

E-ISSN: 2320-7078 P-ISSN: 2349-6800 www.entomoljournal.com

JEZS 2020; 8(2): 1334-1340 © 2020 JEZS Received: 06-01-2020 Accepted: 08-02-2020

Thomas Latha Dept. of Science, Faculty of Science & Technology, University of Belize, Belize

Karen Bautista

Dept. of Science, Faculty of Science & Technology, University of Belize, Belize

Corresponding Author: Thomas Latha Dept. of Science, Faculty of Science & Technology, University of Belize, Belize

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com

J E Entomalogy and Zoology Studies

Preliminary investigation on insect pests of sugarcane in the northern sugar belt region of Belize

Thomas Latha and Karen Bautista

Abstract

A preliminary investigation was done on the insect pests of sugarcane (*Saccharum* sp) in the northern sugar belt region of Belize. Three sites each were identified in the Corozal and Orange Walk districts of Belize which comprised the northern sugar belt region. The study sites were Sugar Industry Research and Development Institute's demonstration plots. Insects were collected by hand picking; sticky traps and bottle traps with bait from each of the study sites. Two such collections were made in the month of January, 2018. Froghopper (*Aneolamia varia*), and sugarcane borer (*Diatraea saccharalis*) were collected from all the sites and was found to cause the most damage to sugarcane crops in the northern sugar belt region. Sugarcane Mealy bug (*Saccharicoccus sacchari*), Weevils (*Apinocis subnudus* and *Sphenophorus incurrens*), Nitudilid beetles and sugarcane lace bug (*Leptodictya tabida*) were also identified. The biology and ecology of these insect pests including control methods for the major pests are reviewed in this paper. Further detailed study over an extended period of time is recommended on the pests of sugarcane and the damage caused by them to the sugar industry of Belize to get a more accurate picture and to plan effective control measures for the same.

Keywords: Sugarcane, insect pests, Belize, Aneolamia varia, Diatraea saccharalis

1. Introduction

The sugar industry in Belize has become a vital component of Belize's economy in the past decade, accounting for an average of 7.8% of the country's GDP and earning about 34% of total foreign exchange in agricultural exports ^[1]. Sugarcane (*Saccharum* sp.) in Belize was first introduced into the northern district of Corozal in 1848 by Mexican immigrants from Yucatan and was grown in small amounts to harvest molasses and sugar by animal-powered mills. The industry took root and grew with the arrival of the American expatriates during the late 1860s and 1870s. By the late 1890s and early 1900s, East Indians brought in as indentured laborers to work in the sugarcane fields further boosted the sugar industry. The industry is largely concentrated in the Northern districts of Corozal and Orange Walk, also called the 'Sugar Belt' with approximately 27,518.6 ha or about 30% of the total agricultural area in the country under sugarcane cultivation ^[2].

The sugarcane fields in the Northern districts are plagued by many different insect pests affecting the quality and quantity of sugar produced ^[3]. In 2006-2007 severe infestation by frog hopper (*Aneolamia varia*) in the northern sugar belt region led to a 10% loss of sugarcane production. Hence, a preliminary investigation was conducted in 2018 in the northern sugar belt region of Belize to document the insect pests prevalent in the sugarcane fields. This would enable Sugar Industry Research and Development Institute (SIRDI) of Belize to take adequate measures to combat these insect pests. The biology and ecology of the insect pests collected and identified in the study including control methods for the two major pests are reviewed in this paper.

2. Materials and methods

2.1 Study sites: A total of six sites (Table 1; Fig. 1A) were selected at random from each of the two districts representing the sugar belt: three in Corozal district and three in Orange Walk district of Belize. The sites were primarily SIRDI demonstration plots (Fig. 1B).

