

# NATURAL HISTORY OF THE RIVERLANDS AND MURRAYLANDS CONTINUED

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## CHAPTER 9. FLORA and VEGETATION

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For tens of thousands of years the natural vegetation of the Murray Mallee and the Riverland was used by Aboriginal people for food, as firewood and for the construction of their wide range of wooden implements. Particularly in the Riverland, this exploitation by the relatively dense human population that lived on the river would have had quite a high impact. Over this long period, a point had been reached where the range of vegetation communities and their associated plants and animals were well adapted to Aboriginal land management practices.

The first European to enter this area was Captain Charles Sturt on his epic journey down the Murray River in 1830 (Sturt 1833). On what is now the South Australian part of the Murray River he was impressed by the towering river red gums on the banks and by the spectacular cliffs that lined the river along part of its length. He also had a good word to say about the country on the eastern bank of the river around present day Murray Bridge. He described it, in the fashion of the time, as follows:

*'The cliffs gave way to undulating hills that were different in appearance from the country we had previously noted down. It would have been impossible for the most tasteful individual to have laid out a pleasure ground to more advantage than Nature had done in planting and disposing various groups of trees along the spine and upon the sides of the elevations that confined the river and bounded the low ground that intervened between it and their base'*

Sturt was however, much less impressed with the majority of the country he encountered away from the river valley, which he described as follows:

*'Again hemmed in by those sterile and sandy tracts upon which the beasts of the field could obtain neither food or water'*

Further downstream he said that:

*'To the north the interior was evidently depressed, it was overgrown with a low scrub and seemed to be barren in the extreme'*

He summarised his impressions as follows:

*'If we accept the partial and alluvial flats on the immediate border and into the neighbourhood of its tributaries and creeks, the Murray might be said to flow through a barren and sandy interior'*

If only this view of the barren nature of the country away from the river had prevailed and the land away from the river valley had not been cleared for agriculture or over-grazed by sheep, the current serious salinity and land management problems of the Murray Darling Basin could have been avoided.

Much of the Murraylands and Riverland were, at the time of European settlement, covered with that most characteristic Australian vegetation community the Mallee (Noble & Bradstock 1989; Noble et al. 1990). 'Mallee' is not easy to define and is perhaps best described as vegetation dominated by mallee growth-form eucalypts. It can also define a particular plant growth form where many stems arise from a large underground woody swelling composed of stem tissue and known as a lignotuber. Thirdly, the word 'mallee' defines a part of Australia around the junction of the three southeastern States (also called the 'Murray Mallee') where mallee eucalypts are a dominant landscape characteristic. Mallee growth form eucalypts also occur widely across Australia in very different vegetation communities from those in the area covered by this natural history (Hill 1989). A

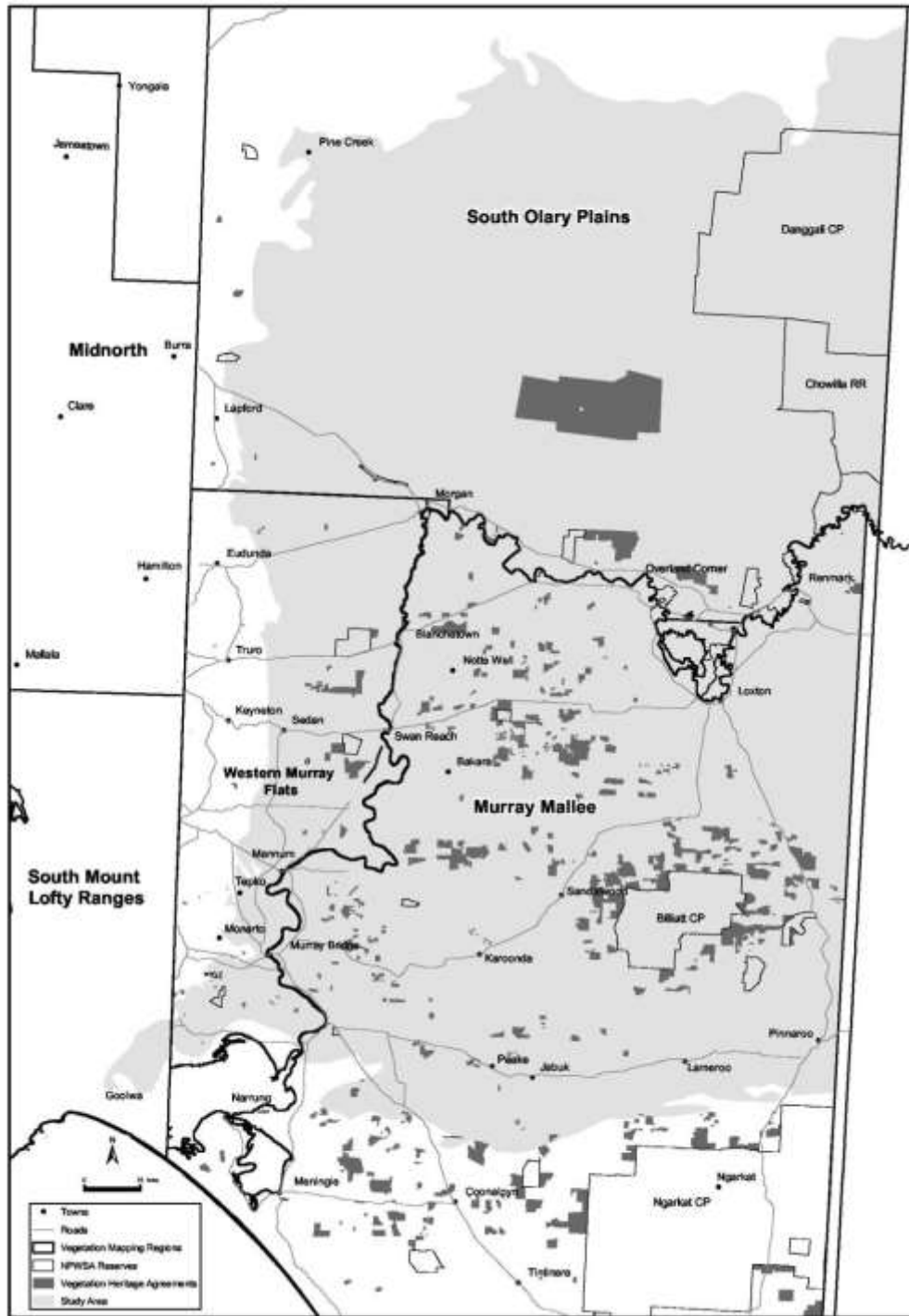
further difficulty arises where even within the mallee that occurs in the Mediterranean climate areas of Australia with mean annual rainfall of 250-400 mm, mallee-form shrubs and trees can occur with a wide variety of different understorey plants making the precise definition of particular vegetation communities very difficult (Sparrow 1989, 1990). To further complicate development of a clear definition of a particular mallee vegetation community studies of the same patch of vegetation over time have demonstrated that all the component plant species are not necessarily visible above the ground in a particular year. Different species of ephemeral understorey plants will germinate following particular combinations of rainfall and temperature, and it can take a number of visits in different conditions before the complete suite of plant species that makes up a particular vegetation community is known (Fox 1990). In spite of these difficulties in recognition of mallee vegetation communities there are some patterns associated predominantly with soil type and rainfall, which can be discerned across the broad landscape (Sparrow 1989). Because of this, clearance of vegetation for agriculture in the Murray Mallee has proceeded chronologically from the best combinations of soil and rainfall for agriculture to those areas that are least suitable. For some communities at the more fertile end there has been so much large-scale clearing and grazing that even the tiny remnants that remain today have been so invaded by weeds that they can be regarded as only semi-natural and quite significantly degraded from their pre- European settlement condition. Conversely the few relatively large areas of mallee vegetation set aside for conservation represent disproportionately the vegetation associations of the poor soils and lower rainfall areas which were perceived to be of little agricultural value.

As annual rainfall further decreases to the north the mallee communities are replaced by a range of Black Oak woodlands and chenopod shrublands and it is these communities which form the basis of the rangeland sheep grazing industry in this area. Although native vegetation is still continuous across this area, there are considerable differences in the health of this vegetation and many areas have changed significantly due to past and, in some cases ongoing, levels of overgrazing by a combination of domestic stock, feral animals and kangaroos.

Cutting through the surrounding plains and dunefields of the Murraylands is the ancient floodplain of the Murray River. In South Australia it can be conveniently divided into three distinct regions. The Mallee Trench from the NSW/Vic. border to Overland Corner is a wide floodplain crossed by the river in a single well-defined channel. Large areas of the floodplain have been cleared of their original vegetation and developed for irrigation of crops of grapes, citrus and stone fruits. The area between Renmark and Morgan is the most important irrigation region in South Australia. The Mallee Gorge extends from Overland Corner to Mannum and here the river has cut down through hard limestone rocks forming a spectacular series of cliffs along the river channel. Along this stretch of the river valley the predominant land use is sheep grazing on both cleared land and areas of remnant natural vegetation. Some irrigated blocks have also been cleared to grow lucerne and vegetables. Along the Lower Murray between Mannum and Wellington much of the valley floor was formerly occupied by wetlands. Most of these have been drained and are now used for dairying. There are only a few remnants of the original vegetation remaining.

### **Vegetation Surveys and Mapping**

From 1990 to 2003 a series of systematic vegetation surveys were carried out across the Murraylands and the Riverland (Table 1). The areas covered by the surveys are shown in Figure 1. The vegetation quadrat data from these four surveys were analysed separately to determine the floristic vegetation communities across each of the survey areas and vegetation maps were then produced from aerial photography. Where possible the floristic vegetation communities were mapped, but a proportion are too restricted or not identifiable from aerial photography. The vegetation maps therefore present a simplified set of vegetation communities when compared with the floristic analyses of the quadrat data from these biological surveys.



**Figure 1.** The Murraylands and Riverland showing areas covered by the four vegetation surveys and the Government conservation areas, private Heritage Agreement areas and other conservation tenures. Only the largest conservation areas are named.

Native vegetation of course does not really occur in discrete discernable units that can be easily mapped. It is actually a series of intergrading communities that often occur in a complex mosaic. Vegetation maps are therefore simplifications of reality and, as the scale of vegetation mapping increases so does the degree of simplification required. Having said this however, vegetation does not change randomly across the landscape and there are clearly close relationships between particular plants or groups of plants and particular soils or landforms. The skill of vegetation mapping is to reflect this variability in a way that makes sense to the observer when he confronts a group of

plants in the field that has been mapped as a particular vegetation community. The vegetation maps for the area covered by this natural history were produced at three separate scales (Table 1). To produce the vegetation mapping for the whole of the Murraylands and Riverland therefore there has been a process of further simplification and amalgamation of these earlier vegetation classifications and mapping. Areas of both existing and pre-European vegetation were calculated for each of the simplified communities. Where vegetation communities were originally mapped as mosaics of two or three communities, statistics for the area calculations were based on ratios of 50/50 or 40/30/30.

**Table 1.** Details of vegetation surveys conducted in the Riverland and Murraylands.

Survey	Type	Date	No of quadrats	Reference	Vegetation map scale
Murray Mallee	Existing	1990	679	Foulkes & Gillen 2000	1:50 000
South Olary Plains	Existing	1991	480	Forward & Robinson 1996	1:100 000
Western Murray Flats	Existing	1992	212	Lock & Goodwins 1993	1:50 000
Murray Valley	Existing	2003	273	Stewart <i>et al.</i> (unpubl.)	1:20 000
Murray Mallee	Pre-European	2004	N/A	Croft unpublished map	1:50 000
Western Murray Flats	Pre-European	2005	N/A	Croft unpublished map	1:50 000

### Pre-European Vegetation Mapping

Mapping of the pre-European settlement vegetation involves the use of historical and current records. Sources include the notes of early explorers, the annotations on the Hundred Plans drawn up when the State was originally surveyed. Remnant trees in paddocks and remaining roadside vegetation also provide clues to what the composition of the pre-European vegetation cover may have been. This information is combined with topographic and soil maps and extrapolation from the remaining remnant natural vegetation patches to produce a vegetation map across the whole landscape. To date pre-European mapping has only been attempted in any detail in some areas of the extensively cleared agricultural regions and for the area covered by this natural history, only the Murray Mallee and the Western Murray Flats have been mapped to date. Only the existing vegetation has been mapped for the Murray River valley and the pastoral rangeland country of the South Olary Plains.

### Vascular Plant Species List

The Biological Databases of South Australia incorporate data on the vascular plants recorded as part of the Biological Survey of South Australia and associated projects. This data is held in the following databases, SURVEY (species lists from all standard plant quadrats collected during standardised biological surveys using the methods outlined in Heard & Channon (1997)), OPPORTUNE (Plant records from geo-coded points across the region), RESERVES (Plant species lists compiled from a variety of sources for the conservation reserves and heritage agreement areas across the region) and PLANT POPULATION (Records of threatened plant populations in the region). In addition the State Herbarium of South Australia has a database of all its plant specimens (ADHERB). Within the boundary of the Murray Mallee and Riverland, all vascular plant records together with their locations and other associated information were extracted. The Biological Databases of South Australia provided 45 501 records while the State Herbarium Database provided 24 601. These lists have been extensively edited to ensure that the final species list produced confirms to the taxonomy in the 2004 Census of Vascular Plants of South Australia (Barker *et al.* 2005), and state and national conservation ratings have also been provided for all the threatened plant species in the area.

A total of 2153 plant taxa are known from the Riverland and Murraylands region (Appendix). 554 (26%) of these have been introduced during the period of European settlement and 64 are proclaimed as Pest Plants due largely to their impact on agricultural production. Many are considered

to be threatened with extinction with 12 species listed Nationally, and a total of 186 species threatened within SA. A selection of some of these threatened species is shown in Figures 2 and 3.



**Figure 2.** Some threatened plant species of the Murraylands and Riverland. Endangered in SA. **A**, Osborn's Eyebright (*Euphrasia collina* ssp. *osbornii*) (Photograph: D. Kraehenbuehl), **B**, Monarto Mintbush (*Prostanthera eurybioides*) (Photograph: L. Heard), **C**, Golden Drumsticks (*Pycnosorus chrysanthes*) (Photograph: P. Lang), **D**, Coloured Spider orchid (*Caladenia colorata*) (Photograph: D. Kraehenbuehl), **E**, Metallic Sun Orchid (*Thelymitra epipactoides*) (Photograph: D. Kraehenbuehl). Vulnerable in SA. **F**, Slender Bell Fruit (*Codonocarpus pyramidalis*) (Photograph: A. Robinson).





**Figure 3.** Some threatened plant species of the Murraylands and Riverland. Vulnerable in SA: **A**, Behr's Swainson-pea (*Swainsona behriana*) (Photograph: P. Lang); **B**, Broombush Templetonia (*Templetonia stenophylla*) (Photograph: P. Lang); **C**, Trailing Hopbush (*Dodonaea procumbens*) (Photograph: P. Lang); **D**, Silver Daisy-bush (*Olearia pannosa* ssp. *pannosa*) (Photograph: A. Robinson). Rare in SA: **E**, Yellow Swainson-pea (*Swainsona pyrophila*) (Photograph: D. Krahenbuehl); **F**, Australian Broomrape (*Orobanche cernua* var. *australiana*) (Photograph: A. Robinson).

### **Vegetation Communities**

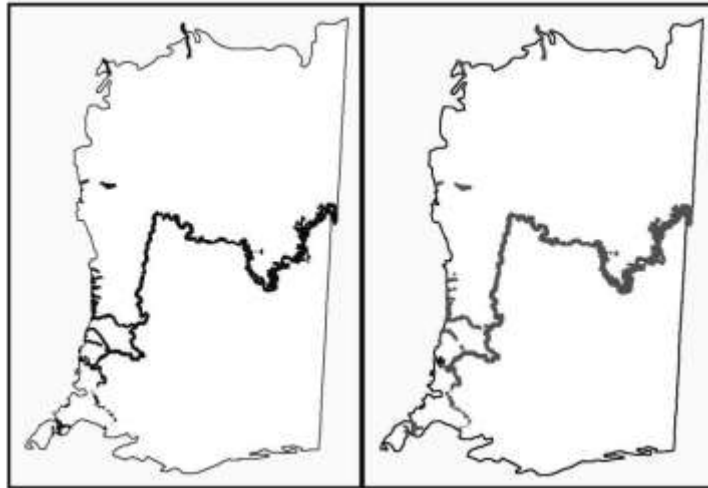
The vegetation communities over the large area of the region are quite complex and varied with 58 recognised vegetation communities. They can be broken down into three major categories, those that are widespread and therefore characteristic of the area (36 communities), those that are still characteristic but which occur over a relatively small area (4 communities) and those which are more characteristic of the surrounding areas and only have a minor intrusion into the area (14 communities). The 36 widespread, or formerly widespread, communities are described below, with a photo of a typical example a map of their pre-European and current extent (remembering that there is no pre-European vegetation mapping available as yet for the Murray Valley and South Olary Plains), a list of characteristic species, the typical landforms and soils on which they occur and some statistics on the area remaining and that conserved in protected areas.



**Characteristic Species**

*Acacia stenophylla*, *Atriplex leptocarpa*, \**Bromus rubens*, *Cyperus gymnocaulos*, *Einadia nutans* ssp. *nutans*, *Enchylaena tomentosa* var. *tomentosa*, *Lachnagrostis filiformis*, *Muehlenbeckia florulenta*, *Senecio glossanthus*, *Setaria jubiflora*, \**Vulpia myuros* f. *myuros*.

**Landform:** River Edge & Floodplain  
**Soils:** Sandy Clay Loam to Medium Clay



**Area (pre-European):** 14327 ha. **Area (present):** 12080 ha **% pre-European:** 84% **Number of patches (size range):** 2461 (<1 to 1005 ha) **Area protected (% of present):** 2607 (22%)

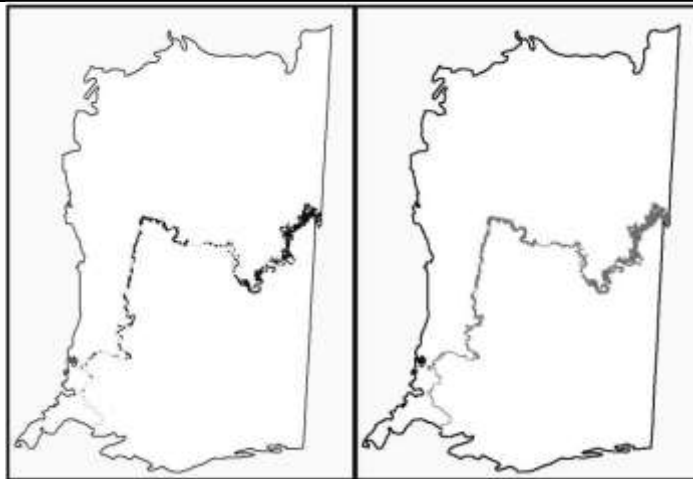
Figure 4. *Eucalyptus camaldulensis* var. *camaldulensis* River Red Gum Forest and Woodland.



**Characteristic Species**

*Acacia stenophylla*, \**Aster subulatus*, \**Cirsium vulgare*, *Cyperus gymnocaulos*, *Eclipta platyglossa*, *Einadia nutans* ssp. *nutans*, *Enchylaena tomentosa* var. *tomentosa*, *Muehlenbeckia florulenta*, *Phragmites australis*, *Setaria jubiflora*, \**Sonchus oleraceus*, *Wahlenbergia fluminalis*.

**Landform:** River Flat & Floodplain  
**Soils:** Sandy Loam to Medium Clay



**Area (pre-European):** 10040 ha. **Area (present):** 10040 ha. **% pre-European:** 100% **Number of patches (size range):** 614 (<1 to 335 ha) **Area protected (% of present):** 4057 (40%)

Figure 5. *Eucalyptus camaldulensis* var. *camaldulensis* River Red Gum, *E. largiflorens* River Box Woodland.



**Characteristic Species**

*Brachycome lineariloba*, \**Bromus rubens*, *Crassula colorata* var. *acuminata*, *Einadia nutans* ssp. *nutans*, *Enchylaena tomentosa* var. *tomentosa*, *Disphyma crassifolium* ssp. *clavellatum*, *Muehlenbeckia florulenta*, *Sclerolaena tricuspis*, *Setaria jubiflora*.

**Landform:** River Flat & Floodplain  
**Soils:** Sandy Loam to Medium Clay



**Area (pre-European):** 17005 ha. **Area (present):** 15824 ha **% pre-European:** 93% **Number of patches (size range):** 16311 (<1 to 319 ha) **Area protected (% of present):** 6561 (41%)

Figure 6. *Eucalyptus largiflorens* River Box Low Woodland.



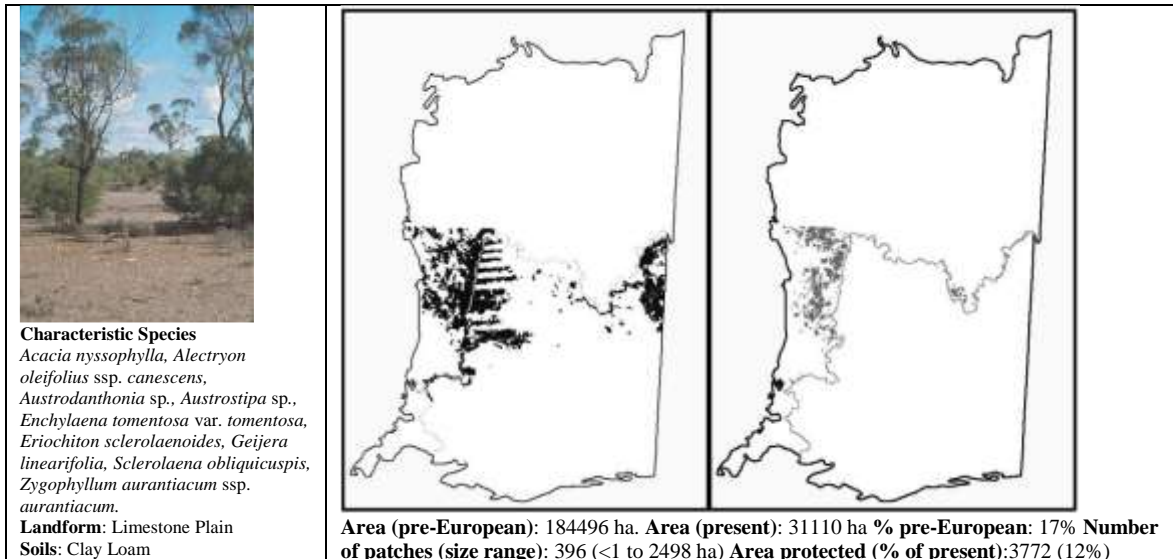


Figure 7. *Myoporum platycarpum* ssp. Sugarwood Low Woodland.

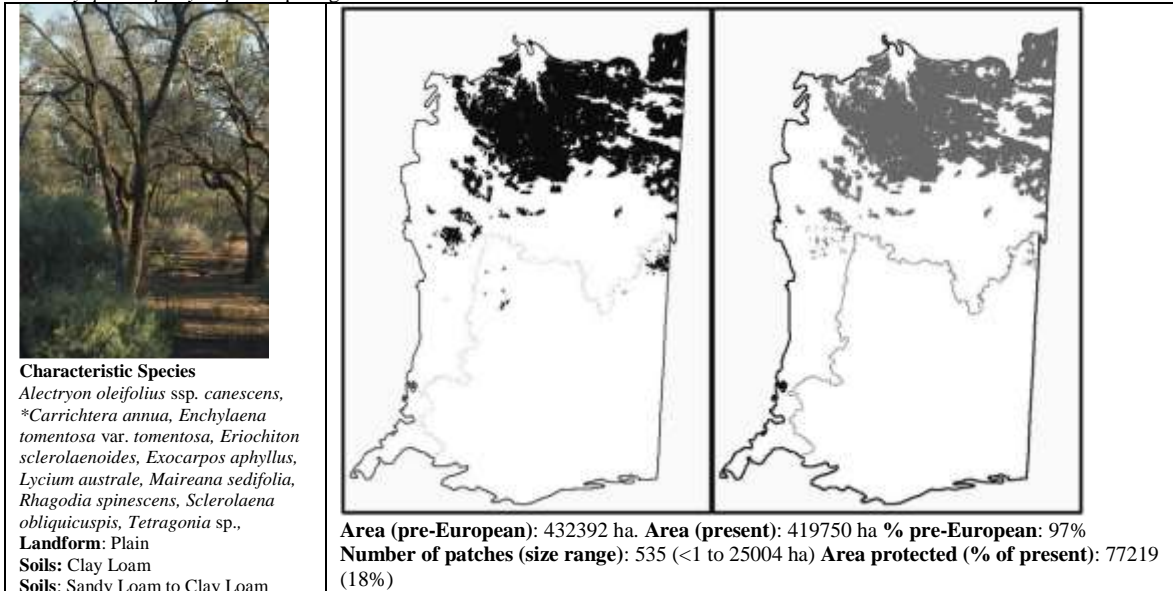


Figure 8. *Casuarina pauper* Black Oak Low Woodland.

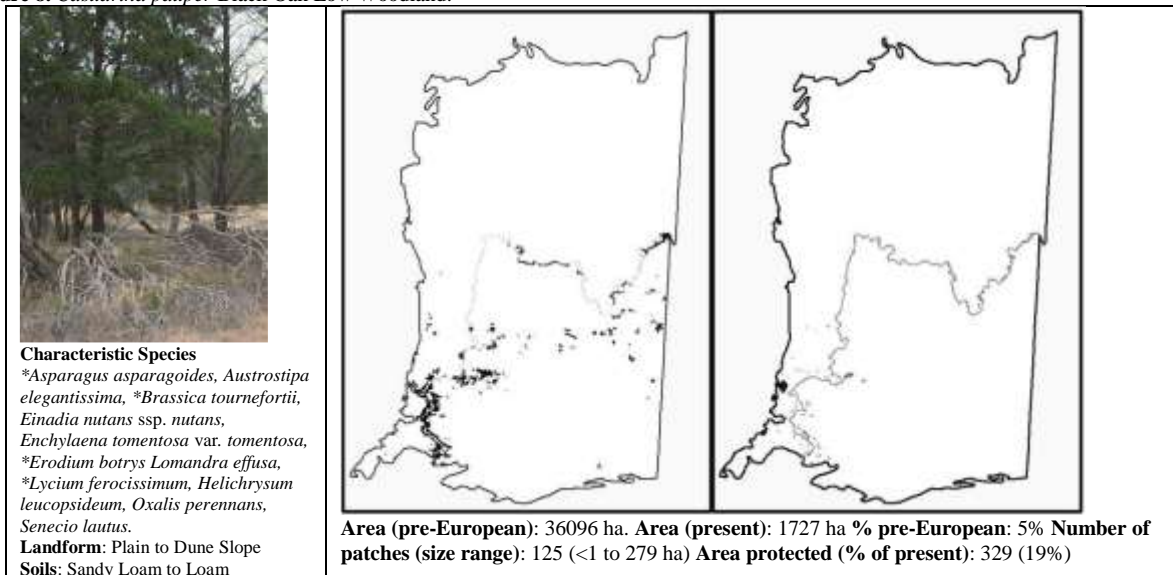


Figure 9. *Callitris gracilis* Southern Cypress Pine Low Open Woodland.

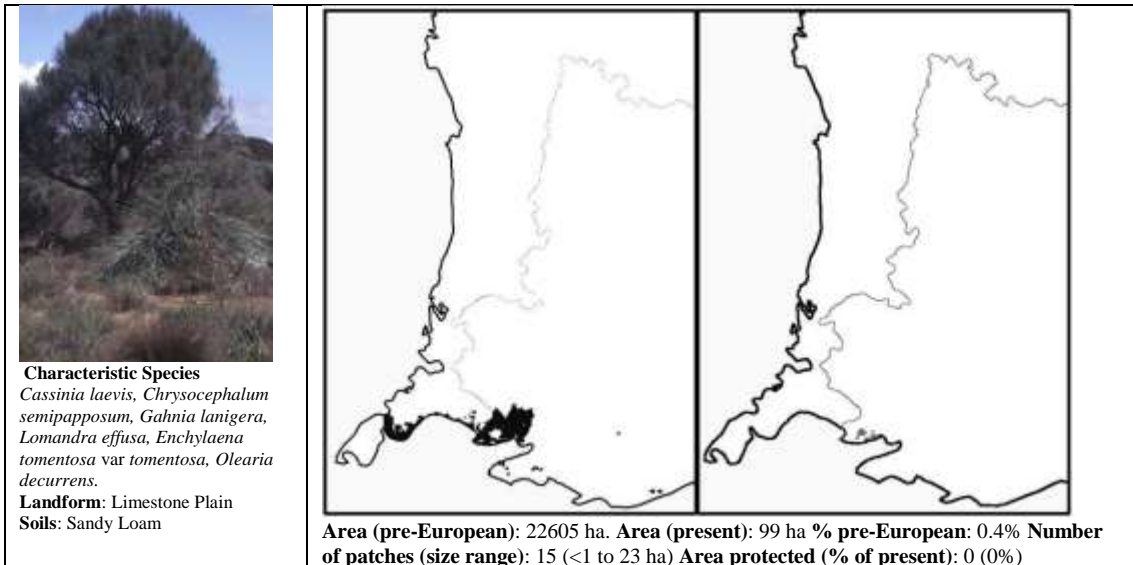


Figure 10. *Allocasuarina verticillata* Drooping Sheoak Low Woodland.

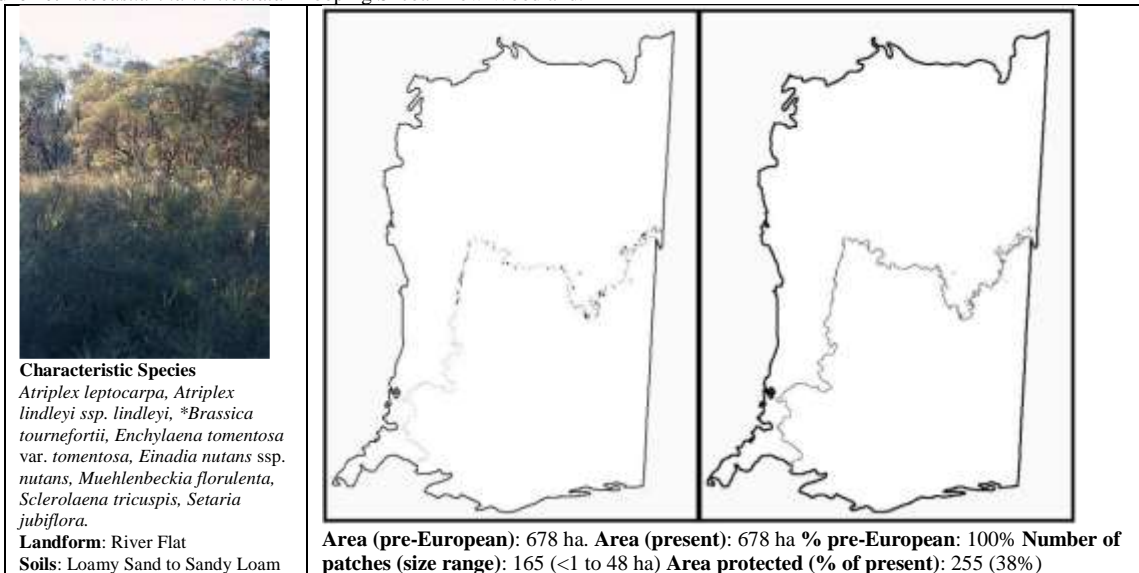


Figure 11. *Acacia stenophylla* River Cooba Low Open Woodland.

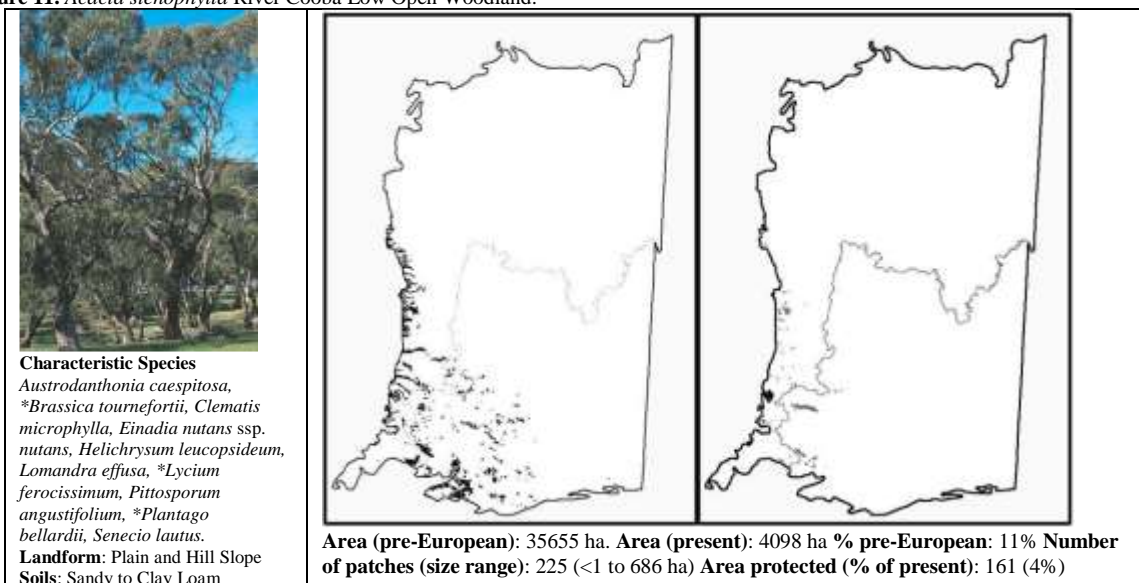
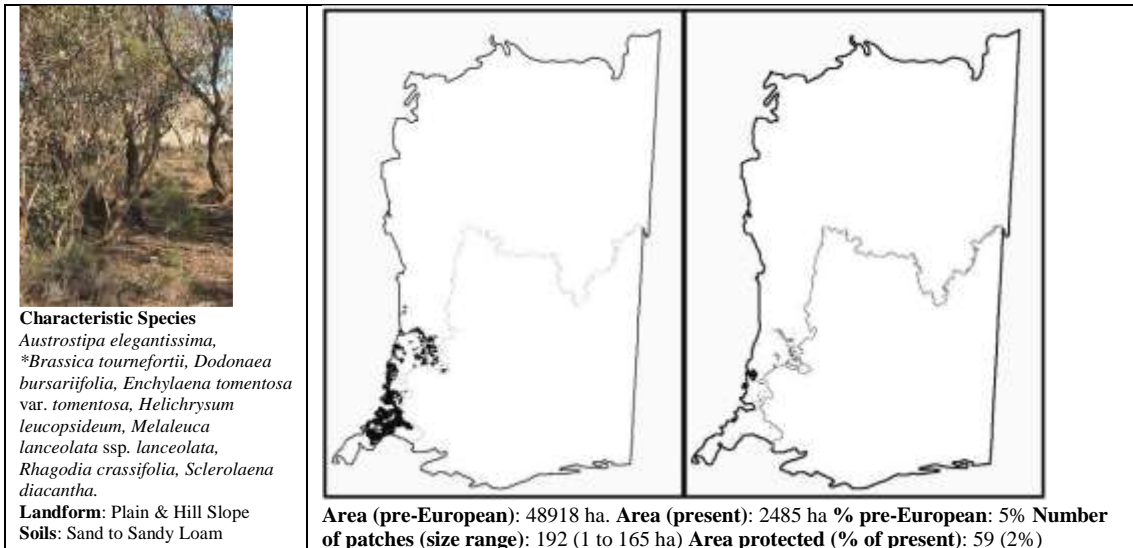
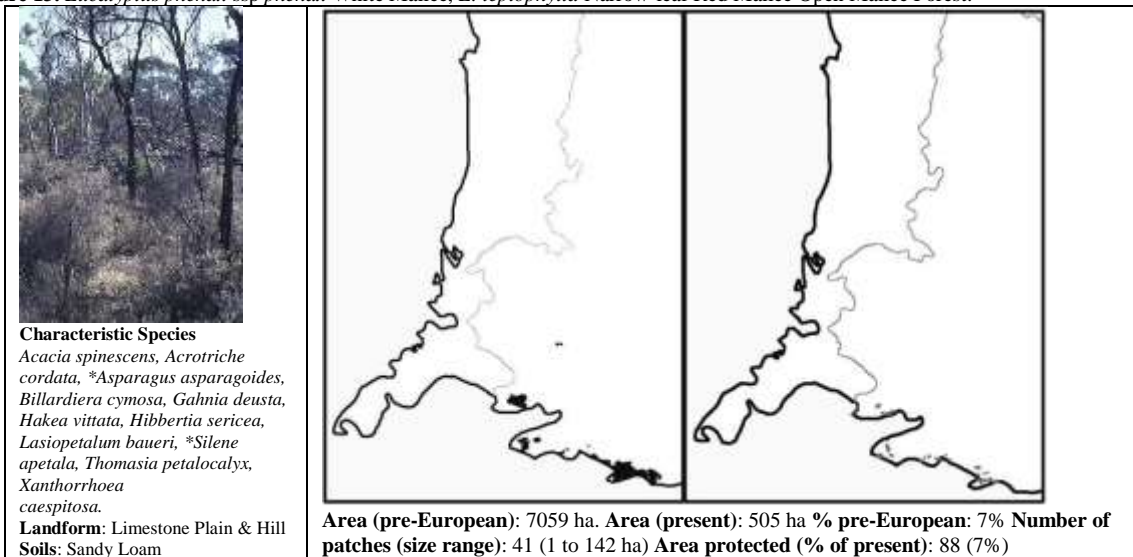


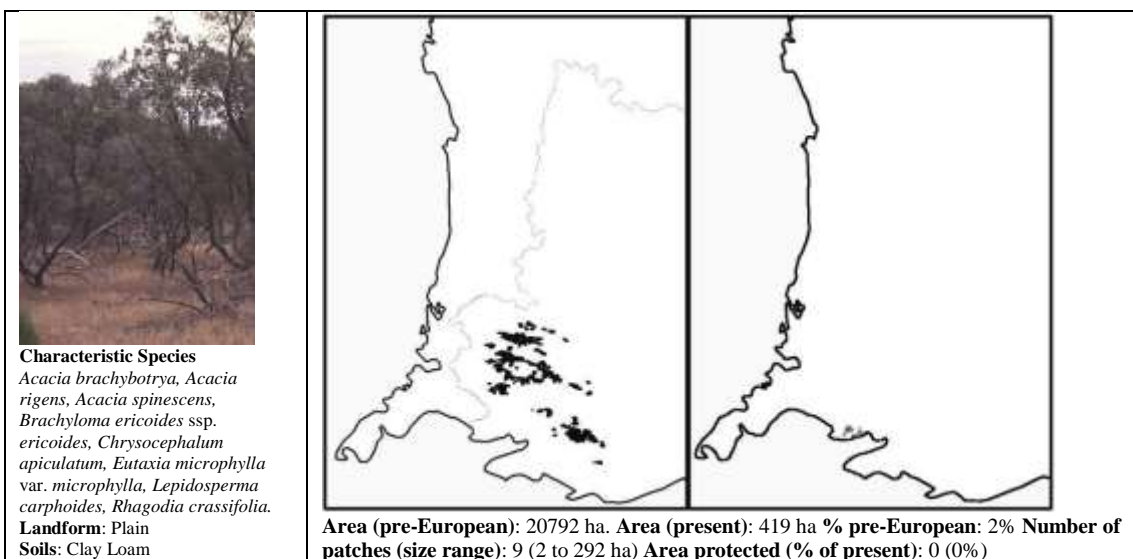
Figure 12. *Eucalyptus porosa* Mallee Box Low Open Woodland.



**Figure 13.** *Eucalyptus phenax* ssp. *phenax* White Mallee, *E. leptophylla* Narrow-leaf Red Mallee Open Mallee Forest.



**Figure 14.** *Eucalyptus diversifolia* Coastal White Mallee Mallee Forest.



**Figure 15.** *Eucalyptus yalataensis* Yalata Mallee, *E. dumosa* White Mallee, *E. gracilis* Yorrell Mallee.



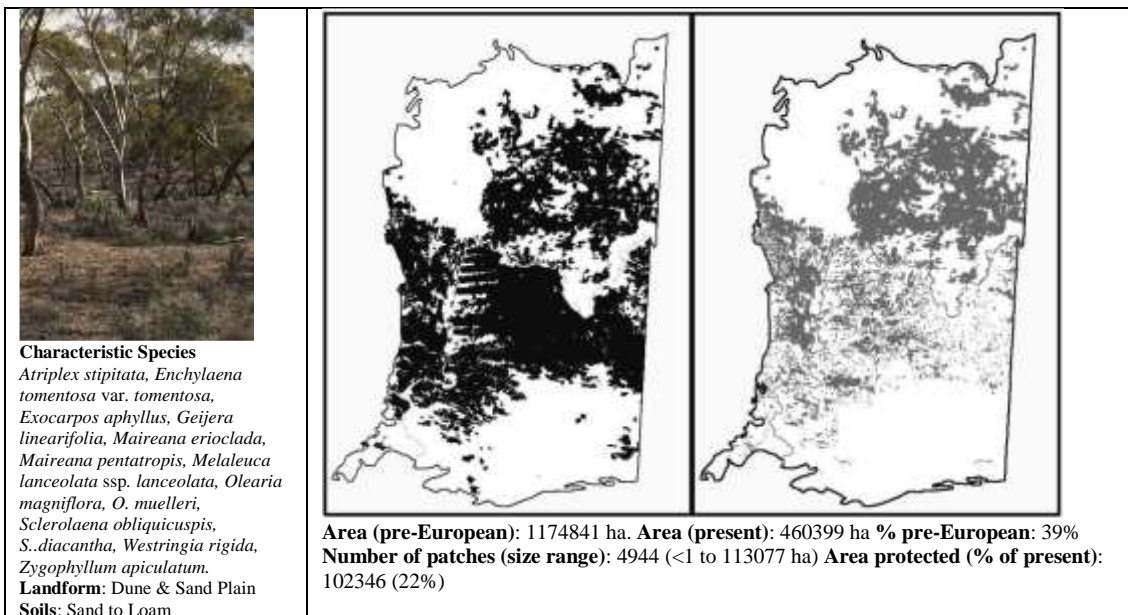


Figure 16. *Eucalyptus gracilis* Yorrell, *E. oleosa* Red Mallee Very Open Mallee.

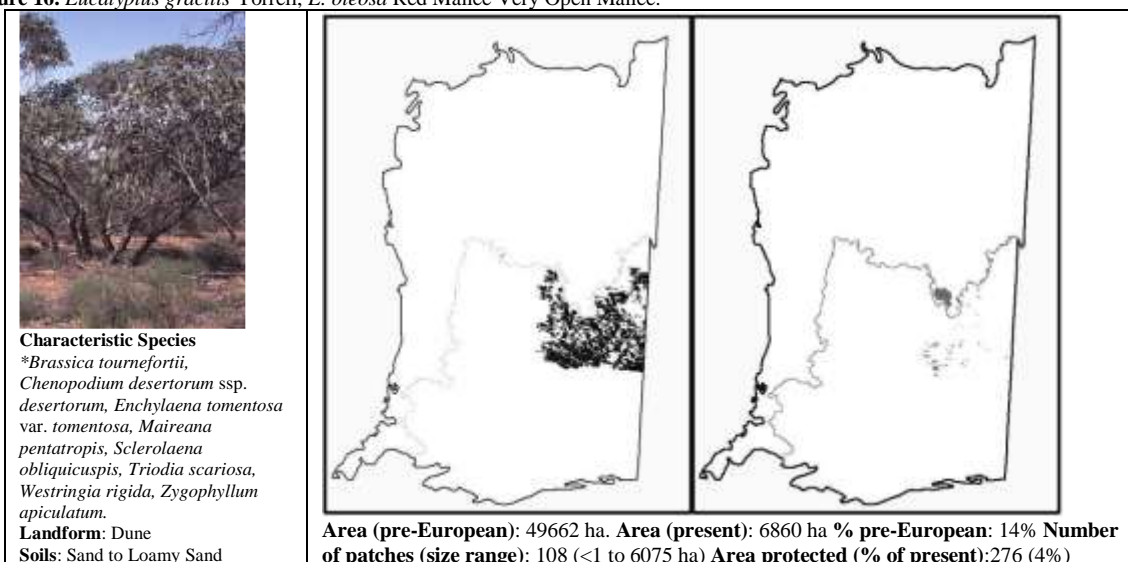


Figure 17. *Eucalyptus cyanophylla* Blue-leaf Mallee Open mallee

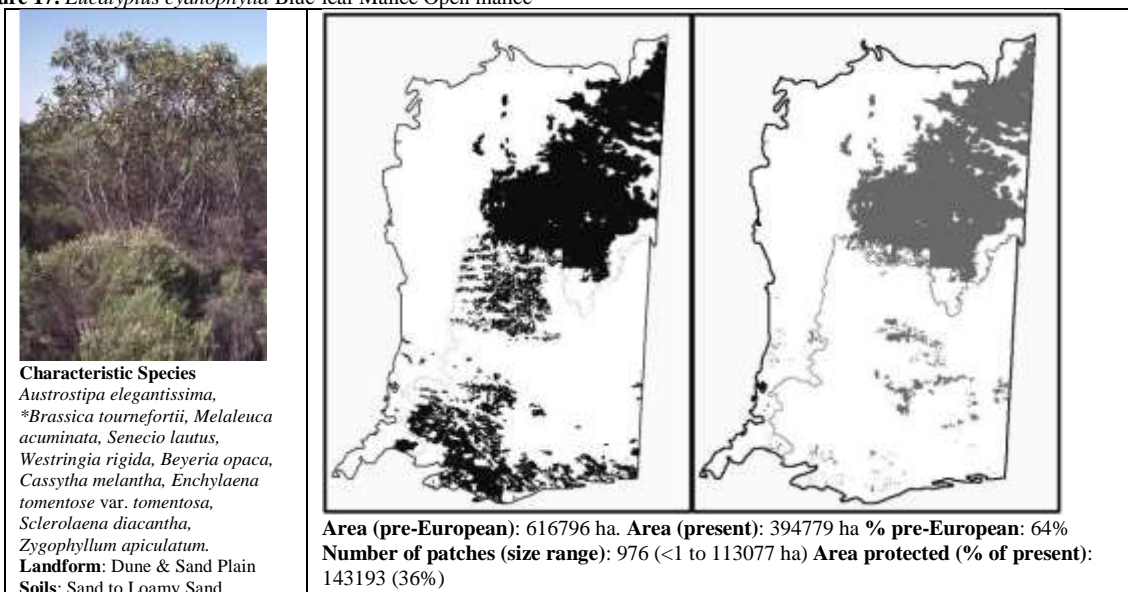
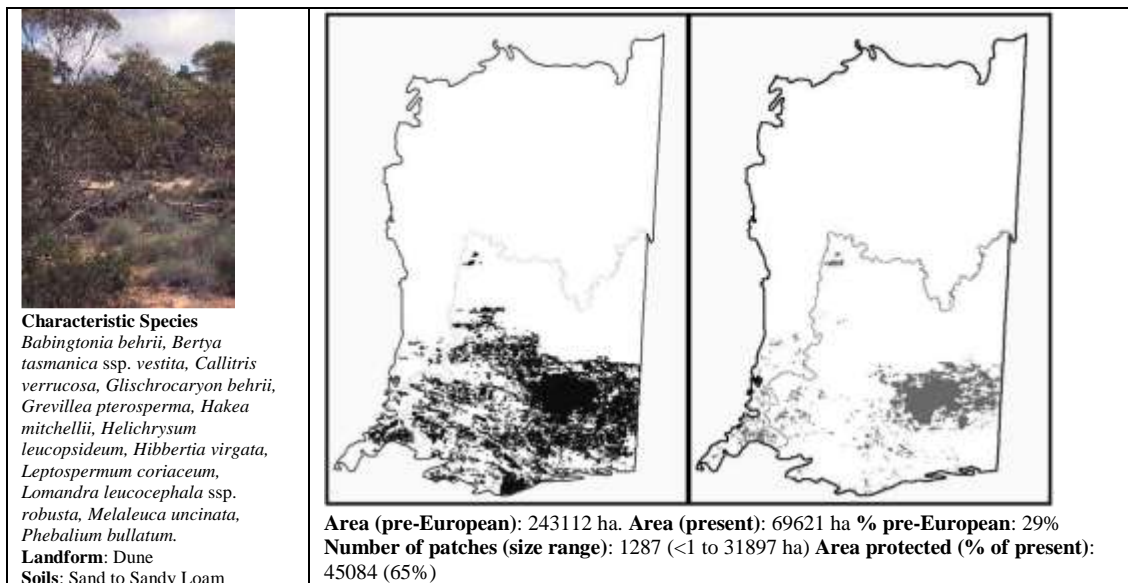
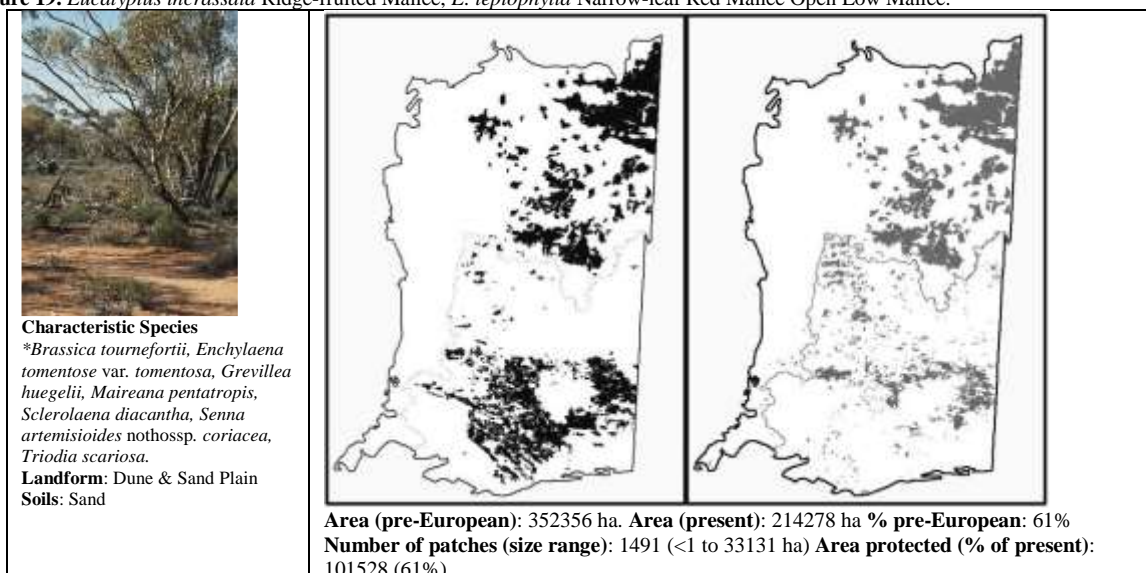


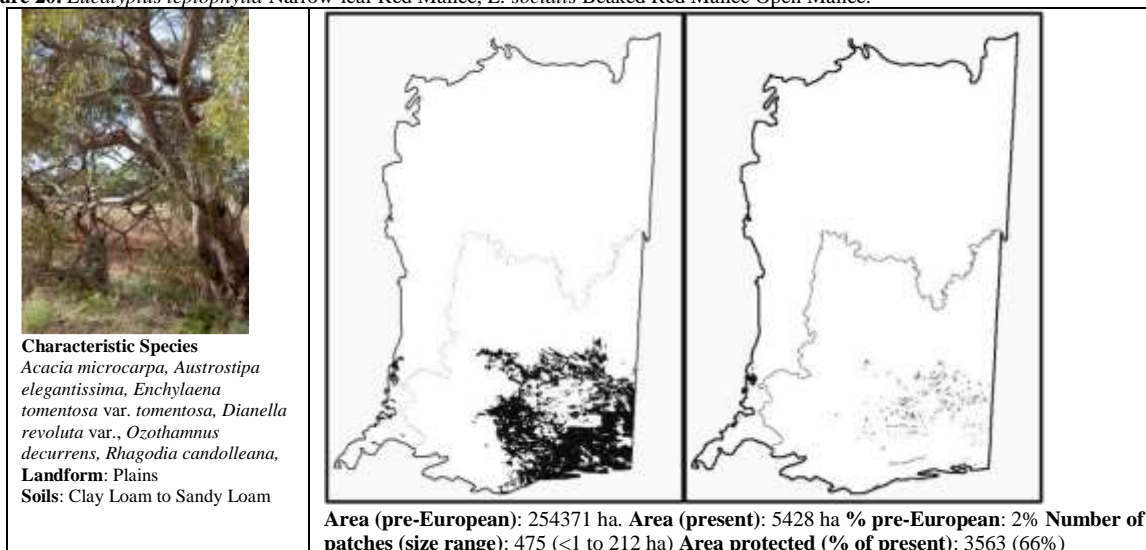
Figure 18. *Eucalyptus dumosa* White Mallee, *E. socialis* Beaked Red Mallee, *E. leptophylla* Narrow-leaf Red Mallee Mallee.



**Figure 19.** *Eucalyptus incrassata* Ridge-fruited Mallee, *E. leptophylla* Narrow-leaf Red Mallee Open Low Mallee.



**Figure 20.** *Eucalyptus leptophylla* Narrow-leaf Red Mallee, *E. socialis* Beaked Red Mallee Open Mallee.



**Figure 21.** *Eucalyptus calycogona* Square-fruit Mallee, *E. dumosa* White Mallee Very Open Mallee.



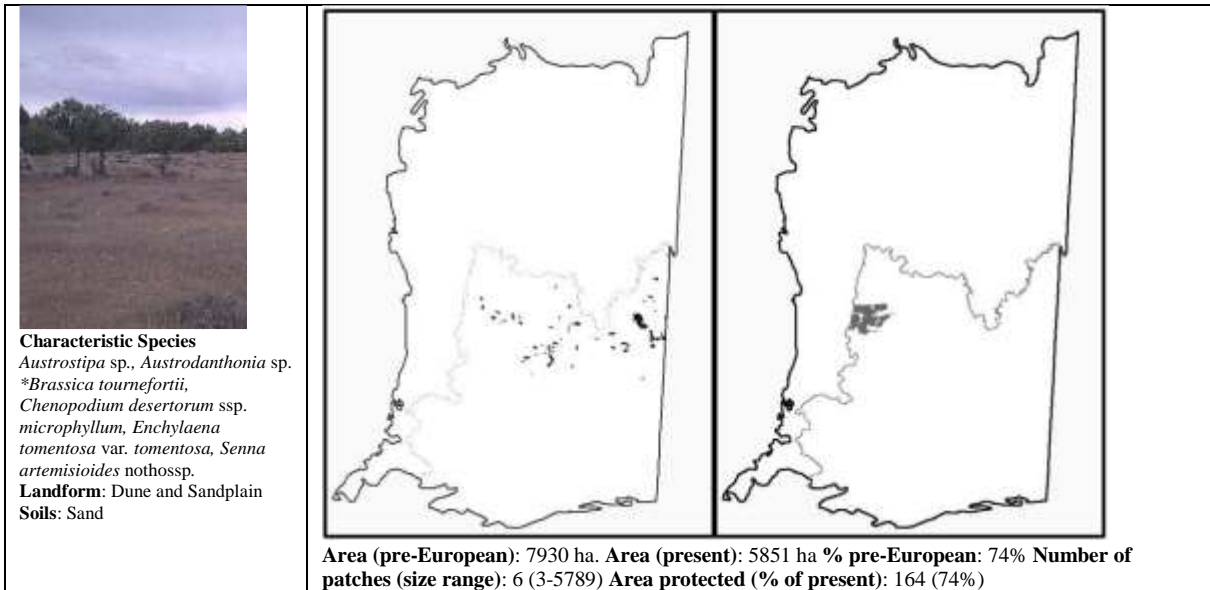


Figure 22. *Alectryon oleifolius* ssp. *canescens* Bullock Bush Tall Shrubland.

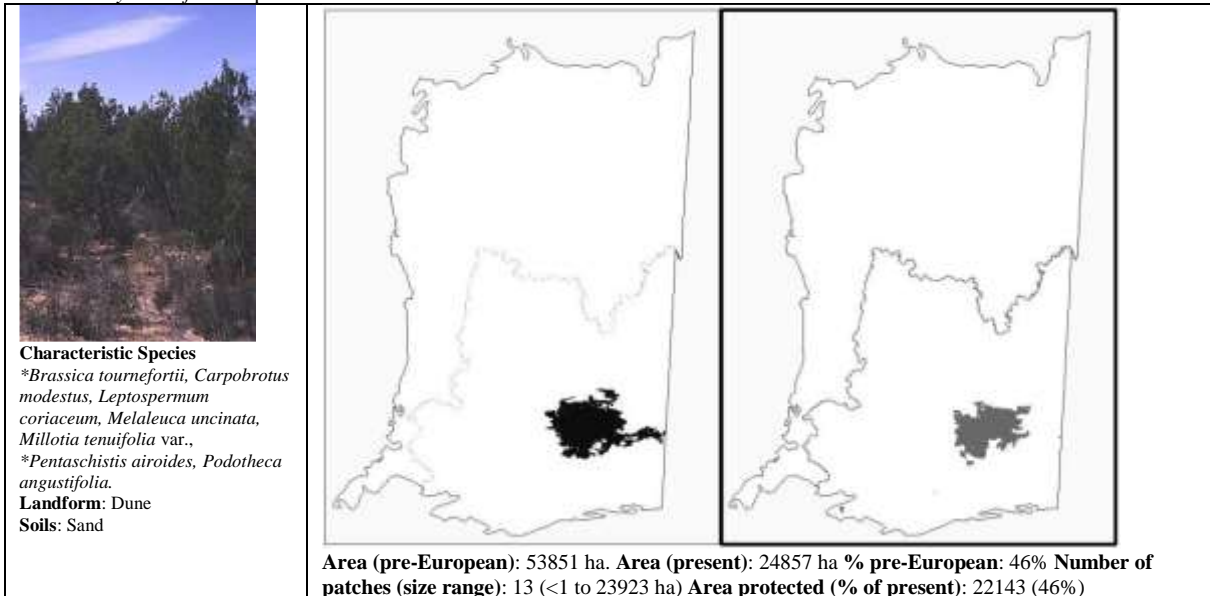


Figure 23. *Callitris verrucosa* Scrub Cypress Pine Tall Open Shrubland.

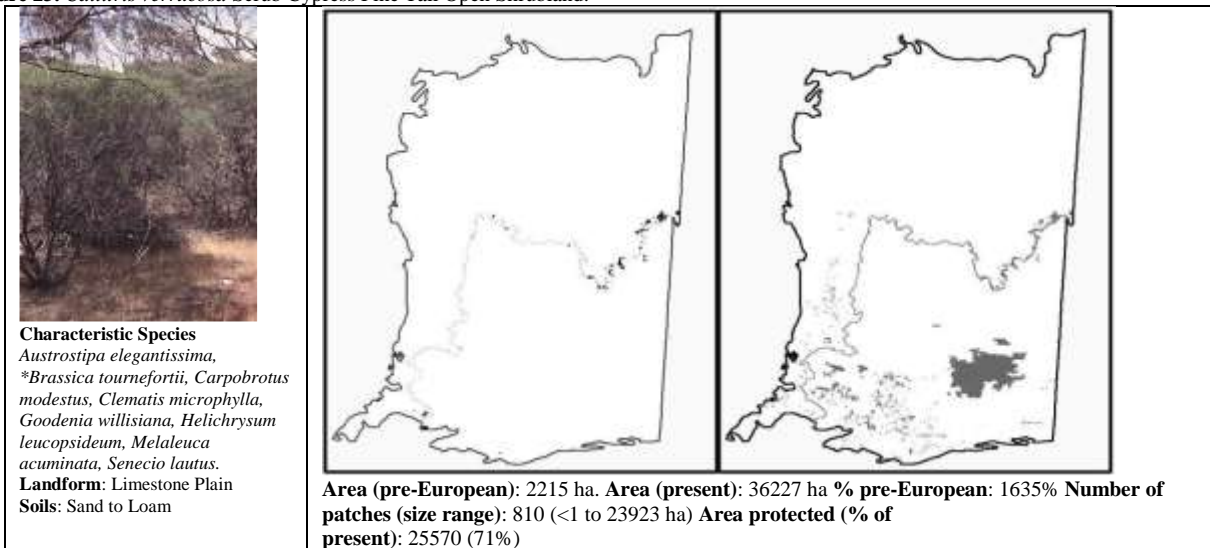


Figure 24. *Melaleuca lanceolata* Dryland Tea-tree Tall Open Shrubland.

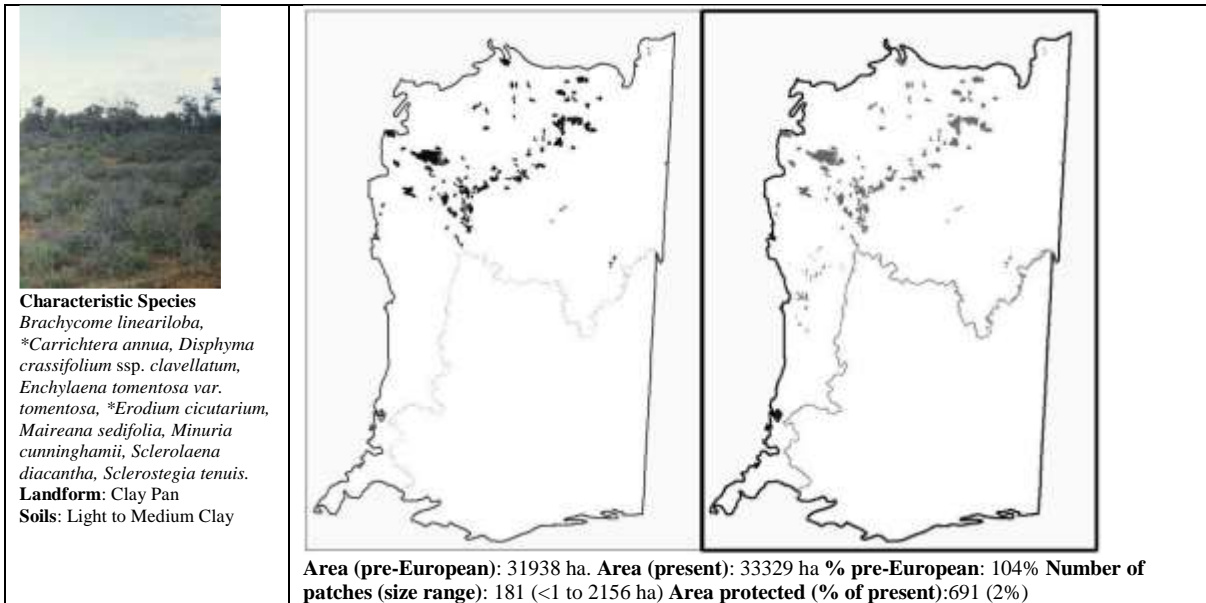


Figure 25. *Lycium australe* Australian Boxthorn, *Nitraria billardieri* Nitre-bush Open Shrubland.

