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Water lily research: Past, present, and future

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Abstract

The water lily order Nymphaeales includes ~100 species and all grow into aquatic herbs. Most of them are not only economic crops but have, for thousands of years, been regarded as cultural symbols, especially in Buddhism and Hinduism. Water lily order forms an early angiosperm branch, occupying vital roles in evolutionary biology. With the advent and rapid development of omic techniques and bioinformatic software, the research on water lily biology has achieved unprecedented success. In this review, we summarized a series of recent and important findings in water lily research. Genomic analyses of water lilies provide insights into details of their evolutionary history and ecological adaptation. The genomes also provide rich resources for genetic studies and molecular breeding. The opening and closing rhythm of flowering is controlled by auxin and the candidate genes are proposed. Part of the genes responsible for floral scents and floral colors in *Nymphaea colorata* have been studied in-depth for their functions. Metabolomic profiling reveals the anthocyanins responsible for the floral color formation and the volatile organic compounds as floral scent molecules. Phenotypic studies surveyed the diverse traits from flowers to leaves and bulb roots, and some of these studies reveal the link between genes and phenotypes. However, genes responsible for some critical traits such as the growth of water lily plants, such as the stem, bulb formation, and the initiation of vivipary are still unknown. Finally, we propose potential future research of water lilies, including genetics, breeding, and industrialization.

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Introduction

The Nymphaeales order, including three families (Hydatellaceae, Cabombaceae, Nymphaeaceae) and 85 species (Fig. 1), are called the water lily order^[1,2], partly due to the dominant character of all these species are aquatic herbs and nearly all of the species have very beautiful flowers and leaves. Hence the water lily is recognized as the common name for all species categorized in the Nymphaeales order^[3–8]. Water lilies are globally distributed, spanning from tropical to cold regions^[9]. They have played vital roles in the economy, ecology, and evolutionary biology.

Economic, cultural, ecological, and evolutionary values of water lilies

Water lilies contain various economic values. The *Nymphaea* spp. flowers are not only good cut flowers, but can be in food and beverages^[11]. For example, some *Nymphaea* water lilies have starchy and edible roots^[12]. The seeds of *Victoria* and *Euryale ferox* are edible^[13]. *Nymphaea* species contain various medicinal compounds, and they have been found to function in anti-bacteria^[14], anti-inflammatory^[15], and anti-depressant^[16] capacaties. According to data from the China Flower Association, at the end of 2019, the annual output of water lilies

in China was 152 million heads, and the combined market size of the lotus and water lily industries reached 10 billion CNY (www.cqla.cn/chinese/news/news_view.asp?id=58006). In Lichuan county from Hubei province, China, the 'hometown of E. ferox in China', its planting area is about 2,000 hectares, and the annual output value exceeds 200 million CNY (http://news.yuanlin.com/detail/2021128/379540.htm). Water lilies have high cultural value and a long consumption history. In ancient Greece and Rome, together with the lotus, some Nymphaea species were regarded as the embodiment of holiness and beauty and were often used as offerings to the gods. In modern times, one of the top impressionist painters Claude Monet has painted hundreds of water lilies^[17]. Many countries have issued stamps or currency with water lily flowers as the theme. Water lilies have a certain ecological value, especially in water purification and heavy metal absorption^[18]. However, some Nymphaea species grow very quickly and are regarded as invasive species. For example, N. odorata and N. capensis encroach heavily on the living space of other species in the same ecosystem and destroy the ecological balance (www.invasive.org/species/aquatic.cfm and www.issg.org/ database/species/ecology.asp?si=985&lang=SC.)

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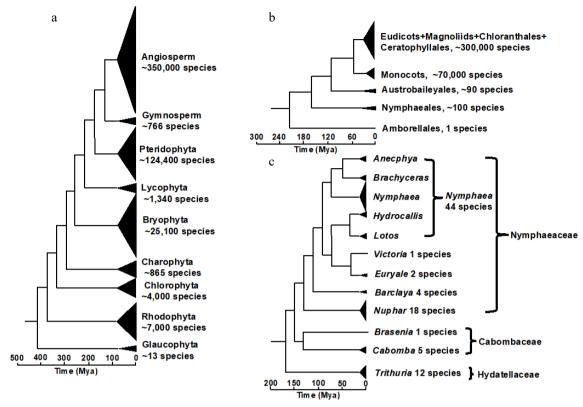


Fig. 1 The evolutionary position of water lilies. (a) Tree of green plant lineage. (b) Tree of angiosperms (flowering plants). (c) Tree of water lily order Nymphaeales. Phylogenetic relationships and timing of the nodes are based on previous phylogenomic research^[9] and phylotranscriptomic research^[10].

