

Effect of the Aqueous Extract of Seeds of *Casimiroa Edulis* and Certain Drug Combinations on Cardiac Contractility, Phosphorilase_a and Adenyl Cyclase Activation in Isolated Perfused Mammalian Heart

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Abstract: *Objective:* To examine the hypotensive effect of the aqueous extract of the seeds of the plant *casimiroa edulis* on mammalian heart and vasculature. *Materials and Methods:* Studies were done on cardiac contractility, phosphorilase and adenyl cyclase in male Dunkin Hartley guinea-pigs weighing between 300-400 g. on perfused heart by the Langendorff technique. *Results:* Positive inotropic and chronotropic effects were observed mediated by cyclic 3', 5' - AMP, which were not abolished by beta adrenergic-blocking agents. These results further indicate that *casimiroa edulis* has the capacity to activate adenyl cyclase in particulate fractions of guinea-pig heart. *Conclusion:* These results suggest that there are separate adenyl cyclase systems for *casimiroa edulis* and nor epinephrine. However, definite proof would require physical separation of the two enzymes. The simultaneous use of H₁-and H₂-receptor agonists and antagonists thus provides a means for effective and selective inhibition of the cardiovascular actions of *casimiroa edulis*.

Keywords: *Casimiroa edulis*, Adenyl cyclase, Mammalian heart

1. Introduction

Plants are described as the major source of medicines, not only as isolated active principle to be dispensed in standardized dosage forms but also as crude drugs [1]. Cardiovascular ailments particularly hypertension is a very widespread condition which is not strictly considered as an illness but if not countered, progressively causes damage to all tissues and loss in their functionality. For this reason the find of new antihypertensive agents is prominent and medicinal plants and their derivatives are valuable for the purpose.

Mode of action of chemical constituents of plants which are having the known effects on hemodynamic and other cardiac parameters is now established. Effects of leaf extract from *Clerodendron Colebrookianum* have been found to decrease blood pressure in dose-dependent manner [2]. Effects of extract from *Cleroden trichotomum* on blood pressure and renal function have been studied by Lu Gw et al [3]. Intravenous injections of gambirine have shown a dose related fall in both systolic and diastolic blood pressure as well as heart rate [4]. Intravenous injections of the active component of the extract of *Salviae miltiorrhizae radix* (Den-Shen) have shown decreased blood pressure in a dose-dependant manner in rats [5]. Yang et al [6], studied the cardiovascular effects of Dehydro evodiamine, an alkaloid isolated from *Evodia ruteacarpajussieu* both *in vivo* & *vitro* experiments. Pharmacodynamic studies on aqueous extract of the root of *Polypodium vulgare* have shown a positive inotropic and chronotropic effect on perfused frog heart by causing hypotension and tachycardia in anesthetized dogs [7].

In the present study we have tried to investigate the

cardiovascular profile of the aqueous extracts of the seeds of *Casimiroa-edulis* Lia Llave et Lex, the "Zapote blanco" (white Zapote) a tropical plant belongs to a species of tropical fruiting tree in the family Rutaceae, native to eastern Mexico and Central America used by Mexican herb medicine since remote times [8]. Regional Mexican studies on the pharmacological effects of the seed's aqueous extract have shown the hypotensive quality of the plant extract [9, 10].

This pioneer phytochemical work was later followed by studies on *Casimiroa edulis* [11, 12, 13, 14, 15, 16, 17, 18] trying to corroborate the hypnotic properties attributed to this sweet fruit.

Lozoya et al [19], Ortega et al [20], have confirmed the vigorous hypotensive effect produced by the aqueous and alcoholic extract from *Casimiroa edulis* seeds on cats, rabbits and dogs, together with the constrictor effect produced on the uterus by *in vitro* experiments on several animal species including human.

From the literature cited above, it is now clear that the seeds and leaves of the plant *Casimiroa edulis* have been used in Folk medicine as an hypotonic and sedative and more recently as an antihypertensive. Effects of aqueous extract of *casimiroa edulis* (Rutacea) on blood pressure and heart rate in albino rats has been reported by Garcia et al [21]. These studies demonstrated a rapid and transitory increase in blood pressure. The amplitude of the blood pressure rise was dose dependent.

Endothelium-dependent vasorelaxing activity of aqueous extracts of lyophilized seeds of *Casimiroa edulis* on rat

mesenteric arterial bed has been reported in the literature by Baisch et al [22].

Phytochemical studies have shown that the leaf essential oil of *Casimiroa edulis* was dominated by sesquiterpene hydrocarbons, predominately germacrene D (16-22%) and (E)-caryophyllene (16-17%), consistent with the traditional use of this plant as a sedative, sleep inducer and hypotensive [23].

The hexanic and methanolic extracts of *Casimiroa edulis* showed vasorelaxation in arterial tissues in rat precontracted by phenylephrine (0.5 μ M); the extracts from seeds always caused a greater relaxation in comparison to those from leaves [24].

Recently age-dependent vasorelaxation of *Casimiroa edulis* and *Casimiroa pubescens* extracts in rat caudal artery in vitro has been reported, thus supporting the traditional use of *Casimiroa* decoctions as antihypertensive, also in elderly [25].

More recently Bertin et al [26] have reported that the phenolic compounds isolated from *Casimiroa* have shown vasorelaxation on rat arterial tissues although with different effectiveness thus suggesting its possible role against hypertension and vasculopathies.

