

## 1 Early evolution of the ecdysozoan body plan

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19

## 20 Abstract

21 **Extant ecdysozoans (moulting animals) are represented by a great**  
22 **variety of vermiform or articulated organisms. However, controversies**  
23 **remain about the nature of their ancestral body plan although the**  
24 **vermiform hypothesis seems to prevail. We describe here *Beretella***  
25 ***spinosa* gen et sp. nov. a tiny ecdysozoan from the early Cambrian,**  
26 **Yanjiahe Formation, South China, with an unusual sack-like appearance,**  
27 **single opening, and spiny ornament. *Beretella* has no equivalent among**  
28 **animals, except *Saccorhytus* from the basal Cambrian. Phylogenetic**  
29 **analyses resolve both forms as a sister group (Saccorhytida) to all**  
30 **known Ecdysozoa, thus suggesting that ancestral ecdysozoans may**  
31 **have been non-vermiform animals. Saccorhytids are likely to represent**  
32 **an early dead-end off-shot along the stem-line Ecdysozoa that possibly**  
33 **evolved through anatomical simplification (e.g. lack of anus). Although**  
34 **extinct during the Cambrian, this animal lineage provides precious**  
35 **insight into the early evolution of Ecdysozoa and the nature (possibly**  
36 **non-vermiform) of the earliest representatives of the group.**

37

## 38 Introduction

39 The Ediacaran–Cambrian transition is marked by the appearance in the fossil  
40 record of a variety of new body plans that prefigure the majority of present-day  
41 animal lineages, including the ecdysozoans a huge clade that encompasses  
42 all invertebrate animals growing through successive moulting stages, such as  
43 panarthropods (Arthropoda, Onychophora, Tardigrada), scalidophoran (incl.

44 Priapulida) and nematoid worms(Erwin, 2020). Altogether ecdysozoans  
45 represent a very high percentage of animal biodiversity and disparity,  
46 inhabiting almost all possible ecological niches on Earth(Brusca et al., 2016).  
47 The nature of the last common ancestor of Ecdysozoa (LCAE) remains largely  
48 unresolved, even though worms are prevalent before the rise of panarthropods  
49 as trace and body fossils in basal Cambrian and late Ediacaran rocks(Buatois  
50 et al., 2014; Liu et al., 2014; Vannier et al., 2010). Some molecular phylogenies  
51 also predict that the most basal ecdysozoans were vermiform  
52 organisms(Howard et al., 2022; Laumer et al., 2019) that possibly diverged in  
53 the Ediacaran (Howard et al., 2022; Rota-Stabelli et al., 2013). Current  
54 reconstruction based on fossil and developmental evidence features the  
55 ancestral ecdysozoan as a millimeter-sized worm(Budd, 2001; Valentine and  
56 Collins, 2000) with a terminal(Ortega-Hernandez et al., 2019) or ventral mouth  
57 (Martín-Durán and Hejnol, 2015; Nielsen, 2019). Clearly, the discovery of  
58 *Saccorhytus*(Han et al., 2017; Liu et al., 2022; Shu and Han, 2020b) in the  
59 basal Cambrian of China (Kuanchuanpu Formation; ca. 535 Ma(Sawaki et al.,  
60 2008)) that is anything but a worm sowed doubt among scientists. *Saccorhytus*  
61 is a sac-like secondarily phosphatized microscopic animal spiked with conical  
62 sclerites and a single opening that was first seen as the earliest known  
63 deuterostome(Han et al., 2017) but is now considered as an ecdysozoan on  
64 more solid grounds(Liu et al., 2022; Shu and Han, 2020b), thus broadening the  
65 anatomical spectrum of the group and its disparity in the Cambrian and  
66 reopening the debate on the nature of LCAE.

67 We describe here *Beretella spinosa* gen. et sp. nov. from Member 5 of the  
68 Yanjiahe Formation (basal Cambrian Stage 2, ca. 529 Ma, Hubei Province,  
69 China) that shares morphological traits with *Saccorhytus* such as an ellipsoidal  
70 body, a pronounced bilaterality, a spiny ornament made of broad-based  
71 sclerites, and a single opening. Cladistic analyses are made to resolve the  
72 position of both *Beretella* and *Saccorhytus* that provide key information on the  
73 early evolution of the group.

74

## 75 **Systematic palaeontology**

76

77 Superphylum Ecdysozoa Aguinaldo et al.(Aguinaldo et al., 1997)

78 Phylum Saccorhytida Han, Shu, Ou and Conway Morris, 2017 stat. nov.

79

80 **Remarks.** Saccorhytida first appeared in the literature as a new stem-group  
81 Deuterostomia that accommodated a single species, *Saccorhytus*  
82 *coronarius*(Han et al., 2017). Since *Saccorhytus* is no longer considered a  
83 primitive deuterostome and, instead, more likely belongs to ecdysozoans,  
84 Saccorhytida became an extinct Order of Ecdysozoa(Liu et al., 2022; Shu and  
85 Han, 2020b). Because both *Saccorhytus* and *Beretella* display major  
86 morphological differences with all other known ecdysozoan phyla (Nematoida,  
87 Scalidophora, and Panarthropoda), Saccorhytida is tentatively elevated here

88 to the rank of phylum within Ecdysozoa.

89

90 **Emended diagnosis.** Microscopic, ellipsoidal body shape with pronounced  
91 bilateral symmetry expressed by paired spiny sclerites. Single, presumably  
92 oral opening on assumed ventral side (no anus).

93

94 **Remarks.** Only two forms, *Saccorhytus* and *Beretella* are currently placed  
95 within Saccorhytida, making it premature to formally define intermediate  
96 taxonomic categories such as an order and a family.

97

98 *Beretella spinosa* Han, Guo, Wang and Qiang, gen. et sp. nov.

99

100 **Etymology.** From ‘béret’, French, that designates a soft, visorless cap  
101 referring to the overall shape of this species, and ‘spinosa’, an adjective (Latin),  
102 alluding to its spiny ornament.

103

104 **Holotype.** CUBar138-12 (Fig. 1a–c).

105 **Paratype.** CUBar171-5 Fig. 1h, i) and CURBar121-8 (Fig. 1j, k).

106

107 **Diagnosis.** Body with a beret-like lateral profile. Convex side (presumably  
108 dorsal) with an elevated (presumably posterior) and lower (presumably  
109 anterior one) end. The opposite side (presumably ventral) flattened. Bilateral  
110 symmetry well expressed in the overall body shape (sagittal plane) and sclerite  
111 distribution. Antero-posterior polarity. Convex side with a slightly elevated  
112 sagittal stripe topped with a single row of four aligned spines (S1) and five  
113 additional spines (S2) on each side. Six broad-based conical sclerites (S3)  
114 distributed in two symmetrical longitudinal rows plus two sagittal ones. Double  
115 rows of six marginal spines (S4 and S5). Flattened side often pushed in and  
116 partly missing, bearing a possible mouth opening. Possible oral spine.

117

118 **Stratigraphy and locality.** *Watsonella crosbyi* Assemblage Zone (Guo et al.,  
119 2021), Member 5 of the Yanjiahe Formation (Cambrian Terreneuvian, Stage 2)  
120 in the Yanjiahe section near Yichang City, Hubei Province, China  
121 (Supplementary Figs. 1, 2).

122

### 123 **Description and comparisons**

124 The body of *Beretella spinosa* is secondarily phosphatized and has a  
125 consistent beret-like three-dimensional shape in the lateral view. Its maximum  
126 length, width, and height range from 1.0–2.9 mm, to 975–2450  $\mu\text{m}$ , and 500–  
127 1000  $\mu\text{m}$ , respectively (Fig. 1, Supplementary Tables 1–3). The ratio of the  
128 maximal length to width is 1.6:1 (Supplementary Fig. 3). As seen in top view, *B.*  
129 *spinosa* shows a small lateral constriction at approximately mid-length (Fig. 1a,  
130 c).