The water lily order occupies a critical evolutionary position. Together with Amborellales and Austrobaileyales, these three orders collectively form the ANA (Amborellales-Nymphaeales-Austrobaileyales) grade, which are the first three flowering plant branches^[19]. The early diverging water lily order provides a unique window to study the origin and early evolutionary features of flowering plants. Besides, the various kinds of floral scents and colors are wonderful traits for ornamental studies. Their genes and related pathways are the focus of the molecular breeding of water lilies and even other ornamental plants.

Growing research on water lilies

Multi-omic tools, including genomics, epigenomics, transcriptomics, proteomics, and metabolomics, are high-throughput and effective tools for plant studies. With the development of high-throughput technologies and bioinformatics tools, various omic research continues to deepen our understanding of plant traits^[20]. Through high-throughput measurement of each omics and data integration, we can not only comprehensively and systematically understand gene functions and promote molecular breeding, but also provide important data and technical approaches for the research of emerging cross disciplines such as quantitative and systems biology.

To date, there are roughly 868 publications indexed in the PubMed database (Fig. 2). The annual number grows rapidly, from eight publications in 2000 to 71 in 2021. The rapid increase reveals the boosted interest in various kinds of water lilies, from Nymphaea to others.

Genome sequences of three water lilies

The genome sequence of Nymphaea colorata

Chen et al. selected the N. colorata, a diploid tropical species that originated from east Africa for genome sequencing due to its relatively small genome size and small plant size^[21]. Relying on the PacBio long-read sequencing technology, a 409 Mb genome is assembled, falling into 14 chromosomes^[9]. The genome assembly has relatively high quality with contig N50 up to 2.1 Mb. Besides, some telomeres and centromeres are very well assembled and characterized. At the same time, the genomes of its mitochondria and chloroplast are successfully assembled using the same PacBio reads^[22]. Based on its genome information, we are able to precisely reveal its evolutionary place (Fig. 3) and identify the genes responsible for the floral scent, floral color^[23], as well as floral morphogenesis^[24]. The expansion of biotic and abiotic stressrelated genes such as WRKY genes and R genes is related to the wide distribution of the Nymphaea water lilies^[25]. Some of the key genes responsible for the stomata development are lost in the N. colorata genome and may explain its unique aquatic lifestyle adaptation^[26]. This is the first report on the water lily genome sequence, which is a milestone showing that water lily research is entering the genomic era.

The genome sequence of *N*. thermarum

Povilus and colleagues from Harvard University assembled a draft genome of a dwarf tropical water lily *N. thermarum*^[27]. This species originates from Rwanda (east Africa). Relying on Illumina short reading sequencing technology, they obtained a

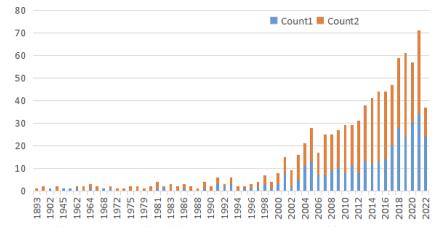


Fig. 2 The growing research publications on water lilies. The searches are based on PubMed from NCBI (https://pubmed.ncbi.nlm.nih.gov/) on May 24, 2022. Count 1 shows the keywords '(nymphaea) OR (waterlily) OR (water lily) OR (euryale) OR (barclaya) OR (nuphar) OR (trithuria) OR (cabomba) OR (brasenia)', while count 2 shows the keywords 'nymphaea'.

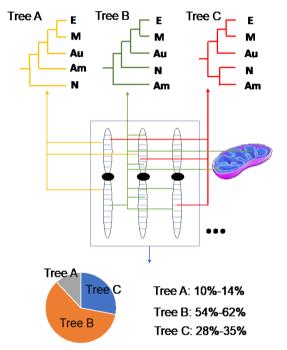


Fig. 3 Schematic diagram showing that the phylogenomic analyses reveal accurate phylogeny of angiosperms. Tree A represents that Nymphaeales, but not Amborellales forms the earliest extant angiosperm branch. Tree B represents Amborellales as the earliest extant angiosperm branch. Tree C represents Amborellales and Nymphaeales are sister lineage as the co-first earliest extant angiosperm branch. The proportion data in the piechart are from our previously published article^[9]. E: Eudicots, M: Monocots, Au: Austrobaileyales, Am: Amborellales, N: Nymphaeales.

368 Mb genome of *N. thermarum*. Analyses of this genome and phenotypes reveal the loss of genes enrolled in the vascular cambium, which might be a key innovation of its adaptation to the aquatic environment. This is the second report on the water lily genome.

