



INTERIM RECEIVING WATERS MONITORING REPORT FOR THE POINT LOMA AND SOUTH BAY OCEAN OUTFALLS

2020

Environmental Monitoring and Technical Services
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June 30, 2021

Mr. David W. Gibson, Executive Officer
California Regional Water Quality Control Board
San Diego Region
2375 Northside Drive, Suite 100
San Diego, CA 92108

Attention: POTW Compliance Unit

Dear Mr. Gibson:

Enclosed is the 2020 Interim Receiving Waters Monitoring Report for the Point Loma and South Bay Ocean Outfalls, as per requirements set forth in the following Orders/Permits:

- (1) Order No. R9-2017-0007 for the City of San Diego's Point Loma Wastewater Treatment Plant (NPDES No. CA0107409).
- (2) Order No. R9-2013-0006 (as amended) for the City's South Bay Water Reclamation Plant (NPDES No. CA0109045).
- (3) Order R9-2014-0009 (as amended) for the United States Section of the International Boundary and Water Commission's South Bay International Wastewater Treatment Plant (NPDES No. CA0108928).

This combined report for the Point Loma and South Bay outfall regions contains data summaries for all portions of the Ocean Monitoring Program conducted during 2020. Additional data in support of this report will be submitted in separate addenda, which will be available online by July 1, 2021.

I certify under penalty of law that this document and all attachments were prepared under my direction or supervision in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on my inquiry of the person or persons who manage the system or those persons directly responsible for gathering the information, the information submitted is, to the best of my knowledge and belief, true, accurate, and complete. I am aware that there are significant penalties for submitting false information, including the possibility of fine and imprisonment for knowing violations.

If you have questions regarding this report, please call Dr. Ryan Kempster, the City's Senior Marine Biologist at (619) 758-2329.

Sincerely,



Peter S. Vroom, Ph.D.
Deputy Public Utilities Director

RK/akl

cc: U.S. Environmental Protection Agency, Region 9
International Boundary and Water Commission, U.S. Section



INTERIM RECEIVING WATERS MONITORING REPORT FOR THE POINT LOMA AND SOUTH BAY OCEAN OUTFALLS 2020

POINT LOMA WASTEWATER TREATMENT PLANT
(ORDER No. R9-2017-0007; NPDES No. CA0107409)

SOUTH BAY WATER RECLAMATION PLANT
(ORDER No. R9-2013-0006 AS AMENDED; NPDES No. CA0109045)

SOUTH BAY INTERNATIONAL WASTEWATER TREATMENT PLANT
(ORDER No. R9-2014-0009 AS AMENDED; NPDES No. CA0108928)

Prepared by:

City of San Diego Ocean Monitoring Program
Environmental Monitoring & Technical Services Division

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June 2021

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Executive Summary

Executive Summary

The City of San Diego (City) conducts an extensive Ocean Monitoring Program to evaluate potential environmental effects associated with the discharge of treated wastewater to the Pacific Ocean via the Point Loma and South Bay Ocean Outfalls (PLOO and SBOO, respectively). Data collected are used to determine compliance with receiving water quality requirements, as specified in National Pollutant Discharge Elimination System (NPDES) permits, and associated orders; these permits and orders are issued by the San Diego Regional Water Quality Control Board (SDRWQCB) and the U.S. Environmental Protection Agency (USEPA) for the City's Point Loma Wastewater Treatment Plant (PLWTP), South Bay Water Reclamation Plant (SBWRP), and the South Bay International Wastewater Treatment Plant (SBIWTP), which is operated by the U.S. Section of the International Boundary and Water Commission (USIBWC). Treated effluent from both the SBWRP and SBIWTP commingle before discharge to the ocean via the SBOO, thus a single monitoring and reporting program, approved by the SDRWQCB and USEPA, is conducted to comply with these two permits.

The principal objectives of the combined ocean monitoring efforts for both the PLOO and SBOO regions include: (1) measure and document compliance with NPDES permit requirements and California Ocean Plan (Ocean Plan) water quality objectives and standards; (2) assess any impact of wastewater discharge, or other anthropogenic inputs, on the local marine ecosystem, including effects on coastal water quality, seafloor sediments, and marine life; (3) monitor natural spatial and temporal fluctuations of key oceanographic parameters, and evaluate the overall health and status of the San Diego marine environment.

Although governed by three separate NPDES permits, this interim report summarizes the purpose, scope, methods, and findings of all receiving waters monitoring conducted for the PLOO and SBOO regions from January through December 2020. A full biennial monitoring and assessment report covering calendar years 2020 and 2021 will be produced and submitted to the San Diego Water Board and USEPA no later than July 1, 2022. Specific details of the main ocean monitoring activities conducted during 2020 are presented in the following five chapters herein, while raw data are presented in Appendices A–D or are available online and by request. Chapter 1 provides a general introduction and overview of the combined PLOO and SBOO program. Chapter 2 presents data characterizing the results of water quality monitoring at 103 different shore or offshore stations located throughout the two regions. This includes measuring concentrations of fecal indicator bacteria in seawater samples and collecting various types of oceanographic data to evaluate dispersal of the PLOO and SBOO wastewater plumes and to assess compliance with Ocean Plan water contact standards. Assessments of benthic sediment quality (e.g., sediment chemistry, particle size distributions) and the ecological status of macrobenthic invertebrate communities at 49 core monitoring stations are presented in Chapter 3. Chapter 4 presents the results of trawling activities conducted at 13 different monitoring stations to assess the health and status of bottom dwelling (demersal) fish and megabenthic invertebrate communities. Contaminants in marine fishes collected from trawl and rig fishing stations are presented in Chapter 5.

Overall, the state of San Diego's coastal ocean waters remained in good condition in 2020 based on the preliminary findings and conclusions summarized in this report. Results for both the PLOO and SBOO regions were consistent with conditions documented in previous years, and there were few changes to local receiving waters, benthic sediments, and marine invertebrate and fish communities that could be attributed

to wastewater discharge or other human activities. Coastal water quality conditions and compliance with Ocean Plan standards were excellent, and there was no evidence that wastewater plumes from the two outfalls were transported into nearshore recreational waters. There were also no clear outfall related patterns in sediment contaminant distributions or differences between invertebrate and fish assemblages at the different monitoring sites. Additionally, benthic habitats surrounding both outfalls, and throughout the entire San Diego region, remained in good overall condition similar to reference conditions for much of the Southern California Bight. Finally, the low levels of contaminant accumulation and general lack of physical anomalies, or other symptoms of disease or stress in local fishes was also indicative of a healthy marine environment off San Diego.

Chapter 1

General Introduction

Chapter 1. General Introduction

PROGRAM REQUIREMENTS & OBJECTIVES

Ocean monitoring within the Point Loma and South Bay outfall regions is conducted by the City of San Diego (City) in accordance with requirements set forth in National Pollution Discharge Elimination System (NPDES) permits and associated orders for the following: the Point Loma Wastewater Treatment Plant (PLWTP), the South Bay Water Reclamation Plant (SBWRP), and the South Bay International Wastewater Treatment Plant (SBIWTP), which is owned and operated by the U.S. Section of the International Boundary and Water Commission (USIBWC) (see Table 1.1). These documents specify the terms and conditions that allow treated effluent to be discharged to the Pacific Ocean via the Point Loma Ocean Outfall (PLOO) and South Bay Ocean Outfall (SBOO). In addition, the Monitoring and Reporting Program (MRP), included within each of these orders, defines the requirements for monitoring ocean (receiving) waters surrounding the two outfalls. These requirements include sampling design, frequency of sampling, field operations and equipment, regulatory compliance criteria, types of laboratory tests and analyses, data management and analysis, statistical methods and procedures, environmental assessment, and reporting guidelines.

The combined ocean monitoring program for these regions is designed to assess the impact of treated wastewater discharged through the PLOO and SBOO on the coastal marine environment off San Diego. The main objectives of the program are to: (1) provide data that satisfy NPDES permit requirements; (2) demonstrate compliance with water-contact standards specified in the California Ocean Plan (Ocean Plan); (3) track movement and dispersion of the wastewater plumes discharged via the outfalls; (4) identify any biological, chemical or physical changes that may be associated with the outfalls and wastewater discharge. These data are used to evaluate and document any potential effects of treated wastewater discharge, or other anthropogenic inputs (e.g., storm water discharge, urban runoff), and natural influences (e.g., seasonality, climate change) on coastal water quality, seafloor sediment conditions, and local marine organisms.

BACKGROUND

Point Loma Ocean Outfall

The City began operation of the PLWTP and original PLOO off Point Loma in 1963, at which time treated effluent was discharged approximately 3.9 km west of the Point Loma peninsula at a depth of around 60 m. The PLWTP operated as a primary treatment facility from 1963 to 1985, after which it was upgraded to advanced primary treatment between mid-1985 and July 1986. This improvement involved the addition of chemical coagulation to the treatment process, which resulted in an increase in removal of total suspended solids (TSS) to about 75%. Since then, the treatment process has continued to be improved with the addition of more sedimentation basins, expanded aerated grit removal, and refinements in chemical treatment, which together further reduced mass emissions from the plant. For example, TSS removals are now consistently greater than the 80%, as required by the NPDES permit.

The structure of the PLOO was significantly modified in the early 1990s when it was extended about 3.3 km farther offshore in order to prevent intrusion of the waste field into nearshore waters and to increase compliance with Ocean Plan standards for water-contact sports areas. Discharge from the original

60-m terminus was discontinued in November 1993 following completion of the outfall extension. Currently, the PLOO extends approximately 7.2 km west of the PLWTP to a depth of around 94 m, where the main outfall pipe splits into a Y-shaped (wye) multiport diffuser system. The two diffuser legs extend an additional 762 m to the north and south, each terminating at a depth of about 98 m. The average discharge of effluent through the PLOO in 2020 was ~144 million gallons per day (mgd).

South Bay Ocean Outfall

The SBOO is located just north of the international border between the United States and Mexico where it terminates approximately 5.6 km offshore and west of Imperial Beach at a depth of around 27 m. Unlike other southern California ocean outfalls that lie on the surface of the seafloor, the SBOO pipeline begins as a tunnel on land that extends from the SBWRP and SBIWTP facilities to the coastline, after which it continues beneath the seabed 4.3 km offshore. The outfall pipe connects to a vertical riser assembly that conveys effluent to a pipeline buried just beneath the surface of the seafloor. This subsurface pipeline then splits into a Y-shaped (wye) multiport diffuser system with the two diffuser legs each extending an additional 0.6 km to the north or south. The SBOO was originally designed to discharge wastewater through 165 diffuser ports and risers, which included one riser at the center of the wye and 82 risers spaced along each diffuser leg. Since discharge began, however, low flow rates have required closure of all ports along the northern diffuser leg and many along the southern diffuser leg in order for the outfall to operate effectively. Consequently, wastewater discharge is restricted primarily to the distal end of the southern diffuser leg and to a few intermediate points at or near the center of the wye. The average discharge of effluent through the SBOO in 2020 was about ~31 mgd, including 4 mgd of secondary and tertiary treated effluent from the SBWRP, and 27 mgd of secondary treated effluent from the SBIWTP.

RECEIVING WATERS MONITORING

The total area for the PLOO and SBOO monitoring program covers approximately 881 km² (~340 mi²) of coastal marine waters from Northern San Diego County into Northern Baja California. Core monitoring for the Point Loma region is conducted at 82 different stations, located from the shore to a depth of around 116 m. Core monitoring for the South Bay region is conducted at a total of 53 stations, ranging from the shore to depths of around 61 m (Figure 1.1). Each of the core monitoring stations is sampled for specific parameters as stated in their respective MRPs. A summary of the results for all quality assurance procedures performed during 2020, in support of these requirements, can be found in City of San Diego (2021a). Data files, detailed methodologies, completed reports, and other pertinent information submitted to the San Diego Regional Water Quality Control Board (SDRWQCB), and the U.S. Environmental Protection Agency (USEPA), during the past year, are available online (City of San Diego 2021b).

Prior to 1994, the City conducted an extensive ocean monitoring program off Point Loma surrounding the original 60-m discharge site. This program was subsequently expanded with the construction and operation of the deeper outfall, as discussed previously. Data from the last year of regular monitoring near the original PLOO discharge site are presented in City of San Diego (1995b), while the results of a 3-year “recovery study” are summarized in City of San Diego (1998). Additionally, a more detailed assessment of spatial and temporal patterns surrounding the original discharge site is available in Zmarzly et al. (1994). From 1991 through 1993, the City also conducted “pre-discharge” monitoring for the new PLOO discharge site in order to collect baseline data prior to wastewater discharge into these deeper waters (City of San Diego 1995a,b). All permit mandated ocean monitoring for the South Bay region has also been performed by the City since wastewater discharge through the SBOO began in 1999; this included pre-discharge monitoring for 3½ years (July 1995–December 1998) in order to provide background information against which

post-discharge conditions could be compared (City of San Diego 2000). Results of NPDES mandated monitoring for the extended PLOO from 1994 to 2019, and the SBOO from 1999 to 2019, are available in previous annual receiving waters monitoring reports (e.g., City of San Diego 2020). Finally, additional detailed assessments of the PLOO region have been completed as part of past modified NPDES permit renewal applications for the PLWTP submitted by the City and subsequent technical decisions issued by the USEPA (e.g., City of San Diego 2015a, USEPA 2017).

The City has also conducted annual region-wide surveys off the coast of San Diego since 1994, either as part of regular outfall monitoring requirements (e.g., City of San Diego 1999, 2020), or as part of larger multi-agency surveys of the entire Southern California Bight (SCB). The latter include the 1994 Southern California Bight Pilot Project (Allen et al. 1998, Bergen et al. 1998, 2001, Schiff and Gossett 1998) and subsequent Bight'98, Bight'03, Bight'08, Bight'13 and Bight'18 programs in 1998, 2003, 2008, 2013 and 2018 respectively (Allen et al. 2002, 2007, 2011, Noblet et al. 2002, Ranasinghe et al. 2003, 2007, 2012, Schiff et al. 2006, 2011, Dodder et al. 2016, Gillett et al. 2017, Walther et al. 2017, BSQPC 2018, SCCWRP 2018). These large-scale surveys are useful for characterizing the ecological health of diverse coastal areas to distinguish reference sites from those impacted by wastewater or storm water discharges, urban runoff, or other sources of contamination. In addition to the above activities, the City participates as a member of the Region Nine Kelp Survey Consortium to fund aerial surveys of all the major kelp beds in San Diego and Orange Counties (e.g., MBC Applied Environmental Sciences 2020).

SPECIAL STUDIES & ENHANCED MONITORING

The City has actively participated in, or supported, numerous important special projects, or enhanced ocean monitoring studies, over the past 10 years or more. Many of these projects to date were identified as part of a scientific review of the City's Ocean Monitoring Program, conducted by the Scripps Institution of Oceanography (SIO) and other participating institutions (SIO 2004). This review evaluated the environmental monitoring needs of the region, and recommended special projects based on priorities identified. Examples of special projects currently underway, or being initiated include:

- San Diego Kelp Forest Ecosystem Monitoring Project: This project represents continuation of a long-term commitment by the City to support important research conducted on local kelp forests by SIO. This work is essential to assessing the health of San Diego's kelp forests and monitoring the effects of wastewater discharge on the local coastal ecosystem relative to other anthropogenic and natural influences (see City of San Diego 2020: Appendix A).
- Real-Time Oceanographic Mooring Systems (RTOMS) for the PLOO and SBOO: This project addresses recommendations that the City should improve monitoring of the fate and behavior of wastewater discharged to the ocean via the SBOO (Terrill et al. 2009) and PLOO (Rogowski et al. 2012a, 2012b, 2013). The project involves the deployment of RTOMS at the terminal ends of the PLOO and SBOO to provide real time data on ocean conditions. The project began in late 2015 with initial deployment of the SBOO mooring in December 2016 and the PLOO mooring in March 2018. This project is being conducted in partnership with SIO, whom presently operate a similar mooring system off Del Mar. The project is expected to significantly enhance the City's environmental monitoring capabilities in order to address current and emerging issues relevant to the health of San Diego's coastal waters, including plume dispersion, subsurface current patterns, ocean acidification, hypoxia, nutrient sources, and coastal upwelling. Additional details are available in the approved Plume Tracking Monitoring Plan for the project (City of San Diego 2018b) and City of San Diego 2020: Appendix B.

- Sediment Toxicity Monitoring of the San Diego Ocean Outfall Regions: This project started with a 3-year pilot study implemented as a new joint regulatory requirement for the Point Loma and South Bay outfall regions in 2015. Findings for the 2016–2018 pilot study (City of San Diego 2015b) were summarized in a final project report (City of San Diego 2019) that included recommendations for continued sampling through 2023. This final project report has been updated to include results from 2019 as City of San Diego 2020: Appendix C.
- Remote Sensing of the San Diego/Tijuana Coastal Region: This project represents a long-term effort, funded by the City and the USIBWC since 2002, to utilize satellite and aerial imagery to better understand regional water quality conditions off San Diego. The project is conducted by Ocean Imaging (Littleton, CO), and is focused on detecting and tracking the dispersion of wastewater plumes from local ocean outfalls and nearshore sediment plumes caused by stormwater runoff or outflows from local bays and rivers (Hess 2019, 2020).
- San Diego Regional Benthic Condition Assessment Project: This multi-phase study represents an ongoing, long-term project designed to assess the condition of continental shelf and slope habitats throughout the entire San Diego region. A preliminary summary of the deeper slope (>200 m) results for data collected between 2003–2013 was included in Appendix C.5 of City of San Diego (2015a), while several publications covering the remainder of the project are planned for completion in late 2021.

REPORT COMPONENTS & ORGANIZATION

This report presents summaries of the results of all receiving waters monitoring activities conducted during January–December 2020 for both the Point Loma and South Bay outfall regions. A more comprehensive assessment, including detailed comparisons of long-term spatial and temporal changes and trends, will be prepared as part of the Biennial Receiving Waters Monitoring and Assessment Report for 2020–2021 to be submitted to the San Diego Water Board and USEPA by July 1, 2022. Included herein are results from all regular core stations that comprise the fixed-site monitoring grids surrounding the two outfalls (Figure 1.1), as well as results from the 2020 summer benthic survey of randomly selected sites that range from near the USA/Mexico border to northern San Diego County (Figure 1.2). Data from the 2018 SCB Regional Monitoring Program are not yet available and are therefore not included herein. The major components of the combined PLOO and SBOO monitoring program are covered in the following chapters and associated appendices of this report: Executive Summary; General Introduction (Chapter 1); Water Quality (Chapter 2, Appendix A); Benthic Conditions (Chapter 3, Appendix B); Demersal Fishes and Megabenthic Invertebrates (Chapter 4, Appendix C); Contaminants in Marine Fishes (Chapter 5, Appendix D).

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CHAPTER 1
FIGURES & TABLES

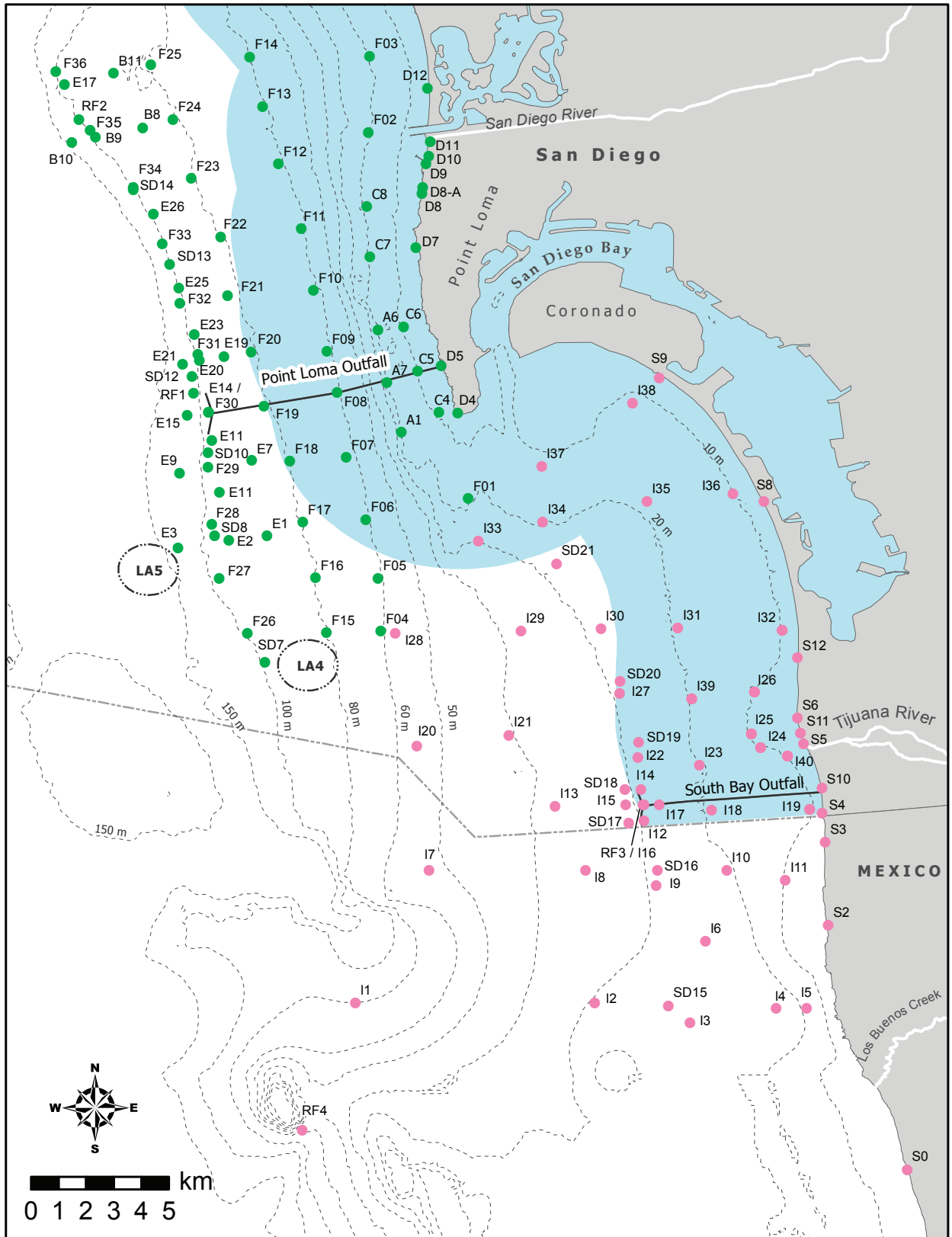


Figure 1.1

Core receiving waters monitoring stations for the PLOO (green) and SBOO (pink) as part of the City of San Diego's Ocean Monitoring Program. Light blue shading represents State of California jurisdictional waters.

Table 1.1

NPDES permits and associated orders issued by the San Diego Water Board for the Point Loma Wastewater Treatment Plant (PLWTP), South Bay Water Reclamation Plant (SBWRP), and South Bay International Wastewater Treatment Plant (SBIWTP) discharges to the Pacific Ocean via the PLOO and SBOO.

Facility	Outfall	NPDES Permit No.	Order No.	Effective Dates
PLWTP	PLOO	CA0107409	R9-2017-0007	October 1, 2017–September 30, 2022
SBWRP	SBOO	CA0109045	R9-2013-0006 ^a	April 4, 2013–April 3, 2018
SBIWTP	SBOO	CA0108928	R9-2014-0009 ^b	August 1, 2014–July 31, 2019

^aOrder R9-2013-0006 amended by Order R9-2014-0071 and R9-2017-0023 (permit administratively extended)

^bOrder R9-2014-0009 amended by Order R9-2014-0094, R9-2017-0024 and R9-2019-0012

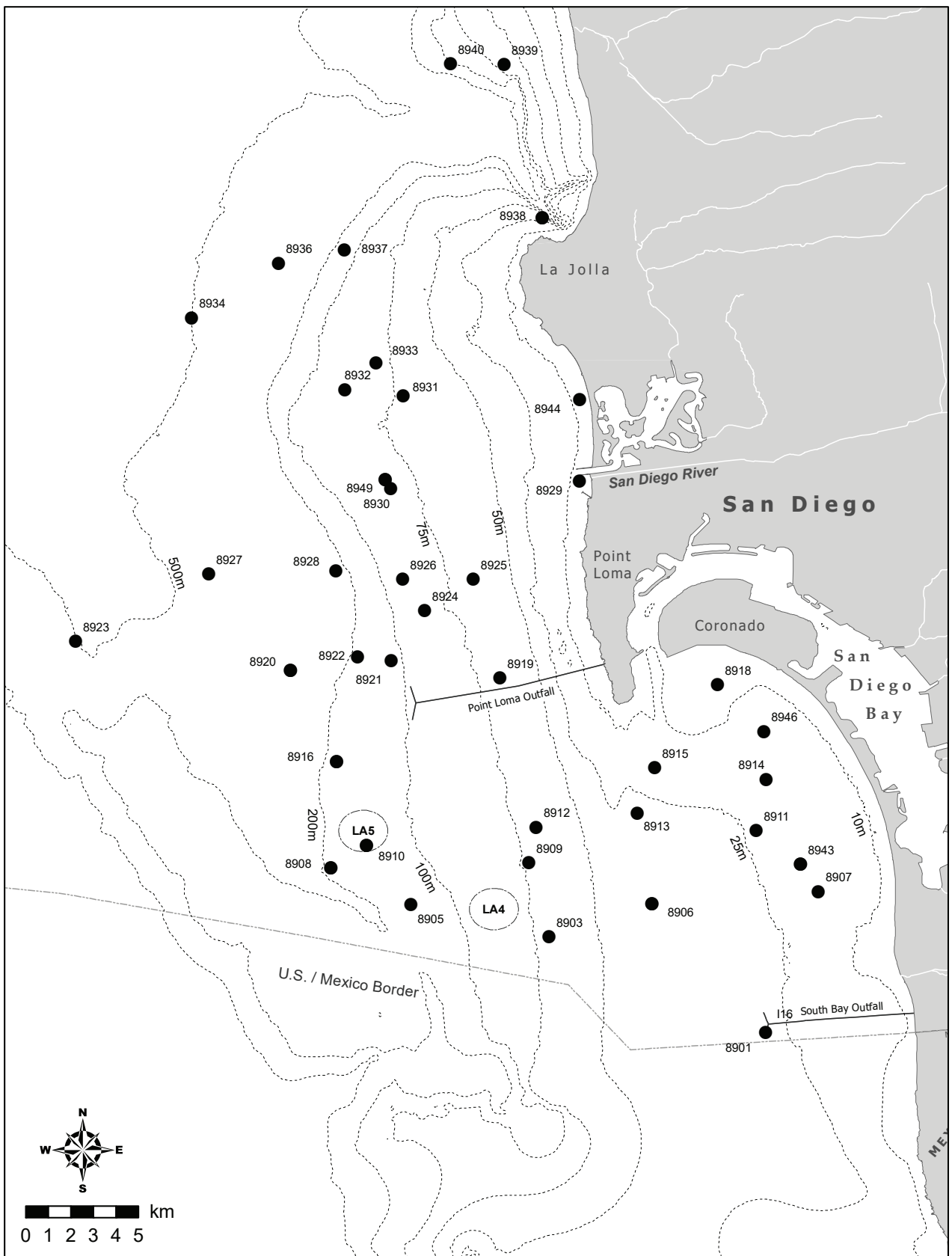


Figure 1.2
 Regional randomly selected benthic survey stations sampled during summer 2020 as part of the City of San Diego's Ocean Monitoring Program.

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Chapter 2

Water Quality

Chapter 2. Water Quality

INTRODUCTION

The City of San Diego conducts extensive monitoring along the shoreline and in offshore coastal waters surrounding the Point Loma and South Bay Ocean Outfalls (PLOO and SBOO, respectively) to characterize regional water quality conditions and to identify possible impacts of wastewater discharge or other contaminant sources on the marine environment. In addition, the City's water quality monitoring efforts are designed to assess compliance with the water contact standards specified in the California Ocean Plan (Ocean Plan) to protect the beneficial uses of California's ocean waters (SWRCB 2015). This chapter presents summaries and preliminary analyses of the oceanographic and microbiological data collected during 2020 at a total of 103 water quality monitoring stations and two real-time oceanographic mooring systems (RTOMS) surrounding the PLOO and SBOO. Raw data summaries supporting these results are presented in Appendix A. A more comprehensive assessment of these results will be presented in the 2020-2021 Biennial Assessment Report to be submitted by July 1, 2022.

MATERIALS AND METHODS

Field Sampling

Shore stations

Seawater samples were collected weekly at 19 shore stations to monitor concentrations of fecal indicator bacteria (FIB) in waters adjacent to public beaches (Figure 2.1). Sixteen of these stations are in California State waters and are therefore subject to Ocean Plan water contact standards (Table 2.1, SWRCB 2015). These include eight PLOO stations (D4, D5, D7, D8/D8-A/D8-B, D9, D10, D11, D12) located from Mission Beach southward to the tip of Point Loma and eight SBOO stations (S4, S5, S6, S8, S9, S10, S11, S12) located between the USA/Mexico border and Coronado. Over the past several years, due to increasing instability in some cliffside areas of Point Loma, City staff have periodically been unable to safely access and sample some stations. As a result, the following modifications to sampling locations, which were fully approved by the Regional Board, have occurred: (1) Station D8 was replaced by alternate station D8-A during July 2016; (2) D8-A was subsequently replaced by station D8-B in March 2018; (3) D8-A sampling resumed in December 2020. The remaining three SBOO shore stations (S0, S2, S3) are located south of the international border and are not subject to Ocean Plan compliance standards.

Seawater samples were collected from the surf zone at each of the above stations in sterile 250-mL bottles, after which they were transported on blue ice to the City's Marine Microbiology Laboratory and analyzed to determine concentrations of three types of FIB (i.e., total coliform, fecal coliform, *Enterococcus* bacteria). In addition, weather conditions and visual observations of water color, surf height, and human/animal activity were recorded at the time of collection. These observations have been previously reported in monthly receiving waters monitoring reports submitted to the San Diego Regional Water Quality Control Board (SDRWQCB) (see City of San Diego 2020–2021a,b), and are available online (City of San Diego 2021b).

Kelp and other offshore stations

Fifteen stations located in relatively shallow waters within or near the Point Loma or Imperial Beach kelp forests (referred to as “kelp” stations herein) were monitored weekly to assess water quality conditions and Ocean Plan compliance in nearshore areas used for recreational activities, such as SCUBA diving, surfing, fishing, and kayaking (Figure 2.1). These included PLOO stations C4, C5, and C6 located along the 9-m depth contour near the inner edge of the Point Loma kelp forest; PLOO stations A1, A6, A7, C7, and C8 located along the 18-m depth contour near the outer edge of the kelp forest; SBOO stations I25, I26, and I39 located at depths of 9–18 m contiguous to the Imperial Beach kelp bed; SBOO stations I19, I24, I32, and I40 located in other nearshore waters along the 9-m depth contour in the SBOO region.

An additional 69 offshore stations were sampled quarterly to monitor water quality conditions and to estimate dispersion of the PLOO and SBOO wastewater plumes. These stations were monitored during February, May, August, and November in 2020 with the 36 PLOO and 33 SBOO stations sampled over three to five days during each survey (Table 2.2). Stations F1–F36 are arranged in a grid surrounding the PLOO along or adjacent to the 18, 60, 80, and 98-m depth contours, while stations I1–I40 are arranged in a grid surrounding the SBOO along the 9, 19, 28, 38, and 55-m depth contours (Figure 2.1). Of these, 15 of the PLOO stations (i.e., F01–F03, F06–F14, F18–F20) and 15 of the SBOO stations (i.e., I12, I14, I16–I18, I22–I23, I27, I31, I33–I38) are located within State jurisdictional waters (i.e., within 3 nautical miles of shore) and therefore subject to Ocean Plan compliance standards.

Seawater samples for FIB analyses were collected from 3 to 5 discrete depths at the kelp and offshore stations as indicated in Table 2.3. These samples were typically collected using a rosette sampler fitted with Niskin bottles surrounding a central conductivity, temperature, and depth instrument (CTD), although replacement samples due to misfires or other causes may have been collected from a separate follow-up cast using stand-alone Van Dorn bottles if necessary. All weekly kelp/nearshore samples and quarterly offshore SBOO samples were analyzed for all three types of FIB, while the quarterly offshore PLOO samples were only analyzed for *Enterococcus* per permit requirements. All FIB samples were refrigerated at sea and then transported on blue ice to the City’s Marine Microbiology Lab for processing and analysis. Oceanographic data were collected simultaneously with the water samples at each station using the central CTD in the rosette sampler (see below). Visual observations of weather, sea conditions, and human/animal activity were also recorded at the time of collection. These observations have been previously reported in monthly receiving waters monitoring reports submitted to the SDRWQCB (see City of San Diego 2020–2021a,b), and are available online (City of San Diego 2021b).

Oceanographic data were collected using a SeaBird SBE 25 Plus CTD. The CTD was lowered through the water column at each station to collect continuous measurements of water temperature, conductivity (used to calculate salinity), pressure (used to calculate depth), dissolved oxygen (DO), pH, transmissivity (a proxy for water clarity), chlorophyll *a* fluorescence (a proxy for phytoplankton), and colored dissolved organic matter (CDOM). Vertical profiles of each parameter were constructed for each station, per survey, by averaging the data values recorded within each 1-m depth bin. This level of data reduction ensures that physical measurements used in subsequent analyses will correspond to discrete sampling depths required for bacterial monitoring.

Real-time oceanographic mooring systems

Two RTOMS were deployed at the terminal ends of the PLOO and SBOO (Figure 2.1). The PLOO RTOMS was anchored at a depth of approximately 100 m, just east of the northern diffuser leg, and the SBOO RTOMS was anchored at a depth of approximately 30 m, just west of the southern diffuser leg terminus. Each mooring was deployed for a period of approximately one year. The second PLOO

deployment occurred from October 7, 2019 to September 29, 2020, and the third SBOO deployment occurred from December 18, 2019 to December 17, 2020. Each RTOMS was outfitted with a series of instruments/sensors at fixed depths (Table 2.4). Critical parameters that were measured on a real-time basis, by both systems, included temperature, conductivity (salinity), total pH, DO, dissolved carbon dioxide ($x\text{CO}_2$), nitrogen (nitrate + nitrite), chlorophyll *a*, CDOM, biological oxygen demand (BOD), and current direction and velocity. Note that pH is reported in total scale from moored instruments with a more accurate calibration and measurement method for seawater, while pH has been reported in National Bureau of Standards (NBS) scale from CTD casts, and it is not recommended to convert between these scales (Marion et al. 2011). All parameters were recorded at 10-minute intervals, with the exception of nitrate + nitrite, which was recorded at 1-hour intervals, and $x\text{CO}_2$, which was recorded at 10-hour intervals. Equipment problems and sensor failures resulted in data gaps, and RTOMS data presented here include only data collected in real-time. For a summary of data issues and additional information on specific sensor issues and challenges experienced, see Addendum 2-1.

Laboratory Analyses

The City's Marine Microbiology Laboratory follows guidelines issued by the U.S. Environmental Protection Agency (USEPA) Water Quality Office, and the California Department of Public Health (CDPH), and Environmental Laboratory Accreditation Program (ELAP) with respect to sampling and analytical procedures (Bordner et al. 1978, APHA 2005, 2012, CDPH 2000, USEPA 2006). All bacterial analyses were initiated within eight hours of sample collection and conformed to standard membrane filtration techniques (APHA 2012).

Plates with FIB counts above or below the ideal counting range were given greater than (>), greater than or equal to (\geq), less than (<), or estimated (e) qualifiers. However, all qualifiers were dropped and densities treated as discrete values when determining compliance with Ocean Plan standards.

Quality assurance tests were performed routinely on bacterial samples to ensure that analyses and sampling variability did not exceed acceptable limits. Laboratory and field duplicate bacteriological samples were processed according to method requirements to measure analyst precision and variability between samples, respectively. Results of these procedures were reported in a separate report (City of San Diego 2021a).

Data Analyses

Oceanographic conditions

Water column parameters measured in 2020 were summarized as quarterly mean values, pooled over all stations, by the following depth layers: PLOO stations = 1–20 m, 21–60 m, 61–80 m, and 81–100 m; SBOO stations = 1–9 m, 10–19 m, 20–28 m, 29–38 m, and 39–55 m. Unless otherwise noted, analyses were performed using R (R Core Team, 2020) and various functions within the following packages: zoo, reshape2, Rmisc, gridExtra, mixOmics, fields, ggplot2, data.table, Hmisc, oce, RODBC, tidyr, dplyr (Zeileis and Grothendieck 2005, Wickham 2007, Hope 2013, Auguie 2017, Rohart et al. 2017, Nychka et al. 2017, Wickham et al. 2018, Dowle and Srinivasan 2020, Harrell et al. 2020, Kelley and Richards 2020, Ripley and Lapsley 2020, Wickham and Henry 2020, Wickham and Francois 2021).

Bacteriological compliance

Compliance with Ocean Plan water contact standards was summarized as the number of times per sampling period that each shore, kelp, and offshore station within State waters exceeded geometric mean or single sample maximum (SSM) standards for total coliforms, fecal coliforms, and *Enterococcus* (Table 2.1,

SWRCB 2015). Compliance calculations were limited to shore, kelp and offshore stations located within State waters, and included resamples. These analyses were performed using R (R Core Team, 2020) and various functions within the following packages: reshape2, Hmisc, RODBC, tidyr (Wickham 2007, Harrell et al. 2020, Ripley and Lapsley 2020, Wickham and Henry 2020).

Wastewater Plume Detection and Out-of-Range Calculations

Presence or absence of the wastewater plume at the PLOO and SBOO offshore stations was estimated by evaluation of a combination of oceanographic parameters (i.e., detection criteria). All stations along the 9-m depth contour were excluded from analyses due to the potential for coastal runoff or sediment resuspension in shallow nearshore waters to confound any CDOM signal that could be associated with plume dispersion from the outfalls. Previous monitoring results have consistently shown that the PLOO plume remains trapped below the pycnocline with no evidence of surfacing throughout the year (City of San Diego 2010a–2014a, 2015a,b, 2016a, 2018, 2020, Rogowski et al. 2012a, b, 2013, Hess 2019, Hess 2020). In contrast, the SBOO plume stays trapped below the pycnocline during seasonal periods of water column stratification, but may rise to the surface when waters become more mixed and stratification breaks down (City of San Diego 2010b–2014b, 2015c, 2016b, 2018, 2020, Terrill et al. 2009, Hess 2019, Hess 2020). Water column stratification and pycnocline depth were quantified using buoyancy frequency (BF, cycles/min) calculations for each quarterly survey. This measure of the water column’s static stability was used to quantify the magnitude of stratification for each survey and was calculated as follows:

$$BF = \sqrt{((g/\rho) \times (d\rho/dz))}$$

where g is the acceleration due to gravity, ρ is the seawater density, and $d\rho/dz$ is the density gradient (Mann and Lazier 1991). The depth of maximum BF was used as a proxy for the depth at which stratification was the greatest. If the water column was determined to be stratified (i.e., maximum BF > 5.5 cycles/min), subsequent analyses were limited to depths below the pycnocline.

Identification of potential plume signal was determined for each quarterly survey at each monitoring station on a combination of CDOM, chlorophyll *a* and salinity levels, as well as a visual review of the overall water column profile. Detection thresholds for the PLOO and SBOO stations were adaptively set for each quarterly sampling period according to the criteria described in City of San Diego (2016a, b). It should be noted that these thresholds are based on observations of ocean properties specific to the distinct PLOO and SBOO monitoring regions and are thus constrained to use within those regions. Finally, water column profiles were visually interpreted to remove stations with spurious signals (e.g., CDOM signals near the sea floor that were likely caused by resuspension of sediments). All analyses were performed using R (R Core Team, 2020) and the various functions within the following packages: reshape2, Rmisc, gridExtra, fields, mixOmics, gtools, oce, RODBC, tidyverse, pracma (Wickham 2007, Hope 2013, Auguie 2017, Nychka et al 2017, Rohart et al 2017, Warnes et al 2018, Kelley and Richards 2020, Ripley and Lapsley 2020, Wickham et al. 2020, Borchers 2021).

The effect of any potential “plume detection” on local water quality was evaluated by comparing mean values of DO, pH, and transmissivity within the possible plume boundaries to thresholds calculated for the same depths from reference stations. Stations with CDOM values below the 85th percentile were considered “reference” (Appendix A.1). Individual non-reference stations were then determined to be out-of-range (OOR) compared to the reference stations if values for the above parameters exceeded narrative water quality standards defined in the Ocean Plan (see Table 2.1). For example, the Ocean Plan defines OOR thresholds for DO as a 10% reduction from that which occurs naturally, for pH as a 0.2 pH unit change, and for transmissivity as below the

lower 95% confidence interval from the mean. For purposes of this report, “naturally” is defined for DO as the mean concentration minus one standard deviation (see Nezlin et al. 2016).

Real-time oceanographic mooring systems

Prior to conducting analyses, all data were subject to a comprehensive suite of quality assurance/quality control (QA/QC) procedures following Quality Assurance of Real-Time Oceanographic Data (QARTOD) methodologies (US IOOS 2017, 2020). Results of QARTOD tests are included in Addenda 2-2, 2-3, and 2-4; see City of San Diego 2020 for details. After review, all flagged data identified as suspect or bad, either manually or from automated tests, were excluded from further analyses and are not presented in this report. A detailed log of data flagged by parameter, site, depth, and date range is available upon request. When possible, additional QA/QC procedures involved analyzing quarterly CTD casts to validate data from RTOMS sensors, and seawater samples to validate nitrate + nitrite results.

Analyses were performed in R (R Development Core Team 2020) using functions within various packages: reshape2, lubridate, Rmisc, mixOmics, gtools, purrr, ggplot2, tidyverse, data.table, pracma, and dplyr (Wickham 2007, Grolemund and Wickham 2011, Hope 2013, Rohart et al. 2017, Warnes et al. 2018, Henry and Wickham 2019, Wickham et al. 2018, Wickham et al. 2019, Dowle and Srinivasan 2020, Borchers 2021, Wickham and Francois 2021). Annual time series of raw and daily-averaged data were plotted at each depth and site for all parameters that passed review, with the exception of ADCP data (ADCPs: Acoustic Doppler Current Profilers; described below). In addition, summary statistics were completed at each depth and site with the following seasonal periods that align with quarterly water quality sampling: winter (January–March); spring (April–June); summer (July–September); fall (October–December). Large data gaps were identified as seasons with <40% data recovery, based on expected number of samples for sensor-specific sampling intervals, and were excluded from summary analyses.

Ocean current data collected by downward-facing surface-mounted RTOMS ADCP instruments (Teledyne RD Instruments 300 kHz Workhorse Broadband) were checked for quality by eliminating those measurements that did not meet echo intensity criteria (i.e., minimum average intensity >100 counts and minimum correlation among the four beams of >70%). Following this initial screening, tidal frequency data were removed using the PL33 filter (Alessi et al. 1984) and compass direction was corrected to true north (+12.8 degrees). For all RTOMS deployments, ADCP data were summarized by season and select depth bins, as described above.

RESULTS

All CTD and bacterial water quality data and associated visual observations for 2020 have been previously reported in monthly receiving waters monitoring reports submitted to the San Diego Water Board and the USEPA (see City of San Diego 2020–2021a, b).

Oceanographic Conditions

Ocean temperature, salinity, DO, pH, transmissivity, and chlorophyll *a* data collected by CTD during 2020 in the PLOO and SBOO monitoring regions are summarized by depth layer for the entire year in Tables 2.5 and 2.6, and by depth layer for each survey in Appendices A.2 and A.3. These same parameters are plotted by depth and survey in Appendices A.4–A.15. Ocean temperature, salinity, DO, total pH, chlorophyll *a*, CDOM, turbidity, nitrate + nitrite, BOD, and xCO₂ data collected by PLOO and SBOO RTOMS during 2020 are summarized

by depth and season in Appendices A.16 and A.17. Ocean current velocity and magnitude are summarized by depth layer and season in Appendices A.18 and A.19. All parameters except current velocity are plotted over time in Appendix A.20.

Bacteriological Compliance

Compliance rates for all Ocean Plan water contact standards are summarized in Tables 2.7 and 2.8. All seawater samples collected from the PLOO and SBOO water quality stations that contained elevated FIB densities are listed in Appendices A.21 and A.22.

Plume Dispersion and Effects

The dispersion of wastewater plumes in 2020 and their effects on natural light, DO and pH levels in local ocean waters off San Diego were assessed by evaluating the results of 164 CTD profile casts performed at the PLOO stations and another 116 CTD casts performed at the SBOO stations. CDOM is plotted by depth and survey in Appendix A.23 and A.24. Potential plume detection results are summarized in Appendices A.25 and A.26.

SUMMARY

During 2020, oceanographic conditions off San Diego were generally within historical ranges reported for the PLOO and SBOO monitoring regions. However, an unprecedented, large spring phytoplankton bloom resulted in higher than normal DO, pH, and chlorophyll *a*, and record low $x\text{CO}_2$, observed near the surface by CTD profiles and RTOMS in April/May 2020, corresponding to a regional red tide event (Anderson and Hepner-Medina, 2020). Conditions typically indicative of coastal upwelling were most evident during the spring months, while maximum stratification or layering of the water column occurred during mid-summer, after which the waters became more mixed in the winter. Decreases in water clarity or transmissivity tended to be associated with terrestrial runoff or outflows from rivers and bays, the re-suspension of nearshore bottom sediments due to waves or storm activity, and to the presence of strong and sustained phytoplankton blooms, particularly in the spring.

The detection of the PLOO and SBOO wastewater plumes and their effects on various water quality indicators, such as natural light levels, DO concentrations, and pH, were low in 2020. Additionally, water quality conditions were mostly consistent with data reported previously for both regions. Compliance with both the SSM and geometric mean standards was higher in the PLOO region, and at kelp and offshore stations compared to stations along the shore. Samples collected from SBOO shore stations showed notably lower compliance rates this year, particularly in March and April 2020. These data coincide with unusually high transboundary flows from the Tijuana River reported for this period (IBWC, 2020). Reduced compliance in both outfall regions tends to occur during the wet season and is significantly influenced by terrestrial outflows, such as the Tijuana River. Finally, there was no evidence that wastewater discharged into the ocean via either outfall reached nearshore recreational waters.

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CHAPTER 2
FIGURES & TABLES

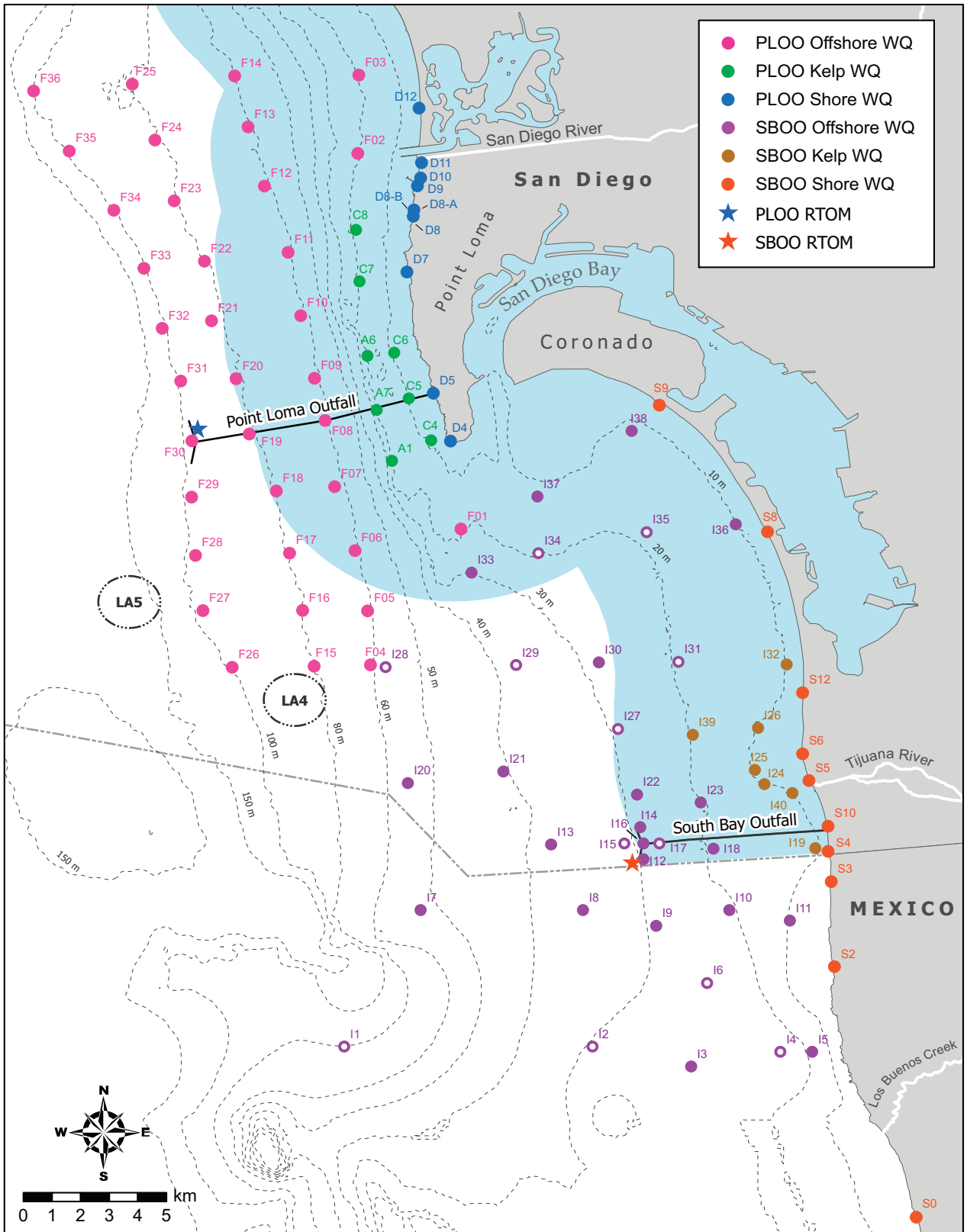


Figure 2.1

Water quality (WQ) monitoring station locations sampled around the PLOO and SBOO as part of the City of San Diego's Ocean Monitoring Program. Light blue shading represents State of California jurisdictional waters. Open circles are sampled by CTD only.

Table 2.1

Water quality objectives for water-contact areas, California Ocean Plan (SWRCB 2015).

A. Bacterial Characteristics – Water Contact Standards; CFU = colony forming units

(a) 30-day Geometric Mean - The following standards are based on the geometric mean of the five most recent samples from each site:

- 1) Total coliform density shall not exceed 1000 CFU/100 mL
- 2) Fecal coliform density shall not exceed 200 CFU/100 mL
- 3) *Enterococcus* density shall not exceed 35 CFU/100 mL

(b) Single Sample Maximum:

- 1) Total coliform density shall not exceed 10,000 CFU/100 mL
- 2) Fecal coliform density shall not exceed 400 CFU/100 mL
- 3) *Enterococcus* density shall not exceed 104 CFU/100 mL
- 4) Total coliform density shall not exceed 1000 CFU/100 mL when the fecal coliform:total coliform ratio exceeds 0.1

B. Physical Characteristics

- (a) Floating particulates and oil and grease shall not be visible
- (b) The discharge of waste shall not cause aesthetically undesirable discoloration of the ocean surface
- (c) Natural light shall not be significantly reduced at any point outside of the initial dilution zone as the result of the discharge of waste

C. Chemical Characteristics

- (a) The dissolved oxygen concentration shall not at any time be depressed more than 10 percent from what occurs naturally, as a result of the discharge of oxygen demanding waste materials
 - (b) The pH shall not be changed at any time more than 0.2 units from that which occurs naturally
-

Table 2.2

Sample dates for quarterly oceanographic surveys conducted during 2020. All stations in each station group were sampled on a single day (see Figure 2.1 for stations and locations).

PLOO Sampling Dates					SBOO Sampling Dates				
Station Group	Feb	May	Aug	Nov	Station Group	Feb	May	Aug	Nov
Kelp WQ	18	18	17	11	Kelp WQ	10	26	27	2
18 & 60-m WQ	20	22	19	12	North WQ	14	29	25	5
80-m WQ	21	21	20	13	Mid WQ	13	28	24	4
98-m WQ	19	20	18	10	South WQ	12	27	26	3

Table 2.3

Depths from which seawater samples are collected for bacteriological analysis from kelp and offshore stations.

Station Contour	PLOO Sample Depth (m)									Station Contour	SBOO Sample Depth (m)							
	1	3	9	12	18	25	60	80	98		2	6	9/11	12	18	27	37	55
<i>Kelp Bed</i>										<i>Kelp Bed</i>								
9-m	x	x	x							9-m	x	x	x ^a					
18-m	x			x	x					19-m	x			x	x			
<i>Offshore</i>										<i>Offshore</i>								
18-m	x			x	x					9-m	x	x	x ^a					
60-m	x					x	x			19-m	x			x	x			
80-m	x					x	x	x		28-m	x				x	x		
98-m	x					x	x	x	x	38-m	x				x			x
										55-m	x				x			x

^a Stations I25, I26, I32, and I40 sampled at 9 m; stations I11, I19, I24, I36, I37, and I38 sampled at 11 m

Table 2.4

Sensor configuration and model type for RTOMS by site and depth during 2020.

Sensor Depth		Parameters Measured (Sensor Types)
PLOO	SBOO	
1 m (surface)	1 m (surface)	Temperature, conductivity, pH (total), DO (Sea-Bird SeapHOx) Ocean currents (RDI 300kHz ADCP) Partial pressure of carbon dioxide (Pro-Oceanus pCO ₂ System) Chlorophyll a, CDOM, turbidity (Sea-Bird ECO triplet) Nitrate + nitrite (Sea-Bird SUNA V2)
10 m	10 m	Temperature, conductivity (Sea-Bird MicroCAT)
	18 m	Temperature, conductivity, DO (Sea-Bird MicroCAT ODO) Chlorophyll a, CDOM, turbidity (Sea-Bird ECO triplet)
20 m		Temperature, conductivity (Sea-Bird MicroCAT)
	26 m (cage)	Temperature, conductivity, pH, DO (Sea-Bird SeapHOx) Chlorophyll a, CDOM, turbidity (Sea-Bird ECO triplet) Nitrate + nitrite (Sea-Bird SUNA V2) BOD (Chelsea UviLux)
30 m (cage-1)		Temperature, conductivity, pH, DO (Sea-Bird SeapHOx) Chlorophyll a, CDOM, turbidity (Sea-Bird ECO triplet) BOD (Chelsea UviLux)
45 m		Temperature, conductivity (Sea-Bird MicroCAT)
60 m		Temperature, conductivity (Sea-Bird MicroCAT)
75 m		Temperature, conductivity (Sea-Bird MicroCAT)
90 m (cage-2)		Temperature, conductivity, pH, DO (Sea-Bird Deep SeapHOx) Chlorophyll a, CDOM, turbidity (Sea-Bird ECO triplet) Nitrate + nitrite (Sea-Bird SUNA V2) BOD (Chelsea UviLux)

Table 2.5

Summary of temperature, salinity, dissolved oxygen (DO), pH, transmissivity, and chlorophyll *a* for various depth layers as well as the entire water column for all PLOO stations during 2020. See Appendix A.2 for sample sizes.

Parameter		Depth (m)				
		1–20	21–60	61–80	81–98	1–98
Temperature (°C)	min	11.2	10.1	10.0	9.7	9.7
	max	23.9	16.4	13.1	12.1	23.9
	mean	16.1	12.6	11.3	10.9	13.3
Salinity (ppt)	min	33.31	33.31	33.48	33.58	33.31
	max	33.98	33.92	33.96	34.04	34.04
	mean	33.59	33.59	33.71	33.80	33.63
DO (mg/L)	min	1.9	2.9	3.0	3.1	1.9
	max	9.3	8.5	6.8	6.3	9.3
	mean	7.1	6.0	4.8	4.2	6.0
pH	min	7.6	7.7	7.7	7.7	7.6
	max	8.4	8.2	8.0	7.9	8.4
	mean	8.1	7.9	7.8	7.8	8.0
Transmissivity (%)	min	22	84	28	55	22
	max	93	94	94	94	94
	mean	85	91	91	90	89
Chlorophyll <i>a</i> (µg/L)	min	0.2	0.1	0.1	0.1	0.1
	max	31.8	5.1	3.7	0.7	31.8
	mean	2.3	1.1	0.4	0.3	1.3

Table 2.6

Summary of temperature, salinity, dissolved oxygen (DO), pH, transmissivity, and chlorophyll *a* for various depth layers as well as the entire water column for all SBOO stations during 2020. See Appendix A.3 for sample sizes.

Parameter		Depth (m)					
		1–9	10–19	20–28	29–38	39–55	1–55
Temperature (°C)	min	12.2	11.6	11.4	11.3	10.5	10.5
	max	23.8	20.7	17.6	15.4	15.1	23.8
	mean	17.3	15.0	14.0	13.4	12.8	15.3
Salinity (ppt)	min	33.27	33.27	33.38	33.38	33.41	33.27
	max	34.28	33.84	33.76	33.76	33.87	34.28
	mean	33.57	33.54	33.54	33.54	33.58	33.55
DO (mg/L)	min	1.3	0.6	4.0	3.9	3.5	0.6
	max	9.9	9.5	8.7	8.5	8.2	9.9
	mean	7.8	7.3	6.8	6.5	6.0	7.2
pH	min	7.6	7.5	7.8	7.8	7.7	7.5
	max	8.3	8.3	8.2	8.1	8.1	8.3
	mean	8.1	8.1	8.0	8.0	7.9	8.1
Transmissivity (%)	min	47	41	73	84	88	41
	max	92	92	92	93	93	93
	mean	81	85	88	90	91	85
Chlorophyll <i>a</i> (µg/L)	min	0.2	0.3	0.4	0.5	0.3	0.2
	max	12.3	7.9	5.4	5.0	3.7	12.3
	mean	2.1	2.3	2.2	1.7	1.1	2.1

Table 2.7

Compliance rates for the three geometric mean water-contact standards and for the four single sample maximum water contact standards for PLOO monitoring stations sampled during 2020. PLOO offshore stations are sampled quarterly, and total and fecal coliform bacteria are not analyzed at these stations; ns = not sampled.

	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
Geometric Mean												
<i>Shore Stations</i>												
Total	100	100	100	100	100	100	100	100	100	100	100	100
Fecal	100	100	100	100	100	100	100	100	100	100	100	100
Enterococci	99	100	100	100	100	100	100	100	100	100	100	97
<i>Kelp Stations</i>												
Total	100	100	100	100	100	100	100	100	100	100	100	100
Fecal	100	100	100	100	100	100	100	100	100	100	100	100
Enterococci	100	100	100	100	100	100	100	100	100	100	100	100
Single Sample Maximum												
<i>Shore Stations</i>												
Total	100	100	100	100	100	100	100	100	100	100	100	100
Fecal	100	100	100	100	100	100	100	100	100	100	100	100
Enterococci	98	100	94	98	100	100	100	100	100	97	100	100
F:T	100	100	100	100	100	100	100	100	100	100	100	100
<i>Kelp Stations</i>												
Total	100	100	100	100	100	100	100	100	100	100	100	100
Fecal	100	100	100	100	100	100	100	100	100	100	100	100
Enterococci	100	100	100	100	100	100	100	100	100	100	100	100
F:T	100	100	100	100	100	100	100	100	100	100	100	100
<i>Offshore Stations</i>												
Enterococci	ns	100	ns	ns	98	ns	ns	98	ns	ns	94	ns

Table 2.8

Compliance rates for the three geometric mean water-contact standards and the four single sample maximum water contact standards for SBOO monitoring stations sampled during 2020. SBOO offshore stations are sampled quarterly; ns=not sampled.

	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec
Geometric Mean												
<i>Shore Stations</i>												
Total	52	63	50	27	54	77	77	88	100	100	100	100
Fecal	52	63	42	23	53	81	76	91	100	100	100	100
Enterococci	30	63	29	4	19	60	60	82	100	100	100	100
<i>Kelp Stations</i>												
Total	68	86	94	69	89	100	100	100	100	100	100	100
Fecal	76	86	93	70	84	100	100	100	100	100	100	100
Enterococci	48	60	83	48	46	100	100	89	100	100	100	100
Single Sample Maximum												
<i>Shore Stations</i>												
Total	73	70	48	43	70	70	78	89	98	100	92	100
Fecal	54	56	37	28	53	70	64	87	87	100	89	100
Enterococci	47	40	35	12	46	57	58	86	87	100	91	100
F:T	52	50	43	27	54	66	69	84	87	100	89	100
<i>Kelp Stations</i>												
Total	90	99	90	90	100	100	100	97	90	100	99	100
Fecal	76	85	80	70	98	100	100	89	81	100	96	100
Enterococci	68	81	87	50	93	100	95	91	81	100	100	100
F:T	72	85	76	68	99	100	99	90	86	100	95	100
<i>Offshore Stations</i>												
Total	ns	100	ns	ns	100	ns	ns	100	ns	ns	100	ns
Fecal	ns	100	ns	ns	100	ns	ns	100	ns	ns	100	ns
Enterococci	ns	100	ns	ns	100	ns	ns	100	ns	ns	100	ns
F:T	ns	100	ns	ns	100	ns	ns	100	ns	ns	100	ns

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Chapter 3

Benthic Conditions

Chapter 3. Benthic Conditions

INTRODUCTION

The City of San Diego conducts extensive monitoring of benthic sediments and communities of small benthic invertebrates (macrofauna) that live within, or on the surface of, soft-bottom seafloor habitats surrounding the Point Loma and South Bay Ocean Outfalls (PLOO and SBOO, respectively). This monitoring helps to characterize regional benthic conditions and identify potential effects of wastewater discharge, or other anthropogenic inputs, on the marine benthic environment. This chapter presents summaries and preliminary analyses of the sediment quality, sediment toxicity, and macrofaunal community data collected during calendar year 2020 at PLOO, SBOO, and San Diego regional benthic monitoring stations. Raw data summaries supporting these results are presented in Appendix B. A more comprehensive assessment of these results will be presented in the 2020-2021 Biennial Assessment Report to be submitted by July 1, 2022.

MATERIALS AND METHODS

Collection and Processing of Samples

Samples were collected at a total of 89 benthic stations to monitor ocean sediments and macrofaunal communities during 2020 (Figure 3.1). These included 22 stations arranged in a grid surrounding the PLOO, along or adjacent to the 88, 98, or 116-m depth contours, and 27 stations arranged in a grid surrounding the SBOO, along or adjacent to the 19, 28, 38, or 55-m depth contours. These stations were sampled during the winter (January) and summer (July). The four stations located within 1000 m of the zone of initial dilution (ZID) for each outfall are considered to represent near-ZID conditions. These include PLOO stations E11, E14, E15, and E17, and SBOO stations I12, I14, I15, and I16. The remaining 40 “regional” stations were selected using a probability-based random stratified sampling design as described in Bergen (1996), Stevens (1997), and Stevens and Olsen (2004). Regional stations were sampled during the summer at depths ranging from 7 to 523 m, including 11 sites along the inner shelf (7–30 m), 17 sites along the mid-shelf (30–120 m), six sites along the outer shelf (120–200 m), and six sites on the upper slope (200–523 m).

Samples were collected using a double 0.1-m² Van Veen grab, with one grab per cast used for sediment quality analysis, one grab per cast used for benthic community analysis, and subsequent grabs used for sediment toxicity testing where required. Visual observations of weather, sea conditions, and human/animal activity were also recorded at the time of sampling (see Addenda 1-1A, 1-1B, 1-1C). Criteria established by the U.S. Environmental Protection Agency (USEPA) to ensure consistency of these types of samples were followed with regard to sample disturbance and depth of penetration (USEPA 1987). Sub-samples for particle size and sediment chemistry analyses were taken from the top 2 cm of the sediment surface and handled according to standard guidelines (USEPA 1987, SCCWRP 2018).

For sediment toxicity samples, a plastic (e.g., high-density polyethylene [HDPE], polycarbonate, Teflon) or stainless-steel scoop was used to collect sediment from the top 2 cm of the undisturbed surface material in the grab. Contact with sediment within 1 cm of the sides of the grab was avoided to minimize cross-contamination. In most cases, multiple grabs were required to obtain enough sediment for toxicity testing (i.e., up to 6 L sediment). If more than one grab was required, sediment from each grab was added to a

Teflon bag and homogenized thoroughly using either a clean Teflon or plastic spoon, or by kneading the sample within the bag. Once collected, the toxicity samples were stored in the dark at 4°C in the laboratory for no longer than four weeks prior to testing.

Samples for infauna analysis were transferred to a wash table aboard ship, rinsed with seawater, and then sieved through a 1.0-mm mesh screen to remove as much sediment as possible. The macroinvertebrates retained on the screen were transferred to sample jars, relaxed for 30 minutes in a magnesium sulfate solution, and then fixed with buffered formalin. The preserved samples were then transferred back to the City's Marine Biology Laboratory. After a minimum of 72 hours, but no more than 10 days, in formalin, each sample was thoroughly rinsed with fresh water and transferred to 70% ethanol for final preservation.

Laboratory Analyses

Sediment Particle Size

All sediment chemistry and particle size analyses were performed at the City's Environmental Chemistry Services Laboratory. Particle size analysis was performed using either a Horiba LA-950V2 Laser Particle Size Analyzer or a set of nested sieves. The Horiba measures particles ranging in size from 0.5 to 2000 µm. Coarser sediments were removed and quantified prior to laser analysis by screening samples through a 2000 µm mesh sieve. These data were later combined with the Horiba results to obtain a complete distribution of particle sizes totaling 100%, and then classified into 11 sub-fractions and four main size fractions based on the Wentworth scale (Folk 1980) (see Appendix B.1). When a sample contained substantial amounts of coarse sand, gravel, shell hash, or other large materials that could damage the Horiba analyzer or where the general distribution of sediments would be poorly represented by laser analysis, a set of nested sieves with mesh sizes of 2000, 1000, 500, 250, 125, 75, and 63 µm was used to divide the samples into eight sub-fractions.

Sediment Chemistry

A detailed description of the analytical protocols can be found in City of San Diego (2021). Briefly, sediment sub-samples were analyzed on a dry weight basis to determine concentrations of various indicators of organic loading (i.e., biochemical oxygen demand, total organic carbon, total nitrogen, total sulfides, total volatile solids), 18 trace metals, 9 chlorinated pesticides, 42 polychlorinated biphenyl compound congeners (PCBs), and 24 polycyclic aromatic hydrocarbons (PAHs). These data were generally limited to values above the method detection limit (MDL) for each parameter (see Appendix B.2). However, concentrations below MDLs were included as estimated values if presence of the specific constituent was verified by mass spectrometry.

Sediment Toxicity Testing

A detailed description of the sediment toxicity testing protocols can be found in City of San Diego (2020b). Briefly, all sediment toxicity testing was conducted by the City of San Diego Toxicology Laboratory (CSDTL) using the marine amphipod *Eohaustorius estuarius*. The 10-day amphipod tests were conducted in accordance with EPA 600/R-94/0925 (USEPA 1994) and the procedures approved for Southern California Bight 2018 Regional Monitoring Program (Parks et al. 2020). Juvenile *E. estuarius* were exposed for 10 days to both test and control sediments. Response criteria included amphipod mortality, emergence from sediment during exposure, and, if considered a measurement of interest, the ability of amphipods to reburial in clean sediment at the end of the bioassay. In addition, a reference toxicant test (using seawater only) was conducted concurrently and under identical environmental conditions as the sediment toxicity tests to determine test organism sensitivity.

Macrobenthic Assemblages

All organisms were separated from the raw material (e.g., sediment grunge, shell hash, debris) and sorted into the following six taxonomic groups by an external contract lab: Annelids (e.g., polychaete and oligochaete worms), Arthropods (e.g., crustaceans and pycnogonids), Molluscs (e.g., clams, snails, scaphopods), non-ophiuroid Echinoderms (e.g., sea urchins, sea stars, sea cucumbers), Ophiuroids (i.e., brittle stars), and other phyla (e.g., flatworms, nemerteans, cnidarians). The sorted macrofaunal samples were then returned to the City's Marine Biology Laboratory where all animals were identified to species, or to the lowest taxon possible, by City Marine Biologists. All identifications followed nomenclatural standards established by the Southern California Association of Marine Invertebrate Taxonomists (SCAMIT 2018).

Data Analyses

Sediment Particle Size and Chemistry

Data summaries for the various sediment parameters included detection rate, mean, minimum and maximum values for all samples by outfall region (i.e., PLOO, SBOO stations) and across the region (i.e., regional stations). All means were calculated using detected values only with no substitutions made for non-detects (i.e., concentrations < MDL). Total DDT (tDDT), total hexachlorocyclohexane (tHCH), total chlordane, total PCB (tPCB), and total PAH (tPAH) were calculated for each sample as the sum of all constituents with reported values for individual constituents. Contaminant concentrations were compared to the Effects Range Low (ERL) and Effects Range Median (ERM) sediment quality guidelines of Long et al. (1995) when available. ERLs represent chemical concentrations below which adverse biological effects are rarely observed, while values above ERLs, but below ERMs, represent levels at which effects occasionally occur. Concentrations above the ERM indicate likely biological effects, although these may not always be validated by toxicity testing (Schiff and Gossett 1998). Analyses were performed using R (R Core Team 2020) and various functions within the dplyr, plyr, reshape2, tidyr, and zoo packages (Zeileis and Grothendieck 2005, Wickham 2007, 2011, Wickham and Henry 2017, Wickham et al. 2017).

Sediment Toxicity Testing

All data were analyzed in accordance with procedures outlined in Sections 12 and 13 of EPA 600/R-94/0925 using the acceptability criterion of $\geq 90\%$ mean control survival at test termination. Additional information and the standard operation procedures for sediment toxicity testing are provided in Appendix B of the CSDTL's Quality Assurance Manual (City of San Diego 2020b).

Macrobenthic Assemblages

Population characteristics were summarized as percent abundance (number of individuals per species/total abundance of all species), frequency of occurrence (percentage of grabs in which a species occurred), and mean abundance per grab (number of individuals per species/total number of grabs). Additionally, the following community structure parameters were calculated for each station and expressed per 0.1-m² grab: species richness (number of species or distinct taxa), abundance (number of individuals), Shannon diversity index (H'), Pielou's evenness index (J'), Swartz dominance index (see Swartz et al. 1986, Ferraro et al. 1994), and benthic response index (BRI; see Smith et al. 2001). Unless otherwise noted, the above analyses were performed using R (R Core Team 2020) and various functions within the reshape2, Rmisc, RODBC, tidyverse, and vegan packages (Wickham 2007, 2017, Hope 2013, Oksanen et al. 2017, Ripley and Lapsley 2017).

Multivariate analyses were performed using PRIMER v7 software to examine spatial and temporal patterns in macrofaunal data collected at the 89 PLOO, SBOO, and regional benthic stations sampled during summer 2020 (see Clarke et al. 2008, Clarke et al. 2014). These included ordination and hierarchical agglomerative clustering (cluster analysis) with group-average linking and similarity profile analysis (SIMPROF) to confirm the non-random structure of the resultant cluster dendrograms. The Bray-Curtis measure of similarity was used as the basis for clustering, and data were square-root transformed to lessen the influence of overly abundant species and increase the importance (or impact) of rare species. Major ecologically-relevant clusters receiving SIMPROF support were retained.

RESULTS

Sediment Quality

Sediment grain size (i.e., main particle size fractions) and chemistry data are summarized for PLOO, SBOO, and regional benthic stations in Tables 3.1 and 3.2. Sediment particle size composition is described for each sample in Appendices B.3, B.4, and B.5. Concentrations of organic loading indicators are listed by sample in Appendices B.6, B.7, and B.8, trace metals are listed by sample in B.9, B.10, and B.11, pesticides are listed by sample in B.12, B.13, and B.14, PCBs are listed by sample in B.15, B.16, and B.17, and PAHs are listed by sample in B.18, B.19, and B.20. Results for sediment toxicity samples collected during 2020 are summarized in Table 3.3.

Macrobenthic Communities

Key community structure parameters, including species richness, abundance, diversity, evenness, dominance, and BRI, are summarized for PLOO, SBOO, and regional benthic stations in Table 3.4. These same parameters are listed by sample in Appendices B.21, B.22, and B.23. The 25 most abundant macroinvertebrate taxa identified at PLOO and SBOO stations during 2020 are summarized by percent abundance, frequency of occurrence, and abundance per grab in Tables 3.5 and 3.6. The 10 most abundant taxa from each depth stratum are summarized for regional stations in Table 3.7. Total numbers of each individual taxon encountered are listed in Appendices B.24, B.25, and B.26. Ordination and cluster analyses were performed to illustrate and quantify the ecological patterns at the macroinvertebrate community level across the San Diego region; these results are presented in Figure 3.2.

SUMMARY

Preliminary analysis of sediment particle size, chemistry, toxicity, and macroinvertebrate data collected in 2020 indicate that wastewater discharged through the PLOO and SBOO has not negatively impacted benthic communities in the coastal waters off San Diego. During the current reporting period, there was no evidence of fine-particle loading related to wastewater discharge via the PLOO or SBOO. Contaminant concentrations at near-ZID stations were generally within the range of variability observed throughout both outfall regions and did not appear to reflect any significant organic enrichment. The quality of PLOO and SBOO sediments in 2020 was similar to previous years (e.g., City of San Diego 2020a), with overall contaminant concentrations remaining relatively low compared to available thresholds or other southern California coastal areas (Schiff and Gossett 1998, Noblet et al. 2002, Schiff et al. 2006, 2011, Maruya and Schiff 2009, Dodder et al. 2016). No evidence of sediment toxicity was observed at any offshore station tested in the San Diego region during 2020, regardless of depth, sediment type, or proximity to either outfall. These results are consistent with findings from previous regional monitoring programs that

have demonstrated minimal sediment toxicity on the southern California continental shelf in contrast to offshore submarine canyons and local embayments (e.g., Bay et al. 2015, Parks et al. 2020). Further, values for most benthic infauna community parameters were similar at stations located both near and far away from the outfall discharge sites. These metrics were within historical ranges reported for the San Diego region (e.g., City of San Diego 2020), and were representative of those characteristic of similar habitats throughout the Southern California Bight (Barnard and Zieshenne 1961, Jones 1969, Fauchald and Jones 1979, Thompson et al. 1987, 1993a,b, Zmarzly et al. 1994, Diener and Fuller 1995, Bergen et al. 1998, 2000, 2001, Ranasinghe et al. 2003, 2007, 2010, 2012, Mikel et al. 2007, Gillett et al. 2017).

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CHAPTER 3
FIGURES & TABLES

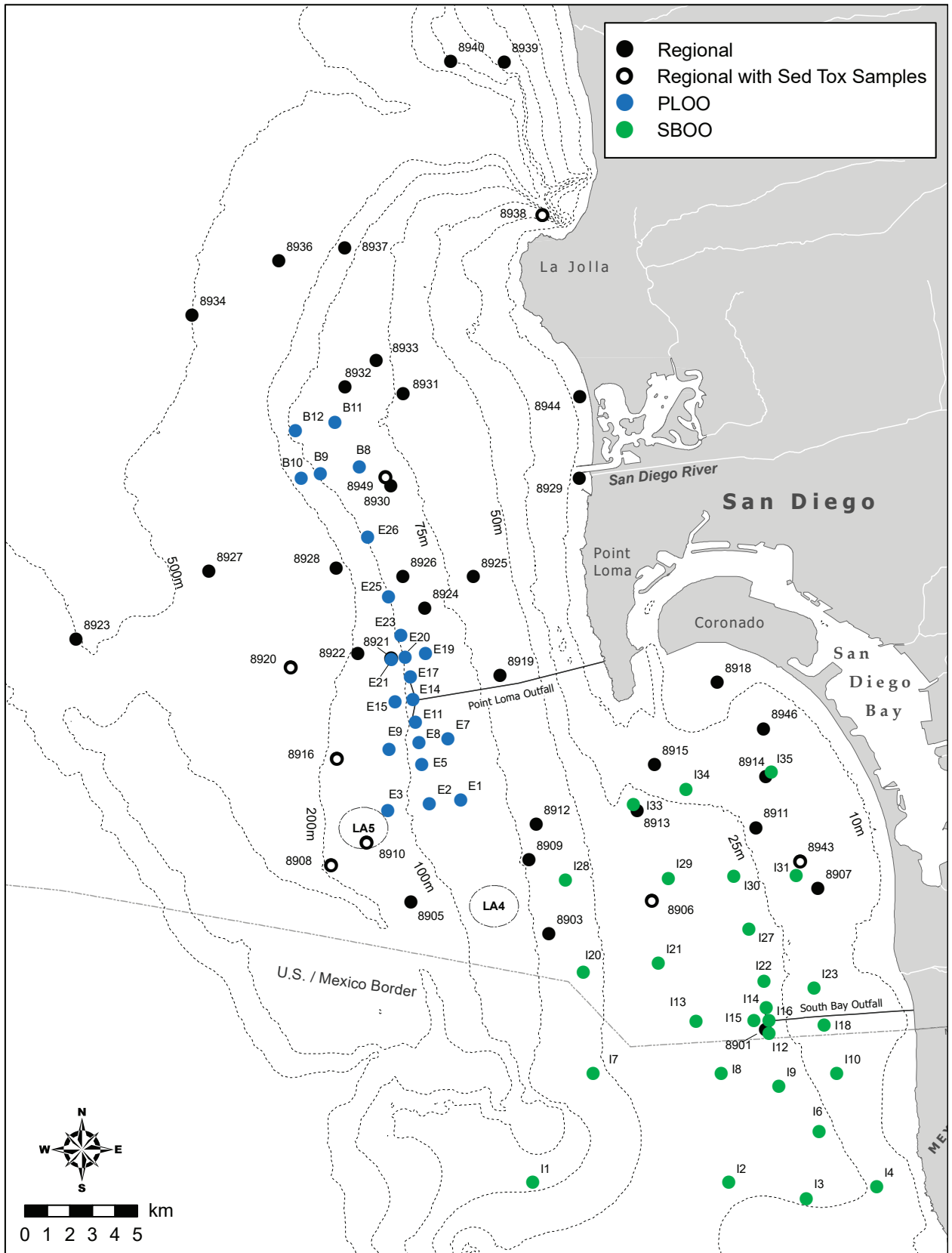


Figure 3.1
 Benthic station locations sampled around the PLOO and SBOO as part of the City of San Diego's Ocean Monitoring Program.

Table 3.1

Summary of particle sizes and chemistry concentrations in sediments from PLOO and SBOO benthic stations sampled during 2020. Data include the detection rate (DR), mean, minimum, and maximum values for each survey area. Minimum and maximum values were calculated using all samples, whereas means were calculated on detected values only; n = number of samples; nd = not detected; na = not analyzed.

Parameter	PLOO				SBOO			
	DR (%)	Mean	Min	Max	DR (%)	Mean	Min	Max
Particle Size (%)								
Coarse Particles	14	0.0	38.6	11.6	41	0.0	46.1	7.8
Med-Coarse Sands	100	0.1	26.0	2.6	94	0.0	92.3	29.2
Fine sands	100	24.3	63.1	39.2	98	0.0	89.2	38.3
Fines	100	17.3	73.1	56.6	96	0.0	100.1	32.9
Organic Indicators								
BOD (ppm)	100	187	546	263	na	—	—	—
Sulfides (ppm)	86	nd	30.80	5.14	46	nd	28.90	5.47
TN (% weight)	100	0.032	0.113	0.051	43	nd	0.062	0.027
TOC (% weight)	100	0.28	3.49	0.77	83	nd	1.15	0.24
TVS (% weight)	100	1.2	3.7	2.1	100	0.4	1.7	0.9
Trace Metals (ppm)								
Aluminum	100	4440	11,000	6847	100	597	7220	3278
Antimony	100	0.8	2.9	1.1	96	nd	1.3	0.6
Arsenic	100	1.79	5.72	2.90	100	0.84	8.95	2.50
Barium	100	13.1	54.1	27.2	100	0.3	38.9	15.1
Beryllium	100	0.1	0.3	0.2	100	0.0	0.1	0.1
Cadmium	50	nd	0.08	0.07	22	nd	0.07	0.04
Chromium	100	10.0	23.9	15.1	100	2.3	13.4	8.4
Copper	100	4.2	17.0	6.6	50	nd	4.9	2.8
Iron	100	6300	18,900	10,369	100	1190	8820	4954
Lead	100	1.9	7.5	3.3	100	0.8	3.2	1.6
Manganese	100	51.3	138.0	81.1	100	6.1	101.0	44.0
Mercury	100	0.008	0.057	0.025	41	nd	0.020	0.009
Nickel	100	3.8	10.0	5.9	100	0.5	5.0	2.0
Selenium	20	nd	0.54	0.32	0	—	—	—
Silver	0	—	—	—	0	—	—	—
Thallium	0	—	—	—	0	—	—	—
Tin	100	0.5	1.5	0.8	100	0.1	1.2	0.4
Zinc	100	16.8	52.5	25.7	100	1.7	41.3	11.0
Pesticides (ppt)								
Total Chlordane	34	nd	248	65	17	nd	169	49
Total DDT	100	93	1213	532	74	nd	3493	281
Dieldrin	2	nd	9	9	2	nd	10	10
Endosulfan Sulfate	2	nd	13	13	0	—	—	—
Hexachlorobenzene	23	nd	167	77	9	nd	179	78
Total HCH	7	nd	19	14	22	nd	218	40
Mirex	0	—	—	—	2	nd	29	29
Total PCB (ppt)	100	95	4563	977	37	nd	1881	351
Total PAH (ppb)	95	nd	199	43	48	nd	53	13

Table 3.2

Summary of particle sizes and chemistry concentrations in sediments from San Diego regional benthic stations sampled during summer 2020. Data include detection rate (DR), minimum, maximum, and mean values for the entire survey area, as well as mean value by depth stratum. Minimum and maximum values were calculated using all samples, whereas means were calculated on detected values only; n=number of samples; nd=not detected.

Parameters	2020 Survey Area				Depth Strata			
	DR (%)	Min	Max	Mean	Inner Shelf n=11	Mid-Shelf n=17	Outer Shelf n=6	Upper Slope n=6
Particle Size (%)								
Coarse particles	13	0.0	14.2	8.1	4.5	10.5	0.0	0.0
Med-coarse sands	100	0.1	77.0	7.0	9.0	10.2	1.2	0.1
Fine sands	100	11.4	84.2	41.1	62.9	35.8	35.4	21.9
Fines	100	7.4	87.7	50.9	27.3	52.1	63.5	78.0
Organic Indicators								
Sulfides (ppm)	95	nd	42.90	13.57	16.41	9.84	13.51	18.83
TN (% weight)	85	nd	0.270	0.081	0.026	0.058	0.085	0.189
TOC (% weight)	98	nd	3.39	0.92	0.16	0.61	1.35	2.63
TVS (% weight)	100	0.5	9.5	2.9	0.9	2.3	3.7	7.3
Trace Metals (ppm)								
Aluminum	100	1390	15,700	6859	3238	6787	8548	12,008
Antimony	95	nd	1.4	0.6	0.3	0.6	0.7	1.0
Arsenic	100	0.83	3.83	2.05	1.36	2.33	2.42	2.15
Barium	100	2.1	120.0	36.4	18.1	31.2	44.0	77.3
Beryllium	50	nd	0.4	0.1	nd	0.1	0.2	0.3
Cadmium	25	nd	0.18	0.08	0.05	0.07	0.13	nd
Chromium	100	4.2	41.9	16.4	7.4	15.5	20.1	32.0
Copper	93	nd	34.5	8.7	2.4	6.8	15.6	16.3
Iron	100	3130	24,100	10,598	4728	10,582	14,203	17,800
Lead	100	0.9	72.7	5.3	1.5	3.9	16.7	4.6
Manganese	100	13.2	158.0	83.6	53.5	82.6	100.4	124.9
Mercury	83	nd	0.107	0.043	0.009	0.039	0.063	0.061
Nickel	100	0.9	21.7	7.0	2.0	6.1	9.2	16.4
Selenium	18	nd	0.87	0.55	nd	0.29	0.44	0.67
Silver	0	—	—	—	—	—	—	—
Thallium	20	nd	0.223	0.177	0.153	0.199	0.211	0.173
Tin	75	nd	2.8	0.8	0.3	0.8	1.4	1.0
Zinc	100	7.1	79.2	28.6	12.2	26.4	43.3	50.2
Pesticides (ppt)								
Total Chlordane	48	nd	918	107	81	80	211	49
Total DDT	98	nd	4231	940	80	637	1219	2951
Dieldrin	8	nd	66	62	56	66	65	nd
Hexachlorobenzene	25	nd	71	46	nd	40	50	51
Total HCH	10	nd	131	63	89	131	12	20
Mirex	8	nd	41	27	21	21	nd	41
Total PCB (ppt)	77	nd	13,651	1876	116	1733	3560	1428
Total PAH (ppb)	90	nd	318	62	14	34	140	121

Table 3.3

Bioassay results (10-day amphipod survival tests) for sediment toxicity testing conducted for San Diego regional benthic stations sampled during summer 2020. Percent fines = percentage of silt + clay combined. Test results are expressed as mean percent survival \pm 1 standard deviation.

Survey	Site/Sample	Depth Stratum	Station Depth (m)	Percent Fines	Sample Date	Test Initiation	% Survival (Mean \pm SD)
Summer 2020	Lab Control	—	—	—	—	7/21/20	95 \pm 6.1
	8943	Inner Shelf	19	37.0	7/13/2020	7/21/2020	98 \pm 2.7
	8938	Inner Shelf	28	54.2	7/14/2020	7/21/2020	99 \pm 2.2
	8906	Mid Shelf	41	13.6	7/13/2020	7/21/2020	96 \pm 4.2
	8930	Mid Shelf	83	55.1	7/14/2020	7/21/2020	98 \pm 4.5
	8910	Outer Shelf	169	51.7	7/13/2020	7/21/2020	93 \pm 6.7
	8916	Outer Shelf	189	53.3	7/13/2020	7/21/2020	96 \pm 8.9
	8908	Outer Shelf	194	51.1	7/13/2020	7/21/2020	92 \pm 8.4
	8920	Upper Slope	270	55.9	7/14/2020	7/21/2020	97 \pm 4.2

Table 3.4

Summary of macrofaunal community parameters for PLOO, SBOO, and San Diego regional benthic stations sampled during 2020. Data for each region include mean, 95% confidence interval (CI), minimum, and maximum values; SR=species richness; Abun=abundance; H'=Shannon diversity index; J'=Pielou's evenness; Dom=Swartz dominance; BRI=benthic response index.

		SR	Abun	H'	J'	Dom	BRI
All PLOO Grabs (n=44)	Mean	82	421	3.6	0.81	23	14
	95% CI	5	29	0.1	0.02	3	2
	Min	54	241	2.9	0.70	11	8
	Max	115	636	4.2	0.90	41	33
All SBOO Grabs (n=54)	Mean	60	226	3.3	0.83	21	16
	95% CI	8	51	0.1	0.02	3	2
	Min	17	30	2.3	0.64	6	1
	Max	160	890	4.3	0.92	52	28
All Regional Grabs (n=40)	Mean	60	265	3.3	0.81	19	17
	95% CI	8	49	0.2	0.04	3	2
	Min	20	48	0.4	0.12	1	8
	Max	113	645	4.1	0.94	40	28

Table 3.5

The 25 most abundant macroinvertebrate taxa collected from PLOO benthic stations during 2020. A total of 44 grabs were collected. PA = percent abundance, FO = frequency of occurrence, MAG = mean abundance per grab.

Taxon	Taxonomic Classification	PA	FO	MAG
<i>Spiophanes duplex</i>	Polychaeta: Spionidae	14	100	57
<i>Axinopsida serricata</i>	Mollusca: Bivalvia	10	95	43
<i>Mediomastus</i> sp	Polychaeta: Capitellidae	5	100	20
<i>Amphiodia urtica</i>	Echinodermata: Ophiuroidea	5	89	19
<i>Prionospio jubata</i>	Polychaeta: Spionidae	4	100	15
<i>Spiophanes kimballi</i>	Polychaeta: Spionidae	3	95	15
<i>Paradiopatra parva</i>	Polychaeta: Onuphidae	3	98	12
Amphiuridae	Echinodermata: Ophiuroidea	2	89	9
<i>Tellina</i> sp B	Mollusca: Bivalvia	2	84	9
<i>Eclysippe trilobata</i>	Polychaeta: Ampharetidae	2	91	8
<i>Prionospio dubia</i>	Polychaeta: Spionidae	2	93	8
<i>Praxillella pacifica</i>	Polychaeta: Maldanidae	2	89	8
<i>Scoloplos armiger</i> Cmplx	Polychaeta: Orbiniidae	2	95	8
Euclymeninae sp A	Polychaeta: Maldanidae	2	89	7
<i>Aphelochaeta glandaria</i> Cmplx	Polychaeta: Cirratulidae	2	93	7
<i>Amphiodia</i> sp	Echinodermata: Ophiuroidea	2	86	7
<i>Phisidia sanctaemariae</i>	Polychaeta: Terebellidae	2	86	6
<i>Rhepoxynius bicuspidatus</i>	Arthropoda: Amphipoda	1	86	6
Maldanidae	Polychaeta: Maldanidae	1	91	5
Heteronemertea sp SD2	Nemertea: Heteronemertea	1	93	4
<i>Tellina carpenteri</i>	Mollusca: Bivalvia	1	68	4
<i>Nuculana</i> sp A	Mollusca: Bivalvia	1	89	4
<i>Glycera nana</i>	Polychaeta: Glyceridae	1	82	3
<i>Rhodine bitorquata</i>	Polychaeta: Maldanidae	1	77	3
<i>Chaetozone hartmanae</i>	Polychaeta: Cirratulidae	1	82	3

Table 3.6

The 25 most abundant macroinvertebrate taxa collected from SBOO benthic stations during 2020. A total of 54 grabs were collected. PA = percent abundance, FO = frequency of occurrence, MAG = mean abundance per grab.

Taxon	Taxonomic Classification	PA	FO	MAG
<i>Spiophanes duplex</i>	Polychaeta: Spionidae	7	78	16
<i>Spiophanes norrisi</i>	Polychaeta: Spionidae	7	89	16
<i>Jasmineira</i> sp B	Polychaeta: Sabellidae	3	15	6
<i>Mediomastus</i> sp	Polychaeta: Capitellidae	3	57	6
<i>Protodorvillea gracilis</i>	Polychaeta: Dorvilleidae	2	33	5
<i>Paramphinome</i> sp	Polychaeta: Amphinomidae	2	4	5
<i>Hesionura coineaui difficilis</i>	Polychaeta: Phyllodocidae	2	7	5
<i>Chondrochelia dubia</i> Cmplx	Arthropoda: Tanaidacea	2	76	5
<i>Pisione</i> sp	Polychaeta: Sigalionidae	2	7	4
<i>Lanassa venusta venusta</i>	Polychaeta: Terebellidae	2	26	4
<i>Dendraster terminalis</i>	Echinodermata: Echinoidea	2	35	3
<i>Sigalion spinosus</i>	Polychaeta: Sigalionidae	1	74	3
<i>Spiochaetopterus costarum</i> Cmplx	Polychaeta: Chaetopteridae	1	65	3
Nematoda	Nematoda	1	63	3
<i>Rhepoxynius heterocuspидatus</i>	Arthropoda: Amphipoda	1	30	3
<i>Gadila aberrans</i>	Mollusca: Scaphopoda	1	48	2
Onuphidae	Polychaeta: Onuphidae	1	26	2
<i>Sthenelanelia uniformis</i>	Polychaeta: Sigalionidae	1	26	2
<i>Tellina modesta</i>	Mollusca: Bivalvia	1	39	2
<i>Euclymeninae</i> sp A	Polychaeta: Maldanidae	1	54	2
<i>Tubulanus polymorphus</i>	Nemertea: Palaeonemertea	1	50	2
<i>Rhepoxynius menziesi</i>	Arthropoda: Amphipoda	1	59	2
<i>Glycera oxycephala</i>	Polychaeta: Glyceridae	1	28	2
<i>Mooreonuphis</i> sp	Polychaeta: Onuphidae	1	19	2
<i>Euphilomedes carcharodonta</i>	Arthropoda: Ostracoda	1	43	2

Table 3.7

The 10 most abundant macroinvertebrate taxa per depth stratum collected from San Diego regional benthic stations sampled during summer 2020. PA = percent abundance, FO = frequency of occurrence, MAG = mean abundance per grab.

