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# EVOLUTION AND DEVELOPMENT OF THE SPIKELET AND FLOWER OF *RHYNCHOSPORA* (CYPERACEAE)

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*Premise of research. Rhynchospora*, one of the biggest genera of sedges, displays several spikelet and flower modifications and is one of a few genera where the pollination strategy changes from wind to insect pollination; however, the mechanisms that have promoted such diversity in the genus are unknown. We have performed a comparative study of spikelet and flower development of *Rhynchospora* from an evolutionary perspective to get insight into the bases of spikelet and flower evolution. Particularly, we investigate correlations between the change from wind to insect pollination that occur in the genus and the spikelet and flower traits.

*Methodology.* The spikelet and flower development of nine species of *Rhynchospora* were studied using SEM images. Parsimony, maximum likelihood, and Bayesian reconstruction analyses were conducted to evaluate the evolutionary history of flower and spikelet developmental programs.

*Pivotal results.* We identified seven morphological and developmental traits that vary among *Rhynchospora* species; ancestral character reconstructions show that the spikelet and flower morphology and their developmental pathways support the diversification of *Rhynchospora* lineages.

*Conclusions.* Our results suggest that the evolution of spikelet and flower developmental programs in *Rhynchospora* may be related to the evolution of its pollination type and mating system. The evolution toward a bisexual spikelet and flowers in which the androecium develops faster than the gynoecium may represent important steps for the transition from a wind- to insect-pollination system. In addition, we found new traits of spikelets and flowers that may help to distinguish natural groups within *Rhynchospora*.

Keywords: Rhynchospora, flower, spikelet, development, evolution, Cyperaceae.

Online enhancements: appendixes.

#### Introduction

Angiosperms display a remarkable diversity of flowers. In the past 2 decades, botanists have started to pay particular attention to how the reproductive structures of angiosperms are developed (Endress 1990; Endress and Doyle 2009; Hileman and Irish 2009; Soltis et al. 2009; Crane et al. 2010). The study of the reproductive structures of flowering plants from early developmental stages to maturity has become a valuable tool not only to asses homology questions on morphological evolution but also to understand functional processes (Marazzi et al. 2007; Reinheimer et al. 2010). Moreover, the use of developmental timing events has gained importance in phylogenetics (Laurin and Germain 2011); in this sense, ontogenetic and phylogenetic approaches have been linked in order to assess the evolution of developmental patterns of flowers, spikelets, and inflorescences in angiosperms (Doust and Kellogg 2002; Doust and Drinnan 2004; Kellogg et al. 2004; Bess et al. 2005; Jabbour et al. 2008, 2009; Feng et al. 2011).

Cyperaceae (sedges) are one of the most species-rich families of flowering plants. They are currently divided into two subfamilies, Cyperoideae and Mapanioideae (Muasya et al. 2009). In Cyperoideae, the spikelet may include only one flower (*Ascolepis*; Goetghebeur 1998) or even more than 10 flowers (*Scleria*; Ahumada 2007). The inflorescence of Cyperoideae has been considered a panicle of spikelets (Goetghebeur 1998), where the spikelets are functionally the reproductive unit of the inflorescence. Flowers in Cyperoideae may be bisexual or unisexual (male or female). A typical Cyperoideae bisexual flower consists of two perianth whorls (exceptions occur in tribes Cypereae and Cariceae, the flowers of which lack a perianth; Goetghebeur 1998), an androecium, and a central gynoecium (Bruhl 1991). These whorls are trimerous, like the typical monocot flower, although there are variations that can

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be understood on the basis of this general model (Vrijdaghs et al. 2009). Flowers in Cyperoideae may vary in the morphology and number of perianth parts when present, the number of stigmatic branches, and the number of stamens (Goetghebeur 1986, 1998; Vrijdaghs et al. 2004, 2005*a*, 2005*b*, 2006, 2009); such modifications could alter the common radial symmetry of the flowers to a bilateral symmetry of each whorl (Prenner et al. 2010).

Subfamily Cyperoideae has ~90 genera, the largest being Carex (more than 1828 species) and Cyperus (more than 702 species; Govaerts et al. 2013); Rhynchospora Vahl, with ~354 species, is the third-largest genus in subfamily Cyperoideae and is mainly distributed in the American savannas (Koyama 1972; Strong 2006; Thomas et al. 2009). The current classification of the genus follows Kükenthal (1949, 1950, 1951, 1952). Kükenthal recognized two subgenera based on the division of the style: a shallowly divided style for Rhynchospora subgenus Haplostyleae and a deeply divided style for Rhynchospora subgenus Diplostyleae (today's subgenus Rhynchospora). Other characters commonly used to circumscribe the species of Rhynchospora were the inflorescence type, the presence/absence of perianth, and the shape and size of the fruit (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1992; Strong 2006). Recently, in a preliminary analysis of the Rhynchosporeae based on molecular data (trnL intron and trnL-F intergenic spacers), Kükenthal's infrageneric division based on the morphology of the style was not supported by the phylogeny (Thomas et al. 2009). According to these results, Rhynchospora may be paraphyletic relative to the genus Pleurostachys (member of the Rhynchosporeae tribe), and the subgenus Haplostyleae (Kükenthal 1949, 1950, 1951, 1952) appears to be divided into several smaller clades instead of being a natural group. Consequently, it was suggested that the clades of Rhynchospora should be recognized as several monophyletic genera (Thomas et al. 2009). Nevertheless, until now, no nomenclatural change has been made for the genus Rhynchospora, and therefore the classification of Kükenthal (1949, 1950, 1951, 1952) is still used.

In terms of reproductive structures, *Rhynchospora* shows a large variation in its inflorescence forms, spikelet and flower morphology, and pollination system. The inflorescence of *Rhynchospora* may be paniculate, capitate, or even reduced to a single spikelet (Kükenthal 1949, 1950, 1951, 1952; Koya-



**Fig. 1** Simplified schemes of *Rhynchospora* spikelet types based on differences in sexual systems. All spikelets have two or three basal sterile glumes. Andromonoecious spikelets (A) consist of a proximal bisexual flower and three to five distal male flowers. Unisexual spikelets (U) have a proximal female flower and three distal male flowers. Bisexual spikelets (B) bear five to six bisexual flowers exclusively.

ma 1972; Thomas 1992; Guaglianone 2001; Strong 2006; Lucero and Vegetti 2012). Each spikelet has two or three basal sterile glumes and several distal fertile glumes. Two distinct spikelet types have been described for the genus: (1) a spikelet that bears exclusively bisexual flowers and (2) a spikelet that consists of a proximal bisexual flower and several distal male flowers (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1992; Guaglianone 2001; Strong 2006). A typical flower of Rhynchospora has a perianth with an outer and an inner whorl of three tepals each (some species may have flowers without a perianth), an androecium consisting of one to three stamens, and a gynoecium with two stigmatic branches (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1992; Guaglianone 2001; Strong 2006). In addition, the genus is characterized by a persistent style base in the fruit (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1992; Guaglianone 2001; Strong 2006).

In regard to the pollination mode, in general, the species of the genus *Rhynchospora* are pollinated by wind or sometimes by insects (Thomas 1984). In this sense, the genus *Rhynchospora* is one of a few genera of Cyperaceae with entomophilous

Species	No. basal sterile glumes	Sexual system	No. stamens at the bisexual flower	Perianth	No. stigmatic branches	Style length	Relative rates of Rhynchospora androecium/gynoecium elongation
R. brittonii	2	В	1	Present	2	Short	Type 3
R. consanguinea	2	А	2	Present	2	Long	Type 2
R. corymbosa	3	Α	3	Present	2	Long	Type 1
R. pilosa	2	А	2	Absent	1	Long	Type 2
R. riedeliana	2	Α	2	Absent	2	Long	Type 2
R. scutellata	3	Α	3	Present	2	Long	Type 2
R. setigera	3	В	3	Absent	2	Short	Type 4
R. tenuis	3	В	3	Absent	2	Short	Type 4
R. terminalis	2	U	NA	Present	2	Long	NA

 Table 1

 Morphological and Developmental Characters of Spikelets and Flowers of Rhynchospora

Note. A, andromoecy; B, bisexual; U, unisexual; NA, not applicable. See text for further explanation of the four relative rates of androecium/gynoecium elongation.



