

“ความหลากหลายทางชีวภาพของจุลินทรีย์ในประเทศไทย”

# Diversity of microbes in the BIOTEC Culture Collection

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NAC 2013

# Thailand

## Birds

World = 9,934  
Thailand = 982

## Plants

World = 250,000  
Thailand = 12,000

## Fungi

World = 80,000  
(Expected No. = 1,500,000)  
Thailand = 6,000  
(Expected No. = 150,000)

## Mammals

World = 4,260  
Thailand = 282

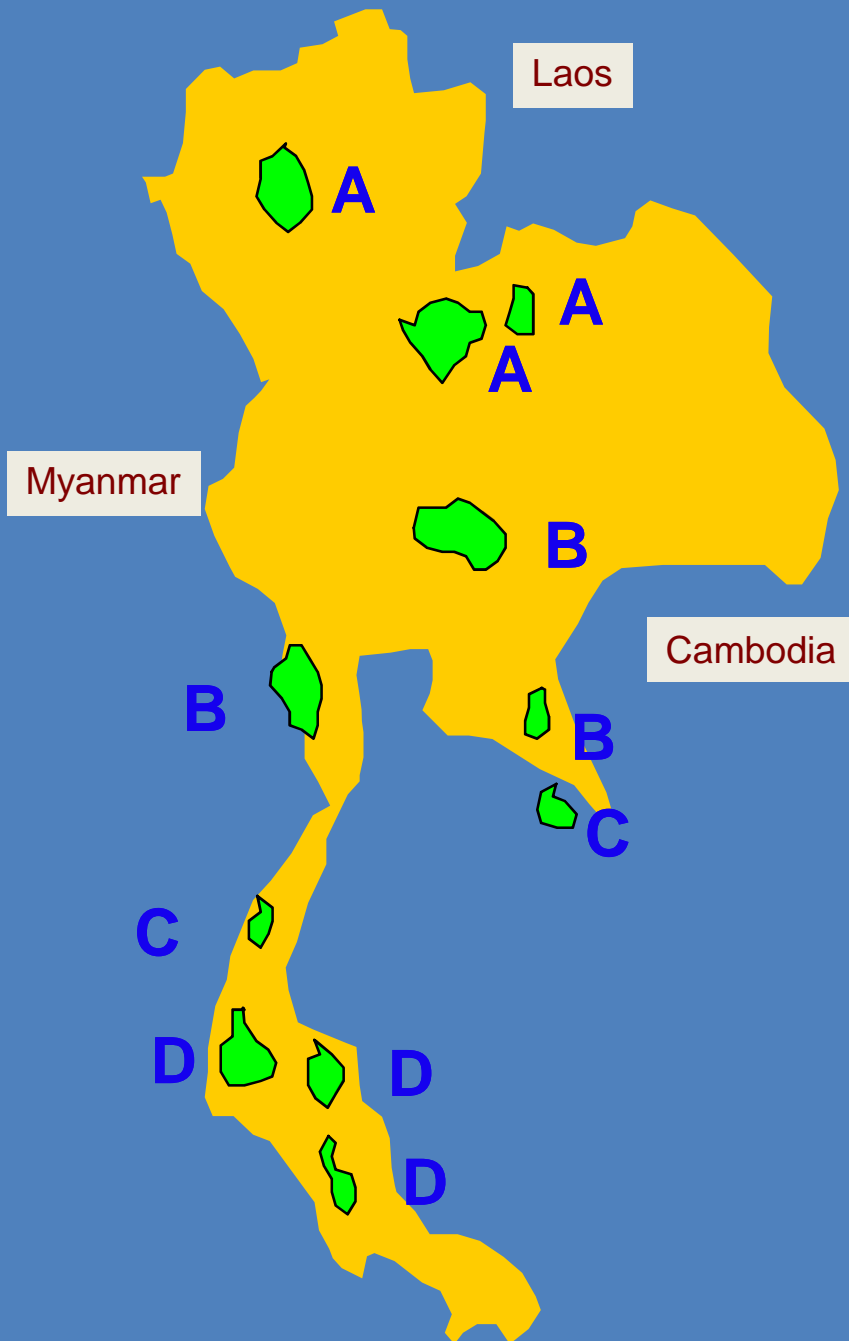
## Other microbes

World = 84,000  
(Expected No. = 1,600,000)  
Thailand = 300  
(Expected No. = 100,000)



Species richness of Thailand is *ca.* 7-10 % of the world's described species

# FOREST TYPES

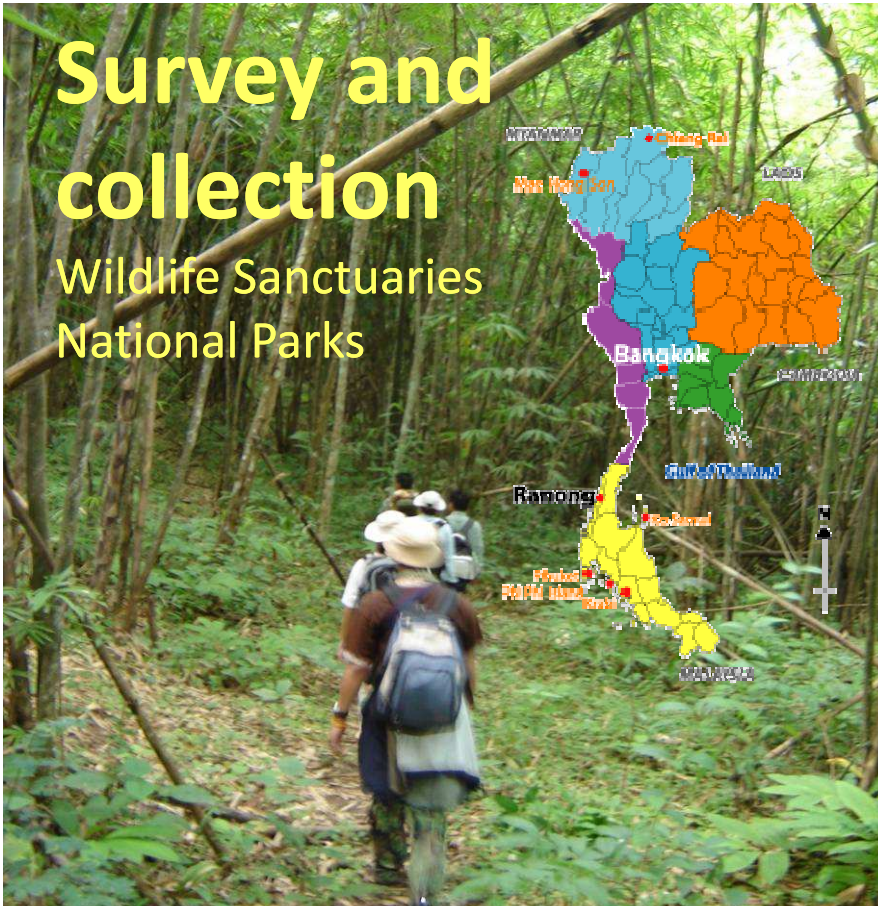


**A. Cool deciduous teak and pine forest**

**B. Evergreen monsoon forest**

**C. Coastal evergreen rain forest**

**D. Hill evergreen rain forest**

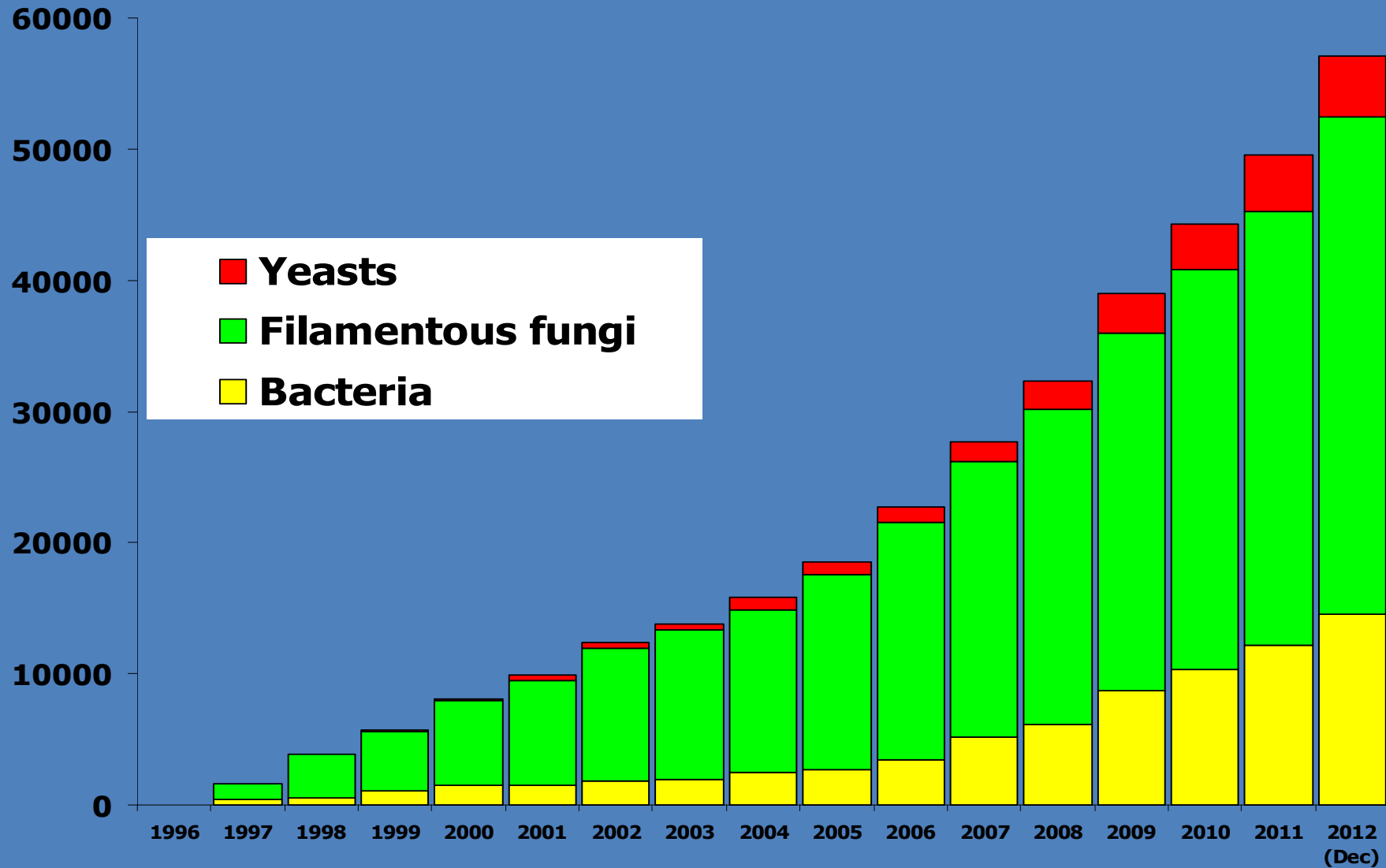


BIOTEC

Thailand Network on Culture Collections (TNCC) was established since 2000. The network consists of four major collections specialized in different groups of microorganisms:

- 1) **BIOTEC Culture Collection (BCC)** holds more than **57,000** strains (1,896 species of 984 genera) of filamentous fungi, yeasts and bacteria isolated from natural environments in Thailand.
- 2) **DMST Culture Collection (DMST)** has in the collection **32,000** strains of filamentous fungi, yeasts, bacteria and cell lines related to human disease control and prevention.
- 3) **DOA Culture Collection (DOA)** houses more than 6,000 strains of agriculturally important algae, bacteria and filamentous fungi.
- 4) **TISTR Culture Collection (TISTR)** collects more than **5,500** strains of agriculturally and industrially useful algae, bacteria, filamentous fungi and yeasts.

