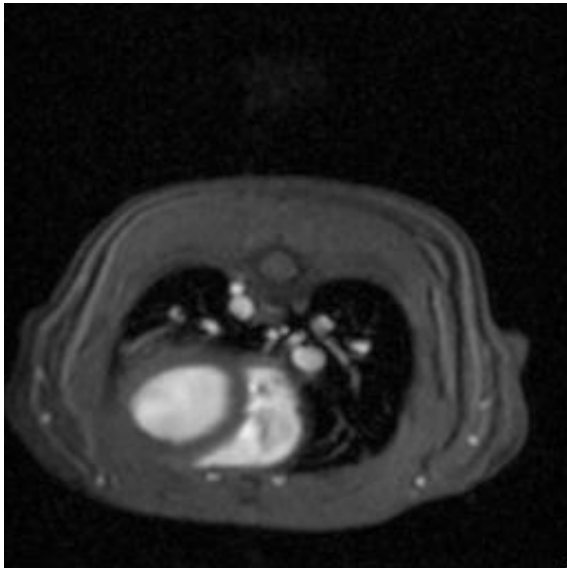


1960: Creation on Nuclear Magnetic Resonance (NMR)

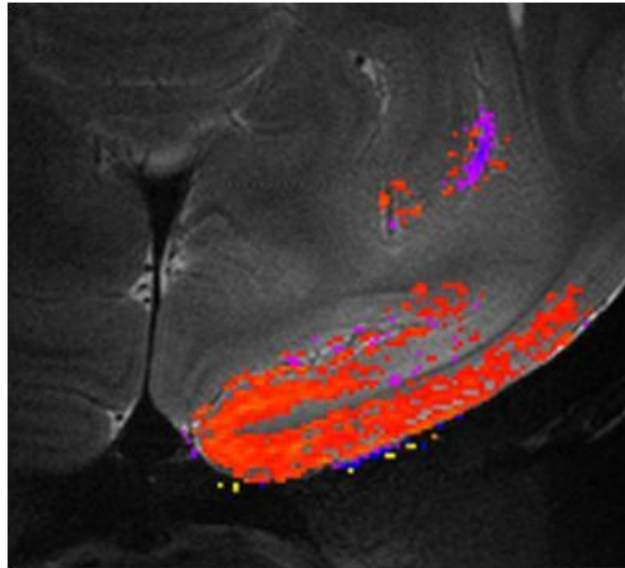


Nowadays: Leader in Pre-clinical Imaging (MRI)

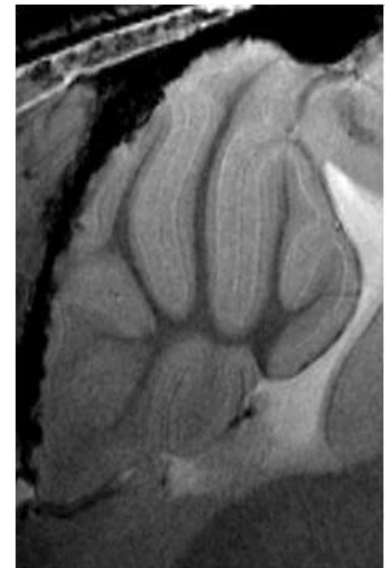
- Non-invasive molecular imaging techniques



Cardiology



Brain functions



Anatomy

Bruker Corporation Divisions



Bruker BioSpin Group



Magnetic Resonance Spectroscopy (MRS)



Preclinical Imaging (PCI)

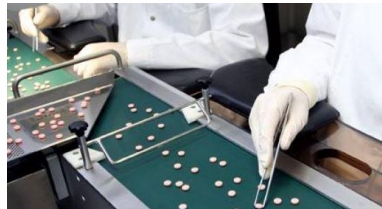


Applied, Industrial and Clinical (AIC)

Bruker CALID Group



Mass spectrometry (Daltonics)



Infrared & Raman spectroscopy (Optics)



CBRNE (Detection)

CALID = Chemical, Applied Markets, Life Science, In-Vitro Diagnostics, Detection

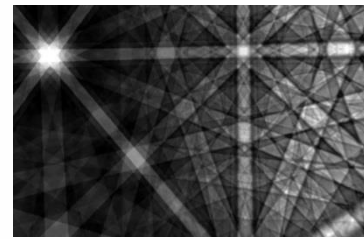
Bruker NANO Group



Bruker Nano Surface



X-ray analysis



Bruker Nano Analytics

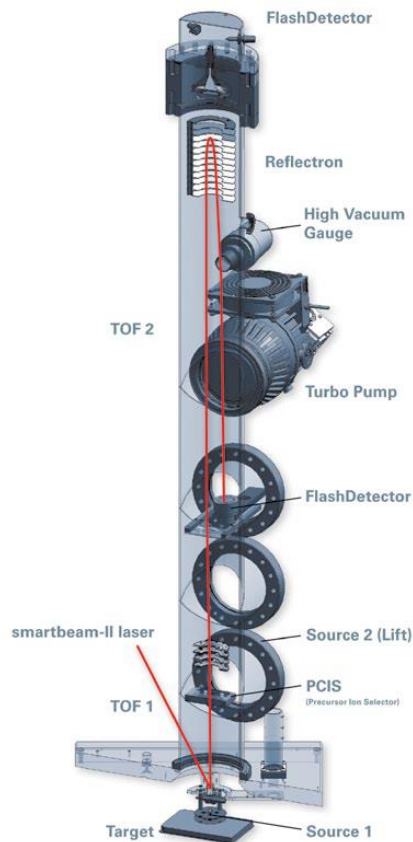
Bruker EST (BEST)



Supercon wire and devices => NMR

BEST: Bruker Energy and Supercon Technologies

Proteomics & Metabolomics Analyses



Tissue Imaging

Top-Down Protein Sequencing

2D-Gels

TLC-MALDI

Polymer Analysis

PTM Analysis



Glycan Analysis

SPRi MALDI Interaction Analysis

FoodOmics

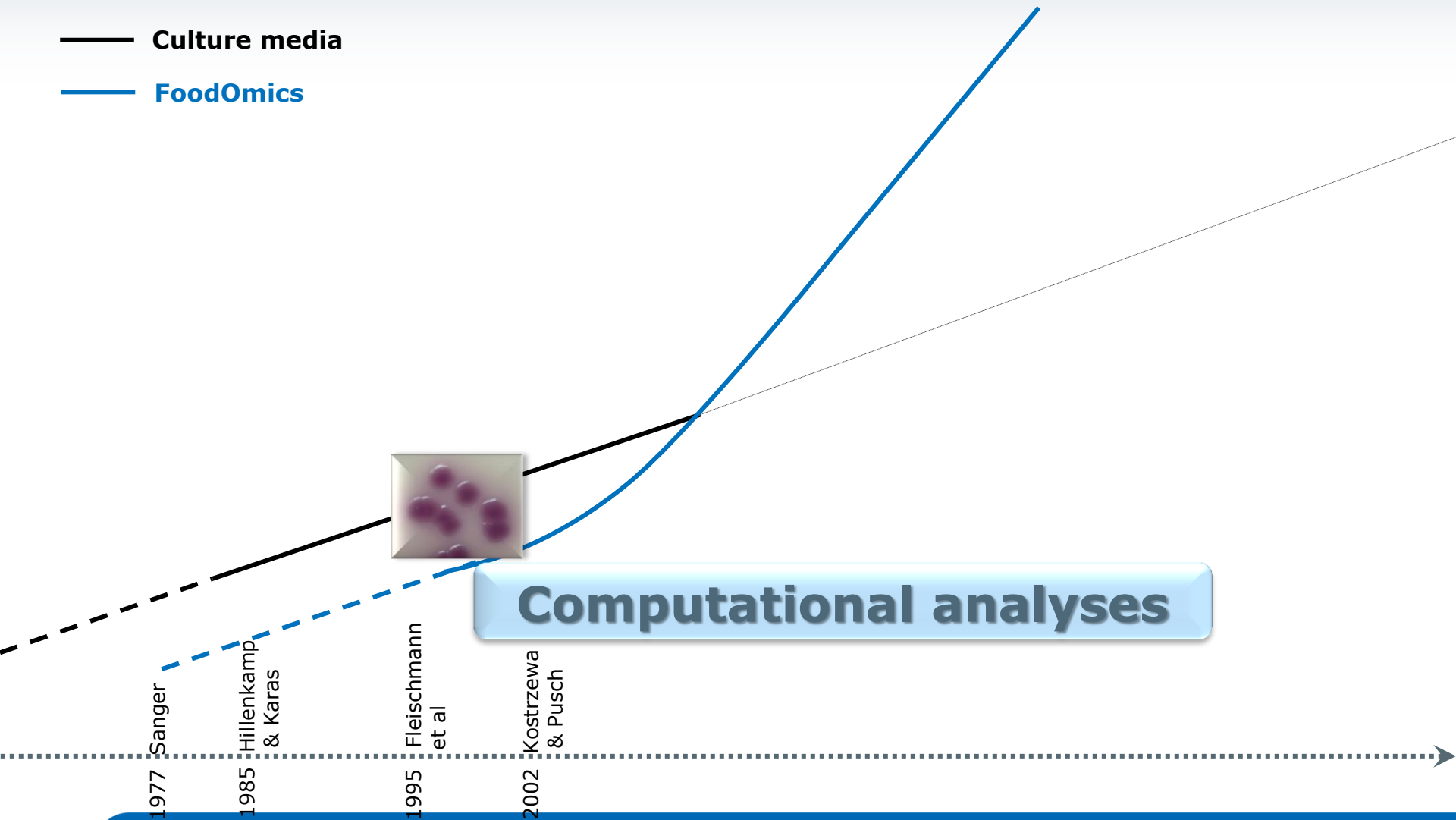
Cifuentes, 2009. J. Chromatogr. A. 1216, 7109

