





<b>SPECIES</b>	<b><i>Eriophyllum confertiflorum</i> (DC.) A. Gray var. <i>confertiflorum</i></b>	
<b>NRCS CODE:</b> <b>ERCOC12</b>  <p>© 2006 Steve Matson</p>	Family: Asteraceae Order: Asterales Subclass: Asteridae Class: <b>Magnoliopsida</b>  <p>seedling, A. Montalvo 2009</p>	  <p>A. Montalvo, Riverside Co.</p>
<b>Subspecific taxa</b>	Of the two varieties, the focus of this profile is on var. <i>confertiflorum</i> . 1. <i>E. c.</i> var. <i>confertiflorum</i> 2. <i>E. c.</i> (DC.) A. Gray var. <i>tanacetiflorum</i> (Greene) Jeps. (FNA 2010, JepsonOnline 2010)	
<b>Synonyms</b>	<i>E. c.</i> (DC.) A. Gray var. <i>discoideum</i> (Rydb.) Munz, <i>E. c.</i> (DC.) A. Gray var. <i>latum</i> H. M. Hall, <i>E. c.</i> (DC.) A. Gray var. <i>laxiflorum</i> A. Gray, <i>E. c.</i> (DC.) A. Gray var. <i>tridactylum</i> (Rydb.) Munz (USDA PLANTS 2010).	
<b>Common name</b>	golden-yarrow, yellow yarrow, long-stemmed golden yarrow	
<b>Taxonomic issues</b>	The FNA (2010), USDA PLANTS (2010), and JepsonOnline (2010) all recognize two varieties of <i>Eriophyllum c.</i> for North America. However, FNA lists only <i>E. c.</i> var. <i>latum</i> and <i>E. c.</i> var. <i>laxiflorum</i> as synonyms of var. <i>confertiflorum</i> , and JepsonOnline reports that the name <i>E. c.</i> var. <i>latum</i> is unresolved. Hybridization may make identification and classification difficult in this morphologically variable species.	
<b>Taxonomic relationships</b>	Based on studies of between-taxon crossability and progeny fertility, <i>E. confertiflorum</i> is hypothesized to be most closely related to <i>E. lanatum</i> (Pursh) Forbes, which shares the same base number of chromosomes (n=8) and with which it hybridizes, and to <i>E. staechadifolium</i> Lag., n=19 (Mooring 1997). Constance (1937) also identified the more northern and coastal <i>E. staechadifolium</i> (Channel Is. northward) as a close relative based on shared morphological traits, including perennial habit, heads with short peduncles, and several other traits of the inflorescence.	
<b>Related taxa in region</b>	<i>E. confertiflorum</i> var. <i>tanacetiflorum</i> (= <i>E. tanacetiflorum</i> Green), an octoploid (Mooring 1994), is native to oak woodlands of the central Sierra Nevada foothills and differs from <i>E. c.</i> var. <i>c.</i> primarily by having only 1-10 heads per inflorescence, longer disk flowers (3.5-4 mm), and wider involucre (5-7 mm) (JepsonOnline 2010). The low-stature annual species, <i>E. multicaule</i> (DC.) A. Gray and <i>E. wallacei</i> (A. Gray) A. Gray overlap in distribution with var. <i>confertiflorum</i> in southern California where they can co-occur in alluvial scrub habitat.	
<b>Other</b>	Extremely variable in degree of compactness and leaf traits (Munz & Keck 1968, Munz 1974).	
<b>GENERAL</b>		
<b>Map</b>	Data provided by the participants of the Consortium of California Herbaria represent 708 records for var. <i>confertiflorum</i> with coordinate data out of 1267 records retrieved; accessed 9/25/10; Some specimens may be misidentified. Berkeley Mapper: <a href="https://ucjeps.berkeley.edu/consortium/">https://ucjeps.berkeley.edu/consortium/</a>  note: the northernmost specimen was collected in 1874 by an unknown collector and with no number. An annotation by L. Constance in 1933 says "!!Locality doubtful!!"	

<b>Geographic range</b>	California, south into Baja CA (Hickman 1993, FNA). It is possible that range extensions have occurred due to its use in roadside seeding projects.
