

1 A new species of deep-water *Lethrinops* (Cichlidae) from Lake Malawi.

2 George F. Turner

3 School of Natural Sciences, Bangor University, Bangor, Gwynedd LL57 2UW, UK  
4 & Vertebrates Division, Natural History Museum, Cromwell Road, London SW7, U.K.  
5 email [bss608@bangor.ac.uk](mailto:bss608@bangor.ac.uk), Orcid: 0000-0003-0099-7261  
6  
7

8 **Abstract**

9 A new species of cichlid fish, *Lethrinops atrilabris* is described from specimens collected by  
10 trawling at a depth of around 90m off Monkey Bay, southern Lake Malawi. It is assigned to  
11 the genus *Lethrinops* on the basis of its vertical flank barring, lack of enlarged cephalic lateral  
12 line canal pores and the form of the lower jaw dental arcade. It can be distinguished from  
13 congeneric species by its male breeding dress of contrasting flank barring and dark ventral  
14 surface, most strikingly on the lips, throat and chest, its relatively small known maximum  
15 size (<75mm SL), large eyes (38-41% head length), laterally compressed body (depth 2.5-2.7  
16 times max head width) and lower gillraker count (13-14).

17 Keywords: New species, Lake Malawi, cichlidae, *Lethrinops*  
18

19 1. INTRODUCTION

20 Lake Malawi hosts an enormous number of endemic cichlid fishes, in one recent guide,  
21 estimated to be over 800 species (Konings, 2016). Although this extraordinary adaptive  
22 radiation is of great interest to evolutionary biologists, conservationists, fishing communities  
23 and aquarium fish enthusiasts, the rate of species description is slow and many species – even  
24 some well-known ones - remain undescribed, rendering them ineligible to receive IUCN  
25 redlisting, or incorporation into standard reference systems such as FishBase, GBIF etc.

26 The genus *Lethrinops*, as currently understood, comprises 22 described and many  
27 undescribed species of sediment-sifting cichlid fishes endemic to the Lake Malawi and  
28 Upper/Middle Shire River catchments. They are characterised by the shape of their lower jaw  
29 dental arcade, in which the outer row curves in posteriorly to end abruptly behind the inner  
30 row(s), if present (Trewavas 1931, Turner 1996, Ngatunga & Snoeks 2004). In the majority  
31 of Malawi endemics, the outer tooth row continues in a relatively straight line, often  
32 dwindling to a few small widely-spaced teeth: referred to as the ‘Haplochromis’-style, as  
33 many of these species were formerly assigned to the genus *Haplochromis*. Two other genera  
34 of Lake Malawi cichlids share the ‘*Lethrinops*-type’ dentition and were separated from  
35 *Lethrinops* by Eccles & Trewavas (1989) on the basis of shared derived characters: the large  
36 long-snouted *Taeniolethrinops* species were reported to have an oblique-striped flank pattern,  
37 while the small short-snouted *Tramitichromis* have a distinctive lower pharyngeal bone shape  
38 and few widely-spaced gillrakers. *Lethrinops*-style dentition is also shown by *Ctenopharynx*  
39 *pictus*, which is placed in its genus on the basis of traits shared with the other two  
40 *Ctenopharynx* species. Therefore, the genus *Lethrinops* is currently defined by a single trait  
41 that appears to have evolved repeatedly and by the absence of other presumed derived traits.  
42 Perhaps not surprisingly, it has been proposed that it is comprised of two or more groups of

43 species that are not particularly closely-related that can be roughly characterised as ‘shallow-  
44 water’ and ‘deep-water’ species, comprised of 12 and 10 described species respectively  
45 (Ngatunga & Snoeks 2004).

46 The aim of the present study is to formally describe an additional deep-water species  
47 conforming to the current definition of the genus *Lethrinops* Regan 1922 (by Eccles &  
48 Trewavas 1989), known informally as *Lethrinops* ‘black chin’ (Turner, 1996).

