

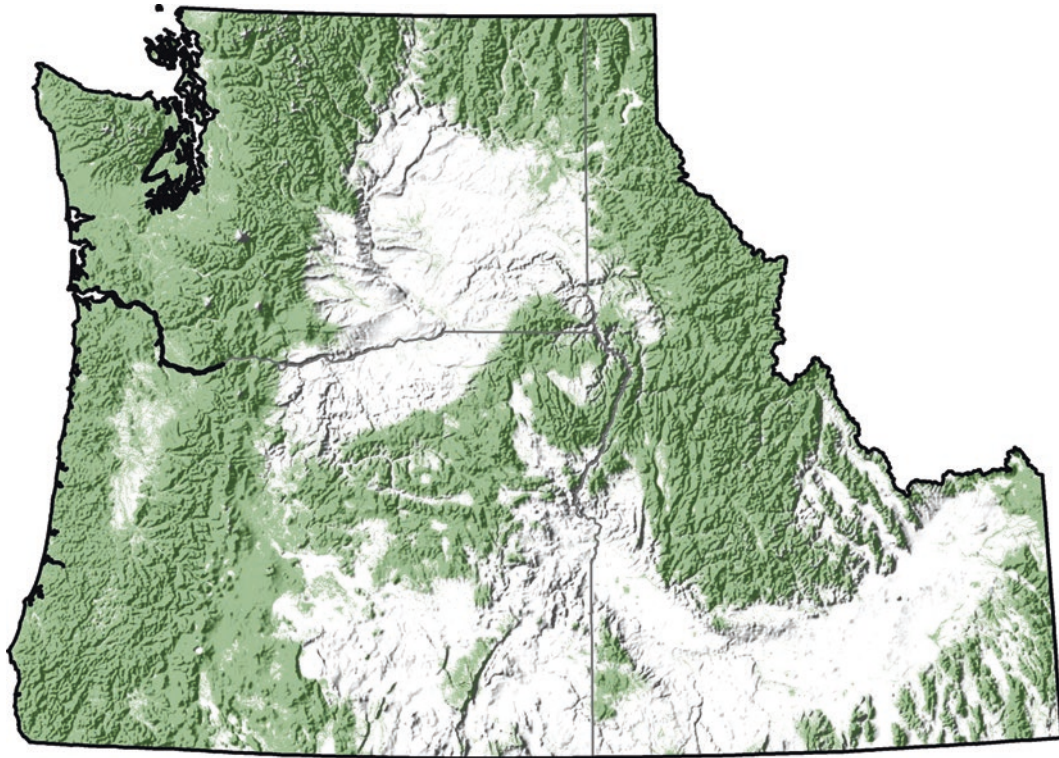
- Ostertag R, Giardina CP, Cordell S (2008) Understory colonization of *Eucalyptus* plantations in Hawaii in relation to light and nutrient levels. *Restor Ecol* 16(3):475–485
- Ostertag R, Cordell S, Michaud J et al (2009) Ecosystem and restoration consequences of invasive woody species removal in Hawaiian lowland wet forest. *Ecosystems* 12:503–515
- Ostertag R, Warman L, Cordell S, Vitousek PM (2015) Using plant functional traits to restore Hawaiian rainforest. *J Appl Ecol* 52:805–809
- Pattison RR, Goldstein G, Ares A (1998) Growth, biomass allocation and photosynthesis of invasive and native Hawaiian rainforest species. *Oecologia* 117:449–459
- Pitt WC, Beasley J, Witmer GW (2017) Ecology and management of terrestrial vertebrate invasive species in the United States. CRC Press, Boca Raton, 415 p
- Pyšek P, Pergl J, Essl F, Lenzner B, Dawson W, Kreft H, Weigelt P et al (2017) Naturalized alien flora of the world. *Preslia* 89(3):203–274
- Reichard SH, White P (2001) Horticulture as a pathway of invasive plant introductions in the United States: Most invasive plants have been introduced for horticultural use by nurseries, botanical gardens, and individuals. *Bioscience* 51(2):103–113. [https://doi.org/10.1641/0006-3568\(2001\)051\[0103:haapoi\]2.0.co;2](https://doi.org/10.1641/0006-3568(2001)051[0103:haapoi]2.0.co;2)
- Schmidt JP, Drake JM (2011) Time since introduction, seed mass, and genome size predict successful invaders among the cultivated vascular plants of Hawaii. *PLoS One* 6(3):e17391. <https://doi.org/10.1371/journal.pone.0017391>
- Schulten JR, Cole TC, Cordell S et al (2014) Persistence of native trees in an invaded Hawaiian lowland wet forest: experimental evaluation of light and water constraints. *Pac Sci* 68(2):267–285. <https://doi.org/10.2984/68.2.7>
- Sin H, Beard KH, Pitt WC (2008) An invasive frog, *Eleutherodactylus coqui*, increases new leaf production and leaf litter decomposition rates through nutrient cycling in Hawaii. *Biol Invasions* 10(3):335–345. <https://doi.org/10.1007/s10530-007-9133-x>
- Smith CW (1985) Impact of alien plants on Hawaii's native biota. In: Stone CP, Scott JM (eds) *Hawaii terrestrial ecosystems: preservation and management*, Cooperative National Park Resources Studies Unit. University of Hawaii, Honolulu, pp 180–250
- Smith CW, Tunison JT (1992) Fire and alien plants in Hawaii: research and management implications for native ecosystems. In: Stone CP, Smith CW, Tunison JT (eds) *Alien plant invasions in native ecosystems of Hawaii: management and research*, Cooperative National Park Resources Studies Unit. University of Hawaii, Honolulu, pp 394–408
- Space J, Falanruw M (1999) Observations on invasive plant species in Micronesia. (Report prepared for the meeting of the Pacific Islands Committee, Council of Western State Foresters, Majuro, Republic of the Marshall Islands)
- Spatz DR, Zilliacus KM, Holmes ND et al (2017) Globally threatened vertebrates on islands with invasive species. *Sci Adv* 3(10):e1603080. <https://doi.org/10.1126/sciadv.1603080>
- Stratton LC, Goldstein G (2001) Carbon uptake, growth and resource-use efficiency in one invasive and six native Hawaiian dry forest tree species. *Tree Physiol* 21:1327–1334
- Timm OE, Giambelluca TW, Diaz HF (2015) Statistical downscaling of rainfall changes in Hawai'i based on the CMIP5 global model projections. *J Geophys Res-Atmos* 120(1):92–112. <https://doi.org/10.1002/2014JD022059>
- Trauernicht C, Pickett E, Giardina CP et al (2015) The contemporary scale and context of wildfire in Hawai'i 1. *Pac Sci* 69(4):427–444
- Vitousek PM (1986) Biological invasions and ecosystem properties: can species make a difference? In: Mooney HA, Drake JA (eds) *Ecology of biological invasions of North America and Hawai'i*. Springer, New York, pp 163–176
- Vorsino AE, Fortini LB, Amidon FA et al (2014) Modeling Hawaiian ecosystem degradation due to invasive plants under current and future climates. *PLoS One* 9(5):e95427. <https://doi.org/10.1371/journal.pone.0095427>
- Wagner WL, Herbst DR, Sohmer SH (1999) *Manual of the flowering plants of Hawaii*. Bishop Museum Special Publication 97. 2 vols. University of Hawaii Press and Bishop Museum Press, Honolulu. 1919 p
- Williams DG, Mack RN, Black RA (1995) Ecophysiology of introduced *Pennisetum setaceum* on Hawaii: the role of phenotypic plasticity. *Ecology* 76(5):1569–1580
- Woodcock D (2003) To restore the watersheds: early twentieth-century tree planting in Hawai'i. *Ann Assoc Am Geogr* 93(3):624–635
- Zou X (1993) Species effects on earthworm density in tropical tree plantations in Hawaii. *Biol Fertl Soils* 15(1):35–38. <https://doi.org/10.1007/bf00336285>

## Northwest Region

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## Introduction

The Northwest region (Idaho, Oregon, and Washington; Figs. A3.1 and A3.2) contains major coastal and inland ports (Coos Bay, Lewiston, Pasco, Portland, Seattle-Tacoma, and The Dalles), waterways (Puget Sound, Columbia River Basin, and Willamette Valley), and major



**Fig. A3.1** The Northwest Region

**Fig. A3.2** The Northwest region of the United States includes Idaho, Oregon, and Washington. (Figure courtesy of Daniel Ryerson, USDA Forest Service Southwestern Region, Forest Health Protection)



highway and rail arteries that provide pathways for invasive plants, pathogens, insects, and vertebrates. Two main pathways in this region for the introduction of invasive forest and horticultural pathogens and insects are shipping infrastructure, especially solid wood packing material from shipping cargo (USDA Forest Service and APHIS 2000), and the live plant trade (Liebhold et al. 2012). Temperate coastal climates, diverse and abundant native vegetation, extensive trade patterns with Pacific Rim nations, and the

border with Canada and its major west coast port of Vancouver have facilitated the repeated introductions of major forest pests such as the European (also known as North American) gypsy moth (*Lymantria dispar dispar*), the Asian gypsy moth (*Lymantria dispar asiatica/japonica*), and the sudden oak death pathogen (*Phytophthora ramorum*). The region has a significant horticultural industry, extensive areas of mesic and dryland agriculture, and abundant urban and native forests whose trees can serve as

adventive hosts or alternate hosts for invasive insects and pathogens as well as disturbed landscapes for invasive plants. Forested lands are regionally vital to the forest industry and are a recreational base for millions in the Northwest. Invasive species have the potential to inflict severe economic hardship on individuals, local governments, and the businesses involved in the forest, horticultural, agricultural, and tourism industries.

The Northwest region has a long history of invasive forest insects and diseases; for example, white pine blister rust, caused by *Cronartium ribicola*, was first introduced on the West Coast and was “Pest #1” on the historical “Quarantine 37” of the Plant Quarantine Act of 1912, and balsam woolly adelgid (*Adelges piceae*) was first noted in this region in the 1920s (Annand 1928; Keen 1952). More modern regulations such as the International Standards for Phytosanitary Measures No. 15 (ISPM 15), an international treaty signed by 200 nations, play a major role in protecting this region from invasive species that may arrive in solid wood packing material (FAO 2009; Strutt et al. 2013). Also, examples of the more profound impacts of climate change and invasion biology are developing in high-elevation ecosystems of this region where five-needle white pines (*Pinus* spp.) and subalpine fir (*Abies lasiocarpa*) trees are experiencing unprecedented rates of mortality (Shoal and Aubry 2006). In these instances the mountain pine beetle, *Dendroctonus ponderosae*, has been a native invader of high-elevation ecotones (Bentz et al. 2010; Logan and Powell 2001; Logan et al. 2003), whereas *C. ribicola* (on pines) and *A. piceae* (on subalpine fir) are significant non-native invaders with expanding elevational ranges (Hrinkevich et al. 2016).

Regional State government departments, interagency collaborative groups such as State Invasive Species Councils, regional USDA Forest Service personnel, and other entities have identified nearly 190 species and species groups as regional invasive or nuisance species of key concern. Altogether, the taxa identified as detected or potential invasive species fall into five categories: 78 plants; 11 plant pathogens and parasites; 93 insect species; 10 aquatic invertebrates (noninsect); and 23 vertebrates (Tables A3.1, A3.2, A3.3, A3.4 and A3.5, Bautista 2017; Flitcroft et al. 2016; Invasive Species of Idaho 2017; OIE 2018; OISC 2015; Prather et al. 2016; WISP 2009). Because of the large number of organisms (most notably plants) treated in this summary (Tables A3.1, A3.2, A3.3, A3.4 and A3.5), scientific names are only provided in the text for the most prominent species. This overview will focus on a subset of the species that are established and have caused resource damage or that have been introduced repeatedly into the Northwest region.

## Plants

Aquatic invasive plants of this region (Table A3.1) include multiple species of submersed aquatic plants (i.e., *Elodea* spp., *Hydrilla* spp., milfoils, and swollen bladderwort), emergent plants (reeds, *Spartina* spp., loosestrife, rushes, giant salvinia, reed canary grass, foxtail barley, yellow flag iris, and water primrose), and floating plants (parrotfeather, curly-leaf pondweed, water chestnut, yellow floating heart, West Indian spongeplant, and dotted duckweed). Two non-plants, a cyanobacterium (toxic blue-green “alga”) and a diatom (rock snot or didymo), are also included in this survey (Table A3.1). Once established, many of these taxa can cause significant impairment of water quality and navigation (by growing as dense mats) (see Chap. 2, Sect. 2.5, for additional discussion of these impacts). Furthermore, established populations may be spread by waterfowl as they move from one location to another, or by human vectors (e.g., boats and fishing gear/tackle). Marine invasive plants are not included in the survey.

Several priority aquatic plants can achieve high densities, leading to ecological problems or nuisance issues for people. Emergent plants can dominate wetland and floodplain areas (i.e., reed canary grass, water primrose, and purple loosestrife), outcompeting or displacing native species. Some invasive aquatic plants were brought initially to the region by the aquarium trade (*Elodea* spp.) or for ornamental use or seeding of wet areas for livestock or waterfowl (reed canary grass (*Phalaris arundinacea*)). Once established, cut fragments of the plants, broken by boat propellers or wildlife, can disperse for colonization. Some species such as *Elodea canadensis* have broad ecological tolerance, which make expansion and invasion a concern. Non-native and native species of water primroses (*Ludwigia* spp.) comprise a plant group whose growth has become problematic in recent years. One species, *L. grandiflora*, native to the Eastern United States, Central America, and South America, is now present in Oregon and Washington (CABI 2018). Physical and chemical alteration of the environment by *L. grandiflora* can cause severe damage to local ecosystems and biodiversity. Dense stands of this plant can reduce floodwater retention, cause hyper-sedimentation and silting, and block slow-moving waterways. The plant also gives off allelopathic elements that impact water quality throughout the year and make it detrimental to vulnerable native flora. Because it can shade out other submersed vegetation, it is generally considered a threat to biodiversity in its introduced range (CABI 2018).

