



<https://gmsmicrofungi.org>: an online database providing updated information of microfungi in the Greater Mekong Subregion

Chaiwan N¹, Gomdola D¹, Wang S¹, Monkai J¹, Tibpromma S^{2,3,7}, Doilom M^{4,5,6}, Wanasinghe DN^{2,3,7}, Mortimer PE^{3,7}, Lumyong S^{4,5,8} and Hyde KD^{1*}

¹Center of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai, 57100, Thailand

²East and Central Asia Regional Office, World Agroforestry Centre (ICRAF), Kunming 650201, Yunnan, People's Republic of China

³Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming 650201, Yunnan, People's Republic of China

⁴Department of Biology, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand

⁵Research Center of Microbial Diversity and Sustainable Utilization, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand

⁶Innovative Institute for Plant Health, Zhongkai University of Agriculture and Engineering, Guangzhou 510225, Guangdong Province, People's Republic of China

⁷Center for Mountain Futures, Kunming Institute of Botany, Chinese Academy of Sciences, Honghe County 654400, Yunnan, China

⁸Academy of Science, The Royal Society of Thailand, Bangkok 10300, Thailand

Chaiwan N, Gomdola D, Wang S, Monkai J, Tibpromma S, Doilom M, Wanasinghe DN, Mortimer PE, Lumyong S, Hyde KD 2021 – <https://gmsmicrofungi.org>: an online database providing updated information of microfungi in the Greater Mekong Subregion. *Mycosphere* 12(1), 1513–1526, Doi 10.5943/mycosphere/12/1/19

Abstract

The Greater Mekong Subregion is a biodiversity hotspot including Yunnan Province, China and Thailand. It is home to an extremely large diversity of microfungi. The highly variable climate and vast range of floral diversity facilitates rapid speciation and diversity in microfungi. During the last few decades, microfungi from the Greater Mekong Subregion on different substrates have been published and reclassified, and many new species have been introduced. However, numerous knowledge gaps concerning species diversity and systematics, challenge the current understanding of the fungi in the region. Basic information of microfungi in the Greater Mekong Subregion, such as taxonomic diversity, molecular phylogeny and host-specificity is incomplete, and available data have not been integrated on a specific platform where all data can be easily retrieved. To address this issue, a website: <https://gmsmicrofungi.org>, focusing on the microfungi reported from the Greater Mekong Subregion has been developed. This website is a portal to comprehensive information on microfungi and updated notes of species reported from the Greater Mekong Subregion, with easily accessible and searchable functions.

Key words – Ascomycota – Chinese mycota – fungi website – taxonomy – Thai mycota

Introduction

Microfungi play crucial roles and are important to natural ecosystems as decomposers that degrade dead organic materials. They are endophytic, pathogenic or saprobic, or they can be epiphytes and symbionts depending on the environment (Hyde et al. 2014). Microfungi are also key

organisms in the nutrient cycling process, which drives sustainable ecosystems (Vandenkoornhuysen et al. 2002, Zhang et al. 2017). They are diverse and ubiquitous heterotrophic organisms (Joshi & Chettri 2019). Mycorrhizal fungi share symbiotic relationships with plants by inhabiting plant roots (Kehri et al. 2018). They are vital for the productivity of farmland by supplying essential nutrients for plant growth (Chen et al. 2018). Saprobic microfungi penetrate substrates and enzymes are released that break down components (Bucher et al. 2004). Owing to the production of various secondary metabolites, microfungi are extremely important to human health and welfare (Jayakumar et al. 2016, Hyde et al. 2019). For example, *Penicillium* spp. are used to produce beta-lactam antibiotics, which are used in penicillin's and cephalosporins (Ozcengiz & Demain 2013). Microfungi are also important in biotechnological applications (Hyde et al. 2019).

In plant pathology, fungal species are responsible for significant economic losses and negative outcomes in agriculture, forestry and natural ecosystems (Jayawardena et al. 2020). Studies of systematics, biology and control of phytopathogenic fungi have not fully clarified lifestyles of pathogenic fungi (Hyde et al. 2014). The diversity of pathogenic fungi leads to a widespread species identification problem. As a part of natural ecosystems, they also play an important role in environments and remain poorly understood.

The current estimated species number of fungi is between 2.2 to 3.8 million (Hawksworth & Lücking 2017). It is important to establish fungal-host specificity and investigate fungal diversity in poorly studied countries or regions (Chethana et al. 2020a, Hyde et al. 2020a).

The Greater Mekong Subregion is a landscape influenced by the Mekong River, comprising Cambodia, the People's Republic of China (PRC, specifically Yunnan Province and Guangxi Zhuang Autonomous Region), Lao People's Democratic Republic (Lao PDR), Myanmar, Thailand and Vietnam (Asian Development Bank 2012). The Greater Mekong Subregion is a biodiversity hotspot spanning temperate to tropical regions (Li et al. 2018, Estoque et al. 2019) and is home to the world's tropical and evergreen forests forming unique ecosystems with high levels of biodiversity (Smith et al. 2002). Moist and dry mixed deciduous forests have different tree communities at the upper and lower elevations. These forests contain valuable timber: teak (*Tectona grandis*) as well as the largest mangrove area around the Mekong River estuaries. Forests have been converted to crop plantations in southern Yunnan (China). The establishment of plantations for the production of rubber, coffee, tea, cashew nuts, cacao, coconut, sugarcane, para rubber and palm oil are the primary drivers of forest loss (Stibig et al. 2010, Senwana et al. 2019, 2020, 2021).

The Greater Mekong Subregion microfungal diversity has been relatively well-studied in Yunnan Province, China and in northern Thailand, whereas other regions lack accurate and comprehensive documentation of fungal diversity. Yunnan Province is geographically circumscribed by the Tibet Autonomous Region to the northwest, Sichuan to the north, Guizhou to the east and the Zhuang Autonomous Region of Guangxi to the southeast. Yunnan shares an international border with Laos and Vietnam at the south and southeast periphery as well as with Myanmar at the west and southwest. Studies into Thailand's fungal diversity have been on the increase since 2015 (Ariyawansa et al. 2015, Liu et al. 2015, Chethana et al. 2020b, Hyde et al. 2020d). Only about 6,000 species descriptions have been reported from Yunnan Province in China of which around 3,000 species were higher fungi (Feng & Yang 2018), accounting for approximately 40% of all fungal taxa in China (Yang 2005). In northern Thailand, Hyde et al. (2018) reported that the percentage of new microfungi is up to 96%, showing a high diversity.

