

Microbiological diagnosis of
Francisella tularensis
and Austrian epidemiology of
tularemia

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Workshop “Dangerous Pathogens” and Leptospirosis, 29 May 2009

Tularemia

- 1. Epidemiology** (Hofer)
- 2. Conventional methods** of diagnosis (Hofer)
- 3. Molecular methods** of diagnosis
(Revilla-Fernández)

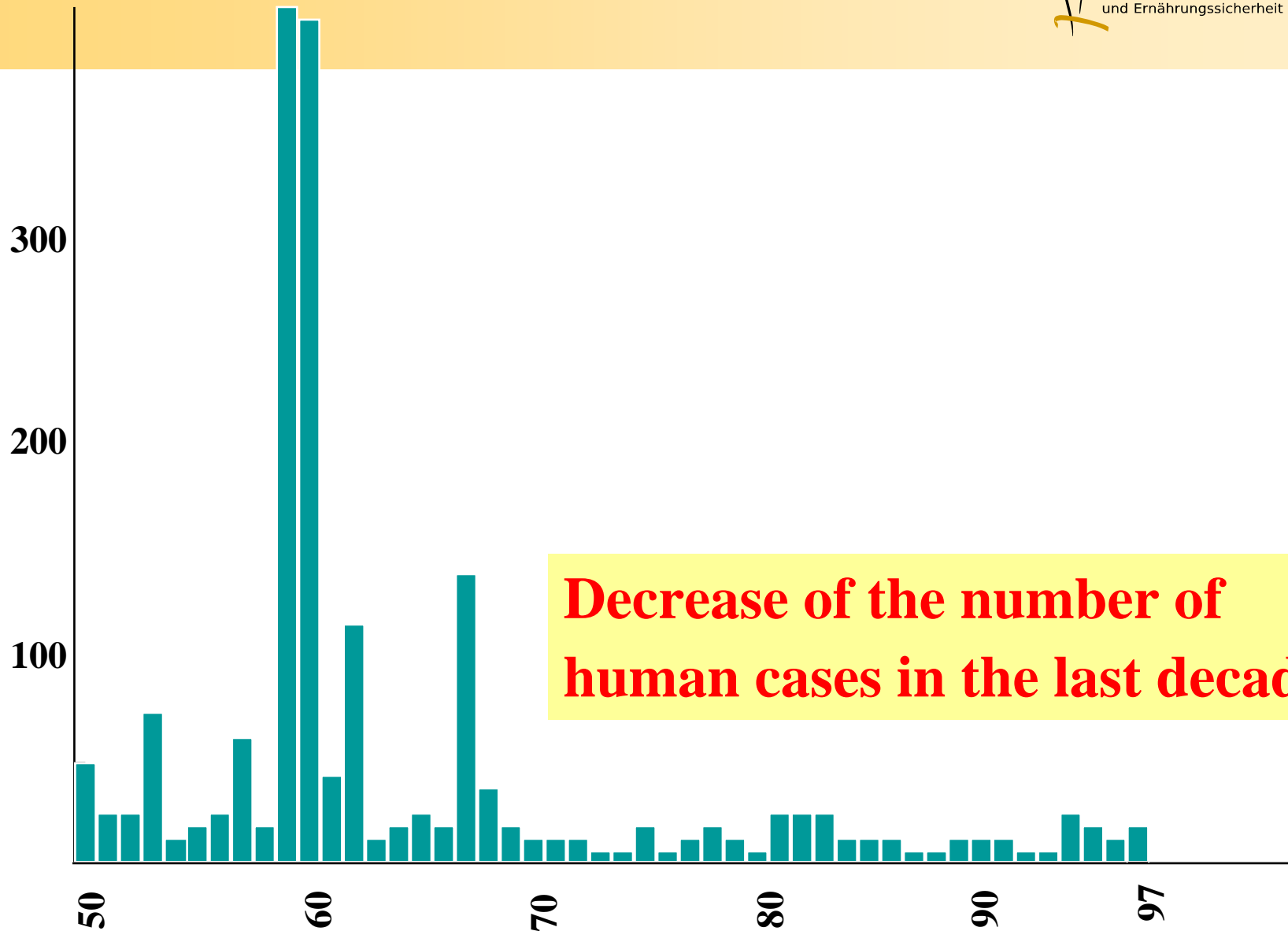
History of tularemia in Austria

1935 First diagnosis in a human in Lower Austria
(infection by skinning a hare)

1936/37 and **1945/46** 2 epidemics, each with about
200 notified cases (most human cases were caused
by **contact with hares**)

1959/60 More than 700 verified cases of pneumonia
in a **sugar factory** in the district of Bruck/Leitha
(inhalation of aerosol by washing contaminated beets
when mice were observed in masses after very dry
weather conditions in autumn)

Human cases from 1950 - 1997



Decrease of the number of human cases in the last decades

Diagnosed tularemia cases 1994 - 2001

Cases	1994	1995	1996	1997	1998	1999	2000	2001
Hare	20	18	5	33	37	27	16	11
Human	26	16	9	16	19	2	5	1

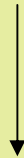
In activated natural foci an increase of the number of **hare** and **human** tularemia cases can be recorded (1994/95, 1997/98)

In **epidemic years** diagnosed human cases can reach the number of 26 (1994)

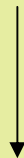
In **inter-epidemic years** diagnosed human cases mostly amount less than 5 (**1999 – 2008**)

Epidemic years

Cyclical increase of the **common vole** *Microtus arvalis*
(dry weather conditions)

