

Status of AM Fungi in Some Medicinal Plants from Panambur Beach Mangalore India

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Abstract: Present study was undertaken to check the present status of AM fungi in four medicinal plants found in costal sand dune (CSD) habitat, from 'Panambur beach' situated in Dakshina Kannada District near Mangalore city, Karnataka India (12° 57' 17" North, 74° 48' 17" East). AM fungal species associated with medicinal plants were belonging to three genera viz., *Acaulospora*, *Gigaspora* and *Glomus* representing 12 species.

Keywords: AM fungi, Coastal Sand dune, CSD, medicinal plants

I. Introduction

The costal biota is under serious threat from human activities that lead to destruction of natural ecosystem at an alarming rate. These has phenomenal increase in vegetation loss during last few decades. The costal regions harbour many economic and medicinal plant species [3] [5]. The local community surrounding the coast depends upon these resources for their livelihood. They collect and utilize many plants for food, timber, fiber, fuel, and medicine. Their high utility, i.e. economic and medicinal, is an important contributing factor to their over exploitation. The continual use of costal plants over many years without these being replanted has resulted in accelerated decline in the abundance or the loss of a wide range of threatened and endangered species [4]. Herbal and traditional medicine in global market is fetching approximately US \$ 60 billion per annum with India's share about US \$ 400-500 million [15]. Due to biopiracy threat, detailed documentation of the untapped costal sand dunes (CSD) resources is of immense value. A wide range of microbes associated with belowground root biomass facilitates the dune stabilization [2]. Amongst the belowground root microbes Arbuscular mycorrhizal fungi (AM) are very significant for uptake of water and nutrients, especially in phosphorus deficient condition, and also help plant establishment in very harsh environments [22]. AM fungi are known to significantly contribute to the development of plant community structure and sand dune stabilization. Coastal sand dunes are of great ecological significance. They act as barriers against the action of waves and tides [1]. Hence, it is necessary to draw attention toward the association of AM fungi and CSD medicinal plants despite of the disturbed or undisturbed nature of dunes.

Most studies on AM fungi of coastal dunes are confined to temperate regions, and subtropical regions [17], a few were performed from the tropical coast of Hawaiian Islands [19] [21], India [12] [13] [24] [25] [26] [27] and Singapore [6]. In last decades, Panambur beach was explored for AM fungal colonization, species richness and diversity associated with *Ipomoea pes-caprae* (L.) R. Br. (with less vegetation) in relation to rhizosphere edaphic features [13]. This study was carried out on severely disturbed dunes because of recreation and harbor activities on west coast of India particularly adjoining sites of Panambur beach. However, costal zone protection initiatives made in past few years has showed good results and this beach is now well maintained. Medicinal efficacy of more frequently growing beach plants viz., *Canavalia rosea* (Sw.) DC (Fabaceae), *I. pes-caprae* (Convolvulaceae), *Launaea sermentosa* (Willd.) Schult-Bip. ex O. Kuntze (Asteraceae) and *Phyla nodiflora* (L) Greene (Verbenaceae) is now well studied. A hot water infusion of the leaves of *C. rosea* with other used in Tonga to treat secondary amenorrhoea postpartum hemorrhage [28]. *I. pes-caprae* leaves are usually used in colic, bedsores and in the treatment of ulcers [23]. It is also used in rheumatism and diuretic [10], gonorrhoea skin diseases and stomachache [4]. *L. sermentosa*, whole plant juice is applied for the treatment of rheumatism [4], it also exhibit antimicrobial property [7] and used in renal disorders [18]. Fresh plant paste or poultice of *P. nodiflora* is applied for boils, swollen cervical glands and chronic indolent ulcers [4]. Thus these are some of the important medicinal plants of sandy beach habitat. Although, AM fungal study was made with *I. pes-caprae* from Panambur beach previously [13], in present study an attempt has made to check present status AM fungi in these medicinal plants found in addition to *I. pes-caprae*.

II. Material And Methods

2.1 AM fungal colonization status

Root samples of medicinal plant species distantly growing approximately 100 meter away from each other were collected from Panambur Beach during July-Sept 2011 and brought to the laboratory. Roots were made free from sand and debris to process and stain [8] [20]. Stained roots were observed under a binocular microscope to evaluate mycorrhizal colonization [14].