49

## 50 2. MATERIALS AND METHODS

51 Specimens were obtained from a research trawl survey carried out by the Monkey Bay  
52 Fisheries Research Station (now known as the Fisheries Research Unit, FRU) of the Malawi  
53 Government, using the trawler Ethelwynn Trewavas, in 1992, intended to estimate standing  
54 stocks of food fishes. The majority of the catch was sold for human consumption, but on this  
55 occasion, a few specimens were preserved for research. These were already dead when  
56 selected and were pinned and photographed before being preserved in formalin, later being  
57 washed and transferred to 70% ethanol for long-term preservation. Counts and measurements  
58 were carried out following the methods of Snoeks (2004), using digital callipers and a low  
59 power magnifying desk lamp and various eye pieces (loupes).

60 Comparison with similar species was based on published (re-)descriptions, largely in Eccles  
61 & Trewavas (1989) for *Lethrinops* and *Ctenochromis* and Hanssens (2004) for  
62 *Placidochromis* along with re-examination of some of the type material, along with  
63 specimens held at Bangor University collected from 1990-2017. Direct comparisons of  
64 morphometric ratios were avoided as diagnoses resulting from generally small samples of  
65 type specimens rarely persist when larger numbers of specimens are examined, particularly  
66 when representing a fuller size range. In the author’s experience differentiation of Lake  
67 Malawi haplochromines is better achieved by overall appearance, aided by verbal  
68 descriptions in combination with meristics, character states such as dentition and male  
69 breeding dress.

### 70 Ethical Statement

71 The study did not involve live animals, as it used preserved specimens that were collected  
72 already dead from a trawl catch carried out as part of a Malawi Government research survey.

73

## 74 3. RESULTS

### 75 *Lethrinops atrilabris* sp. nov.

76 urn:lsid:zoobank.org:pub:101AB870-D407-416D-85F7-D61D73714064

77

78 Holotype: BMNH 2022.4.20.1, male, 72.0mm SL, collected from trawl catch NE of Monkey  
79 Bay, at a reported depth of 84-94m, 13<sup>th</sup> April 1992.

80 Paratypes: BMNH 2022.4.20.2-7, six males 66.2-72.9mm SL, collected with holotype.

81 Diagnosis: the lower jaw dentition ‘*Lethrinops*-type’. Mature males with a melanic pattern of  
82 strongly contrasting dark vertical flank bars on a pale background, and a dark area on the  
83 jaws and the underside of the head and chest. In addition, the species can be identified by its  
84 relatively small adult body size (not known to exceed 73mm SL), large eye, short, rounded  
85 snout, ventrally-placed mouth, 13-14 ceratobranchial gill rakers and laterally compressed  
86 body.

87 Comparisons: The male’s melanic pattern of strongly contrasting vertical flank bars is not  
88 exhibited by any known species of *Ctenochromis*, *Taeniolethrinops* or *Tramitichromis*.  
89 Among the described *Lethrinops* species, males of the shallow-water group (sensu Ngatunga  
90 & Snoeks 2004) do not show such strong vertical flank barring and tend to be less deep-  
91 bodied and laterally compressed and confined to shallower water (generally <50m, compared  
92 to 84-94m for *L. atrilabris*). This group comprises *Lethrinops albus* Regan 1922, *Lethrinops*  
93 *auritus* (Regan 1922), *Lethrinops furcifer* Trewavas 1931, *Lethrinops lethrinus* (Günther  
94 1893), *Lethrinops leptodon* Regan 1922, *Lethrinops lunaris* Trewavas 1931, *Lethrinops*  
95 *macrochir* (Regan 1922), *Lethrinops macrophthalmus* (Boulenger 1908), *Lethrinops*  
96 *marginatus* Ahl 1927, *Lethrinops microstoma* Trewavas 1931, *Lethrinops parvidens*  
97 Trewavas 1931, *Lethrinops turneri* Ngatunga & Snoeks 2003 and a number of undescribed  
98 species. Among the remaining, ‘deep-water’ *Lethrinops* species are 10 described species.  
99 *Lethrinops atrilabris* has a greater number of lower gillrakers (13-14) than *Lethrinops*  
100 *christyi* Trewavas 1931 (8-9), *Lethrinops longipinnis* Eccles & Lewis 1978 (9-10) and  
101 *Lethrinops altus* Trewavas 1931 (10-11). These three species can further be distinguished by  
102 their head and jaw shape: *L. christyi* has small pointed jaws and concave upper profile of  
103 snout v larger jaws set low on a rounded head profile in *L. atrilabris*; *L. longipinnis* has a  
104 much longer snout; *L. altus* has hooked maxillae, showing a markedly curved lower profile,  
105 in contrast to the straight maxillae in *L. atrilabris*. *Lethrinops atrilabris* has fewer lower  
106 gillrakers (13-14) than *Lethrinops micrentodon* (Regan 1922) (15-19), *Lethrinops gossei*  
107 Burgess & Axelrod 1973 (18-19), *Lethrinops stridei* Eccles & Lewis 1977 (19-23),  
108 *Lethrinops macracanthus* Trewavas 1931 (21-24) and *Lethrinops microdon* Eccles & Lewis  
109 1977 (24-29). *Lethrinops mylodon* Eccles & Lewis 1979 generally has fewer lower gillrakers  
110 (10-14 v 13-14 in *L. atrilabris*) and also differs in having a very heavily-built lower  
111 pharyngeal bone with stout molariform teeth (v lightly-built, with small slender teeth in *L.*  
112 *atrilabris*) and in attaining a much larger size (>200mm SL v <80 mm SL in *L. atrilabris*).  
113 *Lethrinops longimanus* Trewavas 1931 generally has a higher count of lower gillrakers:15-19  
114 according to Eccles & Lewis 1979, although Eccles & Trewavas (1989) give 14 as the lower  
115 limit, v 13-14 in *L. atrilabris*. *Lethrinops longimanus* can also be distinguished by its larger  
116 maximum size (150mm SL v <80mm SL) and male breeding dress of a bronze colour,  
117 weakly barred v the strongly barred black and silver of *L. atrilabris*.