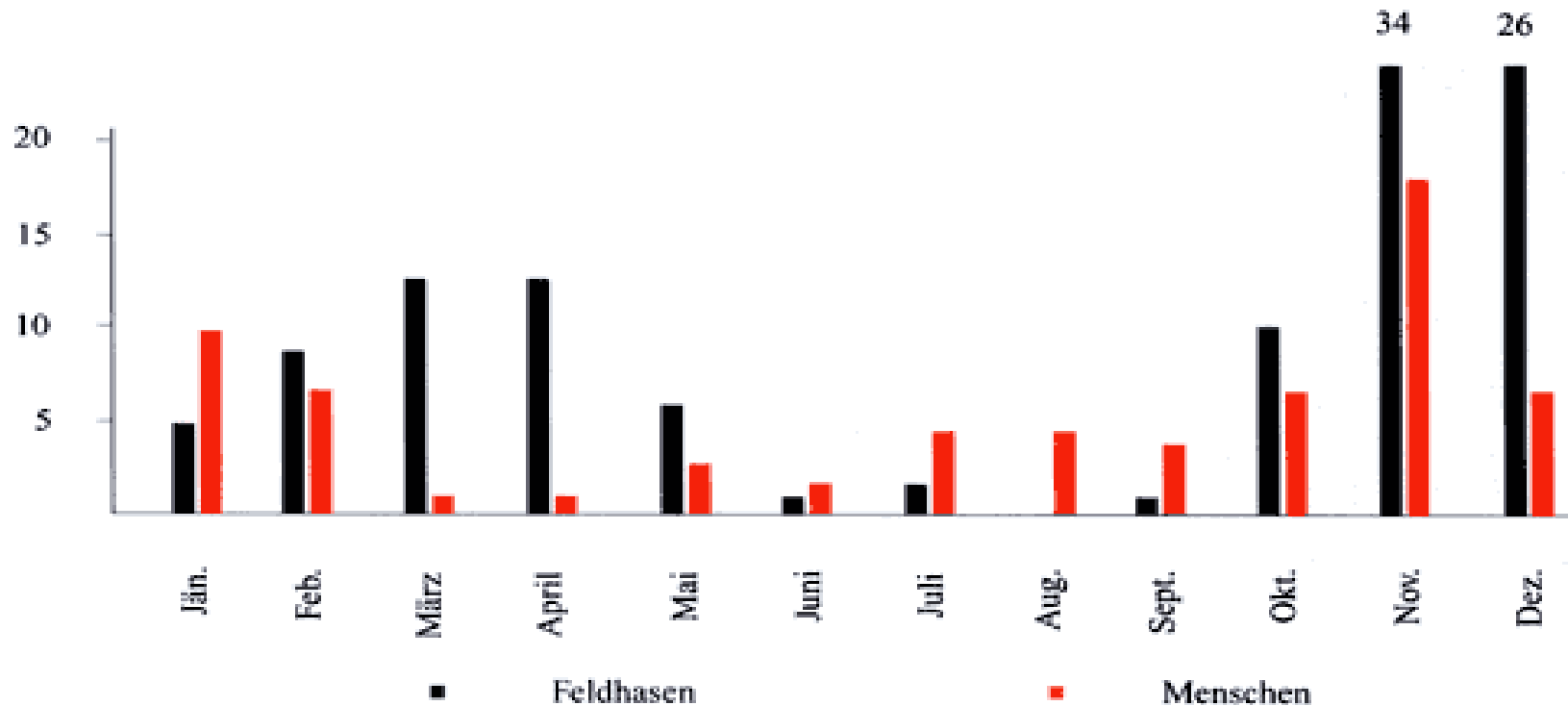


Outbreak in **brown hares** in autumn
(widespread deaths)



Frequent infection of **humans** during the hunting season
(skinning of hares) with increasing notified cases of tularemia
in late autumn

Tularemia cases from Oct. 1994 until March 1998



Most **human cases (red)** were diagnosed from Oct. until Feb., when many septic cases in hares were observed. An increasing number of **hare cases (black)** was also recorded from Feb. until May.

Where is the main reservoir of *Francisella tularensis* in Austria ?

Lepus europaeus

European hare

Brown hare

Field hare



- **infection carrier**
- **source of human infection**
- **latent or chronic** (interepidemic)
- **acute septicemia** (epidemic years)

How healthy are „healthy“ hares ?

1 week old culture

Inoculation with a
section plane
of the organs



Latent infection

Culture of kidney, spleen, liver
from hares **without macroscopic
pathological alterations**

Results

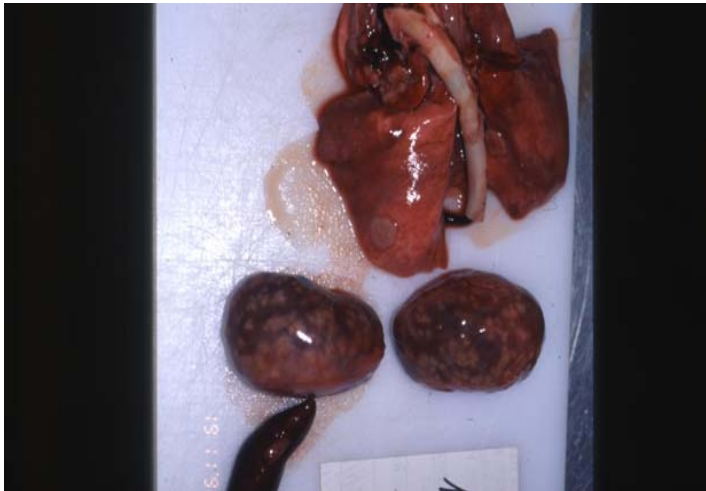
Single colony of *F. tularensis* from
the organs of **2 out of 98 hares**

Hunting bag examination Nov. 2007
TULMON Study
(VUW-FIWI, BMLV, AGES)

Chronically diseased hares excrete *Francisella* via urine !

Chronic tularemia

Necrotic lesions
in kidney and lung
Enlarged spleen



Chronic infection

40 hares (apparently healthy) shot in
December 2008 were examined

10 hares showed characteristic
pathologic findings

Culture of **5 hares** yielded *Francisella*
(small quantity of colonies, agar plate
must be inoculated with the section
plane of the organs)

Hunting bag examination Dec. 2008
TULMON Study
(VUW-FIWI, BMLV, AGES)

Epidemic outbreaks in hares („hare pest“)

Acute tularemia
Enlarged spleen



Photograph: T. Steineck, FIWI-VUW

Septic diseased hares

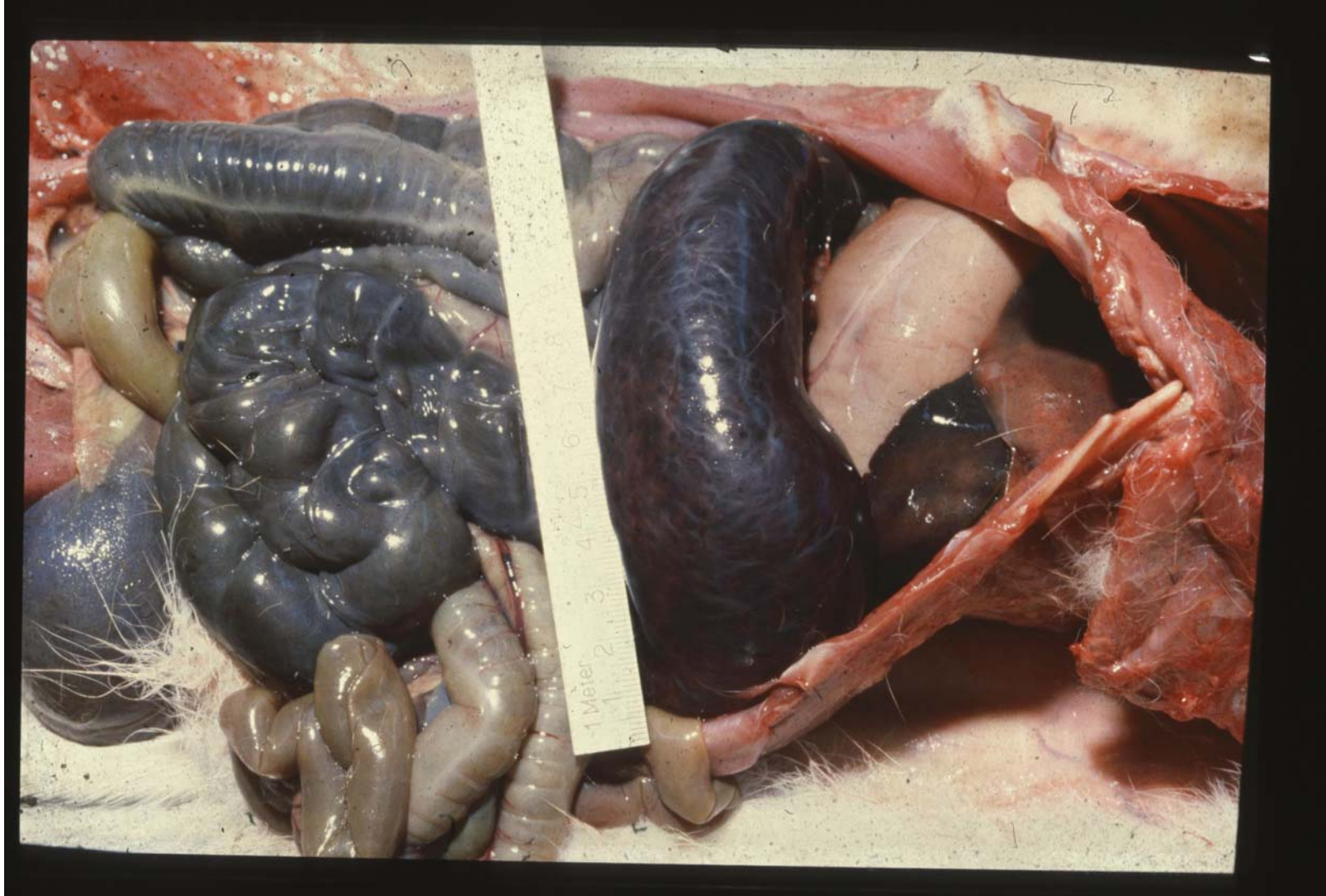
often show an enlarged spleen
exclusively !

