

## **Supplementary Figures and Tables**

### **The Influence of Host Plant Volatiles on the Attraction of Longhorn Beetles to Pheromones**

**R. Maxwell Collignon<sup>1\*</sup>, Ian P. Swift<sup>2</sup>, Yunfan Zou<sup>1</sup>, J. Steven McElfresh<sup>1</sup>, Lawrence M. Hanks<sup>3</sup>, and Jocelyn G. Millar<sup>1</sup>**

*<sup>1</sup> Department of Entomology, University of California, Riverside CA, USA*

*<sup>2</sup> California State Collection of Arthropods, Sacramento, CA, USA*

*<sup>3</sup> University of Illinois at Urbana-Champaign, IL, USA*

## Supplementary Tables

**Table S1.** Total trap catches for the first experiment, testing pheromones with crude host material. Within a row, means with the same letter or no letter are not significantly different (Dunn's Test).

Species	Subfamily	Pher.	Pher. + conifer	Pher. + oak	Conifer material	Oak material	Blank control	Total inds. trapped	Treatment replicates	Kruskal- Wallis
<i>Phymatodes obliquus</i>	Cerambycinae	403a	162b	334ab	2c	3c	1c	905	78	$P < 0.001$
<i>Phymatodes grandis</i>	Cerambycinae	44a	13b	40a	0b	1b	0b	98	45	$P < 0.001$
<i>Brothylus conspersus</i>	Cerambycinae	7ab	4ab	9a	0b	3ab	0b	23	16	$P = 0.003$
<i>Brothylus gemmulatus</i>	Cerambycinae	9a	3ab	8a	0b	0b	0b	20	12	$P < 0.001$
<i>Tetropium abietis</i>	Spondylidinae	6ab	11a	3ab	0b	0b	0b	20	16	$P < 0.001$
<i>Monochamus clamator</i>	Lamiinae	2b	10a	13a	0b	2b	0b	27	21	$P < 0.001$
<i>Tragosoma deparium</i>	Prioninae	8ab	18a	11a	1b	0b	0b	38	26	$P < 0.001$

**Table S2.** Volatiles identified from oak and conifer species that elicited responses from antennae of oak-infesting Cerambycinae species, in coupled gas chromatography-electroantennogram detection (GC-EAD) analyses.

	hexanal	hexanol	(Z)-3-hexenol	(E)-2-hexenol	$\alpha$ -pinene	(E)-2-hexenal	camphene	$\beta$ -pinene	$\beta$ -myrcene	3-carene	limonene	$\beta$ -phellandrene	1,8-cineole	benzaldehyde	borneol	camphor	4-allylanisole	methyl salicylate	(E)- $\beta$ -caryophyllene	unidentified
<i>Brothylus conspersus</i> ♀	x	x	x																	1
<i>Brothylus conspersus</i> ♂	x	x	x	x				x	x				x					x		3
<i>Haplidus testaceus</i> ♀			x		x			x	x	x	x	x					x			16
<i>Haplidus testaceus</i> ♂																				
<i>Molorchus eberneus</i> ♂		x	x	x				x	x	x	x		x		x	x	x	x		1
<i>Neoclytus modestus</i> ♀	x	x	x	x	x	x	x	x	x	x	x	x	x				x	x		8
<i>Neoclytus modestus</i> ♂																				
<i>Phymatodes grandis</i> ♀																				
<i>Phymatodes grandis</i> ♂		x	x					x	x			x		x			x			1
<i>Phymatodes infuscatus</i> ♀		x	x	x	x								x	x			x	x		5
<i>Phymatodes infuscatus</i> ♂																				
<i>Phymatodes obliquus</i> ♀		x	x	x	x	x		x	x	x	x		x		x	x	x	x	x	1
<i>Phymatodes obliquus</i> ♂		x	x	x	x	x							x				x	x		

**Table S3.** Volatiles identified from oak and conifer species that elicited responses from antennae of conifer-infesting cerambycid species, in coupled gas chromatography-electroantennogram detection analyses.