Many studies of microfungi have been carried out in the Greater Mekong Subregion, such as the diversity of terrestrial fungi on woody and leaf litter (Kodsueb et al. 2008, Boonmee et al. 2011, Promputtha et al. 2017), freshwater fungi on submerged wood (Luo et al. 2018a, b, Dong et al. 2020) and microfungi on various hosts – such as bamboo, *Chromolaena odorata*, *Clematis* spp., *Dracaena* spp., *Magnolia* spp., palms, *Pandanus* spp., *Rhododendron* spp. and *Tectona grandis* (teak) (Table 1). Karst fungi and cave fungi have also been studied in recent years (Zhang et al. 2020).

Table 1 Lists of microfungi published on various hosts and substrates in the Greater Mekong Subregion during the past decade.

Host	Location	References
Bamboo	Thailand, Yunnan (China)	Phookamsak et al. (2015), Dai et al. (2017)
<i>Camellia</i> spp.	Yunnan (China)	Abeywickrama et al. (2020)
<i>Chromolaena odorata</i>	Thailand	Mapook et al. (2020)
<i>Clematis</i> spp.	Thailand	Phukhamsakda et al. (2020)
Dead Woody Twigs	Yunnan (China)	Mortimer et al. (2021)
Decaying leaves	Vietnam	Yen et al. (2021)
<i>Dracaena</i> spp.	Thailand	Chaiwan et al. (2020a, b)
Eucalyptus plantations	Yunnan (China)	Li et al. (2020)
Insect fungi	Thailand	Xiao et al. (2019)
Leaves of plants with leaf spots or other lesions	Laos	Phengsintham et al. (2010a, b, 2013)
<i>Magnolia</i> spp.	Yunnan (China)	Wanasinghe et al. (2020)
Grasses	Thailand	Goonasekara et al. (2018), Hyde et al. (2018), Karunarathna et al. (2019, 2020), Brahmanage et al. (2020), Wanasinghe et al. (2020)
Musaceae	Thailand	Samarakoon et al. (2020)
Palms	Thailand	Wanasinghe et al. (2018a), Chen et al. (2020), Konta et al. (2020)
<i>Pandanus</i> spp.	Thailand, Yunnan (China)	Tibpromma et al. (2016, 2018)
Peanut kernels	Myanmar	Chein et al. (2019)
<i>Rhizophora</i> spp.	Thailand	Norphanphoun et al. (2019), Dayarathne et al. (2020)
<i>Rhododendron</i> spp.	Yunnan (China)	Tian et al. (2011), Li et al. (2016), Thiyagaraja et al. (2020)
<i>Rosa</i> spp.	Yunnan (China)	Wanasinghe et al. (2018b)
Submerged wood	Thailand, Yunnan (China)	Luo et al. (2019), Dong et al. (2020)
<i>Tectona grandis</i> (teak)	Thailand	Doilom et al. (2017)
Unidentified leaves	Myanmar	Nozawa et al. (2018)

However, basic information of microfungi, such as taxonomic diversity, molecular phylogeny, ecological roles, biogeographic distributions and host-specificity are either poorly understood or missing in other countries in this region, compared to Thailand and Yunnan, China. To address this issue, it is important to develop a database to collate all of the scattered information. An online platform of microfungi in the Greater Mekong Subregion will enrich our current knowledge and provide an up-to-date record.

Why we need an online database for microfungi in the Greater Mekong Subregion

The Greater Mekong Subregion is a region with a potentially high number of undiscovered fungal species. Studies continue to reveal novel taxa in the Greater Mekong Subregion at a remarkable pace (Tibpromma et al. 2016, 2018, Doilom et al. 2017, Hyde et al. 2018, 2020b, Phookamsak et al. 2019, Dong et al. 2020, Mapook et al. 2020, Phukhamsakda et al. 2020). In the case of microfungi, the numbers are likely to be even higher. This is because there are numerous cryptic species as well as undiscovered genera. The current taxonomic classification and systematics of microfungi in the Greater Mekong Subregion continue to lack up-to-date information, while many published data remain unintegrated. Therefore, it is important to collate all information in to a comprehensive, continuously updated database. The Greater Mekong Subregion website (gmsmicrofungi.org) has accordingly been configured to document all species of microfungi reported from this region. The database provides an updated list of microfungi in the Greater Mekong Subregion, specifically reporting relationships with host plants. The Greater Mekong Subregion website provides notes on each taxon reported from the Greater Mekong Subregion region, keeping abreast of current research. The Greater Mekong Subregion database, so

far, mainly incorporates microfungi reported from northern Thailand and Yunnan Province, China. The database will be further expanded to other parts of the Greater Mekong Subregion as new data becomes available. Data collection will provide a better understanding of microfungal ecology and distribution in different hosts and habitats in the Greater Mekong Subregion, and the website will serve as a directory for fungal biodiversity and host-specificity in the Greater Mekong Subregion.

What is on the website content?

The Greater Mekong Subregion website provides notes on each microfungus taxon reported from this region based on morphological and molecular data, keeping abreast of current research. Description and notes concerning placement and status are provided for each species. The entry represents published data of microfungi species reported from Greater Mekong Subregion, including species name, Faces of fungi number, Index Fungorum/MycoBank number, description, habitat, distribution, material examined, notes, photographic plates, culture, sequence availability and reference (Fig. 1).

We will further expand and include the synopses and keys to genera and species as well as other important data, wherever available through links from other related webpages including “Faces of Fungi” (Jayasiri et al. 2015; <http://www.facesoffungi.org>); “One Stop Shop” (Jayawardena et al. 2019) (<https://onestopshopfungi.org>); “Marine fungi” (Jones et al. 2019) (<http://marinefungi.org>); “Freshwater fungi” (Calabon et al. 2020) (<http://freshwaterfungi.org>); “Sordariomycetes” (Bundhun et al. 2020) (<https://sordariomycetes.org>); “Fungal Genera” (Monkai et al. 2020) (<https://www.fungalgenera.org>) and “Outline of Fungi” (Wijayawardene et al. 2020) (<https://www.outlineoffungi.org>).

Construction

All microfungi in the Greater Mekong Subregion area will be incorporated into the database according to the most recent classifications of Ascomycota (Wijayawardene et al. 2020). The database will be updated regularly as new information becomes available. Outlines, detailed descriptions and notes of each entry on the website will be carefully verified by the curators (Table 2).

