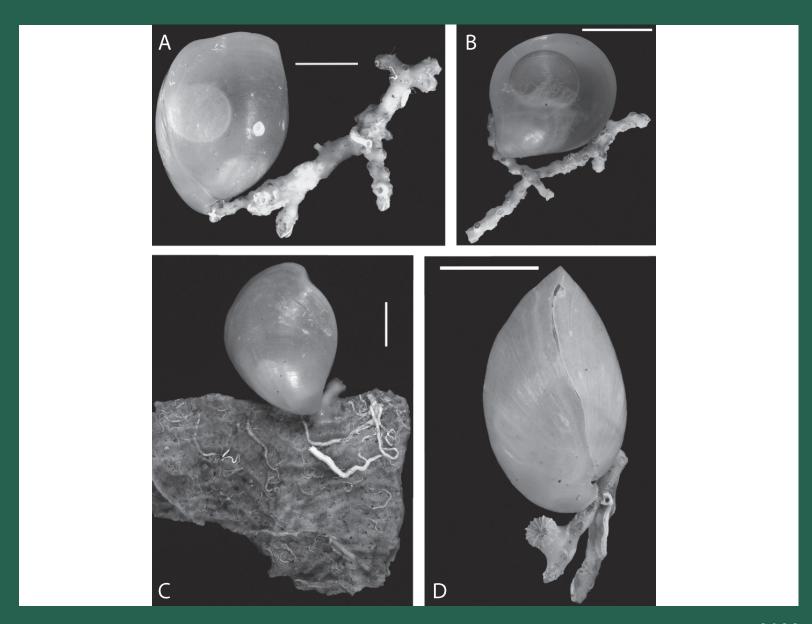
## BULLETIN

OF THE

# FLORIDA MUSEUM OF NATURAL HISTORY





VOL. 59, NO. 1, PP. 1-15

2022

The **FLORIDA MUSEUM OF NATURAL HISTORY** is Florida's state museum of natural history, dedicated to understanding, preserving, and interpreting biological diversity and cultural heritage.

The **BULLETIN OF THE FLORIDA MUSEUM OF NATURAL HISTORY** is an on-line, open-access, peer-reviewed journal that publishes results of original research in zoology, botany, paleontology, archaeology, and museum science. New issues of the Bulletin are published at irregular intervals, and volumes are not necessarily completed in any one year. Volumes contain between 150 and 300 pages, sometimes more. The number of papers contained in each volume varies, depending upon the number of pages in each paper, but four numbers is the current standard. Multi-author issues of related papers have been published together, and inquiries about putting together such issues are welcomed. Address all inquiries to the Editor of the Bulletin.

The electronic edition of this article conforms to the requirements of the amended International Code of Zoological Nomenclature, and hence the new names contained herein are available under that Code. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN.

Roger W. Portell and Richard C. Hulbert Jr., Editors for this issue

#### **Bulletin Committee**

Richard C. Hulbert Jr.
Michelle J. LeFebvre
Jacqueline Miller
Roger W. Portell
Jonathan I. Bloch, *Ex officio Member* 

ISSN: 2373-9991

Copyright © 2022 by the Florida Museum of Natural History, University of Florida. All rights reserved. Text, images and other media are for nonprofit, educational, and personal use of students, scholars, and the public. Any commercial use or republication by printed or electronic media is strictly prohibited without written permission of the museum.

Publication Date: February 1, 2022

This and other issues of the Bulletin can be freely downloaded at:

https://www.floridamuseum.ufl.edu/bulletin/publications/

#### Send communications about this publication to:

Editor of the Bulletin; Florida Museum of Natural History; University of Florida; P.O. Box 117800;

Gainesville, FL 32611-7800 USA

FAX: 352-846-0287; Email: bulletin@flmnh.ufl.edu URL: https://www.floridamuseum.ufl.edu/bulletin/home/

Cover image: Specimens of *Tichosina plicata* attached to deep-water corals.

### OCCURRENCE OF THE BRACHIOPOD *TICHOSINA* IN DEEP-SEA CORAL BOTTOMS OF THE CARIBBEAN SEA AND ITS PALEOENVIRONMENTAL IMPLICATIONS

Alexis Rojas<sup>1</sup>, Adriana Gracia<sup>2</sup>, Ivan Hernández-Ávila<sup>3</sup>, Pedro Patarroyo<sup>4</sup>, and Michał Kowalewski<sup>5</sup>

#### **ABSTRACT**

Despite its importance as the larger component of the modern and Cenozoic brachiopod faunas in the Caribbean region, the ecology and habitat preferences of the terebratulid *Tichosina* remain poorly understood. We compiled field observations from multiple sites in the Caribbean of Colombia (i.e., San Bernado Bank, Bahia Honda-Guajira, Puerto Escondido, Joint Regime Area Jamaica-Colombia) and data from the R/V Pillsbury program, indicating that *Tichosina* may have close ecological ties with deep-water corals. In addition, we reviewed literature sources on Cenozoic sediments in the Dominican Republic and found tentative evidence that such ecological ties could have existed since at least the Pliocene. These observations are reminiscent of the *Gryphus*-anthozoan association observed along the modern Mediterranean continental margin. Understanding to what extent the brachiopod *Tichosina* is linked to deep-water habitats has implications for the recognition of deep-water macrobenthic communities in the Cenozoic rock record of the Caribbean.

Key words: Caribbean Sea, Colombia, deep-coral habitats, brachiopods, Tichosina

#### TABLE OF CONTENTS

Introduction	2
Materials and Methods	2
Results	
Discussion	
Conclusions	
Acknowledgments	12
Literature Cited	

<sup>&</sup>lt;sup>1</sup>Department of Physics, Umeå University, SE-901 87 Umeå, Sweden <alexis.rojas-briceno@umu.se>.

<sup>&</sup>lt;sup>2</sup>Programa de Biología, Universidad del Atlántico, Km 7 Antigua vía Puerto Colombia, Colombia. Instituto de Investigaciones Marinas y Costeras, INVEMAR, Santa Marta, Colombia <a href="mailto:mariaadrianagracia@mail.uniatlantico.edu.co">mailtantico.edu.co</a>.

<sup>&</sup>lt;sup>3</sup>Facultad de Ciencias Naturales, Universidad Autónoma del Carmen, Ciudad del Carmen, México <ihernandez@pampano.unacar.mx>.

<sup>&</sup>lt;sup>4</sup>Departamento de Geociencias, Universidad Nacional de Colombia. A. A. 14490, Bogotá, Colombia; <pcpatarroyog@unal.edu.co>.

<sup>&</sup>lt;sup>5</sup>Florida Museum of Natural History, Division of Invertebrate Paleontology, University of Florida, Gainesville, Florida, 32611, USA;

<sup>&</sup>lt;kowalewski@flmnh.ufl.edu>.