The genome sequence of *Euryale ferox*

The *Euryale* genus only contains one species (*E. ferox*) which is mainly distributed in southern and eastern Asia. Their leaves and flowers are covered in sharp prickles and float on the water's surface. Its edible seeds are usually called fox nuts and are used as food or diet therapy supplies in Asia. Relying on the long-read sequencing platform Oxford Nanopore Technology, Liu and colleagues assembled the genome sequence of *E. ferox*^[28]. The genome assembly has a size of 725.2 Mb and 29 chromosomes. Phylogenomic studies also support that *E. ferox* is an early angiosperm branch, following the *Amborella*, and the incomplete lineage sorting may largely account for phylogenetic inconsistencies of different major lineages in angiosperms. This is the third report on the water lily genome, which provides important resources to reveal the specific evolution of seeds and leaves.

Whole-genome methylation researches

There has been little focus on the DNA methylation in water lily research. Bisulfite sequencing (BS-seq) on *Nuphar advena* did not find CHH type methylation in its genome. Comparative analyses of *N. advena* and other land plants' relative levels of CG methylation are a consistent property across genes, and genic CHG methylation correlates with genome size^[29]. We believe that in the future, more results will emerge as more genome sequences of water lilies become available.

Floral development

Flowering time

Most *Nymphaea* species bloom in the morning and close their flowers in the afternoon. Only a few species bloom at night and close their flowers before the morning. This unique circadian flowering clock is very interesting and attractive. Chen et al. collected flower samples at different time points and sequenced the transcripts^[30]. They uncovered eight expressional gene clusters related to floral circadia. They showed that auxin-related signaling pathway genes are linked to synchronized expression patterns with opening and closure processes.

The bloom of *Nymphaea* flowers usually last three to four days, which largely shortens the shelf life of cut flowers. Li et al. investigated when the senescence process is initiated and how it terminates the movement rhythm^[31]. They identified multiple signaling pathways that were activated at the last stage, the floral closure, on the third day after flowering. Genes related to hydrolase are upregulated on the same day, indicating that petals enter the senescence stage at that time.

Floral fragrance

Unlike species from the Hydatellaceae family and Cabombaceae family, most Nymphaea flowers emit fragrant molecules, volatile organic compounds, to attract various kinds of pollinators. A wide variety of small molecules or metabolites are responsible for the diverse floral colors and scents. Identifying small molecules using gas chromatography-mass spectrometry (GC-MS) is becoming more common. For example, GC-MS-based screening on the flower petals of N. nouchali identified 16 biologically active phytochemicals^[32]. GC-MS on the *N. pubescens*'s crude pedicel and flower also identified 45 bioactive compounds^[33]. However, another study using the same method on the flower of N. pubescens identified 71 bioactive compounds^[34]. A total of 22 VOCs were found in the seven species of Nymphaea subgenus Hydrocallis^[35]. Gasliquid chromatography (GLC) based extraction of N. alba leaf identified 53 bioactive compounds^[36]. The ultra-performance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS) screen on the flower of N. 'Blue Bird' identified 455 metabolites^[37]. A large-scale screen on 56 water lily cultivars identified 117 volatile organic compounds (VOCs), in which alkenes and alkanes were the most abundant, followed by ketones and aldehydes^[38]. The N. colorata flower releases 11 different volatile molecules, mainly terpenoids, fatty acids, and benzenoids as fragrant molecules, among which methyl decanoate is the major fragrance molecule^[9] (Fig. 4).

The Victoria genus only includes species V. amazonica, V. cruziana, and the recently identified species V. boliviana^[39]. Exploration of floral scent composition using GC-MS could provide a large array of floral VOCs in a series of water lilies. In Victoria cruziana, four VOCs were identified from its flower, in which the VOC composition and emission time are also identified^[40].

At the genetic level, transcriptome analyses identified a SABATH gene family member, NC11G0120830, and a terpene synthase gene family member, NC11G0123420, as highly and uniquely expressed in petals and involved in the metabolism of fragrant molecules^[9].

Floral colors

Nymphaea species display a variety of floral petal colors, including white, red, purple, yellow, blue, etc. A few Nymphaea water lilies, such as N. colorata, N. caerulea, N. 'King of Siam', etc., exhibit beautiful blue petals. Flavonoids are the key elements of floral color. Nymphaea species contain a wide variety of flavonoids. For example, a broad sampling of 35 tropical Nymphaea cultivars identified 34 flavonoids^[41]. Transcriptome sequencing was conducted on the colorless stage and the fully colored stage of the blue flower cultivar N. 'King of Siam'. Although no reference genome was available for this species, Wu et al.^[42] still identified 26,206 unigenes, among which 1,581 genes have differential expression values between the two stages. An in-depth survey revealed that 33 genes with significant expressional levels are responsible for color formation. Finally, the authors validated the expressional changes of seven ANTHOCYANIDIN 3-O GLUCOSYLTRANSFERASE (UA3GT) genes using the qRT-PCR method^[42]. N. colorata has very attractive al.^[9] blue petals. Zhang et have identified the Dp3'galloylacetylGal as the main anthocyanin for blue coloration in the petals (Fig. 4).