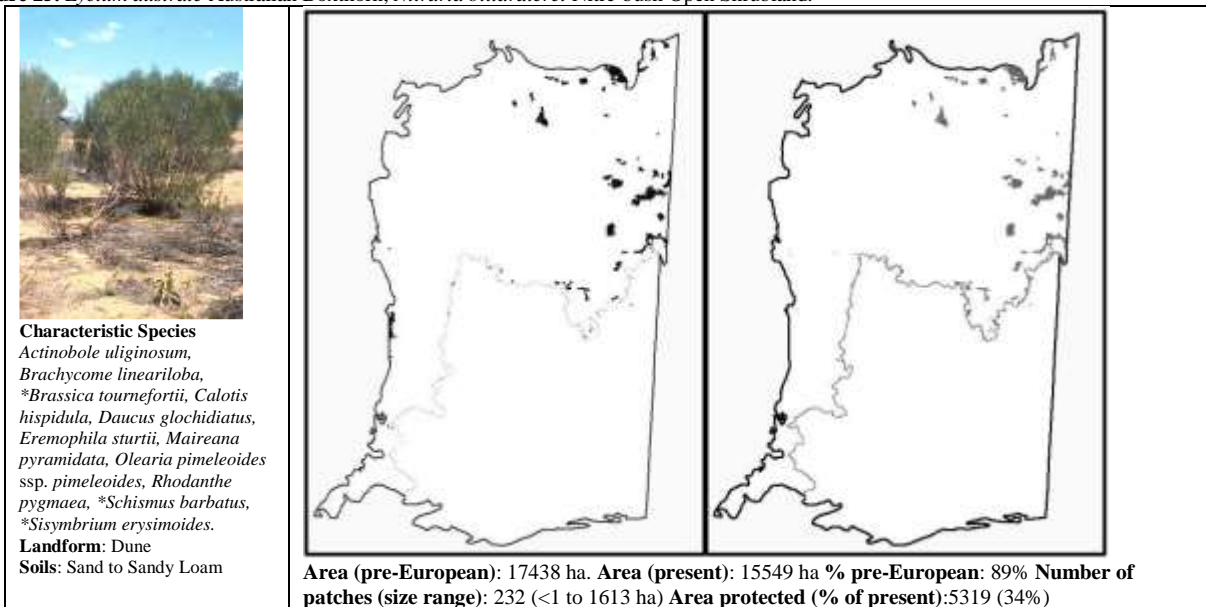


Figure 26. *Dodonaea viscosa* ssp. *angustissima* Narrow-leaf Hop-bush Open Shrubland.

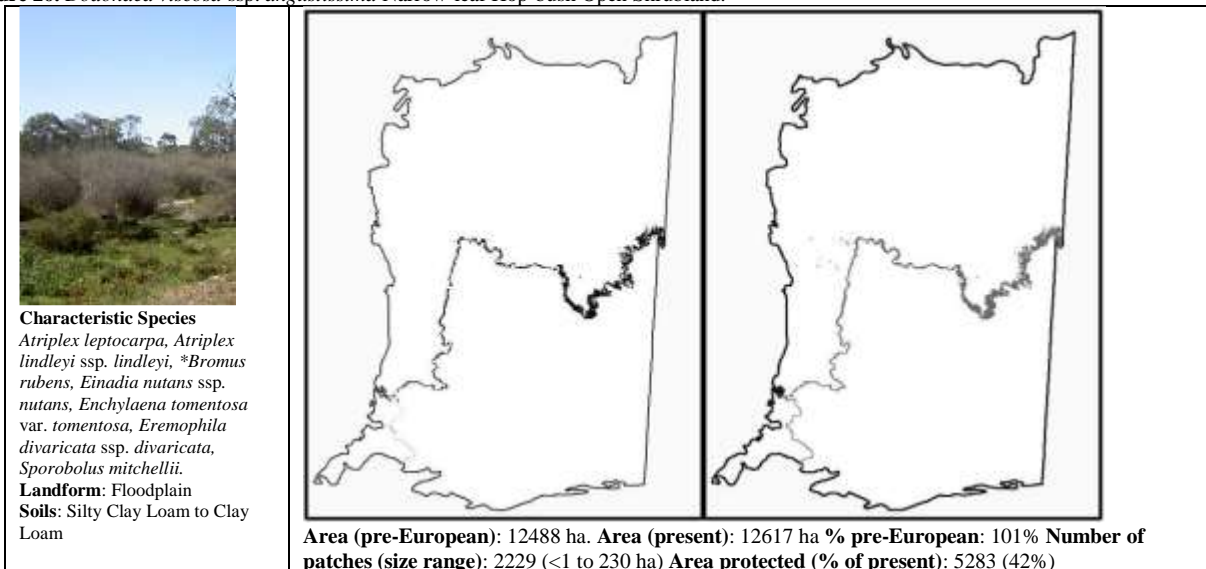


Figure 27. *Muehlenbeckia florulenta* Lignum Shrubland.

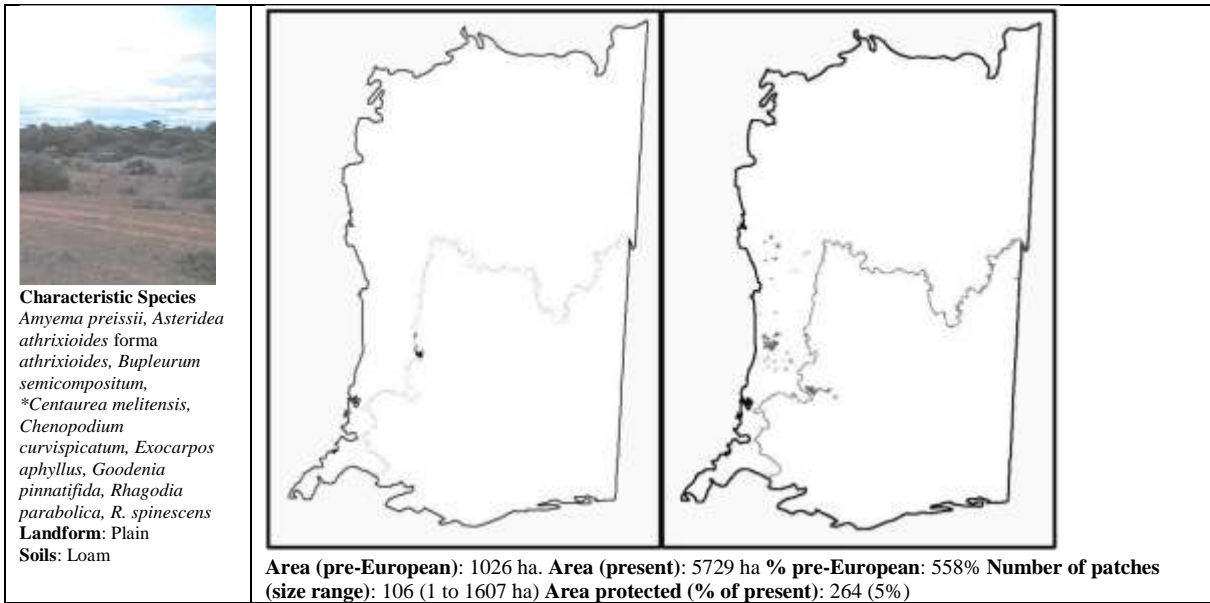


Figure 28. *Acacia nyssophylla* Spine Bush Shrubland.

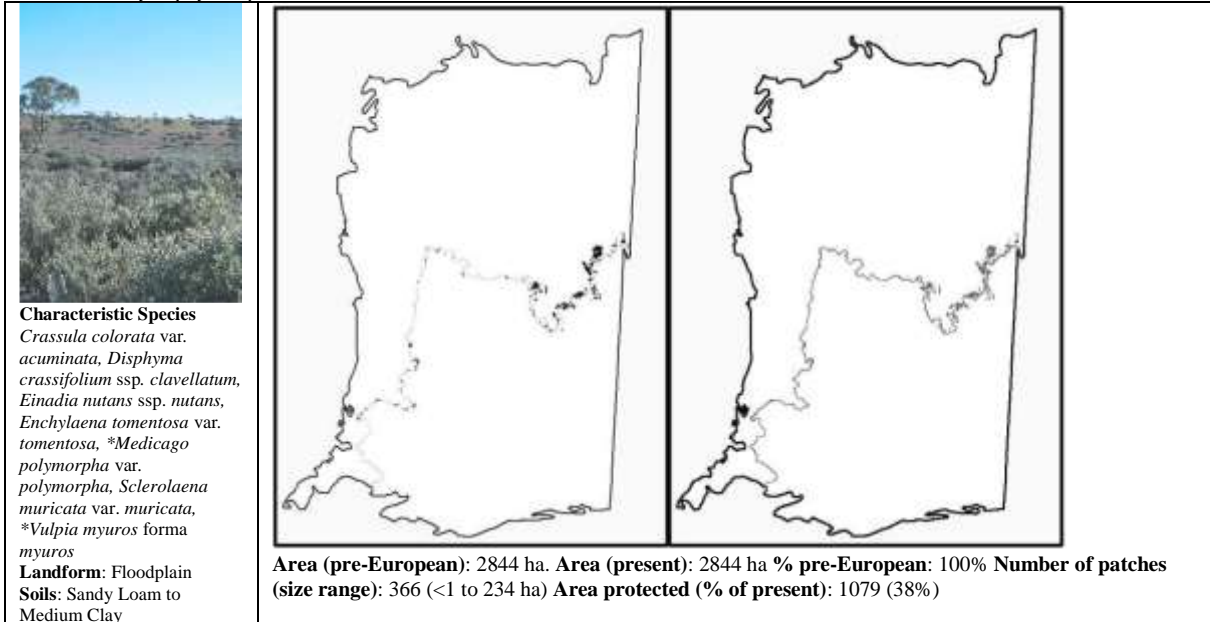


Figure 29. *Atriplex rhagodioides* River Saltbush Shrubland.

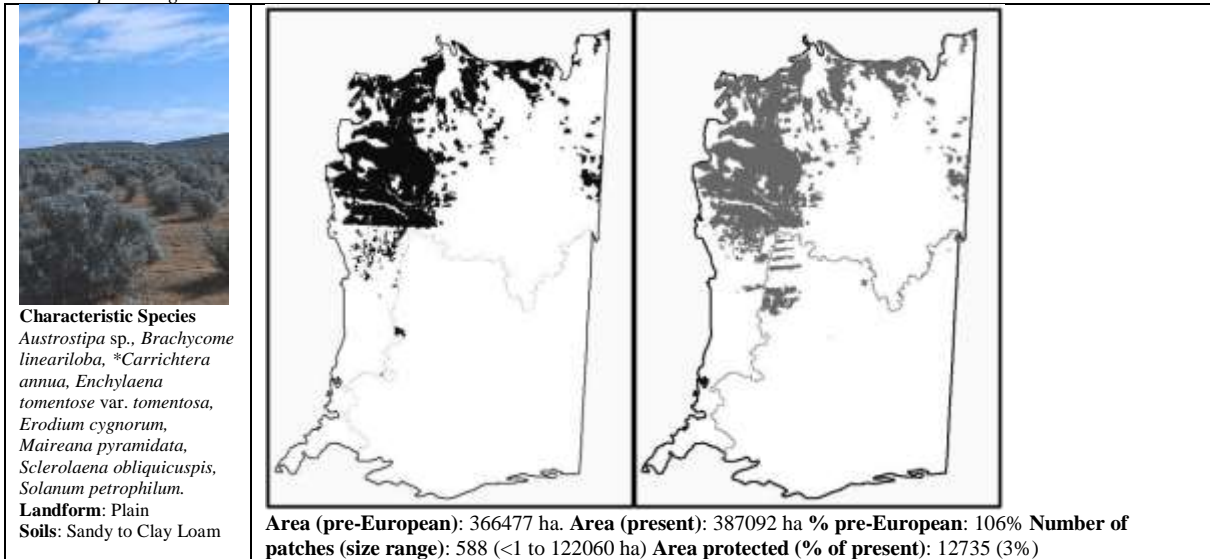
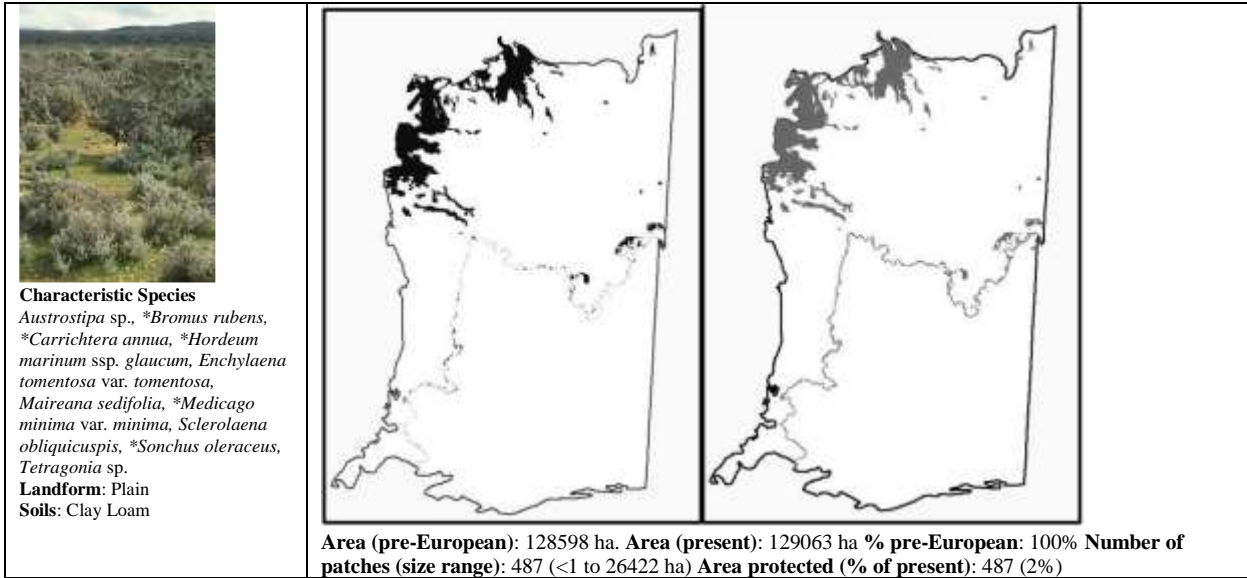
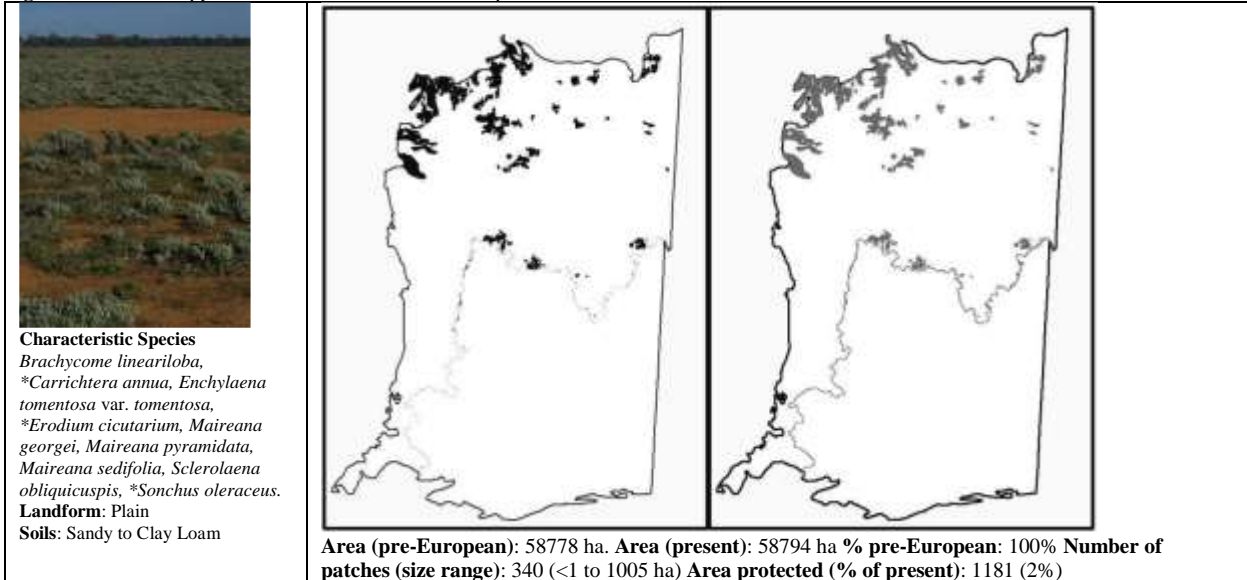


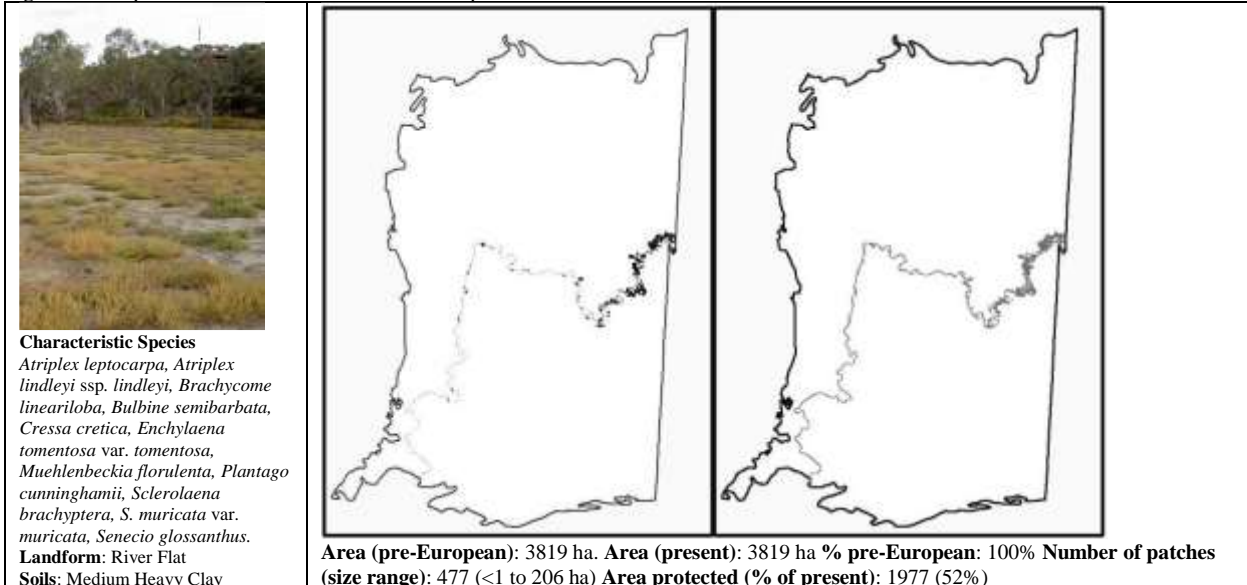
Figure 30. *Maireana sedifolia* Bluebush Low Open Shrubland.



**Figure 31.** *Maireana pyramidata* Black Bluebush Low Open Shrubland



**Figure 32.** *Atriplex vesicaria* Bladder Saltbush Low Open Shrubland.



**Figure 33.** *Atriplex lindleyi* ssp. *lindleyi* Baldoo Low Shrubland.



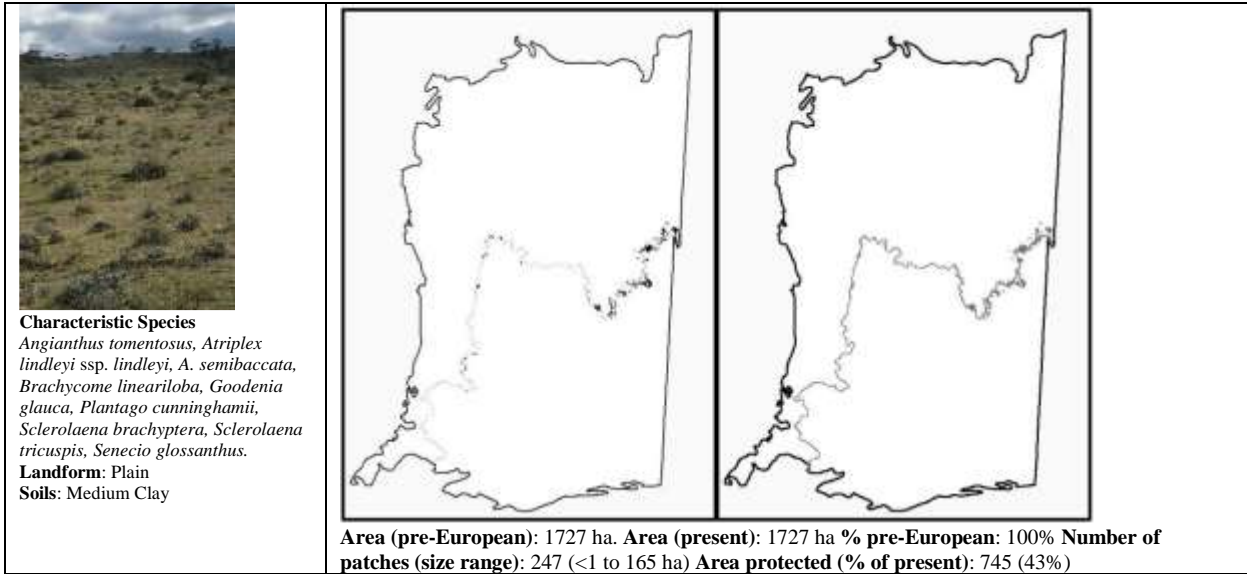


Figure 34. *Sclerolaena* spp. Bindyi Low Open Shrubland.

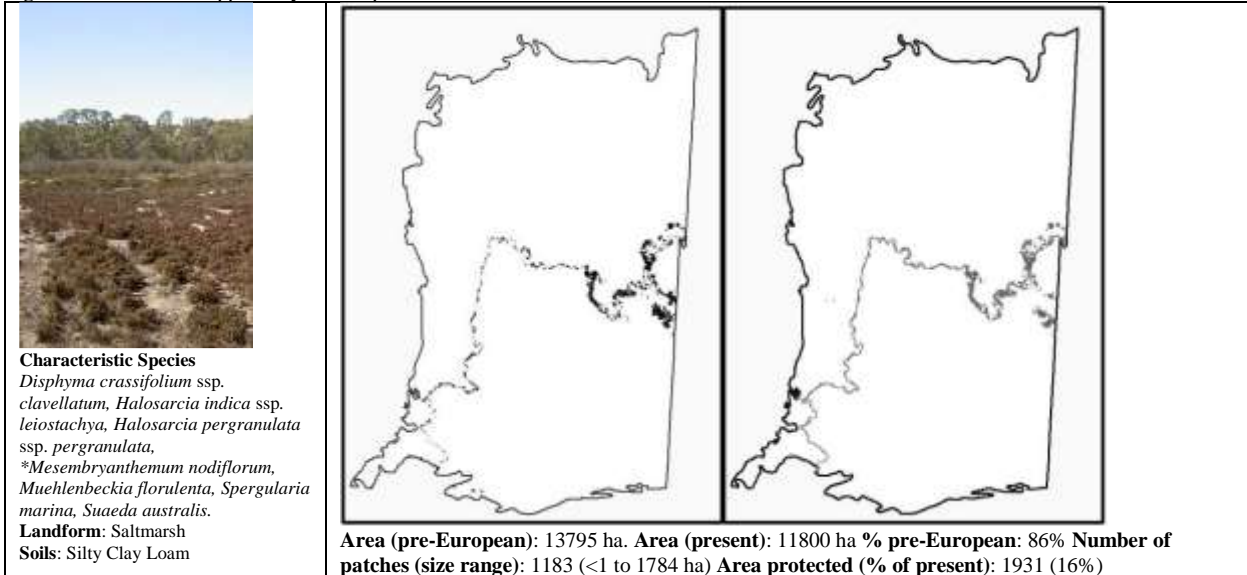


Figure 35. *Halosarcia* spp. Samphire Low Very Open Shrubland.

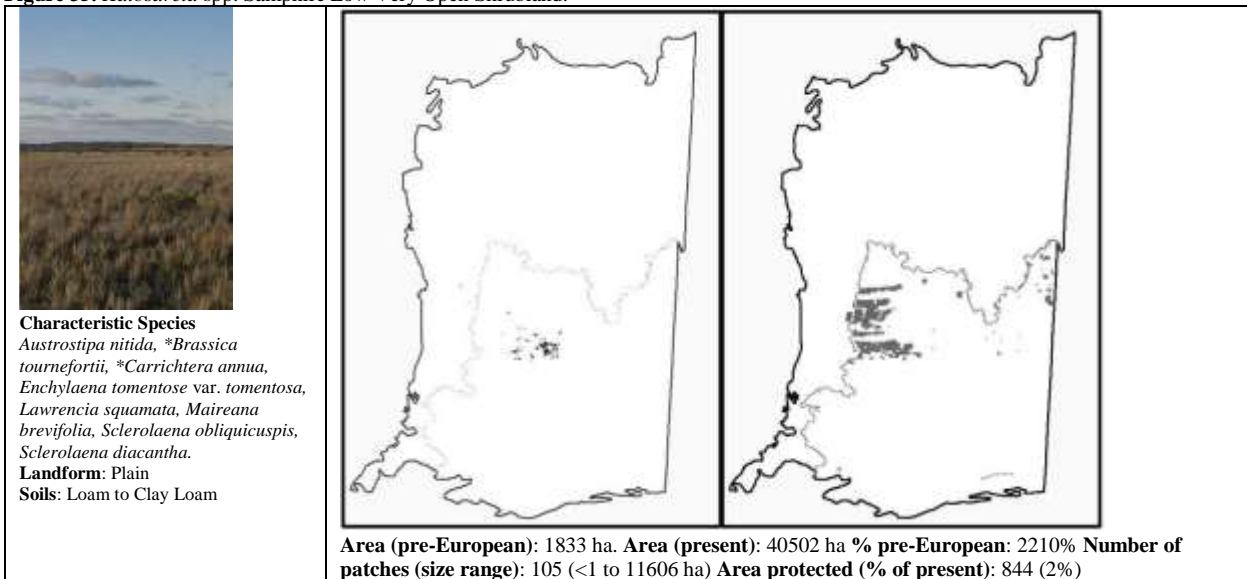
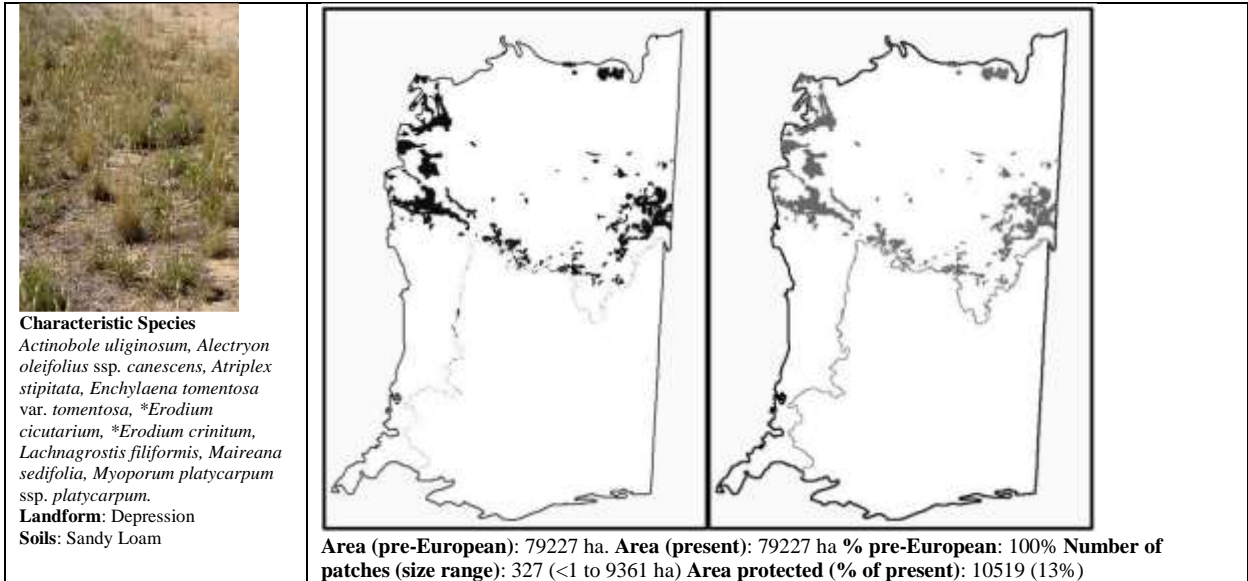
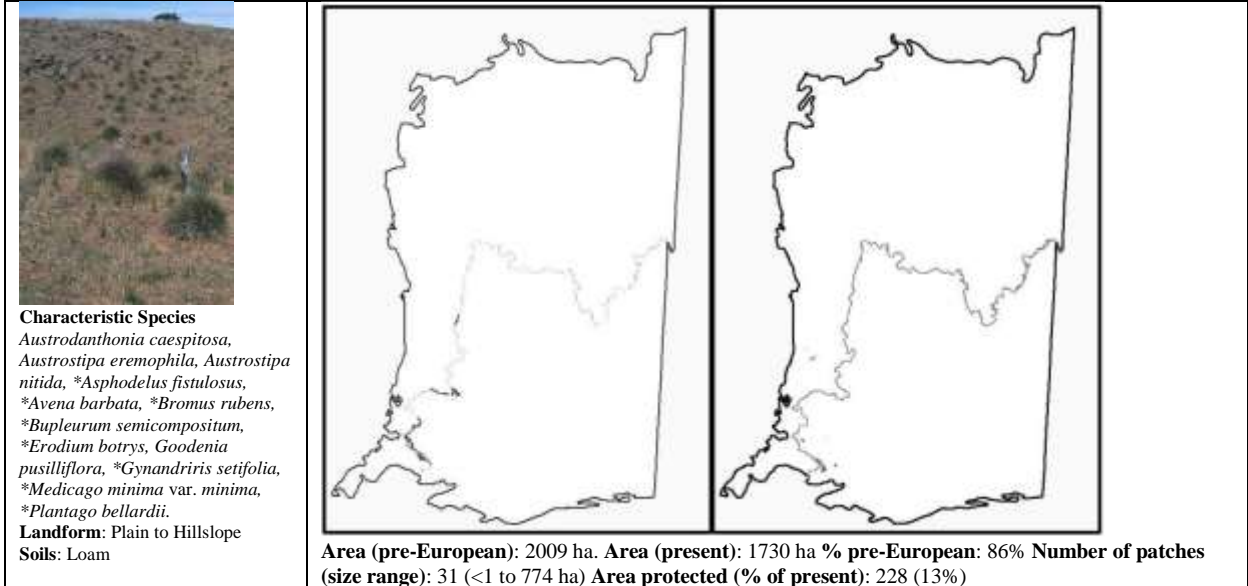


Figure 36. *Austrostipa* spp. Spear-grass Tussock Grassland.

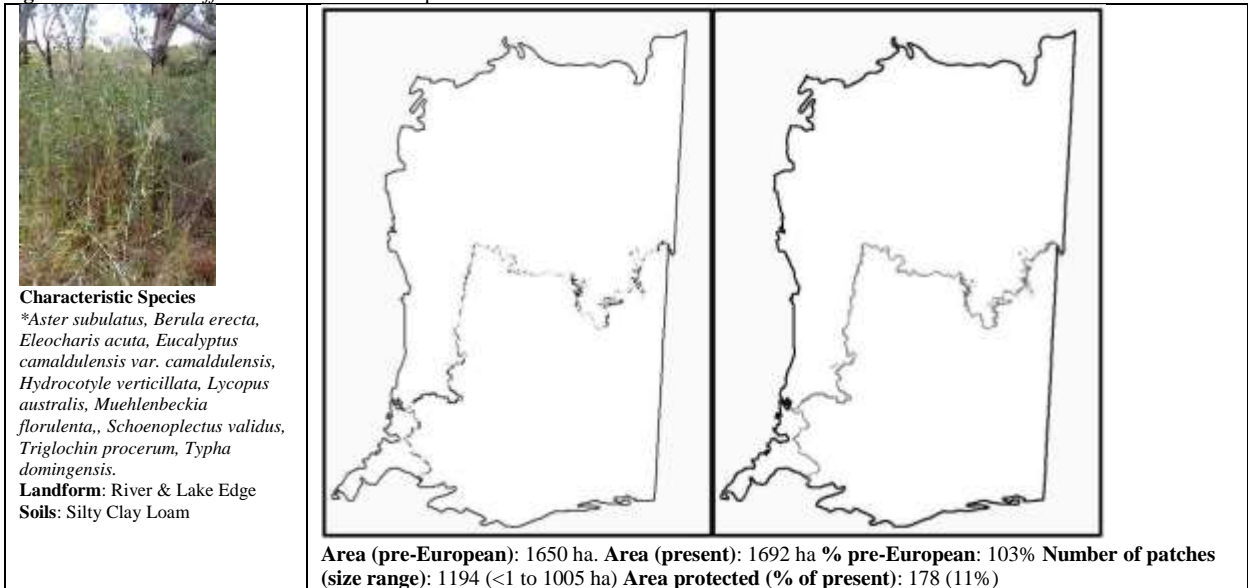




**Figure 37.** *Enneapogon* sp. Umbrella Grass Tussock Grassland.



**Figure 38.** *Lomandra effusa* Scented Mat-rush Open Tussock Grassland.



**Figure 39.** *Phragmites australis* Common Reed, *Typha* spp. Bullrush Sedgeland.

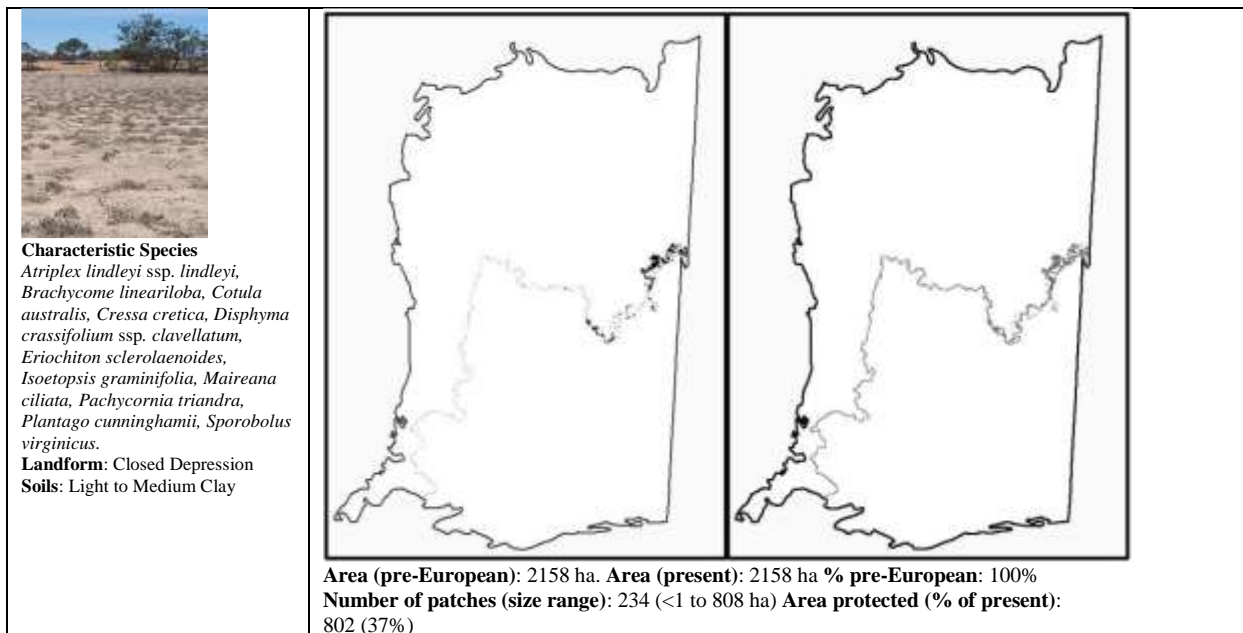


Figure 40. Herbland.

## DISCUSSION

Since European settlement, the vegetation of the Murray Mallee has dramatically changed. A comparison between current and pre-European vegetation cover in Figures 4–40 indicates more favourable agricultural areas, such as those of higher rainfall, more lightly wooded areas or soils of more favourable fertility, have been selectively developed for agriculture (e.g. *Eucalyptus calycogona*, *E. dumosa* Mallee; *E. dumosa*, *E. socialis*, *E. leptophylla* Mallee; and *E. socialis*, *E. leptophylla* Mallee).

Within the pastoral area, the extensive areas of former *Myoporum platycarpum* Woodland noted in the Pre-European mapping have in turn been largely reduced to *Austrostipa* Grassland, *Maireana sedifolia* Low Shrubland or *Alectryon oleifolius* ssp. *canescens* Tall Shrubland, through grazing of domestic stock and rabbits. Some of these badly over-grazed areas have subsequently been re-colonised by spiny and unpalatable shrubs reflected in the expansion of areas of *Acacia nyssophylla* Shrubland.

Along much of the Murray Valley, the former natural vegetation has been completely replaced by areas now utilised for irrigated agriculture, while much of what remains of the once beautiful and extensive areas of *Eucalyptus camaldulensis* var. *camaldulensis* and *E. largiflorens* Forests and Woodlands that so impressed Charles Sturt in 1830 are now either dead from the effects of salinity and altered flood regimes, or are in serious decline. This has also led to a significant expansion of the salt-tolerant vegetation communities such as *Halosarcia* spp. Low Very Open Shrubland and various forms of Herbland.

We are now at a point where much of the native vegetation still remaining is threatened by fragmentation and isolation in unsustainable small blocks. Many areas have salinity problems, environmental weeds and over-grazing by feral animals, domestic stock and the very large numbers of kangaroos that this altered landscape now sustains. The remaining Mallee communities in particular are also threatened by changed fire regimes. As described in the Regional Biodiversity Plan (Kahrimanis et al. 2001), we are now at the point where every tiny remnant of the original natural vegetation of this extensive area must be valued and actively managed. This must be part of a major effort to restore some semblance of connectivity and ecological integrity across the whole landscape by a massive effort at re-vegetation using local native species to reconnect the fragments. Priority should be given to expanding, and where possible, reconnecting the few remaining plant communities on the more fertile agricultural lands before they are lost from the area forever.

## ACKNOWLEDGEMENTS

Our thanks to the hundreds of professional and volunteer biologists who participated in the four major biological surveys in the area covered by this natural history. Without the large number of systematic vegetation quadrats sampled and the subsequent vegetation mapping, the analysis described in this chapter could not have been attempted.

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## APPENDIX. A LIST OF VASCULAR PLANTS RECORDED FROM THE MURRAYLANDS AND RIVERLAND REGIONS

Plant taxonomy follows the Census of South Australian Vascular Plants (Barker et al. 2005). Introduced species are preceded with an asterisk (\*).

Common names are from Jessop and Toelken (1986) and/or the SAFLORA database.

Proclaimed Pest Plants under the Animal and Plant Control (Agricultural Protection and other Purposes) Act 1986 are indicated with a Y.

Conservation Status codes: V = Vulnerable, R = Rare. Codes for Australia are based on the December 2000 listing of species under the Environment Protection and Biodiversity Conservation Act 1999; those for South Australia are based on Schedules of the National Parks and Wildlife Act 1972 (SA) as amended in 2000. There have however been some modifications to the current legislated listings to incorporate more recent taxonomic changes and increases in knowledge of some species status.

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
<b>LYCOPODIACEAE</b>				
<i>Phylloglossum drummondii</i> Kunze	Pygmy Clubmoss			R
* <i>Phyllopodium cordatum</i> (Thunb.)O.M.Hilliard				
<b>ISOETACEAE</b>				
<i>Isoetes drummondii</i> A.Braun ssp. <i>drummondii</i>	Plain Quillwort			
<b>OPHIGLOSSACEAE</b>				
<i>Ophioglossum lusitanicum</i> L.	Austral Adder's-tongue			
<b>ADIANTACEAE</b>				
<i>Adiantum aethiopicum</i> L.	Common Maiden-hair			
<i>Adiantum capillus-veneris</i> L.	Dainty Maiden-hair			V
<i>Cheilanthes austrotenuifolia</i> H.M.Quirk & T.C.Chambers	Annual Rock-fern			
<i>Cheilanthes distans</i> (R.Br.)Mett.	Bristly Cloak-fern			
<i>Cheilanthes lasiophylla</i> Pic.Serm.	Woolly Cloak-fern			
<i>Cheilanthes sieberi</i> Kunze ssp. <i>sieberi</i>	Narrow Rock-fern			
<b>PTERIDACEAE</b>				
<i>Pteridium esculentum</i> (G.Forst.)Cockayne	Bracken			
<i>Pteris tremula</i> R.Br.	Tender Brake			R
<b>ASPLENIACEAE</b>				
<i>Asplenium flabellifolium</i> Cav.	Necklace Fern			
<i>Pleurosorus rutifolius</i> (R.Br.)Fee	Blanket Fern			
<i>Pleurosorus subglandulosus</i> (Hook. & Grev.)Tindale	Clubbed Blanket Fern			
<b>THELYPTERIDACEAE</b>				
<i>Christella dentata</i> (Forssk.)Brownsey & Jermy	Soft Shield-fern			R
<b>MARSILEACEAE</b>				
<i>Marsilea costulifera</i> D.L.Jones	Narrow-leaf Nardoo			
<i>Marsilea drummondii</i> A.Braun	Common Nardoo			
<i>Marsilea hirsuta</i> R.Br.	Short-fruit Nardoo			
<i>Marsilea mutica</i> Mett.	Variable Nardoo			E
<i>Pilularia novae-hollandiae</i> A.Braun	Austral Pillwort			R

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
<b>AZOLLACEAE</b>				
<i>Azolla filiculoides</i>	Pacific Azolla			
<i>Azolla pinnata</i>	Ferny Azolla			
<b>PINACEAE</b>				
* <i>Pinus canariensis</i> C.Smith	Canary Pine			
* <i>Pinus halepensis</i> Mill	Aleppo Pine	Y		
* <i>Pinus pinaster</i> Aiton	Maritime Pine			
* <i>Pinus ponderosa</i> Douglas	Ponderosa Pine			
* <i>Pinus radiata</i> D.Don	Radiata Pine			
<b>CUPRESSACEAE</b>				
<i>Callitris canescens</i> (Parl.)S.T.Blake	Scrubby Cypress Pine			
<i>Callitris glaucophylla</i> Joy Thomps. & L.A.S.Johnson	White Cypress-pine			
<i>Callitris gracilis</i> R.T.Baker	Southern Cypress Pine			
<i>Callitris verrucosa</i> (A.Cunn. ex Endl.)F.Muell.	Scrub Cypress Pine			
<i>Cupressus macrocarpa</i> Hartw. ex Gordon	Monterey Cypress			
<b>CASUARINACEAE</b>				
<i>Allocasuarina luehmannii</i> (R.T.Baker)L.A.S.Johnson	Bull Oak			
<i>Allocasuarina muelleriana</i> (Miq.)L.A.S.Johnson ssp. <i>muelleriana</i>	Common Oak-bush			
<i>Allocasuarina pusilla</i> (Macklin)L.A.S.Johnson	Dwarf Oak-bush			
<i>Allocasuarina verticillata</i> (Lam.)L.A.S.Johnson	Drooping Sheoak			
* <i>Casuarina glauca</i> Sieber ex Spreng.	River Oak			
<i>Casuarina pauper</i> F.Muell. ex L.A.S.Johnson	Black Oak			
<b>SALICACEAE</b>				
* <i>Populus alba</i> L.	White Poplar			
* <i>Salix babylonica</i> L.	Weeping Willow			
* <i>Salix X rubens</i> Schrank	Willow	Y		
<b>CANNABACEAE</b>				
* <i>Cannabis sativa</i> L.	Indian Hemp			
<b>URTICACEAE</b>				
<i>Parietaria cardiostegia</i> Greuter	Mallee Smooth-nettle			
<i>Parietaria debilis</i> G.Forst.	Smooth-nettle			
* <i>Parietaria judaica</i> L.	Wall Pellitory			
<i>Urtica incisa</i> Poir.	Scrub Nettle			
* <i>Urtica urens</i> L.	Small Nettle			
<b>PROTEACEAE</b>				
<i>Adenanthos terminalis</i> R.Br.	Yellow Gland-flower			
<i>Banksia marginata</i> Cav.	Silver Banksia			
<i>Banksia ornata</i> F.Muell. ex Meisn.	Desert Banksia			



Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
<i>Conospermum patens</i> Schldtl.	Slender Smoke-bush			
<i>Grevillea huegelii</i> Meisn.	Comb Grevillea			
<i>Grevillea ilicifolia</i> (R.Br.)R.Br. ssp. <i>ilicifolia</i>	Holly-leaf Grevillea			
<i>Grevillea ilicifolia</i> R.Br. ssp. <i>lobata</i> (F.Muell.)T.L.Downing	Lobed Holly-leaf Grevillea			
<i>Grevillea lavandulacea</i> Schldtl. ssp. <i>lavandulacea</i>	Spider-flower			
<i>Grevillea pterosperma</i> F.Muell.	Dune Grevillea			
<i>Hakea carinata</i> F.Muell. ex Meisn.	Erect Hakea			
* <i>Hakea laurina</i> R.Br.	Pincushion Hakea			
<i>Hakea leucoptera</i> R.Br. ssp. <i>leucoptera</i>	Silver Needlewood			
<i>Hakea mitchellii</i> Meisn.	Silver Needlewood			
<i>Hakea rostrata</i> F.Muell. ex Meisn.	Beaked Hakea			
<i>Hakea rugosa</i> R.Br.	Dwarf Hakea			
<i>Hakea tephrosperma</i> R.Br.	Hooked Needlewood			R
<i>Hakea vittata</i> R.Br.	Limestone Needlebush			
<i>Isopogon ceratophyllus</i> R.Br.	Horny Cone-bush			
<i>Persoonia juniperina</i> Labill.	Prickly Geebung			
<b>SANTALACEAE</b>				
<i>Choretrum glomeratum</i> R.Br. var. <i>chrysanthum</i> (F.Muell.)Benth.	Yellow-flower Sour-bush			R
<i>Choretrum glomeratum</i> R.Br. var. <i>glomeratum</i>	White Sour-bush			
<i>Exocarpos aphyllus</i> R.Br.	Leafless Cherry			
<i>Exocarpos cupressiformis</i> Labill.	Native Cherry			
<i>Exocarpos sparteus</i> R.Br.	Slender Cherry			
<i>Exocarpos strictus</i> R.Br.	Pale-fruit Cherry			R
<i>Leptomeria aphylla</i> R.Br.	Leafless Currant-bush			
<i>Santalum acuminatum</i> (R.Br.)A.DC.	Quandong			
<i>Santalum lanceolatum</i> R.Br.	Plumbush			
<i>Santalum murrayanum</i> (T.L.Mitch.)C.A.Gardner	Bitter Quandong			
<b>LORANTHACEAE</b>				
<i>Amyema linophylla</i> (Fenzl)Tiegh. ssp. <i>orientale</i> Barlow	Casuarina Mistletoe			
<i>Amyema melaleucae</i> (Lehm.ex Miq.)Tiegh.	Tea-tree Mistletoe			
<i>Amyema miquelii</i> (Lehm.ex Miq.)Tiegh.	Box Mistletoe			
<i>Amyema miraculosa</i> (Miq.)Tiegh. ssp. <i>boormanii</i> (Blakely)Barlow	Fleshy Mistletoe			
<i>Amyema pendula</i> (Sieber ex Spreng.)Tiegh. ssp. <i>pendula</i>	Drooping Mistletoe			
<i>Amyema preissii</i> (Miq.)Tiegh.	Wire-leaf Mistletoe			
<i>Lysiana exocarpi</i> (Behr)Tiegh. ssp. <i>exocarpi</i>	Harlequin Mistletoe			
<b>POLYGONACEAE</b>				
* <i>Acetosa vesicaria</i> (L.)A.Love	Rosy Dock			
* <i>Acetosella vulgaris</i> Fourr.	Sorrel			

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
* <i>Emex australis</i> Steinh.	Three-corner Jack	Y		
<i>Muehlenbeckia adpressa</i> (Labill.)Meisn.	Climbing Lignum			
<i>Muehlenbeckia diclina</i> (F.Muell.)F.Muell. ssp. <i>diclina</i>	Twiggy Lignum			
<i>Muehlenbeckia florulenta</i> Meisn.	Lignum			
<i>Muehlenbeckia gunnii</i> (Hook.f.)Endl.	Coastal Climbing Lignum			
<i>Muehlenbeckia horrida</i> H.Gross ssp. <i>horrida</i>	Spiny Lignum			R
* <i>Persicaria capitata</i> (Buch.-Ham. ex D.Don)H.Gross	Japanese Knotweed			
<i>Persicaria decipiens</i> (R.Br.)K.L.Wilson	Slender Knotweed			
<i>Persicaria lapathifolia</i> (L.)Gray	Pale Knotweed			
<i>Persicaria prostrata</i> (R.Br.)Sojak	Creeping Knotweed			
<i>Polygonum aviculare</i> L.	Wireweed			
* <i>Polygonum bellardii</i> All.				
<i>Polygonum plebeium</i> R.Br.	Small Knotweed			
<i>Rumex bidens</i> R.Br.	Mud Dock			
<i>Rumex brownii</i> Campd.	Slender Dock			
* <i>Rumex conglomeratus</i> Murray	Clustered Dock			
* <i>Rumex crispus</i> L.	Curled Dock			
<i>Rumex crystallinus</i> Lange	Glistening Dock			
<i>Rumex dumosus</i> A.Cunn. ex Meisn.	Wiry Dock			R
* <i>Rumex obtusifolius</i> L.	Broad-leaf Dock			
* <i>Rumex pulcher</i> L. ssp. <i>pulcher</i>	Fiddle Dock			
<i>Rumex tenax</i> Rech.f.	Shiny Dock			
<b>GYROSTEMONACEAE</b>				
<i>Codonocarpus cotinifolius</i> (Desf.)F.Muell.	Desert Poplar			
<i>Codonocarpus pyramidalis</i> (F.Muell.)F.Muell.	Slender Bell-fruit		V	E
<i>Gyrostemon australasicus</i> (Moq.)Heimerl Buckbush	Wheel-fruit			
<i>Gyrostemon thesioides</i> (Hook.f.)A.S.George	Broom Wheel-fruit			
<b>NYCTAGINACEAE</b>				
<i>Boerhavia coccinea</i> Mill.	Tar-vine			
<i>Boerhavia dominii</i> Meikle & Hewson	Tar-vine			
<i>Boerhavia schomburgkiana</i> Oliv.	Schomburgk's Tar-vine			
<b>AIZOACEAE</b>				
<i>Carpobrotus modestus</i> S.T.Blake	Inland Pigface			
<i>Carpobrotus rossii</i> (Haw.)Schwantes	Native Pigface			
<i>Carpobrotus</i> sp. Short calyx (S.T.Blake 20451) Toelken				
* <i>Cleretum papulosum</i> (L.f.)L.Bolus ssp. <i>papulosum</i>				
<i>Disphyma crassifolium</i> (L.)L.Bolus ssp. <i>clavellatum</i> (Haw.)Chinnock	Round-leaf Pigface			
* <i>Galenia pubescens</i> (Eckl. & Zeyh.)Druce var. <i>pubescens</i>	Coastal Galenia			

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
* <i>Galenia secunda</i> (L.f.)Sond.	Galenia			
<i>Disphyma crassifolium</i> (L.)L.Bolus ssp. <i>clavellatum</i> (Haw.)Chinnock	Round-leaf Pigface			
<i>Glinus lotoides</i> L.	Hairy Carpet-weed			
<i>Glinus oppositifolia</i> (L.)A.DC.	Slender Carpet-weed			
* <i>Mesembryanthemum aitonis</i> Jacq.	Angled Iceplant			
* <i>Mesembryanthemum crystallinum</i> L.	Common Iceplant			
* <i>Mesembryanthemum nodiflorum</i> L.	Slender Iceplant			
* <i>Psilocaulon granulicaule</i> (Haw.)Schwantes	Match-head Plant			
* <i>Ruschia tumidula</i> (Haw.)Schwantes	Pigface			
<i>Sarcosona praecox</i> (F.Muell.)S.T.Blake ex H.Eichler	Sarcosona			
<i>Tetragonia eremaea</i> Ostenf.	Desert Spinach			
<i>Tetragonia implexicoma</i> (Miq.)Hook.f.	Bower Spinach			
<i>Tetragonia moorii</i> M.Gray				
<i>Tetragonia tetragonioides</i> (Pall.)Kuntze	New Zealand Spinach			
<i>Zaleya galericulata</i> (Melville)H.Eichler ssp. <i>australis</i> (Melville)S.W.L.Jacobs	Hogweed			
<b>PORTULACACEAE</b>				
<i>Calandrinia calypttrata</i> Hook.f.	Pink Purslane			
<i>Calandrinia corrigioloides</i> F.Muell. ex Benth.	Strap Purslane			
<i>Calandrinia eremaea</i> Ewart	Dryland Purslane			
<i>Calandrinia granulifera</i> Benth.	Pigmy Purslane			
<i>Calandrinia volubilis</i> Benth.	Twining Purslane			
<i>Montia australasica</i> (Hook.f.)Pax & K.Hoffm.	White Purslane			R
<i>Portulaca oleracea</i> L.	Common Purslane			
<b>CARYOPHYLLACEAE</b>				
* <i>Arenaria leptoclados</i> (Rchb.)Guss.	Lesser Thyme-leaved Sandwort			
* <i>Cerastium balearicum</i> F.Herm.	Chickweed			
* <i>Cerastium glomeratum</i> Thuill.	Common Mouse-ear Chickweed			
* <i>Gypsophila tubulosa</i> (Jaub. & Spach)Boiss.	Annual Chalkwort			
* <i>Herniaria cinerea</i> DC.	Rupturewort			
* <i>Minuartia mediterranea</i> (Link)K.Maly	Slender Sandwort			
* <i>Moenchia erecta</i> (L.)Gaertn.	Erect Chickweed			
* <i>Paronychia argentea</i> Lam.	Silver Whitlow			
* <i>Petrorhagia dubia</i> (Rafin)G.Lopez & Romo				
* <i>Petrorhagia nanteuilii</i> (Burnat)P.W.Ball & Heywood				
* <i>Polycarpon tetraphyllum</i> (L.)L.	Four-leaf Allseed			
* <i>Sagina apetala</i> Ard.	Annual Pearlwort			
<i>Sagina maritima</i> G.Don	Sea Pearlwort			
* <i>Sagina procumbens</i> L.	Spreading Pearlwort			

Species.	Common name	Proclaimed Pest Plant	PBC Act Status	NPW Act Status
<i>Scleranthus minusculus</i> F.Muell.	Cushion Knawel			
<i>Scleranthus pungens</i> R.Br.	Prickly Knawel			
* <i>Silene apetala</i> Willd.	Sand Catchfly			
* <i>Silene armeria</i> L.	Sweet-william Catchfly			
* <i>Silene gallica</i> L. var. <i>gallica</i>	French Catchfly			
* <i>Silene longicaulis</i> Pourr. ex Lag.	Portuguese Catchfly			
* <i>Silene nocturna</i> L.	Mediterranean Catchfly			
* <i>Silene tridentata</i> Desf.				
* <i>Silene vulgaris</i> (Moench)Garcke	Bladder Campion	Y		
* <i>Spergula arvensis</i> L.	Corn Spurrey			
* <i>Spergularia bocconii</i> (Scheele)Graebn.				
* <i>Spergularia diandra</i> (Guss.)Heldr. & Sartori	Lesser Sand-spurrey			
<i>Spergularia marina</i> (L.)Griseb.	Salt Sand-spurrey			
* <i>Spergularia media</i> (L.)C.Presl mCoast Sand-spurrey				
* <i>Spergularia rubra</i> (L.)J. & C.Presl	Red Sand-spurrey			
<i>Spergularia</i> sp. Butchers Gap (P.Gibbons 234) L.G.Adams & J.G.West				
<i>Spergularia</i> sp. Mt Mulyah (C.W.E.Moore 7046) L.G.Adams				
<i>Stellaria angustifolia</i> Hook.				
<i>Stellaria filiformis</i> (Benth.)Mattf.	Thread Starwort			
* <i>Stellaria media</i> (L.)Vill.	Chickweed			
<i>Stellaria multiflora</i> Hook.	Rayless Starwort			R
* <i>Stellaria pallida</i> (Dumort.)Crep.	Lesser Starwort			
<i>Stellaria 'tenella'</i>	Swamp Starwort			R
* <i>Vaccaria hispanica</i> (Mill.)Rauschert	Cow Soapwort			
<b>CHENOPODIACEAE</b>				
<i>Atriplex acutibractea</i> R.H.Anderson ssp. <i>acutibractea</i>	Pointed Saltbush			
<i>Atriplex acutibractea</i> R.H.Anderson ssp. <i>karoniensis</i> Aellen Pointed Saltbush				
<i>Atriplex acutiloba</i> R.H.Anderson				
<i>Atriplex angulata</i> Benth.	Fan Saltbush			
<i>Atriplex australasica</i> Moq.	Green Saltbush			R
<i>Atriplex eardleyae</i> Aellen	Eardley's Saltbush			
<i>Atriplex elachophylla</i> F.Muell.				
<i>Atriplex holocarpa</i> F.Muell.	Pop Saltbush			
<i>Atriplex leptocarpa</i> F.Muell.	Slender-fruit Saltbush			
<i>Atriplex limbata</i> Benth.	Spreading Saltbush			
<i>Atriplex lindleyi</i> Moq. ssp. <i>inflata</i> (F.Muell.)Paul G.Wilson	Corky Saltbush			
<i>Atriplex lindleyi</i> Moq. ssp. <i>lindleyi</i>	Baldoo			
<i>Atriplex nummularia</i> Lindl. ssp. <i>nummularia</i>	Old-man Saltbush			



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<i>Atriplex paludosa</i> R.Br. ssp. <i>cordata</i> (Benth.)Aellen	Marsh Saltbush			
<i>Atriplex papillata</i> J.H.Willis Coral	Saltbush			E
<i>Atriplex prostrata</i> Boucher ex DC.	Creeping Saltbush			
<i>Atriplex pseudocampanulata</i> Aellen	Spreading Saltbush			
<i>Atriplex pumilio</i> R.Br.	Mat Saltbush			
<i>Atriplex rhagodioides</i> F.Muell.	River Saltbush			
<i>Atriplex semibaccata</i> R.Br.	Berry Saltbush			
<i>Atriplex spongiosa</i> F.Muell.	Pop Saltbush			
<i>Atriplex stipitata</i> Benth.	Bitter Saltbush			
<i>Atriplex suberecta</i> L.Verd.	Lagoon Saltbush			
<i>Atriplex velutinella</i> F.Muell.	Sandhill Saltbush			
<i>Atriplex vesicaria</i> Heward ex Benth.	Bladder Saltbush			
<i>Chenopodium album</i> L.	Fat Hen			
* <i>Chenopodium ambrosioides</i> L. var. <i>ambrosioides</i>	Mexican Tea			
<i>Chenopodium auricomum</i> Lindl.	Golden Goosefoot			
<i>Chenopodium cristatum</i> (F.Muell.)F.Muell.	Crested Goosefoot			
<i>Chenopodium curvispicatum</i> Paul G.Wilson	Cottony Goosefoot			
<i>Chenopodium desertorum</i> (J.M.Black)J.M.Black ssp. <i>desertorum</i>	Mallee Goosefoot			
<i>Chenopodium desertorum</i> (J.M.Black)J.M.Black ssp. <i>microphyllum</i> Paul G.Wilson	Frosted Goosefoot			
<i>Chenopodium desertorum</i> (J.M.Black)J.M.Black ssp. <i>rectum</i> Paul G.Wilson	Erect Goosefoot			
<i>Chenopodium glaucum</i> L.	Glaucous Goosefoot			
<i>Chenopodium melanocarpum</i> (J.M.Black)J.M.Black f. <i>melanocarpum</i>	Black-fruit Goosefoot			
<i>Chenopodium multifidum</i> L.	Scented Goosefoot			
* <i>Chenopodium murale</i> L.	Nettle-leaf Goosefoot			
<i>Chenopodium nitrariaceum</i> (F.Muell.)F.Muell.ex Benth.	Nitre Goosefoot			
<i>Chenopodium pumilio</i> R.Br.	Clammy Goosefoot			
<i>Chenopodium truncatum</i> Paul G.Wilson				
<i>Dissocarpus biflorus</i> F.Muell. var. <i>biflorus</i>	Two-horn Saltbush			
<i>Dissocarpus latifolius</i> (J.M.Black)	Paul G.Wilson			
<i>Dissocarpus paradoxus</i> (R.Br.)F.Muell. ex Ulbr.	Ball Bindyi			
<i>Dysphania glomulifera</i> (Nees)Paul G.Wilson ssp. <i>glomulifera</i>	Globular Crumbweed			
<i>Dysphania platycarpa</i> Paul G.Wilson	Flat-fruit Crumbweed			
<i>Einadia nutans</i> (R.Br.)A.J.Scott ssp. <i>nutans</i>	Climbing Saltbush			
<i>Einadia nutans</i> (R.Br.)A.J.Scott ssp. <i>oxycarpa</i> (Gaub)Paul G.Wilson	Pointed-fruit Climbing Saltbush			
<i>Enchylaena tomentosa</i> R.Br. var. <i>tomentosa</i>	Ruby Saltbush			
<i>Eriochiton sclerolaenoides</i> (F.Muell.)F.Muell. ex A.J.Scott	Woolly-fruit Bluebush			
<i>Halosarcia halocnemoides</i> (Nees)Paul G.Wilson ssp. <i>halocnemoides</i>	Grey Samphire			
<i>Halosarcia indica</i> (Willd.)Paul G.Wilson ssp. <i>bidens</i> (Nees)Paul G.Wilson	Brown-head Samphire			