#### **INTRODUCTION**

Deep-sea brachiopods have been recorded in the Caribbean Sea by several scientific expeditions. The U.S. Coast Survey vessel Blake (late 1870s), the U.S. Fish Commission Albatross (1880s), the R/V Oregon and R/V Oregon II (1950s to 1970s), the R/V Gerda (1960s), and the R/V Pillsbury campaigns (1960s and 1970s) (see also Lutz and Ginsburg, 2007) provided abundant material to describe their taxonomy, general bathymetric and geographic distribution (Cooper, 1977; Logan, 1990). Micromorphic forms (<10 mm in shell length), including Argyrotheca and Terebratulina species, are known to inhabit caves and other cryptic habitats (Jackson et al., 1971; Asgaard and Stentoft, 1984; Logan, 1975; Richardson, 1997) but under low-intensity pressures they can also occur on relatively exposed surfaces (Logan, 1977). In contrast, medium to large size brachiopods, comprising 18 Tichosina species occurring in the Caribbean Sea and the Gulf of Mexico (Cooper, 1977; Rojas et al., 2015), have not been studied extensively and their taxonomy and ecology remain poorly understood (Harper et al., 1997; Oyen et al., 2000; Harper and Donovan, 2007; Emig et al., 2013). These brachiopods are known to occur at depths ranging between 100 and 200 m (Cooper, 1977), most likely below the photic zone, but their potential association with deep-corals habitats, known to occur at depths greater than 70 m (Roberts, 2006; Ramirez-Llodra et al., 2010), has not been evaluated. In contrast, fossil brachiopods in the Caribbean region have been extensively studied with regards to their stratigraphy and taxonomy (Harper, 2002; Harper and Donovan, 2007; Harper and Pickerill, 2008), and biogeography (Shemm-Gregory et al., 2012; Rojas and Sandy, 2018; Sandy and Rojas, 2018).

Although deep-sea corals across the Caribbean Sea have been widely studied during the last decades (Cairns, 1979, 2007; Lutz and Ginsburg, 2007; Davies and Guinotte, 2011; Santodomingo et al., 2013; Hernandez-Avila, 2014), our current knowledge of the brachiopod faunas associated with these coral-supported environments is limited

(Cooper, 1979). These light-independent structures generated by azooxanthellate corals (i.e., corals lacking symbiotic dinoflagellates), including banks, bioherms, lithoherms, and other coral-supported habitats, provide protection and food, as well as breeding, spawning, resting, and nursery areas for several species of invertebrates and vertebrates (Roberts, 2006; Tursi et al., 2004). For instance, a deep-water coral bank located in the southwestern Caribbean of Colombia is known to provide a habitat for at least 118 species of fishes and invertebrates, including echinoderms, crustaceans, gastropods, bivalves, bryozoans, and brachiopods (Reyes et al., 2005; Rojas et al., 2015). However, the distinction of these deep-water macrobenthic communities in the Cenozoic rock record is challenging (Squires, 1964; Mullins et al., 1980; Kano et al., 2007) and water-depth interpretations of some biosediments in the region remains controversial (McNeill et al., 2008).

In this paper, we compiled field observations on brachiopods from multiple deep-shelf settings in the Caribbean of Colombia and available data from the R/V Pillsbury sampling program across the Caribbean (1966 to 1972), indicating that occurrences of *Tichosina* have been recorded consistently in samples containing structure forming and suspected habitat-forming corals. In addition, we discuss literature sources providing tentative evidence that such brachiopod-anthozoan ties could have existed at least since the Pliocene. Understanding to what extent the brachiopod *Tichosina* is linked to deep-coral habitats through the modern Caribbean has implications for both Tertiary geology and paleoecology in this region.

#### **MATERIAL AND METHODS**

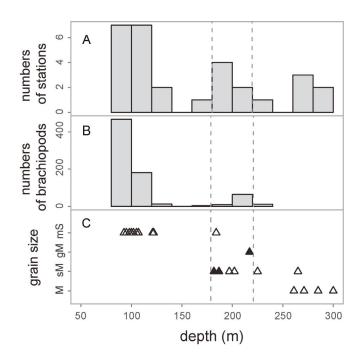
DATA

Three main data sources have been used in our study: literature sources, field collection observations, and databases. First, we examined uncatalogued *Tichosina* shells and fragments deposited at the Museo de Historia Natural Marina of the Instituto de Investigaciones Marinas y Costeras (INVEMAR, Colombia). The materials

were collected during the MACROFAUNA II (2000), MARCORAL (off San Bernardo Archipelago), ANH-I (Bahia Honda-Guajira, off Cartagena and Puerto Escondido), and ANH-Jamaica (northwestern Macondo Guyot) projects that were focused on characterizing deep-water coral and soft-bottom communities and associated populations on the continental shelf of Colombia. These field surveys resulted in the recognition of a deep-water coral structure called San Bernardo Bank (Lutz and Ginsburg, 2007), as well as other coral-dominated habitats (see Reyes et al., 2005; Santodomingo et al., 2007; Santodomingo et al., 2013) on the Caribbean continental shelf of Colombia. Then, we used a dataset on the benthic macrofauna from the R/V Pillsbury sampling program (1966 to 1972), including corals, mollusks, and echinoderms, assembled from databases of the Smithsonian Natural Museum of Natural History (NMNH) and the Marine Invertebrate Museum, Rosenstiel School of Marine and Atmospheric Science (MIN-RSMAS) (Hernández-Ávila, 2014). Brachiopod data from the R/V Pillsbury sampling program were gathered from an extensive monograph on modern Caribbean brachiopods by Cooper (1977). Finally, we inspected Cooper's collection (Cooper, 1977) at the NMNH for direct evidence of Tichosina shells growing on deepwater corals.

#### **Methods**

Brachiopod shells and fragments from the MACROFAUNA II, MARCORAL, ANH-I and ANH-Jamaica projects were handpicked and examined under a binocular microscope. Complete specimens of *Tichosina plicata* were counted and measured. As defined in Rojas and Sandy (2019), a complete specimen includes any shell sufficiently complete to measure its full dimensions (i.e., length and width). A taxonomic description of the brachiopods studied here is provided in Rojas et al. (2015). We used samples from the MARCORAL cruise, carried out between April 22 and May 2, 2005, on board R/V Ancon off the San Bernardo Archipelago (Reyes et al., 2005), to illustrate variations in sampling effort, total number of



**Figure 1.** Sampling data from the INVEMAR MARCORAL project carried out offshore San Bernardo, Colombia. A, Total number of stations; B, Total number of brachiopods, including *Terebratulina cailleti* (503 specimens), *Argyrotheca barrettiana* (208 specimens) and *Tichosina plicata* (6 specimens); C, Sediment grain size per station. Stations were *T. plicata* was recovered are indicated in black. Abbreviations for texture classes are: mud (M), sandy mud (sM), muddy sand (mS), sand (S) and gravelly mud (gM). Depth distribution of the San Bernardo Bank indicated by a segmented line.

brachiopod specimens, and sediment grainsize at each station (Fig. 1). The sampling methodology of this survey was previously described by Rangel-Buitrago and Idárraga-García (2010).

The assembled dataset on the benthic macrofauna from the R/V Pillsbury sampling program was used to reconstruct the habitat of *Tichosina* spp. across the Caribbean Sea. The brachiopod specimens from these expeditions were described in Cooper (1977) and are housed in the Paleobiology Collections of the NMNH. Cooper's monograph was focused on brachiopod taxonomy, and it does not provide information on their interactions. Although all the materials

recovered during the Pillsbury expeditions were divided into major taxonomic groups and placed in specialized collections, AR inspected some available brachiopods in the wet-preserved brachiopod collections at the NMNH for any remaining evidence of the presumed *Tichosina*-anthozoan relationship.

multidimensional Nonmetric scaling (Kruskal, 1964) was used to explore distribution patterns of Tichosina and structure-forming and potential habitat-forming corals across the R/V Pillsbury sampling stations. It was performed using census data at the genus taxonomic-level and the Bray Curtis distance measure after removing rare taxa that provide limited interpretative value (Poos and Jackson, 2012) on the occurrence of Tichosina. This ordination analysis was performed using the R-package vegan version 2.5-1 (Oksanen et al., 2018). In addition, we created a bipartite network representation of the census data gathered from R/V Pillsbury sampling stations (Data S1) and used a clustering analysis called Infomap (Rosvall and Bergstrom, 2008; Calatayud et al. 2019) to identify the potential impact of removing rare taxa in the multivariate analysis and further explore the modular structure of the benthic macrofauna. This community detection approach allows us to identify clusters of highly connected stations and taxa. We performed the network analysis using the MapEquation framework, available online at http:// www.mapequation.org.

