Acta Parasitologica, 1997, 42(3), 149–157 ISSN 1230-2821 Copyright © 1997 W. Stefański Institute of Parasitology, PAS

Digenetic trematode parasites of the flatfishes (Pleuronectiformes) of the Kerala coast, India

A. Bijukumar

Department of Aquatic Biology and Fisheries, University of Kerala, Trivandrum-695 007, Kerala, India

Abstract. Eleven species of digenetic trematodes, including a new one, are reported from 21 species of pleuronectid flatfishes (representing 5 families) inhabiting the Arabian Sea, off the Kerala coast, India. The parasites are *Prosorhynchus* sp. metacercaria; *Stephanostomum* sp. metacercaria; *Decemtestis mehrai* Srivastava, 1936; *Macvicaria cynoglossi* (Madhavi, 1975); *M. longicaudus* (Hafeezullah, 1971); *M. jagannathi* (Gupta et Singh, 1985); *Pseudopecoelus scomberi*

(Hafeezullah, 1971); Lepocreadioides orientalis Park, 1939; didymozoid larva (Type 1); didymozoid larva (Type 2) and Monorcheides hafeezullahi sp. n. There are 36 new host records. A list of digenetic trematodes recorded so far from the flatfishes of India is also given. The adult trematodes collected except L. orientalis showed an endemic distribution to seas around Indian peninsula. The host specificity of parasites is discussed.

Introduction

Pleuronectid flatfishes constitute a substantial fishery in the southwestern (Kerala) coast of India. Considering the great diversity of 91 species of flatfishes inhabiting Indian waters (Menon 1977), there are several reports regarding the digenetic trematodes of the flatfishes of India. Digenetic trematodes reported so far from the flatfishes inhabiting Indian waters include: Neidhartia microrhyncha Chauhan, 1943 from Psettodes erumei; Prosorhynchus arabiana Srivastava, 1938 from Synaptura orientalis; Pleorchis sciaenae Yamaguti, 1938 from Pseudorhombus diacanthus; Lepocreadioides orientalis Park, 1939 from Cynoglossus spp.; L. indicus Srivastava, 1941 from Cynoglossus spp.; Monascus orientalis (Srivastava, 1941) Yamaguti, 1958 from S. orientalis; Opegaster ditrematis Yamaguti, 1942 from P. erumei, Pseudorhombus triocellatus and P. micrognathus; Rhipidocotyle ghanensis Fischthal et Thomas, 1968 from P. erumei; Lepocreadioides srivastavai Gupta et Mehrotra, 1970 from Cynoglossus

cynoglossus and C. lingua; Rhipidocotyle karthai Hafeezullah et Siddigi, 1970 from P. erumei; Plagioporus longicaudus Hafeezullah, 1971 from Cynoglossus lida; Opisthodiplomonorchis elongatus Madhavi, 1974 from P. erumei; Plagioporus cynoglossi Madhavi, 1975 from C. lida; Rhipidocotyle indicus Gupta et Ahmad, 1976 from C. lida; Pleorchis psettodesai Gupta et Gupta, 1976 from P. erumei; Lecithocladium keralense Gupta et Gupta, 1978 from P. erumei; L. psettodi Gupta et Puri, 1982 from P. erumei; Lecithaster bombayensis Ahmad, 1983 from P. diacanthus; Lepocreadioides thaparai Gupta et Govind, 1983 from C. lida; Neonotoporus overstreeti Ahmad et Dhar, 1987 from P. erumei; Podocotyloides sp. from Cynoglossus oligolepis; Allodecemtestis skrjabini Ahmad, 1988 from C. bilineatus and Plagioporus microlepisi Salman et Srivastava, 1990 from C. microlepis.

The aim of the present paper is to provide information on 11 species of digenetic trematodes collected from 21 species of flatfishes inhabiting the

Arabian Sea, off the coast of Kerala. One species, *Monorcheides hafeezullahi* is new to science. There are 36 new host records.

Materials and methods

Twenty-one species of flatfishes belonging to 5 families were collected from trawlers from the Neendakara (08°30'N; 76°53.3'E), Vizhinjam (08°22'N; 76°85'E) and Veli (08°28'N; 76°57'E) landing centres of the Kerala coast from March 1992 to April 1994.

The list of fishes examined and total number of specimens in each species are given in Table I.

Table I. List of flatfishes examined for digenetic trematodes from the Kerala coast, India

Hosts	No. of fish examined
Family Bothidae	
Bothus myriaster (Temminck et Schlegel)	45
Crossorhombus azureus (Alcock)	62
Engyprosopon grandisquamis (Temminck et Schlegel)	55
Laeops nigrescens Lloyd	30
Pseudorhombus arsius (Hamilton)	45
P. elevatus Ogilby	85
P. javanicus (Bleeker)	36
P. triocellatus (Schneider)	60
Family Cynoglossidae	
Cynoglossus arel (Schneider)	254
C. dubius Day	121
C. lida (Bleeker)	118
C. macrostomus Norman	259
C. puncticeps (Richardson)	55
Family Pleuronectidae	
Samaris cristatus Gray	48
Family Psettodidae	
Psettodes erumei (Schneider)	55
Family Soleidae	
Aesopia cornuta Kaup	48
Euryglossa orientalis (Schneider)	65
Solea ovata Richardson	48
Synaptura commersoniana (Lacepede)	52
Zebrias altipinnis (Alcock)	36
Z. synapturoides (Jenkins)	61

The fishes collected frozen from the trawlers were examined both externally and internally for digenetic trematodes. The digenetic trematodes were fixed in 10% neutral buffered formalin, under coverglass pressure, dehydrated in graded concentrations of alcohol and stained in Gower's carmine. The figures were drawn using camera lucida attachment.