Chronical pathologic alterations
can be found additionally in outbreaks

Epicarditis, pneumonia, nephritis
(purulent - necrotic) being most
frequent and necrotising **orchitis**
(not reported in hares before 1999)

Acute tularemia - Enlarged spleen

Photograph: T. Steineck, FIWI-VUW



Tularemia - Necrotising orchitis

Photograph: T. Steineck, FIWI-VUW



Tularemia outbreak in hares 1994/95

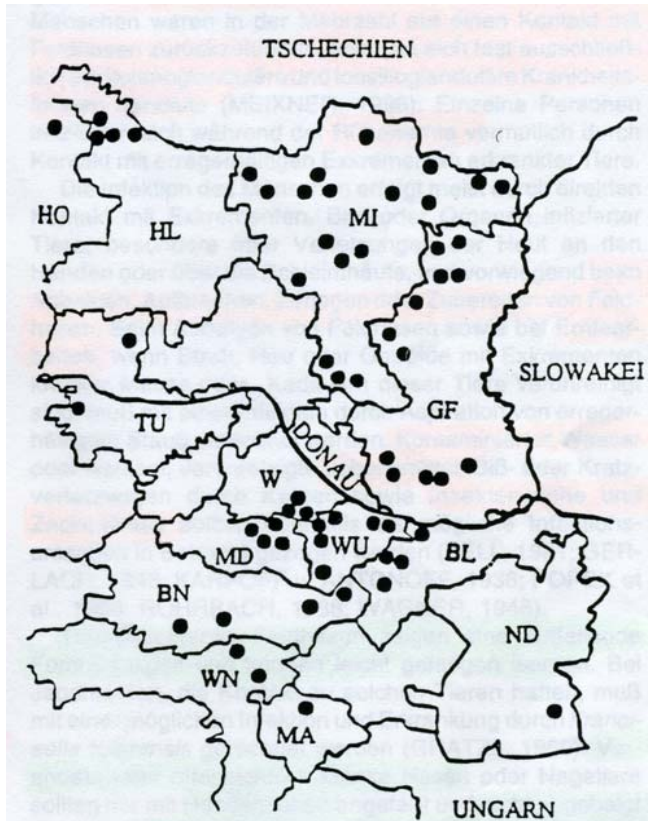


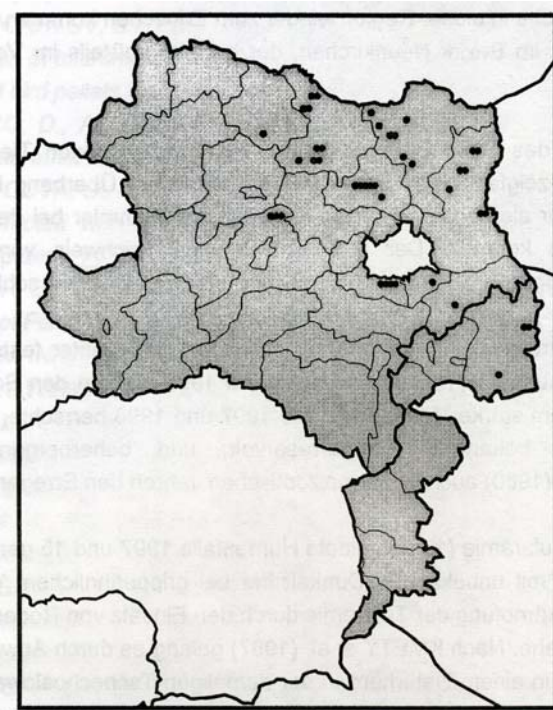
Abb. 5: Geographische Verbreitung der nachgewiesenen Tularämiefälle beim Feldhasen von Oktober 1994 bis Dezember 1996; W = Wien, WN = Wiener Neustadt, MA = Mattersburg, ND = Neusiedl am See, BN = Baden, BL = Bruck/Leitha, WU = Wien-Umgebung, MD = Mödling, TU = Tulln, GF = Gänserndorf, MI = Mistelbach, HL = Hollabrunn, HO = Horn

From Oct.1994 until Dec. 1996
62 cases in hares from 13 districts in
3 federal countries were diagnosed

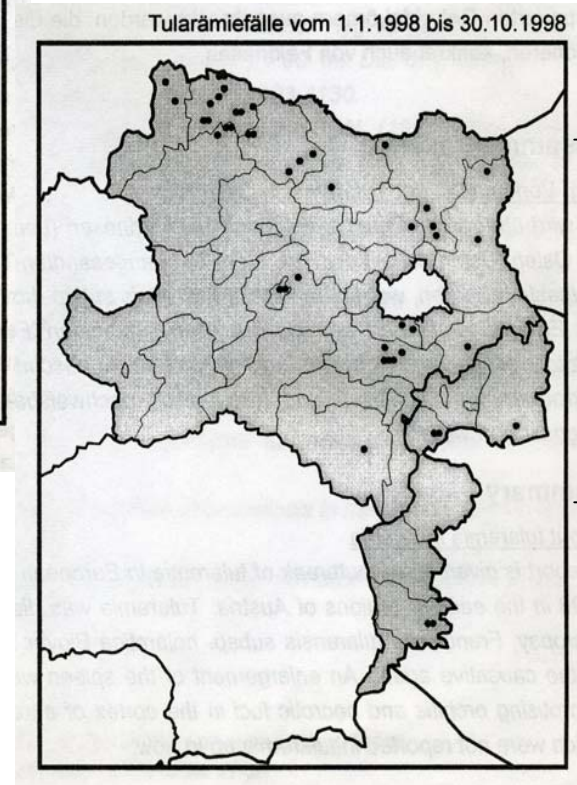
Most cases (hare and human) were
observed 1994 in the **district of
Mistelbach**

