Wing shape evolution in bombycoid moths reveals two distinct strategies for maneuverable flight

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A wide diversity of wing shapes has evolved, but how is aerodynamic strategy coupled to morphological variation? Here we examine how 1 wing shape has evolved across a phylogenetic split between hawkmoths (Sphingidae) and wild silkmoths (Saturniidae), which have divergent 2 life histories, but agile flight behaviors. Combined with kinematics of exemplar species, we find that these two diverse sister families have 3 evolved two distinct strategies for agile flight. Each group has evolved distinct wing shapes in phylogenetic PCA-space. The notoriously 4 agile hawkmoths have not evolved wing shapes typical of maneuverability, but rather ones that reduce power. Instead their kinematics 5 favor maneuverability, primarily through higher wingbeat frequency. In contrast, silkmoths evolved maneuverable wing shapes and use 6 kinematics that reduce power. Therefore, multiple strategies have evolved to achieve similar aerodynamic performance. We suggest flapping 7 wings provide flexible aerodynamics through kinematics and might release morphological constraints, enabling the diversity of wing shapes 8 across extant flyers. 9

wing shape | moth | flight | evolution | aerodynamics | maneuverability

The functional demands associated with the behavioral repertoire of a species can drive the evolution of locomotor morphology and appendage movement. The wings are a prominent and morphologically diverse features of flying animals (Wootton, 1992; Le Roy et al., 2019), which impact moments of inertia and aerodynamics (Dudley, 2000). Wing shape varies across many groups of flying organisms, but unlike fixed wings, flapping wing aerodynamics also depend on active wing movement. These multiple factors make it challenging to link evolutionary patterns of wing shape to the larger strategies of aerodynamic performance employed by animals or to translate these patterns to engineered flapping wing design.

The wing shape of insects, the most speciose clade of extant animals, likely faces strong selective pressures to meet the functional demands of a species (Wootton, 1992). Many flying vertebrates can "morph" wing shape to modulate aerodynamic forces (Lentink et al., 2007; Crandell and Tobalske, 2011; Riskin et al., 2010; Stowers et al., 2017; Baliga et al., 2019). Flexible shapes may reduce selective pressure on wing morphology. Insects do not contain intrinsic wing musculature and cannot morph wing shape, except

through actuation at the hinge. However, interspecific kinematic flexibility does exist across insects, and 14 insects employ a diversity of kinematics (Dudley, 2000). Further, alteration of the expression pattern of 15 a single gene in *Drosophila melanogaster* can advantageously modify wing shape to significantly enhance 16 aerodynamic agility (Ray et al., 2016), suggesting a strong genotype to phenotype link in wing shape on 17 which selection can act. While the aerodynamics of flight can be altered through the modification of wing 18 shape, size, and movement (kinematics), how these traits evolve within insect clades, whether distinct 19 evolutionary groups adopt consistent flight strategies for, and how evolutionary shifts in wing shape and 20 kinematics impact aerodynamic performance remains unclear. 21

Certain features of wing shape have known aerodynamic consequences for maneuverability, force produc-22 tion, and power, most notably: 1) wing loading, $W_s = S/m_t$, where S is the wing area and m_t is the total 23 body mass, 2) wing aspect ratio, AR, $AR = R^2/S$, where R is wing span length, and 3) the distribution of 24 area along the long axis of a wing (radius of the second moment of area, \hat{r}_2). A decrease in W_s , absent other 25 changes, increases maneuverability. W_s is proportional to turning radius in birds (Burns and Ydenberg, 26 2002; Hedenstrom and Rosen, 2001), bats (Aldridge, 1987; Norberg and Rayner, 1987), and Lepidoptera 27 (Betts and Wootton, 1988) and inversely related to rotational speeds and accelerations during maneuvers 28 in birds (Hedenstrom and Rosen, 2001; Dakin et al., 2018) and Lepidoptera (Betts and Wootton, 1988; 29 Berwaerts et al., 2002). W_s is also positively correlated with flight speed in both vertebrates (Norberg 30 and Rayner, 1987) and insects (Betts and Wootton, 1988; Dudley and Srygley, 1994; Dudley, 2002). A 31 reduction in AR increases both power requirements and maneuverability, typically through a reduction 32 in wing moment of inertia as seen in butterflies (Betts and Wootton, 1988; Cespedes et al., 2015; DeVries 33 et al., 2010). High AR reduces the power requirements of flight by reducing induced drag from wing tip 34 vortices (Norberg and Rayner, 1987; Pennycuick, 1968). An increase in \hat{r}_2 can decrease maneuverability 35 through an increase in the moment of inertia of the wing as more wing area, and likely mass, is distributed 36 more distally along the wing in comparison to a wing of a lower \hat{r}_2 . 37

Although AR, W_s , and \hat{r}_2 are often used to infer performance, these relationships between wing shape and 38 performance assume that other factors remain constant; performance might depend as much on differences in 39 wing kinematics as on wing shape. For example, rotational maneuverability can be enhanced by increasing 40 wing beat frequency, n, which increases active torque generation (Hedrick et al., 2009). Indeed, n increases 41 during maneuvers in hovering hawkmoths (Cheng et al., 2011) and hummingbirds (Cheng et al., 2016b). 42 The power requirements of flight are also dependent on wing movement. While it is possible for insects to 43 produce the same flight behavior through different sets of kinematics (Hedrick and Daniel, 2006), insects 44 employ kinematics that minimize energy consumption (Berman and Wang, 2007). Therefore, concomitant 45 changes in wing kinematics may produce very different aerodynamic implications for wing shape. 46

The moth superfamily Bombycoidea, provides a prime opportunity to test how wing shape evolves in a 47 closely related species within a diverse clade of more than 5,000 species (Kitching et al., 2018; Lemaire 48 and Minet, 1998). Within Bombycoidea, the sister families Sphingidae and Saturniidae (Breinholt et al., 49 2018; Kawahara et al., 2019; Hamilton et al., 2019), have ~ 1600 and ~ 3400 described species respectively 50 (Kitching et al., 2018). These two families display an extraordinary diversity of wing shape, and strikingly 51 divergent life history strategies (Tammaru and Haukioja, 1996) and flight behaviors. Within Sphingidae 52 (hawkmoths), Manduca sexta, and more recently Daphnis nerii, Hyles lineata, and Macroglossum stellatarum, 53 are models for studies on flight control (Sponberg et al., 2015; Willmott and Ellington, 1997a; Natesan 54 et al., 2019), vision (Windsor et al., 2014; Stöckl et al., 2017b), olfaction (Hildebrand, 1996; Gage et al., 55 2013; Riffell et al., 2013), and multisensory integration (Roth et al., 2016). Hawkmoths are most known 56 for their ability to sustain long duration bouts of hovering while feeding from flowers (Wasserthal, 1993; 57 Farina et al., 1994; Sprayberry and Daniel, 2007; Sponberg et al., 2015). In addition to the evolution of 58 the hovering behavior, hawkmoths are often described as active, fast-flying, and maneuverable (Callahan, 59 1965; Tuttle, 2007). Some hawkmoths can successfully track flowers oscillating up to frequencies of 14 Hz 60 (Sponberg et al., 2015; Stöckl et al., 2017a), suggesting a high degree of maneuverability. Saturniidae (wild 61 silkmoths) is one model group for studying the predator-prev arms race (Barber et al., 2015; Kawahara 62 and Barber, 2015; Rubin et al., 2018). Many silkmoth species display a flight behavior often described as 63 bobbing, but can also be fast and agile when escaping from predators (Jacobs and Bastian, 2016; Janzen, 64 1984; Lewis et al., 1993). Silkmoths lack functional mouth parts and must rely on the strictly finite energy 65 stores, gathered during the larval period, during their entire reproductive adult life stage (Tuskes et al., 66 1996). 67

Here we use the wing diversity and life history differences between the sister families, Sphingidae and Saturniidae, to test if they adopt distinct flight strategies with correspondingly distinct wing shapes. First, we use museum specimens to explore how wing shape varies in an explicitly phylogenetic context across a diverse group of agile flying insects. We test the hypothesis that wing morphology diverged on two distinct trajectories when hawkmoths and silkmoths split. Alternatively, differences in wing shape and size might arise more within each group, driven by the demands of individual species.

⁷⁴ We next consider measures of wing morphology (AR, W_s , and \hat{r}_2) across Bombycoidea to test if the two ⁷⁵ groups have shapes associated with maneuverable flight. Given their flight performance and maneuverability, ⁷⁶ we hypothesize that both groups evolved forewings of low AR, W_s , and \hat{r}_2 . Alternatively, because most ⁷⁷ hawkmoths (possible exception of the Ambulycini tribe) require high power output in order to sustain long ⁷⁸ bouts of hovering while feeding and because adult stage silkmoths do not feed, we hypothesize that both ⁷⁹ groups have evolved wing shapes that reduce power requirements by increasing wing AR or length or that ⁸⁰ the two groups diverge for separate strategies.

Finally, to assess the potentially subtle interplay of shape, size, and kinematics, we quantified three-81 dimensional wing kinematics during forward flight from live specimens of two species representing the wing 82 shapes of each family. We estimated quasi-steady aerodynamic force production and power requirements 83 using a blade element model (Sane and Dickinson, 2002; Faruque and Humbert, 2010b; Cheng et al., 2016a; 84 Han et al., 2015). Using these two species, we can place the broader differences in wing shape into the 85 context of kinematic differences for these particular species. Determining whether major transitions in 86 flight strategy occur through changes in wing shape, kinematics, or their combination will broadly inform 87 the evolution of animal flight and our ability to infer aerodynamic consequences from wing shape alone. 88

MATERIALS AND METHODS

Phylogenetics. In order to understand the evolution of wing shape across the Bombycoidea, we sampled 90 representatives of all Bombycoidea families. In total, the phylogenetic dataset included 57 species and 91 one outgroup – the Lasiocampidae, the sister lineage to the Bombycoidea. Sampling was highest in the 92 Saturniidae (25 sp.) and Sphingidae (24 sp.). Sequences were a mixture of previously sequenced individuals 93 from prior studies (Breinholt et al., 2018; Rubin et al., 2018) and novel specimens (see Table S1). AHE 94 sequences represented 53 species, while four species had their AHE loci mined from transcriptomic data 95 (Table S1). Nineteen species were newly sequenced for this project, while 38 came from previously published 96 datasets (Table S1). Species were chosen in order to capture the majority of the wing shape diversity 97 throughout the Sphingidae and Saturniidae, based on availability in the collections at the Florida Museum 98 of Natural History, Gainesville, FL, USA (FLMNH). 99

Specimens were obtained from historically preserved dry collections and molecular tissue collections 100 stored at the Florida Museum of Natural History. DNA extraction from pinned museum specimens followed 101 the protocol outlined in (Hamilton et al., 2019). Field-collected specimens were stored in $\geq 95\%$ ethanol, 102 RNAlater (Sigma Aldrich, Saint Louis, MO, United States), or papered and dried with silica gel. Genomic 103 DNA was extracted using OmniPrep Genomic DNA Extraction Kits (G-Biosciences, St. Louis, MO, USA) 104 and DNeasy Blood and Tissue Kits (Qiagen, Valencia, CA, USA). DNA concentration was evaluated 105 through agarose gel electrophoresis and fluorometry using a Qubit 2.0 (Invitrogen, Thermo Fisher Scientific, 106 Carlsbad, CA, USA). 107

We used the Bombycoidea-specific 'BOM1' Anchored Hybrid Enrichment (AHE) probe set to target 571 loci (Hamilton et al., 2019). AHE is a targeted-sequencing methodology designed to capture hundreds of unique orthologous loci (i.e., single copy, phylogenetically-informative markers) from across the genome, for resolving both shallow and deep-level evolutionary relationships (Lemmon et al., 2012; Breinholt et al., 112 2018).

Library preparation, hybridization enrichment, and Illumina HiSeq 2500 sequencing (PE100) was carried out at RAPiD Genomics (Gainesville, FL, USA). Specimen wing vouchering and tissue storage methods follow (Cho et al., 2016). All DNA extracts and specimens preserved in ethanol, RNAlater, or those freshly papered were stored at -80° C at the FLMNH, McGuire Center of Lepidoptera and Biodiversity (MGCL). Historically preserved dry collection specimens were kept in their traditional storage method - pinned in their tray or papered in a box, at the MGCL.

The bioinformatics pipeline of (Breinholt et al., 2018) was used to clean and assemble raw Illumina reads for each AHE locus. The pipeline uses a probe-baited iterative assembly that extends beyond the probe region, checks for quality and cross contamination due to barcode leakage, removes paralogs, and returns a set of aligned orthologs for each locus and taxon of interest. To accomplish these tasks, the pipeline uses the Bombyx mori genome (Xia et al., 2004), and the BOM1 AHE reference library. Previously published scripts (Breinholt et al., 2018) and instructions (Hamilton et al., 2019) on the pipeline are available in Dryad (https://doi.org/10.5061/dryad.5df18fp).

Loci for phylogenetic analysis were selected by applying a cutoff of $\geq 60\%$ sampled taxa recovery (i.e., for 126 a locus to be included in the analysis, the locus had to be recovered in at least 60% of the sampled taxa). 127 The pipeline evaluates density and entropy at each site of a nucleotide sequence alignment. We elected 128 to trim with entropy and density cutoffs only in flanking regions, allowing the probe region (exon) to be 129 converted into amino acid sequences. For a site (outside of the probe region) to remain, that site must then 130 also pass a 60% density and 1.5 entropy (saturation) cutoff, rejecting sites that fail these requirements. A 131 higher first value (60) increases the coverage cutoff (e.g., a site is kept if 60% of all taxa are represented at 132 that site). A higher second value (1.5) increases the entropy cutoff (i.e., entropy values represent the amount 133 of saturation at a site); sites with values higher than 1.5 possess higher saturation and are thus deleted). 134 Following (Hamilton et al., 2019), we built and utilized a Pr+Fl dataset; a concatenated dataset consisting 135 of 605 probe regions + a flanking supermatrix for phylogeny inference. The final dataset comprised 606 loci 136 and 222,369 bp. 137

AliView v1.18 (Larsson, 2014) was used to translate the nucleotides to amino acids, check for frame shifts, and edit sequencing errors or lone/dubious indels. Because flanking sequences are generally non-coding and sites have been deemed homologous, the flanking sequences (i.e., those before and after the probe regions), were separated from the exons, then combined and treated together as an independent partition. Due to the filtering steps in the bioinformatics pipeline (i.e., site orthology, and density and saturation evaluation), the flanking partition can be viewed as a SNP supermatrix, where each site is homologous and independent, but uninformative sites, saturated sites, or sites with large amounts missing data removed. Specimens whose AHE loci were mined from transcriptomes did not have flanking data due to the nature
 of transcriptomic data.

A concatenated supermatrix was assembled using FASconCAT-G v1.02 (Kueck and Longo, 2014). 147 Phylogenetic inference was performed in a maximum likelihood (ML) framework using IQ-TREE v1.5.3 148 (Nguyen et al., 2015). Within this framework we searched for the most appropriate model of nucleotide 149 substitution, as well as 1000 random addition sequence (RAS) replicates, and 1000 replicates each for both 150 ultrafast bootstraps (UFBS) and SH-aLRT tests to find the "best" tree and node support. We classified 151 nodes as well-supported if they were recovered with support values of UFBS >95 and SH-aLRT >80152 (Minh et al., 2013; Nguyen et al., 2015). All branch length estimates are in units of substitutions per 153 site. In the ultrametric tree used for analyses, the branch lengths were scaled relative to each other. All 154 pipeline steps and phylogenomic analyses were conducted on the University of Florida HiPerGator HPC 155 (http://www.hpc.ufl.edu/). All alignment FASTA files, partition files, tree files, and other essential data 156 files used for phylogenetic inference are available as supplementary materials on Drvad. 157

Museum Specimen Imaging. Moth specimens from the collections at the FLMNH were imaged using a 60mm lens (Canon, Tokyo, Japan) mounted to a Canon EOS 7D camera (Canon, Tokyo, Japan). At least one individual of each of the 57 species was imaged. See Table S1 for details on the number and sex of specimens imaged per species. For this analysis, male specimens were analyzed when available (53 of 57 species). We chose to focus on males because they are known to exhibit higher flight activity in comparison to females (Gilchrist, 1990; Le Roy et al., 2019).

Body and Wing Measurements and Morphometrics. The body and wing morphology was digitized from 164 museum images using the StereoMorph package (version 1.6.2) (Olsen and Westneat, 2015) in R (version 165 3.4.2; The R Foundation for Statistical Computing). Eight landmarks were digitized to characterize body 166 size and shape (Fig. 1A). The rostral and caudal bases of both the forewing and hindwing were also digitized. 167 Finally, a series of third order Bezier curves were used to outline the right forewing (Fig. 1A). The curves 168 were then resampled using the StereoMorph package to generate 50 evenly spaced points (semilandmarks) 169 around the wing perimeter. For species with multiple individuals, each wing was aligned at the rostral base 170 of the wing and forewing shape was averaged to produce a mean set of 50 semilandmarks to be used in 171 further analysis. Similarly, for species with multiple individuals, the length and width of each body segment 172 (head, thorax, and abdomen) were averaged to obtain mean values for each species. 173

The digitized output of each moth was further analyzed in MATLAB (version R2018b - 9.5.0.944444). Body mass was estimated by modeling the body as three ellipsoids: one for the head, thorax, and abdomen. The length and width of each body segment were measured as the distance between the minimum and maximum value for that segment. The width of the head was assumed to be equal to its length. The dorso-ventral depth of every segment was assumed to be equal to its width. The density of each body segment was assumed to be equal to water (Ellington, 1984a).

