



Mangrove fish assemblages reflect the environmental diversity of the Galapagos Islands

Denisse Fierro-Arcos^{1,*}, José R. Marín Jarrín^{1,2}, Octavio Aburto-Oropeza³,
Euan S. Harvey⁴, Etienne Rastoin-Laplane^{1,4}, Pelayo Salinas-de-León^{1,5}

¹Charles Darwin Foundation, Charles Darwin Research Station, Puerto Ayora, Santa Cruz Island, Galapagos 200102, Ecuador

²Department of Fisheries Biology, Humboldt State University, Arcata, California 95521, USA

³Marine Biology Research Division, Scripps Institution of Oceanography, San Diego, California 92093, USA

⁴School of Molecular and Life Sciences, Curtin University, Bentley, WA 6845, Australia

⁵Pristine Seas, National Geographic Society, Washington, DC 20036, USA

ABSTRACT: Mangroves are important habitats for a variety of ecologically, commercially and culturally important fishes. However, little is known about their role within the Tropical Eastern Pacific (TEP), and particularly in the Galapagos Archipelago, the westernmost limit for mangroves in the Americas, and the only oceanic islands in the TEP where mangroves are present. We describe patterns of fish composition in the 2 Galapagos bioregions where mangroves are present and assess potential environmental factors influencing fish community composition. Underwater Visual Census and Stereo Baited Remote Underwater Video stations (stereo-BRUVS) were used to sample fish communities. We identified 35 029 fish representing 93 species, 67 genera and 36 families. Pomacentridae, Mugilidae, Haemulidae and Lutjanidae were the most common families. Juveniles made up 43% of the fish, 30 species were of importance to local artisanal fisheries and 80% of species were associated with reef habitats, suggesting mangroves in Galapagos may provide nursery habitats for economically valued species. Fish assemblage composition varied across bioregions, with 6 taxa responsible for driving these differences, including species of economic importance: *Lutjanus novemfasciatus* and *Mycteroperca olfax*. Species richness was 17% higher in the Central-Southeastern than in the Western region; while higher species richness, Shannon-Wiener diversity and Pielou's evenness were detected with BRUVS than through visual censuses. Our results highlight the role of mangroves as a habitat for a unique fish community composed of young, endemic and commercially important species, whose composition is likely driven by the isolation of the islands and its location in a convergence zone.

KEY WORDS: Fisheries · Nursery habitat · Marine protected area · MPA · Underwater Visual Census · UVC · Stereo-Baited Remote Underwater Video stations · Stereo-BRUVS · Tropical Eastern Pacific

Resale or republication not permitted without written consent of the publisher

1. INTRODUCTION

Mangrove forests are considered to be amongst the most productive and diverse ecosystems on Earth (Alongi 2002). Mangroves can be found globally along intertidal areas in tropical and subtropical regions, where they provide a variety of key ecosystem services (Alongi 2002). One such service is the

provision of nursery habitats to various ecologically and economically important species of fish and crustaceans, thus making mangroves an important contributor to the recruitment of fish and invertebrate populations (Aburto-Oropeza et al. 2009, Saenger et al. 2013, Nagelkerken et al. 2015, zu Ermgassen et al. 2020). Despite their ecological importance, an estimated 50% of mangrove forests worldwide have

been lost in the past 50 yr (López-Portillo et al. 2017), largely as a result of human activities (Saenger et al. 2013, Chowdhury et al. 2017). The resilience of mangrove ecosystems are also negatively affected by anthropogenic pressures, making them vulnerable to the effects of climate change (Jennerjahn et al. 2017). It is predicted that by the end of this century the combination of human pressures and climate change impacts will have a significant negative effect not only on mangrove distribution, but also on the ecosystem services they provide (Jennerjahn et al. 2017).

The Tropical Eastern Pacific (TEP), a marine ecoregion extending from southern Baja California (Mexico) to northern Peru (Robertson & Cramer 2009), contains ~7% of global mangrove coverage (Giri et al. 2011). In the TEP, mangroves are found along the continental coastline of the Americas, and on one set of oceanic islands, the Galapagos Islands (Giri et al. 2011). As widely reported in locations across the Caribbean and the Indo-Pacific, mangroves are also known to play an important ecological role in marine ecosystems of the Eastern Pacific (Dorenbosch et al. 2004, 2005, Mumby et al. 2004, Aburto-Oropeza et al. 2008, Nagelkerken et al. 2008). However, very little is known about their influence on the composition of fish community assemblages across most of the TEP, and in particular on mangrove fish communities of the Galapagos Archipelago.

The Galapagos Marine Reserve (GMR) is a multiple use reserve established in 1998 to protect the Galapagos Archipelago and its surrounding waters (Heylings et al. 2002). The GMR covers a total area of 138 000 km² where no industrial fishing is allowed (Heylings et al. 2002). The Galapagos Archipelago is located about 1000 km west of the South American Pacific coast (see Fig. 1), and it is made up of 13 large islands and over 100 islets of volcanic origin (Snell et al. 1996). The Galapagos is the only tropical archipelago located in a convergence zone where various cool and warm water oceanic currents meet (Andersen & Hamann 1986, Edgar et al. 2004a). The distinct oceanographic conditions together with the local geological features result in markedly different environmental conditions across the archipelago (Banks 2002, Schaeffer et al. 2008). This spatial variability in environmental conditions has a strong influence on the distribution of marine fauna, such as shallow reef fishes and sharks (Banks 2002, Edgar et al. 2004a, Palacios et al. 2006, Acuña-Marrero et al. 2018). These patterns in faunal distribution are consistent enough to allow for the separation of the marine areas of the archipelago into 3 main bioregions (see

Fig. 1): (1) far-northern (not shown in Fig. 1), (2) Central-Southeastern (CSE), which includes the northern subregion, and (3) Western, including the Elizabeth subregion (Edgar et al. 2004a).

Mangroves cover about 35% of the Galapagos coastlines, and in contrast with global trends, their coverage on the islands is estimated to have increased by 24% between 2004 and 2014 (Moity et al. 2019). This overall increase was likely the result of the protected status of the Galapagos Islands, and the lack of human interference in mangrove areas (Moity et al. 2019). Despite this upward trend in coverage, mangroves in the Galapagos are particularly vulnerable to climate change because of their location in a convergence zone, where they are regularly subjected to environmental conditions at the edge of their physiological tolerance (Andersen & Hamann 1986, Doney et al. 2012, Moity et al. 2019). Since mangroves are severely understudied in the Galapagos, we cannot be certain of the ecological role they currently play in the marine systems of the archipelago, or how its ecological function will be affected by global climate change. Mangrove fish assemblages have not been fully described yet, and the potential spatial variation of these assemblages across the archipelago has not been examined either. The few studies conducted to date in the archipelago are mostly available as part of the grey literature, and either focus on a small subset of islands (Suárez-Moncada 2012, Aguaiza 2015, Llerena-Martillo et al. 2018), or on a selected group of fish species (Llerena-Martillo et al. 2015). This knowledge gap represents a challenge, as without accurate data about the diversity of mangrove fish populations and the relationship between fish assemblages and mangrove habitat conditions, it is not possible to identify key areas deserving additional protections; or to design, support and improve conservation strategies that are effective in protecting these marine communities (Ley 2005, Banks et al. 2009). An improved understanding about the ecological role of mangroves in the Galapagos could also be used to assess future alterations to this ecosystem brought by a changing climate (Visbeck 2018).

To avoid any negative impacts inside protected areas, such as the GMR, derived from permanently removing individual fish as a result of sampling, the use of non-destructive sampling methods are deemed more suitable (Mallet & Pelletier 2014). Underwater Visual Census (UVC) and Stereo-Baited Remote Underwater Video stations (stereo-BRUVS) are 2 non-destructive techniques widely used to obtain information about the composition of fish

assemblages in a variety of marine systems (Mallet & Pelletier 2014). UVC is a relatively inexpensive and easy method to sample fish communities in clear, shallow water areas; however, observations are influenced by the presence of divers, and it is prone to inconsistencies between observers (Murphy & Jenkins 2010, Mallet & Pelletier 2014).

Advantages of using stereo-BRUVS include a wider range of species sampled due to the presence of bait, higher accuracy and consistency in both identifications and length measurements, as videos can be revised by multiple observers, thus ensuring mistakes are minimised or avoided (Mallet & Pelletier 2014). Nonetheless, stereo-BRUVS also have some disadvantages, including the unknown influence of the bait plume, the time required to analyse videos and the high costs of purchasing required equipment and software (Langlois et al. 2010, Watson et al. 2010).

Here, we combined UVCs and stereo-BRUVS to sample fish communities during the warm season (December to April) of 2015 across mangroves in the GMR. Our study had 5 objectives: (1) describe the Galapagos fish communities across all islands where mangroves are present, (2) compare results about fish community structure obtained with UVC and stereo-BRUVS, (3) compare the composition of fish assemblages across the 2 bioregions where mangroves are present, (4) investigate the environmental factors that may influence fish community composition and (5) explore the potential nursery function of Galapagos mangroves for 3 taxonomic groups important to local artisanal fisheries. We hypothesised that the composition of fish communities in Galapagos mangroves would vary between bioregions due to their consistently different environmental and oceanographic conditions (Edgar et al. 2004a). We also expected community composition to vary between sampling methods, as they have been reported to sample different subsets of the fish community (Colton & Swearer 2010, Langlois et al. 2010). Given that the Galapagos does not have many structured estuarine and coastal areas that may offer nursery habitats for fishes and invertebrates (Beck et al. 2001, Danulat & Edgar 2002, Lefcheck et al. 2019), we anticipate finding evidence supporting the role of Galapagos mangroves as nursery habitats to species of local commercial interest, as has been reported for other areas of the planet (Aburto-Oropeza et al. 2009, Nagelkerken et al. 2015, zu Ermgassen et al. 2020). Furthermore, we expect our results to be used as a reference point to assess the effectiveness of the GMR in protecting fish diversity in mangrove areas

from local human pressures (e.g. tourism and artisanal fishing), and to evaluate the effects of climate change in this ecosystem.

2. MATERIALS AND METHODS

2.1. Study area

The Galapagos represents the westernmost limit for mangroves in the Americas (Lacerda et al. 1993). Here, mangroves are often exposed to environmental conditions at the limit of their physiological tolerance, including little protection from direct wave energy, seasonal exposure to temperatures as low as 18°C, lack of major freshwater sources and low soft sediment availability (Andersen & Hamann 1986, Schaeffer et al. 2008, Moity et al. 2019). The Galapagos has a semidiurnal mesotidal regime (Banks 2002) with an average tidal range spanning from 1.8 to 2.4 m (Wellington 1975), which coincides with the prevailing mesotidal regime in most of the TEP (Castellanos-Galindo & Krumme 2015). Only 4 species of mangrove trees have been reported on the islands, *Rhizophora mangle* (red mangrove), *Avicennia germinans* (black mangrove), *Laguncularia racemosa* (white mangrove) and *Conocarpus erectus* (button mangrove) (Andersen & Hamann 1986). Due to these suboptimal environmental conditions, mangroves on the islands are mostly underdeveloped across the archipelago, and they often form small, narrow vegetated areas growing directly on lava fields. In fact, it was found that over three-quarters of mangrove patches in the archipelago were smaller than 0.25 ha, and 50% of total mangrove coverage was found within 100 m of the coastline (Moity et al. 2019). However, a few enclosed bays exist on Isabela Island, where mangroves are largely sheltered from direct wave energy, and they are able to reach heights of up to 25 m and form well developed forests (Moity et al. 2019).

Mangroves cover 35% of the Galapagos coastlines, with a total area of 3657.10 ha, where 52% are found in the Western bioregion and 48% in the CSE. Mangroves become less common north of the equatorial line, so the northern subregion of the CSE contains <1% of mangrove coverage, and no mangroves are present in the far-northern region (Moity et al. 2019). Due to their remoteness, mangroves in the Galapagos are relatively unaffected by human activities (Lacerda et al. 1993). Mangrove patch size varies greatly across the archipelago as a function of the geological age of the island, with younger islands

having more mangrove cover than older ones (Moity et al. 2019). It is hypothesised that this is due to mangroves acting as pioneer vegetation because of their ability to withstand the difficult and highly dynamic environmental conditions characteristic of intertidal environments (Moity et al. 2019). However, mangroves are not strong competitors, and they are usually outcompeted for light by other species, particularly in the higher parts of the intertidal area, where they are also at risk of desiccation (Duke 2017). Therefore, to ensure an accurate representation of the natural variability of fish communities occupying mangrove habitats in the Galapagos, we sampled a variety of mangrove patch sizes, but concentrated our efforts in the Western and CSE regions, excluding the northern subregion, where the vast majority of mangrove areas are located.

2.2. Sampling of fish communities

Sampling took place during the warm season in daylight hours between 10 and 27 April 2015 in mangrove habitats on 7 islands and across 2 bioregions (Fig. 1) using UVC and stereo-BRUVS. We chose these 2 methods as research on clear water reef sys-

tems suggests that they complement each other by sampling different sections of the fish community under study, thus offering a more complete indication of the overall fish assemblage composition (Colton & Swearer 2010, Langlois et al. 2010, Watson et al. 2010, Walsh et al. 2017). Most surveys took place within 2 h of slack high tide, to ensure mangrove roots were inundated. Whenever possible, individual fish were identified to the finest taxonomical level (i.e. species). Length measurements were also estimated for each individual sampled as detailed below.