Endemic region is mainly the
North-eastern part of Austria
adjoining endemic regions of Czech
Republic, Slovakia, Hungary

Tularemia outbreak in hares 1997/98



Tularämiefälle vom 1.1.1997 bis 31.12.1997



Tularämiefälle vom 1.1.1998 bis 30.10.1998

1997 42 cases in hares from **10 districts** were diagnosed

1998 45 cases in hares from **16 districts** were diagnosed

Remarkable activation of the natural foci in the **south of Burgenland** and the **North of Lower Austria** 1998

Red foxes prefer preying on brown hares

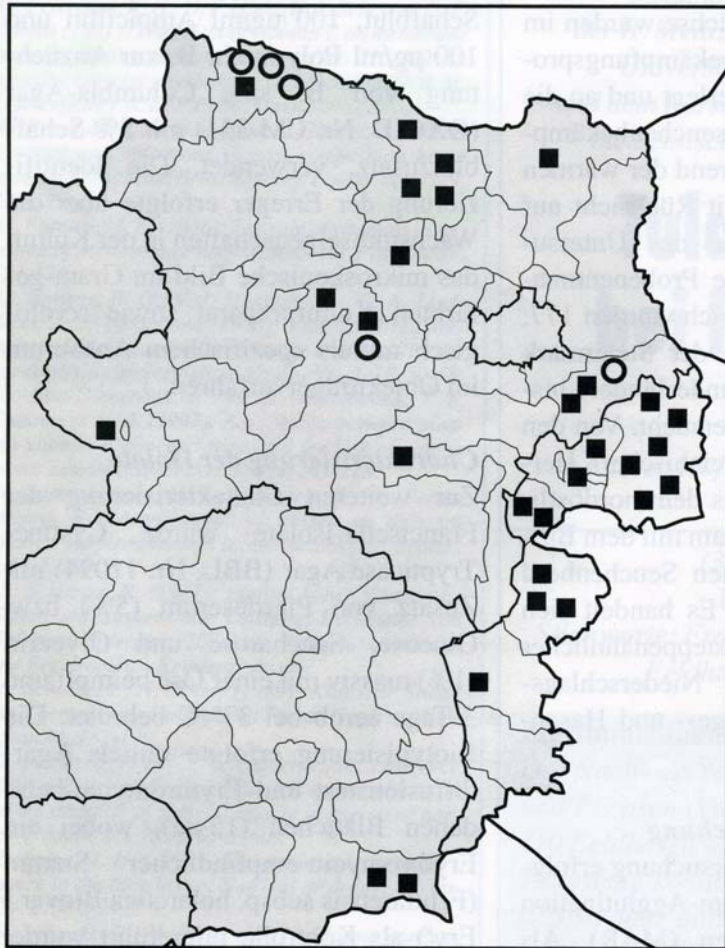


Abb. 2: Geographische Verbreitung der tularämiepositiven Füchse (■) und Feldhasen (○)

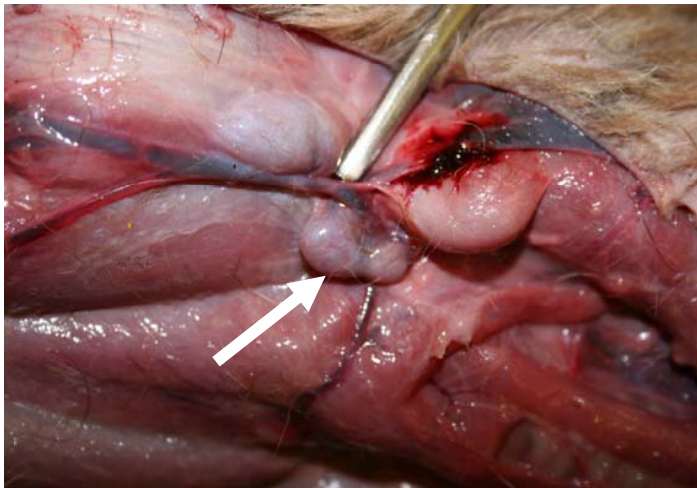
Latent tularemia infection in red foxes (*Vulpes vulpes*)

Serological investigation 1997/98
Tierärztl. Umschau 55, 264-268 (2000)

Activated natural foci of tularemia can be detected serologically by **investigation of blood from red foxes**

It is remarkable, that in the same area **foxes also showed antibodies against *Brucella*** (Hares are also a very important reservoir for *Brucella suis* biovar 2)

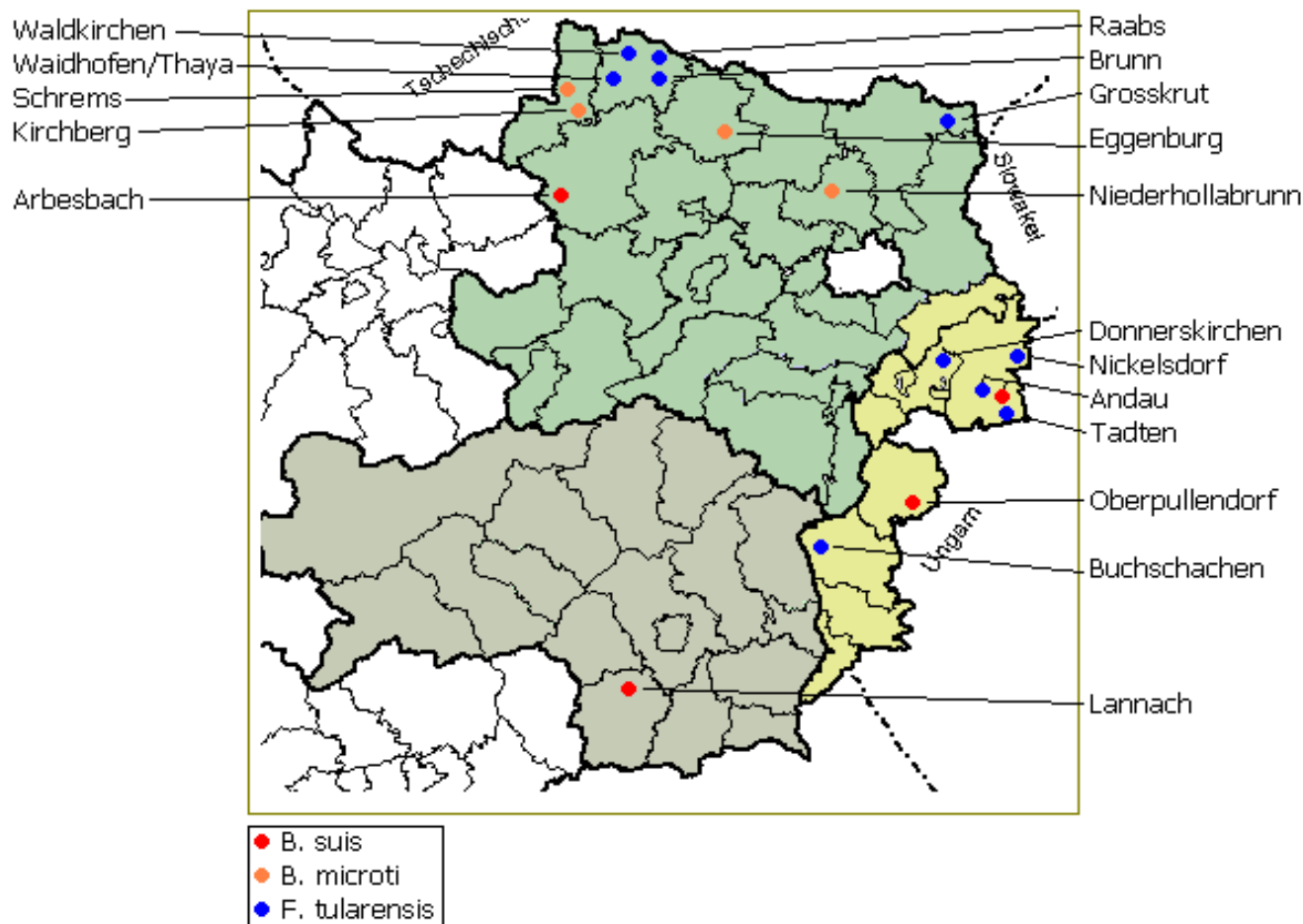
Francisella tularensis can be isolated from red foxes !