Further more phenolic compounds from leaves of *Casimiroa edulis* showed adipogenesis activity as well [27].

In more than fifty years of active research around 20 different substances were obtained from the seeds, leaves and barks of *Casimiroa edulis* [28] and some of their structures were elucidated but the hypogenic active principle was never established and/or published.

As a first step in the characterization of the hypotensive activity of *Casimiroa edulis*, the present study has been carried out using animal model. Included in the study is the determination of the effects of the aqueous seed extract of *casimiroa edulis*, the part of the plant reported to be more active [29, 17, 18] on cardiac contractility, phosphorilase and adenylyl cyclase in male Dunkin Hartley guinea-pigs which may be helpful for the identification of active compounds of the fraction of the crude extract for further pharmacological evaluation to determine its influence in diverse hemodynamic parameters.

2. Materials and Methods

Preparation of *casimiroa edulis* extract

The aqueous extract of *casimiroa edulis* seeds was prepared as described previously [17]. Briefly ripe fruits of *casimiroa edulis* were obtained from local markets and after being defatted, the dry powdered kernels were extracted successfully by maceration at room temperature with hexane, dichloromethane, 4:1 and 1:1 mixtures of dichloromethane-methanol, methanol and finally water. Organic solvents were eliminated in a rotatory evaporator; water was removed by lyophilization. For the pharmacological tests, the solid aqueous extract thus obtained (approximately yield: 5.5%) was dissolved in isotonic

NaCl solution to a concentration of 100

Effects of the aqueous seed extract of *casimiroa edulis* were studied on cardiac contractility, phosphorilase and adenylyl cyclase in male Dunkin Hartley guinea-pigs weighing between 300-400 g. Animals were injected with heparin sodium 8mg/kg, s.c.) 60 minutes prior to sacrifice. The animals were killed and the heart was rapidly removed and perfused by the Langendorff technique with chenoweth-koelle solution by the established method [30], modified by Bell et al [31] at a flow rate of 3.6 ml/min. The flow rate was maintained by a peristaltic pump. The perfusion solution was equilibrated with a 95% O₂-5% CO₂ gas mixture at 37 C⁰. Contractility was monitored by means of a Palmer clip placed in the apex of the heart and connected to a Grass force displacement transducer and recorded on a Grass model 7 polygraph. Diastolic tension was adjusted to 5 g. The hearts were allowed to equilibrate for 25 minutes. At this time two dose-response curves to *casimiroa edulis* were obtained by injecting the extract via a sidearm cannula. The hearts were then perfused with chenoweth-koelle buffer solution containing theophylline (10-3M). Following a 5 minute equilibration period the extract dose-response curves were repeated. The dose-response curves thus obtained were averaged and plotted as a percentage of the maximum response that could be obtained with *casimiroa edulis*.

For the phosphorilase experiments the hearts were perfused as noted above with either buffer or buffer plus 10-3M-theophylline. Extract was then injected via the sidearm cannula and the heart was frozen at the peak of the contractile response by means of a pair of Wollenberger tongs [32] previously chilled in a mixture of alcohol-dry ice. An 80 to 100 mg portion of the apex was cut away and phosphorylase activity measured in the direction of glycogen synthesis as previously described [33].

Since total enzyme activity did not change, all results are expressed as % phosphorylase a which is: [(enzyme activity without AMP)/ (enzyme activity with AMP)] X 100.

For adenylyl cyclase experiments a washed particulate preparation of guinea pig heart was used as the source of the enzyme. Enzyme activity was determined by measuring the conversion of [14C]-ATP to [14C]-cyclic AMP as previously described [34]. The enzyme was prepared and activity measured on the same day. Enzyme activity is expressed as pmol cyclic AMP produced/4/min/mg protein. Protein was determined by the established method [35, 36].

Drugs and chemicals used were, 1-norepinephrine bitartrate (Winthrop Lab.; theophylline (Merck and Co.); heparin sodium (Nutritional Biochemicals Corp.); phenoxybenzamine (SK&F); tolazoline HCl (Ciba Pharm. Co.); sodium flouride (Mallinckrodt); diphenhydramine HCl (Parke Davis and Co.); tripeleminamine HCl (Ciba Pharm. Co.); propranolol (Ayerst Lab. Inc.); 1-epinephrine bitartrate (Winthrop Lab.); imidazole (General Biochemicals); 1-histidine (sigma Chem. Co.); betazole HCl and 3-(B-aminoethyl)-1,2,4-triazole dihydrochloride (Eli Lilly and Co.).

3. Results

As shown in Figure.1, *casimiroa edulis* increased the contractility of the isolated perfused guinea pig heart. The

positive inotropic effect of extract was significantly enhanced ($P < 0.05$) by theophylline in a concentration of either 10^{-4} or 10^{-3} M. *Casimiroa edulis* also increased the activity of phosphorylase_a in the heart (Table.1). Enzyme activation by the *casimiroa edulis* was also enhanced by theophylline.

Casimiroa edulis also proved capable of stimulating cardiac adenylyl cyclase as is shown in Figure 2, whereas epinephrine approximately doubled the enzyme activity.

