

1 **Molecular phylogeny and diversification timing of the Nemouridae family (Insecta,**  
2 **Plecoptera) in the Japanese Archipelago**

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13 Short Title: Molecular phylogeny and diversification timing of Nemouridae

14

15 **Abstract**

16 The generation of the high species diversity of insects in Japan was profoundly influenced by the  
17 formation of the Japanese Archipelago. We explored the species diversification and biogeographical  
18 history of the Nemouridae family in the Japanese Archipelago using mitochondrial DNA and nuclear  
19 DNA markers. We collected 49 species among four genera: *Indonemoura*, *Protonemura*,  
20 *Amphinemura* and *Nemoura* in Japan, China, South Korea and North America. We estimated their  
21 divergence times—based on three molecular clock node calibrations—using Bayesian phylogeography  
22 approaches. Our results suggested that Japanese Archipelago formation events resulted in  
23 diversification events in the middle of the Cretaceous (<120 Ma), speciation in the Paleogene (<50 Ma)  
24 and intra-species diversification segregated into eastern and western Japan of the Fossa Magna region  
25 at late Neogene (20 Ma). The *Indonemoura* samples were genetically separated into two clades—that  
26 of Mainland China and that of Japan. The Japanese clade clustered with the Nemouridae species from  
27 North America, suggesting the possibility of a colonisation event prior to the formation of the Japanese  
28 Archipelago. We believe that our results enhanced the understanding both of the origin of the species  
29 and of local species distribution in the Japanese Archipelago.

30

31 Keywords: stoneflies, Japan, divergence time, molecular phylogeny, speciation

32

## 33 Introduction

34 The East Asian region—and in particular, the Japanese Archipelago—is considered to have  
35 high insect biodiversity [1], [2]. The high degree of Japanese insect biodiversity is a result of several  
36 mechanisms—in particular, the complex geological history. The Japanese Archipelago originated in  
37 the middle of the Miocene [3] as an independent formation of eastern and western Japanese  
38 landmasses. Extensive geographical changes and large-scale climatic changes throughout the islands  
39 facilitated the subsequent connection and disconnection of Japanese landmasses from the Eurasian  
40 continent, and the formation of tectonic lines (as the median tectonic line, MTL; and the Itoigawa-  
41 Shizuoka tectonic line, ISLT) [3], [4], [5]. These geological events—allowing for the colonisation of  
42 insects from the continent and their subsequent diversification as endemic lineages (i.e. new  
43 species)—contributed substantially to the high diversity of insects in Japan [2].

44 The process of species diversification has been intensively explored through  
45 phylogeographical approaches [6], [7]. These approaches have allowed for the observation of the  
46 historical process responsible for the current geographical distribution of individuals [6]. Molecular  
47 approaches to phylogeographic studies, using specific genes—such as mitochondrial DNA (mtDNA) or  
48 nuclear DNA (nDNA)—allow for a better understanding of species diversity by resolving complex  
49 taxonomic groups of species (for instance, cryptic species and species groups) [7]. Molecular  
50 phylogeography has provided valuable insights into the historical process of Japanese Archipelago  
51 formation underlying insect diversification. Previous studies identified genetic differentiation within  
52 species between the Japanese landmasses and the Eurasian continent (for instance, the mayflies  
53 *Isonychia japonica* [8]; caddisflies *Palaeagapetus* spp. [9]; and beetles *Ohomopterus* spp. [10] and the  
54 Carabina subtribe [11]). Dispersal events via land bridges (islands between continents) from the  
55 Eurasian continent to the Japanese Archipelago (of, for instance, the orthopteran *Locusta migratoria*,  
56 [12]; mayflies *Ephron* spp., [13]) or, in reverse, from the Japanese Archipelago to the Eurasian  
57 continent (of, for instance, water bugs *Appasus* spp., [14]) were additionally identified before, during  
58 and after the formation of the Japan Archipelago.

59 Aquatic insects have advantages in the studies of phylogeography, as their specialised  
60 ecological requirements and habitat range make aquatic insect species susceptible to geological  
61 changes. Among the Plecoptera order [15], the family Nemouridae is one of the largest and most  
62 dominant aquatic insect groups. The family comprises 20 genera and more than 400 species  
63 distributed throughout the Northern Hemisphere and across the equator in the Sunda Archipelago  
64 [16]. Several genera of the Nemouridae family have distinct disjunctions in their distribution [15]. For  
65 example, *Ostrocerca*, *Prostoia* and *Soyedina* were found in both the extreme western and the extreme  
66 eastern regions of North America, but they were absent in the central area [17], [18]. Similar

67 disjunctive distributions were also observed among *Protonemura*, *Indonemoura*, *Sphaeronemoura*  
68 and *Illiesonemoura* in the Palaearctic region [19], the western and eastern Himalayan ranges [20] and  
69 North and South India [15]. *Podmosta* and *Zapada* are two interesting cases distributed across the  
70 Nearctic region and East Asia [21], [22]. Previous studies have suggested that their current habitat  
71 distribution could be associated with mountain formation and land bridges. In Japan, the Nemouridae  
72 family is widely distributed with four genera [23] —*Indonemoura*; *Protonemura*; *Amphinemura*; and  
73 *Nemoura*. To date, 30 *Nemoura* species, 17 *Amphinemura* species, 12 *Protonemura* species and 1  
74 *Indonemoura* species have been reported in Japan [16]. However, their evolutionary history in the  
75 Japanese Archipelago remains unknown.