2.2.1. UVCs

We surveyed a total of 22 mangrove bays across 6 islands (Fig. 1) using UVCs. At each site, fish surveys were conducted by 2 researchers snorkelling side by side, at the same speed and parallel to the mangrove fringe, following protocols of previous mangrove surveys within the TEP (Aburto-Oropeza et al. 2007, 2009). Four replicate 50 × 2 m transects were surveyed at each site. Replicates were separated by at least 10 m from one another to avoid detecting the same individual fish across replicates. Total transect

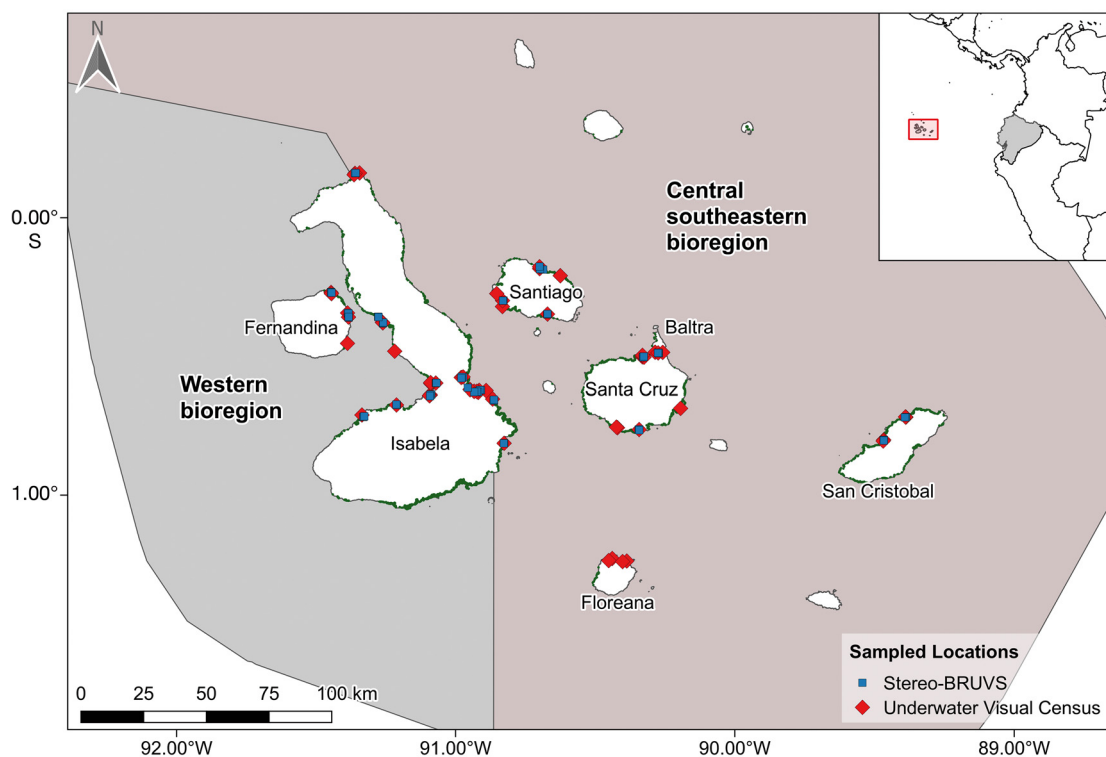


Fig. 1. Mangrove sites where fish communities were surveyed across 2 bioregions of the Galapagos Archipelago (Edgar et al. 2004a). Coastal green areas: mangrove patches across the archipelago

length was measured using a measuring tape, which researchers attached to one end of the mangrove area being sampled. Researchers identified, counted and estimated total length (TL) for all individual fish located within 1 m of either side of the transect, surveying a total area of 100 m² per transect. Surveys were carried out at a maximum depth of 2 m to minimise low visibility conditions, which could affect the accuracy of the data obtained.

Sampling occurred parallel to the mangrove fringe due to the small size and high root density of most mangrove patches in our study area. Researchers were often physically unable to enter mangrove areas and sample among their roots. However, because transects were laid immediately next to the mangrove fringe, fish up to 1 m inside the mangrove roots were included in our surveys.

To reduce observer bias, all UVCs were conducted by only 2 researchers, who were experienced at visually surveying fish assemblages in the TEP and at accurately estimating fish length. Additionally, to ensure TL estimation was consistent between the 2 observers, a plastic transparent ruler and a PVC tube representing 22 size classes, ranging from 2 to 125 cm, were used as reference by researchers when estimating fish sizes (Aburto-Oropeza et al. 2009).

2.2.2. Stereo-BRUVS

Stereo-BRUVS systems were deployed in 20 mangrove bays across 5 islands (Fig. 1). We completed 3 replicate deployments per site. Individual stereo-BRUVS deployments were separated by a minimum distance of 500 m to avoid the overlap of bait plumes and to reduce the likelihood of detecting the same individual fish across multiple replicates (Langlois et al. 2018).

The stereo-BRUVS systems consisted of 2 GoPro Hero 4 black edition cameras mounted inside PVC waterproof housings located 0.70 m apart on a steel base and inwardly converging at a 6° angle (SeaGIS). Each system was deployed from a dinghy at depths ranging from 1 to 4.5 m, and at an average distance of 2 m from the mangrove roots, as due to the small size of mangrove patches, it was often impossible to place stereo-BRUVS among the roots. Stereo systems were left to film for a minimum soaking time of 100 min while floating about 1 m above the sea floor as described by Acuña-Marrero et al. (2018). To keep consistency with previous monitoring programs in the Galapagos, 800 g of yellowfin tuna *Thunnus albacares* was used as bait, which was

placed inside a PVC canister with holes to allow dispersion into the water column.

2.2.3. Video analysis

Stereo-BRUVS systems were calibrated following Harvey & Shortis (1998) using the specialised CAL software (SeaGIS). Videos were analysed using EventMeasure software (SeaGIS). The lead author conducted all video analyses to ensure results were consistent and fish identifications were validated by several fish experts. A total of 90 min of video was analysed per deployment, the first 5 min of footage after the system settled on the seafloor and the last 5 min prior to collection were discarded to minimise any potential disturbance to behaviour of animals in the area. All fish that could be clearly seen by the analyst were counted and identified using Grove & Lavenberg (1997), Robertson & Allen (2015) and Froese & Pauly (2019) as identification guides. MaxN, which is described as the maximum number of individuals of the same taxon present within a single video frame, was used as a measure of relative abundance. MaxN is a conservative measure of abundance, but it allows us to avoid counting the same individual fish multiple times if they re-enter the field of view (Cappo et al. 2003).

Fork length measurements were obtained only from individuals recorded at the time of MaxN for each species whenever allowed by visibility and position of the animal in relation to the cameras (i.e. individual appears as straight as possible in both cameras and their body is parallel to the cameras). Because visibility (i.e. the furthest distance from the cameras to which we could clearly see an object) did vary across individual stereo-BRUVS drops, we only used measurements if the calculated root mean square (RMS), a measure of the performance of our estimates (Skiena 2017), was ≤ 100 , and the precision value was a maximum 10% of the length measured. If either condition was not met, measurements were rejected.

2.3. Collection of environmental data

Based on previous publications, we identified the following variables to be potentially influential on fish community composition: total mangrove area and length of mangrove fringe (Ley 2005, Aburto-Oropeza et al. 2008), tidal stage (Castellanos-Galindo & Krumme 2015, Ramirez-Martínez et al.

2016) and visibility (Gladstone et al. 2012). Furthermore, sea surface temperature (SST) and chlorophyll *a* were identified as another set of potentially influential environmental factors, because they are known to differ greatly across the archipelago (Banks 2002), and can influence the distribution of fishes across space and time (Currie et al. 2004, Tittensor et al. 2010).

Depth, time and location (latitude and longitude) were recorded immediately prior to each deployment and transect, but we were unable to collect environmental data *in situ*. However, open source, remotely sensed environmental data were used to investigate if they had any potential correlations with fish assemblage composition across bioregions. Although, remotely sensed data may not capture the distinctive environmental conditions of sites in waters close to the coastline (Thakur et al. 2018), we decided to use it as an exploratory tool that would allow us to identify the variables that show a potential correlation with the composition of fish communities in mangroves, and are thus worth monitoring over the long-term (LaDeau et al. 2017).

Mean SST and mean chlorophyll *a* concentrations per season (warm season from December 2014 to April 2015, and dry season from June to October 2014) were calculated from Aqua MODIS satellite data, which have a resolution of 4 km (NOAA NMFS SWFSC ERD 2017a,b). Mangrove area and mangrove fringe length were measured in QGIS 3 (QGIS Development Team 2019) using a Galapagos mangrove distribution layer by Moity et al. (2019). Information about tides (incoming or outgoing) were obtained from INOCAR (2017) for the day and time when sampling occurred. Mean maximum visibility was calculated for stereo-BRUVS deployments only, as it varied widely not only across sites, but also with time at the same site.

Visibility measurements were not obtained for UVC transects because the relative short duration of sampling using this method allowed us to put measures in place to ensure visibility was consistent across time and space. As stated in the previous section describing UVCs, sampling occurred at a maximum depth of 2 m to minimise low visibility conditions. To calculate maximum visibility values for stereo-videos, we recorded the distance from the cameras to the furthest object/fish that could be clearly seen in our screen using a 3D point in the EventMeasure software. Distance to the furthest object/fish on the screen was recorded every 10 min for the entire duration of the video being analysed. Finally, a mean maximum visibility was

calculated using all the maximum visibility values per video.

Draftsman plots were used to visually inspect the distribution of the data for each environmental variable described above and to investigate multicollinearity among predictive variables. If any 2 predictive variables were found to be highly correlated ($r > 0.90$), and this correlation was found to be significant, then only 1 variable was kept for further analysis. Plots were created using the GGally package (Schloerke et al. 2018) and significance of correlations were calculated with the corrplot package (Wei & Simko 2017) in R v3.6 (R Core Team 2020). After inspection of the data, a $\log_e(x + 1)$ transformation was applied to mangrove area measurements to reduce skewness (Clarke & Warwick 2001). Environmental data was normalised and a Euclidian similarity matrix was calculated for use in multivariate analyses.

2.4. Statistical analyses

Raw data obtained by UVCs and stereo-BRUVS are not comparable because fish abundance is not measured in the same way by both methods. Since our focus was on comparing results between sampling techniques and bioregions, we standardised results by calculating relative abundances for each fish species. Relative abundance was obtained by dividing the total abundance per species by the total number of all individual fish of all species found in that transect or deployment. Mean relative abundance was calculated per site sampled, and a fourth root transformation was applied to decrease the influence of more dominant species, highlight the relative importance of rare species and to improve normality and reduce heteroscedasticity in our data (Clarke & Warwick 2001). A Bray Curtis dissimilarity matrix was calculated on the transformed relative abundances for use in multivariate analysis.

Fish species were classified into 4 trophic groups: apex, carnivores, herbivores and omnivores, based on trophic level information available at FishBase (Froese & Pauly 2019). Herbivore species had a trophic level between 2.00 and 2.19, omnivores had a trophic level between 2.20 and 2.79, and carnivores had a trophic level ≥ 2.80 (Palomares 2000). The apex predators group included sharks only, as we were interested in finding out if one method was better than the other at detecting them. Due to the lack of assumptions about data and error distribution, univariate permutational multivariate analyses of vari-

ance (PERMANOVA; Anderson 2005) were used to test for differences in the relative proportions of trophic groups between sampling methods and across bioregions. PERMANOVA was performed in the R *vegan* package (Oksanen et al. 2019) using Euclidian distances. Univariate PERMANOVAs based on Euclidean distances were also used to test for differences in total number of individuals detected, species richness, Shannon-Wiener diversity index (H') and Pielou's evenness (J') between methods and across sites.

A 2-way PERMANOVA based on Bray Curtis dissimilarities was used to test for differences in the fish assemblages between stereo-BRUVS and UVCs (method, fixed), and between the CSE and Western (bioregions, fixed), as well as the effect of the interactions between these terms. PERMANOVA tests used 9999 permutations for each term included in the analysis to calculate p-values. If factors were significant ($p < 0.05$), a test of homogeneity of dispersions, PERMDISP (Anderson 2004), was run to ensure differences were due to differences between levels of a factor, instead of due to differences within levels. If significant interactions involving >2 levels were found, these were further explored using pairwise tests in PERMANOVA. A principal coordinates analysis, PCO (Gower 1966), was used to complement PERMANOVA results and to visually detect patterns in fish assemblage composition across methods and bioregions. A 2-way Similarity Percentages (SIMPER) analysis was performed on the transformed data to identify species responsible for differences in the structure of fish assemblages between sampling methods and bioregions. A species was considered to be a discriminant species for a bioregion or method when the ratio between the average dissimilarity ($\bar{\delta}$) and the standard deviation of dissimilarities ($SD \delta$) was >1 , and its percentage contribution was at least 3% (Terlizzi et al. 2005). Relationships between environmental factors and fish communities were investigated using a Distance-based Linear Model (DistLM) which calculates multivariate multiple regressions based on resemblance matrices using permutation methods (Anderson 2016). The most parsimonious model was selected with the corrected Akaike's Information Criterion (AIC_c) using 9999 permutations. To minimise multicollinearity among moderately correlated variables (Spearman correlations < 0.70) and improve results of DistLM, a full model was run that included these moderately correlated variables. From this run only the factor with the highest correlation to the relative abundance data was included in the final model. Factors identified in the

final model were then presented on PCO plots to aid in the interpretation of results. This process was performed using data from both sampling methods, and then separately for UVCs and stereo-BRUVS. All multivariate comparisons were conducted using PRIMER 6 with PERMANOVA+ (PRIMER-E).

Kernel density estimates (KDE) were used to compare length frequency distributions between sampling methods and bioregions for Galapagos sailfin grouper *Mycteroperca olfax*, snappers *Lutjanus* spp. and mullets *Mugil* spp. because of their commercial importance. Length estimates obtained through UVCs were converted to fork length (FL) using length-length conversions from FishBase (Binohlan et al. 2011). KDEs were calculated using the *sm* (Bowman & Azzalini 2018) and *KernSmooth* (Wand 2015) R packages based on a modified script developed by Langlois et al. (2012). The statistical tests applied determined if length distributions were different between methods and/or bioregions. KDE tests were applied to each taxonomic group or species separately. The plots produced by the *sm*.density.compare function from the *sm* R package include a grey polygon representing the null model of no difference between the KDEs constructed for each group under comparison. This polygon has the KDE mean as the centre and it extends to 1 SE either side of the mean, thus it allows us to visually identify the areas of the length frequency distribution that are likely behind a significant result (Langlois et al. 2012). We included a vertical line in these plots representing the approximate size at which individuals change from juveniles to adults. This line was calculated as one-third of the maximum length reported for that species, or the maximum length reported for a taxonomic group following Nagelkerken & van der Velde (2002). The maximum reported lengths were obtained from FishBase (Froese & Pauly 2019). This was done to help us identify if significant differences were driven by the presence of juvenile fish.

