

# Chapter 11

## Monitoring Selected Arthropods

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### Abstract

Arthropod populations were sampled in four study areas in southern Ohio in 1995 to document patterns of arthropod diversity and establish a baseline dataset for long-term monitoring in mixed-oak forests. Pitfall, Malaise, and blacklight traps were operated in 12 treatment units from May through September. Several insect groups were selected for detailed study due to their abundance and potential to reflect community dynamics. These taxa include ground beetles (Family: Carabidae), scarab beetles (Family: Scarabaeidae), long-horned beetles (Family: Cerambycidae), carrion beetles (Family: Silphidae), syrphid flies (Family: Syrphidae), ants (Family: Formicidae), vespid wasps (Family: Vespidae), and nocturnal moths (Order: Lepidoptera). From these 8 groups, a total of 706 species were identified, but only 15 species were relatively abundant and evenly distributed across all treatment units. In addition to these common species, several rare species were collected. This work has added to a growing database documenting forest arthropod diversity in southern Ohio.

### Introduction

Few studies have attempted to document arthropod diversity across several taxonomic groups within a single ecosystem (Dunwiddie 1991; Parsons et al. 1991; Lattin 1993). Although an all-taxa biodiversity inventory was recently initiated in the Great Smoky Mountains National Park, such studies are rare due to the immense diversity of arthropods. With limited resources, it is nearly impossible to adequately sample all arthropods in a community considering the time, labor, funding, and taxonomic expertise necessary to accomplish such a task. To further complicate matters, arthropod samples often contain many scarce species whose presence provides little information about community dynamics.

Our research efforts concentrated on a limited number of taxa based on their potential to provide information on the long-term dynamics of arthropod communities. These selected groups are common, easily collected, and readily identified insect taxa whose biology is relatively well known. Each of these groups also is associated with the forest floor, and thus are more likely to be affected by surface fires than canopy-dwelling arthropods. Prescribed fire might affect microclimatic conditions on the forest floor by increasing soil temperature and/or decreasing soil moisture.

Among surface-inhabiting arthropods, ground beetles (Coleoptera: Carabidae) are commonly monitored to study the dynamics of community change. Ground beetles are important predators of phytophagous and fungivorous insects and populations are sensitive to changes in soil surface conditions. In past studies, these beetles have served as indicators of environmental disturbance (Basedow 1990; Mossakowski et al. 1990), historical land use (Pizzolotto and Brandmayr 1990), and ecological habitat type (Eyre and Luff 1990; Maelfait and Desender 1990). Sampling these beetles via pitfall traps provides reliable estimates of populations based on their activity on the forest floor (den Boer 1985). Fluctuations in abundance and the absence or presence of certain species can provide information on environmental trends (Thiele 1977).

The scarab beetle subfamilies Scarabaeinae and Geotrupinae (Coleoptera: Scarabaeidae) also are closely associated with the soil surface and are common in southern Ohio. Most Scarabaeinae and Geotrupinae adults are dung feeders, though some species prefer fungus and carrion (Howden 1955). Wind, temperature, and humidity on the forest floor strongly affect the microhabitat of dung (biologically and physically) which in turn can affect the fecundity of these beetles (Helgesen 1967).

Another large beetle family commonly found in Ohio's deciduous forests is the long-horned beetles (Coleoptera: Cerambycidae). Most larvae in this family bore through wood and many are destructive to trees and freshly cut logs (Borrer et al. 1989). Increases in long-horned beetle populations can be indicative of increased environmental stress on nearby trees and/or an increase in the amount of dead timber.

Carrion beetles (Coleoptera: Silphidae) are not a major component of the forest floor community but are important as scavengers and are readily sampled by pitfall trapping. We paid special attention to this family in an attempt to document remaining populations of the federally endangered American burying beetle (*Nicrophorus americanus*), last reported from Ohio in a neighboring county in 1974.

In addition to beetles, several insect families also are common and readily sampled in southern Ohio forests. Ants (Hymenoptera: Formicidae) are typically the most numerous arthropods in most terrestrial ecosystems and are closely associated with soil and/or standing and downed timber. Many flies and wasps are easily collected by Malaise trapping and can be useful measures of forest floor conditions. Examples include syrphid flies (Diptera: Syrphidae) and vespid wasps (Hymenoptera: Vespidae). Some syrphids depend on nectar sources and soil surface moisture and, therefore, could be useful in assessing surface conditions. Vespids species that nest in decaying wood may also reflect surface conditions.

Due to the high diversity of moths and butterflies (Lepidoptera), there have been few studies that have comprehensively sample the lepidopteran fauna of deciduous forests (e.g. Butler et al. 1995; Teraguchi and Lublin 1999). However, many lepidopteran species could be important in assessing community changes. Larvae that feed in leaf litter or on low-growing lichens and shrubs are more likely to be affected by environmental changes to the forest floor than species whose larvae feed in tree canopies. Most nocturnal lepidopterans (primarily moths) are easily sampled with blacklight traps; however, species with a relatively short flight range are better indicators of local environmental conditions (Opler and Buckett 1970).

Our research is a component of a large-scale ecosystem management study of prescribed fire in mixed-oak forest ecosystems. Our objective was to document patterns of arthropod abundance and diversity in selected taxonomic groups to establish an adequate baseline for evaluating the effects of prescribed fire.

## Methods

### Study Areas and Experimental Design

The study areas and experimental design are described in detail in Chapter 1. Here a brief overview is provided. The four 75-90 ha study areas are located in Vinton County (Arch Rock and Watch Rock) and Lawrence County (Young's Branch and Bluegrass Ridge). The study areas are within in the Southern Unglaciated Allegheny Plateau, which is characterized by high hills, sharp ridges, and narrow valleys. Sandstones and shales are principle bedrocks. Forests are oak-dominated and the current overstory originated in the late-1800s, after the cessation of clearcutting for the charcoal iron industry. In each study area, three prescribed fire treatments were established, a control unit (CONT), an infrequent burn unit (INFR), and a frequent burn unit (FREQ).

### Field Methods

In 1995, arthropod traps were deployed near the center of each treatment unit in each of the four study areas. Traps were located at or near ridgetops near the center of the treatment units to reduce edge effects.

#### *Pitfall trapping*

In each treatment unit we installed a linear transect of twelve plastic pitfall traps (11-cm diam.) positioned approximately 10 m apart. Each trap was buried with the lip flush with the soil surface and contained 150 ml of ethylene glycol as a killing agent and preservative. A rain cover and mammal-resistant screen were installed over each trap. Traps were in place from May 10 to September 28 (Vinton County) and May 11 to October 10 (Lawrence County). Samples were retrieved and ethylene glycol replaced at weekly intervals until September, thereafter biweekly. All ground beetles (Carabidae), scarab beetles (Scarabaeidae), carrion beetles (Silphidae), and ants (Formicidae) were separated and preserved in 70 percent ethanol for identification to species. Remaining arthropods were preserved in ethanol for future identification.