Forewing measurements began by rotating each forewing so its long axis was perpendicular to the long 180 axis of the body. Wing length (R) was measured as the distance between the minimum and maximum value 18 of the wing outline. All additional wing parameters (AR, W_s , and r_2) were calculated following (Ellington, 182 1984a). The wing was assumed to have a thickness equal to 0.5% mean chord length and a density equal 183 to water. The wing area was then filled with equally distributed points that were each assigned an equal 184 mass fraction. Wing beat frequency (n) was estimated using an equation derived through dimensional 185 analysis of insect body mass and wing area (Table A.1; (Deakin, 2010)). Finally, body mass specific mean 186 inertial power of the wings (P_{acc}) was calculated (see Table A.1) following published methods (Ellington, 187 1984c). Stroke amplitude was not available for every species so the same half stroke amplitude of 60° was 188 used across all species. The potential impact of different kinematics is discussed in the comparison of the 189 exemplar species from which kinematics were obtained. 190

To examine the variation in forewing shape while accounting for phylogeny, we conducted a phylogenetic 191 principal components analysis (pPCA) (Revell, 2009). A pPCA finds the dimensions of wing shape that 192 maximize the covariance of the component shape variables, but corrects the covariance with a relatedness 193 matrix that takes into account the shared evolutionary history of the samples. For each species, the mean 194 forewing semilandmarks were aligned by Procrustes superimposition to obtain size and orientation corrected 195 Procrustes coordinates (implemented with gpagen function in Geomorph R package (version 3.1.2)). Next, 196 the pPCA was conducted on the Procrustes coordinates for each species (implemented with the phyl.pca 197 function in the R package Phytools (version: 0.6-60) (Revell, 2012)). We then projected wing shape back 198 on to the first four pPC axes (Olsen, 2017). To test if clades were significantly clustered in four dimensional 199 pPC morphospace, we conducted a MANOVA where the pPC scores of pPC1, 2, 3 and 4 were used as 200 the response variable and clade (hawkmoths, silkmoths, or ancient bombycoid lineages) was the factor. 20 Additionally, we tested for phylogenetic signal in each wing shape variable across the full phylogeny and 202 within each subclade (hawkmoths, silkmoths, and the clade comprising the hawkmoth-silkmoth sister 203 families). These analyses were conducted in RStudio (version 1.1.383; RStudio, Inc., Boston, MA). 204

Ancestral State Reconstruction. Ancestral state reconstructions were conducted to determine the evolutionary history of each wing trait as well as the wing shape PC scores. Maximum likelihood ancestral state reconstructions (fastAnc in phytools R package (version 0.6.60) (Revell, 2012)) used a Brownian motion model of evolution and estimates the maximum likelihood state estimations for all internal nodes by rerooting the phylogeny at each internal node and computing the contrasts state at the root according to

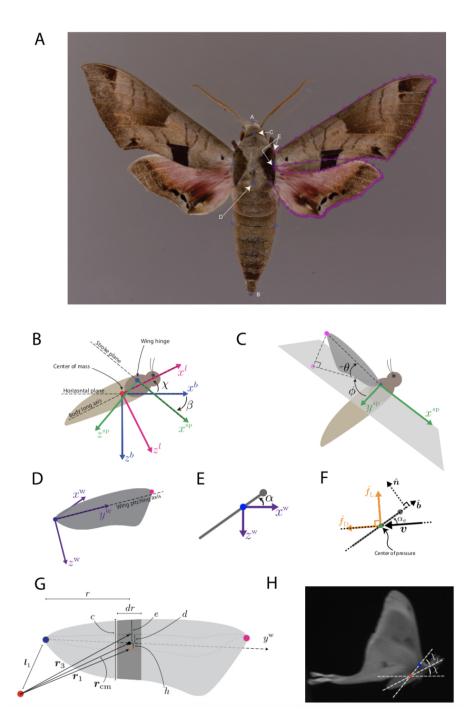


Fig. 1. A Summary of body landmarks and coordinate systems. Eight landmarks (blue points) capture body size and wing attachment. Sagittal landmarks were placed at the tip of the head (1), caudal tip of the abdomen (2), junction between the head and thorax (3), and junction between the thorax and abdomen (4). Parasagittal landmarks were placed at the widest points of the thorax and abdomen and the rostral and caudal base of the forewing (5) and hindwing. Each wing was outlined using a series of Bezier curves (purple points). Each curve was free to vary in perimeter length, and, typically, more points were used to generate the outline around more complex regions of the wing (see distal tip versus leading edge). **B** The body-attached coordinate frame (blue), the body-long coordinate frame (pink) and the stroke-plane frame (green). The red dot represents the location of the center of mass and the blue dot represented the wing hinge. β is the stroke-plane angle and χ is the body angle. **C** Definitions of wing kinematic angles: ϕ and θ defined with respect to the stroke-plane. **D**-**E** The wing-attached coordinate frame and wing-pitching angle α . **F** Relative airflow, effective angle of attack, and lift and drag components of the translational aerodynamic force. **G** Various length parameters relevant to a single wing strip. Red, blue, and pink circles correspond to the body center of mass, wing hinge, and wing tip. Dashed green, orange, and black lines are the quarter-chord, half-chord, and wing pitching axis, respectively. **H** Inclination angle of the wing-hinge point (blue) from the center of mass (red).

the contrasts algorithm (Felsenstein, 1985).

Live specimens. Live specimens were obtained as pupae from a breeder for two species: Actias luna (Saturniidae) and Eumorpha achemon (Sphingidae). These two species were chosen because they were readily available in large numbers and were also widely separated in pPCA morphospace, providing a general representative from each family. Caterpillars of each species were acquired by collecting eggs from local adult moths, and all caterpillars were reared on species-specific host plants. Pupae were stored in an incubator (Darwin Chambers, model: IN034LTDMMP, Saint Louis, MO) set to a temperature of 23° C and a relative humidity of 65%.

Three-dimensional Kinematics. Moths were transferred to the wind tunnel in individual containers with a moist tissue to prevent desiccation. Each individual was dark adapted at the wind tunnel for 1hr prior to the start of filming. Flight experiments were conducted in a 100×60.96 working section of an open-circuit Eiffel-type wind tunnel (ELD, Inc, Lake City, MN). The stream-wise turbulence of the wind tunnel does not exceed 0.5% and the flow speed did not vary by more than 2%. For a detailed overview of the specifications of the wind tunnel see (Matthews and Sponberg, 2018).

Moths were enticed to fly by providing a mild wind speed of 0.7 ms^{-1} . Flight bouts were filmed at 2000 224 frames s^{-1} for *E. achemon* and 1000 frames s^{-1} for *A. luna* using three synchronized Photron high-speed 225 digital video cameras (Mini UX 100; Photron, San Diego, CA, USA) at a resolution of 1280×1024. Two 226 cameras (one upwind and one downwind) were positioned below the wind tunnel test section at a 45° angle 227 relative to the direction of flow. A third camera was placed laterally and orthogonal to plane of the first 228 two cameras. The working section of the wind tunnel was illuminated with six 850Nm IR light (Larson 229 Electronics, Kemp, TX, USA) and a neutral density filter, white LED "moon" light (Neewer CW-126) to 230 control illumination conditions. Videos were digitized and calibrated in XMALab (Knorlein et al., 2016). 231 From the exported 3D points we characterized the wing kinematics by calculating the following variables 232 (Figs. 1 B-E): wing beat frequency (n), stroke plane angle (β) , sweep angle (ϕ) , deviation angle (θ) , and 233 feathering angle (α). The definition of each angle follows those outlined in (Willmott and Ellington, 1997a). 234

Extracting the 3D Time-series Data. We extracted the time series data of 3D coordinates of seven landmark points on a moth's body in the lab frame: head, thorax, abdomen, left wing hinge, right wing hinge, right wing tip and a point on the trailing edge of the forewing. We tracked the coordinates of head, thorax and abdomen to estimate the orientation of the body; and the points on the right wing hinge, right wing tip and trailing edge to estimate the wing kinematics. In videos, we only extracted data from forward flight bouts. From each individual, we digitized 1 complete wingstroke that was contained within a larger set of wingstrokes during forward flight. We never digitized the first or final wingstroke from a forward flight ²⁴² bout. For each wing stroke, at minimum, every other frame was digitized, and, in most videos, we digitized ²⁴³ every frame. To specify the body coordinate system, we defined the absolute horizontal plane using the ²⁴⁴ lateral camera view. Using the horizontal plane, we defined the absolute vertical vector, i.e. the direction of ²⁴⁵ the force of gravity.

Body Coordinate Frames. We defined a body-attached frame to specify the direction of motion, and a 246 body-long frame to keep track of the orientation of the moth's body as shown in Fig. 1B. These frames 247 share a common origin located at the center of mass. The body-long positive x-axis, x^{l} , points from the 248 center of mass towards the center of head; the z^{l} -axis is perpendicular to x^{l} dorsally and lies in a vertical 249 plane that splits the moth's body into symmetric halves; the y^{l} -axis is the cross product of z^{l} and x^{l} 250 according to the right-handed coordinate system. The body-attached positive x-axis, $x^{\rm b}$, starts from the 251 center of mass and points in a direction that is the projection of x^{l} -axis on the absolute horizontal plane; 252 $z^{\rm b}$ points towards the direction of gravity; $y^{\rm b}$ is the cross product of $z^{\rm b}$ and $x^{\rm b}$. This makes the $x^{\rm b}y^{\rm b}$ -plane 253 the absolute horizontal plane irrespective of moth's body orientation. 254

Stroke-plane Coordinate Frame of the Right Wing. We used the definition of stroke-plane in (Willmott and Ellington, 1997a) to estimate the stroke-plane for each wing stroke and then attached a coordinate frame. The origin of a stroke-plane frame is at the wing hinge point as shown in Fig. 1B. Anatomically, we defined the wing hinge point as a single point located at one-third the distance from the rostral to the caudal wing hinge. For the right wing, the positive x^{sp} -axis is in the direction of the downstroke and lies within the $x^{b}-z^{b}$ plane; y^{sp} is outward from the right wing hinge parallel to the y^{l} -axis in a direction from the left wing hinge to the right wing hinge; and z^{sp} is the cross-product of x^{sp} and y^{sp} .

Wing-attached Coordinate Frame of the Right Wing. To calculate the forces on the wing at each time 262 instance, a coordinate frame attached to the wing is required. Our choice of such a coordinate frame is 263 shown in Fig. 1D. The origin of this wing-attached frame is also at the wing hinge point. The y-axis of 264 the wing-attached coordinate frame, y^{w} , is the anatomical wing-pitching axis of the moth, which was set 265 to be perpendicular to the body-long axis and lied in the same plane as the forewing. The wing-attached 266 coordinate frame is obtained by sequentially rotating the stroke-plane frame through wing kinematic angles 267 ϕ and θ about the current z- and x-axis respectively. Hence, the wing-attached frame rotates with the ϕ 268 and θ rotations of the wing. This means that when both ϕ and θ are equal to zero, the stroke-plane and 269 wing-attached coordinate frames are perfectly aligned. For simplicity, our model's wing-attached frame 270 does not rotate with the wing's pitching motion (α). 271

Right Wing Kinematics. The 3D time-series data of the trajectories were linearly interpolated for any frame 272 that was not digitized and then smoothed using a moving-average filter with a window length of 10 frames. 273 Next, we transformed the data from the camera-calibrated frame to the wing-attached frame and split the 274 data into wing strokes starting at a wingtip position that can be kept consistent across different wing strokes. 275 The series of transformations we applied simplified the extraction of the time series data of individual 276 wing strokes from the start of a downstroke to the end of the subsequent upstroke. We applied a series of 277 transformations to transform the data from the camera-calibrated frame to the wing-attached frame and 278 then split the data. The origin was shifted to the right wing hinge through a translation. This allowed the 279 movement of the wing tip to be restricted to just the rotations. 280

The stroke plane was defined for each wing stroke using a best-fit line through the 3D wing tip trajectory and relied on the vector between the left and right wing hinges. The stroke plane was fit to each wing stroke separately. Next, we calculate the two angles of the wing kinematics: the wing sweep angle (ϕ) and the deviation angle (θ) with respect to the wing-stroke, as defined in Fig. 1C. These angles are basically the azimuth and elevation angles, respectively, of the wing tip with respect to the stroke-plane frame following the definitions in (Willmott and Ellington, 1997a). However, we introduced a sign flip to the deviation angle θ to specify it as a (positive) rotation about our x^{w} -axis.

Next, we calculated the wing pitching (feathering) angle α is shown in Fig. 1E, which is the angle 288 of rotation of the wing about the y^{w} -axis. We found this angle by first numerically aligning the wing's 289 surface-plane with the stroke-plane and then calculating the angle between the two planes. The wing's 290 surface-plane was specified by two points tracked on the forewing and the third point at the wing hinge. 291 The alignment was performed by reversing the θ (stroke deviation) rotation at each time instance. The 292 stroke-plane angle β for each wingstroke was calculated as the smaller angle between the stroke-plane and 293 the $x^{b}-y^{b}$ plane, assuming that during one wingstroke the stroke plane did not rotate with respect to the 294 body-attached frame. The body angle specifies the pitch orientation of the body-long coordinate frame 295 with respect to the body-attached frame. The body angle χ for each wing stroke was calculated as the 296 smaller angle between the wing stroke-averaged x^{l} -axis and the $x^{b}-y^{b}$ plane. 297

Fitting a Fourier Series to the Wing Kinematics. In total, we extracted right wing kinematics from three individual moths of both species for a total of three wing strokes from *E. achemon* and three wing strokes from *A. luna*. For each species, we first time-scaled the wing kinematics data from the three wing strokes to their mean time period and calculated the mean ϕ , θ and α . We then, for each species, fit the following third order Fourier series to the species averaged wing kinematic angles:

$$\phi(t) = a_{\phi,0} + \sum_{k=1}^{3} a_{\phi,k} \cos(2\pi knt) + b_{\phi,k} \sin(2\pi knt),$$
[1]

$$\theta(t) = a_{\theta,0} + \sum_{k=1}^{3} a_{\theta,k} \cos(2\pi knt) + b_{\theta,k} \sin(2\pi knt),$$
 [2]

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$$\alpha(t) = a_{\alpha,0} + \sum_{k=1}^{3} a_{\alpha,k} \cos(2\pi knt) + b_{\alpha,k} \sin(2\pi knt),$$
[3]

where *n* is the wingbeat frequency and $a_{\phi,k}$, $a_{\theta,k}$, $a_{\alpha,k}$, $b_{\phi,k}$, $b_{\theta,k}$, $b_{\alpha,k}$ are the Fourier series coefficients. The values of the Fourier coefficients are summarized in Table 2 and the graphs of the Fourier-fitted wing kinematics are shown in Fig. 4.

The blade element model. A blade element model was used to evaluate the quasi-steady aerodynamic forces 309 produced during flapping flight for two species: A. luna and E. achemon (details in the next section). 310 Briefly, the model estimated the contribution of translational, rotational, and added mass to the total 311 aerodynamic force (Sane and Dickinson, 2001, 2002; Faruque and Humbert, 2010a,b; Han et al., 2015; Kim 312 et al., 2015; Cheng et al., 2016a). Although the main focus of this manuscript is on the forewing, to ensure 313 accurate comparisons of aerodynamic forces during flight between these two species, we used the total 314 wing area and shape generated by the overlapping forewing and hindwing as the wing shape in our blade 315 element model. The wing size and shape used for each species was the mean wing size and shape calculated 316 from the museum specimens used in this study. Several configurations of the model were used to assess the 317 relative contribution of wing shape, size, kinematics and flight speed to aerodynamic force production. The 318 specific kinematic and flight speed variables input to each model are outlined in the results. In the base 319 model, Model 1, stroke plane angle and flight velocity (horizontal and vertical) parameters are set equal to 320 that of the species mean obtained from the three-dimensional kinematic analysis. In Model 1, the wing 321 kinematics are those of a third order Fourier series fit to the mean kinematics (three wing strokes from 322 three individuals) of each species. 323

The blade-element model for forward flight includes the effects due to the steady translational motion 324 of the wing as well as some of the unsteady nature of translation (leading edge vortex, for example) and 325 rotation of the wing (Ellington et al., 1996; Hedrick and Daniel, 2006; Kim and Han, 2014). Some of 326 these unsteady effects can be captured by using empirically measured lift and drag coefficients, in addition 327 to considering the effect of force due to the added mass of the air around the airfoil (Han et al., 2015; 328 Usherwood and Ellington, 2002). Therefore, in our model, we assumed the total aerodynamic force as the 329 sum of the translation, rotational, and added mass forces and used empirical coefficients of lift and drag 330 taken from a robotic flapper using dynamically scaled kinematics from the hawkmoth Manduca sexta (Han 331 et al., 2015). 332

First, we provide a summary of the overall approach and then detail the specific formulation of each

step in the blade element model in subsequent sections. All symbols the mathematical notation used in 334 the model are defined in Table A.1. We started by considering the wing of each species as a thin, rigid 335 plate. As is standard in blade element models, we divided the wing into 200 chord-wise strips so that each 336 strip could be treated as an airfoil. To calculate the aerodynamic forces on a strip, we next determined 337 airflow velocity relative to the wing. For this, we modeled the wing motion according to the measured wing 338 kinematics: sweep, deviation and pitching motion relative to a stroke plane inclined at a constant angle. 339 We also consider the contribution of the body translational and rotational velocities to the overall relative 340 airflow velocity, as in previous models (Faruque and Humbert, 2010a; Cheng et al., 2016a). This causes 34 each blade element to have its own relative airflow velocity and effective angle of attack. We calculated 342 the translational and rotational aerodynamic force of each blade element following (Ellington, 1984b; Kim 343 and Han, 2014; Cheng et al., 2016a). We calculated the added-mass force in a non-inertial frame following 344 (Maybury and Lehmann, 2004; Sane and Dickinson, 2001), which considers both the translational and 345 rotational acceleration of the wing as well as the non-inertial nature of the wing-attached frame. We 346 calculated all the aerodynamic forces in a coordinate frame attached to the wing and then summed over all 347 the strips to calculate the total force on the wing. We also calculated aerodynamic moments using moment 348 arms that assume translational force acting at a quarter-chord distance from the leading edge while the 349 rotational and added-mass forces act at a half-chord distance (Ellington, 1984b; Han et al., 2015; Truong 350 et al., 2011). Then we transformed aerodynamic forces and moments to the body-attached frame, which 35 is an inertial frame if the moth is flying at a constant velocity with no body rotations. In the end, we 352 calculated induced power for further comparison of the flight performance of the two species. 353

Relative Airflow Velocity. We defined relative airflow velocity v of a small blade element strip as the velocity of the airflow relative to the strip. This relative airflow is caused by the overall motion of the strip relative to the surrounding air due to its rotation about the wing hinge, body translation and rotation, and possibly wind.

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$$\boldsymbol{v}^{\mathrm{w}} = -\left(\boldsymbol{v}_{\mathrm{b}}^{\mathrm{w}} + \boldsymbol{\omega}_{\mathrm{b}}^{\mathrm{w}} imes \boldsymbol{r}_{\mathrm{cm}}^{\mathrm{w}} + \begin{bmatrix} -r\dot{\phi} \\ 0 \\ r\dot{\theta} \end{bmatrix}\right)$$
[4]

where \boldsymbol{v}_{b}^{w} is the body velocity relative to the wind but measured in the wing-attached frame, $\boldsymbol{\omega}_{b}^{w}$ is the body angular velocity pseudovector in the wing-attached coordinate frame, \boldsymbol{r}_{cm}^{w} is the vector from the body center of mass to the center of the strip, r is the distance from wing hinge to the vertical mid-chord line of the strip (see Fig. 1G), and $\dot{\phi}$ and $\dot{\theta}$ are the stroke positional (sweep) and stroke deviation angular velocities of the wing, respectively. For the calculations performed on the data relevant to this paper, there were no body rotations so $\boldsymbol{\omega}_{b}^{w}$ was equal to zero.

The Effective Angle of Attack. The effective angle of attack of the strip is defined as the angle between the chord line vector from leading edge to the trailing edge and the relative airflow vector. This angle is calculated as

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$$\alpha_{\rm e} = \cos^{-1} \left(-\hat{\boldsymbol{b}}^{\rm w} \cdot \hat{\boldsymbol{v}}^{\rm w} \right), \quad \alpha_e \in [0, \pi).$$
 [5]

where $\hat{\boldsymbol{b}}^{w} = \begin{bmatrix} \cos \alpha & 0 & -\sin \alpha \end{bmatrix}^{'}$ is the unit vector along the chord line in the direction from the trailing edge of the strip to its leading edge, and α is the feathering angle of the strip. We defined a second effective angle of attack α_{r} to set bounds on the values of the effective angle of attack so that it remains between 0 and $\pi/2$ radians. In the calculation of the lift and drag aerodynamic coefficients, the effective angle of attack was restricted between 0 and $\pi/2$ because the coefficients we used from (Han et al., 2015) were experimentally measured for the effective angles of attack only in this range.

$$\alpha_{\rm r} = \begin{cases} \alpha_{\rm e} & 0 \le \alpha_{\rm e} \le \frac{\pi}{2} \\ \pi - \alpha_{\rm e} & \frac{\pi}{2} < \alpha_e \le \pi \end{cases}$$
[6]

Moreover, this definition of the effective angle of attack keeps the lift and drag coefficients positive and simplifies the model because the direction of the lift can just be specified by the lift force direction vector $\hat{f}_{\rm L}$ elaborated in the next section.

