

# A whole-body micro-CT scan library that captures the skeletal diversity of Lake Malawi cichlid fishes

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## ABSTRACT

Here we describe a dataset of freely available, readily processed, whole-body  $\mu$ CT-scans of 56 species (116 specimens) of Lake Malawi cichlid fishes that captures a considerable majority of the morphological variation present in this remarkable adaptive radiation. We contextualise the scanned specimens within a discussion of their respective ecomorphological groupings and suggest possible macroevolutionary studies that could be conducted with these data. We also describe a methodology to efficiently  $\mu$ CT-scan (on average) 23 specimens per hour, limiting scanning time and alleviating the financial cost whilst maintaining high resolution. We demonstrate the utility of this method by reconstructing 3D models of multiple bones from multiple specimens within the dataset. We hope this dataset will enable further morphological study of this fascinating system and permit wider-scale comparisons with other cichlid adaptive radiations.

## Background & Summary

Cichlids are one of the most speciose families of vertebrates, with over 1000 species in the African Rift Valley alone<sup>1,2</sup>. Multiple, independent, adaptive radiations of these fishes have evolved in the Great Lakes of East Africa, their associated satellite water bodies, as well as their connecting riverine systems. The radiations of these fishes (Subfamily: Pseudocrenilabrinae<sup>3</sup>), particularly those associated with Lakes Malawi, Victoria and Tanganyika, have become powerful models for the study of macroevolutionary processes<sup>4-10</sup>, behaviour and physiology<sup>11-15</sup>, and have emerged more recently as models in evolutionary developmental biology<sup>16-19</sup>.

Lake Malawi haplochromine cichlids represent a particularly speciose, phenotypically diverse but genetically homogeneous adaptive radiation of lacustrine fishes. This diversity, comprising approximately 850 species of maternal mouthbrooders, is the most extensive adaptive radiation of vertebrates so far identified<sup>1,9</sup>. Molecular clock analyses estimate the radiation to be approximately 800 thousand years old<sup>4</sup>, a relatively young radiation when compared to the older system of Lake Tanganyika (~10myr) which contains just 250 species<sup>7,20</sup>. Despite their high phenotypic diversity, genetic variation in Lake Malawi cichlids is extremely low. Whole genomic comparisons of representatives from all seven distinct ecomorphological groups within Lake Malawi, estimated an average DNA sequence divergence of just 0.19-0.27%<sup>4</sup> – a range comparable to that within human populations<sup>6</sup>. In addition, a relatively low DNA mutation rate; that alone cannot account for the estimated divergence time of Lake Malawi cichlids<sup>4,6</sup>, overlapping distributions of interspecific pairwise sequence differences and heterozygosity (intraspecific genetic variation)<sup>4</sup> only further complicates this enigmatic adaptive radiation.

East African cichlids, including those belonging to the Lake Malawi radiation, have recently emerged as powerful models in evolutionary developmental biology<sup>17</sup>. Evolutionary modification of embryological mechanisms drives the evolution of novel adaptations and requires genetic variation<sup>21</sup>. Thus, comparing the embryological development of cichlids, which have limited genetic variation, can enable us to identify specific cases where evolution has modified developmental mechanisms. The diversity of feeding habits of Lake Malawi cichlids, and the ability to causally link morphological differences in craniofacial morphology to these ecological niches, has enabled integrative genetic and morphological studies examining the evolution of these traits<sup>22-25</sup>. More recent studies have expanded the scope beyond craniofacial phenotypes, including pigmentation patterning<sup>26-28</sup>; body and fin shape<sup>19,29</sup> and axial elongation<sup>18</sup>. In parallel, aided by developments in whole-genome sequencing

40 technologies<sup>6</sup>, it has been possible to considerably improve our understanding of the phylogenetic relationships among Lake  
41 Malawi cichlids<sup>4,9,30,31</sup>. Previously intractable macroevolutionary studies, such as the convergent evolution of hypertrophied  
42 lips<sup>32</sup> can now take advantage of the relatively robust phylogenies based on whole-genome sequences. Moreover, there are  
43 now opportunities to use this new phylogenetic information to focus on the evolution of other traits, such as the axial and  
44 appendicular skeleton, that is of key importance in teleost diversification<sup>33,34</sup>. However, a whole-body  $\mu$ CT-scan dataset of  
45 Lake Malawi cichlid fishes has not yet been described.

46 Here we present a new database of high-resolution X-ray micro-computed tomography ( $\mu$ CT) scans of Lake Malawi  
47 cichlids, providing 3D data on skeletal morphology for the whole body of 56 species across 26 genera. In total these data  
48 comprise 116 individuals from seven recognized ecomorphological groupings<sup>4</sup> (Table 1), contrasting in multiple aspects of  
49 morphology, size, behaviour, and habitat preference. We demonstrate the resolution and utility of our dataset by illustrating  
50 3D whole-body renderings of several species, and of several skeletal regions of interest. The data will be useful resource  
51 for researchers interested in the emergence of morphological variation, including the macroevolutionary patterns common to  
52 adaptive radiations<sup>35,36</sup>. We argue that where digitisation efforts are being taken to characterise adult cichlid morphology, whole-  
53 body scans should be standard to ensure that other sources of morphological variation can be investigated. We propose methods  
54 that minimise scanning time, thus alleviating financial and time constraints of scanning large numbers of specimens, whilst  
55 maintaining sufficiently high resolution for macroevolutionary studies and geometric morphometric analyses of morphological  
56 variation.

## 57 **Methods**

### 58 **Sample Selection**

59 There are an estimated 850 species of Lake Malawi cichlid fishes<sup>4</sup>. Many of which have not been described, preserved in  
60 museum collections or are available on phylogenies based on whole genome evidence. Therefore, to maximise the utility and  
61 morphological variation captured by our dataset, we focused on species present in published phylogenies<sup>4,9,24</sup>, and sought to  
62 include as many genera as possible in dataset (see Table 1). We prioritised scanning the type species for genera, and avoided  
63 inclusion of species which already had whole body scans freely available online. Our specimens were sourced from the  
64 collections at the Natural History Museum in London (NHMUK), from the School of Biological Sciences of the University of  
65 Bristol (Martin Genner) and from the School of Natural Sciences of Bangor University (George Turner). In total we scanned  
66 116 specimens from 56 species (Supplementary Table 1). Of these 99 were wild-caught, and 17 were laboratory-reared. These  
67 laboratory-reared fish included *Astatotilapia calliptera* (Mbaka River, n=10), *Maylandia zebra* (Boadzulu island, n=5) and  
68 *Rhamphochromis* sp. 'Chilingali' (n=2), all of which died naturally or were euthanised by anaesthetic overdose [Schedule 1;  
69 Animals (Scientific Procedures) Act 1986].

### 70 **$\mu$ CT-Scanning**

71 Since there was already a large collection of specimens present at the Natural History Museum in London (NHMUK) and in the  
72 extensive research collections of Martin J Genner (School of Biological Sciences, University of Bristol) and George F Turner  
73 (School of Natural Sciences, Bangor University) we decided to take advantage of the scanners present in the CT facility of  
74 NHMUK and at the XTM Facility based in the Paleobiology Research Group at the University of Bristol. Of the total 116  
75 individuals scanned (56 species), 56 specimens (28 species) were scanned at the NHMUK Imaging and Analysis Centre and 60  
76 specimens (28 species) were scanned at the XTM Facility at the University of Bristol. A flowchart, describing all the necessary  
77 decisions and required processing steps is provided as Figure 2.

### 78 **Scanning Arrangement**

79 To maximise the utility of our time and the number of species scanned, multiple specimens were scanned in each individual  
80 scan (3). Each batch of specimens was fit to the width of the scan field of view to maximise resolution, and multiple scans were  
81 conducted along a the vertical axis in order to scan the full body length of each specimen. Practically, this meant specimens  
82 were arranged into batches of similar total length. Batch sizes varied between two and five specimens, with the number of  
83 each batch ultimately dependent upon the overall size of the specimens within the batch. Of the 32 batches scanned: 20 were  
84 comprised of four specimens; nine of three specimens, and two and one batch(es) of one and five specimens, respectively. Since  
85 multiple individuals of different species were often scanned together (Figure 3A, B) and it was critical that individuals of the  
86 same species could be readily identified. Therefore, unique, low density objects such as plastic bricks, pipette tips and rubber  
87 bands were placed in physical proximity to each specimen, to act as recognisable markers (Figure 3C) which would readily  
88 resolve in the reconstructed image stacks (see Post-Scanning Processing). These objects were attached to each individual  
89 specimen-containing bag, and the specimens were bundled together, ideally ensuring that the objects faced outwards (Figure  
90 3D-F). Specimens were tightly packed into plastic containers, sealed with tape, and allowed to rest upright (head-up) for at least  
91 ten minutes so the contents could settle to prevent movement during scanning (Figure 3G-K). We were able to scan, on average,

92 23 specimens per hour at maximal efficiency, an efficiency that was primarily the result of having two people per scanning  
93 visit (one scanning and one packing). The scanning rate could be further increased by packaging specimens in advance of the  
94 scanning so that each batch can be scanned continuously.

### 95 **Scanning Procedure**

96 Prior to scanning, a visual inspection of the specimens was made following X-ray exposure to check for internal damage (such  
97 as damage to the vertebral column). If present, and possible, specimens were switched for another specimen of the same batch.

98 To maximise scan resolution, each batch of fish was scanned using the full width of the scanner field of view. This  
99 necessitates that multiple, overlapping scans were conducted along the vertical axis of the scanner (Z-axis) in order to capture  
100 the full body length of each fish. The overlapping scans were subsequently stitched together in the processing stage using the  
101 software, [VGStudio Max 3.2.5 64-bit](#) (see below). The maximise efficiency, individuals with similar body lengths were scanned  
102 together (see above). The fish scanned ranged in standard length of 3.8cm to 33cm, which meant that scanning parameters  
103 varied between batches. The number of projections generated range from 901 to 2001. Exposure times varied between 250 and  
104 500 seconds, with a median of 354 seconds. Similarly, power varied greatly between scans, ranging between 12.48 and 37.995  
105 watts (mode 37.44W). All scanning parameters, for each batch conducted, as well as for each individual scan can be found in  
106 the metadata supplied in Supplementary Table 1.

