northern Gulf of Mexico. Upper slope. 300–512 m.

*Anacanthobatis longirostris* (Bigelow and Schroeder, 1962). Longnose legskate.

Western Central Atlantic: northern Gulf of Mexico and Caribbean. Upper to mid slope. 520–1 052 m.

*Anacanthobatis marmoratus* (von Bonde and Swart, 1924). Spotted legskate.

Western Indian: South Africa and Mozambique. Upper slope. 230–322 m.

*Anacanthobatis nanhaiensis* (Meng and Li, 1981). South China legskate.

South China Sea. Validity uncertain.

*Anacanthobatis ori* (Wallace, 1967). Black legskate.

Western Indian: Mozambique and Madagascar. Mid to deep slope. 1 000–1 725 m.

*Anacanthobatis stenosoma* (Li and Hu, 1982). Narrow legskate.

South China Sea. Validity uncertain.

*Cruriraja andamanica* (Lloyd, 1909). Andaman legskate.

Patchy in the Indian Ocean: Tanzania and the Andaman Sea. Upper slope. 274–511 m.

*Cruriraja atlantis* (Bigelow and Schroeder, 1948). Atlantic legskate.

Western Central Atlantic: Florida (United States) to Cuba. Upper to mid slope. 512–778 m.

*Cruriraja cadenati* (Bigelow and Schroeder, 1962). Broadfoot legskate.

Western Central Atlantic: Florida (United States) and Puerto Rico. Upper to mid slope. 457–896 m.

*Cruriraja durbanensis* (von Bonde and Swart, 1924). Smoothnose legskate.

Southeast Atlantic: South Africa. Mid slope. 860 m. Note types collected off Northern Cape Province of South Africa, not off KwaZulu-Natal as suggested by the specific name.

*Cruriraja parcomaculata* (von Bonde and Swart, 1924). Roughnose legskate

Southeast Atlantic and Western Indian: Namibia and South Africa. Outer shelf and upper slope. 150–620 m.

*Cruriraja poeyi* (Bigelow and Schroeder, 1948). Cuban legskate.

Scattered records throughout the Western Central Atlantic. Upper to mid slope. 366–870 m.

*Cruriraja rugosa* (Bigelow and Schroeder, 1958). Rough legskate.

Western Central Atlantic: Gulf of Mexico and Caribbean. Upper to mid slope. 366–1 007 m.

*Cruriraja triangularis* (Smith, 1964). Triangular legskate.

Western Indian: South Africa and Mozambique. Upper slope. 220–675 m.

*Sinobatis borneensis* (Chan, 1965). Borneo legskate.

Patchy in the Northwest and Western Central Pacific. Upper to mid slope. 475–835 m.

*Sinobatis bulbicauda* (Last and Séret, 2008). Western Australian legskate.

Eastern Indian: western Australia; Western Central Pacific: Indonesia. Outer shelf and upper to mid slope. 150–1 125 m (mostly 400–800 m).

*Sinobatis caerulea* (Last and Séret, 2008). Blue legskate.

Eastern Indian: western Australia. Upper to mid slope. 482–1 168 m.

*Sinobatis filicauda* (Last and Séret, 2008). Eastern Australian legskate.

Western Central Pacific: northeastern Australia. Upper to mid slope. 606–880 m.

*Sinobatis melanosoma* (Chan, 1965). Blackbodied legskate.

Northwest and Western Central Pacific: South China Sea. Mid slope. 900–1 100 m.

#### **Family Plesiobatidae. Giant Stingarees.**

*Plesiobatis daviesi* (Wallace, 1967). Deepwater stingray or giant stingaree.

Patchy in the Indo-West and Central Pacific. Shelf (single record at 44 m off Mozambique) and upper slope. 44–680 m (mostly 275–680 m).

#### **Family Urolophidae. Stingarees.**

*Urolophus expansus* (McCulloch, 1916). Wide stingaree.

Eastern Indian: southern Australia. Outer shelf and upper slope. 130–420 m (mainly 200–300 m).

*Urolophus piperatus* (Séret and Last, 2003). Coral Sea stingaree.

Western Central Pacific: northeastern Australia. Upper slope and deep reefs. 170–370 m.

#### **Family Hexatrygonidae. Sixgill Stingrays.**

*Hexatrygon bickelli* (Heemstra and Smith, 1980). Sixgill stingray.

Wide-ranging but patchy in the Indo-West and Central Pacific. Upper to mid slope (although some shallow water and beach stranding records). 362–1 120 m.

#### **Family Dasyatidae. Whiptail Stingrays.**

*Dasyatis brevicaudata* (Hutton, 1875). Shorttail or smooth stingray.

Patchy in the Indo-West Pacific (New Zealand, Australia and southern Africa). Inshore and shelf in Australia and New Zealand, outermost shelf and upper slope off South Africa. Intertidal–160 m (New Zealand, Australia), 180–480 m (southern Africa).

### **Superorder Galeomorphii. Galeomorph Sharks.**

#### **Order Heterodontiformes. Bullhead Sharks.**

##### **Family Heterodontidae. Bullhead Sharks.**

*Heterodontus ramalheira* (Smith, 1949). Whitespotted bullhead shark.

Western Indian: East Africa and Arabian Sea. Outer shelf and uppermost slope. 40–275 m (mostly >100 m).

## **Order Orectolobiformes. Carpetsharks.**

### **Family Parascylliidae. Collared Carpetsharks.**

*Cirrhoscyllium japonicum* (Kamohara, 1943). Saddled carpetshark.

Northwest Pacific: Japan. Uppermost slope. 250–290 m.

*Parascyllium sparsimaculatum* (Goto and Last, 2002). Ginger carpetshark.

Eastern Indian: Australia. Upper slope. 245–435 m.

## **Order Lamniformes. Mackerel Sharks.**

### **Family Odontaspidae. Sand Tiger Sharks.**

*Odontaspis ferox* (Risso, 1810). Smalltooth sand tiger or bumpytail raggedtooth.

Wide-ranging but patchy in the Atlantic, including the Mediterranean, and the Indo-Pacific. Possibly circumglobal in warm-temperate and tropical waters. Shelf and upper slope. 13–880 m.

*Odontaspis noronhai* (Maul, 1955). Bigeye sand tiger.

Scattered records in the Central and Southwest Atlantic and the Central Pacific. Possibly circumglobal in deep tropical seas. Upper to mid slope as well as mesopelagic. 600–1 000 m.

### **Family Pseudocarchariidae. Crocodile Sharks.**

*Pseudocarcharias kamoharai* (Matsubara, 1936). Crocodile shark.

Cosmopolitan in tropical and warm temperate waters. Epipelagic and mesopelagic. 0–590 m.

### **Family Mitsukurinidae. Goblin Sharks.**

*Mitsukurina owstoni* (Jordan, 1898). Goblin shark.

Wide-ranging but patchy in the Atlantic and the Indo-Pacific. Shelf, upper to mid slope and seamounts. 0–1 300 m (mainly 270–960 m).

### **Family Alopiidae. Thresher Sharks.**

*Alopias superciliosus* (Lowe, 1839). Bigeye thresher.

Cosmopolitan in tropical and warm temperate waters. Shelf, epipelagic and mesopelagic. 0–732 m (mostly > 100 m).

### **Family Cetorhinidae. Basking Sharks.**

*Cetorhinus maximus* (Gunnerus, 1765). Basking shark.

Cosmopolitan in cold to warm-temperate waters. Shelf and upper to mid slope, epipelagic and mesopelagic. 0–904 m.

## **Order Carcharhiniformes. Ground Sharks.**

### **Family Scyliorhinidae. Catsharks.**

*Apristurus albisoma* (Nakaya and Séret, 1999). White catshark.

Western Central Pacific: Norfolk and Lord Howe Ridges. Submarine ridges. 935–1 564 m.

*Apristurus ampliceps* Sasahara, (Sato and Nakaya, 2008). Roughskin catshark.

Eastern Indian and Southwest Pacific: Australia and New Zealand. Mid to deep slope and submarine ridges. 800–1 503 m.

*Apristurus aphyodes* (Nakaya and Stehmann, 1998). White ghost catshark.

Northeast Atlantic: Atlantic Slope. Mid to deep slope. 1 014–1 800 m.

*Apristurus australis* (Sato, Nakaya and Yorozu, 2008). Pinocchio catshark.

Eastern Indian and Western Pacific: Australia. Upper to mid slope and seamounts. 486–1 035 m.

*Apristurus brunneus* (Gilbert, 1892). Brown catshark.

Eastern Pacific: Alaska to Mexico, and possibly Southeast Pacific. Shelf and upper to mid slope. 33–1 298 m.

*Apristurus bucephalus* (White, Last and Pogonoski, 2008). Bighead catshark.

Eastern Indian: western Australia. Mid slope. 920–1 140 m.

*Apristurus canutus* (Springer and Heemstra, in Springer, 1979). Hoary catshark.

Western Central Pacific: Caribbean Islands and Venezuela. Upper to mid slope. 521–915 m.

*Apristurus exsanguis* (Sato, Nakaya and Stewart, 1999). Pale catshark.

Southwest Pacific: New Zealand. Upper to mid slope and submarine ridges. 573–1 200 m.

*Apristurus fedorovi* (Dolganov, 1985). Stout catshark.

Northwest Pacific: Japan. Mid slope. 810–1 430 m.

*Apristurus gibbosus* (Meng, Chu and Li, 1985). Humpback catshark.

Northwest Pacific: South China Sea off southern China. Mid slope. 913 m.

*Apristurus herklotsi* (Fowler, 1934). Longfin catshark.

Northwest and Western Central Pacific: Japan, China and the Philippines. Upper to mid slope. 520–910 m.

*Apristurus indicus* (Brauer, 1906). Smallbelly catshark.

Western Indian: Arabian Sea. Records off southern Africa probably erroneous. Mid to deep slope. 1 289–1 840 m.

*Apristurus internatus* (Deng, Xiong and Zhan, 1988). Shortnose demon catshark.

Northwest Pacific: East China Sea off China. Upper slope. 670 m.

*Apristurus investigatoris* (Misra, 1962). Broadnose catshark.

Eastern Indian: Andaman Sea. Mid slope. 1 040 m.

*Apristurus japonicus* (Nakaya, 1975). Japanese catshark.

Northwest Pacific: Japan. Mid slope. 820–915 m.

*Apristurus kampae* (Taylor, 1972). Longnose catshark.

Eastern Pacific: United States and Mexico. Provisional from the Galapagos Islands. Outer shelf and upper to deep slope. 180–1 888 m.

*Apristurus laurussoni* (Saemundsson, 1922). Iceland catshark.

Wide-ranging but patchy in the North and Central Atlantic. Upper to deep slope. 560–2 060 m. Includes the junior synonyms *Apristurus atlanticus* (Koefoed, 1932) and *Apristurus maderensis* Cadenat and Maul, 1966.

*Apristurus longicephalus* (Nakaya, 1975). Longhead catshark.

Patchy in the Indo-West Pacific. Upper to mid slope and submarine ridges. 500–1 140 m.

*Apristurus macrorhynchus* (Tanaka, 1909). Flathead catshark.

Northwest Pacific: Japan, China and Taiwan. Upper to mid slope. 220–1 140 m.

*Apristurus macrostomus* (Meng, Chu, and Li, 1985). Broadmouth catshark.

Northwest Pacific: South China Sea off China. Mid slope. 913 m.

*Apristurus manis* (Springer, 1979). Ghost catshark.

Patchy records in the North and Southeast Atlantic. Upper to deep slope. 658–1 740 m.

*Apristurus melanospilus* (Iglésias, Nakaya and Stehmann, 2004). Black roughscale catshark.

Patchy in the Atlantic and Indo-West Pacific. Upper to deep slope and submarine ridges. 512–1 520 m.

*Apristurus microps* (Gilchrist, 1922). Smalleye catshark.

Patchy in the North and Southeast Atlantic. Upper to deep slope. 700–2 200 m.

*Apristurus micropterygeus* (Meng, Chu and Li, in Chu, Meng, and Li, 1986). Smalldorsal catshark.

Northwest Pacific: South China Sea off China. Mid slope. 913 m.

*Apristurus nasutus* (de Buen, 1959). Largenose catshark.

Eastern Pacific: patchy off Central and South America. Eastern Central Atlantic record probably erroneous. Upper to mid slope. 400–925 m.

*Apristurus parvipinnis* (Springer and Heemstra, in Springer, 1979). Smallfin catshark.

Western Central Atlantic: Gulf of Mexico, Caribbean and northern South America. Upper to mid slope. 636–1 115 m.

*Apristurus pinguis* (Deng, Xiong and Zhan, 1983). Fat catshark.

Patchy in the Eastern Indian and Western Pacific. Mid to deep slope and submarine ridges. 996–2 057 m.

*Apristurus platyrhynchus* (Tanaka, 1909). Spatulasnout catshark

Patchy in the Eastern Indian and Western Pacific. Upper to mid slope and submarine ridges. 400–1 080 m. Includes the junior synonyms *Apristurus acanutus* Chu, Meng and Li, in Meng, Chu and Li, 1985 and *Apristurus verweyi* (Fowler, 1934).

*Apristurus profundorum* (Goode and Bean, 1896). Deepwater catshark.

Northwest Atlantic off United States, possibly Eastern Central Atlantic off West Africa. Deep slope. ~1 500 m. Records from the Indian Ocean require confirmation.

*Apristurus riveri* (Bigelow and Schroeder, 1944). Broadgill catshark.

Western Central Atlantic: Gulf of Mexico, Caribbean and northern South America. Upper to mid slope. 732–1 461 m.

*Apristurus saldanha* (Barnard, 1925). Saldanha catshark.

Southeast Atlantic: southern Africa. Upper to mid slope. 344–1 009 m.

*Apristurus sibogae* (Weber, 1913). Pale catshark.

Western Central Pacific: Indonesia. Upper slope. 655 m.

*Apristurus sinensis* (Chu and Hu, in Chu, Meng, Hu and Li, 1981). South China catshark.

Eastern Indian and Western Pacific: Australia and South China Sea. Upper to mid slope. 940–1 290 m.

*Apristurus spongiceps* (Gilbert, 1895). Spongehead catshark.

Patchy in the Western and Central Pacific: Indonesia and Hawaii. Upper to mid slope. 572–1 482 m.

*Apristurus stensenii* (Springer, 1979). Panama ghost catshark.

Eastern Central Pacific: Panama. Mid slope. 915–975 m.

*Asymbolus galacticus* (Séret and Last, 2008). New Caledonia spotted catshark.

Western Central Pacific: New Caledonia. Upper slope. 235–550 m.

*Asymbolus pallidus* (Last, Gomon and Gledhill, in Last, 1999). Pale spotted catshark.

Western Central Pacific: northeastern Australia. Upper slope. 225–400 m.

*Bythaelurus? alcocki* (Garman, 1913). Arabian catshark.

Eastern Indian: Arabian Sea. Mid slope. 1 134–1 262 m. Original and only specimen has been lost, and thus placement of the species in this genus is tentative.

*Bythaelurus canescens* (Günther, 1878). Dusky catshark.

Southeast Pacific: Peru and Chile. Upper slope. 250–700 m.

*Bythaelurus clevai* (Séret, 1987). Broadhead catshark.

Eastern Indian: Madagascar. Upper slope. 400–500 m.

*Bythaelurus dawsoni* (Springer, 1971). Dawson's catshark.

Southwest Pacific: New Zealand. Shelf, upper to mid slope. 50–790 m (most common 300–700 m).

*Bythaelurus hispidus* (Alcock, 1891). Bristly catshark.

Eastern Indian: India and the Andaman Islands. Upper to mid slope. 293–766 m.

*Bythaelurus immaculatus* (Chu and Meng, in Chu, Meng, Hu and Li, 1982). Spotless catshark.

Northwest Pacific: South China Sea. Upper to mid slope. 534–1 020 m.

*Bythaelurus incanus* (Last and Stevens, 2008). Dusky catshark.

Eastern Indian: northwestern Australia. Mid slope. 900–1 000 m.

*Bythaelurus lutarius* (Springer and D'Aubrey, 1972). Mud catshark.

Western Indian: patchy off East Africa. Upper to mid slope. 338–766 m.

*Cephaloscyllium albipinnum* (Last, Motomura and White, 2008). Whitefin swellshark.

Eastern Indian and Southwest Pacific: southern Australia. Outer shelf and upper slope. 126–554 m.

*Cephaloscyllium cooki* (Last, Séret and White, 2008). Cook's swellshark.

Eastern Indian: northwestern Australia and Indonesia. Upper slope. 223–300 m.

*Cephaloscyllium fasciatum* (Chan, 1966). Reticulated swellshark.

Patchy in the Western Pacific. Upper slope. 219–450 m.

*Cephaloscyllium hiscosellum* (White and Ebert, 2008). Australian reticulate swellshark.

Eastern Indian: northwestern Australia. Upper slope. 294–420 m.

*Cephaloscyllium isabellum* (Bonnaterre, 1788). Draughtsboard shark.

Southwest Pacific: New Zealand. Shelf and upper slope. Inshore–690 m.

*Cephaloscyllium signourum* (Last, Séret and White, 2008). Flagtail swellshark.

Western Central Pacific: northeastern Australia (and possibly elsewhere in Oceania). Upper slope and submarine ridges. 480–700 m.

*Cephaloscyllium silasi* (Talwar, 1974). Indian swellshark.

Western Indian: India. Upper slope. ~300 m.

*Cephaloscyllium speculum* (Last, Séret and White, 2008). Speckled swellshark.

Eastern Indian: northwestern Australia. Outer shelf and upper slope. 150–455 m.

*Cephaloscyllium sufflans* (Regan, 1921). Balloon shark.

Western Indian: South Africa and Mozambique. Shelf and upper slope. 40–440 m.

*Cephaloscyllium variegatum* (Last and White, 2008). Saddled swellshark.

Western Central and Southwest Pacific: eastern Australia. Outer shelf and upper slope. 114–606 m.

*Cephaloscyllium zebra* (Last and White, 2008). Narrowbar swellshark.

Western Central Pacific: northeastern Australia. Upper slope. 444–454 m.

*Cephalurus cephalus* (Gilbert, 1892). Lollipop catshark.

Eastern Central Pacific: Mexico. Outer shelf and upper to mid slope. 155–927 m.

*Figaro boardmani* (Whitley, 1928). Australian sawtail catshark.

Eastern Indian and Western Pacific: Australia. Outer shelf and upper slope. 150–640 m.

*Figaro striatus* (Gledhill, Last and White, 2008). Northern sawtail catshark.

Western Central Pacific: northeastern Australia. Upper slope. 300–419 m.

*Galeus antillensis* (Springer, 1979). Antilles catshark.

Western Central Atlantic: Straits of Florida and the Caribbean. Upper slope. 293–658 m.

*Galeus arai* (Nichols, 1927). Roughtail catshark.

Northwest and Western Central Atlantic: United States, Gulf of Mexico, Caribbean and Central America. Upper slope. 292–732 m.

*Galeus atlanticus* (Vaillant, 1888). Atlantic sawtail catshark.

Patchy in the Northeast Atlantic and Mediterranean. Upper slope. 400–600 m.

*Galeus cadenati* (Springer, 1966). Longfin sawtail catshark.

Western Central Atlantic: off Panama and Colombia. Upper slope. 439–548 m.

*Galeus eastmani* (Jordan and Snyder, 1904). Gecko catshark.

Patchy in the Northwest Pacific. Habitat information not available.

*Galeus gracilis* (Compagno and Stevens, 1993). Slender sawtail catshark.

Eastern Indian and Western Central Pacific: patchy across northern Australia. Upper slope. 290–470 m.

*Galeus longirostris* (Tachikawa and Taniuchi, 1987). Longnose sawtail catshark.

Northwest Pacific: Japan. Upper slope. 350–550 m.

*Galeus melastomus* (Rafinesque, 1810). Blackmouth catshark.

Wide-ranging in the Northeast and Western Central Atlantic and the Mediterranean. Shelf and upper to mid slope. 55–1 440 m (mainly 200–500 m).

*Galeus mincaronei* (Soto, 2001). Brazilian sawtail catshark.

Southwest Atlantic: southern Brazil. Deep reefs on upper slope. 236–600 m.

*Galeus murinus* (Collett, 1904). Mouse catshark.

Northeast Atlantic: Iceland and the Faeroe Islands. Upper to mid slope. 380–1 250 m.

*Galeus nippensis* (Nakaya, 1975). Broadfin sawtail catshark.

Northwest Pacific: Japan. Upper slope and submarine ridges. 362–540 m.

*Galeus piperatus* (Springer and Wagner, 1966). Peppered catshark.

Eastern Central Pacific: Gulf of California. Upper to mid slope. 275–1 326 m.

*Galeus polli* (Cadenat, 1959). African sawtail catshark.

Wide-ranging in the Eastern Atlantic off West Africa. Outer shelf and upper slope. 159–720 m.

*Galeus priapus* (Séret and Last, 2008).

Western Central Pacific: New Caledonia and Vanuatu. Upper to mid slope. 262–830 m.

*Galeus schultzi* (Springer, 1979). Dwarf sawtail catshark

Western Central Pacific: the Philippines. Shelf (one record) and upper slope. 50–431 m (single record at 50 m, usually 329–431 m).

*Galeus springeri* (Konstantinou and Cozzi, 1998). Springer's sawtail catshark.

Western Central Atlantic: Greater and Lesser Antilles. Upper slope. 457–699 m.

*Holohalaelurus favus* (Human, 2006). Honeycomb or Natal izak.

Western Indian: southern Africa. Upper to mid slope. 200–1 000 m.

*Holohalaelurus grennian* (Human, 2006). East African spotted catshark.

Scattered records in the Western Indian off East Africa. Upper slope. 238–300 m.

*Holohalaelurus melanostigma* (Norman, 1939). Tropical izak catshark.

Western Indian: off East Africa. Upper slope. 607–658 m.

*Holohalaelurus punctatus* (Gilchrist and Thompson, 1914). African spotted catshark

Southeast Atlantic and Western Indian: southern Africa. Upper slope. 220–420 m.

*Holohalaelurus regani* (Gilchrist, 1922). Izak catshark.

Southeast Atlantic and Western Indian: southern Africa. Shelf and upper to mid slope. 40–910 m (mainly ~100–300 m).

*Parmaturus albimarginatus* (Séret and Last, 2007). Whitetip catshark.

Western Central Pacific: New Caledonia. Upper slope. 590–732 m.

*Parmaturus alipenesis* (Séret and Last, 2007). White-clasper catshark.

Western Central Pacific: New Caledonia. Upper slope. 688–732 m.

*Parmaturus bigus* (Séret and Last, 2007). Beige catshark.

Western Central Pacific: northeastern Australia. Upper slope. 590–606 m.

*Parmaturus campechiensis* (Springer, 1979). Campeche catshark.

Western Central Atlantic: Bay of Campeche in the Gulf of Mexico. Mid slope. 1 097 m.

*Parmaturus lanatus* (Séret and Last, 2007). Velvet catshark.

Eastern Indian: Indonesia. Mid slope. 840–855 m.

*Parmaturus macmillani* (Hardy, 1985). New Zealand filetail.

Southwest Pacific: New Zealand; Western Indian: south of Madagascar. Submarine ridges. 950–1 003 m (may also occur deeper).

*Parmaturus melanobranchius* (Chan, 1966). Blackgill catshark.

Western Pacific: East and South China Seas. Upper to mid slope. 540–835 m.

*Parmaturus pilosus* (Garman, 1906). Salamander shark.

Northwest Pacific: Japan and East China Sea. Upper to mid slope. 358–895 m.

*Parmaturus xaniurus* (Gilbert, 1892). Filetail catshark.

Eastern Central Pacific: southern United States and Mexico. Outer shelf and upper to mid slope. Juveniles mesopelagic. 91–1 251 m.

*Pentanchus profundiculus* (Smith and Radcliffe, 1912). Onefin catshark.

Western Central Pacific: the Philippines. Upper to mid slope. 673–1 070 m.

*Schroederichthys maculatus* (Springer, 1966). Narrowtail catshark.

Western Central Atlantic: patchy off Central and northern South America. Outermost shelf and upper slope. 190–410 m.

*Schroederichthys saurisqualus* (Soto, 2001). Lizard catshark.

Southwest Atlantic: southwestern Brazil. Outer shelf (sporadic) and deep reefs on the upper slope. 122–500 m.

*Schroederichthys tenuis* (Springer, 1966). Slender catshark.

Restricted range in the Western Atlantic off Suriname and northern Brazil. Shelf and upper slope. 72–450 m.

*Scyliorhinus boa* (Goode and Bean, 1896). Boa catshark.

Western Central Atlantic: Caribbean and northern South America. Upper slope. 229–676 m.

*Scyliorhinus capensis* (Smith, in Müller and Henle, 1838). Yellowspotted catshark.

Southeast Atlantic and Western Indian: southern Africa. Shelf and upper slope. 26–695 m (mostly 200–400 m).

*Scyliorhinus cervigoni* (Maurin and Bonnet, 1970). West African catshark.

Eastern Atlantic off West Africa. Shelf and upper slope. 45–500 m.

*Scyliorhinus comoroensis* (Compagno, 1989). Comoro catshark.

Western Indian: Comoro Islands. Upper slope. 200–400 m.

*Scyliorhinus haeckelii* (Ribeiro, 1907). Freckled catshark.

Western Atlantic: off South America. Deep reefs on the shelf and upper slope. 37–439 m (mostly >250 m).

*Scyliorhinus hesperius* (Springer, 1966). Whitesaddled catshark.

Western Central Atlantic: Central America and Colombia. Upper slope. 274–457 m.

*Scyliorhinus meadi* (Springer, 1966). Blotched catshark.

Western Central Atlantic: United States, Gulf of Mexico, Cuba and Bahamas. Upper slope. 329–548 m.

*Scyliorhinus retifer* (Garman, 1881). Chain catshark.

Northwest and Western Central Atlantic: United States, Gulf of Mexico and Caribbean. Shelf and upper to mid slope. 75–754 m

*Scyliorhinus torrei* (Howell-Rivero, 1936). Dwarf catshark.

Western Central Atlantic: Florida (United States), Bahamas, Cuba and the Lesser Antilles. Upper slope. 229–550 m (most >366 m).

#### Family Proscylliidae. Finback Catsharks.

*Eridacnis barbouri* (Bigelow and Schroeder, 1944). Cuban ribbontail catshark.

Western Central Atlantic: Florida Straits and Cuba. Upper slope. 430–613 m.

*Eridacnis radcliffei* (Smith, 1913). Pygmy ribbontail catshark.

Wide-ranging but patchy in the Indo-West Pacific. Shelf, upper to mid slope. 71–766 m.

*Eridacnis sinuans* (Smith, 1957). African ribbontail catshark.

Western Indian off East Africa. Outermost shelf and upper slope. 180–480 m.

#### Family Pseudotriakidae. False Catsharks.

*Gollum attenuatus* (Garrick, 1954). Slender smoothhound.

Southwest Pacific: New Zealand. Outer shelf, upper slope and seamounts. 129–724 m (most common 300–600 m).

*Pseudotriakis microdon* (Capello, 1868). False catshark.

Wide-ranging but patchy in the Atlantic, Indo-West and Central Pacific. Shelf (occasional) and upper to deep slope. 200–1 900 m.

#### Family Triakidae. Houndsharks.

*Hemitriakis abdita* (Compagno and Stevens, 1993). Darksnout or deepwater sicklefin houndshark.

Western Central Pacific: northeastern Australia. Upper slope. 225–400 m.

*Iago garricki* (Fourmanoir, 1979). Longnose houndshark.

Patchy in the Eastern Indian and Western Central Pacific. Upper slope. 250–475 m.

*Iago omanensis* (Norman, 1939). Bigeye houndshark.

Northern Indian including the Red Sea; possibly in the Bay of Bengal (although this may represent a distinct species). Shelf and upper to mid (possibly to deep) slope. <110–1 000+ m (possibly to 2 195 m in Red Sea).

*Mustelus canis insularis* (Mitchell, 1815). Dusky smoothhound (island subspecies).

Western Central Atlantic: Caribbean and northern South America. Outer shelf and upper to mid slope. 137–808 m (mostly >200 m). Shelf subspecies (*M. c. canis*) prefers shallow waters (generally <18 m).

***Mustelus stevensi* (White and Last, 2008).** Western spotted gummy shark.

Eastern Indian: northwestern Australia. Outer shelf and upper slope. 121–735 m.

***Mustelus walkeri* (White and Last, 2008).** Eastern spotted gummy shark.

Western Central Pacific: northeastern Australia. Shelf and upper slope. 52–403 m.

#### **Family Carcharhinidae. Requiem Sharks.**

***Carcharhinus altimus* (Springer, 1950).** Bignose shark.

Wide-ranging but patchy in tropical and warm-temperate waters of the Atlantic and Indo-Pacific. Shelf and upper slope. 25–430 m (mostly >90 m, young shallower, to 25 m).

## APPENDIX 2

### Deep-sea chondrichthyan species for FAO Major Fishing Areas<sup>9</sup>

#### Sharks

Family	Species	FAO Major Fishing Areas
<b>Order Hexanchiformes</b>		
Chlamydoselachidae	<i>Chlamydoselachus africana</i>	47, 51
	<i>Chlamydoselachus anguineus</i>	27, 31, 34, 57, 61, 77, 81, 87
Hexanchidae	<i>Heptranchias perlo</i>	21, 27, 31, 34, 37, 41, 47, 51, 57, 61, 71, 81, 87
	<i>Hexanchus griseus</i>	21, 27, 31, 34, 37, 41, 47, 51, 57, 61, 67, 71, 77, 81, 87
	<i>Hexanchus nakamurai</i>	27, 31, 34? , 37, 51, 57, 61, 71, 81
<b>Order Squaliformes</b>		
Echinorhinidae	<i>Echinorhinus brucus</i>	21, 27, 31, 34, 37, 41, 47, 51, 57, 61, 71, 81
	<i>Echinorhinus cookei</i>	57, 61, 71, 77, 81, 87
Squalidae	<i>Cirrhigaleus asper</i>	31, 41, 47, 51, 57?, 77
	<i>Cirrhigaleus australis</i>	57, 81
	<i>Cirrhigaleus barbifer</i>	61, 71
	<i>Squalus albifrons</i>	71, 81
	<i>Squalus altipinnis</i>	57
	<i>Squalus blainville</i>	27, 34, 37, 47, 51, 61?
	<i>Squalus brevirostris</i>	61
	<i>Squalus bucephalus</i>	71, 81
	<i>Squalus chloroculus</i>	57, 81
	<i>Squalus crassispinus</i>	57
	<i>Squalus cubensis</i>	31, 41
	<i>Squalus edmundsi</i>	57
	<i>Squalus grahami</i>	71, 81
	<i>Squalus griffini</i>	81
	<i>Squalus hemipinnis</i>	57
	<i>Squalus japonicus</i>	61, 71
	<i>Squalus lalannei</i>	51
	<i>Squalus megalops</i>	27, 34, 37, 47, 51, 57, 61, 71, 81
	<i>Squalus melanurus</i>	71
	<i>Squalus mitsukurii</i>	31, 34?, 41, 47?, 51, 61, 71, 77, 87
	<i>Squalus montalbani</i>	57, 71, 81
	<i>Squalus nasutus</i>	57, 71
	<i>Squalus notocaudatus</i>	71
	<i>Squalus rancureli</i>	71
	<i>Squalus raoulensis</i>	81
Centrophoridae	<i>Centrophorus acus</i>	57, 61, 71
	<i>Centrophorus atromarginatus</i>	51, 57, 61, 71
	<i>Centrophorus granulosus</i>	27, 31?, 34, 37, 41?, 47, 51, 61, 71
	<i>Centrophorus harrissoni</i>	57, 71, 81
	<i>Centrophorus isodon</i>	51, 57, 61, 71
	<i>Centrophorus lusitanicus</i>	27, 34, 51, 61
	<i>Centrophorus niaukang</i>	57, 61, 71

<sup>9</sup> Area 18: Arctic Sea; Area 21: Atlantic, Northwest; Area 27: Atlantic, Northeast, Area 31: Atlantic, Western Central; Area 34: Atlantic, Eastern Central; Area 37: Mediterranean and Black Sea; Area 41: Atlantic, Southwest; Area 47: Atlantic, Southeast; Area 48: Atlantic, Antarctic; Area 51, Indian Ocean, Western; Area 57: Indian Ocean, Eastern; Area 58: Indian Ocean, Antarctic and Southern; Area 61: Pacific, Northwest; Area 67: Pacific, Northeast; Area 71: Pacific, Western Central; Area 77: Pacific, Eastern Central; Area 81: Pacific, Southwest; Area 87: Pacific, Southeast; Area 88: Pacific, Antarctic.

