Feeding habits of juvenile surface-migratory myctophid fishes (family Myctophidae) in the Kuroshio region of the western North Pacific

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ABSTRACT: We examined the diets and nighttime feeding chronologies of the most abundant juvenile surface-migratory myctophid fishes in the Kuroshio waters of the western North Pacific: Myctophum asperum, M. nitidulum, Symbolophorus evermanni and Centrobranchus brevirostris. Samples were collected from water 0 to 1 m deep at night from January to March during 10 vr between 1961 and 1992. The 4 species exhibited resource partitioning by having different main prey items at night in the top 1 m layer. M. asperum fed mainly on appendicularians, M. nitidulum on copepods and amphipods, S. evermanni on euphausiids and C. brevirostris on pteropods. They also had speciesspecific peak feeding hours, demonstrated by the different hours during which each species had the greatest wet weight of stomach contents (percentage of body weight): 24:00 to 01:00 h (5.1%) and 02:00 to 03:00 h (4.0%) for M. asperum; 03:00 to 04:00 h (7.3%) for M. nitidulum; 04:00 to 05:00 h (3.9%) for S. evermanni; and 20:00 to 21:00 h, 22:00 to 23:00 h and 04:00 to 05:00 h (4.6, 5.3 and 3.8%, respectively) for C. brevirostris. These results suggest that myctophids that migrate to the surface reduce trophic competition by specializing in different food organisms. We observed 2 and 3 feeding peaks for species that feed on gelatinous plankton (M. asperum and C. brevirostris, respectively), compared with 1 peak for crustacean feeders (S. evermanni and M. nitidulum). We also estimated the impact on zooplankton of nocturnal feeding by these 4 species.

KEY WORDS: Myctophid fish \cdot Diel vertical migration \cdot Surface migrant \cdot Feeding habit \cdot Feeding chronology \cdot Resource partitioning \cdot Feeding impact \cdot Kuroshio region

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INTRODUCTION

Surface migratory myctophid fishes that migrate at night from the mesopelagic layer up to the surface are commonly collected there in neuston nets. A 5 to 10 min tow often yields several species (Ogawa 1961, Hattori 1964, Kawaguchi et al. 1972, Watanabe & Kawaguchi 1999). These migrants to surface waters are closely related species and genera of the subfamily Myctophinae (Paxton 1972). This surface migratory

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behavior starts after transformation from larvae to juveniles. The larvae are distributed in the upper 200 m layer both during the daytime and at night (Loeb 1979, 1980, Moser & Smith 1993). Myctophid fishes are thought to migrate to the surface to feed in the productive epipelagic zone, which contributes to their flourishing abundance in the open sea. Generally, the tropical and subtropical species feed mainly in the epipelagic zone at night (Clarke 1978, Hopkins & Baird 1985, Kinzer & Schulz 1985), while the subarctic and transitional water species feed during both night and day (Tyler & Pearcy 1975, Pearcy et al. 1979, Moku et al. 2000). In the Kuroshio region, *Myctophum asperum*,

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Fig. 1. Sampling localities in 1971

M. nitidulum, Symbolophorus evermanni and *Centrobranchus brevirostris* are the 4 most abundant of such migratory species (Hattori 1964, Watanabe 1998). There are considerable data on the feeding habits of myctophid fish that migrate to midwaters (e.g. Merrett & Roe 1974, Baird et al. 1975, Clarke 1978, 1980, Kinzer & Schulz 1985, Hopkins & Gartner 1992), but data for surface migratory species are limited (Gorelova 1975, 1984, Hopkins & Gartner 1992).

In this study, we aimed to determine the main diet and nighttime feeding chronology of the juvenile myctophid species *Myctophum asperum*, *M. nitidulum*, *Symbolophorus evermanni* and *Centrobranchus brevirostris*, which migrate to the upper 1 m layer at night in the Kuroshio Current region. We also estimated the impact of nocturnal feeding by these 4 species on the biomass of epipelagic zooplankton.

