

A whole-body micro-CT scan library that captures the skeletal diversity of Lake Malawi cichlid fishes

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ABSTRACT

Here we describe a dataset of freely available, readily processed, whole-body μ CT-scans of 56 species (116 specimens) of Lake Malawi cichlid fishes that captures a considerable majority of the morphological variation present in this remarkable adaptive radiation. We contextualise the scanned specimens within a discussion of their respective ecomorphological groupings and suggest possible macroevolutionary studies that could be conducted with these data. We also describe a methodology to efficiently μ CT-scan (on average) 23 specimens per hour, limiting scanning time and alleviating the financial cost whilst maintaining high resolution. We demonstrate the utility of this method by reconstructing 3D models of multiple bones from multiple specimens within the dataset. We hope this dataset will enable further morphological study of this fascinating system and permit wider-scale comparisons with other cichlid adaptive radiations.

Background & Summary

Cichlids are one of the most speciose families of vertebrates, with over 1000 species in the African Rift Valley alone^{1,2}. Multiple, independent, adaptive radiations of these fishes have evolved in the Great Lakes of East Africa, their associated satellite water bodies, as well as their connecting riverine systems. The radiations of these fishes (Subfamily: Pseudocrenilabrinae³), particularly those associated with Lakes Malawi, Victoria and Tanganyika, have become powerful models for the study of macroevolutionary processes⁴⁻¹⁰, behaviour and physiology¹¹⁻¹⁵, and have emerged more recently as models in evolutionary developmental biology¹⁶⁻¹⁹.

Lake Malawi haplochromine cichlids represent a particularly speciose, phenotypically diverse but genetically homogeneous adaptive radiation of lacustrine fishes. This diversity, comprising approximately 850 species of maternal mouthbrooders, is the most extensive adaptive radiation of vertebrates so far identified^{1,9}. Molecular clock analyses estimate the radiation to be approximately 800 thousand years old⁴, a relatively young radiation when compared to the older system of Lake Tanganyika (~10myr) which contains just 250 species^{7,20}. Despite their high phenotypic diversity, genetic variation in Lake Malawi cichlids is extremely low. Whole genomic comparisons of representatives from all seven distinct ecomorphological groups within Lake Malawi, estimated an average DNA sequence divergence of just 0.19-0.27%⁴ – a range comparable to that within human populations⁶. In addition, a relatively low DNA mutation rate; that alone cannot account for the estimated divergence time of Lake Malawi cichlids^{4,6}, overlapping distributions of interspecific pairwise sequence differences and heterozygosity (intraspecific genetic variation)⁴ only further complicates this enigmatic adaptive radiation.

East African cichlids, including those belonging to the Lake Malawi radiation, have recently emerged as powerful models in evolutionary developmental biology¹⁷. Evolutionary modification of embryological mechanisms drives the evolution of novel adaptations and requires genetic variation²¹. Thus, comparing the embryological development of cichlids, which have limited genetic variation, can enable us to identify specific cases where evolution has modified developmental mechanisms. The diversity of feeding habits of Lake Malawi cichlids, and the ability to causally link morphological differences in craniofacial morphology to these ecological niches, has enabled integrative genetic and morphological studies examining the evolution of these traits²²⁻²⁵. More recent studies have expanded the scope beyond craniofacial phenotypes, including pigmentation patterning²⁶⁻²⁸; body and fin shape^{19,29} and axial elongation¹⁸. In parallel, aided by developments in whole-genome sequencing

40 technologies⁶, it has been possible to considerably improve our understanding of the phylogenetic relationships among Lake
41 Malawi cichlids^{4,9,30,31}. Previously intractable macroevolutionary studies, such as the convergent evolution of hypertrophied
42 lips³² can now take advantage of the relatively robust phylogenies based on whole-genome sequences. Moreover, there are
43 now opportunities to use this new phylogenetic information to focus on the evolution of other traits, such as the axial and
44 appendicular skeleton, that is of key importance in teleost diversification^{33,34}. However, a whole-body μ CT-scan dataset of
45 Lake Malawi cichlid fishes has not yet been described.

46 Here we present a new database of high-resolution X-ray micro-computed tomography (μ CT) scans of Lake Malawi
47 cichlids, providing 3D data on skeletal morphology for the whole body of 56 species across 26 genera. In total these data
48 comprise 116 individuals from seven recognized ecomorphological groupings⁴ (Table 1), contrasting in multiple aspects of
49 morphology, size, behaviour, and habitat preference. We demonstrate the resolution and utility of our dataset by illustrating
50 3D whole-body renderings of several species, and of several skeletal regions of interest. The data will be useful resource
51 for researchers interested in the emergence of morphological variation, including the macroevolutionary patterns common to
52 adaptive radiations^{35,36}. We argue that where digitisation efforts are being taken to characterise adult cichlid morphology, whole-
53 body scans should be standard to ensure that other sources of morphological variation can be investigated. We propose methods
54 that minimise scanning time, thus alleviating financial and time constraints of scanning large numbers of specimens, whilst
55 maintaining sufficiently high resolution for macroevolutionary studies and geometric morphometric analyses of morphological
56 variation.

57 **Methods**

58 **Sample Selection**

59 There are an estimated 850 species of Lake Malawi cichlid fishes⁴. Many of which have not been described, preserved in
60 museum collections or are available on phylogenies based on whole genome evidence. Therefore, to maximise the utility and
61 morphological variation captured by our dataset, we focused on species present in published phylogenies^{4,9,24}, and sought to
62 include as many genera as possible in dataset (see Table 1). We prioritised scanning the type species for genera, and avoided
63 inclusion of species which already had whole body scans freely available online. Our specimens were sourced from the
64 collections at the Natural History Museum in London (NHMUK), from the School of Biological Sciences of the University of
65 Bristol (Martin Genner) and from the School of Natural Sciences of Bangor University (George Turner). In total we scanned
66 116 specimens from 56 species (Supplementary Table 1). Of these 99 were wild-caught, and 17 were laboratory-reared. These
67 laboratory-reared fish included *Astatotilapia calliptera* (Mbaka River, n=10), *Maylandia zebra* (Boadzulu island, n=5) and
68 *Rhamphochromis* sp. 'Chilingali' (n=2), all of which died naturally or were euthanised by anaesthetic overdose [Schedule 1;
69 Animals (Scientific Procedures) Act 1986].

70 **μ CT-Scanning**

71 Since there was already a large collection of specimens present at the Natural History Museum in London (NHMUK) and in the
72 extensive research collections of Martin J Genner (School of Biological Sciences, University of Bristol) and George F Turner
73 (School of Natural Sciences, Bangor University) we decided to take advantage of the scanners present in the CT facility of
74 NHMUK and at the XTM Facility based in the Paleobiology Research Group at the University of Bristol. Of the total 116
75 individuals scanned (56 species), 56 specimens (28 species) were scanned at the NHMUK Imaging and Analysis Centre and 60
76 specimens (28 species) were scanned at the XTM Facility at the University of Bristol. A flowchart, describing all the necessary
77 decisions and required processing steps is provided as Figure 2.

78 **Scanning Arrangement**

79 To maximise the utility of our time and the number of species scanned, multiple specimens were scanned in each individual
80 scan (3). Each batch of specimens was fit to the width of the scan field of view to maximise resolution, and multiple scans were
81 conducted along a the vertical axis in order to scan the full body length of each specimen. Practically, this meant specimens
82 were arranged into batches of similar total length. Batch sizes varied between two and five specimens, with the number of
83 each batch ultimately dependent upon the overall size of the specimens within the batch. Of the 32 batches scanned: 20 were
84 comprised of four specimens; nine of three specimens, and two and one batch(es) of one and five specimens, respectively. Since
85 multiple individuals of different species were often scanned together (Figure 3A, B) and it was critical that individuals of the
86 same species could be readily identified. Therefore, unique, low density objects such as plastic bricks, pipette tips and rubber
87 bands were placed in physical proximity to each specimen, to act as recognisable markers (Figure 3C) which would readily
88 resolve in the reconstructed image stacks (see Post-Scanning Processing). These objects were attached to each individual
89 specimen-containing bag, and the specimens were bundled together, ideally ensuring that the objects faced outwards (Figure
90 3D-F). Specimens were tightly packed into plastic containers, sealed with tape, and allowed to rest upright (head-up) for at least
91 ten minutes so the contents could settle to prevent movement during scanning (Figure 3G-K). We were able to scan, on average,

92 23 specimens per hour at maximal efficiency, an efficiency that was primarily the result of having two people per scanning
93 visit (one scanning and one packing). The scanning rate could be further increased by packaging specimens in advance of the
94 scanning so that each batch can be scanned continuously.