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
Centrophoridae	<i>Centrophorus seychellorum</i>	51
	<i>Centrophorus squamosus</i>	27, 34, 47, 51, 57, 61, 71, 81
<b>Order Squaliformes</b>		
Centrophoridae	<i>Centrophorus moluccensis</i>	51, 57, 61, 71, 81
	<i>Centrophorus tessellatus</i>	31, 61, 77
	<i>Centrophorus westraliensis</i>	57
	<i>Centrophorus zeehaani</i>	57, 81
	<i>Deania calcea</i>	27, 34, 47, 51, 57, 61, 81, 87
	<i>Deania hystricosum</i>	34?, 47, 61, 81
	<i>Deania profundorum</i>	31, 34, 47, 51, 71
	<i>Deania quadrispinosum</i>	47, 51, 57, 61?, 71, 81
Etmopteridae	<i>Aculeola nigra</i>	87
	<i>Centroscyllium excelsum</i>	61
	<i>Centroscyllium fabricii</i>	21, 27, 31, 34, 47
	<i>Centroscyllium granulatum</i>	87
	<i>Centroscyllium kamoharai</i>	57, 61, 71?, 81
	<i>Centroscyllium nigrum</i>	77, 87
	<i>Centroscyllium ornatum</i>	51, 57
	<i>Centroscyllium ritteri</i>	61
	<i>Etmopterus baxteri</i>	34?, 47, 51, 57, 81
	<i>Etmopterus bigelowi</i>	31, 34, 41, 47, 51, 57, 61, 71, 81, 87
	<i>Etmopterus brachyurus</i>	57, 61, 71
	<i>Etmopterus bullisi</i>	31
	<i>Etmopterus burgesi</i>	61
	<i>Etmopterus carteri</i>	31
	<i>Etmopterus caudistigmus</i>	71
	<i>Etmopterus decacuspidatus</i>	61
	<i>Etmopterus dianthus</i>	71
	<i>Etmopterus dislineatus</i>	71
	<i>Etmopterus evansi</i>	57, 71
	<i>Etmopterus fusus</i>	57
	<i>Etmopterus gracilispinis</i>	21, 31, 41, 47, 51
	<i>Etmopterus granulosus</i>	41, 87
	<i>Etmopterus hillianus</i>	21, 31
	<i>Etmopterus litvinovi</i>	87
	<i>Etmopterus lucifer</i>	41?, 47?, 51?, 57, 61, 71, 81
	<i>Etmopterus molleri</i>	51?, 61, 81
	<i>Etmopterus perryi</i>	31
	<i>Etmopterus polli</i>	31?, 34, 47
	<i>Etmopterus princeps</i>	21, 27, 34
	<i>Etmopterus pseudosqualiolus</i>	71
	<i>Etmopterus pusillus</i>	27, 31, 34, 41, 47, 51, 57, 61, 77, 81
	<i>Etmopterus pycnolepis</i>	87
	<i>Etmopterus robinsi</i>	31
	<i>Etmopterus schultzi</i>	31
	<i>Etmopterus sentosus</i>	51
	<i>Etmopterus spinax</i>	27, 34, 37, 47
	<i>Etmopterus splendidus</i>	61
	<i>Etmopterus unicolor</i>	47, 51, 57, 61, 81
	<i>Etmopterus villosus</i>	77
	<i>Etmopterus virens</i>	31
	<i>Miroscyllium sheikoi</i>	61
	<i>Trigonognathus kabeyai</i>	61, 77
Somniosidae	<i>Centroscymnus coelolepis</i>	21, 27, 31, 34, 37, 41, 47, 51, 57, 61, 71, 81

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Centroscymnus owstoni</i>	27, 31, 34, 41, 47, 57, 61, 81, 87
	<i>Centroselachus crepidater</i>	27, 34, 47, 51, 57, 61, 71, 81, 87
	<i>Proscymnodon macracanthus</i>	87
	<i>Proscymnodon plunketi</i>	51, 57, 81
	<i>Scymnodalatias albicauda</i>	47, 57, 58, 81
	<i>Scymnodalatias garricki</i>	27
	<i>Scymnodalatias sherwoodi</i>	81
	<i>Scymnodon ringens</i>	27, 34, 81?
	<i>Somniosus antarcticus</i>	41, 47, 57, 58, 81, 87
	<i>Somniosus longus</i>	61, 81
	<i>Somniosus microcephalus</i>	21, 27
	<i>Somniosus pacificus</i>	18, 61, 67, 77
	<i>Somniosus rostratus</i>	27, 31?, 34, 37
	<i>Zameus ichiharai</i>	61
	<i>Zameus squamulosus</i>	27, 31, 34, 41, 51, 57, 61, 71, 77, 81
Oxynotidae	<i>Oxynotus bruniensis</i>	57, 81
	<i>Oxynotus caribbaeus</i>	31
	<i>Oxynotus centrina</i>	27, 34, 37, 47
	<i>Oxynotus japonicus</i>	61
	<i>Oxynotus paradoxus</i>	27, 34
Dalatiidae	<i>Dalatias licha</i>	21, 27, 31, 34, 37, 47, 51, 57, 61, 71, 77
	<i>Euprotomicroides zantedeschia</i>	41, 47
	<i>Euprotomicrus bispinatus</i>	41, 47, 51, 57, 61, 71, 77, 81, 87
	<i>Heteroscymnoides marleyi</i>	47, 51, 87
	<i>Isistius brasiliensis</i>	31, 34, 41, 47, 51, 57, 61, 67?, 71, 77, 81, 87
	<i>Isistius labialis</i>	61
	<i>Mollisquama parini</i>	87
	<i>Squaliolus aliae</i>	57, 61, 71, 81
	<i>Squaliolus laticaudus</i>	27, 31, 34, 41, 51, 61, 71
<b>Order Squatiniformes</b>		
Squatiniidae	<i>Squatina aculeata</i>	27, 34, 47
	<i>Squatina africana</i>	51
	<i>Squatina albipunctata</i>	57, 71, 81
	<i>Squatina argentina</i>	41
	<i>Squatina formosa</i>	61, 71
	<i>Squatina pseudocellata</i>	57
	<i>Squatina tergocellata</i>	57
<b>Order Pristiophoriformes</b>		
Pristiophoridae	<i>Pliotrema warreni</i>	47, 51
	<i>Pristiophorus delicatus</i>	71
	<i>Pristiophorus schroederi</i>	31
<b>Order Heterodontiformes</b>		
Heterodontidae	<i>Heterodontus ramalheira</i>	51
<b>Order Orectolobiformes</b>		
Parascylliidae	<i>Cirrhoscyllium japonicum</i>	61
	<i>Parascyllium sparsimaculatum</i>	57
<b>Order Lamniformes</b>		
Odontaspidae	<i>Odontaspis ferox</i>	21, 27, 31, 34, 37, 51, 57, 61, 77, 81
	<i>Odontaspis noronhai</i>	31, 34, 41, 71, 77
Pseudocarchariidae	<i>Pseudocarcharias kamoharai</i>	31, 34, 41, 47, 51, 57, 61, 71, 77, 81, 87
Mitsukurinidae	<i>Mitsukurina owstoni</i>	27, 31, 34, 47, 51, 57, 61, 77, 81
Alopiidae	<i>Alopias superciliosus</i>	21, 27, 31, 34, 37, 41, 47, 51, 57, 61, 71, 77, 81, 87
Cetorhinidae	<i>Cetorhinus maximus</i>	21, 27, 31, 34, 37, 41, 47, 57, 61, 67, 77, 81, 87

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Order Carcharhiniformes</b>		
Scyliorhinidae	<i>Apristurus albisoma</i>	71
	<i>Apristurus ampliceps</i>	57, 81
	<i>Apristurus aphyodes</i>	27
	<i>Apristurus australis</i>	57, 71, 81
	<i>Apristurus brunneus</i>	67, 77, 87?
	<i>Apristurus bucephalus</i>	57
	<i>Apristurus canutus</i>	31
<b>Order Carcharhiniformes</b>		
Scyliorhinidae	<i>Apristurus exsanguis</i>	81
	<i>Apristurus fedorovi</i>	61
Scyliorhinidae	<i>Apristurus gibbosus</i>	61
	<i>Apristurus herklotsi</i>	61, 71
	<i>Apristurus indicus</i>	51
	<i>Apristurus internatus</i>	61
	<i>Apristurus investigatoris</i>	57
	<i>Apristurus japonicus</i>	61
	<i>Apristurus kampae</i>	77, 87?
	<i>Apristurus laurussoni</i>	21, 27, 31, 34
	<i>Apristurus longicephalus</i>	51, 57, 61, 71
	<i>Apristurus macrorhynchus</i>	61
	<i>Apristurus macrostomus</i>	61
	<i>Apristurus manis</i>	21, 27, 47
	<i>Apristurus melanoasper</i>	21, 27, 57, 71, 81
	<i>Apristurus microps</i>	21, 27, 47
	<i>Apristurus micropterygeus</i>	61
	<i>Apristurus nasutus</i>	77, 87
	<i>Apristurus parvipinnis</i>	31
	<i>Apristurus pinguis</i>	57, 61, 81
	<i>Apristurus platyrhynchus</i>	57, 61, 71, 81
	<i>Apristurus profundorum</i>	21, 34?
	<i>Apristurus riveri</i>	31
	<i>Apristurus saldanha</i>	47
	<i>Apristurus sibogae</i>	71
	<i>Apristurus sinensis</i>	57, 61, 71, 81
	<i>Apristurus spongiceps</i>	71, 77
	<i>Apristurus stensenii</i>	77
	<i>Asymbolus galacticus</i>	71
	<i>Asymbolus pallidus</i>	71
	<i>Bythaelurus? alcocki</i>	51
	<i>Bythaelurus canescens</i>	87
	<i>Bythaelurus clevai</i>	51
	<i>Bythaelurus dawsoni</i>	81
	<i>Bythaelurus hispidus</i>	57
	<i>Bythaelurus immaculatus</i>	61
	<i>Bythaelurus incanus</i>	57
	<i>Bythaelurus lutarius</i>	51
	<i>Cephaloscyllium albipinnum</i>	57, 81
	<i>Cephaloscyllium cooki</i>	57
	<i>Cephaloscyllium fasciatum</i>	61, 71
	<i>Cephaloscyllium hiscosellum</i>	57
	<i>Cephaloscyllium isabellum</i>	81
Scyliorhinidae	<i>Cephaloscyllium signourum</i>	71
	<i>Cephaloscyllium silasi</i>	51

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Cephaloscyllium speculum</i>	57
	<i>Cephaloscyllium sufflans</i>	51
	<i>Cephaloscyllium variegatum</i>	71, 81
	<i>Cephaloscyllium zebrum</i>	71
	<i>Cephalurus cephalus</i>	77
	<i>Figaro boardmani</i>	57, 71, 81
	<i>Figaro striatus</i>	71
	<i>Galeus antillensis</i>	31
	<i>Galeus atrae</i>	21, 31
<b>Order Carcharhiniformes</b>		
Scyliorhinidae	<i>Galeus atlanticus</i>	27, 37
	<i>Galeus cadenati</i>	31
	<i>Galeus eastmani</i>	61
	<i>Galeus gracilis</i>	57, 71
	<i>Galeus longirostris</i>	61
	<i>Galeus melastomus</i>	27, 34, 37
	<i>Galeus mincaronei</i>	41
	<i>Galeus murinus</i>	27
	<i>Galeus nipponensis</i>	61
	<i>Galeus piperatus</i>	77
	<i>Galeus polli</i>	34, 47
	<i>Galeus priapus</i>	71
	<i>Galeus schultzi</i>	71
	<i>Galeus springeri</i>	31
	<i>Holohalaelurus favus</i>	51
	<i>Holohalaelurus gennianus</i>	51
	<i>Holohalaelurus melanostigma</i>	51
	<i>Holohalaelurus punctatus</i>	47, 51
	<i>Holohalaelurus regani</i>	47, 51
	<i>Parmaturus albimarginatus</i>	71
	<i>Parmaturus albipenis</i>	71
	<i>Parmaturus bigus</i>	71
	<i>Parmaturus campechiensis</i>	31
	<i>Parmaturus lanatus</i>	57
	<i>Parmaturus macmillani</i>	51, 81
	<i>Parmaturus melanobranchius</i>	61, 71
	<i>Parmaturus pilosus</i>	61
	<i>Parmaturus xaniurus</i>	77
	<i>Pentanchus profundicolus</i>	71
	<i>Schroederichthys maculatus</i>	31
	<i>Schroederichthys saurisqualus</i>	41
	<i>Schroederichthys tenuis</i>	31, 41
	<i>Scyliorhinus boa</i>	31
	<i>Scyliorhinus capensis</i>	47, 51
	<i>Scyliorhinus cervigoni</i>	34, 47
	<i>Scyliorhinus comoroensis</i>	51
	<i>Scyliorhinus haekelii</i>	31, 41
	<i>Scyliorhinus hesperius</i>	31
	<i>Scyliorhinus meadi</i>	31
	<i>Scyliorhinus retifer</i>	21, 31
	<i>Scyliorhinus torrei</i>	31
Proscylliidae	<i>Eridacnis barbouri</i>	31
	<i>Eridacnis radcliffei</i>	51, 57, 61, 71
	<i>Eridacnis sinuans</i>	51

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
Pseudotriakidae	<i>Gollum attenuatus</i>	81
	<i>Pseudotriakis microdon</i>	21, 27, 34, 51, 57, 61, 71, 77, 81
Triakidae	<i>Hemitriakis abdita</i>	71
	<i>Iago garricki</i>	57, 71
	<i>Iago omanensis</i>	51, 57?
	<i>Mustelus canis insularis</i>	31
	<i>Mustelus stevensi</i>	57
	<i>Mustelus walkeri</i>	71
Carcharhinidae	<i>Carcharhinus altimus</i>	31, 34, 37, 41, 51, 57, 61, 71, 77, 81, 87

### Batoids – Order Rajiformes

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
Rhinobatidae	<i>Rhinobatos variegatus</i>	57
Narcinidae	<i>Benthobatis krefftii</i>	41
	<i>Benthobatis marcida</i>	31
	<i>Benthobatis moresbyi</i>	51
	<i>Benthobatis yangi</i>	61, 71
	<i>Narcine lasti</i>	57, 71
	<i>Narcine nelsoni</i>	71
	<i>Narcine tasmaniensis</i>	57, 81
	<i>Heteronarce garmani</i>	51
	<i>Heteronarce mollis</i>	51
	<i>Typhlonarke aysoni</i>	81
	<i>Typhlonarke tarakea</i>	81
Torpedinidae	<i>Torpedo fairchildi</i>	81
	<i>Torpedo fuscomaculata</i>	51
	<i>Torpedo macneilli</i>	57, 71, 81
	<i>Torpedo microdiscus</i>	87
	<i>Torpedo nobiliana</i>	21, 27, 31, 37, 41, 47
	<i>Torpedo puelcha</i>	41
	<i>Torpedo tokionis</i>	57, 61, 71, 81
	<i>Torpedo tremens</i>	77, 87
Arhynchobatidae	<i>Arhynchobatis asperrimus</i>	81
	<i>Bathyraja abyssicola</i>	61, 67, 77
	<i>Bathyraja aguja</i>	87
	<i>Bathyraja albomaculata</i>	41, 87
	<i>Bathyraja aleutica</i>	61, 67, 77
	<i>Bathyraja andriashewi</i>	61
	<i>Bathyraja bergi</i>	61
	<i>Bathyraja brachyurops</i>	41, 87
	<i>Bathyraja caeluronigricans</i>	61
	<i>Bathyraja couesseauae</i>	41
	<i>Bathyraja diplotaenia</i>	61
	<i>Bathyraja eatonii</i>	48, 58, 88
	<i>Bathyraja fedorovi</i>	61
	<i>Bathyraja griseoauda</i>	41, 87
	<i>Bathyraja hesperafricana</i>	34
	<i>Bathyraja interrupta</i>	61, 67, 77
	<i>Bathyraja irrassa</i>	58
	<i>Bathyraja isotrachys</i>	61
	<i>Bathyraja kincaidii</i>	67, 77
	<i>Bathyraja lindbergi</i>	61
	<i>Bathyraja longicauda</i>	87
	<i>Bathyraja maccaini</i>	48, 87
	<i>Bathyraja macloviana</i>	41, 87
	<i>Bathyraja maculata</i>	61, 67
	<i>Bathyraja magellanica</i>	41, 87
	<i>Bathyraja matsubarai</i>	61, 67?
	<i>Bathyraja meridionalis</i>	41, 48
	<i>Bathyraja minispinosa</i>	61, 67
	<i>Bathyraja multispinis</i>	41, 87
	<i>Bathyraja murrayi</i>	48?, 58, 88?
	<i>Bathyraja notoroensis</i>	61
Arhynchobatidae	<i>Bathyraja papilionifera</i>	41

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Bathyraja parmifera</i>	61, 67
	<i>Bathyraja peruviana</i>	87
	<i>Bathyraja richardsoni</i>	21?, 27?, 57, 81
	<i>Bathyraja scaphiops</i>	41
	<i>Bathyraja schroederi</i>	41, 87
	<i>Bathyraja shuntovi</i>	81
	<i>Bathyraja smirnovi</i>	61
	<i>Bathyraja smithii</i>	47
	<i>Bathyraja spinicauda</i>	21, 27
	<i>Bathyraja spinosissima</i>	61?, 77, 87
	<i>Bathyraja trachourus</i>	61
	<i>Bathyraja trachura</i>	61, 67, 77
	<i>Bathyraja tzinovskii</i>	61
	<i>Bathyraja violacea</i>	61, 67
	<i>Brochiraja aenigma</i>	81
	<i>Brochiraja albilabiata</i>	81
	<i>Brochiraja asperula</i>	81
	<i>Brochiraja leviveneta</i>	81
	<i>Brochiraja microspinifera</i>	81
	<i>Brochiraja spinifera</i>	81
	<i>Insetiraja laxipella</i>	71
	<i>Insetiraja subtilispinosa</i>	57, 71
	<i>Notoraja azurea</i>	57, 81
	<i>Notoraja hirticauda</i>	57
	<i>Notoraja lira</i>	57
	<i>Notoraja ochroderma</i>	71
	<i>Notoraja sapphira</i>	71, 81
	<i>Notoraja sticta</i>	57
	<i>Notoraja tobitukai</i>	61
	<i>Pavoraja alleni</i>	57
	<i>Pavoraja arenaria</i>	57
	<i>Pavoraja mosaica</i>	71
	<i>Pavoraja nitida</i>	57, 81
	<i>Pavoraja pseudonitida</i>	71
	<i>Pavoraja umbrosa</i>	71, 81
	<i>Psammobatis scobina</i>	87
	<i>Pseudoraja fischeri</i>	31
	<i>Rhinoraja kujiensis</i>	61
	<i>Rhinoraja longi</i>	67
	<i>Rhinoraja longicauda</i>	61
	<i>Rhinoraja obtusa</i>	61
	<i>Rhinoraja odai</i>	61
	<i>Rhinoraja taranetzi</i>	61, 67
Rajidae	<i>Amblyraja badia</i>	61, 67, 77
	<i>Amblyraja doellojuradoi</i>	41, 87
	<i>Amblyraja frerichsii</i>	41, 87
	<i>Amblyraja georgiana</i>	41, 48, 58, 87, 88
	<i>Amblyraja hyperborea</i>	21, 27, 47, 57, 61, 67, 77, 81
	<i>Amblyraja jensenii</i>	21, 27
	<i>Amblyraja radiata</i>	21, 27, 47
	<i>Amblyraja reversa</i>	51
	<i>Amblyraja robertsi</i>	47
Rajidae	<i>Amblyraja taaf</i>	58
	<i>Breviraja claramaculata</i>	31

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Breviraja colesi</i>	31
	<i>Breviraja marklei</i>	21
	<i>Breviraja mouldi</i>	31
	<i>Breviraja nigriventralis</i>	31
	<i>Breviraja spinosa</i>	31
	<i>Dactylobatus armatus</i>	31
	<i>Dactylobatus clarki</i>	31
	<i>Dipturus acrobelus</i>	57, 81
	<i>Dipturus apricus</i>	71
	<i>Dipturus batis</i>	27, 34, 37
	<i>Dipturus bullisi</i>	31
	<i>Dipturus campbelli</i>	51
	<i>Dipturus canutus</i>	57, 81
	<i>Dipturus cerva</i>	57, 81
	<i>Dipturus crosnieri</i>	51
	<i>Dipturus doutrei</i>	34, 47
	<i>Dipturus endeavouri</i>	71, 81
	<i>Dipturus garricki</i>	31
	<i>Dipturus gigas</i>	61, 71
	<i>Dipturus grahami</i>	71, 81
	<i>Dipturus gadgei</i>	57, 81
	<i>Dipturus healdi</i>	57
	<i>Dipturus innominatus</i>	81
	<i>Dipturus johannisdavesi</i>	51
	<i>Dipturus laevis</i>	21
	<i>Dipturus lanceorostratus</i>	51
	<i>Dipturus leptocaudus</i>	41
	<i>Dipturus linteus</i>	21, 27
	<i>Dipturus macrocaudus</i>	61
	<i>Dipturus melanospilus</i>	71, 81
	<i>Dipturus mennii</i>	41
	<i>Dipturus nidarosiensis</i>	27
	<i>Dipturus oculus</i>	57
	<i>Dipturus oregoni</i>	31
	<i>Dipturus oxyrhynchus</i>	27, 34, 37
	<i>Dipturus polyommata</i>	71
	<i>Dipturus pullopunctata</i>	47
	<i>Dipturus queenslandicus</i>	71
	<i>Dipturus springeri</i>	47, 51
	<i>Dipturus stenorhynchus</i>	47, 51
	<i>Dipturus teevani</i>	31
	<i>Dipturus tenui</i>	61, 71
	<i>Dipturus trachidermus</i>	41, 87
	<i>Dipturus wengi</i>	57, 71, 81
	<i>Fenestrata atripinna</i>	31
	<i>Fenestrata cubensis</i>	31
	<i>Fenestrata ishiyamai</i>	31
	<i>Fenestrata maceachrani</i>	51
	<i>Fenestrata mamillidens</i>	57
	<i>Fenestrata plutonia</i>	31
	<i>Fenestrata sibogae</i>	71
Rajidae	<i>Fenestrata sinusmexicanus</i>	31
	<i>Gurgesiella atlantica</i>	31, 41
	<i>Gurgesiella dorsalifera</i>	31, 41

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Gurgesiella furvescens</i>	87
	<i>Leucoraja compagnoi</i>	47, 51
	<i>Leucoraja fullonica</i>	27, 34, 37
	<i>Leucoraja garmani</i>	21, 31
	<i>Leucoraja lentiginosa</i>	31
	<i>Leucoraja leucosticta</i>	34, 47
	<i>Leucoraja melitensis</i>	37
	<i>Leucoraja naevus</i>	27, 34, 37
	<i>Leucoraja pristispina</i>	57
	<i>Leucoraja wallacei</i>	47, 51
	<i>Leucoraja yucatanensis</i>	31
	<i>Malacoraja kreffti</i>	27
	<i>Malacoraja obscura</i>	41
	<i>Malacoraja senta</i>	21, 31
	<i>Malacoraja spinacidermis</i>	27, 34, 47
	<i>Neoraja africana</i>	34
	<i>Neoraja caerulea</i>	27
	<i>Neoraja carolinensis</i>	31
	<i>Neoraja iberica</i>	27
	<i>Neoraja stehmanni</i>	47
	<i>Okamejei arafurensis</i>	57, 71
	<i>Okamejei heemstrai</i>	51
	<i>Okamejei leptoura</i>	57
	<i>Raja africana?</i>	34, 37
	<i>Raja bahamensis</i>	31
	<i>Raja binoculata</i>	61, 67, 77
	<i>Raja inornata</i>	77
	<i>Raja maderensis</i>	34
	<i>Raja polystigma</i>	37
	<i>Raja rhina</i>	67, 77
	<i>Raja stellulata</i>	77
	<i>Raja straeleni</i>	34, 47
	<i>Rajella annandalei</i>	71
	<i>Rajella barnardi</i>	34, 47
	<i>Rajella bathyphila</i>	21, 27, 34
	<i>Rajella bigelowi</i>	21, 27, 31, 34
	<i>Rajella caudaspinosa</i>	47, 51
	<i>Rajella challengerii</i>	57, 81
	<i>Rajella dissimilis</i>	27, 34, 47
	<i>Rajella eisenhardtii</i>	87
	<i>Rajella fuliginea</i>	31
	<i>Rajella fyllae</i>	21, 27
	<i>Rajella kukujevi</i>	27
	<i>Rajella leopardus</i>	34, 47, 51?
	<i>Rajella nigerrima</i>	87
	<i>Rajella purpuriventralis</i>	31
	<i>Rajella ravidula</i>	34, 47
	<i>Rajella sadowskyii</i>	41, 87
	<i>Rostroraja alba</i>	27, 34, 37, 47, 51
	<i>Zearaja chilensis</i>	41, 87
Rajidae	<i>Zearaja nasutus</i>	81
Anacanthobatidae	<i>Anacanthobatis americanus</i>	31, 41
	<i>Anacanthobatis donghaiensis</i>	61
	<i>Anacanthobatis folirostris</i>	31

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Anacanthobatis longirostris</i>	31
	<i>Anacanthobatis marmoratus</i>	51
	<i>Anacanthobatis nanhaiensis</i>	
	<i>Anacanthobatis ori</i>	51
	<i>Anacanthobatis stenosoma</i>	
	<i>Cruriraja andamanica</i>	51, 57
	<i>Cruriraja atlantis</i>	31
	<i>Cruriraja cadenati</i>	31
	<i>Cruriraja durbanensis</i>	47
	<i>Cruriraja parcomaculata</i>	47, 51
Anacanthobatidae	<i>Cruriraja poeyi</i>	31
	<i>Cruriraja rugosa</i>	31
	<i>Cruriraja triangularis</i>	51
	<i>Sinobatis borneensis</i>	61, 71
	<i>Sinobatis bulbicauda</i>	57, 71
	<i>Sinobatis caerulea</i>	57
	<i>Sinobatis filicauda</i>	71
	<i>Sinobatis melanosoma</i>	61, 71
Plesiobatidae	<i>Plesiobatis daviesi</i>	51, 57, 61, 71, 77, 81
Urolophidae	<i>Urolophus expansus</i>	57
	<i>Urolophus piperatus</i>	71
Hexatrygonidae	<i>Hexatrygon bickelli</i>	47, 51, 57, 61, 71, 77
Dasyatidae	<i>Dasyatis brevicaudata</i>	47, 51, 57, 71, 81

### Chimaeras – Order Chimaeriformes

<b>Family</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
Rhinochimaeridae	<i>Harriotta haekeli</i>	21?, 27, 34, 57, 81
	<i>Harriotta raleighana</i>	21, 27, 34, 41, 47, 51, 57, 61, 71, 77, 81
	<i>Neoharriotta carri</i>	31
	<i>Neoharriotta pinnata</i>	34, 47
	<i>Neoharriotta pumila</i>	51
	<i>Rhinochimaera africana</i>	47, 51, 57, 61
	<i>Rhinochimaera atlantica</i>	21, 27, 31, 34, 41, 47
	<i>Rhinochimaera pacifica</i>	57, 61, 71, 81, 87
Chimaeridae	<i>Chimaera argiloba</i>	57
	<i>Chimaera cubana</i>	31
	<i>Chimaera fulva</i>	57, 81
	<i>Chimaera jordani</i>	51, 61
	<i>Chimaera lignaria</i>	57, 81
	<i>Chimaera macrospina</i>	57, 71, 81
	<i>Chimaera monstrosa</i>	27, 34, 37
	<i>Chimaera obscura</i>	71, 81
	<i>Chimaera owstoni</i>	61
	<i>Chimaera panthera</i>	81
	<i>Chimaera phantasma</i>	61
	<i>Hydrolagus affinis</i>	21, 27, 34
	<i>Hydrolagus africanus</i>	47?, 51
	<i>Hydrolagus alberti</i>	31
	<i>Hydrolagus alphus</i>	87
	<i>Hydrolagus barbouri</i>	61
	<i>Hydrolagus bemisi</i>	81
	<i>Hydrolagus colliei</i>	67, 77
	<i>Hydrolagus homonycteris</i>	57, 81
	<i>Hydrolagus lemures</i>	57, 71, 81
	<i>Hydrolagus lusitanicus</i>	27
	<i>Hydrolagus macrophthalmus</i>	87
	<i>Hydrolagus marmoratus</i>	71, 81
	<i>Hydrolagus matallanasi</i>	41
	<i>Hydrolagus mccoskeri</i>	87
	<i>Hydrolagus melanophasma</i>	77
	<i>Hydrolagus mirabilis</i>	27, 31, 34, 47
	<i>Hydrolagus mitsukurii</i>	61, 71
	<i>Hydrolagus novaezealandiae</i>	81
	<i>Hydrolagus ogilbyi</i>	57, 81
	<i>Hydrolagus pallidus</i>	27
	<i>Hydrolagus purpurescens</i>	61, 77
	<i>Hydrolagus trolli</i>	47, 48, 58?, 71, 81, 88?

### APPENDIX 3

**Deep-sea chondrichthyan species known to be targeted by fisheries and deep-sea chondrichthyan species known to be taken as bycatch in fisheries**

**Deep-sea chondrichthyan species known to be targeted by fisheries**

Order	Family	Species
<b>Sharks</b>		
Squaliformes	Squalidae	<i>Squalus cubensis</i>
		<i>Squalus mitsukurii</i>
	Centrophoridae	<i>Centrophorus atromarginatus</i>
		<i>Centrophorus granulosus</i>
		<i>Centrophorus harrissoni</i>
		<i>Centrophorus isodon</i>
		<i>Centrophorus moluccensis</i>
		<i>Centrophorus squamosus</i>
		<i>Centrophorus zeehaani</i>
		<i>Deania calcea</i>
	Etomopteridae	<i>Etomopterus spinax</i>
		<i>Etomopterus virens</i>
	Somniosidae	<i>Centroscymnus coelolepis</i>
		<i>Centroselachus crepidater</i>
	Dalatiidae	<i>Dalatias licha</i>
Lamniformes	Cetorhinidae	<i>Cetorhinus maximus</i>
Carcharhiniformes	Scyliorhinidae	<i>Galeus melastomus</i>
<b>Batoids</b>		
Rajiformes	Arhynchobatidae	<i>Bathyraja albomaculata</i>
		<i>Bathyraja brachyurops</i>
		<i>Bathyraja courseaue</i>
		<i>Bathyraja griseocauda</i>
		<i>Bathyraja macloviana</i>
		<i>Bathyraja magellanica</i>
		<i>Bathyraja meridionalis</i>
		<i>Bathyraja multispinis</i>
		<i>Bathyraja scaphiops</i>
		<i>Psammobatis scobina</i>
	Rajidae	<i>Amblyraja doellojuradoi</i>
		<i>Amblyraja radiata</i>
		<i>Dipturus trachydermus</i>
		<i>Raja binoculata</i>
		<i>Raja inornata</i>
		<i>Raja rhina</i>
		<i>Zearaja chilensis</i>
<b>Chimaeras</b>		
Chimaeriformes	Chimaeridae	<i>Chimaera monstrosa</i>
		<i>Hydrolagus bemisi</i>
		<i>Hydrolagus novaezealandiae</i>

**Deep-sea chondrichthyan species known to be taken as bycatch in fisheries**

Order	Family	Species
<b>Sharks</b>		
Hexanchiformes	Chlamydoselachidae	<i>Chlamydoselachus anguineus</i>
	Hexanchidae	<i>Heptranchias perlo</i>
		<i>Hexanchus griseus</i>
		<i>Hexanchus nakamurai</i>
Squaliformes	Echinorhinidae	<i>Echinorhinus brucus</i>
	Squalidae	<i>Cirrhigaleus asper</i>
		<i>Cirrhigaleus barbifer</i>
		<i>Squalus blainville</i>
		<i>Squalus chloroculus</i>
		<i>Squalus cubensis</i>
		<i>Squalus edmundsi</i>
		<i>Squalus grahami</i>
		<i>Squalus griffini</i>
		<i>Squalus hemipinnis</i>
		<i>Squalus japonicus</i>
		<i>Squalus lalannei</i>
		<i>Squalus megalops</i>
		<i>Squalus mitsukurii</i>
		<i>Squalus montalbani</i>
		<i>Squalus raoulensis</i>
	Centrophoridae	<i>Centrophorus acus</i>
		<i>Centrophorus atromarginatus</i>
		<i>Centrophorus granulosus</i>
		<i>Centrophorus harrissoni</i>
		<i>Centrophorus isodon</i>
		<i>Centrophorus lusitanicus</i>
		<i>Centrophorus moluccensis</i>
		<i>Centrophorus niaukang</i>
		<i>Centrophorus seychellorum</i>
		<i>Centrophorus squamosus</i>
		<i>Centrophorus tessellatus</i>
		<i>Centrophorus zeehaani</i>
		<i>Deania calcea</i>
		<i>Deania hystricosum</i>
		<i>Deania profundorum</i>
		<i>Deania quadrispinosum</i>
	Etmopteridae	<i>Centroscyllium excelsum</i>
		<i>Centroscyllium fabricii</i>
		<i>Centroscyllium granulatum</i>
		<i>Centroscyllium kamoharai</i>
		<i>Centroscyllium nigrum</i>
		<i>Centroscyllium ornatum</i>
		<i>Centroscyllium ritteri</i>
		<i>Etmopterus baxteri</i>
		<i>Etmopterus bigelowi</i>
		<i>Etmopterus brachyurus</i>
		<i>Etmopterus bullisi</i>

Order	Family	Species
		<i>Etmopterus carteri</i>
		<i>Etmopterus caudistigmus</i>
		<i>Etmopterus decacuspidatus</i>
		<i>Etmopterus dianthus</i>
	Etmopteridae	<i>Etmopterus dislineatus</i>
		<i>Etmopterus evansi</i>
		<i>Etmopterus gracilispinis</i>
		<i>Etmopterus granulosus</i>
		<i>Etmopterus hillianus</i>
		<i>Etmopterus litvinovi</i>
		<i>Etmopterus lucifer</i>
		<i>Etmopterus molleri</i>
		<i>Etmopterus perryi</i>
		<i>Etmopterus princeps</i>
		<i>Etmopterus pusillus</i>
		<i>Etmopterus robinsi</i>
		<i>Etmopterus schultzi</i>
		<i>Etmopterus sentosus</i>
		<i>Etmopterus spinax</i>
		<i>Etmopterus splendidus</i>
		<i>Etmopterus unicolor</i>
		<i>Etmopterus villosus</i>
		<i>Etmopterus virens</i>
		<i>Miroscyllium sheikoi</i>
		<i>Trigonognathus kabeyai</i>
	Somniosidae	<i>Centroscymnus coelolepis</i>
		<i>Centroscymnus owstoni</i>
		<i>Centroselachus crepidater</i>
		<i>Proscymnodon macracanthus</i>
		<i>Proscymnodon plunketi</i>
		<i>Scymnodalatias albicauda</i>
		<i>Scymnodon ringens</i>
		<i>Somniosus antarcticus</i>
		<i>Somniosus longus</i>
		<i>Somniosus microcephalus</i>
		<i>Somniosus pacificus</i>
		<i>Somniosus rostratus</i>
		<i>Zameus ichiharai</i>
		<i>Zameus squamulosus</i>
	Oxynotidae	<i>Oxynotus bruniensis</i>
		<i>Oxynotus caribbaeus</i>
		<i>Oxynotus centrina</i>
	Dalatiidae	<i>Dalatias licha</i>
		<i>Isistius brasiliensis</i>
		<i>Isistius labialis</i>
		<i>Squaliolus aliae</i>
		<i>Squaliolus laticaudus</i>
		<i>Oxynotus japonicus</i>
		<i>Oxynotus paradoxus</i>
	Squatiniidae	<i>Squatina aculeata</i>
		<i>Squatina africana</i>

<b>Order</b>	<b>Family</b>	<b>Species</b>
		<i>Squatina albipunctata</i>
		<i>Squatina tergocellata</i>
Pristiophoriformes	Pristiophoridae	<i>Pristiophorus schroederi</i>
Lamniformes	Odontaspidae	<i>Odontaspis ferox</i>
	Pseudocarchariidae	<i>Pseudocarcharias kamoharai</i>
	Mitsukurinidae	<i>Mitsukurina owstoni</i>
	Alopiidae	<i>Alopias superciliosus</i>
Carcharhiniformes	Scyliorhinidae	<i>Apristurus canutus</i>
		<i>Apristurus laurussoni</i>
		<i>Apristurus manis</i>
		<i>Apristurus microps</i>
		<i>Apristurus parvipinnis</i>
		<i>Apristurus platyrhynchus</i>
		<i>Apristurus profundorum</i>
		<i>Apristurus riveri</i>
		<i>Cephaloscyllium albibinnum</i>
		<i>Cephaloscyllium cooki</i>
		<i>Cephaloscyllium fasciatum</i>
		<i>Cephaloscyllium hiscosellum</i>
		<i>Galeus antillensis</i>
		<i>Galeus arae</i>
		<i>Galeus atlanticus</i>
		<i>Galeus eastmani</i>
		<i>Galeus gracilis</i>
		<i>Galeus longirostris</i>
		<i>Galeus melastomus</i>
		<i>Galeus mincaronei</i>
		<i>Galeus murinus</i>
		<i>Galeus nippensis</i>
		<i>Galeus springeri</i>
		<i>Parmaturus melanobranchius</i>
		<i>Scyliorhinus boa</i>
		<i>Scyliorhinus comoroensis</i>
		<i>Scyliorhinus meadi</i>
		<i>Scyliorhinus retifer</i>
		<i>Scyliorhinus torrei</i>
	Proscylliidae	<i>Eridacnis barbouri</i>
		<i>Eridacnis radcliffei</i>
		<i>Eridacnis sinuans</i>
	Pseudotriakidae	<i>Gollum attenuatus</i>
		<i>Pseudotriakis microdon</i>
	Triakidae	<i>Iago garricki</i>
		<i>Iago omanensis</i>
		<i>Mustelus canis insularis</i>
		<i>Mustelus stevensi</i>
		<i>Mustelus walkeri</i>
	Carcharhinidae	<i>Carcharhinus altimus</i>
<b>Batoids</b>		
Rajiformes	Rhinobatidae	<i>Rhinobatos variegatus</i>
	Narcinidae	<i>Benthobatis krefftii</i>
		<i>Benthobatis marcida</i>

<b>Order</b>	<b>Family</b>	<b>Species</b>
		<i>Benthobatis yangi</i>
		<i>Narcine tasmaniensis</i>
		<i>Heteronarce garmani</i>
		<i>Heteronarce mollis</i>
	Narkidae	<i>Typhlonarke aysoni</i>
		<i>Typhlonarke tarakea</i>
	Torpedinidae	<i>Torpedo fairchildi</i>
		<i>Torpedo fuscomaculata</i>
		<i>Torpedo macneilli</i>
		<i>Torpedo nobiliana</i>
		<i>Torpedo puelcha</i>
		<i>Torpedo tokionis</i>
		<i>Torpedo tremens</i>
	Arhynchobatidae	<i>Arhynchobatis asperrimus</i>
		<i>Bathyraja albomaculata</i>
		<i>Bathyraja aleutica</i>
		<i>Bathyraja bergi</i>
		<i>Bathyraja brachyurops</i>
		<i>Bathyraja caeluronigricans</i>
		<i>Bathyraja couesseauae</i>
		<i>Bathyraja diplotaenia</i>
		<i>Bathyraja eatonii</i>
		<i>Bathyraja fedorovi</i>
		<i>Bathyraja griseoauda</i>
		<i>Bathyraja interrupta</i>
		<i>Bathyraja irrassa</i>
		<i>Bathyraja isotrachys</i>
		<i>Bathyraja kincaidii</i>
		<i>Bathyraja lindbergi</i>
		<i>Bathyraja maccaini</i>
		<i>Bathyraja macloviana</i>
		<i>Bathyraja maculata</i>
		<i>Bathyraja magellanica</i>
		<i>Bathyraja matsubarai</i>
		<i>Bathyraja meridionalis</i>
		<i>Bathyraja minispinosa</i>
		<i>Bathyraja murrayi</i>
		<i>Bathyraja multispinis</i>
		<i>Bathyraja notoroensis</i>
		<i>Bathyraja papilionifera</i>
		<i>Bathyraja parmisera</i>
		<i>Bathyraja peruana</i>
		<i>Bathyraja scaphiops</i>
		<i>Bathyraja shuntovi</i>
		<i>Bathyraja smirnovi</i>
		<i>Bathyraja smithii</i>
		<i>Bathyraja spinicauda</i>
		<i>Bathyraja trachouros</i>
		<i>Bathyraja trachura</i>
		<i>Bathyraja violacea</i>
		<i>Brochiraja albilibiata</i>

<b>Order</b>	<b>Family</b>	<b>Species</b>
		<i>Brochiraja asperula</i>
		<i>Brochiraja leviveneta</i>
		<i>Brochiraja microspinifera</i>
		<i>Brochiraja spinifera</i>
		<i>Notoraja hirticauda</i>
		<i>Pavoraja tobitukai</i>
		<i>Pavoraja alleni</i>
		<i>Pavoraja arenaria</i>
		<i>Pavoraja nitida</i>
		<i>Psammobatis scobina</i>
		<i>Psammobatis kuijensis</i>
		<i>Rhinoraja longi</i>
		<i>Rhinoraja longicauda</i>
		<i>Rhinoraja obtusa</i>
		<i>Rhinoraja odai</i>
		<i>Rhinoraja taranetzi</i>
	Rajidae	<i>Amblyraja doellojuradoi</i>
		<i>Amblyraja frerichsii</i>
		<i>Amblyraja georgiana</i>
		<i>Amblyraja hyperborea</i>
		<i>Amblyraja radiata</i>
		<i>Amblyraja taaf</i>
		<i>Breviraja marklei</i>
		<i>Breviraja nigriventralis</i>
		<i>Dipturus batis</i>
		<i>Dipturus campbelli</i>
		<i>Dipturus cerva</i>
		<i>Dipturus crosnieri</i>
		<i>Dipturus doutrei</i>
		<i>Dipturus gigas</i>
		<i>Dipturus gudgeri</i>
		<i>Dipturus innominatus</i>
		<i>Dipturus laevis</i>
		<i>Dipturus leptocaudus</i>
		<i>Dipturus linteus</i>
		<i>Dipturus macrocaudus</i>
		<i>Dipturus mennii</i>
		<i>Dipturus nidarosiensis</i>
		<i>Dipturus oxyrhynchus</i>
		<i>Dipturus pullopunctata</i>
		<i>Dipturus springeri</i>
		<i>Dipturus stenorhynchus</i>
		<i>Dipturus teevani</i>
		<i>Dipturus tengu</i>
		<i>Dipturus trachydermus</i>
		<i>Dipturus wengi</i>
		<i>Fenestriraja maceachrani</i>
		<i>Gurgesiella atlantica</i>
		<i>Gurgesiella dorsalifera</i>
		<i>Leucoraja fullonica</i>
		<i>Leucoraja garmani</i>

<b>Order</b>	<b>Family</b>	<b>Species</b>
		<i>Leucoraja lentiginosa</i>
		<i>Leucoraja leucosticta</i>
		<i>Leucoraja melitensis</i>
		<i>Leucoraja naevus</i>
		<i>Leucoraja wallacei</i>
		<i>Malacoraja obscura</i>
		<i>Malacoraja senta</i>
		<i>Malacoraja spinacidermis</i>
		<i>Neoraja caerulea</i>
		<i>Neoraja iberica</i>
		<i>Okamejei arafurensis</i>
		<i>Okamejei leptoura</i>
		<i>Raja africana</i>
		<i>Raja binoculata</i>
		<i>Raja maderensis</i>
		<i>Raja polystigma</i>
		<i>Raja stellulata</i>
		<i>Raja straeleni</i>
		<i>Rajella bathyphila</i>
		<i>Rajella caudaspinosa</i>
		<i>Rajella challengerii</i>
		<i>Rajella dissimilis</i>
		<i>Rajella fyllae</i>
		<i>Raja inornata</i>
		<i>Raja kukujevi</i>
		<i>Raja rhina</i>
		<i>Rajella sadowskyii</i>
		<i>Rostroraja alba</i>
		<i>Zearaja chilensis</i>
		<i>Zearaja nasutus</i>
	Anacanthobatidae	<i>Anacanthobatis americanus</i>
		<i>Anacanthobatis donghaiensis</i>
		<i>Cruriraja durbanensis</i>
		<i>Cruriraja parcomaculata</i>
		<i>Cruriraja rugosa</i>
		<i>Sinobatis borneensis</i>
		<i>Sinobatis bulbicauda</i>
		<i>Sinobatis melanosoma</i>
	Plesiobatidae	<i>Plesiobatis daviesi</i>
	Hexatrygonidae	<i>Hexatrygon bickelli</i>
	Dasyatidae	<i>Dasyatis brevicaudata</i>
<b>Chimaeras</b>		
Chimaeriformes	Rhinochimaeridae	<i>Harriotta raleighana</i>
		<i>Neoharriotta pinnata</i>
		<i>Rhinochimaera africana</i>
		<i>Rhinochimaera atlantica</i>
	Chimaeridae	<i>Chimaera cubana</i>
		<i>Chimaera fulva</i>
		<i>Chimaera lignaria</i>
		<i>Chimaera macrospina</i>
		<i>Chimaera monstrosa</i>

<b>Order</b>	<b>Family</b>	<b>Species</b>
		<i>Chimaera phantasma</i>
		<i>Hydrolagus affinis</i>
		<i>Hydrolagus africanus</i>
		<i>Hydrolagus bemisi</i>
		<i>Hydrolagus colliei</i>
		<i>Hydrolagus homonycteris</i>
		<i>Hydrolagus lemures</i>
		<i>Hydrolagus lusitanicus</i>
		<i>Hydrolagus matallanasi</i>
		<i>Hydrolagus mitsukuri</i>
		<i>Hydrolagus novaezealandiae</i>
		<i>Hydrolagus ogilbyi</i>
		<i>Hydrolagus purpureascens</i>
		<i>Hydrolagus trolli</i>

**Global list of cold-water corals (order Scleractinia; sub-order Filifera; sub-class Octocorallia, order Antipatharia) from waters deeper than 200 m, vulnerable species, and draft recommendations for the production of identification guides**

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## **Overview**

Following the FAO terms of reference, four lists of the global distribution (in relation to the 19 FAO Major Fishing Areas) of all cold-water species of scleractinians, Filifera and all potential habitat-forming cold-water octocorallians and antipatharians known to occur in waters deeper than 200 m were compiled and briefly described. A relevant bibliography for each species group was assembled. Finally, draft recommendations for the production of identification guides are suggested.