#### *Malaise trapping*

One standard Townes-type Malaise trap (Townes 1962) fitted with custom wet collecting heads was installed in each treatment unit between May 10 and June 1. The collecting head concentrated specimens into an ethanol-filled jar, which was changed weekly. Traps were in place until September 28 at the Vinton County areas and October 10 at the Lawrence County areas. Contents of collecting jars were sorted and the following insect families were identified to species: long-horned beetles

**Table 1.—Number of species in selected insect families trapped at study areas in 1995 (numbers in parentheses are number of species unique to the study area).**

Family	Study Area <sup>a</sup>			
	AR	BR	WR	YB
<b>Coleoptera:</b>				
Carabidae <sup>b,c</sup>	72(10)	50(2)	64(5)	73(7)
Scarabaeidae <sup>b</sup>	18(0)	15(1)	18(0)	15(0)
Cerambycidae <sup>c,d</sup>	33(5)	29(2)	45(9)	29(5)
Silphidae <sup>c</sup>	6(0)	6(0)	8(0)	6(0)
<b>Diptera:</b>				
Syrphidae <sup>d</sup>	25(4)	15(1)	23(3)	20(3)
<b>Lepidoptera:</b>				
Arctiidae <sup>d</sup>	23(0)	26(2)	24(1)	25(3)
Noctuidae <sup>c</sup>	159(6)	156(19)	184(21)	162(20)
Notodontidae <sup>c</sup>	21(1)	25(3)	31(4)	25(1)
Saturniidae <sup>c</sup>	7(0)	8(2)	9(0)	10(0)
Sphingidae <sup>c</sup>	10(1)	8(2)	9(0)	10(0)

<sup>a</sup> - AR = Arch Rock, Vinton Furnace Experimental Forest, Vinton Township, Vinton County, Ohio. BR = Bluegrass Ridge, Wayne National Forest, Aid Township, Lawrence County, Ohio. WR = Watch Rock, Vinton Furnace Experimental Forest, Vinton Township, Vinton County, Ohio. YB = Young's Branch, Wayne National Forest, Decatur Township, Lawrence County, Ohio.

<sup>b</sup> - species collected by pitfall traps

<sup>c</sup> - species collected by light traps

<sup>d</sup> - species collected by Malaise traps

(Cerambycidae), syrphid flies (Syrphidae), ichneumon wasps (Ichneumonidae), social wasps (Vespidae), and selected Lepidoptera. Horse flies and deer flies (Tabanidae) and tachina flies (Tachinidae) also were separated but not identified to species. The remaining arthropods were preserved in ethanol for future study.

### Light-trap sampling

In each treatment unit, a BioQuip bucket-type light trap with an 8-watt fluorescent ultraviolet lamp and 12-volt timer was operated from 9 p.m. until 5 a.m. one night per week from May to September; there were 13 trap-nights at AR and WR and 10 trap-nights at BR and YB. A 2.5-liter bucket hung beneath each light containing ethyl acetate vapor and dichlorvos-impregnated strips as killing agents. Bucket contents were removed the following day,

placed in plastic bags, and then stored in a freezer for later identification.

## Results

Arthropod taxa collected by the three trapping techniques included spiders (Araneae), harvestmen (Opiliones), mites and ticks (Acari), pillbugs (Isopoda), millipedes (Diplopoda), centipedes (Chilopoda), and insects (Insecta). Of these arthropods, only selected insect families in the orders Coleoptera (beetles), Diptera (true flies), Hymenoptera (ants, bees, and wasps), and Lepidoptera (butterflies and moths) were identified to species (Appendices 1 and 2).

### Beetles (Coleoptera)

Ninety-eight species of ground beetles (Carabidae) were collected (Appendix 1). Carabid richness per study area ranged from 50 at BR to 73 at YB (Table 1). Pitfall sampling collected more than 5,100 carabids representing 45 species. However, only 7 species were captured in all 12 treatment units. Fifty-nine carabid species were collected from the light-trap samples, with 13 occurring in all four study areas. Only 7 species were collected by both trapping methods, indicating that the two techniques sample different members of this important family. Of all species, only *Galerita bicolor*, *Pterostichus tristis*, and *Synuchus impunctatus* were common and evenly distributed across all treatment units (Table 2). Pitfall and blacklight traps also captured *Carabus sylvosus*, *Piesmus submarginatus*, and *Cyclotrachelus incisus*, all previously unrecorded in Ohio (Purington and Stanton 1996). *Carabus sylvosus* was fairly common in the Vinton County pitfall traps.

Twenty-one species of scarab beetles (Scarabaeidae) were identified from pitfall and light traps combined (Appendix 1). Although more than 40 percent of these species were observed at all four study areas, only *Ateuchus histeroides*, *Geotrupes splendidus*, and *Onthophagus striatulus* occurred abundantly and evenly in all study areas (Table 2). *Ateuchus histeroides* was the most abundant scarab species in all the pitfall samples and its abundance was significantly greater at the Vinton County sites than the Lawrence County sites. Among the Geotrupinae, the most abundant species was *Geotrupes splendidus*.

Sixty-two species of long-horned beetles (Cerambycidae) were collected by all three trap types combined (Appendix 1). Cerambycidae richness was greater at WR (45 species) than the other study areas (29-33 species; Table 1). Twelve species were collected at all four study areas but no single species was found in comparable numbers across all treatment units.

**Table 2.—Mean abundance ± s.e. of 15 common insect species across study sites, 1995; means followed by the same letter are not significantly different at p=0.05 (one-way ANOVA); where no letters are shown, means are not significantly different.**

Insect Species	Study Area			
	AR	BR	WR	YB
<b>Coleoptera: Carabidae<sup>a</sup></b>				
<i>Galerita bicolor</i>	181±67.3	66±30	50±17	60.7±5.3
<i>Pterostichus tristis</i>	39.3±18.9	19±8.9	27±3	39.3±25.2
<i>Synuchus impunctatus</i>	256±155.3	27±9.5	215.7±47.1	75±9.7
<b>Coleoptera: Scarabaeidae<sup>a</sup></b>				
<i>Ateuchus histeroideis</i>	342±111.7 <sup>a</sup>	73.7±39.6 <sup>b</sup>	281±24.6 <sup>a</sup>	37±20.2 <sup>b</sup>
<i>Geotrupes splendidus</i>	34.3±2.3	13.7±4.1	50.7±18.7	17.3±4.3
<i>Onthophagus striatulus</i>	73±23.3	22.3±9.7	54.7±21.4	46±8.6
<b>Coleoptera: Silphidae<sup>a</sup></b>				
<i>Nicrophorus orbicollis</i>	240.3±25.7	97.3±54.6	220±29.5	153.7±50.6
<b>Diptera: Syrphidae<sup>b</sup></b>				
<i>Volucella vesicularia</i>	32±11.8	21.3±5.9	39.7±13.6	26.7±4.4
<b>Hymenoptera: Formicidae<sup>a</sup></b>				
<i>Aphaenogaster rudis rudis</i>	366.3±95.5	451±136.2	357.3±26.9	305±52.9
<i>Camponotus</i> spp.	314.3±96.4	377±232.2	807.7±260	485.7±85.1
<b>Lepidoptera: Saturniidae<sup>c</sup></b>				
<i>Actias luna</i>	24.7±6.6	11.3±1.3	23.7±3.3	18±4.5
<i>Automeris io</i>	40±7.5 <sup>a</sup>	18±1.2 <sup>b</sup>	24.7±5.2 <sup>ab</sup>	38.7±2.7 <sup>a</sup>
<i>Dryocampa rubicunda</i>	81±21	31.7±9.5	70±12.1	106±21.8
<b>Lepidoptera: Satyridae<sup>b</sup></b>				
<i>Cyllopsis gemma</i>	71.7±32.7	31±5.9	72.3±44.1	20±7
<b>Lepidoptera: Zygaenidae<sup>b</sup></b>				
<i>Pyromorpha dimidiata</i>	60.7±25.7	24.7±15.6	38.7±21.2	57.7±10.1

<sup>a</sup> specimens collected by pitfall traps

<sup>b</sup> specimens collected by Malaise traps

<sup>c</sup> specimens collected by blacklight trap

Eight species of carrion beetles (Silphidae) were identified, of which the burying beetle *Nicrophorus orbicollis* was most abundant, accounting for 92 percent of all Silphidae collected (Appendix 1). This species was common on all study sites and analysis of variance (ANOVA) revealed no statistically significant differences in numbers within or between areas (Table 2). No *Nicrophorus americanus* were recovered.