95 **Scanning Procedure**

96 Prior to scanning, a visual inspection of the specimens was made following X-ray exposure to check for internal damage (such
97 as damage to the vertebral column). If present, and possible, specimens were switched for another specimen of the same batch.

98 To maximise scan resolution, each batch of fish was scanned using the full width of the scanner field of view. This
99 necessitates that multiple, overlapping scans were conducted along the vertical axis of the scanner (Z-axis) in order to capture
100 the full body length of each fish. The overlapping scans were subsequently stitched together in the processing stage using the
101 software, [VGStudio Max 3.2.5 64-bit](#) (see below). The maximise efficiency, individuals with similar body lengths were scanned
102 together (see above). The fish scanned ranged in standard length of 3.8cm to 33cm, which meant that scanning parameters
103 varied between batches. The number of projections generated range from 901 to 2001. Exposure times varied between 250 and
104 500 seconds, with a median of 354 seconds. Similarly, power varied greatly between scans, ranging between 12.48 and 37.995
105 watts (mode 37.44W). All scanning parameters, for each batch conducted, as well as for each individual scan can be found in
106 the metadata supplied in Supplementary Table 1.

107 **Post-Scanning Processing**

108 All processing and segmentation was conducted on a machine specifically built for image analysis, with the following
109 specifications: 2 x Intel® Xeon® CPU-ES-2640 @2.60GHz, 2601MHz, 8 Core(s) processors, 128GB of dedicated DDR3 RAM
110 running on Microsoft Windows 10 Pro (Build Number: 10.0.19045). Raw, isometric volumes generated from the CT-scanning
111 were imported into [VGStudio Max 3.2.5 64-bit](#) and anterior and posterior halves were subsequently stitched together by
112 defining overlapping regions of interest in both the anterior, middle (if applicable) and posterior volumes. 16-bit tiff stacks were
113 exported from [VGStudio Max 3.2.5 64-bit](#) and imported into FIJI³⁷, a GUI for ImageJ³⁸. In FIJI, individual fish were cropped
114 out of the 16-bit stacks, which were identifiable due to the unique objects associated with each individual (see above). The
115 brightness was adjusted by extending the distribution of pixel values to remove 0 values, and the tiff stacks were converted into
116 8-bit to decrease file size. Total stack file size was further decreased by removing images at the beginning and end of the stacks
117 that did not contain readily identifiable bone.

118 **Data Availability**

119 Cropped 8-bit tiff stacks of all 116 specimens have been deposited into a dedicated project on Morphosource, accessible [here](#),
120 where they are freely available and can be downloaded by anyone. We provide a short discussion about segmenting, including
121 methods to optimise these reconstructed image stacks below. We politely request that a direct reference be made to this
122 paper if any scans from our dataset are used in future analyses. See Supplementary Table 1 for a breakdown of all scanned
123 specimens.

124 **Data Records**

125 Seven major ecomorphological groupings make up the Lake Malawi haplochromine radiation. The *Diplotaxodon* and
126 *Rhamphochromis* groups are comprised of elongate piscivores and zooplanktivores adapted to the deeper and more open
127 water niches within Lake Malawi^{39,40}. The ‘shallow benthic’ group is speciose set of Lake Malawi cichlids⁴¹, comprised of a
128 morphologically diverse array of species found across a range of habitats, but is largely restricted to benthic habitats less than
129 50m. The ‘deep benthic’ is a comprised a group of species specialised for deep water benthic habitats, typically more than 50m
130 in depth. The ‘utaka’ are a group of typically shallow open water shoaling species, primarily represented by members of the
131 genus *Copadichromis*⁴². The ‘mbuna’ are a distinct clade of relatively small-bodied rock-associated species typically found
132 to depths of ~30m, and with a diverse mix of foraging strategies⁴³. The final grouping is represented by only one species,
133 *Astatotilapia calliptera*, that is often present in shallow macrophyte-rich marginal habitats of the Lake Malawi. It is often
134 referred to as a generalist species — consuming phytoplankton, zooplankton and littoral arthropods. It the only representative
135 of the haplochromine radiation that is widespread in rivers and smaller lakes of the Lake Malawi catchment^{44,45}. Given the
136 number of species sampled in our dataset, the inclusion of several undescribed species as well as limited species-specific data
137 for some of the species in our dataset, we are unable to offer descriptions of every specimen within our dataset. We have instead
138 limited our wider discussion of species within some of the ecomorphological groups to a selection of well-studied species that
139 demonstrate the diversity captured within our dataset.

140 ***Diplotaxodon* and *Rhamphochromis***

141 Nine species of the *Diplotaxodon* group (47%; Table 1), including the type species *Diplotaxodon argenteus* (n=1) and
142 *Pallidochromis tokolosh* (n=2) are present within our dataset. We also sampled 10 species of the *Rhamphochromis* group

143 (71%; Table 1), including the type species *Rhamphochromis longiceps* (n=2) and the remarkably large *Rhamphochromis*
144 *woodi* (n=2, see below), that are endemic to Lake Malawi. We were also able to sample two sympatric species from the crater
145 lake, Lake Kingiri, *Rhamphochromis* sp. “Kingiri Dwarf” (n=2) and *Rhamphochromis* sp. “Kingiri Large” (n=2), as well as
146 *Rhamphochromis* sp. ‘Chilingali’ (n=4) from the satellite, Lake Chilingali (Figure 4), which is now presumed extinct in the
147 wild⁴⁶.

148 The *Diplotaxodon* and *Rhamphochromis* groups are two reciprocally monophyletic diverging lineages of Lake Malawi
149 cichlids⁴ that have adapted to the pelagic zone of Lake Malawi⁴⁰. The majority of species in the groups are piscivorous,
150 although several species, including *Diplotaxodon limnothrissa* (n=2) are predominantly zooplanktivorous⁴⁷. Large-bodied
151 *Rhamphochromis* primarily feed on Lake Malawi sardines (usipa; *Engraulicypris sardella*) and endemic cichlids (e.g. utaka).
152 Members of *Diplotaxodon* and *Rhamphochromis* are among the deepest-living of all Lake Malawi cichlids, with representatives
153 of both being caught at depths exceeding 200 metres – the ‘twilight zone’ where light is almost completely absent⁴⁰. Species
154 within the *Diplotaxodon macrops* complex, which is represented in the scanned samples by *Diplotaxodon macrops* (n=1) (Figure
155 4), *Diplotaxodon* sp. ‘macrops north’ (n=2), *Diplotaxodon* sp. ‘macrops black dorsal’ (n=2) and *Diplotaxodon* sp. ‘macrops
156 ngulube’ (n=2), have been found between 100 and 220m, a depth similarly reported to be occupied by *Rhamphochromis* during
157 the day⁴⁸.