The specimens were deposited in the National Museum, Zoological Collections, Zoological Survey of India, Calcutta (ZSINZC), under the accession numbers indicated. Measurements are in micrometers (μ m) unless otherwise indicated. The terms prevalence (%) and mean intensity (given in brackets after the host

name) follow those of Margolis et al. (1982). Asterisk mark before the hosts indicate new host records.

Results

Bucephalidae Poche, 1907

Prosorhynchus sp. metacercaria (Fig. 1)

Hosts: *Crossorhombus azureus (4.8%; 1.7), *Engyprosopon grandisquamis (65.5%; 2.0), Laeops nigrescens (10%; 4.7), *Pseudorhombus elevatus (35.3%; 3.5), *P. javanicus (19.4%; 3.9) (Bothidae); *Cynoglossus arel (21.3%; 1.2), *C. lida (47.5%; 4.6) (Cynoglossidae); *Psettodes erumei (1.8%; 1.0) (Psettodidae); *Aesopia cornuta (43.8%; 2.0), Zebrias altipinnis (16.7%; 1.5) and *Z. synapturoides (31.1%; 1.6) (Soleidae).

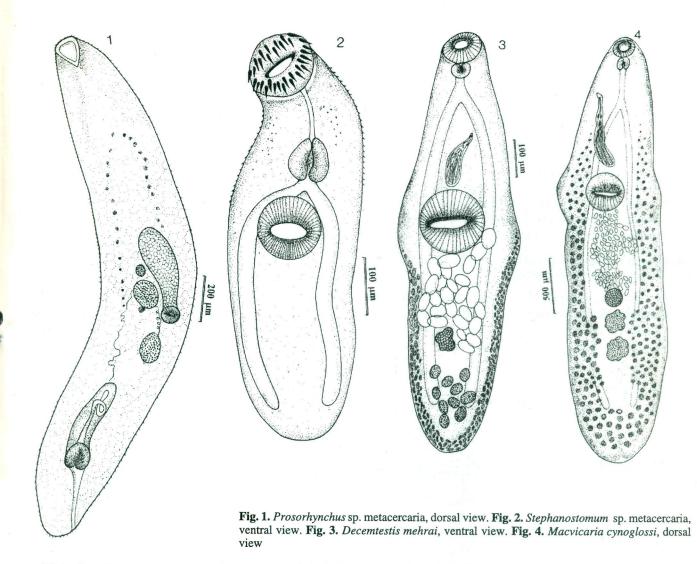
Location: Intestine and muscle.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8276/1, w 8277/1 and w 8278/1.

Description (based on 10 specimens). Body long, cylindrical, 1.33–2.90 × 0.27–0.37mm, rounded at both ends, maximum width in the region of the pharynx. Anterior end with a conical rhynchus, 117–200 × 100–117. Tegument spiny. Pharynx muscular, 57–86 in diameter, situated in the posterior half of the body. Intestinal caecum large, 333–629 × 0.114–0.250, saclike, directed anteriorly and filled with fat globules. Oesophagus, 220–270 long. Rudiments of testes and ovary present close to the intestinal caecum. Cirrus sac short, 243–856 × 57–144. Vitellaria small, rounded, follicular, lateral in anterior half of body, apparently coalescing in midbody anteriorly.

Remarks. Members of the genus Prosorhynchus have been reported from several species of marine fishes of India. P. arabianus and P. manteri were described by Srivastava (1938) from the intestine of marine fishes. Hafeezullah and Siddiqi (1970) reported on P. tsengi Tsin, 1933, P. epinepheli Yamaguti, 1939, P. atlanticus Manter, 1940 and P. chorinemi Yamaguti, 1952 infecting the marine fishes of the east and west coasts of India. Madhavi (1976) described P. atlanticus, P. chorinemi, P. pacificus and P. indicum from the marine fishes off Andhra coast, Bay of Bengal. Recently Madhavi et al. (1994) reported metacercaria of Prosorhynchus sp. from mullets and its cercaria from the bivalve, Modiolus undulatus from Chilka lake. Bilgees (1976) described P. erumenis from the Indian turbot, Psettodes erumei off the Karachi coast.

The present metacercariae occurred free in the intestine or muscles as well as encapsulated in the muscles of the host body. Heavy infection of the muscles could be detected externally by the presence of dark-reddish or nearly black spots amidst a diffuse



reddish discolouration on the white blind side of the fish. More parasites were located in the blind side of the fish.

Stephanostomum sp. metacercaria (Fig. 2)

Hosts: *Cynoglossus arel (57.9%; 13.6), *C. dubius (47.1%; 5.5), *C. lida (54.2%; 4.4) and *C. puncticeps (52.7%; 6.3) (Cynoglossidae).

Location: Fins.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8281/1.