Non-plant toxic “algae” are a health concern for native vertebrates and humans because they create powerful toxins known to kill fish, ducks, geese, marine mammals, and other

**Table A3.1** Priority non-native invasive plants of the Northwest region

Scientific name	Common name	ID	OR	WA	R6 <sup>a</sup>
<b>Aquatic plants</b>					
<i>Lagarosiphon major</i>	African waterweed or African elodea		x		x
<i>Phragmites australis</i>	Common reed	x	x	x	
<i>Potamogeton crispus</i>	Curly-leaf pondweed	x			x
<i>Butomus umbellatus</i>	Flowering rush	x	x		x
<i>Salvinia molesta</i>	Giant salvinia	x	x		
<i>Hydrilla verticillata</i>	Hydrilla, water thyme	x	x	x	x
<i>Myriophyllum</i> spp. including <i>M. spicatum</i> , <i>M. aquaticum</i>	Milfoils: Eurasian, parrotfeather	x		x	x
<i>Lythrum salicaria</i> , <i>Lysimachia vulgaris</i>	Purple loosestrife, garden yellow loosestrife	x		x	x
<i>Phalaris arundinacea</i> ; <i>P. arundinacea</i> var. <i>picta</i>	Reed canary grass; ribbon grass				x
<i>Didymosphenia geminata</i>	Rock snot (Didymo) <sup>b</sup>		x		x
<i>Chondrilla juncea</i>	Rush skeletonweed	x		x	x
<i>Spartina</i> spp. including <i>S. alterniflora</i> , <i>S. densiflora</i>	Spartina (cordgrass)		x	x	
<i>Prymnesium parvum</i> , <i>Cylindrospermopsis raciborskii</i>	Toxic algae (golden, toxic cyanobacteria) <sup>b</sup>		x		
<i>Trapa natans</i>	Water chestnut (European)	x	x	x	
<i>Ludwigia grandiflora</i>	Water primrose <sup>c</sup>		x	x	x
<i>Eichhornia crassipes</i>	Water hyacinth	x			
<i>Egeria densa</i> , <i>Elodea nuttallii</i> , <i>E. canadensis</i> , <i>E. canadensis</i> x <i>E. nuttallii</i> hybrid	Brazilian elodea, western waterweed (elodea)	x		x	
<i>Iris pseudacorus</i>	Yellow flag iris	x			x
<i>Nymphoides peltata</i>	Yellow floating heart	x	x		x
<b>Riparian-terrestrial plants</b>					
<i>Peganum harmala</i>	African rue		x		
<i>Alyssum corsicum</i> , <i>A. murale</i> , <i>Berteroa incana</i>	Alyssums	x	x		x
<i>Hyoscyamus niger</i>	Black henbane	x			
<i>Solanum rostratum</i>	Buffalo bur	x			
<i>Buddleja davidii</i> ; <i>B. globosa</i>	Butterfly bush			x	
<i>Alhagi maurorum</i>	Camelthorn		x		
<i>Bromus tectorum</i>	Cheatgrass				x
<i>Tussilago farfara</i>	Coltsfoot (European)		x		
<i>Anchusa officinalis</i> , <i>A. arvensis</i> , <i>Echium vulgare</i>	Common bugloss, small bugloss, viper's bugloss	x			x
<i>Crupina vulgaris</i>	Common crupina	x		x	x
<i>Isatis tinctoria</i>	Dyer's woad	x			
<i>Linaria dalmatica</i> , <i>L. vulgaris</i>	Toadflax (Dalmatian, yellow)	x		x	x
<i>Hedera helix</i>	English ivy				x
<i>Ammophila arenaria</i>	European beachgrass				x
<i>Brachypodium sylvaticum</i>	False brome				x
<i>Cabomba caroliniana</i>	Fanwort	x			
<i>Azolla pinnata</i>	Feathered mosquito fern	x			
<i>Convolvulus arvensis</i>	Field bindweed	x			
<i>Alliaria petiolata</i>	Garlic mustard			x	x
<i>Geranium robertianum</i> , <i>G. lucidum</i>	Geranium				x
<i>Heracleum mantegazzianum</i>	Giant hogweed	x	x	x	
<i>Aegilops triuncialis</i> , <i>A. ovata</i> , <i>A. cylindrica</i>	Goatgrasses (barbed, ovate, jointed)	x	x		x
<i>Galega officinalis</i>	Goatsrue		x		
<i>Ulex europaeus</i>	Gorse				x
<i>Hieracium piloselloides</i> , <i>H. pratense</i> , <i>H. pilosella</i> , <i>H. aurantiacum</i> , <i>H. floribundum</i> , <i>H. caespitosum</i> , <i>H. lachenalii</i>	Hawkweeds	x	x	x	x
<i>Rubus armeniacus</i>	Himalayan blackberry			x	x
<i>Cynoglossum officinale</i>	Houndstongue	x			x
<i>Cuscuta japonica</i>	Japanese dodder		x		
<i>Sorghum halepense</i>	Johnsongrass	x			
<i>Centaurea virgata</i> ; <i>C. vulgaris</i> ; <i>C. jacea</i> x <i>nigra</i> ; <i>C. stoebe</i>	Knapweeds	x	x	x	x
<i>Fallopia japonica</i> var. <i>japonica</i> ; <i>Polygonum bohemicum</i>	Knotweeds (Japanese, bohemian)	x		x	x
<i>Kochia scoparia</i> ssp. <i>scoparia</i>	Kochia			x	

(continued)

**Table A3.1** (continued)

Scientific name	Common name	ID	OR	WA	R6 <sup>a</sup>
<i>Pueraria montana</i> var. <i>lobata</i>	Kudzu		x	x	
<i>Euphorbia esula</i>	Leafy spurge	x		x	x
<i>Nardus stricta</i>	Matgrass	x	x		
<i>Salvia aethiopsis</i>	Mediterranean sage	x			
<i>Taeniatherum caput-medusae</i>	Medusahead				x
<i>Milium vernale</i>	Millium	x			
<i>Ventenata dubia</i>	North Africa grass				x
<i>Euphorbia oblongata</i>	Oblong spurge		x		
<i>Clematis vitalba</i>	Old man's beard				x
<i>Leucanthemum vulgare</i>	Oxeye daisy	x			
<i>Echium plantagineum</i>	Paterson's curse		x		
<i>Lepidium latifolium</i>	Perennial pepperweed	x			
<i>Sonchus arvensis</i>	Perennial sowthistle	x			
<i>Conium maculatum</i>	Poison hemlock	x			
<i>Impatiens glandulifera</i>	Policeman's helmet	x			x
<i>Tribulus terrestris</i>	Puncturevine	x			x
<i>Cyperus rotundus</i>	Purple nutsedge		x		
<i>Cytisus scoparius</i>	Scotch broom	x		x	x
<i>Solanum elaeagnifolium</i>	Silverleaf nightshade		x		
<i>Centaurea iberica</i> , <i>C. calcitrapa</i> , <i>C. solstitialis</i>	Starthistles	x	x	x	
<i>Potentilla recta</i>	Sulfur cinquefoil				x
<i>Zygophyllum fabago</i>	Syrian bean-caper	x	x		
<i>Tamarix</i> spp.	Tamarix (saltcedar)	x		x	x
<i>Senecio jacobaea</i>	Tansy ragwort	x		x	x
<i>Carduus nutans</i> , <i>C. acanthoides</i> ; <i>Carthamus baeticus</i> , <i>C. lanatus</i> ; <i>Onopordum tauricum</i> , <i>O. acanthium</i>	Thistles	x	x	x	x
<i>Bryonia alba</i>	White bryony	x	x		
<i>Lepidium draba</i>	Whitetop/hoary cress	x			x

<sup>a</sup>R6 = USDA Forest Service Region 6 (OR, WA) priority invasive species

<sup>b</sup>Non-plants (diatom and cyanobacterium, respectively)

<sup>c</sup>Other species of *Ludwigia* (e.g., *L. palustris*) are native to the Northwest region but have become pests

**Table A3.2** Priority non-native invasive plant pathogens and other parasites of the Northwest region

Scientific name	Common name	ID	OR	WA	R6 <sup>a</sup>	Other <sup>b</sup>	Plant or animal host
<i>Phytophthora lateralis</i>	Port-Orford-cedar root disease		x		x		<i>Chamaecyparis lawsoniana</i> , <i>Taxus brevifolia</i>
<i>Phytophthora ramorum</i>	Sudden oak death	x	x		x		<i>Quercus</i> spp., <i>Notholithocarpus</i> spp., many others
<i>Batrachochytrium dendrobatidis</i>	Amphibian chytrid fungus ( <i>Bd</i> )					x	
<i>B. salamandrivorans</i>	Amphibian chytrid fungus ( <i>Bsal</i> )					x	
<i>Ranavirus</i>	Ranavirus					x	
<i>Chronic wasting disease prion</i>	Chronic wasting disease (CWD)		x				
<i>Pseudogymnoascus destructans</i>	White-nose syndrome				x		
<i>Cronartium ribicola</i>	White pine blister rust		x	x	x		<i>Pinus</i> spp.
<i>Melampsora larici-populina</i>	Eurasian poplar leaf rust		x	x			<i>Populus</i> spp.
<i>Lachnellula willkommii</i>	European larch canker	x					
<i>Puccinia graminis</i>	Black stem rust	x					

<sup>a</sup>R6 = US Forest Service Region 6 (OR, WA) priority invasive species

<sup>b</sup>Other = World Organization for Animal Health (Office International des Epizootics) priorities

**Table A3.3** Non-native invasive insects of the Northwest Region include beetles, flies, aphids/adelgids, thrips, wasps, and moths

Scientific name	Common name	Occurrence			Origin	Introduction		Plant or animal host
		ID	OR	WA		Accidental	Intentional	
<b>Coleoptera</b>								
<i>Agrilus cuprescens</i>	Rose stem girdler			x	Europe	•		<i>Rosa</i> spp., <i>Rubus</i> spp.
<i>Agriotes lineatus</i>	Lined click beetle			x	Europe	•		Grass and plant roots, Potatoes, strawberries
<i>Agriotes obscurus</i>	Dusky click beetle			x	Europe	•		Grass and plant roots Potatoes, strawberries
<i>Amphimallon majale</i>	European chafer		x	x	Europe	•		Grass roots
<i>Anobium punctatum</i>	Furniture beetle			x	Europe	•		Wood products
<i>Apion fuscirostre</i>					Europe		•	Grorse ( <i>Ulex europaeus</i> )
<i>Apion ulicis</i>	Grorse weevil	x	x	x	Europe		•	Grorse
<i>Brachyterolus pulicarius</i>	Toadflax flower-feeding beetle	x	x	x	Europe		•	Dalmatian and yellow toadflax, <i>Linaria</i> spp.
<i>Chrysolina hyperici</i>		x	x	x	Europe via Australia		•	Klamathweed ( <i>Hypericum perforatum</i> )
<i>Chrysolina quadrigemina</i>	Klamathweed beetle	x	x	x	Europe via Australia		•	Klamathweed ( <i>Hypericum perforatum</i> )
<i>Cryptorhynchus lapathi</i>	Poplar-and-willow borer	x	x	x	Europe	•		<i>Salix</i> spp. and <i>Populus</i> spp.
<i>Cyclorhipidion bodoanum</i>			x	x	Asia	•		Broad-leaved trees (woodborer)
<i>Hylastes opacus</i>			x	x	Europe	•		<i>Pinus</i>
<i>Ips paraconfusus</i>	California fivespined ips		O	x	California, Oregon	•		<i>Pinus</i> (invasive in Washington, native to Oregon)
<i>Laricobius erichsonii</i>			x	x	Europe		•	<i>Adelges piceae</i>
<i>Larinus minutus</i>	Lesser knapweed flower weevil	x	x	x	Europe		•	Knapweeds, <i>Centaurea</i> sp.
<i>Larinus obtusus</i>	Blunt knapweed flower weevil	x	x	x	Europe		•	Knapweeds, <i>Centaurea</i> sp.
<i>Lasioderma serricorne</i>	Cigarette beetle	x	x	x	Europe (cosmopolitan)	•		Plant products
<i>Lilioceris lili</i>	Scarlet lily beetle			x	Eurasia	•		<i>Fritillaria</i> spp. and <i>Lilium</i> spp. leaves
<i>Lyctus brunneus</i>	Old World lyctus beetle	x	x	x	Europe (cosmopolitan)	•		Wood products
<i>Lyctus linearis</i>	European lyctus beetle	x	x	x	Europe (cosmopolitan)	•		Wood products
<i>Mecinus janthiniformis</i>	Dalmatian toadflax stem-mining weevil	x	x	x	Southeastern Europe		•	Dalmatian toadflax, <i>Linaria dalmatia</i>
<i>Mecinus janthinus</i>	Yellow toadflax stem-mining weevil	x	x	x	Eurasia		•	Yellow toadflax, <i>Linaria vulgaris</i>
<i>Melanotus cete</i>			x	x	Japan	•		Unknown
<i>Micromalthus debilis</i>	Telephone pole beetle		x	x	Eastern North America	•		Decayed wood products
<i>Mogulones crucifer</i>	Houndstongue root weevil	x		x	Europe	•		Houndstongue, <i>Cynoglossum officinale</i>
<i>Nacerdes melanura</i>	Wharf borer	x	x	x	Europe	•		Wood products
<i>Orchestes alni</i>	European elm flea weevil	x	x	x	Europe	•		<i>Ulmus</i> spp.
<i>Otiorhynchus ovatus</i>	Strawberry root weevil	x	x	x	Europe	•		Seedling conifers
<i>Otiorhynchus rugosostriatus</i>	Rough strawberry root weevil	x	x		Europe	•		Seedling conifers
<i>Otiorhynchus sulcatus</i>	Black vine weevil	x	x	x	Europe	•		Seedling conifers
<i>Oulema melanopus</i>	Cereal leaf beetle	x	x	x	Eurasia	•		Cereal crops, wild grasses
<i>Phymatodes testaceus</i>	Tanbark borer			x	Europe, Japan, Middle East, North Africa	•		Wood products/bark

(continued)

**Table A3.3** (continued)

Scientific name	Common name	Occurrence			Origin	Introduction		Plant or animal host
		ID	OR	WA		Accidental	Intentional	
<i>Pityophthorus juglandis</i>	Walnut twig beetle	x	x	x	Arizona/New Mexico, Mexico	•		<i>Juglans</i> spp., <i>Pterocarya</i> spp.
<i>Popillia japonica</i>	Japanese beetle	x	x	x	Japan	•		Broad-leaved trees and many ornamentals
<i>Pullus impexus</i>			x		Europe		•	<i>Adelges piceae</i>
<i>Pyrrhalta viburni</i>	Viburnum leaf beetle			x	Eurasia	•		<i>Viburnum</i> spp.
<i>Rhinusa antirrhini</i>	Toadflax seed-galling weevil	x	x	x	Eurasia, Mediterranean	•		Dalmatian and yellow toadflax, <i>Linaria</i> spp.
<i>Rhinusa neta</i>	Toadflax seed-feeding weevil			x	Europe	•		Dalmatian and yellow toadflax, <i>Linaria</i> spp.
<i>Saperda populnea</i>		x	x	x	Europe	•		<i>Populus</i> spp.
<i>Scolytus multistriatus</i>	Smaller European elm bark beetle	x	x	x	Europe	•		<i>Ulmus</i> spp.
<i>Scolytus rugulosus</i>	Shot hole borer	x	x	x	Europe	•		Broad-leaved trees, Rosaceae
<i>Scolytus schevyrewi</i>	Banded elm bark beetle	x	x	x	Asia	•		<i>Ulmus</i> spp.
<i>Stegobium paniceum</i>	Drugstore beetle	x	x	x	Cosmopolitan	•		Stored products
<i>Tenebroides mauritanicus</i>	Cadelle	x	x	x	Cosmopolitan	•		Stored products
<i>Trypodendron domesticum</i>	European hardwood ambrosia beetle			x	Europe	•		Broad-leaved trees
<i>Xanthogaleruca luteola</i>	Elm leaf beetle	x	x	x	Europe	•		<i>Ulmus</i> spp.
<i>Xestobium rufovillosum</i>	Deathwatch beetle		x		Europe	•		Wood products
<i>Xyleborinus alni</i> = ( <i>attenuates</i> )			x	x	Europe/Asia	•		Broad-leaved trees
<i>Xyleborinus saxeseni</i>	Fruit tree pin-hole borer	x	x	x	Europe	•		Broad-leaved trees and conifers
<i>Xyleborus dispar</i>	European shot hole borer	x	x	x	Europe	•		Broad-leaved trees
<i>Xyleborus pfeili</i>			x		Cosmopolitan	•		Broad-leaved trees
<i>Xylosandrus crassiusculus</i>			x		Africa/Asia	•		Broad-leaved trees
<i>Xylosandrus germanus</i>			x	x	Asia	•		Broad-leaved trees
<i>Xyloterinus politus</i>				x	Eastern North America	•		Broad-leaved trees, rare in conifers
<b>Diptera</b>								
<i>Aphidoletes thompsoni</i>			x	x	Europe		•	<i>Adelges piceae</i>
<i>Compsilura concinnata</i>		?	x	x	Europe		•	<i>Lymantria dispar</i> and other Lepidoptera
<i>Cremifania nigrocellulata</i>			x		Europe		•	<i>Adelges piceae</i>
<i>Delia platura</i>	Seedcorn maggot	x	x	x	Europe	•		Seedling conifers
<i>Drosophila suzukii</i>	Spotted wing Drosophila	x	x	x	Asia	•		Fruits/berries
<i>Leucopis obscura</i>			x	?	Europe		•	<i>Adelges piceae</i>
<i>Phytomyza ilicis</i>	Holly leafminer		x		Europe	•		<i>Ilex</i> spp.
<i>Rhagoletis completa</i>	Walnut husk fly	x	x	x	Eastern USA	•		<i>Juglans</i> spp., <i>Prunus persica</i>
<b>Hemiptera</b>								
<i>Adelges abietis</i>	Eastern spruce gall aphid	x	x	x	Europe	•		<i>Picea</i> spp.