When injected into the perfused guinea pig heart *casimiroa edulis* (0.8Ug) increased the force of contraction. Tripeleennamine (3×10^{-6} M) did not decrease the *casimiroa edulis* effect. The higher concentration of tripeleennamine (10^{-4} M) decreased the extract response. However, at this concentration, tripeleennamine was cardiotoxic and could only be perfused through the heart for 4 minute. Longer persuasion times result in cardiac arrest. (Table 2). The effect of the antihistamine tripeleennamine on *casimiroa edulis* induced activation of cardiac phosphorylase_a is demonstrated in Table 3. Again tripeleennamine (3×10^{-6} M) did not reduce the response while tripeleennamine (10^{-4} M) significantly reduced the activation of the enzyme. Blockade of the extract response by the higher concentration of tripeleennamine appears to be a relatively non-specific phenomenon since the activation of the enzyme by isoproterenol was also blocked by tripeleennamine (10^{-4} M).

All remaining experiments were concerned with the study of various drugs on cardiac adenylyl cyclase in an attempt to characterize the active site for *casimiroa edulis* and to compare these effects with those obtained in the whole heart. Two antihistamines, diphenhydramine and tripeleennamine, were used as potential antagonists of the extract. Diphenhydramine (Table.4) only slightly decreased the stimulation of adenylyl cyclase produced by 10^{-6} M extract when tested at a concentration of 10^{-6} M. Total blockade of the extract response was achieved with a concentration of 10^{-4} M diphenhydramine. Diphenhydramine (10^{-5} M) lowered the maximum response to the extract stimulation of adenylyl cyclase (Figure.3). Similar results were obtained with tripeleennamine except that there was absolutely no decrease in the extract response with 10^{-6} M tripeleennamine (Table.5). The apparent non-specificity of the blockade with these drugs at the higher concentration (10^{-4} M) is demonstrated in the experiments with propranolol (Table. 6). Propranolol produced a decrease in the *casimiroa edulis* response similar to that obtained with the antihistamines. Propranolol, however, proved very capable of specifically blocking the epinephrine stimulation of cardiac adenylyl cyclase (Table.7). In this experiment propranolol (10^{-5} M) blocked the stimulation produced by epinephrine (10^{-4} M) 90% whereas tripeleennamine (10^{-5} M) did not decrease the epinephrine response at all. The alpha--adrenergic blocking agents phenoxy-benzamine and tolazoline also did not decrease the *casimiroa edulis* response when used in a concentration of 10^{-6} M indicating that alpha-receptors are also not involved. None of the blocking agents tested in concentrations of 10^{-5} M affected the stimulation of the enzyme produced by sodium fluoride (10^{-2} M) (Table.8).

4. Discussion

The data presented demonstrate that, in the guinea-pig

Casimiroa edulis has a positive inotropic effect and can increase the activity of cardiac phosphorylase *a* in intact hearts. *Casimiroa edulis* can also increase the activity of cardiac adenylyl cyclase in a washed particulate preparation from guinea-pig. Both the inotropic effect and the phosphorylase activating effect of extract were greatly enhanced by the theophylline. While these data do not prove a cause and effect relationship they are at least suggestive that *Casimiroa edulis* produces its effects on the heart by stimulating adenylyl cyclase thus increasing the intracellular concentration of cyclic AMP as has been suggested by Lein et al [37].

The antihistamine tripeleennamine did not prove to be an effective antagonist of either the mechanical or biochemical effects of *Casimiroa edulis* on the heart. An extremely high concentration (10^{-4} M) of antihistamine was effective in significantly decreasing the inotropic or phosphorylase activating effect of the extract. At this concentration tripeleennamine was cardiotoxic and also blocked the effect of isoproterenol on the heart (Tables 5 and 6). The results support the work of Hattori et al [38] who suggested that the cardiac histamine receptor differed from other histamine receptors since it could not be readily blocked by antihistamines. Similar effects were noted when drug interactions were studied on cardiac adenylyl cyclase. Low doses of antihistamines (10^{-6} M) or propranolol (10^{-6} M) had little or no effect on *casimiroa edulis* extract induced histamine-like-activation of the enzyme. With both the antihistamines and the B-blocker, however, higher concentrations (10^{-4} M) completely blocked the extract response. This latter concentration is cardiotoxic in intact hearts and again demonstrates the non-specificity of the blockade. Similar results have been reported in isolated heart preparations [39]. Phenoxybenzamine and tolazoline, both alpha-adrenergic blocking agents, also failed to decrease the *Casimiroa edulis* response. The blocking agents did not appear to affect stimulation of the enzyme by sodium fluoride (Table-8). This would suggest that their actions are confined to the regulatory subunit since fluoride is believed to activate the catalytic site only [40].

In the present study propranolol was found to be a selective inhibitor of the epinephrine stimulation of cardiac adenylyl cyclase whereas the antihistamine antagonism appeared to be less specific. These data suggest that *Casimiroa edulis* and epinephrine can stimulate adenylyl cyclase by interacting with separate receptors. Further experiments tended to support this hypothesis. Epinephrine and *Casimiroa edulis* when combined together at maximal stimulatory concentrations tended to activate adenylyl cyclase to a slightly greater extent than did extract alone. One possible explanation for this discrepancy is that greater stimulation due to extract alone was found in the present study than was reported by Klein and Levey [41]. Therefore it is possible that a near maximal stimulation due to an interaction of *Casimiroa edulis* with the regulatory subunit of the enzyme prevented a greater interaction when the two agonists were combined. Nevertheless the conclusions from the data are the same as those of previous workers [41] i.e. that separate and distinct receptors exist for the two agonists. This hypothesis was further strengthened by the experiments using the histamine analogs betazole and the triazole derivative. The combination of either of these compounds with epinephrine resulted in an increase in enzyme activity over that obtained with either drug alone. In addition betazole decreased the