FoodOmics

Cifuentes, 2009. J. Chromatogr. A. 1216, 7109



— Culture media
— FoodOmics



Computational analyses

1977 Sanger

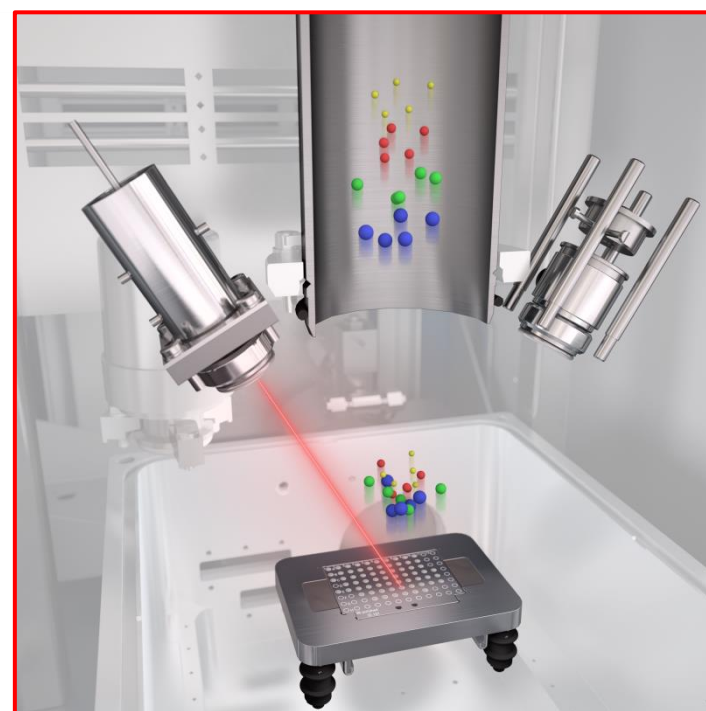
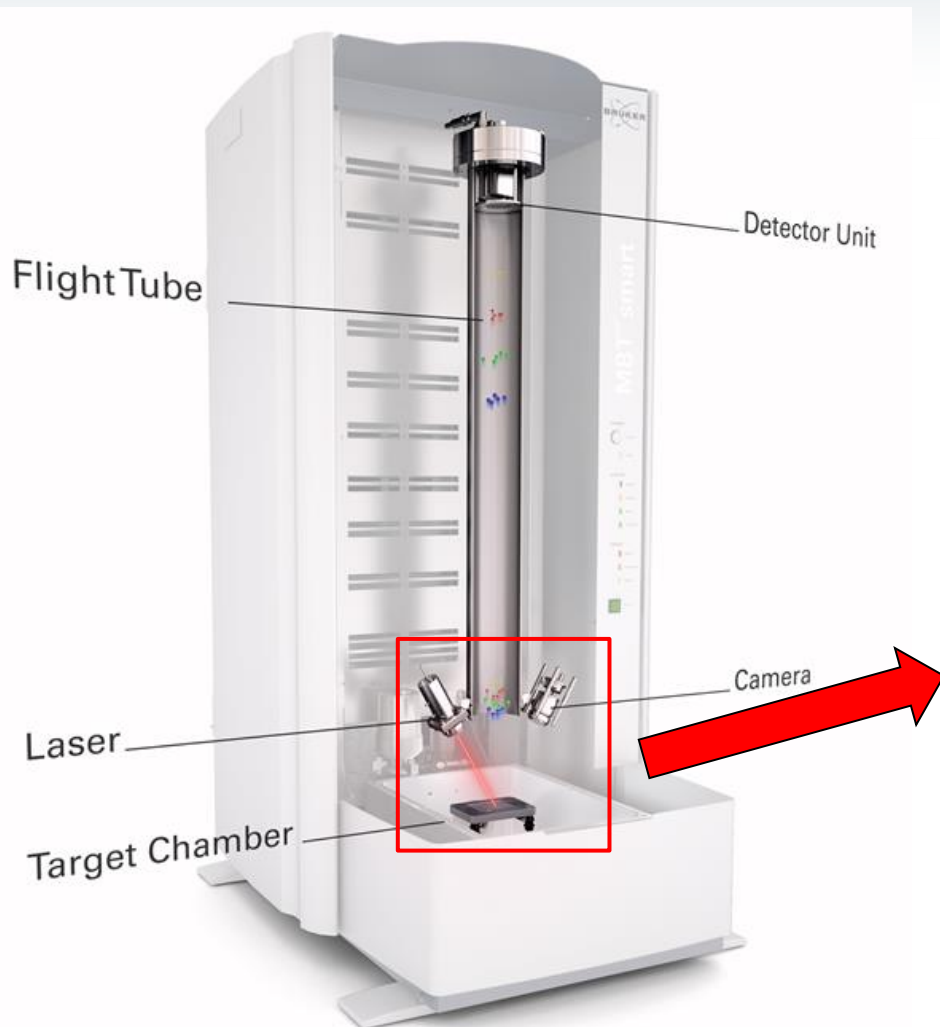
1985 Hillenkamp & Karas

1995 Fleischmann et al

2002 Kostorzewa & Pusch

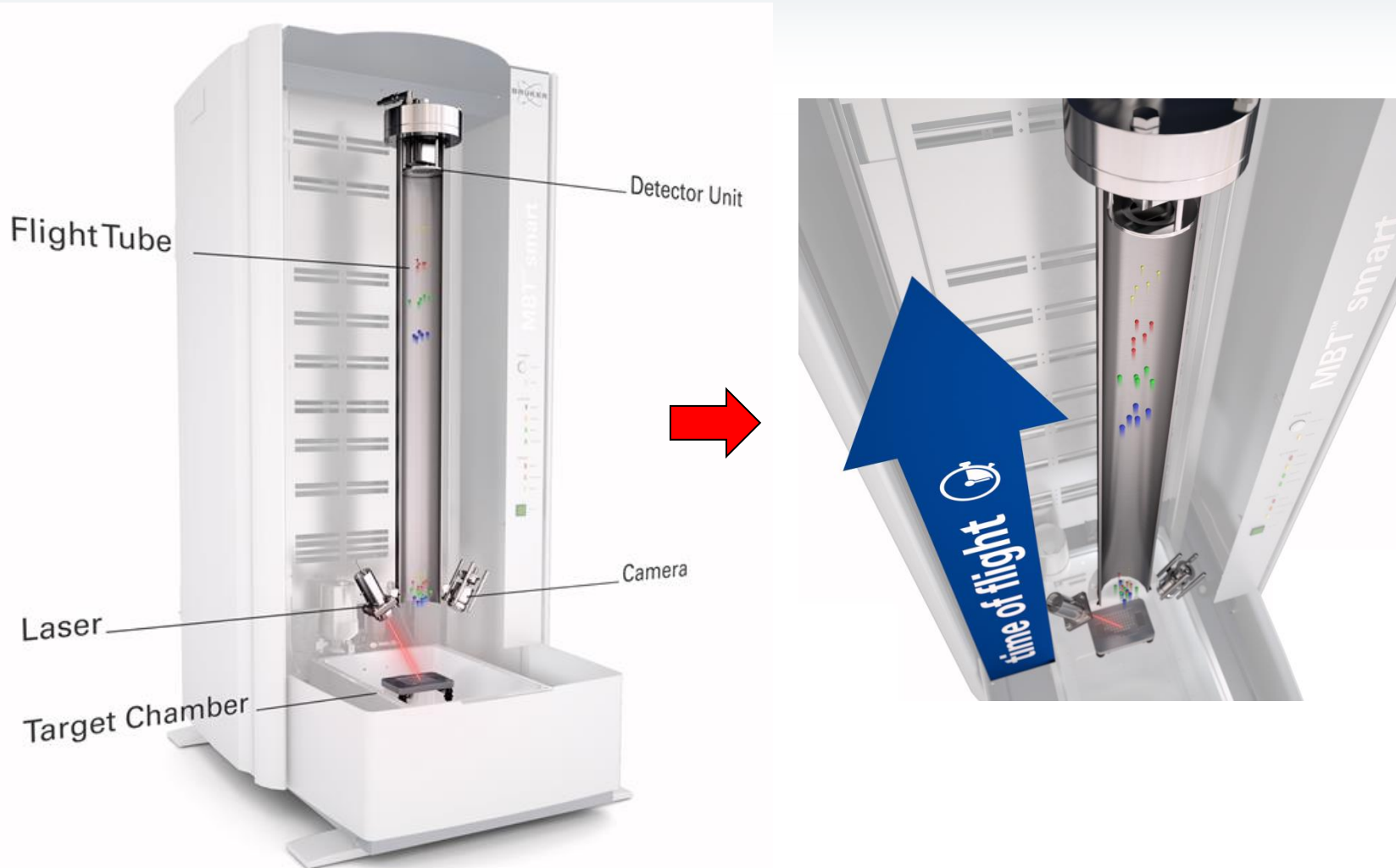
MALDI Biotyper - Basics

Matrix **A**ssisted **L**aser **D**esorption / **I**onization
Time **O**f **F**light Mass Spectrometry