118 The dental arcade trait can be difficult to see without a powerful microscope and appropriate  
119 lighting, so this trait is of little use to fieldworkers. Other deep-bodied, deep-water species  
120 with similar barred patterns are presently classed in the genera *Alticorpus*, *Aulonocara* and  
121 *Placidochromis*. Members of the first two genera are distinguished by having much larger  
122 cephalic lateral line pores, particularly on the underside of the head, than other Malawian  
123 cichlids, including *Lethrinops*. Distinguishing *Placidochromis* species can be more  
124 problematic, as these lack this diagnostic trait. A number of deep-water species were  
125 described by Hanssens in 2004, several superficially resembling *L. atrilabris*. From these, *L.*

126 *atrilabris* can be distinguished by its lower-arch gillraker counts (13-14), which are lower  
127 than those of *Placidochromis chilolae* Hanssens 2004 (14-16), *Placidochromis lukomae*  
128 Hanssens 2004 (14-18), *Placidochromis nigribarbis* Hanssens 2004 (16-18), *Placidochromis*  
129 *obscurus* Hanssens 2004 (18-21) and higher than *Placidochromis domirae* Hanssens 2004 (8-  
130 9), *Placidochromis koningsi* Hanssens 2004 (10), *Placidochromis msakae* Hanssens 2004  
131 (12), *Placidochromis pallidus* Hanssens 2004 (11-12), *Placidochromis rotundifrons*  
132 Hanssens 2004 (11) and *Placidochromis turneri* Hanssens 2004 (9-10). Other species in the  
133 genus can be differentiated quite readily on physical appearance, such as having a shallower  
134 body, smaller eyes, a longer, more pointed snout, larger jaws or a mouth in a more terminal  
135 position or more upwardly-angled (see illustrations in Hanssens 2004 or Konings 2016).

#### 136 Description

137 Body measurements and counts in table 1. *Lethrinops atrilabris* is a small (<80mm SL)  
138 laterally-compressed (maximum body depth 2.5-2.7 times maximum head width) cichlid fish  
139 with a short, rounded snout (27-32% HL), small mouth low down on the head and very large  
140 eyes (38-41% HL). To date, only mature males have been identified and these have  
141 conspicuously barred flanks and a black underside to the head and chest (Figure 1).

142 The size range of the seven specimens is 66-73mm SL. As all specimens collected showed  
143 clear evidence of male breeding dress, it can be assumed that all are adult males, probably  
144 collected on a breeding ground. In haplochromine cichlids, the largest males are typically  
145 larger than the largest females, and there is not usually a great deal of variation in the size of  
146 adult males on breeding grounds. As the specimens were collected from an unselective trawl  
147 catch along with many much larger individuals of other species, it seems likely that the  
148 maximum adult size of this species is less than 80mm SL, at least in the SE Arm of the lake.