extract response to a value intermediate between that obtained with the *Casimiroa edulis* and betazole indicating competition for the same site. In our experiments none of the cardiac effects of the extract were antagonized well by tripeleennamine. In the present study it was found that the diphenhydramine was not that specific in blocking the histamine like activation of adenyl cyclase produced by extract. While a concentration of 10^{-4} M did block the histamine response it also reduced epinephrine stimulation of the enzyme although to a lesser degree. Diphenhydramine (10^{-5} M) appeared to lower the maximum response to *Casimiroa edulis* more than shift the extract dose-response curve to the right. It was also demonstrated that at 10^{-4} M propranolol was capable of reducing the *Casimiroa edulis* response and further supports the hypothesis that the cardiac histamine receptor differs from other histamine receptors [42] and appears that there are at least three regulatory sites on the regulatory subunit of cardiac adenyl cyclase.

Activation of any one of the regulatory sites ultimately leads to activation of the catalytic site. It is postulated that activation of any of the regulatory sites must follow a final common pathway leading to an increase in catalytic site activity since combinations of norepinephrine and glucagon or histamine and epinephrine did not result in additive stimulation as reported previously [43].

Moura et al [44], however, were unable to demonstrate stimulation with thyroid hormone in enzyme obtained from rat heart. The tool to clarify the role of histaminergic effect of *casimiroa edulis* in the cardiovascular system are thus established to some extent and progress in this area can be anticipated with interest.

References

- [1] Aftab K, Ahmed SI, Usmanghari K. Toladitional medicine cassia Absus L (Chaksu) Pharmacological evaluation". *Phytomedicine*. Vol.2, pp.213-219, 1996.
- [2] Gupta M, Upal KM, Sanjib D. "Effect of leaf extract from clerodendron colebrookianum on blood pressure in rats". *Indian J. experimental Biol*. Vol. 32, pp. 216-217, 1994.
- [3] Lu GW, Miura K, Yukimura T, Yamamoto K. "Effect of extract from clerodendron trichotomum on blood pressure and renal function in rats and dogs". *J. Ethnopharmacol*. Vol. 42, pp. 77-82, 1994.
- [4] Mokisli CP, Lee KH, Kam TS, Goh SH. "Cardio vascular responses in the normotensive rat produced by intravenous injection of gambirine isolated from uncaria callophylla BL. Ex Korth." *J. Ethno Pharmacol*. Vol.36, pp. 219-223, 1992.
- [5] Katsuo K, Mariko N, Masabiro N. "Hypotensive effects of lithospermic Acid B-isolated from the extract of *Salviae miltiorrhizae Radix* in the rat". *Gen. Pharmacol*. Vol. 25, pp. 69-73, 1994.
- [6] Yang MC, WU SL, Kuo JS, Chen CF. "The hypotensive and negative chronotropic effects of dehydro evodiamine". *Eur. J. Pharmacol*. Vol. 17, pp. 537-542, 1990.
- [7] Mannan A, Khan RA, Asif M. "Pharmacodynamic Studies on *Polypodium Vulgare*". *Indian J. Exp Biol*. Vol. 27, pp. 556-560, 1989.
- [8] Power FB, Callan T. "The constituents of the seeds of *casimiroa edulis*". *Trans. Chem Soc. (London)* Vol. 99, pp. 183-193, 1911.
- [9] De Lille J. "Nota acerca de la accion de zapote blanco sobrela tension arterial". *Anales del Instituto de Biol*. Vol.5, pp. 45-47, 1934.
- [10] Mendez M. "Pharmacologic data of some Mexican remedies". *J Am Inst Homeopathy*. Vol. 30, pp. 273-285, 1936.
- [11] Kinkle FA, Romo J. "The constituents of *casimiroa-edulis* Llava et Lex. Part 1, The seed". *J Am Chem Soc*. Vol. 76, pp. 4163-4173, 1956.
- [12] Merzels A. "The constituents of *Casimiroa edulis* IV". *J Org Chem*. Vol. 23, pp. 762-764, 1958.
- [13] Djerassi CC. "Partial structure of *casimiroa edine*". *Int J Org Chem*. Vol. 2, pp. 168-170, 1958.
- [14] Sondheimer F. "The constituents of *Casimiroa Edulis-1*". *J Org Chem*. Vol. 23, pp. 762-764, 1958.
- [15] Lozoya X, Rodriguez D, Ortega J, Enriquez R. "Arch Invest Med". Vol. 9, pp, 565-573, 1978.
- [16] Lozoya X, Auglar A, Camacho JR. "Encuesta Sobrle el uso actual de plantas enla medicina tradicional maxicana. *Revista Medica del Instituto Mexicano del seguro social*". Vol. 25, pp. 283-291, 1987.
- [17] Gil AM, Horacio V. "Pharmacology of *Casimiroa edulis*; Part I. Blood Pressure and heart rate effects in anesthetized rat". *Planta. Med*. Vol. 57, pp. 20-24, 1991.
- [18] Gil AM, Horacio V. "Raul E: Pharmacology of *Casimiroa edulis*; III. Relaxation and contractile force in rat aortic ring". *J Ethnopharm*. Vol. 47, pp.1-8, 1995.
- [19] Lozoya X, Romero G, Olmedo M. "Farmacodinamia de los extractos alcoholicos YH acuosos de la semilla de *casimiroa edulis*". *Arch Invest. Med (Mex)*, Vol. 8, 145-152, 1977.
- [20] Ortega J, Lozoya X, Enriquez R. "Aislamiento del Principio Hipotensor de la semilla de *casimiroa edulis*". *Arch. Invest. Med*. Vol. 9, pp. 565-573, 1978.
- [21] García GM, Freer BE, Morales M O. "Effects of *Casimiroa edulis* (Rutacea) on blood pressure and heart rate in albino rats". *Rev Biol Trop*. Vol. 42, pp.115-119, 1994.
- [22] Baisch AL, Urban H, Ruiz AN. "Endothelium-dependent vasorelaxing activity of aqueous extracts of lyophilized seeds of *Casimiroa edulis* (AECe) on rat mesenteric arterial bed". *J Ethnopharmacol*. Vol. 95, pp. 163-7, 2004.
- [23] Miller SL, Haber WA, Setzer WN. "Chemical composition of the leaf essential oil of *Casimiroa edulis* La Llave & Lex. (Rutaceae) from Monteverde, Costa Rica". *Nat Prod Commun*. Vol. 4, pp. 425-426, 2009.
- [24] Froidi G, Bertin R, Secchi E, Zago G, Martínez-Vázquez M, García-Argaéz A. Vasorelaxation by extracts of *Casimiroa* spp. in rat resistance vessels and pharmacological study of cellular mechanisms". *J Ethnopharmacol*. Vol. 12, 134, pp. 637-43, 2011.
- [25] Bertin R, García-Argaéz A, Martínez-Vázquez M, Froidi G. "Age-dependent vasorelaxation of *Casimiroa edulis* and *Casimiroa pubescens* extracts in rat caudal artery in vitro". *J Ethnopharmacol*. Vol. 137, pp. 934-936, 2011.
- [26] Bertin R, Chen Z, Martínez-Vázquez M, García-Argaéz A, Froidi G. "Vasodilation and radical-scavenging activity of imperatorin and selected coumarinic and flavonoid compounds from genus *Casimiroa*". *Phytomedicine*. Vol. 21, pp. 586-94, 2014.
- [27] Nagai H, Tanaka T, Goto T, Kusudo T, Takahashi N, Kawada T. "Phenolic compounds from leaves of *Casimiroa edulis* showed adipogenesis activity". *Biosci*

