

1 **Parallel trophic diversifications in polyploid cyprinid fish from East**
2 **Africa: from preadaptive polymorphism to trophic specialization**

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Abstract

28 Trophic diversification is one of the main mechanisms driving the adaptive radiation. The
29 polyploid lineage of the cyprinid genus *Labeobarbus* represent an excellent model for
30 studying the trophically-based adaptive radiation in either lacustrine or riverine
31 environments. Recently discovered four diversifications in rivers of the Ethiopian Highlands
32 (East Africa) demonstrate independently evolved repeated mouth polymorphisms each
33 represented by four core mouth phenotypes: (i) generalized, (ii) thick-lipped, (iii) scraping,
34 and iv) large-mouthed. Mouth phenotypes in some radiations can be further divided to
35 subtypes representing from four to eight sympatric ecomorphs. Using the stable isotope and
36 gut content analyses we tested hypothesis on trophic resource partitioning within each
37 radiation, revealed disparity in degree of diversification between radiations and tried to
38 reconstruct the process of trophic diversification. Three of four radiations demonstrated
39 partitioning of trophic resources within five trophic niches: i) detritophagy, ii)
40 macrophytophagy, iii) invertivorous benthophagy, iv) periphyton feeding, and v) piscivory.
41 The studied riverine radiations were likely at the different stages of the diversification. One
42 radiation having a similar set of mouth phenotypes was not trophically divergent displaying a
43 remarkable decouple of form and function. A unique case of ecologically non-functional
44 mouth polymorphism at an incipient stage of trophic diversification supports a concept of the
45 plasticity-first evolution. This phenomenon stems from the pre-existing genomic templates of
46 mouth polymorphism ancestrally inherited upon the allopolyploid origin of the *Labeobarbus*
47 lineage. The predetermined and preadaptive mouth polymorphism can be considered a key
48 innovation of the *Labeobarbus* that promoted to resource-based diversification via adaptive
49 radiation.

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51 Running Title: Preadaptive polymorphism and trophic specialization

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53 **Key words:** adaptive radiation, polymorphism, trophic resource partitioning, ecological
54 speciation, vertebrates

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56 **1. Introduction**

57 Trophic divergence is one of the main drivers of ecological speciation and adaptive radiation
58 (Schluter, 2000; Seehausen & Wagner, 2014; Martin & Richards, 2019). The fishes being the
59 most diverse group among vertebrates display numerous examples of trophic diversification
60 during adaptive radiation - not only among textbook examples from cichlids, whitefish,
61 Arctic charrs (Seehausen & Wagner, 2014) - but also in other taxa. Despite tremendous
62 amount of studies focused on the mechanisms underpinning the rapid diversification during
63 adaptive radiation (and achieved progress – e.g., Brawand *et al.*, 2014; McGee *et al.*, 2020;
64 Ronco *et al.*, 2021), the phenomenon is still poorly understood (Martin & Richards, 2019;
65 Gillespie *et al.*, 2020). One of the greatest findings of the last decade – a contribution of the
66 heterogenous genome (usually evolved via past hybridization) into accelerated rates of
67 ecological diversification and speciation (Nosil *et al.*, 2009; Genner & Turner, 2012; Meier *et*
68 *al.*, 2017; Irisarri *et al.*, 2018; Marques *et al.*, 2019; Svardal *et al.*, 2020). One of the
69 mechanisms to increase genetic heterogeneity is a polyploidization. After a round of fish-
70 specific genome duplication (Meyer & Van de Peer, 2005) occurred ca. 350 mya in the ray-
71 finned fish (Actinopterygii) lineage, the numerous further polyploidization events happened
72 within and resulted in >1000 species mostly within Cypriniformes, Salmoniformes and
73 Acipenseriformes (Yang *et al.*, 2022). Contrary to plants, the animal polyploidization is

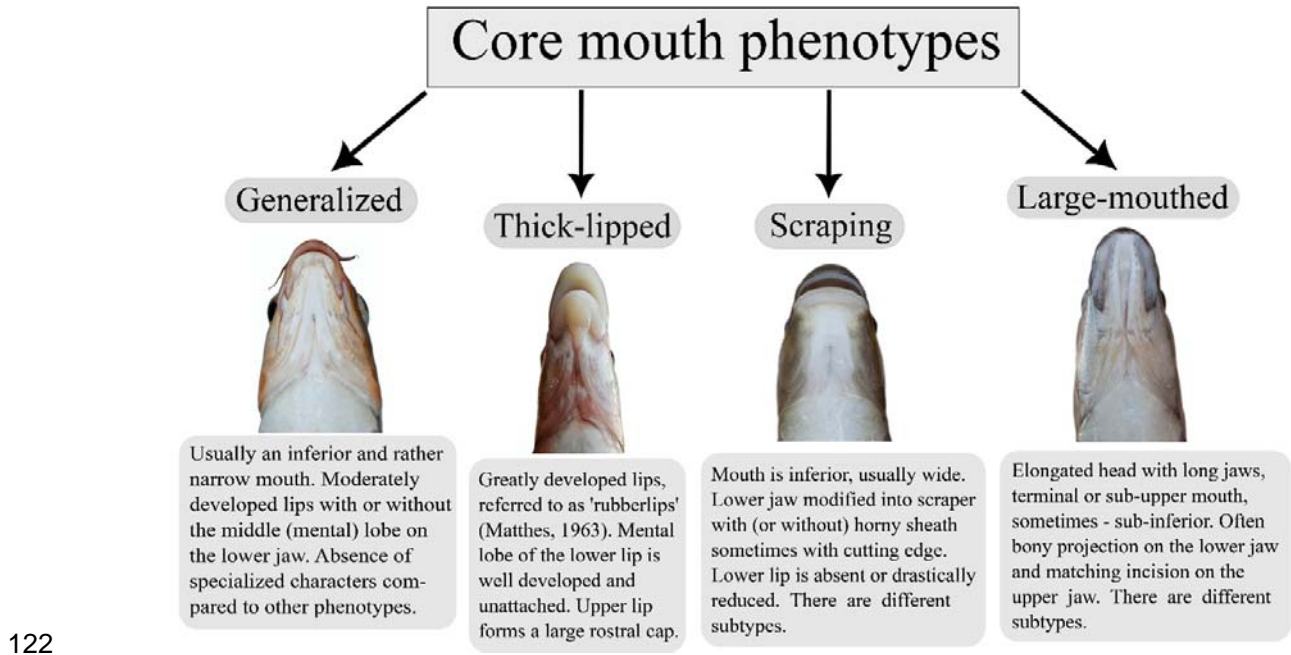
74 realized predominantly *via* allopolyploidization (Gregory & Mable, 2005) that means the
75 most of fish polyploid lineages contains the different subgenomes. Therefore, the polyploid
76 lineages look as perfect candidates to test a hypothesis on role of complex genome in further
77 diversification during adaptive radiation. Herein a polyploidy apart prospective gains also has
78 many restrictions on fish biology that can dismiss the expected benefits (Gregory & Mable,
79 2005; Van de Peer *et al.*, 2017). Nevertheless, the most researchers agreed that polyploid taxa
80 are characterized by increased ecological and phenotypic plasticity that (i) facilitate to
81 overcome the stressful and harsh environment and (ii) can promote to further diversification
82 and speciation that is within ‘flexible stem’ hypothesis (West-Eberhard, 2003; Wund *et al.*,
83 2008; Li & Guo, 2020; Van de Peer *et al.*, 2021). Some polyploid lineages, for instance –
84 Salmonidae and Coregonidae – represent nice examples of phenotypic and ecological
85 plasticity along with possibility to evolve small-scale adaptive radiation especially within
86 genera *Salvelinus*, *Salmo*, and *Coregonus* (Reshetnikov, 1980; Bernatchez *et al.*, 1999;
87 Alekseyev *et al.*, 2002; Markevich *et al.*, 2018; Segherloo *et al.*, 2021; Levin *et al.*, 2022).

88 The cyprinid fishes (fam. Cyprinidae sensu Tan & Armbruster, 2018) being one of the
89 most diverse families of teleostean fishes (> 1780 species - Fricke *et al.*, 2023) are the
90 champions in the number of the polyploid species (ca. 600 species – Yang *et al.*, 2022). Some
91 of cyprinid polyploid lineages display bright examples of adaptive radiation based on trophic
92 polymorphism (Nagelkerke *et al.*, 1994; Roberts & Khaironizam, 2008; Levin *et al.*, 2019;
93 2021; Qiao *et al.*, 2020; Komarova *et al.*, 2021). Among these, the most distinguished one in
94 terms of the morpho-ecological diversification is the cyprinid genus *Labeobarbus*
95 representing species-rich African hexaploid lineage ($2n=150$ – Oellerman & Skelton, 1990;
96 Golubtsov & Krysanov, 1993) that comprises >130 species (Vreven *et al.*, 2016; Fricke *et al.*,
97 2023) and displays numerous trophically-based adaptive radiations (Nagelkerke *et al.*, 1994;
98 2015; Mina *et al.*, 1996; Sibbing & Nagelkerke, 2000; Dimmick *et al.*, 2001; Golubtsov,

99 2010; Shkil *et al.*, 2015; Levin *et al.*, 2020). The *Labeobarbus* is widespread in Africa and
100 presents in each of the ten African ichthyofaunal provinces (Vreven *et al.*, 2016). Based on
101 the recent data on phenotypic, ecological, and genetic data, the taxonomic diversity of the
102 *Labeobarbus* might be greatly underestimated (Levin *et al.*, 2019, 2020; Decru *et al.*, 2022).

103 The Ethiopian Highlands is remarkably distinguished as a hotspot for multiple adaptive
104 radiations of the *Labeobarbus* discovered during last 30 years (see Golubtsov *et al.*, 2021).
105 The most famous example of rapid adaptive radiation among non-cichlid fishes belongs to
106 the lacustrine radiation of the *Labeobarbus* in Lake Tana, Ethiopia, where up to 15
107 species/ecomorphs were described that partitioned trophic resources (Nagelkerke *et al.*, 1994;
108 Mina *et al.*, 1996; Zworykin *et al.*, 2006; Shkil *et al.*, 2015). Apart from the lacustrine
109 radiation strongly predominating among fishes (e.g. Seehausen & Wagner, 2014), there are
110 striking examples of the riverine adaptive radiations in non-cyprinid (Turner *et al.*, 1985;
111 Piálek *et al.*, 2012; Esin *et al.*, 2021; Říčan *et al.*, 2021; Burrell *et al.*, 2022) as well as
112 cyprinid fishes - *Labeobarbus*, in particular (Levin *et al.*, 2019; 2020).

