# Superantigens, superantigen-like proteins and superantigen derivatives for cancer treatment

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Abstract. - OBJECTIVE: Bacterial superantigens (SAgs) are proteins produced by few types of bacteria that have been linked to several human diseases. Due to their potent in vitro and in vivo tumoricidal effects, they are extensively investigated for oncological applications either alone or in combination with classical anticancer drugs. However, the intrinsic toxicity of natural SAgs stimulated the development of more effective and less toxic SAg-based immunotherapy. This review summarizes our current knowledge on SAg-based immunotherapy including SAg-like proteins and SAg derivatives, as well as their potential alone or with other therapeutic modalities including chemotherapy and radiotherapy.

*Key Words:* Superantigen, Superantigen derivative, Superantigen-like, Cancer, Combination therapy

## Introduction

Bacterial Superantigens (SAgs), also commonly known as erythrogenic toxins or streptococcal pyrogenic exotoxins<sup>1</sup>, are the most potent types of T cell mitogens. SAgs are produced by only a few bacterial pathogens, including Staphylococcus aureus<sup>2</sup>. The majority are produced by the Gram-positive organisms Staphylococcus aureus and Streptococcus pyogenes<sup>3,4</sup>. Bacterial SAgs at a very low concentration (typically < 0.1 pg/ml) are sufficient to stimulate the T lymphocytes<sup>5</sup>. In vitro femtomolar concentrations of SAgs are able to stimulate profound proliferation and cytokine production in up to 20% of all peripheral T cells<sup>6</sup>. It was initially thought that the powerful immune response generated by SAgs was due to its binding to Major Histocompatibility Complex class II molecules on antigen-presenting cells and T cell receptors on T cells. Later, it was found that the staphylococcal enterotoxin B (SEB) SAg, also binds the co-stimulatory molecule CD28 sug-

gesting a more complex mechanism of immune response<sup>6</sup>. Indeed, the current view is that SAgs bind to multiple coreceptors forming a costimulatory axis between coreceptors critical for T-cell activation7. CD28 is a homodimer expressed constitutively on T cells that interacts with its B7 coligands expressed on antigen-presenting cells, transducing the signal essential for T cell activation. The staphylococcal superantigen-like protein 1 (SSL1) specifically binds to human extracellular signal-regulated kinase 2 (hERK2), an important stress-activated kinase in mitogen-activated protein kinase signaling pathways<sup>8</sup>. It is now clearer that SAgs induce the release of cytokines and chemokines through multiple pathways as it was recently observed in in vitro experiments<sup>9</sup>.

SAgs can cause severe poisoning and several serious human diseases. For instance, *Staphylococcus aureus* enterotoxins have potent superantigenic activity associated with frequent food poisoning outbreaks<sup>10</sup>, toxic shock syndrome<sup>11,12</sup>, pneumonia<sup>13</sup>, Kawasaki disease<sup>14-17</sup>, nasopharyngeal infections<sup>18</sup>, atopic dermatitis (AD), and chronic rhinosinusitis (CRS) and sepsis-related infections.

SAgs can induce the production of cytokines leading to the hypothesis that viral proteins with a possible superantigen activity may be responsible for the systemic shock, acute respiratory syndrome, multiorgan failure and consequently death observed in patients with COVID-1919. A recent finding from *in silico* studies suggests that small insertions unique to SARS-CoV-2 (SARS-CoV-2 Spike) can display SAg activity and support this hypothesis<sup>20</sup>. It is relatively well established that at least one of the mechanisms by which SAgs contribute to systemic multiorgan failure is the production of a cytokine storm by enhancing the B7-2/CD28 costimulatory receptor interaction<sup>21,22</sup>. The formation of the B7-2/CD28 costimulatory axis is critical for full T-cell activation<sup>21</sup>.

Picomolar amounts of staphylococcal enterotoxin A (SEA) SAg rapidly induced cytotoxic activity against K562 and Raji cells, as well as some natural-killer (NK)-resistant tumour cell lines<sup>23</sup> which could be of value in therapeutic applications. Therefore, despite the serious toxic and adverse effects, SAgs are actively investigated as therapeutic tools for cancer treatment. The aim of this article is to provide a concise review of the potential application of SAgs, SAg-like proteins and SAg derivatives for cancer treatment.

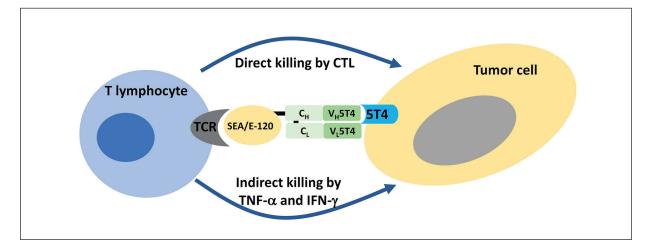
## SAgs for Cancer Treatment

For their ability to potentially activate T lymphocytes, SAgs have been used clinically as an immunomodifier in the treatment of tumors. The SAg Staphylococcal enterotoxin A (SEA) when co-cultured with human peripheral blood mononuclear cells (PBMCs) inhibited the proliferation and induced the death of human lung carcinoma A549 cells<sup>24</sup>. The Staphylococcal enterotoxin B (SEB) is an efficient activator of the antitumor immune response that leads to the eradication of tumor growth and inhibition of metastasis<sup>25</sup>. SEB-Superantigen-activated PBMC (peripheral blood mononuclear cells) significantly induced apoptosis in transitional cell carcinoma cells (TCC)<sup>26</sup>. The Staphylococcal enterotoxin C2 (SEC2) is another classical SAg with potent antitumor activity by activating T lymphocytes