149 All specimens relatively deep-bodied, laterally compressed, deepest part of body generally  
150 well behind first dorsal fin spine. Anterior upper lateral profile convex and gently curving,  
151 without a sharp inflection in curve above the eye. Lower anterior lateral profile also gently  
152 curving, so that tip of snout lies well above insertion of pelvic fins. Mouth relatively small,  
153 low on head, slightly upwardly-angled, snout well below horizontal plane from bottom of  
154 eye. Eye extremely large, circular, generally appearing more or less touching anterior upper  
155 lateral head profile. Lachrymal much wider than deep, 5 openings.

156 Flank scales weakly ctenoid, cteni becoming reduced dorsally, particularly anteriorly above  
157 upper lateral line, where they transition into a cycloid state. Scales on chest are relatively  
158 large, gradually transitioning in size from larger flank scales, as is typical in non-mbuna  
159 Malawian endemic haplochromines (Eccles & Trewavas 1989). A few small scales scattered  
160 on the proximal part of caudal fin.

161 Cephalic lateral line pores inconspicuous, flank lateral line shows the usual cichlid pattern of  
162 separate upper and lower portions.

163 Pectoral fin very long when intact, extending well past first anal spine. Pelvic fins extend past  
164 vent in all specimens and past first anal spine in some: this may be a sexually dimorphic trait,  
165 with female haplochromines often having shorter pelvic fins. Tips of dorsal and anal fins also  
166 prolonged, extending well past the plane through base of caudal fin in some specimens- again  
167 probably a sexually dimorphic trait, exaggerated in males. Tailfin crescentic.

168 Lower jaw relatively small, with thin mandibular bones, but not flattened as it is in some  
169 *Placidochromis*, such as *P. hennydaviesae*. Jaw teeth small, short and erect. Outer series in  
170 both upper and lower jaw largely unequally bicuspid, becoming more equally bicuspid  
171 posteriorly, notably in upper jaw. A single inner series of very small tricuspid teeth.

172 Lower pharyngeal bone small, lightly-built, Y-shaped, and carries small, short, laterally  
173 compressed slightly hooked, blunt, simple teeth. Middle-lying 5-6 teeth on each side of  
174 posterior row slightly larger than others, but molarization lacking. About 12 teeth in midline  
175 row and about 20 on each side on posterior row. Gill rakers simple, erect, fairly long and  
176 well-spaced, with few, if any, reduced to small stubs near anterior part of arch.

177 Colouration of females and immatures is unknown, but from experience of other species from  
178 this habitat, can be expected to be countershaded, sandy-coloured dorsally, with silvery  
179 flanks and probably faint vertical flank bars. All known specimens appear to be males in  
180 breeding dress. Colour notes based on a photograph of a freshly collected type specimen and  
181 an additional specimen collected in 2016, but not yet located in the collection at Cambridge  
182 University (Figure 2). Strong dark brownish vertical flank bars on silvery-white background:  
183 6 bars under dorsal fin, 2 more on caudal peduncle and 1-2 on nape. Head dark brown on  
184 upper surface, but paler laterally, sometimes with a dark lachrymal mark running from eye  
185 toward the mouth. Eye golden brown, darker along the axis of lachrymal stripe. Lips, lower  
186 jaw, throat and chest are black. Dorsal fin dark golden-brown, with series of irregular white  
187 spots or oblique stripes angled forwards from base, with broad black margin and broader  
188 white submarginal band. Pectoral fins translucent, but brownish-tinted. Pelvic fins black,  
189 fading to dark grey on posterior rays. Anal fin black, fading to dark grey basally and marked  
190 with irregular yellowish spots and stripes. Caudal fin with dark grey to black upper and lower  
191 margins, but otherwise dark golden-brown with three thin irregular vertical white bands.

## 192 Distribution

193 Positively known only from the type locality, in the SE Arm of Lake Malawi, NE of Monkey  
194 Bay, at a reported depth of 84-94m. A photograph of a possible specimen of this species was  
195 taken from a trawl catch at 95-105m East of Domwe Island, SE Arm, 4th March 2016. These  
196 two localities are close together, as are the depths.