**Fig. 2** Spikelet development in *Rhynchospora terminalis* as an example of spikelet development in *Rhynchospora*. *A*, Two basal sterile glumes (Gl 1 and Gl 2), with distichous arrangement, are the first organs to be initiated. The floral meristem is emerging in the axil of the third glume (Gl 3), which arises opposite the fourth glume (Gl 4). The rachilla apex (indicated by an asterisk) is visible between the floral meristem and Gl 4. *B*, When the central ovule and the ovary wall of the most proximal flower start to differentiate, two other flower primordia begin their development at the axil of the fifth (Gl 5) and sixth (Gl 6) glumes. Fertile glumes are spirally located (cf. the position of Gl 4 to Gl 7). F = floral meristem (F2, second flower; F3, third flower; F4, fourth flower), Gl = glume, o = ovule, ov = ovary wall, pi = inner perianth part, po = outer perianth part. Scale bars: *A*, *B* = 50  $\mu$ m.

species derived from anemophilous species; other examples were recorded in Cyperus, Carex, Ficinia, and in some Mapanioideae (Wragg and Johnson 2011), given that the ancestral condition of the family is wind pollination (Thomas 1984; Goetghebeur 1998; Givnish et al. 2010; Wragg and Johnson 2011). Species that were recorded to be visited by insects occur in Rhynchospora section Dichromena, whose capitate inflorescences are visited by flour beetles, syrphid flies, and bees (Thomas 1984); species of Rhynchospora section Dichromena have white glumes and sticky pollen, which are traits linked to entomophilous pollination (Thomas 1984). There are no reports of other members of genus Rhynchospora s.s. being insect pollinated. Recently, Wragg and Johnson (2011) found that pollen motility, floral scents, and inflorescence color are traits correlated with the transition from wind pollination to insect pollination in Cyperus. In general, in angiosperms, it is

well documented that floral architecture; sexual system, stamen, and stigma characteristics; and the protandrous or protogynous condition of bisexual flowers are also traits (hereafter meaning the state of a given character) associated with changes in the pollination mode (Sargent and Otto 2004; Friedman and Barrett 2009; Givnish et al. 2010); however, the relationship between these traits and the evolution of an insect-pollination mode has not been examined in Cyperaceae. Given the variability of reproductive structures and the pollination modes mentioned earlier, genus *Rhynchospora* s.l. represents an interesting opportunity to look for correlations between the evolution of the spikelet, floral traits, and pollination mode.

*Rhynchospora* species display several spikelet and flower modifications; however, the mechanisms that have produced such diversity in the genus are unknown, as are their homologies. Furthermore, the correlation between the evolution of spikelet and floral developmental traits of *Rhynchospora* and the transition toward an insect-pollination mode never was explored. To fill these gaps, we determined to (1) investigate the mechanisms that have promoted the morphological diversity of flowers and spikelets in the genus, (2) test the homology of the characters found in the genus, and (3) investigate the relationship between the floral and spikelet traits and pollination mode of *Rhynchospora* species. In order to achieve these goals, we undertook a comparative study of spikelet and flower development using SEM and studied their evolution in a molecular phylogenetic context.

## Material and Methods

#### Developmental Studies (SEM Images)

Nine species were selected depending on material availability, including at least one representative of all the major clades of the genus *Rhynchospora*, according to the most recent phylogeny (Thomas et al. 2009). Four of the species studied here were courtesy of Shirley Martins (Universidade Estadual Paulista, São Paulo, Brazil; app. A; see also app. B, available online). Inflorescences at different stages of development were collected from the field, fixed in FAA (formaldehyde–ethanol– acetic acid–water, 10 : 50 : 5 : 35 v/v) solution for 24 h, and then transferred to 70% ethanol.

Spikelets were dissected under a Nikon SMZ-10 stereoscopic microscope to remove glumes and expose the floral organs. The dissected spikelets were dehydrated in an increasing series of ethanol, up to absolute ethanol. The material was then transferred to pure acetone for desiccation using CO<sub>2</sub> in a critical-point dryer (Emitech K850). Mounted samples were coated with gold/palladium using a Thermo VG Scientific Polaron SC7620 sputter-coater (Zürich) or with gold using a Model 3 Polaron 91000 sputter-coater (Pelco) and then photographed using a Philips XL30 Series (Eindhoven, the Netherlands) scanning electron microscope at the Museo Argentino de Ciencias Naturales Bernardino Rivadavia in Buenos Aires and an EVO40 VP scanning electron microscope (Cambridge) at the Centro Científico Tecnológico Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) Bahía Blanca in Buenos Aires, respectively.

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**Fig. 3** Floral development in *Rhynchospora corymbosa* as an example of floral development in *Rhynchospora*. Three basal sterile glumes were removed. *A*, The initiation of a flower begins with the development of a dome-shaped meristem that arises at the axil of a fertile glume. *B*, Two lateral stamen primordia are the first floral organs to initiate. *C*, Then, a third stamen primordium and the outer whorl of perianth primordia are initiated; outer perianth primordia develop opposite and below the stamens. *D*, Later, the gynoecium primordium appears simultaneously with the inner perianth (arrows); inner perianth primordia develop between stamens. *E*, Then, the ovule and the ovary wall start to differentiate. *F*, The ovary encloses the ovule. *G*, The style elongates surpassing the stamens. *H*, Two shallow stigmas appear. *I*, The style continues growing, whereas stigmas remain brief. *J*, In advanced stages of development, the perianths became scabrid. a = anther, c = connective, f = filament, F = flower meristem, g = gynoecium, Gl = glume, o = ovule, ov = ovary wall, pi = inner perianth part, po = outer perianth part, s = stamen, st = style, sg = stigma, asterisk = rachilla apex. Scale bars: *A*, *F* = 20 µm; *B* = 30 µm; *C*-*E*, *G*, *H* = 50 µm; *I*, *J* = 100 µm.

#### DNA Extraction and Sequencing

Total DNA was extracted from silica-dried tissue using the DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions or the CTAB method (Doyle and Doyle 1987). The plastid trnL (UAA) intron and the intergenic spacer between the trnL (UAA) 3' exon and the trnF (GAA) gene (trnL-F region) were amplified from total DNA using the polymerase chain reaction (PCR) with universal primers c and f (Taberlet et al. 1991). PCR reactions were performed in a final volume of 25  $\mu$ L containing 0.15  $\mu$ L of Tag polymerase (Universidad Nacional de Quilmes), 2.5 µL  $10 \times$  buffer, 2.5  $\mu$ L 5 mM MgCl<sub>2</sub>, 1.5  $\mu$ L 10  $\mu$ M of each primer, 0.25 mM of each dNTP, and 0.75 µL of dimethyl sulfoxide. Amplifications were performed in a TGradient Thermocycler (Biometra, Göttingen, Germany), with an initial denaturation for 5 min at 94°C, followed by 36 cycles at 94°C for 30 s, 48°C for 1 min, 72°C for 1.5 min, and a final extension at 72°C for 10 min.

PCR products were cleaned and sequenced by Macrogen (Seoul) using the ABI PRISM BigDye Terminator Cycle Sequencing Kits with AmpliTaq DNA polymerase (Applied Biosystems, Seoul). Single-pass sequencing was performed using the same primers used for PCR reactions. The sequences were assembled and edited in BioEdit, version 7.0.9.0 (Hall 1999). Sequences were aligned using MUSCLE, version 3.8 (Edgar 2004*a*; 2004*b*), and inspected by eye. GenBank accession numbers are provided in appendixes A, B. Alignment and phylogenetic trees were submitted to TreeBASE (accession no. \$13566).

## Phylogenetic Analysis

The outgroup taxa were selected based on the most recent molecular phylogeny of Cyperaceae (Muasya et al. 2009). Three species were included as outgroup taxa, *Cladium mariscus*, *Scleria distans*, and *Schoenus nigricans*; their sequences were retrieved from GenBank (DQ058298, DQ058299, and AJ295814, respectively). Maximum likelihood (ML) and Bayesian (BI) analyses implemented the GTR+G model of evolution, based on tests for the best-fit model using the Akaike information criterion (AIC) executed in jModelTest 0.1.1 (Posada 2008).

