


**EVOLUTION OF *Fusarium* TAXONOMY:  
MORPHOLOGICAL, BIOLOGICAL  
AND PHYLOGENETIC DIAGNOSTIC  
CONCEPTS**

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Regional Workshop on the prevention and diagnostic of *Fusarium* Wilt (Panama disease) of bananas and plantains caused by *Fusarium oxysporum cubensis* – Tropical Race 4 (TR4)  
Port Spain, Trinidad and Tobago April 28th-May 9th, 2014



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**FUSARIUM**

Maize ear  
Dry rot of potato tubers by *Fusarium* spp.  
Fusarium verticillioides  
Fusarium  
Fusarium

**OMNIPRESENT IN SOIL, WATER,  
AIR AND OTHER MULTIPLE  
SUBSTRATES; ARE  
SAPROPHYTES, PATHOGENS,  
ANTAGONISTS**

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***Fusarium* species as plant pathogens**

- ✓ *Fusarium* species are human, animal and plant pathogens
- ✓ *Fusarium* spp. cause diseases in a wide range of host plants. At least 81 of 101 economically important crop plants of APS list of diseases are caused by *Fusarium*
- ✓ Some of them have a narrow range of hosts; others have a very wide range as *F. oxysporum*
- ✓ They are found in soil, air water and can be transported in vegetable tissues.
- ✓ They can be recovered from the deepest roots in soil until the higher inflorescences of plants.
- ✓ Produce toxins and allergies



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**Diseases caused by different *Fusarium* species**



Sugarcane wilt by *Fusarium sacchari*

Pokkah Boeng in sugarcane by *Fusarium moniliforme*



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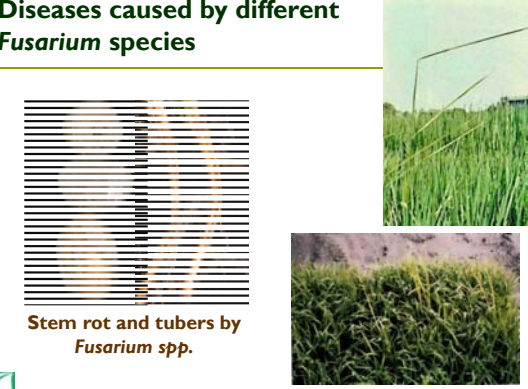
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**Diseases caused by different *Fusarium* species**



Stem rot and tubers by *Fusarium spp.*

Bakanae in rice by *F. fujikuroi*



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**Diseases caused by different *Fusarium* species**



Green point gall of cacao by *Nectria rigidiuscula* (*Fusarium decemcellulare*)

Flowering malformation of mango by (*Fusarium mangifera*)



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**Diseases caused by different *Fusarium* species**



Oil palm wilt by *F. oxysporum* f. sp. *elaeidis*



*F. oxysporum* f. sp. *lycopersici*



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**Diseases caused by different *Fusarium* species**



Panama disease by *F. oxysporum* f. sp. *cubensis*



Crown rot by *F. pallidoroseum*



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**Diseases caused by different *Fusarium* species**



*Gibberella zeae* (*F. graminearum*)



Papaya internal fruit rot by *Fusarium* spp.



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### Fusarium taxonomy

- ✓ The taxonomy of *Fusarium* genus has been affected by species concept changes
- ✓ In this process in the last 100 years have been recognized by taxonomist, as few as 9 and as much as 1000 species.




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### Taxonomy of *Fusarium* genus: Historic review

- 1809. Link create genus *Fusarium*
- 1910. Appel and Wollenweber (1910) grouped all imperfect fungi Tuberculariaceae that have pluricelular macroconidia with croissant shape developed on sporodochia.
- 1935. Wollenweber y Reinking publish the monography "Die *Fusarien*" and split genera in 16 sections and 65 species.
- 1940. Snyder & Hansen questioned sections described by Wollenweber & Reinking and reduce all variants to 10 species. Beside propose that all the forms of *Elegans* section belong to a single specie: *F. oxysporum*
- 1955. V.I. Bilai publish in URSS "The *Fusaria* (Biology and Systematics)"
- 1968. C.M. Messiaen and R. Cassini publish "La systématique des *Fusarium*"; essentially kept the classification of Snyder & Hansen and propose varieties of some species .