Table 1: Study Site Location and Sugarcane crop information

| Site # | Location (District) | Variety | Ratoon | |
|--------|--------------------------------|----------|--------|--|
| 1 | Buena Vista (Corozal) | B79474 | R16 | |
| 2 | Santa Clara (Corozal) | Bbz80240 | R2 | |
| 3 | San Joaquin (Corozal) | B52298 | R20 | |
| 4 | La Savannah (Orange Walk) | B79474 | R15 | |
| 5 | Arnulfo Muñoz (Orange Walk) | B79474 | R5 | |
| 6 | Tower Hill – BSI (Orange Walk) | B79474 | R9 | |

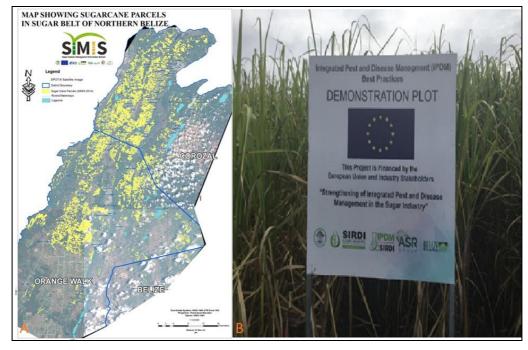


Fig 1: (A) Map showing sugarcane parcels (yellow) in Sugar belt of Northern Belize; (B) SIRDI demonstration plot at Tower Hill

2.2 Sample collection

Samples were collected in the month of January, 2018 on two separate occasions. The pest samples were collected using a combination of hand picking, sticky traps and bottle traps. Three sticky traps (courtesy of SIRDI) were placed in each of the collection sites (Fig. 2 A). Three, 11iter plastic bottles containing an attractant (pineapple) and a preservative fluid at the bottom quarter was used as bottle traps (Fig. 2 B). Four days after the placement of the traps, specimens were collected and preserved in labelled vials containing 70% alcohol and brought to the University of Belize's laboratory for identification.

The insect pests collected were observed, identified and photographed by using stereo microscopes. Identifications were done using morphological characteristics to the closest taxa possible.



Fig 2: (A) Sticky Trap at La Savanna; (B) Bottle trap set up at Tower Hill demonstration plot

3. Results and discussions

The insect pests collected from the Northern Sugar belt region of Belize are summarized in Table 2. Froghopper (*Aeneolamia varia*) and Sugarcane Borer (*Diatraea saccharalis*) were collected from all the sites and are considered as the major pests of sugarcane in northern Belize because of their increased prevalence and damage caused to the sugarcane production. Mealy Bug (*Saccharicoccus sacchari*), Nitidulid beetles, Weevil (*Apinocis subnudus* and *Sphenophorus incurrens*) and Sugarcane lacebug (*Leptodictya tabida*) which were also collected during the study do not cause any significant loss to sugar industry of Belize. The biology and ecology of these insect pests including control methods for the major pests are reviewed in this paper.

Table 2: Table showing location, cane variety, ratoon, trap types and insect pests collected from the sugarcane fields of Northern Belize

| Site # | Location (District) | Variety | Ratoon | Insect pest hand picked | Insect pest bottle trapped | Insect on sticky trap |
|-----------|--------------------------------|----------|--------|---|---|--------------------------|
| 1 | BuenaVista (Corozal) | B79474 | R16 | Froghopper (Aeneolamia varia) | Nitidulid beetles, Sugarcane lace bug (<i>Leptodictya tabida</i>) | |
| 2 | SantaClara (Corozal) | Bbz80240 | R2 | A. varia, Sugarcane borer (Diatraea saccharalis) L. tabida | Nitidulid beetles | |
| 3 | SanJoaquin (Corozal) | B52298 | R20 | A. varia, D. saccharalis | Nitidulid beetles, Weevil (Apinocis subnudus, Sphenophorus incurrens) | |
| 4 | LaSavannah (Orange Walk) | B79474 | R15 | D. saccharalis Sugarcane mealybug (Saccharicoccus sacchari) | Nitidulid beetles, Weevil (A.subnudus, S. incurrens) | A.varia |
| 5 | Arnulfo Muñoz (Orange Walk) | B79474 | R5 | | Nitidulid beetles | A.varia |
| 6 | Tower Hill –BSI +(Orange Walk) | B79474 | R9 | | Nitidulid beetles | A.varia |

3.1 Froghopper (Aeneolamia varia)

Species belonging to *Aeneolamia* are one of the major pests of sugarcane (Fig. 3). They belong to the order Hemiptera, family Cercopidae and subfamily Tomaspidinae. The genus includes eight species and 34 subspecies, which have wide variation in the color pattern on the Tegmina (fore wing) ^[3]. There is also considerable variation in color between specimens of the same species of *Aeneolamia*, found in same or separate localities. But the characters from male genitalia are conservative within the genus and are reliable to identify the species ^[3]. *A. varia* and *A. albofasciatus* are reported from Belize.