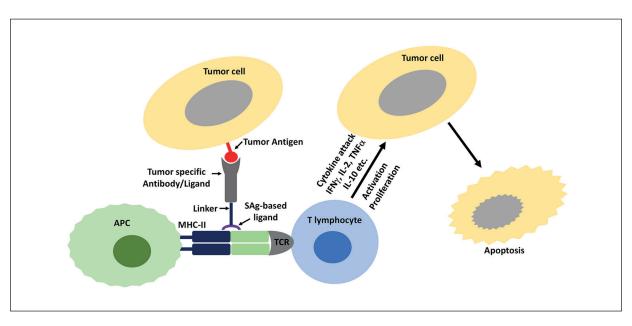
at very low dosage but of limited use due to toxic effects<sup>27</sup>. Natural superantigens have been used for cancer immunotherapy but at the cost of severe side effects due to their ability to induce high systemic levels of a large panel of inflammatory cytokines that may lead to a toxic shock syndrome and therefore attempts to translate the in vitro antitumor effect into clinical trials have been limited. To increase the effect of the SAgs, the concept of tumor-targeted SAgs (TTS) was established with the aim of recruiting a large number of T cells (Figure 1). In addition, because of the differential species susceptibilities, predicting the toxicity of SAgs in humans is difficult. For instance, mice are not valid human disease models because they are significantly less sensitive to toxic shock induced by bacterial SAgs when compared with primates and rabbits<sup>28</sup>. These limitations triggered the evaluation of other SAg alternatives such as SAg-like proteins and derivatives.

### SAg-Like Proteins

The common structure and function of the SAgs are also shared by another group of staphylococcal virulence factors called the superantigen-like proteins (SAg-like proteins)<sup>4</sup>. Staphylococcal enterotoxin-like toxins (SEIs) are proteins with similar amino acid sequences to those of classical SAgs but they exhibit low or no emetic



**Figure 1.** Potential immunotherapy using Tumor-Targeted Superantigen (SAg) (Modified from<sup>61</sup>). A SAg or SAg-based ligand can be linked to a tumor-specific antibody or ligand. The tumor-specific antibody/ligand binds to the tumor antigen whereas the superantigen/ligand crosslink between the major histocompatibility complex class II molecule (MHC-II) and the T cell receptor (TCR) induces T-cell hyperactivation of a T lymphocyte (shown in this figure) and monocytes/macrophages and results in the release of huge amounts of cytokines and chemokines, such as tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin 1 (IL-1), IL-2, interferon  $\gamma$  (IFN- $\gamma$ ). and many others. This leads to T-cell dependent tumor killing likely by apoptosis. APC = Antigen Presenting cell.



**Figure 2.** Potential mechanism of action of ABR-217620 (Naptumomab estafenatox; 5T4Fab-SEA/E-120). Modified from<sup>55</sup>. T lymphocyte activation occurs through its T-cell receptor (TCR) upon binding of the fusion protein ABR-217620 to the 5T4 tumor-associated antigen. The T cell kills tumoral cells directly by its cytotoxic T lymphocyte (CTL) activity and indirectly by producing cytokines (tumor necrosis factor [TNFa] – and interferon [IFNg ).

activity and have anticancer effects by activating lymphocytes particularly CD4<sup>+</sup> and CD8<sup>+</sup> T cells<sup>29</sup>. Treatment with a low concentration of Staphylococcal enterotoxin-like Q (SEIQ) (30 µg/ mouse) could inhibit the growth of tumors by approximately 30% without significant toxicity<sup>29</sup>. Staphylococcal SAg-like protein 6 (SSL6) inhibited CD47 and promoted Sorafenib-induced apoptosis of hepatocellular carcinoma cells Huh-7 and MHCC97H<sup>30</sup> that are widely used as liver cancer models. Other staphylococcal SAg-like protein such as staphylococcal SAg-like protein 7 (SSL7) have been shown to inhibit the formation of the complement membrane attack complex (MAC) and can be useful for treating complement-mediated hemolysis<sup>31</sup>. Recombinant staphylococcus aureus SAg-like protein 7 -that inhibits the complement- decreased the rate of tumor growth in a transplantable murine colon cancer model<sup>32</sup>. Humanized single-domain antibody (sdAb) mimicking the C-terminal domain of SSL7 may also be efficient in inhibiting complement-mediated hemolysis of erythrocytes from patients with paroxysmal nocturnal hemoglobinuria<sup>33</sup>. Staphylococcal SAg-like 10 was found to bind the chemokine receptor CXCR4 expressed on human T acute lymphoblastic leukemia, lymphoma, and cervical carcinoma cell lines inhibiting CXCL12-induced cell migration<sup>34</sup>.