MATERIALS AND METHODS

All samples were collected in the Kuroshio region of the western North Pacific (29 to 36°N, 130 to 144°E) from January to March during each of the 10 yr: 1961, 1964, 1968, 1971, 1973, 1977, 1980, 1984, 1988 and 1992. Typical sampling coverage is shown in Fig. 1. Sampling was conducted at night, at 58 to 131 stations (on average, 95) per year during the Japanese Fisheries Agency's survey of eggs and larvae of commercially important fishes. To collect samples, we used a conical plankton net (mouth opening, 130 cm diameter; Maruchi type A net; Nakai 1962, Watanabe & Kawaguchi 1999) placed within 1 m of the surface at shipside, towed at 1 m s^{-1} (2 knots) for 5 to 10 min. Samplings were made consecutively at 1 to 3 h intervals along the grid line designed to cover the average Kuroshio area, resulting in variation in sampling time and distance from Japan but which were averaged each year.

We studied the feeding habits of individuals that were from 10 to 30 mm in standard length (SL). Fish of this size range were the most frequently caught, and formed the first mode in the size frequency distribution and are considered as 0+ years old (Watanabe 1998). After measuring wet weights (wet wt) of the body of each fish and its stomach contents, we identified each prey item in the stomach to the lowest possible taxonomic level and also measured its wet wt. Additionally, we measured total length and width for prey specimens collected in 1971 and 1988, when more than 25 individuals were collected for each of the 4 myctophid species.

We calculated the stomach content index (SCI), i.e. the mean wet wt of stomach content given as a percentage of body weight, for 23 to 35 individuals for each hour between 19:00 and 05:00 h. For hours during which the sample size was fewer than 20 individuals, we pooled the sample with specimens collected in other years (1972, 1974 to 1976, 1985, 1986). We defined the feeding peak as the hour during which the SCI was statistically higher than the values immediately before or after, and higher than the mean SCI for the night. When the peak spanned 2 h, we tested whether the values before and after the peak were significantly lower.

Lastly, we measured the gill raker interval on the lower branch of the first gill arch for each species except *Centrobranchus brevirostris*, whose degenerated gill rakers are adapted to pteropod feeding (Bekker 1966).

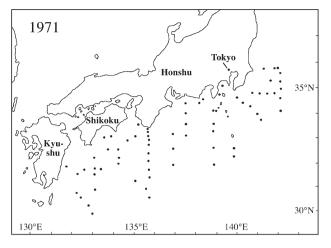
RESULTS

Prey group composition

The 4 species exhibited resource partitioning, although prey composition was sometimes biased due to the small sample size (Fig. 2). No remarkable geographical difference was observed in diets of the 4 species in the years when large sample size was available. Over the 10 yr sampled, unidentifiable stomach content weights were $9.2 \pm 5.4 \%$ (mean \pm SD), $4.3 \pm 3.4 \%$, $8.0 \pm$ 8.1 % and $7.3 \pm 5.9 \%$ of the total stomach contents for *Myctophum asperum, Symbolophorus evermanni, M. nitidulum*, and *Centrobranchus brevirostris*, respectively. These prey items were excluded from Fig. 2.

Myctophum asperum

This species mainly ate appendicularians and a variety of the crustaceans available in the habitat (Fig. 2a). Appendicularians were the most common prey in 6 of



the 10 years (1964, 1968, 1973, 1980, 1984 and 1992) and accounted for 36 to 86 % of the total weight of identified stomach contents (Fig. 2a). In 1961 and 1971, copepods were the most common prey item (53 and 32 %, respectively); the next most common was appendicularians in 1961 and amphipods in 1971 (47 and 29 %, respectively). Euphausiids were the most common prey item in 1977 and 1988 (45 and 41 %, respectively), followed by appendicularians (28 and 32 %, respectively). In 1964, 1973 and 1980, amphipods were the second most common prey item (22 to 34 %) (Fig. 2a).

Symbolophorus evermanni

Euphausiids and amphipods were the main prey items found in identified stomach contents, indicating that this fish depends on larger-sized crustaceans (Fig. 2b). Euphausiids were the most common prey eaten during all 10 years, accounting for 44 to 100 % of the total weight of stomach contents. Amphipods accounted for 0 to 29% and copepods for 0 to 27% (Fig. 2b).

Myctophum nitidulum

This species preyed mostly on copepods and amphipods with occasional euphausiids and appendicularians (Fig. 2c). During the 10 years, copepods and amphipods were the first or second most common prey items, accounting for 66 to 100% of the total weight of identified stomach contents. Euphausiids constituted 6 to 15% in 1971, 1977 and 1980; appendicularians contributed 12 to 34% in 1964, 1984 and 1988 (Fig. 2c).