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### Taxonomy of *Fusarium* genus: Historic review

- 1971. C. Booth publish The Genus *Fusarium*. CMI. (recognize 14 species)
- 1981. Nelson Tousson y Manasas publish "*Fusarium* Species: An Illustrated Manual for Identification".
- 1982. W. Gerlach y H. Nirenberg publish "The genus *Fusarium* – A pictorial Atlas". (recognize 21 species)
- 1980s. The collaboration among taxonomists of EU, Europe, South Africa, and Australia unify criteria of Gerlach and Nirenberg (Germany), Nelson, Tousson y Marasas in (EU) and Burgess et al. (Australia).
- 1994. L.W. Burgess et al., publish "Laboratory Manual for *Fusarium* Research"
- 2006 J. Leslie and B. Summerell publish "The *Fusarium* Laboratory Manual" recognize 70 species on base of morphological, biological and phylogenetic criteria.




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**Relationship among the nine species of Snyder & Hansen, the sections of Wollenweber & Reinking and the species described by Leslie and Summerett (2006)**

Species of Snyder & Hansen	Sections of Wollenweber & Reinking	Summerett & Leslie (2006)
<i>F. episphaeria</i>	Eupionnotes Macroconia	<i>F. dimerum</i> , <i>F. merismoides</i>
<i>F. lateritium</i>	Lateritium	<i>F. lateritium</i>
<i>F. moniliforme</i>	Liseola	<i>F. anthophilum</i> , <i>F. circinatum</i> , <i>F. proliferatum</i> , <i>F. subglutinans</i> , <i>F. thapsinum</i> , <i>F. verticillioides</i> , and other species in
	Fusarium complex	<i>Gibberella fujikuroi</i>
<i>F. nivale</i>	Arachnites	No more consider be Fusarium
<i>F. oxysporum</i>	Elegans	<i>F. oxysporum</i>
<i>F. rigidiuscula</i>	Spicarioides	<i>F. decemcellulare</i>

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**Relationship among the nine species of Snyder & Hansen, the sections of Wollenweber & Reinking and the species described by Leslie and Summerett (2006)**

Species of Snyder & Hansen	Sections of Wollenweber & Reinking	Summerett & Leslie (2006)
<i>F. roseum</i>	Discolor Gibbosum Roseum Arthrosporiella	<i>F. acuminatum</i> , <i>F. armeniacum</i> <i>F. avenaceum</i> , <i>F. compactum</i> ; <i>F. crookwellense</i> , <i>F. culmorum</i> , <i>F. equiseti</i> , <i>F. graminearum</i> , <i>F. longipes</i> , <i>F. heterosporum</i> , <i>F. polyphialidicum</i> , <i>F. pseudograminearum</i> , <i>F. semitectum</i> , <i>F. torulosum</i>
<i>F. solani</i>	Martiella Ventricosum	<i>F. solani</i>
<i>F. tricinctum</i>	Sporotrichiella	<i>F. chlamydosporum</i> , <i>F. poae</i> , <i>F. sporotrichioides</i> , <i>F. tricinctum</i>

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***Fusarium oxysporum* Schlecht. ex Fr**

- ✓ *F. oxysporum* is a complex of anamorphic fungal filamentous species morphologically similars (O'Donnell and Cigelnick, 1998).
- ✓ *F. oxysporum* is omnipresent around the world in different soils.
- ✓ Is the *Fusarium* taxa more economical and agricultural important (Ploetz, 2006)
- ✓ Isolations of *F. oxysporum* are genetically diverse including phytopathogens, saprophytes and biocontrol agents.
- ✓ They include many plant pathogenic representatives. Cause:
  - Vascular wilts,
  - Damping-off problems
  - Root and stalk base rots
- ✓ Have been identified 150 formae speciales with unique pathogenicity or to at an important number of close related hosts




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**Fusarium oxysporum Schlecht. ex Fr**

- ✓ Wilts are important in many vegetable, fiber, ornamentals palm trees and banana.
- ✓ Pastures and cereals are not affected by *F. oxysporum*.
- ✓ Saprophytic populations usually can be aggressive secondary colonizers of diseased plant parts, particularly roots and are morphologically undistinguished from primary colonizers
- ✓ Morphology of *F. oxysporum* colonies are highly variable:
  - Produce a floccose sparse or abundant, white to pale violet mycelia.
  - Usually produce a pale violet to dark magenta pigment in agar media ( some isolates do not produce any pigment)




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**Species definition concepts**

The concepts essentially define criteria to differentiate species. There is three basic concept applied for *Fusarium* :

