

## Astonishing Diversity of Natural Peroxides as Potential Therapeutic Agents

Valery M Dembitsky\*

Institute of Drug Discovery, P.O. Box 45289, Jerusalem 91451, Israel

\*Corresponding author: Dembitsky VM, Institute of Drug Discovery, P.O. Box 45289, Jerusalem 91451, Israel, Tel: +972 526 877 444, E-mail: iddrdo@gmx.com

Received date: January 28, 2015, Accepted date: February 25, 2015, Published date: March 04, 2015

Copyright: © 2015 Dembitsky VM. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Abstract

Peroxides are an interesting group among biological active natural compounds. These metabolites contain a peroxide group (-O-O-) in which each oxygen atom is bonded to the other oxygen and to another atom.  $\beta$ -Oxygen in hydroperoxide group is considered as more active. Present review describes research on more than 230 natural peroxides isolated from plants, algae, and fungi. Intensive searches for new classes of biologically active metabolites produced by terrestrial and marine origin have resulted in the discovery of dozens of compounds possessing high antimalarial, antibacterial, cytotoxic, and other pharmacological activities as an important source of leads for drug discovery.

**Keywords:** Peroxides; Plants; Algae; Fungi; Pharmacological activities

### Introduction

More than 1,000 peroxides (hydro-, endo-, and also acyclic) have been isolated and structurally characterized from natural sources, mainly as constituents of fungi, fungal endophytes, and plants; they also were found in freshwater and marine algae, invertebrates, and other organisms [1-5].

Among naturally occurring hydro- and endoperoxides represented a large group compounds which are shown to possess antimalarial, antibacterial, cytotoxic, and many other activities. In the past several decades, natural peroxides have been isolated from a wide variety of fungi, plants, and marine organisms. Extensive pharmacological screening performed on aquatic and/or terrestrial species resulted in discovery of novel anti-tumor, antibacterial, and mainly antimalarial agents [6-8]. The purpose of this review is to summarize bioactive metabolites of more than 230 natural peroxides, belonging to diverse structural classes: terpenes, steroids, alkaloids, fatty acids, and other compounds.

This paper reviewed more than two hundreds of new and active peroxy natural metabolites produced by plants, algae, fungi, and described their structures, chemistry, and pharmacological activities.

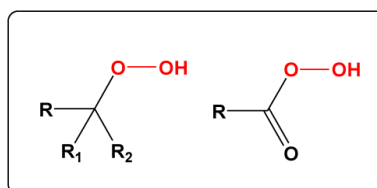
### Fungal Peroxides

#### Hydroperoxides

Mono (or di-) substituted natural products of hydrogen peroxide (dioxidane, HOOH), having the skeleton ROOH (Figure 1), in which R is any organic group(s), called generically as hydroperoxides and are named substitutively by citing the prefix hydroperoxy, and/or peracid(s), respectively. Many different natural hydroperoxides have been isolated from fungi, algae, plant, invertebrates, and animal sources [1-5,9].

Unusual spiro [4.5] decane sesquiterpenes, cordycepol A, B and C (1) (Figure 2), were isolated from the cultured mycelia of parasitic

fungi *Cordyceps ophioglossoides* (family Ophiocordycipitaceae). Isolated compounds showed the cytotoxic activities (IC<sub>50</sub> values in the range of 12-33  $\mu$ g/mL) against HeLa and HepG2 [10].



**Figure 1:** Common structures of mono-hydro peroxide and peracids (the peroxide group is marked in red). R, R1 and R2 mark hydrocarbon moieties.

Eudesmene-type sesquiterpenes, kandenols C (2) and D (3), have been isolated from *Streptomyces* sp. HKI0595 derived from the mangrove plant *Kandelia candel* as weak to moderate inhibitors of *B. subtilis* and *Mycobacterium vaccae* growth [11].

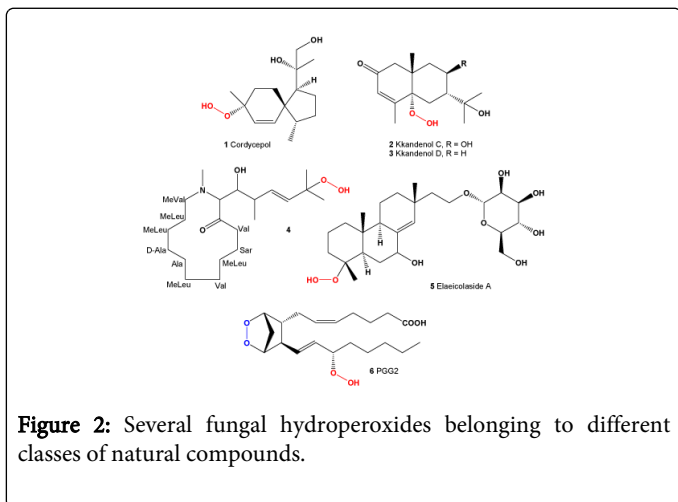
Entomopathogenic species belonging to the genus *Tolypocladium*, *T. terricola*, are known as producers of secondary metabolites and possession of relatively strong mosquitocidal activity [12].

Cyclosporins are produced by certain species of the filamentous fungi, belonging to the genus *Tolypocladium* [2]. Some cyclic peptides and depsipeptides are synthesized in microorganisms by large multienzymes called nonribosomal peptide synthetases. Proven cytotoxic, anti-inflammatory, anticancer, and immunosuppressive activities of some cyclic peptides indicate that these molecules may contribute to the synergistic array of fungal virulence factors and support microbial invasion during fungal infection. Cyclosporin D hydroperoxide (4), was isolated from this cultivated fungus *Tolypocladium terricola* [13].

Several isopimarane-type diterpene glycosides, along with an eremophilane-type sesquiterpene, i.e., elaeicolasides A (5), B and C were isolated from the AcOEt extract of the fermented broth of the ascomycete *Stilbohyphoxylon elaeicola* YMJ173. All these compounds

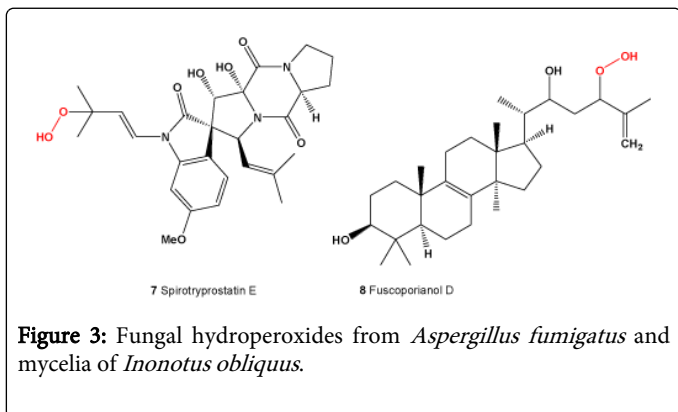
inhibited NO production, detected as nitrite in the culture medium, in activated macrophages without any cytotoxicity at a concentration of 100  $\mu$ M [14].

The opportunistic fungal pathogen *Cryptococcus neoformans* was used for production of several species of prostaglandins (PGE2, PGH2 and 15-keto-PGE2) from arachidonic acid, and including unusual endo- and hydroperoxide PGG2 (6) [15].



**Figure 2:** Several fungal hydroperoxides belonging to different classes of natural compounds.

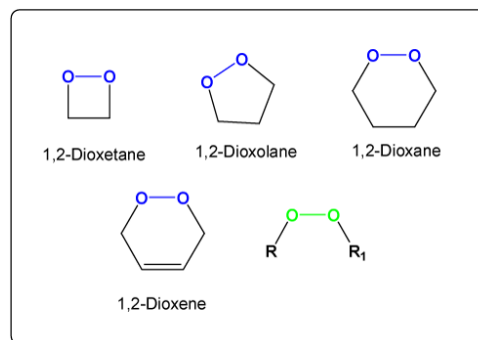
Prenylated indole diketopiperazine alkaloids, spirotryprostatin E (7) has been obtained from the fermentation of *Aspergillus fumigatus* from a holothurian, *Stichopus japonicus* (Lingshan Island, Qingdao, China) [16].



**Figure 3:** Fungal hydroperoxides from *Aspergillus fumigatus* and mycelia of *Inonotus obliquus*.

Sterols are one of the active classes of compounds in *Inonotus obliquus* (known as chaga mushroom) for their effective therapy of many diseases. The results indicated that field-grown mycelia contained lanosterol and inotodiol (45.47% and 25.36% of the total sterols, respectively) and other 10 sterols (comprising the remaining 30.17%) including ergosterol biosynthetic intermediates such as 24-methylene dihydrolanosterol, 4,4-dimethylfecosterol, 4-Me fecosterol, fecosterol and episterol. Column chromatography also led to the isolation of lanosterol, inotodiol, trametenolic acid, fuscoporrianol B and a triterpenoid fuscoporrianol D (8) in field-grown mycelia of *Inonotus obliquus* [17].

**Endoperoxides:** Natural products having the skeleton ROOR<sub>1</sub>, in which R is any organic group(s), called generically as endoperoxides. A lot of different types (Figure 4) natural endoperoxides have been isolated from fungi, algae, plants, and other sources [1-5,9].



**Figure 4:** Common structures of endoperoxides four-membered (1,2-dioxetane), five-membered (1,2-dioxolanes), six-membered (1,2-dioxanes and 1,2-dioxenes), and seven- and more membered endoperoxides (the peroxide group is marked in blue). Acyclic peroxides, R and R<sub>1</sub> marked in green.

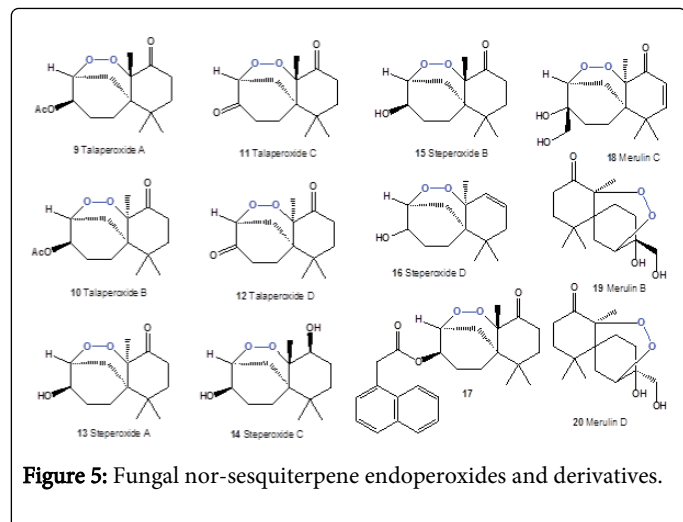
Several nor-sesquiterpene peroxides (Figure 5), named as talaperoxide A (9), B (10), C (11), and D (12), were isolated from culture of fungi *Talaromyces* sp. HN21-3C (family Trichocomaceae). Isolated compounds showed antineoplastic activity against mammary cancer, prostatic carcinoma, uterine cervix carcinoma or hepatic carcinoma [18,19]. Same compounds, talaperoxides A-D (9-12), as well as one known analog, steperoxide B (14, or merulin A), have been isolated from a mangrove endophytic fungus, *Talaromyces flavus* [20]. Talaperoxides (11) and (12) showed cytotoxicity against the five human cancer cell lines with IC<sub>50</sub> values between 0.70 and 2.78  $\mu$ g/mL.

Chamigrane-type metabolites named steperoxides A (13), B (14), C (15) and D (16) were isolated from the hydroid fungus *Steccherinum ochraceum* (family Phanerochaetaceae). Compounds (15 and 16) showed significant antimicrobial activity against *Staphylococcus aureus* at 10 and 5  $\mu$ g/disk [21-23]. Semi-synthetic derivative (17) of the fugally-derived natural product (11) showed the antiparasitic and cytotoxic activity (IC<sub>50</sub>=0.043  $\mu$ M vs. *T. brucei*; IC<sub>50</sub>=13  $\mu$ M vs. Hela cells), respectively [24].

Previously, the same nor-chamigrane endoperoxide, named as merulin A (syn. steperoxide B, 15), B (19), C (18), and D (20), were isolated from the culture broth extract of an endophytic fungus of *Xylocarpus granatum*. Same compounds were isolated from the endophytic fungus *Talaromyces flavus* isolated from the mangrove plant *Sonneratia apetala* [20]. Compounds merulin A and C displayed significant cytotoxicity against human breast (BT474) and colon (SW620) cancer cell lines. Endophytic fungi have been the source of a wide range of structurally interesting and biologically active compounds. The endophytic fungus XG8D, which was isolated from the mangrove plant, *Xylocarpus granatum* (Meliaceae), as the EtOAc extract of this strain showed potent cytotoxic activity against human breast (BT474) and colon (SW620) cancer cell lines. The fungus strain XG8D was classified as a member of the family Meruliaceae (order Polyporales, subclass *Incertae sedis*, class Agaricomycetes, phylum Basidiomycota) from rDNA sequences and LUS phylogeny [25].

Cytosporolides A (21), B (22), and C (23), caryophyllene-derived meroterpenoids with a unique peroxy lactone skeleton, were isolated from cultures of the fungus *Cytospora* sp. All cytosporolides showed

significant antimicrobial activity against the Gram-positive bacteria *Staphylococcus aureus* and *Streptococcus pneumoniae* [18]. Also, compound (23) showed good activity in vitro against dermatophytic fungi and moderate activity against *C. albicans* and *S. aureus*.



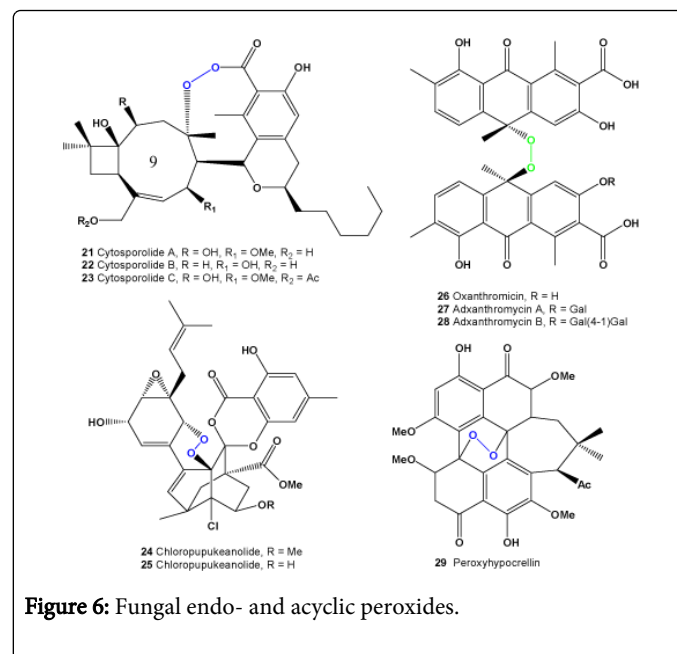
Chloropupekanolides A (24) and B (25), unprecedented spiroketal peroxides, two highly functionalized metabolites featuring a chlorinated pupukeanane core, were isolated from an endophytic fungus *Pestalotiopsis fici*. The compound (24) was reported to have anti-HIV and anti-cancer activities with an inhibitory effect on HIV-1 replication in C8166 cells of 6.9 micromolar (EC<sub>50</sub> value) and IC<sub>50</sub> values of 16.9, 15.5, and 15.9  $\mu$ M against HeLa, MCF-7, and MDA-MB-231 cell lines, respectively [26].

Dimeric anthrone peroxide, named oxanthromycin (26), an antibiotic isolated from an *Actinomadura* sp. SCC 1646 fermentation broth [27,28]. Adxanthromycins A (27) and B (28) are new inhibitors of ICAM-1/LFA-1 mediated cell adhesion molecule isolated from the fermentation broth of *Streptomyces* sp. NA-148. Adxanthromycins A and B inhibited the formation of the JY cell aggregates from 1.5 mg/mL, respectively, in a dose-dependent manner. Components A and B also inhibited a human T cell leukemia cell line SKW-3 adhesion to soluble ICAM-1 in a dose-dependent manner with an IC<sub>50</sub> of 18.8 and 25.0  $\mu$ g/mL, respectively [29,30].

Hypocrellins are dark red pigments having the perylenequinone structure, with photodynamic activity toward microorganisms. These pigments produced by the fungus *Hypocrella bambusae* [31,32], and a parasitic fungus *Shiraia bambusicoia* [33-35]. All isolated metabolites have shown anticancer activities [33,36,37], and antiviral activity against the human immunodeficiency virus (HIV-1) [38]. Natural cytotoxic peroxyhypocrellin (29) was isolated from *S. bambusicoia* [34]. Structures showed in Figure 6.

Bioactive endoperoxide verruculogen [30] (Figure 7) for the first time was isolated from a strain of *Penicillium verruculosum* isolated from peanuts [39]. This compound (30) has also been isolated from a number of other microbiological sources including *Aspergillus caespitosus* [40], *A. fumigatus* [41], *A. fischeri* [42], *Penicillium piscarium* [43], *P. paxilli* [44], *P. piceum*, *P. nigricans*, *P. raistrickii* [43], *P. estinogenum* [45], *P. simplicissimum* [46], *Eupenicillium* sp. [47], and an invasive fungal pathogen *Neosartorya fischeri* (*A. fischerianus*) [48]. A tremorgenic mycotoxin verruculogen (30) is a potent inhibitor of high conductance Ca activated K (maxi-K)

channel, other pharmacological effects of (30) have also been reported [49]. The acetoxy derivative (31) has been isolated from *Penicillium verruculosum* [50]. Other endoperoxide, fumitremorgin A (32), was recovered from a strain of *Aspergillus fumigatus* [51,52]. Fumitremorgin A (32) known as neurotropic metabolite [53] was produced by fungus *Aspergillus fumigatus* [54]. Diketopiperazine alkaloid, 13-oxoverruculogen (33), has been obtained from the fermentation of *Aspergillus fumigatus* from a holothurian, *Stichopus japonicus* (Lingshan Island, Qingdao, China) [55].