### **1. List of relevant corals group**

There are four orders belonging to two cnidarian classes that fit the term “coral”<sup>1</sup>: (i) class Anthozoa, subclass Hexacorallia – order Scleractinia order Zoanthidea, order Antipatharia; subclass Octocorallia and (ii) class Hydrozoa, subclass Hydroidolina – order Anthoathecata. A compilation was made for global distribution (in relation to the 19 FAO Major Fishing Areas) of all cold-water species of scleractinians (797 spp., including undescribed species), filiferans (246 spp.), and all potential habitat-forming cold-water octocorallians (225 spp.) and antipatharians (33 spp.) known to occur in waters deeper than 200 m. Table 1 summarizes the 2 classes down to the family level. See Appendixes 1, 2, 3 and 4 for details on species and their deep-sea depth occurrence. The diversity (number of species) of each selected deep-sea corals group per FAO Major Fishing Area is shown in Table 2.

**Table 1**  
**Global species list for FAO Major Fishing Areas**

<b>Class</b>	<b>Subclass</b>	<b>Order</b>	<b>Suborder</b>	<b>Family</b>	
Anthozoa	Hexacorallia	Scleractinia (see Appendix 1)		<ul style="list-style-type: none"> <li>• Anthemiphylliidae</li> <li>• Caryophylliidae</li> <li>• Dendrophylliidae</li> <li>• Flabellidae</li> <li>• Fungiacyathidae</li> <li>• Gardineriidae</li> <li>• Guynidae</li> </ul>	<ul style="list-style-type: none"> <li>• Micrabaciidae</li> <li>• Oculinidae</li> <li>• Pocilloporidae</li> <li>• Rhizangiidae</li> <li>• Stenocyathidae</li> <li>• Turbinoliidae</li> </ul>
		Antipatharia (see Appendix 2)		<ul style="list-style-type: none"> <li>• Antipathidae</li> <li>• Aphanipathidae</li> <li>• Cladopathidae</li> <li>• Stylopathidae</li> </ul>	<ul style="list-style-type: none"> <li>• Myriopathidae</li> <li>• Schizophathidae</li> <li>• Leiopathidae</li> </ul>
	Octocorallia	Gorgonacea (see Appendix 3)		<ul style="list-style-type: none"> <li>• Chrysogorgiidae</li> <li>• Coralliidae</li> <li>• Isididae</li> </ul>	<ul style="list-style-type: none"> <li>• Paragorgiidae</li> <li>• Plexauridae</li> <li>• Primnoidae</li> </ul>
Hydrozoa	Hydroidolina	Anthoatherata	Filifera (see Appendix 4)	<ul style="list-style-type: none"> <li>• Stylasteridae</li> </ul>	<ul style="list-style-type: none"> <li>• Hydractiniidae</li> </ul>

<sup>1</sup> Definition from Cairn (2007): “animals in the cnidarian classes Anthozoa and Hydrozoa that produce either calcium carbonate (aragonitic or calcitic) secretions resulting in a continuous skeleton or as numerous microscopic, individualized sclerites, or that have a black, horn-like, proteinaceous axis”.

**Table 2**Diversity (number of species) of each selected deep-sea corals group per FAO Major Fishing Area<sup>1</sup>

FAO Major Fishing Areas	Poles			Atlantic					M&B*	Indian			Pacific						
	18	48	88	21	27	31	34	41	47	37	51	57	58	61	67	71	77	81	87
Scleractinia	n/a	8	18	10	69	135	78	73	22	35	122	183	9	136	11	383	94	163	62
Filifera	n/a	7	8	3	13	43	10	17	8	1	8	4	1	14	15	36	16	55	19
Octocorallia	n/a	4	5	15	12	33	8	9	2	5	3	7	1	49	20	41	43	20	4
Antipatharia	n/a	1	1	3	2	11	3	2	0	1	2	4	0	1	5	7	11	3	3

\* Mediterranean and Black Sea.

<sup>1</sup> Area 18: Arctic Sea; Area 21: Atlantic, Northwest; Area 27: Atlantic, Northeast, Area 31: Atlantic, Western Central; Area 34: Atlantic, Eastern Central; Area 37: Mediterranean and Black Sea; Area 41: Atlantic, Southwest; Area 47: Atlantic, Southeast; Area 48: Atlantic, Antarctic; Area 51, Indian Ocean, Western; Area 57: Indian Ocean, Eastern; Area 58: Indian Ocean, Antarctic and Southern; Area 61: Pacific, Northwest; Area 67: Pacific, Northeast; Area 71: Pacific, Western Central; Area 77: Pacific, Eastern Central; Area 81: Pacific, Southwest; Area 87: Pacific, Southeast; Area 88: Pacific, Antarctic.

### Order Scleractinia

Within the approximately 790 species of azooxanthellate Scleractinia, 540 occur in waters deeper than 200 m; the deepest record being a delicate solitary species (*Fungiacyathus marenzelleri* [Vaughan, 1906]) collected from Pacific waters in the Aleutian Trench at 6 328 m (Keller, 1976). Although, most of the azooxanthellate scleractinians are reported from waters ranging in depth from 200 to 1 000 m (the prime depth for this group [Cairns, 2007]), which is the same depth of most cold-water coral reefs and mounds occur (Roberts *et al.*, 2009).

The best-studied areas for deep-sea Scleractinia are found in the north Atlantic waters; the Caribbean and northeast Atlantic (especially Norwegian continental shelf). However, these well-studied regions do not show the scleractinian diversity compared to other less-studied regions. The regions of high diversity are usually found in the western margins of ocean basins or where the continental shelf is “passive” such as the western Indian and the northwestern Atlantic oceans having at least twice as more species compared to the eastern margin of the same oceans. Nevertheless, the northern Indian ocean is considered as one of the poorest known azooxanthellate coral fauna in the world, and its knowledge is based on early expeditionary reports (Alcock, 1893, 1898, 1902a, 1902b, 1902c; Marenzeller, 1907; Gardiner and Waugh, 1938, 1939).

As for polar and sub-polar regions, the scleractinian fauna is depauperate compared to temperate regions. The number of species recorded from tropical Pacific and tropical Atlantic waters is higher than in other regions (Keller, 1998; Cairns, 2007). FAO Major Fishing Area 71 (Pacific, Western Central) being the most diverse area for azooxanthellate Scleractinia.

In Table 3, each one of the 19 FAO Major Fishing Areas is briefly described, and information regarding taxonomic literature, fisheries impact, status on the knowledge, and probability to have additional azooxanthellate Scleractinia records are provided. See Appendix 1 for the extended list of species per FAO Major Fishing Area.

**Table 3**

**Description of 19 FAO Major Fishing Areas and information on taxonomic literature (TL), fisheries impact, status on the knowledge, and probability to have additional records for azooxanthellate Scleractinia**

Area	Description	Fisheries impact	Status on knowledge	Prob. of add. spp. records <sup>1</sup>
18	There is no record of deep-sea scleractinians from this area. It could be related to the depth of the aragonite saturation horizon, which is estimated to be very shallow or even reach surface. The north Pacific region (above 40°N) has one of the shallowest aragonite saturation horizon recorded to date (Guinotte <i>et al.</i> , 2006). TL: None	Unknown	Good	Very small
21	To date there are 10 species of azooxanthellate Scleractinia recorded from this region, 9 of them are solitary and 1 is colonial ( <i>L. pertusa</i> ). This part of the Atlantic includes both Canada and United States exclusives economics zones (EEZs), and has been extensively trawled in the past years (Gianni, 2004). TL: Cairns (1979, 1981, 2000).	High	Good	Small
27	One of the most extensively studied areas for cold-water corals, the northeastern Atlantic ocean (North of 36°N) has 69 recorded species of azooxanthellate scleractinians. 10 of them occur only in waters shallower than 200 m. Within the remaining 59 species, 33 belong to the family Caryophylliidae, 9 to Dendrophylliidae, 6 to Flabellidae, 3 to Fungiacyathidae, 2 to Turbinoliidae, 2 to Pocilloporidae, and 1 representative of each: Guyniidae, Oculinidae, Schizocyathidae, and Stenocyathidae. Large concentrations of <i>L. pertusa</i> , which together with <i>S. variabilis</i> and <i>M. oculata</i> are the main deep-sea framework building species in the region. TL: Squires (1961), Zibrowius (1980, 1985), Fosså <i>et al.</i> (2002).	High	Excellent	Small
31	The central western Atlantic is one of the best known region for deep-sea azooxanthellate scleractinians, studied especially off Florida and Gulf of Mexico. More recently, additional records and new species have been reported from the Colombian Caribbean. Altogether, there are 135 species recorded in this region (30 restricted to waters shallower than 200 m). Within the remaining 105 species, 42 are known only for this region. TL: Pourtalès (1867, 1868, 1871, 1874, 1878, 1880), Cairns (1977, 1978a, 1978b, 1979, 2000), Humann & Deloach (2002), Santodomingo <i>et al.</i> (2007), Reyes <i>et al.</i> (2009).	High	Excellent	Small
34	78 species of azooxanthellate scleractinians belonging to all but Anthemiphylliidae, Gardineriidae, and Micrabaciidae are reported to the tropical realm of the eastern Atlantic Ocean. Fourteen of them are restricted to waters shallower than 200 m. The number of species to be reported is likely to increase with further expeditions. TL: Chevalier (1966a, 1966b), Zibrowius (1980, 1983), and Zibrowius and Gili (1990).	High?	Low	High

Area	Description	Fisheries impact	Status on knowledge	Prob. of add. spp. records <sup>1</sup>
37	Despite being extensively studied, the Mediterranean Sea has just 35 species of azooxanthellate scleractinian known to date of which 12 are restricted to shallow waters. Large deep-sea reefs formed mainly by <i>L. pertusa</i> and <i>M. oculata</i> have been recently described. TL: Zibrowius (1977, 1980, 1983).	High	High	Low/ Medium
41	Considered as a transitional region between the Caribbean and Antarctic azooxanthellate coral fauna, this area has 73 deep-sea scleractinian recorded to date (10 restricted to shallow waters). Amongst the 63 remaining species, large concentrations of <i>L. pertusa</i> , <i>M. oculata</i> , <i>S. variabilis</i> and <i>P. conferta</i> are recorded from southern and southeastern coast off Brazil (Pires <i>et al.</i> , 2004; Castro <i>et al.</i> , 2006; Kitahara, 2006; 2007; Pires, 2007; Kitahara <i>et al.</i> , 2008). However, there are at least 5 potentially rich locations for deep-sea Scleractinia that have been poorly or not studied to date: Rio Grande elevation, St. Peter and St. Paul Rocks, Fernando de Noronha Archipelago, and Trindade seamounts chain. Also, the northeastern and northern Brazilian coast never had extensive deep-sea coral sampling. TL: Pourtales (1874), Tommasi (1970), Cairns (1979, 2000), Zibrowius (1988), and Kitahara (2006, 2007).	High <sup>2</sup>	Medium	High
47	Few species of azooxanthellate scleractinians have been reported to the southeastern (from 6°S to 50°S) Atlantic ocean. From the 22 known occurrences, 3 <i>Balanophyllia</i> , 1 <i>Paraconotrochus</i> , 1 <i>Sclerhelia</i> , and 1 <i>Phyllangia</i> are restricted to waters shallower than 200 m. It is expected that the number of species reported to this area will increase with upcoming collecting campaigns. TL: Gardiner (1904) and Zibrowius and Gili (1990).	Unknown	Low	High
48, 58, 88	Treated as one large region, the three areas (48, 58, and 88) are located within the Antarctic and sub-Antarctic waters. Despite the large collecting effort only 18, 9, and 8 species have been recorded so far for each of these FAO areas respectively. The only species that occurs in waters shallower than 200 m is the Flabellidae <i>Flabellum gardineri</i> (area 48). Other species that apparently are endemic to Antarctic and sub-Antarctic regions are: <i>Caryophyllia eltaninae</i> (Cairns, 1982), <i>C. mabahithi</i> (Gardiner and Waugh, 1938), <i>Flabellum curvatum</i> (Moseley, 1881), <i>F. flexuosum</i> (Cairns, 1982), <i>F. impensum</i> (Squires, 1962), <i>Leptopeneus antarcticus</i> (Cairns, 1989), and <i>Paraconotrochus antarcticus?</i> (Gardiner, 1929). According to Guinotte <i>et al.</i> (2006), Antarctic and sub-Antarctic regions have shallow aragonite saturation horizon, which delay the establishment of many species of azooxanthellate Scleractinia. TL: Squires (1962), Eguchi (1965), and Cairns (1982).	Low	High	Small
57	According to Cairns (2007) the Bay of Bengal is one of the poorest known coral fauna in the world. To date due to extensive collection effort made by Australian institutions around the western and southern Australian coast, this area counts 183 species of azooxanthellate scleractinian (nearly 30% reported only from waters shallower than 200 m). This area counts 5 of the 11 known “facultative” species, and is one of the most diverse regarding the family Turbinoliidae. TL: Alcock (1894, 1898, 1902c), Cairns and Parker (1992), Cairns and Zibrowius (1997), Cairns (1998, 2004), and Venkataraman (2007).	High <sup>3</sup>	Low	High

Area	Description	Fisheries impact	Status on knowledge	Prob. of add. spp. records <sup>1</sup>
61	<p>This area has the fourth highest number of azooxanthellate scleractinian recorded to a single area (136 spp.). However, more than 30% of these species are reported only for waters shallower than 200 m. Within this region, the number of species decreases significantly towards the Arctic, 55°N being the northernmost record.</p> <p>TL: Eguchi (1938, 1942, 1965a, 1965b, 1972, 1973, 1975,), Yabe and Eguchi (1942), Mori (1964), Song (1982), Cairns (1994), Ogawa (2006), Tachikawa (2005, 2008), Lam <i>et al.</i> (2008).</p>	High	High	Low/ Medium
67	<p>This area is depauperate due to the shallow aragonite saturation horizon depth, the northeastern Pacific ocean (from 40°N to north) has only 11 species of azooxanthellate Scleractinia recorded to date, from which all but <i>L. pertusa</i> are solitary.</p> <p>TL: Cairns (1994)</p>	High	High	Low
71	<p>Considered as the most diverse area (hot-spot) for many deep-sea invertebrate groups, the central Indo-Pacific accumulate almost half of the known species of azooxanthellate scleractinians, with over 383 species being reported (including unpublished data from Kitahara and Cairns), of which 86 are restricted to shallow environments. One of the abiotic factors responsible for the presence of so many scleractinian species in this region is the underwater geomorphology which is one of the most complex in the world due to the large numbers of seamounts, ridges, and islands. Statistically, the azooxanthellates from this region, especially those from New Caledonia, have a strong affinity with the New Zealand ones, once these countries are underwater connected by a series of seamounts. Even with this high number of species recorded, species richness estimators (such Jackknife and Bootstrap) shows that this region is more diverse than known to date, and there is a large collection from New Caledonian waters (over 7 000 specimens) that have never been examined at the Muséum National d'Histoire Naturelle – Paris). Interestingly, this is one of the few regions in the world that the presence of <i>L. pertusa</i> is not as extensive as one would imagine, being collected only recently one small fragment of this species in the Philippines (Cairns, personal communication). However, the occurrence of <i>E. rostrata</i>, <i>Dendrophyllia alcocki</i> (Wells, 1954), <i>Madrepora</i> spp., <i>S. variabilis</i>, and other colonial species is very strong around the region. It is important to note that for almost all azooxanthellate scleractinian genera, this region is apparently the most diverse one (i.e. <i>Caryophyllia</i> (~39 spp.), <i>Crispatotrochus</i> (5 spp.), <i>Deltocyathus</i> (14 spp.), <i>Stephanocyathus</i> (6 spp.), <i>Trochocyathus</i> (21 spp.), <i>Gardineria</i> (4 spp.), <i>Balanophyllia</i> (~28 spp.), <i>Flabellum</i> (~21 spp.), <i>Truncatoflabellum</i> (22 spp.), and others).</p> <p>TL: Faustino (1927), Wells (1964), Cairns (1989), Cairns and Zibrowius (1997), Kitahara and Cairns (2008, 2009), Kitahara et al. (2010).</p>	High <sup>4</sup>	High	High

Area	Description	Fisheries impact	Status on knowledge	Prob. of add. spp. records <sup>1</sup>
77	With 94 azooxanthellate scleractinians reported (22 from shallow waters) to date, this area is the most diverse eastern oceanic region of the world. However, due to the high number of islands (i.e. Polynesian islands) never (or poorly) sampled, the number of species is probably much higher than observed. Within this area, the Hawaiian and Cocos islands are the most extensively studied. TL: Vaughan (1907), Wells (1954), Durham (1962, 1966), Keller (1981), and Cairns (1984, 1991, 1994, 2006). Wells (1954), Durham (1962, 1966), Keller (1981), and Cairns (1984, 1991, 1994, 2006).	Unknown	Medium	High
81	The azooxanthellates reported from New Zealand and adjacent waters support some of the most extensive fisheries in the world (orange-roughy). To date there are 163 species reported to this area corresponding to the third most diverse region in the world. TL: Eguchi (1973b), Cairns and Parker (1992), and Cairns (1995, 2004).	High	High	Low
87	Despite of the many taxonomic studies for the azooxanthellates from the Galapagos Archipelago (Durham, 1962; 1966; Wells, 1983; Cairns, 1991), the remaining areas of this FAO area still need sampling effort to be better understood in relation to azooxanthellate scleractinians. Recently, Cairns et al. (2005) reviewed the deep-sea corals from Peru-Chile, and interestingly some of the records of this fauna are very shallow in Chilean coast (i.e. <i>Desmophyllum dianthus</i> [Esper, 1794]). In total there are 62 species recorded from this FAO area (nearly 31% restricted to shallow environments). All the main deep-sea framework colonial species ( <i>L. pertusa</i> , <i>M. oculata</i> , <i>S. variabilis</i> , <i>G. dumosa</i> , and <i>E. rostrata</i> ) are reported to this region. TL: Durham (1962, 1966), Wells (1983), and Cairns (1991).	Unknown	Low/ Medium	Medium/ High

<sup>1</sup> Probability of additional species records.<sup>2</sup> The fisheries impact is high specially in southern Brazilian waters.<sup>3</sup> The fisheries impact is high especially around India and Indonesian islands.<sup>4</sup> The fisheries impact is high in some Philippine and Indonesian regions and probably lower around New Caledonia and northeastern Australia.

## **Order Antipatharia**

With an estimated number of species of 246, the vast majority of antipatharian species is reported from deep-seas (up to 8 600 m) and is widely distributed throughout the world's oceans, including both polar circles. Recently, Roark *et al.* (2009) discovered that some species of *Leiopathes* are among the oldest continuously living organisms on the planet (over 4 000 years). Appendix 2 lists the potential 33 habitat-forming species and follows the recently list published by Roberts *et al.* (2009). According to those authors, this list was based on actual observation of associated organisms and/or by inference based on their large size and local abundance.

## **Subclass Octocorallia**

The sub-class Octocorallia, also known as Alcyonaria and commonly referred as soft-corals, comprises 3 000 estimated species (3 orders, 45 families, and 334 genera)<sup>2</sup> of which nearly 2 400 are reported from waters deeper than 50 m (Cairns, 2007; Roberts *et al.*, 2009). The most diverse octocorallian order, the Alcyonacea, includes 30 families of soft corals (octocorals without a supporting skeletal axis) and gorgonians (octocorals with a supporting skeletal axis of scleroproteinous gorgonin and/or calcite) (Bayer, 1981; Daly *et al.*, 2007). These colonial organisms have an internal skeleton secreted by mesoglea and polyps with eight tentacles and eight mesenteries (8-fold symmetry). According to Daly *et al.* (2007) alcyonacean families are distinguished primarily on the basis of overall colony growth form, presence or absence of a supporting skeletal axis, and details of axial composition. The form and distribution of sclerites (microscopic calcite crystals embedded in the coenenchymal tissue and polyps) are the most important characters used to distinguish genera and species of octocorals, but are less important for familial distinctions.

With the deepest record of 6 620 m, octocorals are ubiquitous in marine environments, occurring from the Arctic to the Antarctic oceans. To date, no comprehensive list of species has been published; Roberts *et al.* (2009) list only potentially habitat-forming deep-sea species. However, it is important to note that some species listed in the Appendix 3 may not have habitat-forming capacity. According to Sánchez (2005), the paragorgiids are amongst the largest and most important sessile benthic invertebrates in the ocean, with some colonies being up to 10 m in height (Smith, 2001) thus providing an ample habitat for numerous fish and invertebrates (Krieger, 1993; Koslow *et al.*, 2001). Amongst the seven genera of primnoids included in the Appendix 3, the most important habitat-forming is probably *Primnoa*, sometimes up to 2 m in height and 7 m wide. The deep-sea species belonging to this genus were observed *in situ* in the Gulf of Alaska at depths between 161 and 365 m and 10 megafaunal groups have been identified that associate with them to feed on the coral, use the coral branches for suspension feeding, or for protection (Krieger & Wing, 2002). Lastly, the family Isididae is cosmopolitan in distribution and is reported as deep as 4 850 m (Bayer & Stefani, 1987).

## **Suborder Filifera**

The family Stylasteridae (lace-corals), sub-order filifera, are the second largest group of calcified cnidarians (246 to 250 spp.). They are ubiquitous in the marine environment from shallow to deep-seas and are recorded from the Antarctica to the Bering Sea (Cairns, 1992; Lindner *et al.*, 2008; Roberts *et al.*, 2009). Stylasterids have an extremely varied morphology ranging from encrusting to massive colonial species with very fine dendritic branches or broad flat plates oriented perpendicular to the current. Generally they have slow growth rates, colored skeleton, long life, and internal fertilization (Brooke and Stone, 2007). It is hypothesized that this group is out-competed by aggressive species in high nutrient waters, finding refuge in low nutrient and less competitive environments (Sanders, 1979; Cairns, 1992). Most species occur in deep-seas, being the most diverse bathymetric range for these corals between 200 to 500 m (Cairns, 1984; 1992). Broch (1942) was the first to comment on the worldwide distribution of this group, being followed by Boschma (1953) who

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<sup>2</sup> See the online research archive: <http://researcharchive.calacademy.org/research/izg/OCTOHT.htm>

concluded that the most diverse faunas of lace-corals were in the “East Indies”, off Japan, and around the Aleutian Islands.

More recently, Cairns (1992) shows the distribution of all known Stylasterids primarily found off small volcanic oceanic islands, atolls, archipelagos, and especially seamounts. This distribution pattern was attributed by Cairns (1991b) as an inability of the group to establish in terrigenous sediment. According to Cairns (1992b), although apparently having a widely and homogenously distribution pattern, the stylasterids are either absent or extremely rare near land masses over 105 000 km<sup>2</sup>, with exception of Honshu, Japan. Nonetheless, islands smaller than 36 000 km<sup>2</sup> generally support this group with some exceptions from high islands surrounded by broad continental shelves. Areas that are particularly favorable for stylasterids are: Norfolk, Kermadec, and Macquarie Ridges, and New Zealand, which altogether present the highest stylasterid species diversity in the world (Cairns, 1991b). More recently, large concentrations have been found around Antarctic and sub-Antarctic regions (especially around south-Georgia island), and in the Aleutian Trench.

To date, the most diverse regions for stylasterids are found in the Caribbean (55 spp.) and around New Zealand (43 spp.). They are extremely rare from northern Indian Ocean, Asian coast to the Sea of Japan, Australia, Africa, northern Indian Ocean, and North, Central, and South America (Cairns, 1991b). However, recently, some shallow and deep-sea species have been collected from Brazilian coast (Lindner, personal communication). According to Cairns (1991b), species of this group have never been found in the Canadian and Russian Arctic oceans, regardless of the island size. The northernmost records of stylasterids in the North Pacific are around 57°N to 58°17'N including the Gulf of Alaska (Fisher, 1938), Okhotsk Sea (Naumov, 1960), and off Pribilof Island in the Bering Sea (Cairns, 1991a). In Atlantic waters, the northernmost record belongs to Norwegian region at 68°30' (Zibrowius & Cairns, 1992).

See Appendix 4 for the extended list of species per FAO major fishing areas.

## 2. Vulnerability of deep-sea corals

Recent studies have highlighted the loss of high-trophic level predatory species (Myers and Worm, 2003) and progressively the lower trophic level fish recorded in landings. Due to escalating demand of food and the collapse of fisheries on continental shelves around the world, the fishing industry has progressively increased efforts on long-lived, slow-growing deep-sea (more than 200 m) demersal fishes (i.e. *Lophius gastrophysus*). According to Roberts *et al.* (2009), this trend brought fishing activity progressively closer to areas where deep-sea corals occur.

Among the four most common demersal fisheries (bottom-trawl, bottom-gillnet, bottom-longline and trap), the bottom-trawl is unquestionably the one causing the most threat and damage to sessile invertebrate communities such as corals and sponges. Furthermore, with the development of robust gears and nets, it is not uncommon for large trawlers to fish over large deep-sea reefs.

Fosså *et al.* (2002) described underwater remotely operated vehicle (ROV) observations on high trawled areas, and in many instances they found that areas previously known as *L. pertusa* habitats, showed only scarred or crushed and broken coral skeleton fragments. They also observed lost trawl warps, nets and furrows ploughed, estimating that up to 50 percent of *L. pertusa* reefs in Norwegian waters have been damaged by trawling. Similar undeniable evidences of large impacts on deep-sea corals have been reported for several countries including Australia (Koslow *et al.*, 2001), Canada (Gass and Williams, 2005), Ireland (Hall-Spencer *et al.*, 2002), (Probert, 1997), United Kingdom (Wheeler *et al.*, 2005), United States (Reed, 2002) and Brazil (Kitahara, 2009).

As well as the obviously direct effects, bottom trawling also disperse seabed sediments, increasing the water-column turbidity. The elevation of turbidity is known to be very harmful for many deep-sea species, especially for lace-corals.

In southern Brazilian waters, Kitahara (2009) showed that not only bottom-trawlers, but also other demersal fisheries were seriously damaging unidentified deep-sea coral reefs. Onboard vessel, scientific observers from the same region concluded that most fishing vessel captains were waiting for signals of corals on their “fish finder” eco sounders to release the fishing gear. Not surprisingly, representatives of scleractinians, octocorallians and antipatharians were usually part of the by-catch. In the same study, Kitahara (2009) compared the main areas used by demersal fisheries and concluded that it overlapped with the known coral distribution of the region.

To date, it is known that not only large colonial species are vulnerable to fishing gear, but also even the very small ones were reported as by-catch. Due to their very low growth rate (mature deep-sea coral reefs take thousands of years to accumulate), their unknown resilience (recovery from trawling impacts, if occurs, is likely to be slow, and where corals were completely destroyed recovery is unlikely [Gianni, 2004]), and their ecological importance (once destroyed, all associated animal community will be lost), it is considered that all known species of deep-sea corals to be “extremely vulnerable” to fishing gear in both, direct and indirect ways.

#### Other threats to deep-sea corals

- Hydrocarbon industries: Mirroring the offshore trend of fisheries, the hydrocarbon industry is exploiting oil and gas reservoirs in progressively deeper waters (Roberts *et al.*, 2009). To date, the risks imposed by this industry is still poorly understood, however, it may be affecting the nearby deep-sea corals with increasingly turbidity and leaking of fluids.
- Mining: According to Roberts *et al.* (2009, 246), improved undersea technologies have brought deep-sea mining into deep-sea coral regions. These initiatives raise concerns that disturbance from mining operations, including sediment plumes and the release of toxic material, could irreparably damage benthic ecosystems.
- Climate change: Probably the biggest threat facing deep-sea corals today is increasing seawater temperatures, progressive ocean acidification, and the raise of the aragonite saturation horizon. If recent predictions are correct (Guinotte *et al.*, 2006), changes of the carbonate chemistry of the ocean will have devastating implications not only for deep-sea corals, but also for all calcareous organisms and species that rely on the habitats formed by them.
- Coral collection: Many species of octocorallians and antipatharians have been collected for hundreds of years and used as material for the production of jewellery and medicine. In Brazil, it is not uncommon to find pet stores selling small to medium sized skeletons of deep-sea scleractinians (collected as bycatch and brought to the stores by the fishermen as additional source of money) for aquarists.

## 7. Recommendations for the production of identification guides

Due to i) high phenotypic plasticity of coral morphology; ii) low number of specimens used for species descriptions (especially before 1970); iii) the utilization of mainly macro-morphological characters to identify specimens to species level (especially for scleractinians); iv) differences of morphology during ontogeny; and v) the subjectivity on the interpretation of descriptions, it is not unusual to find species that have been described at least twice, sometimes by the same taxonomist.

One of the most effective ways to minimize this problem is to i) create an illustrated guide for all structures used in the identification for each particular group; ii) develop an illustrated identification key for families, genera, and species; and iii) compile and produce an illustrated identification guide for each one of the species of cold-water coral known to date including all the new information included in this draft report (with some adjustments).

However, it is expected that some difficulties will be faced when the production of the identification guides start, especially for some species described in the nineteenth century that were not illustrated and the types are presumably lost. Also, there are some species from which only the type specimens are known, and if the exchange of specimens be discouraged or impeded by regulations from international transport of specimens (Convention on International Trade in Endangered Species, [CITES]), it will be important that FAO support museum visits. Additional problems that could cause delays is the extensive work needed for the production of the identification guide regarding the sub-class Octocorallia, which is very diverse if compared within the other three groups mentioned in this report.

Extra efforts will be generated if micro structural data will be included in the scleractinians identification guide due to the limited number of scientists/taxonomists able to perform such examination. Focusing on the octocorallians, antipatharians, and Filifera, extra effort will be generated by the production of their respective plates, since most require scanning electron microscope images to be well illustrated.