- **Morphologic:** (constant observable morphological characters among individuals of the same specie and markedly different between different species).
- **Biological:** (crosses among members of the same specie are sexually fertile with viable and fertile progeny).
- **Phylogenetic:** (ADN sequences of conserved genes are cladistically treated to develop phylogenies of those which are grouped in the same group with a common origin, are considered to be of the same specie)




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**Morphological criteria for specie identification in *Fusarium***

- ✓ **Morphological characteristics of structures in carnation leaf agar)**
  - Macro and microconidia (form, abundance, size, how are produced (better from a CLA culture)
  - Chlamydo spores (present/absent; single, agglomerates)
  - Conidiophores (mono o poliphyalidics)
- ✓ **Cultural characteristics**
  - Growth rate (in PDA)
  - Aerial mycelia absent or present and color (PDA)
  - Presence and color of sporodochia in (PDA and CLA)
  - Color of colonies in the superior and inferior part (PDA)
  - Production of volatile aldehydes (odor; rice grain media)




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**Macroconidia cells** (Leslie y Summerell, 2006)

Basal cells:  
 E) *F. culmorum*; F) *F. crookwellense*;  
 G) *F. avenaceum*; H) *F. culmorum*

Apical cells:  
 I) *F. culmorum*; J) *F. decemcellulare*;  
 K) *F. avenaceum*; L) *F. longipes*

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**Microconidia shapes** (Summerell et al. 2003)

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**Conidiogenic cells of microconidia phialid.**

Microconidia in monophialids  
 Chains  
*F. verticilloides*  
 False head  
*F. solani* *F. oxysporum*

Microconidia in poliphialids  
*F. clamidosporium*  
*F. semitectum*

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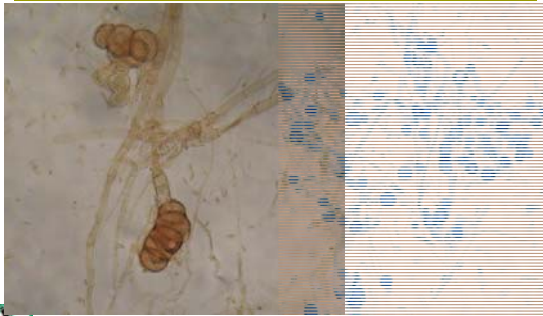
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### Chlamydospores



*F. clamidosporum*

*F. oxysporum f. sp. cubense*

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### Morphology of *Fusarium oxysporum*

Mycelia floccose sparse or abundant varying from white to purple color.

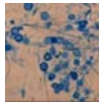
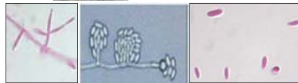
From pale violet to dark magenta pigment in agar (some isolates do not produce any pigment).

Can produce pionnotal colonies

Microconidia without septa produced in false heads in short monophialides

Macroconidia developed in ramified conidiophore on sporodochia

Chlamydospores terminal or intercalated; single or in chains



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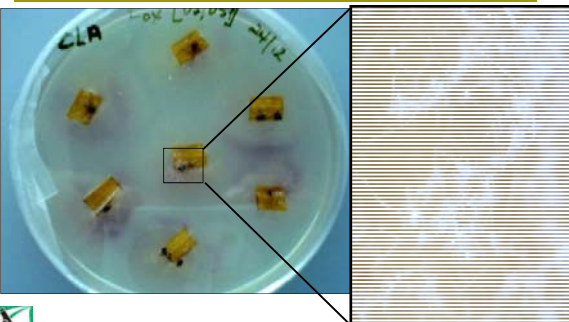
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### Characteristic of *Fusarium* growth in CLA



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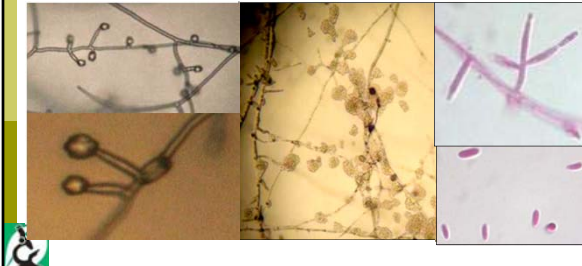
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### Morphology of *F. oxysporum* isolates growth

En CLA and SNA filter paper, produce abundantly microconidia in false heads in short monophyalids developed in hyphae.