## 197 Etymology

198 ‘Atri-‘ from plural of the adjective ‘ater’ (Latin) = black + ‘labris’ from plural of labrum  
199 (Latin)= lip, in reference to the black lips of the males in breeding dress.

200

201

202 TABLE 1. Morphometric and meristic characters of *Lethrinops atrilabris*.

	<b>Holotype</b>	<b>Paratypes (n=6) mean &amp; range</b>
Standard Length (mm)	72.0	69.2 (66.2-72.9)
<b>As % SL</b>		
Maximum Body Depth	38.6	39.2 (38.1-39.8)
Head Length	32.9	32.9 (32.1-33.6)
Dorsal-Fin Base Length	57.2	57.0 (53.7-58.8)
Anal-Fin Base Length	18.8	17.4 (16.7-18.4)
Predorsal Length	39.6	37.5 (36.6-38.1)
Preanal Length	64.4	66.5 (65.4-69.2)
Prepectoral Length	32.8	33.9 (32.3-34.8)
Prepelvic Length	39.9	39.8 (38.1-41.5)
Caudal-Peduncle Length	16.7	16.2 (15.6-16.9)
Caudal-Peduncle Depth	12.1	11.9 (11.6-12.2)
<b>As % Head Length</b>		
Head Width	44.7	46.1 (45.0-47.5)
Interorbital Width	22.8	23.9 (22.1-27.4)
Snout Length	32.1	29.1 (26.7-30.4)
Lower Jaw Length	39.2	39.0 (37.2-41.1)
Premaxillary Pedicel Length	27.0	25.3 (24.2-26.1)
Cheek Depth	16.9	17.3 (16.6-18.2)
Eye Diameter	40.9	39.8 (38.3-40.8)
Lachrymal Depth	21.1	21.1 (20.4-22.9)
<b>Ratios</b>		
Body Depth/Head Width	2.62	2.58 (2.51-2.67)
Caudal-Peduncle Length/Depth	1.38	1.36 (1.30-1.43)
<b>Counts</b>		
	<b>Holotype</b>	<b>Paratypes range</b>
Upper Gillrakers	5	4-5
Lower Gillrakers	14	13-14
Dorsal Fin Rays	XVI, 9	XV-XVI, 9-10
Anal Fin Rays	III, 7	III, 7-9
Longitudinal Line Scales	32	31-34
Cheek Scales	2	2

203

204

205 4. DISCUSSION

206 The cichlid genus *Lethrinops* is endemic to Lake Malawi and its catchment and the  
 207 outflowing Shire River, its expansion in Lake Malombe and continuation to the  
 208 biogeographic barrier represented by the falls on the middle Shire, notably the Kapichira  
 209 rapids, below which the fish fauna is essentially lower Zambezian (Tweddle & Willoughby  
 210 1979). Originally defined by Regan (1922) based on its dentition- principally in having small,

211 weak teeth in narrow bands- the genus originally included just 4 species, including the type *L.*  
212 *lethrinus*. Trewavas (1931) revised the genus, her definition emphasising the semicircular  
213 shape of the lower jaw dental arcade, and increasing the number of included species to 23.  
214 The revision by Eccles & Trewavas (1989) split the genus into three. Five small, short-  
215 snouted species were moved into *Tramitichromis*, characterised by the shape of the lower  
216 pharyngeal bone, in which the upper margin of the blade is turned sharply downwards and the  
217 anterior end of the pharyngeal dental arcade is broad and rounded. In addition, four large,  
218 long-snouted species were grouped into *Taeniolethrinops*, characterised by having an oblique  
219 dark stripe on the flanks of females and immature fishes (although not all species actually  
220 seem to show this in my experience). Thus, *Lethrinops* was left without any defining  
221 synapomorphy: characterised by its dental arcade- shared with *Tramitichromis* and  
222 *Taeniolethrinops*- but lacking the diagnostic traits of the latter two genera.