In **activated endemic regions** *Francisella tularensis* can be isolated from **mandibular lymph nodes** (arrow) with no visible lesions (latent infection)

Besides testing for rabies, additional **screening of red foxes** for the presence of *Francisella* should be conducted in endemic regions in Austria

Latent tularemia infection of foxes indicates natural foci



**Francisella isolation
from red foxes** ●

Autumn/winter 07/08
4 out of 51 foxes
from the district
Waidhofen./Thaya

Spring/summer 2008
1 out of 11 foxes
from the district
Mistelbach

June 07 - March 09
10 out of 494 foxes
from **4 districts of
Burgenland**
(only 5 cases are shown
in the graph)

Subspecies *tularensis* in Austria !

New serious epidemiological situation in Austria !

1990 **Subsp. *tularensis*** (high virulence for humans!)
was isolated from an *Ixodes ricinus* **tick** nearby **Graz** (Styria) in
a laboratory in Bratislava

Gurycova D.: „Characterisation and Classification
of different strains of **Francisella tularensis**
isolated in Central Europe.“

Second International Conference on Tularemia,
Hradec Kralove, Czech Republic, Vojenske Zdravotnicke Listy
– Supplementum, Rocnik LXVI, c. 1 (1997) 24.

Subspecies *tularensis* in Austria !

Schu S4 laboratory strain is the most likely source of the European isolates of *F. tularensis* subsp. *tularensis* and indicate that anthropogenic activities, such as movement of strains or animal vectors, account for the presence of these isolates in Europe.

Given the highly pathogenic nature of this subspecies, **the possibility that it has become established wild in the heartland of Europe carries significant public health implications.**

Citation: Chaudhuri RR, Ren C, Desmond L, Vincent GA, Silman NJ, et al (2007) **Genome Sequencing Shows that European Isolates of *Francisella tularensis* Subspecies *tularensis* Are Almost Identical to US Laboratory Strain Schu S4.** PLoS ONE 2(4): e352. doi:10.1371/journal.pone.0000352

Int J Antimicrob Agents. 2005 Oct;26(4):279-84.

Antimicrobial susceptibilities of Austrian *Francisella tularensis holarctica* biovar II strains. [Tomaso H](#), [Al Dahouk S](#), [Hofer E](#), [Splettstoesser WD](#), [Treu TM](#), [Dierich MP](#), [Neubauer H](#).

50 strains from hares and human patients were examined
24 antimicrobial agents were determined using Etests
All isolates were sensitive to **tetracyclines**,
aminoglycosides, **quinolones**, chloramphenicol and
rifampicin

Resistance was observed in all isolates against
erythromycin, **penicillins** and aztreonam

Data can be applied for the detection and comparison of resistance
development and for the guidance of therapy

Culture of *Francisella tularensis*

**4 – 5 days old culture
acute-septic tularemia
characteristically
dark agar medium
around the colonies**



Cystine Heart – Agar

Sheep blood 10 %

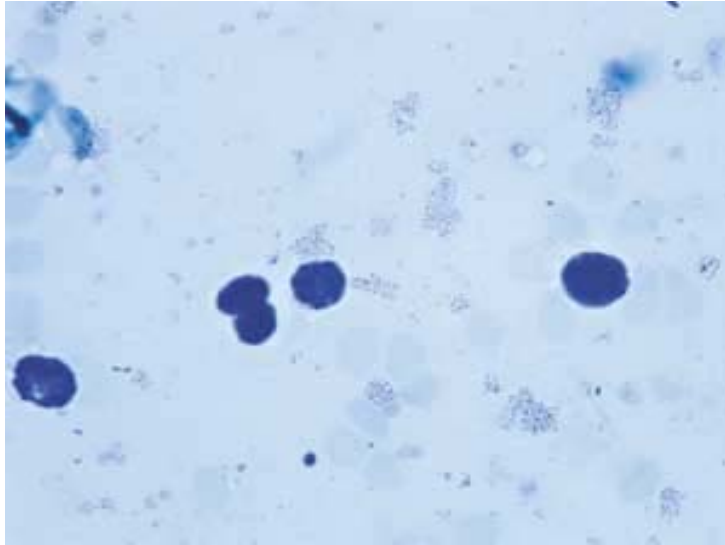
Ampicillin 100 µg/ml

Polymyxin B 100 µg/ml

Confluent growth after 2 - 3 days
incubation in septic cases
(inoculate with a loop)

Single colonies after 4 – 5 days
in chronic cases with small quantity
of bacteria
(inoculate with a section plane)

Identification of *Francisella tularensis*

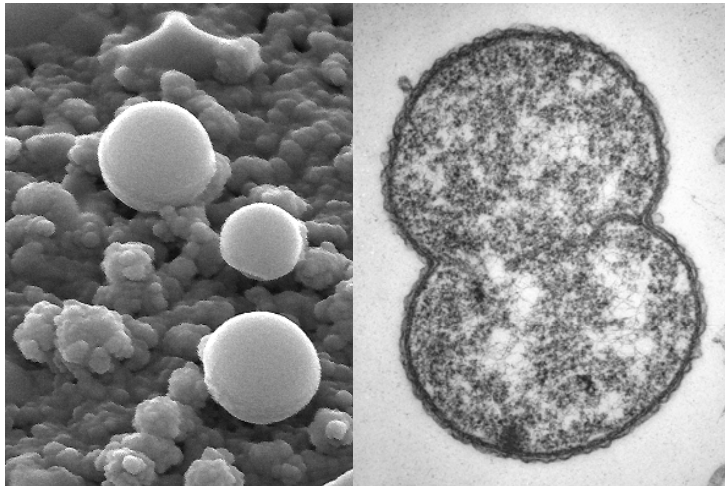


Extremely small!