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### Chlamydoconidia

- ✓ Can be observed in hyphae on agar surface or inside media.
- ✓ Develop on hypha or conidia in old culture of until a month.
- ✓ True chlamydoconidia, pseudochlamydoconidia and swollen cells, can be missed.
  - Chlamydoconidia have a gross verrucose like wall and a light color usually from yellow to brown. Can be presents solitary, in chains aggregates, above or below hypha.
  - Pseudochlamydoconidia (of thin walls, alone or in short chains are present only in *F. andyazi*)
  - Swollen cells are numerous in *Liseola* section




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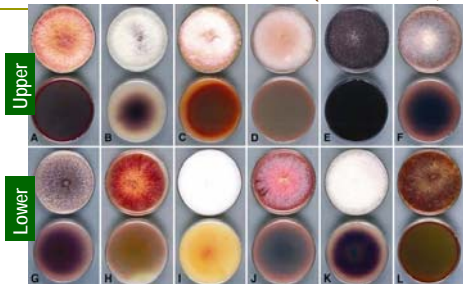
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### Colonies morphology in potato dextrose agar. A considerable variation among *Fusarium* species

(Summerett et al., 2003)



A) *F. poae*; B) *F. oxysporum*; C) *F. acuminatum*; D) *F. nelsonii*; E) *F. subglutinans*; F) *F. nygamai*; G) *F. pseudonygamai*; H) *F. lateritium*; I) *F. thapsinum*; J) *F. decemcellulare*; K) *F. verticillioides*; L) *F. culmorum*.




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### Biological concept

- ✓ Fertility of sexual crosses (teliomorph development)
- ✓ Host range: formae speciales
- ✓ Vegetative compatibility groups.




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### Biologic: fertility of sexual crosses

- ✓ Most fertility analysis are realized for of *Gibberella fujikuroi* and *Haemanectria haematococca* species complex (*Fusarium solani*) differentiation (Leslie and Summerell 2006).
- ✓ This approach can not be used to species that do not develop teliomorpha as *F. oxysporum*.
- ✓ There are two idiomorphic MAT (*MAT-1* and *MAT-2*) that have to be present for mating occurrence (Kroonstadt and Staben, 1987).
  - In *F. oxysporum* has been reported alleles *MAT-1* and *MAT-2* highly similar to those of fertile *Fusarium* species
  - The not fertility can be consequence of mutation of genes that codify indispensable receptors not codified by *MAT* locus.




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### Biologic: fertility of sexual crosses

(Protocol described by por Klittich y Leslie, 1988):

- Female parent is inoculated in carrot agar and allowed colonize plate by a week.
- Is inoculated with a spore suspension from the unknown strain (male). Colony is gently shaken with a glass bar
- Are incubated at < 25°C under light for 4-6 weeks.
- Avoid seal the plate.
- If molecular protocols to determine sexual *MAT-1* and *MAT-2* alleles are available, less crosses are required
- Positive crosses are definitive. Negative ones are not necessarily conclusive.




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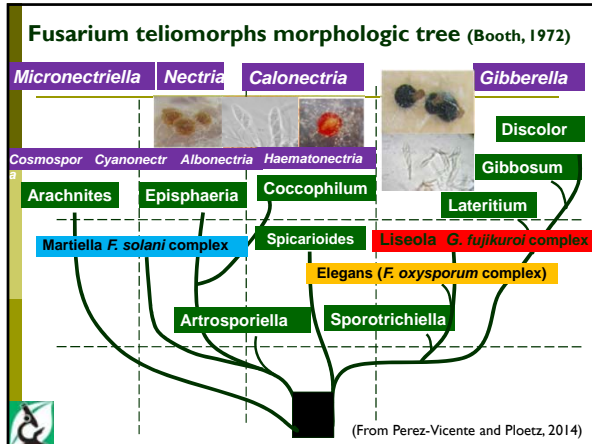
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### Classification in formae speciales by Snyder & Hansen in 1940

THE SPECIES CONCEPT IN FUSARIUM  
W. C. Snyder and H. N. Hansen, 1940

1940

W.N. Hansen

W.C. Snyder

✓ Compress the 16 sections in 9 species  
 ✓ > de 150 formae speciales recognized of *F. oxysporum*  
 ✓ All *Elegans* section species grouped in a single *oxysporum* specie

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### Formae speciales and pathogenic races

- ✓ Most formae speciales attack a single host specie.
- ✓ Some of them attack more than one specie: i.e.; *F. oxysporum* f. sp. *cucumerinum* attack water melon and cucumber (Cafri et al., 2005).
- ✓ Genetic bases regarding host specificity are in general unknown (Baayen et al., 2000) even when there is recent findings regarding genes involved in pathogenicity in chromosomes LS (Ma et al., 2010)
- ✓ It can not be assumed that all individuals in a given single forma special have evolved from a common ancestor. All evidences indicate that monophyletic formae speciales are exceptions (Leslie y Summerell, 2006).

