


Review

Marine Floral Biodiversity, Threats, and Conservation in Vietnam: An Updated Review

Manh-Linh Nguyen ^{1,2}, Myung-Sook Kim ^{3,*}, Nhu-Thuy Nhat Nguyen ⁴, Xuan-Thuy Nguyen ⁴,
Van-Luong Cao ^{1,*}, Xuan-Vy Nguyen ^{2,4} and Christophe Vieira ^{3,*}

- ¹ Institute of Marine Environment and Resources, VAST, Hai Phong 180000, Vietnam
² Faculty of Ecology and Biological Resources, Graduate University of Science and Technology, VAST, Ha Noi 100000, Vietnam
³ Research Institute for Basic Sciences, Jeju National University, Jeju 63000, Republic of Korea
⁴ Institute of Oceanography, VAST, Nha Trang 650000, Vietnam
* Correspondence: myungskim@jejunu.ac.kr (M.-S.K.); luongcv@imer.vast.vn (V.-L.C.); cvcarp@gmail.com (C.V.)

Simple Summary: The present review provides (1) an updated checklist of the Vietnamese marine flora, (2) a review of molecular-assisted alpha taxonomic efforts, (3) an analysis of marine floral biodiversity spatial distribution nationally and regionally (South China Sea), (4) a discussion on the impact of anthropogenic and environmental stressors on the Vietnamese marine flora, and (5) the efforts developed in the last decade for its conservation. The updated checklist consists of 878 species, including 439 Rhodophyta, 156 Ochrophyta, 196 Chlorophyta, 87 Cyanobacteria, and 15 seagrasses. The South Central Coast supports the highest species diversity of marine algae, which coincides with the largest density of coral reefs along the Vietnam coast. Vietnam holds one of the richest marine floras in the South China Sea owing to the country's coastline length and associated marine habitat diversity. However, the Vietnamese marine floral biodiversity is facing critical threats, and present management efforts are yet insufficient for their conservation. A methodical molecular-assisted re-examination of Vietnam marine floral biodiversity is urgently needed, complemented with in-depth investigations of the main threats targeted against it; and finally, conservation measures should be urgently implemented.

Abstract: Part of the Indo-Chinese peninsula and located on the northwest edge of the Coral Triangle in the South China Sea, the Vietnamese coastal zone is home to a wealthy marine biodiversity associated with the regional geological setting and history, which supports a large number of marine ecosystems along a subtropical to tropical gradient. The diversity of coastal benthic marine primary producers is also a key biological factor supporting marine biological diversity. The present review provides: (1) an updated checklist of the Vietnamese marine flora, (2) a review of molecular-assisted alpha taxonomic efforts, (3) an analysis of marine floral biodiversity spatial distribution nationally and regionally (South China Sea), (4) a review of the impact of anthropogenic and environmental stressors on the Vietnamese marine flora, and (5) the efforts developed in the last decade for its conservation. Based on the studies conducted since 2013 and the nomenclatural changes that occurred during this period, an updated checklist of benthic marine algae and seagrasses consisted in a new total of 878 species, including 439 Rhodophyta, 156 Ochrophyta, 196 Chlorophyta, 87 Cyanobacteria, and 15 phanerogam seagrasses. This update contains 54 new records and 5 new species of macroalgae. The fairly poor number of new records and new species identified in the last 10 years in a "mega-diverse" country can be largely attributed to the limited efforts in exploring algal biodiversity and the limited use of genetic tools, with only 25.4% (15 species) of these new records and species made based on molecular-assisted alpha taxonomy. The South Central Coast supports the highest species diversity of marine algae, which coincides with the largest density of coral reefs along the Vietnamese coast. Vietnam holds in the South China Sea one of the richest marine floras, imputable to the country's geographical, geological, and climatic settings. However, Vietnam marine floral biodiversity is under critical threats examined here, and current efforts are insufficient for its conservation. A



Citation: Nguyen, M.-L.; Kim, M.-S.; Nguyen, N.-T.N.; Nguyen, X.-T.; Cao, V.-L.; Nguyen, X.-V.; Vieira, C. Marine Floral Biodiversity, Threats, and Conservation in Vietnam: An Updated Review. *Plants* **2023**, *12*, 1862. <https://doi.org/10.3390/plants12091862>

Academic Editors: Andrey Erst and Yevhen Maltsev

Received: 6 March 2023

Revised: 17 April 2023

Accepted: 19 April 2023

Published: 30 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

methodical molecular-assisted re-examination of Vietnam marine floral biodiversity is urgently needed, complemented with in-depth investigations of the main threats targeting marine flora and vulnerable taxa, and finally, conservation measures should be urgently implemented.

Keywords: algae; Cyanobacteria; Rhodophyta; Ochrophyta; Chlorophyta; conservation; East Vietnam Sea; Biển Đông; molecular-assisted alpha taxonomy; seagrass; seaweed; South China Sea

1. Introduction

Located along the eastern margin of the Indo-Chinese Peninsula, on the northwest edge of the Coral Triangle biodiversity hotspot [1], in the South China Sea (also known as the East Vietnam Sea or Biển Đông), the Vietnamese coastal zone is home to a remarkably rich marine biodiversity [2,3]. Vietnam has been listed among the top 25 most biologically diverse countries in the world [4] and characterized by some authors as a “mega-diverse country” [5,6]. The origin of this diversity is linked to the region’s (Southeast Asia) geological and climatic history, the country’s coastline length covering some 3260 km along a north–south orientation, thus with a wide latitudinal range (stretching from 21°30′ N to 8°25′ N), spanning a subtropical–tropical transition zone, which supports no less than 20 types of marine ecosystems [7]. It is nonetheless important to also point to the role of coastal benthic marine primary producers (e.g., algae, corals, seagrasses) as a key biological factor supporting other forms of marine biological diversity [8]. Benthic marine algae occur across virtually all marine coastal systems from intertidal zone to depths of >200 m, on soft (e.g., sandy) to hard (e.g., rocky) substrates, in a variety of habitats (e.g., lagoons, bays, islands, islets, atolls, and reefs) and ecosystems (e.g., mangroves, seagrasses beds, and coral reefs).

The marine flora of Vietnam, which includes three main classes of macroalgae (Chlorophyta, Ochrophyta, Rhodophyta), marine phanerogams or seagrasses (Alismatales), and Cyanobacteria (Cyanophyceae), has attracted the attention of marine botanists since the 1800s. The first mention of Vietnamese seaweeds appeared in the *Flora Cochinchinensis* [9], with the record of 11 names of marine macroalgal species, later referenced in the work of Agardh [10,11] and De Toni [12,13]. The French institution “Institut océanographique de l’Indochine”, corresponding today to the “Institute of Oceanography”, contributed considerably during the 1930s to the knowledge on the algal diversity of Vietnam, notably owing to the work of Pham-Hoàng Hộ [14], considered to be the first Vietnamese marine algal taxonomists of Vietnam. The first collections from the Spratly Islands Archipelago, by the French naturalist Jean Marie Antoine De Lanessan in 1936 (Gouvernement Général de L’Indochine, 1936) [15]—currently housed at the Museum of Oceanography, Nha Trang City—are worthy of mention (Figure 1). Dawson [16] published the very first checklist on the Vietnamese marine flora of Nha Trang Bay in the province of Khanh Hoa (South Central Coast region) with the “Institut Océanographique de Nha Trang”. Dawson [16] reported a total of 204 species (16 Cyanophyceae, 118 Rhodophyta, 22 Ochrophyta, and 48 Chlorophyta), nearly all of which were new species records for Vietnam. Until 1967, the list of marine macroalgae in South Vietnam consisted of 517 species and subspecies [14]. In north of Vietnam, Nguyen et al. [17] provided a list of 281 species. Subsequent studies focused mainly on understudied taxonomic groups and regions of Vietnam, for example, on the families Sargassaceae [18–20] and Halymeniaceae [21,22] and on the genera *Gracilaria* and *Gracilariopsis* [23,24], *Eucheuma* and *Kappaphycus* [25,26], *Dictyota* [27], and *Laurencia* [28,29]. Regions of Vietnam that were later investigated included archipelagoes or offshore islands, such as the Spratly Archipelago [30–34], Ly Son Island [35], Phu Quy Island [36], and some inland coastal sites, such as Hai Van–Son Cha [37], Nha Trang Bay [38], Ninh Thuan [39], and Con Dao Island [40]. Recently, Belous et al. [41] published a checklist of 702 species including Rhodophyta, Ochrophyta, and Chlorophyta recorded from Southern and Central Vietnam (16°12′ southward) [41]. Earlier, in 2013, Nguyen et al. (2013) published the

first comprehensive marine macroalgal checklist for all Vietnam based on a review of 81 books and publications [42]. The checklist contained a total of 827 species, including 412 Rhodophyta, 180 Chlorophyta, 147 Ochrophyta, and 88 Cyanobacteria. The authors suggested that several taxa needed further investigation to better understand their diversity in Vietnamese waters, and noted the need to combine DNA barcoding and morphological observations to resolve or clarify taxonomic uncertainties. Previously, the checklist of seagrass showed 14 species [43]. In addition, *Halophila major* (Zollinger) Miquel was newly recorded in Vietnamese waters in 2013 [44]. Recently, the phylogenetic analysis inferred from genetic marker and morphological observation revealed the putative hybridization form between two species, *H. ovalis* and *H. major* [45]. No new record or new species of Cyanobacteria were made since the work of Pham-Hoàng Hộ [14].

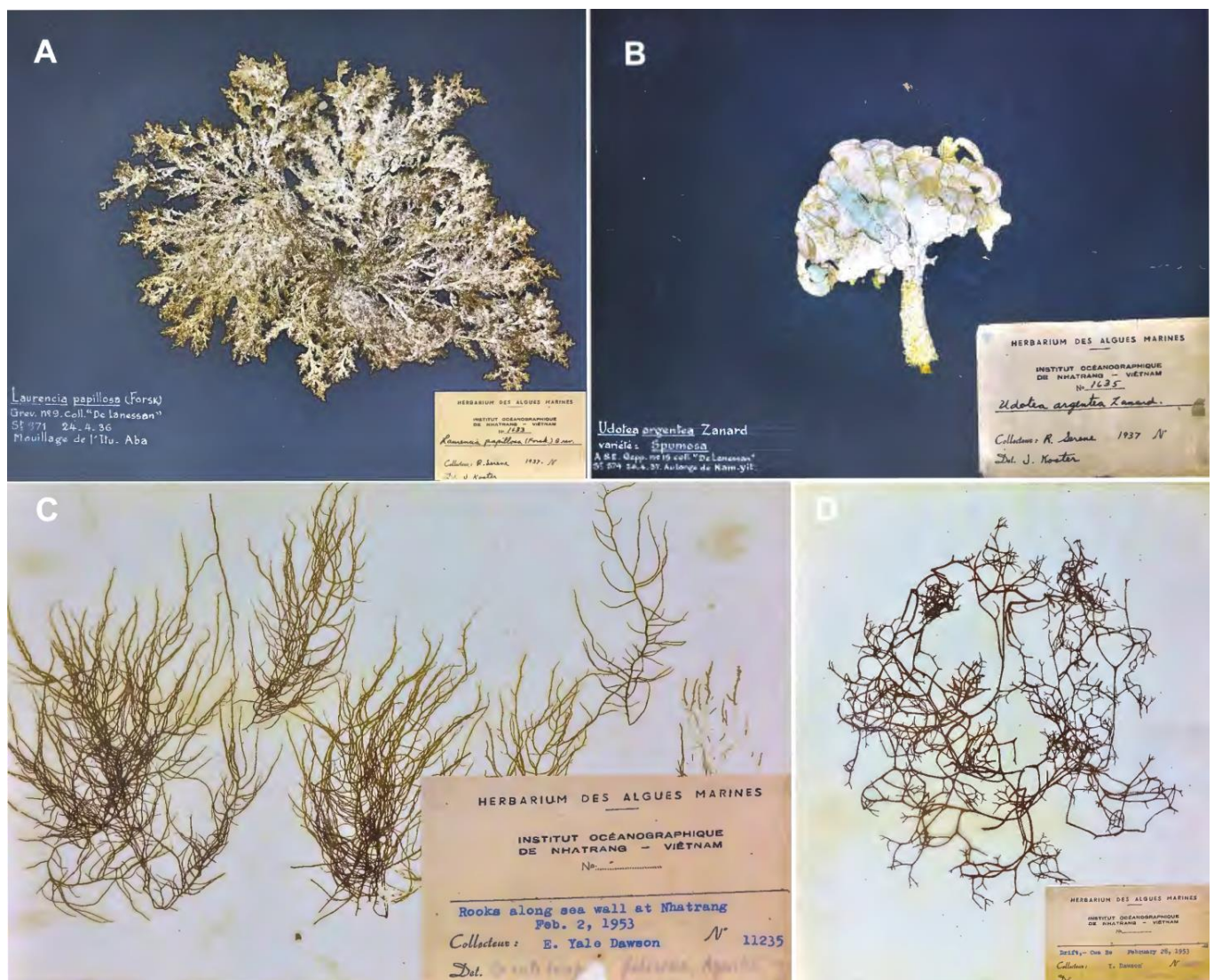


Figure 1. Historical herbarium voucher specimens at Institution of Oceanography—Nha Trang. (A) *Laurencia papillosa*, (B) *Udotea argentea* collected at Spratly Archipelago from De Lanesson [15], (C) *Grateloupia filicina*, and (D) *Chnoospora implexa* collected by Dawson [16] in Nha Trang.

The aims of the present review were multifold: (1) to deliver an updated checklist of the Vietnamese marine flora, (2) to review molecular-assisted alpha taxonomic efforts implemented in its study, (3) to examine marine floral biodiversity spatial patterns across Vietnamese regions and countries in the South China Sea region, (4) to discuss anthro-

pogenic and environmental threats directly affecting Vietnamese marine flora, and (5) the efforts deployed nationally in the last decade for its conservation.

2. Species Diversity Update

Our first objective was to thoroughly review the taxonomic work conducted on the Vietnamese marine flora in the last 10 years since the checklist of Nguyen et al. [42] was published, focusing on 3 main macroalgal phyla/classes (Chlorophyta, Ochrophyta, and Rhodophyta), Cyanobacteria (Cyanophyta), and marine phanerogams or seagrasses (Alismatales). Our literature review of the past 10 years' research on Vietnamese algae and seagrass taxonomy led to the documentation of a total of 54 new records (i.e., species newly recorded from Vietnam in the period between the last checklist and this updated checklist) and 5 new species (i.e., species newly described from Vietnam in the period between the last checklist and this updated checklist) of macroalgae (3 Rhodophyta, 1 Ochrophyta, and 1 Chlorophyta). New species and new records made during the last 10 years are listed in Table 1. No new records of Cyanobacteria or seagrasses were made since 2013. Based on these new data and the nomenclatural changes that occurred in the last 10 years (such as synonymies), we provide an updated checklist of the benthic marine flora, bringing the new total to 878 species of algae in 42 orders, 108 families, and 284 genera, and consisting of 439 Rhodophyta (21 orders, 52 families, and 161 genera), 156 Ochrophyta (7 orders, 13 families, and 36 genera), 196 Chlorophyta (6 orders, 23 families, and 44 genera), 87 Cyanobacteria (8 orders, 18 families, 42 genera), and 15 seagrasses in Alismatales (1 order, 4 families, and 10 genera) listed in Table 2. A total of 112 species of Rhodophyta are updated to the currently accepted names. Three new species and 13 new records were added to the checklist. Four species, including *Gracilaria mammillaris*, *Dasya baillouvoiana*, *Meristotheca papulosa*, and *Mesophyllum erubescens*, were removed from the checklist because they were misidentifications. For Chlorophyta, 17 species were updated to the corrected name; 11 species were newly recorded, mainly in 2 genera, *Ulva* and *Caulerpa*; and only 1 new species was new to science. Two species of Chlorophyta were removed from the checklist: the misspelled name (i.e., orthographic mistake in AlgaeBase) "*Codium tunue*" and *Cladophora adhaerens* Ruprecht, which is an invalidly published name. For Ochrophyta, only 1 new species and 2 new records were added to the checklist, and 7 species were updated to the currently accepted names. Finally, 21 species of Cyanobacteria were updated to currently accepted names. We compared our updated checklist with the data available from AlgaeBase [46] for Cyanobacteria, Rhodophyta, Ochrophyta, and Chlorophyta. AlgaeBase [46] data consisted of a lower number of species with a total of 862 species consisting of 51 Cyanobacteria, 438 Rhodophyta, 171 Ochrophyta, and 202 Chlorophyta. Among 4 phyla, Cyanobacteria showed the lowest richness (87 species, 10%). The checklist of marine macroalgae in Vietnam published by Nguyen et al. [42] did not show any new species and new records of Cyanobacteria [42]. The checklist of marine macroalgae in South Vietnam [41], Nha Trang Bay [38], and Con Dao Island [40] did not list any Cyanobacteria. It indicated that Cyanobacteria have not been studied in detail. Therefore, Cyanobacteria need to be included in future works.

Table 1. List of new species ^(a) and new records ^(b) of marine macroalgae found in Vietnam with support from genetic markers and/or morphological observation since 2013. -/-, as above, na: not available. Ref.: reference.

No.	Phylum	Species	Previous identification in Vietnam	Applied Genetic Markers	Ref.
1 ^b		<i>Amphiroa crassa</i> J.V.Lamouroux		na	[41]
2 ^b		<i>Antithamnion antillanum</i> Børgesen		na	-/-
3 ^b		<i>Antithamnionella breviformis</i> (E.Y.Dawson) Wollaston		na	-/-
5 ^b		<i>Caulacanthus ustulatus</i> (Turner) Kützing		na	-/-
4 ^b		<i>Ceramium borneense</i> Weber Bosse		na	[38]
6 ^b		<i>Chondrophyucus tronoi</i> (Ganzon-Fortes) Nam		<i>rbcL</i>	[47]
8 ^b		<i>Coelothrix irregularis</i> (Harvey) Børgesen		-/-	[48]
7 ^b		<i>Endosiphonia horrida</i> (C.Agardh) P.C.Silva		na	[41]
9 ^b		<i>Gelidium crinale</i> var. <i>perpusillum</i> Piccone & Grunow		na	[38]
10 ^b		<i>Gelidium pusillum</i> var. <i>cylindricum</i> W.R.Taylor		na	[41]
11 ^b		<i>Gelidium pusillum</i> var. <i>pacificum</i> W.R.Taylor		na	[38]
12 ^b		<i>Gloiocladia japonica</i> (Okamura) Yoshida		na	[41]
14 ^a		<i>Gracilaria phuquocensis</i> Le, Muangmai & Zuccarello	<i>Gracilaria mammillaris</i> (Montagne) M. Howe	<i>rbcL</i>	[49]
13 ^b		<i>Gracilaria divergens</i> (C.Agardh) J.Agardh		na	-/-
15 ^b		<i>Halymenia malaysiana</i> Tan, Lim, Lin & Phang		<i>rbcL</i> , COI-5P, 28S	[50]
16 ^b	Rhodophyta	<i>Hypoglossum simulans</i> M.J.Wynne, I.R.Price & D.L.Ballantine		na	[38]
17 ^b		<i>Laurencia natalensis</i> Kylin			[48]
19 ^b		<i>Melanothamnus thailandicus</i> (N.Muangmai & C.Kaewsuralikhit) Díaz-Tapia & Maggs		<i>rbcL</i>	[51]
18 ^a		<i>Meristotheca lysonensis</i> Nguyen, Nguyen, Kittle & McDermid		<i>rbcL</i> , COI-5P	[52]
20 ^b		<i>Parviphycus adnatus</i> (Dawson) Santelices	<i>Gelidiella adnata</i> Dawson	<i>cox1</i> , <i>psaA</i> , <i>rbcL</i>	[53]
21 ^a		<i>Perronella gracilis</i> Boo, Nguyen, Kim & Boo		<i>cox1</i> , <i>psaA</i> , <i>rbcL</i>	-/-
24 ^b		<i>Phycocladia tanegashimensis</i> (I.Shinmura) Santiañez		<i>rbcL</i> , COI-5P, 18S	[54]
22 ^b		<i>Phyllymenia huangiae</i> (Lin & Liang) Lin, Rodríguez-Prieto, De Clerck & Guiry		<i>rbcL</i> , COI-5P	[55]
23 ^b		<i>Phyllymenia taiwanensis</i> (Lin & Liang) Lin, Rodríguez-Prieto, De Clerck & Guiry		<i>rbcL</i> , COI-5P	[56]
25 ^b		<i>Plocamium ovicorne</i> Okamura		na	[41]
26 ^b	<i>Pterocladia bartlettii</i> (Taylor) Santelices		COI-5P, <i>cob</i> , <i>psaA</i> , <i>psbA</i> , <i>rbcL</i>	[57]	
27 ^b	<i>Pterocladia maribagoensis</i> G.H.Boo & P.J.L.Geraldino		-/-	-/-	
28 ^b	<i>Pterocladia musciformis</i> (W.R.Taylor) G.H.Boo & K.A.Miller		-/-	-/-	
29 ^b	<i>Titanophora pikeana</i> (Dickie) Feldmann		-/-	[48]	
30 ^b	<i>Tylopus lichenoides</i> Okamura		-/-	[41]	
31 ^b	<i>Zellera tawallina</i> Martens		<i>rbcL</i> , COI-5P	[55]	

Table 1. Cont.

No.	Phylum	Species	Previous identification in Vietnam	Applied Genetic Markers	Ref.
33 ^b	Ochrophyta	<i>Dictyota grossedentata</i>	De Clerck & Coppejans	<i>rbcL</i> , <i>psbA</i>	[55]
34 ^b		<i>Dictyota hauckiana</i>	Nizamuddin	<i>rbcL</i> , <i>psbA</i>	[58]
32 ^b		<i>Dictyota dichotoma</i> var. <i>intricata</i>	(C.Agardh) Greville	na	[38]
35 ^b		<i>Ectocarpus siliculosus</i>	(Dillwyn) Lyngbye	na	[41]
38 ^b		<i>Lobophora obscura</i>	(Dickie) C.W.Vieira, De Clerck & Payri	na	[59]
36 ^b		<i>Lobophora papenfussii</i>	(W.R.Taylor) Farghaly	-/-	[47]
37 ^a		<i>Lobophora tsengii</i>	Tien & Sun	<i>rbcL</i> , <i>cox3</i>	[60]
39 ^b		<i>Padina japonica</i>	Yamada	na	[41]
40 ^b		<i>Turbinaria turbinata</i>	(Linnaeus) Kuntze	-/-	[41]
41 ^b		<i>Boodleopsis pusilla</i>	(Collins) W.R.Taylor, A.B.Joly & Bernatowicz	-/-	[38]
42 ^b	<i>Caulerpa andamanensis</i>	(Taylor) Draisma, Prudhomme & Sauvage	-/-	[48]	
	Chlorophyta	<i>Caulerpa chemnitzia</i> var. <i>laetevirens</i>	(Montagne) Fernández-García & Riosmena-Rodriguez	-/-	[41]
43 ^b		<i>Caulerpa calcifolia</i>	Harvey & Bailey	-/-	[48]
44 ^b		<i>Caulerpa sertularioides</i> f. <i>longiseta</i>	(Bory) Svedelius	-/-	[41]
45 ^b		<i>Caulerpa oligophylla</i>	Montagne	-/-	[48]
46 ^b		<i>Cladophora gracilis</i>	Kützting	-/-	[41]
47 ^b		<i>Codium intricatum</i>	Okamura	-/-	[38]
48 ^b		<i>Chaetomorpha basiretrorsa</i>	Setchell	-/-	-/-
49 ^b		<i>Parvocaulis exiguus</i>	(Solms-Laubach) S.Berger, Fettweiss, Gleissberg, Liddle, U.Richter, Sawitzky & Zuccarello	-/-	-/-
50 ^a		<i>Ulva vietnamensis</i>	L.-A.T.Tran, Leliaert & De Clerck	ITS, <i>rbcL</i> , <i>tufA</i>	[61]
51 ^b		<i>Ulva aragoënsis</i>	(Bliding) Maggs	-/-	-/-
52 ^b		<i>Ulva chaugulii</i>	Kavale & Kazi	-/-	-/-
53 ^b		<i>Ulva kraftiorum</i>	Huisman	-/-	-/-
54 ^b		<i>Ulva limnetica</i>	Ichihara & Shimada	-/-	-/-
55 ^b	<i>Ulva meridionalis</i>	Horimoto & Shimada	-/-	-/-	
56 ^b	<i>Ulva ohnoi</i>	Hiraoka & Shimada	-/-	-/-	
57 ^b	<i>Ulva tepida</i>	Masakiyo & Shimada	-/-	-/-	
58 ^b	<i>Ulvella leptochaete</i>	(Huber) R.Nielsen, C.J.O'Kelly & B.Wysor	na	[38]	
59 ^b	<i>Ulvella scutata</i>	(Reinke) R.Nielsen, C.J.O'Kelly & B.Wysor	-/-	-/-	

Table 2. Checklist of the marine flora (Alismatales, Chlorophyta, Cyanobacteria, Ochrophyta, Rhodophyta) of Vietnam. The checklist is systematically and alphabetically arranged at the phylum, ordinal, familial, generic, and species levels.

Species Name	Regionally Reported as	References
CYANOBACTERIA		
Chroococcales		
Chroococcaceae		
<i>Chlorogloea</i> Wille		[42]
<i>Chlorogloea endophytica</i> Howe		
<i>Chroococcus</i> Nägeli		
<i>Chroococcus minor</i> (Kützing) Nägeli		[42]
<i>Entophysalis</i> Kützing		
<i>Entophysalis conferta</i> (Kützing) Drouet & Daily		[42]
<i>Entophysalis granulosa</i> Kützing		[42]
<i>Johannesbaptistia</i> G.De Toni		
<i>Johannesbaptistia primaria</i> (N.L.Gardner) G.De Toni		[42]
<i>Limnococcus</i> (Komárek & Anagnostidis) Komárková, Jezberová, O.Komárek & Zapomelová		
<i>Limnococcus limneticus</i> (Lemmermann) Komárková, Jezberová, O.Komárek & Zapomelová	<i>Chroococcus limneticus</i> Lemmermann	[42,62]
Gomphosphaeriaceae		
<i>Gomphosphaeria</i> Kützing		
<i>Gomphosphaeria aponina</i> Kützing		[42,63]
Microcystaceae		
<i>Aphanocapsa</i> Nägeli		
<i>Aphanocapsa litoralis</i> Hansgirg		[42]
<i>Aphanocapsa marina</i> Hansgirg		[42]
<i>Aphanocapsa reinboldii</i> (Richter) Komárek & Anagnostidis	<i>Microcystis reinboldii</i> (Richter) Forti	[42]
<i>Merismopedia</i> Meyen		
<i>Merismopedia glauca</i> (Ehrenberg) Kützing		[42]
Pleurocapsaceae		
<i>Chamaecalyx</i> Komárek & Anagnostidis		
<i>Chamaecalyx swirenkoi</i> (Schirshoff) Komárek & Anagnostidis		[42]
<i>Dermocarpella</i> Lemmermann		
<i>Dermocarpella hemisphaerica</i> (Lemmermann) Lemmermann		[42]
<i>Dermocarpella prasina</i> (Reinsch) Komárek & Anagnostidis		[42]
<i>Hyella</i> É.Bornet & C.Flahault		
<i>Hyella caespitosa</i> Bornet & Flahault		[42]
<i>Stanieria</i> J.Komárek & K.Anagnostidis		
<i>Stanieria sphaerica</i> (Setchell & N.L.Gardner) Anagnostidis & Pantazidou		[42]

Table 2. Cont.