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### Octocorallia literature

A very extensive list of literature on octocorallians is available online at: <http://researcharchive.calacademy.org/research/izg/Octocoralbiblio.htm>

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**APPENDIX 1****Order Scleractinia**

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<b>Family Anthemiphylliidae</b>				
<i>Anthemiphyllia</i>	<i>A. dentata</i>	50	1 050	57, 61, 71, 81
	<i>A. frustum</i>	209	340	61, 71
	<i>A. macrolobata</i>	369	650	71, 77, 81
	<i>A. multidentata</i>	128	270	57, 71, 81
	<i>A. pacifica</i>	205	342	71, 77, 81
	<i>A. patera costata</i>	320	700	71
	<i>A. patera patera</i>	500	700	31
	<i>A. sp. sensu Kitahara &amp; Cairns, unpublished</i>	346	346	71
	<i>A. spinifera</i>	282	650	71, 81
<b>Family Caryophylliidae</b>				
<i>Anomocora</i>	<i>A. carinata</i>	614	785	77
	<i>A. cf. fecunda sensu Cairns, 1995</i>	145	400	77, 81
	<i>A. fecunda</i>	37	640	27, 31, 34, 41, 61
	<i>A. gigas</i>	183	510	51, 71
	<i>A. marchadi</i>	32	229	31, 34, 51, 57, 71
	<i>A. prolifera</i>	30	329	31, 34
<i>Aulocyathus</i>	<i>A. atlanticus</i>	450	1 716	27, 34
	<i>A. juvenescens</i>	182	463	51, 71
	<i>A. matricidus</i>	84	207	61
	<i>A. recidivus</i>	128	1 137	51, 57, 61, 71, 81
<i>Bathycyathus</i>	<i>B. chilensis</i>	26	420	87
<i>Bourneotrochus</i>	<i>B. stellulatus</i>	210	566	71, 77, 81
<i>Caryophyllia</i>	<i>C. abrupta</i>	300	650	71
	<i>C. abyssorum</i>	732	2 000	27, 34
	<i>C. alaskensis</i>	102	399	61, 67
	<i>C. alberti</i>	76	506	27
	<i>C. ambrosia ambrosia</i>	311	1 230	21, 27, 34, 37, 47, 51, 57, 61, 71, 81
	<i>C. ambrosia caribbeana</i>	183	1 646	31, 41
	<i>C. antarctica</i>	87	1 435	41?, 48, 58,
	<i>C. antillarum</i>	150	730	31, 41
	<i>C. arnoldi</i>	40	656	67, 77
	<i>C. aspera</i>	400	400	71
	<i>C. atlantica</i>	298	2 165	27, 34, 37, 57, 71, 77, 81
	<i>C. balaenacea</i>	882	1 112	47
	<i>C. barbadensis</i>	129	249	31, 41
	<i>C. berteriana</i>	91	1 033	31, 41, 51
	<i>C. calveri</i>	91	340	27, 34, 37
	<i>C. cf. calveri sensu Cairns 1999</i>	558	602	71
	<i>C. cincticulata</i>	282	384	51, 71
	<i>C. concreta</i>	215	570	71
	<i>C. cornulum</i>	1 525	2 603	61, 71
	<i>C. corona</i>	60	60	31
	<i>C. corrugata</i>	183	380	31

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>C. crosnieri</i>	133	1 050	51, 57, 71, 81
	<i>C. crypta</i>	12	291	31, 41
	<i>C. cyathus</i>	70	300	27, 34, 37
	<i>C. decamera</i>	124	263	57, 71
	<i>C. dentata</i>	90	263	71
	<i>C. diomedaeae</i>	225	2 200	51, 57, 71, 77, 81, 87
	<i>C. eltaninae</i>	101	814	48
	<i>C. ephyala</i>	420	420	51, 57, 71
	<i>C. foresti</i>	155	950	27
	<i>C. grandis</i>	183	596	51, 57, 71
	<i>C. grayi</i>	37	360	57, 61, 71
	<i>C. hawaiiensis</i>	85	650	61, 71, 77, 81
	<i>C. horologium</i>	55	175	31
	<i>C. huinayensis</i>	16	256	87
	<i>C. inornata</i>	0	100	27, 34, 37
	<i>C. japonica</i>	77	1 680	61, 81
	<i>C. jogashimaensis</i>	52	98	61
	<i>C. karubarica</i>	389	477	71
	<i>C. laevigata</i>	410	1 032	71
	<i>C. lamellifera</i>	89	1 152	51?, 71, 81
	<i>C. mabahithi</i>	278	1 022	48
	<i>C. marmorea</i>	331	500	71, 77
	<i>C. oblonga</i>	670	1 005	71
	<i>C. octonaria</i>	182	622	71
	<i>C. octopali</i>	410	627	71, 77
	<i>C. paradoxa</i>	786	786	51
	<i>C. paucipalata</i>	714	843	31
	<i>C. perculta</i>	54	316	87
	<i>C. planilamellata</i>	128	1 220	57, 71, 81
	<i>C. polygona</i>	700	1 817	31
	<i>C. profunda</i>	35	1 116	47, 48, 51, 57, 81
	<i>C. protei</i>	100	100	31
	<i>C. quadragenaria</i>	54	443	57, 61, 71, 77, 81
	<i>C. quangdongensis</i>	167	179	61
	<i>C. ralphae</i>	270	896	71, 81
	<i>C. rugosa</i>	71	724	51, 57, 61, 71, 77, 81
	<i>C. sarsiae</i>	520	2 200	27, 34, 37
	<i>C. scobinosa</i>	302	2 450	41, 51, 61, 71, 81
	<i>C. secta</i>	220	366	71
	<i>C. seguenziae</i>	1 000	1 400	27, 34
	<i>C. sewelli</i>	366	366	51
	<i>C. smithii</i>	40	400	27, 34, 37, 47, 51, 57?
	<i>C. solida</i>	373	488	87
	<i>C. sp. A sensu Kitahara et al., 2010</i>	416	433	71
	<i>C. sp. A sensu Seig &amp; Ziborwius, 1989</i>	570	610	71
	<i>C. sp. B sensu Seig &amp; Ziborwius, 1989</i>	825	860	71
	<i>C. sp. cf. C. ambrosia sensu Cairns, 1984</i>	56	206	77

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>C. sp. cf. C. scobinosa</i> sensu Cairns, 1994	119	805	61
	<i>C. spinicarens</i>	222	750	71
	<i>C. spinigera</i>	127	347	61, 71
	<i>C. squiresi</i>	406	659	41, 87
	<i>C. stellula</i>	166	420	51, 57
	<i>C. tangaroae</i>	810	1 000	71
	<i>C. transversalis</i>	210	397	71
	<i>C. unicristata</i>	251	620	57, 71
	<i>C. valdiviae</i>	882	2 670	47
	<i>C. versicolorata</i>	215	708	71
	<i>C. zanzibarensis</i>	238	302	51
	<i>C. zopyros</i>	73	618	31
<i>Ceratotrochus</i>	<i>C. franciscana</i>	79	80	77?
	<i>C. laxus</i>	583	678	77
	<i>C. magnaghii</i>	7	400	37
<i>Cladocora</i>	<i>C. debilis</i>	28	480	31, 34, 37, 41, 47
	<i>C. pacifica</i>	45	274	31, 77, 87
<i>Coenocyathus</i>	<i>C. anthophyllites</i>	65	250	27, 34, 37
	<i>C. bowersi</i>	9	302	77
	<i>C. brooki</i>	7	95	81
	<i>C. caribbeana</i>	5	100	31
	<i>C. cylindricus</i>	65	500	34, 37
	<i>C. goreau</i>	2	6	31
	<i>C. humanni</i>	21	21	31
	<i>C. parvulus</i>	79	399	31, 41
<i>Coenosmilia</i>	<i>C. arbuscula</i>	74	622	31, 61, 71
	<i>C. inordinata</i>	244	322	77
<i>Colangia</i>	<i>C. immersa</i>	1	347	31
	<i>C. jamaicaensis</i>	10	20	31
	<i>C. moseleyi</i>	18	54	71
	<i>C. multipalifera</i>	20	20	31
<i>Concentrotheca</i>	<i>C. laevigata</i>	183	772	27, 31, 41
	<i>C. vaughani</i>	313	316	87
<i>Confluphyllia</i>	<i>C. juncta</i>	135	385	71, 81
<i>Conotrochus</i>	<i>C. asymmetros</i>	210	510	71
	<i>C. brunneus</i>	97	1 051	51, 71, 81
	<i>C. funicolumna</i>	80	1 078	61, 71, 77, 81
<i>Crispatotrochus</i>	<i>C. cornu</i>	220	1 097	41?, 81
	<i>C. curvatus</i>	1 373	2 505	81
	<i>C. foxi</i>	82	272	67, 77
	<i>C. galapagensis</i>	84	806	87
	<i>C. gregarius</i>	460	460	71
	<i>C. inornatus</i>	120	400	57, 81
	<i>C. irregularis</i>	549	549	81
	<i>C. niinoui</i>	104	104	61
	<i>C. rubescens</i>	110	634	61, 71, 77
	<i>C. rugosus</i>	142	1 050	57, 71, 81
	<i>C. septumdentatus</i>	187	400	71
	<i>C. sp. cf. cornu</i>	220	241	31
	<i>C. sp.A sensu Cairns 1982</i>	2 305	2 329	81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>C. squiresi</i>	686	822	31
	<i>C. woodsi</i>	77	87	71
<i>Dactylotrochus</i>	<i>D. cervicornis</i>	73	400	61, 71
<i>Dasmosmilia</i>	<i>D. lymani</i>	37	366	27, 31, 34, 41, 61, 81
	<i>D. valida</i>	490	490	51
	<i>D. variegata</i>	110	600	31, 34, 41, 51
<i>Deltocyathus</i>	<i>D. agassizii</i>	494	1 115	31, 41
	<i>D. andamanicus</i>	187	397	51, 57, 71
	<i>D. calcar</i>	81	675	31, 41
	<i>D. cameratus</i>	305	1 175	71, 81
	<i>D. corrugatus</i>	250	600	71, 81
	<i>D. crassiseptum</i>	370	583	71
	<i>D. eccentricus</i>	183	1 000	27, 31, 34, 41
	<i>D. halianthus</i>	46	130	41
	<i>D. heteroclitus</i>	208	600	71
	<i>D. inusitatus</i>	410	777	71
	<i>D. italicus</i>	403	2 634	27, 31, 34, 41, 47
	<i>D. magnificus</i>	88	1 500	57, 61, 71, 81
	<i>D. moseleyi</i>	200	1 372	27, 31, 34, 41
	<i>D. murrayi</i>	1 948	2 312	51
	<i>D. nascornatus</i>	73	2 000	51
	<i>D. ornatus</i>	73	400	71, 81
	<i>D. parvulus</i>	1 940	5 080	87
	<i>D. philippinensis</i>	342	522	71
	<i>D. pourtalesi</i>	311	567	31
	<i>D. rotulus</i>	210	1 300	51, 57, 61, 71, 81
	<i>D. sarsi</i>	44	80	57
	<i>D. sp. A. sensu Cairns &amp; Keller, 1993</i>	207	315	51
	<i>D. sp. sensu Zibrowius, 1974</i>	496	2 278	27
	<i>D. stella</i>	206	597	71, 81
	<i>D. suluensis</i>	142	1 050	57, 71, 81
	<i>D. taiwanicus</i>	320	697	71
	<i>D. varians</i>	655	732	51
	<i>D. vaughani</i>	88	1 097	61, 71
<i>Desmophyllum</i>	<i>D. dianthus</i>	8	2 460	21, 27, 31, 34, 37, 41, 47, 48, 51, 57, 61, 67, 71, 77, 81, 87
	<i>D. quinarium</i>	37	37	71
	<i>D. striatum</i>	277	823	31
<i>Ericiocyathus</i>	<i>E. echinatus</i>	814	1 401	71
<i>Goniocorella</i>	<i>G. dumosa</i>	88	1 488	51, 61, 71, 81, 87
<i>Heterocyathus</i>	<i>H. aequicostatus</i> *	0	20	51?, 57, 61, 71
	<i>H. alternatus</i> *	0	319	57, 71
	<i>H. antoniae</i>	20	70	31
	<i>H. hemisphaericus</i>	2	140	57
	<i>H. japonicus</i>	0	20	61
	<i>H. sulcatus</i> *	11	312	57, 71
<i>Hoplangia</i>	<i>H. durotrix</i>	0	150	27, 34, 37, 81
<i>Labyrinthocyathus</i>	<i>L. quaylei</i>	37	293	77
	<i>L. delicatus</i>	155	1 000	51
	<i>L. facetus</i>	385	402	31
	<i>L. langae</i>	695	810	31

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>L. limatulus</i>	20	508	61, 71, 81
	<i>L.</i> sp. A sensu Cairns & Zibrowius, 1997	210	268	77
	<i>L.</i> sp. sensu Cairns, 1995	665	1 000	81
<i>Lochmaeotrochus</i>	<i>L. gardineri</i>	608	1 175	71
	<i>L. oculatus</i>	240	616	71
<i>Lophelia</i>	<i>L. pertusa</i>	39	3 383?	21, 27, 31, 34, 37, 41, 47, 51, 61, 67, 71, 77, 81, 87
<i>Monohedotrochus</i>	<i>M. capitolii</i>	150	1 200	41
	<i>M. circularis</i>	190	545	57, 71
	<i>M. epithecatus</i>	240	455	71
<i>Nomlandia</i>	<i>N. californica</i>	82	82	77
<i>Oxysmilia</i>	<i>O. corrugata</i>	180	190	71
	<i>O. rotundifolia</i>	46	640	31
<i>Paraconotrochus</i>	<i>P. antarctica</i>	87	728	41, 48, 58,
	<i>P. capense</i>	59	59	47
	<i>P. zeidleri</i>	304	558	57, 71, 81
<i>Paracyathus</i>	<i>P. andersoni</i>	?	?	57
	<i>P. arcuatus</i>	201	343	27, 34
	<i>P. cavatus</i>	?	?	51
	<i>P. conceptus</i>	229	805	51
	<i>P. coronatus</i>	?	?	51
	<i>P. darwinensis</i>	0	26	71
	<i>P. ebonensis</i>	?	?	77?
	<i>P. fulvus</i>	350	433	57
	<i>P. humilis</i>	23	104	87
	<i>P. indicus gracilis</i>	?	?	51
	<i>P. indicus indicus</i>	?	?	57
	<i>P. lifuensis</i>	73	73	71
	<i>P. molokensis</i>	161	260	77
	<i>P. montereyensis</i>	75	146	71, 77
	<i>P. parvulus</i>	73	73	71
	<i>P. persicus</i>	<50	<50	51
	<i>P. porcellanus</i>	?	?	77?
	<i>P. profundus</i>	?	?	51?, 57
	<i>P. pruinosus</i>	150	150	51, 61, 71
	<i>P. pulchellus</i>	17	1 260	27, 31, 34, 37, 41
	<i>P. rotundatus</i>	18	66	57, 71
	<i>P.</i> sp. A sensu Kitahara & Cairns, unpublished	305	370	71
	<i>P.</i> sp. sensu Zibrowius, 1974	80	100	27
	<i>P.</i> sp. sensu Cairns & Zibrowius, 1997	90	397	51, 71
	<i>P. stearnsii</i>	20	134	67, 77
	<i>P. stokesi</i>	?	?	51, 57
	<i>P. vittatus</i>	31	31	57
<i>Phacelocyathus</i>	<i>P. flos</i>	20	255	31, 41
	<i>P.</i> sp. sensu Zibrowius & Grygier, 1985	55	131	51
<i>Phyllangia</i>	<i>P. americana</i>	0	53	31, 41

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>P. americana mouchezi</i>	1	55	27, 34, 37
	<i>P. consagensis</i>	0	82	87
	<i>P. dispersa</i>	<50	<50	77
	<i>P. echinosepes</i>	90	90	61
	<i>P. granulata</i>	?	?	47
	<i>P. hayamaensis</i>	5	5	61
	<i>P. papuensis</i>	88	100	71
	<i>P. pequenatae</i>	48	112	31
<i>Polycyathus</i>	<i>P. andamanensis</i>	?	?	57
	<i>P. atlanticus</i>	?	?	31?
	<i>P. difficilis</i>	?	?	57
	<i>P. fulvus</i>	30	50	71
	<i>P. furanaensis</i>	6	52	71
	<i>P. fuscomarginatus</i>	?	?	57
	<i>P. Hodgsoni</i>	0	35	71
	<i>P. hondaensis</i>	55	64	77?, 87
	<i>P. isabela</i>	14	23	87
	<i>P. marigondoni</i>	35	35	71
	<i>P. mayae</i>	137	309	31
	<i>P. muellerae</i>	0	30	27, 37
	<i>P. norfolkensis</i>	10	20	81
	<i>P. octuplus</i>	90	441	71
	<i>P. palifera</i>	<30	<30	57?
	<i>P. persicus</i>	?	?	51
	<i>P. senegalensis</i>	12	143	31, 34, 41?
	<i>P. verrilli</i>	?	?	51, 57
<i>Pourtalosmilia</i>	<i>P. anthophyllites</i>	25	300	27, 34, 37
	<i>P. conferta</i>	55	191	31, 41
<i>Premocyathus</i>	<i>P. cornuformis</i>	137	2 360	21, 27, 31, 34, 41
	<i>P. dentiformis</i>	22	960	57, 61, 71, 81
<i>Rhizosmilia</i>	<i>R. elata</i>	70	380	71
	<i>R. gerdae</i>	123	549	31
	<i>R. maculata</i>	1	508	31, 41
	<i>R. multipalifera</i>	11	165	57, 71
	<i>R. robusta</i>	66	360	51, 71
	<i>R. sagamiensis</i>	60	98	61, 71
	<i>R. sp. sensu Kitahara &amp; Cairns, unpublished</i>	265	370	71
<i>Solenosmilia</i>	<i>S. variabilis</i>	45	2 165	27, 31, 34, 41, 47, 48, 51, 57, 58, 71, 81, 87
<i>Stephanocyathus</i>	<i>S. campaniformis</i>	882	1 610	47, 51
	<i>S. coronatus</i>	543	1 989	31, 71, 81
	<i>S. crassus</i>	400	1 557	27, 34
	<i>S. diadema</i>	795	2 553	31, 41
	<i>S. explanans</i>	180	1 016	51, 57, 71
	<i>S. imperialis</i>	2 436	2 474	57, 71
	<i>S. isabellae</i>	493	504	31
	<i>S. laevifundus</i>	300	1 158	31
	<i>S. moseleyanus</i>	1 000	2 000	27, 34
	<i>S. nobilis</i>	609	2 200	27, 34, 51
	<i>S. paliferus</i>	220	715	31, 41
	<i>S. platypus</i>	439	1 219	57, 81
	<i>S. regius</i>	563	2 160	57, 71, 81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>S. sp. sensu</i> Cairns & Parker, 1992	520	520	57
	<i>S. spiniger</i>	120	1 188	51, 57, 61, 71, 81
	<i>S. weberianus</i>	700	1 500	57, 61, 71, 81
<i>Sympodangia</i>	<i>S. albatrossi</i>	212	616	71
<i>Tethocyathus</i>	<i>T. cylindraceus</i>	183	649	31, 41, 71, 81
	<i>T. endesa</i>	15	240	87
	<i>T. minor</i>	73	73	71, 77
	<i>T. prahli</i>	303	333	31
	<i>T. recurvatus</i>	320	569	31
	<i>T. sp. sensu</i> Kitahara & Cairns, unpublished	400	400	71
	<i>T. variabilis</i>	250	860	27, 31, 34
	<i>T. virgatus</i>	137	1 200	71, 81
<i>Thalamophyllia</i>	<i>T. gasti</i>	13	200	34, 37
	<i>T. gombergi</i>	188	220	31
	<i>T. riisei</i>	4	914	31
	<i>T. tenuescens</i>	8	360	57, 71, 81
<i>Trochocyathus</i>	<i>T. aithoseptatus</i>	371	454	77
	<i>T. apertus</i>	20	230	57, 71
	<i>T. brevispina</i>	240	560	71
	<i>T. burchae</i>	35	144	57, 71, 77
	<i>T. caryophylloides</i>	115	344	61, 71
	<i>T. cepulla</i>	398	610	71, 81
	<i>T. cf. rawsonii</i> sensu Kitahara & Cairns, unpublished	237	1 100	71
	<i>T. cf. wellsi</i> sensu Kitahara & Cairns, unpublished	237	237	71
	<i>T. cooperi</i>	25	100	61, 71
	<i>T. decamera</i>	70	88	61
	<i>T. discus</i>	240	700	71
	<i>T. efateensis</i>	391	437	71
	<i>T. fasciatus</i>	238	238	31
	<i>T. fossulus</i>	205	775	31
	<i>T. gardineri</i>	274	470	71, 77
	<i>T. gordoni</i>	398	710	81
	<i>T. hastatus</i>	366	710	71, 81
	<i>T. japonicus</i>	150	450	61
	<i>T. laboreli</i>	125	300	41
	<i>T. longispina</i>	326	760	71
	<i>T. maculatus</i>	77	550	71, 81
	<i>T. mauiensis</i>	174	278	77
	<i>T. oahensis</i>	75	571	77
	<i>T. patelliformis</i>	1 010	1 250	71, 77
	<i>T. philippinensis</i>	54	330	57, 61, 71
	<i>T. porphyreus</i>	?	?	57
	<i>T. rawsonii</i>	55	700	31, 41, 47?, 51
	<i>T. rhombcolumna</i>	110	530	51, 71, 77, 81
	<i>T. semperi</i>	38	245	71
	<i>T. sp. A</i> sensu Cairns & Keller, 1993	74	315	51

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>T. sp. cf. aithoseptatus</i> sensu Cairns, 2004	80	86	71
	<i>T. spinosocostatus</i>	1 575	1 650	27, 34
	<i>T. vasiformis</i>	323	650	71
	<i>T. wellsi</i>	75	86	71
<i>Vaughanella</i>	<i>V. concinna</i>	500	3 018	27, 34, 71
	<i>V. margaritata</i>	1 267	1 267	27
	<i>V. multipalifera</i>	920	3 500	71, 81
	<i>V. oreophila</i>	646	1 050	81
	<i>V. sp. cf. oreophila</i> Kitahara & Cairns, unpublished	852	852	71
<b>Family Dendrophylliidae</b>				
<i>Astroides</i>	<i>A. calicularis</i>	0	30	34, 37
<i>Balanophyllia</i>	<i>B. bairdiana</i>	6	548	57, 71, 81
	<i>B. bayeri</i>	274	311	31
	<i>B. bonaespei</i>	?	?	51?, 57?
	<i>B. capensis</i>	27	37	47
	<i>B. caribbeana</i>	33	86	31
	<i>B. carinata</i>	33	124	57, 71
	<i>B. cedrosensis</i>	66	119	77
	<i>B. cellulosa</i>	80	850	27, 34, 37
	<i>B. chnous</i>	140	549	81
	<i>B. corniculans</i>	?	?	71, 77
	<i>B. cornu</i>	60	520	57, 61, 71
	<i>B. crassiseptum</i>	183	250	71
	<i>B. crassitheca</i>	190	508	71, 81
	<i>B. cumingii</i>	65	307	51?, 57, 61, 71
	<i>B. cyathoides</i>	45	494	31
	<i>B. dentata</i>	66	135	57, 71, 81
	<i>B. desmophylloides</i>	95	1 050	57, 71, 77, 81
	<i>B. diademata</i>	165	165	51
	<i>B. difusa</i>	6	274	51
	<i>B. dilatata</i>	?	?	57
	<i>B. dineta</i>	27	274	31, 41
	<i>B. diomedaeae</i>	110	307	77
	<i>B. dubia</i>	55	55	71
	<i>B. elegans</i>	0	293	67, 77
	<i>B. europaea</i>	0	50	37
	<i>B. floridana</i>	13	220	31, 34
	<i>B. galapagensis</i>	18	462	71, 87
	<i>B. gemma</i>	137	522	51, 57, 71
	<i>B. gemmifera</i>	81	81	51
	<i>B. generatrix</i>	96	535	57, 71
	<i>B. gigas</i>	90	640	57, 61, 71, 77, 81
	<i>B. hadros</i>	238	274	31
	<i>B. helenae</i>	18	18	47
	<i>B. imperialis</i>	18	120	57, 71
	<i>B. iwayamaensis</i>	1	3	71?, 77?
	<i>B. japonica</i>	154	237	61
	<i>B. kalakauai</i>	?	?	
	<i>B. laysanensis</i>	238	400	71, 77

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>B. malouinensis</i>	75	1 137	41, 48, 87
	<i>B. merguiensis</i>	?	?	57
	<i>B. palifera</i>	53	708	31
	<i>B. parallela</i>	18	18	71
	<i>B. parvula</i>	192	300	71
	<i>B. pittieri</i>	40	96	31
	<i>B. profundicella</i>	73	73	61, 71
	<i>B. redeviva</i>	90	256	71
	<i>B. regalis</i>	59	59	57
	<i>B. regia</i>	2	100	27, 34, 37
	<i>B. scabra</i>	?	?	57
	<i>B. scabrosa</i>	?	?	71
	<i>B. serrata</i>	190	194	71
	<i>B. sp. 1</i> sensu Kitahara & Cairns, unpublished	311	470	71
	<i>B. sp. 2</i> sensu Kitahara & Cairns, unpublished	192	265	71
	<i>B. sp. 3</i> sensu Kitahara & Cairns, unpublished	235	370	71
	<i>B. sp. 4</i> sensu Kitahara & Cairns, unpublished	400	748	71
	<i>B. sp. 5</i> sensu Kitahara & Cairns, unpublished	265	421	71
	<i>B. sp. 6</i> sensu Kitahara & Cairns, unpublished	248	356	71
	<i>B. sp. A</i> sensu Cairns, 1994	56	100	61
	<i>B. sp. cf. serrata</i> Kitahara & Cairns, unpublished	180	216	71
	<i>B. sp. sensu</i> Cairns 1982	247	253	41
	<i>B. spongiosa</i>	11	18	81
	<i>B. stimpsonii</i>	18	95	51, 57, 71, 81
	<i>B. striata</i>	18	18	47
	<i>B. taprobanae</i>	?	?	51?
	<i>B. tenuis</i>	15	15	71
	<i>B. thalassae</i>	385	1 150	27, 34
	<i>B. vanderhorsti</i>	51	59	51, 61
	<i>B. wellsi</i>	412	575	31
	<i>B. yongei</i>	?	?	71
<i>Bathypsammia</i>	<i>B. fallosocialis</i>	213	805	31
	<i>B. tintinnabulum</i>	183	549	31
<i>Cladopsammia</i>	<i>C. echinata</i>	222	470	71, 77
	<i>C. eguchii</i>	1	85	61, 77, 81, 87
	<i>C. gracilis</i>	0	95	57?, 61, 71, 87
	<i>C. manuelensis</i>	55	366	31, 34, 41
	<i>C. rolandi</i>	12	50	37
	<i>C. willeyi</i>	?	?	71, 77
<i>Dendrophyllia</i>	<i>D. aculeata</i>	11	21	71, 77
	<i>D. alcocki</i>	118	1 200	57, 71, 81
	<i>D. alternata</i>	276	900	27, 31, 41
	<i>D. arbuscula</i>	2	353	51, 57, 61, 71, 81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>D. boschmai bochmai</i>	40	200	57, 61
	<i>D. boschmai cyathoheloides</i>	55	100	61
	<i>D. californica</i>	42	462	77, 87
	<i>D. carleenae</i>	?	?	71
	<i>D. cladonia</i>	49	457	51
	<i>D. cornigera</i>	89	600	27, 34, 37, 51
	<i>D. cibrosa</i>	7	40	34, 61
	<i>D. dilatata</i>	97	132	34, 51
	<i>D. florulenta</i>	69	243	61, 71
	<i>D. futojiku</i>	2	2	61
	<i>D. granosa</i>	91	91	57
	<i>D. ijimai</i>	10	366	51, 61, 71, 81
	<i>D. incisa</i>	?	?	71
	<i>D. indica</i>	5	5	51?, 57
	<i>D. johnsoni</i>	373	462	87
	<i>D. laboreli</i>	1	70	34
	<i>D. minima</i>	2	2	61
	<i>D. minuscula</i>	?	?	51, 57
	<i>D. oldroydae</i>	40	576	77, 87
	<i>D. paragracilis</i>	1	10	61
	<i>D. radians</i>	?	?	
	<i>D. ramea</i>	40	150	27, 34, 37
	<i>D. robusta</i>	?	?	57
	<i>D. sp. A</i>	180	230	71
	<i>D. sp. B</i>	180	180	71
	<i>D. suprarbuscula</i>	90	90	61
	<i>D. velata</i>	?	?	71
<i>Dichopsammia</i>	<i>D. granulosa</i>	20	30	61
	<i>E. cornucopia</i>	91	960	27, 31
	<i>E. fistula</i>	86	910	51, 71, 77, 81
	<i>E. gaditana</i>	30	998	31, 34, 41, 51, 57, 61, 71, 77, 81
	<i>E. japonica</i>	114	1 050	61, 81
	<i>E. serpentina</i>	200	362	77
	<i>E. strigosa</i>	25	77	31
	<i>E. wellsii</i>	32	196	61, 71
	<i>E. profunda</i>	403	1 748	31
	<i>E. pusilla</i>	371	805	27, 34, 71, 81
	<i>E. rostrata</i>	110	2 165	27, 31, 34, 37, 41, 47, 51, 57, 61, 71, 77, 81, 87
<i>Endopachys</i>	<i>E. bulbosa</i>	220	251	57, 71
	<i>E. grayi</i>	37	386	51, 57, 61, 71, 77, 81, 87
<i>Endopsammia</i>	<i>E. philippensis</i>	0	62	51, 57, 71
	<i>E. pourtalesi</i>	?	?	87
	<i>E. regularis</i>	8	73	71
<i>Heteropsammia</i>	<i>H. cochleata</i> *	6	622	51*, 57, 71
	<i>H. eupssammides</i> *	1	38	57, 71
	<i>H. moretonensis</i>	11	48	71
<i>Leptopsammia</i>	<i>L. britannica</i>	160	665	27
	<i>L. chevalieri</i>	70	500	34
	<i>L. column</i>	20	20	57
	<i>L. crassa</i>	22	187	71
	<i>L. formosa</i>	400	900	27, 34

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>L. poculum</i>	90	90	71
	<i>L. pruvoti</i>	3	150	27, 34, 37
	<i>L. queenslandiae</i>	87	87	71, 81
	<i>L. stokesiana</i>	46	69	71
	<i>L. trinitatis</i>	15	40	31
<i>Notophyllia</i>	<i>N. etheridgi</i>	37	238	57, 81
	<i>N. hecki</i>	342	414	71
	<i>N. piscacauda</i>	22	51	57
	<i>N. recta</i>	40	458	57, 81
<i>Pourtalopsammia</i>	<i>P. togata</i>	155	775	51
<i>Rhizopsammia</i>	<i>R. annae</i>	0	80	51
	<i>R. bermudensis</i>	8	12	31
	<i>R. compacta</i>	35	110	51
	<i>R. goesi</i>	5	119	31, 41
	<i>R. minuta</i>	0	176	51, 61, 71
	<i>R. minuta</i> var. <i>bikiniensis</i>	?	?	71
	<i>R. minuta</i> var. <i>mutsuensis</i>	0	2	61
	<i>R. nuda</i>	5.5	220	57, 71
	<i>R. pulchra</i>	?	?	77?
	<i>R. verrilli</i>	2	278	57, 71, 77, 87
	<i>R. wellingtoni</i>	2	43	87
	<i>R. wettsteini</i>	4	4	51
<i>Thecopsammia</i>	<i>T. elongata</i>	271	576	71, 81
	<i>T. socialis</i>	183	879	31
<i>Trochopsammia</i>	<i>T. infundibulum</i>	532	1 472	31
<i>Tubastraea</i>	<i>T. coccinea</i>	0	110	31, 41, 51, 57, 61, 71, 77, 81, 87
	<i>T. diaphana</i>	1	30	51, 57, 61, 71, 77
	<i>T. faulkneri</i>	3	5	61, 71, 87
	<i>T. floreana</i>	2	5	87
	<i>T. micranthus</i>	0	50	34, 51, 57, 61, 71
	<i>T. tagusensis</i>	1	43	41, 87
<b>Family Flabellidae</b>				
<i>Blastotrochus</i>	<i>B. nutrix</i>	11	18	57, 71
<i>Falcatoflabellum</i>	<i>F. raoulensis</i>	366	402	81
<i>Flabellum</i>	<i>F. alabastrum</i>	357	2 000	21, 27, 31, 34, 47
	<i>F. angostomum</i>	?	?	81
	<i>F. angulare</i>	2 000	3 186	21, 27, 31, 34
	<i>F. angustum</i>	?	?	61
	<i>F. aotearoa</i>	130	1 300	71, 81
	<i>F. apertum apertum</i>	220	1 575	41, 48, 51, 58, 61, 71, 81, 87
	<i>F. apertum borealis</i>	307	1 141	61
	<i>F. arcuatile</i>	300	640	61, 71
	<i>F. areum</i>	1 647	2 229	41, 87
	<i>F. atlanticum</i>	357	618	31
	<i>F. australe</i>	36	1 026	57, 71, 81
	<i>F. campanulatum</i>	?	?	71
	<i>F. chunii</i>	200	700	27, 34
	<i>F. conuis</i>	1 994	2 603	61, 71
	<i>F. curvatum</i>	115	1 137	41, 48, 87
	<i>F. daphnense</i>	101	101	87

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>F. deludens</i>	106	1 035	51, 57, 61, 71
	<i>F. flexuosum</i>	101	659	48, 58,
	<i>F. floridanum</i>	80	366	31
	<i>F. folkesoni</i>	124	430	57
	<i>F. gardineri</i>	177	177	48
	<i>F. hoffmeisteri</i>	110	842	57, 71, 81
	<i>F. impensum</i>	46	2 260	41, 48, 58, 67, 81,
	<i>F. japonicum bythios</i>	1 095	1 720	51
	<i>F. japonicum</i> <i>japonicum</i>	128	1 141	51, 57, 61, 71
	<i>F. knoxi</i>	201	1 167	71, 81
	<i>F. lamellulosum</i>	187	490	57, 71, 81
	<i>F. lowekeyesi</i>	278	1 100	51, 57, 81
	<i>F. macandrewi</i>	128	1 260	21, 27, 34
	<i>F. magnificum</i>	225	740	57, 61, 71
	<i>F. marcus</i>	1 050	1 602	71, 77
	<i>F. marenzelleri</i>	179	390	57, 71
	<i>F. messum</i>	368	1 035	51, 57, 71, 81
	<i>F. moseleyi</i>	216	1 097	31
	<i>F. ongulense</i>	570	570	58
	<i>F. patens</i>	204	439	57, 61, 71
	<i>F. pavoninum</i>	73	665	51, 61, 71, 77
	<i>F. politum</i>	40	402	57, 61, 71
	<i>F. sexcostatum</i>	685	1 121	71
	<i>F. sibogae</i>	567	567	51
	<i>F. sp. A</i> sensu Cairns 1991	441	717	87
	<i>F. sp. cf. moseleyi</i> sensu Cairns & Zibrowius, 1997	441	1 058	71
	<i>F. sp. sensu</i> Cairns & Zibrowius, 1997	441	1 058	71, 81
	<i>F. thouarsii</i>	71	305	41
	<i>F. transversale conicum</i>	104	188	61
	<i>F. transversale</i> <i>transversale</i>	55	150	57, 61, 71, 81
	<i>F. transversale</i> <i>triangulare</i>	?	920	58?, 61, 71
	<i>F. tuthilli</i>	347	824	57
	<i>F. vaughani</i>	232	369	77
<i>Javania</i>	<i>J. antarctica</i>	53	1 280	41, 48, 71,
	<i>J. borealis</i>	247	348	61, 87
	<i>J. cailleti</i>	30	2 165	21, 27, 31, 34, 37, 41, 51, 61, 67, 77, 87
	<i>J. californica</i>	62	170	77
	<i>J. erhardti</i>	35	44	71
	<i>J. exserta</i>	91	455	71, 77
	<i>J. fusca</i>	271	1 045	71, 77, 81
	<i>J. insignis</i>	46	1 050	51, 61, 71, 77, 81
	<i>J. lamprotichum</i>	191	881	57, 71, 77, 81
	<i>J. pseudolabastrum</i>	625	1 557	27, 31
	<i>J. sp. 1</i> sensu Kitahara & Cairns, unpublished	>100	>100	71

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>J. sp. 2</i> sensu Kitahara & Cairns, unpublished	310	312	71
	<i>J. sp.</i> sensu Cairns & Zibrowius, 1997	209	291	71
<i>Monomyces</i>	<i>M. pygmaea</i>	5	150	27, 34, 37
	<i>M. rubrum</i>	0	410	81
<i>Placotrochides</i>	<i>P. cf. minuta</i> sensu Kitahara & Cairns, unpublished	426	426	71
	<i>P. cylindrica</i>	1 117	1 402	71
	<i>P. frustum</i>	497	1 378	31, 34, 41
	<i>P. minuta</i>	119	458	71, 77
	<i>P. scaphula</i>	462	1 628	51, 57, 61, 71, 81
<i>Placotrochus</i>	<i>P. laevis</i>	6	289	57, 71
<i>Polymyces</i>	<i>P. fragilis</i>	75	882	31, 41
	<i>P. montereyensis</i>	69	212	77
	<i>P. sp. 1</i> sensu Kitahara & Cairns, unpublished	>100	>100	71
	<i>P. wellsi</i>	355	1 203	31, 57, 71, 77, 81, 87
<i>Rhizotrochus</i>	<i>R. flabelliformis</i>	228	1 050	71, 81
	<i>R. levidensis</i>	1	73	71, 81
	<i>R. tuberculatus</i>	0	73	57
	<i>R. typus</i>	20	1 048	51, 57, 61, 71
<i>Truncatoflabellum</i>	<i>T. aculeatum</i>	11	132	57, 71
	<i>T. angostomum</i>	15	136	57, 71
	<i>T. angustum</i>	195	530	57, 71, 81
	<i>T. arcuatum</i>	350	364	81
	<i>T. australiensis</i>	90	220	57
	<i>T. candeanum</i>	70	290	61, 71
	<i>T. carinatum</i>	30	274	61
	<i>T. crassum</i>	?	?	71
	<i>T. cumingi</i>	46	132	57, 71, 81
	<i>T. dens</i>	286	555	71, 81
	<i>T. formosum</i>	42	933	51, 57, 61, 71
	<i>T. gardineri</i>	138	138	51, 61
	<i>T. inconstans</i>	183	183	51
	<i>T. incrustatum</i>	30	415	71
	<i>T. irregularare</i>	11	42	57, 71
	<i>T. macroeschara</i>	18	201	57, 71
	<i>T. martensii</i>	139	182	71
	<i>T. mortensenii</i>	50	455	57, 71
	<i>T. multispinosum</i>	62	183	51
	<i>T. paripavoninum</i>	394	1 450	51, 57, 71, 81
	<i>T. phoenix</i>	18	441	71, 81
	<i>T. pusillum</i>	85	460	51, 71
	<i>T. sp. A</i> sensu Cairns, 1994	964	3 010	34, 51, 61
	<i>T. sp. A</i> sensu Kitahara & Cairns, unpublished	256	256	71
	<i>T. sp. B</i> sensu Cairns, 1994	80	88	61, 81
	<i>T. sp. B</i> sensu Kitahara & Cairns, unpublished	300	300	71