Strata	Taxon	Taxonomic Classification	PA	FO	MAG
Inner	<i>Prionospio pygmaeus</i>	Polychaeta: Spionidae	27	82	59
Shelf	<i>Mediomastus</i> sp	Polychaeta: Capitellidae	6	64	13
n = 11	<i>Spiophanes duplex</i>	Polychaeta: Spionidae	5	100	12
	<i>Spiophanes norrisi</i>	Polychaeta: Spionidae	4	100	8
	<i>Owenia collaris</i>	Polychaeta: Oweniidae	2	36	5
	<i>Spiochaetopterus costarum</i> Cmplx	Polychaeta: Chaetopteridae	2	73	5
	<i>Gadila aberrans</i>	Mollusca: Scaphopoda	2	55	4
	<i>Carinoma mutabilis</i>	Nemertea: Palaeonemertea	1	73	3
	<i>Callianax baetica</i>	Mollusca: Gastropoda	1	45	3
	<i>Tellina modesta</i>	Mollusca: Bivalvia	1	100	3
Mid-shelf	<i>Spiophanes duplex</i>	Polychaeta: Spionidae	14	100	51
n = 17	<i>Amphiodia urtica</i>	Echinodermata: Ophiuroidea	9	82	35
	<i>Axinopsida serricata</i>	Mollusca: Bivalvia	7	88	24
	<i>Mediomastus</i> sp	Polychaeta: Capitellidae	4	100	15
	Amphiuridae	Echinodermata: Ophiuroidea	4	88	13
	<i>Paradiopatra parva</i>	Polychaeta: Onuphidae	3	88	10
	<i>Amphiodia</i> sp	Echinodermata: Ophiuroidea	2	82	9
	<i>Prionospio jubata</i>	Polychaeta: Spionidae	2	94	8
	<i>Spiophanes kimballi</i>	Polychaeta: Spionidae	2	71	8
	Euclymeninae sp A	Polychaeta: Maldanidae	2	94	7
Outer	<i>Phyllochaetopterus limicolus</i>	Polychaeta: Chaetopteridae	13	100	33
Shelf	<i>Axinopsida serricata</i>	Mollusca: Bivalvia	11	100	28
n = 6	<i>Mediomastus</i> sp	Polychaeta: Capitellidae	7	100	18
	<i>Paraprionospio alata</i>	Polychaeta: Spionidae	5	100	13
	<i>Spiophanes kimballi</i>	Polychaeta: Spionidae	5	100	11
	<i>Paradiopatra parva</i>	Polychaeta: Onuphidae	4	83	11
	<i>Prionospio jubata</i>	Polychaeta: Spionidae	4	83	10
	<i>Spiophanes duplex</i>	Polychaeta: Spionidae	3	67	8
	<i>Laonice nuchala</i>	Polychaeta: Spionidae	3	50	6
	<i>Scoletoma tetraura</i> Cmplx	Polychaeta: Lumbrineridae	2	83	4

Table 3.7 *continued*

Strata	Species	Taxonomic Classification	PA	FO	MAG
<i>Upper</i>	<i>Paraprionospio alata</i>	Polychaeta: Spionidae	17	67	12
<i>Slope</i>	<i>Maldane sarsi</i>	Polychaeta: Maldanidae	9	67	7
n = 6	<i>Prionospio ehlersi</i>	Polychaeta: Spionidae	6	83	4
	<i>Phyllochaetopterus limicolus</i>	Polychaeta: Chaetopteridae	5	50	4
	<i>Pectinaria californiensis</i>	Polychaeta: Pectinariidae	4	67	3
	<i>Eclysippe trilobata</i>	Polychaeta: Ampharetidae	3	50	2
	<i>Mediomastus</i> sp	Polychaeta: Capitellidae	3	50	2
	<i>Yoldiella nana</i>	Mollusca: Bivalvia	2	50	1
	<i>Aphelochaeta monilaris</i>	Polychaeta: Cirratulidae	1	50	1
	<i>Axinopsida serricata</i>	Mollusca: Bivalvia	1	50	1
	<i>Bipalponephrys cornuta</i>	Polychaeta: Nephtyidae	1	33	1
	<i>Fauveliopsis</i> sp SD1	Polychaeta: Fauveliopsidae	1	33	1
	<i>Prionospio jubata</i>	Polychaeta: Spionidae	1	33	1
	<i>Sternaspis affinis</i>	Polychaeta: Sternaspidae	1	33	1

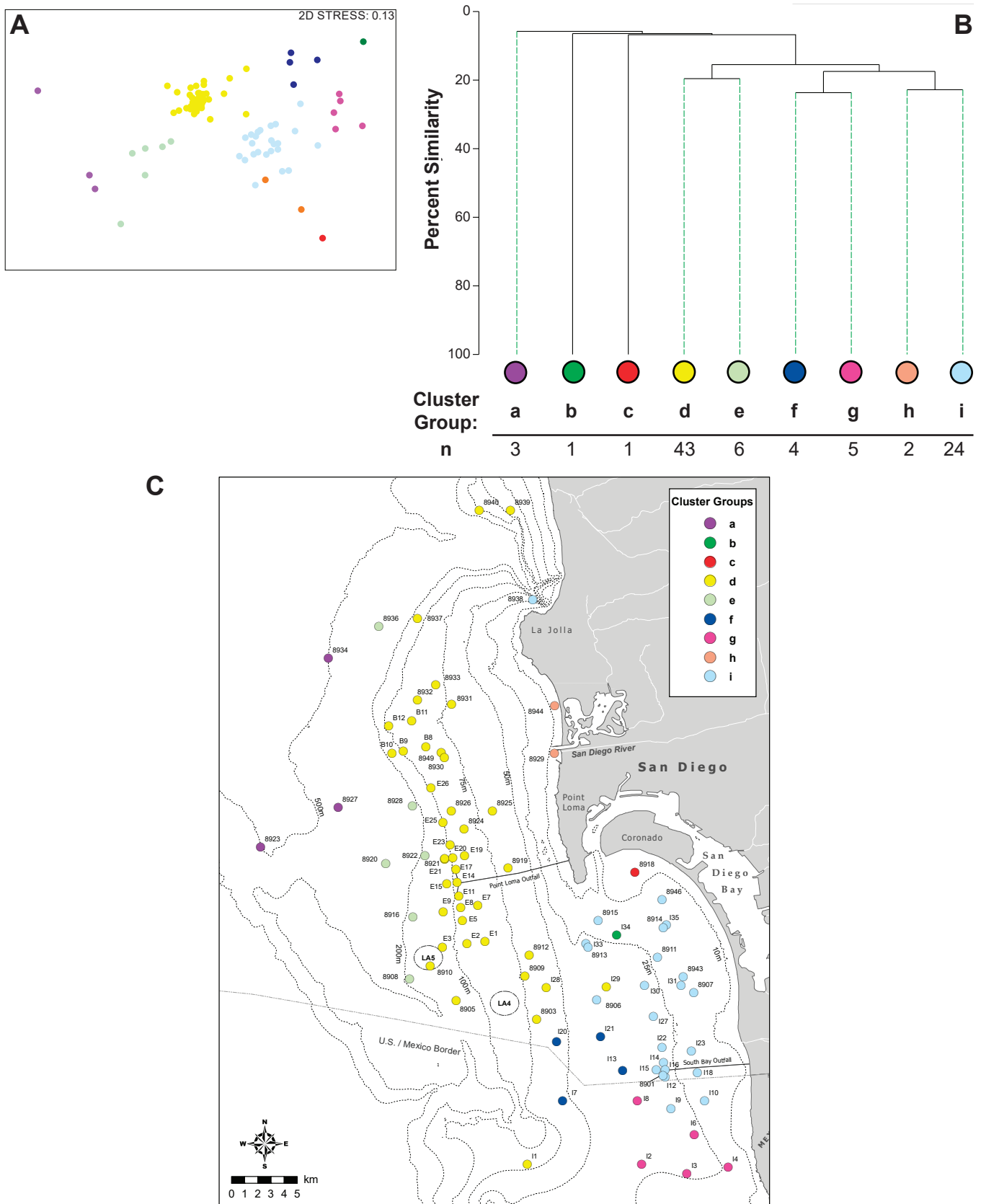


Figure 3.2

Results of ordination and cluster analysis of macrofauna data from PLOO, SBOO, and San Diego regional benthic stations sampled during summer 2020. Results are presented as (A) nMDS ordination; (B) a dendrogram of main cluster groups; (C) a map showing the distribution of cluster groups throughout the region.

Chapter 4

Demersal Fishes and Megabenthic Invertebrates

Chapter 4. Demersal Fishes and Megabenthic Invertebrates

INTRODUCTION

The City of San Diego collects bottom dwelling (demersal) fishes and large (megabenthic) mobile invertebrates by otter trawl to examine potential effects of wastewater discharge, or other disturbances, on the marine environment surrounding the Point Loma and South Bay Ocean Outfalls (PLOO and SBOO, respectively). This chapter presents summaries and preliminary analyses of the demersal fish and megabenthic invertebrate data collected during 2020 at a total of 13 trawl stations surrounding the PLOO and SBOO. Raw data summaries supporting these results are presented in Appendix C. A more comprehensive assessment of these results will be presented in the 2020-2021 Biennial Assessment Report to be submitted by July 1, 2022.

MATERIALS AND METHODS

Field Sampling

Trawls were conducted at 13 stations to monitor demersal fish and megabenthic invertebrate populations during winter (January) and summer (July) 2020 (Figure 4.1). These included six PLOO stations located along the 100-m depth contour (discharge depth), ranging from 9 km south to 8 km north of the outfall, and seven SBOO stations located along the 28-m depth contour (discharge depth), ranging from 7 km south to 8.5 km north of the outfall. The two PLOO stations (SD10 and SD12) and two SBOO stations (SD17 and SD18) located within 1000 m of the outfall structures are considered to represent nearfield conditions. A single trawl was performed at each station, during each survey, using a 7.6-m Marinovich otter trawl fitted with a 1.3-cm cod-end mesh net. Standard sampling procedures require towing the net for a total of 10 minutes bottom time per trawl, at a speed of about 2 knots along a predetermined heading. Pressure-temperature sensors were attached to one of the trawl doors to measure water temperature, depth, and time of the individual trawls. Data collected by these sensors were used to confirm bottom time and depth of each trawl. The catch from each successful trawl was sorted and inspected aboard ship. All individual fish and invertebrates captured were identified to species, or to the lowest taxon possible, based on accepted taxonomic protocols for the region (Eschmeyer and Herald 1998, Page et al. 2013, SCAMIT 2018). If an animal could not be accurately identified to species in the field, it was returned to the laboratory for an attempt at further identification. The total number of individuals and total biomass (kg, wet weight) were recorded for each fish species. Additionally, each fish was inspected for the presence of physical abnormalities (e.g., tumors, lesions, fin erosion, discoloration) and external parasites (e.g., copepods, cymothoid isopods, leeches). The length of each individual fish was measured to the nearest centimeter to determine size class; total length (TL) was measured for cartilaginous fishes, while standard length (SL) was measured for bony fishes (SCCWRP 2018). For trawl-caught invertebrates, only the total number of individuals was recorded for each species. Parasitic invertebrates no longer attached to their hosts, including the cymothoid isopod *Elthusa vulgaris* and leeches in the subclass Hirudinea, were recorded as present/absent, rather than being counted individually, and are not included in the analyses presented herein. Visual observations of weather, sea conditions, and human/animal activity were also recorded at the time of sampling (see Addenda 1-2A, 1-2B).

Data Analyses

Population characteristics of fish and invertebrate species were summarized as percent abundance (number of individuals per species/total abundance of all species), frequency of occurrence (percentage of stations at which a species was collected), mean abundance per haul (number of individuals per species/total number of stations sampled), and mean abundance per occurrence (number of individuals per species/number of stations at which the species was collected). Additionally, the following community structure parameters were calculated per trawl for both fishes and invertebrates: species richness (number of species or distinct taxa), total abundance (number of individuals), and the Shannon Diversity Index (H'). Total biomass was also measured for each fish species. These analyses were performed using R (R Core Team 2018) and various functions within the dplyr, plyr, reshape2, RODBC, sqldf, and vegan packages (Wickham 2007, 2011, Grothendieck 2014, Wickham and Francois 2016, Oksanen et al. 2018, Ripley and Lapsley 2018).

Multivariate analyses were performed in PRIMER v7 software using demersal fish and megabenthic invertebrate data collected from trawls conducted in the PLOO and SBOO regions during 2020 (Clarke 1993, Warwick 1993, Clarke et al. 2014). These analyses included hierarchical agglomerative clustering (cluster analysis) with group-average linking and similarity profile analysis (SIMPROF) to confirm the non-random structure of the resultant cluster dendrogram (Clarke et al. 2008). The Bray-Curtis measure of dissimilarity was used as the basis for the cluster analysis, and abundance data were either square-root (PLOO and SBOO fish, SBOO invertebrates) or log (PLOO invertebrates) transformed to lessen the influence of overly abundant species and increase the importance (or impact) of rare species.

RESULTS

Demersal Fishes

All fish species captured during the 2020 trawl surveys are summarized by percent abundance, frequency of occurrence, mean abundance per haul, and mean abundance per occurrence in Tables 4.1 and 4.2. Species richness, abundance, diversity, and biomass values for each station are summarized in Table 4.3. Total number of individuals, total biomass, minimum and maximum length, and mean length per species are included in Appendices C.1 and C.2. Total abundance and biomass by species for each station are summarized in Appendices C.3–C.6. All abnormalities and parasites found on trawled fish during the reporting period are listed in Appendix C.7. Cluster analyses were performed to evaluate ecological patterns within the demersal fish communities in the San Diego region; these results are presented in Figures 4.2 and 4.3.

Megabenthic Invertebrates

All megabenthic invertebrate species captured during the 2020 trawl surveys are summarized by percent abundance, frequency of occurrence, mean abundance per haul, and mean abundance per occurrence in Tables 4.4 and 4.5. Species richness, abundance, and diversity values for each station are summarized in Table 4.6. The total number of individuals per species is included in Appendices C.8 and C.9. Total abundance by species for each station is summarized in Appendices C.10 and C.11. Cluster analyses were performed to evaluate ecological patterns within the megabenthic invertebrate communities in the San Diego region; these results are presented in Figures 4.4 and 4.5.

SUMMARY

Preliminary analysis of the demersal fish and megabenthic invertebrate data collected in 2020 indicate that treated wastewater discharged through the PLOO and SBOO has not negatively impacted these communities in the coastal waters off San Diego. Values for most community parameters were similar at stations located both near and far away from the outfall discharge sites. Community metrics, such as species richness, abundance, and diversity, were within historical ranges reported for the San Diego region (City of San Diego 2020) and were representative of those characterizing similar habitats throughout the Southern California Bight (Allen et al. 1998, 2002, 2007, 2011, Walther et al. 2017).

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CHAPTER 4
FIGURES & TABLES

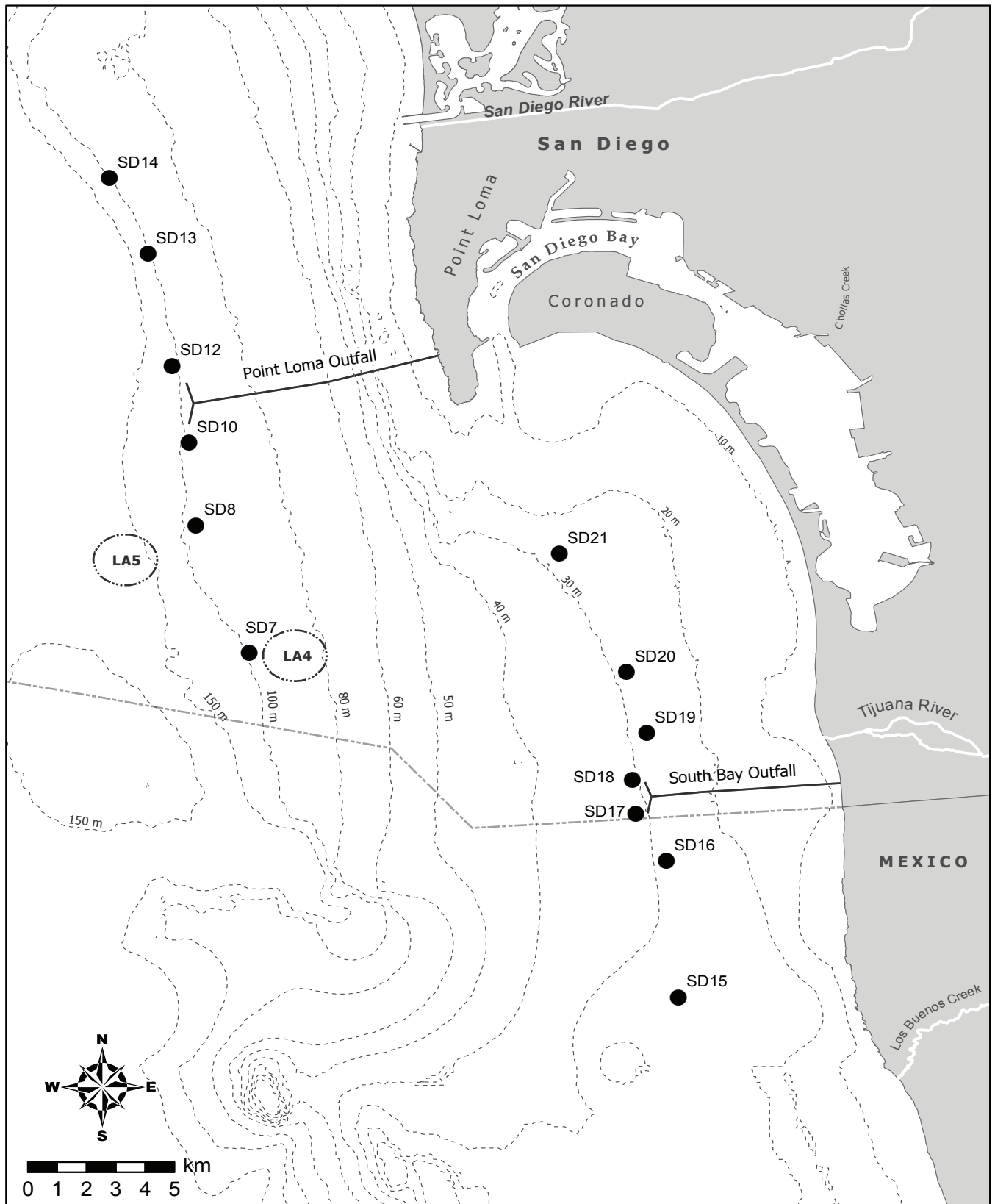


Figure 4.1

Trawl station locations sampled around the PLOO and SBOO as part of the City of San Diego's Ocean Monitoring Program.

Table 4.1

Demersal fish species collected from 12 trawls conducted in the PLOO region during 2020. PA = percent abundance; FO = frequency of occurrence; MAH = mean abundance per haul; MAO = mean abundance per occurrence.

Species	PA	FO	MAH	MAO
Pacific Sanddab	41	100	111	111
Halfbanded Rockfish	23	67	63	94
Dover Sole	10	100	26	26
English Sole	4	92	12	13
Shortspine Combfish	4	100	10	10
Longfin Sanddab	4	83	10	12
Longspine Combfish	3	75	9	12
Plainfin Midshipman	2	67	5	8
Hornyhead Turbot	1	83	4	4
Stripetail Rockfish	1	67	3	4
Pink Seaperch	1	75	3	3
California Tonguefish	1	67	2	3
Slender Sole	1	33	2	6
Yellowchin Sculpin	1	25	2	8
Bigmouth Sole	1	75	2	2
California Scorpionfish	1	67	1	2
Squarespot Rockfish	1	17	1	9
California Lizardfish	< 1	58	1	2
Pacific Argentine	< 1	17	1	5
Specklefin Midshipman	< 1	8	1	10
Rockfish Unidentified	< 1	42	1	2
Spotted Cusk-eel	< 1	67	1	1
White Croaker	< 1	8	< 1	4
California Skate	< 1	25	< 1	1
Curlfin Sole	< 1	17	< 1	2
Greenstriped Rockfish	< 1	17	< 1	2
Vermilion Rockfish	< 1	8	< 1	3
Blackbelly Eelpout	< 1	8	< 1	1
Blacktip Poacher	< 1	8	< 1	1
California Halibut	< 1	8	< 1	1
Fantail Sole	< 1	8	< 1	1
Rosethorn Rockfish	< 1	8	< 1	1
Smooth Stargazer	< 1	8	< 1	1
Spotfin Sculpin	< 1	8	< 1	1
Stripefin Ronquil	< 1	8	< 1	1

Table 4.2

Demersal fish species collected from 14 trawls conducted in the SBOO region during 2020. PA = percent abundance; FO = frequency of occurrence; MAH = mean abundance per haul; MAO = mean abundance per occurrence.

Species	PA	FO	MAH	MAO
Speckled Sanddab	51	100	63	63
Longfin Sanddab	17	71	21	29
California Lizardfish	9	93	11	12
California Tonguefish	5	93	6	6
Pacific Sardine	4	7	4	62
White Croaker	3	21	4	19
Yellowchin Sculpin	3	29	3	12
Pacific Sanddab	2	43	2	6
Hornyhead Turbot	2	57	2	3
Plainfin Midshipman	1	29	1	4
Specklefin Midshipman	1	50	1	2
English Sole	1	29	1	3
Roughback Sculpin	1	43	1	2
California Halibut	1	29	1	3
Fantail Sole	< 1	29	< 1	2
Spotted Turbot	< 1	36	< 1	1
Barcheek Pipefish	< 1	14	< 1	2
Bigmouth Sole	< 1	14	< 1	2
Pipefish Unidentified	< 1	7	< 1	3
Queenfish	< 1	7	< 1	3
Round Stingray	< 1	14	< 1	1
Shovelnose Guitarfish	< 1	14	< 1	1
California Scorpionfish	< 1	7	< 1	1
Giant Kelpfish	< 1	7	< 1	1
Longspine Combfish	< 1	7	< 1	1
Pacific Pompano	< 1	7	< 1	1

Table 4.3

Summary of demersal fish community parameters for PLOO and SBOO trawl stations sampled during 2020. Data are included for species richness, abundance, diversity (H'), and biomass (kg, wet weight).

	Station	<i>Species Richness</i>		<i>Abundance</i>		<i>Diversity</i>		<i>Biomass</i>	
		Winter	Summer	Winter	Summer	Winter	Summer	Winter	Summer
PLOO	SD7	17	14	221	126	2.0	1.6	3.4	2.9
	SD8	22	13	453	229	1.7	1.3	7.2	3.6
	SD10	14	19	184	327	1.5	2.0	9.2	8.0
	SD12	6	17	51	174	1.1	1.8	0.6	8.5
	SD13	12	21	151	929	1.5	1.5	5.3	29.1
	SD14	12	16	119	274	1.7	1.4	5.9	7.7
SBOO	SD15	5	5	48	38	0.5	0.6	1.2	0.5
	SD16	7	9	68	183	0.9	1.2	1.0	3.9
	SD17	7	5	100	19	1.2	1.4	1.2	0.5
	SD18	9	8	66	80	1.5	0.9	6.3	0.8
	SD19	9	12	153	194	1.5	1.2	2.2	3.6
	SD20	8	11	162	299	1.4	1.3	3.1	6.1
	SD21	15	10	144	178	1.9	1.6	7.2	4.9

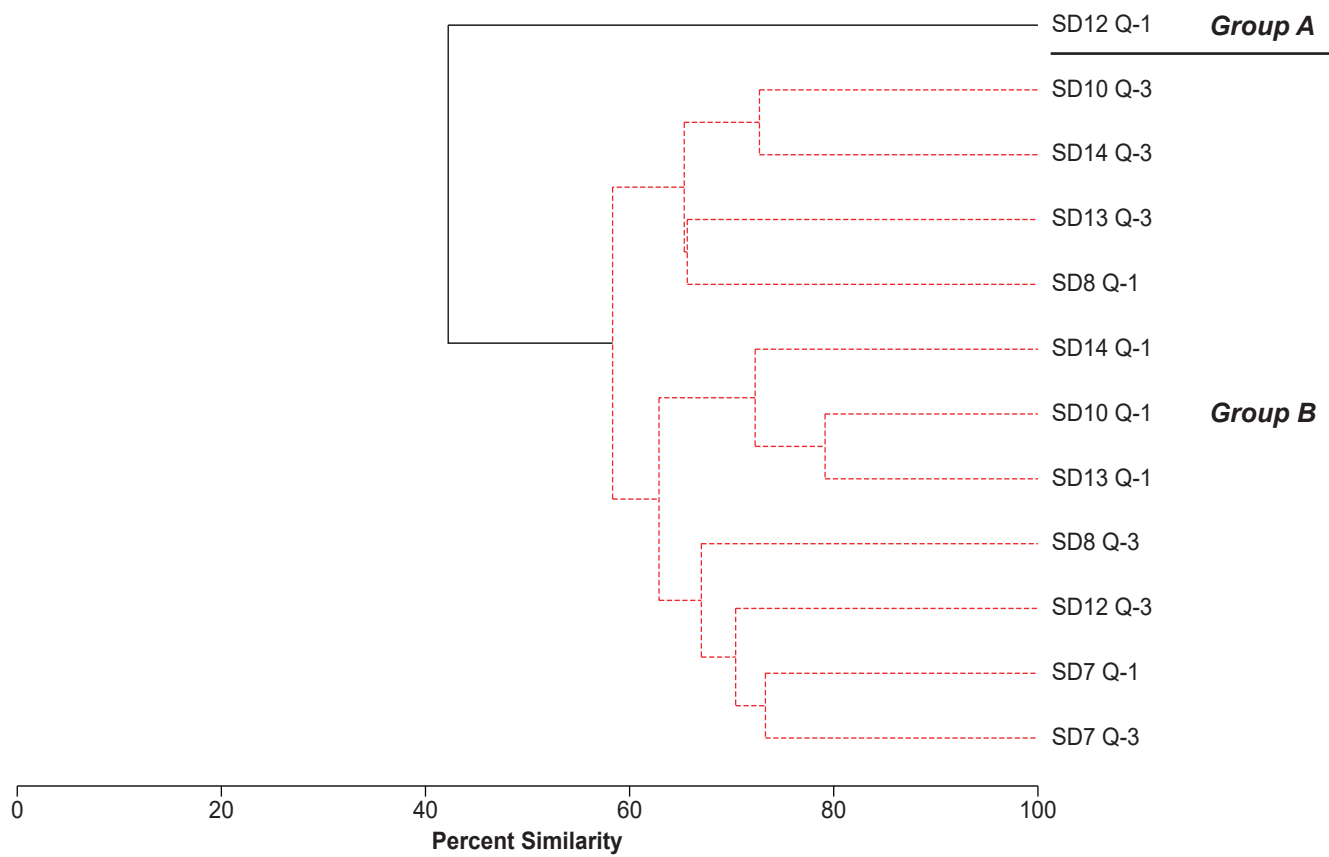


Figure 4.2

Results of cluster analysis of demersal fish data from PLOO trawl stations sampled during 2020. Solid black lines indicate non-random structure of the dendrogram as confirmed by SIMPROF; Q-1 = winter survey, Q-3 = summer survey.

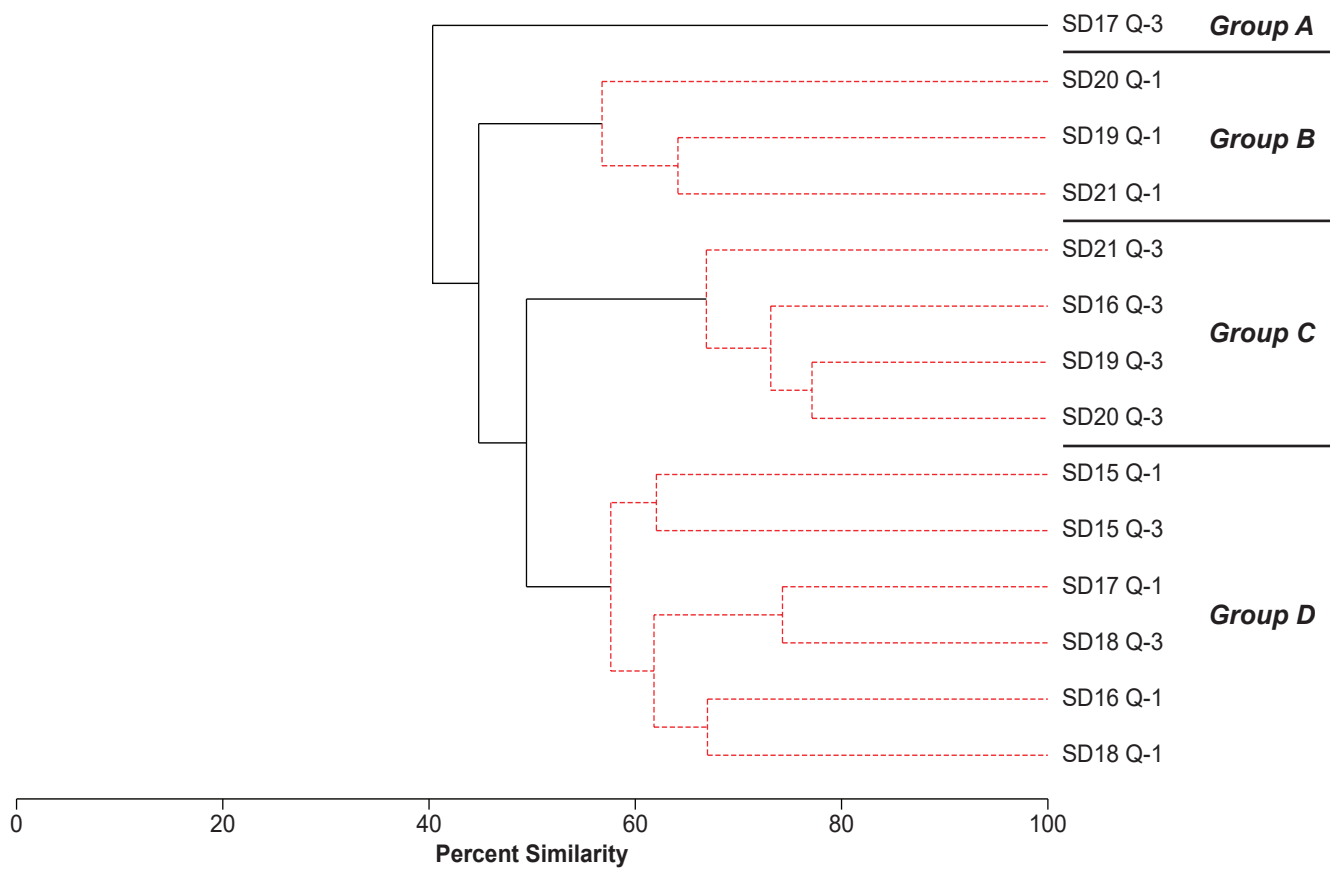


Figure 4.3

Results of cluster analysis of demersal fish data from SBOO trawl stations sampled during 2020. Solid black lines indicate non-random structure of the dendrogram as confirmed by SIMPROF; Q-1 = winter survey, Q-3 = summer survey.

Table 4.4

Megabenthic invertebrate species collected from 12 trawls conducted in the PLOO region during 2020. PA = percent abundance; FO = frequency of occurrence; MAH = mean abundance per haul; MAO = mean abundance per occurrence.

Species	PA	FO	MAH	MAO
<i>Pleuroncodes planipes</i>	97	67	12,635	18,952
<i>Lytechinus pictus</i>	2	100	293	293
<i>Sicyonia ingentis</i>	1	92	93	102
<i>Platymera gaudichaudii</i>	< 1	67	38	57
Paguroidea	< 1	8	3	32
<i>Luidia foliolata</i>	< 1	58	2	3
<i>Astropecten californicus</i>	< 1	75	2	2
<i>Paguristes bakeri</i>	< 1	8	1	13
<i>Apostichopus californicus</i>	< 1	58	1	1
<i>Luidia armata</i>	< 1	17	< 1	3
<i>Ophiopholis bakeri</i>	< 1	17	< 1	2
<i>Ophiothrix spiculata</i>	< 1	17	< 1	2
<i>Ophiura luetkenii</i>	< 1	8	< 1	3
<i>Aphrodita refulgida</i>	< 1	17	< 1	1
<i>Loxorhynchus crispatus</i>	< 1	8	< 1	2
<i>Phimochirus californiensis</i>	< 1	8	< 1	2
<i>Strongylocentrotus fragilis</i>	< 1	17	< 1	1
<i>Acanthoptilum</i> sp	< 1	8	< 1	1
<i>Amphichondrius granulatus</i>	< 1	8	< 1	1
<i>Austrotrophon catalinensis</i>	< 1	8	< 1	1
<i>Calliostoma turbinum</i>	< 1	8	< 1	1
<i>Halichondria</i> sp	< 1	8	< 1	1
<i>Loxorhynchus grandis</i>	< 1	8	< 1	1
<i>Luidia asthenosoma</i>	< 1	8	< 1	1
<i>Octopus rubescens</i>	< 1	8	< 1	1
<i>Pagurus armatus</i>	< 1	8	< 1	1
<i>Tritia insculpta</i>	< 1	8	< 1	1

Table 4.5

Megabenthic invertebrate species collected from 14 trawls conducted in the SBOO region during 2020. PA = percent abundance; FO = frequency of occurrence; MAH = mean abundance per haul; MAO = mean abundance per occurrence.

Species	PA	FO	MAH	MAO
<i>Astropecten californicus</i>	24	79	10	13
<i>Philine auriformis</i>	21	43	9	22
<i>Crangon nigromaculata</i>	20	64	9	14
<i>Sicyonia penicillata</i>	7	50	3	6
<i>Lovenia cordiformis</i>	6	36	3	8
<i>Portunus xantusii</i>	5	50	2	4
<i>Lytechinus pictus</i>	4	43	2	4
<i>Dendraster terminalis</i>	2	14	1	7
<i>Luidia armata</i>	2	43	1	2
<i>Crossata ventricosa</i>	1	36	< 1	1
<i>Pyromaia tuberculata</i>	1	36	< 1	1
<i>Ophiothrix spiculata</i>	1	21	< 1	2
<i>Suberites</i> sp	1	29	< 1	1
<i>Kelletia kelletii</i>	1	14	< 1	2
<i>Loxorhynchus grandis</i>	1	14	< 1	2
<i>Armina californica</i>	< 1	14	< 1	2
<i>Stylatula elongata</i>	< 1	21	< 1	1
<i>Acanthodoris brunnea</i>	< 1	7	< 1	2
<i>Acanthodoris rhodoceras</i>	< 1	14	< 1	1
<i>Hemisquilla californiensis</i>	< 1	14	< 1	1
<i>Platymera gaudichaudii</i>	< 1	14	< 1	1
<i>Sinum scopulosum</i>	< 1	14	< 1	1
<i>Crangon alba</i>	< 1	7	< 1	1
<i>Crassispira semiinflata</i>	< 1	7	< 1	1
<i>Farfantepenaeus californiensis</i>	< 1	7	< 1	1
<i>Latulambrus occidentalis</i>	< 1	7	< 1	1
<i>Moreiradromia sarraburei</i>	< 1	7	< 1	1
<i>Neverita recluziana</i>	< 1	7	< 1	1
<i>Octopus rubescens</i>	< 1	7	< 1	1
<i>Paguristes bakeri</i>	< 1	7	< 1	1
<i>Pagurus spilocarpus</i>	< 1	7	< 1	1
<i>Pleuroncodes planipes</i>	< 1	7	< 1	1
<i>Pteropurpura festiva</i>	< 1	7	< 1	1

Table 4.6

Summary of megabenthic invertebrate community parameters for PLOO and SBOO trawl stations sampled during 2020. Data are included for species richness, abundance, and diversity (H').

	Station	<i>Species Richness</i>		<i>Abundance</i>		<i>Diversity</i>	
		Winter	Summer	Winter	Summer	Winter	Summer
PLOO	SD7	7	9	24	28	1.8	1.7
	SD8	13	9	1092	1010	0.3	0.1
	SD10	9	6	24,242	839	0.1	0.1
	SD12	5	4	46,255	11,558	0.1	0.3
	SD13	7	6	34,371	137	0.0	0.4
	SD14	7	5	36,118	1164	0.1	0.3
SBOO	SD15	10	9	45	46	1.6	1.6
	SD16	7	9	33	49	1.4	1.3
	SD17	13	10	34	75	2.2	0.8
	SD18	7	9	23	47	1.5	1.4
	SD19	7	2	64	48	1.3	0.4
	SD20	5	6	33	41	1.2	0.6
	SD21	6	5	46	32	1.0	0.6

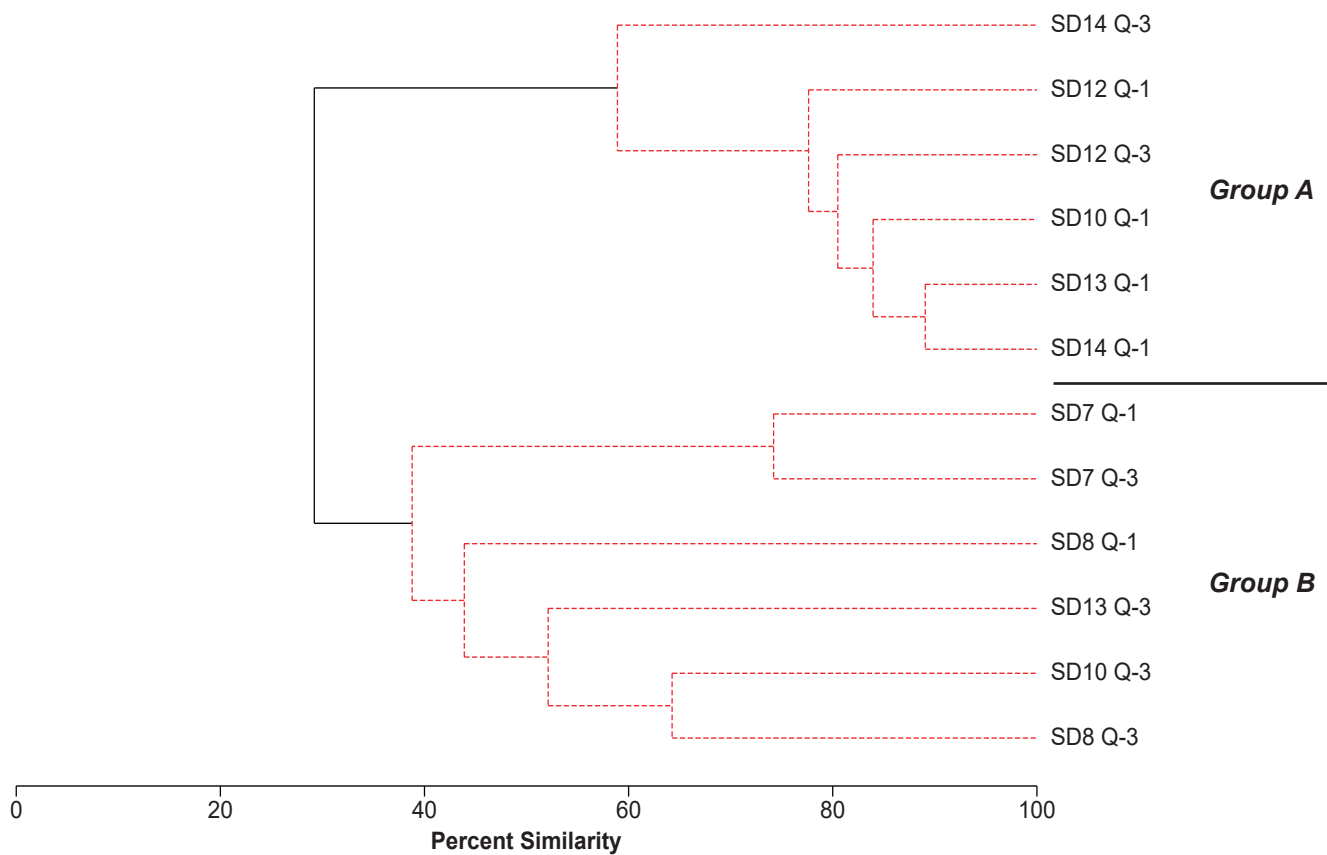


Figure 4.4

Results of cluster analysis of megabenthic invertebrate data from PLOO trawl stations sampled during 2020. Solid black lines indicate non-random structure of the dendrogram as confirmed by SIMPROF; Q-1 = winter survey, Q-3 = summer survey.

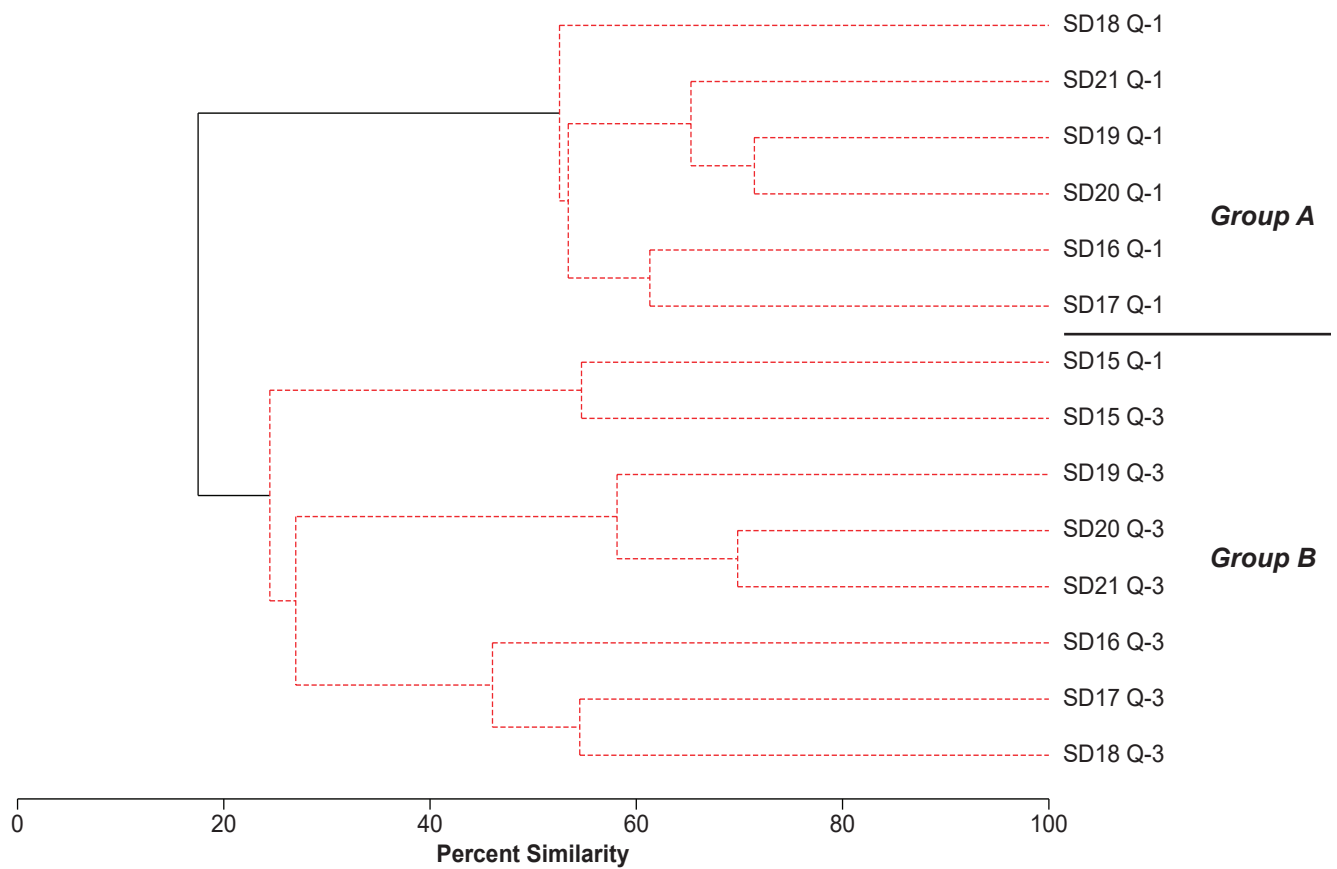


Figure 4.5

Results of cluster analysis of megabenthic invertebrate data from SBOO trawl stations sampled during 2020. Solid black lines indicate non-random structure of the dendrogram as confirmed by SIMPROF; Q-1 = winter survey, Q-3 = summer survey.

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Chapter 5

Contaminants in Marine Fishes

Chapter 5. Contaminants in Marine Fishes

INTRODUCTION

Bottom dwelling (demersal) fishes are collected by the City of San Diego (City) to evaluate the presence of contaminants in their tissues, which may result from the discharge of wastewater from the Point Loma and South Bay Ocean Outfalls (PLOO and SBOO, respectively). Anthropogenic inputs to coastal waters can result in increased concentrations of pollutants within the local marine environment, which may subsequently accumulate in the tissues of fishes and their prey. This portion of the City’s Ocean Monitoring Program consists of two components: (1) analyzing liver tissues from mostly trawl-caught fishes; (2) analyzing muscle tissues from fishes collected by hook and line (rig fishing). All liver and muscle tissue samples collected were analyzed for contaminants specified in the National Pollutant Discharge Elimination System (NPDES) discharge permits that govern monitoring requirements for the PLOO and SBOO regions. This chapter presents summaries and preliminary analyses of all fish tissue data collected during calendar year 2020 at PLOO and SBOO stations. Raw data summaries supporting these results are presented in Appendix D. A more comprehensive assessment of these results will be presented in the 2020-2021 Biennial Assessment Report to be submitted by July 1, 2022.

MATERIALS AND METHODS

Fishes were collected in fall (October) 2020 from a total of nine trawl zones (TZ1–TZ9) and four rig fishing zones (RF1–RF4) that span the PLOO and SBOO monitoring regions (Figure 5.1). Each trawl zone represents an area centered on one or two trawl stations, as specified in Chapter 4. Trawl Zone 1 includes the “nearfield” area within a 1-km radius of PLOO stations SD10 and SD12, which are located just south and north of the outfall discharge site, respectively. Trawl Zone 2 includes the area within a 1-km radius surrounding the northern “farfield” PLOO stations SD13 and SD14. Trawl Zone 3 represents the area within a 1-km radius surrounding the “farfield” PLOO station SD8, which is located south of the outfall near the LA-5 dredged material disposal site. Trawl Zone 4 is the area within a 1-km radius surrounding the “farfield” PLOO station SD7, which is located several kilometers south of the outfall. Trawl Zone 5 includes the area located within a 1-km radius of the SBOO stations SD17 and SD18, which are located just south and north of the outfall discharge site, respectively. Trawl Zone 6 includes the area within a 1-km radius surrounding the northern SBOO stations SD19 and SD20, while Trawl Zone 7 includes the area within a 1-km radius of the northern SBOO station SD21. Trawl Zone 8 represents the area within a 1-km radius surrounding the southern SBOO station SD16, while Trawl Zone 9 represents the area within a 1-km radius surrounding the southern SBOO station SD15. Rig Fishing Zones 1–4 represent the areas within a 1-km radius of the nominal coordinates for stations RF1, RF2, RF3, and RF4. Stations RF1 and RF3 are located within 1 km of the PLOO and SBOO discharge sites, respectively, and are considered the “nearfield” rig fishing sites. In contrast, station RF2 is located approximately 11 km northwest of the PLOO, while station RF4 is located approximately 13 km southeast of the SBOO. These two sites are considered “farfield”, or reference, stations for the analyses herein. Efforts to collect target species by trawl were limited to five 10-minute (bottom time) trawls per site, while rig fishing effort was limited to 5 hours at each station. Occasionally, insufficient numbers of target species are obtained despite this effort; during 2020, this resulted in inadequate amounts of tissue at Trawl Zone 9 to complete three full composite samples.

A total of 14 species of fish were collected for analysis of liver and muscle tissues during the 2020 survey (Table 5.1). Four different species of flatfish were collected from the nine trawl zones for analysis of liver tissues, including Pacific Sanddab (*Citharichthys sordidus*), Longfin Sanddab (*Citharichthys xanhostigma*), Hornyhead Turbot (*Pleuronichthys verticalis*), and Spotted Turbot (*Pleuronichthys ritteri*). These flatfish were collected from regular trawls at the SBOO stations, and by alternative hook and line methods at the PLOO stations. An additional 10 species of fish were collected for analysis of muscle tissues at the rig fishing stations using standard hook and line fishing techniques. These species included California Scorpionfish (*Scorpaena guttata*), Brown Rockfish (*Sebastes auriculatus*), Flag Rockfish (*Sebastes rubrivinctus*), Gopher Rockfish (*Sebastes carnatus*), Olive Rockfish (*Sebastes serranoides*), Speckled Rockfish (*Sebastes ovalis*), Starry Rockfish (*Sebastes constellatus*), Treefish (*Sebastes serripes*), and Vermilion Rockfish (*Sebastes miniatus*).

Only fishes with standard lengths ≥ 11 cm were retained to ensure the collection of sufficient tissue for analysis, while minimizing total catch necessary. These fishes were sorted into three composite samples per station, with a minimum of three individuals in each composite. All fishes were wrapped in aluminum foil, labeled, sealed in re-sealable plastic bags, placed on dry ice, and then transported to the City's Marine Biology Laboratory where they were stored at -20°C prior to dissection and tissue processing.

Tissue Processing and Chemical Analyses

All dissections were performed according to standard techniques for tissue analysis. A brief summary follows, but see City of San Diego (2020b) for additional details. Prior to dissection, each fish was partially defrosted, cleaned with a paper towel to remove loose scales and excess mucus, and the standard length (cm) and weight (g) were recorded (Appendices D.2, D.3). Dissections were carried out on Teflon® pads that were cleaned between samples. The liver or muscle tissues from each fish were removed and placed in separate glass jars for each composite sample, sealed, labeled, and stored in a freezer at -20°C prior to chemical analyses.

All tissue analyses were performed at the City of San Diego's Environmental Chemistry Laboratory. A detailed description of the analytical protocols can be found in City of San Diego (2021). Briefly, all fish tissue samples were analyzed on a wet weight basis to determine the concentrations of 18 different trace metals, nine chlorinated pesticides, 40 polychlorinated biphenyl compound congeners (PCBs), and 24 polycyclic aromatic hydrocarbons (PAHs). Data were generally limited to values above the method detection limit (MDL) for each parameter (Appendix D.1). However, concentrations below MDLs were included as estimated values if the presence of the specific constituent was verified by mass-spectrometry.

Data Analyses

Data summaries for each parameter include detection rate, minimum, maximum, and mean values for all samples combined by species for each outfall region. All means were calculated using detected values only, with no substitutions made for non-detects (analyte concentrations $< \text{MDL}$). Total chlordane, total DDT (tDDT), total hexachlorocyclohexane (tHCH), total PCB (tPCB), and total PAH (tPAH) were calculated for each sample as the sum of all constituents with reported values for individual constituents. Data analyses were performed using R (R Core Team 2020) using various functions within the zoo, reshape2, tidyr, psych, and dplyr packages (Zeileis and Grothendieck 2005, Wickham 2007, Wickham 2018, Revell 2019, Wickham et al. 2020).

Contaminant levels in muscle tissue samples were compared to state, national, and international limits and standards in order to address seafood safety and public health issues. These included: (1) fish contaminant

goals for chlordane, DDT, methylmercury, selenium, and PCBs developed by the California Office of Environmental Health Hazard Assessment (OEHHA) (Klasing and Brodberg 2008); (2) action limits on the amount of mercury, DDT, and chlordane in seafood that can be sold for human consumption, which are set by the U.S. Food and Drug Administration (USFDA) (Mearns et al. 1991); (3) international standards for acceptable concentrations of various metals and DDT (Mearns et al. 1991).

RESULTS

Contaminants in Fish Liver Tissues

Concentrations of trace metals, pesticides, PCBs, and PAHs detected in fish liver tissue samples from PLOO and SBOO trawl zones during 2020 are summarized by species in Tables 5.2 and 5.3. Raw data, including individual constituents summed for total chlordane, tDDT, tHCH, tPCB, and tPAH, are listed by sample in Appendices D.4 – D.7.

Contaminants in Fish Muscle Tissues

Concentrations of trace metals, pesticides, PCBs, and PAHs detected in fish muscle tissue samples from PLOO and SBOO rig fishing zones during 2020 are summarized by species in Tables 5.4 and 5.5. Raw data, including individual constituents summed for total chlordane, tDDT, tHCH, tPCB, and tPAH, are listed by sample in Appendices D.8 – D.11.

SUMMARY

Preliminary analysis of fish tissue data collected in 2020 provide no evidence of contaminant accumulation in PLOO or SBOO fishes associated with wastewater discharge from either outfall, which is consistent with historical findings (City of San Diego 2020a). Concentrations of most contaminants were generally similar across trawl or rig fishing zones, and no relationships with the PLOO or SBOO were evident. These results are consistent with findings of other assessments of bioaccumulation in fishes off San Diego (City of San Diego 2020a, Parnell et al. 2008). Finally, there were no other indications of poor fish health in the region, such as the presence of fin rot or other indicators of disease (see Chapter 4).

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CHAPTER 5
FIGURES & TABLES

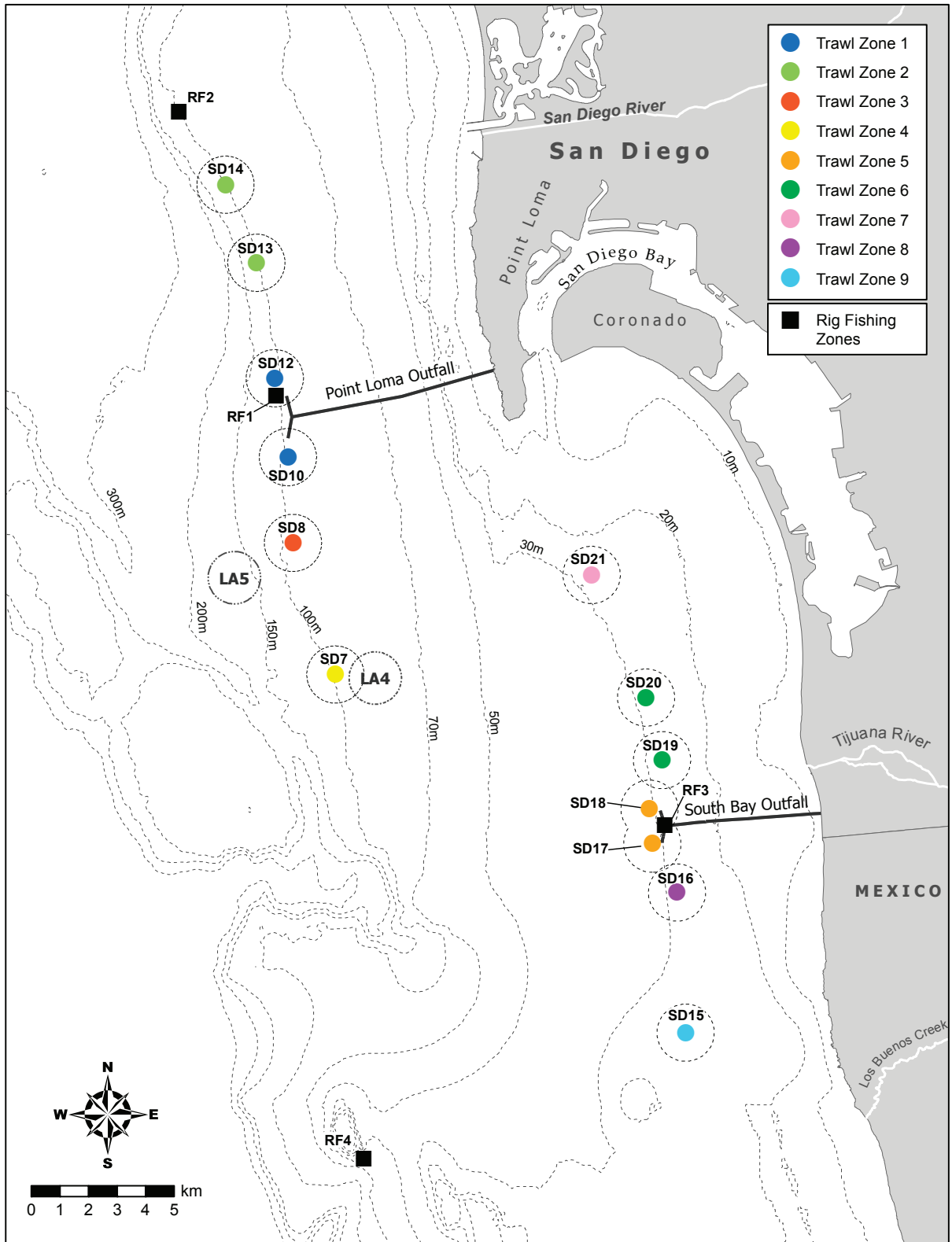


Figure 5.1

Trawl and rig fishing zone locations sampled around the PLOO and SBOO as part of the City of San Diego's Ocean Monitoring Program.

Table 5.1

Species of fish collected from each PLOO and SBOO trawl and rig fishing zone during 2020.

	Zone	Composite 1	Composite 2	Composite 3
PLOO	Rig Fishing Zone 1 (RF1)	Vermilion Rockfish	Vermilion Rockfish	Vermilion Rockfish
	Rig Fishing Zone 2 (RF2)	Starry Rockfish	Starry Rockfish	Mixed Rockfish ^a
	Trawl Zone 1 (TZ1)	Pacific Sanddab	Pacific Sanddab	Pacific Sanddab
	Trawl Zone 2 (TZ2)	Pacific Sanddab	Pacific Sanddab	Pacific Sanddab
	Trawl Zone 3 (TZ3)	Pacific Sanddab	Pacific Sanddab	Pacific Sanddab
	Trawl Zone 4 (TZ4)	Pacific Sanddab	Pacific Sanddab	Longfin Sanddab
SBOO	Rig Fishing Zone 3 (RF3)	California Scorpionfish	Mixed Rockfish ^b	Mixed Rockfish ^c
	Rig Fishing Zone 4 (RF4)	Mixed Rockfish ^b	California Scorpionfish	California Scorpionfish
	Trawl Zone 5 (TZ5)	Longfin Sanddab	Hornyhead Turbot	Spotted Turbot
	Trawl Zone 6 (TZ6)	Longfin Sanddab	Longfin Sanddab	Longfin Sanddab
	Trawl Zone 7 (TZ7)	Longfin Sanddab	Longfin Sanddab	Longfin Sanddab
	Trawl Zone 8 (TZ8)	Hornyhead Turbot	Longfin Sanddab	California Scorpionfish
	Trawl Zone 9 (TZ9)	Spotted Turbot	No sample	No sample

^aIncludes Flag, Speckled, and Starry Rockfish; ^bincludes Gopher Rockfish and Treefish; ^cincludes Brown and Olive Rockfish

Table 5.2

Summary of metals (ppm) in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. Data include the number of detected values (n), minimum, maximum, and mean detected concentrations for each species, and the total number of samples, detection rate, and maximum value for all species within each region. Minimum and maximum values were calculated based on all samples, whereas means were calculated from detected values only; ND = not detected.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Tl	Sn	Zn	
PLOO																			
Longfin Sanddab																			
n	0	0	1	0	0	1	0	1	1	0	1	1	0	0	0	0	0	1	1
value	—	—	11.1	—	—	3.36	—	7.82	135	—	0.606	0.175	—	—	—	—	—	34.6	—
Pacific Sanddab																			
n	0	0	11	0	0	11	0	11	11	0	10	11	1	7	0	0	10	11	11
min	—	—	4.28	—	—	2.11	—	4.32	81.4	—	ND	0.097	ND	ND	—	—	ND	24.4	—
max	—	—	10.8	—	—	4.6	—	8.19	136	—	0.926	0.195	0.121	2.35	—	—	7.96	35.1	—
mean	—	—	7.11	—	—	2.83	—	5.94	101.18	—	0.774	0.138	0.121	2.35	—	—	7.96	29.05	—
Total Samples	12	12	12	12	12	12	12	12	12	12	12	12	12	12	12	12	12	12	12
Detection Rate (%)	0	0	100	0	0	100	0	100	100	0	92	100	8	58	0	0	83	100	100
Max	ND	ND	11.1	ND	ND	4.6	ND	8.19	136	ND	0.926	0.195	0.121	2.35	ND	ND	7.96	35.1	—
SBOO																			
CA Scorpionfish																			
n	0	0	1	0	0	1	0	1	1	0	1	1	0	0	0	0	0	1	1
value	—	—	1.75	—	—	4.3	—	30.1	154	—	0.67	0.174	—	—	—	—	—	141	—
Hornyhead Turbot																			
n	0	0	2	0	0	2	0	2	2	0	2	2	0	0	0	0	0	2	2
min	—	—	3.4	—	—	3.77	—	6.21	67.6	—	0.914	0.095	—	—	—	—	—	65.1	—
max	—	—	3.93	—	—	5.04	—	7.56	78.9	—	1.06	0.108	—	—	—	—	—	75.5	—
mean	—	—	3.67	—	—	4.41	—	6.89	73.25	—	0.987	0.102	—	—	—	—	—	70.3	—
Longfin Sanddab																			
n	0	0	8	0	0	8	0	8	8	0	7	8	0	0	0	0	0	8	8
min	—	—	3.44	—	—	0.45	—	3.12	44.4	—	ND	0.028	—	—	—	—	—	13.5	—
max	—	—	12.1	—	—	2.97	—	8.42	117	—	1.19	0.092	—	—	—	—	—	27	—
mean	—	—	5.95	—	—	0.938	—	5.85	80.01	—	0.866	0.050	—	—	—	—	—	22.06	—
Spotted Turbot																			
n	0	0	2	0	0	2	0	2	2	1	2	2	0	0	0	0	0	2	2
min	—	—	11.4	—	—	2	—	12.5	133	ND	1.49	0.076	—	—	—	—	—	57.4	—
max	—	—	21.2	—	—	4.2	—	19.5	149	0.288	1.58	0.093	—	—	—	—	—	73.6	—
mean	—	—	16.3	—	—	3.1	—	16	141	0.288	1.535	0.085	—	—	—	—	—	65.5	—
Total Samples	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13
Detection Rate (%)	0	0	100	0	0	100	0	100	100	8	92	100	0	0	0	0	0	100	100
Max	ND	ND	21.2	ND	ND	5.04	ND	30.1	154	0.288	1.58	0.174	ND	ND	ND	ND	ND	141	—

Table 5.3

Summary of pesticides (ppb), total PCB (ppb), total PAH (ppb), and lipids (% weight) in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. Data include the number of detected values (n), minimum, maximum, and mean detected concentrations for each species, and the total number of samples, detection rate and maximum value for all species within each region. Minimum and maximum values were based on all samples, whereas means were calculated from detected values only; ND = not detected. See Appendices D.5–D.7 for values of individual constituents summed for total chlordane (tChlor), tDDT, tHCH, tPCB, and tPAH.

	Pesticides					tPCB	tPAH	Lipids	
	tChlor	tDDT	HCB	tHCH	Mirex				
PLOO	Longfin Sanddab								
	n	1	1	0	1	0	1	1	1
	value	8.39	362.08	—	0.52	—	604.9	67.5	35.2
	Pacific Sanddab								
	n	11	11	4	9	5	11	5	11
	min	8.09	329.87	14.1	ND	ND	ND	35.8	19.5
	max	18.41	761.93	126	2.63	5.8	890.49	132.6	38.1
	mean	11.66	492.29	52.2	1.347	2.29	670.32	66.48	31.42
	Total Samples	12	12	4	12	12	12	6	12
	Detection Rate (%)	100	100	100	83	42	100	100	100
Max	18.41	761.93	126	2.63	5.8	890.49	132.6	38.1	
SBOO	CA Scorpionfish								
	n	1	1	0	0	0	1	0	1
	value	3.06	153.5	—	—	—	85.84	—	9.02
	Hornyhead Turbot								
	n	2	2	0	0	0	2	0	2
	min	0.47	32.23	—	—	—	11.48	—	5.08
	max	0.55	33.89	—	—	—	15.41	—	6.78
	mean	0.51	33.06	—	—	—	13.45	—	5.93
	Longfin Sanddab								
	n	8	8	0	8	1	8	0	8
	min	3.54	133.99	—	1.04	ND	93.04	—	29.5
	max	7.58	256.77	—	1.65	2.37	245.07	—	40.4
	mean	4.51	192.83	—	1.45	2.37	145.49	—	35.03
	Spotted Turbot								
	n	1	2	0	0	0	2	0	2
	min	ND	9.79	—	—	—	5.74	—	2.5
	max	0.41	17.94	—	—	—	29.87	—	4.69
	mean	0.41	13.865	—	—	—	17.81	—	3.60
Total Samples	13	13	2	13	13	13	0	13	
Detection Rate (%)	92	100	0	62	8	100	—	100	
Max	7.58	256.77	ND	1.65	2.37	245.07	—	40.4	

Table 5.4

Summary of metals (ppm) in muscle tissues of fishes collected from PLOO and SBOO rig fishing zones during 2020. Data include the number of detected values (n), minimum, maximum, and mean detected concentrations per species and the total number of samples, detection rate, and maximum value for all species within each region. Minimum and maximum values based on all samples, whereas means were calculated from detected values only; ND = not detected.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Tl	Sn	Zn	
PLOO																			
Mixed Rockfish																			
n	0	0	1	0	0	0	0	1	1	0	0	1	0	1	0	0	0	1	
value	—	—	1.28	—	—	—	—	0.213	4.51	—	—	0.122	—	0.878	—	—	—	3.02	
Starry Rockfish																			
n	0	0	2	0	0	0	0	2	2	0	0	2	0	0	0	0	0	2	
min	—	—	1.04	—	—	—	—	0.269	1.02	—	—	0.132	—	—	—	—	—	2.87	
max	—	—	1.1	—	—	—	—	0.299	1.79	—	—	0.233	—	—	—	—	—	3.01	
mean	—	—	1.07	—	—	—	—	0.284	1.405	—	—	0.183	—	—	—	—	—	2.94	
Vermilion Rockfish																			
n	0	0	3	0	0	1	0	3	3	0	0	3	0	1	0	0	0	3	
min	—	—	3.53	—	—	ND	—	0.22	2.67	—	—	0.062	—	ND	—	—	—	3.51	
max	—	—	4	—	—	0.018	—	0.446	3.08	—	—	0.077	—	0.817	—	—	—	4.28	
mean	—	—	3.78	—	—	0.018	—	0.316	2.86	—	—	0.068	—	0.817	—	—	—	3.95	
Total Samples	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	
Detection Rate (%)	0	0	100	0	0	17	0	100	100	0	0	100	0	33	0	0	0	100	
Max	ND	ND	4	ND	ND	0.018	ND	0.446	4.51	ND	ND	0.233	ND	0.878	ND	ND	ND	4.28	
SBOO																			
California Scorpionfish																			
n	0	0	3	0	0	0	0	3	3	0	0	3	0	0	0	0	0	3	
min	—	—	1.73	—	—	—	—	0.241	1.87	—	—	0.164	—	—	—	—	—	3.9	
max	—	—	2.65	—	—	—	—	0.602	4.74	—	—	0.218	—	—	—	—	—	4.59	
mean	—	—	2.21	—	—	—	—	0.430	3.10	—	—	0.186	—	—	—	—	—	4.26	
Mixed Rockfish																			
n	1	0	3	0	0	0	0	3	3	0	0	3	0	0	0	0	0	3	
min	ND	—	1.14	—	—	—	—	0.182	1.29	—	—	0.059	—	—	—	—	—	4.2	
max	2.05	—	1.87	—	—	—	—	0.602	1.91	—	—	0.137	—	—	—	—	—	4.5	
mean	2.05	—	1.43	—	—	—	—	0.329	1.65	—	—	0.109	—	—	—	—	—	4.37	
Total Samples	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	
Detection Rate (%)	17	0	100	0	0	0	0	100	100	0	0	100	0	0	0	0	0	100	
Max	2.05	ND	2.65	ND	ND	ND	ND	0.602	4.74	ND	ND	0.218	ND	ND	ND	ND	ND	4.59	

Table 5.5

Summary of pesticides (ppb), total PCB (ppb), total PAH (ppb), and lipids (% weight) in muscle tissues of fishes collected from PLOO and SBOO rig fishing stations during 2020. Data include the number of detected values (n), minimum, maximum, and mean detected concentrations for each species, and the total number of samples, detection rate and maximum value for all species within region. Minimum and maximum values were based on all samples, whereas means were calculated from detected values only; aEndSul=Alpha Endosulfan; bEndSul=Beta Endosulfan; EndSulSul=Endosulfan Sulfate; ND=not detected. See Appendices D.9–D.11 for values of individual constituents summed for total chlordane (tChlor), tDDT, tHCH, Endrin, aEndSul, bEndSul, EndSulSul, tPCB, tPAH, and Lipids.

	Pesticide											tPAH	Lipids	
	tChlor	tDDT	Dieldrin	HCB	tHCH	Endrin	aEndSul	bEndSul	EndSulSul	tPCB	tPAH			
PLOO														
Mixed Rockfish														
n	1	1	0	1	1	0	0	1	0	1	0	1	1	1
value	0.22	4.81	—	0.14	0.14	—	—	—	—	0.05	—	3.03	340	0.22
Starry Rockfish														
n	1	2	0	2	0	0	0	0	0	0	0	2	2	2
min	ND	1.74	—	0.09	—	—	—	—	—	—	—	0.81	21	0.15
max	0.04	4.63	—	0.11	—	—	—	—	—	—	—	2.93	41.4	0.19
mean	0.04	3.19	—	0.1	—	—	—	—	—	—	—	1.87	31.2	0.17
Vermilion Rockfish														
n	2	3	1	3	2	0	0	0	0	0	0	3	3	3
min	ND	4.3	ND	0.11	ND	—	—	—	—	—	—	3.94	21.6	0.21
max	0.22	6.77	0.08	0.17	0.05	—	—	—	—	—	—	10.15	299	0.39
mean	0.17	5.55	0.08	0.147	0.045	—	—	—	—	—	—	6.21	115.93	0.317
Total samples	6	6	6	6	6	6	6	6	6	6	6	6	6	6
Detection Rate (%)	67	100	17	100	50	0	0	17	0	100	0	100	100	100
Max	0.22	6.77	0.08	0.17	0.14	0	0	0.05	0	10.145	0	10.145	340	0.39
SBOO														
California Scorpionfish														
n	2	3	0	2	1	1	0	1	0	1	0	3	2	3
min	ND	1.44	—	ND	ND	ND	—	ND	—	ND	—	0.57	ND	0.09
max	0.22	13.43	—	0.19	0.04	0.05	—	0.05	—	0.05	—	6.56	63.2	0.2
mean	0.13	5.87	—	0.165	0.04	0.05	—	0.05	—	0.05	—	2.64	39.7	0.15
Mixed Rockfish														
n	3	3	3	1	2	1	1	1	1	1	1	3	3	3
min	0.1	1.57	0.04	ND	ND	ND	ND	ND	ND	ND	ND	0.71	34.2	0.12
max	0.64	2.79	0.26	0.1	0.33	0.67	0.18	0.05	0.06	0.05	0.06	1.88	485.9	0.18
mean	0.29	2.03	0.12	0.1	0.225	0.67	0.18	0.05	0.06	0.05	0.06	1.28	185.45	0.15
Total samples	6	6	6	6	6	6	6	6	6	6	6	6	6	6
Detection Rate (%)	83	100	50	50	50	33	17	33	17	100	17	100	83	100
Max	0.64	13.43	0.26	0.19	0.33	0.67	0.18	0.05	0.06	6.56	0.06	6.56	485.9	0.2

Appendices

Appendix A
Water Quality
2020 Raw Data Summaries

Appendix A.1

Summary of PLOO and SBOO reference stations used during 2020 to calculate out-of-range thresholds (see text for details).

Month	Stations
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February

PLOO A1, A6, A7, C7, C8, F01, F02, F03, F04, F05, F06, F07, F08, F09, F10, F11, F12, F13, F14, F25, F36

SBOO I1, I6, I7, I8, I9, I10, I13, I14, I15, I16, I2, I20, I21, I28, I29, I3, I30, I33, I34, I35

May

PLOO A1, A7, F01

SBOO I1, I3, I6, I7, I8, I9, I10, I13, I18, I2, I21

August

PLOO A1, A6, A7, C8, F01, F02, F03, F04, F05, F06, F07, F09, F10, F11, F12, F13, F14

SBOO I1, I8, I12, I14, I16, I17, I2, I21, I34, I39

November

PLOO A1, A6, C7, F04, F05, F06, F07, F08, F09, F10, F11, F12, F13, F15, F17, F18, F21, F22, F24

SBOO I1, I3, I6, I7, I8, I9, I13, I14, I17, I18, I2, I20, I21, I28, I29

Appendix A.2

Summary of temperature, salinity, DO, pH, transmissivity, and chlorophyll *a* for various depth layers as well as the entire water column for all PLOO stations during 2020. For each quarter: $n \geq 3381$ (1–20 m), $n \geq 5283$ (21–60 m), $n \geq 1851$ (61–80 m), $n \geq 994$ (81–98 m). Sample sizes differed due to variations in bottom depth at individual stations.

		Depth (m)				
		1–20	21–60	61–80	81–98	1–98
Temperature (°C)						
<i>February</i>	min	13.4	11.8	11.7	11.2	11.2
	max	15.9	15.5	13.1	12.1	15.9
	mean	15.4	13.9	12.2	11.6	13.8
<i>May</i>	min	11.2	10.1	10.0	9.7	9.7
	max	19.8	12.4	10.6	10.3	19.8
	mean	14.2	10.8	10.2	10.0	11.7
<i>August</i>	min	12.9	10.9	10.6	10.5	10.5
	max	23.9	16.4	11.9	10.9	23.9
	mean	18.8	12.7	11.0	10.7	14.0
<i>November</i>	min	12.6	11.7	11.2	10.8	10.8
	max	17.7	16.4	12.6	11.8	17.7
	mean	15.9	13.0	11.9	11.2	13.5
Salinity (ppt)						
<i>February</i>	min	33.40	33.42	33.48	33.58	33.40
	max	33.60	33.60	33.67	33.85	33.85
	mean	33.51	33.52	33.59	33.69	33.54
<i>May</i>	min	33.63	33.67	33.83	33.90	33.63
	max	33.98	33.92	33.96	34.04	34.04
	mean	33.73	33.80	33.90	33.96	33.81
<i>August</i>	min	33.31	33.31	33.55	33.68	33.31
	max	33.84	33.68	33.85	33.90	33.90
	mean	33.57	33.50	33.67	33.80	33.57
<i>November</i>	min	33.45	33.44	33.56	33.66	33.44
	max	33.63	33.70	33.79	33.85	33.85
	mean	33.56	33.54	33.66	33.77	33.59

Appendix A.2 *continued*

		Depth (m)				
		1–20	21–60	61–80	81–98	1–98
DO (mg/L)						
<i>February</i>	min	6.5	4.7	4.2	3.9	3.9
	max	8.5	8.3	6.8	6.3	8.5
	mean	7.9	7.2	5.4	4.6	6.9
<i>May</i>	min	1.9	2.9	3.0	3.1	1.9
	max	9.3	5.6	4.0	3.9	9.3
	mean	5.5	4.0	3.7	3.5	4.3
<i>August</i>	min	5.1	4.9	3.9	3.7	3.7
	max	9.2	8.5	6.0	5.2	9.2
	mean	7.9	6.9	5.2	4.3	6.7
<i>November</i>	min	4.7	4.7	4.4	4.0	4.0
	max	7.7	7.4	5.9	5.0	7.7
	mean	7.0	5.9	5.1	4.4	6.0
pH						
<i>February</i>	min	7.9	7.8	7.7	7.7	7.7
	max	8.1	8.1	7.9	7.9	8.1
	mean	8.0	8.0	7.8	7.8	7.9
<i>May</i>	min	7.6	7.7	7.7	7.7	7.6
	max	8.4	8.0	7.8	7.8	8.4
	mean	8.0	7.8	7.8	7.8	7.9
<i>August</i>	min	7.9	7.8	7.8	7.7	7.7
	max	8.3	8.2	8.0	7.9	8.3
	mean	8.2	8.0	7.9	7.8	8.0
<i>November</i>	min	7.8	7.8	7.8	7.8	7.8
	max	8.2	8.1	8.0	7.9	8.2
	mean	8.1	7.9	7.9	7.8	8.0

Appendix A.2 *continued*

		Depth (m)				
		1–20	21–60	61–80	81–98	1–98
Transmissivity (%)						
<i>February</i>	min	79	84	28	55	28
	max	91	92	92	92	92
	mean	89	90	89	84	89
<i>May</i>	min	25	84	77	87	25
	max	93	93	93	93	93
	mean	74	92	91	91	86
<i>August</i>	min	22	84	86	80	22
	max	92	93	94	93	94
	mean	88	91	93	92	91
<i>November</i>	min	76	87	80	85	76
	max	92	94	94	94	94
	mean	88	92	93	93	91
Chlorophyll a (µg/L)						
<i>February</i>	min	0.2	0.3	0.2	0.2	0.2
	max	2.4	2.8	1.1	0.6	2.8
	mean	0.6	1.3	0.6	0.3	0.9
<i>May</i>	min	0.3	0.1	0.1	0.1	0.1
	max	31.8	3.3	3.7	0.7	31.8
	mean	6.2	0.7	0.4	0.3	2.2
<i>August</i>	min	0.3	0.4	0.2	0.2	0.2
	max	5.6	5.1	0.8	0.5	5.6
	mean	1.1	1.5	0.5	0.3	1.1
<i>November</i>	min	0.3	0.3	0.2	0.2	0.2
	max	4.8	3.1	0.6	0.3	4.8
	mean	1.4	0.9	0.3	0.2	0.9

Appendix A.3

Summary of temperature, salinity, DO, pH, transmissivity, and chlorophyll a for various depth layers as well as the entire water column from all SBOO stations during 2020. For each quarter: n≥1440 (1–9 m), n≥1236 (10–19 m), n≥737 (20–28 m), n≥349 (29–38 m), n≥289 (39–55 m). Sample sizes differed due to slight variations in depth at individual stations.

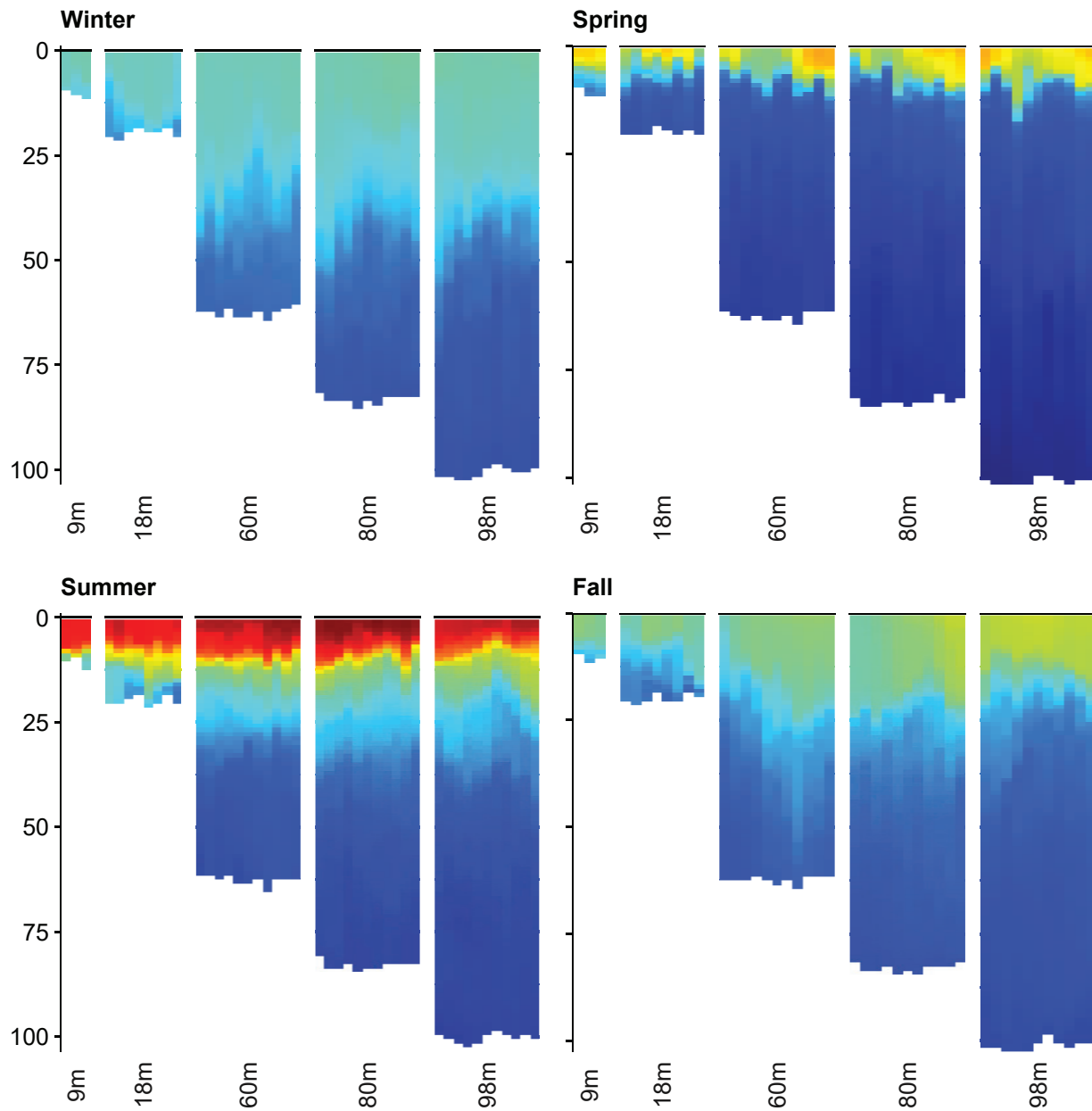
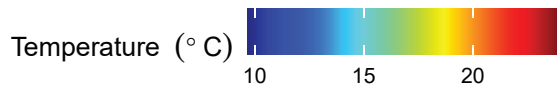
		Depth (m)					
		1–9	10–19	20–28	29–38	39–55	1–55
Temperature (°C)							
<i>February</i>	min	14.2	13.7	13.4	13.4	12.4	12.4
	max	15.5	15.4	15.3	15.4	15.1	15.5
	mean	14.9	14.6	14.3	14.3	13.7	14.6
<i>May</i>	min	12.2	11.6	11.4	11.3	10.5	10.5
	max	20.1	16.7	14.5	13.1	11.6	20.1
	mean	16.9	13.2	12.1	11.6	11.1	14.0
<i>August</i>	min	13.4	12.7	13.4	12.7	11.8	11.8
	max	23.8	20.7	17.6	15.1	14.1	23.8
	mean	19.8	16.1	14.3	13.5	12.9	16.7
<i>November</i>	min	15.1	14.9	14.3	13.4	12.7	12.7
	max	19.2	18.1	16.5	14.9	14.2	19.2
	mean	17.5	16.0	15.0	14.3	13.3	16.0
Salinity (ppt)							
<i>February</i>	min	33.27	33.36	33.46	33.47	33.49	33.27
	max	33.54	33.58	33.57	33.58	33.62	33.62
	mean	33.48	33.50	33.52	33.52	33.55	33.50
<i>May</i>	min	33.55	33.58	33.55	33.53	33.61	33.53
	max	34.28	33.84	33.76	33.76	33.87	34.28
	mean	33.73	33.68	33.67	33.65	33.73	33.70
<i>August</i>	min	33.37	33.27	33.38	33.38	33.41	33.27
	max	33.71	33.63	33.57	33.59	33.64	33.71
	mean	33.59	33.50	33.49	33.49	33.52	33.53
<i>November</i>	min	33.28	33.31	33.45	33.46	33.49	33.28
	max	33.64	33.57	33.49	33.52	33.58	33.64
	mean	33.50	33.47	33.47	33.48	33.53	33.48

Appendix A.3 *continued*

		Depth (m)					
		1–9	10–19	20–28	29–38	39–55	1–55
DO (mg/L)							
<i>February</i>	min	6.9	6.8	6.7	6.6	6.0	6.0
	max	8.3	8.2	7.9	7.9	7.8	8.3
	mean	7.9	7.7	7.4	7.4	6.9	7.6
<i>May</i>	min	1.3	0.6	4.0	3.9	3.5	0.6
	max	8.4	8.6	8.0	6.7	5.9	8.6
	mean	7.0	6.2	5.5	5.3	4.5	6.2
<i>August</i>	min	7.0	6.5	6.9	6.0	5.4	5.4
	max	9.9	9.5	8.7	8.5	8.2	9.9
	mean	8.2	8.3	7.7	7.3	6.7	7.9
<i>November</i>	min	6.4	5.0	6.0	5.4	5.7	5.0
	max	9.3	8.3	7.5	6.7	6.3	9.3
	mean	8.1	7.1	6.6	6.2	6.0	7.2
pH							
<i>February</i>	min	8.0	8.0	8.0	8.0	7.9	7.9
	max	8.1	8.1	8.1	8.1	8.1	8.1
	mean	8.1	8.1	8.0	8.0	8.0	8.1
<i>May</i>	min	7.6	7.5	7.8	7.8	7.7	7.5
	max	8.3	8.2	8.1	8.0	8.0	8.3
	mean	8.1	8.0	7.9	7.9	7.8	8.0
<i>August</i>	min	8.0	7.9	8.0	8.0	7.8	7.8
	max	8.3	8.3	8.2	8.1	8.1	8.3
	mean	8.2	8.2	8.1	8.0	8.0	8.1
<i>November</i>	min	8.1	8.0	8.0	8.0	7.9	7.9
	max	8.3	8.2	8.2	8.1	8.0	8.3
	mean	8.2	8.1	8.1	8.0	8.0	8.1

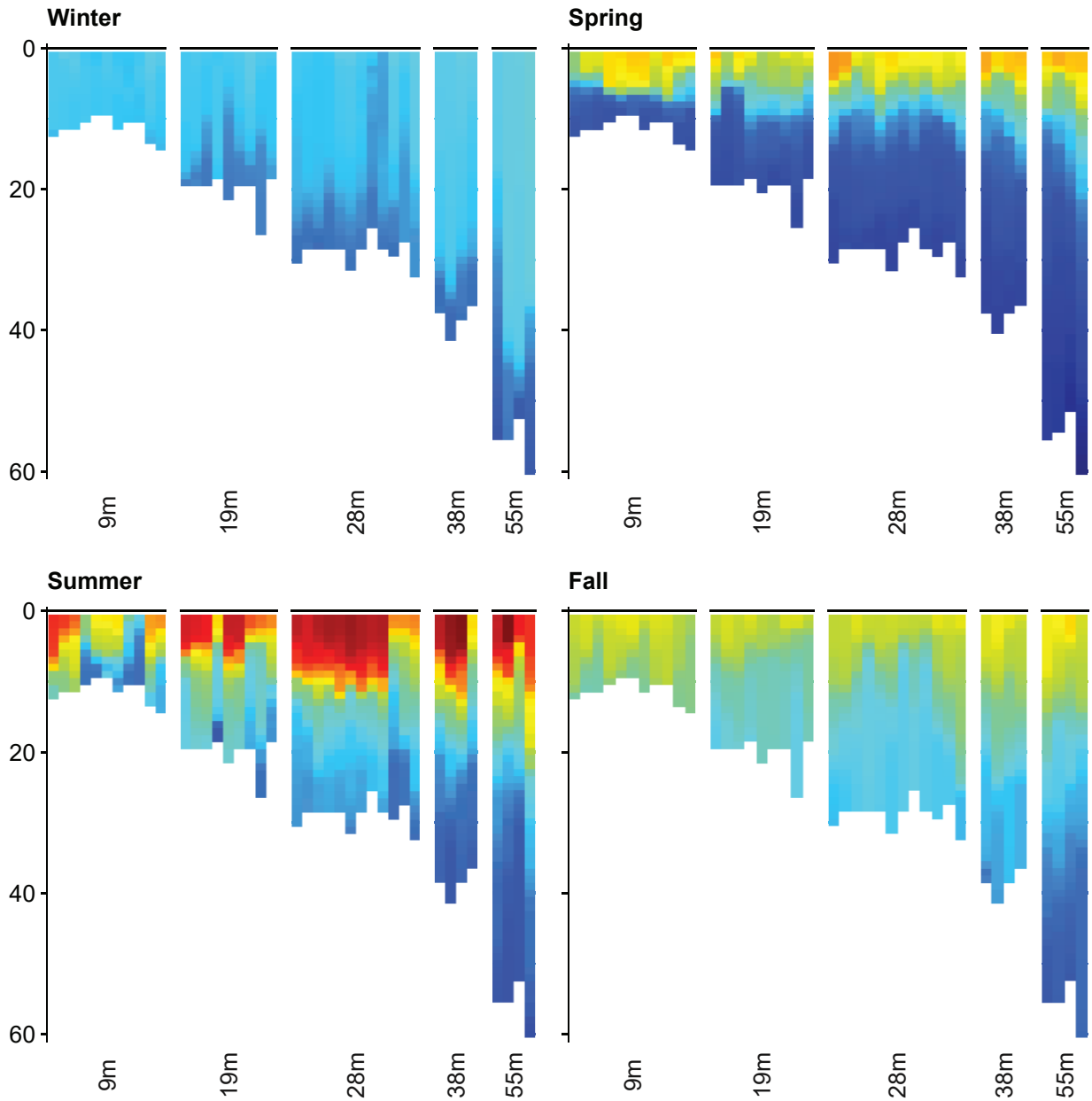
Appendix A.3 *continued*

		Depth (m)					
		1–9	10–19	20–28	29–38	39–55	1–55
Transmissivity (%)							
<i>February</i>	min	49	41	78	85	89	41
	max	91	91	91	91	91	91
	mean	83	86	89	90	90	86
<i>May</i>	min	47	50	73	85	91	47
	max	88	90	92	93	93	93
	mean	77	84	89	91	92	84
<i>August</i>	min	62	75	82	84	88	62
	max	92	92	92	92	92	92
	mean	83	85	87	88	90	85
<i>November</i>	min	69	74	85	87	91	69
	max	90	88	91	92	93	93
	mean	82	85	88	90	92	85
Chlorophyll a (µg/L)							
<i>February</i>	min	0.2	0.3	0.4	0.6	0.8	0.2
	max	3.5	3.0	2.4	2.0	1.5	3.5
	mean	1.2	1.5	1.4	1.2	1.1	1.3
<i>May</i>	min	0.6	0.7	0.8	0.7	0.3	0.3
	max	8.2	4.8	5.4	4.3	1.6	8.2
	mean	2.1	2.2	2.5	1.4	0.6	2.0
<i>August</i>	min	0.5	0.5	0.8	1.0	0.7	0.5
	max	12.3	7.9	5.4	5.0	3.7	12.3
	mean	2.3	2.9	2.8	2.9	1.9	2.6
<i>November</i>	min	0.7	1.1	1.0	0.5	0.4	0.4
	max	5.5	4.0	3.4	2.6	1.1	5.5
	mean	2.7	2.7	2.3	1.5	0.6	2.4



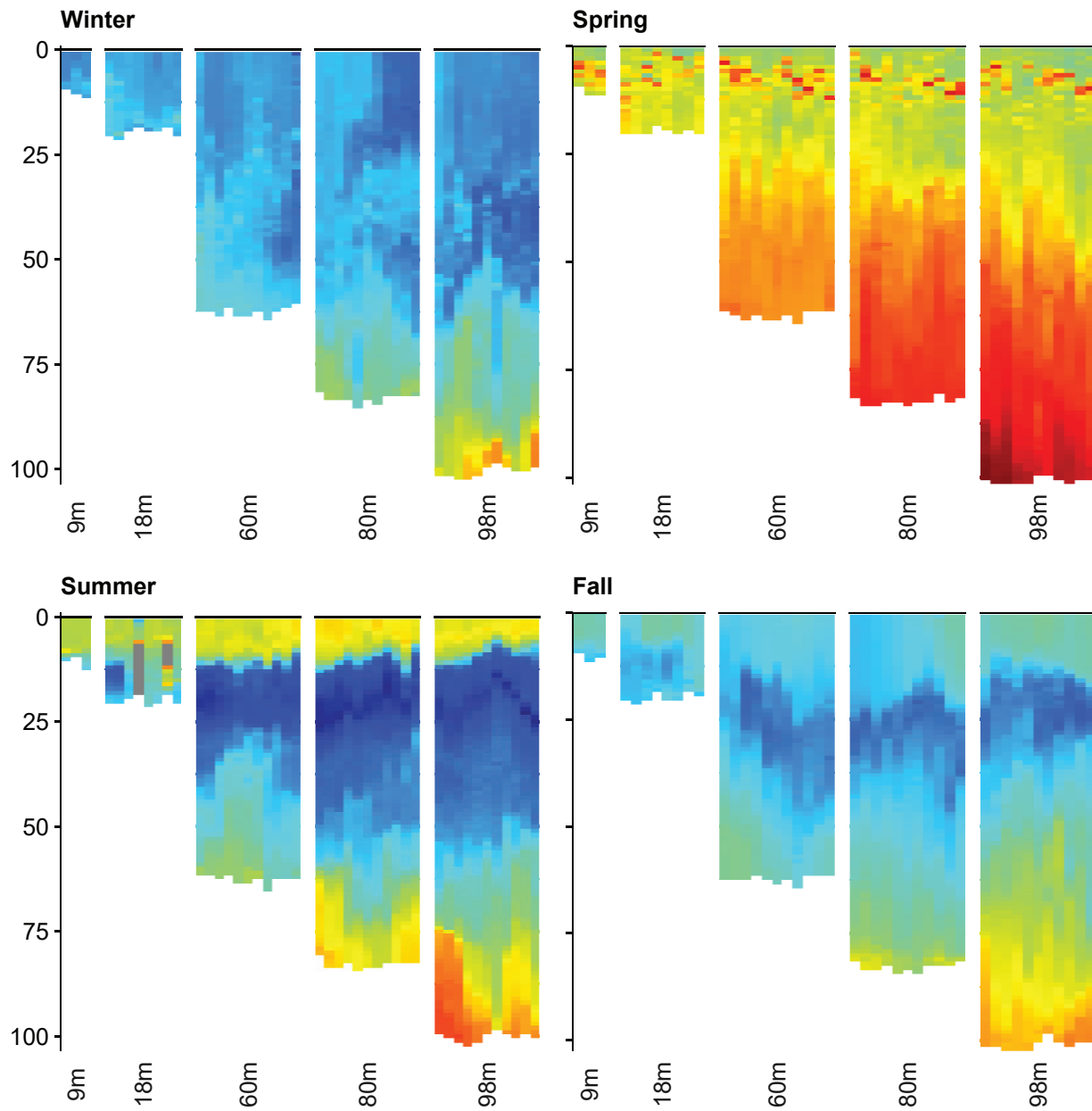
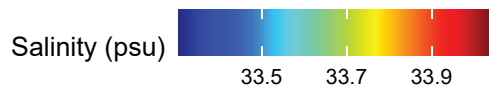
Appendix A.4