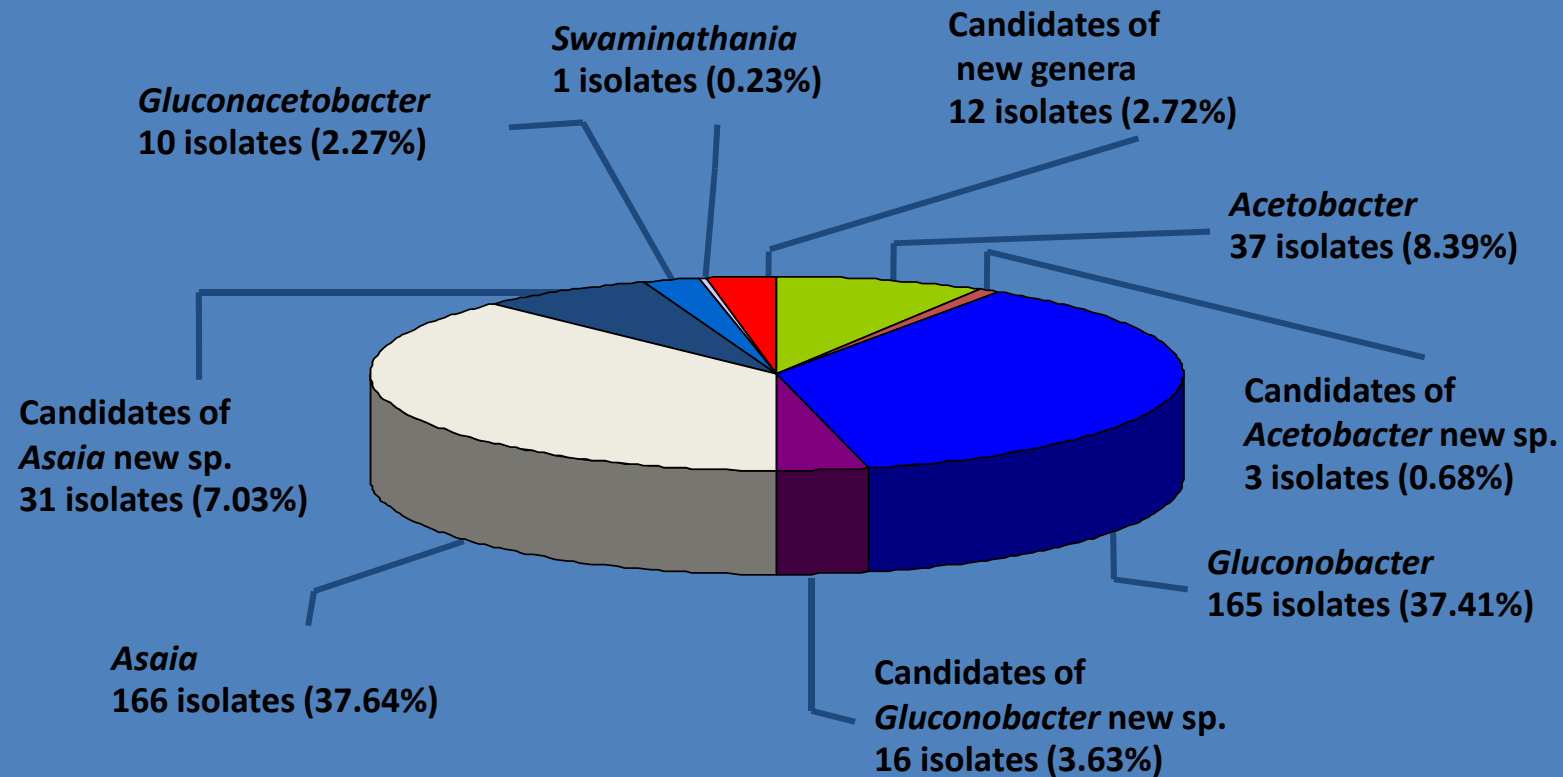
# The number of microorganisms at BCC (57,079 samples)



BCC holdings

Group	No. strains	No. genera	No. species
Bacteria	14,561	155	424
Filamentous fungi	38,012	767	1,158
Yeasts	4,621	62	314
Total	57,194	984	1,896

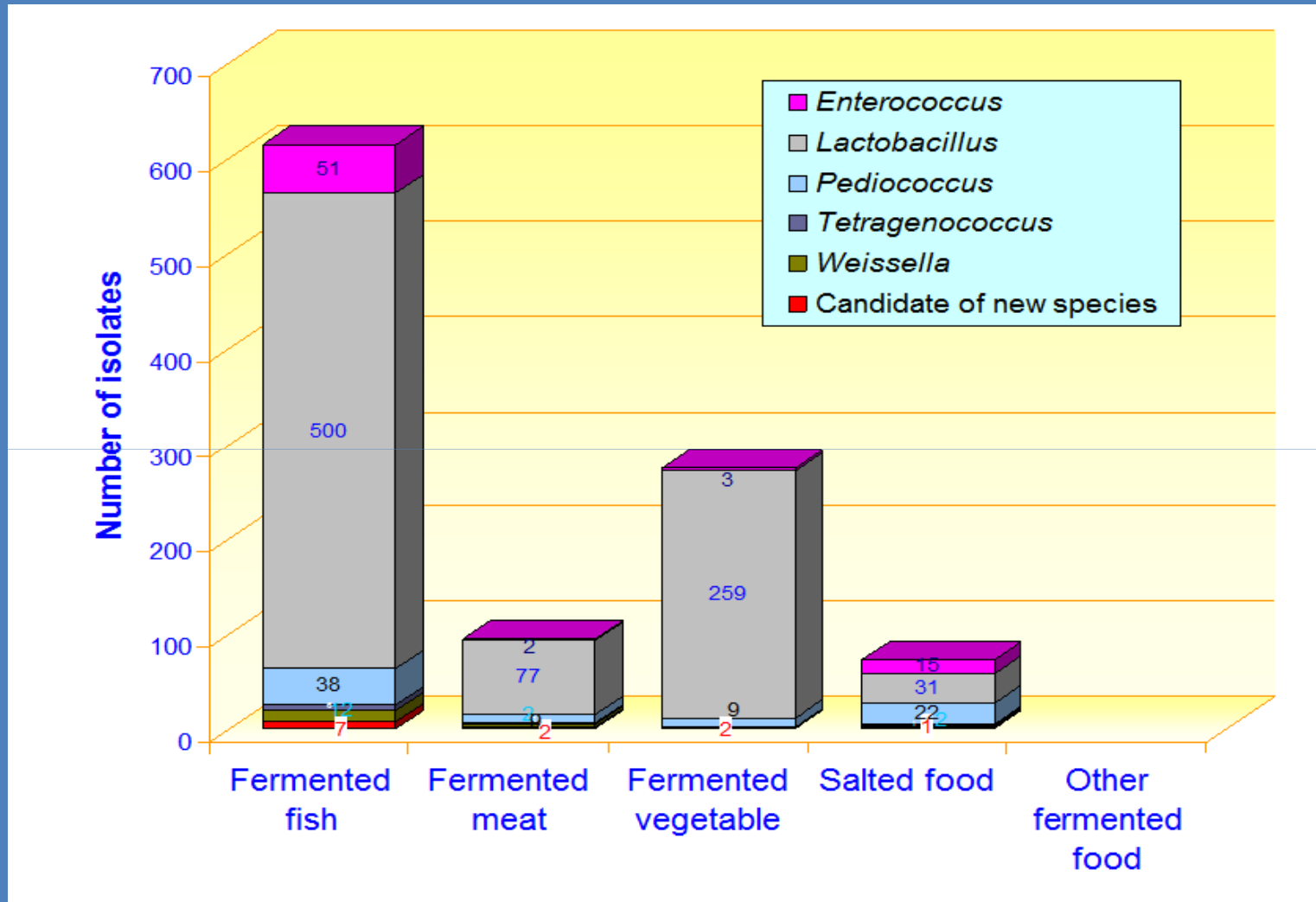
# Taxonomic study of acetic acid bacteria in Thailand



441 strains of acetic acid bacteria from natural sources collected in Thailand

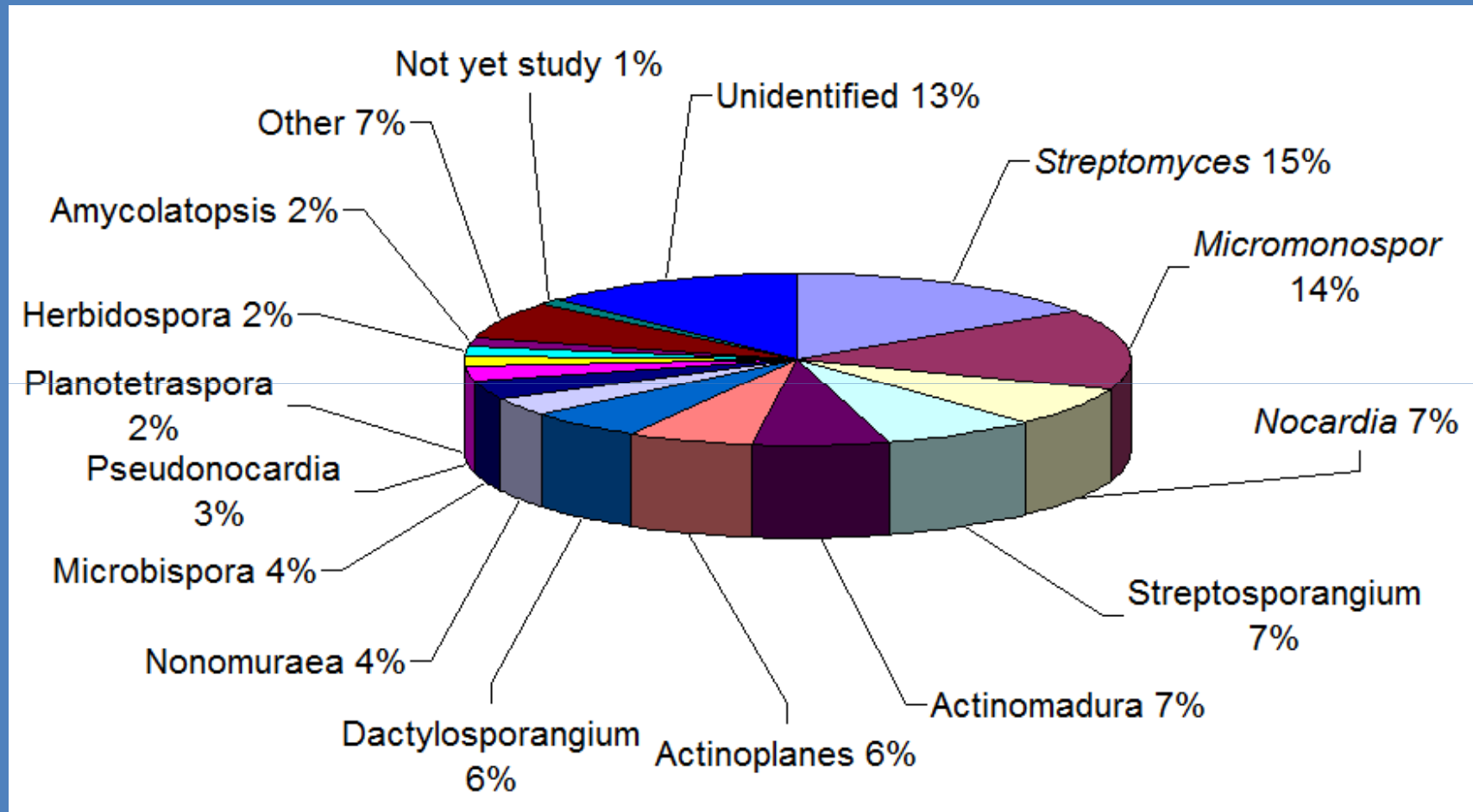


## Taxonomic and ecological study of lactic acid bacteria from fermented foods in Thailand



1,053 strains of lactic acid bacteria from fermented food isolated since 2008

## Filamentous actinomycetes isolated from Thailand for bioactive compound screening



7,984 strains of 43 genera isolated since 2005

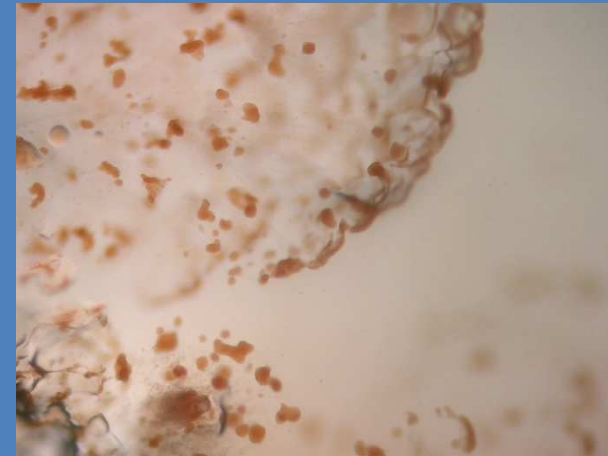
# Myxobacteria isolated from Thailand for bioactive compound screening



*Stigmatella*



*Condromyces*



*Nannocystis*



*Myxococcus*



*Corallococcus*



*Angiococcus*



*Cystobacter*

**105 strains of 7 genera isolated since 2008**

# Fungi



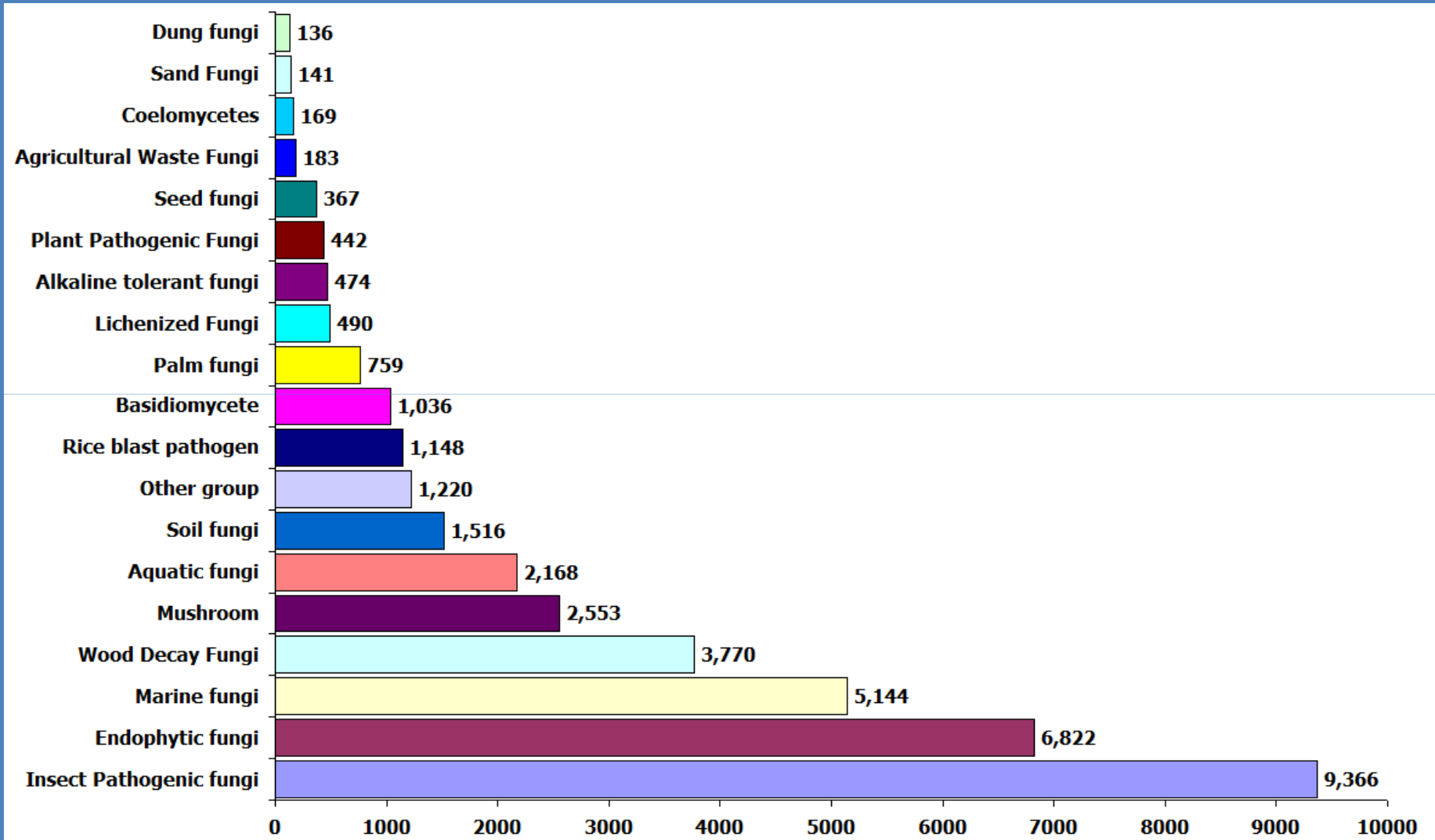


In 1989, BIOTEC started a research on insect-pathogenic fungi. This became a BIOTEC program in 1993 which now concentrates on several taxonomic and ecological fungal groups:

- **Alkalinophilic fungi**
- **Basidiomycetes**
- **Coelomycetes**
- **Endophytes**
- **Dothideomycetes**
- **Dung fungi**
- **Freshwater and Aquatic fungi**
- **Invertebrate-pathogenic fungi**
- **Lichens**
- **Marine fungi**
- **Palm fungi**
- **Plant pathogenic fungi**
- **Seed decay and leaf litter fungi**
- **Thermophilic fungi**
- **Yeasts**

A total of 500-700 species of fungi had been recorded by the late 1980's while this figure now stands at around 6,000 species (Jones *et al.* 2004).

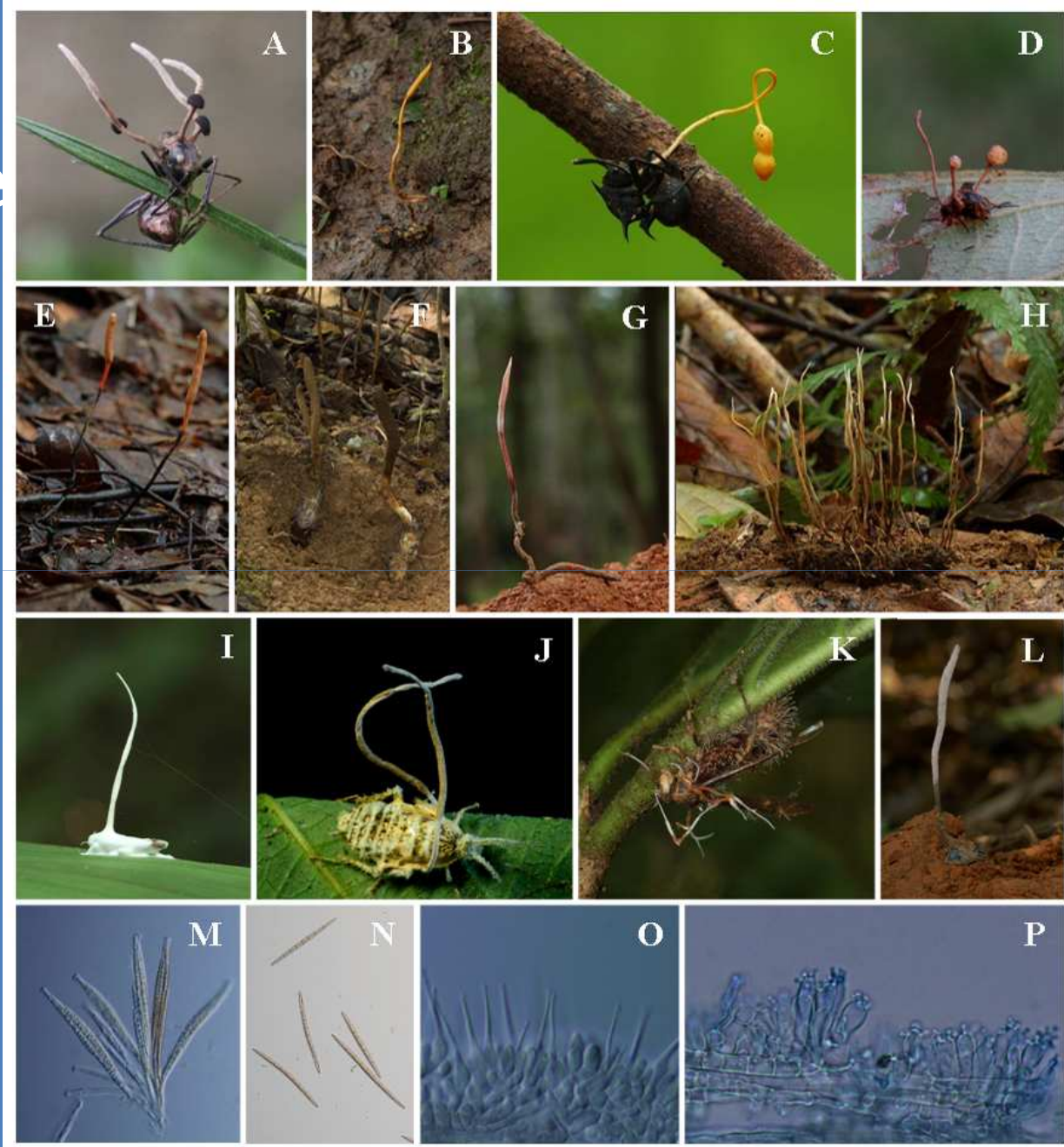
## Group of Fungi in BIOTEC Culture Collection (38,012 Samples)