To assess if Galapagos mangroves are potentially being used as nursery habitats by these fishes of local economic importance, we compared the proportion of juveniles present in mangroves and adjacent rocky reefs, which were sampled as part of a monitoring campaign around the archipelago in 2014 (P. Salinas-de-León unpubl. data). This monitoring campaign was completed using Diver Operated stereo-Video Systems (stereo-DOVS). This method is similar to UVCs because divers survey a transect of a previously specified length. The difference with stereo-DOVS is that instead of recording fish abundance and estimating individual fish sizes *in situ* as in

UVCs, divers film the transect using stereo-video cameras (Harvey et al. 2002, Salinas-de-León et al. 2016, Goetze et al. 2019). The resulting footage is processed offsite using EventMeasure software to collect information on the identity, abundance and length of fishes (Goetze et al. 2019). Both methods, stereo-DOVS and UVCs, have been shown to produce similar results in species richness, relative abundance and biomass of the species being studied (Grane-Feliu et al. 2019). Thus, data obtained by these 2 methods are deemed comparable.

3. RESULTS

3.1. Abundance and diversity

A total of 35 029 individuals belonging to 92 species, 67 genera and 36 families were recorded in our survey using both sampling methods (Table 1). Fishes of local commercial importance, such as groupers, snappers and mullets, were identified as inhabitants of mangrove habitats, including a high proportion of juveniles. It is worth noting that only about 44 % of fish recorded by stereo-BRUVS were identified to species level, in large part due to poor visibility and/or the location of individual fish within the frame. Additionally, approximately 12 % of fish sampled by stereo-BRUVS were categorised as schooling silvery fish because we were unable to identify them up to family level. They were not excluded from the analysis because they represented a large proportion of the stereo-BRUVS assemblage.

There were significant differences in mean abundance between the CSE and Western bioregions, with a higher number of individuals sampled in the CSE (189 ± 29.4 SE) than in the Western bioregion (87 ± 11.4 SE) regardless of the sampling method used (Table 2A, see Fig. S1 in the Supplement at www.int-res.com/articles/suppl/m664p183

[_supp.pdf](#)). There was no significant difference in mean abundance between methods (Table 2A, Fig. S1A) despite the fact that almost twice as many individual fish were recorded using UVCs than stereo-BRUVS (Table 1).

Statistically significant interactions between the effects of sampling methods and bioregions were detected for species richness (Table 2B, Fig. S2C in the Supplement). Mean species richness was 50 % higher in the CSE regardless of the sampling method used, and stereo-BRUVS detected a higher number of species than UVCs.

Neither the Shannon-Wiener diversity index nor evenness were found to significantly differ between bioregions (Table 2, Figs. S3B & S4B in the Supplement). However, the Shannon-Wiener index was significantly higher for stereo-BRUVS (univariate PERMANOVA, Table 2C, Fig. S3A). Additionally, a statistically significant interaction between the effects of sampling methods and bioregions was detected on evenness (Table 2D, Fig. S4C). However, further inspection of the data indicates that the interaction results may have been influenced by the presence of outliers (Fig. S4). In fact, once outliers were removed this particular interaction became non-significant.

Although most fish species were recorded by both methods, there were 23 species uniquely identified by UVCs and 20 species were only recorded by stereo-BRUVS (Table S1 in the Supplement). Family diversity was found to be similar across methods and bioregions (Table 1), but there were 3 families uniquely sampled by UVCs (Cirrhitidae, Apogonidae and Sciaenidae) and 6 only recorded by stereo-BRUVS, including Scombridae, Hemiramphidae, Chanidae, Gobiidae, Centropomidae and Mullidae (Table S1). Mugilidae was the only family to contribute at least 5 % of individuals to the overall fish assemblage across methods and bioregions (Fig. 2). UVCs sampled more individuals belonging to the families of

Table 1. Summary of fish data collected by stereo-Baited Remote Underwater Video Stations (stereo-BRUVS) and Underwater Visual Census (UVC) in Galapagos mangroves during the warm season of 2015

	Stereo-BRUVS			UVC		
	CSE	Western	Total	CSE	Western	Total
Individuals	9307	2420	11727	19786	3516	23302
Species	61	42	69	66	41	72
Unique species sampled per method	19	19	20	24	18	23
Families	29	22	33	29	19	30
Unique families sampled per method	5	5	6	5	2	3

Table 2. Univariate PERMANOVA results based on Euclidean distances testing for differences in (A) abundance, (B) species richness, (C) Shannon-Wiener diversity and (D) Pielou's evenness between methods (UVC and stereo-BRUVS) and across bioregions (CSE and Western) of fish collected in Galapagos mangroves during the warm season of 2015. **Bold** values: significant

	df	SS	MS	F	R ²	p
(A) Abundance						
Bioregion	1	12.262	12.262	9.726	0.042	0.002
Method	1	3.316	3.316	2.630	0.011	0.108
Bioregion:Method	1	2.685	2.685	2.130	0.009	0.144
Residuals	218	274.830	1.261		0.938	
Total	221	293.092			1.000	
(B) Species richness						
Bioregion	1	73.700	73.730	5.716	0.018	0.017
Method	1	1243.000	1243.040	96.380	0.297	<0.001
Bioregion:Method	1	53.300	53.280	4.131	0.013	0.043
Residuals	218	2811.600	12.900		0.672	
Total	221	4181.700			1.000	
(C) Shannon-Wiener diversity						
Bioregion	1	0.523	0.523	2.126	0.009	0.144
Method	1	4.131	4.131	16.794	0.071	<0.001
Bioregion:Method	1	0.100	0.100	0.408	0.002	0.518
Residuals	218	53.626	0.246		0.919	
Total	221	58.380			1.000	
(D) Pielou's evenness						
Bioregion	1	0.010	0.010	0.460	0.002	0.506
Method	1	0.264	0.264	11.945	0.051	0.001
Bioregion:Method	1	0.094	0.094	4.263	0.018	0.039
Residuals	217	4.788	0.022		0.929	
Total	220	5.155			1.000	

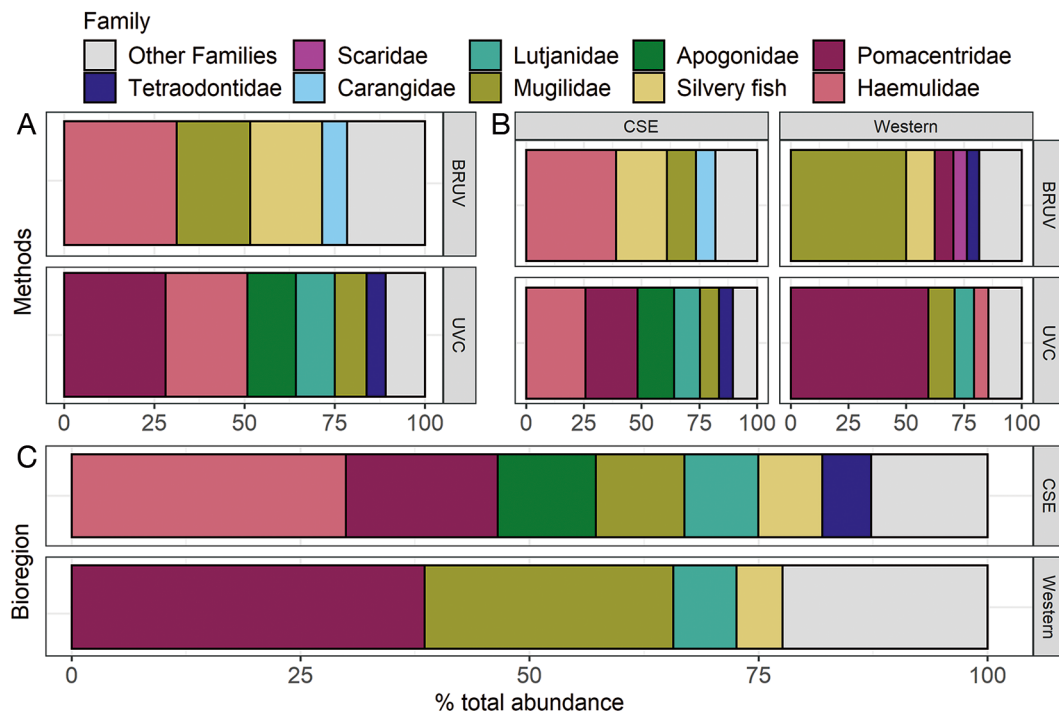


Fig. 2. Relative percentage contributions of fish families to the overall Galapagos mangrove fish assemblage: (A) per sampling method, (B) per method and bioregion, (C) and across bioregions. Families representing <5% of the total assemblage were pooled together under 'Other Families' category. BRUV: stereo-Baited Remote Underwater Video Stations; UVC: Underwater Visual Census; CSE: Central-Southeastern bioregion

Pomacentridae and Apogonidae, which together made up 41.51% of the fish assemblage sampled by this method. Haemulidae, Lutjanidae, Mugilidae and Tetraodontidae were also commonly sampled by this method and were responsible for an additional 47.62% of the fish assemblage (Fig. 2A). The most common families identified by stereo-BRUVS included Haemulidae (31.20%), Mugilidae (20.30%) and Carangidae (6.92%), with silvery fish contributing an additional 19.98% (Fig. 2A). However, dominant families varied within methods and across bioregions and in both cases, it can be seen that the overall pattern seen per method more closely resembles that of the CSE bioregion (Fig. 2B), likely because this bioregion contributes more individuals. The Western bioregion was dominated by 2 families, Mugilidae (27.14%) and Pomacentridae (38.53%) when looking at both methods together, while Lutjanidae contributed just under 7% (Fig. 2C). The most common families for the CSE included Haemulidae, Pomacentridae, Apogonidae, Mugilidae, Lutjanidae and Tetraodontidae, which together represent 80.30% of the fish community of the bioregion as sampled by both methods (Fig. 2C).

3.2. Community composition

Fish assemblage composition was significantly different between sampling methods (PERMANOVA, Table 3). These results were similar to those observed on the PCO biplot for the mean relative fish assemblage composition, which showed clear differences between sampling methods (Fig. 3A). However, it is worth noting that the first 2 PCO axes only explained approximately 31% of the variation, and thus may not reflect all patterns in our data cloud. The PCO also appeared to show that the variance was not homogenous between the 2 sampling meth-

ods, which was validated by PERMDISP results ($F_{1,40} = 12.15$, $p < 0.01$), thus confirming the existence of a dispersion effect (Anderson et al. 2008). When relative abundances from both bioregions were pooled together, the multivariate dispersion across stereo-BRUVS (43.63 ± 1.54) was larger than that from UVCs (36.39 ± 1.40).

Significant differences in fish assemblage composition were also found between bioregions (PERMANOVA, Table 3), and no differences in dispersion were detected by PERMDISP ($F_{1,40} = 1.84$, $p = 0.23$). These results were not readily apparent on the PCO plot with the pooled data, so separate plots were created for each method (Fig. 3B,C). The resulting figures support both PERMANOVA results, as they show a clear separation of fish assemblages between bioregions. Finally, the method:bioregion interaction was not significant, indicating that the effect of the method used to sample fish assemblages was consistent across bioregions.

Five taxa were identified by SIMPER as being important in differentiating fish assemblages sampled by the 2 sampling methods (Table 4A). Two species, *Lutjanus argentiventris* and *Abudefduf troschelii* were almost twice as likely to be sampled by UVCs as by stereo-BRUVS. In addition, the mean abundance recorded by UVCs for *L. novemfasciatus* was 50% more than that recorded by stereo-BRUVS. The remaining 2 taxa were *Eucinostomus* sp. and *Mugil* sp., both of which were more likely to appear on stereo-BRUVS. It is probable that these last 2 taxa were not in fact detected differently by each sampling method, but rather the resulting numbers were due to poor water clarity. This prevented us from identifying individuals to species level, thus we identified them to a coarser taxonomic level, in this case up to genus. Across bioregions, 6 taxa were found to be important in differentiating assemblages (Table 4B), which included species of local economic interest, such as *Myceteroperca olfax*, *L. novemfasciatus* and *Mugil cephalus*. *M. olfax* and *M. cephalus* showed greatest affinity for the Western bioregion, with reported mean abundances of 3.8 and 2.9 times higher than in the CSE, respectively. Mean abundances were also found to be higher in the Western bioregion for all other differentiating species, *Mugil* sp., *Scarus ghobban*, *Archosargus pourtalesii* and *L. novemfasciatus*, than in the CSE (64.20, 31.03, 28.79 and 1.30% higher, respectively).

