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A comparative study of consumption of *Aedes aegypti* and *Aedes albopictus* larvae by ornamental fishes to control their population in West Bengal

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Abstract

Aedes aegypti and Aedes albopictus are the carriers of a variety of diseases such as dengue, dengue hemorrhagic fever (DHF), chikungunya, zika virus, and yellow fever. An affordable way of vector management that effectively suppresses the mosquito population for a long time is the release of larvivorous fish. In this work, ornamental fish species are identified, and their potential larvicidal efficacy for biological control of Aedes mosquito larvae is assessed. The consumption rates of four ornamental fish species (Gold fish, Betta fish, Molly fish, and Guppy) for Aedes aegypti and Aedes albopictus larvae have been observed in the present study, and a comparison between the mean consumption rates of the four fish species for Ae. aegypti and Ae. albopictus has been recorded. The observation indicated practically no variance, and the intake rates for both types of larvae were almost same for the four species of fish. In contrast to guppy fish, the findings indicated that Goldfish, Black molly, and Betta fish had very high feeding rates. Therefore, in the near future, these fish can be thought of as efficient biological control agents for both Aedes aegypti and Aedes albopictus larvae.

Keywords: Aedes aegypti, Aedes albopictus, ornamental fish, biological control, food preference, larvicidal efficacy

Introduction

Mosquitoes are essential insects not just as nuisance biters, but also as disease vectors. Several mosquito species specially *Aedes aegypti* and *Aedes albopictus*, are responsible for the transmission of several important viral infections, including dengue, dengue hemorrhagic fever (DHF), yellow fever, and chikungunya. These diseases have become major public health problems in the world's tropical and subtropical climates ^[1]. Dengue fever is one of the world's fastest spreading diseases, endemic in more than 120 countries, and up to 390 million cases are anticipated to occur each year ^[2]. Dengue-related deaths are significantly less common, although the projected number of apparent cases worldwide in 2013 was 58.4 million ^[3]. Yellow fever, on the other hand, is currently endemic in 47 countries and fatalities between 29,000 and 60,000 people each year ^[4]. Therefore the only ways to prevent dengue transmission and reducing the disease's burden should be the control of mosquitoes, particularly *Aedes aegypti* and *Aedes albopictus*, or eliminating human-vector interaction.

The African subcontinent is most likely the origin of *Aedes aegypti*. It later spread to tropical, subtropical, and temperate areas. Currently, this species can be found in Africa, South and Central America, portions of North America, the Middle East, Southeast Asia, and Oceania, including Northern Australia^[5]. It is primarily an urban mosquito that bites during the day and lays drought-resistant eggs^[6]. They also have a tendency to breed in containers and tiny areas, such as between enormous leaves and stems of vegetation. Female *Aedes aegypti* mosquitos prefer to feed on humans who are in enclosed places, such as houses^[7].

Aedes albopictus, like *Aedes aegypti*, has been increasingly intertwined with urban and periurban areas as its geographic range has grown ^[8]. The species has become increasingly capable of exploiting human-made container habitat and human blood meal hosts inside these landscapes ^[6]. It can resist a wider range of temperatures and prefers cool climates. As a result, this mosquito species may now survive in more temperate climates ^[7]. *Aedes albopictus* mosquitos are currently found throughout Africa, Asia, South America, and the Pacific and Indian Ocean islands ^[9]. *Ae. albopictus* feeds during the day, preferring vegetation-rich habitats that have a more urban and semi-urban appearance ^[10]. Numerous laboratory studies show that *Ae. albopictus* can be equally able to acquire and transmit pathogens as *Ae. aegypti* for a variety of arboviruses, including chikungunya and Zika ^[11, 12, 13].

For reducing the risk of or preventing epidemics, or for responding to outbreaks, respectively, dengue vector control may be aimed against the aquatic larval stages or the adult female mosquito ^[14]. Adult mosquitoes are typically targeted by insecticide fogging or space spraying in response to confirmed dengue transmission, with the aim of reducing biting and transmission as quickly as possible ^[15]. In addition, there is considerable evidence that insecticide-resistant mosquitoes are emerging in dengue-endemic areas ^[16, 17].