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**Fusarium oxysporum diseases in different plant species**

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Palm oil wilt by *F. oxysporum* f. sp. *elaeidis*

*F. oxysporum* f. sp. *lycopersici*



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**Fusarium oxysporum diseases in different plant species**

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Panama disease by *F. oxysporum* f. sp. *cubensis*

*F. oxysporum* f. sp. *ciceris*



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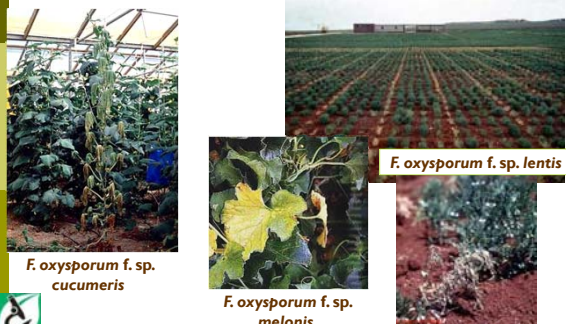
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**Fusarium oxysporum diseases in different plant species**


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*F. oxysporum* f. sp. *cucumeris*

*F. oxysporum* f. sp. *melonis*

*F. oxysporum* f. sp. *lentis*



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**Fusarium oxysporum diseases in different plant species**



*F. oxysporum f. sp. asparagi*



*F. oxysporum f. sp. cepae*



*Fusarium oxysporum f. sp. niveum* en melón

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**Fusarium spp. cultures growth rate (in mm after three days) in PDA at 25 y 30°C in complete darkness (Burgess et al., 1994)**

Species	25°C	30°C	Species	25°C	30°C
<i>F. acuminatum</i>	25-35	5-28	<i>F. decemcellulare</i>	15-25	11-22
<i>F. andiyazi</i>	27-33	27-40	<i>F. dimerum</i>	4-10	5-12
<i>F. anthophilum</i>	25-40	20-45	<i>F. equiseti</i>	34-46	28-44
<i>F. armeniacum</i>	44-58	37-54	<i>F. graminearum</i>	47-61	5-20
<i>F. avenaceum</i>	28-40	5-25	<i>F. heterosporum</i>	28-41	8-30
<i>F. aywerte</i>	40-45	32-42	<i>F. hostae</i>	24-29	20-24
<i>F. babinda</i>	25-37	15-22	<i>F. konzum</i>	21-34	21-38
<i>F. beomiforme</i>	30-39	36-46	<i>F. lateritium</i>	8-20	5-15
<i>F. camptoceras</i>	23-29	20-29	<i>F. longipes</i>	37-54	44-61
<i>F. chlamydosporum</i>	34-46	37-55	<i>F. merismoides</i>	4-10	5-12
<i>F. compactum</i>	41-54	42-58	<i>F. musarum</i>	50-59	44-49
<i>F. crookwellense</i>	54-66	15-25	<i>F. napiforme</i>	20-35	20-32
<i>F. culmorum</i>	55-68	15-25	<i>F. nelsonii</i>	24-39	26-41

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**Fusarium spp. cultures growth rate (in mm after three days) in PDA at 25 y 30°C in complete darkness (Burgess et al., 1994)**

Species	25°C	30°C	Species	25°C	30°C
<i>F. decemcellulare</i>	15-25	11-22	<i>F. nurragi</i>	32-40	6-22
<i>F. dimerum</i>	4-10	5-12	<i>F. nygamai</i>	25-35	32-42
<i>F. equiseti</i>	34-46	28-44	<i>F. oxysporum</i>	25-40	25-40
<i>F. graminearum</i>	47-61	5-20	<i>F. poae</i>	42-54	24-39
<i>F. heterosporum</i>	28-41	8-30	<i>F. polyphialidicum</i>	23-40	20-38
<i>F. hostae</i>	24-29	20-24	<i>F. proliferatum</i>	25-35	25-32
<i>F. konzum</i>	21-34	21-38	<i>F. pseudograminearum</i>	39-51	10-25
<i>F. lateritium</i>	8-20	5-15	<i>F. pseudonygamai</i>	24-30	24-29
<i>F. longipes</i>	37-54	44-61	<i>F. redolens</i>	32-37	33-39
<i>F. merismoides</i>	4-10	5-12	<i>F. sambucinum</i>	24-35	11-21
<i>F. musarum</i>	50-59	44-49	<i>F. semitectum</i>	35-45	16-33
<i>F. napiforme</i>	20-35	20-32	<i>F. scirpi</i>	36-48	36-49
<i>F. nelsonii</i>	24-39	26-41	<i>F. solani</i>	21-29	26-36