- Biotechnol Biochem. Vol. 78, pp. 296-300, 2014.
- [28] Rizvi SH, Kapil RS, Shoeb A. "Alkaloids and Coumarins of *Casimiroa edulis*". J. Natul. Prod. Vol. 48, pp. 146, 1985.
- [29] Flores L. "Encuesta Sobre el uso actual de plantas en la medicina tradicional mexicana". Ann Inst Med Nac. Vol. 9, pp. 391-397, 1907.
- [30] Chenoweth MB, Koelle ES. "An isolated heart perfusion system adapted to the determination of non-gaseous metabolites". J Lab Clin Medicine. Vol. 31, pp. 600-608, 1946.
- [31] Bell RM, Mocanu MM, Yellon DM. "Retrograde heart perfusion: the Langendorff technique of isolated heart perfusion". J Mol Cell Cardiol. Vol.50, pp. 940-950, 2011.
- [32] Wollenberger A, Rista O, G. "Eine einfache technik der extremen schnellen Abkaltung grasserer Gewebstucke". Pfluegers Archiv fur die Gesamte Physiologie des Menschen und der Tiere. Vol. 270, pp. 399-412, 1960.
- [33] Laughlin MR, Petit WA, Jr, Shulman RG, Barrett EJ. "Measurement of myocardial glycogen synthesis in diabetic and fasted rats". Am J Physiol. Vol. 258, pp. E184-90, 1990.
- [34] Roth BL, Nakaki T, Chaung DM, Costa E. "5-hydroxytryptamine 2 receptors coupled to phospholipase α in rate aorta: modulation of Phosphoinositide turnover by phorbol esters". J Pharm Exper Therap. Vol. 238, pp. 480-485, 1996.
- [35] Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. "Protein measurement with the Folin Phenol reagent". J Biol Chem. Vol. 193, pp. 265-275, 1951.
- [36] Phosri S, Arieyawong A, Bunrukchai K, Parichatikanond W, Nishimura A, Nishida M, Mangmool S. "Stimulation of Adenosine A2B Receptor Inhibits Endothelin-1-Induced Cardiac Fibroblast Proliferation and α -Smooth MuscleActin Synthesis Through the cAMP/Epac/PI3K/Akt-Signaling Pathway". Front Pharmacol. Vol. 30, pp. 428, 2017.
- [37] Lein CA, Belmont MR, Abalos A, Eppich L. "The cardio vascular effects and histamine releasing properties of 51W89 in patients receiving anesthesia". Anesthesiology. Vol. 82, pp. 1131-1138, 1995.
- [38] Hattori Y, Hattori K, Matsuda N. "Regulation of the Cardiovascular System by Histamine". Hand book Exp Pharmacol. Vol. 241, pp. 239-258, 2017.
- [39] Kitakaze M. "Clinical Evidence of the Role of Histamine in Heart Failure". J Am Coll Cardiol. Vol. 67, pp. 1553-1555, 2016.
- [40] Fujiwara K, Kitagawa T, Alomso G. "Monoclonal antibodies against glutaraldehyde conjugated histamine. Application to immunocytochemistry". Histochem Cell Biol. Vol. 107, pp. 39-46, 1997.
- [41] Klien I, Levey GS. "Activation of myocardial adenyl cyclase by histamine in guinea pig, cat and human heart". J Clin Invest. Vol. 50, pp. 1012-1015, 1971.
- [42] Seroky JT, Alper CM, Tabari R, Doyle WJ. "Effects of intranasal challenge with histamine, bradykinin and prostaglandin on middle ear pressure and blood flow in cynomolgus monkeys". Acta Otolaryngol (stock). Vol. 115, pp. 83-87, 1995.
- [43] Kuznetsova LA, Plesneva SA, Sharova TS, Pertseva MN, Shpakov AO. "Regulation of adenylcyclase signaling system by insulin, biogenic amines, and glucagon at their separate and combined action in the muscle membranes of the mollusc *Anodonta cygnea*". Zh Evol Biokhim Fiziol. Vol. 49, pp. 111-117.2013.
- [44] Moura AL, Hyslop S, Grassi-Kassisse DM, Spadari RC. "Functional β_2 -adrenoceptors in rat left atria: effect of foot-shock stress". Can J Physiol Pharmacol. Vol. 95, pp. 999-1008. 2017.