### 107 **Post-Scanning Processing**

108 All processing and segmentation was conducted on a machine specifically built for image analysis, with the following  
109 specifications: 2 x Intel® Xeon® CPU-ES-2640 @2.60GHz, 2601MHz, 8 Core(s) processors, 128GB of dedicated DDR3 RAM  
110 running on Microsoft Windows 10 Pro (Build Number: 10.0.19045). Raw, isometric volumes generated from the CT-scanning  
111 were imported into [VGStudio Max 3.2.5 64-bit](#) and anterior and posterior halves were subsequently stitched together by  
112 defining overlapping regions of interest in both the anterior, middle (if applicable) and posterior volumes. 16-bit tiff stacks were  
113 exported from [VGStudio Max 3.2.5 64-bit](#) and imported into FIJI<sup>37</sup>, a GUI for ImageJ<sup>38</sup>. In FIJI, individual fish were cropped  
114 out of the 16-bit stacks, which were identifiable due to the unique objects associated with each individual (see above). The  
115 brightness was adjusted by extending the distribution of pixel values to remove 0 values, and the tiff stacks were converted into  
116 8-bit to decrease file size. Total stack file size was further decreased by removing images at the beginning and end of the stacks  
117 that did not contain readily identifiable bone.

### 118 **Data Availability**

119 Cropped 8-bit tiff stacks of all 116 specimens have been deposited into a dedicated project on Morphosource, accessible [here](#),  
120 where they are freely available and can be downloaded by anyone. We provide a short discussion about segmenting, including  
121 methods to optimise these reconstructed image stacks below. We politely request that a direct reference be made to this  
122 paper if any scans from our dataset are used in future analyses. See Supplementary Table 1 for a breakdown of all scanned  
123 specimens.

### 124 **Data Records**

125 Seven major ecomorphological groupings make up the Lake Malawi haplochromine radiation. The *Diplotaxodon* and  
126 *Rhamphochromis* groups are comprised of elongate piscivores and zooplanktivores adapted to the deeper and more open  
127 water niches within Lake Malawi<sup>39,40</sup>. The ‘shallow benthic’ group is speciose set of Lake Malawi cichlids<sup>41</sup>, comprised of a  
128 morphologically diverse array of species found across a range of habitats, but is largely restricted to benthic habitats less than  
129 50m. The ‘deep benthic’ is a comprised a group of species specialised for deep water benthic habitats, typically more than 50m  
130 in depth. The ‘utaka’ are a group of typically shallow open water shoaling species, primarily represented by members of the  
131 genus *Copadichromis*<sup>42</sup>. The ‘mbuna’ are a distinct clade of relatively small-bodied rock-associated species typically found  
132 to depths of ~30m, and with a diverse mix of foraging strategies<sup>43</sup>. The final grouping is represented by only one species,  
133 *Astatotilapia calliptera*, that is often present in shallow macrophyte-rich marginal habitats of the Lake Malawi. It is often  
134 referred to as a generalist species — consuming phytoplankton, zooplankton and littoral arthropods. It the only representative  
135 of the haplochromine radiation that is widespread in rivers and smaller lakes of the Lake Malawi catchment<sup>44,45</sup>. Given the  
136 number of species sampled in our dataset, the inclusion of several undescribed species as well as limited species-specific data  
137 for some of the species in our dataset, we are unable to offer descriptions of every specimen within our dataset. We have instead  
138 limited our wider discussion of species within some of the ecomorphological groups to a selection of well-studied species that  
139 demonstrate the diversity captured within our dataset.

### 140 ***Diplotaxodon* and *Rhamphochromis***

141 Nine species of the *Diplotaxodon* group (47%; Table 1), including the type species *Diplotaxodon argenteus* (n=1) and  
142 *Pallidochromis tokolosh* (n=2) are present within our dataset. We also sampled 10 species of the *Rhamphochromis* group

143 (71%; Table 1), including the type species *Rhamphochromis longiceps* (n=2) and the remarkably large *Rhamphochromis*  
144 *woodi* (n=2, see below), that are endemic to Lake Malawi. We were also able to sample two sympatric species from the crater  
145 lake, Lake Kingiri, *Rhamphochromis* sp. “Kingiri Dwarf” (n=2) and *Rhamphochromis* sp. “Kingiri Large” (n=2), as well as  
146 *Rhamphochromis* sp. ‘Chilingali’ (n=4) from the satellite, Lake Chilingali (Figure 4), which is now presumed extinct in the  
147 wild<sup>46</sup>.

148 The *Diplotaxodon* and *Rhamphochromis* groups are two reciprocally monophyletic diverging lineages of Lake Malawi  
149 cichlids<sup>4</sup> that have adapted to the pelagic zone of Lake Malawi<sup>40</sup>. The majority of species in the groups are piscivorous,  
150 although several species, including *Diplotaxodon limnothrissa* (n=2) are predominantly zooplanktivorous<sup>47</sup>. Large-bodied  
151 *Rhamphochromis* primarily feed on Lake Malawi sardines (usipa; *Engraulicypris sardella*) and endemic cichlids (e.g. utaka).  
152 Members of *Diplotaxodon* and *Rhamphochromis* are among the deepest-living of all Lake Malawi cichlids, with representatives  
153 of both being caught at depths exceeding 200 metres – the ‘twilight zone’ where light is almost completely absent<sup>40</sup>. Species  
154 within the *Diplotaxodon macrops* complex, which is represented in the scanned samples by *Diplotaxodon macrops* (n=1) (Figure  
155 4), *Diplotaxodon* sp. ‘macrops north’ (n=2), *Diplotaxodon* sp. ‘macrops black dorsal’ (n=2) and *Diplotaxodon* sp. ‘macrops  
156 ngulube’ (n=2), have been found between 100 and 220m, a depth similarly reported to be occupied by *Rhamphochromis* during  
157 the day<sup>48</sup>.

158 Morphological comparisons of *Diplotaxodon* and *Rhamphochromis* with Lake Malawi cichlids from other habitats could  
159 provide valuable insights into convergent adaptation of traits enabling occupation of pelagic niches. Divergence along depth  
160 gradients is associated with the evolution of reproductive isolation in many marine and freshwater species groups, likely a  
161 consequence of the strong selective pressures associated with deeper water, such as the absence of sunlight, greater hydrostatic  
162 pressure, and reduced levels of dissolved oxygen<sup>49,50</sup>. Morphological comparisons of *Diplotaxodon* and *Rhamphochromis*,  
163 against closely related littoral species, could be a powerful model for the study of evolution of convergent phenotypes necessary  
164 for adapting to pelagic environments. Body elongation, supported by increased vertebral counts, is a common adaptive trait in  
165 teleosts adapted to pelagic (and piscivorous) niches<sup>51</sup> and is also the case with *Rhamphochromis*<sup>52</sup>. Vertebral morphology has  
166 also been related to swimming kinematics, body shape and habitat preference<sup>53</sup> and the evolutionary modification of internal  
167 vertebral traits appears to have taken place during adaptation to pelagic environments<sup>34</sup>. A study of these traits using the Lake  
168 Malawi may help to determine the rate at which these phenotypes can become fixed, and provide insight into the role of these  
169 morphological adaptations along the benthic-pelagic speciation axis<sup>49,50</sup>.

170 Remarkable size variation is present within the *Rhamphochromis* genus. *Rhamphochromis woodi* is considered to be  
171 one of the largest Lake Malawi cichlids, measuring up to 40 cm standard length<sup>39</sup>. In contrast, wild-caught specimens of  
172 *Rhamphochromis* sp. ‘Kingiri Dwarf’, endemic to the crater lake Kingiri, do not exceed 7.5 cm standard length<sup>46</sup>. Similarly,  
173 wild caught *Rhamphochromis* sp. ‘Chilingali’ are also small bodied, with maximum observed standard length of 10.6 cm<sup>8</sup>.  
174 Notably *Rhamphochromis* sp. ‘Chilingali’ is relatively amenable to laboratory study, and its elongated body has made it a  
175 useful model in evolutionary developmental biology<sup>17,18</sup>. Given the exceptional size difference in the genus *Rhamphochromis*,  
176 our dataset represents a potentially valuable resource for the study of the evolution of allometric scaling, which has not been  
177 well studied in cichlids<sup>54</sup>.

## 178 Shallow Benthic

179 The shallow benthic species group is extremely speciose, with remarkable amount of morphological diversity<sup>4,41</sup>, and hundreds  
180 of species. The majority of shallow benthic species inhabit relatively shallow inshore habitats of Lake Malawi, such as the sand  
181 or mud lake floor, or sand-rock transitional zones. Our dataset includes 20 shallow benthic species in 12 genera<sup>4</sup> (Table 1),  
182 including several large, ambush predators, as well as a collection of trophic specialists. For a complete list of shallow benthics  
183 see Supplementary Table 1.

184 Large ambush predators represented in the dataset include *Dimidochromis strigatus* (n=1), *Dimidochromis compressiceps*  
185 (n=1), *Tyrannochromis macrostoma* (n=1), *Nimbochromis livingstonii* (n=1) and *Nimbochromis polystigma* (n=2). *Dimi-*  
186 *dochromis compressiceps* has a generalist piscivore lifestyle, and occupies reed-beds of the Lake. *Nimbochromis livingstonii* and  
187 *N. polystigma* are both considered to be ‘sleeper’ (Chichewa: “kaligono”), and have been observed burying themselves within  
188 sandy substrate and snatching prey attracted by the disturbed sediment<sup>41</sup>. Another member of *Nimbochromis*, *Nimbochromis*  
189 *linni* (n=1) has a characteristic downward-projecting snout (Figure 4), enabling it to extract prey from rock crevices<sup>41,55</sup>.

