

PALYNOLOGY OF EL FOYEL GROUP (PALEOGENE), ÑIRIHUAU BASIN, ARGENTINA

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ABSTRACT – This work aims to analyze the paleovegetation, paleoclimates and paleoceanography of the Paleogene of the Río Foyel section (El Foyel Group), Ñirihuau Basin, Argentina. It comprises Troncoso, Salto del Macho and Río Foyel formations. Sporomorph analysis reflects a regional forest dominated by Nothofagaceae, Myrtaceae, Podocarpaceae and Palmae, developed under a temperate to warm-temperate and humid climate. In the studied section, the relation between terrestrial/marine palynomorphs is considered. In the Troncoso Formation, terrigenous palynomorphs dominate over marine elements. The recurrent presence of gymnosperm pollen with Araucariaceae indicates temperate and humid conditions. The Salto del Macho Formation shows a retraction of the gymnosperm forest, associated with the dominance of *Nothofagus* forest, indicating a temperate and humid condition. In the Río Foyel Formation, the sporomorphs indicate the presence of a temperate to warm-humid forest with subordinate open areas and a remarkable input from the coastal environment. In the same formation, the palynological characteristics, which indicate upward shallowing, allow the identification of parasequences. The bounding surfaces of parasequences are defined as surfaces of flooding which represent a relative sea level rise (marine flooding surfaces). The Río Foyel Formation shows an upward shallowing sequence, indicated by the decrease in abundance and diversity of dinocysts. Different marine flooding surfaces were detected, associated with the diversity and abundance of dinocyst. A comparison of these spore-pollen assemblages with others from Patagonia, using multivariate statistic techniques, yields strong similarities between the Troncoso Formation and the Sloggett Formation (late Eocene to ?early Oligocene); and between Salto del Macho and Río Foyel formations with Loreto Formation (late Eocene-Oligocene) and San Julián Formation in cabo Curioso area (Oligocene).

Key words: Paleogene, Palynology, El Foyel Group, Ñirihuau Basin, Argentina.

RESUMO – Este trabalho visa analisar a paleovegetação, os paleoclimas e a paleoceanografia do Paleógeno da região do Río Foyel (Grupo Foyel), bacia Ñirihuau, Argentina, compreendendo as formações Troncoso, Salto del Macho e Río Foyel. A análise de esporomorfos mostra a existência de um bosque regional formado principalmente por Nothofagaceae, Myrtaceae, Podocarpaceae e Palmae, desenvolvido sob clima temperado a temperado-quente e úmido. Na região estudada, foi considerada a relação entre os palinomorfos continentais/marininhos. Há predominância de palinomorfos terrígenos sobre elementos marinhos na Formação Troncoso. A recorrência de pólen de gimnospermas como Araucariaceae indica clima temperado a úmido. A Formação Salto del Macho mostra uma regressão do bosque de gimnospermas, relacionada ao avanço do bosque de *Nothofagus*, sugerindo clima temperado-úmido. Na Formação Río Foyel, os esporomorfos demonstram a presença de um bosque temperado a quente-úmido, com áreas abertas subordinadas e um notável *input* do ambiente costeiro. Na mesma formação, as características palinológicas, que indicam rasamento ascendente, permitem a identificação de parasequências. Os limites das parasequências são definidos como superfícies de inundação, as quais representam um aumento relativo do nível do mar (superfícies de inundação marinha). A Formação Río Foyel mostra uma sequência de rasamento ascendente, indicada pela diminuição de dinocistos. Foram detectadas diferentes superfícies de inundação marinha, associadas à diversidade e abundância de dinocistos. A comparação entre estas associações esporopolínicas com outras da Patagônia, realizada através de métodos estatísticos multivariados, revela uma grande semelhança entre a Formação Troncoso (Eoceno médio a superior) e a Formação Sloggett (Eoceno superior a ?Oligoceno inferior); e entre as formações Salto del Macho e Río Foyel com associações da Formação Loreto (Eoceno superior-Oligoceno) e a Formação San Julián na área do cabo Curioso (Oligoceno).

Palavras-chave: Paleógeno, Palinologia, Grupo El Foyel, bacia Ñirihuau, Argentina.

INTRODUCTION

This paper focuses on the application of palynological studies to determine relationships between palynomorph concentrations and associations of the biosphere to reconstruct plant associations, oceanic conditions and to interpret climatic trends.

