

1 Distribution and genetic diversity of South Florida *Tephrosia* shed light on past cultural use

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22 Running Head: Genetics of *Tephrosia*

23

## 24 **Summary**

- 25 • The genus *Tephrosia* (Fabaceae), the hoary peas, contain high levels of rotenone, which  
26 has a long history of human use as a fish poison. We examine the distribution of  
27 *Tephrosia angustissima*, in South Florida to clarify patterns of genetic relatedness and  
28 shed light on human plant movement before European contact. Several populations of  
29 *Tephrosia angustissima* with a history of taxonomic uncertainty exist in South Florida  
30 and the neighboring Caribbean Islands.
- 31 • To clarify relationships in this group, and to elucidate the conservation status of  
32 populations in Everglades National Park and Big Cypress National Preserve, we used  
33 restriction site associated DNA sequencing (RAD-SEQ) on 94 samples from South  
34 Florida and three locations in southwest Puerto Rico.
- 35 • Analysis of variation in SNP markers by the Bayesian STRUCTURE algorithm and  
36 principal coordinate analysis both separated the samples into three groups. These three  
37 groups were likely separate colonization events of Florida. Genetic diversity is moderate  
38 in all of the groups, with only limited evidence of a bottleneck in some of the disjunct  
39 South Florida populations.
- 40 • Overall, the human association of this group is consistent with a history of human use,  
41 suggesting conservation efforts for these taxa should consider their pre-Columbian  
42 human associations.

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45 **Keywords:** Karst, fish poison, conservation genetics, Florida-Caribbean biogeography

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47 **Societal impact statement:**

48 A great many endangered plant taxa exhibit patterns of edaphic specialization, occurring on  
49 particular substrates such as karst or serpentine soils. Human activities, such as the construction  
50 of shell middens, can create edaphically unique substrates. In the Americas, post-Columbian  
51 land use changes coupled with extensive loss of indigenous cultural knowledge, has created areas  
52 where associations of cultivated plants with human-generated habitats may be lost. Here we use  
53 population genetic approaches to examine rare *Tephrosia* (hoary pea) taxa from South Florida, a  
54 group of plants that produce rotenone that has been used by many indigenous groups as a fish  
55 poison. We find evidence of multiple introductions from the broader Caribbean region and an  
56 association with anthropogenic habitats such as shell middens. In efforts to conserve rare hoary  
57 peas in Florida, an understanding of past use of the landscape by native Americans is essential.

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59

60 **Introduction:**

61 In an era of rapid global development, there has been a decline of indigenous land use practices.  
62 Across the Americas, indigenous land use radically altered landscapes, from the *Terra preta*  
63 “black soils” of central Amazonia to the North American great plains (e.g., Denevan 1992; Mann  
64 2005). With the decline of these land use practices over the past few centuries, many plants that  
65 had been cultivated as food, fuel, fiber, or medicine, now face a range of pressures on their  
66 populations. Although conservation often focuses on preserving unimpacted habitats, preserving  
67 anthropogenic habitats may be essential to their persistence.

68 Many tropical legumes have an indigenous history of human use as fish poisons and as  
69 traditional medicines and are now declining as fish poisoning is practiced less often or are  
70 outlawed. Several legumes produce rotenone, a type of isoflavanoid, that can be effective as fish  
71 poisons. When added to small water bodies, these compounds can stun fish, allowing for easy  
72 collection. In commercial applications, species of *Lonchocarpus* are most widely used, but  
73 species of *Pachyrhizus*, *Derris* and *Tephrosia* have similar chemical profiles. Several *Tephrosia*  
74 species from Asia, Africa, Australia as well as the tropical Americas have been noted as fish  
75 poisons, including the pantropical *Tephrosia purpurea* and African *T. vogelii* (Howe 1930,  
76 Chevalier 1937, Quigley 1956, Sauer, 1968). *T. sinapou* was a widely used fish poison by  
77 Amerindians in Guyana, although less so today now that fish poisoning is illegal and that  
78 traditional hunting practices are being lost (van Andel, 2000). This species also likely has  
79 traditional medicinal uses, and possibly was used as a soap, although these uses are not well-  
80 known (van Andel, 2000). Beyond fish poison, Speck noted that the Catawba people used *T.*  
81 *virginiana* to treat rheumatism (Speck 1937). Austin (2004) describes a variety of medicinal uses  
82 of *T. virginiana* by indigenous groups in the Southeastern US. (Austin, 2004).

83  
84 Narrowleaf hoary pea (*Tephrosia angustissima*) is a Florida endangered species with a very  
85 narrow distribution restricted to southern Florida and Cuba. The flowers of *T. angustissima* have  
86 glabrous styles, which distinguishes this species from all other members of the genus in Florida.  
87 Three varieties of *T. angustissima* are currently considered to have occurred in Florida. Two  
88 varieties, *T. a. var. angustissima* and *T. a. var. corallicola* occurred in pine rockland habitat of  
89 Miami-Dade County, although *T. a. var. angustissima* is currently considered extinct (Gann et al.  
90 2002). The third variety, *T. a. var. curtissii*, is known from coastal strand habitat along the east

91 coast of Florida from Volusia County south to Miami-Dade County. In addition to southern  
92 Florida, *T. a.* var. *corallicola* has also been reported in western Cuba (Beyra Matos 1998). Based  
93 on a single Florida specimen collected in 1919, an additional glabrous-styled species was  
94 described as *T. seminole* (Shinners 1961). That specimen was collected from Godden’s Mission,  
95 an early European mission focused on converting Native Americans to Christianity that would  
96 have had an indigenous community associated with it. Godden’s Mission is believed to have  
97 been situated in eastern Hendry County; the specimen label described the plant as growing “on  
98 prairies” (Isely 1981, 1982). The herbarium labels for the Godden’s mission *Tephrosia* said it  
99 was used to treat nosebleeds and other maladies. A subsequent taxonomic treatment of *Tephrosia*  
100 included *T. seminole* as a synonym for *T. a.* var. *curtissii* (Isely 1981).