190 We sampled several shallow-benthic predators, including *Otopharynx speciosus* (n=2), one of the few piscivores within the  
191 genus *Otopharynx*. Males of this species have been encountered at depths exceeding 25m<sup>41</sup>, suggesting tolerance of relatively  
192 deep water, and suggesting the species may have morphological adaptations enabling occupation of deep-water niches similar  
193 to *Rhamphochromis* and *Diplotaxodon*. Of the approximately 20 species of *Otopharynx*<sup>56</sup> we were able to sample an additional  
194 three species: *Otopharynx lithobates* (n=3, including the holotype NHMUK 1974.7.5.1); *Otopharynx tetrastigma* (n=2); and  
195 the undescribed *Otopharynx* sp. “brooksi nkhatta” (n=1). We also sampled several specialised trophic specialists including the  
196 molluscivores *Mylochromis anaphyrmus* (n=1) and *Trematocranus placodon* (n=1) and the invertebrate picker *Placidochromis*



197 *johnstoni* (n=1, Figure 4). The diet of *T. placodon* predominately comprises the gastropods *Bulinus nyassanus* and *Melanoides*  
198 *tuberculata*<sup>57</sup>. Enlarged sensory pores and lateral lines form a sonar-like detection system that allows *T. placodon* to sense the  
199 movement of these prey within the sediment. Curiously, this strategy and associated morphological characteristics are also  
200 associated with *Aulonocara* and *Lethrinops*, both ‘deep benthics’, suggesting convergent evolution of lateral line phenotypes<sup>9</sup>.  
201 The specimens in our dataset may enable some morphological comparisons to further investigate differences in the sensory  
202 pore characteristics among species.

### 203 ‘Rock-dwelling’ Mbuna

204 The mbuna group dominate the rocky shores of Lake Malawi, and are used as a model system for the study of rapid speciation and  
205 adaptive radiation<sup>25,43,58</sup>. Similar to the shallow-benthics, there are hundreds of species, many of which are undescribed<sup>41,43</sup>.  
206 We aimed to maximise our coverage of the phenotypic diversity in the group by sampling multiple genera, which are largely  
207 differentiated on the basis of head, jaw and tooth morphology<sup>43</sup>. Our dataset includes 7 species (15 individuals) of mbuna,  
208 covering 7 of the 14 described mbuna genera (Table 1).

209 *Cynotilapia* can be distinguished from other genera by the presence of unicuspid (conical) teeth<sup>41,59,60</sup> and is represented in  
210 our dataset by *Cynotilapia axelrodi* (n=1, Figure 4). This is a genus of typically planktivorous species<sup>41</sup> and their relatively  
211 simple dentition may reflect this lifestyle<sup>61</sup>. By contrast, *Maylandia* (*Metriaclima*)<sup>62</sup>, represented by *Maylandia zebra* (n=5),  
212 has closely arranged bicuspid teeth, that is uses for pulling and scraping loose Aufwuchs (periphyton) attached to the rocks  
213 found in their preferred preferred rocky habitats<sup>59,63</sup>. *Tropheops* and *Iodotropheus*, represented by *Tropheops tropheops* (n=2)  
214 and *Iodotropheus sprengerae* (n=2), also have closely packed bicuspid teeth, that they use to feed on epilithic algae which  
215 they pluck with sideways, upwards head jerks, a behaviour likely supported by *Tropheops*’ characteristic steeply sloped vomer  
216 (71-96°)<sup>41</sup>. Members of *Petrotilapia*, represented by *Petrotilapia genalutea* (n=1) have a mixed combination of tricupsid  
217 and unicuspid teeth that they use to comb loose periphyton from rock surfaces<sup>64</sup>. A further represented mbuna genus is the  
218 monotypic *Genyochromis*, represented by *Genyochromis mento* (n=2). Like the majority of mbuna, *G. mento* has prominent  
219 outer bicuspid teeth, and are supported by smaller inner tricupsid teeth<sup>41,59</sup>. In contrast to most other mbuna, however, *G. mento*  
220 is a highly specialised feeder, a lepidophage (scale-eater), that targets the the caudal and anal fins of other cichlids in rocky  
221 habitats<sup>41,59,65</sup>. The preferred striking side of *G. mento* significantly correlates with left-right asymmetry of the dentary, with  
222 right and left-leaning individuals preferring to strike the corresponding side, respectively, of their prey. Interestingly, however,  
223 a comparison of their jaw laterality with *Perissodus microlepis*, a lepidophage endemic to Lake Tanganyika<sup>20</sup>, showed that  
224 laterality in *G. mento* is weaker than in *P. microlepis* – likely a result of phylogenetic constraint from their shorter evolutionary  
225 history and their herbivorous ancestors<sup>65</sup>.

226 The craniofacial bones commonly studied in mbuna, such as the dentary, premaxilla, pharyngeal jaws, as well as their  
227 associated teeth, can be segmented from specimens in the dataset (see *G. mento*, Figure 5). Future sampling should focus  
228 on the seven remaining genera not sampled in our dataset: *Abactochromis*, *Chindongo*, *Cyathochromis*, *Gepyrochromis*,  
229 *Labeotropheus*, *Melanochromis* and *Pseudotropheus*.

### 230 *Astatotilapia calliptera* and Ruaha Catchment

231 *Astatotilapia* is polyphyletic and current members of the genus are widespread across East and North Africa<sup>6,66,67</sup>. Only  
232 one species of *Astatotilapia* is native to Lake Malawi, *Astatotilapia calliptera*, which is also found in East African rivers  
233 flowing eastward to the Indian Ocean, from the Rovuma River in the north, to the Save River in the south. Given the wide  
234 distribution of the species, it is perhaps unsurprising that intraspecific genetic variation within the species is comparable to that  
235 of the whole Lake Malawi radiation<sup>4,6</sup>. Despite their wide distribution and relatively large intraspecific genetic variation, they  
236 phylogenetically cluster within the Lake Malawi radiation (Figure 1), forming a sister clade to the mbuna, with which they  
237 share an excess of alleles<sup>4</sup>. This pattern, alongside a perceived riverine ‘generalist’ lifestyle, has led to the hypothesis that either  
238 Lake Malawi cichlids radiated from an *A. calliptera*-like ancestor or that *A. calliptera* is the sympatric ancestor of all Lake  
239 Malawi cichlids<sup>4,6,41,67</sup>.

240 Given the importance of *A. calliptera* in the Lake Malawi radiation, we sampled multiple individuals from multiple  
241 populations. We were able to scan nine laboratory-reared individuals from the Mbaka river population, which flows into the  
242 northern end of Lake Malawi<sup>68</sup>. We also scanned individuals from Lake Chilwa (an endorheic lake south-east of Lake Malawi<sup>69</sup>;  
243 n=2), Lake ‘Misoko’, presumably Lake Masoko (a crater lake north of Lake Malawi<sup>68</sup>, n=2), and wild-caught individuals from  
244 the main body of Lake Malawi (n=2). Populations of *A. calliptera* differ in life history strategies<sup>70</sup> and are also undergoing  
245 sympatric speciation along a depth gradient in at least one location (Lake Masoko)<sup>8</sup>, where littoral and benthic *A. calliptera*  
246 ecomorphs have diverged in multiple characteristics, including body shape and trophic specialism, in approximately 1000  
247 years<sup>8</sup>. Therefore, it is possible that morphological evaluations of more populations of *A. calliptera* will reveal further diversity,  
248 potentially providing greater insight into the role it has taken in generating the wider Lake Malawi haplochromine radiation.

249 A key part of macroevolutionary studies is the estimation of ancestral state of traits based on the morphology of their  
250 descendants<sup>71</sup>. This relies on a comprehensive understanding of trait diversity across taxa, and such data can also inform

models of morphological evolution, and enable estimates of rates of phenotypic evolution. Since the genetic diversity of the Lake Malawi radiation was possibly seeded by multiple riverine species<sup>5</sup>, we sought to add specimens to the dataset that could enable the morphological reconstruction of the common ancestor of the Lake Malawi radiation. Therefore, we also sampled two additional species of *Astatotilapia*. These included *Astatotilapia gigliolli* (n=2) and *Astatotilapia* sp. ‘Ruaha blue’ (n=2), native to the Great Ruaha River<sup>66,67</sup>. Construction of a mtDNA-based phylogeny initially placed *Astatotilapia* sp. ‘Ruaha blue’ as a sister taxa to the Lake Malawi radiation<sup>66</sup>. However, a phylogeny based on variation within whole-genome sequences has shown *A. gigliolli* and *A. sp. ‘Ruaha blue’*, sister taxa, form a sister clade with both the Lake Malawi and Lake Victoria radiations (see Figure 1) – a topology that is likely the result of an ancestral hybridisation event with the ancestors of both lineages prior to their respective adaptive radiations<sup>5</sup>. Therefore, the addition of species from the Ruaha catchment, may therefore enable a more robust estimation of the ancestral phenotype of Lake Malawi cichlids.

## Deep Benthic and ‘Utaka’

We sampled deep- water benthic species from two genera; *Alticorpus* and *Lethrinops*<sup>9,45</sup> (Table 1). *Alticorpus*, like *Aulonocara* (not represented in the dataset) is characterised by the presence of greatly enlarged cranial sensory openings and lateral line canals used to detect prey in the sediment. Deep-water benthic species are found below 50m, a ‘twilight’ zone with very little visible light. *Alticorpus macrocleithrum* (n=3) is found between 75m and 125m, with abundance peaking above 100m<sup>72</sup>, a depth similarly occupied by deep-water *Lethrinops*<sup>73</sup>, including *Lethrinops gosseii* (n=1). Several species of *Lethrinops*, however, inhabit shallower water<sup>4,41</sup>. We sampled two species of shallow water *Lethrinops*, including *Lethrinops auritus* (n=2), and *Lethrinops albus* (n=2), both of which phylogenetically cluster within the ‘shallow benthic’ lineage (Figure 1).

Our dataset also contains four species of zooplankton-feeding, shoaling cichlids which are commonly referred to as ‘utaka’. Utaka is primarily made up of species belonging to *Copadichromis*<sup>42</sup>, with a small number of species also belonging to *Mchenga* and the older *Nyassachromis*<sup>74</sup>. However, their placement within the utaka is disputed and they have not been considered in our species/genera counts (see Table 1). We sampled four species of *Copadichromis*: *Copadichromis likomae* (n=2), *Copadichromis quadrimaculatus* (n=2), *Copadichromis trimaculatus* (n=2, see Figure 6) and *Copadichromis virginalis* (n=2). Utaka feed in the water column, and can be commonly found close to the shore<sup>41</sup>. *Copadichromis* are generally characterised by their relatively small, highly protrusible mouths, that they use to suck zooplankton into their mouths, as well as numerous long gill rakers which strain plankton from the water that enters their mouths as a result of their sucking feeding mechanism<sup>41,75</sup>.

