

Suckermouth Catfish (*Hypostomus plecostomus*)

Ecological Risk Screening Summary

U.S. Fish and Wildlife Service, Web Version – 1/29/2018



Photo: Ildar Sagdejev. Licensed under Creative Commons BY-SA 4.0 International. Available: https://commons.wikimedia.org/wiki/File:2004-02-02_Plecostomus_on_blue_gravel.jpg. (May 2013).

1 Native Range and Status in the United States

Native Range

From Nico and Neilson (2015):

“South America: Guyana, Suriname [*sic*] and French Guiana, between the Essequibo and Oyapock River basins (Weber et al. 2012)”

Status in the United States

From Nico and Neilson (2015):

“*Hypostomus plecostomus* was collected from Six Mile Creek in Tampa, Florida in 1972 (museum specimens). Various other reports from around the state (Florida FWC 2000), including a borrow pit in Wayside Park in Perrine, Miami-Dade County (Shafland 1976). Collected in

Indian Spring, Nevada, in 1983 (museum specimens). Reported in several watersheds in Texas: the San Antonio River (Texas Parks and Wildlife Department 2001; museum specimens), San Felipe Creek (Gleason 2004), the San Marcos River (museum specimens), Comal River (Howells 1992; Whiteside and Berkhouse 1992), and White Oak Bayou (T. White, personal communication).

A single specimen was collected from Dos Bocas Reservoir and several specimens collected from an irrigation canal in Lajas, Puerto Rico (F. Grana, personal communication)."

"Established in Texas. Reported from Florida, Nevada, and Puerto Rico."

From CABI (2015):

"Although *H. plecostomus* was reported from Indian Spring, Nevada, in 1983, the single specimen was later determined to be an unidentified species of *Hypostomus* and not *H. plecostomus* (USGS NAS, 2015)."

From NatureServe (2017):

"The identity of the species in this genus [*Hypostomus*] that are established in the United States is uncertain."

Means of Introductions in the United States

From Nico and Neilson (2015):

"Aquarium release or escape from aquaculture facilities."

From FAO (2013):

"Reason of Introduction: ornamental"

Remarks

There is some taxonomic uncertainty that interferes with obtaining a clearly defined distribution for *Hypostomus plecostomus*.

2 Biology and Ecology

Taxonomic Hierarchy and Taxonomic Standing

From ITIS (2013):

"Kingdom Animalia
Phylum Chordata
Subphylum Vertebrata
Superclass Osteichthyes
Class Actinopterygii

Subclass Neopterygii
Infraclass Teleostei
Superorder Ostariophysi
Order Siluriformes
Family Loricariidae
Subfamily Hypostominae
Genus *Hypostomus* Lacepède, 1803
Species *Hypostomus plecostomus* (Linnaeus, 1758)”

“Taxonomic Status:
Current Standing: valid”

From Eschmeyer et al. (2017):

“*plecostomus*, *Acipenser* Linnaeus [C.] 1758:238 [Systema Naturae, Ed. X v. 1 [...]] Suriname River. Lectotype: NRM 32 (57.8 mm SL). Paralectotypes: ?NRM 32 (82.0 and 81.4 mm), NRM 32 (smallest). Type catalog and nomenclatural discussion: Fernholm & Wheeler 1983:222 [...], Ferraris 2007:258 [...]. Type information: Isbrücker 1980:30 [...], Wheeler 1989:215 [...]. Neotype designation by Boeseman 1968 (RMNH 18240 is considered to be invalid since at least the smallest specimen in NHRM LP 32 is regarded as part of the type series); Lectotype selected by Weber et al. 2012:210 [...]. •Valid as *Hypostomus plecostomus* (Linnaeus 1758) -- (Isbrücker 1980:30 [...], Burgess 1989:432 [...], Galvis et al. 1997:86 [...], Britski et al. 1998:135 [...], Isbrücker 2001:25, 27 [...], Isbrücker 2002:19 [...], Camargo & Isaac 2001:145 [...], Ferraris 2003:865 [...], Weber in Reis et al. 2003:361 [...], Armbruster 2004:79 [...], Nelson et al. 2004:83 [...], Scharpf 2006:20 [...] as cf. *plecostomus*, Armbruster et al. 2007:67 [...], Ferraris 2007:258 [...], Zawadzki et al. 2010:724 [...], Weber et al. 2012:196 [...], 210, Page et al. 2013:81 [...], Zawadzki et al. 2014:103 [...], Sarmiento et al. 2014:191 [...], Cardoso et al. 2016:22 [...], Zhang et al. 2016:211 [...], Melo et al. 2016:134 [...]). **Current status:** Valid as *Hypostomus plecostomus* (Linnaeus 1758). Loricariidae: Hypostominae.”

Size, Weight, and Age Range

From Froese and Pauly (2013):

“Max length : 50.0 cm SL male/unsexed; [Galvis et al. 1997]; common length: 28.0 cm TL male/unsexed; [Hugg 1996]”

From CABI (2015):

“Limited data are available on the lifespan of *H. plecostomus*. Pectoral fin rays, used in traditional age assessments, may not be accurate due to lumens that form with the growth of the fish (i.e. they become hollow) and due to non-annual formation of growth rings. Lifespans of *Hypostomus* spp. in the wild of range from 7-8 years; however, aquaria specimens are commonly reported to live for 10-15 years (Hoover et al., 2004).”

Environment

From Froese and Pauly (2013):

“Freshwater; demersal; pH range: 6.2 - 8.2; dH range: ? - 28. [...]; 20°C - 28°C [assumed to be recommended aquarium temperature range] [Baensch and Riehl 1985]; [...]”

From CABI (2015):

“*Hypostomus* sp. tolerate brackish water of 6-12 ppt, though are not found in higher adjacent salinities (Barletta et al., 2000; Hoover et al., 2014).”

“Although *Hypostomus* sp. can tolerate hypoxic conditions using accessory breathing, no data are available on oxygen levels necessary to promote this response.”

Climate/Range

From Froese and Pauly (2013):

“Tropical; [...]; 12°N - 25°S, 60°W - 51°W”

From CABI (2015):

“*Hypostomus* sp. are found living in areas where waters reach 32°C (Barletta et al., 2000). *Hypostomus* sp. are tolerant of cooler temperatures (16°C) though at 13°C they exhibit a distinctive reddening of fins due to cold stress (Grier, 1980; Hoover et al., 2014). In controlled laboratory experiments Shafland and Pestrak (1982) determined that a *Hypostomus* spp. reduced feeding at 20.5°C, stopped feeding at 18.7°C and died at 11.2°C. Hoover et al. (2014) suggested a lower lethal temperature of 12-14°C, which was supported by the absence of low temperature ‘winter kills’ above 15°C at Galveston Bay, Texas (Robinson and Culbertson, 2005) and the presence of winter kills at Hillsborough River at 10-12°C (Hoover et al., 2014).”

Distribution Outside the United States

Native

From Nico and Neilson (2015):

“South America: Guyana, Suriname [*sic*] and French Guiana, between the Essequibo and Oyapock River basins (Weber et al. 2012)”

Introduced

From Froese and Pauly (2013):

“Have been introduced to several Asian countries for the aquarium trade [Baensch and Riehl 1985].”

“Established: Bangladesh, Florida and Texas, Thailand, Vietnam, Malaysia, Taiwan, Sri Lanka, Philippines. Introduced: Singapore, Hong Kong, China, UK.”