101 All three varieties of *T. angustissima* are extremely rare. In the wild, *T. a.* var. *curtissii* is the  
102 most widely distributed variety with several extant populations known along the eastern coast of  
103 South Florida and with a recent population estimate of 2000 plants (Wendelberger 2010,  
104 Wendelberger and Maschinski 2016). A single natural population of *T. a.* var. *corallicola* is  
105 currently known in Miami-Dade County. In addition, two populations of this variety were  
106 introduced on conservation land in close proximity to the natural population by staff at Fairchild  
107 Tropical Botanic Garden with their partners in the Miami-Dade County Environmentally  
108 Endangered Lands (EEL) Program (Wendelberger 2010, Wendelberger and Maschinski 2016,  
109 Possley et al. 2022).

110 The taxonomic uncertainty surrounding *Tephrosia* also limits our understanding of its  
111 historical population size and its human use. Although once more abundant, perhaps due to  
112 indigenous human use, the species may have experienced a reduction in population size in the  
113 past. It is likely that *Tephrosia angustissima sensu lato* colonized Florida from geologically  
114 older and higher land in the Caribbean. Known as a prized fish poison to Native Americans,  
115 populations may have been transported to Florida from the Caribbean instead of or in addition to  
116 natural dispersal. Irrespective of how they were introduced, populations in Florida likely  
117 underwent a severe population bottleneck upon arrival that led to reductions in population size  
118 and low genetic diversity. The loss of genetic diversity would limit the capacity of the Florida  
119 populations to respond to subsequent environmental changes, such as sea level rise and the  
120 introduction of invasive species.

121 Here we have developed genetic markers to resolve the relationships of different taxonomic  
122 groups of *Tephrosia* to one another and provide insight into whether the distribution of groups  
123 suggests possible past human movement of this group. Resolving taxonomic groups also allows  
124 us to define management units that are genetically similar. Furthermore, we aimed to elucidate  
125 historical demographic and migration patterns to better understand the likely consequences of  
126 ongoing population loss and the effects of potential management actions.

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## 130 **Methods**

131 *Biological materials:* We first developed a list of plants to collect from a variety of distinct  
132 locations (Figure 1). Our collection sites included all populations of putative *Tephrosia*  
133 *angustissima* in South Florida, as well as *T. florida*, which occurs more widely in South Florida.  
134 We collected from Russell Key in Everglades National Park (a site linked to the Calusa people)  
135 and three sites within the Big Cypress National Preserve, including a location close to the likely  
136 site of Godden's mission visited by Europeans and Seminoles. In Miami Dade County, we  
137 sampled a small population of *T. a. var. curtissii* from Haulover Park and a population of *T. a.*  
138 *var. corallicola* from Chapman Field, a USDA Agricultural Research Service Station.

139 Accessions from Chapman Field are also held as an *ex situ* collection at Fairchild Tropical  
140 Botanic Garden and were used for introductions at two nearby Miami-Dade County preserves:  
141 Ludlam Pineland and the Deering Estate. At Ludlam Pineland we collected wild *T. florida* to  
142 serve as an outgroup for the clustering analysis. We were also able to obtain roadside samples of  
143 *T. cinerea* from a collecting trip to southwestern Puerto Rico in 2017. Material was obtained  
144 from three localities in Municipalidad Cabo Rojo: two more montane sites [**Sierra Bermeja:**  
145 Cabo Rojo NWR “Cerro Mariquita”, 26 Jan 2016, *Lange 16* (MAPR); ‘Finca Escabi’, 27 Jan  
146 2016, *Lange 17* (MAPR); **El Conuco:** ‘Upper Rancho Hugo’, 28 Jan 2016, *Lange 19 w/ Possley*  
147 (FTG); El Conuco ‘Finca Solins’, 28 Jan 2016, *Lange 20* (FTG)] that are fairly close together  
148 and similar and a third along road 301 near the **Cabo Rojo** lighthouse [28 Jan 2016, *Lange 18*  
149 (FTG)]. ).

150 For sequencing, we used a slight variation on the RADseq technique, double digest RAD  
151 (ddRAD), which achieves more reproducible results with as little as 100 ng of genomic DNA

152 (Peterson et al. 2012). DNA from leaf tissues was extracted from all collected specimens using a  
153 Qiagen DNeasy kit (Germantown, MD, USA) and extraction yields were assessed  
154 fluorometrically. The 94 samples with the highest quality DNA, representing at least 8 samples  
155 from all nine *Tephrosia* locations, were processed for restriction-site associated DNA  
156 sequencing. High quality genomic DNA (300 ng) was digested with two restriction enzymes  
157 simultaneously. Adapter sequences, P1 (containing a unique barcode and PCR primer site) and  
158 P2 (containing a PCR primer site) were ligated to the overhanging ends created by the restriction  
159 enzymes. Fragments of the appropriate size for sequencing (~300 bp) were selected using a  
160 Pippin Prep kit (Sage Science), followed by PCR amplification, during which index sequences  
161 were added. All samples were sequenced on an Illumina HiSeq 2500 Sequencing System at the  
162 UC Davis Genome Center. ddRAD sequencing reads underwent quality assessment and SNPs,  
163 genotypes, and haplotypes were called using the GATK software pipeline with recommended  
164 settings (McKenna et. al, 2010). We retained 6278 SNPS after filtering for minimal coverage  
165 (10 reads) and to remove singletons. Sequencing multiple samples from all nine locations gave  
166 us sufficient replication to compare levels of genetic variation in each location, and to infer  
167 historic population sizes. We decided against further sampling so as to not reduce sample sizes  
168 below a threshold where variation cannot be estimated.