ML analyses were performed using PhyML, version 3.0 (Guindon and Gascuel 2003; Guindon et al. 2010), with a BIONJ (an improved version of the neighbor-joining algorithm) tree as a starting point, using nearest neighbor interchange moves for tree searching. Nodal robustness on the ML tree was estimated by the nonparametric bootstrap (BS) with 1000 replicates.

Bayesian phylogenetic estimates were run for 6 million generations in MrBayes, version 3.1.2 (Huelsenbeck and Ronquist 2001). Four Markov chains were run simultaneously in two independent runs, starting with a random tree. Trees were sampled every 1000 generations. Convergence diagnostics for log-likelihood values were assessed visually using Tracer, version 1.5.0 (Rambaut and Drummond 2007). Burn-in for each run was determined independently by plotting –log likelihood against the number of generations. To calculate the Bayesian posterior probabilities (PP), trees prior to stationarity were excluded, and the remaining trees (11,981 trees) were used to generate a 50% majority rule consensus tree.

#### Character Evolution

In order to gain insight into the evolution of the *Rhynchospora* spikelet and flower morphology, we performed ancestral character-state reconstruction studies using the phylogeny generated in this work. We chose several qualitative characters and character states based on SEM observations; to check the suitability of the characters, we examined several specimens (apps. A, B). Although some of the characters studied here were previously used for species description and/or classification (as characters 1, 3, and 4; Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1984, 1992; Guaglianone 2001; Strong 2006), they were never tested properly in a phylogenetic framework as they are here. The seven characters selected and their states were as follows:

1. Number of basal sterile glumes (state 0: two; state 1: three).

2. Spikelet type according to sexual systems (state 0: andromonoecious system; state 1: bisexual system; state 2: unisexual system).

3. Number of stamens in bisexual flowers (state 0: three; state 1: two). Our observations agree with previous literature, except for the number of stamens of flowers of *Rbynchospora brittonii*. Although we observed only one stamen in flowers of *R. brittonii*, we coded this character as polymorphic according to Guaglianone (1980), who found one, two, or even three stamens depending on the studied individual.

4. Presence of flowers with perianth along the spikelet (state 0: all flowers with perianth; state 1: flowers without perianth; state 2: only the proximal flower has perianth).

5. Style length (state 0: long; state 1: short). Our interpretation of the style length differs from previous treatments in that the gynoecium with short style has been considered a twobranched style (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1984, 1992; Guaglianone 2001; Strong 2006). In our study, we considered the style as undivided, differing only in length; the long style is almost double the size of the ovary, while the short style is shorter than the ovary.

6. Number of stigmatic branches (state 0: three; state 1: two; state 2: one at maturity).

7. Relative rates of androecium/gynoecium elongation (state 0: androecium never surpassing the gynoecium [types 1 and 2]; state 1: androecium surpassing the gynoecium sometime during development [types 3 and 4]). The four relative rates of androecium/gynoecium elongation were coded as a binary character (character 7).

Data for outgroup species were obtained from the literature (*S. nigricans*, Vrijdaghs et al. 2007; *C. mariscus*, Vrijdaghs 2006). Given that there are no published ontogenetic reports on *Scleria* species, we used the available morphological description of the adult spikelet of *S. distans* (Ahumada 2007). The inapplicable data were coded as missing data. The data set is presented in appendix C, available online.

Ancestral state reconstruction of spikelet and floral traits was performed using BI, ML, and parsimony methods. The use of different methods for character reconstruction is a common practice to avoid relying on only one method. We use the

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**Fig. 4** Variations on the program of *Rhynchospora* flower development. Basal sterile glumes were removed. *A*, The male flower of *Rhynchospora corymbosa* develops three stamens, whereas the gynoecium and perianths never initiate. *B*, The female flower of *R. terminalis* does not develop stamens. C, The bisexual flower of *R. riedeliana* develops two lateral stamens, but there is no trace of the third stamen or perianths

ML tree generated by PhyML for maximum likelihood and parsimony reconstructions. For the Bayesian analysis, which is the only method that incorporates phylogenetic uncertainty into the analysis, we used BayesTraits, version 1.0 (Pagel and Meade 2006); we included the 11,981 trees obtained with MrBayes. First, we used a maximum likelihood approach to get an approximation of the rate parameters. Second, reversible-jump Markov chain Monte Carlo Bayesian analyses were performed for 10 million generations (sampling every 500 generations), setting values from 0 to 15 as priors for the exponential distribution of the rate parameters and values from 60 to 90 for the ratedev parameter. These settings allowed us to obtain acceptance rates of 20%-40%, as recommended by the authors. Tracer 1.5 (Rambautand and Drummond 2007) was used to check for adequate effective sample sizes (ESS) for all model parameters. Finally, we compared the resulting Bayes factor scores to generate an estimate for the relative support of each alternative hypothesis, using the fossil command implemented in BayesTraits, version 1.0.

Parsimony and maximum likelihood reconstructions were performed using Mesquite, version 2.75 (Maddison and Maddison 2010). The difference between these reconstruction methods is that parsimony incorporates polymorphic states, while maximum likelihood does not. We used the maximum likelihood tree generated by PhyML for this purpose. For the maximum likelihood approach, we used the Markov one-rate (Mk1) model. Characters were treated as unordered for the parsimony analysis.

#### Results

Scanning electron photographs that illustrate in detail the spikelet and flower development of each of the species studied are provided in appendixes A, B. Given space constraints, only the most illustrative scanning electron photographs are presented below. Table 1 summarizes the diversity of *Rhynchospora* spikelet and flower morphology and development found in this study.

### Spikelet Morphology

In the genus *Rhynchospora*, spikelets vary in the number of sterile glumes and the sexuality of the flowers (table 1). Spikelets bear two or three basal sterile glumes depending on the species (e.g., two in *Rhynchospora terminalis* or three in *R. corymbosa*) and several distal fertile glumes. All *Rhynchospora* species examined are monoecious. We found three different spikelet types based on the sex of the flowers (fig. 1). In general, spikelets may have a proximal bisexual flower and three to five distal male flowers (andromonoecious system, A; e.g., *R.* 

*corymbosa*). On the other hand, the spikelet of *R. terminalis* bears a proximal female flower and three distal male flowers (unisexual system, U); this spikelet type is reported here for the first time in *Rhynchospora*. Sometimes spikelets have five to six bisexual flowers (e.g., *R. tenuis*; bisexual system, B).

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#### Spikelet and Flower Development

The spikelet consists of an indeterminate rachilla, along which glumes and flowers develop acropetally (fig. 2). In all the species examined, the basal empty glumes are distichously arranged (fig. 2A) and the distal fertile glumes are placed in a spirodistichous arrangement (fig. 2B). The initiation of a flower primordium begins with a dome-shaped meristem (fig. 3A). The development of a bisexual perfect flower begins with the initiation of two lateral stamen primordia (fig. 3B). Later, the third abaxial stamen arises simultaneously with the outer perianth primordia, which are placed opposite the stamens (fig. 3C). Next, the gynoecium primordium and the inner perianth whorl appear (fig. 3D). During ovule and ovary wall differentiation, the rest of the perianth parts of the inner perianth whorl initiate (fig. 3E). Then, the ovule is covered by the ovary wall, while the stamens and the perianth continue their expansion (fig. 3F). Later, the style elongates (fig. 3G) and two short stigmas initiate (fig. 3H, 3I). Finally, the perianth parts enlarge and become scabrid at the apex (fig. 31).

#### Variation of Floral Developmental Patterns

We found variations on the developmental pattern described above among members of Rhynchospora. Such differences include (1) variations in the number of developed stamens, (2) presence/absence of perianth, (3) number of stigmatic branches, (4) style length, and (5) symmetry of the flower. Male flowers develop three stamens (two lateral stamens and one adaxial and retarded stamen), whereas the gynoecium and perianth are never initiated (fig. 4A). In female flowers, the gynoecium and perianth are formed, but stamens are never initiated (fig. 4B). Bisexual flowers may develop all three stamens (e.g., R. corymbosa; fig. 4A), two lateral stamens (e.g., R. riedeliana; fig. 4C), or only one lateral stamen (R. brittonii; fig. 4D; we did not find rudiments of aborted stamens in those species with a reduced number of such organs. In addition, in some of the species (R. pilosa, R. riedeliana, R. setigera, and R. tenuis), bisexual flowers do not develop perianth (fig. 4C). In the bisexual flower of R. consanguinea, one of the outer perianth parts aborts, leaving a scar visible at advanced developmental stages (fig. 4E, 4F; apps. A, B).