Asperversin A (34) and ergosterol peroxide (35) was obtained from the culture of *Aspergillus versicolor*, an endophytic fungus isolated from the marine brown alga *Sargassum thunbergii*. Compound exhibited antibacterial activities against *Escherichia coli* and *Staphylococcus aureus* [56].

Ergosterol peroxide (35) (Figure 8) has been detected in many fungi and fungal endophytes: *Claviceps purpurea* (ergot fungus), *Ganoderma lucidum*, *G. tsugae* and *G. sichuanense* (lingzhi mushrooms) [2], mushroom *Daedalea quercina* [1], *Piptoporus betulinus* (known as the Birch bracket mushroom), *Cryptoporus volvatus* (known as the Grey-Brown Sap Rot mushroom) [2], *Guignardia laricina* (Botryosphaerales), *Lampteromyces japonicus* (Moonlight mushroom, Australia) [1], a necrotrophic fungus *Botrytis cinerea* [2], *Lactarius uvidus* (North American milk-cap mushroom), *Lactarius volemus*, *Cryptoporus volvatus* (known as the Grey-Brown Sap Rot mushroom), *Aspergillus* sp., *A. niger*, *A. oryzae*, *A. flavus*, *A. terreus*, and *A. fumigatus* [2], *Fusarium moniliforme*, *F. osysporum*, *Penicillium rubrum*, *P. sclerotigenum* [1], *Dictyonema glabratum* (a lichen-forming basidiomycete), *Lasiosphaera nipponica*, *Gloeophyllum odoratum*, *Gymnopilus spectabilis*, *Hericium erinaceus* (medicinal, edible mushroom), *Hypsizigus marmoreus*, *Inonotus obliquus* (known as chaga mushroom), *Inonotus radiatus* (syn. *Mensularia radiata*), *Lenzites betulina* (birch mazelgill or multicolor gill polypore fungus), fungus *Meripilus giganteus*, *Microporus flabelliformis* (syn. *Microporus affinis*), *Naematoloma fasciculare* (yellow mushroom syn. *Hypholoma fasciculare*), *Phellinus pini* (known as Red ring rot or White speck), *Phellinus ribis* (medicinal fungus), *Phellinus torulosus* (known as *Phellinus torulosus*),

*Roseoformis subflexibilis* (wood-rotting fungus), *Pyropolyporus fomentarius* (Syn. *Fomes fomentarius*), *Pisolithus tinctorius* (known in Europe as the Bohemian truffle), *Polyporus tuberaster*, and *Pseudephebe pubescens* (lichenized fungi within the Parmeliaceae family) [2], and from the edible mushroom, *Volvariella volvacea* [57].

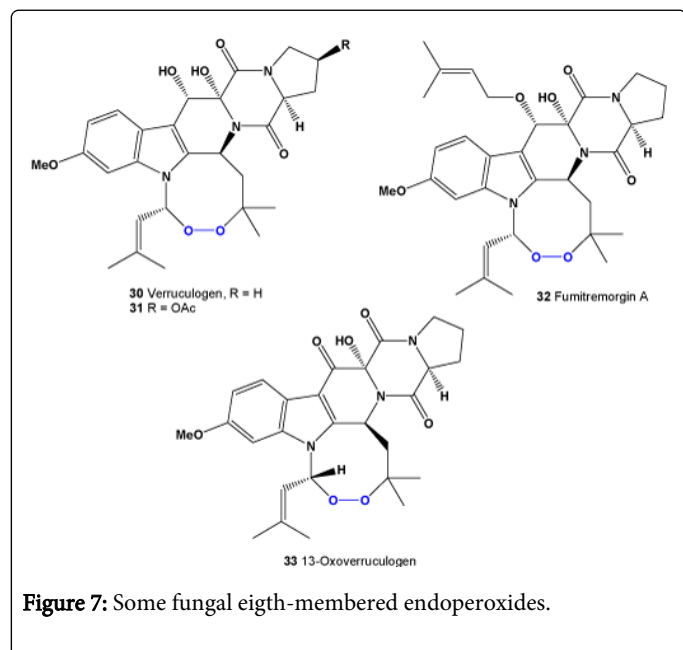


Figure 7: Some fungal eight-membered endoperoxides.

Endophytic fungus No ZZF36 from a brown alga from the South China Sea, produced ergosterol peroxide (26), along with brassicaterol, and ergosterol [58].

The fruiting bodies of the edible mushroom *Gomphus clavatus* (family Gomphaceae) were collected from the wild and extracted with solvents of increasing polarity. Crude extracts were evaluated for their total phenolic content, their antioxidant capacity, and their cytotoxic activity against MCF-7 and PC-3 cancer cell lines. Ergosterol peroxide (26) was one of the most active constituents, with IC<sub>50</sub> values of 35.8 μM and 30.6 μM for MCF-7 and PC-3 cells, respectively, suggesting that the cytotoxic activity of the crude extract could be at least partly attributed to the presence of ergostan derivatives. Those findings suggest that *G. clavatus* can be considered as a medicinal food with antioxidant and chemopreventive activities [59].

Ergosterol peroxide (26) was isolated from the ethanol extract of *Pleurotus eryngii*, an edible mushroom native to Mediterranean regions of Europe, the Middle East, and North Africa, as an inhibitor of osteoclast differentiation. This compound showed an inhibitory effect in a dose-dependent manner and an inhibition rate of up to 62% with low cytotoxicity, even at a concentration as low as 1.0 μg/mL [60]. Ergosterol peroxide (26) isolated from of the deep-sea derived fungus *Aspergillus sp.* CXCTD-06-6a showed activities against P388 and HeLa cell lines [61].

Ergosterol peroxide (26) have also been found in some lichenized ascomycetes: *Cetraria chlorophylla*, *C. islandica*, *Cladonia gracilis*, *Leioderma pycnophorum*, *Pseudocyphellaria encoensis* and *P. pluvialis*, *Lepolichen coccophorus*, *Lobaria pulmonaria*, *Ramalina hierrensis*, *R. tingitana*, *Rhizoplaca melanophthalma*, *Stereocaulon azoreum*, *Peltigera aphthosa* and *P. dolichorrhiza* [1,2].

Methanol extract of fungus *Tremella fuciformis* showed significant neuritogenic activity against PC12 cells. A neuritogenic compounds ergosterol peroxide (26) and 5α,8α-epidioxy-(22E,24R)-ergosta-6,22-diene-3β-ol 3-O-β-D-glucopyranoside (27) were isolated and identified from the methanol extract of fungus *T. fuciformis* [62]. Same compounds were isolated from the fruiting bodies of the Chinese toxic mushroom *Naematoloma fasciculare* [63]. Two peroxides (26 and 28) produced a camphor fungus *Antrodia camphorata* (known as Niu-Chang in Taiwan) [64]. Compound (28) obtained from the fruit bodies of mushroom *Boletus calopus* had weak antifungal action on a few pathogens, such as cucumber wilt disease fungus and wheat scab fungus [65].

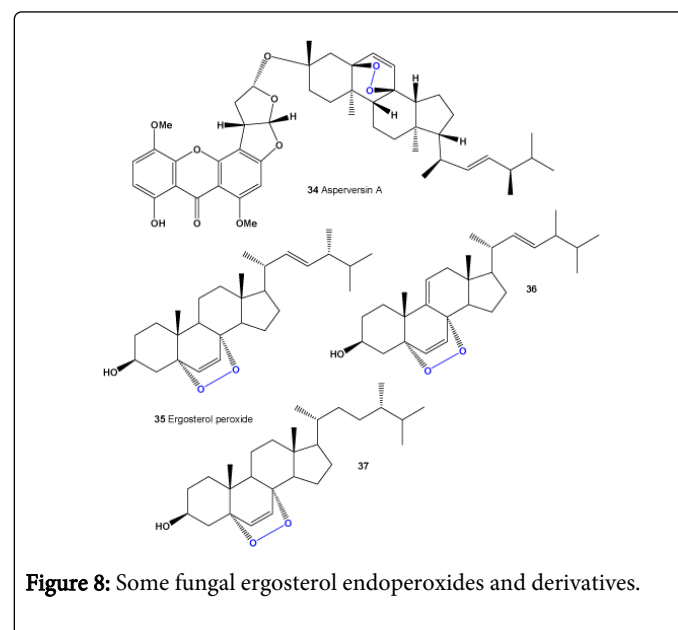


Figure 8: Some fungal ergosterol endoperoxides and derivatives.

Two sterols (31a and 32a) (Figure 9) were isolated from the fruiting body of *Panellus serotinus* also known as Mukitake mushroom (family Mycenaceae). Compound (31a) was also isolated from other edible mushroom *Lepista nuda* (also known as blewit, syn. *Clitocybe nuda*) [66]. Ergostane-type endoperoxy glycosides (33a and 34a) were isolated [67] from the ethanol extract of an attractive mushroom *Lactarius volemus* (family Russulaceae) which inhibits the growth of several tumor cell lines in vitro [68].

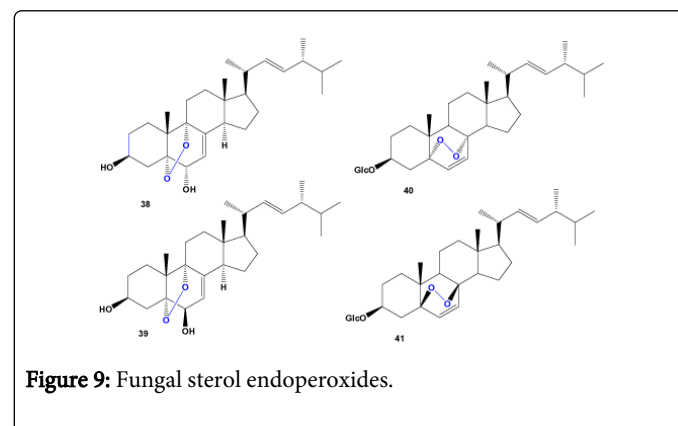


Figure 9: Fungal sterol endoperoxides.

## Peroxides from algal species

**Hydroperoxides:** The (1-ethoxyethyl) hydroperoxide (41) was recovered from several algal species: two brown algae *Cladosiphon okamuranus*, *Analphus japonicus*, and red alga *Gracilariopsis chorda* (Gracilariaceae, Rhodophyta) [69]. Cumene hydroperoxide (42) was detected in green alga *Chlorella pyrenoidosa* [70].

The diterpenoid neoconcinndiol hydroperoxide (43) was found as a constituent of red alga *Laurencia snyderiae*. The suggestion was made that (43) arises from the brominated natural product concinndiol, also from *L. snyderiae*, by solvolytic ring contraction and oxygenation to yield the rearranged allylic hydroperoxide [71]. Laureperoxide (44), cuparene-derived sesquiterpene isolated from the red alga *Laurencia okamurai* (Nanji Island, China) [72]. Halogenated nonterpenoid C15-acetogenin, laurendecumenyne A (45) has been reported from the marine red alga *Laurencia decumbens*. Cytotoxicity against adenocarcinomic human alveolar basal epithelial cells A549 cells was showed [73].

Bioassay-guided fractionation of extracts from a Fijian red alga in the genus *Callophycus* sp. resulted in the isolation of five new compounds of the diterpene-benzoate class. Isolated bromophycoic acids A, B, C (46), D and E display a range of activities against human tumor cell lines, malarial parasites, and bacterial pathogens including low micromolar suppression of MRSA and VREF [74] (Figure 10).

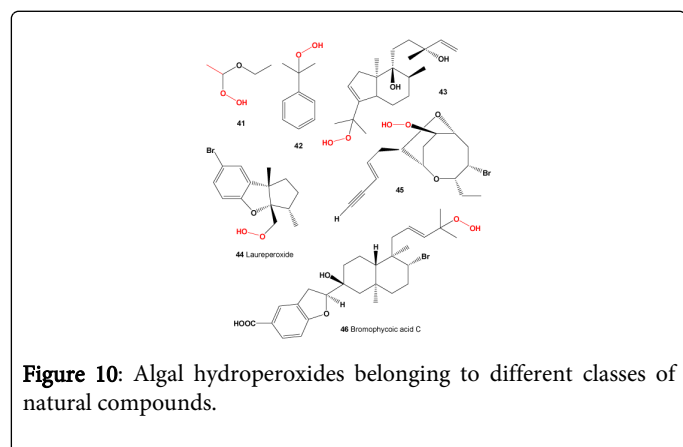


Figure 10: Algal hydroperoxides belonging to different classes of natural compounds.

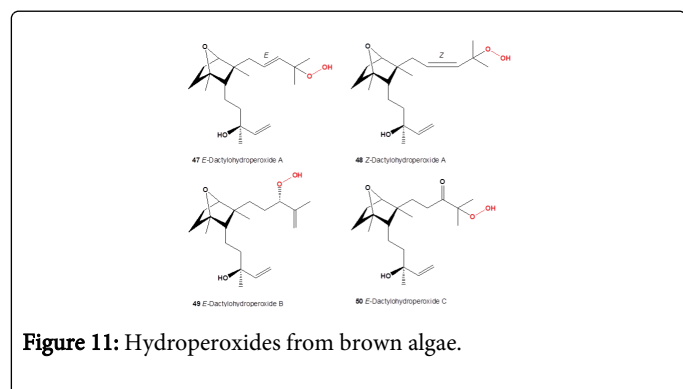


Figure 11: Hydroperoxides from brown algae.

Hydroperoxides have been found in the Russian population of brown alga, *Dictyota dichotoma* (Troitsa Bay, Sea of Japan, Russia), for example, dicyclohydroperoxides A (E-47), A (Z-48), B (49) and C (50) [75] (Figure 11). Some isolated compounds showed moderate cytotoxicity against human cancer cell lines. Also, unstable

hydroperoxide, dactylohydroperoxide C (50) produced by red algae *Laurencia* sp. from Tenerife (Canary Island) [76].

Two bioactive brominated diterpenes, cytotoxic bromoditerpene (51) and antibacterial bromoditerpene 2S-hydroperoxy-12R-hydroxy-isobromosphaerol (52) have been isolated from the marine red alga *Sphaerococcus coronopifolius* (also known as *Hematocelis fissurata*). The structure of the previously reported 12S-hydroxy-bromosphaerodiol (53) and 2S,12S-dihydroxyisobromo-sphaerol (54) were revised [77,78].

Several hydroperoxides as derivatives of the arachidonic metabolites with the lipoxygenase in marine algae have been detected. It was reported that eicosanoids, 12(S)- and 15(S)-hydroperoxyeicosatetraenoic acid were the intermediate product of major aldehyde flavor formation [3(Z)- and 2(E)-nonenal and n-hexanal] in an edible brown alga, *Laminaria angustata* via lipoxygenase (LOX) and hydroperoxide lyase pathway. Three eicosanoids have been found after enzymic formation and identified as 8-, 9-, and 11-hydroperoxy-eicosatetraenoic acids (55-57, respectively, Figure 12) by HPLC. These represented the mechanism of positional selectivity of LOX in this marine alga [79].

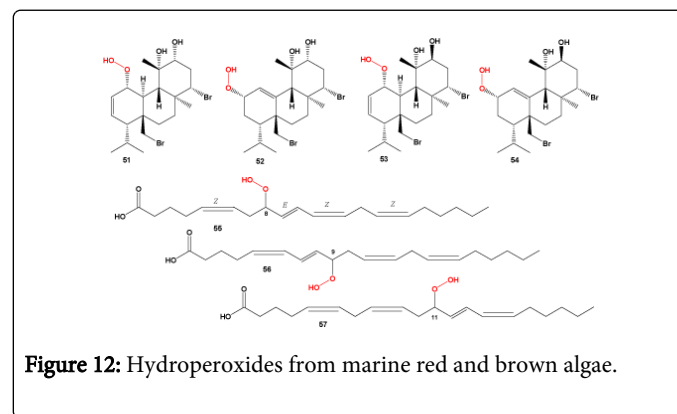


Figure 12: Hydroperoxides from marine red and brown algae.

When long-chain saturated and unsaturated fatty acids were incubated with crude enzyme of marine green alga *Ulva pertusa* (sea lettuce), the corresponding (R)-2-hydroperoxy acids: 2-hydroperoxy-hexadecanoic, 2-hydroperoxy-9(Z)-octadecenoic, 2-hydroperoxy-9(Z), 12(Z)-octadecadienoic acids, respectively) were found to have high enantiomeric excess (>99%). In a similar administration except, the (R)-2-hydroperoxy-acid was obtained from the incubation of palmitic acid with crude enzymes of a variety of marine algae. Thus, authors found that not only green algae but also brown and red algae are capable of enantio-selective 2-hydroperoxylation of palmitic acid [80].

Two bioactive compounds, dicyclohydroperoxide (58) and hydroperoxy-acetoxycrenulide (59), containing hydroperoxyl groups rarely found in algal terpenoids were isolated from the Russian population of brown alga *Dictyota dichotoma*. Isolated compounds showed moderate cytotoxicity against human cancer cell lines [75].

8 $\alpha$ -hydroperoxy- $\alpha$ -tocopherone (60), the primary oxidation product of  $\alpha$ -tocopherol by singlet oxygen, it was isolated from *Chlamydomonas reinhardtii* cultures during high light stress under variety of conditions (presence of inhibitors, an uncoupler, heavy water) [81].