(continued)

Table A3.3 (continued)

Scientific name	Common name	Occurrence			Origin	Introduction		Plant or animal host
		ID	OR	WA		Accidental	Intentional	
<i>Adelges nüsslini</i> (= <i>nordmaneanae</i> )		x	x	x	Europe	•		<i>Picea</i> spp., <i>Abies</i> spp.
<i>Adelges piceae</i>	Balsam woolly adelgid	x	x	x	Europe	•		<i>Abies</i> spp.
<i>Adelges strobilobius</i> (= <i>laricis</i> )	Larch woolly aphid	x	x	x	Europe	•		<i>Picea</i> spp., <i>Larix</i> spp.
<i>Arocatus melanocephalus</i>	Elm seed bug	x	x	x	Europe	•		<i>Ulmus</i> spp.
<i>Asterolecanium minus</i>	Oak pit scale			x	Europe	•		<i>Quercus</i> spp.
<i>Cinara tujafilina</i>		x	x	x	Europe	•		Cupressini
<i>Carulaspis juniperi</i>	Juniper scale	x	x	x		•		
<i>Dialeurodes chittendeni</i>	Rhododendron whitefly				Asia	•		<i>Rhododendron</i> spp.
<i>Elatobium abietinum</i>	Spruce aphid		x	x	Europe	•		<i>Picea</i> spp.
<i>Eriosoma ulmi</i>	European elm leafcurl aphid				Europe	•		<i>Ulmus</i> spp.
<i>Eucallipterus tiliae</i>	Linden aphid	x	x	x		•		<i>Tilia</i> spp.
<i>Euceraphis punctipennis</i>	European birch aphid				Europe	•		<i>Betula</i> spp.
<i>Eulecanium cerasorum</i>	Calico scale	x	x	x		•		
<i>Gossyparia spuria</i>	European elm scale				Europe	•		<i>Ulmus</i> spp.
<i>Halyomorpha halys</i>	Brown marmorated stink bug	x	x	x	Asia	•		Fruit/vegetable crops
<i>Icerya purchasi</i>	Cottony cushion scale	x				•		
<i>Lecanium corni</i>	European fruit lecanium				Europe	•		Broad-leaved trees
<i>Lepidosaphes ulmi</i>	Oystershell scale	x	x	x	Europe	•		Broad-leaved trees
<i>Metopoplax ditomoides</i>			x	x	Europe	•		
<i>Nezara viridula</i>				x		•		
<i>Periphyllus californiensis</i>					Asia	•		<i>Acer</i> spp.
<i>Periphyllus lyropictus</i>	Norway maple aphid	x	x	x	Europe	•		<i>Acer platanoides</i> and other <i>Acer</i> spp.
<i>Periphyllus testudinacea</i>					Europe	•		<i>Acer</i> spp.
<i>Physokermes piceae</i>	Spruce bud scale	x	x	x	Europe	•		<i>Picea</i> spp.
<i>Pineus strobi</i>	Pine bark aphid				Europe	•		<i>Pinus</i> spp.
<i>Quadraspidiotus perniciosus</i>	San Jose scale	x	x	x	Asia	•		Broad-leaved trees
<i>Raglius alboacuminatus</i>			x	x	Europe and the Mediterranean Basin	•		Mint (Lamiaceae) seeds
<i>Rhyarochromus vulgaris</i>	Dirt-colored seed bug			x	Europe	•		
<i>Schizolachnus pineti</i>					Europe	•		<i>Pinus</i> spp.
<i>Stephanitis pyrioides</i>	Azalea lace bug		x	x	Japan	•		<i>Azalea</i> spp., <i>Rhododendron</i> spp.
<b>Thysanoptera</b>								
<i>Taeniothrips inconsequens</i>	Pear thrips		x	x				
<b>Hymenoptera</b>								
<i>Agathis pumila</i>		x	x	x	Europe		•	<i>Coleophora laricella</i>
<i>Apanteles solitarius</i>					Europe		•	<i>Stilpnotia salicis</i>
<i>Caliroa cerasi</i>	Pear sawfly	x	x	x		•		

(continued)



**Table A3.3** (continued)

Scientific name	Common name	Occurrence			Origin	Introduction		Plant or animal host
		ID	OR	WA		Accidental	Intentional	
<i>Chrysocharis laricinellae</i>		x	x	x	Europe		•	<i>Coleophora laricella</i>
<i>Cladius grandis</i>				x	Eurasia	•		<i>Alnus</i> spp., <i>Populus</i> spp., <i>Salix</i> spp.
<i>Cladius gregarius</i>				x	Eastern North America	•		<i>Populus</i> spp.
<i>Craesus alniastri</i>				x	Europe	•		<i>Alnus</i> spp.
<i>Diprion similis</i>	Introduced pine sawfly			x	Europe	•		<i>Pinus</i> spp.
<i>Eupareophora parca</i>				x	Eastern North America	•		<i>Carya</i> spp., <i>Fraxinus</i> spp.
<i>Eriocampa ovata</i>	Alder woolly sawfly				Europe	•		<i>Alnus</i> spp.
<i>Fenusella nana</i>				x	Europe	•		<i>Betula</i> spp.
<i>Fenusa pusilla</i>	Birch leafminer	x	x	x	Europe	•		<i>Betula</i> spp.
<i>Fenusa ulmi</i>				x	Europe	•		<i>Ulmus</i> spp.
<i>Gilpinia hercyniae</i>	European spruce sawfly			x	Eurasia	•		<i>Picea</i> spp.
<i>Halidamia affinis</i>				x	Europe	•		<i>Gallium</i> spp.
<i>Heterarthrus nemoratus</i>				x	Eurasia	•		<i>Betula</i> spp.
<i>Heterarthrus vagans</i>				x	Eurasia	•		<i>Alnus</i> spp.
<i>Kaliofenusa ulmi</i>	Elm leafminer		x	x	Europe	•		<i>Ulmus</i> spp.
<i>Macrophya punctumalbum</i>				x	Europe	•		<i>Ligustrum</i> spp., <i>Syringa</i> spp., <i>Fraxinus</i> spp.
<i>Mesoleius tenthredinis</i>					Europe		•	<i>Pristiphora erichsonii</i>
<i>Metallus lanceolatus</i>				x	Europe	•		<i>Geum macrophyllum</i>
<i>Meteorus versicolor</i>					Europe		•	<i>Stilpnolia salicis</i>
<i>Monophadnus pallescens</i>				x	Europe	•		<i>Ranunculus</i> spp.
<i>Monostegia abdominalis</i>				x	Europe	•		<i>Glaux</i> spp., <i>Lysimachia</i> spp., <i>Anagallis</i> spp.
<i>Monsoma pulveratum</i>	Green alder sawfly	x	x	x	Europe, Asia Minor and North Africa	•		<i>Alnus</i> spp.
<i>Nematus lipovskyi</i>				x	Eastern North America	•		<i>Rhododendron</i> spp.
<i>Neodiprion sertifer</i>	European pine sawfly			x	Europe	•		<i>Pinus</i> spp.
<i>Polistes dominulus</i>	European paper wasp			x	Europe	•		Omnivore
<i>Pristiphora erichsonii</i>	Larch sawfly	x	x	x	Europe	•		<i>Larix</i> spp.
<i>Pristiphora geniculata</i>	Mountain ash sawfly			x	Europe	•		<i>Crataegus</i> spp., <i>Sorbus</i> spp.
<i>Pristiphora rufipes</i>				x	Central Europe	•		<i>Aquilegia</i> spp.
<i>Profenusa thomsoni</i>	Amber-marked birch leafminer			x	Eurasia	•		<i>Betula</i> spp.
<i>Trichiocampus viminalis</i>	Poplar sawfly	x	x	x	Europe	•		<i>Populus</i> spp., <i>Salix</i> spp.
<i>Vespula germanica</i>	German yellowjacket	x			Europe	•		Omnivore
<i>Xiphydria prolongata</i>	Willow wood wasp		x	x	Europe	•		Broad-leaved trees
<b>Lepidoptera</b>								
<i>Aethes rutilana</i>	Pale juniper webworm				Europe	•		<i>Juniperus</i> spp.
<i>Anarsia lineatella</i>	Peach twig borer	x	x	x		•		<i>Prunus</i> spp.
<i>Archips rosanus</i>					Europe	•		Broad-leaved trees

(continued)

**Table A3.3** (continued)

Scientific name	Common name	Occurrence			Origin	Introduction		Plant or animal host
		ID	OR	WA		Accidental	Intentional	
<i>Calophasia lunula</i>	Toadflax defoliating moth	x	x	x	Eurasia		•	Dalmatian and yellow toadflax, <i>Linaria</i> spp.
<i>Caloptilia negundella</i>	Boxelder leafroller				Europe	•		<i>Acer</i> spp.
<i>Caloptilia syringella</i>	Lilac leafminer				Europe	•		Lilac and <i>Fraxinus</i> spp.
<i>Cnephasia longana</i>	Omnivorous leaf-tier				Europe	•		<i>Pseudotsuga</i> spp. and broad-leaved trees
<i>Coleophora laricella</i>	Larch casebearer	x	x	x	Europe	•		<i>Larix</i> spp.
<i>Coleophora serratella</i>	Birch casebearer		x	x	Europe	•		<i>Betula</i> spp.
<i>Dichomeris marginella</i>	Juniper webworm	x	x		Europe	•		<i>Juniperus</i> spp.
<i>Enarmonia formosana</i>	Cherry bark tortrix		x	x	Eurasia	•		<i>Crataegus</i> spp., <i>Malus</i> spp., <i>Prunus</i> spp., <i>Pyrus</i> spp., <i>Sorbus</i> spp.
<i>Epinotia nanana</i>	European spruce needleminer		x	x	Europe	•		<i>Picea</i> spp.
<i>Homadaula anisocentra</i>	Mimosa webworm				Unknown	•		<i>Albizia</i> spp., <i>Gleditsia</i> spp.
<i>Leucoma salicis</i>	Satin moth	x	x	x	Europe	•		<i>Populus</i> spp., <i>Salix</i> spp.
<i>Leucoptera spartifoliella</i>					Europe		•	Scotch broom, <i>Cytisus scoparius</i>
<i>Lymantria dispar dispar</i> <sup>a</sup>	North American gypsy moth				Europe	•		Broad-leaved trees
<i>Lymantria dispar asiatica/japonica</i> <sup>a</sup>	Asian gypsy moth				Asia	•		Broad-leaved trees and conifers
<i>Ocnerostoma piniariellum</i>					Europe	•		<i>Pinus</i> spp.
<i>Operophtera brumata</i>	Winter moth		x	x	Europe	•		Broad-leaved trees and agricultural crops
<i>Pandemis cerasana</i>					Europe	•		Broad-leaved trees
<i>Rhyacionia buoliana</i>	European pine shoot moth	x	x	x	Europe	•		<i>Pinus</i> spp.
<i>Spilonota ocellana</i>	Eyespotted bud moth		x		Europe	•		<i>Quercus</i> spp. and other broad-leaved trees
<i>Synanthedon myopaeformis</i>	Apple clearwing moth			x	Europe, Mediterranean Basin	•		<i>Crataegus</i> spp., <i>Malus</i> spp., <i>Prunus</i> spp., <i>Pyrus</i> spp.
<i>Synanthedon scitula</i>	Eastern dogwood borer			x	Eastern North America	•		Broad-leaved trees esp. <i>Cornus</i> spp., <i>Carya</i> spp., <i>Malus</i> spp.
<i>Tyria jacobaeae</i>	Cinnabar moth		x	x	Europe		•	Tansy ragwort, <i>Senecio jacobaea</i>

<sup>a</sup>Detected repeatedly in the Northwest region, but eradicated. <sup>o</sup> Native range within the region

This list was compiled primarily from Furniss and Carolin (1977) with additions from Acheampong et al. (2016), Bai et al. (2002), Bellows et al. (1998), Childs and Swanson (2003), Doerr et al. (2008), Foote et al. (1993), Gerberg (1957), Hatch (1953, 1962), Hayes and Ragenovich (2001), Hitchcock et al. (2002), Idaho State Department of Agriculture (2013), Ivie (2002), Johnson (1998), Kruse et al. (2010), LaBonte et al. (2005), LaGasa (2006), LaGasa and Murray (2007), Lee et al. (2009, 2011), Looney et al. (2012, 2016a, 2016b), Mudge et al. (2001), Murray et al. (2012, 2013), Nugent (2005), Phillips (2002), Rabaglia et al. (2006, 2019), Rosetta (2013), Sabrosky and Reardon (1976), Seybold and Downing (2009), Sing et al. (2016), Vernon et al. (2001), USDA (1986, 2019), Washington Invasive Species Council (2012), White (1982), and Winston et al. (2014a, b)

US Department of Agriculture 2009. National Agriculture Library. Species Profiles. Accessed online at <http://www.invasivespeciesinfo.gov/plants/main.shtml>

Other terrestrial invertebrates found in Oregon and Washington include *Amyntas agrestis* (Crazy snake/Asian jumping earthworm), giant African snail (*Achatina fulica*), vineyard snail (*Cermeuella virgata*), white garden snail (*Theba pisana*), heath snail (*Xerolenta obvia*)

**Table A3.4** Priority non-native invasive aquatic invertebrates (noninsect) of the Northwest region

Scientific name	Common name	ID	OR	WA	R6 <sup>a</sup>
<i>Potamocorbula amurensis</i>	Asian clam	x	x		x
<i>Radix auricularia</i>	Big-eared radix				x
<i>Eriocheir sinensis</i>	Chinese mitten crab		x	x	
<i>Cipangopaludina chinensis</i> , <i>C. japonica</i>	Chinese mystery snail, Japanese mystery snail	x			x
<i>Orconectes</i> spp., <i>O. virilis</i> ; <i>Procambarus</i> spp.	Crayfish (red swamp, rusty, ringed, virile, marbled, signal, red claw, yabby, marron)	x		x	x
<i>Carcinus maenas</i>	European green crab			x	
<i>Potamopyrgus antipodarum</i>	New Zealand mud snail	x		x	x
<i>Philine auriformis</i>	New Zealand sea slug		x		
<i>Bythotrephes longimanus</i> [ <i>cederstroemi</i> ], <i>Cercopagis pengoi</i>	Water fleas	x	x		
<i>Dreissena polymorpha</i> , <i>D. rostriformis bugensis</i>	Zebra/quagga mussels	x	x	x	x