Incidental observations during this study included several rare beetles. Two specimens of *Megalopinus caelatus* (Staphylinidae) were collected at BR; no previous

records are known from Ohio. *Anelaphus pumilum* (Cerambycidae), collected at WR, may be the first collection in Ohio (T. K. Philips, Western Kentucky University, pers. commun.). The first record of *Pemelus costatus* (Hydrophilidae) in Ohio was collected in pitfall traps near WR and BR (M. Archangelsky, CRILAR, Argentina, pers. commun.). *Onthophilus pleurocostatus*, a hisster beetle collected in a baited pitfall trap, was new record for Ohio (P. Kovarik, Ohio State University, pers. commun.). *Platydemus erythrocerum* (Tenebrionidae), collected in a blacklight trap at WR, was also previously unrecorded in Ohio (C.A. Triplehorn, pers. commun.).

**Table 3.—Mean abundance ± s.e. of common insect families captured by Malaise traps 1 June - 22 Sept. 1995. Means followed by the same letter are not significantly different at p=0.05 (one-way ANOVA); where no letters are shown, means are not significantly different.**

Insect family	Study Area			
	AR	BR	WR	YB
<b>Diptera:</b>				
Syrphidae	31±11.5	21.3±6	39.7±17	26.7±4.4
Tabanidae	547.7±316.6 <sup>a</sup>	205.3±58.3 <sup>b</sup>	250.3±92.8 <sup>b</sup>	564.7±81.3 <sup>a</sup>
Tachinidae	71.7±1	50.3±4.4	77.7±22.9	42.3±5.5
<b>Hymenoptera:</b>				
Ichneumonidae	767.3±134.3 <sup>a</sup>	293.3±48.7 <sup>b</sup>	353.7±100 <sup>b</sup>	362.7±32.9 <sup>b</sup>

### True Flies (Diptera)

Thirty-seven species of syrphid flies (Syrphidae) were identified from Malaise trap samples (Appendix 1). Nine species were collected at all study areas, but only *Volucella vesicularia* was common and evenly distributed (Table 2). Syrphid flies were not as abundant as horseflies and deer flies (Tabanidae) or tachina flies (Tachinidae). There were significant differences in the abundance of tabanids (Tabanidae) among study areas with significantly higher numbers at AR and YB than BR and WR. The abundance of tachinids was evenly distributed across study areas (Table 3).

### Ants and Wasps (Hymenoptera)

Eight species of ants (Formicidae) were recorded from all four study areas (Appendix 1). The most abundant species (Table 2) were in the genera *Aphaenogaster* and *Camponotus* (carpenter ants). Six species of social wasps (Vespidae) were identified but were neither abundant nor evenly distributed (Appendix 1). As a family, the ichneumon wasps (Ichneumonidae) were abundant at all four areas, with significantly greater numbers at AR than the other three study areas (Table 3).

### Butterflies and Moths (Lepidoptera)

A total of 464 species in 26 families of macrolepidoptera were identified (Appendix 2). Fifty-three percent of these species were found at all four study areas. The most diverse family was the Noctuidae, of which WR had the highest richness, at 184 species (Table 1). Despite this diversity, only four moth species [*Actias luna* (Saturniidae), *Automeris io* (Saturniidae), *Dryocampa rubicunda* (Saturniidae), and *Pyromorpha dimidiata* (Zygaenidae)] and one butterfly [*Cyllopsis gemma* (Satyridae)] were abundant and evenly distributed across study areas (Table 2). Of these five species, only

*Automeris io* showed any significant differences among study areas (Table 2).

Two clearwing moths (Sessiidae) collected by light trap were new records for Ohio (Purrington and Horn 1996). *Synanthedon acerni* was collected at all four study areas. *Synanthedon scitula* was collected only at AR.

### Discussion

Overall carabid abundance collected via pitfall trapping was greater than that found in similar studies (Liebherr and Mahar 1979; Lenski 1982; MacLean and Usis 1992). However, the other studies contained fewer study sites and used periodic, rather than continuous, sampling. Likewise, carabid richness collected in these Ohio oak forests was greater than in other studies of similar habitat (Liebherr and Mahar 1979; Lenski 1982). Despite this diversity, many species only occurred in one study area. Only three species were abundant across all treatment units; these species should be important in the future as indicators of environmental change.

Unlike the carabids, species of scarab and silphid beetles were usually found in more than one study area. Species of long-horned beetles were often found in only one study area, though it is possible that more species were present in several study areas but not collected because Malaise and light trapping are not the standard method for collecting these beetles. Of these three families, only four species fit our criteria as potential indicators of environmental change.

The families Syrphidae, Tabanidae, and Tachinidae were abundant at all study areas, but the syrphid flies hold the most promise in assessing population shifts following prescribed fire due to their dependence on nectar sources and soil surface moisture. Within this family, only one species (*Volucella visicularia*) was abundant and evenly-distributed.

Of the hymenoptera, the carpenter ants (*Camponotus*) could be the most useful measure of forest floor surface conditions because they nest in decaying wood. However, their abundance in pitfall traps was determined primarily by the proximity of nests, and thus was highly variable. Therefore *Camponotus* may not be a reliable measure for comparing units or areas.

Many lepidopteran species were found at all study areas. However, many species in the diverse family Noctuidae were collected in only one study area. Of the many species collected, only five were sufficiently abundant and widely distributed to be considered potential indicators.

From these initial patterns of selected insect diversity, it is possible to identify abundant and evenly distributed species to assess the impact of altered microhabitat conditions following prescribed fire. For detecting changes in individual species, fifteen species hold the most promise as environmental indicators following prescribed fire. To detect changes in overall community composition, multivariate analyses (e.g., detrended correspondence analysis) can be used. Also, the species richness values reported here will be important for detecting changes in insect diversity following prescribed fires.

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**Appendix 1-- Insect species in selected families recovered from study sites during 1995. p = pitfall trap; m = Malaise trap; l = light trap; o = observed on site; species newly recorded for Ohio are marked with \*\*\*.**