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<i>Halosarcia indica</i> (Willd.)Paul G.Wilson ssp. <i>leiotachya</i> (Benth.)Paul G.Wilson	Brown-head Samphire			
<i>Halosarcia pergranulata</i> (J.M.Black)Paul G.Wilson ssp. <i>divaricata</i> Paul G.Wilson	Black-seed Samphire			
<i>Halosarcia pergranulata</i> (J.M.Black)Paul G.Wilson ssp. <i>elongata</i> Paul G.Wilson	Black-seed Samphire			
<i>Halosarcia pergranulata</i> (J.M.Black)Paul G.Wilson ssp. <i>pergranulata</i>	Black-seed Samphire			
<i>Halosarcia pruinosa</i> (Paulsen)Paul G.Wilson	Bluish Samphire			
<i>Halosarcia pterygosperma</i> (J.M.Black)Paul G.Wilson ssp. <i>pterygosperma</i>	Winged-seed Samphire			
<i>Halosarcia syncarpa</i> Paul G.Wilson	Fused Samphire			
* <i>Kochia scoparia</i> (L.)Schrad.		Y		
<i>Maireana aphylla</i> (R.Br.)Paul G.Wilson	Cotton-bush			
<i>Maireana appressa</i> Paul G.Wilson	Pale-fruit Bluebush			
<i>Maireana astrotricha</i> (L.A.S.Johnson)Paul G.Wilson	Low Bluebush			
<i>Maireana brevifolia</i> (R.Br.)Paul G.Wilson	Short-leaf Bluebush			
<i>Maireana carnosa</i> (Moq.)Paul G.Wilson	Cottony Bluebush			
<i>Maireana ciliata</i> (F.Muell.)Paul G.Wilson	Hairy Fissure-plant			
<i>Maireana decalvans</i> (Gand.)Paul G.Wilson	Black Cotton-bush			E
<i>Maireana enchylaenoides</i> (F.Muell.)Paul G.Wilson	Wingless Fissure-plant			
<i>Maireana erioclada</i> (Benth.)Paul G.Wilson	Rosy Bluebush			
<i>Maireana excavata</i> (J.M.Black)Paul G.Wilson	Bottle Fissure-plant			V
<i>Maireana georgei</i> (Diels)Paul G.Wilson S	atiny Bluebush			
<i>Maireana integra</i> (Paul G.Wilson)Paul G.Wilson	Entire-wing Bluebush			
<i>Maireana lobiflora</i> (F.Muell.ex Benth.)Paul G.Wilson	Lobed Bluebush			
<i>Maireana oppositifolia</i> (F.Muell.)Paul G.Wilson	Salt Bluebush			
<i>Maireana ovata</i> (Ising)Paul G.Wilson				
<i>Maireana pentagona</i> (R.H.Anderson)Paul G.Wilson	Slender Fissure-plant			R
<i>Maireana pentatropis</i> (Tate)Paul G.Wilson	Erect Mallee Bluebush			
<i>Maireana pyramidata</i> (Benth.)Paul G.Wilson	Black Bluebush			
<i>Maireana radiata</i> (Paul G.Wilson)Paul G.Wilson	Radiate Bluebush			
<i>Maireana rohrlachii</i> (Paul G.Wilson)Paul G.Wilson	Rohrlach's Bluebush			R
<i>Maireana sedifolia</i> (F.Muell.)Paul G.Wilson	Bluebush			
<i>Maireana spongiocarpa</i> (F.Muell.)Paul G.Wilson	Spongy-fruit Bluebush			
<i>Maireana suaedifolia</i> (Paul G.Wilson)Paul G.Wilson Lax	Bluebush			R
<i>Maireana trichoptera</i> (J.M.Black)Paul G.Wilson	Hairy-fruit Bluebush			
<i>Maireana triptera</i> (Benth.)Paul G.Wilson	Three-wing Bluebush			
<i>Maireana turbinata</i> Paul G.Wilson	Top-fruit Bluebush			
<i>Malacocera tricornis</i> (Benth.)R.H.Anderson	Goat-head Soft-horns			
<i>Osteocarpum acropterum</i> (F.Muell. & Tate)Volkens var. <i>acropterum</i>	Tuberculate Bonefruit			
<i>Osteocarpum acropterum</i> (F.Muell. & Tate)Volkens var. <i>deminutum</i> (J.M.Black)Paul G.Wilson	Wingless Bonefruit			R
<i>Osteocarpum salsuginosum</i> F.Muell.	Inland Bonefruit			

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<i>Pachycornia triandra</i> (F.Muell.)J.M.	Black Desert Glasswort			
<i>Rhagodia candolleana</i> Moq. ssp. <i>candolleana</i>	Sea-berry Saltbush			
<i>Rhagodia crassifolia</i> R.Br.	Fleshy Saltbush			
<i>Rhagodia parabolica</i> R.Br.	Mealy Saltbush			
<i>Rhagodia preissii</i> Moq. ssp. <i>preissii</i>	Mallee Saltbush			
<i>Rhagodia spinescens</i> R.Br.	Spiny Saltbush			
<i>Rhagodia ulicina</i> (Gand.)Paul G.Wilson	Intricate Saltbush			
<i>Salsola kali</i> L.	Buckbush			
<i>Sarcocornia quinqueflora</i> (Bunge ex Ung.-Sternb.)A.J.Scott	Beaded Samphire			
<i>Scleroblitum atriplicinum</i> (F.Muell.)Ulbr.	Starry Goosefoot			
<i>Sclerolaena articulata</i> (J.M.Black)A.J.Scott	Jointed Bindyi			
<i>Sclerolaena birchii</i> (F.Muell.)Domin	Galvanised Burr	Y		
<i>Sclerolaena brachyptera</i> (F.Muell.)S.W.L.Jacobs	Short-wing Bindyi			
<i>Sclerolaena brevifolia</i> (Ising)A.J.Scott	Small-leaf Bindyi			
<i>Sclerolaena constricta</i> (Ising)A.J.Scott				
<i>Sclerolaena convexula</i> (R.H.Anderson)A.J.Scott	Tall Bindyi			
<i>Sclerolaena cuneata</i> Paul G.Wilson	Tangled Bindyi			
<i>Sclerolaena decurrens</i> (J.M.Black)A.J.Scott	Green Bindyi			
<i>Sclerolaena diacantha</i> (Nees)Benth.	Grey Bindyi			
<i>Sclerolaena divaricata</i> (R.Br.)Sm.	Tangled Bindyi			
<i>Sclerolaena glabra</i> (F.Muell.)Domin				
<i>Sclerolaena holtiana</i> (Ising)A.J.Scott	Holt's Bindyi			R
<i>Sclerolaena intricata</i> (R.H.Anderson)A.J.Scott	Tangled Bindyi			
<i>Sclerolaena limbata</i> (J.M.Black)Ulbr.	Pearl Bindyi			
<i>Sclerolaena muricata</i> (Moq.)Domin var. <i>muricata</i>	Five-spine Bindyi			
<i>Sclerolaena muricata</i> (Moq.)Domin var. <i>semiglabra</i> (Ising)A.J.Scott	Five-spine Bindyi			
<i>Sclerolaena muricata</i> (Moq.)Domin var. <i>villosa</i> (Benth.)Ulbr.	Five-spine Bindyi			R
<i>Sclerolaena obliquicuspis</i> (R.H.Anderson)Ulbr.	Oblique-spined Bindyi			
<i>Sclerolaena parviflora</i> (R.H.Anderson)A.J.Scott	Small-flower Bindyi			
<i>Sclerolaena patentispis</i> (R.H.Anderson)Ulbr.	Spear-fruit Bindyi			
<i>Sclerolaena stelligera</i> (F.Muell.)S.W.L.Jacobs	Star Bindyi			
<i>Sclerolaena tricuspidis</i> (F.Muell.)Ulbr.	Three-spine Bindyi			
<i>Sclerolaena uniflora</i> R.Br.	Small-spine Bindyi			
<i>Sclerostegia tenuis</i> (Benth.)Paul G.Wilson	Slender Samphire			
<i>Suaeda australis</i> (R.Br.)Moq.	Austral Seablite			
<i>Threlkeldia diffusa</i> R.Br.	Coast Bonefruit			
<b>AMARANTHACEAE</b>				
<i>Alternanthera denticulata</i> R.Br.	Lesser Joyweed			

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<i>Alternanthera nana</i> R.Br.	Hairy Joyweed			
<i>Alternanthera nodiflora</i> R.Br.	Common Joyweed			
* <i>Alternanthera pungens</i> Kunth	Khaki Weed	Y		
* <i>Amaranthus albus</i> L.	Stiff Tumbleweed			
* <i>Amaranthus caudatus</i> L.	Love-lies-bleeding			
<i>Amaranthus grandiflorus</i> (J.M.Black)J.M.Black	Large-flower Amaranth			
<i>Amaranthus macrocarpus</i> Benth. var. <i>macrocarpus</i>				
* <i>Amaranthus muricatus</i> (Moq.)Hieron.	Rough-fruit Amaranth			
* <i>Amaranthus retroflexus</i> L.	Red-root Amaranth			
* <i>Amaranthus viridis</i> L.	Green Amaranth			
* <i>Gomphrena celosioides</i> Mart.				
* <i>Guilleminea densa</i> (Humb. & Bonpl. ex Schult.)Moq.	Small Matweed			
<i>Hemichroa pentandra</i> R.Br.	Trailing Hemichroa			
<i>Ptilotus erubescens</i> Schtdl.	Hairy-tails			R
<i>Ptilotus exaltatus</i> Nees var. <i>exaltatus</i>	Pink Mulla Mulla			
<i>Ptilotus gaudichaudii</i> (Steud.)J.M.Black var. <i>parviflorus</i> (Benth.)Benl	Paper Fox-tail			
<i>Ptilotus nobilis</i> (Lindl.)F.Muell. var. <i>nobilis</i>	Yellow-tails			
<i>Ptilotus obovatus</i> (Gaudich.)F.Muell. var. <i>obovatus</i>	Silver Mulla Mulla			
<i>Ptilotus polystachyus</i> (Gaudich.)F.Muell. var. <i>polystachyus</i>	Long-tails			
<i>Ptilotus seminudus</i> (J.M.Black)J.M.Black	Rabbit-tails			
<i>Ptilotus sessilifolius</i> (Lindl.)Benl var. <i>sessilifolius</i>	Crimson-tails			
<i>Ptilotus spathulatus</i> (R.Br.)Poir. f. <i>spathulatus</i>	Pussy-tails			
<b>CACTACEAE</b>				
* <i>Austrocyllindropuntia cylindrica</i> (Lam.)Backeb.		Y		
* <i>Austrocyllindropuntia subulata</i> (Muehlenpf.)Backeb.	Eve's-pin Cactus	Y		
* <i>Cylindropuntia imbricata</i> (Haw.)F.M.Knuth	Devil's Rope Pear	Y		
* <i>Cylindropuntia leptocaulis</i> (DC.)F.M.Knuth		Y		
* <i>Cylindropuntia tunicata</i> (Lehm.)F.M.Knuth		Y		
* <i>Opuntia elata</i> Salm-Dyck		Y		
* <i>Opuntia elatior</i> Mill.		Y		
* <i>Opuntia engelmannii</i> Salm-Dyck ex Engelm.		Y		
* <i>Opuntia ficus-indica</i> (L.)Mill.	Indian Fig	Y		
* <i>Opuntia linguiformis</i> Griffiths		Y		
* <i>Opuntia microdasys</i> (Lehm.)Pfeiff.	Bunny-ears	Y		
* <i>Opuntia monacantha</i> Haw.		Y		
* <i>Opuntia paraguayensis</i> K.Schum.	Riverina Pear	Y		
* <i>Opuntia polyacantha</i> Haw.		Y		
* <i>Opuntia stricta</i> (Haw.)Haw.	Erect Prickly Pear	Y		



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<i>*Opuntia tomentosa</i> Salm-Dyck	Velvet Pear	Y		
<b>LAURACEAE</b>				
<i>Cassytha glabella</i> R.Br. f. <i>dispar</i> (Schltld.)J.Z.Weber	Slender Dodder-laurel			
<i>Cassytha melantha</i> R.Br.	Coarse Dodder-laurel			
<i>Cassytha pubescens</i> R.Br.	Downy Dodder-laurel			
<b>RANUNCULACEAE</b>				
<i>*Adonis microcarpa</i> DC.	Pheasant's Eye	Y		
<i>*Batrachium trichophyllum</i> (Chaix)Bosch	Water Buttercup			
<i>*Clematis flammula</i> L.				
<i>Clematis microphylla</i> DC. var. <i>microphylla</i>	Old Man's beard			
<i>Myosurus minimus</i> L. var. <i>australis</i> (F.Muell.)Huth	Mousetail			
<i>Ranunculus amphitrichus</i> Colenso S	mall River Buttercup			
<i>Ranunculus inundatus</i> R.Br. ex DC	River Buttercup			R
<i>Ranunculus lappaceus</i> Sm.	Native Buttercup			
<i>*Ranunculus muricatus</i> L.	Pricklefruit Buttercup			
<i>Ranunculus pachycarpus</i> B.G.Briggs	Thick-fruit Buttercup			
<i>Ranunculus papulentus</i> Melville	Large River Buttercup			V
<i>Ranunculus pentandrus</i> J.M.Black var. <i>platycarpus</i> (F.Muell.)H.Eichler	Smooth Buttercup			
<i>Ranunculus pumilio</i> R.Br. ex DC. var. <i>politus</i> Melville	Smooth-fruit Ferny Buttercup			V
<i>Ranunculus pumilio</i> R.Br. ex DC. var. <i>pumilio</i>	Ferny Buttercup			
<i>*Ranunculus sceleratus</i> L. ssp. <i>sceleratus</i>	Celery Buttercup	Y		
<i>Ranunculus sessiliflorus</i> R.Br. ex DC. var. <i>pilulifer</i> (Hook.)Melville	Annual Buttercup			V
<i>Ranunculus sessiliflorus</i> R.Br. ex DC. var. <i>sessiliflorus</i>	Annual Buttercup			
<i>*Ranunculus trilobus</i> Desf.	Three-lobed Buttercup			
<b>NYMPHAEACEAE</b>				
<i>*Nymphaea alba</i> L.	White Water-lily			
<b>CERATOPHYLLACEAE</b>				
<i>Ceratophyllum demersum</i> L.	Hornwort			R
<b>DILLENIACEAE</b>				
<i>Hibbertia crinita</i> Toelken				
<i>Hibbertia</i> sp. <i>Glabriuscula</i> (D.J.Whibley 9012) Toelken				
<i>Hibbertia sericea</i> (R.Br. ex DC.)Benth.	Silky Guinea-flower			
<i>Hibbertia virgata</i> R.Br. ex DC.	Twiggy Guinea-flower			
<b>GUTTIFERAE</b>				
<i>Hypericum gramineum</i> G.Forst.	Small St John's Wort			
<i>Hypericum japonicum</i> Thunb.	Matted St John's Wort			R
<i>*Hypericum perforatum</i> L.	St John's Wort			
<b>DROSERACEAE</b>				

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<i>Drosera auriculata</i> Backh. ex Planch.	Tall Sundew			
<i>Drosera glanduligera</i> Lehm.	Scarlet Sundew			
<i>Drosera macrantha</i> Endl. ssp. <i>planchonii</i> (Hook.f. ex Planch.)N.G.Marchant	Climbing Sundew			
<i>Drosera peltata</i> Thunb.	Pale Sundew			
<i>Drosera pygmaea</i> DC.	Tiny Sundew			
<i>Drosera whittakeri</i> ssp. <i>whittakeri</i>	Scented Sundew			
<b>PAPAVERACEAE</b>				
* <i>Eschscholzia californica</i> Cham.	Californian Poppy			
* <i>Glaucium corniculatum</i> (L.)Rudolph	Horned Poppy			
* <i>Papaver aculeatum</i> Thunb.	Bristle Poppy			
* <i>Papaver dubium</i> L.	Long-headed Poppy			
* <i>Papaver hybridum</i> L.	Rough Poppy			
* <i>Papaver rhoeas</i> L.	Field Poppy			
* <i>Papaver somniferum</i> L.	Small-flower Opium Poppy			
<b>FUMARIACEAE</b>				
* <i>Fumaria bastardii</i> Boreau	Bastard Fumitory			
* <i>Fumaria capreolata</i> L.	White-flower Fumitory			
* <i>Fumaria densiflora</i> DC.	Dense Fumitory			
* <i>Fumaria muralis</i> Sond. ex W.D.J.Koch ssp. <i>muralis</i>	Wall Fumitory			
* <i>Fumaria officinalis</i> L. ssp. <i>officinalis</i>	Common Fumitory			
<i>Fumaria parviflora</i> Lam. var. <i>parviflora</i>	Small-flower Fumitory			
<b>CRUCIFERAE</b>				
* <i>Alyssum linifolium</i> Stephan ex Willd.	Flax-leaf Alyssum			
<i>Arabidella eremigena</i> (F.Muell.)E.A.Shaw	Priddiwalkatji			
<i>Arabidella filifolia</i> (F.Muell.)E.A.Shaw	Thread-leaf Cress			
<i>Arabidella nasturtium</i> (F.Muell.)E.A.Shaw	Yellow Cress			
<i>Arabidella procumbens</i> (Tate)E.A.Shaw	Creeping Cress			
<i>Arabidella trisecta</i> (F.Muell.)O.E.Schulz	Shrubby Cress			
* <i>Brassica elongata</i> Ehrh.				
* <i>Brassica oleracea</i> L.	Cabbage			
* <i>Brassica rapa</i> L. ssp. <i>oleifolia</i> DC.	Turnip Rape			
* <i>Brassica tournefortii</i> Gouan	Wild Turnip			
* <i>Brassica X napus</i> L.				
* <i>Cakile maritima</i> Scop. ssp. <i>maritima</i>	Sea Rocket			
* <i>Capsella bursapastoris</i> (L.)Medik.	Shepherd's Purse			
* <i>Cardamine flexuosa</i> With.	Wood Bitter-cress			
<i>Cardamine moirensis</i> I.Thomps.				
* <i>Carrichtera annua</i> (L.)DC.	Ward's Weed			

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* <i>Diplotaxis muralis</i> (L.)DC. var. <i>muralis</i>	Wall Rocket			
* <i>Diplotaxis tenuifolia</i> (L.)DC.	Lincoln Weed	Y		
* <i>Erophila verna</i> (L.)Chevall. ssp. <i>praecox</i> (Steven)Walters	Whitlow Grass			
* <i>Erophila verna</i> (L.)Chevall. ssp. <i>verna</i>	Whitlow Grass			
* <i>Eruca sativa</i> Mill.	Purple-vein Rocket			
<i>Geococcus pusillus</i> J.L.Drumm. ex Harvey	Earth Cress			
<i>Harmsiodoxa blennodioides</i> (F.Muell.)O.E.Schulz	Hairy-pod Cress			
<i>Harmsiodoxa brevipes</i> (F.Muell.)O.E.Schulz var. <i>brevipes</i>	Short Cress			
* <i>Hirschfeldia incana</i> (L.)Lagr.-Foss.	Hoary Mustard	Y		
* <i>Hornungia procumbens</i> (L.)Hayek	Oval Purse			
<i>Joycea clelandii</i> (Vickery)H.P.Linder				
* <i>Lepidium africanum</i> (Burm.f.)DC.	Common Peppergrass			
* <i>Lepidium bonariense</i> L.	Cut-leaf Peppergrass			
* <i>Lepidium didymum</i> L.				
* <i>Lepidium draba</i> L.		Y		
<i>Lepidium fasciculatum</i> Thell.	Bundled Peppergrass			
<i>Lepidium hyssopifolium</i> Desv.	Small Peppergrass			
<i>Lepidium leptopetalum</i> (F.Muell.)F.Muell.	Shrubby Peppergrass			
<i>Lepidium monoplocoides</i> F.Muell.	Winged Peppergrass		E	E
<i>Lepidium oxytrichum</i> Sprague	Green Peppergrass			
<i>Lepidium papillosum</i> F.Muell.	Warty Peppergrass			
<i>Lepidium phlebopetalum</i> (F.Muell.)F.Muell.	Veined Peppergrass			
<i>Lepidium pseudohyssopifolium</i> Hewson				
<i>Lepidium pseudoruderale</i> Thell.				R
<i>Lepidium pseudotasmanicum</i> Thell.	Shade Peppergrass			V
<i>Lepidium pubescens</i> Desv.	Matted Peppergrass			
<i>Lepidium sagittulatum</i> Thell.	Fine-leaf Peppergrass			
* <i>Lepidium strictum</i> (S.Watson)Rattan				
* <i>Malcolmia flexuosa</i> (Sibth. & Sm.)Sibth. & Sm.				
<i>Menkea australis</i> Lehm.	Fairy Spectacles			
<i>Microlepidium pilosulum</i> F.Muell.	Hairy Shepherd's-purse			R
<i>Pachymitus cardaminoides</i> (F.Muell.)O.E.Schulz	Sand Cress			
<i>Phlegmatospermum eremaeum</i> (J.M.Black)E.A.Shaw	Spreading Cress			R
* <i>Raphanus raphanistrum</i> L.	Wild Radish			
* <i>Rapistrum rugosum</i> (L.)All. ssp. <i>rugosum</i>	Turnip Weed			
<i>Rorippa eustylis</i> (F.Muell.)L.A.S.Johnson	River Bitter-cress			
<i>Rorippa laciniata</i> (F.Muell.)L.A.S.Johnson	Jagged Bitter-cress			R
<i>Rorippa nasturtium-aquaticum</i> (L.)Hayek	Watercress			

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<i>Rorippa palustris</i> (L.)Besser	Yellow Marsh-cress			
* <i>Rorippa sylvestris</i> (L.)Besser ssp. <i>sylvestris</i>	Creeping Yellow-cress			
* <i>Sisymbrium erysimoides</i> Desf.	Smooth Mustard			
* <i>Sisymbrium irio</i> L.	London Mustard			
* <i>Sisymbrium officinale</i>	Hedge Mustard			
* <i>Sisymbrium orientale</i>	Indian Hedge Mustard			
<i>Stenopetalum lineare</i> R.Br. ex DC.	Narrow Thread-petal			
<i>Stenopetalum sphaerocarpum</i> F.Muell.	Round-fruit Thread-petal			
<b>RESEDACEAE</b>				
* <i>Reseda lutea</i> L. Cut-leaf Mignonette		Y		
* <i>Reseda luteola</i> L.	Wild Mignonette			
<b>CRASSULACEAE</b>				
* <i>Cotyledon orbiculata</i> L. var. <i>oblonga</i> (Haw.)DC.	Cotyledon			
<i>Crassula colligata</i> Toelken ssp. <i>Colligata</i>				
<i>Crassula colorata</i> (Nees)Ostenf. var. <i>acuminata</i> (Reader)Toelken	Dense Crassula			
<i>Crassula colorata</i> (Nees)Ostenf. var. <i>colorata</i>	Dense Crassula			
<i>Crassula decumbens</i> Thunb. var. <i>decumbens</i>	Spreading Crassula			
<i>Crassula helmsii</i> (Kirk)Cockayne	Swamp Crassula			
* <i>Crassula natans</i> Thunb. var. <i>minus</i> (Eckl. & Zeyh.)G.D.Rowley	Water Crassula			
<i>Crassula peduncularis</i> )	Purple Crassula			R
* <i>Crassula tetragona</i> L. ssp. <i>robusta</i> (Toelken)Toelken	Crassula			
<i>Crassula tetramera</i> (Toelken)A.P.Druce & Sykes	Australian Stonecrop			
<b>PITTOSPORACEAE</b>				
<i>Marianthus bignoniaceus</i> F.Muell.	Orange Bell-climber			
<i>Billardiera cymosa</i> F.Muell.	Sweet Apple-berry			
<i>Billardiera versicolor</i> F.Muell. ex Klatt	Yellow-flower Apple-berry			
<i>Bursaria spinosa</i> Cav. ssp. <i>lasiophylla</i> (E.M.Benn.)L.Cayzer, Crisp & I.Telford	Downy Bursaria			
<i>Bursaria spinosa</i> Cav. ssp. <i>spinosa</i>	Sweet Bursaria			
<i>Cheiranthra alternifolia</i> E.M.Benn.	Hand-flower			
<i>Pittosporum angustifolium</i> Lodd.	Native Apricot			
<b>ROSACEAE</b>				
<i>Acaena echinata</i> Nees				
<i>Acaena novae-zelandiae</i> Kirk	Biddy-biddy			
<i>Acaena ovina</i> A.Cunn.	Downy Sheep's-burr			
<i>Aphanes australiana</i> (Rothm.)Rothm.	Australian Piert			
* <i>Malus pumila</i> Mill.				
* <i>Prunus armeniaca</i> L.	Apricot			
* <i>Prunus cerasifera</i> Ehrh.	Cherry-plum			

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* <i>Prunus persica</i> (L.)Batsch var. <i>nectarina</i> (R.Br.)Maxim.	Nectarine			
* <i>Prunus persica</i> (L.)Batsch var. <i>persica</i>	Peach			
* <i>Pyrus communis</i> L.	Pear			
* <i>Rosa canina</i> L.	Dog Rose	Y		
* <i>Rosa rubiginosa</i> L.	Sweet Briar	Y		
<i>Rubus parvifolius</i> L.	Native Raspberry			
* <i>Rubus ulmifolius</i> Schott var. <i>anoplothyrsus</i> Sudre	Blackberry	Y		
* <i>Sanguisorba minor</i> Scop. ssp. <i>muricata</i> Briq.	Sheep's Burnet			
<b>LEGUMINOSAE</b>				
<i>Acacia acanthoclada</i> F.Muell. ssp. <i>acanthoclada</i>	Harrow Wattle			
<i>Acacia acinacea</i> Lindl.	Wreath Wattle			
<i>Acacia ancistrophylla</i> C.R.P.Andrews var. <i>lissophylla</i> (J.M.Black)R.S.Cowan & Maslin	Hook-leaf Wattle			
<i>Acacia aneura</i> F.Muell. ex Benth. var. <i>aneura</i>	Mulga			
<i>Acacia argyrophylla</i> Hook.	Silver Mulga-bush			
<i>Acacia beckleri</i> Tindale	Beckler's Rock Wattle			
<i>Acacia brachybotrya</i> Benth.	Grey Mulga-bush			
<i>Acacia burkittii</i> F.Muell. ex Benth.	Pin-bush Wattle			
<i>Acacia calamifolia</i> Sweet ex Lindl.	Wallowa			
<i>Acacia colletioides</i> Benth.	Veined Wait-a-while			
<i>Acacia continua</i> Benth.	Thorn Wattle			
<i>Acacia cupularis</i> Domin	Cup Wattle			
<i>Acacia dodonaeifolia</i> (Pers.)Balb.	Hop-bush Wattle			R
<i>Acacia euthycarpa</i> (J.M.Black)J.M.Black				
<i>Acacia farinosa</i> Lindl.	Mealy Wattle			
<i>Acacia glandulicarpa</i> Reader	Hairy-pod Wattle			
<i>Acacia hakeoides</i> A.Cunn. ex Benth.	Hakea Wattle			
<i>Acacia halliana</i> Maslin	Hall's Wattle			
<i>Acacia ligulata</i> A.Cunn. ex Benth.	Umbrella Bush			E
<i>Acacia lineata</i> A.Cunn. ex G.Don	Streaked Wattle			R
<i>Acacia loderi</i> Maiden	Nealie			R
<i>Acacia longifolia</i> (Andrews)Willd. ssp. <i>sophorae</i> (Labill.)Court	Coastal Wattle			
<i>Acacia melanoxylon</i> R.Br.	Blackwood			
<i>Acacia menzeldii</i> J.M.Black	Menzel's Wattle		V	V
<i>Acacia microcarpa</i> F.Muell.	Manna Wattle			
<i>Acacia montana</i> Benth.	Mallee Wattle			R
<i>Acacia myrtifolia</i> (Sm.)Willd. var. <i>myrtifolia</i>	Myrtle Wattle			
<i>Acacia notabilis</i> F.Muell.	Notable Wattle			
<i>Acacia nyssophylla</i> F.Muell.	Spine Bush			

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<i>Acacia oswaldii</i> F.Muell.	Umbrella Wattle			
<i>Acacia paradoxa</i> DC.	Kangaroo Thorn			
<i>Acacia pendula</i> A.Cunn. ex G.Don	Weeping Myall			V
<i>Acacia pravifolia</i> F.Muell.	Coil-pod Wattle			
<i>Acacia pycnantha</i> Benth.	Golden Wattle			
<i>Acacia retinodes</i> Schltld. var. <i>retinodes</i>	Wirilda			
<i>Acacia rhetinocarpa</i> J.M.Black	Resin Wattle		V	V
<i>Acacia rhigiophylla</i> F.Muell. ex Benth.	Dagger-leaf Wattle			R
<i>Acacia rigens</i> A.Cunn. ex G.Don	Nealie			
<i>Acacia rupicola</i> F.Muell. ex Benth.	Rock Wattle			
<i>Acacia salicina</i> Lindl.	Willow Wattle			
* <i>Acacia saligna</i> (Labill.)H.L.Wendl.	Golden Wreath Wattle			
<i>Acacia sclerophylla</i> Lindl. var. <i>sclerophylla</i>	Hard-leaf Wattle			
<i>Acacia simmonsiana</i> O'Leary & Maslin				R
<i>Acacia spilleriana</i> J.E.Br.	Spiller's Wattle			V
<i>Acacia spinescens</i> Benth.	Spiny Wattle			
<i>Acacia stenophylla</i> A.Cunn. ex Benth.	River Cooba			
<i>Acacia Swamp</i> (N.M.Smith 3022) O'Leary	Swamp Wattle			
<i>Acacia trineura</i> F.Muell.	Three-nerve Wattle			E
<i>Acacia verniciflua</i> A.Cunn.	Varnish Wattle			
<i>Acacia verticillata</i> (L'Her.)Willd. ssp. <i>ovoidea</i> (Benth.)Court	Prickly Moses			
<i>Acacia victoriae</i> Benth. ssp. <i>victoriae</i>	Elegant Wattle			
<i>Acacia wilhelmiana</i> F.Muell.	Dwarf Nealie			
<i>Acacia X grayana</i> J.H.Willis				
* <i>Alhagi maurorum</i> Medik.	Camel Thorn			
<i>Aotus subspinescens</i> (Benth.)Crisp	Mallee Aotus			
* <i>Astragalus sesameus</i> L.	Purple Milk-vetch			
<i>Bossiaea prostrata</i> R.Br.	Creeping Bossiaea			
<i>Bossiaea walkeri</i> F.Muell.	Cactus Pea			
* <i>Caesalpinia gilliesii</i> (Wall. ex Hook.)Benth.				
* <i>Chamaecytisus palmensis</i> (H.Christ)F.A.Bisby & K.Nicholls	Tree Lucerne			
* <i>Coronilla varia</i> L.				
<i>Cullen australasicum</i> (Schltld.)J.W.Grimes	Tall Scurf-pea			
<i>Cullen cinereum</i> (Lindl.)J.W.Grimes	Annual Scurf-pea			
<i>Cullen discolor</i> (Domin)J.W.Grimes	Prostrate Scurf-pea			
<i>Cullen pallidum</i> (N.T.Burb.)J.W.Grimes	White Scurf-pea			
<i>Cullen parvum</i> (F.Muell.)J.W.Grimes				V
<i>Daviesia arenaria</i> Crisp	Sand Bitter-pea			



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<i>Daviesia benthamii</i> Meisn. ssp. <i>acanthoclona</i> (F.Muell.)Crisp	Dryland Bitter-pea			
<i>Daviesia benthamii</i> Meisn. ssp. <i>humilis</i> Crisp	Mallee Bitter-pea			R
<i>Daviesia brevifolia</i> Lindl.	Leafless Bitter-pea			
<i>Daviesia leptophylla</i> A.Cunn. ex G.Don	Narrow-leaf Bitter-pea			
<i>Daviesia ulicifolia</i> Andrews ssp. <i>aridicola</i> G.Chandler & Crisp	Gorse Bitter-pea			
<i>Daviesia ulicifolia</i> Andrews ssp. <i>incarnata</i> G.Chandler & Crisp	Gorse Bitter-pea			
<i>Dillwynia hispida</i> Lindl.	Red Parrot-pea			
<i>Dillwynia sericea</i> A.Cunn.	Showy Parrot-pea			
<i>Dillwynia uncinata</i> (Turcz.)J.M.Black	Silky Parrot-pea			
* <i>Erythrina acanthocarpa</i> E.Mey.	Thorny Coral Tree			
<i>Eutaxia diffusa</i> F.Muell.	Large-leaf Eutaxia			
<i>Eutaxia microphylla</i> (R.Br.)C.H.Wright & Dewar	Common Eutaxia			
<i>Glycine canescens</i> F.J.Herm.	Silky Glycine			
<i>Glycine rubiginosa</i> Tindale & B.E.Pfeil				
<i>Glycyrrhiza acanthocarpa</i> (Lindl.)J.M.Black	Native Liquorice			
* <i>Glycyrrhiza glabra</i> L.	Liquorice			
<i>Gompholobium ecostatum</i> Kuchel	Dwarf Wedge-pea			
<i>Goodia medicaginea</i> F.Muell.	Western Golden-tip			
<i>Hardenbergia violacea</i> (Schneev.)Stearn	Native Lilac			
<i>Indigofera australis</i> Willd. var. <i>australis</i>	Austral Indigo			
<i>Kennedia prorepens</i> (F.Muell.)F.Muell.	Desert Runner			
<i>Kennedia prostrata</i> R.Br.	Scarlet Runner			
<i>Lotus australis</i> Andrews	Austral Trefoil			
<i>Lotus cruentus</i> Court	Red-flower Lotus			
* <i>Lotus preslii</i> Ten.				
* <i>Medicago arabica</i> (L.)Huds.	Spotted Medic			
* <i>Medicago italica</i> (Mill.)Fiori				
* <i>Medicago laciniata</i> (L.)Mill.	Cut-leaf Medic			
* <i>Medicago littoralis</i> Rohde ex Loisel.	Strand Medic			
* <i>Medicago minima</i> (L.)Bartal. var. <i>minima</i>	Little Medic			
* <i>Medicago polymorpha</i> L. var. <i>polymorpha</i>	Burr-medic			
* <i>Medicago praecox</i> DC.	Small-leaf Burr-medic			
* <i>Medicago rugosa</i> Desr.	Gamma Medic			
* <i>Medicago sativa</i> L.	Lucerne			
* <i>Medicago truncatula</i> Gaertn.	Barrel Medic			
* <i>Melilotus albus</i> Medik.	Bokhara Clover			
* <i>Melilotus indicus</i> (L.)All.	King Island Melilot			
<i>Phyllota pleurandroides</i> F.Muell.	Heathy Phyllota			

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<i>Phyllota remota</i> J.H.Willis	Slender Phyllota			
<i>Platylobium obtusangulum</i> Hook.	Holly Flat-pea			
<i>Pultenaea daphnoides</i> J.C.Wendl.	Large-leaf Bush Pea			
<i>Pultenaea densifolia</i> F.Muell.	Dense Bush-pea			
<i>Pultenaea hispidula</i> R.Br. ex Benth.	Rusty Bush-pea			
<i>Pultenaea largiflorens</i> F.Muell. ex Benth.	Twiggy Bush-pea			
<i>Pultenaea pedunculata</i> Hook.	Matted Bush-pea			
<i>Pultenaea prostrata</i> Benth. ex Hook.f.	Silky Bush-pea			
<i>Pultenaea tenuifolia</i> R.Br. & Sims	Narrow-leaf Bush-pea			
* <i>Retama raetam</i> (Forssk.)Webb	White Weeping Broom			
* <i>Robinia pseudoacacia</i> L.	Black Locust			
<i>Senna artemisioides</i> (Gaudich. ex DC.)Randell ssp. X <i>artemisioides</i>	Silver Senna			
<i>Senna artemisioides</i> (Gaudich. ex DC.)Randell ssp. X <i>coriacea</i> (Benth.)Randell	Desert Senna			
<i>Senna artemisioides</i> (Gaudich. ex DC.)Randell ssp. <i>filifolia</i> Randell	Desert Senna			
<i>Senna artemisioides</i> (Gaudich. ex DC.)Randell ssp. <i>petiolaris</i> Randell	Flat-stalk Senna			
<i>Senna artemisioides</i> (Gaudich. ex DC.)Randell ssp. <i>zygophylla</i> (Benth.)Randell	Twin-leaf Desert Senna			
<i>Senna cardiosperma</i> (F.Muell.)Randell ssp. <i>gawlerensis</i>	Randell Gawler Ranges Senna			
<i>Sphaerolobium minus</i> Labill. Leafless	Globe-pea			R
* <i>Sutherlandia frutescens</i> (L.)R.Br.	Bladder Senna			
<i>Swainsona behriana</i> F.Muell. ex J.M.Black	Behr's Swainson-pea			V
<i>Swainsona colutooides</i> F.Muell.	Bladder Swainson-pea			
<i>Swainsona greyana</i> Lindl.	Darling Pea			
<i>Swainsona lessertifolia</i> DC.	Coast Swainson-pea			
<i>Swainsona microphylla</i> A.Gray	Small-leaf Swainson-pea			
<i>Swainsona oroboides</i> F.Muell. ex Benth.	Variable Swainson-pea			
<i>Swainsona oroboides</i> F.Muell. ex Benth.	Dwarf Swainson-pea			
<i>Swainsona purpurea</i> (A.T.Lee)Joy Thomps.	Purple Swainson-pea			
<i>Swainsona pyrophila</i> Joy Thomps.	Yellow Swainson-pea		V	R
<i>Swainsona reticulata</i> J.M.Black				
<i>Swainsona sericea</i> (A.T.Lee)H.Eichler	Silky Swainson-pea			E
<i>Swainsona stipularis</i> F.Muell.	Orange Swainson-pea			
<i>Swainsona tephrotricha</i> F.Muell.	Ashy-haired Swainson-pea			R
<i>Templetonia egena</i> (F.Muell.)Benth.	Broombush Templetonia			
<i>Templetonia stenophylla</i> (F.Muell.)J.M.Black	Broombush Templetonia			
<i>Templetonia sulcata</i> (Meisn.)Benth.	Flat Mallee-pea			
* <i>Trifolium angustifolium</i> L.	Narrow-leaf Clover			
* <i>Trifolium arvense</i> L. var. <i>arvense</i>	Hare's-foot Clover			
* <i>Trifolium campestre</i> Schreb.	Hop Clover			

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* <i>Trifolium cernuum</i> Brot.	Drooping-flower Clover			
* <i>Trifolium cherleri</i> L.	Cupped Clover			
* <i>Trifolium dubium</i> Sibth.	Suckling Clover			
* <i>Trifolium fragiferum</i> L. var. <i>fragiferum</i>	Strawberry Clover			
* <i>Trifolium glomeratum</i> L.	Cluster Clover			
* <i>Trifolium hirtum</i> All.	Rose Clover			
* <i>Trifolium ornithopodioides</i> L.	Bird's-foot Trefoil			
* <i>Trifolium repens</i> L.	White Clover			
* <i>Trifolium scabrum</i> L.	Rough Clover			
* <i>Trifolium striatum</i> L.	Knotted Clover			
* <i>Trifolium subterraneum</i> L. S	ubterranean Clover			
* <i>Trifolium tomentosum</i> L.	Woolly Clover			
<i>Trigonella suavissima</i> Lindl.	Sweet Fenugreek			
* <i>Ulex europaeus</i> L.	Gorse	Y		
* <i>Vicia monantha</i> Retz. ssp. <i>triflora</i> (Ten.)B.L.Burt & P.Lewis	Spurred Vetch			
* <i>Vicia sativa</i> L. ssp. <i>nigra</i> (L.)Ehrh.	Narrow-leaf Vetch			
<b>OXALIDACEAE</b>				
* <i>Oxalis bowiei</i> Ait. ex G.Don Bowie	Wood-sorrel			
* <i>Oxalis flava</i> L.	Finger-leaf Oxalis			
* <i>Oxalis hirta</i> L.	Hairy Wood-sorrel			
<i>Oxalis perennans</i> Haw.	Native Sorrel			
* <i>Oxalis pes-caprae</i> L.	Soursob	Y		
* <i>Oxalis purpurea</i> L.	Downy Wood-sorrel			
<i>Oxalis radicata</i> A.Rich.	Downy Native Sorrel			
<i>Oxalis thompsoniae</i> B.J.Conn & P.G.Richards				
<b>GERANIACEAE</b>				
* <i>Erodium aureum</i> Carolin				
* <i>Erodium botrys</i> (Cav.)Bertol.	Long Heron's-bill			
* <i>Erodium brachycarpum</i> (Godr.)Thell.	Short-fruit Heron's-bill			
* <i>Erodium cicutarium</i> (L.)L'Her. ex Aiton	Cut-leaf Heron's-bill			
<i>Erodium cunitum</i> Carolin	Blue Heron's-bill			
* <i>Erodium moschatum</i> (L.)L'Her. ex Aiton	Musky Herons-bill			
* <i>Geranium dissectum</i> L.	Cut-leaf Geranium			
* <i>Geranium molle</i> L. var. <i>molle</i>	Soft Geranium			
<i>Geranium potentilloides</i> L'Her. ex DC. var. <i>potentilloides</i>	Downy Geranium			
<i>Geranium retrorsum</i> L'Her. ex DC.	Grassland Geranium			
<i>Geranium solanderi</i> Carolin var. <i>solanderi</i>	Austral Geranium			
<i>Geranium</i> sp. Linear segments (K.Preiss 128) W.R.Barker				R

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<i>Pelargonium australe</i> Willd.	Australian Pelargonium			
<b>ZYGOPHYLLACEAE</b>				
<i>Nitraria billardierei</i> DC.	Nitre-bush			
* <i>Peganeum harmala</i> L.	African Rue	Y		
<i>Tribulus minutus</i> Leichh. ex Benth.				
* <i>Tribulus terrestris</i> L.	Caltrop	Y		
<i>Zygophyllum ammophilum</i> F.Muell.	Sand Twinleaf			
<i>Zygophyllum angustifolium</i> H.Eichler	Scrambling Twinleaf			
<i>Zygophyllum apiculatum</i> F.Muell.	Pointed Twinleaf			
<i>Zygophyllum aurantiacum</i> (Lindl.)F.Muell. ssp. <i>simplicifolium</i> H.Eichler ex R.M.Barker				
<i>Zygophyllum aurantiacum</i> (Lindl.)F.Muell. ssp. <i>aurantiacum</i>	Shrubby Twinleaf			
<i>Zygophyllum compressum</i> J.M.Black	Rabbit-ears Twinleaf			
<i>Zygophyllum confluens</i> H.Eichler	Forked Twinleaf			
<i>Zygophyllum crenatum</i> F.Muell.	Notched Twinleaf			
<i>Zygophyllum eremaeum</i> (Diels)Ostenf.	Pale-flower Twinleaf			
<i>Zygophyllum glaucum</i> F.Muell.	Pale Twinleaf			
<i>Zygophyllum iodocarpum</i> F.Muell.	Violet Twinleaf			
<i>Zygophyllum ovatum</i> Ewart & Jean White	Dwarf Twinleaf			
<i>Zygophyllum simile</i> H.Eichler	White Twinleaf			
<b>LINACEAE</b>				
<i>Linum marginale</i> A.Cunn.	Native Flax			
<b>EUPHORBIACEAE</b>				
<i>Adriana quadripartita</i> (Labill.)Mull.Arg.	Coast Bitter-bush			
<i>Adriana tomentosa</i> Gaudich. var. <i>hookeri</i> (F.Muell.)C.L.Gross & M.A.Whalen	Rare Bitter-bush			
<i>Bertya tasmanica</i> (Sond. & F.Muell.)Mull.Arg. ssp. <i>vestita</i> Halford & R.J.F.Hend.	Mitchell's Bertya			
<i>Bertya tasmanica</i> (Sond. & F.Muell.)Mull.Arg. ssp. <i>vestita</i> Halford & R.J.F.Hend.				
<i>Beyeria lechenaultii</i> (DC.)Baill.	Pale Turpentine Bush			
<i>Beyeria opaca</i> F.Muell.	Dark Turpentine Bush			
<i>Chamaesyce australis</i> (Boiss.)D.C.Hassall				
<i>Chamaesyce dallachyana</i> (Boiss.)D.C.Hassall				
<i>Chamaesyce drummondii</i> (Boiss.)D.C.Hassall				
* <i>Eremocarpus setigerus</i> (Hook.)Benth.	Doveweed			
* <i>Euphorbia dendroides</i> L.	Tree Spurge			
<i>Euphorbia parvicaruncula</i> D.C.Hassall	Tree Spurge			
* <i>Euphorbia peplus</i> L.	Petty Spurge			
<i>Euphorbia tannensis</i> Spreng. ssp. <i>eremophila</i> (A.Cunn.)D.C.Hassall	Desert Spurge			
* <i>Euphorbia terracina</i> L.	False Caper	Y		
<i>Phyllanthus fuernrohrii</i> F.Muell.	Sand Spurge			

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<i>Phyllanthus lacunarius</i> F.Muell.	Lagoon Spurge			
<i>Phyllanthus lacunellus</i> Airy Shaw				
<i>Phyllanthus maderaspatensis</i> L. var. <i>angustifolius</i> Benth.				
<i>Phyllanthus saxosus</i> F.Muell.	Rock Spurge			
<i>Poranthera microphylla</i> Brongn.	Small Poranthera			
<i>Poranthera triandra</i> J.M.Black	Three-petal Poranthera			
* <i>Ricinus communis</i> L.	Castor Oil Plant			
<i>Sauropus rigens</i> (F.Muell.)Airy Shaw	Stiff Spurge			
<i>Sauropus trachyspermus</i> (F.Muell.)Airy Shaw	Rough-seed Spurge			
<b>RUTACEAE</b>				
<i>Boronia coerulescens</i> F.Muell. ssp. <i>Coerulescens</i>	Blue Boronia			
<i>Boronia edwardsii</i> Benth.	Edwards' Boronia			
<i>Boronia inornata</i> Turcz. ssp. <i>leptophylla</i> (Turcz.)Burgman	Dryland Boronia			
<i>Correa aemula</i> (Lindl.)F.Muell.	Hairy Correa			R
<i>Correa glabra</i> Lindl. var. <i>turnbullii</i> (Ashby) Paul G.Wilson	Rock Correa			
<i>Correa reflexa</i> (Labill.)Vent. var. <i>scabridula</i> Paul G.Wilson				
<i>Philotheca angustifolia</i> (Paul G.Wilson)Paul G.Wilson ssp. <i>angustifolia</i>	Narrow-leaf Wax-flower			R
<i>Philotheca pungens</i> (Lindl.)Paul G.	Wilson Prickly Wax-flower			
<i>Geijera linearifolia</i> (DC.)J.M.Black	Sheep Bush			
<i>Geijera parviflora</i> Lindl.	Wilga			R
<i>Leonema microphyllum</i> (F.Muell.)Paul G.Wilson	Limestone Phebalium			R
<i>Microcybe multiflora</i> Turcz. ssp. <i>baccharoides</i> (F.Muell.)Paul G.Wilson	Scale-leaf Microcybe			
<i>Microcybe multiflora</i> Turcz. ssp. <i>multiflora</i>	Small-leaf Microcybe			
<i>Microcybe pauciflora</i> Turcz. ssp. <i>pauciflora</i>	Yellow Microcybe			
<i>Phebalium bullatum</i> J.M.Black	Silvery Phebalium			
<i>Zieria veronicea</i> (F.Muell.)Benth. ssp. <i>veronicea</i>	Pink Zieria			R
<b>MELIACEAE</b>				
* <i>Melia azedarach</i> L. var. <i>australasica</i> (A.Juss.)C.DC.	White Cedar			
<b>TREMANDRACEAE</b>				
<i>Tetratheca pilosa</i> Labill. ssp. <i>pilosa</i>	Hairy Pink-bells			
<b>POLYGALACEAE</b>				
<i>Comesperma calymega</i> Labill.	Blue-spike Milkwort			
<i>Comesperma polygaloides</i> F.Muell.	Mauve Milkwort			
<i>Comesperma scoparium</i> J.Drumm.	Broom Milkwort			
<i>Comesperma volubile</i> Labill.	Love Creeper			
<b>ANACARDIACEAE</b>				
* <i>Schinus molle</i> L.	Pepper-tree			

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<i>Alectryon oleifolius</i> (Desf.)S.T.Reynolds ssp. <i>canescens</i> S.T.Reynolds	Bullock Bush			
<i>Dodonaea baueri</i> Endl.	Crinkled Hop-bush			
<i>Dodonaea bursariifolia</i> F.Muell.	Small Hop-bush			
<i>Dodonaea hexandra</i> F.Muell.	Horned Hop-bush			
<i>Dodonaea hexandra</i> F.Muell. x <i>Dodonaea humilis</i> Endl.				
<i>Dodonaea humilis</i> Endl.	Dwarf Hop-bush			
<i>Dodonaea lobulata</i> F.Muell.	Lobed-leaf Hop-bush			
<i>Dodonaea procumbens</i> F.Muell.	Trailing Hopbush			V
<i>Dodonaea stenozyga</i> F.Muell.	Desert Hop-bush			
<i>Dodonaea subglandulifera</i> J.G.West			E	E
<i>Dodonaea tepperi</i> F.Muell. ex Tepper	Streaked Hop-bush			
<i>Dodonaea viscosa</i> Jacq. ssp. <i>angustissima</i> (DC.)J.G.West	Narrow-leaf Hop-bush			
<i>Dodonaea viscosa</i> Jacq. ssp. <i>cuneata</i> (Sm.)J.G.West	Wedge-leaf Hop-bush			
<i>Dodonaea viscosa</i> Jacq. ssp. <i>spatulata</i> (Sm.)J.G.West	Sticky Hop-bush			
<b>STACKHOUSIACEAE</b>				
<i>Stackhousia annua</i> W.R.Barker	Annual Candles			V
<i>Stackhousia aspericocca</i> Schuch. ssp. Cylindrical inflorescence (W.R.Barker 1418) <i>W.R.Barker</i>	Bushy Candles			
<i>Stackhousia aspericocca</i> Schuch. ssp. One-sided inflorescence (W.R.Barker 697) <i>W.R.Barker</i>	One-sided Candles			
<i>Stackhousia megaloptera</i> F.Muell.	Dune Candles			
<i>Stackhousia monogyna</i> Labill.	Creamy Candles			
<b>RHAMNACEAE</b>				
<i>Cryptandra propinqua</i> A.Cunn. ex Fenzl	Silky Cryptandra			
<i>Cryptandra</i> sp. Floriferous (W.R.Barker 4131) <i>W.R.Barker</i>				
<i>Cryptandra</i> sp. Long hypanthium (C.R. Alcock 10626) <i>W.R.Barker</i>				R
<i>Cryptandra tomentosa</i> Lindl.	Heath Cryptandra			
<i>Pomaderris obcordata</i> Fenzl	Wedge-leaf Pomaderris			
<i>Pomaderris paniculosa</i> F.Muell. ex Reissek ssp. <i>paniculosa</i>	Mallee Pomaderris			
* <i>Rhamnus alaternus</i> L.	Blowfly Bush			
<i>Spyridium bifidum</i> (F.Muell.)F.Muell. ex Benth. var. <i>bifidum</i>	Forked Spyridium			
<i>Spyridium eriocephalum</i> Fenzl var. <i>eriocephalum</i>	Heath Spyridium			
<i>Spyridium parvifolium</i> (Hook.)Benth. ex F.Muell.	Dusty Miller			
<i>Spyridium phyllicoides</i> Reissek	Narrow-leaf Spyridium			
<i>Spyridium subochreatum</i> (F.Muell.)Reissek var. <i>laxiusculum</i> J.M.Black	Velvet Spyridium			
<i>Spyridium subochreatum</i> (F.Muell.)Reissek var. <i>subochreatum</i>	Velvet Spyridium			
<i>Stenanthemum leucophractum</i> (Schltdl.)Reissek	White Cryptandra			
<i>Stenanthemum notiale</i> Rye ssp. <i>Notiale</i>				
<i>Trymalium wayae</i> F.Muell. & Tate	Grey Trymalium			
<b>MALVACEAE</b>				



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<i>Abutilon fraseri</i> (Hook.)Hook. ex Walp.	Dwarf Lantern-bush			
<i>Abutilon malvaefolium</i> (Benth.)J.M.Black	Scrambling Lantern-bush			
<i>Abutilon otocarpum</i> F.Muell.	Desert Lantern-bush			
<i>Abutilon theophrasti</i> Medik.	Swamp Lantern-bush			
* <i>Alcea rosea</i> L.	Hollyhock			
<i>Hibiscus krichauffianus</i> F.Muell.	Velvet-leaf Hibiscus			
* <i>Hibiscus trionum</i> L. var. <i>trionum</i>	Bladder Ketmia			
<i>Lawrenzia berthae</i> (F.Muell.)Melville	Showy Lawrenzia			R
<i>Lawrenzia glomerata</i> Hook.	Clustered Lawrenzia			
<i>Lawrenzia squamata</i> Nees	Thorny Lawrenzia			
<i>Malva behriana</i> Schldl.	Australian Hollyhock			
* <i>Malva dendromorpha</i> M.F.Ray	Tree Mallow			
* <i>Malva parviflora</i> L.	Small-flower Marshmallow			
<i>Malvastrum americanum</i> (L.)Torr. var. <i>americanum</i>	Malvastrum			
* <i>Malvella leprosa</i> (Ortega)Krapov.	Alkali Sida	Y		
* <i>Modiola caroliniana</i> (L.)G.Don	Red-flowered Mallow			
* <i>Pavonia hastata</i> Cav.	Pink Pavonia			
<i>Radyera farragei</i> (F.Muell.)Fryxell & S.H.Hashmi	Desert Rose Mallow			
<i>Sida ammophila</i> F.Muell. ex J.H. Willis	Sand Sida			
<i>Sida corrugata</i> Lindl. var. A (N.N.Donner 7573) : W.R.Barker				
<i>Sida corrugata</i> Lindl. var. <i>angustifolia</i> Benth.	Grassland Sida			
<i>Sida corrugata</i> Lindl. var. <i>corrugata</i>	Corrugated Sida			
<i>Sida fibulifera</i> Lindl.	Pin Sida			
<i>Sida intricata</i> F.Muell.	Twiggy Sida			
<i>Sida petrophila</i> F.Muell.	Rock Sida			
<i>Sida</i> sp. Paringa (R.Bates 13800) W.R.Barker				
<i>Sida spodochroma</i> F.Muell.				
<i>Sida trichopoda</i> F.Muell.	High Sida			
<b>STERCULIACEAE</b>				
* <i>Brachychiton populneus</i> (Schott & Endl.)R.Br. ssp. <i>Populneus</i>				
<i>Lasiopetalum baueri</i> Steetz	Slender Velvet-bush			
<i>Lasiopetalum behrii</i> F.Muell.	Pink Velvet-bush			
<i>Rulingia loxophylla</i> F.Muell.	Pur-ar-purpa			
<i>Thomasia petalocalyx</i> F.Muell.	Paper-flower			
<b>THYMELAEACEAE</b>				
<i>Pimelea flava</i> R.Br. ssp. <i>dichotoma</i> (Schldl.)Threlfall	Diosma Riceflower			
<i>Pimelea glauca</i> R.Br.	Smooth Riceflower			
<i>Pimelea humilis</i> R.Br.	Low Riceflower			

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<i>Pimelea imbricata</i> R.Br. var. <i>petraea</i> (Meisn.)Rye	Rock Woolly Riceflower			
<i>Pimelea micrantha</i> F.Muell. ex Meisn.	Silky Riceflower			
<i>Pimelea microcephala</i> R.Br. ssp. <i>microcephala</i>	Shrubby Riceflower			
<i>Pimelea octophylla</i> R.Br.	Woolly Riceflower			
<i>Pimelea phyllicoides</i> Meisn.	Heath Riceflower			
<i>Pimelea serpyllifolia</i> R.Br. ssp. <i>serpyllifolia</i>	Thyme Riceflower			
<i>Pimelea simplex</i> F.Muell. ssp. <i>continua</i> (J.M.Black)Threlfall	Desert Riceflower			
<i>Pimelea simplex</i> F.Muell. ssp. <i>Simplex</i>	Desert Riceflower			
<i>Pimelea stricta</i> Meisn.	Erect Riceflower			
<i>Pimelea trichostachya</i> Lindl.	Spiked Riceflower			
<i>Pimelea williamsonii</i> J.M.Black	Williamson's Riceflower			R
* <i>Thymelaea passerina</i> (L.)Coss. & Germ.	Thymelaea			
<b>VIOLACEAE</b>				
<i>Hybanthus floribundus</i> (Lindl.)F.Muell. ssp. <i>floribundus</i>	Shrub Violet			
<i>Hybanthus monopetalus</i> (Schult.)Domin	Slender Violet			
<i>Hymenanthera dentata</i> R.Br. ex DC.	Tree Violet			
<i>Viola eminens</i> K.R.Thiele & Prober				
<i>Viola sieberiana</i> Spreng.	Tiny Violet			
<b>TAMARICACEAE</b>				
* <i>Tamarix aphylla</i> (L.)H.Karst.	Athel Pine			
* <i>Tamarix parviflora</i> DC.				
<b>FRANKENIACEAE</b>				
<i>Frankenia foliosa</i> J.M.Black	Leafy Sea-heath			
<i>Frankenia pauciflora</i> DC. var. <i>gunnii</i> Summerh.	Southern Sea-heath			
* <i>Frankenia pulverulenta</i> L.	Mediterranean Sea-heath			
<i>Frankenia serpyllifolia</i> Lindl.	Thyme Sea-heath			
<i>Frankenia sessilis</i> Summerh.	Small-leaf Sea-heath			
<b>ELATINACEAE</b>				
<i>Bergia trimera</i> Fisch. & C.A.Mey.	Three-part Water-fire			
<i>Elatine gratioloides</i> A.Cunn.	Waterwort			R
<b>CUCURBITACEAE</b>				
* <i>Citrullus colocynthis</i> (L.)Schrad.	Colocynth			
* <i>Citrullus lanatus</i> (Thunb.)Matsum. & Nakai	Bitter Melon			
<i>Cucumis melo</i> L.	Ulcardo Melon			
* <i>Cucumis myriocarpus</i> Naudin	Paddy Melon			
* <i>Ecballium elaterium</i> (L.)A.Rich.	Squirting Cucumber			
<i>Mukia micrantha</i> (F.Muell.)F.Muell.	Desert Cucumber			
<b>LYTHRACEAE</b>				

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<i>Ammannia multiflora</i> Roxb.	Jerry-jerry			
<i>Lythrum hyssopifolia</i> L.	Lesser Loosestrife			
<i>Lythrum salicaria</i> L.	Purple Loosestrife			R
<b>MYRTACEAE</b>				
<i>Babingtonia behrii</i> (Schltdl.)A.R.Bean	Silver Broombrush			
<i>Baeckea crassifolia</i> Lindl.	Desert Baeckea			
<i>Baeckea ericaea</i> (F.Muell.)Benth.	Mat Baeckea			
<i>Callistemon brachyandrus</i> Lindl.	Prickly Bottlebrush			R
<i>Callistemon rugulosus</i> (D.F.K.Schltdl. ex Link)DC.	Scarlet Bottlebrush			
<i>Callistemon sieberi</i> DC.	River Bottlebrush			
<i>Callistemon teretifolius</i> F.Muell.	Needle Bottlebrush			
<i>Calytrix alpestris</i> (Lindl.)Court	Snow Heath-myrtle			
<i>Calytrix involucrata</i> J.M.Black	Cup Fringe-myrtle			
<i>Calytrix tetragona</i> Labill.	Common Fringe-myrtle			
<i>Eucalyptus arenacea</i> Marginson & Ladiges	Dune Stringybark			
<i>Eucalyptus behriana</i> F.Muell.	Broad-leaf Box			R
<i>Eucalyptus brachycalyx</i> Blakely	Gilja			
<i>Eucalyptus calycogona</i> Turcz. ssp. <i>trachybasis</i> D.Nicolle	Square-fruit Mallee			
<i>Eucalyptus camaldulensis</i> Dehnh. var. <i>camaldulensis</i>	River Red Gum			
<i>Eucalyptus cyanophylla</i> Brooker	Blue-leaf Mallee			
<i>Eucalyptus diversifolia</i> Bonpl. ssp. <i>diversifolia</i>	Coastal White Mallee			
<i>Eucalyptus dumosa</i> A.Cunn. ex Oxley	White Mallee			
<i>Eucalyptus fasciculosa</i> F.Muell.	Pink Gum			R
<i>Eucalyptus gracilis</i> F.Muell.	Yorrell			
<i>Eucalyptus incrassata</i> Labill.	Ridge-fruited Mallee			
<i>Eucalyptus largiflorens</i> F.Muell.	River Box			
<i>Eucalyptus leptophylla</i> F.Muell. ex Miq.	Narrow-leaf Red Mallee			
<i>Eucalyptus leucoxylon</i> F.Muell. ssp. <i>leucoxylon</i>	South Australian Blue Gum			
<i>Eucalyptus leucoxylon</i> F.Muell. ssp. <i>pruinosa</i> (F.Muell. ex Miq.)Boland	Inland South Australian Blue Gum			
<i>Eucalyptus leucoxylon</i> F.Muell. ssp. <i>stephaniae</i> Rule	Scrubby Blue Gum			
<i>Eucalyptus microcarpa</i> (Maiden)Maiden	Grey Box			
<i>Eucalyptus odorata</i> Behr	Peppermint Box			
<i>Eucalyptus oleosa</i> F.Muell. ex Miq. ssp. <i>oleosa</i>	Red Mallee			
<i>Eucalyptus phenax</i> Brooker & Slee ssp. <i>phenax</i>				
<i>Eucalyptus porosa</i> F.Muell. ex Miq.	Mallee Box			
<i>Eucalyptus rugosa</i> R.Br. ex Blakely	Coastal White Mallee			
<i>Eucalyptus socialis</i> F.Muell. ex Miq. ssp. Glossy green (P.J.Lang 804) D.Nicolle				
<i>Eucalyptus socialis</i> F.Muell. ex Miq. ssp. <i>socialis</i>	Beaked Red Mallee			