From Maceda-Veiga et al. (2013):

“Besides the species highlighted in our study, other ornamental species have been recorded in Iberian waters: tinfoil barb (*Barbonymus schwanenfeldi*) in Portugal (Gante et al. 2008); and, Oscar (*Astronotus ocellatus*), red piranha (*Pygocentrus nattereri*), and suckermouth catfish (*Hypostomus plecostomus*) in Spain (Elvira and Almodo´var 2001; Doadrio 2002).”

From Zięba et al. (2010):

“Other released specie of particular note are [...] an armoured suckermouth catfish *Hypostomus plecostomus* (Linnaeus, 1758) in St-John’s Pond of Epping Forest [England]”

From CABI (2015):

“It has been introduced to 17 countries in the Americas, Asia and Europe.”

“It is possible that specimens collected and recorded as *H. plecostomus* from Brazil (Silvano and Begossi, 2001) and Argentina (Lopez et al., 1987) may be introduced populations of *H. plecostomus*, or more likely other *Hypostomus* sp., because these locations are geographically isolated from the natural distribution of *H. plecostomus* (northern South America).”

“Although introduced populations of *H. plecostomus* occur in at least 17 countries, these populations have not been well documented, particularly in many Asian countries. This has been exacerbated by the taxonomic uncertainty of loricariids in general, and *Hypostomus* and *Pterygoplichthys* spp. in particular.”

“In China, *H. plecostomus* was recorded in the Huizhou segment of the Dongjiang River in 2007. It was not recorded in previous surveys in the 1980s (Liu et al., 2011). Ma et al. (2003) reported that *H. plecostomus* was introduced to aquatic habitats in the country in 1990, though provided no further details.”

“In Columbia, introduced populations of *H. plecostomus* are well established in the anthropogenically-impacted upper basin of the Cauca River. Lopez Macias et al. (2009) cited the field collections of Ortega et al. (1999), where it was found that *H. plecostomus* was the most abundant fish species captured. *H. plecostomus* was introduced to Columbia from Guyana (Lopez Macias et al., 2009).”

From Pallewatta et al. (2003):

“[...] not yet considered invasive, but on "watch list"; introduced in 1990s by ornamental fish industry; escaped from breeding ponds into Laguna de Bay/nearby rivers; [...]”

“*Hypostomus plecostomus* (tank cleaner), a species imported to Sri Lanka by the ornamental fish industry, [...]”

Means of Introduction Outside the United States

From FAO (2013):

“Reasons of Introduction: ornamental”

From CABI (2015):

“The majority of nonindigenous populations of *H. plecostomus* are the result of the release of unwanted ornamental fishes (Mendoza-Alfaro et al., 2009; USGS NAS, 2015).”

Short Description

From Froese and Pauly (2013):

“Dorsal spines (total): 1; Dorsal soft rays (total): 7; Anal spines: 1; Anal soft rays: 3 - 5. Body short and robust; caudal peduncle not depressed. Upper parts of head and body encased in longitudinal rows of scutes; lower surface of head and abdomen naked.”

“Adipose fin: present. Pectoral fins: 1 spine, 3-5 soft rays. Pelvic fins: 1 spine 5 soft rays.”

From CABI (2015):

“*H. plecostomus* and other Loricariidae (including *Pterygoplichthys* sp.) can be distinguished from native North American catfishes (Ictaluridae) by the presence of flexible bony plates covering the body (absent in ictalurids) and a ventral suckermouth (terminal in ictalurids) (Nico et al., 2015).”

“In comparison with *Pterygoplichthys* sp., *H. plecostomus* is usually shorter and stouter, the head is broader relative to the length and there are small discrete dark spots on the head (Florida Fish and Wildlife Conservation Commission, 2015).”

“A commonly-introduced species of the latter genus, *Pterygoplichthys multiradiatus*, may also be differentiated from *H. plecostomus* by the connection of the last dorsal ray by a small membrane to the base of the following bony plate. The species also has a granular edge on the snout (Page and Burr, 1991).”

Biology

From CABI (2015):

“Parental care is common in loricariids and many species are cavity builders and nest guards. Male *H. plecostomus* burrow into banks and bottom sediments to create chambers in which females lay eggs. Males guard the mass of eggs (Burgess, 1989) which hatch in 3-5 days (Baensch and Riehl, 1985). Burrows of *H. plecostomus* observed in Florida ponds exhibit a single opening but then subdivide into three or four different tunnels that extend 0.9-1.2 m parallel to the surface of the water (Grier, 1980). In Texas, burrows are reported to be 1.2-1.5 m deep (Texas Parks and Wildlife, 2012). Burrows are typically located in steeply sloping banks

with soils containing almost no gravel, and they are especially evident in highly disturbed urban ponds (Hoover et al., 2014).”

“*H. plecostomus* grows rapidly and may mature at lengths of 150 mm in introduced populations in Florida (Grier, 1980), which is less than half the typical adult size of 400-500 mm (Burgess, 1989). Size at maturity of *H. plecostomus* is comparable with other *Hypostomus* sp. in their native range in South America (Nomura and Mueller, 1980; Mazzoni and Caramaschi, 1995).”

“The total fecundity of *H. plecostomus* is reported to be approximately 3000 eggs (Azevedo, 1938). The batch fecundity of female fish from the San Marcos River in Texas ranged from 871-3367 eggs per ovary (Cook-Hildreth, 2008). Data are similar to those from various *Hypostomus* sp. in their native range, which have total fecundities of several thousand eggs, and batch fecundities of approximately 1000 eggs (Mazzoni and Caramaschi, 1997; Duarte and Araújo, 2002). Egg masses of *H. plecostomus* typically contain 500-700 eggs (Grier, 1980; Hoover et al., 2014).”

“*H. plecostomus* is believed to spawn multiple times throughout a protracted spawning season. In Texas, multiple-sized oocytes, which are indicative of multiple spawning events, are documented for the species (Cook-Hildreth, 2008). The spawning season, based on gonadosomatic indices, is from March through September (Hoover et al., 2014). In their native range, *Hypostomus* sp. also exhibit protracted spawning periods of greater than 5 months, which usually coincides with the warm rainy season (Mazzoni and Caramaschi, 1997).”

“Loricariids have evolved several modifications of their digestive tracts that function as accessory respiratory organs or hydrostatic organs. These modifications include an enlarged stomach in the *Pterygoplichthys* and *Hypostomus* spp., where veins in the stomach walls uptake oxygen into the bloodstream. Loricariids are facultative air breathers and will only breathe air if subject to hypoxia (Armbruster, 1998; Texas Parks and Wildlife, 2012).”

“Loricariid catfish are generally nocturnal (PlecoInvasion, 2015) and non-migratory (Froese and Pauly, 2014). Although not migratory, loricariids exhibit a tendency to disperse throughout and between aquatic habitats. *Hypostomus* spp. can reportedly cross damp land to reach new water bodies if necessary (Texas Parks and Wildlife, 2012; Hoover et al., 2014). According to Gerstner (2007), the dispersal and station-holding ability of *Hypostomus* spp. in flowing water is facilitated by diverse behaviours distinctive to the unusual morphology of the group. These include the ability to hold onto solid substrates using the oral disc (suckermouth), pelvic fin beats, and hooking and bracing using the studded spines of the pectoral fins. These behaviours enable even comparatively small individuals (approximately 80 mm total length) to negotiate flows up to 145 cm/s. Consequently, a single population can quickly colonize adjacent water bodies (Hoover et al., 2014).”