In the 20th century, biological control, notably the utilization of larvivorous fish, was vital to malaria control programmes, especially in urban and periurban regions for immediate application in industrialised and developing countries. It has a very significant role to fulfil in the integrated control methods when both fish or other biotic agents and pesticides have distinct roles to play ^[18]. The WHO has continued to include the use of larvivorous fish as a potential method of reducing dengue vectors; however the capability of fish in preventing or controlling dengue is likely to differ from that of malaria. This is primarily due to the fact that Aedes spp. vectors of dengue typically breed in discrete man-made containers and household water tanks, in contrast to the majority of malaria vectors ^[2]. Job (1940) ^[19] states that larvivorous fish need to be small, hardy, and agile in shallow water among dense weeds, which are find to be ideal breeding sites of mosquitoes. They must be able to survive in both deep and

shallow waters, as well as in drinking water tanks and pools, without contaminating the water, and they must be droughtresistant and must be able to survive harsh handling and long range transportation. Fish that survive must be prolific breeders with a short life cycle. In confined waters, they must effectively and freely reproduce. Even in the presence of other food sources, larvivorous fish should prefer mosquito larvae and be surface feeders and carnivorous by nature. Over many years, various fish species have been studied for their potential to reduce mosquito populations. These include fish reared for human food, such as species of tilapia (family Cichlidae), as well as larvivorous members of the Poeciliidae family (often known as "guppies") ^[20, 21, 22, 23]. Numerous researchers have investigated the larvivorous capabilities of local fishes ^[24, 25]. The ornamental tanks, on the other hand, make for the perfect mosquito breeding environment. These ornamental fish not only have a huge amount of potential for introduction in ornamental tanks, where they would not only provide aesthetic attractiveness but also control mosquito breeding ^[26]. According to Chandra *et al.* (2008) ^[18], Câmara et al. (2017)^[27], Sumithra et al. (2014)^[28], some ornamental fish species, such as Poecilia reticulata (Guppy), Poecilia sphenops (Black Molly), Carassius auratus (Gold Fish), and Betta splendens (Betta Fish), are used as larvivorous However, there hasn't been much research done yet to evaluate the potential of ornamental fish for mosquito larval control. Therefore, the objective of the present laboratory study was to evaluate how effectively these ornamental fishes controlled the populations of Aedes aegypti and Aedes albopictus mosquitoes by eliminating their larvae respectively in the presence of alternative food, and to determine whether there was a difference in the larval consumption rates of these ornamental fishes among the two mosquito species.

Materials and Methods Collection Sites

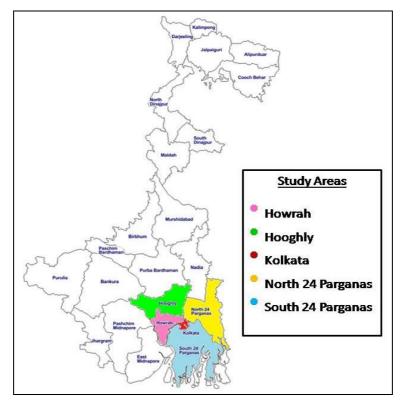


Fig 1: Map representing the collection sites of study area

Description of collection sites

| District | Site of Collections | Collected Species | Latitude | Longitude |
|--------------------|---------------------|--------------------------|-----------|-----------|
| Howrah | Shibpur | Aedes albopictus | 22.5713°N | 88.3109°E |
| nowran | Salkia | Aedes aegypti | 22.6013°N | 88.3313°E |
| Heeshly | Serampore | Aedes albopictus | 22.7505°N | 88.3406°E |
| Hooghly | Uttarpara | Aedes aegypti | 22.6701°N | 88.3355°E |
| Kolkata | New Alipore | Aedes aegypti | 22.5090°N | 88.3329°E |
| Noikata | Hazra | Aedes aegypti | 22.5228°N | 88.3500°E |
| North 24 Parganas | Dumdum | Aedes aegypti | 22.6420°N | 88.4312°E |
| inorui 24 Parganas | Barasat | Aedes aegypti | 22.7248°N | 88.4789°E |
| South 24 Parganas | Diamond Harbour | Aedes albopictus | 22.1927°N | 88.1895°E |

Table 1: Description of collection sites

Collection of Larvae

Larvae of different instar stages of *Aedes aegypti* and *Aedes albopictus* were collected from suspected breeding sites of Howrah, Hooghly, Kolkata, North 24 Parganas, and South 24

Parganas in West Bengal. Plastic containers, discarded jars, tyres, discarded bottles, PVC water reservoirs were considered as primary and major sources of collection.