Description (based on 10 specimens). Body robust, $367-757 \times 100-300$, slightly narrowing towards the anterior tip. Tegument spined; anterior half of the body with 20-25 rows of 8-13 long spines. Oral sucker conspicuous, $43-75\times 84-92$; 33 perioral spines, $17-25\times 2-3$, arranged in two alternating rows. Acetabulum large and rounded, situated more or less in the middle of the body, 72-100 in diameter. Prepharynx, 91-113 long. Pharynx subspherical, nearly anterior to ventral sucker. Oesophagus very short, bifurcating into two long caeca which extend nearly up to the posterior

extremity of the body. Gonads and vitellaria not well developed. Gonads, vitellaria and cirrus sac indistinct.

Remarks. Fifteen species of *Stephanostomum* have been reported from various marine fishes of India (Srivastava 1938, Hafeezullah 1971a, Madhavi 1976, Gupta and Ahmad 1979). But this is the first time that *Stephanostomum* sp. is described from Indian flatfishes. *Stephanostomum* sp. occurred in flatfishes in capsules, attached to the fins. The thick-walled capsules (277–320 × 133–200) were spherical, transparent structures.

Opecoelidae Ozaki, 1925

Decemtestis mehrai Srivastava, 1936 (Fig. 3)

Host: *Cynoglossus lida (0.9%; 1.0) (Cynoglossidae). Location: Intestine.

Locality: Vizhinjam coast, Kerala.

Specimens deposited: ZSINZC No. w 8258/1 and w 8259/1.

Measurements. Body, 1.3×0.45 mm; oral sucker, 125 in diameter; ventral sucker, 250 in diameter; sucker

ratio, 1:2; distance between suckers, 425; pharynx, 150 \times 175; cirrus sac, 250 \times 100; testes, 100 \times 100; ovary, 175 \times 175; eggs, 150–175 \times 100.

Remarks. Decemtestis mehrai was first described by Srivastava (1936) from Sillago sihama. Later Gupta and Mehrotra (1970) collected this species from two marine fishes (Ariodes dussumieri and Triacanthus brevirostris) off Kerala coast, south India. Except for the slight differences in morphometry, the present specimens closely resemble D. mehrai. The present specimens are smaller (667×233) than those described by Srivastava ($2.2-2.7 \times 0.73-0.94$ mm) and Gupta and Mehrotra ($1.77-1.94 \times 0.682-0.686$ mm). Cynoglossus lida is a new host for D. mehrai.

Macvicaria cynoglossi (Madhavi, 1975) (Fig. 4) Syn. Plagioporus cynoglossi Madhavi, 1975

Hosts: *Cynoglossus arel (35%; 4.4), C. lida (5.1%; 1.5) (Cynoglossidae); *Aesopia cornuta (77.1%; 1.6),

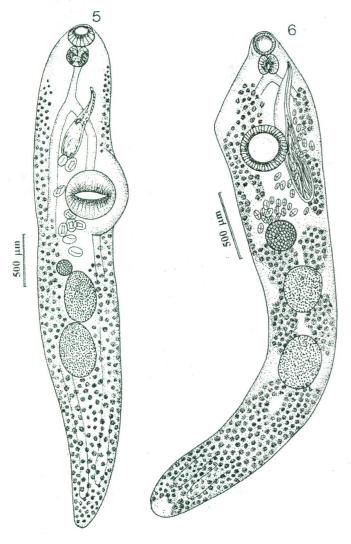


Fig. 5. Macvicaria longicaudus, dorsal view. Fig. 6. Macvicaria jagannathi, ventral view

*Zebrias altipinnis (63.9%;1.4) and Z. synapturoides (68.9%; 3.1) (Soleidae).

Location: Intestine.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8260/1, w 8261/1 and w 8262/1.

Measurements (based on 10 specimens). Body, $1.24-3.96 \times 0.36-0.84$ mm; oral sucker, 117-133 in diameter; ventral sucker, 217-360 in diameter; sucker ratio, 1:1.85-2.88; distance between suckers, 450-950; pharynx, 67-100; anterior testis, 93-107; posterior testis, 107-133; ovary, 80-93; cirrus sac, $480-600 \times 77-100$; eggs, $58-100 \times 20-38$.

Remarks. The specimens agree well with *M. cynoglossi* as described by Madhavi (1975) in all the details. The species was first described from *Cynoglossus lida* off Waltair coast, Bay of Bengal, India, as *Plagioporus cynoglossi*. Gibson and Bray (1982) reorganised the genus *Plagioporus* Stafford, 1904 and erected the new genus *Macvicaria* for accomodating some of the marine *Plagioporus* species. Earlier, *P. cynoglossi* was not placed in *Macvicaria* by Bray (1985) because of the shape of the seminal vesicle (elongated or saccular) and the lobed testis. But later, in 1990, Bray proposed a new combination, *M. cynoglossi*, as the current concept of this genus includes elongated and saccular seminal vesicle and lobed testes.

Macvicaria longicaudus Hafeezullah, 1971 comb. n. (Fig. 5)

Syn. Plagioporus longicaudus Hafeezullah, 1971

Hosts: *Cynoglossus dubius (58.7%; 7.7), *C. macrostomus (23.2%; 2.7) (Cynoglossidae) and *Synaptura commersoniana (51.9%; 1.4) (Soleidae).

Location: Intestine.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8263/1, w 8264/1 and w 8265/1.