113 The most conspicuous phenotypic difference between sympatric ecomorphs of the
114 *Labeobarbus* refers to mouth type variation. The core of mouth polymorphism is composed
115 of four main phenotypes (Figure 1) that were recorded throughout Africa (Worthington,
116 1929; 1933; Matthes, 1963; Banister, 1973; Skelton *et al.*, 1991; Nagelkerke *et al.*, 1994): i)
117 generalized (typical for barbs), ii) thick-lipped ('rubberlip'), iii) scraping ('chiselmouth'), and
118 iv) large-mouthed. Mouth phenotypes are subjected to further sub-diversification and can
119 include subtypes or variants that are recruited for occupying the new ecological niches (Levin
120 *et al.*, 2021b). Moreover, there are intermediate phenotypes possibly of hybrid origin
121 (Banister, 1972; Vreven *et al.*, 2019) that does not exclude their further specialization.



122

123 Figure 1. Pictorial scheme of core mouth phenotypes of the *Labeobarbus*. Photograph
124 examples referred to the *Labeobarbus* from the rivers of the Ethiopian Highlands.

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126 This outstanding mouth polymorphism of the *Labeobarbus* likely stems from ancestral
127 inherited genetic templates since both parental lineages involved in the allopolyploidization
128 event (Yang *et al.*, 2015; 2022) belong to extant lineages and represent different mouth types.
129 The contemporary representatives of the maternal lineage are characterized by generalized
130 and thick-lipped mouth polymorphism (tetraploid *Torinae*, $2n = 100$ - Roberts &
131 Khaironizam, 2008; Walton *et al.*, 2017; Coad, 2021) while paternal diploid lineage
132 (*Cyprinion*, $2n = 50$, Ünlü *et al.*, 1997; Esmaeili & Piravar, 2006) is represented by scraping
133 mouth phenotype (Coad, 2021). Hence, mouth polymorphism of the *Labeobarbus* is
134 ancestrally heritable, likely preadaptive and might re-evolve.

135 The parallel diversifications of the *Labeobarbus* in the riverine environment of the
136 Ethiopian Highlands were recently discovered within the four geographically isolated
137 drainages - i) the Didessa River in Blue Nile basin; ii) the Sore River in White Nile basin; iii)
138 the Gojeb River in Omo-Turkana basin; and iv) the Genale River in Juba-Wabe-Shebelle
139 basin, Indian Ocean catchment (Mina *et al.*, 1998; Golubtsov, 2010; Levin *et al.*, 2019; 2020;
140 Golubtsov *et al.*, 2021). Each diversification includes from four to eight sympatric ecomorphs
141 divergent in mouth phenotype (Table 1) that presumably involves trophic adaptations. The
142 genetic studies showed that all four riverine radiations of *Labeobarbus* were independently
143 evolved from different ancestors although they are very closely-related based on mtDNA data
144 that suggests recent and rapid diversification (Levin *et al.*, 2013; 2019; 2020). Moreover,
145 existing data suggests these radiations might be at various stages of the diversification (Levin
146 *et al.*, 2020; 2023 - in press). Of special interest is that these radiations could repeatedly
147 emerge in the riverine environment that has been considered to be inappropriate for adaptive
148 radiation due to heterogeneous and unpredictable conditions for a long time (reviewed in
149 Burress *et al.*, 2023).

150 A few studies on trophic ecology showed that sympatric riverine ecomorphs of the
151 *Labeobarbus* bearing different mouth phenotypes have partitioned trophic resources - in
152 particular, in the Genale River (Levin *et al.*, 2019). Other riverine radiations of the
153 *Labeobarbus* were not studied comprehensively from the point of trophic divergence. The
154 recent but independent origin of similar diversifications in mouth phenotypes raises several
155 questions. Do all diversifications partition trophic resources according to homologous mouth
156 phenotypes? What is trophic specialization (if present) of various sympatric ecomorphs?
157 Whether riverine diversifications comprising different numbers of ecomorphs / mouth
158 (sub)phenotypes are at various stages of the process? If so, could patterns of the trophic
159 diversification be sorted from more primitive to advanced aiming reconstruction of the

160 trophic radiation? We try to answer these questions in a study of trophic diversification of
 161 sympatric ecomorphs in each of four *Labeobarbus* riverine radiations using the stable isotope
 162 composition and gut content.

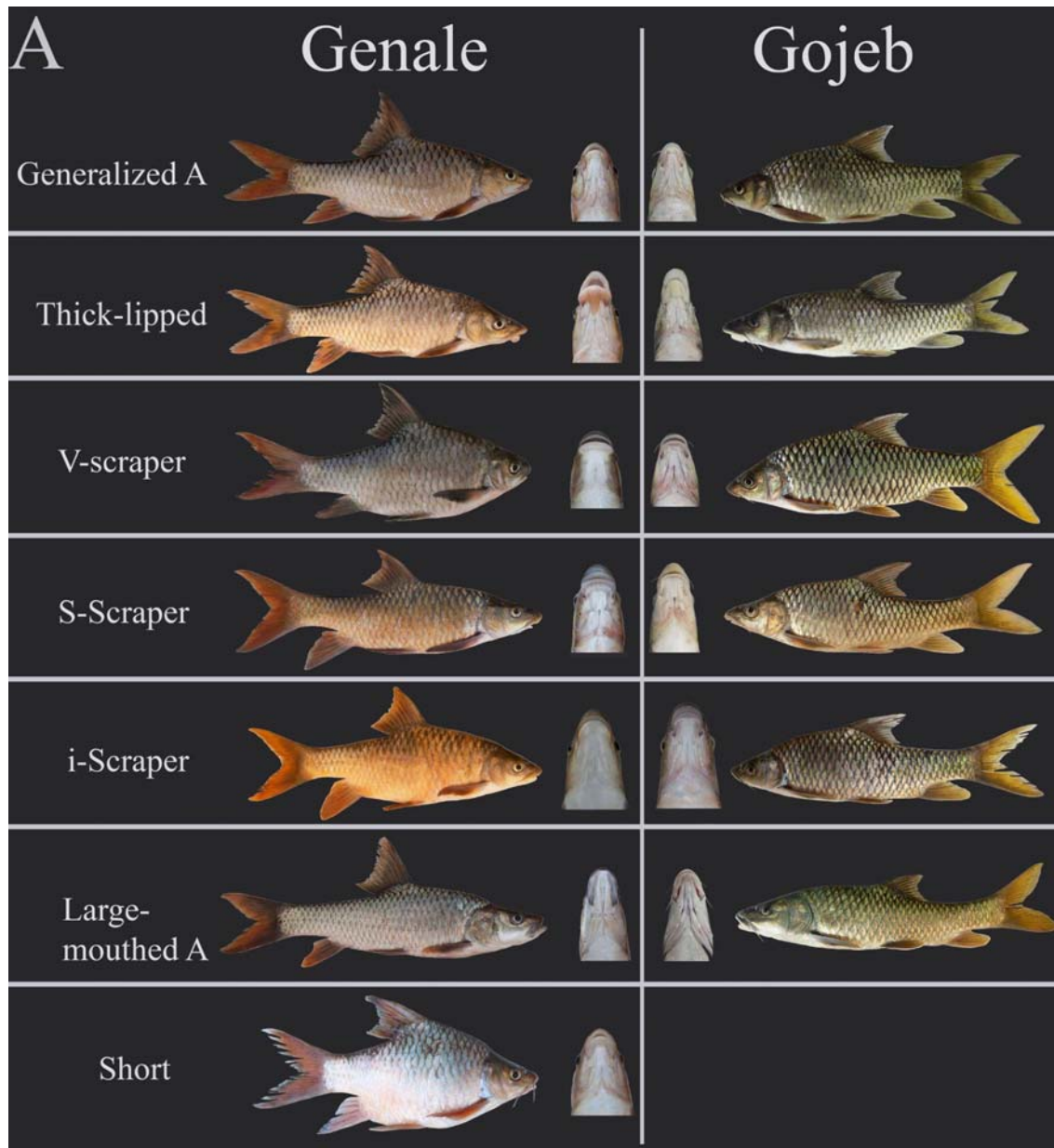
163 Table 1. The composition of riverine sympatric ecomorphs of *Labeobarbus* spp. in
 164 isolated riverine basins in the Ethiopian Highlands (summarized from Golubtsov, 2010;
 165 Levin *et al.*, 2019; 2020; 2021; Mina *et al.*, 1998 and our unpublished data). The photographs
 166 of the fish ecomorphs are given in Figure 2.