### SAg Derivatives

SAg derivatives have been created to reduce systemic toxicity while maintaining profound antitumor effects. SAg side-effects may be caused by nonspecific binding to class II positive cells. To overcome this, several authors created mutated staphylococcal enterotoxin A (SEA). In particular, Hansson et al<sup>35</sup> created a tumor-reactive SAg by engineering a fusion protein composed of a tumor-reactive mAb (C215Fab) and the bacterial SAg staphylococcal enterotoxin A (SEA) (SEA), a well-known SAg. They introduced a point mutation (D227A) at the major MHC class II binding site. The Fab-SEA D227A fusion protein showed profound antitumor effects with a markedly reduced toxicity as compared with the wild-type Fab-SEA fusion protein. Kodama et al<sup>36</sup>, generated mutated SEA by changing Asp at position 227 of native SEA to Ala (mSEA-D227A), which has reduced affinity to MHC class II molecules, but retains the potential for T cell activation. mSEA-D227A was 500 times more tolerated compared to native SEA and had enhanced antitumor activity when conjugated to anti-MUC1 mAb.

SEA, when genetically conjugated to epidermal growth factor inhibited the proliferation of human nasopharyngeal carcinoma cell line CNE2 by promoting the proliferation of human peripheral blood mononuclear cells and enhancing the secretion of several cytokines -such as interferon- $\gamma$ , tumor necrosis factor- $\alpha$ , and interleukin-2 -that have broad antitumor activities<sup>37</sup>. Highly agglutinative staphylococcin (HAS) SAg, a derivative from Staphylococcus aureus, can inhibit and kill a variety of tumors cells as reviewed by Tian et al<sup>38</sup>. His-tagged SEC2 (SEC2-His) a derivative of the Staphylococcal enterotoxin C2 (SEC2) inhibited the growth of human colon adenocarcinoma (Caco-2) cells in vivo27. TGFaL3-SEB is another derivative created by fusing the third loop of transforming growth factor  $\alpha$  (TGF $\alpha$ L3) to staphylococcal enterotoxin type B (SEB) that significantly increased the tumor volume in mice bearing breast cancer without systemic toxicity<sup>39,40</sup>. Bio-products made up of proteins produced by Staphylococcus aureus potently inhibit tumour growth in a murine model of mesothelioma<sup>41</sup>. The SpeC<sub>D203A</sub>-based TTS fusion protein, an engineered human scFv that specifically targets human 5T4 (scFv5T4), was able to control the growth and spread of large tumors in an in vivo humanized mouse model of colon cancer<sup>42</sup>. The Mutant Staphylococcal enterotoxin C2 with lower toxic activity, named 2M-118 (H118A/T20L/ G22E) engineered by site-directed mutagenesis effectively inhibited the growth of S180 sarcoma with acceptable toxicity in the BALB/c mice $^{43}$ . The iRGD peptide fused superantigen mutant exhibited enhanced anti-solid tumor characteristics and induced improved lymphocyte infiltration in mouse B16F10 melanoma cells<sup>44</sup>. SAg derivatives for cancer immunotherapy can be also created by splitting into fragments, individually inactive, until both fragments came into close proximity and reassembled into a biologically active form capable of activating T cell response<sup>45</sup>.

## SAg-Based Treatments in Clinical Trials

Few SAg-based therapies have been tested in clinical trials for cancer. The Fab-SEA fusion protein (PNU-214565) made by fusing the superantigen staphylococcal enterotoxin A (SEA) with the Fab fragment of the monoclonal antibody C242 recognizing human colorectal (CRC) and pancreatic carcinomas (PC) was tested in patients with the aim to determine the maximum tolerated dose and it was found to be safe<sup>46,47</sup>. Similar results were reported in a cohort of patients with advanced gastrointestinal malignancies<sup>48</sup>. A different SEA-based fusion protein, PNU-214936, was evaluated in patients with advanced nonsmall-cell lung cancer to establish the maxi-

mum tolerated dose sing a Bayesian model of Escalation with Overdose Control (EWOC)<sup>49</sup>. Intratumoural injection of superantigen staphylococcal enterotoxin C (SEC) in patients with hepatocellular carcinoma (HCC) after percutaneous microwave ablation (PMWA) was found to be safe and to achieve longer overall survival as well as disease free survival<sup>50</sup>. ABR-214936, a fusion of a Fab recognizing the antigen 5T4, and Staphylococcal enterotoxin was tested in renal cell carcinoma patients. Patients who received higher drug exposure had greater disease control and lived almost twice as long as expected<sup>51</sup>. ABR-217620 (Naptumomab estafenatox; 5T4Fab-SEA/E-120) is an engineered antibody-superantigen fusion protein<sup>52</sup> that induces T-cell mediated killing of tumor cells at concentrations around 10 pM with low toxicity and reduced antigenicity<sup>53</sup>. Naptumomab estafenatox was created by fusing a superantigen (SAg) to the Fab moiety of a tumor-reactive monoclonal antibody. Naptumomab estafenatox targets a 72-kDa oncofetal trophoblast protein (the 5T4 tumor antigen) expressed on many carcinomas, including renal cell carcinoma and has shown anticancer activity in these cells<sup>54</sup>. Fab targeting of ABR-217620 to tumor cells where the SAg portion of the fusion protein elicits a potent tumoricidal cytotoxic T response and production of cytokines (TNF-alpha and IFN-gamma) are the main proposed mechanism of action<sup>55</sup>. At present there are 4 registered clinical trials (www.clinicaltrials.gov: NCT00420888, NCT03983954, NCT00056537, NCT00132379) evaluating the anticancer effects of Naptumomab estafenatox in human cancers. Despite the drug was well tolerated<sup>55</sup> in phase I studies, a phase II/III study to determine the efficacy of naptumomab estafenatox (Nap) + IFN $\alpha$ *vs.* IFN in metastatic renal cell carcinoma (RCC) did not meet its primary endpoint<sup>56</sup>.