Centrobranchus brevirostris

This species fed exclusively on gelatinous zooplanktons, i.e. pteropods and appendicularians (Fig. 2d). Pteropods were the most abundant prey item, being 65 to 100% of the total weight of identified stomach contents in 9 of the 10 years; whereas appendicularians accounted for 2 to 35% in 7 of the 10 years. These 2 prey groups accounted for more than 83% of the total prey in 9 of the 10 years, but only 33 in 1980 (Fig. 2d). Copepods, euphausiids and amphipods were occasionally eaten and accounted for 0 to 67, 0 to 11 and 0 to 3% of the total prey, respectively.

Size of prey species or genera

We identified 11 prey species belonging to 14 genera in 416 stomachs of the 4 myctophid species collected in 1971 and 1988 (Table 1). These species comprise much of the zooplankton community found in the subsurface waters in the Kuroshio region at night (Kidachi & Ito 1979, Kidachi et al. 1983, Hirota et al. 1984, Nakata 1997).

The genera and species of copepods found in *Mycto-phum asperum* and *M. nitidulum* were generally similar. Unique to *M. nitidulum*, however, were *Candacia*

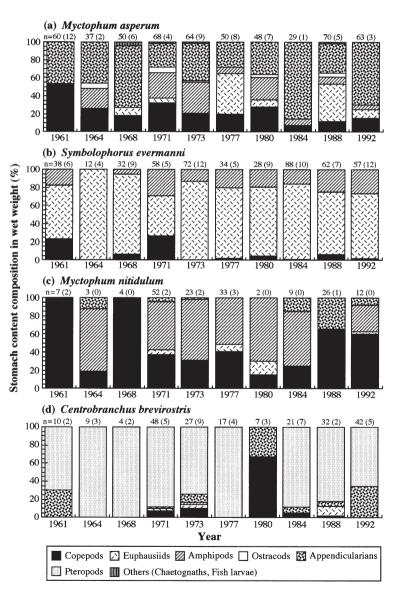
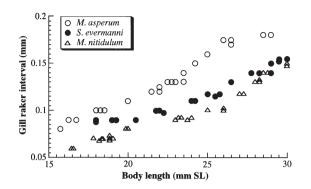


Fig. 2. Wet weight composition of prey groups for the 4 myctophid species in the Kuroshio region: (a) *Myctophum asperum*; (b) *Symbolophorus evermanni*; (c) *Myctophum nitidulum*; and (d) *Centrobranchus brevirostris*. n indicates the number of stomachs examined (number of empty stomachs in parentheses)

Table 1. Size of prey species and genera found in stomachs of *Myctophum asperum, Symbolophorus evermanni, Myctophum nitidulum* and *Centrobranchus brevirostris* in the Kuroshio Current region from January to March in 1971 and 1988