Table 2 List of expert curators with their contact information.

Position	Name	Address	Contact information
Head Curators	Kevin D. Hyde	Center of Excellence in Fungal Research, School of Science Mae Fah Luang University, Chiang Rai, Thailand 57100	kdhyde3@gmail.com
	Peter E. Mortimer	Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming 650201, Yunnan, People’s Republic of China	petermortimer@mac.com
Managing curators	Dhanushka N. Wanasinghe	Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming 650201, Yunnan, People’s Republic of China	dnadeeshan@gmail.com
	Napalai Chaiwan	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	baimai_napalai@hotmail.com
Curators	Jutamart Monkai	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	mjutamart@gmail.com
	Rungtiwa Phookamsak	Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming 650201, Yunnan, People’s Republic of China	rphookamsak@outlook.com

Table 2 Continued.

Position	Name	Address	Contact information
	Mingkwan Doilom	Innovative Institute for Plant Health, Zhongkai University of Agriculture and Engineering, Guangzhou 510225, Guangdong Province, People's Republic of China	j_hammochi@hotmail.com
	Saowaluck Tibpromma	Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming 650201, Yunnan, People's Republic of China	saowaluckfai@gmail.com
	Song Wang	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	WangSong789123@outlook.com
	Deecksha Gomdola	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	deekshagomdola@gmail.com
	Erandi Yasanthika	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	eyasanthika@gmail.com
	Kunthida Phutthacharoen	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	gift_pleiades@hotmail.com
	Rashika Sajith Brahmanage	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	rashika_fernando@yahoo.com
	Guang-Cong Ren	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	guangcong.ren@gmail.com
	Binu Samarakoon	Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100	binusamarakoon2@gmail.com

Database interface and visualization

The website gmsmicrofungi.org is an online platform that compiles published information based on the classification and taxonomy of microfungi in the Greater Mekong Subregion. The website's interface is simple and user-friendly (Fig. 1). The heading provides the nine features and functions of the website. The right side of the webpage lists all recent uploaded genera and species. The search toolbar can be found above 'Recent Genus' section of the webpage. To find the genus or taxon of interest, input relevant information in the search box and a pop-up window will suggest the target fungi, including its taxonomic level (Fig. 2). Clicking on the species name will direct you to the description, notes, photographic plates and phylogenetic tree of the microfungi in Greater Mekong Subregion (Fig. 3). The references used in the description and notes are linked to the original source to obtain information about the species

The nine different features and functions in the uppermost part as following:

- 1) Home page provides the ultimate goals of the webpage, the general information of the website (The menu of the functions including the search toolbar show Home, Host/substrate, Archives, Curators, History, References, News, Contact) and an overview of the Greater Mekong Subregion microfungi. Objectives of launching the website are also provided (Fig. 1) and also the reference of this webpage are show if user using the document of this website. The right side show the recent genus and recent species updated from the webpage.
- 2) Bottom view of the homepage (Fig. 2). This information shows contact details and supporting scholars.

- 3) The Host/Substrate tab when user click to this toolbar the function will provide the data about microfungi species. This tab lists all microfungi that have been reported from Greater Mekong Subregion by fungal classification range (Fig. 3).
- 4) The Archive tab is Greater Mekong Subregion microfungi with the outline of the orders and families of Greater Mekong Subregion microfungi (Fig. 4). When the user opens the “Archives”, the list of orders related to Greater Mekong Subregion microfungi can be visualized. By clicking on a relevant order, the link will navigate to “Read more about the order” or related family list of the order. Inside families, the list of associated genera and species are available.
- 5) Search box (right side), searching species name such as *Phaeoacremonium italicum* and click “Go” to reveal the details about the species (Fig. 5). The information will show the name of microfungi, the Index Fungorum number and Facesoffungi number. The description with illustration of that microfungi, Culture characteristics, Habitat, known hosts or distribution, Material examined, GenBank Accession Number, Notes and reference of the data are also provided.
- 6) The Curators section provides the contact information and affiliated institutions of website curators (Fig. 6).
- 7) The History tab shows a short historical background of Greater Mekong Subregion microfungi, ascomycetes and host list of collection.
- 8) The References tab is a compilation of all published work (e.g., books, reviews, monographs and articles) and other information related to Greater Mekong Subregion microfungi are provided under this heading.
- 9) All activities and news related to mycology are shown in the news tab. Contact to the Home page. The ‘Contact’ section provides contact details for the website and allows users to address any comments and suggestions.

