



International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 5.62

(GIF) Impact Factor: 0.549

IJFAS 2021; 9(1): 272-280

© 2021 IJFAS

www.fisheriesjournal.com

Received: 22-10-2020

Accepted: 02-12-2020

Anju P

School Of Industrial Fisheries,
Cochin University of Science and
Technology, Lakeside Campus,
Foreshore Road, Cochin, Kerala,
India

Mini Sekharan N

School of Industrial Fisheries,
Cochin University of Science and
Technology, Lakeside Campus,
Foreshore Road, Cochin, Kerala,
India

Harikrishnan M

School Of Industrial Fisheries,
Cochin University of Science and
Technology, Lakeside Campus,
Foreshore Road, Cochin, Kerala,
India

Corresponding Author:

Anju P

School Of Industrial Fisheries,
Cochin University of Science and
Technology, Lakeside Campus,
Foreshore Road, Cochin, Kerala,
India

The occurrence of microplastics in gut contents of endemic barb *Sahyadria chalakudiensis* (Menon, Rema Devi & Thobias, 1999) inhabiting river systems of Western Ghats, South India

Anju P, Mini Sekharan N and Harikrishnan M

DOI: <https://doi.org/10.22271/fish.2021.v9.i1d.2411>

Abstract

Sahyadria chalakudiensis, the endemic barb inhabiting rivers of Western Ghats, S. India has been supporting considerable aquarium trade and is known to dwell in upstream areas of these rivers. A concerted study on its feeding biology from 730 fishes has revealed consistent occurrence of microplastics in their guts, pointing to serious plastic pollution affecting riverine ecosystem. This fish has omnivorous feeding habit as evident from percentage index of relative indices of various prey items as animal matter (62%), filamentous algae (26%), sand particles (4%) and other matter (8%). Among other matter, microplastic fibres were consistently encountered in 86 guts (11.8% of total guts examined) collected in all months except September. The monthly occurrence of guts containing microplastic fibres showed significant correlation to guts containing filamentous algae ($r=0.95$, $p<0.05$). Further, the mean \pm s.d of frequency occurrence of guts with microplastic fibres differed significantly between seasons being lowest in pre-monsoon (3 ± 1 guts) and highest in post monsoon (11 ± 5 guts). However, the occurrence of microplastic fibres was not influenced by feeding intensity which indicated that the fish consumed these microplastic fibres inadvertently. The number of microplastic fibres encountered in each gut varied from 1 to 4, however, guts with one fibre only outnumbered others in all seasons. The present results indicated possibility of micro-litter ingestion during feeding from sediments and from shallow stagnant areas in rivers. The paper discusses challenges of aquatic pollution by plastic litter caused by anthropogenic interventions in protected forest areas and suggests mitigation strategies.

Keywords: Micro plastic fibre, river pollution, ornamental fish, habitat management, Western Ghats

1. Introduction

One of the most recent anthropogenic impacts adversely affecting environments is plastic litter which has been identified globally as severe threat to different aquatic habitats. The persisting and buoyant nature of discarded and neglected plastic litter accumulated in nature leads to severe environmental hazards that invite increasing research interest in many parts of the world. Nevertheless, plastic production remains increasing to 353million tonnes^[1] and plastic litter is more of a concern in terrestrial ecosystems which often gets washed off to rivers, estuaries before ending up in marine habitats^[2]. A number of potential hazards of plastic debris in aquatic habitats such as transport of persistent organic pollutants, toxic algae, invasive species etc have been described^[3, 4, 5, 6, 7]. This debris undergoes physical, chemical and biological degradation in such environments^[8, 9]. Apart from being accumulated as mega and macro litter in water bodies, these plastic debris provide large sources of micro- plastics of less than 5mm size^[10].

Anthropogenic sources such as industrial effluents, domestic and urban sewage, consumer products etc., have also been attributed to contain micro plastics which leach out to natural habitats^[11, 12, 13]. Occurrence of micro plastics have been reported from a wide array of aquatic environments; from freshwater^[14,13] to marine^[15], deep sea^[16], Antarctic sea^[17]. Ingestion of these micro-plastics has been reported in many aquatic biota as zooplankton^[18], crustacean^[19], fishes^[20], sea birds^[21], mussel^[22] and clams^[23]. Ingestion of microplastics also result in health hazards in fishes as evidenced from liver toxicity and pathology through bioaccumulation of chemical pollutants that are adsorbed or associated with plastic degradations^[24].

It was recently demonstrated that microplastic ingestion in fishes cause physical abrasions in intestinal wall leading to inflammation due to leukocyte infiltration, hyperaemia and regressive changes in intestinal tissue [25].

Microplastic ingestion by marine fishes have been reported extensively in contrast to freshwater, in particular, riverine fishes (Table.1).

Table 1: Previous works carried out by authors on plastic ingestion in fish species

Authors	Species
[26]	<i>Gadus morhua</i>
[27]	<i>Cathorops spixii</i> <i>Cathorops agassizii</i> <i>Sciades herzbergii</i>
[28]	<i>Gerreidae</i>
[29]	<i>Anguilla anguilla</i>
[30]	<i>Johnius borneensis</i>
[31]	<i>Merlangius merlangus</i> <i>Limanda limanda</i>
[32]	<i>Gobio gobio</i>
[33]	<i>Thunnus albacares</i> , <i>Lethrinus amboinensis</i> , <i>Katsuwonus pelamis</i>
[34]	<i>Cyprinella lutrensis</i> <i>Cyprinella venusta</i> <i>Notropis anabilis</i> <i>Notropis volucellus</i> <i>Pymphales vigilax</i>
[35]	<i>Ammodytes personatus</i> <i>Clupea pallasii</i>
[36]	<i>Galeus melastomus</i>
[37]	<i>Rhizoprionodon terraenovae</i>
[38]	<i>Mullus surmuletus</i>
[39]	<i>Lates niloticus</i> <i>Oreochromis niloticus</i>
[40]	<i>Gonostoma denudatum</i> <i>Serrivomer beanie</i> <i>Lampanyctus macdonaldi</i>
[41]	<i>Lepomis macrochirus</i> <i>Lepomis megalotis</i>
[42]	<i>Myripristis spp.</i> <i>Siganus spp.</i> , <i>Epinephelus merra</i> <i>Cheilopogon simus</i>
[43]	<i>Thamnaconus septentrionalis</i> <i>Carrasius auratus</i> <i>Cyprinus carpio</i> <i>Hemiculter bleekeri</i> <i>Hypophthalmichthys molitrix</i> <i>Megalobrama amblycephala</i> <i>Harpodon nehereus</i> <i>Pampus cinereus</i>
[44]	<i>Hoplosternum littorale</i>
[45]	<i>G. melastomus</i> <i>E. spinax</i>
[46]	<i>Pagellus erythrinus</i> <i>P. bogaraveo</i>