76 We studied the molecular phylogeny of the aquatic insect Nemouridae (Plecoptera) in the  
77 Japanese Archipelago with comprehensive genera-level sampling using mitochondrial cytochrome c  
78 oxidase 1 (*cox1*) and nuclear histone 3 (*H3*) markers. We hypothesised that the Nemouridae family  
79 diversification could be linked to the geological formation of the Japanese Archipelago. Therefore, we  
80 estimated the phylogenetic relationships among Nemouridae species and genera with reference to  
81 their historical biogeography. We focused on geographic events of Japanese Archipelago formation  
82 and their influence on the divergence time among the genera and species using a combination of fossil  
83 records and the Archipelago formation history. Furthermore, to estimate the historical process of the  
84 phylogeography of Nemouridae in Japan, we compared the phylogenetic relationships among the  
85 specimens from South Korea, China and North America, that are assumed to be the potential sources  
86 of Japanese Nemouridae because of the geological formation history of the Japanese Archipelago.

87

88

## 89 **Material and methods**

90

### 91 **Study sites and sample collection**

92 Our sampling sites in Japan comprised 32 sampling sites on Hokkaido Island, 83 on Honshu  
93 Island and 27 on Shikoku Island. None of the Nemouridae species was found on Hokkaido Island during  
94 sampling. All species reported from Hokkaido are known to occur on either Honshu or Shikoku Islands.  
95 Herein, we only reported on the sampling sites wherein specimens were found. We collected 20, 7,  
96 8 and 1 species of the genera *Nemoura*, *Amphinemura*, *Protonemura* and *Indonemoura*, respectively,  
97 on 110 sampling sites in Japan (Fig 1, S1 Table). Additionally, 14 species distributed from 8 sampling  
98 sites of Mainland China and 2 of South Korea (S1 Table, Fig 2) and 100 specimens of the three species  
99 *Zapada columbiana*, *Z. cinctipes* and *Podmosta delicatula* (subfamily Nemourinae) collected from 15  
100 sampling sites of North America (western United States of America and Alaska) were included in our

101 analysis. We added these samples from outside of Japan because of their geographical proximity to  
102 the Japanese Archipelago and their geological formation histories.

103 We collected adult insects using hand nets around riversides. We stored samples in 80%  
104 ethanol in the field, and replaced the ethanol with fresh 99.5% ethanol after morphological  
105 identification. We identified individuals according to the taxonomical keys of [21], [23], [24], [25], [26],  
106 [27], [28], [29], [30], [31] and [32]. Undescribed species resulted in our studied were based on our  
107 taxonomic expertise and inconclusive taxonomic keys.

108

### 109 **DNA extraction, amplification and sequencing**

110 We genetically analysed a total of 289 individuals, out of which 189 were from East Asia  
111 (males, 97; females, 92) and 100 were from North America (males, 92; females, 8). We extracted  
112 genomic DNA individually using DNeasy tissue kits (Qiagen GmbH, Hilden, Germany), following the  
113 manufacturer's instructions. We amplified a 658-bp fragment of mtDNA *cox1* using LCO-1490 and  
114 HCO-2198 primers [33] with an annealing temperature of 38°C and 40 PCR cycles. Further, we  
115 amplified a 328-bp fragment of nDNA marker histone 3 (*H3*) using the universal primers H3F and H3R  
116 [34] with an annealing temperature of 58°C and 40 PCR cycles. We purified the PCR products using  
117 the QIAquick PCR Purification Kit (Qiagen GmbH, Hilden, Germany) and sequenced them in both  
118 directions using the same primers as mentioned above. *Cox1* and *H3* sequences were sequenced by  
119 Eurofins Operon (Tokyo, Japan). All sequence data reported here have been deposited in GenBank.

120

### 121 **Sequence analysis**

122 We assembled and edited forward and reverse sequences using CodonCode Aligner v 3.5  
123 (Codon Code Corporation, Dedham, USA). All sequences were aligned using ClustalW  
124 (<https://www.genome.jp/tools-bin/clustalw>) [35]. We calculated the genetic diversity by the number  
125 of polymorphic sites, number of haplotypes and both mean nucleotide substitution rate (i.e individuals  
126 within species) and pairwise nucleotide substitution rate (i.e between species), with the Kimura 2-  
127 parameter model. We performed all analyses using DnaSp v5.10 [36]. All analyses were performed for  
128 *cox1* and *h3* separately.

129 All sequences of the mtDNA and nDNA markers were compared with the NCBI nucleotide  
130 database using blastn queries (<http://blast.ncbi.nlm.nih.gov>) to corroborate species identification  
131 (DNA barcoding, similarity > 98%) and to discard possible sequence errors.

132

### 133 **DNA species delimitation**

134 To corroborate the morphological species identification match with our molecular data, we  
135 implemented a DNA species delimitation analysis. Putative DNA species were delineated using the  
136 General Mixed Yule Coalescent model (GMYC; [37]). An ultrametric gene tree of *cox1* gene was  
137 constructed using BEST v1.8.3 [38], and the GMYC analysis was performed using the splits package  
138 [39] in R ver. 3.3 (R Core Team).