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**Fusarium spp. cultures growth rate (in mm after three days) in PDA at 25 y 30°C in complete darkness (Burgess et al., 1994)**

Species	25°C	30°C
<i>F. sporotrichioides</i>	51-61	32-42
<i>F. subglutinans</i>	23-37	11-34
<i>F. thapsinum</i>	19-28	14-27
<i>F. tricinctum</i>	29-39	2-15
<i>F. verticillioides</i>	21-30	22-35




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**Biological concept: *F. oxysporum* case; vegetative compatibility groups (VCGs).**

- ✓ *F. oxysporum* is causal agent of an important number of disease with wilt syndrome. The fungus has not known teleomorph.
- ✓ Are morphologically identical, and have been differentiated by pathogenicity to specific hosts in formae speciales
- ✓ In some species, races are not genetically defined as in *Fusarium oxysporum* f. sp. *cubense*. Vegetative compatibility has allowed to determine population structures.
- ✓ In other cases has not been so useful.
- ✓ Further more specific informations on the technique will be provide.




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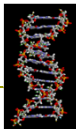
**The phylogenetic concept: sequences of genome's conserved sectors**

In *Fusarium* phylogenetic analysis, genomic sequences of one or several genes in different analysis types have been used :

- ✓  $\beta$ -tubulin (*tub-2*),
- ✓ The translation elongation factor 1- $\alpha$  (*tef-1*),
- ✓ Histone H3
- ✓ Portions of coding region of nuclear or mitochondrial DNA
  - Intergenic spacers (IGS)
  - ITS spaces

Different techniques has been used:

- ✓ RFLP
- ✓ AFLP
- ✓ PCR
- ✓ RAPDs




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### The phylogenetic concept: sequences of genome's conserved sectors 1- $\infty$ (TEF)

The translation elongation factor 1- $\infty$ (TEF) gen codify an essential protein of translation machinery with a high phylogenetic utility due to (Geiser et al., 2006):

- (i) Is highly informative at *Fusarium* species level;
- (ii) Have not been detected orthologous copies of *Fusarium* genera.
- (iii) Appears as a single copy and its sequence has a high polymorphism in genome of many closely related *Fusarium*
- (iv) Has been designed universal primers that work thorough the width of all phylogenic genera.
- (v) These primers (ef1 and ef2) amplify a ~700 bp of TEF region and kept three introns that complete the half of the amplicon length in all *Fusarium* species.




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### The phylogenetic concept: sequences of genome's conserved sectors: IGS

Larger intergenic spacers (IGS) has been very widely used in phylogenetic studies in *Fusarium* due to:

- (i) Appears as the more rapid evolved spacer region in genome
- (ii) Is a very conserved region of nuclear rDNA with sufficient polymorphism to may show considerable differences between close related species and subspecies.
- (iii) Appears in multicopies in genome




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### Fusarium ID database v. 1.0 (Geiser et al., 2006)

Complex of species	Represented species
<i>Gibberella fujikuroi</i> species complex: Excelent representation	<i>Fusarium acutatum</i> , <i>F. andiyazi</i> , <i>F. anthropilum</i> , <i>F. bactridioides</i> , <i>F. begoniae</i> , <i>F. brevicatenulatum</i> , <i>F. bulbicola</i> , <i>F. circinatum</i> , <i>F. circinatum</i> , <i>F. sp. cf. concentricum</i> , <i>F. denticulatum</i> , <i>F. dlamini</i> , <i>F. fractiflexum</i> , <i>F. fujikuroi</i> , <i>F. globosum</i> , <i>F. guttiforme</i> , <i>F. konzum</i> , <i>F. lactis</i> , <i>F. mangiferae</i> , <i>F. napiforme</i> , <i>F. nygamai</i> , <i>F. phyllophilum</i> , <i>F. proliferatum</i> , <i>F. pseudoanthophilum</i> , <i>F. pseudocircinatum</i> , <i>F. pseudonygamai</i> , <i>F. ramigenum</i> , <i>F. sacchari</i> , <i>F. sterilityphosum</i> , <i>F. subglutinans</i> , <i>F. succisae</i> , <i>F. thapsinum</i> , <i>F. udum</i> , <i>F. verticillioides</i> y 20 spp.no descritas
<i>Fusarium</i> type A trichotecene producer species and relatives: Weak representation	<i>Fusarium cerealis</i> (= <i>F. crookwellense</i> ), <i>F. culmorum</i> , <i>F. graminearum</i> y 8 spp. relacionadas; <i>F. lunulosporum</i> , <i>F. pseudograminearum</i>
<i>F. oxysporum</i> and relatives: Good representation.	<i>Fusarium commune</i> , <i>F. hostae</i> , <i>F. miscanthi</i> , 45 linajes de cf. ' <i>F. oxysporum</i> '; <i>F. redolens</i> .
<i>F. solani</i> species complex : Good representation.	<i>Fusarium phaseoli</i> , 25 linajes de cf. ' <i>F. solani</i> '; <i>F. tucumaniae</i> , <i>F. virguliforme</i> .
Other linages: Poor representation	