Prey genera and species	ч	<i>M. asperur</i> Length (mm) Avg Range		n (n = 138) Width (mm) Avg Range	ц	S. e Leng Avg	S. <i>evermanni</i> (n = 120) Length (mm) Width (n Avg Range Avg Rē	<i>i</i> (n = 120) Width (mm) Avg Range	n nge	Ρ	<i>M. nitidulum</i> (n = 78) Length (mm) Width (Avg Range Avg R	ulum (m) V ge A	t (n = 78) Width (mm) Avg Range	8) (mm) Range	ц.	<i>C. b</i> Lengt Avg	<i>C. brevirostris</i> (n = 80) Length (mm) Width (m Avg Range Avg Ra	<i>is</i> (n = Widt] Avg	<i>is</i> (n = 80) Width (mm) Avg Range
Copepods Calanus sinicus	5		3.1-4.2	0.7 0.6-0.8	I :	1	I		4					0.4 - 0.6	1	I.	I	I.	I
Calanus spp. Plenromamma aracile	23	2.6 2.3	2.3-4.1 1 9-2 1	0.6 0.5-0.8	11	3.4	3.2–3.8 –	0.8 0.8-	0.8-0.9 7		2.2 1.9-2.3 18 $17-2.0$		0.5 0.3	0.3-0.6 0.3-0.6	- 1	2.8		0.6	
Pleuromamma spp.	16		1.7 - 2.1	0.4 0.3-0.5	I	I	I							0.3-0.6	8	1.2	0.9 - 1.8	0.3	0.1 - 0.5
Paracalanus spp.	40	1.2 0.8	0.8 - 1.4	0.2 0.2-0.3	36	1.1	0.8 - 1.3		0.2-0.4 57		1.3 0.8-1.5		0.3 0.2	0.2 - 0.4	9	1.2	0.8 - 1.4	0.3	0.2 - 0.4
Candacia ethiopica	Ι	I	I	I	3	2.7	2.5 - 2.7	0.8 0.7-	- 6.0-7.0				I.	I	I	I	I	I	I
Candacia sp.	I	I	I	1	1	ר ו כ				1 2	2.6 –		0.6	I	I	I	I	I	I
Canaacia spp. Euchaeta rimana	1 1				<u>C</u> I	1.7	0.2-0.2			74.	- $ 4.1$ $3.7-4.5$		- 0.9 0.8	$^{-}_{0.8-1.0}$					
<i>Euchaeta</i> sp.	1	4.0	I	1.0 -	I	I	I	I	1					I	I	I	I	I	I
Lucicutia sp.	I	I	I	1	I	I	I	1	J	1 2	2.0 –		0.5	I	I	I	I	I	I
Corycaeus crassicusculus	2	1.3 1.5	1.2 - 1.4	0.3 0.3 - 0.4	I	I	I	1	,	,	1	,	I	I	I	I	I	I	I
<i>Corycaeus</i> spp.	15		1.2 - 1.4	0.3 0.3 - 0.4	I	I	I		- 18		0.9 0.8-1.0		0.1 0.1	0.1 - 0.2	17	0.8	0.7 - 1.0	0.1	0.1 - 0.2
Oithona oculata	2	0.9 0.8	0.8 - 1.0	0.2 0.1 - 0.2	I	I	I		1				I	I	I	I	I	I	I
Oithona spp.	11	0.9 0.8	0.8 - 1.0	0.2 0.1 - 0.2	I	I	I		- 31		0.9 0.7-1.0		0.2 0.1	0.1 - 0.2	I	I	I	I	I
Amphipods Lestrigonus schizogeneios	37	4.2 2.3	2.1 - 5.5	0.9 0.3-1.6	29	3.7	1.7 - 4.3	1.1 0.2-	0.2-1.6 69		2.9 1.1-4.2		0.5 0.3	0.3-0.8	Ţ	3.0	I	0.5	I
Platyscelus serratulus	I	I	I		I	I	I	I	1	7 6	6.1 6.0-6.4		1.5 1.5	1.5 - 1.6	I	I	I	I	I
Euphausiids Euphausia simillis	I	I	I	I	32	15.9	15.4 - 16.5	1.6	1.5-1.7 -	1	1		I	I	I	I	I	I	I
Euphausia nana	I	I	I	I	23	7.6	6.9 - 11.1	1.0	0.8-1.0 -	1	1		I	I	I	I	I	I	I
<i>Euphausia</i> sp.	Ι	I	I	I	I	I	I		1		1		I	I	1	12.3	I	1.4	I
<i>Euphausia</i> spp.	ω	8.1 6.3	6.2 - 11.8	0.9 0.7-1.3	56	11.3	7.2-14.8	1.4	0.8-1.8 (6 10	10.4 9.0-	9.0-11.9 1	1.3 1.1	1.1 - 1.5	I	I	I	I	I
Ostracods <i>Concholeci</i> a spp.	15	2.4 2.0	2.0 - 3.0	0.5 0.4-0.6	I	I	I	I	1	1	I		I	I	I	I	I	I	I
Appendicularians <i>Oikopleura</i> spp.	113	5.3 4.0	4.0-7.3	0.1 0.1-0.2	I	I	I	I	- 61		6.0 5.8-7.3		0.3 0.2	0.2 - 0.4	58	3.5	3.0 - 4.6	0.2	0.1 - 0.3
Pteropods Clio pyramidata	I	I	I	I	I	I	I	1	I	1	1		I	I	74	4.8	4.1 - 8.0	2.1	1.6 - 4.0



sp., Euchaeta rimana and Lucicutia sp. Although the size ranges of Pleuromamma, Paracalanus and Oithona spp. individuals eaten by *M. asperum* and *M.* nitidulum overlapped (Table 1), individuals eaten by *M. asperum* were significantly larger (mean \pm SD, 20.9 \pm 3.3 mm SL) than those eaten by M. nitidulum (20.8 \pm 2.5 mm SL) for 3 prey species: Calanus sinicus (0.6 to 0.8 vs 0.4 to 0.6 mm carapace width), Calanus spp. (0.5 to 0.8 vs 0.3 to 0.6 mm) and Corycaeus spp. (0.3 to 0.4 vs 0.1 to 0.2 mm) (U-test, p < 0.05). All 4 species fed on Calanus and Paracalanus spp., probably because they are very abundant in Kuroshio surface waters (Kidachi & Ito 1979, Nakata 1997).