139

#### 140 **Molecular clock analysis**

141 We estimated the evolutionary history of the family in the Japanese Archipelago according to  
142 the timing of the divergence of the lineages. For this estimation, we implemented a Bayesian  
143 phylogenetic analysis in combination with a molecular clock analysis using BEAST v.2.4.4 [40] with  
144 *Zwicknia bifrons* (Capniidae) as an outgroup for *cox1* and *H3* (own sequences) separately. This outgroup  
145 was selected owing to their close phylogenetic relationship with Nemouridae [41], [42]. To observed  
146 the divergence time, we adopted a relax clock model [43] following a log normal distribution, and  
147 calibrated the phylogenetic tree nodes using three types of molecular clock analysis. The first  
148 calibration was based on fossil records of the Nemouridae family [44]. We calibrated the nodes at 180  
149 million years ago (Ma) and adjusted the parameters with a standard deviation of 20 Ma, as suggested  
150 in previous study [45] for a 95% highest posterior density (HPD). For this analysis, we implemented a  
151 fossilised birth death model [46] for tree prior parameter. The second calibration was based on the  
152 Japanese Archipelago formation events dated from 15 to 30 Ma [3]. We applied several calibrations  
153 from 15, 20, 25 and 30 Ma at all nodes representing taxonomic species. All calibrations were adjusted  
154 to 5 Ma as a standard deviation for a 95% HPD and a fossilised birth death model [46] for tree prior  
155 parameter. Lastly, the third calibration was the time to the most recent common ancestor (TMRCA)  
156 to observe species diversification patterns based on the mean substitution rate of *cox1*. Using a Yule  
157 model tree prior parameter [47], we applied the substitution rate for insect *cox1* of 1.5% [48] and  
158 3.54% [49] per million years for a 95% HPD.

159 For all branch age calibrations (namely, fossil, biogeographic and mtDNA substitution rate),  
160 we performed MCMC for 50 million generations, and log dating trees (BEAST parameters) for every  
161 5000 generations. We tested the output files for convergence after removing a 10% burn-in by  
162 examining the effective sampling size using Tracer v1.5 [50]. We pooled the four resulting output trees  
163 from biogeographical calibration analysis into a single tree. We then pooled the resulting single tree  
164 from biogeographical branch calibration and the single tree from fossil calibration analyses into a  
165 single tree. We performed all pooling analyses using Log Combiner v1.6.1 (BEAST package)  
166 summarised with Tree Annotator (BEAST package) and visualised using FigTree v1.3.1 [51]. We  
167 performed the analyses for *cox1* and *H3* separately. The incongruence length difference test (ILD) [52]

168 was conducted to test the congruence of tree topologies between *cox1* and *H3* using Tree Analysis  
169 Technology (TNT) [53]. ILD test revealed no significant differences in terms of the Bayesian tree  
170 topologies between *cox1* and *H3* ( $P = 0.8$ ); therefore, both markers were pooled into a single tree for  
171 further analysis.

172

### 173 **Phylogenetic analysis between Nemoura from Japan and North America**

174 To observe the phylogenetic relationship between Nemouridae from Japan and North  
175 America (*Zapada columbiana*, *Z. cinctipes* and *Podmosta delicatula*), we analysed the maximum  
176 likelihood (ML) phylogenetic trees of *cox1* and *H3* separately using PhyML 3.1 [54]. The General time-  
177 Reversible (GTR) model and gamma distribution were selected for both markers (*cox1* and *H3*) based  
178 on separate test performed with jModel Test v.3 [55] and using *Zwicknia bifrons* (Capniidae) as an  
179 outgroup as described above. The trees were bootstrapped using 10,000 replications.

180

## 181 **Results**

182

### 183 **Genetic diversity and DNA phylogeny**

184 For studying the phylogeny of Nemouridae in the Japanese Archipelago, we analysed two  
185 molecular markers. *Cox1* sequences were of 658 bp length, with 247 polymorphic sites, 237  
186 parsimony-informative sites, 10 singletons and a mean nucleotide substitution rate of 0.151. *H3*  
187 sequences were of 328 bp length, with 67 polymorphic sites, 54 parsimony-informative sites, 13  
188 singletons and a mean nucleotide substitution rate of 0.051. No gaps were detected for either *cox1* or  
189 *H3* sequences (S1, S2 Fig). In total, for *cox1* and *H3*, we identified 128 and 68 haplotypes, respectively.

190 The GMYC model of *cox1* delimited 61 putative DNA-species (S1 Table). These results agreed  
191 with our 34 morphologically identified and 15 undescribed (five species of *Protonemura*, seven of  
192 *Nemoura*, one of *Indonemoura* and two of *Amphinemura*) species. Eight species (*I. nohirae*, *A.*  
193 *decemseta*, *A. zonata*, *A. longispina*, *A. megaloba*, *N. uenoji*, *N. chinonis* and *N. cf. cercispinosa*) showed  
194 two putative DNA-species. While *A. decemseta* showed multiple putative DNA-species (three putative  
195 DNA-species), *N. sanbena* and *P. kohnoae* showed two putative DNA-species in the same sampling site  
196 suggesting the presence of cryptic species. The congruence of *H3* phylogenetic groups provided  
197 confirmation of DNA-based groups detected by GMYC.

198 We observed the genetic diversity of the species per island (Table 1). Honshu had the highest  
199 number of species (26 species), haplotype richness (63) and mean nucleotide substitution rate  
200 (average 0.027). Five species were found throughout the three Japanese islands (Honshu, Shikoku and  
201 Kyushu), i.e. *A. decemseta*, *A. zonata*, *N. cf. cercispinosa*, *N. chinonis* and *I. nohirae*, with a mean

202 nucleotide substitution rate ranging from 0.011 to 0.126 and a total of 23 haplotypes. *N. sanbena*  
203 haplotypes were observed in two different branches in the phylogenetic tree, both within *N. cf.*  
204 *cercispinosa* and as an isolated branch.

205

## 206 **Divergence dates**

207 The Bayesian phylogenetic trees for *cox1* and *H3* showed tree topology similarity (ILD test,  $P$   
208 = 0.8). Three clades corresponded to the three families—*Protonemura*, *Amphinemura* and  
209 *Nemoura*—whereas *Indonemoura* was divided into two clades—the Mainland China clade, clustered  
210 with *Protonemura*, and the Japanese clade (Fig 2).