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<i>Eucalyptus viminalis</i> Labill. ssp. <i>cygnetensis</i> Boomsma	Rough-bark Manna Gum			
<i>Eucalyptus viridis</i> R.T.Baker ssp. <i>wimmerensis</i> (Rule) Brooker & Slee				R
<i>Eucalyptus yalataensis</i> Boomsma	Yalata Mallee			
<i>Kunzea pomifera</i> F.Muell.	Muntries			
<i>Leptospermum coriaceum</i> (F.Muell. ex Miq.)Cheel	Dune Tea-tree			
<i>Leptospermum lanigerum</i> (Sol. ex Aiton)Sm.	Silky Tea-tree			
<i>Leptospermum myrsinoides</i> Schltl.	Heath Tea-tree			
<i>Melaleuca brevifolia</i> Turcz.	Short-leaf Honey-myrtle			
<i>Melaleuca halmaturorum</i> F.Muell. ex Miq.	Swamp Paper-bark			
<i>Melaleuca lanceolata</i> Otto	Dryland Tea-tree			
<i>Melaleuca pauperiflora</i> F.Muell. ssp. <i>mutica</i> Barlow	Boree			
<i>Melaleuca uncinata</i> R.Br.	Broombush			
<i>Melaleuca xerophila</i> Barlow	Boree			
<i>Micromyrtus ciliata</i> (Sm.)Druce	Fringed Heath-myrtle			R
<i>Thryptomene calycina</i> (Lindl.)Stapf	Grampians Thryptomene			
<b>ONAGRACEAE</b>				
<i>Epilobium billardierianum</i> Ser. ssp. <i>X intermedium</i> P.H.Raven & Engelhorn	Variable Willow-herb			
<i>Epilobium billardierianum</i> Ser. ssp. <i>billardierianum</i>	Robust Willow-herb			
<i>Epilobium hirtigerum</i> A.Cunn.	Hairy Willow-herb			
<i>Epilobium pallidiflorum</i> Sol. ex A.Cunn.	Showy Willow-herb			
* <i>Ludwigia peploides</i> (Kunth)P.H.Raven ssp. <i>montevidensis</i> (Spreng.)P.H.Raven	Water Primrose			
* <i>Oenothera stricta</i> Ledeb. ex Link ssp. <i>stricta</i>	Common Evening Primrose			
<b>HALORAGACEAE</b>				
<i>Glischrocaryon behrii</i> (Schltl.)Orchard	Golden Pennants			
<i>Glischrocaryon flavescens</i> (J.Drumm.)Orchard	Yellow Pennants			
<i>Gonocarpus elatus</i> (A.Cunn. ex Fenzl)Orchard	Hill Raspwort			
<i>Gonocarpus tetragynus</i> Labill.	Small-leaf Raspwort			
<i>Haloragis acutangula</i> F.Muell. f. <i>acutangula</i>	Smooth Raspwort			
<i>Haloragis acutangula</i> F.Muell. f. <i>inflata</i> Orchard	Smooth Raspwort			
<i>Haloragis acutangula</i> F.Muell. f. <i>tetraptera</i> Orchard	Smooth Raspwort			
<i>Haloragis acutangula</i> F.Muell. f. <i>turbinata</i> Orchard	Smooth Raspwort			
<i>Haloragis aspera</i> Lindl.	Rough Raspwort			
<i>Haloragis eichleri</i> Orchard	Eichler's Raspwort			R
<i>Haloragis glauca</i> Lindl. f. <i>glauca</i>	Bluish Raspwort			
<i>Haloragis heterophylla</i> Brongn.	Variable Raspwort			
<i>Haloragis odontocarpa</i> F.Muell. f. <i>pterocarpa</i> Orchard	Mulga Nettle			
* <i>Myriophyllum aquaticum</i> (Vell.)Verdc.				
<i>Myriophyllum caput-medusae</i> Orchard	Coarse Milfoil			

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<i>Myriophyllum crispatum</i> Orchard	Upright Milfoil			V
<i>Myriophyllum integrifolium</i> (Hook.f.)Hook.f.	Tiny Milfoil			R
<i>Myriophyllum papillosum</i> Orchard	Robust Milfoil			R
<i>Myriophyllum simulans</i> Orchard	Amphibious Milfoil			
<i>Myriophyllum variifolium</i> Hook.f.	Varied Milfoil			R
<i>Myriophyllum verrucosum</i> Lindl.	Red Milfoil			
<b>UMBELLIFERAE</b>				
* <i>Ammi majus</i> L.	False Bishop's Weed			
<i>Apium annuum</i> P.S.Short	Annual Celery			
* <i>Apium graveolens</i> L.	Celery			
<i>Apium prostratum</i> Labill. ex Vent. var. <i>filiforme</i> (A.Rich.)Kirk	Native Celery			
<i>Apium prostratum</i> Labill. ex Vent. var. <i>prostratum</i>	Native Celery			
* <i>Berula erecta</i> (Huds.)Coville	Water Parsnip			
* <i>Bupleurum semicompositum</i> L.	Hare's Ear			
<i>Centella asiatica</i> (L.)Urb.	Asian Centella			
* <i>Ciclospermum leptophyllum</i> (Pers.)Sprague	Narrow-leaf Celery			
* <i>Conium maculatum</i> L.	Hemlock			
* <i>Coriandrum sativum</i> L.	Coriander			
* <i>Daucus carota</i> L.	Carrot	Y		
* <i>Foeniculum vulgare</i> Mill.	Fennel			
<i>Hydrocotyle callicarpa</i> Bunge	Tiny Pennywort			
<i>Hydrocotyle capillaris</i> F.Muell. ex Klatt	Thread Pennywort			
<i>Hydrocotyle foveolata</i> H.Eichler	Yellow Pennywort			
<i>Hydrocotyle hirta</i> R.Br. ex A.Rich.	Hairy Pennywort			
<i>Hydrocotyle laxiflora</i> DC.	Stinking Pennywort			
<i>Hydrocotyle pilifera</i> Turcz. var. <i>glabrata</i> Benth.	Buttercup Pennywort			
<i>Hydrocotyle rugulosa</i> Turcz.	Mallee Pennywort			
<i>Hydrocotyle verticillata</i> Thunb.	Shield Pennywort			
<i>Lilaeopsis polyantha</i> (Gand.)H.Eichler	Australian Lilaeopsis			
<i>Trachymene anisocarpa</i> (Turcz.)B.L.Burt	Native Parsnip			R
<i>Trachymene cyanopetala</i> (F.Muell.)Benth.	Purple Trachymene			
<i>Trachymene pilosa</i> Sm.	Dwarf Trachymene			
<i>Xanthosia huegelii</i> (Benth.)Steud.				
<i>Xanthosia leiophylla</i> Klatt				
<b>EPACRIDACEAE</b>				
<i>Acrotriche affinis</i> DC.	Ridged Ground-berry			
<i>Acrotriche cordata</i> (Labill.)R.Br.	Blunt-leaf Ground-berry			
<i>Acrotriche depressa</i> R.Br.	Native Currant			

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<i>Acrotriche patula</i> R.Br.	Prickly Ground-berry			
<i>Acrotriche serrulata</i> (Labill.)R.Br.	Cranberry Heath			
<i>Brachyloma daphnoides</i> (Sm.)Benth.	Daphne Heath			
<i>Brachyloma ericoides</i> (Schltdl.)Sond. ssp. <i>ericoides</i>	Brush Heath			
<i>Leucopogon clelandii</i> Cheel	Cleland's Beard-heath			R
<i>Leucopogon cordifolius</i> Lindl.	Heart-leaf Beard-heath			
<i>Leucopogon costatus</i> (F.Muell.)F.Muell.ex J.M.Black	Twiggy Beard-heath			
<i>Leucopogon ericoides</i> (Sm.)R.Br.	Pink Beard-heath			
<i>Leucopogon rufus</i> Lindl.	Ruddy Beard-heath			
<i>Leucopogon virgatus</i> (Labill.)R.Br. var. <i>virgatus</i>	Common Beard-heath			
<i>Leucopogon woodsii</i> F.Muell.	Nodding Beard-heath			
<i>Lissanthe strigosa</i> (Sm.)R.Br. ssp. <i>subulata</i> (R.Br.)J.M.Powell	Peach Heath			
<i>Styphelia exarrhena</i> (F.Muell.)F.Muell.	Desert Heath			
<b>PRIMULACEAE</b>				
* <i>Anagallis minima</i> (L.)K.Krause	Chaffweed			
* <i>Anchusa arvensis</i> (L.)M.Bieb.	Pimpernel			
* <i>Asterolinon linum-stellatum</i> (L.)Duby	Asterolinon			
<i>Samolus repens</i> (J.R.Forst. & G.Forst.)Pers.	Creeping Brookweed			
<b>LIMONIACEAE</b>				
* <i>Limonium binervosum</i> (G.E.Sm.)C.E.Salmon	Dwarf Sea-lavender			
* <i>Limonium companyonis</i> (Gren. & Billot)Kuntze	Sea-lavender			
* <i>Limonium hyblaenum</i> Brullo				
* <i>Limonium lobatum</i> (L.f.)Kuntze	Winged Sea-lavender			
* <i>Limonium myrianthum</i> (Schrenk)Kuntze				
* <i>Limonium sinuatum</i> (L.)Mill.	Notch-leaf Sea-lavender			
<b>OLEACEAE</b>				
* <i>Fraxinus angustifolia</i> Vahl	Desert Ash			
<i>Jasminum didymum</i> G.Forst. ssp. <i>lineare</i> (R.Br.) P.Green	Native Jasmine			
* <i>Olea europaea</i> L. ssp. <i>europaea</i>	Olive	Y		
<b>LOGANIACEAE</b>				
<i>Logania linifolia</i> Schltdl.	Flax-leaf Logania			
<i>Logania nuda</i> F.Muell.	Leafless Logania			
<i>Logania saxatilis</i> G.Perry ex B.J.Conn	Rock Logania			R
<i>Phyllangium distylis</i> (F.Muell.)Dunlop				R
<i>Phyllangium divergens</i> (Hook.f.)Dunlop	Wiry Mitrewort			
<b>BUDDLEJACEAE</b>				
* <i>Buddleja madagascariensis</i> Lam.	Summer Lilac			
<b>GENTIANACEAE</b>				



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* <i>Centaureum pulchellum</i> (Sw.)Druce				
* <i>Centaureum spicatum</i> (L.)Fritsch ex Janch.	Spike Centaury			
* <i>Centaureum tenuiflorum</i> (Hoffmanns. & Link)Fritsch	Branched Centaury			
* <i>Cicendia filiformis</i> (L.)Delarbre	Slender Cicendia			
<i>Sebaea ovata</i> (Labill.)R.Br.	Yellow Sebaea			
<b>MENYANTHACEAE</b>				
* <i>Nymphaea alba</i> L.	White Water-lily			
<i>Nymphoides crenata</i> (F.Muell.)Kuntze	Wavy Marshwort			R
<i>Villarsia umbricola</i> Aston var. <i>umbricola</i>	Lax Marsh-flower			
<b>APOCYNACEAE</b>				
<i>Alyxia buxifolia</i> R.Br.	Sea Box			
* <i>Vinca major</i> L.	Blue Periwinkle			
<b>ASCLEPIADACEAE</b>				
* <i>Gomphocarpus cancellatus</i> (Burm.f.)Bruyns	Broad-leaf Cotton-bush			
* <i>Gomphocarpus fruticosus</i> (L.)W.T.Aiton	Narrow-leaf Cotton-bush			
<i>Marsdenia australis</i> (R.Br.)Druce	Native Pear			
<i>Rhyncharhena linearis</i> (Decne.)K.L.Wilson	Climbing Purple-star			
<i>Sarcostemma viminalis</i> (L.)R.Br. ssp. <i>australe</i> (R.Br.)P.I.Forst.	Caustic Bush			
<b>RUBIACEAE</b>				
<i>Asperula conferta</i> Hook.f.	Common Woodruff			
<i>Asperula Gemella</i> Airy Shaw & Turnill	Twin-leaf Bedstraw			
* <i>Galium divaricatum</i> Lam.	Slender Bedstraw			
<i>Galium gaudichaudii</i> DC.	Rough Bedstraw			
<i>Galium migrans</i> Ehrend. & McGill.	Loose Bedstraw			
* <i>Galium murale</i> (L.)All.	Small Bedstraw			
* <i>Galium palustre</i> L.				
* <i>Galium spurium</i> L. ssp. <i>ibicinum</i> (Boiss. & Hausskn. ex Boiss.)Ehrend.	Bedstraw			
<i>Opercularia ovata</i> Hook.f.	Broad-leaf Stinkweed			
<i>Opercularia scabrida</i> Schldtl.	Stalked Stinkweed			
<i>Opercularia turpis</i> F.Muell. ex Miq.	Twiggy Stinkweed			
* <i>Sherardia arvensis</i> L.	Field Madder			
<b>CONVOLVULACEAE</b>				
<i>Calystegia sepium</i> (L.)R.Br.	Large Bindweed			
<i>Convolvulus angustissimus</i> R.Br. ssp. <i>angustissimus</i>				
<i>Convolvulus angustissimus</i> R.Br. ssp. <i>peninsularum</i> R.W.Johnson				
<i>Convolvulus clementii</i> Domin				
<i>Convolvulus crispifolius</i> F.Muell.				
<i>Convolvulus eyreanus</i> R.W.Johnson	Silver Bindweed			

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<i>Convolvulus microsepalus</i> R.W.Johnson	Small-flower Bindweed			
<i>Convolvulus recurvatus</i> R.W.Johnson ssp. <i>nullarborensis</i> R.W.Johnson				
<i>Convolvulus recurvatus</i> R.W.Johnson ssp. <i>Recurvatus</i>				
<i>Convolvulus remotus</i> R.Br.	Grassy Bindweed			
<i>Cressa cretica</i> L.	Rosinweed			
* <i>Cuscuta campestris</i> Yunck.	Golden Dodder	Y		
* <i>Cuscuta epithymum</i> (L.)L.	Lesser Dodder			
<i>Dichondra repens</i> J.R.Forst. & G.Forst.	Kidney Weed			
* <i>Ipomoea indica</i> (Burm.)Merr.	Purple Morning-glory			
* <i>Ipomoea pandurata</i> (L.)G.Mey.				
<i>Merremia dissecta</i> (Jacq.)Hallier f.				
<i>Wilsonia rotundifolia</i> Hook.	Round-leaf Wilsonia			
<b>BORAGINACEAE</b>				
* <i>Amsinckia calycina</i> (Moris)Chater	Hairy Fiddle-neck	Y		
* <i>Anchusa arvensis</i> (L.)M.Bieb.	Bugloss			
* <i>Borago officinalis</i> L.	Borage			
* <i>Buglossoides arvensis</i> (L.)I.M.Johnst.	Sheepweed			
<i>Cynoglossum australe</i> R.Br.	Australian Hound's-tongue			
* <i>Echium italicum</i> L.	Italian Bugloss			
* <i>Echium plantagineum</i> L.	Salvation Jane	Y		
* <i>Eryngium campestre</i> L.	White Devil			
<i>Eryngium plantagineum</i> F.Muell.				
<i>Eryngium vesiculosum</i> Labill.	long Eryngium			R
<i>Halgania andromedifolia</i> Behr & F.Muell.	Scented Blue-flower			
<i>Halgania cyanea</i> Lindl.	Rough Blue-flower			
* <i>Heliotropium amplexicaule</i> Vahl	Blue Heliotrope			
<i>Heliotropium asperinum</i> R.Br.	Rough Heliotrope			
* <i>Heliotropium curassavicum</i> L.	Smooth Heliotrope			
* <i>Heliotropium europaeum</i> L.	Common Heliotrope			
* <i>Heliotropium supinum</i> L.	Creeping Heliotrope			
<i>Myosotis australis</i> R.Br.	Austral Forget-me-not			
* <i>Neatostema apulum</i> (L.)I.M.Johnst.	Hairy Sheepweed			
<i>Omphalolappula concava</i> (F.Muell.)Brand	Burr Stickseed			
<i>Plagiobothrys elachanthus</i> (F.Muell.)I.M.Johnst.	Hairy Forget-me-not			
<i>Plagiobothrys plurisepalus</i> (F.Muell.)I.M.Johnst.	White Rochelia			
<b>VERBENACEAE</b>				
* <i>Phyla canescens</i> (Kunth)Greene	Lippia			
* <i>Verbena officinalis</i> L.	Common Verbena			

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* <i>Verbena supina</i> L. var. <i>erecta</i> (Moldenke)Munir	Trailing Verbena			
* <i>Verbena supina</i> L. var. <i>supina</i>	Trailing Verbena			
<b>CHLOANTHACEAE</b>				
<i>Dicrastylis verticillata</i> J.M.Black	Whorled Sand-sage			
<b>CALLITRICHACEAE</b>				
<i>Callitriche sonderi</i> Hegelm.	Matted Water Starwort			R
* <i>Callitriche stagnalis</i> Scop.	Common Water Starwort			
<i>Callitriche umbonata</i> Hegelm.	Water Starwort			V
<b>LABIATAE</b>				
<i>Ajuga australis</i> R.Br. f.A (A.G.Spooner 9058) Toelken	Australian Bugle			
* <i>Ajuga iva</i> (L.)Schreb.	Bugle			
* <i>Lamium amplexicaule</i> L. var. <i>amplexicaule</i>	Deadnettle			
* <i>Lavandula dentata</i> L.	French Lavender			
* <i>Lavandula stoechas</i> L.	Topped Lavender			
<i>Lycopus australis</i> R.Br.	Australian Gipsywort			
* <i>Marrubium vulgare</i> L.	Horehound	Y		
* <i>Melissa officinalis</i> L.	Common Balm			
<i>Mentha australis</i> R.Br.	River Mint			
* <i>Mentha pulegium</i> L.	Pennyroyal			
* <i>Mentha X piperita</i> L. var. <i>citrata</i> (Ehrh.)Briq.	Lemon Mint			
* <i>Mentha spicata</i> L. f.A (R.Bates 3655) Toelken				
* <i>Mentha spicata</i> L. f.B (B.Copley 1119) Toelken				
<i>Prostanthera aspalathoides</i> A.Cunn. ex Benth.	Scarlet Mintbush			
<i>Prostanthera behriana</i> Schldtl.	Downy Mintbush			
<i>Prostanthera chlorantha</i> (F.Muell.)F.Muell. ex Benth.	Green Mintbush			R
<i>Prostanthera eurybioides</i> F.Muell.	Monarto Mintbush		E	E
<i>Prostanthera serpyllifolia</i> (R.Br.)Briq. ssp. <i>microphylla</i> (R.Br.)B.J.Conn	Small-leaf Mintbush			
<i>Prostanthera striatiflora</i> F.Muell.	Striated Mintbush			
* <i>Salvia verbenaca</i> L. var. <i>verbenaca</i>	Wild Sage			
* <i>Salvia verbenaca</i> L. var. <i>vernalis</i> Boiss.	Wild Sage			
<i>Scutellaria humilis</i> R.Br.	Dwarf Skullcap			R
* <i>Stachys arvensis</i> (L.)L.	Stagger Weed			
<i>Teucrium albicaule</i> Toelken	Scurfy Germander			
<i>Teucrium corymbosum</i> R.Br.	Rock Germander			
<i>Teucrium racemosum</i> R.Br.	Grey Germander			
<i>Teucrium sessiliflorum</i> Benth.	Mallee Germander			
<i>Westringia eremicola</i> A.Cunn. ex Benth.	Slender Westringia			
<i>Westringia rigida</i> R.Br.	Stiff Westringia			

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<b>SOLANACEAE</b>				
* <i>Cestrum parqui</i> L'Her.	Green Poison-berry			
<i>Cyphanthera myosotidea</i> (F.Muell.)Haegi	Small-leaf Ray-flower			
* <i>Datura ferox</i> L.	Long-spine Thorn-apple			
* <i>Datura inoxia</i> Mill.	Downy Thorn-apple			
* <i>Datura stramonium</i> L.	Common Thorn-apple			
* <i>Datura wrightii</i> Regel	Hairy Thorn-apple			
<i>Duboisia hopwoodii</i> (F.Muell.)F.Muell.	Pituri			
<i>Grammosolen dixonii</i> (F.Muell. & Tate)Haegi	Dixon's Ray-flower			
<i>Lycium australe</i> F.Muell.	Australian Boxthorn			
* <i>Lycium ferocissimum</i> Miers	African Boxthorn	Y		
* <i>Nicotiana glauca</i> Graham	Tree Tobacco			
<i>Nicotiana goodspeedii</i> H.-M.Wheeler	Small-flower Tobacco			
<i>Nicotiana maritima</i> H.-M.Wheeler	Coast Tobacco			
<i>Nicotiana occidentalis</i> H.-M.Wheeler ssp. <i>obliqua</i> N.T.Burb.	Western Tobacco			
<i>Nicotiana velutina</i> H.-M.Wheeler	Velvet Tobacco			
* <i>Physalis viscosa</i> L.	Sticky Cape Gooseberry			
* <i>Salpichroa origanifolia</i> (Lam.)Thell.	Pampas Lily-of-the-valley			
<i>Solanum capsiciforme</i> (Domin)G.T.S.Baylis	Capsicum Kangaroo-apple			
<i>Solanum cleistogamum</i> Symon	Shy Nightshade			
<i>Solanum coactiliferum</i> J.M.Black	Tomato-bush			
<i>Solanum elaeagnifolium</i> Cav.	Silver-leaf Nightshade	Y		
<i>Solanum ellipticum</i> R.Br.	Velvet Potato-bush			
<i>Solanum esuriale</i> Lindl.	Quena			
<i>Solanum laciniatum</i> Aiton	Cut-leaf Kangaroo-apple			
<i>Solanum lacunarium</i> F.Muell.	Lagoon Nightshade			
* <i>Solanum linnaeanum</i> Hepper & P.-M.L.Jaeger	Apple Of Sodom			
* <i>Solanum marginatum</i> L.f.	White-edged Nightshade			
* <i>Solanum nigrum</i> L. Black	Nightshade			
<i>Solanum opacum</i> A.Braun & Bouche	Green-berry Nightshade			
<i>Solanum petrophilum</i> F.Muell.	Rock Nightshade			
* <i>Solanum physalifolium</i> Rusby var. <i>nitidibaccatum</i> (Bitter)Edmonds				
* <i>Solanum rostratum</i> Dunal	Buffalo Burr			
<i>Solanum simile</i> F.Muell.	Kangaroo Apple			
<i>Solanum sturtianum</i> F.Muell.	Sturt's Nightshade			
* <i>Solanum triflorum</i> Nutt.	Three-flower Nightshade			
<b>SCROPHULARIACEAE</b>				
* <i>Bartsia trixago</i> L.				

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<i>Derwentia decorosa</i> (F.Muell.)B.G.Briggs & Ehrend.	Showy Speedwell			R
* <i>Dischisma capitatum</i> (Thunb.)Choisy	Wooly-head Dichisma			
<i>Euphrasia collina</i> R.Br. ssp. <i>osbornii</i> W.R.Barker	Osborn's Eyebright		E	E
<i>Euphrasia collina</i> R.Br. ssp. <i>tetragona</i> (R.Br.)W.R.Barker	Coast Eyebright			
<i>Glossostigma cleistanthum</i> W.R.Barker	Spoon Mud-mat			
<i>Glossostigma diantrum</i> (L.)Kuntze	Two-anther Mud-mat			
<i>Glossostigma elatinoides</i> (Benth.)Benth. ex Hook.f.	Small Mud-mat			R
<i>Gratiola pedunculata</i> R.Br.	Stalked brooklime			
<i>Gratiola peruviana</i> L.	Austral Brooklime			
<i>Gratiola pumilo</i> F.Muell.	Dwarf Brooklime			R
* <i>Kickxia elatine</i> (L.)Dumort. ssp. <i>crinita</i> (Mabille)Greuter	Twining Toadflax			
<i>Limosella australis</i> R.Br.	Australian Mudwort			
<i>Limosella curdieana</i> F.Muell. var. <i>curdieana</i>	Large Mudwort			
<i>Mimulus prostratus</i> Benth.	Small Monkey-flower R			
<i>Mimulus repens</i> R.Br.	Creeping Monkey-flower			
* <i>Parentucellia latifolia</i> (L.)Caruel	Red Bartsia			
* <i>Parentucellia viscosa</i> (L.)Caruel	Yellow Bartsia			
<i>Peplidium foecundum</i> W.R.Barker	Dwarf Peplidium			
<i>Stemodia florulenta</i> W.R.Barker	Bluerod			
<i>Verbascum virgatum</i> Stokes	Twiggy Mullein			
* <i>Veronica anagallis-aquatica</i> L.	Blue Water-speedwell			
* <i>Veronica hederifolia</i> L.	Ivy-leaf Speedwell			
<i>Veronica hillebrandii</i> F.Muell.	Rigid Speedwell			
* <i>Veronica peregrina</i> L. ssp. <i>xalapensis</i> (Kunth)Pennell	Wandering Speedwell			
* <i>Zaluzianskya divaricata</i> (Thunb.)Walp.	Spreading Night-phlox			
<b>MARTYNIACEAE</b>				
* <i>Proboscidea louisianica</i> (Mill.)Thell.	Purple-flower Devil's Claw			
<b>OROBANCHACEAE</b>				
<i>Orobanche cernua</i> Loefl. var. <i>australiana</i> (F.Muell. ex Tate)J.M.Black ex Beck	Australian Broomrape			R
<i>Orobanche minor</i> L.	Lesser Broomrape	Y		
<i>Orobanche ramosa</i> L.		Y		
<b>LENTIBULARIACEAE</b>				
<i>Utricularia tenella</i> R.Br.	Pink Bladderwort			
<b>MYOPORACEAE</b>				
<i>Eremophila alternifolia</i> R.Br.	Narrow-leaf Emubush			
<i>Eremophila behriana</i> (F.Muell.)F.Muell.	Rough Emubush			
<i>Eremophila bignoniiflora</i> (Benth.)F.Muell.	Bignonia Emubush			
<i>Eremophila crassifolia</i> (F.Muell.)F.Muell.	Thick-leaf Emubush			

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<i>Eremophila deserti</i> (A.Cunn. ex Benth.)Chinnock	Turkey-bush			
<i>Eremophila divaricata</i> (F.Muell.)F.Muell. ssp. <i>divaricata</i>	Spreading Emubush			
<i>Eremophila gibbifolia</i> (F.Muell.)F.Muell.	Coccid Emubush			R
<i>Eremophila glabra</i> (R.Br.)Ostenf. ssp. <i>Glabra</i>				
	Tar Bush			
<i>Eremophila glabra</i> (R.Br.)Ostenf. ssp. Murray (A.G.Spooner 14470) Chinnock	Small Tar Bush			
<i>Eremophila longifolia</i> (R.Br.)F.Muell.	Weeping Emubush			
<i>Eremophila maculata</i> (Ker Gawl.)F.Muell. ssp. <i>maculata</i>	Spotted Emubush			
<i>Eremophila oppositifolia</i> R.Br. ssp. <i>oppositifolia</i>	Opposite-leaved Emubush			
<i>Eremophila polyclada</i> (F.Muell.)F.Muell.	Twiggy Emubush			R
<i>Eremophila scoparia</i> (R.Br.)F.Muell.	Broom Emubush			
<i>Eremophila serrulata</i> (A.Cunn. ex A.DC.)Druce	Green Emubush			
<i>Eremophila</i> sp. Fallax (D.E.Symon 12311) Chinnock				
<i>Eremophila sturtii</i> R.Br.	Turpentine Bush			
<i>Eremophila subfloccosa</i> Benth. ssp. <i>Glandulosa</i> (R.Bates 32961) Chinnock				
<i>Eremophila subfloccosa</i> Benth. ssp. <i>Lanata</i> (R.Bates 33587) Chinnock				
<i>Myoporum brevipes</i> Benth.	Warty Boobialla			
<i>Myoporum insulare</i> R.Br.	Common Boobialla			
<i>Myoporum montanum</i> R.Br.	Native Myrtle			
<i>Myoporum parvifolium</i> R.Br.	Creeping Boobialla			R
<i>Myoporum platycarpum</i> R.Br. ssp. <i>perbellum</i> Chinnock	Mallee Sandalwood			
<i>Myoporum platycarpum</i> R.Br. ssp. <i>platycarpum</i>	False Sandalwood			
<i>Myoporum</i> sp. Petiolatum (R.Taylor 484) Chinnock				
<b>PLANTAGINACEAE</b>				
<i>*Plantago bellardii</i> All.	Hairy Plantain			
<i>*Plantago coronopus</i> L. ssp. <i>commutata</i> (Guss.)Pilg.	Bucks-horn Plantain			
<i>*Plantago coronopus</i> L. ssp. <i>coronopus</i>	Bucks-horn Plantain			
<i>Plantago cunninghamii</i> Decne.	Clay Plantain			
<i>Plantago drummondii</i> Decne.	Dark Plantain			
<i>Plantago gaudichaudii</i> Bameoud	Narrow-leaf Plantain			
<i>Plantago hispida</i> R.Br.	Hairy Plantain			
<i>*Plantago lanceolata</i> L. var. <i>lanceolata</i>	Ribwort			
<i>*Plantago major</i> L.	Greater Plantain			
<i>*Plantago scabra</i> Moench	Rough Plantain			
<i>Plantago</i> sp. B (R.Bates 44765) Toelken	Little Plantain			
<i>Plantago turrifera</i> B.G.Briggs, Carolin & Pulley	Crowned Plantain			
<i>Plantago varia</i> R.Br.	Variable Plantain			
<b>VALERIANACEAE</b>				
<i>*Valerianella discoidea</i> (L.)Loisel.	Lesser Corn-salad			



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<b>DIPSACACEAE</b>				
<i>*Scabiosa atropurpurea</i> L.	Pincushion			
<b>CAMPANULACEAE</b>				
<i>Isotoma petraea</i> F.Muell.	Rock Isotome			
<i>Lobelia alata</i> Labill.	Angled Lobelia			
<i>Lobelia gibbosa</i> Labill.	Tall Lobelia			
<i>Pratia concolor</i> (R.Br.)Druce	Poison Pratia			R
<i>Wahlenbergia communis</i> Carolin	Tufted Bluebell			
<i>Wahlenbergia fluminalis</i> (J.M.Black)E.Wimm. ex H.Eichler	River Bluebell			
<i>Wahlenbergia gracilentia</i> Lothian	Annual Bluebell			
<i>Wahlenbergia litticola</i> P.J.Sm.	Coast Bluebell			
<i>Wahlenbergia luteola</i> P.J.Sm.	Yellow-wash Bluebell			
<i>Wahlenbergia multicaulis</i> Benth.	Tadgell's Bluebell			
<i>Wahlenbergia stricta</i> (R.Br.)Sweet ssp. <i>stricta</i>	Tall Bluebell			
<i>Wahlenbergia tumidifruca</i> P.J.Sm.	Swollen-fruit Bluebell			
<b>GOODENIACEAE</b>				
<i>Dampiera dysantha</i> (Benth.)Rajput & Carolin	Shrubby Dampiera			
<i>Dampiera marifolia</i> Benth.	Velvet Dampiera			
<i>Dampiera rosmarinifolia</i> Schltld.	Rosemary Dampiera			
<i>Goodenia albiflora</i> Schltld.	White Goodenia			
<i>Goodenia blackiana</i> Carolin	Native Primrose			
<i>Goodenia fascicularis</i> F.Muell. & Tate	Silky Goodenia			
<i>Goodenia geniculata</i> R.Br.	Bent Goodenia			
<i>Goodenia glauca</i> F.Muell.	Pale Goodenia			
<i>Goodenia havilandii</i> Maiden & Betche	Hill Goodenia			
<i>Goodenia heteromera</i> F.Muell.	Spreading Goodenia			R
<i>Goodenia ovata</i> Sm.	Hop Goodenia			
<i>Goodenia pinnatifida</i> Schltld.	Cut-leaf Goodenia			
<i>Goodenia pusilliflora</i> F.Muell.	Small-flower Goodenia			
<i>Goodenia robusta</i> (Benth.)K.Krause	Woolly Goodenia			
<i>Goodenia varia</i> R.Br.	Sticky Goodenia			
<i>Goodenia willisiana</i> Carolin	Silver Goodenia			
<i>Scaevola aemula</i> R.Br.	Fairy Fanflower			
<i>Scaevola albida</i> (Sm.)Druce	Pale Fanflower			
<i>Scaevola calendulacea</i> (Andrews)Druce	Dune Fanflower			V
<i>Scaevola depauperata</i> R.Br.	Skeleton Fanflower			
<i>Scaevola humilis</i> R.Br.	Inland Fanflower			
<i>Scaevola spinescens</i> R.Br.	Spiny Fanflower			

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<i>Selliera radicans</i> Cav.	Shiny Swamp-mat			
<i>Velleia arguta</i> R.Br.	Toothed Velleia			
<i>Velleia connata</i> F.Muell.	Cup Velleia			
<i>Velleia paradoxa</i> R.Br.	Spur Velleia			
<b>STYLIDIACEAE</b>				
<i>Levenhookia dubia</i> Sond.	Hairy Stylewort			
<i>Stylidium calcaratum</i> R.Br.	Spurred Trigger-plant			
<i>Stylidium graminifolium</i> Sw. ex Willd.	Grass Trigger-plant			
<i>Stylidium inundatum</i> R.Br.	Hundreds and Thousands			
<b>COMPOSITAE</b>				
<i>Acanthocladium dockeri</i> F.Muell.	Spiny Everlasting		X	E
* <i>Achillea millefolium</i> L.	Yarrow			
* <i>Achillea tomentosa</i> L.	Woolly Yarrow			
* <i>Acroptilon repens</i> (L.)DC.	Creeping Knapweed	Y		
<i>Actinobole uliginosum</i> (A.Gray)H.Eichler	Flannel Cudweed			
<i>Allittia cardiocarpa</i> (F.Muell.ex Benth.)P.S.Short				R
* <i>Ambrosia psilostachya</i> DC.	Perennial Ragweed	Y		
<i>Angianthus preissianus</i> (Steetz)Benth.	Salt Angianthus			
<i>Angianthus tomentosus</i> J.C.Wendl.	Hairy Angianthus			
* <i>Arctotheca calendula</i> (L.)Levyns	Cape Weed			
* <i>Arctotis stoechadifolia</i> P.J.Bergius	White Arctotis			
<i>Argentipallium blandowskianum</i> (Steetz ex Sond.)Paul G.Wilson	Woolly Everlasting			
<i>Argentipallium obtusifolium</i> (F.Muell. & Sond. ex Sond.)Paul G.Wilson	Blunt Everlasting			
* <i>Argyranthemum frutescens</i> (L.)Webb ex Sch.Bip. ssp. <i>foeniculaceum</i> (Pit. & Proust)Humphries	Teneriffe Daisy			
* <i>Argyranthemum frutescens</i> (L.)Webb ex Sch.Bip. ssp. <i>frutescens</i>	Marguerite Daisy			
* <i>Aster subulatus</i> Michx.	Aster-weed			
<i>Asteridea athrixioides</i> (Sond. & F.Muell.)Kroner f. <i>athrixioides</i>	Wirewort			
<i>Asteridea athrixioides</i> (Sond. & F.Muell.)Kroner f. <i>horripes</i> (J.M.Black)D.A.Cooke	Wirewort			
* <i>Asteriscus spinosus</i> (L.)Sch.Bip.	Golden Pallensis			
* <i>Bidens pilosa</i> L.	Cobbler's Pegs			
<i>Blennospora drummondii</i> A.Gray	Dwarf Button-flower			
<i>Brachyscome basaltica</i> F.Muell. var. <i>gracilis</i> Benth.	Swamp Daisy			R
<i>Brachyscome ciliaris</i> (Labill.)Less. var. <i>brachyglossa</i> Gauba	Rayless Variable-daisy			
<i>Brachyscome ciliaris</i> (Labill.)Less. var. <i>ciliaris</i>	Variable Daisy			
<i>Brachyscome ciliaris</i> (Labill.)Less. var. <i>lanuginosa</i> (Steetz)Benth.	Woolly Variable Daisy			
<i>Brachyscome ciliaris</i> (Labill.)Less. var. <i>subintegrifolia</i> G.L.R.Davis	Variable Daisy			R
<i>Brachyscome debilis</i> Sond.	Weak Daisy			
<i>Brachyscome dentata</i> Gaudich.	Lobe-seed Daisy			

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<i>Brachyscome exilis</i> Sond.	Slender Daisy			
<i>Brachyscome goniocarpa</i> Sond. & F.Muell. ex Sond.	Dwarf Daisy			
<i>Brachyscome graminea</i> (Labill.)F.Muell.	Grass Daisy			R
<i>Brachyscome lineariloba</i> (DC.)Druce	Hard-head Daisy			
<i>Brachyscome melanocarpa</i> Sond. & F.Muell. ex Sond.	Black-seed Daisy			V
<i>Brachyscome parvula</i> Hook.f.	Coast Daisy			R
<i>Brachyscome perpussilla</i> (Steetz)J.M.Black	Tiny Daisy			
<i>Brachyscome trachycarpa</i> F.Muell.	Smooth Daisy			
<i>Xerochrysum bracteatum</i> (Vent.)Tzvelev	Golden Everlasting			
* <i>Calendula arvensis</i> L.	Field Marigold			
* <i>Calendula officinalis</i> L.	Garden Marigold			
<i>Calotis cuneifolia</i> R.Br.	Purple Burr-daisy			
<i>Calotis cymbacantha</i> F.Muell.	Showy Burr-daisy			
<i>Calotis erinacea</i> Steetz	Tangled Burr-daisy			
<i>Calotis hispidula</i> (F.Muell.)F.Muell.	Hairy Burr-daisy			
<i>Calotis lappulacea</i> Benth.	Yellow Burr-daisy			R
<i>Calotis latiuscula</i> F.Muell. & Tate	Leafy Burr-daisy			
<i>Calotis scabiosifolia</i> Sond & F.Meull ex Sond. var. <i>scabiosifolia</i>	Rough Burr-daisy			
<i>Calotis scapigera</i> Hook.	Tufted Burr-daisy			R
* <i>Carduus tenuiflorus</i> Curtis	Slender Thistle	Y		
* <i>Carthamus lanatus</i> L.	Saffron Thistle			
<i>Cassinia arcuata</i> R.Br.	Drooping Cassinia			
<i>Cassinia complanata</i> J.M.Black				
<i>Cassinia laevis</i> R.Br.	Curry Bush			
* <i>Centaurea calcitrapa</i> L.	Star Thistle			
* <i>Centaurea eriophora</i> L.				
* <i>Centaurea melitensis</i> L.	Malta Thistle			
* <i>Centaurea nigrescens</i> Willd. ssp. <i>nigrescens</i>				
* <i>Centaurea solstitialis</i> L.	St Barnaby's Thistle			
<i>Centipeda crateriformis</i> N.G.Walsh ssp. <i>compacta</i> N.G.Walsh				
<i>Centipeda crateriformis</i> N.G.Walsh ssp. <i>Crateriformis</i>				
<i>Centipeda cunninghamii</i> (DC.)A.Braun & Asch.	Common Sneezeweed			
<i>Centipeda minima</i> (L.)A.Braun & Asch. ssp. <i>minima</i>	Spreading Sneezeweed			
<i>Centipeda nidiformis</i> N.G.Walsh				
<i>Centipeda pleiocephala</i> N.G.Walsh				
<i>Centipeda thespidioides</i> F.Muell.	Desert Sneezeweed			
* <i>Chondrilla juncea</i> L.	Skeleton Weed	Y		
* <i>Chrysanthemoides monilifera</i> (L.)Norl.	Boneseed	Y		

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* <i>Chrysanthemum coronarium</i> L.				
<i>Chrysocephalum apiculatum</i> (Labill.)Steetz	Common Everlasting			
<i>Chrysocephalum baxteri</i> (A.Cunn. ex DC.)Anderb.	White Everlasting			
<i>Chrysocephalum semipapposum</i> (Labill.)Steetz	Clustered Everlasting			
<i>Chthonocephalus pseudevax</i> Steetz	Ground-heads			
* <i>Cichorium intybus</i> L.	Chicory			
* <i>Cirsium vulgare</i> (Savi)Ten.	Spear Thistle	Y		
* <i>Conyza albida</i> Willd. ex Spreng.	Tall Fleabane			
* <i>Conyza bonariensis</i> (L.)Cronquist	Flax-leaf Fleabane			
<i>Cotula australis</i> (Sieber ex Spreng.)Hook.f.	Common Cotula			
* <i>Cotula bipinnata</i> Thunb.	Ferny Cotula			
* <i>Cotula coronopifolia</i> L.	Water Buttons			
<i>Cotula vulgaris</i> Levyns var. <i>australasica</i> J.H.Willis	Slender Cotula			
<i>Craspedia haplorrhiza</i> J.Everett & Doust				
<i>Craspedia variabilis</i> J.Everett & Doust				
<i>Cratystylis conocephala</i> (F.Muell.)S.Moore	Bluebush Daisy			
<i>Cymbonotus preissianus</i> Steetz	Austral Bear's-ear			
* <i>Dittrichia graveolens</i> (L.)Greuter	Stinkweed			
<i>Eclipta platyglossa</i> F.Muell.	Yellow Twin-heads			
<i>Elachanthus pusillus</i> F.Muell.	Elachanth			
<i>Epaltes australis</i> Less.	Spreading Nut-heads			
<i>Epaltes cunninghamii</i> (Hook.)Benth.	Tall Nut-heads			
<i>Eriochlamys behrii</i> Sond. & F.Muell. ex Sond.	Woolly Mantle			
<i>Erodiophyllum elderi</i> F.Muell.	Koonamore Daisy			
<i>Euchiton collinus</i> Cass.	Creeping Cudweed			
<i>Euchiton involucratu</i> s (G.Forst.)Holub	Star Cudweed			
<i>Euchiton sphaericus</i> (Willd.)Holub	Annual Cudweed			
* <i>Euryops abrotanifolius</i> (L.)DC.	Euryops			
* <i>Gamochoeta americana</i> (Mill.)Wedd.				
* <i>Gazania linearis</i> (Thunb.)Druce	Gazania			
* <i>Gazania rigens</i> (L.)Gaertn.	Gazania			
<i>Gnaphalium indutum</i> Hook.f.	Tiny Cudweed			
* <i>Gnaphalium polycaulon</i> Pers.	Indian Cudweed			
<i>Gnephosis arachnoidea</i> Turcz.	Spidery Button-flower			
<i>Gnephosis tenuissima</i> Cass.	Dwarf Golden-tip			
<i>Haegiela tatei</i> (F.Muell.)P.S.Short & Paul G.Wilson	Small Nut-heads			R
* <i>Hedynois rhagadioloides</i> (L.)F.W.Schmidt	Cretan Weed			
* <i>Helianthus annuus</i> L.	Sunflower			

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<i>Helichrysum adenophorum</i> F.Muell. var. <i>adenophorum</i>	Branched Everlasting			
<i>Helichrysum leucopsideum</i> DC.	Satin Everlasting			
<i>Helichrysum rutidolepis</i> DC.	Pale Everlasting			E
<i>Helichrysum scorpioides</i> Labill.	Button Everlasting			
* <i>Helminthotheca echioides</i> (L.)Holub	Ox-tongue			
<i>Hyalosperma demissum</i> (A.Gray)Paul G.Wilson	Dwarf Sunray			
<i>Hyalosperma glutinosum</i> Steetz ssp. <i>glutinosum</i>	Golden Sunray			
<i>Hyalosperma semisterile</i> (F.Muell.)Paul G.Wilson	Orange Sunray			
<i>Hyalosperma stoveae</i> (D.A.Cooke)Paul G.Wilson	Dwarf Sunray			R
* <i>Hypochoeris glabra</i> L.	Smooth Cat's Ear			
* <i>Hypochoeris radicata</i> L.	Rough Cat's Ear			
<i>Isoetopsis graminifolia</i> Turcz.	Grass Cushion			
<i>Ixodia achillaeoides</i> R.Br. ssp. <i>alata</i> (Schltdl.)Copley	Hills Daisy			
<i>Kippistia suaedifolia</i> F.Muell.	Fleshy Kippistia			
* <i>Lactuca saligna</i> L.	Willow-leaf Lettuce			
* <i>Lactuca serriola</i> L.	Prickly Lettuce			
<i>Lagenophora huegelii</i> Benth.	Coarse Bottle-daisy			
<i>Lemooria burkittii</i> (Benth.)P.S.Short	Wires-and-wool			
<i>Leiocarpa pluriseta</i> (Haegi)Paul G.Wilson				R
<i>Leiocarpa tomentosa</i> (Sond. & F.Muell. ex Sond.)Paul G.Wilson	Woolly Plover-daisy			
<i>Leiocarpa websteri</i> (S.Moore)Paul G.Wilson				
<i>Leinema microphyllum</i> (F.Muell) Paul G. Wilson				
<i>Leptorhynchos baileyi</i> F.Muell.	Bailey's Buttons			
<i>Leptorhynchos elongatus</i> DC.	Lanky Buttons			R
<i>Leptorhynchos squamatus</i> (Labill.)Less. ssp. <i>squamatus</i>	Scaly Buttons			
<i>Leptorhynchos tetrachaetus</i> (Schltdl.)J.M.Black	Little Buttons			
<i>Leptorhynchos waitzia</i> Sond.	Button Immortelle			
<i>Microseris lanceolata</i> (Walp.)Sch.Bip.	Yam Daisy			
<i>Millotia macrocarpa</i> Schodde	Large-fruit Millotia			
<i>Millotia muelleri</i> (Sond.)P.S.Short	Common Bow-flower			
<i>Millotia myosotidifolia</i> (Benth.)Steetz	Broad-leaf Millotia			
<i>Millotia perpusilla</i> (Turcz.)P.S.Short	Tiny Bow-flower			
<i>Millotia tenuifolia</i> Cass. var. <i>nudescens</i> Schodde	Soft Millotia			
<i>Millotia tenuifolia</i> Cass. var. <i>tenuifolia</i>	Soft Millotia			
<i>Minuria cunninghamii</i> (DC.)Benth.	Bush Minuria			
<i>Minuria denticulata</i> (DC.)Benth.	Woolly Minuria			
<i>Minuria integerrima</i> (DC.)Benth.	Smooth Minuria			
<i>Minuria leptophylla</i> DC.	Minnie Daisy			

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<i>Myriocephalus pluriflorus</i> (J.M.Black)D.A.Cooke	Inland Woolly-heads			
<i>Myriocephalus rhizocephalus</i> (DC.)Benth. var <i>rhizocephalus</i>	Woolly-heads			
<i>Olearia brachyphylla</i> (F.Muell. ex Sond.)N.A.Wakef.	Short-leaf Daisy-bush			
<i>Olearia calcarea</i> F.Muell. ex Benth.	Crinkle-leaf Daisy-bush			
<i>Olearia ciliata</i> (Benth.)F.Muell. ex Benth. var <i>.ciliata</i>	Fringed Daisy-bush			
<i>Olearia decurrens</i> (DC.)Benth.	Winged Daisy-bush			
<i>Olearia floribunda</i> (Hook.f.)Benth. var. <i>floribunda</i>	Heath Daisy-bush			
<i>Olearia lanuginosa</i> (J.H.Willis)N.A.Wakef.	Woolly Daisy-bush			
<i>Olearia lepidophylla</i> (Pers.)Benth.	Clubmoss Daisy-bush			
<i>Olearia magniflora</i> (F.Muell.)F.Muell. ex Benth.	Splendid Daisy-bush			
<i>Olearia minor</i> (Benth.)Lander	Heath Daisy-bush			
<i>Olearia muelleri</i> (Sond.)Benth.	Mueller's Daisy-bush			
<i>Olearia pannosa</i> Hook. ssp. <i>pannosa</i>	Silver Daisy-bush		V	V
<i>Olearia passerinoides</i> (Turcz.)Benth. ssp. <i>glutescens</i> (Sond.)D.A.Cooke	Sticky Daisy-bush			R
<i>Olearia passerinoides</i> (Turcz.)Benth. ssp. <i>passerinoides</i>	Feather Daisy-bush			
<i>Olearia picridifolia</i> (F.Muell.)Benth.	Rasp Daisy-bush			R
<i>Olearia pimeleoides</i> (DC.)Benth. ssp. <i>pimeleoides</i>	Pimelea Daisy-bush			
<i>Olearia rudis</i> (Benth.)F.Muell. ex Benth.	Azure Daisy-bush			
<i>Olearia subspicata</i> (Hook.)Benth.	Spiked Daisy-bush			
<i>Olearia teretifolia</i> (Sond.)F.Muell. ex Benth.	Cypress Daisy-bush			
<i>Olearia tubuliflora</i> (Sond. & F.Muell. ex Sond.)Benth.	Rayless Daisy-bush			
* <i>Onopordum acanthium</i> L.	Scotch Thistle			
* <i>Onopordum acaulon</i> L.	Horse Thistle			
* <i>Onopordum illyricum</i> L. ssp. <i>illyricum</i>	Illyrian Thistle			
* <i>Osteospermum clandestinum</i> (Less.)Norl.	Tripteris			
<i>Ozothamnus decurrens</i> F.Muell.	Ridged Bush-everlasting			
<i>Ozothamnus pholidotus</i> F.Muell.	Scaly Haeckeria			V
<i>Ozothamnus retusus</i> Sond. & F.Muell	Notched Bush-everlasting			
* <i>Picnomon acarna</i> (L.)Cass.	Soldier Thistle	Y		
<i>Picris angustifolia</i> DC ssp. <i>angustifolia</i>	Coast Picris			
<i>Picris squarrosa</i> Steetz	Squat Picris			R
<i>Pleuropappus phyllocalymneus</i> F.Muell.	Silver Candles			V
<i>Podolepis canescens</i> A.Cunn. ex DC.	Grey Copper-wire Daisy			
<i>Podolepis capillaris</i> (Steetz)Diels	Wiry Podolepis			
<i>Podolepis jaceoides</i> (Sims)W.Voss	Showy Copper-wire Daisy			R
<i>Podolepis rugata</i> Labill. var. <i>rugata</i>	Pleated Copper-wire Daisy			
<i>Podolepis tepperi</i> (F.Muell.)D.A.Cooke	Delicate Copper-wire Daisy			
<i>Podotheca angustifolia</i> (Labill.)Less.	Sticky Long-heads			

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<i>Pogonolepis muelleriana</i> (Sond.)P.S.Short	Stiff Cup-flower			
<i>Polycalymma stuartii</i> F.Muell.& Sond. ex Sond.	Poached-egg Daisy			
<i>Pseudognaphalium luteoalbum</i> (L.)Hilliard & B.L.Burt	Jersey Cudweed			
<i>Pterocaulon sphacelatum</i> (Labill.)Benth. & Hook.f. ex F.Muell.	Apple-bush			
<i>Pycnosorus globosus</i> F.L.Bauer ex Benth.				V
<i>Quinetia urvillei</i> Cass.	Quinetia			
* <i>Reichardia tingitana</i> (L.)Roth	False Sowthistle			
<i>Rhodanthe corymbiflora</i> (Schltdl.)Paul G.Wilson	Paper Everlasting			
<i>Rhodanthe floribunda</i> (DC.)Paul G.Wilson	White Everlasting			
<i>Rhodanthe laevis</i> (A.Gray)Paul G.Wilson	Smooth Daisy			
<i>Rhodanthe microglossa</i> (Maiden & Betche)Paul G.Wilson	Clustered Everlasting			
<i>Rhodanthe moschata</i> (A.Cunn. ex DC.)Benth.	Musk Daisy			
<i>Rhodanthe polygalifolia</i> (A.Cunn. ex DC.)Paul G.Wilson	Milkwort Everlasting			
<i>Rhodanthe pygmaea</i> (DC.)Paul G.Wilson	Pigmy Daisy			
<i>Rhodanthe stricta</i> (Lindl.)Paul G.Wilson	Slender Everlasting			
<i>Rhodanthe stuartiana</i> (Sond. & F.Muell. ex Sond.)Paul G.Wilson	Clay Everlasting			
<i>Rhodanthe tietkensis</i> (F.Muell.)Paul G.Wilson	Tietken's Daisy			
<i>Rhodanthe uniflora</i> (J.M.Black)Paul G.Wilson	Woolly Daisy			
<i>Rutidosis helichrysoides</i> DC. ssp. <i>helichrysoides</i>	Grey Wrinklewort			
<i>Senecio anethifolius</i> A.Cunn. ex DC. ssp. <i>anethifolius</i>	Feathery Groundsel			
* <i>Senecio angulatus</i> L.	Cape Ivy			
<i>Senecio behrianus</i> Sond. & F.Muell. ex Sond. Behr's	Grounsel			E
<i>Senecio cunninghamii</i> DC. var. <i>cunninghamii</i>	Shrubby Groundsel			
<i>Senecio dolichocephalus</i> I.Thomps.				
* <i>Senecio elegans</i> L.	Purple Grounsel			
<i>Senecio glomeratus</i> Desf. ex Poir. ssp. <i>longifructus</i> I.Thomps.	Swamp Groundsel			
<i>Senecio glomeratus</i> Desf. ex Poir. ssp. <i>glomeratus</i>	Swamp Groundsel			
<i>Senecio glossanthus</i> (Sond.)Belcher	Annual Groundsel			
<i>Senecio gregorii</i> F.Muell.	Fleshy Groundsel			
<i>Senecio helichrysoides</i> F.Muell.				E
<i>Senecio hypoleucus</i> F.Muell. ex Benth.	Pale Groundsel			
<i>Senecio longicollaris</i> I.Thomps.				
<i>Senecio macrocarpus</i> F.Muell. ex Belcher	Large-fruit Groundsel			V
<i>Senecio magnificus</i> F.Muell.	Showy Groundsel			
<i>Senecio megaglossus</i> F.Muell.	Superb Groundsel			E
<i>Senecio odoratus</i> Hornem.	Scented Gounsel			
<i>Senecio phelleus</i> I.Thomps.				
<i>Senecio picridioides</i> (Turcz.)M.E.Lawr.	Purple-leaf Groundsel			

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<i>Senecio pilosicristus</i> I.Thomps.				
<i>Senecio pinnatifolius</i> A.Rich.				
* <i>Senecio pterophorus</i> DC. var. <i>pterophorus</i>	African Daisy			
<i>Senecio quadridentatus</i> Labill.	Cotton Groundsel			
<i>Senecio runcinifolius</i> J.H.Willis	Thistle-leaf Groundsel			
<i>Senecio squarrosus</i> A.Rich.	Squarrose Groundsel			
<i>Siloxerus multiflorus</i> Nees	Small Wrinklewort			
<i>Solenogyne dominii</i> L.G.Adams	Smooth Solenogyne			
* <i>Solidago canadensis</i> L.	Golden Rod			
* <i>Soliva anthemifolia</i> (Juss.)R.Br. ex Less.	Jo Jo			
* <i>Soliva pterosperma</i> (Juss.)Less.	Jo-jo			
* <i>Sonchus asper</i> (L.)Hill ssp. <i>asper</i>	Rough Sow-thistle			
<i>Sonchus hydrophilus</i> Boulos	Native Sow-thistle			
* <i>Sonchus oleraceus</i> L.	Common Sow-thistle			
<i>Stuartina muelleri</i> Sond. Spoon	Cudweed			
* <i>Tagetes minuta</i> L.	Stinking Roger			
* <i>Taraxacum officinale</i> Weber	Red-seed Dandelion			
* <i>Tolpis barbata</i> (L.)Gaertn.	Yellow Hawkweed			
* <i>Tragopogon porrifolius</i> L.	Salsify			
<i>Trichanthodium skirrophorum</i> Sond. & F.Muell. ex Sond.	Woolly Yellow-heads			
<i>Triptilodiscus pygmaeus</i> Turcz.	Small Yellow-heads			
* <i>Urospermum picroides</i> (L.)F.W.Schmidt	False Hawkbit			
* <i>Vellereophyton dealbatum</i> (Thunb.)Hilliard & B.L.Burt	White Cudweed			
* <i>Verbesina encelioides</i> (Cav.)Benth. & Hook. ex A.Gray	Crownbeard			
<i>Vittadinia australasica</i> (Turcz.)N.T.Burb. var. <i>australasica</i>	Sticky New Holland Daisy			
<i>Vittadinia australasica</i> (Turcz.)N.T.Burb. var. <i>oricola</i> N.T.Burb.	New Holland Daisy			V
<i>Vittadinia blackii</i> N.T.Burb.	Narrow-leaf New Holland Daisy			
<i>Vittadinia cervicularis</i> N.T.Burb. var. <i>cervicularis</i>	Waisted New Holland Daisy			
<i>Vittadinia condyloides</i> N.T.Burb.	Club-hair New Holland Daisy			
<i>Vittadinia cuneata</i> DC. f. <i>cuneata</i>	Fuzzy New Holland Daisy			
<i>Vittadinia cuneata</i> DC. var. <i>morrisii</i> N.T.Burb.	New Holland Daisy			
<i>Vittadinia cuneata</i> DC. var. <i>murrayensis</i> N.T.Burb.	Murray New Holland Daisy			
<i>Vittadinia dissecta</i> (Benth.) N.T.Burb. var. <i>hirta</i> N.T.Burb.	Dissected New Holland Daisy			
<i>Vittadinia eremaea</i> N.T.Burb.	Desert New Holland Daisy			
<i>Vittadinia gracilis</i> (Hook.f.)N.T.Burb.	Woolly New Holland Daisy			
<i>Vittadinia megacephala</i> (F.Muell. ex Benth.)J.M.Black	Giant New Holland Daisy			
<i>Waitzia acuminata</i> Steetz var. <i>acuminata</i>	Orange Immortelle			
* <i>Xanthium californicum</i> Greene	Californian Burr			



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* <i>Xanthium occidentale</i> Bertol.	Noogoora Burr	Y		
* <i>Xanthium spinosum</i> L.	Bathurst Burr	Y		
<b>ALISMATACEAE</b>				
* <i>Alisma lanceolatum</i> With.	Water Plantain			
<i>Damasonium minus</i> (R.Br.)Buchenau	Star-fruit			
* <i>Sagittaria platyphylla</i> (Engelm.)J.G.Sm.	Arrowhead			
<b>HYDROCHARITACEAE</b>				
<i>Hydrilla verticillata</i> (L.f.)Royle	Waterthyme			R
<i>Ottelia ovalifolia</i> (R.Br.)L.Rich. ssp. <i>ovalifolia</i>	Swamp Lily			R
<i>Vallisneria americana</i> Michx. var. <i>americana</i>	River Eel-grass			
<b>JUNCAGINACEAE</b>				
<i>Triglochin hexagonum</i> J.M.Black	Six-point Arrowgrass			
<i>Triglochin mucronatum</i> R.Br.	Prickly Arrowgrass			
<i>Triglochin nanum</i> F.Muell.				
<i>Triglochin procerum</i> R.Br.	Water-ribbons			
<i>Triglochin</i> sp. B (J.Z.Weber 1330) Aston				
<i>Triglochin striatum</i> Ruiz & Pav.	Streaked Arrowgrass			
<b>POTAMOGETONACEAE</b>				
<i>Potamogeton crispus</i> L.	Curly Pondweed			
<i>Potamogeton ochreatus</i> Raoul	Blunt Pondweed			R
<i>Potamogeton pectinatus</i> L.	Fennel Pondweed			
<i>Potamogeton tepperi</i> A.Benn.	Tepper's Pondweed			
<i>Potamogeton tricarlinatus</i> F.Muell. & A.Benn. ex A.Benn.	Floating Pondweed			
<i>Ruppia maritima</i> L.	Widgeon Grass			
<i>Ruppia megacarpa</i> R.Mason	Widgeon Grass			
<i>Ruppia polycarpa</i> R.Mason	Widgeon Grass			
<b>ZANNICHELLIACEAE</b>				
<i>Lepilaena australis</i> J.Drumm. ex Harvey	Austral Water-mat			
<i>Lepilaena bilocularis</i> Kirk	Small-fruit Water-mat			
<i>Zannichellia palustris</i> L.				R
<b>NAJADACEAE</b>				
<i>Najas tenuifolia</i> R.Br.	Water Nymph			E
<b>LILIACEAE</b>				
* <i>Allium triquetrum</i> L.	Three-cornered Garlic	Y		
<i>Arthropodium minus</i> R.Br.	Small Vanilla-lily			
<i>Arthropodium strictum</i> R.Br.	Common Vanilla-lily			
* <i>Asparagus asparagoides</i> (L.)Druce	Bridal Creeper	Y		
* <i>Asparagus officinalis</i> L.	Asparagus			

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<i>*Asparagus plumosus</i> Baker				
<i>*Asphodelus fistulosus</i> L.	Onion Weed	Y		
<i>Bulbine bulbosa</i> (R.Br.)Haw.	Bulbine-lily			
<i>Bulbine semibarbata</i> (R.Br.)Haw.	Small Leek-lily			
<i>Burchardia umbellata</i> R.Br.	Milkmaids			
<i>Caesia calliantha</i> R.J.F.Hend.	Blue Grass-lily			
<i>Corynotheca licrota</i> R.J.F.Hend.	Sand Lily			R
<i>Dianella brevicaulis</i> (Ostenf.)G.W.Carr & P.F.	Horsfall Short-stem Flax-lily			
<i>Dianella longifolia</i> R.Br. var. <i>grandis</i> R.J.F.Hend.	Pale Flax-lily			R
<i>Dianella porracea</i> (R.J.F.Hend.)P.F.Horsfall & G.W.Carr	Pale Flax-lily			V
<i>Dianella revoluta</i> R.Br. var. <i>revoluta</i>	Black-anther Flax-lily			
<i>*Ipheion uniflorum</i> (Graham)Raf.	Spring Star-flower			
<i>*Lachenalia aloides</i> (L.f.)Hort.	Lachenalia			
<i>Laxmannia orientalis</i> Keighery	Dwarf Wire-lily			
<i>Lomandra collina</i> (R.Br.)Ewart	Sand Mat-rush			
<i>Lomandra densiflora</i> J.M.Black	Soft Tussock Mat-rush			
<i>Lomandra effusa</i> (Lindl.)Ewart	Scented Mat-rush			
<i>Lomandra fibrata</i> J.M.Black	Mt Lofty Mat-rush			
<i>Lomandra juncea</i> (F.Muell.)Ewart	Desert Mat-rush			
<i>Lomandra leucocephala</i> (R.Br.)Ewart ssp. <i>robusta</i> A.T.Lee	Woolly Mat-rush			
<i>Lomandra micrantha</i> (Endl.)Ewart ssp. <i>micrantha</i> S	mall-flower Mat-rush			
<i>Lomandra multiflora</i> (R.Br.)Britten ssp. <i>dura</i> (F.Muell.)T.D.Macfarl.	Hard Mat-rush			
<i>Lomandra nana</i> (A.T.Lee)A.T.Lee	Small Mat-rush			
<i>Lomandra sororia</i> (F.Muell. ex Benth.)Ewart	Sword Mat-rush			
<i>*Nothoscordum borbonicum</i> Kunth				
<i>*Ornithogalum arabicum</i> L.	Star Of Africa			
<i>*Ornithogalum umbellatum</i> L.				
<i>Thysanotus baueri</i> R.Br.	Mallee Fringe-lily			
<i>Thysanotus juncifolius</i> (Salisb.)J.H.Willis & Court	Rush Fringe-lily			
<i>Thysanotus patersonii</i> R.Br.	Twining Fringe-lily			
<i>Tricoryne elatior</i> R.Br.	Yellow Rush-lily			
<i>Tricoryne tenella</i> R.Br.	Tufted Yellow Rush-lily			
<i>Wurmbea dioica</i> (R.Br.)F.Muell. ssp. <i>dioica</i>	Early Nancy			
<i>Wurmbea dioica</i> (R.Br.)F.Muell. ssp. short upper leaf (S.Williams CBP23) R.J.Bates				
<i>Xanthorrhoea caespitosa</i> Bedford	Sand-heath Yacca			
<i>Xanthorrhoea quadrangulata</i> F.Muell.	Rock Grass-tree			
<i>Xanthorrhoea semiplana</i> F.Muell. ssp. <i>semiplana</i>	Yacca			
<b>AGAVACEAE</b>				