Table 3. PERMANOVA results based on Bray Curtis distances dissimilarities of fourth root-transformed mean relative abundance data testing for differences between methods (UVC and stereo-BRUVS) and across bioregions (CSE and Western) of fish collected in Galapagos mangroves during the warm season of 2015. Significant results are highlighted in **bold**

Source	df	SS	MS	F	p (perm)	Unique
Method	1	11 129	11 129	6.936	<0.001	9908
Bioregion	1	5651	5651	3.522	<0.001	9912
Method:Bioregion	1	2375	2375	1.480	0.105	9915
Residuals	38	60 972	1605			
Total	41	81 648				

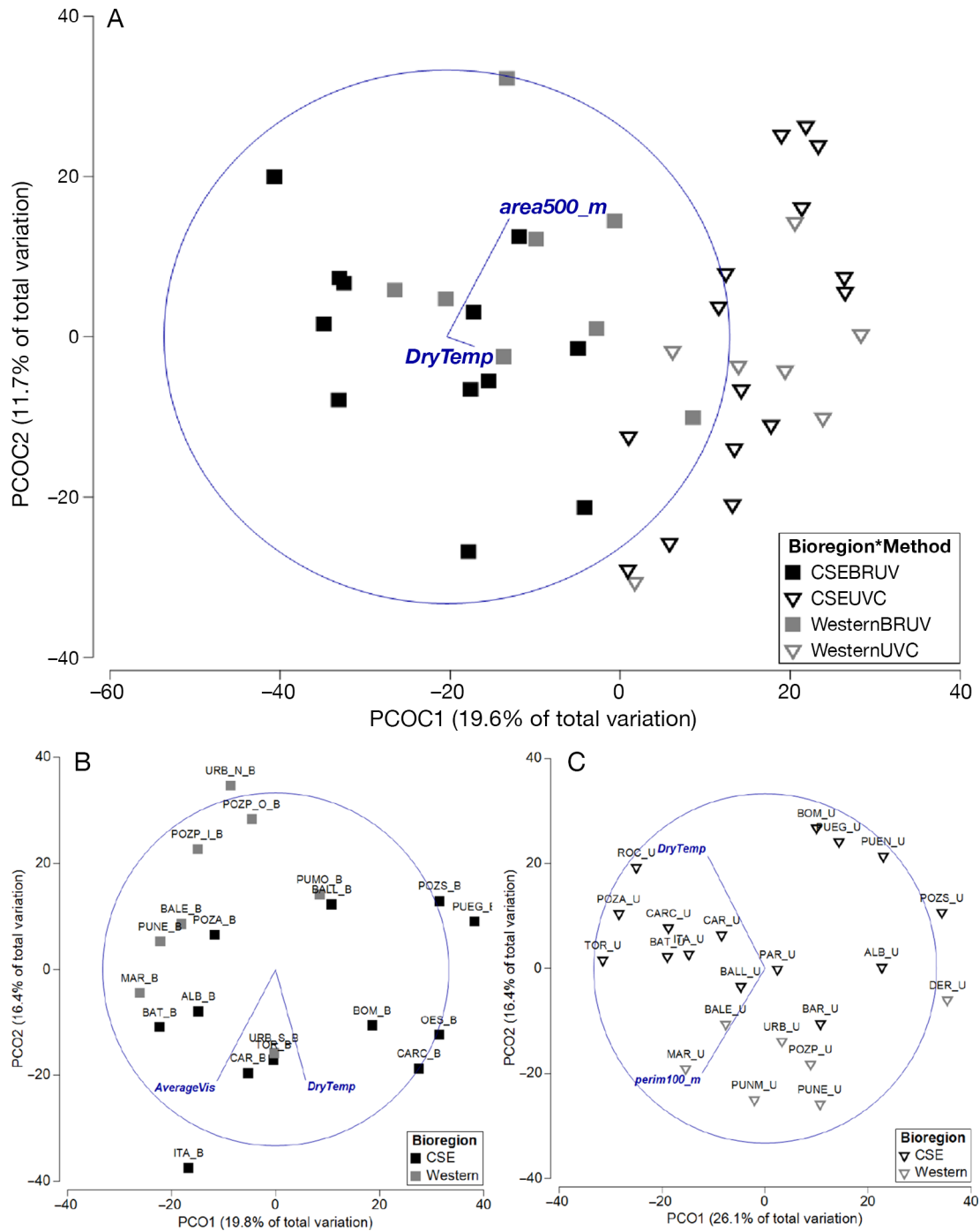


Fig. 3. Principal coordinates (PCO) biplots based on Bray Curtis dissimilarities calculated on fourth root transformed mean relative abundances of fish in mangrove areas of the Galapagos: as recorded by (A) both sampling methods, (B) stereo-Baited Remote Underwater Video Stations (BRUV) and (C) Underwater Visual Census (UVC). Environmental factors identified by distance-based linear models as having a high correlation with biotic data have been overlaid. Blue circles have a radius of one unit and their centers are the origin of the vectors representing correlations. CSE: Central-Southeastern bioregion

The best model from the DistLM analysis using the pooled relative abundance data ($AIC_c = 319.85$, $R^2 = 0.11$) was significant, but represented a poor fit. This model identified the total mangrove area within 500 m

of the site sampled ($F = 2.01$, $p = 0.01$) and mean temperature during the dry season ($F = 2.67$, $p < 0.01$) as having a significant correlation with the fish community structure (Fig. 3A). When the data was divided

Table 4. Results of 2-way SIMPER analysis of average Bray-Curtis dissimilarity calculated on fourth root transformed mean relative abundance data (A) between sampling methods (stereo-BRUVS and UVC) and (B) across bioregions (CSE and Western). Species contributing to $\geq 3\%$ of the assemblage and with a ratio >1 are considered discriminating species, and are highlighted in **bold**. $\bar{\delta}$: mean dissimilarities between groups; δ : dissimilarities between groups. Refer to section 2.4 for more information

Taxon	Mean abundance		$\bar{\delta}$	$\bar{\delta}/SD$	δ	Contribution (%)
	Stereo-BRUVS	UVC				
(A)						
<i>Eucinostomus</i> sp.	1.03	0.00	2.24	2.28	3.38	
<i>Lutjanus argentiventris</i>	0.86	1.77	2.47	1.36	3.72	
<i>Lutjanus novemfasciatus</i>	0.52	1.00	2.21	1.33	3.34	
<i>Eucinostomus dowii</i>	0.38	1.03	1.86	1.30	2.81	
<i>Abudefduf troschelii</i>	0.97	1.87	2.26	1.28	3.42	
<i>Mugil</i> sp.	1.26	0.76	2.38	1.17	3.59	
(B)						
<i>Mycteroperca olfax</i>	0.30	1.14	2.29	1.59	3.84	
<i>Lutjanus novemfasciatus</i>	0.77	0.78	2.06	1.39	3.46	
<i>Chaetodon humeralis</i>	0.56	0.13	1.20	1.30	2.02	
<i>Lutjanus argentiventris</i>	1.33	1.34	1.51	1.27	2.53	
<i>Abudefduf troschelii</i>	1.45	1.43	1.49	1.23	2.50	
<i>Archosargus pourtalesii</i>	0.66	0.85	1.80	1.21	3.02	
<i>Thalassoma lucasanum</i>	0.54	0.61	1.24	1.21	2.08	
<i>Scarus ghobban</i>	0.58	0.76	1.87	1.18	3.14	
<i>Bodianus diplotaenia</i>	0.51	0.39	1.08	1.18	1.81	
<i>Mugil</i> sp.	0.81	1.33	2.42	1.16	4.06	
<i>Sphoeroides annulatus</i>	1.44	1.19	1.54	1.13	2.59	
<i>Eucinostomus dowii</i>	0.89	0.42	1.64	1.12	2.76	
<i>Stegastes arcifrons</i>	1.46	1.79	1.62	1.06	2.72	
<i>Carcharhinus limbatus</i>	0.44	0.18	0.99	1.06	1.66	
<i>Mugil cephalus</i>	0.31	0.91	2.08	1.01	3.50	

based on sampling method, DistLM found that fish assemblages sampled by stereo-BRUVS were best explained by average visibility ($F = 2.09$, $p < 0.01$) and mean temperature during the dry season ($F = 2.16$, $p < 0.01$); however, this model ($AIC_c = 154.45$) explained just over 20% of total variation in the fish communities (Fig. 3B). Total length of the mangrove fringe within 100 m of the site sampled ($F = 2.35$, $p = 0.01$) and mean temperature during the dry season ($F = 2.53$, $p < 0.01$) were identified as factors significantly influencing fish assemblages sampled by UVCs (Fig. 3C). This model explained 22.23% of the variation in the data ($AIC_c = 160.62$).

3.3. Trophic structure

Overall, carnivores made up the largest proportion of mangrove fish assemblages in the archipelago (80.4%), followed by omnivores (12.7%) and herbivores (2.7%). UVCs detected a significantly higher mean relative abundance of carnivores than stereo-BRUVS (Fig. 4, Table S3 in the Supplement). On the

other hand, stereo-BRUVS detected significantly higher mean relative abundances of all other groups than UVCs (Table S3). Across bioregions, only herbivores were found in significantly higher proportions in the Western bioregion (Fig. 4, Table S3). The method:bioregion interaction was not found to significantly influence mean proportions in any trophic group.

3.4. Fish length frequency comparisons

Overall, juveniles in mangroves represented over 43% of all individuals sampled by both methods; however, this percentage varied among species. Mean length frequency distribution and mean FL of *Mycteroperca olfax* were similar between sampling methods and across bioregions (Fig. 5), but a larger size range was recorded by UVCs (50 cm) than by stereo-BRUVS (38 cm) (Table 5). Juveniles of this species represented 98 and 96% of the CSE and Western bioregions, respectively; both sampling methods were just as likely to detect juveniles (stereo-BRUVS 98% and UVC 97%).

Significant differences in the mean FL and length frequency distribution for *Mugil* spp. were detected (Fig. 5). Stereo-BRUVS sampled a large range of sizes for mullets, and, on average, mullets sampled by this method were almost twice as large as those detected by UVCs (Table 5). When comparing bioregions, we found a larger range of sizes and larger mean FL for mullets in the West. Juvenile mullets were found in slightly lower proportion in the Western bioregion (93%) than in the CSE (97%).

Length frequency distributions were also significantly different in their mean and distribution for *Lutjanus* spp. between methods and across bioregions (Fig. 5). UVCs detected a smaller range of sizes as well as a smaller mean FL for snappers than stereo-BRUVS (Table 5). On average snappers were 38% larger in the Western bioregion than those recorded in the CSE, and a slightly higher proportion of juveniles were found in the CSE (91%) than in the Western bioregion (88%). UVCs detected a higher proportion of juveniles (91%) than stereo-BRUVS (71%).

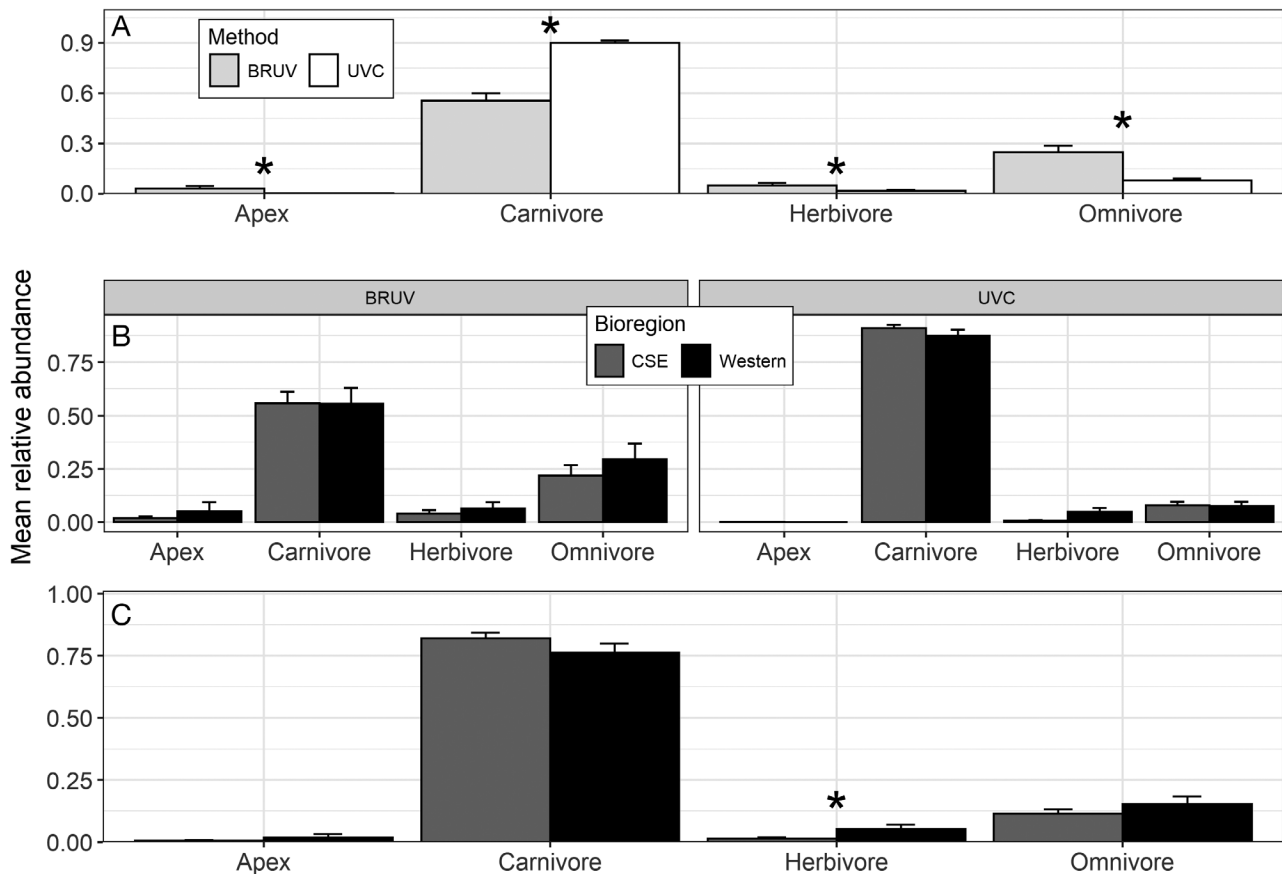


Fig. 4. Proportions of different fish trophic levels comprising mangrove fish assemblages in the Galapagos: (A) per sampling method, (B) per method and bioregion, and (C) across bioregions. Error bars show standard errors; (*) pairs where differences were detected; BRUV: stereo-Baited Remote Underwater Video Stations; UVC: Underwater Visual Census; CSE: Central-Southeastern bioregion

3.5. Juvenile proportion comparisons between mangroves and adjacent reefs

On average in mangrove areas, juveniles of *Myceteroperca olfax*, *L. aratus*, *L. argentiventris*, *L. jordani*, *L. novemfasciatus* and *Lutjanus* sp. represented approximately 85% of all individuals of the aforementioned species, while this proportion dropped to under 40% in adjacent rocky reefs (Table 6). Additionally, some of the snapper species detected in mangrove areas were not present in rocky reefs.