Species Name	Regionally Reported as	References
Tryponema A.Ercegovic		
<i>Tryponema endolithicum</i> Ercegovic		[42]
Chroococciopsidales		
Aliterellaceae		
Gloeocapsopsis Geitler ex Komárek		
<i>Gloeocapsopsis crepidinum</i> (Thuret) Geitler ex Komárek		[42]
Leptolyngbyales		
Leptolyngbyaceae		
Heteroleibleinia (Geitler) Hoffmann		
<i>Heteroleibleinia infixa</i> (Frémy) Anagnostidis et Komárek	<i>Lyngbya infixa</i> Frémy	[42]
Leptolyngbya Anagnostidis & Komárek		
<i>Leptolyngbya gracilis</i> (Lindstedt) Anagnostidis & Komárek	<i>Phormidium gracile</i> (Rabenhorst ex Gomont) Anagnostidis	[42]
<i>Leptolyngbya rivulariarum</i> (Gomont) Anagnostidis & Komárek	<i>Lyngbya rivulariarum</i> Gomont	[42,63]
Phormidesmis Turicchia, Ventura, Komárková & Komárek		
<i>Phormidesmis mollis</i> (Gomont) Turicchia, Ventura, Komárková & Komárek	<i>Phormidium molle</i> Gomont	[42,63]
Planktolyngbya Anagnostidis & Komárek		
<i>Planktolyngbya limnetica</i> (Lemmermann) Komárková-Legnerová & Cronberg	<i>Lyngbya limnetica</i> Lemmermann	[42,63]
Trichocoleusaceae		
Trichocoleus Anagnostidis		
<i>Trichocoleus tenerrimus</i> (Gomont) Anagnostidis	<i>Microcoleus tenerrimus</i> Gomont	[42]
Nostocales		
Aphanizomenonaceae		
Gloeotrichia J.Agardh ex Bornet & Flahault		
<i>Gloeotrichia intermedia</i> (Lemmermann) Geitler		[42,63]
Hydrocoryne H.Schwabe ex É.Bornet & C.Flahault		
<i>Hydrocoryne enteromorphoides</i> (Bornet & Flahault) Umezaki & M.Watanabe	<i>Hormothamnion enteromorphoides</i> Grunow ex Bornet & Flahault	[42,63]
<i>Hydrocoryne soluta</i> (Bornet & Grunow) I.Umezaki	<i>Hormothamnion solutum</i> Bornet & Grunow	[42,63]
Hapalosiphonaceae		
Mastigocoleus Lagerheim ex É.Bornet & C.Flahault		
<i>Mastigocoleus testarum</i> Lagerheim ex Bornet & Flahault		[42,63]
Nostocaceae		
Nostoc Vaucher ex Bornet & Flahault		
<i>Nostoc commune</i> Vaucher ex Bornet & Flahault		[42,63]
Richelia J.Schmidt		
<i>Richelia intracellularis</i> J.Schmidt		[42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Rivulariaceae		
Calothrix C.Agardh ex Bornet & Flahault		
<i>Calothrix aeruginea</i> Thuret ex Bornet & Flahault		[42,63]
<i>Calothrix aeruginosa</i> Woronichin		[42]
<i>Calothrix confervicola</i> C.Agardh ex Bornet & Flahault		[42]
<i>Calothrix contarenii</i> Bornet & Flahault		[42,63]
<i>Calothrix nidulans</i> Setchell & N.L.Gardner		[42,63]
<i>Calothrix parietina</i> Thuret ex Bornet & Flahault		[42]
<i>Calothrix pulvinata</i> C.Agardh ex Bornet & Flahault		[42,63]
<i>Calothrix scopulorum</i> C.Agardh ex Bornet & Flahault		[42,63]
Kyrtuthrix Ercegovic		
<i>Kyrtuthrix maculans</i> (Gomont) I.Umezaki		[42,63]
Microchaete Thuret ex Bornet & C.Flahault		
<i>Microchaete tapahiensis</i> Setchell		[42,63]
<i>Microchaete vitiensis</i> Askenasy ex Bornet & Flahault		[42,63]
Rivularia C.Agardh ex Bornet & Flahault		
<i>Rivularia atra</i> f. <i>hemisphaerica</i> (Bornet & Flahault) Kossinskaja		[42,63]
<i>Rivularia atra</i> var. <i>confluens</i> Bornet		[42,63]
<i>Rivularia australis</i> Harvey ex Bornet & Flahault		[42,63]
Scytonemataceae		
Brachytrichia Zanardini ex Bornet & Flahault		
<i>Brachytrichia lloydii</i> (P.Crouan & H.Crouan) P.C.Silva	<i>Brachytrichia balani</i> Bornet & Flahault	[42,63]
<i>Brachytrichia quoyi</i> Bornet & Flahault		[42,63]
Scytonema C.Agardh ex É.Bornet & C.Flahault		
<i>Scytonema ocellatum</i> Lyngbye ex Bornet & Flahault		[42]
Scytonematopsis Kisseleva		
<i>Scytonematopsis crustacea</i> (Thuret ex Bornet & Flahault) Kováčik & Komárek	<i>Calothrix crustacea</i> Thuret ex Bornet & Flahault	[42]
<i>Scytonematopsis pilosa</i> (Bornet & Flahault) Umezaki & M.Watanabe	<i>Calothrix pilosa</i> Harvey ex Bornet & Flahault	[42,63]
Oscillatoriales		
Coleofasciculaceae		
Coleofasciculus M.A.Siegesmund, J.R.Johansen & T.Friedl		
<i>Coleofasciculus chthonoplastes</i> (Gomont) M.Siegesmund, J.R.Johansen & T.Friedl	<i>Microcoleus chthonoplastes</i> Thuret ex Gomont	[42]
Microcoleaceae		
Leibleinia (M.Gomont) L.Hoffman		
<i>Leibleinia epiphytica</i> (Hieronymus) Compère	<i>Lyngbya epiphytica</i> Hieronymus	[42,63]
Lyngbya C.Agardh ex Gomont		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Lyngbya aestuarii</i> Liebman ex Gomont		[42,63]
<i>Lyngbya agardhii</i> Gomont		[42,63]
<i>Lyngbya confervoides</i> C.Agardh ex Gomont		[42,63]
<i>Lyngbya lutea</i> Gomont	<i>Porphyrosiphon luteus</i> (Gomont) Anagnostidis & Komárek	[42,63]
<i>Lyngbya majuscula</i> Harvey ex Gomont		[42,63]
<i>Lyngbya martensiana</i> f. <i>tenuivaginata</i> Gomont ex Forti		[42,63]
<i>Lyngbya martensiana</i> Meneghini ex Gomont		[42,63]
<i>Lyngbya meneghiniana</i> Gomont		[42,63]
<i>Lyngbya semiplena</i> J.Agardh ex Gomont		[42,63]
<i>Lyngbya sordida</i> Gomont		[42,63]
Planktothrix Anagnostidis & Komárek		
<i>Planktothrix isothrix</i> (Skuja) Komárek & Komárková	<i>Oscillatoria agardhii</i> Gomont	[42,63]
Symploca Kützing ex Gomont		
<i>Symploca hydroides</i> Kützing ex Gomont		[42,63]
Oscillatoriaceae		
Blennothrix Kützing ex Anagnostidis & Komárek		
<i>Blennothrix cantharidosma</i> (Gomont) Anagnostidis & Komárek	<i>Hydrocoleum cantharidosmum</i> (Montagne) Gormont	[42,63]
<i>Blennothrix lyngbyacea</i> (Kützing ex Gomont) Anagnostidis & Komárek	<i>Hydrocoleum lyngbyaceum</i> Kützing ex Gomont	[42,63]
Oscillatoria Vaucher ex Gomont		
<i>Oscillatoria bonnemaisonii</i> P.Crouan & H.Crouan ex Gomont		[42,63]
<i>Oscillatoria corallinae</i> Gomont	<i>Phormidium corallinae</i> (Gomont) Anagnostidis & Komárek	[42,63]
<i>Oscillatoria indica</i> P.C.Silva		[42,48]
<i>Oscillatoria limosa</i> C.Agardh ex Gomont		[42,64]
<i>Oscillatoria margaritifera</i> Kützing ex Gomont		[42,63]
<i>Oscillatoria miniata</i> Hauck ex Gomont		[42,63,64]
<i>Oscillatoria princeps</i> Vaucher ex Gomont		[42,63]
<i>Oscillatoria simplicissima</i> Gomont	<i>Phormidium simplicissimum</i> (Gomont) Anagnostidis & Komárek	[42,63]
<i>Oscillatoria tenuis</i> C.Agardh ex Gomont		[42,63]
Phormidium Kützing ex Gomont		
<i>Phormidium corium</i> Gomont		[42,63,64]
<i>Phormidium feldmannii</i> Frémy		[42,63]
<i>Phormidium jadinianum</i> Gomont		[42]
<i>Phormidium nigroviride</i> (Thwaites ex Gomont) Anagnostidis & Komárek		[42,63]
<i>Phormidium nigrum</i> (Vaucher ex Gomont) Anagnostidis & Komárek		[42,63]
<i>Phormidium salinum</i> (Alten) Anagnostidis & Komárek		[48]
Pleurocapsales		

Table 2. Cont.

Species Name	Regionally Reported as	References
Dermocarpellaceae		
<i>Dermocarpa acervata</i> (Setchell & Gardner) Pham-Hoàng Hô		[42]
Hydrococcaceae		
<i>Hydrococcus</i> Kützing		
<i>Hydrococcus rivularis</i> Kützing		[42]
Spirulinales		
Spirulinaceae		
<i>Spirulina</i> Turpin ex Gomont		
<i>Spirulina major</i> Kützing ex Gomont		[42,63,64]
<i>Spirulina subsalsa</i> Oersted ex Gomont		[42,63]
<i>Spirulina subtilissima</i> Kützing ex Gomont		[42,63]
<i>Spirulina tenerrima</i> Kützing ex Gomont		[42,63]
Pseudanabaenales		
Pseudanabaenaceae		
<i>Pseudanabaena</i> Lauterborn		
<i>Pseudanabaena limnetica</i> (Lemmermann) Komárek		[42]
RHODOPHYTA		
Acrochaetiales		
Acrochaetiaceae		
<i>Acrochaetium</i> Nägeli		
<i>Acrochaetium barbadense</i> (Vickers) Børgesen	<i>Acrochaetium occidentale</i> Børgesen	[14,27,38,41,42]
<i>Acrochaetium catenulatum</i> M.Howe		[14,27,38,41,42,64]
<i>Acrochaetium chaetomorphae</i> (Tanaka & Pham-Hoàng Hô) Heerebout	<i>Erythrocladia chaetomorphae</i> T.Tanaka & Pham-Hoàng Hô	[14,41,65]
<i>Acrochaetium colaconemoides</i> Pham-Hoàng Hô		[14,41,42]
<i>Acrochaetium crassipes</i> (Børgesen) Børgesen		[14,41]
<i>Acrochaetium erectum</i> Børgesen		[41,42]
<i>Acrochaetium liagorae</i> Børgesen		[27,38,41,42]
<i>Acrochaetium microscopicum</i> (Nägeli ex Kützing) Nägeli		[42]
<i>Acrochaetium phuquocense</i> Pham-Hoàng Hô		[14,41,42,66]
<i>Acrochaetium polysporum</i> M.Howe		[42]
<i>Acrochaetium pseudoerectum</i> Pham-Hoàng Hô		[14,38,41,42,66]
<i>Acrochaetium pulchellum</i> Børgesen		[14,41,42]
<i>Acrochaetium sanctaemariae</i> (Darbishire) Hamel		[14,38,41,42]
<i>Acrochaetium sanctithomae</i> Børgesen		[38,41,42]
<i>Acrochaetium secundatum</i> (Lyngbye) Nägeli	<i>Acrochaetium virgatulum</i> (Harvey) Batters	[14,41,42]
<i>Acrochaetium subseriatum</i> Børgesen		[14,38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
Dermocarpellaceae		
<i>Liagorophila</i> Yamada		
<i>Liagorophila endophytica</i> Yamada	<i>Acrochaetium yamadae</i> (Garbary) Y.Lee & I.K.Lee	[38,42]
Ahnfeltiales		
Ahnfeltiaceae		
<i>Ahnfeltia</i> E.M.Fries		
<i>Ahnfeltia plicata</i> (Hudson) E.M.Fries		[41,42]
Bangiales		
Bangiaceae		
<i>Bangia</i> Lyngbye		
<i>Bangia fuscopurpurea</i> (Dillwyn) Lyngbye		[14,38,41,42]
<i>Bangia tanakae</i> Pham-Hoàng Hô	<i>Bangia tanakai</i> Pham H.H.	[14,38,41,42]
<i>Phycocalidia</i> Santiañez & M.J.Wynne		
<i>Phycocalidia tanegashimensis</i> (I.Shinmura) Santiañez		[54]
<i>Phycocalidia suborbiculata</i> (Kjellman) Santiañez & M.J.Wynne	<i>Porphyra suborbiculata</i> Kjellman, <i>Pyropia suborbiculata</i> (Kjellman) J.E.Sutherland, H.G.Choi, M.S.Hwang & W.A.Nelson	[14,38,41,42,67]
<i>Porphyra</i> C.Agardh		
<i>Porphyra tanakae</i> Pham Hoang-Ho	<i>Porphyra tanaka</i> Pham H.H.	[41,42]
<i>Pyropia</i> J.Agardh		
<i>Pyropia pseudolobata</i> (L.-E.Yang, J.Brodie & Q.-Q.Lu) Santiañez & M.J.Wynne	<i>Porphyra suborbiculata</i> Kjellman	[42,67,68]
<i>Pyropia vietnamensis</i> (Tak.Tanaka & P.H.Hô) J.E.Sutherland & Monotilla	<i>Porphyra vietnamensis</i> Tak.Tanaka & P.H.Hô	[38,41,69]
Bonnemaisoniales		
Bonnemaisoniaceae		
<i>Asparagopsis</i> Montagne		
<i>Asparagopsis taxiformis</i> (Delile) Trevisan		[14,27,38,41,64,70–72]
Ceramiales		
Callithamniaceae		
<i>Aglaothamnion</i> Feldmann-Mazoyer		
<i>Aglaothamnion cordatum</i> (Børgesen) Feldmann-Mazoyer	<i>Aglaothamnion neglectum</i> Feldmann-Mazoyer	[27,38,41,42]
<i>Callithamnion</i> Lyngbye		
<i>Callithamnion ramosissimum</i> N.L.Gardner		[42]
<i>Crouania</i> J.Agardh		
<i>Crouania attenuata</i> (C.Agardh) J.Agardh		[27,38,41,42]
<i>Gymnothamnion</i> J.Agardh		
<i>Gymnothamnion elegans</i> (Schousboe ex C.Agardh) J.Agardh		[14,38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
Spyridia Harvey		
<i>Spyridia filamentosa</i> (Wulfen) Harvey		[38,41,42,64,67]
<i>Spyridia hypnoides</i> (Bory) Papenfuss		[38,41,42,64,67]
Ceramiaceae		
Antithamnion Nägeli		
<i>Antithamnion antillanum</i> Børgesen		[38,41,64]
<i>Antithamnion erucacladellum</i> R.E.Norris		[38,41,42,64]
Antithamnionella Lyle		
<i>Antithamnionella basispora</i> (Tokida & Inaba) Cormaci & G.Furnari	<i>Antithamnion basisporum</i> Tokida & Inaba	[14,38,41,42]
<i>Antithamnionella breviramosa</i> (E.Y.Dawson) Wollaston		[41]
<i>Antithamnionella graeffei</i> (Grunow) Athanasiadis		[27,41,42]
<i>Antithamnionella spirographidis</i> (Schiffner) E.M.Wollaston	<i>Antithamnion spirographidis</i> Schiffner	[41,42,63]
Centroceras Kützing		
<i>Centroceras clavulatum</i> (C.Agardh) Montagne		[41,42,68,72]
<i>Centroceras gasparrinii</i> (Meneghini) Kützing	<i>Centroceras inerme</i> Kützing	[14,38,41,42,64]
Ceramium Roth		
<i>Ceramium aduncum</i> Nakamura		[38,41,42,64]
<i>Ceramium borneense</i> Weber Bosse		[41,42,63]
<i>Ceramium cimbricum</i> H.E.Petersen	<i>Ceramium fastigiatum</i> Harvey	[14,38,41,42,64]
<i>Ceramium cingulatum</i> Weber Bosse		[14,38,41,42,64]
<i>Ceramium clarionense</i> Setchell & N.L.Gardner		[14,38,41,42]
<i>Ceramium codii</i> (H.Richards) Mazoyer		[41,42,63]
<i>Ceramium deslongchampsii</i> Chauvin ex Duby		[41,42,63]
<i>Ceramium diaphanum</i> (Lightfoot) Roth	<i>Ceramium gracillimum</i> (Kützing) Zanardini	[41,42,63]
<i>Ceramium macilentum</i> J.Agardh	<i>Ceramium mazatlanense</i> E.Y.Dawson	[27,41,42,64,68,72]
<i>Ceramium maryae</i> Weber Bosse		[14,38,41,42]
<i>Ceramium procumbens</i> Setchell & N.L.Gardner		[38,41,42,64]
<i>Ceramium tenerrimum</i> (G.Martens) Okamura		[41,42,63]
<i>Ceramium vagans</i> P.C.Silva		[41,42,63]
<i>Ceramium vietnamense</i> Pham-Hoàng Hô		[14,38,41,42]
<i>Ceramium zacaе</i> Setchell & N.L.Gardner		[41,42,63]
Corallophila Weber-van Bosse		
<i>Corallophila bella</i> (Setchell & Gardner) R.E.Norris		[27,38,41,42]
<i>Corallophila howei</i> (Weber Bosse) R.E.Norris		[14,38,41,42]
<i>Corallophila huysmansii</i> (Weber Bosse) R.E.Norris	<i>Ceramium huysmansii</i> Weber Bosse	[14,27,38,41,42]
<i>Corallophila kleiwegii</i> Weber Bosse	<i>Corallophila apiculata</i> (Yamada) R.E.Norris	[27,41,42,64]

Table 2. Cont.