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>T. sp. C</i> sensu Kitahara & Cairns, unpublished	216	346	71
	<i>T. sp. sensu</i> Kitahara & Cairns, unpublished	227	332	71
	<i>T. spheniscus</i>	2	174	57, 61, 71
	<i>T. stabile</i>	786	3 010	47, 51, 61, 71
	<i>T. stokesii</i>	12	256	51, 57, 71
	<i>T. trapezoideum</i>	1 630	1 630	77
	<i>T. truncum</i>	595	1 896	41, 48, 87
	<i>T. vanuatu</i>	240	335	71
	<i>T. veroni</i>	15	119	57, 71
	<i>T. vigintifarum</i>	288	1 050	71, 81
	<i>T. zuluense</i>	62	84	51
<b>Family Fungiacyathidae</b>				
<i>Fungiacyathus</i>	<i>F. crispus</i>	183	1 115	27, 31, 34, 41
	<i>F. dennanti</i>	190	770	57, 71, 81
	<i>F. fissidiscus</i>	282	287	71
	<i>F. fissilis</i>	212	503	77
	<i>F. fragilis</i>	200	2 200	21, 27, 34, 57, 71, 77, 81
	<i>F. granulosus</i>	287	1 050	57, 61, 71, 81
	<i>F. hydra</i>	882	886	47
	<i>F. marenzelleri</i>	1 820	6 328	21, 27, 31, 34, 41, 47, 48, 57, 58, 61, 67, 77, 81, 87,
	<i>F. margaretae</i>	440	1 175	71, 81
	<i>F. multicarinatus</i>	348	350	57
	<i>F. paliferus</i>	69	823	51, 57, 61, 71, 81
	<i>F. pliciseptus</i>	480	480	87
	<i>F. pseudostephanus</i>	3 840	5 120	51, 77?, 87
	<i>F. pusillus pacificus</i>	350	1 050	71, 81
	<i>F. pusillus pusillus</i>	285	685	31
	<i>F. sandoi</i>	77	600	71, 81
	<i>F. sibogae</i>	463	1 914	51, 57?, 71
	<i>F. sp. A</i> sensu Cairns 1994	3 175	4 110	61
	<i>F. stephanus</i>	245	2 000	51, 57, 61, 71, 81
	<i>F. symmetricus</i>	183	1 664	31, 41, 51, 57
	<i>F. turbinoliooides</i>	600	930	57, 71, 81
	<i>F. variegatus</i>	84	715	57, 61, 71, 81
<b>Family Gardineriidae</b>				
<i>Gardineria</i>	<i>G. hawaiiensis</i>	142	1 200	57, 71, 77, 81
	<i>G. minor</i>	2	146	31
	<i>G. paradoxa</i>	91	700	31, 71
	<i>G. philippinensis</i>	192	494	57, 71
	<i>G. simplex</i>	46	241	31
	<i>G. sp. 1</i> sensu Kitahara & Cairns, unpublished	265	311	71
<b>New genus sensu Seig &amp; Zibrowius, 1989</b>	sp. nov. sensu Seig & Zibrowius, 1989	315	677	71
<i>Stolarkicyathus</i>	<i>S. pocilliformis</i>	342	367	71
<b>Family Guyniidae</b>				
<i>Guynia</i>	<i>G. annulata</i>	28	653	27, 31, 34, 37, 51, 57, 61, 71, 77

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<b>Family Micrabaciidae</b>				
<i>Leptopenus</i>	<i>L. antarcticus</i>	2 005	2 897	48,
	<i>L. discus</i>	2 842	5 000	31, 41, 48, 57, 58, 61, 67, 77,
	<i>L. hypocoelus</i>	3 949	3 949	87
	<i>L. solidus</i>	2 000	3 250	61, 71
	<i>L. sp. A</i>	221	871	71
<i>Letepsammia</i>	<i>L. fissilis</i>	106	458	57, 71, 81
	<i>L. formosissima</i>	97	500	51, 57, 61, 71, 77, 81
	<i>L. franki</i>	50	650	51, 71
	<i>L. sp. 1 sensu Kitahara &amp; Cairns, unpublished</i>	426	426	71
	<i>L. sp. 2 sensu Kitahara &amp; Cairns, unpublished</i>	237	516	71
	<i>L. superstes</i>	77	710	61, 71, 81
<i>Rhombopsammia</i>	<i>R. niphada</i>	390	804	57, 61, 71, 81
	<i>R. squiresi</i>	622	1 401	57, 71
<i>Stephanophyllia</i>	<i>S. complicata</i>	73	700	51, 57, 71, 81
	<i>S. fungulus</i>	15	653	51, 57, 61, 71
	<i>S. neglecta</i>	49	555	57, 71
	<i>S. sp. sensu Kitahara &amp; Cairns, unpublished</i>	608	608	71
<b>Family Oculinidae</b>				
<i>Bathelia</i>	<i>B. candida</i>	500	1 250	41, 87
<i>Cyathelia</i>	<i>C. axillaris</i>	12	366	51, 57, 61, 71
<i>Madrepora</i>	<i>M. arbuscula</i>	212	658	71
	<i>M. carolina</i>	53	801	31, 41
	<i>M. cf. porcellana</i>	55	238	71
	<i>M. minutiseptum</i>	150	302	61, 71
	<i>M. oculata</i>	55	1 950	27, 31, 34, 37, 41, 51, 57, 61, 71, 77, 81, 87
	<i>M. porcellana</i>	55	516	57, 71
<i>Oculina</i>	<i>O. diffusa</i> *	0	25	31
	<i>O. profunda</i>	119	742	71, 77, 87
	<i>O. tenella</i> *	25	159	31
	<i>O. varicosa</i> *	5	128	31
	<i>O. virgosa</i>	29	1 050	71, 81
<i>Petrophyllia</i>	<i>P. rediviva</i>	0	7	71
<i>Sclerhelia</i>	<i>S. hirtella</i>	18	165	47
<b>Family Pocilloporidae</b>				
<i>Madracis</i>	<i>M. asanoi</i>	110	183	71
	<i>M. asperula</i> *	0	311	31, 34, 41
	<i>M. brueggemannii</i>	51	130	31, 41
	<i>M. hellana</i>	46	46	31, 51, 71
	<i>M. kauaiensis</i>	44	541	71, 77
	<i>M. kauaiensis</i> var. <i>macrocalyx</i>	160	260	77
	<i>M. myriaster</i>	37	1 220	31, 41
	<i>M. p. pharensis</i> *	6	333	27, 31, 34, 37, 41, 71
	<i>M. profunda</i>	112	327	27, 34
	<i>M. sp. A sensu Cairns, 1994</i>	46	110	61, 71

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>M. sp. cf. asperula</i> sensu Cairns, 1991	46	64	87
	<i>M. sp. cf. pharensis</i> (*?) sensu Cairns, 1991	30	343	77, 87
<b>Family Rhizangiidae</b>				
<i>Astrangia</i>	<i>A. atrata</i>	3	51	57, 81
	<i>A. browni</i>	0	5	87
	<i>A. californica</i>	29	29	77
	<i>A. conferta</i>	9	33	77
	<i>A. costata</i>	11	31	77
	<i>A. dentata</i>	0	15	77
	<i>A. equatorialis</i>	0	15	87
	<i>A. haimei</i>	1	53	77
	<i>A. howardi</i>	54	91	77
	<i>A. macrodentata</i>	?	?	27?, 34?
	<i>A. mercatoris</i>	3	6	34?
	<i>A. poculata</i>	0	263	31, 34
	<i>A. rathbuni</i>	5	90	31, 41
	<i>A. solitaria</i>	0	51	31, 41
	<i>A. woodsi</i>	18	18	71, 81
<i>Cladangia</i>	<i>C. exusta</i>	20	20	57, 71
<i>Culicia</i>	<i>C. australiensis</i>	3	378	57, 71
	<i>C. cuticulata</i>	?	?	57?
	<i>C. excavata</i>	?	?	51?, 57?
	<i>C. fragilis</i>	14	20	77?
	<i>C. hoffmeisteri</i>	0	51	57, 71, 81
	<i>C. quinaria</i>	0.5	30	71, 81
	<i>C. rachelfitzhardingeae</i>	2	2	77
	<i>C. rubeola</i>	0	82	57, 61, 81
	<i>C. smithii</i>	?	?	81
	<i>C. sp. cf. rubeola</i> sensu Cairns, 1991	0	27	77, 87
	<i>C. stellata</i>	0	100	61, 71, 87
	<i>C. subaustraliensis</i>	?	?	61
	<i>C. tenella natalensis</i>	?	?	51?
	<i>C. tenella tenella</i>	30	30	51, 71, 81
	<i>C. tenuisepes</i>	0.5	1.5	71
<i>Oulangia</i>	<i>O. bradleyi</i>	0	64	87
	<i>O. cyathiformis</i>	10	20	71
	<i>O. stokesiana</i>	1	22	57, 61, 71
	<i>O. stokesiana</i> var. <i>miltoni</i>	0	135	61
<b>Family Schizocyathidae</b>				
<i>Pourtalocyathus</i>	<i>P. hispidus</i>	349	1 006	31
<i>Schizocyathus</i>	<i>S. fissilis</i>	88	1 300	27, 31, 34, 41
<i>Temnotrochus</i>	<i>T. kermadecensis</i>	321	402	71, 81
<i>Pedicellocyathus</i>	<i>P. keyesi</i>	70	194	81
<i>Stenocyathus</i>	<i>S. vermiformis</i>	110	1 500	27, 31, 34, 37, 41, 51, 57, 61, 71, 77, 81, 87
<i>Truncatoguynia</i>	<i>T. irregularis</i>	80	334	61, 71, 81
<b>Family Turbinoliidae</b>				
<i>Alatotrochus</i>	<i>A. rubescens</i>	180	751	57, 61, 71, 81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<i>Australocyathus</i>	<i>A. vicentinus</i>	16	148	57, 71
<i>Conocyathus</i>	<i>C. formosus</i>	320	367	71
	<i>C. gracilis</i>	22	291	57, 71
	<i>C. zelandiae</i>	4	137	57, 71, 81
<i>Cryptotrochus</i>	<i>C. brevipalpus</i>	466	700	71
	<i>C. carolinensis</i>	320	338	31
	<i>C. javanus</i>	585	585	71
	<i>C. nascornatus</i>	183	457	51
	<i>C. pileus</i>	123	522	57, 61, 71, 81
<i>Deltocyathoides</i>	<i>D. orientalis</i>	44	635	51, 57, 61, 71, 77, 81
	<i>D. stimpsonii</i>	110	600	27, 31, 34, 41
<i>Dunocyathus</i>	<i>D. parasiticus</i>	64	549	57, 71, 81
	<i>D. wallaceae</i>	320	414	71
<i>Endocyathopora</i>	<i>E. laticostata</i>	46	100	57, 71
<i>Foveolocyathus</i>	<i>F. alternans</i>	27	238	57
	<i>F. kitsoni</i>	342	367	57, 71
	<i>F. parkeri</i>	73	183	57
	<i>F. verconis</i>	73	183	57, 71, 81
<i>Holcotrochus</i>	<i>H. crenulatus</i>	40	414	57, 71
	<i>H. scriptus</i>	9	342	57, 71
<i>Idiotrochus</i>	<i>I. alatus</i>	315	450	71, 81
	<i>I. australis</i>	274	274	71
	<i>I. emaciatus</i>	82	238	57
	<i>I. kikutii</i>	97	645	57, 61, 71
<i>Kionotrochus</i>	<i>K. suteri</i>	44	622	81
<i>Lissotrochus</i>	<i>L. curvatus</i>	342	367	71
<i>Notocyathus</i>	<i>N. conicus</i>	34	1 110	61, 71, 81
	<i>N. venustus</i>	70	555	57, 61, 71
<i>Peponocyathus</i>	<i>P. dawsoni</i>	87	988	81
	<i>P. folliculus</i>	30	582	27, 31, 34, 61, 71
	<i>P. minimus</i>	30	903	71
<i>Platytrochus</i>	<i>P. compressus</i>	64	130	81
	<i>P. hastatus</i>	27	148	57
	<i>P. laevigatus</i>	22	165	57
	<i>P. parisepta</i>	40	201	57
	<i>P. venustus</i>	200	397	71, 81
	<i>P. zibrowii</i>	1 137	1 137	71
<i>Pseudocyathoceras</i>	<i>P. avis</i>	91	183	87
<i>Sphenotrochus</i>	<i>S. andrewianus</i> <i>andrewianus</i>	12	105	27, 34, 37
	<i>S. andrewianus moorei</i>	9	42	31
	<i>S. aurantiacus</i>	155	366	51
	<i>S. auritus</i>	15	64	31, 41
	<i>S. cuneolus</i>	42	342	71
	<i>S. evexicostatus</i>	12	73	51
	<i>S. excavatus</i>	?	?	81
	<i>S. gardineri</i>	9	403	41, 87
	<i>S. gilchristi</i>	24	165	51
	<i>S. hancocki</i>	18	274	71, 87
	<i>S. imbricaticostatus</i>	37	347	51
	<i>S. lindstroemi</i>	22	78	31
	<i>S. ralphae</i>	7	104	81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>S. squiresi</i>	66	318	81
<i>Thrypticotrochus</i>	<i>T. petterdi</i>	95	925	51, 71, 81
<i>Trematotrochus</i>	<i>T. corbicula</i>	400	576	31
	<i>T. hedleyi</i>	150	457	71, 81
<i>Tropidocyathus</i>	<i>T. labidus</i>	206	536	57, 61, 71
	<i>T. lessonii</i>	50	421	51, 57, 61, 71
	<i>T. sp.</i>	346	346	71
<i>Turbinolia</i>	<i>T. stephensi</i>	9	32	71

**APPENDIX 2****Order Antipatharia**

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<b>Family Antipathidae</b>				
<i>Antipathes</i>	<i>A. atlantica</i>	21	140	31, 41
	<i>A. caribbeana</i>	11	76	31
	<i>A. curvata</i>	40	604	71, 77
	<i>A. dendrochristos</i>	96	200	77
	<i>A. grandis</i>	30	91	71, 77
<b>Family Aphanipathidae</b>				
<i>Aphanipathes</i>	<i>A. pedata</i>	87	292	21, 31
	<i>A. salix</i>	137	263	31
	<i>A. sarothonmoides</i>	67	111	51, 71
	<i>A. verticellata</i>	79	79	61
<b>Family Cladopathidae</b>				
<i>Chrysopathes</i>	<i>C. formosa</i>	640	950	77, 87
	<i>C. gracilis</i>	640	640	77
	<i>C. speciosa</i>	225	1 168	67
<i>Sibopathes</i>	<i>S. gephura</i>	1 224	1 224	57
	<i>S. macrospina</i>	448	559	31
<i>Trissopathes</i>	<i>T. pseudotristicha</i>	259	2 544	67, 77
	<i>T. tetracrada</i>	375	2 220	34, 57, 77
	<i>T. tristicha</i>	236	1 330	57, 61?, 71, 81
<b>Family Stylopathidae</b>				
<i>Stylopathes</i>	<i>S. americana</i>	159	503	31
<i>Tylopathes</i>	<i>T. atlantica</i>	?	?	34
	<i>T. crispa</i>	732	732	87
<b>Family Myriopathidae</b>				
<i>Plumopathes</i>	<i>P. pennacea</i>	15	190	31, 51
<i>Tanacetopathes</i>	<i>T. hirta</i>	12	262	31, 41
<b>Family Schizopathidae</b>				
<i>Bathypathes</i>	<i>B. alternata</i>	182	4 923	21, 31, 41?, 48, 61?, 67, 71, 81, 87, 88
<i>Dendropathypathes</i>	<i>D. boutillieri</i>	1 689	2 162	61, 67
<i>Lillipathes</i>	<i>L. lillei</i>	?	?	
	<i>L. quadribranchiata</i>	?	?	71
	<i>L. wingi</i>	225	2 083	67
<i>Parantipathes</i>	<i>P. tetrasticha</i>	96	1 165	21, 31
<i>Stauropathes</i>	<i>S. arctica</i>	1 535	1 850	27
	<i>S. punctata</i>	395	395	77
	<i>S. staurocrada</i>	220	1 770	77, 81
<b>Family Leiopathidae</b>				
<i>Adelopora</i>	<i>A. crassilabrum</i>	282	1 169	71, 81
	<i>A. moseleyi</i>	291	360	77, 81
	<i>A. pseudothyron</i>	298	915	41, 81, 87
	<i>A. fragilis</i>	400	1 570	71, 81
<i>Astya</i>	<i>A. aspidopora</i>	590	1 058	81
	<i>A. subviridis</i>	914	914	71
<i>Calyptopora</i>	<i>C. reticulata</i>	93	2 010	81
	<i>C. sinuosa</i>	142	762	71, 81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<i>Cheiloporidion</i>	<i>C. pulvinatum</i>	642	1 137	41
	<i>C. laevis</i>	130	1 052	51?, 61, 71, 81
	<i>C. verrucosa</i>	198	2 626	81, 88
	<i>C. candelabrum</i>	240	1 265	71, 81
	<i>C. tetrastichopora</i>	282	850	71, 81
	<i>C. gigantea</i>	356	544	81
	<i>A. adeta</i>	398	399	71
	<i>C. anthohelia</i>	440	1 175	81
	<i>C. unifacialis</i>	547	547	81
<i>Cryptphelia</i>	<i>C. dura</i>	?	?	51?, 57?
	<i>C. peircei</i>	89	838	31
	<i>C. micropoma</i>	140	140	51
	<i>C. studeri</i>	142	1 058	81
	<i>C. trophostega</i>	150	1 913	61, 67
	<i>C. insolita</i>	159	720	31
	<i>C. papillosa</i>	161	545	31
	<i>C. glebulenta</i>	166	806	87
	<i>C. cymas</i>	169	1 097	81, 87
	<i>C. cryptotrema</i>	170	474	71
	<i>C. glossopoma</i>	198	864	31
	<i>C. japonica</i>	247	551	61
	<i>C. lacunosa</i>	252	806	87
	<i>C. curvata</i>	287	1 058	81
	<i>C. polypoma</i>	290	966	71, 81
	<i>C. dactylopoma</i>	348	813	87
	<i>C. pudica</i>	368	1 633	57?, 61, 71, 81
	<i>C. robusta</i>	374	709	71, 81
	<i>C. vascomarquesi</i>	390	1 520	27, 34
	<i>C. formosa</i>	419	2 044	41
	<i>C. platypoma</i>	469	469	71
	<i>C. gigantea</i>	475	784	87
	<i>C. eueides</i>	475	813	87
	<i>C. floridana</i>	593	823	31
	<i>C. fragilis</i>	647	2 328	57, 71, 81
<i>Distichopora</i>	<i>C. affinis</i>	712	2 790?	27
	<i>C. tenuisepta</i>	761	1 557	27, 31, 34
	<i>C. medioatlantica</i>	861	2 644	27
	<i>C. ramosa</i>	1 165	1 264	71
	<i>C. balia</i>	1 633	1 633	71
	<i>C. clausa</i>	?	?	51?, 57?
	<i>C. stenopoma</i>	?	?	71?
	<i>C. lamellata</i>	83	550	67
	<i>D. sulcata</i>	60	708	31
	<i>D. cervina</i>	68	384	31
	<i>D. barbadensis</i>	108	159	31
	<i>D. contorta</i>	125	368	31
	<i>D. anomala</i>	139	311	31
	<i>D. rosalindae</i>	165	198	31
	<i>D. laevigranulosa</i>	166	806	87
	<i>D. foliacea</i>	174	527	31
	<i>D. asulcata</i>	293	377	77
	<i>D. uniserialis</i>	333	366	31

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>D. anceps</i>	360	736	77
	<i>D. dispar</i>	465	741	81
	<i>D. coccinea</i>	?	?	61?, 71?, 81?
	<i>D. gracilis</i>	?	?	61?, 71?, 81?
	<i>D. livida</i>	?	?	61?, 71?, 81?
	<i>D. profunda</i>	?	?	51?, 57?
	<i>D. providentiae</i>	?	?	51?, 57?
	<i>D. serpens</i>	?	?	51?, 57?
<i>Errina</i>	<i>E. reticulata</i>	79	128	
	<i>E. laterorifa</i>	79	2 273	48, 58, 81
	<i>E. aspera</i>	80	200	27?, 34, 37
	<i>E. japonica</i>	85	85	61
	<i>E. kerguelensis</i>	91	567	81, 88
	<i>E. boschmai</i>	100	686	41, 48, 88?
	<i>E. chathamensis</i>	106	688	81
	<i>E. sinuosa</i>	135	1 075	47, 81
	<i>E. dabneyi</i>	140	2 200	27
	<i>E. gracilis</i>	146	1 226	41, 48, 81, 87, 88
	<i>E. cochleata</i>	194	534	31
	<i>E. altispina</i>	198	309	31
	<i>E. cheilopora</i>	198	659	81
	<i>E. bicolor</i>	252	549	81
	<i>E. cooki</i>	256	380	81
	<i>E. laevigata</i>	333	371	81
	<i>E. macrogastra</i>	549	704	77, 87
	<i>E. atlantica</i>	610	983	27
	<i>E. cyclopora</i>	1 647	2 044	41
	<i>E. capensis</i>	?	?	27?, 34?, 47?
	<i>E. porifera</i>	?	?	61?, 71?, 81?
<i>Errinopora</i>	<i>E. nanneca</i>	63	375	61, 67
	<i>E. stylifera</i>	108	117	61
	<i>E. latifundata</i>	109	109	61
	<i>E. zarhyncha</i>	207	658	67
	<i>E. cestoporina</i>	339	1 240	41
	<i>E. fenestrata</i>	152	622	41, 87
	<i>E. reticulum</i>	250	771	41
<i>Inferiolabiata</i>	<i>I. lowei</i>	164	991	41, 48, 87
	<i>I. spinosa</i>	211	781	81
	<i>I. labiata</i>	238	274	51
<i>Leiopathes</i>	<i>L. acanthphora</i>	1 011	1 020	57
	<i>L. glaberrima</i>	37	2 400	27, 31, 34, 37, 71, 77
<i>Lepidopora</i>	<i>L. clavigera</i>	130	282	31
	<i>L. glabra</i>	183	1 170	21, 31, 41
	<i>L. biserialis</i>	196	370	81
	<i>L. cryptocymas</i>	197	506	81
	<i>L. polystichopora</i>	197	710	31
	<i>L. decipiens</i>	276	670	31
	<i>L. symmetrica</i>	282	1 058	81
	<i>L. dendrostylus</i>	345	975	81
	<i>L. microstylus</i>	405	1 258	71?, 81
	<i>L. eburnea</i>	460	983	27
	<i>L. sarmentosa</i>	502	1 665	71, 81, 88

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>L. acrolophos</i>	659	686	41
	<i>L. concatenata</i>	680	720	87
	<i>L. diffusa</i>	?	?	27?, 34?, 47?
	<i>L. granulosa</i>	?	?	48?, 58?, 88?
<i>Lepidotheca</i>	<i>L. cervicornis</i>	110	110	81
	<i>L. pourtalesi</i>	123	368	31
	<i>L. chauliostylus</i>	130	1 265	71, 81
	<i>L. macropora</i>	169	806	87
	<i>L. tenuistylus</i>	238	274	51
	<i>L. robusta</i>	365	365	81
	<i>L. ramosa</i>	520	520	71
	<i>L. fascicularis</i>	540	2 010	88
	<i>L. brochi</i>	545	864	31
	<i>L. altispina</i>	660	1 570	81
	<i>L. inconsuta</i>	787	1 500	81
	<i>L. horrida</i>	1 089	1 089	71
<i>Paraerrina</i>	<i>P. diciiens</i>	238	274	51
<i>Phalangopora</i>	<i>P. regularis</i>	238	274	51
<i>Pliobonthrus</i>	<i>P. symmetricus</i>	80	1 600	27, 31, 34
	<i>P. echinatus</i>	165	750	31
	<i>P. tubulatus</i>	419	708	31
	<i>P. fistulosus</i>	546	631	77
	<i>P. gracilis</i>	600	700	34
<i>Pseudocryptphelia</i>	<i>P. pachypoma</i>	430	1 089	71
<i>Sporadopora</i>	<i>S. dichotoma</i>	250	1 498	41, 48
	<i>S. martensi</i>	282	282	81
	<i>S. macropora</i>	465	741	81
<i>Stellapora</i>	<i>S. echinata</i>	215	1 846	41
<i>Stenohelia</i>	<i>S. maderensis</i>	110	1 125	27, 34
	<i>S. profunda</i>	159	2 021	31
	<i>S. concinna</i>	166	806	87
	<i>S. pauciseptata</i>	282	514	31
	<i>S. conferta</i>	?	?	61?, 71?, 81?
	<i>S. echinata</i>	?	?	61?, 71?, 81?
	<i>S. tiliata</i>	?	?	61?, 71?, 81?
	<i>S. umbonata</i>	?	?	61?, 71?, 81?
	<i>S. yabei</i>	?	?	61?, 71?, 81?
<i>Stephanohelia</i>	<i>S. praecipua</i>	318	1 277	71, 81
<i>Stylanthecca</i>	<i>S. papillosa</i>	?	?	67?, 77?, 87?
<i>Stylaster</i>	<i>S. brochi</i>	62	582	61, 67
	<i>S. bithalamus</i>	71	71	47
	<i>S. norvegicus</i>	75	1 400	27
	<i>S. brunneus</i>	80	290	71, 81
	<i>S. marenzelleri</i>	84	631	77, 87
	<i>S. verrillii</i>	87	366	67
	<i>S. polyorchis</i>	91	823	67
	<i>S. densicaulis</i>	96	1 362	41, 48
	<i>S. boreopacificus</i>	117	183	61
	<i>S. aurantiacus</i>	123	355	31
	<i>S. moseleyanus</i>	126	518	67
	<i>S. imbricatus</i>	128	1 037	81
	<i>S. miniatus</i>	146	530	31

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>S. erubescens</i>	146	1 440	21, 27, 31
	<i>S. cancellatus</i>	152	582	61, 67
	<i>S. antillarum</i>	174	653	31
	<i>S. filogranus</i>	183	274	31
	<i>S. complanatus</i>	183	707	31
	<i>S. inornatus</i>	198	309	31
	<i>S. elassotomus</i>	210	882	61
	<i>S. horologium</i>	228	1 169	71, 81
	<i>S. galapagensis</i>	252	806	77, 87
	<i>S. gracilis</i>	291	291	81
	<i>S. cocosensis</i>	293	576	77
	<i>S. robustus</i>	300	300	48
	<i>S. griggi</i>	322	583	77
	<i>S. multiplex</i>	372	450	71
	<i>S. spatula</i>	384	549	31
	<i>S. ibericus</i>	500	500	27
	<i>S. bilobatus</i>	520	520	57
	<i>S. infundibuliferus</i>	521	563	77
	<i>S. maroccanus</i>	1 378	1 378	34
	<i>S. alaskanus</i>	?	?	61?, 67
	<i>S. amphiheloides</i>	?	?	47, 48?
	<i>S. bellus</i>	?	?	61?, 71?, 81?
	<i>S. bocki</i>	95	95	61?, 71?, 81?
	<i>S. boschmai</i>	?	?	61?, 71?, 81?
	<i>S. carinatus</i>	?	?	61?, 71?, 81?
	<i>S. crassior</i>	?	?	51?, 57?
	<i>S. dentatus</i>	?	?	61?, 71?, 81?
	<i>S. flabelliformis</i>	?	?	51
	<i>S. granulosus</i>	?	?	61?, 71?, 81?
	<i>S. hattorii</i>	?	?	61?, 71?, 81?
	<i>S. incompletus</i>	?	?	61?, 71?, 81?
	<i>S. incrassitus</i> (not valid?)	?	?	61?, 71?, 81?
	<i>S. microstriatus</i>	?	?	61?, 71?, 81?
	<i>S. nobilis</i>	?	?	47
	<i>S. polymorphus</i>	?	?	
	<i>S. profundus</i>	?	?	21?, 31?, 41?
	<i>S. profundiporus</i>	?	?	61?, 71?, 81?
	<i>S. pulcher</i>	?	?	61?, 71?, 81?
	<i>S. purpuratus</i>	?	?	61?, 71?, 81?
	<i>S. ramosus</i>	?	?	51?, 57?
	<i>S. scabiosus</i>	?	?	61?, 71?, 81?
	<i>S. solidus</i>	?	?	61?, 71?, 81?
	<i>S. stejnegeri</i>	?	?	67?, 77?, 87?
	<i>S. stellulatus</i>	?	?	61?, 71?, 81?
Systemapora	<i>S. ornata</i>	285	725	71, 81

**APPENDIX 3****Subclass Octocorallia (Order Gorgonacea)**

Genus	Species	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<b>Family Chrysogorgiidae</b>				
<i>Iridogorgia</i>	<i>I. bella</i>	748	1 925	77
	<i>I. fontinalis</i>	?	?	31?
	<i>I. magnispiralis</i>	2 143	2 248	31
	<i>I. pourtalesii</i>	1 633	1 633	31, 37
	<i>I. splendens</i>	1 986	1 986	21
<i>Metallogorgia</i>	<i>M. macrospina</i>	?	?	71
	<i>M. melanotrichos</i>	235	2 050	27, 34, 71?, 77, 81
	<i>M. splendens</i>	?	?	
	<i>M. tenuis</i>	490	490	61
<b>Family Coralliidae</b>				
<i>Corallium</i>	<i>C. abyssale</i>	1 829	2 403	77
	<i>C. borneanense</i>	?	?	71
	<i>C. boshuense</i>	670	715	77
	<i>C. ducale</i>	40?	2 000	77
	<i>C. elatius</i>	?	?	61
	<i>C. halmahirensense</i>	?	?	
	<i>C. imperiale</i>	0	2 000	77
	<i>C. johnsoni</i>	3 150	3 150	27
	<i>C. konojoi</i>	?	?	61
	<i>C. maderense</i>	?	?	34?
	<i>C. medea</i>	522	1 426	31, 41
	<i>C. niobe</i>	659	1 646	21, 31
	<i>C. niveum</i>	200	546	77
	<i>C. porcellanum</i>	670	715	77
	<i>C. pusillum</i>	?	?	
	<i>C. regale</i>	68	1 864	61, 67, 77
	<i>C. reginae</i>	1 210	1 260	71
	<i>C. rubrum</i>	?	?	37
	<i>C. secundum</i>	124	1 100	61, 77
	<i>C. sulcatum</i>	800	800	61
	<i>C. tricolor</i>	?	?	
	<i>C. vanderbilti</i>	?	?	
	<i>C. variabile</i>	?	?	
<i>Paracorallium</i>	<i>P. inutile</i>	?	?	
	<i>P. japonicum</i>	?	?	
	<i>P. nix</i>	?	?	
	<i>P. salomonense</i>	?	?	
	<i>P. stylasteroides</i>	?	?	
	<i>P. thrinax</i>	?	?	
	<i>P. tortuosum</i>	0	678	61, 77
<b>Family Isididae</b>				
<i>Acanella</i>	<i>A. africana</i>	>1 000	>1 000	71?
	<i>A. arbuscula</i>	110	4 550	21, 27, 31, 34, 41
	<i>A. chiliensis</i>	324	324	87
	<i>A. dispar</i>	216	1 743	77
	<i>A. eburnea</i>	254	2 881	21, 31, 41, 77

Genus	Species	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>A. furcata</i>	?	?	
	<i>A. microspiculata</i>	?	?	
	<i>A. robusta</i>	>1 000?	>1 000?	57
	<i>A. sibogae</i>	244	1 570	61, 71, 81
	<i>A. verticillata</i>	>1 000?	>1 000?	71
	<i>A. weberi</i>	450	1 767?	71, 77?
<i>Isidella</i>	<i>I. elongata</i>	320	1 582	27?, 31?, 34?, 37
	<i>I. lofotensis</i>	123	>1 000	27
	<i>I. Longiflora</i>	1 700	1 700	27, 41
	<i>I. trichotoma</i>	370	6 300	77
<i>Keratoisis</i>	<i>K. chuni</i>	>1 000?	>1 000?	67
	<i>K. flabellum</i>	415	1 745	77
	<i>K. flexibilis</i>	170	878	21?, 31, 37?, 81
	<i>K. glaesia</i>	406	406	81
	<i>K. gracilis</i>	>50?	>50?	57
	<i>K. grandiflora</i>	384	1 116	71
	<i>K. grandis</i>	1 264	1 563	77?
	<i>K. grayi</i>	1 560	1 560	27, 31?, 37
	<i>K. hikurangiensis</i>	834	951	81
	<i>K. japonica</i>	550	550	61
	<i>K. macrospiculata</i>	1 694	1 694	34
	<i>K. microspiculata</i>	?	?	
	<i>K. palmae</i>	2 082	2 082	34
	<i>K. paucispinosa</i>	348	539	61, 77
	<i>K. philippinensis</i>	567	805	61, 71
	<i>K. profunda</i>	3 230	3 230	67
	<i>K. projecta</i>	743	743	81
	<i>K. siemensii</i>	3 259	3 259	21?, 27?
	<i>K. squatiosa</i>	?	?	
	<i>K. squarrosa</i>	?	?	61
	<i>K. tangentis</i>	406	406	81
	<i>K. wrighti</i>	655	655	71
	<i>K. zelandica</i>	913	1 003	81
<b>Family Paragorgiidae</b>				
<i>Paragorgia</i>	<i>P. alisonae</i>	935	1 180	61, 81
	<i>P. aotearoa</i>	700	900	67, 81
	<i>P. arborea</i>	90	1 798	21, 27, 61, 67, 81
	<i>P. coralloides</i>	602	3 000	71, 77, 81
	<i>P. johnsoni</i>	566	608	21, 31, 41
	<i>P. kaupeka</i>	787	949	61, 81
	<i>P. maunga</i>	545	1 121	61, 81
	<i>P. regalis</i>	332	1 743	67, 77
	<i>P. sibogae</i>	522	522	71
	<i>P. splendens</i>	733	925	57
	<i>P. stephencairnsi</i>	350	490	67, 77
	<i>P. tapachtli</i>	1 950	1 950	77
	<i>P. wahine</i>	890	1 000	61, 81
	<i>P. whero</i>	772	951	61, 81
	<i>P. yutlinux</i>	487	861	77
<b>Family Plexauridae</b>				
<i>Alaskagorgia</i>	<i>A. aluetiana</i>	137	746	67

Genus	Species	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
<i>Paramuricea</i>	<i>P. granids</i>	457	2 180	21, 31
<b>Family Primnoidae</b>				
<i>Callogorgia</i>	<i>C. affinis</i>	113	113	71
	<i>C. americana</i>	37	732	31
	<i>C. chariessa</i>	49	90	71
	<i>C. cristata</i>	160	545	61?, 71
	<i>C. dubia</i>	730	730	57?
	<i>C. elegans</i>	143	143	71
	<i>C. flabellum</i>	402	1 250	71, 77
	<i>C. formosa</i>	80	750	71, 77
	<i>C. gilberti</i>	339	1 000	61, 77
	<i>C. gracilis</i>	51	718	31
	<i>C. grimaldii</i>	?	?	31?
	<i>C. indica</i>	82	490	57
	<i>C. joubini</i>	520	520	71
	<i>C. kinoshitae</i>	220	2 472	77
	<i>C. laevis</i>	?	?	57?
	<i>C. linguimaris</i>	501	1 116	31
	<i>C. minuta</i>	90	90	71
	<i>C. modesta</i>	1 075	1 075	71
	<i>C. pennacea</i>	204	204	71
	<i>C. ramosa</i>	?	?	61
	<i>C. robusta</i>	520	1 301	71
	<i>C. sertosa</i>	256	256	71
	<i>C. similis</i>	90	90	71
	<i>C. ventilabrum</i>	162	162	81
	<i>C. versluysi</i>	203	203	71
	<i>C. verticillata</i>	37	2 472	27, 34, 37
<i>Calyptrophora</i>	<i>C. agassizii</i>	372	1 545	61, 77, 87
	<i>C. angularis</i>	417	1 723	77
	<i>C. antilla</i>	1 110	1 763	21, 31, 77
	<i>C. clarki</i>	216	2 672	61, 67, 77
	<i>C. clinata</i>	1 315	1 842	21
	<i>C. gerdae</i>	229	556	31
	<i>C. japonica</i>	205	2 794	61, 67, 71, 77
	<i>C. juliae</i>	394	729	71
	<i>C. laevispinosa</i>	3 107	3 107	67
	<i>C. microdentata</i>	686	2 247	21, 31
	<i>C. spinosa</i>	216	1 575	71
	<i>C. trilepis</i>	324	911	31
	<i>C. wyvillei</i>	380	1 370	77, 81
<i>Narella</i>	<i>N. abyssalis</i>	4 594	4 594	67
	<i>N. alaskensis</i>	2 377	3 075	67
	<i>N. alata</i>	216	750	77
	<i>N. allmani</i>	622	622	71
	<i>N. alvinae</i>	2 172	3 519	21, 31
	<i>N. ambigua</i>	717	1 463	87
	<i>N. bayeri</i>	3 277	4 091	67
	<i>N. bellissima</i>	439	841	31
	<i>N. biannulata</i>	?	?	61
	<i>N. bowersi</i>	800	2 600	67, 77
	<i>N. clavata</i>	296	296	71

Genus	Species	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>N. compressa</i>	?	?	61
	<i>N. dichotoma</i>	670	1 448	61?, 77
	<i>N. gaussi</i>	2 022	2 897	48
	<i>N. gigas</i>	344	564	77
	<i>N. gilchristi</i>	100	340	51
	<i>N. grandiflora</i>	235	245	61
	<i>N. hawaiiensis</i>	400	1 977	77
	<i>N. horrida</i>	204	204	71
	<i>N. irregularis</i>	188	188	61
	<i>N. japonensis</i>	731	731	61
	<i>N. laxa</i>	3 186	3 186	21
	<i>N. leilae</i>	159	732	71
	<i>N. macrocalyx</i>	1 206	1 807	77
	<i>N. megalepis</i>	224	1 609	57, 61, 71
	<i>N. musikae</i>	215	954	77
	<i>N. obscura</i>	984	984	71
	<i>N. orientalis</i>	235	732	71
	<i>N. ornata</i>	409	1 006	77
	<i>N. parva</i>	1 300	1 633	71
	<i>N. pauciflora</i>	738	1 510	31
	<i>N. regularis</i>	729	729	31
	<i>N. spectabilis</i>	1 485	1 485	31
	<i>N. studeri</i>	1 080	1 264	71, 81
	<i>N. versluysi</i>	659	3 100	21, 27, 31
<i>Paracalyptrophora</i>	<i>P. carinata</i>	514	525	31
	<i>P. duplex</i>	49	575	31
	<i>P. josephinae</i>	201	330	27, 31, 34
	<i>P. kerberti</i>	552	552	61
	<i>P. mariae</i>	520	520	61?, 71
	<i>P. simplex</i>	18	531	31
<i>Plumarella</i>	<i>P. aculeata</i>	350	586	31
	<i>P. acuminata</i>	330	330	61
	<i>P. adhaerans</i>	180?	180?	61
	<i>P. alba</i>	550	550	61
	<i>P. aurea</i>	388	878	31
	<i>P. delicatissima</i>	256	256	41?
	<i>P. dentata</i>	229	494	41, 51?
	<i>P. dichotoma</i>	494	1 064	31
	<i>P. dofleinii</i>	80	250	61
	<i>P. flabellata</i>	188	247	61
	<i>P. gracilis</i>	?	?	61
	<i>P. laevis</i>	90	115	71?
	<i>P. lata</i>	62	68	61
	<i>P. laxiramosa</i>	347	878	31
	<i>P. longispina</i>	198	282	67, 77
	<i>P. pellucida</i>	10	1 200	31
	<i>P. penna</i>	8	10	57
	<i>P. pourtalesii</i>	45	1 435	31
	<i>P. rigida</i>	180	600	61
	<i>P. spicata</i>	779	1 963	61
	<i>P. spinosa</i>	881	881	61
<i>Primnoa</i>	<i>P. notialis</i>	549	1 153	

Genus	Species	Depth range (m)		FAO Major Fishing Areas
		Min	Max	
	<i>P. pacifica</i>	9	800	61, 67, 77
	<i>P. resedaeformis</i>	95	1 020	21, 27, 34?
	<i>P. wingi</i>	110	914	61, 67
<i>Thouarella</i>	<i>T. abies</i>	100	550	48
	<i>T. affinis</i>	106	128	47
	<i>T. alternata</i>	34	138	61
	<i>T. antarctica</i>	118	118	41
	<i>T. bipinnata</i>	82	1 446	31
	<i>T. brucei</i>	102	183	47
	<i>T. chilensis</i>	73	130	41, 48, 87?
	<i>T. clavata</i>	500	500	41?, 47?
	<i>T. coronata</i>	148	148	61?
	<i>T. crenelata</i>	133	686	48
	<i>T. diadema</i>	1 000	1 000	41
	<i>T. grasshoffi</i>	814	1 805	21, 27, 34
	<i>T. hicksoni</i>	110	360	51
	<i>T. hilgendorfi</i>	174	1 760	34, 61, 71, 77
	<i>T. koellikeri</i>	320	730	87
	<i>T. laxa</i>	400	1 644	51, 57, 61, 71
	<i>T. longispinosa</i>	283	284	48
	<i>T. moseleyi</i>	794	1 080	71, 81
	<i>T. parva</i>	?	?	61
	<i>T. pendulina</i>	?	?	
	<i>T. recta</i>	869	924	61
	<i>T. striata</i>	384	642	67
	<i>T. superba</i>	40	384	61, 67
	<i>T. tydemanii</i>	?	?	71?
	<i>T. typica</i>	174	388	61, 77
	<i>T. variabilis</i>	183	366	58
<i>Stylaster</i>	<i>S. hattori</i>	?	?	61?, 71?, 81?
	<i>S. incompletus</i>	?	?	61?, 71?, 81?
	<i>S. incrassatus</i> (not valid?)	?	?	61?, 71?, 81?
	<i>S. microstriatus</i>	?	?	61?, 71?, 81?
	<i>S. nobilis</i>	?	?	47
	<i>S. polymorphus</i>	?	?	
	<i>S. profundus</i>	?	?	21?, 31?, 41?
	<i>S. profundiporus</i>	?	?	61?, 71?, 81?
	<i>S. pulcher</i>	?	?	61?, 71?, 81?
	<i>S. purpuratus</i>	?	?	61?, 71?, 81?
	<i>S. ramosus</i>	?	?	51?, 57?
	<i>S. scabiosus</i>	?	?	61?, 71?, 81?
	<i>S. solidus</i>	?	?	61?, 71?, 81?
	<i>S. stejnegeri</i>	?	?	61?, 71?, 81?
	<i>S. stellulatus</i>	?	?	61?, 71?, 81?