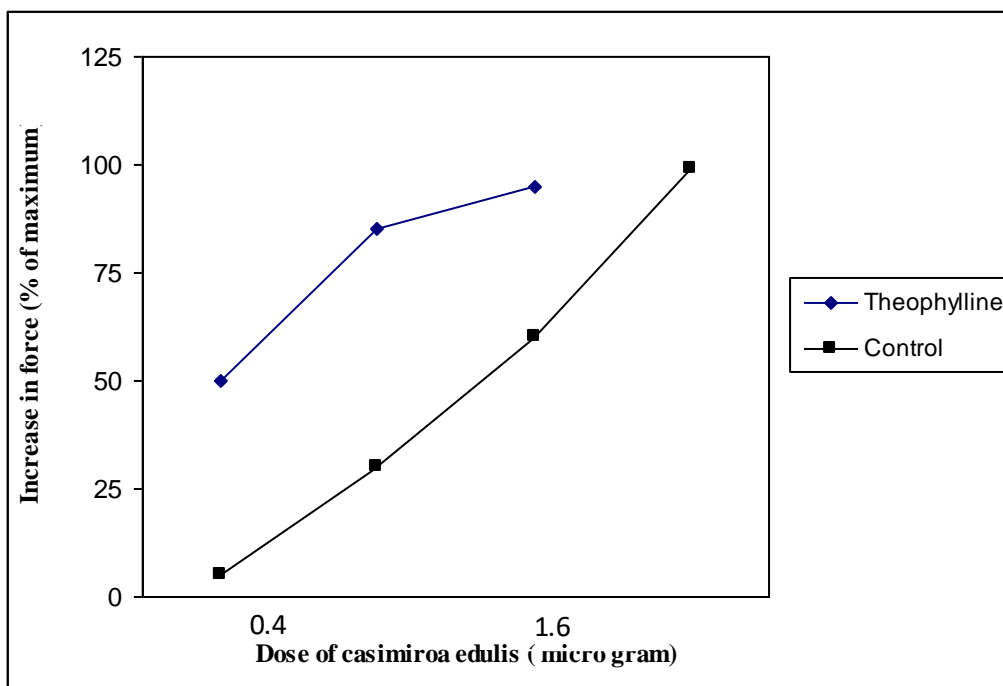


Figure 1A: Effect of casimiroa edulis on cardiac contractile force in the presence and absence of theophylline 10^{-3} M in guinea-pig hearts ($n= 50$). The increase in the force is expressed as a percentage of the maximum response obtained with casimiroa edulis. n represents sample number. The standard errors, in all cases were less than the area covered by the circle.

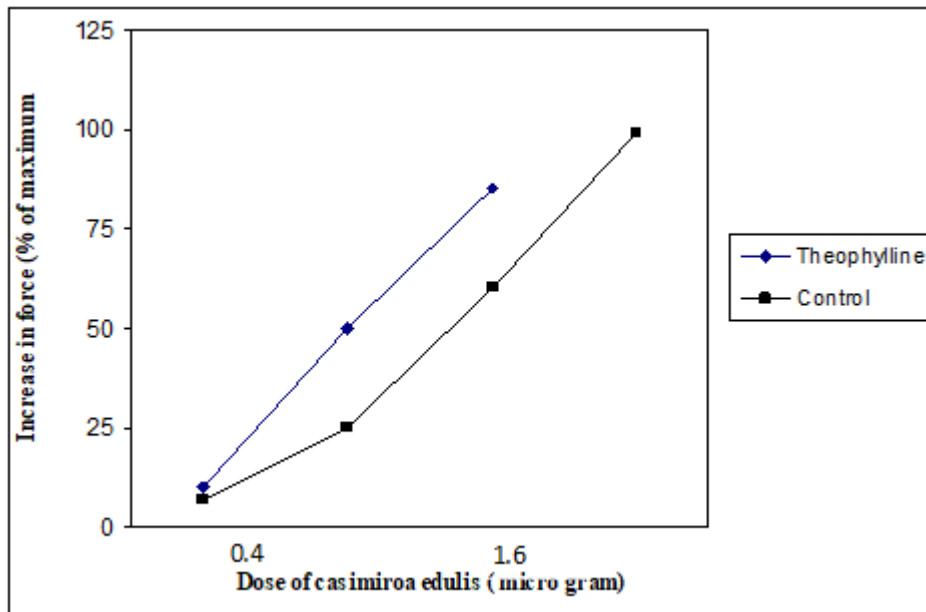


Figure 1B: Effect of casimiroa edulis on cardiac contractile force in the presence and absence of theophylline 10^{-4} M in guinea-pig hearts ($n=50$). The increase in the force is expressed as a percentage of the maximum response obtained with casimiroa edulis. n represents sample number. The standard errors, in all cases were less than the area covered by the circle.