Temperature recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



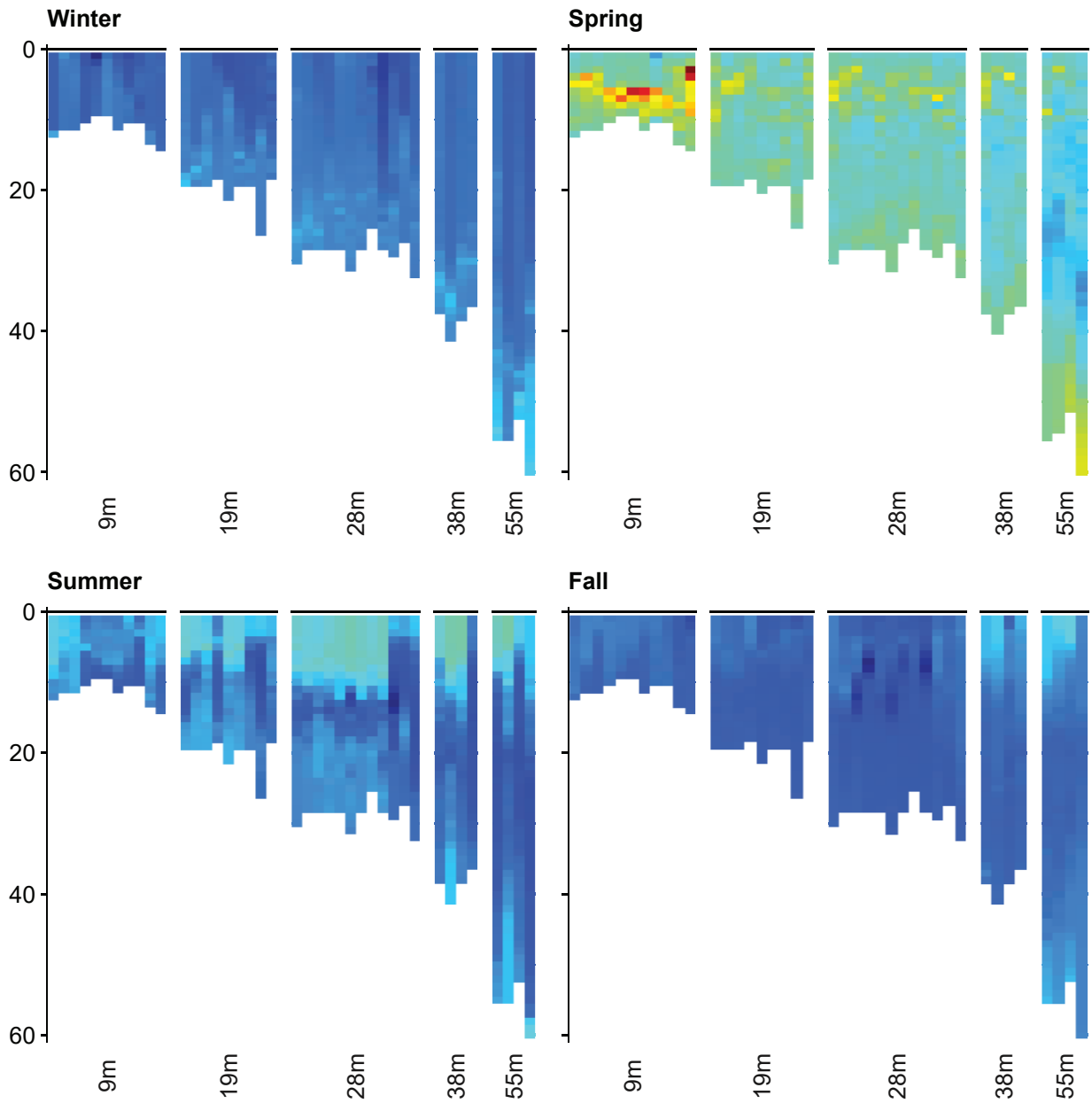
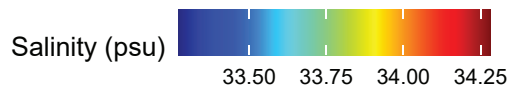
Appendix A.5

Temperature recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



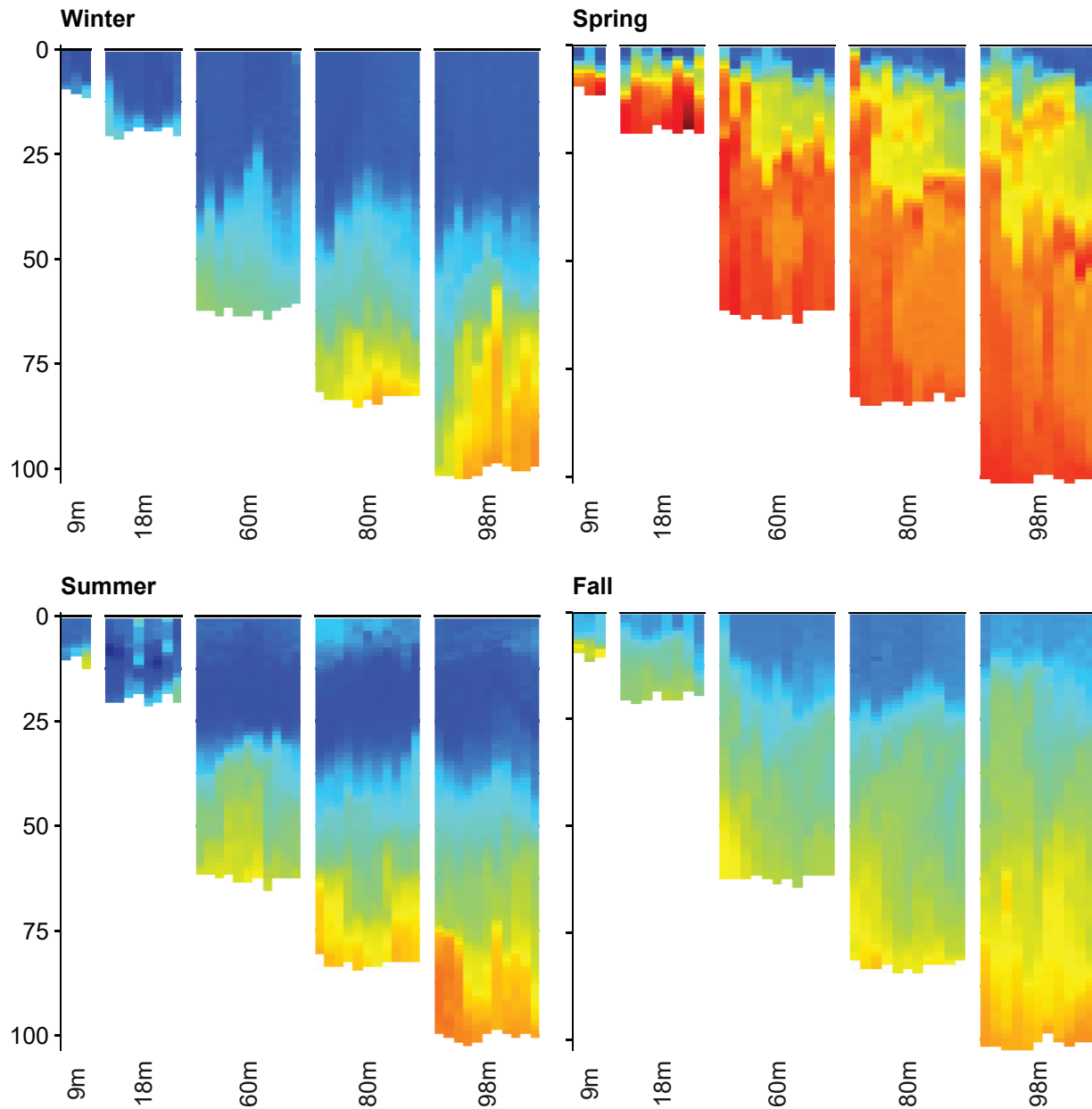
Appendix A.6

Salinity recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



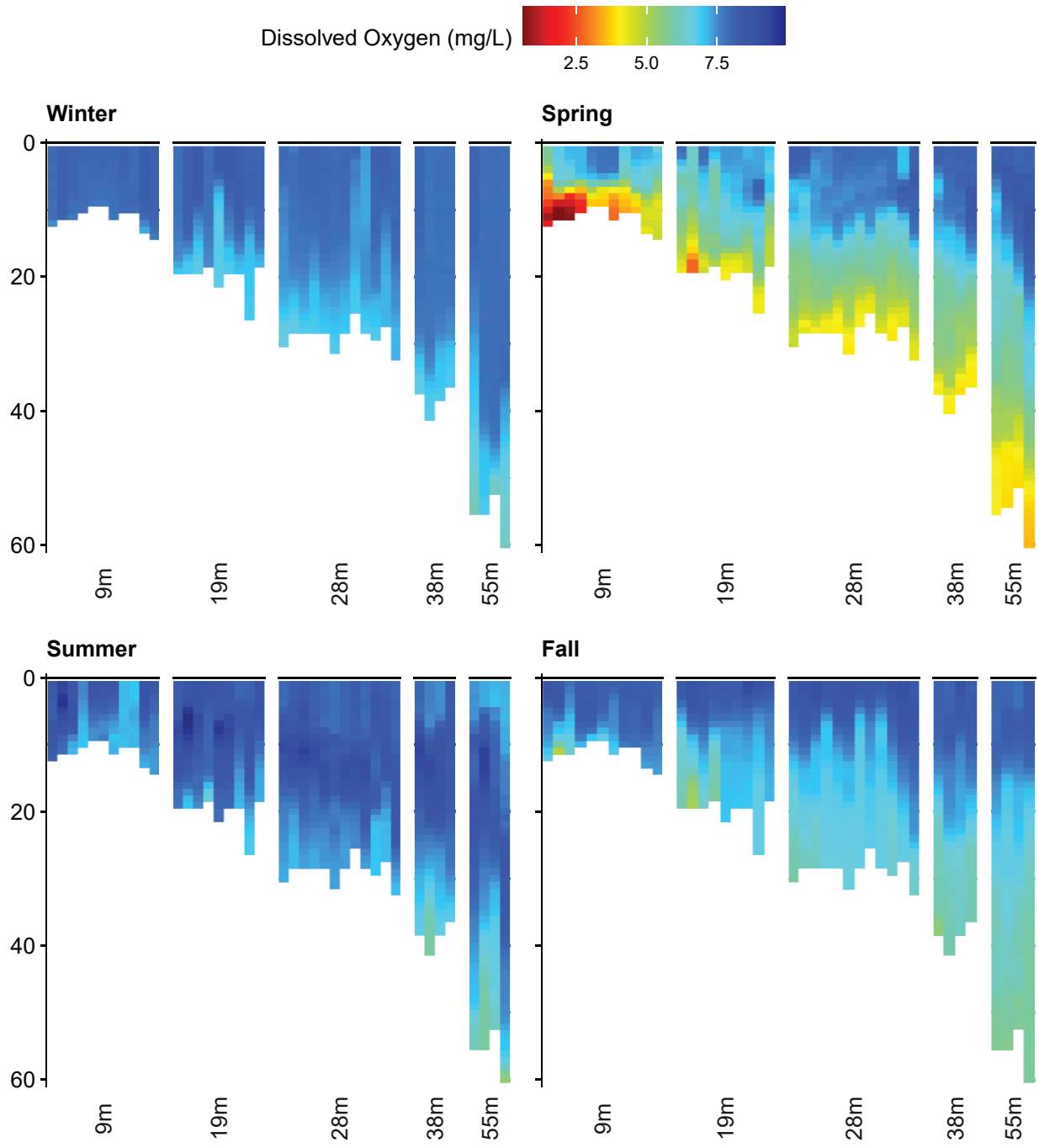
Appendix A.7

Salinity recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



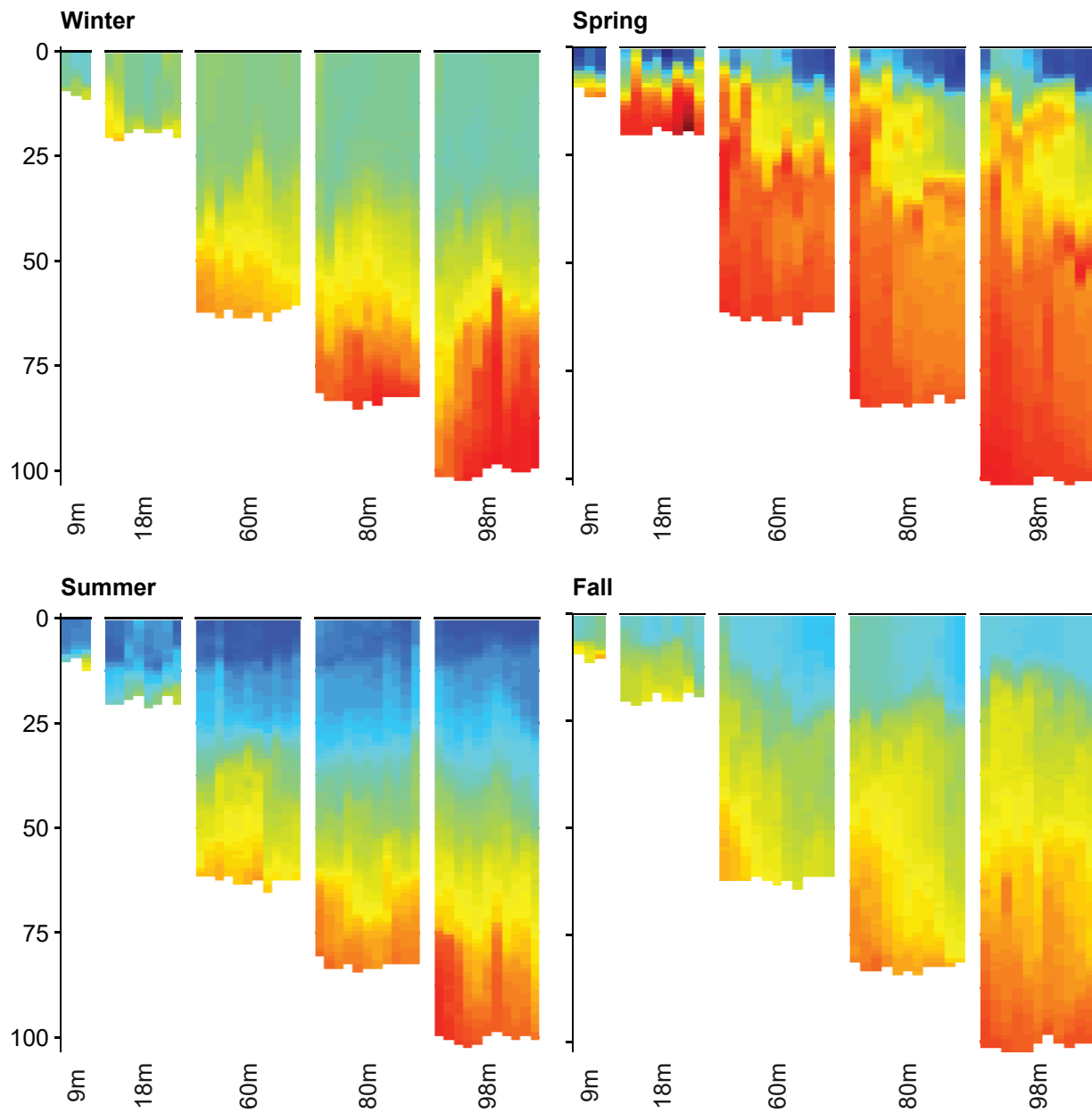
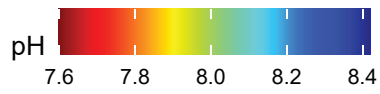
Appendix A.8

Dissolved oxygen recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



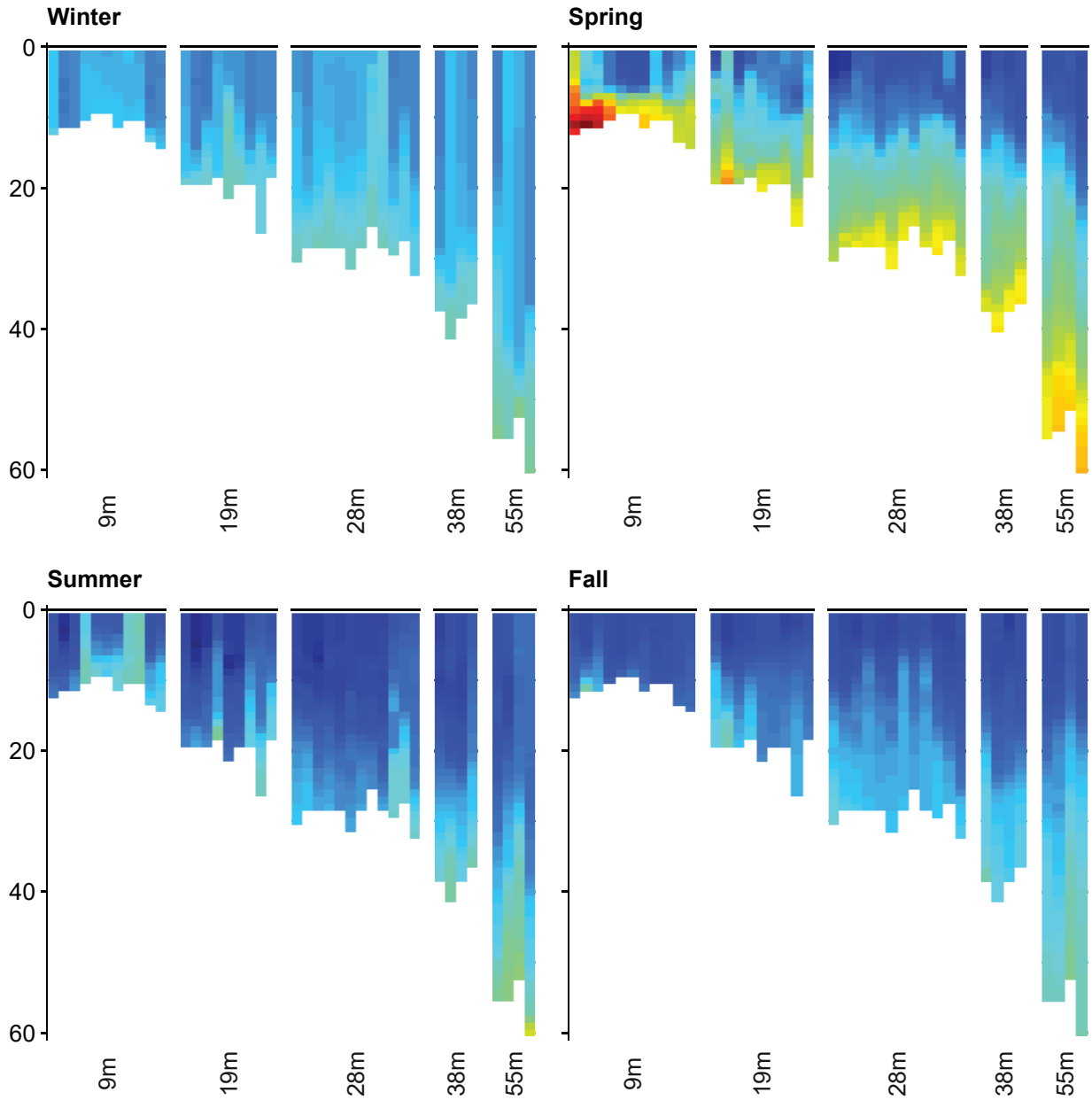
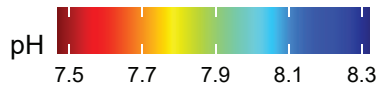
Appendix A.9

Dissolved oxygen recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



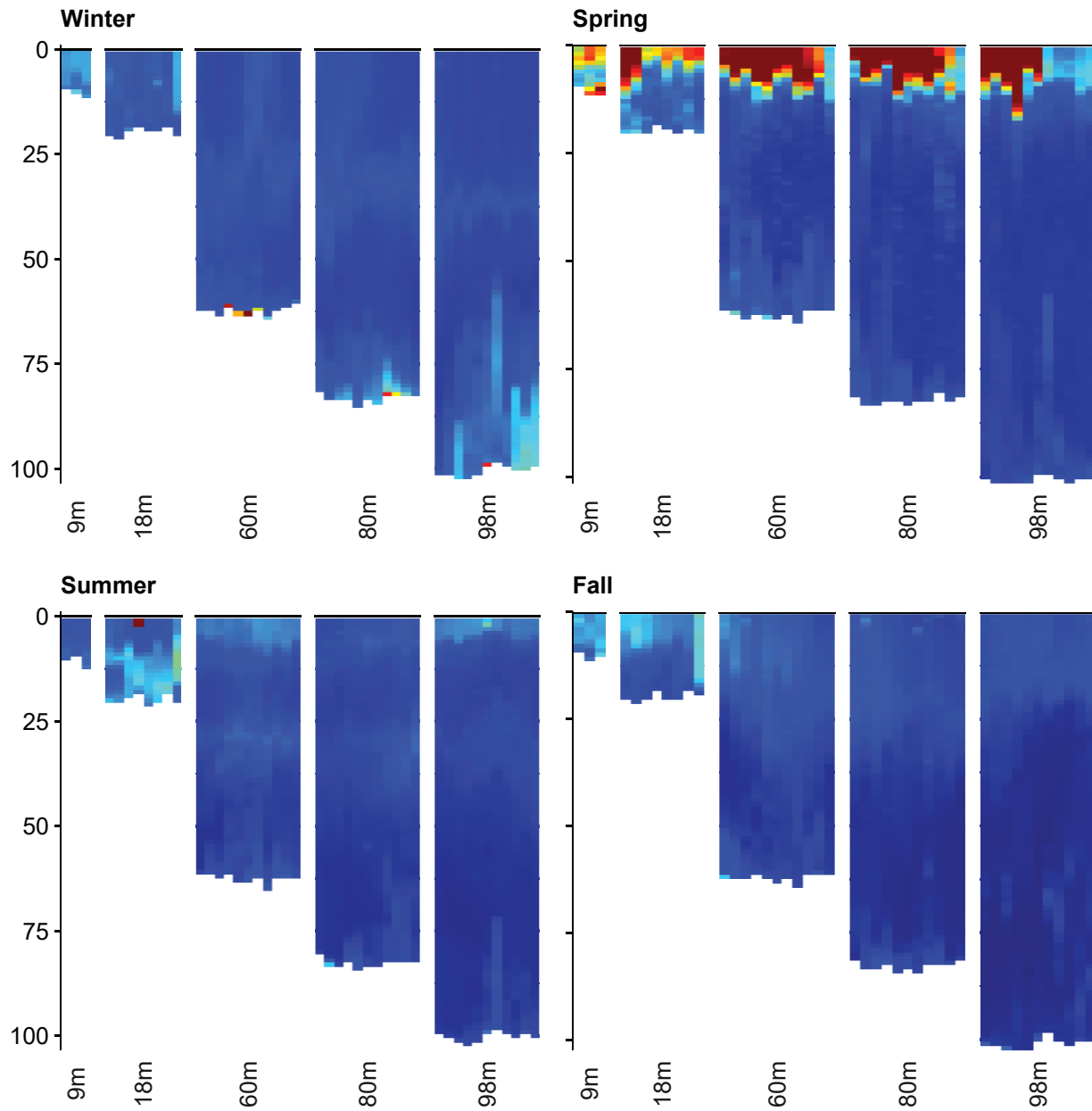
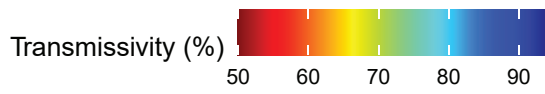
Appendix A.10

Values of pH recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



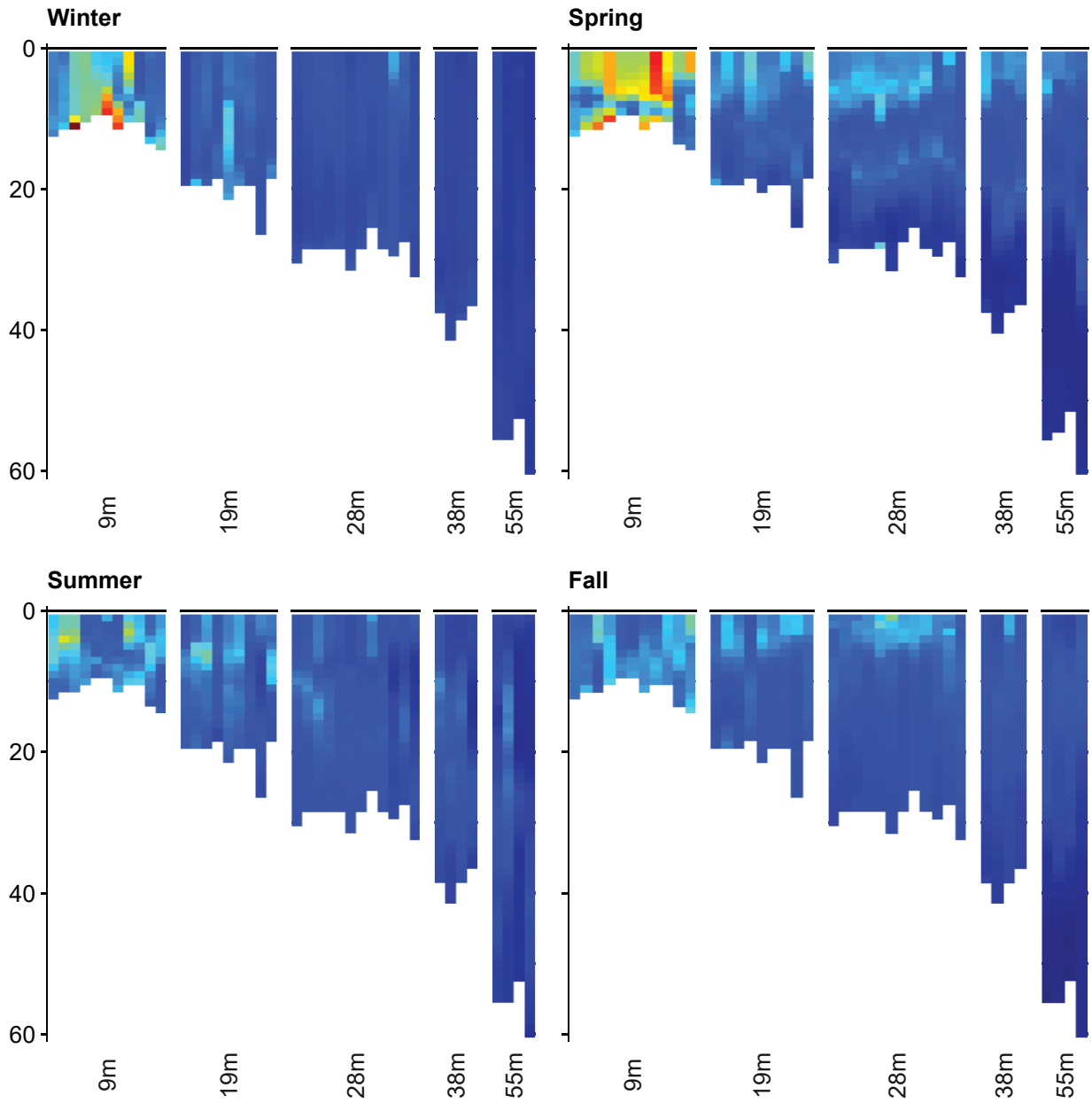
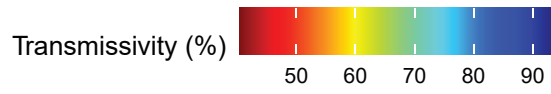
Appendix A.11

Values of pH recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



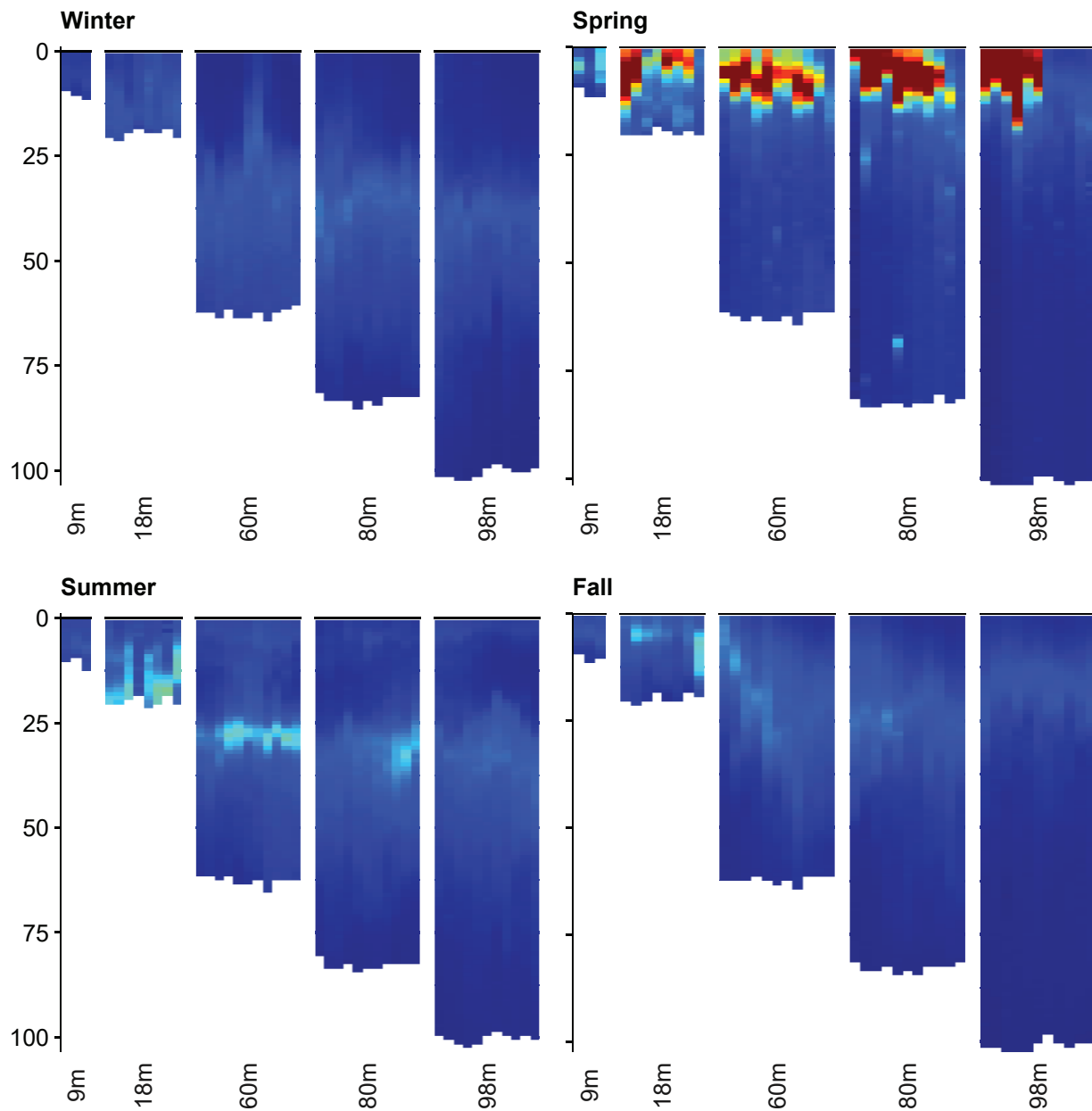
Appendix A.12

Transmissivity recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



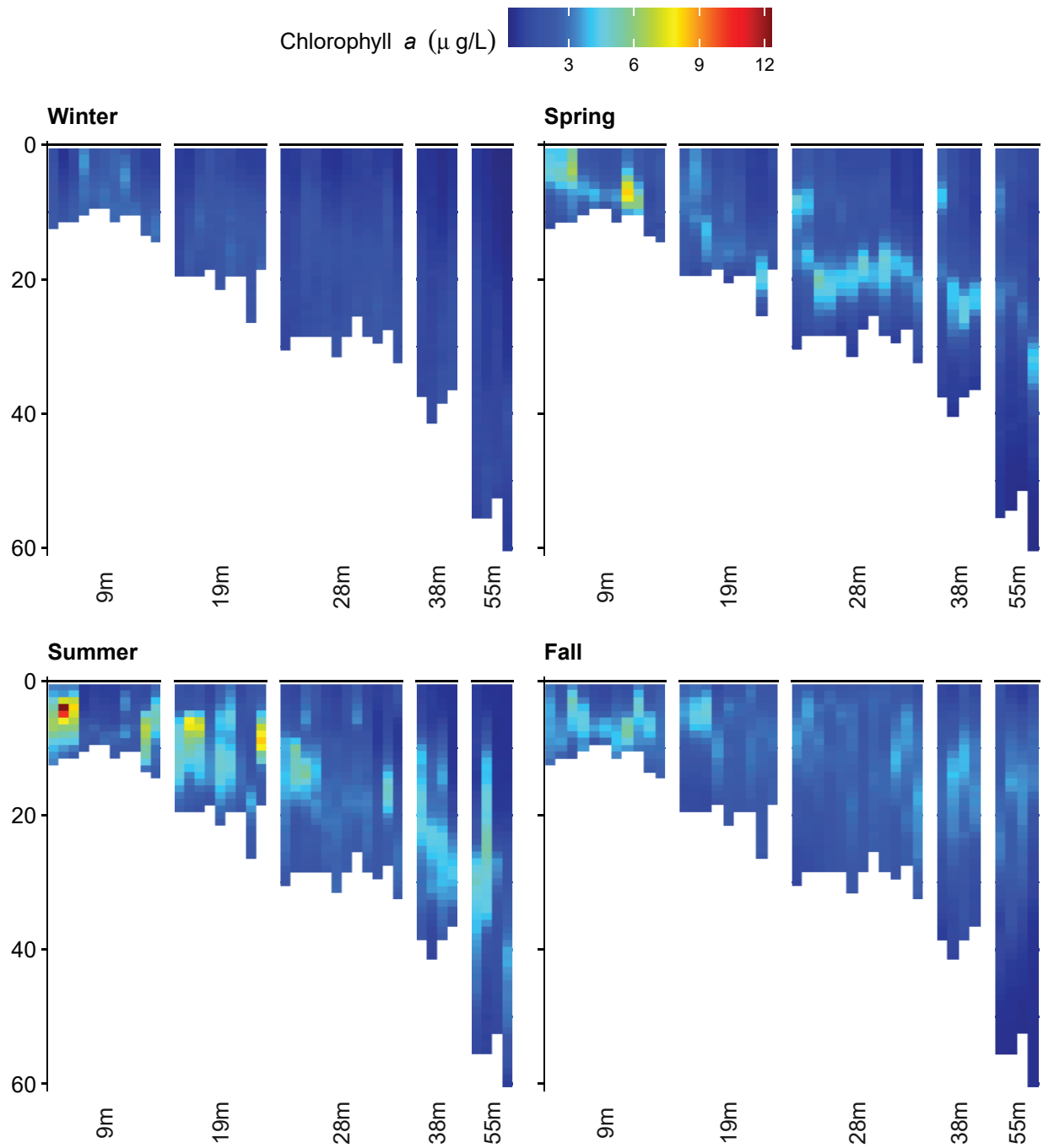
Appendix A.13

Transmissivity recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



Appendix A.14

Chlorophyll a recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



Appendix A.15

Chlorophyll a recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.

Appendix A.16

Summary of temperature, salinity, DO, pH (total), chlorophyll a, CDOM, turbidity, nitrate + nitrite, BOD, and xCO₂ recorded at various depths by the PLOO RTOMS in 2020. Data include mean, minimum, and maximum values, sample size (n), and proportion recovered (n_prop) for each depth by season. Sample sizes differed due to variations in sampling interval, deployment date, and data quality (Addenda 2-1 to 2-4); id=insufficient data (see text). The PLOO RTOMS was not deployed during Fall 2020.

Parameter	Season		1 m	10 m	20 m	30 m	45 m	60 m	75 m	89 m	
Temperature (°C)	Winter	mean	15.93	15.77	15.39	—	13.69	12.68	11.98	11.47	
		min	14.84	13.95	12.56	—	10.92	10.15	9.78	9.61	
		max	17.86	16.63	16.39	—	15.83	14.90	13.87	13.06	
		n	10,103	10,226	10,228	id	10,233	10,229	10,225	8257	
		n_prop	0.78	0.79	0.79	0.35	0.79	0.79	0.79	0.64	
	Spring	mean	18.87	15.82	13.17	—	10.94	10.43	10.19	10.06	
		min	14.76	11.81	10.93	—	10.05	9.84	9.58	9.29	
		max	22.46	20.36	19.03	—	15.07	13.17	12.12	11.22	
		n	12,267	12,430	12,420	0	12,425	12,432	12,434	12,312	
		n_prop	0.94	0.95	0.95	—	0.95	0.95	0.95	0.94	
	Summer	mean	21.21	16.85	13.83	—	11.80	11.22	10.93	10.76	
		min	15.16	11.74	11.13	—	10.42	10.14	9.94	9.84	
		max	25.46	23.27	18.80	—	14.30	13.24	12.74	12.30	
		n	11,949	12,163	12,159	0	12,119	12,109	12,104	11,971	
		n_prop	0.90	0.92	0.92	—	0.91	0.91	0.91	0.90	
	Salinity (psu)	Winter	mean	33.49	33.46	33.47	—	33.51	33.48	33.34	33.31
			min	33.32	33.29	33.34	—	33.32	33.19	32.89	32.97
			max	33.57	33.54	33.57	—	33.65	33.79	33.84	33.72
n			10,100	10,222	10,222	id	6315	10,227	10,224	8252	
n_prop			0.78	0.79	0.79	0.35	0.49	0.79	0.79	0.64	
Spring		mean	33.49	33.45	33.53	—	33.56	33.59	33.61	33.44	
		min	32.12	33.09	33.23	—	33.20	33.22	33.31	33.11	
		max	33.62	33.80	33.86	—	33.80	33.87	33.94	33.70	
		n	12,252	12,241	12,393	0	7916	12,431	12,434	12,312	
		n_prop	0.93	0.93	0.95	—	0.60	0.95	0.95	0.94	
Summer		mean	33.66	33.41	33.43	—	—	33.35	33.36	33.28	
		min	33.45	33.15	33.07	—	—	33.00	33.01	32.97	
		max	33.81	33.87	33.67	—	—	33.69	33.66	33.50	
		n	11,895	12,049	12,157	0	0	12,091	12,094	11,971	
		n_prop	0.90	0.91	0.92	—	—	0.91	0.91	0.90	
DO (mg/L)		Winter	mean	8.38	—	—	—	—	—	—	4.24
			min	8.05	—	—	—	—	—	—	2.20
			max	9.83	—	—	—	—	—	—	5.42
	n		10,159	—	—	id	—	—	—	8305	
	n_prop		0.78	—	—	0.36	—	—	—	0.64	
	Spring	mean	10.38	—	—	—	—	—	—	3.25	
		min	7.07	—	—	—	—	—	—	2.13	
		max	22.49	—	—	—	—	—	—	3.88	
		n	12,384	—	—	0	—	—	—	12,435	
		n_prop	0.95	—	—	—	—	—	—	0.95	

Appendix A.16 *continued*

Parameter	Season		1 m	10 m	20 m	30 m	45 m	60 m	75 m	89 m
pH (total pH)	<i>Summer</i>	mean	8.14	—	—	—	—	—	—	3.76
		min	7.17	—	—	—	—	—	—	3.01
		max	10.05	—	—	—	—	—	—	5.18
		n	12,050	—	—	0	—	—	—	12,104
		n_prop	0.91	—	—	—	—	—	—	0.91
	<i>Winter</i>	mean	8.07	—	—	—	—	—	—	7.75
		min	8.04	—	—	—	—	—	—	7.60
		max	8.15	—	—	—	—	—	—	7.84
		n	10,177	—	—	id	—	—	—	8300
		n_prop	0.79	—	—	0.36	—	—	—	0.64
	<i>Spring</i>	mean	8.21	—	—	—	—	—	—	7.70
		min	8.00	—	—	—	—	—	—	7.58
		max	8.88	—	—	—	—	—	—	7.78
		n	12,383	—	—	0	—	—	—	12,435
		n_prop	0.94	—	—	—	—	—	—	0.95
Chlorophyll a (µg/L)	<i>Summer</i>	mean	8.05	—	—	—	—	—	—	7.69
		min	7.97	—	—	—	—	—	—	7.58
		max	8.16	—	—	—	—	—	—	7.78
		n	12,042	—	—	0	—	—	—	12,104
		n_prop	0.91	—	—	—	—	—	—	0.91
	<i>Winter</i>	mean	—	—	—	0.76	—	—	—	0.07
		min	—	—	—	0.03	—	—	—	0.03
		max	—	—	—	2.23	—	—	—	0.22
		n	id	—	—	7210	—	—	—	9641
		n_prop	0.20	—	—	0.56	—	—	—	0.74
	<i>Spring</i>	mean	2.76	—	—	0.79	—	—	—	0.05
		min	0	—	—	0.05	—	—	—	0.03
		max	29.40	—	—	14.72	—	—	—	0.18
		n	8848	—	—	10,394	—	—	—	12,271
		n_prop	0.68	—	—	0.79	—	—	—	0.94
<i>Summer</i>	mean	—	—	—	—	—	—	—	0.07	
	min	—	—	—	—	—	—	—	0.03	
	max	—	—	—	—	—	—	—	0.33	
	n	0	—	—	0	—	—	—	11,946	
	n_prop	—	—	—	—	—	—	—	0.90	
CDOM (ppb)	<i>Winter</i>	mean	—	—	—	0.92	—	—	—	0.68
		min	—	—	—	0.64	—	—	—	0.36
		max	—	—	—	1.73	—	—	—	1.36
		n	id	—	—	7103	—	—	—	9641
		n_prop	0.19	—	—	0.55	—	—	—	0.74

Appendix A.16 *continued*

Parameter	Season		1 m	10 m	20 m	30 m	45 m	60 m	75 m	89 m
Turbidity (NTU)	<i>Spring</i>	mean	1.64	—	—	0.95	—	—	—	0.56
		min	0	—	—	0.45	—	—	—	0.36
		max	22.08	—	—	1.55	—	—	—	1.00
		n	8702	—	—	10,453	—	—	—	12,279
		n_prop	0.66	—	—	0.80	—	—	—	0.94
	<i>Summer</i>	mean	—	—	—	—	—	—	—	0.54
		min	—	—	—	—	—	—	—	0.36
		max	—	—	—	—	—	—	—	1.00
		n	0	—	—	0	—	—	—	11,949
		n_prop	—	—	—	—	—	—	—	0.90
	<i>Winter</i>	mean	—	—	—	0.11	—	—	—	0.25
		min	—	—	—	0.05	—	—	—	0.02
		max	—	—	—	0.60	—	—	—	1.70
		n	id	—	—	7195	—	—	—	9641
		n_prop	0.20	—	—	0.56	—	—	—	0.74
	<i>Spring</i>	mean	0.81	—	—	0.14	—	—	—	0.13
		min	0	—	—	0.02	—	—	—	0.01
		max	9.79	—	—	1.84	—	—	—	0.65
n		6515	—	—	10,435	—	—	—	12,274	
n_prop		0.50	—	—	0.80	—	—	—	0.94	
<i>Summer</i>	mean	—	—	—	—	—	—	—	0.22	
	min	—	—	—	—	—	—	—	0.05	
	max	—	—	—	—	—	—	—	1.02	
	n	0	—	—	0	—	—	—	8125	
	n_prop	—	—	—	—	—	—	—	0.61	
Nitrate + nitrite (µM)	<i>Winter</i>	mean	—	—	—	—	—	—	—	14.91
		min	—	—	—	—	—	—	—	0
		max	—	—	—	—	—	—	—	40.28
		n	0	—	—	—	—	—	—	1516
		n_prop	—	—	—	—	—	—	—	0.70
	<i>Spring</i>	mean	2.01	—	—	—	—	—	—	17.68
		min	0.74	—	—	—	—	—	—	0.07
		max	3.64	—	—	—	—	—	—	39.23
		n	949	—	—	—	—	—	—	896
		n_prop	0.43	—	—	—	—	—	—	0.41
<i>Summer</i>	mean	1.81	—	—	—	—	—	—	—	
	min	0	—	—	—	—	—	—	—	
	max	3.60	—	—	—	—	—	—	—	
	n	1941	—	—	—	—	—	—	0	
	n_prop	0.88	—	—	—	—	—	—	—	

Appendix A.16 *continued*

Parameter	Season		1 m	10 m	20 m	30 m	45 m	60 m	75 m	89 m
BOD (mg/L)	<i>Winter</i>	mean	—	—	—	0.12	—	—	—	—
		min	—	—	—	0.08	—	—	—	—
		max	—	—	—	0.60	—	—	—	—
		n	—	—	—	9318	—	—	—	id
		n_prop	—	—	—	0.72	—	—	—	0.07
	<i>Spring</i>	mean	—	—	—	0.14	—	—	—	—
		min	—	—	—	0.09	—	—	—	—
		max	—	—	—	0.43	—	—	—	—
		n	—	—	—	7537	—	—	—	id
		n_prop	—	—	—	0.58	—	—	—	0.30
	<i>Summer</i>	mean	—	—	—	—	—	—	—	0.04
		min	—	—	—	—	—	—	—	0
		max	—	—	—	—	—	—	—	0.35
		n	—	—	—	id	—	—	—	5546
		n_prop	—	—	—	0	—	—	—	0.42
xCO2 (ppm)	<i>Winter</i>	mean	370.49	—	—	—	—	—	—	—
		min	301.08	—	—	—	—	—	—	—
		max	390.05	—	—	—	—	—	—	—
		n	171	—	—	—	—	—	—	—
		n_prop	0.79	—	—	—	—	—	—	—
	<i>Spring</i>	mean	271.73	—	—	—	—	—	—	—
		min	59.20	—	—	—	—	—	—	—
		max	422.16	—	—	—	—	—	—	—
		n	212	—	—	—	—	—	—	—
		n_prop	0.97	—	—	—	—	—	—	—
	<i>Summer</i>	mean	384.65	—	—	—	—	—	—	—
		min	303.55	—	—	—	—	—	—	—
		max	476.94	—	—	—	—	—	—	—
		n	207	—	—	—	—	—	—	—
		n_prop	0.94	—	—	—	—	—	—	—

Appendix A.17

Summary of temperature, salinity, DO, pH (total), chlorophyll a, CDOM, turbidity, nitrate + nitrite, BOD, and xCO₂ recorded at various depths by the SBOO RTOMS in 2020. Data include mean, minimum, and maximum values, sample size (n), and proportion recovered (n_prop) for each depth by season. Sample sizes differed due to variations in sampling interval, deployment date, and data quality (Addenda 2-1 to 2-4); id=insufficient data (see text).

Parameter	Season		1 m	10 m	18 m	26 m	
Temperature (°C)	<i>Winter</i>	mean	15.52	15.23	14.84	14.18	
		min	13.61	11.55	10.99	10.81	
		max	17.35	16.80	16.30	16.14	
		n	12,731	12,787	12,801	12,757	
		n_prop	0.98	0.99	0.99	0.98	
	<i>Spring</i>	mean	17.77	13.66	12.50	11.69	
		min	13.70	10.91	10.60	10.56	
		max	22.70	19.71	16.91	15.81	
		n	11,286	11,442	8527	11,345	
		n_prop	0.86	0.87	0.65	0.87	
	<i>Summer</i>	mean	19.75	14.89	—	—	
		min	13.87	11.24	—	—	
		max	24.38	22.70	—	—	
		n	11,258	12,429	0	id	
		n_prop	0.85	0.94	—	0.30	
	<i>Fall</i>	mean	—	15.56	—	—	
		min	—	13.12	—	—	
		max	—	21.31	—	—	
		n	0	10,034	0	0	
		n_prop	—	0.76	—	—	
Salinity (psu)	<i>Winter</i>	mean	33.47	33.44	33.47	33.52	
		min	33.21	33.17	33.05	33.32	
		max	33.56	33.63	33.72	33.76	
		n	12,724	12,778	12,798	12,730	
		n_prop	0.98	0.99	0.99	0.98	
	<i>Spring</i>	mean	33.53	33.48	33.60	33.65	
		min	32.90	33.20	33.24	33.34	
		max	33.77	33.70	33.83	33.84	
		n	11,279	5534	8494	11,338	
		n_prop	0.86	0.42	0.65	0.87	
	<i>Summer</i>	mean	33.61	—	—	—	
		min	33.36	—	—	—	
		max	33.78	—	—	—	
		n	11,210	0	0	id	
		n_prop	0.85	—	—	0.28	
	<i>Fall</i>	n	0	0	0	0	
		<i>Winter</i>	mean	8.22	—	7.76	7.35
			min	6.97	—	4.90	4.86
			max	9.19	—	8.86	8.80
			n	12,650	—	12,811	12,770
n_prop	0.98		—	0.99	0.99		

Appendix A.17 *continued*

Parameter	Season		1 m	10 m	18 m	26 m
pH (total pH)	<i>Spring</i>	mean	9.84	—	5.85	5.03
		min	5.81	—	3.73	3.38
		max	23.31	—	10.21	10.13
		n	11,319	—	6519	11,380
		n_prop	0.86	—	0.50	0.87
	<i>Summer</i>	mean	8.47	—	—	—
		min	6.96	—	—	—
		max	10.95	—	—	—
		n	11,302	—	0	id
		n_prop	0.85	—	—	0.30
	<i>Fall</i>	n	0	—	0	0
	<i>Winter</i>	mean	7.88	—	—	7.79
		min	7.78	—	—	7.61
		max	7.96	—	—	7.89
		n	12,828	—	—	12,862
		n_prop	0.99	—	—	0.99
	<i>Spring</i>	mean	8.01	—	—	7.61
		min	7.69	—	—	7.44
		max	8.61	—	—	8.00
		n	11,365	—	—	11,429
n_prop		0.87	—	—	0.87	
<i>Summer</i>	mean	7.90	—	—	—	
	min	7.78	—	—	—	
	max	8.03	—	—	—	
	n	11,224	—	—	id	
	n_prop	0.85	—	—	0.30	
<i>Fall</i>	n	0	—	—	0	
Chlorophyll a (µg/L)	<i>Winter</i>	mean	0.73	—	1.11	1.16
		min	0	—	0.17	0.36
		max	3.33	—	4.68	4.75
		n	11,931	—	12,673	11,411
		n_prop	0.92	—	0.98	0.88
	<i>Spring</i>	mean	2.08	—	1.56	0.95
		min	0.02	—	0.12	0.10
		max	29.40	—	12.50	12.63
		n	11,183	—	11,252	9718
		n_prop	0.85	—	0.86	0.74
	<i>Summer</i>	mean	—	—	2.43	—
		min	—	—	0.25	—
		max	—	—	9.44	—
		n	id	—	12,327	id
		n_prop	0.36	—	0.93	0.31

Appendix A.17 *continued*

Parameter	Season		1 m	10 m	18 m	26 m	
CDOM (ppb)	<i>Fall</i>	mean	—	—	1.38	—	
		min	—	—	0.09	—	
		max	—	—	5.42	—	
		n	0	—	7393	0	
		n_prop	—	—	0.56	—	
	<i>Winter</i>	mean	0.87	—	1.01	1.13	
		min	0	—	0.54	0.72	
		max	3.16	—	2.53	2.62	
		n	11,874	—	12,574	11,411	
		n_prop	0.92	—	0.97	0.88	
	<i>Spring</i>	mean	0.95	—	1.15	1.09	
		min	0	—	0.72	0.72	
		max	18.35	—	2.89	2.98	
		n	11,184	—	11,251	9718	
		n_prop	0.85	—	0.86	0.74	
<i>Summer</i>	mean	—	—	1.26	—		
	min	—	—	0.81	—		
	max	—	—	2.89	—		
	n	id	—	11,157	id		
	n_prop	0.10	—	0.84	0.31		
Turbidity (NTU)	<i>Fall</i>	n	0	—	0	0	
		<i>Winter</i>	mean	0.21	—	0.19	0.36
			min	0	—	0.06	0.08
			max	3.04	—	1.76	4.46
			n	12,000	—	12,673	11,410
	n_prop		0.93	—	0.98	0.88	
	<i>Spring</i>	mean	0.58	—	0.22	0.22	
		min	0.04	—	0.05	0.05	
		max	9.80	—	1.48	1.36	
		n	5609	—	11,252	9717	
		n_prop	0.43	—	0.86	0.74	
	<i>Summer</i>	mean	—	—	0.30	—	
		min	—	—	0.05	—	
		max	—	—	2.57	—	
		n	0	—	11,528	id	
n_prop		—	—	0.87	0.31		
Nitrate + nitrite (µM)	<i>Fall</i>	n	0	—	0	0	
		<i>Winter</i>	mean	2.43	—	—	4.78
	min		0.17	—	—	0.01	
	max		11.20	—	—	20.77	
	n		2059	—	—	2036	
	n_prop	0.95	—	—	0.94		

Appendix A.17 *continued*

Parameter	Season		1 m	10 m	18 m	26 m
BOD (mg/L)	<i>Spring</i>	mean	1.41	—	—	12.23
		min	0	—	—	0.01
		max	5.87	—	—	25.41
		n	1793	—	—	1773
		n_prop	0.82	—	—	0.81
	<i>Summer</i>	mean	1.22	—	—	—
		min	0	—	—	—
		max	3.65	—	—	—
		n	1949	—	—	id
		n_prop	0.88	—	—	0.29
	<i>Fall</i>	n	id	—	—	0
		n_prop	0	—	—	—
	<i>Winter</i>	mean	—	—	—	0.10
		min	—	—	—	0.04
		max	—	—	—	9.27
n		—	—	—	12,769	
n_prop		—	—	—	0.99	
<i>Spring</i>	mean	—	—	—	0.08	
	min	—	—	—	0.04	
	max	—	—	—	1.67	
	n	—	—	—	11,350	
	n_prop	—	—	—	0.87	
<i>Summer</i>	n	—	—	—	id	
	n_prop	—	—	—	0.30	
<i>Fall</i>	n	—	—	—	id	
	n_prop	—	—	—	0.01	
xCO2 (ppm)	<i>Winter</i>	mean	385.18	—	—	—
		min	317.81	—	—	—
		max	478.34	—	—	—
		n	214	—	—	—
		n_prop	0.99	—	—	—
	<i>Spring</i>	mean	296.49	—	—	—
		min	60.71	—	—	—
		max	564.50	—	—	—
		n	190	—	—	—
		n_prop	0.87	—	—	—
	<i>Summer</i>	mean	359.64	—	—	—
		min	251.88	—	—	—
		max	462.21	—	—	—
		n	217	—	—	—
		n_prop	0.98	—	—	—

Appendix A.17 *continued*

Parameter	Season		1 m	10 m	18 m	26 m
	<i>Fall</i>	mean	398.04	—	—	—
		min	313.57	—	—	—
		max	530.59	—	—	—
		n	169	—	—	—
		n_prop	0.76	—	—	—

Appendix A.18

Summary of current velocity magnitude and direction from the PLOO RTOMS ADCP during 2020. The deployment ended prior to the Fall. Data are presented by depth bin as seasonal recovered observations (n), minimum (min), maximum (max), and means with 95% confidence intervals (CI). Proportion of recovered observations (n_prop) differed due to variations in data quality (Addenda 2-1 to 2-4). Minimum and maximum angles of velocity are not shown due to the circular nature of the measurement.

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
<i>Winter</i>	4	10,168	1.00	0	574	137	2	544	75
	6	10,168	1.00	0	660	159	2	542	74
	8	10,168	1.00	7	498	137	2	550	72
	10	10,168	1.00	1	515	137	2	546	73
	12	10,168	1.00	5	611	151	2	177	75
	14	10,168	1.00	4	590	148	2	175	75
	16	10,168	1.00	14	538	143	2	175	75
	18	10,167	1.00	9	489	130	2	179	74
	20	10,166	1.00	13	510	136	2	174	75
	22	10,166	1.00	14	503	134	2	172	76
	24	10,167	1.00	10	494	130	2	171	75
	26	10,166	1.00	11	484	123	2	171	75
	28	10,166	1.00	6	473	117	2	170	74
	30	10,167	1.00	9	471	115	2	168	74
	32	10,165	1.00	10	462	112	2	168	74
	34	10,165	1.00	4	459	109	2	168	73
	36	10,165	1.00	1	460	105	2	168	73
	38	10,165	1.00	2	458	99	2	168	73
	40	10,165	1.00	4	447	93	1	168	72
	42	10,165	1.00	0	435	89	1	167	72
	44	10,164	1.00	6	421	85	1	165	71
	46	10,164	1.00	3	397	79	1	163	71
	48	10,162	1.00	7	379	75	1	160	70
	50	10,148	1.00	3	346	70	1	156	70
	52	10,128	1.00	0	298	64	1	152	69
	54	10,090	0.99	6	233	59	1	142	69
	56	10,012	0.98	1	229	57	1	128	69
	58	9794	0.96	1	210	56	1	110	69
	60	9440	0.93	0	186	56	1	83	69
	62	8964	0.88	1	170	56	1	64	69
	64	8348	0.82	0	156	55	1	61	69
	66	7677	0.76	0	131	54	1	58	69
	68	6840	0.67	2	124	53	1	60	69
	70	5982	0.59	2	126	54	1	64	70
72	5144	0.51	5	127	53	1	66	71	
74	4407	0.43	0	141	54	1	69	71	

Appendix A.18 *continued*

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
	76	3765	0.37	—	—	—	—	—	—
	78	3251	0.32	—	—	—	—	—	—
	80	2861	0.28	—	—	—	—	—	—
	82	2631	0.26	—	—	—	—	—	—
	84	2386	0.23	—	—	—	—	—	—
	86	2283	0.22	—	—	—	—	—	—
	88	2276	0.22	—	—	—	—	—	—
	90	2337	0.23	—	—	—	—	—	—
	92	2499	0.25	—	—	—	—	—	—
<i>Spring</i>	4	12,413	1.00	2	541	250	2	173	69
	6	12,412	1.00	2	550	270	2	170	69
	8	12,412	1.00	2	498	232	2	173	67
	10	12,411	1.00	8	509	233	2	171	68
	12	12,405	1.00	10	519	260	2	166	69
	14	12,397	1.00	7	501	250	2	165	70
	16	12,396	1.00	6	485	236	2	164	70
	18	12,403	1.00	7	477	207	2	165	70
	20	12,399	1.00	10	475	208	2	162	70
	22	12,396	1.00	10	474	202	2	161	71
	24	12,390	1.00	11	476	191	2	159	71
	26	12,368	1.00	9	474	175	2	157	72
	28	12,349	0.99	6	459	160	2	155	72
	30	12,315	0.99	13	458	158	2	154	73
	32	12,261	0.99	9	450	151	1	151	73
	34	12,204	0.98	5	449	143	1	149	73
	36	12,113	0.98	2	449	135	1	145	73
	38	12,035	0.97	3	445	127	1	141	73
	40	11,919	0.96	3	437	112	1	135	73
	42	11,831	0.95	8	422	108	1	130	72
	44	11,587	0.93	10	407	106	1	124	72
	46	11,365	0.92	3	382	99	1	118	72
	48	11,008	0.89	4	363	94	1	107	72
	50	10,477	0.84	1	326	89	1	92	72
	52	9657	0.78	6	275	83	1	73	72
	54	8669	0.70	12	196	74	1	49	70
	56	7423	0.60	13	172	74	1	44	69
	58	5895	0.47	12	165	72	1	43	67
	60	4457	0.36	—	—	—	—	—	—
	62	3270	0.26	—	—	—	—	—	—
	64	2317	0.19	—	—	—	—	—	—

Appendix A.18 *continued*

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
	66	1590	0.13	—	—	—	—	—	—
	68	995	0.08	—	—	—	—	—	—
	70	617	0.05	—	—	—	—	—	—
	72	354	0.03	—	—	—	—	—	—
	74	210	0.02	—	—	—	—	—	—
	76	146	0.01	—	—	—	—	—	—
	78	117	0.01	—	—	—	—	—	—
	80	94	0.01	—	—	—	—	—	—
	82	100	0.01	—	—	—	—	—	—
	84	110	0.01	—	—	—	—	—	—
	86	110	0.01	—	—	—	—	—	—
	88	115	0.01	—	—	—	—	—	—
	90	149	0.01	—	—	—	—	—	—
	92	175	0.01	—	—	—	—	—	—
<i>Summer</i>	4	12,165	1.00	1	431	171	2	173	60
	6	12,162	1.00	4	440	173	2	167	62
	8	12,160	1.00	7	345	141	1	169	64
	10	12,162	1.00	5	320	133	1	165	65
	12	12,161	1.00	2	379	138	1	159	66
	14	12,161	1.00	6	373	128	1	158	66
	16	12,162	1.00	0	360	120	1	157	67
	18	12,162	1.00	1	312	107	1	153	67
	20	12,160	1.00	6	326	108	1	147	68
	22	12,158	1.00	6	313	106	1	137	68
	24	12,157	1.00	2	287	102	1	124	68
	26	12,156	1.00	3	240	96	1	95	67
	28	12,158	1.00	3	226	92	1	63	68
	30	12,155	1.00	0	220	90	1	59	68
	32	12,150	1.00	1	206	87	1	46	68
	34	12,143	1.00	2	206	85	1	37	69
	36	12,136	1.00	3	206	84	1	32	68
	38	12,110	1.00	4	205	82	1	27	69
	40	12,090	0.99	1	187	81	1	20	69
	42	12,108	1.00	3	200	82	1	20	69
	44	11,990	0.99	3	208	83	1	18	69
	46	11,847	0.97	0	209	82	1	14	69
	48	11,693	0.96	2	200	82	1	11	68
	50	11,465	0.94	3	199	81	1	10	68
	52	11,012	0.91	0	189	81	1	8	67
	54	10,382	0.85	4	193	79	1	5	67
	56	9678	0.80	1	179	79	1	6	67

Appendix A.18 *continued*

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
	58	8536	0.70	0	173	79	1	7	67
	60	7117	0.58	1	172	76	1	8	68
	62	5758	0.47	3	166	73	1	8	67
	64	4266	0.35	—	—	—	—	—	—
	66	3110	0.26	—	—	—	—	—	—
	68	1949	0.16	—	—	—	—	—	—
	70	1221	0.10	—	—	—	—	—	—
	72	714	0.06	—	—	—	—	—	—
	74	469	0.04	—	—	—	—	—	—
	76	314	0.03	—	—	—	—	—	—
	78	249	0.02	—	—	—	—	—	—
	80	204	0.02	—	—	—	—	—	—
	82	163	0.01	—	—	—	—	—	—
	84	157	0.01	—	—	—	—	—	—
	86	150	0.01	—	—	—	—	—	—
	88	156	0.01	—	—	—	—	—	—
	90	161	0.01	—	—	—	—	—	—
	92	163	0.01	—	—	—	—	—	—

Appendix A.19

Summary of current velocity magnitude and direction from the SBOO RTOMS ADCP during 2020. Data are presented by depth bin as seasonal recovered observations (n), minimum (min), maximum (max), and means with 95% confidence intervals (CI). Proportion of recovered observations (n_prop) differed due to variations in data quality (Addenda 2-1 to 2-4). Minimum and maximum angles of velocity are not shown due to the circular nature of the measurement.

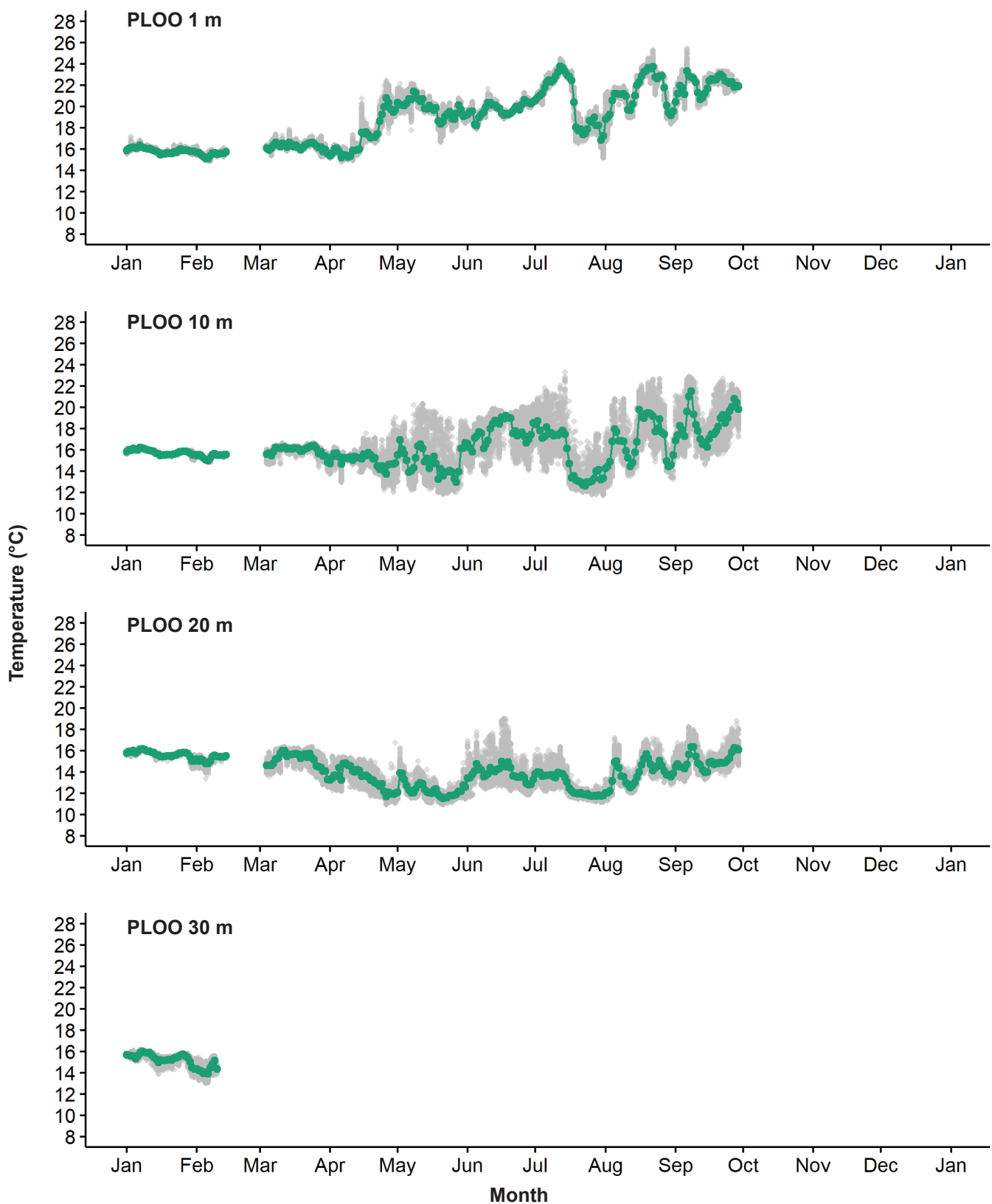
Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle		
				Min	Max	Mean	95% CI	Mean	95% CI	
<i>Winter</i>	3	12,861	1.00	4	271	99	1	180	72	
	4	12,860	1.00	3	330	115	1	174	72	
	5	12,860	1.00	3	331	117	1	173	73	
	6	12,860	1.00	10	317	117	1	172	73	
	7	12,860	1.00	10	307	116	1	170	73	
	8	12,860	1.00	2	302	114	1	170	73	
	9	12,860	1.00	2	149	70	1	561	60	
	10	12,860	1.00	1	242	93	1	174	69	
	11	12,859	1.00	4	292	108	1	164	71	
	12	12,859	1.00	6	277	107	1	162	70	
	13	12,859	1.00	4	266	106	1	160	70	
	14	12,859	1.00	7	258	104	1	158	69	
	15	12,859	1.00	1	231	99	1	159	68	
	16	12,858	1.00	3	184	83	1	166	65	
	17	12,857	1.00	2	172	78	1	172	62	
	18	12,856	1.00	1	213	93	1	152	69	
	19	12,856	1.00	4	207	91	1	152	69	
	20	12,855	1.00	5	195	89	1	149	70	
	21	12,856	1.00	0	188	86	1	148	70	
	22	12,856	1.00	9	151	71	1	152	71	
	23	12,856	1.00	1	115	55	0	173	66	
	24	12,856	1.00	0	119	50	0	576	64	
	25	12,856	1.00	0	160	54	1	170	69	
	26	12,857	1.00	4	153	55	0	147	68	
	27	12,857	1.00	1	135	47	0	145	66	
	28	12,856	1.00	0	125	42	0	156	62	
	<i>Spring</i>	3	11,406	1.00	3	330	125	1	163	75
		4	11,406	1.00	4	352	147	2	163	72
5		11,405	1.00	8	348	144	2	159	72	
6		11,402	1.00	4	351	138	2	155	72	
7		11,404	1.00	12	344	130	1	153	71	
8		11,404	1.00	2	326	124	1	147	72	
9		11,394	1.00	3	197	88	1	589	71	
10		11,395	1.00	4	299	106	1	145	71	
11		11,396	1.00	8	324	110	1	122	70	
12		11,396	1.00	4	318	106	1	104	71	

Appendix A.19 *continued*

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
	13	11,396	1.00	1	310	104	1	88	70
	14	11,396	1.00	7	307	101	1	70	71
	15	11,395	1.00	6	271	94	1	49	72
	16	11,394	1.00	7	210	82	1	10	72
	17	11,394	1.00	1	243	81	1	357	71
	18	11,394	1.00	2	296	92	1	44	71
	19	11,394	1.00	2	283	90	1	47	71
	20	11,394	1.00	1	281	90	1	46	71
	21	11,394	1.00	1	253	88	1	45	72
	22	11,393	1.00	2	202	76	1	28	72
	23	11,393	1.00	2	183	65	1	353	70
	24	11,393	1.00	1	170	64	1	335	69
	25	11,393	1.00	0	192	66	1	2	70
	26	11,392	1.00	9	180	66	1	26	69
	27	11,393	1.00	3	145	56	1	15	68
	28	11,396	1.00	3	145	48	1	5	65
<i>Summer</i>	3	12,669	1.00	6	318	89	1	166	71
	4	12,668	1.00	7	425	108	1	162	71
	5	12,668	1.00	1	450	109	2	156	70
	6	12,668	1.00	3	435	107	1	150	70
	7	12,668	1.00	5	420	103	1	145	71
	8	12,668	1.00	5	399	98	1	140	71
	9	12,668	1.00	4	264	74	1	152	71
	10	12,668	1.00	4	296	83	1	131	71
	11	12,668	1.00	1	371	88	1	128	71
	12	12,667	1.00	2	366	87	1	121	71
	13	12,667	1.00	1	356	84	1	113	71
	14	12,667	1.00	2	339	83	1	95	71
	15	12,667	1.00	0	306	81	1	78	72
	16	12,667	1.00	0	221	74	1	43	72
	17	12,666	1.00	3	241	71	1	34	72
	18	12,666	1.00	0	270	77	1	64	71
	19	12,666	1.00	3	270	77	1	63	71
	20	12,666	1.00	3	259	76	1	59	71
	21	12,666	1.00	0	237	73	1	56	71
	22	12,666	1.00	10	166	65	1	39	70
	23	12,665	1.00	1	156	55	1	19	69
	24	12,665	1.00	2	142	51	1	4	68
	25	12,665	1.00	5	139	53	1	23	67
	26	12,665	1.00	5	132	52	0	30	66
	27	12,664	1.00	6	124	47	0	21	66

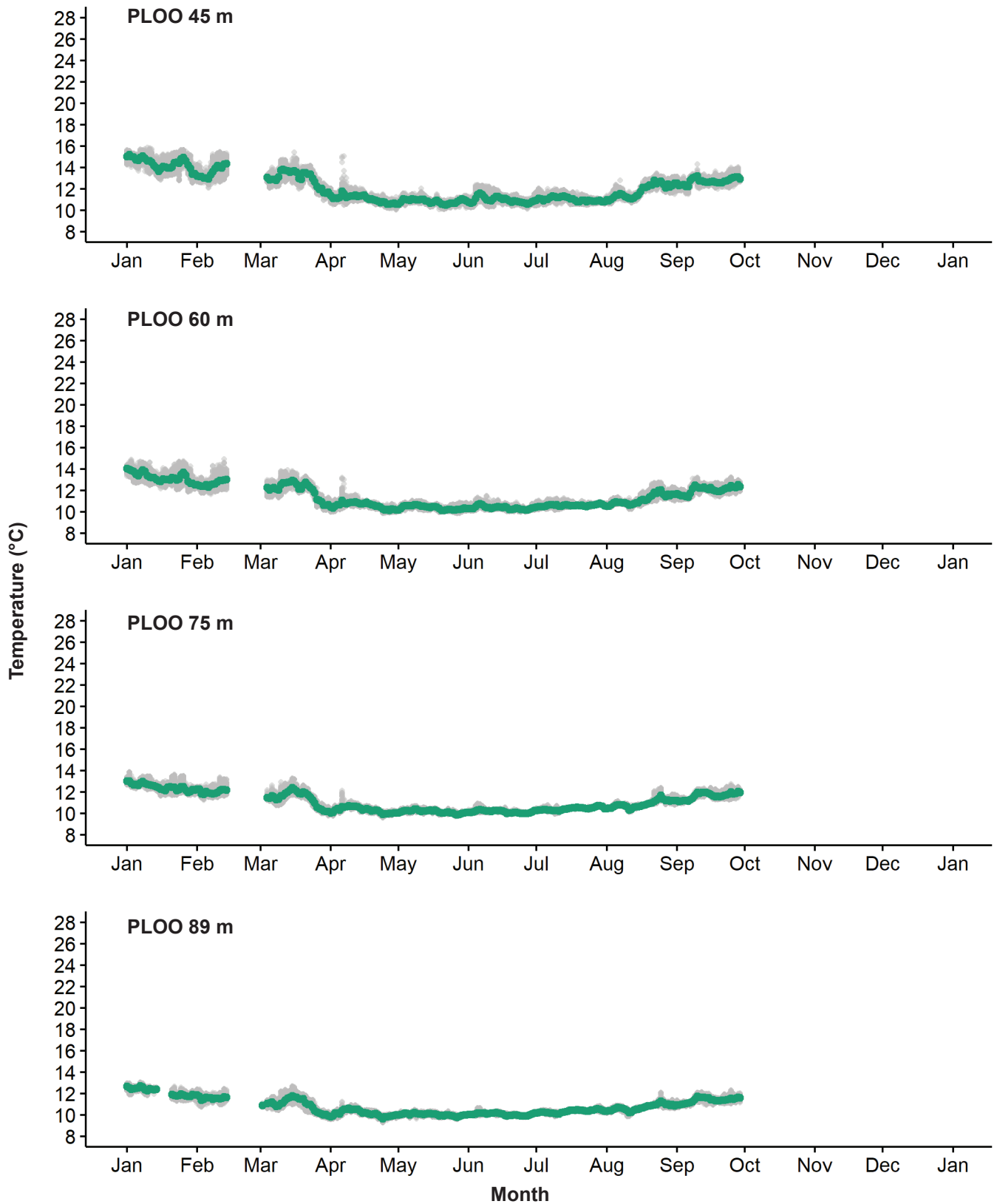
Appendix A.19 *continued*

Season	Depth (m)	n	n_prop	Magnitude (mm/s)				Angle	
				Min	Max	Mean	95% CI	Mean	95% CI
<i>Fall</i>	28	12,664	1.00	0	103	41	0	11	61
	3	10,022	1.00	0	225	51	1	166	46
	4	10,021	1.00	2	239	68	1	153	35
	5	10,021	1.00	4	246	68	1	157	40
	6	10,021	1.00	7	244	65	1	162	46
	7	10,021	1.00	2	246	61	1	167	50
	8	10,021	1.00	0	232	57	1	170	52
	9	10,021	1.00	0	178	47	1	546	53
	10	10,021	1.00	2	205	48	1	545	54
	11	10,021	1.00	2	226	48	1	179	54
	12	10,022	1.00	1	213	48	1	542	56
	13	10,021	1.00	0	214	49	1	549	57
	14	10,020	1.00	2	216	49	1	134	57
	15	10,021	1.00	1	192	49	1	33	57
	16	10,022	1.00	0	142	47	1	8	57
	17	10,021	1.00	1	140	47	1	10	56
	18	10,021	1.00	1	169	51	1	23	56
	19	10,021	1.00	1	164	53	1	19	55
	20	10,021	1.00	0	169	53	1	18	57
	21	10,020	1.00	0	129	50	1	18	57
	22	10,020	1.00	3	109	48	0	13	55
	23	10,019	1.00	3	104	43	0	1	54
	24	10,019	1.00	1	107	40	0	349	56
	25	10,019	1.00	1	96	37	0	0	56
	26	10,020	1.00	1	90	38	0	9	54
	27	10,020	1.00	0	84	35	0	11	55
	28	10,020	1.00	0	81	33	0	353	50

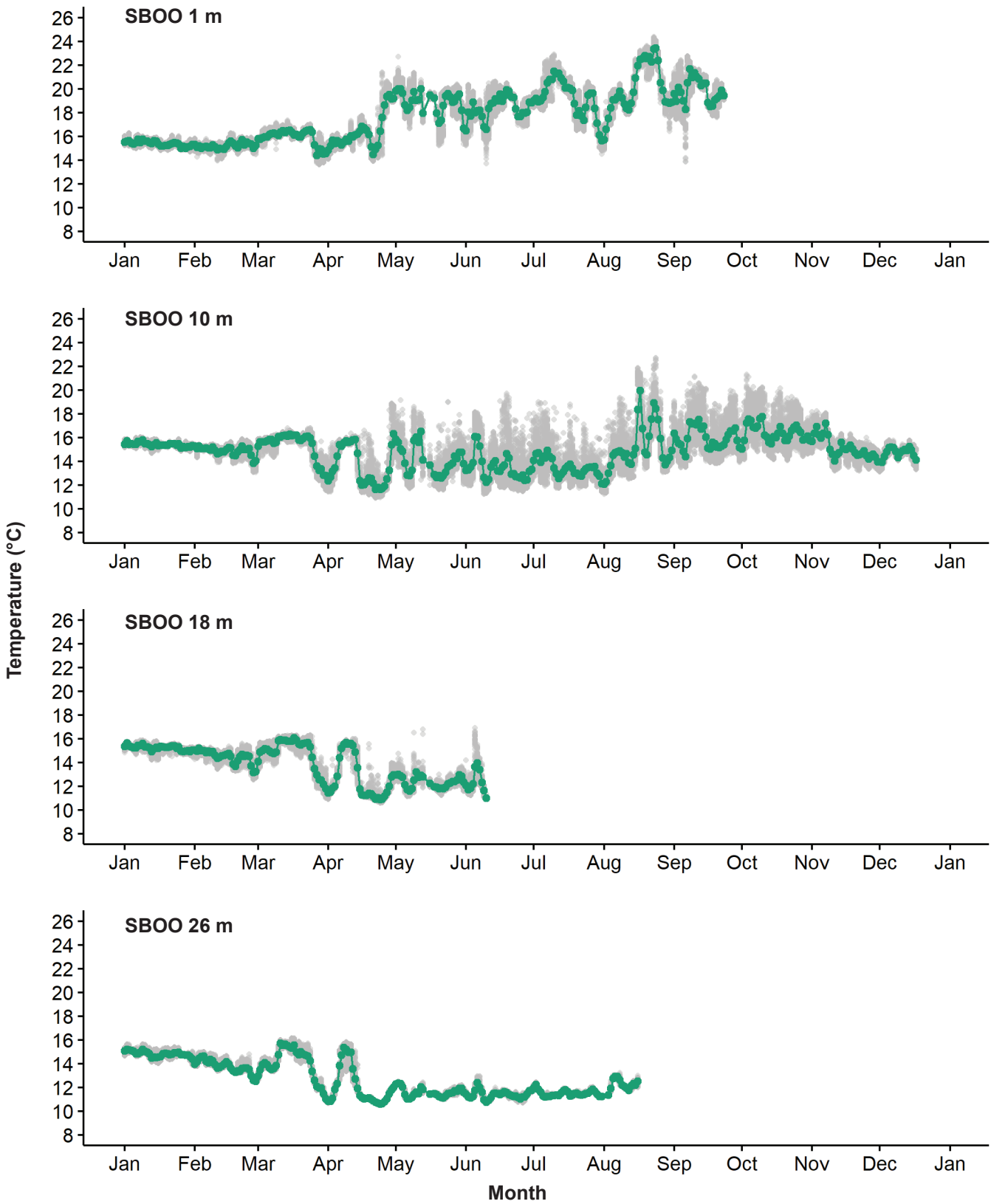


Appendix A.20

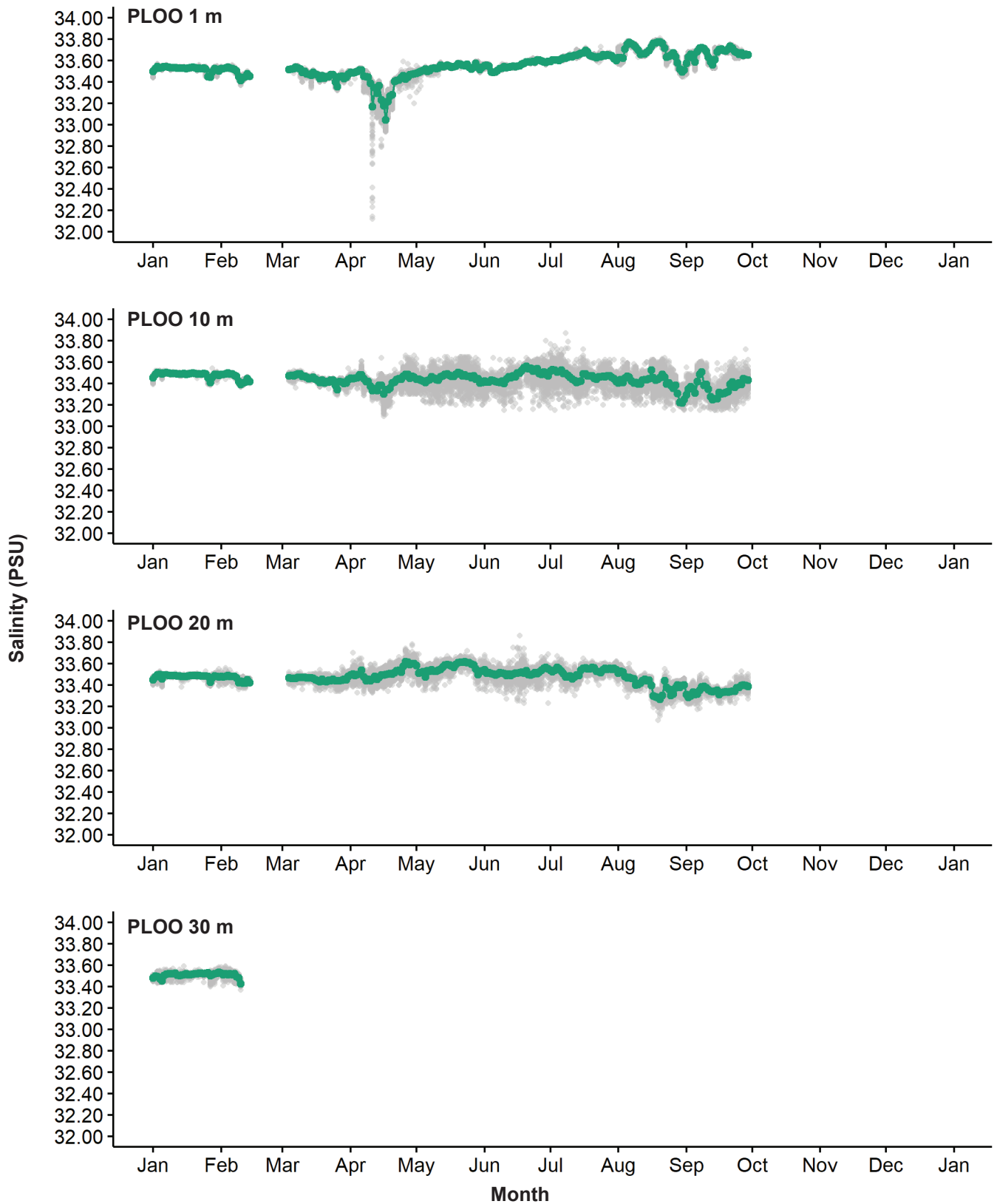
Temperature, salinity, DO, pH (total), chlorophyll a, CDOM, turbidity, nitrate + nitrite, BOD, and xCO₂ recorded at various depths by the PLOO and SBOO RTOMS during 2020. Grey points represent raw data and green points represent daily averaged data.