Species Name	Regionally Reported as	References
Gayliella T.O.Cho, L.M.McIvor & S.M.Boo		
<i>Gayliella fimbriata</i> (Setchell & N.L.Gardner) T.O.Cho & S.M.Boo	<i>Ceramium fimbriatum</i> Setchell & N.L.Gardner	[14,38,41,42]
<i>Gayliella flaccida</i> (Harvey ex Kützing) T.O.Cho & L.M.McIvor	<i>Ceramium flaccidum</i> (Harvey ex Kützing) Ardissonne	[27]
<i>Gayliella mazoyerae</i> T.O.Cho, Fredericq & Hommersand	<i>Ceramium gracillimum</i> var. <i>byssoides</i> Mazoyer	[38,41,42]
<i>Gayliella taylorii</i> (E.Y.Dawson) T.O.Cho & S.M.Boo	<i>Ceramium taylorii</i> E.Y.Dawson	[14,38,41,42]
Reinboldiella De Toni		
<i>Reinboldiella warburgii</i> (Heydrich) Yoshida & Mikami		[41,42,63]
Delesseriaceae		
Acrosorium Zanardini ex Kützing		
<i>Acrosorium polyneurum</i> Okamura		[41,42,63]
Branchioglossum Kylin		
<i>Branchioglossum prostratum</i> C.W.Schneider		[27,38,41,42]
Caloglossa (Harvey) G.Martens		
<i>Caloglossa beccarii</i> (Zanardini) De Toni		[42,73]
<i>Caloglossa bengalensis</i> (G.Martens) R.J.King & Puttock	<i>Caloglossa adnata</i> (Zanardini) De Toni	[41,42,63]
<i>Caloglossa continua</i> (Okamura) R.J.King & Puttock		[41,42,63]
<i>Caloglossa leprieurii</i> (Montagne) G.Martens		[14,38,41,42]
<i>Caloglossa ogasawaraensis</i> Okamura		[41,42,63]
<i>Caloglossa saigonensis</i> Tanaka & Pham-Hoàng Hô		[41,42,74]
<i>Caloglossa stipitata</i> E.Post		[41,42,63]
Claudea J.V.Lamouroux		
<i>Claudea batanensis</i> Tanaka		[38,41,42]
Cottoniella Børgesen		
<i>Cottoniella filamentosa</i> (M.Howe) Børgesen		[41,42,63]
Dasya C.Agardh		
<i>Dasya anastomosans</i> (Weber Bosse) M.J.Wynne	<i>Dasyopsis pilosa</i> Weber Bosse; <i>Dasya pilosa</i> (Weber Bosse) A.J.K.Millar	[14,38,41,42]
<i>Dasya crouaniana</i> J.Agardh		
<i>Dasya pedicellata</i> (C.Agardh) C.Agardh	<i>Dasya baillowiana</i> (S.G.Gmelin) Montagne	[14,38,41,42]
<i>Dasya scoparia</i> Harvey		[41,42,63]
Dictyurus Bory de Saint-Vincent		
<i>Dictyurus occidentalis</i> J.Agardh		[41,42,63]
Hypoglossum Kützing		
<i>Hypoglossum attenuatum</i> N.L.Gardner		[14,38,41,42,64]
<i>Hypoglossum barbatum</i> Okamura		[41,42,63]
<i>Hypoglossum simulans</i> M.J.Wynne, I.R.Price & D.L.Ballantine		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Martensia K.Hering		
<i>Martensia flabelliformis</i> Harvey ex J.Agardh	<i>Neomartensia flabelliformis</i> (Harvey ex J.Agardh) Yoshida & Mikami	[41,42,63]
<i>Martensia fragilis</i> Harvey		[38,41,42]
Nitophyllum Greville		
<i>Nitophyllum adhaerens</i> M.J.Wynne		[27,41,42]
Taenioma J.Agardh		
<i>Taenioma perpusillum</i> (J.Agardh) J.Agardh		[41,42,63]
Zellera G.Martens		
<i>Zellera tawallina</i> G.Martens		[55]
Rhodomelaceae		
Acanthophora J.V.Lamouroux		
<i>Acanthophora muscoides</i> (Linnaeus) Bory		[38,41,42,64,67,68]
<i>Acanthophora spicifera</i> (M.Vahl) Børgesen		[27,41,42,64,68,72]
Acrocystis Zanardini		
<i>Acrocystis nana</i> Zanardini		[14,38,41,42]
Amansia J.V.Lamouroux		
<i>Amansia glomerata</i> C.Agardh	<i>Melanamansia glomerata</i> (C. Agardh) R.E. Norris	[38,41,42,75]
<i>Amansia rhodantha</i> (Harvey) J.Agardh		[41,42,63]
Bostrychia Montagne		
<i>Bostrychia moritziana</i> (Sonder ex Kützing) J.Agardh		[63]
<i>Bostrychia radicans</i> (Montagne) Montagne		[14,38,41,42]
<i>Bostrychia tenella</i> (J.V.Lamouroux) J.Agardh		[38,41,42,67]
Bryocladia F.Schmitz		
<i>Bryocladia cervicornis</i> (Kützing) F.Schmitz		[14,38,41,42]
Chondria C.Agardh		
<i>Chondria armata</i> (Kützing) Okamura		[41,42,63]
<i>Chondria baileyana</i> (Montagne) Harvey		[14,38,41,42]
<i>Chondria dangeardii</i> E.Y.Dawson		[14,27,38,41,42]
<i>Chondria repens</i> Børgesen		[14,38,41,42,64]
<i>Chondria ryukyuensis</i> Yamada		[41,42,63]
<i>Chondria simpliciuscula</i> Weber Bosse		[27,38,41,42]
Chondrophycus (J.Tokida & Y.Saito) Garbary & J.T.Harper		
<i>Chondrophycus articulatus</i> (C.K.Tseng) K.W.Nam	<i>Laurencia articulata</i> C.K.Tseng	[14,38,41,42]
<i>Chondrophycus cartilagineus</i> (Yamada) Garbary & J.T.Harper	<i>Laurencia cartilaginea</i> Yamada	[14,38,41,42]
<i>Chondrophycus tronoi</i> (E.Ganzon-Fortes) K.W.Nam		[47]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Chondrophycus undulatus</i> (Yamada) Garbary & Harper		[41,42,63]
<i>Chondrophycus verticillatus</i> (J.F.Zhang & B.M.Xia) K.W.Nam		[41,42,63]
Endosiphonia Zanardini		
<i>Endosiphonia horrida</i> (C.Agardh) P.C.Silva		[41,42,63]
Epizonaria Díaz-Tapia & Maggs		
<i>Epizonaria prostrata</i> (Harvey) Díaz-Tapia & Maggs	<i>Lophosiphonia prostrata</i> (Harvey) Falkenberg	[27,38,41,42]
Eutrichosiphonia Savoie & G.W.Saunders		
<i>Eutrichosiphonia tapinocarpa</i> (Suringar) D.E.Bustamante & T.O.Cho	<i>Polysiphonia tapinocarpa</i> Suringar	[41,42,63]
Exophyllum Weber-van Bosse		
<i>Exophyllum wentii</i> Weber Bosse		[41,42,63]
Herposiphonia Nägeli		
<i>Herposiphonia caespitosa</i> C.K.Tseng		[41,42,63]
<i>Herposiphonia crassa</i> Hollenberg		[27,38,41,42]
<i>Herposiphonia delicatula</i> Hollenberg		[27,38,41,42]
<i>Herposiphonia insidiosa</i> (Greville ex J.Agardh) Falkenberg		[14,38,41,42]
<i>Herposiphonia parca</i> Setchell		[27,41,42,64]
<i>Herposiphonia tenella</i> (C.Agardh) Ambronn	<i>Herposiphonia secunda</i> f. <i>tenella</i> (C.Agardh) M.J.Wynne	[14,27,38,41,42]
<i>Herposiphonia vietnamica</i> Pham-Hoàng Hô		
Laurencia J.V.Lamouroux		
<i>Laurencia brachyclados</i> Pilger		[41,42,63]
<i>Laurencia caduciramulosa</i> Masuda & S.Kawaguchi		[41,42,63]
<i>Laurencia calliclada</i> Masuda		[41,42,63]
<i>Laurencia corymbosa</i> J.Agardh		[14,38,41,42]
<i>Laurencia decumbens</i> Kützing		[38,41,42]
<i>Laurencia dendroidea</i> J.Agardh	<i>Laurencia majuscula</i> (Harvey) A.H.S.Lucas	[38,41,42,64]
<i>Laurencia filiformis</i> (C.Agardh) Montagne		[41,42,63]
<i>Laurencia galtsoffii</i> M.Howe		[41,42,63]
<i>Laurencia heteroclada</i> Harvey		[41,42,63]
<i>Laurencia intricata</i> J.V.Lamouroux		[41,42,63]
<i>Laurencia lageniformis</i> Masuda & Suzuki		[41,42,63]
<i>Laurencia mariannensis</i> Yamada		[41,42,63]
<i>Laurencia microcladia</i> Kützing		[14,38,41,42]
<i>Laurencia nangii</i> Masuda		[28,38,41,42]
<i>Laurencia natalensis</i> Kylin		[41,42,63]
<i>Laurencia nidifica</i> J.Agardh		[41,42,63]
<i>Laurencia obtusa</i> (Hudson) J.V.Lamouroux		[38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Laurencia obtusa</i> var. <i>densa</i> Yamada		[41,42,63]
<i>Laurencia pinnata</i> Yamada		[38,41,42]
<i>Laurencia silvae</i> J.F.Zhang & B.M.Xia	<i>Laurencia fasciculata</i> J.F.Zhang & B.M.Xia	[41,42,63]
<i>Laurencia similis</i> K.W.Nam & Y.Saito		[41,42,63]
<i>Laurencia tenera</i> C.K.Tseng		[14,38,41,42]
<i>Laurencia tropica</i> Yamada		[38,41,42,67]
Leveillea Decaisne		
<i>Leveillea jungermannioides</i> (Hering & G.Martens) Harvey		[14,27,41,42,67]
Lophosiphonia Falkenberg		
<i>Lophosiphonia obscura</i> (C.Agardh) Falkenberg		[38,41,42]
Melanothamnus Bornet & Falkenberg		
<i>Melanothamnus ferulaceus</i> (Suhr ex J.Agardh) Díaz-Tapia & Maggs	<i>Neosiphonia ferulacea</i> (Suhr ex J.Agardh) S.M.Guimarães & M.T.Fujii	[41,42,63]
<i>Melanothamnus harlandii</i> (Harvey) Díaz-Tapia & Maggs	<i>Neosiphonia harlandii</i> (Harvey) M.S.Kim & I.K.Lee	[38,41,42]
<i>Melanothamnus sparsus</i> (Setchell) Díaz-Tapia & Maggs	<i>Neosiphonia sparsa</i> (Setchell) I.A.Abbott	[27,38,41,42]
<i>Melanothamnus sphaerocarpus</i> (Børgesen) Díaz-Tapia & Maggs	<i>Neosiphonia sphaerocarpa</i> (Børgesen) M.-S.Kim & I.K.Lee	[27,41,42,64]
<i>Melanothamnus tongatensis</i> (Harvey ex Kützing) Díaz-Tapia & Maggs	<i>Neosiphonia tongatensis</i> (Harvey ex Kützing) M.-S.Kim & I.K.Lee	[14,41,42,64]
<i>Melanothamnus upolensis</i> (Grunow) Díaz-Tapia & Maggs	<i>Neosiphonia upolensis</i> (Grunow) M.S.Kim & Boo	[41,42,63]
<i>Melanothamnus infestans</i> (Harvey) Huisman	<i>Polysiphonia infestans</i> Harvey	[41]
<i>Melanothamnus thailandicus</i> (N.Muangmai & C.Kaewsuralikhit) Díaz-Tapia & Maggs		[51]
Neurymenia J.Agardh		
<i>Neurymenia fraxinifolia</i> (Mertens ex Turner) J.Agardh		[41,42,63]
Odonthalia Lyngbye		
<i>Odonthalia corymbifera</i> (S.G.Gmelin) Greville		[41,42,63]
Ohelopapa F.Rousseau, Martin-Lescanne, Payri & L.Le Gall		
<i>Ohelopapa flexilis</i> (Setchell) F.Rousseau, Martin-Lescanne, Payri & L.Le Gall		[39,42]
Palisada K.W.Nam		
<i>Palisada concreta</i> (A.B.Cribb) K.W.Nam		[38,41,42]
<i>Palisada intermedia</i> (Yamada) K.W.Nam		[41,42,63]
<i>Palisada parvipapillata</i> (C.K.Tseng) K.W.Nam		[38,41,42]
<i>Palisada perforata</i> (Bory) K.W.Nam		[41,42,63]
<i>Palisada thuyoides</i> (Kützing) Cassano, Senties, Gil-Rodríguez & M.T.Fujii		[38,41,42]
<i>Palisada yamadana</i> (M.Howe) K.W.Nam		[38,41,42]
Polysiphonia Greville		
<i>Polysiphonia coacta</i> C.K.Tseng		[14,38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Polysiphonia fragilis</i> Suringar		[14,38,41,42]
<i>Polysiphonia kampsaxii</i> Børgesen		[41,42,63]
<i>Polysiphonia nhatrangense</i> Pham-Hoàng Hô		[14,38,41,42]
<i>Polysiphonia poko</i> Hollenberg	<i>Neosiphonia poko</i> (Hollenberg) I.A.Abbott	[38,41,42]
<i>Polysiphonia scopulorum</i> Harvey		[27,38,41,42,64,69]
<i>Polysiphonia sertularioides</i> (Grateloup) J.Agardh		[41,42,63]
<i>Polysiphonia subtilissima</i> Montagne	<i>Neosiphonia subtilissima</i> (Montagne) M.S. Kim & I.K. Lee	[14,27,38,41,42]
<i>Polysiphonia villum</i> J.Agardh	<i>Polysiphonia scopulorum</i> var. <i>villum</i> (J.Agardh) Hollenberg	[38,41,42,64]
Rodriguezella F.Schmitz		
<i>Rodriguezella hongngai</i> Pham-Hoàng Hô		[14,38,41,42]
Symphyocladia Falkenberg		
<i>Symphyocladia marchantioides</i> (Harvey) Falkenberg		[41,42,63]
Tayloriella Kylin		
<i>Tayloriella dictyurus</i> (J.Agardh) Kylin		[41,42,63]
Tolypiocladia F.Schmitz		
<i>Tolypiocladia calodictyon</i> (Harvey ex Kützing) P.C.Silva		[41,42,63]
<i>Tolypiocladia glomerulata</i> (C.Agardh) F.Schmitz		[14,27,41,42,67]
Vertebrata S.F.Gray		
<i>Vertebrata reptabunda</i> (Suhr) Díaz-Tapia & Maggs	<i>Lophosiphonia reptabunda</i> (Suhr) Kylin	[14,38,41,42]
Womersleyella Hollenberg		
<i>Womersleyella herpa</i> (Hollenberg) R.E.Norris	<i>Polysiphonia herpa</i> Hollenberg	[14,38,41,42]
Wrangeliaceae		
Anotrichium Nägeli		
<i>Anotrichium barbatum</i> Nägeli		[41,42,63]
<i>Anotrichium tenue</i> (C.Agardh) Nägeli	<i>Anotrichium tenue</i> var. <i>thyrsigerum</i> (Thwaites ex Harvey) H.S.Kim & I.K.Lee	[27,41,42,64]
Diplothamnion A.B.Joly & Yamagishi		
<i>Diplothamnion jolyi</i> C.Hoek		[41,42,64]
Griffithsia C.Agardh		
<i>Griffithsia heteromorpha</i> Kützing		[38,41,42,64]
<i>Griffithsia japonica</i> Okamura		[14,27,38,41,42,64,67]
<i>Griffithsia metcalfeii</i> C.K. Tseng		[38,41,42,64]
Haloplegma Montagne		
<i>Haloplegma duperreyi</i> Montagne		[41,42,63]
Monosporus Solier		
<i>Monosporus pedicellatus</i> var. <i>tenuis</i> (Feldmann-Mazoyer) Huisman & Kraft	<i>Neomonospora pedicellata</i> var. <i>tenuis</i> Feldmann-Mazoyer	[38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
Pleonosporium Nägeli		
<i>Pleonosporium borrieri</i> (Smith) Nägeli		[14,38,41,42]
Spongoclonium Sonder		
<i>Spongoclonium caribaeum</i> (Børgesen) M.J.Wynne	<i>Mesothamnion caribaeum</i> Børgesen	[41,42,63]
Wrangelia C.Agardh		
<i>Wrangelia argus</i> (Montagne) Montagne		[14,38,41,42]
<i>Wrangelia dumontii</i> (E.Y.Dawson) I.A.Abbott		[27,38,41,42]
<i>Wrangelia tanegana</i> Harvey		[38,41,42]
Colaconematales		
Colaconemataceae		
Colaconema Batters		
<i>Colaconema gracile</i> (Børgesen) Ateweberhan & Prud'homme	<i>Acrochaetium gracile</i> Børgesen	[14,27,38,41,42]
<i>Colaconema gracile</i> var. <i>vietnamense</i> Pham Hoang Ho		[41,42,63]
<i>Colaconema hallandicum</i> (Kyllin) Afonso-Carrillo, Sanson, Sangil & Diaz-Villa		[38,41,42]
<i>Colaconema hypneae</i> (Børgesen) A.A.Santos & C.W.N.Moura		[38,41,42,64]
<i>Colaconema robustum</i> (Børgesen) Huisman & Woelkerling	<i>Acrochaetium robustum</i> Børgesen; <i>Audouinella robusta</i> (Børgesen) Garbary	[14,38,41,42]
<i>Colaconema thuretii</i> (Bornet) P.W.Gabrielson		[38,41,42]
Compsopogonales		
Compsopogonaceae		
Compsopogon Montagne		
<i>Compsopogon caeruleus</i> (Balbis ex C.Agardh) Montagne		[41,42,63]
Corallinales		
Corallinaceae		
Corallina Linnaeus		
<i>Corallina officinalis</i> Linnaeus		[41,42,63]
<i>Corallina pilulifera</i> Postels & Ruprecht		[41,42,63]
Jania J.V.Lamouroux		
<i>Jania acutiloba</i> (Decaisne) J.H.Kim, Guiry & H.-G.Choi		[38,41,42]
<i>Jania cultrata</i> (Harvey) J.H.Kim, Guiry & H.-G.Choi	<i>Cheilosporum cultratum</i> (Harvey) Areschoug	
<i>Jania longiarthra</i> E.Y.Dawson		[14,38,41,42]
<i>Jania micrarthrodia</i> J.V.Lamouroux		[38,41,42,64]
<i>Jania pedunculata</i> var. <i>adhaerens</i> (J.V.Lamouroux) A.S.Harvey, Woelkerling & Rviers	<i>Jania adhaerens</i> J.V.Lamouroux; <i>Jania decussatodichotoma</i> (Yendo) Yendo; <i>Jania capillacea</i> Harvey	[27,41,42,64,68,72]
<i>Jania pumila</i> J.V.Lamouroux		[14,27,38,41,42]
<i>Jania rubens</i> (Linnaeus) J.V.Lamouroux		[14,38,41,42]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Jania spectabilis</i> (Harvey ex Grunow) J.H.Kim, Guiry & H.-G.Choi		[38,41,42]
<i>Jania squamata</i> (Linnaeus) J.H.Kim, Guiry & H.-G.Choi		[41,42,63]
<i>Jania unguolata</i> f. <i>brevior</i> (Yendo) Yendo		[27,38,41,42]
Pneophyllum Kützing		
<i>Pneophyllum confervicola</i> (Kützing) Y.M.Chamberlain	<i>Heteroderma minutulum</i> (Foslie) Foslie); <i>Melobesia confervicola</i> (Kützing) Foslie	[38,41,42]
Hapalidiaceae		
Lithothamnion Heydrich		
<i>Lithothamnion erubescens</i> f. <i>subflabellatum</i> Foslie		[38,41,42]
Hydrolithaceae		
Hydrolithon (Foslie) Foslie		
<i>Hydrolithon boergesenii</i> (Foslie) Foslie	<i>Hydrolithon reinboldii</i> (Weber Bosse & Foslie) Foslie	[14,38,41,42]
<i>Hydrolithon farinosum</i> (J.V.Lamouroux) Penrose & Y.M.Chamberlain		[38,41,42,64]
Lithophyllaceae		
Amphiroa J.V.Lamouroux		
<i>Amphiroa anceps</i> (Lamarck) Decaisne		[41,42,63]
<i>Amphiroa beauvoisii</i> J.V.Lamouroux		[27,41,42,64]
<i>Amphiroa echigoensis</i> Yendo		[41,42,63]
<i>Amphiroa foliacea</i> J.V.Lamouroux, Bory & Eudes-Deslongchamps		[14,27,38,41,42,67]
<i>Amphiroa fragilissima</i> (Linnaeus) J.V.Lamouroux		[27,41,42,64,68,72]
<i>Amphiroa valonioides</i> Yendo		[38,41,42,64]
<i>Amphiroa crassa</i> J.V.Lamouroux, Bory & Eudes-Deslongchamps		[41]
Lithophyllum Philippi		
<i>Lithophyllum okamurae</i> Foslie		[14,38,41,42]
<i>Lithophyllum pygmaeum</i> (Heydrich) Heydrich		[41,42,63]
Titanoderma Nägeli		
<i>Titanoderma pustulatum</i> (J.V.Lamouroux) Nägeli	<i>Lithophyllum pustulatum</i> (J.V.Lamouroux) Foslie	[41,42,63]
Mastophoraceae		
Mastophora Decaisne		
<i>Mastophora pacifica</i> (Heydrich) Foslie	<i>Lithoporella pacifica</i> (Heyrich) Foslie	[38,41,42]
<i>Mastophora rosea</i> (C.Agardh) Setchell		[38,41,42,64,67]
Mesophyllumaceae		
Melyvonnea Athanasiadis & D.L.Ballantine		
<i>Melyvonnea erubescens</i> (Foslie) Athanasiadis & D.L.Ballantine		[41,42,63]
Mesophyllum Me.Lemoine		
<i>Mesophyllum simulans</i> (Foslie) Me.Lemoine	<i>Lithothamnion simulans</i> (Foslie) Foslie	[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Porolithaceae		
<i>Harveylithon</i> A.Rösler, Perfectti, V.Peña & J.C.Braga		
<i>Harveylithon samoëense</i> (Foslie) A.Rösler, Perfectti, V.Peña & J.C.Braga	<i>Hydrolithon samoëense</i> (Foslie) Keats & Y.M. Chamberlain	[38,41,42,67]
<i>Metagoniolithon</i> Weber-van Bosse		
<i>Metagoniolithon stelliferum</i> (Lamarck) Ducker		[41,42,63]
Spongitiaceae		
<i>Neogoniolithon</i> Setchell & L.R.Mason		
<i>Neogoniolithon oblimans</i> (Heydrich) P.C.Silva		[14,38,41,42]
<i>Neogoniolithon trichotomum</i> (Heydrich) Setchell & L.R.Mason		[41,42,63]
Erythropeltales		
Erythrotrichiaceae		
<i>Erythrocladia</i> Rosenvinge		
<i>Erythrocladia irregularis</i> Rosenvinge		[41,42,63]
<i>Erythrotrichia</i> Areschoug		
<i>Erythrotrichia carnea</i> (Dillwyn) J.Agardh		[14,27,38,41,42]
<i>Erythrotrichia parietalis</i> T.Tanaka		[14,38,41,42]
<i>Erythrotrichia parietalis</i> var. <i>majuscula</i> T.Tanaka & Pham-Hoàng Hô		[41,42,63]
<i>Sahlingia</i> Kornmann		
<i>Sahlingia subintegra</i> (Rosenvinge) Kornmann	<i>Erythrocladia subintegra</i> Rosenvinge	[41,42,63]
Gelidiales		
Gelidiaceae		
<i>Gelidium</i> J.V.Lamouroux		
<i>Gelidium corneum</i> (Hudson) J.V.Lamouroux		[41,42,63]
<i>Gelidium crinale</i> (Hare ex Turner) Gaillon		[14,38,41,42]
<i>Gelidium crinale</i> var. <i>perpusillum</i> Piccone & Grunow		[38,41,42]
<i>Gelidium divaricatum</i> G.Martens	<i>Gelidiophycus divaricatus</i> (G.Martens) G.H.Boo, J.K.Park & S.M.Boo	[14,38,41,42,76]
<i>Gelidium fasciculatum</i> Hamel		[41,42,63]
<i>Gelidium minusculum</i> (Weber Bosse) R.E.Norris	<i>Gelidium pusillum</i> var. <i>minusculum</i> Weber Bosse	[41,42,63]
<i>Gelidium pulchellum</i> (Turner) Kützing		[4,27,38,41,42]
<i>Gelidium pusillum</i> (Stackhouse) Le Jolis		[27,41,42,64,68,72]
<i>Gelidium pusillum</i> var. <i>cylindricum</i> W.R.Taylor		[41]
<i>Gelidium pusillum</i> var. <i>pacificum</i> W.R.Taylor		[41,42,63]
<i>Gelidium samoëense</i> Reinbold		[41,42,63]
<i>Gelidium spathulatum</i> (Kützing) Bornet		[14,38,41,42]
<i>Gelidium vietnamense</i> Pham-Hoàng Hô		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Gelidiellaceae		
<i>Gelidiella</i> Feldmann & G.Hamel		
<i>Gelidiella acerosa</i> (Forsskål) Feldmann & Hamel		[14,27,38,41,42,67]
<i>Gelidiella lubrica</i> (Kützing) Feldmann & Hamel		[14,38,41,42]
<i>Millerella</i> G.H.Boo & S.M.Boo		
<i>Millerella myrioclada</i> (Børgesen) G.H.Boo	<i>Gelidiella myrioclada</i> (Børgesen) Feldmann & Hamel	[14,38,41,42]
<i>Millerella pannosa</i> (Feldmann) G.H.Boo & L.Le Gall	<i>Parviphycus pannosus</i> (Feldmann) G.Furnari	[38,41,42,64]
<i>Parviphycus</i> Santelices		
<i>Parviphycus trinitatis</i> (W.R.Taylor) M.J.Wynne		[41,53]
<i>Parviphycus adnatus</i> (E.Y.Dawson) B.Santelices	<i>Gelidiella adnata</i> E.Y.Dawson	[27,38,41,42,53,77]
<i>Perronella</i> G.H.Boo, T.V.Nguyen, J.Y.Kim & S.M. Boo		
<i>Perronella gracilis</i> G.H.Boo, T.V.Nguyen, J.Y.Kim & S.M. Boo		[41,53]
Pterocladaceae		
<i>Pterocladia</i> J.Agardh		
<i>Pterocladia heteroplatos</i> (Børgesen) Umamaheswara Rao & Kaliaperumal	<i>Gelidium heteroplatos</i> Børgesen	[41,42,63]
<i>Pterocladia</i> B.Santelices & Hommersand		
<i>Pterocladia bartlettii</i> (Taylor) Santelices		[57]
<i>Pterocladia maribagoensis</i> G.H.Boo & P.J.L.Geraldino		[57]
<i>Pterocladia musciformis</i> (W.R.Taylor) G.H.Boo & K.A.Miller		[57]
<i>Pterocladia caerulescens</i> (Kützing) Santelices & Hommersand		[27,41,42,64]
<i>Pterocladia caloglossoides</i> (M.Howe) Santelices		[41,42,63]
<i>Pterocladia capillacea</i> (S.G.Gmelin) Santelices & Hommersand	<i>Pterocladia capillacea</i> (S.G. Gmelin) Bornet in Bornet et Thuret	[41,42,63]
<i>Pterocladia tenuis</i> (Okamura) Shimada, Horiguchi & Masuda	<i>Pterocladia pinnata</i> (Hudson) Papenfuss	[41,42,63]
	<i>Pterocladia tenuis</i> Okamura	[41,42,63]
Gigartinales		
Calosiphoniaceae		
<i>Schmitzia</i> P.C.Silva		
<i>Schmitzia japonica</i> (Okamura) P.C.Silva	<i>Bertholdia japonica</i> (Okamura) Segawa	[41,42,63]
Caulacanthaceae		
<i>Catenella</i> Greville		
<i>Catenella impudica</i> (Montagne) J.Agardh		[41,42,63]
<i>Catenella nipae</i> Zanardini		[41,42,63]
<i>Catenella subumbellata</i> C.K.Tseng		[41,42,63]
<i>Caulacanthus</i> Kützing		
<i>Caulacanthus ustulatus</i> (Turner) Kützing		[41,42,63]
<i>Montemaria</i> A.B.Joly & Alveal		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Montemaria horridula</i> (Montagne) A.B.Joly & Alveal		[14,38,41,42]
Clavicloniaceae		
<i>Antrocentrum</i> Kraft & Min-Thein		
<i>Antrocentrum nigrescens</i> (Harvey) Kraft & Min-Thein		[41,42,63]
Cystocloniaceae		
<i>Hypnea</i> J.V.Lamouroux		
<i>Hypnea alopecuroides</i> Kützing		[41,42,63]
<i>Hypnea anastomosans</i> Papenfuss, Lipkin & P.C.Silva		[41,42,63]
<i>Hypnea cenomyce</i> J.Agardh		[14,38,41,42]
<i>Hypnea cervicornis</i> J.Agardh	<i>Hypnea boergesenii</i> T.Tanaka	[14,27,38,41,42]
<i>Hypnea charoides</i> J.V.Lamouroux		[41,42,63]
<i>Hypnea charoides</i> var. <i>indica</i> Weber Bosse		[41,42,63]
<i>Hypnea cornuta</i> (Kützing) J.Agardh		[14,38,41,42]
<i>Hypnea esperi</i> Bory		[14,27,38,41,42]
<i>Hypnea flagelliformis</i> Greville ex J.Agardh		[41,42,63]
<i>Hypnea hamulosa</i> (Esper) J.V.Lamouroux		[41,42,63]
<i>Hypnea japonica</i> Tanaka		[41,42,63]
<i>Hypnea nidulans</i> Setchell		[14,27,38,41,42]
<i>Hypnea pannosa</i> J.Agardh		[41,42,68,72]
<i>Hypnea spinella</i> (C.Agardh) Kützing		[14,27,38,41,42]
<i>Hypnea valentiae</i> (Turner) Montagne		[14,27,38,41,42,67,68]
Dicranemataceae		
<i>Tylopus</i> J.Agardh		
<i>Tylopus lichenoides</i> Okamura		[41]
Dumontiaceae		
<i>Gibsmithia</i> Doty		
<i>Gibsmithia hawaiiensis</i> Doty		[41,42,63]
Endocladaceae		
<i>Gloiopeltis</i> J.Agardh		
<i>Gloiopeltis furcata</i> (Postels & Ruprecht) J.Agardh		[41,42,63]
<i>Gloiopeltis tenax</i> (Turner) Decaisne		[41,42,63]
Gigartinaceae		
<i>Chondracanthus</i> Kützing		
<i>Chondracanthus acicularis</i> (Roth) Fredericq	<i>Gigartina acicularis</i> (Roth) J.V.Lamouroux	[41,42,63]
<i>Chondracanthus intermedius</i> (Suringar) Hommersand		[27,38,41,42,67]
<i>Chondracanthus tenellus</i> (Harvey) Hommersand	<i>Gigartina tenella</i> Harvey	[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Phyllophoraceae		
Ahnfeltiopsis P.C.Silva & DeCew		
<i>Ahnfeltiopsis chnoosporoides</i> (T.Tanaka & Pham-Hoàng Hô) Masuda		[41,42,63]
<i>Ahnfeltiopsis densa</i> (J.Agardh) P.C.Silva & DeCew	<i>Gymnogongrus densus</i> J.Agardh	[41,42,63]
<i>Ahnfeltiopsis flabelliformis</i> (Harvey) Masuda		[27,38,41,42,67,68]
<i>Ahnfeltiopsis pygmaea</i> (J.Agardh) P.C.Silva & DeCew		
<i>Ahnfeltiopsis quinhonensis</i> (Pham-Hoang Ho) Masuda		[27,28,38,41,42,67]
<i>Ahnfeltiopsis serenei</i> (E.Y.Dawson) Masuda		[41,42,63]
Besa Setchell		
<i>Besa divaricata</i> (Holmes) M.S.Calderon & S.M.Boo	<i>Ahnfeltiopsis divaricata</i> (Holmes) Masuda	[41,42,63]
Gymnogongrus C.Martius		
<i>Gymnogongrus griffithsiae</i> (Turner) C.Martius		[41,42,63]
<i>Gymnogongrus johnstonii</i> (Setchell & N.L.Gardner) E.Y.Dawson		[41,42,63]
Rhizophyllidaceae		
Portieria Zanardini		
<i>Portieria hornemannii</i> (Lyngbye) P.C.Silva	<i>Desmia hornemannii</i> Lyngbye	[41,42,63]
<i>Portieria japonica</i> (Harvey) P.C.Silva		[41,42,63]
Solieriaceae		
Betaphycus Doty		
<i>Betaphycus gelatinus</i> (Esper) Doty ex P.C.Silva	<i>Eucheuma gelatinum</i> (Esper) J.Agardh	[14,26,38,41,42,78]
Eucheuma J.Agardh		
<i>Eucheuma edule</i> (Kützting) Weber Bosse		[41,42,63]
Kappaphycopsis Dumilag & Zuccarello		
<i>Kappaphycopsis cottonii</i> (Weber Bosse) Dumilag & Zuccarello	<i>Kappaphycus cottonii</i> (Weber Bosse) Doty ex H.D.Nguyen & Q.N.Huynh	[41,42,63]
Kappaphycus Doty		
<i>Kappaphycus alvarezii</i> (Doty) L.M.Liao	<i>Kappaphycus alvarezii</i> (Doty) Doty ex P.C. Silva	[41,42,63]
<i>Kappaphycus inermis</i> (F.Schmitz) Doty ex H.D.Nguyen & Q.N.Huynh		
<i>Kappaphycus striatus</i> (F.Schmitz) L.M.Liao	<i>Kappaphycus striatus</i> (F. Schmitz) Doty ex P.C. Silva	[38,41,42]
Meristotheca J.Agardh		
<i>Meristotheca lysonensis</i> X.-V.Nguyen, X.-T.Nguyen, Kittle & McDermid		[52]
Mimica Santiañez & M.J.Wynne		
<i>Mimica arnoldii</i> (Weber Bosse) Santiañez & M.J.Wynne	<i>Eucheuma arnoldii</i> Weber Bosse	[41,42,63]
Solieria J.Agardh		
<i>Solieria robusta</i> (Greville) Kylin		[41,42,63]
Wurdemannia Harvey		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Wurdemannia miniata</i> (Sprengel) Feldmann & Hamel		[38,41,42,67]
Gracilariales		
Gracilariaceae		
<i>Gracilaria</i> Greville		
<i>Gracilaria arcuata</i> Zanardini		[14,23,27,38,41,42,79,80]
<i>Gracilaria articulata</i> C.F.Chang & B.M.Xia		[23,42,79]
<i>Gracilaria blodgettii</i> Harvey		[41,42,63]
<i>Gracilaria bursa-pastoris</i> (S.G.Gmelin) P.C.Silva		[41,42,63]
<i>Gracilaria canaliculata</i> Sonder		[38,41,42]
<i>Gracilaria changii</i> (B.M.Xia & I.A.Abbott) I.A.Abbott, J.Zhang & B.M.Xia	<i>Hydropuntia changii</i> (B.M. Xia & I.A. Abbott) M.J. Wynne	[38,42,80]
<i>Gracilaria chondracantha</i> (Kützing) A.J.K.Millar	<i>Gracilaria bangmeiana</i> J.Zhang & I.A.Abbott	[23,41,42,79]
<i>Gracilaria confervoides</i> f. <i>ecortica</i> V.M.May		[41,42,63]
<i>Gracilaria coronopifolia</i> J.Agardh		[41,42,68,72]
<i>Gracilaria cuneifolia</i> (Okamura) I.K.Lee & Kurogi		[38,41,42]
<i>Gracilaria divergens</i> (C.Agardh) J.Agardh		[41]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	<i>Hydropuntia edulis</i> (S.G.Gmelin) Gurgel & Fredericq	[14,27,38,41,42,67,68]
<i>Gracilaria eucheumatoides</i> Harvey	<i>Hydropuntia eucheumatoides</i> (Harvey) Gurgel & Fredericq	[38,41,42,81]
<i>Gracilaria firma</i> C.F.Chang & B.-M.Xia	<i>Crassiphycus firmus</i> (C.F.Chang & B.-M.Xia) Gurgel, J.N.Norris & Fredericq	[23,38,41,42,80]
<i>Gracilaria fisheri</i> (B.M.Xia & I.A.Abbott) I.A.Abbott, J.Zhang & B.M.Xia	<i>Hydropuntia fisheri</i> (B.M.Xia & I.A.Abbott) M.J.Wynne	[38,41,42,80]
<i>Gracilaria foliifera</i> (Forsskål) Børgesen		[41,42,63]
<i>Gracilaria gigas</i> Harvey		[23,42,79]
<i>Gracilaria gracilis</i> (Stackhouse) Steentoft, L.M.Irvine & Farnham		[41,42,63]
<i>Gracilaria hainanensis</i> C.F.Chang & B.M.Xia		[23,42,79]
<i>Gracilaria longirostris</i> Zhang & Wang		[38,41,42]
<i>Gracilaria phuquocensis</i> N.H.Le, N.Muangmai & G.C.Zuccarello	<i>Gracilaria mammillaris</i> (Montagne) M.Howe	[49]
<i>Gracilaria punctata</i> (Okamura) Yamada		[23,42]
<i>Gracilaria rubra</i> C.F.Chang & B.M.Xia		[41]
<i>Gracilaria salicornia</i> (C.Agardh) E.Y.Dawson		[27,41,42,64,67,68,72,80]
<i>Gracilaria spinulosa</i> (Okamura) Chang & B.-M.Xia		[41,42,68,72]
<i>Gracilaria stellata</i> I.A.Abbott, Zhang & B.M.Xia		[41,42,63]
<i>Gracilaria tenuistipitata</i> C.F.Chang & B.-M.Xia	<i>Agarophyton tenuistipitatum</i> (C.F.Chang et B.-M.Xia) Gurgel, J.N.Norris & Fredericq	[27,38,41,42,67,80]
<i>Gracilaria tenuistipitata</i> var. <i>liui</i> Zhang & Xia		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Gracilaria textorii</i> (Suringar) Hariot	<i>Gracilaria textorii</i> (Suringar) De Toni	[23,41,42,79,80]
<i>Gracilaria vermiculophylla</i> (Ohmi) Papenfuss		[41,42,63]
<i>Gracilaria vieillardii</i> P.C.Silva		[42,79]
<i>Gracilaria yamamotoi</i> Zhang & B.M.Xia		[41,42,63]
Gracilariopsis E.Y.Dawson		
<i>Gracilariopsis chorda</i> (Holmes) Ohmi		[41,42,63]
<i>Gracilariopsis heteroclada</i> J.-F.Zhang & B.-M.Xia	<i>Gracilaria heteroclada</i> J.F.Zhang & B.M.Xia <i>Gracilariopsis bailiniae</i> J.Zhang & B.M.Xia	[41,42,63]
<i>Gracilariopsis longissima</i> (S.G.Gmelin) Steentoft, L.M.Irvine & Farnham		[41,42,63]
<i>Gracilariopsis nganii</i> Pham-Hoàng Hô		[41,42,63]
<i>Gracilariopsis nhatrangensis</i> Nhu Hau Le & S.-M.Lin		[38,41,42]
<i>Gracilariopsis phanthietensis</i> Pham-Hoàng Hô		[41,42,63]
<i>Gracilariopsis rhodotricha</i> E.Y.Dawson		[14,38,41,42]
Hydropuntia Montagne		
<i>Hydropuntia divergens</i> (B.M.Xia & I.A.Abbott) M.J.Wynne		[41,42,63]
Halymeniales		
Grateloupiaceae		
Dermocorynus P.Crouan & H.Crouan		
<i>Dermocorynus dichotomus</i> (J.Agardh) Gargiulo, Morabito & Manghisi	<i>Grateloupia dichotoma</i> J.Agardh	[41,42,63]
Grateloupia C.Agardh		
<i>Grateloupia asiatica</i> S.Kawaguchi & H.W.Wang		[41,42,63]
<i>Grateloupia divaricata</i> Okamura		[14,38,41,42]
<i>Grateloupia filicina</i> (J.V.Lamouroux) C.Agardh		[41,42,68,72]
<i>Grateloupia lithophila</i> Bergesen		[41,42,63]
<i>Grateloupia livida</i> (Harvey) Yamada		[41,42,63]
<i>Grateloupia phuquocensis</i> Tanaka & Pham-Hoàng Hô		[41,42,63]
<i>Grateloupia porracea</i> Kützting		[41,42,63]
<i>Grateloupia prolongata</i> J.Agardh		[41,42,63]
<i>Grateloupia ramosissima</i> Okamura		[38,41,42,67]
Yonagunia S.Kawaguchi & M.Masuda		
<i>Yonagunia formosana</i> (Okamura) Kawaguchi & Masuda	<i>Carpopeltis formosana</i> Okamura <i>Prionitis formosana</i> (Okamura) Kawaguchi & Nguyen H. Dinh	[14,38,41,42,78]
<i>Yonagunia ligulata</i> (Harvey ex Kützting) Manghisi, M.Morabito, De Clerck & Le Gall	<i>Polyopes ligulatus</i> (Harvey ex Kützting) De Toni	[41,42,63]
<i>Yonagunia maillardii</i> (Montagne & Maillardet) Showe M.Lin, Y.-C.Chuang & DeClerck	<i>Carpopeltis maillardii</i> (Montagne & Millardet) Chiang	[38,41,42]
Halymeniaceae		
Cryptonemia J.Agardh		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Cryptonemia undulata</i> Sonder		[41,42,63]
Halymenia C.Agardh		
<i>Halymenia dilatata</i> Zanardini		[14,27,38,41,42,64,78]
<i>Halymenia floresii</i> (Clemente) C.Agardh		[41,42,63]
<i>Halymenia harveyana</i> J.Agardh	<i>Halymenia floresii</i> subsp. <i>harveyana</i> (J.Agardh) Womersley & Lewis	[41,42,63]
<i>Halymenia maculata</i> J.Agardh		[14,27,38,41,42,64,78]
<i>Halymenia malaysiana</i> P.-L.Tan, P.-E.Lim, S.-M.Lin & S.-M.Phang		[50]
Phyllymenia J.Agardh		
<i>Phyllymenia huangiae</i> (Showe M.Lin & H.-Y.Liang) Showe M.Lin, Rodríguez-Prieto, De Clerck & Guiry	<i>Grateloupia huangiae</i> Showe M.Lin & H.-Y.Liang	[82]
<i>Phyllymenia taiwanensis</i> (Showe M.Lin & H.-Y.Liang) Showe M.Lin, Rodríguez-Prieto, De Clerck & Guiry	<i>Grateloupia taiwanensis</i> Showe M.Lin & H.Y.Liang	[56]
Prionitis J.Agardh		
<i>Prionitis vietnamensis</i> Pham-Hoàng Hồ		[41,42,63]
Hildenbrandiales		
Hildenbrandiaceae		
Actinotrichia Decaisne		
<i>Actinotrichia fragilis</i> (Forsskål) Børgesen		[14,27,38,41,42,67]
Nemaliales		
Galaxauraceae		
Dichotomaria Lamarck		
<i>Dichotomaria marginata</i> (J.Ellis & Solander) Lamarck		[41,42,63]
<i>Dichotomaria obtusata</i> (J.Ellis & Solander) Lamarck		[38,41,42]
<i>Dichotomaria papillata</i> (Kjellman) Kurihara & Masuda		[41,42,63]
Galaxaura J.V.Lamouroux		
<i>Galaxaura divaricata</i> (Linnaeus) Huisman & R.A.Townsend		[38,41,42]
<i>Galaxaura filamentosa</i> R.C.Y.Chou		[27,41,42,64,68,72]
<i>Galaxaura rugosa</i> (J.Ellis & Solander) J.V.Lamouroux		[38,41,42]
Hildenbrandia Nardo		
<i>Hildenbrandia rubra</i> (Sommerfelt) Meneghini		[38,41,42,64]
Tricleocarpa Huisman & Borowitzka		
<i>Tricleocarpa cylindrica</i> (J.Ellis & Solander) Huisman & Borowitzka	<i>Galaxaura fastigiata</i> Decaisne	[38,41,42,64,67,83]
<i>Tricleocarpa fragilis</i> (Linnaeus) Huisman & R.A.Townsend		[38,41,42,64]
Liagoraceae		
Akalaphycus Huisman, I.A.Abbott & A.R.Sherwood		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Akalaphycus setchelliae</i> (Yamada) Huisman, I.A.Abbott & A.R.Sherwood		[38,41,42]
Dermonema Harvey ex Heydrich		
<i>Dermonema pulvinatum</i> (Grunow) Fan		[27,38,41,42]
<i>Dermonema virens</i> (J.Agardh) Pedroche & Ávila Ortíz		[38,41,42]
<i>Dermonema zinovae</i> Nguyen Huu Dinh		[41,42,63]
Ganonema K.-C.Fan & Y.-C.Wang		
<i>Ganonema farinosum</i> (J.V.Lamouroux) K.-C.Fan & Y.-C.Wang		[38,41,42,64,67]
<i>Ganonema pinnatum</i> (Harvey) Huisman		[41,42,63]
Helminthocladia J.Agardh		
<i>Helminthocladia australis</i> Harvey		[41,42,63]
Hommersandiophycus S.-M.Lin & J.M.Huisman		
<i>Hommersandiophycus samaensis</i> (C.K.Tseng) S.-M.Lin & Huisman	<i>Ganonema samaense</i> (C.K. Tseng) Huisman	[38,41,42]
Izziella Doty		
<i>Izziella orientalis</i> (J.Agardh) Huisman & Schils		[38,42]
Liagora J.V.Lamouroux		
<i>Liagora ceranoides</i> J.V.Lamouroux		[14,27,38,41,42]
<i>Liagora filiformis</i> K.C.Fan & W.H.Li		[41,42,63]
<i>Liagora hawaiiiana</i> Butters		[41,42,63]
Neozziella S.-M.Lin, S.-Y.Yang & Huisman		
<i>Neozziella divaricata</i> (C.K.Tseng) S.-M.Lin, S.-Y.Yang & Huisman		[38,41,42]
Otohimella Mas.Suzuki		
<i>Otohimella japonica</i> (Yamada) Mas.Suzuki, T.Segawa, Hi.Mori & H.Nozaki	<i>Liagora japonica</i> Yamada	[41,42,63]
Titanophycus Huisman, G.W.Saunders & A.R.Sherwood		
<i>Titanophycus validus</i> (Harvey) Huisman, G.W.Saunders & A.R.Sherwood		[41,42,63]
Scinaiceae		
Scinaia Bivona-Bernardi		
<i>Scinaia boergesenii</i> C.K.Tseng		[41,42,63]
Yamadaellaceae		
Platoma Schousboe ex F.Schmitz		
<i>Platoma cyclocolpum</i> (Montagne) F.Schmitz		[41,42,63]
Yamadaella I.A.Abbott		
<i>Yamadaella caenomyce</i> (Decaisne) I.A.Abbott	<i>Liagora caenomyce</i> Decaisne	[27,38,41,42]
Nemastomatales		
Schizymeniaceae		
Titanophora (J.Agardh) Feldmann		
<i>Titanophora pikeana</i> (Dickie) Feldmann		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Titanophora weberae</i> Børgesen	<i>Titanophora pulchra</i> E.Y. Dawson	[38,41,42,67]
Peyssonneliales		
Peyssonneliaceae		
<i>Agissea</i> Pestana, Lyra, Cassano & J.M.C Nunes		
<i>Agissea inamoena</i> (Pilger) Pestana, Lyra, Cassano & J.M.C. Nunes		[27,38,41,42]
<i>Agissea orientalis</i> (Weber Bosse) Pestana, Lyra, Cassano & J.M.C Nunes	<i>Peyssonnelia rubra</i> f. <i>orientalis</i> Weber Bosse	[38,41,42]
<i>Peyssonnelia</i> Decaisne		
<i>Peyssonnelia boergesenii</i> Weber Bosse		[41]
<i>Peyssonnelia caulifera</i> Okamura		[41,42,63]
<i>Peyssonnelia conchicola</i> Piccone & Grunow		[38,41,42,64,67]
<i>Peyssonnelia rubra</i> (Greville) J.Agardh		[14,38,41,42]
<i>Ramicrusta</i> Zhang Derui & Zhou Jinghua		
<i>Ramicrusta calcea</i> (Heydrich) K.R.Dixon & G.W.Saunders	<i>Peyssonnelia calcea</i> Heydrich	[14,38,41,42]
<i>Sonderophycus</i> Denizot		
<i>Sonderophycus capensis</i> (Montagne) M.J.Wynne	<i>Peyssonnelia gunniana</i> J. Agardh	[41,42,63]
Plocamiales		
Plocamiaceae		
<i>Plocamium</i> J.V.Lamouroux		
<i>Plocamium ovicorne</i> Okamura		[41]
Rhodogorgonales		
Rhodogorgonaceae		
<i>Rhodogorgon</i> J.N.Norris & K.E.Bucher		
<i>Rhodogorgon ramosissima</i> J.N.Norris & Bucher	<i>Rhodogorgon carriebowensis</i> J.N. Norris & Bucher	[27,38,41,42]
Rhodymeniales		
Champiaceae		
<i>Champia</i> Desvaux		
<i>Champia parvula</i> (C.Agardh) Harvey		[27,41,42,64,67,68,72]
<i>Champia salicornioides</i> Harvey		[41,42,63]
<i>Champia vieillardii</i> Kützing		[14,27,41]
<i>Coelothrix</i> Børgesen		
<i>Coelothrix irregularis</i> (Harvey) Børgesen		[41,42,63]
Faucheaceae		
<i>Gloiocladia</i> J.Agardh		
<i>Gloiocladia japonica</i> (Okamura) Yoshida		[41]
Hymenocladaceae		
<i>Asteromenia</i> Huisman & A.J.K.Millar		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Asteromenia anastomosans</i> (Weber Bosse) G.W.Saunders, C.E.Lane, C.W.Schneider & Kraft		[38,41,42]
<i>Asteromenia peltata</i> (W.R.Taylor) Huisman & A.J.K.Millar		[41,42,63]
Lomentariaceae		
<i>Ceratodictyon Zanardini</i>		
<i>Ceratodictyon intricatum</i> (C.Agardh) R.E.Norris		[38,41,42,64]
<i>Ceratodictyon repens</i> (Kützing) R.E.Norris	<i>Gelidiopsis repens</i> (Kützing) Weber Bosse	[41,42,63]
<i>Ceratodictyon scoparium</i> (Montagne & Millardet) R.E.Norris		[38,41,42,67]
<i>Ceratodictyon spongiosum</i> Zanardini		[14,27,41,42,67]
<i>Ceratodictyon variabile</i> (J.Agardh) R.E.Norris		[38,41,42]
<i>Yendoa C.C.Santos, Lyra & J.M.C.Nunes</i>		
<i>Yendoa hakodatensis</i> (Yendo) C.C.Santos, Lyra & J.M.C.Nunes	<i>Lomentaria hakodatensis</i> Yendo	[27,38,41,42]
Rhodymeniaceae		
<i>Botryocladia (J.Agardh) Pfeiffer</i>		
<i>Botryocladia leptopoda</i> (J.Agardh) Kylin		[41,42,63]
<i>Botryocladia skottsbergii</i> (Børgesen) Levring		[41,42,63]
<i>Halichrysis (J.Agardh) F.Schmitz</i>		
<i>Halichrysis micans</i> (Hauptfleisch) P.Huvé & H.Huvé		[41,42,63]
<i>Rhodymenia Greville</i>		
<i>Rhodymenia coacta</i> Okamura & Segawa		[41,42,63]
<i>Rhodymenia intricata</i> (Okamura) Okamura		[41,42,63]
<i>Rhodymenia liniformis</i> Okamura		[41,42,63]
Stylonematales		
Stylonemataceae		
<i>Bangiopsis F.Schmitz</i>		
<i>Bangiopsis dumontioides</i> (P.Crouan & H.Crouan) V.Krishnmurthy		[38,41,42]
<i>Chroodactylon Hansgirg</i>		
<i>Chroodactylon ornatum</i> (C.Agardh) Basson		[27,38,41,42]
<i>Stylonema Reinsch</i>		
<i>Stylonema alsidii</i> (Zanardini) K.M.Drew	<i>Goniotrichum alsidii</i> (Zanardini) M. Howe	[27,41,42,64]
OCHROPHYTA		
Dictyotales		
Dictyotaceae		
<i>Canistrocarpus De Paula & De Clerck</i>		
<i>Canistrocarpus cervicornis</i> (Kützing) De Paula & De Clerck		[38,41,42,64,67,68]
<i>Canistrocarpus crispatus</i> (J.V.Lamouroux) De Paula & De Clerck		[38,41,42]
<i>Dictyopteris J.V.Lamouroux</i>		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Dictyopteris delicatula</i> J.V.Lamouroux		[41,42,63]
<i>Dictyopteris plagiogramma</i> (Montagne) Vickers		[41,42,63]
<i>Dictyopteris polypodioides</i> (De Candolle) J.V.Lamouroux		[38,41,42]
<i>Dictyopteris woodwardia</i> (R.Brown ex Turner) C.Agardh		[38,41,42]
Dictyota J.V.Lamouroux		
<i>Dictyota adnata</i> Zanardini		[41,42,63]
<i>Dictyota bartayresiana</i> J.V.Lamouroux		[38,41,42,64,67,68,72]
<i>Dictyota ceylanica</i> var. <i>anastomosans</i> Yamada		[14,38,41,42]
<i>Dictyota ciliolata</i> Sonder ex Kützing		[38,41,42]
<i>Dictyota dichotoma</i> (Hudson) J.V.Lamouroux		[27,41,42,64]
<i>Dictyota dichotoma</i> var. <i>intricata</i> (C.Agardh) Greville		[41,42,63]
<i>Dictyota friabilis</i> Setchell	<i>Dictyota ceylanica</i> var. <i>rotundata</i> Weber-van-Bosse	[27,41,42,64]
<i>Dictyota grossedentata</i> De Clerck & Coppejans		[41,42,63]
<i>Dictyota hauckiana</i> Nizamuddin		[58]
<i>Dictyota implexa</i> (Desfontaines) J.V.Lamouroux	<i>Dictyota divaricata</i> J.V.Lamouroux	[38,41,42,64]
<i>Dictyota mertensii</i> (C.Martius) Kützing		[41,42,63]
<i>Dictyota pinnatifida</i> Kützing		[41,42,63]
<i>Dictyota polyclada</i> Sonder ex Kützing		[41,42,63]
<i>Dictyota spinulosa</i> Hooker f. & Arnott		[38,41,42,64,67]
Distromium Levring		
<i>Distromium decumbens</i> (Okamura) Levring		[41,42,63]
Lobophora J.Agardh		
<i>Lobophora obscura</i> (Dickie) C.W.Vieira, De Clerck & Payri		[59]
<i>Lobophora papenfussii</i> (W.R.Taylor) Farghaly		[47]
<i>Lobophora tsengii</i> D.Tien & Z.Sun		[60]
<i>Lobophora variegata</i> (J.V.Lamouroux) Womersley ex E.C.Oliveira		[14,38,41,42]
Padina Adanson		
<i>Padina antillarum</i> (Kützing) Piccone		[38,41,42]
<i>Padina arborescens</i> Holmes		[41,42,63]
<i>Padina australis</i> Hauck		[14,27,38,41,42]
<i>Padina australis</i> var. <i>cuneata</i> Tak.Tanaka & K.Nozaawa		[41,42,63]
<i>Padina boryana</i> Thivy		[41,42,72]
<i>Padina gymnospora</i> (Kützing) Sonder		[41,42,68,72]
<i>Padina japonica</i> Yamada		[41,42,63]
<i>Padina minor</i> Yamada		[41,42,63]
<i>Padina tetrastromatica</i> Hauck		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Spatoglossum Kützing		
<i>Spatoglossum schroederi</i> (C.Agardh) Kützing		[41,42,63]
<i>Spatoglossum stipitatum</i> (Tanaka & K.Nozawa) Bittner et al.		[38,41,42]
<i>Spatoglossum vietnamense</i> Pham-Hoàng Hô		[14,38,41,42]
Styopodium Kützing		
<i>Styopodium zonale</i> (J.V.Lamouroux) Papenfuss		[41,42,63]
Ectocarpales		
Acinetosporaceae		
Feldmannia Hamel		
<i>Feldmannia filifera</i> (Børgesen) Pham-Hoàng Hô		[41,42,63]
<i>Feldmannia indica</i> (Sonder) Womersley & A.Bailey	<i>Hincksia indica</i> (Sonder) J.Tanaka	[38,41,42]
<i>Feldmannia irregularis</i> (Kützing) Hamel		[14,38,41,42]
<i>Feldmannia mitchelliae</i> (Harvey) H.-S.Kim	<i>Hincksia mitchelliae</i> (Harvey) P.C.Silva	[27,41,42,64]
Herponema J.Agardh		
<i>Herponema zeylanicum</i> (Børgesen) Krishnamurthy & Baluswami		[41,42,63]
Pylaiella Bory de Saint-Vincent		
<i>Pylaiella littoralis</i> (Linnaeus) Kjellman		[41,42,63]
Chordariaceae		
Acrothrix Kylin		
<i>Acrothrix pacifica</i> Okamura & Yamada		[41,42,63]
Chilionema Sauvageau		
<i>Chilionema ocellatum</i> (Kützing) Kornmann		[14,38,41,42]
Hecatonema Sauvageau		
<i>Hecatonema enhali</i> (Børgesen) M.S.Balakrishnan & V.N.Kinkar		[41,42,63]
Kuetzingiella Kornmann		
<i>Kuetzingiella elachistiformis</i> (Heydrich) M.Balakrishnan & Kinkar		[27,38,41,42,67]
Myrionema Greville		
<i>Myrionema strangulans</i> Greville		[41,42,63]
Nemacystus Derbès & Solier		
<i>Nemacystus decipiens</i> (Suringar) Kuckuck		[41,42,63]
Ectocarpaceae		
Ectocarpus Lyngbye		
<i>Ectocarpus siliculosus</i> (Dillwyn) Lyngbye		[41]
<i>Ectocarpus siliculosus</i> f. <i>sporangioramosus</i> A.D.Zinova & Nguyen Huu Dinh		[41,42,63]
<i>Ectocarpus siliculosus</i> var. <i>dasycarpus</i> (Kuckuck) Gallardo		[41,42,63]
<i>Ectocarpus vungtauensis</i> Pham-Hoàng Hô		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
Scytosiphonaceae		
Chnoospora J.Agardh		
<i>Chnoospora minima</i> (Hering) Papenfuss		[38,41,42,67]
Colpomenia (Endlicher) Derbès & Solier		
<i>Colpomenia sinuosa</i> (Mertens ex Roth) Derbès & Solier		[41,42,68,72]
Dactylosiphon Santiañez, K.M.Lee, S.M.Boo & Kogame		
<i>Dactylosiphon bullosus</i> (D.A.Saunders) Santiañez, K.M.Lee, S.M.Boo & Kogame		[41,42,63]
Hydroclathrus Bory		
<i>Hydroclathrus clathratus</i> (C.Agardh) M.Howe		[38,41,42,67]
<i>Hydroclathrus tenuis</i> C.K.Tseng & Lu Baroen		[38,41,42]
Petalonia Derbès & Solier		
<i>Petalonia fascia</i> (O.F.Müller) Kuntze		[41,42,63]
Pseudochnoospora Santiañez, G.Y.Cho & Kogame		
<i>Pseudochnoospora implexa</i> (J.Agardh) Santiañez, G.Y.Cho & Kogame	<i>Chnoospora implexa</i> J.Agardh	[14,27,38,41,42,67]
Rosenvingea Børgesen		
<i>Rosenvingea endiviifolia</i> (Martius) M.J.Wynne		[27,41,42,64,68,72,84]
<i>Rosenvingea fastigiata</i> (Zanardini) Børgesen		[38,41,42]
<i>Rosenvingea nhatrangensis</i> E.Y.Dawson		[38,41,42,67]
<i>Rosenvingea orientalis</i> (J.Agardh) Børgesen		[14,38,41,42,84]
Scytosiphon C.Agardh		
<i>Scytosiphon lomentaria</i> (Lyngbye) Link		[41,42,63]
Fucales		
Sargassaceae		
Hormophysa Kützing		
<i>Hormophysa cuneiformis</i> (J.F.Gmelin) P.C.Silva		[41,42,63]
Sargassum C.Agardh		
<i>Sargassum aemulum</i> var. <i>carpophylloides</i> Grunow		[41,42,63]
<i>Sargassum aemulum</i> var. <i>jouanii</i> Grunow		[41,42,63]
<i>Sargassum angustifolium</i> C.Agardh		[42,85]
<i>Sargassum aquifolium</i> (Turner) C.Agardh		[41,42,85]
<i>Sargassum armatum</i> J.Agardh		[41,42,63]
<i>Sargassum assimile</i> Harvey		[41,42,63]
<i>Sargassum baccularia</i> (Mertens) C.Agardh		[41,42,63]
<i>Sargassum bangmeianum</i> Nguyen Huu Dinh & Huynh Quang Nang		[38,41,42]
<i>Sargassum baorenii</i> Nguyen Huu Dinh & Huynh Quang Nang		[38,41,42]
<i>Sargassum bicornis</i> J.Agardh		[20,38,41,42,85]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Sargassum brevifolium</i> var. <i>pergracile</i> Grunow		[41,42,63]
<i>Sargassum bulbiferum</i> Yoshida		[41,42,63]
<i>Sargassum buui</i> Nguyen Huu Dinh & Huynh Quang Nang		[38,41,42]
<i>Sargassum capillare</i> Kützing		[41,42,63]
<i>Sargassum carpophyllum</i> J.Agardh		[20,38,41,42]
<i>Sargassum carpophyllum</i> var. <i>honomense</i> Nguyen Huu Dinh & Huynh Quang Nang		[41,42,63]
<i>Sargassum carpophyllum</i> var. <i>nhatrangense</i> (Pham-Hoang Ho) Ajisaka		[41,42,63]
<i>Sargassum cinereum</i> J.Agardh		[41,42,63]
<i>Sargassum confusum</i> C.Agardh		[41,42,63]
<i>Sargassum congkinhii</i> Pham-Hoàng Hô		[20,38,41,42,85]
<i>Sargassum cornutifructum</i> Nguyen Huu Dinh & Huynh Quang Nang		[41,42,63]
<i>Sargassum cotoense</i> Nguyen Huu Dai		[41,42,63]
<i>Sargassum cymosum</i> C.Agardh		[42,85]
<i>Sargassum denticarpum</i> Ajisaka		[41,42,63]
<i>Sargassum distichum</i> Sonder		[38,41,42]
<i>Sargassum emarginatum</i> C.K.Tseng & Lu Baroen		[38,41,42]
<i>Sargassum feldmannii</i> Pham-Hoàng Hô		[38,41,42,85]
<i>Sargassum flavicans</i> (Mertens) C.Agardh		[38,41,42,85]
<i>Sargassum glaucescens</i> J.Agardh		[38,41,42,85]
<i>Sargassum gracillimum</i> Reinbold		[41,42,63]
<i>Sargassum graminifolium</i> C.Agardh		[41,42,63]
<i>Sargassum hemiphyllum</i> (Turner) C.Agardh		[14,38,41,42]
<i>Sargassum henslowianum</i> J.Agardh		[38,41,42,85]
<i>Sargassum henslowianum</i> var. <i>bellonae</i> Grunow		[41,42,63]
<i>Sargassum herklotsii</i> Setchell		[41,42,63]
<i>Sargassum hieui</i> Nguyen Huu Dinh & Huynh Quang Nang		[38,41,42]
<i>Sargassum ilicifolioides</i> Tseng & Lu		[41,42,63]
<i>Sargassum ilicifolium</i> (Turner) C.Agardh		[20,38,41,42,85]
<i>Sargassum incanum</i> Grunow		[42,85]
<i>Sargassum kuetzingii</i> Setchell		[20,38,41,42,85]
<i>Sargassum longifructum</i> C.K.Tseng & B.Lu		[38,41,42,85]
<i>Sargassum mclurei</i> f. <i>duplicatum</i> A.D.Zinova & Nguyen Huu Dinh		[41,42,63]
<i>Sargassum mclurei</i> Setchell		[20,41,42,64,68,72]
<i>Sargassum microcystum</i> J.Agardh		[27,41,42,64,68,72,85]
<i>Sargassum miyabei</i> Yendo		[38,41,42,64,67,68]
<i>Sargassum namoense</i> Nguyen Huu Dai		[42,85]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Sargassum natans</i> (Linnaeus) Gaillon		[38,41,42]
<i>Sargassum nipponicum</i> Yendo		[41,42,63]
<i>Sargassum oligocystum</i> Montagne		[20,27,38,41,42,67,85]
<i>Sargassum paniculatum</i> J.Agardh		[42,85]
<i>Sargassum parvifolium</i> (Turner) C.Agardh		[41,42,63]
<i>Sargassum parvivesiculosum</i> C.K.Tseng & B.Lu		[41,42,63]
<i>Sargassum phamhoangii</i> Nguyen Huu Dai		[41,42,63]
<i>Sargassum phyllocystum</i> C.K.Tseng & Lu Baroen		[41,42,63]
<i>Sargassum piluliferum</i> (Turner) C.Agardh		[42,86]
<i>Sargassum piluliferum</i> var. <i>serratifolium</i> Yamada		[42,85]
<i>Sargassum polycystum</i> C.Agardh		[27,41,42,64,67,68,72,85]
<i>Sargassum polyporum</i> Montagne		[42,85]
<i>Sargassum quinhonense</i> Nguyen Huu Dai		[41,42,63]
<i>Sargassum segii</i> Yoshida		[41,42,63]
<i>Sargassum serratum</i> Nguyen Huu Dai		[38,41,42,85]
<i>Sargassum siliquosum</i> J.Agardh		[38,41,42,85]
<i>Sargassum subtilissimum</i> C.K.Tseng & B.Lu		[41,42,63]
<i>Sargassum swartzii</i> C.Agardh		[38,41,42,67,85]
<i>Sargassum tenerrimum</i> J.Agardh		[41,42,63]
<i>Sargassum tsengii</i> Nguyen Huu Dinh & Huynh Quang Nang		[38,41,42]
<i>Sargassum turbinarioides</i> Grunow		[41,42,63]
<i>Sargassum vachellianum</i> Greville		[42,85]
<i>Sargassum vietnamense</i> A.D.Zinova & Dinh		[42,85]
<i>Sargassum virgatum</i> C.Agardh		[38,41,42,85]
Turbinaria J.V.Lamouroux		
<i>Turbinaria conoides</i> (J.Agardh) Kützing		[41,42,63]
<i>Turbinaria decurrens</i> Bory		[41,42,63]
<i>Turbinaria gracilis</i> Sonder		[20,38,41,42,85]
<i>Turbinaria ornata</i> (Turner) J.Agardh		[41,42,68,72]
<i>Turbinaria ornata</i> var. <i>prolifera</i> Pham Hoàng Hô		[41,42,63]
<i>Turbinaria parvifolia</i> C.K.Tseng & Lu Baroen		[41,42,63]
<i>Turbinaria turbinata</i> (Linnaeus) Kuntze		[41,42,63]
Ishigeales		
Petrodermataceae		
<i>Petroderma</i> Kuckuck		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Petroderma vietnamense</i> Pham-Hoàng Hồ		[41,42,63]
Ralfsiales		
Mesosporaceae		
<i>Mesospora</i> Weber-van Bosse		
<i>Mesospora schmidtii</i> Weber Bosse		[38,41,42]
Neoralfsiaceae		
<i>Neoralfsia</i> P.-E.Lim & H.Kawai		
<i>Neoralfsia expansa</i> (J.Agardh) P.-E.Lim & H.Kawai ex Cormaci & G.Furnari		[38,41,42,64]
Pseudoralfsiaceae		
<i>Pseudoralfsia</i> Parente, Fletcher & G.W.Saunders		
<i>Pseudoralfsia verrucosa</i> (Areschoug) Parente, Fletcher & G.W.Saunders	<i>Ralfsia verrucosa</i> (Areschoug) Areschoug	[41,42,63]
Ralfsiaceae		
<i>Ralfsia</i> Berkeley		
<i>Ralfsia fungiformis</i> (Gunnerus) Setchell & N.L.Gardner		[41,42,63]
Scytothamiales		
Asteronemataceae		
<i>Asteronema</i> Delépine & Asensi		
<i>Asteronema breviararticulatum</i> (J.Agardh) Ouriques & Bouzon		[38,41,42,64]
Sphacelariales		
Sphacelariaceae		
<i>Sphacelaria</i> Lyngbye		
<i>Sphacelaria carolinensis</i> Trono		[41,42,63]
<i>Sphacelaria ceylanica</i> Sauvageau		[41,42,63]
<i>Sphacelaria novae-hollandiae</i> Sonder		[14,27,38,41,42]
<i>Sphacelaria rigidula</i> Kützing	<i>Sphacelaria divaricata</i> Montagne	[27,41,42,64]
<i>Sphacelaria solitaria</i> (Pringsheim) Kylin		[41,42,63]
<i>Sphacelaria tribuloides</i> Meneghini		[14,27,38,41,42]
CHLOROPHYTA		
Bryopsidales		
Bryopsidaceae		
<i>Bryopsis</i> J.V.Lamouroux		
<i>Bryopsis chapmanii</i> Molinari & Guiry		[87]
<i>Bryopsis hypnoides</i> J.V. Lamouroux		[41,42,63]
<i>Bryopsis indica</i> A. Gepp & E.S. Gepp		[14,38,41,42,87]
<i>Bryopsis pennata</i> J.V. Lamouroux		[27,38,41,42,64,87]
<i>Bryopsis pennata</i> var. <i>secunda</i> (Harvey) Collins & Hervey		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Bryopsis plumosa</i> (Hudson) C. Agardh		[27,38,41,42,64,87]
Trichosolen Montagne		
<i>Trichosolen mucronatus</i> (Børgesen) W.R. Taylor		[41,42,63]
<i>Trichosolen parvus</i> (E.Y. Dawson) W.R. Taylor	<i>Pseudobryopsis parva</i> E.Y.Dawson	[20,38,41,42,87]
Caulerpaceae		
Caulerpa J.V.Lamouroux		
<i>Caulerpa ambigua</i> Okamura	<i>Caulerpella ambigua</i> (Okamura) Prud'homme & Lokhorst	[14,38,41,42,87]
<i>Caulerpa andamanensis</i> (W.R.Taylor) Draisma, Prudhomme & Sauvage		[41,42,63]
<i>Caulerpa ashmeadii</i> Harvey		[42,87]
<i>Caulerpa brachypus</i> Harvey		[41,42,63]
<i>Caulerpa chemnitzia</i> (Esper) J.V.Lamouroux	<i>Caulerpa peltata</i> J.V.Lamouroux; <i>Caulerpa racemosa</i> var. <i>occidentalis</i> (J.Agardh) Børgesen; <i>Caulerpa racemosa</i> var. <i>peltata</i> (J.V.Lamouroux) Eubank	[41]
<i>Caulerpa chemnitzia</i> var. <i>laetevirens</i> (Montagne) Fernández-García & Riosmena-Rodriguez		[41,42,63]
<i>Caulerpa corynephora</i> Montagne		[41,42,63]
<i>Caulerpa cupressoides</i> (Vahl) C. Agardh		[27,38,41,42,67,87]
<i>Caulerpa cupressoides</i> var. <i>flabellata</i> Børgesen		[41]
<i>Caulerpa cupressoides</i> var. <i>lycopodium</i> Weber Bosse		[41,42,63]
<i>Caulerpa cupressoides</i> var. <i>mamillosa</i> (Montagne) Weber Bosse		[41,42,63]
<i>Caulerpa falcifolia</i> Harvey & Bailey		[41,42,63]
<i>Caulerpa fastigiata</i> Montagne		[14,38,41,42,67,87]
<i>Caulerpa lamourouxii</i> (Turner) C.Agardh	<i>Caulerpa racemosa</i> var. <i>lamourouxii</i> (Turner) Weber Bosse	
<i>Caulerpa lentillifera</i> J. Agardh		[38,41,42,87]
<i>Caulerpa macrodisca</i> Decaisne	<i>Caulerpa peltata</i> var. <i>macrodisca</i> (Decaisne) Weber Bosse	[14,38,41,42,87]
<i>Caulerpa mexicana</i> f. <i>vietnamica</i> Pham H.H.		[41,42,63]
<i>Caulerpa mexicana</i> Sonder ex Kützing		[41,42,63]
<i>Caulerpa microphysa</i> (Weber Bosse) Feldmann		[38,41,42,64,87]
<i>Caulerpa minuta</i> L.N.Hau		[39]
<i>Caulerpa nummularia</i> Harvey ex J. Agardh		[38,41,42,64,67,68]
<i>Caulerpa oligophylla</i> Montagne		[41,42,63]
<i>Caulerpa racemosa</i> (Forsskål) J. Agardh		[14,27,38,41,42,67,87]
<i>Caulerpa racemosa</i> f. <i>vietnamensis</i> A.D. Zinova & Nguyen H. Dinh		[42,87]
<i>Caulerpa racemosa</i> var. <i>macrophysa</i> (Sonder ex Kützing) W.R. Taylor	<i>Caulerpa macrophysa</i> (Sonder ex Kützing) G.Murray	[14,27,38,41,42,67,87]
<i>Caulerpa scalpelliformis</i> (R. Brown ex Turner) C. Agardh		[42,87]
<i>Caulerpa serrulata</i> (Forsskål) J. Agardh	<i>Caulerpa freycinetii</i> C.Agardh	[14,27,38,41,42,67,68,72]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Caulerpa serrulata</i> f. <i>lata</i> (Weber Bosse) C.K. Tseng		[41,42,64]
<i>Caulerpa serrulata</i> var. <i>boryana</i> (J. Agardh) Gilbert		[41,42,63]
<i>Caulerpa sertularioides</i> (S.G. Gmelin) M. Howe		[41,42,63]
<i>Caulerpa sertularioides</i> f. <i>longipes</i> (J. Agardh) Collins		[41]
<i>Caulerpa taxifolia</i> (Vahl) C. Agardh		[41,42,63]
<i>Caulerpa urvilleana</i> Montagne	<i>Caulerpa cupressoides</i> var. <i>urvilleana</i> (Montagne) L.M.Hodgson, P.H. Tri, K.Lewmanomont & K.J. McDermid	[41,42,63]
<i>Caulerpa verticillata</i> J. Agardh		[14,27,38,41,42,87]
<i>Caulerpa verticillata</i> f. <i>charoides</i> Weber Bosse		[41,42,63]
<i>Caulerpa webbiana</i> Montagne		[41,42,63]
<i>Caulerpa webbiana</i> f. <i>tomentella</i> (Harvey ex J. Agardh) Weber Bosse		[41,42,63]
<i>Caulerpa sertularioides</i> f. <i>longiseta</i> (Bory) Svedelius		[41,42,63]
Codiaceae		
Codium Stackhouse		
<i>Codium adhaerens</i> C. Agardh		[14,38,41,42,87]
<i>Codium arabicum</i> Kützing		[14,38,41,42,67,87]
<i>Codium cylindricum</i> Holmes		[41,42,63]
<i>Codium duthieae</i> P.C. Silva		[42,87]
<i>Codium formosanum</i> Yamada		[41,42,63]
<i>Codium geppiorum</i> O.C. Schmidt		[27,38,41,42,64,87]
<i>Codium intricatum</i> Okamura		[41,42,63]
<i>Codium isthmocladum</i> Vickers		
<i>Codium mamillosum</i> Harvey		[38]
<i>Codium repens</i> P.L. Crouan & H.M. Crouan		[38,41,42,64,67,87]
<i>Codium tomentosum</i> Stackhouse		[41,42,63]
Geppella Borgesen		
<i>Geppella prolifera</i> C.K. Tseng & M.L. Dong		[41,42,63]
Derbesiaceae		
Derbesia Solier		
<i>Derbesia attenuata</i> E.Y. Dawson		[14,27,38,41,42,87]
<i>Derbesia marina</i> (Lyngbye) Solier		[42,87]
Halicystis Areschoug		
<i>Halicystis pyriformis</i> Levring		[41,42,63]
Dichotomosiphonaceae		
Avrainvillea Decaisne		
<i>Avrainvillea amadelpa</i> (Montagne) A. Gepp & E. Gepp		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Avrainvillea erecta</i> (Berkeley) A. Gepp & E. Gepp		[14,38,41,42,67,87]
<i>Avrainvillea lacerata</i> Harvey ex J. Agardh		[41,42,63]
<i>Avrainvillea obscura</i> (C. Agardh) J. Agardh	<i>Avrainvillea capituliformis</i> T.Tanaka	[41,42,63]
Halimedaceae		
Boodleopsis A.Gepp & E.S.Gepp		
<i>Boodleopsis pusilla</i> (Collins) W.R.Taylor, A.B.Joly & Bernatowicz		[41,42,63]
Chlorodesmis Harvey & Bailey		
<i>Chlorodesmis hildebrandtii</i> A. Gepp & E. Gepp		[14,38,41,42,87]
Halimeda J.V.Lamouroux		
<i>Halimeda cuneata</i> f. <i>digitata</i> E.S. Barton		[14,38,41,42]
<i>Halimeda cuneata</i> Hering		[41,42,63]
<i>Halimeda cylindracea</i> Decaisne		[41,42,63]
<i>Halimeda discoidea</i> Decaisne		[14,27,41,42,67,87]
<i>Halimeda gracilis</i> Harvey ex J. Agardh		[38,41,42]
<i>Halimeda incrassata</i> (J. Ellis) J.V. Lamouroux		[14,38,41,42,87]
<i>Halimeda macroloba</i> Decaisne		[41,42,63]
<i>Halimeda micronesica</i> Yamada		[41,42,63]
<i>Halimeda opuntia</i> (Linnaeus) J.V. Lamouroux		[14,27,38,41,42,67,68,72,87]
<i>Halimeda taenicola</i> W.R. Taylor		
<i>Halimeda tuna</i> (J. Ellis et Solander) J.V. Lamouroux		[14,38,41,42,87]
<i>Halimeda velasquezii</i> W.R. Taylor		[41,42,63]
<i>Halimeda xishaensis</i> C.K. Tseng & M.L. Dong		[41,42,63]
Penicillus Lamarck		
<i>Penicillus sibogae</i> A. Gepp & E. Gepp		[14,38,41,42,87]
Pseudochlorodesmis Børgesen		
<i>Pseudochlorodesmis furcellata</i> (Zanardini) Børgesen		[14,38,41,42,87]
Rhipidosiphon Montagne		
<i>Rhipidosiphon javensis</i> Montagne	<i>Udotea javensis</i> (Montagne) A.Gepp & E.S.Gepp	[14,38,41,42,87]
Rhipiliopsis A.Gepp & E.S.Gepp		
<i>Rhipiliopsis echinocaulos</i> (A.B. Cribb) Farghaly		[41,42,63]
Tydemanina Weber Bosse		
<i>Tydemanina expeditionis</i> Weber Bosse		[41,42,63]
Udotea J.V.Lamouroux		
<i>Udotea argentea</i> Zanardini		[41,42,63]
<i>Udotea flabellum</i> (J. Ellis & Solander) M. Howe		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Udotea orientalis</i> A. Gepp & E. Gepp		[41,42,63]
<i>Udotea velutina</i> C.K. Tseng & M.L. Dong		[41,42,63]
Ostreobiaceae		
<i>Ostreobium</i> Bornet & Flahault		
<i>Ostreobium quekettii</i> Bornet & Flahault		[38,41,42]
Pseudobryopsidaceae		
<i>Pseudobryopsis</i> Berthold		
<i>Pseudobryopsis hainanensis</i> C.K. Tseng		[41,42,63]
Cladophorales		
Anadyomenaceae		
<i>Anadyomene</i> J.V.Lamouroux		
<i>Anadyomene plicata</i> C.Agardh		[14,38,41,42,67,87]
<i>Anadyomene wrightii</i> Harvey ex J.E. Gray		[14,27,41,42,67,87]
<i>Microdictyon</i> Decaisne		
<i>Microdictyon japonicum</i> Setchell		[27,38,41,42,67]
<i>Microdictyon nigrescens</i> (Yamada) Setchell		[42,87]
<i>Microdictyon okamuriae</i> Setchell		[38,41,42,64,87]
<i>Microdictyon vanbosseae</i> Setchell		[42,87]
Boodleaceae		
<i>Boodlea</i> G.Murray & De Toni		
<i>Boodlea coacta</i> (Dickie) G. Murray & De Toni		[41,42,63]
<i>Boodlea composita</i> (Harvey) F. Brand		[14,27,38,41,42]
<i>Boodlea struveoides</i> M. Howe		[38,41,42,64,87]
<i>Cladophoropsis</i> Børgesen		
<i>Cladophoropsis fasciculata</i> (Kjellman) Wille		[38,41,42,64]
<i>Cladophoropsis membranacea</i> (Hofman ex C. Agardh) Børgesen		[14,27,38,41,42,87]
<i>Cladophoropsis phamhoanghooi</i> Molinari & Guiry		[14,27,38,41,42,87]
<i>Cladophoropsis vaucheriiformis</i> (Areschoug) Papenfuss		[14,27,38,41,42,87]
<i>Phyllodictyon</i> J.E.Gray		
<i>Phyllodictyon anastomosans</i> (Harvey) Kraft & M.J.		[38,41,42,64]
Cladophoraceae		
<i>Chaetomorpha</i> Kützing		
<i>Chaetomorpha aerea</i> (Dillwyn) Kützing	<i>Chaetomorpha crassa</i> (C.Agardh) Kützing	[14,38,41,42,87]
<i>Chaetomorpha antennina</i> (Bory) Kützing		[38,41,42,87]
<i>Chaetomorpha basiretrorsa</i> Setchell		[41,42,63]
<i>Chaetomorpha gracilis</i> Kützing		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Chaetomorpha indica</i> (Kützinger) Kützinger		[38,41,42,64,87]
<i>Chaetomorpha javanica</i> Kützinger		[41,42,63]
<i>Chaetomorpha linum</i> (O.F. Müller) Kützinger		[41,42,63]
<i>Chaetomorpha pachynema</i> (Montagne) Kützinger		[42,87]
<i>Chaetomorpha spiralis</i> Okamura		[38,41,42,87]
<i>Chaetomorpha tortuosa</i> (Dillwyn) Kleen		[42,87]
Cladophora Kützinger		
<i>Cladophora adhaerens</i> Harvey		[41,42,63]
<i>Cladophora albida</i> (Nees) Kützinger		[41,42,63]
<i>Cladophora aokii</i> Yamada		[41,42,63]
<i>Cladophora catenata</i> (Linnaeus) Kützinger		[38,41,42]
<i>Cladophora coelothrix</i> Kützinger		[41,42,63]
<i>Cladophora crispula</i> Vickers		[41,42,63]
<i>Cladophora flexuosa</i> (O.F. Müller) Kützinger		[38,41,42,64]
<i>Cladophora glomerata</i> (Linnaeus) Kützinger		[41,42,63]
<i>Cladophora gracilis</i> Kützinger		[41,42,63]
<i>Cladophora laetevirens</i> (Dillwyn) Kützinger		[27,38,41,42,64,87]
<i>Cladophora papenfussii</i> Pham H.H.		[41,42,63]
<i>Cladophora patentiramea</i> (Montagne) Kützinger		[41,42,63]
<i>Cladophora perpusilla</i> Skottsberg & Levring		[38,41,42,87]
<i>Cladophora prolifera</i> (Roth) Kützinger		[38,41,42]
<i>Cladophora rugulosa</i> G.Martens		[41,42,63]
<i>Cladophora sericea</i> (Hudson) Kützinger		[41,42,63]
<i>Cladophora socialis</i> Kützinger		[14,27,38,41,42]
<i>Cladophora stimpsonii</i> Harvey		[42,87]
<i>Cladophora vagabunda</i> (Linnaeus) C. Hoek	<i>Cladophora uncinella</i> Harvey; <i>Cladophora inserta</i> f. <i>ungulata</i> (Brand) Setchell	[27,38,41,42,67,68]
Lychaete J.Agardh		
<i>Lychaete herpestica</i> (Montagne) M.J.Wynne		[38,42]
<i>Lychaete pellucida</i> (Hudson) M.J.Wynne		[42,87]
<i>Lychaete sakaii</i> (I.A.Abbott) M.J.Wynne		[41,42,63]
Rhizoclonium Kützinger		
<i>Rhizoclonium grande</i> Børgesen		[14,38,41,42,87]
<i>Rhizoclonium riparium</i> (Roth) Harvey	<i>Rhizoclonium implexum</i> (Dillwyn) Kützinger	[38,41,42,64,87]
Siphonocladaceae		
Boergesenia J.Feldmann		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Boergesenia forbesii</i> (Harvey) Feldmann		[14,38,41,42,87]
Dictyosphaeria Decaisne		
<i>Dictyosphaeria cavernosa</i> (Forsskål) Børgesen		[14,27,41,42,67,87]
<i>Dictyosphaeria spinifera</i> C.K. Tseng & C.F. Chang		[42,87]
<i>Dictyosphaeria versluysii</i> Weber Bosse		[14,27,38,41,42]
Valoniaceae		
Valonia C.Agardh		
<i>Valonia aegagropila</i> C. Agardh		[14,38,41,42,67,87]
<i>Valonia fastigiata</i> Harvey ex J. Agardh		[14,38,41,42,67,87]
<i>Valonia macrophysa</i> Kützing		
<i>Valonia utricularis</i> (Roth) C. Agardh		[14,38,41,42,67,87]
<i>Valonia ventricosa</i> J. Agardh		[41,42,63]
Valoniopsis Børgesen		
<i>Valoniopsis pachynema</i> (G. Martens) Børgesen		[41,42,63]
Dasycladales		
Bornetellaceae		
Bornetella Munier-Chalmas		
<i>Bornetella nitida</i> Munier-Chalmas ex Sonder		[41,42,63]
<i>Bornetella oligospora</i> Solms-Laubach		[41,42,63]
<i>Bornetella sphaerica</i> (Zanardini) Solms-Laubach		[14,38,41,42,67,87]
Dasycladaceae		
Neomeris J.V.Lamouroux		
<i>Neomeris annulata</i> Dickie		[14,27,41,42,67,87]
<i>Neomeris bilimbata</i> J.T. Koster		[38,41,42,64,87]
<i>Neomeris vanbosseae</i> M. Howe		[14,27,41,42,67,87]
Polyphysaceae		
Acetabularia J.V.Lamouroux		
<i>Acetabularia caliculus</i> J.V.Lamouroux		[14,38,41,42,67,87]
<i>Acetabularia major</i> G. Martens		[41,42,63]
Parvocaulis S.Berger, U.Fettweiss, S.Gleissberg, L.B.Liddle, U.Richter, H.Sawitzky & G.C.Zuccarello		
<i>Parvocaulis clavatus</i> (Yamada) S. Berger, U. Fettweiss, S. Gleissberg, L.B. Liddle, U. Richter, H. Sawitzky & Zuccarello		[38,41,42]
<i>Parvocaulis exiguus</i> (Solms-Laubach) S.Berger, Fettweiss, Gleissberg, Liddle, U.Richter, Sawitzky & Zuccarello		[41,42,63]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Parvocaulis parvulus</i> (Solms-Laubach) S. Berger, U. Fettweiss, S. Gleissberg, L.B. Liddle, U. Richter, H. Sawitzky & Zuccarello		[38,41,42]
<i>Parvocaulis pusillus</i> (M. Howe) S. Berger, U. Fettweiss, S. Gleissberg, L.B. Liddle, U. Richter, H. Sawitzky & Zuccarello		[38,41,42]
Ulotrichales		
Gayraliaceae		
<i>Gayralia</i> K.L. Vinogradova		
<i>Gayralia oxysperma</i> (Kützing) K.L. Vinogradova ex Scagel, R.F., Gabrielson, P.W., Garbary, D.J., Golden, L., Hawkes, M.W., Lindstrom, S.C., Oliveira, J.C. & Widdowson, T.B.		[41,42,63]
Gomontiaceae		
<i>Gomontia</i> Bornet & Flahault		
<i>Gomontia arrhiza</i> Hariot		[41,42,63]
Monostromataceae		
<i>Monostroma</i> Thuret		
<i>Monostroma nitidum</i> Wittrock	<i>Porphyra crispata</i> Kjellman	[14,38,41,42,87]
Ulotrichaceae		
<i>Ulothrix</i> Kützing		
<i>Ulothrix flacca</i> (Dillwyn) Thuret		[42,87]
<i>Ulothrix subflaccida</i> Wille		[41,42,63]
Ulvales		
Ulvaceae		
<i>Enteromorpha</i> Link		
<i>Enteromorpha stipitata</i> var. <i>catbaensis</i> A.D. Zinova & Nguyen H. Dinh		[87]
<i>Ulva</i> Linnaeus		
<i>Ulva aragoënsis</i> (Bliding) Maggs		[61]
<i>Ulva chaetomorphoides</i> (Børgesen) Hayden, Blomster, Maggs, P.C. Silva, M.J. Stanhope et J.R. Waaland		[38,41,42,64,72]
<i>Ulva chaugulii</i> M.G.Kavale & M.A.Kazi		[61]
<i>Ulva clathrata</i> (Roth) C. Agardh		[38,41,42,88]
<i>Ulva compressa</i> Linnaeus		[38,41,42,64,72]
<i>Ulva conglobata</i> Kjellman		[38,41,42,64,72]
<i>Ulva fenestrata</i> Postels & Ruprecht		[38,41,42,64,72]
<i>Ulva flexuosa</i> subsp. <i>pilifera</i> (Kützing) M.J. Wynne		[41,42,63]
<i>Ulva flexuosa</i> Wulfen		[38,41,42,64,72]
<i>Ulva intestinalis</i> Linnaeus		[38,41,42,64,72]
<i>Ulva kraftiorum</i> Huisman		[61]