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* <i>Agave americana</i> L.	Century Plant			
<b>AMARYLLIDACEAE</b>				
<i>Calostemma luteum</i>	Sims Yellow Garland-lily			
<i>Calostemma purpureum</i> R.Br.	Pink Garland-lily			
<i>Crinum flaccidum</i> Herb.	Murray Lily			
* <i>Sternbergia lutea</i> (L.)Ker Gawl. ex Spreng. ssp. <i>lutea</i>	Autumn Crocus			
<b>HYPOXIDACEAE</b>				
<i>Hypoxis glabella</i> R.Br. var. <i>glabella</i>	Tiny Star			
<i>Hypoxis vaginata</i> Schldtl. var. <i>vaginata</i>	Yellow Star			
<b>PONTEDERIACEAE</b>				
* <i>Eichhornia crassipes</i> (Mart.)Solms	Water Hyacinth	Y		
<b>IRIDACEAE</b>				
* <i>Crocsmia X crocosmiiflora</i> (Lemoine ex E.Morren)N.E.Br.	Montbretia			
* <i>Freesia</i> cultivar	Freesia			
* <i>Gladiolus undulatus</i> L.	Wild Gladiolus			
* <i>Iris albicans</i> Lange				
* <i>Iris foetidissima</i> L.				
* <i>Moraea flaccida</i> (Sweet)Stead.		Y		
* <i>Moraea miniata</i> Andrews		Y		
* <i>Moraea setifolia</i> (L.f.)Druce	Thread Iris			
<i>Patersonia occidentalis</i> R.Br.	Long Purple-flag			
* <i>Romulea minutiflora</i> Klatt	Small-flower Onion-grass			
* <i>Romulea rosea</i> (L.)Eckl. var. <i>australis</i> (Ewart)M.P.de Vos	Common Onion-grass			
* <i>Sparaxis tricolor</i> (Schneev.)Ker Gawl.	Tricolor Harlequin Flower			
* <i>Tritonia squalida</i> (Aiton)Ker Gawl.	Tritonia			
<b>JUNACEAE</b>				
* <i>Juncus acutus</i> L.	Sharp Rush			
<i>Juncus aridicola</i> L.A.S.Johnson	Inland Rush			
* <i>Juncus articulatus</i> L.	Jointed Rush			
<i>Juncus bufonius</i> L.	Toad Rush			
<i>Juncus caespiticius</i> E.Mey.	Grassy Rush			
* <i>Juncus capitatus</i> Weigel	Dwarf Rush			
<i>Juncus flavidus</i> L.A.S.Johnson	Yellow Rush			
<i>Juncus holoschoenus</i> R.Br.	Joint-leaf Rush			
<i>Juncus homalocaulis</i> F.Muell.ex Benth.	Wiry Rush			V
<i>Juncus kraussii</i> Hochst.	Sea Rush			
<i>Juncus pallidus</i> R.Br.	Pale Rush			
<i>Juncus pauciflorus</i> R.Br.	Loose-flower Rush			

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<i>Juncus planifolius</i> R.Br.	Broad-leaf Rush			
<i>Juncus prismatocarpus</i> R.Br.	Branching Rush			E
<i>Juncus sarophorus</i> L.A.S.Johnson				
<i>Juncus subsecundus</i> N.A.Wakef.	Finger Rush			
<i>Juncus usitatus</i> L.A.S.Johnson	Common Rush			
<i>Luzula meridionalis</i> H.Nordensk.	Common Wood-rush			
<i>Luzula ovata</i> Edgar	Clustered Wood-rush			R
<b>RESTIONACEAE</b>				
<i>Apodasmia brownii</i> (Hook.f.)B.G.Briggs & L.A.S.Johnson	Coarse Twine-rush			
<i>Hypolaena fastigiata</i> R.Br.	Tassel Rope-rush			
<i>Lepidobolus drapetocoleus</i> F.Muell.	Scale Shedder			
<i>Leptocarpus tenax</i> (Labill.)R.Br.	Slender Twine-rush			
<b>CENTROLEPIDACEAE</b>				
<i>Aphelia gracilis</i> Sond.	Slender Aphelia			
<i>Aphelia pumilio</i> F.Muell.ex Sond.	Dwarf Aphelia			
<i>Centrolepis aristata</i> (R.Br.)Roem. & Schult.	Pointed Centrolepis			
<i>Centrolepis cephaliformis</i> Reader ssp. <i>cephaloformis</i>	Cushion Centrolepis			R
<i>Centrolepis glabra</i> (F.Muell.ex Sond.)Hieron.	Smooth Centrolepis			R
<i>Centrolepis polygyna</i> (R.Br.)Hieron.	Wiry Centrolepis			
<i>Centrolepis strigosa</i> (R.Br.)Roem. & Schult. ssp. <i>strigosa</i>	Hairy Centrolepis			
<i>Trithuria submersa</i> Hook.f.	Trithuria			
<b>GRAMINEAE</b>				
* <i>Agrostis capillaris</i> L.	Brown Top-bent			
<i>Agrostis venusta</i> Trin.				
* <i>Aira cupaniana</i> Guss.	Small Hair-grass			
* <i>Aira elegantissima</i> Schur	Delicate Hair-grass			
* <i>Alopecurus geniculatus</i> L.	Marsh Fox-tail			
* <i>Alopecurus pratensis</i> L.	Meadow Fox-tail			
<i>Amphibromus archeri</i> (Hook.f.)P.Morris	Pointed Swamp Wallaby-grass			R
<i>Amphibromus nervosus</i> (Hook.f.)Baill.	Veined Swamp Wallaby-grass			
<i>Amphipogon caricinus</i> F.Muell. var. <i>caricinus</i>	Long Grey-beard Grass			
<i>Amphipogon strictus</i> R.Br. var. <i>setifer</i> Benth.	Spreading Grey-beard Grass			
* <i>Anthoxanthum odoratum</i> L.	Sweet Vernal Grass			
<i>Aristida behriana</i> F.Muell.	Brush Wire-grass			
<i>Aristida contorta</i> F.Muell.	Curly Wire-grass			
<i>Aristida holathera</i> Domin var. <i>holathera</i>	Tall Kerosene Grass			
<i>Aristida personata</i> Henrard	Purple Wire-grass			
<i>Austrodanthonia auriculata</i> (J.M.Black)H.P.Linder	Lobed Wallaby-grass			

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<i>Austrodanthonia caespitosa</i> (Gaudich.)H.P.Linder	Common Wallaby-grass			
<i>Austrodanthonia carphoides</i> (F.Muell. ex Benth.)Zotov	Short Wallaby-grass			
<i>Austrodanthonia duttoniana</i> (Cashmore)H.P.Linder	Brown-back Wallaby-grass			R
<i>Austrodanthonia eriantha</i> (Lindl.)H.P.Linder Hill	Wallaby-grass			
<i>Austrodanthonia fulva</i> (Vickery)H.P.Linder	Leafy Wallaby-grass			
<i>Austrodanthonia geniculata</i> (J.M.Black)H.P.Linder	Kneed Wallaby-grass			
<i>Austrodanthonia laevis</i> (Vickery)H.P.Linder	Smooth Wallaby-grass			R
<i>Austrodanthonia pilosa</i> (R.Br.)H.P.Linder	Velvet Wallaby-grass			
<i>Austrodanthonia racemosa</i> (R.Br.)H.P.Linder var. <i>racemosa</i>	Slender Wallaby-grass			
<i>Austrodanthonia setacea</i> (R.Br.)H.P.Linder	Small-flower Wallaby-grass			
<i>Austrodanthonia tenuior</i> (Steud.)Connor & Edgar S	hort-awn Wallaby-grass			R
<i>Austrostipa acrociliata</i> (Reader)S.W.L.Jacobs & J.Everett	Branched Spear-grass			
<i>Austrostipa blackii</i> (C.E.Hubb.)S.W.L.Jacobs & J.Everett	Crested Spear-grass			
<i>Austrostipa curticomis</i> (Vickery)S.W.L.Jacobs & J.Everett	Short-crest Spear-grass			
<i>Austrostipa densiflora</i> (Hughes)S.W.L.Jacobs & J.Everett	Foxtail Spear-grass			R
<i>Austrostipa drummondii</i> (Steud.)S.W.L.Jacobs & J.Everett	Cottony Spear-grass			
<i>Austrostipa elegantissima</i> (Labill.)S.W.L.Jacobs & J.Everett	Feather Spear-grass			
<i>Austrostipa eremophila</i> (Reader)S.W.L.Jacobs & J.Everett	Rusty Spear-grass			
<i>Austrostipa exilis</i> (Vickery)S.W.L.Jacobs & J.Everett	Heath Spear-grass			
<i>Austrostipa flavescens</i> (Labill.)S.W.L.Jacobs & J.Everett	Coast Spear-grass			
<i>Austrostipa gibbosa</i> (Vickery)S.W.L.Jacobs & J.Everett	Swollen Spear-grass			R
<i>Austrostipa hemipogon</i> (Benth.)S.W.L.Jacobs & J.Everett	Half-beard Spear-grass			
<i>Austrostipa macalpinei</i> (Reader)S.W.L.Jacobs & J.Everett	Annual Spear-grass			
<i>Austrostipa mollis</i> (R.Br.)S.W.L.Jacobs & J.Everett	Soft Spear-grass			
<i>Austrostipa multispiculis</i> (J.M.Black)S.W.L.Jacobs & J.Everett	Small-seed Spear-grass			R
<i>Austrostipa mundula</i> (J.M.Black)S.W.L.Jacobs & J.Everett	Neat Spear-grass			
<i>Austrostipa nitida</i> (Summerh. & C.E.Hubb.)S.W.L.Jacobs & J.Everett	Balcarra Spear-grass			
<i>Austrostipa nodosa</i> (S.T.Blake)S.W.L.Jacobs & J.Everett	Tall Spear-grass			
<i>Austrostipa nullanulla</i> (J.Everett & S.W.L.Jacobs)S.W.L.Jacobs & J.Everett	Club Spear-grass			V
<i>Austrostipa pilata</i> (S.W.L.Jacobs & J.Everett)S.W.L.Jacobs & J.Everett	Prickly Spear-grass			V
<i>Austrostipa platychaeta</i> (Hughes)S.W.L.Jacobs & J.Everett	Flat-awn Spear-grass			
<i>Austrostipa puberula</i> (Steud.)S.W.L.Jacobs & J.Everett	Fine-hairy Spear-grass			R
<i>Austrostipa pubinodis</i> (Trin. & Rupr.)S.W.L.Jacobs & J.Everett	Long-shaft grass			
<i>Austrostipa scabra</i> (Lindl.)S.W.L.Jacobs & J.Everett ssp. <i>falcata</i> (Hughes)S.W.L.Jacobs & J.Everett	Slender Spear-grass			
<i>Austrostipa scabra</i> (Lindl.)S.W.L.Jacobs & J.Everett ssp. <i>scabra</i>	Rough Spear-grass			
<i>Austrostipa semibarbata</i> (R.Br.)S.W.L.Jacobs & J.Everett	Fibrous Spear-grass			
<i>Austrostipa setacea</i> (R.Br.)S.W.L.Jacobs & J.Everett	Corkscrew Spear-grass			
<i>Austrostipa stipoides</i> (Hook.f.)S.W.L.Jacobs & J.Everett	Coast Spear-grass			

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<i>Austrostipa tenuifolia</i> (Steud.)S.W.L.Jacobs & J.Everett				R
<i>Austrostipa trichophylla</i> (Benth.)S.W.L.Jacobs & J.Everett				
<i>Austrostipa tuckeri</i> (F.Muell.)S.W.L.Jacobs & J.Everett	Tucker's Spear-grass			R
* <i>Avellinia michelii</i> (Savi)Parl.	Avellinia			
* <i>Avena barbata</i> Pott ex Link	Bearded Oat			
* <i>Avena fatua</i> L.	Wild Oat			
* <i>Avena sativa</i> L.	Cultivated Oat			
<i>Bothriochloa macra</i> (Steud.)S.T.Blake	Red-leg Grass			R
<i>Brachiaria notochthona</i> (Domin)Stapf				
<i>Brachiaria piligera</i> (F.Muell. ex Benth.) Hughes				
* <i>Briza maxima</i> L.	Large Quaking-grass			
* <i>Briza minor</i> L.	Lesser Quaking-grass			
* <i>Bromus alopecuroides</i> Poir.				
<i>Bromus arenarius</i> Labill.	Sand Brome			
* <i>Bromus catharticus</i> Vahl	Prairie Grass			
* <i>Bromus diandrus</i> Roth	Great Brome			
* <i>Bromus hordeaceus</i> L. ssp. <i>hordeaceus</i>	Soft Brome			
* <i>Bromus madritensis</i> L.	Compact Brome			
* <i>Bromus rubens</i> L.	Red Brome			
<i>Catapodium rigidum</i> (L.)C.E.Hubb.	Rigid Fescue			
* <i>Cenchrus ciliaris</i> L.	Buffel Grass			
* <i>Cenchrus incertus</i> M.A.Curtis	Spiny Burr-grass	Y		
* <i>Cenchrus longispinus</i> (Hack.)Fernald S	Spiny Burr-grass	Y		
* <i>Chloris gayana</i> Kunth	Rhodes Grass			
<i>Chloris truncata</i> R.Br.	Windmill Grass			
* <i>Chloris virgata</i> Sw.	Feather-top Rhodes Grass			
* <i>Cortaderia selloana</i> (Schult. & Schult.f.)Asch. & Graebn.	Common Pampas Grass			
<i>Cymbopogon ambiguus</i> A.Camus	Lemon-grass			
<i>Cymbopogon oblectus</i> S.T.Blake	Silky-head Lemon-grass			
* <i>Cynodon dactylon</i> (L.) Pers. var. <i>dactylon</i>	Couch			
<i>Cynodon dactylon</i> (L.) Pers. var. <i>pulchellus</i> Benth.	Couch			
* <i>Cynodon nlemfuensis</i> Vanderyst var. <i>nlemfuensis</i>				
* <i>Cynosurus echinatus</i> L.	Rough Dog's-tail Grass			
* <i>Dactylis glomerata</i> L.	Cocksfoot			
<i>Deyeuxia quadriseta</i> (Labill.)Benth.	Reed bent-grass			
<i>Dichelachne crinita</i> (L.f.)Hook.f.	Long-hair Plume-grass			
<i>Dichelachne inaequiglumis</i> (Hack. ex Cheeseman)Edgar & Connor	Long Plume-grass			
<i>Digitaria ammophila</i> (Benth.)Hughes	Spider Grass			

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<i>Digitaria brownii</i> (Roem. & Schult.)Hughes	Cotton Panic-grass			
* <i>Digitaria ciliaris</i> (Retz.)Koeler	Summer Grass			
<i>Digitaria coenicola</i> (F.Muell.)Hughes	Spider Grass			
* <i>Digitaria ischaemum</i> (Schreb.)Schreb. ex Muhl.	Smooth Summer-grass			
* <i>Digitaria sanguinalis</i> (L.)Scop.	Crab Grass			
<i>Distichlis distichophylla</i> (Labill.)Fassett	Emu-grass			
* <i>Echinochloa colona</i> (L.)Link	Awnless Barnyard Grass			
* <i>Echinochloa crus-galli</i> (L.)P.Beauv.	Common Barnyard Grass			
* <i>Echinochloa crus-pavonis</i> (Kunth)Schult.	Barnyard Grass			
* <i>Echinochloa esculenta</i> (A.Braun) H.Scholz				
<i>Echinochloa lacunaria</i> (F.Muell.)P.W.Michael & Vickery	Lake Millet			E
* <i>Echinochloa pyramidalis</i> (Lam.) Hitchc. & Chase				
* <i>Ehrharta calycina</i> Sm.	Perennial Veldt Grass			
* <i>Ehrharta erecta</i> Lam.	Panic Veldt Grass			
* <i>Ehrharta longiflora</i> Sm.	Annual Veldt Grass			
* <i>Ehrharta villosa</i> (L.f.)Schult.f. ex Schult. & Schult.f. var. <i>maxima</i> Stapf	Pyp Grass			
* <i>Eleusine indica</i> (L.)Gaertn.	Crowsfoot Grass			
<i>Elymus scaber</i> (R.Br.)A.Love var. <i>scaber</i>	Native Wheat-grass			
<i>Elytrophorus spicatus</i> (Willd.)A.	Camus Spike-grass			
<i>Enneapogon avenaceus</i> (Lindl.)C.E.Hubb.	Common Bottle-washers			
<i>Enneapogon caerulescens</i> (Gaudich.)N.T.Burb. var. <i>caerulescens</i>	Blue Bottle-washers			
<i>Enneapogon nigricans</i> (R.Br.)P.Beauv.	Black-head Grass			
<i>Enneapogon polyphyllus</i> (Domin)N.T.Burb.	Leafy Bottle-washers			
<i>Enteropogon acicularis</i> (Lindl.)Lazarides	Umbrella Grass			
<i>Enteropogon ramosus</i> B.K.Simon	Umbrella Grass			
<i>Eragrostis australasica</i> (Steud.)C.E.Hubb.	Cane-grass			
* <i>Eragrostis barrelieri</i> Daveau	Pitted Love-grass			
<i>Eragrostis brownii</i> (Kunth)Nees ex Steud.				
* <i>Eragrostis cilianensis</i> (All.)Vignolo ex Janch.	Stink Grass			
* <i>Eragrostis curvula</i> (Schrud.)Nees	African Love-grass	Y		
<i>Eragrostis dielsii</i> Pilg. var. <i>dielsii</i>	Mulka			
<i>Eragrostis elongata</i> (Willd.)J.Jacq.	Clustered Love-grass			
<i>Eragrostis exigua</i> Lazarides				
<i>Eragrostis falcata</i> (Gaudich.)Gaudich. ex Steud.	Sickle Love-grass			
<i>Eragrostis infecunda</i> J.M.Black	Barren Cane-grass			R
<i>Eragrostis lacunaria</i> F.Muell. ex Benth.	Purple Love-grass			R
<i>Eragrostis laniflora</i> Benth.	Hairy-flower Woollybutt			
<i>Eragrostis leptocarpa</i> Benth.	Drooping Love-grass			

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<i>Eragrostis minor</i> Host	Small Stink-grass			
<i>Eragrostis parviflora</i> (R.Br.)Trin.	Weeping Love-grass			
<i>Eragrostis setifolia</i> Nees	Bristly Love-grass			
<i>Eragrostis xerophila</i> Domin	Knotty-butt Neverfail			
<i>Eriochloa australiensis</i> Stapf ex Thell.	Australian Cupgrass			
<i>Eriochloa crebra</i> S.T.Blake	Tall Cupgrass			
<i>Eriochloa pseudoacrotricha</i> (Stapf ex Thell.)J.M.Black	Perennial Cupgrass			
<i>Eulalia aurea</i> (Bory)Kunth	Silky Brown-top			
* <i>Festuca arundinacea</i> Schreb.	Tall Meadow Fescue			
* <i>Festuca pratensis</i> Huds.	Meadow Fescue			
* <i>Festuca rubra</i> L.	Red Fescue			
* <i>Gastridium phleoides</i> (Nees & Meyen)C.E.Hubb.	Nit-grass			
<i>Glyceria australis</i> C.E.Hubb.	Australian Sweet-grass			
* <i>Hainardia cylindrica</i> (Willd.)Greuter	Common Barb-grass			
<i>Hemarthria uncinata</i> R.Br. var. <i>uncinata</i>	Mat Grass			
* <i>Holcus lanatus</i> L.	Yorkshire Fog			
* <i>Holcus setosus</i> Trin.	Annual Fog			
* <i>Hordeum distichon</i> L.				
* <i>Hordeum glaucum</i> Steud.	Blue Barley-grass			
* <i>Hordeum leporinum</i> Link	Wall Barley-grass			
* <i>Hordeum marinum</i> Huds.	Sea Barley-grass			
* <i>Hordeum vulgare</i> L.	Barley			
<i>Imperata cylindrica</i> (L.)Raeusch.	Blady Grass			
<i>Isachne globosa</i> (Thunb.)Kuntze	Swamp Millet			
<i>Lachnagrostis aemula</i> (R.Br.)Trin.				
<i>Lachnagrostis billardieri</i> (R.Br.)Trin. ssp. <i>Billardieri</i>				
<i>Lachnagrostis filiformis</i> (G.Forst.)Trin.				
<i>Lachnagrostis robusta</i> (Vickery)S.W.L.Jacobs				R
* <i>Lagurus ovatus</i> L.	Hare's Tail Grass			
* <i>Lamarckia aurea</i> (L.)Moench	Toothbrush Grass			
<i>Leptochloa fusca</i> (L.)Kunth ssp. <i>Fusca</i>				
<i>Leptochloa fusca</i> (L.)Kunth ssp. <i>muelleri</i> (Benth.)N.Snow				
* <i>Lolium loliaceum</i> (Bory & Chaub.)Hand-Mazz.	Stiff Ryegrass			
* <i>Lolium perenne</i> L.	Perennial Ryegrass			
* <i>Lolium rigidum</i> Gaudin	Wimmera Ryegrass			
<i>Microlaena stipoides</i> (Labill.)R.Br. var. <i>stipoides</i>	Weeping Rice-grass			
* <i>Miscanthus sinensis</i> Anders.				
<i>Neurachne alopecuroidea</i> R.Br.	Fox-tail Mulga-grass			



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<i>Notodanthonia semiannularis</i> (Labill.)Zotov				
* <i>Panicum antidotale</i> Retz.	Blue Panic			
* <i>Panicum capillare</i> L. var. <i>brevifolium</i> Vasey ex Rydb. & Shear	Witch-grass			
* <i>Panicum coloratum</i> L.	Coolah Grass			
<i>Panicum decompositum</i> R.Br. var. <i>decompositum</i>	Native Millet			
<i>Panicum effusum</i> R.Br. var. <i>effusum</i>	Hairy Panic			
* <i>Panicum hillmanii</i> Chase	Witch-grass			
<i>Panicum laevinode</i> Lindl.	Broom Millet			
* <i>Panicum maximum</i> Jacq. var. <i>trichoglume</i> Eyles ex Robyns				
* <i>Panicum schinzii</i> Hack.	Sweet Panic			
* <i>Parapholis incurva</i> (L.)C.E.Hubb.	Curly Ryegrass			
* <i>Paspalum dilatatum</i> Poir.	Paspalum			
* <i>Paspalum distichum</i> L.	Water Couch			
* <i>Paspalum vaginatum</i> Sw.	Salt-water Couch			
* <i>Pennisetum clandestinum</i> Hochst. ex Chiov.	Kikuyu			
* <i>Pennisetum setaceum</i> (Forssk.)Chiov.	Fountain Grass			
* <i>Pennisetum villosum</i> R.Br. ex Fresen.	Feather-top			
<i>Pentapogon quadrifidus</i> (Labill.)Baill. var. <i>quadrifidus</i> Five-awn	Spear-grass			R
* <i>Pentaschistis airoides</i> (Nees)Stapf	False Hair-grass			
* <i>Pentaschistis pallida</i> (Thunb.)H.P.Linder	Pussy Tail			
* <i>Periballia minuta</i> (L.)Asch. & Graebn.				
* <i>Phalaris aquatica</i> L.	Phalaris			
* <i>Phalaris minor</i> Retz.	Lesser Canary-grass			
* <i>Phalaris paradoxa</i> L.	Paradox Canary-grass			
* <i>Phleum pratense</i> L.	Timothy Grass			
<i>Phragmites australis</i> (Cav.)Trin. ex Steud.	Common Reed			
* <i>Piptatherum miliaceum</i> (L.)Coss.	Rice Millet			
* <i>Poa annua</i> L.	Winter Grass			
* <i>Poa bulbosa</i> L.	Bulbous Meadow-grass			
<i>Poa clelandii</i> Vickery	Matted Tussock-grass			
<i>Poa crassicaudex</i> Vickery	Thick-stem Tussock-grass			
<i>Poa drummondiana</i> Nees	Knotted Poa			R
<i>Poa fax</i> J.H.Willis & Court	Scaly Poa			R
<i>Poa fordeana</i> F.Muell.	Forde's Poa			
<i>Poa labillardieri</i> Steud. var. <i>labillardieri</i>	Common Tussock-grass			
<i>Poa poiformis</i> (Labill.)Druce var. <i>poiformis</i>	Coast Tussock-grass			
* <i>Poa pratensis</i> L.	Kentucky Blue-grass			
* <i>Polypogon maritimus</i> Willd.	Coast Beard-grass			

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* <i>Polypogon monspeliensis</i> (L.)Desf.	Annual Beard-grass			
* <i>Polypogon viridis</i> (Gouan)Breistr.	Water Bent			
<i>Pseudoraphis spinescens</i> (R.Br.)Vickery	Spiny Mud-grass			
* <i>Psilurus incurvus</i> (Gouan)Schinz & Thell.	Bristle-tail Grass			
* <i>Puccinellia distans</i> (Jacq.)Parl.	Reflexed Poa			
* <i>Puccinellia fasciculata</i> (Torr.)E.P.Bicknell	Borrer's Saltmarsh-grass			
<i>Puccinellia stricta</i> (Hook.f.)C.H.Blom var. <i>stricta</i>	Australian Saltmarsh-grass			
* <i>Rostraria cristata</i> (L.)Tzvelev	Annual Cat's-tail			
* <i>Rostraria pumila</i> (Desf.)Tzvelev	Tiny Bristle-grass			
* <i>Schismus barbatus</i> (L.)Thell.	Arabian Grass			
* <i>Sclerochloa dura</i> (L.)P.Beauv.	Hard Meadow-grass			
* <i>Secale cereale</i> L.	Rye			
<i>Setaria basiclada</i> (Hughes)R.D.Webster				
<i>Setaria clementii</i> (Domin)R.D.Webster				
<i>Setaria constricta</i> (Domin)R.D.Webster	Knotty-butt Paspalidium			
* <i>Setaria italica</i> (L.)P.Beauv.	Fox-tail Millet			
<i>Setaria jubiflora</i> (Trin.)R.D.Webster	Warrego Summer-grass			
* <i>Setaria parviflora</i> (Poir.)Kerguelen				
* <i>Setaria pumila</i> (Poir.)Roem. & Schult. ssp. <i>pumila</i>	Pale Pigeon-grass			
* <i>Setaria verticillata</i> (L.)P.Beauv.	Whorled Pigeon-grass			
* <i>Setaria viridis</i> (L.)P.Beauv.	Green Pigeon-grass			
* <i>Sorghum bicolor</i> (L.)Moench	Grain Sorghum			
* <i>Sorghum halepense</i> (L.)Pers.	Johnson Grass			
* <i>Sporobolus africanus</i> (Poir.)Robyns & Tournay				
<i>Sporobolus mitchellii</i> (Trin.)C.E.Hubb. ex S.T.Blake	Rat-tail Couch			
<i>Sporobolus virginicus</i> (L.)Kunth	Salt Couch			
<i>Themeda triandra</i> Forssk.	Kangaroo Grass			
* <i>Thinopyrum elongatum</i> (Host)D.R.Dewey				
<i>Tragus australianus</i> S.T.Blake	Small Burr-grass			
<i>Triodia bunicola</i> (S.W.L.Jacobs)Lazarides	Flinders Ranges Spinifex			
<i>Triodia compacta</i> (N.T.Burb.)S.W.L.Jacobs	Spinifex			
<i>Triodia scariosa</i> N.T.Burb.	Spinifex			
<i>Tripogon loliformis</i> (F.Muell.)C.E.Hubb.	Five-minute Grass			
<i>Triraphis mollis</i> R.Br.	Purple Plume Grass			
* <i>Triticum aestivum</i> L.	Wheat			
* <i>Urochloa panicoides</i> P.Beauv. var. <i>panicoides</i>				
* <i>Vulpia bromoides</i> (L.)Gray	Squirrel-tail Fescue			
* <i>Vulpia ciliata</i> Dumort.	Fringed Fescue			

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* <i>Vulpia fasciculata</i> (Forssk.)Samp.	Sand Fescue			
* <i>Vulpia muralis</i> (Kunth)Nees	Wall Fescue			
* <i>Vulpia myuros</i> (L.)C.C.Gmelin f. <i>megalura</i> (Nutt.)Stace & R.Cotton	Fox-tail Fescue			
* <i>Vulpia myuros</i> (L.)C.C.Gmelin f. <i>myuros</i>	Rat's-tail Fescue			
<i>Whalleya prolata</i> (F.Muell.)K.E.Wills & J.J.Bruhl	Rigid Panic			
<b>PALMAE</b>				
* <i>Phoenix canariensis</i> Hort. ex Chabaud	Canary Island Palm			
<b>ARACEAE</b>				
* <i>Zantedeschia aethiopica</i> (L.)Spreng.	Arum Lily			
<b>LEMNACEAE</b>				
<i>Lemna disperma</i> Hegelm.	Common Duckweed			
<i>Lemna trisulca</i> L.	Ivy-leaf Duckweed			
<i>Spirodela punctata</i> (G.Mey.)C.H.Thomps.	Thin Duckweed			
<i>Wolffia australiana</i> (Benth.)Hartog & Plas	Tiny Duckweed			
<b>TYPHACEAE</b>				
<i>Typha domingensis</i> Pers.	Narrow-leaf Bulrush			
<i>Typha orientalis</i> C.Presl	Broad-leaf Bulrush			
<b>CYPERACEAE</b>				
<i>Baumea arthrophylla</i> (Nees)Boeck.	Swamp Twig-rush			
<i>Baumea articulata</i> (R.Br.)S.T.Blake	Jointed Twig-rush			
<i>Baumea juncea</i> (R.Br.)Palla	Bare Twig-rush			
<i>Bolboschoenus caldwellii</i> (V.J.Cook)Sojak	Salt Club-rush			
<i>Bolboschoenus fluviatilis</i> (Torr)Sojak				R
<i>Bolboschoenus medianus</i> (V.J.Cook)Sojak	Marsh Club-rush			
<i>Carex appressa</i> R.Br.	Tall Sedge			
<i>Carex bichenoviana</i> Boott ex Hook.f.	Notched Sedge			
<i>Carex breviculmis</i> R.Br.	Short-stem Sedge			
<i>Carex divisa</i> Huds.	Divided Sedge			
<i>Carex fascicularis</i> Sol. ex Boott	Tassel Sedge			
<i>Carex gaudichaudiana</i> Kunth	Fen Sedge			
<i>Carex inversa</i> R.Br. var. <i>major</i> Boott	Knob Sedge			R
<i>Carex tereticaulis</i> F.Muell. R	ush Sedge			
<i>Chorizandra enodis</i> Nees	Black Bristle-rush			
<i>Cladium procerum</i> S.T.Blake				R
* <i>Cyperus congestus</i> Vahl	Dense Flat-sedge			
<i>Cyperus difformis</i> L.	Variable Flat-sedge			
* <i>Cyperus eragrostis</i> Lam.	Drain Flat-sedge			
<i>Cyperus exaltatus</i> Retz.	Splendid Flat-sedge			

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<i>Cyperus flaccidus</i> R.Br.	Flaccid Flat-sedge			R
<i>Cyperus gunnii</i> Hook.f. ssp. <i>gunnii</i>	Flecked Flat-sedge			
<i>Cyperus gymnocaulos</i> Steud.	Spiny Flat-sedge			
<i>Cyperus hamulosus</i> M.Bieb.	Curry Flat-sedge			
<i>Cyperus involucratus</i> Rottb.				
<i>Cyperus laevigatus</i> L.	Bore-drain Sedge			
<i>Cyperus nervulosus</i> (Kuk.)S.T.Blake				R
<i>Cyperus pygmaeus</i> Rottb.	Pygmy Flat-sedge			
<i>Cyperus rigidellus</i> (Benth.)J.M.Black	Dwarf Flat-sedge			
<i>Cyperus rotundus</i> L. ssp. <i>rotundus</i>	Nut-grass	Y		
<i>Cyperus sanguinolentus</i> Vahl	Dark Flat-sedge			R
<i>Cyperus sphaeroideus</i> L.A.S.Johnson & O.D.Evans				R
<i>Cyperus squarrosus</i> L.	Bearded Flat-sedge			
<i>Cyperus tenellus</i> L.f.	Tiny Flat-sedge			
<i>Cyperus vaginatus</i> R.Br.	Stiff Flat-sedge			
<i>Cyperus victoriensis</i> C.B.Clarke	Yelka			
<i>Eleocharis acuta</i> R.Br.	Common Spike-rush			
<i>Eleocharis pallens</i> S.T.Blake	Pale Spike-rush			
<i>Eleocharis pusilla</i> R.Br.	Small Spike-rush			
<i>Eleocharis sphacelata</i> R.Br.	Tall Spike-rush			
<i>Fimbristylis aestivalis</i> (Retz.)Vahl	Summer Fringe-rush			R
<i>Fimbristylis dichotoma</i> (L.)Vahl	Common Fringe-rush			
<i>Fimbristylis velata</i> R.Br.	Veiled Fringe-rush			
<i>Gahnia deusta</i> (R.Br.)Benth.	Limestone Saw-sedge			
<i>Gahnia lanigera</i> (R.Br.)Benth.	Black Grass Saw-sedge			
<i>Gahnia trifida</i> Labill.	Cutting Grass			
<i>Isolepis australiensis</i> (Maiden & Betche)K.L.Wilson	Southern Club-rush			
<i>Isolepis cernua</i> (Vahl)Roem. & Schult.	Nodding Club-rush			
<i>Isolepis fluitans</i> (L.)R.Br.	Floating Club-rush			
<i>Isolepis hookeriana</i> Boeck.	Grassy Club-rush			
* <i>Isolepis hystrix</i> (Thunb.)Nees	Awned Club-rush			
<i>Isolepis inundata</i> R.Br.	Swamp Club-rush			
* <i>Isolepis marginata</i> (Thunb.)A.Dietr.	Little Club-rush			
<i>Isolepis nodosa</i> (Rottb.)R.Br.	Knobby Club-rush			
<i>Isolepis platycarpa</i> (S.T.Blake)Sojak	Flat-fruit Club-rush			
* <i>Isolepis trachysperma</i> Nees				
<i>Lepidosperma</i> aff. <i>congestum</i> R.Br.	Clustered Sword-sedge			
<i>Lepidosperma canescens</i> Boeck.	Hoary Rapier-sedge			

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<i>Lepidosperma carphoides</i> F.Muell.ex Benth.	Black Rapier-sedge			
<i>Lepidosperma concavum</i> R.Br.	Spreading Sword-sedge			
<i>Lepidosperma congestum</i> R.Br.	Clustered Sword-sedge			
<i>Lepidosperma curtisiae</i> K.L.Wilson & D.I.Morris	Little Sword-sedge			
<i>Lepidosperma longitudinale</i> Labill.	Pithy Sword-sedge			
<i>Lepidosperma semiteres</i> F.Muell. ex Boeck.	Wire Rapier-sedge			
<i>Lepidosperma viscidum</i> R.Br.	Sticky Sword-sedge			
<i>Schoenoplectus dissachanthus</i> (S.T.Blake)J.Raynal	Inland Club-rush			
<i>Schoenoplectus litoralis</i> (Schrad.)Palla	Shore Club-rush			
<i>Schoenoplectus pungens</i> (Vahl)Palla	Spiky Club-rush			
<i>Schoenoplectus validus</i> (Vahl)A.Love & D.Love	River Club-rush			
<i>Schoenus apogon</i> Roem. & Schult.	Common Bog-rush			
<i>Schoenus breviculmis</i> Benth.	Matted Bog-rush			
<i>Schoenus deformis</i> (R.Br.)Poir. ex Roem. & Schult.	Small Bog-rush			
<i>Schoenus latelaminatus</i> Kuk.	Medusa Bog-rush			V
<i>Schoenus nanus</i> (Nees ex Lehm.)Benth.	Little Bog-rush			
<i>Schoenus nitens</i> (R.Br.)Poir. ex Roem. & Schult.	Shiny Bog-rush			
<i>Schoenus racemosus</i> J.M.Black	Sandhill Bog-rush			
<i>Schoenus subaphyllus</i> Kuk.	Desert Bog-rush			
<b>ORCHIDACEAE</b>				
<i>Acianthus caudatus</i> R.Br.	Mayfly Orchid			
<i>Acianthus pusillus</i> D.L.Jones	Mosquito Orchid			
<i>Caladenia</i> aff. <i>Denticulate</i>				
<i>Caladenia argocalla</i> D.L.Jones	White Beauty Spider-orchid			E
<i>Caladenia bicalliata</i> R.S.Rogers ssp. <i>bicalliata</i>	Limestone Spider-orchid			R
<i>Caladenia capillata</i> D.L.Jones				
<i>Caladenia cardiochila</i> Tate	Heart-lip Spider-orchid			
<i>Caladenia carnea</i> R.Br.	Pink Fingers			
<i>Caladenia colorata</i> D.L.Jones	Coloured Spider-orchid		E	E
<i>Caladenia concolor</i> Fitzg.	Crimson Spider-orchid			
<i>Caladenia flaccida</i> D.L.Jones	Drooping Spider-orchid			V
<i>Caladenia fuscata</i> (Rchb.f.)M.A.Clem. & D.L.Jones				
<i>Caladenia latifolia</i> R.Br.	Pink Caladenia			
<i>Caladenia leptochila</i> Fitzg.	Narrow-lip Spider-orchid			
<i>Caladenia necrophylla</i> D.L.Jones	Late Spider-orchid			R
<i>Caladenia reticulata</i> Fitzg.	Veined Spider-orchid			
<i>Caladenia</i> sp. Monarto South (H.Goldsack 163 AD97708605A) R.J.Bates				
<i>Caladenia</i> sp. Monarto South (H.Goldsack 163) R.J.Bates				

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
<i>Caladenia</i> sp. White (R.Bates 41056) R.J.Bates				
<i>Caladenia stellata</i> D.L.Jones	Star Spider-orchid			R
<i>Caladenia stricta</i> (R.J.Bates)R.J.Bates	Upright Caladenia			
<i>Caladenia tensa</i> G.W.Carr Inland	Green-comb Spider-orchid			
<i>Caladenia tentaculata</i> Schldtl.	King Spider-orchid			
<i>Caladenia toxochila</i> Tate	Bow-lip Spider-orchid			
<i>Caladenia verrucosa</i> G.W.Carr	Yellow-club Spider-orchid			
<i>Caladenia</i> X <i>variabilis</i> Nicholls	Hybrid Spider-orchid			
<i>Calochilus campestris</i> R.Br.	Plains Beard-orchid			R
<i>Calochilus robertsonii</i> Benth.	Purplish beard-orchid			
<i>Corybas diemenicus</i> (Lindl.)Rchb.f.	Veined Helmet -orchid			
<i>Corybas incurvus</i> D.L.Jones & M.A.Clem.	Slaty Helmet-orchid			
<i>Pheladenia deformis</i> (R.Br.)D.L.Jones & M.A.Clem.	Bluebeard Orchid			
<i>Cyrtostylis reniformis</i> R.Br.	Small Gnat-orchid			
<i>Cyrtostylis robusta</i> D.L.Jones & M.A.Clem.	Robust Gnat-orchid			
<i>Dipodium roseum</i> D.L.Jones & M.A.Clem.	Pink Hyacinth-orchid			
* <i>Disa bracteata</i> Sw.	Monadenia			
<i>Diuris behrii</i> Schldtl.	Behr's Cowslip Orchid			V
<i>Diuris orientis</i> D.L.Jones W	allflower Donkey-orchid			
<i>Diuris palustris</i> Lindl.	Little Donkey-orchid			
<i>Diuris pardina</i> Lindl.	Spotted Donkey-orchid			
<i>Diuris</i> X <i>fastidiosa</i> R.S.Rogers	Proud Donkey-orchid			
<i>Diuris</i> X <i>palachila</i> R.S.Rogers	Broad-lip Donkey-orchid			
<i>Eriochilus cucullatus</i> (Labill.)Rchb.f.	Parson's Bands			
<i>Genoplesium nigricans</i> (R.Br.)D.L.Jones & M.A.Clem.	Black Midge-orchid			
<i>Genoplesium rufum</i> (R.Br.)D.L.Jones & M.A.Clem.	Red Midge-orchid			
<i>Glossodia major</i> R.Br.	Purple Cockatoo			
<i>Leporella fimbriata</i> (Lindl.)A.S.George	Fringed Hare-orchid			
<i>Leptoceras menziesii</i> (R.Br.)Lindl.	Hare Orchid			
<i>Microtis arenaria</i> Lindl.	Notched Onion-orchid			
<i>Microtis eremaea</i> R.J.Bates				
<i>Microtis frutetorum</i> Schldtl.				
<i>Microtis parviflora</i> R.Br.	Slender Onion-orchid			
<i>Microtis</i> sp. Short-leaf (R.J.Bates 54342) R.J.Bates				
<i>Orthoceras strictum</i> R.Br.	Horned O rchid			
<i>Prasophyllum constrictum</i> R.S.Rogers	Tawny Leek-orchid			R
<i>Prasophyllum elatum</i> R.Br.	Tall Leek-orchid			
<i>Prasophyllum occidentale</i> R.S.Rogers	Plains Leek-orchid			

Species	Common name	Proclaimed Pest Plant	EPBC Act Status	NPW Act Status
<i>Prasophyllum odoratum</i> R.S.Rogers	Scented Leek-orchid			
<i>Pterostylis boormanii</i> Rupp	Boorman's Greenhood			
<i>Pterostylis cynocephala</i> Fitzg.	Swan-head Greenhood			
<i>Pterostylis dolichochila</i> M.A.Clem. & D.L.Jones	Mallee Shell-orchid			
<i>Pterostylis erythroconcha</i> M.A.Clem. & D.L.Jones	Red Shell-orchid			
<i>Pterostylis excelsa</i> M.A.Clem.	Dryland Greenhood			
<i>Pterostylis mutica</i> R.Br.	Midget Greenhood			
<i>Pterostylis nana</i> R.Br.	Dwarf Greenhood			
<i>Pterostylis plumosa</i> Cady	Bearded Greenhood			
<i>Pterostylis pusilla</i> R.S.Rogers	Small Rusty-hood			
<i>Pterostylis robusta</i> R.S.Rogers	Large Shell-orchid			
<i>Pterostylis sanguinea</i> D.L.Jones & M.A.Clem.	Blood Greenhood			
<i>Pterostylis setifera</i> M.A.Clem., Matthias & D.L.Jones	Bristly Greenhood			E
<i>Pterostylis</i> sp. Veined leaf (R.J.Bates 59781) R.J.Bates				
<i>Pterostylis xerophila</i> M.A.Clem.	Desert Greenhood			V
<i>Pyrorchis nigricans</i> (R.Br.)D.L.Jones & M.A.Clem.	Black Fire-orchid			
<i>Thelymitra albiflora</i> Jeanes				
<i>Thelymitra antennifera</i> (Lindl.)Hook.f.	Lemon Sun-orchid			
<i>Thelymitra arenaria</i> Lindl.				
<i>Thelymitra azurea</i> R.S.Rogers				
<i>Thelymitra bracteata</i> J.Z.Weber ex Jeanes				
<i>Thelymitra brevifolia</i> Jeanes				
<i>Thelymitra epipactoides</i> F.Muell. Metallic	Sun-orchid		E	E
<i>Thelymitra holmesii</i> Nicholls	Blue-star Sun-orchid			V
<i>Thelymitra inflata</i> Jeanes				
<i>Thelymitra juncifolia</i> Lindl.	Spotted Sun-orchid			
<i>Thelymitra luteocilium</i> Fitzg.	Yellow-tuft Sun Orchid			
<i>Thelymitra megalyptra</i> Fitzg.				
<i>Thelymitra nuda</i> R.Br.	Scented Sun-orchid			
<i>Thelymitra pauciflora</i> R.Br.	Slender Sun-orchid			
<i>Thelymitra rubra</i> Fitzg.	Salmon Sun-orchid			
<i>Thelymitra</i> X <i>chasmogama</i> R.S.Rogers	Globe-hood Sun-orchid			
<i>Thelymitra</i> X <i>macmillanii</i> F.Muell.	Crimson Sun-orchid			

## CHAPTER 10. FRESHWATER INVERTEBRATES

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### INTRODUCTION

We usually pay little heed to invertebrate animals in everyday issues of natural history and conservation. There are exceptions, including large species like crayfish and troublesome ones like mosquitoes, but most invertebrates belong to a world that, if not unseen, is often not acknowledged. They are not a formal taxonomic group, but a cluster of convenience — an immensely diverse crowd of distantly-related animals united only by the absence of a “backbone”. Yet they are 99 percent of all animals, and a major part of the biological infra-structure that supports ecosystems.

Freshwater invertebrates abound in the Riverland. This chapter describes some of the major groups and selected species, and also contains short articles on some “macroinvertebrates” (those visible with the naked eye). Adventurous readers will soon discover a much larger world of cryptic, often bizarre species. Guides are available as books (e.g. Williams 1980; Hawking & Smith 1997; Gooderham & Tsyrlin 2002) and on the Internet:

<http://www.mdfrc.org.au/bugguide/index.htm> and [www.lucidcentral.com/keys/lwrrdc/public/aquatics/](http://www.lucidcentral.com/keys/lwrrdc/public/aquatics/)

The chapter concludes with a plea that invertebrates should be given more consideration in conservation issues. In South Australia they generally are not admitted to lists of “threatened” species, although they are regarded as part of ecological communities that could receive such a designation. As the chapter unfolds, you might consider whether they warrant more recognition.

### Environmental diversity

The “Lower Murray River” begins at Wentworth, New South Wales, where the Murray is joined by the Darling. Past the South Australian border, past Renmark, Berri, Loxton and Barmera, the river meanders westward across a broad floodplain with many anabranches, wetlands and woodlands. Near Overland Corner, it enters a limestone gorge before turning southward at Morgan. From Mannum, past Murray Bridge and Taillem Bend to Wellington, where once there were riparian swamplands, the river is separated from its floodplain by earthen levees. At Wellington, the Murray enters Lake Alexandrina, feeding Lake Albert and the Coorong, before meeting the ocean near Goolwa.

The course of the Lower Murray is marked by weirs, wetland regulators, riverbank levees and river-mouth barrages. There are 10 weirs below the Murray-Darling junction, six of them within South Australia (Walker 2006). They are so closely-spaced that the river really is a series of pools rather than a free-flowing stream. The weirs maintain close control over water levels, and while they have benefits for irrigators and local communities, they have disconnected the river and its floodplain, blocking the flow of water and sediment and causing salt to accumulate in the floodplain soil. Earthen levees along the lowermost reaches of the river, between Blanchetown and Wellington, are planted with alien willows (*Salix* spp.) to protect against erosion. The levees were constructed in the early 20th century to reclaim riparian swamplands, of which very little remains. The swamps were a haven for waterfowl, and duck shooters venturing into the water risked “swimmer’s itch” (cercarial dermatitis), a skin irritation caused by trematode parasites whose normal hosts are waterbirds and snails (e.g. Johnston 1941). The reclaimed land is used for dairying and other agriculture.

Along the seaward margins of Lake Alexandrina there are five barrages, constructed in 1939-40 to prevent the ingress of sea water to the lower lakes. They raised the level of the lakes by about 700 mm, and backed water up along the river to Blanchetown. In 2008-2009, as part of works



intended to control the threat of acid sulphate soils (see Chapter 3), the channel linking lakes Alexandrina and Albert was closed, weirs were constructed at Clayton and across the Finnis River, and another is planned for Currency Creek. Another weir has been proposed for the Murray at Pomanda Island, near Wellington, to secure water supplies against drought. The weirs, barrages and other structures mean that no part of the Murray in South Australia is free-flowing.

In the Riverland, the main aquatic habitats are the channel of the Murray and its associated wetlands. The channel includes sparse *benthic* (bottom-living) and *pelagic* (open-water) communities and comparatively rich *littoral* (river-edge) communities that vary with proximity to weirs. Downstream of each weir, where water levels change rapidly and the sediments are unstable, there are few plants. In the middle reaches of the weir pools, there are dense stands of reeds (*Phragmites australis*) and cumbungi (*Typha* spp.), and further downstream, where water levels are held steady by the next weir, dense stands of alien willows (*Salix* spp.) overhang the water (Schulze & Walker 1997). On the floodplain there are anabranches, flood runners and ephemeral, seasonal and permanent backwaters, billabongs and lakes (Sheldon & Walker 1998). Many of the temporary wetlands become saline after flooding (Suter et al. 1993), and some have been annexed as evaporation basins for irrigation runoff (e.g. Berri, Disher Creek, Loveday and Ramco Lagoon).

## **BIODIVERSITY**

### **River and floodplain species**

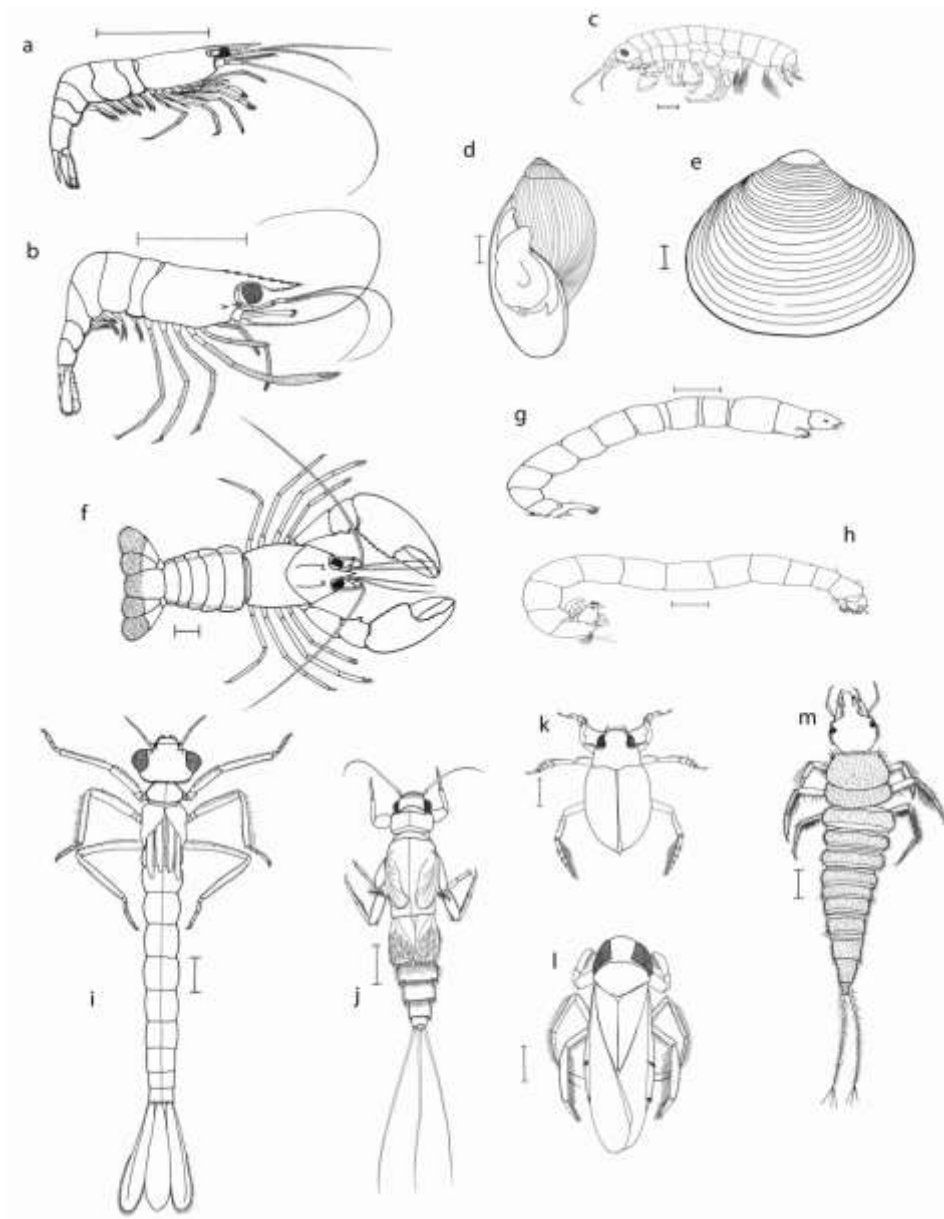
The deep sediments in the Murray channel are sparsely populated by invertebrates, mainly annelid worms, midge larvae (Diptera: Chironomidae) and protists and other microscopic forms. In the open water there are zooplankton, described later, but it is along the margins, in the littoral zone, where most biodiversity resides (Walker et al. 1992). Curiously, the dense stands of cumbungi, reeds and other plants along the river's edge, and many of the animals that live among them, are "refugees" from wetlands. They colonized the channel in the aftermath of weir construction, occupying areas that were the domain of riverine species like the Murray crayfish and river mussel, described later.

More than 200 taxa of macroinvertebrates are recorded from the Murray floodplain in South Australia (e.g. Boulton & Lloyd 1991; Goonan et al. 1992; Suter et al. 1993, 1995; Sheldon & Walker 1998). The most common include prawns (*Macrobrachium australiense*), shrimps (*Paratya australiensis*), amphipods (Ceinidae, Eusiridae), corixids (*Micronecta*), midge larvae (e.g. *Cricotopus*, *Parachironomus*, *Paratanytarsus*, *Procladius*) and oligochaetes (e.g. *Aulodrilus pigueti*, *Nais bretscheri*, *Dero flabelliger*). Some of these common species are illustrated in Figure 1.

Wetland habitats are influenced, of course, by floods from the Murray (Suter et al. 1993, 1995). Early flood assemblages include the corixid *Agraptocorixa eurynome*, the midges *Chironomus* spp. and the alien snail *Physa acuta*. Late flood assemblages—and communities in permanently connected wetlands—include the midge *Cladotanytarsus* and the shrimp *Paratya australiensis*. Isolated wetlands with salinities above 3000 mg L<sup>-1</sup> attract dipteran larvae, including midges (e.g. *Procladius*, *Tanytarsus*) and brine flies (Ephydriidae).

Flows from the highly turbid Darling River also affect invertebrate communities in the Lower Murray channel and wetlands (Bennison et al. 1989; Blanch et al. 1999). When the Darling's contribution is increased by regulation at Lake Victoria, upstream of Renmark, the abundances of many species are much reduced. This may reflect the loss of aquatic plants, and habitats for macroinvertebrates, due to low underwater light.

Tributaries entering the Murray from the Eastern Mt Lofty Ranges include the Marne River and Reedy Creek, which are fresh water, and Burra, Truro, Saunders, Preamimma, Salt, Mitchell, Dry and Rocky Gully creeks, which are episodic and saline. The freshwater streams harbour cumbungi, reeds and sedges, and the saline streams contain submerged charophytes. Most of these streams, fresh and saline, exist as pools that are joined by turbulent riffles in winter, and may flow underground in their lower reaches. They contain some 350 macroinvertebrate taxa, reflecting the diverse habitats provided by alternating pools and riffles. The pool-edge habitats include species like those found in the Murray, and the riffles (which do not occur in the Murray) support blackfly larvae (*Simulium ornatipes*), hydrobiid snails, oligochaetes and amphipods (*Austrochiltonia australis*).



**Figure 1.** Common macroinvertebrates of the Riverland: (a) *Paratya australiensis* (Atyidae); (b) *Macrobrachium australiense* (Palaemonidae); (c) *Austrochiltonia australis* (Ceinidae); (d) *Glyptophysa* sp. (Planorbidae); (e) *Corbiculina australis* (Corbiculidae); (f) *Cherax destructor* (Parastacidae); (g) *Procladius* sp. (Chironomidae); (h) *Chironomus* sp. (Chironomidae); (i) *Ischnura heterosticta* (Coenagrionidae); (j) *Tasmanocoenis tillyardi* (Caenidae); (k) *Necterosoma* sp. adult (Dytiscidae); (l) *Agraptocorixa* sp. (Corixidae); (m) *Necterosoma* sp. larva (Dytiscidae). Scale bars for a,b and f = 10 mm; others = 1 mm. Montage by T. Tan of drawings by J.H. Bradbury, from Wade et al. (2004), with permission from the Environment Protection Authority of South Australia.

### Porifera

Freshwater sponges (Spongillidae) are sessile, filter-feeding, colonial animals that form encrustations on submerged logs, bark, boulders and rocks, and in pipelines (Racek 1969; Williams 1980). The species are distinguished by the size, structure and arrangement of their gemmules (asexual resting propagules) and skeletal spicules.

The only sponge known from the Murray in South Australia is *Eunapius fragilis*, first recorded near Morgan in 1952 (Racek 1969). This species forms small, light tan-coloured clumps on logs and rocks. Like most spongillids, it survives floods without structural damage and is well-

adapted to turbid, sediment-laden water. Unidentified species have been collected from the Chowilla area, from Wongulla Lagoon at the mouth of the Marne River, and from some tributaries.

### **Nemertea**

Nemerteans (proboscis worms) in Australia are represented by a single, possibly introduced freshwater species, *Prostoma graecense* (Williams 1980; Hawking & Smith 1997). They occur throughout the Riverland. They are less than 25 mm long, un-segmented, with three pairs of lateral eyespots, and glide about using cilia. They have a muscular proboscis, used to capture prey, and may be brightly coloured.

### **Platyhelminthes**

Two platyhelminth classes, the Temnocephalidea and Turbellaria, are represented in the Riverland. The former are ectocommensal on crustaceans, including yabbies (*Cherax destructor*). They are oval, 1-12 mm long, with 2-6 finger-like anterior tentacles, and prey on small invertebrates (Williams 1980). Undescribed species of *Temnocephala* occur in the Murray, and wetlands and anabranches from Chowilla to Mannum, and in the Marne and Somme catchments.

Turbellarians (flatworms, or planarians) are free-living, elongate, flattened platyhelminths about 2-10 mm long, found gliding over stones, bark, snags and submerged plants. They are carnivorous, and feed on live or decaying animal matter (Williams 1980). They have been neglected by taxonomists as they are difficult to preserve and identify, often requiring histological examination. Undescribed species have been found along the margins of the Murray and permanent wetlands, and in perennial pools in the creeks of the Eastern Mt Lofty Ranges.

### **Annelida**

Annelids (segmented worms) include the common aquatic oligochaetes (relatives of earthworms), leeches, aphanoneurans and polychaetes (bristle worms).

The Riverland oligochaete fauna is rich in species of Naididae and Tubificidae, most with global or Australasian distributions, except the endemic *Slavina proceriseta* and *Antipodrilus magelensis*. Some Enchytraeidae and Phreodrilidae also may be natives. Most oligochaetes, including *Limnodrilus hoffmeisteri*, feed head-down in the sediments, extending the tail into the water for respiration. Among the Naididae, *Branchiodrilus hortensis* (Fig. 2a) has gills along the body, and species of *Dero* have gills in an anal chamber. Some species, especially members of the Naididae, swim or crawl over the bottom. They ingest mineral particles and decaying organic matter, and obtain their nutrition from the attached micro-organisms. *Chaetogaster diaphanus* is a predator on other small invertebrates, and a near-relative, *C. limnaei*, lives in the shells of certain snails, feeding on algae that grow on the shell. Some of the mobile species, including *Stylaria lacustris* and species of *Nais*, have simple eyes, making them unique among the oligochaetes.

Three families of freshwater leeches are known from South Australia, but only the Glossiphoniidae has been recorded from the Riverland. The adults are oval-shaped, often creamcoloured, and carry juveniles attached to their ventral surface (Hawking & Smith 1997). In *Helobdella papillornata*, described from the Loddon River in Victoria, the young feed on snails captured by the parent. They leave the parent after 2-4 weeks, and forage in small groups until they mature at 2-4 months (Govedich & Davies 1998).

Aphanoneurans resemble oligochaetes and live in the same habitats, but are less strongly segmented. They glide about using cilia, and have brightly-coloured oil glands in the body wall. They are tiny (<2 mm), although their chains of reproductive zooids may be as long as 10 mm. Australian species belong to the genus *Aeolosoma*.

Polychaetes (bristle worms) are mainly marine, but do occur in fresh water, including the Murray channel downstream of Walker Flat. In the lakes near the Murray mouth, the estuarine polychaete *Ficopotamus enigmaticus* has gained notoriety as it has spread rapidly, forming large masses of calcareous tubules on boats, moorings and other submerged surfaces, including the shells of turtles. Many turtles have died as a consequence.



**Figure 2.** (a). Oligochaete, *Branchiodrilus hortensis* (Naididae); (b). water mite, *Arrenurus* sp. (male) (Arrenuridae); (c). micro-caddisfly larva, *Helyethira* sp. (Hydroptilidae); (d). midge larva, *Zavreliella* sp. (Chironomidae); (e). aquatic caterpillar, Pyralidae: Nymphulinae; (f). caddisfly larva and case, *Oecetis* sp. (Leptoceridae). Photographs: (a) A.M. Pinder, Department of Environment and Conservation, Western Australia; (b-f) Chris Madden.