The gynoecium of *Rhynchospora* develops two stigmatic branches, except in *R. pilosa*, which has one stigmatic branch at maturity. Developmental images showed that the stigma of

parts. D, The bisexual flower of R. brittonii has a single stamen and a fully developed whorl of perianths. E, Both perianth whorls of the R. consanguinea flower initiate at an early stage of development. F, One of the outer perianth parts of the R. consanguinea flower arrests its development, leaving a scar (marked with a dagger) at an advanced stage of development. G, The stigma of the R. pilosa flower initially has two branches. H, One of the stigma branches of R. pilosa elongates faster than the other. I, One of the stigma branches of the R. pilosa flower continues the differentiation at advanced stages of development. J, The gynoecium of R. scutellata has a long style and two brief stigmatic branches. K, The gynoecium of R. setigera has a short style and two long stigmatic branches. a = anther, f = filament, Gl = glume, o = ovule, ov = ovary wall, pi = inner perianth part, po = outer perianth part, s = stamen, sg = stigma, st = style, asterisk = rachilla apex, dagger = aborted perianth part. Scale bars: A, G = 30  $\mu$ m; B, F = 100  $\mu$ m; C, E = 20  $\mu$ m; D, H = 50  $\mu$ m; I, K = 300  $\mu$ m; J = 200  $\mu$ m.



**Fig. 5** Schematic representation of variability in the symmetry of *Rhynchospora* flowers. The floral whorl where the symmetry is bilateral is indicated.

*R. pilosa* is first divided into two branches. Later on, one of the stigmatic branches elongates, whereas the other one remains short (fig. 4G, 4H); finally, only one stigma completes its development (fig. 4I). The style may be long (e.g., *R. scutellata*; fig. 4J) or short (e.g., *R. setigera*; fig. 4K).

The symmetry of male flowers is radial. Bisexual and female flowers have bilateral symmetry in the fourth whorl (e.g., *R. tenuis*) given the number of stigmatic branches. In some cases, the third whorl possesses bilateral symmetry given the reduction in the stamen number (e.g., *R. pilosa* and *R. brittonii*). In *R. consanguinea*, the first whorl possesses bilateral symmetry due to the abortion of one perianth part (fig. 5).

#### Relative Rates of Androecium/Gynoecium Elongation

Four different relative rates of androecium/gynoecium development were recognized based on variations of the timing of stamen elongation in comparison with the gynoecium enlargement (table 1). In most flowers, the gynoecium elongates faster than the androecium from early developmental stages (e.g., R. corymbosa; fig. 3C-3I; type 1). In the flower of R. scutellata, elongation of the gynoecium and the androecium occur at a similar rate; only later does elongation of the gynoecium surpass that of the androecium (fig. 6A-6D; type 2). In R. brittonii, the androecium initially surpasses the gynoecium, and then the gynoecium accelerates its growth to surpass the stamens (fig. 6E-6I; type 3). Finally, in some species, something similar occurs to what is observed in R. brittonii, but in these cases the difference between the stamen elongation and the gynoecium elongation is greater; this situation is also inverted after the stamens are fully elongated, when the gynoecium elongates to surpass the androecium (R. tenuis and R. setigera; fig. 6J-6M; type 4).

#### Phylogenetic Analysis and Character Evolution

The phylogeny presented here agrees with the topology published by Thomas et al. (2009; fig. 7). The genus Rhynchospora is monophyletic (PP = 1 and BS = 100; note that we did not include any species of *Pleurostachys* in the analysis), with two main clades that diverged earlier, clade B and E, highly (PP = 1 and BS = 100) and weakly (BS = 58) supported, respectively. Clade B includes only species of subgenus Haplostyleae and is further divided into two subclades containing species R. corymbosa and R. scutellata in clade C (PP = 1 and BS =99) and R. terminalis and R. consanguinea in clade D (PP = 0.96 and BS = 87). Clade E includes some species of subgenus Haplostyleae and all species of the subgenus Rhynchospora; this clade is further divided into subclades F and H. Clade F includes R. brittonii, R. setigera, and R. tenuis (PP = 0.95 and BS = 78), which are representatives of subgenus Rhynchospora. Clade H (PP = 1 and BS = 100) consists of two members of subgenus Haplostyleae (R. riedeliana and R. pilosa). The majority rule consensus tree of the BI produced an identical topology. In general, the topology is supported with BS > 78% and PP > 0.96 values, excepting clade E, which is always weakly supported no matter what method is employed.

In general, the results of the ancestral state reconstruction studies, using parsimony, maximum likelihood, and Bayesian methods, were similar; however, we found a few differences in the timing of some of the evolutionary events and the resolution of the reconstruction, depending on the method used. Ancestral character-state reconstructions are summarized below.

Number of basal sterile glumes. Character 1 (fig. 8A; app. C)—The presence of two basal sterile glumes (state 0) is the most likely state at the base of *Rhynchospora*. Spikelets with three basal sterile glumes (state 1) may appear twice: before the divergence of *R. terminalis*, *R. consanguinea*, *R. scutellata*,



**Fig. 6** Relative rates of androecium/gynoecium elongation of *Rhynchospora* flowers. Types 2, 3, and 4 are documented here, whereas type 1 is illustrated in fig. 3D-3J. Basal sterile glumes were removed. A, Early stage of flower development of *Rhynchospora scutellata*. B, C, Middle developmental stages of the *R. scutellata* flower, in which the androecium and the gynoecium are similar in length. D, Late developmental stage of *R. scutellata*, where the gynoecium surpasses the length of the androecium. E, Early stage of flower development of *R. brittonii*, where stamens surpass the gynoecium of *R. brittonii* surpasses the androecium. J, Early stage of flower development of *R. setigera*, where the stamens are longer than the gynoecium. K, At middle stages of *R. setigera* flower development, the androecium continues to surpass the gynoecium in length. L, When stamens of the *R. setigera* flower are fully elongated, the gynoecium remains shorter than the androecium. M, Finally, at mature stages, the gynoecium of the *R. setigera* flower elongates to surpass the androecium. g = gynoecium, ov = ovary wall, pi = inner perianth part, po = outer perianth part, s = stamen, sg = stigma, st = style, asterisk = rachilla apex. Scale bars: A, B = 30  $\mu$ m; C = 20  $\mu$ m; D, F–H = 100  $\mu$ m; E, J, K = 50  $\mu$ m; I, M = 500  $\mu$ m; L = 300  $\mu$ m.



**Fig. 7** Maximum likelihood tree of *Rhynchospora* obtained with PhyML 3.0. Bayesian posterior probabilities (>95) and likelihood bootstrap support values (>50) are shown below and above branches, respectively. Letters indicate clade names as assigned in this work.

and *R. corymbosa* (node B) and sometime during the diversification of subgenus *Rhynchospora* (node F or node G). Similar results were obtained under parsimony reconstructions; however, parsimony reconstructions predict that the origin of three basal glumes took place at the base of *R. tenuis* and *R. setigera* and before the diversification of *R. scutellata* and *R. corymbosa* (nodes G and C, respectively).

Spikelet type based on sexual systems. Character 2 (fig. 8B; app. C)—Bayesian and maximum likelihood reconstructions show that bisexual spikelets (state 1) represent the most likely state at the base of *Rhynchospora*. Bisexual spikelets were exclusively kept by subgenus *Rhynchospora* (node E and clade F). Andromonoecious spikelets (state 0) may have evolved two independent times: before the divergence of *R. terminalis*, *R. consanguinea*, *R. scutellata*, and *R. corymbosa* (node B) and at the base of the *R. riedeliana* and *R. pilosa* clade (node H). Parsimony reconstruction shows an opposite history, in which the andromonoecious spikelet may be the ancestral state for *Rhynchospora*, whereas the bisexual spikelet evolved once at the base of subgenus *Rhynchospora*. This character showed no homoplasy under parsimony reconstruction.