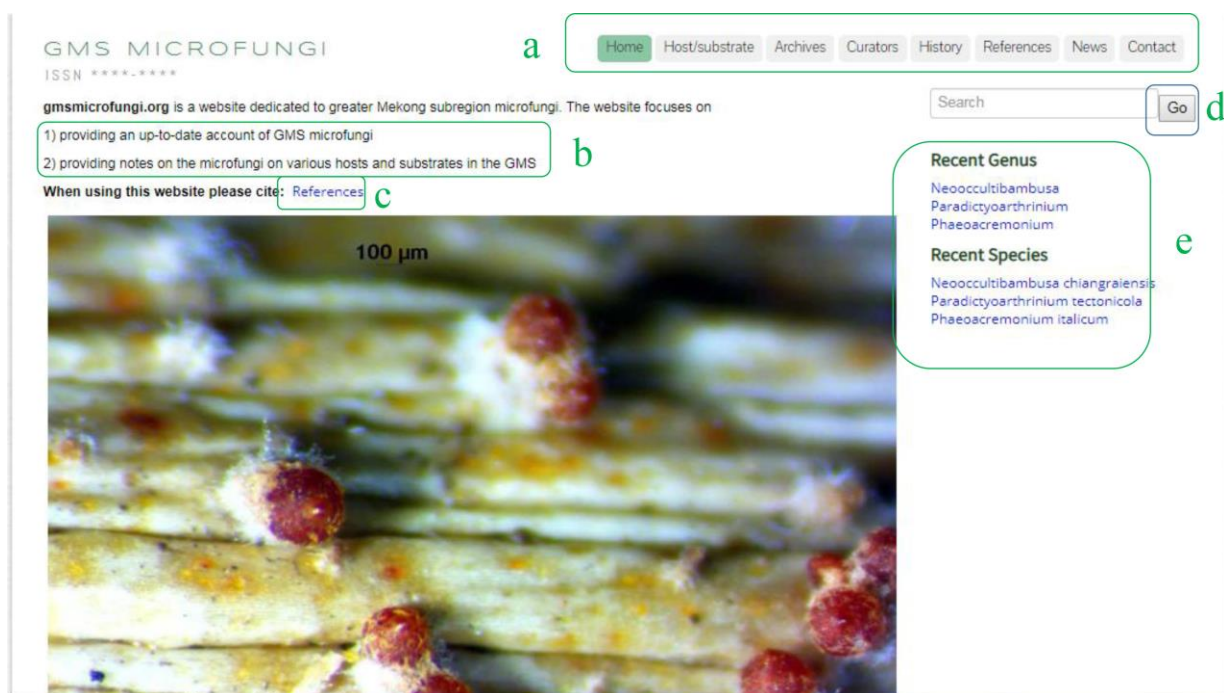


Figure 1 – The homepage view of Greater Mekong Subregion microfungi containing a photo slideshow, with Greater Mekong Subregion microfungi information, and different features and functions including the search toolbar and recent uploaded data. a Headers. b Objectives of the webpage. c Citation of the webpage. d Search box. e Recently updated news, recent genera and species.

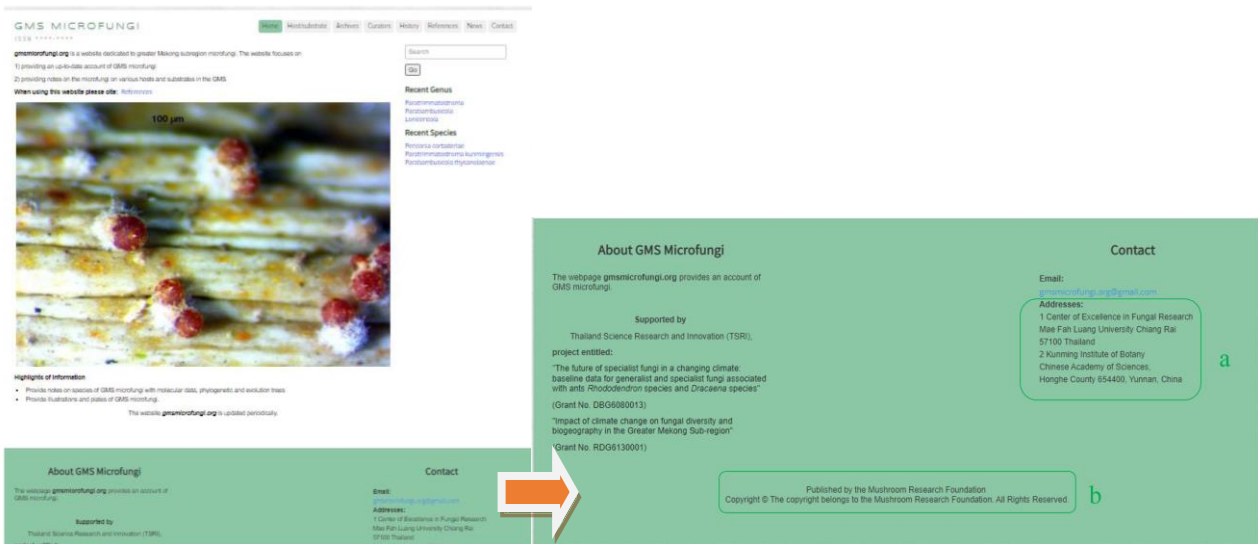


Figure 2 – Go to the bottom of the page. Bottom view of the homepage. a Contact details. b Publisher and copyright information.

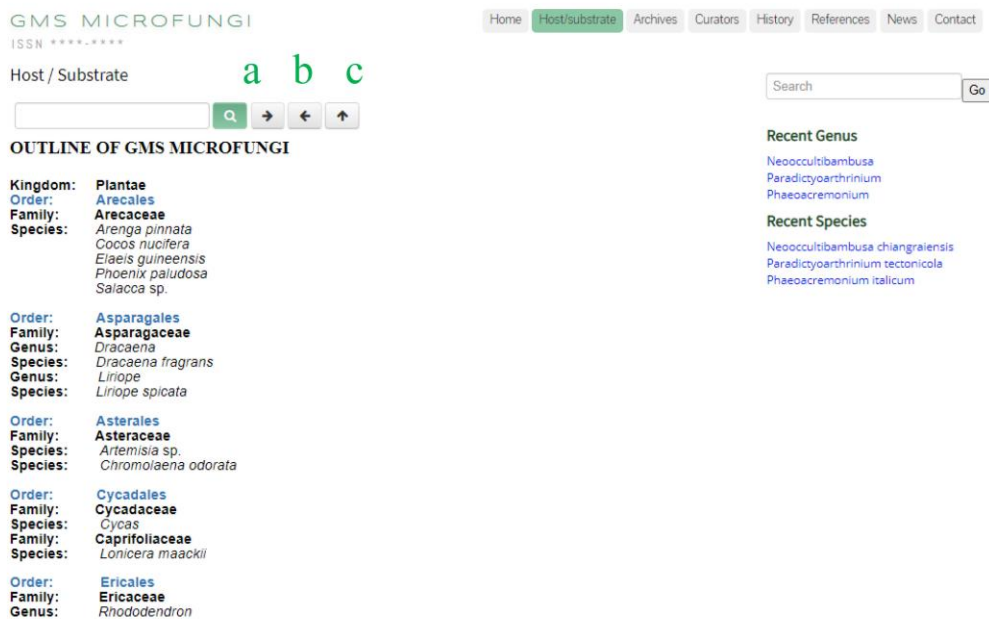


Figure 3 – The section host/substrate of the webpage. a Find next. b Find previous. c Back to top.

GMS Microfungi Hierarchy



Search Go

Recent Genus

[Neooccultibambusa](#)
[Paradictyoarthrinium](#)
[Phaeoacremonium](#)

Recent Species

[Neooccultibambusa chiangraiensis](#)
[Paradictyoarthrinium tectonicola](#)
[Phaeoacremonium italicum](#)

Figure 4 – The Archive tab with the outline of the orders and families of Greater Mekong Subregion microfungi.