Several biological active meroditerpenoids stypohydroperoxide (61), 2 $\beta$ ,3 $\alpha$ -epitaondiol, flabellinol, flabellinone, stypotriolaldehyde, along with known compounds from the marine brown alga

*Styopodium flabelliforme* collected in Papua New Guinea. All of the new metabolites were moderately toxic to murine neuro-2a cells (LC50 2-25  $\mu$ M), and 2 $\beta$ ,3 $\alpha$ -epitaondiol, flabellinol, and flabellinone possessed potent sodium channel blocking activity [82].

Marine cyanobacterium *Lyngbya* sp. led to the isolation of biselyngbyasides A, B, endo-peroxide biselyngbyaside C, and hydrobiselyngbyaside D (62), collected on Tokunoshima Island (Japan) [83,84]. Biselyngbyolide A exhibited strong apoptosis-inducing activity against HeLa S3 (human epithelial carcinoma cell line) and HL60 (human promyelocytic leukemia) cells [83]. Other biselyngbyasides showed growth-inhibitory activity and apoptosis-inducing activity against both HeLa S3 cells and HL60 cells. The fura-2 method revealed that biselyngbyasides increased the cytosolic  $Ca^{2+}$  concentration in HeLa S3 cells [84]. Structures showed in Figure 13.

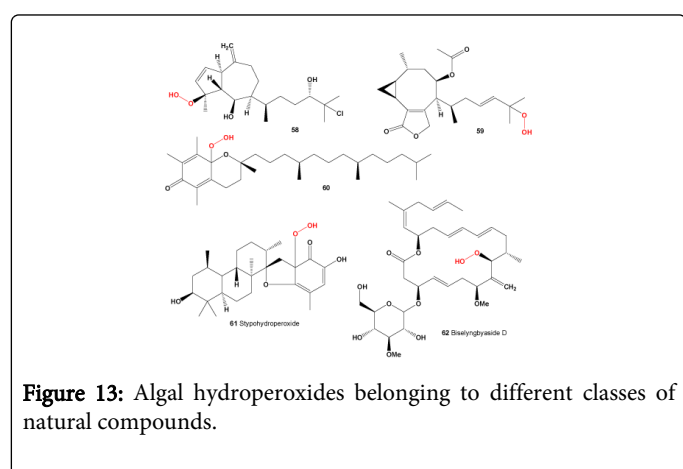


Figure 13: Algal hydroperoxides belonging to different classes of natural compounds.

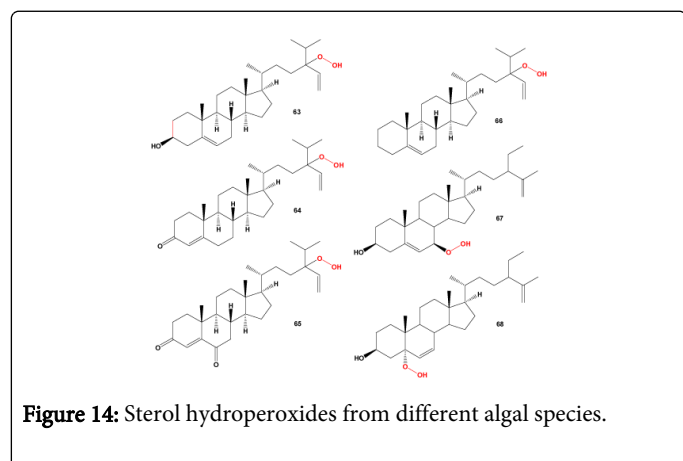


Figure 14: Sterol hydroperoxides from different algal species.

Cytotoxic steroids (63, 64 and 65) have been recovered from the brown alga *Turbinaria conoides*. The cytotoxicity in HeLa cells was expressed in terms of 50% cytotoxic concentration (CC50). These oxygenated steroids exhibited cytotoxicity against HeLa cells with CC50 values ranging from 60.9  $\mu$ g/mL to >100  $\mu$ g/mL [85]. Sterol, 24(R)-hydroxy-24-vinylcholesterol (63) has been isolated from *Sargassum oligocystum* (Heterokontophyta), which it is one of the most abundant algae distributed in the Persian Gulf [86], and in *Sargassum fusiforme* [87]. This compound (63) was also found in red alga *Ceratodictyon spongiosum* (Rhodophyta) [88].

Several unusual glycerolipids, including at 24-ethylcholest-4,24(28)-dien-3 $\beta$ -ol, 24-vinylcholest-5-en-24 $\zeta$ -hydroperoxy (66) were isolated from marine brown alga *Sargassum parvivesiculosum* [89].

Genus *Codium* contains a lot of different lipophilic metabolites [90], and sterols [91]. The two hydroperoxy clerosterols (67 and 68) were isolated from the Indo-Pacific marine green alga *Codium arabicum* [92]. The compounds displayed significant cytotoxicity toward various cancer cell lines.

Ergosterol peroxide (35) has also been identified from some algal species: thus it was isolated from marine red alga *Laurencia cartilaginea* [93], brown alga *Turbinaria conoides* [94], green freshwater alga *Chlorella vulgaris* [95], and halotolerant alga *Dunaliella salina* [96] (Figure 14).

**Endoperoxides:** A few endoperoxides (Figure 15) have been isolated from algal species. Thus, the diastereomeric abietic acid endoperoxides (69) and (70) were isolated from freshwater *Elodea canadensis* [97], and endoperoxide biselyngbyaside C (71) produced by marine *Lyngbya* sp. [84].

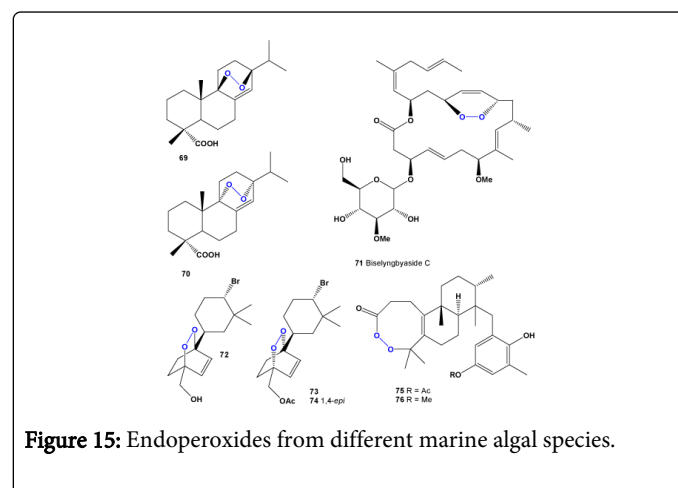


Figure 15: Endoperoxides from different marine algal species.

Several sesquiterpenes, majapolenes A (72), B, majapolone, and majapols A-D, were isolated from a Philippine collection of red alga *Laurencia majuscula*. With the exception of majapolene B, all compounds were isolated as inseparable diastereomeric mixtures. Majapolene A, a dioxabicyclo [2.2.2]-alkene, displayed modest activity in the NCI 60-cell line cytotoxicity screening. Majapolene A (72) was also found as a major component of a Philippine collection of *Laurencia caribica* [98]. Similar majapolane sesquiterpenes with antibacterial activity, as isomeric mixture acetylmajapolene A, (73) and (74), have been found in an extract of an undescribed Malaysian (Pulau Tioman) species of the *Laurencia* genus [99]. Two diterpenoid peroxy lactones, (75) and (76), were isolated from the brown seaweed *Taonia atomaria* [100].

## Novel and bioactive peroxides from plant species

**Hydroperoxides:** Bioactive metabolite (77) (Figure 16) was isolated from the aerial parts of *Salvia sahendica* (family Lamiaceae; it is known that *Salvia* genus showed antibacterial effects on *Klebsiella pneumonia*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*), together with several known compounds, comprising a sesterterpene, a sesquiterpene, a diterpene, triterpenes, steroidal compounds, and flavonoids [101].

A p-menthane hydroperoxide, (1R,4S)-1-hydroperoxy-p-menth-2-en-8-ol acetate (78), as trypanocidal agent against epimastigotes of *Trypanosoma cruzi*, was isolated from dried leaves of an aromatic evergreen tree *Laurus nobilis* (family Lauraceae) [102]. Plant metabolite (79) exhibited antitrypanosomal activity against *Trypanosoma brucei* [103].

The aerial parts of the Mediterranean weed *Carthamus glaucus* (family Compositae) afforded an unusual triglyceride (E-2-crotonyl-sn-1,3-distearoylglycerol), and a series of bisabolane fucopyranosides variously acylated on the sugar moiety, and its peroxy derivatives, such as (80) and (81). Isolated fucopyranosides are as potential an anti-inflammatory cosmetic ingredient in current short supply in its natural form. A comparative investigation of the activity of isolated metabolites involved in inflammation and cancer pathways (NF- $\kappa$ B and STAT-3) showed only marginal activity on NF- $\kappa$ B inhibition for all compounds [104].

Interesting tiglane-type diterpenes, stelleracins A, B, C, D (82) and E (83), were isolated from the roots of a perennial herbaceous plant *Stellera chamaejasme* (Thymelaeaceae), from the Qinghai-Tibet Plateau and in adjacent regions. The isolated compounds showed potent anti-HIV activity (EC<sub>90</sub> 0.00056-0.0068  $\mu$ M) and relatively low or no cytotoxicity (IC<sub>50</sub> 4.4-17.2  $\mu$ M). These compounds represent promising new leads for development into anti-AIDS clinical trial candidates [105] (Figure 16).

A dolabellane diterpene derivative with the naturally rare peroxy function was identified as Me ester of 2,18-O-diacetyl-16-O-(3-hydroxy-3-methylglutaryl)-7-hydroperoxy-dolabella-3,8(17)diene-2,16,18-triol (82) (Figure 17) was isolated from the aerial parts of the herb *Cleome droserifolia* (syn. *Roridula droserifolia*) [106].

A trans-chrysanthemoid monoterpenoid hydroperoxide (83) has been isolated from the aerial parts of *Santolina insularis*, a bush endemic to Sardinia. *S. insularis* is a medicinal plant whose essential oil showed antiviral and anti-bacterial activities and potent and selective cytotoxic activity against the human colon carcinoma cell line. The occurrence of several chemotypes makes the taxonomic identification of *S. insularis* hard to achieve [107]. The biological activity of *S. insularis* was also demonstrated against *Staphylococcus aureus*, *Escherichia coli*, *Candida albicans*, *Candida tropicalis* and *Cryptococcus neoformans* [108].

Three sesquiterpene hydroperoxides (84-86), together with known compounds, germacrone, ent-germacra-4(15),5,10(14)-trien-1 $\alpha$ -ol and teucdiol A were isolated from the aerial parts of *Aster spathulifolius* (family Compositae). The isolated compounds were tested for their cytotoxicity against five human tumor cell lines *in vitro* using a SRB method. The two hydroperoxides (84) and (85), showed moderate cytotoxicity against human cancer cells with ED<sub>50</sub> values ranging from 0.24 to 13.27  $\mu$ g/mL [109].

Hydroperoxy terpene (87) was isolated from *Juniperus przewalskii* (family Cupressaceae; a dominant tree species endemic to the northeast Qinghai-Tibetan Plateau), together with several known terpenes, including, 3 $\alpha$ -hinokiol and 3 $\alpha$ -hydroxymannol which exhibited effective anti-tumor activities to cervical carcinoma (HeLa) and human ovarian carcinoma (HO-8910) cell lines [110].

The lipophilic extract of the fresh water liverwort *Riella helicophylla* yielded several monoterpenes and diterpenes [111]. Several monoterpenes were hydroperoxides (90-93, 95 and 96) [112]. The 11-

hydroperoxy-6,9-eremophiladien-8-one (94), along with oleanolic acid,  $\beta$ -amyrin,  $\beta$ -amyrin acetate and (+)-lupeol, were isolated from the EtOH extracts of *Ligularia kanaitzensis* (family Compositae) [113].

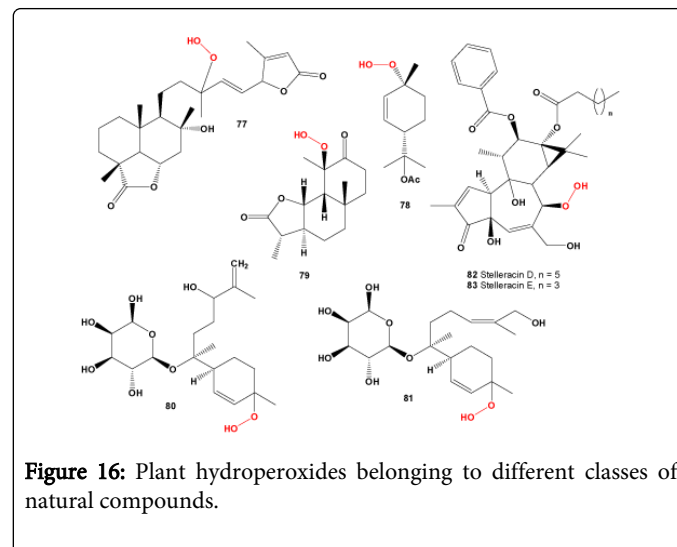


Figure 16: Plant hydroperoxides belonging to different classes of natural compounds.

Monoterpene coumarins, minutin A (97) and B (98) were purified from the citrus plant *Micromelum minutum* (family Rutaceae) leaves. Isolated compounds had some inhibitory activity against one or more lung adenocarcinoma (SBC3 and A549) and leukemia (K562 and K562/ADM) cell lines *in vitro*. Minutin B (77) had the strongest cytotoxic activity against SBC3, A549, K562, and K562/ADM cell lines, with respectively 9.6, 17.5, 8.7, and 6.7  $\mu$ M [114].

Eleven triterpenoids, schincheninins A-H and schincheninlactones A-C together with three known triterpenoids, henrischinins A-C, were isolated from the leaves and stems of *Schisandra chinensis* (also known as five flavor berry; native to forests of Northern China and the Russian Far East) by bioassay-guided fractionation. Schincheninins E and F (99 and 100, Figure 18) are highly oxygenated triterpenoids that contain a hydroperoxyl moiety, which is rare in compound from the *Schisandra* genus. Some compounds showed activities against HSV-2 and adenovirus [115].

Eleven triterpenoids, schincheninins A-H and schincheninlactones A-C together with three known triterpenoids, henrischinins A-C, were isolated from the leaves and stems of *Schisandra chinensis* (also known as five flavor berry; native to forests of Northern China and the Russian Far East) by bioassay-guided fractionation. Schincheninins E and F (99 and 100) (Figure 18) are highly oxygenated triterpenoids that contain a hydroperoxyl moiety, which is rare in compound from the *Schisandra* genus. Some compounds showed activities against HSV-2 and adenovirus [115].

The active constituents of bark and leaves of the traditionally used antimalarial plant *Liriodendron tulipifera* (known as the tulip tree, American tulip tree) by antiplasmodial-assay guided fractionation. Leaves yielded two known sesquiterpene lactones, peroxyferolide (101) and lipiferolide with antiplasmodial activity. The antiplasmodial activity of compound (101) (IC<sub>50</sub>=6.2  $\mu$ g/mL) was reported. This work supports the historical use of *Liriodendron tulipifera* as an antimalarial remedy of the United States and characterizes its antiplasmodial constituents [116].

Several sesquiterpenes, together with compounds (102) and (103) were isolated from leaves of an aromatic evergreen tree *Laurus nobilis*

(also known as Bay leaf). Most of these compounds exhibited moderate to significant cytotoxicity toward K562 leukemia cells [117].

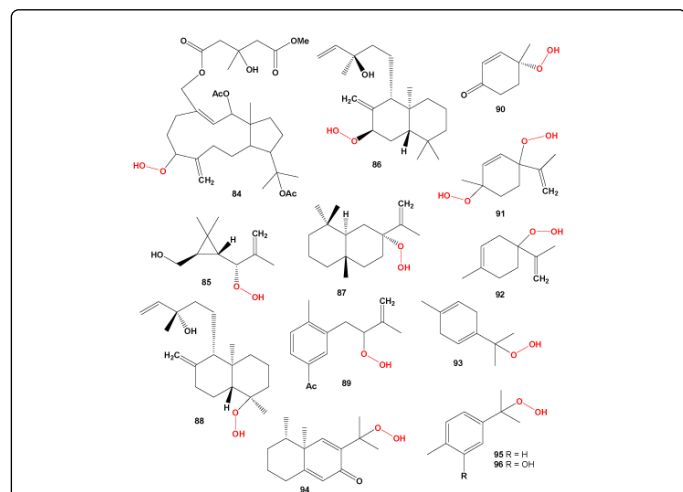


Figure 17: Plant hydroperoxides belonging to terpene, aroma and their derivatives.

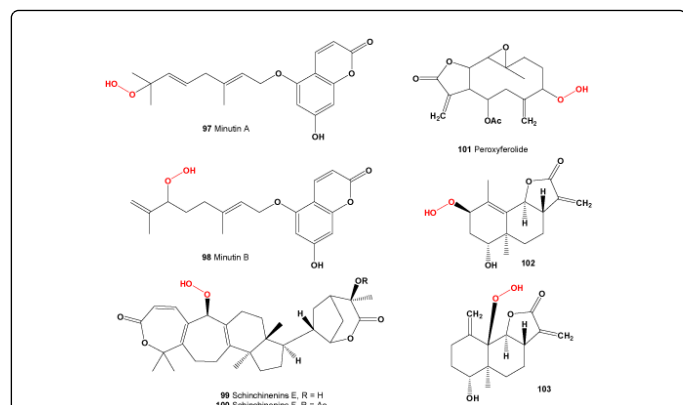


Figure 18: Plant hydroperoxides belonging to different classes of natural compounds

Lindenane sesquiterpenes, spicachlorantins A (104) and C (105) were isolated from the whole dicotyledonous plant of *Chloranthus serratus* (family Chloranthaceae). These isolates were evaluated for their inhibitory effects on lipopolysaccharide-induced nitric oxide production in RAW264.7 cells. Spicachlorantin A and two known compounds, shizukaols B and D, showed significant anti-inflammatory activities, with  $IC_{50}$  values of 0.22, 0.15, and 7.22  $\mu$ M, respectively [118].