<b>Distribution in California; Ecological section and subsection</b>	Common in Northern Baja CA to n. CA except in the Central Valley, North coastal strand, deserts, and interior north west mountains (Munz 1974, Hickman 1993, JepsonOnline); Ecological Units and Subsections (Goudey & Smith 1994, Cleland et al. 2007): North coast ranges (263A), Northern California Coast Range (M261A), Central California Coast Ranges (M262A), Sierra Nevada (M261E) and Foothills (M261F), Central Western CA (M261B,C), SW CA (M261B,M262B), western edge of deserts (322A,C) (Hickman 1993).
<b>Life history, life form</b>	Polycarpic, suffrutescent perennial to subshrub; woody at base to slender stems (Munz 1974).
<b>Distinguishing traits</b>	The short, generally 2-6 dm tall, suffrutescent plants are generally tomentose, with soft whitish hairs flattened against the many erect stems that arise from a woody base. The undersides of the leaves are whitish-tomentose and the upper surface is greenish with sparse hairs. The blades are dissected into narrow, linear lobes with inrolled margins. The plants bear many dense heads of yellow flowers on short peduncles in compound corymbs born on leafy stalks, reminiscent of true yarrow in the genus <i>Achillea</i> ; each head bears 10-35 disk flowers and 4-6 ray flowers (sometimes 0), with corollas about 2-3 mm long. The 4 to 7 ovate and overlapping membranous bracts of the involucre (phyllaries) form an erect cup and are retained on the plant into the next flowering season (Munz & Keck 1968, Clarke et al. 2007).
<b>Root system, rhizomes, stolons, etc.</b>	Taproot (Clarke et al. 2007).
<b>Rooting depth</b>	Roots are generally < 0.5 m in depth (A. Montalvo, pers. obs., Andrew Sanders, curator, UCR Herbarium, pers. com.)
<b>HABITAT</b>	
<b>Plant association groups</b>	Coastal sage scrub, chaparral (Munz 1974). <i>E. c.</i> var. <i>c.</i> is considered to be an indicator of coastal sage scrub (Munz & Keck 1968) but it is also found in many other communities, sometimes as a co-dominant (Sawyer et al. 2009).
<b>Habitat affinity and breadth of habitat</b>	Dry slopes and washes near coast (Munz 1974) and dry slopes of inland valleys and foothills. Plants are a common component of postfire vegetation in coastal sage scrub and chaparral. Postfire populations tend to persist for up to five years (Keeley & Pizzorno 1986), but populations can persist in open areas and along roadsides. In a broad study of coastal sage scrub, Kirkpatrick & Hutchinson (1980) found plants primarily on slopes of northern aspect and more often on slopes under 5 degrees.
<b>Elevation range</b>	Below 3000 m (Hickman 1993).
<b>Soil: texture, chemicals, depth</b>	Disturbed, shallow, rocky soils (Mooring 1994); dry (Hickman 1993); granitic, unconsolidated soils, and other soils (Kirkpatrick & Hutchinson 1980). In the south Coast Ranges, it is associated with serpentine soils and may be an indicator for serpentine or other ultramaphic substrates (Kruckeberg 1984).
<b>Drought tolerance</b>	High (Mooring 1994).
<b>Precipitation</b>	Plants occur in areas that range from 10 to more than 25 inches annual precipitation.
<b>Flooding or high water tolerance</b>	Plants are not adapted to wet soils but do occur in middle to upper terraces of alluvial scrub where inundation is infrequent.
<b>Wetland indicator status for California</b>	None.
<b>Shade tolerance</b>	Withstands full sun to partial shade (O'Brien et al. 2006). Generally a full sun species, requiring at least some full sun in the day (Schmidt 1980).
<b>GROWTH AND REPRODUCTION</b>	
<b>Seedling emergence relevant to general ecology</b>	Seedlings are not found beneath the canopy of chaparral; following fire growth is from germinating seeds, not re-sprouts (Keeley et al. 1981). Seedlings emerge in the rainy season in mid winter.
<b>Growth pattern (phenology)</b>	Plants reach reproductive maturity (flowering) within two years and flower from May to August. Summer drought leads to leaf dormancy and a seasonal dimorphism in leaf growth (dry vs. growing season) (Westman 1981). Small leaves are produced on short shoots that grow from axillary buds along the main stem late in the growing season and persist through the dry season. Larger leaves are produced early in the growing season from the more elongated stems and tend to drop during summer drought. Plants become crowded out as taller shrubs develop in the years following fires (Keeley et al. 1981).