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Ulva kyllinii</i> (Bliding) Hayden, Blomster, Maggs, P.C. Silva, M.J. Stanhope et J.R. Waaland		[38,41,42,64,72]
<i>Ulva lactuca</i> Linnaeus		[38,41,42,64,72]
<i>Ulva limnetica</i> K.Ichihara & S.Shimada		[61]
<i>Ulva meridionalis</i> R.Horimoto & S.Shimada		[61]
<i>Ulva ohnoi</i> M.Hiraoka & S.Shimada		[61]
<i>Ulva papenfussii</i> Pham H.H.		[38,41,42,64,72]
<i>Ulva prolifera</i> O.F. Müller		[38,41,42,64,72]
<i>Ulva ralfsii</i> (Harvey) Le Jolis		[38,41,42,64,72]
<i>Ulva reticulata</i> Forsskål		[27,38,41,42,64,87,88]
<i>Ulva spinulosa</i> Okamura & Segawa		[42,87]
<i>Ulva tepida</i> Y.Masakiyo & S.Shimada		[61]
<i>Ulva torta</i> (Mertens) Trevisan		[41,42,63]
<i>Ulva vietnamensis</i> L.-A.T.Tran, Leliaert & De Clerck		[61]
Ulvellaceae		
<i>Ulvella</i> P.L.Crouan & H.M.Crouan		
<i>Ulvella lens</i> P.L. Crouan et H.M. Crouan		[14,38,41,42,87]
<i>Ulvella leptochaete</i> (Huber) R.Nielsen, C.J.O’Kelly & B.Wysor		[41,42,63]
<i>Ulvella scutata</i> (Reinke) R.Nielsen, C.J.O’Kelly & B.Wysor		[41,42,63]
<i>Ulvella viridis</i> (Reinke) R. Nielsen, C.J. O’Kelly & B. Wysor		[41,42,63]
Vaucheriales		
Vaucheriaceae		
<i>Vaucheria</i> A.P.de Candolle		
<i>Vaucheria piloboloides</i> Thuret		[41,42,63]
TRACHEOPHYTA		
Alismatales		
Cymodoceaceae		
<i>Cymodocea</i> K.D.König		
<i>Cymodocea rotundata</i> Ascherson & Schweinfurth		[89]
<i>Halodule</i> Endlicher		
<i>Halodule pinifolia</i> (Miki) Hartog		[44]
<i>Halodule uninervis</i> (Forsskål) Ascherson		[44]
<i>Oceana</i> Byng & Christenhusz		
<i>Oceana serrulata</i> (R.Brown) Byng & Christenhusz	<i>Cymodocea serrulata</i> (R.Brown) Ascherson & Magnus	[89]
<i>Syringodium</i> Kützing		
<i>Syringodium isoetifolium</i> (Ascherson) Dandy		[44]
<i>Thalassodendron</i> Den Hartog		