131 The body has a convex, assumedly dorsal side with one, presumably posterior

132 end more elevated than the other (Fig. 1b, e, i, k). This elevation is gradual  
133 along the sagittal plane and then becomes more abrupt near the low elevated,  
134 presumably anterior end. The opposite, assumedly ventral side is less well  
135 preserved and seems to have been originally flattened.

136 The convex side bears a complex ornamented pattern made of five sets (S1–  
137 S5) of spiny sclerites directed towards the more elevated end (Figs. 1a, b, d, e,  
138 h–k, 2a, b, d). These sclerites were originally pointed (Figs. 1a, d, e, 2b, k, l,  
139 Supplementary Fig. 4a, b, g), but most of them were broken thus revealing an  
140 internal cavity and an ellipsoidal transverse section (Figs. 1a, b, h–k, 2a–e, g).  
141 The broken sclerites show an inner and outer phosphatic layer (thickness ca.  
142 20 to 50  $\mu\text{m}$ ) often separated by a thin empty space (Fig. 2g–l).

143 The convex side bears six prominent conical sclerites (S3) all with a rounded  
144 to elliptical well-delimited broad base, distributed in two longitudinal  
145 symmetrical pairs with two additional sclerites at both ends of the sagittal plane  
146 (Figs. 1, 2d, Supplementary Figs. 3e–h, 4e–i). A low-relief stripe runs in a  
147 sagittal position and vanishes towards the elevated end. It is topped by a row  
148 of aligned spines (S1, Fig. 1a); the one closer to the more elevated end being  
149 more tubular and longer. This row is flanked on both sides by smaller aligned  
150 spines (S2, Figs. 1a, d, h, 2a–c). Two relatively sinuous rows of six tiny spines  
151 are present parallel to the lateral margins (S4 and S5, Figs. 1b, e, h–j, 2d, e).

152 The convex side bears a polygonal micro-ornament (mesh size ca. 5  $\mu\text{m}$  wide,  
153 Fig. 2f, Supplementary Tables 1–3). However, its exact extension is uncertain  
154 due to coarse secondary phosphatization. Clusters of spherical phosphatized  
155 grains (diameter ca. 20  $\mu\text{m}$ ) occur near the sclerite base (Supplementary Fig.  
156 4b).

157 In most specimens, the flattened side is occupied by a relatively large opening  
158 (1200 and 600 $\mu\text{m}$  in maximal length and width, respectively) with irregularly  
159 defined margins (Fig. 1c, f, see also supplementary information movies 1, 2).  
160 The flattened side is often largely missing and opens into a spacious internal  
161 cavity with no signs of internal organs (e.g. gut and pharynx) (Fig. 1c, f). One  
162 specimen shows a tiny spine on the margin of the flattened side (Fig. 1f, g),  
163 which differs from other spiny sclerites (S1–S5).

164 The length of studied specimens ranges from 1.0 to 2.9 mm (Supplementary  
165 Fig. 3e–h). Whether growth was continuous or instead took place via  
166 successive moulting stages and cuticular renewal (ecdysis) could not be  
167 tested due to the small number of specimens (N=17) available for  
168 measurements. No major morphological variations (e.g. a sclerite pattern) can  
169 be seen between the smallest and largest specimens of *B. spinosa*  
170 (Supplementary Fig. 3e–h).

171

## 172 **Remarks**

### 173 **Body polarities in *Beretella***

174 The anterior-posterior (AP) and dorsal-ventral (DV) polarities of *Beretella* are  
175 uneasy to define because of the lack of modern equivalent among extant

176 animals. In the vast majority of extinct and extant invertebrates for which  
177 antero-posterior polarity is defined on the basis of independent criteria (e.g.  
178 position of mouth), sclerites point backwards (e.g. Cambrian scalidophoran  
179 worms (Han et al., 2007; Huang et al., 2004) and *Wiwaxia* (Zhang et al.,  
180 2015b)). This is most probably also the case with *Beretella* (Fig. 1a, d, j). The  
181 dorsoventral polarity of *Beretella* is supported by the fact that protective  
182 sclerites such as spines most commonly occur on the dorsal side of bilaterians  
183 (Fig. 1a, d, j).

184

### 185 **Comparison with *Saccorhytus* and other ecdysozoans**

186 *Beretella spinosa* has no exact equivalent in any Cambrian animals except  
187 *Saccorhytus coronarius*, an enigmatic, sac-like ecdysozoan (Han et al., 2017;  
188 Liu et al., 2022; Shu and Han, 2020b). Both forms share a tiny, poorly  
189 differentiated ellipsoidal body, and a set of prominent bilaterally arranged spiny  
190 sclerites. Indeed, the broad-based conical sclerites (S3) of *Beretella* are  
191 almost identical to those of *Saccorhytus* (Supplementary Fig. 4c) and have  
192 counterparts among scalidophoran worms (Supplementary Fig. 4d). However,  
193 they differ in number, ornamented structures, shape, and spatial arrangement  
194 (see details in Supplementary Tables 1–3) which makes the hypothesis of  
195 *Saccorhytus* being the larval stage of *Beretella* unlikely. Both *Beretella* and  
196 *Saccorhytus* differ from other known ecdysozoans in the lack of a vermiform  
197 body, introvert, annulations, and through gut (Supplementary Tables 1–3).

198

### 199 **Discussion**

200 **Ventral mouth.** All bilaterian animals have a digestive system with at least one  
201 opening that corresponds to the mouth (Brusca et al., 2016). Although the  
202 presumed oral area of *Beretella* is poorly preserved (ventral side often pushed  
203 in and largely destroyed), its mouth is likely to be found ventrally (see  
204 description), since no other opening occurs on its dorsal side, except those  
205 created by broken sclerites. The well-defined dorsoventral polarity of *Beretella*  
206 would suggest that the animal was resting on its ventral (flattened) side, the  
207 spiny dorsal side playing a protective role. Maintaining ventral contact with a  
208 substrate seems to be very unlikely unless these microscopic ellipsoidal  
209 animals were interstitial.

210

211 **Phylogenetic position of *Beretella*.** *Beretella*'s phylogenetic affinities remain  
212 elusive due to the lack of information concerning its internal anatomy and  
213 ventral side. Its scleritome consists of isolated conical sclerites that were the  
214 cuticular outgrowths of a seemingly rigid integument that covered both sides of  
215 the animal. Such conical sclerites have close counterparts in Cambrian  
216 ecdysozoans such as scalidophoran worms (e. g. *Eokinorhynchus* (Zhang et al.,  
217 2015a)), lobopodians (e.g. *Onychodictyon ferox* (Hou et al., 1991)) and even  
218 more clearly *Saccorhytus* that recent cladistic analyses resolved as a branch  
219 of the total-group Ecdysozoa (Liu et al., 2022). These sclerites unknown in

220 other animal groups, suggest that both *Saccorhytus* and *Beretella* belongs to  
221 Ecdysozoa in the absence of more direct fossil evidence such as exuviae or  
222 features suggesting cuticular moulting(Daley and Drage, 2016; Wang et al.,  
223 2019).