# Cordycipitaceae

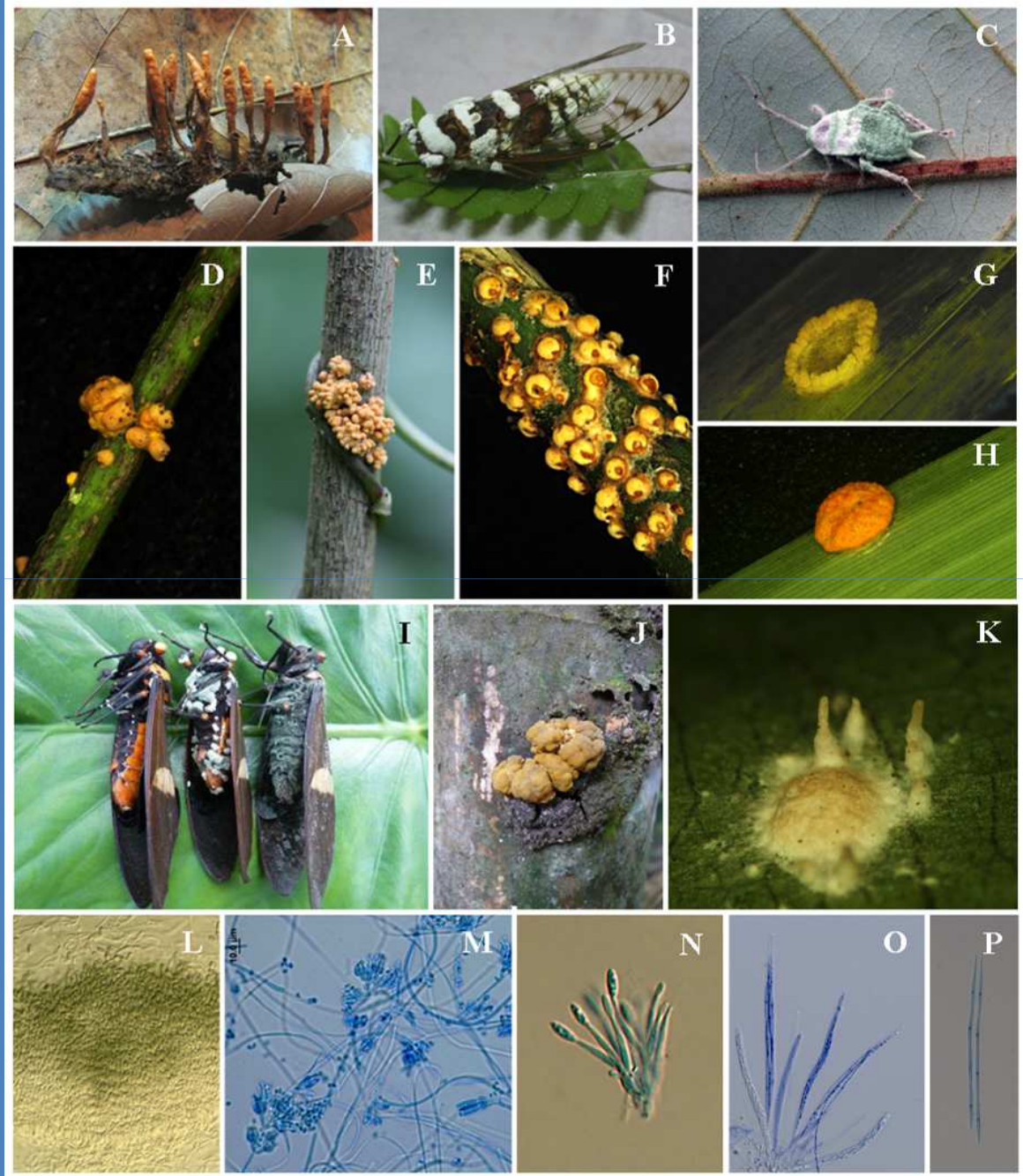


# Ophio- cordycipitaceae



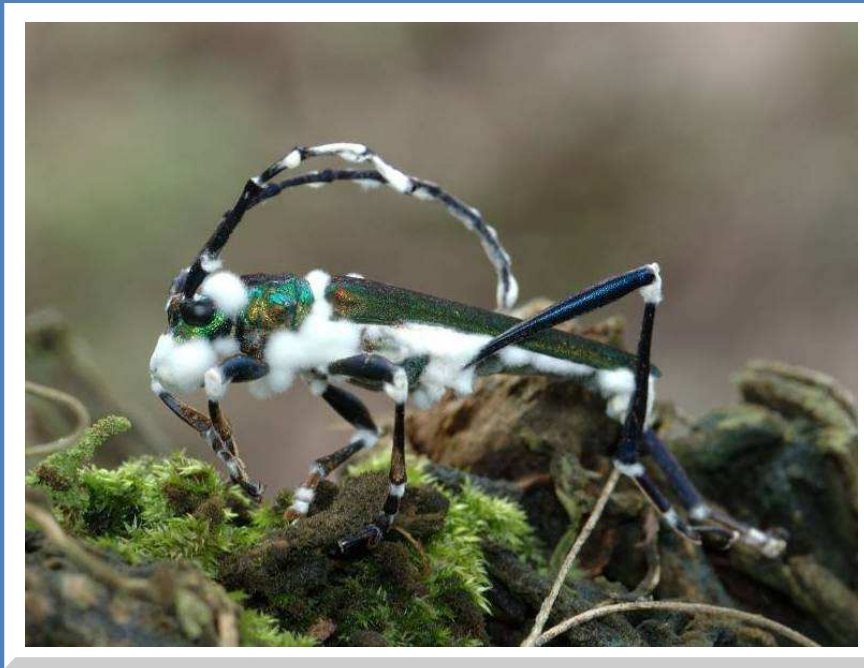


# Clavicipitaceae



# Host specificity

- General opportunistic pathogens
- Fastidious pathogens



# Habitats



# Habitats



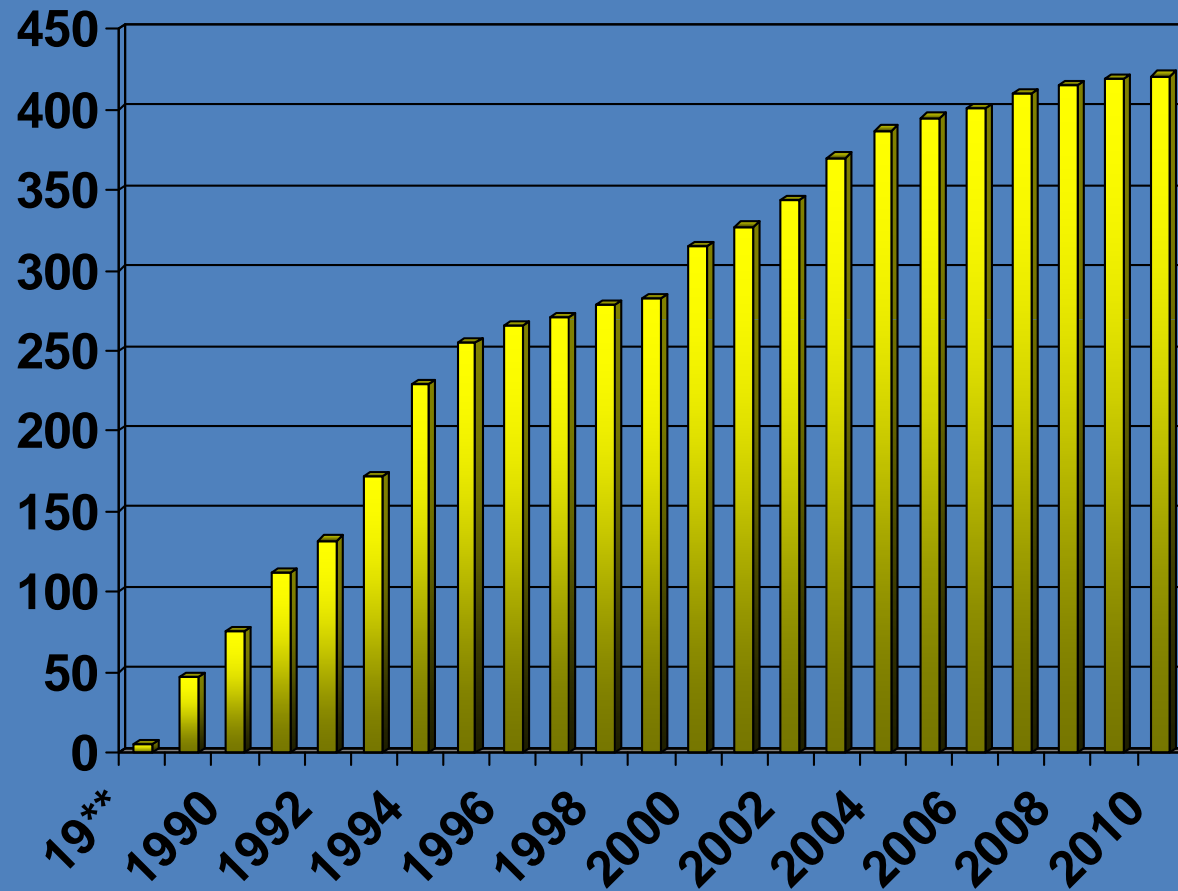
# Habitats



# Habitats



# Numbers of Insect Fungi in Thailand



■ Insect fungi

150 species can  
be put to culture

# Bioactive Substances from Insect Pathogenic Fungi

MASAHIKO ISAKA, PRASAT KITTA KOOP, KANYAWIM KIRTIKARA, NIGEL L. HYWEL-JONES, AND YODHATHAI THEBTARANONTH\*

National Center for Genetic Engineering and Biotechnology (BIOTEC), Thailand Science Park, Klong Luang, Pathumthani 12120, Thailand

Received March 7, 2005

## ABSTRACT

Insect pathogenic fungi have opened up a relatively untapped area of natural product research which, unfortunately, has not received much attention to date. Found in wild abundance in wet tropical Thailand, the insect fungi are shown to contribute not only as controllers of insect populations but also as rich sources of structurally novel biologically active substances.

- Phomalactone – common in various family and genera
- Oosporein - common in various family and genera
- Beauvericin - 7 strains, Coleopterans and lepidopterans
- Cordyropolone – only from *Cordyceps*

Bioactive Substances from Insect Pathogenic Fungi Isaka et al.



FIGURE 3. Excavation revealing the larval host of *Cordyceps sinensis*.



FIGURE 4. *Cordyceps sinensis* stroma on Lepidoptera larvae.

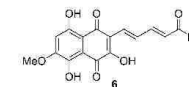
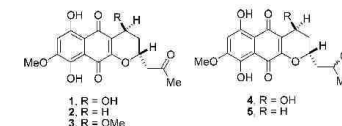


FIGURE 5. *Cordyceps unilateralis* on an ant.



FIGURE 6. *Cordyceps nipponica* on ant lions.

deep red color under acidic conditions but intense purple in basic environments; such color characteristics are attractive to the pigment industry. Production of naphthoquinones by *C. unilateralis*, after optimization of fermentation conditions, can attain yields up to 3 g/L of culture broth.<sup>28</sup>



*Cordyceps nipponica* was originally described from cicadas in Japan and is found infecting both cicadas and ant lions (*Neuroptera*) in Thailand. Two *N*-hydroxy-2-pyridones, cordypridones A (7) and B (8), and two tricyclic *N*-methoxy-2-pyridones, cordypridones C (9) and D (10), were isolated from *Cordyceps nipponica* BCC 1389 (collected from Khao Yai National Park, central Thailand, Figure 6).<sup>29</sup> Cordypridone A (7) is identical to 8-methyl-pyridoxatin, previously isolated from an unidentified fungus OS-F61800,<sup>30</sup> while its atropisomer, cordypridone B (8), was shown to be a metabolite of BCC 1389. A careful study indicated that interconversion between compounds 7 and 8 occurred upon heating the solution, and the absolute configuration of cordypridone 7 (and hence its atropisomer, 8) was later determined using chemical means. Epoxidation of compound 11 (1-*O*-methyl derivative of 7) and subsequent cyclization gave the major product 12, which is the 14-hydroxy derivative of cordypridone C (9). X-ray analysis of 13, the *p*-bromobenzolate derivative of 12, revealed the proposed absolute configuration. Cordypridones A (7) and B (8) exhibited potent antimalarial activity with respective IC<sub>50</sub>

derivatives, 1–6. Interestingly, these naphthoquinones exhibited antimalarial activity with IC<sub>50</sub> values of 2.5–10.1 μg/mL (Table 1). The above naphthoquinones show a



# Conoideocrellide A

JOURNAL OF  
**NATURAL  
PRODUCTS**

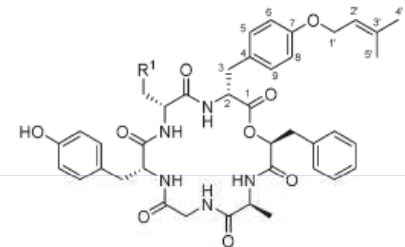
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pubs.acs.org/jnp

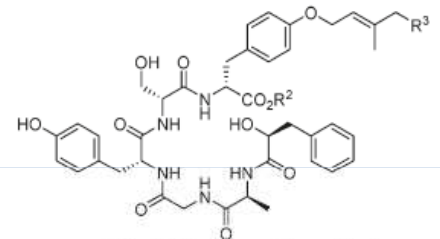
## Bioactive Compounds from the Scale Insect Pathogenic Fungus *Conoideocrella tenuis* BCC 18627

Masahiko Isaka,\* Somporn Palasarn, Sumalee Supothina, Somjit Komwijit, and J. Jennifer Luangsa-ard

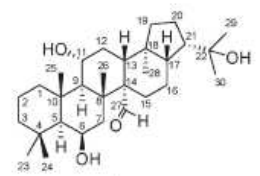
National Center for Genetic Engineering and Biotechnology (BIOTEC), 113 Thailand Science Park, Phaholyothin Road, Klong Luang, Pathumthani 12120, Thailand



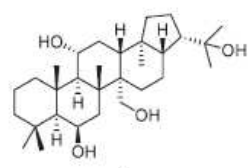
1 R<sup>1</sup> = OH : conoideocrellide A  
16 R<sup>1</sup> = H : gliotide/paecilodepsipeptide A



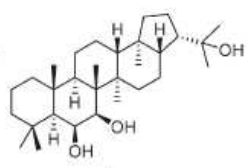
2 R<sup>2</sup> = H, R<sup>3</sup> = H : conoideocrellide B  
3 R<sup>2</sup> = CH<sub>3</sub>, R<sup>3</sup> = H : conoideocrellide C  
4 R<sup>2</sup> = CH<sub>3</sub>, R<sup>3</sup> = OH : conoideocrellide D



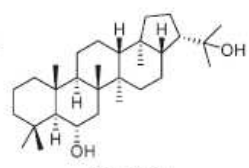
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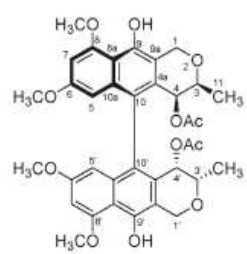
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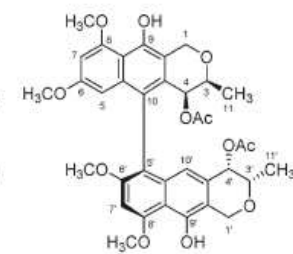
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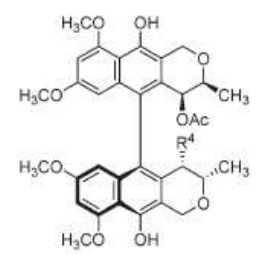
8 : zeorin



9



10



11 R<sup>4</sup> = H : ES-242-1  
12 R<sup>4</sup> = OAc : ES-242-2

# NRPS screening

## Cordycipitaceae:

*Beauveria bassiana*  
*Cordyceps ninchukispora*  
*Isaria tenuipes*  
*Gibellula* sp.



## Clavicipitaceae:

*Aschersonia samoensis*  
*Metarhizium flavoviride*  
*Hypocrella discoidea*



# Marine Fungi

- Marine fungi are an ecological group that are able to grow and sporulate in marine habitats (Kohlmeyer & Kohlmeyer, 1979)
- Marine fungi grow on a variety of substrata.
- Marine fungi are mostly saprobes, and produce a wide range of enzymes. They play a major role in decaying woody materials in the marine habitats.



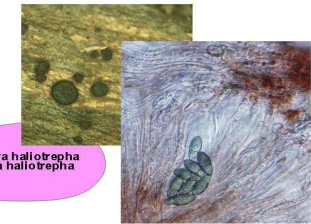
# Marine Fungal Lineages in Kingdom Fungi