3.1.1 Life cycle

Adult *A. varia* females mate immediately after emergence. Females lay around ~150 spindle shaped eggs in soil or decaying plant parts ^[4]. Incubation takes 2-40 weeks depending on the presence or absence of diapause. Photoperiod and soil moisture influences egg development and diapause ^[5, 6, 7, 8, 9]. Nymphs ingest sap from the roots and excrete excess sap to produce spittle mass that protects it from desiccation ^[5, 10]. There are around 5 nymphal instars and life cycle takes about two months. Adult *A. varia* live for approximately 20 days ^[4]. There are generally 2-4 generations per year ^[5] (Fig. 3).

3.1.2 Damage caused

The border parenchyma cells of sugarcane leaves are the primary feeding site of adult A. varia ^[5]. Enzymes in saliva cause necrosis of plant tissue which appears as longitudinal brown streaks; uncontrolled infestations can cause 30-70% reduction of sucrose content ^[5]. On the roots, nymphs are xylem-feeders, although the first and second instars also ingest the contents of parenchyma cells in the cortex of young roots. Cellular damage is extensive in all cases, and occlusion of the xylem elements often occur following feeding by fourth and fifth instars ^[11]. In heavy attacks the leaves turn yellow and then brown and finally wilt and die, a condition called 'froghopper blight' ^[5]. In 2006/2007 season, Aneolamia spp alone was responsible for 10% loss of sugar production in northern Belize. The froghopper insect becomes a problem for sugarcane when it exists in very high numbers, which escalate especially in hot, humid conditions.

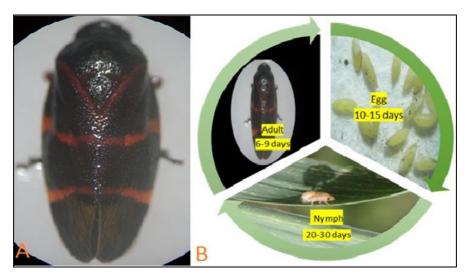


Fig 3: (A) A. varia adult; (B) Life cycle of Aeneolamia sp.

3.1.3 Control

Cultural control methods include maintaining sugarcane fields free of weeds; crop rotation or intercropping; special cultivation methods such as trash lining and retention, interrow tillage and special land preparation techniques before replanting ^[2]. Physical or Mechanical methods include developing improved drainage or irrigation systems; destruction or removal of alternate host species in and around the sugarcane ecosystem; use of clean sugarcane seeds and/or planting materials; use of traps to trap the pest; promotion and encouragement of good harvesting strategies such as minimizing dead and decaying high stump remains and stalk droppings, which constitutes a source of food for some pest species ^[2]. Entomopathogenic fungi, *Metarhizium anisopliae* (Hypocreales: Clavicipitaceae) is used as an effective biological control of froghoppers in Belize. Insecticide Actara WG with active ingredient thiamethoxam, Engeo 247SC with active ingredient Thiamethoxam and Lambda, Jade 0.8 GR with active ingredient Imidacloprid are used as chemical control of froghoppers in Belize^[2].

3.2 Sugarcane Borer (Diatraea saccharalis)

Diatraea spp. commonly called the "moth borer" or the "lesser moth borer" is a major pest of sugarcane in the region (Fig. 5). They are represented by 41 species ^[12]. They belong to the order Lepidoptera, family Pyralidae and subfamily Crambinae. The species *D. saccharalis* is a pest of economic importance in the northern sugar belt region.

3.2.1 Life cycle

Female *D. saccharalis* usually mate only once in their lifetime, but sometimes mates several times in a period of 48 hrs. ^[13]. Mating usually takes place in darkness ^[14]. Eggs are deposited from dusk through night on either side of a leaf in clusters. The eggs are round-oval, flattened with average of 30 eggs per cluster ^[15]. In *D. saccharalis* maximum fecundity may range from 250 to 700 eggs per female ^[4, 16].

The eggs are white at first with microscopic network of depressed lines but later takes on reddish brown color. The eggs of *D. saccharalis* hatch after 4-9 days and the small larvae congregate in the terminal buds of the plants and feed on the tender whorls. Rows of holes appear on the leaves as they expand. Many of the larvae seem to perish during this period partly because of their cannibalistic habits ^[17].