2.2 AM fungal species identification

Spores were isolated from Rhizospheric sand of plant with the help of wet sieving and decanting method [9]. Spores with the same morphology were mounted in water, in polyvinyl-lactoglycerol (PVLG) and in PVLG with a drop of Mezler's reagent for microscopic examination. Spores mounted in Mezler's reagent were crushed in order to observe the staining of the different spore wall layers. At least 20 spores of each of the different morphotypes found were mounted in PVLG and 10 spores mounted in PVLG + Melzer's reagent for morphological identification after the original descriptions [16].

III. Results And Discussion

AM fungi were encountered in the roots of all plants studied from Panambur beach (Table 1, Fig. 1). All the mycorrhizal components (vesicles, arbuscules and hyphae) were found with variation in occurrence intensity in all plants viz., *C. rosea*, *I. pes-caprae*, *L. sermentosa* and *P. nodiflora*. Earlier studies reveals that, *L. sermentosa* exhibited 45% mycorrhizal colonization and 10 AM spores 100 g^{-1} soil from Someshwara West Coast of India [11]. However, in present investigation we have reported 98% colonization and 80 AM spores 100 g^{-1} soil in *L. sermentosa*. *I. pes-caprae* showed drastic improvement in colonization percentage (100%) in comparison with previous study (77.8%) [13]. The spore count in rhizospheric sample was remarkable i.e., total 242 AM spore 100 g^{-1} soil were recorded. The observed AM spore species associated with *I. pes-caprae* agree with earlier findings [13]. However, disappearance of some species was also noticed. The degree of AM fungal colonization was found almost closely similar in some of the plant species such as 98% colonization in *C. rosea* and *L. sermentosa* while *I. pes-caprae* and *P. nodiflora* exhibit 100% colonization. Whereas, earlier study [13] showed that *Canavalia cathartica* and *C. maritime* were colonized at the rate of 83% and 24% respectively from Padubidri region of West coast of India. Although, *P. nodiflora* from Padubidri site in previous study [13] was reported non mycorrhizal, our observations proved that it has mycorrhizal association in present study area which is having similar geographical and climatic conditions like Padubidri. This study indicates the excellent pervasiveness of AM fungi is established in the root system of all four medicinal plants.

In present study, AM fungal species observed in rhizosphere samples of CSD medicinal plants were belonging to three genera viz., *Acaulospora*, *Gigaspora* and *Glomus* which represents 12 species (Table 2). Some of these AM fungal spores are presented in Fig. 2. Most dominating spores of AM fungi in the sand samples from Panambur beach were: *Acaulospora rehmi* Sieverding and Toro, *Acaulospora scrobiculata* Trappe, *Acaulospora* spp, *Gigaspora* spp, *Glomus aggregatum* Schenk and Smith., *Glomus caledonium* (T.H. Nicolson & Gerd.) Trappe & Gerd, *Glomus constrictum* Trappe, *Glomus fasciculatum* (Thaxt.) Gerd. & Trappe, *Glomus flavisporum* Tul. & Tul., *Glomus macrocarpum* Tul. et Tul., *Glomus microcarpum* Tul. et Tul., and *Glomus* spp. Maximum spore count was reported in *I. pes-caprae*, whereas, remaining three plant species showed almost same spore count (Table 2). Our findings in present study, adds significant information to existing data and suggests that, Panmbur beach is nurturing some medicinal plants with good rate of mycorrhizal symbiosis.