## Mollusca

Two classes of molluscs, the Bivalvia and Gastropoda, are well-represented in the Riverland (Williams 1980; Smith 1996). The bivalves include the orb-shell mussels (Corbiculidae) and the freshwater mussels (Hyriidae). All are sedentary animals with bi-valved shells that house plate-like demibranchs (“gills”) used for respiration, filter-feeding and reproduction. Despite their common name, “mussels”, the two families are very different (see *Freshwater mussels*, below).

The Corbiculidae is represented by *Corbiculina australis* (Fig. 1e). The shell of this species is 20-25 mm across, marked with concentric ridges, and is sometimes called a “little basket shell”. It occurs sparsely in the Murray and rarely in wetlands, but sometimes congregates in numbers enough to block irrigation pipes (for example, at Renmark: Woolford 1984). There are records from the Murray at Murtho, Loxton and Woods Point, from Ral Ral, Boat and Eckert creeks, from floodplain sites at Big Mussel, Irwin Flat, Big Toolunka Flat, Lake Carlet, Walker Flat and Mannum Swamp, and from the lower Marne River. Small specimens are easily confused with pea-mussels (Sphaeriidae), but there are no confirmed records of that family.

The second major group of molluscs, the gastropods, include the limpets and the familiar snails (Fig. 1d) (see *Snails*, below). The snails include *caenogastropods* (formerly “prosobranchs”)

with gills and an operculum to close the shell aperture, and *pulmonates* with a vascularised “lung” to aid respiration but without an operculum. All are grazers, equipped with a *radula* to scrape algae, bacteria and detritus from submerged surfaces. In general, the operculate species prefer flowing, well-oxygenated water, and the pulmonates inhabit wetlands where oxygen levels and other conditions vary.

### **Crustacea**

The crustaceans are a diverse group of arthropods, generally with two pairs of antennae and biramous (2-branched) limbs. They include Ostracoda, Copepoda and Cladocera (see *Zooplankton*), and Anostraca, Notostraca, Syncarida, Isopoda, Amphipoda and Decapoda (Williams 1980; Pearson & Hawking 2001). Some of the more striking forms are mentioned here.

The fairy shrimps (Anostraca) are among the most primitive of all living crustaceans, and are elongate, with leaf-like trunk appendages and no carapace (Williams 1980). Regional records include the brine shrimp *Parartemia zietziana* from saline pools at Bookmark Creek, near Renmark, and Waltowa Swamp near Lake Albert, and *Australbranchipus parooensis* from Katarapko Island, near Loxton (Rogers et al. 2007).

The tadpole shrimps (Notostraca) have a prominent, shield-shaped carapace and two long caudal rami (Williams 1980). They live in rainwater pools, fresh and saline lakes, dams and ditches. The only official record from the region is *Lepidurus apus viridis* from Lake Woolpolool, a saline, seasonal swamp on the Calperum floodplain.

The syncarids (Syncarida) are another primitive group, lacking a carapace and with similar-looking thoracic and abdominal appendages (Williams 1980). They occur in permanent lakes and streams and interstitial waters. The only record from the Riverland is of a few *Koonunga* from under dense willows on the Murray at Woods Point (Madden et al. 2003). They usually are part of the hypogean (underground water) fauna.

The isopods (Isopoda) have a distinct head, no carapace, seven pairs of walking legs, a six-segmented abdomen and gills on the abdominal appendages (Williams 1980). *Heterias pusilla* (Janiridae) occurs in low numbers in still or slow-flowing fresh waters throughout the region, including the Murray, anabranch creeks, permanent floodplain wetlands and creeks in the Eastern Mt Lofty Ranges. Another isopod is *Haloniscus searlei* (Oniscidae), which resembles a terrestrial slater and occurs in permanent saline inland waters. It occurs in Logan Creek in the Eastern Mt Lofty Ranges, and in Lake Woolpolool.

Isopods of the family Cirolanidae are represented by *Tachaea caridophaga* (formerly *Austroargathona picta*), an ectoparasite of decapods (e.g. *Paratya*, *Macrobrachium*: Figs 1a-b). It occurs sparsely in the river above Blanchetown (Lock 1).

The inelegantly-named scuds (Amphipoda) are related to the Isopoda, but have gills on the thoracic appendages and their abdominal appendages are used only for swimming (Williams 1980). Four families occur in the Riverland. The most common species is *Austrochiltonia australis* (Ceinidae) (Fig. 1c), in fresh and saline, still and flowing waters. Eusiridae are common in still and slow-flowing water in the Murray and in floodplain habitats, but are less common in the Eastern Mt Lofty Ranges. Corophiidae have been found at Reedy Creek, south of Mannum, and Perthiidae are known from the Murray only at Woods Point (Taylor 2002).

The crabs, crayfish, prawns and shrimps (Decapoda) have a prominent carapace that is fused to form a cephalothorax; they also have eyes on stalks and five pairs of thoracic legs. The crayfish and shrimp-like forms have an elongated cephalothorax and an extended abdomen, and the crab-like forms have a flattened cephalothorax and a reduced abdomen (Williams 1980).

The most common decapod is the shrimp *Paratya australiensis* (Atyidae) (Fig. 1a), found in still and slow-flowing fresh water. There are also two species of *Caridina*, one from Renmark and near Murray Bridge, and a second from Renmark, Berri, Bowhill and Mannum (Page et al. 2005).

Another common decapod from the Murray, and from anabranches and wetlands connected to the river, is the prawn *Macrobrachium australiense* (Fig. 1b), a relative of larger species that are farmed throughout south-eastern Asia. These are a favourite prey for many fish, and fishermen seeking bait know that, when the water is warm, the prawns are easily lured to a tin containing a bar of soap or other fatty material.

The best known decapods are the yabbie, *Cherax destructor* (Fig. 1f) and the Murray crayfish, *Euastacus armatus* (see *Crayfish*, below).

The small spider crab *Amarinus lacustris* (Hymenosomatidae) lives in fresh and slightly saline water (Walker 1969; Johnston & Robson 2005). The crabs live under rocks and among water plants in the river and connected freshwater wetlands, mainly downstream from Walker Flat. They are also known from Mitchell Gully Creek (9000 mg L<sup>-1</sup>) near Mypolonga. *Amarinus lacustris* is distinguished from its marine and estuarine relatives by having direct development, wherein the eggs are retained by the female until they hatch as miniature crabs rather than as planktonic larvae. This species has a curious geographic distribution, including parts of Victoria, Tasmania and New Zealand, King Island in Bass Strait, and Lord Howe and Norfolk Islands (Walker 1969).

### **Acarina**

Mites are represented by four sub-orders, the Astigmata, Oribatida and Prostigmata (Acariformes) and Mesostigmata (Parasitiformes). Oribatid mites (“beetle mites”) especially are common in still and flowing water. *Peza ops* (Pezidae) is also common in the Murray channel and in wetlands, but is apparently not in tributaries (cf. Harvey 1990).

The best known, most diverse group is the Hydracarina, or “water mites”. These have a variety of shapes and forms, and many are brightly coloured. One genus from each of seven families (Anisitsiellidae, Arrenuridae, Eylaidae, Hydrachnidae, Hydrodromidae, Oxidae, Pionidae) has been collected. Species of *Eylais* are conspicuous, with a bright, blood-red colour and soft bodies, whereas those of *Arrenurus* are bright blue or green, with hard, densely-punctured bodies. Female *Arrenurus* generally are globular, but the males can be distorted into bizarre forms (Fig. 2b).

Other mite records include four hygrobatid genera and two genera each from the Hydryphantidae and Unionicolidae. Five local species of unionicolid mites are parasites in freshwater mussels (Viets 1980; Harvey 1998), which in turn have larvae that are parasites on fish (see *Freshwater mussels*, below).

### **Insecta**

The insects are the most diverse group of freshwater invertebrates. Most aquatic forms are the immature stages of species with airborne adults (e.g. dragonflies, mayflies, midges), but there are aquatic adults, especially among the beetles (Coleoptera) and bugs (Hemiptera).

The Ephemeroptera (mayflies) are one of the most primitive groups. The Baetidae are represented by *Cloeon* in river and stream habitats, and the Caenidae by *Tasmanocoenis tillyardi* (Fig. 1j), mainly in wetlands and Eastern Mt Lofty Ranges tributaries, and *T. arcuata*, in the Murray. Another family, the Leptophlebiidae, is typical of cool, flowing water, and its regional representatives, *Atalophlebia* and *Koorrnonga inconspicua*, are common in tributaries.

Damselflies (Odonata) include *Austroagrion watsoni*, *Ischnura heterosticta* (Fig. 1i), *I. aurora*, *Pseudagrion aureofrons* and *Xanthagrion erythroneurum*, all members of the Coenagrionidae. Only *Ischnura* spp. and *X. erythroneurum* occur in tributaries. The Protoneuridae is represented by *Nososticta solida*, found among the root masses of riparian plants (Hawking & Theischinger 1999). The Lestidae includes the salt-tolerant *Austrolestes annulosus* in the river, and *A. analis* and *A. aridus* in tributaries.

Four families of dragonflies (Odonata) are common. Species of *Austrogomphus* (Gomphidae) occur in still waters along the Murray. *Hemicordulia tau* (Corduliidae) and *Hemianax papuensis* (Aeshnidae) and *Orthetrum caledonicum* (Libellulidae) are common in streams. Two other libellulid species, *Diplacodes haematodes* and *D. bipunctata*, also occur.

Stoneflies (Plecoptera) favour cool, flowing water and are rarely found in the Murray, but the gripopterygid *Dinotoperla* occurs in fast-flowing anabranches at Chowilla. Four species from two families are present in the eastern Mt Lofty tributaries. The Gripopterygidae is represented by *Illiesoperla mayi* and two species endemic to South Australia, *Dinotoperla evansi* and *Riekoperla naso*. The latter is unusual in that it occurs in localities prone to dry out in summer (Suter & Bishop 1990). The Notonemouridae is represented by *Austrocerca tasmanica*.

The most abundant insect group, especially in slow-moving or still water, is the bugs (Hemiptera). The Corixidae includes three species each of *Agraptocorixa* (Fig. 1l), *Micronecta* and *Sigara*. *Microvelia peramoena* and *M. oceanica* (Veliidae, Fig. 3c) are common on the water surface,

scavenging for other insects trapped in the surface film. Other surface-dwellers are the water striders (Gerridae); *Aquarius antigone*, for example, is often seen at the river margins.

The Notonectidae or backswimmers, represented by species of *Anisops*, are common in the river. *Enithares* also occurs, but is more common in the tributaries. This may be because *Anisops* (Fig. 3a) is able to attain neutral buoyancy and can rest in mid-water, whereas *Enithares* (Fig. 3b), having more air stored under the wings, readily rises to the surface and must grasp submerged objects to rest (Andersen & Weir 2004).

Other families of aquatic bugs are abundant in vegetated wetland areas rather than the channel. They include *Mesovelia hungerfordi* (Mesoveliidae), *Hydrometra strigosa* (Hydrometridae), *Ranatra dispar* (Nepidae), *Diplonychus eques* (Belostomatidae), *Naucoris congrex* (Naucoridae), *Hebrus axillaris* and *Merragata hackeri* (Hebridae) and *Paraplea* sp. (Pleidae). The semi-aquatic *Saldula* (Saldidae) is often found on the edge of saline wetlands.

Beetles (Coleoptera) are common, particularly members of the Dytiscidae and Hydrophilidae. The former include *Limbodessus* (Fig. 3d), *Antiporus*, *Rhantus*, *Necterosoma*, *Sternopriscus*, *Lancetes lanceolatus* and *Eretes australis*, although the latter is more typical of arid central Australia. The Hydrophilidae include *Berosus* (Fig. 3e), *Enochrus*, *Helochares*, *Paracymus*, *Hydrochus* (Fig. 3f) and *Limnoxenus zelandicus*. Two beetles typical of saline areas are *Necterosoma penicillatum* (Dytiscidae) (Fig. 1k,m) and *Laccobius zietzi* (Hydrophilidae). *Gymnochthebius* and *Hydraena* (Hydraenidae) are also common, especially in wetlands.

Whirligig beetles (Gyrinidae), represented by *Macrogyrus*, are remarkable for having their eyes divided into upper and lower hemispheres – as an adaptation to life on the water surface (e.g. Williams 1980). The species *Coxelmis novemnotata* and *C. v-fasciata* (Elmidae) occur on snags in the river and wetlands. Some beetle taxa occur only in the tributaries, notably two dytiscid species, *Chostonectes nebulosus* and *Platynectes decempunctatus*.

If the Hemiptera (bugs) is the most abundant group of insects in the Riverland, the Diptera (true flies) may be the most diverse. The Chironomidae (midge flies) includes the widespread, salttolerant *Procladius* (Fig. 1g) and freshwater species of *Cricotopus*, *Nanocladius* and *Parakiefferiella*. *Cladotanytarsus* and *Paratanytarsus* occur in the river, and in saline sites *Tanytarsus barbitarsis* often is the only insect present. *Chironomus tepperi* (cf. Fig. 1h) is one of the first insects to colonise newly-inundated wetlands, and *Dicrotendipes*, *Kiefferulus* and *Polypedilum* soon become abundant as food for birds and fish. The larvae of *Xenochironomus* are unusual, living in sponges, and *Zavreliella* (Fig. 2d) also is noteworthy for having a portable case (most midge larvae live in cases fixed to the substratum). The Eastern Mt Lofty Ranges streams have a similar midge fauna, but with added taxa like *Podonomopsis*, *Parametriocnemus* and *Rheotanytarsus*, typical of cool, flowing water.

The mosquitoes (Culicidae) (see *Mosquitoes*, below), are familiar pests in summer. The biting midges (Ceratopogonidae) are no less bothersome, and are represented by species of *Dasyhelea*, *Bezzia* and *Culicoides*. The Simuliidae (blackflies) require flowing water and are rare in wetlands, but *Simulium ornatipes*, *Austrosimulium furiosum* and *Paracnephia* occur in tributaries. Other dipterans are the larvae of Tipulidae, Tabanidae, Stratiomyidae, Empididae, Dolichopididae, Ephydriidae and Psychodidae. Swarms of brine flies (Ephydriidae) are typical of saline areas.

The larvae of Trichoptera (caddisflies) sometimes build elaborate cases that are either portable or fixed to the substratum. The Hydroptilidae (micro-caddisflies) include small (<5 mm) species of *Hellyethira* (Fig. 2c) and *Orthotrichia*, although the latter is rarely found outside the Murray. Leptoceridae is represented by *Oecetis* (Fig. 2f) and *Triplectides*. The predatory Ecnomidae also occur, with *Ecnomus pansus* in the river and *E. cygnitus* in tributaries. *Cheumatopsyche* (Hydropsychidae) builds a net to capture drifting organisms in flowing water; it is rare in the Murray but common in tributaries.

Two curiosities are aquatic caterpillars (Lepidoptera: Pyralidae) with external gills (Fig. 2e), found on wetland plants, and the lacewings (Neuroptera: Sisyridae), which live and feed on sponges.





**Figure 3.** (a). Backswimmer *Anisops* sp. (Notonectidae); (b). backswimmer *Enithares* sp. (Notonectidae); (c). small waterstrider *Microvelia* sp. (Veliidae); (d). diving beetle *Limbodessus* sp. (Dytiscidae); (e). water scavenger beetle *Berosus* sp. (Hydrophilidae); (f). water scavenger beetle *Hydrochus* sp. (Hydrochidae). Photographs: Chris Madden.

## FEATURES

### Invertebrate indicators

The *Australian River Assessment System* (AUSRIVAS), a national biological assessment of river health, is used widely in Australia. Macroinvertebrates are used as indicators because they are common, widely distributed, easily sampled and identified. River health is determined by comparing samples with those from “reference” sites in a comparatively undisturbed state (Simpson & Norris 2000). This provides a ratio of the number of families observed (O) at a site and the number expected (E). If the O/E ratio is near 1, the site earns a high rating (Band A); if the ratio is less than 1, taxa are missing. Sites missing 10-20% of expected families are considered “significantly impaired” (Band B), sites missing 20-40% families are “severely impaired” (Band C), and so on.

There is some dispute over using this system in highly regulated rivers like the Murray, partly for the lack of suitable reference sites. Further, the method works at the family level of taxonomy, not the species level, and some groups (e.g. sponges, crustaceans, molluscs) may not be sampled adequately. Work is underway to evaluate the utility of AUSRIVAS for the Murray, and to improve its value for ecosystem health assessments such as the Murray-Darling Basin Authority’s *Sustainable Rivers Audit* (<http://www.mdbc.gov.au/sra>).

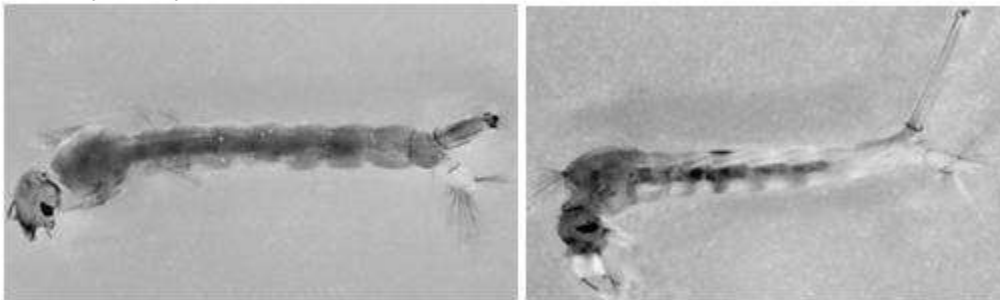


As part of the AUSRIVAS program, nearly 100 sites in the Riverland region have been sampled since 1994. Sites between the State border and Overland Corner generally are near reference condition, but most channel habitats are “significantly impaired”. Among the poorest sites are the Murray at Blanchetown and Morgan, and Disher and Salt Creeks, both affected by saline irrigation drainage. Streams in the Eastern Mt Lofty Ranges are mostly near reference condition, including sites on the Marne River, Reedy, Truro and Saunders Creeks. Sites that rate poorly include Baldina, Salt, Rocky Gully and Long Gully Creeks, which are affected by saline groundwater, and parts of Reedy and Baker Creeks and the Marne River, affected by water diversions and nutrients.

### Mosquitoes

Mosquito larvae (Diptera: Culicidae), or “wigglers”, are aquatic organisms that breathe atmospheric oxygen. This enables them to exploit habitats that are poorly-oxygenated and unattractive to predators (Lee et al. 1982, 1989). They generally use their scraping mouthparts to feed on organic detritus and biofilms, but some eat plant tissue and others are predators on smaller invertebrates. They are sometimes cannibalistic (Clements 1999). After hatching, the larvae progress through four instars and a pupal stage before emerging as adults. The adult females, of course, are usually blood-feeders (“haematophagous”) and a nuisance and disease-risk to humans and other animals (Russell 1993; Russell & Kay 2004).

At least 20 of South Australia’s 49 mosquito species occur in the Riverland, and eight are especially common (Fig. 4). The Riverland species have been studied by the Mosquito Research Laboratory at the University of South Australia since 1996. In 1999, a monitoring and control program was initiated in collaboration with local governments, and adult mosquitoes have been trapped at 30 sites along the Murray in September to May each year since.



**Figure 4.** Common larval mosquitoes. Left: instar IV larva of *Aedes camptorhynchus*, showing the short caudal siphon typical of *Aedes* species. Right: instar IV larva of *Culex annulirostris* showing the long, thin siphon and white antennae with dark apices. Photographs: S.R. Fricker.

Keys to larvae are provided by Russell (1993), and images of larvae and adults are available from the *New South Wales Arbovirus Surveillance and Mosquito Monitoring Program* (<http://www.arbovirus.health.nsw.gov.au>). Mosquito larvae are scarce in the Murray and its tributaries, owing to the presence of predatory fish and insects. Instead, they favour ephemeral and semi-permanent waters in the riparian zone, and natural and man-made habitats containing water from rain, tidal flow, stream flow, irrigation and other sources. Some species tolerate saline water. There are four main kinds of habitats:

- Groundpools (Fig. 5), or pools that collect in depressions and may contain plants;
- Phytotelmata, pools associated with living or dead plants, including pools in leaf axils and treeholes;
- Micropools, very small pools like water-filled hoof-prints created by livestock, and rock-holes that retain water from rain, stream flow or tidal action; and
- Anthropogenic pools, formed in artificial containers like rainwater tanks.



**Figure 5.** Common larval mosquito habitats. Left: a groundpool filled by rainfall and overbank river flow, Berri-Loxton Road: a habitat for *Aedes camptorhynchus*. Right: a water-filled roadside ditch with emergent grass, Renmark: a habitat for *Culex annulirostris*. Photographs: S.R. Fricker.

**Table 1.** Features of habitats for the larvae of mosquito species in the Riverland and Murrayland.

<b>Species</b>	<b>Habitat</b>	<b>Water Source</b>	<b>Salinity Trophic Status</b>	<b>Larval Habit</b>
<i>Aedes alboannulatus</i>	Riparian and non riparian groundpools, floodplains	Rainfall, river flow	Fresh, oligotrophic	Caudal siphon at surface
<i>Aedes camptorhynchus</i>	Non-riparian groundpools, floodplains, samphire swamps	Rainfall, river flow, irrigation, tidal inundation	Brackish, oligotrophic	Caudal siphon at surface
<i>Aedes notoscriptus</i>	Phytotelmata, artificial containers	Rainfall, anthropogenic flow	Fresh, oligotrophic	Caudal siphon at surface
<i>Anopheles annulipes</i>	Riparian groundpools	Rainfall, groundwater, river flow, irrigation	Fresh, oligotrophic	No caudal siphon, body horizontal at surface
<i>Coquillettidia linealis</i>	Riparian groundpools, wetlands	Groundwater, river flow	Fresh, oligotrophic	Siphon pierces submerged plants, body submerged
<i>Culex annulirostris</i> , <i>Culex australicus</i>	Riparian and nonriparian groundpools, floodplains, wetlands	Rainfall, river flow, irrigation	Fresh, oligotrophic	Caudal siphon at surface
<i>Culex quinquefasciatus</i>	Riparian and nonriparian groundpools, floodplains, wetlands, artificial containers	Rainfall, river flow, irrigation, anthropogenic flow	Fresh, oligo- eutrophic	Caudal siphon at surface

Table 1 shows the larval habitats of the eight common mosquito species. Based on these associations, the Lower Murray Valley can be sub-divided into five sub-regions:

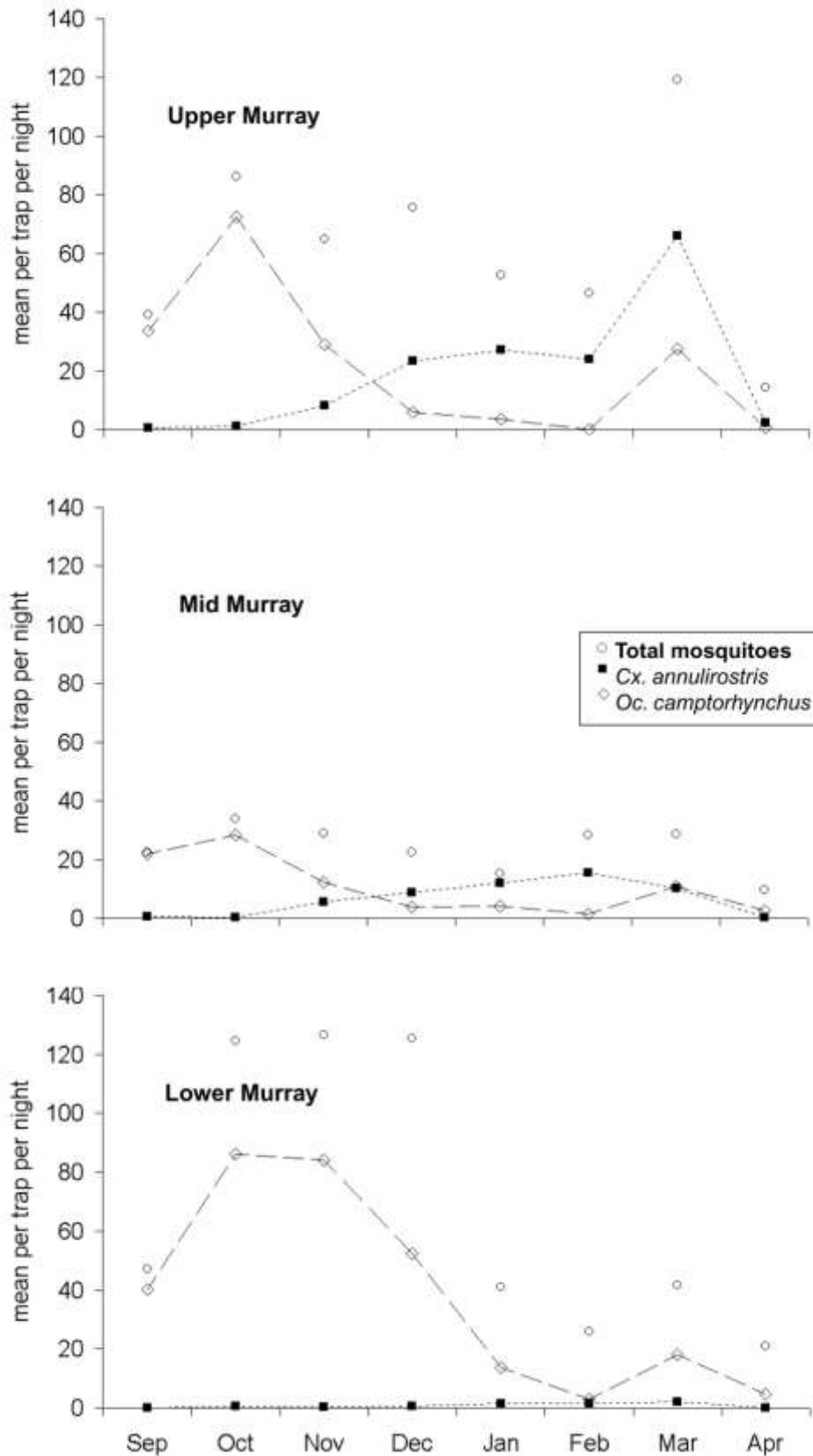
- *Upper Murray* (Renmark–Morgan). A meandering river and broad floodplain with many groundpools. Common species are *Anopheles annulipes* and *Coquillettidia linealis*, in fresh water, and *Cx. annulirostris* and *Aedes camptorhynchus*, in fresh and saline water.
- *Mid-Murray* (Morgan–Mannum). The channel is flanked by cliffs and a narrow floodplain. Dominated by *Ae. camptorhynchus* and, to a lesser extent, *Cx. annulirostris*.
- *Lower Murray* (Mannum–Meningie). Saline samphire swamps and irrigated dairy farms are common habitats, dominated by the salt-tolerant *Ae. camptorhynchus*.
- *Plains* (north of a line through Karoonda and Alawoona). Sandy calcareous mallee, woodlands and grassland areas with sparse numbers of *Cx. australicus* and *Ae. alboannulatus*.
- *Urban*. Cities, towns and settlements providing typical habitats for *Ae. notoscriptus* and *Cx. quinquefasciatus*.

The dominant species in these sub-regions change seasonally (Fig. 6). In cooler months, *Ae. camptorhynchus* is common in Lower Murray areas, and spreads upstream in autumn and spring. In summer, freshwater species like *Cx. annulirostris*, *Cq. linealis* and *An. annulipes* dominate in the Upper- and Mid-Murray. *Cx. annulirostris* develops in high numbers during hot, wet summers when there are overbank flows from the river.

In autumn, winter and spring, *Ae. camptorhynchus*, *Ae. alboannulatus* and other bushland species are prone to attack humans. In summer, *Ae. notoscriptus* and *Cx. annulirostris* (Upper-, Mid-Murray) or *Ae. camptorhynchus* (Lower Murray) attack outdoors, and *Cx. quinquefasciatus* attacks indoors.

Apart from their nuisance value, the main threat posed by mosquitoes to human health is the spread of zoonotic arboviruses. Historically, the most common of these in the Riverland region is Ross River Virus (RRV), which causes a debilitating fever, rash and polyarthritis. It originates in viraemic macropod reservoirs (e.g. kangaroos), and is carried to humans by infective mosquitoes. The main vectors are *Cx. annulirostris* in inland regions and *Ae. camptorhynchus* in coastal regions (Russell and Kay 2004). Local studies are needed, as the ecology of RRV varies and arboviruses have not yet been isolated from mosquitoes in South Australia.

The Riverland and Murrayland regions have overlapping seasonal and regional mosquito communities that could prove useful as indicators of environmental change. For example, salinization may favour salt-tolerant species (e.g. *Ae. camptorhynchus*), as demonstrated in Western Australia (Lindsay et al. 2007). A warmer future climate could promote *Cx. annulirostris*, which grows best in warm conditions, and discourage *Ae. camptorhynchus*, which is favoured by cool temperatures.



**Figure 6.** Mean monthly abundance (1999-2005) of adult female *Culex annulirostris* and *Aedes camptorhynchus* mosquitoes in CO<sub>2</sub>-baited traps at 30 sites along the Murray River. Upper Murray sites are from Renmark to Waikerie, Mid Murray sites are from Morgan to Mannum and Lower Murray sites are from Mypolonga to Meningie. Source: C.R. Williams (unpubl.).

## Crayfish

The Lower Murray is, or rather was, home to two species of freshwater crayfish, the ornately-spined Murray crayfish, *Euastacus armatus* (Fig. 7), and the familiar yabbie, *Cherax destructor*. The Murray crayfish, a riverine species, is now virtually extinct in South Australia, but the yabbie is very common in billabongs, back-waters, swamps, small streams and wetlands. Yabbies also occur in the river channel, in areas impounded by the weirs, but they are not a true big-river species.



**Figure 7.** The Murray crayfish, *Euastacus armatus*. Photograph: M.C. Geddes.

Both crayfish are members of the Parastacidae, found in Australia, New Guinea, New Zealand, South America and Madagascar, as a legacy of Gondwana (Scholtz 1999; Crandall 2002). The family attains its peak diversity in Australia, where there are over 100 species, including highly specialized burrowing forms and the largest crayfish in the world, the giant Tasmanian crayfish *Astacopsis gouldii*, which can attain 5 kg.

The Murray crayfish grows to 3 kg, and is second largest. The Murray crayfish is the only inland representative of more than 30 species of *Euastacus*, typical of mountain streams in eastern Australia. Like its relatives, it has little tolerance of high temperature and low oxygen. Its sensitivity to water quality is illustrated by a widespread “crawl-out” of crayfish recorded in 1992-93, in the middle Murray downstream of the Barmah-Millewa Forest, in response to low-oxygen water (<2 mg L<sup>-1</sup>) (McKinnon 1995).

Murray crayfish breed in autumn (April-May), around the time that the adults moult. The males deposit a sperm package near the base of the female’s second last pair of legs, and the eggs are fertilized as the female extrudes them. The eggs attach to the swimmerets of the female, and are carried for 4-5 months. They hatch in spring, and the juveniles are carried for a further 3-4 weeks. They are slow-growing, mature at about six years, weighing 300-500 g, and attain 1 kg after about 10 years. They may live for several decades.

Murray crayfish were once distributed throughout the Murray and Murrumbidgee rivers, and were an important food for aboriginal people. They have disappeared from 700 km of their former river habitat (Walker 1982; Versteegen & Lawler 1997), including the Lower Murray, although they are a protected species under South Australia’s *Inland Fisheries Act 1995*. Populations remaining in Victoria and New South Wales are subject to strict fishery regulations.

In the first half of the 20th century, there was a small commercial and recreational fishery for Murray crayfish in South Australia. A survey of 20 professional fishers some years ago indicated that the crayfish once occurred as far downstream as Murray Bridge (Geddes et al. 1993). They were caught from May to August (months without an “r”), and were inactive during warmer months. Catches were good in 1940-1960, with crayfish from 0.5-2.5 kg, but declined after 1965. Occasional specimens were caught until the 1980s, and there have been no records since.

The decline of Murray crayfish may be due to a combination of agricultural pesticides, overfishing and environmental changes related to flow regulation and dislocation by weirs. A study to assess the feasibility of re-introducing them to the Lower Murray (Geddes et al. 1993) showed that 500-g berried females held in ponds may hatch up to 500 juveniles that grow to about 5 g after one year, with 30-40% survival over 18 months. This suggests that hatchery-based restocking is feasible, although this would need to be planned carefully to avoid effects on genetic variability in areas where natural populations remain (Versteegen & Lawler 1997).

The yabbie, *Cherax destructor*, is the most widespread of all Australian freshwater crayfish. Although there are taxonomic doubts (e.g. Lawrence et al. 2002), it appears that *C. destructor* is a distinctive species, and the Riverland is part of its natural distribution. It is difficult to be certain about the natural range because yabbies have been translocated by humans, perhaps over thousands of years (Horwitz & Knott 1995). They are found throughout the Murray-Darling Basin, in the Lake Eyre Basin, in western Queensland, in south-western Western Australia and Tasmania, and from coastal areas of south-eastern and eastern Australia.

Like other freshwater crayfish, yabbies are detritivores or herbivores, but may act as predators in some situations. They breed from late spring until autumn. Eggs are attached to the female's swimmerets and develop in 4-8 weeks, depending on temperature. The juveniles grow quickly and mature at about one year, weighing as little as 10-15 g.

One of the remarkable features of yabbies is their ability to survive in dry ponds and billabongs. They dig a burrow down to the groundwater table and live in a puddle of water where they can keep their gills moist and respire. In this way, they can survive for several years between floods. Yabbies were once the basis of an important recreational and commercial fishery in South Australia. Catches of more than 100 tonnes per year were reported from Lake Alexandrina in the early 1970s, but are now only a few tonnes. Although there may have been multiple causes for the "crash" of the fishery, it coincided with the invasion by common carp, *Cyprinus carpio*, which prey on small yabbies.

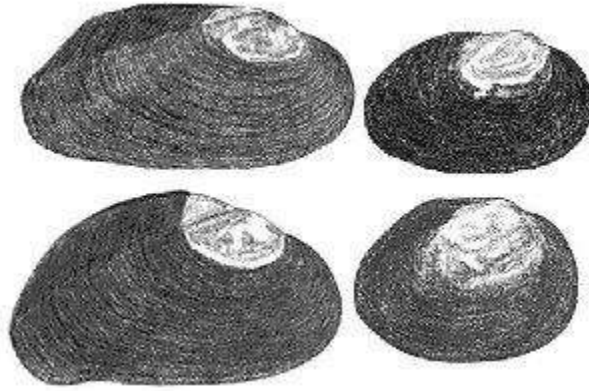
Yabbies have been one of the mainstays of the aquaculture industry in the last 30 years (Geddes & Jones 1997). Most yabbie aquaculture in South Australia has been in the south-east, and on the Fleurieu and Eyre Peninsulas, but there have been small ventures in the Riverland. Yabbies have also been introduced to South Africa, China, south-east Asia and South America. Perhaps the greatest threat to wild populations is the potential for disease and loss of genetic diversity associated with intensive aquaculture.

#### **Freshwater mussels**

There are few places along the Murray where you cannot find the shells of freshwater mussels. In some areas, middens of bleached shells attest to their importance as food for aboriginal people who once lived along the river. In others, there may be small piles of shells left by a predatory water rat, or by a fisherman.

Living mussels are common in wetlands and along the river's margins. They anchor themselves in the sediment, with hind-end protruding and siphons open to draw in the plankton and other organic particles that provide food. They maintain anchorage with a muscular "foot" that extends into the sediment. Within the shell, the foot and visceral mass are enclosed by plate-like demibranchs that function as gills, and also trap incoming particles and transport them, along mucousladen rows of cilia, to palps where edible particles are diverted to the mouth and the residue is expelled as "pseudofaeces".

Although the freshwater mussels superficially resemble those on jetty pilings and seashores, they are very distant cousins. All Australian species belong to the Hyriidae, a family shared with New Guinea, New Zealand and South America, and a legacy of the super-continent of Gondwana (Walker et al. 2001). The Murray-Darling Basin harbours several species (Walker 1981), but only two are likely to be encountered in the Riverland region (Fig. 8). These are the river mussel, *Alathyria jacksoni*, and the floodplain (or billabong) mussel, *Velesunio ambiguus*. The two look similar at first glance, and some experience is needed to tell them apart because the shapes of the shells vary with the strength of the current and the nature of the sediment. Appearances aside, they have quite different ecological and physiological credentials.

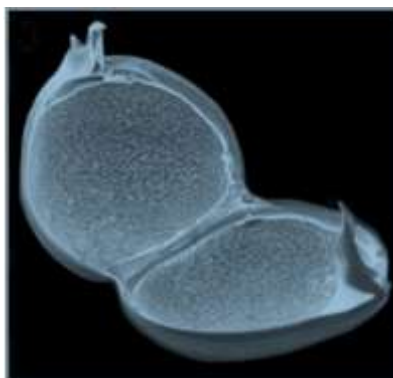


**Figure 8.** Four shells representing common forms of the river mussel *Alathyria jacksoni* and the floodplain mussel *Velesunio ambiguus*. At top left is the shell of a river mussel from a moderate current, and below it the arched shell typical of fast-flowing water. At top right is a typical shell of the floodplain mussel from the Murray, and below it is a variant from wetlands in the Riverland region. The *Alathyria* shell at lower left is 140 mm long; the others are scaled similarly. The shells are right valves, with the anterior end to the right. From Walker et al. (2001).

*Alathyria jacksoni* is the larger of the two, with a heavy, elongate shell up to 150 mm long, and may attain an age of 30 years or more. Shells from moderate currents have a straight dorsal margin, whereas those from the strongest currents have a dorsal arch that allows the foot to extend more deeply into the sediment (Fig. 8) (Balla & Walker 1991). This species is rarely found in billabongs and other still-water environments isolated from the river because it does not tolerate low oxygen, high temperatures and dehydration (Sheldon & Walker 1989). Removed from the water, river mussels die within a few days.

The shells of *V. ambiguus* attain 100 mm length and an age of 10-15 years. They may be fragile and rounded, or more elongate, with thickened, heavy valves – their protean character has caused much confusion for taxonomists, and the species well-deserves its epithet. The floodplain mussel may live out of water for a year or more, tightly closing its valves and switching to anaerobic metabolism when necessary. Its capacity to endure drought, low oxygen and high temperatures equip it for life in billabongs and other capricious wetland environments. It does not inhabit fast-flowing water because it cannot anchor itself against the current.

Notwithstanding their differences, the two mussel species often occur together at sites along the Lower Murray. The channel was once the domain of the river species, but this changed with the construction of weirs some 80 years ago. The weir pools favour the floodplain mussel more than the river species, and the former is now abundant in the shallow water along the banks. A similar invasion has occurred among the crayfish, with the common yabbie (*Cherax destructor*) having become common in the channel at the expense of the Murray crayfish (*Euastacus armatus*). The water plants that flourish along the river margins are also wetland refugees, dependent on the stable pools maintained by the weirs (e.g. Walker 2006).



**Figure 9.** The shell of the glochidium of the river mussel, *Alathyria jacksoni*, measuring 0.27 mm along the hinge between the valves. From Walker et al. (2001).

A remarkable feature of the mussels is that their larvae (glochidia) (Fig. 9) are obligate parasites on fish (Walker 1981). Several native and introduced fish are suitable hosts, but it is not clear whether this is true for the common carp (*Cyprinus carpio*). The glochidia of the floodplain



mussel may also be induced to parasitize tadpoles, and that may account for the occasional discovery of this species in ponds without fish. The male mussels release sperm into the water, to be taken up by the females. Over winter, the females brood their young in a pouch formed within the innermost demibranchs. As the water warms, in spring and summer, they release thousands of tiny (0.25-0.30 mm), bivalved glochidia into the water. If the sensory hairs on a glochidium happen to touch the gills or fins of a likely host, the valves snap shut, a cyst forms and the glochidium metamorphoses, over about three weeks, to become a juvenile mussel. It then breaks free and falls to the sediment, where it burrows for anchorage and protection. After 3-4 years, the young mussel attains sexual maturity and assumes the adult's habit, in water 2-4 m deep, along the river banks. Fish, then, are the agents of dispersal. By comparison, an adult mussel could not move far in three weeks!

Are mussels good to eat? In fact they are bland and unlikely to appeal to most palates, although creative chefs can make chowder and similar dishes. The mussels' propensity to filter organic particles from the water means that they accumulate metals, pesticides and other contaminants, and they are sometimes used to purify water in aquaculture. And, although the Murray species do produce occasional pearls, those few tend to be brown and misshapen (so-called "baroque" pearls), without commercial value. Nor is the shell likely to be useful to seed "cultivated pearls", as it is generally thin and discoloured by mineral inclusions.

### Snails

The aquatic snail fauna of the Lower Murray includes about 18 native species (Walker 1998; Ponder & Walker 2003). At least, that was true years ago. Most species now have very patchy distributions, and some have disappeared entirely from this region.

The tiny freshwater limpet *Ferrissia* (Ancylidae) is scarcely bigger than an "o" printed on this page. It is still widespread and found attached to snags, rocks or submerged plants, especially the broad leaves of water ribbons, *Vallisneria americana*.

The alien European pond snail *Physa acuta* (Physidae), a pulmonate, is another common species in the river and its freshwater wetlands and tributaries. It has a left-handed coiled shell, and a mottled, pigmented mantle that is clearly seen under the thin, transparent shell. The shell is difficult to distinguish from that of the native *Glyptophysa gibbosa* (cf. Fig. 1d), which occurs in the same kinds of habitats. There is evidence that *Physa* suppresses the growth of *Glyptophysa*, and could have contributed to its decline (Zukowski & Walker 2009).

The Planorbidae are pulmonate snails, variously with high-spired, coiled shells or small, flattened shells. They are rarely abundant, and most are patchily distributed. *Glyptophysa gibbosa* occurs throughout the Murray and freshwater wetlands and streams in the region. *Glyptophysa aliciae* occurs in flowing water in the Calperum and Chowilla areas, but has been recorded from the main channel near Tailem Bend. *Isidorella* has a patchy distribution that includes Pilby Creek downstream to Woods Point, and the Marne River and Reedy Creek catchments. The flattened shelled (planospiral) forms are restricted to a few locations from the Murray floodplain below Mypolonga; they include species of *Gyraulus* and *Helicorbis*.

The Lymnaeidae have distinctive right-handed, coiled shells with a wide aperture, and are notable because some species are vectors for sheep liver fluke. Most regional records of the lymnaeids *Austropeplea* and *Pseudosuccinea* are from the Mannum to Tailem Bend area.

Operculate snails include small species of Hydrobiidae. They include the alien *Potamopyrgus antipodarum*, introduced from New Zealand and now common in still and slow-flowing waters throughout the Murray and streams of the Eastern Mt Lofty Ranges. Others include *Posticobia*, found rarely in wetlands from Murray Bridge to Woods Point and Mitchell Gully Creek, and *Austropyrgus*, from the Marne and Somme Rivers and creeks in the Eastern Mt Lofty Ranges.

Pomatiopsidae are known only from Salt Creek in the Eastern Mt Lofty Ranges. In some saline lakes, beyond the Murray Valley, the shells of *Coxiella* are so numerous that they form dunes on beaches.

Thiaridae, including *Thiara balonnensis*, were last recorded from the Murray at Morgan in 1982, although their shells remain along the river banks and, ironically, living snails have become occasional pests in irrigation pipelines. This is true of the river snail *Notopala sublineata hanleyi* (Viviparidae), described below.



The general picture for snails is drastic reductions in range and abundance and regional extinctions of native species, and a few abundant alien species. While the disappearance of a few snail species may attract little attention, the decline of an entire group is evidence of profound changes in the river environment. Possible reasons for the decline include changes in the composition of biofilms, predation by common carp and changes in connectivity between the river and its floodplain wetlands.

Biofilms are mixed communities of bacteria, algae and fungi that appear as brown-green turf on submerged surfaces, including wood, rocks, plants and sediments (Sheldon & Walker 1997). They are at the base of the food chain for grazing invertebrates, including snails, and there is evidence that their composition may have changed in the aftermath of flow regulation. According to this idea, bacterial biofilms were more common prior to regulation, but the water-level regime introduced by the weirs has caused algae to become dominant. If this is correct, it is significant because bacterial biofilms are more nutritious for grazers. This is reflected in the carbon: nitrogen ratio, as low values (say, less than 10) indicate high nutritional value. In the present-day Murray, the C:N ratios generally are high, albeit variable, and so the biofilms may be an inadequate food resource for grazing snails. This is not true of the biofilms that occur in the dark confines of irrigation pipelines, where at least one snail species has found a refuge (see below).

The common carp gained access to the Murray after illegally imported, wild-strain fish escaped to the river in the 1960s (e.g. Koehn 2004). The carp dispersed rapidly and attained very high numbers, especially after floods in 1974-1975, and now are the dominant fish in the Murray-Darling Basin. Carp are indiscriminate feeders on benthic animals, including snails.

Another factor involves lost connections between the river channel and its floodplain wetlands. In general, flows in the Murray are much depleted by diversions for irrigation and other purposes, and the seasonal pattern of flows also is affected. The changed connectivity between river and floodplain has isolated the channel, the main corridor for dispersal of flora and fauna, from the wetlands that are nurseries and refuges for many species, including some snails.

One species, the river snail *Notopala sublineata hanleyi* (Viviparidae) is symbolic of the changed river environment (Fig. 10). Shells are common along the river banks, and we might suppose that the species is common. Yet *Notopala* began to decline after about 1950, when flow regulation began to intensify dramatically, and had virtually disappeared 25 years later.



**Figure 10.** Shells of the river snail *Notopala sublineata hanleyi*, with an operculum for comparison. The spires are about 15 mm long. Photograph: K.F. Walker.

In 1992, river snails were “re-discovered” in the Loveday irrigation pipeline (Sheldon & Walker 1993, 1997; Walker 1997). Not only had they returned from the brink of extinction, they occurred in such numbers that they blocked pipes, pumps and sprinklers and fouled the water. Irrigators were obliged to continually unblock their systems, accumulating large mounds of shells. The irony was taken up by the media, evoking headlines like “*The Revenge of the Gastropods*”. Some seasons are worse than others, and the snails have not been troublesome in recent years.

Why has *Notopala* abandoned its natural environment? Its presence in pipelines suggests that neither salinity nor any other water-borne pollutant is responsible, because the pipelines are fed from the river. Part of the answer may be that they are protected from predatory carp, although some carp do occur in the pipelines. Another may be that the biofilms growing on the inner walls of the pipes are bacterial rather than algal (as there is no light to sustain photosynthesis). These biofilms are

nutritionally superior to those found in the river, and that is significant because *Notopala* is viviparous, energetically a more costly mode of reproduction than laying eggs. The snails may have abandoned the changed river environment because the biofilms there now are less nutritious than they require.

### Zooplankton

Zooplankton, the free-floating, microscopic, animal component of aquatic food webs, includes Protista, Rotifera and “microcrustacean” (Copepoda, Cladocera). The composition and dynamics of zooplankton in the Lower Murray reflect contributions from headwater rivers, mainly the Murray and its tributaries (particularly the Goulburn), with seasonal inputs from the Darling River (Shiel et al. 1982). In the Murray above the Darling confluence, there is a cool-temperate, essentially lacustrine, microcrustacean-dominated community, drawn from numerous impoundments and backwaters. Algal populations are dominated by the diatom *Aulacoseira* (Hötzel & Croome 1996). The less regulated, highly turbid Darling River contributes a warm-temperate, rotifer-dominated riverine plankton. The turbidity restricts algal blooms, and the dominant zooplankters are bacterivores. Below the Murray- Darling confluence, there is a mixed assemblage dominated by rotifers, with fewer cladocerans and fewer still copepods.

Plankton samples taken from the Murray near Morgan in 2002-2003 (M.C. Geddes pers. comm., R.J. Shiel unpubl.) contained virtually the same suite of protists, rotifers and microcrustaceans recorded at Mannum by Shiel et al. (1982), more than 25 years ago. The assemblage included the protists *Stenosemella lacustris* (Ciliophora) and *Diffugia gramen* (Rhizopoda), 20 species of Rotifera (*Anuraeopsis*, *Asplanchna*, *Brachionus*, *Conochilus*, *Filinia*, *Hexarthra*, *Keratella*, *Polyarthra*, *Proalides*, *Synchaeta*, *Trichocerca*) and centropagid calanoid copepods (*Boeckella*, *Calamoecia*).

Other studies relevant to Riverland plankton include the role of turbidity in structuring zooplankton communities in Lake Alexandrina (Geddes 1984), and the role of river backwaters in reconstituting the microfauna of Murray tributaries (Nielsen et al. 2005).

### Rainpool microfauna

Rainpools are the commonest yet least-studied of all standing waters. They include water-filled hoofprints, roadside ditches and shallow depressions on floodplains or rock outcrops, and are often overlooked because they are ephemeral. In fact, they harbour a rich, diverse fauna (e.g. Bishop 1974; Williams 1975; Morton & Bayly 1977; Shiel et al. 1998).

Rainpool inhabitants cope with dry conditions by moving to permanent water, if they are mobile (e.g. insects), or by producing a desiccation-resistant resting stage (cysts, resting eggs, ephippia) which remains in the sediments (e.g. protists, rotifers, microcrustaceans), or by withstanding dehydration as adults (“anhydrobiosis”) (e.g. bdelloid rotifers, nematodes, tardigrades), or by aestivating in an arrested, earlier stage of the life cycle. Some propagules remain viable in sediments for more than 300 years (Hairston et al. 1995). There have been studies on the propagules in sediments of the Murray floodplain in the Riverland (Boulton & Lloyd 1991, 1992; Skinner et al. 2001).

Virtually any dry depression contains an animal “seedbank”, awaiting rain. The diversity of the seedbank depends on which organisms have reached it and survived to produce a resting stage. Resting stage densities of up to 1200 cm<sup>-3</sup> have been recorded from rainpool sediments on the Murray floodplain near Wodonga, Victoria (Shiel et al. 2001). In this survey, nearly 500 taxa were recorded, over half (252) of them Rotifera, but more than half of the species occurred at only one or two sites. Thus, even nearby communities are likely to differ, given the diversity of rainpool habitats and the chance nature of dispersal by wind or water.

The speed of response to wetting is remarkable. Protists and rotifers emerge within hours, microcrustaceans and small macroinvertebrates within a day or two, and copepods or ephippial cladocerans are breeding by the third or fourth day. *Boeckella major*, a predatory calanoid copepod of rainpools on the Murray floodplain, completes its life cycle in 1-3 weeks (Green et al. 1999). Fairy shrimps, shield shrimps, ostracods and other crustaceans emerge by day five. Not all the resting stages of a particular species hatch at the same time, providing some insurance against changeable conditions (e.g. Tan & Shiel 1993).

The macrofauna of rainpools is no less diverse. Insects likely to fly in include members of the Hemiptera, Coleoptera and Odonata. Some crustaceans such as Anostraca, Conchostraca, Notostraca,

and Ostracoda, have life cycles wholly adapted to temporary waters (e.g. Williams 1980). Their eggs remain in dry sediments and are blown about by wind, explaining their tendency to appear suddenly, in pools formed after heavy rainfall.

## CONSERVATION

For the most part, aquatic invertebrates have a low profile among conservation issues, reflecting our empathy with larger, more conspicuous species (the “charismatic megafauna”). This may change as resource managers become more aware of the loss of biodiversity in the Murray-Darling Basin and elsewhere, although there is a trend to identify and manage threats to entire ecological communities rather than individual species. The salient issue, however, is that we need to broaden our perceptions of “conservation” to acknowledge that invertebrates are a very large part of natural biodiversity, that they are no less fascinating than those animals with more popular appeal, and that they too are sensitive to the changing, changeable environment. The only aquatic invertebrate in the Riverland region listed as “endangered” is the Murray crayfish, although it was classed as a fishery species for the purposes of the legislation. The apparent decline of the molluscs reflects environmental changes, and the freshwater mussels in particular illustrate a common “blind spot” in our perceptions of conservation. These relatively long-lived species may fail to reproduce (strictly, to “recruit”) over a number of years, so that when the older individuals die, leaving no successors, it may be too late for remedial action. This lesson has been learned from the demise of related species in other parts of the world. It challenges our tendency to look at the natural world through only a narrow window in space and time.

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## CHAPTER 11. TERRESTRIAL INVERTEBRATES

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In contrast with freshwater invertebrates that are confined almost entirely to the freshwater biome along the Murray River (see Chapter 10), terrestrial invertebrates in the Riverland and Murraylands can be found in a wide variety of habitats, from soil and litter through to living on/in plants and animals.

Invertebrates affect all other forms of life by sheer weight of numbers. They make up the bulk of biodiversity and are essential for maintaining the function of ecosystems in many ways. For example, they form an integral part of the food web, they recycle organic matter, feeding on faeces or dead plants and animals, and they make up the bulk of parasitic species (along with microorganisms such as bacteria) which regulate animal and plant numbers.

Almost all species from the region have much broader distributions and can usually be found in adjacent regions of New South Wales and Victoria. Given that the number and diversity of species in the region is enormous, exemplar taxa are used to indicate the breadth and diversity of terrestrial invertebrates in the region.

### PLATYHELMINTHES (flat worms)

Platyhelminthes are soft-bodied, generally flattened, worm-like, unsegmented animals, with a wide range of body forms. The phylum is divided into three classes: Turbellaria, Trematoda and Cestoda. The majority of aquatic flatworms are turbellarians. Trematodes (flukes) and cestodes (tapeworms) are parasitic although some trematodes have free-living dispersive stages. The Australian terrestrial cestode fauna is poorly documented, with the best-studied host groups being the marsupials. Overall, cestodes have been reported from only 5% of vertebrate species (Beveridge & Jones 2002). Endemism among Australian flatworms is likely to be low in birds and high in parasites of frogs, reptiles and marsupials, and in free-living freshwater forms.

Quite a few trematode and cestode parasites of domestic animals occur in the region, including liver fluke of sheep and cattle (*Fasciola hepatica*), sheep whipworm (*Trichostrongylus axei*), sheep tapeworm (*Moniezia expansa*), and stomach hair worm (*Trichostrongylus axei*), which occurs commonly in sheep, other ruminants, and horses. Other examples include the tapeworm *Echinococcus granulosus* (hydatids), in which humans can act as intermediate hosts. Definitive hosts of this tapeworm are carnivores such as dogs, foxes and dingoes, while intermediate hosts are usually herbivores such as sheep, cattle, kangaroos, wallabies, and wombats. Cestodes are also commonly found in reptiles, birds, and marsupials in the region, and just one of many examples is *Oochoristica trachysauri*, a tapeworm from the sleepy lizard, *Trachysaurus rugosus* (Harvey Johnson 1932).

### NEMATODES (round worms)

Nematodes are unsegmented roundworms, which are not related to earthworms. They are commonly found in soil and freshwater habitats. Others are parasitic in roots and aerial parts of plants, in vertebrates, and in invertebrates such as insects. Free-living forms feed on living organisms such as bacteria rather than on dead organic matter. Discussion here is limited to terrestrial nematodes living in soil and/or parasitising plants. These nematodes are usually microscopic (most less than 3 mm long). Almost all soils contain nematodes, and in Riverland soils with high organic content there may be hundreds per gram of soil. In a survey of soil microinvertebrate assemblages, Bird (1999) found that nematodes dominated both in numbers and diversity in soils from the shores of Lake Alexandrina and the Murray River estuary at Goolwa (87 and 93% of total numbers of microinvertebrates, respectively). There is little knowledge of most Australian soil nematodes, and most species remain undescribed.

Plant parasites are the best known group of soil nematodes, due to their economic importance. These nematodes feed on plant roots, and occasionally within plant leaves. Tylenchids and aphelenchids have a hollow stylet or spear within the head, which they can insert into plant or fungal cells and through which they can suck out some of the cell contents. Other soil nematodes feed on bacteria, algae, fungal hyphae and other nematodes, and invertebrates such as protozoa, rotifers and enchytraeids (Yeates 1993). Rhabditid nematodes usually feed on bacteria, but some are predatory. They have a tube-like mouth, which may have teeth at its base. Rhabditids are important for the recycling of soil nutrients. Mononchids are predators, with teeth that they use to seize their prey. Dorylaimids include a large group of soil and freshwater nematodes, some of which feed on plants, others on bacteria and algae and possibly fungal hyphae, while some are predatory. The size and form of the spear is variable in dorylaimids; with long and slender spears in plant parasites, and short, hollow forms with wide openings in bacterial and algal feeders.

Soil nematodes are generally able to survive long periods of desiccation, important for their survival in many Riverland soils. They may survive dry periods as eggs, or they may coil and become inactive as the soil dries out, entering a state of anhydrobiosis in which metabolism is suspended. Whilst in this state, nematodes are brittle and their cuticle (body covering) is easily damaged by sharp soil particles. However, provided they are not damaged, they recover their normal functions when soil moisture rises again.

Many species of nematodes and a few genera associated with native plants are endemic to Australia, and several species are known only from South Australia. The greatest diversity of species is usually found in areas with moist soils, such as gullies. Only one study of nematodes from soils in the Riverland and Murraylands has been carried out (at Ferries MacDonald Conservation Park at Monarto; Frances Reay, unpublished data). Considering the similarity in climate, soils and vegetation, it is probable that the nematode fauna of the region is similar to that of adjoining regions in Victoria and New South Wales, where some survey work has been done (Sauer, 1985). In broad scale farming, there are many records of economically important tylenchids such as cereal cyst nematode (*Heterodera avenae*) and root lesion nematode (*Pratylenchus neglectus*). Root knot nematodes (*Meloidogyne* spp.), citrus nematode (*Tylenchulus semipenetrans*) and the dorylaimid *Xiphinema* sp. are commonly found in horticultural soils. *Sauertylenchus* sp., *Morulaimus whitei*, and *Paralongidorus eucalypti*, all widespread in Australia (McLeod et al. 1994), have been recorded from Ferries MacDonald Conservation Park. Also recorded were various rhabditids and dorylaimids, including stubby root nematode (*Paratrichodorus* sp.), which has a long stylet and feeds from outside a root, while the tylenchid genera *Radopholus* (burrowing nematodes), *Scutellonema* (spiral nematodes), *Hemicriconemoides*, *Criconemoides* (ring nematodes), and *Hemicycliophora* (sheath nematodes) are also known from this locality (Waite Insect and Nematode Collection, The University of Adelaide).



**Figure 1.** Shoot bud gall of *Fergusonina lockharti* (Photograph: Gary Taylor).



The river red gum (*Eucalyptus camaldulensis*) is host to at least four different combinations of cecidogenic (gall-forming) species of the nematode *Fergusobia* and fly *Fergusonina*. These nematodes and flies together form unique, species-specific mutualisms, in which the nematode relies on the fly for transport, and the fly on the nematode for gall induction (Taylor et al. 2005). Galls vary in form from large shoot bud galls (Fig. 1), which support hundreds of flies and nematodes, to flower bud galls, ‘stem’ galls and small axial bud galls that only support a few flies and accompanying nematodes.

### ANNELIDA (segmented worms; earthworms and leeches)

Apart from leeches (Hirudinea) which are treated in Chapter 10, earthworms (Oligochaeta) are found widely in the Riverland and Murraylands. Introduced earthworms (mostly Lumbricidae), including *Aporrectodea* spp., are more likely to be encountered than native species, especially in disturbed areas such as pastures, orchards, vineyards and gardens. This was the case in a survey of citrus orchards between Waikerie, Loxton and Paringa (Dalby 1999). Lumbricids were introduced into Australia by European settlers, and are native to Britain and other European temperate regions. Once introduced into a new area, they have the ability to breed rapidly and colonise new areas, often surpassing the original native species. The tiger worm, red worm or composting worm (*Eisenia fetida*), which is widely sold for composting, also occurs in the region, although it prefers rotting vegetation, compost, and manure rather than soil.

Megascolecidae is the only indigenous family of earthworms in South Australia, and although the fauna is impoverished, species endemism is relatively high (Jamieson 1974, 1981). Jamieson (1974) suggests that the paucity of the fauna is correlated with the low rainfall over most of the state. Unfortunately, most research in South Australia on earthworms appears to have taken place in higher rainfall areas (e.g. Mt Lofty Ranges, Fleurieu Peninsula and the South-East). Species from the Riverland and Murraylands include *Heteropodrilus shephardi*, collected near Bordertown and extending to the Wimmera River in Victoria (Jamieson 1974; Blakemore 1999, 2000), and *Gemascolex lateralis*, collected near Tailem Bend but more widely distributed in the Mt Lofty Ranges and Fleurieu Peninsula (Jamieson 1974).

### MOLLUSCA (snails and slugs)

Introduced land snails are very common in the region and are often a severe problem. These include the common white snail or vineyard snail (*Cerņuella virgata* – Fig. 2), white Italian snail (*Theba pisana*), conical/pointed snails (*Cochicella acuta* and *C. barbara* – Fig. 3), and small brown snail (*Microxeromagna vestita*). The garden snail (*Helix aspersa*) is also common in areas that are regularly irrigated (e.g. orchards and gardens). *Cerņuella virgata*, *T. pisana* and *Cochicella* spp. can be seen aestivating in the drier months on trees, fence posts, buildings or vegetation (Fig. 2). Aestivation is a form of dormancy where the snails seal off their aperture (opening) with a membrane and remain inactive for long periods of time.



**Figures 2-3.** 2 (left), Common white snail, *Cerņuella virgata*, aestivating on an old headstone at Black Hill Old Cemetery SA (Photograph: Gary Taylor); 3 (right), conical snail, *Cochicella* sp. (Photograph: SARDI Entomology).

Two introduced slug species, *Lehmannia flava* and *Milax gagates*, are common in irrigated situations. The former species is larger and yellowish, whereas the latter species is smaller and black.

### **CRUSTACEANS (slaters and land-hoppers)**

Terrestrial crustaceans such as isopods (Isopoda) (slaters) from the region are poorly known, but include the introduced European woodlouse (*Porcellio laevis*), the common woodlouse or pillbug (*Armadillidium vulgare*) and the delicate slater (*Porcellionides pruinosus*). Slaters are commonly found under debris and leaf litter in moist situations. Land-hoppers (Amphipoda) such as *Austrotriodes crenatus* are also common in shadehouses.

### **MYRIAPODS (millipedes and centipedes)**

Millipedes (Diplopoda) are long, thin arthropods with two pairs of legs on most posterior body segments (Harvey & Yen 1989), which feed largely on decaying plant material. Native millipedes have been poorly studied in the region, however the familiar introduced black Portuguese millipede (*Ommatoiulus moreletii*) occurs throughout the region and is mostly seen during the wetter times of the year.