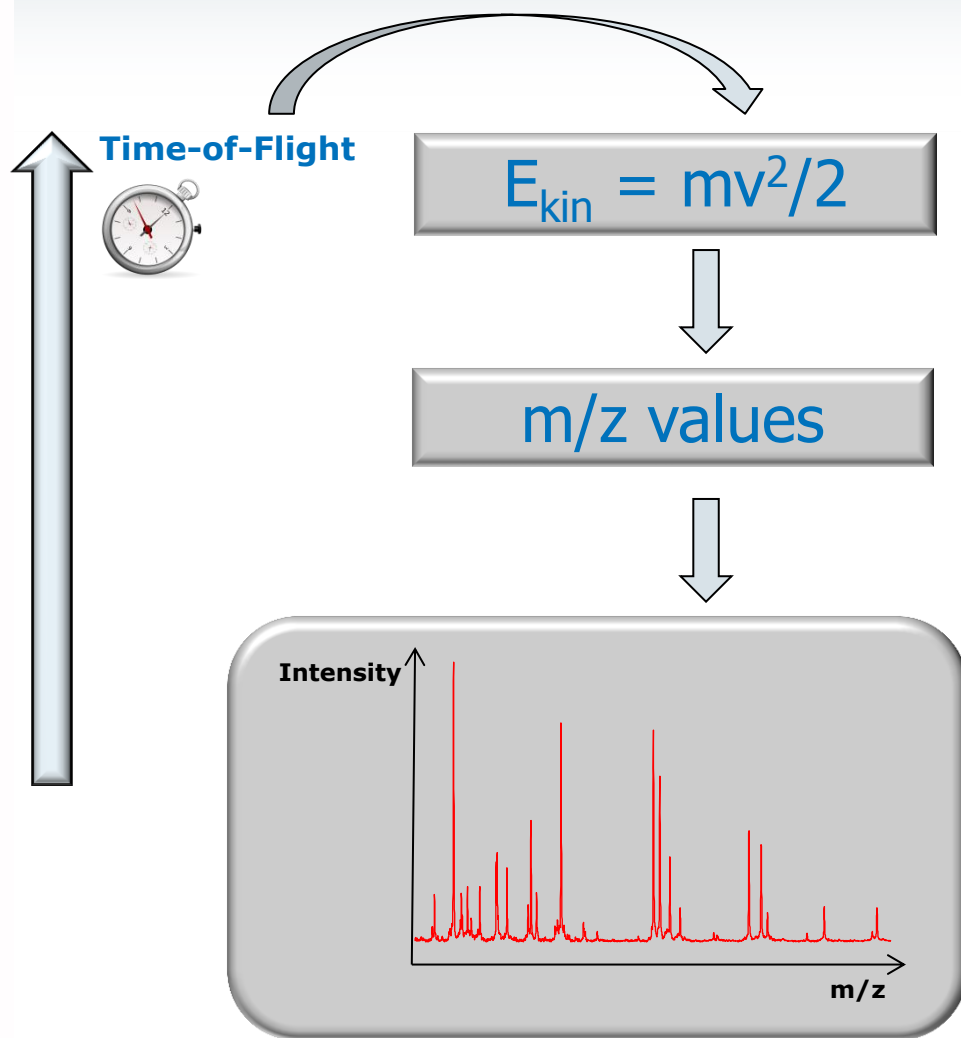
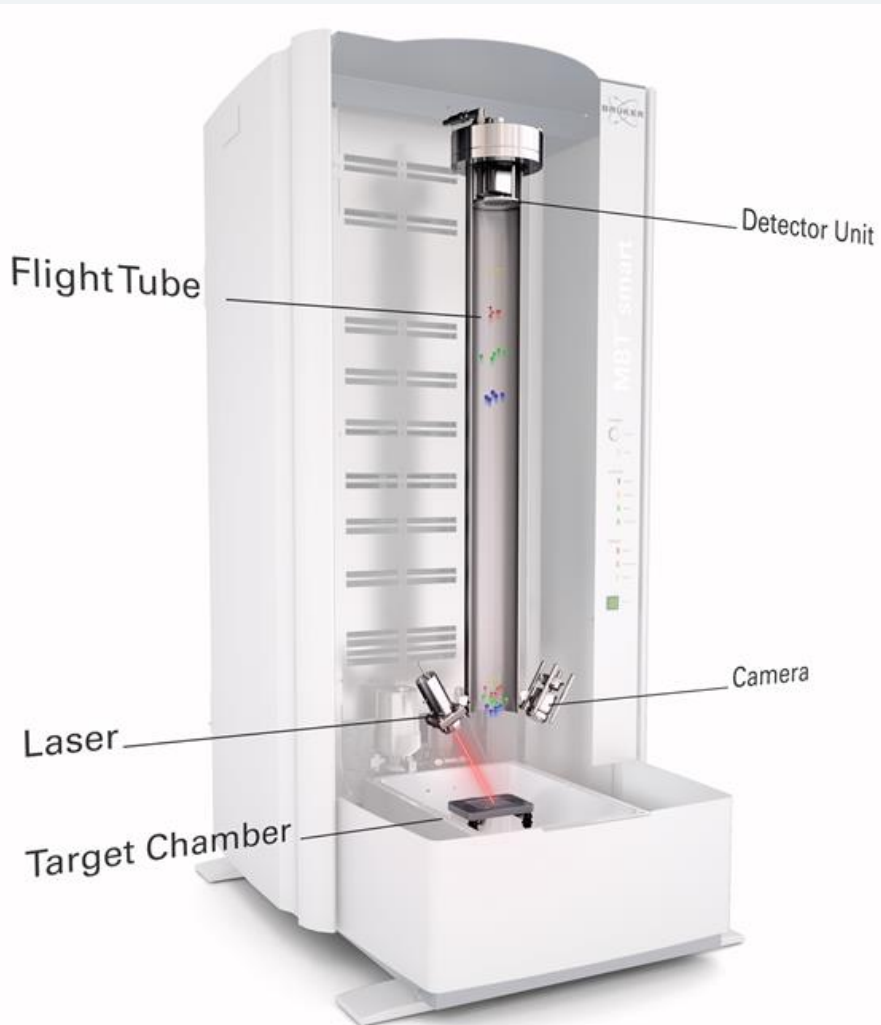
MALDI Biotyper - Basics

Matrix **A**ssisted **L**aser **D**esorption / **I**onization
Time **O**f **F**light Mass Spectrometry



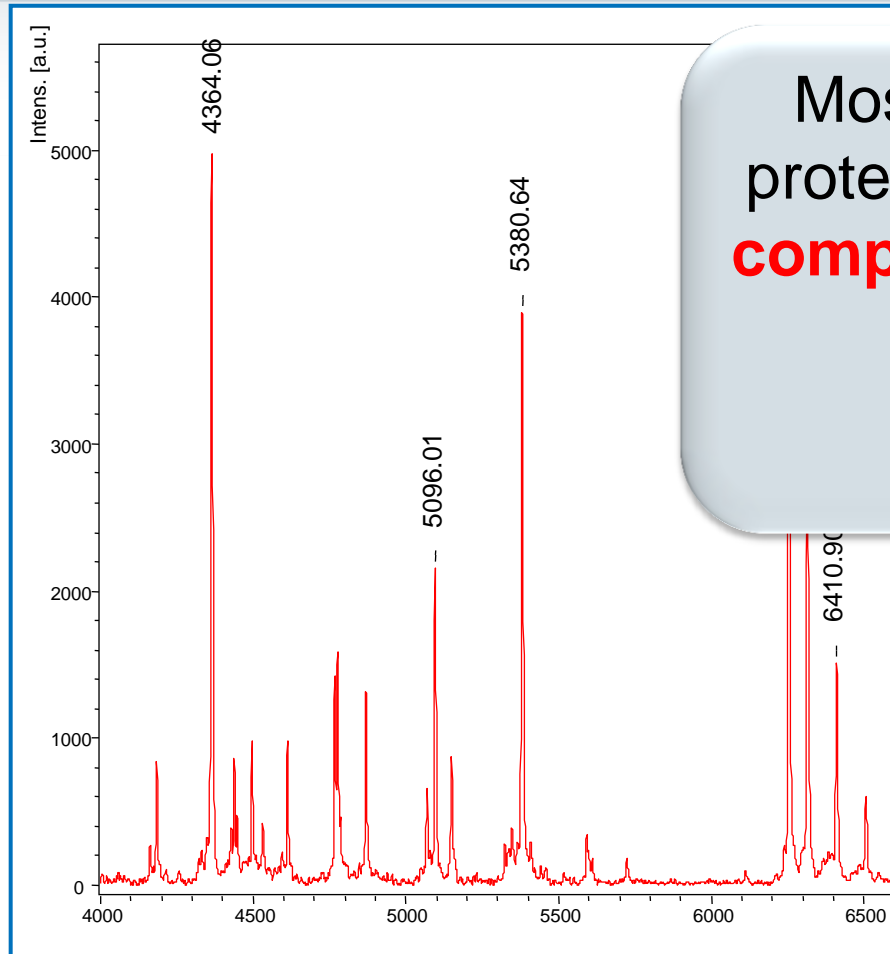
MALDI Biotyper - Basics

Matrix **A**ssisted **L**aser **D**esorption / **I**onization
Time **O**f **F**light Mass Spectrometry