Fig 2: Various collection sites of Aedes larvae

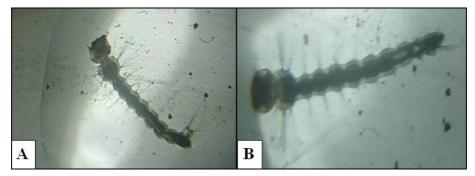


Fig 3: [A] Aedes aegypti larva [B] Aedes albopictus larva

Collection of Fish, Fish pellets and Tubifex sp.

Four species of ornamental fishes, namely Carassius auratus (Goldfish) of 5.8 cm length, Betta splendens (Betta fish) of 6.9 cm length, Poecilia reticulata (Guppy) of 2.7 cm length, and Poecilia sphenops (Black molly) of 3.2 cm length, all under same age group were purchased from ornamental fish shops of Kolkata and Howrah area. Artificial fish pellets (Optimum) and live Tubifex sp. were collected from local aquarium shops in Serampore. Fishes were kept in four 5L glass tanks under the laboratory condition for seven days were supplied with commercial feed. Tubifex sp. were kept in plastic tray submerged in water, and water was changed on a daily basis for both the fishes and *Tubifex* sp.

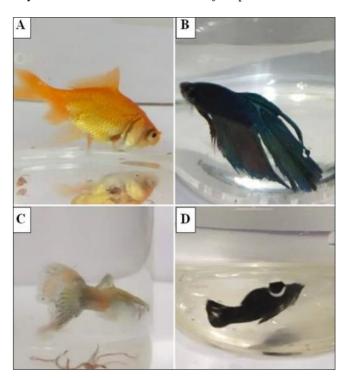


Fig 4: [A] Carassius auratus (Gold Fish), [B] Betta splendens (Betta fish), [C] Poecilia reticulata (Guppy) [D] Poecilia sphenops (Black Molly)

Experimental Design

The fishes were kept for 24 hours without any type of food supply. This was done to make sure that they were in fasting condition before starting the experiments. For each fish, three different sets of experiments were done. In the first set, each fish was given 50 Aedes aegypti larvae, 50 Tubifex sp., and 50 optimum fish pellets; on the second set, each fish was given 50 Aedes albopictus larvae, 50 Tubifex sp. and 50 optimum fish pellets and on the third set of experiment, 50 Aedes aegypti and 50 Aedes albopictus larvae were introduced to each fish species and were observed for 4 hours. The temperature was set at 25 °C.

Each experiment was repeated for ten days to make sure that there is a sufficient time period for the fishes to get used to of the food choice given before starting a new set of combination, so that the larvicidal activity towards the Aedes aegypti and Aedes albopictus larvae could be observed properly. The mean consumption rate of four fishes for Ae. aegypti and Ae. albopictus were taken and observed if any variation exists in the feeding rate of the four fishes for the two species of Aedes.

Statistical Analysis

Two-way ANOVA was performed for each set of experiments followed by Tukey's multiple comparison test for four fish species for ten days of observation using GraphPad Prism (Version-8). Statistical differences were set at p<0.05.

Result and Discussion

Result

The results obtained from the first set of experiment showed significantly higher preference (p<0.0001) for Ae. aegypti larvae than the other two food options for all the fish species. All of the fish species preferred live food than artificial food pellets, but the preference for Aedes aegypti larvae was higher than Tubifex sp. Among the four fishes, Goldfish, Black Molly and Betta fish devoured almost all the larvae in every replicate of the experiments, whereas the feeding rate of guppy was significantly low as compared to the other counterparts. The two way ANOVA showed the results are highly significant at 5% level of confidence.