Both the deep benthics and utaka are currently underrepresented within our dataset and future sampling should aim to add additional species of *Copadichromis*. In addition, sampling missing ‘deep benthic’ genera, such as *Aulonocara* and *Tramitichromis* should be prioritised for future sampling efforts. *Aulonocara stuartgranti* and *Aulonocara steveni* would be particularly interesting future additions and could offer interesting morphological comparisons with deeper living species. Moreover, given that *Lethrinops* is polyphyletic<sup>73,76</sup>, additional sampling of *Lethrinops* species could provide morphological data to support future systematic studies.

## Technical Validation and Usage Notes

As mentioned above, entries for each specimen can be found in Supplementary Table 1 and cropped, reconstructed 8-bit tiff images stacks of all specimens scanned can be accessed and downloaded from a dedicated [project](#) on Morphosource.

To validate our  $\mu$ CT-scan data, we imported the reconstructed image stacks into *Avizo Lite* (v9.3.0), a proprietary software developed by Thermo Fisher Scientific and either generated a full body volume render or rendered 3D models of the whole body. Surfaces from 3D whole-body renderings were exported from AvizoLite as Polygon File Format (.ply) files and imported into *MeshLab* for visualisation and manipulation. We note that there are free, open access alternatives to *Avizo Lite* (v9.3.0) for the segmentation of 3D-image data, such as *3D-Slicer*, which has a large and active community of users<sup>77</sup> and *Dragonfly* which supports the use of deep learning to automatically segment 3D image data and offers non-commercial licenses for academic use free-of-charge. Example whole-body 3D renderings for representatives from each ecomorphological group can be found in Figure 4 and for each specimen in the dataset in the Supplementary Material.

In addition, we manually segmented multiple bones, including the dentary, premaxilla, lower pharyngeal jaw and multiple vertebral types in several species discussed above. This included: the ‘generalist’ *Astatotilapia calliptera*; the ‘mbuna’ and lepidophage, *Genyochromis mento*; the ‘shallow benthic’ and snail crusher, *Trematocranus placodon* (Figure 5); the pelagic piscivores, *Rhamphochromis esox* and *Pallidochromis tokolosh* and the zooplanktivorous (utaka), *Copadichromis trimaculatus* (Figure 6). Lower pharyngeal jaws that are highly variable among species of Lake Malawi cichlids<sup>25</sup> were particularly well resolved. For example, newly erupting teeth were visible on the relatively large, and dense, lower pharyngeal jaw of *Trematocranus placodon* (Figure 5B). Similarly, renderings of multiple vertebral types (Figures 5–6) were also good quality. The zygapophyses and fine structure of the vertebral centra, sometimes including the neural canal, were also well resolved. All 3D-renderings of these bones can be found in the Supplementary Materials as downloadable .ply files.

303 The computer specifications we used for the all analyses (see Methodology) are hard to find on personal, or older machines  
304 and some users may find it difficult to work with some of our larger image stacks. To minimise memory usage during  
305 segmentation and speed up processing, cropped reconstructed stacks can be loaded in multiple increments (note that the Z-voxel  
306 size must be multiplied by said increment). We tested this and found that roughly comparable models could be generated,  
307 although it was clear that finer morphological detail was absent (data not shown). Therefore, where possible, the whole stack  
308 should be used when segmenting regions of interest. In addition, since these regions were manually segmented, many of the  
309 segmentation steps rely on the judgment of the individual segmenting and rendering of the regions of interest. We found that  
310 segmenting from median-filtered reconstructed image stacks drastically lowered the quality of the rendered models (data not  
311 shown) and would suggest refraining from segmenting from a median-filtered image stack. In addition, we found that relatively  
312 low smoothing factors were best for rendering surfaces from segmented regions of interest. In *Avizo Lite (v9.3.0)*, a smoothing  
313 factor between 0-10 (including rational intermediates) can be applied when rendering surfaces of segmented regions of interest.  
314 We rarely found it necessary to use a value above '3'; indeed, all whole-body 3D models were smoothed with a factor of 2.5,  
315 and we suggest, regardless of the tool used to smooth and render segmented surfaces, to use smoothing cautiously. Furthermore,  
316 we were able to scan multiple species of *Copadichromis* (utaka), see above and were able to generate relatively good quality  
317 models (see Figure 6 and the Supplementary Models). However, it is clear that some of the jaw structures did not resolve as  
318 well as in other specimens. It is possible that the jaw bones of these fish species are not particularly dense, which made it  
319 difficult to image them using the same scanning procedure as that was used for all of the other specimens. Therefore, in future,  
320 we would recommend care when segmenting bones from the *Copadichromis* species present in this dataset and also to increase  
321 the power and exposure time for future sampling of *Copadichromis*.

322 Here we have presented the first, comprehensive, freely available, whole-body  $\mu$ CT-scan dataset of Lake Malawi cichlids.  
323 We have described several macroevolutionary studies that could be conducted with this dataset to better understand this  
324 remarkable cichlid adaptive radiation and include suggestions for future sampling. Our Lake Malawi dataset now joins two  
325 other East African adaptive radiation datasets, the recent haplochromine Lake Victoria library<sup>78</sup> and the extensive  $\mu$ CT-scan  
326 dataset of Lake Tanganyika cichlid fishes<sup>7</sup>. Therefore, the addition of our dataset now offers a unique opportunity for much  
327 wider, and systematic, morphological comparisons within East African cichlids. We have also described a methodology to  
328 efficiently  $\mu$ CT-scan multiple specimens simultaneously; reducing scanning time and financial cost, whilst maintaining scan  
329 quality, demonstrating the utility of this method by reconstructing 3D-models of multiple bones from multiple specimens  
330 within our dataset. We hope the availability of these data will inspire people to address some of the many questions left still to  
331 understand this remarkable adaptive radiation, permit wider-scale comparisons with other cichlid adaptive radiations and set a  
332 precedent to make whole-body  $\mu$ CT scans the automatic default for any sampling efforts involving cichlids.

## 333 References

- 334 1. Turner, G. F., Seehausen, O., Knight, M. E., Allender, C. J. & Robinson, R. L. How many species of cichlid fishes are there  
335 in african lakes? *Mol. Ecol.* **10**, 793–806, <https://doi.org/10.1046/j.1365-294x.2001.01200.x> (2001).
- 336 2. Barlow, G. *The Cichlid Fishes: Nature's Grand Experiment In Evolution* (Hachette UK, London, 2008).
- 337 3. Sparks, J. S. & Smith, W. L. Phylogeny and biogeography of cichlid fishes (teleostei: Perciformes: Cichlidae). *Cladistics*  
338 **20**, 501–517, <https://doi.org/10.1111/j.1096-0031.2004.00038.x> (2004).
- 339 4. Malinsky, M. *et al.* Whole-genome sequences of malawi cichlids reveal multiple radiations interconnected by gene flow.  
340 *Nat. Ecol. Evol.* **2**, 1940–1955, <https://doi.org/10.1038/s41559-018-0717-x> (2018).
- 341 5. Svardal, H. *et al.* Ancestral hybridization facilitated species diversification in the lake malawi cichlid fish adaptive radiation.  
342 *Mol. biology evolution* **37**, 1100–1113, <https://doi.org/10.1093/molbev/msz294> (2020).
- 343 6. Svardal, H., Salzburger, W. & Malinsky, M. Genetic variation and hybridization in evolutionary radiations of cichlid fishes.  
344 *Annu. Rev. Animal Biosci.* **9**, 55–79, <https://doi.org/10.1146/annurev-animal-061220-023129> (2021).
- 345 7. Ronco, F. *et al.* Drivers and dynamics of a massive adaptive radiation in cichlid fishes. *Nature* **589**, 76–81, <https://doi.org/10.1038/s41586-020-2930-4> (2021).
- 347 8. Genner, M. J. *et al.* Evolution of a cichlid fish in a lake malawi satellite lake. *Proc. Royal Soc. B: Biol. Sci.* **274**, 2249–2257,  
348 <https://doi.org/10.1098/rspb.2007.0619> (2007).
- 349 9. Genner, M. J. & Turner, G. F. Ancient hybridization and phenotypic novelty within lake malawi's cichlid fish radiation.  
350 *Mol. Biol. Evol.* **29**, 195–206, <https://doi.org/10.1093/molbev/msr183> (2012).
- 351 10. Meier, J. I. *et al.* Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nat. communications* **8**, 14363,  
352 <https://doi.org/10.1038/ncomms14363> (2017).