Usually not visible in a Gram stain, visible in a **Giemsa** or **IF stain** in organs with a high number of bacteria (septic diseased hares)

Extremely coccoid!

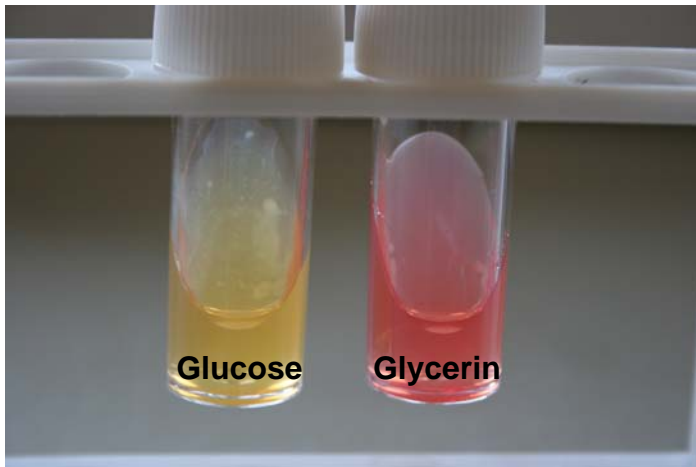
Culture material in a light microscope (phase contrast) or electron microscope



Rapid identification

slide agglutination test with culture material (specific antiserum)

Characterisation of *Francisella tularensis*



Control:
Francisella tularensis
subsp. *holarctica* Biovar I

**Subsp. *holarctica*
biovar I and II
utilize Glucose
but not Glycerin**

With agar diffusion test (15 µg Ery)
only

**Subsp. *holarctica* biovar II
shows resistance** (no inhib. zone)

Only phenotype found in hares and
humans in Austria till now

Molecular methods for the diagnosis and differentiation of *Francisella tularensis*

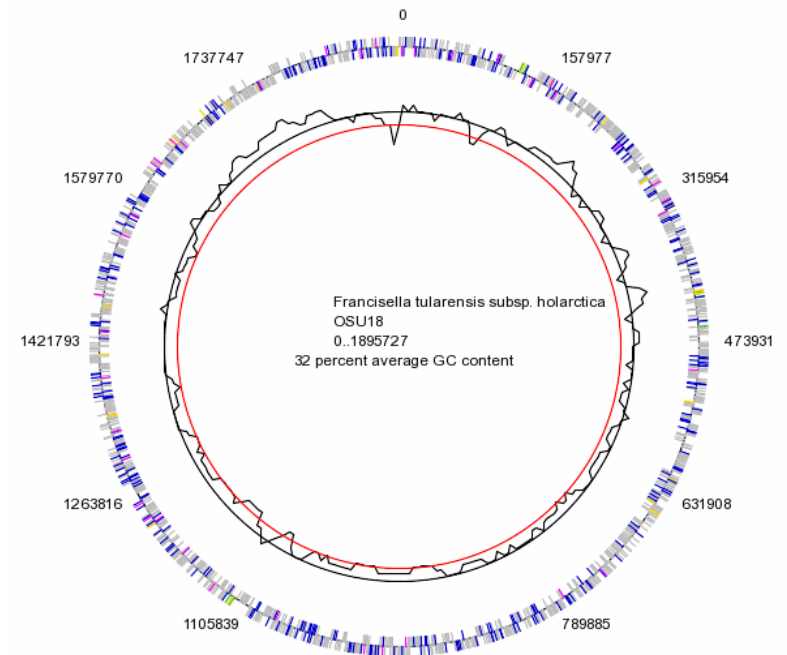
Sandra Revilla-Fernández

Institute for Veterinary Disease Control, Mödling

Workshop “Dangerous Pathogens” and Leptospirosis, 29 May 2009

Francisella tularensis- Taxonomy

- 7 complete genomes sequenced (approx. 1,89 Mb)
- 4 recognised Subspecies:
 - *F. tularensis* subsp. *tularensis*
 - *F. tularensis* subsp. *holarctica*
 - *F. tularensis* subsp. *mediasiatica*
 - *F. tularensis* subsp. *novicida*
- close genetic relationship despite marked variation in virulence in mammals
- *F. tul. tul.* has greater diversity than *F. tul. holar.* despite a less broader geographical prevalence



Francisella Genome Research:
<http://www.francisella.org/>

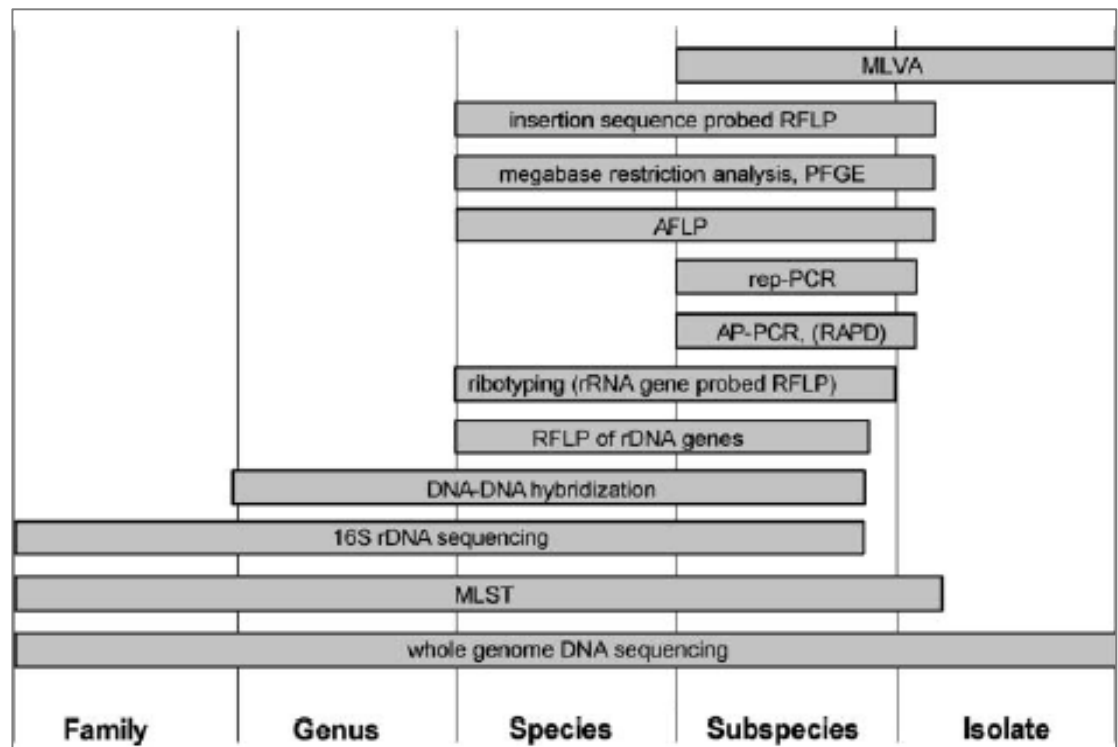
Diversity of molecular methods

➤ **Detection methods:**

- classical PCR
- real-time PCR
- PCR + RFLP
- 16S rRNA

➤ **Relative resolution of DNA-based typing methods**

„a combination of the specific PCR together with one method generating subspecies-specific patterns is suitable as a rapid and relatively simple strategy for discrimination of Francisella species and subspecies.“
(Johansson et al., 2000: J.Clin.Microbiol. 38)