From Nico and Neilson (2015):

“Occurs in quiet, slow-moving waters and swamps of the lower reaches of rivers between the lower falls and the estuarine zone (Weber et al. 2012).”

Human Uses

From Froese and Pauly (2013):

“Fisheries: subsistence fisheries; aquarium: highly commercial”

“Is cultured in ponds in Singapore and Hong Kong for the aquarium trade, where it is very popular [Baensch and Riehl 1985].”

From CABI (2015):

“According to Sterba (1966), the ornamental trade in ‘suckermouth catfishes’ began in 1893 with commercial imports of *H. plecostomus*. *Hypostomus* spp. were common in the ornamental trade in the 1960s and 1970s, when loricariids were exported from Venezuela, Suriname and the Guyanas (the natural distribution of *H. plecostomus*) (PlanetCatfish, 2015).”

“*H. plecostomus* are consumed in parts of their native range (Burgess, 1989) and in Mexico (around the Inferinillo Reservoir) (Hoover et al., 2014).”

“In Mexico, *Hypostomus* and *Pterygoplichthys* sp. have been used to produce collagen, fish paste and fishmeal (Mendoza-Alfaro et al., 2009).”

“During the 1960s, *H. plecostomus* was used to control algae in pools at a zoo in Texas (Barron, 1964). They have also been introduced into the Balsas Basin, Mexico, to control macrophytes and algae (Mendoza-Alfaro et al., 2009). It is not recorded whether these attempts at biological control were successful.”

Diseases

No records of OIE reportable diseases were found.

From Froese and Pauly (2013):

“White spot Disease, Parasitic infestations (protozoa, worms, etc.)
Skin Flukes, Parasitic infestations (protozoa, worms, etc.)
Velvet Disease, Parasitic infestations (protozoa, worms, etc.)”

Threat to Humans

From Froese and Pauly (2013):

“Harmless”

3 Impacts of Introductions

From Nico and Neilson (2015):

“In Texas, Hubbs et al. (1978) reported possible local displacement of algae-feeding native fishes such as *Campostoma anomalum* by *Hypostomus*, and López-Fernández and Winemiller (2005) suggest that reductions in *Dionda diaboli* abundance in portions of San Felipe Creek are the result of population increases of *Hypostomus*. Because of their abundance in Hawaii, introduced *Hypostomus*, *Pterygoplichthys*, and *Ancistrus* may compete for food and space with native stream species (Devick 1989; Sabaj and Englund 1999).”

From Marambe et al. (2011):

“The tank cleaner (*Hypostomus plecostomus*) can out-compete native biota. The species is an omnivore with a diet varying from plankton to plant matter and invertebrates. Further invasion to inland waters may pose a threat to endemic fish species (Wijethunga and Epa 2008). The scrape feeding habits of the tank cleaner could change habitat quality, leading to detrimental effects on co-occurring species (Amarasinghe et al. 2006).”

From CABI (2015):

“Economic impacts of introduced populations of *Hypostomus* and *Pterygoplichthys* sp. have been quantified for commercial tilapia fisheries in Florida and Mexico (Mendoza-Alfaro et al., 2009). During the period 1993-2006, tilapia catch in six lakes in Florida decreased from 45-80% of the total catch to 17-30% of the total catch after *Hypostomus* and *Pterygoplichthys* sp. became established. Concurrently, the representation of loriciids increased to 11-65% of the commercial catch (Hoover et al., 2014).”

“The tilapia catch in a reservoir in Mexico decreased 83% after proliferation of *Hypostomus* and *Pterygoplichthys* sp.. As a result, individual fishermen spend an additional \$1400-\$2600/year to replace damaged nets, work an additional 2 hr/day, and lose more than \$29,000 (US) per year. Total economic losses are approximately \$16.4 million: \$11.63 million from commercial fishing (losses in gear, hours worked, revenue from catch, health status), \$4.74 million from natural capital (water quality, shoreline formation, native fauna), and an unknown quantity from effects on aquarium trade (sale of illegally traded wild-caught *Hypostomus* and *Pterygoplichthys* sp.) (Hoover et al., 2014).”

“The burrows created by *Hypostomus* sp. during reproduction may cause erosion, sedimentation and increased turbidity. Bank failure, shoreline collapse and terracing have been observed in Mexico, Texas, and Florida where burrow densities were high (Hoover et al., 2014).”

“Grazing *H. plecostomus* may reduce algal standing crops and composition. Extensive grazing may promote a change in algal composition from green algae-dominated communities to diatoms (Flecker, 1992) or diatom-dominated communities to blue-green algae (Power, 1984). Resultant impacts include reduced quality of habitat for algae-dwelling invertebrates and fishes, and reduction in food sources for other grazing aquatic organisms (Hoover et al., 2014).”

“Impacts on aquatic biodiversity have been observed as a result of introduced populations of *H. plecostomus* in Texas (San Antonio and San Marcos rivers, and San Felipe Creek). *H. plecostomus* may compete for resources (food and habitat) with sympatric fishes and aquatic organisms, disturb nest sites, eat eggs of native fishes and disrupt trophic flows and nutrient cycling aquatic habitats.”

“In the San Antonio River, *H. plecostomus* has been implicated in the reduced abundance of the algae-eating central stoneroller *Campostoma anomalum* (Hubbs et al., 1978; Hoover et al., 2014).”

“In San Felipe Creek, *H. plecostomus* is believed to be impacting populations of the IUCN endangered Devils River minnow *Dionda diaboli*. *D. diaboli* was once abundant in San Felipe Creek, but the species has undergone a major decrease in abundance concurrent with the dramatic increase in the population of *H. plecostomus* (Howells, 2005). *D. diaboli* is an algivore and is probably subject to resource competition with *H. plecostomus* (Hoover et al., 2014). Other algal-feeding species have also declined, including the native snail *Elimia comalensis* (Howells, 2005).”

“In the San Marcos River, considerable research has been conducted on the biology and ecology of introduced populations of *H. plecostomus*. Pound et al. (2011) investigated the diet of introduced populations of *H. plecostomus* from the San Marcos River using gut contents and stable isotope analyses. They found that *H. plecostomus* primarily consumed amorphous detritus with small quantities of filamentous red algae and picoplankton. They concluded that the large populations of *H. plecostomus* in the San Marcos River probably compete with several native herbivorous fishes and may be disrupting trophic flows and nutrient cycling in spring-influenced streams of central and west Texas.

One of the herbivorous fishes impacted by *H. plecostomus* in the San Marcos River is the IUCN endangered fountain darter *Etheostoma fonticola* (Hoover et al., 2014). *E. fonticola* deposits its eggs on algae and is believed to be impacted by loss of spawning habitat and egg predation. Cook-Hildreth (2008) conducted experiments on the egg survival of *E. fonticola* and the results suggested that survival was reduced in the presence of *H. plecostomus*. The observation of *E. fonticola* eggs in the stomach of *H. plecostomus* indicated that direct predation of eggs also occurs.”

