

BUSHLAND REHABILITATION AND REVEGETATION GUIDELINES

CONSERVATION MANAGEMENT PLAN APPENDIX D

Jandakot Airport Holdings Pty Ltd 16 Eagle Drive Jandakot WA 6164

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1 Introduction

No areas within the Jandakot Airport Conservation Precincts are currently identified as requiring rehabilitation or revegetation. However, in the event that revegetation is required to be undertaken at some future point, these Rehabilitation and Revegetation Guidelines have been developed to assist in planning.

2 Triggers for Revegetation.

The need to undertake rehabilitation or revegetation within the Conservation Precincts of Jandakot Airport may be triggered by:

- Bushfires (where natural regeneration has not been successful).
- Impacts of weeds on vegetation condition are not successfully managed by weed control (i.e. areas defined as degraded in 2011 bushland condition survey show further decline in subsequent 5-yearly survey despite weed treatment).
- Impacts of dieback on vegetation condition are not successfully managed by phosphite and other dieback management measures (i.e. areas defined as degraded in 2011 survey show further decline in subsequent 5-yearly survey despite dieback treatment).
- The closure of surplus or non-essential firebreaks and access tracks.
- Verge impacts from the construction of new roads as detailed in Master Plan 2009.
- The creation of wildlife corridors.

3 Understanding the Area to be Revegetated

It is essential to know as much as possible about the area to be revegetated, including:

- Original vegetation type, including dominant species
- Dieback status
- Threats (e.g. feral and native grazing pressures, susceptibility to erosion, etc.).

4 Species for Use in Revegetation

Only species endemic to the local area that are consistent with the area's original vegetation type may be used for revegetation works. The only exception to this is for revegetation of heavily impacted dieback areas where the original dominant vegetation type is known to be dieback-susceptible, and using these species in revegetation is likely to have a low probability of success.

Plant species endemic to Jandakot Airport are listed in Attachment 1. Nearly half of the species naturally occurring on the site are susceptible to dieback, including many of the tree species.

4.1 Dieback Infested Areas

In dieback areas, only dieback resistant species (see Attachment 1) should be planted. In areas adjacent to dieback infestations or high risk areas for dieback spread (e.g. areas subject to significant earthmoving), primarily dieback resistant species should be planted.

Eucalyptus marginata (Jarrah) is particularly susceptible to dieback; however, some dieback resistant populations have been identified in South West WA. JAH will obtain seed produced by dieback resistant Jarrah trees for revegetation in dieback infested areas where there has been significant Jarrah overstorey decline.

In addition, *Eucalyptus todtiana* (Prickly bark) and *Nuytsia floribunda* (WA Christmas tree) will be considered for planting in infested areas, as these species have demonstrated high levels of natural resistance to dieback. Shearer and Hill (1989) observed that in *Banksia* woodlands of the Bassendean Dune system of the Swan Coastal Plain, most of the dominants and many understorey species are susceptible to *P. cinnamomi. Banksia attenuata*, *B. ilicifolia* and *B. menziesii* are commonly lost from communities, leaving scattered trees of *Eucalyptus todtiana* and *Nuytsia floribunda*, both of which are largely resistant to infection.

4.2 Local Provenance

In addition to using plants or seeds of the right species for revegetation, it is also important to use plants that have the right genes for the local area. It is JAH policy to only use or propagate seed of local provenance, as this maintains the genetic diversity and integrity of a given species. Plants of local provenance are better adapted to local conditions and prove to be more successful in revegetation works. The only exception to this is for revegetation of heavily impacted dieback areas where dieback-resistant strains sourced from other localities may be used.

The hierarchy of provenance seed is summarised below:

- 1st Preference seed collected from airport land
- 2nd Preference seed collected from the Swan Coastal Plain within 20 km of Jandakot Airport
- 3rd Preference seed collected from the Swan Coastal Plain.

5 Revegetation Techniques

Revegetation can be undertaken using three main techniques: regeneration, direct seeding and replanting, or a combination thereof.

The Jandakot Airport Banksia Woodland Revegetation Project, undertaken by the WA Department of Parks and Wildlife utilising offset funds from EPBC 2009/4796, is currently researching the effectiveness of different methods of banksia woodland restoration. The outcome of this research will provide further guidance for future revegetation works in the Conservation Precincts of Jandakot Airport.