Translational Aerodynamic Force. The translational aerodynamic force is the sum of the lift and drag forces on the wing and acts at the center of pressure. We assumed the center of pressure to be located on the wing at a distance one-quarter chord length behind the leading edge (green dashed line in Fig. 1G), because this is the region at which the bound vortex has been regarded to be concentrated according to the thin airfoil theory for both steady and unsteady aerodynamic effects (Ellington, 1984b). The lift and drag forces were calculated using the aerodynamic coefficients of hawkmoth Manduca sexta taken from (Han et al., 2015). The equations of these forces acting on a small wing strip of width dr are as follows (Cheng et al., 2016a).

$$d\boldsymbol{f}_{\mathrm{L}}^{\mathrm{w}} = \frac{1}{2}\rho C_{\mathrm{L}} v^2 c \ dr \ \boldsymbol{\hat{f}}_{\mathrm{L}}^{\mathrm{w}} , \qquad [7]$$

$$d\boldsymbol{f}_{\mathrm{D}}^{\mathrm{w}} = \frac{1}{2}\rho C_{\mathrm{D}} v^2 c \ dr \ \boldsymbol{f}_{\mathrm{D}}^{\mathrm{w}} , \qquad [8]$$

where ρ is the air density, the aerodynamic coefficients (Han et al., 2015)

 $C_{\rm L}(\alpha_{\rm r}) = 1.552\sin(\alpha_{\rm r})\cos(\alpha_{\rm r}) + 1.725\sin^2(\alpha_{\rm r})\cos(\alpha_{\rm r}), \qquad [9]$

390 $C_{\rm D}(\alpha_{\rm r}) = 0.0596 \sin(\alpha_{\rm r}) \cos(\alpha_{\rm r}) + 3.598 \sin^3(\alpha_{\rm r}),$ [10]

v is the relative airflow speed of the strip, c and dr are chord length and width of a strip, and the translational drag and lift unit vectors, $\hat{f}_{\rm L}$ and $\hat{f}_{\rm D}$, are calculated as follows

$$\hat{f}_{\mathrm{L}}^{\mathrm{w}} = rac{q^{\mathrm{w}}}{|q^{\mathrm{w}}|},$$
[11]

$$\hat{f}_{\rm D}^{\rm w} = \hat{\boldsymbol{v}}^{\rm w}, \qquad [12]$$

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$$\boldsymbol{q}^{\mathrm{w}} = \left(\hat{\boldsymbol{v}}^{\mathrm{w}} \cdot \hat{\boldsymbol{n}}^{\mathrm{w}}\right) \left(\left(\hat{\boldsymbol{v}}^{\mathrm{w}} \times \hat{\boldsymbol{n}}^{\mathrm{w}}\right) \times \hat{\boldsymbol{v}}^{\mathrm{w}}\right)$$
[13]

and \hat{n}^{w} is the unit vector normal to the plane of the strip in its dorsal direction (see Fig. 1F). It is imperative to note that $C_{\rm L}$, $C_{\rm D}$, v, c, $\hat{f}_{\rm L}$, $\hat{f}_{\rm D}$ and $\alpha_{\rm r}$ are functions of r. Their values vary for different blade element strips along the span of the wing. Moreover, our calculation of the unit vector $\hat{f}_{\rm L}$ was sufficient to keep track of the direction of the lift force vector, without invoking a sign from the lift coefficient $C_{\rm L}$ outside the range of the effective angle of attack from 0 to $\pi/2$ radians. In the wing-attached coordinate frame, the total translational aerodynamic force on a strip is

$$d\boldsymbol{f}_{\rm tra}^{\rm w} = d\boldsymbol{f}_{\rm L}^{\rm w} + d\boldsymbol{f}_{\rm D}^{\rm w}.$$
[14]

Rotational Aerodynamic Force. In addition to the aerodynamic force due to translation of the wing, we also incorporated the aerodynamic force due to its rotation with angular velocity $\dot{\alpha}$ about the y^{w} -axis (Fung, 1969). This force was assumed to be acting perpendicular to a blade element strip at a distance half-chord behind the leading edge. This was based on the experimental results from (Han et al., 2015) on a dynamically scaled hawkmoth wing. In the wing-attached coordinate frame, the rotational aerodynamic force on a small wing strip of width dr is

$$d\boldsymbol{f}_{\rm rot}^{\rm w} = \rho C_{\rm R} v c^2 \dot{\alpha} \, dr \begin{bmatrix} -\sin\alpha \\ 0 \\ -\cos\alpha \end{bmatrix}$$
[15]

where the rotational aerodynamic coefficient $C_{\rm R} = \pi \left(0.75 - \frac{e}{c}\right)$, e is the distance between the leading edge and wing pitching axis, α is the wing pitching angle and $\dot{\alpha}$ is the the angular velocity of the wing pitching rotation (Cheng et al., 2016a; Ellington, 1984b).

Force due to Added-mass. While the wing is undergoing translational and rotational accelerations during flapping, it experiences an inertial force to accelerate the boundary layer of air around the wing surface. Assuming the moth is flying at a constant velocity (on average), most significant contributions to this force come from the wing accelerations $\ddot{\phi}$ and $\ddot{\alpha}$, and the velocity product $\dot{\phi}\dot{\alpha}$ due to the force being measured in a ⁴¹⁸ non-inertial reference frame. This force acts perpendicular to a blade element strip at a distance half-chord ⁴¹⁹ behind the leading edge because the boundary layer is assumed to be uniformly distributed around a blade ⁴²⁰ element strip (Truong et al., 2011). In the wing-attached coordinate frame, the force due to added-mass on ⁴²¹ a small wing strip of width dr is given by the following equation (Maybury and Lehmann, 2004).

$$d\boldsymbol{f}_{adm}^{w} = \frac{1}{4}\pi\rho\left(\left(\ddot{\phi}\sin\alpha + \dot{\phi}\dot{\alpha}\cos\alpha\right)rc(r)^{2} + \frac{1}{4}\ddot{\alpha}c(r)^{3}\right)dr\begin{bmatrix}\sin\alpha\\0\\\cos\alpha\end{bmatrix},$$
[16]

423 where r is the distance of the wing strip from the wing hinge along the wing pitching axis.

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Sum of Force Components. We numerically integrated each of the translational, rotational and added-mass
aerodynamic force equations along the wing's spanwise length to evaluate the forces on the full right wing.
Then we evaluated the sum of these forces and moments on the right wing in its wing-attached coordinate
frame as follows

$$f_{\text{right}}^{\text{w}} = f_{\text{tra}}^{\text{w}} + f_{\text{rot}}^{\text{w}} + f_{\text{adm}}^{\text{w}}.$$
 [17]

Transformation to the Body-attached Frame. To explore how the aerodynamic forces act on the moth's body and affect its motion, we needed to transform the force vector calculated in the wing-attached frame to the body-attached frame. This was done in two steps. First, we transformed the instantaneous force vector from the wing-attached frame to the stroke-plane frame (through the wing kinematic angles ϕ and θ) as follows

$$\boldsymbol{f}_{\text{right}}^{\text{sp}} = \mathbf{R}_{z}(\phi) \mathbf{R}_{x}(\theta) \boldsymbol{f}_{\text{right}}^{\text{w}} , \qquad [18]$$

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$$\mathbf{R}_{x}(\theta) = \begin{bmatrix} 1 & 0 & 0 \\ 0 & \cos\theta & -\sin\theta \\ 0 & \sin\theta & \cos\theta \end{bmatrix}, \quad \mathbf{R}_{z}(\phi) = \begin{bmatrix} \cos\phi & -\sin\phi & 0 \\ \sin\phi & \cos\phi & 0 \\ 0 & 0 & 1 \end{bmatrix}$$
[19]

⁴³⁷ Second, we transformed from the stroke-plane frame to the body-attached frame (through the stroke-plane ⁴³⁸ angle β , given that there is no body roll rotation) through another rotation matrix

$$\boldsymbol{f}_{\mathrm{right}}^{\mathrm{b}} = \mathbf{R}_{y}(-\beta)\boldsymbol{f}_{\mathrm{right}}^{sp}$$
 [20]

440 where

$$\mathbf{R}_{y}(-\beta) = \begin{bmatrix} \cos\beta & 0 & -\sin\beta \\ 0 & 1 & 0 \\ \sin\beta & 0 & \cos\beta \end{bmatrix}, \quad \boldsymbol{l}_{1}^{\mathrm{b}} = l_{1} \begin{bmatrix} \cos\chi_{e} \\ 0 \\ -\sin\chi_{e} \end{bmatrix}, \quad \chi_{e} = \chi + \chi_{1}$$

The angle χ_e is the inclination angle of the wing hinge from the center of mass with respect to the horizontal plane, and is equal to the sum of body angle χ and the angle χ_1 as shown in Fig. 1H. The overall transformation from the wing-attached frame to the body-attached frame can also be represented as a single transformation matrix \mathbf{R}_{w}^{b} ,

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$$\boldsymbol{f}_{\mathrm{right}}^{\mathrm{b}} = \mathbf{R}_{\mathrm{w}}^{\mathrm{b}} \boldsymbol{f}_{\mathrm{right}}^{\mathrm{w}} ,$$
 [21]

447 where $\mathbf{R}_{w}^{b} = \mathbf{R}_{y}(-\beta)\mathbf{R}_{z}(\phi)\mathbf{R}_{x}(\theta).$

Induced power calculation. The induced power requirement for flight were also calculated for the two representative moth species. Induced power is the power required to impart sufficient momentum to the surrounding fluid in order to offset body weight as well as overcome the induced drag on the wing (Ellington, 1984c). The induced aerodynamic power calculation (Table A.1) was conducted following the method outlined in (Willmott and Ellington, 1997b). Additional details on induced power can be found in the Induced Power Calculation section.

454 **RESULTS**

Phylogeny. Phylogenetic relationships of the 57 bombycoid species used in this study (Fig. 2a; S1A) show similar relationships to those published previously (Breinholt et al., 2018; Hamilton et al., 2019; Barber et al., 2015; Kawahara and Barber, 2015; Rubin et al., 2018). We recovered a monophyletic clade (sister to Bombycidae) of the Sphinigidae and Saturniidae sister-families with a strong branch support values across the tree (Fig. S1A).

Hawkmoths and silkmoths each have diverse, but clustered forewing shapes in morphospace. The 460 forewing shapes of hawkmoths and silkmoths are well separated in morphospace. Most of the varia-461 tion (68%) in forewing shape is explained by the first two pPC axes (44% by pPC1 and 24% by pPC2; 462 Fig. 2B;). pPC three and four explain 14% and 8% of the total variation, respectively (Fig. S2A;). All 463 remaining pPCs each explained less than five percent of shape variation and a total of 10% of the variation. 464 Projections into morphospace reveal that pPC1 generally corresponds to AR, where a low pPC1 value 465 corresponds to a higher ratio between forewing length and width. High values of pPC2 corresponding to 466 large rounded distal tips and low values of pPC2 corresponding to a narrower distal wing tip. 467

We found significant separation in morphospace using a MANOVA where scores of 4 pPCs were the response variables and clade (hawkmoth, silkmoth, ancient bombycoid lineages) was the factor (F = 14.912, $p = 2.33 \times 10^{-14}$). A second MANOVA that only included the hawkmoth and silkmoth clades (instead of all three groups) also reveals significant separation between these two clades (F = 44.42, $p = 6.612 \times 10^{-15}$), demonstrating that the differences are reflected in the split between hawkmoths and silkmoths. Additionally,

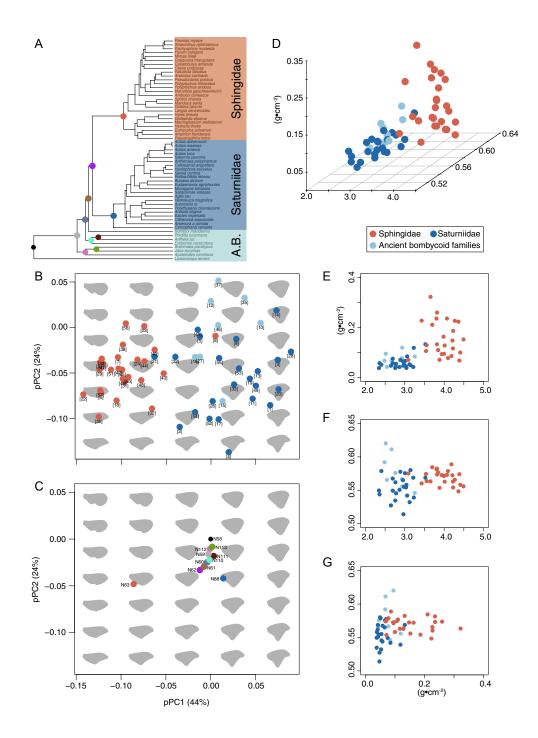


Fig. 2. The evolution and trajectory of forewing shape diversity. (A) The reconstruction of the phylogenetic relationships in bombycoids and outgroups (node labels in Fig. S1B). A.B. refers to Ancient Bombycoid lineages, which do not belong to either the Saturniidae or Sphingidae clades. Ancestral nodes are color coded for plotting in C. (B) Projections of the taxa onto the first two phylogenetic principal components demonstrates the separation of extant hawkmoths and silkmoths. The complete morphospace includes pPC 3 and 4 (Fig. S2A). The species-number key can be found in Fig. S2B (C) The ancestral state reconstruction of pPC1 and pPC2 scores reveals that forewing shape was conserved throughout the early evolutionary history of the Bombycoid superfamily until the ancestral hawkmoth (Node 63) and silkmoth (Node 88) rapidly diverged. (D) Species also cluster along aspect ratio (AR), wing loading (W_s), and radius of the second moment of area (\hat{r}_2). (E-G) Bivariate projections of panel D.

hawkmoths and silkmoths have significant separation along pPC1 (ANOVA, F = 131.94, $p = 3.035 \times 10^{-15}$), pPC3 (ANOVA, F = 27.48, $p = 3.688 \times 10^{-6}$), and pPC4 (ANOVA, F = 99.63, $p = 3.4 \times 10^{-13}$). Finally, both the hawkmoth and silkmoth clades have significantly distinct wing shapes from those of ancient bombycoids in 4D morphospace (MANOVA, Hawkmoths: F = 38.188, $p = 3.65 \times 10^{-11}$; Silkmoths: F =3.3102, p = 0.02555).

We next tested if the hawkmoth and silkmoth clades have wing shape differences once accounting for phylogeny. Across the entire bombycoid phylogeny, wing shape along each pPC has significant phylogenetic signal (Table S2). This indicates that shape varies in some significant way across the phylogeny, but does not indicate where. Scores of pPC1, 3, and 4 have significant phylogenetic signal across a tree pruned to include both hawkmoths and silkmoths (Table S2). However, no phylogenetic signal was found when the tree is pruned to include only either the hawkmoths or the silkmoths (Table S2), indicating that forewing shape has significantly diverged at the node between the two sister families.

Distinct evolutionary trajectories of forewing diversification. We next reconstructed the ancestral trajec-485 tory of wing shape across the phylogeny, focusing on the differences that occur around the split between 486 hawkmoths and silkmoths. Consistent with the phylogenetic signal test, the ancestral state reconstruction 487 reveals that forewing shape was conserved (little variation in pPC score) until a distinct divergence at the 488 nodes where hawkmoths and silkmoths split (Fig. 2C; Table S3). The range of pPC1 and pPC2 scores of 489 the ancient nodes (nodes highlighted in 2 A, C) represent only 6.8% and 17.5% of the total variation in 490 pPC scores of extant bombycoids (Fig. 2C) and are consistent with most of the shapes observed in the 491 long branched "ancient bombycoid" families (Fig. 2B). The ancestral nodes of the hawkmoth (node 63) 492 and silkmoth (node 88) sister families diverged from this cluster, indicated by minimal overlap in the 95%493 confidence interval of the state of these two nodes (Table S3). From these two sister nodes, each group 494 speciated and underwent subsequent diversification. However, while individual hawkmoth or silkmoth 495 species might have somewhat more convergent or divergent wing shapes, the two groups maintain distinct 496 differences, especially in pPC1. 497

Aerodynamic features of the wing and body also separate between clades. The pPCA is a data-driven analysis of overall differences in shape. In order to relate variation in forewing shape to metrics classically used in aerodynamics, we also quantified several specific forewing and body shape variables: wing length (R), wing mean chord length (\bar{c}) , wing area (S), the nondimensional radius of second moment of wing area (\hat{r}_2) , wing aspect ratio (AR), wing loading (W_s) , body length (l_b) , abdomen length (l_{abd}) , the fraction of body length occupied by the abdomen (\hat{l}_{abd}) and thorax (\hat{l}_{tho}) , respectively, and an estimate of total body mass (m_t) (Summary data: Table 1). Before accounting for phylogeny, significant differences were found between hawkmoths and silkmoths for each trait other than m_t (Table 1). Notably, R, \bar{c} , and S are significantly greater in silkmoths than in hawkmoths, while AR, W_s , and \hat{r}_2 are all significantly greater in hawkmoths than in silkmoths. While variation in l_b spans similar ranges within each family, clade average l_b and \hat{l}_{abd} are significantly longer in hawkmoths than silkmoths (Table 1), and \hat{l}_{tho} is generally greater in silkmoths than in hawkmoths, (Table 1).

In order to determine if these morphological variables also diverged between hawkmoths and silkmoths, 510 we again measured phylogenetic signal for each trait. As with the pPCs, significant phylogenetic signal is 511 present when silkmoths and hawkmoths are combined, but not when the tree is pruned to consider each 512 family individually (Table 1; S2). The only exception is W_s , which shows significant phylogenetic signal 513 within only the hawkmoths. The lack of phylogenetic signal within the families means that the primary 514 pattern of divergence occurs at the split of the two clades. However, given the limitations of phylogenetic 515 sampling, specific individual hawkmoth or silkmoth taxa might still significantly diverge from the rest of 516 the larger families. 517

⁵¹⁸ We also reconstructed the ancestral states of the three most commonly used wing shape metrics: AR, ⁵¹⁹ W_s , and \hat{r}_2 (Fig. 3A-C). Each trait was conserved with large confidence intervals at the rootward nodes ⁵²⁰ (Fig. 3; Table S3). However, at the nodes representing the ancestors of the hawkmoth and silkmoth clades, ⁵²¹ significant patterns of divergence are found for AR (no overlap in confidence intervals; Table S3) and W_s ⁵²² (minimal overlap in confidence intervals; Table S3). Taken together both the data driven pPC morphospace ⁵²³ and the specific measures of forewing shape most related to aerodynamics are first conserved and diverge ⁵²⁴ precisely at the split of hawkmoths and silkmoths.