The amphipod Lestrigonus schizogeneios was consumed by all 4 species, accounting for 91 to 100% of the total number of amphipods identified in stomachs (Table 1). The individuals found in *Myctophum nitidulum* stomachs were significantly smaller than those eaten by *M. asperum* (0.9 vs 0.5 mm mean carapace width, *U*-test, p < 0.05), which is similar to the calanoid copepods mentioned above. However, *Platyscelus serratulus*, the size of which was 1.5 to 2 times larger than *L. schizogeneios*, was eaten only by *M. nitidulum*.

Symbolophorus evermanni ate primarily euphausiids, in particular Euphausia similis and E. nana (Table 1). Most of the Euphausia consumed by the other 3 species were less than 12.3 mm in total length and therefore probably E. nana.

The pteropods that we identified were all *Clio pyramidata* and were consumed only by *Centrobranchus brevirostris* (Table 1).

Fig. 3. Slit width of gill rakers on a lower branch of the first gill arch in relation to body length in *Myctophum asperum*, *Symbolophorus evermanni* and *M. nitidulum*

Gill raker interval

Gill raker intervals (slit width) were 0.08 to 0.18 mm, 0.09 to 0.16 mm and 0.06 to 0.15 mm for *Myctophum asperum, Symbolophorus evermanni* and *M. nitidulum* individuals of 15.0 to 30.0 mm SL, respectively (Fig. 3).

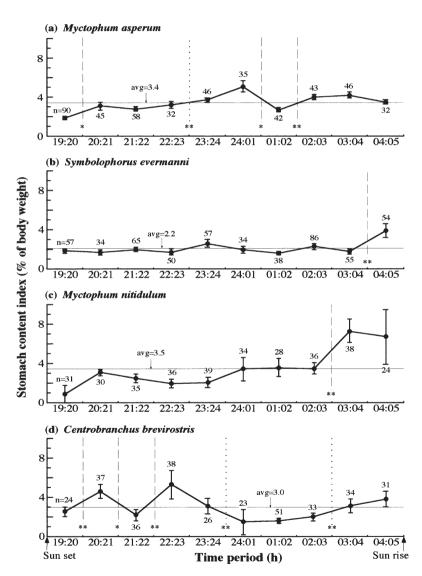
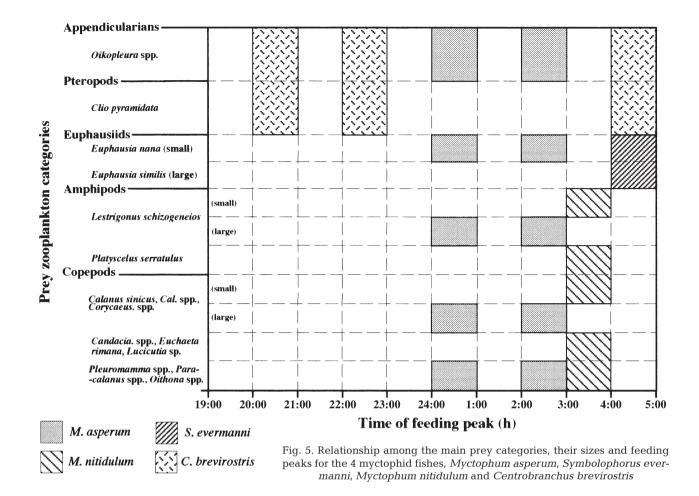


Fig. 4. Temporal change in stomach content weight at night (SCI), given as percentage of fish body weight, for: (a) *Myctophum asperum*; (b) *Symbolophorus evermanni*; (c) *Myctophum nitidulum*; and (d) *Centrobranchus brevirostris*. n indicates the number of stomachs examined. Vertical bars indicate \pm SE. Broken lines indicate a significant difference between both sides of the line; dotted lines indicate a significant difference between the peak value and the values 2 h before or after the peak (U-test, *p < 0.01, **p < 0.05)



These data indicate that the 3 species have gill raker intervals small enough to retain small copepods such as *Paracalanus* and *Oithona* spp. (Table 1). However, some prey species or genera were segregated by size and fish species, indicating that the fishes feed selectively in the top 1 m water layer at night rather than randomly filtering water through gill rakers.