## Basidiomycota



## Eurotiomycetes

## Leotiomycetes

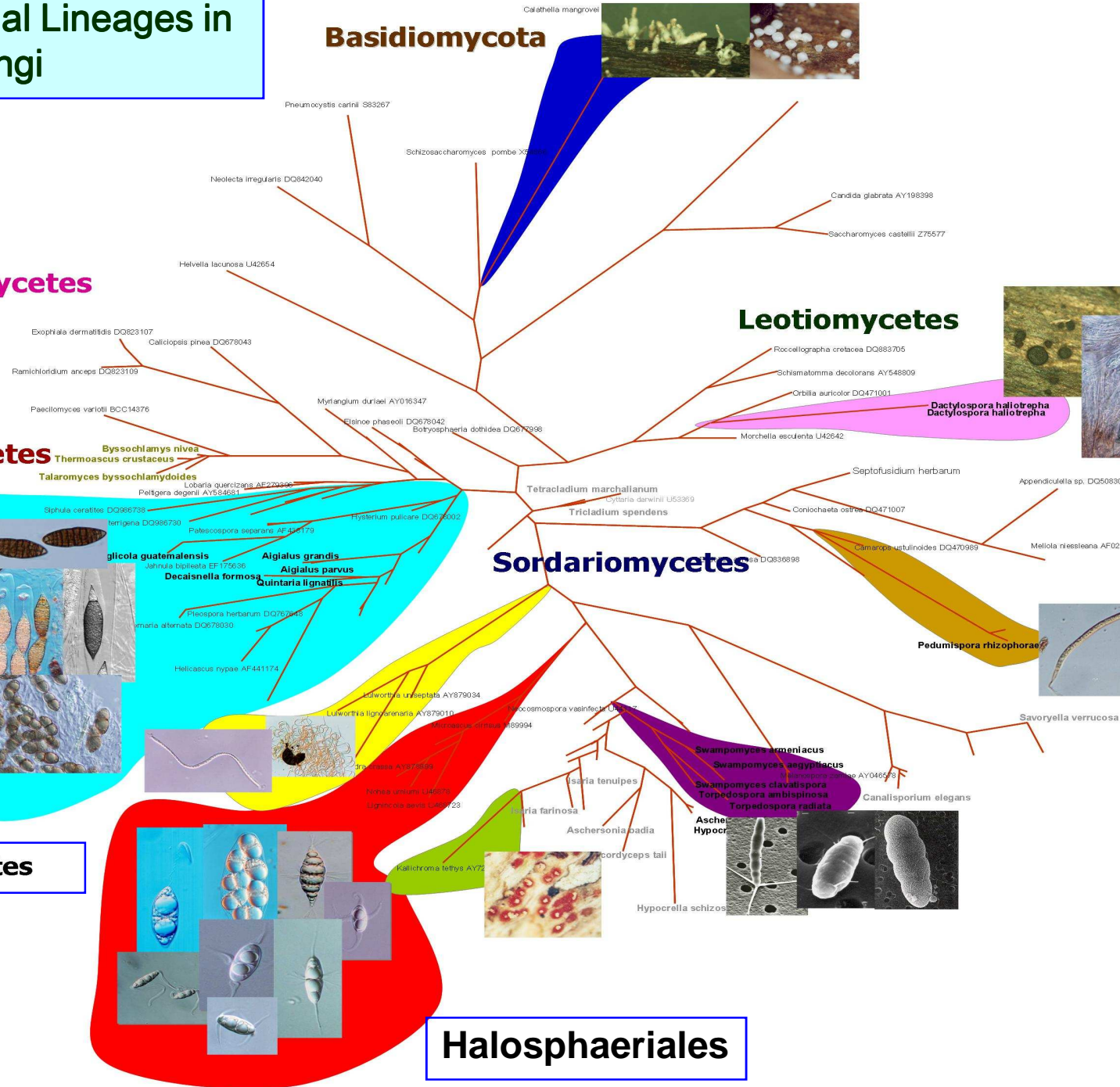


## Lecanoromycetes

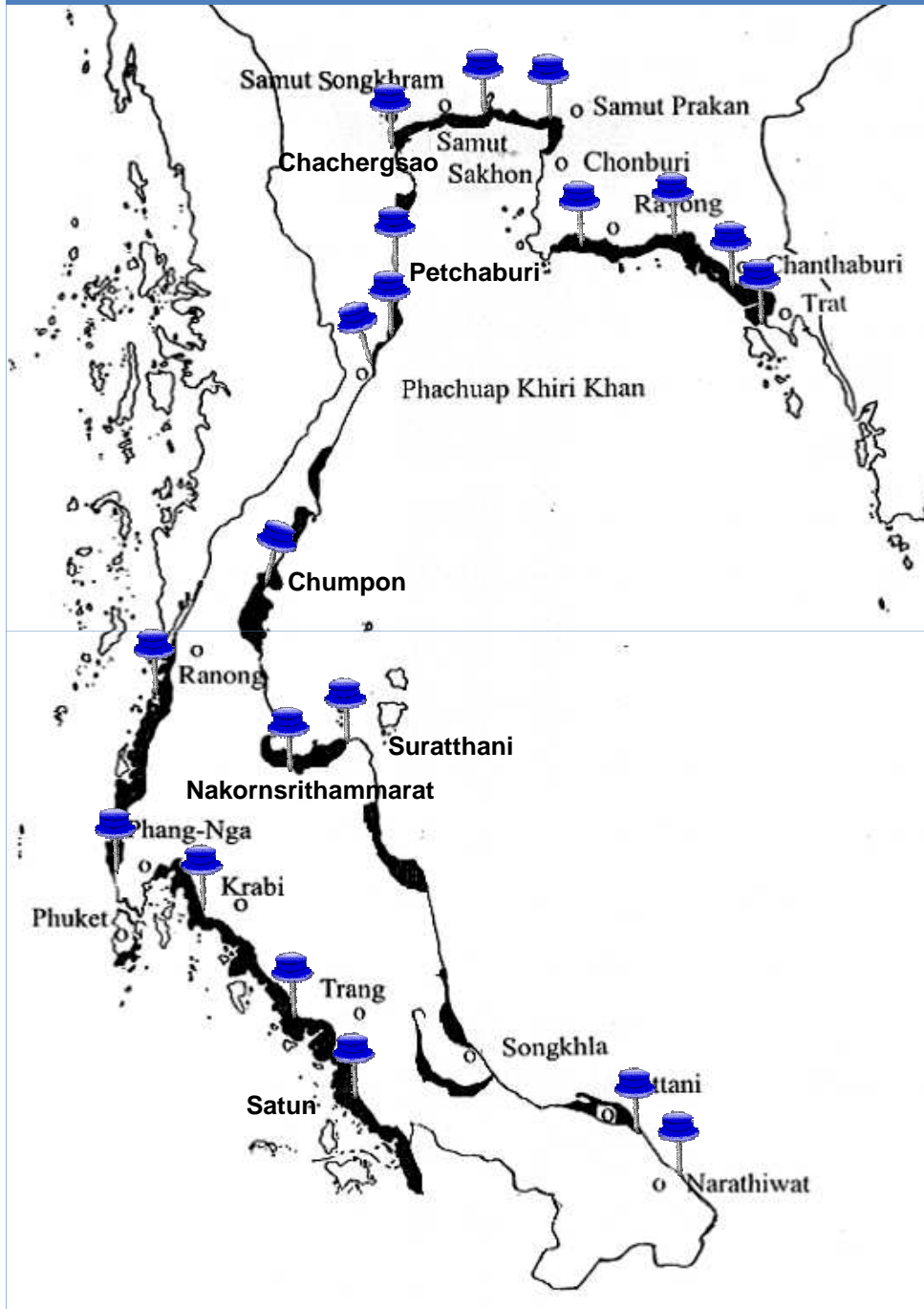
## Sordariomycetes

## Dothideomycetes

## Halosphaeriales



# Thai marine fungal diversity



- Studies of marine fungi in Thailand have been restricted to sandy beaches, sea shores, mangroves and estuarine *Nypa* palm forests along Thai Gulf and Andaman sea.

- Marine and mangrove fungi documented for Thailand: total 163 (Jones *et al.* 2006, Sakayaroj *et al.* 2011).

- A comprehensive classification of the filamentous marine fungi worldwide has outlined 530 sp. (from 321 genera) (Jones *et al.* 2009)

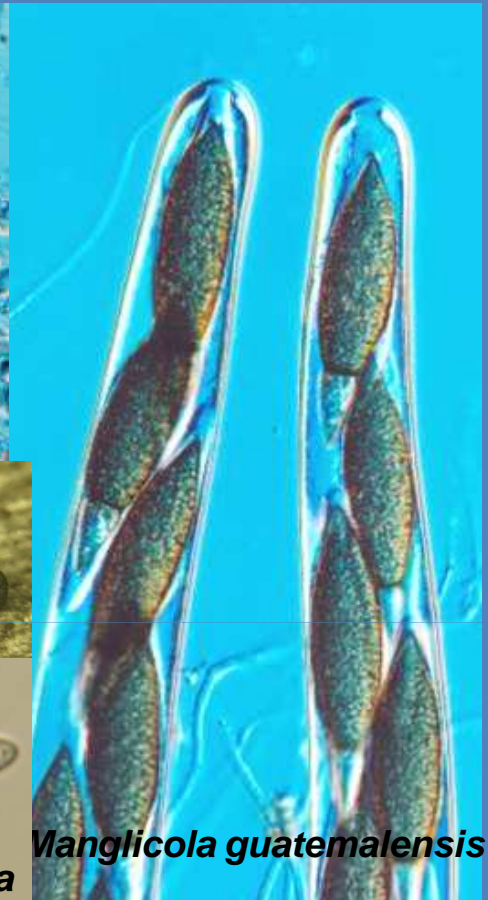
# NEW RECORDS for Thailand



*Lindra thalassiae*



*Swampomyces cf aegyptiacus*



*Manglicola guatemalensis*



*Lautospora simillima*



*Mauritiana rhizophorae*



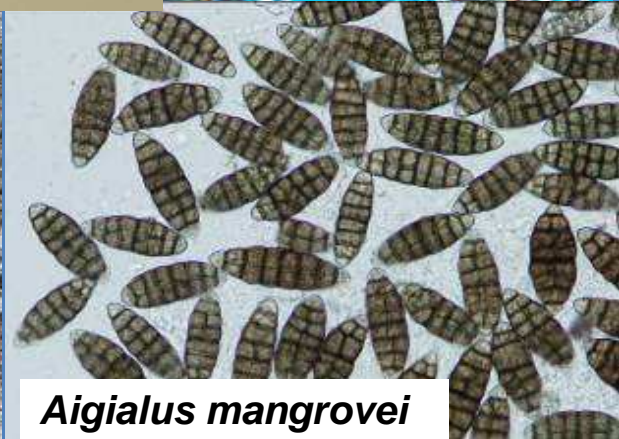
*Patellaria atrata*



*Carinispora nypae*



*Aigialus rhizophorae*



*Aigialus mangrovei*

# Marine fungi as source of bioactive metabolites



- Marine fungi are of particular interest to the pharmaceutical industry
- The most successful marine fungi in producing new compounds are those isolated from mangrove habitats.
- They grow under extreme/stressful conditions and thus may produce a wider range of bioactive compounds.

## *Aigialus parvus*

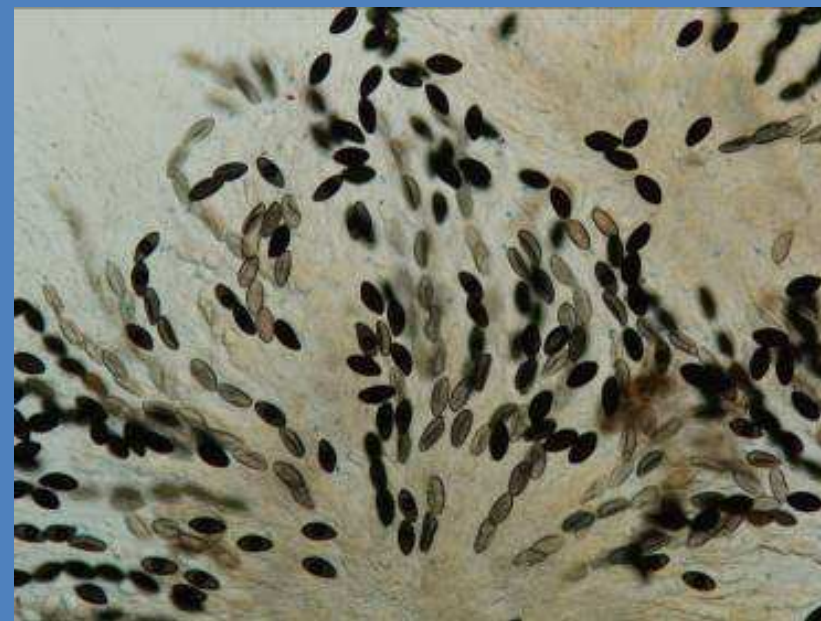
BCC5311



**Aigialomycins A and Hypothemycin showed significant antimalarial and antitumor properties**

Isaka et al., 2002, J.Org. Chem. 67:1561-1566.  
Isaka et al 2009, Tetrahedron 65 (2009) 4396–4403.  
Vongvilai et al., 2009. J. Nat. Prod. 67: 457-460  
Chinworrungsee, M., et. al. J. Chem. Soc.,  
Perkin Trans. 2002; 2004

## *Halorosellinia oceanica* BCC 5149



- **Produced halorosellins A and B,**  
4,8-dihydroxy- 6-methoxy-4,5-dimethyl-3-  
methyleneisochroman-1-one,  
3-acetyl-7-hydroxy-5-methoxy-3,4-dimethyl-  
3Hisobenzofuran-1-one and  
17-dehydroxyhalorosellinic acid

- **Exhibited anti mycobacterial activity**



# Thraustochytrids: A potential PUFA source

- Thraustochytrids are a group of nonphotosynthetic, heterotrophic marine organisms classified under the Labyrinthulomycetes, Kingdom Straminipila.
- Recently, attention has been given to this group of organisms because of their high lipid contents, particularly Poly Unsaturated Fatty Acids (PUFAs).
- With the continuously decreasing resources for traditional fish oils as PUFA sources for aquaculture, thraustochytrids offer a promising alternative and stable source for heterotrophic fatty acid production.