Studies on microplastic pollution and ingestion by fishes in Indian waters are scanty [47,48,49,50]. A concerted study on the feeding biology of an endemic barb *Sahyadria chalakkudiensis* [51] inhabiting rivers originating from the Western Ghats, S.India has revealed consistent ingestion of microplastics by the fish. This paper encompasses extent of microplastic ingestion by endemic fish species inhabiting freshwater rivers of Western Ghats and discusses need for developing habitat management strategies for sustainability

and conservation of this species.

2. Materials and Methods

2.1 Sampling Period and Study area

Two-year sampling was carried out from April, 2015 to March, 2017 in river Pooyamkutty (10°9'39.79"N 76°47'11.94"E) and river Chalakudy (10.2922° N, 76.5149° E) (Fig.1)

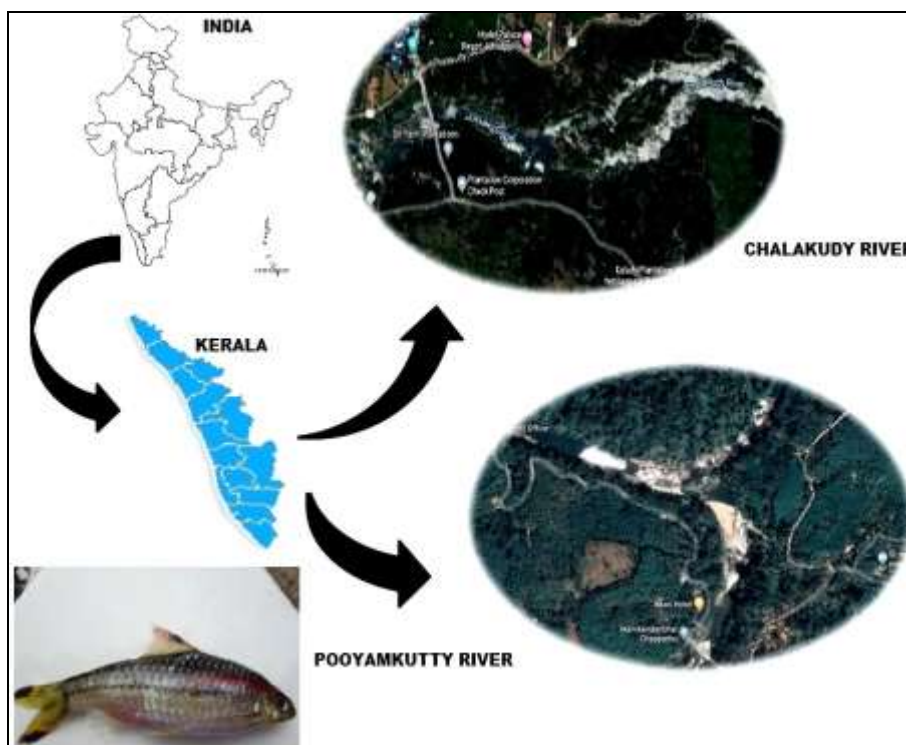


Fig 1: Sampling locations-Chalakudy and Pooyamkutty river of Western Ghats, Kerala, India.

2.2 Study organism

Sahyadria chalakkudiensis is an endemic barb having restricted distribution in Western Ghat river systems. It is a popular food fish among the native communities and is also marketed as a highly valued ornamental fish. *S. chalakkudiensis* was first described by Menon *et al.* (1999) [51] from the upper reaches of river Chalakudy. Apart from Chalakudy river, its presence has also been reported in restricted parts of Achankovil river [52] and Pamba River [53].

2.3 Sample collection

478 fishes were collected from native fish vendors in local markets of river Pooyamkutty which included 167 males and 97 females. 252 fishes were collected from Chalakudy river (Male:147and females:105). The total length (TL) was recorded from tip of snout to end of caudal fin to the nearest millimetre. Weight was recorded to the nearest gram on wet weight basis [54]. A small pierce at the end of belly was given and the fish was preserved in 10% formalin for further analysis.

2.4 Gut content analysis

The total length (mm) and weight (g) of the fish were determined for each individual. The fish was then dried with a tissue paper, put over a box filled with ice. The belly portion was then cut open and the whole alimentary canal was separated from fish. The length of gut was taken and the stomach portion was separated. Length was taken and the stomach was then separated. The gut contents (stomach and intestine) were examined under a stereoscopic microscope (Lawrence and Mayo make) for the identification of food items and micro plastics. All food items were identified to the lowest possible taxonomic level following the protocol of Hynes [55].

2.5 Micro plastic identification

The suspected micro plastic particles were observed to meet all of the following selection criteria, Nor and Obbard (2014) [56]: 1) no visible cellular structures, 2) un segmented nature 3) fibres of consistent width (not tapered) and should have at least two of the additional criteria: 1) brightly coloured coating 2) homogenous texture 3) abnormal shape 4) fibre that remained unbroken if tugged 4) reflective 5) flexible without being brittle.

2.6 Sample digestion

The gut contents along with the suspected particles were taken and transferred to a conical flask in which 10% KOH [57] strong oxidising agent was placed and incubated for 5 days so that all the biological compounds disintegrate and the remaining plastic fibres are separated.

2.7 Confirmation test

Hot Needle Test was performed for further verification. The fibres were held with a forceps and a hot needle was brought near to the plastic fibre and the plastic fibre started to swirl around and it confirmed the material as plastic [58]. Images and measurements (a micrometre in Motic Plus 2.0) of plastic items recovered were taken with a Motic Image Plus 2.0 (Fig.2).