169 We used Genalex 6.05 (Peakall and Smouse 2006, 2012) to calculate a range of population  
170 genetic statistics to understand patterns of variability and make demographic inferences.  
171 Estimates of expected and observed heterozygosity as well as deviations from Hardy-Weinberg  
172 equilibrium genotype frequencies were calculated to establish hypotheses concerning  
173 relationships between population size, genetic structure, and clonal propagation (e.g., Gravuer et  
174 al., 2005). The Haulover population was dropped from these analyses for a small population  
175 size. Analysis of molecular variance (AMOVA, Excoffier et al 1992) was performed to partition  
176 molecular variation among species, varieties, including variation within and between  
177 populations. STRUCTURE, a clustering algorithm, was used to assign individuals to populations  
178 and to identify admixture between populations (Pritchard et al 2000, Evanno et al 2005). A  
179 principal coordinate analysis was used as an alternative clustering approach, which was  
180 implemented in Genalex 6.05 (Peakall and Smouse 20s06, 2012). These two analyses are critical  
181 to defining management units, and circumscribing areas and genotypes that are compatible for  
182 reintroductions or augmentations without risk of hybridization above which occurred in the past.

183 A Treemix analysis (Pickrell and Pritchard, 2012) was performed to detect introgression among  
184 different groups. The relationship between allelic richness and gene diversity, and patterns of  
185 linkage among loci were examined for signs of bottlenecks and multiple colonizations in Florida.

186 We aimed to estimate current and past population sizes in two complementary ways. For  
187 effective population size, we used a program called NeCalculator2 (Do et al., 2014) to estimate  
188 effective population sizes based on the molecular co-ancestry method, since the linkage  
189 disequilibrium and heterozygote excess methods estimated infinite population sizes for these  
190 taxa. We estimated population size over the past thousands of years based on the site frequency  
191 spectrum, SFS (e.g., Ragsdale et al., 2018). RAD-seq. is an imperfect tool for calculating the  
192 absolute value of population size since the calculation depends on the number of invariant sites.  
193 This has always been an issue since that number is not really known with RAD-seq analyses  
194 (e.g., Gattepaille et al., 2013). In our processing steps, we identified sites that are segregating,  
195 but otherwise, we do not know if sites are not sequenced or if they are invariant). However, we  
196 can use the snp loci as “sites of interest”, providing us with 6278 segregating snps among 94  
197 samples.

## 198 **Results**

### 199 *Clustering algorithms*

200 We developed 6278 SNP markers with RAD-seq, after filtering. Both STRUCTURE and PcoA  
201 consistently identified three clusters in the material, consistent with three taxa (Figure 2). At a K  
202 of 3, STRUCTURE separates *T. florida* from Ludlam Pineland into one group, the Big Cypress  
203 location into a second cluster, and Russell Key, Chapman Field, and the Puerto Rican samples  
204 into a third cluster. As interpretation of population assignment can have value at nearby Ks, we  
205 also looked at K=2 and K=4. At K=2, Big Cypress is separated from Russell Key, Chapman  
206 Field, and Puerto Rico, with the Ludlam location admixed. At K=4 the same groupings occur,  
207 but the Florida Russell Key and Chapman field populations are separated from the Puerto Rican  
208 samples. Principal coordinate analysis (Figure 3) gave a similar result. A Treemix analysis  
209 (Figure 4) showed no sign of admixture among the three taxa.

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214 *Diversity metrics and differentiation among populations within taxa*

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216 Estimated standing diversity is similar across the 9 sampled locations, both as gene diversity  
217 ( $H_e$ ) and as Shannon's diversity index (Table 1). Percentage of polymorphic loci are similar  
218 across sampled locations, hovering around 80%, with the highest polymorphism rate in the one  
219 of the restored populations at Chapman Field.

220

221 Fixation index ( $F_{ST}$ ) was high among taxa (above 0.3) and low within taxa ( $< 0.1$ ),  
222 indicating recent divergence among taxa with low diversity within taxonomic groups.  
223 However,  $F_{ST}$  was slightly higher between the two Florida *T. angustissima* locations and the  
224 three Puerto Rican locations ( $\sim 0.01$ - $0.03$ , Supplemental Table 1). AMOVA partitioned 21% of  
225 variation among taxa (Supplemental Figure 1), and 78% within individuals, with none among  
226 populations. This is consistent with the low within taxa  $F_{ST}$  estimates. PcoA within taxa showed  
227 similar patterns, with little differentiation among the three Big Cypress populations.  
228 Nonetheless, PcoA did detect some differentiation among the two Florida *T. angustissima* and 3  
229 Puerto Rican *T. cinerea* populations, even when  $F_{ST}$  values were quite low.

230

231 *Effective population size and historical population size*

232 Estimates of effective population size from NeEstimator are given in Table 2. These estimates  
233 are low with confidence intervals that overlap or nearly overlap with zero. These estimates are  
234 generally low, consistent with population size declines over the past two centuries.

