

Original Research Article

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## Evaluation of the Fertilizing and Nematicidal Effects of Lixivate from Banana Rachis and *Purpureocillium lilacinum* for a Reduction of Fallow Frequency in Dessert Banana Monoculture

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### ABSTRACT

The practice of fallowing for one year represents a viable alternative to chemical control of *Radopholus similis* and *Pratylenchus coffeae* in dessert banana monoculture. Although necessary, fallowing has an economic disadvantage for farmers. The objective of this work is to evaluate the nematicidal and fertilizing effects of lixivate from banana rachis and *Purpureocillium lilacinum* in order to reduce the frequency of fallowing. In this study, we performed a chemical analysis of the lixivate and then compared the two types (plantain lixivate and dessert lixivate) at 25% concentration, Bioact (*Purpureocillium lilacinum*) at 10<sup>6</sup> spores/ml) and two mixed treatments (plantain lixivate + Bioact and dessert lixivate + Bioact) to an untreated control and a reference nematicide, fluopyram at 10% (Verango) during three cycles of cultivation of the dessert banana seedling Grande naine. At the end of each crop cycle, the evolution of the nematode population and the yield of banana plants were evaluated. Chemical analysis revealed a high potassium content in both products and a more remarkable amount of nitrogen in the plantain lixivate. The results showed that Bioact did not show antagonistic effect against nematodes and impact on production. On the other hand, despite less nematicidal activity, the lixivate significantly increased the yield compared to the chemical nematicide treatment. Therefore, only in the plots treated with lixivate, it was possible to carry out several successive crop cycles. This result was more marked with the plantain lixivate. The results presented in this work are encouraging for the development of biological control methods of banana nematodes by lixivate.

#### Keywords

*Radopholus similis*,  
*Pratylenchus coffeae*, fallow land,  
banana plantain  
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## Introduction

Among the production constraints, endoparasitic nematodes (*Radopholus similis* and *Pratylenchus coffeae*) constitute one of the major limiting factors to banana production, causing extensive root damage, resulting in serious economic losses (Queneherve, 2009). According to the "resource concentration hypothesis" (Root 1973), monocultures are more prone to diseases and pest infestations. In the case of banana monocultures, the soils have large populations of *Radopholus similis* and *Pratylenchus coffeae* and a problem of fertility degradation after a few cropping cycles (Risède *et al.*, 2018; Pattison *et al.*, 2011; Gnonhoury and Adiko 2008). This situation forces farmers to turn to prophylactic methods such as fallowing for at least one year to allow the soil to regenerate while eliminating food-deprived nematodes (Tabarant *et al.*, 2011). However, the regular integration of the fallow phase into the farmer's calendar is a practice that does not always lead to economic benefits for most farms: production targets are set and replanted *in vitro* plants are expensive. For the small producer, the practice of fallowing is inconvenient because it requires sufficient acreage to be able to deprive himself of a part of his farm (Chabrier, 2005). These different constraints make it a priority to find solutions that will prevent the development of nematodes and promote long-term soil enrichment. The use of organic matter leads to a series of biological processes related to its degradation, which can reduce plant-parasitic nematode populations and restore the soil's organic matter stock and humus (Oka, 2010). Lixivate, a liquid organic compound, is extracted from plantain and banana rachis. This product has demonstrated fertilizing efficacy in tomato crops and nematicidal efficacy under *in vivo* conditions (Muñoz, 2005; Seri *et al.*, 2018). Another strategy for the sustainable management of nematodes and

improving soil fertility is through the use of biological control agents such as nematophagous fungi (Hernández-Leal *et al.*, 2016). Among nematophagous fungi, *Purpureocillium lilacinum* (former name var *Paecilomyces lilacinum*) is of increasing interest by virtue of its ability to parasitize nematodes and improve soil fertility (Messa, 2020). The objective of the present study is to evaluate the nematicidal and fertilizing effects of lixivate from banana rachis and *Purpureocillium lilacinum* in a context of reduced fallow frequency in dessert banana cv Grande naine plantations.

## Materials and Methods

### Plant material

The plant material consisted of dessert banana cv Grande naine *in vitro* plants. The lixivate was obtained from plantain cv Corne 1, and dessert banana cv Grande naine rachis.

### Fungal material

*Purpureocillium lilacinum*, trade name Bioact DC, was supplied to us by the firm Bayer Crop Science.

### Lixivate preparation

The preparation of the lixivate from plantain (lixp) and banana dessert (lixd) rachis was done using an anaerobic composting process. The slightly modified INRA, (2014) method was used for lixivate preparation. The rachis were fragmented (3 cm thick and 10 cm long). Eight kilograms of cut rachis were mixed with 15 l of distilled water in a 20 l drum, then hermetically sealed and stored at 27°C at the Nematology Laboratory of the CNRA or National Agricultural Research Center. The mixture was stirred manually and vigorously each day for 2 min. After three months of maceration, the resulting solution was filtered

through a sieve (250  $\mu$  mesh diameter) to obtain the lixiviate.