Several bioactive compounds, including (1R,3S,7E,11S,12R)-3-hydroperoxy-dolabella -4(16),7-dien-18-ol (106), was found in leaves extract of the oriental medicinal plant *Aglaia odorata* (family Meliaceae, known as Chinese perfume plant). All isolated compounds possessed potent nitric oxide inhibitory activity with  $IC_{50}$  values ranging from 2.1 to 14.2  $\mu$ M, these being better than that of the positive control, indomethacin ( $IC_{50}$ =14.5  $\mu$ M) [119].

A new hydroperoxyquinolone alkaloid, glycopentaphyllone (107) was isolated from the fruits of lowering plant *Glycosmis pentaphylla*

(family Rutaceae), known commonly as orangeberry and gin berry. Compound showed antibacterial activity against *Escherichia coli* TISTR 780, *Salmonella typhimurium* TISTR 292, *Staphylococcus aureus* TISTR 1466, and Methicillin-resistant *S. aureus* SK1 [120].

Antibacterial acylphloroglucinols, named olympicins A, B, C, D (108), and (E) were isolated and characterized from the aerial parts of the flowering plant *Hypericum olympicum* cf. *uniflorum* (family Hypericaceae). All compounds exhibited min inhibitory concentrations (MICs) of 0.5 to 128 mg/L against *Staphylococcus aureus* strains [121] (Figure 19).

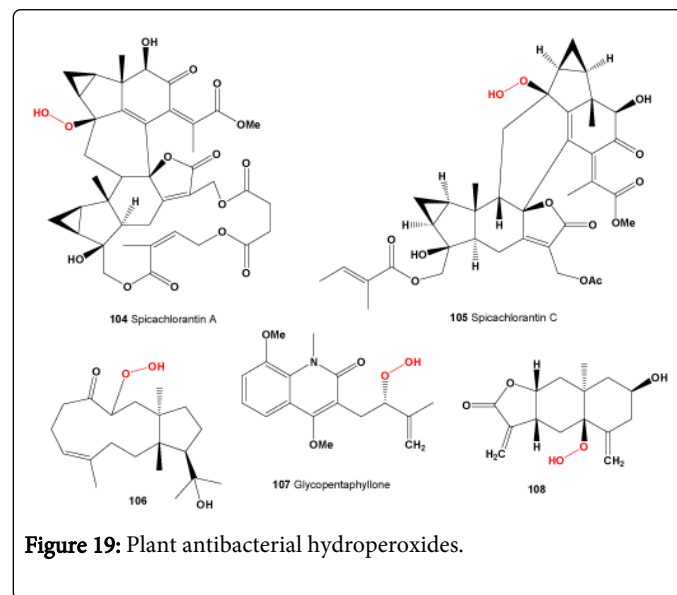


Figure 19: Plant antibacterial hydroperoxides.

Argutins A, B, C, D, E, F (109), G (110), and H (111) highly oxygenated clerodane diterpenes, were isolated from the plant *Casearia arguta* (family Salicaceae) collected in Guatemala. Each of the argutins showed varying levels of synergy with tumor necrosis factor- $\alpha$ -related apoptosis-inducing ligand sensitizers [122]. Antibacterial acylphloroglucinols, named olympicins A, B, C, D (112), and (E) were isolated and characterized from the aerial parts of the plant *Hypericum olympicum* cf. *uniflorum*. All compounds exhibited min inhibitory concentrations (MICs) of 0.5 to 128 mg/L against *Staphylococcus aureus* strains [121].

The genus *Aristolochia* (known as birthwort, pipevine or Dutchman's pipe) is an important source of physiologically active compounds that belong to different chemical classes, and it is the subject of research in numerous pharmacological and chemical studies [123]. Thus, clerodane diterpenoids isolated from *Aristolochia* species, compounds (113 and 114) (Figure 20) were isolated from *A. esperanzae* [124], and compound (115) was identified from *A. chamissonis* [125].

Six new bisabolane-type sesquiterpenes, peroxylippidulcines A-C (116-118) (Figure 21), peroxyepilippidulcine B, and epilippidulcines B and C, have been isolated from the aerial parts of *Lippia dulcis* (perennial herb; native to southern Mexico), along with two known bisabolane-type sesquiterpenes, seven known flavonoids, and a known triterpenoid [126].

The aerial part of *Aster scaber* (known as edible Korean chamchwi; family Compositae) yielded two new monoterpene peroxide glycosides, (121) and (122), and other known compounds [127].



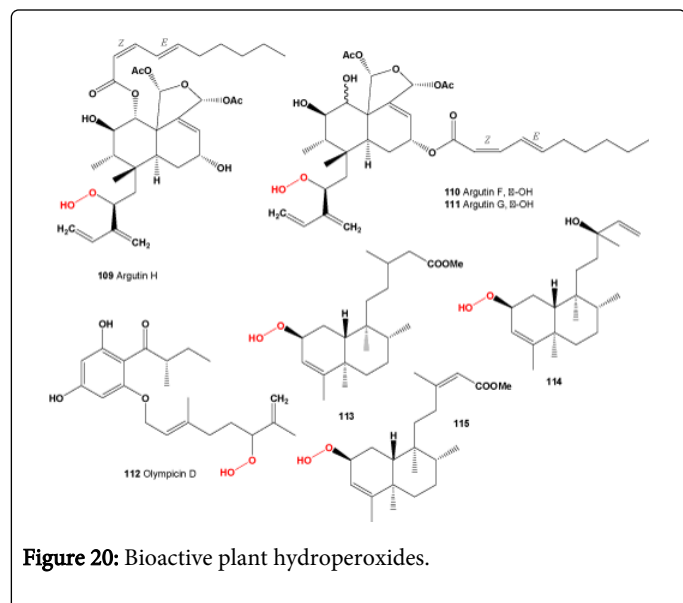


Figure 20: Bioactive plant hydroperoxides.

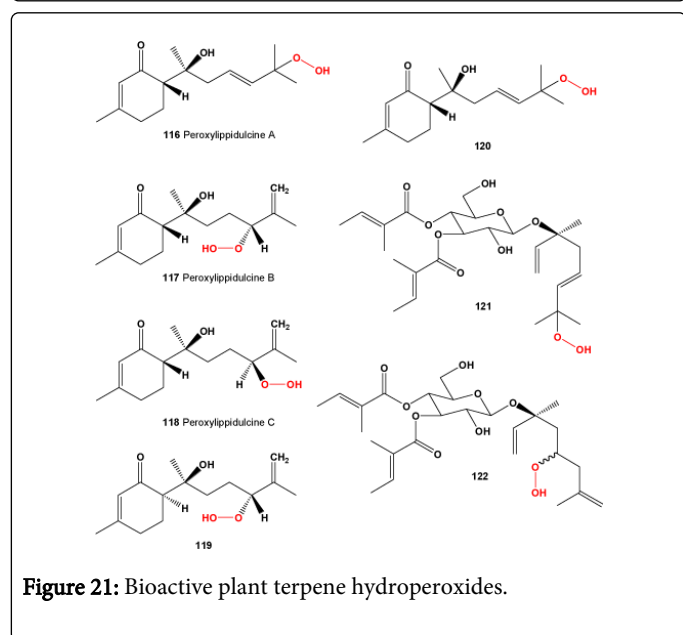


Figure 21: Bioactive plant terpene hydroperoxides.

Several dimeric sesquiterpene lactones (japonicones E-L), including a novel sesquiterpene dimer bearing a rare hydroperoxide group (japonicone E, 123) (Figure 22), were isolated from the aerial part of flowering plant *Inula japonica* (family Asteraceae). Compound (123) displayed strong inhibitory activity against LPS-induced NO production in RAW264.7 macrophages [77]. From the same species afforded additional related dimeric sesquiterpene, japonicone T (124) [78].

Cembrane-type diterpenoids with a trans-fused  $\alpha$ -methylene- $\gamma$ -lactone, including two cembrane hydroperoxides 4-methylene-5 $\beta$ -hydroperoxy-ovatodiolide (125) and 4 $\alpha$ -hydroperoxy-5-enovatodiolide (126) were isolated from a methanol extract of Indian Catmint *Anisomeles indica* (syn: *Nepeta indica*, *Anisomeles ovata*). Compound (126) exhibited cytotoxicity against a small panel of

human cancer cell lines, and showed inhibitory effects on antiplatelet aggregation induced by thrombin [128].

Seven-membered vibsane-type diterpene hydroperoxides named 5-epi-vibsanin K (127), 18-O-methyl-5-epi-vibsanin K (128) as well as their corresponding C-5 epimers (129) and (130) have been isolated from the leaves of *Viburnum awabuki* (syn: *Viburnum odoratissimum* var. *arboricola*; family Adoxaceae, Caprifoliaceae). The occurrence of these seven-membered vibsane-type diterpenes with a cis relationship on the C-5 and C-10 positions in nature have been predicted by conformational analysis of vibsanan B, an eleven-membered vibsane-type diterpene. Some compounds exhibited moderate cytotoxic activities on KB cells [129,130].

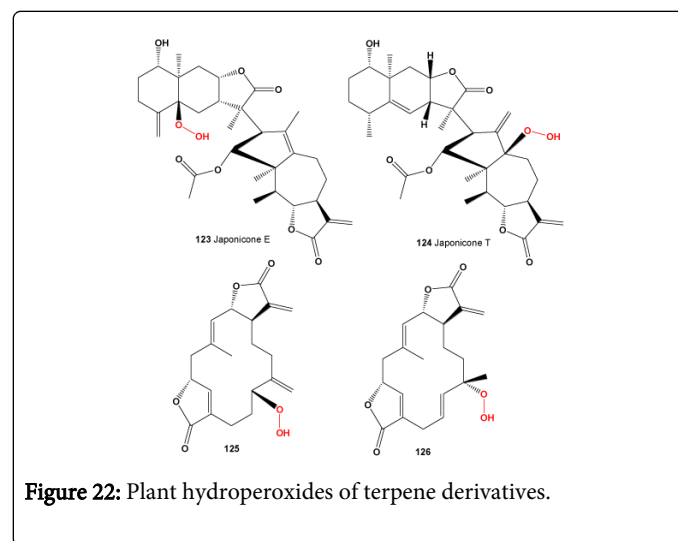


Figure 22: Plant hydroperoxides of terpene derivatives.

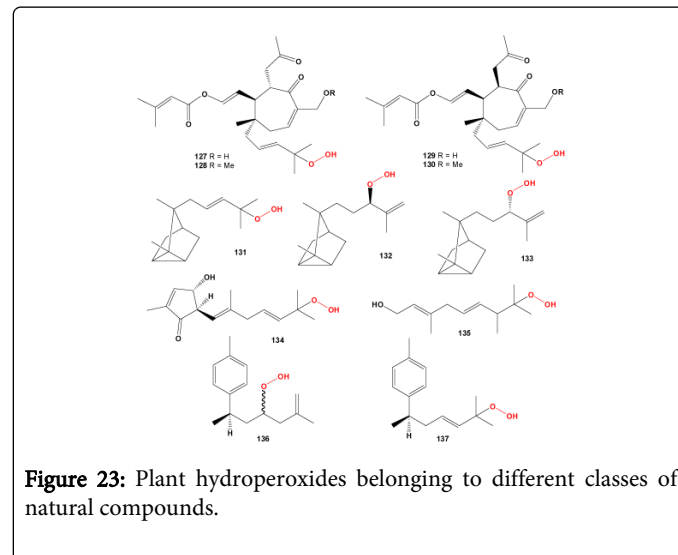


Figure 23: Plant hydroperoxides belonging to different classes of natural compounds.

Several santalane and isocampherenane sesquiterpenes, including three isomeric sesquiterpene hydroperoxides (131-133) have been isolated from *Illicium tsangii* (family Illiciaceae; a poisonous shrub from southern China used in traditional medicine for treating pain). The santalanes may be derived from (-)- $\alpha$ -santalene by oxidation reactions [131,132].

Anti-HIV and cytotoxic activities of litseaverticillol A isolated from the twigs and leaves of shrub *Litsea verticillata* (Lun Ye Mu Jiang Zi in

Chinese; the roots and leaves are used medicinally for treating rheumatism and relieving menstrual cramping and soreness) are known [133]. Synthesis of litseaverticillols B, E (134), I, and J as well as the structural reassignment of litseaverticillol E (134) have been achieved by means of a biomimetic sequence of transformations during which a [4 + 2]-initiated reaction cascade and an ene reaction, both involving singlet oxygen [(1)O(2)], formed key steps. The reassignment of the structure of litseaverticillol E to include an allylic hydroperoxide provides strong support for biogenetic hypothesis was reported [134].

Compound (135) (Figure 23) was isolated from a multi-branched shrub *Heterothalamus alienus* (family Asteraceae; it used in Brazilian and Argentinean folk medicine) [135]. *Senecio* species are used for therapeutic purposes, including the treatment of fungal skin infections [136], antiseptic [137], and pneumonia [138]. The fresh aerial parts of *Senecio selloi* (family Asteraceae) contains two hydroperoxides (136 and 137), which were identified indirectly by isolation, identification and posterior photooxidation of  $\alpha$ -curcumene, their precursor in the plant [139].

**Four- and five-membered endoperoxides:** Four-membered endoperoxides are rare in nature. A few members of this group have been discovered. Thus, a solubilised enzyme fraction of mycelium lyophilisate of *Pleurotus sapidus* convert of  $\beta$ -myrcene to furanoterpenoids through 1,4-endoperoxides, and compound (138) was isolated as intermediate [140].

Few four-membered endoperoxides (Figure 24) have only been found in nature. The unique neolignan mansoxtane (139), was obtained from the methanolic extract of the heartwood from *Mansonia gagei* (plant used in folk medicine in Thailand) [141], and sesquiterpenoid (140) was obtained from marine origin [142].

Two rare four-membered endoperoxides, named pheophytins, bidenphytins A (141) and B (141), with peroxide functionalities on ring E, were isolated from a flowering plant *Biden pilosa* var. *radiata*, a popular Taiwanese folk medicine. Possible biosynthetic pathway for 142 and 141 has been proposed [143].

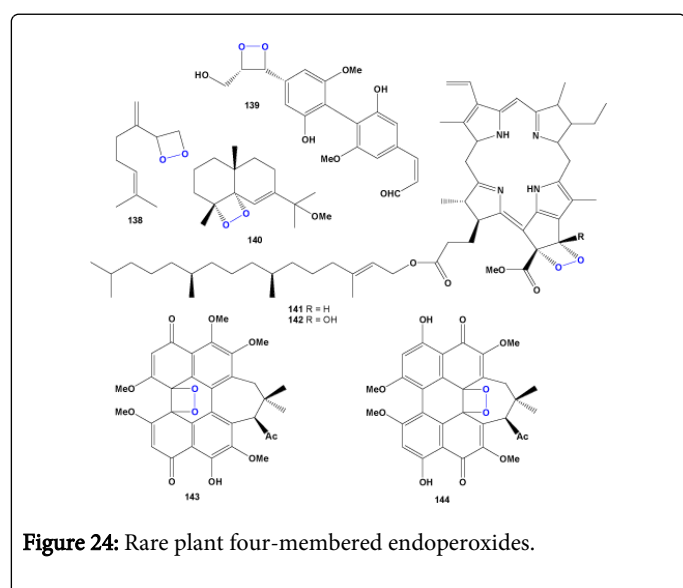


Figure 24: Rare plant four-membered endoperoxides.

Two four-membered cytotoxic peroxyhypocrellins (143 and 144) were obtained by the photooxidation of hypocrellin A [31]. Hypocrellin A has long been known as an excellent photosensitizer

and has gained much attention in recent years because of its light-induced antitumor, antifungal and antiviral activities. It appears to exert photodynamic anticancer and antiviral activities through its abilities to generate reactive oxygen species and to inhibit protein kinase C activity, and it has also been shown to exert antimicrobial and antileishmanial activities *in vitro* [144].

A cyclic monoterpene peroxide (145) (Figure 25) with the irregular santoliny framework was found from aerial parts of *Artemisia fragrans* [145]. A deciduous shrub *Artemisia maritima* is a species of wormwood known as sea wormwood and old woman produced the unusual five-membered ring peroxy hemiketal (146) [146]. Cytotoxic sesquiterpenes, (147-149) with moderate cytotoxicity were obtained from extracts of the aerial parts of *Artemisia abrotanum* [147]. The aerial parts of *Adenosma caeruleum* (family Plantaginaceae) of Vietnamese folk medicine contains cyclic peroxide (150) [148], and known metabolite cetylsaturejol (151) was detected in a small shrub *Satureja gilliesii* (family Lamiaceae) [149].

*Nardostachys chinensis* is an herbaceous member of the Valerian family, and the roots of this plant contains nardosinone (152) and kanshone B (153), and to possess weak antihepatotoxic activity [150]. Compound (152) was also a constituent of *Nardostachys chinensis*, *N. juturnansi*, and Chinese spikenard oil [150,151].

*Curcuma wenyujin* (family Zingiberaceae) is a traditional medicinal herb in China (commonly known as Wen yujin), and compound wenjine (154) was found in extract of this plant [152,153].