In their 3rd and 4th instar, the larva crawls down the outside of the stem to a point near or even below the surface of the ground and bore into the stalk, where they continue to develop ^[17]. Stalk penetration usually occurs through an immature internode ^[18, 19]. Gnawing a hole through the outer layer of the stem, it works its way to the interior of the plant. When full-grown they are about an inch long by one-eighth inch wide. The head is brown and the body white with brown spots ^[16]. The larva of *D. saccharalis* undergoes 5-10 molts on sugarcane ^[20, 21, 22]. Before pupation, the larva enlarges the tunnel, makes an exit opening covered only by a thin layer of plant tissue for the moth to escape on emergence from the pupal stage ^[17]. On molting for the last time, the larva enters the pupal stage, the quiescent period of the insect. The pupa at first is white, but soon changes to dark brown. Duration of the pupal stage can be 6–11 days in *D. saccharalis* ^[20, 22].

The adult is a straw-colored moth, the forewings marked with darker lines. It varies in size, average specimens measuring about an inch across the wings ^[17]. Adult longevity ranges from 5–10 days for *D. saccharalis* ^[23]. *Diatraea* spp. typically produce 6–11 generations per year in the tropics ^[4] (Fig. 4).



Fig 4: *Diatraea saccharalis* life history stages: A. Egg; B. Larva; C. Pupa; D. Adult

3.2.2 Damage caused

Larvae of *D. saccharalis* bores into the tender inner shoot of sugarcane forming a "dead heart." Small plants are killed and larger stalks usually do not die ^[17]. Insect tunneling in stalks can interfere with the movement of nutrients and

photosynthates in later crop stages, increase the level of fiber in the stalk, and decrease its value ^[24, 25]. Injury of stalk can disrupt apical dominance and promote growth of multiple lateral shoots, diverting resources from sucrose synthesis to vegetative growth ^[26]. Multiple entry and exit holes in stalks provide points of entry for microorganisms that can further degrade cane quality and sugar content ^[26, 27, 25]. Stalks bored by the larvae becomes infected with fungus *Colletotrichum falcatum* causing red rot ^[17].

3.2.3 Control

Management of *Diatraea* spp. in many sugarcane regions has largely focused on biological control since the larvae develop inside the sugarcane stalks, which diminishes efficacy of the insecticides ^[28]. Ants, particularly imported red fire ant, *Solenopsis invicta* Buren, are reported to be important predators of sugarcane borer in sugarcane fields, and capable of reducing damage from borers by over 90%. Other ant species such as *Pheidole dentata* Mayr and *Pheidole floridana* Emery (all Hymenoptera: Formicidae) are also important.

Egg parasitoids, *Trichogramma* sp. (Hymenoptera: Trichogrammatidae), are possibly the most important naturally-occurring parasitic insects. Predators could be *Orius* spp. pirate bugs (Hemiptera: Anthocoridae), lacewings (Neuroptera: Chrysopidae), tiger beetles (Coleoptera: Cicindelinae), spiders, and foliage-dwelling ground beetle larvae (Coleoptera: Carabidae). Entomopathogenic fungi, *M. anisopliae* ^[29, 30, 31] has also been used to control *D. saccharalis* larva.

3.3 Sugarcane mealy bug (Saccharicoccus sacchari)

It is a common pest of sugarcane in the warm regions of the world (Fig. 5). They belong to order Hemiptera, family Pseudococcidae. Besides sugarcane they also attack several Poaceae (Graminae), Sorghum, rice.

3.3.1 Life cycle

Female lay upto 1000 eggs under the leaf sheath which hatch quickly in 10-14 hrs. ^[5]. First instar nymphs are generally active, moving to younger parts of the plant or to adjacent plants; older nymphs are less active ^[5]. Females molt three times before attaining maturity and the males molt four times ^[32, 33, 34, 35, 36]. Males cease feeding at the end of the second instar and the third and fourth instar of males are without functional mouth parts ^[37].

Adult female is pink in color, 7mm long, elongate-oval to round in shape with well-developed anal lobes and short legs. Adult male exists as apterous or winged form and is rare. Though parthenogenesis is considered the normal mode of reproduction, it was reported that in Hawaii, *S. sacchari* females failed to produce eggs or produced inviable egg when they did not mate ^[37]. Life cycle takes about 30 days to complete.