158 Morphological comparisons of *Diplotaxodon* and *Rhamphochromis* with Lake Malawi cichlids from other habitats could
159 provide valuable insights into convergent adaptation of traits enabling occupation of pelagic niches. Divergence along depth
160 gradients is associated with the evolution of reproductive isolation in many marine and freshwater species groups, likely a
161 consequence of the strong selective pressures associated with deeper water, such as the absence of sunlight, greater hydrostatic
162 pressure, and reduced levels of dissolved oxygen^{49,50}. Morphological comparisons of *Diplotaxodon* and *Rhamphochromis*,
163 against closely related littoral species, could be a powerful model for the study of evolution of convergent phenotypes necessary
164 for adapting to pelagic environments. Body elongation, supported by increased vertebral counts, is a common adaptive trait in
165 teleosts adapted to pelagic (and piscivorous) niches⁵¹ and is also the case with *Rhamphochromis*⁵². Vertebral morphology has
166 also been related to swimming kinematics, body shape and habitat preference⁵³ and the evolutionary modification of internal
167 vertebral traits appears to have taken place during adaptation to pelagic environments³⁴. A study of these traits using the Lake
168 Malawi may help to determine the rate at which these phenotypes can become fixed, and provide insight into the role of these
169 morphological adaptations along the benthic-pelagic speciation axis^{49,50}.

170 Remarkable size variation is present within the *Rhamphochromis* genus. *Rhamphochromis woodi* is considered to be
171 one of the largest Lake Malawi cichlids, measuring up to 40 cm standard length³⁹. In contrast, wild-caught specimens of
172 *Rhamphochromis* sp. ‘Kingiri Dwarf’, endemic to the crater lake Kingiri, do not exceed 7.5 cm standard length⁴⁶. Similarly,
173 wild caught *Rhamphochromis* sp. ‘Chilingali’ are also small bodied, with maximum observed standard length of 10.6 cm⁸.
174 Notably *Rhamphochromis* sp. ‘Chilingali’ is relatively amenable to laboratory study, and its elongated body has made it a
175 useful model in evolutionary developmental biology^{17,18}. Given the exceptional size difference in the genus *Rhamphochromis*,
176 our dataset represents a potentially valuable resource for the study of the evolution of allometric scaling, which has not been
177 well studied in cichlids⁵⁴.

178 Shallow Benthic

179 The shallow benthic species group is extremely speciose, with remarkable amount of morphological diversity^{4,41}, and hundreds
180 of species. The majority of shallow benthic species inhabit relatively shallow inshore habitats of Lake Malawi, such as the sand
181 or mud lake floor, or sand-rock transitional zones. Our dataset includes 20 shallow benthic species in 12 genera⁴ (Table 1),
182 including several large, ambush predators, as well as a collection of trophic specialists. For a complete list of shallow benthics
183 see Supplementary Table 1.

184 Large ambush predators represented in the dataset include *Dimidochromis strigatus* (n=1), *Dimidochromis compressiceps*
185 (n=1), *Tyrannochromis macrostoma* (n=1), *Nimbochromis livingstonii* (n=1) and *Nimbochromis polystigma* (n=2). *Dimi-*
186 *dochromis compressiceps* has a generalist piscivore lifestyle, and occupies reed-beds of the Lake. *Nimbochromis livingstonii* and
187 *N. polystigma* are both considered to be ‘sleeper’ (Chichewa: “kaligono”), and have been observed burying themselves within
188 sandy substrate and snatching prey attracted by the disturbed sediment⁴¹. Another member of *Nimbochromis*, *Nimbochromis*
189 *linni* (n=1) has a characteristic downward-projecting snout (Figure 4), enabling it to extract prey from rock crevices^{41,55}.

190 We sampled several shallow-benthic predators, including *Otopharynx speciosus* (n=2), one of the few piscivores within the
191 genus *Otopharynx*. Males of this species have been encountered at depths exceeding 25m⁴¹, suggesting tolerance of relatively
192 deep water, and suggesting the species may have morphological adaptations enabling occupation of deep-water niches similar
193 to *Rhamphochromis* and *Diplotaxodon*. Of the approximately 20 species of *Otopharynx*⁵⁶ we were able to sample an additional
194 three species: *Otopharynx lithobates* (n=3, including the holotype NHMUK 1974.7.5.1); *Otopharynx tetrastigma* (n=2); and
195 the undescribed *Otopharynx* sp. “brooksi nkhatta” (n=1). We also sampled several specialised trophic specialists including the
196 molluscivores *Mylochromis anaphyrmus* (n=1) and *Trematocranus placodon* (n=1) and the invertebrate picker *Placidochromis*

197 *johnstoni* (n=1, Figure 4). The diet of *T. placodon* predominately comprises the gastropods *Bulinus nyassanus* and *Melanoides*
198 *tuberculata*⁵⁷. Enlarged sensory pores and lateral lines form a sonar-like detection system that allows *T. placodon* to sense the
199 movement of these prey within the sediment. Curiously, this strategy and associated morphological characteristics are also
200 associated with *Aulonocara* and *Lethrinops*, both ‘deep benthics’, suggesting convergent evolution of lateral line phenotypes⁹.
201 The specimens in our dataset may enable some morphological comparisons to further investigate differences in the sensory
202 pore characteristics among species.

203 ‘Rock-dwelling’ *Mbuna*

204 The mbuna group dominate the rocky shores of Lake Malawi, and are used as a model system for the study of rapid speciation and
205 adaptive radiation^{25,43,58}. Similar to the shallow-benthics, there are hundreds of species, many of which are undescribed^{41,43}.
206 We aimed to maximise our coverage of the phenotypic diversity in the group by sampling multiple genera, which are largely
207 differentiated on the basis of head, jaw and tooth morphology⁴³. Our dataset includes 7 species (15 individuals) of mbuna,
208 covering 7 of the 14 described mbuna genera (Table 1).

209 *Cynotilapia* can be distinguished from other genera by the presence of unicuspid (conical) teeth^{41,59,60} and is represented in
210 our dataset by *Cynotilapia axelrodi* (n=1, Figure 4). This is a genus of typically planktivorous species⁴¹ and their relatively
211 simple dentition may reflect this lifestyle⁶¹. By contrast, *Maylandia* (*Metriaclima*)⁶², represented by *Maylandia zebra* (n=5),
212 has closely arranged bicuspid teeth, that is uses for pulling and scraping loose Aufwuchs (periphyton) attached to the rocks
213 found in their preferred preferred rocky habitats^{59,63}. *Tropheops* and *Iodotropheus*, represented by *Tropheops tropheops* (n=2)
214 and *Iodotropheus sprengerae* (n=2), also have closely packed bicuspid teeth, that they use to feed on epilithic algae which
215 they pluck with sideways, upwards head jerks, a behaviour likely supported by *Tropheops*’ characteristic steeply sloped vomer
216 (71-96°)⁴¹. Members of *Petrotilapia*, represented by *Petrotilapia genalutea* (n=1) have a mixed combination of tricupsid
217 and unicuspid teeth that they use to comb loose periphyton from rock surfaces⁶⁴. A further represented mbuna genus is the
218 monotypic *Genyochromis*, represented by *Genyochromis mento* (n=2). Like the majority of mbuna, *G. mento* has prominent
219 outer bicuspid teeth, and are supported by smaller inner tricupsid teeth^{41,59}. In contrast to most other mbuna, however, *G. mento*
220 is a highly specialised feeder, a lepidophage (scale-eater), that targets the the caudal and anal fins of other cichlids in rocky
221 habitats^{41,59,65}. The preferred striking side of *G. mento* significantly correlates with left-right asymmetry of the dentary, with
222 right and left-leaning individuals preferring to strike the corresponding side, respectively, of their prey. Interestingly, however,
223 a comparison of their jaw laterality with *Perissodus microlepis*, a lepidophage endemic to Lake Tanganyika²⁰, showed that
224 laterality in *G. mento* is weaker than in *P. microlepis* – likely a result of phylogenetic constraint from their shorter evolutionary
225 history and their herbivorous ancestors⁶⁵.