Feeding chronology

Changes in SCI by the hour at night (feeding chronology) showed species-specific patterns of feeding activity (Fig. 4).

Myctophum asperum

Feeding peaks were from 24:00 to 01:00 h (5.1% SCI) and 02:00 to 03:00 h (4.0% SCI) (Fig. 4a). The SCI for 01:00 to 02:00 h (2.7%) was significantly lower than the SCI values immediately before and after (*U*-test, p < 0.01 and p < 0.05).

Symbolophorus evermanni

Feeding activity fluctuated around the mean (2.2% SCI) throughout the night, with a significant peak (3.9%) at dawn (04:00 to 05:00 h) (*U*-test, p < 0.05) (Fig. 4b).

Myctophum nitidulum

SCI values were below the mean (3.5%) before 24:00 h but increased gradually to the peak (7.3%) from 03:00 to 04:00 h (Fig. 4c).

Centrobranchus brevirostris

Two notable peaks, 1.5 to 1.8 times higher than the mean (3.0% SCI), occurred from 20:00 to 21:00 h (4.6%) and from 22:00 to 23:00 h (5.3%) (Fig. 4d). The SCI values decreased to 1.5% from 24:00 to 01:00 h and then gradually increased to 3.8% from 04:00 to 05:00 h (Fig. 4d).

Resource partitioning

Fig. 5 shows the major prey items of the 4 species in relation to the peak feeding hours in Fig. 4. There was no overlap of main prey category or peak feeding time among the 4 species, suggesting that trophic competition is reduced because the species differ in prey type, prey size and peak feeding hours in the upper 1 m layer during the night (Fig. 5).

DISCUSSION

Resource partitioning

Other myctophid species in the subtropical and tropical Atlantic and Pacific Oceans choose the species and size of their prey (Merrett & Roe 1974, Clarke 1980, Roe & Badcock 1984, Hopkins & Baird 1985, Hopkins & Gartner 1992). The myctophid fish studied here appear to behave similarly to such species, except that they do not engage in opportunistic, random filter feeding. Because their gill raker slits are small enough to filter prey of all sizes (as shown by the stomach contents), random filter feeding does not explain why Myctophum asperum, Symbolophorus evermanni and M. nitidulum consume different prey. Microscale spatiotemporal distributions of each prey organism and each myctophid species are required to explain the mechanisms of the observed resource partitioning in the surface layer of the open sea.

Peak feeding hours

The myctophid species in this study differ in peak feeding time, as do fish found off Hawaii, in the tropical western Pacific, in the transitional western North Pacific, and in the subtropical eastern North Atlantic (Merrett & Roe 1974, Gorelova 1975, Clarke 1978, Roe & Badcock 1984, Kawamura & Fujii 1988). However, our nighttime catches of the 4 myctophid species did not significantly differ by the hour (H. Watanabe unpubl. data), suggesting that the density of myctophids that migrate to surface waters is maintained throughout the night in the upper 1 m layer in the waters of the Kuroshio region. Similar results have been reported for Myctophum nitidulum in this study area (Hattori 1964). The nighttime densities of some species of copepods, euphausiids and amphipods in the surface layer, however, are known to change with time according to the species' vertical migration in the equatorial and subarctic areas of the Pacific Ocean and the Southern China Sea (Roger 1971, Roe 1984, Roe et al. 1984, Williams & Conway 1984, Hirota 1987, Hattori

1989). Thus, prey density and composition probably fluctuate temporally during the night in the top 1 m layer of the open sea. The feeding activity of myctophids that migrate vertically over the day and night usually peaks when prey density in the habitat is highest (Clarke 1978, Roe & Badcock 1984, Kinzer & Schulz 1985). Therefore, the chronology of nocturnal feeding by myctophids that migrate to the surface may be affected by temporal changes in zooplankton density and also by the feeding condition (such as being full versus hungry) of the fishes themselves. Notably, M. asperum and Centrobranchus brevirostris, the species that consume gelatinous plankton, had 2 and 3 feeding peaks, respectively (Fig. 4). These species probably digest these low-nutrient, gelatinous prey quickly and thus need to ingest large numbers of them.