The mealybugs secrete a covering of wax, and also produce a syrupy 'honey dew' on which a black, sooty mold usually develops. Cane which has been infested with mealybugs can thus almost always be identified by the presence of wax and sooty mold, even when the insects have disappeared ^[5].

3.3.2 Damage caused

Nymphs and adults suck sap from leaves, nodes, and internodes of canes; severe infestation results in yellowing of leaves, stunting of canes and poor germination in the case of *S. sacchari* attack ^[38]. High populations can produce large

amounts of honeydew and sooty mold growing on the honey dew will disfigure crops. *S. sacchari* is associated with filtration and clarification problems of syrup, lower quality of the syrup and reduced crystallization ^[38, 39, 40] which is related to the production of honeydew and associated polysaccharides and gums and/or the close association of *S. sacchari* with acetic-acid-producing bacteria ^[41]. The pest may also transmit plant viruses ^[42].

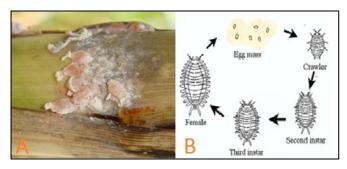


Fig 5: Saccharicoccus sacchari A. adult B. Life cycle

3.4 Nitidulid beetles

Also called sap beetles, belong to the order: Coleoptera, family: Nitidulidae. They are important pest of wide variety of important fruits and grains and also vectors of harmful microorganisms. Nitidulid beetles are 3-4 mm long, small shiny black or dark brown in color with clubbed antennae. The clubbed portion, however, is quite variable within species, being quite distinct or only slightly developed ^[43]. The elytra or wing covers are entire or shortened to expose two or three abdominal segments. The larvae are slender, creamy-white grubs which reach a length of about 6 mm. Larvae and beetles are sometimes found in setts that have failed to germinate. They may be attracted there by fungal growth or by fermenting vegetable matter. They are regarded as a secondary pest.

3.4.1 Damage caused

Pineapple disease of sugarcane are transmitted by Nitidulid beetles ^[44]. It is an economically important sugarcane disease that occurs in almost all countries where sugarcane is grown. The disease is caused by the fungus *Ceratocystis paradoxa*. It primarily affects sugarcane setts in the first weeks of planting. The fungus infects the setts mainly through the cut ends and from there spreads rapidly through the parenchyma. Infected tissue first becomes reddened; the parenchyma then breaks down and the interior of the setts become hollow and blackened. In the early stages of the rotting, the strong odor of overripe pineapples is often present and may help in diagnosing the disease.



Fig 6: Nitidulid beetles

3.5 Weevils

The weevils identified from the bottle traps belonged to the species *Apinocis subnudus* and *Sphenophorus incurrens* (Fig.7A, B). Weevils belongs to the order: Coleoptera, family: Curculionidae. Main damages are caused by weevil larvae rather than adults.

3.5.1 Damage caused

The weevil larva penetrates the base and underground parts of the stem of the cane making galleries in different regions. They can feed on several stems and the rhizome of the strain. The secondary damage caused by the larvae is by the entry of phytopathogenic fungi, *Colletotrichum falcatum* causing decay resulting in red rot disease ^[45].

The larvae of *A. subnudus* usually penetrate the lower part of the stem; the galleries are distinguished by being smaller than those caused by *S. incurrens* and *Metamasius hemipterus*. All species pupate inside the cane generally in the lower and underground parts. Weevils feed on the buds, root primordial, and from the lateral parts of the cut internodes, where the females form galleries and oviposit. The new plant may show symptom of "dead heart" which differs from that caused by lepidoptera such as *Diatraea* spp. and *Eoreuma loftini* because they do not present entry hole in the bud ^[46].



Fig 7: A. Apinocis subnudus B. Sphenophorus incurrens

a) Sugarcane Lace bug (Leptodictya tabida)

They belong to the order: Hemiptera, family: Tingidae.