		hexanal	hexanol	(Z)-3-hexenol	(E)-2-hexenol	$\alpha$ -pinene	(E)-2-hexenal	camphene	$\beta$ -pinene	$\beta$ -myrcene	$\beta$ -carene	limonene	$\beta$ -phellandrene	1,8-cineole	benzaldehyde	borneol	camphor	4-allylanisole	methyl salicylate	(E)- $\beta$ -caryophyllene	unidentified active cmpds
<i>Asemum caseyi</i>	Spondylidinae	x	x	x	x	x			x	x	x	x			x		x	x	x	x	1
<i>Asemum nitidum</i>	Spondylidinae	x	x	x			x											x	x		
<i>Asemum striatum</i>	Spondylidinae										x	x					x	x	x		1
<i>Callidium sp.</i> ♀	Cerambycinae	x	x	x	x		x	x	x	x			x	x			x	x	x		1
<i>Callidium sp.</i> ♂	Cerambycinae	x	x	x	x		x	x	x	x			x		x		x		x		2
<i>Monochamus clamator</i> ♀	Lamiinae				x	x			x	x	x		x	x					x		1
<i>Monochamus clamator</i> ♂	Lamiinae																				
<i>Ortholeptura valida</i>	Lepturinae							x				x		x			x	x			1
<i>Poliaenus oregonus</i> ♀	Lamiinae																				
<i>Poliaenus oregonus</i> ♂	Lamiinae		x	x	x	x					x									x	1
<i>Pseudostylopsis sp.</i>	Lamiinae			x	x	x		x	x	x	x	x			x	x	x			x	17

**Table S4.** Total trap catches for the second experiment, testing pheromones with reconstructed host plant volatiles at three release rates (LR= low release rate, MR=medium release rate, HR=high release rate. Within a row, means with the same letter or no letter are not significantly different (Dunn's Test).

Experiment 2	Subfamily	Pher. + EtOH	Pher. + EtOH + LR conifer	Pher. + EtOH + MR conifer	Pher. + EtOH + HR conifer	Pher. + EtOH + LR oak	Pher. + EtOH + MR oak	Pher. + EtOH + HR oak	EtOH + MR conifer	EtOH + MR oak	EtOH	Total inds. trapped	Trt. rep s	Kruskal- Wallis
<i>Phymatodes obliquus</i>	Cerambycinae	293a	217ab	99bc	27cd	202ab	37cd	8d	0d	6d	7d	896	67	$P < 0.001$
<i>Phymatodes grandis</i>	Cerambycinae	68a	47ab	32ab	17bc	35ab	22bc	18bc	1c	2c	1c	243	45	$P < 0.001$
<i>Brothylus gemmulatus</i>	Cerambycinae	90abc	78ab	42a	22abc	43abc	32abc	10abc	0c	4abc	1bc	322	19	$P < 0.001$
<i>Neocyclus modestus</i>	Cerambycinae	10a	0b	2b	0b	2b	0b	0b	0b	0b	6ab	20	17	$P < 0.001$
<i>Neospondylis upiformis</i>	Spondylidinae	0b	3b	7ab	35a	0b	5b	0b	1b	0b	0b	51	13	$P < 0.001$
<i>Asemum nitidum</i>	Spondylidinae	1	0	2	4	0	2	3	6	0	0	18	13	$P = 0.015$
<i>Asemum striatum</i>	Spondylidinae	2	0	3	3	4	5	2	1	0	0	20	13	$P = 0.059$
<i>Asemum caseyi</i>	Spondylidinae	0	0	0	3	0	1	4	2	1	0	11	5	$P = 0.043$
<i>Xylotrechus albonotatus</i>	Cerambycinae	1	1	1	5	0	0	0	0	0	2	10	7	$P = 0.024$
<i>Monochamus clamator</i>	Lamiinae	1b	1b	5b	10a	0b	0b	1b	0b	0b	0b	18	15	$P < 0.001$

**Table S5.** Total trap catches for the third experiment, testing pheromones with subsets of host plant volatiles. Italicized numbers were not included in the statistical analyses based on experimental objectives. Means with the same letter or no letter are not significantly different (Dunn's Test).