Prey organisms

Some myctophids (such as Lampanyctus alatus, Bolinichthys longipes, Diaphus perspicillatus and Ceratoscopelus warmingi) that migrate to midwaters feed frequently on pigmented copepods belonging to Euchaeta, Pleuromamma, Candacia and Oncaea (Gorelova 1978, Clarke 1980, Hopkins & Baird 1985, Kinzer et al. 1993). This diet has also been reported in the equatorial Pacific for species such as Myctophum spinosum and M. nitidulum that migrate to surface waters (Gorelova 1975). However, the species in this study that migrate to the surface fed mostly on nonpigmented Paracalanus copepods, probably because these copepods are abundant in Kuroshio surface waters (Kidachi & Ito 1979, Nakata 1997).

Centrobranchus also consumes gelatinous zooplankton in the Gulf of Mexico (*C. nigrocellatus*; Hopkins & Gartner 1992) and the tropical western North Pacific (*C. andreae*; Gorelova 1975). The stub-like degenerated gill raker that is diagnostic of this genus (Bekker 1966) very likely evolved as a result of exclusive feeding on gelatinous zooplankton, especially pteropods.

Impact of predation by myctophids on surface zooplankton biomass

Myctophids that migrate to the surface excrete food about 2 to 4 h after feeding in subtropical and tropical waters where temperatures range between 13 and 22° C (Gorelova 1975, 1984, Clarke 1978). Mean surface water temperatures at the stations at which *Myctophum asperum, Symbolophorus evermanni, M. nitidulum* and *Centrobranchus brevirostris* were sampled are 18.2°C (ranging from 11.6 to 22.6°C; n = 3348), 18.4°C (11.5 to 22.3°C; n = 2101), 18.0°C (7.3 to 22.6°C; Table 2. Estimates of the predation impact on zooplankton by the 4 species *Myctophum asperum, Symbolophorus evermanni, Myctophum nitidulum* and *Centrobranchus brevirostris* per night in the top 5 m in the Kuroshio Current region from January to March in 1971, 1973, 1977, 1980 and 1984. *Mean zooplankton biomass (wet weight) in the top 150 m of the studied area from January to March (Kuroda 1991), **biomass (wet weight) of each myctophid species in the top 5 m at night (Watanabe 1998)

	M. asperum	S. evermanni	M. nitidulum	C. brevirostris	Total of the 4 species
Nighttime ration (% of body weight)	8.1-12.5	5.1-8.5	9.7-14.0	7.1-12.2	_
1971					
Zooplankton biomass 111 mg m ⁻³ *					
Myctophid biomass (mg m ⁻³)**	31.2	42.1	14.0	21.8	109.0
Nighttime ration (mg m ⁻³)	2.53 - 3.90	2.15 - 3.58	1.36 - 1.96	1.55 - 2.66	7.59-12.10
Predation impact (% of 111 mg m^{-3})	2.28 - 3.51	1.94 - 3.23	1.23-1.77	1.40 - 2.40	6.84 - 10.90
1973					
Zooplankton biomass 69 mg m ^{-3*}					
Myctophid biomass (mg m^{-3})**	8.7	12.0	1.7	1.7	24.0
Nighttime ration (mg m^{-3})	0.70 - 1.09	0.61 - 1.02	0.16 - 0.24	0.12 - 0.21	1.59 - 2.56
Predation impact (% of 69 mg m^{-3})	1.01 - 1.58	0.88 - 1.48	0.23-0.35	0.17-0.30	2.30-3.71
1977					
Zooplankton biomass 38 mg m ^{-3*}					
Myctophid biomass (mg m ⁻³)**	4.4	3.4	3.3	0.9	12.0
Nighttime ration (mg m ⁻³)	0.36 - 0.55	0.17 - 0.29	0.32 - 0.46	0.06 - 0.11	0.91 - 1.41
Predation impact (% of 38 mg m^{-3})	0.95 - 1.45	0.45 - 0.76	0.84 - 1.21	0.16 - 0.29	2.39 - 3.71
1980					
Zooplankton biomass 53 mg m ⁻³ *					
Myctophid biomass (mg m ⁻³)**	3.6	2.1	0.3	0.5	6.5
Nighttime ration (mg m ⁻³)	0.29 - 0.45	0.11 - 0.18	0.03 - 0.04	0.04 - 0.06	0.47 - 0.73
Predation impact (% of 53 mg m^{-3})	0.55 - 0.85	0.21 - 0.34	0.06 - 0.08	0.08 - 0.11	0.89 - 1.38
1984					
Zooplankton biomass 30 mg m ⁻³ *					
Myctophid biomass (mg m ⁻³)**	3.2	10.0	0.6	1.5	15.3
Nighttime ration (mg m ⁻³)	0.26 - 0.40	0.51-0.85	0.06 - 0.08	0.11 - 0.18	0.94 - 1.51
Predation impact (% of 30 mg m^{-3})	0.87-1.33	1.70 - 2.83	0.20 - 0.27	0.37 - 0.60	3.13-5.03