3.6.1 Life cycle

The adult lace bug (*Leptodictya tabida*) is about 1/8" (3.5 mm) long, flat and light-brown or straw colored (Fig. 8A). The forewings are semi-transparent and finely laced or netlike (Fig. 8A). Five long, erect spines are present on the head. Eggs are laid singly into leaf tissue usually on the underside of the leaves. The tip of the egg is left outside the leaf tissue but is covered with a protective cap secreted by the adult female. Nymphs are flat and whitish in color with many long, branched, erect spines. A generation of lace bugs, from egg to adult, may take 20 to 30 days, with 5 nymphal molts ^{[47, 48][49]}.

3.6.2 Damage caused

Lace bug feeding results in light green or yellow speckles on leaves. Many cultivars develop a red discoloration (russetting) on the leaves that expands beyond the area of feeding (Fig. 8B) ^[50]. Another characteristic diagnostic character of sugarcane lace bug are the small, black, oily fecal deposits (i.e., frass spots) they deposit on the leaves ^[50]. Predators of these plant-sucking insects include earwigs and spiders.



Fig 8: A. Adult Lace bug; B. Adult, nymph and damaged leaf

4. Conclusion

Preliminary investigation on sugarcane pests of northern sugar belt region of Belize revealed that froghopper *Aneolamia varia* and sugarcane borer *Diatraea saccharalis* were the major pests of sugarcane during the 2018 study period. Further detailed study on pests of sugarcane over an extended period of time is recommended to get a more accurate picture of the pests of sugarcane and the damage caused by them to the sugar industry of Belize. This will enable SIRDI to develop effective protocol to control these common pests of sugarcane in the northern sugar belt region of Belize.

Acknowledgement

We wish to thank the Sugar Industry Research and Development Institute (SIRDI), the European Union, and SIRDI EU IPDM Project Unit. Special acknowledgement to the SIRDI extension officers, Dr. Luciano Chi (Research Coordinator / SIRDI EU IPDM Head of Unit) and Mr. Jeffy Gomez (SIRDI EU IPDM Project Manager). We wish to also acknowledge the assistance of our colleagues Dr. Thiagarajan, Dr. Herron and University of Belize for the laboratory resources.

5. References

- 1. Statistical Institute of Belize. Estimated mid-year population 2015 and merchandize trade, population and household. Book Green, London: Halcrow Group Limited, 2015.
- 2. Chi L. Integrated Pest Management of Frog Hopper (*Aeneolamia spp.*) in Sugarcane in Belize. IPDM manual for SIRDI, 2019, 1-26.
- 3. Paladini A, Cavichiolo RR. A new species of *Aeneolamia* (Hemiptera: Cercopidae: Tomaspidinae) from the Neotropical Region. Zoologia. 2003; 30(3):353-355.
- 4. Guagliumi P. The sugarcane pests in Venezuela. Volume 1, Ministry of Agriculture, Center of Agronomic Research, Maracay, Venezuela, 1962, 847p.
- 5. Hill DS. Pests of Crops in Warmer Climates and Their Control, 2008. Springer. 10.1007/978-1-4020-6738-9.
- Urich FW, Pickles A. Studies in the incubation of the eggs of the sugar cane froghopper, *Tomaspis saccharina* Dist. I. Eggs laid in blotting paper. Minut. Proc. Froghopper Invest. Comm. 1930; 3:64-70.
- 7. Urich FW, Pickles A. Studies in the incubation of the eggs of the sugar cane froghopper, *Tomaspis saccharina* Dist. 11. Eggs laid in soil. Minut. Proc. Froghopper

Invest. Comm. 1931; 3:253-255.