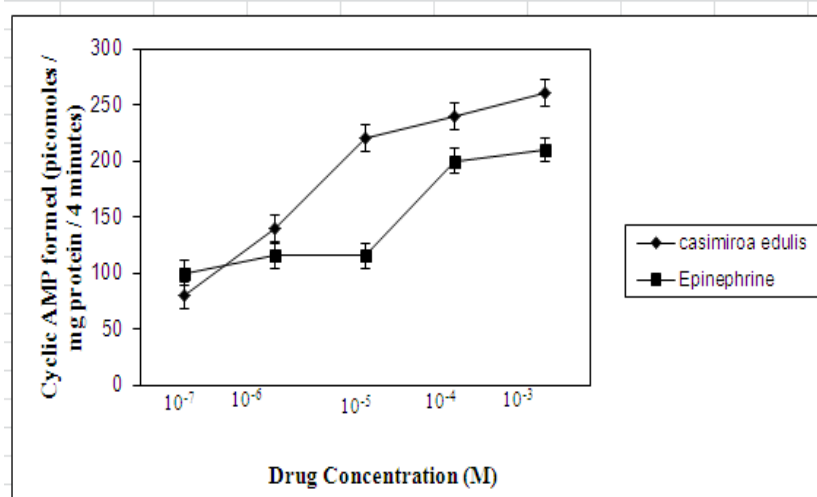


Figure 2: Dose response curves demonstrating the stimulation of guinea-pig heart adenylyl cyclase by casimiroa edulis and epinephrine. ($n=50$). n represents sample number. Vertical bars represent the SE.

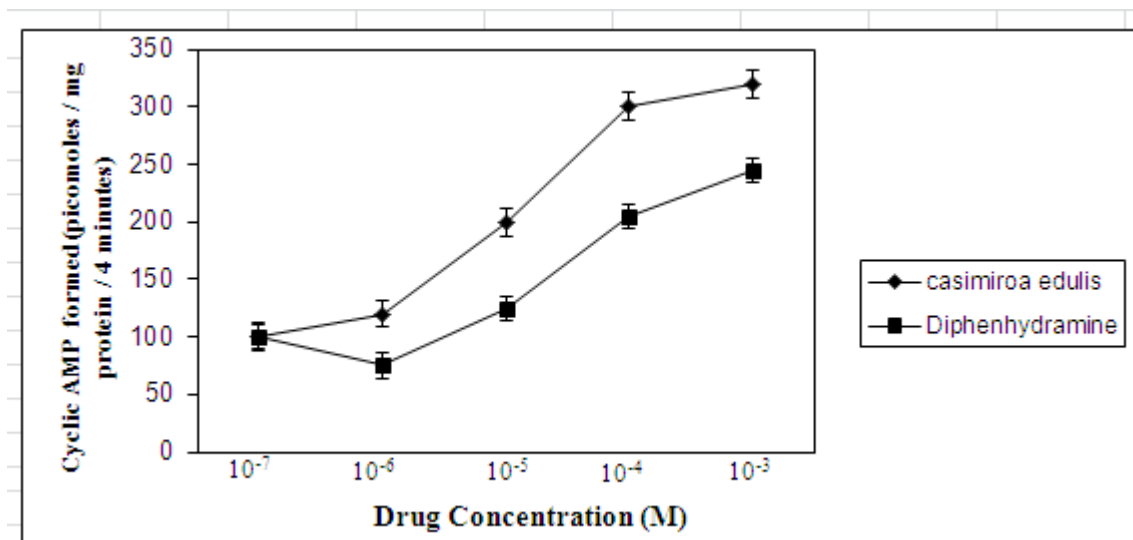


Figure 3: Dose response curves demonstrating the interaction between diphenhydramine (10^{-5} M) and casimiroa edulis on

cardiac adenylyl cyclase in guinea-pig heart. ($n = 50$). n represents sample number. Vertical bars represent the SE.

Table 1: Effect of casimiroa edulis and theophylline on cardiac phosphorilase_a in guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples

Dose of casimiroa edulis (μg)	% Phosphorilase _a \pm SE	
	Buffer perfused	Buffer perfused Plus Theophylline (10^{-3} M)
0	2.6 \pm 1.3	9.7 \pm 2.8 *
0.1	2.9 \pm 1.4	26.3 \pm 5.9 *
0.2	5.8 \pm 1.4	45.8 \pm 2.5 *
0.4	10.5 \pm 1.5 **	44.8 \pm 3.9 *
0.8	27.1 \pm 4.8 **	-
1.6	32.1 \pm 4.0 **	-

* Significantly greater than casimiroa edulis ($p < 0.05$)

** Significantly greater than no drug ($p < 0.5$)

Table 2: Effect of tripeleennamine on casimiroa edulis-induced contractility in the isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples

Treatment	Increase in force (g) obtained with 0.8 μg casimiroa edulis
None	4.5 \pm 0.4
Tripeleennamine (3×10^{-6} M)*	3.5 \pm 0.4
Tripeleennamine (10^{-4} M)**	1.8 \pm 0.3***

*Perfused through the heart for 10 minutes.