Table 2: The mean consumption rate of Aedes aegypti larvae, Tubifex sp. and Optimum fish pellets by four different fishes at the 5% level.

| Name of Fish | Consumption Rate (Mean ± SD) | | | | | |
|--------------------------------|------------------------------|-----------------|-----------------------------|--|--|--|
| Name of Fish | Aedes aegypti Larvae | Tubifex sp. | Optimum Fish pellets | | | |
| Carassius auratus (Gold Fish) | 48.6 ± 2.95 | 45.2 ± 5.21 | 18.9 ± 4.57 | | | |
| Betta splendens (Betta fish) | 45.7 ± 2.62 | 39.2 ± 3.55 | 1.3 ± 3.07 | | | |
| Poecilia sphenops (Molly fish) | 43.2 ± 4.69 | 39.9 ± 4.57 | 11 ± 3.58 | | | |
| Poecilia reticulata (Guppy) | 38.4 ± 3.72 | 21.8 ± 4.21 | 1.1 ± 2.15 | | | |

Table 3: Two way ANOVA of the different fishes on consumption of Aedes aegypti larvae, Tubifex sp. and Optimum fish pellets

| | Sum of Squares | df | Mean Square | F value | P value | Significant |
|--|----------------|----|-------------|---------|----------|-------------|
| Fish | 4540 | 3 | 1513 | 407.9 | P<0.0001 | Yes |
| Aedes aegypti larvae, Tubifex sp. and Optimum fish pellets | 28716 | 2 | 14358 | 3659 | P<0.0001 | Yes |
| Interaction | 1329 | 6 | 221.5 | 56.46 | P<0.0001 | Yes |

Significant at 0.05 level confidence

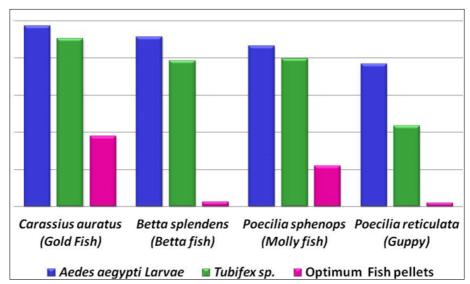


Fig 5: Food preference of four different types of fish for three different types of food items when supplied heterogeneously.

In the second set of experiment also, when the larvae choice was changed, the results showed significantly higher preference (p<0.0001) for *Ae. albopictus* larvae than the other two food options for all the fish species. Here also we observed greater preference for *Ae. albopictus* larvae than the

live *Tubifex* sp. species for all the four fishes. Among the four fishes, Goldfish, Black Molly and Betta fish devoured almost all the larvae in the experiments, but here also, the feeding rate of guppy was comparatively low. The two way ANOVA showed the results are significant at 5% level of confidence.

Table 4: The mean consumption rate of *Aedes albopictus* larvae, *Tubifex* sp. and Optimum fish pellets by four different fishes at the 5% level.

| Nome of Fich | Consumption Rate (Mean ± SD) | | | | | |
|--------------------------------|------------------------------|--------------------|-----------------------------|--|--|--|
| Name of Fish | Aedes albopictus Larvae | <i>Tubifex</i> sp. | Optimum Fish pellets | | | |
| Carassius auratus (Gold Fish) | 49± 2.06 | 45 ± 4.38 | 18.8 ± 4.51 | | | |
| Betta splendens (Betta fish) | 46.3± 5.39 | 38.8 ± 4.01 | 2 ± 3.06 | | | |
| Poecilia sphenops (Molly fish) | 41.8 ± 5.37 | 39.3 ± 4.98 | 11 ± 4.03 | | | |
| Poecilia reticulata (Guppy) | 38.1 ± 3.75 | 20 ± 4.03 | 1.2 ± 2.41 | | | |

Table-5: Two way ANOVA of the different fishes on consumption of Aedes albopictus larvae, Tubifex sp. and Optimum fish pellets

| | Sum of Squares | df | Mean Square | | P value | Significant |
|--|-------------------|----|----------------|-------|------------|-------------|
| Fish | 4871 | 3 | 1624 | 399.6 | P<0.0001 | Yes |
| Aedes aegypti larvae, Tubifex sp. and Optimum fish pellets | 27824 | 2 | 13912 | 2516 | P<0.0001 | Yes |
| Interaction | 1477 | 6 | 246.2 | 60.59 | P<0.0001 | Yes |

*Significant at 0.05 level confidence

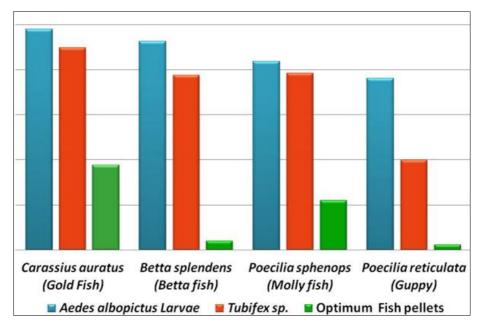


Fig 6: Food preference of four different types of fish for three different types of food items when supplied heterogeneously.