Table 2. Cont.

Species Name	Regionally Reported as	References
<i>Thalassodendron ciliatum</i> (Forsskål) Hartog		[89]
Hydrocharitaceae		
<i>Enhalus</i> L.C. Richard		
<i>Enhalus acoroides</i> (Linnaeus f.) Royle		[89]
<i>Halophila</i> Thouars		
<i>Halophila beccarii</i> Ascherson		[44]
<i>Halophila decipiens</i> Ostenfeld		[44]
<i>Halophila major</i> (Zollinger) Miquel		[44]
<i>Halophila minor</i> (Zollinger) Hartog		[90]
<i>Halophila ovalis</i> (R.Brown) Hooker f.		[44,90]
<i>Thalassia</i> Banks ex König		
<i>Thalassia hemprichii</i> (Ehrenberg) Ascherson		[89]
Ruppiaaceae		
<i>Ruppia</i> Linnaeus		
<i>Ruppia maritima</i> Linnaeus		[91]
Zosteraceae		
<i>Zostera</i> Linnaeus		
<i>Zostera japonica</i> Ascherson & Graebner		[92]

3. Molecular-Assisted Alpha Taxonomy of the Vietnamese Marine Flora

The use of molecular-assisted alpha taxonomy of marine algae is very recent in Vietnam [24,93]. Molecular tools are presently needed among other purposes: (1) validate previous species identification, (2) identify new records and species, and (3) detect introduced species (e.g., [94–96]). Studies combining DNA-based species delimitation techniques and detailed morphological observations have refined our knowledge on Vietnamese species taxonomy and on the individual species' biogeographical ranges. Nevertheless, such efforts have been very limited in the last decade. Among the 59 new records and species made in the last 10 years, only 25.4% (15 species) were based on molecular-assisted alpha taxonomy. Hereafter, we reviewed molecular studies conducted thus far on Vietnamese marine macroalgae, identifying the taxa studied, marker used, and taxonomic results.

3.1. Molecular-Assisted Alpha Taxonomy of Rhodophyta

Molecular-assisted alpha taxonomic studies on Rhodophyta have comprised a total of four markers, analyzed individually or combined, consisting of two chloroplast genes (large subunit of ribulose-1,5-bisphosphate-carboxylase-oxygenase (*rbcL*); photosystem I P700 chlorophyll a apoprotein A1 (*psaA*)), one mitochondrial gene (cytochrome c oxidase I (*cox1*)), and one nuclear gene (LSU rDNA (28S)). The plastidic *rbcL* gene has been mostly used. The 2006 publication by Hau et al. [24] conducted one of the first molecular studies on Vietnamese Rhodophyta, analyzing the phylogenetic relationships among Gracilariaceae using *rbcL*, which revealed a new species of *Gracilariopsis*, *Gracilariopsis nhatrangensis* Le & Lin. Based on *rbcL* alone, Le et al. [49] later showed that *Gracilaria mammillaris* (Montagne) M.Howe had been misidentified as *Gracilaria phuquocensis* Le, Muangmai & Zuccarello, a new species found in Vietnam; Nguyen et al. [56] newly recorded the Halymeniales species *Phyllymenia taiwanensis* (Lin & Liang) Lin, Rodríguez-Prieto, De Clerck & Guiry in Da Nang from Central Vietnam; Nguyen et al. [55] recorded *Phyllymenia huangiae* (Lin & Liang) Lin, Rodríguez-Prieto, De Clerck & Guiry (Figure 2E); and Duy [47] reported the Rhodomelaceae species *Chondrophyucus tronoi* (Ganzon-Fortes) Nam from Vietnam. Analyses based on *cox1*, *psaA*, and *rbcL* sequences allowed the discovery of the Gelidiellaceae species *Perronella gracilis* Boo, Nguyen, Kim & Boo from Nha Trang Bay from Southern Vietnam [53], and the transfer of *Gelidiella adnata* Dawson to *Parviphycus adnatus* (Dawson) Santelices. Analyses based on the concatenated *rbcL* and *cox1* sequences also revealed a new record of Delesseriaceae from Vietnam, *Zellera tawallina* Martens (Figure 2A), previously identified as *Claudea batanensis* Tanaka [55]. Analyses combining *rbcL* and *cox1* sequences allowed the identification of a new species, *Meristotheca lysonensis* Nguyen, Nguyen, Kittle & McDermid, collected at Ly Son Island in the South Central Coast region of Vietnam [52] (Figure 2B). A last worthy account for Rhodophyta is that of the Halymeniaceae species *Halymenia dilatata* Zanardini, a common species in Vietnam, previously reported in several publications [14,16]. Based on phylogenetic analyses using concatenated chloroplast and mitochondrial and nuclear markers (*rbcL*, *cox1*, and LSU rDNA (28S)), Vy et al. [50] showed that *H. dilatata* may have been misidentified as *Halymenia malaysiana* Tan, Lim, Lin & Phang, a study that confirms new distributional records of *Phycocalidia tanegashimensis* along the Chinese and Vietnamese coastline in the South China Sea. The study used molecular sequence data from *rbcL*, COI-5P, and 18S rRNA genes to place *P. tanegashimensis* in a clade with *P. acanthophora*, *P. denticulata*, *P. suborbiculata*, and *P. vietnamensis* as out-groups [54].

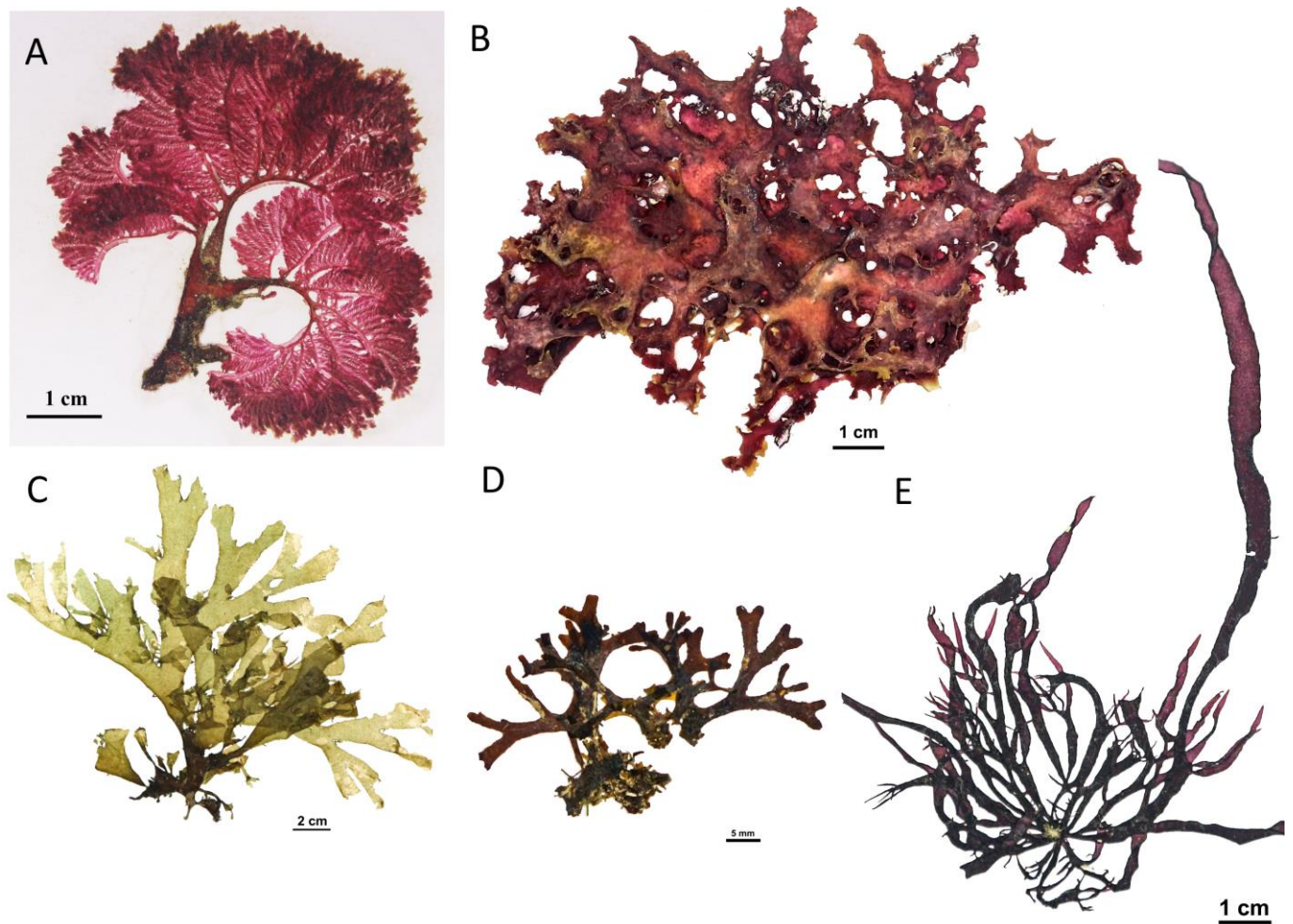


Figure 2. Some recently newly described species and new records made of marine macroalgae from Vietnam. (A) *Zelleria tawallina* Martens collected at Nha Trang Bay; (B) *Meristotheca lysonensis* Nguyen, Nguyen, Kittle & McDermid collected at Ly Son Island; (C) *Dictyota hauckiana* Nizamuddin collected at Ninh Thuan; (D) *Dictyota grossedentata* De Clerck & Coppejans collected at Nha Trang Bay; (E). *Phyllymenia huangiae* (Lin & Liang) Lin, Rodriguez-Prieto, De Clerck & Guiry collected at Da Nang.