Species, river and basin	Ecomorphs	Source
<i>Labeobarbus gananensis</i> complex, Genale River, Wabe-Shebelle basin, Indian Ocean catchment.	1. Generalized A (common) 2. Thick-lipped 3. V-scraeper ¹ (<i>L. jubae</i>) 4. S-scraeper ² 5. i-Scraeper ³ 6. Short ⁴ 7. Large-mouthed A ⁵	Golubtsov, 1993; Mina <i>et al.</i> , 1998; Dimmick <i>et al.</i> , 2001; Levin <i>et al.</i> , 2019; Golubtsov <i>et al.</i> , 2021
<i>Labeobarbus intermedius</i> complex, <i>L. zaphiri</i> , <i>L. beso</i> . Didessa River, Nile basin, Atlantic Ocean catchment.	1. Generalized A 2. Generalized B ⁶ 3. Thick-lipped 4. Semi-thicklipped ⁷ 5. V-scraeper (<i>L. beso</i>) 6. Large-mouthed A1 (<i>L.</i>	Golubtsov, 2010; Mina <i>et al.</i> , 1998; Levin <i>et al.</i> , 2020; 2021; original observations

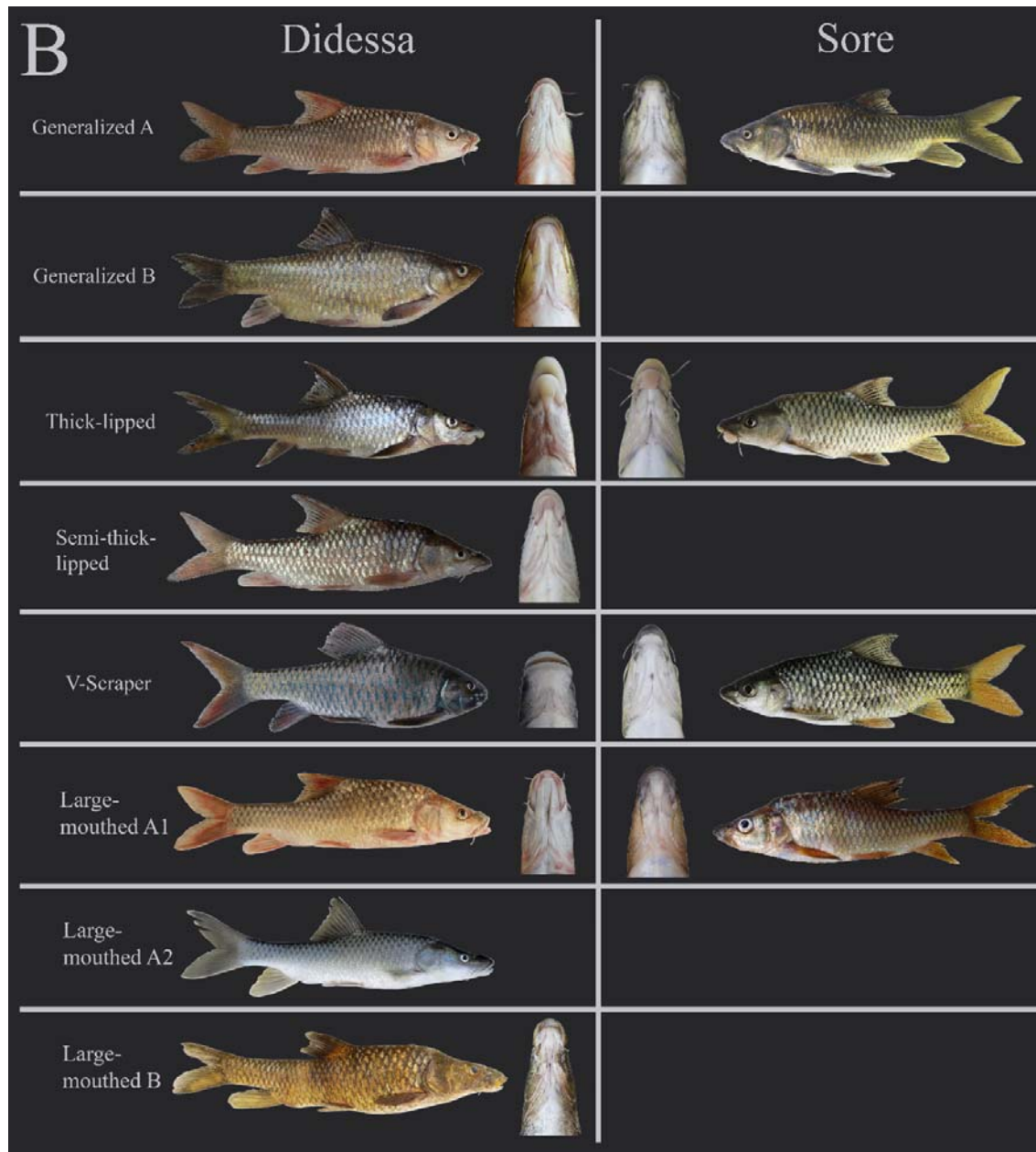
	<i>zaphiri</i>	
	7. Large-mouthed	A2
	<i>(Labeobarbus sp.1)</i> ⁸	
	8. Large-mouthed	B ⁹
	<i>(Labeobarbus sp.2)</i>	

<i>Labeobarbus</i>	1. Generalized A	Golubtsov, 2010; Levin <i>et al.</i> ,
<i>intermedius</i> complex.	2. Thick-lipped	2020; 2021; original
Gojeb River, Omo-	3. V-scraper	observations
Turkana basin, inland	4. S-scraper	
drainage.	5. i-Scraper	
	6. Large-mouthed A	

<i>Labeobarbus</i>	- Generalized A	Golubtsov 2010; Levin <i>et al.</i> ,
<i>intermedius</i> complex.	- Thick-lipped	2020; original observations
Sore River, White Nile	- V-scraper	
basin, Atlantic Ocean	- Large-mouthed ⁹	
catchment.		



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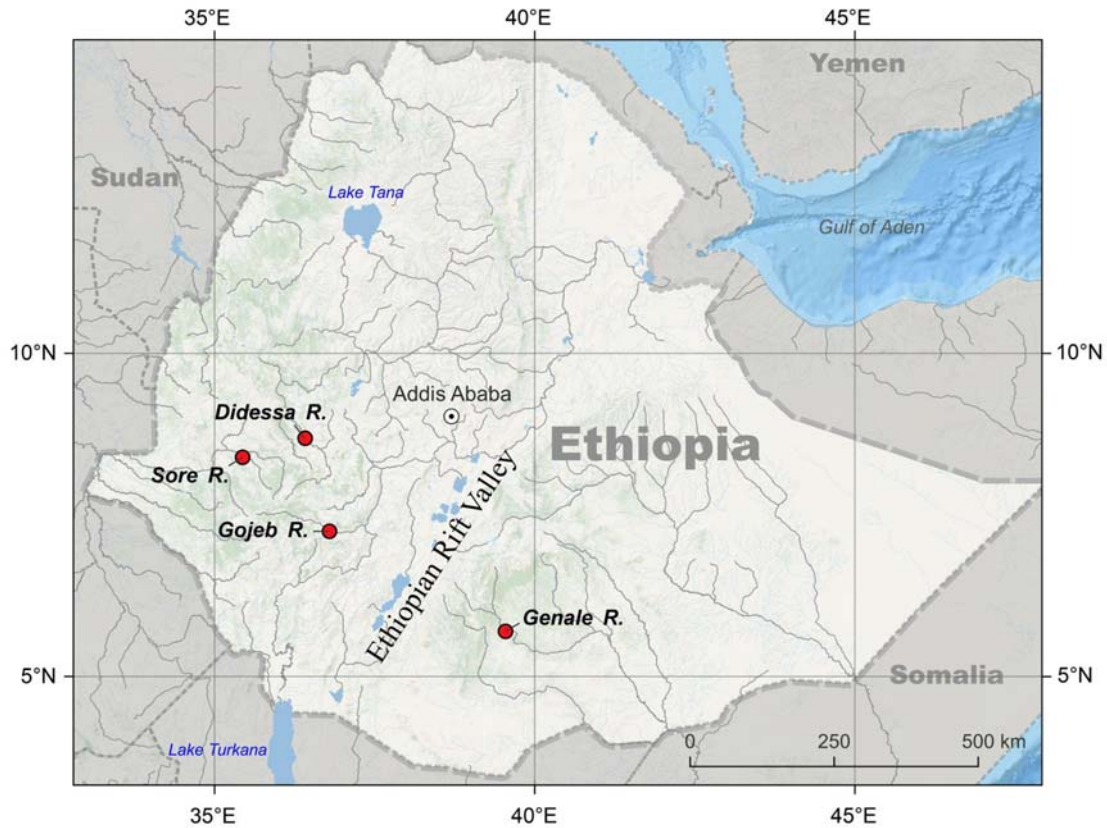
170 Figure 2. Appearance and mouth phenotypes of the ecomorphs of the *Labeobarbus* in
171 different rivers of the Ethiopian Highlands: (A) the Genale and Gojeb Rivers; (B) the Didessa
172 and Sore Rivers. An exceptionally rare representative of the Large-mouthed A2 ecomorph
173 from the Didessa River is presented only from the lateral side.

174

175 **2. Material and Methods**

176 *Ethical approval.* All applicable international, national, and institutional guidelines for the
177 care and use of animals were strictly followed. All animal sample collection protocols
178 complied with the current laws of Russian Federation and Federal Democratic Republic
179 of Ethiopia.

180 *Study sites and material studied.* Samples were collected under the framework of the Joint
181 Ethiopian-Russian Biological Expedition (JERBE) in four rivers draining the Ethiopian
182 Highlands and belonging to four major river basins (Fig. 2): (1) the Juba-Wabe-Shebelle
183 basin in the Indian Ocean catchment – the Genale R. – 5.7025° N 39.5446° E; (2) the Blue
184 Nile basin – the Didessa R., a tributary of the Blue Nile – 8.6921° N 36.4144° E; (3) the
185 Omo-Turkana enclosed basin – the Gojeb R., a tributary of the Omo R. – 7.2539° N 36.7943°
186 E; (4) the White Nile basin – the Sore R. ~35 km downstream City Metu – 8.3987° N,
187 35.4378° E. Fish were caught by gill and cast nets in February–March 2011 (Didessa),
188 March–April 2009, 2019 (Genale), February 2011 (Gojeb), April 2014 (Sore). Fish were
189 killed with an overdose of MS-222 anaesthetic and then photographed using a Canon EOS
190 50D camera (Canon Inc. Tokyo, Japan).



192 Figure 3. Map of sampling sites of the four riverine radiations of the Ethiopian *Labeobarbus*.

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194 Standard length (SL, mm) and gut length (GL, mm) were measured with a ruler. Fish were
195 preserved first in 10% formalin and then transferred to 70 % ethanol. Gut length was studied
196 in 461 individuals. All specimens were deposited at A.N. Severtsov Institute of Ecology and
197 Evolution, the Russian Academy of Sciences, Moscow under provisional labels of JERBE.
198 The detailed information on material studied (gut length, stable isotope composition and gut
199 content) is given in Supporting Table S2.