Species	Study Area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<b>ORDER COLEOPTERA</b>									
<b>Carabidae<sup>a</sup></b>									
<i>Agonum aeruginosum</i>	1		1		<i>Dromius piceus</i>	1	1	1	1
<i>Agonum fidele</i>			1		<i>Elaphropus</i> sp	1			1
<i>Agonum harrisii</i>	1				<i>Galerita bicolor</i>	1p	p	p	p
<i>Agonum placidum</i>				p	<i>Galerita janus</i>	p	p	p	
<i>Agonum punctiforme</i>			1	p	<i>Harpalus compar</i>		1	1	1
<i>Amphasia sericea</i>			1	1	<i>Harpalus erythropus</i>	1		1	1
<i>Anatrichis minuta</i>				1	<i>Harpalus pennsylvanicus</i>				1
<i>Anisodactylus carbonarius</i>				1	<i>Lebia analis</i>				1
<i>Apenes lucidulus</i>	p	p	1p	1p	<i>Lebia atriventris</i>	1			1
<i>Apenes sinuatus</i>	1			1	<i>Lebia fuscata</i>	1		1	1
<i>Badister maculatus</i>			1	1	<i>Lebia grandis</i>		1	1	1
<i>Badister ocularis</i>				1	<i>Lebia ornata</i>				
<i>Bembidion affine</i>	1		1	1	<i>Lebia pulchella</i>	1			
<i>Bembidion rapidum</i>	1		1	1	<i>Lebia solea</i>	1	1		1
<i>Brachinus americanus</i>			p		<i>Lebia tricolor</i>	1	1	1	1
<i>Bradycellus tantillus</i>	1				<i>Lebia viridipennis</i>	1	1		1
<i>Calathus gregarius</i>	p				<i>Lebia viridis</i>	1	1	1	1
<i>Calathus opaculus</i>	p		p	1	<i>Loxandrus velocipes</i>		1	1	1
<i>Calleida viridipennis</i>	1	1	1	1	<i>Loxandrus vitiosus</i>	1		1	1
<i>Calosoma wilcoxi</i>	1				<i>Myas coracinus</i>	p	p	p	p
<i>Carabus goryi</i>	p	p	p	p	<i>Notiobia terminata</i>	1	1	1	1
<i>Carabus sylvosus</i> ***		p		p	<i>Notiophilus aeneus</i>	p	p	p	p
<i>Chlaenius emarginatus</i>	1p	p	1p	p	<i>Oodes amaroides</i>	1		1	1
<i>Chlaenius tricolor</i>	1		1	1	<i>Pasimachus punctulatus</i>	p		p	p
<i>Cicindela sexguttata</i>	mo	o	o	o	<i>Patrobus longicornis</i>			1	
<i>Cicindela unipunctata</i>	p				<i>Pentagonica picticornis</i>		1		1
<i>Clivina americana</i>	1			1	<i>Piesmus submarginatus</i> ***	p			p
<i>Clivina bipustulata</i>	1	1	1	1	<i>Platynus cincticollis</i>	1			1
<i>Clivina dentipes</i>	1			1	<i>Platynus tenuicollis</i>	1	1	1	1
<i>Clivina impressifrons</i>				1	<i>Plochionus timidus</i>	1	1	1	1
<i>Coptodera aerata</i>		1	1	1	<i>Poecilus chalcites</i>			1	
<i>Cyclotrachelus convivus</i>	p	p	p	p	<i>Pterostichus adoxus</i>	p	p	p	p
<i>Cyclotrachelus freitagi</i>			p		<i>Pterostichus atratus</i>	p	p		p
<i>Cyclotrachelus incisus</i> ***	p		p		<i>Pterostichus lachrymosus</i>	p			
<i>Cymindis americanus</i>	p	p	p	p	<i>Pterostichus moestus</i>	p			
<i>Cymindis limbatus</i>	1p	1	1	1p	<i>Pterostichus permundus</i>			p	
<i>Cymindis neglectus</i>	p		p		<i>Pterostichus relictus</i>	p		p	p
<i>Cymindis platicollis</i>		1	1p	1	<i>Pterostichus sayanus</i>	p	p		
<i>Dicaelus ambiguus</i>	p		p	p	<i>Pterostichus stygicus</i>	p	p		
<i>Dicaelus dilatatus</i>		p		p	<i>Pterostichus tristis</i>	p	p	p	p
<i>Dicaelus elongatus</i>	p		p	p	<i>Rhadine caudata</i>	p	p	p	p
<i>Dicaelus furvus</i>		p			<i>Scaphinotus andrewsii mutabilis</i>	p	p		p
<i>Dicaelus politus</i>	p	p	p	p	<i>Scaphinotus unicolor heros</i>	p			
<i>Dicaelus purpuratus</i>	p	p	p	p	<i>Selenophorus hylacis</i>	1		1	1
<i>Dicaelus teter</i>	p	p	p	p	<i>Selenophorus opalinus</i>	1	1	1	1
					<i>Sphaeroderus stenostomus lecontei</i>	p	p	p	p
					<i>Stenolophus comma</i>		1	1	



Appendix 1 cont.

Species	Study Area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<i>Stenolophus lecontei</i>	1	1	1	1	<i>Elaphidion mucronatum</i>		m	m	m
<i>Stenolophus ochropezus</i>	1	1	1	1	<i>Elaphidionoides asperses</i>	m	m	m	m
<i>Synuchus impunctatus</i>	p	p	p	p	<i>Elaphidionoides parallelus</i>			1	
<i>Trichotichus autumnalis</i>		p	p	p	<i>Elaphidionoides villosus</i>	1m	1m	1m	1m
<i>Trichotichnus dichrous</i>	1	1	1	1	<i>Enaphalodes rufulus</i>	1		1	1
<i>Trichotichnus vulpeculus</i>	1	1	1	1	<i>Euderces picepes</i>			m	m
<i>Zuphium americanum</i>	1				<i>Gaurotes cyanipennis</i>	m	m	m	m
<b>Scarabaeidae<sup>c</sup></b>					<i>Goes debilis</i>			m	m
<i>Ateuchus histeroides</i>	1p	1p	1p	1p	<i>Goes pulcher</i>			1	
<i>Canthon chalcites</i>	p		p	p	<i>Goes pulverulentus</i>			1m	1m
<i>Canthon viridis</i>	p		p	p	<i>Goes tessellatus</i>	m	1	1	
<i>Cloeotus globosus</i>	1	1			<i>Goes tigrinus</i>	1		1	1
<i>Copris fricator</i>	p		p	p	<i>Graphisurus fasciatus</i>		m	1m	
<i>Copris minutus</i>		1p	1p	1p	<i>Hesperophanes pubescens</i>			1	
<i>Dichelonyx elongata</i>	1	1	1	1	<i>Heterachthes quadrimaculatus</i>	1		m	
<i>Dichotomius carolinus</i>	1		1		<i>Hetoemis cinerea</i>			1m	
<i>Dynastes tityus</i>		1			<i>Hyperplatys aspersa</i>				m
<i>Eucanthus lazarus</i>	1p	1p	1p	1	<i>Knulliana cincta</i>	1		1	
<i>Geotrupes balyi</i>	p		p		<i>Lepturges angulatus</i>	m			
<i>Geotrupes hornii</i>	1p	p	p		<i>Lepturges confluens</i>			1	
<i>Geotrupes semiopacus</i>		p	p		<i>Mecas pergrata</i>		m		
<i>Geotrupes splendidus</i>	p	p	p	p	<i>Metacmaeops vittata</i>	m	m	m	m
<i>Onthophagus orpheus canadensis</i>	p		p		<i>Monochamus titillator</i>			1	
<i>Onthophagus hecate</i>	p	p	p	p	<i>Necydalis mellita</i>	m			
<i>Onthophagus s. striatulus</i>	1p	p	1p	p	<i>Neoclytus acuminatus</i>	m	m	m	m
<i>Pelidnota punctata</i>	1	1	1	1	<i>Neoclytus mucronatus</i>	m	m	m	
<i>Phileurus valgus</i>	1	1	1	1	<i>Oberea bimaculata</i>			m	
<i>Popillia japonica</i>	o	o	o	o	<i>Oberea ruficollis</i>	m		m	m
<i>Valgus canaliculatus</i>		1	1	1	<i>Oberea tripunctata</i>	m			m
<i>Xyloryctes jamaicensis</i>	1p			p	<i>Orthosoma brunneum</i>	1m	1m	1	1m
<b>Cerambycidae<sup>b</sup></b>					<i>Physocnemum violaceipenne</i>			m	
<i>Aegoschema modesta</i>			m	1	<i>Prionus laticollis</i>	p			
<i>Amniscus macula</i>	1m	m	m		<i>Psyrassa unicolor</i>			m	
<i>Anelaphus pumilus</i>				1	<i>Rhopalophora longipes</i>				m
<i>Analeptura lineola</i>	m	m	m		<i>Saperda candida</i>			1	
<i>Aneflomorpha subpubescens</i>	m	m	m	m	<i>Saperda discoidea</i>		1m	1m	
<i>Astyleiopus variegatus</i>		m			<i>Saperda lateralis</i>	m	m		m
<i>Bellamira scalaris</i>	m	m			<i>Sarosesthes fulminans</i>		m	m	
<i>Brachyleptura rubrica</i>				m	<i>Smodicum cucujiforme</i>	1			
<i>Clytus ruricola</i>		m		m	<i>Stenocorus cinnamopterus</i>		1	1	
<i>Cyrtophorus verrucosus</i>	m				<i>Strangalepta abbreviata</i>	m		m	
<i>Distenia undata</i>	m	m			<i>Strangalia bicolor</i>	m	m	m	m
<i>Eburia quadrigeminata</i>			m	m	<i>Strangalia luteicornis</i>	m	m	m	m
<i>Ecyrus dasycerus</i>	1m		1m		<i>Strophiona nitens</i>	m	m	m	m
					<i>Tylonotus bimaculatus</i>				1
					<i>Typocerus lugubris</i>	m	m	m	m

Appendix 1 cont.