Centipedes (Chilopoda) are long, thin arthropods, which unlike millipedes, never have more than one pair of legs per body segment (Harvey & Yen 1989). They live in moist situations such as in rock crevices, litter or under bark, and come out only at night to hunt and feed on insects, snails, worms and other smaller centipedes. Most species inject venom into their prey and, although the bite may be painful to humans, it usually only results in localised swelling and irritation. Colloff et al. (2005) recently provided the first comprehensive key to Australian centipedes. The widespread genera *Scolopendra*, *Cormocephalus* and *Ethmostigmus* occur in the region, as does the introduced European house centipede (*Scutigera coleoptrata*).

### **ARACHNIDS (spiders, scorpions and their relatives)**

#### *Araneae (spiders)*

The region represents a meeting point for many South Australian arachnids. Here many species from the higher rainfall south-eastern Australian region overlap in distribution with those of the semi-arid north. Additionally there is an influx of species following the riverine habitat from the eastern states and probably the most obvious of these is the common large River Murray huntsman spider (*Holconia murrayensis*) (Sparassidae) associated with the river red gums that provide adequate shelter in the form of hollows, crevices or large loose slabs of bark (Hirst 1991). Other spiders found along the river as an extension from the east include the water spider (*Dolomedes facetus*) and the jumping spider *Astia*.

In general terms most spiders are divided into the Mygalomorphae in which the fangs lie parallel and the Araneomorphae in which the fangs of most species are pincer-like. The Araneomorphae are further divided into primitive and more advanced families. Raven et al. (2002) have provided an interactive key to the identification of spider families.

The Mygalomorphae are known for their long life span and the females are often not seen unless the burrows are excavated. Males are more likely to be encountered as they wander in search of a mate in times of high humidity, usually after rain. Of the Idiopidae, *Blakistonia aurea* occurs to the west of the river and at least one species of *Aganippe* is found in the southern area. Both species have a door to a deep burrow with that of *Blakistonia* being semi-circular with a crenulated lip to both door and burrow entrance, and shaped like a clam shell. Other trapdoor spiders known from the Murraylands include at least three species of mouse spiders (Actinopodidae) which are the only mygalomorphs where the male wanders during daylight hours. The red-headed mouse spider (*Missulena insignis*) (Fig. 4) is common, whereas *M. rutraspina* is found in the extreme north-eastern sector and the smaller *M. dipsaca* is found in the mallee belt through to Western Australia (Faulder 1995). These two species have a black or blue-black head and females of all species are stout, dark brown to black, while that of *M. insignis* can be much larger than the male. Another large black conspicuous species is the brush footed trapdoor (*Idiommata scintillans*), the male of which has a coat of silvery hairs on the head (Raven 1994). The doors to their shallow burrows are well camouflaged and difficult to find.



**Figure 4.** Red-headed mouse spider (*Missulena insignis*) male, near Blanchetown SA (Photograph: Gary Taylor).

*Aname robusta* (Nemesiidae) may have the honour of being the first spider described from the Riverland and Murraylands but its status is obscure. Although the type from Mannum is in the Australian Museum, no other specimens have been attributed to the species. A pale *Aname* species occurs in the mallee country west of Morgan and other similar species are known in the southern areas. Species of *Kwonkan* are known to occur throughout the region but burrows are hard to find as a collar of loose silk is pulled together during the day to cover the entrance. Other species of Nemesiidae may spin a thin sheet of silk over the burrow entrance but none have trapdoors. The black wishbone spider group is represented by one large species, *Aname* sp., and aggressive males may be encountered after rains inciting fear that they may be the infamous funnel-web spider. In the northeastern sector the whistling or so called ‘bird-eating’ spider (*Selenocosmia stirlingi*) is locally common. Grasshoppers, other large insects, frogs, small lizards and the occasional mouse are likely to be a part of its diet as they are generally nocturnal (Kotzman 1990).

Primitive araneomorph families found in the region include Segestriidae (*Ariadna* or *Segestria*) in which some species construct strong dense tubes of silk under the bark of trees with the base of the tube often firmly secured in a borer hole or crevice, while others live in silk-lined burrows in the ground. Of the Oonopidae, also known as six-eyed spiders, the pouched spider (*Grymeus robertsi*) is one of many species found in the area. All are of smooth appearance, yellow or red-brown in colour and are seldom seen due to their small size and habit of living under bark, rocks or in leaf litter (Harvey 1987b).

River red gums provide an important habitat for many tree dwelling spiders found in the region and of these a number of species can be found in large numbers. Close inspection under slabs of old bark will reveal the small cryptic *Wandella murrayensis* (Filistatidae), a small spider with an exceptionally long life and the ability to go long periods with little food or moisture. Hollowed trunks as well as rocky cliffs provide an ideal habitat for the native daddy-long-legs spider (*Wugigara kaurna*) (Pholcidae), while the introduced species *Holocnemus pluchei* and *Pholcus phalangioides*, are more likely to be found in human habitation. Webs of the black house spider (*Badumna insignis*) (Desidae) can be seen in profusion on old red gums that have a thick layer of old bark at the base. Other members of Desidae found in the region include *Forsterina* spp. in sparse webbing under stones or logs, *Phryganoporus davidleei* which usually makes webs in the branches of shrubs, and *P. candidus* which is sometimes communal and produces dense webbing on the foliage of trees and can be a nuisance in orchards (Gray 1983). The much smaller *Callevopthalmus* sp. makes a small web over a depression in the ground and the spider waits for prey in a sheltered crevice at one side. A number of very small desid species are found on tree trunks or in leaf litter, while a pale coloured species inhabits saline areas subject to ephemeral flooding.

Three species of the Nicodamidae, red and black spiders (Harvey 1995), spin sparse webs at the base of tree trunks or in similar sheltered positions. The largest is *Nicodamus peregrinus* and the long legged male is most strikingly coloured. Two smaller species are *Ambicodamus leei* and *Durodamus yeni*. Despite the red colour signifying a warning these spiders are considered harmless. The platform spider (*Corasoides* sp.) (Stiphidiidae), lives in a short burrow in the ground from which a tube of silk rises funnel-shaped into the undergrowth, where it is transformed into a large silken platform above which straggled silk threads act as a knockdown for unwary insects flying through.

The more advanced araneomorph spiders include two species of water spiders (*Dolomedes*) although one species, *D. facetus*, has only been recorded near the Victorian border. Wolf spiders of the genera *Artoria*, *Trochosa* and *Venatrix* commonly occur on the flood plains of the Murray River with *V. speciosa* and *V. goyderi* being found close to water (Framenau & Vink 2001). The common *V. pseudospeciosa* is often found in gardens that enjoy regular watering. *Lycosa leuckartii* is the largest common species to be encountered and often has makeshift burrows in the cracked soil of the northern part (Steggles et al. 2003), but is replaced by *L. gilberta* in the southern part of the river floodplains. In drier areas away from the river wolf spiders are prevalent with at least 20 species known. Several species, one being *Dingosa serrata*, construct a palisade at the burrow entrance either of leaves, grass stems, small twigs or of silk and soil. Another smaller wolf spider, *Venonia* sp., makes horizontal sheet silk webs amongst leaf litter on the ground which are difficult to distinguish, except perhaps for their larger size, from those made by *Callevopthalmus* sp. (Desidae).

Other spiders which make webs on or near the ground include species of *Steatoda* (Theridiidae) and one introduced species, *S. grossa*, which is associated with human habitation. The tiny *Hadrotarsus* is also often found on the ground. The web of the redback (*Latrodectus hasseltii*) is almost a reverse of *Corasoides* with a funnelled retreat above and a tangle of sticky threads below to snare insects and even small reptiles moving on the ground. The grey house spider (*Achaearanea* sp.) is generally associated with buildings while several other species in the genus can be found in more natural situations, often on tree trunks. The related *Euryopis* sp. can be seen at night hanging on a silk line from the branches of tree trunks feeding on ants much larger than itself. On seizing an ant that is walking along the branch then dropping with it on a silk line, the ant is left powerless with nothing to grip onto.

*Tamopsis fickerti* (Hersiliidae) employs a different tactic and, on encountering prey, circles it rapidly playing out streams of silk from long spinnerets (hence the name ‘two-tailed spiders’) rendering the insect helpless. One member of the Thomisidae, *Tmarus* sp., appears to use the same technique as *Euryopis* while other family members, divided between crab spiders (*Stephanopis* and *Tharpyna*) and flower spiders (*Sidymella* and *Diaea cruentata*), are sit-and-wait predators. *Stephanopis cambridgei* (Thomisidae) is found under bark while a second species has been collected more often in leaf litter. Huntsmen spiders can similarly be divided with the sit-and-wait ambush method being employed by *Isopeda leishmanni* (south of Mannum), *Isopedella cerussata* (northern sector) and *I. saundersi* in mallee areas, while adjacent to the Victorian border is *I. frenchi*. *Delena cancerides* is rarely seen as it hunts under large loose slabs of bark rather than on tree trunks. This habit is somewhat emulated by *Holconia nigrigularis*, a close relative of *H. murrayensis*, but which is restricted to the drier north-eastern corner of the region and more likely to be found in stands of large *Casuarina* trees. *Pediana occidentalis* can be found north of the river in mallee vegetation and an intricate pattern of drab colours ensues that it is well camouflaged against the bark. The more active badge huntsmen (*Neosparassus* spp.) may be seen at night moving over the foliage of low shrubs in search of prey. Two species common on the east coast, *N. calligaster* and *N. punctatus*, can be found in the southern Murraylands. Another six or seven species known in the area are undescribed. Several are recorded from only one or two localities, only one is found throughout the area and all are restricted to undisturbed natural vegetation. *Keilira sparsomaculata*, one of South Australia’s smallest huntsmen, with a body length of less than 1 cm, is another active species occurring in heathland in the south.

Sac spiders (Clubionidae) are represented by two common genera. Pale sac spiders (*Cheiracanthium*) prefer foliage and seem to have a preference for prickly bushes or thistles, while *Clubiona* is found under the bark on trees and constructs short silken tube-like nests (Austin 1984). Also found on trees and shrubs are some members of Gnaphosoidea, a large group comprised of several families which include the Gnaphosidae, Galliellienidae (*Meedo mullaroo* and *Oreo renmark*,



both of which are rarely collected) (Platnick 2002), Prodidomidae with unusual long mid ventral spinnerets (Platnick & Baehr 2006), and Trochanteriidae, members of which are referred to as ‘flat rock spiders’ although the common species, *Longrita millawa*, can also be found under bark on tree trunks (Platnick 2002). Liocranidae is little known and one small species of *Orthobula* is recorded by an undescribed species. The notorious white-tailed spider (*Lampona cylindrata*) (Lamponidae) (Fig. 5), naturally occurs under bark or in sheltered rocky areas where it preys on other web building spiders such as the black house spider. There are a number of closely related but smaller relatives, *Lampona gilles*, *L. braemar*, *Lamponina loftia*, *L. asperrima*, *Bigenditia millawa*, *Notosdipus dalby*, *Asadipus bucks* and *Prionosternum nitidiceps* (Platnick 2000). The latter was described in 1909 from Western Australia and represents an element of the Riverland and Murraylands fauna that had a distribution once continuous across South Australia and into Western Australia. Aridification has now confined these once widespread species to the shrinking refuges that remain.



**Figure 5.** White-tailed spider (*Lampona cylindrata*) (Photograph: Mark Harvey).

The Gnaphosidae are well represented with numerous species of *Encoptarthria* and *Hemicloea*, and at least one species in both *Anzacia* and *Eilica*. Many other undescribed genera and species occur in the region. The extremely flattened species of *Hemicloea* are adapted to living between layers of bark but the other genera are found on the ground, under rocks or living in leaf litter.

Other spiders living on the ground include the large family of spotted ant spiders of which *Neostorena victoria* is a burrower and adorns its burrow entrance with a palisade of large leaves. The female is about twice the size of the male and is possibly the largest species in the family. *Storena formosa* is occasionally encountered under objects on the ground and, as with other members of the genus, has large conspicuous yellow spots on the abdomen (Baehr & Jocqué 1994). The much smaller *Pentasteron intermedium* and *P. storosoides* feed on small black ants, while *Habronestes bradleyi* is usually encountered close to nests of meat ants (*Iridomyrmex*). Little is known of the habits of other species including *Cavasteron crassicalcar* and *C. atriceps*, *Holasteron aciculare*, *H. hirsti* and *H. spinosum*, *Australutica quarens*, *Zillimata* sp. and many other undescribed species.

Numerous species of the closely related Zoridae and Miturgidae occur throughout the region but little is known about them. Zorids may be seen during the day as they move around cautiously through the leaf litter layer, but species of *Miturga* (lined spiders) are strictly nocturnal and make silken tube-like nests under stones, logs or even in dense low shrubs near the ground. The white and yellow spotted *Supunna picta* (Corinnidae) is often seen as they dart about in open areas even on sunny days. Other genera such as *Poeciliptra* are more ant-like in appearance and may have metallic hues. An unnamed genus and species with a colourful reddish or even blue anterior body constructs short burrows in the ground with a palisade of silk-bound debris rising from the entrance.

Jumping spiders (Salticidae) may well be the most diverse and numerous group in the region, however most are small and grey or brown coloured and not readily seen. *Ocrisiona melancholica*, *Helpis occidentalis*, *Servaea vestita*, *Breda jovialis* and *Clynotis severus* are larger species commonly found under red gum bark. All may be seen hunting on tree trunks during the day. *Breda jovialis* has a distinctive yellow-gold patch on the abdomen and also preys on the black house spider. Small, flattened species of *Holoplatys* and *Zebraplatys* find their way into narrow crevices between the bark of mostly smaller eucalypts. One of South Australia's largest and brighter patterned jumping spiders, *Sandalodes superbus*, lives exclusively in rolled bark that has dislodged from a tree and been caught in shrubs below. Although recorded from both the northern and southern fringes of the Riverland and Murraylands, it no doubt would be present along the river valley. *Lycidas* is a large genus with a number of species in the area which are usually seen on the ground. The male of *L. chrysomelas* is brightly coloured and looks very unlike the drab female. Both sexes of *Jotus*, in which most species have a distinctive black stripe along the middle of the abdomen, are often seen during summer days in a variety of habitats close to the ground. A species of *Grayenulla*, though commonly collected in pitfall traps, is seldom seen and little is known of its habits. Another family of spiders that have jumping movements, particularly when disturbed, is Oxyopidae. *Oxyopes molarius*, *O. amoenus* and *O. quadrifasciata* are all of similar appearance and live in low herbaceous vegetation. The eggsac is constructed in the upper branches and the female sits astride it guarding against predators.

Many spiders are known to 'balloon' or travel across country on wind-carried silken threads. This practice is not restricted to young spiderlings dispersing from the maternal nest but includes adult midget spiders (*Laetesia* and *Dunedinia* spp.) (Linyphiidae) and even small wolf spiders of the genus *Artoria*. For those species the Murray River is certainly no barrier to dispersal to new suitable habitats. It is likely that the orb-weavers (Araneidae) are also adept at ballooning as at least the larger common garden orb-weavers (*Eriophora heroine*) appears to have established in New Zealand (Court & Forster 1988). Along with *E. biapicata*, this species is found in natural areas or cultivated paddocks, although *E. biapicata* prefers to make a web higher up off the ground whereas *E. heroine* will often have a web lower to the ground in grass. Grass or low shrubs is also preferred by the teardrop spider (*Argiope protensa* and *Gea theridioides*), while the bird-dropping spider (*Celaenia kinbergi*) is more likely to be found hanging in leafy foliage of fruit trees. Orb-weaver species common along the riverbanks that rely on a more humid atmosphere are *Araneus eburnis*, the spiny spider (*Austracantha minax*) which can be seen in semi-communal webs in low shrubs, and the well camouflaged *Larinia phthisica* which makes a small orb web amongst the long grass and sedges (Framenau & Scharff 2008). *Araneus usualis* is normally found on dead trees or branches particularly near water. At least two species of the wrap-around spider (*Dolophones*) are found in the branches of trees and rest during the day with their legs partly tucked beneath the peculiar-shaped body resembling a node on a small branch. *Carepalxis* and a species of *Poltys* have a similar habit but the abdomen of the latter is much higher and its unusual shape can vary considerably (Smith 2006). Numerous orb-weavers of the family Tetragnathidae abound. The long-jawed spiders most often build their orb-webs over water amongst rushes. The golden-orb weaver (*Nephila edulis*) is widespread and can be very common in some seasons (Harvey et al. 2007), while the equally common leaf curlers (*Phonognatha graeffei* and *P. melania*) are less obvious. In borer holes of dead trees *P. dimidiata* shelters during the day emerging at night to make a web. The tiny *Pararchaea* (Pararchaeidae) has been collected from Scott Creek near Morgan. When hunting, the fang bases (chelicerae) are held at right angles to the body and on encountering prey, the fangs likely close upon it rapidly and with some force (Rix 2006).

### **Opiliones**

The paucity of Opiliones in museum collections from the Riverland and Murraylands suggests they may be rare. In fact there have been just four specimens collected from the area. However, it may reflect more their cryptic nature as those four specimens represent three species and three families. The short legged harvestman *Dampetrus gracilis* (Triaenonychidae) was previously known only from Red Cliffs, Victoria (Forster 1949), and following a recent survey near Chowilla is now a new record for South Australia. Two long-legged Opiliones are known, *Megalopsalis* sp. (Megalopsalididae) and the tiny fragile looking *Ballarra* sp. (Neopilionidae) which is considered to be restricted to Gondwanan continental fragments (Hunt & Cokendolpher 1991).

### *Acari (mites and ticks)*

Although many mites are predatory, there are also many species of plant feeders, fungivores, saprophytes, pollen and nectar feeders, microbial filter feeders, and internal and external parasites on a wide range of vertebrate and invertebrate hosts.

Most mites commonly encountered in extensive and intensive agricultural systems are introduced and are usually considered pest species. Among those commonly found in horticultural crops are the pest species two-spotted mite (*Tetranychus urticae*) and bryobia mite (*Bryobia rubriocolus*). Two-spotted mite has a large host range including apple, pear, peach, plum, prune, strawberry, bean, cucurbits, as well as a very wide host range among woody and herbaceous crops and weeds. Both nymphs and adults feed by piercing and sucking the lower leaf surfaces, leading to the development of yellow-green mottling and loss of vigour and yield. The beneficial mite (*Phytoseiulus persimilis*) is common in horticultural regions where it has been introduced. It is a voracious feeder on all stages of two-spotted mite. Bryobia mite can be found on almonds, prunes, plums, pears, apples and apricots, although it is most damaging to almonds, prunes and sometimes pears. This mite damages epidermal cells by piercing the cells and sucking out the contents. The upper leaf surfaces have a stippled green grey appearance. The bronzing typical of two-spotted mite is not produced.

Eriophyid mites (Atopomelidae), grapeleaf blister mite (*Colomerus vitis*) grapeleaf rust mite (*Calepitrimerus vitis*), and bunch mite (*Brevipalpus californicus*) (Tenuipalpidae) can all be found on grape vines in the region. Grapeleaf blister mite feeds on the undersides of leaves producing yellow areas which develop into raised blister-like swellings on the upper surface. The cavities on the underside of the blister are filled with felt-like hairs. The life cycle of the grapeleaf rust mite is similar except that it mainly occurs on the upper side of the leaves, whereas bunch mites are most commonly found in the bunches and on the undersides of adjacent leaves.

A very common pasture and crop pest in the region is redlegged earth mite (*Halotodeus destructor*) (Penthaleidae). This mite feeds on a wide range of field crops, vegetables, flowers, pastures and weeds. It is mainly a pest of broad-leafed plants and feeds by puncturing plant cells and sucking up the sap, producing a mottled effect which gives the leaf a grey or white appearance.

Not all mites in the region are pest species. For example, several families including Erythraeidae and Trombidiiidae have larvae that are parasitic on insects, and have terrestrial adults that are often called red velvet mites, including (*Erythrites reginae*) which is commonly seen in crops and pastures in the region.

The order Sarcoptiformes includes soil mites in the Suborder Oribatida, commonly known as oribatid mites. These are mostly slow-moving, heavily sclerotised mites that are most abundant and diverse in the upper layers of soil and associated organic litter. They feed on dead plant material and associated microfungi. The Sarcoptiformes also includes the Suborder Astigmata, including the common flour mites and cheese mites that infest stored food and which damage stored grain, the house dust mites that cause allergy and asthma in humans, and a wide range of fungivorous and saprophytic species in soil. Many of the Astigmata have a specialised immature form known as the hypopus, which attaches itself to insects for dispersal. The Astigmata also includes feather mites of birds. Domrow (1992) catalogued the species of Astigmata that parasitise Australian vertebrates or occur on their bodies. These include the mites that cause such diseases as scabies (*Sarcoptes scabiei*), scaly leg of chickens (*Knemidokoptes mutans*) and several types of mange in domestic animals (e. g. the ear mite of dogs and cats, *Otodectes cynotis*).

There are about 80 known species of hard ticks, Order Ixodida, in Australia (Roberts 1970; Keirans et al. 1994, 1996). They are all blood-feeding parasites of mammals, reptiles and birds, and are characterised by mouthparts with a large number of backward-directed hooks that are inserted into the host for feeding. Of these, the reptile tick (*Aponomma hydrosauri*), can be found parasitising sleepy lizards (*Trachydosaurus rugosus*) and bluetongues (*Tiliqua* spp.) on the north bank of the Murray River but has not yet been found more than a few kilometres beyond it. The reptile tick, *Amblyomma limbatum*, also occurs in the region north of the 400-mm isohyet (Smyth 1973). Other ticks from reptiles include the goanna ticks *Ap. jimbriatum* and *Ap. undatum* (Smyth 1973), and the common marsupial tick (*Ixodes tasmani*).

### **Scorpions**

The scorpion fauna of southern Australia comprises representatives of three families, Buthidae, Bothriuridae and Urodacidae, all of which are found in the Riverland and Murraylands (Koch 1977). Scorpions are active predators, emerging from their burrows at night to prey upon other invertebrates.

They give birth to live young which are carried on their mothers' back for several weeks prior to dispersal. The buthid fauna of the Murraylands comprises several species of *Lychas* and a single species of *Isometroides*, which are characterized by their slender pedipalps and generally mottled yellowbrown colour. Species of *Isometroides* have a highly specialised diet, as they preferentially hunt burrow-dwelling spiders such as trap-door and wolf spiders. The Bothriuridae contains the single genus *Cercophonius* which tend to be best represented in forested ecosystems. *Cercophonius kershawi* occurs in the Murraylands in leaf litter of mallee trees. The Urodacidae with the sole genus *Urodacus* are restricted to mainland Australia (Koch 1977), and include the largest scorpions in the country. *Urodacus manicatus* lives in shallow depressions under rocks in south-eastern Australia, including the Murraylands. The desert-dwelling *U. yaschenkoi* has been recorded near Renmark, Berri and adjacent regions of Victoria and New South Wales, where it digs deep burrows in the soft soil of dunes (Locket 1990) (Fig. 6). *Urodacus armatus* digs shallower burrows on the dune base and interdune swales (Locket 1990).



**Figure 6.** Scorpion – *Urodacus yaschenkoi* (Photograph: Mark Harvey)

### **Pseudoscorpions**

These are small arachnids, less than 1 cm in length, which resemble scorpions but lack the elongate tail and terminal sting. Although fairly common in most terrestrial ecosystems, they are rarely observed due to their secretive habits. Most research on these animals in South Australia has concentrated on cavernicolous species from the South-East and the Nullarbor Plain. The regional fauna is represented by just a few families. Several species are commonly found under the bark of trees, particularly eucalypts, including the common *Oratemnus punctatus* (Atemnidae), several species of *Conicochernes* (Chernetidae) and at least two species of *Synsphyronus*, *S. dewae* and *S. niger* (Garypidae) (Harvey 1987a). Others occur closer to the ground where they occupy leaf litter and soil habitats. The best known examples include the widespread *Geogarypus taylori* (Geogarypidae), which occurs over much of southern Australia (Harvey 1986), and *Synsphyronus mimulus*, but other families such as the Cheliferidae, Olpiidae and Chthoniidae also occur in the area.

## **INSECTS**

### **Introduced and native insect pests**

Introduced insects are widespread and common in the region, with most being able to be treated conveniently as occurring in either intensive or extensive agricultural systems. Whilst some of the



introduced insects such as the cabbage white butterfly (*Pieris rapae*) and honey bee (*Apis mellifera*) are ubiquitous, most are confined to the agricultural areas, tending not to be so common in natural environments where plant and animal hosts often do not occur.

The intensive agricultural systems include crops such as citrus, stone fruits, grape vines, and vegetables, and also irrigated pastures and some dairying, particularly along the lower reaches of the Murray River. In comparison, extensive systems are mostly dryland where cereals, as well as crops such as canola and field peas, and domestic stock such as sheep and cattle, are farmed.

#### *Introduced insects associated with intensive systems*

The majority of introduced insects found in intensive systems are considered pest species. Many of these can also be found in extensive systems.

There are a large number of introduced pest Hemiptera, and among the more significant are: mealybugs such as citrus mealybug (*Planococcus citri*) and the longtailed mealy bug (*Pseudococcus longispinus*); scale insects such as soft brown scale (*Coccus hesperidum*), grapevine scale (*Parthenolecanium persicae*), black scale (*Saissetia oleae*), red scale (*Aonidiella aurantii*) and yellow scale (*A. citrina*); aphids such as the black peach aphid (*Brachycaudus persicae*), green peach aphid (*Myzus persicae*), black citrus aphid (*Toxoptera aurantii* and *T. citricida*), and cabbage aphid (*Brevicoryne brassicae*); and true bugs such as the green vegetable bug (*Nezara viridula*).

Several thrip species (Thysanoptera) are significant in the region, particularly the western flower thrips (*Frankliniella occidentalis*). This thrip can attack more than 250 plant species including many vegetable and flower crops, and many weed species. Preferred hosts include capsicums, lettuce and chrysanthemums (Mound & Gillespie 1997). It causes flecking, silvering or deformation of flowers, growing tips, young foliage, stems and fruit, and can transmit tomato spotted wilt virus. Other species include Kelly's citrus thrip (*Pezothrips kellyanus*) (Colloff et al. 2003) and tomato thrips (*Frankliniella schultzei*).

Fuller's rose weevil (*Asynonychus cervinus*), the vegetable weevil (*Listroderes difficilis*) and whitefringed weevil (*Naupactus leucoloma*) are among the more important introduced pest beetles. The economic damage caused by Fuller's rose weevil is mainly confined to citrus, but a wide range of plants such as olive, ornamentals such as roses, some vegetables, as well as many weeds are attacked. The vegetable weevil attacks cruciferous plants, especially cabbages and turnips, celery, carrots, lettuce, silver beet and spinach, winter growing flowers such as pansies, and weeds such as marshmallow and capeweed are favoured by the larvae. Adults also damage crucifers, potatoes and tomatoes. Adult whitefringed weevils (Fig. 7) cannot fly and emerge from the soil in summer. The larvae of this weevil are highly polyphagous and chew the roots of a wide range of plants including clovers, lucerne, peas, water melons, tomatoes, potatoes, rye grass, wheat, brassicas and onions. The African black beetle (or lawn beetle) (*Heteronychus arator*) (Scarabaeidae) is also widespread and common. It has a broad host range including maize, lawns, tomato, cabbage, potato, rhubarb and various garden flowers, and other market garden plants, and a wide range of grasses, but not legumes. Most damage is caused by the last larval stage feeding on roots, while young larvae tend to feed on soil organic matter. Adult beetles chew on stems just below ground level.

A number of introduced pest moths and butterflies can be found in the region, among them cabbage white butterfly (*P. rapae*), oriental fruit moth (*Grapholita molesta*), codling moth (*Cydia pomonella*) and diamondback or cabbage moth (*Plutella xylostella*). The cabbage white butterfly is perhaps the most obvious, and its host plants include cruciferous plants, especially cabbages, cauliflowers, brussels sprouts, broccoli, turnips, radish, and mustard, as well as various weeds and garden flowers.

A wide range of beneficial insects has been introduced into the region as biological control agents. Many of these are minute parasitic wasps such as *Aphytis melinus* (Aphelinidae), which is a parasite of armoured scale insects, particularly red scale in citrus. It lays eggs under the scale cover, and the developing larva feeds on the scale insects killing it, and eventually emerging as another adult wasp.



**Figure 7.** Whitefringed weevil adult (*Naupactus leucoloma*) (Photograph: SARDI Entomology).

#### *Introduced insects associated with extensive systems*

The lucerne flea (*Sminthurus viridis*) (Collembola: Sminthuridae) is a native of Mediterranean areas and is well established in southern Australia. Hosts include medics, clovers, beans, cereals and a wide range of mainly broad-leaved crops and weeds. Lucerne flea causes damage by chewing, and results in the leaf surface having an exposed ‘membrane’ or epidermis giving a characteristic ‘window’ on the remote surface. Minute black specks of excreta are also present.

The African mole cricket (*Gryllotalpa africana*) (Gryllotalpidae) are quite distinctive. They are velvety brown, thick-set, about 25-40 mm long, and have strong forelegs well adapted for burrowing. The fore wings are very short, but they can fly quite strongly. The hind wings are large and fan-like when unfolded. Mole crickets damage grasses and sometimes tomatoes, vegetable and flower crops by tunnelling through the roots. Succulent stems may be chewed at ground level. They may occasionally chew into root crops such as carrots and turnips.

Among the various aphid (Aphididae) pests of extensive crops are bluegreen aphid (*Acyrtosiphon kondoi*), which is a native of Korea and Japan. It is chiefly a pest of lucerne, subterranean clover and medics but also peas, lentils, yellow lupins and melilotus can be hosts. The aphids suck sap from the terminal areas of plants; those parts that are tender and succulent. Stems often become severely stunted with small leaves and shortened internodes. The leaves turn yellow, curl and may drop off in severe infestations. Cowpea aphid (*Aphis craccivora*), oat or wheat aphid (*Rhopalosiphum padi*), pea aphid (*Acyrtosiphon pisum*) and spotted alfalfa aphid (*Therioaphis trifolii* f. *maculata*) are also common in extensive crops.

Various introduced beetles are pests. These include sitona weevil (*Sitona discoideus*) (Curculionidae), which was first recorded as a pest in New South Wales in the late 1950s, but now occurs across southern Australia, including Tasmania. It is a significant pest of annual medics and lucerne. The pea weevil (*Bruchus pisorum*) which, despite its common name, is not a weevil, but rather a member of the Bruchidae. It is a cosmopolitan pest of field peas, and was first recorded in South Australia in 1964.

A range of storage insect pests are commonly found in and around grain storage areas on farms, and occasionally in people's houses. These include the lesser grain borer (*Rhyzopertha dominica*), granary weevil (*Sitophilus granarius*), rice weevil (*S. oryzae*), and several species of spider beetles (Anobiidae) including the shiny spider beetle (*Mezium affine*). The Indian meal moth (*Plodia interpunctella*) is also commonplace.

#### *Introduced insects associated with domestick stock*

As well as pests associated with crops and pastures, there is a wide range of introduced pest species associated with domestic stock. These include fleas such as the cat flea (*Ctenocephalides felis*), dog flea (*C. canis*), and poultry stickfast flea (*Echidnophaga gallinacea*). A number of important lice are also found in the region as ectoparasites of domestic stock and include shortnosed cattle louse (*Haematopinus eurysternus*), longnosed cattle louse (*Linognathus vituli*), and sheep body louse (*Bovicola ovis*).

The sheep nasal botfly (*Oestrus ovis*) (Oestridae), is common and widespread. The female fly lays batches of about 50 newly hatched larvae in or on the nostrils of the sheep; the larvae crawl onto the mucous membrane of the nasal passage, where they remain for at least two weeks. Development of the first instar larva may be delayed for one to nine months, and plays a role in the over-wintering cycle. The pupal stage occurs in the ground and lasts one to two months depending upon temperature.

Sneezing and a mucopurulent nasal discharge (snotty nose) may be a result of botfly larvae. The main effects for the host are persistent annoyance and associated debility.

Introduced beneficial insects are found in the region include dung beetles, such as *Aphodius lividus*, which have been introduced to supplement the native dung beetles such as the threehorned dung beetle or Kershaw's burying beetle (*Onthophagus mniszehi*) (Tyndale-Biscoe 1990). As well, various parasitic wasps such as lucerne aphid parasite (*Aphidius ervi*), are widespread and common.

#### *Species commonly associated with humans and their domestic pets*

Apart from those insects included above which can often be found in home gardens, there are a range of insects commonly associated with humans, either in their homes and gardens, or actually attacking humans or their pets.

Several cockroach species (Blattodea) are common in the region, particularly the German cockroach (*Blatta germanica*), Oriental cockroach (*B. orientalis*), American cockroach (*Periplaneta americana*), and the native Australian cockroach (*P. australasiae*). The latter species has been spread by humans so that it is found extensively throughout Australia as well as overseas.

Termites (Isoptera) play an important role in nutrient recycling, but the feeding of some species may be of considerable economic importance with damage occurring to timber in buildings and fences. In the region, common pest termites include *Coptotermes acinaciformis* (Rhinotermitidae), with nests which are subterranean or in stumps or hollowed trunks of dead or living trees, and *Nasutitermes exitiosus*, with largely subterranean nests. Other termites include *Cryptotermes* spp., *Coptotermes frenchi* and *Heterotermes ferox* (Watson and Abbey 1993).

The most widespread introduced earwig (Dermaptera) is the European earwig (*Forficula auriculari*). Earwigs are rather cryptic, small to medium sized insects distinguished from other insects by a pair of forcep- or pincer-like cerci at the end of the abdomen. *Forficula auricularia* can be a pest of gardens and vegetable crops, and is relatively common around homes. In other areas of the state, it has caused damage to the seedlings of emerging canola crops.

The rose aphid (*Macrosiphum rosae*) is the most common insect pest of roses in Australia. Aphids damage plants by sucking sap from plant tissues using highly specialised sucking mouthparts. The usual symptoms of damage are distortion of new leaves and flowers. Aphids also excrete large amounts of honeydew, which can cover the plant, resulting in the growth of sooty moulds and the consequent reduction in photosynthesis and aesthetic appeal. Severe infestations can result in defoliation of the plant and loss of the flowers. As with most aphids, they can breed very rapidly and build up vast numbers, especially in warm, humid weather. The rose aphid parasitoid (*Aphidius rosae*), has been successfully introduced from Italy as a biological control agent (Kitt & Keller 1998).

Recently, bed bugs (*Cimex lectularius*) (Cimicidae), have undergone a dramatic resurgence and worldwide there are reports of increasing numbers of infestations. Bed bugs are wingless bloodsucking insects, oval in shape and 4-5 mm long when fully grown. They are rust brown in colour and change to a deeper red-brown following a blood meal on their human host. Bed bugs are dorsoventrally flattened and being thin means that they can hide in narrow cracks and crevices, making detection often very difficult. The mouthparts of bed bugs are especially adapted for piercing skin and sucking blood, and each nymphal stage requires a blood meal. Like most blood sucking arthropods, they inject saliva during feeding, which has anticoagulant properties. Bed bugs respond to warmth and carbon dioxide of a host and quickly locate a suitable feeding site. Most blood feeding occurs at night, and they generally seek shelter during the day and become inactive while digesting the blood meal. They can survive for long periods without feeding. While their preferred host is human, they will feed on wide variety of other warm-blooded animals including rodents, rabbits, bats, and even birds.

Many species of Muscidae are of economic importance including the cosmopolitan housefly (*Musca domestica*) which is found in association with humans or activities of humans, and was probably introduced with European settlement. It is commonly found around homes and also pig and poultry enterprises, horse stables, and farms. Not only are they a nuisance, but they also can transport disease-causing organisms. Excessive fly populations are obnoxious, and they can pose a public health problem. Other important species include the bush fly (*M. vetustissima*), and the cosmopolitan stable fly (*Stomoxys calcitrans*).

The most common blowfly (Calliphoridae) of the region is the Australian sheep blowfly (*Lucilia cuprina*), which is responsible for initiating most blowfly strikes in sheep. It is, however, not endemic to Australia and was accidentally introduced into Western Australia in the late 1860s, most likely from South Africa (Norris 1990).

Australia's early European settlers introduced honey bees (*A. mellifera*) to ensure a good supply of honey. They are now wild throughout Australia, including the Riverland and Murraylands. Honey bees play an important role as pollinators of crops and wild flowers, but some wild flowers have suffered from the presence of honey bees as these flowers can only be effectively pollinated by native bees or birds (Paton 1997, 2000). As well, by building their hives in places such as hollow tree limbs, birds and small mammals lose places in which to shelter and construct their own nests. Honey bee stings cause intense local pain and swelling, and a severe allergic reaction may occur in some people.

The European wasp (*Vespa germanica*) is native to Europe, North Africa and temperate Asia, and reached the Australian mainland (Melbourne) in 1977. It has since spread rapidly, and can now be found in many towns and cities in south-eastern Australia. Nests are located where shelter is available, with common locations being retaining walls, tree hollows, wall cavities, or underground (Kasper et al. 2008). The nest is made of grey papier mache-like material. If a European wasp is aggravated it may sting, but unlike the honey bee, it can sting multiple times. Some people are allergic to the stings. Through sheer numbers, the predation of European wasps on native insects and honey bees can be deleterious but they can also affect native animals (including birds) through competition for food. The European wasp is sometimes confused with native paper wasps, *Polistes* spp., which are very similar in colour but have antennae that contain yellow, orange or brown markings as well as black. European wasps have antennae that are entirely black. Their nests are also distinctive, being a single 'honeycomb-like' layer which is usually horizontal. If 'honeycomb' cells can be seen it is unlikely to be a European wasp nest.

#### ***Native insects that have pest status***

When environmental conditions are right, native insects can build up in sufficient numbers to become pest species. In intensive systems, this may be a continual problem (i.e. the insect is considered a key pest) or if it occurs spasmodically, then the pest is considered an occasional pest.

The native plague thrips (*Thrips imaginis*) (Thripidae), has been recorded in all states in Australia, and has a host range that includes pears, grapes, peaches, strawberries, roses, and some vegetables. When numerous, they may be found in just about any flowers. Both nymphs and adults cluster inside the flowers and buds where they feed by rasping the surfaces and sucking the exuding sap.

The Australian plague locust (*Chortoicetes terminifera*) (Acrididae) is distributed widely across mainland Australia. Certain conditions lead to migration southward into areas not normally occupied, for example, cereal areas of South Australia. The locust has a wide host range of native grasses, but also attacks cereals, pastures and a range of horticultural crops and weeds. Other native acridid grasshoppers and locusts found in the region include the small plague grasshopper (*Austroicetes crucita*) and wingless grasshopper (*Phaulacridium vittatum*).

The black field cricket (*Teleogryllus commodus*) (Orthoptera: Gryllidae), is found in all southern Australian states, especially where soils are heavy and tend to crack on drying. Young plants of many types may be occasionally damaged. Plants attacked include ryegrass, lucerne, clovers, oat, maize, potato, bean, carrot, strawberry, cotton and sunflower.

Cottony cushion scale (*Icerya purchasi*) (Margarodidae), is considered to be native to Australia, although it was originally described in 1879 from specimens found in New Zealand. Its host range includes citrus, *Acacia* spp., climbing fig (*Ficus* spp.), roses, laburnum, and mulberry. In South Australia, control measures are seldom necessary, as this insect is kept in check by the predacious vedalia ladybird (*Rodolia cardinalis*) and their larvae, and is also parasitised by small flies (*Cryptochaetum iceryae*).

A number of Miridae occur in the region. Although their feeding behaviour is variable, some species are phytophagous and may be pests of crops such as apples, lucerne and stone fruits (e.g. green mirid, *Creontiades dilutus*), but many are beneficial as predators of small soft-bodied insects or insect eggs.

Rutherglen bug (*Nysius vinitor*) (Lygaeidae) is found in all Australian states. It has a broad host range including sunflowers, potatoes, stone and pome fruits, young citrus, grapevines, strawberries, crucifers, grain sorghum, linseed, and a range of weeds as well as native plants. Both nymphs and adults feed by piercing and sucking sap from young tissues that may cause wilting, and blackening of young buds.

Another native insect that has made the transition from native plant hosts to crops is the crusader bug (*Mictis profana*). This insect is found in all Australian states. Native food plants include acacias and sennas. It also attacks citrus, grapes, roses and wisteria. Cultivated sennas, cassias, and wattles are frequently attacked. The closely related *Amorbus* bug causes similar wilting of the terminal growing tips, but this species, although similar in size to the crusader bug, lacks the distinct pale 'cross' on the hemelytra (fore wings) and is restricted to *Eucalyptus*.

Among the commonly found Pyrrhocoridae of the region is the harlequin bug (*Dindymus versicolor*) which may attack a wide range of crop and ornamental plants. The spined citrus bug (*Biprorulus bibax*) is an invader from the eastern states. Both adults and nymphs cause damage to citrus by piercing and sucking with their mouthparts. The result of this damage is a brown stain and gumming on the fruit rind, and premature fruit drop in young fruit or internal damage in mature fruit.

Of the blowflies (Calliphoridae), the hairymaggot blowfly (*Chrysomya rufifacies*) is common, and can cause secondary strike of sheep after initiation by *L. cuprina*, although it generally breeds in animal carcasses. Other blowflies include the lesser brown blowfly (*Calliphora augur*) and the eastern goldenhaired blowfly (*C. stygia*).

Among the beetles, the spinetailed weevil (*Desiantha caudata*) (Curculionidae), can be found attacking grasses, including cereals and subterranean clover, and has been reported on other plants including peas and fruit trees.

False wireworms and wireworms from the beetle families Elateridae and Tenebrionidae may be found attacking a wide range of grasses including cereals, as well as other plants including oil seed crops, grain legumes and vegetables.

Various cockchafers (Scarabaeidae) including the blackheaded pasture cockchafer (*Aphodius tasmaniae*), redheaded pasture cockchafer (*Adoryphorus couloni*), and the honey beetle (*Heteronyx flavus*), are all commonplace in the region. The blackheaded pasture cockchafer can be found in pastures and lawns, including bowling greens and golf courses. Dung of horses, cattle and sheep is the preferred food of adults and larvae. It has also been recorded as attacking wheat, where most damage is caused by the third instar larvae. Their feeding can seriously reduce the quantity of available feed during winter and spring. The larvae forage at night culling off pasture plants near ground level and placing pieces within their tunnels. Foraging occurs whilst the soil surface is moist following rain showers. With the redheaded pasture cockchafer, feeding on the roots of pasture plants results in roots being chopped off just below the soil surface. Grazing stock feeding on these plants simply uproot them. Various birds (e.g. ibis) feeding on the larvae will also uproot the pasture.

Several moths of the family Noctuidae are significant pests in the region. The native budworm (*Helicoverpa punctigera*) can destroy flowers, buds and fruit of tomatoes, maize, lupins, sorghum, beans, crucifers, sunflowers, clovers, medics, and many other field crops, pastures and vegetables, as well as many weeds and garden flowers (Fig. 8). The grapevine moth (*Phalaenoides glycine*) can be found attacking grapes, glory vine, fuchsias, Virginia creeper and some native plants. The build-up of the grapevine moth has been satisfactorily checked by predators on many properties in South Australia. The most common of these is the spined predatory shield bug (*Oechalia schellenbergii*) which spears the caterpillar with its proboscis and sucks out the body fluids. Other noctuids that are important include various armyworms, cutworms and loopers.

The lightbrown apple moth (*Epiphyas postvittana*) (Tortricidae) (Fig. 9) can be found attacking almost all cultivated fruit crops, herbs, ornamentals, *Acacia* and most legumes, as well as a wide range of weeds, including capeweed and docks. The caterpillars are usually pale green with a brown head and grow to 12-20 mm long, moving actively over the host plant and sometimes hanging by silken threads. They tunnel into the undersides of the leaves under a cover of fine webbing or web together leaves and buds whilst eating them away. They may also attack fruits that are in contact with each other or with leaves. When disturbed the caterpillars wriggle actively backwards or forwards out of their shelters. Caterpillars skeletonise the foliage, removing irregular areas to leave a ragged

appearance. Leaf margins are often rolled over. Damage to fruit is usually superficial but can provide an entry point for rotting organisms. Damage usually occurs where two fruits touch, where a leaf has been pulled down over a fruit or in the cavity around the stalk. The larvae always feed under shelter.



**Figures 8-9.** 8 (left) native budworm (*Helicoverpa punctigera*) larvae feeding on lupins; 9 (right) lightbrown apple moth (*Epiphyas postvittana*) (Photographs: SARDI Entomology).

The grapevine hawk moth (*Hippotion celerio*) and the vine hawk moth (*Theretra oldenlandiae*) (Sphingidae) also occur widely in the region, and are occasional pests of grapes. Other sphingid moths are occasionally reported from other plant species. The caterpillars consume the whole leaf and, if present in high numbers, may quickly defoliate the vines.

The larvae of the small citrus butterfly or dainty swallowtail (*Papilio anactus*) (Papilionidae) can be found attacking new foliage growth of citrus and many other plants in the family Rutaceae. The large citrus butterfly (*Papilio aegus aegus*) which is generally confined to New South Wales and Queensland, has been intermittently observed in the region. For example, in 1973-74, when significantly higher rainfall and more humid conditions were recorded, many butterflies were reported from the Upper Murray, Adelaide Plains and Mount Lofty Ranges, but the butterfly has not become established in South Australia.

#### *Native insects in natural habitats*

Apart from the aquatic/semi-aquatic insects described in Chapter 10, the Riverland and Murraylands have an enormous diversity of native insects which are mostly restricted to natural habitats. Given the very large numbers of taxa involved, we have not attempted to cover all orders, but have provided examples to the most common groups.

#### *Minor orders*

Collembola, Diplura and Protura, although not insects, are all hexapods, that is, they have three pairs of legs. Of these, Collembola (springtails) are the most commonly encountered in soil and leaf litter (Greenslade & Greenslade 1989). Most species of springtails feed on decaying vegetable matter, although fungi, algae and lichens are also an important food source for many species. Some species feed on decaying animal matter, such as earthworms, dead flies or other springtails. One common springtail is the purplish coloured *Hypogastrura* spp. which is often seen in large numbers on the surface of pools after rain.

Apart from the introduced cockroaches detailed above, native species include the brightly coloured *Ellipsidion australe* (Blattidae), often seen on flowers, and *Laxta granicollis* and *Calolampra irrorata* (Blaberidae), both of which can be found under bark and in leaf litter. In the latter two species, it is only the males that are winged.

Among the commonly encountered mantids (Mantodea) are *Orthodera ministralis* (green or garden mantid), *Paraoxyphilus tasmaniensis* (Amorphoscelidae) and *Archimantis* spp. (large brown mantids). All three have broad distributions outside of the Riverland and Murraylands.

Several earwig species (Dermaptera) are found in the region. The common brown earwig (*Labidura truncata*) occurs, especially in sandy habitats.

Apart from the locusts and grasshoppers (Orthoptera) detailed above, many other katydids, crickets, grasshoppers and locusts occur in the region (see, for example, Rentz 2003). Conspicuous among these are the inland katydid (*Caedicia simplex*) and the yellow winged locust (*Gastrimargus*



*musicus*). In a recent survey of Calperum Station, three new species of raspy crickets (Gryllacrididae) were collected (Anon. 2007).

Thysanoptera (thrips) are small insects which can be leaf-, flower-, pollen-, and fungalfeeding, although some are carnivorous on small arthropods. Apart from the pest thrips mentioned above, there are many other species from the region including the giant thrips (*Idolothrips spectrum*) and *Haplothrips victoriensis*, a predatory thrips.

Although several families of Neuroptera occur in the region, the most commonly seen are Hemerobiidae (brown lacewings), Chrysopidae (green lacewings) and Myrmeleontidae (antlions). The brown lacewing (*Micromus tasmaniae*), is very common; it feeds on aphids on cereal, pastures and vegetable crops. Green lacewings lay their eggs on stalks for protection, and all life-stages are common. Their larvae are generalist predators, especially of hemipterans such as aphids and scales, and many species cover themselves with debris (including the exoskeletons of their prey). Antlions are mostly ground dwelling, and the larvae of some species construct conical pits as traps for their prey. These pits can often be found in areas protected from rain, such as under raised logs, under cliff ledges along the Murray River, or under or around buildings (Fig. 10). Green lacewing and antlion adults are often attracted to lights.

#### *Hemiptera (bugs, leafhoppers, cicadas, aphids, scale insects, lerps)*

The most characteristic feature of the Hemiptera is the structure of the mouthparts which are adapted to extract the liquid contents of plants or animal prey by suction. They are mostly terrestrial and phytophagous (plant feeding), although the suborder Heteroptera includes groups which are carnivorous (e.g. assassin bugs, Reduviidae), and others which show varying degrees of adaptation to aquatic environments (see also Chapter 10).



**Figures 10-11.** **10** (left) Ant lion pits, Blanchetown SA (Photograph: Gary Taylor); **11** (right) Lerp of *Glycaspis brimblecombei* (Photograph: John Jennings).

Psylloidea (jumping plant lice, lerp insects) are widespread and common in the Riverland and Murraylands. Among these is the white lace lerp (*Cardiaspina albitextura*), which develops regular outbreaks on the river red gum, *E. camaldulensis* (Morgan 1984). Similar outbreaks by the closely related *C. densitexta* occur on pink gum, *E. fasciculosa*, in the South-East, upper South-East and Murraylands. Other species found on river red gums include *Glycaspis brimblecombei*, which produces edible, sugary conical lerps (Fig. 11), *Creiis corniculatus* which forms inedible horn-shaped lerps and *C. costatus* which forms distinctly flattened, white shell-shaped lerps. There are many species that do not produce lerps, most commonly *Acizzia* spp. on *Acacia* and *Cassia*, and *Trioza* spp. on false sandalwood (*Myoporum platycarpum*) and *Casuarina pauper* and various species of *Allocasuarina*. The free-living mistletoe psyllids *Acizzia loranthaceae* and *A. amyemae* feed on the mistletoe *Amyema pendulum* which commonly infests *E. camaldulensis* and black box (*E. largiflorens*) in the region (Taylor 1999).

Common eriococcids (Eriococcidae) found in the region include the gumtree scale (*Eriococcus coriaceus*) which occurs on a range of eucalypts, and *Cylindrococcus spiniferus* which causes galls on *Allocasuarina* spp. Several species of *Apiomorpha*, in which the females form spectacular species-specific galls on eucalypts, occur. *Apiomorpha karschi* is often found on *E. camaldulensis* (Gullan 1984). The woody female gall occurs on stems, with mature galls often aggregating to form large woody swellings which can easily exceed 10 cm in diameter, whereas the small tubular male galls (up to 4.2 mm long) are mostly found massed together on the upper surface of leaves. Other common species include *A. calycina*, in which the female gall is somewhat

cylindrical with a truncate apex. It occurs on *E. dumosa* (white mallee), *E. gracilis* (yorrell), and *E. oleosa* (red mallee) in the region. The female of *A. malleecola* forms an ovoid gall which can be found on *E. dumosa*, *E. oleosa*, and *E. socialis* (red mallee). The female gall of *A. munita* is a distinctive four-sided gall usually with long extensions at each corner of the gall (Gullan 1984), and occurs on a wide range of eucalypt species. The female gall of *A. strombylosa* has a rough surface covered with small protuberances and resembles the seed cones of *Allocasuarina/Casuarina*. As the species name suggests, the female galls of *A. urnalis* are urn-shaped, and in the region occur on *E. dumosa*, *E. gracilis* and *E. largiflorens* (black box).

Whilst many scale insects occur, the armoured scales (Diaspididae) are perhaps more commonly observed. They include *Maskellia globosa* which is unusual in that it is a gall-former and does not produce an external scale. The male galls are small and horn-like and occur either on leaves or associated with the larger rounded female galls on twigs of various eucalypts.

Although cicadas (Cicadidae) are predominantly tropical/subtropical, a number of species have distributions which extend into the Riverland and Murraylands. These include the eastern sandgrinder (*Arenopsaltria nubivena*), the black squeaker (*Pauropsalta encastica*), and the cherrynose (*Macrotristria angularis*) (Moulds 1990). Adults of the latter species have been collected on *Callitris* and *Allocasuarina/Casuarina*. An interesting species is *Marteena rubricanta* which is known from only three disjunct areas; Ravensthorpe WA, Morgan SA and Eurabalong western NSW (Moulds 1990). The adults, and possibly the nymphs, appear to be associated with mallee species. Several *Cicadetta* spp., including *C. adelaida*, can also be found along the Murray River.

A number of Cicadellidae (leafhoppers) are particularly characteristic of the mallee areas of South Australia (as well as WA and western NSW) (Fletcher pers. comm.; Fletcher & Larivière 2008). The Subfamily Austroagalloidinae is endemic to Australia, and comprises eight described species, all on *Eucalyptus*, with two endemic to the Karoonda district. Deltocephalinae: Goniagnathini are arid-zone adapted species, with two of the five Australian species described from Keith. Several ant-attended treehoppers (Eurymelinae: Ipoini) have also been found in the region (Fletcher pers. comm.): *Ipelloides macleayi* from Tailem Bend – this species seems to be endemic to South Australia; *Ipoidea laeta* from Lyrup Flats Reserve, west of Renmark, also been found in western NSW, western Qld and NT; and, *Anacornutipoa lignosa* from Waikerie – also found in WA and eastern Australia. Two species of treehoppers which live in ants' nests (Eurymelinae: Pogonoscopini) are also known from the region. Of the Membracidae (tree hoppers), a number of species, including *Sextius* spp., are associated with *Acacia*. *Rigula calperum* and *Undarella pulleni* were both described from Calperum Station.

Among the Suborder Heteroptera from the region is the widespread Pacific damsel bug (*Nabis kinbergii*) (Nabidae). Eggs are oviposited singly into grass stems and nymphs and adults feed on soft bodied insects, moth eggs, small larvae and mites.

The adults and nymphs of assassin bugs (Reduviidae) are predacious, almost entirely on other arthropods. They are usually solitary insects and are often conspicuous, sitting near the tips of shoots waiting for other insects to come within reach. Most are regarded as beneficial insects, for example *Coranus trabeatus*, a ferocious reduviid which feeds on a wide range of insect pests. Larger species are capable of inflicting a painful 'bite' and may eject a harmful defensive fluid if handled.

The Lygaeidae (cinch bugs) is a large family with most species being seed-feeders. They are usually brown to red with a hard, elongated body ranging from 4-20 mm in length. The coon bug (*Oxycarenus arctatus*), in which the nymphs are small (~3 mm long), black and with blood-red abdomens, is common, and can be locally abundant on fences, at the base of walls and on areas such as ovals.

Coreidae are large bugs with strong repellent odours. Several species are found in the region, including the crusader bug (*M. profana*, see above) on *Acacia*, *Senna* and *Eucalyptus*.





**Figure 12.** Nymph of *Poecilometis* sp. (Photograph: John Jennings).

Many different shield or stink bugs (Pentatomoidae) occur in the region. Scutelleridae include *Austrotichus rugosus*, which has been found in leaf litter at Keith, and *Solenotichus circuliferus* from Morgan and Loxton (Gross 1975). A number of *Poecilometis* spp. (Fig. 12), often found under peeling bark of *Eucalyptus* or on *Callitris* (Gross 1976), also occur. *Poecilometis alienus* has been collected from Morgan, Tailem Bend, and Karoonda to Peebinga, *P. stigatus* from along the Murray River, *P. parilis* from Sinclair Flat, upstream of Lock 1, and *P. vermiculatus* from Mercunda and Mannum (Gross 1976). Several other genera occur in the region (see Gross 1975, 1976), often associated with the common trees of the region such as *Eucalyptus*, *Callitris* and *Allocasuarina/Casuarina*.

#### *Coleoptera* (beetles)

Beetle genera are detailed in the series ‘A Guide to the Genera of Beetles of South Australia’ (Matthews 1980-1997; Matthews & Reid 2002).

Nearly all members of the Suborder Adephaga are nocturnal predators or scavengers, and several significant families occur in the region. Amongst the predatory Carabidae is the green tiger beetle (*Megacephala australis*), which can be locally abundant and hunts prey at dawn and dusk (crepuscular) on the banks of the Murray River. Also common in the region is the iridescent green carab beetle (*Calosoma schayeri*), a large, active species which hunts lepidopteran caterpillars and other slow-moving prey. It flies at night and is attracted to lights. These beetles can give off an unpleasant smell if handled. Fox scats are sometimes composed almost entirely of the remains of these beetles. Many other carabids (ground beetles) can be found, particularly in mallee vegetation. Also, adults of the aquatic families Dytiscidae (predaceous water beetles) and Gyrinidae (whirligig beetles) are often attracted to lights at night (see also Chapter 10). Watts (2002) provided an identification guide to the genera of Australian water beetles.

About 90% of extant beetles belong to the Suborder Polyphaga, and these proportions seem to be exhibited in the beetle fauna of the Riverland and Murraylands. Several families of Staphylinidae are found in the region, especially Staphylinidae (rove beetles). They are mostly predators or carrion feeders living in soil and leaf litter. Most are narrow elongate beetles with the posterior half of their abdomen exposed. As well, the carrion beetle (*Ptomaphila lacrymosa*) (Silphidae), is also common, and is most active in autumn and winter months (Archer 2003).

Of the Scarabaeoidea, the families Trogidae, Lucanidae and Scarabaeidae are the most commonly encountered. Several species of *Trox* (Trogidae) are known from the region. Larvae burrow in the vicinity of carcasses, and both larvae and adults feed mainly on skin, fur and feathers. The larvae of lucanids (stag beetles) feed on dead wood, with the golden stag beetle (*Lamprima aurata*) being occasionally collected from *Leptospermum*, *Acacia*, *Eucalyptus* and sometimes from exotic tree species. Scarabs, however, are far more common and abundant than the other families. Geotrupinae are commonly seen in sandy areas after rain, especially *Blackburnium* spp. They feed on buried humus and underground fungi, and adults stridulate when caught. Aphodiinae, including the pasture cockchafer (*Aphodius tasmaniae*) (see above), are mostly dung-feeding beetles. The five horned dung beetle (*Onthophagus pentacanthus*) is common throughout much of the Murray-Darling Basin and further west in South Australia. Adults construct burrows for their young and provision them with animal dung. Adults are nocturnal and often attracted to lights. Several dung beetles which were introduced to control mainly cattle dung also occur in the region (Tyndale-Biscoe 1990 – see above). Melolonthinae (chafers) have larvae which occur in the soil and feed on roots and humus, whilst adults are mostly short-lived and either do not feed at all, or feed on foliage. Of the many genera occurring in the region, the speciose *Colpochila*, *Heteronyx* and *Liparetrus* are associated with

*Eucalyptus*. Christmas beetles (*Anoplognathus* spp.) can be seen flying in summer. Adults feed on *Eucalyptus* leaves and the larvae feed on rotting wood, humus or grass roots.

Buprestids or jewel beetles (Buprestidae) are widespread throughout the region. Some of the more colourful jewel beetles include *Diadoxus*, whose larvae bore into *Callitris*, *Julidomorpha bakewelli* which can occasionally be encountered in late December in the mallee (Matthews 1985), and *Castiarina* spp. which are mostly collected on flowers of plants such as *Leptospermum* (Barker 2006).

Clerids (Cleridae), many of which are elongate, hairy beetles with distinctive colour patterns, are predacious as both larvae and adults (Matthews 1992). Depending on the species, these beetles can often be collected from flowers (especially *Eucalyptus*, *Bursaria* and *Leptospermum*), or by beating or sweeping vegetation.

Coccinellidae (ladybird beetles) are easily recognised due to their characteristic oval shape and usually bright, often spotted colour pattern. They range in size from 1 to 15 mm in length and have short antennae and legs, which are usually hidden beneath the body when viewed from above (Slipiński 2007). The larvae are soft-bodied, and are often variously coloured with spots, and are usually adorned with spines. Almost all ladybirds are predatory as both larvae and adults, and feed on a variety of small soft-bodied arthropods such as aphids, scales and mites. As such, most ladybirds are viewed as beneficial insects. Apart from *Rodolia cardinalis* (see above), important ladybirds in the region include the transverse ladybird (*Coccinella transversalis*) and common spotted ladybird (*Harmonia conformis*), with adults and larvae of both species being predators of soft-bodied insects. The mite-eating ladybird (*Stethorus* spp.) are also common but often overlooked because of its very small size. In contrast, both larvae and adults of the twentyeight-spotted potato ladybird (*Henosepilachna vigintiseipunctata*) feed on plants such as potato.

The Tenebrionoidea comprises some 28 families occurring in South Australia (Matthews 1987). They are predominantly associated with dead plant matter and the fungi that grow on it, although larvae of Rhipiphoridae and Meloidae are parasitic on other insects. Adults of several families (e.g. Mordellidae and Meloidae) are often collected from flowers. Of the Tenebrionidae, *Lagria grandis* is a common and widespread species with adults found feeding on *Eucalyptus* leaves. *Celibe* spp. and *Helea* spp. (pie-dish beetles) are found in leaf litter, especially in the mallee. *Chalcopteroides* spp. (darkling beetles) are conspicuous blue, green or purple metallic tenebrionids, mainly associated with eucalypts.

The Chrysomeloidea comprises two families, Cerambycidae and Chrysomelidae, both of which are speciose in the Riverland and Murraylands. Numerous longicorn beetles (Cerambycidae) are common, for example, *Phoracantha* spp. which produce oval emergence holes in eucalypts such as the sugar gum. Adults of a number of genera are often seen at lights. These include *Eurynassa* spp. whose larvae bore into the dead wood of *Acacia*. Matthews (1997) has an extensive list of longicorn beetle genera and their host plants in South Australia. Most Chrysomelidae (leaf beetles) are leaf-feeding as both larvae and adults, although larvae may exhibit other habits such as stem or root boring and seed feeding. Matthews and Reid (2002) list chrysomelid beetle genera and their host plants in the state. Amongst the common species are *Paropsis* and *Paropsisterna* which feed mostly on eucalypts.

Many species of weevils (Curculionidae) are found in the region. Common weevils include the large and spectacular diamond beetle (*Chrysolopus spectabilis*) which feeds on various acacias at locations such as Monarto South, elephant weevil (*Orthorhinus cylindrirostris*), Eucalyptus snout weevils (*Gonipterus* spp.), and several species of curculio beetles (*Otiorynchus*), some of which are considered pest species. Amongst the smallest weevils are *Misophrice* spp. which are about 2 mm long and found on *Allocasuarina/Casuarina* where they are believed to be pollen feeders.