200 *Stable isotopes composition.* For stable isotope (SI) analyses, white muscle tissue from the
201 dorsal side of the body under the dorsal fin was sampled from freshly collected specimens.
202 White muscle samples were dried at 60 °C. The samples were weighed using a Mettler

203 Toledo MX5 microbalance (Mettler Toledo, Columbus, OH, United States) with 2 µg
204 accuracy, and wrapped in tin capsules. The weight of the fish tissue samples varied from 250
205 to 500 µg. SI analysis was conducted at the Joint Usage Center of the A.N. Severtsov Institute
206 of Ecology and Evolution RAS, Moscow. Briefly, a Thermo Delta V Plus continuous-flow
207 IRMS was coupled to an elemental analyzer (Flash 1112) equipped with a Thermo No-Blank
208 device. The isotopic composition of N and C was expressed in the δ notation relative to the
209 international standards (atmospheric nitrogen and VPDB, respectively): $\delta X (\text{‰}) =$
210 $[(R_{\text{sample}}/R_{\text{standard}}) - 1] \times 1000$, where R is the molar ratio of the heavier and lighter
211 isotopes. The samples were analyzed with a reference gas calibrated against the International
212 Atomic Energy Agency (IAEA) reference materials USGS 40 and USGS 41 (glutamic acid).
213 The measurement accuracy was $\pm 0.2 \text{ ‰}$. Along with the isotopic analysis, the nitrogen and
214 carbon content (as %) and C/N ratios were determined. In total, 400 white-muscle samples
215 were analyzed.

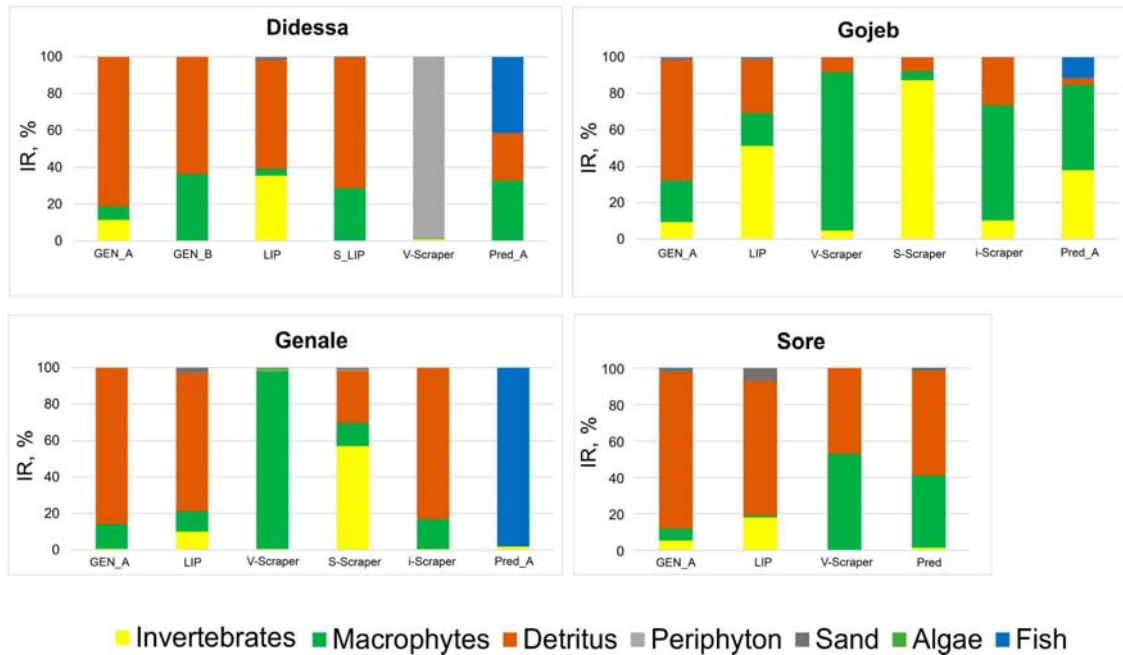
216 *Diet analysis.* Following the best practice for estimation of trophic niches the in tropic rivers
217 (e.g., Davis *et al.*, 2012) we studied gut content in addition to SI composition. Gut content
218 was extracted from preserved specimens, dried on filter paper and weighed using a Pioneer
219 PX84/E balance with 0.0001 g accuracy. The diet particles were identified using Olympus
220 CX41 microscope (100–1000× magnification) and Motic DMW-143-N2GG
221 stereomicroscope (100–400× magnification). The diet components were grouped into: (i)
222 detritus, (ii) invertebrates, (iii) macrophytes, (iv) periphyton, (v) filamentous algae, (vi) fish
223 (body remnants and scales), and (vii) mineral ground. The group ‘Invertebrates’ included
224 mainly the larvae and imago of amphibiotic insects and their fragments as well as imago of
225 aerial insects (Coleoptera, Hymenoptera); rarely Cladocera. The group ‘Macrophytes’
226 included any fragments of helophytic and semi-aquatic plants - such as leaves, stems or
227 seeds. A composite measure of diet, an index of relative importance (IR) [Natarajan &

228 Jhingran, 1961; Popova & Reshetnikov, 2011], was used to assess the contribution of
229 different components to the diet. The IR index was calculated as follows: $IR = (F_i \times$
230 $P_i) / (\sum (F_i \times P_i)) \times 100\%$, where F_i = the frequency of occurrence of each food group, and P_i
231 = its part by weight; the value of i itself changes from 1 to n (n = the part of food organisms
232 in the food bolus). In total, 195 food boluses were analyzed for gut content (Supplementary
233 Table S2).

234 *Statistical analyses.* Several R packages and functions were used for the statistical analyses
235 and plot construction. Basal descriptive statistics was obtained using the *summarytools*
236 library [Comtois, 2022]. Central tendencies in the text are presented as means and 1SD. The
237 Kruskal-Wallis test was applied for comparisons of the values of gut length GL and SI
238 composition using the function *kruskalTest* in each radiation (in library *PMCMRplus*) with
239 subsequent pairwise comparisons using post-hoc Dunn's all-pairs test with Bonferoni
240 adjustment - function *kwAllPairsDunnTest* [R Core Team, 2023]. The violin boxplots were
241 obtained using the *ggplot2* library [Wickham, 2022]. The package SIBER v.2.1.6 [Jackson *et*
242 *al.*, 2011] was used to assess the differences in the isotopic trophic niche features. The total
243 convex hull areas (TA), core trophic niche breadths, and sample size-corrected standard
244 ellipse area (SEAc) were estimated. The trophic overlap for 95% TA was estimated using
245 nicheROVER [Lysy *et al.*, 2021], a method that is insensitive to the sample size and
246 incorporates statistical uncertainty using Bayesian approach [Swanson *et al.*, 2015].

247 **3. Results**

248 *3.1. Diet.* Food spectra of *Labeobarbus* spp. were rather diverse. Feeding of sympatric
249 ecomorphs in all four rivers was divergent but at various degrees as estimated by index of
250 relative importance (Figure 4). Detailed description of the diet for each ecomorph is given in
251 Supplementary material S3.



252

■ Invertebrates ■ Macrophytes ■ Detritus ■ Periphyton ■ Sand ■ Algae ■ Fish

253

Figure 4. Food spectra (IR: the index of relative importance) of the sympatric ecomorphs of

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the *Labeobarbus* spp. from the Didessa, Gojeb, Genale, and Sore rivers. Abbreviations of the

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ecomorphs: GEN_A - Generalized A; GEN_B - Generalized B; LIP - Thick-lipped; S_LIP -

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Semi-thicklipped; Pred_A - Large-mouthed A; Pred_B - Large-mouthed B.

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258

Based on the food spectra, up to five trophic specializations can be detected in the

259

Didessa River: i) detritivore (Generalized A), ii) detritivore-macrophytophage (Generalized B

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and Semi-thicklipped), iii) detritivore-invertivore (Lipped), iv) periphyton feeder (V-scraper),

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and v) piscivore-omnivore (Large-mouthed A and B). Notably, detritus seems to be a core

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food for many ecomorphs except for the V-Scraper and Large-mouthed. A sub-specialization

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within the piscivory strategy is possible taking into account the drastic difference in mouth

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structure (Figure 2B) and diet between the Large-mouthed ecomorphs. Seemingly, up to five

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feeding strategies can be distinguished in the Gojeb River: i) detritivore (Generalized A), ii)

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invertivore-detritivore (Lipped); iii) macrophytophagous (V-Scraper and i-Scraper), iv)