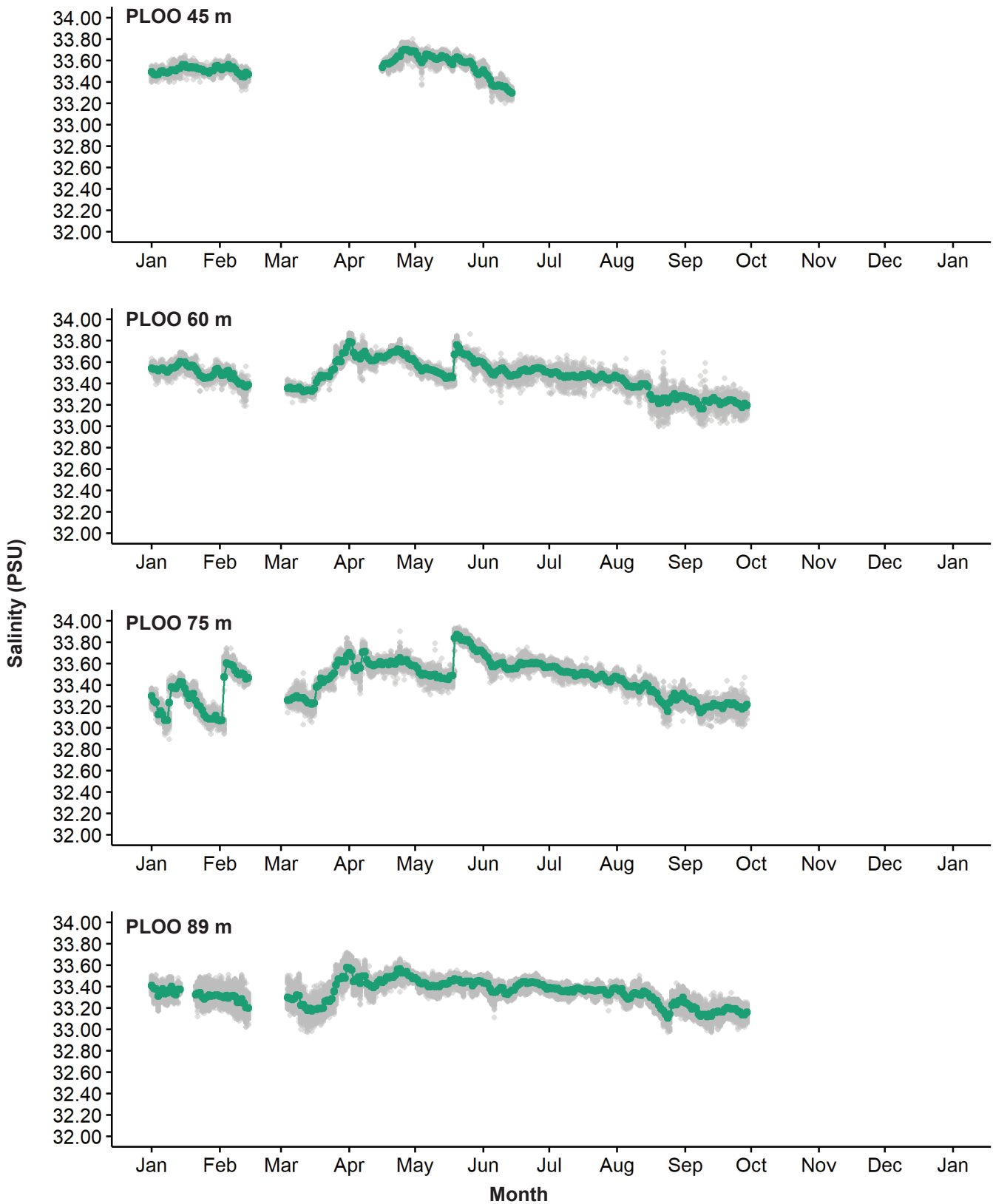
Appendix A.20 *continued*



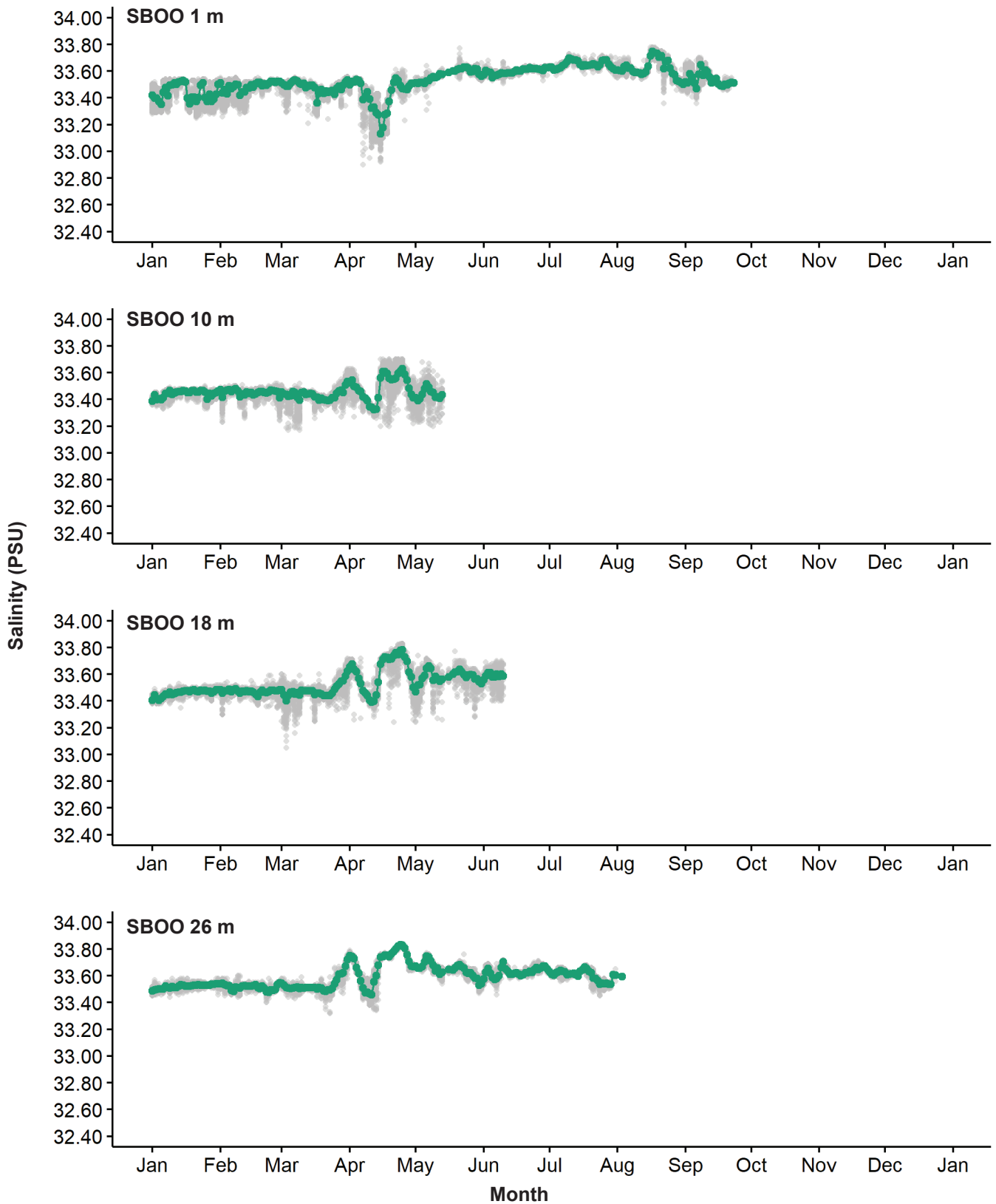
Appendix A.20 *continued*



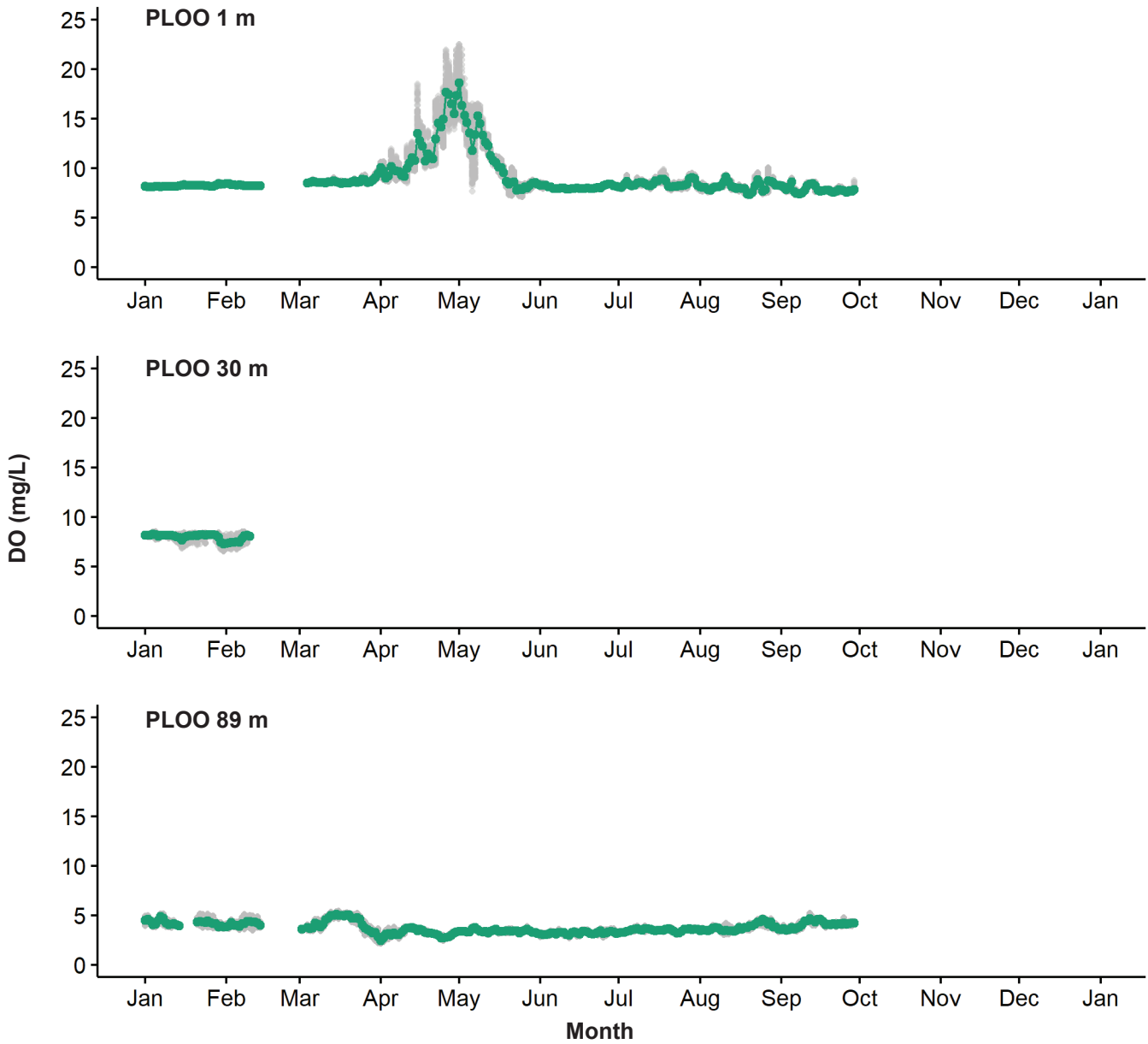
Appendix A.20 *continued*



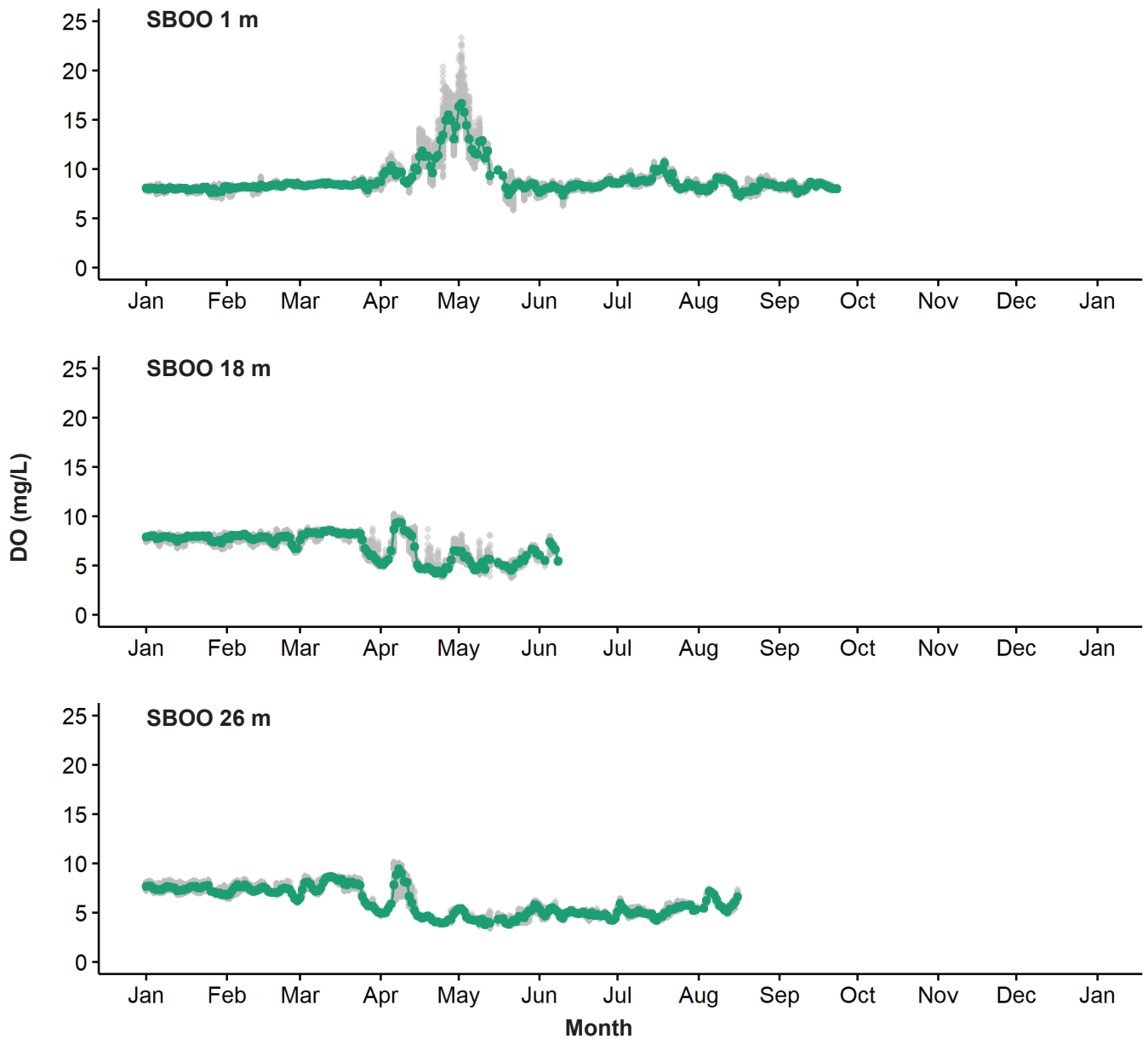
Appendix A.20 *continued*



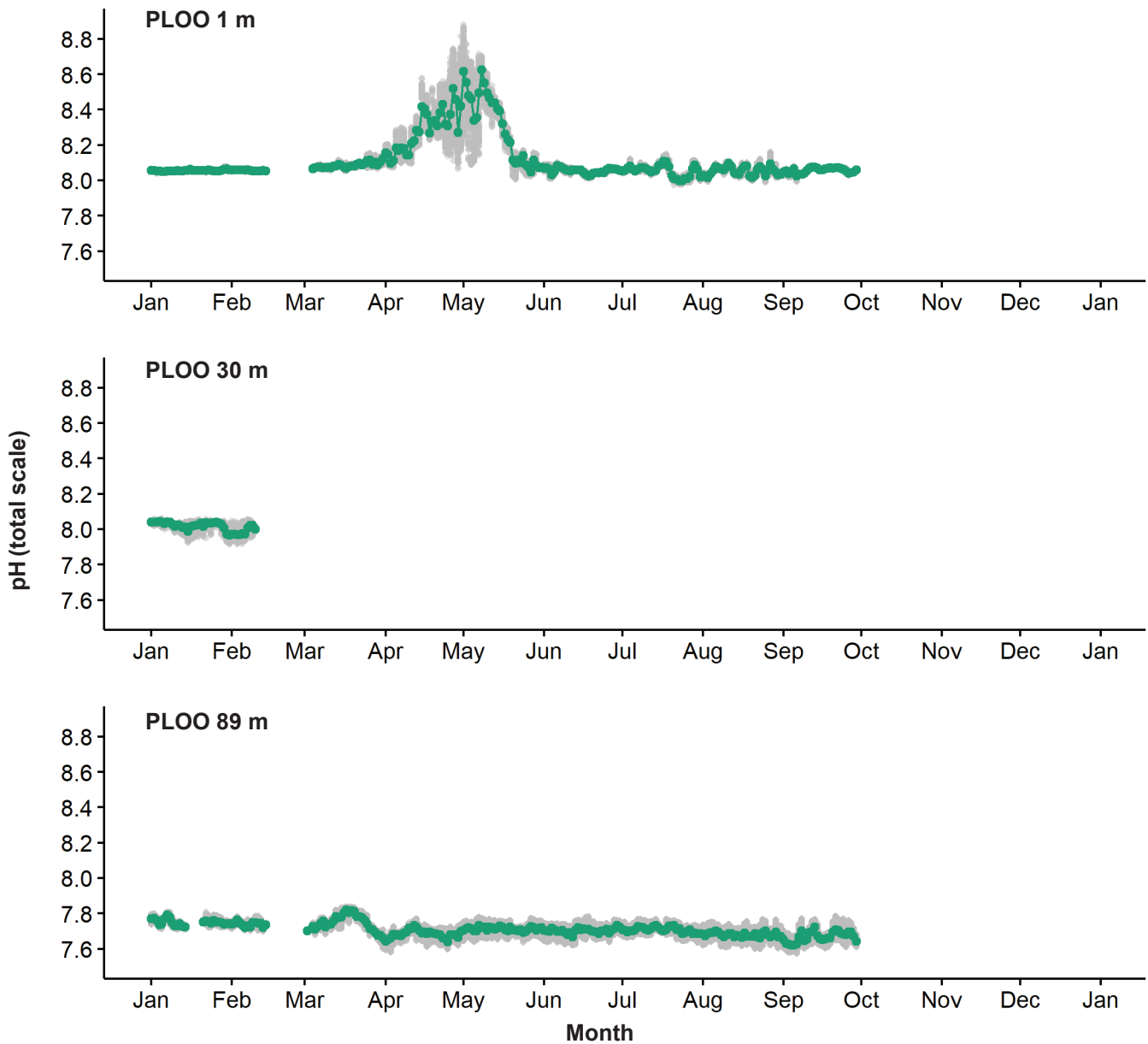
Appendix A.20 *continued*



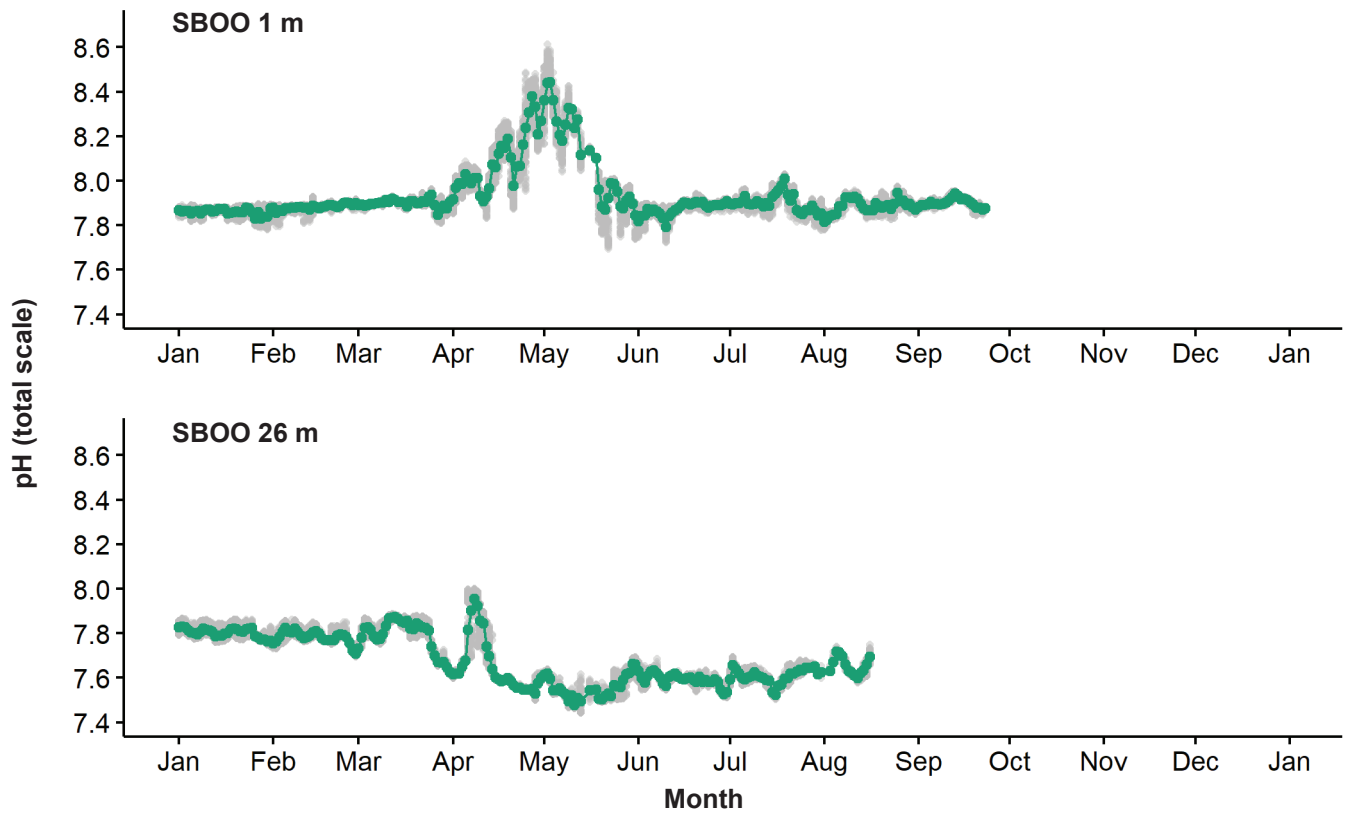
Appendix A.20 *continued*



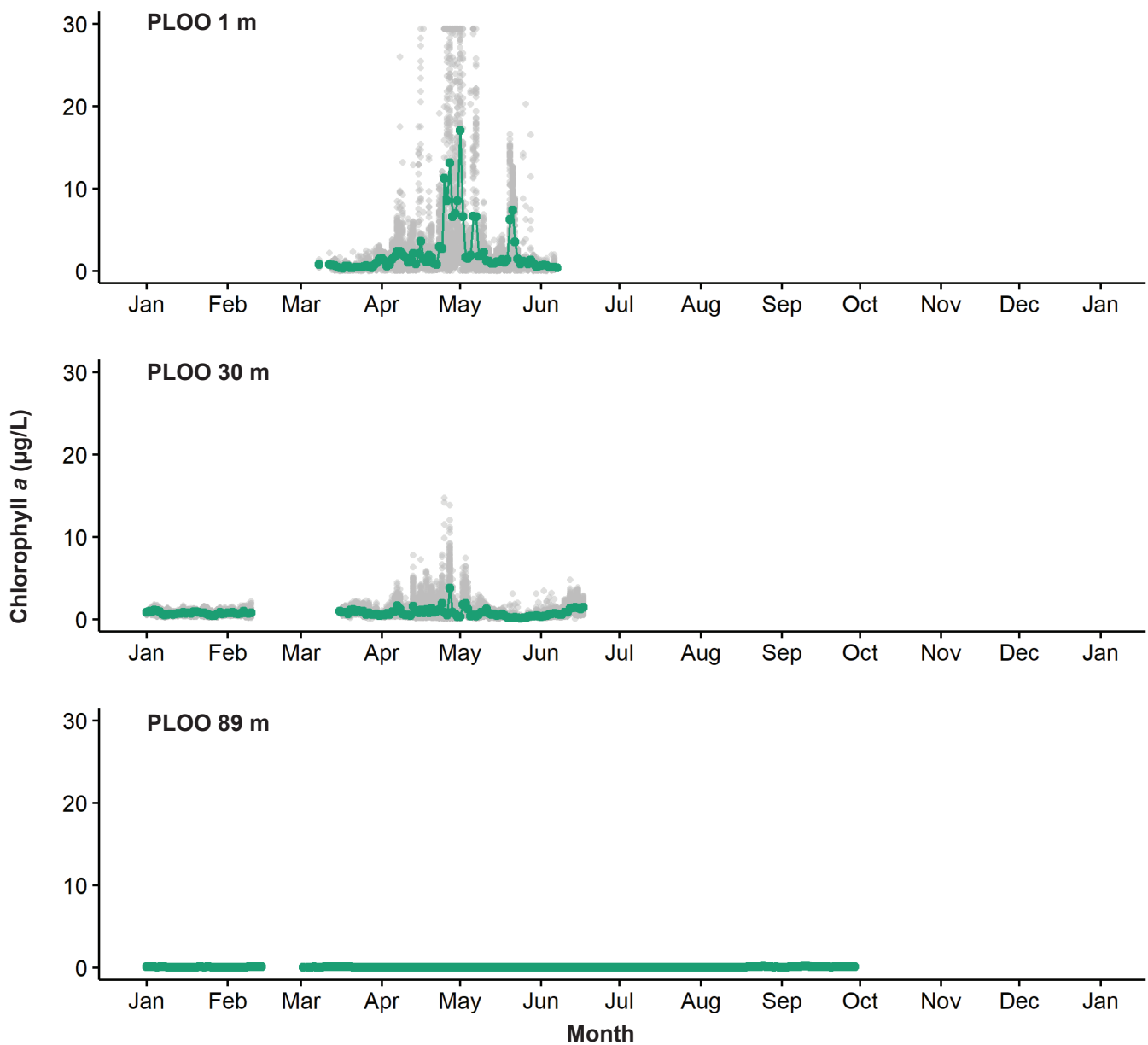
Appendix A.20 *continued*



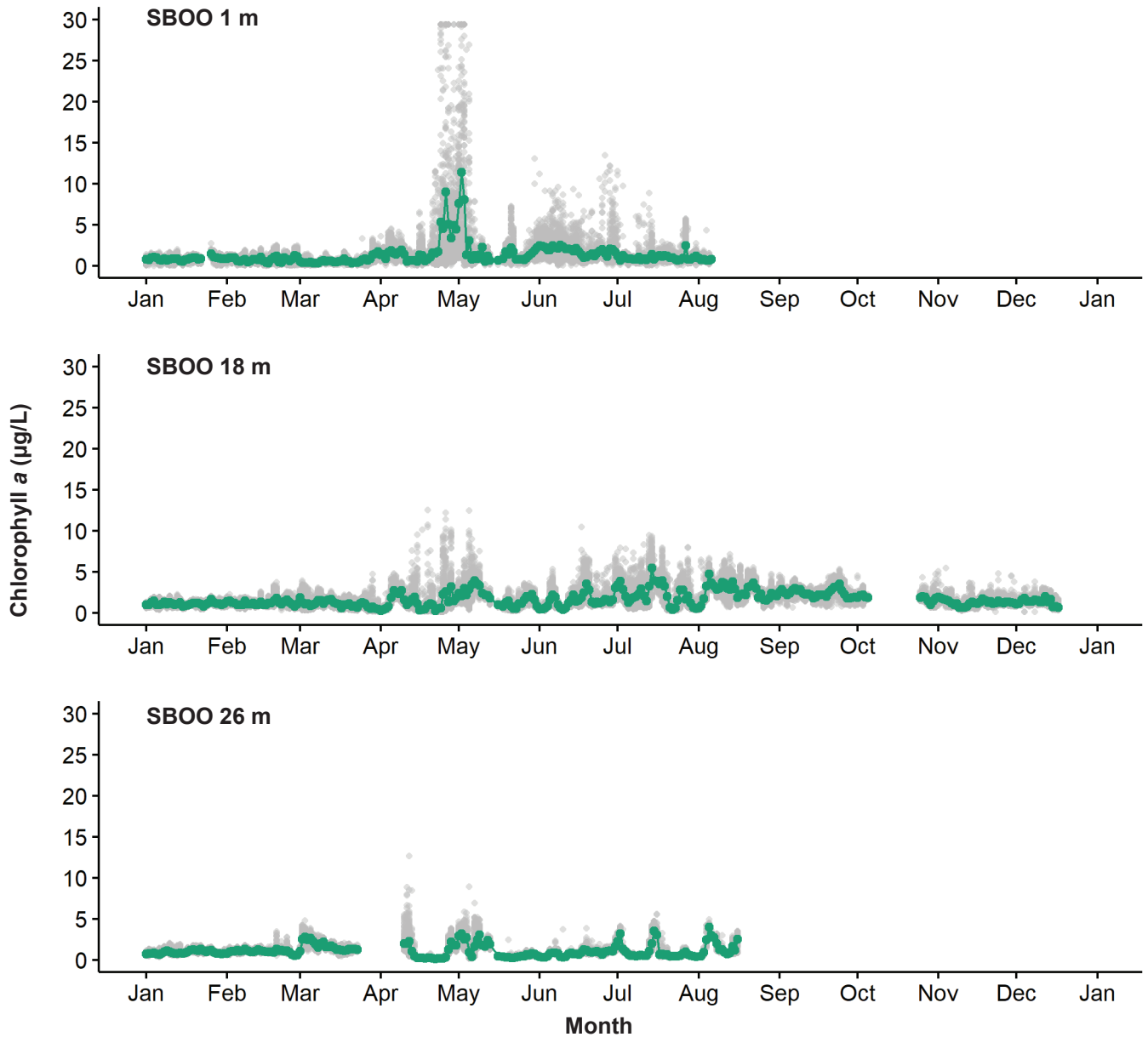
Appendix A.20 *continued*



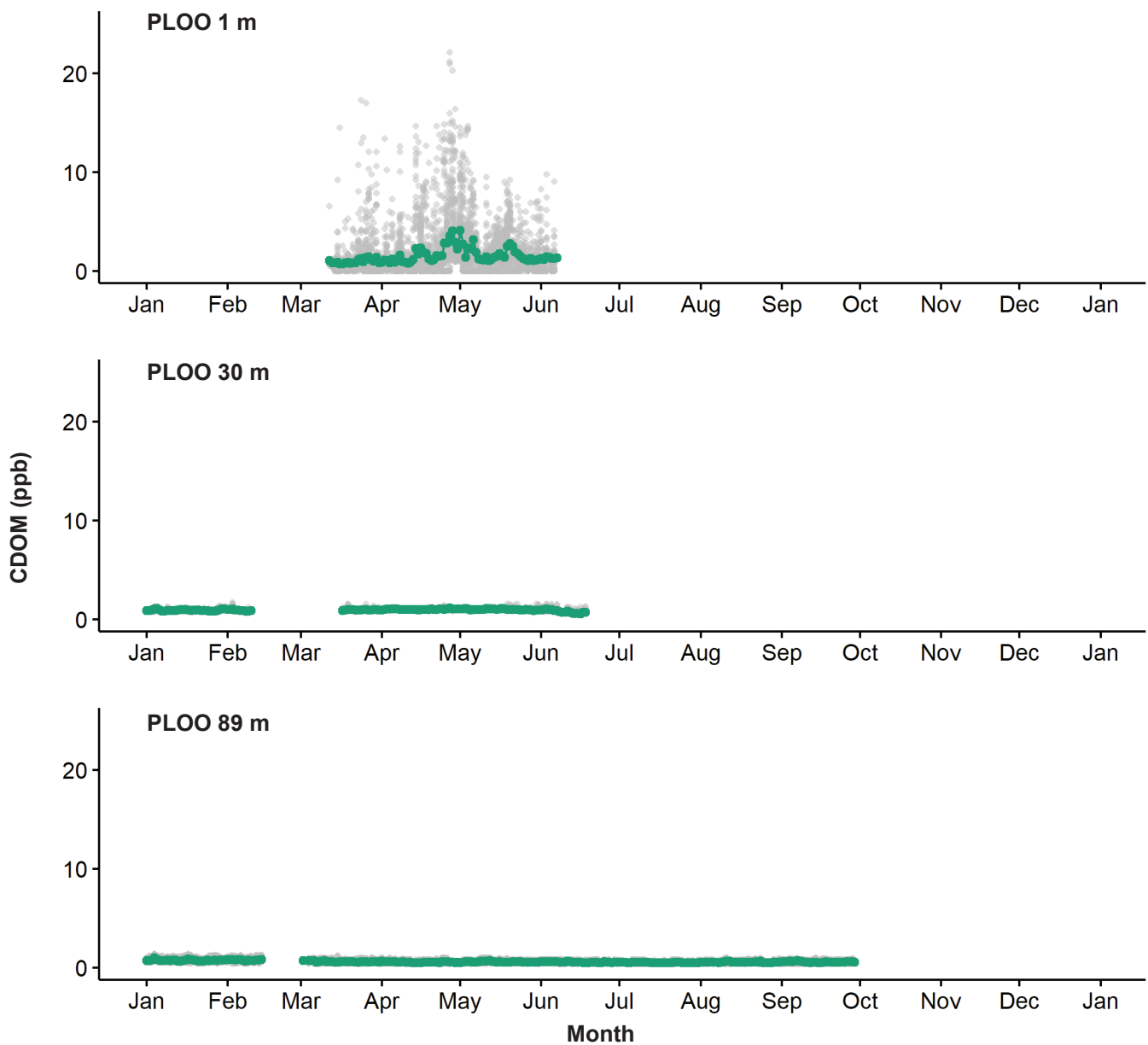
Appendix A.20 *continued*



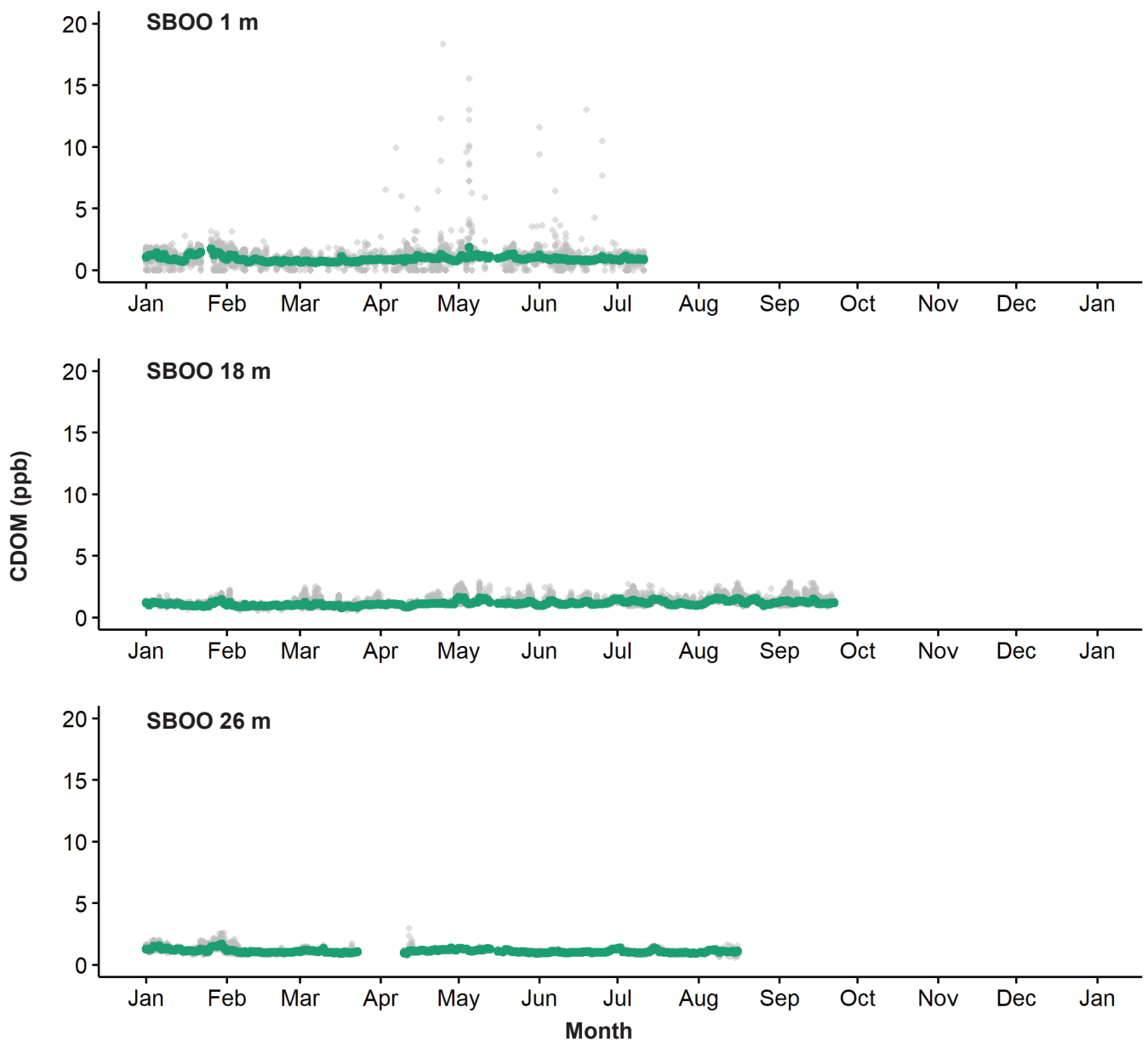
Appendix A.20 *continued*



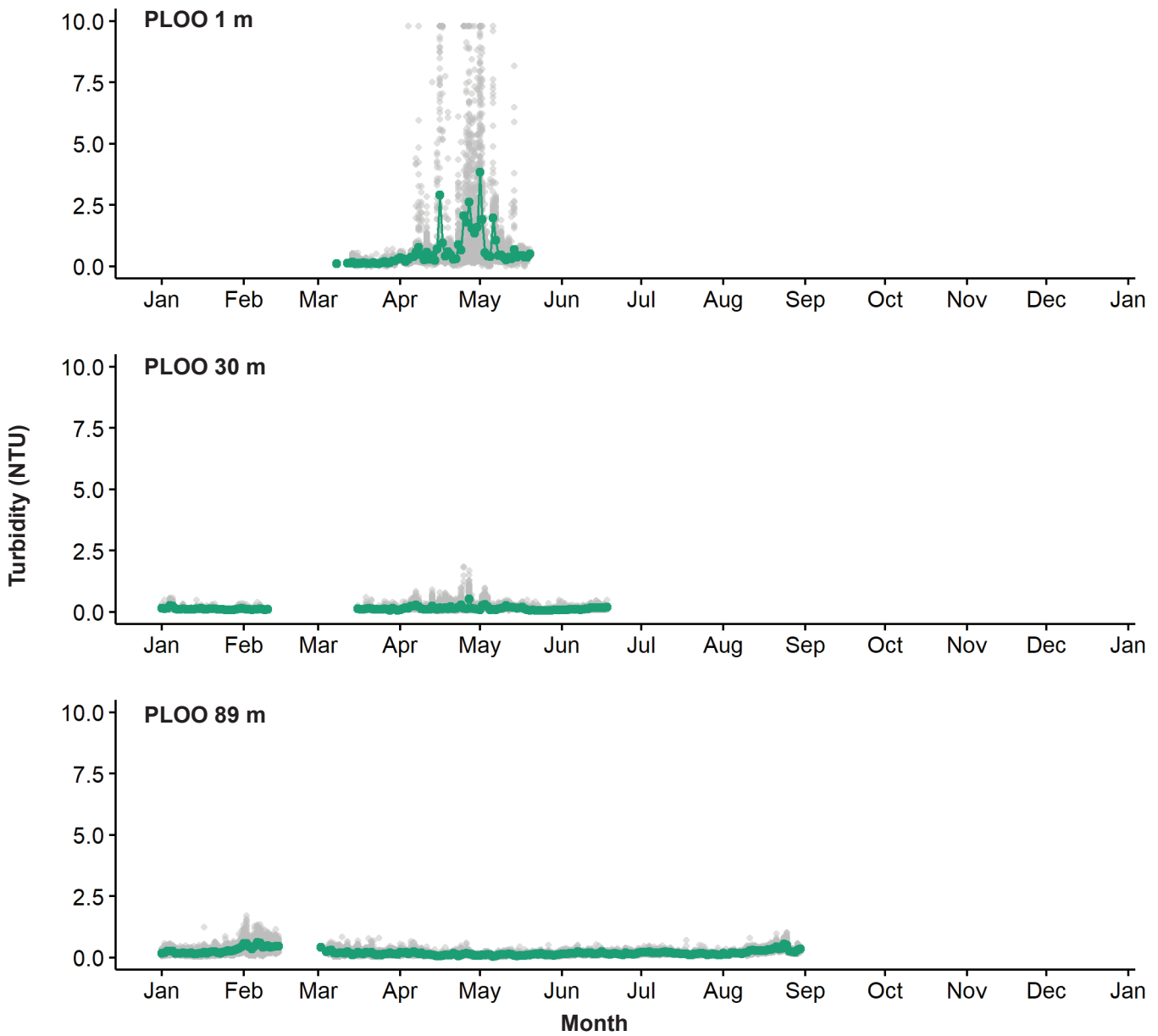
Appendix A.20 *continued*



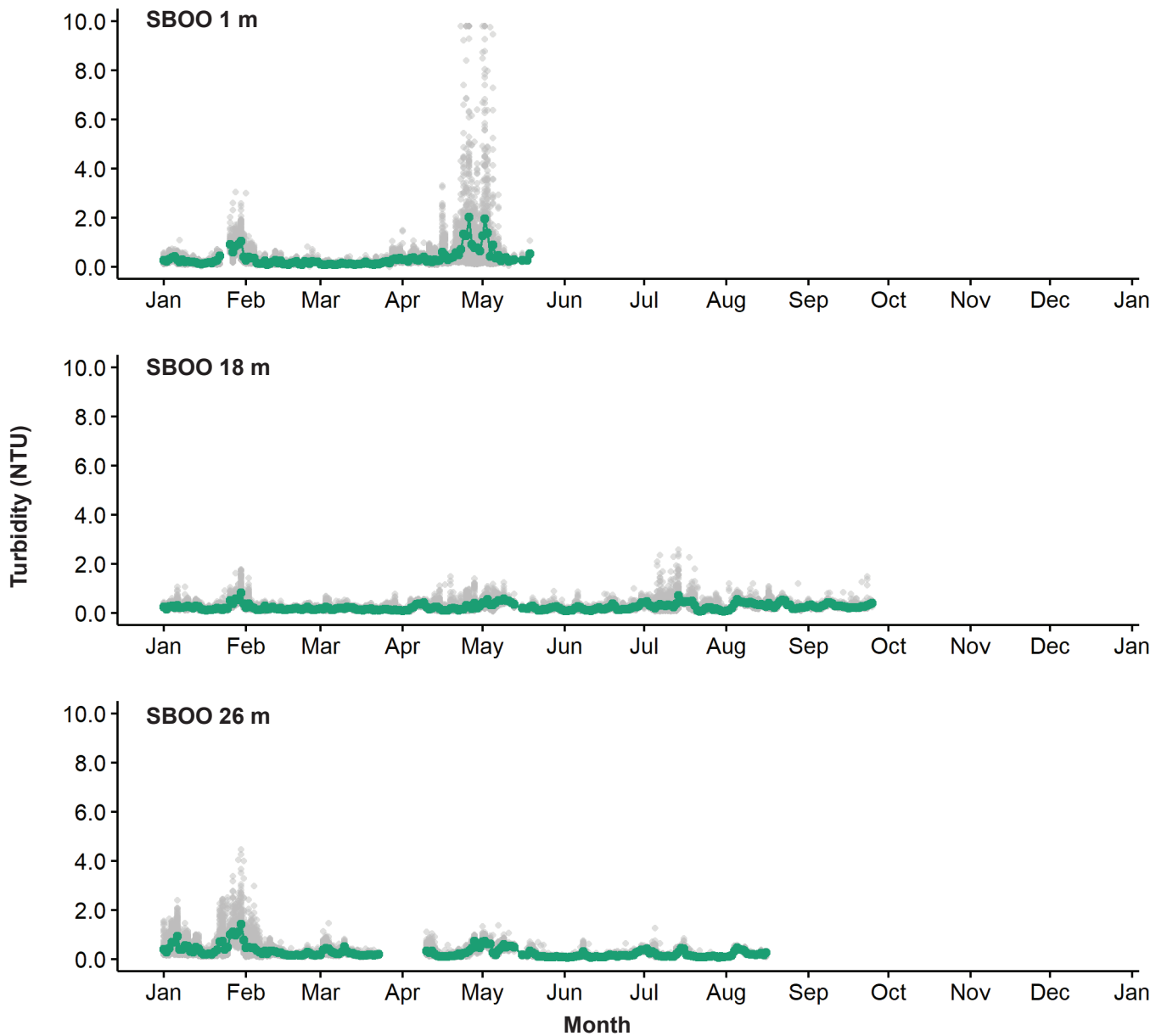
Appendix A.20 *continued*



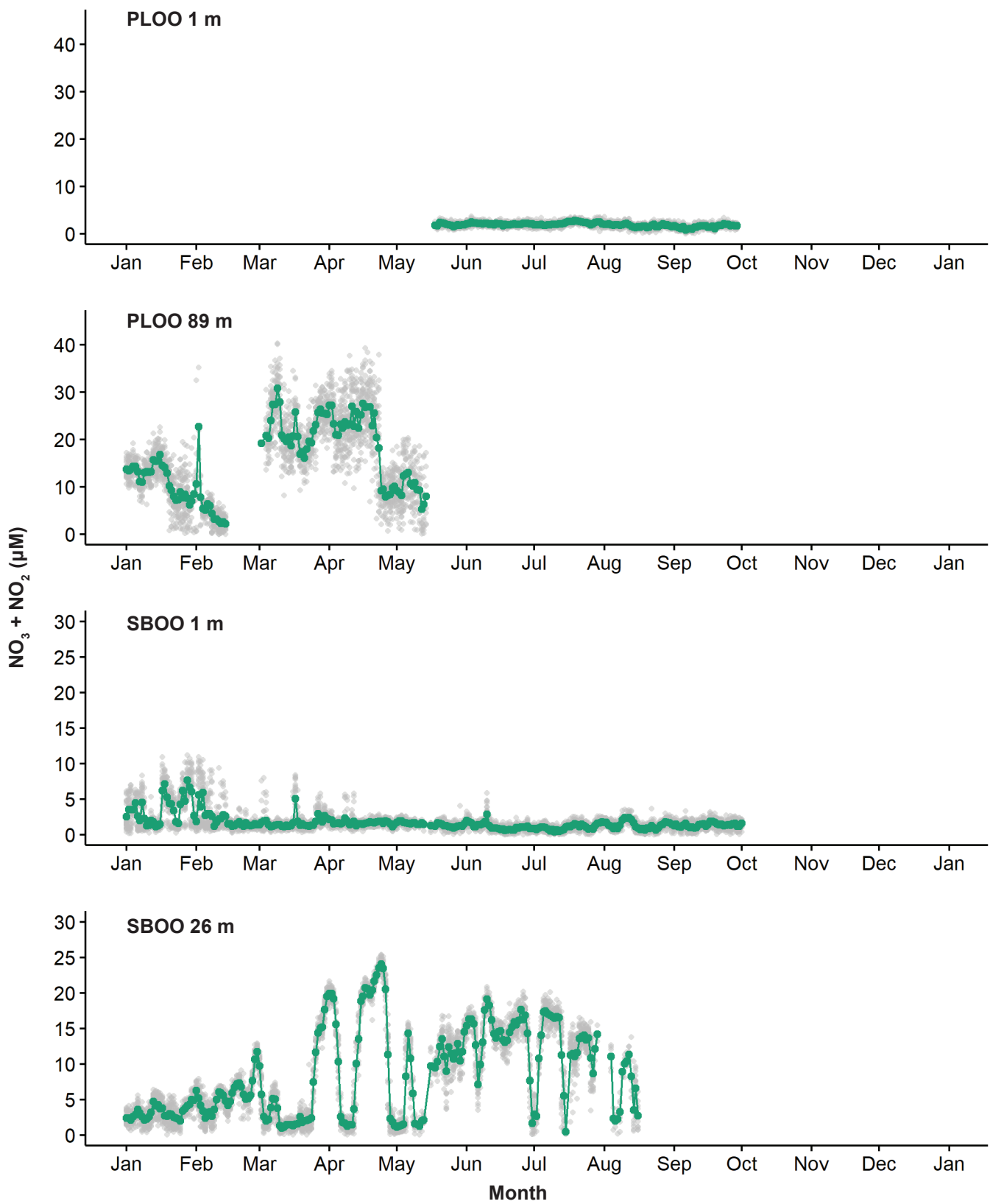
Appendix A.20 *continued*



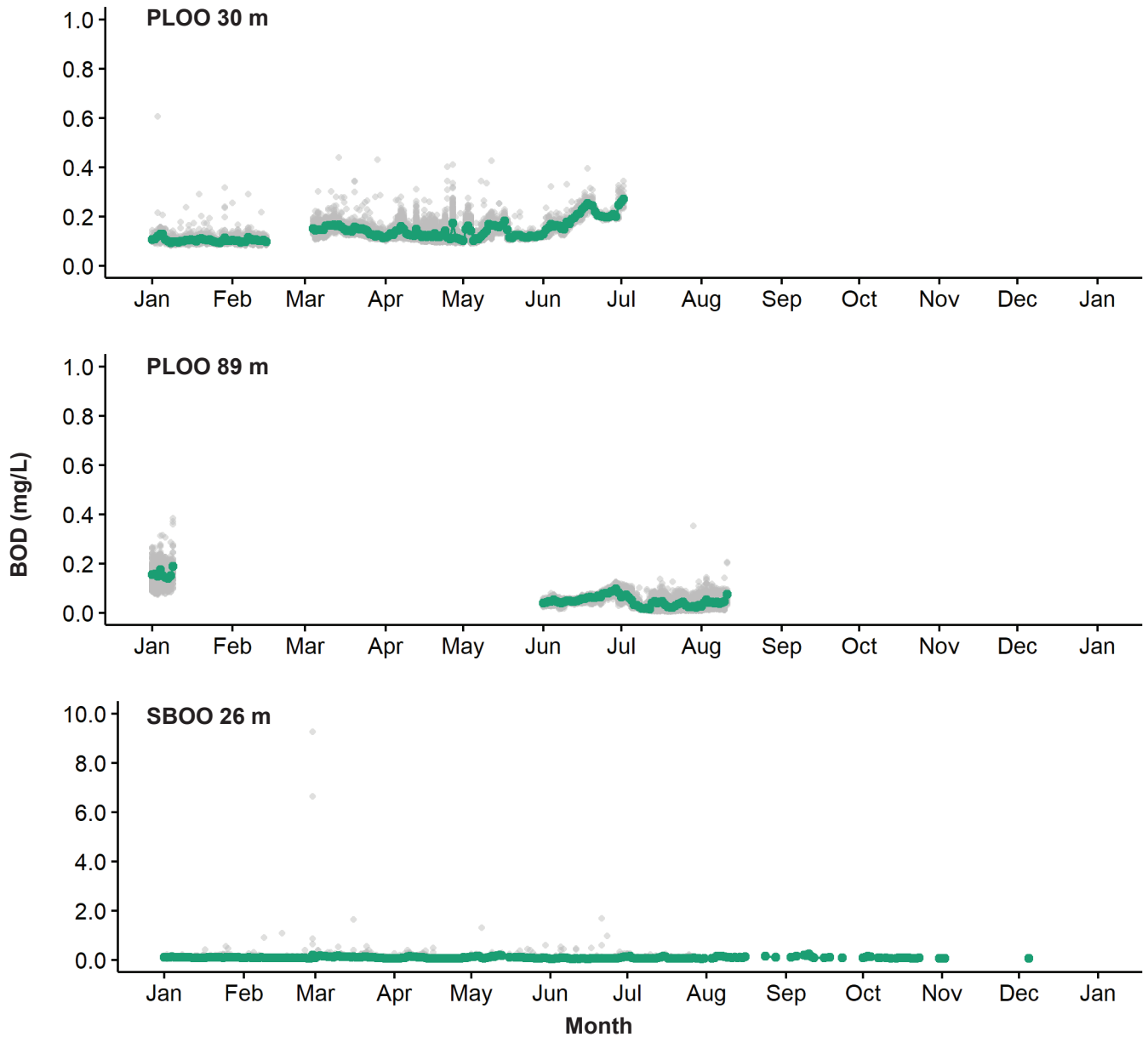
Appendix A.20 *continued*



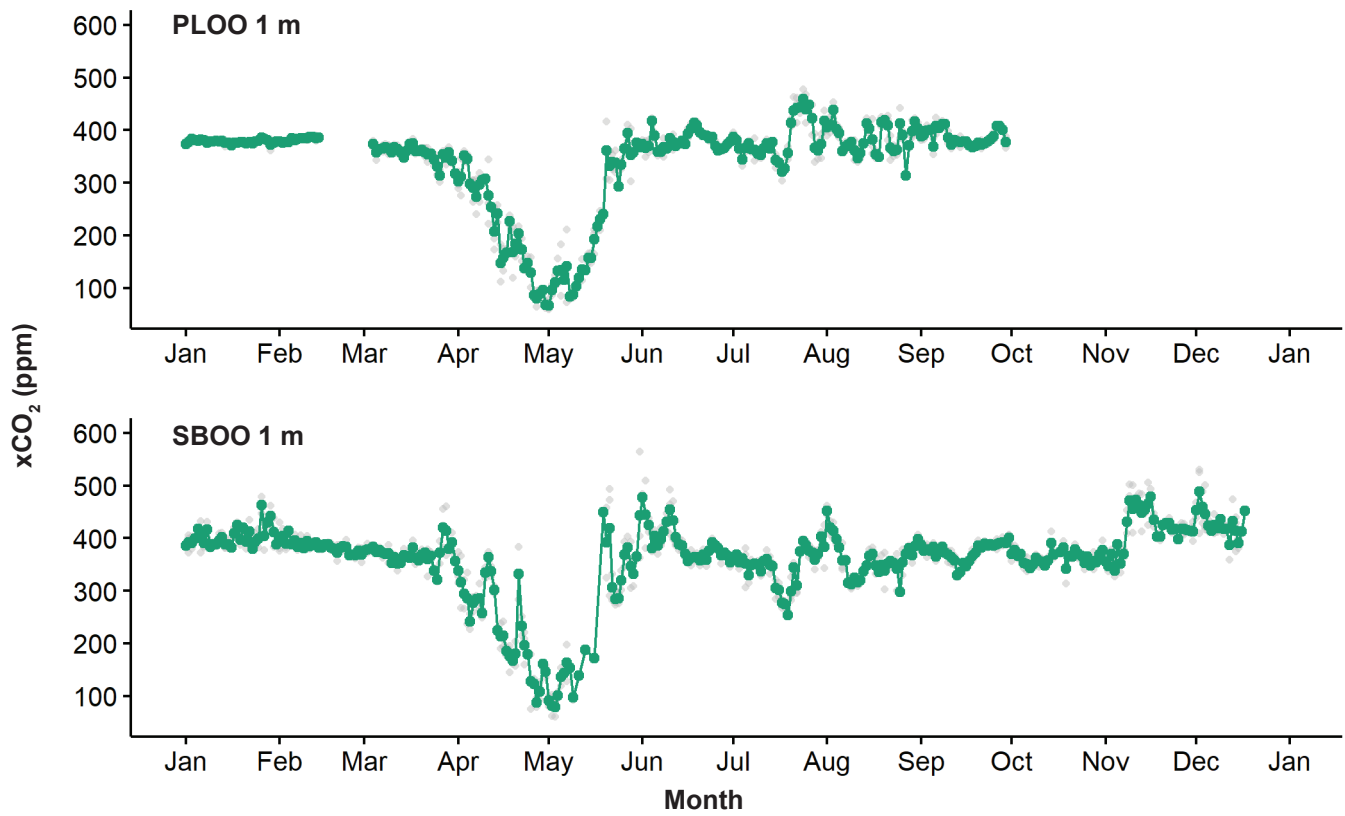
Appendix A.20 *continued*



Appendix A.20 *continued*



Appendix A.20 *continued*



Appendix A.20 *continued*

Appendix A.21

Summary of elevated bacteria densities in samples collected from PLOO shore, kelp, and offshore stations during 2020. Bold values exceed benchmarks for total coliform (> 10,000 CFU/100 mL), fecal coliform (> 400 CFU/100 mL), *Enterococcus* (> 104 CFU/100 mL), or the FTR criterion (total coliforms > 1000 CFU/100 mL and F:T > 0.10).

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
<i>Shore Stations</i>						
D9	22-Jan-20	—	340	96	200	0.28
D11	18-Mar-20	—	5400	200	1000	0.04
D8-B	18-Mar-20	—	200	120	260	0.60
D11	15-Apr-20	—	3000	220	180	0.07
D12	7-Oct-20	—	40	42	140	1.05
<i>Offshore Stations</i>						
F30	19-Feb-20	60	—	—	600	—
F30	19-Feb-20	80	—	—	520	—
F31	19-Feb-20	80	—	—	160	—
F32	19-Feb-20	80	—	—	200	—
F21	21-May-20	80	—	—	200	—
F11	22-May-20	60	—	—	120	—
F29	18-Aug-20	98	—	—	220	—
F30	18-Aug-20	80	—	—	900	—
F30	18-Aug-20	98	—	—	300	—
F31	18-Aug-20	80	—	—	260	—
F17	20-Aug-20	80	—	—	320	—
F19	20-Aug-20	80	—	—	110	—
F26	10-Nov-20	60	—	—	160	—
F27	10-Nov-20	80	—	—	260	—
F27	10-Nov-20	60	—	—	240	—
F28	10-Nov-20	60	—	—	540	—
F29	10-Nov-20	80	—	—	240	—
F15	13-Nov-20	60	—	—	160	—
F18	13-Nov-20	80	—	—	110	—
F19	13-Nov-20	80	—	—	140	—
F20	13-Nov-20	80	—	—	160	—

Appendix A.22

Summary of elevated bacteria densities in samples collected from SBOO shore, kelp, and offshore stations during 2020. Bold values exceed benchmarks for total coliform (> 10,000 CFU/100 mL), fecal coliform (> 400 CFU/100 mL), *Enterococcus* (> 104 CFU/100 mL), or the FTR criterion (total coliforms > 1000 CFU/100 mL and F:T > 0.10).

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
<i>Shorline Stations South of the USA/Mexico Border</i>						
S0	7-Jan-20	—	5000	1200	2000	0.24
S2	7-Jan-20	—	8000	460	82	0.06
S3	7-Jan-20	—	7000	760	110	0.11
S0	14-Jan-20	—	3200	400	180	0.13
S2	14-Jan-20	—	7200	1400	40	0.19
S3	14-Jan-20	—	16000	12000	2200	0.75
S0	21-Jan-20	—	14000	400	1400	0.03
S2	21-Jan-20	—	1200	240	120	0.20
S3	21-Jan-20	—	5800	600	360	0.10
S0	28-Jan-20	—	7000	600	1800	0.09
S2	28-Jan-20	—	6200	1000	540	0.16
S3	28-Jan-20	—	16000	960	60	0.06
S0	4-Feb-20	—	5400	600	1200	0.11
S2	4-Feb-20	—	480	280	140	0.58
S3	4-Feb-20	—	16000	6600	8200	0.41
S0	18-Feb-20	—	4000	500	820	0.13
S0	25-Feb-20	—	8600	1200	2000	0.14
S2	25-Feb-20	—	1800	320	32	0.18
S0	3-Mar-20	—	16000	5400	440	0.34
S2	3-Mar-20	—	800	120	340	0.15
S0	10-Mar-20	—	16000	3800	4800	0.24
S2	10-Mar-20	—	6000	680	760	0.11
S3	10-Mar-20	—	4000	440	480	0.11
S0	17-Mar-20	—	16000	2000	2000	0.13
S2	17-Mar-20	—	3800	480	380	0.13
S3	17-Mar-20	—	4600	640	720	0.14
S0	30-Mar-20	—	6000	2800	600	0.47
S2	30-Mar-20	—	8000	1000	64	0.13

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S3	30-Mar-20	—	16000	9000	500	0.56
S0	7-Apr-20	—	16000	4000	5000	0.25
S2	7-Apr-20	—	3800	500	960	0.13
S3	7-Apr-20	—	16000	12000	12000	0.75
S0	14-Apr-20	—	8800	1400	1000	0.16
S3	14-Apr-20	—	580	160	120	0.28
S0	21-Apr-20	—	16000	2200	2800	0.14
S3	21-Apr-20	—	16000	11000	3000	0.69
S0	28-Apr-20	—	16000	12000	6800	0.75
S3	28-Apr-20	—	600	220	400	0.37
S0	5-May-20	—	6800	1000	480	0.15
S2	5-May-20	—	200	32	140	0.16
S0	19-May-20	—	2200	960	600	0.44
S0	26-May-20	—	13000	5600	880	0.43
S0	9-Jun-20	—	1600	400	220	0.25
S0	23-Jun-20	—	10000	2200	2800	0.22
S0	30-Jun-20	—	440	120	260	0.27
S0	7-Jul-20	—	3800	2600	1100	0.68
S0	14-Jul-20	—	2400	820	480	0.34
S0	21-Jul-20	—	16000	2200	8400	0.14
S0	4-Aug-20	—	4400	1200	880	0.27
S2	4-Aug-20	—	16000	4800	600	0.30
S3	4-Aug-20	—	16000	8600	5000	0.54
S0	11-Aug-20	—	160	60	110	0.38
S0	25-Aug-20	—	10000	240	300	0.02

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S0	1-Sep-20	—	9000	620	320	0.07
S0	15-Sep-20	—	9400	560	560	0.06
S0	22-Sep-20	—	9000	900	820	0.10
S0	29-Sep-20	—	1400	540	300	0.39
S2	29-Sep-20	—	1300	540	240	0.42
S3	29-Sep-20	—	1000	640	100	0.64
S0	13-Oct-20	—	16000	3200	800	0.20
S3	20-Oct-20	—	16000	7000	1200	0.44
S0	27-Oct-20	—	8200	1400	960	0.17
S0	10-Nov-20	—	16000	2800	2000	0.18
S2	10-Nov-20	—	6800	1100	30	0.16
S3	10-Nov-20	—	16000	2000	200	0.13
S0	24-Nov-20	—	16000	12000	12000	0.75
S0	8-Dec-20	—	16000	12000	12000	0.75
S0	15-Dec-20	—	16000	9200	6200	0.58
S0	22-Dec-20	—	16000	12000	12000	0.75
S3	22-Dec-20	—	2600	2400	50	0.92
S0	29-Dec-20	—	16000	3200	3600	0.20
S2	29-Dec-20	—	11000	1000	680	0.09
S3	29-Dec-20	—	1000	140	180	0.14
<i>Shorline Stations North of the USA/Mexico Border</i>						
S5*	1-Jan-20	—	16000	12000	10000	0.75
S6*	1-Jan-20	—	—	—	200	—
S11*	1-Jan-20	—	—	—	220	—
S5*	3-Jan-20	—	16000	12000	12000	0.75
S6*	3-Jan-20	—	—	—	360	—

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S11*	3-Jan-20	—	—	—	300	—
S5*	5-Jan-20	—	16000	12000	12000	0.75
S6*	5-Jan-20	—	—	—	120	—
S4	7-Jan-20	—	7400	800	420	0.11
S10	7-Jan-20	—	2600	900	280	0.35
S4*	9-Jan-20	—	4400	1400	500	0.32
S10*	9-Jan-20	—	6400	980	520	0.15
S4*	11-Jan-20	—	10000	3200	1000	0.32
S10*	11-Jan-20	—	5000	540	240	0.11
S4*	13-Jan-20	—	5600	740	180	0.13
S10*	13-Jan-20	—	4400	500	76	0.11
S4	14-Jan-20	—	15000	3200	360	0.21
S10	14-Jan-20	—	11000	2000	260	0.18
S4*	16-Jan-20	—	16000	12000	4200	0.75
S10*	16-Jan-20	—	16000	12000	12000	0.75
S4*	18-Jan-20	—	16000	4800	2600	0.30
S10*	18-Jan-20	—	16000	12000	12000	0.75
S4*	20-Jan-20	—	2600	400	180	0.15
S10*	20-Jan-20	—	16000	2000	1100	0.13
S4	21-Jan-20	—	16000	6600	2400	0.41
S5	21-Jan-20	—	16000	9000	3000	0.56
S4*	23-Jan-20	—	2600	200	160	0.08
S5*	23-Jan-20	—	16000	12000	12000	0.75
S4*	25-Jan-20	—	—	—	400	—
S5*	25-Jan-20	—	3800	1200	340	0.32
S4*	27-Jan-20	—	—	—	620	—

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S4	4-Feb-20	—	16000	12000	11000	0.75
S5	4-Feb-20	—	200	200	110	1.00
S10	4-Feb-20	—	16000	12000	12000	0.75
S4*	5-Feb-20	—	5600	3600	4200	0.64
S5*	5-Feb-20	—	—	—	12000	—
S10*	5-Feb-20	—	16000	12000	12000	0.75
S4*	7-Feb-20	—	1500	300	240	0.20
S5*	7-Feb-20	—	—	—	3000	—
S10*	7-Feb-20	—	1200	440	140	0.37
S4*	9-Feb-20	—	11000	2200	4200	0.20
S5*	9-Feb-20	—	—	—	9000	—
S10*	9-Feb-20	—	15000	4400	3800	0.29
S5*	12-Feb-20	—	16000	12000	12000	0.75
S5*	14-Feb-20	—	16000	12000	12000	0.75
S5*	16-Feb-20	—	16000	12000	12000	0.75
S4	18-Feb-20	—	8200	400	920	0.05
S5	18-Feb-20	—	16000	12000	12000	0.75
S6	18-Feb-20	—	4600	1200	540	0.26
S10	18-Feb-20	—	16000	7800	5800	0.49
S11	18-Feb-20	—	9000	1800	400	0.20
S4*	20-Feb-20	—	—	—	220	—
S5*	20-Feb-20	—	16000	12000	12000	0.75
S10*	20-Feb-20	—	340	110	200	0.32
S5*	21-Feb-20	—	160	62	120	0.39
S5*	23-Feb-20	—	—	—	2200	—
S4	25-Feb-20	—	12000	2200	240	0.18
S10	25-Feb-20	—	4400	1400	120	0.32
S4*	27-Feb-20	—	8000	3000	1000	0.38

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S10*	27-Feb-20	—	16000	9000	1000	0.56
S4*	28-Feb-20	—	1100	360	240	0.33
S10*	28-Feb-20	—	6400	1600	1400	0.25
S4	3-Mar-20	—	16000	12000	12000	0.75
S5	3-Mar-20	—	16000	12000	12000	0.75
S10	3-Mar-20	—	16000	12000	12000	0.75
S4*	5-Mar-20	—	9000	1200	360	0.13
S5*	5-Mar-20	—	16000	12000	12000	0.75
S10*	5-Mar-20	—	7600	1000	110	0.13
S4*	7-Mar-20	—	16000	12000	8000	0.75
S5*	7-Mar-20	—	16000	12000	9200	0.75
S10*	7-Mar-20	—	16000	12000	10000	0.75
S4*	9-Mar-20	—	16000	4800	960	0.30
S5*	9-Mar-20	—	5600	1000	540	0.18
S10*	9-Mar-20	—	13000	3800	860	0.29
S4	10-Mar-20	—	2200	740	580	0.34
S5	10-Mar-20	—	16000	5000	4800	0.31
S6	10-Mar-20	—	12000	3400	3800	0.28
S8	10-Mar-20	—	15000	4600	2200	0.31
S9	10-Mar-20	—	440	120	340	0.27
S10	10-Mar-20	—	3200	800	980	0.25
S11	10-Mar-20	—	10000	3800	2000	0.38
S12	10-Mar-20	—	16000	12000	12000	0.75
S4*	12-Mar-20	—	400	46	140	0.12
S5*	12-Mar-20	—	16000	12000	4000	0.75
S6*	12-Mar-20	—	16000	12000	12000	0.75
S8*	12-Mar-20	—	11000	2000	320	0.18
S10*	12-Mar-20	—	1400	94	220	0.07
S11*	12-Mar-20	—	16000	12000	12000	0.75
S12*	12-Mar-20	—	16000	12000	9000	0.75
S4*	14-Mar-20	—	—	—	1200	—
S5*	14-Mar-20	—	16000	12000	12000	0.75

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S6*	14-Mar-20	—	16000	12000	12000	0.75
S8*	14-Mar-20	—	16000	12000	4600	0.75
S10*	14-Mar-20	—	—	—	820	—
S11*	14-Mar-20	—	16000	12000	12000	0.75
S12*	14-Mar-20	—	16000	12000	7400	0.75
S5*	16-Mar-20	—	16000	12000	12000	0.75
S6*	16-Mar-20	—	16000	4600	800	0.29
S11*	16-Mar-20	—	16000	12000	11000	0.75
S12*	16-Mar-20	—	8200	1600	120	0.20
S4	17-Mar-20	—	16000	4400	2000	0.28
S5	17-Mar-20	—	16000	12000	12000	0.75
S6	17-Mar-20	—	16000	12000	12000	0.75
S8	17-Mar-20	—	14000	800	300	0.06
S10	17-Mar-20	—	16000	6200	4600	0.39
S11	17-Mar-20	—	16000	12000	12000	0.75
S12	17-Mar-20	—	16000	1000	500	0.06
S4*	19-Mar-20	—	16000	12000	8000	0.75
S5*	19-Mar-20	—	16000	12000	12000	0.75
S10*	19-Mar-20	—	16000	12000	12000	0.75
S4*	20-Mar-20	—	7200	800	1100	0.11
S5*	20-Mar-20	—	11000	1200	1000	0.11
S10*	20-Mar-20	—	8400	900	840	0.11
S5	24-Mar-20	—	16000	1200	700	0.08
S6	24-Mar-20	—	16000	5800	980	0.36
S8	24-Mar-20	—	16000	820	140	0.05
S11	24-Mar-20	—	16000	1000	380	0.06
S12	24-Mar-20	—	16000	8600	1200	0.54
S4	30-Mar-20	—	16000	12000	6200	0.75
S5	30-Mar-20	—	16000	12000	12000	0.75
S10	30-Mar-20	—	16000	12000	12000	0.75
S4*	1-Apr-20	—	2800	340	140	0.12
S5*	1-Apr-20	—	16000	12000	12000	0.75
S10*	1-Apr-20	—	6600	540	320	0.08

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S4*	3-Apr-20	—	820	300	340	0.37
S5*	3-Apr-20	—	16000	12000	12000	0.75
S10*	3-Apr-20	—	—	420	600	—
S4*	5-Apr-20	—	—	—	280	—
S5*	5-Apr-20	—	16000	12000	12000	0.75
S10*	5-Apr-20	—	—	80	180	—
S4	7-Apr-20	—	16000	5600	4000	0.35
S5	7-Apr-20	—	12000	2000	3400	0.17
S6	7-Apr-20	—	16000	8600	8200	0.54
S8	7-Apr-20	—	2400	1800	980	0.75
S9	7-Apr-20	—	2400	1100	740	0.46
S10	7-Apr-20	—	16000	4600	3600	0.29
S11	7-Apr-20	—	16000	12000	8200	0.75
S12	7-Apr-20	—	6000	1600	800	0.27
S4*	9-Apr-20	—	13000	3200	1600	0.25
S5*	9-Apr-20	—	7600	2400	1100	0.32
S6*	9-Apr-20	—	8400	1100	840	0.13
S9*	9-Apr-20	—	16000	6800	1800	0.43
S10*	9-Apr-20	—	3200	240	480	0.08
S11*	9-Apr-20	—	8200	1000	1400	0.12
S12*	9-Apr-20	—	16000	12000	7200	0.75
S4*	11-Apr-20	—	16000	8000	3600	0.50
S5*	11-Apr-20	—	16000	12000	12000	0.75
S6*	11-Apr-20	—	16000	12000	12000	0.75
S8*	11-Apr-20	—	16000	6000	4600	0.38
S9*	11-Apr-20	—	16000	6000	3600	0.38
S10*	11-Apr-20	—	—	—	5000	—
S11*	11-Apr-20	—	16000	12000	12000	0.75
S12*	11-Apr-20	—	16000	12000	12000	0.75
S4*	13-Apr-20	—	16000	7400	1800	0.46
S5*	13-Apr-20	—	16000	12000	12000	0.75
S6*	13-Apr-20	—	16000	12000	5400	0.75
S8*	13-Apr-20	—	16000	8200	1000	0.51
S10*	13-Apr-20	—	—	—	3000	—
S11*	13-Apr-20	—	16000	12000	4600	0.75

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S12*	13-Apr-20	—	16000	4200	1100	0.26
S4	14-Apr-20	—	4800	1600	620	0.33
S5	14-Apr-20	—	16000	12000	12000	0.75
S6	14-Apr-20	—	460	60	160	0.13
S10	14-Apr-20	—	6800	2000	660	0.29
S11	14-Apr-20	—	16000	7200	4800	0.45
S5*	16-Apr-20	—	16000	12000	12000	0.75
S6*	16-Apr-20	—	—	—	12000	—
S11*	16-Apr-20	—	16000	12000	12000	0.75
S5*	17-Apr-20	—	16000	12000	12000	0.75
S6*	17-Apr-20	—	—	—	12000	—
S11*	17-Apr-20	—	16000	12000	12000	0.75
S5*	19-Apr-20	—	16000	3600	740	0.23
S6*	19-Apr-20	—	—	—	760	—
S11*	19-Apr-20	—	2200	380	1000	0.17
S4	21-Apr-20	—	16000	7000	820	0.44
S5	21-Apr-20	—	16000	12000	4200	0.75
S6	21-Apr-20	—	3000	600	480	0.20
S10	21-Apr-20	—	16000	12000	2200	0.75
S11	21-Apr-20	—	16000	9000	2200	0.56
S4*	23-Apr-20	—	16000	4000	920	0.25
S5*	23-Apr-20	—	16000	12000	3200	0.75
S6*	23-Apr-20	—	240	20	340	0.08
S10*	23-Apr-20	—	16000	11000	3000	0.69
S11*	23-Apr-20	—	600	60	520	0.10
S4*	24-Apr-20	—	15000	9600	3200	0.64
S5*	24-Apr-20	—	16000	12000	4200	0.75
S10*	24-Apr-20	—	16000	12000	6200	0.75
S11*	24-Apr-20	—	—	—	220	—
S4*	26-Apr-20	—	1000	580	500	0.58
S5*	26-Apr-20	—	16000	12000	12000	0.75
S10*	26-Apr-20	—	4800	2800	840	0.58

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S11*	26-Apr-20	—	—	—	3000	—
S4	28-Apr-20	—	600	2	260	0.00
S5	28-Apr-20	—	16000	12000	12000	0.75
S6	28-Apr-20	—	16000	3800	1600	0.24
S8	28-Apr-20	—	200	24	120	0.12
S9	28-Apr-20	—	200	40	220	0.20
S10	28-Apr-20	—	200	20	120	0.10
S11	28-Apr-20	—	16000	6600	3200	0.41
S12	28-Apr-20	—	600	340	420	0.57
S4*	30-Apr-20	—	—	—	12000	—
S5*	30-Apr-20	—	16000	12000	12000	0.75
S6*	30-Apr-20	—	400	220	600	0.55
S10*	30-Apr-20	—	—	—	12000	—
S11*	30-Apr-20	—	1200	520	600	0.43
S12*	30-Apr-20	—	—	—	180	—
S4*	2-May-20	—	—	—	5200	—
S5*	2-May-20	—	16000	12000	12000	0.75
S10*	2-May-20	—	—	—	12000	—
S11*	2-May-20	—	20	2	160	0.10
S4*	4-May-20	—	—	—	520	—
S5*	4-May-20	—	16000	12000	12000	0.75
S10*	4-May-20	—	—	—	12000	—
S4	5-May-20	—	4800	2600	700	0.54
S5	5-May-20	—	11000	1800	820	0.16
S10	5-May-20	—	6200	1400	720	0.23
S4*	7-May-20	—	16000	8400	2600	0.53
S5*	7-May-20	—	2000	720	400	0.36
S10*	7-May-20	—	16000	12000	8400	0.75
S5*	9-May-20	—	16000	12000	12000	0.75
S5*	11-May-20	—	16000	12000	12000	0.75
S5	12-May-20	—	16000	12000	12000	0.75

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S6	12-May-20	—	4200	1200	4800	0.29
S8	12-May-20	—	1200	100	460	0.08
S11	12-May-20	—	16000	2800	9400	0.18
S12	12-May-20	—	6200	800	4400	0.13
S5*	14-May-20	—	16000	12000	12000	0.75
S6*	14-May-20	—	80	60	260	0.75
S11*	14-May-20	—	720	600	800	0.83
S5*	15-May-20	—	7800	7200	12000	0.92
S4	19-May-20	—	200	30	180	0.15
S5	26-May-20	—	16000	12000	12000	0.75
S6	26-May-20	—	8400	2400	1600	0.29
S11	26-May-20	—	12000	1200	5200	0.10
S5*	28-May-20	—	16000	12000	12000	0.75
S6*	28-May-20	—	2600	1000	800	0.38
S11*	28-May-20	—	12000	6000	7000	0.50
S5*	29-May-20	—	16000	12000	12000	0.75
S6*	29-May-20	—	9800	4000	2800	0.41
S11*	29-May-20	—	16000	9000	7800	0.56
S5*	31-May-20	—	16000	6800	6800	0.43
S6*	31-May-20	—	280	140	140	0.50
S11*	31-May-20	—	2800	300	360	0.11
S5	9-Jun-20	—	16000	12000	12000	0.75
S11	9-Jun-20	—	840	260	120	0.31
S5*	11-Jun-20	—	16000	12000	12000	0.75
S11*	11-Jun-20	—	—	—	6400	—
S5*	12-Jun-20	—	16000	12000	12000	0.75
S11*	12-Jun-20	—	—	—	3000	—
S5*	14-Jun-20	—	16000	2800	9000	0.18

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S12	16-Jun-20	—	1300	300	260	0.23
S5	23-Jun-20	—	16000	12000	5400	0.75
S6	23-Jun-20	—	800	280	160	0.35
S11	23-Jun-20	—	16000	6000	4600	0.38
S5*	25-Jun-20	—	16000	12000	12000	0.75
S6*	25-Jun-20	—	—	—	3000	—
S11*	25-Jun-20	—	16000	10000	5600	0.63
S5*	27-Jun-20	—	16000	12000	12000	0.75
S6*	27-Jun-20	—	—	—	580	—
S11*	27-Jun-20	—	16000	12000	6600	0.75
S5*	29-Jun-20	—	16000	12000	12000	0.75
S6*	29-Jun-20	—	—	—	5400	—
S11*	29-Jun-20	—	16000	12000	12000	0.75
S5	30-Jun-20	—	16000	12000	12000	0.75
S6	30-Jun-20	—	16000	7000	4200	0.44
S11	30-Jun-20	—	16000	12000	11000	0.75
S12	30-Jun-20	—	1200	200	180	0.17
S5*	2-Jul-20	—	240	50	220	0.21
S5*	4-Jul-20	—	—	—	12000	—
S5*	6-Jul-20	—	—	—	2800	—
S5	7-Jul-20	—	16000	12000	12000	0.75
S6	7-Jul-20	—	1400	180	960	0.13
S11	7-Jul-20	—	1600	580	1800	0.36
S5*	9-Jul-20	—	16000	12000	12000	0.75
S6*	9-Jul-20	—	16000	2600	1600	0.16
S11*	9-Jul-20	—	16000	12000	11000	0.75
S5*	11-Jul-20	—	16000	12000	12000	0.75
S6*	11-Jul-20	—	1000	520	300	0.52
S11*	11-Jul-20	—	16000	12000	11000	0.75

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S5*	13-Jul-20	—	16000	12000	12000	0.75
S5	14-Jul-20	—	16000	12000	12000	0.75
S5*	16-Jul-20	—	4200	960	540	0.23
S5*	17-Jul-20	—	16000	12000	12000	0.75
S5*	19-Jul-20	—	16000	12000	12000	0.75
S5	21-Jul-20	—	5000	2600	500	0.52
S10	21-Jul-20	—	7000	1000	640	0.14
S5*	23-Jul-20	—	9000	600	1000	0.07
S5*	24-Jul-20	—	—	12000	12000	—
S5	28-Jul-20	—	16000	12000	12000	0.75
S11	28-Jul-20	—	6800	680	380	0.10
S5*	30-Jul-20	—	6800	600	340	0.09
S4	4-Aug-20	—	6000	1100	780	0.18
S5	4-Aug-20	—	16000	3400	1100	0.21
S6	4-Aug-20	—	16000	6800	2200	0.43
S10	4-Aug-20	—	1400	320	280	0.23
S11	4-Aug-20	—	16000	4400	840	0.28
S12	18-Aug-20	—	11000	1200	80	0.11
S4	8-Sep-20	—	8800	1800	880	0.20
S5	8-Sep-20	—	11000	5000	800	0.45
S6	8-Sep-20	—	7200	2400	980	0.33
S10	8-Sep-20	—	5200	2000	1100	0.38
S11	8-Sep-20	—	6200	2600	600	0.42
S12	8-Sep-20	—	2400	440	200	0.18
S4	10-Nov-20	—	16000	4000	110	0.25
S5	10-Nov-20	—	16000	12000	12000	0.75
S10	10-Nov-20	—	16000	12000	720	0.75

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
S5*	12-Nov-20	—	6400	660	50	0.10
<i>Kelp Stations</i>						
I19	2-Jan-20	6	2200	760	200	0.35
I19	2-Jan-20	2	16000	3200	720	0.20
I19	2-Jan-20	11	2400	320	180	0.13
I24	2-Jan-20	11	2400	520	180	0.22
I24	2-Jan-20	6	9600	1400	320	0.15
I24	2-Jan-20	2	7000	1000	340	0.14
I25	2-Jan-20	6	3200	620	180	0.19
I25	2-Jan-20	9	800	68	220	0.09
I26	2-Jan-20	6	3400	680	460	0.20
I26	2-Jan-20	9	3400	480	600	0.14
I26	2-Jan-20	2	2600	360	860	0.14
I32	2-Jan-20	6	200	48	480	0.24
I32	2-Jan-20	9	320	64	620	0.20
I32	2-Jan-20	2	340	58	300	0.17
I40	2-Jan-20	9	2400	400	260	0.17
I40	2-Jan-20	6	2800	380	200	0.14
I19	7-Jan-20	6	9200	1800	580	0.20
I19	7-Jan-20	11	8800	1400	440	0.16
I19	7-Jan-20	2	11000	900	240	0.08
I26	7-Jan-20	9	60	20	400	0.33
I40	7-Jan-20	2	1400	380	50	0.27
I19	13-Jan-20	11	1400	340	94	0.24
I19	13-Jan-20	2	11000	2000	280	0.18
I19	13-Jan-20	6	7400	1200	280	0.16
I24	13-Jan-20	11	1500	200	58	0.13
I24	13-Jan-20	6	16000	2000	300	0.13
I24	13-Jan-20	2	16000	1800	660	0.11
I25	13-Jan-20	9	3200	420	74	0.13
I25	13-Jan-20	6	10000	1100	120	0.11
I25	13-Jan-20	2	14000	1200	140	0.09
I40	13-Jan-20	6	5800	780	60	0.13
I40	13-Jan-20	2	14000	1000	160	0.07
I19	21-Jan-20	2	5400	720	160	0.13
I24	21-Jan-20	2	16000	600	2200	0.04
I26	21-Jan-20	9	20	2	200	0.10

^aResample

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
I40	21-Jan-20	2	12000	5000	960	0.42
I40	21-Jan-20	6	11000	1200	800	0.11
I40	21-Jan-20	9	5600	200	220	0.04
I19	28-Jan-20	6	3200	540	160	0.17
I19	28-Jan-20	11	1600	200	88	0.13
I19	5-Feb-20	6	3600	1000	520	0.28
I19	5-Feb-20	11	4800	1100	360	0.23
I19	5-Feb-20	2	4200	600	380	0.14
I40	5-Feb-20	2	1800	560	880	0.31
I24	10-Feb-20	2	600	520	100	0.87
I26	10-Feb-20	9	600	440	340	0.73
I26	10-Feb-20	2	1800	740	1100	0.41
I26	10-Feb-20	6	2000	400	440	0.20
I32	10-Feb-20	9	1800	860	480	0.48
I32	10-Feb-20	2	200	84	300	0.42
I32	10-Feb-20	6	2800	680	640	0.24
I40	10-Feb-20	2	4200	980	760	0.23
I19	18-Feb-20	2	580	260	340	0.45
I19	18-Feb-20	6	8200	2400	640	0.29
I19	18-Feb-20	11	16000	3800	1200	0.24
I40	18-Feb-20	6	4400	1100	360	0.25
I40	18-Feb-20	2	2000	400	1100	0.20
I19	5-Mar-20	2	16000	12000	820	0.75
I19	5-Mar-20	11	1200	560	2	0.47
I19	5-Mar-20	6	16000	1600	380	0.10
I24	9-Mar-20	2	640	160	110	0.25
I25	9-Mar-20	2	1500	240	100	0.16
I40	9-Mar-20	2	520	360	240	0.69
I24	16-Mar-20	2	16000	11000	1000	0.69
I24	16-Mar-20	6	3400	740	32	0.22
I24	16-Mar-20	11	2400	460	24	0.19
I25	16-Mar-20	2	16000	8600	560	0.54
I25	16-Mar-20	6	4400	1400	92	0.32
I25	16-Mar-20	9	6000	520	42	0.09

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
I32	16-Mar-20	6	1800	300	18	0.17
I32	16-Mar-20	9	2400	320	68	0.13
I39	16-Mar-20	2	2000	440	34	0.22
I40	16-Mar-20	2	16000	8000	700	0.50
I40	16-Mar-20	9	1400	200	18	0.14
I24	24-Mar-20	2	300	480	16	1.60
I25	24-Mar-20	6	2800	420	28	0.15
I25	24-Mar-20	9	1400	200	12	0.14
I25	24-Mar-20	2	6400	800	42	0.13
I26	24-Mar-20	2	2800	1600	26	0.57
I32	24-Mar-20	2	16000	4400	160	0.28
I39	24-Mar-20	2	15000	3200	340	0.21
I40	24-Mar-20	9	1400	360	420	0.26
I40	24-Mar-20	6	11000	400	220	0.04
I19	1-Apr-20	2	4200	980	240	0.23
I19	1-Apr-20	6	3000	340	110	0.11
I19	1-Apr-20	11	1800	120	320	0.07
I40	1-Apr-20	2	14000	5800	600	0.41
I40	1-Apr-20	6	16000	5000	1000	0.31
I40	1-Apr-20	9	4800	700	110	0.15
I19	10-Apr-20	11	7200	1400	1800	0.19
I19	10-Apr-20	2	5800	800	600	0.14
I19	10-Apr-20	6	4600	600	1200	0.13
I24	10-Apr-20	6	1000	300	160	0.30
I24	10-Apr-20	11	980	280	120	0.29
I24	10-Apr-20	2	1400	200	320	0.14
I25	10-Apr-20	9	500	180	200	0.36
I25	10-Apr-20	6	620	140	180	0.23
I26	10-Apr-20	6	480	100	240	0.21
I26	10-Apr-20	2	440	60	120	0.14
I26	10-Apr-20	9	500	40	120	0.08
I32	10-Apr-20	2	16000	12000	11000	0.75
I32	10-Apr-20	9	16000	12000	3200	0.75
I32	10-Apr-20	6	16000	6400	2000	0.40
I40	10-Apr-20	9	400	340	620	0.85
I40	10-Apr-20	6	1100	320	400	0.29
I40	10-Apr-20	2	1000	140	320	0.14

Appendix A.22 *continued*

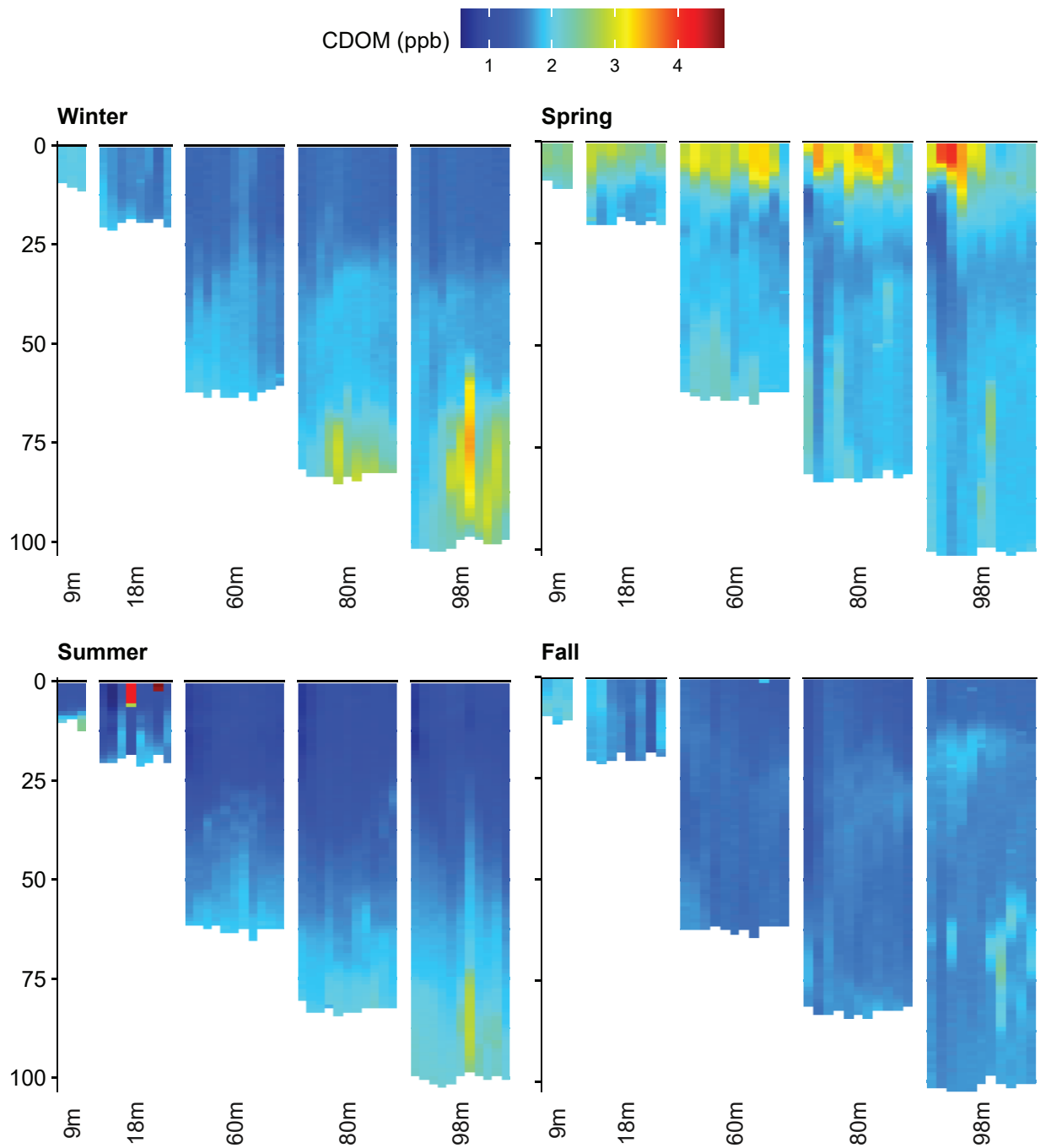
Station	Date	Depth (m)	Total	Fecal	Entero	F:T
I19	14-Apr-20	2	1200	720	220	0.60
I19	14-Apr-20	11	3400	840	340	0.25
I19	14-Apr-20	6	5800	1400	240	0.24
I24	14-Apr-20	11	400	68	320	0.17
I40	14-Apr-20	2	6800	1000	1100	0.15
I40	14-Apr-20	9	7400	1000	1200	0.14
I40	14-Apr-20	6	11000	1100	1800	0.10
I19	21-Apr-20	2	11000	1800	40	0.16
I19	21-Apr-20	6	6800	840	160	0.12
I19	21-Apr-20	11	1600	180	280	0.11
I24	21-Apr-20	6	16000	3800	560	0.24
I24	21-Apr-20	2	16000	3800	320	0.24
I24	21-Apr-20	11	7600	200	300	0.03
I32	21-Apr-20	6	200	76	240	0.38
I32	21-Apr-20	9	140	50	120	0.36
I40	21-Apr-20	2	16000	7600	520	0.48
I40	21-Apr-20	6	6800	1800	280	0.26
I40	21-Apr-20	9	5400	1200	360	0.22
I19	28-Apr-20	2	5400	2000	3000	0.37
I19	28-Apr-20	6	4400	1400	800	0.32
I19	28-Apr-20	11	1200	160	280	0.13
I24	28-Apr-20	6	400	60	160	0.15
I26	28-Apr-20	2	16000	8400	5000	0.53
I26	28-Apr-20	9	2800	460	240	0.16
I32	28-Apr-20	6	1200	800	480	0.67
I32	28-Apr-20	9	800	240	320	0.30
I32	28-Apr-20	2	4400	600	200	0.14
I40	28-Apr-20	6	200	160	110	0.80
I40	28-Apr-20	2	1000	600	600	0.60
I40	28-Apr-20	9	5200	920	440	0.18
I19	5-May-20	6	1000	560	200	0.56
I19	5-May-20	2	6600	3200	140	0.48
I40	5-May-20	9	800	160	260	0.20
I24	12-May-20	11	20	8	120	0.40
I32	12-May-20	6	140	46	110	0.33
I40	12-May-20	9	20	6	110	0.30

Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
I24	21-Jul-20	2	400	96	460	0.24
I24	21-Jul-20	6	2800	300	800	0.11
I40	21-Jul-20	2	400	140	320	0.35
I19	28-Jul-20	2	680	56	120	0.08
I19	4-Aug-20	2	900	560	90	0.62
I19	4-Aug-20	11	16000	9800	2200	0.61
I19	4-Aug-20	6	6000	820	320	0.14
I24	4-Aug-20	6	6400	2200	560	0.34
I24	4-Aug-20	11	4200	660	140	0.16
I25	4-Aug-20	6	9000	2000	580	0.22
I25	4-Aug-20	9	4000	820	140	0.21
I26	4-Aug-20	6	4800	800	100	0.17
I26	4-Aug-20	9	4000	540	100	0.14
I40	4-Aug-20	9	16000	8400	1400	0.53
I40	4-Aug-20	6	16000	4800	1100	0.30
I40	4-Aug-20	2	8400	960	140	0.11
I19	8-Sep-20	2	12000	8600	2000	0.72
I19	8-Sep-20	6	8600	2400	3200	0.28
I19	8-Sep-20	11	11000	1000	900	0.09
I24	8-Sep-20	11	6000	1200	540	0.20
I24	8-Sep-20	2	7000	840	500	0.12
I24	8-Sep-20	6	8000	720	620	0.09
I25	8-Sep-20	9	8200	3600	540	0.44
I25	8-Sep-20	6	11000	1800	780	0.16
I25	8-Sep-20	2	9600	980	720	0.10
I26	8-Sep-20	2	12000	4200	820	0.35
I26	8-Sep-20	6	14000	4400	2000	0.31
I26	8-Sep-20	9	16000	4000	900	0.25
I39	8-Sep-20	2	20	460	360	23.00
I40	8-Sep-20	6	9000	2000	1100	0.22
I40	8-Sep-20	2	13000	2400	1400	0.18
I40	8-Sep-20	9	12000	1000	960	0.08
I25	11-Nov-20	9	2200	280	2	0.13
I25	11-Nov-20	2	4000	420	8	0.11
I25	11-Nov-20	6	11000	840	22	0.08
I40	11-Nov-20	2	1200	200	2	0.17

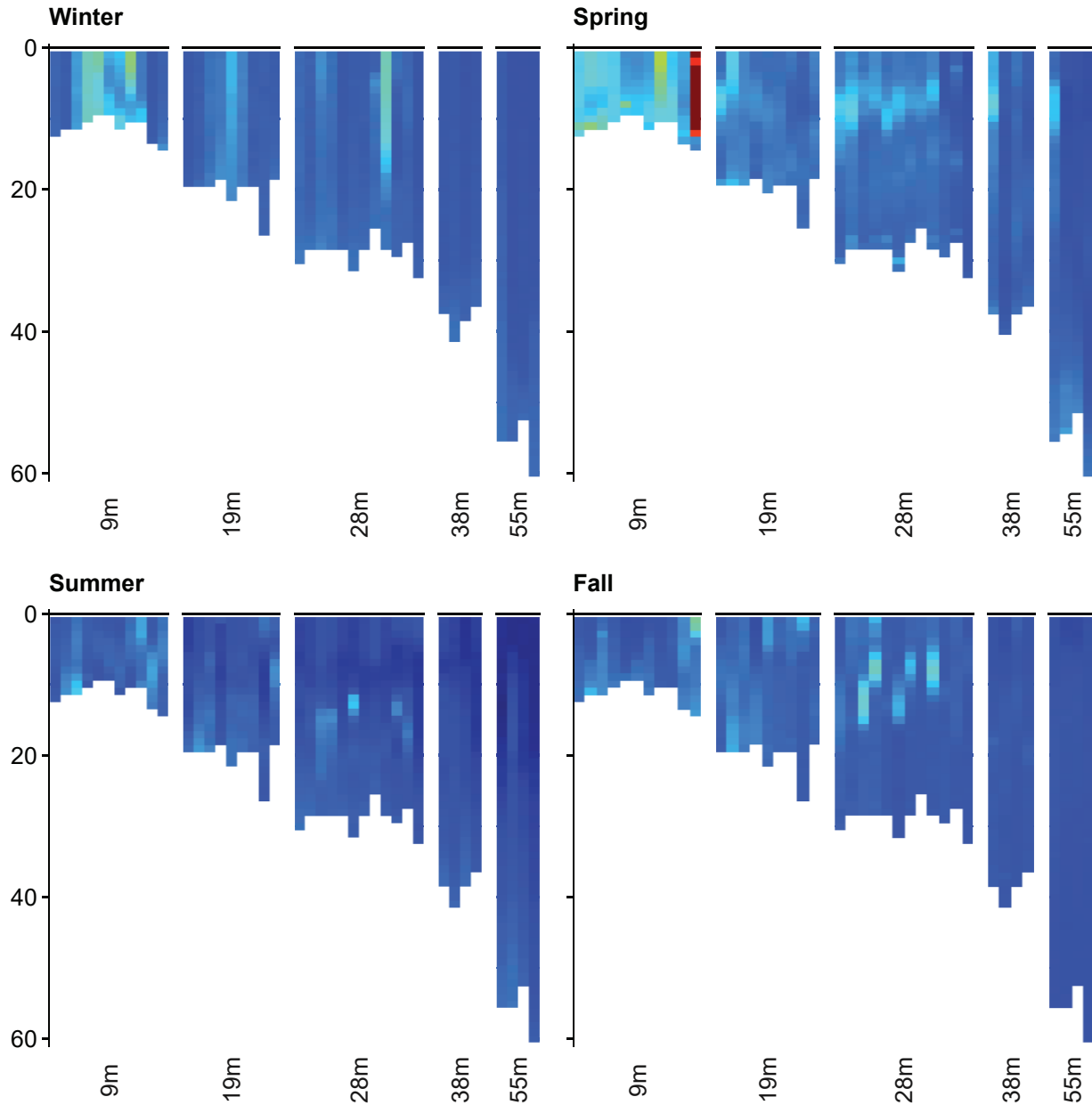
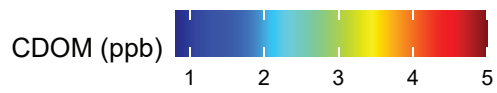
Appendix A.22 *continued*

Station	Date	Depth (m)	Total	Fecal	Entero	F:T
I40	11-Nov-20	6	1600	180	8	0.11
I40	11-Nov-20	9	10000	580	4	0.06
<i>Offshore Stations</i>						
I5	3-Nov-20	2	16000	9600	5400	0.60
I5	3-Nov-20	6	4000	600	120	0.15
I5	3-Nov-20	11	3400	500	140	0.15
I11	3-Nov-20	6	6200	680	280	0.11



Appendix A.23

Concentrations of CDOM recorded in the PLOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.