Experiment 3	Subfamily	Pher.	Pher. + EtOH	Pher. + HR $\alpha$ - pinene	Pher. + EtOH + HR $\alpha$ - pinene	Pher. + HR conifer	Pher. + EtOH + HR conifer	Total inds. trapped	Treatment replicates	Kruskal- Wallis
<i>Phymatodes obliquus</i>	Cerambycinae	147	143	<i>61</i>	<i>66</i>	<i>14</i>	<i>27</i>	458	45	$P = 0.39$
<i>Phymatodes grandis</i>	Cerambycinae	28b	67a	<i>19</i>	<i>55</i>	<i>10</i>	<i>29</i>	208	45	$P = 0.004$
<i>Brothylus conspersus</i>	Cerambycinae	1	2	<i>1</i>	<i>3</i>	<i>2</i>	<i>1</i>	10	8	$P = 0.54$
<i>Brothylus gemmulatus</i>	Cerambycinae	41	40	<i>14</i>	<i>66</i>	<i>6</i>	<i>17</i>	184	17	$P = 0.97$
<i>Neospondylis upiformis</i>	Spondylidinae	<i>0</i>	<i>0</i>	7b	4b	26ab	59a	96	11	$P = 0.001$
<i>Asemum nitidum</i>	Spondylidinae	3	<i>0</i>	75a	37ab	18b	9b	142	29	$P < 0.001$
<i>Asemum striatum</i>	Spondylidinae	<i>0</i>	<i>0</i>	14a	9ab	1b	1b	25	11	$P = 0.002$
<i>Tetropium abietis</i>	Spondylidinae	6	<i>0</i>	3	1	0	3	13	9	$P = 0.28$
<i>Xylotrechus albonotatus</i>	Cerambycinae	<i>0</i>	3	1	1	0	3	8	6	$P = 0.20$
<i>Monochamus clamator</i>	Lamiinae	2	3	3ab	2b	13a	11ab	34	21	$P = 0.010$

## List of Supplementary Figures

**Fig. S1.** Mean ( $\pm 1$  SE) numbers of beetles caught in traps for three conifer-infesting spondylidine species caught in traps baited with the pheromone blend, low (LR), medium (MR), and high (HR) release rates of reconstructed blends of host plant compounds, and ethanol. There were no significant means separations among the treatments (Dunn's Test).

**Fig. S2.** Mean ( $\pm 1$  SE) numbers of beetles caught in traps for two cerambycine oak-infesting species. There were no significant means separations via Dunn's Test, but both species showed a similar increased response to treatments with ethanol versus the same treatment without (i.e.,  $\alpha$ -pinene with ethanol vs.  $\alpha$ -pinene without ethanol, and conifer blend with ethanol vs. conifer blend without ethanol).

Fig. S1

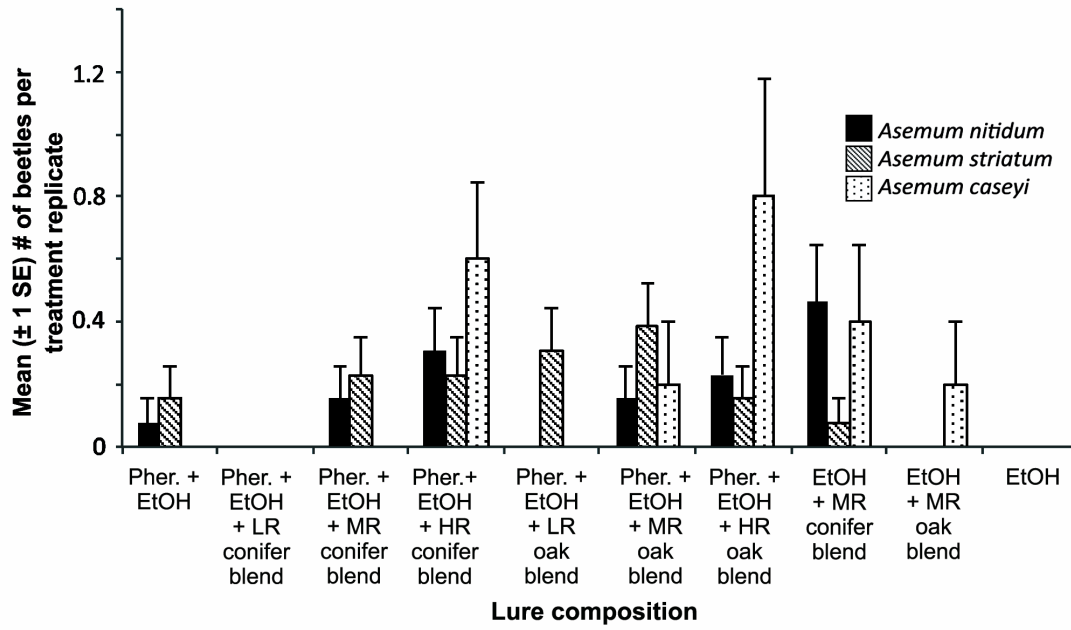




Fig. S2