Despite the scarcity of published clinical trials on SAg-based treatment in cancer patients, this therapeutic modality seems to be safe, well tolerated and more importantly they showed promising antitumor effects as reported in Phase I and II clinical trials.

## Combination Therapy

Tumors are complex and genetically and phenotypically heterogeneous and some cancers may not be treatable with just one strategy. A combination of traditional anticancer drugs with SAgbased therapies can help to overcome the drug resistance, lessen the symptoms and improve overall survival. Thus, effectiveness of SAgs, SAg-like proteins and SAg derivatives can be increased by combination therapy with traditional FDA-approved anticancer drugs that have been proven to give superior results when used in combination<sup>57</sup>. Additionally, this combination therapy can be added to tumors that are routinely treated with radiotherapy such as gliomas<sup>58</sup> and esophageal cancer<sup>59</sup>. Table I shows examples of combinations therapy using SAg-based therapies with traditional chemotherapy +/- radiotherapy.

## Conclusions

Although SAgs have been associated with several diseases and serious adverse effects, they are extensively investigated for oncological applications either alone or in combination with classical anticancer drugs. The potent antitumor activity of natural SAgs is limited by their high intrinsic toxicity. However, a plethora of SAg-like proteins and derivatives with lower toxicity are already available and new ones are under development.

SAg	Adjuvant	Cell type/tumor model	Effect	Ref.
SAg	Doxorubicin	Urothelial urinary bladder cancer	Iincreased CD4+ T cell activation	62
Tumor-targeted superantigen	Tasquinimod	Melanoma cells	Increased Tumor-specific CD8(+)	63
Microbiota-derived Staphylococcal superantigen-like protein 6 (SSL6)	Sorafenib (SFN)	Hepatocellular Carcinoma	Enhances SFN sensitivity	30
Staphylococcal enterotoxin-B	Chimeric antigen receptor (CAR) T cell	Mice bearing established E0771-Her2 tumors	Tumor-growth inhibition	64
Superantigen staphylococcal enterotoxin B	Ipilimumab and Nivolumab	Mouse MC38 and CT26 colorectal tumor models	Increased anti-Tumor activity	65
Staphylococcal enterotoxin C (SEC)	Surgery + Chemotherapy + Radiotherapy	Glioma patients	Increased "effective rate"	58
ABR-217620	Docetaxel	Patients with non-small-cell lung cancer (NSCLC), pancreatic cancer (PC), and renal cell cancer (RCC)	Evidence of immunological and antitumor activity.	55
Staphylococcal enterotoxin A (SEA) in fusion with an anti-tumor Fab-fragment	Docetaxel	B16-C215 tumors growing in the lung of C57Bl/6 mice	Prolonged long term survival	66
C215Fab-SEA fusion protein	Linomide	Syngenic B16 melanoma cells transfected with GA733-2 (a human colon cancer cell surface antigen)	> 99% reduction of liver metastasis in C57/Bl6 mice.	67
Staphylococcal enterotoxin A (SEA) in fusion with an anti-tumor Fab-fragment	Interferon alpha	B16-C215 tumors growing in the lung of C57Bl/6 mice	Synergistic anti-tumor effects, prolonged survival	68
Staphylococcal enterotoxin-A (SEA),	Staphylococcal protein-A (PA)	Animals carrying the Ehrlich ascites tumor.	Long-term survival as compared with PA or SEA alone	69
Fab-SEA	Fab-IL-2	B16 melanoma	Cure in 90% of tumo -bearing animals	70

**Table I.** Combination therapy using SAg-based therapy.

Another approach to decrease the toxicity of SAgbased immunotherapy is to create SAg variants with less vasodilation effect but more tolerable for cancer immunotherapy as has been reported by Bashraheel et al<sup>60</sup>. In summary, the potent antitumor activity of naturally occurring SAgs can be exploited by identifying and developing less toxic SAg-like proteins and SAg derivatives and especially by using them in combination therapy. At present the effectiveness of this approach has been tested in few clinical trials, mostly limited to phase I and phase II studies with encouraging results in terms of safety and antitumor activity that warrants further evaluation in larger phase III studies.

#### **Conflict of Interest**

The Authors declare that they have no conflict of interests.