Togniniales » Togniniaceae » Phaeoacremonium
Phaeoacremonium italicum

Phaeoacremonium italicum A. Carlucci & M.L. Raimondo., in Raimondo et al., Mycologia 106(6), 1123 (2014)

Facesoffungi number: FoF01876

Associated with stem wilt disease of *T. grandis*. **Sexual morph:** Undetermined. **Asexual morph:** Structures on MEA: Mycelium 1–2.6 μm broad, branched, septate, single or in bundles, partly superficial, partly immersed, hyaline to pale brown, verruculose. Conidiophores up to 40 μm long, 1–2.8 diam, erect to slightly curved, up to 2–3-septate, swollen at the base, tapering at the apex, occasionally bearing 2 lateral phialides next to the terminal phialide, wide, hyaline to pale brown, each newly proliferated segment swollen at the base. Phialides terminal or lateral, mostly monophalidic, occasion- ally polyphalidic, type I phialides (14–)16–19(–26) high × (2.2–)2.3–2.4(–2.7) μm (*x* = 18 × 2.4 μm), adelophalidic, subcylindrical, no basal septum; type II phialides (14–) 20–24 (–31) high × (1.9–)2.3–2.5(–2.9) μm (*x* = 20 × 2.4 μm), predominant, elongate-ampulliform attenu- ated at the base to subcylindrical, constricted at the base, ta- pering towards the apex; type III phialides (13–)22–29(–41) high × (2–)2.5–2.7(–3.1) μm (*x* = 25 × 2.6 μm), subcylindrical to navicular. Conidia (1–)3.6–3.9(–4.7) × (1–)1.5–1.7(–2) μm (*x* = 3.6 × 1.6 μm, *n* = 30), allantoid to oblong ellipsoidal, smooth to verrucose, hyaline.

Habitat: Known to inhabit *Vitis vinifera* (Raimondo et al. 2014) and associated with stem wilt disease of *T. grandis* (current study).

Known distribution: Italy (Raimondo et al. 2014) and Thailand (current study).

Material examined: THAILAND, Chiang Rai Province, Muang District, associated with stem wilt of *T. grandis*, 25 January 2013, M. Dollom & E. Wongsakul, MFLU 15–3427, dry culture, living culture MFLUCC 13–0336, MKT 104/1, GenBank Accession No: Actin: KU194224, TUB: KU194225.

Notes: It would appear that it is unlikely that *Phaeoacremonium italicum* (MFLUCC 13–0336) is the same as *Pm. italicum* as reported in Raimondo et al. (2014) due to differences in host and geography. In addition, it did not pro- duce a yellow pigment on MEA and PDA. Differences are evident also in the conidiophores of *Pm. italicum* (MFLUCC 13–0336) which are unbranched to branched, but unbranched for ex-type culture (Raimondo et al. 2014). However, the type II phialides in the collections in the present study predominate and the isolate MFLUCC 13–0336 is closely related to *Pm. italicum* based on DNA sequence data.



Recent Genus

[Neooccultibambusa](#)
[Paradictyoarthrinium](#)
[Phaeoacremonium](#)

Recent Species

[Neooccultibambusa chiangraiensis](#)
[Paradictyoarthrinium tectonicola](#)
[Phaeoacremonium italicum](#)

Figure 5 – Clicking a species name such as *Phaeoacremonium italicum* reveals detailed description.

Curators



Head Curator: Emeritus Prof. Kevin D. Hyde
The Director of the Center of Excellence in Fungal Research, School of Science, Mae Fah Luang University, Chiang Rai, Thailand 57100
Email: kdhyde3@gmail.com



Head Curator: Prof. Peter E. Mortimer
Key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, 65201 P.R. China
Email: petermortimer@mac.com



Curator: Dr. Jutamart Monkai



Curator: Rungtiwa Phookamsak

 Recent Genus

[Neooeclembus](#)
[Paradicthartrium](#)
[Phaeoacremonium](#)

Recent Species

[Neooeclembus chiangraensis](#)
[Paradicthartrium tectonicola](#)
[Phaeoacremonium italicum](#)

Figure 6 – Website curator information.

Discussion

Taxonomy links various databases that store information on different organisms. Several global fungal nomenclatural and taxonomic databases can be sourced to find the information about distribution of fungi. While knowledge on fungi and their hosts in tropical Southeast Asia is poor, this database will enrich our knowledge of fungi in Greater Mekong Subregion especially in Yunnan Province (China) and Thailand. The website provides basic information for mycologists to understand fungal distribution that can be exploited further.

The Greater Mekong Subregion is an ideal location to conduct diverse research on micro-fungal occurrences on specific areas/hosts. Yunnan Province of China and Thailand have tropical and subtropical climates that are favorable for fungal growth and reproduction. These areas are reported having a rich fungal diversity. More than 700 new fungal species have been described in Thailand (Hyde et al. 2018, 2020a, b, c). The idea of the webpage is to gather all this scattered published information in to one user-friendly platform.

The Greater Mekong Subregion microfungi website provides fungal classification information, host specific fungi and other related useful data in the Greater Mekong Subregion, which will enhance current understanding and ultimately enable mycologists to gain better and updated insights into the current fungal diversity in the Greater Mekong Subregion. In addition, the database also allows access to comprehensive data including descriptions of fungi, locations and specific plant-host information of fungi. This is a user-friendly database providing an up-to-date account of Greater Mekong Subregion microfungi and notes of microfungi on various hosts and substrates. The information includes Ascomycota and Eurotiomycetes (comprise four main classes Chaetothyriomycetidae, Dothideomycetes, Leotiomycetes and Sordariomycetes) in various hosts and substrates in the Greater Mekong Subregion. The webpage is managed by 12 experienced curators who upload and update information of fungi in the Greater Mekong Subregion. The present paper introduces the Greater Mekong Subregion microfungi database and provides classification and identification of the main fungal groups in the Greater Mekong Subregion. The presented data are reported from the Yunnan Province in China and Thailand.

Acknowledgements

This work was financed by the Mushroom Research Foundation (MRF), Thailand. Napalai Chaiwan is grateful to the Thailand Research Fund (PHD60K0147). Kevin D. Hyde thanks Chiang Mai University for the award of Visiting Professor. Kevin D. Hyde thanks the grants Thailand Science Research and Innovation (TSRI), project entitled: 1 the future of specialist fungi in a changing climate: baseline data for generalist and specialist fungi associated with ants, *Rhododendron* species and *Dracaena* species (Grant number: DBG6080013) and 2. The climate changes grant: Impact of climate change on fungal diversity and biogeography in the Greater Mekong Subregion (Grant number: RDG613001). Dhanushka Wanasinghe would like to thank the CAS President's International Fellowship Initiative (number 2019PC0008), the 64th batch of China Postdoctoral Science Foundation (grant no. 2018M643549), Postdoctoral Fund from Human Resources and Social Security Bureau of Yunnan Province. Saowaluck Tibpromma would like to thank the International Postdoctoral Exchange Fellowship Program (number Y9180822S1), CAS President's International Fellowship Initiative (PIFI) (number 2020PC0009), China Post doctoral Science Foundation, and the Yunnan Human Resources and Social Security Department Foundation for funding her postdoctoral research. Mingkwan Doilom and Saisamorn Lumyong thanks Chiang Mai University for their partial support of this research.