4. DISCUSSION

This is the first study to characterise mangrove fish assemblages at the only oceanic archipelago within the TEP where mangroves naturally occur at the edge of their biogeographical range (Giri et al. 2011). Based on our results, we can conclude that fish assemblages vary between bioregions during the

warm season. We also found that the sampling method chosen affects the end results regardless of the bioregion sampled, because they detect a different cross section of the mangrove fish community, as has been reported elsewhere (Colton & Swearer 2010, Langlois et al. 2010, Watson et al. 2010, Walsh et al. 2017). Furthermore, our results suggest mangroves in the Galapagos are used by a higher proportion of juveniles of commercially important fish families when compared to rocky reefs. This indicates that mangroves are potentially functioning as nursery habitats.

4.1. Mangrove fish assemblages

Using both sampling methods, we identified 92 species of fish, most of which were detected in low numbers. The low evenness values calculated from our observations suggest that there is a high prevalence of rare species with only a handful of common

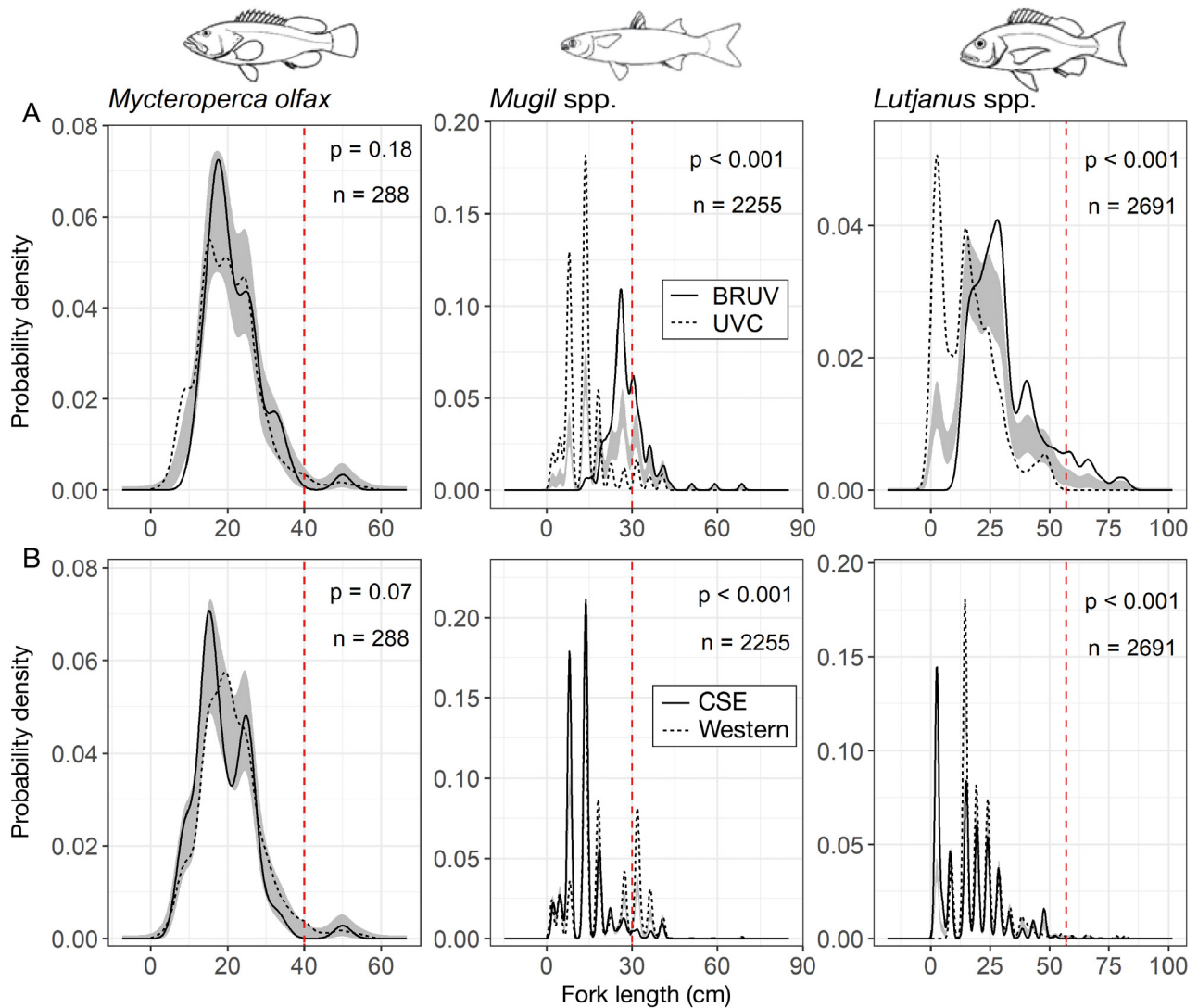


Fig. 5. Comparison of kernel density estimates (KDEs) (A) between methods and (B) across bioregions for *Mycteroperca olfax* (left), *Mugil* spp. (centre) and *Lutjanus* spp. (right) sampled in mangrove areas of the Galapagos. Red dashed line: reproductive size (i.e. juveniles are to the left and adults to the right); Grey shading: 1 SE to either side of the null model. BRUV: stereo-Baited Remote Underwater Video Stations; UVC: Underwater Visual Census; CSE: Central-Southeastern bioregion; n: number of individual fish measurements included in the KDE calculation. Differences in the location and shape of the length-frequency distributions are considered significant at $p < 0.05$ based on permutation tests

species dominating mangrove fish assemblages in the Galapagos. When comparing richness and evenness of Galapagos mangrove fish communities to values reported for other mangrove areas elsewhere in the TEP, we expected that our results would be similar to those from continental Ecuador due to the closer proximity, or northern Mexico as environmental conditions are similar to those found in the archipelago (e.g. low freshwater input, high salinity, smaller mangrove coverage). However, our species richness results were almost 3 times higher than those from northern Mexico and continental Ecuador (Castellanos-Galindo et al. 2013). Instead, we found

a similar number of species to central Mexico and Costa Rica, which included sampling sites with a much larger mangrove coverage and a much higher fishing pressure than in the Galapagos (Feutry et al. 2010, Castellanos-Galindo et al. 2013, de la Lanza Espino & Hernandez Pulido 2017, Alms & Wolff 2019). When comparing to areas outside the TEP, species richness in the Galapagos was found to be similar to that reported in subtropical mangrove systems of Southeast Asia, as well as tropical systems in both the West and East Atlantic regions, where at least 100 species of fish have been identified (Ley 2005, Nagelkerken et al. 2008, Castellanos-Galindo

Table 5. Fork lengths (cm) summary for 3 commercially important taxonomic groups of the Galapagos. Length summaries broken down by method (stereo-BRUVS and UVC) and across bioregions (CSE and Western)

	N	Mean \pm SE (min.–max.)
Stereo-BRUVS		
<i>Mycteroperca olfax</i>	55	21.70 \pm 0.96 (12.00–49.90)
<i>Mugil</i> spp.	139	28.40 \pm 0.62 (13.50–68.50)
<i>Lutjanus</i> spp.	132	31.40 \pm 1.28 (13.00–81.80)
UVC		
<i>Mycteroperca olfax</i>	233	20.40 \pm 0.53 (5.00–55.00)
<i>Mugil</i> spp.	2116	15.10 \pm 0.19 (2.00–45.00)
<i>Lutjanus</i> spp.	2559	16.10 \pm 0.25 (2.00–55.00)
CSE		
<i>Mycteroperca olfax</i>	65	19.0 \pm 0.91 (9.00–49.90)
<i>Mugil</i> spp.	1777	14.50 \pm 0.19 (2.00–59.10)
<i>Lutjanus</i> spp.	2314	16.00 \pm 0.27 (2.00–72.10)
Western		
<i>Mycteroperca olfax</i>	223	21.10 \pm 0.53 (5.00–55.00)
<i>Mugil</i> spp.	478	21.3 \pm 0.51 (2.00–68.50)
<i>Lutjanus</i> spp.	377	22.00 \pm 0.56 (9.00–81.80)

Table 6. Comparison of juvenile proportions found in mangrove areas and nearby rocky reefs. Fish data obtained from fish monitoring campaigns using Diver Operated stereo-Video Systems (stereo-DOVS) in rocky reefs across the Galapagos Marine Reserve (P. Salinas-de-León unpubl data). No individuals of the *Mugil* genus were found in rocky reefs

Species	% Juveniles	
	Mangroves	Rocky reefs
<i>Lutjanus aratus</i>	72.70	Not found
<i>Lutjanus argentiventris</i>	80.20	35.58
<i>Lutjanus jordani</i>	78.00	Not found
<i>Lutjanus novemfasciatus</i>	98.50	Not found
<i>Lutjanus</i> sp.	100.00	50.00
<i>Mycteroperca olfax</i>	84.20	32.89

& Krumme 2015). Evenness in the Galapagos had a similar pattern to that reported for northern Mexico and continental Ecuador, where few species dominate the fish assemblage (Castellanos-Galindo et al. 2013).

The reason that our results differed greatly from those previously reported for areas that are geographically close or have similar environmental conditions in the wider TEP is again likely to be the sampling method chosen. Studies using just 1 sampling method or gear type reported 50 species or less; while those using a combination of methods and/or gear type identified at least 75 species (Shervette et al. 2007, Castellanos-Galindo et al. 2013, Castellanos-Galindo & Krumme 2015, Lee et al. 2017).

There are other possible explanations, including the location of the Galapagos in a convergence zone (Banks 2002), the unique oceanographic and environmental conditions its mangroves are usually subjected to (Moity et al. 2019) and the spatio-temporal coverage of the study, or a combination of all of these factors.

Although the composition of mangrove fish communities can vary across the TEP with total mangrove coverage and as a result of local environmental conditions (Castellanos-Galindo et al. 2013), there are some families that are commonly found throughout the region, including: Lutjanidae, Tetraodontidae, Mugilidae, Gerreidae, Centropomidae, Ariidae and Clupeidae (Lacerda et al. 1993, Shervette et al. 2007, Castellanos-Galindo et al. 2013, Castellanos-Galindo & Krumme 2015). In the Galapagos mangroves, we found 4 of these families: Lutjanidae, Mugilidae, Gerreidae and Tetraodontidae, and they were among the 10 most abundant and speciose families (7 species for Lutjanidae and 5 species for each of the other 3 families). The Centropomidae family was also found in the mangroves of the archipelago, however it was uncommon, with only 3 individuals of the same species, *Centropomus viridis*, identified at Fernandina Island.

Two families commonly found in mangroves across the TEP, Ariidae and Clupeidae, were notably absent from our surveys. The Clupeidae family does occur in the Galapagos, in fact 5 genera and 6 species, 1 of which is endemic to the islands, have been reported in the archipelago (Robertson & Allen 2015). Given that most of species of the Clupeidae family are reported to occur in coastal shallow waters, often forming large schools, the schools of silvery fish detected in the stereo-BRUVS could possibly have comprised fishes of this family. Due to their distance from the camera and low visibility, we were unable to confirm this, but it is possible that this family does occupy mangroves in the Galapagos. The Ariidae family, on the other hand, has not been reported in the Galapagos, despite the fact that this family is commonly found along the entire continental coastline of the TEP, predominantly in marine and brackish waters (Castellanos-Galindo & Krumme 2015, Robertson & Allen 2015). A likely explanation for their absence in the Galapagos is the lack of brackish waters, as there are no rivers on the islands, and also because the Ariidae family prefers sheltered bays with sandy or muddy substrate (Robertson & Allen 2015), which are not a common feature of mangrove areas of the Galapagos (Moity et al. 2019). Another notable difference is the relative dominance of the

Apogonidae and Pomacentridae families in mangrove fish assemblages of the Galapagos. Although both these families have been reported in mangroves of the TEP, they are mostly associated with rocky and coral reefs (Robertson & Allen 2015). It is likely that these families are common in the Galapagos because mangroves across the archipelago mostly grow directly on lava fields (Moity et al. 2019), thus offering a hard substrate with plenty of crevices that both of these families prefer (Robertson & Allen 2015).

Carnivores were the most common trophic group found in Galapagos mangroves, which is likely the result of local conditions: low soft sediment availability and the absence or shortage of other food sources, such as detritus and benthic producers. These findings are similar to those reported for Colombian mangroves (Castellanos-Galindo & Krumme 2015).

Finally, 80% of the species observed in our study ($n = 74$) are associated with reef habitats, which suggests a strong connectivity between mangrove and reef habitats in the Galapagos, as has been reported for other parts of the world such as the Caribbean and Indo-Pacific regions (Mumby et al. 2004, Unsworth et al. 2008, Saenger et al. 2013, Nagelkerken et al. 2015). However, further research needs to be conducted to establish the strength of this potential link between these habitats.

4.2. Spatial variability of mangrove fish assemblages

Our results suggest that mangrove fish community composition was significantly different across bioregions, which are known to endure different environmental conditions (Banks 2002, Schaeffer et al. 2008). Fish communities in the CSE were found to have an average of 50% more species and a higher proportion of endemic species (17.63%) than the Western bioregion (7.06%). However, fish assemblages in the West had a significantly higher proportion of herbivores than the CSE.

We identified 6 taxa that were responsible for driving dissimilarities in community composition between bioregions. The discriminating species included 2 taxa of interest to fisheries, the Sailfin grouper *Mycteroperca olfax* and mullets *Mugil* spp., both of which had significantly higher abundances in the Western bioregion than in the CSE. This finding highlights the importance of mangroves in the western side of the archipelago in supporting local fisheries. Another group which just missed the threshold to be considered a discriminating species, but that is

worth noting, is juvenile blacktip sharks *Carcharhinus limbatus*. This species appeared to have a higher affinity for the CSE bioregion, where abundance was on average 2.4 times that recorded in the West. This result keeps in line with previous reports of juvenile *C. limbatus* as regular users of mangrove areas in the central area of the Galapagos Archipelago (Llerena-Martillo et al. 2015). *C. limbatus* is a charismatic species with a high economic value as a tourist attraction, thus this result emphasises the importance of mangroves in the CSE in supporting the local tourism industry (Llerena-Martillo et al. 2015, Lynham et al. 2015).