#### References

- 1) McCormick JK, Yarwood JM, Schlievert PM. Toxic shock syndrome and bacterial superantigens: an update. Annu Rev Microbiol 2001; 55: 77-104.
- Xu SX, McCormick JK. Staphylococcal superantigens in colonization and disease. Front Cell Infect Microbiol 2012; 2: 52.
- Alouf JE, Müller-Alouf H. Staphylococcal and streptococcal superantigens: molecular, biological and clinical aspects. Int J Med Microbiol 2003; 292: 429-440.
- Fraser JD, Proft T. The bacterial superantigen and superantigen-like proteins. Immunol Rev 2008; 225: 226-243.
- 5) Proft T, Fraser JD. Bacterial superantigens. Clin Exp Immunol 2003; 133: 299-306.
- 6) Fraser JD. Clarifying the mechanism of superantigen toxicity. PLoS Biol 2011; 9: e1001145.
- Kaempfer R, Popugailo A, Levy R, Arad G, Hillman D, Rotfogel Z. Bacterial superantigen toxins induce a lethal cytokine storm by enhancing B7-2/ CD28 costimulatory receptor engagement, a critical immune checkpoint. Receptors Clin Investig 2017; 4: e1500.
- Dutta D, Mukherjee D, Mukherjee IA, Maiti TK, Basak A, Das AK. Staphylococcal superantigen-like proteins interact with human MAP kinase signaling protein ERK2. FEBS Lett 2020; 594: 266-277.
- Liu X, Wen Y, Wang D, Zhao Z, Jeffry J, Zeng L, Zou Z, Chen H, Tao A. Synergistic activation of Src, ERK and STAT pathways in PBMCs for Staphylococcal enterotoxin A induced production of cytokines and chemokines. Asian Pac J Allergy Immunol 2020; 38: 52-63.

- Fisher EL, Otto M, Cheung GYC. Basis of Virulence in Enterotoxin-Mediated Staphylococcal Food Poisoning. Front Microbiol 2018; 9: 436.
- Krakauer T. Staphylococcal Superantigens: Pyrogenic Toxins Induce Toxic Shock. Toxins (Basel) 2019; 11: 178.
- 12) Tilahun AY, Chowdhary VR, David CS, Rajagopalan G. Systemic inflammatory response elicited by superantigen destabilizes T regulatory cells, rendering them ineffective during toxic shock syndrome. J Immunol 2014; 193: 2919-2930.
- 13) Karau MJ, Tilahun ME, Krogman A, Osborne BA, Goldsby RA, David CS, Mandrekar JN, Patel R, Rajagopalan G. Passive therapy with humanized anti-staphylococcal enterotoxin B antibodies attenuates systemic inflammatory response and protects from lethal pneumonia caused by staphylococcal enterotoxin B-producing Staphylococcus aureus. Virulence 2017; 8: 1148-1159.
- 14) Blankier S, McCrindle BW, Ito S, Yeung RS. The role of atorvastatin in regulating the immune response leading to vascular damage in a model of Kawasaki disease. Clin Exp Immunol 2011; 164: 193-201.
- Leung DYM, Schlievert PM. Kawasaki syndrome: role of superantigens revisited. FEBS J 2020. Aug 08. doi.org/101111/febs.15512.
- 16) Natividad MF, Torres-Villanueva CA, Saloma CP. Superantigen involvement and susceptibility factors in Kawasaki disease: profiles of TCR Vβ2+ T cells and HLA-DRB1, TNF-α and ITPKC genes among Filipino patients. Int J Mol Epidemiol Genet 2013; 4: 70-76.
- Suenaga T, Suzuki H, Shibuta S, Takeuchi T, Yoshikawa N. Detection of multiple superantigen genes in stools of patients with Kawasaki disease. J Pediatr 2009; 155: 266-270.
- 18) Kasper KJ, Zeppa JJ, Wakabayashi AT, Xu SX, Mazzuca DM, Welch I, Baroja ML, Kotb M, Cairns E, Cleary PP, Haeryfar SM, McCormick JK. Bacterial superantigens promote acute nasopharyngeal infection by Streptococcus pyogenes in a human MHC Class II-dependent manner. PLoS Pathog 2014; 10: e1004155.
- Scaglioni V, Soriano ER. Are superantigens the cause of cytokine storm and viral sepsis in severe COVID-19? Observations and hypothesis. Scand J Immunol 2020; 92: e12944.
- 20) Cheng MH, Zhang S, Porritt RA, Arditi M, Bahar I. An insertion unique to SARS-CoV-2 exhibits superantigenic character strengthened by recent mutations. bioRxiv [Preprint]. 2020 May 21:2020.05.21.109272.
- Levy R, Rotfogel Z, Hillman D, Popugailo A, Arad G, Supper E, Osman F, Kaempfer R. Superantigens hyperinduce inflammatory cytokines by enhancing the B7-2/CD28 costimulatory receptor interaction. Proc Natl Acad Sci U S A 2016; 113: E6437-e6446.
- 22) Popugailo A, Rotfogel Z, Supper E, Hillman D, Kaempfer R. Staphylococcal and Streptococcal

Superantigens Trigger B7/CD28 Costimulatory Receptor Engagement to Hyperinduce Inflammatory Cytokines. Front Immunol 2019; 10: 942.