References

- Abeywickrama PD, Wanasinghe DN, Karunaratna SC, Jayawardena RS et al. 2020 – A new host report of *Diaporthe manihotia* (Diaporthales, Ascomycota) from *Camellia* sp. in Yunnan province, China. *Asian Journal of Mycology* 3, 295–306.
- Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B et al. 2015 – Fungal diversity notes 111–252 – taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 75, 27–274.
- Asian Development Bank. 2012 – Greater Mekong Subregion Atlas of the Environment 2nd Edition. Manila, Philippines.
- Brahmanage RS, Dayarathne MC, Wanasinghe DN, Thambugala KM et al. 2020 – Taxonomic novelties of saprobic Pleosporales from selected dicotyledons and grasses. *Mycosphere* 11, 2481–2541.
- Boonmee S, Zhang Y, Chomnunti P, Chukeatirote E et al. 2011 – Revision of lignicolous Tubeufiaceae based on morphological reexamination and phylogenetic analysis. *Fungal Diversity* 51, 63–102.
- Bucher VVC, Hyde KD, Pointing SB, Reddy CA. 2004 – Production of wood decay enzymes, mass loss and lignin solubilization in wood by marine ascomycetes and their anamorphs. *Fungal Diversity* 15, 1–14.
- Bundhun D, Maharachchikumbura SSN, Jeewon R, Senanayake IC et al. 2020 – <https://sordariomycetes.org/>, a platform for the identification, ranking and classification of taxa within Sordariomycetes. *Asian Journal of Mycology* 3, 13–21.
- Calabon MS, Hyde KD, Jones EBG, Chandrasiri S et al. 2020 – www.freshwaterfungi.org, an online platform for the taxonomic classification of freshwater fungi. *Asian Journal of Mycology* 3, 419–445.
- Chen M, Arato M, Borghi L, Nouri E et al. 2018 – Beneficial services of arbuscular mycorrhizal fungi – From ecology to application. *Frontiers in Plant Science* 9, 1–14.
- Chen YJ, Jayawardena RS, Bhunjun CS, Harishchanra DL et al. 2020 – *Pseudocercospora dypsidis* sp. nov. (Mycosphaerellaceae) on *Dypsis lutescens* leaves in Thailand. *Phytotaxa* 474, 218–234.
- Chaiwan N, Maharachchikumbura SSN, Wanasinghe DN, Doilom M et al. 2020a – First sexual morph record of *Sarcopodium vanilla*. *Mycotaxon* 134, 707–717.
- Chaiwan N, Wanasinghe DN, Mapook A, Jayawardena RS et al. 2020b – Novel species of *Pestalotiopsis* fungi on *Dracaena* from Thailand. *Mycology* 11, 306–315.

- Chein SH, Sadiq MB, Datta A, Anal AK. 2019 – Prevalence and identification of *Aspergillus* and *Penicillium* species isolated from peanut kernels in central Myanmar. *Journal of Food Safety* 39, e12686.
- Chethana KWT, Jayawardena RS, Hyde KD. 2020a – Hurdles in fungal taxonomy: effectiveness of recent methods in discriminating taxa. *Megataxa* 1, 114–122.
- Chethana KWT, Niranjana M, Dong W, Samarakoon MC et al. 2020b – AJOM new records and collections of fungi: 101-150. *Asian Journal of Mycology* 4, 113–260.
- Doilom M, Dissanayake AJ, Wanasinghe DN, Boonmee S et al. 2017 – Microfungi on *Tectona grandis* (teak) in northern Thailand. *Fungal Diversity* 82, 107–182.
- Dai DQ, Phookamsak R, Wijayawardene NN, Li WJ et al. 2017 – Bambusicolous fungi. *Fungal Diversity* 82, 1–105.
- Dayarathne MC, Jones EBG, Maharachchikumbura SSN, Devadatha B et al. 2020 – Morpho-molecular characterization of microfungi associated with marine based habitats. *Mycosphere* 11, 1–188.
- Dong W, Wang B, Hyde KD, McKenzie EHC et al. 2020 – Freshwater Dothideomycetes. *Fungal Diversity* 105, 319–575.
- Estoque RC, Ooba M, Avitabile V, Hijioka Y et al. 2019 – The future of Southeast Asia's forests. *Nature Communication* 10, 1–12.
- Feng B, Yang Z. 2018 – Studies on diversity of higher fungi in Yunnan, southwestern China: A review. *Plant Diversity* 40, 165–171.
- Goonasekara ID, Jayawardene RS, Saichana N, Hyde KD et al. 2018 – Checklist of microfungi on grasses in Thailand (excluding bambusicolous fungi). *Asian Journal of Mycology* 1, 88–105.
- Hawksworth DL, Lücking R. 2017 – Fungal diversity revisited: 2.2 to 3.8 million species. *Microbiology Spectrum* 5, 79–95.
- Hyde KD, Nilsson RH, Alias SA, Ariyawansa HA et al. 2014 – One stop shop: backbones trees for important phytopathogenic genera: I (2014). *Fungal Diversity* 67, 1–105.
- Hyde KD, Norphanphoun C, Chen J, Dissanayake AJ et al. 2018 – Thailand's amazing diversity: up to 96% of fungi in northern Thailand may be novel. *Fungal Diversity* 93, 215–239.
- Hyde KD, Jeewon R, Chen YJ, Bhunjun CS et al. 2020a – The numbers of fungi: is the descriptive curve flattening?. *Fungal Diversity* 103, 219–271.
- Hyde KD, Dong Y, Phookamsak R, Jeewon R et al. 2020b – Fungal diversity notes 1151–1276: taxonomic and phylogenetic contributions on genera and species of fungal taxa. *Fungal Diversity* 100, 5–277.
- Hyde KD, Chethana KWT, Jayawardena RS, Luangharn T et al. 2020c – The rise of mycology in Asia. *ScienceAsia* 46S, 1–11.
- Hyde KD, de Silva NI, Jeewon R, Bhat DJ et al. 2020d – AJOM new records and collections of fungi: 1–100. *Asian Journal of Mycology* 3, 22–294.
- Jayakumar K, Babu PG, Theja CP, Kumari SG et al. 2016 – Biomedical potentials of *Talaromyces tratensis*-A new endolichenic fungi associated with high altitude crustose lichen *Lecanora* sp. *Research Journal of Pharmaceutical Biological & Chemical Sciences* 7, 1412–1419.
- Jayasiri SC, Hyde KD, Ariyawansa HA, Bhat DJ et al. 2015 – The Faces of Fungi database: fungal names linked with morphology, phylogeny and human impacts. *Fungal Diversity* 74, 3–18.
- Jayawardena RS, McKenzie EHC, Chen YJ, Phillips AJL et al. 2019 – <https://onestopshopfungi.org/>, a webpage to enhance identification of phytopathogenic genera. *Asian Journal of Mycology* 2, 281–286.
- Jayawardena RS, Hyde KD, Chen YJ, Papp V et al. 2020 – One stop shop IV: taxonomic update with molecular phylogeny for important phytopathogenic genera: 76–100. *Fungal Diversity* 103, 87–218.
- Jones EBG, Pang KL, Abdel-Wahab MA, Scholz B et al. 2019 – An online resource for marine fungi. *Fungal Diversity* 96, 347–433.
- Joshi SR, Chetri U. 2019 – Fungi in hypogean environment: Bioprospection perspective. In *Advancing Frontiers in Mycology & Mycotechnology*, 539–561.