#### **RESULTS**

TICHOSINA IN DEEP-SEA CORAL BOTTOMS OF THE COLOMBIAN CARIBBEAN SEA

The combined *Tichosina* material from the INVEMAR projects comprises live and dead specimens, including 55 conjoined shells and 24 disarticulated valves. This shell material represents ten sampling stations from five different sites along the Caribbean continental shelf of Colombia. These sites are located offshore of Guajira Province, Bolivar Province, Córdoba Province, and the northwestern Macondo Guyot in the Joint Regime Area (JRA) Jamaica-Colombia (Gutiérrez-

Salcedo et al., 2015) (Table 1; Fig. 1). Deep-sea Tichosina were recorded at depths ranging between 120 and 500 m and all studied localities registered a single species, Tichosina plicata (Rojas et al., 2015). Well-preserved dead specimens were found in sandy and gravelly litho-bioclastic mud (Rangel-Buitrago and Idárraga-García, 2010) (Fig. 2). Although the dominant coral species in the San Bernardo Bank is Madracis myriaster (Reyes et al., 2005; Santodomingo et al., 2007), a few live specimens were found attached to dead and live branching corals Madracis asperula, Madracis sp., and in fragments of Agaricia sp. and Anomocora fecunda (Fig. 2). Dead specimens of Tichosina were observed attached to the stony coral Caryophyllia sp. Offshore San Bernardo Archipelago, the deep-sea brachiopod fauna also included three micromorphic forms, Terebratulina cailleti (503 specimens), Argyrotheca barrettiana (208 specimens), and Terebratulina latifrons (one specimen) (Rojas et al., 2015). These brachiopods were common on the continental shelf with only a few occurring on the shelf break and in association with the deep-water coral structures.

Dead and living specimens of *Tichosina* sp. in environments dominated by solitary scleractinians were also observed in Bahia Honda-Guajira (265 m ANH I Project, Sts. E253 and E254), off Cartagena (500 m ANH I Project, St. E264), off Puerto Escondido (500 m ANH I Project, St. E267) and the northwestern Macondo Guyot (320 m ANH-Jamaica, St. E296). Geological and biological evidence suggest the presence of deep-coral structures in all those sites.

 $\it Tichosina$  and the Benthic Megafauna from the R/V Pillsbury Sampling Stations

A total of five species of *Tichosina* were reported from 28 stations of the R/V Pillsbury sampling program, including *T. plicata* (=*T. dubia*), *T. obesa, T. truncata, T. bartletti*, and *T. cubensis*. These five species represent 28% of all species assigned to this genus that have been reported from the Caribbean and the Gulf of Mexico (Cooper, 1977). Because most of the species of this genus appear to be synonymous (Emig et al., 2013;

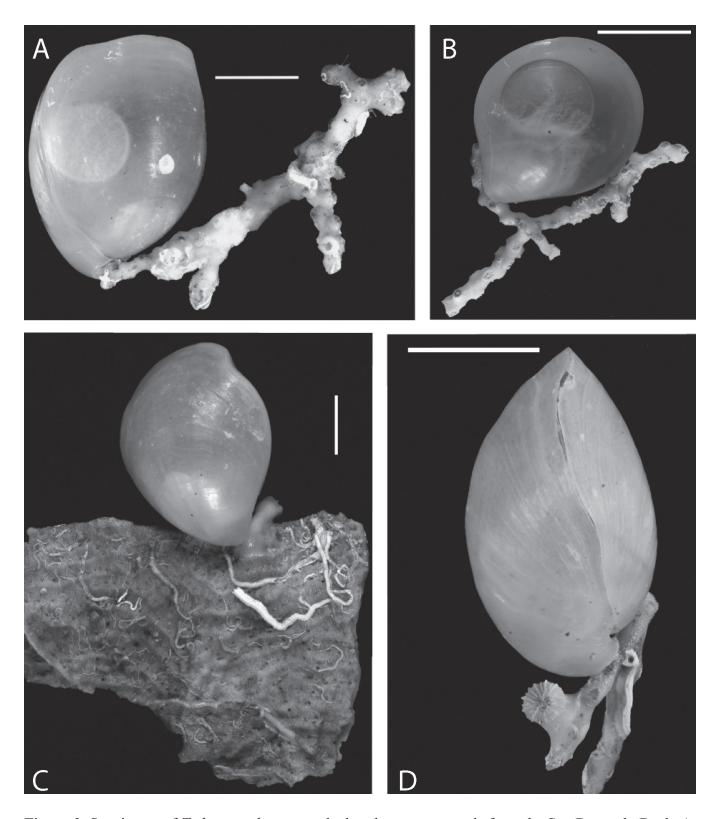
**Table 1**. Information on sampling stations were *Tichosina* species have been recovered in the Caribbean Sea by the INVEMAR projects and the R/V Pillsbury campaigns. Brachiopod data of the R/V Pillsbury campaigns from Cooper (1977).

# spec.	Species	Depth (m)	Station ID	Site	Project Source		
	T. plicata	265	E253	Bahia Honda-Guajira	INVEMAR	ANH-1	
10	T. plicata	265	E254	Bahia Honda-Guajira	_	ANH-1	
1	T. plicata	500	E264	Offshore-Cartagena	_	ANH-1	
3	T. plicata	Offshore-Puerto Escondido E267 500 T. pla		Offshore-Puerto Escondido	_	ANH-1	
2	T. plicata	320	E296	Northwestern Macondo Guyot	_	ANH-Jamaica	
33	T. plicata	160	E155	Tolú	_	MACROFAUNA II	
8	T. plicata	155	E156	Tolú	_	MACROFAUNA II	
4	T. plicata	Offshore-San Bernardo D3 210–217		Offshore-San Bernardo	_	MARCORAL	
1	T. plicata	181-184	D128*	Offshore-San Bernardo	_	MARCORAL	
1	T. plicata	178–180	D28*	Offshore-San Bernardo	_	MARCORAL	
4	T. bahamiensis	619	2655	Grand Bahama	Cooper	Albatross	
2	T. plicata*	458	147	St. Kitts	_	Blake	
1	T. martiinicensis	309	193	Martinique	_	Blake	
3	T. elongata	641	450	La Isabela	_	Combat	
Ç	T. subtriangulata	403-412	6070	Mayaguez	_	Fishhawk	
3	T. erecta	622–695	694	Grand Bahama	_	Gerda	
(	T. rotundovata	201–210	482	Key West	_	Gerda	
3	T. plicata*	201	3608	East Nicaragua	_	Oregon	
1	T. floridensis	183	955	West Florida Shelf	_	Oregon	
10	T. floridensis	119	1025	West Florida Shelf	_	Oregon	
2	T. obesa	366	1408	New Orleans	_	Oregon	
11	T. ovata						
2	T. plicata*	183	10513	Georgetown	_	Oregon II	
2	T. labiata	231–258	876	St Vincent	_	Pillsbury	
3	T. obesa	60-73	737	Northeast Caracas	_	Pillsbury	
2	T. obesa	60-68	734	Northwest Guanta	_	Pillsbury	
3	T. pillsburyae	130–165	1387	Macoris	_	Pillsbury	
2	T. truncata	348-353	584	Mujeres Island	_	Pillsbury	
	T. expansa						
1	T. obesa						
1	T. expansa	549	3499	Great Iguana	_	Silver Bay	

