

## Article

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### A new species of *Lethrinops* (Cichliformes: Cichlidae) from a Lake Malawi satellite lake, believed to be extinct in the wild.

GEORGE F. TURNER<sup>1</sup>, DENISE A. CRAMPTON<sup>2</sup> & MARTIN J. GENNER<sup>3</sup>

<sup>1</sup> School of Natural Sciences, Bangor University, Bangor, Gwynedd LL57 2UW, United Kingdom & Vertebrates Division, Natural History Museum, Cromwell Road, London SW7, UK.

Corresponding author: email [bss608@bangor.ac.uk](mailto:bss608@bangor.ac.uk)

ID: <https://orcid.org/0000-0003-0099-7261>

<sup>2</sup> School of Natural Sciences, Bangor University, Bangor, Gwynedd LL57 2UW, United Kingdom; present address: School of Biological & Environmental Sciences, Liverpool John Moores University, Liverpool, L3 3AF, UK; email: [D.Crampton@2023.ljmu.ac.uk](mailto:D.Crampton@2023.ljmu.ac.uk)

ID: <https://orcid.org/0000-0002-2877-5209>

<sup>3</sup> School of Biological Sciences, University of Bristol, Life Sciences Building, 24 Tyndall Avenue, Bristol, BS8 1TQ, United Kingdom. Email: [m.genner@bristol.ac.uk](mailto:m.genner@bristol.ac.uk)

ID: <https://orcid.org/0000-0003-1117-9168>

#### Abstract

A new species of cichlid fish, *Lethrinops chilingali* is described from specimens collected from Lake Chilingali, near Nkhotakota, Malawi. It is assigned to the genus *Lethrinops* based on the form of the lower jaw dental arcade and by the absence of traits diagnostic of the phenotypically similar *Ctenopharynx*, *Taeniolethrinops* and *Tramitichromis*. It also lacks the enlarged cephalic lateral line canal pores found in species of *Alticorpus* and *Aulonocara*. The presence of a broken horizontal stripe on the flanks of females and immature/non-territorial males of *Lethrinops chilingali* distinguishes them from all congeners, including *Lethrinops lethrinus*, in which the stripe is typically continuous. *Lethrinops chilingali* also has a relatively shorter snout, shorter lachrymal bone and less ventrally positioned mouth than *Lethrinops lethrinus*. It appears likely that *Lethrinops chilingali* is now extinct in the wild, as this narrow endemic species has not been positively recorded in the natural environment since 2009. Breeding populations remain in captivity.

**Keywords:** African cichlid, haplochromine, Lake Chilingali, morphology.

## 39 Introduction

40 Satellite lakes are small lakes lying in the catchment of much larger lakes, formerly or  
41 sometimes intermittently connected (Kaufman & Ochumba 1993; Mwanja et al. 2001; Genner  
42 et al. 2007). Their presence has been proposed to enhance the generation of biodiversity by  
43 isolating populations and facilitating allopatric speciation. Their role in the generation of  
44 African cichlid fish diversity was highlighted by the discovery of unique haplochromine cichlid  
45 fishes in Lake Nabugabo in the Lake Victoria catchment (Greenwood, 1965). Subsequently,  
46 several satellite lakes around Lake Malawi have also been shown to be inhabited by unique  
47 haplochromine cichlid fish populations (Turner *et al.*, 2019). These satellite water bodies  
48 include Lake Chilingali, a small lake lying on the Kaombe River which flows into the middle  
49 of the western shoreline of Lake Malawi near Nkhotakota, from which a phenotypically distinct  
50 haplochromine species informally referred to as *Lethrinops* sp. “chilingali” (Tyers et al. 2014;  
51 Turner et al. 2019) has been sampled.

52 The genus *Lethrinops* Regan 1922 is currently used for haplochromine cichlids endemic to the  
53 Lake Malawi catchment distinguished by the semicircular shape of the dental arcade of the  
54 outer series of lower jaw teeth, which curves round to end abruptly behind the inner row(s), if  
55 present (Trewavas 1931, Turner 1996, Ngatunga & Snoeks 2004). This character is also found  
56 in the genera *Taeniolethrinops* Eccles & Trewavas 1989 and *Tramitichromis* Eccles &  
57 Trewavas 1989 which were split off from *Lethrinops* by Eccles & Trewavas (1989). The  
58 character is also known in a single species of the genus *Ctenopharynx* Eccles & Trewavas 1989  
59 [*Ctenopharynx pictus* (Trewavas 1935)]. All of these taxa have ventrally positioned mouths,  
60 and relatively flat lower jaws with thin mandibular bones and small teeth. This jaw structure is  
61 believed to be associated with their feeding behaviour, which, where known, largely consists  
62 of ‘sediment-sifting’ or ‘winnowing’ (Weller *et al.* 2022), whereby loose sand or mud is picked  
63 up in the mouth, tumbled briefly and then ejected through the mouth and / or operculum,  
64 presumably with prey retained and swallowed (Fryer 1959; Fryer & Iles 1972; Konings 2016).  
65 Species in the genus *Lethrinops* are largely distinguished from *Taeniolethrinops*,  
66 *Tramitichromis* and *Ctenopharynx* by their lack of traits that distinguish those genera (Eccles  
67 & Trewavas 1989, Turner 2022). Not surprisingly, *Lethrinops* is currently believed to be  
68 polyphyletic (Ngatunga & Snoeks 2004; Malinsky et al. 2018; Masonick et al. 2022).  
69 Currently, the genus is ‘operational’, in the sense that it is possible to determine whether newly  
70 discovered taxa fall within its definition.

71 The purpose of the current work is to describe the Lake Chilingali species previously referred  
72 to as *Lethrinops* sp. ‘chilingali’ (Tyers et al. 2014; Turner et al. 2019) as *Lethrinops chilingali*,  
73 and to compare it with its presumed sister species from the main body of Lake Malawi, the  
74 morphologically similar *Lethrinops lethrinus* (Günther, 1893). The distributions of both  
75 species are discussed, and the current conservation status of *L. chilingali* is reviewed.

## 76 Materials and methods

77 Specimens of the new species were obtained from fishermen on the shores of Lake Chilingali  
78 from 22-24 June 2009, euthanised with MS-222 (if still alive) and fixed in 10% formalin before  
79 being transferred to 70% alcohol (Industrial Methylated Spirit, IMS) for long term  
80 preservation. Additional specimens obtained from a captive strain kept at Bangor University  
81 euthanised in 2020 were preserved directly in IMS. These were used to investigate allometric

82 comparisons between the two species, as they had grown to larger sizes than field-collected  
83 material. These captive bred fishes were excluded from the type series, but were included in  
84 statistical tests.

85 Comparative material of *L. lethrinus* included the holotype, and material from collections that  
86 were made in 1991-1992. These specimens were fixed in formalin and preserved in alcohol,  
87 along with some specimens collected in 2017 that were preserved directly in alcohol.  
88 Information on other congeneric species was obtained from literature, notably Trewavas  
89 (1931), Eccles & Lewis (1978), Eccles & Trewavas (1989), Turner (1996) and Ngatunga &  
90 Snoeks (2004). Counts and linear measurements were carried out following the methods of  
91 Snoeks (2004), and analysed using SPSS v27 (IBM, NY).

92 Geometric morphometric analyses were carried out on preserved specimens, photographed  
93 against a standard grey background with a scale for calibration. An initial tps file was  
94 constructed using image file names with tpsUtil v1.82 (Rohlf, 2015). A total of 15 landmarks  
95 (Figure 1) were then placed using tpsDig2 v2.32 (Rohlf, 2015): 1 anterior tip of upper jaw; 2  
96 posterior tip of upper jaw (maxilla); 3-6 anterior, posterior, lower and upper point of eye; 7-8  
97 beginning and end of dorsal fin; 9-10 beginning and end of anal fin; 11 anterior origin of pelvic  
98 fin; 12-13 lower and upper insertion of pectoral fin, 14 posterior margin of upper insertion of  
99 the operculum, 15 base of isthmus. The posterior of the caudal peduncle was not landmarked  
100 due to the upward flexion of the peduncle in several *L. lethrinus* specimens. Landmark data  
101 from the tps file were imported to MorphoJ v1.07 (Klingenberg 2011), where a Procrustes  
102 analysis was used to transpose, rotate and scale them into comparable Procrustes coordinates.  
103 These were analysed using SPSS v27 (IBM, NY).

104 Observations of live fish were collected from stocks descended from wild-caught fish obtained  
105 from Lake Chilingali between 2004 and 2009. Information on diets was taken from previous  
106 publications (Tyers et al. 2014; Turner et al. 2019), and an additional three wild-caught  
107 specimens of the new species were dissected to inspect stomach contents. Data and images  
108 used in analyses are available at: <https://doi.org/10.5281/zenodo.8007304>.