<sup>a</sup>R6 = USDA Forest Service Region 6 (OR, WA) priority invasive species

**Table A3.5** Priority non-native invasive vertebrates of the Northwest region

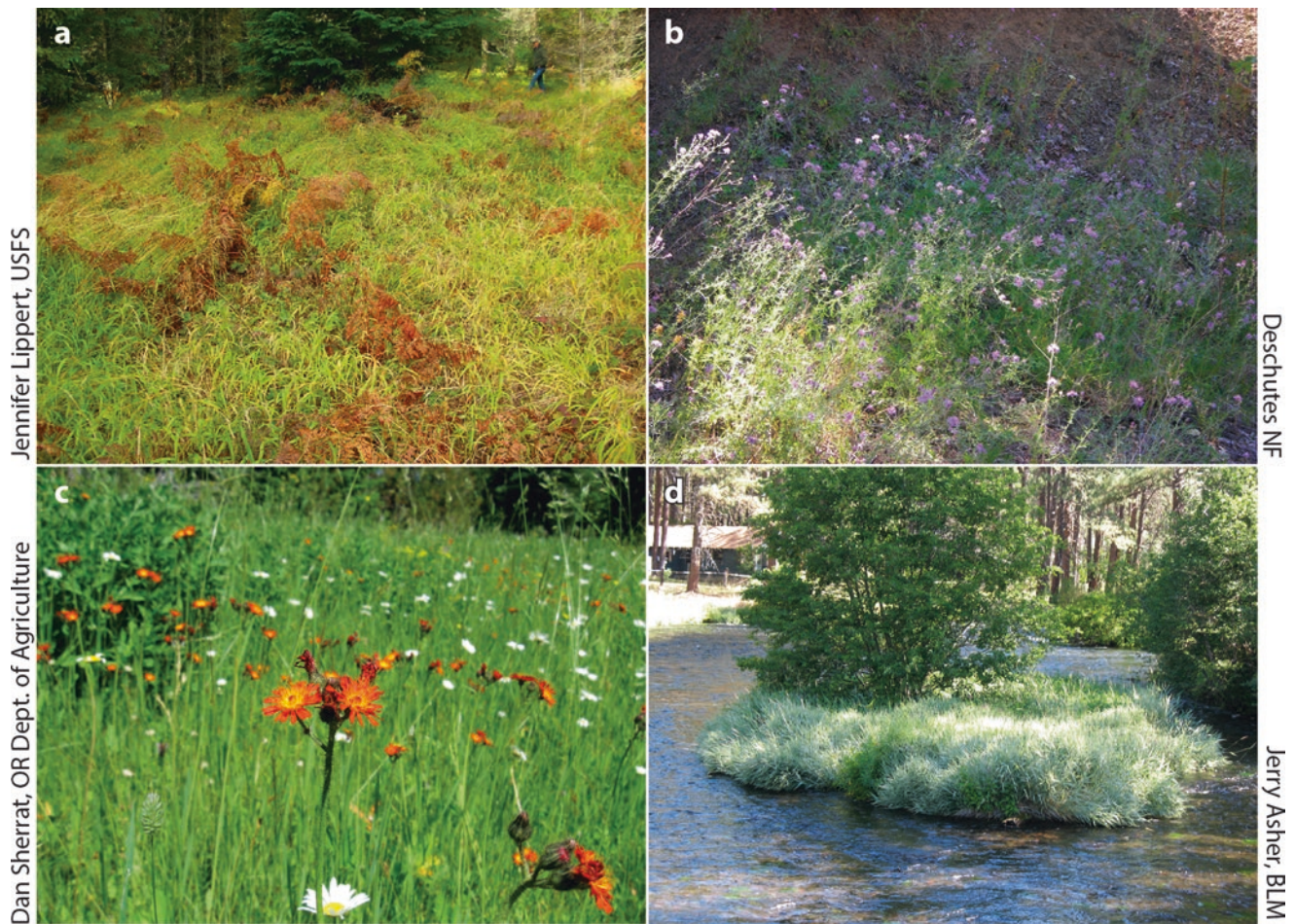
Scientific name	Common name	ID	OR	WA	R6 <sup>a</sup>
<b>Aquatic vertebrates</b>					
<i>Lithobates catesbeianus</i> ( <i>Rana catesbeiana</i> )	American bullfrog	x		x	x
<i>Amia calva</i>	Bowfin	x			
<i>Hypophthalmichthys</i> spp., <i>Mylopharyngodon piceus</i>	Carp (Asian, black, big head, diploid grass, silver)	x	x	x	
<i>Salmo salar</i>	Atlantic salmon		x	x	
<i>Didemnum vexillum</i>	Didemnum tunicate			x	
<i>Chelydra serpentina serpentina</i>	Eastern snapping turtle	x	x		
<i>Lepisosteidae</i> spp.	Gar	x			
<i>Neogobius melanostomus</i> , <i>Rhinogobius brunneus</i> , <i>Tridentiger bifasciatus</i>	Goby	x	x		
<i>Notemigonus crysoleucas</i>	Golden shiner		x		
<i>Acipenser medirostris</i>	Green sturgeon	x			
<i>Leuciscus idus</i>	Ide	x			
<i>Esox</i> spp.	Muskellunge/northern pike		x		
<i>Serrasalmus</i> spp., <i>Rosseveltella</i> spp., <i>Pygocentrus</i> spp.	Piranhas	x			
<i>Trachemys scripta elegans</i>	Red-eared slider	x			
<i>Taricha granulosa</i>	Rough-skinned newt	x			
<i>Scardinius erythrophthalmus</i>	Rudd	x			
<i>Gymnocephalus cernuus</i>	Ruffe	x	x		
<i>Channa</i> spp.	Snakehead	x	x	x	
<i>Dorosoma petenense</i>	Threadfin shad (yellowtails)		x		
<i>Clarias</i> spp.	Walking catfish	x			
<b>Terrestrial vertebrates</b>					
<i>Sus scrofa</i>	Feral swine		x	x	x
<i>Cygnus olor</i>	Mute swan		x		
<i>Myocastor coypus</i>	Nutria	x	x	x	x

<sup>a</sup>R6 = USDA Forest Service Region 6 (OR, WA) priority invasive species

wildlife (Edwards 1999). Toxic algal blooms are enhanced by high water temperatures and fertilizer runoff. The diatom rock snot (didymo) is actually native to the Pacific Northwest, but in the mid-1980s, it became more prolific in its distribution and began to impact recreational activity.

Three invasive terrestrial plants—Himalayan blackberry (*Rubus armeniacus*), Japanese knotweed (*Fallopia japonica*), and giant hogweed (*Heracleum mantegazzianum*)—are problematic in both upland and riparian environments in the Marine West Coast Forest ecoregion (Fig. A3.3a). These

very abundant species were introduced intentionally as ornamentals or crops. They tend to shade out smaller native plants, reducing plant diversity and limiting habitat and food sources for both birds and native wildlife. Native to Western Europe, Himalayan blackberry is an evergreen woody vine whose canes have large stiff prickles and form dense thickets. Its growth form approximates a shrub in terms of how animals use it and its height in the environment. The plant is very common in the Northwest region, providing prolific berries that are collected recreationally



**Fig. A3.3** Invasive plants in the Northwest region include (a) false brome (*Brachypodium* spp.), on the Willamette National Forest, OR; (b) roadside infestation of spotted knapweed (*Centaurea stoebe*) on the Deschutes National Forest, OR; (c) orange hawkweed (*Hieracium*

*aurantiacum*) on the Willamette National Forest, OR; (d) ribbon grass, a striped horticultural variety of reed canary grass (*Phalaris arundinacea*), on an islet in the Metolius River, Deschutes National Forest, OR

and commercially (Stannard 2014). The thickets, mounds, or banks can completely and permanently exclude other plant species and pose a potential fire hazard. Japanese knotweed, native originally to Asia, but introduced to the Netherlands in 1829 (The Knotweed Company Ltd. 2018), and giant hogweed, native to the Caucasus region of Eurasia, can grow as tall as 15–20 ft. and spread rapidly. Japanese knotweed is known globally as one of the world’s most destructive invasive species because its large underground root system can damage structures, walls, and architectural sites, as well as reduce channel capacity. Giant hogweed is considered a public health hazard because it causes a phototoxic reaction when animal skin is exposed to sap and ultraviolet radiation.

Many of the invasive plants in the Northwest region were imported initially as ornamentals to either the area, or, more generally, North America. Some examples are English ivy (native to the United Kingdom), old man’s beard (native to the United Kingdom), orange hawkweed (native to Europe),

yellow archangel (native to Europe/Western Asia), garlic mustard (native to Europe and Asia), Scotch broom (native to Europe and North Africa), purple loosestrife (native to Europe and Asia), and saltcedar (*Tamarix* spp., native to Europe and Asia) (Table A3.1). Saltcedars (see Southwest region summary) are riparian shrubs or small trees that are aggressively invasive. Populations of these plants are prevalent in the warm and dry riparian corridors of the Blue Mountains and Cold Basins Ecoregions of eastern Oregon and Washington along the Owyhee, Snake, and John Day Rivers, and in the Columbia Plateau Ecoregion (Fig. A3.2) (Thorson et al. 2003). These riparian trees are known to decrease stream flows, lower biodiversity, and create salinization issues. Purple loosestrife is a wetland forb that can rapidly establish and replace native vegetation with a dense, homogeneous stand that reduces local biodiversity, endangers rare species, and provides little value to wildlife. English ivy and old man’s beard are both vigorous creepers that not only cover terrestrial surfaces but will climb high into trees

and smother competing vegetation. Garlic mustard was initially introduced to the East Coast of North America as a medicinal herb, but it has spread through forest understories and competes with native species. Orange hawkweed and yellow archangel are also understory shade plants that spread vigorously and smother competing native plants.

Many of the remaining invasive upland plants (Table A3.1) are weed species that can become dominant in meadow, range, or forest habitats, often creating monocultures. Most of these species were introduced accidentally, although some were introduced as ornamentals (i.e., Dalmatian toadflax). They can severely degrade the quality of range habitats for cattle and other domestic and native ungulates (i.e., cheatgrass, knapweeds). Although they are not toxic specifically to livestock as are some other species (i.e., some knapweeds, tansy ragwort), they can cause mechanical damage to the animals. In addition, invasive annual grasses like cheatgrass, medusahead, and ventenata are considered highly detrimental in the interior Western United States because they have the potential to alter wild-fire regimes in some areas (see Chap. 2, Box 2.1; and the Southwest region summary for additional discussion of these issues). These species are problematic largely in the Blue Mountains and Cold Basins Ecoregions of the Northwest region (Fig. A3.2). Western juniper (*Juniperus occidentalis*) is an example of a native species that is often managed as an invasive species due to emergent concerns of its ecological and economic impact in the interior ecoregions, including degrading habitat for the threatened greater sage-grouse (*Centrocercus urophasianus*). Juniper management may be problematic, however, because removal can encourage exotic weedy grasses like cheatgrass to invade cut areas.

### Plant Pathogens

In the Pacific Northwest, invasive pathogens (fungi, water molds, bacteria, nematodes, and viruses) are a significant forestry problem for wood production as well as for urban and rural landscaping (Table A3.2). Two of the most damaging invasive species are in the genus *Phytophthora*, “water molds” in the kingdom Straminipila (formerly Chromista), which includes aquatic organisms such as diatoms and kelp (Dick 2001). These fungi-like microbes thrive in wet conditions, so hydric or mesic native ecosystems west of the Cascade Mountains and well-irrigated nurseries of the Northwest region are conducive for their growth and reproduction. Notably, all three of the destructive pathogens described below were introduced on imported nursery stock, illustrating the significance of the nursery pathway (Liebhold et al. 2012) in the Northwest region.

**Sudden Oak Death** (*Phytophthora ramorum*) First discovered in this region in coastal forests of southwest Oregon in

2001, *Phytophthora ramorum* causes sudden oak death and other diseases. It is lethal to tanoak (*Notholithocarpus densiflorus*) and threatens this species throughout its range in Oregon (Kanaskie et al. 2017) (see Chap. 2, Box 2.5; Chap. 6, Sect. 6.4.2; Chap. 7, Sect. 7.4.2; and the Southwest regional summary for additional discussion of this pathogen). The pathogen also infects *Rhododendron*, *Viburnum*, and other important plant species in Pacific Northwest horticultural nurseries so the Federal and State quarantines affect the nursery industry throughout Oregon and Washington as well as forestry interests in part of Oregon’s Curry County. The pathogen was inadvertently introduced to Oregon forests via infested nursery stock that originated in either California or Oregon (Kamvar et al. 2015).

From 2001 to 2012, a Federal and State interagency team attempted to eradicate the pathogen from Oregon, supported by the Oregon quarantine that required destruction of infected and nearby uninfested host plants. Although eradication treatments eliminated this disease from many infested sites, the pathogen continued to spread slowly. The wildland quarantine area has expanded from 22 km<sup>2</sup> (9 mi<sup>2</sup>) in 2001 to 1333 km<sup>2</sup> (515 mi<sup>2</sup>) in 2017, affecting over 30% of Curry County. Hundreds of thousands of tanoaks have died from *P. ramorum* in southwest Oregon (Kanaskie et al. 2013).

In 2015, a second significant introduction of the pathogen was detected in this region—the first find of the *P. ramorum* EU1 lineage (European Union Lineage One) in a US forest. The detection (on a tanoak tree) was approximately 1.6 km (1 mi) north of a small private nursery (now closed) near the Pistol River which, once again, underscores the importance of the nursery pathway for long-distance invasive pathogen movement (Grünwald et al. 2016). In Europe, the EU1 lineage of *P. ramorum* damages and kills several conifer species of significance to the Northwest region, including larch (*Larix* spp.), Douglas fir (*Pseudotsuga menziesii*), western hemlock (*Tsuga heterophylla*), and grand fir (*Abies grandis*) (Webber et al. 2010). In 2016, the EU1 strain was detected in an Oregon rural forest for a second time, less than 1.6 km (1 mi) south of the original 2015 detection (California Oak Mortality Task Force 2016).

Current management of *P. ramorum* focuses on early detection and rapid response. To support the overall goal of containment, new infections outside of the generally infested area are eradicated. With detection of new infections on the rise, management strategies will need to include a general, integrated pest management approach to minimize the impact to urban and rural forests susceptible to *P. ramorum* in the Northwest region.

**Port-Orford-Cedar Root Disease** (*Phytophthora lateralis*) Port-Orford-cedar or Lawson’s cypress

(*Chamaecyparis lawsoniana*) is a large, beautiful conifer, endemic to southwestern Oregon and northwestern California (Zobel 1990). It is planted frequently in urban areas of the Pacific Northwest. However, the introduction and spread of the exotic fungus-like pathogen *Phytophthora lateralis* has caused high mortality levels on high-risk sites in old-growth Port-Orford-cedar forests (Fig. A3.4) and in ornamental landscapes (Hansen 2011) (see the Southwest regional summary for additional discussion of this pathogen).

The pathogen was introduced on infected nursery stock near Seattle, WA, in the 1920s and moved southward on horticultural plantings until it reached native southwest Oregon forest stands in 1952 (Hansen 2011). Port-Orford-cedar and Pacific yew (*Taxus brevifolia*) are the only native North American tree species known to be susceptible to *P. lateralis* (DeNitto and Kliejunas 1991), but the pathogen can persist in soil for more than 5 years (Hansen and Hamm 1996). The pathogen moves via transport of infected nursery plants, infested soil, and contaminated runoff water, and disease spread is correlated with proximity to roads and rivers (Hansen et al. 2000).

Several management practices are recommended to minimize the impact of Port-Orford-cedar root disease. The best management strategy involves a combination of appropriate techniques tailored for use in a specific site or landscape. These include planting resistant trees, closing forest roads during the wet season, limiting activities involving heavy equipment (e.g., timber harvesting, road maintenance) to the summer dry season, washing vehicles before they enter uninfested areas, paving road surfaces, and

using only pathogen-free water for dust abatement and fire-fighting (Betlejewski et al. 2011; Hansen et al. 2000).

**White Pine Blister Rust** (*Cronartium ribicola*) In the Northwest region, white pine blister rust (*Cronartium ribicola*) threatens, damages, or kills western white pine (*Pinus monticola*), sugar pine (*P. lambertiana*), and other high-elevation white (five-needle) pines, i.e., whitebark (*P. albicaulis*) and limber (*P. flexilis*). The future of whitebark pine in the Pacific Northwest is a serious concern due to *C. ribicola* infection and the effects of colonization by mountain pine beetle (*Dendroctonus ponderosae*), wildfire, climate change, and other factors (Aubry et al. 2008).

*Cronartium ribicola* is one of the most damaging invasive pathogens in US forests and parks (Benedict 1981; Boyce 1938; Vitousek et al. 1996) (see also Chap. 7, Sect. 7.3.2; and the Southwest regional summary for additional discussion of this pathogen). The rust, which is native to Asia, was introduced to Western North America around 1910, on nursery stock from France imported into Vancouver, British Columbia, Canada (Liebhold et al. 2012). The 1912 Plant Quarantine Act was prompted by its introduction, with US Quarantine No. 1 prohibiting import of five-needle pines (Maloy 1997).

In the Northwest region, surveys for *C. ribicola* began around Puget Sound, WA, in 1917, with the first detection in Washington reported on black currant (*Ribes* spp.) and western white pine in 1921 (Detwiler 1922, as cited by Geils et al. 2010). Over the next several decades, the pathogen had spread on that host throughout much of Washington and

**Fig. A3.4** Port-Orford-cedar killed by *Phytophthora lateralis* on the Gold Beach Ranger District, Rogue River-Siskiyou National Forest. (Figure courtesy of Ellen Goheen, USDA Forest Service Northwestern Region, Forest Health Protection)



Oregon, but was also detected on whitebark pine (Geils et al. 2010).