Measurements (based on 10 specimens). Body, $3.48-4.6 \times 0.6-0.84$ mm; oral sucker, $133-217 \times 167-217$; ventral sucker, 267-333 in diameter; sucker ratio, 1:1.67-2.00; distance between suckers, 750-950; pharynx, $83-200 \times 117-217$; testes, $167-383 \times 167-300$; ovary, 100-133; cirrus sac, $500-583 \times 67-133$; eggs, $57-72 \times 34-49$.

Remarks. The present material closely resembles *P. longicaudus* described by Hafeezullah (1971b) from *C. lida* from the Gulf of Mannar, Tuticorin, India. Gibson and Bray (1982) erected a new genus, *Macvicaria*, incorporating many members of the genus *Plagioporus* and considered that the latter is restricted to freshwater fishes. Considering the close proximity of the present species to *Macvicaria*, it is transferred into this genus. *C. dubius*, *C. macrostomus* and *S. commersoniana* are new host records for *M. longicaudus*.

Macvicaria jagannathi Gupta et Singh, 1985 comb. n. (Fig. 6)

Syn. Plagioporus jagannathi Gupta et Singh, 1985

Hosts: *Bothus myriaster (53.3%; 2.8), *Engyprosopon grandisquamis (30.9%; 1.9), *Pseudorhombus elevatus (34.1%; 1.8) and *P. javanicus (33.3%; 2.2) (Bothidae).

Location: Intestine.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8266/1, w 8267/1 and w 8268/1.

Measurements (based on 10 specimens). Body, $1.175-3.25\times0.325-0.5$ mm; oral sucker, 80-130; ventral sucker, 150-250; sucker ratio, 1:1.85-2.02; distance between suckers, 150-250; pharynx, 60-100; anterior testis, $130-240\times120-180$; posterior testis, $150-250\times120-180$; ovary, 60-130; cirrus sac, $400-533\times60-80$; eggs, $60-80\times30-50$.

Remarks. Except for the slightly larger size, the present material resembles *P. jagannathi* as described

by Gupta and Singh (1985) from *Pseudodax moluc-canus* from Puri, Bay of Bengal, India. Based on the reorganization of the genus *Macvicaria* proposed by Gibson and Bray (1982) this species is transferred to the genus *Macvicaria* as *M. jagannathi*.

Pseudopecoelus scomberi (Hafeezullah, 1971) (Fig. 7) Syn. *Pseudopecoeloides scomberi* Hafeezullah, 1971

Host: *Engyprosopon grandisquamis (1.8%; 1.0) (Bothidae).

Location: Intestine.

Locality: Vizhinjam coast, Kerala.

Specimens deposited: ZSINZC No. w 8269/1.

Measurements. Body, 3.96×0.6 mm; oral sucker, 117×117 ; ventral sucker, 183×183 ; distance between suckers, 217; pharynx, 80×80 ; anterior testis, 120×120 ; posterior testis, 133×120 ; ovary, 120×173 ; eggs, $33-50 \times 17-33$.

Remarks. The three species of *Pseudopecoelus* reported from India are *P. scomberi* (Hafeezullah, 1971), *P. bilqeesae* Ahmad et Dhar, 1987, and *P.*

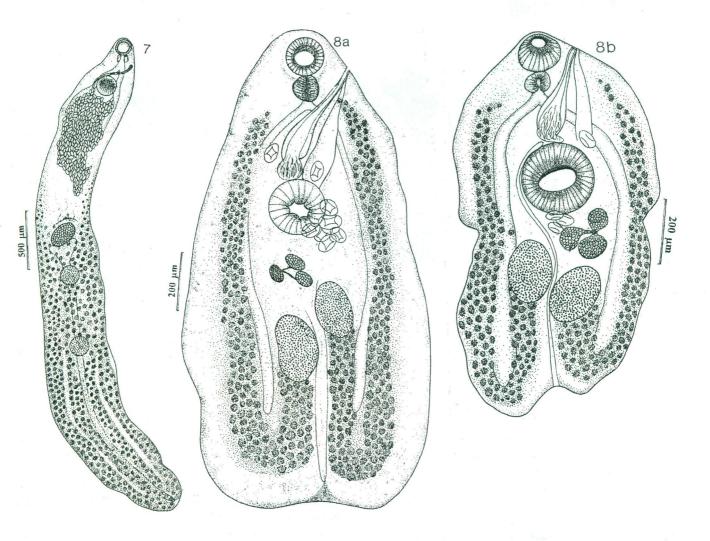


Fig. 7. Pseudopecoelus scomberi, dorsal view. Fig. Sa, b. Lepocreadioides orientalis, ventral view

dollfusi Ahmad et Dhar, 1987. The present material closely resembles *P. scomberi* described by Hafeezullah (1971b). Hafeezullah first described this species as *Pseudopecoeloides scomberi* from the carangid fish, *Scomberoides tol.* But later, Madhavi (1975) transferred it to the genus *Pseudopecoelus*. Engyprosopon grandisquamis is a new host for *P. scomberi*.