- 23) Lando PA, Hedlund G, Dohlsten M, Kalland T. Bacterial superantigens as anti-tumour agents: induction of tumour cytotoxicity in human lymphocytes by staphylococcal enterotoxin A. Cancer Immunol Immunother 1991; 33: 231-237.
- 24) Liu X, Zeng L, Zhao Z, He J, Xie Y, Xiao L, Wang S, Zhang J, Zou Z, He Y, Tao A, Zhang J. PBMC activation via the ERK and STAT signaling pathways enhances the anti-tumor activity of Staphylococcal enterotoxin A. Mol Cell Biochem 2017; 434: 75-87.
- 25) Heidary MF, Mahmoodzadeh Hosseini H, Mehdizadeh Aghdam E, Nourani MR, Ranjbar R, Mirnejad R, Imani Fooladi AA. Overexpression of Metastatic Related MicroRNAs, Mir-335 and Mir-10b, by Staphylococcal Enterotoxin B in the Metastatic Breast Cancer Cell Line. Adv Pharm Bull 2015; 5: 255-259.
- 26) Perabo FG, Willert PL, Wirger A, Schmidt DH, Von Ruecker A, Mueller SC. Superantigen-activated mononuclear cells induce apoptosis in transitional cell carcinoma. Anticancer Res 2005; 25: 3565-3573.
- 27) Zhao W, Li Y, Liu W, Ding D, Xu Y, Pan L, Chen S. Transcytosis, Antitumor Activity and Toxicity of Staphylococcal Enterotoxin C2 as an Oral Administration Protein Drug. Toxins (Basel) 2016; 8: 185.
- 28) Yeung RS, Penninger JM, Kündig T, Khoo W, Ohashi PS, Kroemer G, Mak TW. Human CD4 and human major histocompatibility complex class II (DQ6) transgenic mice: supersensitivity to superantigen-induced septic shock. Eur J Immunol 1996; 26: 1074-1082.
- 29) He Y, Sun Y, Ren Y, Qiao L, Guo R, Du J, Zhu X, Liu Y, Lin J. The T cell activating properties and antitumour activity of Staphylococcal Enterotoxin-like Q. Med Microbiol Immunol 2019; 208: 781-792.
- 30) Zhang X, Wu L, Xu Y, Yu H, Chen Y, Zhao H, Lei J, Zhou Y, Zhang J, Wang J, Peng J, Jiang L, Sheng H, Li Y. Microbiota-derived SSL6 enhances the sensitivity of hepatocellular carcinoma to sorafenib by down-regulating glycolysis. Cancer Lett 2020; 481: 32-44.
- Li Y, Clow F, Fraser JD, Lin F. Therapeutic potential of staphylococcal superantigen-like protein 7 for complement-mediated hemolysis. J Mol Med (Berl) 2018; 96: 965-974.
- 32) Downs-Canner S, Magge D, Ravindranathan R, O'Malley ME, Francis L, Liu Z, Sheng Guo Z, Obermajer N, Bartlett DL. Complement Inhibition: A Novel Form of Immunotherapy for Colon Cancer. Ann Surg Oncol 2016; 23: 655-662.
- 33) Yatime L, Merle NS, Hansen AG, Friis NA, Østergaard JA, Bjerre M, Roumenina LT, Thiel S, Kristensen P, Andersen GR. A Single-Domain Antibody Targeting Complement Component C5 Acts

as a Selective Inhibitor of the Terminal Pathway of the Complement System and Thus Functionally Mimicks the C-Terminal Domain of the Staphylococcus aureus SSL7 Protein. Front Immunol 2018; 9: 2822.

- 34) Walenkamp AM, Boer IG, Bestebroer J, Rozeveld D, Timmer-Bosscha H, Hemrika W, van Strijp JA, de Haas CJ. Staphylococcal superantigen-like 10 inhibits CXCL12-induced human tumor cell migration. Neoplasia 2009; 11: 333-344.
- 35) Hansson J, Ohlsson L, Persson R, Andersson G, Ilbäck NG, Litton MJ, Kalland T, Dohlsten M. Genetically engineered superantigens as tolerable antitumor agents. Proc Natl Acad Sci U S A 1997; 94: 2489-2494.
- 36) Kodama H, Suzuki M, Katayose Y, Shinoda M, Sakurai N, Takemura S, Yoshida H, Saeki H, Ichiyama M, Tsumoto K, Asano R, Kumagai I, Imai K, Hinoda Y, Matsuno S, Kudo T. Mutated SEA-D227A-conjugated antibodies greatly enhance antitumor activity against MUC1-expressing bile duct carcinoma. Cancer Immunol Immunother 2001; 50: 539-548.
- 37) Liu X, Zeng L, Zhao Z, Xie Y, Wang S, Zhang J, He Y, Zou Z, Zhang J, Tao A. Construction, Expression, and Characterization of rSEA-EGF and In Vitro Evaluation of its Antitumor Activity Against Nasopharyngeal Cancer. Technol Cancer Res Treat 2018; 17: 1533033818762910.
- 38) Tian XL, Yan Z, Chen J, Zhao WH, Guo W. Clinical application of highly agglutinative staphylococcin in cancer treatment updates of the literature. Eur Rev Med Pharmacol Sci 2016; 20: 2718-2725.
- 39) Yousefi F, Mousavi SF, Siadat SD, Aslani MM, Amani J, Rad HS, Fooladi AA. Preparation and In Vitro Evaluation of Antitumor Activity of TG-FαL3-SEB as a Ligand-Targeted Superantigen. Technol Cancer Res Treat 2016; 15: 215-226.
- 40) Yousefi F, Siadat SD, Saraji AA, Hesaraki S, Aslani MM, Mousavi SF, Imani Fooladi AA. Tagging staphylococcal enterotoxin B (SEB) with TGFaL3 for breast cancer therapy. Tumour Biol 2016; 37: 5305-5316.
- 41) Lansley SM, Varano Della Vergiliana JF, Cleaver AL, Ren SH, Segal A, Xu MY, Lee YC. A commercially available preparation of Staphylococcus aureus bio-products potently inhibits tumour growth in a murine model of mesothelioma. Respirology 2014; 19: 1025-1033.
- 42) Patterson KG, Dixon Pittaro JL, Bastedo PS, Hess DA, Haeryfar SM, McCormick JK. Control of established colon cancer xenografts using a novel humanized single chain antibody-streptococcal superantigen fusion protein targeting the 5T4 oncofetal antigen. PLoS One 2014; 9: e95200.
- 43) Zhang J, Cai YM, Xu MK, Song ZH, Li CY, Wang HR, Dai HH, Zhang ZP, Liu CX. Anti-tumor activity and immunogenicity of a mutated staphylococcal enterotoxin C2. Pharmazie 2013; 68: 359-364.
- 44) Song Y, Xu M, Li Y, Li Y, Gu W, Halimu G, Fu X, Zhang H, Zhang C. An iRGD peptide fused supe-