211 The evolutionary divergence between the Nemouridae and Capniidae families was settled at  
212 180 Ma, with a 95% HPD interval of 160 to 198 Ma, in the Jurassic geological period (Fig 2, S3 and S4  
213 Fig). Genus-level diversifications within Nemouridae occurred in the early and middle Cretaceous.  
214 *Indonemoura* from Japan at 119.0 Ma (95% HPD, 125.8 to 100.2 Ma), *Indonemoura* from Mainland  
215 China at 112.0 Ma (95% HPD, 90.2 to 115.0 Ma), *Protonemura* at 112.7 Ma (95% HPD, 98.0 to 121.3  
216 Ma), *Nemoura* at 107.0 Ma (95% HPD, 98.8 to 110.1 Ma) and *Amphinemura* at 80.0 Ma (95% HPD,  
217 75.1 to 92.0 Ma). The speciation process occurred between 25 Ma (early Paleogene) and 90 Ma (late  
218 Crustaceous). Out of 35 events of speciation (i.e. nodes), 16 (45%) occurred during late Crustaceous  
219 and 19 (54%) occurred during early Paleogene, broadly overlapping with the formation time of the  
220 Japanese Archipelago (15 to 30 Ma). We observed intra-species diversification in *I. nohirae*, *A.*  
221 *decemseta*, *A. zonata*, *A. longispina*, *A. megaloba*, *N. chinonis*, *N. uenoi* and *N. cf. cercispinosa* (GMYC  
222 > 1 species, S1 Table). These species were divided into two clades (S5 Fig), spatially segregated into  
223 eastern and western Japan of the Fossa Magna region during the late Neogene period (20 to 22 Ma).  
224 Recent diversifications for *Nemoura* and *Amphinemura* species within either eastern or western  
225 Japanese branches were additionally revealed by TMRCA analysis of *cox1* (see Methods). *A. decemseta*  
226 ranging from 3 to 3.5 Ma (95% HPD, 2.8 to 4.1 Ma); *A. zonata*, ranging from 3 to 4 Ma (95% HPD, 3.5  
227 to 5 Ma); *A. longispina*, ranging from 3.6 to 4.5 Ma (95% HPD, 3.9 to 5 Ma); *A. megaloba*, ranging from  
228 3.5 to 4 Ma (95% HPD, 2.8 to 4 Ma); *N. uenoi*, ranging from 3 to 4 Ma (95% HPD, 3.5 to 4.2 Ma) and *N.*  
229 *cf. cercispinosa*, ranging from 3.5 to 4.1 Ma (95% HPD, 3 to 5 Ma), for 1.5% Ma and 3.54% Ma analysis  
230 respectively.

231

## 232 **Phylogeographic pattern between Nemoura from Japan and North America**

233 DNA sequences in the Japanese clade of *Indonemoura* (single species, *I. nohirae*) showed a  
234 high homology with those in the Alaskan species of *Z. columbiana* (*COI*: KM874174; >93% sequence  
235 similarity) and *Z. cinctipes* (*H3*: EF622600; >98% sequence similarity) based on blastn results. The ML



236 phylogenetic trees for both *cox1* and *H3* (Fig 3) showed that the *Indonemoura* Japanese clade  
237 clustered with three North American species (*Z. columbiana*, *Z. cinctipes* and *P. delicatula*) and the  
238 *Indonemoura* Mainland China clade clustered with the East Asian Nemouridae genera (*Nemoura*,  
239 *Protonemura* and *Amphinemura*). The pairwise nucleotide substitution rate based on *cox1* between  
240 the *Indonemoura* Japanese clade and *Zapada* spp. or *P. delicatula* from North America ranged from  
241 0.13 to 0.15, whereas a higher pairwise nucleotide substitution rate based on *cox1* of 0.26 was  
242 observed between the *Indonemoura* Japanese and Mainland China clades (Table 2).

243

## 244 Discussion

245 We studied mitochondrial *cox1* and nuclear *H3* gene sequences to determine the patterns of  
246 diversification and phylogenetic relationships of species belonging to four genera of stoneflies of the  
247 Nemouridae family in the Japanese Archipelago. We estimated the divergence among *Nemoura*,  
248 *Amphinemura*, *Indonemoura* and *Protonemura* to have occurred in the early and mid-Cretaceous  
249 (around 100 Ma), which is compatible with previous studies based on fossil records [56], [57]. Our  
250 results suggested that these four genera might have dispersed and colonised different areas of the  
251 Eurasian continent—including the Japanese landmasses—when they were still connected to the  
252 Eurasian continent. Among the four genera, the diversification of *Indonemoura* occurred earlier (120  
253 Ma) than that of the other genera (<100 Ma), suggesting that it is an ancient genus. The geological  
254 isolation of colonised areas [58], the long evolutionary time [59] and poor dispersal ability of  
255 *Indonemoura* [23], [60] might have accounted for their ancient diversification.

256 Based on the phylogenetic relationships of both molecular markers (*cox1* and *H3*), we  
257 observed that the three genera *Nemoura*, *Amphinemura* and *Protonemura* were monophyletic and  
258 clustered as three independent groups, as previously observed by morphological systematics [16].  
259 However, *Indonemoura* was paraphyletic. This genus was divided into two clades corresponding to  
260 the Mainland China and the Japanese clades. Surprisingly, the Japanese clade of *Indonemoura* (single  
261 species, *I. nohirae*) clustered together with North American species (*Z. columbiana*, *Z. cinctipes* and *P.*  
262 *delicatula*), with a low pairwise nucleotide substitution rate (<0.15). The distribution range of these  
263 two North America genera covers North America and Eastern Asia. Previous studies suggested that  
264 their distribution could be related to the land connection (i.e. the islands) between Alaska and Eastern  
265 Asia [15]. Dispersal by island connectivity between Alaska, the Aleutian Islands, the Kamchatka  
266 peninsula and the Kuril islands has been observed in other stonefly families (for instance, *Arcynopteryx*  
267 *dichroa*, *Capnia nearctica*, *Mesocapnia variabilis* and *Nemoura arctica*) [61]. However, the distribution  
268 of *Indonemoura* on these islands is unknown.