224 Cladistic analyses were performed to test the relation of *Beretella* and  
225 *Saccorhytus* to other ecdysozoan groups and, more generally, their  
226 phylogenetic relationships with other bilaterian groups (see details in  
227 Supplementary Table 4). Both taxa join in a clade (Saccorhytida, Fig. 3a–c)  
228 that is resolved as stem species within total-group Ecdysozoa and as the sister  
229 group of Cycloneuralia plus Panarthropoda, i.e. crown-group Ecdysozoa (Figs.  
230 3d, 4, Supplementary Figs. 6-9). These results are consistent with the body  
231 plan of Saccorhytida being markedly different from that of crown-group  
232 ecdysozoans that all have a vermiform body and differentiated structures such  
233 as the introvert and pharyngeal complex (Fig. 4).

234

### 235 **The ancestral ecdysozoan body plan**

236 Molecular clock analyses often place the divergence of Ecdysozoa relatively  
237 deep into the Ediacaran (Howard et al., 2022; Rota-Stabelli et al., 2013), thus  
238 highlighting major discrepancy with the known fossil record of the group.  
239 Potential ecdysozoans occur in the late Precambrian as suggested by 1)  
240 sclerites resembling those of extant priapulids, found in Ediacaran Small  
241 Carbonaceous Fossils assemblages(Moczyłowska et al., 2015) and 2)  
242 locomotion traces presumably made by scalidophoran worms (Buatois et al.,  
243 2014; Vannier et al., 2010). In the absence of fossil data for other vermiform  
244 groups such as nematoids, scalidophorans are potentially the oldest known  
245 representatives of Ecdysozoa. Recent Bayesian analyses based on a large  
246 molecular data set obtained from the 8 extant ecdysozoan phyla recover  
247 Scalidophora as the sister-group to Nematoida + Panarthropoda and suggest  
248 that ecdysozoans probably diverged in the Ediacaran possibly some 23 million  
249 years before the oldest fossil occurrence (trace fossils) of the group (Howard  
250 et al., 2022). Although this study does not speculate on the nature of the last  
251 common ancestor of Ecdysozoa, it is consistent with the view that the earliest  
252 representatives of the group were probably vermiform. Howard et al. (Howard  
253 et al., 2022) drew comparable conclusions based on *Acosmia*, an assumed  
254 stem-ecdysozoan worm from early Cambrian Chengjiang Lagerstätte.  
255 However, the re-evaluation of the morphological characteristics of this worm  
256 rather suggests a less basal position either within the total-group Cycloneuralia  
257 (Fig. 3d, Supplementary Figs. 6, 7) or among crown-group Ecdysozoa  
258 (Supplementary Figs. 8, 9). The non-vermiform nature of saccorhytids and  
259 their position as the sister group of the crown-group Ecdysozoa clearly  
260 reopens the debate on the nature of the ancestral ecdysozoan (Fig. 4) and  
261 suggests exploring various evolutionary hypotheses, in particular: 1) does the  
262 enigmatic saccorhytid body plan results from anatomical simplification? 2) to  
263 what extent may these animals shed light on the nature of the earliest

264 ecdysozoans?

265

### 266 **Do saccorhytids result from simplification?**

267 A relatively simple body plan and tiny size is often seen as resulting from  
268 anatomical simplification (e.g. digestive system) and miniaturization  
269 (micrometric size) in possible relation with the adaptation to specialized  
270 ecological niches or parasitism (Hanken and Wake, 1993). For example, some  
271 extant scalidophoran worms living in interstitial (meiobenthic) habitats such as  
272 loriciferans have a miniaturized body (Kirstensen, 1983) compared with their  
273 macroscopic counterparts (e.g. *Priapul*(Schmidt-Rhaesa, 2013b)). However,  
274 they retain a through gut and a functional introvert and show no sign of drastic  
275 internal simplification (Schmidt-Rhaesa, 2013a). Anatomical reduction is a  
276 typical feature of parasitism(Hanken and Wake, 1993) that is well-represented  
277 among extant ecdysozoans such as nematodes(Schmidt-Rhaesa, 2014).  
278 Although relatively small (ca. 0.1-2.5 mm long), nematodes underwent no  
279 simplification of their digestive system. Saccorhytids have no specialized  
280 features (e.g. anchoring or piercing structures) that would point to any  
281 adaptation to ecto- or endo-parasitic lifestyles(Cong et al., 2017). *Saccorhytus*  
282 has been interpreted<sup>16</sup> as a possible interstitial animal based on its micrometric  
283 size which corresponds to that the of the extant meiofauna. If we accept the  
284 hypothesis of saccorhytids resulting from simplification, then we need to  
285 determine its origin. Simplification of saccorhytids from a vermiform animal  
286 (e.g. cycloneuralian worm with a through gut and terminal mouth) is difficult to  
287 conceive because it would involve considerable anatomical transformations  
288 such as the loss of vermiform organization, introvert and pharynx in addition to  
289 that of the digestive system (Fig. 4). Alternative options to consider are  
290 ancestral and not necessarily vermiform ecdysozoans.

291

### 292 **Early evolution of ecdysozoans: a new scenario**

293 We propose here an alternative evolutionary hypothesis (Fig. 4) in which  
294 saccorhytids are replaced within the broader framework of the origin and early  
295 diversification of moulting animals. Saccorhytids are seen as an early off-shot  
296 from the stem-line Ecdysozoa (see cladistic analysis above) that possibly  
297 retained important features of the body plan of ancestral non-vermiform  
298 ecdysozoans (see ancestral character state reconstruction in Supplementary  
299 Table 4). This scenario must be considered as a working hypothesis whose  
300 aim is to stimulate research in this key area of animal evolution.

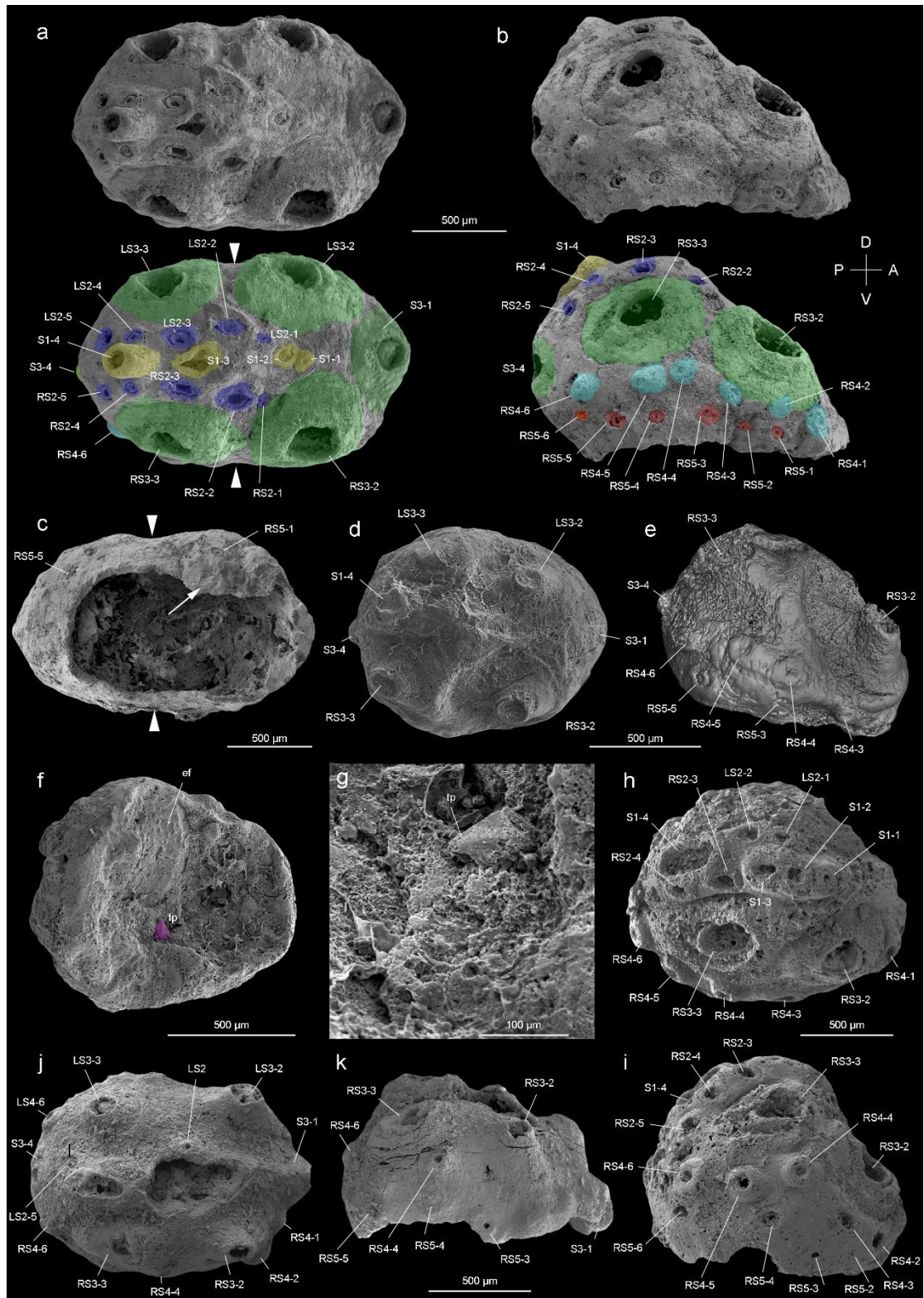
301 The cuticular secretion and the loss of cilia (Valentine and Collins, 2000) are  
302 seen as the first of a series of evolutionary events (Fig. 4) that led to the rise of  
303 Ecdysozoa. Moulting (shedding of the old cuticle via apolysis and its renewal)  
304 further reconciled body growth and cuticular protection (Schmidt-Rhaesa,  
305 2007). Cuticle secretion and moulting may have been quasi-simultaneous  
306 innovations that took place over a relatively short time interval. The nature of  
307 the very first ecdysozoans is hypothetical and lacks fossil evidence. However,