<sup>\*</sup>Species name updated as per Rojas et al. (2015)

Rojas et al., 2015), our analyses are performed at the genus level. Occurrences of the brachiopod *Tichosina* in the R/V Pillsbury data is limited to only 28 out of 318 stations with coral records. The scarcity of *Tichosina* may likely reflect the sampling method (e.g., otter trawls), which was not

optimal for capturing small and fragile specimens. Remarkably, all sampling stations from the R/V Pillsbury sampling program recording *Tichosina* brachiopods, as reported in Cooper (1977), also register structure-forming and potential habitat-forming corals in our compilation. The megafauna



**Figure 2.** Specimens of *Tichosina plicata* attached to deep-water corals from the San Bernardo Bank. A, *Madracis asperula*; B, *Madracis* sp.; C, Dead *Agaricia* sp. and recruit of Caryophylliidae; D, *Madracis* sp. and recruit of Caryophylliidae. Scale bars 10 mm.

**Table 2.** Major components of the benthic megafauna from R/V Pillsbury stations where *Tichosina* species have been recovered in the Caribbean Sea.

Corals	% occ.	Echinoids	% occ.	Crinoids	% occ.	Asteroids	% occ.	Gastropods	% occ.
Ellisella sp.	32.1	Echinolampas depressa	25.0	Comactinia echinoptera	28.6	Cheiraster echinulatus	21.4	Naticidae indet.	14.3
Thesea sp.	32.1	Coelopleurus floridanus	25.0	Crinometra brevipinna	21.4	Echinaster modestus	14.3	Cocculinidae indet.	10.7
Acanthogorgia sp.	25.0	Agassizia excentrica	21.4	Neocomatella alata	21.4	Anthenoides peircei	14.3	Lamellitrochus lamellosus	10.7
Placogorgia sp.	21.4	Araeosoma belli	17.9	Comissia venustus	17.9	Circeaster americanus	10.7	Conus cancellatus	10.7
Siphonogorgia sp.	17.9	Cidaris rugosa	17.9	Hypalometra defecta	17.9	Goniaster tessellatus	10.7	Conus sp.	10.7
Bebryce sp.	17.9	Genocidaris maculata	17.9	Comactinia meridionalis	s 10.7	Rosaster alexandri	10.7	Conus villepinii	10.7
Antipathes lenta	17.9	Stylocidaris affinis	17.9	Stylometra spinifera	10.7	Other 31 spp.	< 10	Other 57 spp.	< 10
Trochocyathus rawsonii	17.9	Stylocidaris lineata	17.9	Endoxocrinus parrae	10.7				
Other 83 spp.	< 15	Other 44 spp.	<15	Other 7 spp.	<10				

% occ: percentage of occurrence for each species across stations

associations at those stations represent relatively diverse deep-water benthic communities, including corals (91 species), gastropods (63 species), echinoids (52 species), asteroids (37 species) and crinoids (15 species) (Table 2). The diverse material collected from those stations includes the structure-forming coral Madrepora oculata, as well as a number of potential habitat-forming corals (sensu Lutz and Ginsburg, 2007), including Acanella sp., Diodogorgia sp., Riisea paniculata, Swiftia exserta, Parantipathes sp., Coenosmilia arbuscula, Madracis asperula, M. formosa, Nicella americana, N. guadalupensis, Neospongodes sp., Oxysmilia rotundifolia, Thalamophyllia riisei, Eguchipsammia cornucopia, and Polymyces fragilis. Each of those coral species occurs in 4 to 11% of the total stations.

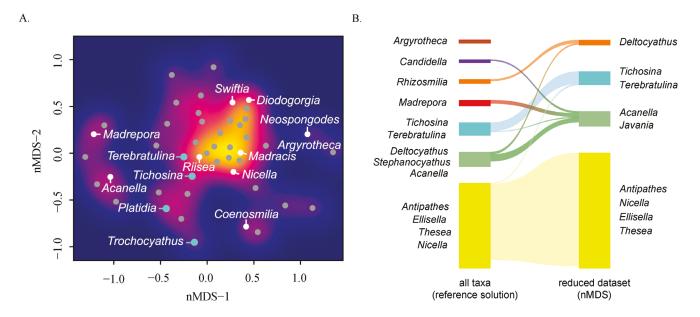
The non-metric multidimensional scaling ordination (Fig. 3A) does not show a clear distinction between habitat-forming corals and other anthozoans across the R/V Pillsbury sampling stations. However, Tichosina ordinates in close proximity to habitat-forming (i.e., Madracis) and potential habitat-forming (i.e., Swiftia, Riisea, and Nicella) taxa. The well-known structure-forming coral Madrepora plots outside this region. Estimation of the Besag's L-function (Besag, 1977) indicates significant clustering for the point pattern resulted from the ordination analysis. Network clustering analyses based on the reduced dataset used in the multivariate analysis as well as the complete dataset that includes rare

taxa show a similar modular structure; i.e., despite removing rare taxa, we were able to retrieve similar modules or communities (Fig. 3B). The reference solution, obtained by clustering the assembled network representing the R/V Pillsbury campaign, includes 29 modules that comprise both sampling stations and genera (Data S2). In this network partition, Tichosina is clustered with deep-water brachiopods as well as solitary, attached, or freeliving corals, including the potential habitatforming Parantipathes and Polymyces (sensu Lutz and Ginsburg, 2007). The three larger modules delineating in the structure of this benthic megafauna are shown in Fig. 4. Finally, inspection of Cooper's collection (Cooper, 1977) uncovered a single wet sample from Venezuela (uncataloged, Pillsbury sampling station P736) of Tichosina growing on an undetermined coral fragment (Fig. 4).

#### **DISCUSSION**

TICHOSINA IN DEEP-SEA CORAL BOTTOMS OF THE COLOMBIAN CARIBBEAN SEA

The MACROFAUNA II and MARCORAL sampling programs targeted deep-sea corals structures and the sampling methodologies were not suitable for recovering fragile material such as shells of *Tichosina*. Consequently, the available material is represented largely by shell fragments and its abundance, based on complete specimens, is likely severely underestimated. In contrast, the relatively



**Figure 3.** Ordination and network-based clusters for combined data. A. Non-metric multidimensional scaling plot for combined coral and brachiopod genus census data from the R/V Pillsbury campaigns. Only common genus (recorded in three or more samples) and samples in which three or more species are recorded were used in this ordination. The stress value for the three-dimensional analysis is 0.13, and the *R*-squared value for the linear fit of observed dissimilarity and ordination distance is 0.9. B. Alluvial diagram showing the differences between the two partitions derived from network representations of the total census data from R/V Pillsbury sampling stations and the reduced dataset generated after removing rare taxa.

sturdy shells of the micromorphic brachiopods in our samples, Argyrotheca and Terebratulina, were dominated by conjoined specimens. This negative effect of some sampling methodologies on selected taxa has been observed in modern ecological studies (Costello et al., 2017). Remarkably, despite this sampling bias, complete specimens and abundant fragments of T. plicata were still recovered at sites where branching *Madracis* spp. and other potential habitat-forming corals were present. The azooxanthellate Madracis corals are considered the main framework builder of deepwater coral structures in the Colombian Caribbean (Santodomingo et al., 2007). Other scleractinian corals considered to represent habitat-forming species and a potential habitat to Tichosina include Anomocora fecunda, Coenosmillia arbuscula, *Eguchipsammia* cornucopia, Thalamophyllia riisei, and Javania cailleti (Reyes et al., 2005; Santodomingo et al., 2007). The consistent cooccurrence of live specimens of both *Tichosina* and deep-water corals points to a potential *Tichosina*-anthozoan association in the Caribbean continental shelf of Colombia. Although lack of *in situ* observations prevent a detailed description of such an interaction, it is likely that *Tichosina* occurs on the periphery of the coral structures, i.e., off-mound (Rosso et al., 2010). Well-preserved specimens of *T. rotundovata* have been also reported in association with deep-water coral structures in the little Bahama Bank at 485–496 m water depth (Cooper, 1977:21).