## 109 **Results**

### 110 *Quantitative comparisons*

111 Geometric morphometric data were ordinated using a Principal Component Analysis, with the  
112 primary axis (PC1) and secondary axis (PC2) capturing 34.2 and 19.6% of the variation,  
113 respectively. Overall, there was highly significant differentiation between *L. chilingali* and *L.*  
114 *lethrinus* on PC1 (General Linear Model;  $F_{1,47} = 39.25$ ,  $P < 0.001$ ), but not PC2 ( $F_{1,47} = 0.52$ ,  
115  $P = 0.60$ ). The respective type specimens were among the most clearly differentiated  
116 individuals (Figure 1). The wireframe plots showed that the *L. lethrinus* specimens have a  
117 relatively more ventrally positioned mouth than *L. chilingali*, leading to a longer snout, and a  
118 deeper body at the anterior insertion of the dorsal fin.

119 Comparisons of linear morphometric measurements revealed significant differences in slopes  
120 of head length, anal fin base length and caudal peduncle length when regressed on standard  
121 length (Table 1). Assuming equal slopes, and using standard length as a covariable, *L. lethrinus*  
122 had significantly relatively greater body depth, interorbital width, snout length, lower jaw  
123 length, lachrymal bone depth, pre-pelvic length and caudal peduncle depth than *L. chilingali*

124 (Table 1). The clearest differences were in snout length and lachrymal bone depth, followed by  
125 interorbital width.

126 Comparisons of meristic counts showed that *L. lethrinus* had significantly more cheek scale  
127 rows than *L. chilingali* (K-S test,  $Z = 2.001$ ,  $P = 0.001$ ). Meanwhile *L. chilingali* had  
128 significantly more dorsal rays (K-S test,  $Z = 1.805$ ,  $P = 0.003$ ), upper gillrakers (K-S test,  $Z =$   
129  $1.682$ ,  $P = 0.007$ ) and lower gillrakers (K-S test,  $Z = 2.903$ ,  $P < 0.001$ ) than *L. lethrinus* (Tables  
130 2 & 3). There were no differences between the species in dorsal spines (K-S test,  $Z = 1.05$ ,  $P =$   
131  $0.221$ ), anal spines (always 3), anal rays (K-S test,  $Z = 0.265$ ,  $P \sim 1.00$ ) or lateral line scales  
132 (K-S test,  $Z = 0.98$ ,  $P = 0.292$ ) (Tables 2 & 3).

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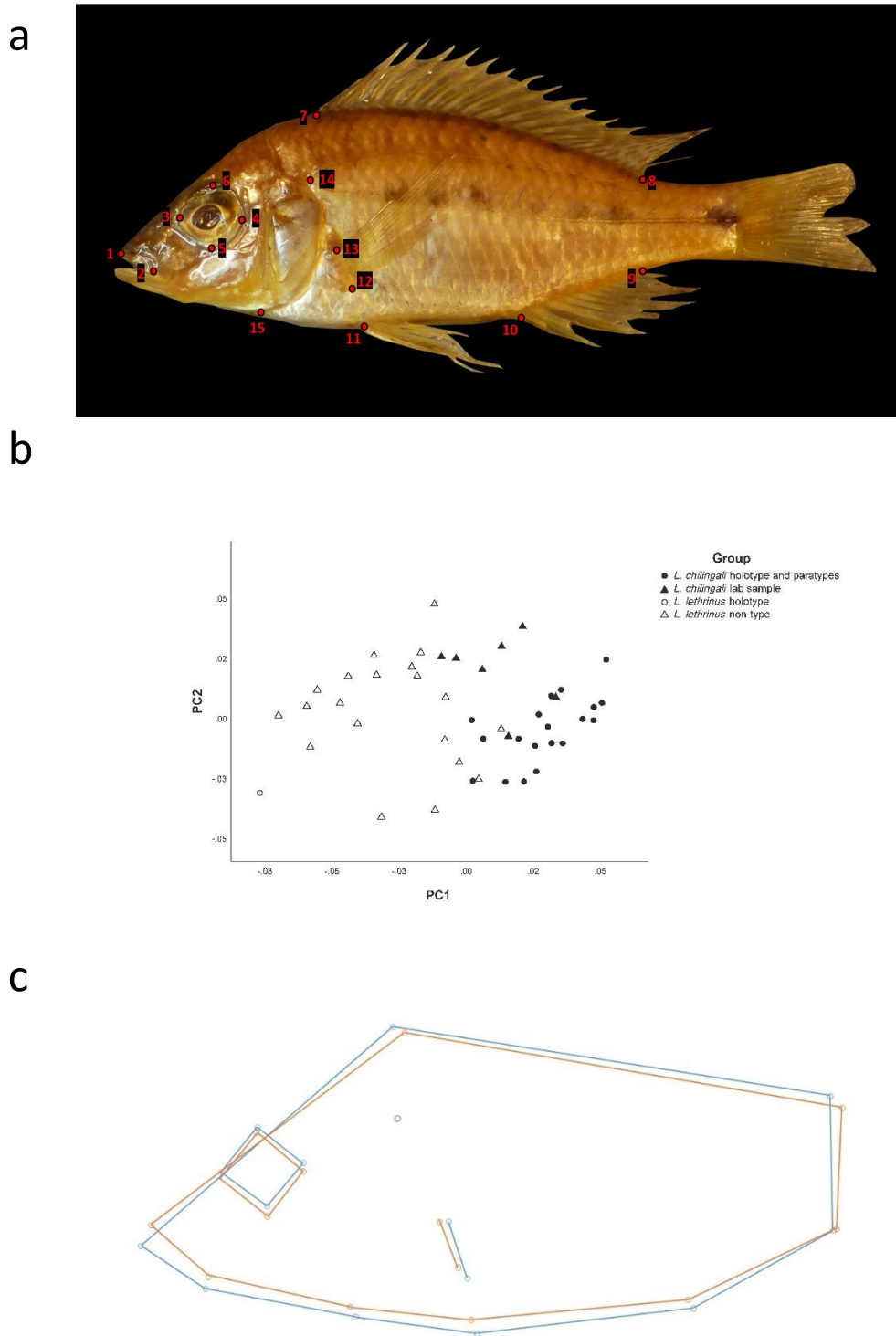
134 **TABLE 1.** Comparison of linear morphometric measurements between *Lethrinops chilingali*  
135 (including captive-bred specimens) and *Lethrinops lethrinus* using General Linear Models and  
136  $\log_{10}$  transformed data. Bold indicates statistically significant differences between the species.  
137 \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ .  
138

Measurement	Slope		Elevation	
	$F_{1,51}$	$P$	$F_{1,52}$	$P$
Maximum body depth	0.07	0.794	<b>4.08</b>	<b>0.049*</b>
Head length	<b>4.55</b>	<b>0.038*</b>	0.62	0.435
Head width	2.09	0.155	1.41	0.240
Interorbital width	1.73	0.198	<b>10.80</b>	<b>0.002**</b>
Snout length	0.72	0.401	<b>15.11</b>	<b>&lt; 0.001***</b>
Lower jaw length	0.93	0.340	4.21	0.045
Premaxillary pedicel length	1.09	0.301	1.56	0.218
Eye diameter	1.02	0.317	1.79	0.187
Lachrymal depth	0.27	0.614	<b>17.69</b>	<b>&lt; 0.001***</b>
Dorsal fin base length	0.00	0.995	2.09	0.155
Anal fin base length	<b>5.87</b>	<b>0.019*</b>	0.87	0.354
Predorsal length	3.42	0.070	0.17	0.686
Preanal length	1.20	0.279	0.06	0.815
Prepectoral length	0.06	0.808	<b>5.41</b>	<b>0.024*</b>
Prepelvic length	1.08	0.303	2.51	0.119
Caudal peduncle length	<b>4.96</b>	<b>0.030*</b>	0.92	0.341
Caudal peduncle depth	2.48	0.122	<b>4.40</b>	<b>0.041*</b>

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142 **FIGURE 1.** Geometric morphometric analyses of *Lethrinus lethrinus* and *Lethrinus chilingali*  
143 **a.** Landmarks used to quantify shape variation of preserved specimen (see Materials and  
144 methods for details). **b.** Principal Component Analysis indicates strong separation of *L.*  
145 *lethrinus* and *L. chilingali* on PC1, with clear differentiation of the respective holotypes. **c.**  
146 Wireframe plots of mean body shapes of *L. lethrinus* (blue) and *L. chilingali* (orange), showing  
147 the more ventrally placed mouth, longer snout, and higher back in specimens of *L. lethrinus*  
148 relative to *L. chilingali*



149 ***Lethrinops chilingali* new species.**

150 **Holotype:** BMNH 2023.1.11.1, female, 70.9 mm SL, collected from seine catches, Lake  
151 Chilingali (12.94°S, 34.21°E), 22-24 June 2009.