3.2. Molecular-Assisted Alpha Taxonomy of Ochrophyta

Molecular-assisted alpha taxonomic studies on Ochrophyta have comprised a total of four molecular markers, analyzed individually or combined, consisting of two chloroplast genes (*rbcL* and the PSII thylakoid protein D1 (*psbA*)), one mitochondrial gene (cytochrome c oxidase subunit III (*cox3*)), and one nuclear encoded ribosomal cistron (ITS 2 rDNA). Tu [97] used ITS2 rDNA and *cox3* sequences to reassess *Sargassum* species diversity from Vietnam. The order Dictyotales has received particular attention in recent years. Using *rbcL* and *psbA* markers, Nguyen-Nhat et al. [58] newly identified *Dictyota hauckiana* Nizamuddin from Ninh Thuan (Figure 2C). One additional species of *Dictyota* was newly recorded from Vietnam, *Dictyota grossedentata* De Clerck & Coppejans [55] (Figure 2D). Molecular phylogenetic analyses based on concatenated *rbcL* and *cox3* sequences led to the description of the new species *Lobophora tsengii* Tien & Sun from Bach Long Vy [60], although morphological and molecular analyses did not conclusively rule out its conspecificity with *Lobophora rosacea* C.W.Vieira, Payri & De Clerck.

3.3. Molecular-Assisted Alpha Taxonomy of Chlorophyta

For Chlorophyta, the only molecular-assisted alpha taxonomic study reported until now is that by Tran et al. [61], who reassessed the species diversity in Vietnam of the Ulvaceae genus *Ulva* based on *rbcL* and the elongation factor Tu (*tufA*). The study revealed seven new records of *Ulva* from Vietnam and identified a new species, *U. vietnamensis* L-A. T. Tran, Leliaert & De Clerck.

3.4. Molecular-Assisted Alpha Taxonomy of Cyanobacteria and Alismatales

For Alismatales (seagrasses), the concatenated *rbcL* and *matK* were applied to assess the species diversity of *Halophila* [98]. Based on the genetic marker ITS, a later study by Nguyen et al. [44] showed that *Halophila major* was the correct name for the collections of *Halophila ovalis* from Nha Trang Bay. All seagrass species from Vietnam were confirmed with molecular markers, and samples previously labeled as “*Halophila johnsonii*” were reidentified as *H. ovalis*. Therefore, *Halophila johnsonii* was removed from the seagrass checklist of Vietnam [99]. *Halophila major* was found in most offshore islands, whereas *H. ovalis* occurred in lagoons in Vietnamese waters [100]. In contrast, no molecular-assisted alpha taxonomic study on Cyanophyceae was yet conducted in Vietnam.

3.5. Intraspecific Genetic Diversity Studies

Several DNA fingerprinting have been applied to investigate the genetic relationships among individuals within or among populations of the same species [101,102]. In a global study of *Gracilaria salicornia* (Agardh) Dawson from Southeast Asia, Yang et al. [103] distinguished a lineage of the Philippines from other Southeast Asian countries (e.g., Malaysia and Thailand). For another Rhodophyta, *Phycocalidia acanthophora* (Oliveira & Coll) Santiañez, the dataset of *rbcL* indicated that there is no haplotype sharing between populations in the Philippines and other nearby areas, including Taiwan, Japan, and Hong Kong [104]. In Vietnam, the red algae species in *Kappaphycus* and *Euचेuma* are important economically and were widely cultivated in the South Central. So far, based on a combined *cox2–3* and *rbcL* dataset, Zuccarelo et al. [105] compared the genetic variation among cultivated *Kappaphycus alvarezii* (Doty) Liao farming worldwide, including a strain from Vietnam; the authors indicated that there is no genetic variation among samples collected in Vietnam and other Southeast Asian countries, such as the Philippines, Malaysia, and Indonesia. However, *Kappaphycus alvarezii* collected from Africa and Hawai'i showed significant differences from populations in Southeast Asian countries. By using random amplified polymorphic DNA (RAPD) markers, Hong et al. [106] also revealed the genetic variation among strains of *Kappaphycus* spp. and *Euचेuma* spp. in Vietnam, *Kappaphycus striatus* (Schmitz) Liao. The analyses of the mitochondrial *cox2–3* spacer of *Kappaphycus* spp. and *Euचेuma* spp. showed that there are two haplotypes of *K. alvarezii*, and an unidentified *Kappaphycus* sp. was also found in Vietnam and the Philippines [107]. A later study by Tan et al. [108] indicated that the aring-aring strain was described as the new species *Kappaphycus malesianus* Tan, Lim & Phang. There is no evidence of occurrence of this species in Vietnamese waters. The biogeography of *Halymenia malaysiana* was studied in more detail. Our previous study showed that the common haplotype in Vietnam is R1, and three new haplotypes were added to *H. malaysiana* for Southeast Asia (Figure 3). There are statistically significant genetic differences between Sunda Shelf (Vietnam and Malaysia) populations and those in Philippine waters [50]. For another economic species, *Gracilaria tenuistipitata* Chang & Xia, Song et al. [109] found that there is only one haplotype (T5) in Vietnam. Compared with other haplotypes in Thailand, Malaysia, and Singapore, a haplotype of *Gracilaria tenuistipitata* collected in Vietnam showed from one to eight mutational steps. Recently, the *tufA* gene was applied to find the haplotype and genetic diversity of the green algae *Halimeda* spp.; Nguyen et al. [110] concluded that the genetic variation in *H. macroloba* Decaisne is very low, and *H. opuntia* (Linnaeus) J.V.Lamouroux tends to form a distinct group in Vietnamese waters.

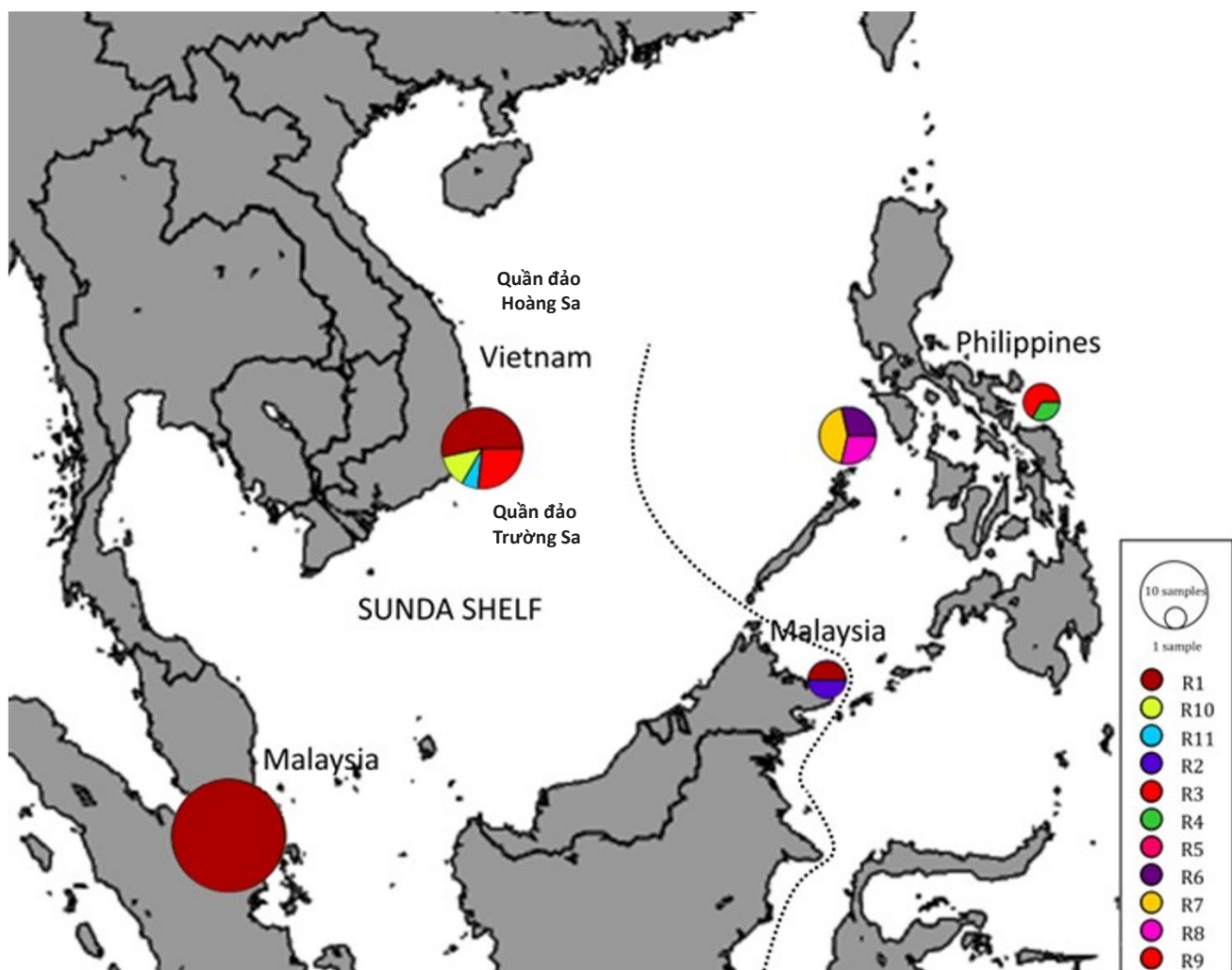


Figure 3. Distribution of haplotypes of *Halymenia malaysiana* in Sunda Shelf (Malaysia and Vietnam) and the Philippines [50]. Adapted from Nguyen et al. [50].

4. Biodiversity Distribution Patterns

4.1. Marine Floral Biodiversity across the South China Sea

Located on the northwest edge of the Coral Triangle biodiversity hotspot, the South China Sea is one of the most productive marine regions in the world [111,112]. The sea is bordered by twelve states and territories, including Brunei, Cambodia, (mainland) China, Hong Kong, Indonesia, Macao, Malaysia, the Philippines, Singapore, Taiwan, Thailand, and Vietnam. Phang et al. [63] documented 1412 species of marine algae from the South China Sea (119 Cyanophyceae, 305 Chlorophyta, 258 Ochrophyta, and 730 Rhodophyta) from six countries bordering the South China Sea (Indonesia, Malaysia, the Philippines, Singapore, Thailand, and Vietnam). Their analyses showed similarity in the marine algal floras of Malaysia, Singapore, and Thailand and those of Vietnam and the Philippines. We present here an overview of species diversity from states and territories bordering the South China Sea based on AlgaeBase [46] for four algae classes (Cyanobacteria, Rhodophyta, Ochrophyta, and Chlorophyta) and seagrasses. No data were available on AlgaeBase for Brunei, Hong Kong, Macau, Cambodia, and Taiwan. Data for seagrasses were retrieved from different sources indicated in Figure 4 and Table 3. The Gulfs of Thailand and Tonkin were included in the South China Sea. It should be noted that with the exception of Vietnam, the species numbers provided here are not restricted to the South China Sea, but are all-inclusive for each country (i.e., not restricted to the South China Sea), and retrieving data restricted to the South China Sea was not possible. In comparison with other South

China Sea bordering states/territories, Vietnam supports the fourth highest marine floral diversity with 877 species, according to AlgaeBase [46] (but 881 species according to our updated checklist). However, taking into account the all-inclusiveness of the number for other countries, Vietnam possibly holds the highest diversity in the South China Sea. In fact, the South China Sea coastlines of the three other species-rich states (China, the Philippines, Indonesia) represent only a fraction of these countries.

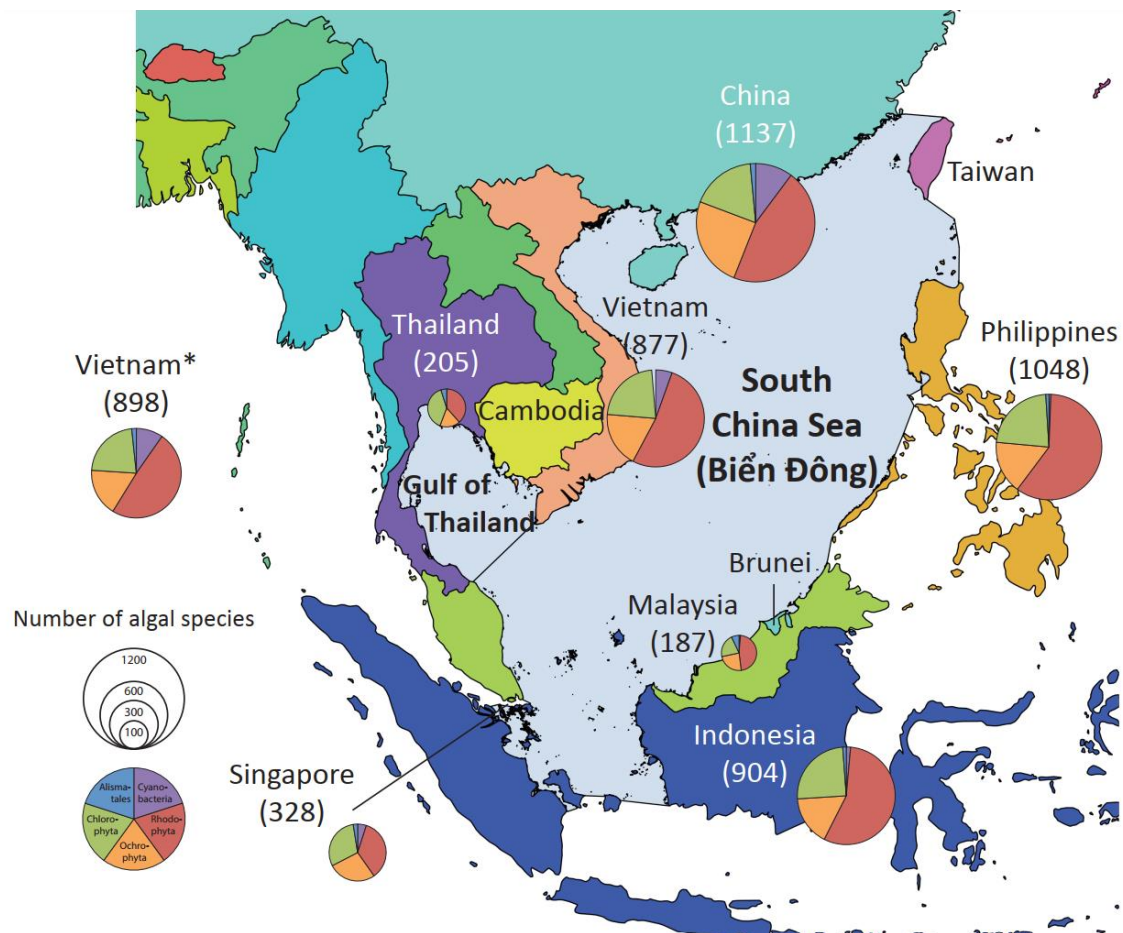


Figure 4. Spatial variation in marine floral biodiversity (Cyanobacteria, Rhodophyta, Ochrophyta, and Chlorophyta) across countries bordering the South China Sea (East Vietnam Sea). Marine floral biodiversity data based on AlgaeBase [46]. * Data for the Vietnam marine flora based on the current updated checklist. Colors mean blue is Alismatales, green is Chlorophyta, red is Rhodophyta, brown is Ochrophyta, and purple is Cyanobacteria.

Table 3. Species diversity of marine floral groups in countries bordering the South China Sea (East Vietnam Sea). Data based on AlgaeBase [46]. * References for seagrasses (Alismatales).

Countries	Cyanobacteria	Rhodophyta	Ochrophyta	Chlorophyta	Alima-Stales	Ref. *
China	114	495	301	205	22	[113]
Indonesia	11	483	165	232	13	[114]
Malaysia	2	85	45	39	16	[115]
Philippines	6	597	183	244	18	[116]
Singapore	15	114	92	97	10	[117]
Thailand	0	76	40	77	12	[118]
Vietnam	51	438	171	202	15	[119]

Biodiversity numbers should nevertheless be interpreted cautiously as they may under-represent the actual floral diversity of each country and the region, since they are for the most part established on morphological-based identification, and additionally, some countries have received much lesser attention than others (e.g., Brunei, Cambodia, and Malaysia). Notwithstanding, the high floral biodiversity in the South China Sea documented so far from Vietnam can be attributed to its geographical location, situated along the southeastern margin of the Indo-Chinese Peninsula, comprising the largest area of the peninsula and the longest coastline in the South China Sea.

We examined the similarity of the marine floras in seven of the states and territories bordering the South China Sea using a Bray–Curtis similarity index [120] multivariate analysis implemented in Primer V.6 software [121] based on compiled data for the region. Results showed that the Vietnamese marine flora was most similar to that of China, followed by those of Indonesia and the Philippines (Figure 5), and that the marine floras of Malaysia, Singapore, and Thailand were very similar (Figure 5), consistent with previous findings by Phang et al. [63].

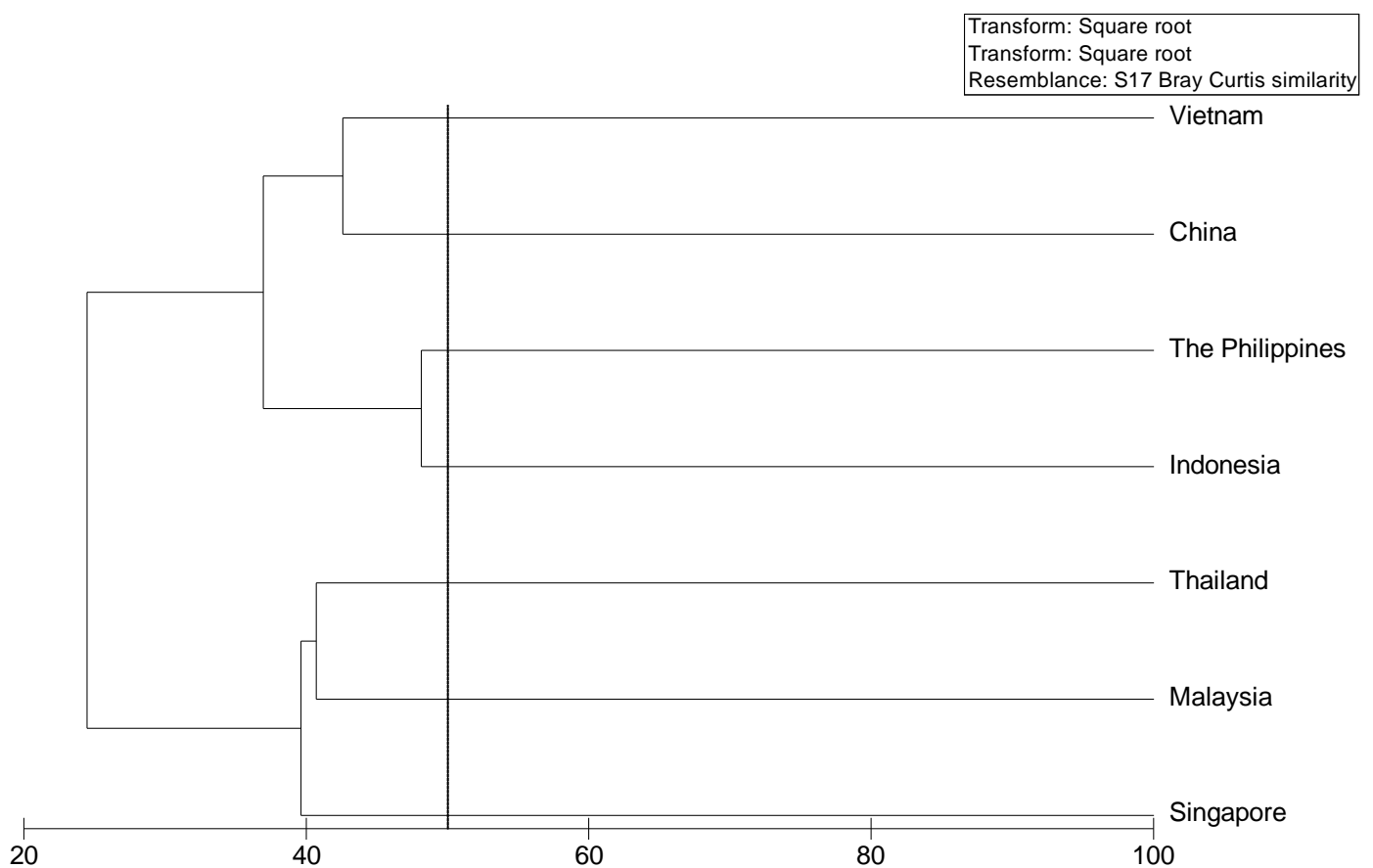


Figure 5. Cluster analyses of similarity (Bray–Curtis index) for the marine floras of the countries bordering the South China Sea. Data for similarity analyses range between 20 and 100. Hierarchical clustering is based on square-root-transformed presence data and on a resemblance matrix calculated using S17 Bray–Curtis similarity.

4.2. Marine Floral Biodiversity across Vietnam Regions

The Vietnamese coastline is divided into three “Geographical Regions” (Northern Vietnam, Central Vietnam, Southern Vietnam) subdivided into eight “Administrative Regions” (Northeast, Northwest, Red River Delta, North Central Coast, South Central Coast, Central Highlands, Southeast, Mekong River Delta) (Figure 6A). Based on our updated checklist, we show marine floral biodiversity across the “Administrative Regions”, excluding the Northwest and Central Highlands regions, which have no coast-

line. The geographical distribution of the marine floral biodiversity is uneven across Vietnamese regions. The South Central Coast holds the highest diversity by far, with a total of 587 species, followed by the Southeast (243), Red River Delta (210), North Central Coast (204), and Mekong River Delta (203) regions. The Northeast region is the least species region with 160 species. The high marine floral biodiversity documented in the South Central Coast coincides with the largest coral reef density along the coastline of Vietnam (Figure 6B) and a high diversity of marine environments [3]. It is worth mentioning that Spalding [122] proposed five marine ecoregions (Gulf of Thailand, Gulf of Tonkin, Southern Vietnam, Sunda Shelf/Java Sea, South China Sea (East Vietnam Sea) Ocean Islands) along the Vietnamese coastline (Figure 6A).

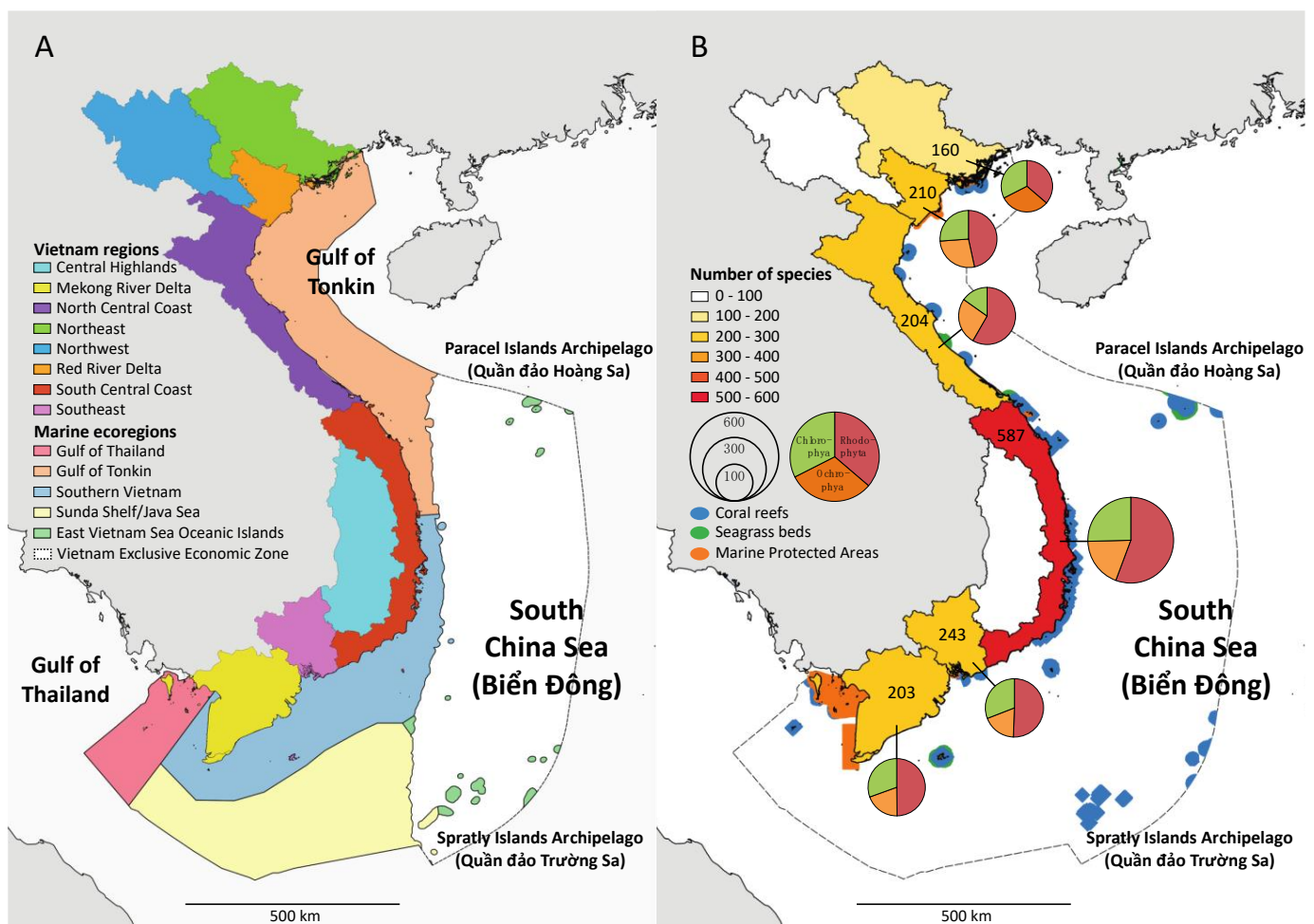


Figure 6. Spatial variation in marine floral biodiversity across Vietnamese regions. (A) Illustration of the Vietnamese administrative regions and the marine ecoregions sensu Spalding et al. [122] within the Vietnamese Exclusive Economic Zone (VEEZ). (B) Representation of species diversity across marine macroalgal classes (Cyanobacteria, Rhodophyta, Ochrophyta, and Chlorophyta) and depiction of coral reefs, seagrass beds, and marine protected area distributions within the VEEZ; numbers in each region represent the total number of species.

According to present data, with only 45 endemic species, Vietnam would seem to contain a very low level of endemism of marine flora (5.01%, 45 spp.; Table 4). However, this number most likely under-represents the actual level of endemism for the marine flora of Vietnam, since molecular-assisted alpha taxonomic efforts, needed to obtain accurate taxonomic data, have been very limited.

Table 4. Endemic marine algae species from Vietnam.

No.	Phylum	Genus	Name Species	
1	Rhodophyta		<i>Acrochaetium gracile</i> var. <i>vietnamense</i> Pham-Hoàng Hộ	
2		<i>Acrochaetium</i> Nägeli	<i>Acrochaetium phuquocense</i> Pham-Hoàng Hộ	
3			<i>Acrochaetium pseudoerectum</i> Pham-Hoàng Hộ	
4		<i>Ahnfeltiopsis</i> P.C.Silva & DeCew	<i>Ahnfeltiopsis quinhonensis</i> (Pham-Hoàng Hộ) Masuda	
5		<i>Bangia</i> Lyngbye	<i>Bangia tanakae</i> Pham-Hoàng Hộ	
6		<i>Caloglossa</i> (Harvey) G.Martens	<i>Caloglossa saigonensis</i> Tanaka & Pham-Hoàng Hộ	
7		<i>Ceramium</i> Roth	<i>Ceramium vietnamense</i> Pham-Hoàng Hộ	
8			<i>Ectocarpus siliculosus</i> f. <i>sporangiogramosus</i> A.D.Zinova & Nguyen Huu Dinh	
9		<i>Ectocarpus</i> Lyngbye	<i>Ectocarpus vungtauensis</i> Pham-Hoàng Hộ	
10		<i>Erythrotrichia</i> Areschoug	<i>Erythrotrichia parietalis</i> var. <i>majuscula</i> T.Tanaka & Pham-Hoàng Hộ	
11		<i>Gelidium</i> J.V.Lamouroux	<i>Gelidium vietnamense</i> Pham-Hoàng Hộ	
12			<i>Gracilariopsis nganii</i> Pham-Hoàng Hộ	
13		<i>Gracilariopsis</i> E.Y.Dawson	<i>Gracilariopsis nhatrangensis</i> Nhu Hau Le & S.-M.Lin	
14			<i>Gracilariopsis phanthietensis</i> Pham-Hoàng Hộ	
15		<i>Herposiphonia</i> Nägeli	<i>Herposiphonia vietnamica</i> Pham-Hoàng Hộ	
16		<i>Meristotheca</i> J.Agardh	<i>Meristotheca lysonensis</i> X.-V.Nguyen, X.-T.Nguyen, Kittle & McDermid	
17		<i>Porphyra</i> C.Agardh	<i>Porphyra tanakae</i> Pham-Hoàng Hộ	
18		<i>Prionitis</i> J.Agardh	<i>Prionitis vietnamensis</i> Pham-Hoàng Hộ	
19		<i>Pyropia</i> J.Agardh	<i>Pyropia vietnamensis</i> (Tak.Tanaka & P.H.Hồ) J.E.Sutherland & Monotilla	
20		<i>Rodriguezella</i> F.Schmitz	<i>Rodriguezella hongngai</i> Pham-Hoàng Hộ	
21		<i>Lobophora</i> J.Agardh	<i>Lobophora tsengii</i> D.Tien & Z.Sun	
22		<i>Petroderma</i> Kuckuck	<i>Petroderma vietnamensis</i> Pham-Hoàng Hộ	
23		Ochrophyta		<i>Sargassum bangmeianum</i> Nguyen Huu Dinh & Huynh Quang Nang
24				<i>Sargassum baoreni</i> Nguyen Huu Dinh & Huynh Quang Nang
25				<i>Sargassum buui</i> Nguyen Huu Dinh & Huynh Quang Nang
26				<i>Sargassum carpophyllum</i> var. <i>honomense</i> Nguyen Huu Dinh & Huynh Quang Nang
27				<i>Sargassum carpophyllum</i> var. <i>nhatrangense</i> (Pham-Hoàng Hộ) Ajisaka
28				<i>Sargassum congkinhii</i> Pham-Hoàng Hộ
29			<i>Sargassum</i> C.Agardh	<i>Sargassum cornutifractum</i> Nguyen Huu Dinh & Huynh Quang Nang
30				<i>Sargassum cotoense</i> Nguyen Huu Dai
31				<i>Sargassum hieui</i> Nguyen Huu Dinh & Huynh Quang Nang
32				<i>Sargassum mcclurei</i> f. <i>duplicatum</i> A.D.Zinova & Nguyen Huu Dinh
33				<i>Sargassum namoense</i> Nguyen Huu Dai
34				<i>Sargassum phamhoangii</i> Nguyen Huu Dai
35				<i>Sargassum quinhonense</i> Nguyen Huu Dai
36				<i>Sargassum serratum</i> Nguyen Huu Dai
37				<i>Sargassum tsengii</i> Nguyen Huu Dinh & Huynh Quang Nang
38				<i>Sargassum vietnamense</i> A.D.Zinova & Dinh
39			<i>Spatoglossum</i> Kützing	<i>Spatoglossum vietnamense</i> Pham-Hoàng Hộ
40			<i>Turbinaria</i> J.V.Lamouroux	<i>Turbinaria ornata</i> var. <i>prolifera</i> Pham-Hoàng Hộ

Table 4. Cont.