<b>Vegetative propagation</b>	None.

<b>Regeneration after fire or other disturbance</b>	Fire stimulates seed germination and plants become most abundant, sometimes in mass, after wildfire in coastal sage scrub and chaparral (Keeley & Nitzberg 1984, Keeley & Pizzorno 1986, Zammit & Zedler 1994).
<b>Pollination</b>	Pollen is dispersed by a diverse assemblage of insects, many of which are generalists. Mooring (1994) reports "bees and beetles" while Moldenke (1976) reports a diverse assemblage of insects, including butterflies, wasps, and many kinds of bees as flower visitors and potential pollinators.
<b>Seed dispersal</b>	Seeds (achenes) are very small and autodispersed over the summer (Keeley 1991).
<b>Breeding system, mating system</b>	Self-incompatible (Mooring 1975, 1994). Flowers that were enclosed in bags produced very few fruits, and only 0.2% germinated. Artificial self-pollinations were not attempted.
<b>Hybridization potential</b>	High; natural hybridization with <i>E. lanatum</i> var. <i>arachnoideum</i> (Hickman 1993; Mooring 1994; Mooring 2001). Visitation by generalist pollinating insects may increase potential for hybridization in areas where related taxa overlap in flowering.
<b>Inbreeding and outbreeding effects</b>	Crosses among geographically distant, diploid populations of <i>E. c.</i> var. <i>confertiflorum</i> resulted in outbreeding depression, with more distant crosses having lower seed germination than close crosses (Mooring 1994). However, germination of first generation (F <sub>1</sub> ) seeds and pollen viability of progeny (an indicator of hybrid sterility) did not differ significantly between intra- and inter-variety crosses between <i>E. c.</i> var. <i>confertiflorum</i> and <i>E. c.</i> var. <i>tanacetiflorum</i> (Mooring 1994). Hybridization with the related <i>E. lanatum</i> yielded on the average, <65% pollen viability in offspring while intra-variety hybridization yielded 75% pollen viability.
<b>BIOLOGICAL INTERACTIONS</b>	
<b>Competitiveness</b>	Although frequent after fire, plants die out as the canopies of larger, woody shrubs close in (A. Montalvo, pers. obs.).
<b>Herbivory, seed predation, disease</b>	"Insect larvae" consume ~25% of fruits (Mooring 1994). A tephritid fly, <i>Paroxyyna genalis</i> , consumes ovules and achenes of <i>Eriophyllum lanatum</i> and other Asteraceae in southern California, presumably including <i>E. confertiflorum</i> (Goeden et al. 1994).
<b>Palatability, attractiveness to animals; response to grazing</b>	
<b>Mycorrhizal?</b>	Plants are thought to be associated with vesicular mycorrhizal fungi (L. Legerton-Warburton, pers. com), and the closely related <i>E. lanatum</i> has been shown to be colonized (Ingham & Wilson 1999).
<b>ECOLOGICAL GENETICS</b>	
<b>Ploidy</b>	Ploidy is variable but generally consistent within a single population. The base chromosome number is n=8 (Mooring 1966); diploids and tetraploids can be equally frequent and hexaploid and octoploid populations have been reported (based on 130 populations, Mooring 1994, 2n = 16, 32, 48, 64).
<b>Plasticity</b>	Seeds are polymorphic: 25% from a given pool will germinate readily while the other 75% require stimulation (Keeley & Pizzorno 1986). Plants are seasonally dimorphic and produce two kinds of leaves through the growing season (Westman 1981).
<b>Geographic variation (morphological and physiological traits)</b>	Plants are highly variable across the species range. Height, size, leaf, and flower differences tend to be similar within locations and may be influenced by light and soil variability (Mooring 1994). There are geographic patterns in diploid vs. tetraploid chromosome number (Mooring 1994). In the south, diploid populations are associated with xeric sites in coastal sage scrub, chaparral, and woodland. North of Santa Barbara and the Tejon Pass, xeric diploid populations extend northward in the Interior Coast Ranges, but in Coast Live Oak and Blue Oak Woodlands, diploid populations tend to be associated with more mesic sites. Tetraploid populations are concentrated in northern California, where they are found in the more xeric sites. Mooring (1994) hypothesized that the formation of polyploids may be induced in harsh conditions and that this may explain the segregation of the two types in the north, but not in the south. Constance (1934) mentions the possibility of genetic races at higher elevations.