A dimeric guaiane peroxide named vielanin C (155) was obtained from the leaves of *Xylopiamelan* (family Annonaceae), which it used in traditional medicine in Vietnam (Kamperdick et al., 2001). Eudesmane peroxide (156) to be present in extract from the roots of *Vladimiria souliei* (**pharmaceutical name: Radix Aucklandiae seu Vladimiriae**) [154].

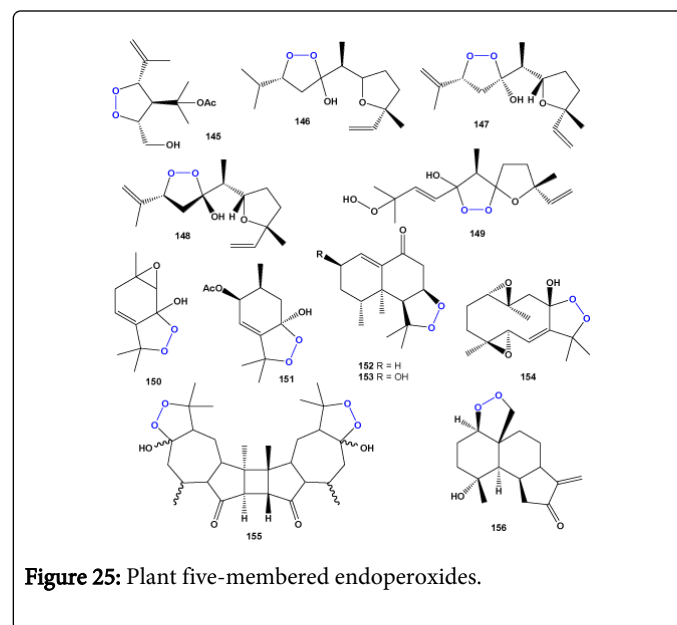


Figure 25: Plant five-membered endoperoxides.

Dimeric distansolides A (157) and B (158) were obtained from the flower heads of *Achillea distans* [155]. *Achillea* is a genus of over 100 flowering plants, and belonging to the family Asteraceae. A group of guaiane 6,10-endoperoxides (159-176) have been isolated from this genus: *A. asplenifolia*, *A. clavennae*, *A. collina*, *A. distans*, *A. ligustica*,

*A. millefolium*, *A. depressa*, *A. crithmifolia*, and *A. pseudoaleppica* [155-157]. Many of these compounds showed cytotoxic, antibacterial and antifungal activities [157,158].

Sesquiterpene peroxide (177) has been found from the aerial parts of *Croton arboreus* [159]. *Artemisia filifolia* (known by common names including sand sagebrush, sand sage and sandhill sage) was the source for an interesting longibornan endoperoxide (178) [160]. Tehranolide (179) was a constituent of extract with the antimalarial activity of *Artemisia diffusa* [161] (Figure 26).

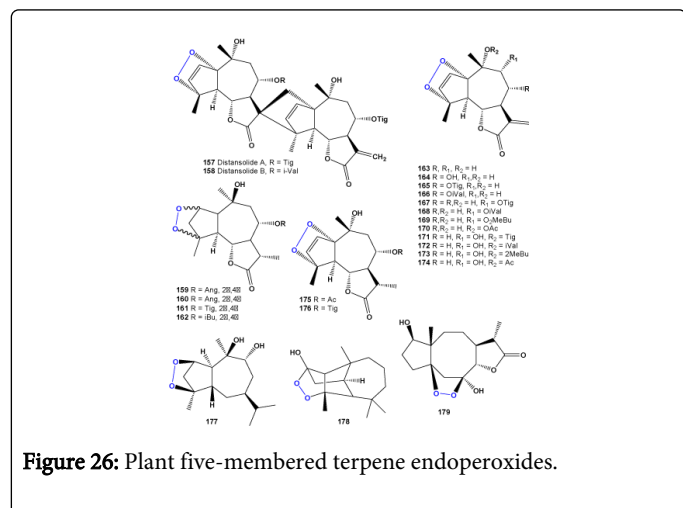


Figure 26: Plant five-membered terpene endoperoxides.

The seeds of the tree *Mammea americana* (family Calophyllaceae; commonly known as mamee, mamee apple, mamey, and mamey apple) contain two peroxy coumarin derivatives were isolated from the extract. The hydroperoxides (179) and (180) were optically active. The authors do not know whether the products result from metabolism within the seeds or are formed during isolation. Seeds of *M. americana* showed insecticidal activity (Structures showed in Figure 27).

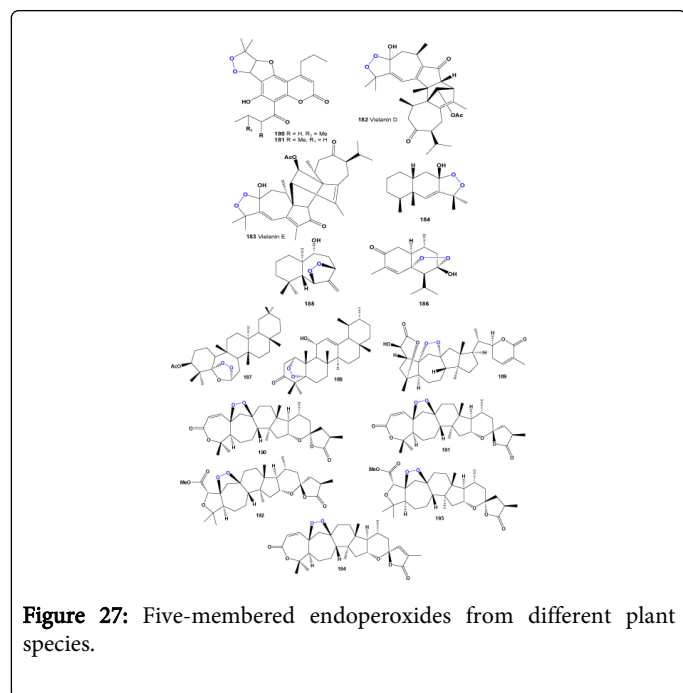


Figure 27: Five-membered endoperoxides from different plant species.

From the leaves of *Xylopia vielana* (Annonaceae, the custard apple family) two dimeric guaianes named vielanins D (182) and E (183) were isolated. These compounds consist of bridged ring systems formally representing the Diels-Alder products from the hypothetical guaiane-type monomers [162].

Three species of the genus *Ligularia* (also known as leopard plant) perennial plants in the family Asteraceae: *L. kanaitzensis*, *L. subspicata*, and *L. veitchiana*, provided the eremophilane peroxide (184) [55,113,163].

A poisonous shrub *Illicium tsangii* (family Illiciaceae) from southern China used in traditional medicine for treating pain. Extract of this shrub contain allohimachalane peroxide (185) [131,132]. The aerial parts of the flowering plant *Ageratina adenophora* (syn. *Eupatorium adenophorum*) contain the sesquiterpene (186) [164]. Aceranol acetate (187), a 5,6-cleaved glutinane derivative, has been found in *Acer mandshuricum* (also known as Manchurian Maple) [165].

An ursane triterpenoid, 1 $\alpha$ ,5 $\alpha$ -dioxo-11 $\alpha$ -hydroxyurs-12-en-3-one (188) was isolated from the rhizome of *Vladimiria muliensis*. Isolated peroxide exhibited modest antimicrobial activity against *Escherichia coli*, *Candida albicans*, *Pseudomonas aeruginosa*, *Enterococcus faecalis*, *Bacillus cereus*, and *Staphylococcus aureus* [166]. Anticancer triterpenoid schinalactone A (189), an endoperoxide with an unusual contracted ring A, has been isolated from the roots and stems of a deciduous climber *Schisandra sphenanthera*, it showed significant cytotoxicity against PANC-1 cell lines with a IC50 value of 5.9  $\mu$ M [167]. Three triterpene peroxides, pseudolarolides Q (190), R (191), pseudolarolides T1 (192), T2 (193), and Q2 (194) were discovered from the leaves of *Pseudolarix kaempferi* (family Pinaceae) [168,169].

**Six-membered endoperoxides:** Ascaridole (195) and endoperoxy derivatives (196-198) (Figure 28) thereof were identified from essential oil of the fresh *Chenopodium ambrosioides* [112,170,171]. Ascaridole was first isolated in 1895 by a German pharmacist living in Brazil and it has been attributed with most of the vermifuge (worm-expelling) actions of the plant. In the early 1900's it was one of the major antihelmintics used to treat ascarids and hookworms in humans, cats, dogs, horses, and pigs [172-175]. Ascaridole (also known as ascarisin; 1,4-epidioxy-p-menth-2-ene) is a bicyclic monoterpene that has an unusual bridging peroxide functional group. Ascaridole has been documented with sedative and pain-relieving properties as well as antifungal effects [176]. Ascaridole was found to be a potent inhibitor in vitro development of *Plasmodium falciparum* [177], *Trypanosoma cruzi* [178], and *Leishmania amazonensis* [179]. Ascaridole also showed activity against different tumor cell lines in vitro (CCRF-CEM, HL60, MDA-MB-231). The findings are the first hint that ascaridole may be an interesting novel candidate drug for cancer treatment [180]. A few review articles devoted to pharmaceutical application of ascaridole have been published [4,181,182].

The aquatic liverwort *Riella helicophylla* yielded two six-membered monoterpenes (199 and 200) [112]. The phytochemical investigation of the roots *Bombax anceps* (locally known as *Ngiu paa*; family Malvaceae) led to the isolation of dihydrobenzodioxine derivative, bombaxoin (201) [184]. Echinobithiophene A (202), a peroxide bithiophene with significant antimicrobial activity, was isolated from flowering plant *Echinops ritro* (family Asteraceae) [23]. Enzymatic extract from the leaves of *Piper crassinervium* (Piperaceae) afforded prenylated hydroquinone (203) [184].

An antitumor substance, the bisabolane sesquiterpenoid endoperoxide, 3,6-epidioxy-1,10-bisaboladiene (204), was isolated from the edible wild-plant, *Cacalia delphiniifolia* (Sanshoku). It showed cytotoxicity toward human chronic myelogenous leukemia K562 and human prostate carcinoma LNCaP cell lines with IC<sub>50</sub> values of 9.1 μM and 23.4 μM, respectively. DNA fragmentation and condensation of chromatin, the hallmarks of apoptosis, appeared in K562 cells after an 18-h treatment with compound (204). The results indicate the importance of the endoperoxide structure within endoperoxide (204) to its anti-tumor activity *in vitro* [185]. This metabolite (204) has also been isolated from other plants including *Chamaemelum fuscatum* [186,187], *Encelia canescens* [188], *Eupatorium refescens*, *Heterothalamus alienus*, *Senecio selloii* [189,190], *H. psiadioides* [191], *Ligularia speciosa* [192], *Relhania genistifolia* [193] and *Smallanthus macvaughii* [194].

The buds of *Lonicera japonica* contained a cyclic peroxide named shuangkangsu (205) with significant antiviral activities [195]. A cembrane endoperoxide (206) was isolated from the flowers of Greek cultivated tobacco, *Nicotiana tabacum* [196].

Three compounds with a unique skeleton, i.e., apigenosylides A-C (207-209, respectively) (Figure 29) recovered from the EtOH extract of the leaves of *Machilus japonica* var. kusanoi. They possess an unprecedented skeleton comprising the adduct of a butenolide moiety and apigenin glycoside linked via a 1,2-dioxane moiety. Some of these compounds possess moderate inhibitory activity against α-glucosidase [197].

Steenkrotin B (210) has been isolated from an ethanol extract of the leaves of *Croton steenkampianus* (family Euphorbiaceae). Isolated compound showed cytotoxicity against african green monkey Vero cells by MTT assay, and antiplasmodial activity against chloroquine-susceptible *Plasmodium falciparum* D6, Dd2, w2, D10 [198]. This plant commonly known as Marsh Fever-berry and Tonga Croton, is a shrub or tree endemic to restricted areas of central and eastern parts of Africa. Various medicinal uses of the genus *Croton* are reported in countries all over the world, and many species are used to treat bleeding, bleeding gums, chest complaints, coughs, fever, indigestion, malaria, and rheumatism [199].

Unusual agent with anti-inflammatory activity named kramecyne (211), was isolated from a MeOH extract of *Krameria cystisoides* (Krameriaceae). This compound exhibited good anti-inflammatory activity in the 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced mouse ear edema (51.8% inhibition) and carrageenan-induced rat paw edema models at doses of 50 mg/kg. It also significantly reduced edema to 63.1% after 1.0 h and the effect was unchanged for 5h. Kramecyne (211) did not present acute toxicity, even at doses of 5 g/kg [200].

Several diterpenoids, abiesadines A-Y were isolated from the aerial parts of conifer *Abies georgei* (family Pinaceae), including three endoperoxides (212-214) (Figure 30). Some compounds showed the strongest effect against LPS-induced NO production in RAW264, anti-inflammatory activity, anticancer activity against LOVO cells [201]. Diterpenoid, caniojane (215), has found in the roots of flowering plant *Jatropha curcas* (family Euphorbiaceae) [202].

Taibaihenryiin T (216) was isolated from *Phlomis umbrosa* (sogdan is traditionally used as herbal medicines for healthy pregnancy and easy delivery in China, Japan, and Korea). The infrequency of the C-O-O-C group is manifested in this molecular configuration, and hydrogen bonding assembles the molecules into a three-dimensional

networking structure in the crystal [203]. An unusual peroxy-multiflorane triterpene ester, (217), was isolated from the processed seeds of a flowering plant in the family Cucurbitaceae, *Trichosanthes kirilowii*. Compound (217) showed *in vitro* cytotoxicity against human tumor cell lines (Hela, HL-60, and MCF-7) and anti-inflammatory activity (LPS-induced B lymphocyte cells) [204].

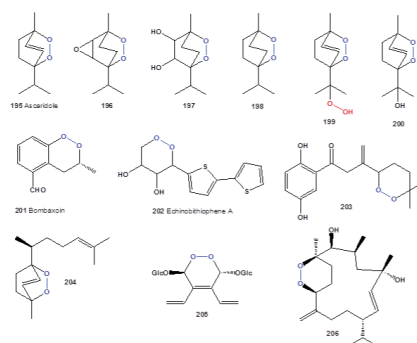


Figure 28: Six-membered endoperoxides from different plant species.

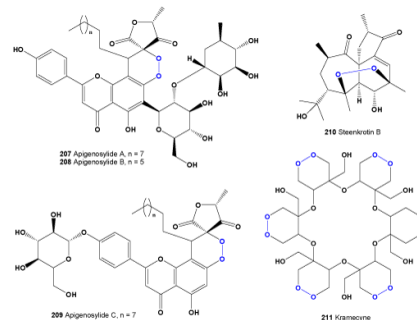


Figure 29: Six-membered endoperoxides from different plant species.

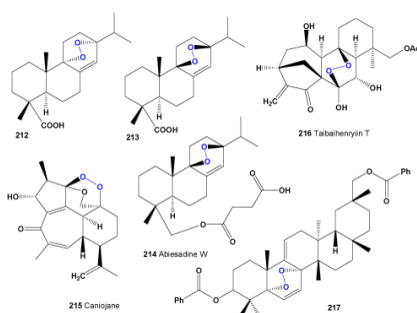


Figure 30: Plant six-membered terpene endoperoxides.

**Seven-membered endoperoxides:** Two prenylated benzophenone peroxide derivatives, peroxy-sampsones A (218) and B (219) (Figure 31), together with a known compound, plukenetione C, were isolated from the roots of the Chinese medicinal plant *Hypericum sampsonii*. These compounds are the unusual peroxides of polyprenylated

benzophenone derivatives, containing the unique caged moiety of 4,5-dioxatetracyclo (9.3.1.19,13.01,7) hexadecane-12,14,15-trione. Unusual peroxide (218) showed comparable activity with norfloxacin against a NorA overexpressing multidrug-resistant strain of *S. aureus* SA-1199B [205]. Sesquiterpene, 4-hydroxy-1,15-peroxyeudesmane (220) was found in hexane extract from *Xylopia emarginata* fruits [206].

10,12-peroxycalamenene (221) was obtained from the dried tubers of *Cyperus rotundus* [207], and this compound has good *in vitro* antimalarial activity. Other antimalarial compound (222), is a constituent of cardamom, herb *Amomum krervanh* (family Zingiberaceae) [208]. *Artemisia annua* (known as sweet wormwood), afforded a rare seven-membered endoperoxide lactone arteannuin H (223) [209]. A labdane diterpene, coronarin B (224), and b-hydroxycoronarin B (225) were isolated from *Hedychium coronarium* (known also as white ginger lily) [210], and coronarin B has also been found in *Alpinia chinensis* [211].

Unusual metabolite (227) was discovered from *Xylopia emarginata* (family Annonaceae) and *Ecdysanthera rosea* (syn: *Urceola rosea*), respectively [206,212]. The eremophilane peroxide (228) was found in three species of the *Ligularia* genus (family Asteraceae): *L. subspicata*, *L. kanaitzensis*, and *L. veitchiana*, respectively [55,111,163].

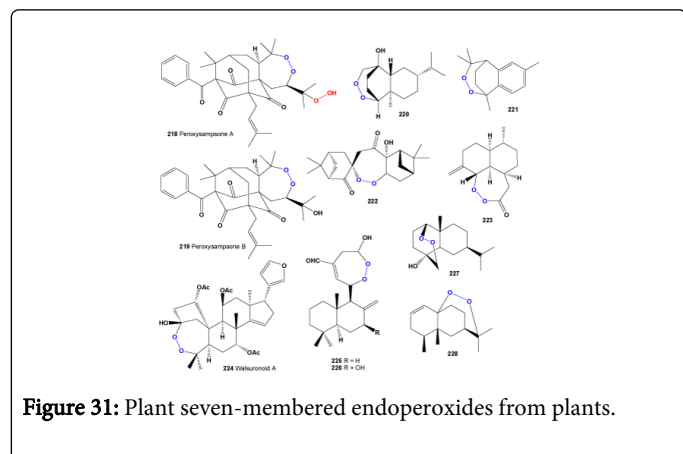


Figure 31: Plant seven-membered endoperoxides from plants.