The unisexual spikelet appears at least once in *R*. *terminalis* from an andromonoecious spikelet.

Number of stamens in the bisexual flower. Character 3 (fig. 8C; app. C)—Three stamens (state 0) is the ancestral state reconstructed using a Bayes approach. Bisexual flowers with two stamens evolved at least twice: before the divergence of *R. riedeliana* and *R. pilosa* (node H) and in *R. consanguinea*. The bisexual flower with one stamen has arisen at least once in *R. brittonii* from flowers with three stamens. Under the parsimony method, the reconstruction of the nodes of interest is uncertain. The maximum likelihood method cannot be used for the reconstruction of this character due to the polymorphic nature of *R. brittonii*.

Presence of flowers with perianth along the spikelet. Character 4 (fig. 8D; app. C)—Spikelets that have flowers without perianth (state 1) represent the most likely state at the base of *Rhynchospora* (node A), as well as at node E. Inside *Rhynchospora*, the perianth appears twice but in different spikelet types. Spikelets with perianth in the proximal flower and distal flowers without perianth (state 2) appear at node B, whereas spikelets that have all flowers with perianth are exclusive of



Fig. 8 Ancestral character-state reconstruction of *Rhynchospora* spikelet and flower morphology and development. Character names and their states are indicated in shaded boxes. Species mapped with gray circles represent missing data. We used the maximum likelihood tree of

*R. brittonii* (state 0). Parsimony reconstruction at the base of *Rhynchospora* is uncertain; for the rest of the nodes, the reconstruction is identical to the Bayesian and maximum likelihood analyses.

*Style length.* Character 5 (fig. 8*E*; app. C)—Bayesian and maximum likelihood methods indicate that *Rhynchospora* ancestors have flowers with a short style. Long styles appear twice at nodes B and H. Parsimony reconstruction is uncertain for nodes A and E, so the direction of the evolution of the style length cannot be assessed.

Number of stigmatic branches. Character 6 (fig. 8F; app. C)—The three different methods employed confirm a reduction in the number of stigmatic branches. Three stigmatic branches (state 0) is the ancestral state of the subfamily Cyperoideae. The number of stigmatic branches has been reduced to two (state 1) at the base of *Rhynchospora* and later to one (state 2) in *R. pilosa*.

*Relative rates of androecium/gynoecium elongation.* Character 7 (fig. 8*G*; app. C)—At nodes A, E, and F, flowers with an androecium that surpasses the gynoecium sometime during flower development (state 1) represent the most likely state, according to the Bayesian and maximum likelihood approaches. The three reconstruction methods agree that the most likely state at nodes B and H is a flower in which the androecium never surpasses the gynoecium (state 0). Parsimony reconstructions are ambiguous at the base of *Rhynchospora*; on the other hand, this method indicates that an androecium that surpasses the gynoecium sometime during flower development (state 1) is a synapomorphy of node F.

# Discussion

In *Rhynchospora*, the spikelet consists of an indeterminate rachilla bearing lateral flowers in the axil of glumes. Recently, Vrijdaghs et al. (2010) addressed the structure of the spikelet based on an ontogenetic approach in subfamily Cyperoideae (Cyperaceae) and concluded that the spikelet is indeterminate (monopodial); our observations on *Rhynchospora* agree with this model of spikelet development. Furthermore, our results contrast with the conclusion of Browning and Gordon-Gray (1995); these authors, studying mature spikelets of *Rhynchospora gracillima* ssp. *subquadrata*, suggest that flowers are not axillary to glumes.

The spikelet of subfamily Cyperoideae has been described as having sterile glumes distichously or spirally arranged depending on the genus (Eiten 1976; Goetghebeur 1998; Vrijdaghs et al. 2010). The spikelet of Rhynchospora may have two or three basal sterile glumes and several distal fertile glumes. In systematic treatments of *Rhynchospora*, the glume arrangement in the spikelet has been considered spiral (Thomas 1992; Guaglianone 2001; Strong 2006); however, we found variations in the glume phyllotaxis within the spikelet. Interestingly, in Rhynchospora, basal sterile glumes develop distichously, whereas fertile glumes switch to a spirodistichous disposition. A similar phenomenon of glume phyllotaxis shift along the same spikelet rachis was found in Scleria (Ahumada and Vegetti 2009) and Abildgaardia ovata (Reutemann et al. 2009). An inverse case was observed in Scirpus falsus (Vrijdaghs et al. 2010; now named Dracoscirpoides falsa [Muasya



**Fig. 8 (continued)** fig. 7 for character optimization. Pie charts indicate the proportional likelihood obtained under the Markov one-rate model using Mesquite, version 2.01. Character 3 was reconstructed under only the parsimony criterion using Mesquite, version 2.01, due to the presence of polymorphic states. For the rest of the characters, the corresponding parsimony reconstruction is presented in appendix C, available online. Posterior probabilities (values over 20%) obtained with BayesTraits are shown at nodes of interest. The Bayes factor was performed to test the support of one particular state over the other state(s): three asterisks (BF values >5) indicate strong support; two asterisks (BF values >2) indicate positive support; one asterisk (BF values 2 > 1) indicates low support. Letters indicate node names as in fig. 7.



**Fig. 9** Summary of *Rhynchospora* spikelet and flower character evolution. Characters that became synapomorphic under any of the reconstruction methods employed are indicated. Gray lines indicate those states inferred by Bayesian analysis, maximum likelihood, and parsimony, and black lines indicate states inferred by parsimony only. Section *Dichromena* is included in the tree (in blue) based on the topology proposed by Thomas et al. (2009); this section has only insect-pollinated species (Thomas 1984), which is illustrated by a syrphid over the blue branch. The shaded square indicates the hypothetical origin of insect-pollinated species at the base of subgenus *Rhynchospora*. Letters indicate node names as in figure 7.

et al. 2012]), where basal sterile glumes develop spirally, whereas fertile glumes switch to a distichous disposition.

Flowers of subfamily Cyperoideae develop following the scirpoid model (Vrijdaghs et al. 2005a, 2009). Typically, in the scirpoid model, a perfect flower begins as a primordium in the axil of a glume; two lateral stamens are the first organs initiated, followed by a third abaxial stamen. Then, three outer (opposite the stamens) and three inner (alternate with stamens) tepals appear simultaneously with the gynoecium. This tetracyclic scheme with trimerous whorls has a radial symmetry. Vrijdaghs et al. (2005a) found variations to this general model of flower development in Eriophorum and Dulichium due to differences in the tetracyclic scheme. In general, Rhynchospora flowers follow the scirpoid pattern proposed for the subfamily Cyperoideae, except for variations in the symmetry and timing of floral organ development. Rhynchospora flowers with two stigmatic branches or a reduced number of stamens have a bilateral symmetry instead of a radial one, as the model predicts; only Rhynchospora male flowers show a radial symmetry. Interestingly, in Rhynchospora, the outer whorl of perianth initiates before the gynoecium, whereas in the scirpoid model, the gynoecium appears simultaneously with the two whorls of perianth.

## Evolution of Spikelet and Flower Morphology and Development in Rhynchospora

The phylogeny herein produced (fig. 7) is consistent with the phylogeny proposed by Thomas et al. (2009). When the morphological and developmental characters were analyzed on the *Rhynchospora* tree, we found that spikelet and floral traits indeed are consistent with the evolution of several clades. In addition, some of the spikelet and floral character modifications may be correlated with the evolution of *Rhynchospora* mating and pollination systems and fruit dispersal mode, as explained below (fig. 9).

We observed that the variation in the number of basal sterile glumes is correlated with the molecular phylogeny of *Rhynchospora*. The ancestor of the genus may have had a spikelet with two sterile glumes. Later, the number of sterile glumes has increased to three at least two independent times, supporting the origin or diversification of subgenus *Rhynchospora* and the lineage consisting of *R. terminalis*, *R. consanguinea*, *R. scutellata*, and *R. corymbosa*.

