

Novel methods for early detection of the bloody red shrimp, *Hemimysis anomala*

Meghan Brown, Hobart and William Smith Colleges

Brent Boscarino & *Sonomi Oyagi*, Poughkeepsie Day School

Mike Tibbetts & Carla Sanchez, Bard College



HOBART AND WILLIAM SMITH COLLEGES



CPG

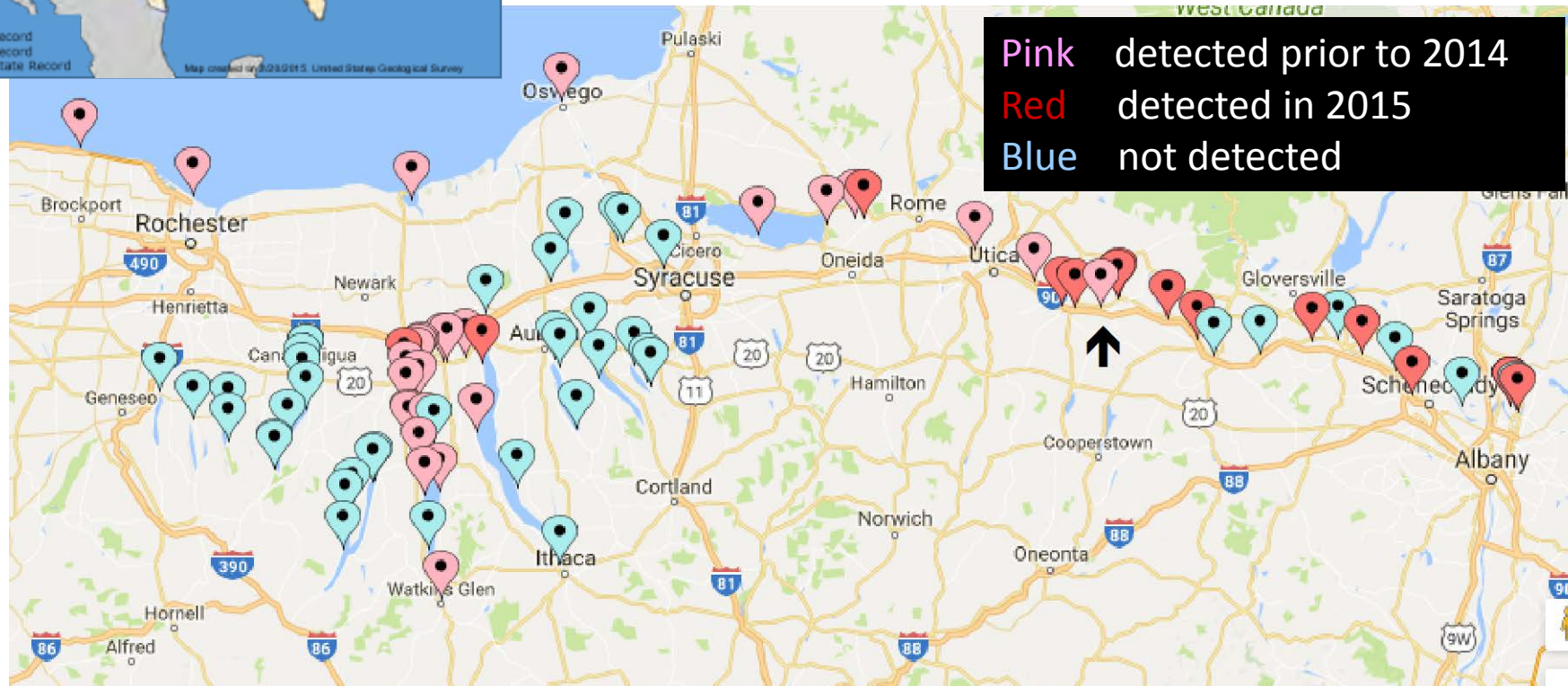
An emerging invader: The Bloody Red Shrimp, *Hemimysis anomala*