308 they are tentatively represented here as small epibenthic or interstitial  
309 slow-moving non-vermiform animals from which saccorhytids may have  
310 evolved via an assumed anatomical simplification (i.e. loss of anus seen  
311 details in Supplementary Table 4, Fig. 4).

312 In our scenario, this ancestral ecdysozoan stock would have also given rise to  
313 vermiform ecdysozoans through stepwise anatomical transformations such as  
314 the body elongation, the differentiation of key morpho-functional structures  
315 such as the pharynx and the introvert and the shift of the ventral mouth to a  
316 terminal position (Martín-Durán and Hejnol, 2015) (Fig. 4). This mouth shift  
317 from ventral to terminal arising in crown ecdysozoans is consistent with the  
318 chronology of divergence of animal lineages and the fact that the mouth of  
319 most spiralian is ventral (Martín-Durán and Hejnol, 2015; Nielsen, 2019;  
320 Ortega-Hernandez et al., 2019). Developmental studies show that embryos of  
321 extant cycloneuralians have a ventral mouth that moves to a terminal position  
322 towards the adult stage (Martín-Durán and Hejnol, 2015; Nielsen, 2019).  
323 These assumed major anatomical changes (e.g. functional introvert) must be  
324 placed in the ecological context of Cambrian animal radiation. Important  
325 changes in the functioning of marine ecosystems occurred in the early  
326 Cambrian such as interactive relationships between animal species,  
327 exemplified by predation (Vannier and Chen, 2005; Vermeij, 1977) may have  
328 acted as drivers in the evolution of early ecdysozoans, in promoting burrowing  
329 into sediment and the colonization of endobenthic habitats for the first time  
330 (Vannier et al., 2010). Burrowing into the sediment could be seen as the  
331 evolutionary response of epibenthic animals such as ancestral ecdysozoans to  
332 escape visual predation (Daley et al., 2013; Vannier and Chen, 2005). This  
333 migration to endobenthic shelters was made possible by the development of a  
334 resistant cuticular layer (Fig. 4) that strongly reduced physical damage caused  
335 by friction with the sediment and provided anchoring points (e.g. scalids and  
336 sclerites). Whereas saccorhytids became rapidly extinct during the Cambrian,  
337 worms massively colonized endobenthic habitats, resulting in bioturbation and  
338 ecological turnover.