One of the main concerns of botanists since Darwin's time has been the changes in the sexuality of flowers and sexual systems of angiosperms. Although it is not clear whether the bisexual flower is the angiosperm ancestral state (Endress and Doyle 2009), it is known that unisexual flowers have originated multiple times, from bisexual flowers, in almost 10% of angiosperms (Barret 2002; Mitchell and Diggle 2005; Diggle et al. 2011). In this work, two different histories on the evolution of flower sexuality were recovered, depending on the methodology. Indeed, Bayesian analyses and maximum likelihood reconstructions estimated that the ancestral state of Rhynchospora is a bisexual spikelet (having only bisexual flowers). By contrast, the parsimony method suggested that the ancestral state of the genus may be an andromonoecious spikelet (consisting of a proximal bisexual flower and several distal male flowers). Several spikelet types, based on the sex of the flowers, have been described for Cyperoideae. Two distinct spikelet types had previously been characterized for Rhynchospora, the andromonoecious and the bisexual (Kükenthal 1949, 1950, 1951, 1952; Koyama 1972; Thomas 1992; Guaglianone 2001; Strong 2006). We have found an additional Rhynchospora spikelet type, named here a unisexual spikelet, that has unisexual flowers exclusively. The ancestral state reconstruction analyses may indicate that the unisexual spikelet evolved only later in the history of the genus from an andromonoecious spikelet; this conclusion needs further corroboration with a broader sampling. While in some of the Cyperoideae genera, one spikelet type predominates (e.g., Cyperus or Eleocharis; Eiten 1976), others such as Rhynchospora, Carex (Goetghebeur 1998), and Scleria (Ahumada and Vegetti 2009) have diversified their spikelet structure, changing the sex of the flowers. Given the uncertainties of the ancestral state reconstruction at the base of the genus, we were not able to infer the direction of the evolution of such changes in Rhynchospora; however, considering the importance of sexual systems for reproduction, we presume that the evolution of the spikelet types of Rhynchospora may denote an evolution of the Rhynchospora pollination mode, mating system, and even fruit dispersal mode.

The loss of stamens seems to be correlated with the sex of the flower of *Rhynchospora*. Bisexual flowers may develop one, two, or three stamens, and male flowers always have three stamens. The reconstruction of this floral trait was equivocal at several nodes. Usually in Cyperoideae, the stamen number does not vary among flowers of different sexes. Few exceptions

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to this general trend have been reported. For instance, the male flower of *Pycreus pumilus* (Vrijdaghs et al. 2009) has more stamens than bisexual flowers. The reduction of stamens in the bisexual flower might possibly also reduce the chance of autogamy and promote outcrossing.

Some of the genera of Cyperaceae are characterized by species that bear flowers with a perianth, whereas other genera or entire tribes have no perianth. It has been postulated that in sedges, the perianth may play an important role in fruit dispersion (Bruhl 1995; Goetghebeur 1998). Only a few genera of sedges, such as Rhynchospora, include species both with and without perianth; other examples were registered in Eleocharis, Fuirena, and Schoenus (Goetghebeur 1998). According to our results, flowers with a perianth evolved at least twice, while the ancestor of Rhynchospora may have flowers lacking a perianth. This character supports the divergence of the two main lineages of Rhynchospora; the presence of perianth at the proximal flower of the spikelet is a synapomorphy of clade B, whereas flowers without perianth belong to clade E. The abortion of only one tepal from the outer whorl in R. consanguinea is another interesting and new finding for Cyperaceae; given that we examined only one specimen, we cannot be certain that this is a stable condition, and further observations in this and other species of the genus and family are needed to confirm the occurrence of organ abortions in Cyperaceae.

In our study, we interpreted the style as short or long and undivided. Here the short style is a synapomorphy of subgenus *Rhynchospora*, and it seems to be correlated with the sexual system. While andromonoecious and unisexual spikelets have flowers with a long style, bisexual spikelets (exclusive of subgenus *Rhynchospora*) develop flowers with a short style. Whether this trait has implications for the mating system in *Rhynchospora* is an open question.

The trimerous condition of floral organs in Cyperoideae has faced reductional trends in several genera (Vrijdaghs et al. 2004, 2005*a*, 2005*b*, 2006, 2009). In *Rhynchospora*, the number of stigmatic branches was reduced to two branches from the Cyperoideae ancestor, which had three stigmatic branches. In this work, we have reported an additional and extreme case of stigmatic branch reduction in *R. pilosa*, in which only one stigmatic branch reaches maturity, while the other one aborts. Similarly, Vrijdaghs et al. (2011) have reported that *Pycreus flavescens* (which is placed in the tribe Cypereae) has two stigmatic branches growing at different rates (one is retarded with respect to the other), but finally both branches complete their development.

Our finding of different relative rates of androecium/gynoecium elongation in *Rhynchospora* stresses the importance of a comparative approach in close relatives, also in Cyperaceae. According to the parsimony reconstruction, an androecium that surpasses the gynoecium sometime during flower development is a synapomorphy of subgenus *Rhynchospora*. Flowers in which the gynoecium is never surpassed by the androecium during development appear two independent times. The flower may be protogynous when the androecium is always shorter than the gynoecium during development, whereas it may be protandrous when the androecium surpasses the gynoecium at some stages of development. Interestingly, protogynous species are associated with wind pollination, whereas protandrous species are commonly animal pollinated (Sargent and Otto 2004; Friedman and Barrett 2008, 2009). Given this, the recognition of this trait may be an important first step toward understanding the pollination system in *Rhynchospora*.

#### Systematic Implications

Although the previous subgeneric classification based on the morphology of the style is not consistent with the phylogeny of Thomas et al. (2009) and the one presented here, our work showed that some morphological and developmental characters are indeed correlated with the molecular phylogeny of *Rhynchospora* (fig. 9). The presence of a perianth in the proximal flower of andromonoecious spikelets at node B and the absence of perianth in all flowers of the spikelets at node E support the divergence of these clades. The short style, the spikelet with only bisexual flowers, and the flowers where the androecium surpasses the gynoecium sometime during development are synapomorphies for the clade that includes *R. brittonii*, *R. setigera*, and *R. tenuis* (node F). We presume that these characters would be of help in a future revision of the systematics of *Rhynchospora*.

## Spikelets and Floral Traits That Are Related with an Entomophilous Pollination in Rhynchospora

The pollination system of Rhynchospora has changed from anemophilous to entomophilous at least once in the derived clade that includes all Dichromena species (Thomas et al. 2009). We found similarities among insect-pollinated Dichromena species and other members of subgenus Rhynchospora studied in this work. Indeed, Dichromena species have bisexual spikelets (producing three or more achenes per spikelet) and have flowers with short styles, such as R. brittonii, R. setigera, and R. tenuis (clade F). Moreover, R. setigera is sister to Dichromena (Thomas et al. 2009), both having capitate inflorescences, white glumes, and flowers without perianth (fig. 9). The floral similarities between Dichromena species and R. brittonii, R. setigera, and R. tenuis indicate that at least part of the subgenus Rhynchospora may have diversified from the rest of Rhynchospora through changes in its spikelet and flower morphology and specialization of its pollination mode. Several floral and developmental traits studied here, such as the presence of a short style, the relative rate of androecium/gynoecium elongation, bisexual spikelets, and flowers without perianth, may support the origin of subgenus Rhynchospora (fig. 9); interestingly, the bisexuality of flowers and the protandrous condition of flowers are related to entomophilous pollination (Sargent and Otto 2004; Friedman and Barrett 2008, 2009; Givnish et al. 2010; Soza et al. 2012). In particular, Wragg and Johnson (2011) suggested that it is not unexpected that insect-pollinated Cyperaceae evolve from wind-pollinated plants that have both sexes in close proximity in space and time; other insect-pollinated Cyperoideae species were reported to have bisexual spikelets (Thomas 1984; Wragg and Johnson 2011); however, it must be remarked that so far the main focus was put on the subfamily Cyperoideae and that subfamily Mapanioideae should receive more attention in the future. In this sense, the presence of bisexual spikelets (in which both sexes are in close proximity) in subgenus Rhynchospora

may also represent a step toward an entomophilous pollination. Consequently, sexual systems in the Cyperaceae spikelet could be considered another relevant trait correlated with the change to entomophilous pollination in the family. According to our observations, the presence of protandrous flowers may occur in some species of subgenus *Rhynchospora* where the androecium develops faster than the gynoecium. The implication of flowers without perianth and the short style in insectpollinated species of Cyperaceae has not been investigated so far (Thomas 1984; Friedman and Barrett 2008; Wragg and Johnson 2011).