### **Mineral composition and chemical properties of lixiviate**

The constituents were quantified by various methods:

total nitrogen was determined by the Kjeldhal method (Goyal *et al.*, 2005);

potassium (K) and calcium (Ca) were determined by flame spectrophotometry after incineration and mineralization with hydrochloric acid;

phosphoric acid was determined by the colorimetric method on the hydrochloric solution of solution of the ashes thanks to the yellow coloration that it gives with the vanadomolybdic reagent;

pH of the lixiviate was measured with a pH meter.

### **Experimental device**

The trial was conducted on a SAKJ plot, ready to be fallowed, which had a high inoculum (21000 endoparasitic nematodes/100 g root) and a low productivity (15 t/ha). The trial started three months after planting the in vitro plants and was conducted over three growing cycles. The preliminary nematological analysis allowed to verify the homogeneity of the initial infestation within the plot (6000 endoparasitic nematodes/100 g of roots).

Both types of lixiviate (plantain lixp and dessert lixd) at the 25% concentration, Bioact DC at  $10^6$  spores/ml and two mixed treatments [plantain lixiviate (lixp) + Bioact and banana dessert lixiviate (lixd) + Bioact) were compared to an untreated control and a reference nematicide, Fluopyram at 10%

(Verango). Lixiviate treatments were applied monthly until the end of the trial at 500 ml/plant. Bioact was applied every two months for six months at 82 ml/plant. For the mixed treatments (lixiviate + Bioact), the two products were applied alternately every month for six months and subsequent applications were only to lixiviate every month until the end of the trial. Fluopyram was applied once every four months at an amount of 50 ml/plant. The device chosen was a complete randomized block with five replications. Each plot was 275 m<sup>2</sup> (11 m x 25 m) and consisted of 48 plants, 24 of which were studied, surrounded by a belt of 24 border plants.

### **Nematological analysis**

The nematode population was assessed at flowering of each crop cycle after extraction of nematodes by the double centrifugation technique (Coolen and d'Herde, 1972). Nematode numbers were expressed as the number of nematodes per 100 g of roots.

### **Evaluation of vegetative and production parameters**

The vegetative parameters that were measured at flowering of each crop cycle were the height (H) and circumference (C) of banana plants. They are expressed in cm.

The following production parameters were evaluated: planting-flowering interval (IPF), flowering-harvest interval (IFR), number of hands (Nm), bunch mass (M) and proportion of bunch harvested (Pr). The gross yield (Rb) of each cycle was calculated according to the following formula:

$$Rb = (Mm \cdot PR \cdot D \cdot 365) / Cy$$

Mm: average mass of a bunch, Pr: proportion of bunches harvested, D: density of plants (plants/ha) and Cy: cycle length in days.

### Statistical analysis

ANOVAs were performed to compare the means of the variables by treatment. In case of significant difference at the 5% level, Fisher's LSD test was used to compare the means. Nonparametric tests were also performed to test the significance ( $P < 0.05$ ) of some means per treatment. Nematode populations were transformed to  $\log_{10}(x+1)$  decimal and percentages were transformed using the arcsine ( $x/100$ ) before statistical analysis.

### Results and Discussion

#### Mineral content and chemical properties of lixiviate

Analysis of the content of mineral constituents [nitrogen (N), phosphorus (P), potassium (K), calcium (Ca)] of the different lixiviate shows a significant difference for the average values of nitrogen and calcium (Table 1). The plantain lixiviate is richer in nitrogen (0.9 vs. 0.3). For calcium, the dessert lixiviate has the highest content (41 vs. 17). The pH values of the two lixiviate are statistically close.

#### Effects of lixiviate and Bioact on *Radopholus similis* and *Pratylenchus coffeae*

Population results of *R. similis* and *P. coffeae* nematodes at flowering for the three crop cycles are recorded in Table 2. In the control and Bioact treated plots, the development of infestations was statistically similar over the three cycles. At flowering of the first cycle, the population level was around 14000 nematodes/100 g of roots. Infestations increased steadily with the favorable rainfall conditions at flowering of the 2nd and 3rd cycle (20000 nematodes/100 g of roots). This result shows that Bioact did not have a nematocidal effect. At all lixiviate plots (lixp, lixd, lixp + Bioact and lixd + Bioact), nematode population densities were statistically similar. The level of infestation (6000 nematodes/100 g roots) was significantly lower in the control and Bioact plots but higher than in the plots that received the chemical treatment (2200 nematodes/100 g roots). These results show a more effective nematocidal action of fluopyram compared to lixiviate.