5.1 Regeneration

Regeneration involves the natural regrowth of endemic vegetation, using existing seed bank in the soil or from nearby vegetation sources. It guarantees that vegetation is endemic and possesses the local genes to be successful. Where practical, natural regeneration of native species, in preference to replanting, is encouraged in the first instance as the revegetation method.

Regeneration, however, is most appropriate for areas that have only recently been cleared, where topsoil is intact and seed stock is available. Areas of long-term disturbance or landuse may not be appropriate. The transfer of topsoil from recently cleared areas of the same vegetation type may be beneficial in promoting regeneration. Topsoil may be collected from cleared areas for reuse in revegetation as long as materials from dieback infested areas are kept within the infestation boundaries.

However, this technique is unlikely to be successful for species that do not readily propagate via a soil-based seed bank and may need to be augmented by direct seeding or replanting.

5.2 Direct Seeding

Direct seeding involves the sowing of seeds, either by hand or machine, directly to a revegetation area. Direct seeding is considered more cost and labour efficient than planting,

excluding time required for seed collection, and allows for a higher plant density, which provides shelter to seedlings and reduces the potential for smothering by weeds. Direct seeding also results in a more natural mix of trees, shrubs and groundcovers than can be achieved through planting.

Seeds for use in direct seeding projects should be collected from Jandakot Airport bushland (preferably from areas scheduled to be cleared) well in advance of being required and stored in managed 'seed banks'. Prior to use, it is beneficial to undertake germination trials to determine the viability of the seed and thereby calculate the amount required to achieve the desired density of key species.

In dieback infested areas, it may be appropriate to revegetate using direct seeding instead of planting seedlings as seedlings are known to be particularly susceptible to death from dieback, even if they are not of a dieback susceptible species.

5.3 Replanting

Replanting involves the direct planting of endemic seedlings or tubestock. Plants are propagated in a Nursery Industry Accreditation Scheme Australia (NIASA)-accredited nursery using either provenance seed or cuttings.

The number of plants required is dependent factors such as the required vegetation type and survival rates.

Transplants, generally sourced from areas scheduled for clearing, can also be used, with the greatest success expected from seedlings and juvenile plants as opposed to mature individuals. Some species are also known to transplant more readily, with others not tolerating root disturbance. Transplants should not be collected from dieback infested areas. Seed can be collected from dieback infested areas as long as appropriate dieback clean-down procedures are implemented for all shoes, vehicles and tools.

6 Management Techniques to Ensure Success

Planning, site preparation and ongoing management techniques will increase the probability of successful revegetation. The type of techniques used will be dependent on the revegetation technique used.

6.1 Planning

The table below provides indicative timing for which key activities should be undertaken.

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Activity	Timing	
Begin propagating plants in nursery from seed or cuttings.	October – November	
Collect local seed	Seasonally dependent. Usually November – February.	
Site Preparation	March/April. Note areas with heavy weed infestations may require 12-18 months of weed control prior to revegetation occurring.	
Planting	May – July.	

6.2 Site preparation

Prior to revegetating an area, a range of site preparation works will be required. The following activities are recommended during site preparation.

Activity	Purpose	
Weed Control	To prevent weeds smothering revegetation.	
Fencing	To prevent damage by:	
Ripping	To encourage root penetration to aid seedling establishment. Care required in areas prone to erosion.	
Mounding	To aid seedling survival in damp areas prone to waterlogging.	
Mulch	Mulching after planting may be of benefit in areas used for direct planting. Mulching may help plants establish by protecting the roots and preventing excessive moisture loss from the surrounding soil, but it may also help to stabilise areas prone to erosion (e.g. bunds and slopes).	
	Mulching (with the exception of hydromulching) is not recommended in areas where direct seeding is undertaken or regeneration from transferred topsoil is being promoted.	
Mulch may be collected from cleared areas for reuse in revegetation a materials from dieback infested areas are kept within the infestation bound		

6.3 Ongoing Management

Once an area is regenerating or has been replanted, some ongoing monitoring and management will be required to ensure high survival rates. This may include watering seedlings during dry periods for the first year, ongoing weed control, rabbit control and infilling bare areas.