In the third set of experiment, the comparison of the mean consumption rate of *Ae. aegypti* and *Ae. albopictus* of four fishes showed almost no variation and the consumption rate for both the types of larvae was almost at par with each other

for all the four fish species. However, the data indicated that rate of feeding of Goldfish, Black molly and Betta fish were very high, as compared to the Guppy fish.

Table 6: The mean consumption rate of Aedes aegypti and Aedes albopictus larvae, by four different fishes at the 5% level.

| Name of Fish | Consumption Rate (Mean ± SD) | | | | | |
|--------------------------------|-------------------------------------|-------------------------|--|--|--|--|
| Ivalle of FISH | Aedes aegypti Larvae | Aedes albopictus Larvae | | | | |
| Carassius auratus (Gold Fish) | 48.6± 2.65 | 48.2 ± 3.43 | | | | |
| Betta splendens (Betta fish) | 44.8± 3.55 | 45.4 ± 4.35 | | | | |
| Poecilia sphenops (Molly fish) | 40.2 ± 3.67 | 40.3 ± 4.03 | | | | |
| Poecilia reticulata (Guppy) | 37.5 ± 4.26 | 36.1 ± 5.34 | | | | |

Table 7: Two way ANOVA of the different fishes on consumption of Aedes aegypti and Aedes albopictus larvae

| Sum of Squares | df | Mean Square | F value | P value | Significant |
|----------------|---------------|------------------|----------------------------|---|---|
| 1581 | 3 | 527 | 96.28 | P<0.0001 | Yes |
| 1.513 | 1 | 1.513 | 0.5442 | P=0.4655 | No |
| 10.94 | 3 | 3.646 | 1.312 | P=0.2855 | No |
| | 1581 1.513 | 1581 3 1.513 1 | 1581 3 527 1.513 1 1.513 | 1581 3 527 96.28 1.513 1 1.513 0.5442 | 1581 3 527 96.28 P<0.0001 1.513 1 1.513 0.5442 P=0.4655 |

*Significant at 0.05 level confidence

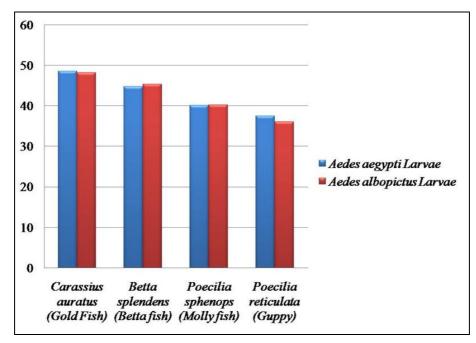


Fig 7: Comparisons between consumption of Aedes aegypti and Aedes albopictus by four different types of fish

Discussion

In this present study we evaluate the potentiality of these four ornamental fish species. In contrast to Poecilia reticulata (Guppy); Poecilia sphenops (Black Molly), Carassius auratus (Gold Fish), and Betta splendens (Betta Fish) all had shown strong larvivorous potentiality. In the laboratory, these fish species are used to reduce the number of mosquito larvae. The present findings are consistent with those of earlier research reports in various ecosystems conducted by Datta et al., 2022 ^[29]; Tilak et al., 2007 ^[26]; Câmara et al., 2017 ^[27]; Sumithra et al., 2014 [28] and Gupta et al., 2009 [30]. The results of the present study make it clear that all four species of fish prefer live food, particularly mosquito larvae, than artificial food, and that this preference is substantially greater than that of other fish species (Table-3; Table-5; Fig-5; Fig-6). The consumption rates of both types of larvae were near about equal for all four fish species, and the observational data also confirmed this result. Therefore these four ornamental fishes have no particular preference for any of the larvae of the two different species of Aedes (Table-7; Fig-7). Production of ornamental fish has become a popular and economically viable industry in several nations throughout the world. Some of these larvivorous fish are also taken directly out of the river for decorative purposes and sold to the nearby aquarium stores, which benefits all parties involved ^[1]. The widespread distribution of these four fish species further supports their sustainability and ability to adapt to a variety of ecological circumstances ^[18, 27]. Since the results of a laboratory study have previously been proven, it is anticipated that using the same approach in a natural setting will be successful.