n = 525) and 18.9°C (14.3 to 22.3°C; n = 522), respectively (Watanabe 1998). At these temperatures, the time to excretion would be about 2 to 4 h after feeding. We assumed a time to excretion of 4 h and estimated the impact of predation on zooplankton biomass by summing the highest SCI values observed from 19:00 to 23:00 h, 23:00 to 03:00 h and 03:00 to 05:00 h. We estimated a maximum predation impact of 12.5% for M. asperum at night, based on interval estimates of 3.2% (from 19:00 to 23:00 h), 5.1% (from 23:00 to 03:00 h) and 4.2% (from 03:00 to 05:00 h). Similarly, we used sums of the lowest SCI values (1.9%, 2.7% and 3.5%)over these same 3 time periods to estimate 8.1% as the minimum predation impact. Thus, we estimate that M. asperum consumed 8.1 to 12.5% of its body weight in zooplankton biomass per night. Based on similar calculations, nighttime zooplankton rations were estimated as 5.1 to 8.5% for S. evermanni, 9.7 to 14.0% for M. nitidulum and 7.1 to 12.2% for C. brevirostris. Considering that migratory myctophid fishes generally feed mainly at night in subtropical and tropical waters (Gorelova 1973, Baird et al. 1975, Clarke 1978, Hopkins & Baird 1985, Kinzer & Schulz 1985), these values are valid estimates of the daily rations for these species. Gorelova (1984) reported that daily rations of juvenile surface migratory myctophids such as *M. asperum, M. spinosum* and *Hygophum proximum* (17 to 28 mm SL) in the tropical waters of the Pacific are approximately 10 to 20 % of the body wet wt. This estimate is almost 1.5 to 2 times of magnitude higher than our estimate for the Kuroshio region in the western North Pacific.

The biomass of myctophids that migrate to surface waters was estimated for the top 5 m in the studied area from 1971 to 1984 (Watanabe 1998). Zooplankton biomass in the upper 150 m was measured at each myctophid sampling station (Kuroda 1991). Based on these biomass data, we estimated the zooplankton biomass consumed by the 4 myctophid species per night, and their feeding impact, for 5 years between 1971 and 1984 (Table 2). Our calculations indicate that, by consuming individuals of 10 to 30 mm SL, the 4 species removed 2.30 to 3.71%, 2.39 to 3.71%, 0.89 to 1.38% and 3.13 to 5.05% of the total zooplankton biomass per night in the top 150 m during 1973, 1977, 1980 and 1984, respectively (Table 2). Hopkins &

Gartner (1992) reported that predation by myctophids removes 2% of the zooplankton biomass per night in the upper 200 m of the eastern Gulf of Mexico. This value takes into account predation by species that migrate to both surface and midwaters, so the impact on zooplankton of nocturnal predation by myctophids that migrate to the surface is higher in the Kuroshio region of the western North Pacific than in the Gulf of Mexico. In 1971, zooplankton biomass from January to March in the study area was 1.5 to 3 times greater than in any other year from 1972 to 1988 (Kuroda 1991) and biomass of the 4 myctophid species was 4 to 15 times greater in 1971 than in any other year from 1957 to 1988 (Watanabe 1998). Thus, our estimate of predation impact of 6.84 to 10.90% by the 4 species combined for 1971 (Table 2) is probably so high because recruitment was high; Japanese sardine recruitment was also high in 1971, the year that the biomass began to rebound.

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