<b>Genetic variation and population structure</b>	
<b>Phenotypic or genotypic variation in interactions with other organisms</b>	

<b>Local adaptation</b>	This is a highly variable and widespread species in California that occurs over a large range of elevation and precipitation gradients. It is common for plants to develop adaptive genetic differences between populations along steep environmental gradients. The variation in ploidy that correlates with habitat suggests there may be other important genetic and adaptive differences among populations. However, the outcrossing nature of this plant and potentially high gene flow potential suggest that the scale of adaptive differences will be on the scale of ecological sections and possibly subsections when there are large steps in elevation among subsections. Studies are needed to examine the scale of genetic differentiation and how that may affect translocation within vs. among ecological sections.
<b>Translocation risks</b>	The observed drop in seed germination after experimental crosses among different varieties of <i>E. confertiflorum</i> and among geographically distant populations of diploid var. <i>confertiflorum</i> suggests that there is a risk to geographically distant translocations. The scale of the distances that are associated with losses in fitness have not been studied. It would be prudent to only move seeds within geographic regions of adaptation and to avoid mixing diploid and tetraploid populations in fields used for seed increase. In addition, avoid mixing <i>E. confertiflorum</i> and <i>E. lanatum</i> in the same plant palette for restoration except when planting in specific locations where they occur together naturally
<b>SEEDS</b>	Rancho Santa Ana Botanic Garden Seed Program, seed image: <a href="http://www.hazmac.biz/050314/050314EriophyllumConfertiflorum.html">http://www.hazmac.biz/050314/050314EriophyllumConfertiflorum.html</a>
<b>General</b>	Standards for minimum purity and germination have been supplied by various seed companies: 30% purity, 60% germination (S&S Seeds, pers. com.). For <i>Eriophyllum</i> taxa in CA, seed germination ranges from 50-95% (Mooring 1975). Achenes (one seeded fruit) are small (2-3 mm long). Each achene has a pappus of short (< 1mm long) paleae that vary in size (Munz & Keck 1968). Seeds accumulate in soil seedbank. In chaparral in San Diego Co., Zammit & Zedler (1994) found that seeds in the soil seed bank did not respond significantly to fire treatments and that soil seed densities increased significantly with crown cover of perennial shrubs.
<b>Seed longevity</b>	<i>Eriophyllum</i> taxa in CA, seeds stored at room temperature have germinated after 8 years (Mooring 1975). Unlike many Asteraceae, seeds are generally long-lived in soil seed banks of this fire-follower.
<b>Seed dormancy</b>	Up to 25% of seeds will germinate readily without pretreatment (Mirov & Kraebel 1939, Keeley et al. 1985, Keeley & Pizzorno 1986, Emery 1988). The other 75% require dormancy breaking treatment. Light stimulates germination, but is not necessary when seeds are exposed to extracts from charred wood, which can increase germination to over 80% (Keeley & Pizzorno 1986). Direct heating of seeds does not increase germination (Christensen & Muller 1975, Keeley & Keeley 1982), but application of charred wood extract (i.e., charate made by running water through powdered, charred <i>Adenostoma fasciculatum</i> stems) significantly increases germination. When heat (120°C five minutes) is combined with charate, there is a small synergistic effect (Keeley & Keeley 1982, Keely et al. 1985). Heated lignin and cellulose or tannic acid with pH of 7 also enhanced germination. Keeley and Nitzberg (1984) showed that simply heating wood to over 175°C produces water soluble compounds that significantly stimulate germination.
<b>Seed maturation</b>	Timing of seed maturity depends on location and the pattern of rainfall. Mirov and Kraebel (1939) report May-September for seed collection.
<b>Seed collecting</b>	When the phyllaries are dry, and achenes are ripe, the whole top of the inflorescence can be harvested with clippers and placed in open container or breathable bag (e.g., cloth, paper).