The *Victoria* species display large flowers, showing white petals on the first day, turning pink or red on the second day.

Relying on the combination method of high-performance liquid chromatography with photodiode array detection (HPLC-DAD) and ultra-performance liquid chromatography coupled with tandem mass spectrometry (UPLC-MS/MS), Wu et al. identified 14 flavonoids, including 10 flavonols and four anthocyanins, in the petals of two *Victoria* species^[43].

Seed biology

The large and starchy seeds of *E. ferox* are both a nutritious food and a medicine in East Asia. By transcriptomic analyses of different seed stages, Liu et al. found increased expression of *P450* and *PAL* genes in phenylpropanoid metabolism, which might be involved in the maturation of its seeds in aspects of seed size, color, harness, and accumulation of secondary metabolites^[13]. Specifically, the authors found the changed expressional pattern of flavonoids during the seed development, involving ABA and SA phytohormones. Candidate genes in regulating the two phytohormones in regulating the flavonoids are identified^[44].

Leaf vivipary: rapid vegetative propagation strategy by tropical water lilies

Some *Nymphaea* species are leaf vivipary by producing seedlings on the leaves. As an interesting horticultural trait, the leaf vivipary is not only a supplement to breeding programs, but also an opportunity to study stem cell initiation in mature organs. By comparing the transcriptomes of four stages of leaf development between the non-vivipary *N. colorata* and the vivipary *N. micrantha*, Su et al. found four potential regulators, including the transcription factors ERF1B, ERF105, RAP2-3, and WRKY22^[45]. In the future, through functional studies of genes, such as CRISPR-Cas9 mediated gene knockout and transgenic technologies, it is possible to gain more knowledge of the origin and evolution of vivipary in plants.

Floral vivipary: branching flowers

Nymphaea prolifera is the water lily currently known to produce floral vivipary, an amazing trait called branching flowers. Instead of fertile flowers, *N. prolifera* usually produces tuberiferous and sterile flowers that act as vegetative organs. *N. prolifera* can produce two levels and sometimes even three levels of branching flowers, giving rise to dozens and often more than 100 vegetative propagules^[46]. This unique trait is, however, still under researched. In the future, its potential in general biology will perhaps show its significance, at least in breeding or maintaining important mutations.

The water lily rhizome and tuber bulb

A large number of angiosperm crops, including the *Allium*, *Iris*, *Tulipa*, and *Begonia*, have bulbs. All Nymphaeaceae species contain starchy tuber bulbs, although ranging dramatically in size from one centimeter to a foot. It seems that the bulbs have evolved independently many times during evolution. *Nymphaea* water lilies develop four major types of bulbs, including Marliac-type rhizome, pineapple-type rhizome, ordorata-type rhizome, and tuberosa-type rhizome. These bulbs are rich in starch and nutrients, making them a good alternative food in some tropical regions. At the genetic level,

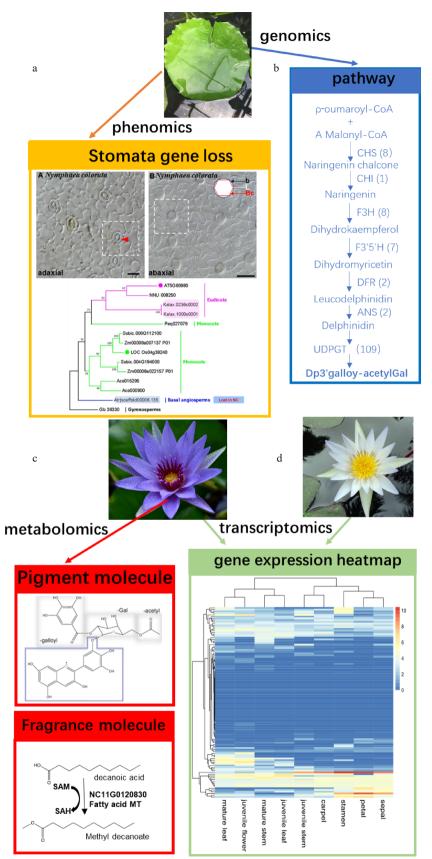


Fig. 4 (a) Phenomics, (b) genomics, (c) metabolomics, and (d) transcriptomics, guided gene screening related to blue anthocyanin in the petal of *N. colorata* and the stomata development. The genomic and phenomic studies identified loss of genes associated with stomatal development in the lower abaxial side leaf of *N. colorata*. Genomic, transcriptomic, and metabolomic studies identified floral fragrance-related genes and pathways associated with blue anthocyanin synthesis.