**APPENDIX 4****Order Anthoathecata (Suborder Filifera)**

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
<b>Family Hydractiniidae</b>				
<i>Hydrocorella</i>	<i>H. africana</i>	18	57	47
<i>Janaria</i>	<i>J. mirabilis</i>	7	137	71, 77
<i>Polyhydra</i>	<i>P. calcarea</i>	?	?	27?
<b>Family Stylasteridae</b>				
<i>Adelopora</i>	<i>A. crassilabrum</i>	282	1 169	71, 81
	<i>A. moseleyi</i>	291	360	77, 81
	<i>A. pseudothyron</i>	298	915	41, 81, 87
	<i>A. fragilis</i>	400	1 570	71, 81
<i>Astya</i>	<i>A. aspidopora</i>	590	1 058	81
	<i>A. subviridis</i>	914	914	71
<i>Calyptopora</i>	<i>C. reticulata</i>	93	2 010	81
	<i>C. sinuosa</i>	142	762	71, 81
<i>Cheiloporidion</i>	<i>C. pulvinatum</i>	642	1 137	41
<i>Conopora</i>	<i>C. laevis</i>	130	1 052	51?, 61, 71, 81
	<i>C. verrucosa</i>	198	2 626	81, 88
	<i>C. candelabrum</i>	240	1 265	71, 81
	<i>C. tetrastichopora</i>	282	850	71, 81
	<i>C. gigantea</i>	356	544	81
	<i>A. adeta</i>	398	399	71
	<i>C. anthohelia</i>	440	1 175	81
	<i>C. unifacialis</i>	547	547	81
	<i>C. dura</i>	?	?	51?, 57?
<i>Cryptphelia</i>	<i>C. peircei</i>	89	838	31
	<i>C. micropoma</i>	140	140	51
	<i>C. studeri</i>	142	1 058	81
	<i>C. trophostega</i>	150	1 913	61, 67
	<i>C. insolita</i>	159	720	31
	<i>C. papillosa</i>	161	545	31
	<i>C. glebulenta</i>	166	806	87
	<i>C. cymas</i>	169	1 097	81, 87
	<i>C. cryptotrema</i>	170	474	71
	<i>C. glossopoma</i>	198	864	31
	<i>C. japonica</i>	247	551	61
	<i>C. lacunosa</i>	252	806	87
	<i>C. curvata</i>	287	1 058	81
	<i>C. polypoma</i>	290	966	71, 81
	<i>C. dactylopoma</i>	348	813	87
	<i>C. pudica</i>	368	1 633	57?, 61, 71, 81
	<i>C. robusta</i>	374	709	71, 81
	<i>C. vascomarquesi</i>	390	1 520	27, 34
	<i>C. formosa</i>	419	2 044	41
	<i>C. platypoma</i>	469	469	71
	<i>C. gigantea</i>	475	784	87

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
	<i>C. eueides</i>	475	813	87
	<i>C. floridana</i>	593	823	31
	<i>C. fragilis</i>	647	2 328	57, 71, 81
	<i>C. affinis</i>	712	2 790?	27
	<i>C. tenuisepta</i>	761	1 557	27, 31, 34
	<i>C. medioatlantica</i>	861	2 644	27
	<i>C. ramosa</i>	1 165	1 264	71
	<i>C. balia</i>	1 633	1 633	71
	<i>C. clausa</i>	?	?	51?, 57?
	<i>C. stenopoma</i>	?	?	71?
	<i>C. lamellata</i>	83	550	67
<i>Distichopora</i>	<i>D. violacea</i>	0	122	34, 47, 51
	<i>D. irregularis</i>	6	46	31
	<i>D. borealis</i>	10	881	61, 67, 71
	<i>D. vervoorti</i>	15	20	71
	<i>D. nitida</i>	22	22	71
	<i>D. yucatanensis</i>	39	261	31
	<i>D. sulcata</i>	60	708	31
	<i>D. cervina</i>	68	384	31
	<i>D. barbadensis</i>	108	159	31
	<i>D. contorta</i>	125	368	31
	<i>D. anomala</i>	139	311	31
	<i>D. rosalindae</i>	165	198	31
	<i>D. laevigranulosa</i>	166	806	87
	<i>D. foliacea</i>	174	527	31
	<i>D. asulcata</i>	293	377	77
	<i>D. uniserialis</i>	333	366	31
	<i>D. anceps</i>	360	736	77
	<i>D. dispar</i>	465	741	81
	<i>D. coccinea</i>	?	?	61?, 71?, 81?
	<i>D. gracilis</i>	?	?	61?, 71?, 81?
	<i>D. livida</i>	?	?	61?, 71?, 81?
	<i>D. profunda</i>	?	?	51?, 57?
	<i>D. providentiae</i>	?	?	51?, 57?
	<i>D. serpens</i>	?	?	51?, 57?
<i>Errina</i>	<i>E. antarctica</i>	25	903	41, 87
	<i>E. novaezelandiae</i>	30	117	81
	<i>E. dendyi</i>	30	383	81
	<i>E. hicksoni</i>	37	157	81
	<i>E. fissurata</i>	46	1 775	48, 88
	<i>E. reticulata</i>	79	128	81
	<i>E. laterorifa</i>	79	2 273	48, 58, 81, 88
	<i>E. aspera</i>	80	200	27?, 34, 37
	<i>E. japonica</i>	85	85	61
	<i>E. kerguelensis</i>	91	567	81, 88
	<i>E. boschmai</i>	100	686	41, 48
	<i>E. chathamensis</i>	106	688	81
	<i>E. sinuosa</i>	135	1 075	47, 81

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
	<i>E. dabneyi</i>	140	2 200	27
	<i>E. gracilis</i>	146	1 226	41, 48, 81, 87, 88
	<i>E. cochleata</i>	194	534	31
	<i>E. altispina</i>	198	309	31
	<i>E. cheilopora</i>	198	659	81
	<i>E. bicolor</i>	252	549	81
	<i>E. cooki</i>	256	380	81
	<i>E. laevigata</i>	333	371	81
	<i>E. macrogastera</i>	549	704	77, 87
	<i>E. atlantica</i>	610	983	27
	<i>E. cyclopora</i>	1 647	2 044	41
	<i>E. capensis</i>	?	?	27?, 34?, 47?
	<i>E. porifera</i>	?	?	61?, 71?, 81?
<i>Errinopora</i>	<i>E. pourtalesi</i>	49	183	77
	<i>E. nanneca</i>	63	375	61, 67
	<i>E. stylifera</i>	108	117	61
	<i>E. latifundata</i>	109	109	61
	<i>E. zarhyncha</i>	207	658	67
	<i>E. cestoporina</i>	339	1 240	41
	<i>E. fenestrata</i>	152	622	41, 87
	<i>E. reticulum</i>	250	771	41
<i>Gyropora</i>	<i>G. africana</i>	22	22	47
<i>Inferiolabiata</i>	<i>I. lowei</i>	164	991	41, 48, 87
	<i>I. spinosa</i>	211	781	81
	<i>I. labiata</i>	238	274	51
<i>Lepidopora</i>	<i>L. carinata</i>	60	494	31
	<i>L. clavigera</i>	130	282	31
	<i>L. glabra</i>	183	1 170	21, 31, 41
	<i>L. biserialis</i>	196	370	31
	<i>L. cryptocymas</i>	197	506	81
	<i>L. polystichopora</i>	197	710	81
	<i>L. decipiens</i>	276	670	31
	<i>L. symmetrica</i>	282	1 058	81
	<i>L. dendrostylus</i>	345	975	81
	<i>L. microstylus</i>	405	1 258	71?, 81
	<i>L. eburnea</i>	460	983	27
	<i>L. sarmentosa</i>	502	1 665	71, 81, 88
	<i>L. acrolophos</i>	659	686	41
	<i>L. concatenata</i>	680	720	87
	<i>L. diffusa</i>	?	?	27?, 34?, 47?
	<i>L. granulosa</i>	?	?	48?, 58?, 88?
<i>Lepidothecea</i>	<i>L. cervicornis</i>	110	110	81
	<i>L. pourtalesi</i>	123	368	31
	<i>L. chauliostylus</i>	130	1 265	71, 81
	<i>L. macropora</i>	169	806	87
	<i>L. tenuistylus</i>	238	274	51
	<i>L. robusta</i>	365	365	81
	<i>L. ramosa</i>	520	520	71

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
	<i>L. fascicularis</i>	540	2 010	88
	<i>L. brochi</i>	545	864	31
	<i>L. altispina</i>	660	1 570	81
	<i>L. inconsuta</i>	787	1 500	81
	<i>L. horrida</i>	1 089	1 089	71
<i>Paraerrina</i>	<i>P. diciiens</i>	238	274	51
<i>Phalangopora</i>	<i>P. regularis</i>	238	274	51
<i>Pliobonthrus</i>	<i>P. symmetricus</i>	80	1 600	27, 31, 34
	<i>P. echinatus</i>	165	750	31
	<i>P. tubulatus</i>	419	708	31
	<i>P. fistulosus</i>	546	631	77
	<i>P. gracilis</i>	600	700	34
<i>Pseudocryptethelia</i>	<i>P. pachypoma</i>	430	1 089	71
<i>Sporadopora</i>	<i>S. dichotoma</i>	250	1 498	41, 48
	<i>S. martensi</i>	282	282	81
	<i>S. macropora</i>	465	741	81
<i>Stellapora</i>	<i>S. echinata</i>	215	1 846	41
<i>Stenohelia</i>	<i>S. maderensis</i>	110	1 125	27, 34
	<i>S. profunda</i>	159	2 021	31
	<i>S. concinna</i>	166	806	87
	<i>S. pauciseptata</i>	282	514	31
	<i>S. conferta</i>	?	?	48?, 58?, 88?
	<i>S. echinata</i>	?	?	61?, 71?, 81?
	<i>S. tiliata</i>	?	?	61?, 71?, 81?
	<i>S. umbonata</i>	?	?	61?, 71?, 81?
	<i>S. yabei</i>	?	?	61?, 71?, 81?
<i>Stephanohelia</i>	<i>S. praecipua</i>	318	1 277	71, 81
<i>Stylanthesca</i>	<i>S. prophrya</i>	0	1	77
	<i>S. petrograpta</i>	0	15	67
	<i>S. papillosa</i>	?	?	67?, 77?, 87?
<i>Stylaster</i>	<i>S. marshae</i>	0	13	57
	<i>S. tenisonwoodsi</i>	1	90	57, 71
	<i>S. roseus</i>	2	494	31, 41
	<i>S. sanguineus</i>	3	20	71, 77
	<i>S. papuensis</i>	6	20	71
	<i>S. blatteus</i>	10	10	34
	<i>S. californicus</i>	12	91	67, 77
	<i>S. asper</i>	13	133	71
	<i>S. corallium</i>	13	298	31
	<i>S. laevigatus</i>	20	1 170	31
	<i>S. subviolacea</i>	22	22	47
	<i>S. campyleucus</i>	23	518	61, 67
	<i>S. eguchii</i>	24	1 520	71, 81
	<i>S. venustus</i>	27	108	67, 77
	<i>S. lonchitis</i>	30	113	51
	<i>S. rosaceus</i>	37	37	34
	<i>S. gemmascens</i>	40	665	21, 27
	<i>S. eximius</i> (valid?)	40	823	31, 71

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
	<i>S. duchassaingi</i>	40	915	31, 41?
	<i>S. divergens</i>	55	549	87
	<i>S. brochi</i>	62	582	61, 67
	<i>S. bithalamus</i>	71	71	47
	<i>S. norvegicus</i>	75	1 400	27
	<i>S. brunneus</i>	80	290	71, 81?
	<i>S. marenzelleri</i>	84	631	77, 87
	<i>S. verrillii</i>	87	366	67
	<i>S. polyorchis</i>	91	823	67
	<i>S. densicaulis</i>	96	1 362	41, 48
	<i>S. boreopacificus</i>	117	183	61
	<i>S. aurantiacus</i>	123	355	31
	<i>S. moseleyanus</i>	126	518	67
	<i>S. imbricatus</i>	128	1 037	81
	<i>S. miniatus</i>	146	530	31
	<i>S. erubescens</i>	146	1 440	21, 27, 31
	<i>S. cancellatus</i>	152	582	61, 67
	<i>S. antillarum</i>	174	653	31
	<i>S. filogranus</i>	183	274	31
	<i>S. complanatus</i>	183	707	31
	<i>S. inornatus</i>	198	309	31
	<i>S. elassotomus</i>	210	882	61
	<i>S. horologium</i>	228	1 169	71, 81
	<i>S. galapagensis</i>	252	806	77, 87
	<i>S. gracilis</i>	291	291	81
	<i>S. cocosensis</i>	293	576	77
	<i>S. robustus</i>	300	300	48
	<i>S. griggi</i>	322	583	77
	<i>S. multiplex</i>	372	450	71
	<i>S. spatula</i>	384	549	31
	<i>S. ibericus</i>	500	500	27
	<i>S. bilobatus</i>	520	520	57
	<i>S. infundibuliferus</i>	521	563	77
	<i>S. maroccanus</i>	1 378	1 378	34
	<i>S. alaskanus</i>	?	?	61?, 67?
	<i>S. amphiheloides</i>	?	?	47, 48?
	<i>S. bellus</i>	?	?	61?, 71?, 81?
	<i>S. bocki</i>	95	95	61?, 71?, 81?
	<i>S. boschmai</i>	?	?	61?, 71?, 81?
	<i>S. carinatus</i>	?	?	61?, 71?, 81?
	<i>S. crassior</i>	?	?	51?, 57?
	<i>S. dentatus</i>	?	?	61?, 71?, 81?
	<i>S. flabelliformis</i>	?	?	51
	<i>S. granulosus</i>	?	?	61?, 71?, 81?
	<i>S. hattorii</i>	?	?	61?, 71?, 81?
	<i>S. incompletus</i>	?	?	61?, 71?, 81?
	<i>S. incrassitus</i> (not valid?)	?	?	61?, 71?, 81?
	<i>S. microstriatus</i>	?	?	61?, 71?, 81?

Genus	Species (* facultative species)	Depth range (m)		FAO Major Fishing Areas
		Max	Min	
	<i>S. nobilis</i>	?	?	47
	<i>S. polymorphus</i>	?	?	
	<i>S. profundus</i>	?	?	21?, 31?, 41?
	<i>S. profundiporus</i>	?	?	61?, 71?, 81?
	<i>S. pulcher</i>	?	?	61?, 71?, 81?
	<i>S. purpuratus</i>	?	?	61?, 71?, 81?
	<i>S. ramosus</i>	?	?	51?, 57?
	<i>S. scabiosus</i>	?	?	61?, 71?, 81?
	<i>S. solidus</i>	?	?	61?, 71?, 81?
	<i>S. stejnegeri</i>	?	?	67?, 77?, 87?
	<i>S. stellulatus</i>	?	?	61?, 71?, 81?
<i>Systemapora</i>	<i>S. ornata</i>	285	725	71, 81

## Towards the development of an identification guide of vulnerable deep-sea sponges

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### Overview

Following the FAO terms of reference, a list of the global distribution (in relation to the 19 FAO Major Fishing Areas) of the most relevant deep-sea sponge groups: the entire class Hexactinellida; orders Lithistida and Astrophorida within Demospongiae; some other genera within Demospongiae, was compiled. Also, a relevant bibliography for general taxonomy, historical campaigns, each species group and main international sponge initiatives was assembled. Finally, draft recommendations for the production of identification guides are suggested.

### 1. List of relevant sponge groups

A compilation of a global species list (for FAO Fishing Areas) of the most relevant deep-sea sponge groups: the entire class Hexactinellida; orders Lithistida and Astrophorida within Demospongiae; some other genera within the Orders Poecilosclerida, Halichondrida and Haplosclerida (Demospongiae), is presented in Table 1. These are the groups that are most likely to be impacted by fisheries.

**Table 1**  
**Global species list for FAO Major Fishing Areas**

Class	Subclass	Order	Family
Hexactinellida Schmidt, 1870	Amphidiscophora Schulze, 1886	Amphidiscosida Schrammen, 1924	<ul style="list-style-type: none"> <li>• Hyalonematidae Gray, 1857</li> <li>• Monorhaphididae Ijima, 1927</li> <li>• Pheronematidae Gray, 1870</li> </ul>
	Hexasterophora Schulze, 1886	Aulocalycoida Tabachnick and Reiswig, 2000	<ul style="list-style-type: none"> <li>• Aulocalycidae Ijima, 1927</li> <li>• Uncinateridae Reiswig, 2002</li> </ul>
		Fieldingida Tabachnick and Janussen, 2004	<ul style="list-style-type: none"> <li>• Fieldingiidae Tabachnick and Janussen, 2004</li> </ul>
		Hexactinosida Schrammen, 1912	<ul style="list-style-type: none"> <li>• Aphrocallistidae Gray, 1867</li> <li>• Craticulariidae Rauff, 1893</li> <li>• Cribrospongiidae Roemer, 1864</li> <li>• Dactylocalycidae Gray, 1867</li> <li>• Euretidae Zittel, 1877</li> <li>• Farreidae Gray, 1872</li> <li>• Tretodictyidae Schulze, 1886</li> </ul>
		Lychniscosida Schrammen, 1903	<ul style="list-style-type: none"> <li>• Aulocystidae Sollas, 1887</li> <li>• Diapleuridae Ijima, 1927</li> </ul>
		Lyssacinosida Zittel, 1877	<ul style="list-style-type: none"> <li>• Euplectellidae Gray, 1867</li> <li>• Leucopsacidae Ijima, 1903</li> </ul>
		Rossellidae Schulze, 1885	
Demospongiae Sollas, 1885		Astrophorida Sollas, 1887	<ul style="list-style-type: none"> <li>• Ancorinidae Schmidt, 1870</li> <li>• Calthropellidae Lendenfeld, 1907</li> <li>• Geodiidae Gray, 1867</li> <li>• Pachastrellidae Carter, 1875</li> <li>• Thrombidae Sollas, 1888</li> </ul>

Class	Subclass	Order	Family
		Hadromerida Topsent, 1894	<ul style="list-style-type: none"> <li>● Stylocordylidae Topsent, 1892</li> <li>● Suberitidae Schmidt, 1870</li> <li>● Tethyidae Gray, 1848</li> </ul>
		Lithistida Schmidt, 1870	<ul style="list-style-type: none"> <li>● Scleritodermidae Sollas, 1888</li> <li>● Corallistidae Sollas, 1888</li> <li>● Pleromidae Sollas, 1888</li> <li>● Theonellidae Lendenfeld, 1903</li> <li>● Siphonidiidae Lendenfeld, 1903</li> <li>● Neopeltidae Sollas, 1888</li> <li>● Azoricidae Sollas, 1888</li> <li>● Desmathidae Topsent, 1894</li> <li>● Vetulinidae Lendenfeld, 1903</li> <li>● Phymatellidae Schrammen, 1910</li> <li>● Isoraphiniidae Schrammen, 1924</li> <li>● Macandrewiidae Schrammen, 1924</li> <li>● Phymaraphiniidae Schrammen, 1924</li> </ul>
		Poecilosclerida Topsent, 1928	<ul style="list-style-type: none"> <li>● Cladorhizidae Dendy, 1922</li> <li>● Acarnidae Dendy, 1922</li> <li>● Esperiopsidae Hentschel, 1923</li> <li>● Podospongidae de Laubenfels, 1936</li> <li>● Isodictyidae Dendy, 1924</li> </ul>
		Halichondrida Gray, 1867	<ul style="list-style-type: none"> <li>● Axinellidae Carter, 1875</li> <li>● Halichondriidae Gray, 1867</li> </ul>
		Haplosclerida Topsent, 1928	<ul style="list-style-type: none"> <li>● Chalinidae Gray, 1867</li> <li>● Phloeodictyidae Carter, 1882</li> <li>● Petrosiidae Van Soest, 1980</li> </ul>

A summary of the diversity (number of species) of each selected sponge group per FAO area<sup>1</sup> is presented in Table 2. See Appendixes 1, 2, 3, 4, 5, 6 and 7 for details on species and their deep-sea depth occurrence.

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<sup>1</sup> Area 18: Arctic Sea; Area 21: Atlantic, Northwest; Area 27: Atlantic, Northeast, Area 31: Atlantic, Western Central; Area 34: Atlantic, Eastern Central; Area 37: Mediterranean and Black Sea; Area 41: Atlantic, Southwest; Area 47: Atlantic, Southeast; Area 48: Atlantic, Antarctic; Area 51, Indian Ocean, Western; Area 57: Indian Ocean, Eastern; Area 58: Indian Ocean, Antarctic and Southern; Area 61: Pacific, Northwest; Area 67: Pacific, Northeast; Area 71: Pacific, Western Central; Area 77: Pacific, Eastern Central; Area 81: Pacific, Southwest; Area 87: Pacific, Southeast; Area 88: Pacific, Antarctic.

**Table 2**  
Diversity (number of species) of each selected sponge group per FAO area<sup>1</sup>

	Poles		Atlantic					Med	Indian			Pacific						
	48, 88	18	21	27	31	34	41	47	37	51	57	58	61	67	71	77	81	87
Hexactinellida	59	9	18	29	44	2	13	4	3	28	35	7	83	33	181	53	21	7
Astrophorida	6	3	3	60	40	28	25	38	43	37	34	8	67	4	73	37	25	10
Hadromerida	4	5	1	8	5	0	2	1	3	2	1	2	10	1	3	2	2	1
Lithistida	1	0	0	11	38	10	5	7	9	33	10	1	10	0	57	2	32	1
Poecilosclerida	37	18	11	32	6	6	9	10	3	4	4	16	23	8	18	10	10	8
Halichondrida	1	10	3	17	6	0	2	0	3	1	1	0	6	2	3	3	1	0
Haplosclerida	3	1	3	19	7	0	0	1	2	0	1	0	0	1	6	1	4	0

<sup>1</sup> Here, the Antarctic is considered as one region, ANT, and not as two different regions, i.e., 48 and 88.

## 2. Historical campaigns

A list of the main historical campaigns that yielded extensive reports on deep-sea sponge fauna is presented in Table 3.

**Table 3**  
Main historical campaigns on deep-sea sponge fauna

Expedition	Year	Area	Authors
Porcupine	1869 and 1870	Northeast Atlantic	Carter. 1874, 1876.
Norma	1870	Spain	Kent. 1870 a, b.
Challenger	1872 to 1876	World	Sollas. 1888; Ridley & Dendy. 1887; Polejaeff. 1883, 1884; Schulze. 1887.
Blake	1877-1879	Caribbean	Schmidt. 1879, 1880.
Vega	1878-1880	Arctic	Fristedt. 1887.
Willem Barents	1878-1884	Arctic	Vosmaer. 1882, 1885.
Alert	1880-1881	Patagonia, Australia, Seychelles	Ridley. 1881, 1884
Albert Monaco (Hirondelle)	1888	Northeast Atlantic, Mediterranean	Topsent. 1892.
Caudan	1895	Gulf of Biscay	Topsent. 1896.
Ingolf	1895-96	North Atlantic, Arctic	Lundbeck. 1902, 1905, 1910; Burton. 1928.
Valdivia	1898-1899	World	Lendenfeld. 1907. Schulze. 1904.
Belgian Antarctic	1897 to 1899	Antarctic	Topsent. 1902.
Siboga	1899-1900	Indonesia	Ijima. 1927; Burton. 1939; Vosmaer. 1911; Vosmaer & Vernhout. 1902.
Albert Monaco (Alice)	1900	Azores	Topsent. 1904.
Investigator	1900	Indian Ocean	Schulze. 1902; Dendy & Burton. 1926; Burton. 1928.
Swedish Antarctic	1901-1903	Antarctic	Burton. 1934a.
Südpolar	1901-03	Antarctic	Hentschel. 1914; Lendenfeld. 1907; Schulze & Kirkpatrick. 1910.
Discovery 1	1901-1904	Antarctic	Kirkpatrick. 1907, 1908; Jenkin. 1908.
1st French Antarctic	1903-05	Antarctic	Topsent. 1908.
Scottish Antarctic	1903-04	South Africa, Antarctic	Topsent. 1915; Stephens, 1915.

Expedition	Year	Area	Authors
Sealark	1905	West Indian Ocean	Dendy. 1913, 1916, 1922.
Albert Monaco (Alice2)	1906-07	North Atlantic	Topsent. 1913.
Denmark E Greenland	1906-07	East Greenland	Brøndsted. 1917.
2nd French Antarctic	1908-10	Antarctic	Topsent. 1917.
Albert Monaco (Alice3)		Northeast Atlantic	Topsent. 1928.
Terra Nova	1910	Antarctic	Dendy. 1924; Burton. 1929.
Michael Sars	1910	Northeast Atlantic	Arnesen. 1920.
Discovery 2	1925-27	Falklands, Antarctic	Burton. 1932.
Albatross 1	1888	North America	Schulze. 1899.
Albatross 2	1891-1905	East Pacific	Wilson. 1904; Lendenfeld. 1910 a, b, 1915.
Albatross 3	1907-10	Philippines	Wilson. 1925; Okada. 1932.
Australasian Antarctic	1911-1914	Antarctic	Dendy. 1918.
Fisheries Ireland	1914	Ireland	Stephens. 1915, 1921.
Norwegian E Greenland	1930-31	East Greenland	Burton. 1934b.
BANZARE	1929-1931	Antarctic	Koltun. 1976.
Allan Hancock	1930-31	East Pacific	Dickinson. 1945.
John Murray	1933-34	Arabian Gulf	Burton. 1959.
Rosaura	1938	Atlantic	Burton. 1954.
Galathea	1950-52	World	Lévi. 1964.
Théodore Tissier	1955-56	Northeast Atlantic	Lévi & Vacelet. 1958.
Soviet Antarctic	1955-1958	Antarctic	Koltun. 1964.
Kuriles-Kamtschatka	1966	North Pacific	Koltun. 1970.
CANCAP I-VII	1981-1987	East Atlantic	de Weerdt & van Soest. 1986.
Vauban	1983	New Caledonia	Lévi & Lévi. 1983 a, b.
Balgim	1984	Mediterranean	Boury-Esnault <i>et al.</i> 1994.
BIOCAL	1985	New Caledonia	Lévi. 1993.
Thalassa	1987	Gulf of Biscay	Borojevic & Boury-Esnault. 1987.
MUSORSTOM	1987	Philippines	Lévi & Lévi. 1989.
MAR-ECO	2002-2004	Mid North Atlantic	Tabachnick & Collins. 2008.
BIOSYS	2005-2007	North Atlantic	van Soest <i>et al.</i> 2007.

### 3. Recommendations for the production of the identification guide

Sponge taxonomy and systematics is mainly based on skeletal structures that require microscopic examination, which greatly limits the number of species that can be included in a field guide. Therefore, and in order to assure maximum geographic and taxonomic coverage, the approach proposed for the sponge guide is to select 15–25 species representing the different deep-sea sponge groups for each FAO area (global total of 300–450 spp.).

Criteria for species selection should include:

- Morphological distinctness – select species that due to macro-morphological characteristics are more likely to be confidently and accurately identified in the field.
- Vulnerability – with exception of the abyssal species (>3 500 m depth) all deep-sea sponge species are potentially vulnerable to fishing. However, larger species or those with more elaborate growth forms (massive, cup-shaped, branching) are more likely to be by-caught in fishing operations. Background knowledge on which species are more commonly represented in the by-catch would also be very useful for this selection.
- Structural role – all species known to form dense aggregations should always be included in the guide.

A Working Group encompassing sponge taxonomists with expertise in each of the main deep-sea sponge groups (orders/families) and focal geographical areas should be built. The working group should meet regularly during workshops organized by FAO in collaboration with its members.

The Northeast Atlantic is one of the areas for which the deep-sea sponge fauna has been most comprehensively studied, both in historic as well as more recent times. This area should therefore be selected as pilot for the production of the first regional guide.

An image bank comprising deep-sea ROV, submersible or drop-down camera images and films as well as illustrations in historical monographs should be assembled to assist the selection of species.

Each species sheet should contain scaled *in situ* (optional) and on-deck (obligatory) pictures as well as a description and line drawings highlighting the main macro-morphological diagnostic characteristics (shape, consistency, texture, size, surface ornamentation). Most species lack common names and therefore some effort could be made in either creating suitable common names or attributing unique codes to each species.

Each species sheet should include a distribution map, habitat and depth range. The distribution map should include both an expert map as well as all point records available, especially the species type locality. For some species it would be possible to build predictive habitat-suitability maps through ecological modelling techniques. This would require extra effort but it would be well worth for the structural species, i.e. species known to form dense aggregations (sponge grounds or reefs).

Some species commonly form a characteristic multispecific assemblage (e.g. *Geodia*, *Stryphnus*, *Isops*) or are associated with a particular substrate type (e.g. *Thenea* spp. are always in soft sediment) and therefore this information should be included in the respective sheets.

For regional guides, organizing species by growth form (e.g. massive spherical, branching, vase-shaped, etc) may be more practical than following the classification system.

Some species have large spicules that protrude the sponge surface and can be harmful or at least painful if not handled with care. A small note should be included in these species' sheets.

Potential limitations:

There is a general lack of information on species geographic/bathymetric distribution, and abundance. Many records are unique for a given species and constitute the species type locality. Except for some regions, the deep-sea sponge fauna is largely unstudied and therefore many of the vulnerable species may constitute new (undescribed) species.

There is a strong bias in both quality and quantity of available information towards the areas that were more comprehensively studied in the past or those that are the focus of more intensive study in recent times (e.g. Northeast Atlantic, Brazil, New Zealand).

Last but not least, as in most taxonomic groups also in Porifera there is a general shortage of taxonomists especially in some deep-sea groups.