No.	Phylum	Genus	Name Species
41	Chlorophyta		<i>Caulerpa mexicana</i> f. <i>vietnamica</i> Pham-Hoàng Hộ
42		<i>Caulerpa</i> J.V.Lamouroux	<i>Caulerpa minuta</i> L.N.Hau
43			<i>Caulerpa racemosa</i> f. <i>vietnamensis</i> A.D. Zinova & Nguyen H. Dinh
44		<i>Cladophoropsis</i> Børgesen	<i>Cladophoropsis phamhoanghooi</i> Molinari & Guiry
45		<i>Ulva</i> Linnaeus	<i>Ulva vietnamensis</i> L.-A.T.Tran, Leliaert & De Clecrck

4.3. Seaweed Biodiversity Loss, Threats, and Conservation

Some species of marine algae in Vietnam have experienced declines in their populations due to a variety of factors. Several threats to marine algae exist in Vietnam, including but not restricted to pollution, climate change, overharvesting, invasive species, and habitat destruction. Marine algae are vulnerable to pollution from a variety of sources, including agricultural runoff, industrial discharge, and sewage. Pollution can harm marine algae directly and also make their habitat less suitable for growth [123]. Marine algae are sensitive to changes in temperature, salinity, and other environmental conditions, and they may be negatively impacted by climate change [124]. Some species of marine algae are in high demand for use in food, cosmetics, and other products, and overharvesting can lead to a decline in their populations [125]. Non-native species of marine algae that are introduced to new areas can outcompete native species and reduce their populations [126]. Marine algae rely on specific types of habitat for growth, and the destruction of these habitats can negatively impact their populations [127]. The biodiversity (marine and terrestrial) of Vietnam has decreased quickly [128]. Some of the known factors in Vietnam are land conversion without a proper scientific base, quick reduction of natural forests, infrastructure developments (e.g., dams, roads, and new urban and rural human settlements), and overexploitation of natural resource/illegal exploitations in fishing, hunting, forestry [2,129,130]. It is difficult to quantify the extent of marine algal diversity loss in Vietnam, as there are limited data available on this topic. Titlyanov et al. [38] quantified seaweed community changes in Nha Trang Bay and investigated the factors associated with these changes. Collections sampled between 1953 and 1968 and 1982 and 1987 did not change significantly in either the species diversity nor the floristic composition. However, the species composition assessed between 2002 and 2010 showed changes in the species diversity composition, with an increase in Chlorophyta and a reduction of Rhodophyta and Ochrophyta species. In Con Dao Island, significant changes in marine floral species composition were observed between 1998 and 2008, with a proportional species replacement in each taxonomic group over the last two decades [40]. Since the 1970s, several species have not been observed, such as *Erythrocladia irregularis* Rosenvinge, *Acrochaetium crassipes* (Børgesen) Børgesen, *Metagoniolithon stelliferum* (Lamarck) Ducker, and *Exophyllum wentii* Weber Bosse [41]. Similarly, the species diversity of *Sargassum* was previously well studied at Nha Trang Bay, with the identification of 21 species between 1950 and 1970. Between 1980 and 2000, 9 of the previously identified species were not recorded, while an additional 15 species were newly added to *Sargassum*. However, based on the most recent collection, in 2020 in Nha Trang Bay, *Sargassum* was represented by 14 species, including 7 species found in the previous two surveys and 7 new additions. Overall, 24% (149 species) of algal species in South Vietnam recorded between 1980 and 2000 could not be found between 2000 and 2020 [41]. A report by Vy et al. [131] indicated that nearly 50% of the *Sargassum* beds at Hon Chong (Khanh Hoa Province) have disappeared because of loss of substratum, and the species *Sargassum crassifolium*, once a dominant species in this site, disappeared. In Nha Trang Bay, seawater pollution resulting from dissolved organic and inorganic compounds of nitrogen and phosphorus may lead to an increasing larger number of green algae and their biomass as well as population density. The green algae may displace fleshy and foliose forms of red and brown macroalgae from communities [38].

Another threat to seaweed biodiversity is harvesting of natural stocks. Local harvesters collect large quantities of *Sargassum* for production of alginates, Asian herbal medicine, and various human foods [132]. Young populations of *Sargassum* are commonly harvested prior to reaching sexual maturity and reproduction, thus affecting natural stock renewal (authors' pers. obs.). Another case of overexploitation of natural stocks was reported in the edible red seaweed *Betaphycus gelatinus*, now very rare due to harvesting by locals at Ninh Thuan Province. The Vietnam Red data Book [133] shows 8 and 5 species of Rhodophyta and Ochrophyta, respectively. Among them, *Crytonemia undulata* is in the critically endangered category. Six species including 5 Rhodophyta and 3 Ochrophyta are in the endangered category. The 6 remaining species are in the vulnerable category. There are 12 marine protected areas (MPAs) from 10 provinces/cities in Vietnam. Large seaweed beds in Khanh Hoa, Ninh Thuan, are out of the core zone of MPAs and therefore under threat. The natural stock of *Hydropuntia eucheumatoides*, *Betaphycus gelatinus*, and *Sargassum* spp. (endangered category) is still collected by local people due to lack of Red Data Book. Like seagrasses, the management models of marine macroalgal ecosystems in Vietnam are mostly integrated into coastal management models to solve the problems of weaknesses that exist in the management, exploitation, and use of natural resources and environmental protection in coastal areas.

5. Conservation Efforts

Conservation efforts are needed to protect and conserve marine floral diversity in Vietnam. It is important to address these threats in order to maintain the health and resilience of Vietnam marine environments and the economic and cultural value of these resources. This may involve measures such as habitat conservation, sustainable harvesting, pollution reduction, and invasive species management. There are several conservation efforts underway in Vietnam to protect the marine flora and the marine environments they are a part of. Some of these efforts are reviewed below.

5.1. Habitat Conservation

Many conservation efforts in Vietnam focus on protecting and preserving the habitats that support marine flora. This may involve establishing marine protected areas (MPAs) or other types of conservation zones, which are designated areas of the ocean that are set aside for the protection and conservation of marine life. Recognizing the importance of marine protected areas in the protection of marine biodiversity, the prime minister released Decision No. 742/QĐ-TTg on 26 May 2010, authorizing the preparation for the marine biodiversity scheme. Marine conservation in Vietnam will continue until 2020 (this deadline has been extended), with the aim of preserving habitats and marine species of economic and scientific importance. It aimed to contribute to the development of marine economy and improve the livelihoods of fishermen communities in coastal localities. The Fisheries Law of 2017 was passed by the 14th National Assembly, which includes provisions for the protection and growth of aquatic resources, including marine conservation, in the sense of sustainable fisheries development and international integration. The Communist Party of Vietnam's Central Committee released Resolution No. 36-NQ/TW on the Strategy for Sustainable Development of Vietnam's Marine Economy to 2030 with a Vision to 2045 on 22 October 2018. The document stated on the matter: "Sustainable development of the marine economy on the basis of green growth, biodiversity protection, and marine environment conservation; ensure harmony between economic and natural environments, conservation and development, promoting the sea's potentials and advantages, and creating a driving force for national economic development"; and that the specific target was to "Well maintain and protect aquatic, coastal, and island ecosystems; raise the area of marine and coastal protected areas to at least 6% of the national marine area". Currently, the Ministry of Agriculture and Rural Development and other cities have created and operationalized 12 marine protected areas (Figure 4B, Table 5). These 12 MPAs amount to a total of 243,023 ha (ca. 2430 km²), which corresponds to ca. 0.17% of the total surface (ca.

1,395,096 km²) of the Vietnamese Marine Exclusive Economic Zone. Among the 12 MPAs, 3 typically contain seaweeds and 9 seagrasses.

Table 5. Checklist MPAs in Vietnam.

No.	MPA Name	Province	Regions	Area (ha)	Typical Species
1	Co To-Dao Tran	Quang Ninh	Southeast	18,400	Coral, seaweed, seagrass
2	Bai Tu Long	Quang Ninh	Southeast	96,500	Coral, seagrass
3	Cat Ba	Hai Phong	Red River Delta	10,900	Coral, seaweed, seagrass
4	Bach Long Vi	Hai Phong	Red River Delta	27,000	Coral, abalone
5	Con Co	Quang Tri	North Central Coast	2140	Red coral, seagrass
6	Cu Lao Cham	Quang Nam	South Central Coast	6716	Coral, seaweed, seagrass
7	Ly Son	Quang Ngai	South Central Coast	7925	Coral, seagrass
8	Nha Trang Bay	Khanh Hoa	South Central Coast	12,000	Coral, seagrass
9	Nui Chua	Ninh Thuan	South Central Coast	7352	Coral, sea turtle
10	Hon Cau	Binh Thuan	South Central Coast	12,390	Coral, seagrass, rock crab, shrimp
11	Con Dao	Ba Ria–Vung Tau	South Central Coast	23,000	Dugong, sea turtle, coral
12	Phu Quoc	Kien Giang	Mekong River Delta	18,700	Coral, seagrass
Total area (ha)				243,023	-

Moreover, there are other nature reserves along the coast of Vietnam [134] (Table 6). The Ministry of Agriculture and Rural Development has developed comprehensive plans for the establishment of four MPAs, which have been submitted to provincial people's committees for approval: Hon Me/Thanh Hoa, Nam Yet/Khanh Hoa, Phu Quy/Binh Thuan, and Hai Van–Son Cha/Da Nang–Hue.

Table 6. Nature reserves and national parks in Vietnam.

Nature Reserve	Province	Regions	Area (ha)
Dong Rui Wetland Conservation Area	Quang Ninh	Southeast	15,750
Cat Ba Biosphere Reserve	Hai Phong	Red River Delta	26,588
Thai Thuy Wetland Conservation Area	Thai Binh	Red River Delta	6560
Red River Delta Biosphere Reserve	Thai Binh–Nam Dinh–Ninh Binh	Red River Delta	105,558
Tam Giang–Cau Hai Wetland Nature Reserve	Thua Thien Hue	North Central Coast	2071
Son Tra Nature Reserve	Da Nang	North Central Coast	4400
Nui Chua National Park	Ninh Thuan	South Central Coast	29,865
Binh Chau–Phuoc Buu Nature Reserve	Ba Ria–Vung Tau	Southeast Coast	10,537
Can Gio Biosphere Reserve	Ho Chi Minh	Southeast Coast	75,740
Mui Ca Mau National Park	Ca Mau	Mekong River Delta	41,862

Despite the fact that the Ministry of Agriculture and Rural Development has organized a mission to inspect, guide, and have several documents to direct and inform, the provincial people's committees have not yet approved the establishment after more than 5 years of handover of Vietnam's coastal area has high biodiversity: 13 out of 28 national parks, 22 out of 55 nature reserves, and 17 out of 34 forests of cultural, historical, and environmental significance are located in coastal areas and islands.

5.2. Sustainable Harvesting

Some species of seaweed in Vietnam are harvested for use in food, cosmetics, and other products. In order to ensure the sustainable use of these resources, there are efforts to establish sustainable harvesting practices and to manage fisheries to ensure that seaweed populations are not overharvested. Vietnam adopted international standards of the sanitary and phytosanitary (SPS) agreement-based regulation, which includes seaweeds. This established regulation covers a wide range of standards, including ensuring that the seaweeds are disease-, pathogen-, toxin-free, and furthermore that seaweeds meet permissible levels for heavy metals and other contaminants (e.g., pesticides). Vietnam has national regulations for controlling the movement of aquatic aquaculture organisms (quarantine), which also includes the import of live seaweed. For example, Vietnam provided a technical guideline for importing live seaweed, e.g., *Gracilaria* species [135], and technical requirements for *Kappaphycus alvarezii* (Table 7).

Table 7. Biosecurity components adopted in the national seaweed policies and regulations in the main *Gracilaria* and *Kappaphycus alvarezii* production.

Biosecurity Component	Policy/Regulation	Description
Established aquatic animal/plant regulation	Regulation No. 434/2021	National plan for prevention and control of some dangerous diseases on aquaculture
Disease surveillance system	Plant Protection and Quarantine Law No. 41/2013	Law on activities of preventing and combating organisms harmful to plants, phytosanitary, management of pesticides
National regulation of live seaweed movement	Fisheries Law No. 18/2017	Law on fisheries activities, state management of fisheries
National statutory contingency plan	Regulation No. 1664/2021	Law on fisheries activities, state management of fisheries
National standards of seaweed cultivation, production process, food safety	- TCN 108:1998; - TCN 155:2000; - TCVN 10371:2014	- Ministry standard for <i>Gracilaria asiatica</i> planting material; - Ministry standard for technical process of cultivation of <i>Gracilaria asiatica</i> to produce 2 tons dry seaweed/ha/year; - Technical requirements for <i>Kappaphycus alvarezii</i> .

5.3. Pollution Reduction

Seaweeds are vulnerable to pollution from a variety of sources, and efforts are being made to reduce pollution in Vietnam's coastal waters in order to protect these ecosystems. This may involve measures such as improving wastewater treatment, regulating industrial discharge, and reducing agricultural runoff. However, no particular reports and regulations were found on pollution reduction in Vietnam's coastal waters.

5.4. Invasive Species Management

Non-native species of seaweed that are introduced to new areas can outcompete native species and reduce their populations. To address this threat, efforts are being made to control the spread of invasive seaweed species in Vietnam. Circular No. 35/2018/TT-BTNMT dated 28 December 2018 of MONRE stipulates the criteria for the identification and promulgation of a list of invasive alien species. However, the subject only focuses on species that have been announced under the guidance of Circular 35, and the assessed ecosystems are only terrestrial and aquatic. There is almost no information about groups of marine organisms, including seaweed and seagrasses, more specifically, foreign species that are invasive in the sea and by shipping route; there has not been a specific study in Vietnam. In order to prevent the entry of alien organisms in the ballast water environment transported by ships from other sea areas, affecting the ecosystem, economy, and human health and strengthening measures to protect the marine environment, IMO ratified the

BWM Convention on 13 February 2004, and the convention met the conditions to enter into force on 8 September 2017. By 8 September 2024, all ships are required to use a ballast water management system (D2). Vietnam is in the process of completing the procedures to join the convention. The basic legal documents related to the activities of dumping garbage and discharging wastewater and ballast water are specified in Article 117 of Decree 58/2017/ND-CP guiding the Vietnam Maritime Code on the management of cargo operations [136].

6. Conclusions: Challenges and Future Directions

Studies conducted in the last decade effectively illustrated the need to combine molecular tools with morphological observations (i.e., habit view, vegetative and reproductive morphology) in (1) the reassessment of marine floral species diversity, (2) previous species names' validation, (3) misidentification detection, and (4) new species discovery. However, the fairly poor number (15 taxa; 25.4% of the new records and species) of new records and species made over the last 10 years in a “mega-diverse” country raised worrying concerns on the efforts put into the study of marine floral biodiversity. Past molecular-assisted taxonomic efforts have been focused on a limited number of taxa and localities. Currently, three main institutions, including (1) the Institute of Marine Environment and Resources in the North, (2) the Institute of Oceanography in the Central, and (3) the Institute of Tropical Biology in the South, are conducting most studies on marine algae taxonomy nationally. Considering the important length of the Vietnamese coastline (>3200 km), an exhaustive exploration of the Vietnamese marine flora represents a Herculean task for these institutions alone. In addition, the limited number of algal taxonomists in Vietnam and limited funding availability represent a major challenge to the study of marine floral biodiversity. Methodical molecular-assisted re-examination of Vietnam marine floral biodiversity is urgently needed in order to get an accurate picture of biodiversity and endemism, and thereby obtain baseline data for the marine floral management and protection. In particular, future efforts will need to be directed towards specific taxa and regions of Vietnam. Data provided in this review on species diversity, groups targeted with molecular-assisted alpha taxonomic approaches, and spatial variation in biodiversity offer valuable data to orientate future efforts. Finally, a more in-depth investigation of the threats targeting the marine flora of Vietnam is needed, and urgent implementation of measures for its conservation is called for, in particular, the increase in marine protected areas across Vietnam, which represent now less than 1% of the Vietnamese Marine Exclusive Economic Zone.

Author Contributions: Conceptualization, C.V., X.-V.N. and M.-L.N.; methodology, M.-L.N., C.V. and X.-V.N.; software, M.-L.N. and C.V.; validation, C.V., M.-L.N. and X.-V.N.; formal analysis, M.-L.N. and C.V.; investigation, M.-L.N., X.-V.N. and C.V.; resources, M.-L.N.; data curation, M.-L.N., M.-S.K., N.-T.N.N., X.-T.N., V.-L.C., X.-V.N. and C.V.; writing—original draft preparation, C.V., M.-L.N. and X.-V.N.; writing—review and editing, C.V., X.-V.N. and M.-L.N.; visualization, C.V., X.-V.N. and M.-L.N.; supervision, C.V. and X.-V.N.; project administration, C.V., X.-V.N. and M.-S.K.; funding acquisition, V.-L.C., X.-V.N., C.V. and M.-S.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Vietnam Academy of Science and Technology, grant number VAST06.01/22-23; TĐĐTĐ0.04/21-23. This research was supported by the Basic Science Research Program (2019R1A6A1A10072987) through the National Research Foundation of Korea (NRF) funded by the Ministry of Education of Korea.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: This article is part of the results of a PhD program at the Graduate University of Science and Technology so we are deeply indebted to Graduate University of Science and Technology, Vietnam Academy of Science and Technology (VAST), Ha Noi, Vietnam; and all staff at the Institute of Marine Environment and Resources, VAST, Hai Phong, Vietnam. We thank project ĐL000.01/23-24; QTRU04.03/18-19; QTRU02.08/21-22 of Vietnam Academy of Science and Technology for sharing information; Vingroup Innovation Foundation—VinIF grant number VINIF.2022.DA00079 and JSPS CREPSUM collaborative research and education project in Southeast Asia for sustainable use of marine ecosystems. This paper is a contribution to celebrate the 65th anniversary of the Institute of Marine Environment and Resources, Hai Phong, VAST.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Hoeksema, B.W. Delineation of the Indo-Malayan Centre of Maximum Marine Biodiversity: The Coral Triangle. In *Biogeography, Time, and Place: Distributions, Barriers, and Islands*; Renema, W., Ed.; Springer: Dordrecht, The Netherlands, 2007; Volume 29, pp. 117–178. ISBN 978-1-4020-6374-9.
2. Do Cong, T. Research Results of Marine Biodiversity and Biological Resources in Vietnam: Current Status, Threats, Proposed Solutions for Sustainable Use. *Vietnam J. Mar. Sci. Technol.* **2022**, *22*, 1–20. [\[CrossRef\]](#)
3. Nhung, N.; Armstrong, C.W.; Kim Anh, N.T.; Ngoc, Q.T.K.; Hai Anh, N. Incorporating Fisheries Management into Biodiversity Conservation Policies to Enhance Effectiveness of MPAs: A Case Study in Cu Lao Cham MPA, Vietnam. *Fish People* **2011**, *9*, 39–49.
4. Groombridge, B.; Jenkins, M.D. *World Atlas of Biodiversity Earth's Living Resources*; University of California Press: Berkeley, CA, USA, 2002; ISBN 0520236688.
5. Larsen, P.B. Linking Livelihoods and Protected Area Conservation in Vietnam: Phong Nha Kẻ Bàng World Heritage, Local Futures? In *People, Protected Areas, and Global Change: Participatory Conservation in Latin America, Africa, Asia and Europe*; Galvin, M., Haller, T., Larsen, P.B., Eds.; NCCR North-South: Bern, Switzerland, 2008; Volume 3, pp. 431–470. ISBN 978-3-905835-06-9.
6. Ziegler, T.; Vu, T.N. Ten Years of Herpetodiversity Research in Phong Nha—Ke Bang National Park, Central Vietnam. In *Phong Nha—Ke Bang National Park and Cologne Zoo, 10 Years of Cooperation (1999–2009)*; Phong Nha—Ke Bang National Park and Cologne Zoo: Laos, Vietnam, 2009; pp. 103–124.
7. Hung, N.Q.; Chieu, H.D.; Dung, D.T.; Son, L.T.; Duc, V.T.; Duy, D.A. Climate Change Impacts on Marine Ecosystems in Vietnam. In *Environmental Management of Marine Ecosystems*; CRC Press: Boca Raton, FL, USA, 2018; pp. 209–235. ISBN 9781315153933.
8. Chavez, F.P.; Messié, M.; Pennington, J.T. Marine Primary Production in Relation to Climate Variability and Change. *Annu. Rev. Mar. Sci.* **2011**, *3*, 227–260. [\[CrossRef\]](#)
9. de Loureiro, J. *Flora Cochinchinensi: Sistens Plantas in Regno Cochinchina Nascentes: Quibus Accedunt Aliae Observatae in Sinensi Imperio, Africa Orientali, Indiaeque Locis Variis: Omnes Dispositae Secundum Systema Sexuale Linnaeanum*; Typis, et expensis Academicis: Ulyssipone; 1790; Volume 1. [\[CrossRef\]](#)
10. Georg, A.J. *Species, Genera et Ordines Algarum: Seu Descriptiones Succinctae Specierum, Generum et Rodinum, Quibus Algarum Regnum Constituitur, Lundae, apud C.W.K. Gleerum, 1848–1901*; BHL: Washington, DC, USA, 1851; Volume 2.
11. S., F.A.; Agardh, J.G. Species Genera et Ordines Algarum. *Taxon* **1978**, *27*, 101. [\[CrossRef\]](#)
12. Toni, G.B.d.; De' Toni, E.; Boston Society of Natural History. *Sylloge Algarum Omnium Hucusque Cognitarum*; Patavii, Sumptibus auctoris, 1889–1924; 1895; Volume 1. [\[CrossRef\]](#)
13. De Toni, G.B. *Sylloge Algarum Omnium Hucusque Cognitarum. Volume IV. Florideae. Sectio IV. Pdf. Patavii* **1905**, *4*, 476.
14. Ho, P.H. *Vietnam Seaweed—Southern Part*; Ministry of Education and Youth, Saigon Learning Resource Center: Ho Chi Minh City, Vietnam, 1969.
15. Peñailillo Núñez, T.C.; Medina Valmaseda, A.A.; Pomponi, S.A.; Rodríguez, S.; Ross, N.A.; Aagaard, J.; Aagaard, J.; Aahangarzadeh, M.; Aahmadi, B.; Aakbarinasab, M.; et al. *Campagnes Du «De Lanessan» (1925–1929): Liste Des Stations*; Gouvernement Général de L'Indochine: New Delhi, India, 1936; Volume 17.
16. Dawson, E.Y. Marine Plants in the Vicinity of the Institute Oceanographique de Nhatrang, Vietnam. *Pac. Sc.* **1954**, *8*, 373–481.
17. Dinh, N.H.; Nang, H.Q.; But, T.N.; Van Tien, N. *Vietnam Seaweed—Northern Part*; Hanoi Science and Technology Publishing: Ha Noi, Vietnam, 1993.
18. Dai, N.H. *Sargassum (Sargassacea) Vietnam*; Agriculture Publishing House: Ho Chi Minh City, Vietnam, 1997.
19. Nang, H.Q. *Current Status and Economic Resources of Seaweed in the South Coast of Vietnam*; ELMAR: Cam Giang District, Vietnam, 1999.
20. Van Tu, N.; Boo, S.M. Distribution Patterns and Biogeography of *Sargassum* (Fucales, Phaeophyceae) along the Coast of Vietnam. *Bot. Mar.* **2020**, *63*, 463–468. [\[CrossRef\]](#)
21. Kawaguchi, S.; Dinh, N.H. Taxonomic Studies of the Halymeniaceae (Halymeniales, Rhodophyta) from Vietnam, I. A Transfer of *Carpopeltis Formosana* Okamura to *Prionitis*. *Bot. Mar.* **1998**, *41*, 391–397. [\[CrossRef\]](#)
22. Kawaguchi, S. Morphology of *Halymenia Maculata* J. Agardh from Vietnam. Section V. *Halymenia* Species. In *Taxonomy of Economic Seaweeds with Reference to Some Pacific Species*; University of California: California, CA, USA, 2002; pp. 259–266.

23. Dinh, N.H. Vietnamese Species of *Gracilaria* and *Gracilariopsis*. In *Species, Taxonomy of Economic Seaweeds with Reference to Some Pacific Species*; Abbott, I.A., Ed.; California Sea Grant College Program: California, CA, USA, 1992; pp. 207–210.
24. Hau, L.N.; Lin, S. *Gracilariopsis nhatrangensis* (Gracilariaceae, Rhodophyta), a New Marine Red Alga from Nhatrang, Southern Vietnam. *Bot. Stud.* **2006**, *47*, 329–337.
25. Dai, N.H.; Tri, P.H. Some New Records of Marine Algae From Vietnam. *Collect. Mar. Res. Work.* **2002**, Volume XII, 149–158.
26. Dinh, N.H.; Nang, H.Q. Species of *Eucheuma* and *Kappaphycus* in Vietnam. In *Proceedings of the Taxonomy of Economic Seaweeds with Reference to Some Pacific Species*; Abbott, I.A., Ed.; California Sea Grant College Program: California, CA, USA, 1995; pp. 43–51.
27. Abbott, I.A.; Fisher, J.; McDermid, K.J. Newly Reported and Revised Marine Algae from the Vicinity of Nha Trang, Vietnam. *Taxon. Econ. Seaweeds Ref. Some Pac. Species* **2002**, *8*, 291–321.
28. Masuda, M.; Kawaguchi, S.; Takahashi, Y.; Matsuo, Y.; Susuki, M. A Taxonomic Study of the Genus *Laurencia* (Ceramiales, Rhodophyta) from Vietnam. I. *Laurencia Caduciramulosa* Masunda et Kawaguchi, Sp. Nov. *Cryptogam. Algol.* **1997**, *18*, 1–10.
29. Masuda, M.; Takahashi, Y.; Matsuo, Y.; Suzuki, M. A Taxonomic Study of the Genus *Laurencia* (Ceramiales, Rhodophyta) from Vietnam. II. *Laurencia lageniformis* Sp. Nov. *Cryptogam. Algol.* **1997**, *18*, 163–174.
30. Dam, D.T.; Nguyen, V.T. Species Composition and Distribution of Seaweed in Truong Sa Island-Spratly Islands. *Collect. Mar. Res. Work.* **1996**, *3*, 243–252.
31. Tri, P.H. Contributing to the Study of Seaweed in Truong Sa Archipelago. *Collect. Mar. Res. Work.* **1996**, 147–162.
32. Tri, P.H. Marine Macroalgae Resources at Son Ca and Song Tu Tay Island—Spratly Islands. *Collect. Mar. Res. Work.* **1999**, 205–215.
33. Hau, L.N. Notes on Some New Records of Marine Algae from Truong Sa Islands (Vietnam). *Collect. Mar. Res. Work.* **2001**, *11*, 115–120.
34. Tien, D.D.; Cuong, D.H. Species Composition and Distribution of Seaweeds From Some Small Islands (Nam Yet, Son Ca, Song Tu Tay, Sinh Ton) of Truong Sa Archipelago. *J. Mar. Sci. Technol.* **2016**, *16*, 297–305. [[CrossRef](#)]
35. Dai, N.H.; Tri, P.H. Seaweed Resources of Ly Son Island. *Collect. Mar. Res. Work.* **2001**, *11*, 121–134.
36. Dai, N.H.; Tri, P.H.; Vy, N.X. Species Composition and Distribution of Seaweed, Segrass in Phu Quy Island (Cu Lao Thu), Binh Thuan Province. *Collect. Mar. Res. Work.* **2009**, 225–243.
37. Tien, D.D. Seaweed Resources of Hai Van—Son Cha. In *Proceedings of the Scientific, Technological and Economic Conference Serving the Cause of Industrialization and Modernization of the Country*; Vietnam Union of Science and Technology Associations: Ha Noi, Vietnam, 2006; pp. 164–169.
38. Titlyanov, E.A.; Titlyanova, T.V.; Belous, O.S. Checklist of the Marine Flora of Nha Trang Bay (Vietnam, South China Sea) and Decadal Changes in the Species Diversity Composition between 1953 and 2010. *Bot. Mar.* **2015**, *58*, 367–377. [[CrossRef](#)]
39. Hau, L.N. Notes on Some New Species of Marine Algae from Ninh Thuan Province (South Vietnam). *Collect. Mar. Res. Work.* **2000**, Volume X, 141–148.
40. Titlyanov, E.A.; Titlyanova, T.V.; Belous, O.S.; Pham, T.H.; Dam, D.T. An Inventory and Decadal Changes of the Benthic Marine Flora on the Con Dao Islands, South China Sea, Vietnam. *Russ. J. Mar. Biol.* **2020**, *46*, 166–180. [[CrossRef](#)]
41. Belous, O.S.; Titlyanov, E.A.; Titlyanova, T.V. *Decadal Comparison (1950–2020) of Benthic Marine Flora from Central and Southern Vietnam*; 2021; Volume 521, ISBN ISBN 0000000329. [[CrossRef](#)]
42. Van Nguyen, T.; Le, N.H.; Lin, S.M.; Steen, F.; De Clerck, O. Checklist of the Marine Macroalgae of Vietnam. *Bot. Mar.* **2013**, *56*, 207–227. [[CrossRef](#)]
43. Tien, N.V.; Thanh, D.N.; Dai, N.H. *Seagrasses in Viet Nam*; Ha Noi Science and Technology Publishing House: Ha Noi, Vietnam, 2002.
44. Vy, N.X.; Holzmeyer, L.; Papenbrock, J. New Record of the Seagrass Species *Halophila major* (Zoll.) Miquel in Vietnam: Evidence from Leaf Morphology and ITS Analysis. *Bot. Mar.* **2013**, *56*, 313–321. [[CrossRef](#)]
45. Nguyen, X.-V.; Lau, V.-K.; Nguyen-Nhat, N.-T.; Nguyen, T.-H.; Phan, K.-H.; Dao, V.-H.; Ho-Dinh, D.; Hayashizaki, K.; Fortes, M.D.; Papenbrock, J. Update of Seagrass Cover and Species Diversity in Southern Viet Nam Using Remote Sensing Data and Molecular Analyses. *Reg. Stud. Mar. Sci.* **2021**, *44*, 101803. [[CrossRef](#)]
46. Guiry, M.D.; Guiry, G. M AlgaeBase. World-Wide Electronic Publication. National University of Ireland, Galway. Available online: <http://www.algaebase.org> (accessed on 6 January 2023).
47. Duy, D.A. Research on Seaweed Resources and Biodiversity in Nam Du Archipelago, Vietnam. Ph.D. Thesis, Research Institute for Marine Fisheries, 2022.
48. Hau, L.N.; Ly, B.M.; Huynh, T.V.; Trung, V.T. Some New Records of Marine Algae From Vietnam. *J. Mar. Sci. Technol.* **2015**, *15*, 35–44. [[CrossRef](#)]
49. Le, H.N.; Muangmai, N.; Kheauthong, S.; Sun, Z.; Zuccarello, G.C. *Gracilaria phuquocensis* Sp. Nov., a New Flattened *Gracilaria* Species (Gracilariales, Rhodophyta), Previously Recognized as *G. mammillaris*, from the Southern Coast of Vietnam. *Phycol. Res.* **2020**, *68*, 50–56. [[CrossRef](#)]
50. Vy, N.X.; Thuy, N.N.N.; Thuy, N.X.; Hieu, N.T.; Thanh, T.S.H.; Ha, D.V.; Duy, D.A.; McDermid, K.J. New Record of *Halymenia malaysiana* (Halymeniaceae, Rhodophyta) from Viet Nam, and Its Genetic Diversity in the Western Pacific. *Bot. Mar.* **2023**, *66*, 113–123. [[CrossRef](#)]
51. Binh, D.T.; An, K.T.; Cam, V.H.; Tuan, T.V. New Records and Evolutionary Relationship of Epiphyte (*Melanothamnus thailandicus*) on Red Algae (*Kappaphycus alvarezii*) in Khanh Hoa. *J. Fish Sci. Technol.* **2020**, *2*, 2–10.