Species	Study Area			
	AR	BR	WR	YB
<i>Typocerus velutinus</i>	m	m	m	
<i>Urgleptes querci</i>		m	m	
<i>Xylotrechus colonus</i>	m	m	m	m
<b>Silphidae<sup>d</sup></b>				
<i>Necrodes surinamensis</i>			1	1
<i>Necrophila americana</i>	1p	p	p	p
<i>Nicrophorus orbicollis</i>	1p	1p	1p	1p
<i>Nicrophorus pustulatus</i>	1	1	1p	1
<i>Nicrophorus sayi</i>	1p	1p	1p	1p
<i>Nicrophorus tomentosus</i>	p	p	p	p
<i>Oiceoptoma inaequale</i>			p	p
<i>Oiceoptoma noveboracense</i>	p		p	
<b>ORDER DIPTERA</b>				
<b>Syrphidae<sup>e</sup></b>				
<i>Ceriana abbreviata</i>	m		m	
<i>Chilosia pallipes</i>				m
<i>Chilosia dives</i>	m	m	m	m
<i>Chrysotoxum perplexum</i>	m	m	m	m
<i>Dasysyrphus amolopsus</i>	m			
<i>Didea fuscipes</i>	m		m	
<i>Epistrophe</i> sp.	m	m		
<i>Eristalis arbustorum</i>		m		
<i>Eristalis bastardi</i>	m			
<i>Eristalis flavipes</i>	m		m	m
<i>Eristalis obscurus</i>	m	m	m	m
<i>Ferdinandea dives</i>	m	m	m	
<i>Melanostoma obscurum</i>			m	m
<i>Merodon equestris</i>				m
<i>Mesogramma geminata</i>	m		m	
<i>Mesogramma marginata</i>	m	m	m	m
<i>Metasyrphus emarginatus</i>	m			m
<i>Milesia virginensis</i>	m	m	m	m
<i>Myiolepta nigra</i>		m		m
<i>Myiolepta varipes</i>				m
<i>Platycheirus peltitoides</i>	m			
<i>Rhingia nasica</i>				m
<i>Somula decora</i>	m			m
<i>Sphaerophoria scripta</i>			m	
<i>Sphecomomyia vittata</i>			m	
<i>Sphegina</i> sp.	m			
<i>Spilomyia hamifera</i>			m	m

Species	Study Area			
	AR	BR	WR	YB
<i>Spilomyia longicornis</i>	m		m	
<i>Syrphus</i> sp.	m	m	m	m
<i>Temnostoma balyras</i>	m		m	
<i>Temnostoma trifa</i>		m	m	
<i>Toxomerus geminatus</i>	m	m	m	m
<i>Volucella vesicularia</i>	m	m	m	m
<i>Xanthogramma flavipes</i>	m	m	m	m
<i>Xylota chalybea</i>		m		m
<i>Xylota flavitibia</i>	m		m	
<i>Xylota pigra</i>	m			m
<b>ORDER HYMENOPTERA</b>				
<b>Formicidae<sup>f</sup></b>				
<i>Aphaenogaster rudis rudis</i>	p	p	p	p
<i>Camponotus americanus</i>	p	p	p	p
<i>Camponotus ferrugineus</i>	p	p	p	p
<i>Camponotus pennsylvanicus</i>	p	p	p	p
<i>Camponotus subbarbatus</i>	p	p	p	p
<i>Crematogaster lineolata</i>	p	p	p	p
<i>Formica subsericea</i>	p	p	p	p
<i>Prenolepis imparis imparis</i>	p	p	p	p
<b>Vespidae<sup>g</sup></b>				
<i>Dolichovespula maculata</i>	m	m	m	m
<i>Polistes metricus</i>	m		m	
<i>Vespa crabro</i>			m	
<i>Vespula germanica</i>	m		m	m
<i>Vespula maculifrons</i>	m	m	m	m
<i>Vespula squamosa</i>	m	m	m	m

<sup>a</sup> Specific determinations by F. F. Purrington.

<sup>b</sup> Specific determinations by T. K. Philips, D. J. Horn, D. M. Osborne and P. Blades.

<sup>c</sup> Specific determinations by T. K. Philips, F. F. Purrington and A. E. Smith.

<sup>d</sup> Specific determinations by D. J. Horn, G. D. Keeney and F. F. Purrington.

<sup>e</sup> Tentative determinations by E. Johnson and D. J. Horn.

<sup>f</sup> Specific determinations by G. Coovert, W. D. Raby and C. Ranger.

<sup>g</sup> Specific determinations by D. J. Horn.

**Appendix 2—Lepidopteran species recovered from study sites during 1995; m = Malaise trap; l = light trap; o = observed on site; species newly recorded for Ohio are marked with \*\*\*.**

Species <sup>a</sup>	Study area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<b>Oecophoridae</b>					<i>Polygonia comma</i>			o	
<i>Antaeotrichia schlaegeri</i>	l	l	l	l	<i>Nymphalis antiopa</i>	o		o	
<b>Sesiidae</b>					<i>Vanessa atalanta</i>	o	o	o	o
<i>Synanthedon scitula</i> ***		l			<i>Speyeria cybele</i>	mo	m	o	
<i>Synanthedon acerni</i> ***	l	l	l	l	<i>Chlosyne nycteis</i>			o	
<b>Cossidae</b>					<i>Phyciodes tharos</i>	o	m	mo	m
<i>Prionoxystus robiniae</i>		l	l	l	<i>Limenitis astyanax</i>	mo	m	mo	
<b>Tortricidae</b>					<b>Satyridae</b>				
<i>Argyrotaenia alisellana</i>		l		l	<i>Enodia portlandia</i>	m		l	
<b>Hesperiidae</b>					<i>Cyllopsis gemma</i>	m	m	m	m
<i>Epargyreus clarus</i>	o		o		<i>Hermeuptychia sosybius</i>		m		
<i>Thorybes pylades</i>	m				<i>Megisto cymela</i>	mo	m	m	m
<i>Erynnis brizo</i>	m	m	m	m	<i>Cercyonis pegala</i>	m	m	lm	
<i>Erynnis juvenalis</i>	m	m	m	m	<b>Zygaenidae</b>				
<i>Erynnis horatius</i>	m	m	m	m	<i>Pyromorpha dimidiata</i>	m	lm	m	m
<i>Pholisora catullus</i>	m				<b>Megalopygidae</b>				
<i>Ancyloxipha numitor</i>		m			<i>Lagoa crispata</i>	l	l	l	l
<i>Thymelicus lineola</i>			m		<i>Norape ovina</i>		l	l	l
<i>Polites peckius</i>	m	m	m	m	<b>Limacodidae</b>				
<i>Poanes hobomok</i>			m		<i>Apoda yinversum</i>	l	l	l	l
<b>Papilionidae</b>					<i>Apoda biguttata</i>	l	l	l	l
<i>Papilio polyxenes</i>	o		o		<i>Prolimacodes badia</i>	l	l	l	l
<i>Papilio glaucus</i>	o	o	o	o	<i>Natada nasoni</i>			l	
<i>Papilio troilus</i>	mo	m	mo	m	<i>Euclea delphinii</i>	l	l	l	l
<i>Eurytides marcellus</i>		o	o		<i>Parasa chloris</i>	l	l	l	l
<b>Pieridae</b>					<b>Pyralidae</b>				
<i>Pieris rapae</i>	o	m	o	o	<i>Ostrinia nubilalis</i>	l	l	l	l
<i>Anthocharis midea</i>	o		o		<i>Desmia funeralis</i>	l	l	l	l
<i>Colias philodice</i>			o		<i>Palipita magniferalis</i>	l	l	l	l
<b>Lycaenidae</b>					<i>Pantographa limata</i>	l	l	l	l
<i>Feniseca tarquinius</i>	m		m	m	<i>Euzophera ostricolorata</i>		l		
<i>Satyrium calanus</i>		m			<b>Thyatiridae</b>				
<i>Calycopis cecrops</i>	m	m	m	m	<i>Habrosyne scripta</i>	l			
<i>Incisalia henrici</i>		o	o		<i>Pseudothyatira cymatophoroides</i>	l	l	l	l
<i>Glaucopsyche lygdamus</i>		m			<b>Drepanidae</b>				
<i>Everes comyntas</i>	m	m	m	m	<i>Drepana arcuata</i>	l		l	l
<i>Celastrina ladon</i>	mo	mo	mo	mo	<i>Oreta rosea</i>	l	l	l	l
<b>Nymphalidae</b>									
<i>Polygonia interrogationis</i>	o		o	o					

Appendix 2 cont.