MALDI Biotyper - Basics

Robust identification method, as it relies on the highly abundant proteins



E. coli

Mostly intracellular, hydrophilic proteins and **primarily ribosomal components** or other noncatalytic, structural complexes


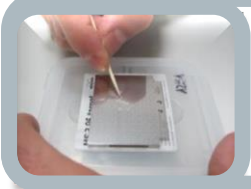


Mass range calibrated
2,000 – 20,000 Da

➤ Routine testing

Easy and short workflow



1 room is enough

-  **1 isolate**
-  **1 spot**
-  **1 drop**
-  **1 clic**

☞ **Easy and short handling**
Direct transfer

1 isolate	Approx 20 sec
48 isolates	Approx 10 min
95 isolates	Approx 18 min

↘ Routine testing

Easy and short workflow




Spectrum acquisition

Rank	Matched Pattern	Score Value
1	Pseudomonas aeruginosa DSM 1117 DSM	2.18
2	Pseudomonas aeruginosa 8147_2CHB	2.17
3	Pseudomonas aeruginosa ATCC 27853 THL	1.98
4	Pseudomonas aeruginosa AD7_08_Pudu FLR	1.94
5	Pseudomonas aeruginosa 19955_1CHB	1.82
6	Pseudomonas aeruginosa DSM 50071THAM	1.78

Unsupervised Main Spectra & Updated Library

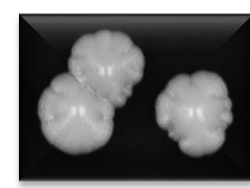
YEAR

Update

- ⌚ **Short time-to-result**, including
- . Bacterial test standard
 - . Drying of matrix
 - . Mass spectrum acquisition

1 isolate	Approx 15 min
48 isolates	Approx 35 min
95 isolates	Approx 60 min

Bacteria, yeasts and molds

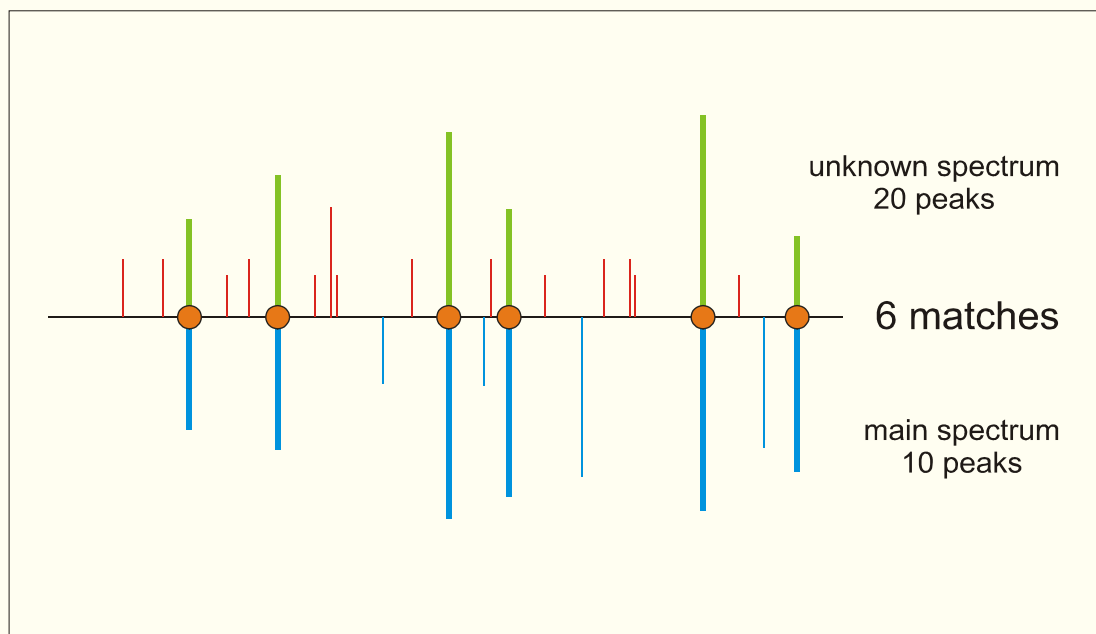


5 to 10% alternative sample preps

MALDI Biotyper - Bioinformatics

Unsupervised **Main Spectra (MSP)** concept

Score base pattern matching



Tested isolate is matched against each **Main Spectrum (MSP)** in the library

Calculation of a matching score based on:

Matches MSP to unknown

% matches of the reference spectrum (e.g. 6/10)

Matches unknown to MSP

% matches of the unknown spectrum (e.g. 6/20 = 3/10)

Correlation of intensities

value of intensity correlation

Range	Interpretation	Symbols	Color
2.000 - 3.000	High Confidence Identification	(+++)	green
1.700 - 1.999	Low Confidence Identification	(+)	yellow
0.000 - 1.699	No Organism Identification Possible	(-)	red

MALDI Biotyper - Bioinformatics

Unsupervised Main Spectra Concept



Rank (Quality)	Matched Pattern	Score Value	NCBI Identifier
1 (+++)	<i>Clostridium perfringens</i> B 1968_NCTC 3110_BOG	2.514	1502
2 (+++)	<i>Clostridium perfringens</i> B 1038_NCTC 4964_BOG	2.454	1502
3 (+++)	<i>Clostridium perfringens</i> B 1971_ATCC 3626_BOG	2.305	1502
4 (++)	<i>Clostridium perfringens</i> A 1037_NCTC 8237_BOG	2.254	37763
5 (++)	<i>Clostridium perfringens</i> D 2150_NCTC 8346_BOG	2.253	107819
6 (++)	<i>Clostridium perfringens</i> C 1041_NCTC 10720_BOG	2.11	79668
7 (-)	<i>Comamonas testosteroni</i> DSM 50244 HAM	1.308	285
8 (-)	Listeria grayi murrei DSM 20596 DSM	1.262	1641
9 (-)	Bacillus atrophaeus DSM 675 DSM	1.163	1452
10 (-)	<i>Clostridium beijerinckii</i> 1072_ATCC 25752_BOG	1.136	1520

MALDI Biotyper RUO library

Updates since 2010



Clinical, environment, industrial, and veterinary and isolates provided by collaborating partners, round robin strains and strains from isolates from accredited ISO 9001:2008 certified strain collections

rDNA and relevant housekeeping genes sequencing According to molecular taxonomy

	Spring 2010	^	Sep 2010	^	Jan 2011	^	Nov 2011	^	Aug 2012	^	Jan 2014	^	July 2015	^	April 2016
N° of MSP in total	3476		3740		3995		4110		4613		5627		5989		6903
N° of genus	315		318		368		341		364		390		410		424
N° of species	1850		1947		2012		2026		2185		2297		2371		2461
N° of total added MSP		336		319		147		515		1020		377		938	
N° of total deleted MSP		-72		-64		-32		-12		-3 (-3)		-15		-24	
N° of MSP for new genus/species entries		111		42		32		151		220		106		161	
N° of MSP for additional species entries		224		231		102		359		797		261		766	
N° of MSP replaced		1		46		13		5		3		10		11	
Further overall improvements	Continuous nomenclature updates according International Journal of Systematic and Evolutionary Microbiology (IJSEM) referenced at www.bacterio.net (LPSN = List of prokaryotic names with standing in nomenclature).														