152 **Paratypes:** BMNH 2023.1.11.2-21, twenty specimens 59.3-81.2 mm SL, collected with  
153 holotype.

154 **Other material (excluded from the type series):** BMNH 2023.1.11.22-28; seven specimens  
155 56.8-98.7mm SL, laboratory bred from specimens collected at Lake Chilingali

156 **Etymology:** ‘chilingali’ from Lake Chilingali, the type locality, used as a noun in apposition.

157 **Diagnosis:** The outer tooth row of the lower jaw curves smoothly to end just behind the inner  
158 tooth rows (*Lethrinops*-type dentition), distinguishing the species from other Lake Malawi  
159 haplochromines apart from species of the genera *Ctenopharynx*, *Lethrinops*, *Taeniolethrinops*  
160 or *Tramitichromis*. *Lethrinops chilingali* can be distinguished from other species in the genera  
161 *Ctenopharynx*, *Lethrinops*, *Taeniolethrinops* and *Tramitichromis* by the presence of a  
162 conspicuous horizontal series of dark grey to black spots along the middle of the flanks behind  
163 the head, linked to form one or two longer dashes, in total comprising 3-6 separate elements.  
164 *Lethrinops lethrinus* has a similar horizontal dark midlateral band, but it is typically more  
165 continuous, particularly posterior to the first anal spine, rather than broken into shorter spots  
166 and dashes. The horizontal melanic elements are generally not exhibited in dominant  
167 reproductively active males, however. *L. chilingali* also typically has a less ventrally placed  
168 mouth and shorter snout than *L. lethrinus* (snout as % of head length: 31.1-41.8 in *L. chilingali*,  
169 37.6-50.0 in *L. lethrinus*).

170 **Description.** Body measurements and counts are presented in Table 2. *L. chilingali* is a small  
171 (<85mm SL in wild-caught specimens) moderately laterally compressed (maximum body  
172 depth 2.0-2.4 times maximum head width) cichlid fish with a moderately long snout (31.1-41.8  
173 % head length). Females and immature males have distinctive melanic markings in the form of  
174 a horizontal row of dark spots and dashes (fig. 3b, d) and also have a thin red dorsal fin margin,  
175 while mature males are brilliant metallic green with a red dorsal fin margin above broader black  
176 and white bands (fig. 3f).

177 All specimens are relatively deep-bodied and laterally compressed, with the deepest part of the  
178 body generally well behind the first dorsal fin spine. The anterior upper lateral profile is almost  
179 straight from the tip of the snout to the plane of the posterior margin of the eye, occasionally  
180 with a slight concavity above the eye, gentle sloping at an angle of about 40-degrees to the  
181 horizontal plane. There is no inflection to the angle of the profile above the eye (in contrast to  
182 *Tramitichromis* and *Tropheops* Trewavas 1984) and the premaxillary pedicel makes little or no  
183 interruption to the profile. The tip of the snout lies at about the same level in a horizontal plane  
184 as the upper margin of the insertion of the pectoral fin and at or below the level of the lowermost  
185 margin of the eye. The lower anterior lateral profile is also almost straight as far as the insertion  
186 of the pelvic fins, gently angled to the horizontal plane (about 12-15-degrees) and with little  
187 inflection at the posterior angle of the lower jaw even when the mouth is fully closed. The  
188 lower profile is more or less horizontal between the pelvic and anal fins. The mouth is relatively  
189 small and moderately upwardly-angled (gape about 40-degrees to horizontal). The caudal  
190 peduncle is relatively slender (1.4-1.8 times longer than deep). The pectoral fins are relatively

191 long, extending past the first anal spine, but the pelvic fins are generally short of this, except  
192 in the largest mature males. The dorsal and anal fins, when folded, end well short of the caudal  
193 fin insertion, except in large mature males. The caudal fin is crescentic. The eye is large and  
194 circular and almost touches the upper lateral profile in perpendicular lateral view.

195 The flank scales are weakly ctenoid, with the ctenii becoming reduced dorsally, particularly  
196 anteriorly above the upper lateral line, where they transition into a cycloid state. The scales on  
197 the chest are relatively large and there is a gradual transition in size from the larger flank scales,  
198 as is typical in non-mbuna Lake Malawi endemic haplochromines (Eccles & Trewavas 1989).  
199 A few small scales are scattered on the proximal part of the caudal fin.

200 The cephalic lateral line pores are inconspicuous and the flank lateral line shows the usual  
201 cichlid pattern of separate upper and lower portions. The lachrymal bone is about as wide as  
202 deep and the lateral line pores are heavily overgrown with skin.

203 The lower jaw is relatively small, with thin mandibular bones. The jaw teeth are small, short  
204 and erect. The outer series in both the upper and lower jaw are short, blunt, erect and largely  
205 unequally bicuspid. There is a single inner series of small, pointed tricuspid teeth.

206 The lower pharyngeal bone (fig. 4a) is small, lightly built, Y-shaped, and carries small, slender,  
207 widely-spaced simple teeth, as illustrated for *L. lethrinus* by Eccles & Lewis (1978, figure 5).  
208 The teeth gradually increase in size from lateral to medial positions, but there are no distinctly  
209 differentiated enlarged medial teeth. There are approximately nine teeth in the midline row and  
210 17-18 on each side on the posterior row. The gill rakers are short and blunt, generally with the  
211 most anterior rakers in the lower and upper arches reduced to small stubs.

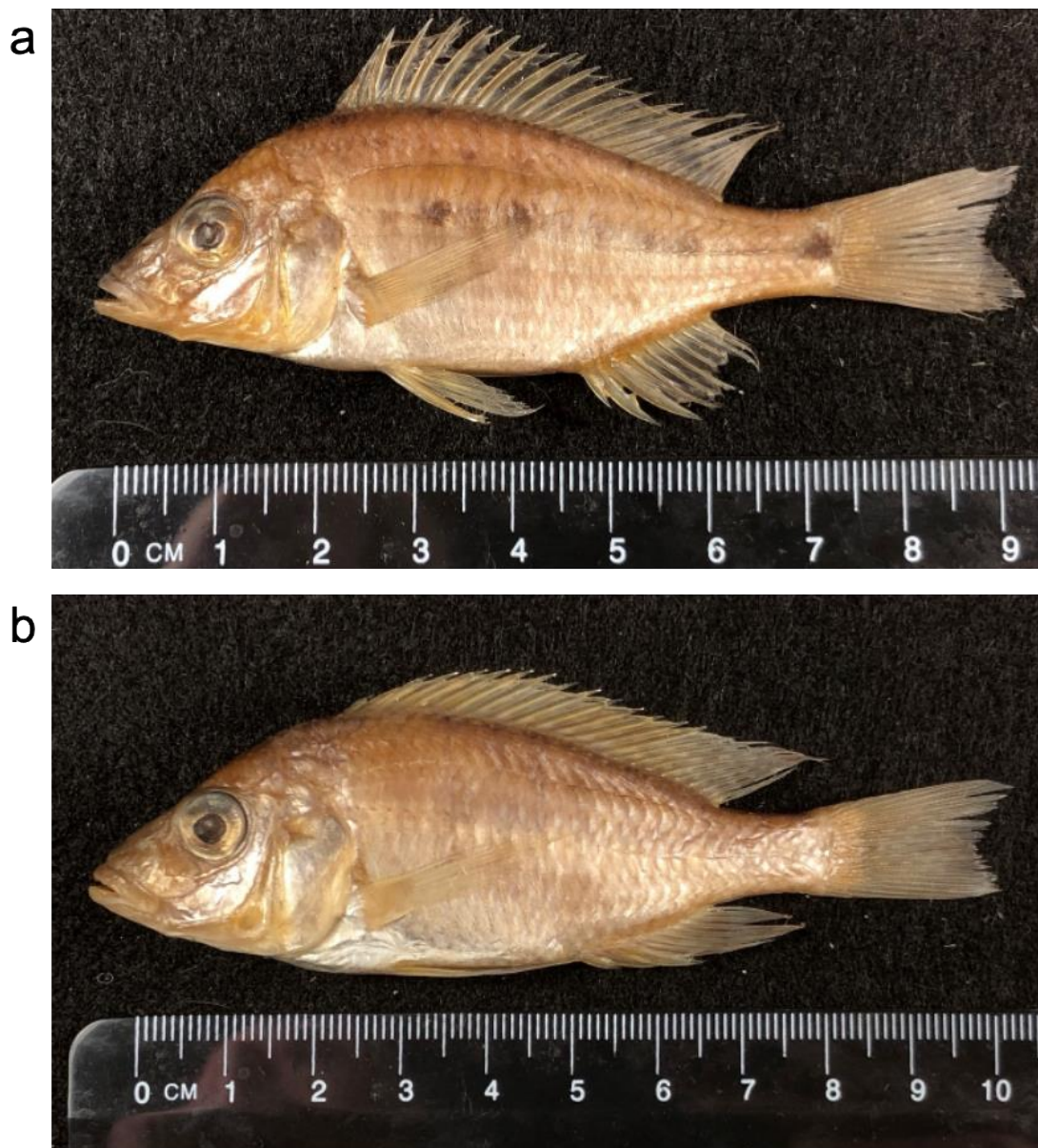
212 Female and immature fish (fig. 3d) are countershaded, pale sandy-brown dorsally, pale silvery  
213 on the flanks and underside. The flanks are marked by a midlateral horizontal row of dark spots  
214 and stripes extending from just behind the upper part of the operculum to the caudal peduncle.  
215 This varies between individuals, but generally comprises three to six separate melanic  
216 elements. A series up to six dark blotches is sometimes visible at the base of the dorsal fin, and  
217 element of a thin longitudinal dark stripe sometimes appears about half-way between the  
218 midlateral stripe and the base of the dorsal fin, usually starting a little behind the head and  
219 ending well before the caudal peduncle. The dorsal fin has a thin red outer margin and  
220 occasionally shows some faint dark spotting on both spinous and soft portions. Occasionally  
221 there is a pale submarginal band and anteriorly a thicker dark band. The caudal fin is usually  
222 translucent, sometimes with faint spotting. The anal fin sometimes shows a few faint yellowish  
223 spots.