(Johansson et al., 2004, APMIS 112)

Applied methods

		Target gene	Specificity	Publication
1	<i>Francisella tularensis</i> LC Real-time PCR with hybridisation probes	<i>tul4 (lpnA)</i> 17 kDa protein	conserved in <i>Francisella tularensis</i>	In-house Oligos (Plicka, MoD/ARWT) not published
2	<i>Francisella tularensis</i> SYBR Green real-time PCR	Ft-M19	differentiation of <i>F. tularensis</i> subsp. <i>holarctica/ tularensis</i>	In-house Oligos (Plicka, MoD/ARWT) not published
3	DNA Sequencing	<i>tul4</i> protein 16s rRNA	confirmation, not discriminative	
4	<i>Francisella</i> MLVA analysis (in process) cooperation AGES-MoD	sequencing of 6 VNTR Markers of <i>F. tularensis</i>	Phylogenetical clustering, diversity, strain linkage	Johansson et al., 2004 Byström et al., 2005
Other typing methods used within international cooperation				
5	<i>Francisella</i> MLVA analysis (in process) cooperation AGES- FLI Jena	sequencing of 6 VNTR Markers of <i>F. tularensis</i>	Phylogenetical clustering, diversity, strain linkage	Johansson et al., 2004 Byström et al., 2005
6	DNA sequencing cooperation AGES-Bundeswehr Munich	5 housekeeping genes 2 protein genes	Phylogenetical clustering for <i>tularensis/holarctica</i>	Nübel et al., 2006

DNA-typing methods allow safe strain characterisation from killed bacterial preparations!!

Laboratorial Procedure

human, fox, hare
tissue, wound

phenotypic profiling

bacterial
culture

DNA isolation

High Pure Template
PCR Kit

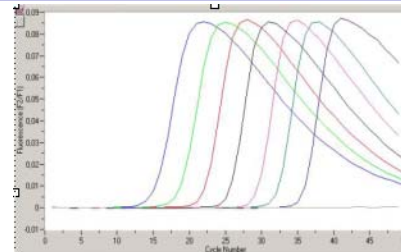
positiv

Francisella tul4
real-time PCR

negativ

Differentiation Ft-
M19 real-time PCR

F. tul. tularensis
F. tul. holarctica



Gel
Electrophoresis

sequencing

16S rDNA sequencing

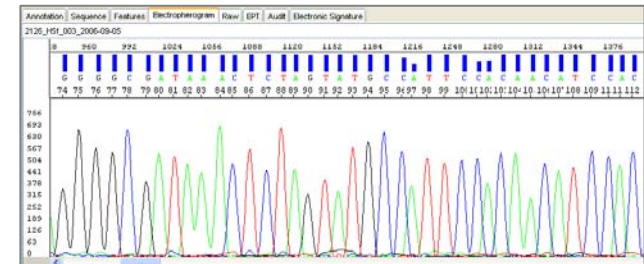
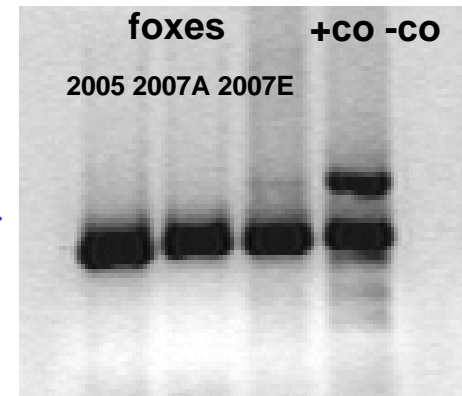
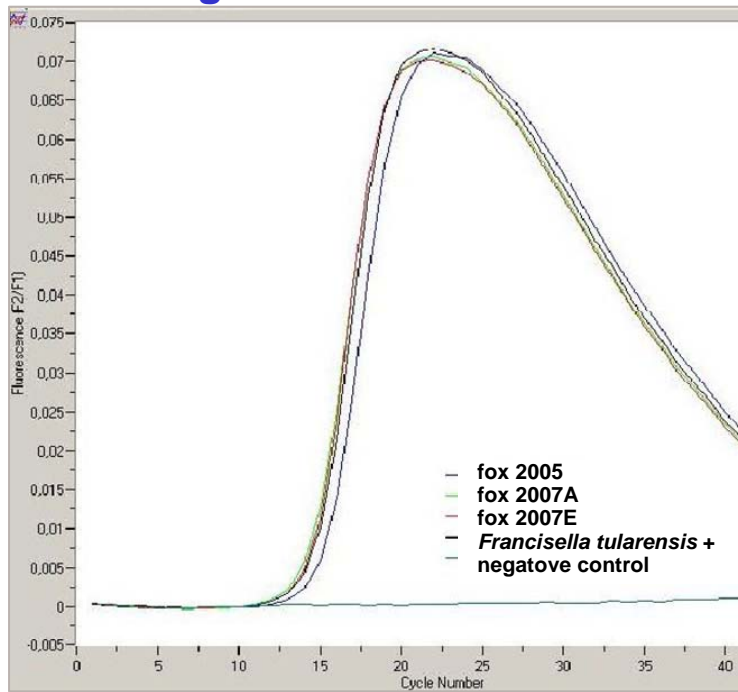
Bacterial
Species
determination