- 353 **11.** Keller-Costa, T., Canário, A. V. & Hubbard, P. C. Chemical communication in cichlids: a mini-review. *Gen. comparative*  
354 *endocrinology* **221**, 64–74, <https://doi.org/10.1016/j.ygcen.2015.01.001> (2015).
- 355 **12.** Faber-Hammond, J. J. & Renn, S. C. Transcriptomic changes associated with maternal care in the brain of mouthbrooding  
356 cichlid *astatotilapia burtoni* reflect adaptation to self-induced metabolic stress. *J. Exp. Biol.* **226**, jeb244734, <https://doi.org/10.1242/jeb.244734> (2023).
- 357  
358 **13.** Plenderleith, M., Oosterhout, C. v., Robinson, R. L. & Turner, G. F. Female preference for conspecific males based on  
359 olfactory cues in a lake malawi cichlid fish. *Biol. Lett.* **1**, 411–414, <https://doi.org/10.1098/rsbl.2005.0355> (2005).
- 360 **14.** Morita, M. *et al.* Bower-building behaviour is associated with increased sperm longevity in tanganyikan cichlids. *J.*  
361 *evolutionary biology* **27**, 2629–2643, <https://doi.org/10.1111/jeb.12522> (2014).
- 362 **15.** McKaye, K. R. & Kocher, T. Head ramming behaviour by three paedophagous cichlids in lake malawi, africa. *Animal*  
363 *Behav.* **31**, 206–210, [https://doi.org/10.1016/S0003-3472\(83\)80190-0](https://doi.org/10.1016/S0003-3472(83)80190-0) (1983).
- 364 **16.** Woltering, J. M., Holzem, M., Schneider, R. F., Nanos, V. & Meyer, A. The skeletal ontogeny of *astatotilapia burtoni*—a  
365 direct-developing model system for the evolution and development of the teleost body plan. *BMC developmental biology*  
366 **18**, 1–23, <https://doi.org/10.1186/s12861-018-0166-4> (2018).
- 367 **17.** Santos, M. E., Lopes, J. F. & Kratochwil, C. F. East african cichlid fishes. *EvoDevo* **14**, 1, <https://doi.org/10.1186/s13227-022-00205-5> (2023).
- 368  
369 **18.** Marconi, A., Yang, C. Z., McKay, S. & Santos, M. E. Morphological and temporal variation in early embryogenesis  
370 contributes to species divergence in malawi cichlid fishes. *Evol. & Dev.* **25**, 170–193, <https://doi.org/10.1111/ede.12429>  
371 (2023).
- 372 **19.** Navon, D., Olearczyk, N. & Albertson, R. C. Genetic and developmental basis for fin shape variation in african cichlid  
373 fishes. *Mol. Ecol.* **26**, 291–303, <https://doi.org/10.1111/mec.13905> (2017).
- 374 **20.** Ronco, F., Büscher, H. H., Indermaur, A. & Salzburger, W. The taxonomic diversity of the cichlid fish fauna of ancient  
375 lake tanganyika, east africa. *J. Gt. Lakes Res.* **46**, 1067–1078, <https://doi.org/10.1016/j.jglr.2019.05.009> (2020).
- 376 **21.** Arthur, W. The emerging conceptual framework of evolutionary developmental biology. *Nature* **415**, 757–764, <https://doi.org/10.1038/415757a> (2002).
- 377  
378 **22.** Albertson, R. C. & Kocher, T. D. Assessing morphological differences in an adaptive trait: a landmark-based morphometric  
379 approach. *The J. Exp. Zool.* **289**, 385–403, <https://doi.org/10.1002/jez.1020> (2001).
- 380 **23.** Adams, D., Yamaoka, K. & Kassam, D. Functional significance of variation in trophic morphology within feed-  
381 ing microhabitat-differentiated cichlid species in lake malawi. *Animal Biol.* **54**, 77–90, <https://doi.org/10.1163/157075604323010060> (2004).
- 382  
383 **24.** Hulsey, C. D., Alfaro, M. E., Zheng, J., Meyer, A. & Holzman, R. Pleiotropic jaw morphology links the evolution of  
384 mechanical modularity and functional feeding convergence in lake malawi cichlids. *Proc. Royal Soc. B: Biol. Sci.* **286**,  
385 20182358, <https://doi.org/10.1098/rspb.2018.2358> (2019).
- 386 **25.** Conith, A. J. & Albertson, R. C. The cichlid oral and pharyngeal jaws are evolutionarily and genetically coupled. *Nat.*  
387 *Commun.* **12**, 5477, <https://doi.org/10.1038/s41467-021-25755-5> (2021).
- 388 **26.** Kratochwil, C. F. *et al.* Agouti-related peptide 2 facilitates convergent evolution of stripe patterns across cichlid fish  
389 radiations. *Science* **362**, 457–460, <https://doi.org/10.1126/science.aao6809> (2018).
- 390 **27.** Gerwin, J., Urban, S., Meyer, A. & Kratochwil, C. F. Of bars and stripes: A malawi cichlid hybrid cross provides insights  
391 into genetic modularity and evolution of modifier loci underlying colour pattern diversification. *Mol. Ecol.* **30**, 4789–4803,  
392 <https://doi.org/10.1111/mec.16097> (2021).
- 393 **28.** Clark, B. *et al.* Oca2 targeting using crispr/cas9 in the malawi cichlid *astatotilapia calliptera*. *Royal Soc. Open Sci.* **9**,  
394 220077, <https://doi.org/10.1098/rsos.220077> (2022).
- 395 **29.** DeLorenzo, L. *et al.* Genetic basis of ecologically relevant body shape variation among four genera of cichlid fishes. *Mol.*  
396 *ecology* **32**, 3975–3988, <https://doi.org/10.1111/mec.16977> (2023).
- 397 **30.** Darrin Hulsey, C., Keck, B. P., Alamillo, H. & O’Meara, B. C. Mitochondrial genome primers for lake malawi cichlids.  
398 *Mol. Ecol. Resour.* **13**, 347–353, <https://doi.org/10.1111/1755-0998.12066> (2013).
- 399 **31.** McGee, M. D. *et al.* The ecological and genomic basis of explosive adaptive radiation. *Nature* **586**, 75–79, <https://doi.org/10.1038/s41586-020-2652-7> (2020).
- 400



- 401 **32.** Masonick, P., Meyer, A. & Hulsey, C. D. Phylogenomic analyses show repeated evolution of hypertrophied lips among  
402 lake malawi cichlid fishes. *Genome Biol. Evol.* **14**, evac051, <https://doi.org/10.1093/gbe/evac051> (2022).
- 403 **33.** Price, S. A., Friedman, S. T. & Wainwright, P. C. How predation shaped fish: the impact of fin spines on body form  
404 evolution across teleosts. *Proc. Royal Soc. B: Biol. Sci.* **282**, 20151428, <https://doi.org/10.1098/rspb.2015.1428> (2015).
- 405 **34.** Baxter, D., Cohen, K. E., Donatelli, C. M. & Tytell, E. D. Internal vertebral morphology of bony fishes matches the  
406 mechanical demands of different environments. *Ecol. Evol.* **12**, e9499, <https://doi.org/10.1002/ece3.9499> (2022).
- 407 **35.** Todd Streebman, J. & Danley, P. D. The stages of vertebrate evolutionary radiation. *Trends Ecol. & Evol.* **18**, 126–131,  
408 [https://doi.org/10.1016/S0169-5347\(02\)00036-8](https://doi.org/10.1016/S0169-5347(02)00036-8) (2003).
- 409 **36.** Gavrillets, S. & Losos, J. B. Adaptive radiation: Contrasting theory with data. *Science* **323**, 732–737, <https://doi.org/10.1126/science.1157966> (2009).
- 411 **37.** Schindelin, J. *et al.* Fiji: an open-source platform for biological-image analysis. *Nat. methods* **9**, 676–682, <https://doi.org/10.1038/nmeth.2019> (2012).
- 413 **38.** Schneider, C. A., Rasband, W. S. & Eliceiri, K. W. Nih image to imagej: 25 years of image analysis. *Nat. methods* **9**,  
414 671–675, <https://doi.org/10.1038/nmeth.2089> (2012).
- 415 **39.** Turner, G., Robinson, R., Shaw, P., Carvalho, G. & Snoeks, J. *Identification and biology of Diplotaxodon, Rhamphochromis*  
416 *and Pallidochromis* (Cichlid Press, 2004).
- 417 **40.** Hahn, C., Genner, M. J., Turner, G. F. & Joyce, D. A. The genomic basis of cichlid fish adaptation within the deepwater  
418 “twilight zone” of lake malawi. *Evol. Lett.* **1**, 184–198, <https://doi.org/10.1002/evl3.20> (2017).
- 419 **41.** Konings, A. *Malawi Cichlids in their Natural Habitat* (Cichlid Press, 2016), 5th edition edn.
- 420 **42.** Anseeuw, D., Nevado, B., Busselen, P., Snoeks, J. & Verheyen, E. Extensive introgression among ancestral mtDNA  
421 lineages: Phylogenetic relationships of the utaka within the lake malawi cichlid flock. *Int. J. Evol. Biol.* **2012**, 1–9,  
422 <https://doi.org/10.1155/2012/865603> (2012).
- 423 **43.** Genner, M. J. & Turner, G. F. The mbuna cichlids of lake malawi: a model for rapid speciation and adaptive radiation.  
424 *Fish fisheries* **6**, 1–34, <https://doi.org/10.1111/j.1467-2679.2005.00173.x> (2005).
- 425 **44.** Nichols, P. *et al.* Secondary contact seeds phenotypic novelty in cichlid fishes. *Proc. Royal Soc. B: Biol. Sci.* **282**, 20142272,  
426 <https://doi.org/10.1098/rspb.2014.2272> (2015).
- 427 **45.** Joyce, D. A. *et al.* Repeated colonization and hybridization in lake malawi cichlids. *Curr. Biol.* **21**, R108–R109,  
428 <https://doi.org/10.1016/j.cub.2010.11.029> (2011).
- 429 **46.** Turner, G., Ngatunga, B. P. & Genner, M. J. The natural history of the satellite lakes of lake malawi. Preprint at  
430 <https://doi.org/10.32942/osf.io/sehdq> (2019).
- 431 **47.** Turner, G. F. Description of a commercially important pelagic species of the genus diplotaxodon (pisces: Cichlidae) from  
432 lake malawi, africa. *J. Fish Biol.* **44**, 799–807, <https://doi.org/10.1111/j.1095-8649.1994.tb01256.x> (1994).
- 433 **48.** Lowe-McConnell, R. Recent research in the african great lakes: fisheries, biodiversity and cichlid evolution. Accessible  
434 here =<https://aquadocs.org/handle/1834/22270> (2003).
- 435 **49.** Wilson, G. D. & Hessler, R. R. Speciation in the deep sea. *Annu. Rev. Ecol. Syst.* **18**, 185–207, <https://doi.org/10.1146/annurev.es.18.110187.001153> (1987).
- 437 **50.** Jennings, R. M., Etter, R. J. & Ficarra, L. Population differentiation and species formation in the deep sea: the potential  
438 role of environmental gradients and depth. *PLoS One* **8**, e77594, <https://doi.org/10.1371/journal.pone.0077594> (2013).
- 439 **51.** Neat, F. & Campbell, N. Proliferation of elongate fishes in the deep sea. *J. Fish Biol.* **83**, 1576–1591, <https://doi.org/10.1111/jfb.12266> (2013).
- 441 **52.** Stiassny, M. Phylogenetic versus convergent relationships between piscivorous cichlid fishes from lakes malawi and  
442 tanganyika. *Bull. Br. Mus. (Natural Hist. Zool.* **40**, 67 – 101 (1981).
- 443 **53.** Donatelli, C. M. *et al.* Foretelling the flex—vertebral shape predicts behavior and ecology of fishes. *Integr. Comp. Biol.* **61**,  
444 414–426, <https://doi.org/10.1093/icb/icab110> (2021).
- 445 **54.** Fujimura, K. & Okada, N. Shaping of the lower jaw bone during growth of nile tilapia oreochromis niloticus and a  
446 lake victoria cichlid haplochromis chilotes: A geometric morphometric approach. *Dev. Growth & Differ.* **50**, 653–663,  
447 <https://doi.org/10.1111/j.1440-169X.2008.01063.x> (2008).