**Table.1** Mineral content (N, P, K, Ca) and pH of the lixiviate

| Lixiviate | Nt %  | P   | K   | Ca   | pH  |
|-----------|-------|-----|-----|------|-----|
| Plantain  | 0,9 a | 1,6 | 471 | 17 b | 7,4 |
| Dessert   | 0,3 b | 1,5 | 475 | 41 a | 7,1 |

In the same column, the averages affected with different letters indicate significant differences between treatments according to Student T test ( $P=0,05$ ).

**Table.2** Population of endoparasitic nematodes at flowering in each cycle

| Cycles  | Treatments |           |         |        |        |               |               | Meaning |
|---------|------------|-----------|---------|--------|--------|---------------|---------------|---------|
|         | Control    | Fluopyram | Bioact  | Lixp   | Lixd   | Lixp + Bioact | Lixd + Bioact |         |
| Cycle 1 | 13349 a    | 2123 c    | 14920 a | 5575 b | 6111 b | 5573 b        | 5518 b        | S       |
| Cycle 2 | 20471 a    | 1837 c    | 18766 a | 5668 b | 7221 b | 5094 b        | 6359 b        | S       |
| Cycle 3 | 22076 a    | 2668 c    | 19023 a | 6271 b | 5394 b | 6165 b        | 6010 b        | S       |
| Moyenne | 17965 a    | 2209 c    | 17570 a | 5838 b | 6242 b | 5610 b        | 5963 b        | S       |

In the same row, the averages affected with different letters indicate significant differences between treatments ( $P=0,05$ )

Plantain lixiviate : lixp ; lixiviate dessert: lixd. Data are averages of five replicates

**Table.3** Vegetative and production parameters at flowering of each cycle

| Cycles | Parameters | Treatments |           |         |         |         |               |               |    |
|--------|------------|------------|-----------|---------|---------|---------|---------------|---------------|----|
|        |            | Control    | Fluopyram | Bioact  | Lixp    | Lixd    | Lixp + Bioact | Lixd + Bioact |    |
| 1      | H (cm)     | 206,8 ± b  | 223,8 a   | 207,9 b | 227,4 a | 222,7 a | 228,2 a       | 225,5 a       | S  |
|        | C (cm)     | 41,8 b     | 47,5 a    | 42,3 b  | 48,1 a  | 47,6 a  | 48,1 a        | 47,6 a        | S  |
|        | IPF 1 (j)  | 226,5 b    | 218,3 a   | 225,2 b | 215,5 a | 218,1 b | 214,5 b       | 216,8 b       | S  |
|        | IP1R1 (j)  | 316,5 b    | 308,3 a   | 315,2 b | 305,5 a | 308,1 a | 304, 5 a      | 306, 8 a      | S  |
|        | Nm         | 8,3        | 8,4       | 8,3     | 8,5     | 8,5     | 8,5           | 8,5           | NS |
|        | M (Kg)     | 17,4 b     | 19,8 a    | 17,5 b  | 21,2 a  | 21 a    | 21,4 a        | 21,2 a        | S  |
|        | Pr (%)     | 60 c       | 91,4 a    | 63 c    | 86 b    | 86 b    | 86 b          | 86 b          | S  |
|        | Rb (t)     | 20,50 b    | 36,40 a   | 21,70 b | 37,00 a | 36,40 a | 37,50 a       | 36,90 a       | S  |
| 2      | H (cm)     | 218,5 b    | 233,1 ab  | 214,3 b | 257,1 a | 254,9 a | 259,7 a       | 255,6 a       | S  |
|        | C (cm)     | 44,1 c     | 50,2 b    | 45,2 c  | 62,3 a  | 61,6 a  | 62,7 a        | 62 a          | S  |
|        | IR1F2 (j)  | 116,2 b    | 107,4 ab  | 114,3 b | 100,7 a | 101,6 a | 100,4 a       | 101,8 a       | S  |
|        | IR1R2 (j)  | 206,2 b    | 197,4 b   | 204,2 b | 186,3 a | 187,6 a | 185,6 a       | 186,8 a       | S  |
|        | Nm         | 8,1        | 8,4       | 8,1     | 8,8     | 8,6     | 8,8           | 8,8           | NS |
|        | M (Kg)     | 15,04 c    | 18,1 b    | 15,6 c  | 22,5 a  | 22,1 a  | 22,9 a        | 22,6 a        | S  |
|        | Pr (%)     | 63 c       | 88,6 a    | 63 c    | 83 b    | 83 b    | 83 b          | 83 b          | S  |
|        | Rb (t)     | 28,5 c     | 50,4 b    | 29,8 c  | 62,2 a  | 60,6 a  | 63,5 a        | 62,3 a        | S  |
| 3      | H (cm)     | 257,4 c    | 303,6 b   | 261,4 c | 322,2 a | 319 a   | 323,1 a       | 322 a         | S  |
|        | C (cm)     | 43,9 c     | 50 b      | 44,8 c  | 64,8 a  | 63,6 a  | 65,7 a        | 63,8 a        | S  |
|        | IR2F3 (j)  | 187,3 b    | 185 b     | 184,3 b | 166,9 a | 168,3 a | 164,4 a       | 167,7 a       | S  |
|        | IR2R3 (j)  | 277,3 b    | 275 b     | 274,3 b | 250,9 a | 253,3 a | 249,4 a       | 252,7 a       | S  |
|        | Nm         | 7,9 b      | 8 b       | 8 b     | 9,6 a   | 9,4 a   | 9,7 a         | 9,6 a         | S  |
|        | M (Kg)     | 14,1 c     | 17,1 b    | 14,6 c  | 24,6 ab | 22,2 b  | 26,9 a        | 24,9 ab       | S  |
|        | Pr (%)     | 43 d       | 86 a      | 48 c    | 83 b    | 83 b    | 83 b          | 83 b          | S  |
|        | Rb (t)     | 13,6 d     | 33,2 c    | 15,8 d  | 50,5 ab | 45,1 b  | 55,5 a        | 50,7 ab       | S  |