TICHOSINA AND THE BENTHIC MEGAFAUNA OF THE R/V PILLSBURY SAMPLING PROGRAM

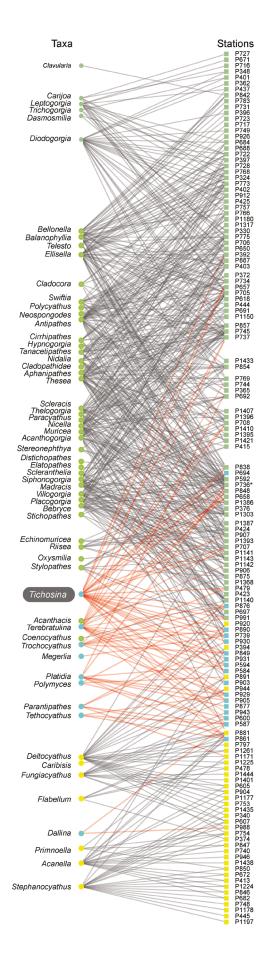
Our data compilation suggests that *Tichosina* spp. are associated with deep-water benthic communities dominated by corals and other characteristic megafaunal elements including gastropods, echinoids, asteroids, and crinoids. The

**Figure 4.** Bipartite representation of the census data from R/V Pillsbury sampling stations with nodes, species, and stations, colored by their module affiliation. (\*) Sample with *Tichosina* shell growing on undetermined coral fragment in the Cooper's collection. Only the three largest modules are illustrated.

coral material reported in those sampling stations includes structure-forming and potential habitatforming corals. Those corals appear to provide the optimum structural habitat for *Tichosina* species to grow across the Caribbean. The number of azooxanthellate corals in the Caribbean region is higher on the continental slope (Hernández-Ávila, 2014; Hernández-Ávila et al., 2018), which coincides with the depth distribution of Tichosina species (Cooper, 1977; Harper et al., 1995; Harper, 2002). The commensal relationship between T. floridensis and benthic foraminifera occurring at depths of 120-180 m on the west Florida shelf illustrates the relationship of these brachiopods with the deep-water benthos (Zumwalt and Delaca 1980). Although the original material recovered from the R/V Pillsbury survey has been divided into major zoological groups and housed in specialized collections, we inspected some available wet samples of brachiopods at the NMNH and found remains at station P736 from Venezuela that are reminiscent of the substrate recovered at the San Bernardo Bank.

Different deep-coral species appear to offer a suitable habitat to *Tichosina* across the Caribbean (Fig. 4). Despite the fact, that all samples from the R/V Pillsbury survey containing *Tichosina* brachiopods also include known or potential habitat-forming corals, results from the NMS indicates there is not a preferred coral species by *Tichosina* to use as a habitat. However, network clustering analysis points to both *Parantipathes* and *Polymyces* as the most suitable corals as habitat for *Tichosina*. This finding should be further corroborated with field observations.

Coral-brachiopod associations in deepwater environments are known from temperate



zones, where deep-coral banks provide widespread habitats for abundant and diverse benthic organisms (Teichert, 1958; Squires, 1964; Logan, 1979; Rosso et al., 2010). Similarly, based on direct observations and dredging, Emig (1987, 1989) reported that Mediterranean continental-margin environments (depths of 110-250 m) with strong bottom currents, low sedimentation rates, and high nutrient delivery harbor higher density populations of Gryphus vitreus. The San Bernardo Bank is similar to the Mediterranean settings in that it is influenced by an undercurrent called the Panama-Colombia Countercurrent (Andrade et al., 2003). The local nutrient enrichment potentially associated with this current is unknown. Moreover, Gryphus and Tichosina represent Cenozoic genera that are similar in shape and size (Harper and Pickerill, 2008). Gryphus is well represented in the Eocene of Cuba but is absent in the modern Caribbean Sea (Cooper, 1979). A recent study of modern surface sediments in a deep-water coral mound located at Santa Maria di Leuca (Mediterranean) confirmed the association of Gryphus vitreus with coralsupported structures (Rosso et al., 2010). These previous studies and the findings reported here suggest that populations of *Tichosina* are likely to concentrate along the continental margins of the Caribbean, where suitable habitats for deep-water corals exist (Hernandez-Avila, 2014).

TICHOSINA FROM DEEP-WATER CORAL HABITATS IN NEOGENE DEPOSITS IN THE DOMINICAN REPUBLIC

In a detailed stratigraphic survey, Saunders et al. (1986) observed several sedimentological (e.g., channeling, load features, flame structures, and silt casts) and biological (e.g., deep-water assemblages dominated by pteropod mollusks and deepwater corals) features indicating a deeper-water environment (>100 m in depth) for the base of Mao Formation at the Rio Gurabo section. A taxonomic study of the corals carried out by Cairns and Wells (1987), based on 1590 specimens representing 20 species, suggested an even deeper depositional environment (>200 m in depth) for certain parts of the Mao Formation. Those potentially deep-water environments include localities NMB 15823 and

15827-15029 from stratigraphic horizons between 650 and 750 m in the Rio Gurabo section (Cairns and Wells, 1987, table 1; Saunders et al., 1986, fig. 4). A deep-water origin for those rocks is supported by the occurrence of *Pourtalocyathus hispidus* (Pourtalès 1878), a deep-water coral species in the modern Caribbean (Cairns and Wells, 1987), and the composition of the mollusk assemblages (Saunders et al., 1982).

Terebratulid brachiopods mentioned briefly by Saunders et al. (1986) were later identified by Logan (1987) as Tichosina? lecta, a species described by Cooper (1979) based on Eocene material from Trinidad. The foramen shape and size of *Tichosina? lecta* is similar to those reported for other species of Tichosina, including the Eocene Tichosina? trinitatensis and a number of modern species including T. pillsburyae, T. obesa, T. labiate, T. expansa, and T.? bartletti (Cooper, 1977, 1979). The uniplicate anterior margin of *T.? lecta* is also a common feature in several modern species of Tichosina (Rojas et al., 2015). Logan (1987) also described Tichosina fragments gathered at the upper part of the Mao Adentro Limestone Member on the Rio Cana section, locality NMB 16884 (Saunders et al., 1987; Schultz and Budd 2008). This locality contained a diverse coral assemblage including genera recorded in modern deep-water habitats of the Caribbean (e.g., Madracis). Coral species common in modern shallow waters were also present in the upper part of the Mao Adentro Limestone member in the Rio Cana section (Budd and Klaus, 2008). However, only a few of those coral remains appear to have been preserved in life position (Saunders et al., 1982). Complete specimens of T.? lecta showing post-mortem distortion and fracturing were also reported at the very bottom of the Mao Formation in the Rio Gurabo section, locality TU 1300 (Saunders et al., 1987; Logan, 1987). Saunders et al. (1986) identified an increase in the richness of planktonic foraminifera from the upper Gurabo Formation through the lower Mao Formation in the Rio Gurabo section. The latter include the stratigraphic horizons with the Tichosina remains mentioned above.