226 The craniofacial bones commonly studied in mbuna, such as the dentary, premaxilla, pharyngeal jaws, as well as their
227 associated teeth, can be segmented from specimens in the dataset (see *G. mento*, Figure 5). Future sampling should focus
228 on the seven remaining genera not sampled in our dataset: *Abactochromis*, *Chindongo*, *Cyathochromis*, *Gepyrochromis*,
229 *Labeotropheus*, *Melanochromis* and *Pseudotropheus*.

230 *Astatotilapia calliptera* and Ruaha Catchment

231 *Astatotilapia* is polyphyletic and current members of the genus are widespread across East and North Africa^{6,66,67}. Only
232 one species of *Astatotilapia* is native to Lake Malawi, *Astatotilapia calliptera*, which is also found in East African rivers
233 flowing eastward to the Indian Ocean, from the Rovuma River in the north, to the Save River in the south. Given the wide
234 distribution of the species, it is perhaps unsurprising that intraspecific genetic variation within the species is comparable to that
235 of the whole Lake Malawi radiation^{4,6}. Despite their wide distribution and relatively large intraspecific genetic variation, they
236 phylogenetically cluster within the Lake Malawi radiation (Figure 1), forming a sister clade to the mbuna, with which they
237 share an excess of alleles⁴. This pattern, alongside a perceived riverine ‘generalist’ lifestyle, has led to the hypothesis that either
238 Lake Malawi cichlids radiated from an *A. calliptera*-like ancestor or that *A. calliptera* is the sympatric ancestor of all Lake
239 Malawi cichlids^{4,6,41,67}.

240 Given the importance of *A. calliptera* in the Lake Malawi radiation, we sampled multiple individuals from multiple
241 populations. We were able to scan nine laboratory-reared individuals from the Mbaka river population, which flows into the
242 northern end of Lake Malawi⁶⁸. We also scanned individuals from Lake Chilwa (an endorheic lake south-east of Lake Malawi⁶⁹;
243 n=2), Lake ‘Misoko’, presumably Lake Masoko (a crater lake north of Lake Malawi⁶⁸, n=2), and wild-caught individuals from
244 the main body of Lake Malawi (n=2). Populations of *A. calliptera* differ in life history strategies⁷⁰ and are also undergoing
245 sympatric speciation along a depth gradient in at least one location (Lake Masoko)⁸, where littoral and benthic *A. calliptera*
246 ecomorphs have diverged in multiple characteristics, including body shape and trophic specialism, in approximately 1000
247 years⁸. Therefore, it is possible that morphological evaluations of more populations of *A. calliptera* will reveal further diversity,
248 potentially providing greater insight into the role it has taken in generating the wider Lake Malawi haplochromine radiation.

249 A key part of macroevolutionary studies is the estimation of ancestral state of traits based on the morphology of their
250 descendants⁷¹. This relies on a comprehensive understanding of trait diversity across taxa, and such data can also inform

models of morphological evolution, and enable estimates of rates of phenotypic evolution. Since the genetic diversity of the Lake Malawi radiation was possibly seeded by multiple riverine species⁵, we sought to add specimens to the dataset that could enable the morphological reconstruction of the common ancestor of the Lake Malawi radiation. Therefore, we also sampled two additional species of *Astatotilapia*. These included *Astatotilapia gigliolli* (n=2) and *Astatotilapia* sp. ‘Ruaha blue’ (n=2), native to the Great Ruaha River^{66,67}. Construction of a mtDNA-based phylogeny initially placed *Astatotilapia* sp. ‘Ruaha blue’ as a sister taxa to the Lake Malawi radiation⁶⁶. However, a phylogeny based on variation within whole-genome sequences has shown *A. gigliolli* and *A. sp. ‘Ruaha blue’*, sister taxa, form a sister clade with both the Lake Malawi and Lake Victoria radiations (see Figure 1) – a topology that is likely the result of an ancestral hybridisation event with the ancestors of both lineages prior to their respective adaptive radiations⁵. Therefore, the addition of species from the Ruaha catchment, may therefore enable a more robust estimation of the ancestral phenotype of Lake Malawi cichlids.

Deep Benthic and ‘Utaka’

We sampled deep- water benthic species from two genera; *Alticorpus* and *Lethrinops*^{9,45} (Table 1). *Alticorpus*, like *Aulonocara* (not represented in the dataset) is characterised by the presence of greatly enlarged cranial sensory openings and lateral line canals used to detect prey in the sediment. Deep-water benthic species are found below 50m, a ‘twilight’ zone with very little visible light. *Alticorpus macrocleithrum* (n=3) is found between 75m and 125m, with abundance peaking above 100m⁷², a depth similarly occupied by deep-water *Lethrinops*⁷³, including *Lethrinops gosseii* (n=1). Several species of *Lethrinops*, however, inhabit shallower water^{4,41}. We sampled two species of shallow water *Lethrinops*, including *Lethrinops auritus* (n=2), and *Lethrinops albus* (n=2), both of which phylogenetically cluster within the ‘shallow benthic’ lineage (Figure 1).

Our dataset also contains four species of zooplankton-feeding, shoaling cichlids which are commonly referred to as ‘utaka’. Utaka is primarily made up of species belonging to *Copadichromis*⁴², with a small number of species also belonging to *Mchenga* and the older *Nyassachromis*⁷⁴. However, their placement within the utaka is disputed and they have not been considered in our species/genera counts (see Table 1). We sampled four species of *Copadichromis*: *Copadichromis likomae* (n=2), *Copadichromis quadrimaculatus* (n=2), *Copadichromis trimaculatus* (n=2, see Figure 6) and *Copadichromis virginalis* (n=2). Utaka feed in the water column, and can be commonly found close to the shore⁴¹. *Copadichromis* are generally characterised by their relatively small, highly protrusible mouths, that they use to suck zooplankton into their mouths, as well as numerous long gill rakers which strain plankton from the water that enters their mouths as a result of their sucking feeding mechanism^{41,75}.

Both the deep benthics and utaka are currently underrepresented within our dataset and future sampling should aim to add additional species of *Copadichromis*. In addition, sampling missing ‘deep benthic’ genera, such as *Aulonocara* and *Tramitichromis* should be prioritised for future sampling efforts. *Aulonocara stuartgranti* and *Aulonocara steveni* would be particularly interesting future additions and could offer interesting morphological comparisons with deeper living species. Moreover, given that *Lethrinops* is polyphyletic^{73,76}, additional sampling of *Lethrinops* species could provide morphological data to support future systematic studies.

Technical Validation and Usage Notes

As mentioned above, entries for each specimen can be found in Supplementary Table 1 and cropped, reconstructed 8-bit tiff images stacks of all specimens scanned can be accessed and downloaded from a dedicated [project](#) on Morphosource.

To validate our μ CT-scan data, we imported the reconstructed image stacks into [Avizo Lite \(v9.3.0\)](#), a proprietary software developed by Thermo Fisher Scientific and either generated a full body volume render or rendered 3D models of the whole body. Surfaces from 3D whole-body renderings were exported from AvizoLite as Polygon File Format (.ply) files and imported into [MeshLab](#) for visualisation and manipulation. We note that there are free, open access alternatives to [Avizo Lite \(v9.3.0\)](#) for the segmentation of 3D-image data, such as [3D-Slicer](#), which has a large and active community of users⁷⁷ and [Dragonfly](#) which supports the use of deep learning to automatically segment 3D image data and offers non-commercial licenses for academic use free-of-charge. Example whole-body 3D renderings for representatives from each ecomorphological group can be found in Figure 4 and for each specimen in the dataset in the Supplementary Material.