Wing beat frequency, but not inertial power, diverges between hawkmoths and silkmoths. Wing beat 525 frequency and the power required to accelerate the mass of the wings each wingstroke (inertial power – 526 P_{acc}) are both important values that depend on wing size. Lacking wing kinematics for all the museum 527 specimens, n was estimated from scaling relationships (Table A.1; (Deakin, 2010)). In our analysis, n528 is distinct from wing shape, but not independent of wing size variation. The P_{acc} calculation relies on 529 variables of m_t and m_w , which were estimated from museum specimens by assuming the body has a density 530 equal to water (Ellington, 1984a). Mean n is significantly greater in hawkmoths (mean \pm SD: 33.92 \pm 11.23 531 Hz) compared to silkmoths (mean \pm SD: 17.47 \pm 5.56 Hz; p < 0.0001), but there is no significant difference 532 in P_{acc} between the hawkmoth and silkmoth clades (hawkmoths: $48.65 \pm 12.97 \text{ w kg}^{-1}$, silkmoths: $41.57 \pm$ 533 8.50 w kg⁻¹; p > 0.05; Table 1). n diverges between hawkmoths and silkmoths early in the evolutionary 534 history of bombycoids (Fig. 3E). n at the ancestral nodes for the hawkmoth and silkmoth clades are 535 estimated to be 34.28 Hz (95% confidence interval: 27.10 - 41.47) and 22.88 Hz (95% confidence interval: 536 14.35 – 31.41), respectively. Prior to that split, the ancestral state of P_{acc} is equivocal at rootward nodes 537

	A	verage ± stde	9V		T-test	
	Ancient bombycoids	Silkmoths	Hawkmoths	р (АВ-S)	р (АВ-Н)	р (S-H)
<i>R</i> (m)	0.0349 ± 0.017151	0.0516 ± 0.01638	0.04197 ± 0.0150	0.02012	0.26813	0.03706
<i>ī</i> (m)	0.0128 ± 0.006359	0.0179 ± 0.0056	0.01058 ± 0.0036	0.04038	0.22018	<0.0001
<i>S</i> (m²)	0.0005 ± 0.000524	0.0010 ± 0.00058	0.00049 ± 0.00033	0.05346	0.77188	0.00034
\hat{r}_2	0.5792 ± 0.026389	0.5539 ± 0.01769	0.56927 ± 0.01034	0.00478	0.12092	0.00046
AR	2.7600 ± 0.265587	2.88959 ± 0.28949	3.96152 ± 0.337358	0.27537	<0.0001	<0.0001
l_b (m)	0.0272 ± 0.010591	0.02852 ± 0.007602	0.03699 ± 0.010579	0.69850	0.02837	0.00260
m_t (g)	0.7855 ± 0.78914	1.19349 ± 0.83900	1.59744 ± 1.419522	0.23925	0.13448	0.23920
<i>n</i> (Hz)	26.5094 ± 9.630467	17.4708 ± 5.56408	33.9163 ± 11.22714	0.00291	0.10248	<0.0001
<i>W</i> s (g∙cm ⁻²)	0.0749 ± 0.025196	0.05971 ± 0.024333	0.15295 ± 0.069721	0.14164	0.00427	<0.0001
<i>l_{tho}</i>	0.3008 ± 0.059889	0.32948 ± 0.032573	0.29120 ± 0.029230	0.09752	0.53970	<0.0001
<i>l_{abd}</i>	0.6351 ± 0.06688	0.61704 ± 0.04127	0.64521 ± 0.035400	0.37452	0.57503	0.01334
l_{abd} (m)	0.0175 ± 0.007939	0.01771 ± 0.00534	0.023979 ± 0.007354	0.94196	0.04087	0.00150
pPC1	0.0114 ± 0.0244	0.0222 ± 0.0412	-0.0961 ± 0.0306	0.493	<0.0001	<0.0001
pPC2	-0.0031 ± 0.043370	-0.0563 ± 0.03847	-0.0465 ± 0.025442	0.002	0.001	0.2910
pPC3	-0.0004 ± 0.014623	0.0016 ± 0.02405	-0.0366 ± 0.026721	0.815	0.001	<0.0001
pPC4	-0.0050 ± 0.016029	-0.0181 ± 0.02798	0.0452 ± 0.015287	0.224	<0.0001	<0.0001
P _{acc}	33.0317 ± 11.13033	41.5706 ± 8.49548	48.6594 ± 12.97370	0.03143	0.00434	0.0306

Table 1. Clade averaged wing and body measurements with corresponding statistics. Adjusted alpha = 0.0027778

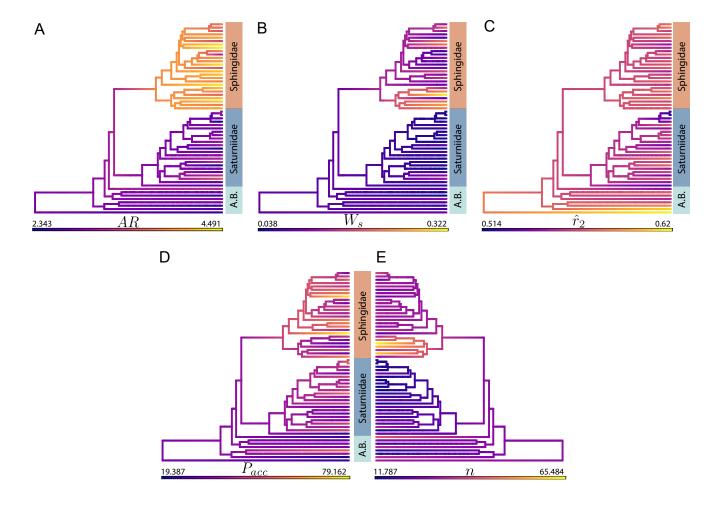


Fig. 3. Maximum likelihood ancestral state reconstructions of morphological variables. Ancestral state reconstructions of (A) aspect ratio (AR) and (B) wing loading (W_s) diverge between the hawkmoth and silkmoth sister clades. The reconstruction of (C) radius of second moment of wing area (\hat{r}_2) is more conserved across bombycoids, but is generally shifted towards higher values in hawkmoths than silkmoths. Finally, (D) inertial power (P_{acc}) is conserved across the superfamily with no divergence between hawkmoths and silkmoths. The reconstruction of (D) wing beat frequency (n) also reveals strong divergence between the hawkmoth and silkmoth clades. The hawkmoth subfamily, Macroglossinae (Node 82), is a notable example of particularly high n.

(Fig. 3D; Table S3), and remains generally conserved over time. Finally, significant phylogenetic signal is found in n over the tree pruned to include only silkmoths and hawkmoths, and is also found when the tree is pruned to only include either silkmoths or hawkmoths (Table 1, S2). However, no phylogenetic signal is found for P_{acc} (Table S2).

Three-dimensional body and forewing kinematics are different between the exemplar hawkmoth and silkmoth. To determine the impact of wing movement on aerodynamic force production, we chose a species of hawkmoth (*E. achemon taxa #29* in Fig. 2B) and silkmoth (*A. luna – taxa #3* in Fig. 2B) that represent the more divergent regions of morphospace (Fig. 4A, B; Fig. 2B). Three individuals from each species were recorded conducting steady forward flight (Fig. 4C, D). *A. luna* had a lower *n*, more steeply inclined stroke plane, β , and larger wingstroke sweep, ϕ , and deviation, θ , amplitudes (Table 2; Fig. 4C, D). In comparison, *E. achemon* utilizes a greater $\overline{\phi}$ and α amplitude throughout a wing stroke (Fig. 4C, D).

Wingstroke averaged forces are comparable between species. Given that the A. luna and E. achemon 549 use their wings in different ways in addition to having different wing shapes, we next assessed the overall 550 aerodynamic force and power implications between the two species. Despite an interspecific difference in 551 wing area of a factor of 3.12, the peak and wingstroke averaged body centered forces are similar in magnitude 552 between species (Table 3). The within wingstroke force profiles do show different patterns, especially in f_x^b 553 during the first half of wing stroke and f_z^b during the second half of wing stroke; in both cases, these forces 554 are predominately positive in A. luna and negative in E. achemon (Fig. 5a). Although the magnitude of 555 rotational (f_{rot}^b) and added mass (f_{adm}^b) forces are generally larger in A. luna in comparison to E. achemon 556 (Fig. 6A), these forces tend to act in opposition, and interspecific differences in the total body forces (f_{tot}^b) 557 are primarily due to translational force, f_{trans}^b (Fig. 6A). To assess if flight speed might contribute to 558 difference in aerodynamics, we conducted an additional model where the horizontal and vertical velocities 559 are set to the species average recorded during free flight. Doing so resulted in the same patterns of force 560 (Fig. S3A-B). 561

Actias luna and Eumorpha achemon have similar induced aerodynamic power requirements. While *E. achemon* and *A. luna* have nearly the same magnitude wing stroke averaged f_{tot}^b , the *n* of *E. achemon* is approximately double that of *A. luna* (*E.a* = 31.50; *A.l* = 14.20). Therefore, when comparing wing-stroke averaged force production per unit time, *E. achemon* produces approximately twice the force in a given amount of time in comparison to *A. luna*. These interspecific differences in force production per unit time are paralleled by an interspecific difference in the induced power (P_{ind}) requirement. The P_{ind} requirement of *E. achemon* is 13.758 W kg⁻¹, while the P_{ind} requirement of *A. luna* is 5.52 W kg⁻¹ (Table 3).

			Measure	ed wing kinema	atics			
		Eumorpho	a achemon			Actia	s luna	
	<i>E.a</i> . 1	E.a. 2	E.a. 3	Mean ± stdev.	<i>A.I.</i> 1	A.I. 2	A.I. 3	Mean ± stdev.
<i>n</i> (Hz)	31.75	33.33	29.41	31.50 ± 1.97	15.38	13.33	13.89	14.20 ± 1.06
eta (deg.)	42.04	22.94	36.93	33.97 ± 9.89	88.81	74.70	64.14	75.88 ± 12.37
χ_{p-p} (deg.)	3.95	9.91	4.05	5.97 ± 3.41	30.02	25.80	22.32	26.05 ± 3.86
$\overline{\chi}$ (deg.)	26.83	36.35	34.48	32.55 ± 5.04	12.57	27.76	23.14	23.14 ± 9.18
ϕ_{p-p} (deg.)	115.34	96.39	120.64	109.51 ± 12.74	129.01	132.55	135.78	127.80 ± 3.38
$ar{\phi}$ (deg.)	24.03	31.82	6.28	19.87 ± 13.14	9.22	10.76	15.67	11.03 ± 3.42
ϕ_{min} (deg.)	-30.95	-15.55	-49.32	-31.70 ± 16.91	-52.72	-51.60	-50.86	-51.44 ± 0.94
ϕ_{max} (deg.)	84.38	80.84	71.31	77.81 ± 6.76	76.29	80.95	84.92	76.36 ± 4.32
$ heta_{p-p}$ (deg.)	6.23	9.04	22.24	11.41 ± 8.55	17.41	22.84	22.81	19.64 ± 3.12
$ar{ heta}$ (deg.)	-2.71	-0.95	1.90	-0.56 ± 2.36	2.98	-3.20	3.81	1.18 ± 3.89
$ heta_{min}$ (deg.)	-4.92	-5.05	-6.47	-4.71 ± 0.86	-6.63	-18.86	-8.03	-10.52 ± 6.69
$ heta_{max}$ (deg.)	1.27	3.99	15.77	6.70 ± 6.29	10.78	3.89	14.78	9.13 ± 5.46
$lpha_{p-p}$ (deg.)	70.61	92.09	90.11	80.91 ± 11.87	53.00	49.98	54.87	45.08 ± 2.47
\overline{lpha} (deg.)	77.08	87.67	88.74	84.56 ± 6.43	88.37	76.85	81.44	82.14 ± 5.82
$lpha_{min}$ (deg.)	44.12	54.43	47.04	50.14 ± 5.31	60.43	51.77	56.53	58.49 ± 4.34
$lpha_{max}$ (deg.)	115.46	146.41	136.78	133.02 ± 15.84	112.76	102.18	108.66	101.54 ±

Fourier coefficients fit to species mean wing kinematics

		E. ach	nemon	A. I	una
Kinematic variables	k	a	b	a	b
	0	0.4006	0	0.2165	0
+ (+)	1	0.9480	-0.0711	1.0910	-0.0309
$\phi(t)$	2	0.0074	-0.0839	0	0
	3	0	0	0	0
	0	-0.0133	0	0.022	0
0(4)	1	-0.0168	-0.0742	-0.0025	-0.1448
$\theta(t)$	2	-0.0403	-0.0064	0.0337	0.0102
	3	-0.0082	0.0152	-0.0037	0.0261
	0	1.4723	0	1.4360	0
~ (t)	1	-0.0525	-0.6877	0.0022	-0.3856
$\alpha(t)$	2	-1.597	-0.0620	0.0419	0.0101
	3	0	0	0.0059	-0.0123

Table 2. Summary of comparative wing kinematics for the representative hawkmoth and silkmoth species.

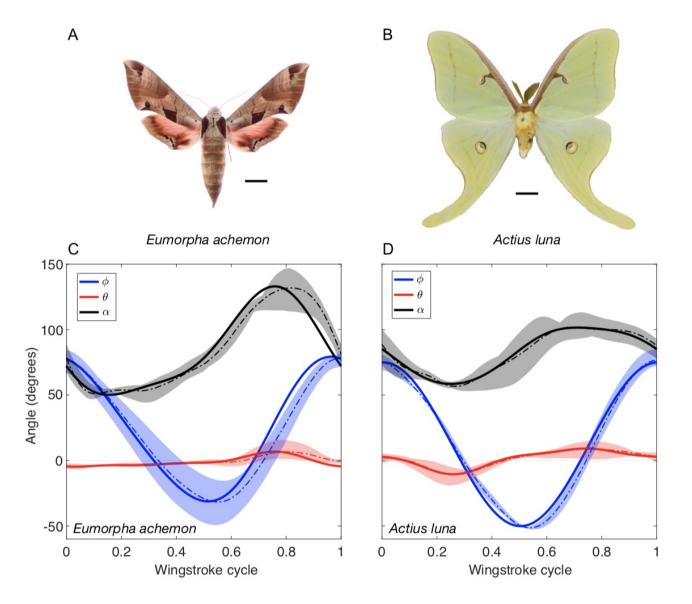


Fig. 4. Three-dimensional wing kinematics of a representative hawkmoth and silkmoth. *E. achemon* (A) *A. luna* (B) were flown in a wind tunnel to obtain three-dimensional kinematics (C and D). Scale bars are equal to 1 centimeter. For both species, we measured sweep (ϕ , blue), deviation (θ , red), and morphological feathering (α , black) angles. The shaded region of each curve represents the range of values recorded, the dashed lines represent the species mean, and the solid line represents the third order Fourier fit, used in all aerodynamic models. N = 3 individuals per species and one wing stroke per individual. The mean *n* is 14.2 in *A. luna* and 31.5 in *E. achemon* (Table 2).

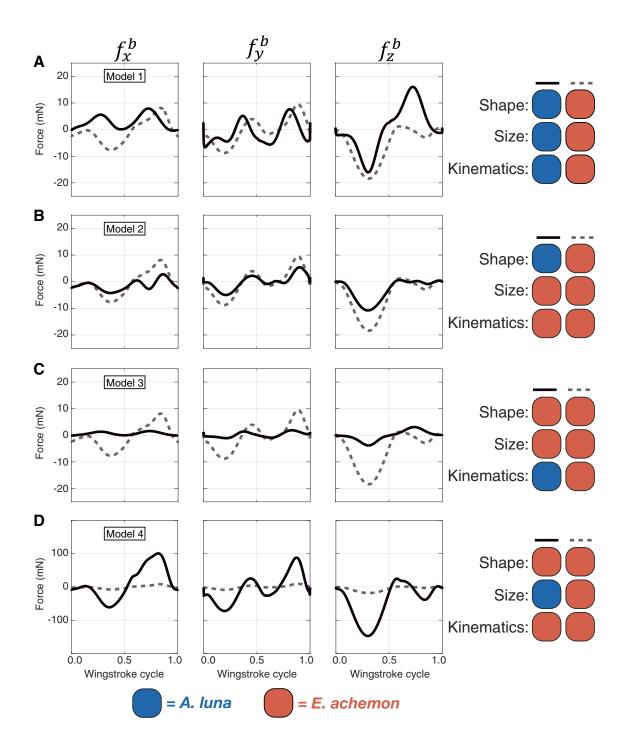


Fig. 5. Quasi-steady aerodynamic force production by the right wing in the body-centered coordinate system. Column one, two, and three display the f_x^b , f_y^b , and f_z^b , respectively. The two wings of each model are identified by solid and dashed lines, respectively. The dashed line is the same in each model. The key on the right side of each model can be used to determine the specific wing shape and movement parameters used for each wing. Red represents variables from *A. luna* and blue represents variables from *E. achemon*. (A) Model 1 compares interspecific aerodynamics between *A. luna* (solid line) and *E. achemon* (dashed line). Models 2 (B), 3 (C), and 4 (D) investigate how aerodynamics are impacted by changes in wing shape, movement, and size, respectively. All forces are only presented for a single right wing. The negative f_z^b direction points upward and the positive f_x^b direction points forward in the coordinate system.

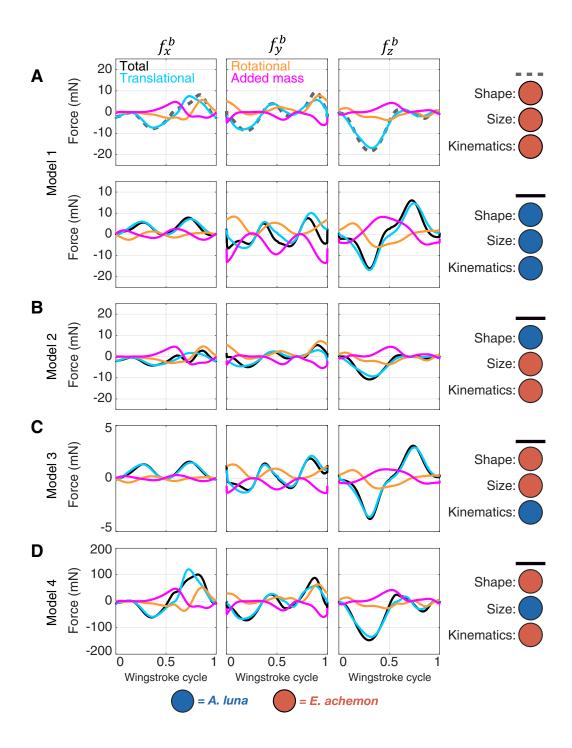


Fig. 6. Translational, rotational, and added mass components of aerodynamics force. Details of the four models are identical to those in Figure 5. Color schemes for component are the same for both species. Black represents the total force, cyan represents the translational force component (f_{trans}^b) , gold represents the rotational force component (f_{rot}^b) , and pink represents the added mass force component (f_{adm}^b) . Column one, two, and three display the f_x^b , f_y^b , and f_z^b , respectively. All forces are only presented for a single right wing. In all four models for each species, f_{trans}^b drives the majority of the pattern in total force throughout the wing stroke.

			f_{tot}^{b} (N)			f_{trans}^{b} (N)			f_{rot}^{b} (N)			f_{adm}^{b} (N)		Induced Power
Model	Species	x	у	Ζ	х	у	Ζ	х	у	Ζ	х	у	Ζ	(W/kg)
1	E. a.	-0.545	-0.282	-5.714	-0.692	-0.606	-5.513	-0.243	1.974	-1.162	0.39	-1.65	0.96	13.758
1	A. I.	2.915	-1.023	0.402	3.097	0.771	-0.47	-0.244	3.421	-0.422	0.062	-5.216	1.294	5.52
2	E. a.	-0.545	-0.282	-5.714	-0.692	-0.606	-5.513	-0.243	1.974	-1.162	0.39	-1.65	0.96	13.758
2	A. I.	-1.342	-0.022	-3.022	-1.192	-0.394	-3.099	-0.269	1.891	-1.088	0.119	-1.519	1.165	7.747
2	E. a.	-0.545	-0.282	-5.714	-0.692	-0.606	-5.513	-0.243	1.974	-1.162	0.39	-1.65	0.96	13.758
3	A. I.	0.648	0.193	-0.093	0.66	0.212	-0.15	-0.039	0.546	-0.075	0.027	-0.564	0.132	8.962
4	E. a.	-0.545	-0.282	-5.714	-0.692	-0.606	-5.513	-0.243	1.974	-1.162	0.39	-1.65	0.96	13.758
4	A. I.	12.085	-4.914	-43.778	9.627	-3.588	-44.082	-1.351	14.777	-9.07	3.809	-16.103	9.374	11.629

Table 3. Wing stroke averaged total and component forces produced by each species across models.