224 Males in breeding dress (fig. 3f) are iridescent metallic green to pale blue. The horizontal  
225 melanic markings are occasionally exhibited when individuals are caught in fishing gear, or  
226 defeated in aggressive contests (seen in aquaria). Sometimes a series of faint dark vertical bars  
227 are visible. Patches of flank scales sometimes exhibit a metallic orange section anteriorly. The  
228 dorsal fin has a broad scarlet margin, underlain with a white submarginal band: these bands are  
229 narrower on the soft dorsal area. On the spinous dorsal, the red and white bands are underlain  
230 with a broad black band which extends to the base of the dorsal fin on the first inter-radial  
231 membrane, but as the membranes become longer posteriorly, the band overlies a series of  
232 orange spots extending onto the soft dorsal area, where they can be up to 10 spots between the  
233 longest rays. The membranes between the spots are pale grey to white. The caudal fin continues

234 this pattern of orange spots with white/grey areas between. Sometimes the white inter-spot  
235 areas are missing, resulting in spots merging into stripes parallel to the fin rays. Occasionally,  
236 the white areas merge into stripes too. The upper and lower parts of the caudal fin can  
237 sometimes appear a bit darker, particularly on the basal section closer to the body, and  
238 particularly during dominant/courting behaviour. The pelvic fins are dark grey to black with a  
239 thin white anterior edge. The anal fin is greyish to black depending on mood, with a wide pink  
240 to red lower margin. A variable number (4-18) of large pale yellow 'egg-spots' are visible in  
241 one to two rows on the membranes behind the third anal spine. The colour of the iris varies  
242 from silvery to dark gold, with a darker spot above and below the lens continuing the line of a  
243 dark lachrymal stripe from the corner of the mouth. This stripe is very variable in intensity,  
244 showing up very prominently during territorial defence and courtship phases. The lower surface  
245 of the head and chest can turn dark grey during courtship and territorial behaviour but is  
246 otherwise pale greyish.

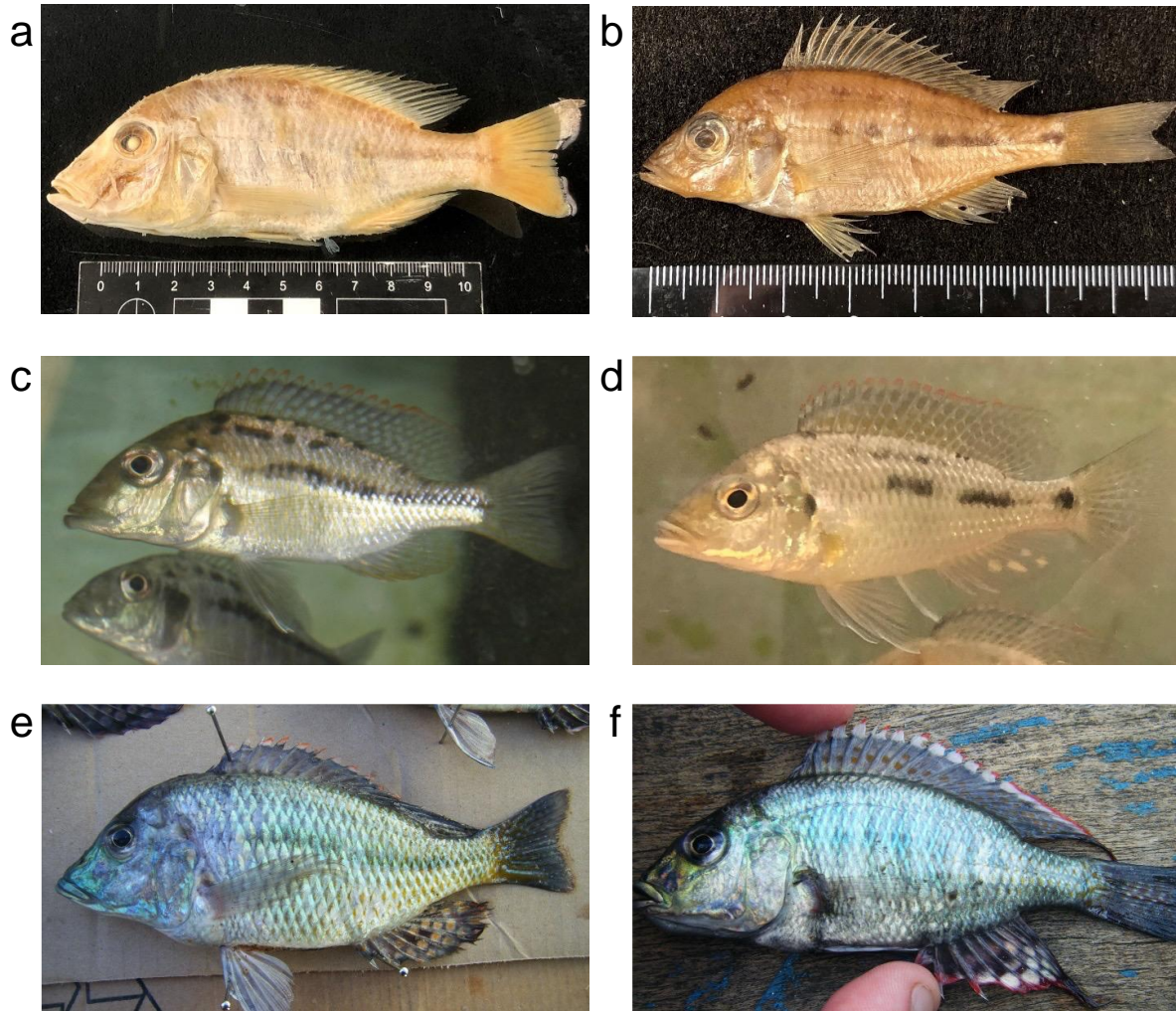
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249 **FIGURE 2.** *Lethrinops chilingali*. **a.** Holotype, BMNH 2023.1.11.1; female 70.9mm SL. **b.**  
250 Paratype, BMNH 2023.1.11.2-21; mature male, 81.2mm SL.



251

252 **FIGURE 3.** Comparisons of *Lethrinops lethrinus* and *Lethinops chilingali*. **a.** holotype of *L.*  
253 *lethrinus*, BMNH 1893.15.15., 118.5mm SL. **b.** paratype of *L. chilingali*, BMNH 2023.1.11.2-  
254 21, female, 60.7mm SL; **c.** *L. lethrinus* apparent female alive in aquarium. **d.** *L. chilingali*  
255 apparent immature male alive in aquarium. **e.** mature male *L. lethrinus*. **f.** mature male *L.*  
256 *chilingali*. The shorter snout *L. chilingali* is evident, and the more broken midlateral stripe can  
257 be seen in the live specimens.

258

259

260 **TABLE 2.** Morphometric and meristic characters of *Lethrinops chilingali*.