#### *Diptera (mosquitoes, midges, flies)*

Apart from the mosquitoes (Culicidae), midges (Chironomidae) and other aquatic flies treated in Chapter 10, many other families and species are found in the region. Flies are distinguished by having a single pair of wings on the thorax and a pair of halteres or balancing organs (tiny reduced hind wings).

Tabanidae (March flies and horse flies) are medium to large biting flies, and several species are known from the region. Larvae are carnivorous, preying on insects and snails, and may be found living in damp soil or rotting logs, as well as in aquatic environments. Adults of most Australian

species are blood-suckers, but some are flower-feeders (e.g. *Scaptia*). The blood-suckers are voracious feeders, often hovering before landing and piercing the skin with their needle-like proboscis and absorbing a considerable amount of blood.

Robber flies (Asilidae) are a widely distributed group of predatory flies which are diverse in semi-arid and arid regions. Adult robber flies prey on insects, although some capture spiders. Several species of *Bathypogon*, *Cerdistus*, *Colepia* and *Mauropteron* are known from the region (Lavigne 2003).

Beeflies (Bombyliidae) and hover flies (Syrphidae) are common and widespread in the region. Amongst the more colourful beeflies (Exoprosopini) are *Balaana centrosa*, *Kapua adelaidica* and *K. irwini* collected near Calperum Station, which have distinct clear and black wing patterns (Lambkin et al. 2003). Hover flies of the region include members of the common genera *Simosyrphus* and *Eristalis*, the larvae of which prey on soft-bodied insects.

Tachinid flies (Tachinidae) can be small to large with wingspans up to 35 mm depending on the species. Most have stout bodies covered in bristles and are either grey or black, although some have a metallic appearance and, as a consequence, superficially resemble blowflies. The larvae of tachinids are parasitoids of other insects.

#### *Lepidoptera (moths and butterflies)*

Many of the moths listed for the Adelaide Region in McQuillan and Forrest (1985) occur in the Riverland and Murraylands, and here are highlighted just some of the more common species. In autumn, various ghost moths (Hepialidae) can be seen, including the Bardi (Barti) moth (*Trictena argentata*) (Fig. 13), which is common along the banks of the Murray River at Waikerie when they emerge from their pupal cases in the ground near trees. Larvae, pupae and adults were eaten by Aboriginals (see Chapter 6).

The giant wood moth (*Xyleutes liturata*) (Cossidae), is the largest moth in South Australia with a wingspan of up to 200 mm in females and 130 mm in males. Its larvae bore into the woody tissue of eucalypts, particularly the river red gum. Several species of case moth (Psychidae) occur in the region, including the faggot case moth (*Clania ignobilis*) whose older larvae form distinctive larval cases surrounded by twigs of their eucalypt food-plant.



**Figure 13.** *Trictena argentata* (Photograph: Claudia Taylor).

An interesting moth occurring in the region is *Stathmopoda melanochra* (Stathmopodidae), which deposits eggs amongst colonies of gum tree scales (e.g. *Eriococcus coriaceus*) where the larvae devour them. A number of geometrid moths are common including the red-lined geometrid (*Crypsiophona ocutaria*), angled satin moth (*Thalaina angulosa*) and hakea moth (*Oenochroma vinaria*). The gumleaf skeletonizer (*Uraba lugens*), often found on river red gum, is interesting in that the caterpillars have a characteristic 'spike' on top of their head, comprising old head capsules which are shed as they grow. One noctuid, the granny moth (*Dasypodia selenophora*), whose larvae feed on *Acacia*, is commonly seen in late spring resting during daylight hours under eaves or on ceilings.

Butterflies can generally be distinguished from moths by a number of characters, including their day-flying habits, bright colours, presence of a clubbed antenna and the ability to close their wings dorsally over the thorax (Braby 2004). However, there are exceptions! Butterflies belong to two superfamilies, Hesperioidea and Papilionoidea (several families), with most Australian species belonging to the latter. Braby (2004) covers the described butterflies of the region, providing descriptions, habits and larval food plants. As well, South Australian butterflies

are extensively treated by Grund (2008). Several skippers (Hesperiidae) can be found in the Riverland and Murraylands, including the sciron ochre or mallee ochre (*Trapezites sciron*), whose males can be observed hilltopping on sand dune ridges. In addition to the small citrus butterfly (*P. anactus*) and occasionally the large citrus butterfly (*P. aegus aegus*) (see above), the only other Papilionidae seen in the region is the chequered swallowtail (*P. demoleus*). Pieridae (whites and yellows) include the cabbage white butterfly (see above), the white migrant (*Catopsilia pyranthe*), which occasionally migrates into the region but does not breed, the common small grass yellow (*Eurema smilax*) (Fig. 14), and the spotted jezebel (*Delias aganippe*).

Nymphalidae are medium to large butterflies and are commonly brown or orange. The common brown (*Heteronympha merope*) is widespread in the region and its larval food plants include *Poa* spp. and kangaroo grass (*Themeda trianda*) (Braby 2004). The meadow argus (*Junonia villida*), Australian painted lady (*Vanessa kershawi*), yellow admiral (*V. itea*), lesser admiral (*Danaus chrysippus*), and monarch or wanderer butterfly (*D. plexippus*), are all widespread and common.



**Figures 14-15.** 14 (left), small grass yellow, *Eurema smilax*; 15 (right), inland form of the amaryllis azure (*Ogyris amaryllis meridionalis*,) female (Photographs: Roger Grund - <http://users.sa.chariot.net.au/~rbg/>).

Lycaenidae (blues and azures) are small to medium-sized butterflies, often with wings which have an iridescent blue, purple or green upper and brown or white patterned underside of their wings. Numerous species occur in the region. Perhaps the most often encountered is the common grass-blue (*Zizina labradus*) which occurs throughout Australia. The larvae eat the flowers, leaves and immature seeds within developing seed pods of a wide variety of native and introduced legumes, including lucerne. Other species include the amaryllis azure (*Ogyris amaryllis meridionalis*) (Fig. 15), whose larvae feed on various mistletoes (*Amyema* spp.) growing on *Acacia* and *Casuarina/Allocasuarina*. They can be attended by a range of ant species, including *Iridomyrmex* and *Camponotus*. The amethyst hairstreak (*Jalmenus icilius*) also occurs in the region. Its larvae feed on *Acacia* and *Senna* and are attended by *Iridomyrmex*. Other blues include the two-spotted lined blue (*Nacaduba biocellata*) whose larvae live on flower buds and flowers of *Acacia*, wattle blue (*Theclinesstes miskini*) whose larvae feed mainly on *Acacia*, saltbush blue (*T. serpentata*) whose larvae feed on chenopods such as *Atriplex* and *Rhagodia*, and the long-tailed pea-blue (*Lampides boeticus*) whose larvae feed on Fabaceae.

#### *Hymenoptera* (sawflies, wasps, ants and bees)

As with other megadiverse orders (Coleoptera, Diptera Lepidoptera and Hemiptera), several thousand species of Hymenoptera occur in the region. Many of the parasitic wasps are 2 mm or less in length, and are difficult to observe and identify even to family level without the aid of a stereo-microscope and specialist taxonomic keys (see Stevens et al. 2007). The order is divided traditionally into Symphyta and Apocrita, with the former being the ‘primitive’ sawflies. Apocritans include wasps, bees and ants, and are distinguished from the Symphyta by the narrow ‘waist’ (petiole) formed between the first two segments of the actual abdomen (Stevens et al. 2007).

Of the sawfly families (symphytans), Pergidae is the most common, with members of the genus *Perga* being the most abundant. Their larvae or ‘spitfires’ (Fig. 16), so called because the aggregating larvae rear up when disturbed and regurgitate eucalypt oils, are common on *Eucalyptus*. Adult pergids are, however, rarely seen. Leaf blister sawflies (*Phylacteophaga*) are also common, and mine the young leaves of river red gum and other eucalypts.



Among the apocritan wasps, two families belonging to the Superfamily Evanioidea (Evaniidae and Gasteruptiidae) are widespread in the Riverland and Murraylands. Evaniidae (hatchet or ensign wasps) parasitise cockroach oothecae and adults of several species of *Acanthinevania* and *Szepligetella* can be collected on various flowers, including eucalypts, *Melaleuca* and *Bursaria*, as adults use nectar as an energy source. The larvae of Gasteruptiidae occur in the nests of solitary bees and wasps where they kill the host egg or larvae and then consume the pollen store provided by the host for its developing larvae (i.e. predator-inquilines). As with evaniid wasps, adult gasteruptiids can be found on flowers with an abundant nectar supply or can be seen searching for host nests on tree trunks, banks or ground (Fig. 17). One gasteruptiid, *Pseudofoenus morganensis*, has only been collected near Morgan (Jennings & Austin 2002).



**Figures 16-17.** **16** (left) ‘Spitfires’ of *Perga* sp. (Photograph: John Jennings); **17** (right) *Gasteruption* sp., female (Photograph: G. Weber).

Ichneumonid wasps (Ichneumonidae) can be small to very large and are commonly seen at flowers although some species are attracted to light, including *Enicospilus* spp., which are ectoparasitoids of large lepidopteran larvae or symphytan wasps. Whilst many ichneumonids parasitise lepidopteran larvae, others parasitise Hymenoptera, Neuroptera and Coleoptera. Braconids (Braconidae) are minute to large wasps, and amongst the genera in the region are *Apanteles*, *Chelonus*, *Cotesia*, and *Bassus* (parasitoids of moth larvae), *Jarra* (parasitoids of beetle larvae), *Aspilota* and *Opius* (parasitoids of dipteran larvae) and *Aphidius* (parasitoids of aphids).

Platygastrid wasps (Platygastroidea) include species which parasitise Cecidomyiidae (Diptera) leaf and stem galls on various plants including *Eucalyptus* and *Acacia*. Amongst the Scelionidae, is the genus *Scelio*, which are parasitoids of locust and grasshopper eggs. Several species are known from the region, including *S. fulgidus* (Murray Bridge - bred from eggs of the Australian plague locust), *S. fulvithorax* (Renmark and Bookmark Biosphere Reserve), *S. imporcerus* (near Keith), and *S. parvicornis* (Waikerie) (Dangerfield et al. 2001).

Many of the ~19 Australian families classified in the Superfamily Chalcidoidea can be found in the region. They are mostly small to minute parasitic wasps. Of these families, species of Pteromalidae have been reared from *Apiomorpha* galls (Hemiptera), Aphelinidae including *Aphytis* (parasitoids of diaspid scales including red scale on citrus) and *Aphelinus* (aphid parasitoids), and Trichogrammatidae (*Trichogramma* spp. which parasitise moth eggs, including *Helicoverpa*) are common. Several chrysidid or cuckoo wasps (Chrysididae) (Fig. 18) occur in the region. The large metallic blue *Stilbum cyanurum* parasitises mud dauber wasps (*Sceliphron*) and potter wasps (*Abispa*) and has probably been introduced.



**Figures 18-19.** **18** (left) Cuckoo wasp (Chrysididae) (Photograph: John Jennings); **19** (right) *Polistes* nest (Photograph: SARDI Entomology).

The Superfamily Vespoidea (7 families) is very well represented. Pompilidae (spider wasps) are small to large predatory wasps that all hunt spiders or, more rarely, other pompiliids. Several velvet ant species (Mutillidae) occur in the region. The wingless females, which superficially resemble large hairy ants, run around on the ground, tree trunks or walls searching for nests of Vespidae and Apidae. Female velvet ants are capable of delivering a painful sting if disturbed. Numerous species of flower wasps (Tiphiidae) also occur in the region. Most are believed to be parasitic on soil-dwelling beetle larvae, and all species have wingless females. In these species, the male often carries the female from flower to flower while mating. One obvious species is the ‘blue ant’ (*Diamma bicolor*) which parasitises mole crickets. The female is about 23 mm long, has a polished blue-green body and reddish legs, and can deliver a painful sting if disturbed. Some species of flower wasps are also implicated in the pollination of sexually deceptive orchids such as the endangered coloured spider orchid, *Caladenia colorata* (Anon. 2005). Vespid wasps (Vespidae) are commonly encountered, including potter wasps (*Abispa*) and paper wasps (*Polistes*). *Polistes* nests are common under ledges on the cliffs along the Murray River (e.g. at Blanchetown and Waikerie) and under eaves of buildings (Fig. 19).

Many genera and species of ants (Formicidae) are represented (Greenslade 1979; Shattuck 1999). Whilst the majority of ants are general predators or scavengers, feeding on a wide range of prey including other arthropods, many are seed feeders. Some also tend sap-sucking Hemiptera and other insects. Among the commonly encountered ants in the region are the meat ants (*Iridomyrmex* spp.), which have nests that can be of an enormous size with tens of thousands of workers. They clear vegetation from the surface around their nest and cover the area with small stones. Some *Iridomyrmex* are associated with butterflies, including the satin azure (*Ogyris amaryllis*). Another common genus is *Camponotus* (sugar ants). They can be seen foraging on the ground and on vegetation by day but more frequently by night. Adults derive their sustenance from the excreta of sap-sucking insects, floral nectaries and seeds. Several species are known to have a mutualistic relationship with *Ogyris* butterflies. Other common ant genera in the region include *Melophorus*, *Meranoplus* (especially in the mallee), *Myrmecia* (bulldog ants and inch ants – see Fig. 20), *Pheidole* and *Rhytidoponera*.



**Figures 20-21.** **20** (left) inch ant (*Myrmecia* sp.) (Photograph: Brett Smith); **21** (right) male bluebanded bee (*Amegilla* sp.) (Photograph: Remko Leijs). Apoidea from the region include *Sphex* (Sphecidae) which provisions its underground nest with crickets (Gryllidae) and katydids (Tettigonidae), the common mud dauber (*Sceliphron laetum*)

(Sphecidae) which provisions its mud nest with spiders, and *Bembix* (Crabronidae) which provisions its underground nests with flies or occasionally bees or other wasps. Bees all belong to the Family Apidae, and all native bees in the region are solitary (not social as is the case with the honey bee). They are commonly seen collecting nectar and/or pollen on a wide range of flowers, and include the genera *Ctenocolletes*, *Euryglossa*, *Hylaeus*, *Leioproctus* and *Nomia*. Some of the more conspicuous bees are the solitary blue-banded bees (*Amegilla* spp.) (Fig. 21), which have pale blue bands on a black abdomen. At least one species of these bees shows potential as a pollinator of glasshouse grown tomatoes (Hogendoorn et al. 2006).

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## CHAPTER 12. FISHES

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### INTRODUCTION

The Murray River is a lifeline for aquatic biodiversity within a generally dry State. Its variety of freshwater habitats including deep river channel, anabranches, floodplain/wetlands, billabongs and stream tributaries has the richest freshwater fish community of the five major biogeographical (drainage) divisions within South Australia (Hammer & Walker 2004). The region under consideration here includes the Murray River and associated habitats and stream tributaries between the SA border and its confluence with Lake Alexandrina just downstream of Wellington. Treatments for nearby associated habitats of the Eastern Mount Lofty Ranges, Lakes Alexandrina and Albert and the Coorong are provided elsewhere (Eckert & Robinson 1990; Sim et al. 2000; Wedderburn & Hammer 2003; Hammer 2004).

Overall, information for the fish fauna of the region is lacking, particularly on historic composition and abundance, but also concerning changes through time and species interactions with the local environment. While the River has attracted general interest, no detailed, systematic historical surveys were undertaken for fishes and records are patchy at best (e.g. Zeitz 1902; Waite 1923). Information on population trends over the past 50 years is limited to a few large-bodied species such as Murray cod and golden perch (callop), using commercial catches as an approximate index (Reynolds 1976a; Ye 2005; Ye & Zampatti 2007). Until recent years, there has been little documented fish ecology research specific to the South Australian Murray, and much of the information is at the moment inferred from interstate studies or anecdotal information (but see Reynolds 1983; Lloyd & Walker 1986; Lloyd 1987; Puckridge 1990; Vilizzi 1998; Vilizzi & Walker 1999; Hammer 2001; Leigh 2002; Smith & Walker 2003ab, 2004ab; Bice & Zampatti 2005; Barrett 2007; Wedderburn et al. 2007; Leigh et al. 2008; Cheshire & Ye 2008ab; Wedderburn 2008; Ye et al. 2008).

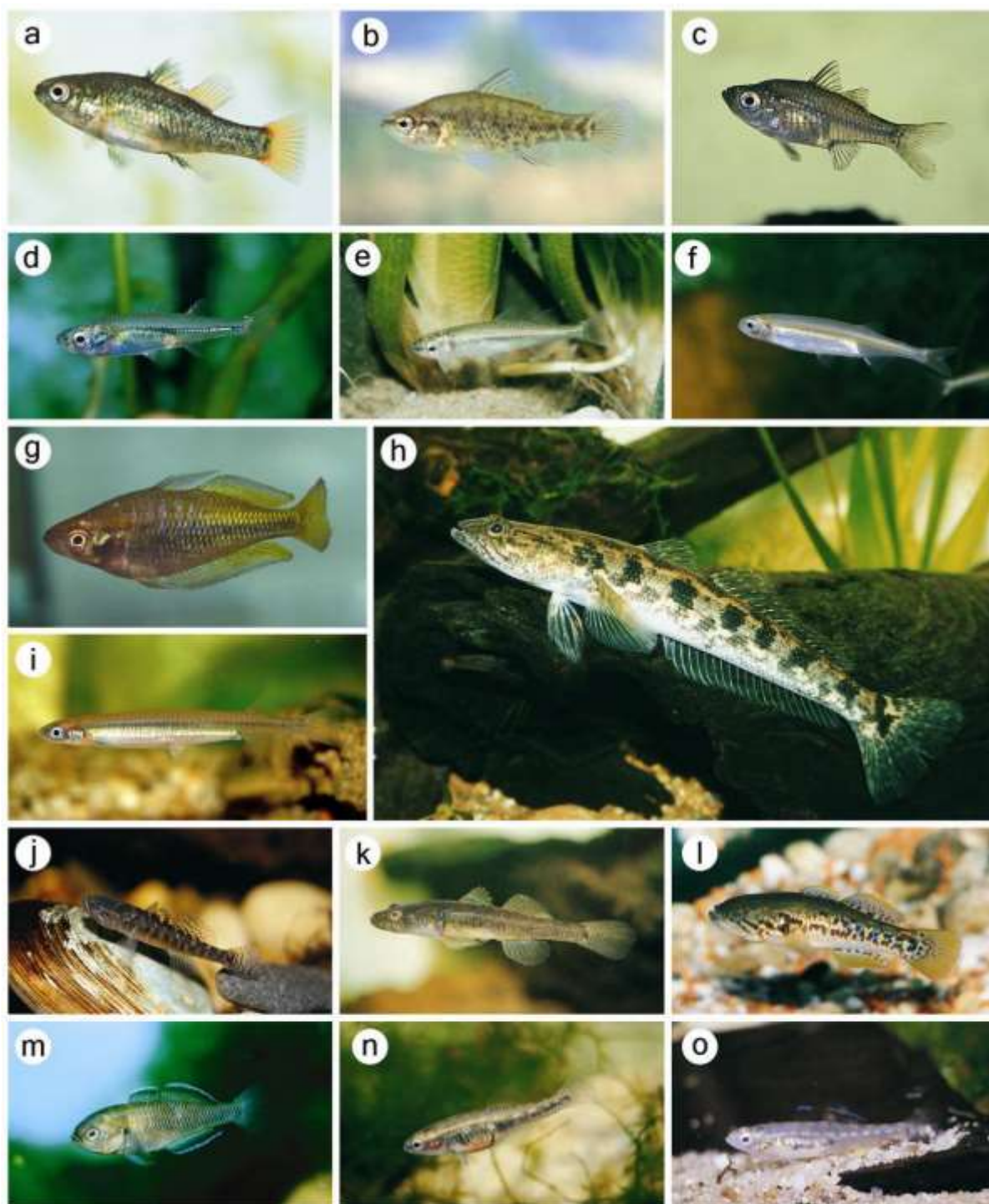
Hence there is still much to be discovered about the past and present distribution, biology and status of fishes in the region. Also, in light of highly regulated flow and major modifications to habitat (e.g. river transformed to a series of weir pools), opportunities also need to be taken to conserve remaining natural values.

### DIVERSITY OF FISHES

Representative native and exotic species are pictured in Figures 1, 2 & 3. For further taxonomic and general ecological information see: Lake 1971; Scott et al. 1974; Koehn & O'Connor 1990; McDowall 1996a; Young 2001; Allen et al. 2002; Lintermans 2007.

**Native fishes** A total of 35 native freshwater fish species representing 15 families have been recorded in the region (see Appendix, Figs 1 & 2). These fishes are predominantly temperate/subtropical species endemic to southeastern Australia and all occur more widely in the Murray-Darling Basin or nearby coastal systems; however the region forms a considerable portion of the Murray-Darling Basin range of at least seven species (see Appendix). The best-represented families are the Percichthyidae (cods, basses and pygmy perches), Eleotridae (gudgeons), Galaxiidae (galaxiids) and Atherinidae (hardyheads).

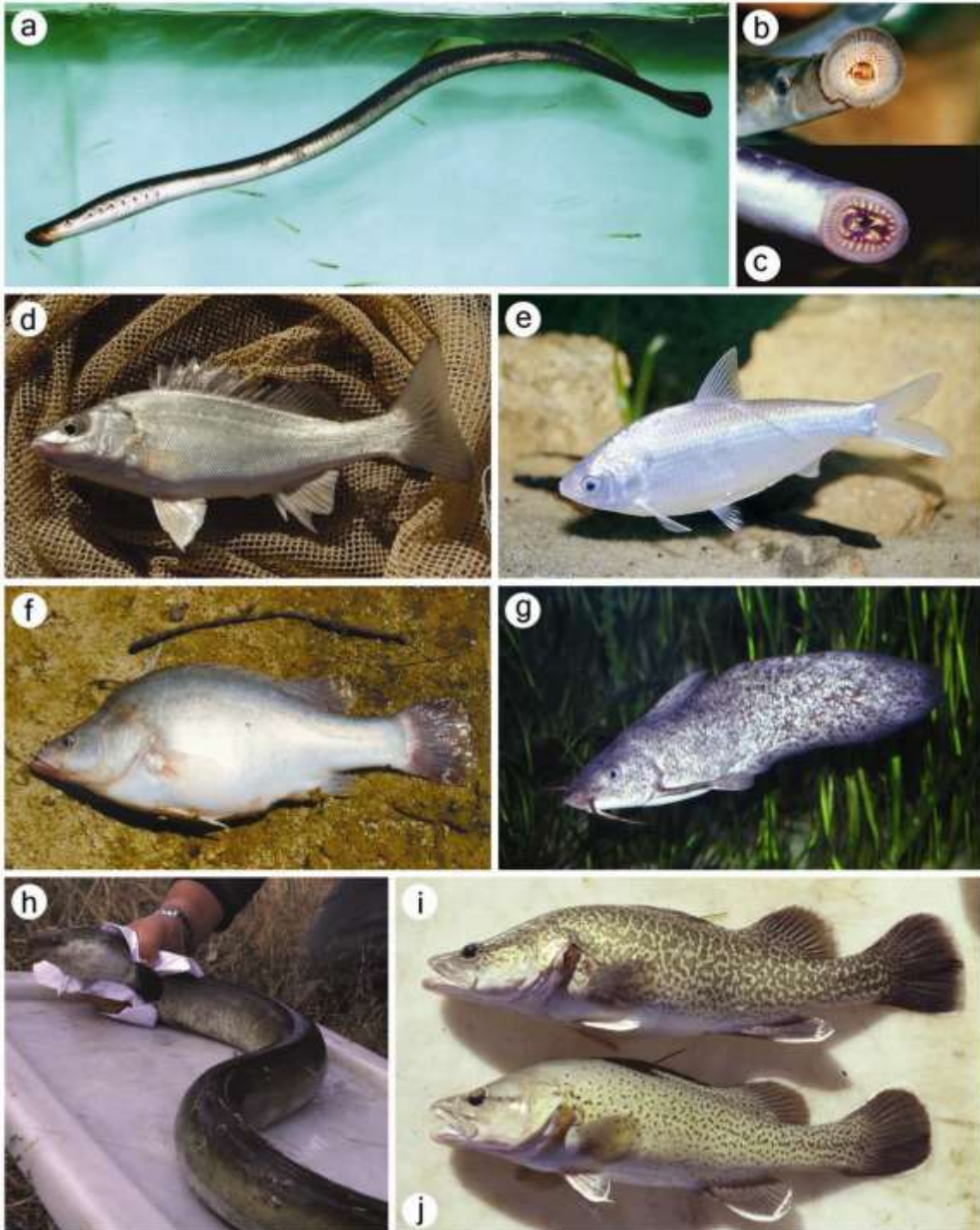
Twenty-six of the 35 listed species are exclusively freshwater forms and thus most can be expected to complete their life cycle in the region. These include the iconic Murray cod and golden perch; protected freshwater catfish, river blackfish, pygmy perches, silver perch and southern purplespotted gudgeon; historically recorded Murray galaxias, chanda perch, trout cod and Macquarie perch; and widely occurring and usually abundant bony herring, smelt and carp gudgeons. The freshwater fishes also include some occasional downstream visitors from the upper Murray-Darling Basin like the spangled perch (which as a member of the Terapontidae is more correctly known as a grunter).



**Figure 1.** Small-bodied native fishes (i.e. growing no larger than 35 cm, most < 10 cm) of the Lower River Murray, South Australia with indicative common size range: **a)** southern pygmy perch, 3-6 cm; **b)** Yarra pygmy perch, 4-6 cm; **c)** chanda perch, 3-4 cm; **d)** Murray hardyhead, 3-6 cm; **e)** unspcked hardyhead, 3-6 cm; **f)** smelt, 5-7 cm; **g)** Murray rainbowfish, 3-6 cm; **h)** congolli, 11-17 cm; **i)** common galaxias, 5-13 cm; **j)** dwarf flathead gudgeon, 2-4 cm; **k)** flathead gudgeon, 3-9 cm; **l)** southern purple-spotted gudgeon, 4-7 cm; **m)** Midgley's carp gudgeon (male), 3-4 cm; **n)** Midgley's carp gudgeon (female), 3-4 cm; **o)** western carp gudgeon (male), 3-4 cm.

There are seven diadromous species which migrate between fresh and saltwater habitats for one or more life cycle components or for the purposes of reproduction: pouched lamprey, shortheaded lamprey, shortfinned eel, climbing galaxias, common galaxias, estuary perch and congolli. The remaining three species (small-mouthed hardyhead, bluespot goby and lagoon goby) are euryhaline (broad salinity tolerance), and are extended populations from downstream lentic and estuarine conditions.





**Figure 2.** Large-bodied native fishes (i.e. potentially growing larger than 40 cm) of the Lower River Murray, South Australia with indicative common size range: **a)** pouched lamprey, 50-60 cm; **b)** oral disk of pouched lamprey; **c)** oral disk of shortheaded lamprey; **d)** silver perch, 15-40 cm; **e)** bony herring, 10-40 cm; **f)** Murray- Darling golden perch, 25-50 cm; **g)** freshwater catfish, 20-40 cm; **h)** shortfinned eel, 60-100 cm; **i)** Murray cod, 50-150 cm; **j)** trout cod, 40-100 cm.

Other essentially marine/estuarine species have not been specifically included here, although species such as black bream (*Acanthopagrus butcheri*), Australian salmon (*Arripis truttaceus*), mulloway (*Argyrosomus japonicus*), sea mullet (*Mugil cephalus*) and sandy sprat (*Hyperlophus vittatus*) have been reported upstream of Wellington on occasions (Scott et al. 1974; Smith et al. 2007; SARDI Fisheries Statistics; commercial fisher H. Jones pers. comm.). Such species would have had better access to the Lower Murray before the construction of barrages, although predominantly fresh water conditions prior to flow regulation (Sim & Muller 2004) suggest natural habitat would not have been that suitable.

The eventual number of native species from the region may be slightly higher, and our perceptions of conservation status and ecology are sure to change. Taxonomic issues exist with mountain galaxias, smelt and carp gudgeons (e.g. Bertozzi et al. 2000; Raadik 2001; Hammer et al. 2007), and probably other groups. For example prior to the 1980s carp gudgeons in the Murray- Darling Basin were commonly referred to a single common species the “western carp gudgeon”. Recent research however, indicates that at least four taxa are actually represented and that hybrids (potential unisexual lineages) between three of these are common (Bertozzi et al. 2000). Preliminary observations suggest subtle biological and ecological differences between different taxa (Unmack 2000).

### **Alien species**

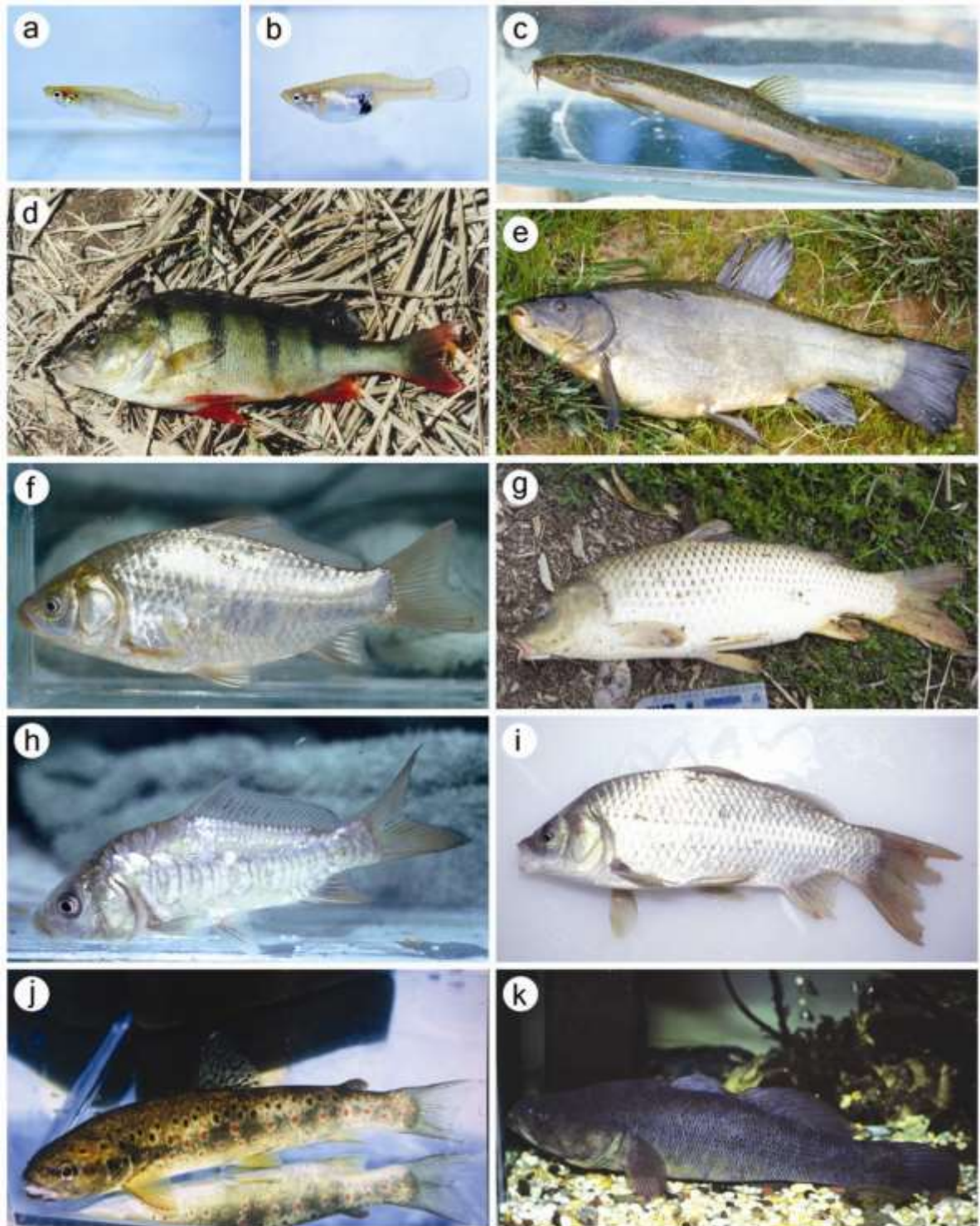
Alien species include species exotic to Australia and Australian native species translocated outside their natural range. Eleven alien species representing seven families have so far been recorded in the region (Appendix). It is possible that more species have been introduced (e.g. via bait bucket translocations, discarded aquarium specimens, other stockings: Lintermans 2004) but captures or established populations have not been reported.

Of the exotic species, common carp have been in Australia for over a century, and have spread widely throughout the Murray-Darling Basin since the release of the “Boolara strain” near Mildura in 1968 (Brown 1996). Rapid proliferation and range expansion of carp occurred during floods in the early 1970s (the first record in South Australia was in 1970: Reynolds 1976a) and they often now dominate the biomass of inland fish communities (Gehrke et al. 1995) including the Lower Murray (Davies et al. 2008). Carp have been implicated in causing damage to riverbanks and aquatic macrophytes, and competing with native fish for food and habitat (Fletcher et al. 1985; Koehn et al. 2000, Sim et al. 2000). The related goldfish was an early introduction (also known as golden carp) and remains common, with occasional hybrids between carp and goldfish occurring. A third cyprinid, tench, was common prior to the arrival of carp (Reynolds 1976a) but is now very rare in the region.

Gambusia is a small species (< 5 cm) that was introduced to Australia in the 1920s originally for use in aquaria and released into the wild for mosquito control (McDowall 1996b). It is now widespread in many areas and is ubiquitous to warm shallow waters associated with regulated Lower Murray wetlands (Lloyd & Walker 1986; Smith et al. 2007). This species is an aggressive competitor that can multiply rapidly and is subsequently thought to have contributed to the decline of some native fish species (Lloyd 1987; Arthington & Lloyd 1989). Redfin perch, brown trout and rainbow trout are generally cooler water species, which were introduced to Australia around the mid to late 1800s (McDowall 1996a).

Redfin perch are widely distributed in the region and they appear to have cycles of abundance (e.g. Fig. 4). Trout are occasionally reported in the main stream of the Murray and have been stocked into various tributary streams in the region (see Appendix). These three large predators are implicated in the decline of small native fishes (e.g. Crowl et al. 1992; Morgan et al. 2002). Redfin perch also carry a virus damaging to some native species and in general alien fishes are potential vectors for pathogens and parasites (e.g. Langdon & Humphrey 1987). There have been isolated records of Atlantic salmon in the mid 1990s comprising large fish netted near Renmark (SA Museum records). A single unverified report of oriental weatherloach has been made in the region (Hammer & Walker 2004); however, the arrival of this extremely hardy and prolific species is likely in the near future as it spreads downstream along the Murray (already common in central Victoria).





**Figure 3.** Alien fishes of the Lower River Murray, South Australia with indicative common size range: **a)** gambausia (male), 1.5-3 cm; **b)** gambausia (female), 2.5-4 cm; **c)** weather loach, 5-7 cm; **d)** redfin, 10-30 cm; **e)** tench, 15-40 cm; **f)** goldfish, 5-12 cm; **g)** common carp, 25-60 cm; **h)** common carp (mirror form), 25-50 cm; **i)** goldfish x common carp hybrid, 20-50 cm; **j)** brown trout, 15-40 cm; **k)** sleepy cod, 12-30 cm.

Two Australian native fishes translocated outside their natural range have been recorded in the region. Namely, Australian bass, a species endemic to coastal drainages of eastern Australia, was netted near Loxton in 1992, and sleepy cod, native to north and northeastern Australian waters, was reported near Swan Reach in the 1990s (SA Museum records; Hammer & Walker 2004). One sleepy cod captured from the River lived in a fish tank at the Swan Reach Hotel for several years (Fig. 3). Neither appears to have established populations in the region.

## **General ecology**

The unique and variable environment of the River Murray supports a diverse fish assemblage in terms of basic body morphologies, life history modes, and idiosyncratic fish adaptations and behaviours (see Schiller & Harris 2001; Lintermans 2007 for a broader coverage).

## **Size and age**

Fishes in the region range from the tiny (e.g. 3-4 cm adult size for carp gudgeons) to the mighty - Murray cod is Australia's, and one of the world's, largest freshwater fish reported to grow up to 1.8 m and 113 kg (Harris & Rowland 1996). Other large-bodied native fishes (i.e. potentially reaching a size of over 40 cm) include freshwater catfish, bony herring, other percichthyids (trout cod, golden perch, Macquarie perch, estuary perch) and silver perch. Most of the other species like galaxiids, smelt, Murray rainbowfish, hardyheads, gudgeons and gobies grow no larger than 10-15 cm. Longevity information is limited for many species in the region. Some large-bodied fishes such as Murray cod, golden perch and silver perch are long-lived. Murray cod has been aged up to 48 years (Anderson et al. 1992) and golden perch to 26 years (Mallen-Cooper & Stuart 2003). Many small fishes are presumably short-lived or have populations dominated by younger fish.

## **Habitat & movement**

Within exclusively freshwater species different ecologies are displayed. Some species are pelagic, living in open water, but the majority are demersal, associated with structure and cover. The adult stages of several large fishes are river specialists, particularly Murray cod which inhabit slow flowing rivers, anabranches and creeks, often in or near deep holes, and prefer habitats containing cover (e.g. rocks, clay banks, woody debris, snags, or overhanging vegetation: see Harris & Rowland 1996; Ye et al. 2000). A number of small fishes such as Murray rainbowfish, unspotted hardyhead, and carp gudgeons are wetland specialists, but given the lentic nature of the main channel of the Lower Murray (series of weir pools) they can also be found in river edge habitats (Smith et al. 2007; Davies et al. 2008). Other species such as pelagic bony herring and smelt are more generalists, having a broad distributional range across various habitat types.

Many native fish undergo longitudinal (i.e. up or down-stream) or lateral (i.e. between the main channel and floodplains) movement. Diadromous species recorded in the region have undergone determined migrations from the sea at particular life stages to colonise or move through the region (e.g. juvenile common galaxias and congolli, adult shortheaded lamprey). Some large-bodied species undertake long-distance longitudinal migrations as demonstrated by the seminal study of Reynolds (1983) who tagged several species in SA and recorded recaptures of golden perch and silver perch at various points up to 2300 km along the Murray and Darling rivers during floods (some fish also swam downstream to the Lower Lakes). Radio telemetry data for Murray cod in Victoria indicate that upstream spawning migrations of up to 80-100 km can occur, with fish then returning to the original snag within their home range/habitat (Koehn and Nicol 1998). Studies of fish moving through or accumulated below fishways on Murray weirs provide particular insight, and suggest general mobility across different life-stages for the aforementioned large-bodied species, plus movement of smaller species (e.g. smelt, carp gudgeons), especially during small rises in river flows/height (Mallen-Cooper 1996; Stuart et al. 2008). Nevertheless several benthic or habitat specialists including freshwater catfish, southern pygmy perch and southern purple-spotted gudgeon appear relatively sedentary (Reynolds 1983; Hammer 2001; Hammer 2008).

## **Reproduction**

The primary environmental conditions for stimulating reproduction in fishes of the Murray-Darling Basin are considered to be flow (increases for some species, low flow for others) and increasing temperature (Koehn & O'Connor 1990; Humphries et al. 1999). Many species also utilise particular habitat components for spawning within diverse breeding strategies (see Appendix). A number display parental care where the male will guard and tend to eggs while they develop inside hollow logs or other concealed places (e.g. Murray cod, river blackfish, gudgeon species). In the case of freshwater catfish they build a circular to oval nest around 0.6-2.0 m in diameter constructed from gravel and pebbles (Lake 1967b; Merrick & Midgley 1981). Other species like Murray rainbowfish and pygmy perches, attach or distribute adhesive eggs to a structure (vegetation or substratum), but do not guard the eggs/young. Golden perch and silver perch produce a large number of non-adhesive



semi-buoyant pelagic eggs which are at the disposal of currents (Lake 1967c; Rowland 1983). The sole representative of a live-bearing reproductive strategy is the exotic gambusia.

### **Feeding and diet**

The large-bodied fishes such as freshwater catfish, Murray cod and golden perch are mostly opportunistic carnivores as adults. Their diets include crustaceans, insects, molluscs and fish (Lake 1967a; Cadwallader 1977, 1979; Davis 1977; Llewellyn & MacDonald 1980). However, the larvae and small juveniles of these fish feed primarily on zooplankton such as cladocerans, copepods, and chironomids (Lake 1967a; Arumugam & Geddes 1987, 1992; Arumugam 1990; Culver & Geddes 1993; Rowland 1992). Several smaller native fish are also either benthic or nektonic (underwater surfaces) feeding carnivores (Lloyd 1987). Smelt, Murray rainbowfish, hardyheads and silver perch are omnivores as adults (additionally consuming algae and other plant matter: Lake 1967a; Lloyd 1987). Bony herring are mainly herbivores, concentrating on attached filamentous algae, but also feed on muddy detritus and biofilms on the surface of substrates (Merrick & Schmida 1984; Puckridge and Walker 1990).

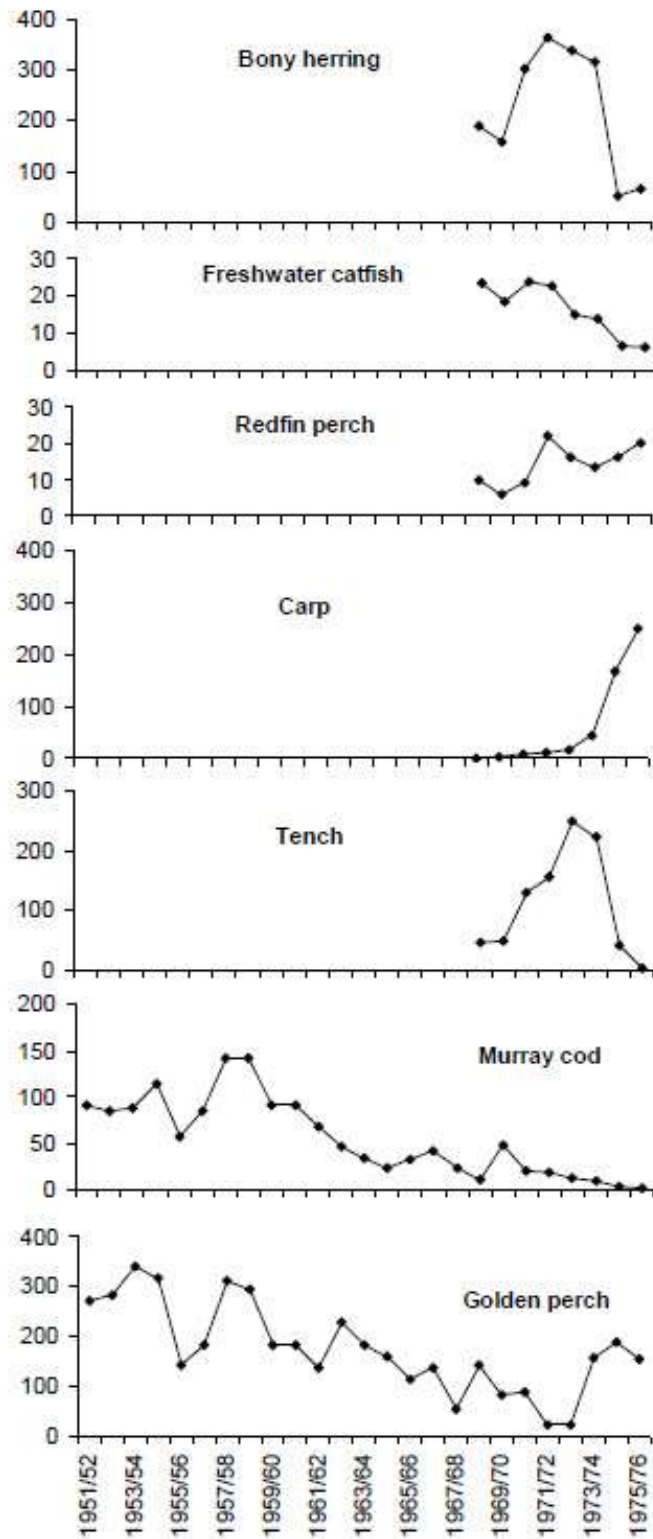
Floods provide enhanced opportunities for recruitment due to the increase in available aquatic resources (space and food). At first feeding many fish species are often restricted by mouth gape and poor swimming and pursuit abilities, and the presence of high concentrations of suitable-sized food is pivotal to larval survival and subsequent recruitment (Arumugam 1986; Arumugam & Geddes 1987; King et al. 2003).

### **FISHERY**

**Commercial fishery** A commercial fishery began shortly after European settlement and was well established by 1850 (Leslie 1995). Initially the fishery was concentrated on upriver population centres (e.g. Renmark and Morgan), and the majority of production was sold to meet local demand or shipped to Adelaide markets (Pillar 1980). In about 1908 the establishment of rail linkages allowed export via shipping to the higher-value Melbourne market (Leslie 1995). A "Reach" fishery was first established in 1923 (Poole 1984) whereby commercial fishers were responsible for the administration of their own reaches. The reach system in various forms continued until June 2003 (Poole 1984; Rohan 1987, 1988, 1989; Olsen 1991; Presser 1996). Fishing techniques primarily involved drum nets and gill nets. Drum nets work more effectively during flow events which trigger directional movement of fish, whereas gill nets take fish during more static conditions. A third commercial gear involved set lines comprising multiple hook-lines baited and set almost invariably near snags.

Catch statistics of key commercial species from the South Australian Inland Waters Fishery between 1951/52 and 1975/76 are shown in Figure 4. Catches of native species varied greatly, with production generally peaking during flood years, but overall they show a declining trend over time. A dramatic decline is evident in Murray cod production from late 1950s to mid 1970s (Reynolds 1976a). A similar downward trend is shown for golden perch, although less pronounced. The catches of catfish and silver perch have also undergone a significant decline (Reynolds 1976a; Pillar 1980; Rohan 1987). In contrast, a population explosion is apparent for carp since their introduction to South Australia in 1970 and they have become a large component of the catch.

As late as 2002/03, the annual commercial catches of Murray cod and golden perch were 27 and 7 tonnes, respectively. Sizable catches of bony herring and carp were mostly sold for marine lobster bait, whereas Murray cod and golden perch were palatable top-priced freshwater fishes. Detailed fishery assessment reports for Murray cod and golden perch in later years of fishery are provided by Ye et al. (2000), Ye (2005) and Ye & Zampatti (2007).



**Figure 4.** The annual commercial catches of key fish freshwater species from the South Australian River Murray and Lower Lakes between 1951/52 and 1975/76 (Data source: Reynolds 1976b; SA Fisheries Statistics). *Natural History of the Riverland and Murraylands* 342

## **Recreational fishing**

The historical development of the SA inland recreational fishery has closely paralleled that of the commercial sector (Leslie 1995). Besides the practical need for protein, fishing was a sporting diversion and leisure activity for the early European settlers, and the River Murray has remained the most popular inland recreational fishing region in SA (Philipson et al. 1986). A survey of recreational fishing in 1982/83 indicated that the Murray region received 200,000 visits through that year of which 66% (132,000) were for the purpose of fishing (Philipson et al. 1986), and the National Recreational and Indigenous Fishing Survey, undertaken between May 2000 and April 2001 (Henry & Lyle 2003), estimated that the total recreational catches of Murray cod, golden perch and carp from the River Murray were 2,233, 211,299 and 421,790 in numbers, equivalent to 22, 222 and 239 tonnes, respectively. The release rates were 10% for Murray cod and 62% for golden perch.

The recreational fishery is currently restricted to rod/handline and yabby traps (drum nets and set lines were historically permitted). A number of fish species from the River Murray are totally protected including silver perch, freshwater catfish, trout cod, river blackfish, and fish of the genera *Ambassis*, *Nannoperca* and *Mogurnda*. Other current restrictions for recreational fishing include closed seasons, legal minimum/maximum size and bag limits (i.e. for Murray cod and callop).

## **AN ALTERED ENVIRONMENT**

In various ways humans have exploited the fishes and resources of the Murray and affected its ecology. An intimate interaction with the River Murray stretches from aboriginal history and culture (e.g. Eyre 1815-1901 – Journals of Edward John Eyre, an account of the manners and customs of the aborigines and the state of their relations with Europeans: <http://ibiblio.org/gutenberg/etext04/xpcsc10.txt>; Walker 1986). However, the recent and ever increasing demands on the environment have undoubtedly had a major impact on native fish populations.

### **Threats**

Environmental degradation following European settlement has caused serious declines in native fish populations in the Murray-Darling Basin, and the impact has increased with levels of development (Cadwallader 1977; Walker 1985; Gehrke et al. 1995; Harris & Gehrke 1997). The declining productivity of the fisheries has been coincident with increased regulation of the water resource and reduced structural complexity of the ecosystem (Lake 1994; Reid et al. 1997). Abstraction for irrigation and water supply removes around 2/3 of the flow which naturally flowed to South Australia, and zero flows now occur 1 in 2 years compared with 1 in 20 under natural conditions (Maheshwari et al. 1995). A reduction in flow volume has been accompanied by a drastically altered natural flow regime including the reduced frequency and magnitude of floods, most notable in the loss of smaller floods and the altered timing of high and low flows (Walker 1993).

Broader changes are compounded by local regulation through the construction of ten lowlevel weirs in 1922-1935 (six of them within SA) and the Murray barrages in the 1940s (Walker 1985, 1992; Walker et al. 1992; Walker & Thoms 1993; Maheshwari et al. 1995). Furthermore, these major structures act as physical barriers to fish movement and migration (Mallen-Cooper 1993; Harris 2001). Recently, record drought has exacerbated the problems of regulation, and brings new potential reactions to water shortage such as another weir proposed for the Lower River reach at Wellington (DEH 2009). Some other key threats to native fish have been identified as habitat degradation and removal (e.g. de-snagging), lowered water quality, barriers, alien species, exploitation, diseases, translocation and stocking (Cadwallader 1978; Walker 1983, 2006; Lintermans 2007).

### **Status**

Considering environmental changes, the current threatened state of native fish is not surprising. Many are conservation-listed or protected at the State and national level. Lloyd & Walker (1986) considered nine species from the Lower Murray to be under threat and that no species could be considered secure. More recent assessments list around half of the 35 fish given in the Appendix in higher extinction risk categories (DEH 2003; Hammer et al. 2007). Broadly, fish populations in the Murray- Darling Basin are currently estimated to be about 10% of their pre-European settlement levels (MDBC 2004), and for many species in the study region this figure would seem generous.

A range of species occurred historically but are now presumably long extinct from the region including Murray galaxias, trout cod, Macquarie perch, and southern and Yarra pygmy perch. A more

recent loss includes the chanda perch, last recorded in 1983 from the Marne River mouth (Lloyd & Walker 1986). The southern purple-spotted gudgeon, was in 2004 rediscovered from a single wetland near Murray Bridge after a 30 year absence of records, but now with critical habitat drying is again likely to be locally extinct in the wild (Hammer 2008). Limited information points to significant decline in larger species (see Fishery), and certainly anecdotal information suggests species commonly encountered by anglers at certain times (especially freshwater catfish) have disappeared. Diadromous species including lampreys, shortfinned eel, estuary perch and congolli were once more abundant and widespread prior to the construction of the barrages (Sim et al. 2000; H. Jones personal communication), but have also undoubtedly suffered from wider ecosystem change. Other species including river blackfish and Murray hardyhead currently display highly restricted distributions in the region (Lloyd & Walker 1986; Hammer et al. 2007; Wedderburn et al. 2007).

## CONSERVATION

Native fish conservation and rehabilitation is an important part of environmental restoration as fishes are a significant yet threatened component of local biodiversity; they respond to change in aquatic habitats and hence are key indicators of ecosystem health; fishes have intrinsic value which extends from adaptation to local environments, links in food chains, and a role in culture and use (angling, food, aquarium fish); and they are iconic species (e.g. Harris 1995; Jackson et al. 2001).

Any broad measures to protect and restore flows and habitats will improve the prognosis for native fish (e.g. restoration of smaller floods, re-snagging, wetland habitat rehabilitation). For example, recent research work suggests that a within channel flow pulse may be sufficient to provide suitable conditions for spawning and enhance larval survivorship and recruitment of the large-bodied native fish, such as golden perch, in the lower River Murray (Cheshire & Ye 2008b; Ye et al. 2008). Continual improvements in fish passage both along the main stem and to off-channel habitats will facilitate fish movement, migration and dispersion, and give fish greater access to habitats (e.g. Mallen-Cooper 2001; Bice and Zampatti 2005; Leigh & Zampatti 2005; Barrett 2007). Finally science based decision tools and community involvement are key drivers for sustaining fish populations into the future.

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Table 1. Freshwater fishes recorded from the Riverland and Murraylands in South Australia. Information is based largely on collections housed at the South Australian Museum as well as relevant literature (e.g. Scott et al. 1974; Lloyd & Walker 1986; Hammer & Walker 2004; Smith & Hammer 2005; Hammer *et al.* 2007) and SARDI Fisheries Statistics. Taxonomy follows Hammer & Walker (2004). \* Distribution in study region forms a significant part of the Murray-Darling Range of the species. [*Life history strategy*: D; diadromous – migrates between fresh and saltwater, E; euryhaline – estuarine species comfortable in fresh and salt water, F; complete lifecycle in fresh water. *Reproductive strategy*: n; parental care – guard eggs and/or build a nest, a; use structure by attaching or distributing eggs within (e.g. aquatic vegetation), no parental care, r; distribute demersal eggs randomly, p; spawn surface drifting (pelagic) eggs, l; bear live young.].

Family	Taxon	Common name	Ecology	Comments on status in the River Murray
NATIVE SPECIES				
Geotriidae	<i>Geotria australis</i> Grey, 1851	Pouched lamprey	D, r	Declined significantly from historic abundance, endangered
Mordaciidae	<i>Mordacia mordax</i> (Richardson, 1846)	Shortheaded lamprey	D, r	Still recorded in the Lower Murray and upstream near Yarrawonga (Vic.), migrations with large numbers of individuals no longer occur, endangered
Anguillidae	<i>Anguilla australis</i> Richardson, 1841	Shortfinned eel	D, p?	The Murray-Darling Basin lies at the western edge of the species' distribution. Larval migration to the region is spasmodic – naturally rare but may have declined
Plotosidae	<i>Tandanus tandanus</i> Mitchell, 1838	Freshwater catfish	F, n	Historically very common, significant declines in the 1970s & 80s. Remains at a low population level
Clupeidae	<i>Nematalosa erebi</i> (Günther, 1868)	Bony herring	F, p	One of the most common species in the region
Retropinnidae	<i>Retropinna semoni</i> (Weber, 1895)	Smelt	F, r	One of the most common species in the region
Galaxiidae	<i>Galaxias brevipinnis</i> Günther, 1866	Climbing galaxias	D, a	A migratory species that probably occasionally enters the region as juveniles
	<i>Galaxias maculatus</i> (Jenyns, 1842)	Common galaxias	D/F, a	Common in the lower reaches
	<i>Galaxias olidus</i> Günther, 1866	Mountain galaxias	F, a	A species complex. Recorded from a tributaries (Marne River and Reedy Creek) and historically from swamps near Murray Bridge
	<i>Galaxias rostratus</i> Klunzinger, 1872	Murray galaxias	F, r	A floodplain/wetland species, only early records from near Murray Bridge but probably occurred patchily right along the River. Likely extinct
Melanotaeniidae	<i>Melanotaenia fluviatilis</i> (Castelnau, 1878)	Murray rainbowfish	F, a	Still common in wetland environments
Atherinidae	<i>Atherinosoma microstoma</i> (Günther, 1861)	Small-mouthed hardyhead	E, a	Occasionally recorded from the lower reaches

	<i>Craterocephalus fluviatilis</i> McCulloch, 1912		Murray hardyhead	F, a	*Poorly known species. Historically widespread, known remaining habitats in region include wetlands near Murray Bridge and saline habitat near Renmark/Berri
	<i>Craterocephalus fulvus</i> Ivanstoff, Crowley & Allen, 1987	<i>stercusmuscarum</i>	Unspecked hardyhead	F, a	Common but patchy distribution in wetland, river edge habitats
Ambassidae	<i>Ambassis agassizii</i> 1867	Steindachner,	Chanda perch	F, a	Poorly known species. Does not appear to have been common historically, last record was a single individual in 1983 from the Marne R mouth
Percichthyidae	<i>Gadopsis marmoratus</i> 1848	Richardson,	River blackfish	F, n	Historic reports from the main channel, now endangered and remaining only in a small pocket in a tributary (Marne R)
	<i>Maccullochella macquariensis</i> (Cuvier, 1829)		Trout cod	F, n	Also known as rock cod locally. Confused with Murray cod in the past, but indications that it occurred not-uncommonly prior to river regulation. Presumed extinct
	<i>Maccullochella peelii peelii</i> (Mitchell, 1838)		Murray cod	F, n	Historically very common. Large declines, now at a low population level
	<i>Macquaria ambigua ambigua</i> (Richardson, 1845)		Murray-Darling golden perch	F, p	Remains common
	<i>Macquaria australasica</i> Cuvier, 1830		Macquarie perch	F, a	Poorly known species in SA. Some indication that it occurred in the upper section of the region historically either as a local population or as occasional downstream immigrants. Suitable habitat no longer occurs – extinct
	<i>Macquaria colonorum</i> 1863)	(Günther,	Estuary perch	D, p	*Poorly known species in SA. More common prior to the construction of the barrages, now only occasional records. Unknown if locally reproducing population remains or if now only migrants to the area
	<i>Nannoperca australis</i> Günther, 1861		Southern pygmy perch	F, a	Common in swamps downstream of Mannum historically, potentially also occurred patchily in wetland/billabong habitats along the River. Now extinct from region (still in other tributaries to Lake Alexandrina)
<i>Nannoperca obscura</i> 1872)	(Klunzinger,	Yarra pygmy perch	F, a	*Poorly known species in SA, confused historically with southern pygmy perch. Some early records for the lower reaches, presumed extinct	

Terapontidae	<i>Bidyanus bidyanus</i> (Mitchell, 1838)	Silver perch		F, p	Historically very common. Significant declines since the 1970s, remains rare. Juveniles are common encountered by anglers in some upstream sections of the SA Murray
	<i>Leiopotherapon unicolor</i> (Günther, 1859)	Spangled perch		F, r	Occasional records of fish that come down the Darling R during floods
Pseudaphritidae	<i>Pseudaphritis urvillii</i> (Valenciennes, 1832)	Congolli		r?	*Historically common. Large declines in the region, now rarely recorded
Eleotridae	<i>Hypseleotris klunzingeri</i> (Ogilby, 1898)	Western gudgeon	carp	F, n	Poorly known in SA. Considerable taxonomic confusion with the carp gudgeons. Mainly along river edge
	<i>Hypseleotris</i> sp. 1 (undescribed)	Midgley's gudgeon	carp	F, n	A common species of wetlands and the river edge
	<i>Hypseleotris</i> sp. 3 (undescribed)	Murray-Darling gudgeon	carp	F, n	A not uncommon species of wetlands, also in tributaries (Marne R, Saunders Ck)
	<i>Hypseleotris</i> spp. (species complex)	Hybrid forms (e.g. Lake's carp gudgeon)		F, n	Considerable taxonomic confusion with the carp gudgeons extends to a range of F1 hybrid forms which may prove to be unisexual lineages. Combined hybrid forms are common in the region
	<i>Mogurnda adpersa</i> (Castelnau, 1878)	Southern purple-spotted gudgeon		F, n *	Historically common in wetlands and sheltered river edge habitat until the early 1970s. Rediscovered from a single wetland near Murray Bridge in 2004, but has again disappeared with a major drop in river level. Captive breeding underway
	<i>Philypnodon grandiceps</i> (Krefft, 1864)	Flathead gudgeon		F, n	Remains common
	<i>Philypnodon</i> sp. (undescribed)	Dwarf gudgeon	flathead	F, n	*Widespread distribution but occurs normally in low numbers
Gobiidae	<i>Pseudogobius olorum</i> (Sauvage, 1880)	Western goby	bluespot	E, n	*Probably occurred in lower reaches historically, now confined to below Wellington
	<i>Tasmanogobius lasti</i> Hoese, 1991	Lagoon goby		E, n	*Still occasionally recorded from lower reaches (e.g. Swanport, Lake Carlet, Rocky Gully)
TRANSLOCATED AUSTRALIAN NATIVE SPECIES					
Percichthyidae	<i>Macquaria novemaculeata</i> (Steindachner, 1866)	Australian bass		D, p S	single specimen netted near Loxton in 1992. Native to coastal drainages of eastern Australia
Eleotridae	<i>Oxyeleotris lineolata</i> (Steindachner, 1867)	Sleepy cod		F, n	Two independent reports from near Swan Reach in the 1990s. Native to coastal drainages of northern and north east Australia

EXOTIC SPECIES				
Cyprinidae	<i>Carassius auratus</i> (Linnaeus, 1758)	Goldfish	F, a	Common since its introduction soon after European settlement (also known as golden carp) Arrived in the region in early 1970s from whence its populations expanded rapidly. Remains common An early introduction to the region. Reportedly common until the arrival of carp
	<i>Cyprinus carpio</i> Linnaeus, 1758	Common carp	F, a	
	<i>Tinca tinca</i> (Linnaeus, 1758)	Tench	F, a	
Cobitidae	<i>Misgurnus anguillicaudatus</i> (Cantor, 1842)	Oriental weatherloach	F, r	Unverified report for near Murray Bridge in the 1980s, an imminent arrival as it spreads down the Murray from Victoria and New South Wales
Salmonidae	<i>Oncorhynchus mykiss</i> (Walbaum, 1792)	Rainbow trout	F, n	Occasional records from the region (e.g. Renmark in 1917), has been stocked in tributaries like the Marne R, Burra Ck and Baldina Ck A few records from the mid 1990s of large fish netted near Renmark Occasional records from the region (e.g. Murray Bridge 1972), has been stocked in tributaries like the Marne R, Reedy Ck and Burra Ck
	<i>Salmo salar</i> Linnaeus, 1758	Atlantic salmon	D, n	
	<i>Salmo trutta</i> Linnaeus, 1758	Brown trout	F, n	
Poeciliidae	<i>Gambusia holbrooki</i> Girard, 1859	Gambusia	F, l	Present in the region since at least the 1950s, has become a dominant species in still, shallow and degraded wetland areas
Percidae	<i>Perca fluviatilis</i> Linnaeus, 1758	Redfin perch	F, a	Present since at least the 1950s, reached plague proportions at times, now less common