Conclusion

In general, a variety of methods have been used to reduce diseases spread by mosquitoes. These techniques either stop the parasite from growing inside the mosquito or kill the mosquito carrier. However, dependency on chemical vector control strategies, a lack of infrastructure and resources, and inadequate management strategies all reduce the effectiveness of controlling vector-borne diseases. Additionally, environmental alterations and variations in the behaviours of many mosquito species, such as insecticide resistance among mosquito strains and pest resurgence, lead to the failure of chemical insecticide-based mosquito control methods. Even though it is more difficult to use and sustain, bio-control with ornamental fishes is preferable to chemical insecticides. One of its most significant benefits is that it is a method that protects the environment by not introducing toxins.

Therefore, it is recommended to step up research on many facets of fish biology and its function. It is important to adopt strategies like regular monitoring and surveying, identifying breeding areas, and raising public awareness. There is a lack of field research and studies on these fish's effectiveness as prospective larval control agents. Additionally, studies designed to evaluate the viability and effectiveness of these fish species in the wild may be useful in developing vector management strategies to combat *Aedes* mosquito population.

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References

- Das MK, Rao MRK, Kulsreshtha AK. Native larvivorous fish diversity as a biological control agent against mosquito larvae in an endemic malarious region of Ranchi district in Jharkhand, India. J Vector Borne Dis. 2018 Jan-Mar;55(1):34-41. Doi: 10.4103/0972-9062.234624. PMID: 29916446.
- Han WW, Lazaro A, McCall PJ, George L, Runge-Ranzinger S, Toledo J, *et al.* Efficacy and community effectiveness of larvivorous fish for dengue vector control. Trop Med Int Health. 2015 Sep;20(9):1239-1256. Doi: 10.1111/tmi.12538. Epub 2015 Jun 3. PMID: 25962851.
- Stanaway JD, Shepard DS, Undurraga EA, Halasa YA, Coffeng LE, Brady OJ, *et al.* The global burden of dengue: an analysis from the Global Burden of Disease Study 2013. Lancet Infectious Diseases. 2016;16(6):712-23.
- 4. World Health Organization. Yellow fever fact sheet. Updated May; c2016. http://www.who.int/mediacentre/factsheets/ fs100/en/ (accessed 29 June 2017).
- Powell JR, Tabachnick WJ. History of domestication and spread of *Aedes aegypti*: A review. Mem Inst Oswaldo Cruz. 2013;108(Suppl 1):11-7. Doi: 10.1590/0074-0276130395. PMID: 24473798; PMCID: PMC4109175.
- Manore CA, Ostfeld RS, Agusto FB, Gaff H, LaDeau SL. Defining the Risk of Zika and Chikungunya Virus Transmission in Human Population Centers of the Eastern United States. PLoS Negl Trop Dis. 2017;11(1):e0005255.

https://doi.org/10.1371/journal.pntd.0005255

 Weeratunga P, Rodrigo C, Fernando SD, Rajapakse S. Control methods for *Aedes albopictus* and *Aedes aegypti*. Cochrane Database Syst Rev. 2017 Aug 10;(8):CD012759. Doi: 10.1002/14651858.CD012759. PMCID: PMC6483697. Rochlin I, Ninivaggi DV, Hutchinson ML, Farajollahi A. Climate Change and Range Expansion of the Asian Tiger Mosquito (*Aedes albopictus*) in Northeastern USA: Implications for Public Health Practitioners. PLoS ONE. 2013;8(4):e60874.