the *FLOWERING LOCUS T* (*FT*) genes are found to be involved in bulb formation in onions^[47]. Zhang et al. found a total of five *FT* genes in the genome of *Nymphaea colorata*^[9]. The five *FT* genes are clustered into two groups, which are produced by *Pi*-whole genome duplication^[48]. This suggests that the two groups may have different functions, with one group controlling the flowering time and the other controlling the bulb formation. However, due to the very limited research on water lilies, we still do not know which *FT* gene is responsible for its bulb initiation and development.

The runner stem in water lilies

A few water lily species produce stems that grow flat along the ground called runners. It will root at the node and form a new plant, thereby spread rapidly. The *Nymphaea mexicana* and the *Brasenia schrebri* produce runners and become an invasive species in many aquatic environments. The runner is also common in monocot plants such as wild rice and *Iris* spp., and eudicots such as strawberry and fish mint. Runners could be utilized for plant propagation. Although we now know that the phytohormone GA and the *DELLA* gene are involved in runner formation^[49], the genetics of the runner is completely unknown in water lilies.

Germplasm studies

Hybrid breeding within and across various *Nymphaea* species is relatively easy and popular. To date, there are hundreds of *Nymphaea* cultivars available. Different *Nymphaea* cultivars display various floral colors, floral scents, flowering times, leaf shapes, and bulb sizes. Various traits of water lily populations have been collected for a long time and many of them have been well described and statistically analyzed. Dąbrowska et al. studied nine traits, including leaf length, length of outer petals, and maximum leaf width, of four *Nymphaea* species^[50]. Pan et al. collected 86 *Nymphaea* cultivars and 45 traits. These traits include floral colors, length of the leafstalk, leaf size, bud shape, and bulb shape. Results show that extensive variations occur in this population, in which floral color is the most highly diverse trait^[51].

Xu et al. find that *Nymphaea* species have an asymmetrical distribution of stomata on the leaf, with normal stomata on the adaxial side and abnormal ones on the abaxial side. Relying on the reference genome sequence of *N. colorata*, a series of core stomata regulators, including *EPF2*, *AP2C3*, *MPK6*, as well as stomatal polarity regulators *BASL* and *POLAR* genes, are absent in its genome, indicating that gene loss is a major innovation in its aquatic adaptation. This study provides an example that gene linkage phenotype analyses could be a trend in future studies^[26].

Water lilies are promising model plants for studying plant evolution and floral biology

Water lilies have some advantageous characteristics as potential model plants for genetic and genomic studies (Fig. 5). All water lilies grow into aquatic herbs, and most *Nymphaea* water lilies have small sizes, with some species even stretching up to 0.5 m in diameter. Some *Nymphaea* water lilies (i) have rapid life (around three to four months from seed to seed), and

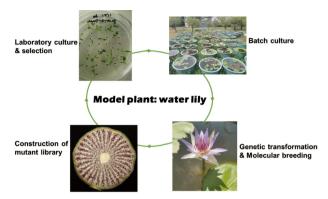


Fig. 5 The life cycle and characteristics of *Nymphaea* water lily make it a suitable model plant. Genetic transformation through its flowers is available in water lilies, similar to that in Arabidopsis. The numerous seeds in one *Nymphaea* fruit pod provide a good opportunity for the construction of a mutant library. By laboratory culture of seeds or organs, water lily could be ideal for lab research. Batch culture of water lilies is relatively easy as shown in the illustration.

bloom all the year in tropical regions, (ii) have lots of flowers in a single plant and each flower contains ~3,000 fertile seeds. These characteristics make them ideal candidates for model plants. In addition, the release of the genome sequence of *N. colorata*, together with the genetic transformation pipeline of *Nymphaea* species^[4], all allow *Nymphaea* water lilies to be ideal model plants for studying the evolution of angiosperms and floral characteristics.

Cabomba plants also have some excellent traits and growth characteristics, suitable as model plants for studying the origin of angiosperms. For example, *C. aquatica* and *C. caroliniana* can be efficiently cultivated in the laboratory^[52]. Their flowering can be induced by light, and their small size as an aquatic herb with a smaller than average genome of the ANA grade, all make *Cabomba* a suitable model plant^[52].