Aerodynamic force production is impacted by interspecific differences in wing size, shape, and move-569 ment. To assess how size, shape and kinematics impact the aerodynamics, we created several intermediate 570 models that separate their contributions. The base comparison reported above (Model 1 – Figs. 5A, 6A) 571 uses each species' own wing shape, size, kinematics (ϕ , θ , and α), n, and β , with forward flight velocities 572 set to 2 m/s. In Model 2 (Figs. 5B, 6B) we set wing kinematics and size to be that of *E. achemon* in both 573 cases, leaving interspecific differences only in wing shape. In Model 3 (Figs. 5C, 6C), we set wing shape 574 and size to be that of *E. achemon*, leaving interspecific differences only in wing kinematics. Finally, in 575 Model 4 (Figs. 5D, 6D), we set wing shape and kinematics to be that of *E. achemon*, leaving interspecific 576 differences only in wing size. 577

Wing shape: The E. achemon wing shape produces larger net aerodynamic forces than A. luna shaped 578 wings (Figs. 5B; 6B). The primary determinant of this increase in f_{tot}^b is the greater f_{trans}^b (Fig. 6B). While 579 different in average and peak magnitude, the pattern of the f_{trans}^{b} (as well as the rotational and added mass 580 forces) throughout the wing stroke was generally similar in both wing shape cases (Fig 6B; Table 3). The 581 small interspecific differences that exist in the shape of f_x^b and f_z^b during the second half of wing stroke (Fig. 582 5B) occur because the magnitude of the translational force component of A. luna shaped wings is decreased 583 relative to the magnitude of the added mass and rotational force components (Fig. 6B). The increase in 584 the relative contribution of $f_{\rm rot}^b$ and $f_{\rm adm}^b$ in A. luna is responsible for the interspecific differences in the 585 shape of f_x^b and f_z^b during the second half of wing stroke (Figs. 5B, 6B). 586

⁵⁸⁷ Wing kinematics:. The most apparent interspecific difference in aerodynamics due to kinematics alone (Model ⁵⁸⁸ 3) is that A. luna produces much lower overall forces than E. achemon (Fig. 5C). The reduction in total ⁵⁸⁹ body forces is again determined primarily by differences in translational force (Fig. 6C). The main cause of ⁵⁹⁰ this difference is that n, and hence wing velocity, of E. achemon is 2.2 times greater than A. luna (Table 2. ⁵⁹¹ Interspecific differences in kinematics are also responsible for interspecific differences in the sign of f_x^b and ⁵⁹² f_z^b in Model 1 during the first half and second half of wing stroke, respectively (Fig. 5A). To break this down further, we separated the contributions of stroke plane angle (β), wing angles (ϕ , θ , and α) and n(Fig. S3). The interspecific sign flip in f_z^b (Fig. 5A) occurs due to a combination of stroke plane angle and angle of attack (Fig. S3A), and the f_x^b sign flip (Fig. 5A) occurs primarily from the more vertical stroke plane (larger β) of A. luna (Fig. S3C).

Wing size: A. luna has larger wings and, if all other variables are equal (Model 4), it is not surprising that 597 A. luna produces larger overall forces (Figs. 5D, 6D). As before, differences in total force are primarily due 598 to differences in the translational force (Fig. 6D). However, the magnitude of the rotational and added 599 mass components of f_x^b , f_y^b , and f_z^b are also nearly an order of magnitude larger in A. luna sized wings 600 compared to E. achemon (Fig. 6D). Overall, wing size has the predictable effect of scaling all of the 601 force components. Taken together, the three knobs (shape, kinematics, and size) that evolution can tinker 602 with means that even though the wing shapes are quite distinct between hawkmoths and silkmoths, these 603 differences converge on similar overall wingstroke averaged forces (but different within wing stroke patterns) 604 from different combinations of morphology and kinematics. 605

606 **DISCUSSION**

Wing shape can have a strong influence on the aerodynamics and maneuverability of flying animals. The first goal of this study was to test if hawkmoths and silkmoths diverged signifcantly in wing shape across the moth superfamily Bombycoidea. We find that early in the evolutionary history of Bombycoidea, wing shape was generally conserved until the ancestors of the hawkmoth and silkmoth sister clades rapidly separated in morphospace (Fig. 2C). The evolutionary split between these two families has been dated to occur between 57 and 75 MYA (Kawahara et al., 2019), suggesting that these wing shape trajectories have been evolving since then.

The distinct trajectory in wing shape change between hawkmoths and silkmoths was followed by a 614 subsequent shape diversification within each group. Even specific species within each group that subsequently 615 converged in life history did not fully converge to employ overlapping wing shapes. For example, while the 616 majority of adult hawkmoths are known for their hovering nectaring behavior, members of the hawkmoth 617 subfamily, Smerinthinae (Node 67; Figs. S1B, 2A), have lost the ability to feed as adults (Tuskes et al., 618 1996), convergent with all species of silkmoth. However, the wing shape of Smerinthinae species remains 619 divergent with silkmoths with the exception of Andriasa contraria (Taxon 8). While we chose species 620 to broadly cover the groups within bombycoids, sampling is far from complete. Therefore, we remain 621 conservative in our interpretation, focusing on the split between hawkmoths and silkmoths for which we 622 were able to accumulate broad sampling for our analysis. In sum, these data provide phylogenetic evidence 623 supporting our hypothesis that there have been distinct wing shape trajectories in these two groups of 624

625 bombycoids.

Sillkmoth wing shapes are more favorable to maneuverability than hawkmoths. The evolution of forewing 626 shape does not support hypotheses based on the presumed maneuverability of hawkmoths. We find that 627 hawkmoths employ wings of high AR, W_s , and \hat{r}_2 ; all metrics typically associated with low degrees of 628 maneuverability. In contrast, we find that silkmoths are employing wings of low AR, W_s , and \hat{r}_2 ; all metrics 629 typically associated with higher degrees of maneuverability. However, these relationships between wing 630 shape and maneuverability are typically used in fixed wing aircraft theory or when the kinematics of the 631 flapping wing are held constant. In species employing flapping flight, wing shape and movement could 632 undergo correlated evolution to produce species-specific performance metrics that could not be predicted by 633 either wing shape or movement alone. 634

⁶⁰⁵ The high aspect ratio wings of hawkmoths and larger wing stroke amplitude of silkmoths lead to similar

induced power requirements. In comparison to silkmoths, hawkmoths evolved high AR wings (Figs. 2, 636 3; Table 1) which might reduce maneuverability; however, high AR also reduce the induced power (P_{ind}) 637 requirements of flight (Norberg and Rayner, 1987; Pennycuick, 1968). P_{ind} is the power required to overcome 638 induced drag on the wing and impart sufficient downwards momentum to the surrounding air to offset body 639 weight. The reduced maneuverability of high AR wings is due to an increase in the moments of inertia of 640 the wing pair (Betts and Wootton, 1988), but the hawkmoth AR increase is concomitant with a reduction 641 in wing area. Hawkmoths have evolved high AR by reducing mean chord length, \bar{c} , rather than through 642 an increase in wing span, R (Table 1; Fig 2B). Therefore, while selection for economical flight (increased 643 AR) might often reduce maneuverability, the evolution of short, high AR wings in hawkmoths (achieved 644 through a reduction in \overline{c}) could act to increase economy while not necessarily sacrificing maneuverability. 645 The potential cost is that proportionally smaller wings could reduce wing stroke-averaged aerodynamic 646 force production, if wing movement remains constant. However, our aerodynamic model (Figs. 5A; 6A) 647 reveals that the changes in wing kinematics, and particularly the higher wingbeat frequencies of hawkmoths, 648 result in very similar wing stroke averaged forces between E. achemon and A. luna. 649

⁶⁵⁰ Despite employing lower AR wings, the representative silkmoth, *A. luna*, does not have a greater ⁶⁵¹ induced power (P_{ind}) requirement than the representative hawkmoth, *E. achemon*. The higher AR wings of ⁶⁵² hawkmoths leads to the prediction that hawkmoths would require less P_{ind} than silkmoths (Pennycuick, ⁶⁵³ 1968). However, P_{ind} required for flight is also inversely proportional to both *R* and wing stroke amplitude ⁶⁵⁴ (ϕ_{p-p}). The kinematics from our two representative species show that the silkmoth (*A. luna*) employs a ⁶⁵⁵ greater ϕ_{p-p} in comparison to the hawkmoth (*E. achemon*) (Fig. 4; Table 2). The net effect of AR, *R*, ⁶⁵⁶ and ϕ_{p-p} is that there is little difference in the P_{ind} requirement between these two species, with *A. luna* 657 actually being somewhat lower.

The employment of smaller, high AR wings by hawkmoths and the production of a large amplitude 658 wingstrokes by silkmoths are two different strategies that can act to reduce the $P_{\rm ind}$ requirements of flight. 659 Many hawkmoth species are known for their ability to sustain long duration bouts of hovering, often 660 associated with nectaring from flowers (Sprayberry and Daniel, 2007; Wasserthal, 1993, 1998), which require 661 a high-power output. In contrast, all silkmoths employ rudimentary mouth parts and do not feed as adults 662 (Janzen, 1984; Tuskes et al., 1996). Therefore, in hawkmoths, the selective pressure to reduce power while 663 not sacrificing maneuverability has likely driven the evolution of short, high AR wings used at a high n. In 664 silkmoths, the selective pressure to increase lifespan (endurance) has likely driven the evolution of large 665 wing beat amplitudes used at a low n. Kinematic differences (particularly n and ϕ_{p-p}) parallel the broader 666 clade specific wing shape differences in AR. Hawkmoths and silkmoths have therefore evolved different 667 strategies to reduce P_{ind} . 668

While P_{ind} is one of three components of the total aerodynamic power requirement, at slower flight speeds, 669 which were employed by our exemplar species, the majority of the total aerodynamic cost in Manduca 670 sexta, a species of hawkmoth, is from $P_{\rm ind}$ (Willmott and Ellington, 1997b). Inertial power requirements 671 are similar between our exemplar species and the similar body size between the exemplar species suggests 672 similar parasitic power requirements. The difference in wing area between our exemplar species does suggest. 673 however, that A. luna will incur greater profile power costs in comparison to E. achemon. However, we 674 suggest that the reduced P_{ind} of A. luna in comparison E. achemon ultimately leads to similar overall 675 aerodynamic power requirements between the two species. 676

Hig wing beat frequency might act to offset high wing loading (W_s) in hawkmoths. The lower W_s of 677 silkmoths suggests they are more maneuverable than hawkmoths, but, as before, this assumes all other 678 things are equal. The reduced W_s of silkmoths comes primarily from a larger S rather than smaller body 679 size compared to hawkmoths (Table 1). Higher W_s reduces mass-specific wing-stroke averaged forces. 680 However, many species use increased flight speeds or high n to offset a high W_s in terms of aerodynamic 681 force production (Ahmad, 1984; Byrne et al., 1988). Therefore, one selective pressure that could have led 682 to the evolution of a higher n in hawkmoths is the need to offset the greater magnitude of W_s relative to 683 silkmoths and other bombycoid families. 684

The greater radius of second moment of area (\hat{r}_2) of hawkmoths can augment force production. The high \hat{r}_2 of hawkmoth wings again suggests that silkmoths should be more maneuverable than hawkmoths (Le Roy et al., 2019; Ellington, 1984a). For a wing of uniform thickness and density, larger \hat{r}_2 means more mass is concentrated distally along the wingspan, corresponding to an increase in moments of inertia. In fixed wing aircrafts, larger wing moments of inertia hinder yaw and roll maneuvers (Etkin and Reid, 1996). However, in flapping or revolving wings, when all other things are equal, the larger \hat{r}_2 of hawkmoths would also increase their magnitude of torque production relative to silkmoths. The velocity of a wing section increases with its distance from the axis of rotation, and aerodynamic force production is proportional to velocity squared. Shifting more area distally (increasing \hat{r}_2) means more of the wing is moving at higher speeds enhancing production of aerodynamic forces and torques (e.g. Muijres et al. (2017); Fernandez et al. (2017)).

We can see this pattern when comparing aerodynamic force production between our representative hawkmoth (*E. achemon*) and silkmoth (*A. luna*) species. In Model 2, where wing area and all kinematic parameters are equal between species, the f_x^b , f_y^b , and f_z^b traces follow similar trajectories across the wing stroke and the magnitude of every force is greater in *E. achemon* shaped wings than in *A. luna* shaped wings (Fig. 5B; 6B).

Large, slow wings might produce bobbing flight in silkmoths. We find that the complex interaction 701 between wing shape, size, and kinematics are tightly linked to produce the within wingstroke aerodynamic 702 forces of A. luna and E. achemon (Figs. 5A, 6A), and might contribute to the differences in flight behavior 703 between the families. A. luna has more variation in forces during the wingstroke even though it produces 704 approximately the same average force as *E. achemon*. This is especially noticable in the switch from 705 negative f_z^b to positive during the wingstroke in A. luna. Large force fluctuations and asymmetry (Figs. 706 5A, 6A) should lead to greater fluctuation in body vertical velocity and are likely the source of the bobbing, 707 erratic motions that are characteristic of silkmoths and may be useful in predator avoidance. 708

⁷⁰⁹ Evolution of high wing beat frequency (*n*) enables hawkmoth maneuverability with wing shape to reduce

power. The evolution of n parallels the divergence of wing shape between hawkmoths and silkmoths. High 710 n may be the key to conducting high speed maneuvers in small flapping flyers like hawkmoths. Although 711 hawkmoths have not evolved forewing shapes thought to be advantageous for maneuverability, it is clear 712 that hawkmoths have evolved a means to accomplish rapid maneuvers while foraging (Wasserthal, 1993; 713 Sponberg et al., 2015; Stöckl et al., 2017a). As opposed to fixed-wing cases, maneuverability of flapping 714 flight relies on the generation of aerodynamic forces from wing movement to initiate directional change 715 (Warrick et al., 1998). Therefore, an increase in n would allow for more frequent modification of force vectors, 716 which could increase maneuverability. Further, increasing n will also enhance maneuverability by increasing 717 the force and torque produced by a wing of similar shape and area (Hedrick et al., 2009), which is also 718 exemplified in Model 3 of this study (Fig. 5C). The diversification of n could therefore contribute to 719 interspecific variation in flight control and maneuverability across species. Although n was inferred from 720

scaling in this study, the estimate closely matches the observed frequency of the species considered here that have a known n. To the best that we can assess, n has strongly diverged between hawkmoths and silkmoths, and mean n is nearly double for a hawkmoth in comparison to a silkmoth of similar body size (Table 1, 2). Therefore, we suggest that high n is one of the aspects of flight control that evolved in hawkmoths allowing for the completion of high frequency maneuvers while employing wings of high AR, W_s , and \hat{r}_2 .

Two strategies for agile flight. Silkmoths and hawkmoths have evolved two distinct strategies for agile 726 flapping flight. Species of both families have evolved mechanisms for maneuverability and power reduction, 727 but have distinctly different wing shapes. Hawkmoths achieve maneuverability through high n (movement) 728 and reduce power by employing wings of high AR (shape), while silkmoths achieve maneuverability 729 through favorable wing shapes and a reduction of power through the production of high amplitude wing 730 strokes. The evolutionary trajectories of forewing shape have diverged between the two families, but the 731 distinct flight strategies arise in part due to how differently they use their wings. The metrics of forewing 732 shape, historically derived for fixed-winged aircraft, are not consistent with initial expectations of flight 733 maneuverability based on the life history of species in each clade. Instead, we find that aerodynamic 734 performance emerges from the interaction of wing shape, size, and movement (kinematics), demonstrating 735 an example of parallel evolution between the components of a complex locomotor system (Aiello et al., 2017). 736 The ability for natural selection to act both on wing shape and movement to impact the power requirements 737 and maneuverability of an animal demonstrates the potential decoupling of animal locomotor performance 738 metrics. The employment of flapping flight therefore provides the flexibility to tune aerodynamics through 739 kinematics. Thus, we suggest the complex interplay between wing shape, size, and movement in resultant 740 performance reduces morphological constraints that would drive the convergent evolution of wings to meet 741 one or few advantageous shapes, leading to the diversity of wing shapes seen across extant aerial animals. 742

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754 APPENDIX

Symbol	Definition
A.B.	ancient bombycoids
$AR = R^2/S$	aspect ratio
$(a_{\phi,k},b_{\phi,k})$	Fourier series coefficients of Fourier fits of ϕ
$(a_{ heta,k},b_{ heta,k})$	Fourier series coefficients of Fourier fits of θ
$(a_{lpha,k},b_{lpha,k})$	Fourier series coefficients of Fourier fits of α
\hat{b}	trailing edge to leading edge unit vector
CD	aerodynamic coefficient of the drag force
$C_{ m L}$	aerodynamic coefficient of the lift force
$C_{ m R}$	coefficient of the rotational aerodynamic force
с	chord length
\overline{c}	Mean chordwise wing length
d	distance between the wing-attached y -axis (wing-pitching axis)
	and the quarter-chord line on the wing
dr	width of an infinitesimal blade element strip
e	distance between the leading edge and the wing-pitching axis

Table A.1. List of symbols in the alphabetical order.