Female with developing embryos removed from the marsupium

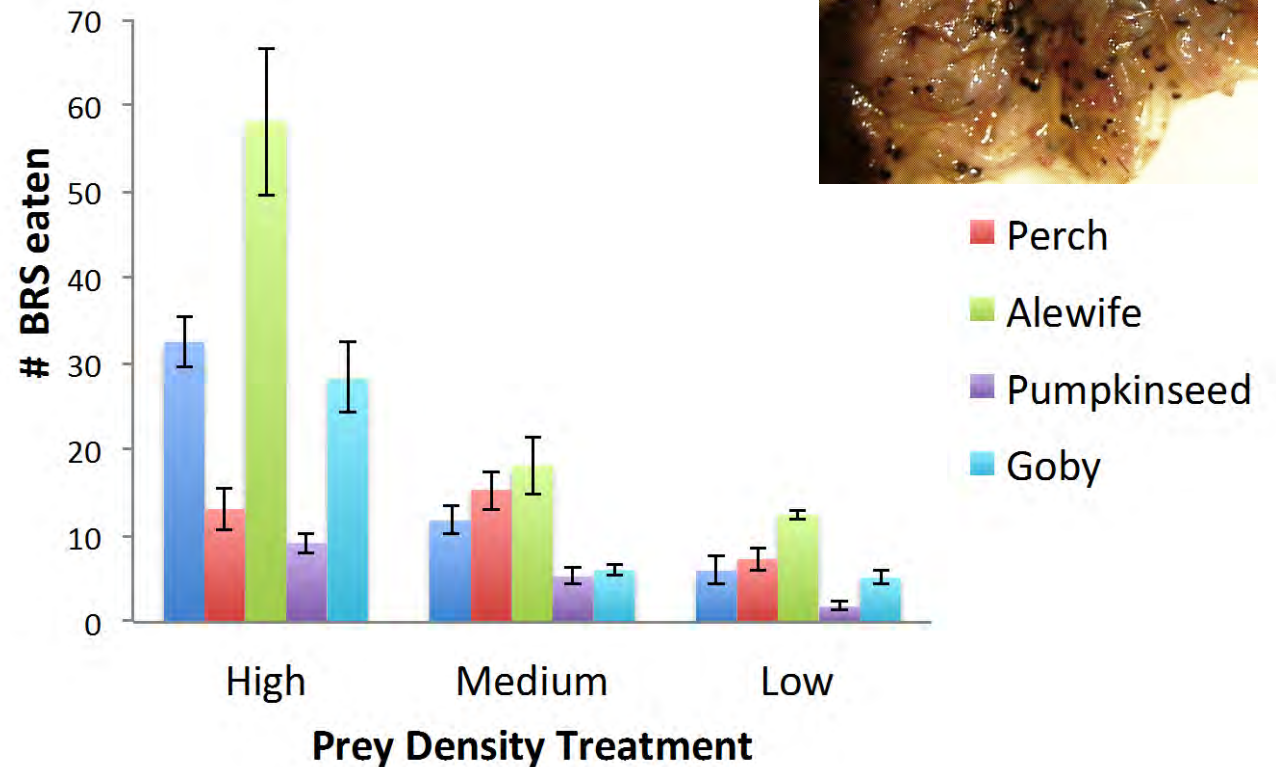
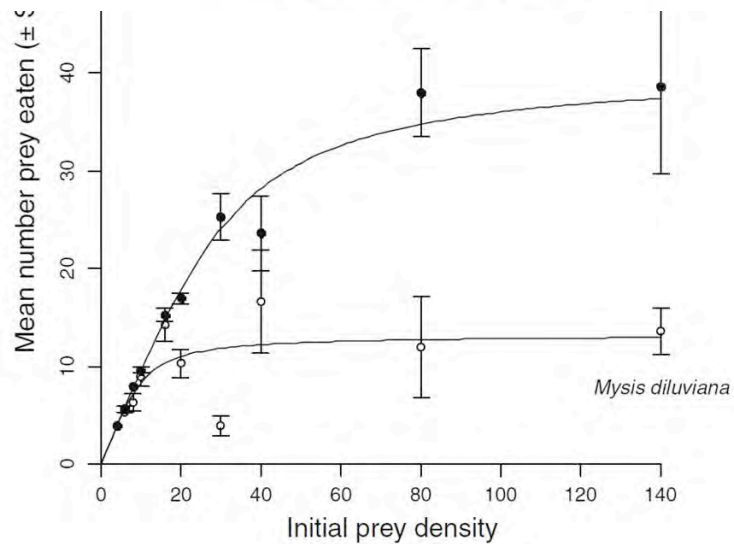
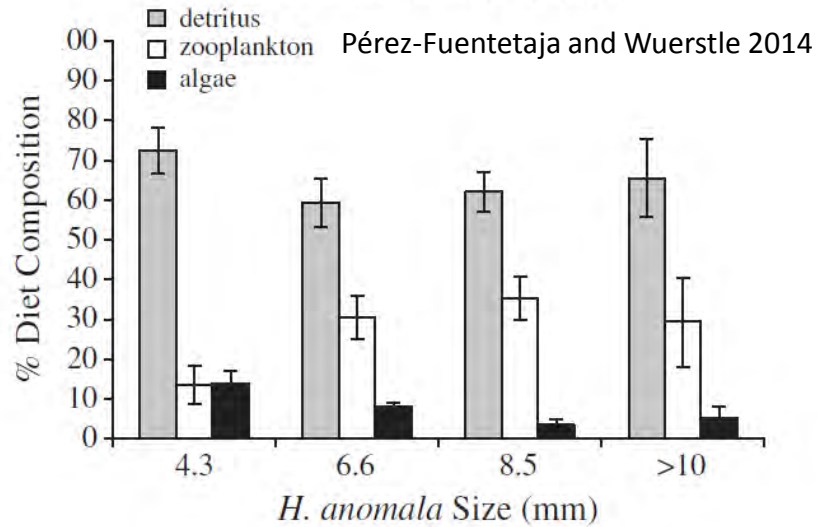