Fig 2: Micro plastics obtained from gut and measured using a micrometre in Motic Plus 2.0

2.8 Statistical analysis

A descriptive analysis was carried out to describe types of collected micro plastics according to shape categories, size classes and colour opacity. Inferential Analyses were performed and graphical representations were generated with the statistical software SPSS and PAST. Images and measurements of plastic items recovered were taken with a Motic Image Plus 2.0

3. Results

A total of 730 guts were dissected for examining the contents. The percentage IRI values worked out in respect of various prey items are depicted in Fig.3.

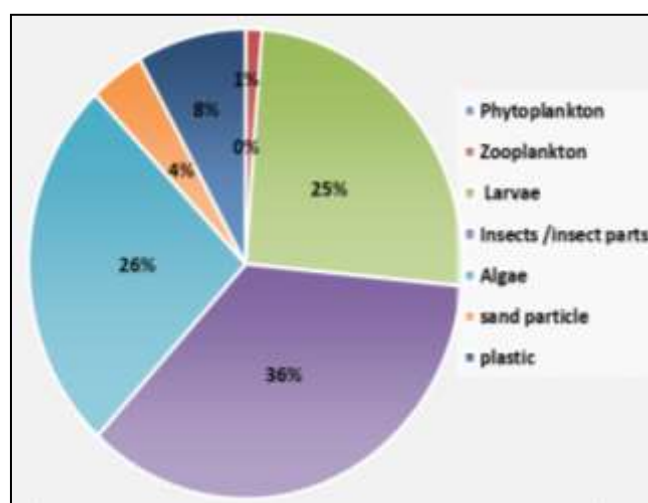


Fig 3: Percentage IRI recorded in major food items of *S. chalakkudiensis*

It could be noted that the fish feeds mainly on animal matter comprised mainly of insect parts (36%) and larvae (25%). Higher percentage IRI could also be recorded in filamentous algae (26%) and sand particles (4%). Other minor matter included very small proportions of phytoplankton, zooplankton and undigested matter.

Among undigested matter, a number of fibres like matter could be observed, which on further examination were identified as microplastic fibres. Such fibres were observed in 86 guts which formed 11.8% of total guts examined. Elongated pale red coloured fibres of approximately 2.6mm could be identified in the guts collected in all months except September. The lowest % occurrence of microplastic fibres among gut contents could be noticed in March (7%) while it was highest in October (25%). The mean \pm s.d gut frequency having microplastic fibres in pre-monsoon (February – May), monsoon (June to September) and post monsoon (October to January) are depicted in Fig.4.

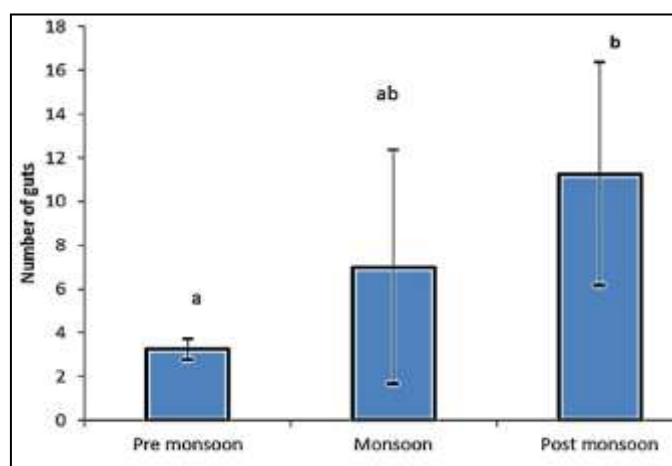


Fig 4: Season wise percentage of guts containing micro plastic fibres.

It could be noticed that mean number of guts with occurrence of microplastic fibres was significantly different between seasons ($F= 5.49, p<0.05$) which increased from pre-monsoon (3 ± 1 guts) to monsoon (7 ± 5 guts) and post monsoon (11 ± 5 guts). It could also be noted that the occurrence of microplastic fibres was not influenced by feeding intensity of the fish as these fibres were present in guts with considerable amount of food and guts having traces of food. The proportion of guts with food and plastic and guts with trace of food and plastic were not statistically significant in pre-monsoon ($\chi^2=3.6, p>0.05$), monsoon ($\chi^2=3.7, p>0.05$) and in post monsoon ($\chi^2=1.0, p>0.05$).

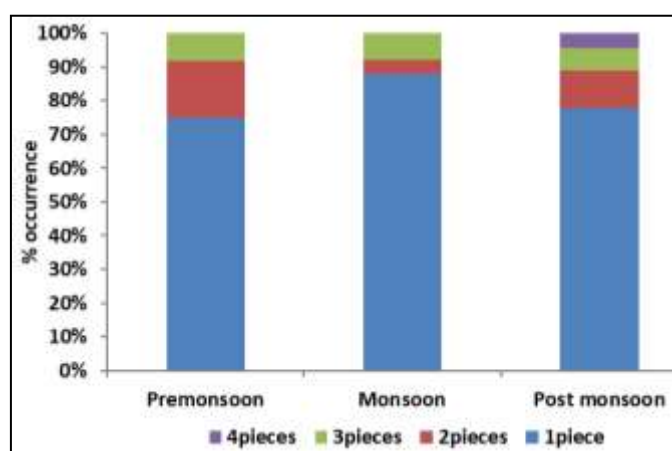


Fig 5: Percentage frequency of occurrence of microplastic fibres obtained from guts in different seasons.

The number of micro plastic fibres encountered in each gut varied from 1 to 4 and the frequency of guts with one fibre

predominated in all seasons with 75% of guts in pre-monsoon, 88% of guts in monsoon and in 77% of guts in post monsoon (Fig.5). The number of guts with 2 fibres each were high in pre-monsoon (16%) followed by post monsoon (11%) and lowest in monsoon (4%). Number of guts containing 3 micro plastic fibres each did not vary considerably between seasons with 8%, 7% and 8% in pre-monsoon, monsoon and post monsoon seasons. However, guts containing four microplastic fibres were encountered only in post monsoon season.