Appendix A.24

Concentrations of CDOM recorded in the SBOO region during 2020. Within each contour, stations are ordered north to south. Data were collected over four to five days during each survey.

Appendix A.25

Summary of oceanographic data within potential detected plume at PLOO offshore stations and corresponding reference values during 2020. Plume depth is the minimum depth at which CDOM exceeds the 95th percentile while plume width is the number of meters across which that exceedance occurs. Out-of-range values are indicated with an asterisk. DO = dissolved oxygen; XMS = transmissivity; SD = standard deviation; CI = confidence interval.

Station	Date	Potential Plume						Reference ^b			
		Depth (m)	Width (m)	Mean DO	Mean pH	Mean XMS	DO (Mean -SD)	pH (Mean)	XMS (Mean -95% CI)	XMS (Mean -95% CI)	
F27	19-Feb-20	72	21	4.3*	7.7	85	5.5	7.8	77	77	
F28	19-Feb-20	82	16	4.2*	7.7	81*	5.8	7.8	91	91	
F29	19-Feb-20	87	10	4.4*	7.7	84*	5.6	7.8	91	91	
F30	19-Feb-20	59	36	4.3*	7.7	83	5.7	7.9	83	83	
F31	19-Feb-20	71	24	4.5*	7.7	85	5.5	7.8	78	78	
F32	19-Feb-20	78	10	4.5*	7.8	86	5.9	7.9	86	86	
F19 ^a	21-Feb-20	78	4	4.4*	7.7	84	5.8	7.9	79	79	
F21	21-Feb-20	71	12	4.7*	7.8	87	5.4	7.8	70	70	
F30	20-May-20	61	15	3.5	7.8	89	—	—	—	—	
F27	18-Aug-20	84	13	4.3	7.8	92	—	—	—	—	
F28	18-Aug-20	83	15	4.3	7.8	93	—	—	—	—	
F29	18-Aug-20	85	12	4.3	7.8	92	—	—	—	—	
F30	18-Aug-20	72	24	4.4	7.8	90	—	—	—	—	
F31	18-Aug-20	81	16	4.6	7.8	92	—	—	—	—	
F32	18-Aug-20	94	4	4.2	7.8	93	—	—	—	—	
F33	18-Aug-20	97	3	4.0	7.8	93	—	—	—	—	
F34	18-Aug-20	91	7	3.9	7.8	93	—	—	—	—	
F35	18-Aug-20	90	7	3.8	7.8	93	—	—	—	—	
F36	18-Aug-20	86	11	3.7	7.7	93	—	—	—	—	
F26	10-Nov-20	62	12	5.1	7.9	93	5.1	7.9	93	93	
F27	10-Nov-20	55	8	5.0	7.9	93	5.1	7.9	92	92	
F28	10-Nov-20	55	10	5.0	7.9	93	5.2	7.9	92	92	
F29	10-Nov-20	61	27	4.7	7.8	93	5.0	7.9	92	92	
F30	10-Nov-20	64	8	4.8	7.8	92*	5.2	7.9	93	93	
F34	10-Nov-20	64	9	4.7*	7.8	93	5.5	7.9	92	92	

^aStation located within State jurisdictional waters ^bNo reference values possible at plume depths in May or August

Appendix A.26

Summary of oceanographic data within potential detected plume at SBOO offshore stations and corresponding reference values during 2020. Plume depth is the minimum depth at which CDOM exceeds the 95th percentile while plume width is the number of meters across which that exceedance occurs. Out-of-range values are indicated with an asterisk. DO = dissolved oxygen; XMS = transmissivity; SD = standard deviation; CI = confidence interval.

Station	Date	Potential Plume							Reference			
		Depth (m)	Width (m)	Mean DO	Mean pH	Mean XMS	DO (Mean - SD)	pH (Mean)	XMS (Mean - 95% CI)			
I12 ^a	13-Feb-2020	2	16	7.3	8.0	87	7.7	8.1	86			
I23 ^a	13-Feb-2020	2	11	7.5	8.0	78*	7.8	8.1	86			
I12 ^a	28-May-2020	6	1	7.6	8.1	79*	7.5	8.2	82			
I14 ^a	28-May-2020	9	2	7.5	8.1	77*	7.2	8.1	84			
I34 ^a	29-May-2020	6	3	6.2*	8.0	79*	7.5	8.2	83			
I15	24-Aug-2020	12	3	8.4	8.2	85	8.1	8.2	84			
I22 ^a	24-Aug-2020	14	5	8.2	8.2	83*	8.0	8.2	84			
I27 ^a	24-Aug-2020	14	9	8.2	8.2	82*	7.9	8.2	84			
I31 ^a	25-Aug-2020	14	3	8.0	8.1	83*	8.0	8.2	84			
I35 ^a	25-Aug-2020	16	1	7.8	8.1	84	8.1	8.2	84			
I9	26-Aug-2020	13	2	8.0	8.1	89	8.1	8.2	84			
I12 ^a	4-Nov-2020	5	7	7.1	8.1	85	7.6	8.2	83			
I15	4-Nov-2020	11	4	7.3	8.1	85	7.2	8.2	84			
I16 ^a	4-Nov-2020	6	4	6.9*	8.1	86	7.7	8.2	83			
I22 ^a	4-Nov-2020	6	5	7.1	8.1	85	7.6	8.2	83			
I27 ^a	4-Nov-2020	9	7	7.2	8.1	85	7.2	8.2	84			
I35 ^a	5-Nov-2020	15	2	5.6*	8.0	87	7.0	8.1	85			

^aStation located within State jurisdictional waters

Appendix B

Benthic Conditions

2020 Raw Data Summaries

Appendix B.1

Particle size classification schemes (based on Folk 1980) used in the analysis of sediments during 2020. Included is a subset of the Wentworth scale presented as “phi” categories with corresponding Horiba channels, sieve sizes, and size fractions.

Wentworth Scale					
Phi size	Horiba^a		Sieve Size	Sub-Fraction	Fraction
	Min μm	Max μm			
-1	—	—	SIEVE_2000	Granules	Coarse Particles
0	1000	2000	SIEVE_1000	Very coarse sand	Coarse Particles
1	500	1000	SIEVE_500	Coarse sand	Med-Coarse Sands
2	250	500	SIEVE_250	Medium sand	Med-Coarse Sands
3	125	250	SIEVE_125	Fine sand	Fine Sands
3.5	88	125	SIEVE_75	Very fine sand	Fine Sands
4	62.5	88	SIEVE_63	Very fine sand	Fine Sands
5	31	62.5	SIEVE_0 ^b	Coarse silt	Fine Particles ^c
6	15.6	31	—	Medium silt	Fine Particles ^c
7	7.8	15.6	—	Fine silt	Fine Particles ^c
8	3.9	7.8	—	Very fine silt	Fine Particles ^c
9	\leq	3.9	—	Clay	Fine Particles ^c

^aValues correspond to Horiba channels; particles >2000 μm measured by sieve

^bSIEVE_0=sum of all silt and clay, which cannot be distinguished for samples processed by nested sieves

^cFine particles also referred to as percent fines

Appendix B.2

Constituents and method detection limits (MDL) used for the analysis of sediments during 2020. NA=not available.

Parameter	MDL	Parameter	MDL
Organic Indicators			
Biological Oxygen Demand (BOD, ppm)	2	Total Sulfides (ppm)	0.68-1.50
Total Nitrogen (TN, % wt.)	0.008	Total Volatile Solids (TVS, % wt.)	0.11
Total Organic Carbon (TOC, % wt.)	0.07		
Metals (ppm)			
Aluminum (Al)	3.1	Lead (Pb)	0.1
Antimony (Sb)	0.17	Manganese (Mn)	0.061
Arsenic (As)	0.152	Mercury (Hg)	0.003-0.0145
Barium (Ba)	0.155	Nickel (Ni)	0.28
Beryllium (Be)	0.003	Selenium (Se)	0.213
Cadmium (Cd)	0.018	Silver (Ag)	0.133
Chromium (Cr)	0.049	Thallium (Tl)	0.122
Copper (Cu)	1.19	Tin (Sn)	0.059
Iron (Fe)	1.88-9.40	Zinc (Zn)	0.384
Chlorinated Pesticides (ppt)			
<i>Hexachlorocyclohexane (HCH)</i>			
HCH, Alpha isomer (HCH-A)	26.4-110	HCH, Delta isomer (HCH-D)	71.8-147
HCH, Beta isomer (HCH-B)	37.6-76.9	HCH, Gamma isomer (HCH-G)	14.1-147
<i>Total Chlordane</i>			
Alpha (cis) Chlordane (A(c)C)	48.6-95	Heptachlor epoxide (HeptEpox)	33.2-67.9
Cis Nonachlor (cNon)	46.5-95.1	Methoxychlor (Methoxy)	91.1-847
Gamma (trans) Chlordane (G(t)C)	27-126	Oxychlordane (Oxychlor)	70-143
Heptachlor (Hept)	100-344	Trans Nonachlor (tNon)	29.3-125
<i>Total Dichlorodiphenyltrichloroethane (DDT)</i>			
o,p-DDD	36.4-111	p,p-DDE	17.2-138
o,p-DDE	14.1-115	p,p-DDMU	37.5-76.6
o,p-DDT	31.6-96.2	p,p-DDT	21.3-130
p,p-DDD	30.3-69.7		
<i>Miscellaneous Pesticides</i>			
Aldrin	30.4-62.1	Endrin	97.6-200
Alpha Endosulfan	78.2-329	Endrin aldehyde (EndAld)	153-313
Beta Endosulfan	50.9-1020	Hexachlorobenzene (HCB)	50.9-35
Dieldrin	53.5-109	Mirex	27.9-57.1
Endosulfan Sulfate	41.5-308		

Appendix B.2 *continued*

Parameter	MDL	Parameter	MDL
Polychlorinated Biphenyl Congeners (PCBs) (ppt)			
PCB 8	40.9	PCB 126	56.7-116
PCB 18	55.2-113	PCB 128	43.3-77.2
PCB 28	25.6-122	PCB 138	49.3-101
PCB 37	26.1-150	PCB 149	20.6-112
PCB 44	22.3-87.8	PCB 151	38.1-66.2
PCB 49	27.2-85.7	PCB 153/168	87-266
PCB 52	25.4-92.6	PCB 156	29.7-146
PCB 66	21.9-83.5	PCB 157	18.8-174
PCB 70	18.9-87.8	PCB 158	30.5-184
PCB 74	13.2-87.1	PCB 167	17.5-106
PCB 77	28-114	PCB 169	42-85.9
PCB 81	30.1-61.5	PCB 170	35.2-62.7
PCB 87	25.7-83.9	PCB 177	23.2-83.8
PCB 99	19.6-71	PCB 180	38.2-67.9
PCB 101	40.4-82.6	PCB 183	25.9-52.1
PCB 105	37.9-136	PCB 187	32.2-65.8
PCB 110	35.4-104	PCB 189	17-167
PCB 114	50.6-103	PCB 194	21.4-91.7
PCB 118	17.6-97.7	PCB 195	29.2-29.2
PCB 119	29.6-142	PCB 201	42.9-87.7
PCB 123	20.4-134	PCB 206	20.6-97.3
Polycyclic Aromatic Hydrocarbons (PAHs) (ppb)			
1-methylnaphthalene	7.78-14.5	Benzo[G,H,I]perylene	5.12-9.55
1-methylphenanthrene	5.22-9.74	Benzo[K]fluoranthene	4.69-8.76
2,3,5-trimethylnaphthalene	5.62-10.5	Biphenyl	NA
2,6-dimethylnaphthalene	6.74-12.6	Chrysene	4.97-9.28
2-methylnaphthalene	7.25-13.5	Dibenzo(A,H)anthracene	4.77-8.89
3,4-benzo(B)fluoranthene	5.3-9.89	Fluoranthene	6.01-9.37
Acenaphthene	9.72-18.1	Fluorene	9.86-18.4
Acenaphthylene	7.02-13.1	Indeno(1,2,3-CD)pyrene	4.44-8.29
Anthracene	9.61-17.9	Naphthalene	5.9-11
Benzo[A]anthracene	4.88-9.1	Perylene	5.16-9.62
Benzo[A]pyrene	5.58-10.4	Phenanthrene	7.83-14.6
Benzo[e]pyrene	4.49-8.39	Pyrene	6.47-12.1

Appendix B.3

Summary of particle size parameters (%) for each PLOO station sampled during winter 2020. Visual observations are from sieved “grunge” (i.e., particles retained on 1-mm mesh screen and preserved with infauna for benthic community analysis). Gran = Granules; VCS = Very Coarse Sand; CS = Coarse Sand; MS = Medium Sand; FS = Fine Sand; VFS = Very Fine Sand; CFS = Coarse Silt; MSi = Medium Silt; FSi = Fine Silt; VFSi = Very Fine Silt.

	Coarse Particles			Med-Coarse Sands			Fine Sands			Fine Particles			Total Visual Observations		
	Gran	VCS	Total	CS	MS	Total	FS	VFS	Total	MSi	FSi	VFSi		Clay	
<i>88-m Stations</i>															
B11	0.0	0.0	0.0	0.0	0.8	0.8	8.0	19.7	27.7	18.3	28.0	6.7	0.1	71.5 shell hash; gravel; cobble	
B8	0.0	0.0	0.0	0.0	0.1	0.1	4.7	26.3	31.0	27.0	20.6	5.3	0.1	69.0 none	
E19	0.0	0.0	0.0	0.0	0.1	0.1	6.9	33.7	40.6	25.5	16.8	4.2	0.0	59.3 none	
E7	0.0	0.0	0.0	0.0	0.1	0.1	6.5	32.5	39.0	23.9	18.9	4.6	0.0	61.0 none	
E1	0.0	0.0	0.0	0.0	0.7	0.7	9.1	22.8	31.9	19.6	24.7	5.7	0.0	67.4 shell hash	
<i>98-m Stations</i>															
B12	0.0	0.0	0.0	0.0	2.0	2.0	10.4	16.3	26.7	13.6	17.6	31.5	8.6	0.2	71.5 shell hash; gravel; cobble
B9	0.0	0.0	0.0	0.0	0.1	0.1	7.0	26.8	33.8	20.5	15.7	24.0	5.8	0.0	66.0 pea gravel
E26	0.0	0.0	0.0	0.0	0.1	0.1	6.6	30.6	37.2	23.5	15.1	19.8	4.3	0.0	62.7 shell hash
E25	0.0	0.0	0.0	0.0	0.5	0.5	10.1	32.8	42.9	20.3	12.7	18.9	4.6	0.0	56.5 shell hash; gravel
E23	0.0	0.0	0.0	0.0	0.1	0.1	7.4	29.0	36.4	20.7	14.3	22.8	5.8	0.0	63.6 none
E20	0.0	0.0	0.0	0.0	0.2	0.2	9.2	32.4	41.6	19.6	13.1	20.5	5.1	0.0	58.3 none
E17 ^a	0.0	0.0	0.0	0.0	0.2	0.2	10.0	33.7	43.7	20.1	12.9	18.7	4.4	0.0	56.1 none
E14 ^{a,s}	20.7	17.9	38.6	8.8	1.3	10.0	1.0	33.2	34.2	—	—	—	—	17.3	gravel
E11 ^a	0.0	0.0	0.0	0.0	0.6	0.6	12.6	36.6	49.2	17.7	11.1	17.3	4.2	0.0	50.3 black sand; organic debris
E8	0.0	0.0	0.0	0.0	0.8	0.8	13.2	34.6	47.8	17.8	11.8	17.6	4.3	0.0	51.5 none
E5	0.0	0.0	0.0	0.0	0.8	0.8	10.2	26.5	36.7	17.5	14.7	24.2	6.1	0.0	62.5 none
E2 ^s	0.0	0.0	0.0	0.0	1.3	1.3	10.0	25.0	35.0	20.4	16.2	22.0	5.1	0.0	63.7 shell hash; gravel; organic debris
<i>116-m Stations</i>															
B10	0.0	0.0	0.0	0.0	1.2	1.2	15.6	33.5	49.1	16.7	12.1	16.8	4.1	0.0	49.7 shell hash
E21	0.0	0.0	0.0	0.0	0.2	0.2	9.8	32.7	42.5	18.6	12.8	20.6	5.3	0.0	57.3 none
E15 ^a	0.0	0.0	0.0	0.0	0.2	0.2	9.1	28.7	37.8	17.3	14.1	24.1	6.3	0.1	61.9 organic debris
E9	0.0	0.0	0.0	0.0	1.3	1.3	10.3	26.6	36.9	17.9	13.4	23.3	7.0	0.2	61.8 none
E3	0.0	0.0	0.0	0.0	1.5	1.5	9.5	16.0	25.5	15.1	19.8	31.3	6.9	0.0	73.1 shell hash; gravel; organic debris

^aNear-ZID station

^smeasured by sieve (not Horiba; silt and clay fractions are indistinguishable)

Appendix B.3 *continued*

Summary of particle size parameters (%) for each PLOO station sampled during summer 2020. Visual observations are from sieved “grunge” (i.e., particles retained on 1-mm mesh screen and preserved with infauna for benthic community analysis). Gran = Granules; VCS = Very Coarse Sand; CS = Coarse Sand; MS = Medium Sand; FS = Fine Sand; VFS = Very Fine Sand; CSi = Coarse Silt; MSi = Medium Silt; FSi = Fine Silt; VFSi = Very Fine Silt; ns = not sampled.

	Coarse Particles			Med-Coarse Sands			Fine Sands			Fine Particles			Total	Visual Observations		
	Gran	VCS	Total	CS	MS	Total	FS	VFS	Total	CSi	MSi	FSi			VFSi	Clay
<i>88-m Stations</i>																
B11	0.0	0.0	0.0	0.0	0.8	0.8	8.0	20.2	28.2	18.0	17.1	28.0	7.6	0.2	70.9	gravel; shell hash
B8	0.0	0.0	0.0	0.0	0.1	0.1	4.7	26.1	30.8	28.4	17.5	19.3	4.0	0.0	69.2	none
E19	0.0	0.0	0.0	0.0	0.2	0.2	7.4	32.3	39.7	24.4	12.9	18.0	4.9	0.0	60.2	worm tubes; shell hash
E7	0.0	0.0	0.0	0.0	0.2	0.2	7.7	32.1	39.8	24.0	13.8	18.2	4.2	0.0	60.2	shell hash
E1	0.0	0.0	0.0	0.0	1.5	1.5	11.8	26.5	38.3	20.2	14.2	20.5	5.4	0.0	60.3	shell hash
<i>98-m Stations</i>																
B12	0.3	5.9	6.2	14.0	9.5	23.5	10.4	13.9	24.3	10.4	11.2	18.8	5.5	0.1	46.0	black sand; shell hash
B9	0.0	0.0	0.0	0.0	0.7	0.7	8.7	28.3	37.0	20.8	15.0	21.4	5.1	0.0	62.3	pea gravel; shell hash
E26	0.0	0.0	0.0	0.0	0.2	0.2	7.6	30.5	38.1	24.3	15.2	18.4	3.9	0.0	61.8	shell hash
E25	0.0	0.0	0.0	0.0	0.5	0.5	9.6	33.6	43.2	22.0	12.8	17.1	4.3	0.0	56.2	shell hash
E23	0.0	0.0	0.0	0.0	0.6	0.6	10.1	36.5	46.6	21.8	11.0	15.9	4.1	0.0	52.8	shell hash
E20	0.0	0.0	0.0	0.0	0.2	0.2	10.3	38.2	48.5	20.9	11.2	15.5	3.7	0.0	51.3	worm tubes; shell hash
E17 ^a	0.0	0.0	0.0	0.0	0.6	0.6	12.8	38.2	51.0	18.7	10.5	15.4	3.8	0.0	48.4	worm tubes; shell hash
E14 ^a	0.0	0.0	0.0	0.0	1.2	1.2	18.5	44.6	63.1	13.2	7.1	11.8	3.6	0.0	35.7	black sand; shell hash
E11 ^a	0.4	8.9	9.3	4.1	1.1	5.2	11.5	34.4	45.9	16.0	8.3	12.1	3.2	0.0	39.6	none
E8	0.0	0.0	0.0	0.0	0.6	0.6	11.2	33.7	44.9	20.6	12.7	17.1	4.1	0.0	54.5	none
E5	0.0	0.0	0.0	0.0	0.9	0.9	10.8	30.0	40.8	20.5	13.9	19.4	4.5	0.0	58.3	none
E2 ^s	0.0	6.8	6.8	17.0	9.0	26.0	8.5	17.1	25.6	14.3	9.2	13.4	4.4	0.1	41.5	shell hash
<i>116-m Stations</i>																
B10	0.0	0.0	0.0	0.0	0.8	0.8	14.7	35.3	50.0	15.6	11.2	17.8	4.5	0.0	49.1	shell hash
E21	0.0	0.0	0.0	0.0	0.2	0.2	11.7	42.5	54.2	18.8	9.1	14.0	3.7	0.0	45.6	none
E15 ^a	0.0	0.0	0.0	0.0	0.6	0.6	13.1	38.9	52.0	18.4	10.7	14.9	3.4	0.0	47.4	worm tubes; shell hash
E9	0.3	8.1	8.5	9.7	2.8	12.5	7.4	21.6	29.0	16.1	11.7	17.5	4.9	0.0	50.2	shell hash
E3	0.0	0.1	0.1	5.4	9.6	15.1	12.5	16.0	28.5	13.6	15.2	22.4	5.2	0.0	56.4	shell hash

^aNear-ZID station

Appendix B.4

Summary of particle size parameters (%) for each SBOO station sampled during winter 2020. Visual observations are from sieved “grunge” (i.e., particles retained on 1-mm mesh screen and preserved with infauna for benthic community analysis). Gran=Granules; VCS=Very Coarse Sand; CS=Coarse Sand; MS=Medium Sand; FS=Fine Sand; VFS=Very Fine Sand; CSi=Coarse Silt; MSi=Medium Silt; FSi=Fine Silt; VFSi=Very Fine Silt.

	Coarse Particles										Fine Sands										Fine Particles										Total	Visual Observations																									
	Gran					VCS					CS					MS					FS					VFS							CSi					MSi					FSi					VFSi					Clay				
	Total	VCS	Total	CS	MS	Total	MS	CS	FS	VFS	Total	FS	VFS	Total	CSi	MSi	FSi	VFSi	Total	CSi	MSi	FSi	VFSi	Total	CSi	MSi	FSi	VFSi	Total	Clay			Total	Visual Observations																							
19-m Stations	0.0	0.0	0.0	0.0	0.0	2.1	2.1	14.0	27.1	41.1	21.5	15.3	16.7	3.3	0.0	56.8	organic debris																																								
I35	19.3	26.8	46.1	21.5	20.0	41.5	20.0	10.4	0.3	10.7	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1.7	red relict sand; shell hash																									
I31	0.0	0.0	0.0	0.0	0.9	0.9	23.1	61.2	84.3	5.1	1.2	4.7	3.6	0.1	14.7	none																																									
I23	0.0	0.5	0.5	16.5	15.6	32.1	14.1	33.6	47.7	8.7	2.9	5.4	2.7	0.0	19.7	none																																									
I18	0.0	0.0	0.0	0.0	3.1	3.1	26.7	51.6	78.3	7.0	2.4	6.0	3.3	0.0	18.7	none																																									
I10	2.1	14.9	17.0	20.7	6.8	27.5	17.7	23.4	41.1	4.4	3.0	5.4	1.7	0.0	14.5	none																																									
I4	0.0	0.0	0.0	4.0	65.2	69.2	19.3	1.5	20.8	1.0	2.1	4.8	2.0	0.0	9.9	shell hash																																									
28-m Stations	0.0	0.0	0.0	0.0	3.1	3.1	19.7	21.4	41.1	10.5	15.5	24.9	4.9	0.0	55.8	none																																									
I30	0.0	0.0	0.0	0.0	1.6	1.6	18.0	43.4	61.4	12.9	7.9	13.1	3.2	0.0	37.1	none																																									
I27	0.0	0.0	0.0	0.0	0.9	0.9	17.8	60.1	77.9	10.5	1.9	5.0	3.7	0.1	21.2	none																																									
I22	0.0	0.0	0.0	3.8	30.7	34.5	15.3	11.7	27.0	10.7	10.5	13.3	4.0	0.0	38.5	none																																									
I14 ^a	0.0	0.0	0.0	0.0	1.7	1.7	20.6	55.5	76.1	10.9	2.2	5.4	3.5	0.1	22.2	none																																									
I16 ^a	0.0	1.0	1.0	14.9	43.2	58.1	26.6	6.1	32.7	2.0	2.3	3.2	0.7	0.0	8.2	gravel; shell hash																																									
I15	0.0	10.0	10.0	52.1	26.9	79.0	3.6	2.3	5.9	1.4	1.3	1.9	0.6	0.0	5.2	none																																									
I12 ^a	0.0	0.0	0.0	3.5	31.8	35.3	28.6	6.3	34.9	5.2	8.2	13.0	3.4	0.0	29.8	shell hash																																									
I9	0.0	0.0	0.0	0.0	0.9	0.9	16.1	58.6	74.7	13.5	2.4	5.2	3.3	0.1	24.5	none																																									
I6	0.0	0.0	0.0	8.4	71.5	79.9	12.5	1.0	13.5	0.7	1.4	3.1	1.4	0.0	6.6	shell hash																																									
I2	0.0	0.1	0.1	12.4	59.8	72.2	24.4	2.0	26.4	0.1	0.4	0.9	0.1	0.0	1.4	none																																									
I3	0.0	0.0	0.0	0.0	0.1	0.1	2.1	6.0	8.0	10.6	21.8	37.7	17.4	4.5	92.0	shell hash																																									
38-m Stations	0.0	0.0	0.0	0.0	0.8	0.8	12.6	41.7	54.3	20.5	8.9	12.3	3.2	0.0	44.9	red relict sand; gravel																																									
I29	0.0	8.9	8.9	56.4	23.4	79.8	2.2	1.4	3.5	1.4	2.1	3.5	0.9	0.0	7.8	none																																									
I13	0.0	3.9	3.9	43.7	31.8	75.5	2.6	1.4	4.1	1.7	3.9	8.3	2.7	0.0	16.6	shell hash																																									
I8	0.0	0.0	0.0	0.0	0.1	0.1	2.4	5.8	8.2	10.4	25.8	44.9	10.5	0.2	91.8	none																																									
55-m Stations	0.0	0.0	0.0	0.0	0.8	0.8	7.1	11.9	19.0	12.6	19.6	36.0	11.5	0.5	80.2	black gravel; shell hash																																									
I28	0.0	0.0	0.0	0.0	0.0	0.0	1.8	3.0	4.8	4.9	9.8	15.0	2.9	0.0	32.6	red relict sand																																									
I20	0.0	5.3	5.3	47.3	10.1	57.4	0.3	1.4	1.7	4.4	19.5	53.2	20.1	1.1	98.3	red relict sand; shell hash																																									
I7	0.0	0.0	0.0	0.0	0.0	0.0	23.6	29.4	53.0	9.7	11.7	19.1	4.1	0.0	44.6	none																																									
I1	0.0	0.0	0.0	0.0	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	none																																									

^a Near-ZID station

^s measured by sieve (not Horiba; silt and clay fractions are indistinguishable)

Appendix B.4 *continued*

Summary of particle size parameters (%) for each SBOO station sampled during summer 2020. Visual observations are from sieved “grunge” (i.e., particles retained on 1-mm mesh screen and preserved with infauna for benthic community analysis). Gran = Granules; VCS = Very Coarse Sand; CS = Coarse Sand; MS = Medium Sand; FS = Fine Sand; VFS = Very Fine Sand; CSj = Coarse Silt; MSj = Medium Silt; FSi = Fine Silt; VFSj = Very Fine Silt; not sampled.

	Coarse Particles			Med-Coarse Sands			Fine Sands			Fine Particles			Total	Visual Observations		
	Gran	VCS	Total	CS	MS	Total	FS	VFS	Total	CSi	MSi	VFSi			Clay	
19-m Stations	0.0	0.0	0.0	0.0	1.5	1.5	15.3	40.4	55.7	21.8	9.0	9.6	2.4	0.0	42.8	organic debris
I34 ^s	11.7	16.7	28.4	29.0	31.1	60.1	9.0	0.3	9.3	—	—	—	—	—	2.2	red relict sand; shell hash
I31	0.0	0.0	0.0	0.0	0.6	0.6	19.2	68.8	88.0	5.0	0.2	2.8	3.3	0.2	11.4	none
I23	0.0	0.0	0.0	0.0	2.1	2.1	22.2	50.9	73.1	9.2	3.3	8.0	4.2	0.1	24.8	none
I18	0.0	0.0	0.0	0.0	0.9	0.9	24.2	65.0	89.2	4.3	0.2	2.4	2.8	0.2	9.9	shell hash
I10	0.0	0.6	0.6	8.9	8.4	17.3	18.7	38.1	56.8	9.3	4.0	8.0	3.8	0.1	25.2	worm tubes
I4	0.0	3.3	3.3	50.4	41.9	92.3	4.1	0.4	4.4	0.0	0.0	0.0	0.0	0.0	0.0	shell hash
28-m Stations	0.0	0.0	0.0	0.0	2.7	2.7	30.4	37.9	68.3	7.2	5.2	11.7	4.9	0.1	29.1	worm tubes
I30	0.0	0.0	0.0	0.0	0.8	0.8	15.6	58.4	74.0	14.1	2.9	5.3	2.8	0.1	25.2	none
I27	0.0	0.0	0.0	0.0	0.6	0.6	15.0	61.2	76.2	12.7	2.1	4.9	3.4	0.1	23.2	none
I22	0.0	0.4	0.4	10.1	10.7	20.8	3.3	3.7	7.0	6.9	19.2	36.1	9.3	0.2	71.7	none
I14 ^a	0.0	0.0	0.0	0.0	1.0	1.0	17.3	60.8	78.1	11.1	1.7	4.6	3.4	0.1	21.0	none
I16 ^a	0.0	0.1	0.1	8.2	33.7	41.9	40.9	11.2	52.1	1.8	1.5	2.1	0.6	0.0	6.0	shell hash
I15	0.0	2.3	2.3	24.1	47.3	71.4	16.4	4.3	20.7	1.7	1.4	2.0	0.5	0.0	5.5	none
I12 ^a	0.0	0.0	0.0	1.3	13.8	15.1	30.7	36.8	67.5	7.9	2.6	4.7	2.2	0.0	17.4	none
I9	0.0	0.0	0.0	0.0	1.1	1.1	16.5	58.5	75.0	13.9	2.3	4.7	2.8	0.1	23.8	organic debris
I6	0.0	4.3	4.3	51.5	38.9	90.4	4.8	0.4	5.2	0.0	0.0	0.0	0.0	0.0	0.0	shell hash
I2	0.0	0.0	0.0	0.1	6.2	6.3	6.1	6.6	12.8	10.0	24.7	38.7	7.5	0.0	80.9	none
I3	0.0	0.0	0.0	0.0	0.0	0.0	0.9	5.2	6.1	12.5	28.9	43.0	9.3	0.2	93.8	red relict sand; shell hash
38-m Stations	0.0	3.0	3.0	5.0	5.0	9.9	13.0	28.9	41.9	16.3	10.0	14.9	3.9	0.0	45.1	organic debris
I21	0.0	8.1	8.1	59.4	28.2	87.6	2.6	0.2	2.8	0.0	0.5	0.9	0.1	0.0	1.6	shell hash
I13	0.0	2.6	2.6	15.3	10.7	26.0	5.7	5.5	11.2	7.2	16.5	29.4	7.0	0.1	60.2	shell hash
I8	0.0	2.6	2.6	35.9	51.1	87.0	8.3	0.8	9.1	0.0	0.4	0.8	0.1	0.0	1.3	none
55-m Stations	2.2	14.4	16.6	30.7	4.6	35.3	4.6	9.7	14.3	7.7	7.8	13.5	4.7	0.1	33.8	black gravel
I20	0.0	7.8	7.8	39.6	8.9	48.5	2.4	3.7	6.0	5.5	11.3	17.5	3.3	0.0	37.6	red relict sand
I7	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	1.3	23.0	61.0	14.6	0.2	100.1	red relict sand; shell hash
I1	0.0	0.0	0.0	0.0	6.2	6.2	41.9	29.0	70.9	5.8	4.7	9.0	3.4	0.0	22.9	none

^aNear-ZID station

^smeasured by sieve (not Horiba; silt and clay fractions are indistinguishable)

Appendix B.5

Summary of particle size parameters (%) for sediments from the San Diego regional benthic stations sampled during summer 2020. Visual observations are from sieved "grunge" (i.e., particles retained on 1-mm mesh screen and preserved with infauna for benthic community analysis). Gran = Granules; VCS = Very Coarse Sand; CS = Coarse Sand; MS = Medium Sand; FS = Fine Sand; VFS = Very Fine Sand; CSI = Coarse Silt; MSI = Medium Silt; VFSi = Fine Silt; VFSi = Very Fine Silt.

Station	Depth (m)	Coarse Particles			Med-Coarse Sands			Fine Sands			Fine Particles					Total	Visual Observations
		Gran	VCS	Total	CS	MS	Total	FS	VFS	Total	CSI	MSI	VFSi	Clay			
<i>Inner Shelf</i>																	
8929	7	0.0	0.0	0.0	1.2	14.4	15.6	44.8	29.8	74.6	3.9	1.8	2.9	1.2	0.0	9.8	none
8944	7	0.0	2.7	2.7	5.9	9.0	15.0	33.7	38.5	72.2	4.0	1.2	3.1	1.8	0.0	10.2	none
8918	8	0.0	0.0	0.0	2.5	12.1	14.6	11.5	10.1	21.6	9.2	17.0	30.9	6.7	0.0	63.8	shell hash
8946	15	0.0	0.0	0.0	0.5	12.4	12.9	38.7	16.9	55.6	6.7	8.7	12.5	3.6	0.0	31.5	none
8915	16	1.5	4.7	6.3	8.6	15.0	23.6	40.1	17.9	58.0	2.3	2.4	5.3	2.2	0.0	12.2	gravel; shell hash
8907	18	0.0	0.0	0.0	0.0	1.3	1.3	23.5	60.7	84.2	5.5	1.1	4.2	3.6	0.1	14.5	none
8943	19	0.0	0.0	0.0	0.0	0.7	0.7	19.0	52.3	71.3	9.0	3.6	10.0	5.3	0.1	28.0	none
8914	20	0.0	0.0	0.0	0.0	1.7	1.7	15.7	38.1	53.8	21.9	10.0	10.1	2.5	0.0	44.5	none
8911	24	0.0	0.0	0.0	0.0	0.2	0.2	12.8	60.2	73.0	14.3	3.8	6.5	2.3	0.0	26.8	none
8938	28	0.0	0.0	0.0	0.0	5.8	5.8	32.1	25.6	57.7	8.3	8.9	14.4	4.8	0.1	36.5	organic debris
8901	29	0.0	0.0	0.0	0.1	7.8	7.8	28.5	40.9	69.4	10.4	3.3	6.0	3.1	0.1	22.9	none
<i>Mid Shelf</i>																	
8913	31	0.0	0.0	0.0	0.0	1.5	1.5	29.5	48.7	78.2	6.2	2.6	7.3	4.1	0.1	20.2	none
8906	41	0.0	4.3	4.3	33.8	43.2	77.0	9.3	2.1	11.4	1.4	2.0	3.1	0.8	0.0	7.4	none
8939	65	0.0	0.0	0.0	0.0	2.1	2.1	17.1	29.8	46.9	14.5	12.4	19.4	4.7	0.0	51.0	none
8903	66	0.0	13.0	13.0	45.7	7.7	53.4	8.0	7.9	15.9	4.4	4.2	7.0	2.3	0.0	17.8	none
8912	67	0.0	0.0	0.0	0.0	0.7	0.7	9.9	31.5	41.4	21.4	13.7	18.6	4.3	0.0	58.0	organic debris
8925	67	0.0	0.0	0.0	0.0	0.1	0.1	5.0	28.4	33.4	25.3	16.1	20.6	4.6	0.0	66.6	none
8919	68	0.0	0.0	0.0	0.0	0.2	0.2	8.1	30.9	39.0	23.9	14.8	18.2	4.0	0.0	60.9	gravel; shell hash
8909	73	0.0	0.0	0.0	0.0	3.6	3.6	16.4	25.3	41.7	15.8	13.0	20.7	5.2	0.0	54.7	shell hash
8931	74	0.0	0.0	0.0	0.0	0.1	0.1	5.3	28.8	34.1	28.3	16.9	17.4	3.2	0.0	65.8	none
8933	80	0.0	0.0	0.0	0.0	0.1	0.1	4.9	26.4	31.3	27.7	17.6	19.3	3.9	0.0	68.5	none
8930	83	0.0	0.0	0.0	0.0	0.1	0.1	4.2	22.5	26.7	26.8	19.7	22.4	4.4	0.0	73.3	none
8949	83	0.0	0.0	0.0	0.0	0.1	0.1	4.5	24.3	28.8	25.7	17.9	22.5	5.0	0.0	71.1	none
8924	85	0.0	0.0	0.0	0.0	0.1	0.1	5.0	30.2	35.2	26.6	14.3	19.1	4.7	0.0	64.7	organic debris
8926	87	0.0	0.0	0.0	0.0	0.2	0.2	7.7	35.3	43.0	28.5	12.7	12.9	2.8	0.0	56.9	none
8932	89	0.0	0.0	0.0	0.0	0.1	0.1	5.3	25.9	31.2	24.6	16.7	22.3	5.2	0.0	68.8	gravel; organic debris
8940	108	2.9	11.3	14.2	29.8	4.8	34.6	5.0	14.0	19.0	9.4	7.5	11.9	3.4	0.0	32.2	gravel
8921	116	0.0	0.0	0.0	0.0	0.2	0.2	11.3	40.1	51.4	18.6	9.9	15.8	4.2	0.0	48.5	none

Appendix B.5 *continued*

Station	Depth (m)	Coarse Particles			Med-Coarse Sands			Fine Sands			Fine Particles					Visual Observations	
		Gran	VCS	Total	CS	MS	Total	FS	VFS	Total	CSI	MSi	FSi	VFSi	Clay		Total
<i>Outer Shelf</i>																	
8905	134	0.0	0.0	0.0	0.0	0.2	0.2	9.0	31.5	40.5	19.3	13.2	21.3	5.6	0.0	59.4	shell hash
8937	137	0.0	0.0	0.0	0.0	0.6	0.6	9.3	34.2	43.5	22.0	12.3	17.3	4.4	0.0	56.0	cobble; worm tubes
8910	169	0.0	0.0	0.0	0.0	5.8	5.8	17.5	21.1	38.6	14.0	12.5	21.5	7.4	0.3	55.7	organic debris
8922	185	0.0	0.0	0.0	0.0	0.1	0.1	4.9	28.3	33.2	26.8	15.7	19.8	4.5	0.0	66.8	organic debris; worm tubes
8916	189	0.0	0.0	0.0	0.0	0.1	0.1	5.2	21.3	26.5	22.8	19.7	25.3	5.6	0.0	73.4	organic debris; worm tubes
8908	194	0.0	0.0	0.0	0.0	0.2	0.2	6.4	23.8	30.2	21.3	17.6	25.0	5.8	0.0	69.7	organic debris; shell hash
<i>Upper Slope</i>																	
8928	223	0.0	0.0	0.0	0.0	0.1	0.1	4.3	24.2	28.5	26.9	18.7	21.4	4.4	0.0	71.4	organic debris; shell hash
8920	270	0.0	0.0	0.0	0.0	0.1	0.1	4.0	19.0	23.0	24.8	22.3	25.1	4.8	0.0	77.0	none
8936	292	0.0	0.0	0.0	0.0	0.1	0.1	5.7	23.2	28.9	21.2	18.0	25.8	5.9	0.0	70.9	none
8927	460	0.0	0.0	0.0	0.0	0.1	0.1	3.8	13.1	16.9	17.4	24.2	34.6	6.8	0.0	83.0	none
8934	491	0.0	0.0	0.0	0.0	0.1	0.1	2.7	9.4	12.1	15.2	26.6	38.5	7.4	0.0	87.7	none
8923	523	0.0	0.0	0.0	0.0	0.5	0.5	6.8	15.0	21.8	12.8	19.6	36.7	8.5	0.1	77.8	none

Appendix B.6

Concentrations of organic loading indicators detected in sediments from PLOO stations sampled during winter and summer 2020. See Appendix B.1 for MDLs; ND=not detected; NR=not reportable.

	Winter					Summer				
	BOD ^b (ppm)	Sulfides (ppm)	TN (% wt)	TOC (% wt)	TVS (% wt)	BOD (ppm)	Sulfides (ppm)	TN (% wt)	TOC (% wt)	TVS (% wt)
<i>88-m Depth Contour</i>										
B11	—	ND	0.113	2.27	NR	—	18.40	0.092	2.11	3.7
B8	—	2.67	0.078	0.80	NR	—	12.10	0.082	0.81	3.0
E19	—	2.87	0.057	0.59	NR	—	3.04	0.057	0.56	2.1
E7	—	1.85	0.052	0.51	NR	—	5.82	0.052	0.48	2.1
E1	—	ND	0.049	0.55	NR	—	1.80	0.054	0.51	2.1
<i>98-m Depth Contour</i>										
B12	426	5.58	0.059	3.49	NR	546	3.88	0.053	2.86	2.7
B9	240	5.41	0.063	0.83	2.5	297	2.37	0.058	0.81	2.7
E26	187	4.33	0.053	0.51	2.1	255	10.20	0.052	0.55	2.3
E25	202	4.98	0.044	0.53	1.9	251	ND	0.041	0.41	1.9
E23	204	1.71	0.052	0.53	1.9	254	ND	0.047	0.46	2.1
E20	204	1.98	0.043	0.43	1.8	233	1.65	0.045	0.43	1.8
E17 ^a	212	3.62	0.042	0.40	1.6	271	7.08	0.034	0.28	1.5
E14 ^a	371	5.27	0.032	0.33	1.2	200	30.80	0.035	0.34	1.4
E11 ^a	213	1.70	0.039	0.43	1.9	314	1.64	0.041	0.46	1.9
E8	214	NA	0.034	0.39	1.7	216	3.42	0.041	0.46	2.0
E5	191	0.87	0.045	0.53	2.0	208	1.83	0.049	0.58	2.3
E2	396	3.00	0.062	0.76	NR	208	3.20	0.050	0.66	2.1
<i>116-m Depth Contour</i>										
B10	—	2.62	0.051	1.01	NR	—	10.90	0.048	0.99	2.4
E21	—	1.76	0.040	0.42	NR	—	7.49	0.043	0.43	1.7
E15 ^a	—	3.90	0.037	0.45	NR	—	ND	0.044	0.49	1.8
E9	—	3.05	0.050	1.33	NR	—	6.30	0.051	1.19	2.3
E3	—	0.91	0.052	0.63	NR	—	ND	0.038	0.36	1.6
Detection Rate (%)	100	90	100	100	100	100	82	100	100	100

^aNear-ZID station

^bBOD only sampled at PLOO primary core stations

Appendix B.7

Concentrations of organic indicators detected in sediments from SBOO stations sampled during winter and summer 2020. See Appendix B.1 for MDLs; ND = not detected.

	Winter				Summer			
	Sulfides (ppm)	TN (% wt)	TOC (% wt)	TVS (% wt)	Sulfides (ppm)	TN (% wt)	TOC (% wt)	TVS (% wt)
<i>19-m Stations</i>								
I35	9.78	0.029	0.25	NR	4.25	0.032	0.28	1.3
I34	0.74	ND	1.15	NR	ND	ND	0.86	0.6
I31	1.66	ND	0.09	NR	8.15	0.020	0.14	0.8
I23	2.52	ND	0.10	NR	ND	0.022	0.14	0.9
I18	0.69	ND	0.08	NR	2.22	ND	0.11	0.8
I10	1.82	ND	0.09	NR	5.91	ND	0.11	0.8
I4	ND	ND	0.27	NR	ND	ND	0.09	0.4
<i>28-m Stations</i>								
I33	1.09	ND	0.25	NR	14.70	0.035	0.43	1.5
I30	1.78	0.024	0.18	NR	ND	0.027	0.20	1.3
I27	3.44	0.010	0.15	NR	1.57	0.026	0.19	1.0
I22	ND	ND	0.11	NR	5.07	0.023	0.16	0.9
I14 ^a	5.06	ND	0.18	NR	2.74	0.027	0.20	1.0
I16 ^a	ND	ND	ND	NR	ND	ND	0.10	0.7
I15	ND	ND	ND	NR	ND	ND	0.11	0.6
I12 ^a	ND	ND	ND	NR	28.90	0.026	0.20	1.0
I9	0.94	0.019	0.19	NR	8.72	0.026	0.19	1.5
I6	ND	0.021	0.17	NR	ND	ND	0.08	0.5
I2	ND	ND	ND	NR	ND	ND	0.08	0.5
I3	ND	ND	ND	NR	ND	ND	ND	0.4
<i>38-m Stations</i>								
I29	1.86	0.028	0.31	NR	18.10	0.041	0.44	1.7
I21	ND	ND	0.12	NR	ND	0.019	0.14	0.6
I13	ND	ND	ND	NR	ND	ND	ND	0.5
I8	ND	ND	0.12	NR	ND	ND	0.09	0.5
<i>55-m Stations</i>								
I28	3.31	0.052	0.86	NR	1.61	0.062	0.96	1.5
I20	ND	ND	0.17	NR	ND	0.023	0.15	0.5
I7	ND	ND	0.08	NR	ND	ND	ND	0.5
I1	ND	0.009	0.13	NR	ND	0.026	0.18	1.0
Detection Rate (%)	48	30	78	NR	44	56	89	100

^aNear-ZID station

Appendix B.8

Concentrations of organic indicators detected in sediments from the San Diego regional benthic stations sampled during summer 2020. See Appendix B.1 for MDLs; ND=not detected.

	Station	Depth (m)	Sulfides (ppm)	TN (% wt)	TOC (% wt)	TVS (% wt)
Inner Shelf	8929	7	6.39	ND	0.30	0.9
	8944	7	8.97	0.027	0.20	1.0
	8918	8	ND	ND	0.07	0.5
	8946	15	16.40	ND	ND	0.8
	8915	16	1.58	ND	0.09	0.8
	8907	18	29.00	ND	0.10	0.6
	8943	19	7.21	ND	0.11	0.6
	8914	20	28.90	0.034	0.27	1.7
	8911	24	30.30	0.022	0.17	1.0
	8938	28	7.02	0.022	0.13	0.7
	8901	29	28.30	0.026	0.19	1.2
Mid Shelf	8913	31	38.40	0.026	0.20	1.2
	8906	41	ND	0.018	0.15	0.6
	8939	65	37.10	0.039	0.32	1.6
	8903	66	8.07	0.037	0.34	1.1
	8912	67	10.90	0.058	0.66	2.6
	8925	67	4.20	0.073	0.80	3.1
	8919	68	1.68	0.067	0.70	2.8
	8909	73	2.95	0.055	0.56	2.1
	8931	74	1.59	0.080	0.79	2.9
	8933	80	1.59	0.080	0.77	2.9
	8930	83	17.60	0.086	0.89	3.2
	8949	83	6.73	0.072	0.75	3.1
	8924	85	8.43	0.068	0.69	2.8
	8926	87	2.66	0.050	0.51	2.2
	8932	89	5.63	0.078	0.83	3.2
8940	108	8.16	0.057	0.96	1.3	
8921	116	1.75	0.038	0.40	1.7	
Outer Shelf	8905	134	3.41	0.060	0.95	2.8
	8937	137	19.40	0.056	1.21	2.2
	8910	169	3.46	0.051	0.60	2.5
	8922	185	7.74	0.088	1.23	3.9
	8916	189	38.90	0.120	1.74	5.3
	8908	194	8.12	0.136	2.38	5.6
Upper Slope	8928	223	15.70	0.133	1.78	5.2
	8920	270	42.90	0.187	2.64	7.1
	8936	292	20.50	0.137	1.86	5.5
	8927	460	18.00	0.211	2.78	8.7
	8934	491	12.10	0.270	3.33	9.5
	8923	523	3.75	0.196	3.39	7.6
Detection Rate (%)			95	85	98	100

Appendix B.9

Concentrations of trace metals (ppm) detected in sediments from PLOO stations sampled during winter 2020. See Appendix B.1 for MDLs and translation of periodic table symbols; DR = detection rate; ND = not detected.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Ti	Sn	Zn
<i>88-m Depth Contour</i>																		
B11	9360	1.4	4.78	49.7	0.28	ND	22.3	9.1	18000	4.1	134.0	0.032	7.8	0.54	ND	ND	1.0	39.1
B8	10700	1.4	3.58	39.5	0.24	ND	21.0	8.8	13800	4.7	114.0	0.035	9.1	0.33	ND	ND	1.2	36.4
E19	8910	1.1	2.61	35.9	0.19	ND	16.6	6.7	11100	3.0	96.7	0.022	7.2	0.34	ND	ND	0.9	27.3
E7	7910	1.0	2.94	29.6	0.17	ND	15.4	6.9	10200	3.6	87.6	0.027	6.5	ND	ND	ND	1.0	25.5
E1	7330	1.1	3.07	34.6	0.17	ND	14.6	17.0	10400	4.4	83.3	0.042	6.2	ND	ND	ND	1.0	26.2
<i>98-m Depth Contour</i>																		
B12	5540	1.2	5.72	16.4	0.28	ND	23.9	7.1	18900	2.5	133.0	0.014	5.3	ND	ND	ND	0.7	32.8
B9	7170	1.2	3.28	35.3	0.21	ND	18.6	6.1	12700	3.4	85.9	0.024	7.1	ND	ND	ND	0.8	29.7
E26	7790	1.1	2.84	27.7	0.18	ND	15.5	6.2	10300	3.6	85.4	0.025	6.6	0.28	ND	ND	1.0	25.3
E25	7000	1.0	2.14	26.4	0.16	ND	13.9	5.8	9280	3.0	79.0	0.019	5.7	0.31	ND	ND	0.8	23.7
E23	7310	1.1	2.92	29.7	0.17	ND	15.0	6.4	9840	3.3	84.8	0.023	6.6	ND	ND	ND	0.9	24.0
E20	6370	1.0	2.77	25.6	0.15	ND	13.4	5.3	8770	3.0	74.6	0.019	5.8	ND	ND	ND	0.7	20.9
E17 ^a	6130	1.0	2.70	23.9	0.14	ND	13.1	5.2	8500	2.8	71.4	0.016	5.5	ND	ND	ND	0.7	20.5
E14 ^a	4930	0.9	3.21	21.7	0.12	ND	14.6	5.0	7540	2.3	85.4	0.012	10.0	ND	ND	ND	0.6	18.4
E11 ^a	5490	0.8	2.71	20.5	0.13	ND	12.0	4.8	7850	2.5	64.5	0.016	5.0	ND	ND	ND	0.7	18.9
E8	5780	0.8	1.85	20.7	0.14	ND	12.4	4.8	7980	2.6	65.7	0.016	4.8	0.29	ND	ND	0.6	19.9
E5	6300	0.8	2.36	22.9	0.15	ND	12.7	5.4	8720	2.8	69.0	0.020	5.1	0.27	ND	ND	0.7	21.5
E2	8850	1.3	3.58	39.3	0.20	ND	17.2	9.1	12500	4.1	96.8	0.040	7.2	ND	ND	ND	1.1	30.0
<i>116-m Depth Contour</i>																		
B10	6190	0.9	2.39	20.2	0.19	ND	16.6	5.0	11200	2.6	64.9	0.016	4.9	0.28	ND	ND	0.6	26.2
E21	5300	0.9	2.27	16.8	0.13	ND	12.1	4.6	7680	2.7	61.5	0.009	4.9	ND	ND	ND	0.6	18.8
E15 ^a	5520	0.8	2.40	17.7	0.14	ND	12.2	4.7	7870	2.5	61.2	0.014	4.5	0.24	ND	ND	0.6	19.0
E9	5370	2.9	2.42	17.3	0.15	ND	15.0	7.8	10200	3.0	60.3	0.008	4.7	ND	ND	ND	0.7	32.5
E3	7350	1.2	2.28	42.4	0.15	ND	12.6	9.0	10700	5.3	86.9	0.057	5.1	ND	ND	ND	0.9	24.9
DR (%)	100	100	100	100	100	0	100	100	100	100	100	100	100	41	0	0	100	100
ERL ^b	—	—	8.2	—	—	1.2	81	34	—	46.7	—	0.15	20.9	—	1.0	—	—	150
ERM ^b	—	—	70.0	—	—	9.6	370	270	—	218.0	—	0.71	51.6	—	3.7	—	—	410

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.9 *continued*

Concentrations of trace metals (ppm) detected in sediments from PLOO stations sampled during summer 2020. See Appendix B.1 for MDLs and translation of periodic table symbols; DR = detection rate; nd = not detected.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Ti	Sn	Zn
<i>88-m Depth Contour</i>																		
B11	8960	1.3	3.68	33.8	0.28	0.08	20.9	8.8	16100	3.8	138.0	0.035	7.5	ND	ND	ND	0.9	35.8
B8	11000	1.6	3.92	43.3	0.24	0.07	21.1	9.2	13700	4.6	113.0	0.044	9.2	ND	ND	ND	1.1	34.6
E19	8300	1.3	3.11	34.1	0.18	0.07	15.9	6.8	10700	3.6	95.2	0.030	6.8	ND	ND	ND	1.0	25.9
E7	7660	1.1	2.77	30.5	0.16	0.07	14.7	6.5	9760	3.4	88.8	0.029	6.3	ND	ND	ND	0.9	23.8
E1	7230	1.0	2.89	28.8	0.15	0.06	13.7	7.0	9720	4.7	87.7	0.047	5.6	ND	ND	ND	1.5	25.1
<i>98-m Depth Contour</i>																		
B12	5940	1.3	5.20	13.1	0.28	0.07	23.4	6.0	18600	2.7	51.3	0.029	5.1	ND	ND	ND	0.6	32.5
B9	8220	1.5	3.48	54.1	0.24	0.05	19.8	6.3	13900	3.2	90.2	0.026	7.0	ND	ND	ND	0.7	31.4
E26	7680	1.3	2.69	29.0	0.17	0.06	14.8	5.9	9750	3.2	84.4	0.037	6.6	ND	ND	ND	0.7	24.5
E25	6280	1.0	2.53	23.5	0.14	0.07	12.7	4.8	8370	2.6	71.6	0.019	5.4	ND	ND	ND	0.5	20.3
E23	6750	1.0	2.74	26.7	0.15	0.05	13.7	5.5	9000	3.0	76.1	0.028	5.8	ND	ND	ND	0.7	22.5
E20	6010	0.9	2.50	21.5	0.14	0.07	12.4	4.9	8110	2.9	68.7	0.020	5.2	ND	ND	ND	0.6	20.0
E17 ^a	5480	0.8	2.47	19.6	0.13	0.06	11.4	5.0	7650	2.5	65.3	0.016	4.7	ND	ND	ND	0.6	18.4
E14 ^a	4440	0.8	2.08	16.3	0.11	0.08	10.0	4.2	6300	1.9	57.5	0.014	4.0	ND	ND	ND	0.6	16.8
E11 ^a	5710	1.2	2.48	18.8	0.13	0.07	11.7	4.7	7780	2.3	66.1	0.020	4.7	ND	ND	ND	0.7	18.6
E8	6330	1.7	2.13	22.8	0.15	0.06	12.8	5.1	8480	2.6	73.8	0.021	5.2	ND	ND	ND	0.8	20.3
E5	6490	1.1	2.94	25.6	0.16	0.08	13.4	6.3	9250	3.4	73.1	0.027	5.4	ND	ND	ND	1.1	23.0
E2	8060	1.1	2.92	33.4	0.17	0.04	14.5	7.7	11400	3.6	94.6	0.034	5.7	ND	ND	ND	1.4	27.4
<i>116-m Depth Contour</i>																		
B10	5700	1.0	2.37	19.5	0.17	0.08	14.7	4.8	9680	2.5	60.9	0.020	4.7	ND	ND	ND	0.5	22.6
E21	5430	0.9	2.29	19.2	0.13	0.07	11.6	4.5	7470	2.6	61.3	0.020	4.9	ND	ND	ND	0.5	18.8
E15 ^a	5450	0.8	2.20	17.7	0.13	0.07	11.8	4.4	7650	2.4	63.3	0.019	4.4	ND	ND	ND	0.6	18.6
E9	5490	0.9	3.41	17.8	0.15	0.07	15.2	7.5	9760	3.0	59.7	0.020	4.5	ND	ND	ND	0.8	34.8
E3	6070	1.2	1.79	33.5	0.13	0.04	12.3	12.8	9090	7.5	81.1	0.040	3.8	ND	ND	ND	0.9	52.5
DR (%)	100	100	100	100	100	100	100	100	100	100	100	100	100	0	0	0	100	100
ERL ^b	—	—	8.2	—	—	1.2	81	34	—	46.7	—	0.15	20.9	—	1.0	—	—	150
ERM ^b	—	—	70.0	—	—	9.6	370	270	—	218.0	—	0.71	51.6	—	3.7	—	—	410

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.10

Concentrations of trace metals (ppm) detected in sediments from SBOO stations sampled during winter 2020. See Appendix B.1 for MDLs and translation of periodic table symbols; nd = not detected.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Ti	Sn	Zn
<i>19-m Stations</i>																		
I35	7110	1.3	1.83	38.9	0.13	ND	13.4	4.9	8820	2.9	96.9	0.016	4.5	ND	ND	ND	0.8	25.4
I34	1110	0.2	2.76	5.4	0.02	ND	2.8	2.4	2620	1.1	32.0	ND	0.6	ND	ND	ND	0.2	4.8
I31	3000	0.6	0.99	20.1	0.06	ND	7.0	1.4	3370	1.3	47.8	ND	1.5	ND	ND	ND	0.3	8.1
I23	4080	0.4	1.72	19.4	0.08	ND	8.3	ND	4710	1.5	53.2	ND	2.2	ND	ND	ND	0.3	10.9
I18	4200	0.4	1.29	34.9	0.09	ND	10.6	ND	6050	1.5	67.9	ND	2.2	ND	ND	ND	0.4	11.9
I10	4150	0.5	1.55	24.5	0.08	ND	9.8	ND	5660	1.5	61.6	ND	2.2	ND	ND	ND	0.4	11.5
I4	918	0.6	1.22	3.2	0.03	ND	5.2	ND	2650	1.6	42.3	ND	0.7	ND	ND	ND	0.3	3.4
<i>28-m Stations</i>																		
I33	4190	0.7	2.19	19.0	0.08	ND	8.0	2.8	5570	2.6	64.4	0.011	2.4	ND	ND	ND	0.6	14.2
I30	5840	0.9	1.93	27.1	0.10	ND	10.9	3.3	6260	1.8	62.5	0.006	3.4	ND	ND	ND	0.5	16.6
I27	6080	0.9	1.47	29.0	0.10	ND	10.3	1.5	6100	1.5	66.7	0.004	3.2	ND	ND	ND	0.5	15.8
I22	1620	0.3	1.48	4.5	0.05	ND	5.0	ND	2510	1.3	18.9	ND	0.9	ND	ND	ND	0.2	5.2
I14 ^a	6100	0.9	1.37	30.6	0.11	ND	10.6	1.4	6320	1.4	72.1	ND	3.3	ND	ND	ND	0.5	16.2
I16 ^a	2430	0.3	1.41	9.2	0.06	ND	6.3	ND	3990	1.3	40.6	ND	1.2	ND	ND	ND	0.2	9.2
I15	1760	0.4	2.39	5.5	0.06	ND	8.2	ND	4200	1.5	21.8	ND	0.9	ND	ND	ND	0.2	7.9
I12 ^a	1970	0.3	1.55	9.8	0.05	ND	6.0	ND	3400	1.2	27.9	ND	1.0	ND	ND	ND	0.2	7.0
I9	7140	1.0	1.53	36.5	0.12	ND	11.9	1.7	7330	1.3	79.4	0.004	4.0	ND	ND	ND	0.5	19.3
I6	1320	0.5	5.55	3.4	0.05	ND	8.3	ND	4130	1.6	12.4	ND	1.1	ND	ND	ND	0.2	4.4
I2	1090	0.2	0.84	2.4	0.03	ND	5.5	ND	1310	0.9	10.9	ND	0.7	ND	ND	ND	0.1	2.5
I3	695	0.3	1.09	1.2	0.02	ND	5.2	ND	1220	0.8	6.1	ND	0.5	ND	ND	ND	0.1	1.7
<i>38-m Stations</i>																		
I29	7020	1.1	1.92	35.4	0.13	ND	12.9	4.8	7850	2.5	78.5	0.012	5.0	ND	ND	ND	0.7	20.7
I21	1220	0.7	7.69	1.9	0.07	ND	11.4	ND	7240	2.9	13.6	ND	0.7	ND	ND	ND	0.2	6.4
I13	862	0.6	6.32	1.7	0.06	ND	9.4	ND	5540	2.3	14.9	ND	0.5	ND	ND	ND	0.2	4.8
I8	1700	0.5	2.13	4.2	0.05	ND	8.3	ND	3820	1.1	20.6	ND	1.1	ND	ND	ND	0.2	6.7
<i>55-m Stations</i>																		
I28	5040	0.8	2.11	24.1	0.12	ND	9.7	4.6	6630	2.6	56.5	0.020	4.9	ND	ND	ND	0.6	16.8
I20	1400	0.4	3.08	3.0	0.06	ND	5.3	ND	4900	1.5	18.4	ND	0.9	ND	ND	ND	0.3	6.0
I7	1200	0.8	6.84	2.1	0.06	ND	8.6	ND	6780	2.3	20.5	ND	0.7	ND	ND	ND	0.2	5.9
I1	2430	0.3	0.97	9.0	0.06	0.04	6.6	ND	3350	1.5	36.3	0.005	2.3	ND	ND	ND	0.2	7.3
DR (%)	100	100	100	100	100	4	100	37	100	100	100	30	100	0	0	0	100	100
ERL ^b	—	—	8.2	—	—	1.2	81	34	—	46.7	—	0.15	20.9	—	1.0	—	—	150
ERM ^b	—	—	70.0	—	—	9.6	370	270	—	218.0	—	0.71	51.6	—	3.7	—	—	410

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.10 continued

Concentrations of trace metals (ppm) detected in sediments from SBOO stations sampled during summer 2020. See Appendix B.1 for MDLs and translation of periodic table symbols; nd = not detected; ns = not sampled.

	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Ti	Sn	Zn
19-m Stations																		
I35	6920	1.0	2.29	35.1	0.13	0.07	12.5	4.4	8600	2.7	101.0	0.019	4.1	ND	ND	ND	1.2	25.8
I34	1010	ND	2.70	3.0	0.02	0.03	2.3	1.6	2470	1.0	23.9	ND	0.5	ND	ND	ND	0.2	4.0
I31	3530	0.6	1.92	15.5	0.06	0.04	7.4	1.6	4090	1.2	62.7	ND	1.8	ND	ND	ND	0.6	9.3
I23	4660	0.7	2.01	24.8	0.08	0.04	8.6	2.3	5200	1.5	67.9	ND	2.5	ND	ND	ND	0.7	12.6
I18	4380	0.8	1.82	29.3	0.08	ND	10.9	2.2	5960	1.4	70.5	ND	2.4	ND	ND	ND	0.4	12.2
I10	4900	0.8	1.67	24.8	0.09	ND	9.4	2.4	5740	1.3	63.8	ND	2.6	ND	ND	ND	0.4	14.1
I4	738	0.2	1.45	1.9	0.03	ND	4.3	ND	1540	1.1	10.9	ND	0.6	ND	ND	ND	0.1	2.5
28-m Stations																		
I33	4540	0.8	2.15	17.5	0.09	0.04	8.5	4.0	6160	2.7	76.3	0.020	2.6	ND	ND	ND	0.9	16.0
I30	5810	0.7	1.71	26.0	0.10	0.05	10.3	2.9	6060	1.7	66.8	0.006	3.1	ND	ND	ND	0.8	18.0
I27	5510	0.8	1.83	27.3	0.10	ND	10.2	3.0	5820	1.6	61.1	0.005	3.2	ND	ND	ND	0.5	15.9
I22	4600	0.8	1.55	25.0	0.09	ND	9.5	2.5	4990	1.5	57.1	0.005	2.9	ND	ND	ND	0.4	41.3
I14 ^a	5860	0.8	1.70	29.5	0.10	ND	10.4	3.0	6120	1.5	69.4	0.004	3.4	ND	ND	ND	0.5	16.6
I16 ^a	3100	0.5	1.46	11.4	0.07	0.02	7.3	1.3	4130	1.2	43.6	0.004	1.6	ND	ND	ND	0.3	10.5
I15	2150	0.6	1.99	5.6	0.06	ND	8.5	ND	4020	1.4	28.7	0.004	1.2	ND	ND	ND	0.2	8.2
I12 ^a	5540	0.8	1.84	31.5	0.10	ND	9.8	2.6	6320	1.3	69.1	0.004	3.0	ND	ND	ND	0.5	17.3
I9	7220	1.0	2.17	38.1	0.12	ND	12.2	3.9	7790	1.4	80.6	0.005	4.4	ND	ND	ND	0.5	21.5
I6	855	0.4	4.86	1.9	0.04	ND	7.6	ND	3560	1.5	7.5	ND	0.5	ND	ND	ND	0.1	3.4
I2	1010	0.2	0.93	2.2	0.02	ND	5.4	ND	1190	0.9	8.3	ND	0.7	ND	ND	ND	0.1	2.5
I3	597	ND	1.77	1.1	0.02	ND	4.7	ND	1610	0.8	25.8	ND	0.6	ND	ND	ND	0.1	2.0
38-m Stations																		
I29	5250	0.7	2.46	25.5	0.11	ND	11.2	3.8	6780	2.4	60.2	0.014	3.9	ND	ND	ND	0.6	17.5
I21	1260	0.7	8.95	0.3	0.07	0.06	11.3	ND	7660	3.2	15.8	ND	1.0	ND	ND	ND	0.2	7.1
I13	952	0.5	5.94	0.3	0.05	0.03	8.6	ND	5070	2.2	15.7	0.004	0.7	ND	ND	ND	0.2	5.1
I8	1590	0.4	2.47	4.3	0.05	ND	8.7	ND	3810	1.2	18.0	ND	1.0	ND	ND	ND	0.2	7.1
55-m Stations																		
I28	4540	0.6	2.50	16.9	0.11	0.06	8.1	3.7	6400	2.4	58.7	0.016	4.2	ND	ND	ND	0.8	14.6
I20	1310	0.4	3.24	1.6	0.06	0.04	5.2	ND	4810	1.6	15.9	ND	0.9	ND	ND	ND	0.2	5.6
I7	1140	0.5	5.19	2.7	0.05	ND	8.3	ND	5960	2.0	17.8	ND	0.9	ND	ND	ND	0.2	5.5
I1	2390	0.5	1.18	7.7	0.06	ND	6.6	1.4	3330	1.5	35.2	0.006	2.4	ND	ND	ND	0.3	8.3
DR (%)	100	93	100	100	100	41	100	63	100	100	100	52	100	0	0	0	100	100
ERL ^b	—	—	8.2	—	—	1.2	81	34	—	46.7	—	0.15	20.9	—	1.0	—	—	150
ERM ^b	—	—	70.0	—	—	9.6	370	270	—	218.0	—	0.71	51.6	—	3.7	—	—	410

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.11

Concentrations of trace metals (ppm) detected in sediments from the San Diego regional benthic stations sampled during summer 2020. See Appendix B.1 for MDLs; ND= not detected.

	Station	Depth (m)	Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe
<i>Inner Shelf</i>	8929	7	3920	0.3	0.83	30.6	ND	ND	8.2	2.0	6610
	8944	7	2280	ND	0.88	14.8	ND	ND	5.1	ND	3560
	8918	8	1700	ND	1.24	10.6	0.02	ND	4.2	ND	3130
	8946	15	2660	0.2	1.27	9.4	ND	ND	5.3	1.4	3490
	8915	16	2100	0.2	1.74	11.8	0.04	ND	6.2	2.6	4560
	8907	18	2790	0.3	1.15	12.1	0.02	0.04	7.2	1.6	3520
	8943	19	3010	0.3	0.96	12.1	ND	ND	7.1	1.5	3510
	8914	20	4910	0.4	1.78	29.2	0.08	ND	11.1	4.2	7300
	8911	24	4310	0.3	2.10	19.0	0.05	0.07	9.6	2.8	5250
	8938	28	2410	0.3	1.16	12.8	ND	ND	6.5	1.6	3930
8901	29	5530	0.5	1.85	36.3	0.05	0.05	11.3	3.6	7150	
<i>Mid Shelf</i>	8913	31	3480	0.2	1.62	17.5	0.06	ND	8.2	3.3	5510
	8906	41	1390	0.4	2.66	2.1	0.06	0.03	7.3	ND	4490
	8939	65	5860	0.6	1.68	30.0	ND	ND	12.8	4.0	9210
	8903	66	2570	0.4	1.83	9.4	0.05	0.05	6.8	2.3	4180
	8912	67	7990	0.8	3.29	39.7	0.12	0.12	17.1	9.8	12400
	8925	67	9390	0.6	2.73	48.1	ND	ND	19.9	9.1	13300
	8919	68	7330	0.6	3.09	43.2	0.14	ND	16.9	8.6	12700
	8909	73	5730	0.6	3.45	27.5	0.09	0.09	13.6	7.1	8990
	8931	74	9450	0.7	2.75	42.9	ND	ND	19.6	8.4	13400
	8933	80	9380	0.7	2.55	40.4	ND	ND	19.7	8.1	13400
	8930	83	10200	0.6	2.04	44.6	ND	ND	21.1	9.5	14100
	8949	83	9930	0.7	1.64	44.7	ND	ND	20.2	8.9	13300
	8924	85	8730	0.6	2.13	39.5	ND	ND	18.6	8.1	12600
	8926	87	6930	0.6	1.67	30.6	ND	ND	15.1	6.2	10200
	8932	89	9520	0.9	2.25	36.0	ND	ND	26.1	8.3	18500
8940	108	3550	0.4	2.04	16.6	ND	ND	9.3	3.2	6540	
8921	116	3950	0.3	2.13	16.7	0.09	ND	10.5	4.2	7080	
<i>Outer Shelf</i>	8905	134	6560	0.5	2.21	28.0	0.12	0.09	15.6	8.0	9790
	8937	137	6360	0.5	1.42	27.8	ND	ND	15.1	5.9	9830
	8910	169	10300	0.9	3.21	62.4	0.13	0.11	19.4	34.5	18000
	8922	185	7270	0.6	2.14	36.9	0.17	ND	19.0	9.6	12700
	8916	189	10100	0.8	2.50	57.7	0.24	ND	25.6	19.6	17600
	8908	194	10700	0.9	3.02	51.3	0.22	0.18	26.1	15.9	17300
<i>Upper Slope</i>	8928	223	11200	0.7	1.43	51.9	ND	ND	24.8	12.6	14400
	8920	270	10400	0.9	3.83	59.1	0.26	ND	27.1	15.5	17300
	8936	292	11600	1.0	1.23	53.9	ND	ND	27.3	13.4	15400
	8927	460	13400	0.9	1.36	91.3	ND	ND	33.1	18.0	16700
	8934	491	15700	1.4	1.49	120.0	ND	ND	38.0	21.5	18900
	8923	523	9750	1.3	3.55	87.6	0.38	ND	41.9	16.5	24100
Detection Rate (%)			100	95	100	100	50	25	100	93	100
ERL ^a :			—	—	8.2	—	—	1.2	81	34	—
ERM ^a :			—	—	70.0	—	—	9.6	370	270	—

^aFrom Long et al. 1995

Appendix B.11 *continued*

	Station	Depth (m)	Pb	Mn	Hg	Ni	Se	Ag	Tl	Sn	Zn
<i>Inner Shelf</i>	8929	7	1.5	87.5	ND	1.9	ND	ND	ND	0.3	18.2
	8944	7	0.9	56.2	ND	1.1	ND	ND	ND	ND	8.8
	8918	8	0.9	31.6	ND	0.9	ND	ND	ND	0.1	7.1
	8946	15	1.4	38.0	0.007	1.4	ND	ND	0.19	ND	8.8
	8915	16	2.7	43.1	0.010	1.5	ND	ND	ND	0.3	10.4
	8907	18	1.2	42.2	ND	1.6	ND	ND	ND	0.2	8.4
	8943	19	1.0	45.1	ND	1.5	ND	ND	0.14	ND	8.2
<i>Mid Shelf</i>	8914	20	2.9	71.9	0.018	3.6	ND	ND	ND	0.5	21.7
	8911	24	1.8	49.8	0.008	2.9	ND	ND	ND	0.3	13.9
	8938	28	1.0	51.0	ND	2.0	ND	ND	0.13	ND	9.2
	8901	29	1.5	72.2	0.004	3.6	ND	ND	ND	0.4	19.5
	8913	31	2.7	58.6	0.018	2.5	ND	ND	ND	0.6	15.8
	8906	41	1.5	13.2	ND	1.0	ND	ND	ND	0.1	7.9
	8939	65	2.5	80.3	0.009	4.2	ND	ND	0.22	ND	23.1
	8903	66	2.1	34.2	0.011	3.0	ND	ND	ND	0.3	9.6
	8912	67	5.6	97.1	0.056	7.4	ND	ND	ND	1.0	36.7
	8925	67	5.5	115.0	0.098	8.4	ND	ND	ND	1.2	35.0
	8919	68	4.3	106.0	0.041	7.0	ND	ND	ND	1.1	33.5
	8909	73	4.9	70.5	0.063	5.9	ND	ND	ND	0.8	23.8
	8931	74	5.2	110.0	0.041	7.9	ND	ND	ND	1.1	33.9
	8933	80	4.9	107.0	0.038	7.9	ND	ND	ND	1.1	33.1
<i>Outer Shelf</i>	8930	83	5.3	115.0	0.048	9.1	ND	ND	ND	1.2	36.5
	8949	83	4.3	113.0	0.075	8.6	ND	ND	0.18	ND	33.9
	8924	85	4.4	103.0	0.037	8.0	ND	ND	ND	1.0	31.7
	8926	87	3.8	82.9	0.027	6.4	0.29	ND	ND	0.7	25.2
	8932	89	5.1	102.0	0.039	8.5	ND	ND	ND	1.1	37.0
	8940	108	2.1	47.3	0.010	3.6	ND	ND	ND	ND	14.8
	8921	116	2.7	49.4	0.019	4.3	ND	ND	ND	0.4	17.9
	8905	134	72.7	75.9	0.041	7.5	ND	ND	ND	0.9	25.7
	8937	137	3.0	75.3	0.025	5.6	ND	ND	0.21	ND	24.3
	8910	169	7.8	128.0	0.081	6.7	ND	ND	ND	1.1	55.4
	8922	185	4.2	90.2	0.046	9.4	ND	ND	ND	0.8	33.1
	8916	189	7.6	121.0	0.107	12.5	0.38	ND	ND	2.8	79.2
	8908	194	5.0	112.0	0.079	13.6	0.51	ND	ND	1.3	42.3
<i>Upper Slope</i>	8928	223	4.6	119.0	0.057	12.8	0.42	ND	ND	1.1	42.0
	8920	270	4.8	121.0	0.066	16.2	0.64	ND	ND	1.0	47.8
	8936	292	4.4	123.0	0.053	12.8	ND	ND	0.17	ND	42.3
	8927	460	4.7	136.0	0.059	18.9	0.76	ND	ND	1.0	54.7
	8934	491	5.1	158.0	0.078	21.7	ND	ND	0.17	ND	62.6
	8923	523	4.4	92.6	0.053	15.8	0.87	ND	ND	0.9	52.0
Detection Rate (%)			100	100	83	100	18	0	20	75	100
ERL ^a :			46.7	—	0.15	20.9	—	1.0	—	—	150
ERM ^a :			218.0	—	0.71	51.6	—	3.7	—	—	410

^aFrom Long et al. 1995

Appendix B.12

Concentrations of pesticides (ppt) detected in sediments from PLOO stations sampled during 2020. See Appendix B.1 for MDLs and abbreviations; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

Winter	Chlordane										DDT									
	2020	tChlord	A(C)C	cNon	G(t)C	Hept	HeptEpo	Methoxy	Oxychlor	tNon	tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT		
<i>88-m Depth Contour</i>																				
B11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	1034.2	21.4 E	39	21.6 E	45.8 E	53.9	784	68.5		
B8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	966.8	23.2 E	37.3	20.9 E	53	60.9	699	72.5		
E19	9.93	ND	ND	ND	ND	ND	ND	ND	9.93 E	ND	583.6	18.2 E	23.2	21 E	36.9 E	37.7 E	391	55.6		
E7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	638.5	17.5 E	18	18 E	32.5 E	47.8	449	55.7		
E1	116.5	25.4 E	13.8 E	45.9	ND	ND	ND	ND	31.4 E	ND	723.5	19.1 E	16.4	18.6 E	39.4 E	58.6	518	53.4		
<i>98-m Depth Contour</i>																				
B12	11.4	ND	ND	11.4 E	ND	ND	ND	ND	ND	ND	485.9	ND	18.1	ND	20.3 E	20.5 E	408	19 E		
B9	11.9	ND	ND	11.9 E	ND	ND	ND	ND	ND	ND	704.1	14.1 E	22.2	18.4 E	36.2 E	39.3 E	527	46.9		
E26	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	1024.7	16.7 E	19.6	19.8 E	32.1 E	47.5	440	449		
E25	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	324.3	ND	10.9 E	ND	19.1 E	19.9 E	251	23.4 E		
E23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	523.3	16.5 E	15.7 E	13.8 E	28.8 E	36.9 E	372	39.6		
E20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	400.1	13.4 E	12.3 E	ND	20 E	25.3 E	299	30.1		
E17 ^a	9.96	ND	ND	9.96 E	ND	ND	ND	ND	ND	ND	530.29	ND	9.79 E	29.5 E	18.5 E	24.5 E	297	151		
E14 ^a	14.4	14.4 E	ND	ND	ND	ND	ND	ND	ND	ND	276.5	11 E	11.3 E	ND	20.3 E	17.9 E	216	ND		
E11 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	382.02	12.4 E	15.5 E	9.82 E	22.9 E	25.4 E	268	28		
E8	21.86	12 E	ND	9.86 E	ND	ND	ND	ND	ND	ND	399.11	9.22 E	14.3 E	9.99 E	28 E	25 E	286	26.6		
E5	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	418.8	ND	13.7 E	ND	29.5 E	25 E	328	22.6 E		
E2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	1120.7	25.3 E	27	65	43.8 E	60.6	678	221		
<i>116-m Depth Contour</i>																				
B10	36.2	12.3 E	ND	11.4 E	ND	ND	ND	ND	12.5 E	ND	578.2	13.3 E	26	11.7 E	31.1 E	28 E	439	29.1		
E21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	340.24	9.14 E	11.9 E	ND	17.8 E	21.6 E	258	21.8 E		
E15 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	358.2	10.8 E	12.2 E	ND	18.3 E	25.3 E	268	23.6 E		
E9	20.78	11 E	ND	9.78 E	ND	ND	ND	ND	ND	ND	380.6	11.8 E	17.4	ND	20.8 E	22 E	284	24.6		
E3	195.6	44.9 E	27.2 E	81.7	ND	ND	ND	ND	41.8	ND	606.2	27 E	11.1 E	13.5 E	29.1 E	128	311	86.5		
DR (%)	45	27	9	36	0	0	0	0	18	100	100	82	100	64	100	100	100	95		
ERL ^b	—	—	—	—	—	—	—	—	—	1580	—	—	—	—	—	—	2200	—		
ERM ^b	—	—	—	—	—	—	—	—	—	46,100	—	—	—	—	—	—	2700	—		

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.12 *continued*

Winter 2020	HCH							Endosulfan			HCB	Mirex		
	tHC	HCH-A	HCH-B	HCH-D	HCH-G	Aldrin	Dieldrin	Alpha	Beta	Sulfate			Endrin	EndAld
<i>88-m Depth Contour</i>														
B11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	135	ND
B8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E19	18.8	ND	18.8 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E1	ND	ND	ND	ND	ND	ND	ND	ND	13.3 E	ND	ND	ND	ND	ND
<i>98-m Depth Contour</i>														
B12	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
B9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	19.1 E	ND
E26	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E25	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	21.7 E	ND
E23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E17 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	22.7 E	ND
E14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E11 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E5	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
<i>116-m Depth Contour</i>														
B10	15.2	ND	15.2 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	167	ND
E21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
E15 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E9	8.88	ND	8.88 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	73.2	ND
E3	ND	ND	ND	ND	ND	ND	9.11 E	ND	ND	ND	ND	ND	ND	ND
DR (%)	14	0	14	0	0	0	5	0	0	5	0	0	40	0
ERL ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—
ERM ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.12 *continued*

Summer	Chlordane										DDT							
	2020	tChlord	A(c)C	cNon	G(t)C	Hept	HeptEpo	Methoxy	Oxychlor	tNon	tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT
<i>88-m Depth Contour</i>																		
B11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	602.2	12.3 E	20.4 E	16 E	21.7 E	30.8 E	440	61
B8	34.9	13.2 E	ND	11.4 E	ND	ND	ND	ND	10.3 E	918.5	28.2 E	28.9 E	21 E	42.9 E	67.6	656	73.9 E	
E19	ND	ND	ND	ND	ND	ND	ND	ND	ND	526.9	12.9 E	18 E	16.1 E	24.5 E	61.5	348	45.9	
E7	ND	ND	ND	ND	ND	ND	ND	ND	ND	539.1	16.8 E	15.1 E	17.5 E	28.6 E	40.9 E	375	45.2 E	
E1	248.4	84.6	29.4 E	77.1	ND	ND	ND	ND	57.3 E	1212.8	46.7 E	24.1 E	62.7 E	43.3 E	133	720	183	
<i>98-m Depth Contour</i>																		
B12	17.3	ND	ND	17.3 E	ND	ND	ND	ND	ND	636.5	17.1 E	36.7 E	ND	25 E	36.7 E	415	106	
B9	ND	ND	ND	ND	ND	ND	ND	ND	ND	749	23.8 E	25.7 E	20.6 E	32.8 E	72.6	489	84.5	
E26	ND	ND	ND	ND	ND	ND	ND	ND	ND	434.5	12.8 E	14.9 E	ND	23.7 E	30.3 E	323	29.8 E	
E25	ND	ND	ND	ND	ND	ND	ND	ND	ND	433.5	11 E	11.7 E	21.1 E	19.6 E	27.7 E	276	66.4 E	
E23	ND	ND	ND	ND	ND	ND	ND	ND	ND	281.6	ND	10.6 E	11 E	ND	18.9 E	213	28.1 E	
E20	ND	ND	ND	ND	ND	ND	ND	ND	ND	548.7	11.3 E	27 E	ND	28.4 E	110	320	52	
E17 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	196.3	ND	ND	ND	12.9 E	13.3 E	152	18.1 E	
E14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	93.4	ND	ND	ND	ND	ND	93.4	ND	
E11 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	218	ND	ND	ND	11.8 E	12.2 E	171	23 E	
E8	ND	ND	ND	ND	ND	ND	ND	ND	ND	381.1	12.8 E	12.2 E	12.8 E	19.7 E	33.7 E	252	37.9 E	
E5	ND	ND	ND	ND	ND	ND	ND	ND	ND	588.5	16.9 E	17.7 E	18 E	28.5 E	43.2	403	61.2 E	
E2	36.2	13 E	ND	11.6 E	ND	ND	ND	ND	11.6 E	626.6	20.2 E	18.1 E	17.6 E	32.4 E	56.6	429	52.7 E	
<i>116-m Depth Contour</i>																		
B10	ND	ND	ND	ND	ND	ND	ND	ND	ND	313.6	ND	12.3 E	ND	14.3 E	12.5 E	258	16.5 E	
E21	ND	ND	ND	ND	ND	ND	ND	ND	ND	190.2	ND	ND	ND	10.8 E	16 E	138	25.4 E	
E15 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	307.47	9.47 E	ND	16.7 E	10.7 E	15.6 E	213	42	
E9	ND	ND	ND	ND	ND	ND	ND	ND	ND	355.9	13.2 E	12.2 E	ND	17.3 E	40	251	22.2 E	
E3	187.8	54.7 E	25.4 E	67.7 E	ND	ND	ND	ND	40 E	462.8	30.9 E	10.3 E	ND	23.5 E	147	224	27.1 E	
DR (%)	23	18	9	23	0	0	0	0	18	100	73	77	55	91	95	100	95	
ERL ^b	—	—	—	—	—	—	—	—	—	1580	—	—	—	—	—	2200	—	
ERM ^b	—	—	—	—	—	—	—	—	—	46,100	—	—	—	—	—	2700	—	

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.12 *continued*

Summer 2020	HCH										Endosulfan			HCB	Mirex
	tHCH	HCH-A	HCH-B	HCH-D	HCH-G	Aldrin	Dieldrin	Alpha	Beta	Sulfate	Endrin	EndAld			
<i>88-m Depth Contour</i>															
B11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
B8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E19	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>98-m Depth Contour</i>															
B12	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
B9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E26	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E25	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	98.2 E	ND	ND
E17 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E11 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E5	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
E2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
<i>116-m Depth Contour</i>															
B10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E15 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
DR (%)	0	0	0	0	0	0	0	0	0	0	0	0	7	0	0
ERL ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
ERM ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—

^a Near-ZID station

^b From Long et al. 1995

Appendix B.13

Concentrations of pesticides (ppt) detected in sediments from SBOO stations sampled during 2020. See Appendix B.1 for MDLs and abbreviations; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

Winter 2020	Chlordane							DDT										
	tChlor	A(c)C	cNon	G(t)C	Hept	HeptE	pox	Methoxy	Oxychlor	tNon	tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT
<i>19-m Stations</i>																		
I35	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	191.7	ND	ND	ND	18.1 E	16.7 E	140	16.9 E
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I31	28.17	ND	ND	18.6 E	ND	ND	ND	ND	9.57 E	10.6	10.6	ND	ND	10.6 E	ND	NR	NR	ND
I23	ND	ND	ND	ND	ND	ND	ND	ND	ND	43.5	43.5	ND	ND	ND	ND	43.5	ND	ND
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	35.88	35.88	ND	ND	9.38 E	ND	26.5	ND	ND
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	26.4	26.4	ND	ND	ND	ND	26.4	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	10.8	10.8	ND	ND	ND	ND	10.8 E	ND	ND
<i>28-m Stations</i>																		
I33	55.4	14.8 E	ND	13.4 E	16 E	ND	ND	ND	11.2 E	105.7	105.7	ND	12.2 E	18.3 E	ND	75.2	ND	ND
I30	10.1	ND	ND	10.1 E	ND	ND	ND	ND	ND	124.6	124.6	ND	ND	ND	9.9 E	103	11.7 E	ND
I27	9.16	ND	ND	9.16 E	ND	ND	ND	ND	ND	96.87	96.87	ND	ND	10.9 E	ND	76.6	9.37 E	ND
I22	99.6	29.8 E	ND	47.7	22.1 E	ND	ND	ND	ND	126.2	126.2	ND	ND	ND	31.5 E	73.4	21.3 E	ND
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	422.5	422.5	32.3 E	10.5 E	16.6 E	73.1	290	ND	ND
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	97.06	97.06	ND	8.36 E	14.9 E	16.2 E	57.6	ND	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	13.9	13.9	ND	ND	ND	ND	13.9 E	ND	ND
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	15.6	15.6	ND	ND	ND	ND	15.6 E	ND	ND
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	83.4	83.4	ND	ND	ND	ND	70.8	12.6 E	ND
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	13	13	ND	ND	ND	ND	13 E	ND	ND
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	12.3	12.3	ND	ND	ND	ND	12.3 E	ND	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	15	15	ND	ND	ND	ND	15 E	ND	ND
<i>38-m Stations</i>																		
I29	24.3	13.4 E	ND	10.9 E	ND	ND	ND	ND	ND	3493.1	3493.1	121	41.1	122	243	2200	613	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	11.6	11.6	ND	ND	ND	ND	11.6 E	ND	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	18.5	18.5	ND	ND	ND	ND	18.5	ND	ND
<i>55-m Stations</i>																		
I28	ND	ND	ND	ND	ND	ND	ND	ND	ND	2183.5	2183.5	58.7	31	68.8	136	1120	587	ND
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND
I7	ND	ND	ND	ND	ND	ND	ND	ND	ND	14.7	14.7	ND	ND	ND	ND	14.7 E	ND	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	54	54	ND	ND	ND	ND	54	ND	ND
DR (%)	22	11	0	19	11	0	0	0	7	89	89	11	19	7	33	26	100	26
ERL ^b	—	—	—	—	—	—	—	—	—	1580	1580	—	—	—	—	2200	—	—
ERM ^b	—	—	—	—	—	—	—	—	—	46,100	46,100	—	—	—	—	2700	—	—

^a Near-ZID station; ^b From Long et al. 1995

Appendix B.13 continued

Winter 2020	HCH							Endosulfan				Mirex		
	tHCH	HCH-A	HCH-B	HCH-D	HCH-G	Aldrin	Dieldrin	Alpha	Beta	Sulfate	Endrin		EndAld	HCB
<i>19-m Stations</i>														
I35	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I31	47.2	23 E	NR	ND	24.2	ND	ND	ND	ND	ND	ND	ND	NR	ND
I23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18	10.7	ND	10.7 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>28-m Stations</i>														
I33	17.9	NR	NR	ND	17.9	ND	ND	ND	ND	ND	ND	ND	NR	ND
I30	11.2	ND	NR	ND	11.2 E	ND	ND	ND	ND	ND	ND	ND	NR	ND
I27	9.28	ND	ND	ND	9.28 E	ND	ND	ND	ND	ND	ND	ND	NR	ND
I22	217.7	34.5	150	ND	33.2	ND	ND	26.5 E	ND	ND	ND	ND	NR	ND
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	179	ND
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	34.5 E	ND
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	9.07	ND	9.07 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3	31.8	ND	15.7 E	16.1 E	ND	ND	9.96 E	ND	ND	ND	ND	ND	21.6 E	ND
<i>38-m Stations</i>														
I29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>55-m Stations</i>														
I28	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	30	8	17	4	19	0	4	4	0	0	0	0	20	0
ERL ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—
ERM ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—

^aNear-ZID station; ^bFrom Long et al. 1995

Appendix B.13 *continued*

Summer 2020	Chlordane										DDT						
	tChlord	A(c)C	cNon	G(t)C	Hept	HeptEpo	Methoxy	Oxychlor	tNon	tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT
<i>19-m Stations</i>																	
I35	ND	ND	ND	ND	ND	ND	ND	ND	ND	189.5	ND	ND	ND	15.1 E	23.3 E	134	17.1 E
I34	ND	ND	ND	ND	ND	ND	ND	ND	43.95	ND	ND	ND	ND	ND	35	NR	8.95 E
I31	ND	ND	ND	ND	ND	ND	ND	ND	30.1	ND	ND	ND	ND	ND	13.9 E	NR	16.2 E
I23	ND	ND	ND	ND	ND	ND	ND	ND	94.37	ND	ND	ND	ND	9.01 E	9.26 E	76.1 E	ND
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
<i>28-m Stations</i>																	
I33	ND	ND	ND	ND	ND	ND	ND	ND	111.2	ND	ND	ND	ND	10.4 E	ND	86.8	14 E
I30	169.3	30.9 E	33.1 E	26.2 E	ND	52.1 E	ND	27 E	308.3	31.6 E	25.8 E	35.5 E	10.8 E	39.4 E	119	46.2 E	
I27	33.43	11.5 E	ND	12 E	9.93 E	ND	ND	ND	138.66	ND	9.31 E	ND	13.8 E	9.15 E	94.7	11.7 E	
I22	ND	ND	ND	ND	ND	ND	ND	ND	86.7	ND	ND	ND	ND	ND	ND	86.7	ND
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	167.82	ND	ND	9.48 E	9.44 E	15.5 E	101	32.4 E	
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	45.6	ND	ND	ND	ND	ND	ND	45.6 E	ND
I9	ND	ND	ND	ND	ND	ND	ND	ND	131	ND	ND	ND	ND	ND	10 E	106	15 E
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
<i>38-m Stations</i>																	
I29	ND	ND	ND	ND	ND	ND	ND	ND	1903.6	75 E	38.2 E	25.3 E	109	182	1400	74.1 E	
I21	ND	ND	ND	ND	ND	ND	ND	ND	9.66	ND	ND	ND	ND	ND	NR	9.66 E	
I13	ND	ND	ND	ND	ND	ND	ND	ND	140.26	ND	10.3 E	ND	9.36 E	46	60.9 E	13.7 E	
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND
<i>55-m Stations</i>																	
I28	ND	ND	ND	ND	ND	ND	ND	ND	568.3	16.1 E	12 E	18.1 E	25.1 E	43.9	403	50.1 E	
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND
I7	9.49	ND	ND	9.49 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	68.4	ND	ND	ND	ND	ND	68.4 E	ND	ND
DR (%)	11	7	4	11	4	0	4	0	59	11	19	15	33	41	100	44	
ERL ^b	—	—	—	—	—	—	—	—	1580	—	—	—	—	—	2200	—	
ERM ^b	—	—	—	—	—	—	—	—	46,100	—	—	—	—	—	2700	—	

^aNear-ZID station; ^bFrom Long et al. 1995

Appendix B.13 *continued*

Summer 2020	HCH				Endosulfan				Mirex						
	tHCH	HCH-A	HCH-B	HCH-D	HCH-G	Aldrin	Dieldrin	Alpha		Beta	Sulfate	Endrin	EndAld	HCB	
<i>19-m Stations</i>															
I35	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I31	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I23	8.74	8.74 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
<i>28-m Stations</i>															
I33	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I30	47.5	ND	26.4 E	21.1 E	ND	ND	ND	ND	ND	28.5 E	ND	ND	ND	ND	28.8 E
I27	37.1	12.6 E	11.6 E	ND	12.9 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I22	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>38-m Stations</i>															
I29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>55-m Stations</i>															
I28	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	35.8	10.9 E	12.8 E	ND	12.1 E	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	15	11	11	4	7	0	0	0	0	4	0	0	0	0	4
ERL ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
ERM ^b	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—

^a Near-ZID station; ^b From Long et al. 1995

Appendix B.14

Concentrations of pesticides (ppt) detected in sediments from the San Diego regional benthic stations sampled during summer 2020. See Appendix B.1 for MDLs and abbreviations; E= estimated; ND=not detected; NR=not reportable.