- Karunaratna A, Phookamsak R, Jayawardena RS, Cheewangkoon R. 2019 – The holomorph of *Neoroussoella alishanense* sp. nov. (Roussoellaceae, Pleosporales) on *Pennisetum purpureum* (Poaceae). *Phytotaxa* 406, 218–236.
- Karunaratna A, Phookamsak R, Jayawardena RS, Hyde KD et al. 2020 – *Kwanghwana miscanthi* Karun., C.H. Kuo & K.D. Hyde, gen. et sp. nov. (Phaeosphaeriaceae, Pleosporales) on *Miscanthus floridulus* (Labill.) Warb. ex K. Schum. & Lauterb. (Poaceae). *Cryptogamie Mycologie* 46, 119–132.
- Kehri HK, Akhtar O, Zoomi I, Pandey. 2018 – Arbuscular mycorrhizal fungi: Taxonomy and its systematics. *International Journal of Life Sciences Research* 6, 58–71.
- Kodsueb R, McKenzie EHC, Lumyong S, Hyde KD. 2008 – Diversity of saprobic fungi on Magnoliaceae. *Fungal Diversity* 30, 37–53.
- Konta S, Hyde KD, Phookamsak R, Xu JC et al. 2020 – Polyphyletic genera in Xylariaceae (Xylariales): *Neoxylaria* gen. nov. and *Stilbohypoxyton*. *Mycosphere* 11, 2629–2651.
- Li J, Phukhamsakda R, Mapook A, Boonmee S et al. 2016 – *Seifertia shangrilaensis* sp. nov. (Melanommataceae), a new species from Southwest China *Phytotaxa* 273, 034–042.
- Li H, Guo J, Karunaratna SC, Ye L et al. 2018 – Native Forests Have a Higher Diversity of Macrofungi Than Comparable Plantation Forests in the Greater Mekong Subregion. *Forests*: 9, 402.
- Li G, Slippers B, Wingfield MJ, Chen S. 2020 – Variation in Botryosphaeriaceae from Eucalyptus plantations in YunNan Province in southwestern China across a climatic gradient. *IMA Fungus* 11, 22–71.
- Liu JK, Hyde KD, Jones EBG, Ariyawansa HA et al. 2015 – Fungal diversity notes 1–110: taxonomic and phylogenetic contributions to fungal species. *Fungal Diversity* 72, 1–197.
- Luo ZL, Hyde KD, Liu JK, Bhat DJ et al. 2018a – Lignicolous freshwater fungi from China II: Novel *Distoseptispora* (Distoseptisporaceae) species from northwestern Yunnan Province and a suggested unified method for studying lignicolous freshwater fungi. *Mycosphere* 9, 444–461.
- Luo ZL, Hyde KD, Bhat DJ, Jeewon R et al. 2018b – Morphological and molecular taxonomy of novel species Pleurotheciaceae from freshwater habitats in Yunnan, China. *Mycological Progress* 17, 511–530.
- Luo ZL, Hyde KD, Liu JK, Maharachchikumbura SSN et al. 2019 – Freshwater Sordariomycetes. *Fungal Diversity* 99, 451–660.
- Mapook A, Hyde KD, McKenzie EHC, Jones EBG et al. 2020 – Taxonomic and phylogenetic contributions to fungi associated with the invasive weed *Chromolaena odorata* (Siam weed). *Fungal Diversity* 101, 1–175.
- Monkai J, McKenzie EHC, Phillips AJL, Hongsanan S et al. 2020 – <https://fungalgenera.org/>: a comprehensive database providing web-based information for all fungal genera. *Asian Journal of Mycology* 2, 297–304.
- Mortimer PE, Jeewon R, Xu JC, Lumyong S et al. 2021 – Morpho-phylo taxonomy of novel dothideomycetous fungi associated with dead woody twigs in Yunnan Province, China. *Frontiers in Microbiology* 12, 654–683.
- Norphanphoun C, Jayawardena RS, Chen Y, Wen TC et al. 2019 – Morphological and phylogenetic characterization of novel pestalotioid species associated with mangroves in Thailand. *Mycosphere* 10, 531–578.
- Nozawa S, Ando K, Phay N, Watanabe K. 2018 – *Pseudopestalotiopsis dawaina* sp. nov. and *Ps. kawthaungina* sp. nov.: two new species from Myanmar. *Mycological Progress* 17, 865–870.
- Ozcengiz G, Demain AL. 2013 – Recent advances in the biosynthesis of penicillins, cephalosporins and clavams and its regulation. *Biotechnology Advances* 31, 287–311.
- Phengsintham P, Chukeatirote E, Abdelsalam KA, Hyde KD et al. 2010a – *Cercospora* and allied genera from Laos 2. *Current Research in Environmental & Applied Mycology* 3, 34–158.
- Phengsintham P, Chukeatirote E, Bahkali AH, Moslem MA et al. 2010b – *Cercospora* and allied genera from Laos 3. *Cryptogamie, Mycologie* 31, 305–322.