235 The Big Cypress population (Figure 5, Blue color, T1), shows a drop in  $N_e$  (effective  
236 population size) at about 7K years ago, steadying off at  $N_e = 800K$  (Figure 5). The Ludlam  
237 Pineland *T. florida* population (T3, red color) started decreasing about 12K years ago and has  
238 been decreasing since then, only steadying off about 200 years ago. The Russel Key/Chapman  
239 Field/Puerto Rican group shows higher and consistent population size (Black color, T2).  
240 Although the  $N_e$  values should not be interpreted as numerically precise, qualitatively they are  
241 consistent with population bottlenecks around the time arriving in Southern Florida since the last  
242 glacial maxima. The Puerto Rican samples (T2) did not converge and showed little shift in  
243 population size over the past several hundred thousand years.

244

245 **Discussion**

246 Our sampling provides some insight into the relationship of *Tephrosia angustissima sensu lato* in  
247 South Florida. Our analyses clearly indicate that plants from Big Cypress are quite distinct from  
248 populations in Russell Key and Chapman Field as well as those from Puerto Rico. The Russell  
249 Key site is a location associated with the Calusa people, one of the pre-Columbian groups of  
250 Florida. The Big Cypress sites are close to the historical location of Godden's mission, where  
251 trade and cultural exchange occurred between Europeans and the Seminole, a group that fled to  
252 South Florida after wars and persecution in the mid-19<sup>th</sup> century. Consequently, we recognize  
253 that the current taxonomic classification of *T. angustissima sensu lato* represents at least 2  
254 distinct taxonomic units. Due to DNA degradation issues, we were unable to get similar types of  
255 sequence data from herbarium samples. Therefore, with the apparent extinction of *T.*  
256 *angustissima* var. *angustissima*, we cannot directly address the subspecies question with  
257 complete confidence. However, we note that some of the differences could be due to  
258 environmental as well as genetic differences. The relatively low amount of differentiation  
259 between the Russell Key/Chapman field populations and the three Puerto Rican populations,  
260 relative to the more highly diverged Big Cypress population, are consistent with *Tephrosia*  
261 *seminole* being a distinct entity, and *Tephrosia angustissima* being part of a more widespread  
262 Caribbean species. These patterns are also consistent with multiple introductions of *Tephrosia* to  
263 South Florida, perhaps facilitated by separate groups of humans, although we have limited  
264 capacity to make inferences about that.

265 Without broader sampling of the Caribbean populations, in particular Cuba, determining the  
266 number of introductions of these taxa to Florida is not possible. Broader sampling efforts with  
267 voucher specimens are critical to unravel a more thorough demographical history of this group.  
268 We hope in the future such sampling is possible. We know that many taxa in South Florida have  
269 close relatives in Cuba, yet, at the very least our analysis suggest that the Russell Key location is  
270 genetically very similar to locations from southeast Florida Pine Rockland and coastal strand,  
271 which was unexpected. Our findings point to a potential single introduction of this taxa into  
272 Florida, that would have involved long distance dispersal within southern Florida. The presence  
273 of these taxa in both pine rocklands and coastal strands of the eastern coast of Florida and a shell  
274 mound in Southwestern Florida suggest a potential cultural link between the two regions.

275 Lastly, our results revealed very small effective population sizes for all *Tephrosia* locations  
276 sampled. In our collection sites, all of the sampled locations had fewer than 50 stems, indicative  
277 of very small populations that are widely isolated from one another. As a result, it is likely that  
278 these groups exhibit persistent small population sizes that are exacerbated by genetic bottlenecks  
279 during introduction to new locations. We argue that continued conservation of all these locations  
280 is essential especially for populations in unusual and threatened habitats, such as shell middens,  
281 which are of particular interest due to their disjunct distribution, human history, and unique  
282 adaptations to challenging environmental conditions. The Chapman Field site, which has been  
283 used as a seed source for reintroduction plantings, has high observed levels of diversity, and  
284 should be managed as an important germplasm repository for further reintroductions.

285 Native American populations in the study region reflect an unfortunate and sad history since  
286 European arrival, with near total annihilation of indigenous populations in Cuba, declines and  
287 significant cultural shifts in the Tequesta, Calusa and other groups that have been in South  
288 Florida since Spanish colonial times, and Miccosukee and Seminole groups that fled to the  
289 Everglades region following eviction and genocide during the Trail of Tears and Seminole wars  
290 of the mid-19<sup>th</sup> century (Wasserman, 2009). Written records from colonial times of the Tequesta  
291 and Calusa peoples that inhabited South Florida before European contact are limited, with most  
292 of the survivors of the groups perishing when evacuated to Cuba when Florida was transferred to  
293 British rule in 1763. Consequently, ethnographic approaches that document use of *Tephrosia* as a  
294 fish poison in Florida may not be able to uncover past use. However, the distribution of these  
295 taxa is highly suggestive of a pattern of human dispersal to anthropogenic sites such as shell  
296 middens and former mission sites. Furthermore, as a taxon closely associated with indigenous  
297 human use, they likely declined as disease, 19<sup>th</sup> century wars, and 20<sup>th</sup> century development  
298 altered the indigenous anthropogenic habitat where they thrived.

299 The distribution of *T. angustissima*, which is geographically restricted in the US and includes  
300 anthropogenic sites such as shell middens and former mission sites, is highly suggestive of  
301 movement by humans. However, written records of plant use by South Florida's early  
302 indigenous groups from colonial times are limited to a few accounts and only include a few  
303 species. Consequently, ethnographic approaches that document use of *Tephrosia*, whether as a  
304 fish poison or other use in Florida, were generally not available as a means of understanding the  
305 current distribution of this species. The approach taken in this study offers a novel means of

306 assessing the potential role of humans in the distribution of plant species where this information  
307 is lacking. Our work demonstrated that plants collected in Big Cypress were genetically distinct  
308 when compared to the rest of the populations. Otherwise, there was not clear genetic evidence to  
309 suggest human movement in the other taxa.