**Rare acyclic peroxides:** Acyclic peroxides differ the other two types of peroxides: a) hydroperoxides and b) endoperoxides by the presence of the peroxy (-O-O-) linkage between the fragments of the same and/or other molecular structure. This bridge on the chemical structures marked in green. Several compounds were found in nature. Thus, acyclic peroxide oxanthromycin (26) (Figure 32) was isolated from an *Actinomadura* sp. fermentation broth [27,28], and two adxanthromycins A (27) and B (28) were found in the fermentation broth of *Streptomyces* sp. NA-148.

Dimethyl peroxide (229) was detected among the volatile components of *Basella rubra* (family Basellaceae) [213], and three other acyclic bis(1-hydroxyalkyl)peroxides (230–232) in the essential oil of Japanese citrus fruit, *Citrus iyo* (iyokan, also known as anadomikan) [214,215]. Extract of the Brazilian medicinal plant *Kielmeyera coriacea* (family Clusiaceae) afforded a  $\delta$ -tocotrienol peroxy-dimer (234) [216], other peroxide dimer named bungein A (235) was found in the aerial parts of the medicinal plant *Clerodendrum bungei* (also known as Mexicali Rose, Mexican Hydrangea and/or Cashmere Bouque) [217]. Leucoperoxyterpene (236) with good antibacterial activity has been isolated from extract of

aerial parts of the medicinal plant *Leucoscepttrum canum* (family Laminaceae) [218].

Dimeric dihalenaquinolides A (237) and B (238), from marine origin, have a peroxide linkage between two meroterpenoid units [219]. Lactucin-8-O-p-methoxyphenyl acetate (239), a cytotoxic sesquiterpene lactone, has been detected in *Mulgedium tataricum* (family Compositae) [220].

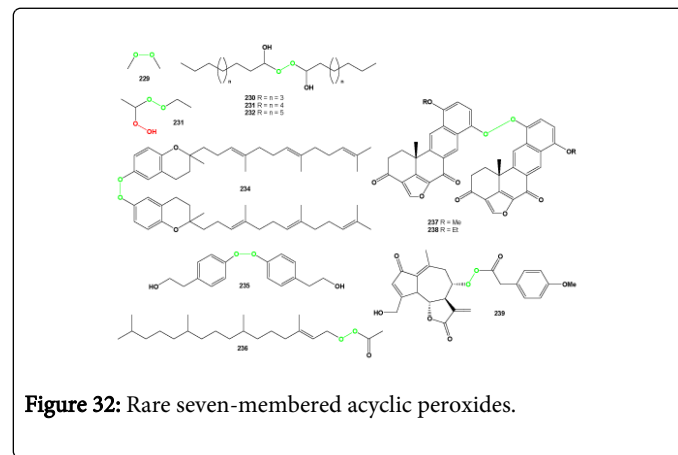


Figure 32: Rare seven-membered acyclic peroxides.

## Conclusions

During the last 10-15 years, there has been an unprecedented growth in the chemistry of natural as well as synthetic peroxides. Currently, the rapid progress in chemistry of organic peroxides is to a large degree determined by their high biological activity. In medicinal chemistry of peroxides, particular emphasis is given to the design of compounds having activity against causative agents of malaria and human helminth infections. In medicinal chemistry of peroxides, for example, ascaridole and artemisinin a natural peroxides exhibiting high antimalarial activity, is the most important drug in use for approximately 30 years [1-5,8,9,221-225]. This review also emphasizes the role of peroxides from fungi, fungal endophytes, algae, plants, lichens, and bryophytes as an important source of leads for drug discovery.

## References

1. Casteel DA (1992) Peroxy natural products. Nat Prod Rep 9: 289-312.
2. Casteel DA (1999) Peroxy natural products. Nat Prod Rep 16: 55-73.
3. Dembitsky VM, Glorizova TA, Poroikov VV (2007) Natural peroxy anticancer agents. Mini Rev Med Chem 7: 571-589.
4. Dembitsky VM (2008) Bioactive peroxides as potential therapeutic agents. Eur J Med Chem 43: 223-251.
5. Liu DZ, Liu JK (2013) Peroxy natural products. Nat Prod Bioprospect 3: 161-206.
6. Lange BM, Ahkami A (2013) Metabolic engineering of plant monoterpenes, sesquiterpenes and diterpenes--current status and future opportunities. Plant Biotechnol J 11: 169-196.
7. Liu QM, Jiang JG (2012) Antioxidative activities of medicinal plants from TCM. Mini Rev Med Chem 12: 1154-1172.
8. Terent'ev AO, Borisov DA, Vil' VA, Dembitsky VM (2014) Synthesis of five- and six-membered cyclic organic peroxides: Key transformations into peroxide ring-retaining products. Beilstein J Org Chem 10: 34-114.
9. Jung M, Kim H, Lee K, Park M (2003) Naturally occurring peroxides with biological activities. Mini Rev Med Chem 3: 159-165.

10. Sun Y, Zhao Z, Feng Q, Xu Q, Lue L, et al. (2013) Unusual spirodecane sesquiterpenes and a fumagillol analog from *Cordyceps ophioglossoides*. *Helv Chim Acta* 96: 76-84.
11. Ding L, Maier A, Fiebig HH, Lin WH, Peschel G, et al. (2012) Kandenols A-E, eudesmenes from an endophytic *Streptomyces* sp. of the mangrove tree *Kandelia candel*. *J Nat Prod* 75: 2223-2227.
12. Weiser J, Matha V, Jegorov A (1991) *Tolypocladium terricola* sp. N., a new mosquito-killing species of the genus *Tolypocladium* Gams (Hyphomycetes). *Folia Parasitol (Praha)* 38: 363-369.
13. Sedmera P, Halicek V, Jegorov A, Serge AL (1995) Cyclosporin D hydroperoxide a new metabolite of *Tolypocladium terricola*. *Tetrahedron Lett* 36: 6953-6956.
14. Wang G-J, Liang W-L, Ju Y-M, Yang W-B, Chang Y-W, et al. (2012) Inhibitory effects of terpenoids from the fermented broth of the ascomycete *Stilbohypoxylon elaeicola* YMJ173 on nitric oxide production in RAW264.7 macrophages. *Chem Biodiver* 9: 131-138.
15. Erb-Downward JR, Nogge RM, Williamson PR, Huffnagle GB (2008) The role of laccase in prostaglandin production by *Cryptococcus neoformans*. *Mol Microbiol* 68: 1428-1437.
16. Wang FZ, Fang YC, Zhu TJ, Zhang M, Lin AQ, et al. (2008) Seven new prenylated indole diketopiperazine alkaloids from holothurian-derived fungus *Aspergillus fumigatus*. *Tetrahedron* 7986-7991.
17. Zheng WF, Liu T, Xiang XY, Gu Q (2007) Sterol composition in field-grown and cultured mycelia of *Inonotus obliquus*. *Yao Xue Xue Bao* 42: 750-756.
18. Li Y, Niu S, Sun B, Liu S, Liu X, et al. (2010) Cytosporolides A-C, antimicrobial meroterpenoids with a unique peroxy lactone skeleton from *Cytospora* sp. *Org Lett* 12: 3144-3147.
19. She Z, Li H, Li M, Zhu X, Lin Y, et al. (2011) Norsesquiterpenoid peroxide with antitumor activity and preparation and application thereof. Chinese Patent: CN 2011-10031487: 6.
20. Li H, Huang H, Shao C, Huang H, Jiang J, et al. (2011) Cytotoxic norsesquiterpene peroxides from the endophytic fungus *Talaromyces flavus* isolated from the mangrove plant *Sonneratia apetala*. *J Nat Prod* 74: 1230-1235.
21. Liu DZ, Luo MH (2010) Two new chamigrane metabolites from fermentation broth of *Steccherinum ochraceum*. *Fitoterapia* 81: 1205-1207.
22. Liu D-Z, Dong Z-J, Wang F, Liu J-K (2010) Two novel norsesquiterpene peroxides from basidiomycete *Steccherinum ochraceum*. *Tetrahedron Lett* 3152-3153.
23. Li L-B, Ren J, Lai R, Cheng Z-M, Zhu H-J (2011) Natural cyclic peroxide echinobithiophene A with anti-microbial activity from *Echinops ritro* L. *Gaodeng Xuexiao Huaxue Xuebao* 32: 891-896.
24. Lington R, Navarro G, Pudhom K, McKerrow J (2013) Novel semi-synthetic small molecules for the treatment parasitic disease. Patent: WO 2012-US487: 21.
25. Chokpaiboon S, Sommit D, Teerawatananond T, Muangsin N, Bunyapaiboonsri T, et al. (2010) Cytotoxic nor-chamigrane and chamigrane endoperoxides from a basidiomycetous fungus. *J Nat Prod* 73: 1005-1007.
26. Liu L, Niu S, Lu X, Chen X, Zhang H, et al. (2010) Unique metabolites of *Pestalotiopsis fici* suggest a biosynthetic hypothesis involving a Diels-Alder reaction and then mechanistic diversification. *Chem Commun (Camb)* 46: 460-462.
27. Patel M, Horan AC, Gullo VP, Loebenberg D, Marquez JA, et al. (1984) Oxanthromycin, a novel antibiotic from *Actinomadura*. *J Antibiot (Tokyo)* 37: 413-415.
28. Wright JJK, Merrill Y, Puar M, McPhail AT (1984) Structure of oxanthromycin (antibiotic 16-550), a novel dimeric anthrone peroxide. *J Chem Soc Chem Commun* 473-474.
29. Takahashi S, Nakano T, Koiwa T, Noshita T, Funayama S, et al. (2000) Adxanthromycins A and B, new inhibitors of ICAM-1/LFA-1 mediated cell adhesion molecule from *Streptomyces* sp. NA-148. II. Physico-chemical properties and structure elucidation. *J Antibiotic (Tokyo)* 53: 163-170.
30. Nakano T, Koiwa T, Takahashi S, Nakagawa A (2000) Adxanthromycins A and B, new inhibitors of ICAM-1/LFA-1 mediated cell adhesion molecule from *Streptomyces* sp. NA-148. I. Taxonomy, production, isolation and biological activities. *J Antibiotic (Tokyo)* 53: 12-18.
31. Chen WS, Chen YT, Wan XY, Friedrichs E, Puff H, et al. (1981) Structure of hypocrellin and its photooxidation product peroxyhypocrellin. *Liebigs Ann Chem* 10: 880-885.
32. Zhenjun D, Lown JW (1990) Hypocrellins and their use in photosensitization. *Photochem Photobiol* 52: 609-616.
33. Fang LZ, Qing C, Shao HJ, Yang YD, Dong ZJ, et al. (2006) Hypocrellin D, a cytotoxic fungal pigment from fruiting bodies of the ascomycete *Shiraia bambusicola*. *J Antibiot (Tokyo)* 59: 351-354.
34. Shen YX, Rong XG, Gao ZH (2002) [Studies on the chemical constituents of *Shiraia bambusicola*]. *Zhongguo Zhong Yao Za Zhi* 27: 674-676.
35. Kishi T, Tahara S, Taniguchi N, Tsuda M, Tanaka C, et al. (1991) New perylenequinones from *Shiraia bambusicola*. *Planta Med* 57: 376-379.
36. Diwu Z (1995) Novel therapeutic and diagnostic applications of hypocrellins and hypericins. *Photochem Photobiol* 61: 529-539.
37. Diwu Z, Zhang C, Lown JW (1993) Photosensitization with anticancer agents. 18. Perylenequinonoid pigments as potential photodynamic therapeutic agents: preparation and photodynamic properties of amino-substituted hypocrellin derivatives. *Anticancer Drug Des* 8: 129-143.
38. Hudson JB, Zhou J, Chen J, Harris L, Yip L, et al. (1994) Hypocrellin, from *Hypocrella bambuase*, is phototoxic to human immunodeficiency virus. *Photochem Photobiol* 60: 253-255.
39. Cole RJ, Kirksey JW, Moore JH, Blankenship BR, Diener UL, et al. (1972) Tremorgenic toxin from *Penicillium veruculosum*. *Appl Microbiol* 24: 248-250.
40. Schroeder HW, Cole RJ, Hein H Jr, Kirksey JW (1975) Tremorgenic mycotoxins from *Aspergillus caespitosus*. *Appl Microbiol* 29: 857-858.
41. Dorner JW, Cole RJ, Hill RA (1984) Tremorgenic mycotoxins produced by *Aspergillus fumigatus* and *Penicillium crustosum* isolated from molded corn implicated in a natural intoxication of cattle. *J Agric Food Chem* 32: 411-413.
42. Patterson DS, Shreeve BJ, Roberts BA, MacDonald SM (1981) Verruculogen Produced by soil fungi in England and Wales. *Appl Environ Microbiol* 42: 916-917.
43. Gallagher RT, Latch GC (1977) Production of the Tremorgenic Mycotoxins Verruculogen and Fumitremorgin B by *Penicillium piscarium* Westling. *Appl Environ Microbiol* 33: 730-731.
44. Cockrum PA, Culvenor CC, Edgar JA, Payne AL (1979) Chemically different tremorgenic mycotoxins in isolates of *Penicillium paxilli* from Australia and North America. *J Nat Prod* 42: 534-536.
45. Day JB, Mantle PG, Shaw BI (1980) Production of verruculogen by *Penicillium estinogenum* in stirred fermenters. *J Gen Microbiol* 117: 405-410.
46. Day JB, Mantle PG (1982) Biosynthesis of radiolabeled verruculogen by *Penicillium simplicissimum*. *Appl Environ Microbiol* 43: 514-516.
47. Horie Y, Yamazaki M (1985) Production of carcinogenic mycotoxins, sterigmatocystin and its allied compounds, by *Emericella* species. *Trans Mycol Soc Japan* 26: 411-419.
48. Nielsen PV, Beuchat LR, Frisvad JC (1988) Growth of and fumitremorgin production by *Neosartorya fischeri* as affected by temperature, light, and water activity. *Appl Environ Microbiol* 54: 1504-1510.
49. Nishiyama M, Kuga T (1986) Pharmacological effects of the tremorgenic mycotoxin fumitremorgin A. *Jpn J Pharmacol* 40: 481-489.
50. Uramoto M, Tanabe M, Hirotsu K, Clardy J (1982) A new tremorgenic metabolite related to verruculogen from *Penicillium verruculosum*. *Heterocycles* 17: 349-358.
51. Yamazaki M, Fujimoto H, Kawasaki T (1980) Chemistry of tremorgenic metabolites. I. Fumitremorgin A from *Aspergillus fumigatus*. *Chem Pharm Bull (Tokyo)* 28: 245-254.