339

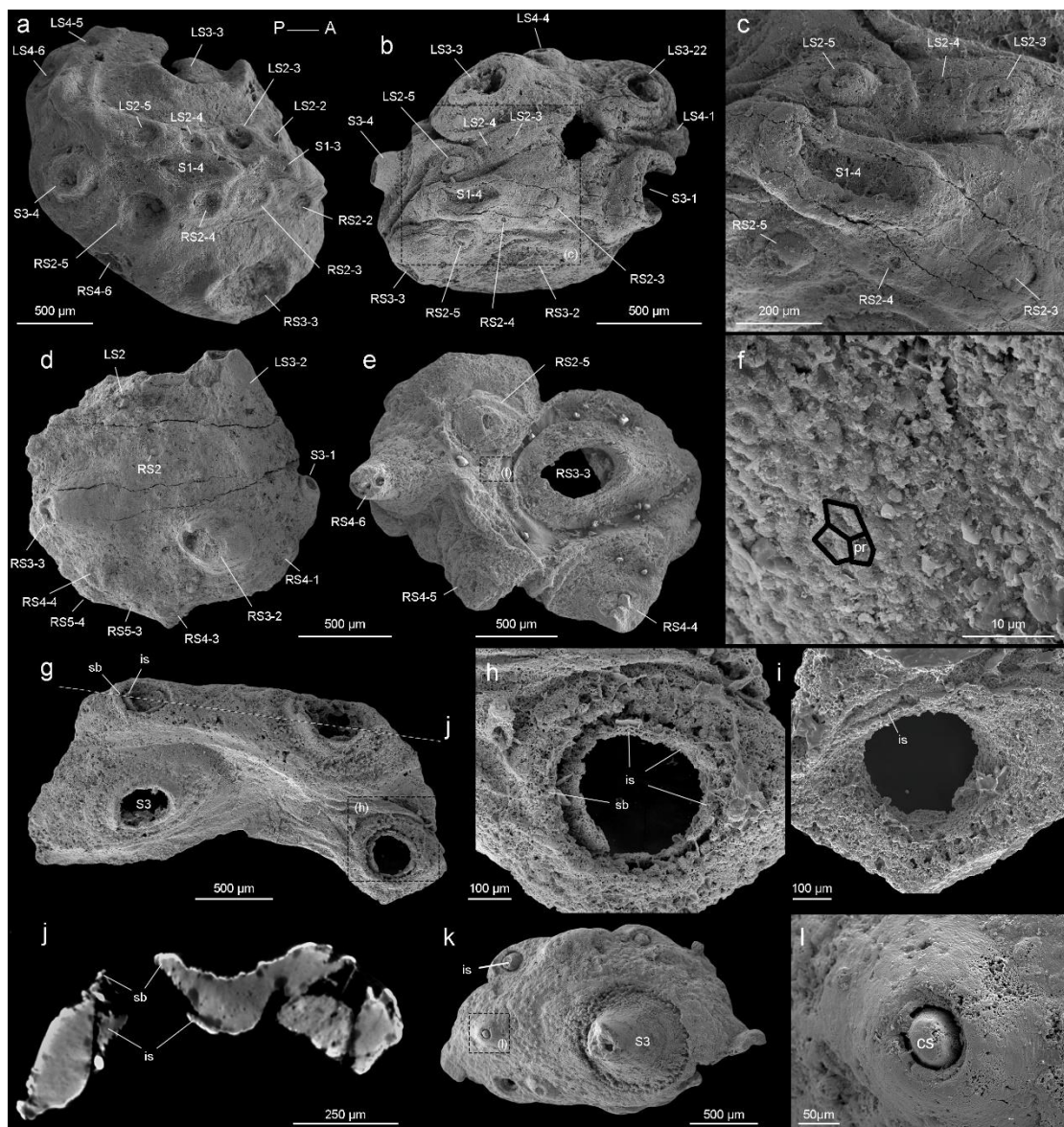




340

341 **Fig. 1. *Beretella spinosa* gen. et sp. nov. from Member 5 of the Yanjiahe**  
 342 **Formation (Cambrian Stage 2), Yichang, Hubei Province, China. a–c,**  
 343 **Holotype, CUBar138-12. a, Dorsal view showing the external ornament: (five**  
 344 **sclerites at the midline in yellow (S1); flanked by two rows of sclerites in blue**  
 345 **(S2); large broad-based conical sclerites in two dorsolateral pairs and one**  
 346 **antero-posterior pairs in green (S3)); white arrows indicate lateral constriction.**

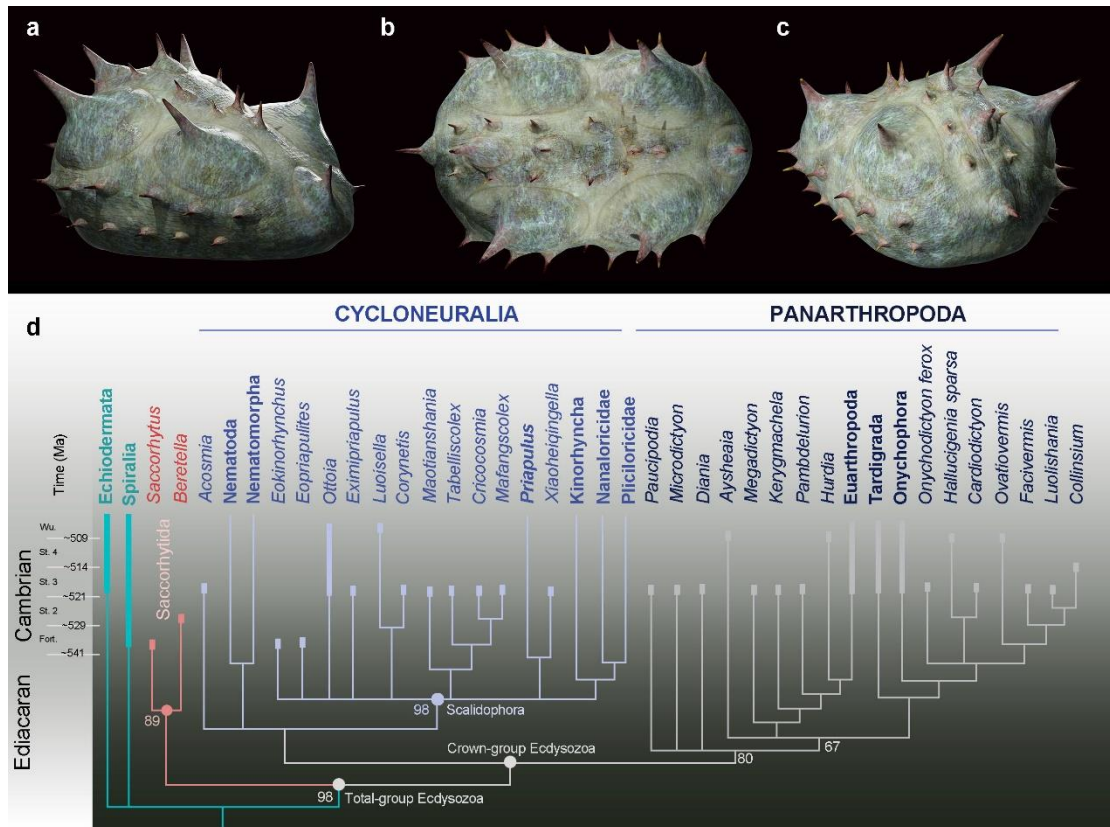
347 **b**, Right lateral view showing two additional rows of six sclerites (S4 and S5, in  
 348 light blue and pink, respectively). **c**, Ventral view showing a large opening that  
 349 may have accommodated the mouth (see the text) and an empty body cavity.  
 350 **d–g**, CUBar75-45. **d**, Dorsal view showing a broken S3. **e**, Micro-CT image,  
 351 right lateral view displaying S4. **f**, Ventral view depicting a tiny projection in  
 352 purple. **g**, An enlargement of the projection of **f**. **h–i**, Paratype, CUBar171-5. **h**,  
 353 Right dorsal view showing S1–S4. **i**, Right-lateral view showing S4 and S5. **j–k**,  
 354 Paratype CUBar121-8. **j**, Dorsal view showing poorly preserved S1 and S2. **k**,  
 355 Right-lateral view showing S3–S5. A, assumed anterior end (see text); ef,  
 356 exotic fragment; D, assumed dorsal side; L, left; P, posterior end; R, right; tp,  
 357 tiny spine; V, ventral side. The same abbreviations are used throughout the  
 358 manuscript including Supplementary materials.  
 359



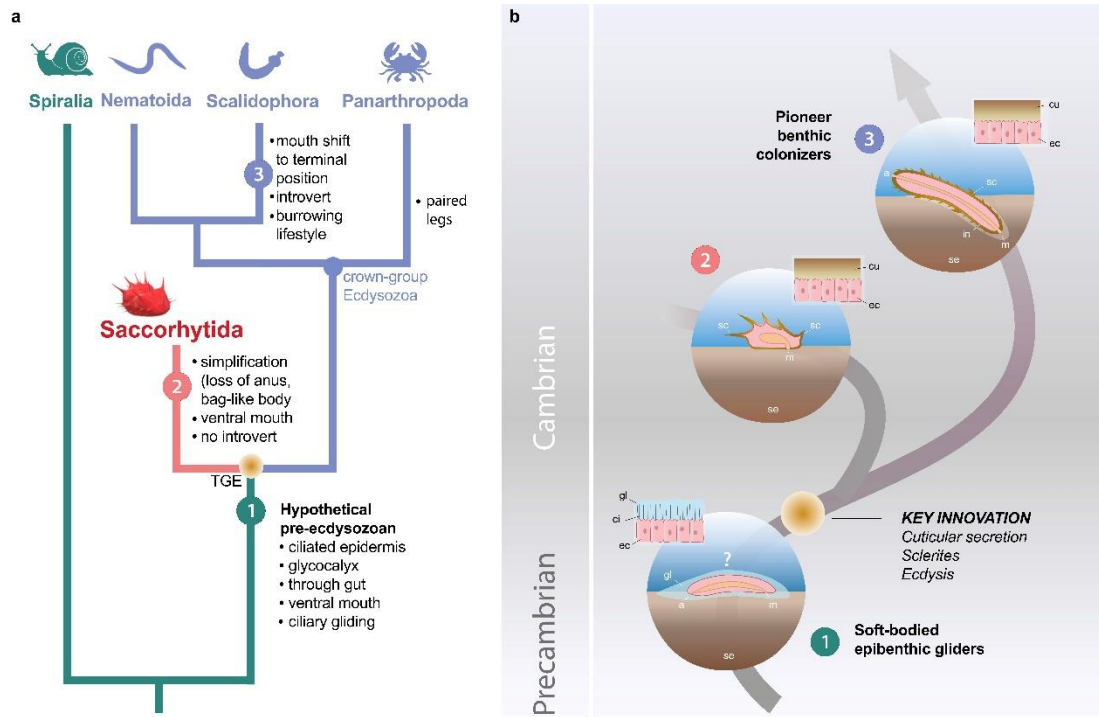
360

361 **Fig. 2. *Beretella spinosa* gen. et sp. nov.** a, CUBar99-19, dorsal view  
 362 showing an ornament S1–S4. **b–c**, CUBar136-9, general dorsal view and