H: height; C: circumference; IPF1: planting-flowering interval 1; IPR1: planting-harvest interval 1; IR1F2: harvest 1-flowering 2 interval; IR1R2: harvest 1-harvest 2 interval; IR2F3: harvest 2-flowering 3 interval; IR2R3: harvest 2-harvest 3 interval; Nm: number of hands; M: mass; Pr: percentage of bunch harvested; Rb: gross annual yield; S: significant; NS: not significant. Data are averages of five replicates

### **Influence of lixivate and Bioact on vegetative parameters and production**

Plant growth and productivity in the control and Bioact-treated plots were significantly lower than in the other treatments during all three growing cycles and flowering was late. Many plants fell (approximately 37% in the first and second cycles and approximately 45% of plant harvested in the third cycle) between flowering and harvest, and the average mass of harvested bunches was low, resulting in low yields. In the third cycle, the yield was three times lower, for the control (13.6 t) and Bioact (15.8 t) against approximately 50 t for the lixivate treatments. The chemical nematicide treatment of the plots resulted in an increase in the number of harvested plants (approximately 90%), however their productivity was significantly lower than that of the lixivate-treated plots. Indeed, the application of lixivate resulted in an early flowering and an increase in the mass of harvested bunches.

The present study results show an improvement in production in the presence of lixivate. Of all the lixivate treatments, productivity was better in the plantain lixivate plots especially when combined with Bioact. This result shows a more pronounced fertilizing activity of the plantain lixivate.

The low yields observed over the three cycles in the control and Bioact plots can be explained by the high parasite pressure in these plots. Indeed, the growth and yield of a banana plant depend on the health of its roots (Sen, 2005). Our results corroborate those of Mukasa *et al.*, (2006), who observed a 40% decrease in above-ground biomass and yield on banana after nematode attack. Chemical treatment provided better nematicide treatment compared to lixivate treatment, however yield was significantly better for lixivate treatments. This result suggests a tolerance mechanism in the plots that received the lixivate. Nutrients in quality and quantity in

the lixivate, especially potassium and nitrogen, compensated for the damage inflicted on the plant by nematodes by improving soil fertility. Such effects of organic matter were observed with McIntyre *et al.*, (2000). In addition, other studies have shown that the input of lixivate to the soil positively influenced microbial activity (Muñoz, 2005; Bautista *et al.*, 2015) including endomycorrhizae which are potential agents of plant tolerance to nematodes (Elsen *et al.*, 2003). The more pronounced fertilizing activity of plantain lixivate could be explained by its higher nitrogen content. This observation was also highlighted in the work of Herrera (2018) who advocates the use of plantain lixivate for better fertilization. Based on the production criteria (level of productivity and nematode inoculum present) in the 3rd cycle, increasing the number of production cycles was only feasible for the lixivate treated plots. Following of the control and Bioact plots was imminent due to the low production level and high nematode inoculum. The same was true for the Fluopyram-treated plots, where the cost of repeated application was becoming prohibitive for such declining productivity.

In the present study, plantain and dessert lixivate showed fertilizing and nematicidal effects. These properties were more optimized with the plantain lixivate especially when combined with Bioact. Bioact alone had no effect on the evolution of the nematode population, but seemed to improve soil enrichment. In a context of reduced fallow frequency, the application of lixivate preferentially from plantain could contribute to increasing the number of crop cycles in intensive dessert banana cv Gandenaine monoculture.

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