4. Discussion

S. chalakudiensis is a fish used for food as well as for aquarium purpose. Other than exploiting live fishes for aquarium purpose, many other important reasons add up to the sustainability issues faced by this fish. The present study has revealed another important issue of environmental pollution, affecting endemic indigenous fishes inhabiting rivers flowing through protected forest areas after originating in Western Ghats of India. It has been reported that about 70% to 80% of marine litter, most of it plastics, originate from inland sources and are emitted by rivers to the oceans [59]. Most of the available information of reports of plastic debris ingestion is for marine species, however studies on freshwater fishes, especially those used as food resource by humans are limited [60].

A primary field report of freshwater fish ingesting microplastics described 12% microplastic in *Gobio gobio* [32, 61]. However, it may be pointed out that such data on micro plastic intake by freshwater fishes is very limited [14]. During the present study, it could be noted that fishes collected from rivers are mostly consumed by natives and tribes and are also sold in nearby markets, which indicated the possibility of microplastic ingested fishes making their way to the next level of consumers. Fishes with microplastics to the tune of 20% have been reported in market-purchased freshwater fish [39]. With each trophic level, bioaccumulation of ingested plastics soon leads to bio magnification, ultimately risking human health [62, 63]. Plastic pollution is carcinogenic to human, it can also cause birth defects, damage immune system, endocrine and reproductive system [64]. Recent studies show how fish health and overall wellbeing of fish are affected by consuming microplastics [65, 66, 67, 68].

If plastic particles become nano-sized, they can cross the blood-brain barrier and can cause brain damage resulting in behavioural changes in organisms [69, 70]. Predators, preferring a greater number of fish with full guts, will generally have higher exposure to microplastics due to simply ingesting more material [71]. A recent study found that microplastic burden varied significantly between species depending up on feeding habits and trophic transfer and top predators contained the highest load of microplastics [72].

In the present study, it could be noted that this fish feeds mainly on filamentous algae and larva and has preference to slender elongated objects. It may also be inferred that the fish seeks food in shallow stagnant pools and river banks where filamentous algae and insect larvae can be available in plenty. As these areas are shallow and without considerable water flow, permit sediments to accumulate, increasing settlement of microplastic fibres. It is reasonable to infer from present results that *S. chalakudiensis* adopts bottom feeding as indicated by presence of sand and higher proportion of insect larvae, algae and occurrence of microplastic among gut contents could be indicative of extent of plastic pollution in the rivers. Plastic debris would have mixed with sediment and

when fish forage on the bottom, the microplastic fibres could have been unintentionally consumed [73]. Wang et al. [74] found that most microplastic obtained from fish species were coloured and fibrous in nature [75, 76]. More than 80% of microplastics had a size of 2mm. In the present study also, all microplastic encountered were fibrous in nature and were on average 2.6mm in size. Generally, 1- 2 ingested microplastic pieces were encountered in fish guts [77]. In the present study, up to 4 pieces of microplastic fibres could be collected from individual guts. Predators preferably ingest microplastics with colours resembling their prey [78, 79, 80]. The microplastic fibres obtained in present study were red in colour. Coloured plastics have been detected from organisms as well as from habitat [81, 82].

4.1 Potential pathways of microplastics in tropical river system

The microplastic pollution in the two rivers of Western Ghat were from untreated sewage, fishing, tourism and industrial waste (Fig.6).

It may be pointed out that urbanisation and population growth are the major reasons for microplastic pollution [83, 84, 85]. House hold waste is a potential source of microplastic fibres as untreated house hold waste consisted of partially digested

bags, paints, withered plastic utensils, cosmetics, cloths etc. Untreated sewage may also be an important carrier that conveys fibres to the aquatic system via effluent discharge or surface runoff [86]. Clothing and packaging from surrounding residential areas might be potential sources for these coloured items in the studied areas as reported by Wang et al [74]. In both rivers of present study, natives wash their cloths in river on a regular basis. Washing and dumping of garments discharge microplastic fibres [87, 86, 88]. Lack of proper waste management plan for discharged sewage water from nearby houses contribute to the existing problem of fabric dumping [89].

The tribes and natives inhabiting river banks depend largely on fishing as livelihood means and do supply fish to local markets. Use of modern fishing netting materials is common among them which they frequently replace owing to tearing loss from using in fast flowing waters in rocky areas. Such damaged netting materials are abandoned in rivers which succumb to weathering and biodegradation. Smaller fibres from withered fishing nets are likely to get ingested by fishes. The improper waste management in plantations (rubber, cocoa, plantain and pineapple) result in run off of microplastics during monsoon, result in accumulation of fibres in river waters.