Species <sup>a</sup>	Study area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<b>Geometridae</b>					<b>Mimallonidae</b>				
<i>Itame pustularia</i>	1	1	1	1	<i>Lacosoma chiridota</i>		1		1
<i>Semiothisa promiscuata</i>	1	1	1	1	<i>Cicinnus melsheimeri</i>	1	1	1	1
<i>Semiothisa granitata</i>	1	1	1	1	<b>Apatelodidae</b>				
<i>Semiothisa quadrinotaria</i>	1	1	1	1	<i>Apatelodes torrefacta</i>	1	1	1	1
<i>Glena cribrataria</i>		1	1	1	<i>Olceclostera angelica</i>	1	1	1	1
<i>Iridopsis larvaria</i>	1	1	1	1	<b>Lasiocampidae</b>				
<i>Anavitrinella pampinaria</i>	1	1	1	1	<i>Malacosoma disstria</i>	1	1	1	1
<i>Epemecis hortaria</i>	1	1	1	1	<i>Malacosoma americanum</i>	1	1	1	1
<i>Melanolophia canadaria</i>		1		1	<b>Saturniidae</b>				
<i>Melanolophia signataria</i>	1	1	1	1	<i>Eacles imperialis</i>	1	1	1	1
<i>Biston betularia</i>	1	1	1	1	<i>Citheronia regalis</i>	1	1	1	1
<i>Hypagirtis unipuncta</i>	1	1	1	1	<i>Citheronia sepulcralis</i>		1		
<i>Lomographa vestaliata</i>	1		1		<i>Sphingicampa bisecta</i>			1	1
<i>Lytrosia unitaria</i>	1	1	1	1	<i>Dryocampa rubicunda</i>	1	1	1	1
<i>Euchlaena amoenaria</i>	1		1	1	<i>Anisota stigma</i>	1	1	1	1
<i>Euchlaena tigrinaria</i>	1	1	1	1	<i>Anisota virginiensis</i>		1	1	
<i>Euchlaena irrorata</i>	1	1	1	1	<i>Automeris io</i>	1	1	1	1
<i>Nacophora quernaria</i>	1	1	1	1	<i>Antheraea polymphemus</i>	1	1	1	1
<i>Campaea perlata</i>	1	1	1	1	<i>Actias luna</i>	1	1	1	1
<i>Metarranthis hypochraria</i>				1	<i>Callosamia promethea</i>		1		1
<i>Probole nyssaria</i>			1	1	<i>Callosamia angulifera</i>	1	1	1	1
<i>Probole amicularia</i>	1	1	1	1	<b>Sphingidae</b>				
<i>Plagodis serinaria</i>	1		1	1	<i>Manduca jaminearum</i>		1		
<i>Plagodis kuetzingi</i>	1	1	1	1	<i>Dolba hyloeus</i>		1		
<i>Plagodis phlogosaria</i>	1	1	1	1	<i>Ceratonia undulosa</i>	1	1	1	1
<i>Plagodis alchoolaria</i>	1	1	1	1	<i>Lapara coniferarum</i>		1	1	
<i>Plagodis fervidaria</i>	1	1	1	1	<i>Smerinthus jamaicensis</i>			1	
<i>Besma endopriaria</i>	1		1		<i>Paonias excaecatus</i>	1	1	1	1
<i>Besma quercivoraria</i>	1	1	1	1	<i>Paonias myops</i>		1	1	1
<i>Tetracis crocallata</i>	1	1	1	1	<i>Laotioe juglandis</i>	lm	m	lm	m
<i>Tetracis cachexiata</i>	1	1	1	1	<i>Amphion floridana</i>		m		
<i>Prochoerodes transversata</i>			1	1	<i>Pachysphinx modesta</i>	1		1	
<i>Nematocampa limbata</i>	1		1	1	<i>Sphexcodina abbottii</i>		1		1
<i>Nemoria bistriaria</i>	1	1	1	1	<i>Deidamia inscripta</i>	1	1	1	1
<i>Nemoria mimosaria</i>	1	1	1	1	<i>Darapsa myron</i>		1	1	1
<i>Dichorda iridaria</i>	1	1	1	1	<i>Darapsa pholus</i>	1	1	1	
<i>Cyclophora packardi</i>	1		1	1	<b>Notodontidae</b>				
<i>Scopula limboundata</i>	1	1	1	1	<i>Clostera albostigma</i>			1	1
<i>Eulithis diversilineata</i>	1	1	1	1	<i>Clostera inclusa</i>	1	1	1	1
<i>Ecliptopera atricolorata</i>	1		1	1	<i>Datana ministra</i>		1		1
<i>Hydriomena transfigurata</i>	1	1	1	1	<i>Datana angusii</i>	1	1	1	1
<i>Hydria prunivorata</i>	1	1	1	1	<i>Datana drexelii</i>	1	1	1	1
<i>Orthonama obstipata</i>	1	1	1	1	<i>Datana major</i>		1		1
<i>Orthonama centrostrigaria</i>	1	1	1	1					
<i>Eubaphe mendica</i>	1	1	1	1					
<i>Eupithecia miserulata</i>	1	1	1	1					
<i>Dyspteris abortivaria</i>	1	1	1	1					

Appendix 2 cont.