MALDI Biotyper

RUO Reference library Bacteria & Yeasts



Bruker MBT Compass Library
(version April 2017)

7,311 MSPs
2,509 different species

**Every Microorganism
could be of Relevance**

Abiotrophia defective **Acetobacter** *aceti* **Acetobacter** *cerevisiae* **Acetobacter** *xyloxydans* **Acidaminococcus** *fermentans* **Acidaminococcus** *fermentans* **Acinetobacter** *bouvetii* **Acinetobacter** *calcoaceticus* **Acinetobacter** *gomeri* **Acinetobacter** *schindleri* **Acinetobacter** *sp* **Acinetobacter** *tandoii* **Acinetobacter** *sp* **Actinobacillus** *ureae* **Actinobaculum** *massiliense* **Actinobaculum** *schaalii* **Actinomyces** *denticolens* **Actinomyces** *europaeus* **Actinomyces** *funkei* **Actinomyces** *ge* **Actinomyces** *nasicola* **Actinomyces** *neuui* **Actinomyces** *odontolyticus* **Actinomyces** *viscosus* **Actinomyces** *weissii* **Adlercreutzia** *equolifaciens* **Advenella** *ince* **Aeromonas** *encheleia* **Aeromonas** *enteropelogenes* **Aeromonas** *eucr* **Aeromonas** *sobria* **Aeromonas** *sp[2]* **Aeromonas** *veronii* **Afiopia** *broomeae* **Afiopia** *felis* **Agromyces** *humatus* **Agromyces** *italicus* **Agromyces** *lapidis* **Agromyces** *mediolanus* **Agromyces** *contaminans* **Alicyclobacillus** *cycloheptanicus* **Alicyclobacillus** *fastidiosus* **Alicyclobacillus** *vulcanalis* **Aliivibrio** *fischeri* **Alishewanella** *fetalis* **Alistipes** *fingoldii* **Alisiphaea** *coloradensis* **Amycolatopsis** *fastidiosa* **Amycolatopsis** *japonica* **Amycolatopsis** *sp* **Anaerobiospirillum** *succiniciproducens* **Anaerococcus** *hydrogenalis* **Anaerococcus** *sp* **Anaerotruncus** *migulanus* **Aquincola** *tertiaricarbonis* **Arcanobacterium** *canis* **Arcanobacterium** *nitrofigillis* **Arcobacter** *skirrowii* **Aromatoleum** *alkani* **Aromatoleum** *anaerobium* **Aromatoleum** *toluovorans* **Arsenicicoccus** *bolidensis* **Arsenicicoccus** *derm* **Arsenicicoccus** *citreus* **Arthrobacter** *creatinolyticus* **Arthrobacter** *crystallopoietes* **Arthrobacter** *monumenti* **Arthrobacter** *mysorens* **Arthrobacter** *nasiphocae* **Arthrobacter** *protophormiae* **Arthrobacter** *psychrolactophilus* **Arthrobacter** *psychrophend* **Arthrobacter** *tumbae* **Arthrobacter** *uratoxydans* **Arthrobacter** *ureafaciens* **Arthrobacter** *sp* **Avibacterium** *gallinarum* **Avibacterium** *volantium* **Azoarcus** *communis* **Azoarcus** *arsenicus* **Bacillus** *asahii* **Bacillus** *atrophaeus* **Bacillus** *azotoformans* **Bacillus** *cohnii* **Bacillus** *decolorationis* **Bacillus** *drentensis* **Bacillus** *endophy* **Bacillus** *hemicellulosilyticus* **Bacillus** *horikoshii* **Bacillus** *horneckiae* **Bacillus** *horti* **Bacillus** *mannanilyticus* **Bacillus** *marisflavi* **Bacillus** *megaterium* **Bacillus** *m* **Bacillus** *pseudofirmus* **Bacillus** *pseudomycooides* **Bacillus** *psychrosaccharo* **Bacillus** *subterraneus* **Bacillus** *subtilis* **Bacillus** *thermoamylovorans* **Bacillus** *coagulans* **Bacteroides** *coprocola* **Bacteroides** *coprophilus* **Bacteroides** *eg* **Bacteroides** *plebeius* **Bacteroides** *pyogenes* **Bacteroides** *salyersiae* **Bacteroides** *stercor* **Bacteroides** *angulatum* **Bifidobacterium** *animalis* **Bifidobacterium** *asteroids* **Bifidobacterium** *longum* **Bifidobacterium** *magnum* **Bifidobacterium** *merycicum* **Bifidobacterium** *sp* **Bifidobacterium** *thermophilum* **Bilophila** *sp* **Bilophila** *wadsworthia* **Blastobacterium** *sp* **Borrelia** *burdorferi* **Borrelia** *parinii* **Borrelia** *spielmanii* **Brachybacterium** *sp*

MALDI Biotyper library

Fungi Library



45 genus, 129 species – 366 strains

<i>Absidia coerulea</i>	<i>Botrytis cinerea</i>	<i>Fusarium solani</i>	<i>Penicillium citrinum</i>	<i>Phoma sorghina</i>
<i>Acremonium strictum</i>	<i>Chaetomium funicola</i>	<i>Fusarium sporotrichoides</i>	<i>Penicillium commune</i>	<i>Rhizomucor pusillus</i>
<i>Alternaria alternata</i>	<i>Chaetomium globosum</i>	<i>Fusarium tabacinum</i>	<i>Penicillium corylophilum</i>	<i>Rhizopus microsporus</i>
<i>Arthrinium phaeospermum</i>	<i>Chrysosporium keratinophilum</i>	<i>Fusarium verticillioides</i>	<i>Penicillium crustosum</i>	<i>Rhizopus oryzae</i>
<i>Arthrographis_kalrae</i> [ana] (<i>Eremomyces_langeronii</i> [teleo])	<i>Cladosporium cladosporioides</i>	<i>Geomyces pannorum</i>	<i>Penicillium daleae</i>	<i>Rhizopus stolonifer</i>
<i>Aspergillus candidus</i>	<i>Cladosporium herbarum</i>	<i>Geosmithia argillaceae</i>	<i>Penicillium dierckxii</i>	<i>Scedosporium prolificans</i>
<i>Aspergillus clavatus</i>	<i>Cladosporium sp</i>	<i>Kerstesia gyiorum</i>	<i>Penicillium digitatum</i>	<i>Scedosporium_apiospermum</i> [ana] <i>Pseudallescheria_boydii</i> [teleo]
<i>Aspergillus flavus</i>	<i>Cladosporium sp</i>	<i>Lecythophora hoffmannii</i>	<i>Penicillium discolor</i>	<i>Schizophyllum commune</i>
<i>Aspergillus fumigatus</i>	<i>Cunninghamella elegans</i>	<i>Lichtheimia corymbifera</i>	<i>Penicillium expansum</i>	<i>Scopulariopsis acremonium</i>
<i>Aspergillus glaucus</i>	<i>Curvularia clavata</i>	<i>Microsporum canis</i>	<i>Penicillium funiculosum</i>	<i>Scopulariopsis brevicaulis</i>
<i>Aspergillus niger</i>	<i>Curvularia lunata</i>	<i>Microsporum cookei</i>	<i>Penicillium glabrum</i>	<i>Scopulariopsis brumptii</i>
<i>Aspergillus nomius</i>	<i>Curvularia pallescens</i>	<i>Microsporum equinum</i>	<i>Penicillium italicum</i>	<i>Scytalidium lignicola</i>
<i>Aspergillus ochraceus</i>	<i>Curvularia verruculosa</i>	<i>Microsporum fulvum</i>	<i>Penicillium olsonii</i>	<i>Sporothrix schenckii</i>
<i>Aspergillus oryzae</i>	<i>Epicoccum nigrum</i>	<i>Microsporum gypseum</i>	<i>Penicillium pseudostromaticum</i>	<i>Syncephalastrum racemosum</i>
<i>Aspergillus parasiticus</i>	<i>Epidermophyton floccosum</i>	<i>Microsporum persicolor</i>	<i>Penicillium purpurogenum</i>	<i>Thanatephorus cucumeris</i>
<i>Aspergillus sclerotiorum</i>	<i>Fennellia flavipes</i>	<i>Microsporum praecox</i>	<i>Penicillium roqueforti</i>	<i>Trichoderma koningii</i>
<i>Aspergillus sydowi</i>	<i>Fusarium aquaeductuum</i>	<i>Monilinia laxa</i>	<i>Penicillium rugulosum</i>	<i>Trichoderma longibrachiatum</i>
<i>Aspergillus tamarii</i>	<i>Fusarium cerealis</i>	<i>Mucor circinelloides</i>	<i>Penicillium sp</i>	<i>Trichophyton eboreum</i>
<i>Aspergillus terreus</i>	<i>Fusarium chlamydosporum</i>	<i>Mucor ramosissimus</i>	<i>Penicillium striatisporum</i>	<i>Trichophyton equinum</i>
<i>Aspergillus unguis</i>	<i>Fusarium culmorum</i>	<i>Paecilomyces farinosus</i>	<i>Penicillium turbatum</i>	<i>Trichophyton interdigitale</i>
<i>Aspergillus ustus</i>	<i>Fusarium dimerum</i>	<i>Paecilomyces lilacinus</i>	<i>Penicillium verrucosum</i>	<i>Trichophyton rubrum</i>
<i>Aspergillus versicolor</i>	<i>Fusarium equiseti</i>	<i>Paecilomyces marquandii</i>	<i>Phaeoacremonium sp</i>	<i>Trichophyton tonsurans</i>
<i>Aspergillus_amstelodami</i> [ana] <i>Eurotium_amstelodami</i> [teleo]	<i>Fusarium incarnatum</i>	<i>Paecilomyces variotii</i>	<i>Phialemonium sp</i>	<i>Trichophyton violaceum</i>
<i>Aspergillus_nidulans</i> [ana] <i>Emericella_nidulans</i> [teleo]	<i>Fusarium moniliforme</i>	<i>Penicillium brevicompactum</i>	<i>Phialophora bubakii</i>	<i>Trichophyton_mentagrophytes_var_erinacei</i> [ana] <i>Arthroderma benhamiae</i> [teleo]
<i>Aureobasidium pullulans</i>	<i>Fusarium oxysporum</i>	<i>Penicillium chrysogenum</i>	<i>Phoma glomerata</i>	<i>Trichurus sp</i>
<i>Beauveria bassiana</i>	<i>Fusarium proliferatum</i>	<i>Penicillium citreonigrum</i>	<i>Phoma herbarum</i>	