<b>Seed processing</b>	For seed banking, Wall and MacDonald (2009) recommend rubbing the dry flower heads over a medium screen, sifting through #18 and #25 sieves, and then using an Oregon Seed Blower unit at speed 1.5 to clean. The Bend Seed Extractory used a Hoffman Hand Debearder to open the capsule-like heads and then air screened to clean to 96% purity and 96% filled seeds (see NPNPP protocol link below). For direct seeding and bulk seed storage, the dry heads can be placed in a blender with tape over the blades to break up heads, then sieved to separate large chaff from the tiny seeds (Montalvo, pers obs.).
<b>Seed storage</b>	Seeds can be stored at room temperature several years (Mooring 1975). Cold, dry storage is likely to increase longevity in storage.
<b>Seed germination</b>	Seeds take about 11 days to germinate after moistened (Mirov & Kraebel 1939). Germination response varies with seed source and test conditions (Keeley & Pizzorno 1986), but about a quarter of seeds tend to germinate in light at 20°C with no treatment (Atwater 1980).
<b>Seeds/lb</b>	2,750,000 seeds/lb (S&S Seeds 2009) 2,600,000 seeds/lb (Mirov & Kraebel 1939)
<b>Planting</b>	The tiny achenes are planted shallowly or on the surface. They do well when hydrosseeded or planted with shallow sowing methods.

<b>Seed increase activities or potential</b>	Rolle (2004) reported success with four fall plantings of the related <i>Eriophyllum lanatum</i> . Good seed crops were produced in the first year and for several years.
<b>Recommendations for seed production</b>	Data on outbreeding depression, variation in ploidy within species, and lowered pollen viability of among species (inter-specific) hybrids suggest that there would be undesirable consequences if populations from widely different habitats or locations were mixed for restoration or seed increase purposes. Due to ease of hybridization, fields of <i>E. confertiflorum</i> and <i>E. lanatum</i> (or different varieties of the same species) should not be placed in close proximity. These species are highly outcrossing and some of their pollinators can forage over long distances. We recommend that isolation distances between seed increase fields be large enough to discourage natural hybridization between fields of different species and different populations of the same species. This may require placing fields on different farms. Seeding with mixtures of genetically and geographically disparate populations is discouraged, especially if the planting site is on the drier end of the species range.
<b>USES</b>	
<b>Revegetation and erosion control</b>	This species germinates in the winter, grows rapidly, and is good for erosion control on slopes. Plants are frequently recommended and used successfully in seeding mixtures to stabilize dry slopes from northwestern California to southern California (Newton & Claassen 2003, A. Montalvo pers. obs).
<b>Habitat restoration</b>	Used widely in restoration seeding of coastal sage scrub vegetation (Montalvo pers. obs.) and other habitats (Newton & Claassen 2003). Seeds are easily collected from wildlands and are frequently available from native seed companies. This species is a good choice for planting on bare, sterile soils and for kick-starting the early successional stages of a restored shrubland community.
<b>Horticulture or agriculture</b>	Horticulture: The masses of golden-yellow flowers make it an attractive ornamental for home gardens and ecological landscaping and can provide color into the summer, especially in rock gardens and along borders (Schmidt 1980, Keator 1990, 1994, Clarke & Toogood 1994). Agriculture/Restoration: Plants can be easily cultivated from seed in well-drained soil and do best in full sun for at least part of the day (Schmidt 1980). Container plants are easy to produce from seeds, and plants transplant well (Montalvo pers. obs.).
<b>Wildlife value</b>	Flowers offer a food resource for many nectar-foraging insects (Moldenke 1976). In a study in the San Gabriel Mountains, <i>E. confertiflorum</i> made up 5.6% of the plants browsed by mountain sheep, as determined in fall fecal samples (Perry et al. 1987).
<b>Plant material releases by NRCS and cooperators</b>	None.
<b>Ethnobotanical</b>	Yaeger (1941) reported that southern California tribes used the white wooly hairs as a remedy for rheumatism by rolling the hairs into a ball, placing it on the affected part, and lighting it on fire. No verification of this has been found. The Cahuilla parched seeds and ground them into a flour (NAE).