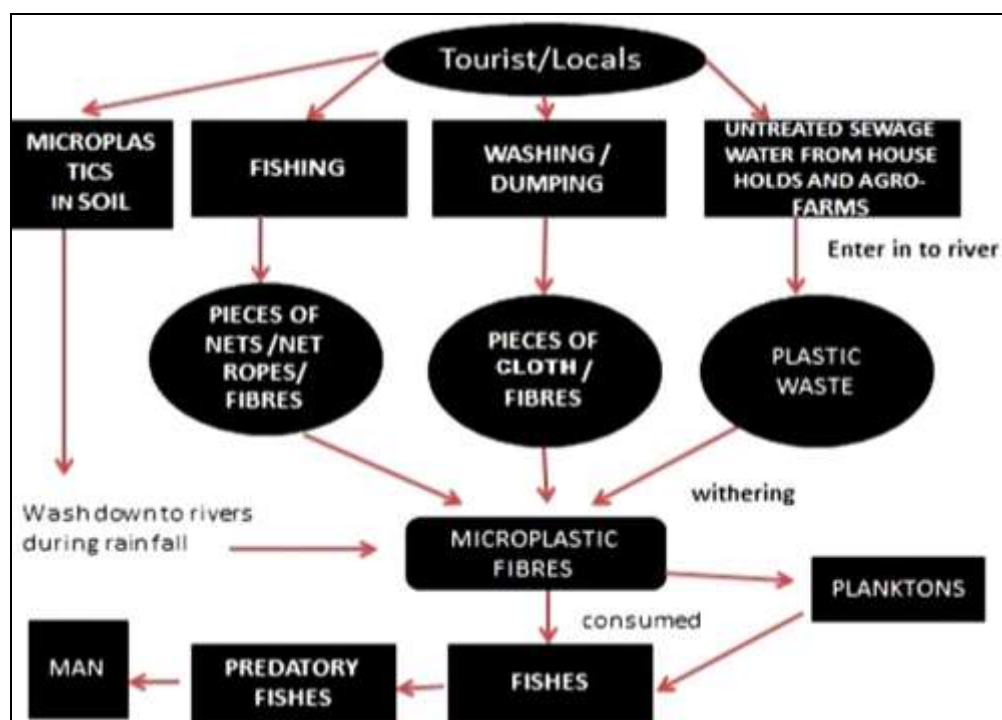


Fig 6: Potential pathways of microplastic debris in freshwater fishes of tropical river system

There are some important tourist spots in the rivers Chalakudy and Pooyamkutty. These areas are known for large quantity of plastic debris which due to human negligence gets dumped into these rivers (ie. Cloths, bags, bottles). Need for sustainable eco-tourism plans by giving more thrust on habitat management for healthy sustaining of life have well been recognized by authorities. Reduce, reuse, and recycle strategy might help to resolve the microplastics issue [48].

4.2 Strategies to reduce microplastic pollution

The study provides evidence to the fact that even when rivers flow through protected forest they face serious challenge from plastic pollution. Hence, it is important to propose waste management strategies for reducing microplastic pollution. Based on each of the channels of microplastic transmission, habitat management strategies for sustainable maintenance of healthy ecosystem were formulated (Table 2).

Table 2: Strategies to reduce microplastic pollution in Western Ghat rivers

Waste management strategies for reducing plastic fibre discard from household & tourism	Strategies for reducing micro plastic fibre load during fishing	Strategies for reducing micro plastic fibre load originating from textiles
<ol style="list-style-type: none"> Educate the communities and official in charge of the area through local government bodies & NGO about the sources of microplastics, its hazards and better management practices during visit and fishing. Support plastic litter free zones in upper streams of river and participate in regular river clean-up. Reduce the use of single-use plastics and avoid using cosmetics containing microbeads and paints (road, wall paints) containing microfibers. Encourage the use of natural paints. Government must provide effective wastewater treatment facilities in each locality for industrial/household sewage treatment. Advanced biofilter and reverse osmosis filters can be used in public sewage treatment plant. Guppy friend washing bags should be introduced to people and its effectiveness and acceptance in rural areas of Kerala need to be subjected to further studies. 	<ol style="list-style-type: none"> Use bio degradable netting/line materials, in fishing gears for fishing. Make the stakeholders (native men, fish collectors, tribe's, forest officials) aware about the micro plastics, its potential pathways, probable hazards and good management practices. During the months when the water is less the exposed river bottoms must survey and plastic materials should be cleared A fishery manager should be entrusted to look after the duties of habitat management in each river. 	<ol style="list-style-type: none"> Encourage the use of natural fibres like silk, cotton, jute and hemp. Encourage the installation of ultra-filters in washing machines. Create awareness among the people about hazards of washing /dumping cloths in open river

5. Conclusion

The results of present study reveal that even rivers flowing through protected forest are prone to serious challenges from plastic pollution. This paper throws light on the microplastic pollution in Western Ghat river system and consequent challenges to its biota, which necessitate their conservation through implementing better management practices in fish collection and sustainable eco-tourism plans, by giving more thrust on habitat management. Maintaining health of ecosystems is the best way to sustain healthy fish stock in river ecosystems.

6. Acknowledgements

This study was conducted under School of industrial fisheries, CUSAT. The first author would like to extend deep gratitude to the Director of school of industrial fisheries and guide for providing all facilities for the study. The authors would also like to thank fishermen and fish vendors of Pooyamkutty and Chalakudy River who whole heartedly supported by providing information, as well as samples for this study. This research did not receive any specific grant from funding agencies in the public, commercial, or non-for-profit sectors.

7. Funding Information

This study was carried out as a part of the authors thesis work. The expenses are met with the fellowship availed for the PhD programme, from Cochin university of science and technology, Kerala

8. References

- Plastics Europe, Plastics the Facts Available at: <http://www.plasticseurope.org/>(Accessed 1st March 2020), 2019. doi: https://www.plasticseurope.org/application/files/9715/7129/9584/FINAL_web_version_Plastics_the_facts2019_14102019.pdf
- Andrady L. In Plastics and the environment (ed. Andrady A. L., editor.). West Sussex, England: John Wiley and Sons 2003, 792.
- Mato Y, Isobe T, Takada H, Kanehiro H, Ohtake C, Kaminuma T, Plastic resin pellets as a transport medium for toxic chemicals in the marine environment. Environ. Sci. Technol 2001;35:318-324.
- Barnes DKA. Invasions by marine life on plastic debris. Nature 2002;416:808-809.
- Masó M, Garcés J, Pagès F, Camp J. Drifting plastic debris as a potential vector for dispersing Harmful Algal Blooms (HAB) species, Sci. Mar. 2003;67:107-111.
- Barnes DKA, Milner P. Drifting plastic and its consequences for sessile organism dispersal in the Atlantic Ocean. Mar. Biol. 2005;146:815-825.
- Gregory MR. Environmental implications of plastic debris in marine settings-entanglement, ingestion, smothering, hangers-on, hitch-hiking and alien invasions, Philos. Trans. R. Soc. B Biol. Sci 2009;364:2013-2025.
- Barnes DK, Galgani F, Thompson RC, Barlaz M. Accumulation and fragmentation of plastic debris in global environments, Philos. T. Roy. Soc. B 2009;364(1526):1985-1998.
- O'Brine T, Thompson RC. Degradation of plastic carrier bags in the marine environment, Mar. Pollut. Bull 2010;60:2279-2283.
- Cauwemberghe LV, Vanreusel A, Mees JCR, Janssen, Microplastic pollution in deep sea sediments, Environ. Pollut 2013;XXX:1-5.
- Browne MA. Sources and pathways of microplastic to habitats. In M. Bergmann, L. Gutow, & M. Klages (Eds.), Marine anthropogenic litter, Springer, Berlin 2015, 229-244.
- Dris R, Gasperi J, Rocher V, Saad M, Renault N, Tassin B. Microplastic contamination in an urban area: a case study in Greater Paris. Environ. Chem 2015;12(5).
- McCormick TJ, Hoellein MG, London J, Hittie JW, Scott JJ, Kelly. Microplastic in surface waters of urban rivers: Concentration, sources, and associated bacterial assemblages. Ecosphere 2016;7(11).
- Wagner M, Scherer C, Alvarez-Muñoz D, Brennholt N, Bourrain X, Buchinger S, et al. Reifferscheid, Microplastics in freshwater ecosystems: what we know and what we need to know, Environ. Sci. Eur 2014;26:1-9.
- Lusher, Microplastics in the marine environment: Distribution, interactions and effects. (Ed. by Bergmann M. Gutow L. Klages M. Springer, Berlin), In Marine Anthropogenic Litter 2015, 245-308.
- Woodall LC, Sanchez-Vidal A, Canals M, Paterson GLJ, Coppock R, Sleight V et al. Thompson, The deep sea is a major sink for microplastic debris. Roy. Soc. Open. Sci 2014;1(4):140-317.
- Barnes DK, Walters A, Gonclaves L. Macroplastics at