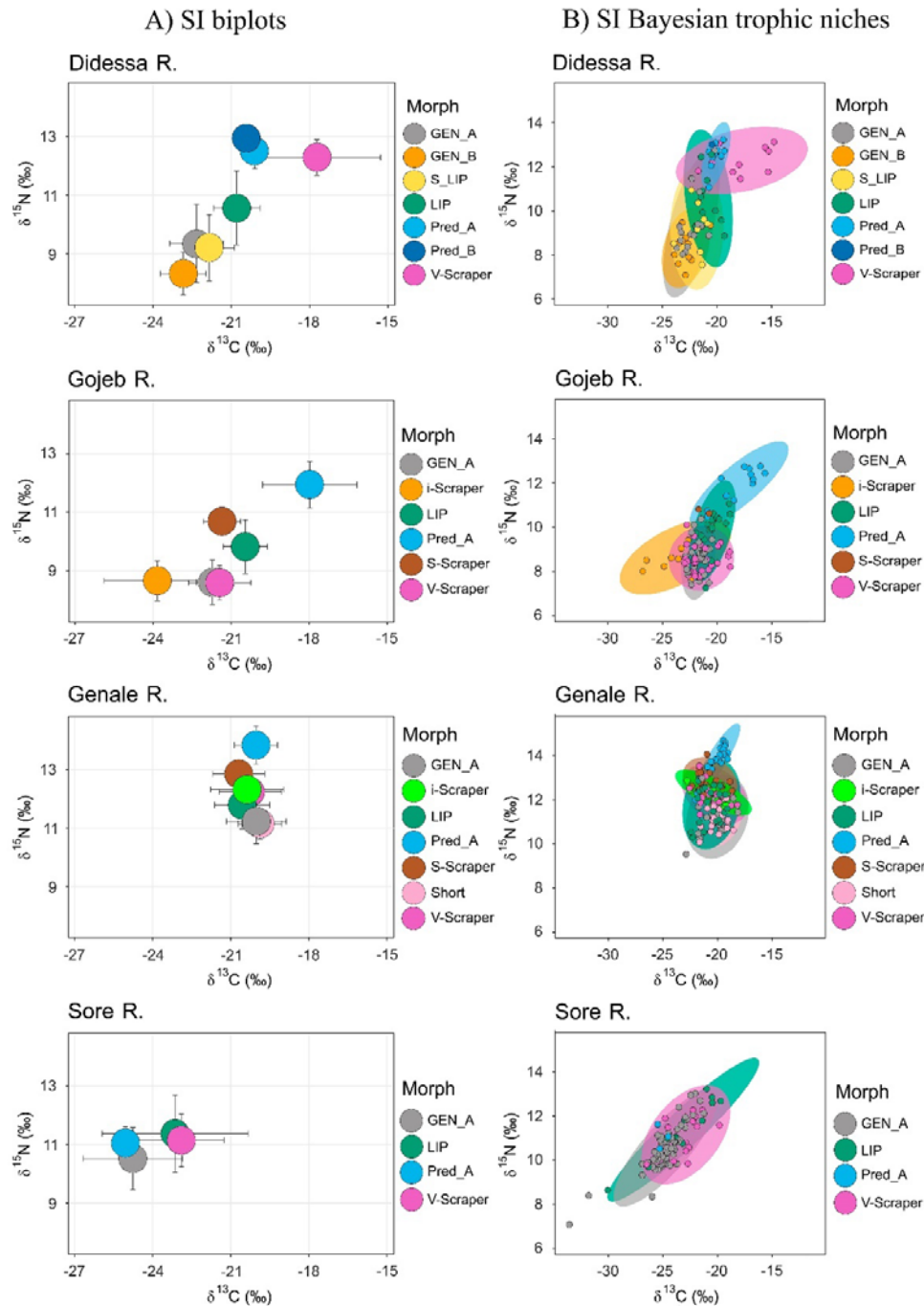
267 invertivore (S-scraper), and v) omnivore-piscivore (Large-mouthed A). Compared to the
268 Didessa, a detritus was a core food only for the Generalized A ecomorph. Large-mouthed
269 ecomorph is not predominantly piscivorous but rather omnivorous with a small portion of
270 fish food. In the Genale River, up to four feeding strategies can be detected – i) detritivore
271 (Generalized A, Lipped, and i-Scraper), ii) macrophytophagous (V-Scraper), iii) invertivore
272 (S-scraper), and iv) piscivore (Large-mouthed A). Remarkably, a detritus was a core food for
273 three ecomorphs in the Gojeb River, while the Large-mouthed ecomorph is a strongly
274 piscivorous specialist. In contrast, little diversification in food spectra was detected in the
275 Sore River. Briefly, two feeding strategies can be recognized: i) detritivore (Generalized A,
276 Lipped, and i-Scraper), and ii) macrophytophagous-detritivore (V-Scraper and Large-
277 mouthed). Remarkably detritus was a core food for three of four ecomorphs although a
278 notable portion (almost 20 %) of benthic invertebrates was detected in the Lipped ecomorph.
279 Large-mouthed ecomorph is not a piscivorous specialist in the Sore River.

280 *3.2. Stable Isotope Composition.* Basic statistics for both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values and data on
281 the total area (TA), standard ellipse area (SEA), and corrected standard ellipse area (SEAc)
282 are given in Supplementary Materials S4. Detailed results are presented in subsequent
283 sections.

284 *3.2.1. Didessa Radiation.* Among seven ecomorphs, several were significantly divergent in SI
285 values (six pairwise comparisons in both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ - Table S4). The largest values of
286 $\delta^{15}\text{N}$ were detected in two Large-mouthed (piscivory) ecomorphs (12.5 ± 0.6 and 12.9%) as
287 well as in highly specialized V-scraper *L. beso* ($12.3\pm 0.6\%$). The V-scraper was also most
288 enriched in ^{13}C ($\delta^{13}\text{C} -17.7\pm 2.4\%$) (Figure 5). The lowest values of both $\delta^{15}\text{N}$ ($8.33\pm 0.7\%$)
289 and $\delta^{13}\text{C}$ ($-22.8\pm 0.9\%$) were in the Generalized B ecomorph. Other ecomorphs, Generalized
290 A, Thick-lipped, and Semi-thicklipped, had intermediate $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values, although

291 Thick-lipped ecomorph had somewhat higher $\delta^{15}\text{N}$ values ($10.6\pm 1.3\text{‰}$) compared to both
292 Generalized A and Semi-thicklipped (8.3 ± 1.3 and $9.2\pm 1.1\text{‰}$, respectively). The maximum
293 difference in mean $\delta^{15}\text{N}$ values between ecomorphs in the Didessa was 4.61‰. The highest
294 difference between ecomorphs in the mean $\delta^{13}\text{C}$ values was 5.10‰. The isotopic niches (as
295 assessed by the standard ellipses) were almost fully separated between Generalized B and V-
296 Scraper (overlap <1%), between Generalized B and Large-mouthed A (overlap <1%), and
297 weakly overlapped between Semi-Thicklipped and V-scraper, Semi-Thicklipped and Large-
298 mouthed A, and Generalized A and V-Scraper (overlap <6%, <10 %, and <11%,
299 respectively) (see details in Supplementary File S4). The most overlapping trophic niches
300 were between Generalized A and B (57 and 90%) and between Generalized A and Semi-
301 Thicklipped (77 and 79%) (Supplementary File S4).

302 *3.2.2. Gojeb Radiation.* Among six ecomorphs, some were significantly divergent from
303 others in SI values (five and six pairwise comparisons in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values, respectively;
304 Table S4). The largest values of $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ were found in Large-mouthed (piscivory)
305 ecomorph ($11.94\pm 0.8\text{‰}$ and $-17.98 \pm 1.8\text{‰}$, respectively) that was notably (sometimes
306 significantly - Figure 5, Table S4) higher than in all other sympatric ecomorphs. The minimal
307 values of $\delta^{15}\text{N}$ were detected in the V-scraper ($8.60\pm 0.6\text{‰}$) and Generalized ($8.61\pm 0.8\text{‰}$)
308 ecomorphs while in the i-Scraper ecomorph the $\delta^{13}\text{C}$ value was at the minimum (-23.84 ± 2.0
309 ‰). The total divergence between ecomorphs was lower in $\delta^{15}\text{N}$ but higher in $\delta^{13}\text{C}$ values
310 compared to the Didessa radiation. In particular, the highest difference between ecomorphs in
311 the mean $\delta^{15}\text{N}$ values achieved 3.34‰, while the same for the mean $\delta^{13}\text{C}$ values was 5.86‰
312 (Figure 5, Table S4). The isotopic niches were almost fully separated between Large-
313 mouthed and V-Scraper (overlap <2%) and between Generalized and Large-mouthed (<4%)
314 while the most overlapping trophic niches were between Generalized and V-Scraper (overlap
315 70 and 91%) (Supplementary File S4).



316

317 Figure. 5. Stable isotope biplots (A) showing mean values and 1 SD, and Bayesian ellipses
318 showing trophic niche widths and overlaps (B) in sympatric ecomorphs of the *Labeobarbus*
319 spp. from the Didessa, Gojeb, Genale, and Sore Rivers. Ellipses with 95% confidence
320 intervals are based on standard ellipses corrected for small sample sizes (SEAc; isotopic
321 niche metrics; SIBER package). Each point corresponds to the individual isotopic value.

322 Abbreviations of the ecomorphs: GEN_A - Generalized A; GEN_B - Generalized B; LIP -
323 Thick-lipped; S_LIP - Semi-thicklipped; Pred_A - Large-mouthed A; Pred_B - Large-
324 mouthed B.