- 448 **55.** Gatumu, E. M. Redescription of the genera nimbochromis and tyrannochromis (teleostei: Cichlidae) from lake malawi,  
449 africa. [https://solo.bodleian.ox.ac.uk/permalink/44OXF\\_INST/ao2p7t/cdi\\_proquest\\_journals\\_305235338](https://solo.bodleian.ox.ac.uk/permalink/44OXF_INST/ao2p7t/cdi_proquest_journals_305235338) (2003).
- 450 **56.** Oliver, M. K. Six new species of the cichlid genus otopharynx from lake malawi (teleostei: Cichlidae). *Bull. Peabody Mus.*  
451 *Nat. Hist.* **59**, 159–197, <https://doi.org/10.3374/014.059.0204> (2018).
- 452 **57.** Evers, B. N., Madsen, H., McKaye, K. M. & Stauffer, J. R. The schistosome intermediate host, bulinus nyassanus, is  
453 a 'preferred' food for the cichlid fish, trematocranus placodon, at cape maclear, lake malawi. *Annals Trop. Medicine &*  
454 *Parasitol.* **100**, 75–85, <https://doi.org/10.1179/136485906X78553> (2006).
- 455 **58.** Albertson, R. C. Morphological divergence predicts habitat partitioning in a lake malawi cichlid species complex. *Copeia*  
456 **2008**, 689–698, <https://doi.org/10.1643/CG-07-217> (2008).
- 457 **59.** Ribbink, A., Marsh, B., Marsh, A., Ribbink, A. & Sharp, B. A preliminary survey of the cichlid fishes of rocky habitats in  
458 lake malawi. *South Afr. J. Zool.* **18**, 149–310, <https://doi.org/10.1080/02541858.1983.11447831> (1983).
- 459 **60.** Kassam, D., Seki, S., Rusuwa, B., Ambali, A. J. & Yamaoka, K. Genetic diversity within the genus cynotilapia and its  
460 phylogenetic position among lake malawi's mbuna cichlids. *Afr. J. Biotechnol.* **4**, <https://doi.org/10.4314/ajb.v4i10.71319>  
461 (2005).
- 462 **61.** Genner, M., Turner, G., Barker, S. & Hawkins, S. Niche segregation among lake malawi cichlid fishes? evidence from  
463 stable isotope signatures. *Ecol. Lett.* **2**, 185–190, <https://doi.org/10.1046/j.1461-0248.1999.00068.x> (1999).
- 464 **62.** Stauffer Jr, J. R., Bowers, N. J., Kellogg, K. A. & McKaye, K. R. A revision of the blue-black pseudotropheus zebra  
465 (teleostei: Cichlidae) complex from lake malawi, africa, with a description of a new genus and ten new species. *Proc. Acad.*  
466 *Nat. Sci. Phila.* 189–230, <http://www.jstor.org/stable/4065053> (1997).
- 467 **63.** Holzberg, S. A field and laboratory study of the behaviour and ecology of pseudotropheus zebra (boulenger), an endemic  
468 cichlid of lake malawi (pisces; cichlidae). *J. Zool. Syst. Evol. Res.* **16**, 171–187, <https://doi.org/10.1111/j.1439-0469.1978.tb00929.x>  
469 (1978).
- 470 **64.** Marsh, A. A taxonomic study of the fish genus petrotilapia (pisces: Cichlidae) from lake malawi. *Ichthyol. Bull. J.L.B.*  
471 *Smith Inst. Ichthyol.* **48**, 1–14 (1983).
- 472 **65.** Takeuchi, Y. *et al.* Specialized movement and laterality of fin-biting behaviour in genyochromis mento in lake malawi. *J.*  
473 *Exp. Biol.* **222**, jeb191676, <https://doi.org/10.1242/jeb.191676> (2019).
- 474 **66.** Genner, M. J., Ngatunga, B. P., Mzighani, S., Smith, A. & Turner, G. F. Geographical ancestry of lake malawi's cichlid fish  
475 diversity. *Biol. Lett.* **11**, 20150232, <https://doi.org/10.1098/rsbl.2015.0232> (2015).
- 476 **67.** Turner, G., Ngatunga, B. P. & Genner, M. J. Astatotilapia species (teleostei, cichlidae) from malawi, mozambique and  
477 tanzania, excluding the basin of lake victoria. Preprint at <https://doi.org/10.32942/osf.io/eu6rx> (2021).
- 478 **68.** Malinsky, M. *et al.* Genomic islands of speciation separate cichlid ecomorphs in an east african crater lake. *Science* **350**,  
479 1493–1498, <https://doi.org/10.1126/science.aac9927> (2015).
- 480 **69.** Njaya, F. *et al.* The natural history and fisheries ecology of lake chilwa, southern malawi. *J. Gt. Lakes Res.* **37**, 15–25,  
481 <https://doi.org/10.1016/j.jglr.2010.09.008> (2011).
- 482 **70.** Parsons, P. J., Bridle, J. R., Rüber, L. & Genner, M. J. Evolutionary divergence in life history traits among populations  
483 of the lake malawi cichlid fish astatotilapia calliptera. *Ecol. evolution* **7**, 8488–8506, <https://doi.org/10.1002/ece3.3311>  
484 (2017).
- 485 **71.** Omland, K. E. The assumptions and challenges of ancestral state reconstructions. *Syst. biology* **48**, 604–611, <https://doi.org/10.1080/106351599260175>  
486 (1999).
- 487 **72.** Duponchelle, F., Ribbink, A., Msukwa, A., Mafuka, J. & Mandere, D. Depth distribution and breeding patterns of  
488 the demersal species most commonly caught by trawling in the south west arm of lake malawi. Available at [https://malawicichlids.com/duponchelle\\_ch2.pdf](https://malawicichlids.com/duponchelle_ch2.pdf) (2000).
- 489 **73.** Turner, G. F. A new species of deep-water lethrinops (cichlidae) from lake malawi. *J. Fish Biol.* **101**, 1405–1410,  
490 <https://doi.org/10.1111/jfb.15208> (2022).
- 491 **74.** Stauffer, J. & Konings, A. Review of copadichromis (teleostei: Cichlidae) with the description of a new genus and six new  
492 species. *Ichthyol. Explor. Freshwaters* **17**, 9–42 (2006).
- 493 **75.** Turner, G. F., Crampton, D. A., Rusuwa, B., Hooft van Huysduynen, A. & Svardal, H. Taxonomic investigation of  
494 the zooplanktivorous lake malawi cichlids copadichromis mloto (iles) and c. virginalis (iles). *Hydrobiologia* 1–11,  
495 <https://doi.org/10.1007/s10750-022-05025-1> (2022).
- 496

- 497 **76.** Turner, G. F., Crampton, D. A. & Genner, M. J. A new species of lethinops (cichliformes: Cichlidae) from a lake malawi  
 498 satellite lake, believed to be extinct in the wild. Preprint at <https://doi.org/10.1101/2023.03.17.533142> (2023).
- 499 **77.** Kikinis, R., Pieper, S. D. & Vosburgh, K. G. *3D Slicer: A Platform for Subject-Specific Image Analysis, Visualization, and*  
 500 *Clinical Support* (Springer New York, New York, NY, 2014).
- 501 **78.** Haberthür, D. *et al.* Microtomographic investigation of a large corpus of cichlids. *Plos one* **18**, e0291003, <https://doi.org/10.1371/journal.pone.0291003> (2023).

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## 508 Author contributions statement

509 C.V.B. and B.V. conceived the study. C.V.B and R.B. designed the data acquisition pipeline and dataset curation methodology.  
 510 C.V.B. and R.B. conducted the imaging. C.V.B. performed the sample collection, processed all the raw image data, and wrote  
 511 the manuscript. B.V., R.B. and M.J.G. edited the manuscript. J.M. organised specimens at the NHMUK. G.F.T. and M.J.G.  
 512 contributed specimens to be imaged. All authors reviewed the manuscript.