- sea around Antarctica. *Mar. Environ. Res* 2010;70:250-252.
18. Cole M, Lindeque P, Fileman E, Halsband C, Goodhead R, Moger J *et al.* Microplastic ingestion by zooplankton, *Envir. Sci. Technol* 2013;47(12):6646-6655.
 19. Murray F, Cowie PR. Plastic contamination in the decapod crustacean *Nephrops norvegicus* (Linnaeus, 1758), *Mar. Pollut. Bull* 2011;62(6):1207-1217.
 20. Lusher L, McHugh M, Thompson RC. Occurrence of microplastics in the gastrointestinal tract of pelagic and demersal fish from the English Channel, *Mar. Pollut. Bull* 2013;67(1):94-99.
 21. Trevail AM, Gabrielsen GW, Kühn S, Van Franeker JA. Elevated levels of ingested plastic in a high Arctic seabird, the northern fulmar (*Fulmarus glacialis*), *Polar Biol* 2015, 1-7.
 22. Moos VN, Burkhardt-Holm P, Köhler A. Uptake effects of microplastics on cells and tissue of the blue mussel *Mytilus edulis* L. after an experimental exposure, *Environ. Sci. Technol* 2012;46:11327-11335.
 23. Mukhopadhyay P, Edappazham G. Incidence of microplastics in *Villorita cyprinoides* from Vembanad lakes, COMAD, MDT037, 11-12 April Kochi 2018, 85.
 24. Rochman CM, Hoh E, Kurobe T, Teh SW. Ingested plastic transfers hazardous chemicals to fish and induces hepatic stress. *Sci. Rep* 2013;3:32-63.
 25. Ahrendt C, Venegasa DJP, Urbinad M, Gonzalez C, Echevestee P, Aldanah M *et al.* Microplastic ingestion cause intestinal lesions in the intertidal fish *Girella laevis*. *J Mar. Pol. Bul* 2019, 1-6.
 26. Dos Santos J, Jobling M. Gastric emptying in cod, *Gadus morhua* L.: emptying and retention of indigestible solids, *J Fish. Biol* 1991;38:187-197.
 27. Possatto FE, Barletta M, Costa MF, Ivar do Sul JA, Dantas DV. Plastic debris ingestion by marine catfish: an unexpected fisheries impact, *Mar. Pollut. Bull* 2011;62:1098-1102.
 28. Ramos JA, Barletta M, Costa MF. Ingestion of nylon threads by Gerreidae while using a tropical estuary as foraging grounds, *Aquat. Biol* 2012;17:29-34.
 29. Oliveira M, Ribeiro A, Hylland K, Guilhermino L. Single and combined effects of microplastics and pyrene on juveniles (0p group) of the common goby *Pomatoschistus microps* (Teleostei, Gobiidae), *Ecol. Indicat* 2013;34:641-647.
 30. Luís LG, Ferreira P, Fonte E, Oliveira M, Guilhermino L. Does the presence of microplastics influence the acute toxicity of chromium (VI) to early juveniles of the common goby (*Pomatoschistus microps*)? A study with juveniles from two wild estuarine populations, *Aquat. Toxicol* 2015;164:163-174.
 31. Ferreira P, Fonte E, Soares ME, Carvalho F, Guilhermino L. Effects of multistressors on juveniles of the marine fish *Pomatoschistus microps*: gold nanoparticles, microplastics and temperature, *Aquat. Toxicol* 2016;170:89-103.
 32. Sanchez W, Bender C, Porcher JM. Wild gudgeons (*Gobio gobio*) from French rivers are contaminated by microplastics: preliminary study and first evidence, *Environ. Res* 2014;128:98-100.
 33. Mattsson K, Ekvall MT, Hansson LA, Linse S, Malmendal A, Cedervall T. Altered Behavior, Physiology, and Metabolism in Fish Exposed to Polystyrene Nanoparticles, *Environ. Sci. Technol.*, 2015;49:553-561.
 34. Phillips MB, Bonner TH. Occurrence and amount of microplastic ingested by fishes in watersheds of the Gulf of Mexico, *Mar. Pollut. Bull.* 2015;100:264-269.
 35. Avio CG, Gorbi S, Regoli F. Experimental development of a new protocol for extraction and characterization of microplastics in fish tissues: first observations in commercial species from Adriatic Sea, *Mar. Environ. Res* 2015;111:18-26.
 36. Mazurais D, Ernande B, Quazuguel P, Severe A, Huelvan C, Madec L *et al.* Zambonino-Infante, Evaluation of the impact of polyethylene microbeads ingestion in European sea bass (*Dicentrarchus labrax*) larvae, *Mar. Environ. Res* 2015;112:78-85,
 37. De Sá LC, Luís LG, Guilhermino L. Effects of microplastics on juveniles of the common goby (*Pomatoschistus microps*): confusion with prey, reduction of the predatory performance and efficiency, and possible influence of developmental conditions, *Environ. Pollut* 2015;196:359-362.
 38. Lu Y, Zhang Y, Deng Y, Jiang W, Zhao Y, Geng J *et al.* Uptake and Accumulation of Polystyrene Microplastics in Zebrafish (*Danio rerio*) and Toxic Effects in Liver, *Environ. Sci. Technol* 2016;50(7):4054-4060.
 39. Biginagwa FJ, Mayoma BS, Shashoua Y, Syberg K, Khan FR. First evidence of microplastics in the African great lakes: recovery from Lake Victoria Nile perch and Nile tilapia. *J Gt. Lakes. Res* 2015;42(1):146-149.
 40. Peda C, Caccamo L, Fossi MC, Gai F, Andaloro F, Genovese L *et al.* Intestinal alterations in European sea bass *Dicentrarchus labrax* (Linnaeus, 1758) exposed to microplastics: Preliminary results, *Environ. Pollut* 2016;212:251-256.
 41. Peters CA, Bratton SP. Urbanization is a major influence on microplastic ingestion by sunfish in the Brazos River Basin, Central Texas, USA, *Environ. Pollut* 2016;210:380-387.
 42. Grigorakis S, Mason SA, Drouillard KG. Determination of the gut retention of plastic microbeads and microfibers in goldfish (*Carassius auratus*), *Chemosphere* 2017;169:233-238.
 43. Jabeen K, Su L, JN Li, Yang DQ, Tong CF, Microplastics and mesoplastics in fish from coastal and fresh waters of China, *Environ. Pollut* 2017;221:141-149.
 44. Silva-Cavalcanti JS, Silva JD, França EJ, Araújo MC, Gusmão F. Microplastic's ingestion by a common tropical freshwater fishing resource, *Environ. Pollut* 2017;221:218-226.
 45. Valente T, Sbrana A, Scacco U, Jacomini C, Bianchi J, Palazzo L *et al.* Exploring microplastic ingestion by three deep-water elasmobranch species: a case study from the Tyrrhenian Sea, *Environ. Pollut* 2019, 18.
 46. Savoca S, Capillo G, Mancuso M, Bottari T, Crupi R, Branca C, *et al.* Span`o, Microplastics occurrence in the tyrrhenian waters and in the gastrointestinal tract of two congener species of seabreams, *Environ. Toxicol. Pharmacol* 2019.
 47. Kripa VN, Preetha G, Dhanya AM, Pravitha VP, Abhilash KS, Mohammed AA *et al.* Prema, Microplastics in the gut of anchovies caught from the mud bank area of Alappuzha, Kerala. *Marine Fisheries Information Service, Technical and Extension Series* 2014;219:27-28.
 48. Sruthy S, Ramaswami EV. Microplastic pollution in Vembanad lake Kerala, India: the first report of