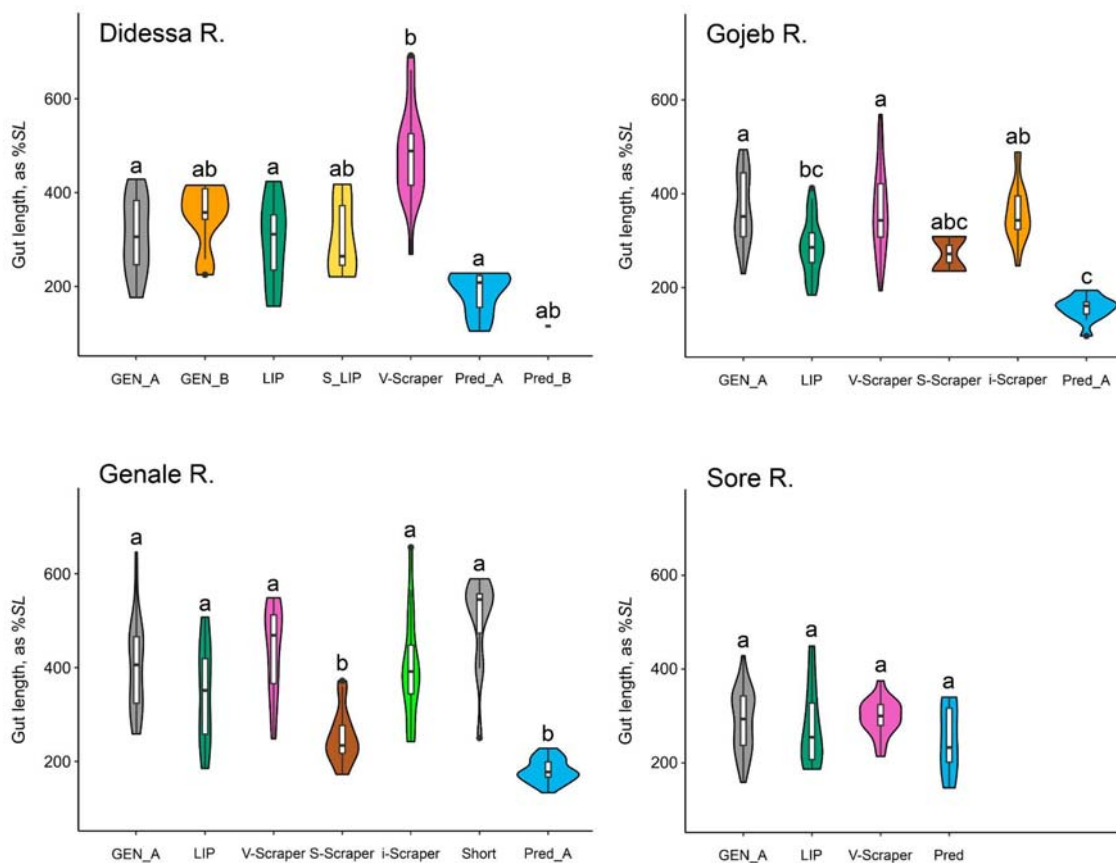
325

326 *3.2.3. Genale Radiation.* Among seven ecomorphs, some were significantly divergent from
327 others in $\delta^{15}\text{N}$ values (eight pairwise comparisons), but not in $\delta^{13}\text{C}$ values (Table S4). The
328 largest value of $\delta^{15}\text{N}$ was found in the Large-mouthed (piscivory) ecomorph ($13.83\pm 0.7\text{‰}$)
329 (Figure 5, Table S4). The minimal values of $\delta^{15}\text{N}$ were detected in the Short and Generalized
330 ecomorphs ($11.15\pm 0.5\text{‰}$ and $11.23\pm 0.8\text{‰}$, respectively). The range between largest and
331 lowest mean $\delta^{13}\text{C}$ values was small, from $-19.87\pm 0.8\text{‰}$ (Short) to $-20.69\pm 1.0\text{‰}$ (S-Scraper).
332 The total divergence between ecomorphs was lower in $\delta^{15}\text{N}$ values than in both the Didessa
333 and Gojeb radiations. In particular, the highest difference between ecomorphs in mean $\delta^{15}\text{N}$
334 values was 2.73‰ , and only 0.82‰ in $\delta^{13}\text{C}$ values. In the isotopic niches the most divergent
335 ecomorph was the Large-Mouthed, which overlap with other ecomorphs ranged from 0.06%
336 (vs. Short) to 38 and 43% (vs. benthophagous S-Scraper). Among other comparisons a low
337 overlap was detected between S-Scraper and Short (7 and 8%) (Supplementary File S4).

338 *3.2.4. Sore Radiation.* Among four ecomorphs, a few were significantly divergent in $\delta^{13}\text{C}$
339 values only (two pairwise comparisons - Generalized vs. Thick-Lipped and Generalized vs.
340 V-Scraper; Table S4). The highest difference between ecomorphs in $\delta^{15}\text{N}$ values was 0.85‰
341 only, while in the mean $\delta^{13}\text{C}$ values it was 1.86‰ . The isotopic niches greatly overlapped
342 ($67\text{...}82\%$) in all comparisons (Supplementary File S4).

343 *3.3. Gut Length.* The relative gut length varied significantly within each radiation except for
344 the Sore River (Figure 6). In three of four radiations, there were significant differences
345 between some ecomorphs (Figure 6; Supplementary File S5). Longest guts (up to $548\text{-}692\%$

346 SL) were detected among V-scrapers (in the Didessa, Genale, and Gojeb Rivers). The large-
347 mouthed ecomorphs showed shortest guts (highest values were up 194-228% SL) except for
348 the Sore River, where it reached $251 \pm 75\%$. Other ecomorphs had intermediate gut length;
349 sometimes they differed from each other like Generalized and Thick-lipped ecomorphs in the
350 Gojeb River (Figure 6). In spite of diverse mouth phenotypes in the Sore River, the
351 ecomorphs were not divergent from each other representing the middle length of the gut
352 (means varied within 251-290% SL).



353

354 Figure 6. Violin plots of relative gut length distribution in sympatric ecomorphs from the four
355 riverine radiations of the *Labeobarbus*. Min-max values (whiskers), 1st and 3rd quartiles
356 (white vertical bars), median values (black horizontal bars), and outliers (black points) are
357 indicated. Letters above the violin plots indicate significant differences between ecomorphs

358 (p < 0.05, Kruskal–Wallis test with Dunn’s post hoc test). Abbreviations of the ecomorphs:
359 GEN_A - Generalized A; GEN_B - Generalized B; LIP - Thick-lipped; S_LIP - Semi-
360 thicklipped; Pred_A - Large-mouthed A; Pred_B - Large-mouthed B.

361

362 **4. Discussion**

363 Our results show that parallel trophic diversifications occurred within the polyploid lineage,
364 the genus *Labeobarbus*. These diversifications differ in degrees of specialization within the
365 same set of core mouth phenotypes (generalized, thick-lipped, scraping, and large-mouthed),
366 indicating various stages of the process. We discuss the results obtained in the context of
367 trophic specializations of certain mouth phenotypes, the preadaptive nature of the ancestral
368 discrete mouth polymorphism, which likely originated via allopolyploidization of the
369 *Labeobarbus* lineage, and attempt to reconstruct the process of trophic diversification by
370 analyzing four repeated cases discovered in the riverine environment of the Ethiopian
371 Highlands.

372 *4.1. Trophic resource partitioning and trophic specializations.*

373 Our study revealed that differences in diet between ecomorphs of the *Labeobarbus* in each
374 river were generally confirmed by SI analysis. For instance, the large-mouthed ecomorphs
375 which had piscivorous diet were also significantly enriched in $\delta^{15}\text{N}$ values (Figures 4 and 5).
376 In contrast, the ecomorphs with detritivorous mode of feeding had lowest $\delta^{15}\text{N}$ values. The
377 periphyton feeder that have been recorded in the Didessa River had a high level of $\delta^{15}\text{N}$
378 values comparable with such in piscivorous ecomorphs but was characterized by enriched
379 $\delta^{13}\text{C}$ values (Figure 5). The invertivorous ecomorphs whose diet was based on the larvae of
380 amphibiotic insects had intermediate levels of $\delta^{15}\text{N}$ values between detritivores and

381 piscivores. The data on gut length were less informative although the piscivorous ecomorphs
 382 had significantly shortened gut (Figure 6) that is in line with literature data for both
 383 *Labeobarbus* (Nagelkerke et al., 1994; Levin et al., 2019) and other fishes (e.g., Hugueny &
 384 Pouilly, 1999; Wagner et al., 2009). We summarized obtained data on the trophic
 385 specialization of sympatric ecomorphs from all rivers based on the diet, gut length, and SI
 386 composition in Table 2.

387

388 Table 2. Correlation between mouth phenotype and trophic specialization in the *Labeobarbus*
 389 spp. from four riverine radiations. The expected trophic niche is based on prediction from the
 390 mouth phenotype and published data on trophic ecology*. Observed trophic niche - data
 391 obtained in this study.

Mouth phenotype (core/subtype)	Expected trophic niche	River and observed trophic niches			
		Didessa	Gojeb	Genale	Sore
1. Generalized A	Omnivore	Detritivore	Detritivore	Detritivore	Detritivore
2. Generalized B	Omnivore	Detritivore- macrophyto phage	-	-	-
3. Thick-lipped	Benthic invertivore	Detritivore- invertivore	Invertivore- detritivore	Detritivore	Detritivore
4. Thick-lipped / Semi-thicklipped	Invertivore ?	Detritivore- macrophyto	-	-	-

	phag				
5. Scraping / V- Scraper	Periphyton feeder	Periphyton feeder	Macrophyto phage	Macrophyto phage	Detritivore- macrophyto phage
6. Scraping / S- Scraper	Periphyton feeder	-	Benthic invertivore	Benthic invertivore	-
7. Scraping / i- Scraper	Periphyton feeder?	-	Macrophyto phage	Detritivore	-
8. Large-mouthed A	Piscivore	Piscivore- omnivore	Omnivore- piscivore	Piscivore	Detritivore- macrophyto phage
9. Large-mouthed B	Piscivore	Piscivore?	-	-	-
No. trophic niches	-	5	5	4	2

392 * References used: Matthes, 1963; Nagelkerke et al., 1994; Golubtsov, 2010; Levin et al., 2019; Teshome et al.,
393 2023; Levin et al., [in press](#).

394

395 Results of this study suggest five main trophic strategies among sympatric
396 ecomorphs of the *Labeobarbus*: i) detritivory, ii) macrophytophagy, iii) invertivorous
397 benthophagy, iv) periphyton feeding, and v) piscivory. Apart those, some ecomorphs had
398 mixed modes of feeding - e.g., detritivory-macrophytophagy, detritivory-invertivory, and
399 piscivory-omnivory (also with inclusion of a noted portion of detritus). Their more flexible

400 trophic specialization might be an adaptive strategy for living in mountain rivers with
401 unstable hydrological regimes (Jepsen & Winemiller, 2002). Although the core of mouth
402 phenotypes (generalized, thick-lipped, scraping, and large-mouthed) was the same in four
403 riverine radiations of the *Labeobarbus*, some phenotypes display more diversified sets of
404 ecologically relevant mouth subtypes. Scraping mouth phenotype was represented by three
405 subtypes, which occupy four different trophic niches (periphyton feeding, detritophagy,
406 invertivorous benthophagy, and macrophytophagy). It is an outstanding example of trophic
407 diversification of scraping mouth phenotype that previously was considered as adapted for
408 feeding via scraping the algal periphyton from stones and rocks (Matthes, 1963; see also
409 Vreven et al., 2016). Recent studies uncovered similar patterns of diversifications within
410 other phylogenetically distant cyprinid lineages bearing the scraping mouth phenotype - the
411 *Garra* and *Schyzopygopsis*, in particular (Komarova et al., 2021; 2022; Levin et al., 2021a).
412 Contrary to the *Labeobarbus* radiations that likely initiated from the generalized ancestor as
413 most ubiquitous throughout the generic range, the highly specialized scraper lineages (*Garra*
414 and *Schyzopygopsis*) could give a trophic radiation outside the ancestral narrow
415 specialization.