This rust fungus is an obligate, biotrophic (requires a live host) pathogen with a complex life cycle that requires an alternate host, primarily currants, along with the white pine host for the disease to occur (Geils et al. 2010). The Civilian Conservation Corps undertook extensive efforts to control the disease by removing *Ribes* species in the 1930s (Benedict 1981), but contemporary management favors resistance breeding programs (Kegley et al. 2012; Schoettle et al. 2012), as well as pruning young stands to minimize infections in the lower crowns and protecting larger trees from other mortality agents, such as fire and mountain pine beetle (Goheen and Goheen 2014) (see also Chap. 7, Table 7.2).

### Insects

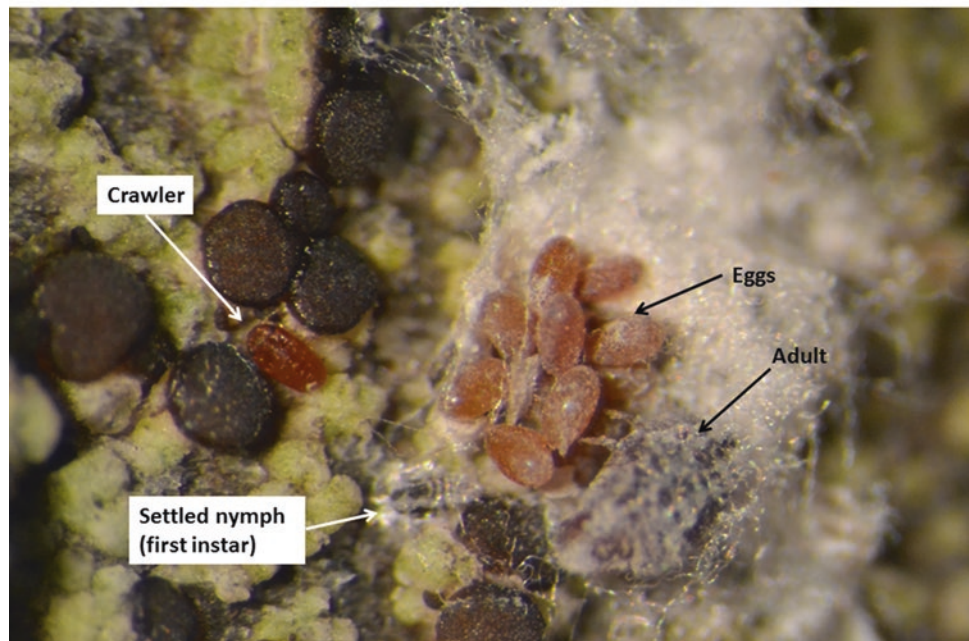
A late twentieth-century summary of extra-continental forest insects known to have been established in Western North America suggested that 75 species had been introduced and that 17 of these were purposeful introductions—releases as biological agents for the control of insects or noxious plants (Furniss and Carolin 1977). Many of these historical introduced species, as well as some new species, have established populations in the Northwest region (Table A3.3) (Furniss and Carolin 1977; Hayes and Ragenovich 2001; LaBonte et al. 2005; Mudge et al. 2001; Rabaglia et al. 2019; Seybold and Downing 2009). A large number of additional purposeful introductions have been made in the Northwest region for the biological control of urban and wildland forest

pests (Bellows et al. 1998) or invasive plants (Sing et al. 2016; Winston et al. 2014a, b). Though not invasive species in the strict sense, we include these purposefully introduced taxa (Table A3.3) to maintain continuity with the original summary by Furniss and Carolin (1977). The European (North American) gypsy moth (*Lymantria dispar dispar*) was included in the historical summary (Furniss and Carolin 1977), but the Asian gypsy moth (*Lymantria dispar asiatica/japonica*) has been detected recently in the vicinity of the coastal ports of the Northwest region (see below). Furthermore, this region has sustained new invasions by pest insects that threaten valuable horticultural crops. Several insect species that already damage or threaten to damage forest trees and horticultural crops in the Northwest region are highlighted below.

**Balsam Woolly Adelgid (*Adelges piceae*)** A tiny (about 1-mm-long) sap-sucking insect, the balsam woolly adelgid (*Adelges piceae*) (Fig. A3.5), is probably the most prominent invasive pest of forest trees in the Northwest region (Ragenovich and Mitchell 2006) (see Southeast and Caribbean regional summary for further discussion of this species). It first appeared on the West Coast in the late 1920s (Annand 1928; Keen 1952). In this region, it infests primarily subalpine fir, Pacific silver fir (*Abies amabilis*), and grand fir (*A. grandis*) (Ragenovich and Mitchell 2006). Other susceptible hosts include noble fir (*A. procera*), Shasta fir (*A. magnifica*), and white fir (*A. concolor*). Subalpine fir and Pacific silver fir are impacted to a greater degree in

**Fig. A3.5** Life stages of the balsam woolly adelgid (*Adelges piceae*). (Figure courtesy of USDA Forest Service Northwestern Region, Forest Health Protection)

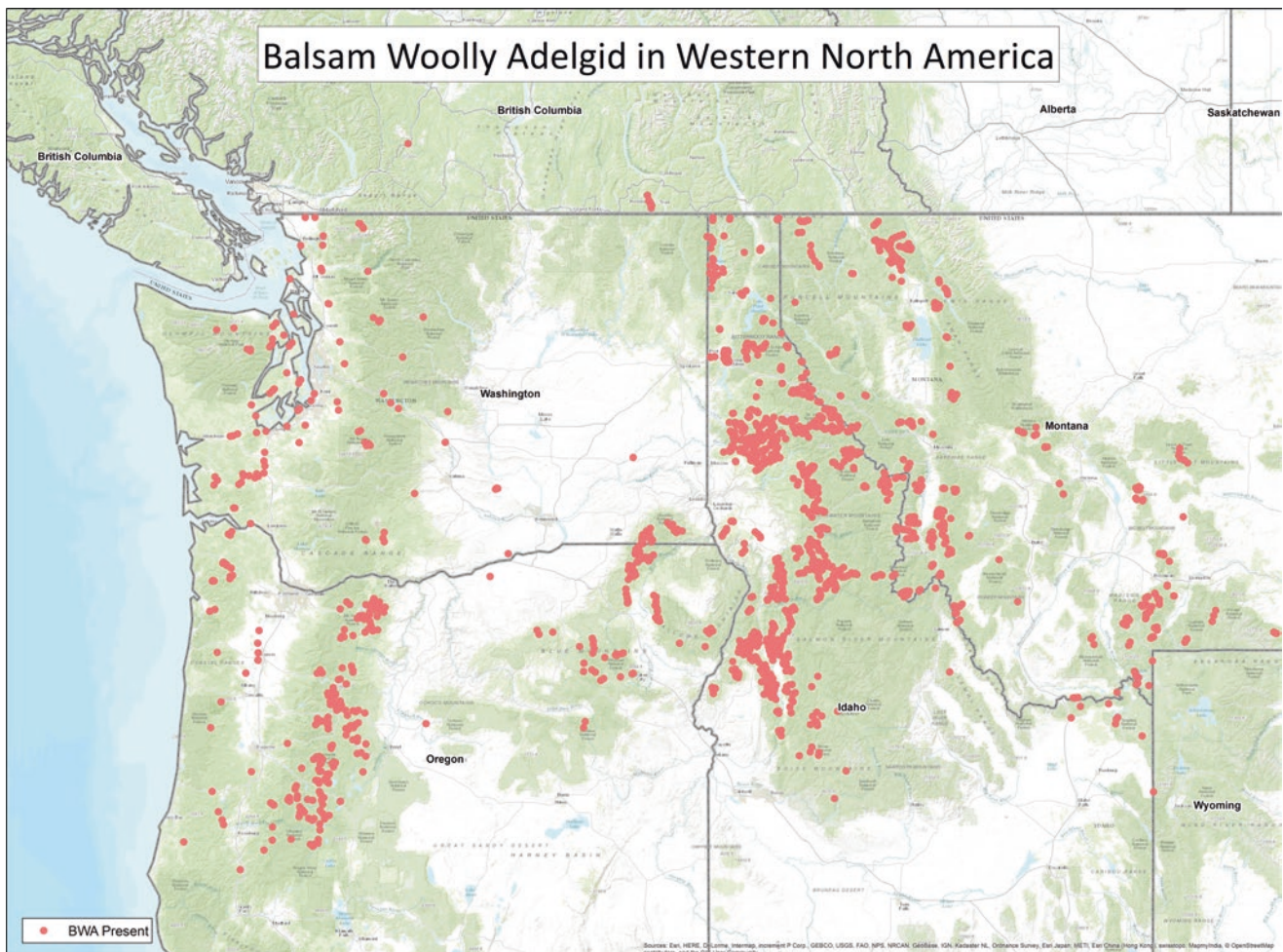
### Life Stages of Balsam Woolly Adelgid



mountainous areas, whereas grand fir is impacted to a greater degree in lowland valleys. The symptoms of attack by balsam woolly adelgid, especially on young fir trees, include buds failing to open and twigs becoming enlarged at the nodes and around the buds. Feeding by the insect results in stem and twig injury whereby the adelgid injects a substance into the inner bark, resulting in abnormal cell division and differentiation in the inner bark and newly formed wood. In the 1950s, a major balsam woolly adelgid outbreak killed or “seriously weakened” over 3,539,606 m<sup>3</sup> (1.5 billion board feet) of mature Pacific silver fir trees across 161,874 ha (400,000 ac) in southwestern Washington (Johnson et al. 1963). The life cycle of this species (and related adelgids) is unusual (Havill and Foottit 2007; Havill et al. 2011). North American populations are composed entirely of females; thus reproduction is parthenogenetic (i.e., without mating and fertilization). Adults are dark purple to black, nearly spherical, and wingless (Fig. A3.5). They produce a thick

mass of a white waxy wool-like material that covers the body and protects the adult and her eggs (Fig. A3.5).

The range of balsam woolly adelgid has been expanding in the eastern portion of the Northwest region (Fig. A3.6) (Gast et al. 1990; Hrinkevich et al. 2016; Lowrey and Davis 2017). In apparent synergy with climate change effects, the invasive threat here is to high-elevation stands of subalpine fir, which play an important ecological function in regulation of snow melt and wildlife habitat and have a modest, yet measureable, timber value (Alexander 1987; Steele et al. 1981). Although 20 predaceous insects were introduced to the West for the biological control of balsam woolly adelgid (Table A3.3) (Bellows et al. 1998; Mitchell and Wright 1967), only 5 species were considered established in 1998 (*Aphidoletes thompsoni*, *Pullus impexus*, *Laricobius erichsonii*, *Cremifania nigrocellulata*, and *Leucopis obscura*), and Bellows et al. (1998) characterized the



**Fig. A3.6** Distribution of the balsam woolly adelgid (*Adelges piceae*) in the Northwest region of the United States and Canada as of spring 2016 based on georeferenced collection records and posi-

tive identification of specimens. (Figure courtesy of Amy Gannon, Montana Department of Natural Resources and Conservation, Missoula, MT)



effectiveness of the biological control as “variable and somewhat limited.” It is unknown whether any of these species (released in Oregon, Washington, and British Columbia) may have spread into the expanding range of balsam woolly adelgid in the eastern portion of the Northwest region (Lowrey and Davis 2017).

**Hemlock Woolly Adelgid (*Adelges tsugae*)** Hemlock woolly adelgid (*Adelges tsugae*) is an economically and ecologically important invasive pest of hemlock trees (*Tsuga* spp.) in eastern North America (McClure et al. 1996) that has only had minor impacts to ornamentals and other plantings in the Northwest region (Furniss and Carolin 1977) (see the Southeast and Caribbean regional summary for further discussion of this species). Western hosts include western hemlock, *T. heterophylla*, and mountain hemlock (*T. mertensiana*), though the latter has only been colonized in adventive plantings (Havill et al. 2016). Recent molecular analyses (microsatellite and mitochondrial DNA sequences) have revealed that populations of hemlock woolly adelgid in the Northwest region are a consequence of an ancient colonization event from an ancestor whose host was an Asian hemlock (*T. sieboldii*) (Havill et al. 2016). Colonization of Western North America was estimated to have occurred prior to the last glacial period by adelgids directly ancestral to those in southern Japan, perhaps carried by birds. Havill et al. (2011) report that the earliest North American specimens were collected in 1907 from South Bend, WA (US National Collection of Insects, Beltsville, MD). Other early records from the West include a report of damage to western hemlocks in Vancouver, British Columbia (Chrystal 1916), and specimens collected in Oregon and California that were used to formally describe hemlock woolly adelgid as a new species (Annand 1924). Populations of adelgids in the Northwest Region have served as important sources of natural enemies for the biological control program for hemlock woolly adelgid in the Eastern United States (McClure 2001; Reardon et al. 2004). Two non-native species in this program, a beetle, *Laricobius erichsonii*, and a fly, *Leucopis obscura*, were introduced originally into the Northwest region to control hemlock woolly adelgid (Furniss and Carolin 1977). However, a native species of beetle, *Laricobius nigrinus*, and two native species of flies, *Leucopis argenticollis* and *Leucopis piniperda*, have been recognized as predators of hemlock woolly adelgid in Idaho (*L. nigrinus* only), Oregon, and Washington and were moved subsequently by specialists to the Eastern United States for biological control (Kohler et al. 2008, 2016; Mausel et al. 2011).

An ensemble of other sapsucking insects has invaded the Northwest region and has, at times, threatened the health of native trees (Table A3.3). These include larch woolly aphid,

eastern spruce gall aphid, spruce aphid, European elm scale, European birch aphid, and pine bark aphid.

**Larch Casebearer (*Coleophora laricella*)** Larch casebearer (*Coleophora laricella*) is a small moth whose larvae feed in the needles of western larch (*Larix occidentalis*), in the Northwest region. The species was introduced from Europe first to the Eastern United States in 1886 (Bellows et al. 1998) and detected in western larch near St. Maries, ID, in 1957 (Tunnock and Ryan 1985). In the 1970s, it caused significant damage to western larch in the Blue Mountains (OR and WA). Its invaded range in this region is now coincident with that of western larch, which is present in all three States of the Northwest region, as well as in Montana and southern Canada (Tunnock and Ryan 1985). Beginning in the 1960s, ten parasitoid wasps were introduced into the region from eastern North America, Europe, and Japan (Bellows et al. 1998); two of the parasitoids, *Agathis pumila* and *Chrysocharis laricinellae*, established populations in the Blue Mountains and appeared to significantly lower the population density of larch casebearer to the point that “the species is no longer considered a pest in the West” (Bellows et al. 1998; Ryan 1997). However, Hayes and Ragenovich (2001) suggested that the level of control of larch casebearer in eastern Oregon relaxed in the late 1990s such that insect population densities and western larch defoliation increased. A subsequent survey in this area for both larch casebearer and the two principal parasitoids revealed that both the defoliator and the introduced natural enemies were widespread, that parasitism rates ranged from 1.8% to 53.4%, and that moth population density was negatively correlated with percentage parasitism by *A. pumila* (Shaw and Oester 2010).

**Gypsy Moths (*Lymantria* spp.)** Larvae of two non-native invasive moths are a constant and recurring threat for introduction and establishment in the Northwest region. These defoliators are the European (North American) gypsy moth (*Lymantria dispar dispar*) and the Asian gypsy moth (*Lymantria asiatica/japonica*) (see Chap. 2, Sect. 2.3; Chap. 7, Sect. 7.4.1; and the Southwest and Southeast and Caribbean regional summaries for additional discussion of this insect complex). Potential sources of European gypsy moth include populations from northeastern and North central North America (introduced and established first in Medford, MA, in 1869) and ancestral locations in Europe; sources of Asian gypsy moth include Siberia, the Russian Far East, Korea, China, and Japan. The Canadian province of British Columbia has also experienced frequent introductions and attempted eradications of these moths (Myers et al. 2000). These ecologically and behaviorally distinct subspecies were distinguished initially by mitochondrial and nuclear DNA sequencing techniques, including microsatellite DNA analy-

sis (Bogdanowicz et al. 1993, 1997, 2000). Other workers have used more sophisticated approaches and methodology, including real-time multiplex PCR (Djoumad et al. 2017; Islam et al. 2015; Stewart et al. 2016). With its broader host range (that includes conifers) and the flight capacity of females, Asian gypsy moth is considered a greater threat to the forests of the Western United States. Other potentially invasive moth defoliators that threaten forest trees in the Northwest region are the nun moth (*L. monacha*), the pink gypsy moth (*L. mathura*), and the Siberian moth (*Dendrolimus superans sibiricus*) (Hayes and Ragenovich 2001). Three other *Lymantria* species in Japan are also recognized as threats to Northwest region forests, *L. albescens*, *L. umbrosa*, and *L. postalba*, which are all listed as quarantine pests by the North Atlantic Plant Protection Organization (NAPPO).