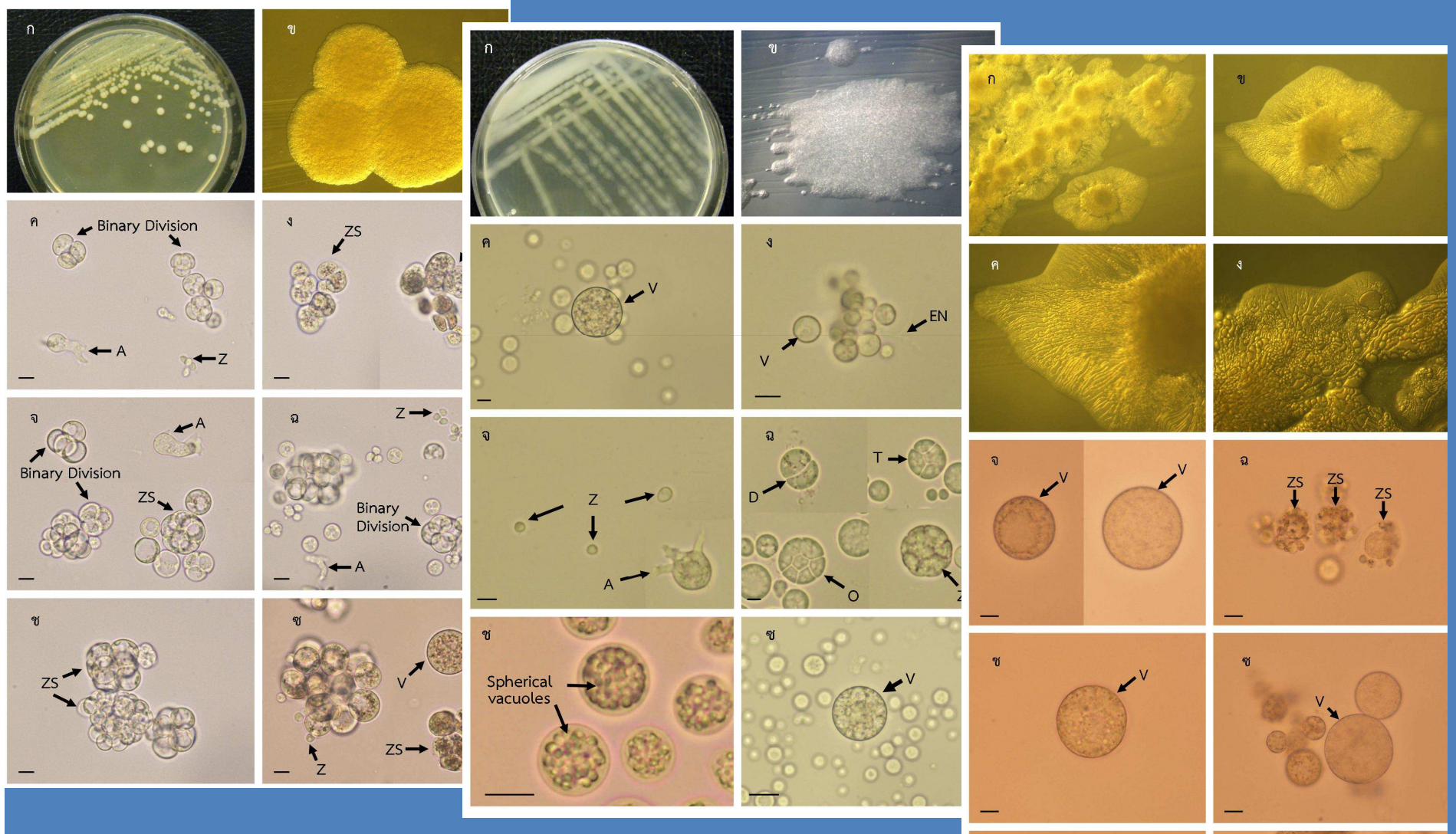


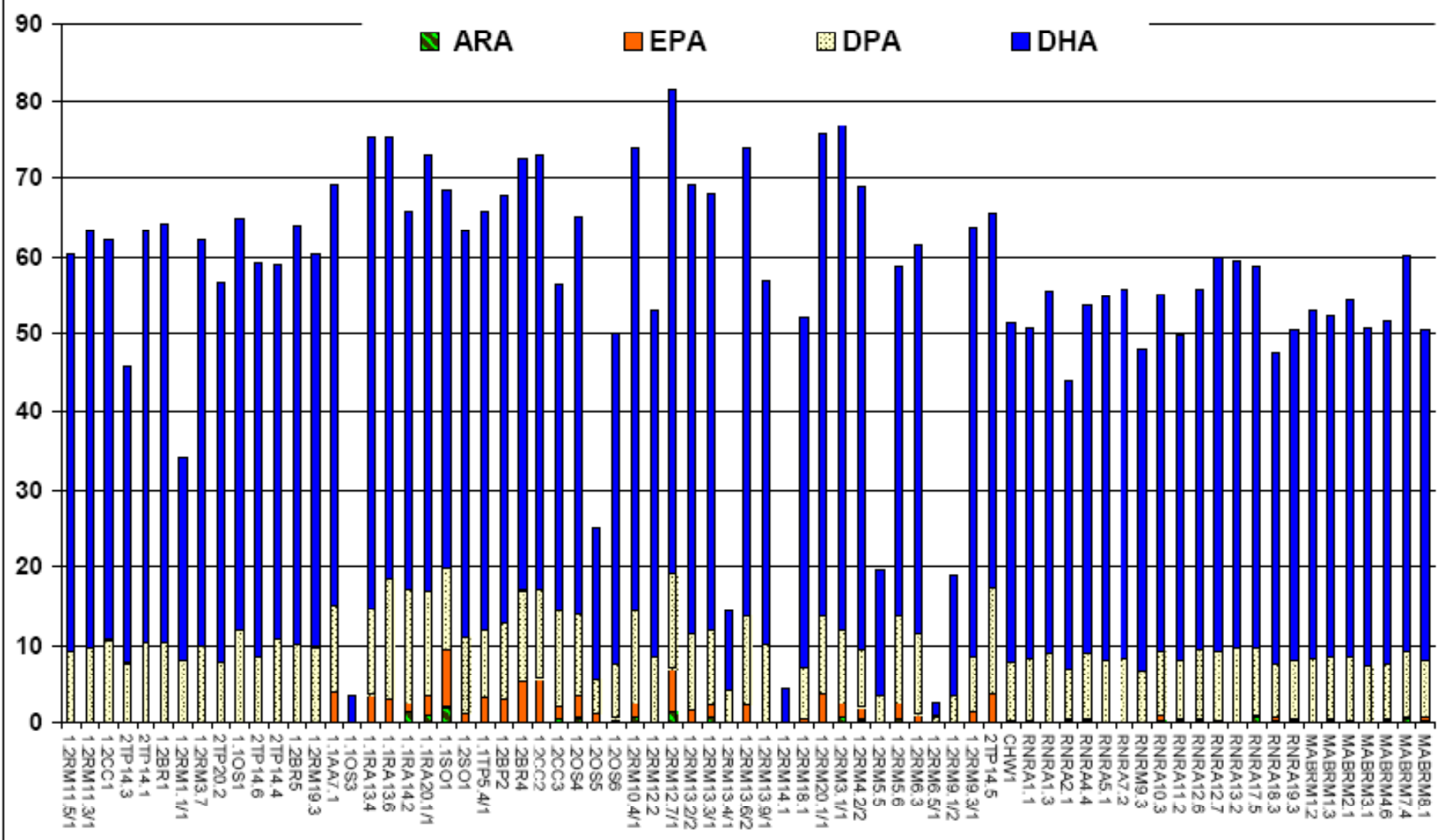
Marine and estuarine habitats: seawater, marine sediments, littoral algae, seaweeds, invertebrates, fallen mangrove leaves



# Colony appearance, dividing cells, zoosporangium, zoospores and mature cells of thraustochytrids.

(The most common genera are *Aurantiochytrium* and *Schizochytrium*.)





# Marine Dothideomycetes



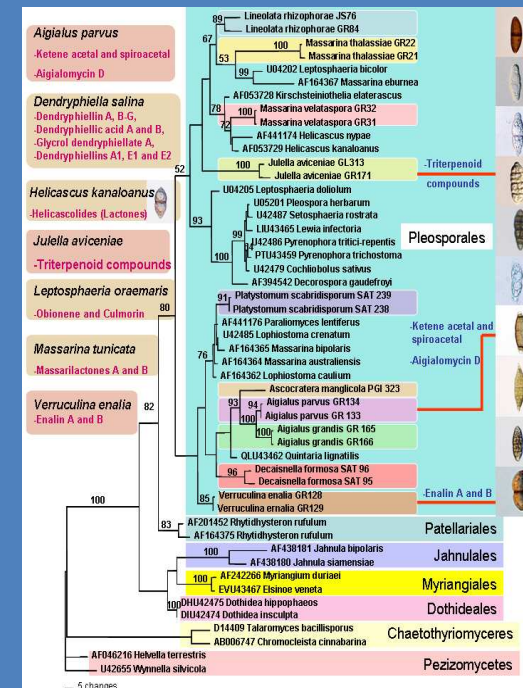
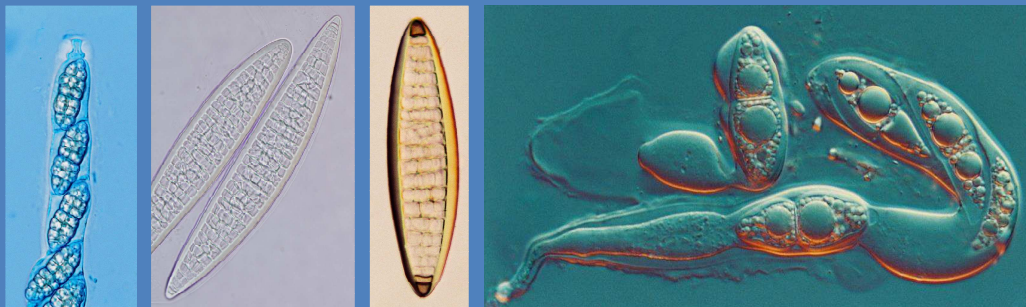
*Nypa fruticans*



Mangrove wood



Rhizomes of Sea grasses



**Classification of marine Ascomycota, anamorphic taxa and Basidiomycota**

Jones, E.B.G.<sup>1\*</sup>, Sakayaroj, J.<sup>1</sup>, Suetrong, S.<sup>1,3</sup>, Somrithipol, S.<sup>1</sup> and Pang, K.L.<sup>2</sup>

<sup>1</sup>Bioresources Technology Unit, Phylogenetics Laboratory, National Center for Genetic Engineering and Biotechnology, 113 Paholyothin Road, Khlong 1, Khlong Luang, Pathum Thani 12120, Thailand

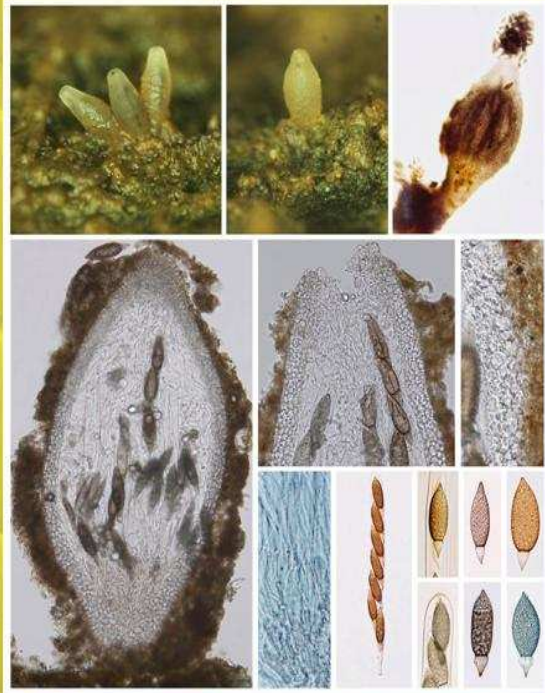
<sup>2</sup>Institute of Marine Biology, National Taiwan Ocean University, No. 2 Pei-Ning Road, Keelung 20224, Taiwan

<sup>3</sup>Department of Microbiology, Faculty of Science, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand.

# Fungal Diversity

Volume 35, March 2009

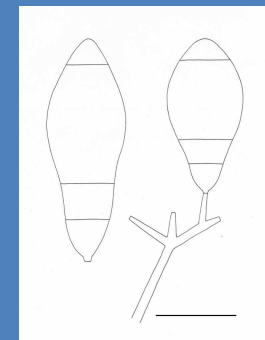
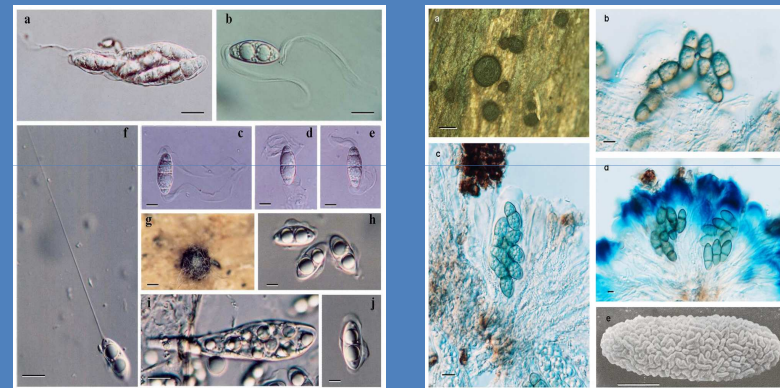
An International Journal of Mycology





Kunming University of Science and Technology



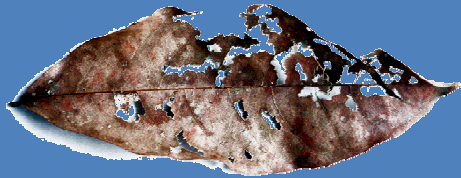
Web site <http://www.fungaldiversity.org>



## Examples of bioactive compounds from Thai marine fungi

Species	Activity	Chemical structure	References
<i>Aigialus parvus</i> 	Anti-malarial	Aigialomycin D, Aigialone and aigialospirol	Isaka et al 2002; Vongvilai et al 2004
<i>Halorosellinia oceanica</i>	Weak anti-mycobacterial	Halorosellinic acid	Chinworrungsee et al, 2001
<i>Halorosellinia oceanica</i> 	Mild anti-mycobacterial	Halorosellins A, B	Chinworrungsee et al, 2004

## FUNGI ON DECAYING SEEDS AND LEAF LITTER



Leaf litter fungi  
(Twelve new described species)

- *Berkleasmium typhae*
- *Calcarisporium phaeopodium*
- *Dictyoarthrinium synnematicum*
- *Digitoramispora lageniformis*
- *Falcocladium turbinatum*
- *Infundibulomyces cupulata*
- *Infundibulomyces oblongisporus*
- *Lauriomyces cylindricus*
- *Lauriomyces ellipticus*
- *Melanogrophium proliferum*
- *Pseudoacrodictys dimorphospora*
- *Pseudorobillarda siamensis*

One fungus produces a new bioactive compound.

- *Kionochaeta ramifera*  
→ Ramiferin A



Fungi on decaying seed  
(Two new described species)

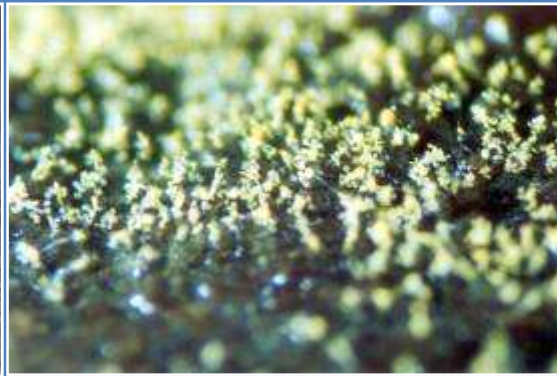
- *Cirrenalia nigrospora*
- *Lauriomyces sakaeratensis*

Two fungi produce 2 new bioactive compounds.

- *Kionochaeta pughii*  
→ Pughinin A
- *Menisporopsis theobromae*  
→ Menisporopsin A



## Fungal characteristics under dissection microscope



## Common species on decaying fruits and seeds



*Dictyochoaeta* sp.



*Thozetella nivea*



*Cryptophialoidea*  
*secunda*



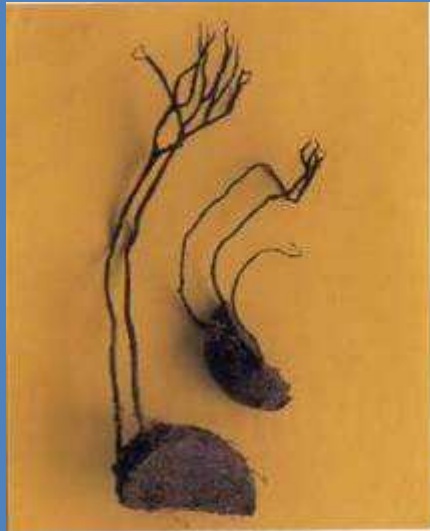
*Dinemasporium*  
*lanatum*

## Anamorphic fungi on decaying fruits and seeds



Somrithipol, S., Hywel-Jones, N.L. and Jones, E.B.G., and 2004. Seed fungi. In *Thai Fungal Diversity*. (E.B.G Jones, M. Tanticharoen and K.D. Hyde, eds.). BIOTEC, Thailand. pp. 129-140.