363 details. **d**, CUBar136-11, dorsal view showing S1–S5. **e–f**, CUBar73-15  
 364 general view and details of the cuticular polygonal reticulation in black. **g–j**,  
 365 CUBar128-27. **g–i**, general view and details of the bi-layered structure of the  
 366 cuticular wall as seen in broken conical sclerites. **j**, Micro-CT section showing  
 367 possibly sclerite infilling. **k–l**, CUBar99-18, cuticular fragment, general view and  
 368 details of large sclerite (central feature represents possible phosphatic infilling).  
 369 is, infilling sclerite; pr, polygonal reticulation; sb, sclerite base.  
 370  
 371



372  
 373 **Fig. 3. Position of *Beretella spinosa* in the animal tree based on cladistic**  
 374 **analysis.** **a–c**, artistic three-dimensional reconstructions of *Beretella spinosa*  
 375 in the anterolateral, dorsal, and posterolateral views. **d**, Animal tree obtained  
 376 from cladistic analyses using maximum likelihood tree obtained from cladistic  
 377 analyses using maximal likelihood (IQTREE). *Saccorhytus* and *Beretella* join in  
 378 a clade (new phylum Saccorhytida) resolved as the sister-group of all other  
 379 ecdysozoans; numbers at key nodes denote probability. Fossil and extant taxa  
 380 are in italics and bold, respectively. Known fossil record indicated by thicker  
 381 bars (after (Shu and Han, 2020a)).



382

383 **Fig. 4. Possible evolutionary scenario to explain the origin and early**  
 384 **evolution of ecdysozoans. a**, Summary tree (see Supplementary Figs. 6-9)  
 385 showing saccorhytids as a sister-group of Cycloneuralia (Nematoida plus  
 386 Scalidophora) + Panarthropoda; main morphological features of each group  
 387 listed along each branch. **b**, Potential evolutionary pathway to evolve  
 388 Saccorhytida and crown-group Ecdysozoa. Numbers in green, red and blue  
 389 circles designate pre-ecdysozoan (Spiralia), Saccorhytida and Cycloneuralia,  
 390 respectively. Light brown gradient (circle) to emphasize ecdysis and sclerite  
 391 secretion seen as key evolutionary steps. 1, Hypothetical pre-ecdysozoan  
 392 animal with a ciliated epidermis and glycocalyx. 2, Saccorhytid exemplified by  
 393 *Beretella* with a cuticle bearing sclerites and a simplified internal organization  
 394 (e.g. loss of anus). 3, Crown-group ecdysozoan exemplified by a  
 395 scalidophoran worm with an elongated shape, a differentiated head (introvert)  
 396 and trunk, sclerites, a through gut, a terminal mouth and abilities to burrow into  
 397 bottom sediment. Animals not to scale. Abbreviations: a, anus; ci, cilia; cu,  
 398 cuticle; ec, epidermal cell; gl, glycocalyx (mucous layer); m, mouth; in, introvert;  
 399 sc, sclerite; se, sediment; TGE, total-group Ecdysozoa. Silhouettes from  
 400 phylopic.org.

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## 409 **Methods**

### 410 **Material**

411 Fourteen specimens of *Beretella spinosa* were recovered from samples  
412 (siliceous-phosphatic, intraclastic limestone) collected from Member 5 of the  
413 Yanjiahe Formation, Yanjiahe section near Yichang City, Hubei Province,  
414 China (Guo et al., 2021) (Supplementary Tables 1–3). These were obtained by  
415 digesting the rocks in 10% acetic acid. Faunal elements associated with  
416 *Beretella spinosa* (Supplementary Tables 1–3) in residues are mainly tiny  
417 molluscs (CUBar21-4 and CUBar206-6). Comparisons were made with 10  
418 specimens of *Saccorhytus coronarius* (ELIXX25-62, ELIXX34-298,  
419 ELIXX45-20, ELIXX48-64, ELIXX58-336, ELIXX61-27, ELIXX65-116,  
420 ELIXX65-296, ELIXX99-420) and one coeval scalidophoran specimen  
421 (ELIXX57-320) all from Bed 2 of the Kuanchuanpu Formation, Zhangjiagou  
422 section near Xixiang County, south Shaanxi Province, China. All specimens of  
423 *Beretella* are deposited in the paleontological collections of Chang'an  
424 University, Xi'an (CU), those of scalidophoran, and *Saccorhytus* at Northwest  
425 University, Xi'an (ELI), China.

426

### 427 **Scanning electron microscopy (SEM)**

428 All specimens were coated with gold and then imaged using a FEI Quanta 400  
429 FEG SEM at Northwest University and a FEI Quanta 650 at Chang'an  
430 University.

431

### 432 **X-ray computed microtomography and 3D reconstruction**

433 Micro-CT-images (tiff format, with pixel size 1.1  $\mu\text{m}$ ) of *Beretella* (CUBar75-45,  
434 CUBar128-27, CUBar138-12) and *Saccorhytus* (ELIXX65-116, ELIXX99-420)  
435 were acquired using the Zeiss Xradia 520 at Northwest University (NWU),  
436 Xi'an, China, at an accelerating voltage of 50 kV and a beam current of 80  $\mu\text{A}$ .  
437 Micro-CT data were processed using VGStudio Max 3.2 for 3D volume  
438 rendering.

439

### 440 **Measurements**

441 Measurements of the length, width, and height of *Beretella* and *Saccorhytus*  
442 were obtained from Micro-CT and SEM images by using tipDig2 v.2.16.

443

### 444 **Phylogenetic analysis**

445 We built our matrix with 55 taxa coded using 191 morphological characteristics  
446 (Supplementary Texts 1, 2). It is largely based on the data published by  
447 Howard et al. (Howard et al., 2020), Vinther and Parry (Vinther and Parry, 2019)  
448 and Ou et al. (Ou et al., 2017), although emended and supplemented by recent  
449 updates and new observations (Supplementary Text 1). Three characters (37.  
450 Through gut, 38. U-shaped gut, and 40. Ventral mouth) in matrix were coded  
451 as “? (uncertain)”, “?”, and “?”, respectively. Because although we can infer a

452 ventral mouth and no anus of *Beretella*, these anatomic structures are invisible  
453 in fossils. We analyzed the data matrix using parsimony (TNT), likelihood  
454 (IQTREE) and Bayesian inference (MrBayes). Parsimony analysis was  
455 implemented in TNT under equal and implied (k=3) weight. Parameters are  
456 default (Goloboff et al., 2008; Goloboff and Catalano, 2016). The  
457 maximum-likelihood tree search was conducted in IQ-TREE(Nguyen et al.,  
458 2015), and support was assessed using the ultrafast phylogenetic bootstrap  
459 replication method (Hoang et al., 2018; Minh et al., 2013) to run 50,000  
460 replicates. Bayesian inference was conducted in with MrBayes v3.2.6a with  
461 default priors and Markov chain Monte Carlo settings(Ronquist et al., 2012).  
462 Two independent runs of 7,000,000 Markov chain Monte Carlo generations  
463 were performed, each containing four Markov chains under the Mkv +  $\Gamma$  model  
464 for the discrete morphological character data(Lewis, 2001). In each run (N=2),  
465 trees were collected at a sampling frequency of every 5,000 generations and  
466 with the first 25% samples discarded as burn-in. The convergence of chains  
467 was checked by effective sample size (ESS) values over 1,000 in Tracer  
468 v.1.7(Rambaut et al., 2018), 1.0 for the potential scale reduction factor  
469 (PSRF)(Gelman and Rubin, 1992), and by an average standard deviation of  
470 split frequencies below 0.007.