“Scott et al. (2012) reported that *H. plecostomus* has a wide range and occurs in high densities in the San Marcos River. They conducted mesocosm experiments to determine the impacts of *H. plecostomus* on aquatic ecosystem function and found that it impacted on ecosystems by decreasing periphyton biomass, altering periphyton nutrient ratios, and facilitating detrital decomposition. The presence of *H. plecostomus* altered the aquatic invertebrate community composition in leaf packs and produced ecosystem engineering effects by altering the benthic habitat. Mesocosm experiments by Hoover et al. (2013) demonstrated that *Hypostomus* sp. and *Pterygoplichthys* sp. did not impact water quality or an insectivorous fish after three months, but reduced the abundance of a floating macrophyte, increased phytoplankton-based turbidity and eliminated periphyton.”

“Hoover et al. (2014) theorized that *H. plecostomus* can monopolize nutrient resources in the San Marcos River due to the species rapid maturation, high densities and longevity. The large size and high density of *H. plecostomus* may constitute a significant phosphorus sink in the oligotrophic San Marcos River system. This may lead to reduced primary productivity in the form of a reduction in algal standing crops, which may in turn may impact secondary productivity and invertebrate standing crops.”

From Pallewatta et al. (2003):

“*Hypostomus plecostomus* (tank cleaner), a species imported to Sri Lanka by the ornamental fish industry, has been observed to attach itself by its ventral sucker to the bodies of larger fish. When it detaches, the slime layer covering the outside of the fish which acts as a protective covering is also removed, making the host susceptible to diseases (Bambaradeniya et al., 2001).”

4 Global Distribution



Figure 1. Known global distribution of *Hypostomus plecostomus*. Map from GBIF Secretariat (2017).

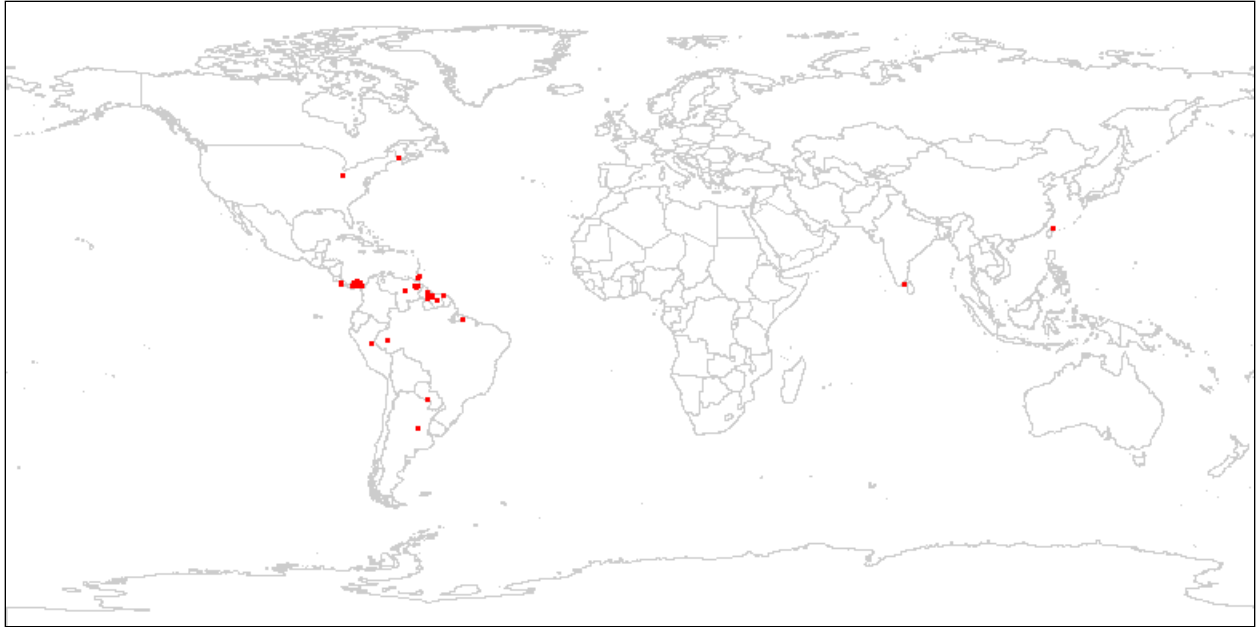


Figure 2. Known global distribution of *Hypostomus plecostomus*. Map from Froese and Pauly (2015).

Locations reported in Canada and Ohio were from aquarium collections and not wild observations (GBIF Secretariat 2017). Locations in South America that were south of the described native range were not included due to confusion about the species identification of those populations (CABI 2015). The record England (Zięba et al. 2010) did not have more specific location data. None of these locations were used as source points in the climate match.

5 Distribution Within the United States



Figure 3. Known distribution of *Hypostomus plecostomus* within the United States. Map from USGS NAS Database (Nico and Neilson 2015).

The record in Nevada did not represent an established population (CABI 2015) and was not used as a source point in the climate match.

6 Climate Matching

Summary of Climate Matching Analysis

The climate match for *Hypostomus plecostomus* was very high through Texas, Florida, the southern Atlantic coast, the southern Pacific Coast, and along the border with Mexico. Parts of the Middle Atlantic States and the Great Lakes Basin had a medium to high match. Elsewhere had a low match. The Climate 6 score (Sanders et al. 2014; 16 climate variables; Euclidean distance) for the contiguous U.S. was 0.205, high, and individually high in Arizona, Arkansas, California, Florida, Georgia, Louisiana, Maryland, Mississippi, New Mexico, North Carolina, Oklahoma, South Carolina, Texas, and Virginia.