Among the environmental drivers of differences in community composition, mean water temperature during the dry season appeared to be one of the most influential factors, as it was identified as a significant variable by all our exploratory models (Fig. 3). This result indicates that there may be a lag effect of temperature on mangrove fish communities. Similar lags between environmental and ecological variables have been described in other marine systems across the planet (Olden & Neff 2001, Atkinson et al. 2004, Moraes et al. 2012), which further underpins the importance of assessing this lag effect of temperature on fish communities in future studies. Total mangrove area and total length of the mangrove fringe were 2 other factors identified as significant by our exploratory models, both of which have been found to be highly influential on fish communities of the TEP (Aburto-Oropeza et al. 2008, 2009). Visibility was also determined to have a significant, though minor, effect on the community structure detected by stereo-BRUVS. There are 2 potential explanations for this result: (1) visibility does in fact affect the composition of fish assemblages through a decline in visual predators present in the area (Rodríguez & Lewis 1997, Nowicki et al. 2019), or (2) it hinders the ability of researchers to detect fish in the video footage (Mallet & Pelletier 2014). In either case, we recommend further investigation of the effect of visibility of mangrove fish communities.

Although not included in our models, we suggest the influence of soft sediment availability on the fish assemblages is further investigated. Researchers involved in UVC sampling reported that the amount of soft sediment appeared to vary between bioregions. Sites on the western side of the archipelago, including Isabela and Fernandina Islands, were mostly dominated by lava fields, while sites on the eastern side of the archipelago had more soft sediment available. High variability in soft sediment has been previously reported across the archipelago, but no

clear spatial pattern has been identified (Costa et al. 2019, Moity et al. 2019). Researchers have hypothesised that variability in soft sediment availability may be the result of local bathymetry and geomorphology, which shelter coastal areas from physical disturbances caused by wave action (Costa et al. 2019). Moity et al. (2019), on the other hand, proposed that soft sediment may be related to island age, as they found that the younger islands on the western side of the archipelago had a higher proportion of lava cover above the water line than older islands to the east. Given the reported relationship between soft sediment and bioregions, we recommend that future studies include soft sediment as an abiotic factor potentially influencing fish community composition in Galapagos mangroves.

So far, our results related to differences in fish assemblages across bioregions reflect observations from previous studies of marine ecosystems of the Galapagos. However, the higher proportion of endemic species we detected in the CSE directly opposes findings by Edgar et al. (2004a), whose study was used to first define the bioregions of the Galapagos Archipelago. Edgar et al. (2004a) identified a total of 10 endemic species, 5 of which were not detected during our surveys: *Paralabrax albomaculatus*, *Lythrypnus gilberti*, *Lepidonectes corallicola*, *Acanthemblemaria castroi* and *Odontoscion eurymesops*. Of these undetected species, only *P. albomaculatus* has been reported to be a mangrove dweller, while the rest are more closely associated with rocky and coral reefs (Robertson & Allen 2015). Thus, it is reasonable to assume that the patterns detected by Edgar et al. (2004a) reflect the spatial distribution of endemic species associated with rocky reefs, and these do not necessarily replicate in mangrove habitats. We suggest that studies in other marine areas, such as the oceanic pelagic zone, are carried out to establish if the patterns in fish assemblage composition detected by Edgar et al. (2004a) are common to other marine habitats of the Galapagos.

The discrepancy in the distribution of endemics could also be explained by major differences between studies, including the habitats surveyed (mangroves vs. rocky reefs), methods used for fish sampling (UVCs and stereo-BRUVS vs. UVCs), depths sampled (<5 m vs. up to 20 m) and the temporal scale of the study (1 survey vs. multiple surveys over 18 mo). Furthermore, sampling for these studies occurred at opposing phases of the El Niño Southern Oscillation (ENSO). While Edgar et al. (2004a) sampled fish communities during a relatively cold period associated with La Niña conditions, our study took place

exclusively during the warm season and during a warm ENSO period (NOAA 2019). In fact, an El Niño event was officially declared just a few weeks after our sampling campaign was completed (Bureau of Meteorology 2015). It is likely that a combination of the factors described above resulted in significantly different environmental conditions during sampling. ENSO phases, in particular, are linked to strong changes in water temperature across the archipelago (Banks 2002), which would in turn influence the composition of fish assemblages (Attrill & Power 2002, Edgar et al. 2004a, Collie et al. 2008).

Finally, it is worth emphasising that our study investigated mangrove fish assemblage composition during daylight hours, at similar tidal stages and during the warm season. Thus, our findings should be taken as representative of these conditions, but they could be used as a base to further examine the factors that are likely having a global effect on the composition of fish communities of Galapagos mangroves. We suggest future studies examine the effect of tides, time of day (including dusk and dawn) and seasonality on assemblage composition to determine if these factors become more or less influential across time and space.

4.3. Differences between sampling methods

Our results suggest that the sampling method chosen (UVCs vs. stereo-BRUVS) determined the mangrove fish communities we observed. This effect was consistent across bioregions, with stereo-BRUVS detecting a higher number of species, as well as a more even distribution of species across all sites. When looking at average abundance per deployment/transect, stereo-BRUVS detected a slightly higher, though non-significant, abundance than UVCs, regardless of the bioregion sampled. The higher mean abundance detected by stereo-BRUVS could be due to the absence of a diver, which is known to influence results in UVCs (Mallet & Pelletier 2014). The use of bait, longer sampling time and local hydrographic conditions (e.g. tide and current speed) affect the total area of influence of the bait plume, which could also result in higher mean abundance as fish are attracted over a larger area (Taylor et al. 2013).

We also found that although the vast majority of species identified in this study were sampled by UVCs and stereo-BRUVS, there were some species uniquely identified by just 1 method (Table S1). Stereo-BRUVS consistently sampled significantly higher proportions of apex predators (i.e. sharks),

herbivores and omnivores, with mullets *Mugil* spp. detected as a discriminating species for this method (Table 4). UVCs, however, were better at detecting carnivores, in particular *Lutjanus argentiventris* and *L. novemfasciatus*, with recorded abundances twice as high in UVCs. We did not expect stereo-BRUVS to detect a lower proportion of carnivores as this contradicts previous studies (Harvey et al. 2007, Langlois et al. 2010, Andradi-Brown et al. 2016). However, previous work in marine protected areas have found that carnivorous fishes, like snappers, are actively attracted to divers (Cole et al. 1990).

We also found that variability of our data was not homogenous, with stereo-BRUVS showing a larger dispersion in comparison to UVCs. When reviewing PCO results (Fig. 3A), we saw a clear separation of data points based on the sampling method. The difference in the dispersion of the data between methods was also readily apparent in this plot. Thus, the differences detected between methods by the PERMANOVA are likely due to a combination of differences between methods and the heterogeneity of data dispersions (Anderson et al. 2008). These results confirm findings from previous studies suggesting each method chosen in this study samples a different subset of the overall population (Langlois et al. 2010, Watson et al. 2010). This finding is particularly relevant when designing monitoring strategies of mangrove fish assemblages in the future, as it shows that although each sampling method has its biases, they will remain constant regardless of differences in environmental conditions as long as the same habitat is sampled (Harvey et al. 2007).

Given that these 2 methods sampled different subsets of the mangrove fish communities, it is recommended that future sampling campaigns use them together as they will provide a more complete view of the fish assemblages under study.

4.4. Potential nursery role of Galapagos mangroves

Studies across the world have determined that structured, coastal and estuarine areas near the coast provide important nursery habitats to a variety of vertebrate and invertebrate species. These areas include seagrass meadows, marshes, kelp beds, coral reefs and mangroves (Nagelkerken et al. 2000, Beck et al. 2001, Dorenbosch et al. 2004, Lefcheck et al. 2019). Considering that most of the previously mentioned nursery habitats are not found in the Galapagos (Danulat & Edgar 2002), and that mangroves are

used as nurseries in other parts of the TEP (Shervette et al. 2007, Aburto-Oropeza et al. 2009, Ramirez-Martínez et al. 2016), we hypothesised that mangroves in the archipelago may provide nursery grounds to local fish communities. Our results support this hypothesis, with juveniles found to represent approximately 43% of all individuals sampled in our study. This juvenile proportion is similar to that reported for mangroves of tropical regions, such as the Caribbean and the Indo-Pacific (Dorenbosch et al. 2005).

We further explored the role of Galapagos mangroves as potential nursery habitats for 6 species of economic interest to local fisheries, including *Mycteroperca olfax* and 5 species of the *Lutjanus* genus, by comparing juvenile proportions between mangroves and adjacent rocky reefs. We focused on these species because together they contribute about US\$1.3 million per year to the local economy (Tanner et al. 2019), thus potentially providing an economic incentive to protect mangrove areas. In the mangrove areas, we found that approximately 85% of individuals belonging to these taxa were juveniles. This represented over twice the juvenile proportion found in nearby shallow rocky reefs (Table 6). These findings are similar to those of Aguaiza (2015), who found juveniles of *L. argentiventris* and *M. olfax* in significantly higher proportions in mangrove habitats than in nearby shallow rocky reefs.

Mangroves also appeared to work as nursery grounds for blacktip sharks *C. limbatus*. All blacktip sharks we detected in mangrove areas were considered to be juveniles based on their size. Our findings are in line with previous studies in the archipelago, which found evidence of mangroves being used as nurseries by blacktip sharks, as well as 2 more shark species: *Triaenodon obesus* and *Sphyrna lewini* (Llerena-Martillo 2009, Ketchum et al. 2014).

4.5. Management implications

The current coastal zoning plan for the Galapagos only includes waters up to 2 nautical miles from the coastline. This zoning was approved for the GMR in 2001 with the aim of allowing multiple users to coexist with minimal conflict (Heylings et al. 2002, Moity 2018). However, the 2001 plan gives full protection to <1% of the marine area inside the reserve (~1323 km²) (Moity 2018), with only ~5% of mangrove habitats receiving this full protection against fishing (Moity 2018, Moity et al. 2019). This lack of protection is a problem as mangrove bays are commonly fished by

artisanal fishers to obtain baitfish for the handline fishery, and to catch mullets and grunts for local markets (Andrade & Murillo 2002, Heel 2012, Llerena-Martillo et al. 2015). At least 26 fish species are used as bait, including at least 4 endemics, all of which are classified as either Vulnerable or Endangered by the IUCN Red List of Threatened Species (Heel 2012). Additionally, the fishing gear used has very low selectivity, thus a variety of fish species and size ranges are usually caught (Andrade & Murillo 2002, Zimmerhackel et al. 2015). This lack of selectivity is concerning because we have identified a number of species using mangroves during their juvenile stage. Poorly selective fishing may risk these juveniles being removed from populations prior to reproduction. Removal can further complicate the management of fish stocks that are already showing signs of overexploitation, including most notably *M. olfax*, a species also listed as Vulnerable by the IUCN (Burbano et al. 2014, Salinas-de-León et al. 2015, Schiller et al. 2015, Zimmerhackel et al. 2015, Usseglio et al. 2016, Eddy et al. 2019, Ramírez-González et al. 2019).

We recommend that mangrove protection should be expanded across the archipelago. This protection should include areas in the western part of the archipelago, which may be providing important nursery habitats for sailfin grouper, as well as areas in the central part of the archipelago where we have found an important proportion of juveniles for snappers and juvenile blacktip sharks, which are important species to fisheries and the tourism sector in the Galapagos. We also recorded a variety of other non-fish species and charismatic megafauna, such as green turtles *Chelonia mydas*, Galapagos penguins *Spheniscus mendiculus*, Galapagos sea lions *Zalophus wollebaeki* and the flightless cormorant *Phalacrocorax harrisi*, that actively use mangrove forests to rest, feed and breed. This provides evidence that Galapagos mangroves are an important habitat not only to a vast number of fishes, but also to some marine mammals, sea birds and invertebrates (Table S2). Our results show mangroves in the Galapagos support highly diverse fish communities, including species of local commercial and of tourism interest.

Although not a baseline study, results from this first assessment of fish communities in mangrove habitats can be used as a reference point to evaluate the effectiveness of the current zoning plan in protecting mangroves and its inhabitants, and to identify key areas (i.e. due to the provision of habitat to species of interest for fisheries and/or tourism) that are not currently protected from extractive use. This assessment

should be considered a pressing issue, as questions have been raised about the influence of some users in the design of the different management zones in the GMR (Edgar et al. 2004b).

Our work has resulted in an improvement in our understanding of a habitat that has been largely understudied in the Galapagos. Our results support the development of management strategies that aim to protect fish biodiversity and ensure the long-term sustainability of local artisanal fisheries under a changing climate (Harvey et al. 2012, Schiller et al. 2015, Salinas-de-León et al. 2020). Furthermore, identifying the environmental factors with the highest influence on the composition of fish communities could help us recognise areas that are more vulnerable to anthropogenic pressures or global climate change (Moore et al. 2016), thus worthy of additional protection measures.

Acknowledgements. We are grateful to the Charles Darwin Foundation and the Galapagos National Park Directorate for the institutional support. We are also grateful to Juan José Cota, Ismael Mascareñas, Yanella Tutivén and Cristina Aguaiza for their assistance during data collection. Special thanks to the crew of the expedition yacht 'Queen Mabel'. This study was authorised by the Galapagos National Park under permit PC-16-15 to Dr. Pelayo Salinas de León of the Charles Darwin Foundation. We are grateful to the Helmsley Charitable Trust and the Gordon and Betty Moore Foundation for providing the necessary funding for this study. Finally, we would like to thank the anonymous reviewers whose comments helped improve the manuscript. This is contribution 2379 of the Charles Darwin Research Station for the Galapagos Islands.