Microphotograph of a bloody red shrimp. Length approximately 1/4 to 1/2-inch.



Hemimysis is an omnivore with high consumption rates.

And, a prey item for fish.



Limitations of plankton net collections.



Light-based traps are an effective detection method



Contents lists available at ScienceDirect

Journal of Great Lakes Research

journal homepage: www.elsevier.com/locate/jglr



The light at the end of the funnel?: Using light-based traps for the detection and collection of a nearshore aquatic, invasive invertebrate, *Hemimysis anomala*

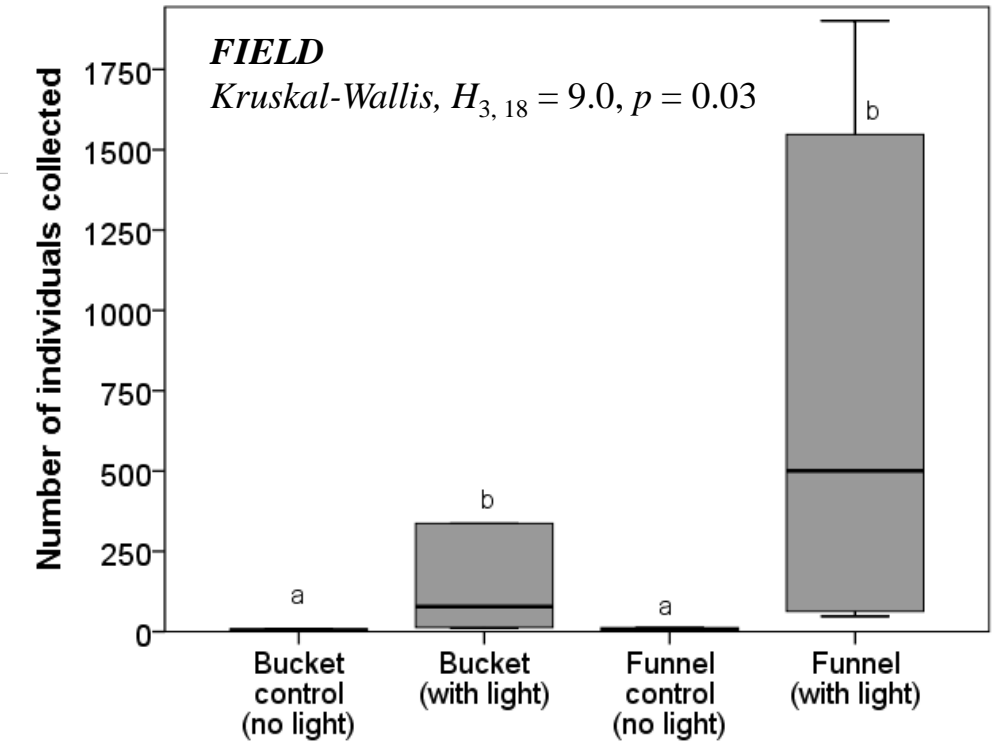
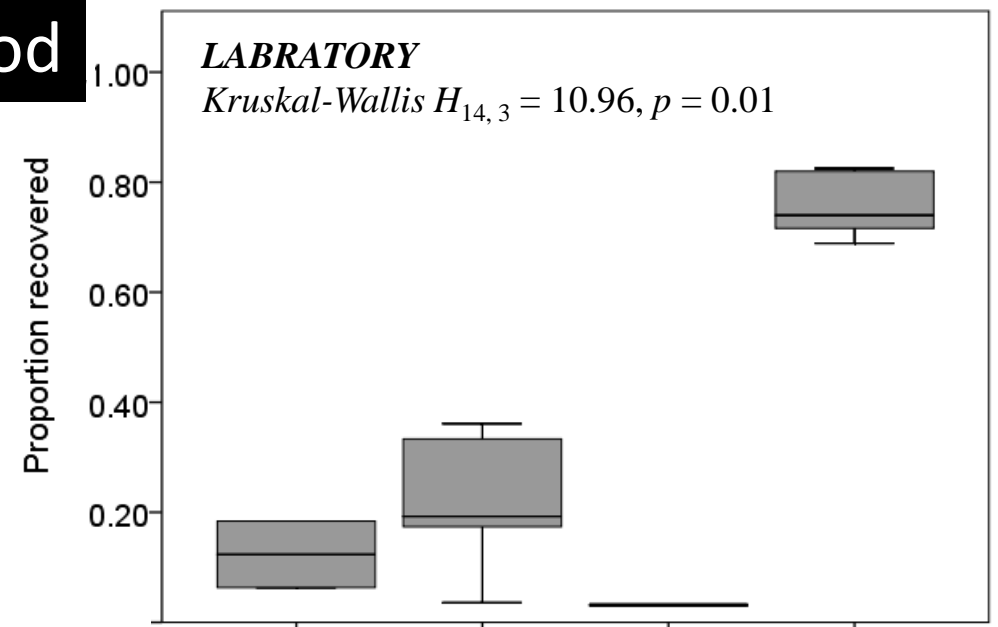
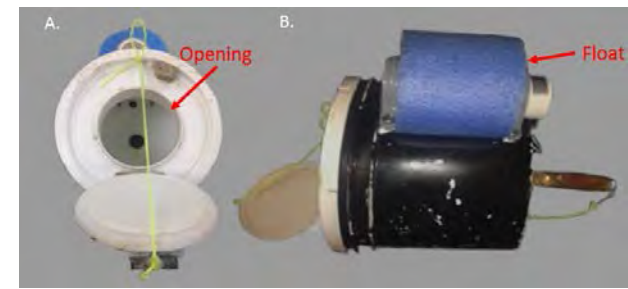
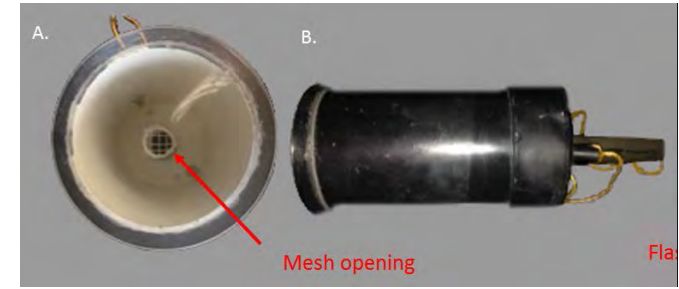
Meghan Brown ^{a,*}, Jamila Roth ^b, Bruce Smith ^c, Brent Boscarino ^d

^a Hobart and William Smith Colleges, Department of Biology, 300 Pulteney Street, Geneva, NY 14456, USA

^b Skidmore College, Departments of Biology and Environmental Science, 815 N Broadway, Saratoga Springs, NY 12833, USA