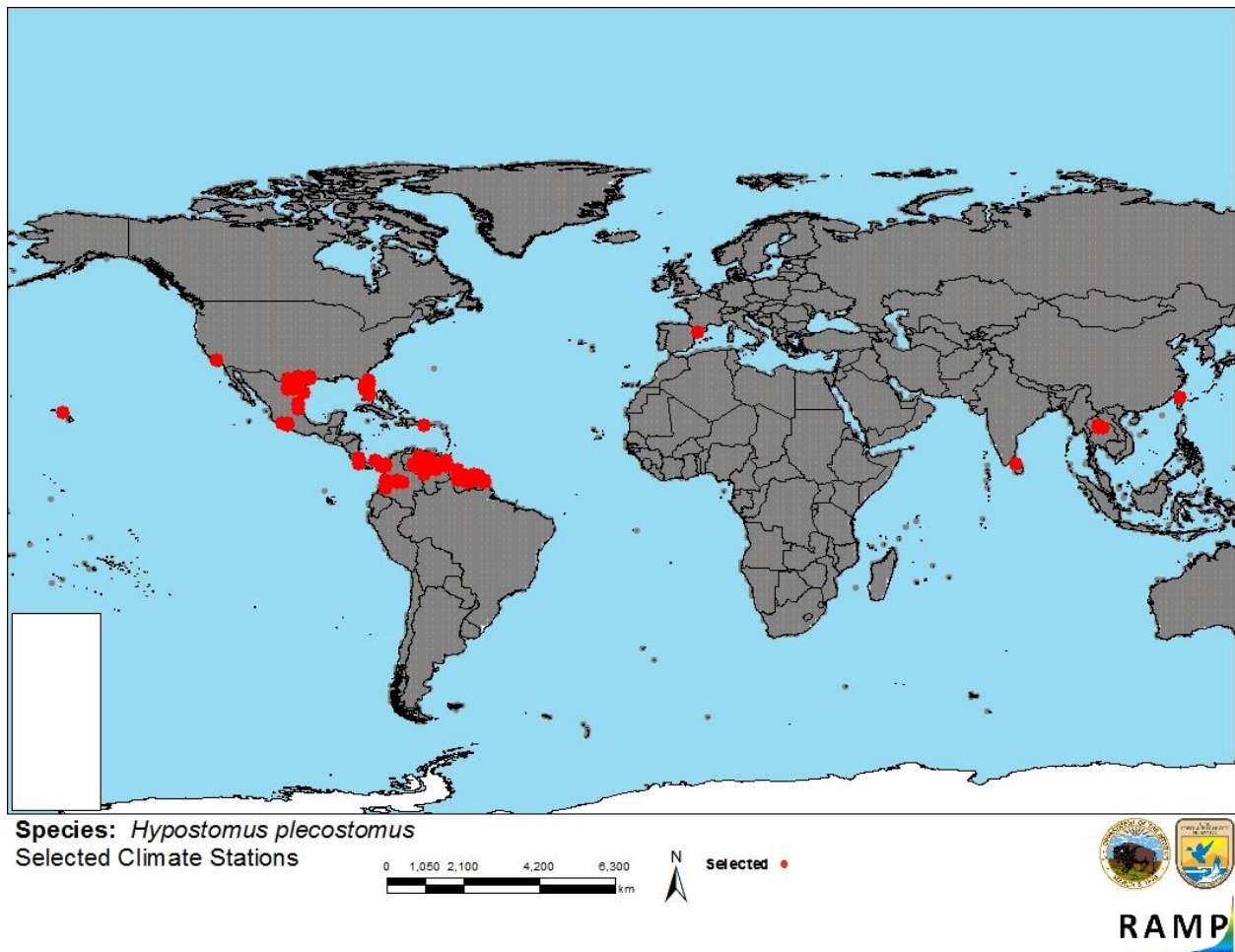


Figure 4. RAMP (Sanders et al. 2014) source map showing weather stations selected as source locations (red) and non-source locations (grey) for *Hypostomus plecostomus* climate matching. Source locations from CABI (2015), Froese and Pauly (2015), USGS NAS Database (Nico and Neilson 2015), and GBIF Secretariat (2017).

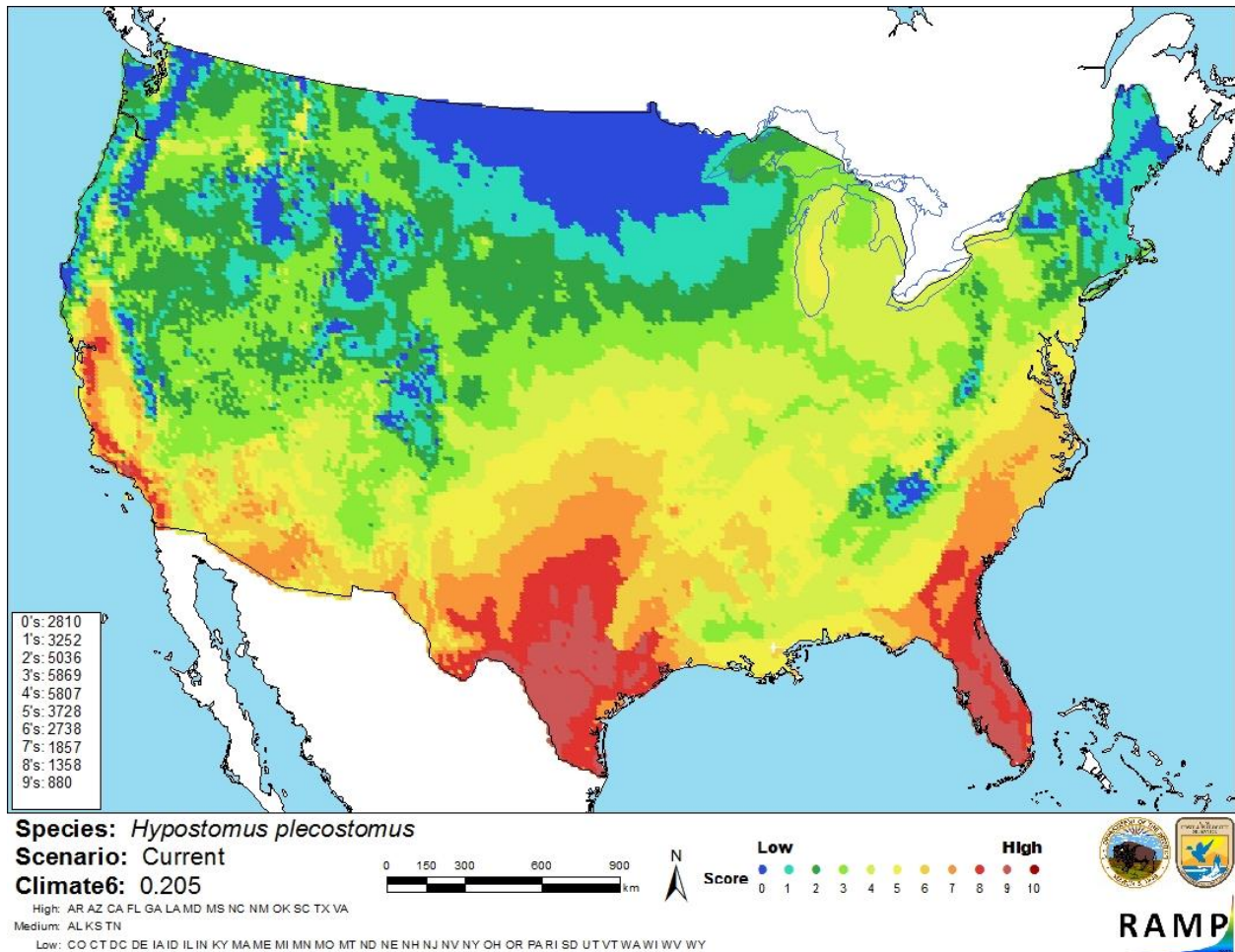


Figure 5. Map of RAMP (Sanders et al. 2014) climate matches for *Hypostomus plecostomus* in the contiguous United States based on source locations reported by CABI (2015), Froese and Pauly (2015), USGS NAS Database (Nico and Neilson 2015), and GBIF Secretariat (2017). 0 = Lowest match, 10 = Highest match.

The High, Medium, and Low Climate match Categories are based on the following table:

Climate 6: Proportion of (Sum of Climate Scores 6-10) / (Sum of total Climate Scores)	Climate Match Category
$0.000 \leq X < 0.005$	Low
$0.005 < X < 0.103$	Medium
≥ 0.103	High

7 Certainty of Assessment

The certainty of assessment is medium. There was a good amount of information available from reliable sources for this species. Many records of introductions and impacts of introductions were found for *Hypostomus plecostomus*. There is some doubt to the native distribution of *H.*

plecostomus. Many records were incomplete and doubt has been cast on the species identification of populations in South America outside of *H. plecostomus*' native range.

8 Risk Assessment

Summary of Risk to the Contiguous United States

The history of invasiveness for *Hypostomus plecostomus* is high. Established populations easily expand their range in favorable conditions. There are many well documented ecological and economic impacts due to introductions of *Hypostomus plecostomus*. The climate match of this species is high. Most of the contiguous United States had at least a medium match, many places had high matches. The certainty of assessment is medium. The overall risk assessment category is high.