Attachment 1. Jandakot Airport Flora Species Dieback Susceptibility

S = Dieback Susceptible; R = Dieback Resistant Taken from information compiled by E. Groves, G. Hardy and J. McComb, Murdoch University. Species list reviewed by Mark Brundrett, 2011.

Acacia applanata Acacia huegelii R Acacia pulchella R Acacia saligna R Acacia stenoptera S Acacia willdenowiana Adenanthos cygnorum S Adenanthos obovatus S Allocasuarina fraseriana S Allocasuarina humilis S Amphipogon laguroides Amphipogon turbinates Anigozanthos humilis Anigozanthos manglesii R Aotus sp. procumbent Arnocrinum preissii R Astroloma xerophyllum S Austrodanthonia occidentalis Austrodanthonia pilosa Austrostipa compressa Baeckea camphorosmae R Banksia ilicifolia S Banksia littoralis S Banksia littoralis S Banksia nivea S Baufortia elegans Beaufortia squarrosa Boronia crenulata R Boronia ramosa Bossiaea eriocarpa S Brachyloma preissii R Caladenia flava		
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Caladenia huegelii	
Caladenia longicauda	
Calectasia narragara	
Calytrix angulata	
Calytrix flavescens	R
Calytrix fraseri	
Calytrix strigosa	
Cassytha flava	R
Cassytha glabella	R
Cassytha racemosa	
Centrolepis aristata	
Centrolepis drummondiana	
Centrolepis humillima	
Chamaescilla corymbosa var. corymbosa	
Chordifex microcodon	
Comesperma calymega	R
Conospermum triplinervium	S
Conostephium minus	
Conostephium pendulum	S
Conostephium preisii	
Conostylis aculeata	R
Conostylis aurea	
Conostylis caricina ssp. Caricina	
Conostylis juncea	
Conostylis setigera ssp. Setigera	R
Crassula colorata	
Croninia kingiana	
Cryptostylis ovata	R
Cyanicula gemmata	
Cyathochaeta avenacea	R
Dampiera linearis	R
Danthonia pilosa	
Dasypogon bromeliifolius	S
Daviesia gracilis	
Daviesia incrassata	S
Daviesia juncea	
Daviesia nudiflora	
Daviesia physodes	S
Daviesia triflora	

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Desmocladus fasciculatus	R
Desmocladus flexuosus	R
Dianella revoluta	S
Dielsia stenostachya	
Diuris corymbosa	
Diuris emarginata	
Diuris laxiflora	
Drosera erythrorhiza	R
Drosera macrantha	
Drosera menziesii ssp. Penicillaris	
Drosera paleacea ssp. Paleacea	
Drosera pulchella	
Eremaea asterocarpa	
Eremaea pauciflora	
Eriostemon spicatus	R
Eucalyptus gomphocephala	R
Eucalyptus marginata	S
Eucalyptus rudis	R
Eucalyptus todtiana	S
Euchilopsis linearis	
Euchiton sphaericus	
Eutaxia virgata	
Gastrolobium capitatum	
Gnaphaluim sphaericum	
Gompholobium confertum	
Gompholobium scabrum	
Gompholobium tomentosum	R
Gonocarpus pithyoides	
Goodenia pulchella	
Haemodorum paniculatum	R
Haemodorum spicatum	
Hardenbergia comptoniana	R
Hemiandra pungens	R
Hensmania turbinata	
Hibbertia aurea	
Hibbertia huegelii	S
Hibbertia hypericoides	S
Hibbertia pachyrrhiza	
Hibbertia racemosa	R
Hibbertia subvaginata	
Homalosciadium homalocarpum	
Hovea trisperma	R
Hyalosperma cotula	
Hypocalymma angustifolium	R
Hypocalymma robustum	S
τηροσαιγιπιτα τουαδιαπί	J