^c Ithaca College, Biology Department, 953 Danby Rd., Ithaca, NY 14850, USA

^d Poughkeepsie Day School, 260 Boardman Road, Poughkeepsie, NY 12603, USA



eDNA as a potential early-detection tool

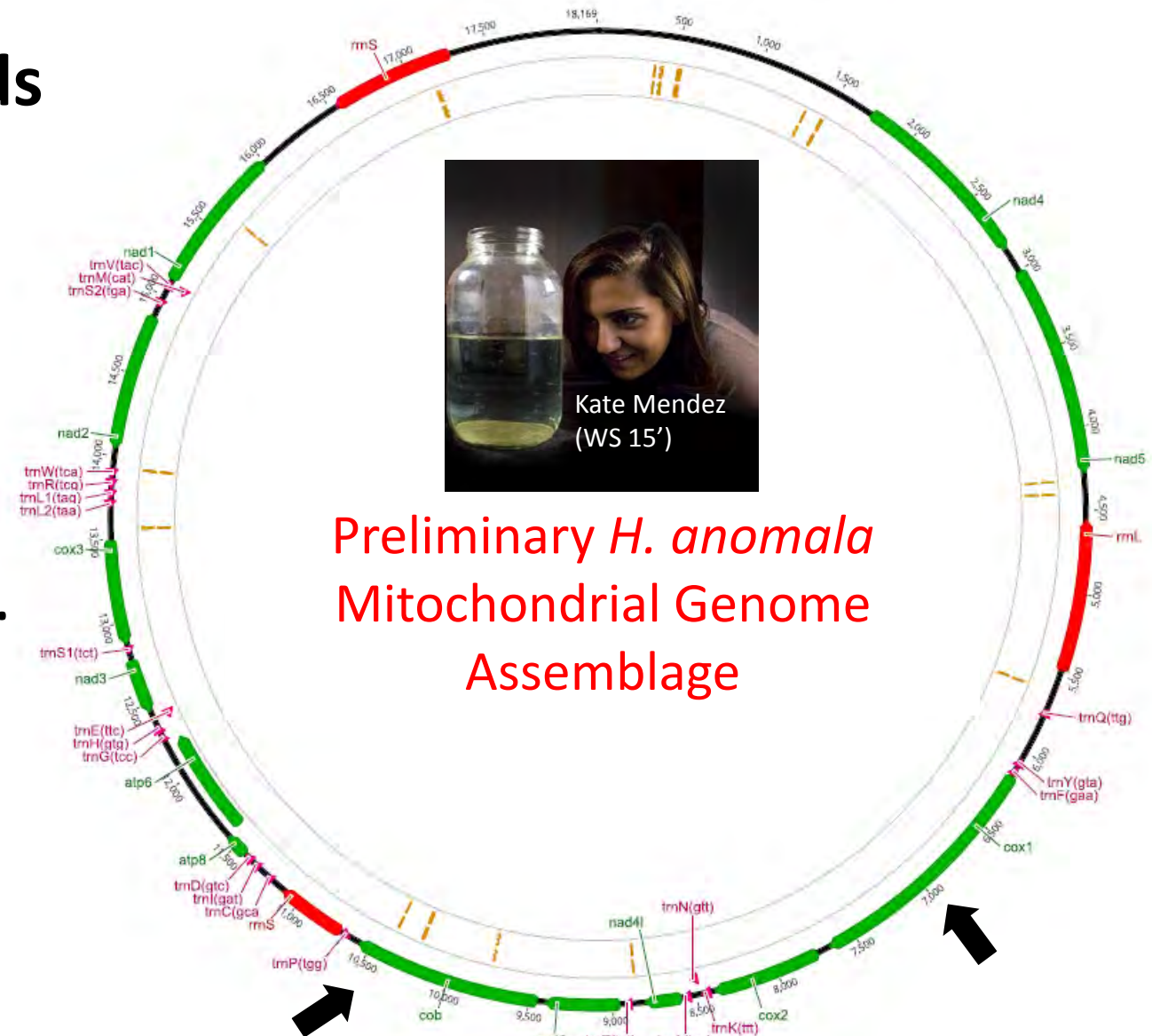
DNA extraction and qPCR methods

Field studies to compare

- habitat types,
- times of day, and
- locations in the water column.

Lab experiments to compare

- *Hemimysis* density and
- incubation time.



What are the most suitable primer for qPCR detection of *Hemimysis*?

mt-DNA, subunit I of cytochrome oxidase gene

HAc01-4AF & HAc01-4BF (two variants)