52. Vy, N.X.; Thuy, N.X.; Ronald, P.K.I.; McDermid, K.J. *Meristotheca lysonensis* sp. nov. (Solieriaceae, Rhodophyta), a New Flattened Species from Vietnamese Waters. *Phytotaxa* **2022**, *574*, 137–148. [[CrossRef](#)]
53. Boo, G.H.; Van Nguyen, T.; Kim, J.Y.; Gall, L.L.; Rico, J.M.; Bottalico, A.; Boo, S.M. A Revised Classification of the Gelidiellaceae (Rhodophyta) with Descriptions of Three New Genera: *Huismaniella*, *Millerella* and *Perronella*. *Taxon* **2016**, *65*, 965–979. [[CrossRef](#)]
54. Zhao, W.; Dong, L.; Hong, D.D.; Brodie, J.; Chen, W.-Z.; Tien, D.D.; Zhou, W.; Lu, Q.-Q.; Zhang, M.-R.; Yang, L.-E. Haplotype Networks of Phycocalidia Tanegashimensis (Bangiales, Rhodophyta) Indicate a Probable Invasion from the South China Sea to Brazil. *Mar. Biodivers.* **2021**, *51*, 33. [[CrossRef](#)]
55. Nguyen, X.-V.; Nguyen-Nhat, N.-T.; Nguyen, X.-T.T.; Nguyen, M.-N.T.; Dao, V.-H.; McDermid, K.J. Three New Records of Marine Macroalgae from Viet Nam Based on Morphological Observations and Molecular Analyses. *Pac. Sci.* **2021**, *75*, 497–512. [[CrossRef](#)]
56. Nguyen, X.V.; Nguyen, T.H.; Dao, V.H.; Liao, L. New Record of *Grateloupia taiwanensis* S.-M. Lin et H.-Y. Liang in Vietnam: Evidence of Morphological Observation and RbcL Sequence Analysis. *Biodiversitas* **2019**, *20*, 688–695. [[CrossRef](#)]
57. Boo, G.H.; Leliaert, F.; Le Gall, L.; Coppejans, E.; De Clerck, O.; Van Nguyen, T.; Payri, C.E.; Miller, K.A.; Yoon, H.S. Ancient Tethyan Vicariance and Long-Distance Dispersal Drive Global Diversification and Cryptic Speciation in the Red Seaweed Pterocladia. *Front. Plant Sci.* **2022**, *13*, 849476. [[CrossRef](#)] [[PubMed](#)]
58. Nguyen-Nhat, N.T.; Dao, V.H.; Nguyen, X.V. New Record of the Rare Brown Alga *Dictyota hauckiana* from Vietnam. *Bot. Mar.* **2019**, *30*, 599–603. [[CrossRef](#)]
59. Van Tu, N.; Trang, L.T.; Phuong Thao, T.T. Diversity and Distribution of the Brown Macroalgae (Phaeophyceae Kjellman, 1891) in Cham Islands, Quang Nam Province, Vietnam. *Acad. J. Biol.* **2021**, *43*, 37–45. [[CrossRef](#)]
60. Sun, Z.; Dao, M.D.; Tran, Q.T.; Dam, D.T. A New Species *Lobophora tsengii* sp. nov. (Dictyotales; Phaeophyceae) from Bach Long Vy (Bailongwei) Island. *J. Oceanol. Limnol.* **2021**, *39*, 2363–2369. [[CrossRef](#)]
61. Tran, L.T.; Leliaert, F.; Vieira, C.; Tran, T.V.; Nguyen, T.V.; Dam, T.D.; De Clerck, O. Molecular Assessment of *Ulva* (Ulvales, Chlorophyta) Diversity in Vietnam Including the New Species *U. vietnamensis*. *Phycol. Res.* **2022**, *71*, 3–24. [[CrossRef](#)]
62. Nguyet, P.M.; Tan, H.T.W.; Mitrovic, S.; Yeo, H.H.T. *A Checklist of the Algae of Singapore*; Raffles Museum of Biodiversity Research: Singapore, 2011; ISBN 9789810701833.
63. Phang, S.M.; Yeong, H.Y.; Ganzon-Fortes, E.T.; Lewmanomont, K.; Prathep, A.; Hau, L.N.; Gerung, G.S.; Tan, K.S. Marine Algae of the South China Sea Bordered by Indonesia, Malaysia, Philippines, Singapore, Thailand and Vietnam. *Raffles Bull. Zool.* **2016**, *2016*, 13–59. [[CrossRef](#)]
64. Kristiansen, J.; Lind, J.F. On the Taxonomic Relation between *Synura curtispina* and *S. favus* (Synurophyceae). *Nord. J. Bot.* **1995**, *15*, 443–448. [[CrossRef](#)]
65. Tanaka, T.; Ho, P.H. Notes on Some Marine Algae from Viet Nam—I. *Mem. Fac. Fish.* **1962**, *11*, 23–40.
66. Garbary, D.J. *The Acrochaetiaceae (Rhodophyta): An Annotated Bibliography*; Koeltz Botanical Books: Darmstadt, Germany, 1987; Volume 77, ISBN 978-3-443-60004-4.
67. Tsutsui, I.; Nang, H.Q.; Dinh, N.H.; Airai, S.; Yoshida, T. The Common Marine Plants of Southern Vietnam. *Umi no Kenkyu (Oceanography in Japan)* **2006**, *15*, 299.
68. Titlyanov, E.A.; Titlyanova, T.V. *Marine Plants of the Asian Pacific Region Countries, Their Use and Cultivation*; Dalnauka: Vladivostok, Russia, 2012.
69. Sutherland, J.E.; Lindstrom, S.C.; Nelson, W.A.; Brodie, J.; Lynch, M.D.J.; Hwang, M.S.; Choi, H.-G.; Miyata, M.; Kikuchi, N.; Oliveira, M.C.; et al. A New Look at an Ancient Order: Generic Revision of the Bangiales (Rhodophyta). *J. Phycol.* **2011**, *47*, 1131–1151. [[CrossRef](#)]
70. Gropman, A.L. Neuroimaging in Mitochondrial Disorders. *NeuroTherapeutics* **2013**, *10*, 273–285. [[CrossRef](#)] [[PubMed](#)]
71. Ní Chualáin, F.; Maggs, C.A.; Saunders, G.W.; Guiry, M.D. The Invasive Genus *Asparagopsis* (Bonnemaisoniaceae, Rhodophyta): Molecular Systematics, Morphology, and Ecophysiology of *Falkenbergia* Isolates. *J. Phycol.* **2004**, *40*, 1112–1126. [[CrossRef](#)]
72. Titlyanov, E.A.; Kiyashko, S.I.; Titlyanova, T.V.; Huyen, P.V.; Yakovleva, I.M. Identifying Nitrogen Sources for Macroalgal Growth in Various Polluted Coastal Areas of Southern Vietnam. *Bot. Mar.* **2011**, *54*, 367–376. [[CrossRef](#)]
73. Zuccarello, G.C.; Kamiya, M.; Ootsuki, R.; De Goër, S.L.; Pedroche, F.F.; West, J.A. New Records of Red Algae from Mangroves in El Salvador and Pacific Mexico, Combining Culture and Molecular Observations. *Bot. Mar.* **2012**, *55*, 101–111. [[CrossRef](#)]
74. Wynne, M. *The Red Algal Families Delesseriaceae and Sarcomeniaceae*; Koeltz Scientific Books: Königstein, Germany, 2014; ISBN 978-3-87429-443-0.
75. Yeurt, A.D.R.N. A Revision of *Amansia glomerata* C. Agardh, *Amansia rhodantha* (Harvey) J. Agardh and *Melanamansia glomerata* (C. Agardh) R. E. Norris (Rhodophyta: Rhodomelaceae). *Bot. Mar.* **2002**, *45*, 231–242. [[CrossRef](#)]
76. Boo, G.H.; Qiu, Y.-X.; Kim, J.Y.; Ang, P.O.J.; Bosch, S.; De Clerck, O.; He, P.; Higa, A.; Huang, B.; Kogame, K.; et al. Contrasting Patterns of Genetic Structure and Phylogeography in the Marine Agarophytes *Gelidiophycus divaricatus* and *G. freshwateri* (Gelidiales, Rhodophyta) from East Asia. *J. Phycol.* **2019**, *55*, 1319–1334. [[CrossRef](#)] [[PubMed](#)]
77. Santelices, B. *Parviphycus*, a New Genus in the Gelidiellaceae (Gelidiales, Rhodophyta). *Cryptogam. Algol.* **2004**, *25*, 313–326.
78. Kawaguchi, S.; Shimada, S.; Wang, H.W.; Masuda, M. The New Genus *Yonagunia* Kawaguchi et Masuda (Halymeniaceae, Rhodophyta), Based on *Y. tenuifolia* Kawaguchi et Masuda Sp. Nov. from Southern Japan and Including *Y. formosana* (Okamura) Kawaguchi et Masuda Comb. Nov. from Southeast Asia. *J. Phycol.* **2004**, *40*, 180–192. [[CrossRef](#)]
79. Tseng, C.K.; Xia, B.-M. On the *Gracilaria* in the Western Pacific and the Southeastern Asia Region. *Bot. Mar.* **1999**, *42*, 209–218. [[CrossRef](#)]

80. Ohno, M.; Terada, R.; Yamamoto, H. The Species of *Gracilaria* from Vietnam. In *Taxonomy of Economic Seaweeds with Reference to Some Pacific Species Vol. VII*; Abbott, I.A., Ed.; California Sea Grant College System: La Jolla, CA, USA, 1999; Volume 7, pp. 99–111.
81. Vo, T.D.; Nishihara, G.N.; Kitamura, Y.; Shimada, S.; Kawaguchi, S.; Terada, R. The Effect of Irradiance and Temperature on the Photosynthesis of *Hydropuntia edulis* and *Hydropuntia euclideanoides* (Gracilariaceae, Rhodophyta) from Vietnam. *Phycologia* **2015**, *54*, 24–31. [CrossRef]
82. Vy, N.X.; Thu, N.N.N.; Hieu, N.T.; Thuy, N.T.X. Selection of Suitable Fragment from RbcL Gene for DNA Barcode Analysis of Family Halymeniaceae, Rhodophyta. *Vietnam J. Mar. Sci. Technol.* **2019**, *19*, 201–213. [CrossRef]
83. Titlyanova, T.V.; Titlyanov, E.A.; Xia, B.; Bartsch, I. New Records of Benthic Marine Green Algae (Chlorophyta) for the Island of Hainan (China). *Nova Hedwig.* **2012**, *94*, 441–470. [CrossRef]
84. Lee, K.M.; Hong, D.D.; Boo, S.M. Phylogenetic Relationships of *Rosenvingea* (Scytosiphonaceae, Phaeophyceae) from Vietnam Based on Cox3 and PsaA Sequences. *Algae* **2014**, *29*, 289–297. [CrossRef]
85. Dai, N.H. Sargasseae. In *Vietnamese Plants*; Hanoi Science and Technology Publishing: Ha Noi, Vietnam, 2007; pp. 1–117.
86. Yoshida, T.; Dai, N.H.; Ajisaka, T.; Noro, T. Verification of Sargassum Species Identified with Subgenera *Bactrophyucus*, *Phylotrichia* and *Schizophycus* in Vietnam. In *Taxonomy of Economic Seaweeds with Reference to Some Pacific Species. Vol. VIII*; Abbott, I.A., Mcdermid, K.J., Eds.; California Sea Grant College: La Jolla, CA, USA, 2002; Volume 8, pp. 95–102.
87. Van Tien, N. *Chlorophyta Pascher (Marine Taxons)*; Hanoi Science and Technology Publishing: Ha Noi, Vietnam, 2007.
88. Linh, N.M.; Quang, P.V.; Luong, C.V.; Hung, V.M.; Tien, D.D. Diversity of Green Macroalgae Genus *Ulva* (Chlorophyta) in Hai Phong. *VNU J. Sci. Nat. Sci. Technol.* **2022**, *38*, 61–69. [CrossRef]
89. Fortes, M.D.; Ooi, J.L.S.; Tan, Y.M.; Prathep, A.; Bujang, J.S.; Yaakub, S.M. Seagrass in Southeast Asia: A Review of Status and Knowledge Gaps, and a Road Map for Conservation. *Bot. Mar.* **2018**, *61*, 269–288. [CrossRef]
90. Phan, T.T.H.; Stiers, I.; Nguyen, T.T.H.; Pham, T.T.; Ton, T.P.; Luong, Q.D.; Triest, L. Spatial and Temporal Distribution of Submerged Aquatic Vegetation in a Tropical Coastal Lagoon Habitat in Viet Nam. *Bot. Mar.* **2018**, *61*, 213–224. [CrossRef]
91. Novak, A.B.; Short, F.T. Leaf Reddening in Seagrasses. *Bot. Mar.* **2010**, *53*, 93–97. [CrossRef]
92. Luong, C.V.; Thao, N.V.; Komatsu, T.; Ve, N.D.; Tien, D.D. Status and Threats on Seagrass Beds Using GIS in Vietnam. *Remote Sens. Mar. Environ. II* **2012**, 8525, 852512. [CrossRef]
93. Hong, D.D.; Bach, N.D.; Hai, L.Q.; Thu, N.H.; Tien, D.D.; Van Dong, N. Phylogenetic Characterization of Some Vietnamese Algae (*Gracilaria*, *Hypnea*, *Caulerpa*, *Scenedesmus*) Based on Their ITS-1 Sequences. In Proceedings of the Basic Research Issues in Life Sciences: Scientific Report of the Second National Conference, Basic Research in Biology, Agriculture, Medicine; 2003; pp. 913–916. Available online: https://www.researchgate.net/publication/285816725_Phylogenetic_characterization_of_some_Vietnamese_algae_based_on_their ITS-1_sequences (accessed on 6 January 2023).
94. Bringloe, T.T.; Sjøtun, K.; Saunders, G.W. A DNA Barcode Survey of Marine Macroalgae from Bergen (Norway). *Mar. Biol. Res.* **2019**, *15*, 580–589. [CrossRef]
95. Vieira, C.; De Ramon N'Yeurt, A.; Rasoamanendrika, F.A.; D'Hondt, S.; Tran, L.-A.T.; Van den Spiegel, D.; Kawai, H.; De Clerck, O. Marine Macroalgal Biodiversity of Northern Madagascar: Morpho-Genetic Systematics and Implications of Anthropogenic Impacts for Conservation. *Biodivers. Conserv.* **2021**, *30*, 1501–1546. [CrossRef]
96. Saunders, G.W. Routine DNA Barcoding of Canadian Gracilariales (Rhodophyta) Reveals the Invasive Species *Gracilaria vermiculophylla* in British Columbia. *Mol. Ecol. Resour.* **2009**, *9* (Suppl. S1), 140–150. [CrossRef] [PubMed]
97. Van Tu, N. Seaweed Diversity in Vietnam, with an Emphasis on the Brown Algal Genus *Sargassum*. Ph.D. Thesis, Ghent University, Ghent, Belgium, 2014. Available online: <http://hdl.handle.net/1854/LU-6940554> (accessed on 6 January 2023).
98. Vy, N.X.; Bujang, J.S.; Papenbrock, J. Variability of Leaf Morphology and Marker Genes of Members of the *Halophila* Complex Collected in Viet Nam. *Aquat. Bot.* **2013**, *110*, 6–15. [CrossRef]
99. Nguyen, X.-V.; Höfler, S.; Glasenapp, Y.; Thangaradjou, T.; Lucas, C.; Papenbrock, J. New Insights into DNA Barcoding of Seagrasses. *Syst. Biodivers.* **2015**, *13*, 496–508. [CrossRef]
100. Nguyen, X.V.; Nguyen-Nhat, N.T.; Nguyen, X.T.; Dao, V.H.; Liao, L.M.; Papenbrock, J. Analysis of rDNA Reveals a High Genetic Diversity of *Halophila major* in the Wallacea Region. *PLoS ONE* **2021**, *16*, e0258956. [CrossRef]
101. Trivedi, S.; Ansari, A.A.; Ghosh, S.; Rehman, H. *DNA Barcoding in Marine Perspectives: Assessment and Conservation of Biodiversity*; Springer: Berlin/Heidelberg, Germany, 2016.
102. Trivedi, S.; Aloufi, A.A.; Ansari, A.A.; Ghosh, S.K. Role of DNA Barcoding in Marine Biodiversity Assessment and Conservation: An Update. *Saudi J. Biol. Sci.* **2016**, *23*, 161–171. [CrossRef]
103. Yang, M.Y.; Geraldino, P.J.L.; Kim, M.S. DNA Barcode Assessment of *Gracilaria salicornia* (Gracilariaceae, Rhodophyta) from Southeast Asia. *Bot. Stud.* **2013**, *54*, 27. [CrossRef] [PubMed]
104. Dumilag, R.V.; Aguinaldo, Z.-Z.A. Genetic Differentiation and Distribution of *Pyropia*, *Acanthophora* (Bangiales, Rhodophyta) in the Philippines. *Eur. J. Phycol.* **2017**, *52*, 104–115. [CrossRef]
105. Zuccarello, G.C.; Critchley, A.T.; Smith, J.; Sieber, V.; Lhonneur, G.B.; West, J.A. Systematics and Genetic Variation in Commercial Shape *Kappaphycus* and Shape *Euclidean* (Solieriaceae, Rhodophyta). *J. Appl. Phycol.* **2006**, *18*, 643–651. [CrossRef]
106. Hong, D.D.; Hien, H.T.M.; Thu, N.H.; Nam, H.S.; Nang, H.Q.; Duc, T.M. Taxonomy of Southeast Asian seaweeds. *Phycologia* **2008**, *51*, 475–476. [CrossRef]
107. Lim, P.E.; Tan, J.; Phang, S.M.; Nikmatullah, A.; Hong, D.D.; Sunarpi, H.; Hurtado, A.Q. Genetic Diversity of *Kappaphycus* Doty and *Euclidean* J. Agardh (Solieriaceae, Rhodophyta) in Southeast Asia. *J. Appl. Phycol.* **2014**, *26*, 1253–1272. [CrossRef]

108. Tan, J.; Lim, P.E.; Phang, S.M.; Rahiman, A.; Nikmatullah, A.; Sunarpi, H.; Hurtado, A.Q. *Kappaphycus malesianus* Sp. Nov.: A New Species of *Kappaphycus* (Gigartinales, Rhodophyta) from Southeast Asia. *J. Appl. Phycol.* **2014**, *26*, 1273–1285. [CrossRef]
109. Song, S.L.; Lim, P.E.; Poong, S.W.; Phang, S.M. Genetic Variation in *Gracilaria tenuistipitata* (Rhodophyta) from Northern Singapore and Neighbouring Countries. *Raffles Bull. Zool.* **2015**, *2015*, 16–23.
110. Nguyen, T.H.; Nguyen Nhat, N.T.; Nguyen, X.T.; Nguyen, V.X. Morphological Variation and Haplotype Diversity of *Halimeda macroloba* and *H. opuntia* (Chlorophyta: Halimedaceae) from Southern Vietnam. *Vietnam J. Mar. Sci. Technol.* **2022**, *22*, 165–176. [CrossRef]
111. Sherman, K.; Hempel, G. The UNEP Large Marine Ecosystem Report: A Perspective on Changing Conditions in LMEs of the World's Regional Seas. In Proceedings of the UNEP Regional Seas Report and Studies; United Nations Environment Programme: Nairobi, Kenya, 2008; pp. 297–308.
112. Morton, B.; Blackmore, G. South China Sea. *Mar. Pollut. Bull.* **2001**, *42*, 1236–1263. [CrossRef]
113. Zheng, F.; Qiu, G.; Fan, H.; Zhang, W. Diversity, distribution and conservation of Chinese seagrass species. *Biodivers. Sci.* **2013**, *21*, 517–526. [CrossRef]
114. Tanto, T.A.; Putra, A.; Hermon, D.; Damanhuri, H. Suitability of Seagrass Ecosystem for Marine Ecotourism in Padang City, West Sumatera Province. *Forum Geogr.* **2018**, *32*, 88–95. [CrossRef]
115. Bujang, J.S.; Zakaria, M.H.; Short, F.T. Seagrass in Malaysia: Issues and Challenges Ahead. In *The Wetland Book: II: Distribution, Description, and Conservation*; Finlayson, C.M., Milton, G.R., Prentice, R.C., Davidson, N.C., Eds.; Springer: Dordrecht, The Netherlands, 2018; pp. 1875–1883. ISBN 978-94-007-4001-3.
116. Fortes, M.D. A Review: Biodiversity, Distribution and Conservation of Philippine Seagrasses. *Philipp. J. Sci.* **2013**, *142*, 95–111.
117. McKenzie, L.J.; Yaakub, S.M.; Tan, R.; Seymour, J.; Yoshida, R.L. Seagrass Habitats of Singapore: Environmental Drivers and Key Processes. *Raffles Bull. Zool.* **2016**, *34*, 60–77.
118. UNEP. National Action Plan for Seagrass Thailand. In Proceedings of the Sixth Meeting of the Regional Working Group for the Seagrass Sub-component of the UNEP/GEF Project: “Reversing Environmental Degradation Trends in the South China Sea and Gulf of Thailand”, Philippines, 2005; pp. 1–8. Available online: <http://www.unepscs.org/remository/startdown/270.html> (accessed on 6 January 2023).
119. Van Luong, C. Study on Characteristics of Seagrass Communities and Their Carbon Storage Capacity in Some Typical Lagoons in Central Vietnam. Ph.D. Thesis, Institute of Ecology and Biology Resources, Ha Noi, Vietnam, 2019.
120. Bray, J.R.; Curtis, J.T. An Ordination of the Upland Forest Communities of Southern Wisconsin. *Ecol. Monogr.* **1957**, *27*, 325–349. [CrossRef]
121. Clarke, K.R.; Gorley, R.N. PRIMER v6: User Manual/Tutorial (Plymouth Routines in Multivariate Ecological Research), PRIMER-e, Plymouth 2006, 192. Available online: https://www.researchgate.net/publication/285668711_PRIMER_v6_user_manualtutorial_PRIMER-E_Plymouth (accessed on 6 January 2023).
122. Spalding, M.D.; Fox, H.E.; Allen, G.R.; Davidson, N.; Ferdaña, Z.A.; Finlayson, M.; Halpern, B.S.; Jorge, M.A.; Lombana, A.; Lourie, S.A.; et al. Marine Ecoregions of the World: A Bioregionalization of Coastal and Shelf Areas. *BioScience* **2007**, *57*, 573–583. [CrossRef]
123. Contreras-Porcía, L.; Meynard, A.; López-Cristoffanini, C.; Latorre, N.; Kumar, M. Marine Metal Pollution and Effects on Seaweed Species. In *Proceedings of the Systems Biology of Marine Ecosystems*; Springer: Cham, Switzerland, 2017; pp. 35–48.
124. Harley, C.D.G.; Anderson, K.M.; Demes, K.W.; Jorve, J.P.; Kordas, R.L.; Coyle, T.A.; Graham, M.H. Effects Of Climate Change On Global Seaweed Communities. *J. Phycol.* **2012**, *48*, 1064–1078. [CrossRef]
125. Tano, S.A.; Halling, C.; Lind, E.; Buriyo, A.; Wikström, S.A. Extensive Spread of Farmed Seaweeds Causes a Shift from Native to Non-Native Haplotypes in Natural Seaweed Beds. *Mar. Biol.* **2015**, *162*, 1983–1992. [CrossRef]
126. Schaffelke, B.; Hewitt, C.L. Impacts of Introduced Seaweeds. *Bot. Mar.* **2007**, *50*, 397–417. [CrossRef]
127. Walker, D.I.; Kendrick, G.A. Threats to Macroalgal Diversity: Marine Habitat Destruction and Fragmentation, Pollution and Introduced Species. *Bot. Mar.* **1998**, *41*, 105–112. [CrossRef]
128. Loc, P.K.; Yen, M.D.; Averyanov, L. Biodiversity in Vietnam. In *Global Biodiversity. Selected Countries in Asia*; Pullaiah, T., Ed.; Apple Academic Press: Ottawa, Canada, 2018; Volume 1, p. 30. ISBN 9780429487743.
129. Ca, V.T.; Hieu, P.V.; Luong, C.V.; Tien, D.D. Application of a Quantitative Assessment Method for the Onservation Potential of the Seagrass Ecosystem in a Pilot Study at Ly Son Islands, Quang Ngai. *J. Mar. Sci. Technol.* **2011**, *11*, 47–56. [CrossRef]
130. WWF. *Root Causes of Biodiversity Loss in Vietnam Summary*; WWF: New York, NY, USA, 1998.
131. Vy, N.X.; Dai, N.H. Re-Assessment of *Sargassum* Beds at Hon Chong Area, Nha Trang Bay, Vietnam. *Seto Mar. Biol. Lab.* **2011**, *41*, 63–69.
132. Hong, D.D.; Ha, N.C. Seaweeds of Vietnam: Opportunities for Commercial Production. In *Sustainable Global Resources of Seaweeds Volume 1: Bioresources, Cultivation, Trade and Multifarious Applications*; Ranga Rao, A., Ravishankar, G.A., Eds.; Springer International Publishing: Cham, Switzerland, 2022; pp. 109–127. ISBN 978-3-030-91955-9.
133. Averyanov, L.V.; Anh, T.; Tran, B.; Ban, N.; Bich, L.; Bien, L.; Binh, N.; Chieu, N.; Duc, P.; Dung, V.; et al. *Vietnam Red Data Book. Part II. Plants. Orchidaceae*; Natural Science and Technology Publishing House: Ha Noi, Vietnam, 2007; p. 611.
134. PEMSEA. Viet Nam Administration for Seas and Islands (VASI) and Ministry of Environment and Natural Resources (MONRE). In *Proceedings of the National State of Oceans and Coasts 2018: Blue Economy Growth of Viet Nam. Partnerships in Environmental Management for the Seas of East Asia (PEMSEA)*; PEMSEA: Quezon City, Philippines, 2018; p. 232.

135. Mantri, V.A.; Kambey, C.S.B.; Cottier-Cook, E.J.; Usandizaga, S.; Buschmann, A.H.; Chung, I.K.; Liu, T.; Sondak, C.F.A.; Qi, Z.; Lim, P.E.; et al. Overview of Global Gracilaria Production, the Role of Biosecurity Policies and Regulations in the Sustainable Development of This Industry. *Rev. Aquac.* **2023**, *15*, 801–819. [[CrossRef](#)]
136. News. Vietnam Maritime Administration. Available online: <https://www.vinamarine.gov.vn/en> (accessed on 10 January 2023).

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.