McNeill et al. (2008) re-interpreted the

Mao Formation as a progressive shallowing sequence based on a regional interpretation of the Cibao Basin and the Bahamas, with both regions experiencing shallowing during the late Pliocene. The water-depth interpretation of the Mao Formation by McNeill et al. (2008) was established on the Rio Cana section, which is located more than 15 km west of the Rio Gurabo section. The Mao Formation at the Rio Gurabo section differs in terms of thickness of sediments, proportion of different facies, and paleontological records when compared to the Rio Cana section. The Mao Adentro Limestone sensu Saunders et al. (1982) in the Rio Cana section comprises ~350 m of thick bedded coral limestone and massive limestones with the bivalves Ostrea, Lithophaga, and shallow-water reef forming corals. In contrast, this member has not been identified satisfactorily in the Rio Gurabo section (see Saunders et al., 1982, fig. 3; Saunders et al., 1986, fig. 4). The potential sediments corresponding to this member in the Rio Gurabo section are ~100 m thick sequence of intercalated limestones and siltstones obscured by covered outcrop intervals and faults. As indicated above, those deposits are characterized by deep-water taxa including *Limopsis* sp., and the corals Pourtalocyathus hispidus, Dendrophyllia cornucopia, and Trochocyathus duncani (Saunders et al., 1982, fig. 2; Cairns and Wells, 1987). The stony cup coral Dendrophyllia may not be a member of the deep-coral communities because its bathymetric distribution is generally lower (Emig pers. comm., 2018). The genus Trochocyathus was clustered together with Tichosina in the network analysis of the modern Caribbean data (Fig. 4).

Based on these observations, the sediments from the Rio Gurabo section bearing *Tichosina* were likely deposited in deep-water settings (>100 m) below the photic zone. Our re-evaluation of the paleontological data of the Mao Formation suggests that *Tichosina* has likely been associated with deep-water coral communities since at least the Pliocene. Large terebratulids of the genus *Tichosina* have been reported in post-Paleocene sediments across the Caribbean Basin (see Harper, 2002). Typically, the setting of those records is interpreted

as a fore reef, an extension of the shallow-water coral environments (Harper et al., 1995; Harper et al., 1997; Harper et al., 2002). However, depth estimations for those deposits frequently exceed the deepest limit of the bathymetric distribution of modern zooxanthellate corals (Englebert et al., 2014). Fossil assemblages with *Tichosina* species, such as those recently described by Donovan et al. (2015), may represent depth-water benthic communities developed below the photic zone.

#### **CONCLUSIONS**

Our observations from multiple sites on the Caribbean continental shelf of Colombian suggest that T. plicata inhabit deep-water coral habitats dominated by known and potential habitat-forming corals (i.e., Madracis spp., Anomocora fecunda). Furthermore, compiled data from the R/V Pillsbury sampling program indicates that *Tichosina* species may have close ecological ties with deep-water corals as they were recovered consistently across trawl samples structure-forming (i.e., Madrepora oculata) and potential habitat-forming corals (e.g., Acanella sp., Diodogorgia sp., Riisea paniculata, Swiftia exserta, Parantipathes sp., Coenosmilia arbuscula, Madracis asperula, M. formosa, Nicella americana, N. guadalupensis, Neospongodes sp., Oxysmilia rotundifolia, Thalamophyllia riisei, Eguchipsammia cornucopia, and Polymyces fragilis). Furthermore, networks analysis points to Parantipathes and Polymyces as important habitatforming corals for Tichosina growth. Deep-water benthic communities dominated by solitary, attached, and free-living corals appear to provide the optimum structural habitat for *Tichosina* spp. growth across the continental shelves of the modern Caribbean Sea. A re-evaluation of the paleontological data from carbonates of the Mao Adentro Formation (Rio Gurabo section) in the Dominican Republic, suggests that Tichosina brachiopods may have been tied to deep-water coral habitats since at least the Pliocene (~3 Ma). Understanding to what extent the brachiopod Tichosina is linked to deep-coral habitats through the modern Caribbean Sea has several implications for both Tertiary geology and paleoecology in the region. Although the distinction of deep-water communities in the Cenozoic rock record is challenging, this potential brachiopod-anthozoan association could be useful to distinguish those communities in the Cenozoic rock record and thus it would enhance our understanding of the sealevel history in the Caribbean basin.

#### **ACKNOWLEDGMENTS**

We thank Nadia Santodomingo (Natural History Museum, London) for identification of corals and Miguel Martelo and Erika Montoya for support at the Marine Natural History Museum of Colombia. AR thanks Roger W. Portell (Florida Museum of Natural History) for advice and support. We thank Gustav Paulay (Florida Museum of Natural History), Liang Mao (Department of Geography, University of Florida), and Christian C. Emig (BrachNet) for their comments on earlier drafts of this report. AR is grateful to Elizabeth Sandoval for her continuous support. AR was supported by a Jon L. and Beverly A. Thompson Endowment Fund. MARCORAL project was funded by Invemar -Colciencias (Project Code 2115-09-16649). Coral database from the Pillsbury expedition were provided with the permission of the Smithsonian Natural Museum of Natural History, the Marine Invertebrate Museum, and the Rosenstiel School of Marine and Atmospheric Science (MIN-RSMAS). IHA thanks Nancy Voss for providing the MIM-RSMAS data. IHA is supported by a Fundayacucho PhD grant E-223-85-2012-2, and Campus France grant 796045K.

#### LITERATURE CITED

- Álvarez F., C. C. Emig, and J. Tréguier. 2017. Brachiopodes actuels: historique et révision de la collection D.-P. Œhlert (Laval). Carnets de Géologie, Madrid, CG2017\_B02, 386 p.
- Andrade, C. A. 2003. Evidence for an eastward flow along the Central and South American Caribbean Coast. Journal of Geophysical Research 108(6):1–11. <doi: 10.1029/2002JC001549>.

- Asgaard, U., and N. Stentoft, 1984. Recent micromorph brachiopods from Barbados: palaeoecological and evolutionary implications. Geobios 17(8):29–37.
- Besag, J. 1977. Contribution to the discussion on Dr. Ripley's paper. Journals of the Royal Statistical Society, 39:193–195.
- Cairns, S. 2007. Deep-water corals: an overview with special reference to diversity and distribution of deep-water scleractinian corals. Bulletin of Marine Science 81(3):311–322.
- Cairns, S., and J. W. Wells. 1987. Neogene paleontology in the northern Dominican Republic, 5. The suborders Caryophylliina and Dendrophylliina (Anthozoa, Scleractinia). Bulletins of American Paleontology 93(328):23–43.
- Calatayud, J., R. Bernardo, M. Neuman, A. Rojas, and M. Rosvall. 2019. Exploring the solution landscape enables more reliable network community detection. Physical Review E 100(5):052308. <doi:10.1103/PhysRevE.100.052308>.
- Cooper, A. 1977. Brachiopods from the Caribbean Sea, Gulf of Mexico and Adjacent Waters. University of Miami Press, Miami, 211 p.
- Cooper, G. A. 1979. Tertiary and Cretaceous brachiopods from Cuba and Caribbean. Smithsonian Contributions to Paleobiology 37:1–45.
- Costello, M. J., Z. Basher, L. McLeod, I. Asaad, S. Claus, L. Vandepitte, M. Yasuhara, H. Gislason, M. Edwards, W. Appeltans, H. Enevoldsen, G. J. Edgar, P. Miloslavich, S. De Monte, I. S. Pinto, D. Obura, and A. E. Bates. 2017. Methods for the Study of Marine Biodiversity. Pp. 129–163 *in* M. Walters and R. J. Scholes, eds. The GEO Handbook on Biodiversity Observation Networks. Springer International Publishing, Cham.
- Davies, A. J., and J. M. Guinotte. 2011. Global habitat suitability for framework-forming cold-water corals. PLoS ONE 6(4):e18483. <doi:10.1371/journal.pone.0018483>.
- Donovan, S. K., and D. A. T. Harper. 1998. Diving deep on a Pleistocene reef in eastern Jamaica.