52. Yamazaki M, Suzuki S, Miyaki K (1971) Tremorgenic toxins from *Aspergillus fumigatus* Fres. Chem Pharm Bull (Tokyo) 19: 1739-1740.
53. Sanfilippo C, Patti A, Nicolosi G (1999) Enzymatic resolution of ( $\pm$ )-conduritol-B, a key intermediate for the synthesis of glycosidase inhibitors. Tetrahedron: Asymm 10: 3273-3276.
54. Dembitsky VM (2003) Oxidation, epoxidation and sulfoxidation reactions catalysed by haloperoxidases. Tetrahedron 59: 4701-4720.
55. Wang CF (2008) Studies on the chemical constituents about three plants of compositae and one plant of Thymelaeaceae. PhD thesis. Zhejiang University, China.
56. Miao FP, Li XD, Liu XH, Cichewicz RH, Ji NY (2012) Secondary metabolites from an algicolous *Aspergillus versicolor* strain. Mar Drugs 10: 131-139.
57. Mallavadhani UU, Sudhakar AVS, Satyanarayana KVS, Mahapatra A, Li W, et al. (2006) Chemical and analytical screening of some edible mushrooms. Food Chemistry 95: 58-64.
58. Yang RY, Li CY, Lin YC, Peng GT, Zhou SN (2006) [Study on the sterols from a brown alga endophytic fungus (NO. ZZ36) from the South China Sea]. Zhong Yao Cai 29: 908-909.
59. Makropoulou M, Aliogiannis N, Gonou-Zagou Z, Pratsinis H, Skaltsounis AL, et al. (2012) Antioxidant and cytotoxic activity of the wild edible mushroom *Gomphus clavatus*. J Med Food 15: 216-221.
60. Yokoyama S, Bang TH, Shimizu K, Kondo R (2012) Osteoclastogenesis inhibitory effect of ergosterol peroxide isolated from *Pleurotus eryngii*. Nat Prod Commun 7: 1163-1164.
61. Ji C, Sun S, Cai S, Zhu T, Gu Q, et al. (2011) Studies on the active secondary metabolites from the deep-sea-derived fungus *Aspergillus* sp. CXCTD-06-6a. Zhongguo Haiyang Yaowu 30: 1-6.
62. Zhang Y, Pei L, Gao L, Huang Q, Qi J (2011) [A neurotogenic compound from *Tremella fuciformis*]. Zhongguo Zhong Yao Za Zhi 36: 2358-2360.
63. Shi XW, Li XJ, Gao JM, Zhang XC (2011) Fasciculol H and I, two lanostane derivatives from Chinese mushroom *Naematoloma fasciculare*. Chem Biodivers 8: 1864-1870.
64. Huang HC, Liaw CC, Yang HL, Hseu YC, Kuo HT, et al. (2012) Lanostane triterpenoids and sterols from *Antrodia camphorata*. Phytochemistry 84: 177-183.
65. Zuo W, Luo D-Q (2010) Research on the chemical components of the fruit bodies of *Boletus calopus*. Anhui Nongye Kexue 38: 2356-2357.
66. Yaoita Y, Matsuki K, Iijima T, Nakano S, Kakuda R, et al. (2001) New sterols and triterpenoids from four edible mushrooms. Chem Pharm Bull (Tokyo) 49: 589-594.
67. Yue JM, Chen SN, Lin ZW, Sun HD (2001) Sterols from the fungus *Lactarium volemus*. Phytochemistry 56: 801-806.
68. Zang M, Ying JZ (1994) Economic fungi in the South West of China, Scientific Press. Beijing 307.
69. Fusetani N, Hashimoto K (1981) Diethyl peroxides. Probably responsible for mozuku poisoning. Bull Jpn Soc Sci Fish 47: 1059-1063.
70. Ikawa M, Sasner JJ, Haney JF (2001) Activity of cyanobacterial and algal odor compounds found in lake waters on green alga *Chlorella pyrenoidosa* growth. Hydrobiologia 443: 19-22.
71. Howard BM, Fenical W, Finer J, Hirotsu K, Clardy J (1977) Neoconcinndiol hydroperoxide, a novel marine diterpenoid from the red alga *Laurencia*. J Am Chem Soc 99: 6440-6441.
72. Yu XQ, He WF, Liu DQ, Feng MT, Fang Y, et al. (2014) A seco-laurane sesquiterpene and related laurane derivatives from the red alga *Laurencia okamurai* Yamada. Phytochemistry 103: 162-170.
73. Ji NY, Li XM, Li K, Wang BG (2007) Laurendecumallenes A-B and laurendecumenynes A-B, halogenated nonterpenoid C(15)-acetogenins from the marine red alga *Laurencia decumbens*. J Nat Prod 70: 1499-1502.
74. Teasdale ME, Shearer TL, Engel S, Alexander TS, Fairchild CR, et al. (2012) Bromophycoic acids: bioactive natural products from a Fijian red alga *Callophycus* sp. J Org Chem 77: 8000-8006.
75. Kolesnikova SA, Lyakhova EG, Kalinovsky AI, Dmitrenok PS, Dyshlovoy SA, et al. (2009) Diterpenoid hydroperoxides from the Far-Eastern brown alga *Dictyota dichotoma*. Aust J Chem 62: 1185-1188.
76. Fernandez JJ, Souto ML, Gil LV, Norte M (2005) Isolation of naturally occurring dactylomelane metabolites as *Laurencia* constituents. Tetrahedron 61: 8910-8915.
77. Qin JJ, Jin HZ, Zhu JX, Fu JJ, Hu XJ, et al. (2010) Japonicones E-L, dimeric sesquiterpene lactones from *Inula japonica* Thunb. Planta Med 76: 278-283.
78. Zhu JX, Qin JJ, Jin HZ, Zhang WD (2013) Japonicones Q-T, four new dimeric sesquiterpene lactones from *Inula japonica* Thunb. Fitoterapia 84: 40-46.
79. Boonprab K, Matsui K, Akakabe Y, Yotsukura N, Kajiwara T (2004) Arachidonic acid conversion by lipoxygenase in the brown alga, *Laminaria angustata*. Kasetsart J Nat Sci 38: 72-77.
80. Akakabe Y, Matsui K, Kajiwara T (2001) Enantioselective 2-hydroperoxylation of long-chain fatty acids in marine algae. Fish Sci 67: 328-332.
81. Nowicka B, Kruk J (2012) Plastoquinol is more active than  $\alpha$ -tocopherol in singlet oxygen scavenging during high light stress of *Chlamydomonas reinhardtii*. Biochim Biophys Acta 1817: 389-394.
82. Sabry OM, Andrews S, McPhail KL, Goeger DE, Yokochi A, et al. (2005) Neurotoxic meroditerpenoids from the tropical marine brown alga *Styopodium flabelliforme*. J Nat Prod 68: 1022-1030.
83. Morita M, Ohno O, Suenaga K (2012) Biselyngbyolide A, a novel cytotoxic macrolide from the marine cyanobacterium *Lyngbya* sp. Chem Lett 41: 165-167.
84. Morita M, Ohno O, Teruya T, Yamori T, Inuzuka T, et al. (2012) Isolation and structures of biselyngbyasides B, C, and D from the marine cyanobacterium *Lyngbya* sp., and the biological activities of biselyngbyasides. Tetrahedron 68: 5984-5990.
85. Kumar SS, Jayendra K (2012) Cytotoxicity of marine algal steroids in HeLa cells - 2D & 3D QSAR approach. Int J Pharm Bio Sci 3: 204-212.
86. Perme P, Saeidnia S, Mashinchian-Moradi A, Gohari AR (2012) Sterols from *Sargassum oligocystum*, a brown algae from the Persian Gulf, and their bioactivity. Nat Prod Res 26: 774-777.
87. Wang W, Li H, Wang Y, Xia X, Okada Y, et al. (2008) Chemical constituents from brown alga *Sargassum fusiforme*. Zhongcaoyao 39: 657-661.
88. Lo J-M, Wang W-L, Chiang Y-M, Chen C-M (2001) Ceramides from the Taiwan red alga *Ceratodictyon spongiosum* and symbiotic sponge *Sigmadocia symbiotica*. J Chinese Chem Soc (Taipei, Taiwan) 48: 821-826.
89. Qi SH, Zhang S, Huang JS, Xiao ZH, Wu J, et al. (2004) Glycerol derivatives and sterols from *Sargassum parvivesiculosum*. Chem Pharm Bull (Tokyo) 52: 986-988.
90. Dembitsky VM, Rezankova H, Rezanka T, Hanus LO (2003) Variability of the fatty acids of the marine green algae belonging to the genus *Codium*. Biochem Syst Ecol 31: 1125-1145.
91. He Z, Zhang A, Ding L, Lei X, Sun J, et al. (2010) Chemical composition of the green alga *Codium Divaricatum* Holmes. Fitoterapia 81: 1125-1128.
92. Sheu J-H, Liaw C-C, Duh C-Y (1995) Oxygenated clerosterols isolated from the marine alga *Codium arabicum*. J Nat Prod 58: 1521-1526.
93. Yang X-Y, Xu S-H, Guo S-H (2005) Study on sterols from *Laurencia cartilaginea*. Guangpu Shiyanshi 22: 12-16.
94. Sheu J-H, Sung P-J (1991) Isolation of 24-hydroperoxy-24-vinylcholesterol and fucosterol from the brown alga *Turbinaria conoides*. J Chinese Chem Soc 38: 501-503.
95. Yasukawa K, Akihisa T, Kanno H, Kaminaga T, Izumida M, et al. (1996) Inhibitory effects of sterols isolated from *Chlorella vulgaris* on 12-O-tetradecanoylphorbol-13-acetate-induced inflammation and tumor promotion in mouse skin. Biol Pharm Bull 19: 573-576.

96. Sheffer M, Fried A, Gottlieb HE, Tietz A, Avron M (1986) Lipid composition of the plasma membrane of the halotolerant alga, *Dunaliella salina*. *Biochim Biophys Acta* 857: 165-172.
97. Monaco P, Parrilli M, Previtera L (1987) Two endoperoxide diterpenes from *Elodea canadensis*. *Tetrahedron Lett* 28: 4609-4610.
98. Erickson KL, Beutler JA, Gray GN, Cardellina JH 2nd, Boyd MR (1995) Majapolene A, a cytotoxic peroxide, and related sesquiterpenes from the red alga *Laurencia majuscula*. *J Nat Prod* 58: 1848-1860.
99. Vairappan CS, Suzuki M, Ishii T, Okino T, Abe T, et al. (2008) Antibacterial activity of halogenated sesquiterpenes from Malaysian *Laurencia* spp. *Phytochemistry* 69: 2490-2494.
100. Gonzalez AG, Alvarez MA, Martin JD, Norte M, Perez C, et al. (1982) Diterpenoids of mixed biogenesis in phaeophyta biogenetic-type interconversions. *Tetrahedron* 38: 719-728.
101. Moghaddam FM, Farimani MM, Seirafi M, Taheri S, Khavasi HR, et al. (2010) Sesterterpenoids and other constituents of *Salvia sahendica*. *J Nat Prod* 73: 1601-1605.
102. Uchiyama N, Matsunaga K, Kiuchi F, Honda G, Tsubouchi A, et al. (2002) Trypanocidal terpenoids from *Laurus nobilis* L. *Chem Pharm Bull (Tokyo)* 50: 1514-1516.
103. Otoguro K, Iwatsuki M, Ishiyama A, Namatame M, Nishihara-Tukashima A, et al. (2011) In vitro antitrypanosomal activity of plant terpenes against *Trypanosoma brucei*. *Phytochemistry* 72: 2024-2030.
104. Tagliatela-Scafati O, Pollastro F, Cicione L, Chianese G, Bellido ML, et al. (2012) STAT-3 inhibitory bisabolanes from *Carthamus glaucus*. *J Nat Prod* 75: 453-458.
105. Asada Y, Sukemori A, Watanabe T, Malla KJ, Yoshikawa T, et al. (2013) Isolation, structure determination, and anti-HIV evaluation of tiglane-type diterpenes and biflavonoid from *Stellera chamaejasme*. *J Nat Prod* 76: 852-857.
106. Aboushoer MI, Fathy HM, Abdel-Kader MS, Goetz G, Omar AA (2010) Terpenes and flavonoids from an Egyptian collection of *Cleome droserifolia*. *Nat Prod Res* 24: 687-696.
107. Fattorusso E, Santelia FU, Appendino G, Ballero M, Tagliatela-Scafati O (2004) Polyoxygenated eudesmanes and trans-chrysanthemanes from the aerial parts of *Santolina insularis*. *J Nat Prod* 67: 37-41.
108. Cherchi G, Deidda D, De Gioannis B, Marongiu B, Pompei R, et al. (2001) Extraction of *Santolina insularis* essential oil by supercritical carbon dioxide: influence of some process parameters and biological activity. *Flavour Fragr J* 16: 35-43.
109. Lee SO, Choi SZ, Choi SU, Kim GH, Kim YC, et al. (2006) Cytotoxic terpene hydroperoxides from the aerial parts of *Aster spathulifolius*. *Arch Pharm Res* 29: 845-848.
110. Wang WS, Li EW, Jia ZJ (2002) Terpenes from *Juniperus przewalskii* and their antitumor activities. *Pharmazie* 57: 343-345.
111. Dembitsky VM (1993) Lipids of bryophytes. *Prog Lipid Res* 32: 281-356.
112. Becker H, Martini U (1999) Studies on the chemistry and biology of mosses, No. 140. Terpenoids from the in vitro cultured liverwort *Riella helicophylla*. *Zeitsch Naturfor C: J Biosci* 54: 997-1004.
113. Li Y, Wang Z, Zhang M, Luo S, Chen J (2002) Study on the terpene compounds from *Ligularia kanaitzensis*. *J Chinese Pharm Sci* 11: 115-117.
114. Sakunpak A, Matsunami K, Otsuka H, Panichayupakaranant P (2013) Isolation of new monoterpene coumarins from *Micromelum minutum* leaves and their cytotoxic activity against *Leishmania major* and cancer cells. *Food Chem* 139: 458-463.
115. Song QY, Jiang K, Zhao QQ, Gao K, Jin XJ, et al. (2013) Eleven new highly oxygenated triterpenoids from the leaves and stems of *Schisandra chinensis*. *Org Biomol Chem* 11: 1251-1258.
116. Graziose R, Rathinasabapathy T, Lategan C, Poulev A, Smith PJ, et al. (2011) Antiplasmodial activity of aporphine alkaloids and sesquiterpene lactones from *Liriodendron tulipifera* L. *J Ethnopharmacol* 133: 26-30.
117. Julianti E, Jang KH, Lee S, Lee D, Mar W, et al. (2012) Sesquiterpenes from the leaves of *Laurus nobilis* L. *Phytochemistry* 80: 70-76.
118. Zhang M, Iinuma M, Wang JS, Oyama M, Ito T, et al. (2012) Terpenoids from *Chloranthus serratus* and their anti-inflammatory activities. *J Nat Prod* 75: 694-698.
119. Yodsoue O, Sonprasit J, Karalai C, Ponglimanont C, Tewtrakul S, et al. (2012) Diterpenoids and triterpenoids with potential anti-inflammatory activity from the leaves of *Aglaia odorata*. *Phytochemistry* 76: 83-91.
120. Sripisut T, Ritthiwigrom T, Promgool T, Yossathera K, Deachathai S, et al. (2012) Glycopentaphyllone: The first isolation of hydroperoxyquinolone from the fruits of *Glycosmis pentaphylla*. *Phytochemistry Lett* 5: 379-381.
121. Shiu WK, Rahman MM, Curry J, Stapleton P, Zloh M, et al. (2012) Antibacterial acylphloroglucinols from *Hypericum olympicum*. *J Nat Prod* 75: 336-343.
122. Whitson EL, Thomas CL, Henrich CJ, Sayers TJ, McMahon JB, et al. (2010) Clerodane diterpenes from *Casearia arguta* that act as synergistic TRAIL sensitizers. *J Nat Prod* 73: 2013-2018.
123. Pacheco AG, Machado de Oliveira P, Piló-Veloso D, Flávio de Carvalho Alcântara A (2009) <sup>13</sup>C-NMR data of diterpenes isolated from *Aristolochia* Species. *Molecules* 14: 1245-1262.
124. Lopes LMX, Bolzani VS, Trevisan LMV (1987) Clerodane diterpenes from *Aristolochia* species. *Phytochemistry* 26: 2781-2784.
125. Bomm MD, Zukerman-Schpector J, Lopes LMX (1999) Rearranged (4→2)-abeo-clerodane and clerodane diterpenes from *Aristolochia chamissonis*. *Phytochemistry* 50: 455-461.
126. Ono M, Tsuru T, Abe H, Eto M, Okawa M, et al. (2006) Bisabolane-type sesquiterpenes from the aerial parts of *Lippia dulcis*. *J Nat Prod* 69: 1417-1420.
127. Jung CM, Kwon HC, Seo JJ, Ohizumi Y, Matsunaga K, et al. (2001) Two new monoterpene peroxide glycosides from *Aster scaber*. *Chem Pharm Bull (Tokyo)* 49: 912-914.
128. Chen YL, Lan YH, Hsieh PW, Wu CC, Chen SL, et al. (2008) Bioactive cembrane diterpenoids of *Anisomeles indica*. *J Nat Prod* 71: 1207-1212.
129. Minami H, Anzaki S, Kubo M, Kodama M, Kawazu K, et al. (1998) Structures of new seven-membered ring vibsane-type diterpenes isolated from leaves of *Viburnum awabuki*. *Chem Pharm Bull* 46: 1194-1198.
130. Fukuyama Y, Minami H, Matsuo A, Kitamura K, Akizuki M, et al. (2002) Seven-membered vibsane-type diterpenes with a ,10-cis relationship from *Viburnum awabuki*. *Chem Pharm Bull (Tokyo)* 50: 368-371.
131. Ngo KS, Brown GD (1999) Allohimachalane, seco-allohimachalane and himachalane sesquiterpenes from *Illicium tsangii*. *Tetrahedron* 55: 759-770.
132. Ngo KS, Brown GD (1999) Santalane and isocampherene sesquiterpenoids from *Illicium tsangii*. *Phytochemistry* 50: 1213-1216.
133. Zhang HJ, Tan GT, Hoang VD, Hung NV, Cuong NM, et al. (2003) Natural anti-HIV agents. Part IV. Anti-HIV constituents from *Vatica cinerea*. *J Nat Prod* 66: 263-268.
134. Vassilikogiannakis G, Margaros I, Montagnon T (2004) Biomimetic total synthesis of litseaverticillols B, E, I, and J and structural reassignment of litseaverticillol E. *Org Lett* 6: 2039-2042.
135. Huber H, Frohlike E (1972) A new spray-reagent for the detection and quantitative estimation of peroxides. *Chromatographia* 5: 256-257.
136. Portillo A, Vila R, Freixa B, Adzet T, Cañigueral S (2001) Antifungal activity of Paraguayan plants used in traditional medicine. *J Ethnopharmacol* 76: 93-98.
137. Bah M, Bye R, Pereda-Miranda R (1994) Hepatotoxic pyrrolizidine alkaloids in the Mexican medicinal plant *Packera candidissima* (Asteraceae: Senecioneae). *J Ethnopharm* 43: 19-30.
138. Hammond GB, Fernández ID, Villegas LF, Vaisberg AJ (1998) A survey of traditional medicinal plants from the Callejón de Huaylas, Department of Ancash, Perú. *J Ethnopharmacol* 61: 17-30.
139. RÄ¼cker G, Manns D, Schenkel EP, Hartmann R, Heinzmann BM (1999) Sesquiterpene peroxides from *Senecio selloi*. *Phytochemistry* 52: 1587-1591.