Trithuria, a small genus with 12 species from Australia, New Zealand, and India, grows to very a small size (~2 cm in height) and flowers very quickly in the laboratory^[53]. Its seeds can be stored conveniently^[53]. In summary, *Trithuria* spp. may provide a good opportunity for both reverse and forward genetic screening.

Future development of water lily studies

Combination of omic tools and bioinformatic techniques

The water lily order contains about 100 species with a relatively wide geographical distribution, and different species possess a large number of different metabolites. In the future, multi-omic tools will be combined in water lily research. Eventually, all water lily genome sequences will be deciphered by relying on long-length or even ultra-long read-length sequencing technology with high accuracy. Based on this, the micro-evolution of all water lilies will be deeply revealed, such as more phenotypically related genes can be mined using genome-wide association studies (GWAS) technology. Using epigenomic technology, we can deepen our understanding of gene regulation. More metabolites can be mined using metabolomics, while the association between genes and phenotypes can be studied by phenomics. In conclusion, more

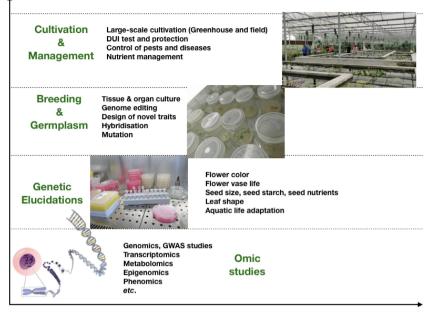


Fig. 6 Future roadmap of water lily research.

and more technologies will contribute significantly to the understanding and application of water lilies.

From genetics to breeding and industry

In the near future, we are confident that all water lily genomes will be decoded (Fig. 6). The pan-genome of water lilies will be established. Genome-wide association studies and genome editing will accelerate the genetics of water lilies. The various genes and pathways responsible for important traits will be uncovered. Afterward, these genes will be employed for intelligent molecular design breeding. Excellent varieties of water lilies will emerge, coupled with large-scale cultivation for application in the water lily industry.

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Conflict of interest

Fei Chen, Zhiqiang Xia, and Liangsheng Zhang are the Editorial Board members of Journal *Tropical Plants*. They were blinded from reviewing or making decisions on the manuscript. The article was subject to the journal's standard procedures, with peer-review handled independently of these Editorial Board members and their research groups.

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REFERENCES

 Borsch T, Löhne C, Wiersema J. 2008. Phylogeny and evolutionary patterns in Nymphaeales: integrating genes, genomes and morphology. TAXON 57:1052–4E

- Les DH, Garvin DK, Wimpee CF. 1991. Molecular evolutionary history of ancient aquatic angiosperms. *PNAS* 88:10119–23
- Saarela JM, Rai HS, Doyle JA, Endress PK, Mathews S, et al. 2007. Hydatellaceae identified as a new branch near the base of the angiosperm phylogenetic tree. *Nature* 446:312–15
- Yu C, Qiao G, Qiu W, Yu D, Zhou S, et al. 2018. Molecular breeding of water lily: engineering cold stress tolerance into tropical water lily. *Horticulture Research* 5:73
- Schneider E, Tucker SC, Williamson PS. 2003. Floral Development in the Nymphaeales. *International Journal of Plant Sciences* 164:S279–S292
- Friis EM, Pedersen KR, Crane PR. 2001. Fossil evidence of water lilies (Nymphaeales) in the Early Cretaceous. *Nature* 410:357–60
- Friedman WE. 2008. Hydatellaceae are water lilies with gymnospermous tendencies. *Nature* 453:94–97
- Pellicer J, Kelly LJ, Magdalena C, Leitch IJ. 2013. Insights into the dynamics of genome size and chromosome evolution in the early diverging angiosperm lineage Nymphaeales (water lilies). *Genome* 56:437–49
- 9. Zhang L, Chen F, Zhang X, Li Z, Zhao Y, et al. 2020. The water lily genome and the early evolution of flowering plants. *Nature* 577:79–84
- Wickett NJ, Mirarab S, Nguyen N, Warnow T, Carpenter E, et al. 2014. Phylotranscriptomic analysis of the origin and early diversification of land plants. *PNAS* 111:E4859–E4868
- Schimpf M, Ulmer T, Hiller H, Barbuto AF. 2021. Toxicity from blue lotus (*Nymphaea* caerulea) after ingestion or inhalation: A case series. *Military Medicine* 11:usab328
- Devi SA, Thongam B, Handique PJ. 2015. Nymphaea rubra Roxb. ex Andrews cultivated as an ornamental, food and vegetable in the North Eastern region of India. Genetic Resources and Crop Evolution 62:315–20
- Liu X, He Z, Yin Y, Xu X, Wu W, et al. 2018. Transcriptome sequencing and analysis during seed growth and development in *Euryale ferox* Salisb. *BMC Genomics* 19:343
- Dash BK, Sen MK, Alam K, Hossain K, Islam R, et al. 2013. Antibacterial activity of Nymphaea nouchali (Burm. f) flower. Annals of Clinical Microbiology and Antimicrobials 12:27
- Hsu CL, Fang SC, Yen GC. 2013. Anti-inflammatory effects of phenolic compounds isolated from the flowers of Nymphaea mexicana Zucc. Food & Function 4:1216–22
- Fajemiroye JO, Adam K, Jordan KZ, Alves CE, Aderoju AA. 2018. Evaluation of Anxiolytic and Antidepressant-like Activity of Aqueous Leaf Extract of Nymphaea Lotus Linn. in Mice. Iranian Journal of Pharmaceutical Research 17:613–26