Lepocreadiidae (Odhner, 1905) Nicoll, 1935

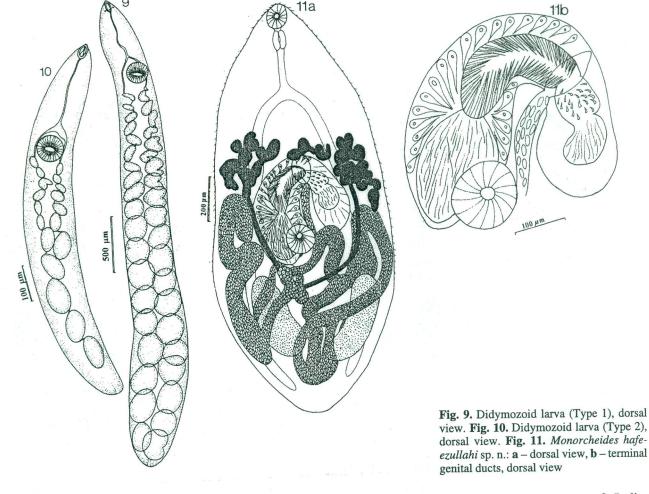
Lepocreadioides orientalis Park, 1939 (Fig. 8a, b) Syn. Lepocreadioides indicus Srivastava, 1941; Lepocreadioides srivastavai Gupta et Mehrotra, 1970; Bicaudum otolithi Bilqees, 1971; Lepocreadioides thaparai Gupta et Govind, 1984

Hosts: Cynoglossus arel (78.3%; 9.9), C. dubius (22.3%; 5.4), C. lida (21.2%; 1.9), C. macrostomus (40.2%;

Measurements (based on 10 specimens). Body, $1.099-2.966\times0.267-0.5$ mm; oral sucker, 67-107; ventral sucker, 133-267; sucker ratio, 1:1.45-1.65; distance between suckers, 267-333; pharynx, 33-67; testes, $200-300\times67-100$; ovary, 80-167; cirrus sac, 233×100 ; eggs, 67×33 .

Remarks. Lepocreadioides orientalis was originally described by Park (1939) from the flatfish, Areliscus joyneri from Simmi Island, North Tyosen, Korea.

Lepocreadioides indicus was reported by Srivastava (1941) from Platycephalus insidator from Puri and Karachi. Hafeezullah (1970) recorded this species from various flatfishes of the family Cynoglossidae (C. bilineatus, C. dubius, C. lida, C. lingua, C. macrolepidotus, C. puncticeps and C. sindensis), oc-



2.4), *C. puncticeps (21.8%; 1.4) (Cynoglossidae); *Aesopia cornuta (12.5%; 1.8), *Zebrias altipinnis (19.4%; 1.0) and *Z. synapturoides (11.5%; 1.1) (Soleidae).

Location: Stomach, intestine.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8270/1, w 8271/1, w 8272/1 and w 8273/1.

curring along the east and west coasts of India. Madhavi (1972) also recorded this species from C. lida off the Waltair coast, India. Gupta and Mehrotra (1970) recorded L. srivastavai from Cynoglossus cynoglossus and C. lingua off the Kerala coast. Gupta and Govind (1983) recorded L. thaparai from the fish C. lida from Puri coast, Orissa, India.

Recent investigations by Hafeezullah and Chakrabarti (1994) consider the genus Bicaudum Bilqees, 1971 to be a synonym of Lepocreadioides Yamaguti, 1936 and L. srivastavai, L. indicus and Bicaudum otolithi are synonyms of L. orientalis Park, 1939. Thus, so far L. orientalis has been reported from 9 species of the flatfish belonging to the genus Cynoglossus. Considering the high prevalence of L. orientalis in flatfishes of the family Cynoglossidae, it may be presumed that this parasite is the most common digenean inhabiting the intestine of Cynoglossus spp.

Two morphological variants of this species (one with foliate body with posterior median and lateral emarginations and the second with more or less oval body) were encountered during the present study (Fig. 8b and a, respectively).

Didymozoidae Poche, 1907

Didymozoid larva (Type 1) (Fig. 9)

Hosts: *Cynoglossus arel (12.2%; 1.8), *C. lida (24.6%; 2.1) and C. macrostomus (2.7%; 1.4) (Cynoglossidae).

Location: Intestine, muscle.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8279/1.

Description (based on 10 specimens). Body long, narrow, $1.633-3.199 \times 0.133-0.367$ mm, tapering at anterior end and rounded at posterior end, covered with smooth tegument. Oral sucker small, oval, 96-115 × 29-88; ventral sucker large, subspherical, 157-171 in diameter; distance between suckers, 314–386. Oesophagus sinuous and narrow, bifurcating just anterior to ventral sucker. Behind the ventral sucker, each caecum formed into 19-21 chambers; anterior chambers small, oval to irregular, posterior ones large and rounded. One pair of eye spots, crescentic, on either side of the oral sucker. Parenchymal gland cells fill the entire body.

Didymozoid larva (Type 2) (Fig. 10)

Hosts: *Pseudorhombus arsius (6.7%; 2.3), *P. elevatus (7.1%; 1.5), P. javanicus (2.8%; 1.0) and P. triocellatus (5%; 1.7) (Bothidae).

Location: Intestine.

Locality: Neendakara and Vizhinjam coasts, Kerala. Specimens deposited: ZSINZC No. w 8280/1.

Description (based on 5 specimens). Body robust but narrow, $1.0-1.484 \times 83-133$, smoothly tapering towards anterior and poterior ends. Tegument smooth. Oral sucker oval, $34-43 \times 17-23$, projecting slightly from the body at the anterior end. Ventral sucker large, subspherical, $80-86 \times 80-86$. Distance between suckers, 584-717. Oesophagus narrow, indistinct, bifurcating just in front of ventral sucker into unequal

branches; proximal parts of branches dilated. Right caecum divided into 11-13 chambers and left into 6-8 chambers. Eye spots absent.