	<b>Holotype</b>	<b>Paratypes (n=20) mean (range)</b>	<b>Captive strain (n=7) mean (range)</b>
Standard length (mm)	70.9	65.7 (59.3-81.2)	80.4 (56.8-98.7)
<b>As % Standard length</b>			
Maximum body depth	36.2	35.2 (33.1-36.8)	34.1 (31.1-36.7)
Head length	34.4	33.6 (32.1-35.9)	35.9 (34.7-38.9)
Dorsal fin base length	53.9	53.0 (51.0-55.7)	53.0 (50.8-56.7)
Anal fin base length	18.8	19.6 (16.9-21.5)	18.0 (17.1-18.8)
Predorsal length	39.2	37.5 (35.0-39.3)	39.1 (35.2-42.5)
Preanal length	65.3	64.1 (62.5-66.5)	64.6 (61.1-67.8)
Prepectoral length	36.4	35.5 (33.5-38.0)	36.1 (33.8-38.0)
Prepelvic length	40.2	39.9 (37.1-43.1)	41.5 (38.2-44.1)
Caudal peduncle length	19.2	17.9 (16.1-20.0)	17.2 (16.1-20.4)
Caudal peduncle depth	11.0	11.5 (10.4-12.4)	11.4 (10.9-12.3)
<b>As % Head length</b>			
Head width	47.1	45.6 (40.9-50.0)	43.7 (40.4-47.4)
Interorbital width	21.1	21.8 (18.8-24.5)	22.7 (20.4-27.2)
Snout length	33.3	35.2 (31.1-38.2)	38.7 (34.6-41.8)
Lower jaw length	40.9	39.2 (35.3-42.9)	39.2 (37.3-44.2)
Premaxillary pedicel length	29.8	29.7 (25.7-35.9)	30.0 (24.9-35.4)
Eye diameter	31.1	31.8 (28.2-37.7)	29.1 (25.7-33.0)
Lachrymal depth	21.5	21.4 (18.0-25.9)	23.8 (21.0-27.7)
<b>Ratios</b>			
Body depth/Head width	2.25	2.30 (2.11-2.41)	2.18 (1.99-2.34)
Caudal peduncle length/depth	1.74	1.56 (1.37-1.80)	1.51 (1.37-1.76)
<b>Counts</b>			
Upper gill rakers	3	3-4	3-4
Lower gill rakers	10	9-11	10-12
Dorsal fin	XV, 10	XIV-XVI, 9-10	XIV-XV, 10-11
Anal fin	III, 8	III, 8-10	III, 8-9
Longitudinal line scales	31	31-33	30-33
Cheek scales	3	2-4	2-4

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263 **Distribution.** Known only from Lake Chilingali in the Lake Malawi catchment (fig. 4b).

264 **Behaviour and Ecology.** The diet of *L. chilingali* specimens sampled in 2009 consisted largely  
265 of chaoborid (midge) larvae and pupae, along with cladocerans and other larger invertebrates,  
266 including odonatan nymphs and caridinid shrimps, but with little detritus, perhaps suggesting  
267 more midwater feeding than is usual in *Lethrinops* species. The behaviour of the species in the  
268 wild has not been observed, as the water of Lake Chilingali was highly turbid when visited  
269 between 2004 and 2009.

270 In captivity, *L. chilingali* females, non-territorial males and juveniles tend to aggregate in loose  
271 groups, feeding not only in the sediment, but on objects such as rocks or plants, or even at the  
272 surface. When attempts are made to catch the fish, they show a strong tendency to dive into the  
273 sand, turning sideways and completely burying themselves. This same behaviour has been  
274 reported to occur in the wild in *Fossorochromis rostratus* (Boulenger 1899), another cichlid  
275 from the Lake Malawi radiation (Fryer & Iles 1972, p. 207).

276 Dominant male *L. chilingali* are territorial and actively court females in typical haplochromine  
277 style: fins wide open, body horizontal or head-up, making rapid darts to the spawning site and  
278 back to the female, with spawning taking place amid bouts of circling and quivering, while  
279 alternating head-to-anal-fin ‘T-positions’ on the substrate. It is notable that dominant male  
280 coloration and aggression vary a lot, appearing to peak when females are approaching  
281 spawning, but are otherwise often quite subdued. During persistent bouts of courtship or  
282 aggression, the melanic elements of the male colour are emphasised, particularly the  
283 lachrymal/eye stripe, dark pelvic and anal fins, dark upper and lower margins of the caudal fin  
284 and even faint vertical barring on the flanks. Even in a large tank with a high density of fish,  
285 there is usually just a single dominant male: this is similar to *Astatotilapia* Pellegrin 1904,  
286 which tend to be solitary breeders. Communal lek breeders, such as *Oreochromis* Günther 1889  
287 will usually divide up a tank into numerous smaller territories and engage in frequent boundary  
288 disputes. This suggests that *Lethrinops chilingali* are not communal lek breeders in the wild.

289 There is little indication of bower construction in *L. chilingali* when a sand or gravel substrate  
290 is provided: dominant males usually try to lead females to a slight depression near to an object  
291 such as a rock or piece of wood: in a bare tank, the focus would probably be the tank bottom  
292 near one of the corners or a wall near a heater or filter inlet. This is in marked contrast to reports  
293 of *L. lethrinus* where complex bowers have been recorded in the field, out over open substrate  
294 (Konings 2016, p. 369). In *L. chilingali*, the construction of the depression seems almost  
295 haphazard: males have not been observed to show consistent bouts of digging, but spend most  
296 of their time chasing, then returning to the territory focus next to the object, during which they  
297 make occasional ‘feeding movement’ of picking up a mouthful of substrate, moving forwards  
298 and ejecting it through the mouth and/or opercular openings at a slight distance away. This  
299 occurs all over the vicinity of the side of the object they are defending, but there seems to be a  
300 slight bias towards a certain point up against the object, which thereby becomes a shallow  
301 depression.

302 Female *L. chilingali* are maternal mouthbrooders, brooding young until they are capable of  
303 independent feeding. As fry complete the absorption of the yolk, they show through the  
304 female’s buccal membrane as a dark area, but females do not develop the ‘warpaint’ typical of  
305 fry guarders, such as known *Astatotilapia* or *Oreochromis*: dark eyes, lachrymal stripes and  
306 forehead stripes. There is no indication that females guard free-swimming fry or permit them

307 to return to their mouths. This non-guarding behaviour is similar to other known shallow-water  
308 *Lethrinops* species.

309

310 ***Lethrinops lethrinus* (Günther, 1893)**

311 **Holotype:** *Lethrinops lethrinus* (Günther, 1893): BMNH 1893.11.15.15, 116.1 mm SL, coll.  
312 A. Whyte, Upper Shire River at Fort Johnston (Mangochi), March 1892,

313 **Other material examined:**

314 BMNH 2023.1.11.29, 1 specimen 130.1mm SL, collected by G.F. Turner from experimental  
315 trawl at depth of 5-18m, between Namiasi and Palm Beach (approximately 14.38°S, 35.22°E),  
316 SE Arm of Lake Malawi, 30 July 1991.

317 BMNH 2023.1.11.30, 1 specimen, 120.6 mm SL, collected by G.F. Turner, trawled at 5-18m  
318 depth between Namiasi and Malindi (approximately 14.34°S, 35.22°E), SE Arm of Lake  
319 Malawi, 30<sup>th</sup> July 1991.

320 BMNH 2023.1.11.31, 1 specimen, 101.4mm SL, collected by G.F. Turner from kambuzi seine  
321 fisherman, West shore of Lake Malombe, probably at Chimwala (14.64°S, 35.18°E), 26 June  
322 1992,

323 BMNH 2023.1.11.32-36, 5 specimens, 63.2-66.6 mm SL, collected by G.F. Turner from Lake  
324 Malombe, probably at Chimwala (14.64°S, 35.18°E), 23 July 1992.

325 BMNH 2023.1.11.37, 1 specimen 90.2 mm SL, collected by G. F. Turner, Middle Shire River,  
326 probably at Liwonde Barrage (15.06°S, 35.22°E), 20<sup>th</sup> May 1992.

327 BMNH 2023.1.11.38-43, 6 specimens 129.2-152.6 mm SL, collected by G. F. Turner  
328 unspecified sites in southern Lake Malawi, 1990-1992.

329 BMNH 2023.1.11.44-46, 3 specimens 97.9-116.1 mm SL, collected by G. F. Turner, trawled  
330 at 18-21m at Ulande 1a station (14.23°S, 35.21°E), SE Arm Lake Malawi, 1991.

331 BMNH 2023.1.11.47-48, 2 specimens 106.4-130.2 mm SL, collected by David Bavin, from  
332 seine fishermen, Lake Malombe (14.64°S, 35.18°E), 6<sup>th</sup> July 2009.

333 BMNH 2023.1.11.49-50, 2 specimens 121.7-128.0 mm SL, collected by G. F. Turner, trawled  
334 at 26m depth at Michesi station (14.32°S, 35.19°E), SE Arm of Lake Malawi, 1992.

335 BMNH 2023.1.11.51-53, 3 specimens 109.4-123.2 mm SL, collected by G. F. Turner, from  
336 seine net fishermen, Palm Beach (14.41°S, 35.23°E), SE Arm of Lake Malawi, 23 Jan 2017.

337 BMNH 2023.1.11.54, 1 specimen 120.7 mm SL, collected by G. F. Turner, from seine net  
338 fishermen, Palm Beach (14.41°S, 35.23°E), SE Arm of Lake Malawi, 22 Jan 2017.