MBT Library

No impact of the updates



BDD v2

BDD v1

Rank	Matched Pattern	Score Value
1	Pseudomonas aeruginosa ATCC 27853 THL	1.983
2	Pseudomonas aeruginosa A07_08_Pudu FLR	1.943
3	Pseudomonas aeruginosa 19955_1 CHB	1.821
4	Pseudomonas aeruginosa DSM 50071T HAM	1.78
5	Pseudomonas indica DSM 14015T HAM	1.55
6	Pseudomonas jinjuensis LMG 21316T HAM	1.49
7	Pseudomonas resinovorans LMG 2274T HAM	1.417
8	Pseudomonas citronellolis DSM 50332T HAM	1.393
9	Pseudomonas taetrolens LMG 2336T HAM	1.349
10	Pseudomonas libanensis CIP 105460T HAM	1.282

Rank	Matched Pattern	Score Value
1	Pseudomonas aeruginosa DSM 1117 DSM	2.188
2	Pseudomonas aeruginosa 8147_2 CHB	2.179
3	Pseudomonas aeruginosa ATCC 27853 THL	1.983
4	Pseudomonas aeruginosa A07_08_Pudu FLR	1.943
5	Pseudomonas aeruginosa 19955_1 CHB	1.821
6	Pseudomonas aeruginosa DSM 50071T HAM	1.78
7	Pseudomonas indica DSM 14015T HAM	1.55
8	Pseudomonas jinjuensis LMG 21316T HAM	1.49
9	Pseudomonas resinovorans LMG 2274T HAM	1.417
10	Pseudomonas citronellolis DSM 50332T HAM	1.393

MALDI Biotyper

Open microbiology concept

MicrobeNet by CDC



Collaboration with U.S. Centers for Disease Control and Prevention, **CDC**

Bruker Collaborates with CDC to Advance Infectious Disease Surveillance

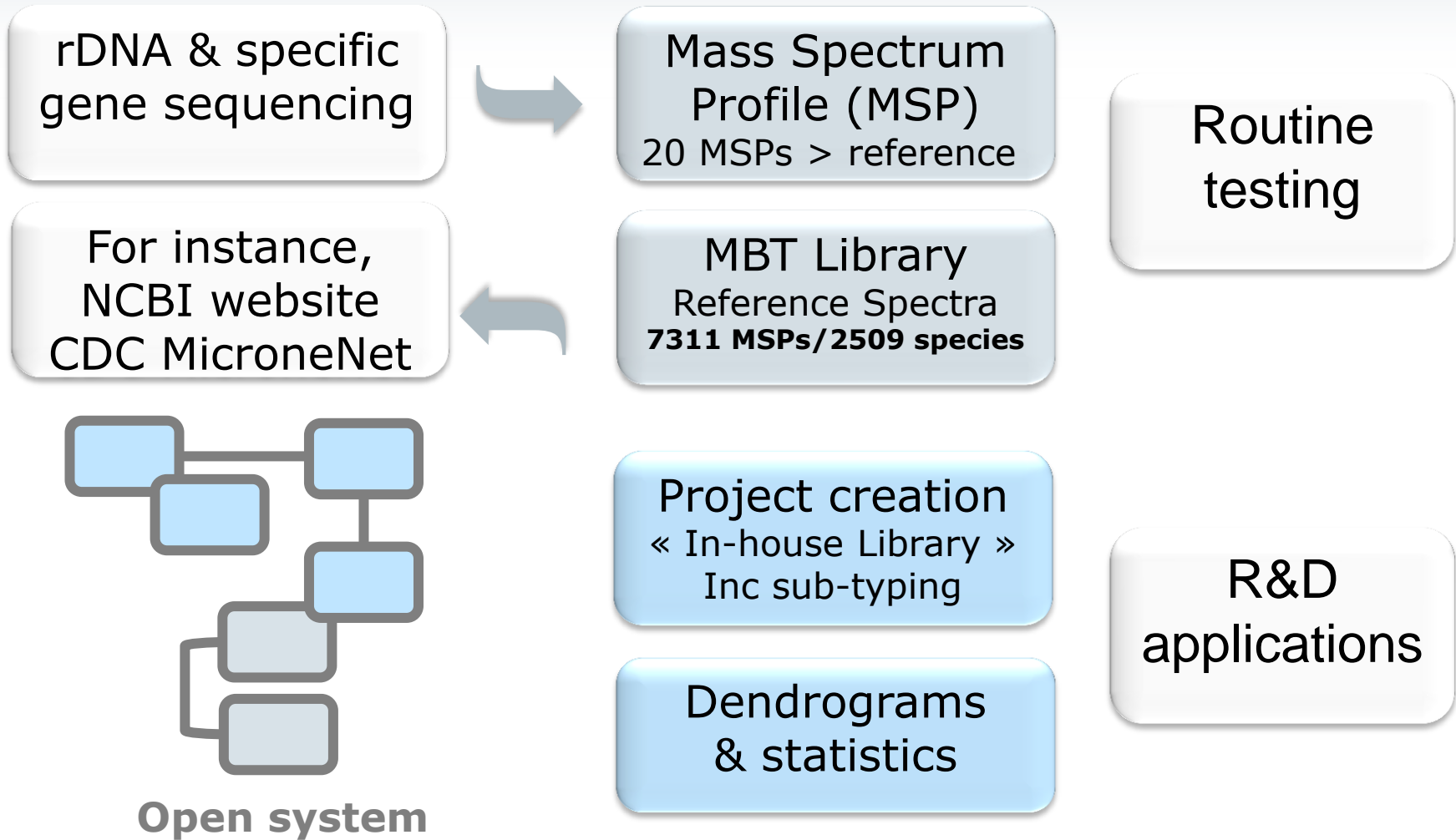
06/16/2016

BOSTON, June 16, 2016 /PRNewswire/ – At the American Society for Microbiology (ASM) Microbe 2016 conference, Bruker today announces the results of a successful collaboration with the Special Bacteriology Reference Laboratory at the U.S. Centers for Disease Control and Prevention (CDC) in Atlanta, Georgia to create an expanded microorganism reference library for the Bruker *MALDI Biotyper*.

This new library, encompassing many rare and emerging pathogens, is now searchable through MicrobeNet (<https://microbenet.cdc.gov>). Developed by the CDC, MicrobeNet is a database curated by the pathogen subject matter experts and accessible for all public health and clinical laboratories. The *MALDI Biotyper* system has been rapidly adopted by microbiology laboratories worldwide as fast and affordable, and as the most comprehensive microbial identification tool available today, replacing conventional biochemical and molecular assays. The supplementary microbe knowledge base generated by the CDC not only provides expanded capabilities, but access to a wealth of other information and references for *MALDI Biotyper* users, from different parts of the world, in the global public health arena. It will help researchers identify a wide variety of microbial pathogens more rapidly in order to better serve public health needs.

The MBT “**Open Microbiology**” concept enables this initiative!

➤ SUMMARY





ELSEVIER

Available online at www.sciencedirect.com

ScienceDirect

MALDI-TOF MS identification and tracking of food spoilers and food-borne pathogens