	Continuation of Table A.1
Symbol	Definition
$egin{array}{c} egin{array}{c} f_x \ f_y \ f_z \end{array} \end{bmatrix}$	force vector
$f_{ m tra}$	translational aerodynamic force vector
$f_{ m D}$	drag component vector of the translational aerodynamic force
$\hat{f}_{ m D}$	drag component unit vector of the translational aerodynamic
	force
$f_{ m L}$	lift component vector of the translational aerodynamic force
$\hat{f}_{ m L}$	lift component unit vector of the translational aerodynamic force
$f_{ m rot}$	rotational aerodynamic force vector
$f_{ m adm}$	aerodynamic force vector due to the added mass
$f_{ m right}$	vector of the total aerodynamic force on right wing
h	distance between the wing-attached y -axis (wing-pitching axis)
	and the half-chord line on the wing
$\boldsymbol{I} = \begin{bmatrix} I_{xx} & -I_{xy} & -I_{xz} \\ -I_{xy} & I_{yy} & -I_{yz} \\ -I_{xz} & -I_{yz} & I_{zz} \end{bmatrix}$	body's moment of inertia tensor
l_1	position vector from body center of mass to the wing hinge point
$l_{ m b}$	body length
$l_{ m abd}$	abdomen length
$\hat{l}_{ m abd} = l_{ m abd}/l_{ m b}$	fraction of body length occupied by abdomen
$\hat{l}_{ m tho} = l_{ m tho}/l_{ m b}$	fraction of body length occupied by thorax
MYA	million years ago
$M_{ m tra}$	translational aerodynamic moment pseudovector
$M_{ m rot}$	rotational aerodynamic moment pseudovector
$M_{ m adm}$	aerodynamic moment pseudovector due to added-mass force
$M_{ m right}$	total aerodynamic moment pseudovector of right wing
$m_{ m b}$	body mass
$m_{ m w}$	wing mass
$m_{\rm t} = m_{\rm b} + 2m_{\rm w}$	Total (body+wings) mass
n	wingbeat frequency

	Continuation of Table A.1
Symbol	Definition
$n = 187 m_{\rm t}^{0.3} S^{-0.7}$	wingbeat frequency
\hat{n}	unit vector normal to the dorsal surface of the wing
$P_{\rm pro} = \frac{1}{T} \int_0^T \int_0^R \frac{1}{2} C_{\rm D} \rho c v^3 \ dr \ dt/m_{\rm t}$	body mass-specific mean profile power
$P_{\rm ind} = w \left(m_{\rm t} g - f_{{\rm D},z} \right) / m_{\rm t}$	body mass-specific mean induced power
$P_{\rm acc} = 8\pi^2 \phi_{\rm p-p}^2 n^2 R^2 \hat{r}_2 m_{\rm w}/m_{\rm t}$	body mass-specific mean inertial power
pPCn	$n^{\rm th}$ axis of the phylogenetic principal components
$\mathbf{R}_{z}(\phi)$	transformation matrix for rotation of ϕ radians about the $z\text{-axis}$
$\mathbf{R}_x(heta)$	transformation matrix for rotation of θ radians about the x-axis
$\mathbf{R}_{y}(eta)$	transformation matrix for rotation of β radians about the $y\text{-axis}$
$\mathbf{R}^{\mathrm{b}}_{\mathrm{w}}$	transformation matrix for rotating the coordinate system from
	wing-attached to body-attached frame
R	spanwise wing length
$r_{ m cm}$	position vector from the body center of mass to a blade element
	strip of the wing
$r_{ m wh}$	position vector from the wing hinge to a blade element strip of
	the wing
r	distance of a blade element wing strip from the wing hinge along
	the y^{w} axis
r_3	position vector from body center of mass to the quarter-chord
	line on a blade-element wing strip
$ r_1 $	position vector from body center of mass to the half-chord line
	on a blade-element wing strip
$\hat{r}_2 = \sqrt{\int_0^1 \hat{c} \hat{r}^2 d\hat{r}}$	nondimensional radius of second moment of area
S	wing area
T = 1/n	wingbeat time period
t	time variable during a wingstroke, where $t = 0$ corresponds to
	the start of the downstroke
v	relative airflow speed
v	relative airflow velocity vector
\hat{v}	relative airflow velocity unit vector
$v_{ m b}$	body linear velocity vector

	Continuation of Table A.1
Symbol	Definition
$W_{\rm s}=m_{\rm t}/S$	wing loading
w_0	induced airflow speed
$x^{\mathrm{b}}y^{\mathrm{b}}z^{\mathrm{b}}$	body-attached coordinate frame
$x^ly^lz^l$	body-long coordinate frame
$x^{\mathrm{sp}}y^{\mathrm{sp}}z^{\mathrm{sp}}$	stroke-plane coordinate frame
$x^{\mathbf{w}}y^{\mathbf{w}}z^{\mathbf{w}}$	wing-attached coordinate frame
$\hat{oldsymbol{y}}^{ ext{w}}$	unit vector along the wing-attached y -axis (wing-pitching axis)
α	wing pitching angle (feathering angle)
\dot{lpha}	wing pitching angular velocity
$\alpha_{ ext{p-p}}$	peak-to-peak amplitude of the feathering angle
$\overline{\alpha}$	mean feathering angle
$lpha_{ m e}$	effective angle of attack
$\alpha_{ m r}$	effective angle of attack bound between 0 and 90°
β	stroke-plane angle
θ	stroke deviation angle
$\dot{ heta}$	stroke deviation angular velocity
$\theta_{\mathrm{p-p}}$	peak-to-peak deviation angle
$\overline{ heta}$	mean deviation angle
ρ	density of air
ϕ	stroke positional angle (sweep angle)
$\dot{\phi}$	stroke positional angular velocity
$\phi_{\rm p-p}$	peak-to-peak amplitude of the stroke positional (sweep) angle
$\overline{\phi}$	mean sweep angle
χ	body angle
χ_1	angle of inclination of the wing hinge from the center of mass
	with respect to the body-long xy -plane
χ_e	angle of inclination of the wing hinge from the center of mass
	with respect to the horizontal plane
$oldsymbol{\omega}_{\mathrm{w}}$	wing angular velocity pseudovector due to wing kinematic motion
$\omega_{ m b}$	body angular velocity pseudovector

	Continuation of Table A.1
Symbol	Definition
Superscripts:	
b	measured with respect to the body-attached coordinate frame
l	measured with respect to the body-long frame
sp	measured with respect to the stroke-plane frame
w	measured with respect to the wing-attached coordinate frame

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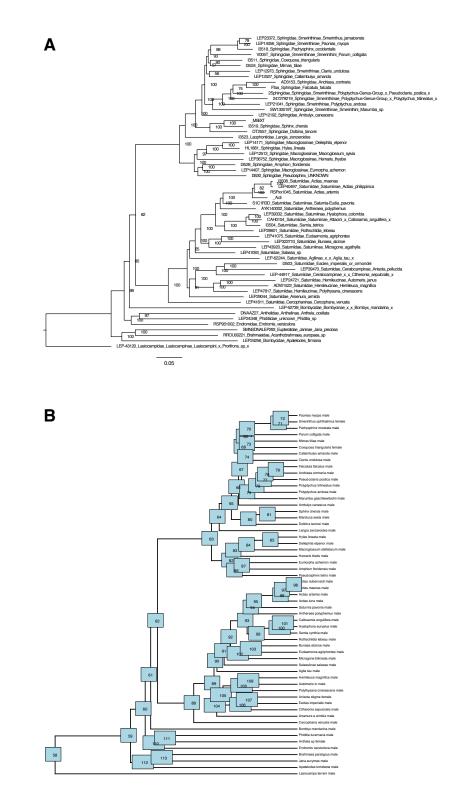
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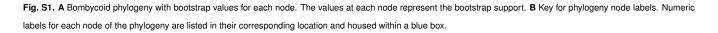
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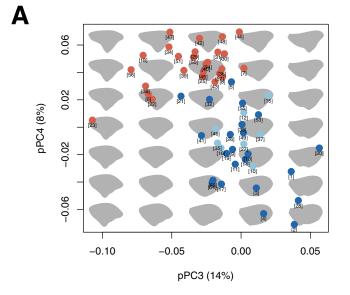
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Supplemental Information







В

Species-Number key for morphospace pPCA

 Sphingidae 	 Saturniid 	ae • Early div	erging bombycoids
• 1. Actias_artemis	• 16. Brahmaea_paratypus	• 31. Hemaris_thetis	• 46. Phiditia_lucernaria
• 2. Actias_dubernardi	• 17. Bunaea_alcinoe	• 32. Hemileuca_magnifica	• 47. Polyptychus_andosa
 3. Actias_luna 	18. Callambulyx_amanda	• 33. Hyalophora_euryalus	48. Polyptychus_trilineatus
• 4. Actias_maenas	19. Callosamia_angulifera	34. Hyles_lineata	• 49. Polythysana_cinerascens
● 5. Aglia_tau	• 20. Cercophana_venusta	35. Jana_eurymas	• 50. Pseudoclanis_postica
• 6. Ambulyx_canescus	• 21. Citheronia_sepulcralis	• 36. Langia_zenzeroides	• 51. Pseudosphinx_tetrio
• 7. Amphion_floridensis	• 22. Clanis_undulosa	• 37. Lasiocampa_terreni	• 52. Rothschildia_lebeau
• 8. Andriasa_contraria	 23. Coequosa_triangularis 	• 38. Macroglossum_stellatarum	• 53. Salassiinae_salassa
9. Anisota_stigma	• 24. Deilephila_elpenor	• 39. Manduca_sexta	• 54. Samia_cynthia
10. Anthela_sp	• 25. Dolbina_tancrei	40. Marumba_gaschkewitschii	• 55. Saturnia_pavonia
• 11. Antheraea_polyphemus	• 26. Eacles_imperialis	• 41. Micragone_bilineata	• 56. Smerinthus_ophthalmica
12. Apatelodes_torrefacta	• 27. Endromis_versicolora	• 42. Mimas_tiliae	• 57. Sphinx_chersis
13. Arsenura_a_armida	 28. Eudaemonia_agriphontes 	• 43. Pachysphinx_modesta	
• 14. Automeris_io	• 29. Eumorpha_achemon	• 44. Paonias_myops	
15. Bombyx_mandarina	• 30. Falcatula_falcatus	45. Parum_colligata	

Fig. S2. A Forewing morphospace across the pPC3 and pPC4 axes. Projections of the taxa onto the third and fourth phylogenetic principal components also demonstrates the separation of extant hawkmoths and silkmoths. B Key for species names in morphospace. This figure contains the number and corresponding scientific name of each species in morphospace. These numbers are used in panel A and in Fig. 2

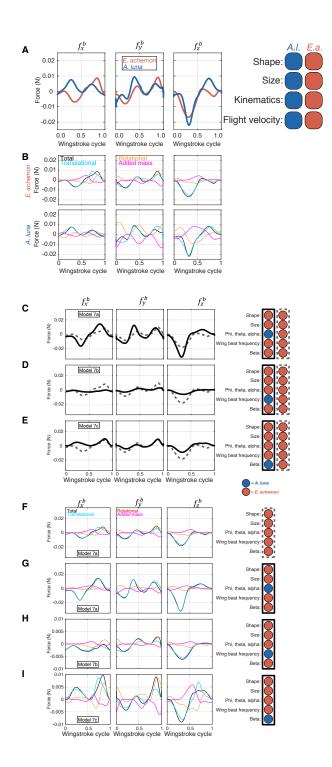


Fig. S3. A-B The total, translational, rotational, and added mass components of aerodynamic force during recorded flight speeds. (A) The total forces for *A. luna* (blue) and *E. achemon* (red) modeled at their natural flight velocities. (B) The component forces for each model using recorded flight velocities of each species. **C-I** The role of kinematic parameters in shaping the aerodynamics of each species. This set of models investigates the contribution of wing kinematics (C), wing beat frequency (D), and stroke plane angle (E) to total aerodynamic force production. In each panel, the two models are distinguished by solid and dashed lines. The variables used in each model can be found to the right of the data and are outlined in a corresponding solid or dashed line. The color of each circle represents the species from which each variable was measured. The components of the total aerodynamic force generated in each model are presented in panels **F-I**. The component forces for the dashed line model are only presented once (Panel E) because they are the same in each model.

Tip Name	Family	Genus	Species	Data	Dataset	Recovery	Geuns_species_of_imaged_moth	N Females	N Males	N Total
Acti	Saturniidae	Actias	luna	transcriptome	Kawahara & Breinholt 2014	776	Actias luna	0	2	2
MSEXT	Sphingidae	Manduca	sexta	transcriptome	Kawahara & Breinholt 2014	790	Manduca_sexta	0	2	2
247279219 Sphingidae Smerinthinae Polyptychus-Genus-Group x Polyptychus trilineatus x	Sphingidae	Polyptychus	trilineatus	AHE	NEW	244	Polyptychus trilineatus	0	2	2
2Sphingidae_Sphingidae_Smerinthinae_Polyptychus-Genus-Group_x_Pseudoclanis_postica_x	Sphingidae	Pseudoclanis	postica	AHE	NEW	496	Pseudoclanis postica	0	2	2
AD3153_Sphingidae_Andriasa_contraria	Sphingidae	Andriasa	contraria	AHE	Hamilton et al., 2019	720	Andriasa_contraria	ō	2	2
ADW1522 Saturniidae Hemileucinae Hemileuca magnifica	Saturniidae	Hemileuca	magnifica	AHE	Hamilton et al., 2019	402	Hemileuca magnifica	ō	3	3
AYK140002 Saturniidae Antheraea polyphemus	Saturniidae	Antheraea	polyphemus	AHE	Rubin & Hamilton et al., 2018	750	Antheraea polyphemus	0	5	5
CAH0104 Saturniidae Saturniinae Attacini x Callosamia angulifera x	Saturniidae	Callosamia	angulifera	AHE	NEW	564	Callosamia angulifera	ő	5	5
DNAAZ27 Anthelidae Anthelinae Anthela ocellata	Anthelidae	Anthela	ocellata	AHE	Hamilton et al., 2019	442	Anthela_sp	1	0	1
DT2557_Sphingidae_Dolbina_tancrei	Sphingidae	Dolbina	tancrei	AHE	Hamilton et al., 2019	774	Dolbina tancrei	0	2	2
Ffaa Sphingidae Falcatula falcata	Sphingidae	Falcatula	falcata	AHE	Hamilton et al., 2019	761	Falcatula falcatus	0	1	1
HL1881 Sphingidae Hyles lineata	Sphingidae	Hyles	lineata	AHE	NEW	761	Hyles lineata	0	2	2
	Saturniidae	Eacles	imperialis	AHE	Hamilton et al., 2019	738	Eacles imperialis	0	3	3
13503_Saturniidae_Eacles_imperialis_or_ormondei								0	5	5
I3504_Saturniidae_Samia_tetrica	Saturniidae	Samia	tetrica	AHE	Rubin & Hamilton et al., 2018	753	Samia_cynthia	0		
I3508_Saturniidae_Actias_maenas	Saturniidae	Actias	maenas	AHE	Rubin & Hamilton et al., 2018	766	Actias_maenas	-	2	2
I3511_Sphingidae_Coequosa_triangularis	Sphingidae	Coequosa	triangularis	AHE	Hamilton et al., 2019	766	Coequosa_triangularis	1	0	1
I3518_Sphingidae_Pachysphinx_occidentalis	Sphingidae	Pachysphinx	imperator	AHE	NEW	776	Pachysphinx_modesta	0	3	3
I3519_Sphingidae_Sphinx_chersis	Sphingidae	Sphinx	chersis	AHE	Hamilton et al., 2019	775	Sphinx_chersis	0	2	2
I3523_Laophontidae_Langia_zenzeroides	Sphingidae	Langia	zenzeroides	AHE	Hamilton et al., 2019	763	Langia_zenzeroides	0	2	2
I3524_Sphingidae_Mimas_tiliae	Sphingidae	Mimas	tiliae	AHE	NEW	780	Mimas_tiliae	0	2	2
I3528_Sphingidae_Amphion_floridensis	Sphingidae	Amphion	floridensis	AHE	NEW	766	Amphion_floridensis	0	2	2
I3530_Sphingidae_Pseudosphinx_UNKNOWN	Sphingidae	Pseudosphinx	tetrio	AHE	Hamilton et al., 2019	771	Pseudosphinx_tetrio	0	2	2
LEP-43120_Lasiocampidae_Lasiocampinae_Lasiocampini_x_Prorifrons_sp_x	Lasiocampidae	Prorifrons	sp	AHE	NEW	497	Lasiocampa_terreni	0	2	2
LEP-44917_Saturniidae_Ceratocampinae_x_x_Citheronia_sepulcralis_x	Saturniidae	Citheronia	sepulcralis	AHE	NEW	567	Citheronia_sepulcralis	0	2	2
LEP-52739_Bombycidae_Bombycinae_x_x_Bombyx_mandarina_x	Bombycidae	Bombyx	mandarina	AHE	NEW	586	Bombyx_mandarina	0	2	2
LEP-62244_Saturniidae_Agliinae_x_x_Aglia_tau_x	Saturniidae	Aglia	tau	AHE	NEW	565	Aglia_tau	0	3	3
LEP12192 Sphingidae Ambulyx canescens	Sphingidae	Ambulyx	canescens	AHE	Hamilton et al., 2019	723	Ambulyx canescus	0	2	2
LEP12513 Sphingidae Macroglossinae Macroglossum sylvia	Sphingidae	Macroglossum	sylvia	AHE	Hamilton et al., 2019	532	Macroglossum stellatarum	0	1	1
LEP12527 Sphingidae Callambulyx amanda	Sphingidae	Callambulyx	amanda	AHE	Hamilton et al., 2019	741	Callambulyx amanda	0	1	1
LEP12973 Sphingidae Smerinthinae Clanis undulosa	Sphingidae	Clanis	undulosa	AHE	Hamilton et al., 2019	595	Clanis undulosa	0	2	2
LEP14056 Sphingidae Smerinthinae Paonias myops	Saturniidae	Paonias	myops	AHE	NEW	618	Paonias myops	0	3	3
LEP14171 Sphingidae Macroglossinae Deilephila elpenor	Sphingidae	Deilephila	elpenor	AHE	NEW	589	Deilephila elpenor	0	2	2
LEP14407 Sphingidae Macroglossinae Eumorpha achemon	Saturniidae	Eumorpha	achemon	AHE	NEW	597	Eumorpha achemon	0	2	2
LEP21041 Sphingidae Smerinthinae Polyptychus andosa	Sphingidae	Polyptychus	andosa	AHE	Hamilton et al., 2019	548	Polyptychus andosa	ō	2	2
LEP23372 Sphingidae Smerinthinae Smerinthus jamaicensis	Sphingidae	Smerinthus	jamaicensis	AHE	Hamilton et al., 2019	605	Smerinthus ophthalmica	2	0	2
LEP24256 Bombycidae Apatelodes firmiana	Apatelodidae	Apatelodes	firmiana	AHE	Hamilton et al., 2019	703	Apatelodes torrefacta	0	3	3
LEP24348 Phiditiidae unknown Phiditia sp	Phiditiidae	Phiditia	sp	AHE	NEW	547	Phiditia lucernaria	0	2	2
LEP24731_Saturniidae_Hemileucinae_Automeris_janus	Saturniidae	Automeris	ianus	AHE	NEW	614	Automeris io	0	2	2
LEP28601 Saturnidae Rothschildia lebeau	Saturniidae	Rothschildia	lebeau	AHE	Rubin & Hamilton et al., 2018	533	Rothschildia lebeau	0	4	4
LEP2904 Saturnidae Arsenura armida	Saturniidae	Arsenura	armida	AHE	Rubin & Hamilton et al., 2018	766	Arsenura a armida	0	3	3
LEP32044_saturnidae_Alsendra_annua	Saturniidae	Bunaea	alcinoe	AHE	Rubin & Hamilton et al., 2018	700	Bunaea alcinoe	0	2	2
LEP36752 Sphingidae Macroglossinae Hemaris thysbe	Sphingidae	Hemaris	thysbe	AHE	Hamilton et al., 2019	592	Hemaris thetis	0	1	1
	Saturniidae		colombia	AHE	NEW	592		0	3	3
LEP39332_Saturniidae_Saturniinae_Hyalophora_colombia		Hyalophora					Hyalophora_euryalus	1	3	3
LEP39470_Saturniidae_Ceratocampinae_Anisota_pellucida	Saturniidae	Anisota	pellucida	AHE	Hamilton et al., 2019	585	Anisota_stigma	-	2	-
LEP40497_Saturniidae_Saturniinae_Actias_philippinica	Saturniidae	Actias	philippinica	AHE	Rubin & Hamilton et al., 2018	570	Actias_dubernardi	0		2
LEP41075_Saturniidae_Eudaemonia_agriphontes	Saturniidae	Eudaemonia	agriphontes	AHE	Rubin & Hamilton et al., 2018	760	Eudaemonia_agriphontes	0	3	3
LEP41093_Saturniidae_Salassa_sp	Saturniidae	Salassa	sp	AHE	Rubin & Hamilton et al., 2018	748	Salassiinae_salassa	0	3	3
LEP41811_Saturniidae_Cercophaninae_Cercophana_venusta	Saturniidae	Cercophana	venusta	AHE	Hamilton et al., 2019	598	Cercophana_venusta	0	2	2
LEP43920_Saturniidae_Saturniinae_Micragone_agathylla	Saturniidae	Micragone	agathylla	AHE	Rubin & Hamilton et al., 2018	561	Micragone_bilineata	0	1	1
LEP47817_Saturniidae_Hemileucinae_Polythysana_cinerascens	Saturniidae	Polythysana	cinerascens	AHE	Hamilton et al., 2019	525	Polythysana_cinerascens	0	2	2
RROU00221_Brahmaeidae_Acanthobrahmaea_europaea_sp	Brahmaeidae	Brahmaea	europaea	AHE	Hamilton et al., 2019	502	Brahmaea_paratypus	0	1	1
RSP951002_Endromidae_Endromis_versicolora	Endromidae	Endromis	versicolora	AHE	Hamilton et al., 2019	769	Endromis_versicolora	0	2	2
RSPxx1045_Saturniidae_Actias_artemis	Saturniidae	Actias	artemis	AHE	Rubin & Hamilton et al., 2018	764	Actias_artemis	0	2	2
S1C1R3D_Saturniidae_Saturniinae_Saturnia-Eudia_pavonia	Saturniidae	Saturnia	pavonia	AHE	Hamilton et al., 2019	588	Saturnia_pavonia	0	3	3
SMNSDNALEP293_Eupterotidae_Janinae_Jana_preciosa	Eupterotidae	Jana	preciosa	AHE	Hamilton et al., 2019	575	Jana_eurymas	0	2	2
SW130019T_Sphingidae_Smerinthinae_Smerinthini_Marumba_sp	Sphingidae	Marumba	sp	transcriptome	NEW	719	Marumba_gaschkewitschii	0	2	2
V005T_Sphingidae_Smerinthinae_Smerinthini_Parum_colligata	Sphingidae	Parum	colligata	transcriptome	NEW	723	Parum colligata	0	2	2