MLVA-VNTR typing

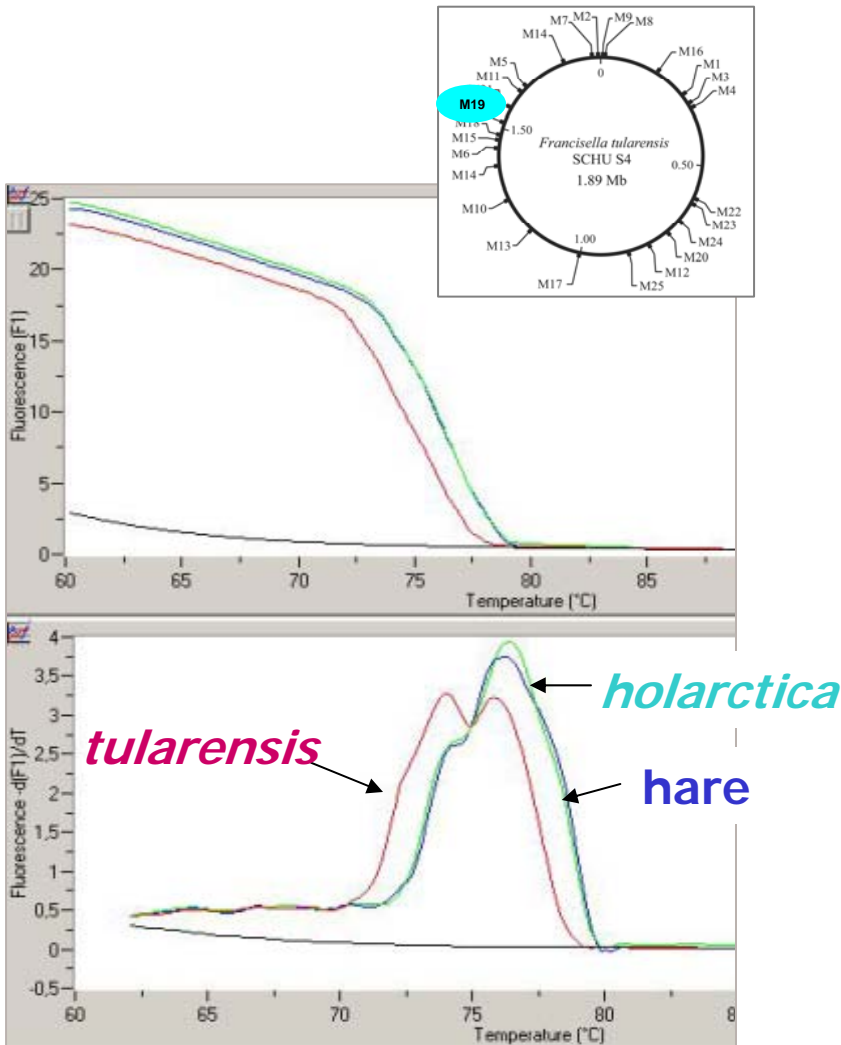
Francisella tularensis : detection

- **Positive cases (2004-2008):** wild hares (organs, culture)
2 human samples (lymph node, FFPE)
red foxes (culture)
- **Results:**

tul4 gene real-time PCR

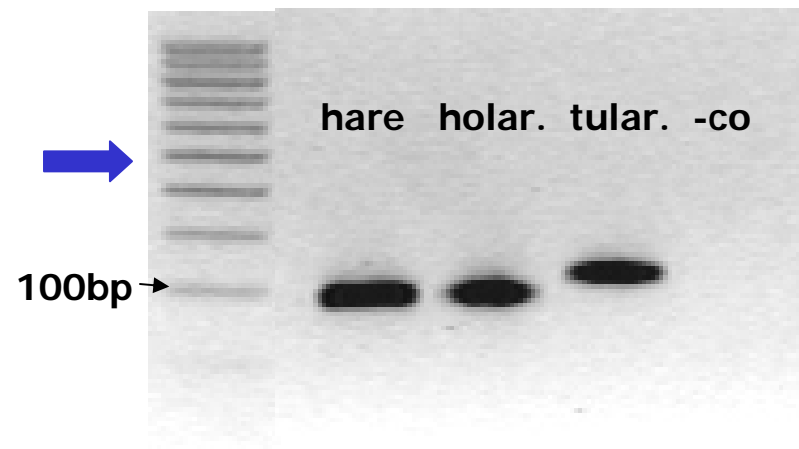


F. tularensis subsp. *holarctica*: Subspecies differentiation



Real-time PCR and melting curve analysis of Ft-M19 fragments:

The assay rapidly identifies the two main subspecies since a 30-bp sequence is deleted only in isolates of subspecies *holarctica*.

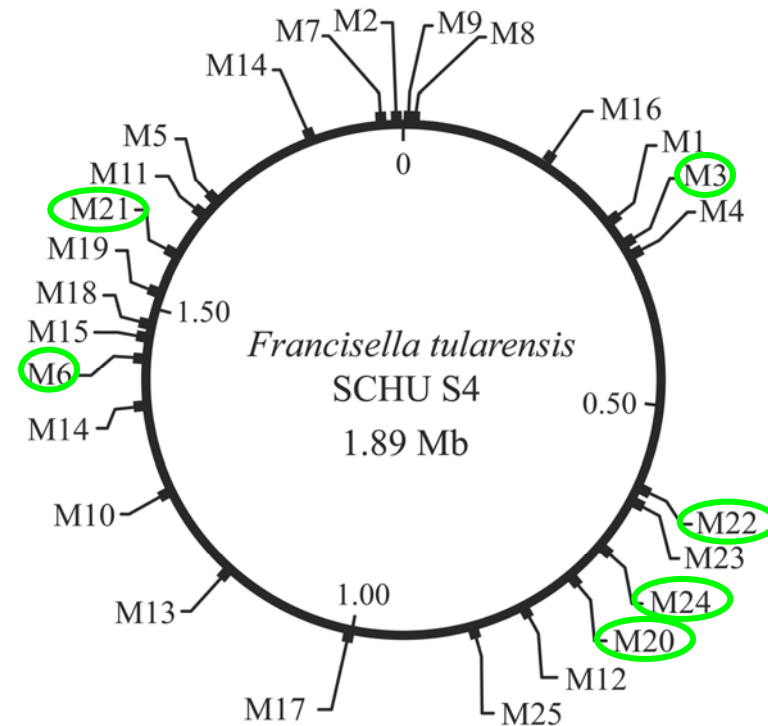


DNA-fragments of approximately 101-bp are amplified only from isolates of subspecies *holarctica*, whereas fragments of 131-bp are amplified only from isolates of subspecies *tularensis*.

MLVA-VNTR analysis of Austrian isolates 1997 til 2009 (1)

❖ Cooperation AGES-MoD/ARWT (Plicka)

- 23 *F. tul. holar.* strains from hare, fox and human samples (limited geographical distribution)
- Project start: May 09
- MLVA-VNTR: Sequencing of 6 discriminative VNTR-loci of *F. tularensis* and phylogenetical analysis
- Future analysis of repeat units of the VNTR loci will be validated with GeneMapper Analysis software and ABI DNA Sequencer



[\(Johansson et al., 2004, APMIS 112\)](#)

MLVA-VNTR analysis of Austrian isolates 1997 til 2009 (2)

Preliminary results:

- MLVA analysis of **Ft-M3** locus of 23 Austrian isolates of *F. tul. holarctica*, which is the most discriminative, generated at least **3 different alleles**.
- The little genetic diversity found in *F. tul. holarctica* may be in accordance with previous global analysis
- Interspecies transmission between hare and fox is shown
- The genetic relationship to strains from other Austrian regions remains under study

MLVA-VNTR typing of Austrian isolates 2005 til 2009

❖ Cooperation AGES- Dr.Tomaso (FLI, Jena)

- 49 strains of *F. tularensis* subsp. *holarctica* sent in Februar 09 for MLVA-VNTR-Analysis (hare, fox, human samples) from a wider geographical distribution
- Preliminary analysis of Ft-M3 and Ft-M6 sequences reveals linkage of the Austrian strains (99-100% homology)
- Analysis still in process, not published

Acknowledgements:

We thank all laboratory staff and contributors from the Institute for Veterinary Disease Control in Mödling (K. Reisp), the Austrian Ministry for Health (A. Höflechner-Pörtl), Austrian Ministry of Defence (H. Plicka) and the VUW-FIWI (T. Steinek) for their excellent work and assistance

Thank you very much for your attention



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