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### WebPages of the main international sponge initiatives

- World Porifera Database (WPD): [www.marinespecies.org/porifera](http://www.marinespecies.org/porifera)  
Sponge Barcoding Project (SBP): [www.spongebarcoding.org](http://www.spongebarcoding.org)  
Porifera Tree of Life Project (PorToL): [www.portol.org](http://www.portol.org)

**APPENDIX 1****Hexactinellida**

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
<b>Order Amphidiscosida</b>				
<b>Family Hyalonematidae</b>				
	<i>Hyalonema</i>			
		( <i>Cyliconema</i> )	<i>abyssale</i>	51
		( <i>Leptonema</i> )	<i>acuferum</i>	71
		( <i>Ijimaonema</i> )	<i>aculeatum</i>	57
		( <i>Oonema</i> )	<i>aequatoriale</i>	77
		( <i>Onconema</i> )	<i>agassizi</i>	77
		( <i>Prionema</i> )	<i>agujanum</i>	77
		( <i>Paradisconema</i> )	<i>alcocki</i>	57
		( <i>Cyliconema</i> )	<i>apertum apertum</i>	61
		( <i>Cyliconema</i> )	<i>apertum simplex</i>	61
		( <i>Prionema</i> )	<i>azuerone</i>	77
		( <i>Oonema</i> )	<i>bianchoratum bianchor</i>	77
		( <i>Oonema</i> )	<i>bianchoratum pinulina</i>	77
		( <i>Oonema</i> )	<i>bipinnulum</i>	81
		( <i>Phialonemiella</i> )	<i>brevancora</i>	77
		( <i>Corynonema</i> )	<i>calix</i>	57
		( <i>Leptonema</i> )	<i>campanula campanula</i>	77
		( <i>Leptonema</i> )	<i>campanula longispicula</i>	47
		( <i>Ijimaonema</i> )	<i>cebuense</i>	71
		( <i>Leptonema</i> )	<i>choaniferum</i>	51
		( <i>Corynonema</i> )	<i>clathratum</i>	61
		( <i>Ijimaonema</i> )	<i>clavigerum</i>	58
		( <i>Cyliconema</i> )	<i>coniforme</i>	51
		( <i>Coscinonema</i> )	<i>conus</i>	81
		( <i>Oonema</i> )	<i>crassipinulum</i>	77
		( <i>Prionema</i> )	<i>crassum</i>	77
		( <i>Corynonema</i> )	<i>cupressiferum</i>	77
		( <i>Cyliconema</i> )	<i>curvisclera</i>	51
		( <i>Oonema</i> )	<i>densem</i>	77
		( <i>Corynonema</i> )	<i>depressum</i>	77
		( <i>Leptonema</i> )	<i>divergens</i>	71
		( <i>Cyliconema</i> )	<i>drygalskii</i>	(48, 88)
		( <i>Coscinonema</i> )	<i>elegans</i>	71
		( <i>Cyliconema</i> )	<i>eupinnulum</i>	51
		( <i>Prionema</i> )	<i>fimbriatum</i>	77
		( <i>Leptonema</i> )	<i>flagelliferum</i>	71
		( <i>Oonema</i> )	<i>geminatum</i>	77
	<i>Hyalonema</i>	( <i>Cyliconema</i> )	<i>globiferum</i>	57

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
		( <i>Ijimaonema</i> )	<i>globus</i>	71
		( <i>Coscinonema</i> )	<i>gracile</i>	71
		( <i>Corynonema</i> )	<i>grandancora</i>	77
		( <i>Ijimaonema</i> )	<i>heideri</i>	57
		( <i>Oonema</i> )	<i>henshawi</i>	77
		( <i>Corynonema</i> )	<i>hercules</i>	77
		( <i>Cyliconema</i> )	<i>hozawai</i>	61
		( <i>Coscinonema</i> )	<i>indicum andamanense</i>	57
		( <i>Coscinonema</i> )	<i>indicum indicum</i>	57
		( <i>Coscinonema</i> )	<i>indicum laccadivense</i>	57
		( <i>Cyliconema</i> )	<i>infundibulum</i>	27
		( <i>Hyalonema</i> )	<i>intermedium</i>	71
		( <i>Corynonema</i> )	<i>intersubgenerica</i>	71
		( <i>Paradisconema</i> )	<i>investigatoris</i>	51
		( <i>Hyalonema</i> )	<i>keianum</i>	71
		( <i>Cyliconema</i> )	<i>keiense</i>	71
		( <i>Coscinonema</i> )	<i>kenti</i>	31
		( <i>Coscinonema</i> )	<i>kirkpatricki globosum</i>	61
		( <i>Coscinonema</i> )	<i>kirkpatricki kirkpatricki</i>	71
		( <i>Coscinonema</i> )	<i>lamella</i>	57
		( <i>Cyliconema</i> )	<i>lanceolata</i>	57
		( <i>Leptonema</i> )	<i>lusitanicum</i>	21
		( <i>Cyliconema</i> )	<i>madagascarensse</i>	51
		( <i>Cyliconema</i> )	<i>martabanense</i>	57
		( <i>Cyliconema</i> )	<i>masoni</i>	57
		( <i>Oonema</i> )	<i>microstauractina</i>	71
		( <i>Cyliconema</i> )	<i>molle</i>	51
		( <i>Corynonema</i> )	<i>natalense</i>	51
		( <i>Cyliconema</i> )	<i>nicobaricum</i>	57
		( <i>Onconema</i> )	<i>obtusum</i>	77
		( <i>Coscinonema</i> )	<i>ovatum</i>	61
		( <i>Cyliconema</i> )	<i>ovatum</i>	67
		( <i>Oonema</i> )	<i>ovichela</i>	51
		( <i>Leptonema</i> )	<i>ovuliferum</i>	67
		( <i>Corynonema</i> )	<i>owstoni</i>	67
		( <i>Halonema</i> )	<i>parallela</i>	71
		( <i>Coscinonema</i> )	<i>pateriferum</i>	77
		( <i>Oonema</i> )	<i>pedunculatum</i>	77
		( <i>Halonema</i> )	<i>pellucidum</i>	61
		( <i>Prionema</i> )	<i>pinulifusum</i>	77
	<b>Halonema</b>	( <i>Corynonema</i> )	<i>pirum</i>	57
		( <i>Prionema</i> )	<i>poculum</i>	77

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
		( <i>Cyliconema</i> )	<i>polycaulm</i>	77
		( <i>Coscinonema</i> )	<i>polycoelum</i>	71
		( <i>Corynonema</i> )	<i>populiferum</i>	67
		( <i>Hyalonema</i> )	<i>proximum</i>	71
		( <i>Cyliconema</i> )	<i>rapa</i>	57
		( <i>Prionema</i> )	<i>repletum</i>	31
		( <i>Oonema</i> )	<i>robustum</i>	61
		( <i>Corynonema</i> )	<i>rotundum</i>	71
		( <i>Coscinonema</i> )	<i>schmidti</i>	31
		( <i>Oonema</i> )	<i>sequoia</i>	77
		( <i>Hyalonema</i> )	<i>sieboldi</i>	61
		( <i>Cyliconema</i> )	<i>simile</i>	51
		( <i>Hyalonema</i> )	<i>soelae</i>	57
		( <i>Leptonema</i> )	<i>solutum</i>	71
		( <i>Cyliconema</i> )	<i>somaticum</i>	51
		( <i>Leptonema</i> )	<i>spatha</i>	71
		( <i>Prionema</i> )	<i>spinosum</i>	77
		( <i>Cyliconema</i> )	<i>tasmani</i>	81
		( <i>Cyliconema</i> )	<i>tenerum</i>	71
		( <i>Coscinonema</i> )	<i>tenue</i>	41
		( <i>Corynonema</i> )	<i>tenuifusum</i>	77
		( <i>Thamnonemiella</i> )	<i>thamnophorum</i>	71
		( <i>Cyliconema</i> )	<i>thomsoni exiguum</i>	21
		( <i>Cyliconema</i> )	<i>thomsoni thomsoni</i>	21
		( <i>Cyliconema</i> )	<i>timorense</i>	71
		( <i>Ijimaonema</i> )	<i>topsentii</i>	71
		( <i>Coscinonema</i> )	<i>toxeres</i>	31
		( <i>Oonema</i> )	<i>trifidum</i>	71
		( <i>Cyliconema</i> )	<i>tulipa</i>	71
		( <i>Corynonema</i> )	<i>tylostylum</i>	77
		( <i>Oonema</i> )	<i>umbraculum</i>	77
		( <i>Onconema</i> )	<i>uncinata</i>	71
		( <i>Leptonema</i> )	<i>urna</i>	37
		( <i>Cyliconema</i> )	<i>valdiviae</i>	58
		( <i>Prionema</i> )	<i>validum</i>	51
		( <i>Paradisconema</i> )	<i>vosmaeri</i>	71
		( <i>Corynonema</i> )	<i>weltneri</i>	57
<b>Platella</b>				
			<i>polybasalia</i>	71
	<b>Compsocalyx</b>		<i>gibberosa</i>	57
	<b>Lophophysema</b>			
			<i>australicum</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>gilchristi</i>	51
			<i>inflatum</i>	57
	<i>Chalaronema</i>			
			<i>sibogae</i>	71
<b>Monorhaphididae</b>				
	<i>Monorhaphis</i>			
			<i>chuni</i>	71
<b>Pheronematidae</b>				
	<i>Pheronema</i>			
			<i>amphorae</i>	81
			<i>annaee</i>	31
			<i>barbulosclera</i>	71
			<i>carpenteri</i>	27
			<i>conicum</i>	71
			<i>echinatum</i>	71
			<i>giganteum</i>	71
			<i>gigas</i>	81
			<i>globosum globosum</i>	71
			<i>globosum kagoshimensis</i>	61
			<i>hemisphaericum</i>	71
			<i>ijimai</i>	61
			<i>megaglobosum</i>	71
			<i>nascaniensis</i>	
			<i>pilosum</i>	71
			<i>pseudogiganteum</i>	71
			<i>raphanus</i>	57
			<i>semiglobosum</i>	71
			<i>surugense</i>	61
			<i>weberi</i>	71
	<i>Platylistrum</i>			
			<i>platessa</i>	51
	<i>Poliopogon</i>			
			<i>amadou amadou</i>	27
			<i>amadou pacifica</i>	71
			<i>claviculus</i>	71
			<i>maitai</i>	71
			<i>mendocino</i>	67
			<i>micropentactinus</i>	71
			<i>zonecus</i>	71
	<i>Schulzeviella</i>			
			<i>gigas</i>	81
	<i>Semperella</i>			
			<i>abyssalis</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>alba</i>	71
			<i>crosnieri</i>	71
			<i>cucumis</i>	57
			<i>schultzei</i>	71
			<i>similis</i>	71
			<i>spicifera</i>	71
			<i>stomata</i>	67
			<i>varioactina</i>	71
	<i>Sericolophus</i>			
			<i>calsubus</i>	71
			<i>cidaricus</i>	71
			<i>hawaiicus</i>	71
			<i>neocalaledonicus</i>	71
			<i>reflexus</i>	67
Order Aulocalycoida				
<b>Aulocalycidae</b>				
	<i>Aulocalyx</i>			
			<i>irregularis</i>	51
			<i>serialis</i>	51
	<i>Rhabdodictyum</i>			
			<i>delicatum</i>	31
			<i>kurens</i>	71
	<i>Leioplegma</i>			
			<i>polyphyllon</i>	31
	<i>Euryplegma</i>			
			<i>auriculare</i>	81
Order Fieldingida				
<b>Uncinateridae</b>				
	<i>Uncinatera</i>			
			<i>plicata</i>	(48, 88)
	<i>Tretopleura</i>			
			<i>candelabrum</i>	71
			<i>styloformis</i>	71
<b>Fieldingiidae</b>				
	<i>Fieldingia</i>			
			<i>lagettoides</i>	27
			<i>valentini tizardi</i>	71
			<i>valentini valentini</i>	71
Order Hexactinosida				
<b>Aphrocallistidae</b>				
	<i>Aphrocallistes</i>			
			<i>beatrix beatrix</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>beatrix orientalis</i>	71
			<i>bocagei</i>	27
			<i>intermedia</i>	61
			<i>subglobosa</i>	71
			<i>vastus</i>	67
			<i>yatsui</i>	61
	<i>Heterochone</i>			
			<i>aleutiana</i>	61
			<i>calyx</i>	61
			<i>hamata</i>	81
			<i>incognita</i>	61
			<i>tenera</i>	67
<b>Craticulariidae</b>				
	<i>Laocoetis</i>			
			<i>perion</i>	71
<b>Cribrospongiidae</b>				
	<i>Stereochlamis</i>			
			<i>incertum</i>	71
<b>Dactylocalycidae</b>				
	<i>Auloplax</i>			
			<i>auricularis</i>	27
			<i>filholi</i>	27
	<i>Iphiteon</i>			
			<i>compressa</i>	31
			<i>panicea</i>	31
	<i>Dactylocalyx</i>			
			<i>crispus</i>	31
			<i>patella</i>	27
			<i>potatorum</i>	31
			<i>pumiceus</i>	31
			<i>subglobosus</i>	31
<b>Euretidae</b>				
	<i>Lefroyella</i>			
			<i>ceramensis</i>	71
	<i>Calyptorete</i>			
			<i>ijimai</i>	61
	<i>Eurete</i>			
			<i>atlanticum</i>	41
	<i>Eurete</i>		<i>bowerbanki</i>	61
			<i>freelandi</i>	71
			<i>irregularis</i>	61
			<i>lamellina</i>	71
			<i>marshalli</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>nipponicum</i>	61
			<i>sacculiforme</i>	61
			<i>schmidti kampeni</i>	71
			<i>schmidti schmidti</i>	71
			<i>schmidti treubi</i>	71
			<i>simplicissimum</i>	71
			<i>spinosum</i>	77
			<i>trachydocus</i>	71
	<i>Conorete</i>			
			<i>erectum erectum</i>	77
			<i>erectum gracile</i>	77
			<i>erectum mucronatum</i>	77
			<i>erectum tubulifer</i>	77
	<i>Pityrete</i>			
			<i>azoricum</i>	(48, 88)
	<i>Endorete</i>			
			<i>pertusum</i>	27
	<i>Pararete</i>			
			<i>baliense</i>	71
			<i>carteri</i>	71
			<i>farreopsis fragiferum</i>	71
			<i>farreopsis jakosalemi</i>	71
			<i>farreopsis</i>	71
			<i>farrreopsis subglobosum</i>	71
			<i>freeri</i>	71
			<i>gerlachei</i>	(48, 88)
			<i>kangeanganum</i>	71
			<i>semperi</i>	71
	<i>Gymnoretete</i>			
			<i>alicei</i>	(48, 88)
	<i>Heterorete</i>			
			<i>pulchrum</i>	51
	<i>Chonelasma</i>			
			<i>choanoides</i>	(48, 88)
			<i>doederleini</i>	61
			<i>ijimai</i>	(48, 88)
			<i>lamella</i>	81
	<i>Bathyxiphus</i>			
			<i>subtilis</i>	67
	<i>Myliusia</i>			
			<i>callocyathus</i>	31
			<i>challengeri</i>	71
			<i>conica</i>	31

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>lanterna</i>	31
			<i>seriata</i>	31
			<i>verrucosa</i>	71
	<i>Periphragella</i>			
			<i>antarctica</i>	(48, 88)
			<i>challengeri</i>	71
			<i>elisae</i>	71
			<i>irregularis</i>	71
			<i>lusitanica</i>	27
			<i>parva</i>	71
	<i>Pleurochorium</i>			
			<i>annandalei</i>	(48, 88)
			<i>cornutum</i>	71
	<i>Tretochone</i>			
			<i>duplicata</i>	27
	<i>Verrucocoeloidea</i>			
			<i>burtoni</i>	71
<b>Farreidae</b>				
	<i>Sarostegia</i>			
			<i>oculata</i>	34
	<i>Asceptrulum</i>			
			<i>axialis</i>	67
	<i>Farrea</i>			
			<i>aculeata</i>	31
			<i>aculeata</i>	67
			<i>beringiana</i>	61
			<i>convolvulus</i>	61
			<i>deanea</i>	31
			<i>fistulata</i>	31
			<i>foliascens</i>	21
			<i>gassioti</i>	31
			<i>hanitschi</i>	71
			<i>herdendorfi</i>	21
			<i>inermis</i>	31
			<i>intermedia</i>	71
			<i>irregularis</i>	37
	<i>Farrea</i>		<i>kurilensis</i>	61
			<i>laevis</i>	31
			<i>laminaris</i>	21
			<i>lendenfeldi</i>	71
			<i>mexicana</i>	77
			<i>nodulosa</i>	71
			<i>occa claviformis</i>	77

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>occa clavigera</i>	71
			<i>occa cuspidata</i>	71
			<i>occa erecta</i>	71
			<i>occa mammillata</i>	71
			<i>occa microclavula</i>	71
			<i>occa occa</i>	61
			<i>occa ouwensi</i>	71
			<i>occa polyclavula</i>	71
			<i>occa scutella</i>	77
			<i>occa subclavigera</i>	71
			<i>parasitica</i>	31
			<i>perarmata</i>	31
			<i>pocillum</i>	31
			<i>robusta</i>	31
			<i>seiri</i>	58
			<i>sollasi sollasi</i>	61
			<i>sollasi yakushimensis</i>	61
			<i>spinifera</i>	31
			<i>spinulenta</i>	37
			<i>spirifera</i>	71
			<i>vosmaeri</i>	61
			<i>watasei</i>	61
			<i>weltneri</i>	(48, 88)
			<i>woodwardi</i>	27
	<i>Claviscopulia</i>			
			<i>facunda</i>	31
	<i>Lonchiphora</i>			
			<i>inversa</i>	71
	<i>Aspidoscopulia</i>			
			<i>furcillata</i>	71
Tretodictyidae				
	<i>Anomochone</i>			
			<i>expansa</i>	71
			<i>globosa</i>	71
	<i>Cyrtaulon</i>		<i>sigsbeei</i>	31
			<i>solutus</i>	71
	<i>Tretocalyx</i>		<i>polae</i>	51
	<i>Sclerothamnopsis</i>		<i>compressa</i>	77
			<i>schulzei</i>	71
	<i>Sclerothamnus</i>			

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>clausi</i>	71
	<b><i>Tretodictyum</i></b>			
			<i>labyrinthicum</i>	77
			<i>montereyense</i>	67
			<i>pumicosum</i>	71
			<i>schrammeni</i>	71
			<i>tubulosum</i>	61
	<b><i>Psilocalyx</i></b>			
			<i>wilsoni</i>	71
	<b><i>Hexactinella</i></b>			
			<i>carolinensis</i>	21
			<i>divergens</i>	41
			<i>grimaldii</i>	27
			<i>lata</i>	71
			<i>lingua</i>	71
			<i>minor</i>	57
			<i>monticularis</i>	77
			<i>rugosa</i>	71
			<i>spongiosa</i>	71
			<i>ventilabrum</i>	61
			<i>vermiculosa</i>	71
Order Lychniscosida				
<b>Aulocystidae</b>				
	<b><i>Lychnocystis</i></b>			
			<i>superstes</i>	31
	<b><i>Neoaulocystis</i></b>			
			<i>grayi</i>	31
			<i>polae</i>	71
			<i>zitteli sibogae</i>	71
			<i>zitteli</i>	71
<b>Diapleuridae</b>				
	<b><i>Scleroplegma</i></b>			
			<i>herculeum</i>	31
			<i>lanterna</i>	31
	<b><i>Scleroplegma</i></b>		<i>maasi</i>	71
			<i>seriatum</i>	31
Order Lyssacinosida				
<b>Euplectellidae</b>				
	<b><i>Placopegma</i></b>			
			<i>plumicomum</i>	71
			<i>solutum</i>	57
	<b><i>Acoelocalyx</i></b>			

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>brucei</i>	(48, 88)
	<i>Malacosaccus</i>			
			<i>coatsi</i>	(48, 88)
			<i>erectus</i>	81
			<i>floricomatus</i>	(48, 88)
			<i>heteropinularia</i>	47
			<i>pedunculatus</i>	(48, 88)
			<i>unguiculatus</i>	34
			<i>vastus</i>	58
	<i>Docosaccus</i>			
			<i>ancoratus</i>	(48, 88)
	<i>Euplectella</i>			
			<i>aspera</i>	57
			<i>aspergillum aspergillum</i>	71
			<i>aspergillum indonesicum</i>	71
			<i>aspergillum regalis</i>	81
			<i>crassistellata</i>	71
			<i>cucumer</i>	51
			<i>curvistellata</i>	(48, 88)
			<i>gibbsa</i>	21
			<i>imperialis</i>	67
			<i>jovis</i>	31
			<i>marshalli</i>	67
			<i>nobilis</i>	27
			<i>nodosa</i>	31
			<i>oweni</i>	61
			<i>paratettractina</i>	57
			<i>plumosum</i>	71
			<i>simplex</i>	57
			<i>suberea</i>	27
			<i>timorensis</i>	71
	<i>Holascus</i>			
			<i>ancoratus</i>	77
			<i>belyaevi</i>	61
	<i>Holascus</i>		<i>edwardsi</i>	77
			<i>euonyx</i>	77
			<i>fibulatus</i>	81
			<i>obesus</i>	(48, 88)
			<i>polejaevi</i>	(48, 88)
			<i>pseudostellatus</i>	(48, 88)
			<i>ridleyi</i>	71
			<i>robustus</i>	57

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>stellatus</i>	41
			<i>taraxacum</i>	77
			<i>tener</i>	57
			<i>tenuis</i>	(48, 88)
			<i>undulatus</i>	18
	<i>Trachycaulus</i>			
			<i>gurlitti</i>	81
	<i>Caledoniella</i>			
			<i>caulophacoides</i>	71
	<i>Caulocalyx</i>			
			<i>tener</i>	47
	<i>Saccocalyx</i>			
			<i>careyi</i>	(48, 88)
			<i>pedunculatus</i>	57
	<i>Bolosoma</i>			
			<i>biocalum</i>	71
			<i>cavum</i>	71
			<i>charcoti</i>	71
			<i>cyanae</i>	71
			<i>meridionale</i>	71
			<i>musorstromum</i>	71
			<i>paradictyum</i>	61
			<i>volsmarum</i>	71
	<i>Vityaziella</i>			
			<i>renki</i>	57
	<i>Amphidiscella</i>			
			<i>atlantica</i>	61
			<i>caledonica</i>	71
			<i>monai</i>	41
	<i>Hyalostylus</i>			
			<i>dives</i>	81
			<i>monomicrosclerus</i>	71
	<i>Hertwiggia</i>			
			<i>falcifera</i>	31
	<i>Pseudoplectella</i>			
			<i>dentatum</i>	
	<i>Corbitella</i>			
			<i>discasterosa</i>	71
			<i>elegans</i>	71
			<i>pulchra</i>	71
			<i>speciosa</i>	71
	<i>Dictyaulus</i>			
			<i>elegans</i>	57

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>marecoi</i>	21
			<i>starmeri</i>	71
	<i>Heterotella</i>			
			<i>corbicula</i>	51
			<i>midatlantica</i>	21
			<i>pacifica</i>	71
			<i>pomponae</i>	31
	<i>Regadrella</i>			
			<i>cylindrica</i>	71
			<i>decora</i>	57
			<i>delicata</i>	77
			<i>heterotelliformis</i>	71
			<i>komeyamai</i>	61
			<i>okinoseana</i>	67
			<i>peru</i>	
			<i>phoenix</i>	31
			<i>rhizophora</i>	71
	<i>Atlantisella</i>			
			<i>incognita</i>	27
	<i>Walteria</i>			
			<i>flemmingi</i>	81
			<i>leuckarti leuckarti</i>	67
			<i>leuckarti longipina</i>	71
	<i>Dictyocalyx</i>			
			<i>gracilis</i>	71
			<i>lifousantalis</i>	71
	<i>Symplectella</i>			
			<i>rowi</i>	81
	<i>Ijimaiella</i>			
			<i>beringiana</i>	18
	<i>Rhabdopectella</i>			
			<i>tintinnus</i>	31
Leucopsacidae				
	<i>Oopsacas</i>			
			<i>minuta</i>	27
	<i>Leucopsacus</i>			
			<i>distantus</i>	71
			<i>ingolfi</i>	27
			<i>orthodocus</i>	61
			<i>scoliodocus retroscissus</i>	21
			<i>scoliodocus scoliodocus</i>	61
	<i>Chaunoplectella</i>			
			<i>cavernosa</i>	67

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>spinifera</i>	61
			<i>stelleta</i>	71
<b>Rosselidae</b>				
	<b><i>Crateromorpha</i></b>			
		( <i>Craterochone</i> )	<i>bermudensis</i>	71
		( <i>Caledochone</i> )	<i>caledoniensis</i>	71
		( <i>Aulochone</i> )	<i>cylindrica</i>	81
			<i>haliprum</i>	71
		( <i>Neopsacas</i> )	<i>krinovi discoli</i>	77
		( <i>Neopsacas</i> )	<i>krinovi krinovi</i>	21
		( <i>Crateromorpha</i> )	<i>lankesteri</i>	51
		( <i>Aulochone</i> )	<i>lilium</i>	71
		( <i>Crateromorpha</i> )	<i>meyeri corrugata</i>	61
		( <i>Crateromorpha</i> )	<i>meyeri meyeri</i>	71
		( <i>Crateromorpha</i> )	<i>meyeri rugosa</i>	61
		( <i>Crateromorpha</i> )	<i>meyeri tuberosa</i>	61
		( <i>Crateromorpha</i> )	<i>meyeri tubulosa</i>	61
		( <i>Crateromorpha</i> )	<i>murrayi</i>	71
		( <i>Neopsacas</i> )	<i>obi</i>	51
		( <i>Crateromorpha</i> )	<i>pachyactina</i>	61
		( <i>Aulochone</i> )	<i>pedunculata</i>	71
		( <i>Crateromorpha</i> )	<i>thierfelderi</i>	71
		( <i>Crateromorpha</i> )	<i>tumida</i>	71
		( <i>Neopsacas</i> )	<i>variata</i>	27
	<b><i>Caulophacella</i></b>			
			<i>tenuis</i>	77
	<b><i>Caulophacus</i></b>			
		( <i>Caulophacus</i> )	<i>abyssalis</i>	41
		( <i>Caulophacus</i> )	<i>agassizi</i>	21
		( <i>Caulophacus</i> )	<i>antarcticus</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>arcticus</i>	18
		( <i>Caulophacus</i> )	<i>basispinosus</i>	51
	<b><i>Caulophacus</i></b>	( <i>Caulodiscus</i> )	<i>brandti</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>cyanae</i>	71
		( <i>Caulophacus</i> )	<i>discohexactinus</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>discohexaster</i>	71
		( <i>Caulophacus</i> )	<i>elegans</i>	61
		( <i>Caulophacus</i> )	<i>galatheae</i>	51
		( <i>Caulophacus</i> )	<i>hadalis</i>	81
		( <i>Caulophacus</i> )	<i>instabilis</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>latus</i>	58
		( <i>Caulophacus</i> )	<i>lotifolium similis</i>	71
		( <i>Caulodiscus</i> )	<i>onychohexactinus</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
		( <i>Caulodiscus</i> )	<i>otifolium</i>	61
		( <i>Caulophacus</i> )	<i>oviformis</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>pipetta</i>	(48, 88)
		( <i>Caulodiscus</i> )	<i>polyspicula</i>	47
		( <i>Caulophacus</i> )	<i>schulzei</i>	77
		( <i>Caulophacus</i> )	<i>scotiae</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>valdiviae</i>	(48, 88)
		( <i>Caulophacus</i> )	<i>variens juvenilis</i>	71
		( <i>Caulophacus</i> )	<i>variens</i>	71
		( <i>Oxydiscus</i> )	<i>weddelli</i>	(48, 88)
	<i>Asconema</i>			
			<i>bispiculigastrum</i>	18
			<i>foliatum</i>	18
			<i>fristedti fristedti</i>	21
			<i>fristedti islandiense</i>	27
			<i>fristedti nordazoriense</i>	27
			<i>megaatrialia biacorica</i>	21
			<i>megaatrialia</i>	
			<i>megaatrialia</i>	27
			<i>megaatrialia nordiense</i>	18
			<i>megaatrialia seamounti</i>	21
			<i>setubalense</i>	27
			<i>topsentii</i>	27
	<i>Aphorme</i>			
			<i>horrida</i>	61
	<i>Trichasterina</i>			
			<i>borealis</i>	27
			<i>sagittaria</i>	(48, 88)
	<i>Bathydorus</i>			
			<i>fimbriatus</i>	61
			<i>laevis ciliatus</i>	(48, 88)
			<i>laevis laevis</i>	57
	<i>Bathydorus</i>		<i>laevis spinosus</i>	77
			<i>servatus</i>	27
			<i>spinosissimus</i>	77
			<i>spinosus</i>	58
			<i>uncifer</i>	67
	<i>Vitrollula</i>			
			<i>fertilis</i>	61
	<i>Schaudinnia</i>			
			<i>rosea</i>	18
	<i>Hyalascus</i>			
			<i>anisoactinus</i>	71

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>attenuatus</i>	61
			<i>baculifer</i>	81
			<i>giganteus</i>	61
			<i>hodgsoni</i>	(48, 88)
			<i>pinulohexactinus</i>	71
			<i>sagamiensis</i>	67
			<i>similis</i>	61
			<i>stellatus</i>	41
	<i>Vazella</i>			
			<i>pourtalesi</i>	31
	<i>Acanthascus</i>			
		( <i>Staurocalyptus</i> )	<i>affinis</i>	61
		( <i>Acanthascus</i> )	<i>alani</i>	61
		( <i>Rhabdocalyptus</i> )	<i>arcticus</i>	18
		( <i>Rhabdocalyptus</i> )	<i>asper</i>	67
		( <i>Rhabdocalyptus</i> )	<i>australis</i>	(48, 88)
		( <i>Rhabdocalyptus</i> )	<i>baculifer</i>	51
		( <i>Rhabdocalyptus</i> )	<i>bidentatus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>borealis</i>	61
		( <i>Acanthascus</i> )	<i>cactus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>capillatus</i>	61
		( <i>Staurocalyptus</i> )	<i>celebesianus</i>	71
		( <i>Rhabdocalyptus</i> )	<i>dawsoni</i>	67
		( <i>Staurocalyptus</i> )	<i>dowlingi</i>	67
		( <i>Staurocalyptus</i> )	<i>entacanthus</i>	61
		( <i>Staurocalyptus</i> )	<i>fasciculatus</i>	67
		( <i>Staurocalyptus</i> )	<i>fuca</i>	71
		( <i>Staurocalyptus</i> )	<i>glaber</i>	61
		( <i>Staurocalyptus</i> )	<i>hamatus</i>	77
		( <i>Staurocalyptus</i> )	<i>heteractinus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>heteraster</i>	61
	<i>Acanthascus</i>	( <i>Staurocalyptus</i> )	<i>microchetus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>mirabilis</i>	67
		( <i>Rhabdocalyptus</i> )	<i>mollis</i>	61
		( <i>Rhabdocalyptus</i> )	<i>monstraster</i>	51
		( <i>Rhabdocalyptus</i> )	<i>nodulosus</i>	67
		( <i>Acanthascus</i> )	<i>pachyderma</i>	61
		( <i>Acanthascus</i> )	<i>platei</i>	67
		( <i>Staurocalyptus</i> )	<i>pleorhaphides</i>	61
		( <i>Rhabdocalyptus</i> )	<i>plumodigitatus</i>	51
		( <i>Staurocalyptus</i> )	<i>roeperi</i>	
		( <i>Staurocalyptus</i> )	<i>rugocruciatus</i>	61
		( <i>Staurocalyptus</i> )	<i>solidus</i>	67

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
		( <i>Rhabdocalyptus</i> )	<i>tener</i>	67
		( <i>Rhabdocalyptus</i> )	<i>tenuis</i>	67
		( <i>Staurocalyptus</i> )	<i>tubulosus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>unguiculatus</i>	61
		( <i>Rhabdocalyptus</i> )	<i>victor</i>	61
	<b><i>Rossella</i></b>			
			<i>antarctica</i>	(48, 88)
			<i>aperta</i>	(48, 88)
			<i>dubia</i>	
			<i>fibulata</i>	(48, 88)
			<i>gaini</i>	(48, 88)
			<i>ijimai</i>	81
			<i>inermis</i>	(48, 88)
			<i>levis</i>	(48, 88)
			<i>longstaffi</i>	(48, 88)
			<i>lychnophora</i>	(48, 88)
			<i>mixta</i>	(48, 88)
			<i>mortensi</i>	27
			<i>nodastrella</i>	27
			<i>nuda</i>	(48, 88)
			<i>pilosa</i>	(48, 88)
			<i>podagrosa tenuis</i>	(48, 88)
			<i>podagrosa</i>	(48, 88)
			<i>racovitzae hexactinophila</i>	(48, 88)
			<i>racovitzae microdiscina</i>	(48, 88)
			<i>racovitzae racovitzae</i>	(48, 88)
			<i>schulzei</i>	(48, 88)
			<i>vanhoeffeni armata</i>	(48, 88)
			<i>vanhoeffeni vanhoeffeni</i>	(48, 88)
			<i>villosa</i>	(48, 88)
	<b><i>Rossella</i></b>		<i>vitiosa</i>	(48, 88)
	<b><i>Anoxycalyx</i></b>			
		( <i>Anoxycalyx</i> )	<i>ijimai</i>	(48, 88)
		( <i>Scolymastra</i> )	<i>joubini</i>	(48, 88)
		( <i>Anoxycalyx</i> )	<i>laceratus</i>	61
	<b><i>Aulosaccus</i></b>			
			<i>albatrossi</i>	61
			<i>fissuratus shimushirensis</i>	61
			<i>fissuratus</i>	61
			<i>ijimae</i>	67
			<i>mitsukurii</i>	61
			<i>pinularis</i>	61

Order	Genus	Subgenus	Species	FAO Major Fishing Areas
			<i>schulzei</i>	67
			<i>solaster</i>	61
	<i>Scyphidium</i>			
			<i>australiense</i>	81
			<i>chilense</i>	
			<i>jamatai</i>	61
			<i>longispinum</i>	67
			<i>namiyei</i>	61
			<i>septentrionale</i>	18
			<i>tuberculatum</i>	61
	<i>Calycosoma</i>			
			<i>validum</i>	21
	<i>Sympagella</i>			
			<i>anomala</i>	61
			<i>cantharellus</i>	77
			<i>clavipinula</i>	71
			<i>gracilis</i>	71
			<i>johnstoni</i>	58
			<i>multihexastera</i>	57
			<i>nux</i>	31
	<i>Doconesthes</i>			
			<i>sessilis</i>	27
	<i>Lanuginella</i>			
			<i>pupa</i>	31
	<i>Lanugonychia</i>			
			<i>flabellum</i>	77
	<i>Lophocalyx</i>			
			<i>atlantiensis</i>	41
			<i>biogasi</i>	41
			<i>brasiliensis</i>	41
	<i>Lophocalyx</i>		<i>moscalevia</i>	71
			<i>oregoni</i>	41
			<i>philippinensis</i>	71
			<i>profunda</i>	(48, 88)
			<i>pseudovalida</i>	41
			<i>reiswigi</i>	41
			<i>spinosa</i>	57
			<i>suluana</i>	71
			<i>topsendi</i>	(48, 88)
	<i>Mellonymphha</i>			
			<i>velata</i>	21

**APPENDIX 2****Astrophorida**

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Ancorinidae</b>			
	<i>Stelletta</i>		
		<i>addita</i>	37
		<i>aeruginosa</i>	81
		<i>agglutinans</i>	57
		<i>agulhana</i>	47
		<i>agulhana paucistella</i>	47
		<i>anancora</i>	41
		<i>anasteria</i>	41
		<i>arenaria</i>	81
		<i>aruensis</i>	71
		<i>aspera</i>	71
		<i>atrophia</i>	61
		<i>beae</i>	41
		<i>bocki</i>	57
		<i>boglicii</i>	37
		<i>brevidens</i>	71
		<i>brevioxea</i>	51
		<i>brevis</i>	57
		<i>calyx</i>	61
		<i>capensis</i>	47
		<i>carolinensis</i>	21
		<i>cavernosa</i>	51, 57, 58
		<i>centroradiata</i>	71
		<i>centrotyla</i>	57
		<i>ceylonica</i>	57
		<i>clarella</i>	77
		<i>clavosa</i>	71
		<i>communis</i>	57, 71, 81
		<i>conulosa</i>	81
		<i>crassicula</i>	51
		<i>crassispicula</i>	34, 41
		<i>crater</i>	81
		<i>cyathoides</i>	47
		<i>cylindrica</i>	51
		<i>debilis</i>	71
		<i>debilis stenospiculata</i>	31
		<i>digitifera</i>	34
		<i>discolor</i>	51
		<i>dorsigera</i>	37

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Stelletta</i>	<i>durissima</i>	71
		<i>estrella</i>	77
		<i>farcimen</i>	47
		<i>fibrosa</i>	31
		<i>fibulifera</i>	37
		<i>freitasi</i>	51
		<i>gigantea</i>	61
		<i>gigas</i>	41
		<i>globulariformis</i>	31
		<i>grubii</i>	27, 37
		<i>grubiooides</i>	47
		<i>hajdui</i>	41
		<i>herdmani</i>	47, 57
		<i>herdmani robusta</i>	51
		<i>hispida</i>	34, 37
		<i>horrens</i>	47
		<i>horrens subcylindrica</i>	47
		<i>hyperoxea</i>	71
		<i>incrustans</i>	27
		<i>incrustata</i>	31
		<i>individua</i>	31
		<i>inermis</i>	27
		<i>japonica</i>	61
		<i>jonesi</i>	51
		<i>kallitetilla</i>	31
		<i>kundukensis</i>	61
		<i>lactea</i>	27, 37
		<i>latiancora</i>	34
		<i>lithodes</i>	81
		<i>longicladus</i>	57
		<i>mamilliformis</i>	57
		<i>maori</i>	81
		<i>mauritiana</i>	51
		<i>maxima</i>	61, 81
		<i>mediterranea</i>	37
		<i>misakensis</i>	61
		<i>morikawai</i>	61
		<i>moseleyi</i>	57, 71, 81
		<i>naseana</i>	61
		<i>normani</i>	21, 27
		<i>novaezealandiae</i>	81
		<i>orientalis</i>	61
		<i>orthotriaena</i>	61

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Stelletta</i>	<i>osculifera</i>	51
		<i>ovalae</i>	61
		<i>pachydermata</i>	77
		<i>parva</i>	51
		<i>parvispicula</i>	71
		<i>paucistellata</i>	34
		<i>phialimorpha</i>	81
		<i>phrissens</i>	87
		<i>pisum</i>	61
		<i>plagioreducta</i>	71
		<i>porosa</i>	71
		<i>pudica</i>	31
		<i>pulchra</i>	71
		<i>pumex</i>	37
		<i>purpurea</i>	51, 57, 61, 71, 81
		<i>pygmaeorum</i>	31
		<i>pyriformis</i>	71
		<i>radicifera</i>	71
		<i>radicifera robusta</i>	71
		<i>retroclada</i>	47
		<i>rhaphidiophora</i>	27
		<i>ridleyi</i>	71
		<i>ruetzleri</i>	41
		<i>rugosa</i>	47
		<i>sandalinum</i>	81
		<i>siemensi</i>	51
		<i>sigmatriaena</i>	71
		<i>simplicissima</i>	37
		<i>solida</i>	31
		<i>solida</i>	61
		<i>solidissima</i>	31
		<i>soteropolitana</i>	41
		<i>sphaerica</i>	47
		<i>sphaeroides</i>	71
		<i>spinulosa</i>	61
		<i>splendens</i>	61
		<i>stellata</i>	37
		<i>stellata</i>	37
		<i>stellifera</i>	71
		<i>subtilis</i>	61
		<i>tenuispicula</i>	31
		<i>teres</i>	61
		<i>tethyopsis</i>	57