#### Concluding Remarks

In this work, we provide sequences of floral organogenesis of nine species of *Rhynchospora* that differ in the presence/ absence of perianth, number of stamens in bisexual flowers, number of stigmatic branches, style length, and sexuality of spikelets. We found four different relative rates of androecium

Voucher information of studied taxa. Specimens are deposited at the "Ingeniero A. Ragonese" herbarium (SF). Taxon: voucher specimen (collection country), accession number. *R. brittonii* Gale: LL 63 (Argentina), KC11177. *R. consanguinea* (Kunth) Boeck.: SM 341 (Brasil), KC111770. *R. corymbosa* (L.) Britton: AGR 140 (Argentina), KC111768; LL 14 (Argentina). *R. pilosa* Boeck.: SM 342 (Brasil),

- Ahumada O 2007 Las especies argentinas y paraguayas del género *Scleria* P. J. Bergius (Cyperaceae): estudio taxonómico y tipología de la sinflorescencia. PhD diss. Universidad Nacional del Litoral, Santa Fe, Argentina.
- Ahumada O, AC Vegetti 2009 Inflorescence structure in species of Scleria subgenus Hypoporum and subgenus Scleria (Sclerieae-Cyperaceae). Plant Syst Evol 281:115–135.
- Barrett SCH 2002 The evolution of plant sexual diversity. Nat Rev Genet 3:274–284.
- Bess EC, AN Doust, EA Kellogg 2005 A naked grass in the "bristle clade": a phylogenetic and developmental study of *Panicum* section *bulbosa* (Paniceae: Poaceae). Int J Plant Sci 166:371–381.
- Browning J, KD Gordon-Gray 1995 Studies in Cyperaceae in southern Africa 28: spikelet morphology in *Rhynchospora gracillima* ssp. *subquadrata*. S Afr J Bot 61:319–324.
- Bruhl JJ 1991 Comparative development of some taxonomically critical floral/inflorescence features in Cyperaceae. Aust J Bot 39:119– 127.
- 1995 Sedge genera of the world: relationships and a new classification of the Cyperaceae. Aust Syst Bot 8:125–305.
- Crane PR, EM Friis, WG Chaloner 2010 Darwin and the evolution of flowers. Philos Trans R Soc B 365:347–350.
- Diggle PK, VS Di Stilio, AR Gschwend, EM Golenberg, RC Moore, JRW Russell, JP Sinclair 2011 Multiple developmental processes underlie sex differentiation in angiosperms. Trends Genet 27:368– 376.
- Doust AN, AN Drinnan 2004 Floral development and molecular phylogeny support the generic status of *Tasmannia* (Winteraceae). Am J Bot 91:321–331.
- Doust AN, EA Kellogg 2002 Inflorescence diversification in the pan-

and gynoecium elongation, which may indicate the presence of protandrous and protogynous species within the genus. The phylogenetic analysis carried out here indicates that some morphological and ontogenetic features of flowers and spikelets support the diversification of several lineages. The bisexual spikelet and flowers in which the androecium develops faster than the gynoecium may represent important steps for the transition from a wind- to insect-pollination system in the genus.

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#### Appendix A

KC111775. **R.** *riedeliana* **C. B.** Clarke: SM 344 (Brasil), KC111776. **R.** *setigera* (Kunth) Boeck.: AGR 151 (Argentina), KC111773; AGR 154 (Argentina); AGR 165 (Argentina). **R.** *scutellata* Griseb: LL 03 (Argentina), KC111769; AGR 58 (Argentina). **R.** *tenuis* Link: AGR 131 (Argentina), KC111774; LL 20 (Argentina). **R.** *terminalis* (Nees) Steud: SM 343 (Brasil), KC111771.

## Literature Cited

icoid "bristle grass" clade (Paniceae, Poaceae): evidence from molecular phylogenies and developmental morphology. Am J Bot 89: 1203–1222.

- Doyle JJ, JL Doyle 1987 A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull 19:11–15.
- Edgar RC 2004*a* MUSCLE: a multiple sequence alignment method with reduced time and space complexity. BMC Bioinform 5:113.
- 2004b MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res 32:1792–1797.
- Eiten LT 1976 Inflorescence units in the Cyperaceae. Ann Mo Bot Gard 63:81–112.
- Endress PK 1990 Patterns of floral construction in ontogeny and phylogeny. Biol J Linn Soc 39:153–175.
- Endress PK, JA Doyle 2009 Reconstructing the ancestral angiosperm flower and its initial specializations. Am J Bot 96:22–66.
- Feng C, Q Xiang, RG Franks 2011 Phylogeny-based developmental analyses illuminate evolution of inflorescence architectures in dogwoods (*Cornus* s.l., Cornaceae). New Phytol 191:850–869.
- Friedman J, SCH Barrett 2008 A phylogenetic analysis of the evolution of wind pollination in the angiosperms. Int J Plant Sci 169: 49–58.
- 2009 Wind of change: new insights on the ecology and evolution of pollination and mating in wind-pollinated plants. Ann Bot 103:1515–1527.
- Givnish TJ, M Ames, JR McNeal, MR McKain, PR Steele, CW Depamphilis, SW Graham, et al 2010 Assembling the tree of the monocotyledons: plastome sequence phylogeny and evolution of poales. Ann Mo Bot Gard 97:584–616.

Goetghebeur P 1986 Genera Cyperacearum: een bijdrage tot de ken-

nis van de morfologie, systematiek en fylogenese van de Cyperaceae genera. PhD thesis. Groep Plantkunde, Rijksuniversiteit Gent.

- 1998 Cyperaceae. Pages 141–190 *in* K Kubitzki, ed. The families and genera of vascular plants. Vol 4. Springer, Berlin.
- Govaerts R, J Dransfield, SF Zona, DR Hodel, A Henderson 2013 World checklist of Cyperaceae. Facilitated by the Royal Botanic Gardens, Kew. http://apps.kew.org/wcsp/.
- Guaglianone ER 1980 Contribución al estudio del género Rhynchospora Vahl (Cyperaceae) II. Darwiniana 22:499-510.
- 2001 Contribución al estudio del género Rhynchospora (Cyperaceae) V. Sección Longirostres en América austral. Darwiniana 39:287–342.
- Guindon S, JF Dufayard, V Lefort, M Anisimova, W Hordijk, O Gascuel 2010 New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol 59:307–321.
- Guindon S, O Gascuel 2003 A simple, fast and accurate algorithm to estimate large phylogenies by maximum likelihood. Syst Biol 52: 696–704.
- Hall TA 1999 BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symp Ser 41:95–98.
- Hileman LC, VF Irish 2009 More is better: the uses of developmental genetic data to reconstruct perianth evolution. Am J Bot 96:83–95.
- Huelsenbeck JP, F Ronquist 2001 MrBayes: Bayesian inference of phylogenetic trees. Bioinformatics 17:754–755.
- Jabbour F, C Damerval, S Nadot 2008 Evolutionary trends in the flowers of Asteridae: is polyandry an alternative to zygomorphy. Ann Bot 102:153–165.
- Jabbour F, LP Ronse De Creane, S Nadot, C Damerval 2009 Establishment of zygomorphy on an ontogenic spiral and evolution of perianth in the tribe Delphinieae (Ranunculaceae). Ann Bot 104: 809–822.
- Kellogg EA, KM Hiser, AN Doust 2004 Taxonomy, phylogeny, and inflorescence development of the genus *Ixophorus* (Panicoideae: Poaceae). Int J Plant Sci 165:1089–1105.
- Koyama T 1972 The botany of the Guayana highland. IX. *Cypera*ceae-Rhynchosporeae and Cladieae. Mem N Y Bot Gard 23:23-89.
- Kükenthal G 1949 Vorarbeiten zu einer Monographie der Rhynchosporoideae. Bot Jahrb Syst 74:375-509.
- 1950 Vorarbeiten zu einer Monographie der Rhynchosporoideae. Bot Jahrb Syst 75:90—195.
- 1951 Vorarbeiten zu einer Monographie der Rhynchosporoideae. Bot Jahrb Syst 75:273–314.
- 1952 Vorarbeiten zu einer Monographie der Rhynchosporoideae: pleurostachys. Bot Jahrb Syst 75:451–497.
- Laurin M, D Germain 2011 Developmental characters in phylogenetic inference and their absolute timing information. Syst Biol 60: 630–644.
- Lucero LE, AC Vegetti 2012 Inflorescence structure in *Rhynchospora* Vahl (Cyperaceae). Flora 207:47–56.
- Maddison WP, DR Maddison 2010 Mesquite: a modular system for evolutionary analysis. Version 2.74. http://mesquiteproject.org.
- Marazzi B, E Conti, PK Endress 2007 Diversity in anthers and stigmas in the buzz-pollinated genus *Senna* (Leguminosae, Cassiinae). Int J Plant Sci 168:371–391.
- Mitchell CH, PK Diggle 2005 Evolution of unisexual flowers: morphological and functional convergence results from diverse developmental transitions. Am J Bot 92:1068–1076.
- Muasya AM, M Reynders, P Goetghebeur, DA Simpson, A Vrijdaghs 2012 Dracoscirpoides (Cyperaceae): a new genus from Southern Africa, its taxonomy and floral ontogeny. S Afr J Bot 78:104–115.
- Muasya AM, DA Simpson, GA Verboom, P Goetghebeur, RFC Naczi, MW Chase, E Smets 2009 Phylogeny of Cyperaceae based on DNA sequence data: current progress and future prospects. Bot Rev 75: 2–21.