Table S1. Tip names and data source of each species in phylogeny as well as sex and count of individuals used

in shape analysis for each species

		All clades	ades		2	cmouns and Ha	Silkmoths and Hawkmoths only	< 		SIIKmoths only	sonly			Hawkmoths only	ins only	
VARIABLE	Lambda	logL	logL0	q	Lambda	logL	logL0	q	Lambda	logL	log L0	q	Lambda	logL	logL0	q
Wing length	6.73E-01	154.77413	152.72192	4.28E-02	5.97E-01	134.6286	132.7487	5.25E-02	4.78E-01	62.89071	78	0.339669	7.53E-01	72.96265	72.68433	0.455618
Mean chord length of wing	8.04E-01	222.56847	211.63032	2.91E-06	7.17E-01	193.8313	182.1356	1.32E-06	5.92E-01	87.87921		0.173243	2.23E-01			0.705753
Wing Area	6.61E-01	355.15542	349.67178	9.27E-04	5.76E-01	306.6241	300.5873	5.11E-04	3.86E-01	139.564 139.2045		0.396474	5.63E-01	171.762	171.774	1
Radius of second moment of area	3 8.94E-01	156.6572	146.81576	9.14E-06	7.62E-01	139.3126	133.2992	5.24E-04	7.98E-01	61.96275	60.67493	0.10852	7.95E-05	82.46353	82.46368	1
Aspect ratio	8.66E-01	-27.37978	-54.46312	1.84E-13	7.94E-01	-21.4874	-45.943	2.68E-12	6.70E-05	-3.61346	-3.61313	1	7.95E-05	-8.13094	-8.1307	1
Body length	4.93E-01	182.05982	180.31661	6.19E-02	3.30E-01	157.8792	155.8999	4.66E-02	6.70E-05	80.09822	80.09846	1	9.94E-01	81.85932	81.88774	1
Body mass	6.65E-05	-88.65496	-88.6541	1.00E+00	6.67E-05	-77.504	-77.5037	1.00E+00	6.70E-05	-28.0873	-28.0871	1	8.98E-01	-45.8953	-45.4909	1
Wing beat frequency	9.68E-01	-202.68686	-221.00342	1.43E-09	9.63E-01	-171.949	-191.554	3.81E-10	8.88E-01	-68.9056		0.020265	9.00E-01	-95.5285		0.006303
Wing loading	9.25E-01	88.49541	73.49837	4.33E-08	1.01E+00	77.64915	60.71952	5.92E-09	9.89E-01	55.15641		0.056712	8.01E-01	35.38334		0.024728
Thorax length (% body length)	7.41E-01	109.0667	103.55681	9.01E-04	4.11E-01	99.19072	93.71724	9.38E-04	5.95E-05	46.63365	46.63371	1	7.95E-05	55.46341	55.46362	1
Abdomen length (% body length)	8.02E-01	101.83411	97.28912	2.57E-03	5.92E-01	91.86848	88.17052	6.54E-03	4.29E-01	41.57689		0.379065	7.25E-01	52.01277	50.48391	0.080354
Abdomen length (meters)	7.67E-01	202.6083	200.03252	2.32E-02	8.76E-01	176.1017	172.9968	1.27E-02	6.70E-05	88.18174	88.18192	1	9.23E-01	91.28054	91.34096	1
Inertial power	7.88E-01	-220.16556	-222.89423	1.95E-02	7.70E-01	-187.489	-188.932	8.93E-02	1.79E-01	-81.3109		0.830876	8.72E-01	-101.994	-103.019	0.152355
pPC1	7.92E-01	98.08059	73.38695	2.10E-12	7.23E-01	85.00891	61.67966	8.45E-12	6.70E-05	41.22814	41.22848	1			54.26087	0.525214
pPC2	6.41E-01	110.485	106.707	5.98E-03	8.20E-02	99.33557	99.25253	6.84E-01	3.04E-01		42.8051	0.396353	7.95E-05	59.07206	59.07238	1
pPC3	5.89E-01	126.1117	118.483	9.38E-05	5.77E-01	107.7102	99.98249	8.45E-05	7.99E-01	54.79132	53.60685	0.123771	7.95E-05	57.79694	57.79727	1
5004	8.62E-01	133.768	107.5375	4.39E-13	8.49E-01	115.9159	90.28858	8.11E-13	7.81E-01	51.58913	50.12487	0.087028	7.95E-05	72.31542	72.31576	1

Table S2. Phylogenetic signal for each variable using the entire phylogeny and various pruned phylogenies

| Node number
58

 | Ancestral state
-5.67E-16 | pPC1
Upper 95% CI
-0.114783617

 | Lower 95% Ci
 | Ancestral state
-0.114783617 | pPC2
Upper 95% CI
-0.08438098 | Lower 95% CI | Ancestral state
0.03530532
 | Wing length
Upper 95% CI
-0.00562559 | Lower 95% CI | Mean
Ancestral state
0.012481993
 | chord length of
Upper 95% CI
-0.00133569 | wing
Lower 95% CI
0.02629967
 | Ancestral state
0.000524736 | Wing area
Upper 95% CI
-1.03E-03
 | Lower 95% CI | Radius of ti
Ancestral state
0.585369 | Upper 95% CI
0.5471965
 | Lower 95% CI
0.6235415
 | Ancestral state
2.848957 | Aspect ratio
Upper 95% CI
2.033795
 | Lower 95% CI
3.664119 | Ancestral state
0.02829903 | Body length
Upper 95% CI
0.00343255 | Lower 95% CI |

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--|--|---|--|
| 59
60

 | -2.56E-03
-4.93E-03 | -0.0673981
-0.061524517

 | 0.06227856
 | -0.0673981
-0.061524517 | -0.06372247
-0.06647971 | 0.031606843 0.016732345 | 0.03754315
 | 0.015316293
0.017361202 | 0.0577251 | 0.013242219 0.012628399
 | 0.005815267 | 0.02104747 0.01944153
 | 0.000609192
0.000561362 | -2.68E-04
-2.05E-04
 | 0.001486778 0.001327399 | 0.574515
0.5689185 | 0.5529524 0.5500966
 | 0.5960777
 | 2.953081
3.029427 | 2.492617
2.627493
 | 3.413545
3.431362 | 0.02934259 0.02831734 | 0.01529615
0.01605633 | 0.04338902 |
| 61
62

 | -7.75E-03
-1.22E-02 | -0.064090637
-0.068810748

 | 0.04859222 0.04446395
 | -0.064090637
-0.068810748 | -0.07104259
-0.07473722 | 0.011794048 0.008534501 | 0.03776706
 | 0.017676165 0.01867789 | 0.05785796 | 0.012499847
0.012638435
 | 0.005717452 0.005820417 | 0.01928224 0.01945645
 | 0.000560794
0.000580437 | -2.02E-04
-1.86E-04
 | 0.001323375
0.001347023 | 0.5671469 0.5659436 | 0.54841
0.5471082
 | 0.5858839 0.5847789
 | 3.083352
3.141861
3.862847 | 2.683231
2.739638
 | 3.483474
3.544084 | 0.02846948 0.02914303 | 0.0162638 0.01687324 | 0.04067517
0.04141283 |
| 63
64
65

 | -8.65E-02
-9.25E-02
-9.69E-02 | -0.134235498
-0.138663194
-0.136811536

 | -0.03885561
-0.04643063
-0.05697497
 | -0.134235498
-0.138663194
-0.136811536 | -0.08310554
-0.08575808
-0.07999142 | -0.012988837
-0.01795508
-0.021301098 | 0.0425123
0.04500151
0.04530894
 | 0.02550645
0.028556813
0.031074403 | 0.05951815
0.0614462 | 0.011062137
0.011453115
0.011444204
 | 0.00532121
0.005901626
0.006638831 | 0.01680306
0.0170046
0.01624958
 | 0.000540471 0.000580611 0.000569982 | -1.05E-04
-4.36E-05
2.97E-05
 | 0.001185954
0.001204794
0.001110276 | 0.568373
0.567345
0.5671425 | 0.5525132
0.5520086
0.5538672
 | 0.5842327
0.5826815
0.5804177
 | 3.862847
3.930447
3.962204 | 3.524166
3.602941
3.678716
 | 4.201528
4.257952
4.245693 | 0.03648822
0.0379693
0.03858273 | 0.02615677 | 0.04681966
0.04795983
0.04723054 |
| 66
67

 | -9.64E-02
-9.48E-02 | -0.136811536
-0.131761302
-0.127286235

 | -0.05697497
-0.06106687
-0.06237826
 | -0.131761302
-0.127286235 | -0.07999142
-0.07612637
-0.07265989 | -0.021301098
-0.024156717
-0.024944035 | 0.0460027
0.04603033
 | 0.031074403
0.03339817
0.034457497 | 0.05860723 | 0.011444204
0.011596277
0.011634879
 | 0.00734117
0.00772806 | 0.01585138 0.0155417
 | 0.000582517
0.000585938 | 1.04E-04
1.47E-04
 | 0.001110276
0.001060941
0.001025203 | 0.5662414
0.5664753 | 0.5538672
0.55544863
0.5556824
 | 0.5779965
0.5772682
 | 3.962204
3.973969
3.965812 | 3.722942
3.735332
 | 4.245693
4.224995
4.196291 | 0.03858273
0.03890117
0.03886534 | 0.03124363 0.03183458 | 0.04655872
0.0458961 |
| 68
69

 | -9.57E-02 | -0.127873038
-0.127568098

 | -0.06345158
 | -0.127873038 | -0.07222731 | -0.02486911
-0.02173919 | 0.0470675
 | 0.035581414 | 0.05855359 | 0.011883204
 | 0.008005668 | 0.01576074
 | 0.000612391 0.00061053 | 1.76E-04
1.61E-04
 | 0.001048363 | 0.5662655 | 0.5555535
 | 0.5769775
 | 3.972439 | 3.743687
 | 4.201191 4.180703 | 0.03943944 | 0.03246137 | 0.0464175 |
| 70
71

 | -9.31E-02
-7.64E-02 | -0.129699843
-0.107746723

 | -0.0564343
-0.04497875
 | -0.129699843
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0.011716752
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 | 0.5802672 0.5853492
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 | 4.185868
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 | 0.005438609
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0.005512112 | 0.01444427
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 | 4.209024
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79
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 | 4.102499
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82

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| 83
84

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86

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88

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 | 0.5872046
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 | 4.327964
3.374421 | 0.03615124 0.02783606 | 0.02578284 | |
| 89
90
91

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 | 0.032564619
0.033066373
0.035761897 | 0.06282393 | 0.01625912
0.016473141
0.017516935
 | 0.011280896
0.011450281
0.012444598 | 0.02123734
0.021496
0.02258927
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92
93

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97

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99
100

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| 102
103

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105

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| 113

 | 1.58E-03 | -0.07353989

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 | -0.07353989 | -0.06344292 | 0.047010041 | 0.04440533
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 | 0.006758769 |
 | | -1.96E-04
 | 0.001837207 | 0.5781853 | 0.5532018
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|

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 | 0.01/0104/3 | |
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 | 0.0031000
 | 2.830240 | 2.322/31
 | | 0.03341132 | | |
|

 | continued. | Body mass

 |
 | • | ving beat frequer | ncy | 1
 | Wing loading | 1 | Thorax I
 | ength (% body |
 | Abdomen | length (% bod
 | | Abdo | men length (me
 | ters)
 | | Inertial power
 | | 0.03341132 | | |
| Node number
58

 | Ancestral state
0.8760748
0.9991875 | Body mass
Upper 95% Cl
-1.76942793

 | Lower 95% Cl
3.521577
2.493564
 | W
Ancestral state
27.37922 | Ving beat frequer
Upper 95% Cl
10.073798 | ncy
Lower 95% CI
44.68465 | Ancestral state
0.08755236
 | Wing loading
Upper 95% CI
-0.01409863 | Lower 95% CI
0.18920334 |
 | ength (% body
Upper 95% Cl
0.2193811 |
 | | length (% bod
Upper 95% CI
0.4966009
 | | Abdo
Ancestral state
0.01757996 | men length (me
Upper 95% Cl
0.002480124
 | ters)
Lower 95% Cl
0.03267979
 | Ancestral state
33.78903 | Inertial power
Upper 95% CI
8.468984
 | Lower 95% CI
59.10908 | 0.03341132 | | |
|

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323 | Body mass
Upper 95% CI
-1.76942793
-0.49518934
-0.35088294
-0.32071012

 | 3.521577
2.493564
2.257972
2.276375
 | M
Ancestral state
27.37922
26.45973
27.30315
27.75656 | Ving beat frequer
Upper 95% Cl
10.073798
16.684342
18.770298
19.262208 | Lower 95% CI
44.68465
36.23513
35.83599
36.25091 | Ancestral state
0.08755236
0.08529203
0.08795014
0.09051374
 | Wing loading
Upper 95% CI
-0.01403863
0.02787198
0.03782874
0.04061845 | Lower 95% CI
0.18920334
0.14271207
0.13807155
0.14040902 | Thorax I
Ancestral state
0.3195407
0.3009029
0.3037193
0.3066549
 | ength (% body
Upper 95% CI
0.2193811
0.2443253
0.2543333
0.2574916 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215 | length (% bod
Upper 95% CI
0.4966009
0.5718666
0.5786124
0.5763337
 | y length)
Lower 95% Cl
0.7240282
0.7003345
0.6907507
0.6879662 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182212
0.01826141 | men length (mer
Upper 95% Cl
0.010426745
0.010775868
0.010849669
 | ters)
Lower 95% CI
0.03267979
0.02748576
0.02566652
0.02567315
 | Ancestral state
33.78903
36.44301
36.7972
37.19876 | Inertial power
Upper 95% CI
8.468984
22.140364
24.312557
24.770435
 | Lower 95% CI
59.10908
50.74566
49.28185
49.62708 | 0.03341132 | | |
| Node number
58
59
60
61
62
63

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323
1.0435646
1.5779134 | Body mass
Upper 95% Cl
-1.76942793
-0.49518934
-0.35088294
-0.32071012
-0.26179817
0.47876808

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
 | W
Ancestral state
27.37922
26.45973
27.30315
27.75656
27.68571
34.27996 | Upper 95% CI
10.073798
16.684342
18.770298
19.262208
19.246747
27.089959 | Lower 95% Cl
44.68465
36.23513
35.83599
36.25091
36.22468
41.46997 | Ancestral state
0.08755236
0.08529203
0.08795014
0.09051374
0.09398024
0.15135673
 | Wing loading
Upper 95% CI
-0.01409863
0.02787198
0.03782874
0.04061845
0.04382289
0.10912309 | Lower 95% Cl
0.18920334
0.14271207
0.13807155
0.14040902
0.14413759
0.19359037 | Thorax I
Ancestral state
0.3195407
0.3009029
0.3037193
0.306549
0.3072389
0.2961875
 | ength (% body
Upper 95% Cl
0.2193811
0.2443253
0.2543333
0.2574916
0.2578174
0.2545735 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
0.3558604
0.3378015
 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215
0.6316738
0.6385046 | length (% bod
Upper 95% Cl
0.4966009
0.5718666
0.5786124
0.5763337
0.5755644
0.5912592
 | y length)
Lower 95% Cl
0.7240282
0.7003345
0.6907507
0.6879662
0.6877832
0.68575 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182612
0.01826141
0.01867781
0.02352193 | men length (mei
Upper 95% Cl
0.002480124
0.010426745
0.010775868
0.010849669
0.011227146
0.017248298
 | ters)
Lower 95% CI
0.03267979
0.02748576
0.0256652
0.02567315
0.02612848
0.02979556
 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294 | Inertial power
Upper 95% CI
8.468984
22.140364
24.312557
24.770435
25.826327
37.923047
 | Lower 95% CI
59.10908
50.74566
49.28185
49.62708
50.81352
58.96284 | 0.03341132 | | |
| Node number
58
59
60
61
62
63
64
65

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029 | Body mass
Upper 95% CI
-1.76942793
-0.49518934
-0.35088294
-0.32071012
-0.26179817
-0.47876808
0.59741823
0.80026392

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
2.72317
2.640316
 | W
Ancestral state
27.37922
26.45973
27.30315
27.75656
27.68571
34.27996
32.29742
30.94837 | Ving beat frequer
Upper 95% Cl
10.073798
16.684342
18.770298
19.262208
19.146747
27.089959
25.344668
24.930064 | Lower 95% Cl
44.68465
36.23513
35.83599
36.25091
36.22468
41.46997
39.25017
36.96667 | Ancestral state
0.08755236
0.08529203
0.08795014
0.09051374
0.09338024
0.15135673
0.14467611
0.14110493
 | Wing loading
Upper 95% Cl
-0.01409863
0.02787198
0.02787198
0.04061845
0.04382289
0.10912309
0.1033609
0.10575379 | Lower 95% Cl
0.18920334
0.14271207
0.13807155
0.14040902
0.14413759
0.19359037
0.18551613
0.17645608 | Thorax A
Ancestral state
0.3195407
0.3009029
0.3037193
0.3066549
0.3072389
0.2944671
0.2944671
0.2906338
 | length (% body
Upper 95% Cl
0.2193811
0.2443253
0.2543333
0.2574916
0.2578174
0.2545735
0.2542263
0.2558013 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
0.3566604
0.3378015
0.334708
0.3254663
 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215
0.6316738
0.64385046
0.6438504
0.643842
0.6479501 | length (% bod
Upper 95% Cl
0.4966009
0.5718666
0.5786124
0.5786124
0.5763337
0.5755644
0.5981556
0.608404
 | y length)
Lower 95% Cl
0.7240282
0.7003345
0.6907507
0.68779662
0.6877832
0.68575
0.68575
0.6855284
0.6874963 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182212
0.01826141
0.01867781
0.02352193
0.02463604
0.0251024 | men length (met
Upper 95% Cl
0.002480124
0.010426745
0.01075868
0.010849669
0.011227146
0.011227146
0.01228298
0.018569421
0.018551129
 | ters)
Lower 95% Cl
0.03267979
0.02748576
0.02566652
0.02567315
0.02612848
0.02979556
0.03070265
0.03070265
0.03035366
 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.85431 | Inertial power
Upper 95% CI
8.468984
22.140364
24.312557
24.770435
25.826327
37.923047
41.283372
43.078762
 | Lower 95% Cl
59.10908
50.74566
49.28185
49.62708
50.81352
58.96284
61.6289
60.68986 | 0.03341152 | | |
| Node number
58
59
60
61
62
63
64

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029
1.8385855
1.8648451 | Body mass
Upper 95% CI
-1.76942793
-0.49518934
-0.35088294
-0.32071012
-0.26179817
0.47876808
0.59741823
0.80026392
1.02391222
1.11685412