## Teleomorphic fungi on decaying fruits and seeds



*Xylaria* species  
**ASCOMYCETES**

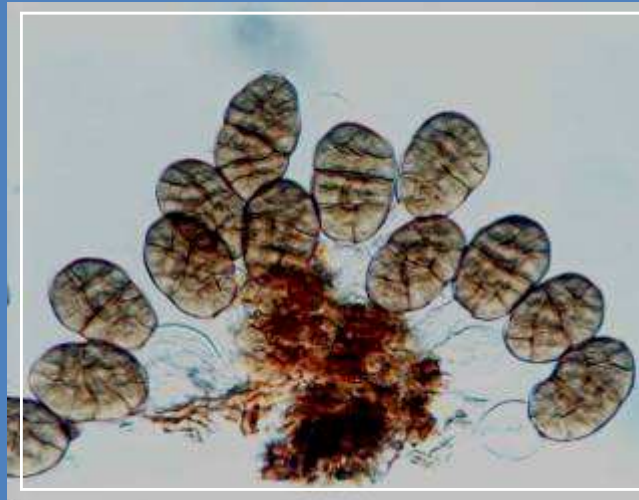


*Coprinus* species  
**BASIDIOMYCETES**

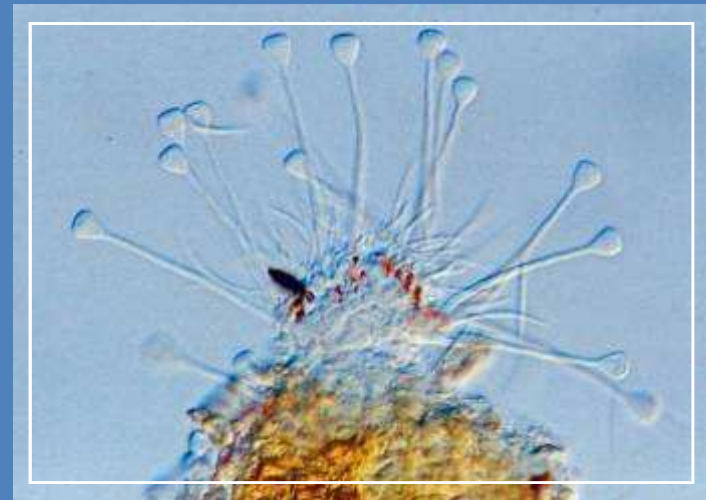
## Some new fungi on seed and leaf litter



*Cirrenalia nigrospora*



*Berkleasmium typhae*



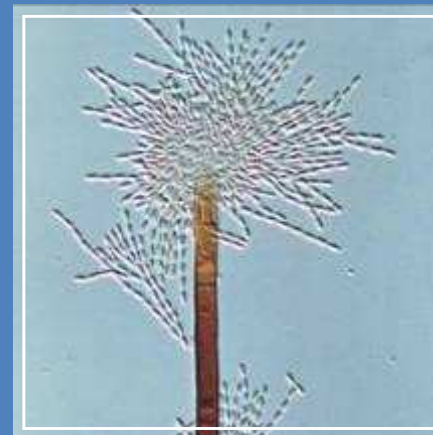
*Falcocladium turbinatum*



*Lauriomyces sakaeratensis*



*Lauriomyces cylindricus*

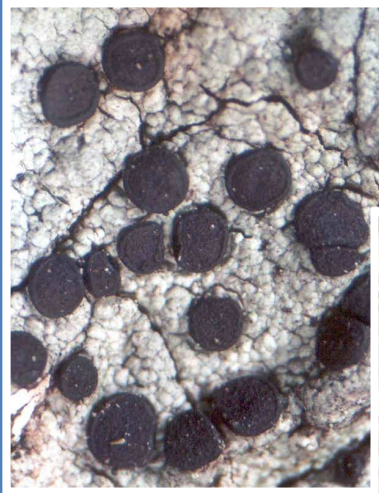


*Lauriomyces ellipticus*



*Melanographium proliferum*

# Lichens



## Lichen herbarium

3,515 specimens comprising

3 orders

14 families

33 genera and ~ 250 species

## Lichenized fungi

- 152 strains

- 36 strains showed the bioactive screening against anti-*Candida albicans* (30%), anti-*Herpes simplex* virus (50%), anti-tuberculosis (25-50  $\mu\text{g/ml}$ ), anti-cancer (5%) and anti-malaria at  $\text{ED}_{50} = 0.84\text{-}12.68 \mu\text{g/ml}$

- 2 new compounds were found

# Freshwater fungi

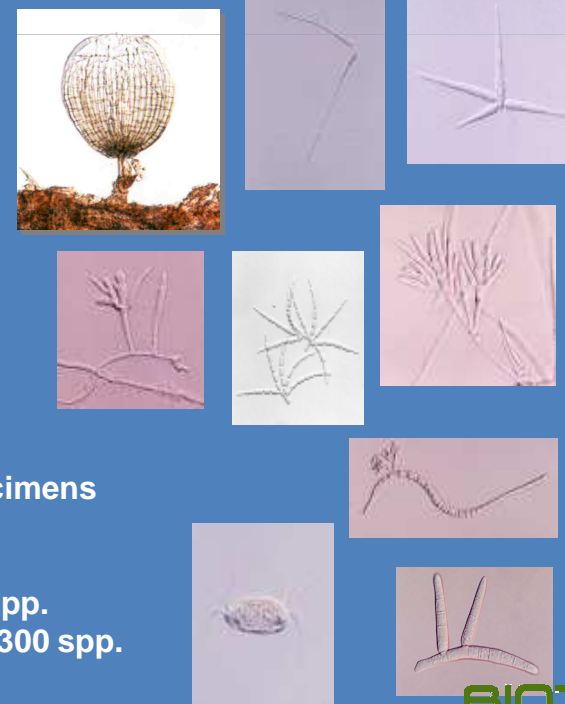
## Lignicolous Freshwater fungi



Groups encountered

- Ascomycota
- Basidiomycota
- Mitosporic (excluding Ingoldian fungi) : >300 spp.
- Aquatic & aero-aquatic fungi : ~ 200-300 spp.

## Aquatic & aero-aquatic fungi



# Identification of Xylariaceae by inducing the formation of fruiting bodies





# Palm Fungi

## Selected palms to study

- |                                 |   |                                 |
|---------------------------------|---|---------------------------------|
| 1. <i>Licuala longicalycata</i> | } | <b>Peat<br/>swamp<br/>palms</b> |
| 2. <i>Nenga pumila</i>          |   |                                 |
| 3. <i>Eleiodoxa conferta</i>    |   |                                 |
| 4. <i>Metroxylon sagu</i>       | } | <b>Terrestrial<br/>palms</b>    |
| 5. <i>Calamus spp.</i>          |   |                                 |
| 6. <i>Arenga sp.</i>            |   |                                 |
| 7. <i>Elaeis guineensis</i>     |   |                                 |

## Potential application of fungal isolates from palms

- Weight loss from basidiomycetes
- Antagonistic fungi from palms
- Edible mushroom cultivation from oil palm waste

## Results

- Saprophytes
- ✦ Ascomycetes 251 taxa
  - Basidiomycetes 298 collections
  - Anamorphic fungi 293 taxa
- Endophytes
- ✦ 717 morpho types (4 different trips)

## New Taxa

- ✦ 18 new species were described including 13 genera (5 new genera, 13 new species)
- ✦ 15 new species await descriptions

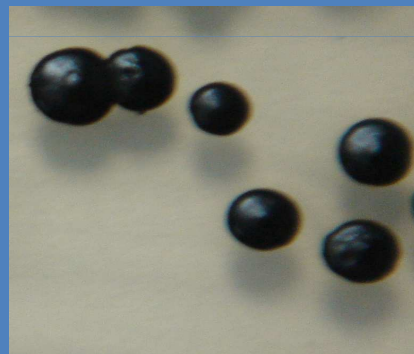
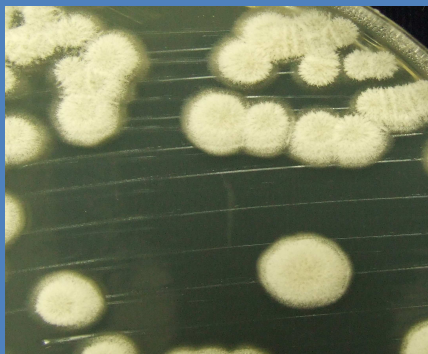
# Sources of yeasts



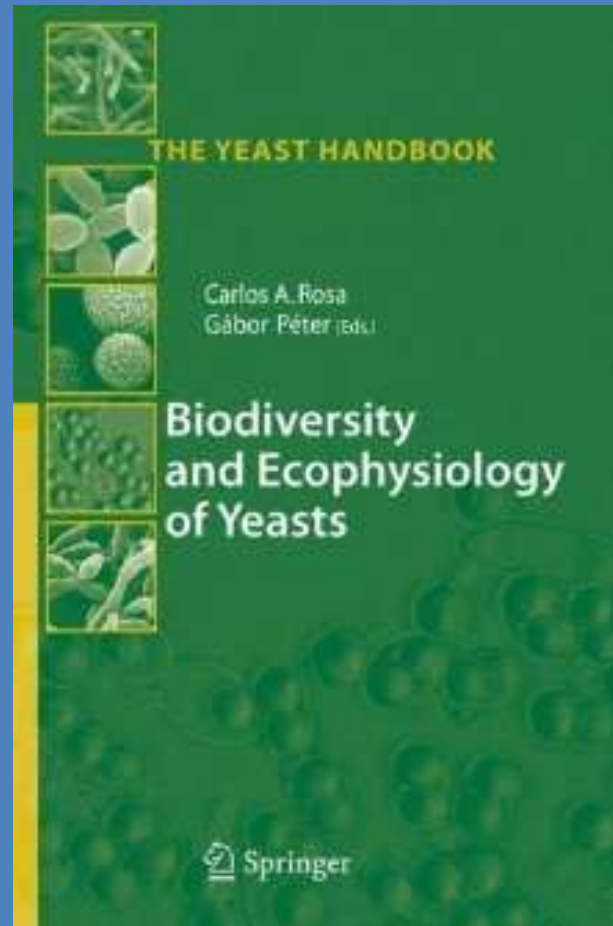
Insect frass



# Yeasts colony



# Biodiversity of Yeasts in Thailand and Asia



## Yeast biodiversity in tropical forest of Asia

by T. Nakase and S. Jindamorakot published in

## **The Yeast Handbook “Biodiversity and Ecophysiology of Yeasts”**

Edited by Carlos A. Rosa and Gábor Péter  
Springer January 2006

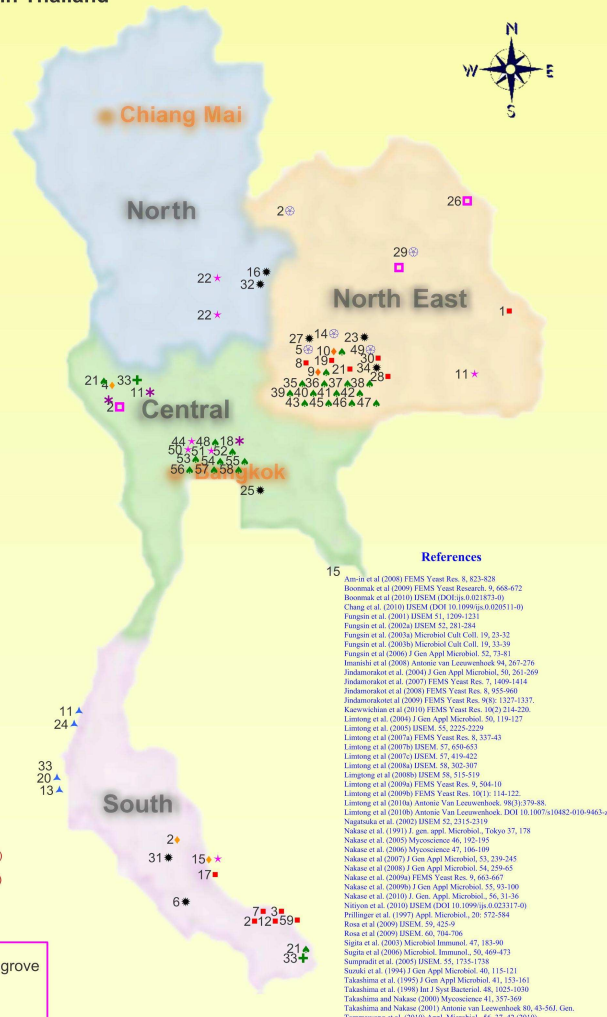
<http://www.cplbookshop.com/contents/C2426.htm>

# New Yeast Species Found in Thailand

Thailand have long been considered to be rich in microbial biodiversity. Since the first report describing new yeast species, *Kockovaella thailandica* and *K. imperatae* from Thailand (Nakase et al., 1991), the describing new yeast taxa have been increased year and year and 59 new yeast species from Thailand are currently recognized, 34 ascomycetous yeasts and 25 of basidiomycetous yeasts. These studies suggested that a vast number of unknown yeast species are living in the natural environment of Thailand.