In addition, we manually segmented multiple bones, including the dentary, premaxilla, lower pharyngeal jaw and multiple vertebral types in several species discussed above. This included: the ‘generalist’ *Astatotilapia calliptera*; the ‘mbuna’ and lepidophage, *Genyochromis mento*; the ‘shallow benthic’ and snail crusher, *Trematocranus placodon* (Figure 5); the pelagic piscivores, *Rhamphochromis esox* and *Pallidochromis tokolosh* and the zooplanktivorous (utaka), *Copadichromis trimaculatus* (Figure 6). Lower pharyngeal jaws that are highly variable among species of Lake Malawi cichlids²⁵ were particularly well resolved. For example, newly erupting teeth were visible on the relatively large, and dense, lower pharyngeal jaw of *Trematocranus placodon* (Figure 5B). Similarly, renderings of multiple vertebral types (Figures 5–6) were also good quality. The zygapophyses and fine structure of the vertebral centra, sometimes including the neural canal, were also well resolved. All 3D-renderings of these bones can be found in the Supplementary Materials as downloadable .ply files.

303 The computer specifications we used for the all analyses (see Methodology) are hard to find on personal, or older machines
304 and some users may find it difficult to work with some of our larger image stacks. To minimise memory usage during
305 segmentation and speed up processing, cropped reconstructed stacks can be loaded in multiple increments (note that the Z-voxel
306 size must be multiplied by said increment). We tested this and found that roughly comparable models could be generated,
307 although it was clear that finer morphological detail was absent (data not shown). Therefore, where possible, the whole stack
308 should be used when segmenting regions of interest. In addition, since these regions were manually segmented, many of the
309 segmentation steps rely on the judgment of the individual segmenting and rendering of the regions of interest. We found that
310 segmenting from median-filtered reconstructed image stacks drastically lowered the quality of the rendered models (data not
311 shown) and would suggest refraining from segmenting from a median-filtered image stack. In addition, we found that relatively
312 low smoothing factors were best for rendering surfaces from segmented regions of interest. In *Avizo Lite (v9.3.0)*, a smoothing
313 factor between 0-10 (including rational intermediates) can be applied when rendering surfaces of segmented regions of interest.
314 We rarely found it necessary to use a value above '3'; indeed, all whole-body 3D models were smoothed with a factor of 2.5,
315 and we suggest, regardless of the tool used to smooth and render segmented surfaces, to use smoothing cautiously. Furthermore,
316 we were able to scan multiple species of *Copadichromis* (utaka), see above and were able to generate relatively good quality
317 models (see Figure 6 and the Supplementary Models). However, it is clear that some of the jaw structures did not resolve as
318 well as in other specimens. It is possible that the jaw bones of these fish species are not particularly dense, which made it
319 difficult to image them using the same scanning procedure as that was used for all of the other specimens. Therefore, in future,
320 we would recommend care when segmenting bones from the *Copadichromis* species present in this dataset and also to increase
321 the power and exposure time for future sampling of *Copadichromis*.

322 Here we have presented the first, comprehensive, freely available, whole-body μ CT-scan dataset of Lake Malawi cichlids.
323 We have described several macroevolutionary studies that could be conducted with this dataset to better understand this
324 remarkable cichlid adaptive radiation and include suggestions for future sampling. Our Lake Malawi dataset now joins two
325 other East African adaptive radiation datasets, the recent haplochromine Lake Victoria library⁷⁸ and the extensive μ CT-scan
326 dataset of Lake Tanganyika cichlid fishes⁷. Therefore, the addition of our dataset now offers a unique opportunity for much
327 wider, and systematic, morphological comparisons within East African cichlids. We have also described a methodology to
328 efficiently μ CT-scan multiple specimens simultaneously; reducing scanning time and financial cost, whilst maintaining scan
329 quality, demonstrating the utility of this method by reconstructing 3D-models of multiple bones from multiple specimens
330 within our dataset. We hope the availability of these data will inspire people to address some of the many questions left still to
331 understand this remarkable adaptive radiation, permit wider-scale comparisons with other cichlid adaptive radiations and set a
332 precedent to make whole-body μ CT scans the automatic default for any sampling efforts involving cichlids.

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503 Acknowledgements

504 We thank Vincent Fernandez, CT facility manager at the NHMUK and Liz Martin-Silverstone at the XTM Facility at the
 505 University of Bristol for organising access to their respective imaging facilities. Thank you to the fish whose lives were
 506 sacrificed for this work. This research was partly funded by a Biotechnology and Biological Sciences Research Council
 507 (BBSRC) studentship (Grant Number: 2445747).

508 Author contributions statement

509 C.V.B. and B.V. conceived the study. C.V.B and R.B. designed the data acquisition pipeline and dataset curation methodology.
 510 C.V.B. and R.B. conducted the imaging. C.V.B. performed the sample collection, processed all the raw image data, and wrote
 511 the manuscript. B.V., R.B. and M.J.G. edited the manuscript. J.M. organised specimens at the NHMUK. G.F.T. and M.J.G.
 512 contributed specimens to be imaged. All authors reviewed the manuscript.