TABLES

Table 1. Status of AM fungi in some medicinal plants from Panambur Beach

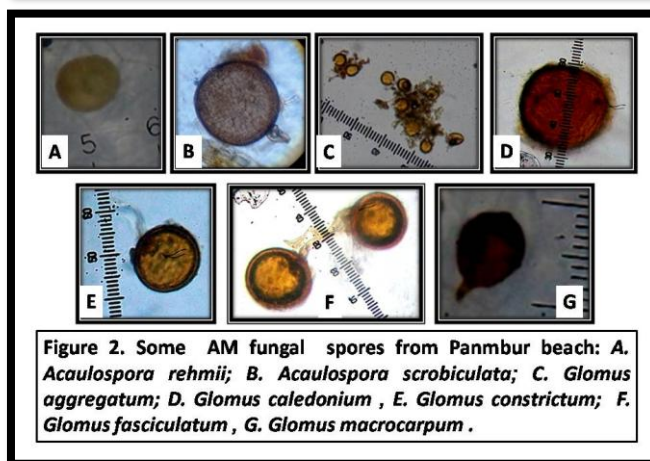
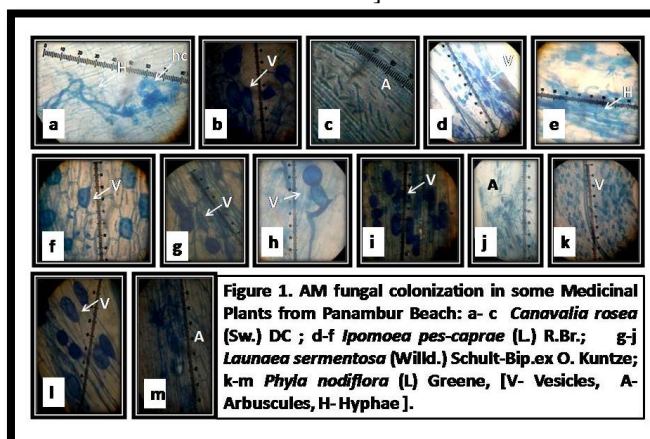
| Sr No. | Medicinal plant species | #Occurrence intensity of AM fungi | | | Percentage colonization |
|--------|---|-----------------------------------|-----|------|-------------------------|
| | | V | A | H | |
| 1. | <i>Canavalia rosea</i> (Sw.) DC (Fabaceae) | ++++ | +++ | ++++ | 98% |
| 2. | <i>Ipomoea pes-caprae</i> (L.) R. Br. (Convolvulaceae) | ++++ | + | ++++ | 100% |
| 3. | <i>Launaea sermentosa</i> (Willd.) Schult-Bip.ex O. Kuntze (Asteraceae) | ++++ | ++ | ++++ | 98% |
| 4. | <i>Phyla nodiflora</i> (L) Greene (Verbenaceae) | ++++ | + | ++++ | 100% |

[Vesicles (V); Arbuscules (A); Hyphae (H); (+)1-25%: Poor; (++)25-50%: Moderate; (+++)50-75%: good; (++++)more than 75%: Excellent.]

Table 2. AM fungal species in some medicinal plants from Panambur Beach

| Sr. No. | AM fungal species | * Occurrence of AM fungal spores in Medicinal plant species | | | |
|---|--|---|------------|-----------|-----------|
| | | Cr | Ip | Ls | Pn |
| 1. | <i>Acaulospora rehmsii</i> Sieverding and Toro | + | - | - | - |
| 2. | <i>Acaulospora scrobiculata</i> Trappe | - | - | - | + |
| 3. | <i>Acaulospora</i> spp | - | - | - | + |
| 4. | <i>Gigaspora</i> spp | - | - | + | - |
| 5. | <i>Glomus aggregatum</i> Schenk and Smith | - | - | + | - |
| 6. | <i>Glomus caledonium</i> (T.H. Nicolson & Gerd.) Trappe & Gerd | - | + | - | - |
| 7. | <i>Glomus constrictum</i> Trappe | - | + | - | - |
| 8. | <i>Glomus fasciculatum</i> (Thaxt.) Gerd. & Trappe | - | + | - | - |
| 9. | <i>Glomus flavisporum</i> Tul. & Tul. | + | - | - | - |
| 10. | <i>Glomus macrocarpum</i> Tul. et Tul. | + | - | - | - |
| 11. | <i>Glomus microcarpum</i> Tul. & Tul. | - | - | + | - |
| 12. | <i>Glomus</i> spp | + | - | - | - |
| Total No. of AM Spores 100 g⁻¹ rhizosphere sample | | 86 | 242 | 80 | 83 |

* [Cr: *Canavalia rosea*; Ip: *Ipomoea pes caprae*; Ls: *Launaea sermentosa*; Pn: *Phyla nodiflora*; (+) present; (-) absent]



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