## List of new yeast species found in Thailand

1. *Candida asiatica* (Limtong et al., 2010b)
2. *Candida easanensis* (Jindamorakot et al., 2004)
3. *Candida chanthaburiensis* (Limtong et al., 2010a)
4. *Candida gokubei* (Rosa et al., 2009)
5. *Candida jaronii* (Manshi et al., 2008)
6. *Candida gosongica* (Chang et al., 2010)
7. *Candida hasegawae* (Nakase et al., 2007)
8. *Candida kanchanaburiensis* (Nakase et al., 2008)
9. *Candida khaoyaiensis* (Jindamorakot et al., 2008)
10. *Candida kungkrabaensis* (Limtong et al., 2010)
11. *Candida krabiensis* (Limtong et al., 2004)
12. *Candida kazae* (Nakase et al., 2007)
13. *Candida lignicola* (Jindamorakot et al., 2007)
14. *Candida morakotiae* (Nakase et al., 2009)
15. *Candida nakhonratthasimensis* (Jindamorakot et al., 2004)
16. *Candida nonsortophila* (Nakase et al., 2008)
17. *Candida pattaniensis* (Jindamorakot et al., 2004)
18. *Candida phangngensis* (Limtong et al., 2008)
19. *Candida ratchasimensis* (Jindamorakot et al., 2008)
20. *Candida sesii* (Limtong et al., 2009b)
21. *Candida santii* (Limtong et al., 2009b)
22. *Candida saraburiensis* (Nityon et al., 2010)
23. *Candida siamensis* (Boonmak et al., 2008)
24. *Candida silbepensis* (Limtong et al., 2004)
25. *Candida songkhaensis* (Manshi et al., 2008)
26. *Candida stellimalicola* (Suzuki et al., 1994)
27. *Candida suralensis* (Limtong et al., 2010)
28. *Candida suwanarii* (Limtong et al., 2009b)
29. *Candida thailandica* (Jindamorakot et al., 2007)
30. *Candida thaimueangensis* (Limtong et al., 2007b)
31. *Candida wancheriae* (Nakase et al., 2009)
32. *Candida xylarijica* (Boonmak et al., 2010)
33. *Citeromyces siamensis* (Nagatsuka et al., 2002)
34. *Geotrichum phuruaeensis* (Kaeuwichian et al., 2010)
35. *Geotrichum siamensis* (Kaeuwichian et al., 2010)
36. *Hanseniaspora singularis* (Jindamorakot et al., 2009)
37. *Hanseniaspora thailandica* (Jindamorakot et al., 2009)
38. *Kazachstania siamensis* (Limtong et al., 2007c)
39. *Kloeckera hatyaiensis* (Jindamorakot et al., 2009)
40. *Kluyveromyces siamensis* (Am-in et al., 2008)
41. *Milleromyces petchaburiensis* (Tammawong et al., 2010)
42. *Ogataea chonburiensis* (Limtong et al., 2008)
43. *Ogataea nakhonphanomensis* (Limtong et al., 2008)
44. *Pichia jaronii* (Limtong et al., 2009a)
45. *Pichia koratensis* (Nakase et al., 2007)
46. *Pichia siamensis* (Limtong et al., 2004)
47. *Pichia nongkratonensis* (Nakase et al., 2005)
48. *Pichia thermomethanolicola* (Limtong et al., 2005)
49. *Tetraplopora namnaensis* (Sunpradit et al., 2005)
50. *Torulasporea mateae* (Limtong et al., 2007a)
51. *Vanderwaltozyma tropicalis* (Nakase et al., 2010)
52. *Wickerhamomyces edaphicus* (Limtong et al., 2009)
53. *Bullera arundinariae* (Fungsin et al., 2002a)
54. *Bullera koratensis* (Fungsin et al., 2006)
55. *Bullera lagerstroemiae* (Fungsin et al., 2006)
56. *Bulleringia musae* (Takashima et al., 1995)
57. *Bullera patii* (Fungsin et al., 2003a)
58. *Bullera penniseticola* (Takashima et al., 1998)
59. *Bullera sakaratica* (Fungsin et al., 2003b)
60. *Bullera siamensis* (Fungsin et al., 2003a)
61. *Berlingeria thailandica* (Fungsin et al., 2001)
62. *Fellomyces thailandicus* (Prüllinger et al., 1997)
63. *Kockovaella barringtoniae* (Fungsin et al., 2002a)
64. *Kockovaella imperatae* (Nakase et al., 1991)
65. *Kockovaella thailandica* (Nakase et al., 1991)
66. *Kockovaella sacchari* (Takashima et al., 1998)
67. *Moniliella fonsaeae* (Rosa et al., 2009)
68. *Pseudozyma parantarctica* (Sugita et al., 2003)
69. *Pseudozyma thailandica* (Sugita et al., 2003)
70. *Sporobolomyces nylandii* (Takashima and Nakase, 2000)
71. *Sporobolomyces ponoosokiae* (Takashima and Nakase, 2000)
72. *Sporobolomyces burmese* (Takashima and Nakase, 2000)
73. *Sporobolomyces verticillatus* (Takashima and Nakase, 2000)
74. *Tilletopsis dextri* (Takashima and Nakase, 2001)
75. *Tilletopsis oryzoicola* (Takashima and Nakase, 2001)
76. *Tilletopsis penniseti* (Takashima and Nakase, 2001)
77. *Trichosporon siamense* (Nakase et al., 2004)



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# Diversity of yeasts in Thailand

Many new species of yeasts have been isolated from Thailand and described in recent years.

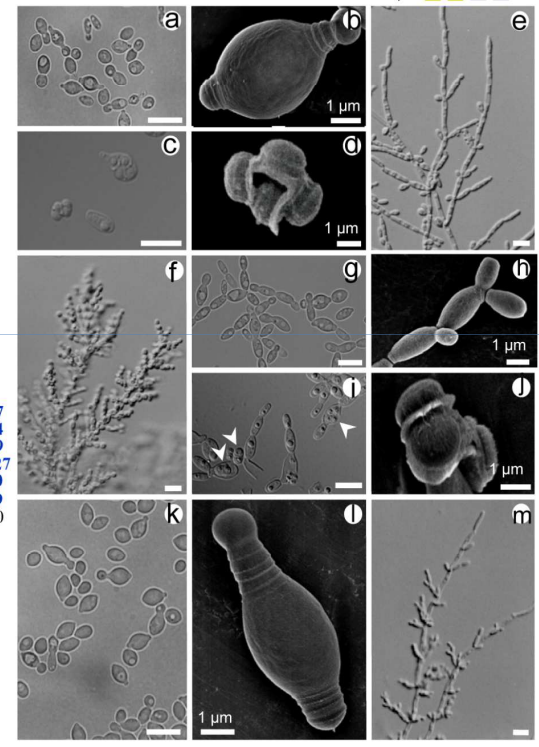
More than 80 species of ascomycetous yeasts have been described as new species from various kinds of substrates in Thailand from 1991-2012.

# Many new yeast species published from Thailand

## Three new *Hanseniaspora*/*Kloeckera*

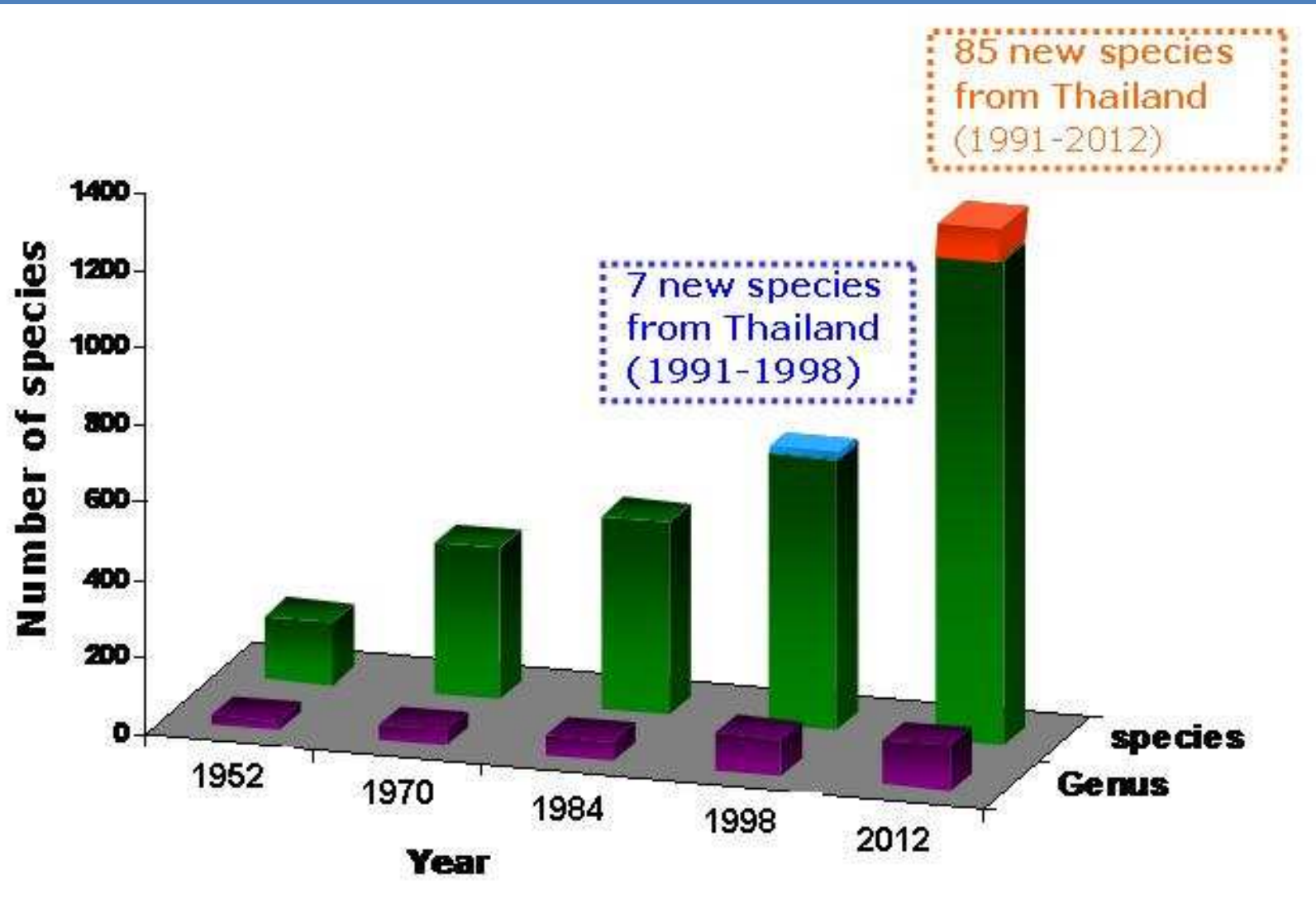


- Hanseniaspora opuntiae* (ST-9)
- Hanseniaspora opuntiae* (ST-8)
- Hanseniaspora opuntiae* (ST-5)
- Hanseniaspora opuntiae* (ST-481)
- Hanseniaspora opuntiae* (ST-448)
- Hanseniaspora opuntiae* AY267820
- Hanseniaspora opuntiae* AJ512451
- Hanseniaspora opuntiae*<sup>1</sup> AJ512453
- Hanseniaspora lachancei* AJ512457
- Hanseniaspora lachancei*<sup>1</sup> AJ512459
- Hanseniaspora guilliermondii* EF449520
- Hanseniaspora guilliermondii*<sup>1</sup> U84230
- Hanseniaspora pseudoguilliermondii*<sup>1</sup> AJ512455
- Hanseniaspora pseudoguilliermondii* (ST-398)
- Hanseniaspora meyeri*<sup>1</sup> AJ512454
- Hanseniaspora meyeri* AJ512460
- Hanseniaspora meyeri* AJ512461
- Hanseniaspora meyeri* AJ512458
- Hanseniaspora clermontiae* AJ512456
- Hanseniaspora clermontiae*<sup>1</sup> AJ512452
- Hanseniaspora iwarum* AF257273
- Hanseniaspora iwarum* AM160628
- Hanseniaspora iwarum*<sup>1</sup> U84229
- Hanseniaspora meyeri* (G4p4) AY305671
- Hanseniaspora iwarum* (G10p1) AY305681
- Hanseniaspora thailandica* (ST-613) DQ414467
- Hanseniaspora thailandica* (ST-306) DQ404494
- Hanseniaspora thailandica* (ST-250) DQ404489
- Hanseniaspora thailandica*<sup>1</sup> (ST-464) DQ404527
- Hanseniaspora thailandica* (ST-493) DQ404519
- Hanseniaspora thailandica* (ST-391) DQ404519
- Hanseniaspora meyeri* (G4p1) AY305670
- Kloeckera songkhlaensis*<sup>1</sup> (ST-476) DQ404528
- Kloeckera lindneri*<sup>1</sup> U84226
- Kloeckera lindneri* EF460514
- Kloeckera lindneri* EF653913
- Kloeckera lindneri* (ST-484)
- Hanseniaspora valbyensis* AJ966341
- Hanseniaspora valbyensis*<sup>1</sup> U73596
- Hanseniaspora singularis*<sup>1</sup> (ST-244) FJ391977
- Hanseniaspora occidentalis*<sup>1</sup> U84225
- Hanseniaspora osmophila*<sup>1</sup> U84228
- Hanseniaspora osmophila* AF533065
- Hanseniaspora vineae*<sup>1</sup> U84224
- Hanseniaspora vineae* (ST-515)
- Saccharomyces cerevisiae*<sup>1</sup> U44806



# The increasing number of recognized species

(The Yeast, A Taxonomic study 5<sup>th</sup> edition)



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Thank you for your attention