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317

### 318 **Author Contribution:**

319 The project was conceived by JS and EvW, with input from JP. Funding was obtained by EvW  
320 and JS. Collections were performed by JS, EvW, JP, and JL. Laboratory work was performed  
321 by NCG, and analyses were performed by PC, EO, and EvW. EvW pulled together the  
322 manuscript, with help from all authors who wrote subsections and revised multiple versions.

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### 324 **Data Availability Statement:**

325 All sequence data will be available on NCBI upon acceptance of the manuscript. Processed files  
326 are available on the open science foundation page for this project, <https://osf.io/q5wzu/>.

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### 328 **Conflict of Interest Statement:**

329 The authors declare no conflict of interest.

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### 332 **References:**

333

334 Allendorf, F.W. and G. Luikart. (2007). Conservation and the Genetics of Populations.  
335 Blackwell Press, New York, NY.

336

- 337 Austin, D.F. 2004. Florida Ethnobotany. CRC Press. Boca Raton. 909 pgs  
338
- 339 Baird, N. A., Etter, P. D., Atwood, T. S., Currey, M. C., Shiver, A. L., Lewis, Z. A., ... &  
340 Johnson, E. A. (2008). Rapid SNP discovery and genetic mapping using sequenced RAD  
341 markers. *PloS one*, 3(10), e3376.  
342
- 343 Beyra Matos A (1998) Las Leguminosas (Fabaceae) de Cuba, II. Tribus Crotalarieae,  
344 Aeschynoimeneae, Millettieae y Robinieae. Institute Botanica. Collectanea Botanica  
345 24(1998):149–332.  
346
- 347 Chevalier, A. 1937. Plantes ichtyotoxiques des genres *Tephrosia* et *Mundulea*; leur idspersion,  
348 leur culture, et leuers proprietes insecticides. *Revue de botanique appliquee et d'agriculture*  
349 *tropicale*. 17: 9-27.  
350
- 351 Davey, J. W., Hohenlohe, P. A., Etter, P. D., Boone, J. Q., Catchen, J. M., & Blaxter, M. L.  
352 (2011). Genome-wide genetic marker discovery and genotyping using next-generation  
353 sequencing. *Nature Reviews Genetics*, 12(7), 499-510.  
354
- 355 Do, C., Waples, R. S., Peel, D., Macbeth, G. M., Tillett, B. J., & Ovenden, J. R. (2014).  
356 NeEstimator v2: re-implementation of software for the estimation of contemporary effective  
357 population size (Ne) from genetic data. *Molecular Ecology Resources*, 14(1), 209-214.  
358
- 359 Ellstrand, N. C. and M. L. Roose. (1987). Patterns of genotypic diversity in clonal plant species.  
360 *American Journal of Botany* 74: 123-131.  
361
- 362 Evanno G., S. Regnau, and J. Goudet. (2005). Detecting the number of clusters of individuals  
363 using the software STRUCTURE: a simulation study. *Mol Ecol* 14: 2611–2620.  
364
- 365 Excoffier, L., Smouse, P.E., and Quattro, J.M. (1992). Analysis of molecular variance inferred  
366 from metric distances among DNA haplotypes: application to human mitochondrial DNA  
367 restriction data. *Genetics* 131: 479-491.

368

369 Excoffier L. (2001). Analysis of population subdivision, p. 271-307. In: D.J. Balding, M. Bishop,  
370 and C. Cannings, eds. *Handbook of statistical genetics*. Chichester (UK): John Wiley & Sons.

371

372 Fotinos TD, Maschinski J, and von Wettberg EJ. 2014. Saving the endangered Florida Key Tree  
373 cactus (*Pilosocereus robinii*) using new genetic tools. *Palmetto*, 31-12-15,

374

375 Gattepaille, L.M., Jakobsson, M. and Blum, M.G., 2013. Inferring population size changes with  
376 sequence and SNP data: lessons from human bottlenecks. *Heredity*, 110(5), pp.409-419.

377

378 Gann, G.D., K.A. Bradley, and S.W. Woodmansee. 2002. Rare Plants of South Florida: Their  
379 History, Conservation, and Restoration. The Institute for Regional Conservation, Miami, Florida.

380

381 Gravuer, K., von Wettberg, E., & Schmitt, J. (2005). Population differentiation and genetic  
382 variation inform translocation decisions for *Liatris scariosa* var. *novae-angliae*, a rare New  
383 England grassland perennial. *Biological Conservation*, 124(2), 155-167.

384

385 Hedrick, P.W. (2006). Genetic polymorphism in heterogenous environments: the age of  
386 genomics. *Annual review of ecology and systematics*. 37: 67-93.

387

388 Hohenlohe, P. A., Bassham, S., Etter, P. D., Stiffler, N., Johnson, E. A., & Cresko, W. A. (2010).  
389 Population genomics of parallel adaptation in threespine stickleback using sequenced RAD  
390 tags. *PLoS Genet*, 6(2), e1000862.

391

392 Hohenlohe, P. A., Day, M. D., Amish, S. J., Miller, M. R., Kamps-Hughes, N., Boyer, M. C., ...  
393 & Luikart, G. (2013). Genomic patterns of introgression in rainbow and westslope cutthroat trout  
394 illuminated by overlapping paired-end RAD sequencing. *Molecular Ecology*, 22(11), 3002-3013.

395

396 Howes, F.N., 1930. Fish-poison plants. *Bulletin of Miscellaneous Information (Royal Botanic  
397 Gardens, Kew)*, 1930(4), pp.129-153.