339 **Remarks:** *L. lethrinus* was selected as the type of the genus *Lethrinops* by Regan (1922). It  
340 was originally described as *Chromis lethrinus* from a single specimen, but was redescribed  
341 from additional material by Regan (1922), Trewavas (1931), Eccles & Lewis (1978) and Eccles  
342 & Trewavas (1989). It was also included in a key to the shallow-water *Lethrinops* species by  
343 Ngatunga & Snoeks (2004). The original illustration in Günther (1893) shows a specimen with  
344 a continuous horizontal midlateral stripe beginning at the eye and extending to the base of the

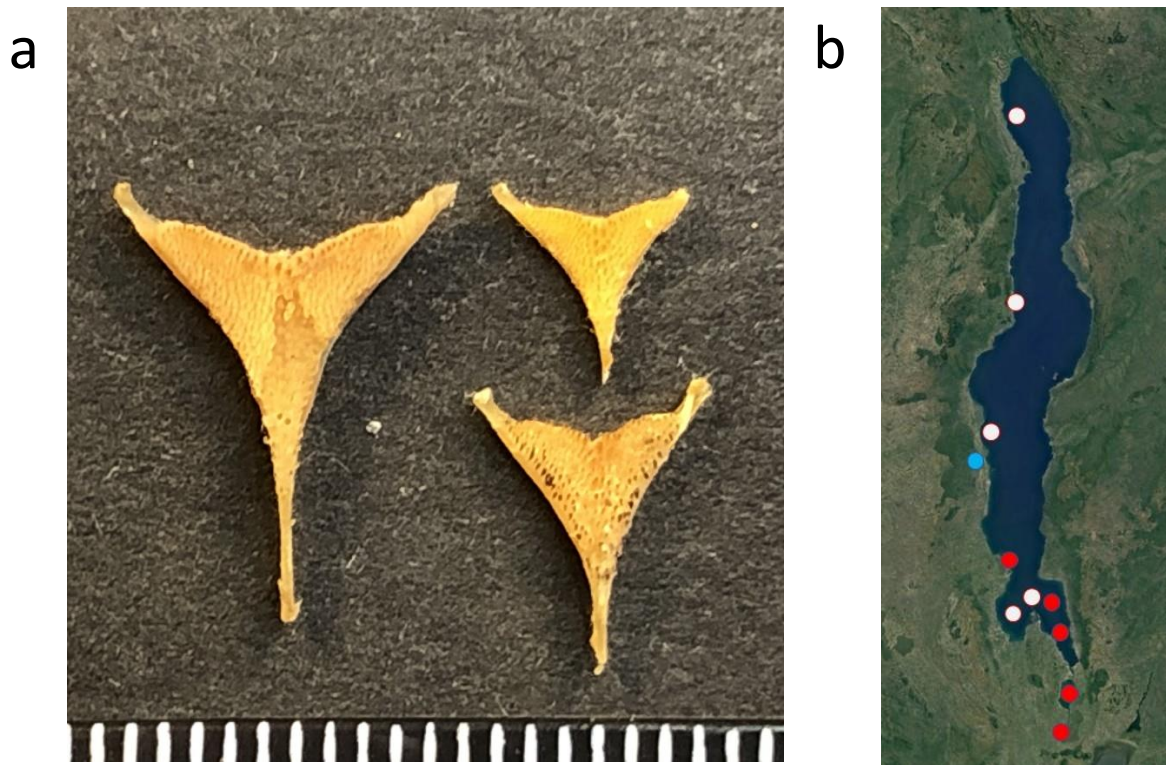
345 caudal fin. This is reprinted in Eccles & Trewavas (1989), where the imaged specimen is  
346 erroneously referred to as the lectotype (it is the holotype). The redescription by Eccles &  
347 Lewis (1978) includes a drawing of a non-type specimen in which the horizontal midlateral  
348 stripe is composed of a series of about 15 spots running from just behind the origin of the pelvic  
349 fin to the base of the caudal fin. Anteriorly, the first five spots are separate, but the gaps between  
350 them are much narrower than the length of the spots. Posteriorly, all of the spots overlap, to  
351 form a continuous, albeit irregular, blotchy line. Eccles & Lewis (1978) stated they examined  
352 (but did not measure) the type and there seems little doubt that the non-type material they  
353 studied (uncatalogued, Monkey Bay Fisheries Research Unit, Malawi, status unknown)  
354 corresponds to this species.

355 *Lethrinops lethrinus* is readily diagnosed based on its typical *Lethrinops*-type dentition,  
356 horizontal melanic flank markings and long snout. Mature males show a metallic blue-green  
357 breeding dress, with a prominent red and white dorsal fin margin and numerous large eggspots  
358 on the anal fin (Figure 3, see also Konings 2016). *L. lethrinus* appears to be confined to shallow  
359 waters with muddy bottoms, often river mouths with extensive beds of reeds and other  
360 macrophytes, feeding on invertebrates and other edible material obtained from the sediment  
361 (Turner 1996). Konings (2016) reports a lakewide distribution and it has been recorded from  
362 Lake Malombe and the Upper and Middle Shire Rivers (Turner 1996), but records from Domira  
363 Bay northwards are based on juveniles that are hard to distinguish from *L. chilingali* or lack  
364 available voucher material (fig. 4b). Counts and measurements of the material we examined  
365 are presented on Table 3.

366



367



368

369 **FIGURE 4.** a. Lower pharyngeal bones of *Lethrinops lethrinus*, 128mm SL, BMNH  
370 2023.1.11.49-50 (left); *Lethrinops lethrinus* 84mm SL (unregistered, bottom right); *Lethrinops*  
371 *chilingali* 69mmSL (unregistered, top right); b. Distribution of *Lethrinops lethrinus* specimens  
372 examined (●), unconfirmed records: juveniles or not examined (○) and *Lethrinops chilingali*  
373 (●).

374 **TABLE 3.** Morphometric and meristic characters of *Lethrinops lethrinus*.

	<b>Holotype</b>	<b>Non-types (n=26) mean (range)</b>
Standard Length (mm)	118.5	110.7 (62.9-152.6)
<b>As % Standard length</b>		
Maximum body depth	36.4	37.2 (33.0-41.0)
Head length	34.5	35.1 (33.1-39.1)
Dorsal fin base length	53.9	53.4 (49.9-56.2)
Anal fin base length	17.7	18.7 (16.5-21.0)
Predorsal length	37.7	39.5 (37.1-42.3)
Preanal length	66.2	64.9 (61.5-68.6)
Prepectoral length	35.1	37.1 (33.9-40.1)
Prepelvic length	42.5	42.2 (35.7-46.4)
Caudal peduncle length	17.9	17.5 (14.7-20.2)
Caudal peduncle depth	12.5	12.1 (10.8-13.4)
<b>As % Head length</b>		
Head width	46.2	44.8 (41.0-50.1)
Interorbital width	24.2	22.6 (18.0-26.9)
Snout length	42.5	44.4 (37.6-50.0)
Lower jaw length	40.1	41.0 (37.0-43.5)
Premaxillary pedicel length	31.1	31.0 (25.4-34.3)
Eye diameter	29.5	28.4 (25.2-34.7)
Lachrymal depth	29.5	30.7 (21.6-34.8)
<b>Ratios</b>		
Body depth/Head width	2.29	2.36 (2.14-2.67)
Caudal peduncle length/depth	1.43	1.45 (1.17-1.66)
<b>Counts</b>		
Upper gill rakers	3	2-4
Lower gill rakers	9	8-10
Dorsal fin	XV, 11	XIV-XVI, 8-12
Anal fin	III, 9	III, 8-9
Longitudinal line scales	31	30-36
Cheek scales	3	3-4

375

376



377

378 **FIGURE 5.** Comparative material. **a.** Three small specimens (BMNH 1935.6.14.2077-9) from  
379 Lupembe in northern Lake Malawi match *Lethrinops lethrinus*, in melanin pattern and low  
380 position of mouth on the head. **b.** A syntype of *Lethrinops leptodon* BMNH 1921.9.6.201-207,  
381 showing two oblique stripes thickened and fused together to form a midlateral blotch. This  
382 pattern is distinguishable from those of *L. chilingali* and *L. lethrinus*, but is similar to the  
383 Nkhata Bay population reported by Eccles & Lewis (1978) and assigned by them to *L.*  
384 *lethrinus*.