Chris G de Koster¹ and Stanley Brul²

Current Opinion in Food Science 2016, 10:1–6

This review comes from a themed issue on *Foodomics Technologies*

Edited by Stanley Brul

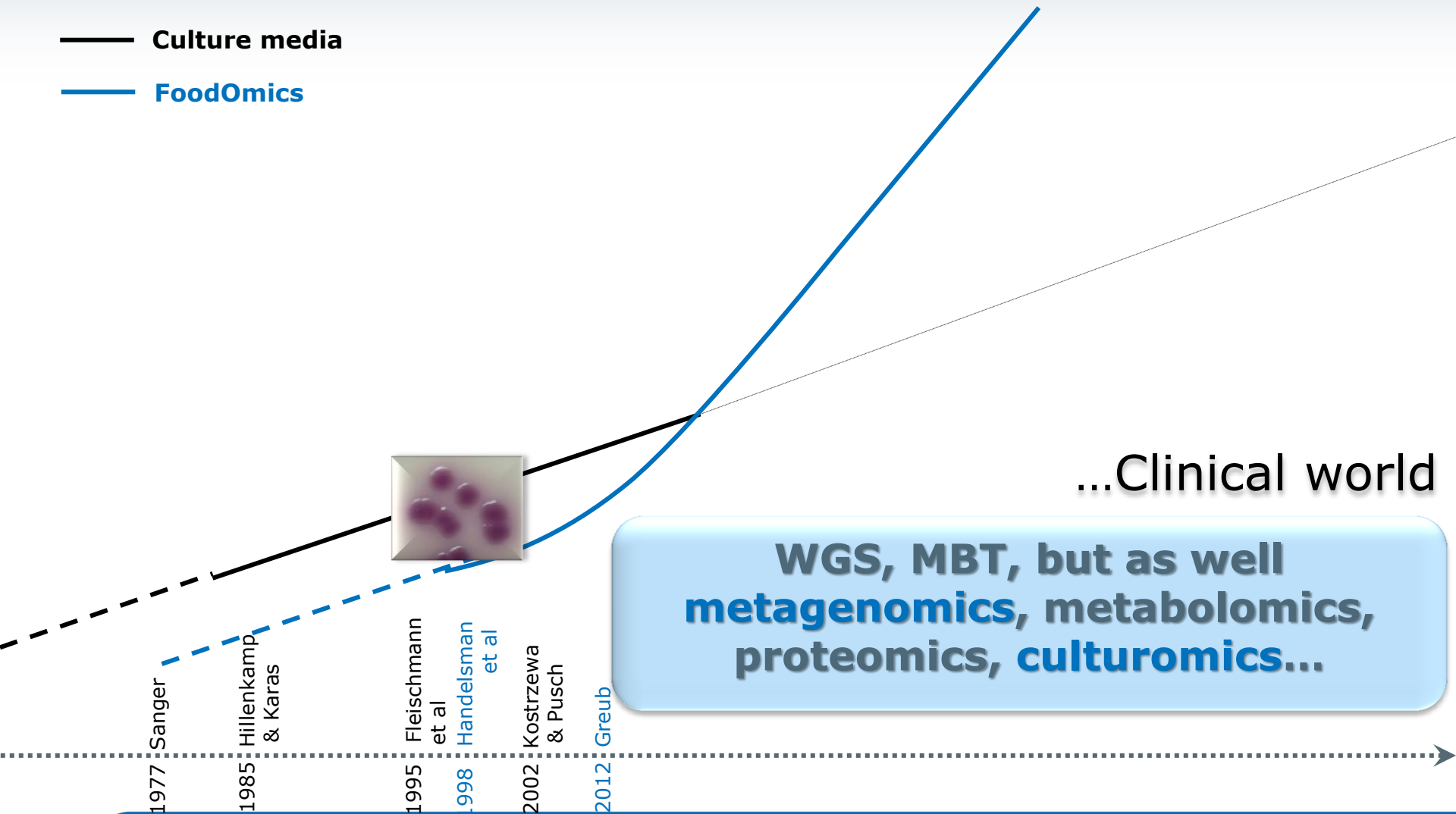


FoodOmics

Cifuentes, 2009. J. Chromatogr. A. 1216, 7109



— Culture media
— FoodOmics





RESEARCH ARTICLE

MALDI-TOF Mass Spectrometry Enables a Comprehensive and Fast Analysis of Dynamics and Qualities of Stress Responses of *Lactobacillus paracasei* subsp. *paracasei* F19

Ann-Sophie Schott¹, Jürgen Behr^{2*}, Jennifer Quinn¹, Rudi F. Vogel¹

¹ Lehrstuhl für Technische Mikrobiologie, Technische Universität München, Freising, Germany, ² Bavarian Center for Biomolecular Mass Spectrometry, Technische Universität München, Freising, Germany



Complementarity WGS & MALDI Biotyper



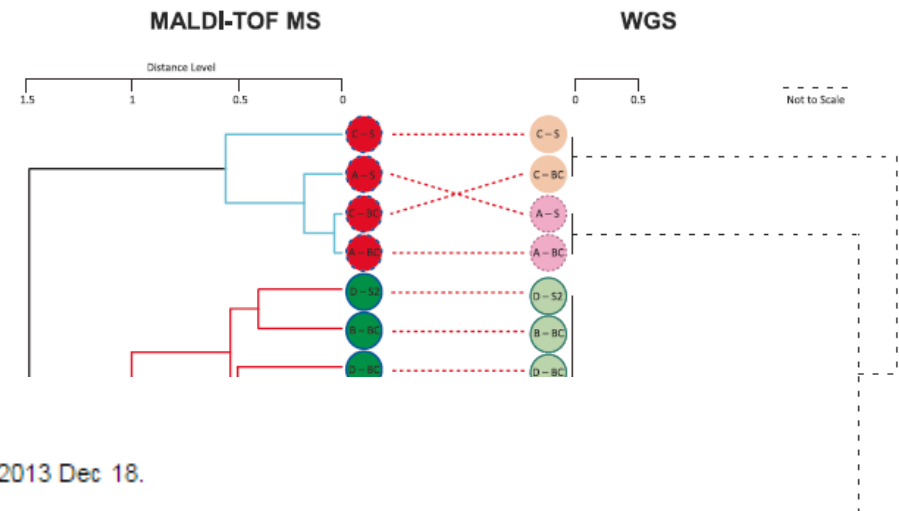
Eur J Clin Microbiol Infect Dis
DOI 10.1007/s10096-016-2824-4



ORIGINAL ARTICLE

MALDI-TOF MS meets WGS in a VRE outbreak investigation

S. Schlebusch^{1,2} • G. R. Price¹ • R. L. Gallagher¹ • V. Horton-Szar¹ • L. D. H. Elbourne³ •
P. Griffin^{2,4,5} • D. J. Venter^{1,2,6} • S. O. Jensen^{7,8} • S. J. Van Hal⁹



See 1 citation found using an alternative search:

[J Dairy Sci.](#) 2014 Feb;97(2):632-41. doi: 10.3168/jds.2013-7147. Epub 2013 Dec 18.

Comparison of methods for the microbiological identification and typing of Cronobacter species in infant formula.

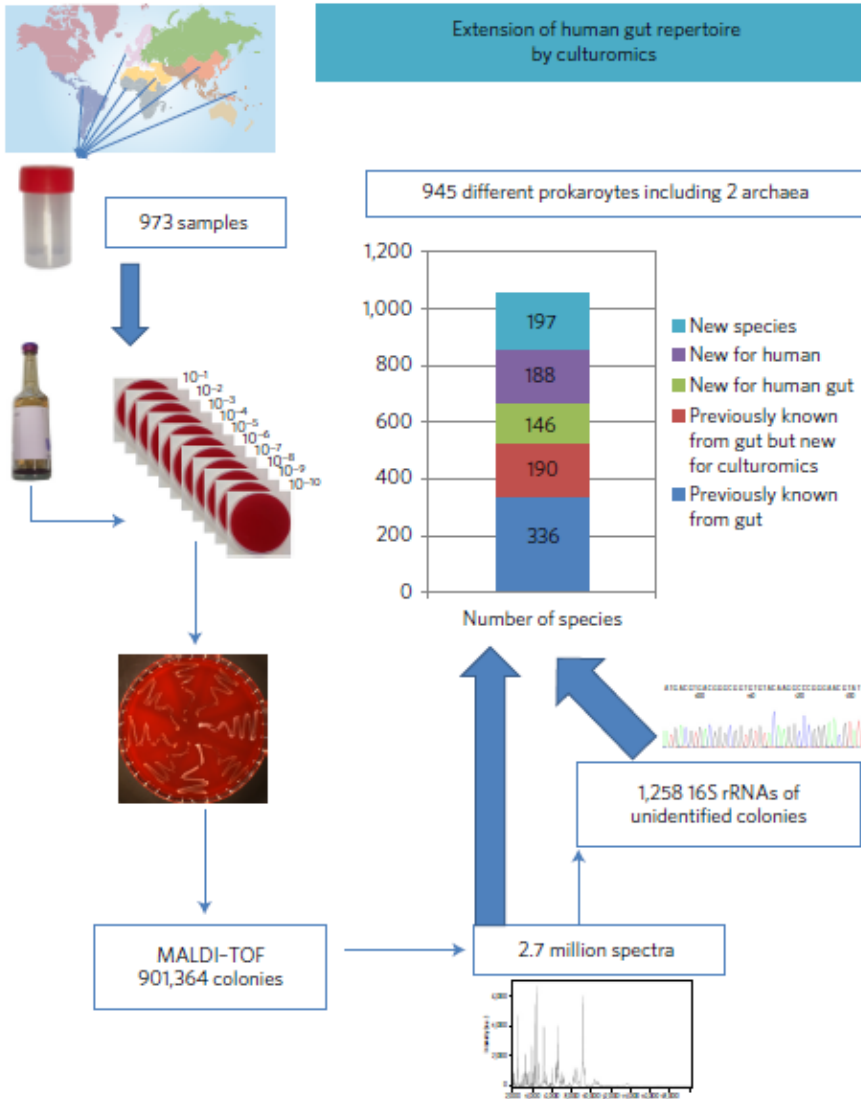
[Lu Y¹](#), [Chen Y²](#), [Lu XA²](#), [Lv J²](#), [Man CX³](#), [Chai YL⁴](#), [Jiang YJ⁵](#).