Increased trade with the Russian Far East and other parts of Asia has greatly increased the frequency of encounters that regulatory entomologists face with Asian gypsy moth and other lymantriid moths. In 1990 and 1991, an introduced population of Asian gypsy moth was detected in pheromone-baited traps primarily around Tacoma, WA (Bogdanowicz et al. 1993), which resulted in a 47,146-ha (116,500-ac) eradication program. In a similar scenario in 2015, trap captures of ten Asian gypsy moth males in Washington (primarily in the southern half of the Puget Sound area) and two Asian gypsy moth males near Portland, OR, revealed that an incipient population of Asian gypsy moth with gravid adult females may have occurred on the U.S. mainland. Potential pathways included egg masses or pupae attached to ship cargo from Asia and then moved inland as the cargo was offloaded or dispersal of young larvae or adults from cargo on board or from the ship superstructure to sites on shore. These pathways are likely as Asian gypsy moth egg masses are intercepted annually on ships and cargo in Washington and Oregon ports. One male Asian gypsy moth has been trapped as far inland as northern Idaho (Kootenai County), with speculation that the insect was transported on a shipping container along a nearby rail line (Lech and Livingston 2004; Pederson et al. 2004). To mitigate introductions, USDA entomologists have long pursued a cooperative assistance program with their foreign counterparts in and around ports in the Russian Far East, Japan, Korea, and China to detect and manage these potential invaders (Freyman 2015; Humble et al. 2013; USDA 1993, 2016). To further reduce interceptions of Asian gypsy moth and other lymantriids in North America, NAPPO developed Regional Standards for Phytosanitary Measures (RSPM 33: Guidelines for Regulating the Movement of Vessels from Areas Infested with the Asian Gypsy Moth).

In response to the 2015 detection of Asian gypsy moth, a technical working group proposed a combination of insecticide treatment and delimitation trapping in the areas

where moths were detected. Three- and six-mile (4.8- and 9.7-km) radius zones were established around each of six locations. A microbially based material, *Bacillus thuringiensis* var. *kurstaki* (Btk), was applied aerially three times within the designated treatment areas of more than 4047 ha (10,000 acres) in Washington and 2833 ha (7000 ac) in Oregon. Approximately 11,000 delimitation traps were placed in Washington, and approximately 3100 traps were placed in Oregon. The estimated cost for the 2016 Asian gypsy moth eradication program was \$5 million. Detections of Asian gypsy moth in the Northwest region are continually being made against a backdrop of detections of European gypsy moth, presumably from introductions from eastern North America. In 2015, a population of European gypsy moth found in the Seattle area raised the concern that this population might mix with the contemporaneously detected Asian gypsy moth populations.

**Sawfly Defoliators (Hymenoptera, Symphyta)** A major survey for native and non-native invasive sawflies has revealed a large number of new defoliators in Washington and Oregon (Table A3.3) (Looney et al. 2012, 2016a, b). At least 20 species were found when field trap catches or museum collections were evaluated, and these were primarily free feeding or leaf-mining forms on hardwoods (alder, birch, elm, etc.), shrubs, and herbaceous vegetation. Many of the species were originally from Europe, but had first invaded eastern North America before they were introduced into the Northwest region (Looney et al. 2016b). Some key forest pests of note are the green alder sawfly (*Monsoma pulveratum*), which has also been damaging alder in Alaska (Kruse et al. 2010), the introduced and European pine sawflies (*Diprion similis* and *Neodiprion sertifer*, respectively), the European spruce sawfly (*Gilpinia hercyniae*), and various leafminers on birch (*Fenususa pusilla*, *Fenusella nana*, and *Profenususa thomsoni*). The ultimate impact of these new species to the region—mostly collected since the mid-2000s—is unknown, but the local biology, feeding behavior, and interactions within this assemblage of invading defoliators bear future scrutiny.

**Spotted Wing Drosophila (*Drosophila suzukii*)** The spotted wing drosophila (SWD) (*Drosophila suzukii*) was first found in August 2008 in the US mainland in California (Hauser 2011). Infestations were reported soon thereafter in 2009 in Oregon and Washington (Lee et al. 2011) and in 2012 in Idaho. While SWD is a primary economic concern in blackberry, blueberry, cherry, and raspberry crops, this insect species has a wide host range that includes many wild and ornamental hosts, which enables it to persist in woodland areas (Kenis et al. 2016; Lee et al. 2015). In a 2-year study at 35 farms, SWD were captured earlier in farms that had more woodland area surrounding the farm (Pelton et al. 2016).

These woodland habitats often contained wild blackberry and wild cherry in mid-summer and other fall-bearing fruit that may provide SWD with a late-season host after the crop has been harvested. A study that marked wild blackberry borders revealed that SWD moved from there and into the raspberry crop in the summer (Klick et al. 2015). Within agricultural areas, SWD is primarily managed with insecticides (Bruck et al. 2011). Classical biological control agents for SWD from Asia are being evaluated and are still under quarantine. Endemic parasitoids, such as the pupal parasitoid *Pachycrepoideus vindemiae* and the larval parasitoid *Leptopilina heterotoma*, have been found to attack sentinel SWD in mixed farm, raspberry, blueberry, and riparian habitats of Oregon (Miller et al. 2015). Although parasitism counts were low, conserving or augmenting these biological control agents may lead to greater pest suppression in unmanaged areas.

**Brown Marmorated Stink Bug (*Halyomorpha halys*)** The brown marmorated stink bug (BMSB) (*Halyomorpha halys*) was first found in Portland, OR, in 2004 and in Vancouver, WA, in 2010 (ODA 2010; Wiman and Lowenstein 2017). As with the invasion process in the East Coast, BMSB was first a nuisance pest reported by homeowners and then appeared in agricultural crops and spread to every county in the Willamette Valley of Oregon, Hood River, and southern Oregon. By 2013, economic losses from BMSB were reported in the northern Willamette Valley (Wiman and Lowenstein 2017). Although BMSB is a primary economic concern to stone fruit, vegetables, and field crop and nut tree growers (Rice et al. 2014), it also has a wide host range that includes many trees: elm, hawthorn, holly, linden, maples, and tree of heaven (Bergman et al. 2014; Leskey and Nielsen 2018). Many ornamental hosts are partial hosts meaning that one or more of BMSB's life stages can develop on the host but not all stages (Bergman et al. 2014, 2016), and these partial hosts may facilitate the movement of BMSB across a landscape. In agricultural areas, BMSB are often treated with insecticides although some may have limited efficacy (Leskey et al. 2012; Leskey and Nielsen 2018). The classical biological control agent, *Trissolcus japonicus*, is being studied for potential release. Genetic analyses of the *T. japonicus* found in the field in Maryland and Oregon-Washington revealed that these populations were not related to the populations in quarantine (stopBMSB.org; Mortenson 2016). In Oregon, the Oregon Department of Agriculture has approved releases of the *T. japonicus* reared from field sources, and efforts are underway in Washington to make similar releases.

### Noninsect Invertebrates

Aquatic invasive invertebrates (Table A3.4) include several mollusks (Asian clam, big-eared radix, Chinese mystery snail, New Zealand mud snails, New Zealand sea slug, and

zebra/quagga mussels) and crustaceans (crayfish, crabs, and water fleas). Many of the invasive mollusks of concern in the Northwest spread rapidly and can attain large population sizes that displace native species. These taxa can accrue prey resources rapidly, affecting foundation levels of food webs (algae, phytoplankton) in aquatic systems. Along with abundant populations come abundant waste products that can affect environmental systems. In some systems, for example, the tissues or waste products of zebra mussels may accumulate contaminants to 300,000 times the level available in the environment, with subsequent effects on their environment, including contaminating their predators (Snyder et al. 1997) (see Chap. 2, Sect. 2.6, and Box 2.6 for additional discussion of the impacts of zebra mussels). Another concern is that large numbers of mollusks can foul human structures. Introductions of some species are likely tied to inadvertent human transmission, such as in ship ballast water or in boats or fishing gear (i.e., zebra/quagga mussels, spiny water flea (*Bythotrephes longimanus*), and green crabs). The deliberate introduction and consequent escape of some species are also associated with food and medical markets (i.e., Chinese mystery snails, crayfish, and mitten crabs). Nearly all terrestrial invertebrates are insects (see above and Table A3.3).

### Vertebrates

Two frogs (American bullfrog, red-legged frog (Ranidae)), a tunicate (*Didemnum vexillum*), a turtle, and 11 fishes (Table A3.5) have established populations in the Northwest region as a consequence of human activities. For example, American bullfrogs are native to the Eastern United States, but were brought to the West to establish food farms and out of nostalgia for their calls. Bullfrogs are carriers of the amphibian chytrid fungus, *Batrachochytrium dendrobatidis* (Bd) (Table A3.2), but do not always exhibit disease symptoms and hence may serve as a reservoir species of the pathogen. Invasive fishes include a mix of species introduced for human food, as bait for recreational fisheries, or from the aquarium or ornamental industry (Table A3.5). They are of concern primarily because of their ecological effects on native ecosystems. Atlantic salmon are native to the North Atlantic Ocean, where they are anadromous, occurring in the ocean and returning to spawn in rivers. Farms in Washington and British Columbia are thought to be the origin of the species found elsewhere in the Northwest, as these fish stray from "natal" streams—even as far as Alaska. Competition with native salmonids, pollution from the farms, and the potential for farm-raised animals to carry pathogens to native stocks are all ecological impacts of these invaders. Certain species of gobies (Table A3.5) are of Asian origin and occur in fresh and brackish water. They are thought to have been introduced to the Northwest region by ballast water and may compete with or prey upon native species. Golden shiners

are from the Eastern United States and are pond-cultured fishes that are also used as bait. Golden shiners may displace native species. The carpet sea squirt or ascidian (*Didemnum vexillum*) is a colonial tunicate in the phylum Chordata (hence its inclusion under vertebrates). It seems to be native to Japan, but it has been detected along the Washington coast since 2009 and in two Oregon bays since 2010. It is a fouling organism in marine and estuarine systems, growing rapidly and covering vast surfaces as mats, displacing native biota and encrusting dock pilings and aquatic equipment. It can be introduced in ballast water, or it may hitchhike on the hulls of boats, or with commercial shellfish stock or equipment.

The category of terrestrial invasive vertebrates in the Northwest region contains only six species (Table A3.5), but they can have extensive effects, ecologically and socioeconomically. Three species are strongly associated with wetland and riparian environments (feral swine, mute swan, nutria). Feral swine are escaped domestic pigs whose rooting behavior can have several effects: waterway habitat degradation, provision of an invasion pathway for non-native plants, and damage to agricultural crops and lands. The mute swan was introduced from New York for scenic enjoyment. These aggressive, large (11.3–13.6 kg, 25–30 lb) birds may consume significant quantities of aquatic plants, competing with native birds for food and habitat supplies. Nutria were initially brought to the Pacific Northwest as part of fur farming in the 1920s. Escaped and released animals (after the collapse of this element of the fur industry) subsequently spread throughout the region. They burrow into the banks of streams and agricultural canals, destabilizing natural stream systems and human agricultural infrastructure. See the Pacific Southwest regional summary for additional discussion of nutria. In addition to these three species, European starling and rock doves are strong competitors for nest space and food sources with native birds. The Norway rat is common in urban settings in the Northwest region and is closely affiliated with human structures. These animals may cause extensive damage to human structures and are known carriers of pathogens that may affect human health.

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## Literature Cited

- Acheampong S, Strong WB, Schwartz MD et al (2016) First Canadian records for two invasive seed-feeding bugs, *Arocatus melanocephalus* (Fabricius, 1798) and *Raglius alboacuminatus* (Goeze, 1778), and a range extension for a third species, *Rhyparochromus vulgaris* (Schilling, 1829) (Hemiptera: Heteroptera). *J Entomol Soc Br Columbia* 113:74–78
- Alexander RR (1987) Ecology, silviculture and management of the Engelmann spruce-subalpine fir type in the central and southern Rocky Mountains. *Agricultural handbook* 659. U.S. Department of Agriculture, Forest Service, Washington, DC, 144 p
- Annand PN (1924) A new species of *Adelges* (Hemiptera, Phylloxeridae). *Pan Pac Entomol* 1:79–82
- Annand PN (1928) A contribution toward a monograph of the Adelginae (Phylloxeridae) of North America. *University Series in Biological Sciences*, vol 6, no. 1. Stanford University Publication, Palo Alto. 146 p
- Aubry C, Goheen D, Shoal R, Ohlson T, Lorenz T et al (2008) Whitebark pine restoration strategy for the Pacific northwest region 2009–2013. USDA Forest Service, Pacific Northwest Region, Portland, 96 p
- Bai BB, Worth RA, Johnson KJR, Brown G (2002) Distribution and phenology of the cereal leaf beetle, *Oulema melanopus* (L.) (Coleoptera: Chrysomelidae) in Oregon. *Proceedings 61st Annual Pacific Northwest Insect Management Conference*, Portland. 4 p. <https://research.libraries.wsu.edu:8443/xmlui/bitstream/handle/2376/2253/2002%20Oregon%20CLB%20Survey%20-%20Bai%20et%20al.pdf?sequence=1&isAllowed=y>
- Bautista S (2017) Personal communication on April 19, 2017 to Rebecca Flitcroft, U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station, 3200 SW Jefferson Way, Corvallis, OR 97330
- Bellows TS, Meisenbacher C, Reardon RC (1998) Biological control of arthropod forest pests of the western United States: a review and recommendations. Publication FHTET-96-21. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Morgantown, 121 p