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
2.72317
2.640316
2.653259
2.612836
 | W
Ancestral state
27.37922
26.45973
27.30315
27.75656
27.68571
34.27996
32.29742
30.94837
30.174
29.79839 | Ving beat frequer
Upper 95% CI
10.073798
16.684342
19.262208
19.146747
27.089959
25.344668
24.930064
24.844658
24.930064 | Lower 95% Cl
44.68465
36.23513
35.83599
36.25091
36.22468
41.46997
39.25017
36.96667
35.50315
34.69134 | Ancestral
state
0.08755236
0.08759203
0.08795014
0.09051374
0.09051374
0.0938024
0.15135673
0.1447611
0.14410493
0.1415014
0.13941814 | Wing loading
Upper 95% C1
-0.01409863
0.02787198
0.03782874
0.04061845
0.04382289
0.04382289
0.10375379
0.10375379
0.11024708
0.11067728 | Lower 95% CI
0.18920334
0.14271207
0.13807155
0.14040902
0.14413759
0.19359037
0.18551613
0.17645608
0.17285321
0.16815899 | Thorax I
Ancestral state
0.3195407
0.3009029
0.3037193
0.3066549
0.3072389
0.3072389
0.2961875
0.2944671
0.2906338
0.2890027
0.2876816
 | length (% body
Upper 95% C1
0.2193811
0.2443253
0.2543333
0.2574916
0.2578174
0.2542263
0.2558013
0.2558013
0.25581589
0.2558524 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
0.356604
0.358015
0.334708
0.3254663
0.3198465
0.3180008
 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215
0.6316738
0.6438046
0.6438042
0.6439021
0.6521146
0.6552146 | length (% bod
Upper 95% CI
0.4966009
0.5718666
0.5786124
0.5763337
0.5755644
0.5912592
0.5981556
0.608404
0.6170969
0.6228982
 | y length)
Lower 95% Cl
0.7240282
0.7003345
0.6907507
0.6877632
0.6877832
0.6877832
0.68575
0.6895284
0.6871823
0.6871323 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182212
0.01826141
0.01867781
0.02352193
0.02463604
0.0251024
0.025423 | men length (me
Upper 95% Cl
0.002480124
0.010426745
0.01075868
0.011227146
0.011227146
0.011227146
0.011227146
0.018561421
0.018561421
0.021258648
 | ters)
Lower 95% Cl
0.03267979
0.02748576
0.02566652
0.02657315
0.02612848
0.02979556
0.03070265
0.0303366
0.03003224
0.02979732
 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.88431
52.12407
51.77322 | Inertial power
Upper 95% CI
8.468384
22.140364
24.312557
24.770435
25.826327
37.923047
41.283372
43.078762
44.326846
44.614214
 | Lower 95% CI
59.10908
50.74566
49.28185
49.62708
50.81352
58.96284
61.6289
60.68986
59.92129
58.93222 | 003341152 | | |
| Node number
58
59
60
61
62
63
64
65
66
65
66
67
68
69

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029
1.8385855
1.8648451
2.0012356
2.0164672 | Body mass
Upper 95% (-1
-1.76942793
-0.49518934
-0.35088294
-0.35088294
-0.26179817
-0.47876808
0.59741823
0.80026392
1.02391222
1.11685412
1.25825109
1.25972931

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
2.72317
2.640316
2.653259
2.612836
2.74362
2.782205
 | W
Ancestral state
27.37922
26.45973
27.30315
27.75656
32.29742
30.94837
30.174
29.79839
29.50299
29.51254 | Ving beat frequer
Upper 95% CI
10.073798
16.684342
18.770298
19.262208
19.146747
27.089959
25.344668
24.930064
24.844658
24.905447
24.646715
24.503502 | Lower 95% Cl
44.68465
36.23513
35.83599
36.25091
36.22468
41.46697
39.25017
36.96667
35.50315
34.69134
34.35926
34.59124 | Ancestral
state
0.08755236
0.085755246
0.08795014
0.09051374
0.09051374
0.0513567
0.15135674
0.15135674
0.15135014
0.1415014
0.14252769
0.14252769 | Wing loading
Upper 95% Cl
-0.01403863
0.02787198
0.03782874
0.04061845
0.04061845
0.10312309
0.10375379
0.11027708
0.11067728
0.11067728
0.1124829 | Lower 95% CI
0.18920334
0.14271207
0.13807155
0.14040902
0.144413759
0.19359037
0.18551613
0.17645608
0.17285321
0.16815899
0.17105312
0.1719383 | Thorax
Ancestral state
0.3195407
0.3009029
0.3007193
0.3066549
0.3072389
0.2961875
0.2944671
0.2906338
0.2890027
0.2876816
0.2873079
0.2877666
 | ength (% body
Upper 95% C
0.2193811
0.2443253
0.2574916
0.2578174
0.2542263
0.2542263
0.2558013
0.2581589
0.2593624
0.2593624
0.259201
0.2592755 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
0.3556604
0.337805
0.334708
0.3254663
0.3198463
0.3196008
0.31954148
0.31617577
 | Abdomen
Ancestral state
0.6103145
0.6346105
0.6346816
0.63215
0.6316738
0.6385046
0.643842
0.6479501
0.6551496
0.6550496
0.6554839 | length (% bod
Upper 95% Cl
0.4966009
0.5718666
0.5786124
0.576337
0.5755644
0.5912592
0.6981556
0.608404
0.6170969
0.6228982
0.62245735
0.62243386
 | y length)
Lower 95% Cl
0.7240282
0.7003345
0.6907507
0.6879662
0.6877832
0.68755
0.6895284
0.6874963
0.6871323
0.6871201
0.6883944
0.6882671 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182512
0.0182512
0.018267181
0.02352193
0.02463604
0.0251024
0.02552798
0.02552798
0.02552798 | men length (met
Upper 95% Cl
0.002480124
0.010426745
0.01077868
0.011227146
0.011227146
0.0118569421
0.0188569421
0.0128569421
0.020792357
0.0221258648
0.0221730434
0.0221254508
 | ters)
Lower 95% Cl
0.03267979
0.02748576
0.0256652
0.02567315
0.02612848
0.03070265
0.03070265
0.0303366
0.0309324
0.02979732
0.0302051
0.02999577
 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.88431
52.12407
51.77322
52.39662
51.8438 | Inertial power
Upper 95% CI
8.468984
22.140364
24.312557
24.770435
25.826327
41.283372
43.078762
44.326846
44.614214
45.291276
44.514936
 | Lower 95% CI
59.10908
50.74566
49.22708
50.81352
58.96284
61.6289
60.68986
59.92129
58.93222
59.50197
59.17266 | 003341132 | | |
| Node number
58
59
60
61
62
63
64
65
66
65
66
67
68
69
70
71

 | Ancestral state
0.8760748
0.9991875
0.9535444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029
1.835855
1.85648451
2.0012356 | Body mass
Upper 95% C1
-1.76942793
-0.49518934
-0.35088294
-0.32071012
-0.26179817
-0.47376808
0.59741823
0.80026392
-1.02391222
1.11685412
1.2585109
1.25072931
1.06853064
0.97446042

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
2.72317
2.640316
2.653259
2.612836
2.74362
 | M
Ancestral state
27.37922
26.45973
27.30315
27.75656
32.29742
30.94837
30.174
29.79839
29.50299 | Ving beat freques
Upper 95% Cl
10.073798
16.684342
18.770298
19.262208
19.146747
27.089959
25.344668
24.930064
24.844558
24.905447
24.646715
24.503502
24.198745
26.246505 | Lower 95% CI
44.68465
36.23513
35.83599
36.225091
36.22468
41.46997
39.25017
36.96667
35.50315
34.69134
34.35926 | Ancestral
state
0.08755236
0.08755236
0.08795014
0.09398024
0.09398024
0.09398024
0.15135673
0.14457611
0.14110491
0.14155014
0.14217106
0.14048241
0.14374997 | Wing loading
Upper 95% Cl
-0.01403863
0.02787198
0.02787198
0.02382874
0.04082289
0.10912309
0.10355379
0.11024708
0.11067728
0.11407226
0.11274829
0.10255669 | Lower 95% CI
0.18920334
0.14271207
0.13807155
0.14040902
0.14413759
0.1935037
0.1855161
0.17645608
0.17285321
0.16815899
0.17105312 | Thorax
Ancestral state
0.3195407
0.3009029
0.3037193
0.3066549
0.3072389
0.2961875
0.2944671
0.290632
0.2890027
0.2876816
0.2873079
0.2877666
0.2851196
0.2851196
 | length (% body
Upper 95% CI
0.2193811
0.2443253
0.2543333
0.2574916
0.2574916
0.2578174
0.2584263
0.2581589
0.2583624
0.2583624
0.2583755
0.2581755
0.2583755 | length)
0.4197003
0.3574805
0.3531054
0.358181
0.3566604
0.3378015
0.3254663
0.3198465
0.3198465
0.3196408
 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215
0.6316738
0.64385046
0.6438504
0.64379501
0.6521146
0.6550496
0.6550496 | length (% bod
Upper 95% CI
0.4966009
0.5718666
0.5786124
0.5756347
0.5755644
0.5912592
0.5981556
0.608404
0.6170969
0.6228982
0.6224385
0.6234386
0.6234226
 | y length)
Lower 95% CI
0.7240282
0.7003345
0.6907507
0.6877622
0.6877832
0.68575
0.6855284
0.6872011
0.6882011
0.68828144
0.68282671
0.66820521
0.6692052
0.6948339 | Abdo
Ancestral state
0.01757996
0.01825212
0.01825141
0.028252193
0.02463604
0.0251423
0.02542423
0.02552798
0.02552798
0.02556777
0.02562514
0.02552128 | men length (me
Upper 95% CI
0.002480124
0.010426745
0.01075868
0.011227146
0.011227146
0.0112748298
0.01856129
0.020792357
0.021258648
 | ters)
Lower 95% CI
0.03267979
0.02748576
0.02566652
0.0267315
0.02612848
0.02979556
0.03070254
0.03009224
0.02979732
0.0302051
 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.88431
52.12407
51.77322
52.39662 | Inertial power
Upper 95% CI
8.468384
22.140364
24.312557
24.770435
25.826327
37.923047
41.283372
43.078762
44.326846
44.614214
45.291276
 | Lower 95% CI
59.10908
50.74566
49.28185
49.62708
50.81352
58.96284
61.6289
60.68986
59.92129
58.93222
59.50197
59.17266
59.24242
53.24745 | 003341132 | | |
| Node number
58
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 | Ancestral state
0.8760748
0.9991875
0.935444
0.9778323
1.0435646
1.77029
1.8385855
1.8648451
2.0012356
2.0164672
1.9132331
1.6697903 | Body mass
Upper 95% C1
-1.76942793
-0.49518934
-0.32071012
-0.226179817
-0.47576808
0.59741823
0.80026392
1.02391222
1.116655412
1.25885109
1.25072931
1.06893064

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.677059
2.72317
2.640316
2.653259
2.612836
2.74362
2.782205
2.775736
2.42112
 | M
Ancestral state
27.37922
26.45973
27.30315
27.75656
27.68571
34.27996
32.29742
30.174
29.79839
29.50299
29.51254
29.772171
30.97813 | Ving beat frequen
Upper 95% CI
10.073798
16.684342
18.770298
19.146747
27.089959
25.344668
24.930064
24.944658
24.930064
24.944658
24.905447
24.646715
24.503502
24.198745 | Lower 95% Cl
44.68465
36.23513
35.83599
36.25091
36.22468
41.46997
39.25017
36.96667
35.50315
34.69134
34.3926
34.52158
35.24467
35.70976 | Ancestral
state
0.08755236
0.08529203
0.09051374
0.09051374
0.09308024
0.15135673
0.1446761
0.14110493
0.14455014
0.13941814
0.13941814
0.14252769
0.14217106 | Wing loading
Upper 95% CI
-0.01409863
0.02787188
0.03782874
0.04061845
0.04061845
0.040362289
0.04382289
0.04382289
0.10383609
0.103575379
0.11024708
0.11024708
0.11024778
0.11204778
0.11204778 | Lower 95% CI
0.18220334
0.14271207
0.13807155
0.14040902
0.13807155
0.14040902
0.138551613
0.17645568
0.17265321
0.168158993
0.17165132
0.17165132
0.17169383
0.17129383
0.17292334
0.18154325
0.18460287
0.17620547 | Thorax I
Ancestral state
0.3195407
0.3009029
0.3007193
0.3066549
0.3072389
0.2961875
0.2944671
0.2906338
0.2890027
0.2876816
0.2873079
0.2877666
 | Lipper 95% Cl
0.2193811
0.2443253
0.2574916
0.2574916
0.25578174
0.25452263
0.2558013
0.2581589
0.259201
0.2592755
0.2592755
0.253154 | length)
Lower 95% Cl
0.4197003
0.3574805
0.3531054
0.3558181
0.3558664
0.358664
0.3254663
0.3198465
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 | Abdomen
Ancestral state
0.6103145
0.6361005
0.6346816
0.63215
0.6316738
0.6385046
0.643842
0.6479501
0.6521146
0.6550496
0.6550496
0.65563529
0.65599139
0.6599139 | length (% bod
Upper 95% CI
0.4966009
0.5786124
0.5786124
0.59812592
0.5981556
0.608404
0.6170969
0.6228982
0.6245735
0.6234386
0.6234386
 | y length)
Lower 95% CI
0.7240282
0.7003345
0.6907507
0.6877962
0.6877832
0.6877832
0.68774963
0.6871323
0.6872011
0.6883944
0.6892671
0.692652 | Abdo
Ancestral state
0.01757996
0.01895625
0.0182212
0.01826141
0.02857781
0.02463604
0.0251024
0.0255423
0.02552778
0.02562517
0.02562517 | men length (mei
Upper 95% Cl
0.0102480124
0.010426745
0.01075868
0.010275868
0.011227146
0.011227146
0.0118269421
0.018569421
0.01985129
0.020792357
0.021258648
0.021730434
0.021254508
0.0205602192
 | ters)
Lower 95% (1
0.03267379
0.02748576
0.0256652
0.02567315
0.02612248
0.02979556
0.03070265
0.03003236
0.03003236
0.0300324
0.030299577
0.0302454
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 | Ancestral state
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.88431
52.12407
51.77322
52.39662
51.8438
51.16162
52.39662
51.8438
51.16162 | Inertial power
Upper 95% CI
8.468984
22.140364
24.312557
24.770435
25.826327
37.923047
41.283375
43.078752
43.078752
43.078752
44.326846
44.614214
45.291276
44.514936
43.08082
39.401505
 | Lower 95% CI
59.10908
50.74566
49.28185
49.62708
50.81352
58.96284
61.6289
60.68986
59.92129
58.93222
59.50197
59.17266
59.24242 | 0.0334132 | | |
| Node number
58
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 | Ancestral state
0.8760748
0.9931875
0.9335444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029
1.8385855
1.8548451
2.0012356
2.0164672
1.9132231
1.669828
2.1413467
2.1930682
1.4593551
1.073758 | Body mass
Upper 95% C1
-1.76942793
-0.459518934
-0.35088294
-0.35088294
-0.26179817
-0.47876808
0.59741823
0.80026392
1.02391222
1.11685412
1.25885109
1.25072931
1.06893064
0.97446042
0.92911735
1.28616155
1.34065705
0.60173694
0.21165613

 | 3.521577
2.493564
2.257972
2.276375
2.348927
2.67059
2.772317
2.600316
2.653259
2.612836
2.74362
2.782205
2.77536
2.42112
2.304648
2.996532
3.045479
2.316973
1.93586
 | Ancestral state
27.37922
26.45973
27.30325
27.30315
27.75656
32.256571
32.25742
30.94837
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27.75555555555 | Ving beat frequer
Upper 35% (1
10.073798)
16.654342
19.166747
19.262208
19.166747
24.93064
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24.93064
24.930745
26.246505
27.23268
23.732868
23.242743
24.427745
24.4707852 | Lower 95% C1
44.68465
36.23513
35.83599
36.25091
36.22468
41.46997
39.25017
36.96667
35.50315
34.69134
34.35926
34.55158
35.24467
35.70976
36.64901
34.91157
36.64901
34.91157
35.49738
35.98676 | Ancestral
state
0.08755236
0.08755286
0.08755246
0.08755014
0.09051374
0.04875614
0.14110493
0.141155014
0.14110493
0.14155014
0.14217106
0.14048241
0.13941344577
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Upper 95% CI
-0.01403863
0.02787198
0.03782374
0.04081280
0.10312209
0.10312209
0.10042209
0.10047230
0.110677537
0.110677537
0.110677537
0.110677537
0.110677537
0.110678669
0.11274829
0.1013174937
0.11058669
0.11528576 | Lower 95% C
0.18920334
0.14271207
0.13807155
0.14040902
0.14413759
0.139359037
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 | Ancestral date
33.78903
36.44301
36.7972
37.19876
38.31992
48.44294
51.45613
51.85431
52.12407
51.77322
52.39643
51.16162
46.32248
51.16162
46.32248
51.73265
46.372376 | Inertial power
Upper 95%
(C
8.468384
22.140364
42.312557
24.770435
25.826527
37.923047
41.283372
43.078762
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44.514293
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45.10505
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39.40 | Lower 95% C1
59.10908
50.74566
49.28185
49.28185
49.62708
50.81352
58.90224
61.6289
59.92129
59.92129
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59.92129
59.50197
59.24242
59.324745
59.91762
56.96201
55.462413 | 0.0394134 | | |
| Node number
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 | Ancestral state
0.8760748
0.9991875
0.95335444
0.9778323
1.0435646
1.5779134
1.6602942
1.72029
1.8385855
1.8644851
1.9612356
2.01646772
2.1612362
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2.141 | Body mass
-1,7642793
-0,49518934
-0,49518934
-0,35088294
-0,35088294
-0,35071912
0,26179817
0,47876808
0,59741823
0,80026392
1,02391222
1,11685412
1,02891222
1,11685412
1,06893064
0,979446042
0,92911735
1,28616156
1,34065705
0,60173694
0,21165613
0,00988399
0,079541344

 | 3321577
2493564
2257972
2276375
2348927
2677059
272317
2660316
2653259
2612836
278250
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242112
2304648
2996532
3145479
2316973
193586
1726465
1693111
 | Accestral atua
27.37922
26.45973
27.3032
27.30315
27.76566
32.276557
34.27996
32.29742
30.174
30.298437
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25.50299
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25.51254 | Upper 75% C1
10.073738
16.664342
19.262208
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19.146747
27.099599
24.3446475
24.84455
24.940547
24.84455
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24.546475
24.546475
26.246675
27.265055
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23.723268
23.2422743
24.427736
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Lower 95% C1
44.65465
38.25513
38.25091
38.22468
41.46997
38.95667
38.25128
35.50315
34.69124
34.35926
35.24467
35.24467
35.24473
35.24473 | Ancestral
state
0.08755236
0.08755286
0.08755246
0.08755014
0.08795014
0.1938024
0.1411043
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0.1421706
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0.142776 | Wing loading
Upper 95% CI
-0.01403863
0.02787198
0.03782374
0.0408212309
0.103422309
0.103422309
0.10347230
0.10575379
0.11067723
0.10467234
0.11067723
0.1024708
0.11274829
0.102595609
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 | Abdomen
Anostrial state
0.6303145
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8.468384
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0.11274829
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0.074116455
0.09294785 | Lower 95% CI
0.18920334
0.14271207
0.13897155
0.14040902
0.13807155
0.1413759
0.1255321
0.177645608
0.17265321
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0.17159383
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0.17159283
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 | length (% bod
Upper 95% CI
0.4966009
0.577812466
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22:140364
24:312557
37:9230477
41:283372
43:078762
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44:5324936
44:54214
44:326846
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44:53291276
38:276388
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Red shading represents ancestral node of Sphingidae clade Blue shading represents ancestral node of the Saturniidae clade

Table S3. Ancestral state condition and the corresponding confidence interval at each node.