- Geology Today 14(1):26–30. <doi:10.1046/j.1365-2451.1998.014001026.x>.
- Donovan, S. K., D.A.T. Harper, and R.W. Portell. 2015. In deep water: a crinoid-brachiopod association in the Upper Oligocene of Antigua, West Indies. Lethaia 48(3):291–298. <doi:10.1111/let.12105>.
- Emig, C. C. 1987. Offshore brachiopods investigated by submersible. Journal of Experimental Marine Biology and Ecology 108(3):261–273. <doi:10.1016/0022-0981(87)90089-X>.
- Emig, C. C. 1989. Distributional patterns along the Mediterranean continental margin (upper bathyal) using *Gryphus vitreus* (Brachiopoda) densities. Palaeogeography, Palaeoclimatology, Palaeoecology 71:253–256. <doi:10.1016/0031-0182(89)90053-9>.
- Emig, C. C. 1997. Bathyal zones on the Mediterranean continental slope: an attempt. Publicaciones Especiales. Instituto Español de Oceanografía 23:23–33.
- Emig, C. C., M. A. Bitner, and F. Alvarez. 2013. Phylum Brachiopoda. Zootaxa 3703(1):75–78. <doi:10.11646/zootaxa.3703.1.15>.
- Englebert, N., P. Bongaerts, P. Muir, K. B. Hay, and O. Hoegh-Guldberg. 2014. Deepest zooxanthellate corals of the Great Barrier Reef and Coral Sea. Marine Biodiversity 45(1):1–2. <doi:10.1007/s12526-014-0221-8>.
- Harper, D. A. T. 2002. Fossil Brachiopoda of the Caribbean region: biodiversity patterns. Pp. 139–148 *in* T. Jackson ed., Caribbean Geology into the Third Millennium. Transactions of the 15th Caribbean Geological Conference. The Press, University of the West Indies, Kingston, Jamaica.
- Harper, D. A. T., and S. K. Donovan. 2007. Fossil brachiopods from the Pleistocene of the Antilles. Scripta Geologica 135:213–239.
- Harper, D. A. T., and R. K. Pickerill. 2008. Generation of brachiopod-dominated shell beds in the Miocene rocks of Carriacou, Lesser Antilles. Geological Journal 43(5):573–581. <doi: doi.org/10.1002/gj.1131>.
- Harper, D. A. T., E. N. Doyle, and S. K. Donovan. 1995. Palaeoecology and palaeobathymetry of

- Pleistocene brachiopods from the Manchioneal Formation of Jamaica. Proceedings of the Geologists Association 106(3):219–227. <doi:10.1016/S0016-7878(08)80025-9>.
- Harper, D. A. T., S. K. Donovan, and R. W. Portell. 1997. The brachiopods *Tichosina* and *Terebratulina* from the Miocene of Jamaica. Caribbean Journal of Science 33:117–119.
- Hernández-Ávila, I. 2014. Patterns of deep-water coral diversity in the Caribbean Basin and adjacent southern waters: an approach based on records from the R/V Pillsbury Expeditions. PLoS ONE 9(3):e92834. <doi:10.1371/journal.pone.0092834>.
- Hernández-Ávila, I., E. Guerra-Castro, C. Bracho, M. Rada, F. A. Ocaña, and D. Pech. 2018. Variation in species diversity of deep-water megafauna assemblages in the Caribbean across depth and ecoregions. PLoS ONE 13(8): e0201269. <doi:10.1371/journal.pone.0201269>.
- Jackson, J. B. C., T. F. Goreau, and W. D. Hartman. 1971. Recent brachiopod-coralline sponge communities and their paleoecological significance. Science 173(3997):623–625. <doi:10.1126/science.173.3997.623>.
- Kahng, S. E., J. R. Garcia-Sais, H. L. Spalding, E. Brokovich, D. Wagner, E. Weil, L. Hinderstein, and R. J. Toonen, 2010. Community ecology of mesophotic coral reef ecosystems. Coral Reefs 29(2):255–275. <doi:10.1007/s00338-010-0593-6>.
- Kano, A., T. G. Ferdelman, T. Williams, J. P. Henriet, T. Ishikawa, N. Kawagoe, C. Takashima, Y. Kakizaki, K. Abe, S. Sakai, E. L. Browning, X. Li, and Integrated Ocean Drilling Program Expedition 307 Scientists. 2007. Age constraints on the origin and growth history of a deep-water coral mound in the northeast Atlantic drilled during Integrated Ocean Drilling Program Expedition 307. Geology 35(11):1051–1054. doi:10.1130/G23917A.1
- Kruskal, J. 1964. Nonmetric multidimensional scaling: a numerical method. Psychometrika 29(2):115–129. <doi:10.1007/BF02289694>.

- Logan, A. 1975. Ecological observations on the recent articulate brachiopod *Argyrotheca bermudana* Dall from the Bermuda Platform. Bulletin of Marine Science 25:186–204.
- Logan, A. 1979. The recent Brachiopoda of the Mediterranean Sea. Bulletin de l'Institut Océanographique de Monaco 72(1434b):1–112.
- Logan, A. 1987. The phylum Brachiopoda. Bulletins of American Paleontology 93(328):44–52.
- Logan, A. 1990. Recent Brachiopoda from the Snellius and Luymes expeditions to the Surinam-Guyana shelf, Bonaire-Curacao, and Saba bank, Caribbean Sea, 1966 and 1969-1972. Zoologische Mededelingen 63:123–136.
- Lutz, S., and R. Ginsburg. 2007. State of deep coral ecosystems in the Caribbean Region: Puerto Rico and the U.S. Virgin Islands. Pp. 307–365 *in* E. Lumsden, T. Hourigan, A. Bruckner, and G. Dorr, eds. The State of Deep Coral Ecosystems of the United States, National Oceanic and Atmospheric Administration, Silver Spring.
- McNeill, D. F., J. S. Klaus, C. C. Evans, and A. F. Budd. 2008. An overview of the regional geology and stratigraphy of the Neogene deposits of the Cibao Valley, Dominican Republic. Pp. 21–45 *in* R. H. Nehm and A. F. Budd, eds. Evolutionary Stasis and Change in the Dominican Republic Neogene. Springer Netherlands, Dordrecht.
- Mullins, H. T., A. C. Neumann, R. J. Wilber, and M. R. Boardman. 1980. Nodular carbonate sediment on Bahamian slopes: possible precursors to nodular limestones. Journal of Sedimentary Research 50(1):117–131. <doi:10.1306/212F797D-2B24-11D7-8648000102C1865D>.
- Nehm, R. H., and A. F. Budd. 2008. Palaeobiological research in the Cibao Valley of the northern Dominican Republic. Pp. 1–19 *in* R. H. Nehm and A. F. Budd, eds. Evolutionary stasis and change in the Dominican Republic Neogene. Springer Netherlands, Dordrecht.
- Oksanen, J., F. Blanchet, M. Friendly, R. Kindt, P. Legendre, D. McGlinn, P. Minchin, R.