269           The complex history of the geological formation of the Japanese Archipelago may provide a  
270 possible alternative explanation. The ancestral Japanese landmasses were located on the borders of  
271 four major tectonic plates, of which two are continental plates—the Eurasian plate and the North  
272 American plate [4] (S6 Fig). The eastern Japanese landmass was located on the North American plate,  
273 whereas the western Japanese landmass was located on the Eurasian plate [5]. The dispersal and  
274 colonisation of *Indonemoura* might have occurred from the North American plate to the Eurasian  
275 continent or vice versa (from the Eurasian continent to the North American plate) before their  
276 geographic separation in an ancient time (around 70 to 80 Ma) [62]. Dispersal events between  
277 Eurasian and Japanese landmasses are commonly reported for aquatic insects [10], [63]. Particularly,  
278 a dispersal event between North America and the Japanese Archipelago was detected by the  
279 phylogenetic relationship of the monophyletic group of caddisflies, *Palaeagapetus* spp. [9]. However,  
280 no prior studies have observed speciation events of aquatic insects associated with geological events  
281 that occurred in ancient times (>12 Ma). Our result suggests an ancient divergence time and a  
282 distribution pattern of *Indonemoura*, consistent with a hypothesis of an ancient colonisation  
283 influenced by the connection of the Japanese landmass with the North American plate in the Eurasian  
284 continent.

285           Nemouridae species diversification, as has been observed in other species of aquatic insects,  
286 such as beetles [10], caddisflies [9], water bugs [14] and mayflies [8], [13], was also observed to be  
287 affected by the geological formation of the Japanese Archipelago. The diversification of the  
288 Nemouridae species occurred during the Paleogene period (<50 Ma). This geological period is  
289 consistent with the movement of landmasses (S6 Fig) about 70 Ma ago [4] and the active geological  
290 formation of the Japanese Archipelago around 20 Ma ago [5], which could be the cause of the  
291 Nemouridae diversification, as previously reported for the mayfly *Dipteromimus flavipterus* (35 Ma)  
292 [2].

293           *Indonemoura nohirae* is the single species of *Indonemoura* on the Japanese Archipelago [25],  
294 [26]. The morphology of their terminalia resembles that of *Protonemura* rather than of *Indonemoura*,  
295 but the characteristic gill formula justifies their taxonomical classification in *Indonemoura* [25], [26].  
296 To date, there are 24 *Indonemoura* species from China [16], [24] and 30 species belonging to the  
297 Himalayan and Oriental regions in East Asia [15], [20]. These species are morphologically different  
298 from *I. nohirae* in Japan [15], [16], [20], [24], [25], [26]. We hypothesise that the *Indonemoura* species  
299 of East Asia could be forming separate phylogenetic clades clustered by geographical regions. For the  
300 hypothesis testing, further collection of molecular data on *Indonemoura* from wider areas such as  
301 Northeast China, Southeast China, Mongolia, Russia and other countries in Asia is needed in future  
302 studies.

303           Eight species (*I. nohirae*, *A. decemseta*, *A. zonata*, *A. longispina*, *A. megaloba*, *N. chinonis*, *N.*  
304 *uenoi* and *N. cercispinosa*) showed interesting patterns of intra-species separation into two genetic  
305 groups corresponding to eastern and western areas of the Fossa Magna region of Honshu Island (S5  
306 Fig). Honshu is the centre of insect biodiversity [10]; apart from its extensive territorial space, it is the  
307 main island with a geological history [3], [4], [5]. We found supporting evidence on the genetic  
308 diversity of these eight species. We found a larger mean nucleotide substitution rate and haplotype  
309 number in the Honshu region than in other islands (Table 1). The mean nucleotide substitution rate  
310 and haplotype diversity are indications of biodiversity [64], which could lead to evidence of speciation  
311 [65]. Out of eight species, the diversification of six species (*A. decemseta*, *A. zonata*, *A. longispina*, *A.*  
312 *megaloba*, *N. uenoi* and *N. cercispinosa*) occurred during the late Neogene period (20 to 22 Ma). This  
313 event corresponded with the double-door (i.e. the union of eastern and western Japan; S6 Fig)  
314 geological model and the formation of the Itoigawa-Shizuoka tectonic line (ISLT) at around 20 Ma [5],  
315 [66]. The speciation of aquatic insects was often observed to be influenced by these two geological  
316 events [2]. Additionally, species diversification—from eastern or western Japan of the Fossa Magna  
317 region—showed recent diversification events (3 to 5 Ma) corresponding with the formation of the  
318 small islands in northeastern or southwestern edge areas of Japan (Fig 1). The northeastern islands  
319 created land bridges between the Japanese Archipelago and China or Korea, whereas the  
320 southwestern islands connected Taiwan or the Philippines with the Japanese Archipelago [5], [66].  
321 This connectivity promoted immigration events in Japan that might have contributed to the formation  
322 of the current genetic diversity, as previously observed in mayflies [13] and beetles [10].