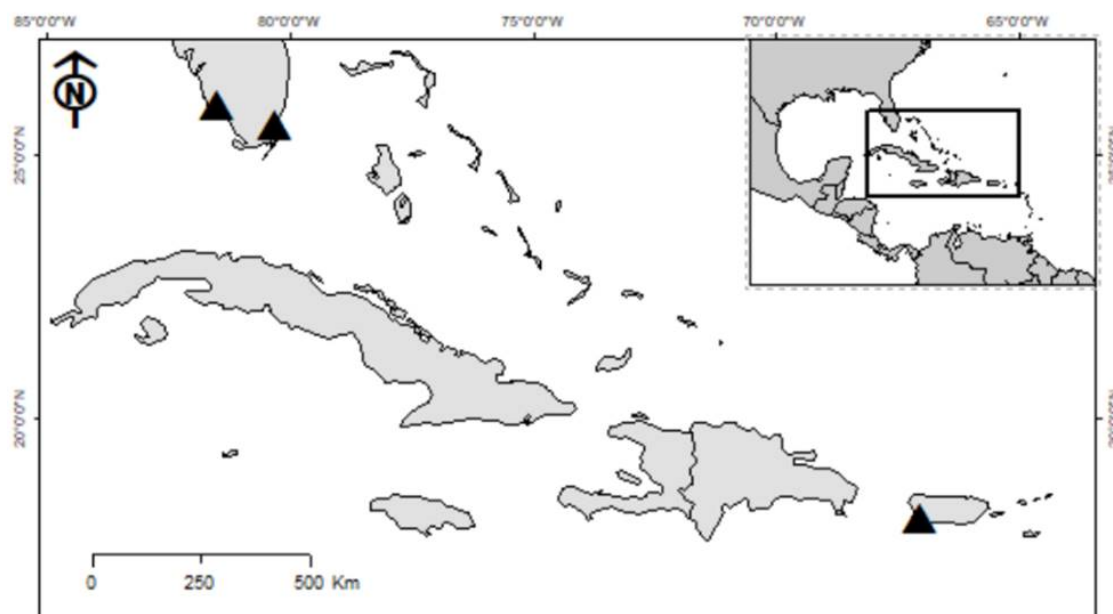
398

- 399 Isely, D. 1981. Leguminosae of the United States III. Subfamily Papilionoideae: Tribes,  
400 Sophoreae, Podalyriaceae, Loteae. Mem New York Bot. Gard. 25(3): 1–264.  
401
- 402 Isely, D. (1982). New combinations and one new variety among the genera *Indigofera*, *Robinia*,  
403 and *Tephrosia* (Leguminosae). *Brittonia*, 34(3), 339-341.  
404
- 405 McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytzky, A., ... &  
406 DePristo, M. A. (2010). The Genome Analysis Toolkit: a MapReduce framework for analyzing  
407 next-generation DNA sequencing data. *Genome research*, 20(9), 1297-1303.  
408
- 409 Peakall, R. O. D., & Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population  
410 genetic software for teaching and research. *Molecular ecology notes*, 6(1), 288-295.  
411
- 412 Peakall PE, Smouse R. (2012) GENALEX. Genetic analysis in Excel. Population genetic  
413 software for teaching and research—an update." *Bioinformatics* 28: 2537-2539.  
414
- 415 Peterson, B. K., Weber, J. N., Kay, E. H., Fisher, H. S., & Hoekstra, H. E. (2012). Double digest  
416 RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and non-  
417 model species. *PloS one*, 7(5), e37135.  
418
- 419 Possley, J., L. Cuni, E. Guinan, S. Wintergerst, N. Frade, B. Harding and D. Champney. 2022.  
420 Conservation of South Florida Endangered and Threatened Flora: 2021-2022 program at  
421 Fairchild Tropical Botanic Garden. Final report for Contract #027818, Florida Department of  
422 Agriculture and Consumer Services, Division of Plant Industry, Gainesville, FL.  
423
- 424 Pritchard J.K., M. Stephens, P. Donnelly. (2000). Inference of population structure using  
425 multilocus genotype data. *Genetics* 155: 945–959.  
426
- 427 Quigley, C., 1956. Aboriginal fish poisons and the diffusion problem. *American*  
428 *Anthropologist*, 58(3), pp.508-525.  
429

- 430 Ragsdale, A.P., Moreau, C. and Gravel, S., 2018. Genomic inference using diffusion models and  
431 the allele frequency spectrum. *Current Opinion in Genetics & Development*, 53, pp.140-147.  
432
- 433 Rowe, H. C., Renaut, S., & Guggisberg, A. (2011). RAD in the realm of next-generation  
434 sequencing technologies. *Molecular Ecology*, 20(17), 3499-3502.  
435
- 436 Sauer, CO. (1968). *Agricultural Origins and Dispersals - The Domestication of Animals and*  
437 *Foodstuffs*. 2<sup>nd</sup> Edition. MIT Press, Cambridge, MA USA.  
438
- 439
- 440 Shinnery, L. H. (1962). Key to southeastern glabrous-styled *Tephrosia* (Leguminosae). *SIDA*,  
441 *Contributions to Botany*, 1(1), 60-62.  
442
- 443
- 444 Wasserman A. 2009. *A People's History of Florida 1513-1876: How Africans, Seminoles,*  
445 *Women, and Lower Class Whites Shaped the Sunshine State*. 4<sup>th</sup> Edition. CreateSpace  
446 Independent Publishing Platform. 978-1442167094. 634 pps.  
447
- 448 Wendelberger, K. S. (2010). *Assessing microsite and regeneration niche preferences when*  
449 *introducing endangered species*. THE UNIVERSITY OF NORTH CAROLINA AT CHAPEL  
450 HILL.  
451
- 452 Wendelberger, K.S. and Maschinski, J., 2016. Assessing microsite and regeneration niche  
453 preferences through experimental reintroduction of the rare plant *Tephrosia angustissima* var.  
454 *corallicola*. *Plant ecology*, 217(2), pp.155-167.  
455