	Station	Depth (m)	Chlordane								tNon
			tChlord	A(c)C	cNon	G(t)C	Hept	HeptEpox	Methoxy	Oxychlor	
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8944	7	11.5	5.65 E	ND	5.85 E	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8911	24	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8938	28	150.7	34 E	ND	48.6	ND	34.9 E	ND	ND	33.2 E
	8901	29	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8903	66	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8912	67	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8925	67	71.1	30.3 E	ND	28 E	ND	ND	ND	ND	12.8 E
	8919	68	44.9	16.2 E	ND	15.8 E	ND	ND	ND	ND	12.9 E
	8909	73	197.6	49.9 E	ND	115	ND	ND	ND	ND	32.7 E
	8931	74	25.6	ND	ND	13.6 E	ND	ND	ND	ND	12 E
	8933	80	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8930	83	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8949	83	159.7	ND	ND	72.6	ND	42.6 E	ND	ND	44.5 E
	8924	85	10.6	ND	ND	10.6 E	ND	ND	ND	ND	ND
	8926	87	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8932	89	51.1	18.5 E	ND	18.5 E	ND	ND	ND	ND	14.1 E
8940	108	ND	ND	ND	ND	ND	ND	ND	ND	ND	
8921	116	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>Outer Shelf</i>	8905	134	10.6	ND	ND	10.6 E	ND	ND	ND	ND	ND
	8937	137	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8910	169	918	234	106	381	ND	ND	ND	ND	197
	8922	185	42.6	ND	10.1 E	16.5 E	ND	ND	ND	ND	16 E
	8916	189	48.7	ND	21.6 E	ND	ND	ND	ND	ND	27.1 E
	8908	194	37	ND	ND	37 E	ND	ND	ND	ND	ND
<i>Upper Slope</i>	8928	223	32.4	ND	ND	16 E	ND	ND	ND	ND	16.4 E
	8920	270	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8936	292	60.6	22.5 E	ND	19.1 E	ND	ND	ND	ND	19 E
	8927	460	63.7	25.9 E	ND	21 E	ND	ND	ND	ND	16.8 E
	8934	491	28.1	28.1 E	ND	ND	ND	ND	ND	ND	ND
	8923	523	59.1	19.1 E	ND	20.1 E	ND	ND	ND	ND	19.9 E
Detection Rate (%)			48	28	8	43	0	5	0	0	35
ERL ^a :			—	—	—	—	—	—	—	—	—
ERM ^a :			—	—	—	—	—	—	—	—	—

^aFrom Long et al. 1995

Appendix B.14 *continued*

	Station	Depth (m)	DDT							
			tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT
<i>Inner Shelf</i>	8929	7	16.9	ND	ND	ND	ND	ND	16.9 E	ND
	8944	7	13.5	ND	ND	ND	ND	ND	13.5 E	ND
	8918	8	44.43	ND	4.73 E	ND	ND	18.2 E	21.5 E	ND
	8946	15	24.4	ND	ND	ND	ND	ND	24.4 E	ND
	8915	16	ND	ND	ND	ND	ND	ND	NR	ND
	8907	18	89.5	ND	ND	ND	10.1 E	ND	79.4	ND
	8943	19	25.7	ND	ND	ND	ND	ND	25.7 E	ND
	8914	20	196.4	ND	ND	ND	10.2 E	17 E	108	61.2
	8911	24	93	ND	ND	ND	11.9 E	ND	81.1 E	ND
	8938	28	141.6	19.5 E	34.6 E	ND	ND	17.8 E	51.5 E	18.2 E
8901	29	154.2	ND	ND	ND	10.7 E	15.6 E	115	12.9 E	
<i>Mid Shelf</i>	8913	31	46.4	ND	ND	ND	ND	ND	46.4 E	ND
	8906	41	69.3	ND	ND	ND	ND	ND	69.3 E	ND
	8939	65	197.41	ND	10.1 E	ND	10.6 E	10.3 E	157	9.41 E
	8903	66	192.43	ND	ND	ND	8.83 E	ND	164	19.6 E
	8912	67	976.9	37.1 E	21.3 E	ND	46.9 E	124	674	73.6
	8925	67	662.9	23.8 E	17.7 E	18.6 E	34.2 E	56	455	57.6 E
	8919	68	761.9	34.2 E	21 E	25.1 E	46 E	85	482	68.6 E
	8909	73	1499	52.2	25.8 E	34.2 E	55.8	166	948	217
	8931	74	798.1	21.5 E	24.9 E	27 E	40.7 E	62.6	544	77.4 E
	8933	80	827.3	23.1 E	26.6 E	24.5 E	41 E	65.1	565	82 E
	8930	83	1096.5	29.1 E	46 E	40.2 E	52.3 E	71.9	731	126
	8949	83	955.9	37.2 E	63	52.5	35.1 E	56.5	623	88.6
	8924	85	828.4	25 E	24.7 E	20.8 E	45.8 E	76.9	571	64.2 E
	8926	87	468.5	14.1 E	13.4 E	17.1 E	23.2 E	36.7 E	317	47 E
	8932	89	1040.7	28.2 E	38.8 E	36.5 E	48.7 E	77.5	706	105
8940	108	164	ND	ND	ND	9 E	ND	155	ND	
8921	116	251.85	ND	5.05 E	ND	15.1 E	19 E	196	16.7 E	
<i>Outer Shelf</i>	8905	134	604.3	13.4 E	21.4 E	10.8 E	26.1 E	42.8	448	41.8 E
	8937	137	461	10.4 E	18.5 E	ND	23.3 E	25.9 E	356	26.9 E
	8910	169	1954.1	38.5 E	16.9 E	25 E	50.7 E	182	411	1230
	8922	185	1373.7	38.7 E	48.6 E	31.9 E	63.2	95.5	999	96.8
	8916	189	1639.3	35.5 E	57.6 E	ND	83.9	163	1210	89.3
	8908	194	1283.7	24.1 E	49.3 E	20.3 E	49.7 E	91.9	971	77.4
<i>Upper Slope</i>	8928	223	1794.1	46.9 E	66 E	36.1 E	74.1	143	1320	108
	8920	270	2486.9	54.9 E	99	112	113	138	1820	150
	8936	292	3448.7	73.7 E	120	192	109	187	2030	737
	8927	460	3123.5	95 E	178	33.7 E	113	203	2410	90.8 E
	8934	491	4231.2	110 E	284	49.2 E	206	253	3180	149
	8923	523	2618.9	78.9 E	141	36.7 E	98.3	170	1960	134
Detection Rate (%)			98	60	68	50	78	73	100	73
ERL ^a :			1580	—	—	—	—	—	2200	—
ERM ^a :			46,100	—	—	—	—	—	2700	—

^aFrom Long et al. 1995

Appendix B.14 *continued*

	Station	Depth (m)	HCH					Aldrin	Dieldrin
			tHCH	HCH-A	HCH-B	HCH-D	HCH-G		
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	ND
	8944	7	ND	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	ND	ND	ND	ND	ND
	8911	24	ND	ND	ND	ND	ND	ND	ND
	8938	28	88.6	ND	44.3 E	44.3 E	ND	ND	56.1 E
8901	29	ND	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	ND	ND	ND	ND
	8903	66	ND	ND	ND	ND	ND	ND	ND
	8912	67	ND	ND	ND	ND	ND	ND	ND
	8925	67	ND	ND	ND	ND	ND	ND	ND
	8919	68	ND	ND	ND	ND	ND	ND	ND
	8909	73	ND	ND	ND	ND	ND	ND	ND
	8931	74	ND	ND	ND	ND	ND	ND	ND
	8933	80	ND	ND	ND	ND	ND	ND	ND
	8930	83	ND	ND	ND	ND	ND	ND	ND
	8949	83	130.6	ND	60.9	47.4 E	22.3 E	ND	65.6 E
	8924	85	ND	ND	ND	ND	ND	ND	ND
	8926	87	ND	ND	ND	ND	ND	ND	ND
	8932	89	ND	ND	NR	ND	ND	ND	ND
8940	108	ND	ND	ND	ND	ND	ND	ND	
8921	116	ND	ND	ND	ND	ND	ND	ND	
<i>Outer Shelf</i>	8905	134	ND	ND	ND	ND	ND	ND	ND
	8937	137	ND	ND	ND	ND	ND	ND	ND
	8910	169	ND	ND	ND	ND	ND	ND	64.8 E
	8922	185	ND	ND	ND	ND	ND	ND	ND
	8916	189	ND	ND	ND	ND	ND	ND	ND
	8908	194	11.8	ND	11.8 E	ND	ND	ND	ND
<i>Upper Slope</i>	8928	223	ND	ND	NR	ND	ND	ND	ND
	8920	270	19.8	ND	19.8 E	ND	ND	ND	ND
	8936	292	ND	ND	ND	ND	ND	ND	ND
	8927	460	ND	ND	ND	ND	ND	ND	ND
	8934	491	ND	ND	ND	ND	ND	ND	ND
	8923	523	ND	ND	NR	ND	ND	ND	ND
Detection Rate (%)			10	0	11	5	3	0	8
ERL ^a :			—	—	—	—	—	—	—
ERM ^a :			—	—	—	—	—	—	—

^aFrom Long et al. 1995

Appendix B.14 *continued*

	Station	Depth (m)	Endosulfan			Endrin	EndAld	HCB	Mirex
			Alpha	Beta	Sulfate				
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	
	8944	7	ND	ND	ND	ND	ND	ND	
	8918	8	ND	ND	ND	ND	ND	ND	
	8946	15	ND	ND	ND	ND	ND	ND	
	8915	16	ND	ND	ND	ND	ND	ND	
	8907	18	ND	ND	ND	ND	ND	ND	
	8943	19	ND	ND	ND	ND	ND	ND	
	8914	20	ND	ND	ND	ND	ND	ND	
	8911	24	ND	ND	ND	ND	ND	ND	
	8938	28	ND	ND	ND	ND	ND	20.8 E	
	8901	29	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND	
	8906	41	ND	ND	ND	ND	ND	ND	
	8939	65	ND	ND	ND	ND	ND	ND	
	8903	66	ND	ND	ND	ND	ND	ND	
	8912	67	ND	ND	ND	ND	ND	ND	
	8925	67	ND	ND	ND	ND	ND	ND	
	8919	68	ND	ND	ND	ND	ND	ND	
	8909	73	ND	ND	ND	ND	NR	ND	
	8931	74	ND	ND	ND	ND	50.9 E	ND	
	8933	80	ND	ND	ND	ND	ND	ND	
	8930	83	ND	ND	ND	ND	ND	ND	
	8949	83	ND	ND	ND	ND	ND	20.7 E	
	8924	85	ND	ND	ND	ND	38.3 E	ND	
	8926	87	ND	ND	ND	ND	ND	ND	
8932	89	ND	ND	ND	ND	27.3 E	ND		
8940	108	ND	ND	ND	ND	ND	ND		
8921	116	ND	ND	ND	ND	44.6 E	ND		
<i>Outer Shelf</i>	8905	134	ND	ND	ND	ND	ND	ND	
	8937	137	ND	ND	ND	ND	ND	ND	
	8910	169	ND	ND	ND	ND	ND	ND	
	8922	185	ND	ND	ND	ND	NR	ND	
	8916	189	ND	ND	ND	ND	49.6 E	ND	
	8908	194	ND	ND	ND	ND	NR	ND	
<i>Upper Slope</i>	8928	223	ND	ND	ND	ND	37.4 E	ND	
	8920	270	ND	ND	ND	ND	NR	ND	
	8936	292	ND	ND	ND	ND	51 E	ND	
	8927	460	ND	ND	ND	ND	46.2 E	40.8 E	
	8934	491	ND	ND	ND	ND	ND	ND	
	8923	523	ND	ND	ND	ND	71.2 E	ND	
Detection Rate (%)			0	0	0	0	0	25	8
ERL ^a :			—	—	—	—	—	—	—
ERM ^a :			—	—	—	—	—	—	—

^aFrom Long et al. 1995

Appendix B.15

Concentrations of PCBs (ppt) detected in sediments from PLOO stations sampled during 2020. See Appendix B.1 for MDLs; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

	PCB Congener																					
	2020	tPCB	18	28	37	44	49	52	66	70	74	77	81	87	99	101	105	110	114	118	119	
<i>88-m Depth Contour</i>																						
B11	1074	21.8 E	33 E	ND	20.7 E	26.2 E	33.3 E	29	26.7	12.7 E	ND	ND	22.4 E	43.5	61.1	27.7 E	56.1	ND	ND	70.8	ND	
B8	1097.8	14.4 E	22 E	ND	20.4 E	22.7 E	35.1	31.8	30.8	12.1 E	ND	ND	30.2 E	49.1	76.3	31.8 E	82.6	ND	ND	82.4	ND	
E19	751.4	11.3 E	22 E	15.1 E	16.4 E	22.3 E	20.5 E	26.8	21.6 E	15.3 E	ND	ND	17.7 E	34.2	43.2 E	19.1 E	39.6 E	ND	ND	47.4	11.8 E	
E7	832.6	ND	13 E	ND	ND	17.4 E	NR	23.4 E	16.9 E	ND	ND	ND	17.7 E	43.6	48.4 E	21 E	54.9	ND	ND	63	ND	
E1	2716.26	ND	20.1 E	ND	30.3	45.2	61.4	51.4	49.6	18.8	9.46 E	ND	58.6	89	170	69.7	160	ND	ND	180	13.9 E	
<i>98-m Depth Contour</i>																						
B12	301.81	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	16.2 E	16.6 E	10.7 E	18 E	ND	ND	25.6	ND	
B9	475.65	ND	ND	ND	ND	ND	11.1 E	14.4 E	11.3 E	ND	ND	ND	12.8 E	21.2 E	24.4 E	15.1 E	29.2 E	ND	ND	38	ND	
E26	1026.1	ND	10.5 E	ND	ND	13.3 E	NR	19.4 E	14.2 E	ND	ND	ND	13 E	30.3	43.8 E	15.8 E	39.4 E	ND	ND	47.4	ND	
E25	231.81	ND	ND	ND	ND	ND	ND	10.4 E	ND	ND	ND	ND	ND	14.8 E	16.5 E	ND	19.1 E	ND	ND	23.7	ND	
E23	629.6	ND	ND	ND	ND	13.7 E	NR	18.8 E	16.6 E	ND	ND	ND	14.8 E	31.3	42.7 E	20.8 E	45.5	ND	ND	52.8	ND	
E20	358.27	ND	ND	ND	ND	9.77 E	ND	12.5 E	ND	ND	ND	ND	ND	20 E	23.5 E	13.3 E	26.6 E	ND	ND	32.2	ND	
E17 ^a	449.9	ND	9.2 E	ND	ND	ND	15.1 E	11.8 E	12.3 E	ND	ND	ND	16.4 E	19.4 E	32.8 E	15 E	36.8 E	ND	ND	37.6	ND	
E14 ^a	279	ND	ND	ND	ND	ND	NR	11.3 E	ND	ND	ND	ND	ND	17.9 E	20.3 E	ND	21.1 E	ND	ND	27	ND	
E11 ^a	453.65	ND	ND	ND	ND	ND	NR	13.8 E	9.91 E	ND	ND	ND	11.2 E	25	30.2 E	14 E	28.7 E	ND	ND	35.8	ND	
E8	596.07	ND	ND	ND	ND	9.07 E	16 E	14 E	13.7 E	ND	ND	ND	20 E	26.7	44.5 E	22 E	50.1	ND	ND	52.2	ND	
E5	639.9	ND	ND	ND	ND	11.2 E	12.1 E	17.4 E	12.4 E	ND	ND	ND	12.2 E	29.9	35.8 E	21.9 E	44.4	ND	ND	49	ND	
E2	2123.8	14.5 E	23.6 E	ND	26.3	51.2	56.5	54.5	45	13.6 E	ND	ND	45.9	110	150	51.8	150	ND	ND	170	ND	
<i>116-m Depth Contour</i>																						
B10	809.7	ND	15.1 E	13.6 E	17.6 E	19.8 E	27 E	22.8 E	24.6	14.9 E	ND	ND	28.6 E	45.7	59.8	23.8 E	55.7	ND	ND	59.2	ND	
E21	481.2	ND	ND	ND	ND	10.3 E	NR	14.2 E	13.5 E	ND	ND	ND	15.9 E	22.4 E	36.8 E	17.5 E	39.6 E	ND	ND	43.3	ND	
E15 ^a	549.7	9.3 E	ND	ND	ND	12 E	NR	14 E	11 E	ND	ND	ND	12 E	28	33 E	15 E	33 E	ND	ND	39	ND	
E9	1191.6	11 E	16.1 E	10 E	25	25.9 E	41.2	27.4	33.1	15.6	ND	ND	38.2	57.4	90	35.5 E	89	ND	ND	84.8	8.8 E	
E3	4437.76	39.6 E	56.5	13.7 E	94	110	200	140	160	56.8	17 E	ND	130	180	330	140	430	ND	ND	370	20.4 E	
DR (%)	100	32	50	18	36	73	80	95	82	36	9	0	82	100	100	91	100	0	0	100	18	

^a Near-ZID station

Appendix B.15 continued

Winter	PCB Congener																					
	2020	123	126	128	138	149	151	153/168	156	157	158	167	169	170	177	180	183	187	189	194	201	206
<i>88-m Depth Contour</i>																						
B11	ND	ND	30.4 E	98.6	65.9	15 E	141	14.3 E	ND	ND	ND	ND	ND	34.4 E	24.2 E	63.9	16.2 E	57.7	ND	27.4 E	ND	NR
B8	ND	ND	30.1 E	95.8	76.5	17.6 E	120	13.5 E	ND	11.1 E	ND	ND	ND	28.8 E	19.9 E	51.6	16.7 E	49	ND	25.5 E	ND	NR
E19	ND	ND	19.7 E	54.2 E	46.9	14.6 E	90.6 E	ND	ND	ND	ND	ND	ND	20.2 E	14.9 E	41.6 E	12.9 E	34.8 E	ND	16.7 E	ND	NR
E7	ND	ND	24.6 E	79.5	63.6	14.3 E	129	11.3 E	ND	ND	ND	ND	ND	27.5 E	20 E	55.9	16.2 E	48.3	ND	23.1 E	ND	NR
E1	18 E	ND	59.1	240	220	58.7	380	43	ND	23.2 E	13.6 E	ND	ND	100	70.6	200	51.3	160	ND	51.3	ND	NR
<i>98-m Depth Contour</i>																						
B12	ND	ND	12.5 E	40.8 E	30	ND	49.8 E	ND	ND	ND	ND	ND	ND	10.9 E	14.7 E	18.7 E	ND	27.8 E	ND	9.51 E	ND	NR
B9	ND	ND	16.1 E	49.7 E	38.1	ND	63.7 E	ND	ND	ND	ND	ND	ND	18.6 E	16.9 E	35.7 E	9.65 E	33.8 E	ND	15.9 E	ND	NR
E26	ND	ND	20.9 E	79.1	70.1	19 E	138	10.1 E	ND	ND	ND	ND	ND	62	38.4	160	37	85.3	ND	59.1	ND	NR
E25	ND	ND	ND	31.6 E	24.5	ND	37.7 E	ND	ND	ND	ND	ND	ND	10.7 E	11.8 E	ND	ND	21.7 E	ND	9.31 E	ND	NR
E23	ND	ND	17.3 E	65.9	43.2	10 E	91.9 E	ND	ND	ND	ND	ND	ND	19.5 E	14.1 E	44.5 E	12.4 E	35 E	ND	18.8 E	ND	NR
E20	ND	ND	12.7 E	45.3 E	32	ND	63.7 E	ND	ND	ND	ND	ND	ND	ND	10.7 E	28.5 E	ND	27.5 E	ND	ND	ND	NR
E17 ^a	ND	ND	16.1 E	49.3 E	37.4	ND	56.3 E	ND	ND	ND	ND	ND	ND	12.2 E	12.5 E	28.6 E	ND	20.6 E	ND	10.5 E	ND	NR
E14 ^a	ND	ND	9.3 E	32.2 E	24.2	ND	49.2 E	ND	ND	ND	ND	ND	ND	11.1 E	ND	22.1 E	ND	22.5 E	ND	10.8 E	ND	NR
E11 ^a	ND	ND	14.3 E	53.1 E	36.8	ND	72.4 E	ND	ND	ND	ND	ND	ND	15.8 E	12.7 E	28.8 E	9.64 E	28.3 E	ND	13.2 E	ND	NR
E8	ND	ND	19.7 E	68.1	47.3	10.4 E	76.9 E	ND	ND	ND	ND	ND	ND	16.3 E	14.9 E	32.3 E	ND	30.7 E	ND	11.2 E	ND	NR
E5	ND	ND	20.1 E	71.9	55.4	10.6 E	82.4 E	ND	ND	ND	ND	ND	ND	20.6 E	21.9 E	41.6 E	13.6 E	39.7	ND	15.8 E	ND	NR
E2	19.9 E	ND	53.2	190	160	39.9 E	290	22 E	ND	19.5 E	10.5 E	ND	ND	51.5	38.9	100	31.8	97	ND	36.7	ND	NR
<i>116-m Depth Contour</i>																						
B10	ND	ND	21.1 E	68.6	54.7	16.6 E	87.9 E	9.6 E	ND	ND	ND	ND	ND	15.4 E	15.5 E	32.7 E	10.3 E	34.3 E	ND	14.8 E	ND	NR
E21	ND	ND	14.5 E	51.7 E	36.2	ND	73.2 E	ND	ND	ND	ND	ND	ND	14.8 E	10.4 E	26.6 E	ND	26.6 E	ND	13.7 E	ND	NR
E15 ^a	ND	ND	15 E	53 E	39	9.4 E	76 E	ND	ND	ND	ND	ND	ND	18 E	14 E	41 E	11 E	35 E	ND	32	ND	NR
E9	11.1 E	ND	32 E	100	79.4	19.4 E	150	15.2 E	ND	11.4 E	ND	ND	ND	27.4 E	20.5 E	53.5	14.9 E	47.8	ND	ND	ND	NR
E3	37.9	ND	89	330	310	66.5	440	41.5	8.84 E	36.5	18.4 E	ND	ND	72.1	63.9	190	46.4	140	ND	50	8.72 E	NR
DR (%)	18	0	95	100	100	64	100	41	5	23	14	0	95	95	95	95	68	100	0	91	5	NR

^a Near-ZID station

Appendix B.15 *continued*

Summer	PCB Congener																					
	2020	tPCB	18	28	37	44	49	52	66	70	74	77	81	87	99	101	105	110	114	118	119	
88-m Depth Contour																						
B11	421.1	ND	ND	ND	ND	ND	ND	11.9 E	14.4 E	10.7 E	ND	ND	ND	11.2 E	19.5 E	24.3 E	15.9 E	26.8 E	ND	37.2 E	ND	
B8	976.22	ND	ND	ND	10.4 E	19 E	18.3 E	30.8 E	22.1 E	9.82 E	ND	ND	ND	17.2 E	40.8 E	54.6	30.5 E	56.6 E	ND	65.2 E	ND	
E19	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	
E7	943.4	ND	ND	ND	12.3 E	19.1 E	27.2 E	25.8 E	23.1 E	ND	ND	ND	ND	22.6 E	41.6 E	67.6	31.2 E	67.2 E	ND	68.5 E	ND	
E1	4027	12.8 E	ND	ND	54.8 E	76.9 E	130	92	96	34.3 E	ND	ND	ND	120	170	340	130 E	320	ND	300	24.3 E	
98-m Depth Contour																						
B12	318.3	ND	ND	ND	ND	ND	ND	11.2 E	ND	ND	ND	ND	ND	ND	15.8 E	23.7 E	11.3 E	22.5 E	ND	27.5 E	ND	
B9	492.85	ND	ND	ND	ND	13.3 E	NA	18.5 E	10.7 E	ND	ND	ND	ND	10.2 E	25.9 E	34.2 E	16.5 E	31.2 E	ND	39.8 E	ND	
E26	664.6	ND	ND	ND	ND	ND	13.3 E	18.3 E	13.4 E	ND	ND	ND	ND	14.9 E	28.4 E	45.1 E	18.5 E	45.3 E	ND	49.7 E	ND	
E25	310.69	ND	ND	ND	ND	11.8 E	11.6 E	15.1 E	9.99 E	ND	ND	ND	ND	ND	16.6 E	25 E	11.2 E	21.7 E	ND	30.4 E	ND	
E23	215.7	ND	ND	ND	ND	ND	ND	10.6 E	ND	ND	ND	ND	ND	ND	12.9 E	16.6 E	ND	15.6 E	ND	22.6 E	ND	
E20	276.8	ND	11.9 E	ND	ND	ND	ND	ND	13.5 E	ND	ND	ND	ND	ND	14.3 E	17.8 E	ND	20.3 E	ND	26.8 E	ND	
E17 ^a	177.68	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	9.54 E	14 E	ND	13.9 E	ND	19.9 E	ND	
E14 ^a	94.6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	11.4 E	ND	9.5 E	ND	13.7 E	ND	
E11 ^a	262.7	ND	ND	ND	ND	ND	ND	10.6 E	ND	ND	ND	ND	ND	ND	13.1 E	21.1 E	11.1 E	22.2 E	ND	27.1 E	ND	
E8	490	ND	ND	ND	ND	ND	11.1 E	NA	16.4 E	10.5 E	ND	ND	ND	ND	25.4 E	33.1 E	17.1 E	33.4 E	ND	41.7 E	ND	
E5	1148.53	ND	ND	ND	14.4 E	25.7 E	31 E	28.7 E	24.1 E	ND	ND	ND	ND	24.5 E	57.1 E	79.2	30.4 E	73.6 E	ND	79.3 E	ND	
E2	2404.8	10.8 E	20.2 E	ND	30.5 E	51.9 E	67.4 E	59 E	51 E	17.3 E	ND	ND	ND	59 E	110	190	70.9 E	190	ND	180	16.7 E	
116-m Depth Contour																						
B10	167.26	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	10.3 E	12.8 E	ND	13.5 E	ND	19.4 E	ND	
E21	789.4	ND	ND	ND	26.2 E	ND	57.5 E	18.1 E	34.2 E	ND	ND	ND	ND	38.5	34.3 E	82.7	35.7 E	87.1	ND	80.5	ND	
E15 ^a	342.2	ND	ND	ND	ND	ND	15.7 E	11 E	10.5 E	ND	ND	ND	ND	10.8 E	14.7 E	30.5 E	13.1 E	30.6 E	ND	29.8 E	ND	
E9	1402.34	9.14 E	ND	ND	30.3 E	33.1 E	59.1 E	37.4 E	44.5 E	17.4 E	ND	ND	ND	42.8 E	60.1	120	58.1 E	110	ND	100	ND	
E3	4563.3	49.2 E	61.3 E	13.1 E	120	120	220	150	170	64.2 E	13.6 E	ND	ND	140	180	390	140	410	ND	330	24.1 E	
DR (%)	100	19	14	5	38	48	68	76	71	24	5	0	57	95	100	76	100	100	0	100	14	

^aNear-ZID station

Appendix B.15 *continued*

		PCB Congener																					
Summer	2020	123	126	128	138	149	151	153/168	156	157	158	167	169	170	177	180	183	187	189	194	201	206	
<i>88-m Depth Contour</i>																							
B11	ND	ND	ND	55.3 E	33.8 E	ND	ND	67.6 E	ND	ND	ND	ND	ND	16.7 E	12.1 E	25.1 E	ND	25.9 E	ND	12.7 E	ND	ND	
B8	ND	ND	29.9 E	89.2	56.8 E	15.3 E	133 E	11.4 E	ND	ND	ND	ND	ND	31 E	22.6 E	57.1	17.4 E	51.4	ND	26.6 E	ND	59.2 E	
E19	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
E7	ND	ND	25.7 E	89.4	58.2 E	14.9 E	125 E	10.9 E	ND	10 E	ND	ND	ND	29.6 E	19.9 E	62.7	18.9 E	45.9	ND	26.1 E	ND	NR	
E1	28.2 E	ND	96	370	260	69.7	520	43.9 E	10.2 E	42 E	20.5 E	ND	87.9	58	190	59.9	140	ND	55.8 E	10 E	63.8 E		
<i>98-m Depth Contour</i>																							
B12	ND	ND	ND	41.8 E	35.2 E	12 E	56.5 E	ND	ND	ND	ND	ND	ND	10.7 E	12.5 E	15.5 E	ND	22.1 E	ND	ND	ND	NR	
B9	ND	ND	17.9 E	58 E	37 E	ND	72.7 E	ND	ND	ND	ND	ND	ND	15.5 E	13.4 E	28.7 E	9.65 E	26.7 E	ND	13 E	ND	NR	
E26	ND	ND	22.6 E	72.8	52.5 E	11.7 E	101 E	9.6 E	ND	ND	ND	ND	ND	24 E	15 E	49.8	11.7 E	29.4 E	ND	17.6 E	ND	NR	
E25	ND	ND	12.4 E	40.9 E	24.6 E	ND	35 E	ND	ND	ND	ND	ND	ND	ND	ND	23.6 E	ND	20.8 E	ND	ND	ND	NR	
E23	ND	ND	ND	31.3 E	22.1 E	ND	38.4 E	ND	ND	ND	ND	ND	ND	11.8 E	ND	17.2 E	ND	16.6 E	ND	ND	ND	NR	
E20	ND	ND	ND	32.8 E	24 E	ND	43.4 E	ND	ND	ND	ND	ND	ND	13.3 E	ND	27.1 E	ND	17.5 E	ND	14.1 E	ND	NR	
E17 ^a	ND	ND	9.13 E	25.3 E	17.3 E	ND	33.2 E	ND	ND	ND	ND	ND	ND	9.41 E	ND	14.2 E	ND	11.8 E	ND	ND	ND	ND	
E14 ^a	ND	ND	ND	17.1 E	12.3 E	ND	21.4 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	9.2 E	ND	ND	ND	ND	
E11 ^a	ND	ND	10.1 E	34.1 E	22.7 E	ND	45.1 E	ND	ND	ND	ND	ND	ND	11.8 E	ND	15.9 E	ND	17.8 E	ND	ND	ND	ND	
E8	ND	ND	17.8 E	58.7 E	38.6 E	9.3 E	79.7 E	ND	ND	ND	ND	ND	ND	11.8 E	ND	29.6 E	10.4 E	30 E	ND	13.8 E	ND	NR	
E5	9.83 E	ND	30.7 E	100	91.2 E	19.7 E	162 E	11.7 E	ND	11.2 E	ND	ND	ND	31 E	24.6 E	73.6	22.2 E	63.2	ND	29.6 E	ND	NR	
E2	21 E	ND	63.5	220	150	37.9 E	310	25.8 E	ND	24.8 E	13.2 E	ND	51.9	39.4 E	110	35.2	95	ND	34.3 E	ND	48.1 E		
<i>116-m Depth Contour</i>																							
B10	ND	ND	ND	30.6 E	19.3 E	ND	35.7 E	ND	ND	ND	ND	ND	ND	ND	ND	9.46 E	ND	16.2 E	ND	ND	ND	ND	
E21	ND	ND	19 E	67.1 E	50.3 E	12.1 E	76.4 E	11.7 E	ND	11.9 E	ND	ND	ND	11.6 E	ND	21.3 E	ND	13.2 E	ND	ND	ND	ND	
E15 ^a	ND	ND	14 E	40 E	29.3 E	ND	50.5 E	ND	ND	ND	ND	ND	ND	10.3 E	ND	15.8 E	ND	15.6 E	ND	ND	ND	ND	
E9	10.9 E	ND	34.9 E	120	84.7 E	21.9 E	165 E	14.6 E	ND	13.8 E	ND	ND	ND	34.9 E	22.5 E	60.4	19.6 E	53.1	ND	24.1 E	ND	NR	
E3	31.2 E	ND	96	360	280	67.6	470	40.2 E	10.1 E	39.6 E	19.6 E	ND	76.5	51.8 E	160	45	120	ND	48.4 E	ND	51.8 E		
DR (%)	24	0	71	100	100	52	100	43	10	33	14	0	81	57	95	48	100	0	57	5	33		

^aNear-ZID station

Appendix B.16

Concentrations of PCBs (ppt) detected in sediments from SBOO stations sampled during 2020. See Appendix B.1 for MDLs; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

	PCB Congener																					
	1PCB	18	28	37	44	49	52	66	70	74	77	81	87	99	101	105	110	114	118	119		
Winter 2020																						
<i>19-m Stations</i>																						
I35	490.9	ND	ND	ND	11 E	10.1 E	22 E	12.4 E	17 E	ND	ND	ND	19.9 E	27.2	45.7 E	24.4 E	49.2	ND	46	ND	ND	
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I31	163.85	16.6 E	17.9 E	18.1 E	12.1 E	11.8 E	13.7 E	11.5 E	10.3 E	11.1 E	ND	ND	ND	ND	9.95 E	ND	ND	ND	ND	ND	ND	
I23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I18	9.43	ND	ND	ND	ND	ND	9.43 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>28-m Stations</i>																						
I33	494.17	14.1 E	16.6 E	16.7 E	14.3 E	15.9 E	16.8 E	18 E	14.8 E	14 E	9.11 E	ND	11.4 E	18.6 E	21.7 E	ND	16.8 E	ND	17.8 E	10.1 E	ND	
I30	30.8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I27	131.14	12.3 E	ND	10.4 E	14.5 E	13.6 E	16.8 E	11.7 E	11.6 E	ND	ND	ND	ND	ND	9.51 E	ND	ND	ND	ND	ND	ND	
I22	59.21	12.2 E	ND	9.04 E	9.1 E	9.63 E	10.7 E	8.54 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I16 ^a	350.86	10.5 E	13.8 E	9.29 E	14 E	13 E	17.6 E	13.4 E	15 E	10.4 E	ND	ND	14.5 E	14.6 E	26.1 E	10.4 E	26 E	ND	22.9	ND	ND	
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I2	25.63	ND	ND	ND	8.83 E	ND	8.42 E	8.38 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>38-m Stations</i>																						
I29	455.1	13.7 E	15.5 E	ND	ND	14.8 E	15.1 E	14.2 E	10.5 E	ND	ND	ND	ND	20.5 E	26.9 E	ND	22.7 E	ND	27.7	ND	ND	
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>55-m Stations</i>																						
I28	1211.22	ND	ND	9.72 E	13.8 E	35.7	27 E	30.7	21.8	11.4 E	ND	ND	20.6 E	70.3	69.1	34.7 E	62.3	ND	77.5	ND	ND	
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
DR (%)	41	22	15	22	30	30	33	37	26	15	4	0	15	19	26	11	19	0	19	0	19	4

^aNear-ZID station

Appendix B.16 *continued*

		PCB Congener																				
Winter 2020		123	126	128	138	149	151	153/168	156	157	158	167	169	170	177	180	183	187	189	194	201	206
<i>19-m Stations</i>																						
I35		ND	ND	14 E	44.5 E	43.7	ND	86.2 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	17.6 E	ND	ND	ND	NR
I34		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31		ND	ND	ND	ND	10 E	ND	20.8 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR
I23		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR
I10		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR
I4		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>28-m Stations</i>																						
I33		ND	ND	9.46 E	23.8 E	28.6	11.3 E	65.8 E	ND	ND	ND	ND	ND	ND	11.6 E	26.7 E	10.1 E	30.6 E	ND	29.5	ND	NR
I30		ND	ND	ND	ND	10.1 E	ND	20.7 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I27		ND	ND	ND	ND	9.93 E	ND	20.8 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I22		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I14 ^a		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I16 ^a		ND	ND	ND	27.3 E	22	9.94 E	35.1 E	ND	ND	ND	ND	ND	ND	15.3 E	ND	9.73 E	ND	ND	ND	ND	NR
I15		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I9		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I6		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>38-m Stations</i>																						
I29		ND	ND	13.6 E	40 E	40.4	ND	91.6 E	ND	ND	ND	ND	ND	12.9 E	12.8 E	25.7 E	ND	24.3 E	ND	12.2 E	ND	NR
I21		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR
<i>55-m Stations</i>																						
I28		14.4 E	ND	32.4 E	92	120	18.4 E	230	16.8 E	ND	16.9 E	9.6 E	ND	24.4 E	21.5 E	47.5	17 E	45.3	ND	20.4 E	ND	NR
I20		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR
I1		ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)		4	0	15	19	30	11	30	4	0	4	4	0	7	11	15	7	19	0	11	0	0

^aNear-ZID station

Appendix B.16 *continued*

Summer 2020		PCB Congener																			
	tPCB	18	28	37	44	49	52	66	70	74	77	81	87	99	101	105	110	114	118	119	
<i>19-m Stations</i>																					
I35	619.5	ND	ND	ND	NR 16.2 E	36.1 E	18.7 E	22.5 E	ND	ND	ND	ND	26.9 E	28.5 E	63.1	20.3 E	59.4 E	ND	59.5 E	ND	
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I31	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I23	18.59	ND	ND	ND	ND 9.13 E	NR	ND	ND	ND	ND	ND	ND	ND	ND 9.46 E	ND	ND	ND	ND	ND	ND	
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I4	ND	ND	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>28-m Stations</i>																					
I33	399.45	ND	11.7 E	ND	NR 14 E	NR	18 E	NR	ND	ND	ND	ND	10.1 E	18.4 E	28.2 E	ND	21.5 E	ND	24.3 E	ND	
I30	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I27	130.5	NR 13.9 E	9.9 E	NR 13.1 E	NR 12.5 E	NR	ND	NR	ND	ND	ND	ND	ND	12.6 E	18.3 E	ND	10.8 E	ND	10.8 E	ND	
I22	48.31	ND	ND	ND	ND 9.91 E	13.5 E	11.2 E	13.7 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>38-m Stations</i>																					
I29	472.7	ND	ND	ND	NR 14.9 E	NR 17.1 E	NR	NR	ND	ND	ND	ND	12 E	21 E	34 E	10.9 E	27 E	ND	33.6 E	ND	
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I13	18.28	ND	9.34 E	ND	8.94 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>55-m Stations</i>																					
I28	1880.76	ND	ND	ND	30.7 E	32.1 E	77.8 E	26 E	43.3 E	10.9 E	ND	ND	61.8 E	65.6	140	50.5 E	150	ND	130	ND	
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I7	19.24	NR	ND	ND	NR 10.8 E	NR 8.44 E	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
DR (%)	33	0	11	4	9	30	14	26	13	4	0	0	15	19	22	11	19	0	19	0	

^a Near-ZID station

Appendix B.16 *continued*

		PCB Congener																			
Summer 2020	123	126	128	138	149	151	153/168	156	157	158	167	169	170	177	180	183	187	189	194	201	206
<i>19-m Stations</i>																					
I35	ND	ND	12.1 E	58.7 E	44.6 E	NR	77 E	ND	ND	ND	ND	ND	13.4 E	9.5 E	23.4 E	ND	18.5 E	ND	11.1 E	ND	ND
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>28-m Stations</i>																					
I33	ND	ND	ND	ND	NR	ND	51.6 E	ND	ND	ND	ND	ND	10.3 E	9.65 E	53.4	10.9 E	33 E	ND	38.6 E	ND	45.8 E
I30	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I27	ND	ND	ND	10.4 E	NR	ND	18.2 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I22	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I14 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I9	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>38-m Stations</i>																					
I29	ND	ND	11.7 E	48.5 E	36 E	10.7 E	74.1 E	ND	ND	ND	ND	ND	17.2 E	12.3 E	37.8 E	11.1 E	26.5 E	ND	16.3 E	ND	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>55-m Stations</i>																					
I28	12.3 E	ND	35.6 E	130	150	43.8	189 E	16.6 E	ND	16.9 E	8.86 E	ND	62.2	43 E	160	41.4	100	ND	52.4 E	ND	NR
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	ND	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	4	0	11	15	13	8	19	4	0	4	4	0	15	15	15	11	15	0	15	0	4

^aNear-ZID station

Appendix B.17

Concentrations of PCBs (ppt) detected in sediments from the San Diego regional benthic stations sampled during summer 2020. See Appendix B.1 for MDLs; E = estimated; ND = not detected; NR = not reportable.

	Station	Depth (m)	PCB Congener											
			tPCB	8	18	28	37	44	49	52	66	70	74	77
<i>Inner Shelf</i>	8929	7	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8944	7	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8915	16	180.8	ND	ND	ND	ND	ND	ND	8.7 E	10.4 E	ND	ND	ND
	8907	18	ND	ND	NR	ND	ND	ND	ND	NR	ND	ND	ND	ND
	8943	19	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8914	20	137	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8911	24	29.21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8938	28	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
8901	29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	38.11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8939	65	87.68	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8903	66	148.5	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8912	67	13650.8	ND	37.5 E	43.4	10.8 E	290	370	800	250	420	77.9	23.9 E
	8925	67	1100.1	NR	ND	20.1 E	ND	12.4 E	23.9 E	22.7 E	35.7 E	20.9 E	12 E	ND
	8919	68	1612.2	NR	18 E	31.4 E	ND	23.6 E	42.6 E	40.3 E	52.8 E	33.3 E	16.6 E	ND
	8909	73	4005.7	ND	12.9 E	23.1 E	ND	48.4 E	92	130	88.9	84.9	25.3 E	12.1 E
	8931	74	816.9	NR	ND	18 E	ND	10.8 E	22.1 E	19.6 E	32.2 E	20.1 E	ND	ND
	8933	80	775.2	NR	ND	ND	ND	ND	19 E	18.3 E	28.8 E	19.3 E	ND	ND
	8930	83	901.6	NR	ND	19.2 E	ND	ND	ND	26.9 E	35.1 E	27.9 E	11.8 E	ND
	8949	83	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	8924	85	943.67	NR	ND	15.5 E	ND	11.6 E	24.5 E	23.9 E	33 E	20.2 E	ND	ND
	8926	87	493.04	NR	ND	ND	ND	ND	10.9 E	NR	18.4 E	9.84 E	ND	ND
	8932	89	1050.5	NR	ND	18.7 E	10.7 E	15 E	27.3 E	28 E	37 E	25.9 E	13.3 E	12.5 E
8940	108	57.43	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
8921	116	311	NR	ND	ND	ND	ND	ND	ND	12 E	4.5 E	ND	ND	
<i>Outer Shelf</i>	8905	134	1460.32	ND	ND	14 E	ND	18 E	29 E	34.5 E	33 E	24 E	10.4 E	ND
	8937	137	608.93	NR	ND	ND	ND	9.33 E	10.7 E	19.8 E	16.2 E	15 E	ND	ND
	8910	169	8336.7	NR	29 E	62.7	ND	140	190	330	220	210	68.6	21.9 E
	8922	185	1754.9	NR	ND	16.1 E	ND	18.9 E	33.2 E	38 E	41.4 E	34.1 E	14.4 E	ND
	8916	189	6152.3	NR	ND	38.4 E	17.5 E	51 E	97.8	110	120	97.5	42.9 E	25.3 E
	8908	194	3047.9	ND	ND	25.5 E	11.4 E	39.5 E	53.6	82.4 E	61.6 E	66.6 E	23.6 E	14.4 E
<i>Upper Slope</i>	8928	223	1807	NR	ND	19.2 E	ND	20.2 E	39 E	46.4 E	47.7 E	40.3 E	15.4 E	ND
	8920	270	1392.2	NR	ND	19.4 E	ND	ND	ND	34.7 E	46 E	32.8 E	15.8 E	12.4 E
	8936	292	1381	NR	ND	ND	ND	17.4 E	27.1 E	30.1 E	42.6 E	32.2 E	17.6 E	ND
	8927	460	1224.7	NR	ND	19.2 E	ND	23.9 E	35.4 E	40.2 E	47.2 E	35.5 E	18.7 E	ND
	8934	491	1650	NR	ND	ND	ND	31.9 E	45.7 E	52.2 E	55.3 E	46.5 E	21.9 E	ND
	8923	523	1113.6	NR	ND	17.4 E	ND	18.7 E	33.2 E	34.4 E	38.7 E	28.9 E	14.7 E	ND
Detection Rate (%)			77	0	11	44	10	46	51	59	62	59	44	18

Appendix B.17 *continued*

	Station	Depth (m)	PCB Congener									
			81	87	99	101	105	110	114	118	119	123
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8944	7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	14.2 E	18.6 E	ND	16.4 E	ND	14.3 E	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	11.1 E	13.6 E	ND	12.9 E	ND	16.4 E	ND	ND
	8911	24	ND	ND	ND	ND	ND	ND	ND	9.61 E	ND	ND
	8938	28	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
8901	29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND	ND	9.51 E	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	12.9 E	ND	9.18 E	ND	12.7 E	ND	ND
	8903	66	ND	ND	ND	10.5 E	ND	9.68 E	ND	13.2 E	ND	ND
	8912	67	ND	500	600	1400	430	1600	22.6 E	1300	71.4	100
	8925	67	ND	19 E	52.3 E	63	32.8 E	65.2 E	ND	73.5 E	ND	ND
	8919	68	ND	30.8 E	78.5	100	43.1 E	100	ND	120	ND	11.3 E
	8909	73	ND	110	180	340	110	370	ND	360	22.1 E	33.4 E
	8931	74	ND	ND	36.7 E	44.6 E	24.3 E	44.7 E	ND	68.3 E	ND	ND
	8933	80	ND	17.8 E	37 E	44.9 E	24.8 E	44.6 E	ND	60.6 E	ND	ND
	8930	83	ND	19.8 E	32.6 E	53.2 E	28.5 E	60	ND	71.6 E	ND	11.1 E
	8949	83	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	8924	85	ND	17.3 E	44.5 E	60.9	27.7 E	59.3 E	ND	76.6 E	ND	ND
	8926	87	ND	ND	28 E	25.9 E	15.1 E	30.9 E	ND	39.3 E	ND	ND
	8932	89	ND	24.2 E	47.6 E	60	33.3 E	59.2 E	ND	78.2 E	ND	12.4 E
	8940	108	ND	ND	ND	9.12 E	ND	ND	ND	10.3 E	ND	ND
8921	116	ND	4.9 E	19 E	25 E	11 E	22 E	ND	29 E	ND	ND	
<i>Outer Shelf</i>	8905	134	ND	29.6 E	63.7	88.3	41.9 E	110	ND	110	ND	12.5 E
	8937	137	ND	21.2 E	29 E	54.7	23.1 E	57.8 E	ND	52.7 E	ND	ND
	8910	169	ND	200	420	710	190	700	ND	670	38.4 E	58.5 E
	8922	185	ND	38.1 E	75.3	110	55.3 E	120	ND	120	ND	12.5 E
	8916	189	ND	140	230	390	180	440	ND	490	25.4 E	46.5 E
	8908	194	ND	82.1	120	210	97.5	250	ND	260	ND	25.7 E
<i>Upper Slope</i>	8928	223	ND	40.1 E	72.2	120	60.9 E	110	ND	140	ND	15.3 E
	8920	270	ND	27.4 E	52.4 E	81.8	48.5 E	89.8	ND	116	ND	14.6 E
	8936	292	ND	30.2 E	60.7 E	89.3	46.7 E	84.2 E	ND	100	ND	11.3 E
	8927	460	ND	34.6 E	54.6 E	90.2	48 E	88.2 E	ND	97.8 E	ND	ND
	8934	491	ND	45.4 E	78.9 E	122	60.6 E	124 E	ND	127 E	ND	ND
	8923	523	ND	28 E	45.1 E	72.7	45.8 E	72.3 E	ND	90.4 E	ND	ND
Detection Rate (%)			0	54	64	72	59	69	3	77	10	33

Appendix B.17 *continued*

	Station	Depth (m)	PCB Congener										
			126	128	138	149	151	153/168	156	157	158	167	
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8944	7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	20.5 E	19.4 E	ND	32.5 E	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	23.6 E	16.1 E	ND	30.1 E	ND	ND	ND	ND	ND
	8911	24	ND	ND	ND	NR	ND	19.6 E	ND	ND	ND	ND	ND
	8938	28	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
8901	29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	ND	ND	ND	10.2 E	ND	18.4 E	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	16.7 E	12.5 E	ND	23.7 E	ND	ND	ND	ND	ND
	8903	66	ND	ND	23 E	17.4 E	ND	29.3 E	ND	ND	ND	ND	ND
	8912	67	ND	310	1100	930	180	1300	150	38.3 E	140	57 E	
	8925	67	ND	31 E	100	80.2 E	18.2 E	149 E	12.1 E	ND	10.7 E	ND	
	8919	68	ND	41 E	140	94.8 E	21.7 E	194 E	16.7 E	ND	14.8 E	ND	
	8909	73	ND	110	380	300	66.9	490	48.4 E	ND	41.6 E	20.5 E	
	8931	74	ND	27.1 E	87.2	53.4 E	12.3 E	83.9 E	ND	ND	ND	ND	
	8933	80	ND	26.1 E	89	53.1 E	12.1 E	78.2 E	ND	ND	ND	ND	
	8930	83	ND	ND	82	62.8 E	17.1 E	112 E	13.3 E	ND	12 E	ND	
	8949	83	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	8924	85	ND	29.4 E	91.1	69.3 E	15 E	89.4 E	9.59 E	ND	9.58 E	ND	
	8926	87	ND	14.7 E	53.6 E	34 E	ND	76.1 E	ND	ND	ND	ND	
	8932	89	ND	34.2 E	100	71.8 E	18.9 E	103 E	11 E	ND	ND	ND	
	8940	108	ND	ND	9.61 E	10.2 E	ND	18.2 E	ND	ND	ND	ND	
8921	116	ND	12 E	38 E	27 E	ND	50 E	ND	ND	ND	ND		
<i>Outer Shelf</i>	8905	134	ND	49.7 E	160	140	31.8 E	210 E	13.9 E	ND	11 E	9.62 E	
	8937	137	ND	22 E	70.8	49.2 E	10.5 E	81.8 E	ND	ND	ND	ND	
	8910	169	ND	180	680	640	120	1100	69.7 E	ND	66 E	34.4 E	
	8922	185	ND	48.6 E	180	118 E	29.6 E	249 E	19.7 E	ND	13.4 E	ND	
	8916	189	ND	190	630	530	120	870	71.5 E	ND	ND	36.6 E	
	8908	194	ND	82.4	290	250	56.3 E	400	30.4 E	11.7 E	20.7 E	17.3 E	
<i>Upper Slope</i>	8928	223	ND	51 E	180	128 E	35 E	165 E	18.4 E	ND	13.7 E	11.7 E	
	8920	270	ND	45 E	150	113 E	30.6 E	186	ND	ND	ND	ND	
	8936	292	ND	45.3 E	140	94.2 E	22.6 E	183 E	13.5 E	ND	ND	ND	
	8927	460	ND	34.9 E	126	91.4 E	20.8 E	93.7 E	ND	ND	ND	ND	
	8934	491	ND	43.5 E	160	109 E	26.5 E	202 E	ND	ND	ND	ND	
	8923	523	ND	32.3 E	114	80.8 E	18.6 E	83.6 E	ND	ND	ND	ND	
Detection Rate (%)			0	56	72	76	54	77	36	5	28	18	

Appendix B.17 *continued*

	Station	Depth (m)	PCB Congener										
			169	170	177	180	183	187	189	194	195	201	206
<i>Inner Shelf</i>	8929	7	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
	8944	7	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
	8915	16	ND	ND	ND	14.2 E	ND	11.6 E	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
	8914	20	ND	ND	ND	ND	ND	13.2 E	ND	ND	ND	ND	ND
	8911	24	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8938	28	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
8901	29	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
	8903	66	ND	9.22 E	ND	20.1 E	ND	16.1 E	ND	ND	ND	ND	ND
	8912	67	ND	180	130	320	92.6	210	ND	85	ND	NR	80.4 E
	8925	67	ND	35.2 E	24.7 E	73.9	22.2 E	58.6	ND	30.8 E	NR	ND	NR
	8919	68	ND	38.2 E	27.1 E	72.3	23.6 E	63	ND	29.1 E	NR	ND	93.6
	8909	73	ND	83.6	49.4 E	170	42.4	110	ND	NR	ND	ND	49.8 E
	8931	74	ND	33.2 E	22.7 E	60.4	18.4 E	46.3	ND	30.6 E	NR	ND	NR
	8933	80	ND	31.5 E	23 E	55.5	16.6 E	48.4	ND	26.6 E	NR	ND	NR
	8930	83	ND	30.6 E	22.5 E	57.1 E	20.2 E	49.2 E	ND	25.1 E	NR	ND	NR
	8949	83	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	8924	85	ND	32.6 E	22.9 E	66.2	19.4 E	49.3	ND	24.9 E	NR	ND	NR
	8926	87	ND	20.5 E	13.9 E	40.9 E	12 E	32.5 E	ND	16.5 E	NR	ND	NR
	8932	89	ND	32 E	26.8 E	51.4	18.9 E	53	ND	26.2 E	NR	ND	NR
	8940	108	ND	ND	ND	ND	ND	ND	ND	ND	NR	ND	ND
8921	116	ND	11 E	ND	22 E	ND	19 E	ND	4.6 E	NR	ND	ND	
<i>Outer Shelf</i>	8905	134	ND	36.1 E	29.5 E	65.4	14.2 E	70.2	ND	NR	ND	ND	NR
	8937	137	ND	12.1 E	11.8 E	16.8 E	ND	24.4 E	ND	ND	NR	ND	ND
	8910	169	ND	180	120	350	86.7	310	11.5 E	110	NR	19.3 E	NR
	8922	185	ND	44.9 E	39.2 E	76.3	24.7 E	92.7	ND	36.9 E	NR	ND	54.6 E
	8916	189	ND	170	120	340	81.4	310	ND	120	NR	20.5 E	NR
	8908	194	ND	65.3	56.5 E	110	29.9 E	130	ND	NR	ND	ND	73.5 E
<i>Upper Slope</i>	8928	223	ND	52.2	40.6 E	98.4	24.6 E	94.5	ND	40.7 E	NR	ND	66.5 E
	8920	270	ND	44.5 E	31.9 E	67.7 E	20.7 E	79	ND	32.2 E	NR	ND	NR
	8936	292	ND	32.1 E	32.9 E	65.1	14.4 E	76.6	ND	27.6 E	NR	ND	44.3 E
	8927	460	ND	28.3 E	26.1 E	42.9 E	ND	57 E	ND	20.7 E	NR	ND	49.4 E
	8934	491	ND	28.7 E	29.4 E	55.7 E	16.5 E	71.4	ND	34.9 E	NR	ND	61 E
	8923	523	ND	27.9 E	26.2 E	54.6 E	13.3 E	55.5	ND	25.1 E	NR	ND	41.4 E
Detection Rate (%)			0	62	56	64	51	67	3	53	0	5	36

Appendix B.18

Concentrations of PAHs (ppb) detected in sediments from PLOO stations sampled during 2020. See Appendix B.1 for MDLs; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

Winter 2020	tPAH	1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo[A] anthracene	Benzo[A] pyrene
<i>88-m Depth Contour</i>												
B11	57.96	ND	ND	ND	ND	6.84 E	ND	ND	ND	ND	ND	ND
B8	31.16	ND	ND	ND	ND	6.1 E	ND	ND	ND	ND	ND	ND
E19	10.73	ND	ND	ND	ND	4.21 E	ND	ND	ND	ND	ND	ND
E7	22.86	ND	ND	ND	ND	6.98 E	ND	ND	ND	ND	ND	ND
E1	111.35	ND	ND	ND	ND	6.56 E	18.9	ND	ND	ND	ND	16.2
<i>98-m Depth Contour</i>												
B12	11.01	ND	ND	ND	ND	3.94 E	ND	ND	ND	ND	ND	ND
B9	32.5	ND	2.87 E	3.05 E	ND	8.35 E	ND	ND	2.45 E	ND	ND	ND
E26	15.61	ND	ND	ND	ND	6.68 E	ND	ND	ND	ND	ND	ND
E25	12.94	ND	ND	ND	ND	6.64 E	ND	ND	ND	ND	ND	ND
E23	15.95	ND	ND	ND	ND	6.1 E	ND	ND	ND	ND	ND	ND
E20	13.2	ND	ND	ND	ND	5.95 E	ND	ND	ND	ND	ND	ND
E17 ^a	13.81	ND	ND	ND	ND	5.74 E	ND	ND	ND	ND	ND	ND
E14 ^a	13.75	ND	ND	ND	ND	4.45 E	ND	ND	ND	ND	ND	ND
E11 ^a	10.76	ND	ND	ND	ND	5.37 E	ND	ND	ND	ND	ND	ND
E8	12.35	ND	ND	ND	ND	5.48 E	ND	ND	ND	ND	ND	ND
E5	26.59	ND	ND	ND	ND	6.96 E	ND	ND	ND	ND	ND	ND
E2	199.14	ND	ND	ND	ND	7.01 E	32.6	ND	5.59 E	ND	ND	26.5
<i>116-m Depth Contour</i>												
B10	11.02	ND	ND	ND	ND	5.11 E	ND	ND	ND	ND	ND	ND
E21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E15 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E9	13.88	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
E3	146.73	ND	ND	ND	ND	4.41 E	25	ND	2.98 E	ND	ND	17.6
DR (%)	91	0	5	5	0	86	14	0	14	0	0	14
ERL ^b	4022	—	—	70	—	—	—	16	44	85.3	261	430
ERM ^b	44,792	—	—	670	—	—	—	500	640	1100	1600	1600

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.18 *continued*

Winter 2020	Benzo[e] pyrene	Benzo[G,H,I] perylene	Benzo[K] fluoranthene	Biphenyl	Chrysene	Dibenzo(A,H) anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-CD) pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
<i>88-m Depth Contour</i>													
B11	8.1	11.2	ND	NR	4.22 E	ND	6.92 E	ND	9.64	3.49 E	ND	ND	7.55 E
B8	ND	6.3 E	ND	NR	3.96 E	ND	7.13 E	ND	ND	ND	ND	ND	7.67 E
E19	ND	ND	ND	NR	ND	ND	2.7 E	ND	ND	ND	ND	ND	3.82 E
E7	ND	ND	ND	NR	ND	ND	4.87 E	ND	5 E	ND	ND	ND	6.01 E
E1	10.8	12.5	7.42	NR	ND	ND	10.9	ND	9.53	ND	ND	7.04 E	11.5
<i>98-m Depth Contour</i>													
B12	ND	ND	ND	NR	ND	ND	2.97 E	ND	ND	ND	ND	ND	4.1 E
B9	ND	ND	ND	NR	ND	ND	5.49 E	ND	ND	5.06 E	ND	ND	5.23 E
E26	ND	ND	ND	NR	ND	ND	4.21 E	ND	ND	ND	ND	ND	4.72 E
E25	ND	ND	ND	NR	ND	ND	2.92 E	ND	ND	ND	ND	ND	3.38 E
E23	ND	ND	ND	NR	ND	ND	4.85 E	ND	ND	ND	ND	ND	5 E
E20	ND	ND	ND	NR	ND	ND	3.85 E	ND	ND	ND	ND	ND	3.4 E
E17 ^a	ND	ND	ND	NR	ND	ND	4.07 E	ND	ND	ND	ND	ND	4 E
E14 ^a	ND	ND	ND	NR	ND	ND	5.18 E	ND	ND	ND	ND	ND	4.12 E
E11 ^a	ND	ND	ND	NR	ND	ND	1.88 E	ND	ND	ND	ND	ND	3.51 E
E8	ND	ND	ND	NR	ND	ND	3.36 E	ND	ND	ND	ND	ND	3.51 E
E5	5.73	ND	4.61 E	NR	5 E	ND	ND	ND	ND	ND	ND	ND	4.29 E
E2	16.3	16.7	12	NR	16	ND	17	ND	13.9	3.94 E	7.52	6.38 E	17.7
<i>116-m Depth Contour</i>													
B10	ND	ND	ND	NR	ND	ND	2.82 E	ND	ND	ND	ND	ND	3.09 E
E21	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
E15 ^a	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
E9	ND	ND	ND	NR	4.05 E	ND	3.67 E	ND	ND	ND	ND	ND	6.16 E
E3	14.4	13.9	10.2	NR	9.81	ND	12.8	ND	11.7	ND	ND	9.73	14.2
DR (%)	23	23	18	NR	27	0	86	0	23	14	5	14	91
ERL ^b	—	—	—	—	384	63.4	600	19	—	160	—	240	665
ERM ^b	—	—	—	—	2800	260	5100	540	—	2100	—	1500	2600

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.18 *continued*

Summer 2020	tPAH	1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo(A) anthracene	Benzo(A) pyrene
88-m Depth Contour												
B11	25.43	ND	ND	ND	ND	8.21 E	ND	ND	ND	ND	ND	ND
B8	29.56	ND	ND	ND	ND	9.11 E	ND	ND	ND	ND	ND	ND
E19	75.71	3.61 E	6.27 E	7.24	ND	17.1	8.46 E	8.27 E	ND	ND	ND	ND
E7	141.28	5.34 E	ND	ND	ND	9.02	5.98 E	5.01 E	7.24 E	ND	ND	11.5
E1	93.41	ND	ND	ND	ND	9	ND	ND	ND	ND	ND	12.8
98-m Depth Contour												
B12	38.61	ND	6.6	6.23 E	ND	8.11 E	ND	2.57 E	ND	ND	ND	ND
B9	23.08	ND	ND	ND	ND	7.36 E	ND	ND	ND	ND	ND	ND
E26	134.93	5.22 E	ND	ND	ND	8.45 E	12.1	4.26 E	5.7 E	ND	ND	11
E25	12.78	ND	ND	ND	ND	5.86 E	ND	ND	ND	ND	ND	ND
E23	11.03	ND	ND	ND	ND	6.67 E	ND	ND	ND	ND	ND	ND
E20	44.78	ND	ND	ND	ND	8.07 E	ND	2.73 E	ND	ND	ND	ND
E17 ^a	14.05	ND	ND	ND	ND	9.4	ND	ND	ND	ND	ND	ND
E14 ^a	15.37	ND	ND	ND	ND	7.82 E	ND	ND	ND	ND	ND	ND
E11 ^a	26.4	ND	ND	ND	ND	8.03 E	1.73 E	ND	ND	ND	ND	ND
E8	9.92	ND	ND	ND	ND	5.91 E	ND	ND	ND	ND	ND	ND
E5	16.88	ND	ND	ND	ND	6.9 E	ND	ND	ND	ND	ND	ND
E2	111.13	ND	ND	ND	ND	8.79	17.4	4.14 E	6.81 E	ND	ND	11.7
116-m Depth Contour												
B10	21.43	ND	ND	ND	ND	8.18 E	ND	ND	ND	ND	ND	ND
E21	20.04	ND	ND	ND	ND	9.58	ND	ND	ND	ND	ND	ND
E15 ^a	17.36	ND	ND	ND	ND	8.51	ND	ND	ND	ND	ND	ND
E9	22.2	ND	ND	ND	ND	4.24 E	ND	ND	ND	ND	ND	ND
E3	103.65	ND	ND	ND	ND	4.93 E	18.6	4.45 E	4.45 E	ND	ND	13.1
DR (%)	100	14	9	9	0	100	23	18	27	18	0	23
ERL ^b	4022	—	—	70	—	—	—	16	44	85.3	261	430
ERM ^b	44,792	—	—	670	—	—	—	500	640	1100	1600	1600

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.18 *continued*

Summer 2020	Benzo[e] pyrene	Benzo[G,H,I] perylene	Benzo[K] fluoranthene	Biphenyl	Chrysene	Dibenzo(A,H) anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-CD) pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
<i>88-m Depth Contour</i>													
B11	ND	6.32 E	ND	NR	ND	ND	5.15 E	ND	ND	ND	ND	ND	5.75 E
B8	ND	5.99 E	ND	NR	ND	ND	6.62 E	ND	ND	ND	ND	ND	7.84 E
E19	ND	ND	ND	NR	ND	ND	6.87 E	ND	ND	9.32	ND	ND	8.57
E7	10.1	11.8	11.7	NR	8.18	9.59	9.82	ND	10.7	ND	ND	ND	11.8
E1	9.35	12.2	ND	NR	6.58	ND	7.9	ND	9.07	ND	ND	ND	9.81
<i>98-m Depth Contour</i>													
B12	ND	ND	ND	NR	ND	ND	ND	ND	ND	9.06	ND	ND	3.04 E
B9	ND	ND	ND	NR	ND	ND	4.14 E	ND	ND	5.99 E	ND	ND	5.59 E
E26	11	13.4	9.84	NR	8.44	10.8	9.11	ND	11.7	3.97 E	ND	ND	9.94
E25	ND	ND	ND	NR	ND	ND	3.27 E	ND	ND	ND	ND	ND	3.65 E
E23	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	4.36 E
E20	ND	9.76	ND	NR	4.16 E	ND	4.77 E	ND	9.05	ND	ND	ND	6.24 E
E17 ^a	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	4.65 E
E14 ^a	ND	ND	ND	NR	ND	ND	3.28 E	ND	ND	ND	ND	ND	4.27 E
E11 ^a	ND	5.72 E	ND	NR	2.83 E	ND	3.58 E	ND	ND	ND	ND	ND	4.51 E
E8	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	4.01 E
E5	ND	ND	ND	NR	ND	ND	4.67 E	ND	ND	ND	ND	ND	5.31 E
E2	ND	9.47	9.11	NR	6.85	ND	12.3	ND	7.68	4.48 E	ND	ND	12.4
<i>116-m Depth Contour</i>													
B10	ND	ND	ND	NR	ND	ND	5.96 E	ND	ND	ND	ND	ND	7.29 E
E21	ND	ND	ND	NR	ND	ND	ND	ND	ND	5.99 E	ND	ND	4.47 E
E15 ^a	ND	ND	ND	NR	ND	ND	4.26 E	ND	ND	ND	ND	ND	4.59 E
E9	ND	5.94 E	ND	NR	ND	ND	3.64 E	ND	4.39 E	ND	ND	ND	3.99 E
E3	9.39	12.4	ND	NR	6.73 E	ND	8.9 E	ND	9.79	ND	ND	3.36 E	12
DR (%)	18	45	14	NR	32	9	77	0	32	27	0	5	100
ERL ^b	—	—	—	—	384	63.4	600	19	—	160	—	240	665
ERM ^b	—	—	—	—	2800	260	5100	540	—	2100	—	1500	2600

^aNear-ZID station

^bFrom Long et al. 1995

Appendix B.19

Concentrations of PAHs (ppb) detected in sediments from SBOO stations sampled during 2020. See Appendix B.1 for MDLs; DR = detection rate; E = estimated; ND = not detected; NR = not reportable.

Winter 2020	tPAH	1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo(A) anthracene	Benzo(A) pyrene
<i>19-m Stations</i>												
I35	17.82	ND	ND	ND	ND	6.01 E	ND	ND	ND	ND	ND	ND
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I23	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	1.56	ND	ND	ND	ND	1.56 E	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>28-m Stations</i>												
I33	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I30	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I27	24.62	ND	3.49 E	3.81 E	ND	5.56 E	3.85 E	3.95 E	ND	ND	ND	ND
I22	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I14 ^a	3.1	ND	ND	ND	ND	3.1 E	ND	ND	ND	ND	ND	ND
I16 ^a	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	5.91	ND	ND	ND	ND	ND	1.51 E	1.65 E	ND	ND	ND	ND
I9	3.38	ND	ND	ND	ND	3.38 E	ND	ND	ND	ND	ND	ND
I6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>38-m Stations</i>												
I29	16.67	ND	ND	ND	ND	8.67	ND	ND	ND	ND	ND	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>55-m Stations</i>												
I28	16.27	ND	ND	ND	ND	7.97	ND	ND	ND	ND	ND	ND
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	30	0	4	4	0	26	0	7	7	0	0	0
ERL ^b	4022	—	—	70	—	—	—	16	44	85.3	261	430
ERM ^b	44,792	—	—	670	—	—	—	500	640	1100	1600	1600

^aNear-ZID station; ^bFrom Long et al. 1995

Appendix B.19 *continued*

Winter 2020	Benzo[e]pyrene	Benzo[G,H,I]perylene	Benzo[K]fluoranthene	Biphenyl	Chrysene	Dibenzo(A,H)anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-CD)pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
19-m Stations													
I35	ND	ND	ND	NR	ND	ND	5.99 E	ND	ND	ND	ND	ND	5.82 E
I34	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I23	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I18	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
28-m Stations													
I33	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I30	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I27	ND	ND	ND	NR	ND	ND	ND	ND	3.96 E	ND	ND	ND	ND
I22	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I14 ^a	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I16 ^a	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	ND	ND	ND	NR	ND	ND	ND	ND	2.75 E	ND	ND	ND	ND
I9	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I6	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I3	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
38-m Stations													
I29	ND	ND	ND	NR	ND	ND	3.89 E	ND	ND	ND	ND	ND	4.11 E
I21	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
55-m Stations													
I28	ND	ND	ND	NR	ND	ND	4.31 E	ND	ND	ND	ND	ND	3.99 E
I20	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	0	0	0	NR	0	0	11	0	0	7	0	0	11
ERL ^b	—	—	—	—	384	63.4	600	19	—	160	—	240	665
ERM ^b	—	—	—	—	2800	260	5100	540	—	2100	—	1500	2600

^a Near-ZID station; ^b From Long et al. 1995

Appendix B.19 *continued*

Summer 2020	fPAH	1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo[A] anthracene	Benzo[A] pyrene
<i>19-m Stations</i>												
I35	23.07	ND	ND	ND	ND	13.2	ND	ND	ND	ND	ND	ND
I34	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31	4.03	ND	ND	ND	ND	4.03 E	ND	ND	ND	ND	ND	ND
I23	10.88	ND	ND	ND	ND	7.34 E	ND	ND	ND	ND	ND	ND
I18	4.88	ND	ND	ND	ND	4.88 E	ND	ND	ND	ND	ND	ND
I10	3.49	ND	ND	ND	ND	3.49 E	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>28-m Stations</i>												
I33	10.73	ND	ND	ND	ND	8.26 E	ND	ND	ND	ND	ND	ND
I30	10.8	ND	ND	ND	ND	10.8	ND	ND	ND	ND	ND	ND
I27	8.33	ND	ND	ND	ND	8.33 E	ND	ND	ND	ND	ND	ND
I22	23.39	ND	ND	ND	ND	9.2	4.4 E	2.56 E	ND	ND	ND	ND
I14 ^a	15.29	ND	ND	ND	ND	9.2	ND	ND	ND	ND	ND	ND
I16 ^a	7.83	ND	ND	ND	ND	7.83 E	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	21.83	3.8 E	ND	ND	ND	8.34	3.59 E	ND	ND	ND	ND	ND
I9	12.4	ND	ND	ND	ND	12.4	ND	ND	ND	ND	ND	ND
I6	1.22	ND	ND	ND	ND	1.22 E	ND	ND	ND	ND	ND	ND
I2	11.27	ND	ND	ND	ND	2.84 E	ND	ND	ND	ND	ND	ND
I3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>38-m Stations</i>												
I29	12.6	ND	ND	ND	ND	12.6	ND	ND	ND	ND	ND	ND
I21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>55-m Stations</i>												
I28	53.02	ND	5.86	5.47 E	ND	11.4	6.87 E	4.84 E	ND	ND	ND	ND
I20	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	9.32	ND	ND	ND	ND	2.95 E	ND	ND	ND	ND	ND	ND
DR (%)	67	4	4	4	0	67	11	7	0	0	0	0
ERL ^b	4022	—	—	70	—	—	16	44	85.3	261	430	430
ERM ^b	44,792	—	—	670	—	—	500	640	1100	1600	1600	1600

^a Near-ZID station; ^b From Long et al. 1995

Appendix B.19 *continued*

Summer 2020	Benzo[e] pyrene	Benzo[G,H,I] perylene	Benzo[K] fluoranthene	Biphenyl	Chrysene	Dibenzo(A,H) anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-CD) pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
19-m Stations													
I35	ND	ND	ND	NR	ND	ND	4.19 E	ND	ND	5.68 E	ND	ND	ND
I34	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I31	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I23	ND	ND	ND	NR	ND	ND	2.3 E	ND	ND	ND	ND	1.24 E	ND
I18	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I10	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I4	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
28-m Stations													
I33	ND	ND	ND	NR	ND	ND	2.47 E	ND	ND	ND	ND	ND	ND
I30	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I27	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I22	ND	ND	ND	NR	ND	ND	2.96 E	ND	ND	4.27 E	ND	ND	ND
I14 ^a	ND	ND	ND	NR	ND	ND	2.71 E	ND	ND	ND	ND	ND	3.38 E
I16 ^a	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I15	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I12 ^a	ND	ND	ND	NR	ND	ND	2.77 E	ND	ND	ND	ND	ND	3.33 E
I9	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I6	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I2	ND	ND	ND	NR	ND	ND	3.95 E	ND	ND	ND	ND	ND	4.48 E
I3	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
38-m Stations													
I29	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I21	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I13	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I8	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
55-m Stations													
I28	ND	ND	ND	NR	ND	ND	6.21 E	ND	ND	7.23	ND	ND	5.14 E
I20	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I7	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
I1	ND	ND	ND	6.37 E	ND	ND	ND	ND	ND	ND	ND	ND	ND
DR (%)	0	0	0	100	0	0	30	0	0	11	0	4	15
ERL ^b	—	—	—	—	384	63.4	600	19	—	160	—	240	665
ERM ^b	—	—	—	—	2800	260	5100	540	—	2100	—	1500	2600

^a Near-ZID station; ^b From Long et al. 1995

Appendix B.20

Concentrations of PAHs (ppb) detected in sediments from the San Diego regional benthic stations sampled during summer 2020. See Appendix B.1 for MDLs; E = estimated; ND = not detected; NR = not reportable.

	Station	Depth (m)	tPAH	1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene
<i>Inner Shelf</i>	8929	7	55.41	2.41 E	ND	ND	ND	ND	5.84 E
	8944	7	5.12	ND	ND	ND	ND	5.12 E	ND
	8918	8	ND	ND	ND	ND	ND	ND	ND
	8946	15	3.9	ND	ND	ND	ND	3.9 E	ND
	8915	16	ND	ND	ND	ND	ND	ND	ND
	8907	18	2.86	ND	ND	ND	ND	2.86 E	ND
	8943	19	ND	ND	ND	ND	ND	ND	ND
	8914	20	22.64	ND	ND	ND	ND	10.8	ND
	8911	24	9.27	ND	ND	ND	ND	9.27	ND
	8938	28	1.56	ND	ND	ND	ND	1.56 E	ND
	8901	29	15.19	ND	ND	ND	ND	8.24 E	ND
<i>Mid Shelf</i>	8913	31	27.88	ND	ND	ND	ND	7.04 E	ND
	8906	41	ND	ND	ND	ND	ND	ND	ND
	8939	65	14.9	ND	ND	ND	ND	14.9	ND
	8903	66	4.32	ND	ND	ND	ND	4.32 E	ND
	8912	67	41.02	ND	ND	ND	ND	14.8	ND
	8925	67	30.39	ND	ND	ND	ND	15.3	ND
	8919	68	37.98	ND	ND	ND	ND	13.1	ND
	8909	73	50.67	ND	ND	ND	ND	16.2	ND
	8931	74	42.75	ND	ND	ND	ND	9.38 E	ND
	8933	80	28.22	ND	ND	ND	ND	10	ND
	8930	83	23.01	ND	ND	ND	ND	7.2 E	ND
	8949	83	20.07	ND	ND	ND	ND	6.96 E	ND
	8924	85	50.33	ND	ND	ND	ND	13.3	ND
	8926	87	111.7	ND	7.35	8.43	ND	17.1	ND
	8932	89	18.19	ND	ND	ND	ND	6.29 E	ND
8940	108	21.98	ND	3.12 E	3.45 E	ND	7.66	ND	
8921	116	22.1	ND	ND	4.65 E	ND	10.9	ND	
<i>Outer Shelf</i>	8905	134	41.3	ND	ND	ND	ND	8.34 E	ND
	8937	137	10.7	ND	ND	ND	ND	10.7	ND
	8910	169	318.33	ND	ND	ND	ND	5.23 E	66.8
	8922	185	57.21	ND	ND	ND	ND	14.3	ND
	8916	189	269.84	ND	ND	ND	ND	11.6	46.6
	8908	194	140.58	ND	ND	ND	ND	7.97 E	27.2
<i>Upper Slope</i>	8928	223	114.43	ND	ND	ND	ND	11.6	16.4
	8920	270	277.87	ND	ND	ND	ND	10.9 E	27
	8936	292	132.49	ND	ND	ND	ND	11.4	18.2
	8927	460	51.28	ND	ND	ND	ND	6.73 E	10.9
	8934	491	23.22	ND	ND	ND	ND	ND	ND
	8923	523	125.42	ND	ND	ND	ND	12.7	11.2
Detection Rate (%)			90	3	5	8	0	85	23
ERL ^a :			4022	—	—	70	—	—	—
ERM ^a :			44,792	—	—	670	—	—	—

^aFrom Long et al. 1995

Appendix B.20 *continued*

	Station	Depth (m)	Acenaphthene	Acenaphthylene	Anthracene	Benzo[A]anthracene	Benzo[A]pyrene	Benzo[e]pyrene
<i>Inner Shelf</i>	8929	7	4.38 E	ND	4.5 E	ND	5.06 E	4.93 E
	8944	7	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	ND	ND	ND	ND
	8911	24	ND	ND	ND	ND	ND	ND
	8938	28	ND	ND	ND	ND	ND	ND
	8901	29	ND	ND	ND	ND	ND	ND
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	7.95	ND
	8906	41	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	ND	ND	ND
	8903	66	ND	ND	ND	ND	ND	ND
	8912	67	ND	ND	ND	ND	ND	ND
	8925	67	ND	ND	ND	ND	ND	ND
	8919	68	ND	ND	ND	ND	ND	ND
	8909	73	ND	ND	ND	ND	ND	ND
	8931	74	ND	ND	ND	ND	ND	ND
	8933	80	ND	ND	ND	ND	ND	ND
	8930	83	ND	ND	ND	ND	ND	ND
	8949	83	ND	ND	ND	ND	ND	ND
	8924	85	ND	ND	ND	ND	ND	ND
	8926	87	ND	9.07	9.55 E	ND	ND	ND
	8932	89	ND	ND	ND	ND	ND	ND
8940	108	ND	2.96 E	ND	ND	ND	ND	
8921	116	ND	ND	ND	ND	ND	ND	
<i>Outer Shelf</i>	8905	134	ND	ND	ND	ND	ND	ND
	8937	137	ND	ND	ND	ND	ND	ND
	8910	169	ND	5.79 E	9.77 E	ND	40.8	27
	8922	185	ND	ND	ND	ND	ND	ND
	8916	189	ND	5.15 E	8.45 E	ND	33.8	24.9
	8908	194	ND	ND	ND	ND	17.2	13.4
<i>Upper Slope</i>	8928	223	ND	ND	ND	ND	12.7	10.8
	8920	270	ND	8.14 E	12.4 E	ND	22.1	13.4
	8936	292	ND	ND	ND	ND	14.6	13.5
	8927	460	ND	ND	ND	ND	ND	ND
	8934	491	ND	ND	ND	ND	ND	ND
	8923	523	ND	ND	ND	ND	10.2	10.4
Detection Rate (%)			3	13	13	0	23	20
ERL ^a :			16	44	85.3	261	430	—
ERM ^a :			500	640	1100	1600	1600	—

^aFrom Long et al. 1995

Appendix B.20 *continued*

	Station	Depth (m)	Benzo[G,H,I] perylene	Benzo[K] fluoranthene	Biphenyl	Chrysene	Dibenzo(A,H) anthracene	Fluoranthene
<i>Inner Shelf</i>	8929	7	ND	ND	NR	3.42 E	ND	3.77 E
	8944	7	ND	ND	NR	ND	ND	ND
	8918	8	ND	ND	NR	ND	ND	ND
	8946	15	ND	ND	NR	ND	ND	ND
	8915	16	ND	ND	NR	ND	ND	ND
	8907	18	ND	ND	NR	ND	ND	ND
	8943	19	ND	ND	NR	ND	ND	ND
	8914	20	ND	ND	NR	ND	ND	5.98 E
	8911	24	ND	ND	NR	ND	ND	ND
	8938	28	ND	ND	NR	ND	ND	ND
8901	29	ND	ND	NR	ND	ND	3.71 E	
<i>Mid Shelf</i>	8913	31	6.15	ND	6.74 E	ND	ND	ND
	8906	41	ND	ND	NR	ND	ND	ND
	8939	65	ND	ND	NR	ND	ND	ND
	8903	66	ND	ND	NR	ND	ND	ND
	8912	67	ND	ND	NR	5.22 E	ND	10.1
	8925	67	ND	ND	NR	ND	ND	7.52 E
	8919	68	8.38	ND	NR	ND	ND	6.6 E
	8909	73	ND	ND	8.59 E	5.28 E	ND	10.5
	8931	74	ND	ND	NR	4.62 E	ND	8.1
	8933	80	ND	ND	NR	ND	ND	6.66 E
	8930	83	ND	ND	NR	ND	ND	5.8 E
	8949	83	ND	ND	NR	ND	ND	5.77 E
	8924	85	10.3	ND	NR	5.94 E	ND	5.91 E
	8926	87	9.35	ND	NR	10.2	ND	7.46
8932	89	ND	ND	NR	ND	ND	5.25 E	
8940	108	ND	ND	NR	ND	ND	2.36 E	
8921	116	ND	ND	NR	ND	ND	ND	
<i>Outer Shelf</i>	8905	134	10.5	ND	NR	7.4	ND	7.23
	8937	137	ND	ND	NR	ND	ND	ND
	8910	169	32.8	24.4	NR	17.8	ND	24.9
	8922	185	11.3	ND	NR	6.75	ND	7.34 E
	8916	189	29.1	18.2	NR	18.8	ND	21.3
	8908	194	15.4	6.11 E	NR	13.2	ND	13.6
<i>Upper Slope</i>	8928	223	12.5	8.74	NR	6.11 E	ND	10
	8920	270	18.2	10.1	NR	19.4	ND	41.9
	8936	292	15.9	ND	NR	8.25	16.5	6.54 E
	8927	460	ND	ND	NR	6.01 E	ND	9.92 E
	8934	491	ND	ND	NR	ND	ND	9.12 E
	8923	523	12.5	8.61	NR	7.5 E	ND	9.29 E
Detection Rate (%)			33	15	100	40	3	65
ERL ^a :			—	—	—	384	63.4	600
ERM ^a :			—	—	—	2800	260	5100

^aFrom Long et al. 1995

Appendix B.20 *continued*

	Station	Depth (m)	Fluorene	Indeno(1,2,3-CD) pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
<i>Inner Shelf</i>	8929	7	ND	6.36 E	3 E	2.19 E	5.26 E	4.29 E
	8944	7	ND	ND	ND	ND	ND	ND
	8918	8	ND	ND	ND	ND	ND	ND
	8946	15	ND	ND	ND	ND	ND	ND
	8915	16	ND	ND	ND	ND	ND	ND
	8907	18	ND	ND	ND	ND	ND	ND
	8943	19	ND	ND	ND	ND	ND	ND
	8914	20	ND	ND	ND	ND	ND	5.86 E
	8911	24	ND	ND	ND	ND	ND	ND
	8938	28	ND	ND	ND	ND	ND	ND
	8901	29	ND	ND	ND	ND	ND	3.24 E
<i>Mid Shelf</i>	8913	31	ND	ND	ND	ND	ND	ND
	8906	41	ND	ND	ND	ND	ND	ND
	8939	65	ND	ND	ND	ND	ND	ND
	8903	66	ND	ND	ND	ND	ND	ND
	8912	67	ND	ND	ND	ND	ND	10.9
	8925	67	ND	ND	ND	ND	ND	7.57 E
	8919	68	ND	ND	ND	ND	ND	9.9
	8909	73	ND	ND	ND	ND	ND	10.1
	8931	74	ND	ND	6.21 E	ND	4.88 E	9.56
	8933	80	ND	ND	4.08 E	ND	ND	7.48 E
	8930	83	ND	ND	2.72 E	ND	ND	7.29 E
	8949	83	ND	ND	ND	ND	ND	7.34 E
	8924	85	ND	7.87	ND	ND	ND	7.01 E
	8926	87	ND	8.12	10	ND	7.76 E	7.31 E
	8932	89	ND	ND	ND	ND	ND	6.65 E
8940	108	ND	ND	ND	ND	ND	2.43 E	
8921	116	ND	ND	6.55 E	ND	ND	ND	
<i>Outer Shelf</i>	8905	134	ND	ND	ND	ND	ND	7.83
	8937	137	ND	ND	ND	ND	ND	ND
	8910	169	ND	25.9	ND	ND	7.54 E	29.6
	8922	185	ND	9.05	ND	ND	ND	8.47
	8916	189	ND	21.4	4.64 E	ND	ND	25.9
	8908	194	ND	12.5	ND	ND	ND	14
<i>Upper Slope</i>	8928	223	ND	11.1	ND	ND	4.18 E	10.3
	8920	270	ND	13.6	5.83 E	ND	30.9	44
	8936	292	ND	14.4	ND	ND	ND	13.2
	8927	460	ND	ND	ND	ND	5.72 E	12
	8934	491	ND	ND	ND	ND	ND	14.1
	8923	523	ND	12.2	5.05 E	9.96	4.81 E	11
Detection Rate (%)			0	28	23	5	20	65
ERL ^a :			19	—	160	—	240	665
ERM ^a :			540	—	2100	—	1500	2600

^aFrom Long et al. 1995

Appendix B.21

Macrofaunal community parameters by grab for PLOO benthic stations sampled during 2020. SR= species richness; Abun= abundance; H'=Shannon diversity index; J'=Pielou's evenness; Dom= Swartz dominance; BRI= benthic response index.

Depth Contour	Station	Survey	SR	Abun	H'	J'	Dom	BRI	
88-m	B11	winter	74	284	3.8	0.87	25	18	
		summer	104	294	4.2	0.90	41	9	
	B8	winter	75	407	3.3	0.75	17	10	
		summer	87	373	3.7	0.82	23	9	
	E19	winter	70	477	3.3	0.78	14	10	
		summer	79	575	3.3	0.75	18	13	
	E7	winter	70	438	3.5	0.82	21	8	
		summer	86	619	3.6	0.82	22	12	
	E1	winter	80	450	3.5	0.80	21	8	
		summer	101	549	3.7	0.81	27	12	
	98-m	B12	winter	94	422	3.9	0.85	30	9
			summer	109	354	4.2	0.89	41	9
		B9	winter	85	380	3.8	0.85	25	14
			summer	93	318	3.9	0.87	35	12
E26		winter	72	397	3.4	0.80	20	11	
		summer	89	359	3.6	0.81	25	14	
E25		winter	68	393	3.4	0.80	18	8	
		summer	77	550	3.4	0.79	20	11	
E23		winter	70	471	3.4	0.79	18	13	
		summer	91	578	3.4	0.74	18	12	
E20		winter	64	377	3.2	0.76	13	15	
		summer	66	414	3.0	0.72	14	12	
E17 ^a		winter	65	457	3.1	0.73	15	19	
		summer	91	536	3.4	0.75	19	18	
E14 ^a		winter	72	558	3.4	0.80	15	33	
		summer	61	322	3.3	0.81	16	31	
E11 ^a		winter	54	456	3.1	0.76	12	18	
		summer	69	337	3.2	0.76	17	16	
E8		winter	67	318	3.5	0.83	20	12	
		summer	81	470	3.7	0.84	25	13	
E5		winter	79	399	3.6	0.83	23	8	
		summer	85	337	3.7	0.83	26	12	
E2		winter	109	468	3.9	0.84	32	15	
		summer	101	488	3.8	0.83	29	12	

^aNear-ZID station

Appendix B.21 *continued*

Depth Contour	Station	Survey	SR	Abun	H'	J'	Dom	BRI
116-m	B10	winter	70	284	3.6	0.85	22	16
		summer	86	241	3.9	0.88	33	14
	E21	winter	58	394	3.0	0.75	12	17
		summer	60	407	2.9	0.70	11	16
	E15 ^a	winter	62	329	3.1	0.76	12	17
		summer	80	636	3.2	0.73	14	15
	E9	winter	111	420	4.1	0.87	41	12
		summer	115	505	3.9	0.81	33	11
	E3	winter	105	373	4.2	0.89	38	15
		summer	102	325	4.1	0.90	39	9

^aNear-ZID station

Appendix B.22

Macrofaunal community parameters by grab for SBOO benthic stations sampled during 2020. SR = species richness; Abun = abundance; H' = Shannon diversity index; J' = Pielou's evenness; Dom = Swartz dominance; BRI = benthic response index.

Depth	Contour	Station	Survey	SR	Abun	H'	J'	Dom	BRI	
19-m	I35	winter		71	186	3.9	0.91	30	28	
		summer		99	390	3.9	0.84	32	27	
	I34	winter		49	764	2.5	0.64	6	24	
		summer		64	875	3.1	0.74	10	21	
	I31	winter		37	121	3.0	0.84	13	14	
		summer		60	213	3.5	0.87	22	16	
	I23	winter		46	109	3.2	0.85	19	21	
		summer		96	352	3.8	0.83	28	17	
	I18	winter		46	87	3.5	0.91	25	15	
		summer		48	178	3.4	0.88	19	17	
	I10	winter		60	169	3.4	0.83	22	15	
		summer		57	136	3.7	0.91	25	20	
	I4	winter		20	44	2.6	0.86	10	2	
		summer		29	70	3.1	0.91	13	5	
	28-m	I33	winter		74	171	3.9	0.91	32	26
			summer		88	225	4.0	0.90	36	24
		I30	winter		59	129	3.6	0.89	27	23
			summer		70	246	3.6	0.84	23	23
I27		winter		30	73	2.8	0.83	12	18	
		summer		73	281	3.5	0.82	24	20	
I22		winter		29	92	2.7	0.79	10	18	
		summer		64	145	3.7	0.90	29	23	
I14 ^a		winter		69	174	3.8	0.89	30	24	
		summer		86	241	3.9	0.88	34	20	
I16 ^a		winter		28	109	2.3	0.70	7	13	
		summer		67	301	3.0	0.71	16	16	
I15		winter		17	30	2.5	0.89	10	8	
		summer		48	128	2.9	0.75	17	22	
I12 ^a		winter		26	80	2.3	0.71	10	1	
		summer		93	245	4.1	0.91	40	20	
I9		winter		61	141	3.7	0.90	27	26	
		summer		59	141	3.6	0.89	25	24	
I6		winter		31	108	2.8	0.81	10	6	
		summer		35	226	2.7	0.75	9	7	
I2	winter		31	104	2.9	0.84	11	14		
	summer		41	173	2.6	0.69	8	18		
I3	winter		26	119	2.4	0.75	6	7		
	summer		60	277	3.1	0.76	14	19		

^aNear-ZID station

Appendix B.22 *continued*

Depth Contour	Station	Survey	SR	Abun	H'	J'	Dom	BRI
38-m	I29	winter	105	366	3.9	0.83	31	14
		summer	120	424	4.2	0.87	43	15
	I21	winter	40	87	3.3	0.90	19	10
		summer	63	238	3.5	0.85	20	11
	I13	winter	40	95	3.3	0.89	17	11
		summer	46	200	3.1	0.82	15	6
	I8	winter	35	86	3.2	0.90	16	19
		summer	39	131	2.8	0.77	15	17
55-m	I28	winter	160	575	4.3	0.85	52	16
		summer	116	434	4.1	0.87	37	15
	I20	winter	91	890	3.2	0.72	16	11
		summer	77	291	3.4	0.77	22	8
	I7	winter	47	130	3.2	0.84	18	1
		summer	57	145	3.7	0.92	26	4
	I1	winter	72	271	3.5	0.83	24	18
		summer	73	194	3.7	0.85	29	14

Appendix B.23

Macrofaunal community parameters by grab for the San Diego regional benthic stations sampled during summer 2020. SR = species richness; Abun = abundance; H' = Shannon diversity index; J' = Pielou's evenness; Dom = Swartz dominance; BRI = benthic response index.