323           The evolutionary divergence of the Nemouridae family was promoted by the complex  
324 geological formation of the Japanese Archipelago. Despite the different evolutionary rate of both  
325 molecular markers, Bayesian analysis found congruence between both markers; however, failed to  
326 find congruence with their morphological taxonomy. The main morphological character used for  
327 identification of adult stoneflies species is its genital morphology. The evolution of genital morphology  
328 is, however, governed by within-population sexual selection rather than environmental or geological  
329 history of the locations [67]. Conversely, the genetic variation of natural populations has been  
330 observed to be directly associated with environmental [68] and geological variations [2]. Therefore,  
331 the genetic variation could reflect an independent course in the evolutionary history of *Indonemoura*  
332 than do the morphological characters used for their taxonomy. However, we detected that *N. sanbena*  
333 shared haplotypes from different lineages, revealing a possible introgression or incomplete sorting of  
334 ancestral polymorphisms [10]. This is an often reported phenomenon in stoneflies [40], [69], [70],  
335 which remains as unresolved species. Resolving the problems between the process of evolution of

336 morphological characters and the genetic variation within species will improve our future  
337 understanding of the origin of the species and the local species distribution.

338 Finally, our inference of divergence time was based on the coalescent simulation approach.  
339 Despite the frequent use of this approach, a biased sampling of lineages and extreme state-dependent  
340 molecular substitutions rate heterogeneity are known to potentially cause erroneous inference of  
341 divergence time [71]. Therefore, combining node calibrations generated by more than one calibration  
342 analyses is recommended [71], [72]. A cautious method such as the combined uses of fossil records  
343 and biogeographic ages as employed in our analysis may minimise the risk of such erroneous  
344 inference.

345

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350

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523  
524

525 **Table 1.** Regional distribution of sample size (n), haplotype richness (h) and mean nucleotide  
 526 substitution rate of Nemouridae species among the three main islands in Japan, based on  
 527 mitochondrial DNA (*cox1*) sequences. Total species richness was 26, 23 and 6 for Honshu, Shikoku and  
 528 Kyushu, respectively.  
 529

Genus	Species	Honshu			Shikoku			Kyushu		
		n	h	Nucleotide substitution	n	h	Nucleotide substitution	n	h	Nucleotide substitution
<i>Amphinemura</i>	<i>A. bulla</i>	5	4	0.006						
	<i>A. decemseta</i> *	19	13	0.014	9	5	0.02	3	1	0
	<i>A. dentifera</i>	2	1	0	2	2	0.01			
	<i>A. flavostigma</i>	3	3	0.005	3	2	0.01			
	<i>A. longispina</i>	2	1	0						
	<i>A. megaloba</i>	4	2	0.091	3	1	0			
	<i>A. zonata</i> *	2	2	0.053	1	1		1	1	
	<i>A. sp. n.</i>	1	1		3	3	0.03			
<i>Indonemoura</i>	<i>I. nohirae</i> *	9	4	0.059	4	2	0.01	2	1	0
<i>Nemoura</i>	<i>N. akagii</i>	2	1	0						
	<i>N. cf. cercispinosa</i> *	2	2	0.011	13	7	0.01	2	1	0
	<i>N. chinonis</i> *	2	2	0.126	5	3	0.09	1	1	
	<i>N. fulva</i>	3	3	0.043						
	<i>N. cf. hikosan</i>				2	2	0.1			
	<i>N. longicercia</i>	2	1	0	7	5	0			
	<i>N. naraiensis</i>				2	1	0			
	<i>N. ovocercia</i>	1	1							
	<i>N. redimiculum</i>	1	1		3	3	0.01			
	<i>N. sanbena</i>				2	2	0.5			
	<i>N. shikokuensis</i>				4	1	0			
	<i>N. stratum</i>	2	2	0.027						
	<i>N. speciosa</i>	2	2	0.003						
	<i>N. transversospinosa</i>				6	4	0.01			
	<i>N. uenoi</i>	1	1		2	2	0.01			
	<i>N. yakushimana</i>							2	1	0
	<i>N. sp. n. 1</i>				3	1	0			
	<i>N. sp. n. 2</i>	2	2	0.003						
<i>N. sp. n. 3</i>	1	1								
<i>N. sp. n. 4</i>	1	1								
<i>Protonemura</i>	<i>P. kohnoae</i>	6	3	0.043						
	<i>P. orbiculata</i>	6	6	0.029						
	<i>P. sp. n.</i>				2	2	0.01			
	<i>P. sp. n. 1</i>	1	1							
	<i>P. sp. n. 2</i>	2	1	0						
	<i>P. sp. n. 3</i>	1	1							
	<i>P. sp. n. 4</i>						1	1		

530 (\*) Species found on the three Japanese islands.



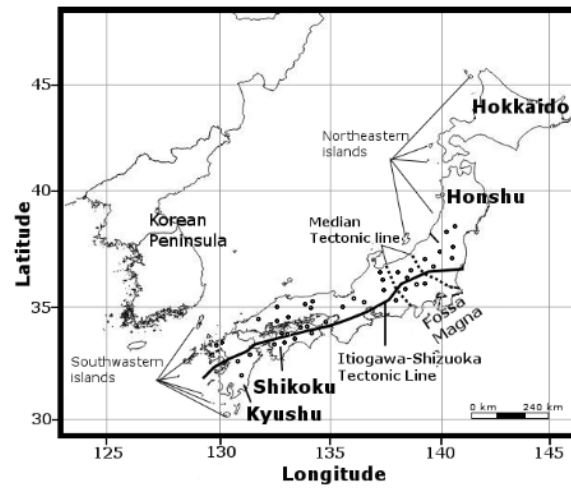
531 **Table 2.** Pairwise nucleotide substitution rate based on *cox1* between the East Asian Nemouridae and North American (western USA and Alaskan) species.