Species <sup>a</sup>	Study area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<i>Datana contracta</i>	1				<i>Estigmene acrea</i>		1		
<i>Datana integerrima</i>		1	1		<i>Spilosoma latipennis</i>		1		1
<i>Nadata gibbosa</i>	1	1	1	1	<i>Spilosoma congrua</i>	1	1	1	1
<i>Hyperaeschra georgica</i>	1	1	1	1	<i>Spilosoma virginica</i>	1	1	1	
<i>Peridea basitriens</i>	1	1	1	1	<i>Hyphantria cunea</i>	1	1	1	1
<i>Peridea angulosa</i>	1	1	1	1	<i>Ecpantheria scribonia</i>	1	1	1	1
<i>Pheosia rimosa</i>	1		1		<i>Apantesis phalerata</i>			1	
<i>Odontotia elegans</i>	1		1		<i>Apantesis carlota</i>				1
<i>Nerice bidentata</i>		1		1	<i>Grammia anna</i>	1	1	1	1
<i>Ellida caniplaga</i>	1	1	1	1	<i>Grammia figurata</i>	1	1	1	1
<i>Gluphisia septentrionis</i>			1	1	<i>Grammia parthenice</i>	1	1	1	1
<i>Furcula borealis</i>				1	<i>Grammia virgo</i>				1
<i>Furcula cinerea</i>			1		<i>Halysidota tessellaris</i>	1	1	1	1
<i>Symmerista albifrons</i>	1	1	1	1	<i>Lophocampa caryae</i>	1	1	1	1
<i>Dasylophia anguina</i>	1	1	1	1	<i>Cycnia tenera</i>	1	1	1	1
<i>Dasylophia thyatiroides</i>	1		1	1	<i>Euchaetis egle</i>	1	1	1	1
<i>Misogada unicolor</i>		1			<i>Cisseps fulvicollis</i>	1	1	1	1
<i>Marcurocampa marthesia</i>	1	1	1	1					
<i>Heterocampa obliqua</i>	1	1	1	1	<b>Lymantriidae</b>				
<i>Heterocampa subrotata</i>	1				<i>Dasychira tephra</i>	1	1	1	1
<i>Heterocampa umbrata</i>	1	1	1	1	<i>Dasychira basiflava</i>	1	1	1	1
<i>Heterocampa guttivitta</i>	1	1	1	1	<i>Dasychira obliquata</i>	1	1	1	1
<i>Heterocampa biundata</i>		1	1	1	<i>Orgyia definita</i>			1	1
<i>Lochmaeus manteo</i>		1	1	1	<i>Orgyia leucostigma</i>	1	1	1	1
<i>Lochmaeus bilineata</i>	1	1	1	1					
<i>Schizura ipomoeae</i>	1	1	1	1	<b>Noctuidae</b>				
<i>Schizura unicornis</i>			1		<i>Idia americalis</i>	1	1	1	1
<i>Schizura concinna</i>			1		<i>Idia aemula</i>	1	1	1	1
<i>Schizura leptinoides</i>		1	1	1	<i>Idia majoralis</i>				1
<i>Oligocentria semirufescens</i>		1	1		<i>Idia rotundalis</i>	1	1	1	1
<i>Oligocentria lignicolor</i>	1	1	1	1	<i>Idia forbesi</i>	1	1	1	1
<i>Hyparpax aurora</i>				1	<i>Idia julia</i>	1	1	1	
<b>Arctiidae</b>					<i>Idia diminuendis</i>	1	1	1	1
<i>Crambidia pallida</i>	1	1	1	1	<i>Idia scobialis</i>	1		1	1
<i>Crambidia uniformis</i>	1	1	1	1	<i>Idia denticulalis</i>		1		1
<i>Crambidia cephalica</i>		1	1	1	<i>Idia lubricalis</i>			1	1
<i>Cisthene plumbea</i>	1	1	1	1	<i>Zanclognatha laevigata</i>	1			
<i>Cisthene packardii</i>		1	1		<i>Zanclognatha obscuripennis</i>		1		1
<i>Lycomorpha pholus</i>	1		1	1	<i>Zanclognatha pedipilalis</i>			1	
<i>Hypoprepia miniata</i>		1			<i>Zanclognatha martha</i>				1
<i>Hypoprepia fucosa</i>	1	1	1	1	<i>Zanclognatha cruralis</i>	1	1	1	1
<i>Clemensia albata</i>	1	1	1	1	<i>Zanclognatha jacchusalis</i>	1		1	
<i>Pagara simplex</i>				1	<i>Zanclognatha ochreipennis</i>		1	1	1
<i>Haploa clymene</i>	1	1	1	1	<i>Chytolita morbidalis</i>		1	1	
<i>Haploa contigua</i>	1	1	1	1	<i>Macrochilo absorptalis</i>			1	
<i>Haploa lecontei</i>	1	1	1	1	<i>Phalaenostola larentioides</i>			1	
<i>Holomelina opella</i>	1	1	1	1	<i>Tetanolita floridana</i>				1
<i>Pyrrharctia isabella</i>	1		1	1	<i>Bleptina caradrinalis</i>	1	1	1	1
					<i>Renia salusalis</i>		1	1	

Appendix 2 cont.

Species <sup>a</sup>	Study area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<i>Renia factorialis</i>				1	<i>Caenurgina erechtea</i>	1	1	1	1
<i>Renia nemoralis</i>				1	<i>Mocis texana</i>	1		1	1
<i>Renia discoloralis</i>	1	1	1	1	<i>Celipetra frustulum</i>	1	1	1	1
<i>Renia sobrialis</i>		1	1		<i>Argyrostroma anilis</i>	1			1
<i>Renia adspergillus</i>		1			<i>Catocala piatrix</i>	1			
<i>Palthis angulalis</i>	1	1	1	1	<i>Catocala habilis</i>	1		1	1
<i>Palthis asopialis</i>	1	1	1	1	<i>Catocala robinsoni</i>	1		1	
<i>Redectis vitrea</i>		1			<i>Catocala flebilis</i>	1		1	1
<i>Oxycilla malaca</i>				1	<i>Catocala angusi</i>			1	1
<i>Hypenodes fractilinea</i>			1		<i>Catocala obscura</i>	1	1	1	1
<i>Dyspyralis puncticosta</i>				1	<i>Catocala resecta</i>	1	1	1	1
<i>Nigetia formosalis</i>		1	1		<i>Catocala residua</i>	1	1	1	1
<i>Bomolocha manalis</i>	1	1	1	1	<i>Catocala insolabilis</i>	1			
<i>Bomolocha baltimoralis</i>	1	1	1	1	<i>Catocala vidua</i>	1	1	1	1
<i>Bomolocha bijugalis</i>	1	1	1	1	<i>Catocala paleogama</i>	1	1	1	1
<i>Bomolocha palparia</i>	1	1	1	1	<i>Catocala subnata</i>	1		1	1
<i>Bomolocha abalienalis</i>				1	<i>Catocala neogama</i>	1	1	1	1
<i>Bomolocha deceptalis</i>				1	<i>Catocala ilia</i>	1	1	1	1
<i>Bomolocha madefactalis</i>	1	1	1	1	<i>Catocala sordida</i>		1		
<i>Bomolocha sordidula</i>				1	<i>Catocala andromedae</i>	1	1	1	1
<i>Plathmpena scabra</i>	1	1	1	1	<i>Catocala coccinata</i>			1	1
<i>Spargaloma sexpunctata</i>		1			<i>Catocala miranda</i>				1
<i>Pangrapta decoralis</i>	1	1	1	1	<i>Catocala ultronia</i>	1	1	1	1
<i>Metalectra discalis</i>		1	1		<i>Catocala grynea</i>		1		
<i>Metalectra quadrisignata</i>				1	<i>Catocala dulciola</i>		1		
<i>Metalectra richardsi</i>		1			<i>Catocala clintoni</i>			1	
<i>Arugisa latiorella</i>		1		1	<i>Catocala similis</i>	1	1		1
<i>Scolecocampa liburna</i>	1	1	1	1	<i>Catocala micronympha</i>	1	1	1	1
<i>Physopropus callitrichoides</i>	1	1	1	1	<i>Catocala amica</i>	1	1	1	1
<i>Hypsoropha monilis</i>		1			<i>Catocala lineella</i>	1		1	
<i>Hypsoropha hormos</i>	1	1	1	1	<i>Pseudoplusia includens</i>	1	1	1	1
<i>Scoliopteryx libatix</i>				1	<i>Allagrapha aerea</i>		1	1	
<i>Calyptra canadensis</i>	1	1	1	1	<i>Polychrisia morigera</i>		1		
<i>Panopoda rufimargo</i>	1	1	1	1	<i>Chrysanympa formosa</i>	1	1	1	1
<i>Panopoda carneicosta</i>	1	1	1	1	<i>Eosphoroptermx thyatiroides</i>	1			1
<i>Cissusa spadix</i>	1		1		<i>Autographa precatationis</i>	1	1	1	
<i>Lesmone detrahens</i>		1			<i>Anagrapha falcifera</i>	1	1	1	1
<i>Zale lunata</i>	1	1	1	1	<i>Marathyssa inficita</i>			1	
<i>Zale galbanata</i>	1		1		<i>Paectes oculatrix</i>	1	1	1	1
<i>Zale aeruginosa</i>		1			<i>Paectes abrostollela</i>	1	1	1	
<i>Zale undularis</i>			1		<i>Paectes abrostoliodes</i>		1		
<i>Zale minerea</i>	1	1	1	1	<i>Eutelia pygmaea</i>		1		
<i>Zale lunifera</i>	1	1	1	1	<i>Eutelia pulcherrima</i>	1			
<i>Zale unilineata</i>	1		1		<i>Baileya ophthalmica</i>	1	1	1	1
<i>Zale horrida</i>	1		1	1	<i>Baileya dormitans</i>	1	1	1	1
<i>Euparthenos nubilis</i>	1	1	1	1	<i>Baileya levitans</i>	1	1	1	
<i>Allotria elonympha</i>	1	1	1	1	<i>Baileya australis</i>		1		
<i>Parallelia bistriaris</i>	1	1	1	1	<i>Characoma nilotica</i>		1		
<i>Euclidia cuspidata</i>	1	1	1		<i>Meganola phylla</i>	1	1	1	1

Appendix 2 cont.