- Evans DE. Interim report. Caroni Research Station, Waterloo Estate, Carapichaima, Trinidad and Tobago, 1971.
- 9. Vreugdenhil A. Estudio sobre la diapausa dela can del ilia (Zona Yaritagua, Estado Yaracuy, 1973-1976). II. Seminario sobre los problemas de la candelilla y el taladrador en cana de azucar y pastos Barquisimeto, Lara, Venezuela. 1984; 4:1-10.
- Valério JR, Cardona MC, Peck DC, Sotelo G, Kelemu S. Spittlebugs: Bioecology, host plant resistance and advances in IPM. In: International Grassland Congress (Sao Paulo, Brazil), 2001.
- 11. Hagley EAC, Blackman JA. Site of Feeding of the Sugarcane froghopper, Aeneolamia varia saccharina (Homoptera: Cercopidae). Annals of the Entomological Society of America. 1966; 59(6):1289-1291.
- Solis MA, Metz MA. An illustrated guide to the identification of the known species of *Diatraea* Guilding (Lepidoptera, Crambidae, Crambinae) based on genitalia. Zookeys. 2016; 565:73-121. doi:10.3897/zookeys.565.6797
- Linares BA. Influencia de la temperatira en el desarrollo de *Diatraea saccharalis* Fabricius. Cana de Azucar. 1987; 6:43-66.
- Batista-Pereira LG, Santangelo EM, Stein K, Unelius CR, Eiras AE, Correa AG. Electrophysiological studies and identification of possible sex pheromone components of Brazillian populations of the sugarcane borer, *Diatraea* saccharalis. Verlag der Zeitschrift Naturforschung. 2002; 57:753-758.
- 15. Pastrana CE, Gomez LA, Zuluaga JI. Life Cycle of *Diatraea indigenella* under different diets. Revista Colombiana de Entomologia. 1993; 19:101-106.
- Bessin RT, Reagan TE. Fecundity of sugarcane borer (Lepidoptera: Pyralidae), as affected by larval development on gramineous host plants. Environmental Entomology. 1990; 19: 635-639.
- 17. Holloway TE, Haley WE, Loftin UC, Heinrich C. The sugar-cane moth borer in the United States. 1928. United States Dept of Agriculture, US Govt. Printing Office, Washington DC.
- 18. Peairs FB, Saunders JL. *Diatraea lineolate* and *D. saccharalis*; a review in relation to corn. Agronomia Costarricense. 1980; 4:123-135.
- 19. White WH. Movement and establishment of sugarcane borer (Lepidoptera: Pyralidae) larvae on resistant and susceptible sugar cane. Florida Entomologist. 1993; 76:456-473.
- 20. Taylor DJ. Life History studies of the sugarcane moth borer, *Diatreaea saccharalis* Linn. The Florida Entomologist. 1944; 27:10-13.
- 21. King EG, Miles R, Martin DE. Advances in rearing *Lixophaga diatraeae*. Entomophaga. 1975; 20:307-311.
- 22. Pastrana CE, Gómez LA, Zuluaga JI. Life cycle of *Diatraea indigenella* under different diets. Revista Colombiana de Entomología. (in Spanish). 1993; 19:101-106.
- Walker DW, Alemany A. Biology of the sugarcane borer, *Diatraea saccharalis* (Fabr.) II. Longevity of adults. 1965, 1469-1471. In Proceedings of the 12th International Congress of Sugarcane Technology, San Juan, Puerto Rico. 1965; 28(10):

- 24. Milligan SB, Balzarini M, White WH. Broad-sense heritabilities, genetic correlations, and selection indices for sugarcane borer resistance and their relation to yield loss. Crop Science. 2002; 47:1729-1735.
- White WH, Viatorv RP, Dufrene EO, Dalley CD, Richard EP Jr, Tew TL. Reevaluation of sugarcane borer (Lepidoptera: Crambidae) bioeconomics in Louisiana. Crop Protection. 2008, 27:1256-1261. doi:10.1016/j.cropro.2008.03.011
- Metcalfe JR. The estimation of loss caused by sugar cabe, In William, J.R.; Metcalfe, J.R.; Mungomery, R.W.; Mathes, R. [eds.]. Pest of Sugarcane. Elsevier, Amsterdam. The Netherlands, 1969, 61-79.
- 27. White WH, Hensley SD. Techniques to quantify the effect of *Diatraea saccharalis* (Lepidoptera: Pyralidae) on sugarcane quality. Field Crops Research. 1987; 15:341-248.
- Pereira FF, Kassab SO, Calado VRF, Vargas EL, Oliveira HN, Zanuncio JC. Parasitism and Emergence of *Tetrastichus howardi* (Hymenoptera: Eulophidae) on *Diatraea saccharalis* (Lepidoptera: Crambidae) Larvae, Pupae and Adults. Florida Entomologist. 2015; 98: 384-387. 10.1653/024.098.0164.
- Alves SB, Rossi LS, Lopes RB, Tamai MA, Pereira RM. "Beauveria bassiana yeast phase on agar medium and its pathogenicity against Diatraea saccharalis (Lepidoptera: Crambidae) and Tetranychus urticae (Acari: Tetranychidae)," Journal of Invertebrate Pathology. 2002; 81(2):70-77.
- Zambrano K, Davila M, Castillo MA. Detecci ' on de ' fragmentos de AND de hongos y su posible relacion con la ' s'intesis de prote'inas de actividad entomopatogena," ' Revista de la Facultad De Agronom'ia (LUZ). 2002; 19:185-193.
- 31. Estrada ME, Romero M, Rivero MJ, Barroso, F. Natural presence of *Beauveria bassiana* (Balsamo) Vuillemin in the sugar cane (Saccharum sp. hybrid) in Cuba," Revista Iberoamericana de Micologia. 2004; 21(1):42-43.
- Matheson R. Number of moults of the female of Dactylopius citri. Canadian Entomologist. 1909; 39:284-287.
- Myers LE. Two economic greenhouse mealybugs of Mississippi. Journal of Economic