- 35 thermal cycles, most reliable time frame and avoided primer dimers
- 54 -55°C ideal for annealing

```
HA -----Gggttagtaggttcttctttgagaattttaatt
MD ctctatthttgtgtttggggcttgggctggaatagttggatcttctttaagagttttaatt
      ** ****  ** *****  ** *****

HA cgacttgaattgggtcagcccggtaggttaattggggatgatcagattataacgtgatt
MD cggttagagttaggccaacctgggcatttgattggggacagacaactttataacgtaatt
      ** * ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** **

HA gtgacagcgcacgtcttggtaataatthttttatagttataccaaccataattgggtgt
MD gttactgcgcacgtcttggtaataatthttctttatagttatacctattacagttgggtgg
      ** ** *****  ***** ***** ***** * * * *****

HA ttcggaactggtagttcttthaataattggggccccagatatagcgtttctcgaata
MD ttggaaattggtagtaccgttgatattaggagctcctgatagcttttctcgtata
      ** ** ** ***** ** ** * ** ** * ** ***** ***** **

HA aataatataagatthttgattattacctccatcggttaagattgctthtttagctagagggtta
MD aataatataaggthttgactthttgccaccttctttagctcttataactaataagaggata
      ***** ***** * ** * ** * ** * * * * * ** * ** ***** **

HA gtggaggggggggttggtaggggtggacagtctaccctcccttagcgggtaacgtgtca
MD gtagagagaggggttagggactggttgaacgthttatccaccattggcttcaaacattgct
      ** *** * ***** ** ** ** * ** * ** * ** * ** * ** * ** * **

HA cacatgggtgcagctgtggatataggaatthtttctttgcatttagctggcgcctcttct
MD catgcaggggcagcagtagatataggaatthtttctttacatttagctggggcttcttca
      **      ** ***** ** ***** ***** ***** ***** ** *****

HA atthttaggtgctgttaatthtttctacagttattaataacgtgctgtaggaataggg
MD atthttaggtgctgtaaatthtttcaactgttattaataacgggcacctggggtaggt
      ***** ***** ** ***** ***** **      ** ****

HA ttgacagtataccactatthttgtttggtctgtgthttactgctatthttactacttctg
MD atagatagactacctctthttgtgtggtaatthtttattacagcaattctthttactt
      * ** ** ***** ** ***** ***** * ***** ** ** * ** **

HA tctttacctgthtttagcaggggctattactatgctt-----
MD tctttaccagtgttagcaggggctattacaatactthtaacagaccgtaatthtaact
      ***** ** ***** ***** ** **

HA -----
MD tctttctttgacctgtaggtgggggtgacctta
```

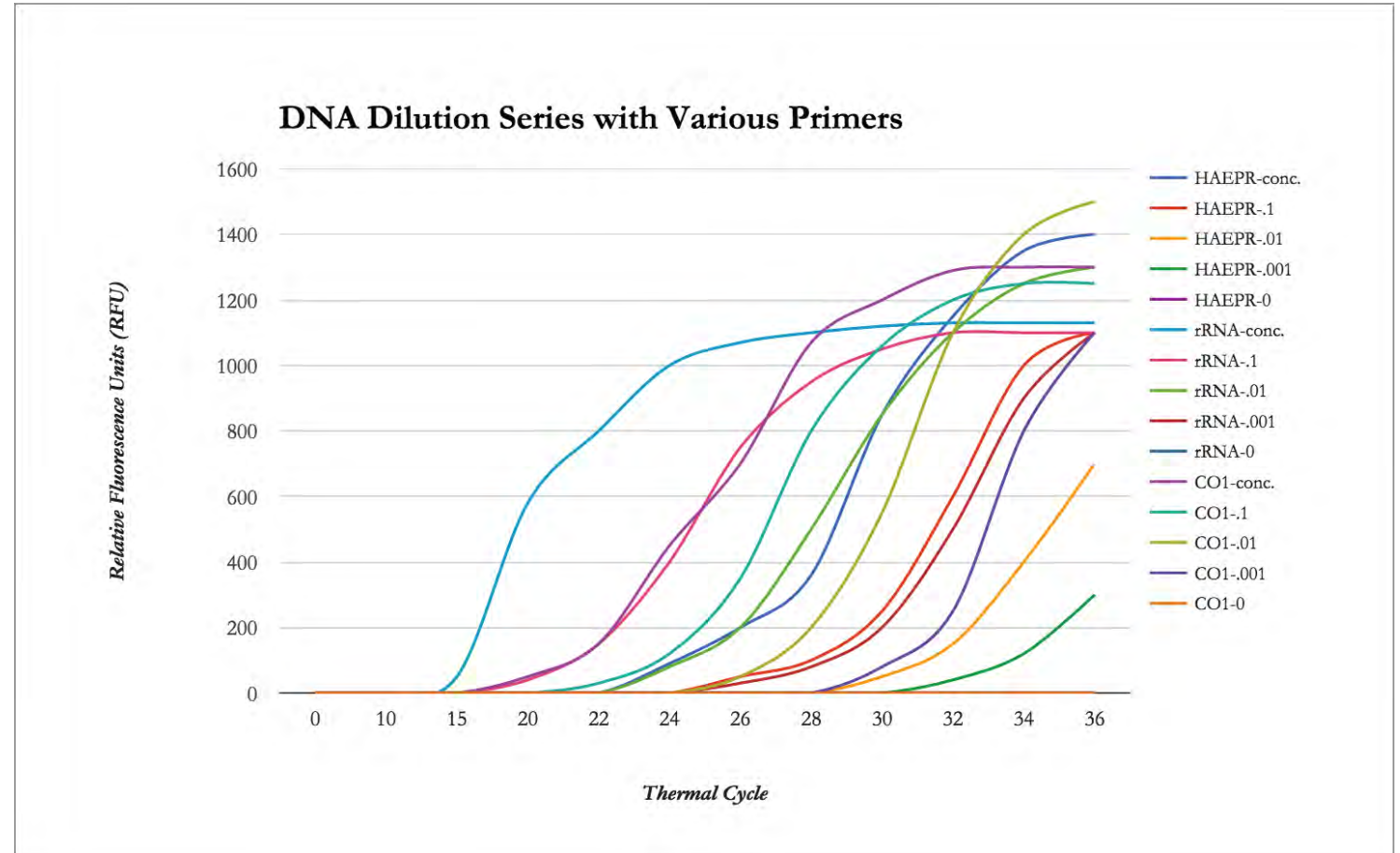
What densities and consequent concentration of eDNA are detectable?