Assessment Elements

- **History of Invasiveness (Sec. 3): High**
- **Climate Match (Sec. 6): High**
- **Certainty of Assessment (Sec. 7): Medium**
- **Remarks/Important additional information** The identity of the species in this genus that is established in the United States is uncertain (NatureServe (2017)).
- **Overall Risk Assessment Category: High**

9 References

Note: The following references were accessed for this ERSS. References cited within quoted text but not accessed are included below in Section 10.

CABI. 2015. *Hypostomus plecostomus*. [original text by M. Maddern]. In Invasive Species Compendium. CAB International, Wallingford, UK. Available: <http://www.cabi.org/isc/datasheet/114927>. (December 2015).

Eschmeyer, W. N., R. Fricke, and R. van der Laan, editors. 2017. Catalog of fishes: genera, species, references. Available: <http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>. (May 2017).

FAO (Fisheries and Agriculture Organization of the United Nations). 2013. Database on introductions of aquatic species. FAO, Rome. Available: <http://www.fao.org/fishery/introsp/search/en>. (May 2013).

Froese, R., and D. Pauly, editors. 2013. *Hypostomus Plecostomus* (Linnaeus, 1758). FishBase. Available: <http://www.fishbase.org/summary/Hypostomus-plecostomus.html>. (May 2013).

- GBIF Secretariat. 2017. GBIF backbone taxonomy: *Hypostomus Plecostomus* (Linnaeus, 1758). Global Biodiversity Information Facility, Copenhagen. Available: <http://www.gbif.org/species/5202178>. (May 2017).
- ITIS (Integrated Taxonomic Information System). 2013. *Hypostomus Plecostomus* (Linnaeus, 1758). Integrated Taxonomic Information System, Reston, Virginia. Available: http://www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=164342. (May 2013).
- Maceda-Veiga, A., J. Escribano-Alacid, A. de Sostoa, and E. García-Berthou. 2013. The aquarium trade as a potential source of fish introductions in southwestern Europe. *Biological Invasions*. [online] DOI 10.1007/s10530-013-0485-0.
- Marambe, B., P. Silva, S. Ranwala, J. Gunawardena, D. Weerakoon, S. Wijesundara, L. Manawadu, N. Atapattu, and M. Kurukulasuriya. 2011. Invasive alien fauna in Sri Lanka: national list, impacts and regulatory framework. Pages 445–450 in C. R. Veitch, M. N. Clout, and D. R. Towns, editors. *Island invasions: eradication and management*. IUCN, Gland, Switzerland.
- NatureServe. 2017. NatureServe explorer: an online encyclopedia of life. Version 7.1. NatureServe, Arlington, Virginia. Available: <http://explorer.natureserve.org>. (May 2017).
- Nico, L., and M. Neilson. 2015. *Hypostomus plecostomus* (Linnaeus, 1758). U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, Florida. Available: <http://nas.er.usgs.gov/queries/FactSheet.aspx?speciesID=761>. (December 2015).
- Pallewatta, N., J. K. Reaser, and A. T. Gutierrez, editors. 2003. *Invasive alien species in south-southeast Asia: national reports & directory of resources*. Global Invasive Species Programme, Cape Town, South Africa.
- Sanders, S., C. Castiglione, and M. Hoff. 2014. Risk assessment mapping program: RAMP. U.S. Fish and Wildlife Service.
- Zięba, G., G. H. Copp, G. D. Davies, P. Stebbing, K. J. Wesley, and J. R. Britton. 2010. Recent releases and dispersal of non-native fishes in England and Wales, with emphasis on sunbleak *Leucaspius delineatus* (Heckel, 1843). *Aquatic Invasions* 5(2):155–161.

10 References Quoted But Not Accessed

Note: The following references are cited within quoted text within this ERSS, but were not accessed for its preparation. They are included here to provide the reader with more information.

- Amarasinghe, U. S., R. R. A. R. Shirantha, and M. J. S. Wijeyaratne. 2006. Some aspects of ecology of endemic freshwater fishes of Sri Lanka. In C.N.B. Bambaradeniya, editor. *The fauna of Sri Lanka: status of taxonomy, research and consecration*. The World Conservation Union (IUCN), Colombo, Sri Lanka.

- Armbruster, J. W. 1998. Modifications of the digestive tract for holding air in Loricariid and Scoloplacid Catfishes. *Copeia* 1998(3):663–675.
- Armbruster, J. W. 2004. Phylogenetic relationships of the suckermouth armoured catfishes (Loricariidae) with emphasis on the Hypostominae and the Ancistrinae. *Zoological Journal of the Linnean Society* 141(1):1–80.
- Armbruster, J. W., L. A. Tansey, and N. K. Lujan. 2007. *Hypostomus rhanthos* (Siluriformes: Loricariidae), a new species from southern Venezuela. *Zootaxa* 1553:59–68.
- Azevedo, P. 1938. O cascudo dos açudes nordestinos *Plecostomus plecostomus*. *Arquivos do Instituto Biológico* 211–224.
- Baensch, H. A., and R. Riehl. 1985. *Aquarien atlas. Band 2.* Mergus, Verlag für Natur-und Heimtierkunde GmbH, Melle, Germany.
- Bambaradeniya, C. N. B., S. P. Ekanayake, and J. Gunawardena. 2001. Preliminary observations on the status of alien invasive biota in natural ecosystems of Sri Lanka. *In* P. Balakrishna, editor. *Report of the Workshop on Alien Invasive Species.* Global Biodiversity Forum, South and Southeast Asia Session, October 1999, Colombo, Sri Lanka.
- Barletta, M., U. Saint-Paul, A. Barletta-Bergan, W. Ekau, and D. Schories. 2000. Spatial and temporal distribution of *Myrophis punctatus* (Ophichthidae) and associated fish fauna in a northern Brazilian intertidal mangrove forest. *Hydrobiologia* 426(1/3):65–74.
- Barron, J. C. 1964. Reproduction and apparent overwinter survival of the suckermouth armor catfish, *Plecostomus* sp., in the headwaters of the San Antonio River. *Texas Journal of Science* 16(4):449–450.
- Britski, H. A., K. Z. de Sz. de Silimon, and B. S. Lopes. 1999. *Peixes do Pantanal. Manual de identificação.* Embrapa. Serviço de Produção -SPI, Brasília, DF, Brazil.
- Burgess, W. E. 1989. *An atlas of freshwater and marine catfishes. A preliminary survey of the Siluriformes.* T.F.H. Publications, Neptune City, New Jersey.
- Camargo, M., and V. Isaac. 2001. Os peixes estuarinos da região norte do Brasil: lista de espécies e considerações sobre sua distribuição geográfica. *Boletim do Museu Paraense Emilio Goeldi, Nova Serie, Zoologia* 17(2):133–157.
- Cardoso, Y. P., F. Brancolini, A. Paracampo, M. Lizarralde, R. Covain, and J. I. Montoya-Burgos. 2016. *Hypostomus formosae*, a new catfish species from the Paraguay River basin with redescription of *H. boulengeri* (Siluriformes: Loricariidae). *Ichthyological Exploration of Freshwaters* 27(1):9–23