Entomology. 1932; 25:891-896.

- 34. Uichanco LB, Villanueva FE. Biology of the pink mealybug of sugar cane, *Trhnymus sacchari* (Cockerell) in the Philippines. Philippine Agriculturist. 1932; 21(4):205-267.
- 35. Basinger AJ. Reproduction in mealybugs. Annals of the Entomological Society of America.1934; 27:17-20.
- 36. James HC. On the pre-adult instars of *Pseudococcus longispinus* Targ., with special reference to characters of possible generic significance. Transactions of the Royal Entomological Society of London. 1937; 86:73-84.
- Beardsley JW. Notes on the Biology of the Pink Sugar Cane Mealybug, *Saccharicoccus sacchari* (Cockerell), in Hawaii (Homoptera: Pseudococcidae). 1962; 18(1):55-60.
- Dick J. The Mealybugs of sugarcane. In William J.R. *et al.* Pests of Sugarcane. (Coccids p. 343-365). Elsevier Pub. Co., Amsterdam, London, New York. 1969; 1(12):568p.
- 39. Hall WJ. Observations on the Coccidae of Egypt. Bulletin, Ministry of Agriculture, Egypt, Technical and

Scientific Service. 1922, 22:1-54.

- 40. Dymond GC. Mealybugs. Their effect on cane culture and manufacture in Natal. Proceedings of the South African Sugarcane Technologists Association. 1929; 3:51-53.
- 41. Ashbolt NJ, Inkerman PA. Acetic Acid Bacterial Biota of the Pink Sugar Cane Mealybug, *Saccharococcus sacchari*, and Its Environs. Applied Environmental Microbiology. 1990; 56(3):707-712.
- 42. Lokhart BEL, Autrey LJC, Comstock JC. Partial purification and serology of sugarcane mild mosaic virus, a mealy bug- transmitted closterolike virus. Phytopathology. 1992; 82:691-695.
- 43. Parsons CT. A revision of Nearctic Nitidulidae (Coleoptera). Bulletin of Comparative Zoology. 1943; 92:121-248.
- 44. Chang VCS, Jensen L. Transmission of the Pineapple Disease Organism of Sugarcane by Nitidulid Beetles in Hawaii. Journal of Economic Entomology. 1974; 67(2):190-192.
- 45. Butler EJ. Fugus diseases of sugarcane in Bengal. Indian Dept. of Agriculture, Memoirs of Department of Agriculture India, Botanical Series. 1906; 1(3):2-24.
- 46. Ruiz-Montiel C, Illescas-Riquelme CP, Altamirano-Hernández U, Jones RW. Nuevos registros de picudos (Coleoptera: Curculionidae) afectando caña de azúcar (*Saccharum officinarum* L.) en Veracruz, México. Southwestern Entomologis. 2015; 40:427-432.
- 47. Heidemann O. The sugar-cane tingid from Mexico. Journal of Economic Entomology. 1913; 6:249-251.
- 48. Hall DG. Sugarcane Lacebug *Leptodictya tabida*, an insect pest new to Florida. Florida Entomologist, Scientific notes. 1991; 74(1):148-149.
- 49. Nguyen R, Hall DG. Sugarcane Lace Bug, *Leptodictya tabida*; Publication EENY-044, Florida Cooperative Extension Service, University of Florida: Gainesville, FL, USA, 1998.
- 50. Chang VCS. The sugarcane lacebug: a new insect pest in Hawaii. Annual Conference. Report, Hawaiian Sugar Technology. 1985; 44:A27-A29.