Dilutions of pure *Hemimysis* DNA

1/10

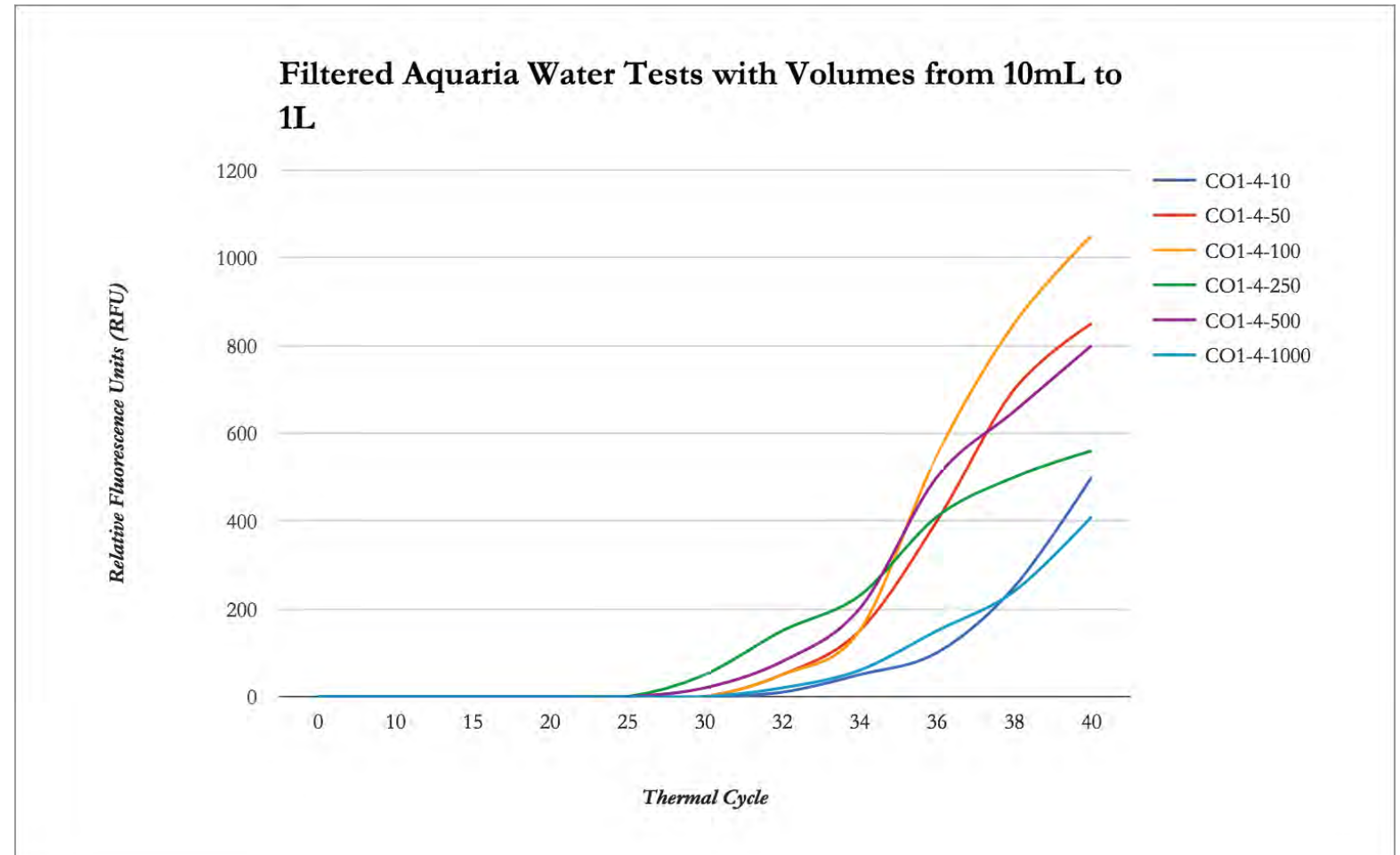
1/100

1/1000



What volume of water is needed for reliable detection?

Water volumes from aquaria holding moderate density of *Hemimysis*
10-1000 mL



Field experiments

Sites: Seneca Lake (established by 2010)
Erie Canal Lock #5 (established by 2014)

Location : Water and sediment surface

Time mid-day and midnight

Controls: Skaneateles Lake, field and lab controls



Preliminary Results!

# reps with cq < 35		Seneca Lake		NYS Canal	Controls	
		Filter size (μm)			Skaneateles Lake	Field controls
Night	Deep	0.45	3/3		0/3	0/3
		0.65	3/3			
		1.2	3/3			
	Shallow	0.45	1/3		0/3	
		0.65	2/3			
		1.2	1/3			
Day	Deep	0.45	2/3			0/3