The smaller size, lesser and unequal number of chambers of caeca, dilated proximal parts of caeca and absence of eye spots distinguish didymozoid larva Type 2 from Type 1.

Remarks. Køie and Lester (1985) are of the opinion that "with the present knowledge, it is impossible to classify the larval didymozoids to generic or taxa levels and the larval didymozoids should not be given Latin names to avoid double taxonomic system, one for the larval and other for the mature didymozoids".

Several workers have reported on the occurrence of didymozoid larva in marine fishes and suggested that fishes play a role of paratenic hosts in the transfer of didymozoid metacercariae indicating that crustaceans act as the second intermediate hosts to these parasites (Køie and Lester 1985). As the majority of flatfishes are benthophagous and since crustaceans form a major component of their food (Ramanathan et al. 1977, Radhamanyamma 1988), it is quite probably that these fishes act as a paratenic hosts for didymozoids.

In the present study two types of didymozoid larvae were collected from flatfishes; the first type occurred in three species of Cynoglossus and the second type in four species of Pseudorhombus, suggesting a comparatively high degree of host-specificity for the two types.

Though Type 1 larvae were collected both from the intestine and muscles of cynoglossid fishes, they were more abundant in the muscles. Didymozoid larvae with eye spots have been reported earlier from marine fishes of Australia by Køie and Lester (1985). According to these authors, the eye spots present in larval didymozoids may help them in their migration inside the final host, especially when the infection site of the parasite is close to the surface of the fish where some light may penetrate.

Monorchiidae Odhner, 1911

Monorcheides hafeezullahi sp. n. (Fig. 11a, b)

Host: Pseudorhombus triocellatus (33.3%; 1.7) (Bothidae).

Location: Pyloric caeca.

Locality: Vizhinjam coast, Kerala.

Specimens deposited: ZSINZC holotype No. w 8274/1;

paratype No. w 8275/1.

Description (based on 10 specimens). Body suboval, widest in hindbody, slightly narrowing anteriorly, $0.875-1.75 \times 0.625-0.8$ mm, somewhat opaque, reddish in colour. Tegument spined up to the level of ovary (about 2/3 distance from anterior tip). Eye spot pigment absent. Suckers poorly developed. Oral sucker rounded, subterminal, 120-133 in diameter. Ventral

sucker, 133-147, slightly postequtorial in position; sucker ratio, 1:1.11-1.18; distance between suckers, 867–900. Prepharynx short; pharynx oval, distinct, $50-83 \times 32-50$. Oesophagus, 167–200; caecal bifurcation nearer to oral sucker than to ventral sucker; caeca slender, extending nearly of posterior end of the body, terminating blindly. Testes elliptical, $325-375 \times 125-$ 150, symmetrically postequatorial, in posterior third of body. Genital pore median, preacetabular at midway between caecal bifurcation and ventral sucker. Cirrus sac large, $450-625 \times 175-225$, proximal end of it situated slightly posterior to ventral sucker, containing oval seminal vesicle, short pars prostatica surrounded by numerous prostatic cells and long, protrusible cirrus armed with long, needle-like spines. Genital atrium unarmed. Ovary small, $150-225 \times 75-125$, 2-4lobed, submedian to right, between the testes. Receptaculum seminis absent. Uterus voluminous, occupying all available space in postacetabular part of body, ultimately joining metraterm and metraterm sac at its distal part. Metraterm sac well differentiated, smaller than cirrus sac, divisible into two parts, posterior part muscular, thick-walled, unarmed and anterior part with triangular recurved spines. Vitellaria in lateral bunches of irregularly lobed follicles, postacetabular, extending up to level of genital pore or slightly anterior thereto. Vitelline ducts wide, converging towards vitelline reservoir near ovary. Eggs oval, $8-12 \times 7-8$. Excretory pore subterminal; vesicle apparently Y-shaped.

Remarks. The following species have been described in the genus Monorcheides Odhner, 1905: M. diplorchis Odhner, 1905; M. soldatovi Issaitschikow, 1928; M. petrowi Layman, 1930; M. cumingae (Martin, 1938) Martin, 1939; M. decapteri Manter et Pritchard, 1961; and *M. alexanderi* Arai, 1962. Martin (1940) expressed the opinion that M. petrowi may not belong to this genus. Margolis and Ching (1965) transferred it to the genus *Pentagramma* Chulkova, 1939. Among other species, M. alexanderi probably does not belong to this genus. With its postacetabular vitellaria and tubular excretory bladder, it readily fits into the genus Diplomonorchis Hopkins, 1941. M. alexanderi thus becomes D. alexanderi (Arai, 1962) comb. n. M. cumingae also has postacetabular vitellaria and in this and other characters it also fit into the genus Diplomonorchis Hopkins, 1941.

Monorcheides hafeezullahi sp. n. closely resembles M. decapteri Manter et Pritchard, 1961 in appearance, but differs in the robust body, in having four-lobed rather than three-lobed ovary, in the character and distribution of vitelline follicles and in the armature of terminal genitalia. In M. decapteri vitelline follicles are ovoid to elongate overlapping the acetabulum; the cirrus is armed with needle-like and thorn-like spines and metraterm with needle-like spines. In the new

species, the cirrus spines are of uniform type, being needle-like and metraterm has thorn-like spines.