- O'Hara, G. Simpson, P. Solymos, M. Stevens, E. Szoecs, and H. Wagner. 2018. *Vegan: Community Ecology Package*.
- Oyen, C., R. Fountain, R. W. Portell, and G. McClellan. 2000. Occurrence of Plio-Pleistocene phosphatic macro-invertebrates from the Upper West Florida Slope, Eastern Gulf of Mexico. Bulletin of the Florida Museum of Natural History 42(5):219–252.
- Poos, M. S., and D. A. Jackson. 2012. Addressing the removal of rare species in multivariate bioassessments: the impact of methodological choices. Ecological Indicators 18:82–90. <doi:10.1016/j.ecolind.2011.10.008>.
- Ramirez-Llodra, E., A. Brandt, R. Danovaro, B. De Mol, E. Escobar, C. R. German, L. A. Levin, P. Martinez-Arbizu, L. Menot, P. Buhl-Mortensen, B. E. Narayanaswamy, C. R. Smith, D. P. Tittensor, P. A. Tyler, A. Vanreusel, and M. Vecchione. 2010. Deep, diverse and definitely different: unique attributes of the world's largest ecosystem. Biogeosciences 7:2851–2899. <doi:10.5194/bg-7-2851-2010>.
- Rangel-Buitrago, N., and J. Idárraga-García. 2010. Geología general, morfología submarina y facies sedimentarias en el margen continental y fondos oceánicos del Caribe colombiano. Pp. 29–52 *in* INVEMAR. Biodiversidad Del Margen Continental Del Caribe Colombiano. Marquillas S.A., Santa Marta, Colombia.
- Reyes, J., N. Santodomingo, A. Gracia, G. Borrero-Pérez, G. Navas, L. M. Mejía-Ladino, A. Bermúdez, and M. Benavides. 2005. Southern Caribbean azooxanthellate coral communities off Colombia. Pp. 309–330 *in* A. Freiwald and J. M. Roberts, eds. Cold-Water Corals and Ecosystems. Springer-Verlag, Berlin/ Heidelberg.
- Richardson, J. R. 1997. Ecology of articulate brachiopods. Pp. 441–462 *in* R. L. Kaesler ed. Treatise on Invertebrate Paleontology. Part H, revised. Brachiopoda Vol. 1. Geological Society of America and University of Kansas, Boulder, Colorado, and Lawrence, Kansas.
- Roberts, J. M. 2006. Reefs of the deep: the biology and geology of cold-water coral ecosystems.

- Science 312(5773):543–547. <doi:10.1126/science.1119861>.
- Rojas, A., A. Gracia, and P. Patarroyo. 2015. Brachiopods from off the San Bernardo Archipelago (Colombian Caribbean), with comments on specific synonymies in *Tichosina*, Cooper, 1977. Zootaxa 3914(1):55–63. <doi:10.11646/zootaxa.3914.1.3>.
- Rojas, A., and M. R. Sandy. 2019. Early Cretaceous (Valanginian) brachiopods from the Rosablanca Formation, Colombia, South America: biostratigraphic significance and paleogeographic implications. Cretaceous Research 96:184–195. <doi: 10.1016/j.cretres.2018.12.011>.
- Rosso, A., A. Vertino, I. Di Geronimo, R. Sanfilippo, F. Sciuto, R. Di Geronimo, D. Violanti, C. Corselli, M. Taviani, F. Mastrototaro, and A. Tursi. 2010. Hard- and soft-bottom thanatofacies from the Santa Maria di Leuca deep-water coral province, Mediterranean. Deep Sea Research Part II: Topical Studies in Oceanography 57:360–379. <doi: 10.1016/j. dsr2.2009.08.024>.
- Rosvall, M., D. Axelsson, and C. T. Bergstrom. 2009. The map equation. The European Physical Journal Special Topics 178:13–23. <doi:10.1140/epjst/e2010-01179-1>.
- Sandy,M.R.,andA.Rojas.2019.Paleobiogeographic affinities, biostratigraphic potential, and taxonomy of Cretaceous terebratulide brachiopods from Colombia. Geological Society of America Abstracts with Programs 50(6). <doi:10.1130/abs/2018AM-325121>.
- Santodomingo, N., J. Reyes, A. Gracia, A. Martinez, G. Ojeda, and C. Garcia. 2007. Azooxanthellate *Madracis* coral communities off San Bernardo and Rosario Islands (Colombian Caribbean). Pp. 273–287 *in* Conservation and Adaptive Management of Seamount and Deep-Sea Coral Ecosystems, Vol. 81. University of Miami Rosenstiel School of Marine and Atmospheric Science, Miami, Florida.
- Santodomingo, N., J. Reyes, P. Flórez, I. C. Chacón-Gómez, L. P. van Ofwegen, and B. W.

- Hoeksema. 2013. Diversity and distribution of azooxanthellate corals in the Colombian Caribbean. Marine Biodiversity 43:7–22. <doi:10.1007/s12526-012-0131-6>.
- Saunders, J. B., P. Jung, and B. Biju-Duval. 1986. Neogene palaeontology of the northern Dominican Republic, 1, field surveys, lithology, environment, and age. Bulletins of American Paleontology 89:1–79.
- Saunders, J. B., P. Jung, J. Geister, and B. Biju-Duval. 1982. The Neogene of the south bank of the Cibao Valley, Dominican Republic: a stratigraphic study. Transcripts of Caribbean Geological Conference 9(2):151–160.
- Schemm-Gregory, M., A. Rojas-Briceño, P. Patarroyo, and C. Jaramillo. 2012. First report of *Hadrosia* Cooper, 1983 in South America and its biostratigraphical and palaeobiogeographical implications. Cretaceous Research 34:257–267. <doi:10.1016/j.cretres.2011.11.005>.
- Schultz, H. A., and A. F. Budd. 2008. Neogene evolution of the reef coral species complex *Montastraea* "cavernosa". Pp. 147–170 in R. H. Nehm and A. F. Budd, eds. Evolutionary stasis and change in the Dominican Republic Neogene, Vol. 30. Springer Netherlands, Dordrecht.
- Squires, D. F. 1964. Fossil coral thickets in Wairarapa, New Zealand. Journal of Paleontology 38(5):904–915.
- Teichert, C. 1958. Cold and deep-water coral banks. Bulletin American Association of Petroleum Geologists 42(5):1064–1082. <doi:10.1306/0BDA5AF5-16BD-11D7-8645000102C1865D>.
- Tursi, A., F. Mastrototaro, A. Matarrese, P. Maiorano, and G. D'onghia. 2004. Biodiversity of the white coral reefs in the Ionian Sea (Central Mediterranean). Chemistry and Ecology 20:107–116. <doi:0.1080/027575403 10001629170>.
- Zumwalt, G., and T. Delaca. 1980. Utilization of brachiopod feeding currents by epizoic foraminifera. Journal of Paleontology 54(2):477–4