385

386



## 387 4. DISCUSSION

388

### 389 Relationship of *L. chilingali* to other taxa in the Lake Malawi radiation

390 The present study has assumed that *L. lethrinus* is both the most likely sister taxon for *L.*  
391 *chilingali* and the species most likely to interbreed with it, should habitat barriers be broken  
392 down. The former proposition is based on their overall similar appearance, including very  
393 similar male breeding dress, and similar – although distinct- melanin patterns in the females  
394 and juveniles. They are the only two known *Lethrinops* species to share a largely horizontally-  
395 banded melanin pattern. Other Lake Malawi cichlids also share some of these features, notably  
396 species of *Protomelas* Eccles & Trewavas 1989 found in similar shallow weedy/muddy  
397 habitats, including *Protomelas kirkii* (Günther 1894), *Protomelas similis* (Regan 1922) and  
398 *Protomelas labridens* (Trewavas 1935) (Eccles & Trewavas 1989, Konings 2016, Turner  
399 1996). These three species also have females/immatures with a sandy/countershaded  
400 appearance, with a strong horizontal dark band running along the flank. Males are also metallic  
401 blue-green, with a red and white dorsal fin margin. These species have shorter snouts and more  
402 upwardly-angled mouths than *L. lethrinus*, but so does *L. chilingali*, which is arguably  
403 morphologically intermediate between them. The genera *Protomelas* and *Lethrinops* can be  
404 distinguished by the shape of the lower jaw dental arcade, and it is presently assumed that this  
405 is a phylogenetically informative trait (Eccles & Trewavas 1989), although this requires  
406 confirmation from a phylogenetic analysis, ideally based on genome-scale sequence data. A  
407 published phylogenomic analysis places *L. lethrinus* in the middle of a clade of shallow water  
408 *Lethrinops*, *Taeniolethrinops* and *Tramitichromis* (Masonick et al. 2022), thus grouping these  
409 genera showing *Lethrinops*-type dentition (Eccles & Trewavas 1989). However, *P. kirkii*, *P.*  
410 *similis* and *P. labridens* were not included in that analysis (Masonick et al. 2022). Notably,  
411 however, an additional group of deep-water *Lethrinops* appears in a separate part of the  
412 phylogenetic tree, suggesting that the *Lethrinops*-type dentition is prone to parallelism. Thus,  
413 we conclude that available evidence does not conflict with *L. chilingali* being a sister species  
414 to *L. lethrinus*, but this requires detailed phylogenetic investigation for confirmation. If *L.*  
415 *lethrinus* shows relatively high levels of population structure, it could be paraphyletic  
416 (ancestral) with respect to *L. chilingali*.

417

### 418 Distributions of *L. chilingali* and *L. lethrinus*

419 *Lethrinops chilingali* has only been positively recorded from Lake Chilingali, but here we  
420 consider whether it may have a broader distribution in Lake Malawi, possibly extending to the  
421 central to northern part of the lake as an allopatric sister species to *L. lethrinus*. Although a  
422 lake-wide distribution has been claimed for *L. lethrinus* (Konings 2016), the great majority of  
423 records backed by preserved specimens or photographs come from the southern arms, Lake  
424 Malombe and the Shire River (Eccles & Lewis 1978, Turner 1996, Konings 2016). On the  
425 Global Biodiversity Information Facility website (GBIF 2023), there is a record of *Lethrinops*  
426 *lethrinus* from co-ordinates indicating a collection site off the Tanzanian shore near Ngkuyo  
427 Island, Mbamba Bay (11.334°S, 34.769°E), based on specimens at the South African Institute  
428 for Aquatic Biodiversity (SAIAB). An offshore location near a rocky headland seems an  
429 unlikely collecting site for *Lethrinops lethrinus*, which favours shallow sheltered vegetated

430 habitats and the locality label is given as ‘Lifuwu’, which probably corresponds to the vicinity  
431 of Lifuwu village (13.69°S, 34.60°E) just north of Salima, suggesting that the co-ordinates  
432 have been recorded in error. The single small specimen shows no melanic markings (faded  
433 post-preservation?), but the head shape is consistent with *Lethrinops lethrinus* rather than *L.*  
434 *chilingali*. Another GBIF record from co-ordinates 13.35°S, 33.4°E would suggest specimens  
435 were collected from the Rusa River, a tributary of the Bua River, which joins Lake Malawi  
436 near Lake Chilingali. The site is far upstream, around 97km West of the Lake Malawi shore at  
437 Benga, and initially we thought this might suggest specimens of *L. chilingali* could be  
438 widespread in the river catchment. However, the collection label indicates the specimens were  
439 collected from Lake Malawi at Foo, which is a trawling station in the SE Arm of Lake Malawi  
440 (also sometimes written as Fowo), which is at approximately 14.14°S, 35.18°E, again  
441 suggesting an error in the co-ordinates. Photographs of the specimens show typical *Lethrinops*  
442 *lethrinus*, with long snouts and strong horizontal melanic markings. The catalogue of the  
443 Natural History Museum in London contains a single accession of three specimens labelled as  
444 *L. lethrinus* from Lupembe Sand Bar, collected by Cuthbert Christy in 1925 (BMNH  
445 1935.6.14.2077-9; Figure 4). The electronic catalogue suggests that this location is in Tanzania,  
446 perhaps following Eccles & Trewavas (1989) who suggested it might represent a site at the  
447 mouth of the ‘Kivira River’. However, the town at the mouth of the Kiwira River (as presently  
448 named) is currently known as Itungi Port. It is more likely that the 1925 collection site is  
449 Lupembe on the Malawian lakeshore, just south of Karonga (10.055°S, 33.99°E). Notably,  
450 recent satellite images show a conspicuous sandbar (Google Earth). Examination of the  
451 unpublished diary of Cuthbert Christy held at the Natural History Museum shows a single  
452 accession from Lupembe following an extensive collection of several hundred accessions from  
453 Vua / Deep Bay (Chilumba area) and immediately before another extensive collection from  
454 Mwaya in Tanzania, on the far north coast of the lake (itemising various river mouths visited).  
455 No other accessions were made from Lupembe. This suggests that the location was visited en-  
456 route from Chilumba to Tanzania, which would fit well with the location near Karonga.  
457 Unfortunately, the specimens (fig. 5a) are very small (44.8-50.9 mm SL) which makes  
458 morphological comparisons difficult with the larger specimens examined for this study, due to  
459 allometric effects. For example, they have notably relatively large eyes, making snout  
460 measurements relatively small. However, the low position of the mouth on the head and the  
461 largely continuous midlateral stripe, fit far better with *L. lethrinus* than with *L. chilingali*. Thus,  
462 available museum specimens strongly support the occurrence of typical *Lethrinops lethrinus*  
463 only in the southern arms of the lake, but tentatively indicate that they may also occur just north  
464 of Senga Bay and indeed almost at the northernmost extremity of the lake, but do not provide  
465 evidence for the occurrence of *L. chilingali* or any other similar form within Lake Malawi,

466 Other published records are not backed by specimens available to examine or photographic  
467 evidence. Eccles & Lewis (1978) reported that they had found *L. lethrinus* at Nkhata Bay,  
468 which is well to the north of Lake Chilingali. However, they reported an unusual melanin  
469 pattern: “the dark line along the middle of the flank curves upwards anteriorly to merge with  
470 the lower of the two rows of spots and the spots themselves may run together posteriorly to  
471 form a stripe”. The occurrence of specimens with dramatically different stripe patterns at  
472 Nkhata Bay might lend credence to the idea that *L. lethrinus* represents a complex of allopatric  
473 taxa, which might increase the probability that *L. chilingali* might persist in the main Lake  
474 Malawi. Eccles & Lewis provided no illustration of this ‘Nkhata Bay variant’. Their specimens  
475 were deposited in the collection of the Monkey Bay Fisheries Research Unit, Malawi and their

476 present status is unknown. The pattern described is reminiscent of that seen on some of the  
477 type specimens of *L. leptodon* Regan 1922 (fig. 5b). In the same 1978 paper, Eccles & Lewis  
478 redescribed that species based on a single specimen collected from Chintheche in the NW of  
479 the lake, near Nkhata Bay, but their illustration of that specimen showed a clear midlateral  
480 blotch on the upper part of the flank. They reported examining, but not measuring, three of the  
481 type specimens of *L. leptodon*, which are held at the Natural History Museum in London  
482 (BMNH 1921.9.6.201-207, collected by Wood from somewhere in ‘Lake Nyasa’). Thus, it  
483 seems unclear whether the reported Nkhata Bay populations represent *L. lethrinus* or *L.*  
484 *leptodon*, or indeed something else. In summary, the status of the northern populations of  
485 *Lethrinops* of this group is unclear but is consistent with the hypothesis that *L. lethrinus* is  
486 found in suitable habitats throughout Lake Malawi, and that *L. chilingali* is a satellite lake  
487 endemic extinct in the wild.