Monorcheides diplorchis differs from M. hafeezullahi sp. n. in having a large three-lobed, dome-shaped ovary, largely intercaecal vitellaria, genital pore immediately posterior to caecal bifurcation and oral sucker distinctly larger than ventral sucker. M. soldatovi has a large three-lobed ovary, the vitellaria are distributed in the lateral fields between intestinal bifurcation and anterior margin of testes and the cirrus sac does not extend posterior to ventral sucker.

The species name is given in honour of Dr. M. Hafeezullah.

Conclusions

The pleuronectiform flatfishes of the Kerala coast harbour a characteristic, diverse, though not rich, trematode fauna. All the adult parasites collected during the present study except *Lepocreadioides orientalis* Park, 1939 have a distribution restricted to the seas around the Indian subcontinent. This is in conformity to the greater endemicity of the fish parasite fauna of the Indo-Pacific region (Rohde 1993).

Stephanostomum sp. metacercaria, Macvicaria jagannathi, didymozoid larva (Type 1) and didymozoid larva (Type 2) restricted their occurrence to a single host family. Of these, Stephanostomum sp. metacercaria and didymozoid metacercariae were stenoxenic. Macvicaria cynoglossi, M. longicaudus and Lepocreadioides orientalis infected two families of flat-fishes, Cynoglossidae and Soleidae.

Prosorhynchus sp. metacercarial infection was noted in four (Bothidae, Cynoglossidae, Psettodidae and Soleidae) out of the five host families examined. This parasite, however, was a cosmopolitan form recorded from flatfishes of several parts of the world (Yamaguti 1971).

Acknowledgements. I am deeply grateful to my supervising teacher Dr. S. Radhakrishnan, Department of Aquatic Biology and Fisheries for his valuable guidance in the present study. The work was carried out under a junior research fellowship of U.G.C. Respectful and sincere thanks are due to Prof. R. Madhavi, Andhra University, for her prompt help in various ways during the study and manuscript preparation. I am highly obliged to Dr. M. Hafeezullah for critically going through the manuscript.

References

Ahmad J. 1983. Studies on digenetic trematodes of the family Lecithasteridae from marine fishes from Bombay coast, Arabian Sea. *Rivista di Parassitologia*, 44, 119–124.

Ahmad J. 1988. Two new species of Allodecemtestis (Digenea: Opecoelidae) from marine fishes of Bay of Bengal. *Angewandte Parasitologie*, 29, 235–239.

Ahmad J., Dhar R. L. 1987. Studies on digenetic trematodes of marine fishes from the Puri coast of Bay of Bengal. Part III. Family Monorchiidae and Opecoelidae. *Pakistan Journal of Zoology*, 19, 184–196.

- Bilqees F. M. 1976. Two trematodes of the genus *Prosorhynchus* Odhner, 1905 (Bucephalidae) including a new species *P. erumenis* from the fish *Psettodes erumei* (Bl. & Sch.) off the Karachi Coast. *Norwegian Journal of Zoology*, 24, 345–348.
- Bray R. A. 1985. Macvicaria taksengi n. sp. (Digenea: Opecoelidae) in marine teleosts from Pinang, Malaysia. Systematic Parasitology, 7, 75–80.
- Bray R. A. 1990. Two new opecoelids (Digenea) in the flatfish *Pseudorhombus jenynsi* (Bleeker) from Shark Bay, Western Australia. *Systematic Parasitology*, 15, 33–40.
- Chauhan B. S. 1943. Trematodes from Indian marine fishes. Part II. On some trematodes of the gasterostome family Bucephalidae Poche, 1907; with description of four new species. *Proceedings of Indian Academy of Sciences, B*, 17, 97–117.
- Gibson D. I., Bray R. A. 1982. A study and reorganisation of *Plagioporus* Stafford, 1904 (Digena: Opecoelidae) and related genera, with special reference to forms from European Atlantic waters. *Journal of Natural History*, 16, 529–559.
- Gupta N. K., Mehrotra V. 1970. Observations on some already known digenetic trematodes of marine fishes of Ernakulam (south India). Research Bulletin (N.S.), Punjab University, 21, 87-99.
- Gupta P. C., Govind H. 1983. On a new trematode species *Lepocreadioides thaparai* n. sp. from a marine fish, *Cynoglossus lida* Gunther, from Bay of Bengal, off the Puri coast, Orissa. *Proceedings of Indian Science Congress*, 70, 128.
- Gupta P. C., Singh R. B. 1985. Four new digenetic trematodes from marine fishes off Puri coast, Bay of Bengal. *Indian Journal of Parasitology*, 9, 25–34.
- Gupta R. C., Gupta S. P. 1978. Five new trematodes of the genus Lecithocladium Luhe, 1901 (Hemiuridae) from marine fishes. Folia Parasitologica, 25, 223–229.
- Gupta S. P., Gupta R. C. 1976. On four new species of trematodes from marine fishes. *Indian Journal of Helminthology*, 28, 114–126.
- Gupta V., Ahmad J. 1976. Digenetic trematodes of marine fishes. On some new and known digenetic trematodes of the family Bucephalidae Poche, 1907 from marine fishes of Puri, Orissa, India. Anales del Instituto de Biologia. Universidad National de Mexico, Ser. Zoologia, 47 (2), 9–18.
- Gupta V., Ahmad J. 1979. Digenetic trematodes of marine fishes of Bay of Bengal, Puri, Orissa. *Helminthologia*, 16, 161–171.
- Gupta V., Puri M. 1982. Six new species of the genus *Lecithocladium* Luhe, 1901 (Hemiuridae Luhe, 1901) from marine fishes of Puri, Orissa. *Indian Journal of Helminthology*, 13, 58–78.
- Hafeezullah M. 1970. Lepocreadiid trematodes of marine fishes of India. *Parasitology*, 61, 345–356.
- Hafeezullah M. 1971a. On some new and known digenetic trematodes from marine fishes of India. *Indian Journal of Helminthology*, 45, 73–88.
- Hafeezullah M. 1971b. Opecoelid trematodes of marine fishes of India. *Parasitology*, 62, 321–329.
- Hafeezullah M., Chakrabarti S. 1994. Studies on variations and consequent systematic positions of various Indian species of *Lepocreadioides* Yamaguti, 1936 (Trematoda: Lepocreadioidae). Records of Zoological Survey of India, 94, 1–13.
- Hafeezullah M., Siddiqi A. H. 1970. Digenetic trematodes of marine fishes of India. Part I. Bucephalidae and Cryptogonimidae. *Indian Journal of Helminthology*, 22, 1–2.
- Køie M., Lester R. J. G. 1985. Larval didymozoids (Trematoda) in fishes from Moreton Bay, Australia. Proceedings of Helminthological Society of Washington, 54, 196–203.