223 Early molecular studies using mitochondrial DNA restriction fragment analyses placed the  
224 deep-water *Lethrinops gossei* in a surprising grouping with the mbuna species, along with a  
225 number of *Aulonocara* species, and not with the major ‘Haplochromis’ or ‘sand-dweller’  
226 group from sandy or muddy habitats (Moran et al. 1994). However, later studies placed a  
227 number of shallow-water *Lethrinops* and a *Taeniolethrinops* species in the ‘sand-dweller’  
228 group, suggesting the genus to be polyphyletic (Joyce et al. 2011, Genner & Turner 2012). In  
229 addition, the deep-water species were shown to have an affinity with *Alticorpus* and some  
230 deep-water *Placidochromis* species. Early nuclear gene analyses presented rather inconsistent  
231 pictures, but whole genome sequencing (Malinsky et al. 2018; Masonick et al. 2022) has  
232 continued to support the distinctness of the deep-water and shallow-water *Lethrinops* species,  
233 and the affinity of the former to *Aulonocara* and *Alticorpus* (deep-water *Placidochromis* were  
234 not investigated).

235 On the basis of the emerging mitochondrial data, Ngatunga and Snoeks (2004) informally  
236 split the genus into deep-water and shallow-water groups, with the type species, *Lethrinops*  
237 *lethrinus* clearly a member of the latter, suggesting that the deep-water species will be in need  
238 of a new generic classification. However, this has yet to be attempted and at present the  
239 distinction is unclear.

240 Generally, the deep-water species mostly occur at depths of 50m or more and seem to be  
241 relatively deep-bodied and laterally compressed. Males in breeding dress tend to express  
242 strong vertical barring on their flanks, as do species of *Alticorpus*, *Aulonocara* and  
243 *Placidochromis* from the same habitat, while shallow-water *Lethrinops* males are usually  
244 unbarred or weakly-barrred with a range of bright colours including red, orange, yellow, blue  
245 and green: see illustrations in Konings (2016), for example. A few species, such as *L. altus*,  
246 *L. christyi*, *L. longimanus*, *L. longipinnis* and *L. micrentodon* are more problematic, with  
247 forms exhibiting a mix of traits, and often being found at depths of 20-60m. However,  
248 *Lethrinops atrilabris* is unambiguously a member of the deep-water group, with its strongly  
249 barred males and relatively deep, laterally compressed body. The species shows superficial  
250 similarities to a number of species of the genus *Placidochromis*, which also includes a  
251 number of deep-water, vertically-barrred species. From these, it can be distinguished by the  
252 shape of the lower jaw dental arcade (Hanssens 2004). However, it is not clear whether this  
253 trait really has much phylogenetic significance: this will probably require extensive whole  
254 genome sequencing and phylogenetic analysis.

255 An additional case of evolution of the *Lethrinops*-style dentition appears to have occurred in  
256 *Ctenopharynx pictus*, which, like known species of *Lethrinops*, *Taeniolethrinops* and  
257 *Tramitichromis*, is a sediment-sifting species. Eccles & Trewavas (1989) placed this species  
258 in *Ctenopharynx* on the basis of its spotted melanin pattern, large number of gillrakers and  
259 ‘weak’ jaws and dentition. This classification is supported by recent genome-wide analysis  
260 (Masonick et al. 2022), although the specimen is mistakenly labelled as ‘*Otopharynx pictus*’,  
261 possibly following a period of usage of *Ctenopharynx* as a subgenus of *Otopharynx* in some  
262 publications in the 1990s (e.g. Konings 1990).

263

## 264 ACKNOWLEDGEMENTS

265 I am grateful to all my colleagues and collaborators in Malawi during the late 1980s and early  
266 1990s who helped me with collecting and identifying the fishes we sampled during the ODA  
267 (now DFID)-funded Lidole Project and the FAO Chambo Fisheries Project, carried out in  
268 collaboration with the Malawi Government Fisheries Research Unit at Monkey Bay, and I  
269 thank James Maclaine at the Natural History Museum in London for curating the specimens.

270

## 271 REFERENCES

272 Eccles, D.H. & Lewis, D. S. C. (1979). A taxonomic study of the genus *Lethrinops* Part 3.  
273 *Ichthyological Bulletin of Rhodes University* **38**, 1-25.

274

275 Eccles, D. H. & Trewavas, E. (1989). *Malawian cichlid fishes. The classification of some*  
276 *Haplochromine genera*. Lake Fish Movies, Herten, Germany, 335 pp.

277

278 Genner, M. J. & Turner, G. F. (2012). Ancient hybridisation and phenotypic novelty within  
279 Lake Malawi’s cichlid fish radiation. *Molecular Biology and Evolution* **29**, 195-206.