471

#### 472 **Ancestral character state reconstructions**

473 Ancestral character state reconstructions for six morphological characters  
474 were performed on the ecdysozoan total group node, the ecdysozoan crown  
475 group node and saccorhytid node. Characters selected for ancestral state  
476 reconstruction represent traits inferred as ecdysozoan plesiomorphies  
477 (ancestral characters) from studies of crown group taxa. These characters  
478 included the presence or absence of: (1) through gut; (2) ventral mouth; (3)  
479 introvert (see Supplementary Table 4).

480 This was carried out individually for the selected character in MrBayes. This  
481 was employed to calculate the posterior probability of the presence (1) and  
482 absence (0) of the selected characters at the selected nodes. Analyses used  
483 the MK + gamma model, and always converged after 2 million generations.  
484 Average deviation of split frequencies (< 0.01), ESS scores (> 200), and PSRF  
485 values (= approx. 1.00) assessed convergence of the MCMC chains (Howard  
486 et al., 2020).

487

#### 488 **Data availability**

489 The data that support the findings of this study are available in the recent  
490 paper and its Supplementary Information.

491

#### 492 **References and Notes**

493

494 Aguinaldo, A.M.A., Turbeville, J.M., Linford, L.S., Rivera, M.C., Garey, J.R.,  
495 Raff, R.A. and Lake, J.A., 1997. Evidence for a clade of nematodes,

- 496 arthropods and other moulting animals. *Nature*, 387(6632): 489-93.
- 497 Brusca, R.C., Moore, W. and Shuster, S.M., 2016. *Invertebrates*. Sinauer  
498 Associates, Inc., Sunderland Massachusetts USA, 639-910 pp.
- 499 Buatois, L.A., Narbonne, G.M., Mangano, M.G., Carmona, N.B. and Myrow, P.,  
500 2014. Ediacaran matground ecology persisted into the earliest  
501 Cambrian. *Nat Commun*, 5: 3544.
- 502 Budd, G.E., 2001. Why are arthropods segmented? *Evolution & Development*,  
503 3(5): 332–342.
- 504 Cong, P., Ma, X., Williams, M., Siveter, D.J., Siveter, D.J., Gabbott, S.E., Zhai,  
505 D., Goral, T., Edgecombe, G.D. and Hou, X., 2017. Host-specific  
506 infestation in early Cambrian worms. *Nature Ecology & Evolution*, 1(10):  
507 1465-1469.
- 508 Daley, A.C. and Drage, H.B., 2016. The fossil record of ecdysis, and trends in  
509 the moulting behaviour of trilobites. *Arthropod Structure & Development*,  
510 45(2): 71-96.
- 511 Daley, A.C., Paterson, J.R., Edgecombe, G.D., García-Bellido, D.C., Jago, J.B.  
512 and Donoghue, P., 2013. New anatomical information on *Anomalocaris*  
513 from the Cambrian Emu Bay Shale of South Australia and a  
514 reassessment of its inferred predatory habits. *Palaeontology*, 56:  
515 971-990.
- 516 Erwin, D.H., 2020. The origin of animal body plans: a view from fossil evidence  
517 and the regulatory genome. *Development*, 147(4): dev182899.
- 518 Gelman, A. and Rubin, D.B., 1992. Inference from iterative simulation using  
519 multiple sequences (with discussion). *Statistical Science*, 7: 457-472.
- 520 Goloboff, P.A., Carpenter, J.M., Arias, J.S. and Esquivel, D.R.M., 2008.  
521 Weighting against homoplasy improves phylogenetic analysis of  
522 morphological data sets. *Cladistics*, 24(5): 758-773.
- 523 Goloboff, P.A. and Catalano, S.A., 2016. TNT version 1.5, including a full  
524 implementation of phylogenetic morphometrics. *Cladistics*, 32: 221-238.
- 525 Guo, J.-F., Li, G.-X., Qiang, Y.-Q., Song, Z.-C., Zhang, Z.-F., Han, J. and Wang,  
526 W.-Z., 2021. *Watsonella crosbyi* from the lower Cambrian (Terreneuvian,  
527 Stage 2) Yanjiahe Formation in Three Gorges Area, South China.  
528 *Palaeoworld*, 30(1): 1-19.
- 529 Han, J., Conway Morris, S., Ou, Q., Shu, D.G. and Huang, H., 2017.  
530 Meiofaunal deuterostomes from the basal Cambrian of Shaanxi (China).  
531 *Nature*, 542(7640): 228-231.
- 532 Han, J., Liu, J.N., Zhang, Z.F., Zhang, X.L. and Shu, D.G., 2007. Trunk  
533 ornament on the palaeoscolecid worms *Cricocosmia* and *Tabelliscolex*  
534 from the Early Cambrian Chengjiang deposits of China. *Acta*  
535 *Palaeontologica Polonica*, 52(2): 423–431.
- 536 Hanken, J. and Wake, D.B., 1993. Miniaturization of body size: Organismal  
537 consequences and evolutionary significance. *Annual Review of Ecology*  
538 *and Systematics*, 24: 501-519.
- 539 Hoang, D.T., Chernomor, O., von Haeseler, A., Minh, B.Q. and Vinh, L.S., 2018.

- 540 UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Mol Biol*  
541 *Evol*, 35(2): 518-522.
- 542 Hou, X.G., Ramskold, L. and Bergstrom, J., 1991. Composition and  
543 preservation of the Chengjiang fauna -a Lower Cambrian soft-bodied  
544 biota. *Zoologica Scripta*, 20(4): 395-411.
- 545 Howard, R.J., Edgecombe, G.D., Shi, X., Hou, X. and Ma, X., 2020. Ancestral  
546 morphology of Ecdysozoa constrained by an early Cambrian stem  
547 group ecdysozoan. *BMC Evolutionary Biology*, 20(1): 156.
- 548 Howard, R.J., Giacomelli, M., Lozano-Fernandez, J., Edgecombe, G.D.,  
549 Fleming, J.F., Kristensen, R.M., Ma, X., Olesen, J., Sørensen, M.V.,  
550 Thomsen, P.F., Wills, M.A., Donoghue, P.C.J. and Pisani, D., 2022. The  
551 Ediacaran origin of Ecdysozoa: integrating fossil and phylogenomic  
552 data. *Journal of the Geological Society*, 179: jgs2021-107.
- 553 Huang, D., Vannier, J. and Chen, J., 2004. Anatomy and lifestyles of Early  
554 Cambrian priapulid worms exemplified by *Corynetis* and *Anningvermis*  
555 from the Maotianshan Shale (SW China). *Lethaia*, 37(1): 21-33.
- 556 Kirstensen, R.M., 1983. Loricifera, a new phylum with Aschelminthes  
557 characters from the meiobenthos. *Zeitschrift für zoologische Systematik*  
558 *und Evolutionsforschung*, 21(3): 163-180.
- 559 Laumer, C.E., Fernandez, R., Lemer, S., Combosch, D., Kocot, K.M., Riesgo,  
560 A., Andrade, S.C.S., Sterrer, W., Sorensen, M.V. and Giribet, G., 2019.  
561 Revisiting metazoan phylogeny with genomic sampling of all phyla.  
562 *Proceedings of the Royal Society B*, 286(1906): 20190831.
- 563 Lewis, P.O., 2001. A likelihood approach to estimating phylogeny from discrete  
564 morphological character data. *Systematic biology*, 50(6): 913-925.
- 565 Liu, A.G., Matthews, J.J., Menon, L.R., Mcllroy, D. and Brasier, M.D., 2014.  
566 *Hootia quadriformis* n. gen., n. sp., interpreted as a muscular cnidarian  
567 impression from the Late Ediacaran period (approx. 560 Ma). *Proc Biol*  
568 *Sci*, 281(1793).
- 569 Liu, Y., Carlisle, E., Zhang, H., Yang, B., Steiner, M., Shao, T., Duan, B.,  
570 Marone, F., Xiao, S. and Donoghue, P.C.J., 2022. *Saccorhytus* is an  
571 early ecdysozoan and not the earliest deuterostome. *Nature*, 609:  
572 541-546.
- 573 Martín-Durán, J.M. and Hejnol, A., 2015. The study of *Priapulid caudatus*  
574 reveals conserved molecular patterning underlying different gut  
575 morphogenesis in the Ecdysozoa. *BMC Biology*, 13: 29.
- 576 Minh, B.Q., Nguyen, M.A. and von Haeseler, A., 2013. Ultrafast approximation  
577 for phylogenetic bootstrap. *Mol Biol Evol*, 30(5): 1188-95.
- 578 Moczyłowska, M., Budd, G.E. and Agić, H., 2015. Ecdysozoan-like sclerites  
579 among Ediacaran microfossils. *Geological Magazine*, 152(06):  
580 1145-1148.
- 581 Nguyen, L.T., Schmidt, H.A., Von Haeseler, A. and Minh, B.Q., 2015. IQ-TREE:  
582 a fast and effective stochastic algorithm for estimating  
583 maximum-likelihood phylogenies. *Molecular biology and evolution*,