https://doi.org/10.1371/journal.pone.0060874

- Hawley WA. The biology of *Aedes albopictus*. Journal of the American Mosquito Control Association. 1988;1:1-39.
- 10. Smith LB, Kasai S, Scott JG. Pyrethroid resistance in *Aedes aegypti* and *Aedes albopictus*: Important mosquito vectors of human diseases. Pesticide Biochemistry and Physiology. 2016;133:1-12.
- Jansen CC, Williams CR, van den Hurk AF. The Usual Suspects: Comparison of the Relative Roles of Potential Urban Chikungunya Virus Vectors in Australia. Plos One. 2015;10(8):e0134975. pmid:26247366
- Vega-Rua A, Zouache K, Girod R, Failloux AB, Lourenco-de-Oliveira R. High Level of Vector Competence of *Aedes aegypti* and *Aedes albopictus* from Ten American Countries as a Crucial Factor in the Spread of Chikungunya Virus. Journal of Virology. 2014;88(11):6294–6306. pmid:24672026
- Wong PSJ, Li MZI, Chong CS, Ng LC, Tan CH. Aedes (Stegomyia) albopictus (Skuse): A Potential Vector of Zika Virus in Singapore. Plos Neglected Tropical Diseases. 2013 Aug 1;7(8):e2348.
- 14. World Health Organization. Global Strategy for Dengue Prevention and Control 2012–2020 (in IRIS). World Health Organization: Geneva; c2012.
- Esu E, Lenhart A, Smith L, Horstick O. Effectiveness of peridomestic space spraying with insecticide on dengue transmission; systematic review. Trop Med Int Health. 2010 May;15(5):619-31. Doi: 10.1111/j.1365-3156.2010.02489.x. Epub 2010 Mar 8. PMID: 20214764.
- Flores AEPG, Silva BG, Gutierrez SM, Bobadilla C, Lopez B, Mercado R, *et al.* Wide spread cross resistance to pyrethroids in *Aedes aegypti* (Diptera: Culicidae) from Veracruz state Mexico. J Econ Entomol 2013:106(2):959-969.
- 17. Ranson HB, Burhani J, Lumjuan N, Black WC IV. Insecticide resistance in dengue vectors. 2010;1(1):01-12. (Available from: http://journal.tropika.net).
- Chandra G, Bhattacharjee I, Chatterjee SN, Ghosh A. Mosquito control by larvivorous fish 2008; Indian J Med Res. 2008 Jan 1;127(1):13.
- 19. Job TJ. An investigation of the nutrition of the perches of the Madras coast. Rec Mind Mus. 1940;42:289-364.
- Sharma VP, Sharma RC. Community based bioenvironmental control of malaria in Kheda district, Gujarat, India. J Amer Mosq Contr Assoc 1989;5(4):514-521.
- 21. Nelson S, Keenan L. Use of an indigenous fish species, *Fundulus zebrinus*, in a mosquito abatement program: a field comparison with the mosquitofish, Gambusia affinis. Am Mosq Control Assoc. 1992 Sep 1;8(3):301-304.
- 22. Sitaraman NL, Mahadevan S, Swami S. Biological control of Anopheles stephensi larvae in wells by Poecilia reticulata in Greater Hyderabad City, India J Com Dis. 1976;8(3):315319.
- Alio A, Isaq A, Delfini L. Field Trial on the Impact of Oreochromis spilurus Spilurus on Malaria Transmission in Northern Somalia, 1985. WHO:

(http://apps.who.int/iris/handle/ 10665/65921) 1985.

- 24. Menon PKB, Rajagopalan PK. Mosquito control potential of some species of indigenous fishes in Pondicherry. IJMR. 1977;66(5):765-771.
- 25. Yadav RS, Padhan K, Sharma VP. Fishes of District Sundargarh, Orissa, with special reference to their potential in mosquito control. Indian J Malariol 1992;29(5):225-233.
- 26. Tilak R, Dutta J, Dutta Gupta KK. Prospects for the Use of Ornamental Fishes for Mosquito Control: A Laboratory Investigation, Indian Journal of Public Health. 2007 Jan 1;51(1):54.
- 27. Câmara Miraldo, Marcel & PECORA, Iracy. Efficiency of Brazilian native ornamental fishes as mosquito larvae predators. Boletim do Instituto de Pesca, 2017;43:93-98. 10.20950/1678-2305.2017.93.98.
- Sumithra V, Janakiraman A, Altaff K. Bio-Control of Mosquito Larvae Through The Black Molly, Poecilia Sphenops. International Journal of Pure and Applied Zoology. 2014 Jan 1;2:270-274.
- 29. Datta A, Sen R, Banerjee R, Banerjee PK. Larvicidal efficacy of ornamental fishes to control *Aedes aegypti* in West Bengal. Int J Mosq Res. 2022;9(4):39-46. DOI: https://doi.org/10.22271/23487941.2022.v9.i4a.621
- 30. Gupta S, Banerjee S. Food preference of goldfish (*Carassius auratus* (Linnaeus, 1758)) and its potential in mosquito control. Electronic Journal of Ichthyology. 2009;5:47-58.