280

281 Hanssens, M. (2004). The deep-water *Placidochromis*. Snoeks, J. (ed) *The Cichlid Diversity*  
282 *of Lake Malawi/Nyasa/Niassa: Identification, Distribution and Taxonomy*. Cichlid Press, El  
283 Paso, TX, 104-197.

284

285 Joyce, D. A., Lunt, D. H., Genner, M. J., Turner, G. F., Bills, R. & Seehausen, O. (2011).  
286 Repeated colonization and hybridization in Lake Malawi cichlids. *Current Biology* **21**, R108-  
287 109.

288

289 Konings, A. (1990) *Konings’s Book of Cichlids and All The Other Fishes of Lake Malawi*.  
290 TFH Publications, Neptune City, NJ.

291

292 Konings, A. (2016). *Lake Malawi Cichlids in their Natural Habitat. 5<sup>th</sup> Edn*. Cichlid Press, El  
293 Paso TX.

294

295 Malinsky, M., Svardal, H., Tyers, A. M., Miska, E.A., Genner, M. J. Turner, G. F. & Durbin,  
296 R. (2018). Whole genome sequences of Malawi cichlids reveal multiple radiations  
297 interconnected by gene flow. *Nature Ecology & Evolution* **2**, 1940-1955.

298



- 299 Masonick, P., Meyer, A. & Hulsey, C. D. (2022) Phylogenomic analyses show repeated  
300 evolution of hypertrophied lips among Lake Malawi cichlid fishes. *Genome Biology &*  
301 *Evolution* **14**, evac051.  
302
- 303 Moran, P., Kornfield, I. & Reinthal, P. N. (1994). Molecular systematics and radiation of the  
304 haplochromine cichlids (Teleostei: Perciformes) of Lake Malawi. *Copeia* **1994**, 274-288.  
305
- 306 Ngatunga, B. P. & Snoeks, J. (2004). Key to the shallow water *Lethrinops sensu lato*. Snoeks,  
307 J. (ed) *The Cichlid Diversity of Lake Malawi/Nyasa/Niassa: Identification, Distribution and*  
308 *Taxonomy*. Cichlid Press, El Paso, TX, 252-260.  
309
- 310 Regan, C. T. (1922). The cichlid fishes of Lake Nyasa. *Proceedings of the Zoological Society*  
311 *of London* **1921**, 675-727.  
312
- 313 Snoeks, J. (2004). Materials and Methods. In Snoeks, J. (ed) *The Cichlid Diversity of Lake*  
314 *Malawi/Nyasa/Niassa: Identification, Distribution and Taxonomy*. Cichlid Press, El Paso,  
315 TX: 12-19.  
316
- 317 Trewavas, E. (1931). A revision of the cichlid fishes of the genus *Lethrinops*. *Annals and*  
318 *Magazine of Natural History* (**10**) **7**, 133-153.  
319
- 320 Trewavas, E. (1935). A synopsis of the cichlid fishes of Lake Nyasa. *Annals and Magazine of*  
321 *Natural History* (**10**) **16**, 65-118.  
322
- 323 Turner, G. F. (1996). *Offshore Cichlids of Lake Malawi*. Cichlid Press, Lauenau. 240 pp.  
324
- 325 Tweddle, D. & Willoughby, N. G. (1979). The nature of the barrier separating the Lake  
326 Malawi and Zambezi fish faunas. *Ichthyological Bulletin of Rhodes University* **39**, 1-9.  
327  
328

329



330

331

332

333

Figure 1: *Lethrinops atrilabris* sp. nov. Above: holotype: BMNH 2022.4.20.1, male, 72mm SL, collected from trawl catch NE of Monkey Bay, at a reported depth of 84-94m, 13<sup>th</sup> April 1992; Below: the full type series, holotype labelled 1, collecting information as holotype.



334



335

336

337 Figure 2: *Lethrinops atrilabris* sp. nov. Fresh coloration. Above: one of the type specimens  
338 photographed shortly after capture. Below: probable *L. atrilabris*, collected from trawl catch  
339 at 95-105m depth, East of Domwe Island, SE Arm, 4th March 2016. Cambridge University  
340 collection, identification not confirmed.

341