- Cook-Hildreth, S. L. 2008. Exotic armoured catfishes in Texas: reproductive biology and effects of foraging on egg survival of native fishes (*Ethoestoma fonticola*, endangered, and *Dionda diabola*, threatened). Master's thesis. Texas State University, San Marcos.
- Courtenay, W. R., Jr., and J. R. Stauffer. 1990. The introduced fish problem and the aquarium fish industry. *Journal of the World Aquaculture Society* 21(3):145–159.
- Devick, W. S. 1989. Disturbances and fluctuations in the Wahiawa Reservoir ecosystem. Project Number F-14-R-13, Job 4, Study I. Division of Aquatic Resources, Hawaii Department of Land and Natural Resources.
- Doadrio, I. 2002. Atlas y libro rojo de los peces continentales de Espana. Direccion General de Conservacion de la Naturaleza. Ministerio de Medio Ambiente, Madrid.
- Durante, S., and F. G. Araujo. 2002. Fecundity of the *Hyposomus affinis* (Siluriformes, Loricariidae) in the Lajes Reservoir, Rio de Janeiro, Brazil. *Revista de biologia tropical* 50(1):193–197.
- Elvira, B., and A. Almodovar. 2001. Freshwater fish introductions in Spain: facts and figures at the beginning of the 21st century. *Journal of Fish Biology* 59:323–331.
- Fernholm, B., and A. C. Wheeler. 1983. Linnaean fish specimens in the Swedish Museum of Natural History, Stockholm. *Zoological Journal of the Linnean Society* 78(3):199–286.
- Ferraris, C. J., Jr. 2003. Auchenipteridae, Pimelodidae, Aspredinidae, and Loricariidae. Pages 853–865 in K. E. Carpenter. *The living marine resources of the Western Central Atlantic. Volume 2: bony fishes part 1 (Acipenseridae to Grammatidae)*. FAO species identification guide for fishery purposes and American Society of Ichthyologist and Herpetologists, Special Publication 5, FAO, Rome.
- Ferraris, C. J., Jr. 2007. Checklist of catfishes, recent and fossil (Osteichthyes: Siluriformes), and catalogue of siluriform primary types. *Zootaxa* 1418:1–628.
- Flecker, A. S. 1992. Fish trophic guilds and the structure of a tropical stream: weak vs. strong indirect effects. *Ecology* 73:927–940.
- Florida Fish and Wildlife Conservation Commission (Florida FWC). 2000. Florida freshwater fisheries database via MARIS. Florida Fish and Wildlife Conservation Commission, Tallahassee.
- Florida Fish and Wildlife Conservation Commission. 2015. Florida Fish and Wildlife Conservation Commission. Available: <http://myfwc.com/>.
- Froese, R., and D. Pauly. 2014. FishBase. Available: <http://www.fishbase.org>.

- Galvis, G., J. I. Mojica, and M. Camargo. 1997. Peces del Catatumbo. Asociación Cravo Norte, Santafé de Bogotá, D.C.
- Gante, H. F., L. Morira da Costa, J. Micael, and M. J. Alves. 2008. First record of *Barbonymus schwanenfeldii* (Bleeker) in the Iberian Peninsula. *Journal of Fish Biology* 72:1089–1094.
- Gerstner, C. L. 2007. Effect of oral suction and other friction-enhancing behaviors on the station-holding performance of suckermouth catfish (*Hypostomus* spp.). *Canadian Journal of Zoology* 85(1):133–140.
- Gleason, K. 2004. Where have all the minnows gone? *Del Rio News-Herald* (July 18).
- Grier, H. 1980. *Plecostomus*. *Freshwater and Marine Aquarium* 3(8):23–26, 85.
- Hoover, J. J., N. M. Hahn, and J. A. Collins. 2013. Demonstrating the ecosystem effects of armored suckermouth catfishes (Loricariidae): a feasibility study using mesocosms. ANSRP Technical Notes Collection, ERDC/TN ANSRP-13-2, Vicksburg, Mississippi.
- Hoover, J. J., K. J. Killgore, and A. F. Cofrancesco. 2004. Suckermouth catfishes: threats to aquatic ecosystems of the United States? *Aquatic Nuisance Species Research Program Bulletin* 1-13.
- Hoover, J. J., C. E. Murphy, and J. Killgore. 2014. Ecological impacts of suckermouth catfishes (Loricariidae) in North America: a conceptual model. *Aquatic Nuisance Species Research Program Bulletin* 14-1:1–13.
- Howells, R. G. 1992. Annotated list of non-native fishes, mollusks, crustaceans, and aquatic plants, in Texas waters. Texas Parks and Wildlife Department, Management Data Series 78, Austin.
- Howells, R. G. 2005. Exotic suckermouth catfishes (family Loricariidae) in Texas waters. Texas Parks and Wildlife Department.
- Hubbs, C., R. J. Edwards, and G. P. Garrett. 1991. An annotated checklist of freshwater fishes of Texas, with key to identification of species. *Texas Journal of Science Supplement* 43(4):1–56.
- Hubbs, C., T. Luciere, G. P. Garrett, R. J. Edwards, S. M. Dean, and E. Marsh. 1978. Survival and abundance of introduced fishes near San Antonio, Texas. *The Texas Journal of Science* 30(4):369–376.
- Hugg, D. O. 1996. MAPFISH georeferenced mapping database. Freshwater and estuarine fishes of North America. Life Science Software. Dennis O. and Steven Hugg, Edgewater, Maryland.

- Isbrücker, I. J. H. 1980. Classification and catalogue of the mailed Loricariidae (Pisces, Siluriformes). Verslagen en Technische Gegevens, Instituut voor Taxonomische Zoölogie, Universiteit van Amsterdam 22:1–181.
- Isbrücker, I. J. H. 2001. Nomenklator der Gattungen und Arten der Harnischwelse, Familie Loricariidae Rafinesque, 1815 (Teleostei, Ostariophysi). DATZ-Sonderheft Harnischwelse 2:25–32.
- Isbrücker, I. J. H. 2002. Nomenclator of the 108 genera with 692 species of the mailed catfishes, family Loricariidae Rafinesque, 1815 (Teleostei, Ostariophysi). Cat Chat, Journal of the catfish study group (UK) 3(1):11–30.
- Linnaeus, C. 1758. Systema Naturae, Ed. X. (Systema naturae per regna tria naturae, secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis. Tomus I. Editio decima, reformata.) Holmiae 1:1–824.
- Liu, Y., X. T. Lin, J. Sun, P. F. Zhang, and G. Z. Chen. 2011. Fish community changes in Huizhou segment of Dongjiang River. Chinese Journal of Zoology 46(2):1–11.
- Lopez, H. L., R. C. Menni, and A. M. Miguelarena. 1987. List of freshwater fish of Argentina. (Lista de los peces de agua dulce de la Argentina.) Biología Acuática 12.
- López-Fernández and Winemiller. 2005. [Source material did not give full citation for this reference.]
- Lopez Macias, J. N., F. Garcia Vallejo, E. Rubio Rincón, A. Castillo Giraldo, and F. Cerón. 2009. Genetic diversity of Bocachico (*Prochilodus reticulatus*) of the Cuenca Alta of Río Cauca (Colombia). (Diversidad genética del Bocachico (*Prochilodus reticulatus*) de la Cuenca Alta del Río Cauca (Colombia).) Acta Biológica Paranaense 38(1/4):113–138.
- Ma, X., X. Bangxi, W. Yindong, and W. Mingxue. 2003. Intentionally introduced and transferred fishes in China's inland waters. Asian Fisheries Society 16(3/4):279–290.
- Mazzoni, R., and E. P. Caramaschi. 1995. Size structure, sex ratio and onset of sexual maturity of two species of *Hypostomus*. Journal of Fish Biology 47(5):841–849.
- Melo, B. F., R. C. Benine, R. Britzke, C. S. Gama, and C. Oliveira. 2016. An inventory of coastal freshwater fishes from Amapá highlighting the occurrence of eight new records for Brazil. ZooKeys 606:127–140.
- Mendoza-Alfaro, R. E., B. Cudmore, R. Orr, J. P. Fisher, S. C. Balderas, W. R. Courtenay, and P. K. Osorio. 2009. Trinational risk assessment guidelines for aquatic alien invasive species - test cases for the snakeheads (Channidae) and armoured catfishes (Loricariidae) in North American inland waters. CEC Project Report. Commission on Environmental Cooperation, Montreal.