	Station	Depth (m)	SR	Abun	H'	J'	Dom	BRI ^a
<i>Inner Shelf</i>	8929	7	39	248	2.9	0.78	10	—
	8944	7	32	108	3.2	0.92	15	—
	8918	8	21	645	0.4	0.12	1	—
	8946	15	42	140	3.3	0.88	18	26
	8915	16	59	145	3.8	0.93	27	14
	8907	18	44	116	3.5	0.92	22	15
	8943	19	41	87	3.3	0.90	20	19
	8914	20	113	413	4.0	0.84	37	28
	8911	24	64	216	3.5	0.85	23	23
	8938	28	46	150	3.5	0.91	21	27
	8901	29	64	162	3.8	0.91	28	19
<i>Mid Shelf</i>	8913	31	105	329	4.1	0.88	37	25
	8906	41	63	172	3.6	0.86	24	16
	8939	65	53	220	2.9	0.73	15	20
	8903	66	105	323	4.0	0.86	40	12
	8912	67	83	329	3.7	0.83	24	14
	8925	67	80	423	3.4	0.77	18	15
	8919	68	82	398	3.5	0.79	21	17
	8909	73	86	413	3.6	0.81	22	8
	8931	74	72	389	3.3	0.77	18	9
	8933	80	85	442	3.5	0.80	22	13
	8930	83	76	302	3.5	0.81	21	8
	8949	83	89	402	3.5	0.78	20	12
	8924	85	88	531	3.5	0.78	20	10
	8926	87	82	541	3.4	0.78	18	11
	8932	89	66	233	3.5	0.84	22	10
8940	108	66	332	3.5	0.82	20	21	
8921	116	58	466	2.8	0.68	10	19	
<i>Outer Shelf</i>	8905	134	67	238	3.5	0.82	19	15
	8937	137	84	290	3.8	0.85	28	19
	8910	169	64	348	3.3	0.79	18	22
	8922	185	39	220	2.5	0.69	7	24
	8916	189	33	215	2.3	0.65	5	25
	8908	194	54	172	3.4	0.86	19	16
<i>Upper Slope</i>	8928	223	31	79	3.0	0.87	14	—
	8920	270	20	69	2.0	0.68	5	—
	8936	292	32	88	2.9	0.85	12	—
	8927	460	26	48	3.1	0.94	15	—
	8934	491	36	90	2.9	0.80	14	—
	8923	523	25	51	3.0	0.94	13	—

^aBRI statistic not calculated for stations located at depths < 10 m or > 200 m

Appendix B.24

Summary taxonomic listing of benthic infauna taxa identified from PLOO stations during 2020. Data are total number of individuals (n). Taxonomic arrangement follows SCAMIT (2018). ** indicates taxon is not within previous family

Phylum	Class	Family	Taxon	n
Cnidaria	Hydrozoa	Corymorphidae	<i>Corymorpha bigelowi</i>	3
			<i>Euphysa</i> sp A	5
	Anthozoa	Virgulariidae	<i>Stylatula</i> sp A	1
			<i>Stylatula</i> sp	2
			** <i>Actiniaria</i>	1
			Edwardsiidae	25
			<i>Edwardsia olguini</i>	1
			<i>Scolanthus triangulus</i>	72
			Halcampidae	8
			<i>Halianthella</i> sp A	8
Platyhelminthes	Rhabditophora	Plehnidae	<i>Diplehnia caeca</i>	2
	Nemertea	Anopla	Palaeonemertea	6
Carinomidae			<i>Carinoma mutabilis</i>	3
Tubulanidae			7	
<i>Tubulanus cingulatus</i>			10	
<i>Tubulanus polymorphus</i>			49	
<i>Tubulanus</i> sp A			3	
Tubulanidae sp D			4	
** Heteronemertea			3	
Lineidae			32	
<i>Cerebratulus californiensis</i>			2	
Lineidae sp SD1			3	
<i>Lineus bilineatus</i>			29	
<i>Maculaura alaskensis</i> Cmplx			1	
<i>Zygeupolia rubens</i>			3	
** Heteronemertea sp SD2			164	
Enopla			Hoplonemertea	2
Emplectonematidae			<i>Paranemertes californica</i>	7
Amphiporidae			<i>Amphiporus flavescens</i>	1
** Hoplonemertea sp A	1			
Hoplonemertea sp B	1			
Hoplonemertea sp SD3	1			
Mollusca	Caudofoveata	Chaetodermatidae	Chaetodermatida	1
			<i>Falcidens longus</i>	1
	Gastropoda		<i>Solariella peramabilis</i>	10
			<i>Lirobittium rugatum</i> Cmplx	3
			<i>Neverita draconis</i>	1
			<i>Caecum crebricinctum</i>	10
			<i>Caesia perpinguis</i>	1
			<i>Boreotrophon</i> sp	1
			<i>Kurtzia arteaga</i>	2
			<i>Kurtzina beta</i>	5
			<i>Rictaxis punctocaelatus</i>	1
			<i>Odostomia</i> sp	16
			<i>Turbonilla chocolata</i>	2
			<i>Turbonilla santarosana</i>	4

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
			<i>Turbonilla</i> sp A	8
			<i>Turbonilla</i> sp SD5	4
			<i>Turbonilla</i> sp SD6	1
		Dotoidae	<i>Doto</i> sp	1
		Rhizoridae	<i>Volvulella californica</i>	9
			<i>Volvulella cylindrica</i>	1
			<i>Volvulella panamica</i>	1
			<i>Volvulella</i> sp	2
		Acteocinidae	<i>Acteocina cerealis</i>	3
		Philinidae	<i>Philine auriformis</i>	13
		Aglajidae	<i>Aglaja ocelligera</i>	1
		Gastropteridae	<i>Gastropteran pacificum</i>	3
		Cylichnidae	<i>Cylichna diegensis</i>	8
		**	<i>Bullomorpha</i> sp A	1
	Bivalvia			11
		Nuculidae	<i>Acila castrensis</i>	1
			<i>Ennucula tenuis</i>	31
		Solemyidae	<i>Solemya pervernicosa</i>	3
		Nucinellidae	<i>Huxleyia munita</i>	2
		Nuculanidae	<i>Nuculana hamata</i>	8
			<i>Nuculana</i> sp A	155
		Mytilidae	<i>Amygdalum pallidulum</i>	1
		Limidae	<i>Limatula saturna</i>	1
		Carditidae	<i>Cyclocardia ventricosa</i>	1
		Lucinidae	<i>Parvilucina tenuisculpta</i>	85
			<i>Lucinoma annulatum</i>	23
		Thyasiridae	<i>Adontorhina cycليا</i>	12
			<i>Axinopsida serricata</i>	1902
			<i>Thyasira flexuosa</i>	11
		Lasaeidae	<i>Kurtiella tumida</i>	2
			<i>Kurtiella</i> sp D	1
		Cardiidae	<i>Keenaea centifilosum</i>	27
		Tellinidae	<i>Tellina carpenteri</i>	164
			<i>Tellina</i> sp B	383
			<i>Tellina</i> sp	5
			<i>Macoma</i> sp	5
		Hiatellidae	<i>Saxicavella nybakkeni</i>	6
			<i>Saxicavella pacifica</i>	1
		Veneridae	<i>Nutricola ovalis</i>	2
		Lyonsiidae		1
		Thraciidae	<i>Thracia trapezoides</i>	1
		Cuspidariidae		1
			<i>Cuspidaria parapodema</i>	2
	Scaphopoda			2
		Dentaliidae	<i>Dentalium vallicolens</i>	3
		Gadilidae	<i>Polyschides quadrifissatus</i>	18
			<i>Gadila aberrans</i>	1
		**	<i>Compressidens stearnsii</i>	1
Sipuncula				1
	Sipunculidea	Golfingiidae	<i>Nephasoma diaphanes</i>	5

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n	
Annelida	Polychaeta	Phascolionidae	<i>Thysanocardia nigra</i>	1	
			<i>Phascolion</i> sp A	30	
			Thalassematidae	<i>Listriolobus pelodes</i>	2
				Amphinomidae	<i>Chloeia pinnata</i>
			Lumbrineridae	<i>Eranno bicirrata</i>	3
				<i>Eranno lagunae</i>	4
				<i>Lumbrineris cruzensis</i>	26
				<i>Lumbrineris index</i>	1
				<i>Lumbrineris latreilli</i>	7
				<i>Lumbrineris ligulata</i>	5
				<i>Lumbrineris</i> sp Group I	24
				<i>Lumbrineris</i> sp Group II	1
				<i>Lumbrineris</i> sp	2
				<i>Scoletoma tetraura</i> Cmplx	33
		<i>Scoletoma</i> sp		1	
		Oeonidae	<i>Drilonereis falcata</i>	16	
			<i>Drilonereis</i> sp A	1	
			<i>Drilonereis</i> sp	13	
			<i>Notocirrus californiensis</i>	6	
			Onuphidae		24
		<i>Mooreonuphis exigua</i>		5	
		<i>Mooreonuphis nebulosa</i>		8	
		<i>Mooreonuphis segmentispadix</i>		12	
		<i>Mooreonuphis</i> sp SD1		7	
		<i>Mooreonuphis</i> sp SD2		3	
		<i>Mooreonuphis</i> sp		32	
		<i>Nothria occidentalis</i>		3	
		<i>Nothria</i> sp		8	
		<i>Onuphis eremita parva</i>		2	
		<i>Onuphis iridescens</i>		1	
		<i>Onuphis</i> sp A		15	
		<i>Onuphis</i> sp		2	
		<i>Paradiopatra parva</i>		544	
		Acoetidae		<i>Acoetes pacifica</i>	2
		Aphroditidae			1
		Polynoidae		<i>Aphrodita</i> sp	2
			<i>Malmgreniella baschi</i>	4	
			<i>Malmgreniella liei</i>	1	
			<i>Malmgreniella sanpedroensis</i>	5	
			<i>Malmgreniella</i> sp A	9	
			<i>Subadyte mexicana</i>	5	
			<i>Tenonia priops</i>	11	
		Pholoidae		1	
Sigalionidae	<i>Pholoe glabra</i>	62			
	<i>Pholoides asperus</i>	2			
	<i>Sigalion spinosus</i>	35			
	<i>Sthenelais tertiaglabra</i>	20			
	<i>Sthenelais</i> sp	1			
	<i>Sthenelanella uniformis</i>	21			
Glyceridae		1			

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
			<i>Glycera americana</i>	6
			<i>Glycera nana</i>	153
			<i>Glycera</i> sp	1
		Goniadidae	<i>Glycinde armigera</i>	23
			<i>Goniada brunnea</i>	13
			<i>Goniada maculata</i>	84
		Hesionidae	<i>Podarkeopsis glabrus</i>	7
			<i>Podarkeopsis</i> sp A	1
		Nereididae	<i>Nereis</i> sp A	14
			<i>Platynereis bicanaliculata</i>	1
		Pilargidae	<i>Sigambra setosa</i>	1
		Syllidae	<i>Eusyllis blomstrandii</i> Cmplx	4
			<i>Eusyllis habeii</i>	1
			<i>Paraehlersia articulata</i>	5
			<i>Exogone dwisula</i>	1
			<i>Exogone lourei</i>	23
			<i>Syllis heterochaeta</i>	6
		Nephtyidae		1
			<i>Aglaophamus verrilli</i>	8
			<i>Nephtys caecoides</i>	5
			<i>Nephtys ferruginea</i>	63
		Phyllodoceidae	<i>Eteone brigitteae</i>	1
			<i>Eteone pigmentata</i>	1
			<i>Eulalia</i> sp SD4	1
			<i>Eumida longicornuta</i>	1
			<i>Sige</i> sp A	16
			<i>Nereiphylla</i> sp 2	1
			<i>Nereiphylla</i> sp SD1	1
			<i>Paranaitis polynoides</i>	4
			<i>Phyllodoce cuspidata</i>	1
			<i>Phyllodoce groenlandica</i>	2
			<i>Phyllodoce hartmanae</i>	34
			<i>Phyllodoce longipes</i>	13
			<i>Phyllodoce pettiboneae</i>	12
		Oweniidae		1
			<i>Galathowenia pygidialis</i>	18
			<i>Myriochele gracilis</i>	18
			<i>Myriochele olgae</i>	1
			<i>Myriochele striolata</i>	3
			<i>Myriowenia californiensis</i>	1
		Sabellariidae	<i>Neosabellaria cementarium</i>	1
		Sabellidae		1
			<i>Acromegalomma pigmentum</i>	1
			<i>Acromegalomma splendidum</i>	3
			<i>Dialychone albocincta</i>	56
			<i>Dialychone trilineata</i>	129
			<i>Dialychone veleronis</i>	5
			<i>Euchone arenae</i>	2
			<i>Euchone hancocki</i>	14
			<i>Euchone incolor</i>	73

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
			<i>Euchone</i> sp A	12
			<i>Euchone</i> sp	1
			<i>Jasmineira</i> sp B	7
			<i>Myxicola</i> sp	5
			<i>Paradialychone ecaudata</i>	1
			<i>Paradialychone harrisae</i>	3
			<i>Paradialychone paramollis</i>	4
			<i>Potamethus</i> sp A	8
		Apistobranchidae	<i>Apistobranchus ornatus</i>	1
		Longosomatidae	<i>Heterospio catalinensis</i>	4
		Magelonidae		10
			<i>Magelona berkeleyi</i>	3
			<i>Magelona hartmanae</i>	1
			<i>Magelona hobsonae</i>	1
			<i>Magelona</i> sp B	5
		Spionidae	<i>Dipolydora giardi</i>	1
			<i>Dipolydora socialis</i>	2
			<i>Laonice cirrata</i>	27
			<i>Laonice nuchala</i>	37
			<i>Microspio pigmentata</i>	130
			<i>Paraprionospio alata</i>	95
			<i>Prionospio dubia</i>	342
			<i>Prionospio jubata</i>	653
			<i>Prionospio lighti</i>	6
			<i>Prionospio</i> sp	1
			<i>Scolecopsis (Parascolecopsis) texana</i>	1
			<i>Spio filicornis</i>	1
			<i>Spio maculata</i>	12
			<i>Spiophanes berkeleyorum</i>	41
			<i>Spiophanes duplex</i>	2527
			<i>Spiophanes kimballi</i>	641
			<i>Spiophanes norrisi</i>	4
			<i>Spiophanes wigleyi</i>	12
			<i>Spiophanes</i> sp	1
		Cirratulidae	<i>Aphelochaeta glandaria</i> Cmplx	306
			<i>Aphelochaeta monilaris</i>	64
			<i>Aphelochaeta phillipsi</i>	50
			<i>Aphelochaeta tigrina</i>	23
			<i>Aphelochaeta williamsae</i>	5
			<i>Aphelochaeta</i> sp HYP2	3
			<i>Aphelochaeta</i> sp LA1	22
			<i>Aphelochaeta</i> sp SD3	3
			<i>Aphelochaeta</i> sp SD18	1
			<i>Aphelochaeta</i> sp	22
			<i>Chaetozone hartmanae</i>	152
			<i>Chaetozone setosa</i> Cmplx	2
			<i>Chaetozone</i> sp SD1	1
			<i>Chaetozone</i> sp SD3	2
			<i>Chaetozone</i> sp SD5	3
			<i>Chaetozone</i> sp SD7	27

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
			<i>Chaetozone</i> sp	12
			<i>Kirkegaardia cryptica</i>	10
			<i>Kirkegaardia serratseta</i>	3
			<i>Kirkegaardia sibilina</i>	109
			<i>Kirkegaardia tessellata</i>	55
			<i>Kirkegaardia</i> sp SD9	6
			<i>Kirkegaardia</i> sp	4
		Fauveliopsidae	<i>Fauveliopsis</i> sp SD1	35
		Flabelligeridae	<i>Brada pluribranchiata</i>	1
			<i>Pherusa neopapillata</i>	1
		Sternaspidae	<i>Sternaspis affinis</i>	119
		Ampharetidae		5
			<i>Amage anops</i>	4
			<i>Amage scutata</i>	15
			<i>Ampharete finmarchica</i>	18
			<i>Ampharete labrops</i>	4
			<i>Ampharete</i> sp	1
			Ampharetidae sp SD1	8
			<i>Amphicteis scaphobranchiata</i>	10
			<i>Amphisamytha bioculata</i>	1
			<i>Anobothrus gracilis</i>	29
			<i>Eclysippe trilobata</i>	373
			<i>Lysippe</i> sp A	25
			<i>Lysippe</i> sp B	46
			<i>Sabellides manriquei</i>	1
			<i>Samytha californiensis</i>	1
			<i>Sosane occidentalis</i>	9
			<i>Melinna oculata</i>	1
		Pectinariidae	<i>Pectinaria californiensis</i>	131
		Terebellidae		1
			<i>Polycirrus californicus</i>	30
			<i>Polycirrus</i> sp A	96
			<i>Polycirrus</i> sp OC1	113
			<i>Polycirrus</i> sp SD3	2
			<i>Polycirrus</i> sp	83
			<i>Lanassa venusta venusta</i>	134
			<i>Phisidia sanctaemariae</i>	282
			<i>Pista brevibranchiata</i>	6
			<i>Pista estevanica</i>	36
			<i>Pista wui</i>	4
			<i>Pista</i> sp	2
			<i>Proclea</i> sp A	27
		Trichobranchidae	<i>Terebellides californica</i>	5
		Chaetopteridae	<i>Phyllochaetopterus limicolus</i>	5
			<i>Phyllochaetopterus</i> sp	1
			<i>Spiochaetopterus costarum</i> Cmplx	65
		Capitellidae		1
			<i>Capitella teleta</i>	29
			<i>Decamastus gracilis</i>	33
			<i>Decamastus</i> sp	2

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
			<i>Mediomastus</i> sp	863
			<i>Notomastus hemipodus</i>	75
			<i>Notomastus latericeus</i>	3
			<i>Notomastus</i> sp	13
		Cossuridae	<i>Cossura bansei</i>	1
			<i>Cossura candida</i>	9
			<i>Cossura</i> sp A	3
		Maldanidae		215
			Euclymeninae	38
			<i>Axiothella</i> sp	6
			<i>Clymenella complanata</i>	1
			<i>Clymenura gracilis</i>	137
			Euclymeninae sp A	313
			<i>Isocirrus longiceps</i>	2
			<i>Petaloclymene pacifica</i>	57
			<i>Praxillella gracilis</i>	1
			<i>Praxillella pacifica</i>	338
			<i>Notoproctus pacificus</i>	24
			Maldaninae	4
			<i>Maldane sarsi</i>	90
			<i>Metasychis disparidentatus</i>	10
			<i>Petaloproctus neoborealis</i>	1
			<i>Rhodine bitorquata</i>	153
		Opheliidae	<i>Armandia brevis</i>	3
			<i>Ophelina acuminata</i>	1
			<i>Ophelina</i> sp SD1	2
			<i>Scoloplos acmeiceps</i>	1
			<i>Scoloplos armiger</i> Cmplx	335
		Paraonidae		1
			<i>Aricidea (Acmira) catherinae</i>	47
			<i>Aricidea (Acmira) lopezi</i>	14
			<i>Aricidea (Acmira) rubra</i>	1
			<i>Aricidea (Acmira) simplex</i>	30
			<i>Aricidea (Acmira)</i> sp SD1	1
			<i>Aricidea (Aricidea) pseudoarticulata</i>	1
			<i>Aricidea (Aricidea) wassi</i>	9
			<i>Aricidea (Aricidea)</i> sp	4
			<i>Aricidea (Strelzovia) antennata</i>	63
			<i>Aricidea (Strelzovia) hartleyi</i>	2
			<i>Aricidea (Strelzovia)</i> sp A	21
			<i>Cirrophorus furcatus</i>	1
			<i>Levinsenia gracilis</i>	21
			<i>Levinsenia kirbyae</i>	4
			<i>Paradoneis</i> sp SD1	1
			<i>Paradoneis</i> sp	3
		Scalibregmatidae	<i>Scalibregma californicum</i>	16
		Travisiidae	<i>Travisia brevis</i>	131
Arthropoda	Ostracoda	Cylindroleberididae	<i>Xenoleberis californica</i>	1
		Philomedidae	<i>Euphilomedes carcharodonta</i>	5
			<i>Euphilomedes producta</i>	24

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
	Malacostraca	Nebaliidae	<i>Nebalia pugettensis</i> Cmplx	1
		Caprellidae	<i>Caprella kennerlyi</i>	1
			<i>Mayerella banksia</i>	1
		Ischyroceridae	<i>Erichthonius brasiliensis</i>	9
		Photidae	<i>Photis bifurcata</i>	3
			<i>Photis californica</i>	9
			<i>Photis lacia</i>	5
			<i>Photis</i> sp	2
			<i>Podoceropsis ociosa</i>	4
		Aoridae	<i>Aoroides</i> sp A	3
			<i>Aoroides</i> sp	3
		Corophiidae	<i>Protomedeia articulata</i> Cmplx	12
		Oedicerotidae		1
			<i>Americhelidium shoemakeri</i>	12
			<i>Bathymedon pumilus</i>	13
			<i>Deflexilodes norvegicus</i>	9
			<i>Monoculodes emarginatus</i>	9
			<i>Westwoodilla tone</i>	8
		Eusiridae	<i>Rhachotropis</i> sp A	1
		Liljeborgiidae	<i>Listriella goleta</i>	2
			<i>Listriella melanica</i>	1
		Pleustidae	<i>Dactylopleustes</i> sp A	1
		Pardaliscidae	<i>Halicoides synopiae</i>	4
			<i>Nicippe tumida</i>	2
			<i>Pardaliscella symmetrica</i>	1
		Ampeliscidae	<i>Ampelisca agassizi</i>	6
			<i>Ampelisca brevisimulata</i>	59
			<i>Ampelisca</i> cf <i>brevisimulata</i>	6
			<i>Ampelisca careyi</i>	98
			<i>Ampelisca hancocki</i>	14
			<i>Ampelisca indentata</i>	2
			<i>Ampelisca lobata</i>	1
			<i>Ampelisca pacifica</i>	45
			<i>Ampelisca pugetica</i>	91
			<i>Ampelisca</i> sp	1
			<i>Byblis millsii</i>	4
		Argissidae	<i>Argissa hamatipes</i>	1
		Urothoidae	<i>Urothoe elegans</i> Cmplx	8
		Phoxocephalidae	<i>Foxiphalus similis</i>	2
			<i>Rhepoxynius bicuspidatus</i>	274
			<i>Rhepoxynius menziesi</i>	52
			<i>Rhepoxynius stenodes</i>	1
			<i>Eyakia robusta</i>	19
			<i>Heterophoxus oculatus</i>	34
		**	Lysianassoidea	1
		Lysianassidae	<i>Aruga holmesi</i>	1
			<i>Aruga oculata</i>	1
		Tryphosidae	<i>Hippomedon</i> sp A	2
		Acidostomatidae	<i>Acidostoma hancocki</i>	1
		Pakynidae	<i>Pachynus barnardi</i>	3

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
		Gnathiidae	<i>Caecognathia crenulatifrons</i>	26
		Anthuridae	<i>Haliophasma geminata</i>	14
		Serolidae	<i>Heteroserolis carinata</i>	2
		**	Tanaidacea	2
		Akanthophoreidae	<i>Chauliopleona dentata</i>	3
		Anarthruridae	<i>Siphonolabrum californiensis</i>	1
		Leptocheliidae	<i>Chondrochelia dubia</i> Cmplx	32
		Tanaellidae	<i>Araphura breviararia</i>	33
			<i>Araphura</i> sp SD1	10
			<i>Tanaella propinquus</i>	10
		Typhlotanaidae	<i>Typhlotanais williamsae</i>	3
		Tanaopsidae	<i>Tanaopsis cadieni</i>	12
		Leuconidae	<i>Eudorella pacifica</i>	1
		Nannastacidae	<i>Campylaspis canaliculata</i>	1
			<i>Procampylaspis caenosa</i>	14
		Diastylidae	<i>Diastylis crenellata</i>	14
			<i>Leptostylis abditis</i>	1
		**	Paguroidea	1
		Paguridae	<i>Pylopagurus holmesi</i>	1
		Cyclodorippidae	<i>Deilocerus planus</i>	1
		Pinnotheridae	<i>Pinnixa occidentalis</i> Cmplx	1
			<i>Pinnixa</i> sp	1
Nematoda				21
Echinodermata	Asteroidea			12
		Astropectinidae	<i>Astropecten californicus</i>	2
	Ophiuroidea			18
		Ophiuridae	<i>Ophiura luetkenii</i>	18
		Amphiuridae		415
			<i>Amphichondrius granulatus</i>	36
			<i>Amphiodia digitata</i>	39
			<i>Amphiodia urtica</i>	841
			<i>Amphiodia</i> sp	292
			<i>Amphioplus</i> sp	2
			<i>Amphiura arcystata</i>	11
			<i>Dougaloplus amphacanthus</i>	4
			<i>Dougaloplus</i> sp A	5
	Echinoidea			3
		Toxopneustidae	<i>Lytechinus pictus</i>	2
		**	Spatangoida	1
		Brissidae	<i>Brissopsis pacifica</i>	1
	Holothuroidea	Phylloporidae		1
		Synaptidae	<i>Leptosynapta</i> sp	10
		Chiridotidae	<i>Chiridota</i> sp	11
Phoronida				11
		Phoronidae	<i>Phoronis</i> sp SD1	4
			<i>Phoronis</i> sp	15
Chordata	Enteropneusta			4
		Ptychoderidae	<i>Balanoglossus</i> sp	1
		Spengelidae	<i>Schizocardium</i> sp	1

Appendix B.24 *continued*

Phylum	Class	Family	Taxon	n
		Harrimaniidae	<i>Saccoglossus</i> sp	2
			<i>Stereobalanus</i> sp	13
	Ascidiacea	Molgulidae	<i>Molgula pugetiensis</i>	1

Appendix B.25

Summary taxonomic listing of benthic infauna taxa identified from SBOO stations during 2020. Data are total number of individuals (n). Taxonomic arrangement follows SCAMIT (2018).** indicates taxon is not within previous family

Phylum	Class	Family	Taxon	n	
Cnidaria	Hydrozoa	Corymorphidae	<i>Corymorpha bigelowi</i>	2	
			<i>Euphysa</i> sp A	5	
		Campanulariidae	<i>Laomedea calceolifera</i>	5	
	Anthozoa		Pennatulacea		1
			Virgulariidae	<i>Stylatula</i> sp A	1
				<i>Virgularia</i> sp	1
		**		Ceriantharia	8
			Arachnactidae	<i>Arachnanthus</i> sp A	2
		**		Actiniaria	5
			Edwardsiidae		32
				<i>Edwardsia juliae</i>	14
				<i>Edwardsia olguini</i>	7
				<i>Scolanthus triangulus</i>	21
				Edwardsiidae sp SD1	1
				Halcampidae	10
				<i>Halcampa decemtentaculata</i>	10
				Isanthidae	8
				<i>Zaolutus actius</i>	8
				Limnactiniidae	5
		Limnactiniidae sp A	5		
		Haloclavidae	2		
		<i>Anemonactis</i> sp A	2		
Platyhelminthes	Rhabditophora	Callioplanidae	<i>Koinostylochus burchami</i>	1	
		Stylochidae	<i>Stylochus exiguus</i>	3	
		Cryptocelidae	<i>Cryptocelis occidentalis</i>	4	
		Plehnidae	<i>Diplehnia caeca</i>	2	
		**		Rhabditophora sp A	1
				Rhabditophora sp C	3
					3
					5
Nemertea	Anopla	Cephalotrichidae	<i>Cephalothrix</i> sp	38	
		**		Palaeonemertea	20
		Carinomidae	<i>Carinoma mutabilis</i>	57	
		Tubulanidae		6	
			<i>Tubulanus cingulatus</i>	13	
			<i>Tubulanus polymorphus</i>	91	
			<i>Tubulanus</i> sp A	1	
		**		<i>Heteronemertea</i>	1
			Lineidae		57
				<i>Cerebratulus californiensis</i>	2
				<i>Cerebratulus</i> sp	3
				Lineidae sp SD1	9
				<i>Lineus bilineatus</i>	19
				<i>Macaulaura alaskensis</i> Cmplx	12
				<i>Micrura wilsoni</i>	1
				<i>Zygeupolia rubens</i>	1
		**		Heteronemertea sp SD2	39
			Enopla		3
			Hoplonemertea	19	
	Emplectonematidae	<i>Cryptonemertes actinophila</i>	1		
		<i>Paranemertes californica</i>	8		

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
		Prosorhochmidae	<i>Prosorhochmus albidus</i>	3
		Oerstedidae	<i>Oerstedea dorsalis</i> Cmplx	17
		Amphiporidae	<i>Amphiporus flavescens</i>	5
		Tetrastemmatidae	<i>QuasitetraSTEMMA nigrifrons</i>	2
			<i>TetraSTEMMA candidum</i>	4
		**	Hoplonemertea sp A	1
			Hoplonemertea sp D	3
			Hoplonemertea sp SD3	1
Mollusca	Caudofoveata		Chaetodermatida	2
		Chaetodermatidae	<i>Chaetoderma pacificum</i>	1
			<i>Falcidens longus</i>	3
	Gastropoda	Trochidae	<i>Halistylus pupoideus</i>	2
		Cerithiidae	<i>Lirobittium rugatum</i> Cmplx	4
			<i>Lirobittium</i> sp	1
		Calyptraeidae		1
			<i>Calyptraea fastigiata</i>	1
		Naticidae	<i>Neverita recluziana</i>	1
		Rissoidae	<i>Alvania compacta</i>	1
		Barleeiidae	<i>Lirobarleeia kelseyi</i>	5
		Caecidae	<i>Caecum crebricinctum</i>	42
		Bursidae	<i>Crossata ventricosa</i>	1
		Epitoniidae	<i>Epitonium bellastriatum</i>	1
			<i>Epitonium sawinae</i>	2
		Eulimidae	<i>Balcis oldroydae</i>	15
			<i>Eulima raymondi</i>	1
			<i>Polygireulima rutila</i>	26
		Nassariidae	<i>Caesia perpinguis</i>	1
		Olivellidae	<i>Callianax baetica</i>	17
		Borsoniidae	<i>Ophiodermella inermis</i>	2
		Mangeliidae	<i>Kurtzia arteaga</i>	7
			<i>Kurtziella plumbea</i>	2
			<i>Kurtzina beta</i>	2
		Pseudomelatomidae	<i>Crassispira semiinflata</i>	3
		Terebridae	<i>Terebra pedroana</i>	5
		Acteonidae	<i>Rictaxis punctocaelatus</i>	1
		Pyramidellidae	<i>Odostomia</i> sp	2
			<i>Turbonilla chocolata</i>	1
			<i>Turbonilla santarosana</i>	1
			<i>Turbonilla</i> sp SD5	1
	<i>Turbonilla</i> sp SD7		3	
<i>Turbonilla</i> sp	1			
Onchidorididae	<i>Acanthodoris rhodoceras</i>	1		
Rhizoridae	<i>Volvulella cylindrica</i>	1		
	<i>Volvulella</i> sp	1		
Acteocinidae	<i>Acteocina cerealis</i>	2		
	<i>Acteocina culcitella</i>	2		
	<i>Acteocina harpa</i>	2		
Philinidae	<i>Philine auriformis</i>	36		
Gastropteridae	<i>Gastropteron pacificum</i>	5		

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
		Philinoglossidae	<i>Philinoglossa</i> sp A	3
		Cylichnidae	<i>Cylichna diegensis</i>	8
		Diaphanidae	<i>Diaphana californica</i>	4
	Bivalvia			3
		Nuculidae	<i>Ennucula tenuis</i>	1
		Solemyidae	<i>Solemya pervernica</i>	3
		Nuculanidae	<i>Nuculana hamata</i>	1
			<i>Nuculana taphria</i>	12
		Glycymerididae	<i>Glycymeris septentrionalis</i>	1
		Mytilidae	<i>Crenella decussata</i>	43
			<i>Solamen columbianum</i>	1
			Modiolinae	8
			<i>Amygdalum pallidulum</i>	4
		Pectinidae		5
			<i>Leptopecten latiauratus</i>	30
		Carditidae	<i>Cyclocardia ventricosa</i>	1
			<i>Cyclocardia</i> sp	2
		Lucinidae	<i>Lucinisca nuttalli</i>	1
			<i>Parvilucina tenuisculpta</i>	28
			<i>Lucinoma annulatum</i>	5
		Thyasiridae	<i>Axinopsida serricata</i>	5
			<i>Thyasira flexuosa</i>	4
		Lasaeidae	<i>Kurtiella grippi</i>	5
			<i>Kurtiella tumida</i>	31
		Cardiidae	<i>Keenaea centifilosum</i>	23
			<i>Trachycardium quadragenarium</i>	1
		Tellinidae		2
			<i>Tellina bodegensis</i>	3
			<i>Tellina carpenteri</i>	1
			<i>Tellina meropsis</i>	1
			<i>Tellina modesta</i>	99
			<i>Tellina</i> sp B	25
			<i>Macoma yoldiformis</i>	45
			<i>Macoma</i> sp	3
		Semelidae	<i>Semele venusta</i>	1
		**	Solenioidea	2
		Solenidae	<i>Solen sicarius</i>	5
		Pharidae	<i>Ensis myrae</i>	4
		Veneridae	Venerinae	1
			<i>Compsomyax subdiaphana</i>	6
		Petricolidae	<i>Cooperella subdiaphana</i>	14
		Mactridae	<i>Simomactra falcata</i>	16
		Lyonsiidae		10
			<i>Lyonsia californica</i>	6
		Thraciidae		2
		Periplomatidae	<i>Periploma discus</i>	1
		Cuspidariidae	<i>Cardiomya pectinata</i>	2
	Scaphopoda			7
		Dentaliidae	<i>Dentalium vallicolens</i>	1
		Gadilidae	<i>Polyschides quadrifissatus</i>	35

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Gadila aberrans</i>	135
Sipuncula				14
	Sipunculidea	Golfingiidae	<i>Thysanocardia nigra</i>	24
		Phascolionidae	<i>Phascolion</i> sp A	16
		Sipunculidae	<i>Siphonosoma ingens</i>	2
			<i>Sipunculus nudus</i>	1
	Phascolosomatidea	Phascolosomatidae	<i>Apionsoma misakianum</i>	51
Annelida	Polychaeta	Amphinomidae	<i>Chloeia pinnata</i>	12
			<i>Paramphinome</i> sp	280
			<i>Pareurythoe californica</i>	44
		Dorvilleidae		2
			<i>Dorvillea (Dorvillea)</i> sp	1
			<i>Meiodorvillea</i> sp SD1	1
			<i>Ophryotrocha</i> sp	2
			<i>Parougia caeca</i>	5
			<i>Protodorvillea gracilis</i>	293
		Eunicidae		6
			<i>Leodice americana</i>	6
			<i>Marphysa disjuncta</i>	4
			<i>Marphysa</i> sp	1
		Lumbrineridae		2
			Lumbrineridae Group III	1
			<i>Lumbrinerides platypygos</i>	33
			<i>Lumbrineris cruzensis</i>	12
			<i>Lumbrineris japonica</i>	1
			<i>Lumbrineris latreilli</i>	40
			<i>Lumbrineris ligulata</i>	31
			<i>Lumbrineris limicola</i>	1
			<i>Lumbrineris</i> sp Group I	3
			<i>Lumbrineris</i> sp	2
			<i>Scoletoma tetraura</i> Cmplx	11
		Oeonidae	<i>Arabella iricolor</i> Cmplx	1
			<i>Drilonereis falcata</i>	2
			<i>Drilonereis</i> sp	3
		Onuphidae		125
			<i>Diopatra ornata</i>	6
			<i>Diopatra splendidissima</i>	5
			<i>Diopatra tridentata</i>	7
			<i>Diopatra</i> sp	28
			<i>Mooreonuphis exigua</i>	2
			<i>Mooreonuphis nebulosa</i>	18
			<i>Mooreonuphis</i> sp SD1	51
			<i>Mooreonuphis</i> sp SD2	9
			<i>Mooreonuphis</i> sp	87
			<i>Nothria occidentalis</i>	1
			<i>Nothria</i> sp	5
			<i>Onuphis affinis</i>	1
			<i>Onuphis elegans</i>	2
			<i>Onuphis eremita parva</i>	8
			<i>Onuphis iridescens</i>	9

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Onuphis</i> sp A	40
			<i>Onuphis</i> sp	5
			<i>Paradiopatra parva</i>	61
			<i>Rhamphobrachium longisetosum</i>	1
		Aphroditidae		5
			<i>Aphrodita</i> sp	2
		Polynoidae	<i>Lepidasthenia longicirrata</i>	1
			<i>Lepidonotus</i> cf <i>squamatus</i>	1
			<i>Malmgreniella macginitiei</i>	2
			<i>Malmgreniella scriptoria</i>	1
			<i>Malmgreniella</i> sp A	1
			<i>Malmgreniella</i> sp	1
			<i>Tenonia priops</i>	14
		Sigalionidae	<i>Pholoides asperus</i>	1
			<i>Pisione</i> sp	198
			<i>Sigalion spinosus</i>	180
			<i>Sthenelais tertiaglabra</i>	4
			<i>Sthenelais verruculosa</i>	3
			<i>Sthenelanella uniformis</i>	123
		Glyceridae	<i>Glycera americana</i>	3
			<i>Glycera nana</i>	20
			<i>Glycera oxycephala</i>	87
			<i>Glycera tessellata</i>	2
			<i>Glycera</i> sp	30
			<i>Hemipodia borealis</i>	3
		Goniadidae	<i>Glycinde armigera</i>	68
			<i>Goniada acicula</i>	1
			<i>Goniada littorea</i>	14
			<i>Goniada maculata</i>	12
			<i>Goniada</i> sp	1
		Chrysopetalidae	<i>Chrysopetalum occidentale</i>	1
		Hesionidae	<i>Gyptis brunnea</i>	6
			<i>Heteropodarke heteromorpha</i>	3
			<i>Microphthalmus hystrix</i>	22
			<i>Micropodarke dubia</i>	26
			<i>Oxydromus pugettensis</i>	14
			<i>Podarkeopsis glabrus</i>	1
		Nereididae	<i>Nereis</i> sp A	39
			<i>Platynereis bicanaliculata</i>	38
		Pilargidae	<i>Ancistrosyllis groenlandica</i>	1
			<i>Hermundura fauveli</i>	1
			<i>Hermundura ocularis</i>	1
		Syllidae	<i>Epigamia-Myrianida</i> Cmplx	3
			<i>Eusyllis blomstrandii</i> Cmplx	3
			<i>Eusyllis habeii</i>	20
			<i>Eusyllis transecta</i>	11
			<i>Eusyllis</i> sp SD2	29
			<i>Eusyllis</i> sp	5
			<i>Odontosyllis phosphorea</i>	33
			<i>Opisthodonta</i> sp SD2	1

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Paraehlersia articulata</i>	1
			<i>Exogone dwisula</i>	3
			<i>Exogone lourei</i>	25
			<i>Parexogone acutipalpa</i>	2
			<i>Parexogone breviseta</i>	2
			<i>Parexogone molesta</i>	1
			<i>Salvatoria mediodentata</i>	1
			<i>Sphaerosyllis californiensis</i>	11
			<i>Syllis farallonensis</i>	4
			<i>Syllis heterochaeta</i>	23
			<i>Syllis</i> sp SD1	10
			<i>Syllis</i> sp SD2	14
			<i>Xenosyllis</i> sp	1
		Nephtyidae	<i>Bipalponephtys cornuta</i>	3
			<i>Nephtys caecoides</i>	13
			<i>Nephtys ferruginea</i>	4
			<i>Nephtys simoni</i>	19
			<i>Nephtys</i> sp SD2	9
			<i>Nephtys</i> sp	1
		Sphaerodoridae	<i>Sphaerodoropsis biserialis</i>	1
		Phyllodocidae		1
			<i>Eteone brigitteae</i>	1
			<i>Eteone pigmentata</i>	1
			<i>Eulalia californiensis</i>	2
			<i>Eulalia levicornuta</i> Cmplx	1
			<i>Eulalia</i> sp SD1	34
			<i>Eumida longicornuta</i>	9
			<i>Hesionura coineaui difficilis</i>	264
			<i>Sige</i> sp A	19
			<i>Nereiphylla ferruginea</i> Cmplx	1
			<i>Nereiphylla</i> sp 2	3
			<i>Nereiphylla</i> sp SD1	2
			<i>Nereiphylla</i> sp	1
			<i>Phyllodoce groenlandica</i>	1
			<i>Phyllodoce hartmanae</i>	29
			<i>Phyllodoce longipes</i>	27
			<i>Phyllodoce medipapillata</i>	19
			<i>Phyllodoce pettiboneae</i>	27
		Fabriciidae	<i>Pseudofabriciola californica</i>	9
		Oweniidae	<i>Galathowenia pygidialis</i>	6
			<i>Myriochele gracilis</i>	22
			<i>Myriochele olgae</i>	1
			<i>Myriochele striolata</i>	21
			<i>Owenia collaris</i>	8
		Sabellidae		1
			<i>Acromegalomma pigmentum</i>	7
			<i>Acromegalomma splendidum</i>	1
			<i>Bispira</i> sp	1
			<i>Dialychone albocincta</i>	7
			<i>Dialychone trilineata</i>	11

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Dialychnone veleronis</i>	37
			<i>Euchone arenae</i>	41
			<i>Euchone hancocki</i>	4
			<i>Euchone incolor</i>	53
			<i>Euchone</i> sp	2
			<i>Jasmineira</i> sp B	347
			<i>Paradialychnone bimaculata</i>	10
			<i>Paradialychnone ecaudata</i>	5
			<i>Paradialychnone harrisae</i>	45
			<i>Paradialychnone paramollis</i>	8
			<i>Potamethus</i> sp A	1
		Magelonidae	<i>Magelona hartmanae</i>	10
			<i>Magelona pitelkai</i>	2
		Poecilochaetidae	<i>Poecilochaetus johnsoni</i>	17
			<i>Poecilochaetus</i> sp	1
		Spionidae		3
			<i>Aonides</i> sp	11
			<i>Dipolydora socialis</i>	15
			<i>Dispio</i> sp SD1	23
			<i>Laonice cirrata</i>	9
			<i>Laonice nuchala</i>	1
			<i>Laonice</i> sp	1
			<i>Malacoceros indicus</i>	1
			<i>Microspio pigmentata</i>	13
			<i>Paraprionospio alata</i>	75
			<i>Polydora cirrosa</i>	1
			<i>Prionospio dubia</i>	29
			<i>Prionospio jubata</i>	75
			<i>Prionospio lighti</i>	3
			<i>Prionospio pygmaeus</i>	71
			<i>Scolelepis (Scolelepis) occidentalis</i>	7
			<i>Spio maculata</i>	25
			<i>Spiophanes berkeleyorum</i>	37
			<i>Spiophanes duplex</i>	867
			<i>Spiophanes kimballi</i>	42
			<i>Spiophanes norrisi</i>	850
			<i>Spiophanes wigleyi</i>	1
			<i>Spiophanes</i> sp	1
		Acrocirridae	<i>Macrochaeta</i> sp A	3
		Cirratulidae	<i>Aphelochaeta glandaria</i> Cmplx	9
			<i>Aphelochaeta monilaris</i>	5
			<i>Aphelochaeta tigrina</i>	6
			<i>Aphelochaeta williamsae</i>	3
			<i>Aphelochaeta</i> sp LA1	9
			<i>Aphelochaeta</i> sp	4
			<i>Caulleriella hamata</i>	1
			<i>Caulleriella pacifica</i>	11
			<i>Caulleriella</i> sp SD2	3
			<i>Caulleriella</i> sp	12
			<i>Chaetozone armata</i>	7

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Chaetozone bansei</i>	1
			<i>Chaetozone columbiana</i>	4
			<i>Chaetozone commonalis</i>	1
			<i>Chaetozone corona</i>	39
			<i>Chaetozone hartmanae</i>	40
			<i>Chaetozone lunula</i>	2
			<i>Chaetozone</i> sp SD2	8
			<i>Chaetozone</i> sp SD5	13
			<i>Chaetozone</i> sp SD7	2
			<i>Chaetozone</i> sp	9
			<i>Cirratulus</i> sp	3
			<i>Kirkegaardia cryptica</i>	7
			<i>Kirkegaardia sibilina</i>	77
			<i>Kirkegaardia tessellata</i>	21
		Fauveliopsidae	<i>Fauveliopsis</i> sp SD1	1
		Flabelligeridae		1
			<i>Pherusa neopapillata</i>	6
			<i>Piromis</i> sp	2
		Sternaspidae	<i>Sternaspis affinis</i>	6
		Ampharetidae		8
			<i>Amage anops</i>	6
			<i>Amage scutata</i>	4
			<i>Ampharete acutifrons</i>	1
			<i>Ampharete finmarchica</i>	7
			<i>Ampharete labrops</i>	70
			<i>Ampharete</i> sp	1
			<i>Ampharetidae</i> sp SD1	7
			<i>Amphicteis scaphobranchiata</i>	16
			<i>Anobothrus gracilis</i>	9
			<i>Asabellides lineata</i>	6
			<i>Eclysippe trilobata</i>	23
			<i>Lysippe</i> sp A	9
			<i>Lysippe</i> sp B	3
			<i>Lysippe</i> sp	1
			<i>Sabellides manriquei</i>	51
			<i>Samytha californiensis</i>	3
			<i>Schistocomus</i> sp A	1
			<i>Schistocomus</i> sp	6
			<i>Melinna oculata</i>	48
		Pectinariidae	<i>Pectinaria californiensis</i>	12
		Terebellidae		2
			<i>Amaeana occidentalis</i>	11
			<i>Polycirrus californicus</i>	1
			<i>Polycirrus</i> sp I	5
			<i>Polycirrus</i> sp A	49
			<i>Polycirrus</i> sp SD3	58
			<i>Polycirrus</i> sp	41
			<i>Artacama coniferi</i>	2
			<i>Eupolymnia heterobranchia</i>	1
			<i>Lanassa venusta venusta</i>	190

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Phisidia sanctaemariae</i>	9
			<i>Pista brevibranchiata</i>	6
			<i>Pista estevanica</i>	55
			<i>Pista wui</i>	43
			<i>Pista</i> sp	3
			<i>Streblosoma crassibranchia</i>	7
			<i>Streblosoma</i> sp B	7
			<i>Streblosoma</i> sp C	33
			<i>Streblosoma</i> sp	8
		Trichobranchidae	<i>Terebellides californica</i>	1
		Chaetopteridae	<i>Chaetopterus variopedatus</i> Cmplx	1
			<i>Mesochaetopterus</i> sp	1
			<i>Phyllochaetopterus prolifica</i>	3
			<i>Spiochaetopterus costarum</i> Cmplx	160
		Capitellidae		3
			<i>Capitella teleta</i>	13
			<i>Decamastus gracilis</i>	1
			<i>Mediomastus acutus</i>	3
			<i>Mediomastus</i> sp	341
			<i>Notomastus hemipodus</i>	6
			<i>Notomastus latericeus</i>	12
			<i>Notomastus lineatus</i>	8
		Cossuridae	<i>Cossura</i> sp A	2
		Maldanidae		57
			Euclymeninae	25
			<i>Axiothella</i> sp	3
			<i>Clymenella complanata</i>	1
			<i>Clymenella</i> sp A	1
			<i>Clymenella</i> sp	2
			<i>Clymenura gracilis</i>	1
			Euclymeninae sp A	98
			<i>Petaloclymene pacifica</i>	53
			<i>Praxillella pacifica</i>	68
			<i>Maldane sarsi</i>	11
			<i>Metasychis disparidentatus</i>	34
			<i>Praxillura maculata</i>	5
			<i>Rhodine bitorquata</i>	1
		Opheliidae		2
			<i>Armandia brevis</i>	1
			<i>Ophelia pulchella</i>	23
		Orbiniidae		3
			<i>Leitoscoloplos pugettensis</i>	10
			<i>Naineris uncinata</i>	6
			<i>Naineris</i> sp	1
			<i>Scoloplos acmeceps</i>	11
			<i>Scoloplos armiger</i> Cmplx	67
		Paraonidae	<i>Aricidea (Acmira) catherinae</i>	7
			<i>Aricidea (Acmira) cerrutii</i>	12
			<i>Aricidea (Acmira) lopezi</i>	1
			<i>Aricidea (Acmira) rubra</i>	1

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n		
Arthropoda			<i>Aricidea (Acmira) simplex</i>	16		
			<i>Aricidea (Aedicira) pacifica</i>	1		
			<i>Aricidea (Aricidea) wassi</i>	5		
			<i>Aricidea (Aricidea) sp</i>	2		
			<i>Aricidea (Strelzovia) antennata</i>	3		
			<i>Aricidea (Strelzovia) hartleyi</i>	2		
			<i>Cirrophorus furcatus</i>	5		
			<i>Levinsenia gracilis</i>	12		
			<i>Levinsenia kirbyae</i>	2		
			<i>Paradoneis</i> sp SD1	23		
			<i>Paradoneis</i> sp	2		
			Scalibregmatidae	<i>Scalibregma californicum</i>	64	
			Travisiidae	<i>Travisia brevis</i>	6	
				<i>Travisia gigas</i>	7	
				<i>Travisia pupa</i>	4	
				Polygordiidae	<i>Polygordius</i> sp	9
				Saccocirridae	<i>Saccocirrus</i> sp	1
			Clitellata		Oligochaeta	4
			Pycnogonida	Phoxichilidiidae	<i>Anoplodactylus erectus</i>	1
			Ostracoda	Cylindroleberididae	<i>Leuroleberis sharpei</i>	2
					<i>Xenoleberis californica</i>	1
				Philomedidae	<i>Euphilomedes carcharodonta</i>	85
				Sarsiellidae	<i>Eusarsiella thominx</i>	2
			Malacostraca	Nebaliidae	<i>Nebalia daytoni</i>	14
					<i>Nebalia pugettensis</i> Cmplx	7
				Mysidae		1
					<i>Neomysis kadiakensis</i>	3
					<i>Metamysidopsis elongata</i>	9
				Caprellidae		3
					<i>Caprella mendax</i>	13
					<i>Hemiproto</i> sp A	1
				Ischyroceridae	<i>Notopoma</i> sp A	72
				Kamakidae	<i>Amphideutopus oculatus</i>	18
				Photidae	<i>Ampelisciphotis podophthalma</i>	14
					<i>Gammaropsis thompsoni</i>	13
					<i>Photis bifurcata</i>	6
					<i>Photis brevipes</i>	19
					<i>Photis californica</i>	43
					<i>Photis lacia</i>	5
					<i>Photis macinerneyi</i>	1
		<i>Photis</i> sp C	5			
		<i>Photis</i> sp OC1	42			
		<i>Photis</i> sp	9			
	Aoridae	<i>Aoroides intermedia</i>	1			
		<i>Aoroides</i> sp	2			
	Unciolidae	<i>Rudilemboides stenopropodus</i>	5			
	Corophiidae	<i>Laticorophium baconi</i>	16			
	Megaluropidae	<i>Gibberosus myersi</i>	8			
		<i>Megaluropidae</i> sp A	9			
	Oedicerotidae	<i>Americhelidium shoemakeri</i>	5			

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Americhelidium</i> sp SD1	10
			<i>Americhelidium</i> sp SD4	1
			<i>Deflexilodes norvegicus</i>	3
			<i>Hartmanodes hartmanae</i>	5
			<i>Westwoodilla tone</i>	1
		Liljeborgiidae	<i>Listriella albina</i>	1
			<i>Listriella goleta</i>	2
		Stenothoidae	<i>Metopa dawsoni</i>	1
		Melphidippidae	<i>Melphisana bola</i> Cmplx	1
		Ampeliscidae	<i>Ampelisca agassizi</i>	23
			<i>Ampelisca brachycladus</i>	64
			<i>Ampelisca brevisimulata</i>	71
			<i>Ampelisca cf brevisimulata</i>	2
			<i>Ampelisca careyi</i>	28
			<i>Ampelisca cristata cristata</i>	63
			<i>Ampelisca cristata microdentata</i>	83
			<i>Ampelisca hancocki</i>	2
			<i>Ampelisca milleri</i>	1
			<i>Ampelisca pugetica</i>	40
			<i>Ampelisca</i> sp	20
			<i>Byblis millsii</i>	20
		Synopiidae	<i>Tiron biocellata</i>	2
		Argissidae	<i>Argissa hamatipes</i>	6
		Platyischnopidae	<i>Tiburonella viscana</i>	2
		Urothoidae	<i>Urothoe elegans</i> Cmplx	5
		Phoxocephalidae	<i>Foxiphalus golfensis</i>	8
			<i>Foxiphalus obtusidens</i>	45
			<i>Rhepoxynius fatigans</i>	2
			<i>Rhepoxynius heterocuspoidatus</i>	156
			<i>Rhepoxynius lucubrans</i>	13
			<i>Rhepoxynius menziesi</i>	89
			<i>Rhepoxynius stenodes</i>	31
			<i>Rhepoxynius variatus</i>	2
			<i>Rhepoxynius</i> sp	2
			<i>Eyakia robusta</i>	1
			<i>Metaphoxus frequens</i>	1
		Lysianassidae	<i>Aruga holmesi</i>	1
			<i>Aruga oculata</i>	4
		Tryphosidae	<i>Hippomedon columbianus</i>	1
			<i>Hippomedon zetesimus</i>	7
			<i>Lepidepecreum serraculum</i>	1
			<i>Orchomenella pinguis</i>	1
		Acidostomatidae	<i>Acidostoma hancocki</i>	3
		Pakynidae	<i>Pachynus barnardi</i>	1
		Cirolanidae	<i>Eurydice caudata</i>	15
		Gnathiidae	<i>Caecognathia crenulatifrons</i>	22
		Anthuridae	<i>Haliophasma geminata</i>	18
		Idoteidae	<i>Edotia sublittoralis</i>	12
			<i>Edotia</i> sp B	3
		Sphaeromatidae	<i>Exosphaeroma</i> sp	1

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
		Joeropsididae	<i>Joeropsis dubia</i>	1
		Leptocheliidae	<i>Chondrochelia dubia</i> Cmplx	260
		Bodotriidae	<i>Cyclaspis nubila</i>	10
		Nannastacidae	<i>Campylaspis canaliculata</i>	5
			<i>Campylaspis maculinodulosa</i>	2
			<i>Procampylaspis caenosa</i>	5
		Lampropidae	<i>Hemilamprops californicus</i>	40
			<i>Mesolamprops bispinosus</i>	5
		Diastylidae	<i>Anchicolurus occidentalis</i>	2
			<i>Diastylis californica</i>	5
			<i>Diastylopsis tenuis</i>	3
			<i>Oxyurostylis pacifica</i>	15
		**	Decapoda	1
		Crangonidae	<i>Crangon alaskensis</i>	7
		Diogenidae	<i>Paguristes turgidus</i>	1
			<i>Paguristes ulreyi</i>	1
		Paguridae	<i>Pylopagurus holmesii</i>	1
		Albuneidae	<i>Lepidopa californica</i>	2
		Cyclodorippidae	<i>Deilocerus planus</i>	1
		Cancriidae		1
		Pinnotheridae	<i>Pinnixa franciscana</i>	2
			<i>Pinnixa longipes</i>	5
			<i>Pinnixa</i> sp	65
Nematoda				158
Echinodermata	Asteroidea			14
		Astropectinidae	<i>Astropecten armatus</i>	1
			<i>Astropecten californicus</i>	9
			<i>Astropecten</i> sp	5
	Ophiuroidea			5
		Ophiuridae	<i>Ophiura luetkenii</i>	41
		Ophioscolecidae	<i>Ophiuroconis bispinosa</i>	41
		Amphiuridae		56
			<i>Amphiodia digitata</i>	6
			<i>Amphiodia psara</i>	5
			<i>Amphiodia urtica</i>	4
			<i>Amphiodia</i> sp	23
			<i>Amphioplus</i> sp A	1
			<i>Amphipholis squamata</i>	27
			<i>Dougaloplus</i> sp A	3
		Ophiotrichidae	<i>Ophiotrix spiculata</i>	1
	Echinoidea			10
		Toxopneustidae	<i>Lytechinus pictus</i>	1
		Dendrasteridae	<i>Dendraster terminalis</i>	184
		**	Spatangoida	2
		Maretiidae	<i>Nacospatangus laevis</i>	1
		Loveniidae	<i>Lovenia cordiformis</i>	13
	Holothuroidea	Phylloporidae		4
		Synaptidae	<i>Leptosynapta</i> sp	1
Phoronida				3
		Phoronidae	<i>Phoronis</i> sp SD1	8

Appendix B.25 *continued*

Phylum	Class	Family	Taxon	n
			<i>Phoronis</i> sp	50
			<i>Phoronopsis</i> sp	7
Brachiopoda	Lingulata	Lingulidae	<i>Glottidia albida</i>	30
Chordata	Enteropneusta			2
		Spengeliidae	<i>Schizocardium</i> sp	12
		Harrimaniidae	<i>Saccoglossus</i> sp	3
			<i>Stereobalanus</i> sp	4
	Ascidiacea			1
		Agneziidae	<i>Agnezia septentrionalis</i>	1
		**	Stolidobranchiata	1
		Styelidae	<i>Cnemidocarpa rhizopus</i>	14
		Molgulidae	<i>Eugyra glutinans</i>	2
	Leptocardii	Branchiostomatidae	<i>Branchiostoma californiense</i>	14

Appendix B.26

Summary taxonomic listing of benthic infauna taxa identified from the San Diego regional benthic stations sampled during summer 2020. Data are total number of individuals (n). Taxonomic arrangement follows SCAMIT (2018).** indicates taxon is not within previous family

Phylum	Class	Family	Taxon	n	
Cnidaria	Hydrozoa	Corymorphidae	<i>Euphysa</i> sp A	3	
		Anthozoa	Virgulariidae	<i>Stylatula</i> sp A	2
				<i>Stylatula</i> sp	1
			**	Ceriantharia	2
				Actiniaria	6
			Edwardsiidae		30
				<i>Edwardsia juliae</i>	17
				<i>Edwardsia olguini</i>	6
				<i>Edwardsia profunda</i>	1
				<i>Scolanthus triangulus</i>	39
			Halcampidae	<i>Halcampa decemtentaculata</i>	1
				<i>Halianthella</i> sp A	3
			Isanthidae	<i>Zaolutus actius</i>	23
			Limnactiniidae	Limnactiniidae sp A	1
	Platyhelminthes	Rhabditophora	Stylochidae	<i>Stylochus exiguus</i>	4
			<i>Stylochus</i> sp	1	
Plehnidae			<i>Diplehnia caeca</i>	5	
Nemertea	Anopla	**	Leptoplanoidea	1	
			<i>Palaeonemertea</i>	10	
		Carinomidae	<i>Carinoma mutabilis</i>	39	
		Tubulanidae		5	
			<i>Tubulanus cingulatus</i>	8	
			<i>Tubulanus polymorphus</i>	58	
			<i>Tubulanus</i> sp A	5	
		**	Heteronemertea	2	
		Lineidae		33	
			<i>Cerebratulus californiensis</i>	2	
			<i>Cerebratulus</i> sp	1	
			Lineidae sp SD1	11	
			<i>Lineus bilineatus</i>	8	
			<i>Maculaura alaskensis</i> Cmplx	2	
			<i>Zygeupolia rubens</i>	5	
	**	Heteronemertea sp SD2	41		
	Enopla	Hoplonemertea	5		
		Emplectonematidae	<i>Paranemertes californica</i>	9	
		Prosorhochmidae	<i>Prosorhochmus albidus</i>	1	
		Oerstedidae	<i>Oerstedtia dorsalis</i> Cmplx	1	
		Tetrastemmatidae	<i>Tetrastemma candidum</i>	1	
		**	Hoplonemertea sp A	1	
			Hoplonemertea sp D	1	
Mollusca	Caudofoveata	Chaetodermatidae	<i>Chaetoderma marinelli</i>	3	
			<i>Chaetoderma nanulum</i>	1	
			<i>Chaetoderma pacificum</i>	5	
			<i>Falcidens longus</i>	5	
			Limifossoridae	<i>Limifossor fratula</i>	1
			Polyplacophora	Leptochitonidae	<i>Leptochiton rugatus</i>

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
	Gastropoda			1
		Solariellidae	<i>Solariella peramabilis</i>	2
		Calyptraeidae	<i>Calyptraea fastigiata</i>	1
		Naticidae	<i>Neverita recluziana</i>	2
		Eulimidae	<i>Balcis oldroydae</i>	2
			<i>Eulima raymondi</i>	1
			<i>Polygireulima rutila</i>	1
		Nassariidae	<i>Tritia insculpta</i>	2
		Muricidae		1
		Olivellidae	<i>Callianax baetica</i>	38
		Mangeliidae	<i>Kurtziella plumbea</i>	3
			<i>Kurtzina beta</i>	2
		Terebridae	<i>Terebra pedroana</i>	1
		Acteonidae	<i>Rictaxis punctocaelatus</i>	1
		Pyramidellidae	<i>Odostomia</i> sp	6
			<i>Turbonilla santarosana</i>	1
			<i>Turbonilla</i> sp A	6
			<i>Turbonilla</i> sp SD5	1
			<i>Turbonilla</i> sp SD9	2
		Rhizoridae	<i>Volvulella californica</i>	3
			<i>Volvulella cylindrica</i>	4
			<i>Volvulella panamica</i>	3
		Philinidae	<i>Philine auriformis</i>	24
		Gastropteridae	<i>Gastropteran pacificum</i>	4
		Cylichnidae	<i>Cylichna diegensis</i>	6
		Diaphanidae	<i>Diaphana californica</i>	1
	Bivalvia			6
		Nuculidae	<i>Acila castrensis</i>	2
			<i>Ennucula tenuis</i>	14
		**	Nuculanida	1
		Nuculanidae	<i>Nuculana conceptionis</i>	2
			<i>Nuculana hamata</i>	5
			<i>Nuculana taphria</i>	10
			<i>Nuculana</i> sp A	65
			<i>Nuculana</i> sp B	1
			<i>Nuculana</i> sp	1
		Yoldiidae	<i>Yoldiella nana</i>	8
		Mytilidae	<i>Solamen columbianum</i>	1
			<i>Dacrydium pacificum</i>	1
			Modiolinae	6
			<i>Amygdalum pallidulum</i>	4
		Pectinidae	<i>Delectopecten vancouverensis</i>	3
			<i>Leptopecten latiauratus</i>	1
		Lucinidae	<i>Parvilucina tenuisculpta</i>	63
			<i>Lucinoma annulatum</i>	7
		Thyasiridae	<i>Adontorhina cyclica</i>	12
			<i>Axinopsida serricata</i>	577
			<i>Thyasira flexuosa</i>	7
		Galeommatidae	<i>Waldo arthuri</i>	1

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
		Lasaeidae	<i>Kurtiella grippi</i>	3
			<i>Kurtiella tumida</i>	5
		Cardiidae	<i>Keenaea centifilosum</i>	8
		Tellinidae		2
			<i>Tellina carpenteri</i>	36
			<i>Tellina modesta</i>	38
			<i>Tellina</i> sp B	98
			<i>Tellina</i> sp	1
			<i>Macoma carlottensis</i>	5
			<i>Macoma yoldiformis</i>	21
			<i>Macoma</i> sp	1
		Solenidae	<i>Solen sicarius</i>	1
		Pharidae	<i>Ensis myrae</i>	2
		Hiatellidae	<i>Saxicavella pacifica</i>	4
		Veneridae	<i>Compsomyax subdiaphana</i>	2
		Petricolidae	<i>Cooperella subdiaphana</i>	11
		Mactridae	<i>Simomactra falcata</i>	3
		Lyonsiidae	<i>Lyonsia californica</i>	1
		**	Thracioidea	2
		Periplomatidae	<i>Periploma</i> sp	1
		Cuspidariidae	<i>Cuspidaria parapodema</i>	5
Mollusca	Scaphopoda			3
		Dentaliidae	<i>Dentalium vallicolens</i>	2
		Rhabdidae	<i>Rhabdus rectius</i>	4
		Gadilidae	<i>Polyschides quadrifissatus</i>	20
			<i>Cadulus californicus</i>	3
			<i>Gadila aberrans</i>	69
		**	<i>Compressidens stearnsii</i>	2
Sipuncula	Sipunculidea	Golfingiidae	<i>Nephasoma diaphanes</i>	2
			<i>Thysanocardia nigra</i>	2
		Phascolionidae	<i>Phascolion</i> sp A	32
		Sipunculidae	<i>Siphonosoma ingens</i>	4
Annelida	Polychaeta		<i>Echiura</i>	1
		Thalassematidae	<i>Listriolobus hexamyotus</i>	1
			<i>Listriolobus pelodes</i>	1
		Amphinomidae	<i>Chloeia pinnata</i>	4
			<i>Pareurythoe californica</i>	1
		Dorvilleidae	<i>Meiodorvillea</i> sp SD1	1
		Eunicidae		1
			<i>Leodice americana</i>	1
			<i>Marphysa disjuncta</i>	1
		Lumbrineridae	<i>Eranno lagunae</i>	1
			<i>Lumbrinerides platypygos</i>	1
			<i>Lumbrineris cruzensis</i>	18
			<i>Lumbrineris latreilli</i>	3
			<i>Lumbrineris ligulata</i>	3
			<i>Lumbrineris</i> sp Group I	9
			<i>Lumbrineris</i> sp Group II	3
			<i>Lumbrineris</i> sp	3

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Ninoe tridentata</i>	2
			<i>Scoletoma tetraura</i> Cmplx	62
			<i>Scoletoma</i> sp	21
		Oeononidae		2
			<i>Drilonereis falcata</i>	2
			<i>Drilonereis</i> sp	3
		Onuphidae		4
			<i>Diopatra ornata</i>	3
			<i>Diopatra splendidissima</i>	4
			<i>Diopatra tridentata</i>	1
			<i>Diopatra</i> sp	12
			<i>Mooreonuphis exigua</i>	2
			<i>Mooreonuphis nebulosa</i>	6
			<i>Mooreonuphis</i> sp SD1	3
			<i>Mooreonuphis</i> sp SD2	1
			<i>Mooreonuphis</i> sp	12
			<i>Nothria occidentalis</i>	11
			<i>Onuphis affinis</i>	4
			<i>Onuphis eremita parva</i>	3
			<i>Onuphis iridescens</i>	6
			<i>Onuphis</i> sp A	21
			<i>Paradiopatra parva</i>	245
		Acoetidae	<i>Acoetes pacifica</i>	3
		Polynoidae		1
			<i>Eucranta anoculata</i>	2
			<i>Malmgreniella bansei</i>	1
			<i>Malmgreniella baschi</i>	4
			<i>Malmgreniella macginitiei</i>	3
			<i>Malmgreniella nigralba</i>	2
			<i>Malmgreniella sanpedroensis</i>	1
			<i>Malmgreniella scriptoria</i>	2
			<i>Malmgreniella</i> sp A	5
			<i>Subadyte mexicana</i>	5
			<i>Tenonia priops</i>	12
		Pholoidae	<i>Pholoe glabra</i>	58
		Sigalionidae	<i>Pholoides asperus</i>	2
			<i>Pisione</i> sp	1
			<i>Sigalion spinosus</i>	49
			<i>Sthenelais tertiaglabra</i>	4
			<i>Sthenelais verruculosa</i>	5
			<i>Sthenelanella uniformis</i>	15
		Glyceridae	<i>Glycera americana</i>	3
			<i>Glycera macrobranchia</i>	7
			<i>Glycera nana</i>	53
			<i>Glycera oxycephala</i>	15
		Goniadidae	<i>Glycinde armigera</i>	35
			<i>Goniada brunnea</i>	5
			<i>Goniada littorea</i>	12
			<i>Goniada maculata</i>	63

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
		Hesionidae	<i>Heteropodarke heteromorpha</i>	1
			<i>Micropodarke dubia</i>	1
			<i>Podarkeopsis glabrus</i>	6
		Nereididae	<i>Ceratocephale loveni</i>	1
			<i>Nereis</i> sp A	18
			<i>Nicon moniloceras</i>	1
			<i>Platynereis bicanaliculata</i>	4
		Pilargidae	<i>Ancistrosyllis groenlandica</i>	3
			<i>Hermundura fauveli</i>	1
			<i>Hermundura ocularis</i>	1
			<i>Sigambra bassi</i>	1
			<i>Sigambra setosa</i>	1
		Syllidae	<i>Epigamia-Myrianida</i> Cmplx	1
			<i>Eusyllis habeii</i>	1
			<i>Eusyllis longicirrata</i>	12
			<i>Eusyllis transecta</i>	1
			<i>Odontosyllis phosphorea</i>	2
			<i>Paraehlersia articulata</i>	1
			<i>Exogone lourei</i>	3
			<i>Parexogone molesta</i>	1
			<i>Syllis heterochaeta</i>	8
		Nephtyidae	<i>Aglaophamus erectans</i>	1
			<i>Aglaophamus verrilli</i>	5
			<i>Bipalponephtys cornuta</i>	12
			<i>Nephtys caecoides</i>	9
			<i>Nephtys ferruginea</i>	23
			<i>Nephtys</i> sp SD2	5
		Phyllodocidae	<i>Eteone brigitteae</i>	1
			<i>Eteone leptotes</i>	1
			<i>Eteone pigmentata</i>	1
			<i>Eulalia levicornuta</i> Cmplx	2
			<i>Eumida longicornuta</i>	10
			<i>Sige</i> sp A	6
			<i>Nereiphylla ferruginea</i> Cmplx	1
			<i>Nereiphylla</i> sp 2	1
			<i>Paranaitis polynoides</i>	1
			<i>Phyllodoce groenlandica</i>	1
			<i>Phyllodoce hartmanae</i>	36
			<i>Phyllodoce longipes</i>	17
			<i>Phyllodoce pettiboneae</i>	10
		Oweniidae	<i>Myriochele gracilis</i>	3
			<i>Myriochele olgae</i>	2
			<i>Myriochele striolata</i>	3
			<i>Owenia collaris</i>	55
		Sabellariidae	<i>Neosabellaria cementarium</i>	1
		Sabellidae	<i>Acromegalomma pigmentum</i>	3
			<i>Acromegalomma splendidum</i>	2
			<i>Bispira</i> sp	1
			<i>Dialychone albocincta</i>	11

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Dialychone trilineata</i>	16
			<i>Dialychone veleronis</i>	12
			<i>Euchone arenae</i>	2
			<i>Euchone hancocki</i>	6
			<i>Euchone incolor</i>	14
			<i>Euchone</i> sp A	10
			<i>Euchone</i> sp	1
			<i>Jasmineira</i> sp B	9
			<i>Myxicola</i> sp	2
			<i>Paradialychone bimaculata</i>	2
			<i>Paradialychone harrisae</i>	1
			<i>Potamethus</i> sp A	24
		Longosomatidae	<i>Heterospio catalinensis</i>	1
		Magelonidae	<i>Magelona berkeleyi</i>	2
			<i>Magelona hartmanae</i>	1
			<i>Magelona</i> sp B	2
		Poecilochaetidae	<i>Poecilochaetus johnsoni</i>	3
			<i>Poecilochaetus martini</i>	2
		Spionidae		1
			<i>Carazziella</i> sp A	6
			<i>Dipolydora socialis</i>	10
			<i>Dispia</i> sp SD1	2
			<i>Laonice cirrata</i>	18
			<i>Laonice nuchala</i>	46
			<i>Malacoceros indicus</i>	1
			<i>Microspio pigmentata</i>	42
			<i>Paraprionospio alata</i>	191
			<i>Polydora cirrosa</i>	1
			<i>Polydora</i> sp	2
			<i>Prionospio dubia</i>	123
			<i>Prionospio ehlersi</i>	33
			<i>Prionospio jubata</i>	219
			<i>Prionospio lighti</i>	8
			<i>Prionospio pygmaeus</i>	668
			<i>Pseudopolydora paucibranchiata</i>	1
			<i>Spiophanes anoculata</i>	1
			<i>Spiophanes berkeleyorum</i>	18
			<i>Spiophanes duplex</i>	1037
			<i>Spiophanes fimbriata</i>	3
			<i>Spiophanes kimballi</i>	205
			<i>Spiophanes norrisi</i>	123
			<i>Spiophanes wigleyi</i>	2
		Cirratulidae	<i>Aphelochaeta elongata</i>	1
			<i>Aphelochaeta glandaria</i> Cmplx	46
			<i>Aphelochaeta monilaris</i>	42
			<i>Aphelochaeta phillipsi</i>	5
			<i>Aphelochaeta tigrina</i>	25
			<i>Aphelochaeta</i> sp LA1	1
			<i>Aphelochaeta</i> sp	2

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Caulleriella lajolla</i>	1
			<i>Chaetozone corona</i>	16
			<i>Chaetozone hartmanae</i>	25
			<i>Chaetozone senticosa</i>	1
			<i>Chaetozone</i> sp SD2	6
			<i>Chaetozone</i> sp SD3	1
			<i>Chaetozone</i> sp SD4	1
			<i>Chaetozone</i> sp SD5	10
			<i>Chaetozone</i> sp	1
			<i>Kirkegaardia cryptica</i>	32
			<i>Kirkegaardia serratiseta</i>	1
			<i>Kirkegaardia sibilina</i>	77
			<i>Kirkegaardia tessellata</i>	23
			<i>Kirkegaardia</i> sp SD9	3
		Fauveliopsidae	<i>Fauveliopsis glabra</i>	1
			<i>Fauveliopsis</i> sp SD1	6
			<i>Fauveliopsis</i> sp	2
		Flabelligeridae	<i>Brada pilosa</i>	4
			<i>Brada pluribranchiata</i>	2
			<i>Pherusa neopapillata</i>	2
		Sternaspidae	<i>Sternaspis affinis</i>	55
		Ampharetidae		16
			<i>Amage anops</i>	4
			<i>Amage scutata</i>	4
			<i>Ampharete acutifrons</i>	1
			<i>Ampharete finmarchica</i>	2
			<i>Ampharete labrops</i>	26
			Ampharetidae sp SD1	3
			<i>Amphicteis scaphobranchiata</i>	9
			<i>Anobothrus gracilis</i>	9
			<i>Eclysippe trilobata</i>	90
			<i>Lysippe</i> sp A	4
			<i>Lysippe</i> sp B	19
			<i>Sabellides manriquei</i>	2
			<i>Samytha californiensis</i>	1
			<i>Sosane occidentalis</i>	3
			<i>Melinna heterodonta</i>	2
			<i>Melinna oculata</i>	14
		Pectinariidae	<i>Cistenides granulata</i>	9
			<i>Pectinaria californiensis</i>	64
		Terebellidae	<i>Amaeana occidentalis</i>	4
			<i>Polycirrus californicus</i>	2
			<i>Polycirrus</i> sp A	11
			<i>Polycirrus</i> sp OC1	11
			<i>Polycirrus</i> sp	9
			<i>Lanassa venusta venusta</i>	6
			<i>Phisidia sanctaemariae</i>	69
			<i>Pista brevibranchiata</i>	3
			<i>Pista estevanica</i>	54

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Pista moorei</i>	1
			<i>Pista wui</i>	8
			<i>Proclea</i> sp A	2
			<i>Streblosoma</i> sp B	3
			<i>Streblosoma</i> sp SF1	6
			<i>Streblosoma</i> sp	1
		Trichobranchidae	<i>Terebellides californica</i>	9
		Chaetopteridae		1
			<i>Phyllochaetopterus limicolus</i>	223
			<i>Spiochaetopterus costarum</i> Cmplx	103
		Capitellidae		1
			<i>Capitella teleta</i>	81
			<i>Decamastus gracilis</i>	13
			<i>Mediomastus acutus</i>	1
			<i>Mediomastus</i> sp	520
			<i>Notomastus hemipodus</i>	16
			<i>Notomastus latericeus</i>	7
			<i>Notomastus</i> sp	2
		Cossuridae	<i>Cossura bansei</i>	9
			<i>Cossura candida</i>	11
			<i>Cossura</i> sp A	4
		Maldanidae		50
			Euclymeninae	16
			<i>Axiothella</i> sp	1
			<i>Clymenella complanata</i>	3
			<i>Clymenella</i> sp A	1
			<i>Clymenura gracilis</i>	25
			Euclymeninae sp A	166
			<i>Petaloclymene pacifica</i>	60
			<i>Praxillella pacifica</i>	118
			<i>Maldane sarsi</i>	73
			<i>Metasychis disparidentatus</i>	23
			<i>Petaloproctus neoborealis</i>	1
			<i>Rhodine bitorquata</i>	51
		Opheliidae	<i>Armandia brevis</i>	8
		Orbiniidae	<i>Leitoscoloplos pugettensis</i>	9
			<i>Scoloplos acmeceps</i>	2
			<i>Scoloplos armiger</i> Cmplx	122
		Paraonidae		1
			<i>Aricidea (Acmira) catherinae</i>	33
			<i>Aricidea (Acmira) cerrutii</i>	4
			<i>Aricidea (Acmira) horikoshii</i>	1
			<i>Aricidea (Acmira) lopezi</i>	15
			<i>Aricidea (Acmira) rubra</i>	2
			<i>Aricidea (Acmira) simplex</i>	12
			<i>Aricidea (Aricidea) wassi</i>	4
			<i>Aricidea (Strelzovia) antennata</i>	13
			<i>Aricidea (Strelzovia) hartleyi</i>	2
			<i>Aricidea (Strelzovia) sp A</i>	17

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Cirrophorus branchiatus</i>	4
			<i>Cirrophorus furcatus</i>	1
			<i>Levinsenia gracilis</i>	31
			<i>Levinsenia oculata</i>	1
			<i>Levinsenia kirbyae</i>	6
			<i>Paradoneis</i> sp SD1	1
			<i>Paradoneis</i> sp	1
			<i>Paraonides platybranchia</i>	3
		Scalibregmatidae	<i>Scalibregma californicum</i>	21
		Travisiidae	<i>Travisia brevis</i>	62
Arthropoda	Pycnogonida	Callipallenidae	<i>Anoropallene palpida</i>	23
		Phoxichilidiidae	<i>Anoplodactylus erectus</i>	1
	Ostracoda	Cypridinidae	<i>Vargula tsujii</i>	1
		Cylindroleberididae	<i>Xenoleberis californica</i>	3
		Philomedidae	<i>Euphilomedes carcharodonta</i>	12
			<i>Euphilomedes producta</i>	3
		Sarsiellidae	<i>Eusarsiella thominx</i>	1
	Malacostraca	Nebaliidae	<i>Nebalia daytoni</i>	1
			<i>Nebalia pugettensis</i> Cmplx	2
		Mysidae	<i>Alienacanthomysis macropsis</i>	1
		Caprellidae		3
			<i>Mayerella banksia</i>	2
		Ischyroceridae	<i>Notopoma</i> sp A	12
		Kamakidae	<i>Amphideutopus oculatus</i>	8
		Photidae	<i>Gammaropsis thompsoni</i>	2
			<i>Photis brevipes</i>	2
			<i>Photis californica</i>	1
			<i>Photis lacia</i>	1
			<i>Photis macinerneyi</i>	5
			<i>Photis</i> sp OC1	8
			<i>Photis</i> sp	1
		Aoridae	<i>Aoroides exilis</i>	1
			<i>Aoroides</i> sp A	1
			<i>Aoroides</i> sp	1
		Unciolidae	<i>Rudilemboides stenopropodus</i>	1
			<i>Rudilemboides</i> sp	8
		Corophiidae		8
			<i>Protomedeia articulata</i> Cmplx	2
		Maeridae	<i>Maera jerrica</i>	12
		Megaluropidae	<i>Gibberosus myersi</i>	16
			<i>Megaluropidae</i> sp A	1
		Oedicerotidae	<i>Americhelidium shoemakeri</i>	15
			<i>Americhelidium</i> sp SD4	1
			<i>Bathymedon pumilus</i>	15
			<i>Deflexilodes norvegicus</i>	3
			<i>Hartmanodes hartmanae</i>	3
			<i>Monoculodes latissimanus</i>	1
			<i>Westwoodilla tone</i>	3
		Eusiridae	<i>Rhachotropis</i> sp SD1	1

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
		Liljeborgiidae	<i>Listriella diffusa</i>	1
			<i>Listriella goleta</i>	1
		Pardaliscidae	<i>Nicippe tumida</i>	3
		Ampeliscidae	<i>Ampelisca agassizi</i>	5
			<i>Ampelisca brachycladus</i>	3
			<i>Ampelisca brevisimulata</i>	31
			<i>Ampelisca cf brevisimulata</i>	4
			<i>Ampelisca careyi</i>	60
			<i>Ampelisca cristata cristata</i>	8
			<i>Ampelisca cristata microdentata</i>	13
			<i>Ampelisca hancocki</i>	9
			<i>Ampelisca indentata</i>	3
			<i>Ampelisca pacifica</i>	22
			<i>Ampelisca pugetica</i>	56
			<i>Ampelisca romigi</i>	6
			<i>Ampelisca unsocalae</i>	2
			<i>Ampelisca sp</i>	6
			<i>Byblis millsii</i>	1
		Argissidae	<i>Argissa hamatipes</i>	2
		Urothoidae	<i>Urothoe elegans</i> Cmplx	1
		Phoxocephalidae	<i>Foxiphalus obtusidens</i>	5
			<i>Rhepoxynius bicuspidatus</i>	125
			<i>Rhepoxynius fatigans</i>	1
			<i>Rhepoxynius heterocuspoidatus</i>	3
			<i>Rhepoxynius lucubrans</i>	9
			<i>Rhepoxynius menziesi</i>	24
			<i>Rhepoxynius stenodes</i>	5
			<i>Rhepoxynius variatus</i>	11
			<i>Eyakia robusta</i>	5
			<i>Harpiniopsis fulgens</i>	1
			<i>Heterophoxus ellisi</i>	2
			<i>Heterophoxus oculatus</i>	23
			<i>Heterophoxus sp</i>	3
		Lysianassidae	<i>Aruga oculata</i>	3
		Tryphosidae	<i>Orchomenella decipiens</i>	1
		Gnathiidae	<i>Caecognathia crenulatifrons</i>	10
		Anthuridae	<i>Haliophasma geminata</i>	7
		Idoteidae	<i>Edotia sublittoralis</i>	6
		Munnopsidae	<i>Ilyarachna profunda</i>	3
		Anarthruridae	<i>Siphonolabrum californiensis</i>	1
		Leptocheliidae	<i>Chondrochelia dubia</i> Cmplx	18
		Tanaellidae	<i>Araphura breviararia</i>	12
			<i>Araphura sp</i> SD1	3
			<i>Araphura sp</i>	1
		Tanaopsidae	<i>Tanaopsis cadieni</i>	1
		Bodotriidae	<i>Leptocuma forsmani</i>	6
		Leuconidae	<i>Eudorella pacifica</i>	4
			<i>Leucon declivis</i>	1
		Nannastacidae	<i>Campylaspis rubromaculata</i>	1

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
			<i>Procampylaspis caenosa</i>	3
		Lampropidae	<i>Hemilamprops californicus</i>	23
			<i>Hemilamprops</i> sp A	1
			<i>Mesolamprops bispinosus</i>	5
		Diastylidae	<i>Anchicolurus occidentalis</i>	3
			<i>Diastylis crenellata</i>	6
			<i>Diastylis pellucida</i>	1
			<i>Diastylopsis tenuis</i>	7
			<i>Oxyurostylis pacifica</i>	1
		Crangonidae	<i>Crangon alaskensis</i>	1
		Upogebiidae	<i>Upogebia lepta</i>	2
		Hippidae	<i>Emerita analoga</i>	3
		Cancriidae	<i>Romaleon jordani</i>	2
		Pinnotheridae	<i>Pinnixa franciscana</i>	1
			<i>Pinnixa longipes</i>	4
			<i>Pinnixa occidentalis</i> Cmplx	7
			<i>Pinnixa</i> sp	24
	Hexanauplia	Scalpellidae	<i>Hamatoscalpellum californicum</i>	1
Nematoda				16
Echinodermata	Asteroidea			4
		Astropectinidae	<i>Astropecten californicus</i>	2
	Ophiuroidea			10
		Ophiuridae	<i>Ophiura luetkenii</i>	6
		Amphiuridae		234
			<i>Amphichondrius granulatus</i>	6
			<i>Amphiodia digitata</i>	9
			<i>Amphiodia urtica</i>	596
			<i>Amphiodia</i> sp	150
			<i>Amphioplus strongyloplax</i>	4
			<i>Amphipholis squamata</i>	1
			<i>Amphiura arcystata</i>	8
			<i>Dougaloplus amphacanthus</i>	9
			<i>Dougaloplus</i> sp	1
	Echinoidea			4
			Spatangoida	2
		Schizasteridae	<i>Brisaster latifrons</i>	2
			<i>Brisaster townsendi</i>	1
			<i>Brisaster</i> sp	1
		Brissidae	<i>Brissopsis pacifica</i>	3
	Holothuroidea			1
		Phylloporidae	<i>Thyone benti</i>	1
		Ypsilothuriidae	<i>Ypsilothuria bitentaculata</i>	1
		Synaptidae	<i>Leptosynapta</i> sp	8
		Chiridotidae	<i>Chiridota</i> sp	8
Phoronida				10
		Phoronidae	<i>Phoronis</i> sp SD1	11
			<i>Phoronis</i> sp	35
Brachiopoda	Lingulata	Lingulidae	<i>Glottidia albida</i>	9
Chordata	Enteropneusta			1

Appendix B.26 *continued*

Phylum	Class	Family	Taxon	n
		Spengeliidae	<i>Schizocardium</i> sp	3
		Harrimaniidae	<i>Saccoglossus</i> sp	8
			<i>Stereobalanus</i> sp	29
	Ascidiacea	Molgulidae	<i>Molgula regularis</i>	1

Appendix C

Demersal Fishes and Megabenthic Invertebrates

2020 Raw Data Summaries

Appendix C.1

Taxonomic listing of demersal fish species captured at PLOO trawl stations during 2020. Data are total number of fish (n), biomass (BM, wet weight, kg), minimum (Min), maximum (Max), and mean length (standard length, cm). Taxonomic arrangement follows Eschmeyer and Herald (1998) and Page et al. (2013).

Taxonomic Classification	Common Name	n	BM	Length (cm)			
				Min	Max	Mean	
RAJIFORMES							
Rajidae	<i>Raja inornata</i>	California Skate ^a	3	1.9	24	55	38
ARGENTINIFORMES							
Argentinidae	<i>Argentina sialis</i>	Pacific Argentine	10	0.2	5	8	7
AULOPIFORMES							
Synodontidae	<i>Synodus lucioceps</i>	California Lizardfish	12	1.1	17	34	22
OPHIDIIFORMES							
Ophidiidae	<i>Chilara taylori</i>	Spotted Cusk-eel	8	0.8	11	18	15
BATRACHOIDIFORMES							
Batrachoididae	<i>Porichthys myriaster</i>	Specklefin Midshipman	10	0.1	4	12	8
	<i>Porichthys notatus</i>	Plainfin Midshipman	62	1.5	8	15	11
SCORPAENIFORMES							
Scorpaenidae	<i>Scorpaena guttata</i>	California Scorpionfish	17	2.7	10	21	16
Sebastidae	<i>Sebastes elongatus</i>	Greenstriped Rockfish	3	0.2	9	10	10
	<i>Sebastes helvomaculatus</i>	Rosethorn Rockfish	1	0.1	8	8	8
	<i>Sebastes hopkinsi</i>	Squarespot Rockfish	17	0.6	9	12	11
	<i>Sebastes miniatus</i>	Vermilion Rockfish	3	0.5	15	20	17
	<i>Sebastes saxicola</i>	Stripetail Rockfish	33	0.8	6	10	8
	<i>Sebastes semicinctus</i>	Halfbanded Rockfish	752	16.4	5	18	10
	<i>Sebastes</i> sp	Rockfish Unidentified	8	0.5	3	5	4
	<i>Sebastes</i> sp	Rockfish Unidentified	8	0.5	3	5	4
Hexagrammidae	<i>Zaniolepis frenata</i>	Shortspine Combfish	119	1.6	8	18	11
	<i>Zaniolepis latipinnis</i>	Longspine Combfish	105	1.7	7	15	11
Cottidae	<i>Icelinus quadriseriatus</i>	Yellowchin Sculpin	23	0.3	3	7	6
	<i>Icelinus tenuis</i>	Spotfin Sculpin	1	0.1	3	3	3
Agonidae	<i>Xeneretmus latifrons</i>	Blacktip Poacher	1	0.1	14	14	14
PERCIFORMES							
Sciaenidae	<i>Genyonemus lineatus</i>	White Croaker	4	0.1	16	18	16
Embiotocidae	<i>Zalemnius rosaceus</i>	Pink Seaperch	30	0.9	5	12	9
Bathymasteridae	<i>Rathbunella alleni</i>	Stripefin Ronquil	1	0.1	10	10	10
Zoarcidae	<i>Lycodes pacificus</i>	Blackbelly Eelpout	1	0.1	18	18	18
Uranoscopidae	<i>Kathetostoma averruncus</i>	Smooth Stargazer	1	0.1	8	8	8
PLEURONECIFORMES							
Paralichthyidae	<i>Citharichthys sordidus</i>	Pacific Sanddab	1333	30.9	3	26	10
	<i>Citharichthys xanthostigma</i>	Longfin Sanddab	115	3.6	4	24	13
	<i>Hippoglossina stomata</i>	Bigmouth Sole	18	1.6	11	23	15
	<i>Paralichthys californicus</i>	California Halibut	1	1.8	52	52	52
	<i>Xystreurys liolepis</i>	Fantail Sole	1	1.0	35	35	35
Pleuronectidae	<i>Lyopsetta exilis</i>	Slender Sole	24	0.5	9	17	14
	<i>Microstomus pacificus</i>	Dover Sole	308	8.1	6	19	12
	<i>Parophrys vetulus</i>	English Sole	142	9.0	11	23	16
	<i>Pleuronichthys decurrens</i>	Curlfin Sole	3	0.2	11	16	14
	<i>Pleuronichthys verticalis</i>	Hornyhead Turbot	42	1.4	10	19	13
Cynoglossidae	<i>Symphurus atricaudus</i>	California Tonguefish	26	0.8	9	16	13

^aLength measured as total length, not standard length (see text)

Appendix C.2

Taxonomic listing of demersal fish species captured at SBOO trawl stations during 2020. Data are total number of fish (n), biomass (BM, wet weight, kg), minimum (Min), maximum (Max), and mean length (standard length, cm). Taxonomic arrangement follows Eschmeyer and Herald (1998) and Page et al. (2013).