LITERATURE CITED

- ✦ Aburto-Oropeza O, Sala E, Paredes G, Mendoza A, Ballesteros E (2007) Predictability of reef fish recruitment in a highly variable nursery habitat. *Ecology* 88:2220–2228
- ✦ Aburto-Oropeza O, Ezcurra E, Danemann G, Valdez V, Murray J, Sala E (2008) Mangrove in the Gulf of California increase fishery yields. *Proc Natl Acad Sci USA* 105: 10456–10459
- ✦ Aburto-Oropeza O, Dominguez-Guerrero I, Cota-Nieto J, Plomozo-Lugo T (2009) Recruitment and ontogenetic habitat shifts of the yellow snapper (*Lutjanus argentiventris*) in the Gulf of California. *Mar Biol* 156: 2461–2472
- ✦ Acuña-Marrero D, Smith ANH, Salinas-de-León P, Harvey ES, Pawley MDM, Anderson MJ (2018) Spatial patterns of distribution and relative abundance of coastal shark species in the Galapagos Marine Reserve. *Mar Ecol Prog Ser* 593:73–95
- Aguaiza C (2015) The role of mangroves as nursery habitats for coral reef fish species in the Galapagos Islands. MSc thesis, University of Queensland, Brisbane
- ✦ Alms V, Wolff M (2019) The Gulf of Nicoya (Costa Rica) fisheries system: two decades of change. *Mar Coast Fish* 11: 139–161

- Alongi DM (2002) Present state and future of the world's mangrove forests. *Environ Conserv* 29:331–349
- Andersen SW, Hamann O (1986) Manglares de las Islas Galápagos. *Rev Inst Geogr Mil Ecuador* 23:101–122
- Anderson MJ (2004) PERMDISP: a FORTRAN computer program for permutational analysis of multivariate dispersions (for any two-factor ANOVA design) using permutation tests. Department of Statistics, University of Auckland, Auckland
- Anderson MJ (2005) PERMANOVA: a FORTRAN computer program for permutational multivariate analysis of variance. Department of Statistics, University of Auckland, Auckland
- Anderson MJ (2016) DISTLM v. 5: a FORTRAN computer program to calculate a distance-based multivariate analysis for a linear model. Department of Statistics, University of Auckland, Auckland
- Anderson MJ, Gorley RN, Clarke KR (2008) PERMANOVA+ for PRIMER: guide to software and statistical methods. PRIMER-E, Plymouth
- Andrade R, Murillo JC (2002) Lisas. In: Danulat E, Edgar GJ (eds) Reserva Marina de Galápagos. Línea base de la biodiversidad. Fundación Charles Darwin/Servicio Parque Nacional Galápagos, Santa Cruz, Galápagos, p 166–175
- Andradi-Brown DA, Macaya-Solis C, Exton DA, Gress E, Wright G, Rogers AD (2016) Assessing Caribbean shallow and mesophotic reef fish communities using baited-remote underwater video (BRUV) and diver-operated video (DOV) survey techniques. *PLOS ONE* 11:e0168235
- Atkinson A, Siegel V, Pakhomov EA, Rothery P (2004) Long-term decline in krill stock and increase in salps within the Southern Ocean. *Nature* 432:100–103
- Attrill MJ, Power M (2002) Climatic influence on a marine fish assemblage. *Nature* 417:275–278
- Banks S (2002) Ambiente físico. In: Danulat E, Edgar GJ (eds) Reserva Marina de Galápagos. Línea base de la biodiversidad. Fundación Charles Darwin/Servicio Parque Nacional Galápagos, Santa Cruz, Galápagos, p 22–37
- Banks S, Vera M, Chiriboga A (2009) Establishing reference points to assess long-term change in zooxanthellate coral communities of the northern Galapagos coral reefs. *Galapagos Res* 66:43–64
- Beck MW, Heck KL, Able KW, Childers DL and others (2001) The identification, conservation, and management of estuarine and marine nurseries for fish and invertebrates. *Bioscience* 51:633–641
- Binohlan C, Froese R, Pauly D, Reyes R (2011) The LENGTH-LENGTH table in FishBase. https://www.fishbase.in/manual/english/PDF/FB_Book_CBinohlan_Length-Length_RF_JG.pdf (accessed 27 July 2020)
- Bowman AW, Azzalini A (2018) R package 'sm': nonparametric smoothing methods. <https://cran.r-project.org/web/packages/sm/sm.pdf> (accessed 24 June 2020)
- Burbano DV, Mena CF, Guarderas P, Vinueza L, Reck G (2014) Shifting baselines in the Galapagos white fin fishery, using fisher's anecdotes to reassess fisheries management: the case of the Galapagos grouper. In: Denkinger J, Vinueza L (eds) The Galapagos Marine Reserve: a dynamic social-ecological system. Springer, New York, p 227–246
- Bureau of Meteorology (2015) El Niño in the tropical Pacific. www.bom.gov.au/climate/enso/archive/ensowrap_20150512.pdf (accessed 12 November 2019)
- Cappo M, Harvey ES, Malcolm HA, Speare P (2003) Potential of video techniques to monitor diversity, abundance and size of fish in studies of Marine Protected Areas. In: Beumer J, Grant A, Smith D (eds) Aquatic protected areas—what works best and how do we know? Proc World Congress on Aquatic Protected Areas, August 2002, Cairns. Australian Society for Fish Biology, Perth, WA, p 455–464
- Castellanos-Galindo GA, Krumme U (2015) Tides, salinity, and biogeography affect fish assemblage structure and function in macrotidal mangroves of the neotropics. *Ecosystems* 18:1165–1178
- Castellanos-Galindo GA, Krumme U, Rubio EA, Saint-Paul U (2013) Spatial variability of mangrove fish assemblage composition in the tropical eastern Pacific Ocean. *Rev Fish Biol Fish* 23:69–86
- Chowdhury RR, Uchida E, Chen L, Osorio V, Yoder L (2017) Anthropogenic drivers of mangrove loss: geographic patterns and implications for livelihoods. In: Rivera-Monroy VH, Lee SY, Kristensen E, Twilley RR (eds) Mangrove ecosystems: a global biogeographic perspective. Springer International Publishing, Cham, p 275–300
- Clarke KR, Warwick RM (2001) Change in marine communities: an approach to statistical analysis and interpretation. PRIMER-E, Plymouth
- Cole RG, Ayling TM, Creese RG (1990) Effects of marine reserve protection at Goat Island, northern New Zealand. *NZ J Mar Freshw Res* 24:197–210
- Collie JS, Wood AD, Jeffries HP (2008) Long-term shifts in the species composition of a coastal fish community. *Can J Fish Aquat Sci* 65:1352–1365
- Colton MA, Swearer SE (2010) A comparison of two survey methods: differences between underwater visual census and baited remote underwater video. *Mar Ecol Prog Ser* 400:19–36
- Costa MT, Salinas-de-León P, Aburto-Oropeza O (2019) Storage of blue carbon in isolated mangrove forests of the Galapagos' rocky coast. *Wetlands Ecol Manage* 27: 455–463
- Currie DJ, Mittelbach GG, Cornell HV, Field R and others (2004) Predictions and tests of climate-based hypotheses of broad-scale variation in taxonomic richness. *Ecol Lett* 7:1121–1134
- Danulat E, Edgar GJ (eds) (2002) Reserva Marina de Galápagos. Línea base de la biodiversidad. Fundación Charles Darwin, Servicio Parque Nacional Galápagos, Santa Cruz, Galápagos
- de la Lanza Espino G, Hernandez Pulido S (2017) Natural and induced space/time environmental changes in the Teacapán-Agua Brava Lagoon System, NW Mexico. *J Aquac Mar Biol* 5:00140
- Doney SC, Ruckelshaus M, Emmett Duffy J, Barry JP and others (2012) Climate Change Impacts on Marine Ecosystems. *Annu Rev Mar Sci* 4:11–37
- Dorenbosch M, van Riel MC, Nagelkerken I, van der Velde G (2004) The relationship of reef fish densities to the proximity of mangrove and seagrass nurseries. *Estuar Coast Shelf Sci* 60:37–48
- Dorenbosch M, Grol MGG, Christianen MJA, Nagelkerken I, van der Velde G (2005) Indo-Pacific seagrass beds and mangroves contribute to fish density and diversity on adjacent coral reefs. *Mar Ecol Prog Ser* 302:63–76
- Duke NC (2017) Mangrove floristic and biogeography revisited: further deductions from biodiversity hot spots, ancestral discontinuities, and common evolutionary pro-

- cesses. In: Rivera-Monroy VH, Lee SY, Kristensen E, Twilley RR (eds) Mangrove ecosystems: a global biogeographic perspective. Springer International Publishing, Cham, p 17–53
- ✦ Eddy TD, Friedlander AM, Salinas-de-León P (2019) Ecosystem effects of fishing & El Niño at the Galapagos Marine Reserve. *PeerJ* 7:e6878
- ✦ Edgar GJ, Banks S, Fariña JM, Calvopiña M, Martínez C (2004a) Regional biogeography of shallow reef fish and macro-invertebrate communities in the Galapagos archipelago. *J Biogeogr* 31:1107–1124
- ✦ Edgar GJ, Bustamante RH, Fariña JM, Calvopiña M, Martínez C, Toral-Granda MV (2004b) Bias in evaluating the effects of marine protected areas: the importance of baseline data for the Galapagos Marine Reserve. *Environ Conserv* 31:212–218
- ✦ Feutry P, Hartmann HJ, Casabonnet H, Umaña G (2010) Preliminary analysis of the fish species of the Pacific Central American mangrove of Zancudo, Golfo Dulce, Costa Rica. *Wetlands Ecol Manage* 18:637–650
- Froese R, Pauly D (2019) FishBase. www.fishbase.org (accessed 2 July 2020)
- ✦ Giri C, Ochieng E, Tieszen LL, Zhu Z and others (2011) Status and distribution of mangrove forests of the world using earth observation satellite data. *Glob Ecol Biogeogr* 20:154–159
- ✦ Gladstone W, Lindfield S, Coleman M, Kelaher B (2012) Optimisation of baited remote underwater video sampling designs for estuarine fish assemblages. *J Exp Mar Biol Ecol* 429:28–35
- ✦ Goetze JS, Bond T, McLean DL, Saunders BJ and others (2019) A field and video analysis guide for diver operated stereo-video. *Methods Ecol Evol* 10:1083–1090
- ✦ Gower JC (1966) Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika* 53:325–338
- Grane-Feliu X, Bennett S, Hereu B, Aspillaga E, Santana-Garcón J (2019) Comparison of diver operated stereo-video and visual census to assess targeted fish species in Mediterranean marine protected areas. *J Exp Mar Biol Ecol* 520:151205
- Grove JS, Lavenberg RJ (1997) The fishes of the Galapagos Islands. Stanford University Press, Stanford, CA
- Harvey ES, Shortis M (1998) Calibration stability of an underwater stereo-video system: implications for measurement accuracy and precision. *Mar Technol Soc J* 32: 3–17
- ✦ Harvey ES, Fletcher D, Shortis M (2002) Estimation of reef fish length by divers and by stereo-video: a first comparison of the accuracy and precision in the field on living fish under operational conditions. *Fish Res* 57:255–265
- ✦ Harvey ES, Cappo M, Butler JJ, Hall N, Kendrick GA (2007) Bait attraction affects the performance of remote underwater video stations in assessment of demersal fish community structure. *Mar Ecol Prog Ser* 350: 245–254
- ✦ Harvey ES, Newman SJ, McLean DL, Cappo M, Meeuwig JJ, Skepper CL (2012) Comparison of the relative efficiencies of stereo-BRUVs and traps for sampling tropical continental shelf demersal fishes. *Fish Res* 125-126: 108–120
- Heel LC (2012) Assessment of a bait fishery in the Galapagos Marine Protected Area: implications for sustainable management. Leuphana University, Lüneburg
- Heylings P, Bensted-Smith R, Altamirano M (2002) Zonificación e historia de la reserva marina de Galápagos. In: Danulat E, Edgar GJ (eds) Reserva Marina de Galápagos. Línea base de la biodiversidad. Fundación Charles Darwin/Servicio Parque Nacional Galápagos, Santa Cruz, Galápagos, p 10–21
- INOCAR (2017) Tabla de mareas puertos del Ecuador. www.inocar.mil.ec/web/index.php/productos/tabla-mareas (accessed January 2017)
- Jennerjahn TC, Gilman E, Krauss KW, Lacerda LD, Nordhaus I, Wolanski E (2017) Mangrove ecosystems under climate change. In: Rivera-Monroy VH, Lee SY, Kristensen E, Twilley RR (eds) Mangrove ecosystems: a global biogeographic perspective. Springer International Publishing, Cham, p 211–244
- ✦ Ketchum JT, Hearn AR, Klimley AP, Peñaherrera C and others (2014) Inter-island movements of scalloped hammerhead sharks (*Sphyrna lewini*) and seasonal connectivity in a marine protected area of the eastern tropical Pacific. *Mar Biol* 161:939–951
- Lacerda LD, Conde JE, Alarcon C, Alvarez-León R and others (1993) Mangrove ecosystems of Latin America and the Caribbean: A summary. In: Lacerda LD (ed) Conservation and sustainable utilization of mangrove forests in Latin America and Africa regions. International Society for Mangrove Ecosystems and International Tropical Timber Organization Project PD114/90(F), Yokohama
- ✦ LaDeau SL, Han BA, Rosi-Marshall EJ, Weathers KC (2017) The next decade of big data in ecosystem science. *Ecosystems* 20:274–283
- ✦ Langlois TJ, Harvey ES, Fitzpatrick B, Meeuwig JJ, She-drawi G, Watson DL (2010) Cost-efficient sampling of fish assemblages: comparison of baited video stations and diver video transects. *Aquat Biol* 9:155–168
- ✦ Langlois TJ, Fitzpatrick BR, Fairclough DV, Wakefield CB and others (2012) Similarities between line fishing and baited stereo-video estimations of length-frequency: novel application of kernel density estimates. *PLOS ONE* 7:e45973
- Langlois TJ, Williams J, Monk J, Bouchet P and others (2018) Marine sampling field manual for benthic stereo BRUVS (baited remote underwater video). In: Przeslawski R, Foster S (eds) Field manuals for marine sampling to monitor Australian waters, Version 1. National Environmental Science Programme (NESP), Marine Biodiversity Hub, Canberra, p 82–104
- Lee SY, Jones EBG, Diele K, Castellanos-Galindo GA, Nordhaus I (2017) Biodiversity. In: Rivera-Monroy VH, Lee SY, Kristensen E, Twilley RR (eds) Mangrove ecosystems: a global biogeographic perspective. Springer, Cham, p 55–86
- Lefcheck JS, Hughes BB, Johnson AJ, Pfirrmann BW and others (2019) Are coastal habitats important nurseries? A meta-analysis. *Conserv Lett* 12:e12645
- ✦ Ley JA (2005) Linking fish assemblages and attributes of mangrove estuaries in tropical Australia: criteria for regional marine reserves. *Mar Ecol Prog Ser* 305: 41–57
- Llerena-Martillo Y (2009) Identificación de tiburones juveniles y caracterización de sus hábitats en las zonas costeras de pesca de la isla San Cristóbal - Reserva Marina de Galápagos. Bióloga, Universidad de Guayaquil, Guayaquil
- Llerena-Martillo Y, Peñaherrera-Palma C, Espinoza E, Hirschfeld M, Wolff M, Vinuesa L (2015) Áreas de cri-