** Perfused through the heart for 4 minutes.

*** Significantly less than no drug ($p < 0.05$)

Table 3: Effect of tripeleennamine on drug-induced increase in cardiac phosphorilase_a in the isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples.

Treatment	Phosphorilase _a
Saline	1.0 \pm 1.1
Tripeleennamine 10^{-4} M	3.9 \pm 1.4
Casimiroa edulis 0.8 μg	29.1 \pm 3.6
Casimiroa edulis 0.8 μg + Tripeleennamine 10^{-4} M	4.8 \pm 1.1*
Casimiroa edulis 0.8 μg + Tripeleennamine 3×10^{-6} M	30.5 \pm 3.6*
Isoproterenol 0.2 μg	31.7 \pm 3.3
Isoproterenol 0.2 μg + Tripeleennamine 10^{-4} M	0.0 \pm 0.0*

*Significantly less than casimiroa edulis or isoproterenol alone. Tripeleennamine perfused through the heart.

Table 4: Effect of casimiroa edulis and diphenhydramine on adenylyl cyclase in isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples

Treatment	Experiment No. 1	Experiment No. 2	Diphenhydramine (10^{-4} M) Mean Activity
	Diphenhydramine (10^{-6} M) Mean Activity	Diphenhydramine (10^{-6} M) Mean Activity	
Control	181 \pm 5.0	102 \pm 1.2	125 \pm 2.0
Casimiroa edulis (10^{-6} M)	235 \pm 6.0	150 \pm 3.6	156 \pm 3.2
Diphenhydramine	157 \pm 2.0	105 \pm 2.3	132 \pm 8.2
Casimiroa edulis (10^{-6} M) + Diphenhydramine	191 \pm 2.0	141 \pm 4.1	113 \pm 1.2

Adenylyl cyclase activity expressed as picomoles of cyclic AMP formed / mg protein / 4.0 minutes.

Table 5: Effect of casimiroa edulis and tripeleennamine on adenylyl cyclase in isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples.

Treatment	Experiment No. 1	Experiment No. 2	Tripeleennamine (10^{-4} M) Mean Activity
	Tripeleennamine (10^{-6} M) Mean Activity	Tripeleennamine (10^{-6} M) Mean Activity	
Control	181 \pm 5.0	102 \pm 1.2	125 \pm 2.0
Casimiroa edulis (10^{-6} M)	235 \pm 6.0	150 \pm 3.6	156 \pm 3.2
Tripeleennamine	151 \pm 5.0	106 \pm 2.2	110 \pm 4.1
Casimiroa edulis (10^{-6} M) + Tripeleennamine	208 \pm 2.8	154 \pm 7.1	99 \pm 1.3

Adenylyl cyclase activity expressed as picomoles of cyclic AMP formed / mg protein / 4.0 minutes.

Table 6: Effect of casimiroa edulis and propranolol on adenylyl cyclase in isolated guinea-pig heart
 Values are Mean \pm SE, ($n = 50$); n represents total number of samples.

Treatment	Experiment No. 1	Experiment No. 2	Propranolol (10^{-4} M) Mean Activity
	Propranolol (10^{-6} M) Mean Activity	Propranolol (10^{-6}) Mean Activity	
Control	181 \pm 5.0	102 \pm 1.2	125 \pm 2.0
Casimiroa edulis (10^{-6} M)	235 \pm 6.0	150 \pm 3.6	156 \pm 3.2
Propranolol	118 \pm 3.0	100 \pm 5.2	128 \pm 4.1
Casimiroa edulis (10^{-6} M)+Propranolol	163 \pm 9.8	163 \pm 3.1	129 \pm 7.1

Adenylyl cyclase activity expressed as picomoles of cyclic AMP formed / mg protein / 4.0 minutes.

Table 7: Effect of epinephrine, propranolol and tripeleennamine on adenylyl cyclase in isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples

Treatment	Mean Activity
Control	111 \pm 5.0
Epinephrine (10^{-4} M)	246 \pm 9.1
Propranolol (10^{-5} M)	79 \pm 1.0
Tripeleennamine(10^{-5} M)	77 \pm 7.2
Epinephrine (10^{-4} M) + Tripeleennamine(10^{-5} M)	231 \pm 8.0
Epinephrine (10^{-4} M) + Propranolol (10^{-5} M)	93 \pm 6.2

Adenylyl cyclase activity expressed as picomoles of cyclic AMP formed / mg protein / 4.0 minutes.

Table 8: Effect of blocking agents on sodium fluoride (NaF) stimulation of adenylyl cyclase in isolated guinea-pig heart. Values are Mean \pm SE, ($n = 50$); n represents total number of samples

Treatment	Mean Activity
None	142 \pm 7.0
NaF (10^{-2} M)	723 \pm 11
Tripeleennamine(10^{-5} M) + NaF	715 \pm 19
Diphenhydramine (10^{-5} M) + NaF	740 \pm 15
Propranolol (10^{-5} M) + NaF	754 \pm 18

Adenylyl cyclase activity expressed as picomoles of cyclic AMP formed / mg protein / 4.0 minutes.