- Nelson, J. S., E. J. Crossman, H. Espinosa Pérez, L. T. Findley, C. R. Gilbert, R. N. Lea, and J. D. Williams. 2004. Common and scientific names of fishes from the United States, Canada, and Mexico, 6th edition. American Fisheries Society, Special Publication 29, Bethesda, Maryland.
- Nico, et al. 2015. [Source material did not give full citation for this reference.]
- Nomura, H., and I. M. de Mueller. 1980. Biology of the armored catfish, *Plecostomus hermanni* Ihering, 1905 Mogi Guacu, Sao Paulo (Osteichthyes, Loricariidae). (Biologia do cascudo, *Plecostomus hermanni* Ihering, 1905 do Rio Mogi Guaçu, São Paulo (Osteichthyes, Loricariidae).) Revista Brasileira de Biologia 40(2):267–275.
- Ortega, A., O. Murillo, M. Y. Pimienta, and J. Sterling. 1999. Characterization of the native fish fauna of rivers in the upper basin of the Rio Cauca department of Valle del Cauca (Caracterización de la ictiofauna nativa de los ríos de la cuenca alta del Río Cauca en el departamento del Valle del Cauca, Corporación Autónoma Regional del Valle del Cauca, CVC). Corporacion Autonoma Regional del Valle del Cauca, CVC.
- Page, L. M., and B. M. Burr. 1991. A field guide to freshwater fishes of North America north of Mexico. The Peterson Field Guide Series, volume 42. Houghton Mifflin, Boston.
- Page, L. M., H. Espinosa-Pérez, L. D. Findley, C. R. Gilbert, R. N. Lea, N. E. Mandrak, R. L. Mayden, and J. S. Nelson. 2013. Common and scientific names of fishes from the United States, Canada, and Mexico, 7th edition. American Fisheries Society, Special Publication 34, Bethesda, Maryland.
- PlanetCatfish. 2015. PlanetCatfish. Aquatic Republic Network. Available: <http://www.planetcatfish.com/>.
- PlecoInvasion. 2015. General biology and invasion information. Available: http://www.plecoinvasion.org/index_files/Page386.htm.
- Pound, K. L., W. H. Nowlin, D. G. Huffman, and T. H. Bonner. 2011. Trophic ecology of a nonnative population of suckermouth catfish (*Hypostomus plecostomus*) in a central Texas spring-fed stream. Environmental Biology of Fishes 90(3):277–285.
- Power, M. E. 1984. Habitat quality and the distribution of algae-grazing catfish in a Panamanian stream. Journal of Animal Ecology 53:357–374.
- Reis, R. E., S. O. Kullander, and C. J. Ferraris, Jr., editors. 2003. Check list of the freshwater fishes of South and Central America. CLOFFSCA. EDIPUCRS, Porto Alegre, Brazil.
- Robinson, L., and J. Culbertson. 2005. A synoptic survey for nonindigenous ichthyofauna in selected tidal bayous of Galveston Bay. Report prepared for the Texas Commission on Environmental Quality, Houston.

- Sabaj, M. H., and R. A. Englund. 1999. Preliminary identification and current distribution of two suckermouth armored catfishes (Loricariidae) introduced to Oahu streams. Bishop Museum Occasional Papers 59:50–55.
- Sarmiento, J., R. Bigorne, F. M. Carvajal-Vallejos, M. Maldonado, E. Leciak, and T. Oberdorff, editors. 2014. Peces de Bolivia/Bolivian fishes. IRD-Biofresh (EU).
- Scharpf, C. 2006. Annotated checklist of North American freshwater fishes, including subspecies and undescribed forms. Part II: Catostomidae ... [through] Mugilidae. American Currents 32(4):1–40.
- Scott, S. E., C. L. Pray, W. H. Nowlin, and Y. X. Zhang. 2012. Effects of native and invasive species on stream ecosystem functioning. Aquatic Sciences 74(4):793–808.
- Shafland, P. L. 1976. The continuing problem of non-native fishes in Florida. Fisheries 1(6):25.
- Shafland, P. L., and J. M. Pestrak. 1982. Lower lethal temperatures for 14 non-native fishes in Florida. Environmental Biology of Fishes 7:149–156.
- Silvano, R. A. M., and A. Begossi. 2001. Seasonal dynamics of fishery at the Piracicaba River (Brazil). Fisheries Research 51(1):69–86.
- Sterba, G. 1966. Freshwater fishes of the world. T.H.F Publications, Hong Kong.
- Texas Parks and Wildlife Department. 2001. Fish records: water body - all tackle. Texas Parks and Wildlife Department, April 24, 2001.
- Texas Parks and Wildlife. 2012. Freshwater aquarium hobbyists and invasive species in the Houston-Galveston region. Final Project Report. Houston Advanced Research Center (HARC), Texas.
- USGS NAS. 2015. USGS Nonindigenous Aquatic Species Database. Gainesville, Florida. Available: <http://nas.er.usgs.gov/>.
- Weber, C., R. Covain, and S. Fisch-Muller. 2012. Identity of *Hypostomus plecostomus* (Linnaeus, 1758), with an overview of *Hypostomus* species from the Guianas (Teleostei: Siluriformes: Loricariidae). Cybium 36(1):195–227.
- Wheeler, A. C. 1989. Further notes on the fishes from the collection of Laurens Theodore Gronovius (1730-1777). Zoological Journal of the Linnean Society 95(3):205–218.
- Whiteside, B. G., and C. Berkhouse. 1992. Some new collections locations for six fish species. The Texas Journal of Science 44(4):494.
- Wijesekera, G. A. W., and C. N. B. Bambardeniya. 2007. Invasive alien species. The National Atlas of Sri Lanka, 2nd edition. Survey Department, Colombo, Sri Lanka.

- Zawadzki, C. H., L. F. C. Tencatt, and O. Froehlich. 2014. A new unicuspid-toothed species of *Hypostomus* Lacépède, 1803 (Siluriformes: Loricariidae) from the rio Paraguai basin. *Neotropical Ichthyology* 12(1):97–104.
- Zawadzki, C. H., C. Weber, and C. S. Pavanelli. 2010. A new dark-saddled species of *Hypostomus* (Siluriformes: Loricariidae) from the upper rio Paraguay basin. *Neotropical Ichthyology* 8(4):719–725.
- Zhang, C.-G., Y.-H. Zhao, et al. 2016. Species diversity and distribution of inland fishes in China. [Source material did not give full citation for this reference.]