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**Results influenced by the concept approach**

Eg.: *Fusarium moniliforme*: By decades considered causal agent stalk and ear rot of corn; stem and grain rot of sorghum; bakanae in rice; pokkah boeng in sugarcane.

According the concept:

- ✓ **Morphologic:** *Fusarium moniliforme sensu lato* (wide sense)
- ✓ **Biologic:**
  - *F. verticillioides*: stalk and ear corn rot (produce fumonisin)
  - *F. taylorii*: stalk and grain sorghum rot (produce few fumonisin and high amounts of moniliformin).
- ✓ **Phylogenetic:**
  - *F. andiyazi* (beside *F. verticillioides* and *F. taylorii*), few or any aproduction of fumonisin and moniliformin and not toxic to dugs.




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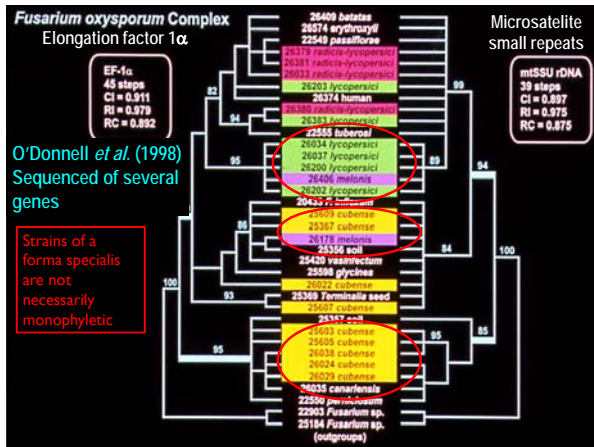
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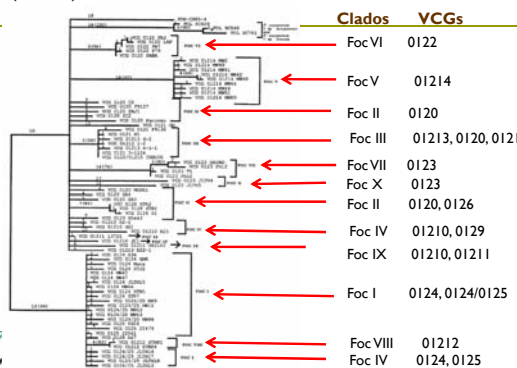
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**Phylogenetic relationship between VCGs in *Foc* (RFLP) (Koenig et al., 1997)**




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**PCR-RFLP analysis of rRNA IGS region, and digestion of IGS amplicon with enzymes (AvaI, BceAI, BbvI, Csp6I, BsrDI) to differentiate *Foc* lineages (Fouere et al., 2009)**

**8 lineages were differentiated:**

- I (VCG 01219)
- II (VCGs 1210, 0126)
- III (VCG 01211)
- IV (VCGs 0120, 0122, 01215)
- V (VCGs 0121, 01213)
- VI (VCGs 0124, 0125, 0128, 01220)
- VII (VCGs 0123, 01217, 01218)
- VIII (VCGs 01214)

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***Fusarium* taxonomy evolution since the introduction of phylogenetic concept**

Specie complex → Complex of species

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***Fusarium oxysporum* f. sp. *cubense* population grouping according different characterization methods**