<b>ACKNOWLEDGMENTS</b>	
Partial funding for production of this plant profile was provided by the U.S. Department of Agriculture, Forest Service, Pacific Southwest Region Native Plant Materials Program.	
<b>CITATION</b>	
Montalvo, A. M., and J. L. Beyers. 2010. Plant Profile for <i>Eriophyllum confertiflorum</i> var. <i>confertiflorum</i> . Native Plant Recommendations for Southern California Ecoregions. Riverside-Corona Resource Conservation District and U.S. Department of Agriculture, Forest Service, Pacific Southwest Research Station, Riverside, CA. Online: <a href="https://www.rcrec.org/plant-profiles">https://www.rcrec.org/plant-profiles</a> .	
<b>LINKS TO REVIEWED DATABASES &amp; PLANT PROFILES</b> (updated 3/23/2020)	
<b>Fire Effects Information System (FEIS)</b>	No matches: <a href="https://www.feis-crs.org/feis/">https://www.feis-crs.org/feis/</a>
<b>Jepson Flora, Herbarium (JepsonOnline)</b>	<a href="https://ucjeps.berkeley.edu/cgi-bin/get_cpn.pl?2814">https://ucjeps.berkeley.edu/cgi-bin/get_cpn.pl?2814</a>
<b>Jepson Flora, Herbarium, 2nd Edition (JepsonOnline 2nd Ed.)</b>	<a href="https://ucjeps.berkeley.edu/eflora/eflora_display.php?tid=2814">https://ucjeps.berkeley.edu/eflora/eflora_display.php?tid=2814</a>
<b>USDA PLANTS</b>	<a href="https://plants.usda.gov/core/profile?symbol=ERCOC12">https://plants.usda.gov/core/profile?symbol=ERCOC12</a>
<b>Native Plant Network Propagation Protocol Database (NPNPP)</b>	<a href="https://nnp.rngr.net/propagation/protocols">https://nnp.rngr.net/propagation/protocols</a>

Native Seed Network	<a href="https://nativeseednetwork.org/">https://nativeseednetwork.org/</a>
GRIN	no matches: <a href="https://npgsweb.ars-grin.gov/gringlobal/search.aspx?">https://npgsweb.ars-grin.gov/gringlobal/search.aspx?</a>
Flora of North America (FNA) (online version)	<a href="http://www.efloras.org/florataxon.aspx?flora_id=1&amp;taxon_id=250066707">http://www.efloras.org/florataxon.aspx?flora_id=1&amp;taxon_id=250066707</a>
Native American Ethnobotany (NAE)	<a href="http://naeb.brit.org/uses/search/?string=Eriophyllum+confertiflorum">http://naeb.brit.org/uses/search/?string=Eriophyllum+confertiflorum</a>
Calflora	<a href="https://www.calflora.org/">https://www.calflora.org/</a>
Rancho Santa Ana Botanic Garden Seed Program, seed photos	<a href="http://www.hazmac.biz/rsabghome.html">http://www.hazmac.biz/rsabghome.html</a>
<b>USE OF IMAGE WITH COPYRIGHT</b>	Matson photo copyright: This photo may not be used except with express written permission from Steve Matson. To obtain permission for personal, academic, commercial, or other uses, or to inquire about high resolution images, prints, fees, or licensing, or if you have other questions, contact Steve Matson <a href="mailto:ssmat@sbcbglobal.net">ssmat@sbcbglobal.net</a> . (Replace the [AT] with the @ symbol before sending an email.)

## Bibliography for *Eriophyllum confertiflorum* var. *confertiflorum*

- Atwater, B. R. 1980. Germination, dormancy and morphology of the seeds of herbaceous ornamental plants. *Seed Science and Technology* **8**:523-573.
- Christensen, N. L., and C. H. Muller. 1975. Relative importance of factors controlling germination and seedling survival in *Adenostoma* chaparral. *The American Midland Naturalist* **93**:71-78.
- Clarke, G., and A. Toogood. 1994. *The Complete Book of Plant Propagation*. Ward Lock Limited, London, England.
- Clarke, O. F., D. Svehla, G. Ballmer, and A. Montalvo. 2007. *Flora of the Santa Ana River and Environs with References to World Botany*. Heyday Books, Berkeley, CA.