140. Krügener S, Schaper C, Krings U, Berger RG (2009) Pleurotus species convert monoterpenes to furanoterpenoids through  $\alpha$ -endoperoxides. *Bioresour Technol* 100: 2855-2860.
141. Tiew P, Takayama H, Kitajima M, Aimi N, Kokpol U, et al. (2003) A novel neolignan, mansoxetane, and two new sesquiterpenes, mansonones R and S, from *Mansonia gagei*. *Tetrahedron Lett* 44: 6759-6761.
142. Cheng SY, Wang SK, Wen ZH, Dai CF, Duh CY (2009) Three new eudesmanoids from the Formosan soft coral *Nephtea erecta*. *J Asian Nat Prod Res* 11: 967-973.
143. Lee T-H, Lu C-K, Kuo Y-H, Lo JM, Lee CK (2008) Unexpected novel pheophytin peroxides from the leaves of *Biden pilosa*. *Helvetica Chim. Acta* 91: 79-84.
144. Park S, Im SA, Kim KH, Lee CK (2011) Immunomodulatory Effects of Hypocrellin A on MHC-restricted Antigen Processing. *Immune Netw* 11: 412-415.
145. Marco JA, Sanz-Cervera JF, Ropero FJ, Batlle N, Guara M, et al. (1998) Germacranolides and a monoterpene cyclic peroxide from *Artemisia fragrans*. *Phytochemistry* 47: 1417-1419.
146. Růžek G, Breitmaier E, Mayer R, Manns D (1987) Peroxide als Pflanzeninhaltsstoffe, 4. Mitt. Ein cyclisches  $\alpha$ -ungliedriges Sesquiterpen-Peroxyhalbketal aus *Artemisia maritima*. *Archiv Pharmazie* 320: 437-441.
147. Rucker G, Manns D, Wilbert S (1993) Peroxide als Pflanzeninhaltsstoffe, 12. Mitt.: Peroxide vom Davanon-Typ aus der Eberraute (*Artemisia abrotanum*) und ihre Darstellung. *Archiv Pharmazie* 326: 457-460.
148. Adam G, Porzel A, Sung TV, Schmidt J (1992) A monoterpene peroxide from *Adenosma caeruleum*. *Phytochemistry* 31: 2885-2887.
149. Labbe C, Castillo M, Connolly J (1993) Mono and sesquiterpenoids from *Satureja gilliesii*. *Phytochemistry* 34: 441-444.
150. Bagchi A, Oshima Y, Hikino H (1988) Kanshones A and B, sesquiterpenoids of *Nardostachys chinensis*. *Phytochemistry* 27: 1199-1201.
151. Schulte KE, Růžek G, Glauch G (1967) [On some components of underground parts from *Nardostachys chinensis* Batalin]. *Planta Med* 15: 274-281.
152. Gao J-F, Xie J-H, Iitaka Y, Inayama S (1989) The stereostructure of wenjine and related (1S,10S), (4S,5S)-germacrone-1(10),4-diepoxy isolated from *Curcuma wenyujin*. *Chem Pharm Bull* 37: 233-236.
153. Gao J-F, Xie J-H, Harimaya H, Kawamata T, Iitaka Y, et al. (1991) The absolute structure and synthesis of Wenjine isolated from *Curcuma wenyujin*. *Chem Pharm Bull* 39: 854-856.
154. Xu J, Zhao X, Guo YQ, Zhang S (2009) A novel sesquiterpene from the roots of *Vladimiria souliei*. *Pharmazie* 64: 623-624.
155. Todorova M, Trendafilova A, Mikhova B, Vitkova A, Duddeck H (2007) Terpenoids from *Achillea distans* Waldst. & Kit. ex Willd. *Biochem Syst Ecol* 35: 852-858.
156. Trifunovic SS (2006) Comparative investigation of the chemical composition of some plant species of the genus *Achillea* L. Dissertation. Belgrade. University of Belgrade, Faculty of Chemistry.
157. Si XT, Zhang ML, Shi QW, Kiyota H (2006) Chemical constituents of the plants in the genus *Achillea*. *Chem Biodivers* 3: 1163-1180.
158. Trifunovic S, Isakovic AM, Isakovic A, Vuckovic I, Mandic B, et al. (2014) Isolation, characterization, and in vitro cytotoxicity of new sesquiterpenoids from *Achillea clavennae*. *Planta Med* 80: 297-305.
159. Aguilar-Guadarrama AB, Rios MY (2004) Three new sesquiterpenes from *Croton arboreus*. *J Nat Prod* 67: 914-917.
160. Bohlmann F, Zdero C, Jakupovic J, Greger H (1983) Types of sesquiterpenes from *Artemisia filifolia*. *Phytochemistry* 22: 503-507.
161. Rustaiyan A, Sigari H, Jakupovic J, Grenz M (1989) A sesquiterpene lactone from *Artemisia diffusa*. *Phytochemistry* 28: 2723-2725.
162. Kamperdick C, Phuong NM, Van Sung T, Adam G (2001) Guaiane dimers from *Xylophia vielana*. *Phytochemistry* 56: 335-340.
163. Saito Y, Hattori M, Iwamoto Y, Takashima Y, Mihara K, et al. (2011) Overlapping chemical and genetic diversity in *Ligularia lamarum* and *Ligularia subspicata*. Isolation of ten new eremophilanes and a new seco-bakkane compound. *Tetrahedron* 67: 2220-2231.
164. Zhao X, Zheng GW, Niu XM, Li WQ, Wang FS, et al. (2009) Terpenes from *Eupatorium adenophorum* and their allelopathic effects on *Arabidopsis* seeds germination (dagger). *J Agric Food Chem* 57: 478-482.
165. Ding Y, Liang C, Kim JH, Lee YM, Hyun JH, et al. (2010) Triterpene compounds isolated from *Acer mandshuricum* and their anti-inflammatory activity. *Bioorg Med Chem Lett* 20: 1528-1531.
166. Chen JJ, Fei DQ, Chen SG, Gao K (2008) Antimicrobial triterpenoids from *Vladimiria muliensis*. *J Nat Prod* 71: 547-550.
167. He F, Pu JX, Huang SX, Wang YY, Xiao WL, et al. (2010) Schinalactone A, a new cytotoxic triterpenoid from *Schisandra sphenanthera*. *Org Lett* 12: 1208-1211.
168. Zhou T, Zhang H, Zhu N, Chiu P (2004) New triterpene peroxides from *Pseudolarix kaempferi*. *Tetrahedron* 60: 4931-4936.
169. Tan JM, Qiu YH, Tan XQ, Tan CH (2011) Three new peroxy triterpene lactones from *Pseudolarix kaempferi*. *Helv Chim Acta* 94: 1697-1702.
170. Dembitsky V, Shkrob I, Hanus LO (2008) Ascaridole and related peroxides from the genus *Chenopodium*. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 152: 209-215.
171. Cavalli JF, Tomi F, Bernardini AF, Casanova J (2004) Combined analysis of the essential oil of *Chenopodium ambrosioides* by GC, GC-MS and <sup>13</sup>C-NMR spectroscopy: quantitative determination of ascaridole, a heat-sensitive compound. *Phytochem Anal* 15: 275-279.
172. Duviau G (1953) [Treatment of intestinal parasitosis]. *J Med Bord* 130: 44-51.
173. Guimaraes F (1927) Parasitotropism of essence of *Chenopodium* for Entozoarians. *Comp Rend Sean Soc Biol* 96: 1251-1253.
174. Faust EC (1937) The use of anthelmintics. *J Am Med Assoc* 108: 386-392.
175. Wautie A (1946) Prophylaxis and treatment of the chief parasitic diseases of the gastrointestinal tract of the horse. *Parasitica* 2: 44-67.
176. Pare PW, Zajicek J, Ferracini VL, Melo IS (1993) Antifungal terpenoids from *Chenopodium ambrosioides*. *Biochem Syst Ecol* 21: 649-653.
177. Pollack Y, Segal R, Golenser J (1990) The effect of ascaridole on the in vitro development of *Plasmodium falciparum*. *Parasitol Res* 76: 570-572.
178. Kiuchi F, Itano Y, Uchiyama N, Honda G, Tsubouchi A, et al. (2002) Monoterpene hydroperoxides with trypanocidal activity from *Chenopodium ambrosioides*. *J Nat Prod* 65: 509-512.
179. Monzote L, Montalvo AM, Almanonni S, Scull R, Miranda M, et al. (2006) Activity of the essential oil from *Chenopodium ambrosioides* grown in Cuba against *Leishmania amazonensis*. *Chemotherapy* 52: 130-136.
180. Efferth T, Olbrich A, Sauerbrey A, Ross DD, Gebhart E, et al. (2002) Activity of ascaridol from the anthelmintic herb *Chenopodium anthelminticum* L. against sensitive and multidrug-resistant tumor cells. *Anticancer Res* 22: 4221-4224.
181. Roth HJ (1968) Photochemistry and pharmaceuticals. *Farm Glasnik* 24: 349-359.
182. Sekera A, Rahm J (1952) [Antural althelminthic drugs; askaridol]. *Cesk Farm* 1: 471-477.
183. Sichaem J, Siripong P, Khumkratok S, Tip-Pyang S (2010) Chemical constituents from the roots of *Bombax anceps*. *J Chilean Chem Soc* 55: 325-327.
184. Danelutte AP, Lago JH, Young MC, Kato MJ (2003) Antifungal flavanones and prenylated hydroquinones from *Piper crassinervium* Kunth. *Phytochemistry* 64: 555-559.
185. Nishikawa K, Aburai N, Yamada K, Koshino H, Tsuchiya E, et al. (2008) The bisabolane sesquiterpenoid endoperoxide,  $\alpha$ -epidioxy-,10-bisaboladiene, isolated from *Cacalia delphiniifolia* inhibits the growth of human cancer cells and induces apoptosis. *Biosci Biotech Biochem* 72: 2463-2466.
186. De Pascual-Teresa J, Caballero E, Caballero C, Anaya J, Gonzalez MS (1983) Four aliphatic esters of *Chamaemelum fuscum* essential oil. *Phytochemistry* 22: 1757-1759.

187. De Pascual-Teresa J, Anaya J, Caballero E, Caballero MC (1988) Sesquiterpene lactones and aliphatic ester from *Chamaemelum fuscum*. *Phytochemistry* 27: 855-860.
188. Bohlmann F, Tsankova E, Jakupovic J, King RM, Robinson H (1983) Dimeric chromenes and mixed dimers of a chromene with euparin from *Encelia canescens*. *Phytochemistry* 22: 557-560.
189. Rucker G, Schenkel EP, Manns D, Mayer R, Heiden K, et al. (1996) Sesquiterpene peroxides from *Senecio selloi* and *Eupatorium rufescens*. *Planta Med* 62: 565-566.
190. Rucker G, Schenkel E, Manns D, Falkenberg M, Marek A (1996) Peroxides and other constituents from *Heterothalamus alienus*. *Phytochemistry* 41: 297-300.
191. Rucker G, Walter RD, Manns D, Mayer R (1991) Antimalarial activity of some natural peroxides. *Planta Med* 57: 295-296.
192. Bohlmann F, Fritz U (1980) Isofukinone, an eremophilane from *Ligularia speciosa*. *Phytochemistry* 19: 2471-2472.
193. Tschritzis F, Jakupovic J (1990) Diterpenes and other constituents from *Relhania* species. *Phytochemistry* 29: 3173-3187.
194. Castro V, Jakupovic J, Dominguez XA (1989) Melampolides from *Melampodium* and *Smallanthus* species. *Phytochemistry* 28: 2727-2729.
195. Yu DQ, Chen RY, Huang LJ, Xie FZ, Ming DS, et al. (2008) The structure and absolute configuration of Shuangkangsu: a novel natural cyclic peroxide from *Lonicera japonica* (Thunb.). *J Asian Nat Prod Res* 10: 851-856.
196. Arndt R, Wahlberg I, Enzell CR, Berg J-E (1988) Tobacco chemistry. 68. Structuredetermination and biomimetic synthesis of two new tobacco cembranoids. *Acta Chem Scand B* 42: 294-298.
197. Lee SS, Lin YS, Chen CK (2009) Three adducts of butenolide and apigenin glycoside from the leaves of *Machilus japonica*. *J Nat Prod* 72: 1249-1252.
198. Adelekan AM, Prozesky EA, Hussein AA, Ureña LD, van Rooyen PH, et al. (2008) Bioactive diterpenes and other constituents of *Croton steenkampianus*. *J Nat Prod* 71: 1919-1922.
199. Pooley E (1993) *The Complete Field Guide to Trees of Natal, Zululand and Transkei*. Natal Flora Publications Trust, Durban, pp. 512.
200. Pérez-Gutiérrez S, Sánchez-Mendoza E, Martínez-González D, Zavala-Sánchez MA, Pérez-González C (2012) Kramecyne-a new anti-inflammatory compound isolated from *Krameria cysisoides*. *Molecules* 17: 2049-2057.
201. Yang XW, Feng L, Li SM, Liu XH, Li YL, et al. (2010) Isolation, structure, and bioactivities of abiesadines A-Y, 25 new diterpenes from *Abies georgei* Orr. *Bioorg Med Chem* 18: 744-754.
202. Kong LY, Min ZD, Shi JX (1996) Chemical constituents from roots of *Jatropha curcas*. *Acta Bot Sinica* 38: 161-166.
203. Tian G-H, Liu CF, Lai P-H, Zhao H, Nie F (2010) Configuration and conformation of taibaihenryiin T isolated from *Phlomis umbrosa*. *Chem Nat Comp* 46: 219-221.
204. Ma Y-P, Li N, Gao J, Fu K-L, Qin Y, et al. (2011) A new peroxy-multiflorane triterpene ester from the processed seeds of *Trichosanthes kirilowii*. *Helvetica Chim Acta* 94: 1881-1887.
205. Xiao ZY, Zeng YH, Mu Q, Shiu WK, Gibbons S (2010) Prenylated benzophenone peroxide derivatives from *Hypericum sampsonii*. *Chem Biodivers* 7: 953-958.
206. Moreira IC, Roque NF, Contini K, Lago JHG (2007) Sesquiterpenes and hydrocarbons from the fruits of *Xylopia emarginata* (Annonaceae). *Revista Brasil Farmac* 17: 55-58.
207. Thebtaranonth C, Thebtaranonth Y, Wanauppathamkul S, Yuthavong Y (1995) Antimalarial sesquiterpenes from tubers of *Cyperus rotundus*: structure of 10,12-peroxycalamenene, a sesquiterpene endoperoxide. *Phytochemistry* 40: 125-128.
208. Kamchonwongpaisan S, Nilanonta C, Tarmchampoo B, Thebtaranonth C, Thebtaranonth Y, et al. (1995) An antimalarial peroxide from *Amomum krervanh* Pierre. *Tetrahedron Lett* 36: 1821-1824.
209. Sy LK, Brown GD (1997) Labdane Diterpenoids from *Alpinia chinensis*. *J Nat Prod* 60: 904-908.
210. Nakatani N, Kikuzaki H, Yamaji H, Yoshio K, Kitora C, et al. (1994) Labdane diterpenes from rhizomes of *Hedychium coronarium*. *Phytochemistry* 37: 1383-1388.
211. Sy LK, Brown GD, Haynes R (1998) A novel endoperoxide and related sesquiterpenes from *Artemisia annua* which are possibly derived from allylic hydroperoxides. *Tetrahedron* 54: 4345-4356.
212. Zhu X, Zhang Q, Kong L, Wang F, Luo S (2010) New hydroquinone diglycoside acyl esters and sesquiterpene and apocarotenoid from *Ecdysanthera rosea*. *Fitoterapia* 81: 906-909.
213. Kameoka H, Kubo K, Miyazawa M (1991) Volatile flavor components of malabar-nightsshade (*Basella rubra* L.). *J Food Compos Anal* 4: 315-321.
214. Hiroi M, Takaoka D (1973) Studies on the peel oil of *Citrus iyo*. *Nippon Kagaku Kaishi* 1339-1341.
215. Kobayashi A, Nohara K, Ohsumi E, Yamanishi T (1990) Identification of di(l-oxyalkyl)-peroxides in the essential oil of *Citrus iyo*. *Agric Biol Chem* 54: 561.
216. de Mesquita ML, Araújo RM, Bezerra DP, Filho RB, de Paula JE, et al. (2011) Cytotoxicity of 6-tocotrienols from *Kielmeyera coriacea* against cancer cell lines. *Bioorg Med Chem* 19: 623-630.
217. Yang H, Hou AJ, Mei SX, Sun HD, Che CT (2002) Constituents of *Clerodendrum bungei*. *J Asian Nat Prod Res* 4: 165-169.
218. Devkota KP, Lenta BN, Wansi JD, Sewald N (2010) Antibacterial constituents from *Leucosceptrum canum*. *Phytochemistry Lett* 3: 24-28.
219. Shen YC, Prakash CVS, Guh JH (2004) New pentacyclic polyketide dimeric peroxides from a Taiwanese marine sponge *Petrosia elastica*. *Tetrahedron Lett* 45: 2463-2466.
220. Wang XX, Lin CJ, Jia ZJ (2006) Triterpenoids and Sesquiterpenes from *Mulgedium tataricum*. *Planta Med* 72: 764-767.
221. Dembitsky VM (2004) Chemistry and biodiversity of the biologically active natural glycosides. *Chem Biodivers* 1: 673-781.
222. Youns M, Hoheisel JD, Efferth T (2010) Traditional Chinese medicines (TCMs) for molecular targeted therapies of tumours. *Curr Drug Discov Technol* 7: 37-45.
223. Angus B1 (2014) Novel anti-malarial combinations and their toxicity. *Expert Rev Clin Pharmacol* 7: 299-316.
224. Farhi M, Kozin M, Duchin S, Vainstein A (2013) Metabolic engineering of plants for artemisinin synthesis. *Biotechnol Genet Eng Rev* 29: 135-148.
225. Pastre JC, Browne DL, Ley SV (2013) Flow chemistry syntheses of natural products. *Chem Soc Rev* 42: 8849-8869.