- microplastics in lake and estuarine sediments in India, *Environ. Pollut* 2017;222:315-322.
49. Rao BM. Microplastics in the aquatic environment: implications for post-harvest fish quality, *Indian J Fish* 2019;66(1):142-152.
 50. Sulochanan B, Bhat GS, Lavanya S. Marine litter in the coastal environment of Mangalore, *Mar. Fish. Infor. Serv. T & E Ser* 2011;208:18-19.
 51. Menon AGK, Remadevi K, Thobias MP. *Puntius chalakkudiensis*, a new colourful species of *Puntius* (family: Cyprinidae) fish from Kerala, south India, *Rec. zool. Surv. India* 1999;97(4):61-63.
 52. Baby F, Tharian J, Philip S, Ali A, Raghavan R. Checklist of the fishes of the Achankovil forests, Kerala, India with notes on the range extension of an endemic cyprinid *Puntius chalakkudiensis*, *JoTT Short Communication* 2011;3(7):1936-1941.
 53. Arunachalam M, Raja M, Nandagopal S, Chandran A. Range Extension of an Endemic Ornamental Fish Species *Puntius halakkudiensis* from Southern Kerala River, India, *RRJZS: e-ISSN: 2321-6190, p-ISSN: 2347-2294* 2014;2(1):20-24.
 54. Jayaram KC. The freshwater fishes of the Indian region. Narendra Pub. House, Delhi 1999, 396,
 55. Hynes HBN. The food of freshwater sticklebacks (*Gasterosteus aculeatus* and *Pygosteus pungitius*) with a review of methods used in studies of the food of fishes. *J Anim. Ecol* 1950;19:36-58.
 56. Nor NH, Obbard JP. Microplastics in Singapore's coastal mangrove ecosystems, *Mar. Pollut. Bull* 2014;79:278-283.
 57. Foekema EM, De Groot C, Mergia MT, van Franeker JA, Murk AJ, Koelmans AA. Plastic in North Sea fish, *Environ. Sci. Technol* 2013;47(15):8818-8824.
 58. De Witte B, Devriese L, Bekaert K, Hoffman S, Vandermeersch G, Cooreman K *et al.* Quality assessment of the blue mussel (*Mytilus edulis*): Comparison between commercial and wild types, *Mar. Pollut. Bull* 2014;85(1):146-155.
 59. Gesamp (Imo/Fao/Unesco-Ioc/Unido/Wmo/Iaea/Un/Unep Joint group of experts on the scientific Aspects of Marine environmental Protection); T. Bowmer and P. J. Kershaw, (eds.), Proceedings of the GESAMP international Workshop on plastic particles as a vector in transporting persistent, bio-accumulating and toxic substances in the oceans. GESAMP rep. stud. no. 2010;82:1-68.
 60. Eerkes-Medrano D, Thompson RC, Aldridge DC. Microplastics in freshwater systems: a review of the emerging threats, identification of knowledge gaps and prioritisation of research needs, *Water Res* 2015;75:63-82.
 61. Pinheiro C, Oliveira U, Vieira M. Occurrence and impacts of microplastics in freshwater fish. *J Aquac. Mar. Biol* 2017;5:1-5.
 62. Bakir SJ, Rowland RC. Thompson, Competitive sorption of persistent organic pollutants onto microplastics in the marine environment, *Mar. Pollut. Bull* 2012;64:2782-2789.
 63. Zarfl Matthias M. Are marine plastic particles transport vectors for organic pollutants to the Arctic? *Mar. Pollut. Bull* 2010;60:1810-1814.
 64. Pavani P, Rajeswari TR. Impact of heavy metals on environmental pollution. *J of Chem. And Pharmaceu. Sci* 2014;94(3):87-93.
 65. Mattsson K, Ekvall MT, Hansson LA, Linse S, Malmendal A, Cedervall T. Altered behavior, physiology, and metabolism in fish exposed to polystyrene nanoparticles, *Environ. Sci. Technol* 2014;49:553-561.
 66. Rochman CM, Kurobe T, Flores I, Teh SJ. Early warning signs of endocrine disruption in adult fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the marine environment, *Sci. Total Environ* 2014;493:656-661.
 67. Greven AC, Merk T, Karag oz F, Mohr K, Klapper M, Jovanovi CB *et al.* Polycarbonate and polystyrene nanoplastic particles act as stressors to the innate immune system of fathead minnow (*Pimephales promelas*), *Environ. Toxicol. Chem* 2016;35:3093-3100.
 68. Espinosa C, Esteban M A, Cuesta A. Microplastics in Aquatic Environments and Their Toxicological Implications for Fish, *Toxicol. New Aspects to This Scientific Conundrum* 2016, 113.
 69. Van Cauwenberghe L, Janssen CR. Microplastics in bivalves cultured for human consumption, *Environ. Pollut* 2014;193:65-70.
<http://dx.doi.org/10.1016/j.envpol.2014.06.010>.
 70. Mattsson K, Johnson EV, Malmendal A, Linse S, Hansson LA, Cedervall T. Brain damage and behavioural disorders in fish induced by plastic nanoparticles delivered through the food chain, *Sci. Rep* 2017;7:11452.
 71. Halstead JE, Smith JA, Carter EA, Lay PA, Johnston EL. Assessment tools for microplastics and natural fibres ingested by fish in an urbanised estuary, *Environ. Pollut* 2018;234(2018):552-561.
 72. Campbell SH, Williamson PR, Hall BD. Microplastics in the gastrointestinal tracts of fish and the water from an urban prairie creek, *Facets* 2017;2:395-409.
 73. Horton A, J rgens MD, Lahive E, van Bodegom PM, Vijver MG. The influence of exposure and physiology on microplastic ingestion by the freshwater fish *Rutilus rutilus* (roach) in the River Thames, UK. *Environ. Pollut* 2018;236:188-194.
 74. Wang W, Ndungu AW, Li Z, Wang J. Microplastic's pollution in inland freshwaters of China: a case study in urban surface waters of Wuhan, China. *Sci. Total Environ* 2016.
 75. Zhao S, Zhu L, Wang T, Li D. Suspended microplastics in the surface water of the Yangtze estuary system, China: first observations on occurrence distribution, *Mar Pollut. Bull* 2014;86(1, 2):562-568.
 76. Lusher L, Hernandez-Milian G, O'Brien J, Berrow S, O'Connor IR. Officer, Microplastic and macroplastic ingestion by a deep diving, oceanic cetacean: The True's beaked whale *Mesoplodon mirus*, *Environmental Pollution* 2015;199:185-191.
 77. Lusher PCH, Hollman JJ, Mendoza-Hill. Microplastics in fisheries and aquaculture: status of knowledge on their occurrence and implications for aquatic organisms and food safety, In *FAO Fisheries and Aquaculture Technical Rome, Italy* 2017, 615.
 78. Moser ML, Lee DS. A fourteen-year survey of plastic ingestion by western North Atlantic seabirds. *Colon Water bird* 1992;15(1):83-94.
 79. Moore CJ. Synthetic polymers in the marine environment: a rapidly increasing, longterm threat, *Environ. Res* 2008;108(2):131-139.