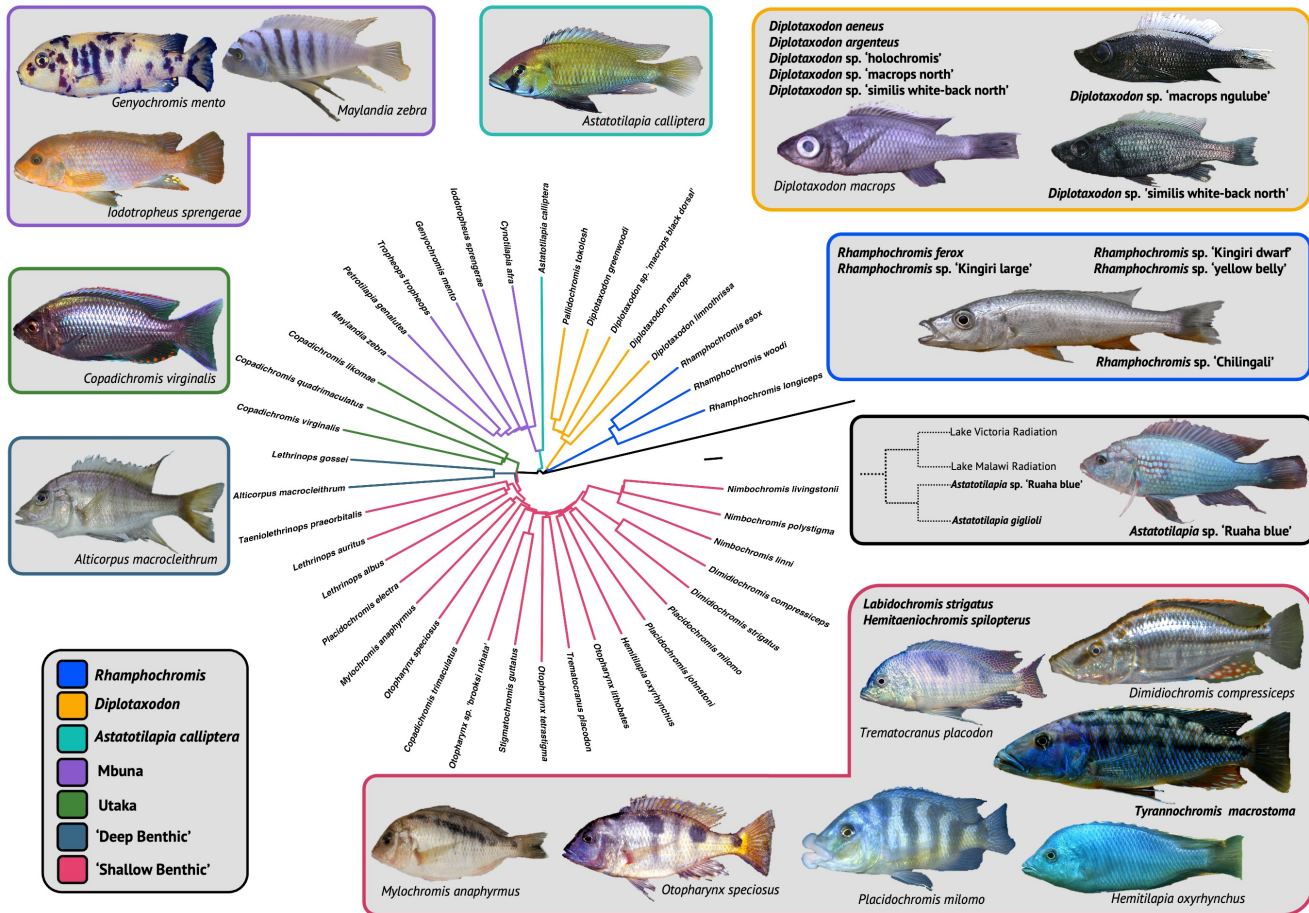
513 Competing interests

514 The authors declare no competing interests.

515 Figures & Tables

Ecomorphology	Genera Sampled	Genera Sampled (%)	Species Sampled	Species Sampled (%)
<i>Astatotilapia (calliptera)</i>	1 of 1	100%	3*	N/A
Deep Benthic	2 of 5	40%	3 (5 [†]) of 150	2.00% (3.33%)
<i>Diplotaxodon</i>	2 of 2 [‡]	100%	9 of 19	47.37%
Mbuna	7 of 12	58.33%	7 of 328	2.13%
<i>Rhamphochromis</i>	1 of 1	100%	10 of 14	71.43%
Shallow Benthic	12 of 32	37.50%	20 of 287	6.97%
Utaka	1 of 1	100%	4 [§] of 55	7.27%
Total	26 of 54	48.15%	56 of 856	6.54%

Table 1. Genera and species represented within the dataset. The number of genera and species for each ecomorphological group are the same as those used in Malinsky et al., 2018⁴. **Astatotilapia calliptera*, *Astatotilapia* sp. ‘Ruaha blue’ and *Astatotilapia gigliolli*. [†]If considering the addition of *Lethrinops albus* and *Lethrinops auritus* that cluster within the ‘Shallow Benthics’. [‡]Includes *Pallidochromis*. [§]Includes *Copadichromis trimaculatus* which clusters within the shallow benthics in the phylogeny depicted in Figure 1.



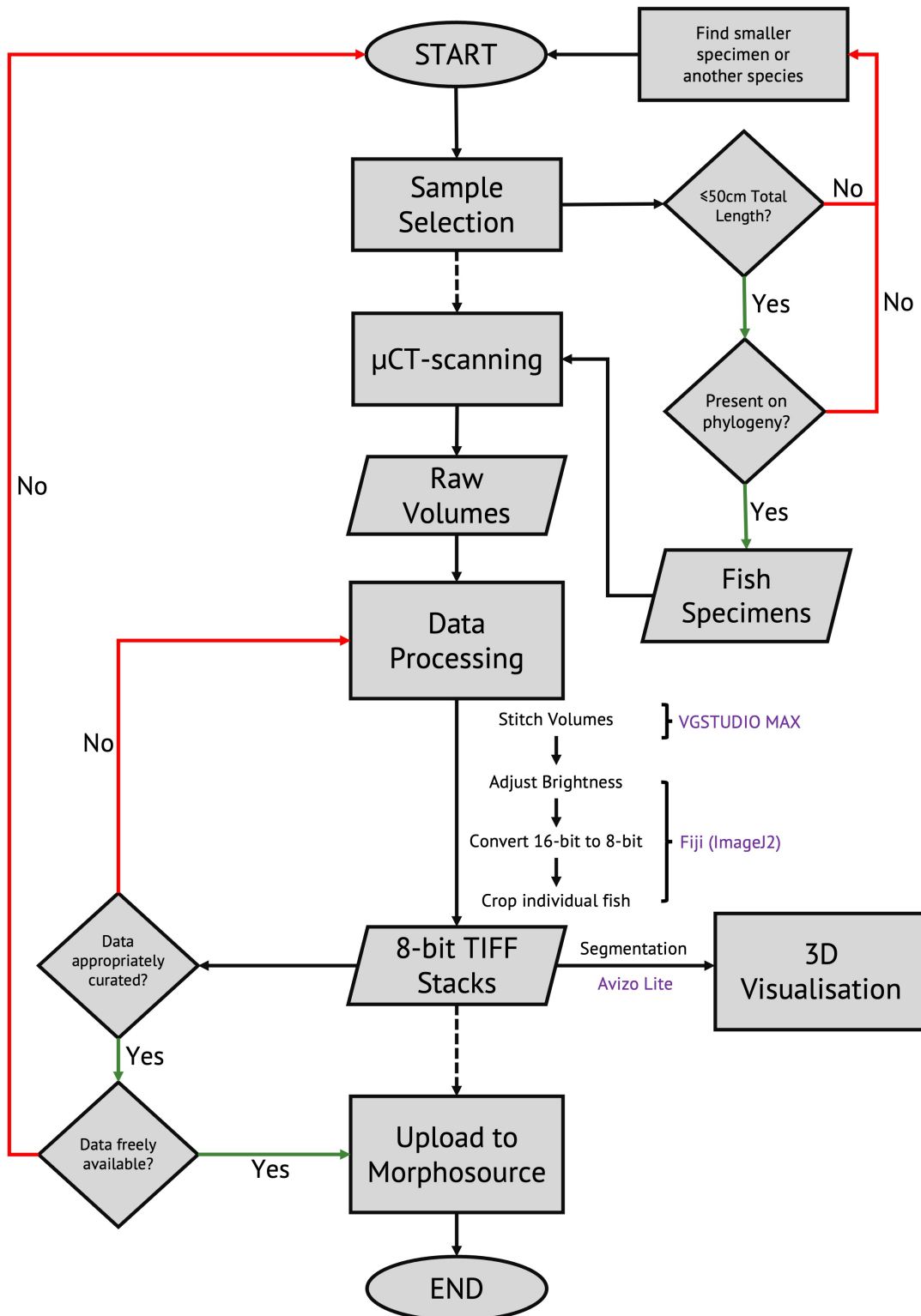


Figure 2. Flowchart of μ CT-scanning, image processing and segmentation methodology. The flowchart outlines the necessary decisions that were made during collation of the described μ -CT scan dataset. Rectangles represent processes; parallelograms represent inputs or outputs; diamonds represent decisions. It is sufficiently generalised that it can be reused for future data collection. We were focused on generating data for a specific macroevolutionary study, so we restricted the dataset to species with known phylogenetic placements. Software associated with data processing steps are indicated in purple.