- Pagel M, A Meade 2006 Bayesian analysis of correlated evolution of discrete characters by reversible-jump Markov chain Monte Carlo. Am Nat 167:808–825.
- Posada D 2008 jModelTest: phylogenetic model averaging. Mol Biol Evol 25:1253–1256.
- Prenner G, RM Bateman, PJ Rudall 2010 Floral formulae updated for routine inclusion in formal taxonomic descriptions. Taxon 59: 241–250.
- Rambaut A, AJ Drummond 2007 Tracer, version 1.5. http://beast .bio.ed.ac.uk/Tracer.
- Reinheimer R, FO Zuloaga, AC Vegetti, R Pozner 2010 Changes in floret development patterns that may correlate with sex determination in the PCK clade (Poaceae). Int J Plant Sci 171:24–33.
- Reutemann AG, NJ Guarise, AC Vegetti 2009 Structure of the inflorescences of selected South American species of *Abildgaardia* Vahl, *Bulbostylis* Kunth, and *Fimbristylis* Vahl (Abildgaardieae-Cyperoideae-Cyperaceae). Plant Syst Evol 283:93–110.
- Sargent RD, SP Otto 2004 A phylogenetic analysis of pollination mode and the evolution of dichogamy in angiosperms. Evol Ecol Res 6:1183–1199.
- Soltis PS, SF Brockington, MJ Yoo, A Piedrahita, M Latvis, MJ Moore, AS Chanderbali, DE Soltis 2009 Floral variation and floral genetics in basal Angiosperms. Am J Bot 96:110–128.
- Soza VL, J Brunet, A Liston, PS Smith, VS Di Stilio 2012 Phylogenetic insights into the correlates of dioecy in meadow-rues (*Thalictrum*, Ranunculaceae). Mol Phylogenet Evol 63:180–192.
- Strong MT 2006 Taxonomy and distribution of *Rhynchospora* (Cyperaceae) in the Guianas, South America. Contrib U S Nat Herb 53:1–225.
- Taberlet P, L Gielly, G Pautou, J Bouvet 1991 Universal primers for amplification of three non-coding regions of chloroplast DNA. Plant Mol Biol 17:1105–1109.
- Thomas WW 1984 The systematics of *Rhynchospora* sect. *Dichromena*. Mem N Y Bot Gard 37:1–116.
- 1992 A synopsis of *Rhynchospora* (Cyperaceae) in Mesoamerica. Brittonia 44:14–44.
- Thomas WW, AC Araujo, MV Alves 2009 A preliminary molecular phylogeny of the Rhynchosporeae (Cyperaceae). Bot Rev 75:22–29.
- Vrijdaghs A 2006 A floral ontogenetic approach to homology questions in non-mapanioid Cyperaceae. PhD thesis. Laboratory of Plant Systematics, K U Leuven, Belgium.
- Vrijdaghs A, P Caris, P Goetghebeur, E Smets 2005a Floral ontogeny in *Scirpus, Eriophorum*, and *Dulichium* (Cyperaceae), with special reference to the perianth. Ann Bot 95:1199–1209.
- Vrijdaghs A, P Goetghebeur, AM Muasya, P Caris, E Smets 2005*b* Floral ontogeny in *Ficinia* Schrad. and *Isolepis* R. Br. (Cyperaceae), with focus on the nature and origin of the gynophore. Ann Bot 96: 1247–1264.
- Vrijdaghs A, P Goetghebeur, AM Muasya, E Smets, P Caris 2004 The nature of the perianth in *Fuirena* (Cyperaceae). S Afr J Bot 70:587– 559.
- Vrijdaghs A, P Goetghebeur, E Smets, P Caris 2007 The Schoenus spikelet: a rhipidium? a floral ontogenetic answer. Aliso 23:204– 209.
- Vrijdaghs A, P Goetghebeur, E Smets, AM Muasya 2006 The floral scales in *Hellmuthia* (Cyperaceae, Cyperoideae) and *Paramapania* (Cyperaceae, Mapanioideae): an ontogenetic study. Ann Bot 98: 619–630.
- Vrijdaghs A, AM Muasya, P Goetghebeur, P Caris, A Nagels, E Smets 2009 A floral ontogenetic approach to homology questions within the Cyperoideae (Cyperaceae). Bot Rev 75:30–51.
- Vrijdaghs A, M Reynders, I Larridon, AM Muasya, P Goetghebeur, E Smets 2011 Morphology and development of spikelets and flowers in *Cyperus* and *Pycreus* (Cyperaceae). Plant Ecol Evol 144:44–63.
- Vrijdaghs A, M Reynders, I Larridon, AM Muasya, E Smets, P Goetghebeur 2010 Spikelet structure and development in Cyperoideae

(Cyperaceae): a monopodial general model based on ontogenetic evidence. Ann Bot 105:555–571.

Wragg PD, SD Johnson 2011 Transition from wind pollination to

insect pollination in sedges: experimental evidence and functional traits. New Phytol 191:1128-1140.

## QUERIES TO THE AUTHOR

- **q1.** I edited your article for clarity and agreement with journal style, and minor revisions were made without comment. Please review the text and advise whether any of my changes are unacceptable. Any layout problems or errors you see will be corrected after the proof is returned.
- **q2.** Your appendix 1 was split into print appendix A, containing the voucher information, and online appendix B, containing all of the figures. I have updated each original reference to appendix 1 to include both appendix A and appendix B, but please update as necessary if you do not need to cite both appendixes in each case. Appendix C now contains the character and state information and the accompanying figure.
- **q3.** In the Material and Methods section, please be sure the name of the manufacturer and the manufacturer's location is provided for all of the equipment used.
- **q4.** Figure 7 and table C1 both mention "Cladium marisculus," which appears to be a misspelling. Okay to correct those to match the spelling here, "Cladium mariscus"?
- q5. Should this be "Bayesian inference (BI)"? If not, what does BI stand for?
- **q6.** Is citation of appendixes A and B correct here, or should it also include appendix C?
- **q7.** In the table 1 note, figure 8, and figure C1*B*, should "andromoecy/andomoecious" read "andromonoecy/andromonoecious"?
- **q8.** The figure 3 legend references "arrows" in panel D, although panel D does not include any. Please review and advise re. the appropriate edit.
- **q9.** The figure 9 legend references a "blue branch," but the image is grayscale. How would you like revise the legend? Also, in the graphic, which should it be, "only the proximal flower possesses perianth" or "only the proximal flowers possess perianth"?