rantigen mutant induced tumor-targeting and T lymphocyte infiltrating in cancer immunotherapy. Int J Pharm 2020; 586: 119498.

- Golob-Urbanc A, Rajčević U, Strmšek Ž, Jerala R. Design of split superantigen fusion proteins for cancer immunotherapy. J Biol Chem 2019; 294: 6294-6305.
- 46) Giantonio BJ, Alpaugh RK, Schultz J, McAleer C, Newton DW, Shannon B, Guedez Y, Kotb M, Vitek L, Persson R, Gunnarsson PO, Kalland T, Dohlsten M, Persson B, Weiner LM. Superantigen-based immunotherapy: a phase I trial of PNU-214565, a monoclonal antibody-staphylococcal enterotoxin A recombinant fusion protein, in advanced pancreatic and colorectal cancer. J Clin Oncol 1997; 15: 1994-2007.
- 47) Nielsen SE, Zeuthen J, Lund B, Persson B, Alenfall J, Hansen HH. Phase I study of single, escalating doses of a superantigen-antibody fusion protein (PNU-214565) in patients with advanced colorectal or pancreatic carcinoma. J Immunother 2000; 23: 146-153.
- 48) Alpaugh RK, Schultz J, McAleer C, Giantonio BJ, Persson R, Burnite M, Nielsen SE, Vitek L, Persson B, Weiner LM. Superantigen-targeted therapy: phase I escalating repeat dose trial of the fusion protein PNU-214565 in patients with advanced gastrointestinal malignancies. Clin Cancer Res 1998; 4: 1903-1914.
- 49) Cheng JD, Babb JS, Langer C, Aamdal S, Robert F, Engelhardt LR, Fernberg O, Schiller J, Forsberg G, Alpaugh RK, Weiner LM, Rogatko A. Individualized patient dosing in phase I clinical trials: the role of escalation with overdose control in PNU-214936. J Clin Oncol 2004; 22: 602-609.
- 50) Zhou P, Liang P, Dong B, Yu X, Han X, Wang Y, Han Z. Long-term results of a phase II clinical trial of superantigen therapy with staphylococcal enterotoxin C after microwave ablation in hepatocellular carcinoma. Int J Hyperthermia 2011; 27: 132-139.
- 51) Shaw DM, Connolly NB, Patel PM, Kilany S, Hedlund G, Nordle O, Forsberg G, Zweit J, Stern PL, Hawkins RE. A phase II study of a 5T4 oncofoetal antigen tumour-targeted superantigen (ABR-214936) therapy in patients with advanced renal cell carcinoma. Br J Cancer 2007; 96: 567-574.
- 52) Hedlund G, Eriksson H, Sundstedt A, Forsberg G, Jakobsen BK, Pumphrey N, Rödström K, Lindkvist-Petersson K, Björk P. The tumor targeted superantigen ABR-217620 selectively engages TRBV7-9 and exploits TCR-pMHC affinity mimicry in mediating T cell cytotoxicity. PLoS One 2013; 8: e79082.
- 53) Forsberg G, Skartved NJ, Wallén-Ohman M, Nyhlén HC, Behm K, Hedlund G, Nederman T. Naptumomab estafenatox, an engineered antibody-superantigen fusion protein with low toxicity and reduced antigenicity. J Immunother 2010; 33: 492-499.
- 54) Eisen T, Hedlund G, Forsberg G, Hawkins R. Naptumomab estafenatox: targeted immunother-

apy with a novel immunotoxin. Curr Oncol Rep 2014; 16: 370.