Taxonomic Classification	Common Name	n	BM	Length (cm)		
				Min	Max	Mean
RAJIFORMES						
Rhinobatidae	<i>Rhinobatos productus</i>	2	0.5	32	41	37
MYLIOBATIFORMES						
Urolophidae	<i>Urobatis halleri</i>	2	0.5	24	33	29
CLUPEIFORMES						
Clupeidae	<i>Sardinops sagax</i>	62	1.0	11	14	12
AULOPIFORMES						
Synodontidae	<i>Synodus lucioceps</i>	158	3.0	9	27	13
BATRACHOIDIFORMES						
Batrachoididae	<i>Porichthys myriaster</i>	14	1.0	6	29	10
	<i>Porichthys notatus</i>	16	0.4	4	6	5
GASTEROSTEIFORMES						
Syngnathidae	<i>Syngnathus exilis</i>	4	0.2	13	18	17
	<i>Syngnathus</i> sp	3	0.1	13	20	16
SCORPAENIFORMES						
Scorpaenidae	<i>Scorpaena guttata</i>	1	0.1	18	18	18
Hexagrammidae	<i>Zaniolepis latipinnis</i>	1	0.1	16	16	16
Cottidae	<i>Chitonotus pugetensis</i>	11	0.6	7	9	8
	<i>Icelinus quadriseriatus</i>	46	0.4	6	7	7
PERCIFORMES						
Sciaenidae	<i>Genyonemus lineatus</i>	58	3.0	12	18	14
	<i>Seriphus politus</i>	3	0.2	14	14	14
Clinidae	<i>Heterostichus rostratus</i>	1	0.1	12	12	12
Stromateidae	<i>Peprilus simillimus</i>	1	0.1	9	9	9
PLEURONECTIFORMES						
Paralichthyidae	<i>Citharichthys sordidus</i>	33	0.6	3	10	5
	<i>Citharichthys stigmaeus</i>	882	5.5	4	12	8
	<i>Citharichthys xanthostigma</i>	291	10.6	4	20	12
	<i>Hippoglossina stomata</i>	4	0.3	17	24	21
	<i>Paralichthys californicus</i>	10	9.2	25	63	35
	<i>Xystreurys liolepis</i>	6	0.7	10	22	18
Pleuronectidae	<i>Parophrys vetulus</i>	11	1.0	16	23	19
	<i>Pleuronichthys ritteri</i>	6	0.7	13	19	17
	<i>Pleuronichthys verticalis</i>	26	1.3	7	18	11
Cynoglossidae	<i>Symphurus atricaudus</i>	80	1.3	5	15	11
	<i>Paralichthys californicus</i>	1	1.8	52	52	52
	<i>Xystreurys liolepis</i>	1	1.0	35	35	35
Pleuronectidae	<i>Lyopsetta exilis</i>	24	0.5	9	17	14
	<i>Microstomus pacificus</i>	308	8.1	6	19	12
	<i>Parophrys vetulus</i>	142	9.0	11	23	16
	<i>Pleuronichthys decurrens</i>	3	0.2	11	16	14
	<i>Pleuronichthys verticalis</i>	42	1.4	10	19	13
Cynoglossidae	<i>Symphurus atricaudus</i>	26	0.8	9	16	13

^aLength measured as total length, not standard length (see text)

Appendix C.3

Total abundance by species and station for demersal fish collected at PLOO trawl stations during 2020.

Species	Winter 2020						Species Abundance by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
Pacific Sanddab	99	207	106	33	91	64	600
Halfbanded Rockfish	7	112	2	5			126
Dover Sole	25	23	17	8	6	4	83
Longfin Sanddab	6	9	28		15	15	73
Shortspine Combfish	14	37	4	2	4	4	65
English Sole	12	22	2		12	4	52
Longspine Combfish	7	7	8		13		35
Yellowchin Sculpin	15	7					22
Stripetail Rockfish	2	6			1	10	19
Hornyhead Turbot	1	1	9		4	4	19
California Scorpionfish		2	2	2	2	6	14
California Tonguefish	9	2					11
Specklefin Midshipman	10						10
Pink Seaperch	2	1	2		1	3	9
Pacific Argentine	9						9
California Lizardfish	1	2	1		1	3	8
Bigmouth Sole	1	4			1		6
White Croaker		4					4
Spotted Cusk-eel	1		1	1			3
Plainfin Midshipman		2				1	3
Squarespot Rockfish		1					1
Smooth Stargazer			1				1
Rockfish Unidentified						1	1
Greenstriped Rockfish		1					1
Fantail Sole		1					1
California Skate		1					1
California Halibut			1				1
Blacktip Poacher		1					1
Survey Total	221	453	184	51	151	119	1179

Appendix C.3 *continued*

Species	Summer 2020						Species Abundance by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
Pacific Sanddab	63	144	126	76	153	171	733
Halfbanded Rockfish		6	47		571	2	626
Dover Sole	27	41	54	35	41	27	225
English Sole	1	1	27	15	35	11	90
Longspine Combfish	4		21	10	23	12	70
Plainfin Midshipman	1	1	4	1	25	27	59
Shortspine Combfish	13	14	10	3	7	7	54
Longfin Sanddab	6	9		4	22	1	42
Slender Sole			2	15	4	3	24
Hornyhead Turbot		1	9	4	6	3	23
Pink Seaperch	4		8	3	6		21
Squarespot Rockfish					16		16
California Tonguefish	2	6	4	1	1	1	15
Stripetail Rockfish	1		8	1		4	14
Bigmouth Sole	1	1	1	1	7	1	12
Rockfish Unidentified		3	1		1	2	7
Spotted Cusk-eel	1	1	1	1	1		5
California Lizardfish				1	3		4
Vermilion Rockfish					3		3
Curlfin Sole			1		2		3
California Scorpionfish			1		1	1	3
Greenstriped Rockfish				2			2
California Skate				1	1		2
Yellowchin Sculpin	1						1
Stripefin Ronquil			1				1
Spotfin Sculpin		1					1
Rosethorn Rockfish			1				1
Pacific Argentine	1						1
Blackbelly Eelpout						1	1
Survey Total	126	229	327	174	929	274	2059
Annual Total	347	682	511	225	1080	393	3238

Appendix C.4

Total abundance by species and station for demersal fish collected at SBOO trawl stations during 2020.

Species	Winter 2020							Species Abundance by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
Speckled Sanddab	43	52	58	37	70	38	42	340
California Lizardfish	1	5	19	5	41	32	34	137
Pacific Sardine						62		62
White Croaker				2		26	30	58
California Tonguefish		5	1	4	9	1	15	35
Longfin Sanddab			18	10	3		3	34
Plainfin Midshipman	1	1			13		1	16
Specklefin Midshipman		1	2	3	5	1	1	13
Hornyhead Turbot			1		8	1	3	13
California Halibut				3			4	7
Roughback Sculpin		3		1				4
Barcheek Pipefish					3	1		4
Spotted Turbot	2		1					3
Queenfish							3	3
Pipefish Unidentified							3	3
Fantail Sole		1					2	3
Shovelnose Guitarfish					1		1	2
Round Stingray	1			1				2
Pacific Pompano							1	1
Giant Kelpfish							1	1
Survey Total	48	68	100	66	153	162	144	741

Appendix C.4 *continued*

Species	Summer 2020							Species Abundance by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
Speckled Sanddab	32	119	7	62	120	159	43	542
Longfin Sanddab		27	5	8	49	88	80	257
Yellowchin Sculpin		4			3	26	13	46
California Tonguefish	2	5	2	3	7	7	19	45
Pacific Sanddab		22	4	2	2	1	2	33
California Lizardfish		3	1	1	1	6	9	21
Hornyhead Turbot	1			2	2		8	13
English Sole				1	4	5	1	11
Roughback Sculpin	2	1			1	3		7
Bigmouth Sole					3	1		4
Spotted Turbot	1			1			1	3
Fantail Sole						1	2	3
California Halibut		1				2		3
Specklefin Midshipman		1						1
Longspine Combfish					1			1
California Scorpionfish					1			1
Survey Total	38	183	19	80	194	299	178	991
Annual Total	86	251	119	146	347	461	322	1732

Appendix C.5

Biomass (kg) by species and station for demersal fish collected at PLOO trawl stations during 2020.

Species	Winter 2020						Species Biomass by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
Pacific Sanddab	0.8	1.2	5.3	0.1	3.0	2.3	12.7
English Sole	0.7	1.2	0.2		0.9	0.2	3.2
Longfin Sanddab	0.2	0.1	0.8		0.4	0.7	2.2
California Scorpionfish		0.4	0.1	0.1	0.1	1.5	2.2
Halfbanded Rockfish	0.1	1.5	0.1	0.1			1.8
California Halibut			1.8				1.8
Shortspine Combfish	0.3	0.3	0.1	0.1	0.1	0.1	1.0
Fantail Sole		1.0					1.0
California Lizardfish	0.1	0.1	0.1		0.1	0.5	0.9
Dover Sole	0.2	0.1	0.2	0.1	0.1	0.1	0.8
Hornyhead Turbot	0.1	0.1	0.1		0.2	0.1	0.6
Pink Seaperch	0.1	0.1	0.1		0.1	0.1	0.5
Stripetail Rockfish	0.1	0.1			0.1	0.1	0.4
Longspine Combfish	0.1	0.1	0.1		0.1		0.4
Spotted Cusk-eel	0.1		0.1	0.1			0.3
Bigmouth Sole	0.1	0.1			0.1		0.3
Yellowchin Sculpin	0.1	0.1					0.2
Plainfin Midshipman		0.1				0.1	0.2
California Tonguefish	0.1	0.1					0.2
White Croaker		0.1					0.1
Squarespot Rockfish		0.1					0.1
Specklefin Midshipman	0.1						0.1
Smooth Stargazer			0.1				0.1
Rockfish Unidentified						0.1	0.1
Pacific Argentine	0.1						0.1
Greenstriped Rockfish		0.1					0.1
California Skate		0.1					0.1
Blacktip Poacher		0.1					0.1
Survey Total	3.4	7.2	9.2	0.6	5.3	5.9	31.6

Appendix C.5 *continued*

Species	Summer 2020						Species Biomass by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
Pacific Sanddab	0.7	1.2	2.3	4.0	5.8	4.2	18.2
Halfbanded Rockfish		0.1	0.7		13.7	0.1	14.6
Dover Sole	1.0	1.2	1.3	0.9	1.8	1.1	7.3
English Sole	0.1	0.1	1.5	1.2	2.3	0.6	5.8
California Skate				1.0	0.8		1.8
Longfin Sanddab	0.1	0.2		0.1	0.9	0.1	1.4
Plainfin Midshipman	0.1	0.1	0.1	0.1	0.5	0.4	1.3
Longspine Combfish	0.1		0.4	0.1	0.5	0.2	1.3
Bigmouth Sole	0.1	0.1	0.1	0.1	0.8	0.1	1.3
Hornyhead Turbot		0.1	0.4	0.1	0.1	0.1	0.8
Shortspine Combfish	0.1	0.1	0.1	0.1	0.1	0.1	0.6
California Tonguefish	0.1	0.1	0.1	0.1	0.1	0.1	0.6
Vermilion Rockfish					0.5		0.5
Squarespot Rockfish					0.5		0.5
Spotted Cusk-eel	0.1	0.1	0.1	0.1	0.1		0.5
Slender Sole			0.1	0.2	0.1	0.1	0.5
California Scorpionfish			0.2		0.1	0.2	0.5
Stripetail Rockfish	0.1		0.1	0.1		0.1	0.4
Rockfish Unidentified		0.1	0.1		0.1	0.1	0.4
Pink Seaperch	0.1		0.1	0.1	0.1		0.4
Curfin Sole			0.1		0.1		0.2
California Lizardfish				0.1	0.1		0.2
Yellowchin Sculpin	0.1						0.1
Stripefin Ronquil			0.1				0.1
Spotfin Sculpin		0.1					0.1
Rosethorn Rockfish			0.1				0.1
Pacific Argentine	0.1						0.1
Greenstriped Rockfish				0.1			0.1
Blackbelly Eelpout						0.1	0.1
Survey Total	2.9	3.6	8.0	8.5	29.1	7.7	59.8
Annual Total	6.3	10.8	17.2	9.1	34.4	13.6	91.4

Appendix C.6

Biomass (kg) by species and station for demersal fish collected at SBOO trawl stations during 2020.

Species	Winter 2020							Species Biomass by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
California Halibut				4.5			2.2	6.7
White Croaker				0.3		1.0	1.7	3.0
Speckled Sanddab	0.3	0.4	0.5	0.4	0.7	0.3	0.4	3.0
California Lizardfish	0.1	0.1	0.2	0.1	0.4	0.4	0.9	2.2
Pacific Sardine						1.0		1.0
Longfin Sanddab			0.1	0.6	0.2		0.1	1.0
Specklefin Midshipman		0.1	0.1	0.1	0.1	0.1	0.4	0.9
Hornyhead Turbot			0.1		0.2	0.1	0.2	0.6
California Tonguefish		0.1	0.1	0.1	0.1	0.1	0.1	0.6
Shovelnose Guitarfish					0.3		0.2	0.5
Round Stingray	0.4			0.1				0.5
Fantail Sole		0.1					0.4	0.5
Spotted Turbot	0.3		0.1					0.4
Plainfin Midshipman	0.1	0.1			0.1		0.1	0.4
Roughback Sculpin		0.1		0.1				0.2
Queenfish							0.2	0.2
Barcheek Pipefish					0.1	0.1		0.2
Pipefish Unidentified							0.1	0.1
Pacific Pompano							0.1	0.1
Giant Kelpfish							0.1	0.1
Survey Total	1.2	1.0	1.2	6.3	2.2	3.1	7.2	22.2

Appendix C.6 *continued*

Species	Summer 2020							Species Biomass by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
Longfin Sanddab		0.6	0.1	0.1	1.8	3.7	3.3	9.6
Speckled Sanddab	0.1	0.7	0.1	0.1	0.4	0.8	0.3	2.5
California Halibut		2.0				0.5		2.5
English Sole				0.1	0.4	0.4	0.1	1.0
California Lizardfish		0.1	0.1	0.1	0.1	0.1	0.3	0.8
Hornyhead Turbot	0.1			0.1	0.1		0.4	0.7
California Tonguefish	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.7
Pacific Sanddab		0.1	0.1	0.1	0.1	0.1	0.1	0.6
Yellowchin Sculpin		0.1			0.1	0.1	0.1	0.4
Roughback Sculpin	0.1	0.1			0.1	0.1		0.4
Spotted Turbot	0.1			0.1			0.1	0.3
Bigmouth Sole					0.2	0.1		0.3
Fantail Sole						0.1	0.1	0.2
Specklefin Midshipman		0.1						0.1
Longspine Combfish					0.1			0.1
California Scorpionfish					0.1			0.1
Survey Total	0.5	3.9	0.5	0.8	3.6	6.1	4.9	20.3
Annual Total	1.7	4.9	1.7	7.1	5.8	9.2	12.1	42.5

Appendix C.7

Summary of demersal fish abnormalities and parasites at trawl stations sampled during 2020. PE=eye parasite; PG=gill parasite; PL=leech; D=skeletal deformity; O=other; T=tumor.

Region	Survey	Station	Species	Size Class	Count	Type	Abnormalities/Parasite
PLOO	winter	SD7	Pacific Sanddab	8	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD8	Pacific Sanddab	6	2	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD8	Fantail Sole	35	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD8	Pacific Sanddab	9	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD8	Pacific Sanddab	8	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD8	Pacific Sanddab	7	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	summer	SD8	Dover Sole	8	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD12	Pacific Sanddab	13	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	winter	SD10	Pacific Sanddab	13	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	summer	SD12	Pacific Sanddab	11	1	PG	<i>Elthusa vulgaris</i>
PLOO	summer	SD13	Pacific Sanddab	6	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	summer	SD14	Pacific Sanddab	9	1	PE	<i>PhrEXOcephalus cininnatus</i>
PLOO	summer	SD14	Pacific Sanddab	16	1	PE	<i>PhrEXOcephalus cininnatus</i>
SBOO	winter	SD17	Speckled Sanddab	9	1	PG	<i>Elthusa vulgaris</i>
SBOO	winter	SD18	California Halibut	63	1	PL	Leech
SBOO	winter	SD21	Speckled Sanddab	7	1	D	Spinal deformity
SBOO	summer	SD19	California Tonguefish	10	1	O	Dent on body, ventral eyeside
SBOO	summer	SD20	Speckled Sanddab	9	1	T	Tumor

Appendix C.8

Taxonomic listing of megabenthic invertebrate taxa captured at PLOO trawl stations during 2020. Data are number of individuals (n). Taxonomic arrangement follows SCAMIT (2018).

Taxonomic Classification				n
SILICEA				
	Demospongiae	Halichondriidae	<i>Halichondria</i> sp	1
CNIDARIA				
	Anthozoa	Virgulariidae	<i>Acanthoptilum</i> sp	1
MOLLUSCA				
	Gastropoda	Calliostomatidae	<i>Calliostoma turbinum</i>	1
		Nassariidae	<i>Tritia insculpta</i>	1
		Muricidae	<i>Austrotrophon catalinensis</i>	1
	Cephalopoda	Octopodidae	<i>Octopus rubescens</i>	1
ANNELIDA				
	Polychaeta	Aphroditidae	<i>Aphrodita refulgida</i>	2
ARTHROPODA				
	Malacostraca	Sicyoniidae	<i>Sicyonia ingentis</i>	1120
		(no family)	Paguroidea	32
		Diogenidae	<i>Paguristes bakeri</i>	13
		Paguridae	<i>Pagurus armatus</i>	1
			<i>Phimochirus californiensis</i>	2
		Munididae	<i>Pleuroncodes planipes</i>	151,616
		Calappidae	<i>Platymera gaudichaudii</i>	452
		Epialtidae	<i>Loxorhynchus crispatus</i>	2
			<i>Loxorhynchus grandis</i>	1
ECHINODERMATA				
	Asteroidea	Luidiidae	<i>Luidia armata</i>	5
			<i>Luidia asthenosoma</i>	1
			<i>Luidia foliolata</i>	24
	Ophiuroidea	Astropectinidae	<i>Astropecten californicus</i>	18
		Ophiuridae	<i>Ophiura luetkenii</i>	3
		Amphiuridae	<i>Amphichondrius granulatus</i>	1
		Ophiopholidae	<i>Ophiopholis bakeri</i>	4
		Ophiotrichidae	<i>Ophiotrix spiculata</i>	4
		Echinoidea	Toxopneustidae	<i>Lytechinus pictus</i>
		Strongylocentrotidae	<i>Strongylocentrotus fragilis</i>	2
	Holothuroidea	Stichopodidae	<i>Apostichopus californicus</i>	10

Appendix C.9

Taxonomic listing of megabenthic invertebrate taxa captured at SBOO trawl stations during 2020. Data are number of individuals (n). Taxonomic arrangement follows SCAMIT (2018).

Taxonomic Classification				n	
SILICEA					
	Demospongiae	Suberitidae	<i>Suberites</i> sp	5	
CNIDARIA					
	Anthozoa	Virgulariidae	<i>Stylatula elongata</i>	3	
MOLLUSCA					
	Gastropoda	Naticidae	<i>Neverita recluziana</i>	1	
			<i>Sinum scopulosum</i>	2	
		Bursidae	<i>Crossata ventricosa</i>	6	
		Buccinidae	<i>Kelletia kelletii</i>	4	
		Muricidae	<i>Pteropurpura festiva</i>	1	
		Pseudomelatomidae	<i>Crassispira semiinflata</i>	1	
		Onchidorididae	<i>Acanthodoris brunnea</i>	2	
			<i>Acanthodoris rhodoceras</i>	2	
			Arminidae	<i>Armina californica</i>	3
			Philineidae	<i>Philine auriformis</i>	131
	Cephalopoda	Octopodidae	<i>Octopus rubescens</i>	1	
ARTHROPODA					
	Malacostraca	Hemisquillidae	<i>Hemisquilla californiensis</i>	2	
			Penaeidae	<i>Farfantepenaeus californiensis</i>	1
			Sicyoniidae	<i>Sicyonia penicillata</i>	43
			Crangonidae	<i>Crangon alba</i>	1
				<i>Crangon nigromaculata</i>	123
			Diogenidae	<i>Paguristes bakeri</i>	1
			Paguridae	<i>Pagurus spilocarpus</i>	1
			Munididae	<i>Pleuroncodes planipes</i>	1
			Dromiidae	<i>Moreiradromia sarraburei</i>	1
			Calappidae	<i>Platymera gaudichaudii</i>	2
			Epialtidae	<i>Loxorhynchus grandis</i>	4
			Inachoididae	<i>Pyromaia tuberculata</i>	6
			Parthenopidae	<i>Latulambrus occidentalis</i>	1
			Portunidae	<i>Portunus xantusii</i>	28
ECHINODERMATA					
		Asteroidea	Luidiidae	<i>Luidia armata</i>	12
			Astropectinidae	<i>Astropecten californicus</i>	145
	Ophiuroidea	Ophiotrichidae	<i>Ophiothrix spiculata</i>	5	
	Echinoidea	Toxopneustidae	<i>Lytechinus pictus</i>	25	
		Dendrasteridae	<i>Dendraster terminalis</i>	13	
		Loveniidae	<i>Lovenia cordiformis</i>	39	

Appendix C.10

Total abundance by species and station for megabenthic invertebrates captured at PLOO trawl stations during 2020.

Species	Winter 2020						Species Abundance by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
<i>Pleuroncodes planipes</i>			23,890	45,712	34,238	35,763	139,603
<i>Lytechinus pictus</i>	6	1026	1	429	7	6	1475
<i>Sicyonia ingentis</i>	3	4	341	86	76	336	846
<i>Platymera gaudichaudii</i>		2	3	23	47	9	84
Paguroidea		32					32
<i>Luidia foliolata</i>	5	4	1	5		2	17
<i>Paguristes bakeri</i>		13					13
<i>Astropecten californicus</i>	4	2	2		1	1	10
<i>Apostichopus californicus</i>	1		2		1	1	5
<i>Ophiura luetkenii</i>	3						3
<i>Ophiothrix spiculata</i>		3					3
<i>Phimochirus californiensis</i>		2					2
<i>Luidia armata</i>	2						2
<i>Strongylocentrotus fragilis</i>					1		1
<i>Octopus rubescens</i>		1					1
<i>Luidia asthenosoma</i>		1					1
<i>Loxorhynchus grandis</i>			1				1
<i>Austrotrophon catalinensis</i>		1					1
<i>Aphrodita refulgida</i>			1				1
<i>Acanthoptilum</i> sp		1					1
Survey Total	24	1092	24,242	46,255	34,371	36,118	142,102

Appendix C.10 *continued*

Species	Summer 2020						Species Abundance by Survey
	SD7	SD8	SD10	SD12	SD13	SD14	
<i>Pleuroncodes planipes</i>		1	3	10,925		1084	12,013
<i>Lytechinus pictus</i>	12	997	832	2	124	77	2044
<i>Platymera gaudichaudii</i>	1			366		1	368
<i>Sicyonia ingentis</i>	1	1		265	6	1	274
<i>Astropecten californicus</i>	3	3	1			1	8
<i>Luidia foliolata</i>	5	2					7
<i>Apostichopus californicus</i>	1	2			2		5
<i>Ophiopholis bakeri</i>			1		3		4
<i>Luidia armata</i>	3						3
<i>Loxorhynchus crispatus</i>		2					2
<i>Tritia insculpta</i>					1		1
<i>Strongylocentrotus fragilis</i>			1				1
<i>Pagurus armatus</i>			1				1
<i>Ophiothrix spiculata</i>	1						1
<i>Halichondria</i> sp					1		1
<i>Calliostoma turbinum</i>	1						1
<i>Aphrodita refulgida</i>		1					1
<i>Amphichondrius granulatus</i>		1					1
Survey Total	28	1010	839	11,558	137	1164	14,736
Annual Total	52	2102	25,081	57,813	34,508	37,282	156,838

Appendix C.11

Total abundance by species and station for megabenthic invertebrates captured at SBOO trawl stations during 2020.

Species	Winter 2020							Species Abundance by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
<i>Crangon nigromaculata</i>	1	19	8	11	34	19	28	120
<i>Sicyonia penicillata</i>		4	7	1	9	7	13	41
<i>Portunus xantusii</i>	1	4	3	2	12	4	2	28
<i>Lovenia cordiformis</i>	23							23
<i>Astropecten californicus</i>	5		1	1	6	2		15
<i>Lytechinus pictus</i>	7		1	5				13
<i>Philine auriformis</i>			4	2				6
<i>Suberites</i> sp	1	2	1					4
<i>Ophiothrix spiculata</i>		2	2					4
<i>Dendraster terminalis</i>	4							4
<i>Loxorhynchus grandis</i>			3					3
<i>Stylatula elongata</i>	1		1					2
<i>Sinum scopulosum</i>	1		1					2
<i>Pyromaia tuberculata</i>		1	1					2
<i>Hemisquilla californiensis</i>				1			1	2
<i>Pagurus spilocarpus</i>					1			1
<i>Paguristes bakeri</i>							1	1
<i>Octopus rubescens</i>					1			1
<i>Moreiradromia sarraburei</i>		1						1
<i>Luidia armata</i>						1		1
<i>Kelletia kelletii</i>					1			1
<i>Farfantepenaeus californiensis</i>							1	1
<i>Crossata ventricosa</i>			1					1
<i>Crangon alba</i>	1							1
Survey Total	45	33	34	23	64	33	46	278

Appendix C.11 *continued*

Species	Summer 2020							Species Abundance by Survey
	SD15	SD16	SD17	SD18	SD19	SD20	SD21	
<i>Astropecten californicus</i>	16	8		1	42	36	27	130
<i>Philine auriformis</i>		31	61	27	6			125
<i>Lovenia cordiformis</i>	13		1	1		1		16
<i>Lytechinus pictus</i>	3		1	8				12
<i>Luidia armata</i>		1	6	2		1	1	11
<i>Dendraster terminalis</i>	9							9
<i>Crossata ventricosa</i>			1	2		1	1	5
<i>Pyromaia tuberculata</i>		2	1			1		4
<i>Kelletia kelletii</i>				3				3
<i>Crangon nigromaculata</i>		2		1				3
<i>Armina californica</i>	1	2						3
<i>Sicyonia penicillata</i>							2	2
<i>Platymera gaudichaudii</i>	1					1		2
<i>Acanthodoris rhodoceras</i>	1	1						2
<i>Acanthodoris brunnea</i>				2				2
<i>Suberites</i> sp		1						1
<i>Stylatula elongata</i>			1					1
<i>Pteropurpura festiva</i>	1							1
<i>Pleuroncodes planipes</i>							1	1
<i>Ophiothrix spiculata</i>		1						1
<i>Neverita recluziana</i>	1							1
<i>Loxorhynchus grandis</i>			1					1
<i>Latulambrus occidentalis</i>			1					1
<i>Crassispira semiinflata</i>			1					1
Survey Total	46	49	75	47	48	41	32	338
Annual Total	91	82	109	70	112	74	78	616

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Appendix D

Contaminants in Marine Fishes

2020 Raw Data Summaries

Appendix D.1

Constituents and method detection limits (MDL) used for the analysis of liver and muscle tissues of fishes collected during 2020; NA= not applicable.

Parameter	MDL		Parameter	MDL	
	Liver	Muscle		Liver	Muscle
Metals (ppm)					
Aluminum (Al)	5.02-5.02	1.67-1.67	Lead (Pb)	0.277-0.277	0.092-0.092
Antimony (Sb)	0.531-0.531	0.177-0.177	Manganese (Mn)	0.526-0.526	0.142-0.142
Arsenic (As)	0.579-0.579	0.193-0.193	Mercury (Hg)	0.002-0.008	0.003-0.003
Barium (Ba)	1.07-1.07	0.357-0.357	Nickel (Ni)	0.109-0.109	0.036-0.036
Beryllium (Be)	0.011-0.011	0.004-0.004	Selenium (Se)	2.35-2.35	0.176-0.176
Cadmium (Cd)	0.053-0.053	0.018-0.018	Silver (Ag)	0.524-0.524	0.175-0.175
Chromium (Cr)	0.449-0.449	0.15-0.15	Tin (Sn)	7.96-7.96	2.65-2.65
Copper (Cu)	0.287-0.287	0.096-0.096	Zinc (Zn)	1.74-1.74	0.581-0.581
Iron (Fe)	2.96-2.96	0.988-0.988			
Chlorinated Pesticides (ppb)					
<i>Hexachlorocyclohexane (HCH)</i>					
HCH, Alpha isomer	2.26-2.68	0.26-0.27	HCH, Delta isomer	4.03-4.93	0.48-0.5
HCH, Beta isomer	5.6-6.11	0.59-0.61	HCH, Gamma isomer	2.26-2.77	0.27-0.28
<i>Total Chlordane</i>					
Alpha (cis) chlordane	3.11-3.45	0.33-0.35	Heptachlor epoxide	2.09-2.56	0.25-0.26
Cis nonachlor	2.68-3.14	0.3-0.32	Methoxychlor	1.88-2.3	0.22-0.23
Gamma (trans) chlordane	2.52-3.08	0.3-0.31	Oxychlordane	2.66-3.26	0.32-0.33
Heptachlor	3.22-3.93	0.38-0.4	Trans nonachlor	1.95-2.39	0.23-0.24
<i>Total Dichlorodiphenyltrichloroethane (DDT)</i>					
o,p-DDD	4.81-5.7	0.55-0.57	p,p-DDD	6.86-7.45	0.73-0.75
o,p-DDE	3.49-3.78	0.37-0.38	p,p-DDE	12.4-14.9	1.47-1.52
o,p-DDT	2.87-3.51	0.34-0.35	p,p-DDT	5.81-6.34	0.62-0.63
p,-p-DDMU	2.76-3.38	0.33-0.33			
<i>Miscellaneous Pesticides</i>					
Aldrin	4.97-6.07	0.59-0.61	Endrin	4.15-5.08	0.49-0.51
AlphaEndosulfan	3.45-4.22	0.41-0.42	Endrin aldehyde	7.57-9.26	0.9-0.93
BetaEndosulfan	7.93-9.7	0.94-0.97	Hexachlorobenzene (HCB)	429-466	46-46.6
Dieldrin	1.14-1.39	0.14-0.14	Mirex	6.59-8.07	0.78-0.81
EndosulfanSulfate	2.39-2.92	0.28-0.29			

Appendix D.1 *continued*

Parameter	MDL		Parameter	MDL	
	Liver	Muscle		Liver	Muscle
Polychlorinated Biphenyls Congeners (PCBs) (ppb)					
PCB 18	5.26-6.43	0.62-0.64	PCB 126	1.76-2.15	0.21-0.22
PCB 28	2.68-3.13	0.3-0.31	PCB 128	4.93-5.94	0.58-0.6
PCB 37	2.12-2.59	0.25-0.26	PCB 138	5.32-6.4	0.64-0.65
PCB 44	2.56-3.13	0.3-0.31	PCB 149	4.31-5.02	0.5-0.51
PCB 49	2.51-2.98	0.29-0.3	PCB 151	6.83-7.53	0.74-0.75
PCB 52	2.52-3.04	0.29-0.3	PCB 153/168	12.6-15.1	1.54-1.54
PCB 66	4.23-5.1	0.49-0.51	PCB 156	2.23-2.69	0.26-0.27
PCB 70	4.58-5	0.48-0.5	PCB 157	2.67-2.96	0.29-0.3
PCB 74	4.59-5.01	0.49-0.5	PCB 158	5.85-6.44	0.62-0.65
PCB 77	2.56-3.13	0.3-0.31	PCB 167	3.87-4.59	0.44-0.46
PCB 81	3.61-4.41	0.43-0.44	PCB 169	2.65-3.25	0.31-0.33
PCB 87	2.06-2.44	0.24-0.24	PCB 170	2.77-3.34	0.33-0.34
PCB 99	4.54-5.55	0.56-0.56	PCB 177	5.03-5.96	0.59-0.6
PCB 101	2.19-2.63	0.27-0.27	PCB 180	5.41-6.5	0.66-0.66
PCB 105	3.72-4.48	0.44-0.45	PCB 183	5.89-6.98	0.69-0.7
PCB 110	4.19-5.06	0.5-0.5	PCB 187	5.98-7.19	NA
PCB 114	4.7-5.75	0.56-0.58	PCB 189	3.66-4.47	0.43-0.45
PCB 118	5.19-6.24	0.64-0.64	PCB 194	6.47-7.67	0.75-0.77
PCB 119	4.19-5.13	0.5-0.51	PCB 201	8.11-9.48	0.92-0.95
PCB 123	5.22-6.18	0.61-0.62	PCB 206	6.16-7.3	0.71-0.73
Polycyclic Aromatic Hydrocarbons (PAHs) (ppb)					
1-methylnaphthalene	227-232	224-231	Benzo[G,H,I]perylene	431-441	425-440
1-methylphenanthrene	295-299	297-301	Benzo[K]fluoranthene	426-436	420-435
2,3,5-trimethylnaphthalene	318-326	314-325	Biphenyl	NA	220-220
2,6-dimethylnaphthalene	244-250	241-249	Chrysene	188-192	185-192
2-methylnaphthalene	189-189	184-190	Dibenzo(A,H)anthracene	376-384	370-383
3,4-benzo(B)fluoranthene	451-461	444-460	Fluoranthene	243-249	240-248
Acenaphthene	200-204	197-203	Fluorene	120-122	118-122
Acenaphthylene	183-187	180-187	Indeno(1,2,3-CD)pyrene	349-356	344-355
Anthracene	212-217	209-216	Naphthalene	NA	NA
Benzo[A]anthracene	232-237	228-236	Perylene	324-332	320-330
Benzo[A]pyrene	358-366	353-365	Phenanthrene	427-437	421-436
Benzo[e]pyrene	437-447	431-446	Pyrene	230-235	226-234

Appendix D.2

Lengths and weights of fishes used for each composite (Comp) tissue sample from PLOO trawl and rig fishing zones during 2020. Data are summarized as number of individuals (n), minimum, maximum, and mean values.

Zone	Comp	Species	n	Length (cm, size class)			Weight (g)		
				Min	Max	Mean	Min	Max	Mean
RF1	1	Vermilion Rockfish	3	22	26	24	282	479	358
RF1	2	Vermilion Rockfish	3	24	27	26	360	629	525
RF1	3	Vermilion Rockfish	3	24	27	25	372	545	438
RF2	1	Starry Rockfish	3	18	26	21	197	595	331
RF2	2	Starry Rockfish	3	22	24	23	224	377	326
RF2	3	Mixed Rockfish ^a	3	21	28	25	242	574	423
TZ1	1	Pacific Sanddab	4	16	24	20	74	252	168
TZ1	2	Pacific Sanddab	6	17	20	19	74	134	105
TZ1	3	Pacific Sanddab	10	14	20	16	33	123	60
TZ2	1	Pacific Sanddab	7	13	21	17	30	155	88
TZ2	2	Pacific Sanddab	8	13	21	18	22	155	84
TZ2	3	Pacific Sanddab	6	16	24	19	51	211	118
TZ3	1	Pacific Sanddab	6	11	20	16	26	177	90
TZ3	2	Pacific Sanddab	7	12	19	16	23	121	69
TZ3	3	Pacific Sanddab	7	14	21	17	41	131	77
TZ4	1	Pacific Sanddab	5	14	20	17	46	163	99
TZ4	2	Pacific Sanddab	7	14	21	16	30	135	62
TZ4	3	Longfin Sanddab	6	13	18	16	32	125	76

^aIncludes Flag, Speckled, and Starry Rockfish

Appendix D.3

Lengths and weights of fishes used for each composite (Comp) tissue sample from SBOO trawl and rig fishing stations during 2020. Data are summarized as number of individuals (n), minimum, maximum, and mean values.

Zone	Comp	Species	n	Length (cm, size class)			Weight (g)		
				Min	Max	Mean	Min	Max	Mean
RF3	1	California Scorpionfish	3	26	27	26	587	705	642
RF3	2	Mixed Rockfish ^a	3	22	26	25	287	585	484
RF3	3	Mixed Rockfish ^b	3	18	27	21	172	458	268
RF4	1	Mixed Rockfish ^c	3	21	26	23	252	390	326
RF4	2	California Scorpionfish	3	25	29	27	567	812	686
RF4	3	California Scorpionfish	3	24	26	25	424	571	521
TZ5	1	Longfin Sanddab	11	12	20	14	26	145	50
TZ5	2	Hornyhead Turbot	6	14	20	17	51	177	130
TZ5	3	Spotted Turbot	12	15	20	16	57	169	94
TZ6	1	Longfin Sanddab	7	12	16	14	26	91	52
TZ6	2	Longfin Sanddab	7	12	16	14	35	94	58
TZ6	3	Longfin Sanddab	11	12	14	13	28	62	40
TZ7	1	Longfin Sanddab	10	12	15	13	29	70	40
TZ7	2	Longfin Sanddab	9	12	15	13	24	67	37
TZ7	3	Longfin Sanddab	10	11	15	13	24	71	37
TZ8	1	Hornyhead Turbot	8	13	19	14	60	178	94
TZ8	2	Longfin Sanddab	9	11	15	12	32	85	47
TZ8	3	California Scorpionfish	3	20	23	22	278	320	299
TZ9	1	Spotted Turbot	5	12	23	17	40	335	137
TZ9	2	No sample	—	—	—	—	—	—	—
TZ9	3	No sample	—	—	—	—	—	—	—

^aIncludes Gopher Rockfish and Treefish; ^bIncludes Brown and Olive Rockfish; ^cIncludes Gopher Rockfish and Treefish

Appendix D.4

Concentrations of metals (ppm) detected in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. See Appendix D.1 for MDLs; ND = not detected; NR = not reportable.

Zone	Comp Species	Trace Metals																	
		Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Tl	Sn	Zn
TZ1	1 Pacific Sanddab	ND	ND	8.44	ND	ND	2.11	ND	4.89	82.7	ND	0.131	ND	ND	ND	ND	ND	7.96	24.4
	2 Pacific Sanddab	ND	ND	6.41	ND	ND	2.61	ND	5.71	98.7	ND	0.842	ND	ND	ND	ND	ND	7.96	26.5
	3 Pacific Sanddab	ND	ND	9.11	ND	ND	2.11	ND	5.54	112	ND	0.749	0.099	ND	2.35	ND	ND	7.96	26.1
TZ2	1 Pacific Sanddab	ND	ND	10.8	ND	ND	3.24	ND	8.19	101	ND	0.806	0.154	0.121	2.35	ND	ND	7.96	32.1
	2 Pacific Sanddab	ND	ND	8.89	ND	ND	3.06	ND	5.79	136	ND	0.926	0.129	ND	2.35	ND	ND	7.96	32
	3 Pacific Sanddab	ND	ND	10.5	ND	ND	4.6	ND	7.43	100	ND	0.923	0.195	ND	2.35	ND	ND	7.96	30.9
TZ3	1 Pacific Sanddab	ND	ND	4.28	ND	ND	3.61	ND	7.08	81.4	ND	0.825	0.149	ND	ND	ND	ND	7.96	26.4
	2 Pacific Sanddab	ND	ND	4.7	ND	ND	2.26	ND	4.32	96.1	ND	0.767	0.097	ND	2.35	ND	ND	7.96	26.9
	3 Pacific Sanddab	ND	ND	6.02	ND	ND	2.48	ND	4.45	118	ND	0.716	0.131	ND	2.35	ND	ND	7.96	35.1
TZ4	1 Pacific Sanddab	ND	ND	4.67	ND	ND	2.83	ND	7.14	94.6	ND	0.657	0.146	ND	2.35	ND	ND	7.96	31.8
	2 Pacific Sanddab	ND	ND	4.39	ND	ND	2.2	ND	4.79	92.5	ND	0.526	0.126	ND	ND	ND	ND	ND	27.3
	3 Longfin Sanddab	ND	ND	11.1	ND	ND	3.36	ND	7.82	135	ND	0.606	0.175	ND	ND	ND	ND	ND	34.6
Detection Rate (%)		0	0	100	0	0	100	0	100	100	0	92	100	8	58	0	0	83	100
TZ5	1 Longfin Sanddab	ND	ND	12.1	ND	ND	2.97	ND	8.42	117	ND	0.843	0.092	ND	ND	ND	ND	ND	25
	2 Hornyhead Turbot	ND	ND	3.93	ND	ND	5.04	ND	6.21	78.9	ND	0.914	0.095	ND	ND	ND	ND	ND	75.5
	3 Spotted Turbot	ND	ND	11.4	ND	ND	4.2	ND	12.5	149	ND	1.49	0.076	ND	ND	ND	ND	ND	57.4
TZ6	1 Longfin Sanddab	ND	ND	7.16	ND	ND	0.766	ND	6.74	84.6	ND	1.18	0.055	ND	ND	ND	ND	ND	27
	2 Longfin Sanddab	ND	ND	5.98	ND	ND	0.837	ND	6.47	84.8	ND	0.88	0.065	ND	ND	ND	ND	ND	21.8
	3 Longfin Sanddab	ND	ND	3.81	ND	ND	0.606	ND	4.83	75.1	ND	0.786	0.031	ND	ND	ND	ND	ND	18.8
TZ7	1 Longfin Sanddab	ND	ND	5.23	ND	ND	0.638	ND	5.96	78.5	ND	0.608	0.049	ND	ND	ND	ND	ND	25.1
	2 Longfin Sanddab	ND	ND	3.44	ND	ND	0.45	ND	3.12	44.4	ND	ND	0.028	ND	ND	ND	ND	ND	13.5
	3 Longfin Sanddab	ND	ND	4.39	ND	ND	0.504	ND	5.22	71	ND	0.576	0.036	ND	ND	ND	ND	ND	20.8
TZ8	1 Hornyhead Turbot	ND	ND	3.4	ND	ND	3.77	ND	7.56	67.6	ND	1.06	0.108	ND	ND	ND	ND	ND	65.1
	2 Longfin Sanddab	ND	ND	5.45	ND	ND	0.732	ND	6.07	84.7	ND	1.19	0.045	ND	ND	ND	ND	ND	24.5
	3 CA Scorpionfish	ND	ND	1.75	ND	ND	4.3	ND	30.1	154	ND	0.67	0.174	ND	ND	ND	ND	ND	141
TZ9	1 Spotted Turbot	ND	ND	21.2	ND	ND	2	ND	19.5	1330.288	1.58	0.093	ND	ND	ND	ND	ND	ND	73.6
	2 No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
	3 No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
Detection Rate (%)		0	0	100	0	0	100	0	100	100	8	92	100	0	0	0	0	0	100

Appendix D.5

Concentrations of pesticides (ppb) and lipids (% weight) detected in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. See Appendix D.1 for MDLs and abbreviations; E = estimated; ND = not detected; NR = not reportable.

		Chlordane									
Zone	Comp	Species	tChlordane	A(c)C	cNon	G(t)C	Hept	HeptEpo	Methoxy	Oxychlor	tNon
TZ1	1	Pacific Sanddab	11.79	2.87 E	1.43 E	ND	ND	ND	ND	ND	7.49
	2	Pacific Sanddab	11.66	2.39 E	1.46 E	ND	ND	ND	ND	ND	7.81
	3	Pacific Sanddab	18.41	2.93 E	2.48 E	ND	ND	ND	ND	ND	13.00
TZ2	1	Pacific Sanddab	10.77	1.91 E	1.73 E	ND	ND	ND	ND	ND	7.13
	2	Pacific Sanddab	8.64	1.29 E	1.51 E	ND	ND	ND	ND	ND	5.84
	3	Pacific Sanddab	13.68	3.18	ND	0.4 E	ND	ND	ND	ND	10.10
TZ3	1	Pacific Sanddab	11.47	1.81 E	2.36 E	ND	ND	ND	ND	ND	7.30
	2	Pacific Sanddab	10.99	1.67 E	2.58 E	ND	ND	ND	ND	ND	6.74
	3	Pacific Sanddab	12.67	2.16 E	2.77 E	ND	ND	ND	ND	ND	7.74
TZ4	1	Pacific Sanddab	8.09	1.92 E	1.4 E	ND	ND	ND	ND	ND	4.77
	2	Pacific Sanddab	10.12	2.15 E	1.9 E	0.47 E	ND	ND	ND	ND	5.60
	3	Longfin Sanddab	8.39	2.27 E	1.83 E	ND	ND	ND	ND	ND	4.29
		Detection Rate (%)	100	100	92	17	0	0	0	0	100
TZ5	1	Longfin Sanddab	7.58	2.04 E	1.42 E	ND	ND	ND	ND	ND	4.12
	2	Hornyhead Turbot	0.47	ND	ND	ND	ND	ND	ND	ND	0.47 E
	3	Spotted Turbot	ND	ND	ND	ND	ND	ND	ND	ND	ND
TZ6	1	Longfin Sanddab	3.68	1.39 E	ND	ND	ND	ND	ND	ND	2.29 E
	2	Longfin Sanddab	4.41	1.51 E	ND	ND	ND	ND	ND	ND	2.90
	3	Longfin Sanddab	3.54	1.34 E	ND	ND	ND	ND	ND	ND	2.20
TZ7	1	Longfin Sanddab	5.09	1.7 E	0.93 E	ND	ND	ND	ND	ND	2.46
	2	Longfin Sanddab	3.64	1.6 E	ND	ND	ND	ND	ND	ND	2.04 E
	3	Longfin Sanddab	4.07	1.62 E	ND	ND	ND	ND	ND	ND	2.45
TZ8	1	Hornyhead Turbot	0.55	ND	ND	ND	ND	ND	ND	ND	0.55 E
	2	Longfin Sanddab	4.04	1.36 E	0.7 E	ND	ND	ND	ND	ND	1.98 E
	3	California Scorpionfish	3.06	ND	ND	ND	ND	ND	ND	ND	3.06
TZ9	1	Spotted Turbot	0.41	ND	ND	ND	ND	ND	ND	ND	0.41 E
	2	No sample	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—
		Detection Rate (%)	92	62	23	0	0	0	0	0	92

Appendix D.5 *continued*

Zone	Comp	Species	DDT										Endosulfan	
			tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT	Alpha	Beta	Sulfate	
TZ1	1	Pacific Sanddab	555.21	ND	1.29 E	0.73 E	15.4	3.91 E	531	2.88 E	ND	ND	ND	
	2	Pacific Sanddab	540.055	ND	1.05 E	0.545 E	12.4	2.59 E	521	2.47 E	ND	ND	ND	
	3	Pacific Sanddab	761.93	ND	1.99 E	1.37 E	15.3	3.27 E	736	4 E	ND	ND	ND	
TZ2	1	Pacific Sanddab	512.82	ND	1.3 E	ND	10.9	2.41 E	495	3.21 E	ND	ND	ND	
	2	Pacific Sanddab	352.56	ND	0.96 E	ND	8.17	1.59 E	340	1.84 E	ND	ND	ND	
	3	Pacific Sanddab	619.24	ND	2.1 E	ND	15.2	6.03 E	592	3.91 E	ND	ND	ND	
TZ3	1	Pacific Sanddab	455.99	ND	2.1 E	ND	10.3	2.71 E	438	2.88 E	ND	ND	ND	
	2	Pacific Sanddab	460.28	ND	2.13 E	ND	10.2	2.9 E	442	3.05 E	ND	ND	ND	
	3	Pacific Sanddab	471.49	ND	1.66 E	ND	14.3	2.7 E	450	2.83 E	ND	ND	ND	
TZ4	1	Pacific Sanddab	329.87	ND	0.87 E	ND	NR	NR	327	2 E	ND	ND	ND	
	2	Pacific Sanddab	355.75	ND	1.54 E	ND	NR	NR	352	2.21 E	ND	ND	ND	
	3	Longfin Sanddab	362.08	0.88 E	6.44	ND	NR	NR	352	2.76 E	ND	ND	ND	
		Detection Rate (%)	100	8	100	25	100	100	100	0	0	0	0	
TZ5	1	Longfin Sanddab	256.77	0.58 E	3.07 E	ND	7.91	3.57 E	239	2.64 E	ND	ND	ND	
	2	Hornyhead Turbot	32.23	ND	0.39 E	ND	1.73 E	0.41 E	29.7	ND	ND	ND	ND	
	3	Spotted Turbot	9.79	ND	ND	ND	ND	ND	9.79 E	ND	ND	ND	ND	
TZ6	1	Longfin Sanddab	200.34	0.63 E	3.45 E	ND	7.9	2.46 E	184	1.9 E	ND	ND	ND	
	2	Longfin Sanddab	228.525	0.655 E	4	ND	8.37	2.68 E	211	1.82 E	ND	ND	ND	
	3	Longfin Sanddab	173.83	0.53 E	2.55 E	ND	6.27	2.02 E	161	1.46 E	ND	ND	ND	
TZ7	1	Longfin Sanddab	156.87	0.55 E	2.85 E	ND	6.28	2.75 E	143	1.44 E	ND	ND	ND	
	2	Longfin Sanddab	133.99	0.52 E	2.29 E	ND	5.22	2.16 E	122	1.8 E	ND	ND	ND	
	3	Longfin Sanddab	175.59	0.6 E	3.25 E	ND	6.92	2.49 E	161	1.33 E	ND	ND	ND	
TZ8	1	Hornyhead Turbot	33.89	ND	0.52 E	ND	2.14 E	0.53 E	30.7	ND	ND	ND	ND	
	2	Longfin Sanddab	216.76	0.49 E	3.01 E	ND	6.59	2 E	203	1.67 E	ND	ND	ND	
	3	California Scorpionfish	153.5	ND	ND	ND	1.55 E	0.95 E	151	ND	ND	ND	ND	
TZ9	1	Spotted Turbot	17.94	ND	ND	ND	0.54 E	ND	17.4	ND	ND	ND	ND	
	2	No sample	—	—	—	—	—	—	—	—	—	—	—	
	3	No sample	—	—	—	—	—	—	—	—	—	—	—	
		Detection Rate (%)	100	62	77	0	92	85	100	62	0	0	0	

Appendix D.5 continued

HCH

Zone	Comp	Species	HCH						Mirex	Lipids					
			tHCH	Alpha	Beta	Delta	Gamma	Aldrin			Dieldrin	Endrin	EndAld	HCB	
TZ1	1	Pacific Sanddab	2.63	0.64 E	1.61 E	ND	0.38 E	ND	ND	ND	ND	NR	NR	ND	33.7
	2	Pacific Sanddab	2.05	0.485 E	1.16 E	ND	0.405 E	ND	ND	ND	ND	NR	NR	ND	31.1
	3	Pacific Sanddab	1.09	ND	1.09 E	ND	ND	ND	ND	ND	ND	NR	NR	1.76 E	28.4
TZ2	1	Pacific Sanddab	1.55	ND	1.09 E	ND	0.46 E	ND	ND	ND	ND	14.1 E	14.1 E	1.19 E	30.9
	2	Pacific Sanddab	0.71	ND	0.71 E	ND	ND	ND	ND	ND	ND	27.2 E	27.2 E	ND	19.5
	3	Pacific Sanddab	1.04	ND	1.04 E	ND	ND	ND	ND	ND	ND	NR	NR	5.8 E	26.5
TZ3	1	Pacific Sanddab	0.83	ND	0.83 E	ND	ND	ND	ND	ND	ND	NR	NR	1.32 E	28.4
	2	Pacific Sanddab	0.9	ND	0.9 E	ND	ND	ND	ND	ND	ND	41.5 E	41.5 E	1.4 E	35.5
	3	Pacific Sanddab	1.32	ND	1.32 E	ND	ND	ND	ND	ND	ND	126 E	126 E	ND	37.4
TZ4	1	Pacific Sanddab	ND	ND	NR	ND	ND	ND	ND	ND	ND	NR	NR	ND	36.1
	2	Pacific Sanddab	ND	ND	NR	ND	ND	ND	ND	ND	ND	NR	NR	ND	38.1
	3	Longfin Sanddab	0.52	ND	NR	ND	0.52 E	ND	ND	ND	ND	NR	NR	ND	35.2
		Detection Rate (%)	83	17	100	0	33	0	0	0	0	100	100	42	100
TZ5	1	Longfin Sanddab	1.49	0.54 E	0.95 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	33
	2	Hornyhead Turbot	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND	5.08
	3	Spotted Turbot	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	2.5
TZ6	1	Longfin Sanddab	1.54	0.53 E	1.01 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	38.7
	2	Longfin Sanddab	1.49	0.51 E	0.98 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	40.4
	3	Longfin Sanddab	1.28	0.38 E	0.9 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	32.8
TZ7	1	Longfin Sanddab	1.65	0.49 E	1.16 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	34.7
	2	Longfin Sanddab	1.52	0.5 E	1.02 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	29.5
	3	Longfin Sanddab	1.61	0.57 E	1.04 E	ND	ND	ND	ND	ND	ND	NR	NR	ND	35.7
TZ8	1	Hornyhead Turbot	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND	6.78
	2	Longfin Sanddab	1.04	ND	1.04 E	ND	ND	ND	ND	ND	ND	NR	NR	2.37 E	35.4
	3	California Scorpionfish	ND	ND	ND	ND	ND	ND	ND	ND	ND	NR	NR	ND	9.02
TZ9	1	Spotted Turbot	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	4.69
	2	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—
		Detection Rate (%)	62	54	62	0	0	0	0	0	0	0	0	8	100

Appendix D.6

Concentrations of PCBs (ppb) detected in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. See Appendix D.1 for MDLs; E = estimated; ND = not detected; NR = not reportable.

		PCB Congener										
Zone	Comp	Species	tPCB	18	28	37	44	49	52	66	70	74
TZ1	1	Pacific Sanddab	566.43	ND	1.7 E	ND	0.58 E	2.52 E	2.81	7.93	1.59 E	1.99 E
	2	Pacific Sanddab	550.245	0.36 E	1.34 E	ND	0.595 E	2.08 E	2.41 E	6.04	1.51 E	1.8 E
	3	Pacific Sanddab	890.49	ND	1.69 E	ND	0.63 E	2.38 E	3.05	8.71	1.85 E	2.76 E
TZ2	1	Pacific Sanddab	660.47	ND	1.02 E	ND	0.54 E	1.8 E	2.07 E	4.98	1.37 E	2.18 E
	2	Pacific Sanddab	630.23	ND	0.7 E	ND	0.43 E	1.31 E	1.77 E	3.12 E	1.22 E	1.3 E
	3	Pacific Sanddab	743.2	0.39 E	2.12 E	ND	1.98 E	7.3	9.93	8.41	3.03 E	2.99 E
TZ3	1	Pacific Sanddab	770.11	0.57 E	1.65 E	ND	1.37 E	5.51	6.96	9.01	2.93 E	3.47 E
	2	Pacific Sanddab	801.09	0.55 E	1.58 E	ND	1.35 E	5.62	6.92	8.92	3.06 E	3.58 E
	3	Pacific Sanddab	703.99	0.37 E	1.48 E	ND	1.13 E	4.5	6.28	6.96	3.21 E	3 E
TZ4	1	Pacific Sanddab	495.45	ND	0.86 E	ND	0.72 E	3.87	4.38	4.36 E	1.68 E	1.66 E
	2	Pacific Sanddab	561.81	0.38 E	1.33 E	ND	1.37 E	4.32	5.94	5.6	3.18 E	2.44 E
	3	Longfin Sanddab	604.9	0.65 E	1.86 E	ND	1.41 E	6.79	9.81	7.77	3.52 E	2.97 E
Detection Rate (%)			100	58	100	100	100	100	100	100	100	100
TZ5	1	Longfin Sanddab	127.12	ND	0.6 E	ND	ND	0.71 E	1.38 E	1.81 E	0.61 E	0.95 E
	2	Hornyhead Turbot	11.48	ND	ND	ND	ND	0.38 E	0.5 E	0.33 E	ND	ND
	3	Spotted Turbot	5.74	ND	ND	ND	ND	ND	ND	ND	ND	ND
TZ6	1	Longfin Sanddab	114.53	ND	0.74 E	ND	ND	0.8 E	1.3 E	1.75 E	0.65 E	0.87 E
	2	Longfin Sanddab	113.155	ND	ND	ND	ND	0.83 E	1.52 E	1.81 E	0.52 E	0.905 E
	3	Longfin Sanddab	93.04	ND	0.58 E	ND	ND	0.83 E	1.15 E	1.6 E	0.55 E	0.76 E
TZ7	1	Longfin Sanddab	245.07	ND	2.1 E	ND	0.51 E	2.1 E	2.98	4.32	1.3 E	1.83 E
	2	Longfin Sanddab	170.75	ND	1.4 E	ND	0.48 E	1.59 E	2.02 E	3.16 E	1.17 E	1.52 E
	3	Longfin Sanddab	200.26	ND	1.78 E	ND	0.52 E	2.05 E	2.89	3.95 E	1.18 E	1.79 E
TZ8	1	Hornyhead Turbot	15.41	ND	ND	ND	ND	0.41 E	0.49 E	0.53 E	ND	ND
	2	Longfin Sanddab	99.97	ND	0.6 E	ND	ND	0.67 E	1.21 E	1.52 E	0.48 E	0.89 E
	3	California Scorpionfish	85.84	ND	ND	ND	ND	0.52 E	0.68 E	1.32 E	0.33 E	0.51 E
TZ9	1	Spotted Turbot	29.87	ND	ND	ND	ND	0.47 E	0.58 E	0.56 E	0.37 E	ND
	2	No sample	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—
Detection Rate (%)			100	0	54	0	23	92	92	92	77	69

Appendix D.6 *continued*

PCB Congener

Zone	Comp	Species	77	81	87	99	101	105	110	114	118	119
TZ1	1	Pacific Sanddab	0.31 E	ND	1.45 E	27.6	9.97	17.8	6.11	1.09 E	52.3	1.62 E
	2	Pacific Sanddab	ND	ND	1.25 E	24.1	8.64	14.6	5.01	0.85 E	41.8	1.39 E
	3	Pacific Sanddab	ND	ND	1.69 E	39.8	11.6	26.5	7.57	1.8 E	77.2	2.06 E
TZ2	1	Pacific Sanddab	ND	ND	1.14 E	28.5	8.18	21.8	4.29 E	1.33 E	64.2	1.02 E
	2	Pacific Sanddab	ND	ND	1.09 E	19	6.65	11.3	4.41 E	0.72 E	34.1	0.87 E
	3	Pacific Sanddab	ND	ND	3.38	38.2	23.5	19.9	13	1.27 E	62.6	2.33 E
TZ3	1	Pacific Sanddab	ND	ND	3.64	41.1	22.5	20.2	16.8	1.36 E	67.2	2.29 E
	2	Pacific Sanddab	ND	ND	3.74	43.4	22.4	22.3	17.5	1.43 E	70.5	2.38 E
	3	Pacific Sanddab	ND	ND	4.06	35.5	18.2	19.6	17.5	1.22 E	66	2.27 E
TZ4	1	Pacific Sanddab	ND	ND	1.8 E	26.2	13.7	12.9	8.02	0.97 E	38.7	2.06 E
	2	Pacific Sanddab	ND	ND	3.88	29.8	17.3	15.2	15.1	1.09 E	48.8	1.97 E
	3	Longfin Sanddab	ND	ND	3.95	34.2	23.8	15.8	19	1.05 E	53.8	1.66 E
Detection Rate (%)			8	0	100	100	100	100	100	100	100	100
TZ5	1	Longfin Sanddab	ND	ND	0.72 E	7.69	4.41	2.74 E	1.91 E	ND	10.9	ND
	2	Hornyhead Turbot	ND	ND	ND	1.27 E	0.99 E	ND	ND	ND	1.32 E	ND
	3	Spotted Turbot	ND	ND	ND	ND	0.6 E	ND	ND	ND	0.79 E	ND
TZ6	1	Longfin Sanddab	ND	ND	0.62 E	7.14	3.84	2.49 E	2.05 E	ND	9.46	ND
	2	Longfin Sanddab	ND	ND	0.61 E	7.83	4.54	2.73 E	1.92 E	ND	9.97	ND
	3	Longfin Sanddab	ND	ND	0.72 E	6.8	3.81	2.24 E	1.44 E	ND	8.97	ND
TZ7	1	Longfin Sanddab	ND	ND	1.23 E	16.2	8.48	5.54	5.22	0.55 E	23.2	0.59 E
	2	Longfin Sanddab	0.33 E	ND	1.04 E	10.2	5.89	3.74 E	3.1 E	0.47 E	15.6	0.55 E
	3	Longfin Sanddab	ND	ND	1.41 E	13.5	8.04	3.97	4.26 E	0.41 E	17.6	0.73 E
TZ8	1	Hornyhead Turbot	ND	ND	ND	1.44 E	1.3 E	ND	0.46 E	ND	1.82 E	ND
	2	Longfin Sanddab	ND	ND	0.62 E	6.59	3.41	2.32 E	1.77 E	ND	8.87	ND
	3	California Scorpionfish	ND	ND	ND	5.54	3.4	2.24 E	1.23 E	ND	8.63	ND
TZ9	1	Spotted Turbot	ND	ND	ND	2.2 E	1.68 E	0.71 E	ND	ND	3.86 E	ND
	2	No sample	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—
Detection Rate (%)			8	0	62	92	100	77	77	23	100	23

Appendix D.6 *continued*

		PCB Congener										
Zone	Comp	Species	123	126	128	138	149	151	153/168	156	157	158
TZ1	1	Pacific Sanddab	6.98	ND	17	79.3	3.43 E	2.08 E	125	9.21	2.54 E	3.79 E
	2	Pacific Sanddab	5.72 E	ND	15.8	73.2	3.5 E	1.84 E	131	8.07	2.28 E	3.43 E
	3	Pacific Sanddab	11.5	ND	29.1	134	5.35	2.99 E	216	15.9	4.22	6
TZ2	1	Pacific Sanddab	6.47	ND	20.1	95.6	2.74 E	1.9 E	143	11.4	3.09	5.05 E
	2	Pacific Sanddab	4.16 E	ND	13.3	72	2.63 E	2.54 E	138	7.47	1.74 E	4.78 E
	3	Pacific Sanddab	7.63	ND	21.1	99.8	9.6	5.97 E	164	10.5	3	3.85 E
TZ3	1	Pacific Sanddab	7.75	ND	20.3	106	11.8	6.8 E	177	10.7	2.81 E	6.07 E
	2	Pacific Sanddab	8.2	ND	21.8	110	11.6	7.26	180	11	3.05	6.37
	3	Pacific Sanddab	7.07	ND	20.4	99.6	8.38	6.31 E	152	10.2	2.56 E	6.28
TZ4	1	Pacific Sanddab	5.07 E	ND	12.7	59.3	4.91 E	3.44 E	117	6.74	1.84 E	3.71 E
	2	Pacific Sanddab	5.51	ND	15.6	76	10.2	6.21 E	118	8.44	2.05 E	5.12 E
	3	Longfin Sanddab	6.03	ND	15.8	78.3	16.3	8.97	120	8.24	2.17 E	5.86 E
Detection Rate (%)			100	0	100	100	100	100	100	100	100	100
TZ5	1	Longfin Sanddab	1.27 E	ND	3.13 E	17.9	NR	NR	31.5	1.37 E	0.43 E	0.99 E
	2	Hornyhead Turbot	ND	ND	ND	2 E	NR	NR	3.77 E	ND	ND	ND
	3	Spotted Turbot	ND	ND	ND	1.59 E	NR	ND	2.76 E	ND	ND	ND
TZ6	1	Longfin Sanddab	1.08 E	ND	2.91 E	15.1	NR	NR	25.9	1.27 E	ND	0.83 E
	2	Longfin Sanddab	1.14 E	ND	3.3 E	17.8	NR	NR	30.6	1.27 E	0.44 E	0.96 E
	3	Longfin Sanddab	1.01 E	ND	2.46 E	15.4	NR	NR	23.2	1.08 E	0.33 E	0.78 E
TZ7	1	Longfin Sanddab	2.45 E	ND	6.54	33.4	NR	3.62 E	54.1	3	0.84 E	2.11 E
	2	Longfin Sanddab	1.85 E	ND	4.46 E	22.9	NR	NR	37	1.81 E	0.63 E	1.34 E
	3	Longfin Sanddab	2.35 E	ND	4.99 E	26.5	NR	3.67 E	43.5	2.04 E	0.71 E	1.57 E
TZ8	1	Hornyhead Turbot	ND	ND	0.45 E	2.7 E	NR	NR	4.73 E	ND	ND	ND
	2	Longfin Sanddab	0.8 E	ND	2.71 E	15.3	NR	NR	26	1.16 E	0.42 E	0.91 E
	3	California Scorpionfish	1.14 E	ND	2.53 E	13.8	NR	NR	23	1.21 E	0.47 E	0.72 E
TZ9	1	Spotted Turbot	0.38 E	ND	0.47 E	5.81 E	NR	NR	9.25 E	0.42 E	ND	ND
	2	No sample	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—
Detection Rate (%)			77	0	85	100	NR	67	100	77	62	69

Appendix D.6 *continued*

PCB Congener

Zone	Comp	Species	167	169	170	177	180	183	187	189	194	201	206
TZ1	1	Pacific Sanddab	6.09	ND	22	6.02	53.6	14.6	36.3	1.64 E	20.7	1.18 E	17.6
	2	Pacific Sanddab	5.52	ND	24.1	6.66	60.7	15.5	38.4	1.59 E	22	1.26 E	15.9
	3	Pacific Sanddab	9.64	ND	37.5	9.01	84.1	21.9	55.6	2.43 E	30.9	1.96 E	23.1
TZ2	1	Pacific Sanddab	6.31	ND	26.8	5.5 E	78.6	17.5	41.9	1.67 E	28.7	1.42 E	18.3
	2	Pacific Sanddab	4.44	ND	34.4	11.1	109	27.1	58.6	1.71 E	32.6	2.57 E	14.1
	3	Pacific Sanddab	7.35	ND	27.1	8.21	65	17.3	50	1.71 E	22.9	1.85 E	16
TZ3	1	Pacific Sanddab	7.13	ND	25.9	9.21	65	17.5	49.5	1.76 E	21.3	2.12 E	14.9
	2	Pacific Sanddab	7.79	ND	27.2	9.56	67.5	18.4	53.4	1.81 E	22.2	2.22 E	16.5
	3	Pacific Sanddab	6.83	ND	23.5	8.41	59.8	16.1	45.7	1.59 E	20.9	2.08 E	15
TZ4	1	Pacific Sanddab	4.53	ND	17.8	5.05 E	46.7	12.9	39.7	1.35 E	17.8	1.33 E	11.7
	2	Pacific Sanddab	5.26	ND	18.7	6.28	49	13.2	35.8	1.17 E	15.3	1.57 E	10.7
	3	Longfin Sanddab	5.49	ND	18.6	7.22	48	13.5	36	1.29 E	14.2	1.56 E	9.33
		Detection Rate (%)	100	0	100	100	100	100	100	100	100	100	100
TZ5	1	Longfin Sanddab	1.07 E	ND	NR	2.61 E	8.93	3.37 E	15.1	ND	2.78 E	0.51 E	1.73 E
	2	Hornyhead Turbot	ND	ND	NR	ND	NR	0.47 E	NR	ND	0.45 E	ND	ND
	3	Spotted Turbot	ND	ND	ND	ND	NR	ND	NR	ND	ND	ND	ND
TZ6	1	Longfin Sanddab	1 E	ND	NR	2.45 E	11.2	2.81 E	12.4	ND	3.77 E	ND	2.1 E
	2	Longfin Sanddab	0.95 E	ND	NR	2.67 E	NR	2.57 E	13.8	ND	2.68 E	ND	1.79 E
	3	Longfin Sanddab	0.82 E	ND	NR	2.16 E	NR	2.13 E	10.7	ND	2.08 E	ND	1.44 E
TZ7	1	Longfin Sanddab	2.35 E	ND	7.3	4.51 E	14.1	4.63 E	21.8	ND	4.44 E	0.64 E	3.09 E
	2	Longfin Sanddab	1.73 E	ND	5.23	3.48 E	10.8	3.62 E	16.8	ND	3.64 E	0.72 E	2.48 E
	3	Longfin Sanddab	1.66 E	ND	5.79	3.76 E	11	3.65 E	17.4	ND	4.14 E	0.67 E	2.78 E
TZ8	1	Hornyhead Turbot	ND	ND	NR	ND	NR	0.62 E	NR	ND	0.46 E	ND	ND
	2	Longfin Sanddab	0.9 E	ND	NR	2.25 E	NR	2.91 E	12.5	ND	3.01 E	0.48 E	1.67 E
	3	California Scorpionfish	0.89 E	ND	NR	1.93 E	NR	2.34 E	8.55	ND	2.78 E	0.31 E	1.77 E
TZ9	1	Spotted Turbot	ND	ND	NR	0.43 E	NR	0.94 E	NR	ND	1.06 E	ND	0.68 E
	2	No sample	—	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—	—
		Detection Rate (%)	69	0	75	77	100	92	100	0	92	46	77

Appendix D.7

Concentrations of PAHs (ppb) detected in liver tissues of fishes collected from PLOO and SBOO trawl zones during 2020. See Appendix D.1 for MDLs; E = estimated; ND = not detected; NR = not reportable.

Zone	Comp	Species	tPAH																							
			1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo(A) anthracene	Benzo(A) pyrene													
TZ1	1	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR					
	2	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR				
	3	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR			
TZ2	1	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR			
	2	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR		
	3	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR		
TZ3	1	Pacific Sanddab	35.8	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND		
	2	Pacific Sanddab	41.2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
	3	Pacific Sanddab	67.2	ND	ND	20.3 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
TZ4	1	Pacific Sanddab	55.6	ND	ND	14.6 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
	2	Pacific Sanddab	132.6	ND	25.2 E	36.2 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
	3	Longfin Sanddab	67.5	ND	ND	17.9 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
		Detection Rate (%)	100	0	17	67	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
TZ5	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	
	2	Hornyhead Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Spotted Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ6	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ7	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ8	1	Hornyhead Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	California Scorpionfish	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ9	1	Spotted Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
		Detection Rate (%)	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR

Appendix D.7 continued

Zone	Comp	Species	Benzo[e]pyrene	Benzo[G,H,I]perylene	Benzo[<i>k</i>]fluoranthene	Biphenyl	Chrysene	Dibenzo[A,H]anthracene	Fluoranthene	Fluorene	Indeno[1,2,3- <i>cd</i>]pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene	
TZ1	1	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ2	1	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Pacific Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ3	1	Pacific Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	35.8 E	ND	ND	ND	ND
	2	Pacific Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	41.2 E	ND	ND	ND	ND
	3	Pacific Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	46.9 E	ND	ND	ND	ND
TZ4	1	Pacific Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	41 E	ND	ND	ND	ND
	2	Pacific Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	71.2 E	ND	ND	ND	ND
	3	Longfin Sanddab	ND	ND	ND	NR	ND	ND	ND	ND	ND	49.6 E	ND	ND	ND	ND
Detection Rate (%)			0	0	0	NR	0	0	0	0	0	100	0	0	0	0
TZ5	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Hornyhead Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Spotted Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ6	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ7	1	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ8	1	Hornyhead Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	Longfin Sanddab	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	3	California Scorpionfish	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
TZ9	1	Spotted Turbot	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
	2	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—
	3	No sample	—	—	—	—	—	—	—	—	—	—	—	—	—	—
Detection Rate (%)			NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR

Appendix D.8

Concentrations of metals (ppm) detected in muscle tissues of fishes collected from PLOO and SBOO rig fishing zones during 2020. See Appendix D.1 for MDLs; ND = not detected.

Zone	Comp Species	Trace Metals																	
		Al	Sb	As	Ba	Be	Cd	Cr	Cu	Fe	Pb	Mn	Hg	Ni	Se	Ag	Tl	Sn	Zn
RF1	1	Vermillion Rockfish	ND	ND	3.53	ND	ND	0.018	ND	0.22	2.67	ND	ND	0.077	ND	ND	ND	ND	3.51
	2	Vermillion Rockfish	ND	ND	3.82	ND	ND	ND	0.283	2.82	ND	ND	0.066	ND	0.817	ND	ND	ND	4.28
	3	Vermillion Rockfish	ND	ND	4	ND	ND	ND	0.446	3.08	ND	ND	0.062	ND	ND	ND	ND	ND	4.05
RF2	1	Starry Rockfish	ND	ND	1.04	ND	ND	ND	0.299	1.79	ND	ND	0.233	ND	ND	ND	ND	ND	3.01
	2	Starry Rockfish	ND	ND	1.1	ND	ND	ND	0.269	1.02	ND	ND	0.132	ND	ND	ND	ND	ND	2.87
	3	Mixed Rockfish	ND	ND	1.28	ND	ND	ND	0.213	4.51	ND	ND	0.122	ND	0.878	ND	ND	ND	3.02
Detection Rate (%)			0	0	100	0	0	17	0	100	100	0	0	100	0	33	0	0	100
RF3	1	California Scorpionfish	ND	ND	1.73	ND	ND	ND	0.241	4.74	ND	ND	0.218	ND	ND	ND	ND	ND	3.9
	2	Mixed Rockfish	ND	ND	1.87	ND	ND	ND	0.203	1.29	ND	ND	0.137	ND	ND	ND	ND	ND	4.42
	3	Mixed Rockfish	2.05	ND	1.14	ND	ND	ND	0.182	1.91	ND	ND	0.059	ND	ND	ND	ND	ND	4.2
RF4	1	Mixed Rockfish	ND	ND	1.29	ND	ND	ND	0.602	1.75	ND	ND	0.132	ND	ND	ND	ND	ND	4.5
	2	California Scorpionfish	ND	ND	2.26	ND	ND	ND	0.602	2.7	ND	ND	0.175	ND	ND	ND	ND	ND	4.59
	3	California Scorpionfish	ND	ND	2.65	ND	ND	ND	0.446	1.87	ND	ND	0.164	ND	ND	ND	ND	ND	4.29
Detection Rate (%)			17	0	100	0	0	0	100	100	100	0	0	100	0	0	0	0	100
OEHHA ^a			—	—	—	—	—	—	—	—	—	—	0.22	—	7.4	—	—	—	—
USFDA Action Limit ^b			—	—	—	—	—	—	—	—	—	—	1.0	—	—	—	—	—	—
Median IS ^b			—	—	1.4	—	—	1.0	1.0	20	—	2.0	—	0.50	—	0.3	—	—	175

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.9

Concentrations of pesticides (ppb) and lipids (% weight) detected in muscle tissues of fishes collected from PLOO and SBOO rig fishing zones during 2020. See Appendix D.1 for MDLs and abbreviations; E = estimated; ND = not detected; na = not available.

		Chlordane									
Zone	Comp	Species	tChlordane	A(c)C	cNon	G(t)C	Hept	HeptEpo	Methoxy	Oxychlor	tNon
	RF1	1	Vermilion Rockfish	ND	ND	ND	ND	ND	ND	ND	ND
	RF1	2	Vermilion Rockfish	0.12	ND	ND	ND	ND	ND	ND	0.12 E
O	RF1	3	Vermilion Rockfish	0.22	0.06 E	0.05 E	ND	ND	ND	ND	0.11 E
O	RF2	1	Starry Rockfish	0.04	ND	ND	ND	ND	ND	ND	0.04 E
al	RF2	2	Starry Rockfish	ND	ND	ND	ND	ND	ND	ND	ND
	RF2	3	Mixed Rockfish	0.22	0.06 E	ND	0.07 E	ND	ND	ND	0.09 E
			Detection Rate (%)	67	33	17	17	0	0	0	67
	RF3	1	California Scorpionfish	0.22	0.04 E	ND	0.04 E	ND	0.04 E	ND	0.1 E
	RF3	2	Mixed Rockfish	0.64	0.12 E	ND	0.3 E	0.05 E	0.17 E	ND	ND
O	RF3	3	Mixed Rockfish	0.13	ND	ND	0.09 E	ND	ND	0.04 E	ND
O	RF4	1	Mixed Rockfish	0.1	ND	ND	0.05 E	ND	ND	ND	0.05 E
ss	RF4	2	California Scorpionfish	0.04	ND	ND	ND	ND	ND	ND	0.04 E
	RF4	3	California Scorpionfish	ND	ND	ND	ND	ND	ND	ND	ND
			Detection Rate (%)	83	33	0	67	17	33	0	17
			OEHHA ^a	5.6	—	—	—	—	—	—	—
			USFDA Action Limit ^b	300	—	—	—	—	—	—	—
			Median IS ^b	100	—	—	—	—	—	—	—

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.9 *continued*

Zone	Comp Species	DDT										Endosulfan		
		tDDT	o,p-DDD	o,p-DDE	o,p-DDT	p,p-DDMU	p,p-DDD	p,p-DDE	p,p-DDT	Alpha	Beta	Sulfate		
O L 1	RF1	4.3	ND	ND	ND	0.15 E	0.04 E	4.07	0.04 E	ND	ND	ND	ND	
	RF1	6.77	ND	0.05 E	ND	0.225 E	ND	6.42	0.075 E	ND	ND	ND	ND	
	RF1	5.58	ND	0.07 E	0.03 E	0.23 E	0.07 E	5.12	0.06 E	ND	ND	ND	ND	
O L 2	RF2	4.63	ND	ND	ND	0.13 E	0.05 E	4.4	0.05 E	ND	ND	ND	ND	
	RF2	1.74	ND	ND	ND	0.05 E	ND	1.69	ND	ND	ND	ND	ND	
	RF2	4.81	ND	0.06 E	ND	0.11 E	0.04 E	4.6	ND	ND	0.05 E	ND	ND	
Detection Rate (%)		100	0	50	17	100	67	100	67	0	17	0	0	
O L 3	RF3	13.43	ND	0.04 E	ND	0.15 E	0.1 E	13.1	0.04 E	ND	ND	ND	ND	
	RF3	1.74	ND	ND	ND	0.04 E	0.1 E	1.51 E	0.09 E	0.18 E	ND	0.06 E	ND	
	RF3	1.57	ND	0.07 E	0.04 E	0.11 E	0.04 E	1.27 E	0.04 E	ND	ND	ND	ND	
O L 4	RF4	2.79	ND	ND	ND	0.04 E	0.05 E	2.7	ND	ND	0.05 E	ND	ND	
	RF4	2.73	ND	ND	ND	0.04 E	0.04 E	2.65	ND	ND	0.05 E	ND	ND	
	RF4	1.44	ND	ND	ND	ND	ND	1.44 E	ND	ND	ND	ND	ND	
Detection Rate (%)		100	0	33	17	83	83	100	50	17	33	17	17	
OEHA ^a		21	—	—	—	—	—	—	—	—	—	—	—	
USFDA Action Limit ^b		5000	—	—	—	—	—	—	—	—	—	—	—	
Median IS ^b		5000	—	—	—	—	—	—	—	—	—	—	—	

^a From the California OEHA (Klasing and Brodberg 2008)

^b From Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.9 *continued*

Zone	Comp	Species	HCH										Mirex	Lipids		
			tHCH	Alpha	Beta	Delta	Gamma	Aldrin	Dieldrin	Endrin	EndAld	HCB				
RF1	1	Vermilion Rockfish	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	0.11 E	ND	0.21
RF1	2	Vermilion Rockfish	0.05	ND	0.05 E	ND	ND	ND	ND	ND	ND	ND	ND	0.17 E	ND	0.35
RF1	3	Vermilion Rockfish	0.04	ND	0.04 E	ND	ND	ND	ND	0.08 E	ND	ND	ND	0.16 E	ND	0.39
RF2	1	Starry Rockfish	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	0.09 E	ND	0.19
RF2	2	Starry Rockfish	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	0.11 E	ND	0.15
RF2	3	Mixed Rockfish	0.14	0.04 E	0.06 E	0.04 E	ND	ND	ND	ND	ND	ND	ND	0.14 E	ND	0.22
Detection Rate (%)			50	17	50	17	0	0	0	17	0	0	0	100	0	100
RF3	1	California Scorpionfish	0.04	ND	0.04 E	ND	ND	ND	ND	ND	ND	ND	ND	0.19 E	ND	0.2
RF3	2	Mixed Rockfish	0.33	0.07 E	0.18 E	ND	0.08 E	ND	0.26	0.67	ND	ND	ND	ND	ND	0.12
RF3	3	Mixed Rockfish	0.12	ND	0.07 E	ND	0.05 E	ND	0.06 E	ND	ND	ND	ND	0.1 E	ND	0.18
RF4	1	Mixed Rockfish	ND	ND	ND	ND	ND	ND	0.04 E	ND	ND	ND	ND	ND	ND	0.15
RF4	2	California Scorpionfish	ND	ND	ND	ND	ND	ND	ND	0.05 E	ND	ND	ND	ND	ND	0.16
RF4	3	California Scorpionfish	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	0.14 E	ND	0.09
Detection Rate (%)			50	17	50	0	33	0	50	33	0	0	0	50	0	100
OEHHA ^a			na	—	—	—	—	—	0.46	na	—	—	na	—	—	—
USFDA Action Limit ^b			na	—	—	—	—	—	300	300	—	—	300	100	100	—
Median IS ^b			na	—	—	—	—	—	na	na	—	—	100	—	—	—

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.10

Concentrations of PCBs (ppb) detected in muscle tissues of fishes collected from PLOO and SBOO rig fishing zones during 2020. See Appendix D.1 for MDLs; E = estimated; ND = not detected; na = not available.

Zone	Comp	Species	PCB Congener											74
			18	28	37	44	49	52	66	70	70			
RF1	1	Vermilion Rockfish	3.94	ND	ND	ND	ND	0.05 E	0.07 E	ND	ND	ND	ND	
RF1	2	Vermilion Rockfish	10.145	ND	ND	ND	ND	0.085 E	ND	ND	ND	ND	ND	
RF1	3	Vermilion Rockfish	4.54	ND	ND	ND	ND	ND	0.09 E	0.04 E	ND	ND	ND	
RF2	1	Starry Rockfish	2.93	ND	0.05 E	ND	0.06 E	ND	0.08 E	ND	ND	ND	ND	
RF2	2	Starry Rockfish	0.81	ND	ND	ND	ND	0.04 E	ND	ND	ND	ND	ND	
RF2	3	Mixed Rockfish	3.03	0.04 E	0.06 E	0.04 E	ND	0.08 E	0.08 E	0.06 E	ND	ND	ND	
Detection Rate (%)			100	17	33	17	17	0	83	50	33	0	0	
RF3	1	California Scorpionfish	6.56	ND	0.05 E	0.03 E	0.05 E	ND	0.09 E	0.12 E	ND	0.07 E	0.07 E	
RF3	2	Mixed Rockfish	1.25	0.04 E	0.06 E	0.04 E	0.05 E	ND	0.06 E	0.06 E	0.04 E	ND	ND	
RF3	3	Mixed Rockfish	1.88	0.05 E	0.07 E	0.07 E	0.07 E	ND	0.08 E	0.08 E	0.07 E	0.07 E	0.07 E	
RF4	1	Mixed Rockfish	0.71	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
RF4	2	California Scorpionfish	0.79	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
RF4	3	California Scorpionfish	0.57	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	
Detection Rate (%)			100	33	50	50	50	0	50	50	33	33	33	
OEHHA ^a			3.6	—	—	—	—	—	—	—	—	—	—	
USFDA Action Limit ^b			na	—	—	—	—	—	—	—	—	—	—	
Median IS ^b			na	—	—	—	—	—	—	—	—	—	—	

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.10 *continued*

		PCB Congener										
Zone	Comp Species	77	81	87	99	101	105	110	114	118	119	
RF1	Vermilion Rockfish	ND	ND	ND	0.21 E	0.18 E	0.1 E	0.1 E	ND	0.34 E	ND	
RF1	Vermilion Rockfish	ND	ND	0.05 E	0.56	0.27	0.45	0.32 E	ND	1.29	ND	
RF1	Vermilion Rockfish	ND	ND	0.06 E	0.24 E	0.23 E	0.13 E	0.14 E	ND	0.42 E	ND	
RF2	Starry Rockfish	ND	ND	0.05 E	0.13 E	0.21 E	0.07 E	0.12 E	ND	0.21 E	ND	
RF2	Starry Rockfish	ND	ND	ND	0.05 E	0.08 E	0.04 E	0.04 E	ND	0.09 E	ND	
RF2	Mixed Rockfish	ND	ND	0.06 E	0.17 E	0.17 E	0.09 E	0.09 E	ND	0.25 E	ND	
	Detection Rate (%)	0	0	67	100	100	100	100	0	100	0	
RF3	California Scorpionfish	ND	ND	0.06 E	0.3 E	0.23 E	0.16 E	0.13 E	ND	0.55 E	ND	
RF3	Mixed Rockfish	ND	ND	ND	0.08 E	0.06 E	0.05 E	0.03 E	ND	0.12 E	ND	
RF3	Mixed Rockfish	0.05 E	0.04 E	0.05 E	0.1 E	0.1 E	0.05 E	0.06 E	ND	0.11 E	0.04 E	
RF4	Mixed Rockfish	ND	ND	ND	0.06 E	0.05 E	ND	ND	ND	0.1 E	ND	
RF4	California Scorpionfish	ND	ND	ND	0.06 E	0.06 E	ND	ND	ND	0.11 E	ND	
RF4	California Scorpionfish	ND	ND	ND	0.04 E	0.04 E	ND	ND	ND	0.08 E	ND	
	Detection Rate (%)	17	17	33	100	100	50	50	0	100	17	
	OEHHA ^a	—	—	—	—	—	—	—	—	—	—	
	USFDA Action Limit ^b	—	—	—	—	—	—	—	—	—	—	
	Median IS ^b	—	—	—	—	—	—	—	—	—	—	

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.10 *continued*

		PCB Congener										
Zone	Comp	Species	123	126	128	138	149	151	153/168	156	157	158
RF1	1	Vermilion Rockfish	0.04 E	ND	0.12 E	0.51 E	0.15 E	0.04 E	0.89 E	0.04 E	ND	ND
RF1	2	Vermilion Rockfish	0.095 E	ND	0.35 E	1.55	0.2 E	0.065 E	2.15	0.2 E	ND	0.145 E
RF1	3	Vermilion Rockfish	0.05 E	ND	0.12 E	0.55 E	0.2 E	0.06 E	0.95 E	0.06 E	ND	ND
RF2	1	Starry Rockfish	ND	ND	0.07 E	0.33 E	0.26 E	0.09 E	0.59 E	ND	ND	ND
RF2	2	Starry Rockfish	ND	ND	ND	0.12 E	0.06 E	ND	0.19 E	ND	ND	ND
RF2	3	Mixed Rockfish	0.04 E	ND	0.08 E	0.34 E	0.14 E	0.05 E	0.58 E	0.03 E	ND	ND
Detection Rate (%)			67	0	83	100	100	83	100	67	0	17
RF3	1	California Scorpionfish	0.08 E	ND	0.14 E	0.72	0.13 E	0.09 E	1.29 E	0.06 E	ND	0.63 E
RF3	2	Mixed Rockfish	ND	ND	ND	0.13 E	0.05 E	ND	0.25 E	ND	ND	ND
RF3	3	Mixed Rockfish	0.04 E	ND	0.05 E	0.13 E	0.07 E	0.04 E	0.24 E	ND	ND	ND
RF4	1	Mixed Rockfish	ND	ND	ND	0.13 E	ND	ND	0.23 E	ND	ND	ND
RF4	2	California Scorpionfish	ND	ND	ND	0.14 E	ND	ND	0.24 E	ND	ND	ND
RF4	3	California Scorpionfish	ND	ND	ND	0.09 E	ND	ND	0.19 E	ND	ND	ND
Detection Rate (%)			33	0	33	100	50	33	100	17	0	17
OEHHA ^a			—	—	—	—	—	—	—	—	—	—
USFDA Action Limit ^b			—	—	—	—	—	—	—	—	—	—
Median IS ^b			—	—	—	—	—	—	—	—	—	—

^aFrom the California OEHHA (Klasing and Brodberg 2008)

^bFrom Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.10 continued

Zone	Comp	Species	PCB Congener												
			167	169	170	177	180	183	187	189	194	201	206		
RF1	1	Vermilion Rockfish	ND	ND	0.14 E	0.06 E	0.31 E	0.1 E	0.29 E	ND	0.1 E	ND	0.1 E	ND	0.1 E
RF1	2	Vermilion Rockfish	0.1 E	ND	0.33 E	0.105 E	0.75 E	0.18 E	0.56 E	ND	0.18 E	ND	0.2 E	ND	0.14 E
RF1	3	Vermilion Rockfish	0.04 E	ND	0.15 E	0.07 E	0.34 E	0.1 E	0.32 E	ND	0.1 E	ND	0.1 E	ND	0.08 E
RF2	1	Starry Rockfish	ND	ND	0.07 E	0.06 E	0.15 E	0.06 E	0.23 E	ND	0.06 E	ND	0.04 E	ND	ND
RF2	2	Starry Rockfish	ND	ND	ND	ND	0.04 E	ND	0.06 E	ND	ND	ND	ND	ND	ND
RF2	3	Mixed Rockfish	ND	ND	0.08 E	0.03 E	0.2 E	0.05 E	0.17 E	ND	0.05 E	ND	0.05 E	ND	ND
		Detection Rate (%)	33	0	83	83	100	83	100	0	83	0	83	0	50
RF3	1	California Scorpionfish	0.05 E	ND	0.19 E	0.12 E	0.44 E	0.12 E	0.45 E	ND	0.12 E	ND	0.12 E	ND	0.09 E
RF3	2	Mixed Rockfish	ND	ND	0.03 E	ND	0.05 E	ND	0.05 E	ND	ND	ND	ND	ND	ND
RF3	3	Mixed Rockfish	ND	ND	ND	ND	0.06 E	ND	0.09 E	ND	ND	ND	ND	ND	ND
RF4	1	Mixed Rockfish	ND	ND	ND	ND	0.08 E	ND	0.06 E	ND	ND	ND	ND	ND	ND
RF4	2	California Scorpionfish	ND	ND	0.03 E	ND	0.08 E	ND	0.07 E	ND	ND	ND	ND	ND	ND
RF4	3	California Scorpionfish	ND	ND	ND	ND	0.06 E	ND	0.07 E	ND	ND	ND	ND	ND	ND
		Detection Rate (%)	17	0	50	17	100	17	100	0	17	0	17	0	17
		OEHHA ^a	—	—	—	—	—	—	—	—	—	—	—	—	—
		USFDA Action Limit ^b	—	—	—	—	—	—	—	—	—	—	—	—	—
		Median IS ^b	—	—	—	—	—	—	—	—	—	—	—	—	—

^a From the California OEHHA (Klasing and Brodberg 2008)

^b From Mearns et al. 1991. USFDA action limits for mercury and all international standards are for shellfish, but are often applied to fish

Appendix D.11

Concentrations of PAHs (ppb) detected in muscle tissues of fishes collected from PLOO and SBOO rig fishing zones during 2020. See Appendix D.1 for MDLs; E = estimated; ND = not detected; NR = not reportable.

Zone	Comp	Species	tPAH														
			1-methyl naphthalene	1-methyl phenanthrene	2-methyl naphthalene	2,3,5-trimethyl naphthalene	2,6-dimethyl naphthalene	3,4-benzo(B) fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benzo(A) anthracene	Benzo(A) pyrene				
PLOO	RF1	Vermilion Rockfish	299	ND	299	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF1	Vermilion Rockfish	27.2	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF1	Vermilion Rockfish	21.6	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
SBOO	RF2	Starry Rockfish	41.4	ND	10.3 E	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF2	Starry Rockfish	21	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF2	Mixed Rockfish	340	ND	153 E	NR	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND
Detection Rate (%)			100	0	50	0	0	0	0	0	0	0	0	0	0	0	0
SBOO	RF3	California Scorpionfish	16.2	ND	16.2 E	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF3	Mixed Rockfish	485.9	ND	81.5 E	NR	ND	52.2 E	ND	ND	ND	ND	20.7 E	ND	ND	ND	ND
	RF3	Mixed Rockfish	36.25	ND	8.85 E	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
SBOO	RF4	Mixed Rockfish	34.2	ND	21.6 E	NR	ND	12.6 E	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF4	California Scorpionfish	ND	ND	ND	NR	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
	RF4	California Scorpionfish	63.2	ND	35.9 E	NR	ND	27.3 E	ND	ND	ND	ND	ND	ND	ND	ND	ND
Detection Rate (%)			83	0	83	0	0	50	0	0	0	0	17	0	0	0	0

Appendix D.11 continued

Zone	Comp	Species	Benzo[e]pyrene	Benzo[G,H,I]perylene	Benzo[K]fluoranthene	Biphenyl	Chrysene	Dibenzo[A,H]anthracene	Fluoranthene	Fluorene	Indeno[1,2,3-cd]pyrene	Naphthalene	Perylene	Phenanthrene	Pyrene
PO	RF1	1	Vermilion Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	NR	ND	ND	ND
	RF1	2	Vermilion Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	27.2 E	ND	ND	ND
	RF1	3	Vermilion Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	21.6 E	ND	ND	ND
P	RF2	1	Starry Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	31.1 E	ND	ND	ND
	RF2	2	Starry Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	21 E	ND	ND	ND
	RF2	3	Mixed Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	187 E	ND	ND	ND
			Detection Rate (%)	0	0	0	0	0	0	0	0	100	0	0	0
SBO	RF3	1	California Scorpionfish	ND	ND	NR	ND	ND	ND	ND	ND	NR	ND	ND	ND
	RF3	2	Mixed Rockfish	ND	ND	227	ND	ND	40 E	ND	ND	NR	ND	42.3 E	22.2 E
	RF3	3	Mixed Rockfish	ND	ND	NR	ND	27.4 E	ND	ND	ND	NR	ND	ND	ND
SBO	RF4	1	Mixed Rockfish	ND	ND	NR	ND	ND	ND	ND	ND	NR	ND	ND	ND
	RF4	2	California Scorpionfish	ND	ND	NR	ND	ND	ND	ND	ND	NR	ND	ND	ND
	RF4	3	California Scorpionfish	ND	ND	NR	ND	ND	ND	ND	ND	NR	ND	ND	ND
			Detection Rate (%)	0	0	100	0	17	0	17	0	NR	0	17	17