- 584 32(1): 268-274.
- 585 Nielsen, C., 2019. Was the ancestral panarthropod mouth ventral or terminal?  
586 *Arthropod Structure & Development*, 49: 152-154.
- 587 Ortega-Hernandez, J., Janssen, R. and Budd, G.E., 2019. The last common  
588 ancestor of Ecdysozoa had an adult terminal mouth. *Arthropod*  
589 *Structure & Development*, 49: 155-158.
- 590 Ou, Q., Han, J., Zhang, Z., Shu, D., Sun, G. and Mayer, G., 2017. Three  
591 Cambrian fossils assembled into an extinct body plan of cnidarian  
592 affinity. *Proceedings of the National Academy of Science*, 114(33):  
593 8835-8840.
- 594 Rambaut, A., Drummond, A., Xie, D., Baele, G. and Suchard, M., 2018.  
595 Posterior summarisation in Bayesian phylogenetics using Tracer 1.7.  
596 *Systematic Biology*, 67(5): 901-904.
- 597 Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Höhna, S.,  
598 Larget, B., Liu, L., Suchard, M.A. and Huelsenbeck, J.P., 2012.  
599 MrBayes 3.2: efficient Bayesian phylogenetic inference and model  
600 choice across a large model space. *Systematic Biology*, 61(3): 539-42.
- 601 Rota-Stabelli, O., Daley, A.C. and Pisani, D., 2013. Molecular timetrees reveal  
602 a Cambrian colonization of land and a new scenario for ecdysozoan  
603 evolution. *Current Biology*, 23(5): 392-8.
- 604 Sawaki, Y., Nishizawa, M., Suo, T., Komiya, T., Hirata, T., Takahata, N., Sano,  
605 Y., Han, J., Kon, Y. and Maruyama, S., 2008. Internal structures and U–  
606 Pb ages of zircons from a tuff layer in the Meishucunian formation,  
607 Yunnan Province, South China. *Gondwana Research*, 14(1-2):  
608 148-158.
- 609 Schmidt-Rhaesa, A., 2007. *The evolution of organ systems*. Oxford university  
610 press, 54-73 pp.
- 611 Schmidt-Rhaesa, A., 2013a. Nematomorpha. In: A. Schmidt-Rhaesa (Editor),  
612 *Gastrotricha, Cycloneuralia and Gnathifera*. De Gruyter, Germany, pp.  
613 29-146.
- 614 Schmidt-Rhaesa, A., 2013b. Priapulida. In: A. Schmidt-Rhaesa (Editor),  
615 *Gastrotricha, Cycloneuralia and Gnathifera*. De Gruyter, Germany, pp.  
616 147-180.
- 617 Schmidt-Rhaesa, A., 2014. *Handbook of Zoology. Gastrotricha, Cycloneuralia*  
618 *and Gnathifera*, 2. De Gruyter, Germany, 5-12 pp.
- 619 Shu, D.G. and Han, J., 2020a. The core value of Chengjiang fauna: the  
620 formation of the animal kingdom and the birth of basic human organs.  
621 *Earth Science Frontiers*, 27: 382-412.
- 622 Shu, D.G. and Han, J., 2020b. The core value of Chengjiang fauna: the  
623 information of animal kingdom and the birth basic human organs. *Earth*  
624 *Science Frontiers*, 27: 1-32.
- 625 Valentine, J.W. and Collins, A.G., 2000. The significance of moulting in  
626 ecdysozoan evolution. *Evolution & Development*, 2(3): 152–156.
- 627 Vannier, J., Calandra, I., Gaillard, C. and Żylińska, A., 2010. Priapulid worms:

- 628 Pioneer horizontal burrowers at the Precambrian-Cambrian boundary.  
629 *Geology*, 38(8): 711-714.
- 630 Vannier, J. and Chen, J., 2005. Early Cambrian Food Chain: New Evidence  
631 from Fossil Aggregates in the Maotianshan Shale Biota, SW China.  
632 *Palaios*, 20(1): 3-26.
- 633 Vermeij, G.J., 1977. The Mesozoic marine revolution: evidence from snails,  
634 predators and grazer. *Paleobiology*, 3: 245-258.
- 635 Vinther, J. and Parry, L.A., 2019. Bilateral jaw elements in *Amiskwia*  
636 *sagittiformis* bridge the morphological gap between gnathiferans and  
637 chaetognaths. *Current Biology*, 29(5): 881-888 e1.
- 638 Wang, D., Vannier, J., Schumann, I., Wang, X., Yang, X.G., Komiya, T., Uesugi,  
639 K., Sun, J. and Han, J., 2019. Origin of ecdysis: fossil evidence from  
640 535-million-year-old scalidophoran worms. *Proceedings of the Royal*  
641 *Society B*, 286(1906): 20190791.
- 642 Zhang, H., Xiao, S., Liu, Y., Yuan, X., Wan, B., Muscente, A.D., Shao, T., Gong,  
643 H. and Cao, G., 2015a. Armored kinorhynch-like scalidophoran animals  
644 from the early Cambrian. *Scientific Reports*, 5: 16521.
- 645 Zhang, Z., Smith, M.R. and Shu, D., 2015b. New reconstruction of the *Wiwaxia*  
646 scleritome, with data from Chengjiang juveniles. *Sci Rep*, 5: 14810.

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661 prepared all the specimens, photographs, figures except Figure 4 (J.V.). J.S.  
662 performed the analysis by Micro-CT and visualization with Micro-CT data. D.W.  
663 and J.H. performed phylogenetic analyses. Y.Y. performed morphospace  
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668

669 **Competing interests:** The authors declare no competing interests.

670

671 **Additional information**

672 **Supplementary information** The online version contains supplementary  
673 material (tomographic data of *Beretella* and *Saccorhytus*, and movies of  
674 3D-animation of the holotype of *Beretella*) available at  
675 <https://figshare.com/s/054f31fc22567a590d7f>.

676

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680