- 17. Harris JC. 2007. The Water-lily pond-symphony in green. *Archives* of *General Psychiatry* 64:1347
- Lavid N, Schwartz A, Yarden O, Tel-Or E. 2001. The involvement of polyphenols and peroxidase activities in heavy-metal accumulation by epidermal glands of the waterlily (Nymphaeaceae). *Planta* 212:323–31
- The Angiosperm Phylogeny Group. 2009. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Botanical Journal of the Linnean Society* 161:105–21
- 20. Zhang LS. 2019. Advance of Horticultural Plant Genomes. *Hortic Plant J* 5:229–30
- 21. Chen F, Liu X, Yu C, Chen Y, Tang H, et al. 2017. Water lilies as emerging models for Darwin's abominable mystery. *Horticulture Research* 4:17051
- 22. Dong S, Zhao C, Chen F, Liu Y, Zhang S, et al. 2018. The complete mitochondrial genome of the early flowering plant *Nymphaea* colorata is highly repetitive with low recombination. *BMC* Genomics 19:614
- 23. Lyu J. 2020. Encoding beauty. Nature Plants 6:50
- Chen F, Zhang X, Liu X, Zhang L. 2017. Evolutionary Analysis of MIKC^C-Type MADS-Box Genes in Gymnosperms and Angiosperms. *Frontiers in Plant Science* 8:895
- 25. Chen F, Hu Y, Vannozzi A, Wu K, Cai H, et al. 2017. The WRKY Transcription Factor Family in Model Plants and Crops. *Critical Reviews in Plant Sciences* 36:311–35
- 26. Xu M, Chen F, Qi S, Zhang L, Wu S. 2018. Loss or duplication of key regulatory genes coincides with environmental adaptation of the stomatal complex in *Nymphaea* colorata and *Kalanchoe laxiflora*. *Horticulture Research* 5:42
- Povilus RA, DaCosta JM, Grassa C, Satyaki PRV, Moeglein M, et al. 2020. Water lily (*Nymphaea* thermarum) genome reveals variable genomic signatures of ancient vascular cambium losses. *PNAS* 117:8649–56
- Yang Y, Sun P, Lv L, Wang D, Ru D, et al. 2020. Prickly waterlily and rigid hornwort genomes shed light on early angiosperm evolution. *Nature Plants* 6:215–22
- 29. Takuno S, Ran JH, Gaut BS. 2016. Evolutionary patterns of genic DNA methylation vary across land plants. *Nature Plants* 2:15222
- Ke M, Gao Z, Chen J, Qiu Y, Zhang L, et al. 2018. Auxin controls circadian flower opening and closure in the waterlily. *BMC Plant Biology* 18:143
- 31. Li Z, Zhou W, Wang P, Chen Y, Huo S, et al. 2021. Transcriptome analysis reveals the senescence process controlling the flower opening and closure rhythm in the waterlilies (*Nymphaea* L.). *Frontiers in Plant Science* 12:701633
- Dias O, Tungare K, Palamthodi S, Bhori M. 2021. Nymphaea nouchali burm. f. flowers as a potential food additiveand revitalizer: A biochemico-toxicological insight. Journal of Food Processing and Preservation 45:e15045
- 33. Acharya J, De B. 2016. Bioactivity-guided fractionation to identify β-glucuronidase inhibitors in Nymphaea pubescens flower extract. Cogent Food & Agriculture 2:1134379
- Acharya J, Dutta M, Chaudhury K, De B. 2018. Metabolomics and chemometric study for identification of acetylcholinesterase inhibitor(s) from the flower extracts of Nymphaea pubescens. Journal of Food Biochemistry 42:e12575
- 35. Maia ACD, de Lima CT, Navarro DMAF, Chartier M, Giulietti AM, et al. 2014. The floral scents of Nymphaea subg. Hydrocallis (Nymphaeaceae), the New World night-blooming water lilies, and their relation with putative pollinators. *Phytochemistry* 103:67–75
- Bakr RO, El-Naa MM, Zaghloul SS, Omar MM. 2017. Profile of bioactive compounds in *Nymphaea* alba L. leaves growing in Egypt: hepatoprotective, antioxidant and anti- inflammatory activity. *BMC Complementary and Alternative Medicine* 17:52