	Shallow	0.45	2/3			

Methods:

Triplicate 250 mL water samples filtered (0.45 μm pore-diameter acetate filter paper)

Filters preserved in 70% ethanol

Equipment cleaned between replicates (50% bleach and DI rinse)

Field controls were deionized water transferred to sample bottles in the field and processed the same as field samples

Lab experiments

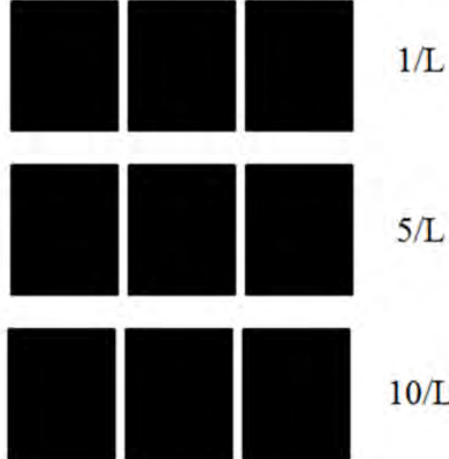
Mysis Control
6L buckets with 30 organisms/L



Well Water Control
6L buckets



Hemimysis anomala control
6 L buckets with varied densities



Water samples at

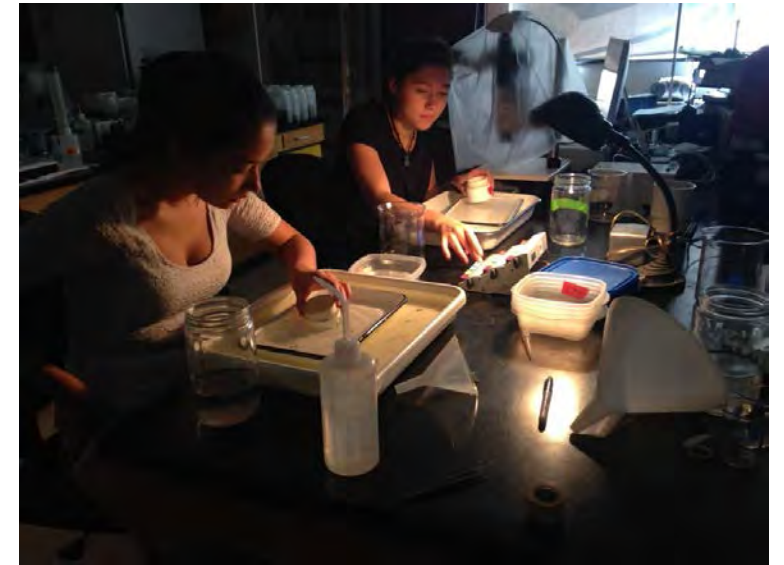
- 3, 5, 7, 9, 11, 13 days
- 12 hr and 2 days after *Hemimysis* removed



500 mL water samples



Filter Samples



Questions?