- anza de tiburones punta negra (*Carcharhinus limbatus*) en zonas de manglar en la parte central del archipiélago de Galápagos. Informe Galápagos 2013-2014. DPNG, CGREG, FCD y GC, Puerto Ayora, Galápagos
- ✦ Llerena-Martillo Y, Peñaherrera-Palma C, Espinoza ER (2018) Fish assemblages in three fringed mangrove bays of Santa Cruz Island, Galapagos Marine Reserve. *Rev Biol Trop* 66:674–687
- López-Portillo J, Lewis RR, Saenger P, Rovai A and others (2017) Mangrove forest restoration and rehabilitation. In: Rivera-Monroy VH, Lee SY, Kristensen E, Twilley RR (eds) *Mangrove ecosystems: a global biogeographic perspective*. Springer International Publishing, Cham, p 301–345
- Lynham J, Costello C, Gaines S, Sala E (2015) Economic valuation of marine- and shark-based tourism in the Galapagos Islands. *National Geographic Pristine Seas*, Puerto Ayora, Galápagos
- ✦ Mallet D, Pelletier D (2014) Underwater video techniques for observing coastal marine biodiversity: a review of sixty years of publications (1952–2012). *Fish Res* 154:44–62
- ✦ Moity N (2018) Evaluation of no-take zones in the Galapagos Marine Reserve, zoning plan 2000. *Front Mar Sci* 5: 244
- ✦ Moity N, Delgado B, Salinas-de-León P (2019) Mangroves in the Galapagos islands: distribution and dynamics. *PLOS ONE* 14:e0209313
- ✦ Moore CH, Radford BT, Possingham HP, Heyward AJ and others (2016) Improving spatial prioritisation for remote marine regions: optimising biodiversity conservation and sustainable development trade-offs. *Sci Rep* 6:32029
- ✦ Moraes LE, Paes E, Garcia A, Möller O Jr, Vieira J (2012) Delayed response of fish abundance to environmental changes: a novel multivariate time-lag approach. *Mar Ecol Prog Ser* 456:159–168
- ✦ Mumby PJ, Edwards AJ, Arias-González JE, Lindeman KC and others (2004) Mangroves enhance the biomass of coral reef fish communities in the Caribbean. *Nature* 427:533–536
- ✦ Murphy HM, Jenkins GP (2010) Observational methods used in marine spatial monitoring of fishes and associated habitats: a review. *Mar Freshw Res* 61:236–252
- ✦ Nagelkerken I, van der Velde G (2002) Do non-estuarine mangroves harbour higher densities of juvenile fish than adjacent shallow-water and coral reef habitats in Curaçao (Netherlands Antilles)? *Mar Ecol Prog Ser* 245: 191–204
- ✦ Nagelkerken I, van der Velde G, Gorissen MW, Meijer GJ, Van't Hof T, den Hartog C (2000) Importance of mangroves, seagrass beds and the shallow coral reef as a nursery for important coral reef fishes, using a visual census technique. *Estuar Coast Shelf Sci* 51:31–44
- ✦ Nagelkerken I, Blaber SJM, Bouillon S, Green P and others (2008) The habitat function of mangroves for terrestrial and marine fauna: a review. *Aquat Bot* 89:155–185
- ✦ Nagelkerken I, Sheaves M, Baker R, Connolly RM (2015) The seascape nursery: a novel spatial approach to identify and manage nurseries for coastal marine fauna. *Fish Fish* 16:362–371
- NOAA (2019) Cold & warm episodes by season. https://origin.cpc.ncep.noaa.gov/products/analysis_monitoring/ensostuff/ONL_v5.php (accessed 12 November 2019)
- NOAA NMFS SWFSC ERD (2017a) Chlorophyll-a, Aqua MODIS, NPP, 0.025°, Pacific Ocean, EXPERIMENTAL, 2006-present (1 Day Composite), Lon+/-180. https://coastwatch.pfeg.noaa.gov/erddap/griddap/erdMBchla1day_LonPM180.html
- NOAA NMFS SWFSC ERD (2017b) SST, Aqua MODIS, NPP, 0.025 degrees, Pacific Ocean, Daytime, 2006-present (1 Day Composite). https://coastwatch.pfeg.noaa.gov/erddap/griddap/erdMBSstd1day_LonPM180.html
- Nowicki R, Heithaus M, Thomson J, Burkholder D, Gastrich K, Wirsing A (2019) Indirect legacy effects of an extreme climatic event on a marine megafaunal community. *Ecol Monogr* 89:e01365
- ✦ Oksanen J, Blanchet FG, Friendly M, Kindt R and others (2019) *vegan: Community Ecology Package*. <https://cran.r-project.org/web/packages/vegan/vegan.pdf> (accessed 24 June 2020)
- ✦ Olden JD, Neff BD (2001) Cross-correlation bias in lag analysis of aquatic time series. *Mar Biol* 138:1063–1070
- ✦ Palacios DM, Bograd SJ, Foley DG, Schwing FB (2006) Oceanographic characteristics of biological hot spots in the North Pacific: a remote sensing perspective. *Deep Sea Res II* 53:250–269
- Palomares ML (2000) The ECOLOGY table. In: Froese R, Pauly D (eds) *FishBase 2000: concepts, design and data sources*. ICLARM, Los Baños, Laguna
- QGIS Development Team (2019) QGIS Geographic Information System. Open Source Geospatial Foundation Project
- R Core Team (2020) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Ramírez-González J, Marín Jarrín JR, Andrade-Vera S, Tanner MK, Salinas-de-León P, Barragán MJ (2019) ¿Cómo lograr pesquerías sustentables de peces en Galápagos? In: Hojnowski C, Cruz D (eds) *Informe Galápagos 2017-2018*. Galapagos Conservancy, Fairfax
- ✦ Ramirez-Martínez GA, Castellanos-Galindo GA, Krumme U (2016) Tidal and diel patterns in abundance and feeding of a marine-estuarine-dependent fish from macrotidal mangrove creeks in the Tropical Eastern Pacific (Colombia). *Estuaries Coasts* 39:1249–1261
- Robertson DR, Allen GR (2015) *Shorefishes of the Tropical Eastern Pacific: online information system*. Version 2.0 Smithsonian Tropical Research Institute, Balboa. <http://biogeodb.stri.si.edu/sfstep/en/pages> (accessed June 2020)
- ✦ Robertson DR, Cramer KL (2009) Shore fishes and biogeographic subdivisions of the Tropical Eastern Pacific. *Mar Ecol Prog Ser* 380:1–17
- ✦ Rodríguez MA, Lewis WM (1997) Structure of fish assemblages along environmental gradients in floodplain lakes of the Orinoco River. *Ecol Monogr* 67:109–128
- Saenger P, Gartside D, Funge-Smith S (2013) A review of mangrove and seagrass ecosystems and their linkage to fisheries and fisheries management. Food and Agriculture Organization of the United Nations Regional Office for Asia and the Pacific, Bangkok
- ✦ Salinas-de-León P, Rastoin E, Acuña-Marrero D (2015) First record of a spawning aggregation for the tropical eastern Pacific endemic grouper *Mycteroperca olfax* in the Galapagos Marine Reserve. *J Fish Biol* 87:179–186
- ✦ Salinas-de-León P, Acuña-Marrero D, Rastoin E, Friedlander AM, Donovan MK, Sala E (2016) Largest global shark biomass found in the northern Galapagos Islands of Darwin and Wolf. *PeerJ* 4:e1911
- ✦ Salinas-de-León P, Andrade S, Arnés-Urgellés C, Bermudez JR and others (2020) Evolution of the Galapagos in the Anthropocene. *Nat Clim Chang* 10:380–382

- Schaeffer BA, Morrison JM, Kamykowski D, Feldman GC and others (2008) Phytoplankton biomass distribution and identification of productive habitats within the Galapagos Marine Reserve by MODIS, a surface acquisition system, and in-situ measurements. *Remote Sens Environ* 112:3044–3054
- Schiller L, Alava JJ, Grove J, Reck G, Pauly D (2015) The demise of Darwin's fishes: Evidence of fishing down and illegal shark finning in the Galapagos Islands. *Aquat Conserv* 25:431–446
- Schloerke B, Crowley J, Cook D, Briatte F and others (2018) GGally: Extension to 'ggplot2'. <https://cran.r-project.org/web/packages/GGally/GGally.pdf> (accessed 24 June 2020)
- Shervette VR, Aguirre WE, Blacio E, Cevallos R, Gonzalez M, Pozo F, Gelwick F (2007) Fish communities of a disturbed mangrove wetland and an adjacent tidal river in Palmar, Ecuador. *Estuar Coast Shelf Sci* 72: 115–128
- Skiena SS (2017) *The data science design manual*. Springer Nature, Cham
- Snell HM, Stone PA, Snell HL (1996) A summary of geographical characteristics of the Galapagos Islands. *J Biogeogr* 23:619–624
- Suárez-Moncada JM (2012) *Comunidades biológicas en los manglares de Isabela, Santa Cruz y San Cristóbal*. Bióloga, Universidad de Guayaquil, Guayaquil
- Tanner MK, Moity N, Costa MT, Marin Jarrin JR, Aburto-Oropeza O, Salinas-de-León P (2019) Mangroves in the Galapagos: ecosystem services and their valuation. *Ecol Econ* 160:12–24
- Taylor MD, Baker J, Suthers IM (2013) Tidal currents, sampling effort and baited remote underwater video (BRUV) surveys: are we drawing the right conclusions? *Fish Res* 140:96–104
- Terlizzi A, Benedetti-Cecchi L, Bevilacqua S, Frascchetti S, Guidetti P, Anderson MJ (2005) Multivariate and univariate asymmetrical analyses in environmental impact assessment: A case study of Mediterranean subtidal sessile assemblages. *Mar Ecol Prog Ser* 289:27–42
- Thakur KK, Vanderstichel R, Barrell J, Stryhn H, Patanasatienkul T, Revie CW (2018) Comparison of remotely-sensed sea surface temperature and salinity products with *in situ* measurements from British Columbia, Canada. *Front Mar Sci* 5:121
- Tittensor DP, Mora C, Jetz W, Lotze HK, Ricard D, Berghe EV, Worm B (2010) Global patterns and predictors of marine biodiversity across taxa. *Nature* 466:1098–1101
- Unsworth RKF, Salinas-de-León P, Garrard SL, Jompa J, Smith DJ, Bell JJ (2008) High connectivity of Indo-Pacific seagrass fish assemblages with mangrove and coral reef habitats. *Mar Ecol Prog Ser* 353:213–224
- Usseglio P, Friedlander AM, Koike H, Zimmerhackel J, Schuhbauer A, Eddy T, Salinas-de-León P (2016) So long and thanks for all the fish: overexploitation of the regionally endemic Galapagos grouper *Mycteroperca olfax* (Jenyns, 1840). *PLOS ONE* 11:e0165167
- Visbeck M (2018) Ocean science research is key for a sustainable future. *Nat Commun* 9:690
- Walsh AT, Barrett NS, Hill NA (2017) Efficacy of baited remote underwater video systems and bait type in the cool-temperature zone for monitoring 'no-take' marine reserves. *Mar Freshw Res* 68:568–580
- Wand M (2015) KernSmooth: functions for kernel smoothing supporting Wand & Jones (1995). <https://cran.r-project.org/web/packages/KernSmooth/KernSmooth.pdf> (accessed 24 June 2020)
- Watson DL, Harvey ES, Fitzpatrick BM, Langlois TJ, She-drawi G (2010) Assessing reef fish assemblage structure: how do different stereo-video techniques compare? *Mar Biol* 157:1237–1250
- Wei T, Simko V (2017) R package 'corrplot': visualization of a correlation matrix (Version 0.84). <https://cran.r-project.org/web/packages/corrplot/corrplot.pdf> (accessed 24 June 2020)
- Wellington GM (1975) *Medio ambientes marinos costeros de Galápagos: un informe de recursos al Departamento de Parques Nacionales y Vida Silvestre*. Departamento de Parques Nacionales y Vida Silvestre, Quito
- Zimmerhackel JS, Schuhbauer AC, Usseglio P, Heel LC, Salinas-de-León P (2015) Catch, bycatch and discards of the Galapagos Marine Reserve small-scale handline fishery. *PeerJ* 3:e995
- zu Ermgassen PSE, Grove T, Nagelkerken I (2020) Global affiliation of juvenile fishes and invertebrates with mangrove habitats. *Bull Mar Sci* 96:403–414

Editorial responsibility: Alejandro Gallego, Aberdeen, UK
Reviewed by: 3 anonymous referees

Submitted: November 12, 2019
Accepted: January 4, 2021
Proofs received from author(s): March 28, 2021