Method	I	II
✓ Actual race classification (Stover, 1962 b; Su et al. 1977)	Race 4	Races 1 and 2.
✓ Formación de lacinia en K2 (Sun et al., 1978).	Present (not always)	Not present
✓ Vegetative compatibility (Ploetz, 1990 a y b; Brake et al., 1990; Pegg et al., 1993; Moore et al., 1993).	0120, 0121, 0122, 0126, 0129, 01211, 01213/16, 01215, 01219	0123, 0124, 0125 0128, 01210*, 01212, 01214, 01217, 01218, 01220
✓ Volatile production (Moore, 1994).	Odoratum	inodoratum (not always)
✓ RAPD-PCR (Sorensen, et al. 1993; Sorensen et al., 1994)	RAPD-PCR group 1	RAPD-PCR group 2
✓ Chromosomic number genome size. (Boehm et al., 1994)	Ek I high	Ek II low

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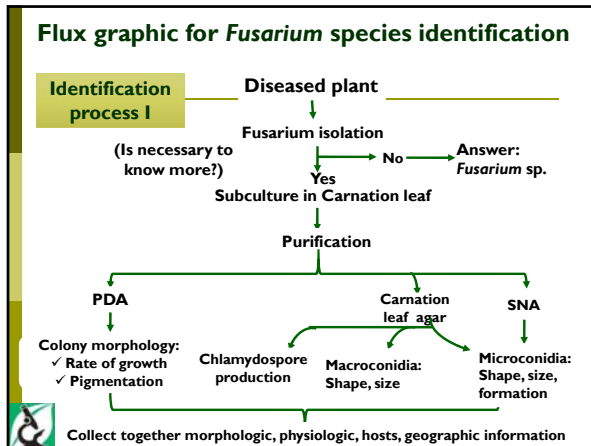
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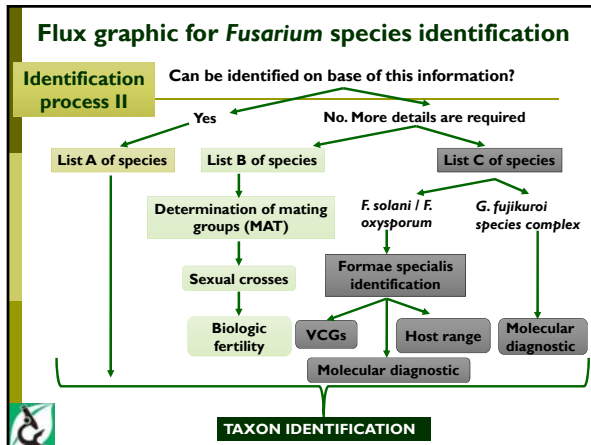
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### Species more commonly isolated that can be identified by different data types

Only morphology SPECIES LIST A	Fertility in sexual crosses SPECIES LIST B	DNA data SPECIES LIST C
<i>F. acuminatum</i> , <i>F. avenaceum</i> , <i>F. chlamidosporum</i> , <i>F. compactum</i> , <i>F. crookwellense</i> , <i>F. culmorum</i> , <i>F. decemcellulare</i> , <i>F. dimerum</i> , <i>F. equiseti</i> , <i>F. graminearum</i> , <i>F. longipes</i> , <i>F. merismoides</i> , <i>F. oxysporum</i> , <i>F. poae</i> , <i>F. pseudograminearum</i> , <i>F. scirpi</i> , <i>F. semitectum</i> , <i>F. solani</i> , <i>F. sporotrichoides</i> , <i>F. torulosum</i> , <i>F. tricinctum</i>	<i>F. circinatum</i> , <i>F. fujikuroi</i> , <i>F. proliferatum</i> , <i>F. sacchari</i> , <i>F. sambucinum</i> , <i>F. subglutinans</i> , <i>F. thapsinum</i> , <i>F. verticilloides</i>	Remanent species of <i>Gibberella fujikuroi</i> complex Many formae specialis of <i>F. oxysporum</i> and <i>F. solani</i> .

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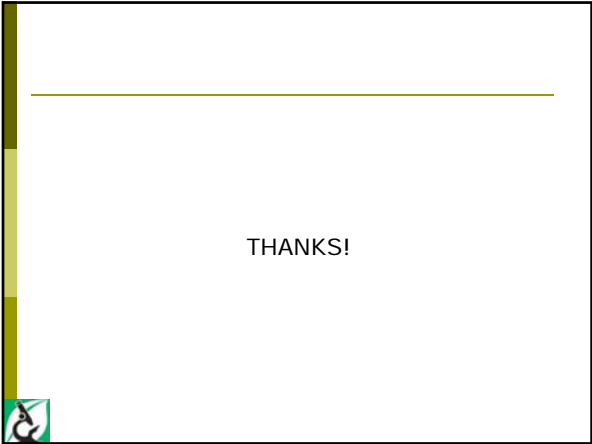
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