Species <sup>a</sup>	Study area				Species	Study Area			
	AR	BR	WR	YB		AR	BR	WR	YB
<i>Meganola spodia</i>	1	1	1	1	<i>Polygrammate hebraeicum</i>	1	1	1	1
<i>Tripudia quadrifera</i>			1		<i>Eudryas unio</i>		1		1
<i>Oruza albocostaliata</i>	1	1	1	1	<i>Eudryas grata</i>	1	1	1	1
<i>Hyperstrotia pervertens</i>	1		1	1	<i>Apamea cristata</i>				1
<i>Hyperstrotia villificans</i>	1	1	1	1	<i>Apamea vulgaris</i>			1	
<i>Hyperstrotia secta</i>	1	1	1	1	<i>Apamea sordens</i>				1
<i>Thioptera nigrofimbria</i>	1	1	1	1	<i>Agroperina helva</i>			1	
<i>Lithacodia muscosula</i>	1	1	1	1	<i>Crymodes devastator</i>				1
<i>Lithacodia musta</i>	1		1	1	<i>Oligia modica</i>				1
<i>Lithacodia carneola</i>	1	1	1	1	<i>Oligia fractilina</i>	1			1
<i>Cerma cerintha</i>	1	1	1	1	<i>Oligia crytora</i>	1			
<i>Leuconycta diphteroides</i>	1	1	1	1	<i>Oligia semicana</i>			1	
<i>Leuconycta lepidula</i>			1		<i>Oligia obtusa</i>		1		
<i>Tarachidia erastrioides</i>			1	1	<i>Archanara oblonga</i>				1
<i>Panthea furcilla</i>	1		1		<i>Amphipoea velata</i>				1
<i>Colocasia flavicornis</i>	1	1	1	1	<i>Bellura obliqua</i>				1
<i>Colocasia propinquilinea</i>	1	1	1	1	<i>Euplexia benesimilis</i>	1	1	1	1
<i>Charadra deridens</i>	1	1	1	1	<i>Phlogophora periculosa</i>	1	1	1	1
<i>Raphia frater</i>	1	1	1	1	<i>Chytonix palliatricula</i>	1	1	1	1
<i>Acronicta americana</i>	1	1	1	1	<i>Dypterygia rozmani</i>	1		1	
<i>Acronicta dactylina</i>	1		1		<i>Phosphila turbulenta</i>	1	1	1	1
<i>Acronicta lepusculina</i>				1	<i>Phosphila miselioides</i>	1	1	1	1
<i>Acronicta betulae</i>			1		<i>Callopietria mollissima</i>	1	1	1	1
<i>Acronicta radcliffei</i>	1		1		<i>Amphipyra pyramidoides</i>	1	1	1	1
<i>Acronicta funeralis</i>	1	1	1	1	<i>Anorthodes tarda</i>	1	1	1	1
<i>Acronicta vinnula</i>	1	1	1	1	<i>Balsa malana</i>	1		1	
<i>Acronicta superans</i>				1	<i>Balsa tristrigella</i>	1	1	1	1
<i>Acronicta laetifica</i>		1	1		<i>Balsa labecula</i>	1	1	1	1
<i>Acronicta hasta</i>	1	1	1	1	<i>Elaphria versicolor</i>	1	1	1	1
<i>Acronicta spinigera</i>	1	1	1	1	<i>Elaphria festivooides</i>	1	1	1	1
<i>Acronicta morula</i>	1	1	1	1	<i>Elaphria grata</i>		1		1
<i>Acronicta interrupta</i>		1	1		<i>Galgula partita</i>	1	1	1	1
<i>Acronicta lobeliae</i>	1	1	1	1	<i>Perigea xanthioides</i>		1	1	
<i>Acronicta heitzmani</i>			1	1	<i>Platysenta videns</i>	1	1	1	1
<i>Acronicta exilis</i>			1		<i>Ogdoconta cinereola</i>			1	
<i>Acronicta ovata</i>	1	1	1	1	<i>Stiroides obtusa</i>				1
<i>Acronicta modica</i>	1	1	1	1	<i>Plagiomimicus pityochromus</i>			1	
<i>Acronicta haesitata</i>	1	1	1	1	<i>Basilodes pepita</i>	1		1	1
<i>Acronicta tristis</i>	1		1		<i>Cosmia calami</i>	1	1	1	1
<i>Acronicta hamamelis</i>	1				<i>Amolita fessa</i>		1		
<i>Acronicta increta</i>	1	1	1	1	<i>Lithophane petulca</i>	1		1	
<i>Acronicta inclarata</i>		1			<i>Lithophane hemina</i>	1		1	
<i>Acronicta retardata</i>	1	1	1	1	<i>Lithophane antennata</i>	1		1	
<i>Acronicta afflictata</i>	1	1	1	1	<i>Pyreferra hesperidago</i>			1	
<i>Acronicta impleta</i>	1	1	1	1	<i>Sideridis congermana</i>			1	
<i>Acronicta lithospila</i>	1	1	1	1	<i>Polia detracta</i>	1	1	1	
<i>Acronicta oblinita</i>		1			<i>Polia goodelli</i>	1			1
<i>Agriopodes fallax</i>	1	1	1	1	<i>Polia latex</i>	1	1	1	1
<i>Agriopodes teratophora</i>		1	1	1	<i>Melanchra adjuncta</i>	1	1	1	1

Appendix 2 cont.

Species <sup>a</sup>	Study area			
	AR	BR	WR	YB
<i>Hadena ectypa</i>			1	
<i>Lacinipolia renigera</i>	1	1	1	1
<i>Lacinipolia lorea</i>		1	1	1
<i>Lacinipolia implicata</i>	1	1	1	1
<i>Pseudaletia unipuncta</i>	1	1	1	
<i>Leucania linda</i>	1	1	1	1
<i>Leucania ursula</i>	1	1	1	1
<i>Leucania pseudargyria</i>	1	1	1	1
<i>Orthosia rubescens</i>	1			1
<i>Crocigrapha normani</i>	1		1	1
<i>Egira alternans</i>				1
<i>Achatia distincta</i>	1		1	1
<i>Morrisonia confusa</i>	1		1	
<i>Nephelodes minians</i>	1		1	1
<i>Homorthodes furfurata</i>	1	1	1	1
<i>Ulolonche culea</i>	1	1	1	1
<i>Pseudorthodes vecors</i>	1		1	
<i>Orthodes crenulata</i>	1	1	1	
<i>Orthodes cynica</i>	1	1	1	1
<i>Tricholita signata</i>			1	

Species <sup>a</sup>	Study area			
	AR	BR	WR	YB
<i>Agrotis ipsilon</i>	1	1	1	1
<i>Feltia jaculifera</i>	1	1	1	
<i>Ochropleura plecta</i>	1	1	1	1
<i>Trichosilia geniculata</i>		1		
<i>Euagrotis illapsa</i>	1			
<i>Peridroma saucia</i>	1	1	1	
<i>Xestia adela</i>	1		1	
<i>Xestia dolosa</i>	1	1	1	1
<i>Xestia normaniana</i>			1	
<i>Xestia smithii</i>	1		1	1
<i>Xestia bicarnea</i>	1		1	
<i>Protolampra brunneicollis</i>		1	1	
<i>Abagrotis alternata</i>		1	1	
<i>Helicoverpa zea</i>	1	1		1
<i>Schinia arcigera</i>	1		1	1

<sup>a</sup> - Specific determinations by E. H. Metzler, M. Gilligan, T. Gilligan, D. J. Horn, J. W. Peacock, R. D. Watkins and R. Zebold

Species sequence is standard from Hodges et al. (1983)