- Benedict WV (1981) History of white pine blister rust control: a personal account. FS-355. U.S. Department of Agriculture, Forest Service, Washington, DC, 47 p
- Bentz BJ, Régnière J, Fettig CJ et al (2010) Climate change and bark beetles of the western US and Canada: direct and indirect effects. *Bioscience* 60:602–613
- Bergmann EJ, Bernhard KM, Bernon G et al (2014) Host plants of the brown marmorated stink bug in the U.S. [StopBMSB.org](http://StopBMSB.org). USDA-NIFA SCRI Coordinated Agricultural Project Northeastern IPM Center. [www.stopbmsb.org/where-is-bmsb/host-plants/](http://www.stopbmsb.org/where-is-bmsb/host-plants/)
- Bergmann EJ, Venugopal PD, Martinson HM et al (2016) Host plant use by the invasive *Halyomorpha halys* (Stal) on woody ornamental trees and shrubs. *PLoS One* 11:e0149975
- Betlejewski F, Goheen DJ, Angwin PA, Sniezko RA (2011) Port-Orford-cedar root disease. Forest Insect & Disease Leaflet 131 (revised). US Department of Agriculture, Forest Service, Portland. 12 p
- Bogdanowicz SM, Mastro VC, Prasher DC, Harrison RG (1997) Microsatellite DNA variation among Asian and North American gypsy moths (Lepidoptera: Lymantriidae). *Ann Entomol Soc Am* 90:768–775
- Bogdanowicz SM, Schaefer PW, Harrison RG (2000) Mitochondrial DNA variation among worldwide populations of gypsy moths, *Lymantria dispar*. *Mol Phylogenet Evol* 15:487–495
- Bogdanowicz SM, Wallner WE, Bell J et al (1993) Asian gypsy moths (Lepidoptera: Lymantriidae) in North America: evidence from molecular data. *Ann Entomol Soc Am* 86:710–715
- Boyce JS (1938) Forest pathology, 1st edn. McGraw-Hill, New York/London, 600 p
- Bruck DJ, Bolda M, Tanigoshi L et al (2011) Laboratory and field comparisons of insecticides to reduce infestation of *Drosophila suzukii* in berry crops. *Pest Manag Sci* 67:1375–1385
- CABI Invasive Species Compendium (2018) *Ludwigia grandiflora* (water primrose). <https://www.cabi.org/isc/datasheet/109148#3C51E072-69A3-4DD8-9920-0E5C0E57CE27>
- California Oak Mortality Task Force (2016) November monthly report. 4 p. <http://www.suddenoakdeath.org/wp-content/uploads/2016/11/COMTF-Report-November-2016.pdf>
- Childs RD, Swanson DC (2003) The winter moth, (*Operophtera brumata* (L.)). University of Massachusetts Extension Report, Amherst. 2 p. [http://www.massaudubon.org/content/download/7334/133282/file/winter\\_moth.pdf](http://www.massaudubon.org/content/download/7334/133282/file/winter_moth.pdf)
- Chrystal RN (1916) The life-history of *Chermes cooleyi* Gillette, in Stanley Park, Vancouver, B.C. 46th annual report of the entomological society of Ontario 1915, pp 123–130
- DeNitto GA, Kliejunas JT (1991) First report of *Phytophthora lateralis* on Pacific yew. *Plant Dis* 75:968
- Detwiler SB (1922) Blister rust appears in the Puget Sound region. *Am For* 28:97–98
- Dick MW (2001) Straminipilous fungi: systematics of the Peronosporomycetes including accounts of the marine Straminipilous protists, the Plasmodiophorids and similar organisms. Kluwer Academic Publishers, Dordrecht, 660 p
- Djoumad A, Nisole A, Zahiri R et al (2017) Comparative analysis of mitochondrial genomes of geographic variants of the gypsy moth, *Lymantria dispar*, reveals a previously undescribed genotypic entity. *Sci Rep* 7:1–12. <https://doi.org/10.1038/s41598-017-14530-6>
- Doerr MD, Brunner JF, Smith TJ (2008) Biology and management of bark beetles (Coleoptera: Curculionidae) in Washington cherry orchards. *J Entomol Soc BC* 105:69–81
- Edwards N (1999) Anatoxin. University of Sussex at Brighton. Updated 1 September 1999
- Flitcroft R, Hansen B, Capurso J, Christiansen K (2016) Coverage of aquatic invasive risk assessment in USFS Region 6. Report to U.S. Forest Service Region 6. US Department of Agriculture, Forest Service, Pacific Northwest Research Station, Corvallis. Available at: <https://www.fs.fed.us/pnw/lwm/aem/people/flitcroft.html>
- FAO (Food and Agriculture Organization) (2009) International standards for phytosanitary measures: revision of ISPM No. 15, Regulation of wood packaging material in international trade. Publication No. 15. Food and Agriculture Organization of the United Nations, Rome. [https://en.wikipedia.org/wiki/ISPM\\_15](https://en.wikipedia.org/wiki/ISPM_15)
- Foote RH, Blanc FL, Norrbom AL (1993) Handbook of the fruit flies (Diptera: Tephritidae) of America north of Mexico. Cornell University Press, Ithaca, 571 p
- Freyman T (2015) The monitoring of Asian gypsy moth, pink gypsy moth and nun moth at the ports of Vladivostok (Russky Island), Nakhodka, Votochny, Slavyanka, Olga, Vanino, Plastun, Pos'et, Zarubino, Kozmino, Korsakov in 2015. OFO-PR-15-01. Federal Service for Veterinary and Phytosanitary Surveillance, Federal State Budgetary Institution, All-Russia Plant Quarantine Service, Ogden. 51 p
- Furniss RL, Carolin VM (1977) Western forest insects. Miscellaneous Publication 1339. U.S. Department of Agriculture, Forest Service, Washington, DC. 654 p
- Gast S, Beckman DP, Livingston RL, Gustin J (1990) Distribution of the balsam woolly adelgid in Idaho. Report 90–5. USDA Forest Service Northern Region, Missoula. 8 p

- Geils BW, Hummer KE, Hunt RS (2010) White pines, *Ribes*, and blister rust: a review and synthesis. For Pathol 40(3–4):147–185
- Gerberg EJ (1957) A revision of the New World species of powder-post beetles belonging to the family Lyctidae. United States Department of Agriculture, Technical Bulletin No.1157, 55 p
- Goheen EM, Goheen DJ (2014) Status of sugar and western white pines on federal forest lands in southwest Oregon: Inventory query and natural stand survey results. SWOFIDSC-14-01. US Department of Agriculture, Forest Service, Pacific Northwest Region, Portland. 71 p. [https://www.fs.usda.gov/Internet/FSE\\_DOCUMENTS/stelprdb5447311.pdf](https://www.fs.usda.gov/Internet/FSE_DOCUMENTS/stelprdb5447311.pdf)
- Grünwald NJ, Larsen MM, Kamvar ZN et al (2016) First report of the EU1 clonal lineage of *Phytophthora ramorum* on tanoak in an Oregon forest. Plant Dis 100:1024
- Hansen EM (2011) *Phytophthora lateralis*, Species profile. For Phytophthoras. <https://doi.org/10.5399/osu/fp.1.1.1816>. <http://journals.library.oregonstate.edu/ForestPhytophthora/article/view/1816/1811>
- Hansen EM, Goheen DJ, Jules ES, Ullian B (2000) Managing port-Orford-cedar and the introduced pathogen *Phytophthora lateralis*. Plant Dis 84:4–14
- Hansen EM, Hamm PB (1996) Survival of *Phytophthora lateralis* in infected roots of Port Orford cedar. Plant Dis 80:1075–1078
- Hatch MH (1953) The beetles of the Pacific Northwest. Part 1: introduction and Adephaga. University of Washington Publications in Biology 16(1). University of Washington Press, Seattle. 340 p
- Hatch MH (1962) The beetles of the Pacific Northwest. Part 3: Pselaphidae and Diversicornia. University of Washington Publications in Biology 16(3). University of Washington Press, Seattle. 503 p
- Hauser M (2011) A historic account of the invasion of *Drosophila suzukii* (Matsumura) (Diptera: Drosophilidae) in the continental United States, with remarks on their identification. Pest Manag Sci 67:1352–1357
- Havill NP, Footitt RG (2007) Biology and evolution of Adelgidae. Annu Rev Entomol 52:325–349. <https://doi.org/10.1146/annurev.ento.52.110405.091303>
- Havill NP, Montgomery ME, Keena M (2011) Hemlock woolly adelgid and its hemlock hosts: a global perspective. In: Onken B, Reardon R (eds) Implementation and status of biological control of the hemlock woolly adelgid. Publication FHTET-2011-04. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Morgantown, pp 3–14
- Havill NP, Shiyake S, Galloway AL et al (2016) Ancient and modern colonization of North America by hemlock woolly adelgid, *Adelges tsugae* (Hemiptera: Adelgidae), an invasive insect from East Asia. Mol Ecol 25:2065–2080. <https://doi.org/10.1111/mec.13589>
- Hayes JL, Ragenovich I (2001) Non-native invasive forest insects of eastern Oregon and Washington. Northwest Sci 75:77–84
- Hitchcock ME, Miller S, Pike K, Gould MC (2002) Cereal leaf beetle survey and biocontrol activities in Washington State, 2002. Olympia, Washington: 2002 Entomology Project Report—WSDA Pub 077 (N/8/02), Plant Protection Division, Pest Program Washington State Department of Agriculture. 10 p. <https://research.libraries.wsu.edu/xmlui/bitstream/handle/2376/2228/2002%20Washington%20State%20CLB%20Activity%20Report%20-%20Hitchcox%20et%20al.pdf?sequence=6&isAllowed=y>
- Hrinkevich KH, Progar RA, Shaw DC (2016) Climate risk modelling of balsam woolly adelgid damage severity in subalpine fir stands of western North America. PLoS One. <https://doi.org/10.1371/journal.pone.0165094>
- Humble LM, Mastro V, Munson AS (2013) Asian gypsy moth, it's back! In: McManus KA, Gottschalk KW (eds) Proceedings of the 24th USDA interagency research forum on invasive species; 2013 January 8–11; Annapolis, MD: Publication FHTET-13-01. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Fort Collins. 124 p
- Idaho State Department of Agriculture (2013) Elm seed bug, *Arocatus melanocephalus*: an exotic invasive pest new to the U.S. Fact Sheet, Boise. 4 p. [http://extension.oregonstate.edu/malheur/sites/default/files/spring\\_2013\\_esb\\_fact\\_sheet.pdf](http://extension.oregonstate.edu/malheur/sites/default/files/spring_2013_esb_fact_sheet.pdf)
- Invasive Species of Idaho (2017) <http://invasivespecies.idaho.gov/pests/>
- Islam MS, Barr NB, Braswell WE et al (2015) A multiplex real-time PCR assay for screening gypsy moths (Lepidoptera: Erebidae) in the United States for evidence of an Asian genotype. J Econ Entomol 108:2450–2457
- Ivie MA (2002) 69. Bostrichidae. In: Arnett RH, Frank JH, Thomas MC, Skelley PE (eds) American beetles, volume II: Polyphaga: Scarabaeoidea through Curculionoidea. CRC Press LLC, Boca Raton, pp 233–244
- Johnson NE, Mitchell RG, Wright KH (1963) Mortality and damage to Pacific silver fir by the balsam woolly aphid in southwestern Washington. J For 61:854–860
- Johnson PJ (1998) *Melanotus cete* Candèze, a second adventitious species of *Melanotus* Eschscholtz in North America (Coleoptera: Elateridae). Pan Pac Entomol 74:118–119
- Kamvar ZN, Larsen MM, Kanaskie AM et al (2015) Spatial and temporal analysis of populations of the sudden oak death pathogen in Oregon forests. Phytopathology 105:982–989

- Kanaskie A, Hansen E, Goheen EM et al (2013) Detection and eradication of *Phytophthora ramorum* from Oregon forests, 2001–2011. In: Frankel SJ, Kliejunas JT, Palmieri KM, Alexander JM (tech. coords) Proceedings of the sudden oak death fifth science symposium. General Technical Report GTR-PSW-243. U.S. Department of Agriculture, Forest Service, Pacific Southwest Research Station, Albany. 169 p
- Kanaskie A, Wiese R, Norlander D et al (2017) Slowing spread of sudden oak death in Oregon forests, 2001–2015. In: Frankel SJ, Harrell KM (tech. coords) Proceedings of the sudden oak death sixth science symposium. General Technical Report GTR-PSW-255. U.S. Department of Agriculture, Forest Service, Pacific Southwest Research Station, Albany. 106 p
- Keen FP (1952) Insect enemies of western forests. Miscellaneous Publication 273. US Department of Agriculture, Forest Service, Washington, DC. 280 p
- Kegley A, Sniezko RA, Danchok R, Savin DP (2012) Blister rust resistance among 19 families of whitebark pine, *Pinus albicaulis*, from Oregon and Washington—early results from an artificial inoculation trial. In: Sniezko RA, Yanchuk AD, Kliejunas JT, Palmieri KM, Alexander JM, Frankel SJ (tech. coords) Proceedings of the fourth international workshop on the genetics of host-parasite interactions in forestry: disease and insect resistance in forest trees. General Technical Reports. PSW-GTR-240. Pacific Southwest Research Station, Forest Service, U.S. Department of Agriculture, Albany, pp 311–315
- Kenis M, Tonina L, Eschen R et al (2016) Non-crop plants used as hosts by *Drosophila suzukii* in Europe. *J Pest Sci* 89:735–748
- Klick J, Yang WQ, Walton VM et al (2015) Distribution and activity of *Drosophila suzukii* in cultivated raspberry and surrounding vegetation. *J Appl Entomol* 140:37–46
- Kohler GR, Stiefel VL, Wallin KF, Ross DW (2008) Predators associated with the hemlock woolly adelgid (Hemiptera: Adelgidae) in the Pacific northwest. *Environ Entomol* 37:494–504
- Kohler GR, Wallin KF, Ross DW (2016) Seasonal phenology and abundance of *Leucopis argenticollis*, *Leucopis piniperda* (Diptera: Chamaemyiidae), *Laricobius nigrinus* (Coleoptera: Derodontidae) and *Adelges tsugae* (Hemiptera: Adelgidae) in the Pacific Northwest USA. *Bull Entomol Res* 106:546–550. <https://doi.org/10.1017/S0007485316000250>
- Kruse JJ, Zogas K, Hard J, Lisuzzo N (2010) New pest in Alaska and Washington—The green alder sawfly—*Monsoma pulveratum* (Retzius). Pest Alert, R10-PR-022, Anchorage, AK: USDA Forest Service, State and Private Forestry. 2 p
- LaBonte JR, Mudge AD, Johnson KJR (2005) Non-indigenous woodboring Coleoptera (Cerambycidae, Curculionidae: Scolytinae) new to Oregon and Washington, 1999–2002: consequences of the intracontinental movement of raw wood products and solid wood packing materials. *Proc Entomol Soc Wash* 107:554–564
- LaGasa E (2006) New pest alert and update: Introduced exotic seed-bugs are new and increasing nuisance problems in areas of western Washington—*Rhyparochromis vulgaris* and *Raglius alboacuminatus*. AGR PUB 805–158, Olympia, WA: 2006 Entomology Pest Alert Pest Program, Plant Protection Division Washington State Department of Agriculture. 1 p. [https://agr.wa.gov/PlantsInsects/InsectPests/Exotics/Surveys/seedbugs\\_06.pdf](https://agr.wa.gov/PlantsInsects/InsectPests/Exotics/Surveys/seedbugs_06.pdf)
- LaGasa E, Murray T (2007) Exotic seed-bugs (Lygeoidea: Rhyparochromidae & Oxycaenidae) new to the Pacific Northwest. In: Proceedings 66th annual pacific northwest insect management conference, January 8–9, 2007, pp 5–6. [http://www.ipmnet.org/PNWIMC/2007\\_PNW\\_Conference\\_Proceedings.pdf](http://www.ipmnet.org/PNWIMC/2007_PNW_Conference_Proceedings.pdf)
- Lech G, Livingston RL (2004) State of Idaho gypsy moth survey trapping program summary report 2004. Report No. IDL 04–2, Boise
- Lee JC, Aguayo I, Aslin R et al (2009) Co-occurrence of the invasive banded and European elm bark beetles (Coleoptera: Scolytidae) in North America. *Ann Entomol Soc Am* 102:426–436
- Lee JC, Bruck DJ, Dreves AJ et al (2011) In focus: spotted wing drosophila, *Drosophila suzukii*, across perspectives. *Pest Manag Sci* 67:1349–1351
- Lee JC, Dreves AJ, Cave AM et al (2015) Infestation of wild and ornamental noncrop fruits by *Drosophila suzukii* (Diptera: Drosophilidae). *Ann Entomol Soc Am* 108:117–129
- Leskey TC, Nielsen AL (2018) Impact of the invasive brown marmorated stink bug in North America and Europe: history, biology, ecology, and management. *Annu Rev Entomol* 63:599–618
- Leskey TC, Lee D-H, Short BD, Wright SE (2012) Impact of insecticides on the invasive *Halyomorpha halys* (Hemiptera: Pentatomidae): analysis of insecticide lethality. *J Econ Entomol* 105:1726–1735
- Liebhold AM, Brockerhoff EG, Garrett LJ et al (2012) Live plant imports: the major pathway for forest insect and pathogen invasions of the U.S. *Front Ecol Environ* 10:135–143
- Logan JA, Powell JA (2001) Ghost forests, global warming, and the mountain pine beetle. *Am Entomol* 47:160–173
- Logan JA, Régnière J, Powell JA (2003) Assessing the impacts of global warming on forest pest dynamics. *Front Ecol Environ* 1:130–137
- Looney C, Sheehan K, Bai B et al (2012) The distribution of a potential new forest pest, *Monsoma pulveratum*