East Asia	<i>Amphinemura</i> spp.	0							
	<i>Nemoura</i> spp.	0.187	0						
	<i>Protonemura</i> spp.	0.197	0.19	0					
	<i>Indonemoura</i> spp. (China)	0.213	0.193	0.197	0				
	<i>Indonemoura</i> spp. (Japan)	0.197	0.183	0.175	0.260	0			
North America	<i>Zapada columbiana</i>	0.178	0.165	0.182	0.190	0.145	0		
	<i>Zapada cinctipes</i>	0.170	0.154	0.156	0.179	0.149	0.133	0	
	<i>Podmosta delicatula</i>	0.202	0.196	0.191	0.205	0.135	0.185	0.201	0

532

533 **Figures**

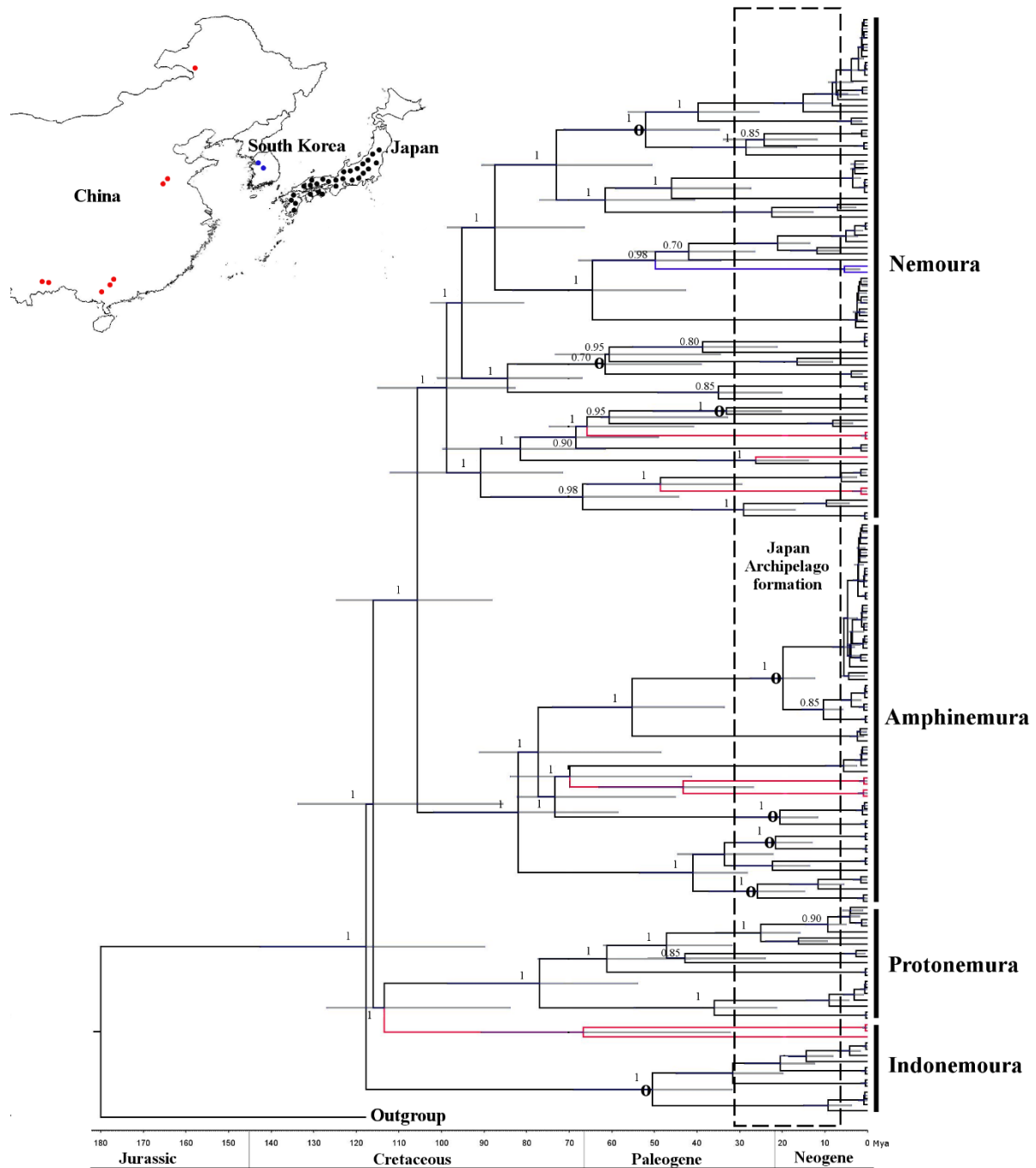
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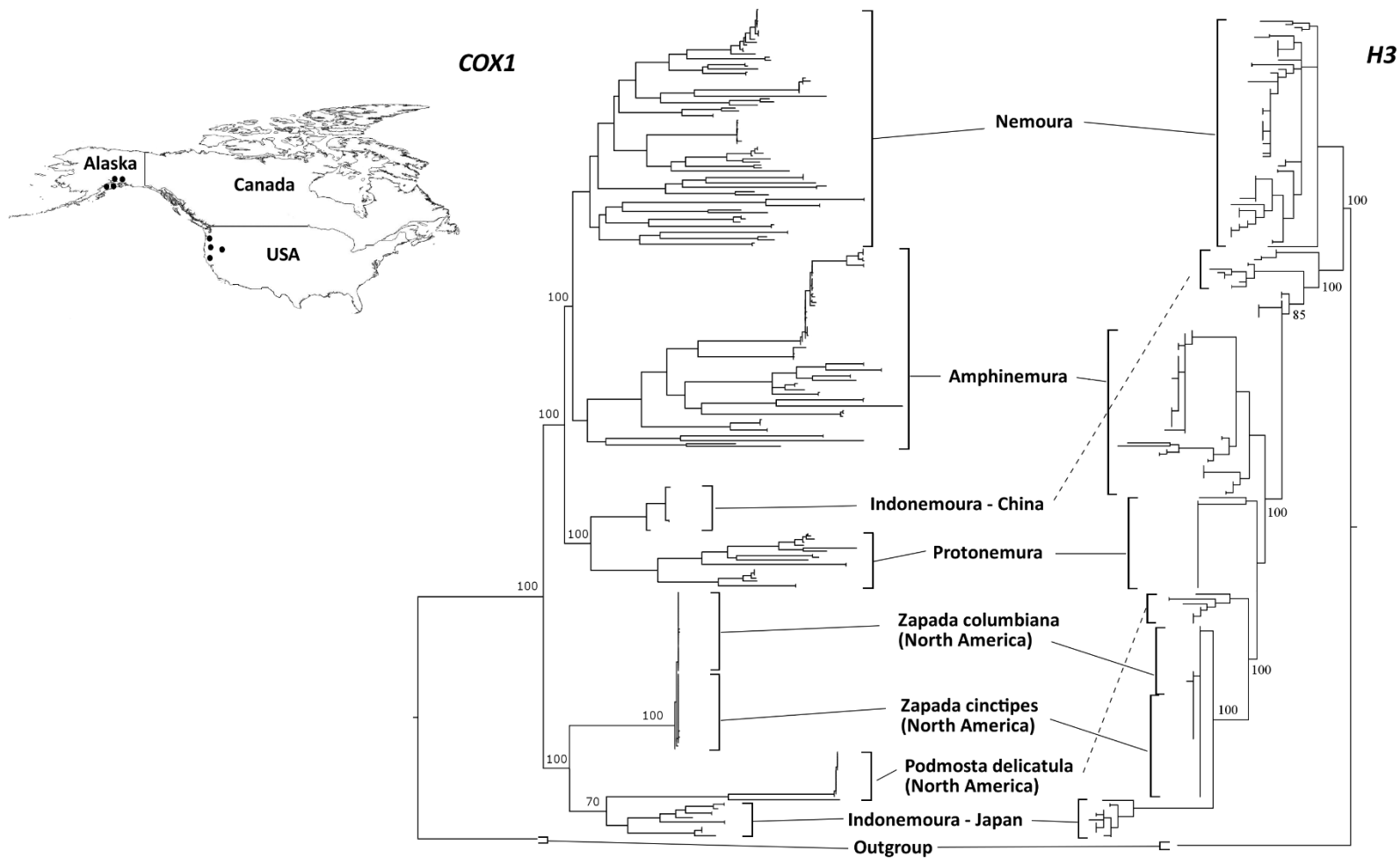
536 **Fig 1.** The Japanese islands and distribution of 110 sampling sites from where Nemouridae samples