- 55) Borghaei H, Alpaugh K, Hedlund G, Forsberg G, Langer C, Rogatko A, Hawkins R, Dueland S, Lassen U, Cohen RB. Phase I dose escalation, pharmacokinetic and pharmacodynamic study of naptumomab estafenatox alone in patients with advanced cancer and with docetaxel in patients with advanced non-small-cell lung cancer. J Clin Oncol 2009; 27: 4116-4123.
- 56) Hawkins RE, Gore M, Shparyk Y, Bondar V, Gladkov O, Ganev T, Harza M, Polenkov S, Bondarenko I, Karlov P, Karyakin O, Khasanov R, Hedlund G, Forsberg G, Nordle Ö, Eisen T. A Randomized Phase II/III Study of Naptumomab Estafenatox + IFNα versus IFNα in Renal Cell Carcinoma: Final Analysis with Baseline Biomarker Subgroup and Trend Analysis. Clin Cancer Res 2016; 22: 3172-3181.
- 57) Palmer AC, Sorger PK. Combination Cancer Therapy Can Confer Benefit via Patient-to-Patient Variability without Drug Additivity or Synergy. Cell 2017; 171: 1678-1691.e1613.
- 58) Wang F, Huang Q, Zhou LY. Analysis of the treatment of gliomas with SEC therapy combined with radiochemotherapy. Eur Rev Med Pharmacol Sci 2015; 19: 2400-2405.
- 59) Heidecke CD, Weighardt H, Feith M, Fink U, Zimmermann F, Stein HJ, Siewert JR, Holzmann B. Neoadjuvant treatment of esophageal cancer: Immunosuppression following combined radiochemotherapy. Surgery 2002; 132: 495-501.
- 60) Bashraheel SS, AlQahtani AD, Rashidi FB, Al-Sulaiti H, Domling A, Orie NN, Goda SK. Studies on vascular response to full superantigens and superantigen derived peptides: Possible production of novel superantigen variants with less vasodilation effect for tolerable cancer immunotherapy. Biomed Pharmacother 2019; 115: 108905.
- 61) Bashraheel SS, Domling A, Goda SK. Update on targeted cancer therapies, single or in combination, and their fine tuning for precision medicine. Biomed Pharmacother 2020; 125: 110009.
- 62) Zirakzadeh AA, Kinn J, Krantz D, Rosenblatt R, Winerdal ME, Hu J, Hartana CA, Lundgren C, Bergman EA, Johansson M, Holmström B, Hansson J, Sidikii A, Vasko J, Marits P, Sherif A, Winqvist O. Doxorubicin enhances the capacity of B cells to activate T cells in urothelial urinary bladder cancer. Clin Immunol 2017; 176: 63-70.
- 63) Shen L, Sundstedt A, Ciesielski M, Miles KM, Celander M, Adelaiye R, Orillion A, Ciamporcero E, Ramakrishnan S, Ellis L, Fenstermaker R, Abrams SI, Eriksson H, Leanderson T, Olsson A, Pili R. Tasquinimod modulates suppressive myeloid cells and enhances cancer immunotherapies in murine models. Cancer Immunol Res 2015; 3: 136-148.
- 64) von Scheidt B, Wang M, Oliver AJ, Chan JD, Jana MK, Ali Al, Clow F, Fraser JD, Quinn KM, Darcy PK, Kershaw MH, Slaney CY. Enterotoxins can

support CAR T cells against solid tumors. Proc Natl Acad Sci U S A 2019; 116: 25229-25235.

- 65) Selby MJ, Engelhardt JJ, Johnston RJ, Lu LS, Han M, Thudium K, Yao D, Quigley M, Valle J, Wang C, Chen B, Cardarelli PM, Blanset D, Korman AJ. Preclinical Development of Ipilimumab and Nivolumab Combination Immunotherapy: Mouse Tumor Models, In Vitro Functional Studies, and Cynomolgus Macaque Toxicology. PLoS One 2016; 11: e0161779.
- 66) Sundstedt A, Celander M, Ohman MW, Forsberg G, Hedlund G. Immunotherapy with tumor-targeted superantigens (TTS) in combination with docetaxel results in synergistic anti-tumor effects. Int Immunopharmacol 2009; 9: 1063-1070.
- 67) Liu Q, Klintman D, Corbascio M, Ekberg H, Hedlund G, Forsberg G, Thorlacius H. Linomide and

antibody-targeted superantigen therapy abolishes formation of liver metastases in mice. Eur Surg Res 2003; 35: 457-463.

- 68) Sundstedt A, Celander M, Hedlund G. Combining tumor-targeted superantigens with interferon-alpha results in synergistic anti-tumor effects. Int Immunopharmacol 2008; 8: 442-452.
- 69) Mondal TK, Bhatta D, Biswas S, Pal P. Repeated treatment with S. aureus superantigens expands the survival rate of Ehrlich ascites tumor bearing mice. Immunol Invest 2002; 31: 13-28.
- 70) Rosendahl A, Kristensson K, Carlsson M, Skartved NJ, Riesbeck K, Søgaard M, Dohlsten M. Long-term survival and complete cures of B16 melanoma-carrying animals after therapy with tumor-targeted IL-2 and SEA. Int J Cancer 1999; 81: 156-163.

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