## 513 Competing interests

514 The authors declare no competing interests.

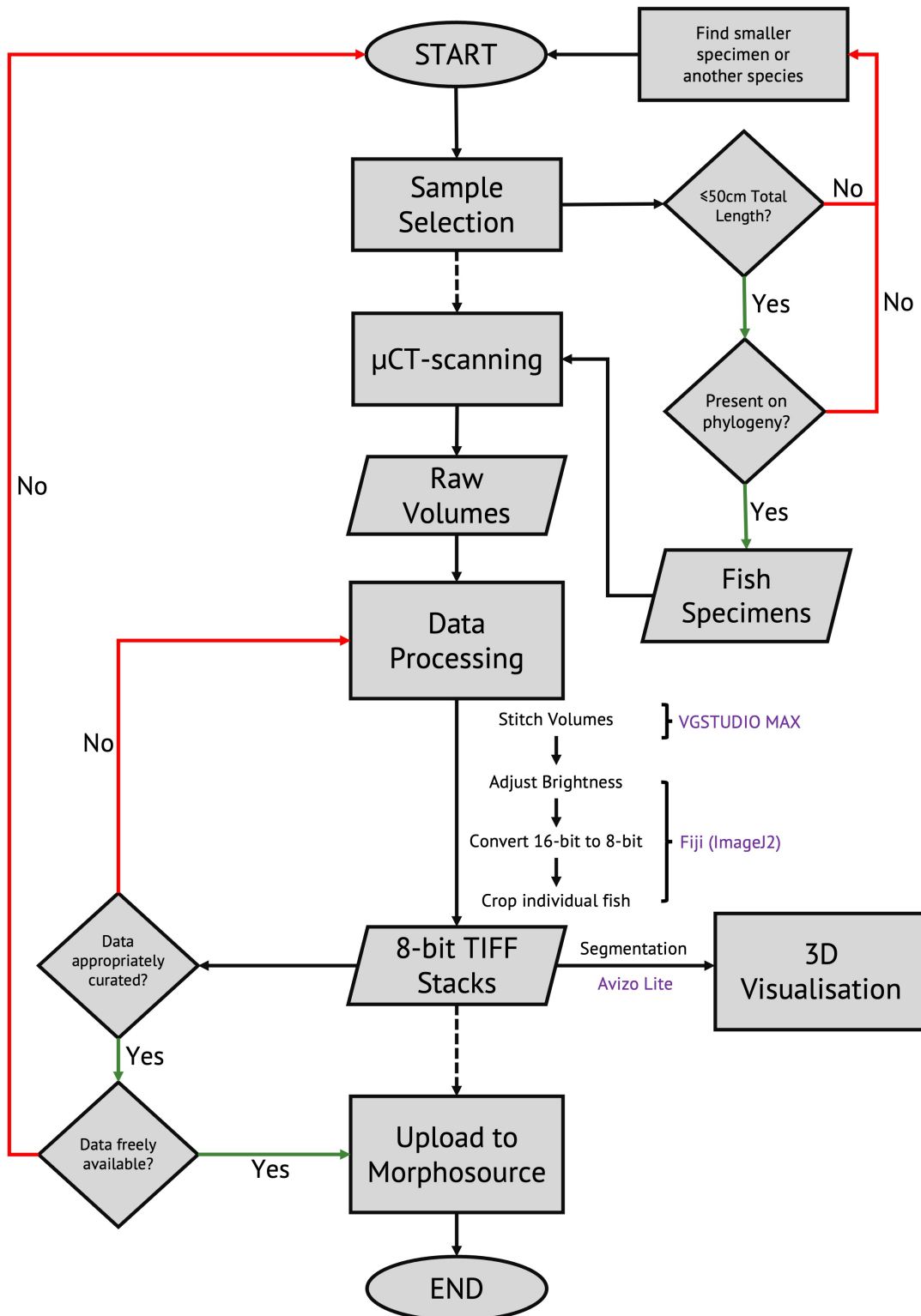
## 515 Figures & Tables

<b>Ecomorphology</b>	<b>Genera Sampled</b>	<b>Genera Sampled (%)</b>	<b>Species Sampled</b>	<b>Species Sampled (%)</b>
<i>Astatotilapia (calliptera)</i>	1 of 1	100%	3*	N/A
Deep Benthic	2 of 5	40%	3 (5 <sup>†</sup> ) of 150	2.00% (3.33%)
<i>Diplotaxodon</i>	2 of 2 <sup>‡</sup>	100%	9 of 19	47.37%
Mbuna	7 of 12	58.33%	7 of 328	2.13%
<i>Rhamphochromis</i>	1 of 1	100%	10 of 14	71.43%
Shallow Benthic	12 of 32	37.50%	20 of 287	6.97%
Utaka	1 of 1	100%	4 <sup>§</sup> of 55	7.27%
<b>Total</b>	<b>26 of 54</b>	<b>48.15%</b>	<b>56 of 856</b>	<b>6.54%</b>

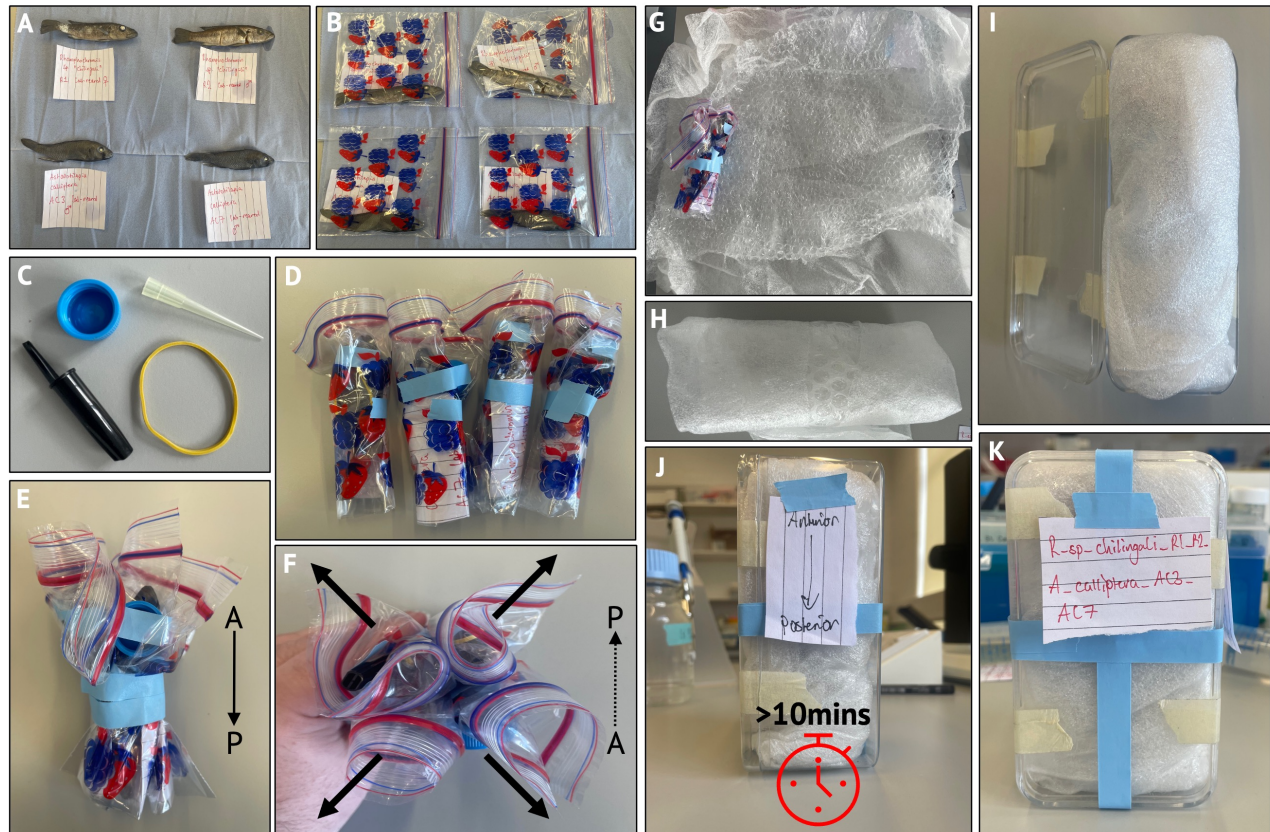
**Table 1. Genera and species represented within the dataset.** The number of genera and species for each ecomorphological group are the same as those used in Malinsky et al., 2018<sup>4</sup>. \**Astatotilapia calliptera*, *Astatotilapia* sp. ‘Ruaha blue’ and *Astatotilapia gigliolli*. <sup>†</sup>If considering the addition of *Lethrinops albus* and *Lethrinops auritus* that cluster within the ‘Shallow Benthics’. <sup>‡</sup>Includes *Pallidochromis*. <sup>§</sup>Includes *Copadichromis trimaculatus* which clusters within the shallow benthics in the phylogeny depicted in Figure 1.



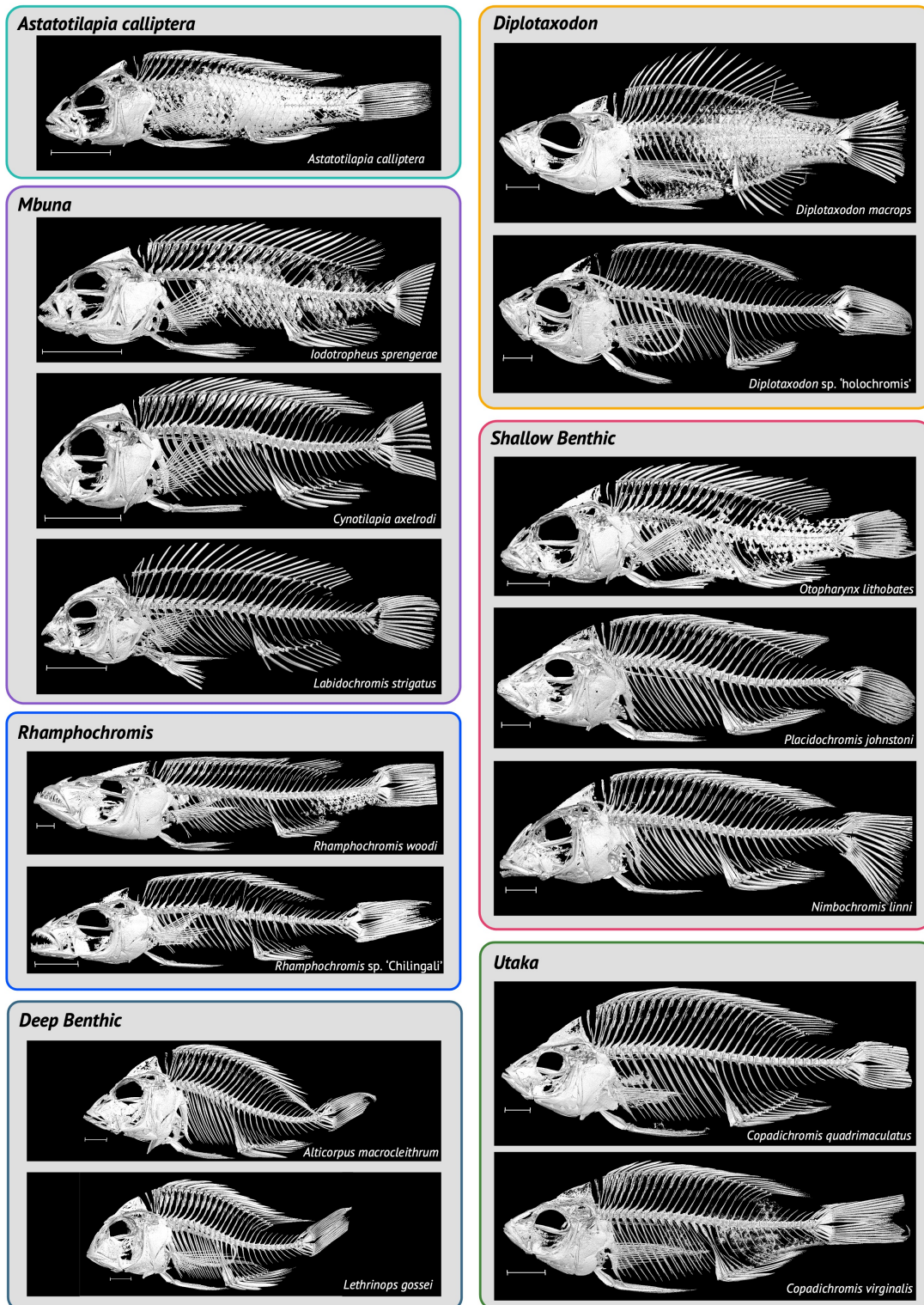




**Figure 2. Flowchart of  $\mu$ CT-scanning, image processing and segmentation methodology.** The flowchart outlines the necessary decisions that were made during collation of the described  $\mu$ -CT scan dataset. Rectangles represent processes; parallelograms represent inputs or outputs; diamonds represent decisions. It is sufficiently generalised that it can be reused for future data collection. We were focused on generating data for a specific macroevolutionary study, so we restricted the dataset to species with known phylogenetic placements. Software associated with data processing steps are indicated in purple.