537 were collected (open circles).



538

539 **Fig 2.** Concatenated Bayesian phylogeny (*cox1* + *H3*) of the East Asian Nemouridae family. The  
540 phylogenetic tree nodes were calibrated using 180 Ma based on fossil records + 15 to 30 Ma based on  
541 the Japanese Archipelago formation. Calibration and geological time are shown at the bottom of the  
542 tree. A 95% HPD is indicated as a horizontal grey bar and posterior probabilities are shown for each  
543 node. Circle symbol (o) in the nodes indicates intra-species diversification based on the eastern and  
544 western Japanese boundaries of the Fossa Magna region. Inserted upper map shows sample site  
545 locations for Japan (black), China (red) and South Korea (blue) as dots. Colour branches indicate  
546 sample location distribution as shown in the map.



547

548 **Fig 3.** Maximum likelihood trees based on both *cox1* and *H3* markers for comparison between the East Asia Nemouridae family and three North American  
 549 Nemourinae species: *Zapada cinctipes*, *Z. columbiana* and *Podmosta delicatula*. Inserted upper map shows sampling site locations in North America (western  
 550 USA and Alaska) as black dots.

## Supplementary information

**S1 Table.** Location information of samples of East Asia Nemouridae. Numbers of individuals (N), presences of male (M), female (F) and imago (im), DNA-species delimitation (GMYC).

**S1 Fig.** Multiple alignment of *cox1* region for the 54 Nemouridae genotypes.

**S2 Fig.** Multiple alignment of *H3* regions for the 54 Nemouridae genotypes.

**S3 Fig.** *Cox1* Bayesian trees using three types of node calibration. A – fossil, B – island formation and D – TMRCA.

**S4 Fig.** *H3* Bayesian trees using two types of node calibration. A – fossil and B – islands formation.

**S5 Fig.** Concatenated Bayesian phylogeny (*cox1* + *H3*) for East Asian Nemouridae family enlarging intra-species diversification in *I. nohirae*, *A. decemseta*, *A. zonata*, *A. longispina*, *A. megaloba*, *N. chinonis*, *N. uenoi* and *N. cf. cercispinosa* (GMYC = 2 species).

**S6 Fig.** Putative formation of the Japanese Archipelago [2], [3], [4], [5]. (A) Around 30 to 130 Ma, the Japanese landmasses were located in two major tectonic plates from the Eurasian continent. (B) Around 15 to 30 Ma, the Japanese landmasses began to separate from Eurasia and the North American Plates began to separate from the Eurasian continent, and remained separated by a sea zone called Fossa Magna—a geological event called double-door. (C) Current map of the Japanese Archipelago in East Asia, where the names of the main four Japanese islands and the two tectonic lines are shown.