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Stelletta</i>	<i>tetrafurcata</i>	61
		<i>toxiastra</i>	81
		<i>trichotriaena</i>	57
		<i>trisclera</i>	47
		<i>tuba</i>	61
		<i>tuberculata</i>	57
		<i>tuberosa</i>	27
		<i>tulearensis</i>	51
		<i>vaceleti</i>	81
		<i>validissima</i>	61
		<i>variabilis</i>	31
		<i>variohamata</i>	71
		<i>ventricosa</i>	27
		<i>vestigium</i>	51
		<i>vosmaeri</i>	87
	<i>Tethyopsis</i>		
		<i>brondstedi</i>	(48, 88)
		<i>calcifera</i>	81
		<i>longispinus</i>	(48, 88)
		<i>patriciae</i>	71
		<i>radiata</i>	81
	<i>Ancorina</i>		
		<i>diplococcus</i>	81
		<i>multistella</i>	34
	<i>Stryphnus</i>		
		<i>fortis</i>	27
		<i>mucronatus</i>	37
		<i>ponderosus</i>	27
		<i>progressus</i>	47
	<i>Penares</i>		
		<i>cortiusorientalis</i>	61
		<i>foliaformis</i>	77
		<i>mastoidea</i>	31
		<i>micraster</i>	71
		<i>palmatoclada</i>	71
		<i>sclerobesa</i>	27
		<i>sphaera</i>	47
<b>Calthropellidae</b>			
	<i>Calthropella</i>		
		<i>simplex</i>	27
		<i>simplex durissima</i>	27
	<i>Pachataxa</i>		
		<i>enigmatica</i>	71

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Pachataxa</i>	<i>lithistina</i>	31
		<i>n.sp.</i>	77
<b>Geodiidae</b>			
	<i>Erylus</i>		
		<i>aleuticus</i>	67
		<i>amissus</i>	71
		<i>amorphus</i>	47
		<i>amphiastera</i>	31
		<i>aspidodiscus</i>	37
		<i>bahamensis</i>	31
		<i>burtoni</i>	71
		<i>caliculatus</i>	77
		<i>cantabricus</i>	27
		<i>carteri</i>	57
		<i>circus</i>	71
		<i>citrus</i>	71
		<i>corneus</i>	41
		<i>cornutus</i>	71
		<i>cylindriger</i>	51
		<i>decumbens</i>	71
		<i>deficiens</i>	34
		<i>diminutus</i>	41
		<i>discastera</i>	77
		<i>discophorus</i>	27, 37
		<i>euastrum</i>	37
		<i>expletus</i>	27
		<i>fibrillosus</i>	71
		<i>formosus</i>	41
		<i>fromontae</i>	57
		<i>geodiooides</i>	57
		<i>gilchristi</i>	47
		<i>goffrilleri</i>	31
		<i>inaequalis</i>	71
		<i>incrustans</i>	31
		<i>lendenfeldi</i>	57
		<i>mamillaris</i>	27, 37
		<i>monticularis</i>	71
		<i>nobilis</i>	71
		<i>nummulifer</i>	27
		<i>oblongus</i>	27
		<i>oxyaster</i>	87
		<i>papillatus</i>	27
		<i>philippinens</i>	71

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Erylus</i>	<i>placenta</i>	61
		<i>polyaster</i>	47
		<i>proximus</i>	57
		<i>rotundus</i>	77
		<i>soesti</i>	41
		<i>sollasi</i>	77
		<i>topsentii</i>	27
		<i>toxiformis</i>	41
		<i>transiens</i>	27, 31
		<i>trisphaerus</i>	31
	<i>Caminus</i>		
		<i>albus</i>	71
		<i>awashimensis</i>	61
		<i>chinensis</i>	61
		<i>sphaeroconia</i>	41
		<i>vulcani</i>	34, 37
	<i>Pachymatisma</i>		
		<i>areolata</i>	57
		<i>bifida</i>	51
		<i>geodiformis</i>	31
		<i>johnstonia</i>	27
		<i>monaena</i>	47
		<i>normani</i>	27
	<i>Geodia</i>		
		<i>acanthylastra</i>	77
		<i>agassizi</i>	77
		<i>alba</i>	71
		<i>alba minor</i>	71
		<i>amadaiba</i>	61
		<i>amphistomyla</i>	87
		<i>arabica</i>	51
		<i>areolata</i>	57
		<i>arripiens</i>	71
		<i>ataxastra</i>	77
		<i>ataxastra angustana</i>	77
		<i>ataxastra latana</i>	77
		<i>auroristella</i>	51
		<i>australis</i>	41
		<i>barretti</i>	27, 34, 37
		<i>basilea</i>	47

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Geodia</i>	<i>berryi</i>	61
		<i>breviana</i>	77
		<i>carteri</i>	57
		<i>cidaris</i>	37
		<i>composita</i>	51
		<i>conchilega</i>	37
		<i>cooksoni</i>	87
		<i>crustosa</i>	51
		<i>cumulus</i>	31
		<i>cydonium</i>	27, 37
		<i>cydonium aegagropila</i>	37
		<i>cylindrica</i>	61
		<i>dendyi</i>	47
		<i>depressa</i>	37
		<i>distincta</i>	71
		<i>divaricans</i>	34
		<i>dura</i>	71
		<i>dysoni</i>	77
		<i>echinastrella</i>	27
		<i>eosaster</i>	27
		<i>erinacea</i>	57
		<i>exigua</i>	61
		<i>gallica</i>	47
		<i>geodina</i>	37
		<i>gibberella</i>	77
		<i>gibberosa</i>	31
		<i>gigas</i>	37
		<i>glariosa</i>	41
		<i>globosa</i>	47
		<i>globostellifera</i>	51, 57
		<i>globus</i>	27
		<i>hilgendorfi</i>	61
		<i>hilgendorfii</i>	61
		<i>hirsuta</i>	71
		<i>hyotania</i>	61
		<i>inconspicua</i>	71
		<i>isabella</i>	77
		<i>japonica</i>	61
		<i>japonica spherulifera</i>	71
		<i>kuekenthali</i>	71
		<i>labyrinthica</i>	47
		<i>libera</i>	47, (48, 88)
		<i>lindgreni</i>	57, 71

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Geodia</i>	<i>littoralis</i>	47, (48, 88)
		<i>lophotriaena</i>	81
		<i>macandrewi</i>	27
		<i>magellani</i>	87
		<i>media</i>	77
		<i>media leptorhaphes</i>	31
		<i>megaster</i>	47
		<i>megastrella</i>	27, 34
		<i>megastrella laevispina</i>	27
		<i>mesotriaena</i>	77
		<i>mesotriaena</i>	18
		<i>mesotriaenella</i>	77
		<i>micropora</i>	87
		<i>micropunctata</i>	51
		<i>nigra</i>	57
		<i>nodastrella</i>	27, 34
		<i>normani</i>	27
		<i>orthomesotriaena</i>	61
		<i>ovifractus</i>	47
		<i>ovifractus cyathoides</i>	47
		<i>ovis</i>	67
		<i>papyracea</i>	31
		<i>pergamentacea</i>	27
		<i>peruncinata</i>	51
		<i>philippensis</i>	71
		<i>piriformis</i>	51
		<i>placenta</i>	37
		<i>pleiades</i>	51
		<i>poculata</i>	51
		<i>punctata</i>	57
		<i>ramodigitata</i>	51
		<i>ramosa</i>	34
		<i>regina</i>	81
		<i>reniformis</i>	61
		<i>reniformis robusta</i>	61
		<i>riograndensis</i>	41
		<i>robusta</i>	47
		<i>robusta carolae</i>	67
		<i>robusta megaclada</i>	67
		<i>robusta megasterra</i>	77
		<i>rovinciensis</i>	37
		<i>senegalensis</i>	34
		<i>simplex</i>	27

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Geodia</i>	<i>simplicissima</i>	27
		<i>sparsa</i>	71
		<i>sphaeroides</i>	71
		<i>spheranthesstra</i>	51
		<i>spherastrea</i>	31
		<i>spherastrella</i>	27
		<i>spherastrosa</i>	71
		<i>splendida</i>	41
		<i>stellata</i>	47
		<i>stellosa</i>	37
		<i>stromatodes</i>	31
		<i>strongyla</i>	61
		<i>thomsoni</i>	31
		<i>tuber</i>	37
		<i>tuberculosa</i>	77
		<i>tuberosa</i>	37
		<i>tylastra</i>	41
		<i>variospiculosa</i>	61
		<i>variospiculosa clavigera</i>	61
		<i>variospiculosa intermedia</i>	61
		<i>variospiculosa micraster</i>	61
		<i>variospiculosa aapta</i>	61
		<i>vaubani</i>	71
		<i>vestigifera</i>	81
	<i>Isops</i>		
		<i>apiarum</i>	31
		<i>canaliculata</i>	37
		<i>contorta</i>	71
		<i>imperfecta</i>	71
		<i>loricata</i>	37
		<i>maculosus</i>	37
		<i>micraster</i>	51
		<i>nigra</i>	71
		<i>obscura</i>	61
		<i>ostracomorpha</i>	71
		<i>pachydermata</i>	27, 31
		<i>perarmata</i>	51
		<i>phlegraei</i>	18, 27
		<i>sollasi</i>	71
		<i>velata</i>	61
	<i>Sidonops</i>		
		<i>angulata</i>	77

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Sidonops</i>	<i>atlantica</i>	27
		<i>bicolor</i>	77
		<i>californica</i>	77
		<i>corticostylifera</i>	41
		<i>flemingi</i>	81
		<i>microspinosa</i>	71
		<i>neptuni</i>	41
		<i>nitida</i>	41
		<i>oxyastræ</i>	77
		<i>picteti</i>	71
		<i>reticulata</i>	77
		<i>vosmaeri</i>	41
<b>Pachastrellidae</b>			
	<i>Characella</i>		
		<i>abbreviata</i>	71
		<i>agassizi</i>	31
		<i>aspera</i>	41
		<i>capitolii</i>	41
		<i>connectens</i>	31
		<i>enæ</i>	31
		<i>flexibilis</i>	71
		<i>ijimai</i>	61
		<i>laevis</i>	61
		<i>pachastrelloides</i>	27, 34
		<i>reticulata</i>	61
		<i>stellettodes</i>	61
		<i>tripodaria</i>	37
	<i>Cladothenea</i>		
		<i>andriashevi</i>	(48, 88)
	<i>Pachastrella</i>		
		<i>abyssi</i>	27, 31
		<i>caliculata</i>	47
		<i>chuni</i>	34
		<i>cribrum</i>	61
		<i>dilifera</i>	31
		<i>echinorhabda</i>	37
		<i>fusca</i>	61
		<i>incrustata</i>	81
		<i>isorrhopa</i>	47
		<i>monilifera</i>	27, 34, 37, 47
		<i>multipora</i>	77
		<i>ovisternata</i>	27
		<i>scrobiculosæ</i>	61

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Poecillastra</i>		
		<i>amygdalooides</i>	27, 34,
		<i>ciliata</i>	71
		<i>compressa</i>	27, 34, 37, 47
		<i>compressa antarctica</i>	(48, 88)
		<i>crassiuscula</i>	27, 34
		<i>cribraria</i>	87
		<i>eccentrica</i>	57
		<i>fragilis</i>	37
		<i>incrustans</i>	47
		<i>japonica</i>	61
		<i>laminaris</i>	71
		<i>nana</i>	57
		<i>rickettsi</i>	77
		<i>saxicola</i>	37
		<i>schulzei</i>	58
		<i>stipitata</i>	71
		<i>symbiotica</i>	27
		<i>tenuilaminaris</i>	71
		<i>tenuirhabda</i>	47
		<i>tuberosa</i>	47
		<i>wondoensis</i>	61
	<i>Thenea</i>		
		<i>abyssorum</i>	18
		<i>andamanensis</i>	57
		<i>bojeadori</i>	34
		<i>calyx</i>	61
		<i>centrotyla</i>	58
		<i>compacta</i>	61
		<i>compressa</i>	61
		<i>corallophila</i>	57
		<i>delicata</i>	58
		<i>echinata</i>	77
		<i>echinata</i>	21
		<i>fenestrata</i>	31
		<i>grayi</i>	71
		<i>grayi lateralis</i>	61
		<i>grayi sulcata</i>	71
		<i>hemisphaerica</i>	61
		<i>irregularis</i>	61
		<i>lamelliformis</i>	77
		<i>levis</i>	27
		<i>malindiae</i>	51

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
	<i>Thenea</i>	<i>megaspina</i>	58
		<i>megastrella</i>	34
		<i>mesotriaena</i>	58
		<i>microclada</i>	34
		<i>microspina</i>	58
		<i>microspirastra</i>	71
		<i>multiformis</i>	58
		<i>muricata</i>	27, 31, 34, 37
		<i>nicobarensis</i>	57
		<i>novaehollandiae</i>	81
		<i>nucula</i>	61
		<i>pendula</i>	51
		<i>pyriformis</i>	77
		<i>rotunda</i>	51
		<i>schmidti</i>	27
		<i>shimodensis</i>	61
		<i>tyla</i>	51
		<i>valdiviae</i>	27
		<i>wrightii</i>	87
		<i>wyvillei</i>	71
	<i>Vulcanella</i>		
		<i>annulata</i>	57
		<i>ornata</i>	27, 34
		<i>schmidti</i>	31
		<i>verruculosa</i>	37
		<i>aberrans</i>	37
		<i>acanthoxea</i>	61
		<i>armata</i>	27
		<i>bifacialis</i>	71
		<i>cribrifera</i>	34
		<i>cribriporosa</i>	61
		<i>doederleini</i>	61
		<i>gracilis</i>	27, 34
		<i>horrida</i>	27, 31, 34
		<i>linaresi</i>	27
		<i>netheides</i>	61
		<i>orthotriaena</i>	81
		<i>osculanigera</i>	77
		<i>porosa</i>	61
		<i>theneides</i>	51
		<i>tricornis</i>	87

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Thrombidae</b>	<i>Thrombus</i>	<i>abyssi</i>	27
		<i>abyssi niger</i>	27
		<i>challengeri</i>	71
		<i>kitttoni</i>	31
		<i>ornatus</i>	51

**APPENDIX 3****Hadromerida**

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Stylocordylidae</b>			
	<i>Stylocordyla</i>		
		<i>borealis</i>	18, 21, 27, 61
		<i>borealis acuata</i>	(48, 88)
		<i>borealis eos</i>	18, 61
		<i>borealis globosa</i>	58
		<i>borealis irregularis</i>	(48, 88)
		<i>borealis typica</i>	27
		<i>chupachups</i>	41, (48, 88)
		<i>fragilis</i>	81
		<i>longissima</i>	18, 61
		<i>muta</i>	31
		<i>pellita</i>	27
<b>Suberitidae</b>			
	<i>Homaxinella</i>		
		<i>balfourensis</i>	(48, 88), 58
		<i>subdola</i>	18, 27, 61
	<i>Rhizaxinella</i>		
		<i>arborescens</i>	61
		<i>australiensis</i>	51
		<i>biseta</i>	27
		<i>burtoni</i>	61
		<i>cervicornis</i>	61
		<i>clava</i>	31
		<i>clavata</i>	61
		<i>dichotoma</i>	71
		<i>durissima</i>	81
		<i>elevata</i>	61
		<i>elongata</i>	27
		<i>gadus</i>	67
		<i>gracilis</i>	37
		<i>incrassata</i>	61
		<i>nuda</i>	71
		<i>pyrifera</i>	37
		<i>ramulosa</i>	51, 77
		<i>ramulosa cilidifera</i>	77
		<i>schaudinni</i>	18
		<i>shikmonae</i>	37
		<i>spiralis</i>	87
		<i>uniseta</i>	27
<b>Tethyidae</b>	<i>Halicometes</i>		

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>cometes</i>	31
		<i>elongata</i>	27
		<i>hooperi</i>	71
		<i>minuta</i>	41
		<i>pediculata</i>	47
		<i>stellata</i>	31
		<i>stonei</i>	57
		<i>thyris</i>	31

**APPENDIX 4****Lithistida**

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Scleritodermidae</b>			
	<i>Aciculites</i>		
		<i>ciliata</i>	71
		<i>cribrophora</i>	31
		<i>higginsi</i>	31
		<i>manawatawhi</i>	81
		<i>mediterranea</i>	37
		<i>orientalis</i>	57, 71
		<i>oxytylota</i>	71
		<i>papillata</i>	71
		<i>pulchra</i>	81
		<i>spinosa</i>	51
		<i>sulcus</i>	81
		<i>tulearensis</i>	51
	<i>Amphibleptula</i>		
		<i>madrepora</i>	31
	<i>Microscleroderma</i>		
		<i>chonelleides</i>	61
		<i>herdmani</i>	57, 71
		<i>hirsutum</i>	58
		<i>lamina</i>	37
		<i>novaehelandiae</i>	81
		<i>spirophora</i>	34
		<i>stoneae</i>	71
	<i>Scleritoderma</i>		
		<i>camusi</i>	71
		<i>cyanea</i>	31
		<i>flabelliforme</i>	71, 81
		<i>nodosum</i>	71
	<i>Setidium</i>		
		<i>obtectum</i>	31
<b>Corallistidae</b>			
	<i>Awhiowhio</i>		
		<i>osheai</i>	81
		<i>sepulchrum</i>	81
		<i>unda</i>	81
	<i>Corallistes</i>		
		<i>aculeata</i>	51
	<i>Corallistes</i>	<i>australis</i>	71
		<i>bispiraster</i>	41

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>bowerbanki</i>	34
		<i>elegantior</i>	27
		<i>elegantissima</i>	51
		<i>fulvodesmus</i>	71, 81
		<i>isabela</i>	87
		<i>masoni</i>	34, 37
		<i>microstylifer</i>	71
		<i>microtuberculatus</i>	34
		<i>multituberculatus</i>	71
		<i>paratypus</i>	31
		<i>thomasi</i>	71
		<i>tubulatus</i>	31
		<i>typus</i>	31
		<i>undulatus</i>	71
		<i>verrucosa</i>	51
	<b><i>Herengeria</i></b>		
		<i>auriculata</i>	71, 81
		<i>vasiformis</i>	71, 81
	<b><i>Isabella</i></b>		
		<i>mirabilis</i>	71
	<b><i>Neophrissospongia</i></b>		
		<i>nana</i>	37
		<i>nolitangere</i>	27, 31
	<b><i>Neoschrammeniella</i></b>		
		<i>antarctica</i>	(48, 88), 81
		<i>castrum</i>	71
		<i>moreti</i>	71
		<i>norfolfki</i>	71
<b>Pleromidae</b>			
	<b><i>Anaderma</i></b>		
		<i>rancureli</i>	71
	<b><i>Pleroma</i></b>		
		<i>aotea</i>	81
		<i>menoui</i>	71, 81
		<i>torquilla</i>	31
		<i>turbinatum</i>	71, 81
	<b><i>Discodermia</i></b>		
		<i>aspera</i>	51
		<i>calyx</i>	61
		<i>claviformis</i>	71
	<b><i>Discodermia</i></b>	<i>discifera</i>	51, 57
		<i>discifurca</i>	81
		<i>dissoluta</i>	31
		<i>dubia</i>	51

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>emarginata</i>	51
		<i>emarginata</i>	71
		<i>gorgonoides</i>	57
		<i>inscripta</i>	31
		<i>interspersa</i>	57
		<i>irregularis</i>	61
		<i>japonica</i>	61
		<i>jogashima</i>	61
		<i>kiiensis</i>	61
		<i>laevidiscus</i>	51
		<i>natalensis</i>	51
		<i>ornata</i>	
		<i>panoplia</i>	71
		<i>papillata</i>	51
		<i>polydiscus</i>	31
		<i>proliferans</i>	71, 81
		<i>ramifera</i>	27
		<i>simillima</i>	
		<i>sinuosa</i>	51
		<i>stylifera</i>	51
		<i>tuberosa</i>	51
		<i>vermicularis</i>	61
		<i>verrucosa</i>	31, 34
<b>Theonellidae</b>			
	<i>Manihinea</i>		
		<i>conferta</i>	51
	<i>Racodiscula</i>		
		<i>asteroides</i>	31
		<i>clava</i>	27, 31, 34
		<i>incrustans</i>	51
		<i>sceptrellifera</i>	51
		<i>sceptrellifera</i>	51
		<i>sceptrellifera</i>	51
		<i>spinispirulifera</i>	51
	<i>Siliquariaspongia</i>		
		<i>japonica</i>	61
	<i>Theonella</i>		
		<i>annulata</i>	34
	<i>Theonella</i>	<i>atlantica</i>	31
		<i>complicata</i>	51
		<i>conica</i>	51, 71
		<i>cupola</i>	51
		<i>cylindrica</i>	71
		<i>ferruginea</i>	

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>incerta</i>	71
		<i>invaginata</i>	71
		<i>lacerata</i>	71
		<i>levior</i>	57
		<i>mirabilis</i>	71
		<i>pulchrifolia</i>	51, 57
		<i>swinhoei</i>	51, 57, 61
		<i>swinhoei var verrucosa</i>	71
<b>Siphoniidae</b>			
	<i>Gastrophanella</i>		
		<i>cavernicola</i>	41
		<i>implexa</i>	31, 41
		<i>mammilliformis</i>	47
		<i>phoeniciensis</i>	37
		<i>primore</i>	77
	<i>Lithobactrum</i>		
		<i>forte</i>	47
	<i>Siphonidium</i>		
		<i>capitatum</i>	71
		<i>dendyi</i>	57
		<i>dubium</i>	34
		<i>geminum</i>	31
		<i>ramosum</i>	27, 31, 37
<b>Neopeltidae</b>			
	<i>Callipelta</i>		
		<i>cavernicola</i>	51
		<i>mixta</i>	51
		<i>ornata</i>	71
		<i>punctata</i>	71, 81
		<i>sollasi</i>	71
		<i>thoosa</i>	51
	<i>Daedalopelta</i>		
		<i>nodosa</i>	31
	<i>Homophymia</i>		
		<i>lamellosa</i>	51
		<i>pollubrum</i>	71
	<i>Homophymia</i>	<i>stipitata</i>	81
	<i>Neopelta</i>		
		<i>amphiaster</i>	31
		<i>imperfecta</i>	31
		<i>perfecta</i>	31
		<i>plinthosellina</i>	71
		<i>pulvinus</i>	81

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Azoricidae</b>			
	<i>Desmascula</i>		
		<i>desdemona</i>	31
	<i>Jereicopsis</i>		
		<i>graphidophora</i>	71
	<i>Leiodermatium</i>		
		<i>colini</i>	81
		<i>crassiusculum</i>	
		<i>dampieri</i>	81
		<i>deciduum</i>	31
		<i>heteroformis</i>	61
		<i>linea</i>	81
		<i>lynceus</i>	27, 31, 34, 51
		<i>marginatum</i>	71
		<i>paniceus</i>	71
		<i>pfeifferae</i>	27, 31
<b>Desmantidae</b>			
	<i>Desmanthus</i>		
		<i>incrustans</i>	31, 37
		<i>levii</i>	77
		<i>meandroides</i>	41
		<i>rhabdophorus</i>	71
		<i>topsendi</i>	71
	<i>Paradesmanthus</i>		
		<i>macphersoni</i>	47
	<i>Petromica</i>		
		<i>ciocalyptoides</i>	31
		<i>citrina</i>	41
		<i>pacifica</i>	71
		<i>digitata</i>	47
		<i>grimaldii</i>	27, 37
		<i>massalis</i>	57
		<i>plumosa</i>	47
		<i>tubulata</i>	47
	<i>Sulcastrella</i>	<i>clausa</i>	31
		<i>leviorum</i>	31
		<i>tenens</i>	31, 37
<b>Vetulinidae</b>			
	<i>Vetulina</i>		
		<i>stalactites</i>	31
<b>Phymatellidae</b>			
	<i>Neoaulaxinia</i>		
		<i>clavata</i>	71, 81

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>persicum</i>	81
		<i>zingiberadix</i>	81
	<i>Neosiphonia</i>		
		<i>fruticosa</i>	71
		<i>motukawanui</i>	81
		<i>schmidti</i>	31
		<i>superstes</i>	71, 81
	<i>Reidispongia</i>		
		<i>coerulea</i>	71, 81
		<i>tuberculata</i>	71
<b>Isoraphiniidae</b>			
	<i>Costifer</i>		
		<i>vasiformis</i>	71
		<i>wilsoni</i>	71, 81
<b>Macandrewiidae</b>			
	<i>Macandrewia</i>		
		<i>auris</i>	47
		<i>azorica</i>	27, 34
		<i>clavatella</i>	31
		<i>nodosa</i>	31
		<i>ramosa</i>	27
		<i>rigida</i>	71
		<i>robusta</i>	27
		<i>spinifoliata</i>	71, 81
<b>Phymaraphiniidae</b>			
	<i>Exsuperantia</i>		
		<i>clava</i>	31
	<i>Kaliapsis</i>		
		<i>cidaris</i>	71
		<i>incrustans</i>	51
		<i>permollis</i>	51
	<i>Lepidothenea</i>		
		<i>incrustans</i>	81

## APPENDIX 5

## Poecilosclerida

Family	Genus	Species	FAO Major Fishing Areas
<b>Cladorhizidae</b>			
	<i>Asbestopluma</i>		
		<i>agglutinans</i>	87
		<i>pennatula</i>	18, 27,
		<i>formosa</i>	71
		<i>stylivarians</i>	34
		<i>belgicae</i>	(48, 88)
		<i>bilamellata</i>	71
		<i>biserialis</i>	71
		<i>biserialis californiana</i>	77
		<i>callithrix</i>	(48, 88)
		<i>calyx</i>	(48, 88)
		<i>comata</i>	27
		<i>cupressiformis</i>	18, 27
		<i>flabellum</i>	61
		<i>furcata</i>	27
		<i>globularis</i>	77
		<i>gracilior</i>	31
		<i>gracilis</i>	61
		<i>hadalis</i>	81
		<i>hydra</i>	27
		<i>hypogea</i>	37
		<i>infundibulum</i>	18, 27
		<i>infundibulum orientalis</i>	61
		<i>lebedi</i>	61
		<i>lycopodium</i>	18, 21, 27, 61
		<i>minuta</i>	27
		<i>occidentalis</i>	67
		<i>quadririalis</i>	27
		<i>ramosa</i>	61
		<i>robusta</i>	18
		<i>wolffi</i>	81
	<i>Chondrocladia</i>		
		<i>occulta</i>	67
		<i>stipitata</i>	58
		<i>tasmaniensis</i>	57
		<i>turbiformis</i>	81
		<i>albatrossi</i>	41
		<i>amphactis</i>	31
	<i>Chondrocladia</i>	<i>antarctica</i>	(48, 88)
		<i>arctica</i>	27

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>arenifera</i>	61
		<i>asigmata</i>	81
		<i>burtoni</i>	34
		<i>clavata</i>	71
		<i>concrescens</i>	31
		<i>crinita</i>	71
		<i>dichotoma</i>	51
		<i>fatimae</i>	58
		<i>gigantea</i>	21, 27
		<i>gracilis</i>	51
		<i>guiteli</i>	27
		<i>koltuni</i>	61
		<i>lampadiglobus</i>	87
		<i>levii</i>	47
		<i>magna</i>	61
		<i>michaelsarsi</i>	34
		<i>multichela</i>	51
		<i>nani</i>	58
		<i>nicolae</i>	47
		<i>nucleus</i>	27
		<i>pulvinata</i>	71
		<i>scolionema</i>	71
		<i>vaceleti</i>	47
		<i>verticillata</i>	31
		<i>virgata</i>	27, 37
		<i>yatsui</i>	61
	<i>Cladorhiza</i>		
		<i>abyssicola</i>	27, 34
		<i>arctica</i>	18,
		<i>bathycrinoides</i>	61
		<i>corona</i>	67
		<i>corticocancellata</i>	27
		<i>depressa</i>	71
		<i>ephyrula</i>	47
		<i>flosabyssi</i>	34
		<i>fristedti</i>	21
		<i>gelida</i>	18, 21, 27
		<i>grandis</i>	21,
		<i>grimaldii</i>	34
		<i>inversa</i>	41
	<i>Cladorhiza</i>	<i>linearis</i>	81
		<i>longipinna</i>	77
		<i>mani</i>	(48, 88)
		<i>methanophila</i>	31

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>microchela</i>	61
		<i>mirabilis</i>	87
		<i>moruliformis</i>	57
		<i>nematophora</i>	57
		<i>nobilis</i>	27,
		<i>oxeata</i>	27,
		<i>pteron</i>	77
		<i>rectangularis</i>	61
		<i>schistochela</i>	71
		<i>segonzaci</i>	77
		<i>septemdentalis</i>	61
		<i>similis</i>	77
		<i>tenuisigma</i>	18, 27
		<i>thomsoni</i>	47
		<i>tridentata</i>	21
<b>Acarnidae</b>			
	<b><i>Iophon</i></b>		
		<i>abnormalis</i>	(48, 88), 58
		<i>aceratum</i>	(48, 88)
		<i>cheliferum</i>	21, 47, 58, 67
		<i>chilense</i>	87
		<i>cylindricum</i>	81
		<i>dubium</i>	18, 21, 27
		<i>flabellodigitatum</i>	(48, 88)
		<i>flabellodigitatum gaussi</i>	(48, 88)
		<i>frigidum</i>	18, 21, 27
		<i>frigidum gracile</i>	18,
		<i>gaussi</i>	(48, 88)
		<i>hesperidesi</i>	(48, 88)
		<i>indentatum</i>	77
		<i>lamella</i>	77
		<i>lamella indivisa</i>	77
		<i>laminale</i>	58
		<i>omnivorus</i>	81
		<i>ostiamagna</i>	77
		<i>piceum</i>	18, 27, 61
		<i>piceum abipocillus</i>	61
		<i>piceum orientale</i>	61
	<b><i>Iophon</i></b>	<i>piceum pacificum</i>	18,
		<i>pluricorne</i>	(48, 88)
		<i>pluricorne trulliferum</i>	(48, 88)
		<i>proximum</i>	41, (48, 88), 57, 58, 87
		<i>radiatum</i>	(48, 88) 87
		<i>spatulatum</i>	41, (48, 88)

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>terranovae</i>	(48, 88)
		<i>unicorn</i>	(48, 88), 58
<b>Esperiopsidae</b>			
	<i>Esperiopsis</i>		
		<i>bathyalis</i>	41
		<i>challengeri</i>	71
		<i>challengeri meangensis</i>	71
		<i>chindoensis</i>	61
		<i>decora</i>	27
		<i>desmophora</i>	71
		<i>diasolenia</i>	71
		<i>flagellum</i>	21, 27
		<i>flagrum</i>	67
		<i>flava</i>	71
		<i>heardi</i>	58
		<i>incognita</i>	27
		<i>inodes</i>	71
		<i>koltuni</i>	61
		<i>lesliei</i>	47
		<i>lingua</i>	61
		<i>macrosigma</i>	27
		<i>macrosigma novaezealandiae</i>	81
		<i>magnifolia</i>	71
		<i>megachela</i>	81
		<i>plumosa</i>	61
		<i>polymorpha</i>	27,
		<i>praedita</i>	27
		<i>profunda</i>	58
		<i>pulchella</i>	58, 71
		<i>rugosa</i>	41, 47, (48, 88), 58, 87
		<i>scotiae</i>	(48, 88)
		<i>stipula</i>	18, 61
		<i>strongylophora</i>	37
		<i>symmetrica</i>	58
		<i>varia</i>	87
		<i>villosa</i>	18, 27, (48, 88)
<b>Podospongidae</b>	<i>Podospongia</i>		
		<i>india</i>	31
		<i>loveni</i>	27
		<i>natalensis</i>	51
		<i>similis</i>	71
<b>Isodictyidae</b>	<i>Isodictya</i>		

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>alata</i>	47
		<i>bentarti</i>	(48, 88)
		<i>cavicornuta</i>	(48, 88) , 81
		<i>chichatouzae</i>	47
		<i>delicata megachela</i>	(48, 88)
		<i>doryphora</i>	(48, 88)
		<i>dufresni</i>	58
		<i>echinata</i>	(48, 88)
		<i>erinacea</i>	(48, 88)
		<i>histodermella</i>	18,
		<i>kerguelensis</i>	41, (48, 88) , 58
		<i>lankesteri</i>	(48, 88)
		<i>maeandrina</i>	(48, 88)
		<i>microchela</i>	41, (48, 88)
		<i>obliquidens</i>	41, (48, 88)
		<i>palmata</i>	21, 27
		<i>pulviliformis</i>	61
		<i>quatsinoensis</i>	18, 67
		<i>rigida</i>	18, 67
		<i>setifera</i>	(48, 88)
		<i>spinigera</i>	(48, 88)
		<i>toxophila</i>	(48, 88)
		<i>trigona</i>	(48, 88)
		<i>vancouverensis</i>	67
		<i>verrucosa</i>	(48, 88) , 58

**APPENDIX 6****Halichondrida**

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Axinellidae</b>			
	<i>Axinella</i>		
		<i>anamesa</i>	77
		<i>antarctica</i>	(48, 88)
		<i>arctica</i>	18, 27
		<i>australiensis</i>	
		<i>bidderi</i>	51
		<i>blanca</i>	18, 61
		<i>brondstedi</i>	
		<i>damicornis</i>	27, 31, 37
		<i>elegans</i>	81
		<i>flustra</i>	27
		<i>hirondellei</i>	27
		<i>hispida</i>	18
		<i>infundibuliformis</i>	
		<i>lifouensis</i>	71
		<i>profunda</i>	61
		<i>profunda kurushima</i>	61
		<i>pyramidata</i>	27
		<i>rugosa</i>	18, 27, 61
		<i>setosa</i>	18, 27
		<i>solenoeides</i>	77
		<i>trichophora</i>	18
		<i>vasonuda</i>	27
		<i>vellerea</i>	27
	<i>Phakellia</i>		
		<i>bowerbanki</i>	18, 21, 27
		<i>connexiva</i>	31, 41
		<i>dalli</i>	18, 61, 67
		<i>folium</i>	31
		<i>lambei</i>	61, 67
		<i>lamelligera</i>	77
		<i>plumosa</i>	71
		<i>robusta</i>	18, 27, 37
		<i>stelliderma</i>	71
		<i>sur</i>	41
		<i>ventilabrum</i>	18, 21, 27, 37
<b>Halichondriidae</b>			
	<i>Spongisorites</i>		
		<i>andamensis</i>	57
		<i>annandalei</i>	27
	<i>Spongisorites</i>	<i>arenatus</i>	31

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>coralliphaga</i>	27
		<i>dendyi</i>	27
		<i>difficilis</i>	21, 27
		<i>placenta</i>	27
		<i>ruetzleri</i>	31
		<i>siliquaria</i>	31

**APPENDIX 7****Haplosclerida**

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
<b>Chalinidae</b>			
	<i>Cladocroce</i>		
		<i>fibrosa</i>	27
		<i>gaussiana</i>	(48, 88)
		<i>incurvata</i>	71
		<i>osculosa</i>	27
		<i>parenchyma</i>	21, 27
		<i>spathiformis</i>	27
		<i>ventilabrum</i>	27, (48, 88)
<b>Phloeodictyidae</b>			
	<i>Oceanapia</i>		
		<i>aberrans</i>	81
		<i>aerea</i>	27
		<i>amphirhiza</i>	31
		<i>arcifera</i>	81
		<i>ascidia</i>	31
		<i>atlantica</i>	47
		<i>bacillifera</i>	77
		<i>bartschi</i>	31
		<i>cancap</i>	27
		<i>coriacea</i>	27
		<i>elongata</i>	27
		<i>fibulata</i>	31
		<i>fistulosa</i>	57
		<i>kirkpatricki</i>	(48, 88)
		<i>media</i>	27
		<i>nodulosa</i>	27
		<i>putridosa</i>	27, 81
		<i>reticulata</i>	27
		<i>robusta</i>	18, 21, 27
		<i>tuber</i>	21, 27
		<i>vera</i>	71
		<i>viridescens</i>	31
<b>Petrosiidae</b>			
	<i>Petrosia</i>		
		<i>australis</i>	81
		<i>borealis</i>	67
		<i>brachysclera</i>	71
		<i>canariensis</i>	27
		<i>crassa</i>	27
	<i>Petrosia</i>	<i>pluricristata</i>	71

<b>Family</b>	<b>Genus</b>	<b>Species</b>	<b>FAO Major Fishing Areas</b>
		<i>punctata</i>	71
		<i>raphida</i>	27, 37
		<i>hartmani</i>	31
		<i>mamillata</i>	71
		<i>stoneae</i>	31
		<i>vansoesti</i>	27, 37

This is the report of the FAO Workshop on Deep-sea Species Identification held in Rome, Italy, from 2 to 4 December, 2009. This meeting was organized in response to the need for a strategy for deep-sea species identification guides. The report includes an overview of the presentations and discussions held during the Workshop, and it presents the conclusions and recommendations agreed upon by participants. The three background documents on deep-sea sharks, corals and sponges drafted for the Workshop are included in Part 2 of this report. The Workshop identified specific activities that will be needed to advance the development of the identification guides.

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