- Phengsintham P, Chukeatirote E, McKenzie EHC, Hyde KD et al. 2013 – Monograph of Cercosporoid fungi from Laos. *Cryptogamie, Mycologie* 31, 161–181.
- Phookamsak R, Norphanphoun C, Tanaka K, Dai DQ et al. 2015 – Towards a natural classification of *Astrosphaeriella*-like species; introducing *Astrosphaeriellaceae* and *Pseudoastrosphaeriellaceae* fam. nov. and *Astrosphaeriellopsis*, gen. nov. *Fungal Diversity* 74, 143–197.
- Phookamsak R, Hyde KD, Jeewon R, Bhat DJ et al. 2019 – Fungal diversity notes 929–1035: taxonomic and phylogenetic contributions on genera and species of fungi. *Fungal Diversity* 95, 1–273.
- Phukhamsakda C, McKenzie EHC, Phillips AJL, Jones EBG et al. 2020 – Microfungi associated with *Clematis* (Ranunculaceae) with an integrated approach to delimiting species boundaries. *Fungal diversity* 120, 1–203.
- Promptutha I, Mckenzie EHC, Tennakoon DS, Lumyong S et al. 2017 – Succession and natural occurrence of saprobic fungi on leaves of *Magnolia liliifera* in a tropical forest. *Cryptogamie Mycologie* 38, 213–225.
- Samarakoon BC, Phookamsak R, Wanasinghe DN, Chomnunti P et al. 2020 – Taxonomy and phylogenetic appraisal of *Spegazzinia musae* sp. nov. and *S. deightonii* (Didymosphaeriaceae, Pleosporales) on Musaceae from Thailand. *Mycosphere* 70, 19–37.
- Senwana C, Hongsanan S, Phookamsak R, Tibpromma S et al. 2019 – *Muyocopron heveae* sp. nov. and *M. dipterocarpi* appears to have host-jumped to rubber. *Mycological Progress* 18, 741–752.
- Senwana C, Hongsanan S, Hyde KD, Cheewangkoon R et al. 2020 – First report of the sexual morph of *Pseudofusicoccum adansoniae* Pavlic, T.I. Burgess & M.J. Wingf. on Para Rubber. *Cryptogamie Mycologie* 41, 133.
- Senwana C, Mapook A, Samarakoon MC, Karunarathna A et al. 2021 – Ascomycetes on Para rubber (*Hevea brasiliensis*). *Mycosphere* 12(1), 1230–1408.
Doi 10.5943/mycosphere/12/1/18
- Smith J, Molina R, Huso MM, Luoma D et al. 2002 – Species richness, abundance, and composition of hypogeous and epigeous ectomycorrhizal fungal sporocarps in young, rotation-age, and old-growth stands of Douglas-fir (*Pseudotsuga menziesii*) in the Cascade Range of Oregon, USA. *Canadian Journal of Botany* 80, 186–204.
- Stibig HJ, Achard F, Carboni S, Rasi R et al. 2010 – Change in tropical forest cover of Southeast Asia from 1990 to 2010. *Biogeosciences* 11, 247–258.
- Thiyagaraja V, Hyde KD, Wanasinghe DN, Worthy FR et al. 2020 – Addition to Melanommataceae: a new geographical record of *Alpinaria rhododendri* from Shangri La, China. *Asian Journal of Mycology* 3, 335–344.
- Tian W, Zhang CQ, Qiao P, Milne R. 2011 – Diversity of culturable ericoid mycorrhizal fungi of *Rhododendron decorum* in Yunnan, China. *Mycologia* 103, 703–709.
- Tibpromma S, Hyde KD, McKenzie EHC, Bhat DJ et al. 2018 – Fungal diversity notes 840–928: micro-fungi associated with Pandanaceae. *Fungal Diversity* 93, 1–160.
- Tibpromma S, McKenzie EHC, Karunarathna SC, Mortimer PE et al. 2016 – *Muyocopron Garethjonesii* sp. nov. (Muyocopronales, Dothideomycetes) on *Pandanus* sp. *Mycosphere*, 7(9), 1480–1489.
- Vandenkoornhuyse P, Baldauf SL, Leyval C, Straczek J et al. 2002 – Extensive fungal diversity in plant roots. (Evolution). *Science* 295, 2051–2052.
- Yang ZL. 2005 – Evolutionary genetics of fungal diversity and biogeography of higher fungi in China. *Horizon Bioscience*, 35–62.
- Yen LTH, Yamaguchi K, Hop DV, Tsurumi Y, et al. 2021 – Phylogeny and a new species of *Polylobatispora*. *Mycoscience* 62, 176–181.
- Wanasinghe DN, Jeewon R, Jones EBG, Boonmee S et al. 2018a – Novel palmicolous taxa within Pleosporales: multigene phylogeny and taxonomic circumscription. *Mycological Progress* 17, 571–590.

- Wanasinghe DN, Phukhamsakda C, Hyde KD, Jeewon R et al. 2018b – Fungal diversity notes 709–839: taxonomic and phylogenetic contributions to fungal taxa with an emphasis on fungi on Rosaceae. *Fungal Diversity* 89, 1–236.
- Wanasinghe DN, Wijayawardene NN, Xu J, Cheewangkoon R et al. 2020 – Taxonomic novelties in *Magnolia*-associated pleosporalean fungi in the Kunming Botanical Gardens (Yunnan, China). *Plos One* 15, e0235855.
- Wijayawardene NN, Hyde KD, Al-Ani LKT, Tedersoo L et al. 2020 – Outline of fungi and fungus-like taxa. *Mycosphere* 11, 1060–1456.
- Xiao YP, Hongsanan S, Hyde KD, Brooks S et al. 2019 – Two new entomopathogenic species of *Ophiocordyceps* in Thailand. *MycKeys* 47, 53–74.
- Zhang K, Adams JM, Shi Y, Yang T et al. 2017 – Environment and geographic distance differ in relative importance for determining fungal community of rhizosphere and bulk soil. *Environmental Microbiology* 19, 3649–3659.
- Zhang ZF, Zhou SY, Eurwilaichitr L, Ingsriswang S et al. 2020 – Culturable mycobiota from Karst caves in China II, with descriptions of 33 new species. *Fungal Diversity* 106, 29–136.