- Cleland, D. T. F., J. A. Freeouf, J. E. Keys, G.J. Nowacki, C. A. Carpenter, W. H. McNab. 2007. Ecological Subregions: Sections and Subsections for the conterminous United States. Gen. Tech. Report WO-76D [Map on CD-ROM] (A.M. Sloan, cartographer). U.S. Department of Agriculture, Forest Service, Washington, DC.
- Constance, L. 1934. A preliminary revision of the perennial species of *Eriophyllum*. *Proceedings of the National Academy of Sciences* **20**:409-413.
- Constance, L. 1937. A systematic study of the genus *Eriophyllum* Lag. *University of California Publications in Botany* **18**:69-136.
- Emery, D. E. 1988. *Seed Propagation of Native California Plants*. Santa Barbara Botanical Garden, Santa Barbara, CA. 23 p.
- FNA. 2010. Editorial Committee. 1993+. *Flora of North America North of Mexico*. 10+ volumes. New York and Oxford. Available online: [http://beta.floranorthamerica.org/Published\\_Volumes](http://beta.floranorthamerica.org/Published_Volumes). Accessed September 29, 2010.
- Goeden, R. D., D. H. Headrick, and J. A. Teerink. 1994. Life history and description of immature stages of *Paroxyna genalis* (Thomson) (Diptera: Tephritidae) on native Asteraceae in southern California. *Proceedings of the Entomological Society of Washington* **96**:612-629.
- Goudey, C. B., and D. W. Smith. 1994. *Ecological units of California: Subsections (map)*. San Francisco, CA: U.S. Department of Agriculture, Forest Service. Pacific Southwest Region. Scale 1:1,000,000; colored.
- Hickman, J. C. 1993. *The Jepson Manual: Higher Plants of California*. University of California Press, Berkeley, CA.
- Ingham, E. R., and M. V. Wilson. 1999. The mycorrhizal colonization of six wetland plant species at sites differing in land use history. *Mycorrhiza* **9**:233-235.
- JepsonOnline. 2010. *The Jepson Manual Higher Plants of California; Online Version with 2nd edition notes*. <http://ucjeps.berkeley.edu/jepman.html>. (updated to Jepson eFlora: <https://ucjeps.berkeley.edu/eflora/>)
- JepsonOnline2ndEd. 2010. J. S. Mooring and D. E. Johnson. 2011, in press. *Eriophyllum*. in B. G. Baldwin and others, editors. *The Jepson Manual: Vascular Plants of California, 2nd Edition*. University of California Press., Berkeley, CA. Online: <http://ucjeps.berkeley.edu/jepman.html>. Accessed: September 30, 2010. (Updated to: <https://ucjeps.berkeley.edu/eflora/>)
- Keator, G. 1990. *Complete Garden Guide to the Native Perennials of California*. Chronicle Books, San Francisco, CA.
- Keator, G. 1994. *Complete Garden Guide to the Native Shrubs of California*. Chronicle Books, San Francisco, CA.
- Keeley, J. E. 1991. Seed germination and life history syndromes in the California chaparral. *The Botanical Review* **57**:81-116.

- Keeley, S. C., and J. E. Keeley. 1982. The role of allelopathy, heat, and charred wood in the germination of chaparral herbs. Pages 128-134 in C. E. Conrad and W. C. Oechel, editors. Proceedings of the Symposium on Dynamics and Management of Mediterranean-Type Ecosystems. General Technical Report PSW-58. U.S. Department of Agriculture, Forest Service, Pacific Southwest Forest and Range Experiment Station, Berkeley, CA.
- Keeley, S. C., J. E. Keeley, S. M. Hutchinson, and A. W. Johnson. 1981. Postfire succession of the herbaceous flora in southern California chaparral. *Ecology* **62**:1608-1621.
- Keeley, J. E., B. A. Morton, A. Pedrosa, and P. Trotter. 1985. Role of allelopathy, heat and charred wood in the germination of chaparral herbs and suffrutescents. *Journal of Ecology* **73**:445-458.
- Keeley, J. E., and M. E. Nitzberg. 1984. Role of charred wood in the germination of the chaparral herbs *Emmenanthe penduliflora* (Hydrophyllaceae) and *Eriophyllum confertiflorum* (Asteraceae). *Madroño* **31**:208-218.