- 37. Zhao Y, Zhou W, Chen Y, Li Z, Song X, et al. 2021. Metabolite analysis in Nymphaea 'Blue Bird' petals reveal the roles of flavonoids in color formation, stress amelioration, and bee orientation. Plant Science 312:11025
- Yuan R, Li S, Zheng X, Wu Q, Zhang H, et al. 2014. Determination of Volatiles in Water Lily Flowers Using Gas Chromatography-Mass Spectrometry. *Analytical Letters* 47:1541–51
- 39. Smith LT, Magdalena C, Przelomska NAS, et al. 2022. Revised species delimitation in the giant water lily genus Victoria (Nymphaeaceae) confirms a new species and has implications for its conservation. Frontiers in Plant Science 13:883151
- 40. Jiang Y, Liu G, Zhang W, Zhang C, Chen X, et al. 2021. Biosynthesis and emission of methyl hexanoate, the major constituent of floral scent of a night-blooming water lily *Victoria cruziana*. *Phytochemistry* 191:112899
- Zhu M, Zheng X, Shu Q, Li H, Zhong P, et al. 2012. Relationship between the Composition of Flavonoids and Flower Colors Variation in Tropical Water Lily (*Nymphaea*) Cultivars. *PLoS One* 7:e34335
- 42. Wu Q, Wu J, Li S, Zhang H, Feng C, et al. 2016. Transcriptome sequencing and metabolite analysis for revealing the blue flower formation in waterlily. *BMC Genomics* 17:897
- Wu Q, Li P, Zhang H, Feng C, Li S, et al. 2018. Relationship between the flavonoid composition and flower colour variation in Victoria. *Plant Biology* 20:674–81
- 44. Wu P, Liu A, Li L. 2021. Metabolomics and transcriptome analysis of the biosynthesis mechanism of flavonoids in the seeds of *Euryale ferox* Salisb at different developmental stages. *Molecular Genetics and Genomics* 296:953–70
- Su Q, Tian M, Li C, Li X, Lu J, et al. 2021. Transcriptome sequencing analysis of leaf vivipary in water lily. *Chinese Journal of Tropical Crops* 42:3443–50
- 46. Grob V, Moline P, Pfeifer E, Novelo AR, Rutishauser R. 2006. Developmental morphology of branching flowers in Nymphaea prolifera. Journal of Plant Research 119:561–70
- 47. Manimaran P, Ghosh S, Priyanka R. 2017. Bulb size and growth regulators on the growth and performance of bulbous ornamental crops A review. *Chemical Science Review and Letters* 6:1277–84
- Zhang L, Wu S, Chang X, Wang X, Zhao Y, et al. 2020. The ancient wave of polyploidization events in flowering plants and their facilitated adaptation to environmental stress. *Plant, Cell & Environment* 43:2847–56
- Caruana JC, Sittmann JW, Wang W, Liu Z. 2018. Suppressor of Runnerless encodes a DELLA protein that controls runner formation for asexual reproduction in strawberry. Molecular Plant 11:230–33
- 50. Dąbrowska MA, Rola K, Volkova P, Suda J, Zalewska-Gałosz J. 2015. Genome size and phenotypic variation of Nymphaea (Nymphaeaceae) species from Eastern Europe and temperate Asia. Acta Societatis Botanicorum Poloniae 84:277–86
- Pan Q, Fu Y, Gu J, Sheng Y, Li Q, et al. 2021. Analysis of phenotypic diversity of Nymphaea L. in Hainan, China. Chinese Journal of Tropical Crops 42:2777–88
- 52. Vialette-Guiraud ACM, Alaux M, Legeai F, Finet C, Chambrier P, et al. 2011. Cabomba as a model for studies of early angiosperm evolution. *Annals of Botany* 108:589–98
- Tuckett RE, Merritt DJ, Rudall PJ, Hay F, Hopper SD, et al. 2010. A new type of specialized morphophysiological dormancy and seed storage behaviour in Hydatellaceae, an early-divergent angiosperm family. *Annals of Botany* 105:1053–61

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