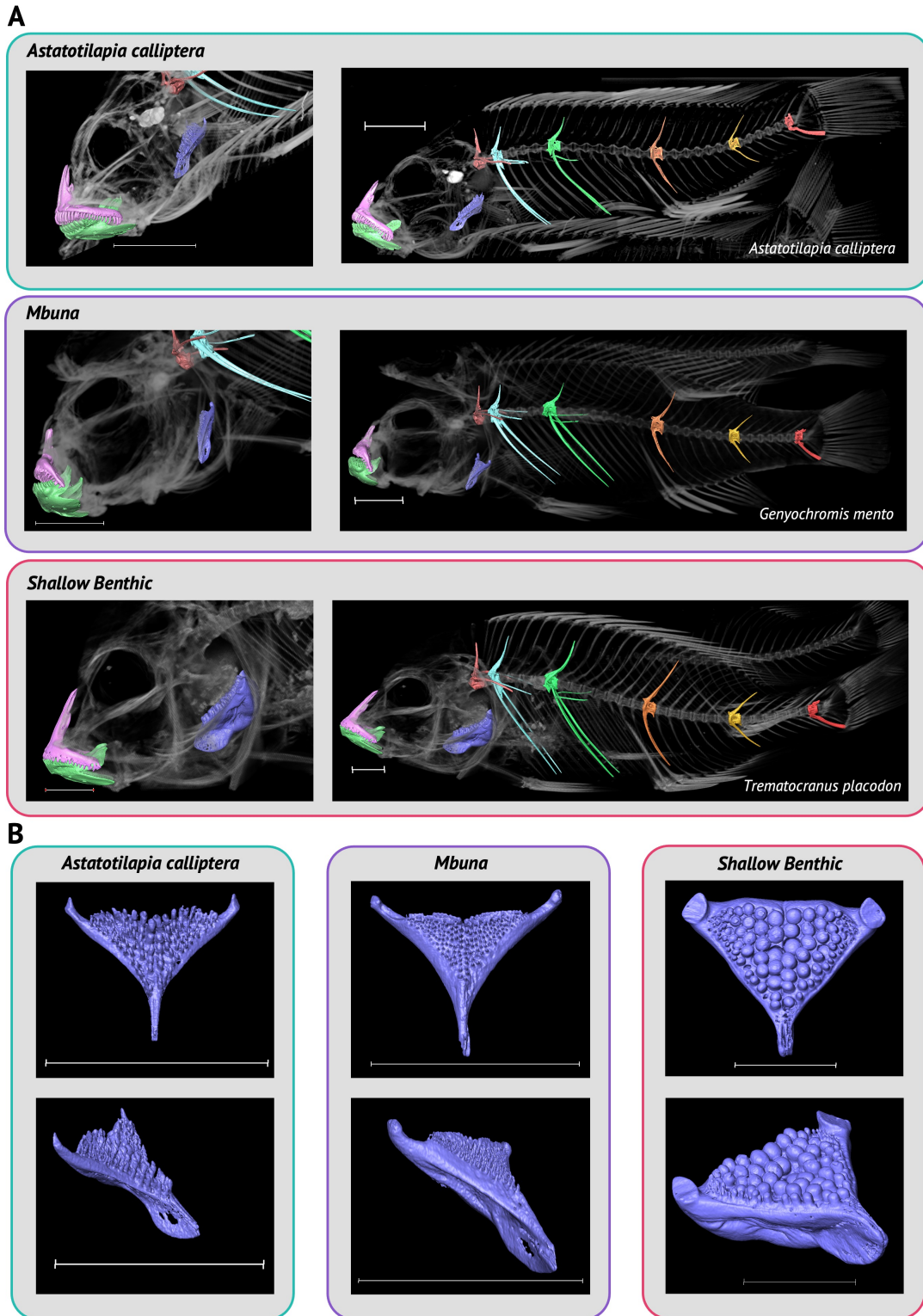


**Figure 3. Specimen preparation for  $\mu$ CT-scanning.** Multiple fish were scanned at the same time. (A-B) Individual fish were labelled and placed in separate plastic bags (so they could be correctly identified and correctly stored after imaging). Unique objects (C) that would be readily identifiable were attached to the outside of these bags, ideally close to the heads, positioned outwards (F, arrows), and bundled together with tape (D-F) all with the same orientation (head-up). Bundles were then wrapped in bubble wrap and other packaging material and tightly sealed inside a plastic container, again head-up. Containers were left for at least ten minutes to settle to prevent movement during scanning.



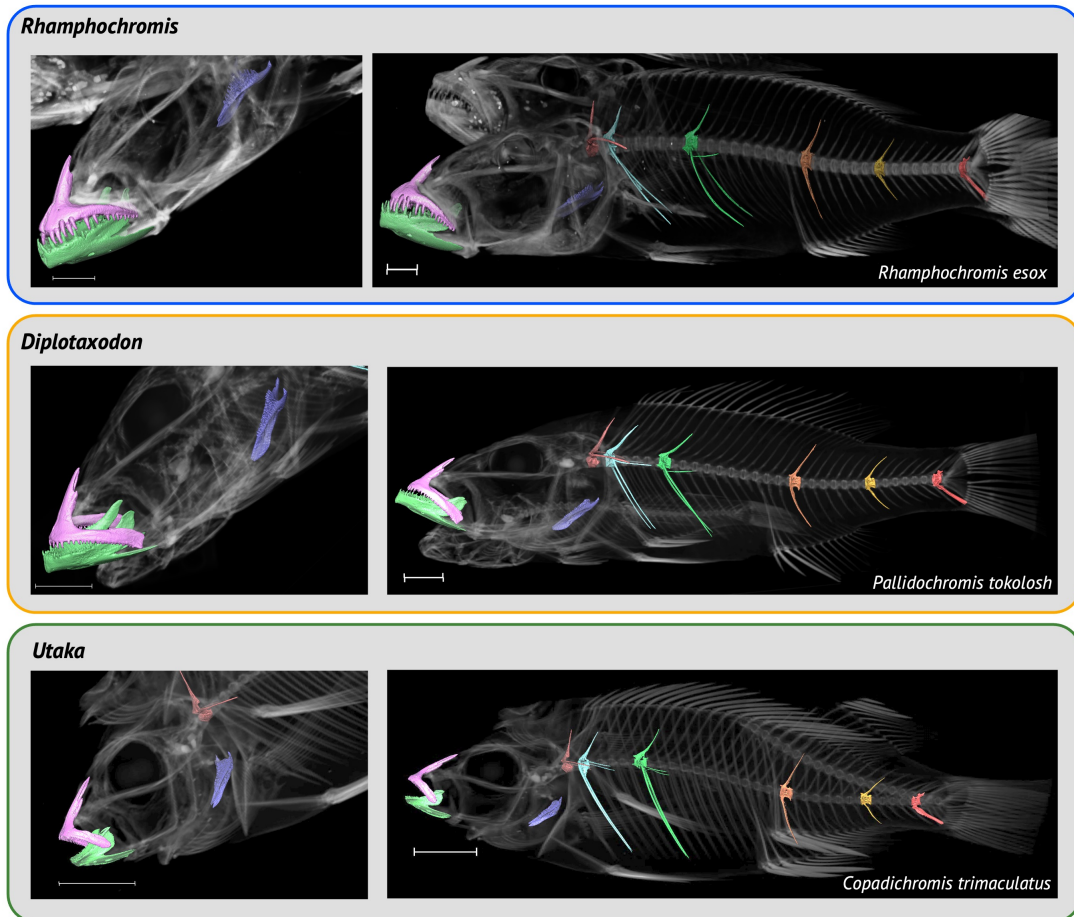
**Figure 4. Whole-body 3D models of select specimens from the dataset.** Scale is 1cm. Specimens are arranged according to the ecomorphological group they belong to. Species names are indicated. Specimens used are indicated in Supplementary Table 1. Some of the specimens have been rendered with their scales, mainly as we found that several specimens had particularly thick scales making it difficult to render whole-body models without including the scales. It is worth noting, however, that the lateral lines have resolved quite well in the models with scales present and with a sufficiently high threshold value scales could resolve well. The ring structure in *Diplotaxodon* sp. 'holochromis' is a rubber band used for identification purposes.





**Figure 5. Segmented Bones from *Astatotilapia calliptera*, *Genyochromis mento* (mbuna) and *Trematocranus placodon* (shallow benthic).** (A, left) A close up, lateral view of the head of each species (species name indicated on right), showing the dentary (green), premaxilla (pink) and lower pharyngeal jaw (purple) positioned within the a volume render of the head. (A, right) A whole body lateral view showing the aforementioned jaw bones, as well as the first non-rib-bearing vertebra (orange), the first rib-bearing (precaudal) vertebra (light blue), precaudal vertebra 8 (green), caudal vertebra 3 (orange), caudal vertebra 10 (gold) and the pre-urostyle (final caudal) vertebra (red). (B) Anterior (top) and anterolateral (bottom) view of the lower pharyngeal jaws for each species in (A). Scale for all images is shown as 1cm. See Supplementary Table 1 for details of the specimens used.





**Figure 6. Segmented Bones from *Rhamphochromis esox* (*Rhamphochromis*), *Pallidochromis tokolosh* (*Diplotaxodon*) and *Copadichromis trimaculatus* (*Utaka*).** A close up, lateral view of the head of each species (species name indicated on right), showing the dentary (green), premaxilla (pink) and lower pharyngeal jaw (purple) positioned within a volume render of the head is shown for each specimen on the left. On the right are whole body lateral views showing the aforementioned jaw bones, as well as the first non-rib-bearing vertebrae (orange), the first rib-bearing (precaudal) vertebrae (light blue), precaudal vertebrae 8 (green), caudal vertebrae 3 (orange), caudal vertebrae 10 (gold) and the pre-urostyle (final caudal) vertebrae (red) within a volume rendering of the whole body. Scale for all images is shown as 1cm. See Supplementary Table 1 for details of the specimens used.