- Madhavi R. 1972. Digenetic trematodes from marine fishes of Waltair coast, Bay of Bengal. I. Family Lepocreadiidae. Journal of Parasitology, 58, 217-225.
- Madhavi R. 1974. Digenetic trematodes from marine fishes of Waltair coast, Bay of Bengal. Family Monorchiidae. *Rivista di Parassitologia*, 36, 87–98.
- Madhavi R. 1975. Digenetic trematodes from marine fishes of Waltair coast, Bay of Bengal. Family Opecoelidae. Rivista di Parassitologia, 36, 153–164.
- Madhavi R. 1976. Digenetic trematodes from marine fishes of Waltair coast, Bay of Bengal. Family Bucephalidae. *Rivista di Parassitologia*, 35, 189–199.
- Madhavi R., Jhansi Lakshmibai K., Narasimha Rao N. 1994. Cercarial fauna of molluscs from Chilka lake: a new bucephalid cercaria and its metacercaria. *Journal of Parasitology and Applied Animal Biology*, 3, 79–82.
- Margolis L., Ching H. L. 1965. Review of the trematode genera Bacciger and Pentagramma (Fellodistomatidae) and description of P. petrowi (Layman, 1930) n. comb. from marine fishes from the Pacific coast of Canada. Canadian Journal of Zoology, 43, 381–405.
- Margolis L., Esch G. W., Holmes J. C., Kuris A. M., Schad G. A. 1982. The use of ecological terms in parasitology. *Journal of Parasitology*, 68, 131–133.
- Martin W. E. 1940. Studies on the trematodes of Woods Hole. III. The life cycle of *Monorcheides* (Martin) with special reference to its effect on the invertebrate host. *Biological Bulletin*, 79, 131–144.
- Menon A. G. K. 1977. A systematic monograph of the tongue sole of the genus *Cynoglossus* Hamilton-Buchanan (Pisces: Cynoglossidae). *Smithsonian Contributions to Zoology*, No. 238, 1–129.
- Park J. T. 1939. Trematodes of fishes from Tyosen. III. Some new trematodes of the family Allocreadiidae Stossich, 1904 and the genus *Macrolecithus* Hasegawa and Ozaki, 1926. *Keyzo Journal of Medicine*, 10, 52–62.
- Radhamanyamma P. 1988. Studies on Pleuronectiformes occurring along the southwest coast of India. Ph.D. thesis, University of Kerala, India.
- Ramanathan N., Vijaya P., Ramaiyan V., Natarajan R. 1977. On the biology of the large-scaled tongue sole *Cynoglossus macro*lepidotus (Bleeker). *Indian Journal of Fisheries*, 24, 83–89.
- Rohde K. 1993. Ecology of marine parasites. CAB International, Oxon, U.K.
- Salman A. S., Srivastava H. D. 1990. New and previously described digenetic trematodes (Plagioporinae Manter, 1947) from Indian marine fishes. Acta Parasitologica Polonica, 35, 11–18.
- Srivastava H. D. 1936. New allocreadiids (Trematoda) from Indian fishes. Part II. New parasites of the genus *Decemtestis* Yamaguti, 1934. *Proceedings of National Academy of Sci*ences, India, 6, 187–195.
- Srivastava H. D. 1938. Studies on the gasterostomatous trematodes of Indian food fishes. *Indian Journal of Veterinary Science and Animal Husbandry*, 8, 247–248.
- Srivastava H. D. 1941. New allocreadiids (Trematoda) from Indian marine food fishes. Part V. A new parasite of the genus Lepocreadioides Yamaguti, 1936. Indian Journal of Veterinary Science and Animal Husbandry, 11, 52–54.
- Yamaguti S. 1971. Synopsis of digenetic trematodes of vertebrates. Keigaku Publishing Co., Tokyo.