416 The Large-mouthed phenotype of the *Labeobarbus* demonstrated further sub-
417 specialization within the adaptive zone of the piscivory in the Didessa River while in other
418 rivers this phenotype had only one feeding strategy that varied from obligate piscivory
419 (Genale) via piscivory-omnivory (Gojeb) to non-piscivory mode of feeding (Sore). The sub-
420 specialization of the Large-mouthed phenotype in the riverine environment is similar in some
421 extent to the more diversified set of specializations discovered in Lake Tana where up to
422 seven sympatric large-mouthed species/ecomorphs were described (Nagelkerke et al., 1994),
423 whose piscivory specialization subdivided in benthic and pelagic zones (de Graaf et al.,
424 2010).

425 A large incongruence between mouth phenotype and expected trophic niche is noted
426 not only for the trophic specialists. The Generalized ecomorph, including Ethiopian species
427 and populations, was considered omnivorous (Matthes, 1963; Teshome et al., 2023), but
428 according to our data it is detritivorous in all studied rivers (Levin et al., 2019; 2023 - **in**
429 **press**; this study). At the same time further diversification within Generalized mouth
430 phenotype (A and B subtypes) have been found. It is the emergence of an ecomorph with
431 horseshoe shape of the lower jaw without mental lobe and deep body in the Didessa River
432 (Generalized B – Figure 2B). This ecomorph had lowest both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values among
433 other sympatric ecomorphs occupying a niche of detritivore-macrophytophage (Figures 4-5).

434 Noticeably, a detritus was the main food not only for generalists but often for thick-
435 lipped phenotype and also for all ecomorphs from the Sore River regardless their mouth
436 phenotypes. We consider detritus as the most available and permanent trophic resource in
437 riverine ecosystems of the Ethiopian Highlands with an unstable hydrological regime. In fact,
438 it is a core food for the Generalized ecomorph and an important food resource for other
439 ecomorphs.

440 Apart from the core mouth phenotypes, the intermediate ones were also found. For
441 instance, i-Scraper phenotype is intermediate between the Scraping and Generalized. Its
442 hybrid origin in the Genale River (by S-scraper and Generalized) was confirmed by genetic
443 data (Levin et al., 2019). Isotopic niche of i-Scraper from the Genale was between parental
444 species (Figure 5) but the gut content was similar to sympatric Generalized ecomorph.
445 Supposedly, i-Scraper intermediate phenotype might have its own trophic sub-niche. Another
446 example of intermediate mouth phenotype that seemingly occupies its own sub-niche refers
447 to the Semi-thicklipped ecomorph (see Figures 4-5) in the Didessa River. Obtained results are
448 consistent with the syngameon hypothesis, proposing that hybridization between members of

449 the radiation can promote further niche expansion and diversification (see Seehausen, 2004;
450 Frei et al., 2022). Intermediate mouth phenotypes were reported among the *Labeobarbus* –
451 between scraping ‘*Varicorhinus*’-like and generalized phenotypes by Banister (1972) and
452 Nagelkerke and Sibbing (1996) as well as between scraping and thick-lipped mouth
453 phenotypes by Vreven et al. (2019). In the last case the intermediate phenotype resembled the
454 generalized one (Generalized B in our study). Authors revealed a hybrid nature of this
455 phenotype by genetics (Vreven et al., 2019) but there was no data on trophic ecology or
456 specialization. Actually, the recruiting of novel trophic niches via hybridization is an
457 intriguing issue that is yet weakly studied (but see experimental study of Selz and Seehausen,
458 2019).

459 In summary, the *Labeobarbus* trophic radiations involve a mixture of pelagic
460 predator (piscivore) and a wide assortment of benthic-oriented specialists. The benthic-
461 pelagic habitat axis is most frequent in cyprinid and cichlid adaptive radiations in both lakes
462 and rivers (Nagelkerke et al., 1994; Cooper et al. 2010; Levin et al., 2019; Burress et al.,
463 2023).

464 4.2. *Decoupled form and function – preadaptive phenotypes and their functionalization.*

465 Our results show that the same mouth phenotypes in the *Labeobarbus* may occupy different
466 trophic niches. A mismatch between expected and observed trophic specializations may have
467 different nature: i) specialized phenotype has unspecialized (generalized or omnivorous)
468 feeding mode that is known as Liem’s paradox (Liem, 1980; Robinson et al., 1998), and ii)
469 specialized phenotype has unexpected (biased) trophic specialization possibly due to
470 insufficient knowledge on trophic specializations in certain lineages. We focus in this study
471 on the first phenomenon. Liem’s paradox was established for cichlid fishes (e.g., Liem, 1980;
472 Sturmbauer et al., 1992; Wagner et al., 2009; Binning et al., 2009; Torres-Dowdall & Meyer,

473 2021) but it was also detected in other taxonomic groups (e.g., in cyprinids – Lammens et al.,
474 1991; Levin et al., 2021a; Komarova et al., 2022). The mismatch between form and function
475 might be i) a temporary phenomenon for proper specialist explained by plasticity of its diet in
476 some circumstances (ecological release that may be a base for evolutionary re-specialization
477 – see above-mentioned example of highly specialized scraping periphyton feeders of the
478 genus *Garra* that could re-specialize in other trophic specializations – Komarova et al., 2022)
479 or ii) heterochronous decouple of form and function when the emerging phenotype is not yet
480 functionalized, i.e. phenotype is preadaptive (expectation on its trophic adaptation based on
481 phenotypic features) but not yet involved in trophic resource partitioning. This is exactly the
482 case of the mouth diversity in the Sore River, where detritus was a main food for all mouth
483 phenotypes including the Large-mouthed, i.e. mouth divergence was uncorrelated with the
484 use of food resources. In other words, mouth polymorphism in the Sore River is ecologically
485 almost non-functional compared to other riverine diversifications. This contradicts the
486 ‘ecological theory’ of adaptive radiation in neglecting the ‘habitat first rule’ in particular (see
487 Schluter, 2000) but fits the flexible stem hypothesis (Wund et al., 2008; Gibert, 2017) or its
488 variant known as the ‘plasticity-first’ evolution (Levis & Pfennig, 2016; 2019). According to
489 this hypothesis, an adaptive phenotypic plasticity in an ancestral population could precede
490 adaptation to a new environment through the process of genetic assimilation. Remarkably, the
491 case of decoupled form and function in the *Labeobarbus* is shared to some extent with such
492 in riverine adaptive radiations in the pike cichlids, genus *Crenicichla* (Burress et al., 2023)
493 that may be a general feature for young radiations in riverine environments.

494 As already mentioned, the repeated mouth polymorphisms of the *Labeobarbus* may
495 be predetermined due to its polyploid origin (Yang et al., 2022). Its maternal lineage is
496 characterized by generalized / thick-lipped mouth polymorphism widely distributed among
497 Torinae in the genera *Tor* and *Neolissochilus* (Hoang et al., 2015; Walton et al., 2017) – so

498 the generalized/thick-lipped polymorphism is persistent within a Torinae lineage. Paternal
499 lineage referred to contemporary genus *Cyprinion* is represented by scraping mouth
500 phenotype (Coad, 2021). Hence, mouth polymorphism of the *Labeobarbus* is ancestrally
501 heritable and re-evolved under particular ecological circumstances. Given this, the thick-
502 lipped and scraping mouthed phenotypes of the *Labeobarbus* in the Sore River are seemingly
503 ‘preadaptive’ upon emergence *de-novo*.

504 Modular-assembling genome of the *Labeobarbus* consisted of the different
505 subgenomes (Yang et al., 2022) that is able to produce discrete mouth phenotypes inherent to
506 parental lineages is formally still within ‘flexible stem hypothesis’. However, from another
507 hand, it might be also in frame of ‘transporter process’ (Schluter & Conte, 2009; Marques et
508 al., 2019; Martin & Richards, 2019) meaning that the adaptive alleles and genetic
509 architectures differentiating each species within a rapid radiation are older than the radiation
510 itself. The oldest paleorecords of the *Labeobarbus* in East Africa are dated by Late Miocene
511 (Stewart & Murray, 2017). Some trophic specialists, for instance, with scraping phenotype
512 are remarkably older than the recently emerged repeated radiations in the Ethiopian
513 Highlands under consideration. Their age is dated as Pleistocene according to molecular
514 clocks (Beshera et al., 2016). It suggests that similar mouth diversification might evolved
515 repeatedly before the diversifications we observe in this study. Therefore, the genomes of the
516 modern *Labeobarbus* might be rather experienced, i.e. may have the standing genetic
517 variation contributing to the ecological speciation and diversification. Regardless which is the
518 certain prerequisite for adaptive radiation of the *Labeobarbus* – ‘flexible stem’ or ‘transporter
519 process’ (or their combination) – it might significantly facilitate the diversification.

520 *4.3. Possible scenario of evolution of trophic radiation: from incipient to matured*

521 Our results demonstrate that four riverine assemblages of the *Labeobarbus* in the Ethiopian
522 Highlands being similar in the core mouth phenotypes and direction of trophic diversification
523 are at various stages of the evolution. The number of mouth phenotypes together with
524 subtypes and their ecologic functionality in the *Labeobarbus* varied significantly from river
525 to river. We tried to range the cases from simplest to most matured based on the obtained
526 results – diversity of ecomorphs/mouth phenotypes and their functionality (as estimated by
527 the gut content and isotopic niche) within each radiation (Table 3; Figures 4-5).

528 Table 3. Ranking the diversifications by number of ecological niches and other aspects of
529 trophic differentiation. Number of points in rows 3 and 4 corresponds to the number of
530 significant differences in pairwise comparisons between sympatric ecomorphs (taken from
531 Supplementary File S4).