456 Figure 1. Map of the Caribbean showing sampling locations in Florida and Puerto Rico.



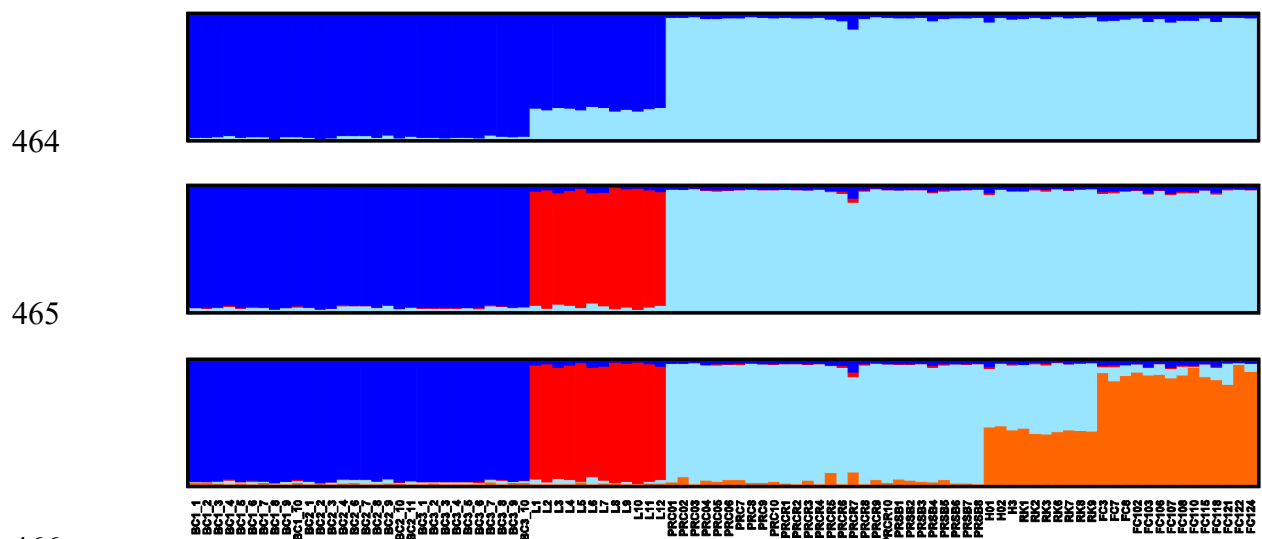
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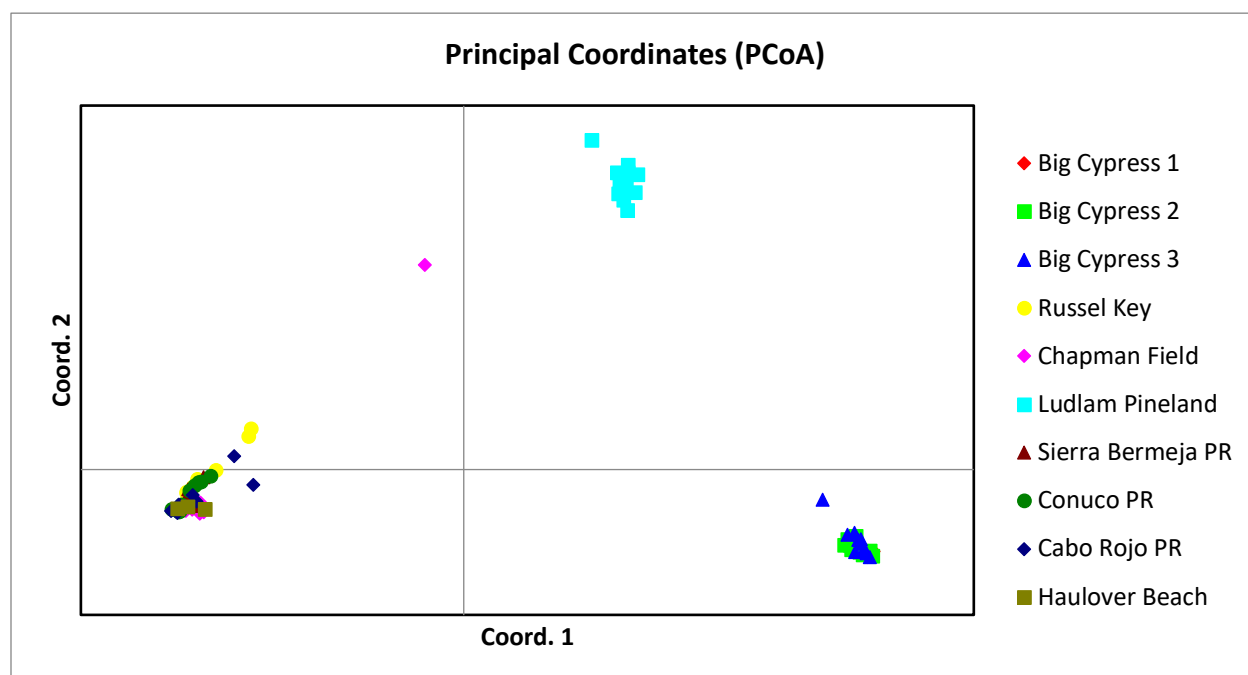
461 Figure 2. Results of a STRUCTURE analysis on 6272 SNPs, with population numbers (K) of K  
462 =2, K=3 and K=4 shown. The Evanno method determined K=3 to best describe the data. K=2  
463 and K=4 are shown to contextualize our interpretation of assignment of samples of populations.