#### 488 **Conservation status of *Lethrinops chilingali***

489 Lake Chilingali is approximately 5km in length and a maximum of 1km in width, and is  
490 characterised by two deeper basins of approximately 5m depth separated by a shallower  
491 plateau (Turner et al. 2019). It has a single outflow, the Kaombe River, which meanders for  
492 approximately 12km before reaching the main body of Lake Malawi (Genner et al. 2007).  
493 The lake is a natural water body, and the two basins of the modern lake are represented on  
494 early European exploration maps, as two separate bodies of water, Lake Chikukutu to the  
495 south, and Lake Chilingali to the north (Turner et al. 2019). The lake level was raised when a  
496 dam was constructed across the single outflow for irrigation purposes, initially in the 1950s,  
497 before being modified in the early 1970s (Denys et al. 2013). The dam collapsed early in  
498 2012 (Denys et al. 2013), and the single lake disappeared, reforming the two separate smaller  
499 shallow basins. In 2016 these basins were estimated to be only ~1m deep and fringed with  
500 macrophytes. The lake was refilled to approximately its pre-collapse-level in June-July 2019  
501 following the construction of a new dam.

502 During the period 2004 to 2011, before the collapse of the dam, *L. chilingali* was periodically  
503 and reliably sampled from the lake, alongside another apparently endemic haplochromine  
504 cichlid, the undescribed *Rhamphochromis* sp. “chilingali” (Genner et al. 2007; Turner et al.  
505 2019). To our knowledge, the last sampling event where *L. chilingali* was recorded in the  
506 wild was on 25 June 2009 (by G. Turner), while representatives of *R. sp.* “chilingali” were  
507 last collected from an artisanal fishing catch on 12 January 2011 (by M. Genner). During  
508 sampling in February 2016, neither of the species was encountered in a survey of the main  
509 northern and southern basins of Lake Chilingali (Turner et al. 2019). A survey in April 2022  
510 also failed to sample any either *L. chilingali* or *R. sp.* “chilingali” but did find that Lake  
511 Malawi endemic *Otopharynx tetrastigma* (Günther 1894) was abundant (H. Svardal, pers  
512 comm). This species was absent between 2004 and 2016 and is likely to have been introduced  
513 during restocking after the lake was refilled in 2019 (H. Svardal, pers comm). Although  
514 further surveys of Lake Chilingali and the Kaombe river are warranted to determine if  
515 remnant populations of either *L. chilingali* or *R. sp.* “chilingali” persist, on the basis of the  
516 current evidence, we consider it most likely that both species are no longer present in the  
517 natural environment. Breeding populations of *L. chilingali* or *R. sp.* “chilingali” are, however,  
518 maintained in captivity, and may be candidates for reintroduction. On the basis of the  
519 evidence discussed above, we recommend that *L. chilingali* is attributed the status of Extinct



520 in the Wild (EW) on the International Union for Conservation of Nature (IUCN) Red List of  
521 Threatened Species.

522

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535

## 536 **REFERENCES**

- 537 Denys, F. J. M., Hagen, D. J., Stephens, T., Brown, R. & Namukopwe V. (2013). Lake  
538 Chilingali Dam Rehabilitation. Presentation at SANCOLD Conference November 2013.  
539 Available at: <https://tinyurl.com/5epzav8z>  
540
- 541 Eccles, D. H. & Lewis, D. S. C. (1978). A taxonomic study of the genus *Lethrinops* Part 2.  
542 *Ichthyological Bulletin of Rhodes University*, 37, 1-11.  
543
- 544 Eccles, D. H. & Trewavas, E. (1989). *Malawian cichlid fishes. The classification of some*  
545 *Haplochromine genera*. Lake Fish Movies, Herten, Germany, 335 pp.  
546
- 547 Fryer, G. (1959). The trophic interrelationships and ecology of some littoral communities of  
548 Lake Nyasa with especial reference to the fishes, and a discussion of the evolution of a group  
549 of rock-frequenting Cichlidae. *Proceedings of the Zoological Society of London*, 132, 153-281.  
550
- 551 Fryer, G. A. & Iles, T. D. (1972). *The Cichlid Fishes of the Great Lakes of Africa*. Oliver &  
552 Boyd, Edinburgh.  
553
- 554 GBIF (2023) Global Biodiversity Information Facility. [www.gbif.org](http://www.gbif.org) (last accessed 12 March  
555 2023)  
556
- 557 Genner, M. J., Nichols, P., Carvalho, G. R., Robinson, R. L., Shaw, P. W., Smith, A. & Turner,  
558 G. F. (2007) Evolution of a cichlid fish in a Lake Malawi satellite lake. *Proceedings of the*  
559 *Royal Society of London, Series B*, 274, 2249-2257  
560
- 561 Greenwood, P. H. (1965) The cichlid fishes of Lake Nabugabo, Uganda. *Bulletin of the British*  
562 *Museum (Natural History), Zoology*, 12, 315–357.  
563

- 564 Günther, A. (1894) Second report on the reptiles, batrachians, and fishes transmitted by Mr. H.  
565 H. Johnston, C. B., from British Central Africa. *Proceedings of the Zoological Society of*  
566 *London* 1894, 616-628, Pls. 53-57  
567
- 568 Kaufman, L., & Ochumba, P. (1993) Evolutionary and conservation biology of cichlid fishes  
569 as revealed by faunal remnants in northern Lake Victoria. *Conservation Biology*, 7, 719– 730.  
570
- 571 Konings, A. (2016) *Lake Malawi Cichlids in their Natural Habitat. 5<sup>th</sup> Edn.* Cichlid Press, El  
572 Paso TX.  
573
- 574 Malinsky, M., Svardal, H., Tyers, A. M., Miska, E. A., Genner, M. J., Turner, G. F. &  
575 Durbin, R. (2018) Whole-genome sequences of Malawi cichlids reveal multiple radiations  
576 interconnected by gene flow. *Nature Ecology and Evolution*, 2, 1940-1955.  
577
- 578 Masonick, P., Meyer, A. & Hulsey, C. D. (2022) Phylogenomic analyses show repeated  
579 evolution of hypertrophied lips among Lake Malawi Cichlid Fishes. *Genome Biology &*  
580 *Evolution*, 14, evac051.  
581
- 582 Mwanja, W. W., Armodlian, A. S., Wandera, S. B., Kaufman, L., Wu, L., Booton, G. C., &  
583 Fuerst, P. A. (2001) The bounty of minor lakes: the role of small water bodies in evolution and  
584 conservation of fishes in the Lake Victoria Region, East Africa. *Hydrobiologia*, 458, 55–62  
585
- 586 Ngatunga, B. P. & Snoeks, J. (2004). Key to the shallow water *Lethrinops sensu lato*. In:  
587 Snoeks, J. (ed) *The Cichlid Diversity of Lake Malawi/Nyasa/Niassa: Identification,*  
588 *Distribution and Taxonomy.* Cichlid Press, El Paso, Texas, 252-260.  
589
- 590 Palmer, R. (2019) <https://www.inaturalist.org/observations/22139001>  
591
- 592 Regan, C. T. (1922). The cichlid fishes of Lake Nyasa. *Proceedings of the Zoological Society*  
593 *of London*, 1921, 675-727.  
594
- 595 Rohlf, F. J. (2015) The tps series of software. *Hystrix - The Italian Journal of Mammalogy*, 26,  
596 9-12.  
597
- 598 Snoeks, J. (2004) Materials and Methods. In Snoeks, J. (ed) *The Cichlid Diversity of Lake*  
599 *Malawi/Nyasa/Niassa: Identification, Distribution and Taxonomy.* Cichlid Press, El Paso, TX:  
600 12-19.  
601
- 602 Trewavas, E. (1931). A revision of the cichlid fishes of the genus *Lethrinops*. *Annals and*  
603 *Magazine of Natural History*, (10) 7, 133-153.  
604
- 605 Trewavas, E. (1935). A synopsis of the cichlid fishes of Lake Nyasa. *Annals and Magazine of*  
606 *Natural History* (10) 16, 65-118.  
607
- 608 Turner, G. F. (1996). *Offshore Cichlids of Lake Malawi.* Cichlid Press, Lauenau. 240 pp.  
609
- 610 Turner, G. F. (2022). A new species of deep-water *Lethrinops* (Cichlidae) from Lake Malawi.  
611 *Journal of Fish Biology*, 101, 1405-1410.  
612

- 613 Turner, G. F., Ngatunga, B. P. & Genner, M. J. (2019) The natural history of the satellite lakes  
614 of Lake Malawi. *EcoevoRxiv*. <https://ecoevorxiv.org/sehdq/> 131pp.  
615
- 616 Tyers, A. M., Bavin, D., Cooke, G. M., Griggs, C. & Turner, G. F. (2014). Peripheral isolate  
617 speciation of a Lake Malawi cichlid fish from shallow-muddy habitats. *Evolutionary Biology*,  
618 41, 439-451.  
619
- 620 Weller, H., López-Fernández, H., McMahan, C. D. & Brainerd, E. L. (2022). Relaxed feeding  
621 constraints facilitate the evolution of mouthbrooding in Neotropical cichlids. *American*  
622 *Naturalist*, 199, E197-E210.  
623  
624