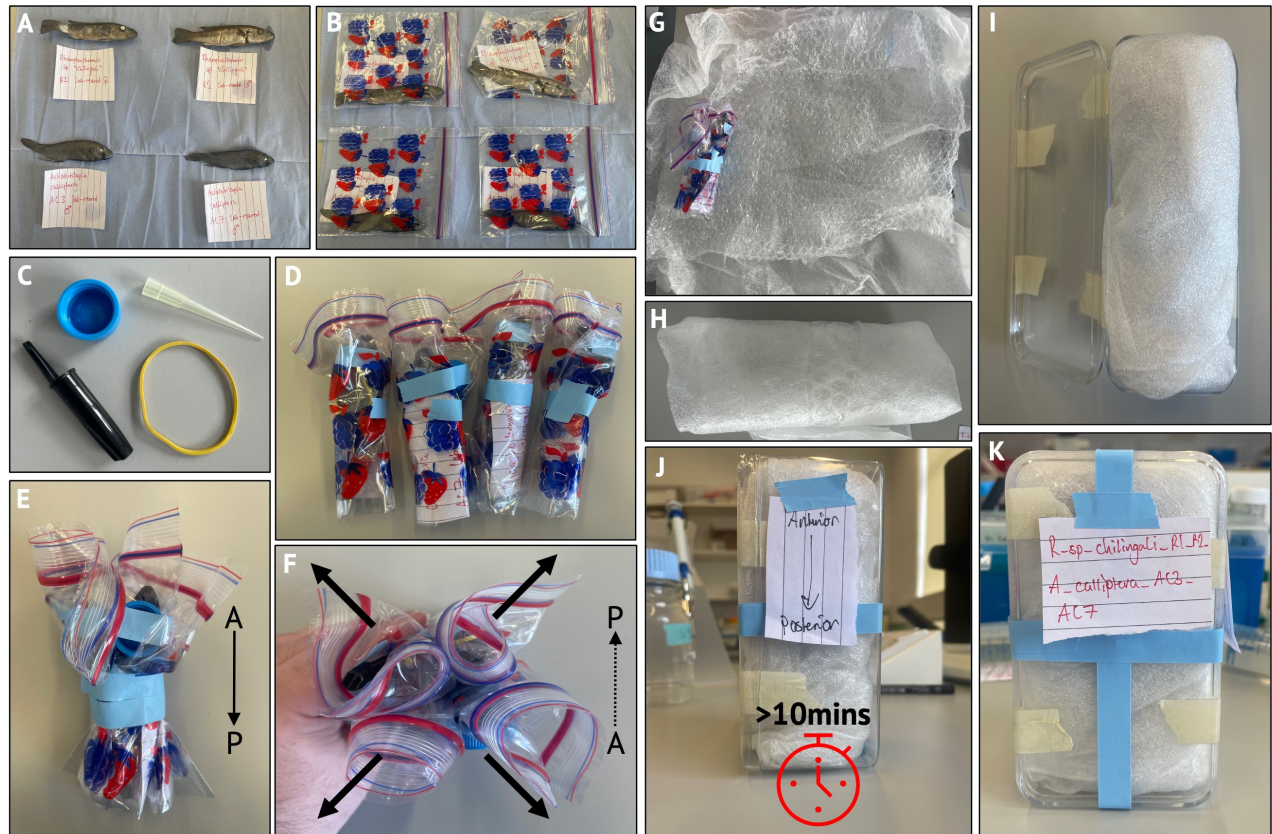


Figure 3. Specimen preparation for μ CT-scanning. Multiple fish were scanned at the same time. (A-B) Individual fish were labelled and placed in separate plastic bags (so they could be correctly identified and correctly stored after imaging). Unique objects (C) that would be readily identifiable were attached to the outside of these bags, ideally close to the heads, positioned outwards (F, arrows), and bundled together with tape (D-F) all with the same orientation (head-up). Bundles were then wrapped in bubble wrap and other packaging material and tightly sealed inside a plastic container, again head-up. Containers were left for at least ten minutes to settle to prevent movement during scanning.

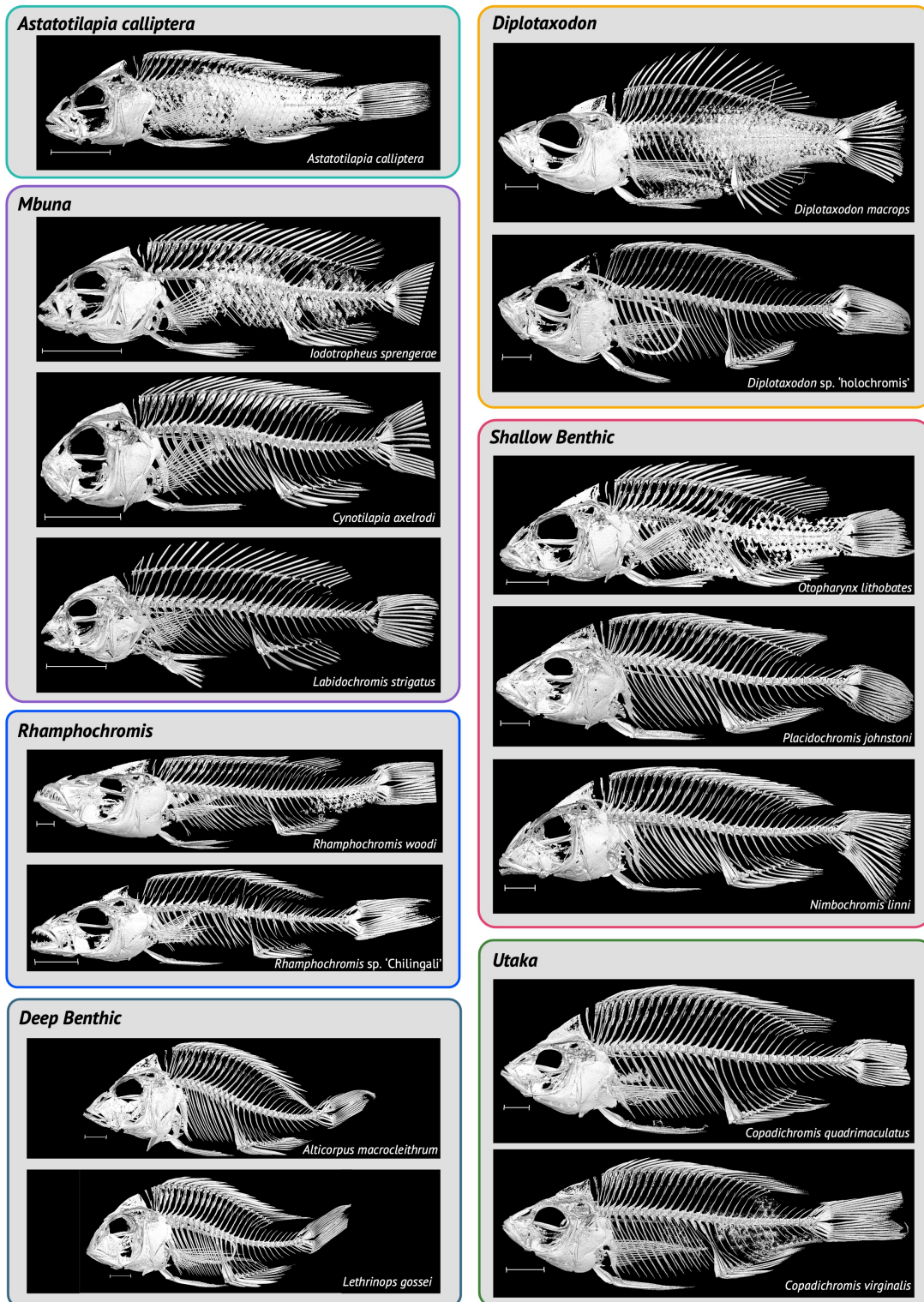


Figure 4. Whole-body 3D models of select specimens from the dataset. Scale is 1cm. Specimens are arranged according to the ecomorphological group they belong to. Species names are indicated. Specimens used are indicated in Supplementary Table 1. Some of the specimens have been rendered with their scales, mainly as we found that several specimens had particularly thick scales making it difficult to render whole-body models without including the scales. It is worth noting, however, that the lateral lines have resolved quite well in the models with scales present and with a sufficiently high threshold value scales could resolve well. The ring structure in *Diplotaxodon* sp. 'holochromis' is a rubber band used for identification purposes.