471 =

472 Figure 3. Principal coordinate analysis. Populations 1-3 are from Big Cypress (lower right  
473 corner). Population 4 is Russell Key, Population 5 is Chapman Field. Population 6 is Ludlam  
474 Pineland. Population 7-9 are from Puerto Rico. PCoA coordinate axes one and two represent  
475 42.6 and 19.3 percent of diversity, respectively.

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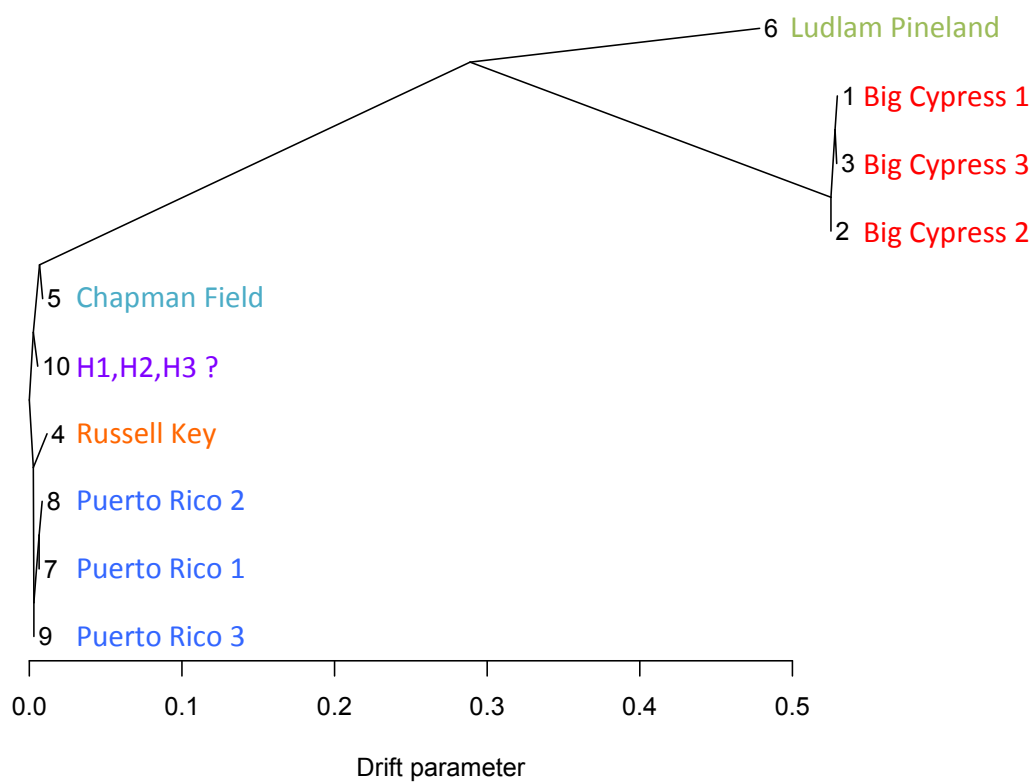
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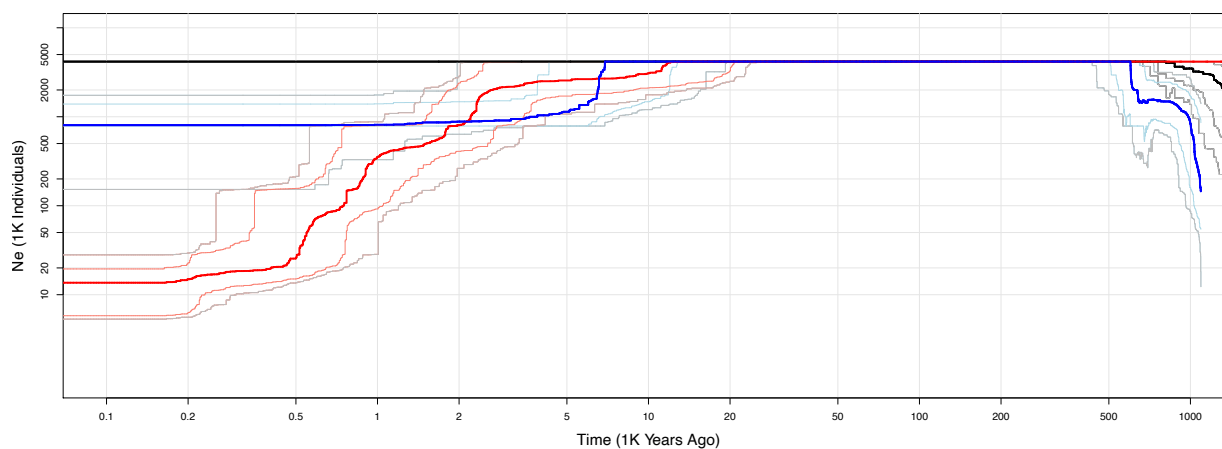
481 Figure 4. Treemix analysis showing a lack of introgression among the three South Florida  
482 *Tephrosia* taxa.  
483

Stratification level: Population



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487 Figure 5. Historical effective population size estimates based on the site frequency spectrum.  
488 Blue color indicates the Big Cypress population (T1), Red the *T. florida* population from Ludlum  
489 (T3), and black the Russel Key/Chapman Field, Puerto Rican group ((T2). Solid lines represent  
490 the estimated population size, and dotted lines the 75 and 95% confidence intervals around the  
491 estimate.  
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