80. Boerger CM, Lattin GL, Moore SL, Moore CJ. Plastic ingestion by planktivorous fishes in the North Pacific central gyre, *Mar. Pollut. Bull* 2010;60(12):2275-2278.
81. Hoarau L, Ainley L, Jean C, Ciccione S. Ingestion and defecation of marine debris by loggerhead sea turtles, *Caretta caretta*, from by-catches in the South- West Indian ocean, *Mar. Pollut. Bull* 2014;84(1):90-96.
82. Zhao S, Zhu L, Li D. Microplastic in three urban estuaries, China. *Environ. Pollut* 2015;206:597-604.
83. Andrady AL. Microplastics in the marine environment, *Mar. Pollut. Bull* 2011;62(8):1596-1605.
84. Eriksen M, Mason S, Wilson S, Box C, Zellers A, Edwards W *et al.* Microplastic pollution in the surface waters of the Laurentian Great Lakes. *Mar. Pollut. Bull* 2013;77(1):177-182.
85. Yonkos LT, Friedel EA, Perez-Reyes AC, Ghosal S, Arthur CD. Microplastics in four estuarine rivers in the Chesapeake Bay, USA, *Environ. Sci. Technol* 2014;48(24):14195-14202.
86. Browne MA, Crump P, Niven SJ, Teuten E, Tonkin A, Galloway T *et al.*, Accumulation of microplastic on shorelines worldwide: sources and sinks, *Environ. Sci. Technol* 2011;45:9175-9179.
87. Harner T, Shoeib M, Diamond M, Stern G, Rosenberg B. Using passive air samplers to assess urban-rural trends for persistent organic pollutants: polychlorinated biphenyls and organochlorine pesticides, *Environ. Sci. Technol* 2004;38:4474-4483.
88. Napper IE, Thompson RC. Release of synthetic microplastic plastic fibres from domestic washing machines: effects of fabric type and washing conditions, *Mar. Poll. Bull* 2016;112:39-45.
89. Free CM, Jensen OP, Mason SA, Eriksen M, Williamson NJ, Boldgiv B. High-levels of microplastic pollution in a large, remote, mountain lake, *Mar. Pollut. Bull* 2014;85(1):156-163.