- (Hymenoptera: Tenthredinidae), in the Pacific northwest states. *Northwest Sci* 86:342–345
- Looney C, Murray T, Lagasa E, Hellman WE, Passoa SC (2016a) Shadow surveys: how non-target identifications and citizen outreach enhance exotic pest detection. *Am Entomol* 62:247–254
- Looney C, Smith DR, Collman SJ, Langor DW, Peterson MA (2016b) Sawflies (Hymenoptera, Symphyta) newly recorded from Washington state. *J Hymenopt Res* 49:129–159
- Lowrey L, Davis G (2017) Management and research goals for balsam woolly adelgid in the Interior West. USDA Forest Service, Northern Region Forest Health Protection, Unpublished report, Missoula. 9 p
- Maloy OC (1997) White pine blister rust control in North America: a case history. *Annu Rev Phytopathol* 35:87–109
- Mausel DL, Van Driesche RG, Elkinton JS (2011) Comparative cold tolerance and climate matching of coastal and inland *Laricobius nigrinus* (Coleoptera: Derodontidae), a biological control agent of hemlock woolly adelgid. *Biol Control* 58:96–102
- McClure MS (2001) Biological control of the hemlock woolly adelgid in the eastern United States. Publication HTET-2000-08. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Morgantown. 10 p
- McClure MS, Salom SM, Shields KS (1996) Hemlock woolly adelgid. Publication FHTET-96-35. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Morgantown. 18 p
- Miller B, Anfora G, Buffington M et al (2015) Seasonal occurrence of resident parasitoids associated with *Drosophila suzukii* in two small fruit production regions of Italy and the USA. *Bull Insect* 68:255–263
- Mitchell RG, Wright KH (1967) Foreign predator introductions for control of the balsam woolly aphid in the Pacific northwest. *J Econ Entomol* 60:140–147
- Mortenson M (2016) Stinkbugs' natural predator has arrived in the Pacific Northwest. Capital Press, July 28, 2016. <http://www.capitalpress.com/Oregon/20160728/stinkbugs-natural-predator-has-arrived-in-the-pacific-northwest>
- Mudge AD, LaBonte JR, Johnson KJR et al (2001) Exotic woodboring Coleoptera (Micromalthidae, Scolytidae) and Hymenoptera (Xiphidriidae) new to Oregon and Washington. *Proc Entomol Soc Wash* 103:1011–1019
- Murray T, LaGasa E, Glass J (2012) Pest alert: Red lily leaf beetle. Home Gardening Series, Washington State University Extension Publication, Pullman. 2 p. <http://oregonstate.edu/dept/nurspest/RLLB.pdf>
- Murray TA, Kohler GR, Wilhite EA (2013) *Ips paraconfusus* Lanier (Coleoptera: Curculionidae): new records of the California fivespined Ips from Washington state and the Columbia River gorge in Oregon. *Coleopt Bull* 67:28–31
- Myers JH, Simberloff D, Kuris AM, Carey JR (2000) Eradication revisited: dealing with exotic species. *Trends Ecol Evol* 15:316–320
- Nugent M (2005) Oregon invasive species action plan. Oregon Department of Fish and Wildlife. Portland. Accessed online at: <http://www.oregon.gov/OISC/index.shtml>
- ODA (Oregon Department of Agriculture) (2010) Pest Alert: Brown marmorated stink bug. <https://www.oregon.gov/ODA/shared/Documents/Publications/IPPM/BrownMarmoratedStinkBugPestAlert.pdf>. Updated July 2016
- OIE (World Organization for Animal Health [Office International des Epizooties]) (2018) OIE-listed diseases, infections and infestations in force in 2018. Available at: <http://www.oie.int/animal-health-in-the-world/oie-listed-diseases-2018/>
- OISC (2015) 100 worst list. Oregon Invasive Species Council. Available at: [https://static1.squarespace.com/static/58740d57579fb3b4fa5ce66f/t/5891577c579fb38e735f0f83/1485920126222/OISC\\_top100\\_2015.pdf](https://static1.squarespace.com/static/58740d57579fb3b4fa5ce66f/t/5891577c579fb38e735f0f83/1485920126222/OISC_top100_2015.pdf)
- Pederson LA, Beckman DP, Halsey RL, Stipe LE (2004) Idaho forest insect and disease conditions report. Idaho Department of Lands USDA Forest Service, Northern and Intermountain Regions IDL Report No. 04–3, Boise. 39 p. [https://www.fs.usda.gov/Internet/FSE\\_DOCUMENTS/fsbdev3\\_015300.pdf](https://www.fs.usda.gov/Internet/FSE_DOCUMENTS/fsbdev3_015300.pdf)
- Pelton E, Gratton C, Isaacs R et al (2016) Earlier activity of *Drosophila suzukii* in high woodland landscapes but relative abundance is unaffected. *J Pest Sci* 89:725–733
- Philips TK (2002) 70. Anobiidae. In: Arnett RH, Frank JH, Thomas MC, Skelley PE (eds) American beetles, volume II: Polyphaga: Scarabaeoidea through Curculionioidea. CRC Press LLC, Boca Raton, pp 245–260
- Prather T, Robins S, Morishita D (2016) Idaho's Noxious Weeds, 8th edn. University of Idaho Extension, Moscow. Available at: <http://www.cals.uidaho.edu/edComm/pdf/BUL/BUL816.pdf>
- Rabaglia RJ, Cognato AI, Hoebeke ER et al (2019) Early detection and rapid response, a ten-year summary of the USDA Forest Service program of surveillance for non-native bark and ambrosia beetles. *Am Entomol* 65:29–42
- Rabaglia RJ, Dole SA, Cognato AI (2006) Review of American Xyleborina (Coleoptera: Curculionidae: Scolytinae) occurring north of Mexico, with an illustrated key. *Ann Entomol Soc Am* 99:1034–1056
- Ragenovich IR, Mitchell RG (2006) Balsam woolly adelgid. Forest Insect & Disease Leaflet 118 (revised). US Department of Agriculture, Forest Service, Washington, DC. 11 p



- Reardon R, Onken B, Cheah C et al (2004) Biological control of the hemlock woolly adelgid. Publication FHTET-2004-04. US Department of Agriculture, Forest Service, Forest Health Technology Enterprise Team, Morgantown. 22 p
- Rice KB, Bergh CJ, Bergmann EJ et al (2014) Biology, ecology, and management of brown marmorated stink bug (Hemiptera: Pentatomidae). *J Integr Pest Manag* 5:1–13
- Rosetta R (2013) Azalea lace bug: biology and management in commercial nurseries and landscapes. Oregon State University Extension Service, Publication EM 9066, Corvallis. 6 p. <https://catalog.extension.oregonstate.edu/em9066>
- Ryan RB (1997) Before and after evaluation of biological control of the larch casebearer (Lepidoptera: Coleophoridae) in the Blue Mountains of Oregon and Washington, 1972–1995. *Environ Entomol* 26:703–715
- Sabrosky C, Reardon RC (1976) Tachinid parasites of the gypsy moth, *Lymantria dispar*, with keys to adults and puparia. *Misc Publ Entomol Soc Am* 10(2). 126 p
- Schoettle AW, Klutsch JG, Sniezko RA (2012) Integrating regeneration, genetic resistance, and timing of intervention for the long-term sustainability of ecosystems challenged by non-native pests—a novel proactive approach. In: Sniezko RA, Yanchuk AD, Kliejunas JT, Palmieri KM, Alexander JM, Frankel SJ (tech. coords) Proceedings of the fourth international workshop on the genetics of host-parasite interactions in forestry: disease and insect resistance in forest trees. General Technical Report. PSW-GTR-240. Pacific Southwest Research Station, Forest Service, U.S. Department of Agriculture, Albany, pp 112–123
- Seybold SJ, Downing M (2009) What risk do invasive bark beetles and woodborers pose to forests of the western U.S.? A case study of the Mediterranean pine engraver, *Orthotomicus erosus*. In: Hayes JL, Lundquist JE (compilers) The western bark beetle research group: a unique collaboration with Forest Health Protection. Proceedings of a symposium at the 2007 Society of American Foresters Conference, October 23–28, 2007, Portland, Oregon. General Technical Report, GTR-PNW-784, 134 p. Forest Service, U.S. Department of Agriculture, Portland, pp 111–134
- Shaw D, Oester P (2010) Are the introduced parasites of larch casebearer (*Coleophora laricella*), still present in the Blue Mountains, Oregon? USDA Forest Service Western Wildland Environmental Threat Assessment Center, Prineville. WWETAC Project FY10TS74. 1 p. <https://www.fs.fed.us/wwetac/old/projects/shaw2.html>
- Shoal R, Aubry CA (2006) Assessment of whitebark pine health on eight national forests in Oregon and Washington. Pacific Northwest Albicaulis project. US Department of Agriculture, Forest Service, Pacific Northwest Region, Olympia. 22 p
- Sing SE, De Clerck-Floate RA, Hansen RW et al (2016) Biology and control of Dalmatian and yellow toadflax. USDA Forest Service, Forest Health Technology Enterprise Team, Morgantown. FHTET-2016-01. 141 p. <https://www.fs.usda.gov/treearch/pubs/52446>
- Snyder FL, Hilgendorf MB, Garton DW (1997) Zebra mussels in North America: the invasion and its implications. Ohio State University, Ohio Sea Grant, Columbus. 4 p
- Stannard ME (2014) Plant guide for Himalayan blackberry (*Rubus armeniacus*). USDA Natural Resources Conservation Service. Plant Materials Center, Pullman. 3 p. [https://plants.usda.gov/plantguide/pdf/pg\\_ruar9.pdf](https://plants.usda.gov/plantguide/pdf/pg_ruar9.pdf)
- Steele R, Pfister RD, Ryker RA, Kittams JA (1981) Forest habitat types of central Idaho. General Technical Report INT-114. U.S. Department of Agriculture, Forest Service, Intermountain Forest and Range Experiment Station, Ogden. 138 p
- Stewart D, Zahiri R, Djoumad A et al (2016) A multi-species TaqMan PCR assay for the identification of Asian gypsy moths (*Lymantria* spp.) and other invasive lymantriines of biosecurity concern to North America. *PLoS One* 11:e0160878
- Strutt A, Turner JA, Haack RA, Olson L (2013) Evaluating the impacts of an international phytosanitary standard for wood packaging material: global and United States trade implications. *Forest Policy Econ* 27:54–64
- The Knotweed Company Ltd (2018) Japanese knotweed history and biology. <http://www.knotweed-removal.co.uk/history-and-biology-of-japanese-knotweed.php>
- Thorson TD, Bryce SA, Lammers DA et al (2003) Ecoregions of Oregon (color poster with map, descriptive text, summary tables, and photographs). U.S. Geological Survey (map scale 1:1,500,000), Reston. <http://people.oregonstate.edu/~muiRP/FuelsReductionSWOregon/ToolsResources/EcoregionsOregonLevelIVEPA.pdf>
- Tunnock S, Ryan RB (1985) Larch casebearer in western larch. Forest Insect & Disease Leaflet 96 (revised). US Department of Agriculture, Forest Service, Washington, DC. 8 p
- USDA Agricultural Research Service (1986) Stored-grain insects. Agricultural Handbook No. 500. 57 p
- USDA Forest Service and APHIS (1993) Russia and United States Pest Monitoring Program, 2 p
- USDA Forest Service and APHIS (2000) Pest risk assessment for importation of solid wood packing materials into the United States, 275 p
- USDA APHIS (2016) Pest alert: Asian gypsy moth. APHIS 81–35-027, Issued April 2016, 2 p
- USDA Forest Service, Northern Research Station and Forest Health Protection (2019) Alien Forest Pest Explorer—

- species map. Database last updated 28 July 2016. <https://www.nrs.fs.fed.us/tools/afpe/>
- Vernon RS, LaGasa EH, Phillip H (2001) Geographic and temporal distribution of *Agriotes obscurus* and *A. lineatus* (Coleoptera: Elateridae) in British Columbia and Washington as determined by pheromone trap surveys. *J Entomol Soc Br Columbia* 98:257–265
- Vitousek PM, Antonio CM, Loope LL, Westbrooks R (1996) Biological invasions as global environmental change. *Am Sci* 84:468–478
- Washington Invasive Species Council (2012) Stop the invasion: Bark boring moths. Fact Sheet, Washington State Recreation and Conservation Office, Washington Invasive Species Council. 2 p. <https://www.yumpu.com/en/document/read/12118488/bark-borking-moths-fact-sheet-washington-invasive-species-council>
- Webber JF, Mullett M, Brasier CM (2010) Dieback and mortality of plantation Japanese larch (*Larix kaempferi*) associated with infection by *Phytophthora ramorum*. *New Dis Rep* 22:19. <https://doi.org/10.5197/j.2044-0588.2010.022.019>
- White RE (1982) A catalog of the Coleoptera of America north of Mexico. Family: Anobiidae. USDA Agricultural Handbook No. 529–70, 59 p
- Wiman NG, Lowenstein DM (2017) Emerging pest: brown marmorated stink bug—a pending threat to Pacific northwest agriculture. In: Hollingsworth CS (ed) Pacific Northwest insect management handbook. Pacific Northwest Extension Publication. Oregon State University, Corvallis. unpaginated. <https://pnwhandbooks.org/node/12634/print>
- Winston R, Randall CB, De Clerck-Floate R et al (2014a) Field guide for the biological control of weeds. University of Idaho Extension. Publication FHTET-2014-08. USDA Forest Service, Forest Health Technology Enterprise Team, Morgantown. 338 p. <https://www.ibiocontrol.org/westernweeds.pdf>
- Winston RL, Schwarzländer M, Hinz HL et al (2014b) Biological control of weeds: a world catalogue of agents and their target weeds, 5th edn. Publication FHTET-2014-04. USDA Forest Service, Forest Health Technology Enterprise Team, Morgantown. 838 p. [https://www.ibiocontrol.org/catalog/JulienCatalogueFHTET\\_2014\\_04.pdf](https://www.ibiocontrol.org/catalog/JulienCatalogueFHTET_2014_04.pdf)
- WISP (2009) Top priorities. Washington Invasive Species Council. Available at: <https://invasivespecies.wa.gov/find-a-priority-species/>
- Zobel DB (1990) *Chamaecyparis lawsoniana* (A. Murr.) Parl. Port-Orford-Cedar. In: Burns RM, Honkala BH (tech. coords) *Silvics of North America: 1. Conifers; 2. Hardwoods*. Agriculture Handbook 654. U.S. Department of Agriculture, Forest Service, Washington, DC. 877 p

## Southwest Region

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### Introduction

The Southwest region (Arizona, California, Colorado, Nevada, New Mexico, and Utah) (Figs. A4.1 and A4.2) is marked by Mediterranean, montane, and desert climates/ecosystems that provide unique and amenable conditions and habitats for invading plants, pathogens, insects, and vertebrates. Aridity is perhaps the dominant climatic feature framing the forest ecosystems of the Southwest (Peterson 2012). Extreme elevational gradients and the intervening desert landscapes in this region (Fig. A4.2) create pronounced biogeographical boundaries and refugia for endemic species of plants and animals. The southern edge of this region has an extensive, but ecologically contiguous, border with Mexico that facilitates biological invasions. Future climate conditions projected for the southern portion of this region predict a trend of increasing temperature and decreasing precipitation (Cayan et al. 2010; Peterson 2012; Williams et al. 2010). Changing climate will likely place water stress on native trees and other plants, perhaps accelerating the establishment of invasive species (Peterson 2012) and amplifying outbreaks of native pest species (Breshears et al. 2005). These changes may also facilitate the spread of invasive species northward across this international border (e.g., Billings et al. 2014; Moser et al. 2005). The rate of spread of invasive species across this border may be increased by instances of unregulated movement of humans and cargo.

This region also features a wide range of non-native ornamental plants in urban and rural areas, enormously productive and diverse agroecosystems, and huge tracts of public lands with grazing impacts that favor the establishment and spread of invasive plants and pathogens by wild and domestic ungulates and other animals. High property values and residences in and near this region's forests make the impacts of invasive species particularly expensive and difficult to manage, as they often range across varied and numerous land ownerships. From a sociological perspective, the diverse human population of the region provides linkages to many overseas source populations of invasive species, whereas numerous maritime and overland ports-of-entry as well as U.S. and international tourism in response to the attractive natural features and mild winter climate may also enhance the introduction, establishment, and spread of invading organisms.

### Plants

Terrestrial invasive plants in the Southwest region include annual, biennial, and perennial species of grasses, forbs,