Nos.	Parameters	Sore	Genale	Gojeb	Didessa
1	Nos. mouth phenotypes	4	6	6	7 ¹
2	Nos. trophic niches ²	2	4	5	5
3	Nos. dif. in $\delta^{15}\text{N}$	0	8	5	6
4	Nos. dif. in $\delta^{13}\text{C}$	2	0	6	6
5	Nos. trophic levels ³	0.36	1.18	1.45	2.0

532 ¹Eight ecomorphs were detected in the Didessa but only seven were included in the study due to absence of data
533 on the exceptionally rare Large-mouthed ecomorph. ² Number of trophic niches is taken from Table 2. ³ One
534 trophic level is arbitrary taken as 2.3‰ between mean $\delta^{15}\text{N}$ values according to McCutchan et al. (2003) and
535 Vanderklift & Ponsard (2003): the number of trophic levels was counted by dividing the difference between
536 minimal and maximal mean $\delta^{15}\text{N}$ values within a pool of sympatric ecomorphs by 2.3.
537

538 The trophic diversifications could be ranged from the most undifferentiated in the Sore River
539 via the Genale and Gojeb Rivers to the most diversified in the Didessa River (Table 3, Figure
540 7). Next stages of the trophic diversification can be provisionally outlined (see scheme on
541 Figure 7):

542 (1) Burst of mouth polymorphism. A set of core mouth phenotypes quickly emerges but
543 their ecological functionalization is immature (preadaptive phenotypes).
544 Morphological phenotypes are trophically irrelevant or there is a weak phenotype-
545 trophic correlation (Liem's paradox). At this stage phenotypic diversity strongly
546 dominates over the ecological diversity - see the case of the Sore River (four
547 phenotypes vs. two trophic niches).

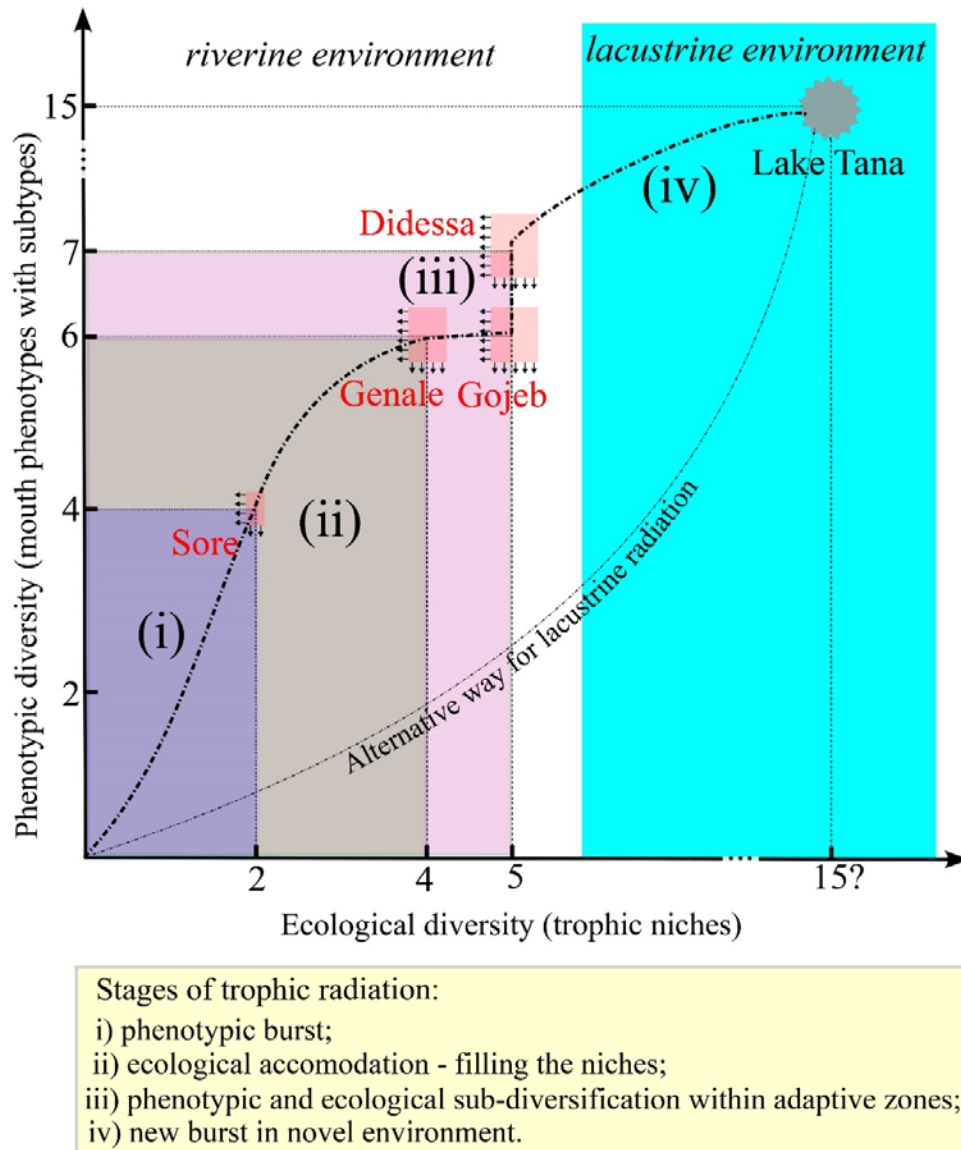
548 (2) Filling the trophic niches (ecological accommodation). Preadaptive phenotypes began
549 to be functionalized by occupation of the trophic niches best matched to the mouth
550 type. This process might be named as ecological accommodation of phenotypic trait –
551 analogously to term 'phenotypic accommodation' sensu West-Eberhard (2003) The
552 numbers of phenotypes and trophic niches are almost equal. Probably it is a very
553 rapidly realizing process else it is hard to explain what are reasons to sustain non-
554 functional mouth polymorphism.

555 (3) Continued diversification within the core mouth phenotypes with corresponding
556 increase of the ecological diversity. Upon filling the most evident trophic niches, a
557 further sub-diversification occurs in some radiations. It might go by various scenarios
558 - *via* ecological sub-specialization of certain phenotypes or *via* hybridization and
559 functionalization of intermediate phenotypes. At matured stages of the diversification,
560 the biotic interactions may serve as an additional ecological opportunity ('diversity
561 begets diversity' – Martin & Richards, 2019). Significant increase of the populations
562 size during morpho-ecological diversification (due to utilization of various trophic

563 resources in a river) makes the piscivory strategy more reliable that may result in sub-
564 diversification within this adaptive zone - like in the Didessa River (see also
565 cascading speciation - Broderson et al., 2017; Bracewell et al., 2018).

566 (4) A new diversification burst provoked by colonization of the novel environment might
567 occur as exemplified from the lacustrine burst of the *Labeobarbus* in Lake Tana
568 (Ethiopia) resulted in evolution of 15-16 species/ecomorphs (Nagelkerke et al., 1994;
569 Mina et al., 1996; Sibbing, Nagelkerke, 1998; Nagelkerke et al., 2015; Beshera and
570 Harris, 2023). It is still unknown whether Lake Tana radiation evolved from a single
571 generalized ancestor or stemmed from already diversified set of riverine radiation.

572



573

574 Figure 7. Schematic reconstruction of process of morpho-ecological diversification of the
 575 *Labeobarbus* in the Ethiopian Highlands. The number of little arrows at each box correspond
 576 to the numbers of mouth phenotypes (horizontal arrows) and numbers of trophic niches
 577 (vertical arrows).

578

579 The most immature diversification has been detected in the Sore River where a form
 580 (mouth phenotype) and its ecological function were largely decoupled. The ecologically non-

581 functional or non-matured mouth polymorphism in the Sore River might be explained by
582 several reasons. First, it might be due to a very young age of this radiation (at stage of
583 incipient diversification) that is supported by low genetic diversity and absence of mtDNA
584 haplotype sorting (Levin et al., 2020). Another reason refers to insufficient ecological
585 opportunities in the river to realize the existing potential (mouth polymorphism).
586 Heterogeneous and unpredictable environment in the rivers (flow regimes, turbidity, depth,
587 water chemistry, temperature, food availability, etc.) may weaken the divergent selection
588 necessary to drive specialization, instead favoring opportunistic ecological roles such as
589 detritivory-omnivory (Jepsen and Winemiller, 2002; Burress et al., 2023). But the Sore River
590 is a home for another bright adaptive radiation of hillstream cyprinid fish of the genus *Garra*
591 that is represented by six genetically, morphologically and trophically divergent young
592 species (Golubtsov et al., 2012; Levin et al., 2021a; Komarova et al., 2022). Possible
593 overlapping in the trophic specializations (predator, periphyton feeding, benthic invertivore)
594 with more advantageous cyprinid radiation in shared environment can acutely suppress or
595 constrain the diversification process in the Sore population of the *Labeobarbus* resulting in
596 freezing the non-functional mouth polymorphism stage for a while. Simultaneously, other
597 studied riverine populations of the *Labeobarbus* have represented more advanced trophic
598 diversification stages in absence of other sympatric radiations (as far as we know) and
599 characterized by matured or ecologically functional mouth phenotypes.

600

Conclusions

601 Using the polyploid African barbs of the genus *Labeobarbus* as model, we showed the
602 parallel morpho-ecological diversifications being similar in this phenomenon with other
603 prominent examples of adaptive radiation among vertebrate animals like cichlids (Seehausen,
604 2015; Torres-Dowdall & Meyer, 2021; Burress et al., 2023) and Caribbean *Anolis* (Losos,

605 2009; Stroud & Losos, 2020). The most striking difference from above-mentioned examples
606 is the sharply preadaptive nature of inherited mouth polymorphism as a starter of radiation.
607 Likely, the diversification of the *Labeobarbus* starts from the onset of genomic pre-existing
608 templates of mouth polymorphism. The emerged mouth phenotypes are preadaptive but not
609 (or weakly) functional at the incipient stage of trophic diversification demonstrating
610 decoupled form and function. Enhancement of ecological functions of preadaptive
611 phenotypes (ecological accommodation) increases upon filling the trophic niches and result
612 in maturation of trophic specialization that is often continued by further sub-diversification of
613 mouth types and sub-specialization within previously occupied adaptive zones. The
614 preadaptive mouth polymorphism can be considered a key innovation of the *Labeobarbus*
615 that makes the bridge between phenotypic diversification and ecological opportunities
616 remarkably shorter that might accelerate diversification rates. It apparently explains why this
617 lineage overcomes the obstacles in unstable riverine environments unfavored for adaptive
618 radiations by other fish lineages.

619

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634 **Data availability.** Raw isotopic data are available as Supplementary data.

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639 preparation and their analysis; A.S.K. performed diet analysis; All authors wrote the
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