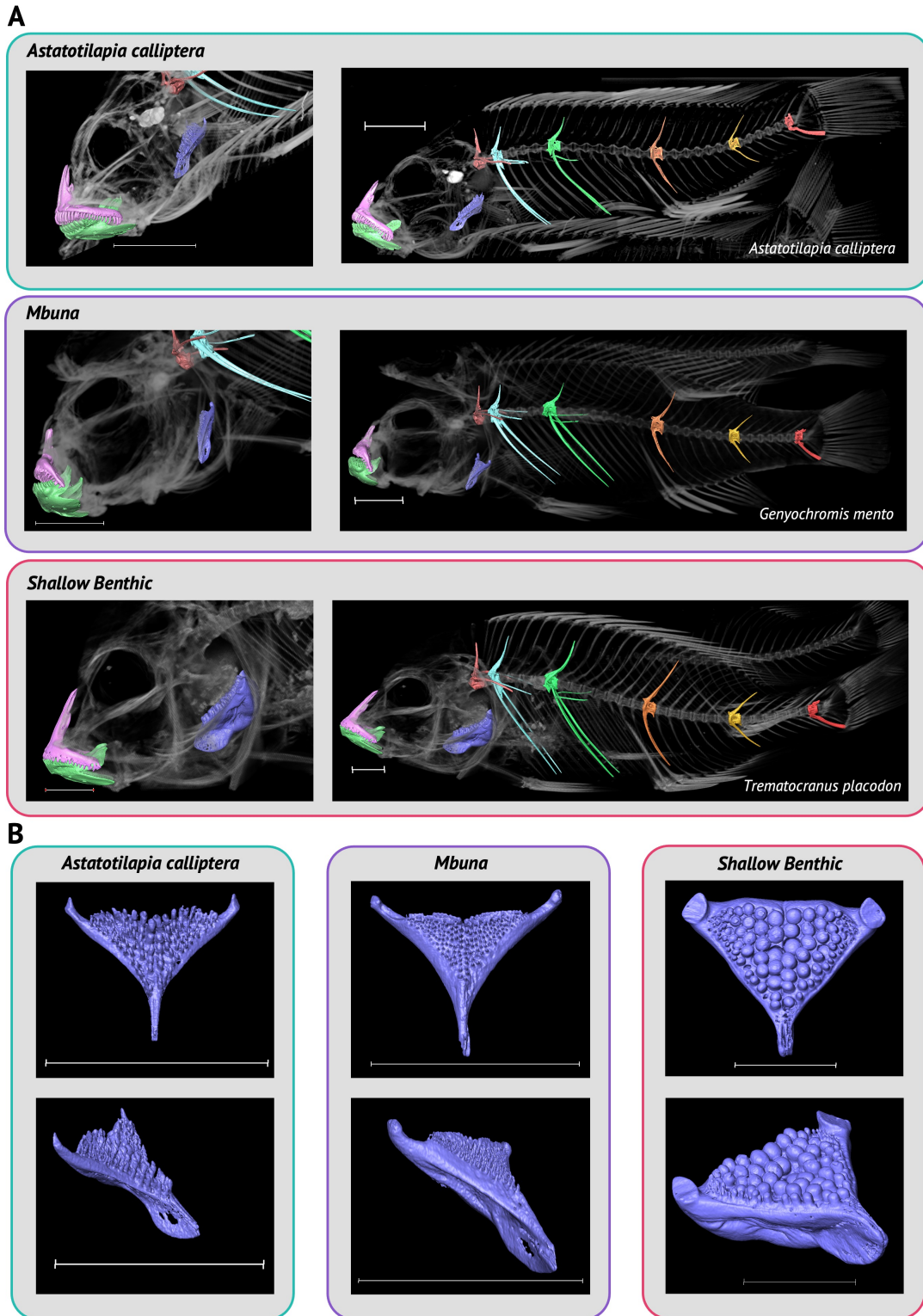


Figure 5. Segmented Bones from *Astatotilapia calliptera*, *Genyochromis mento* (mbuna) and *Trematocranus placodon* (shallow benthic). (A, left) A close up, lateral view of the head of each species (species name indicated on right), showing the dentary (green), premaxilla (pink) and lower pharyngeal jaw (purple) positioned within the a volume render of the head. (A, right) A whole body lateral view showing the aforementioned jaw bones, as well as the first non-rib-bearing vertebra (orange), the first rib-bearing (precaudal) vertebra (light blue), precaudal vertebra 8 (green), caudal vertebra 3 (orange), caudal vertebra 10 (gold) and the pre-urostyle (final caudal) vertebra (red). (B) Anterior (top) and anterolateral (bottom) view of the lower pharyngeal jaws for each species in (A). Scale for all images is shown as 1cm. See Supplementary Table 1 for details of the specimens used.

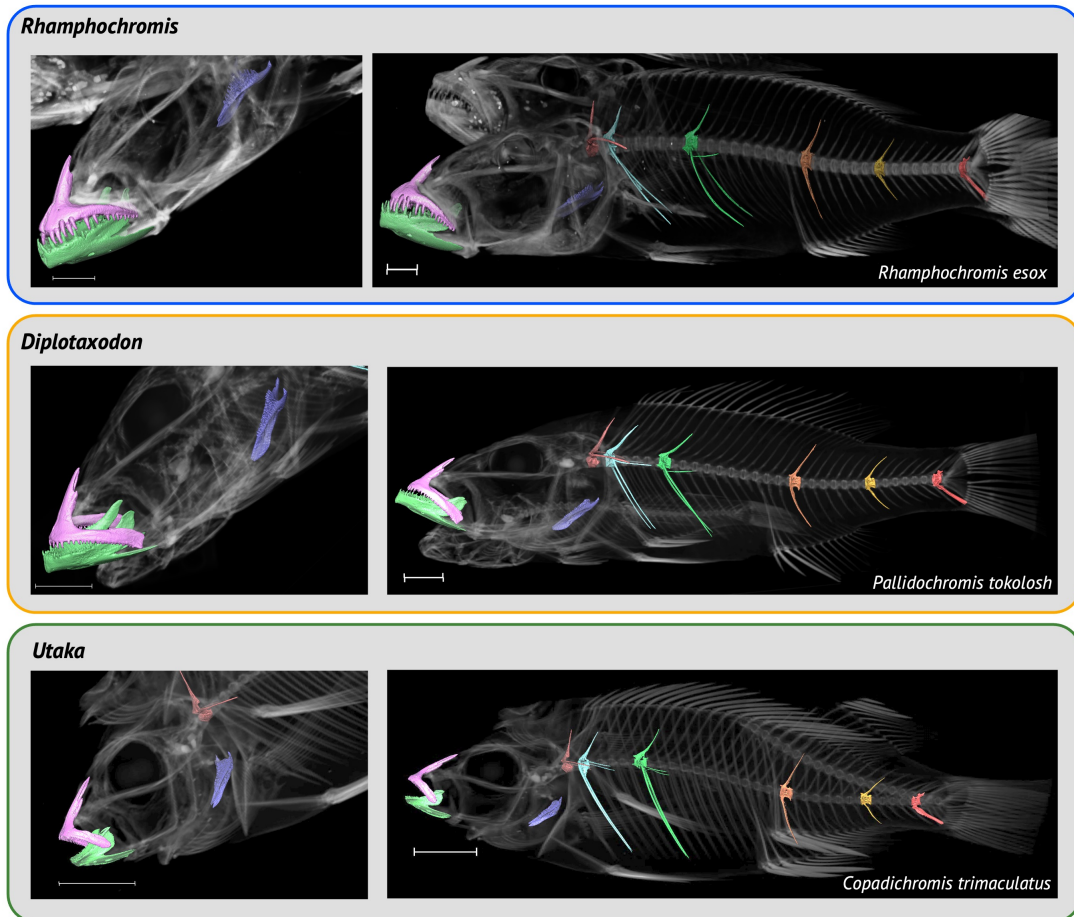


Figure 6. Segmented Bones from *Rhamphochromis esox* (*Rhamphochromis*), *Pallidochromis tokolosh* (*Diplotaxodon*) and *Copadichromis trimaculatus* (*Utaka*). A close up, lateral view of the head of each species (species name indicated on right), showing the dentary (green), premaxilla (pink) and lower pharyngeal jaw (purple) positioned within a volume render of the head is shown for each specimen on the left. On the right are whole body lateral views showing the aforementioned jaw bones, as well as the first non-rib-bearing vertebrae (orange), the first rib-bearing (precaudal) vertebrae (light blue), precaudal vertebrae 8 (green), caudal vertebrae 3 (orange), caudal vertebrae 10 (gold) and the pre-urostyle (final caudal) vertebrae (red) within a volume rendering of the whole body. Scale for all images is shown as 1cm. See Supplementary Table 1 for details of the specimens used.