ST203

Culture of previously uncultured members of the human gut microbiota by culturomics

Jean-Christophe Lagier¹, Saber Khelaifia¹, Maryam Tidjani Alou¹, Sokhna Ndongo¹, Niokhor Dione¹, Perrine Hugon¹, Aurelia Caputo¹, Frédéric Cadoret¹, Sory Ibrahima Traore¹, El Hadji Seck¹, Gregory Dubourg¹, Guillaume Durand¹, Gaël Mourembou¹, Elodie Guilhot¹, Amadou Togo¹, Sara Bellali¹, Dipankar Bachar¹, Nadim Cassir¹, Fadi Bittar¹, Jérémy Delerce¹, Morgane Mailhe¹, Davide Ricaboni¹, Melhem Bilen¹, Nicole Prisca Makaya Dangui Niekou¹, Ndeye Mery Dia Badiane¹, Camille Valles¹, Donia Mouelhi¹, Khoudia Diop¹, Matthieu Million¹, Didier Musso², Jônatas Abrahão³, Esam Ibraheem Azhar⁴, Fehmida Bibi⁴, Muhammad Yasir⁴, Aldiouma Diallo⁵, Cheikh Sokhna⁵, Felix Djossou⁶, Véronique Vitton⁷, Catherine Robert¹, Jean Marc Rolain¹, Bernard La Scola¹, Pierre-Edouard Fournier¹, Anthony Levasseur¹ and Didier Raoult^{1*}

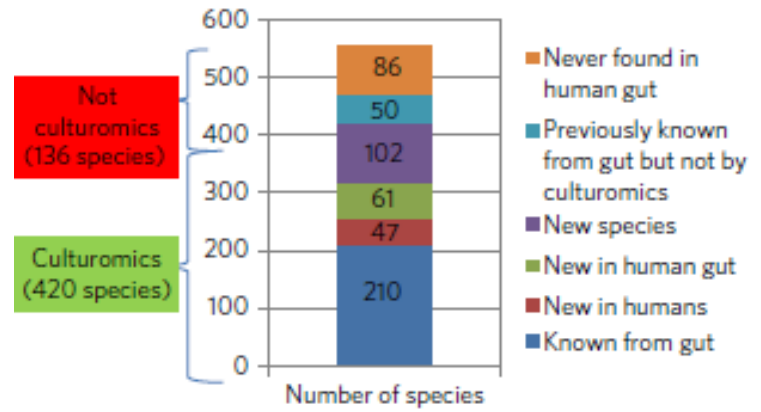
CultuOmics



Comparison of 84 samples analysed by metagenomics and cultuomics

(4) Among the 200 16S rRNAs of the new species: 102 recovered 827 times (average 9.8 per stool)

(5) Analysis of the species with a cut off of 20 reads = 4,158 OTU and 556 species



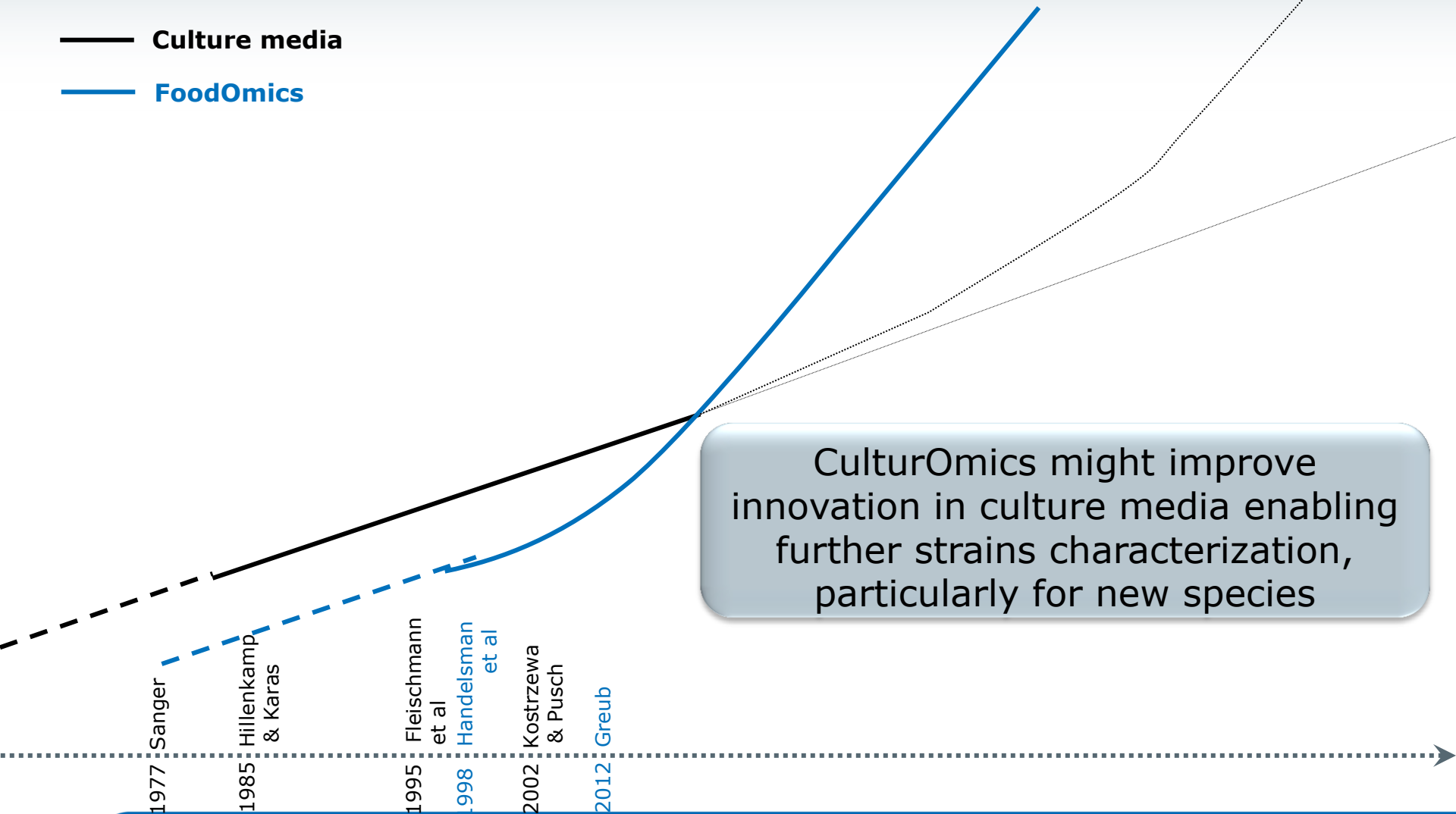
Food microbiomes, e.g fermentation and ripening microbiomes, biofilms, etc...

FoodOmics

Cifuentes, 2009. J. Chromatogr. A. 1216, 7109



— Culture media
— FoodOmics



CulturOmics might improve innovation in culture media enabling further strains characterization, particularly for new species

Are Microbiologists Mutating into Chemists?



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ScienceDirect



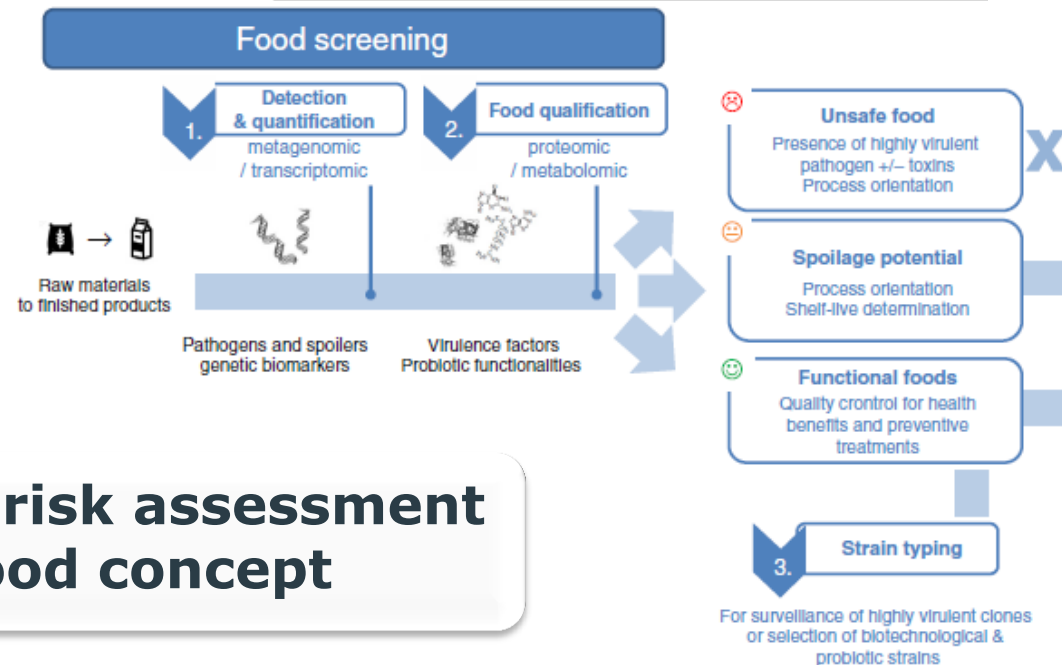
Current Opinion in Food Science 2016, 10:1–6

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Edited by **Stanley Brul**

A typical day working in a laboratory in 2050: are microbiologists becoming chemists and serene workers?

Daniele Sohier, Armelle Riou and Florence Postollec



- 1. Improve microbial risk assessment
- 2. Develop pharma-food concept

Thank you, and see you in