Hypolaena exsulca Hypolaena pubescens	
Hypolaena pubescens	
. If postaria postario	
Jacksonia furcellata	S
Jacksonia sternbergiana	S
Juncus kraussii	
Kennedia prostrata	R
Kunzea ericifolia	S
Laxmannia ramosa	
Laxmannia squarrosa	
Lechenaultia biloba	R
Lechenaultia expansa	
Lechenaultia floribunda	
Lepidosperma angustatum	
Lepidosperma effusum	
Lepidosperma longitudinale	
Lepidosperma scabrum	R
Lepidosperma squamatum	R
Lepidosperma tenue	R
Leporella fimbriata	R
Leptocarpus canus	
Leptocarpus tenax	
Leptomeria empetriformis	
Leptospermum erubescens	R
Lepyrodia muirii	
Leucopogon australis	S
Leucopogon constephioides	S
Leucopogon insularis	
Leucopogon kingianus	
Leucopogon nutans	S
Leucopogon oxycedrus	S
Leucopogon pendulus	R
Leucopogon polymorphus	S
Leucopogon propinquus	S
Leucopogon pulchellus	S
Leucopogon racemulosus	
Leucopogon strictus	
Levenhookia stipitata	
Lobelia tenuior	
Lomandra caespitosa	
Lomandra caespitosa Lomandra endlicheri	
Lomandra endlicheri	
Lomandra endlicheri Lomandra hermaphrodita	R
Lomandra endlicheri Lomandra hermaphrodita Lomandra micrantha	R S

Lomandra purpurea	
Lomandra suaveolans	
Loxocarya cinerea	S
Loxocarya pubescens	
Loxycarya fasciculata	
Loxycarya flexuosa	
Lyginia barbata	
Lyperanthus nigricans	
Lysinema ciliatum	S
Lysinema elegans	
Macrozamia riedlei	S
Melaleuca incana	
Melaleuca preissiana	R
Melaleuca scabra	S
Melaleuca seriata	R
Melaleuca systena	
Melaleuca thymoides	S
Mesomelaena pseudostygia	
Mesomelaena stygia	R
Mesomelaena tetragona	R
Microtis media	
Millotia tenuifolia	R
Mitrasacme paradoxa	
Monotaxis grandiflora	
Neurachne ps.	
Nuytsia floribunda	R
Patersonia occidentalis	S
Perricalymma ellipticum	S
Persoonia saccata	R
Petrophile linearis	S
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Philotheca spicata	R
Phlebocarya ciliata	Λ.
Philippointe no redouum	
Phyllangium paradoxum	
Pimelea angustifolia	
Pimelea imbricata	
Pimelea rosea	
Pimelea sulphurea	
Pithocarpa pulchella	+-
Platysace compressa	S
Platytheca galioides	
Podotheca angustifolia	
Podotheca chrysantha	
Poranthera microphylla	
Prasophyllum parvifolium	

Pterostylis pyramidalis	
Pterostylis recurva	
Pterostylis vittata/sanguinea?	
Pultenaea reticulata	
Pyrorchis nigricans	
Quinetia urvillei	
Regelia ciliata	
Regelia inops	
Restio microcodon	
Restio stenostachyus	
Ricinocarpos glaucus	
Scaevola paludosa	
Scaevola repens	
Schoenus brevisetis	
Schoenus curvifolius	R
Schoenus efoliatus	
Schoenus globifes	
Schoenus rodwayanus	
Scholtzia involucrata	S
Senecio lautis ssp. Maritimus	
Siloxerus humifusus	
Sowerbaea laxiflora	
Stackhousia monogyna	
Stirlingia latifolia	S
Stylidium brunonianum	R
Stylidium carnosum	
Stylidium guttatum	
Stylidium junceum	S
Stylidium piliferum	R
Stylidium repens	
Stylidium schoenoides	S
Synaphea spinulosa	
Synaphea sp.	
Tetratheca setigera	S
Thelymitra campanulata	
Thelymitra crinita	
Thelymitra fuscolutea	
Thysanotus arbuscula	
Thysanotus manglesianus	
Thysanotus multiflorus	
Thysanotus patersonii	
Thysanotus sparteus	
Thysanotus thyrsoideus	S
Thysanotus triandrus	
Trachymene pilosa	

Tricoryne elatior	R
Tricoryne tenella	
Tripterococcus brunonis	
Verticordia drummondii	
Wahlenbergia preissii	
Waitzia suaveolens	
Xanthorrhoea preissii	S
Xanthosia huegelii	R

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