- Keeley, S. C., and M. Pizzorno. 1986. Charred wood stimulated germination of two fire-following herbs of the California chaparral and the roll of hemicellulose. *American Journal of Botany* **73**:1289-1297.
- Kirkpatrick, J. B., and C. F. Hutchinson. 1980. The environmental relationships of California coastal sage scrub and some of its component communities and species. *Journal of Biogeography* **7**:23-28.
- Kruckeberg, A. R. 1984. California serpentine: Flora, vegetation, geology, soils, and management problems. University of California Publications in Botany **78**:1-180.
- Mirov, N. T., and C. J. Kraebel. 1939. Collecting and handling seeds of wild plants. Civilian Conservation Corps, Forestry Publication No. 5. 42 p.
- Moldenke, A. R. 1976. California pollination ecology and vegetation types. *Phytologia* **34**:305-361.
- Mooring, J. S. 1966. Chromosome numbers in the *Eriophyllum lanatum* (Compositae, Helenieae) complex. *Madroño* **18**:236-239.
- Mooring, J. S. 1975. A cytogeographic study of *Eriophyllum lanatum* (Compositae, Helenieae). *American Journal of Botany* **62**:1027-1037.
- Mooring, J. S. 1994. A cytogenetic study of *Eriophyllum confertiflorum* (Compositae, Helenieae). *American Journal of Botany* **81**:919-926.
- Mooring, J. S. 1997. A new base chromosome number and phylogeny for *Eriophyllum* (Asteraceae, Helenieae). *Madroño* **44**:364-373.
- Mooring, J. S. 2001. Barriers to interbreeding in the *Eriophyllum lanatum* (Asteraceae, Helenieae) species complex. *American Journal of Botany* **88**:285-312.
- Munz, P. A. 1974. A Flora of Southern California. University of California Press, Berkeley, CA.
- Munz, P. A., and D. D. Keck. 1968. A California Flora with Supplement. University of California Press, Berkeley, CA.
- Newton, G. A., and V. Claassen. 2003. Rehabilitation of Disturbed Lands in California: A Manual for Decision-Making. California Department of Conservation, California Geological Survey.
- O'Brien, B., B. Landis, and E. Mackey. 2006. Care & Maintenance of Southern California Native Plant Gardens. Rancho Santa Ana Botanic Garden, Claremont, CA.
- Perry, B. 1992. Landscape plants for western regions: An illustrated guide to plants for water conservation. Land Design Publishing, Claremont, CA.
- Perry, W. M., J. W. Dole, and S. A. Holl. 1987. Analysis of the diets of mountain sheep from the San Gabriel Mountains, California. *California Fish and Game* **73**:156-162.
- Rolle, W. 2004. Average per-acre seed yields and lifespans of native seed crops grown at J. Herbert Stone Forest Service Nursery. Summary for all native seed crops from 1992 through 2002. USDA Forest Service.



- S&S Seeds. 2009. S & S Seeds Seed Selection Guide:  
[http://www.ssseeds.com/media/218482/ssseeds\\_guide.pdf](http://www.ssseeds.com/media/218482/ssseeds_guide.pdf).
- Sawyer, J. O., T. Keeler-Wolf, and J. M. Evens. 2009. A Manual of California Vegetation, 2nd edition. California Native Plant Society Press, Sacramento, CA.
- Schmidt, M. G. 1980. Growing California Native Plants. University of California Press, Los Angeles.
- USDA PLANTS. 2010. The PLANTS Database (<https://plants.usda.gov/java/>). National Data Center, Baton Rouge, LA 70874-4490 USA.
- Wall, M., and J. Macdonald. 2009. Processing Seeds of California Native Plants for Conservation, Storage, and Restoration. Rancho Santa Ana Botanic Garden Seed Program, Claremont, CA. Available online: <http://www.hazmac.biz/seedhome.html>.
- Westman, W. E. 1981. Seasonal dimorphism of foliage in Californian coastal sage scrub. *Oecologia* **51**:385-388.
- Yaeger, E. C. 1941. Desert Wildflowers. Stanford University Press, Stanford, CA.
- Zammit, C., and P. H. Zedler. 1994. Organisation of the soil bank in mixed chaparral. *Vegetatio* **111**:1-16.