Its goal is to contribute to understand the paleovegetation, paleoclimates and paleoceanographic conditions of El Foyel Group (Paleogene) at the Río Foyel type section, in the Ñirihuau Basin. The Troncoso, Salto del Macho and Río Foyel formations of El Foyel Group are here included.

The major vegetation types may be recognized from the pollen assemblage. Many Tertiary pollen types show affinities with living taxa, hence it should be possible to use the ecological tolerance of today to reconstruct Tertiary climates; though not always satisfactorily due to the fact that ecological tolerances may have changed over the time as suggested by Martin (1987, 1997). Among the most important proxies that were used to detect paleoenvironmental changes are the organic-walled dinoflagellate cysts (dinocysts). Dinocyst assemblages in sediments provide information pertinent to the ecology of the water in which the dinoflagellates lived.

PREVIOUS INVESTIGATIONS ON PALEONTOLOGY

The Río Foyel Formation contains an abundant fauna of marine invertebrates, which have been studied since early 20th century (von Ihering, 1904, 1907, 1914; Gonzalez Bonorino, 1944). In samples from the same section studied here, but from the upper levels, Bertels (1980, 1993, 1994a,b) found strong similarities between the assemblages of foraminifera and those from the Monte León Formation (Santa Cruz Province, Atlantic coast). Based in this bio-correlation, Bertels (1980, 1993, 1994a, b) postulated an Oligocene age for the Río Foyel Formation. Recently, Malumián *et al.* (2008) found moulds of *Transversigerina* cf. *T. transversa* (Cushman) at 18 m from the base of the section under study. This genus appeared in the late Oligocene and the compared species in the early Miocene. Chiesa & Camacho (2001), based on the megafauna, proposed a middle Eocene age for the lower part of this unit.

The megaflora collected by Washburne between 1911 and 1912 in the Ñirihuau Formation was the basis of Berry's (1928) studies about South American flora. Romero & Dibbern (1984) provide a detailed report on the megaflora in the studied area.

Previous palynological studies correspond to the Huitrera, Salto del Macho and Río Foyel formations (Meléndez *et al.*, 2003; Barreda *et al.*, 2003; Martínez *et al.*, 2008) and in the "Serie Andesítica" (Sepúlveda, 1980). The Huitrera Formation (Meléndez *et al.*, 2003) presents microfloras without *Nothofagidites* of the early Eocene and microfloras with *Nothofagidites* ("brassi" and "fusca" groups) of the middle to late Eocene. Based on the fungal assemblage, Sepúlveda (1980) associated the "Serie Andesítica", in the Cordón

Oriental del Futalaufquen, to the middle Eocene. Other samples from the upper levels of this same section (not studied here) (Barreda *et al.*, 2003) recognize a similar palynological assemblages, but less diverse and they postulated a late Oligocene to early Miocene age taking into account the presence of *Tuberculodinium vancampoae* (Rossignol) Wall. However, this species has already been accurately recognized as corresponding to the early Oligocene (see Head *et al.*, 1989, p. 440). Martínez *et al.* (2008) provide the sedimentology, environmental conditions and an introduction of the plant communities of the Salto del Macho Formation.

STRATIGRAPHY

The fill of the Tertiary Ñirihuau Basin, which is located on the eastern Subandean flank of the Northern Patagonian Cordillera, is characterized by two major volcanic and sedimentary cycles. The Base Cycle (late Paleocene to early Oligocene) involves the Pilcaniyeu Volcanic Belt (Huitrera Formation) and the Cerro Bastión, Río Foyel ("Patagonian") and Rincón de Cholila formations. The Top Cycle (early Oligocene to Pliocene) is represented by the Nahuel Huapi Group and consists, from base to top, of the El Maitén Volcanic Belt (Ventana Formation) and the Ñorquinco, Ñirihuau and Collón Cura formations (Asensio *et al.*, 2010). Structurally, the western part of the basin has taken part in the deformation process that gave rise to the Ñirihuau fold and thrust belt, whereas the eastern border accounts for the basin platform showing poor or no-deformation, with three distinct regions: North (Ñirihuau), South (Ñorquinco) and West (El Bolsón).

El Foyel Group (Asensio *et al.*, 2005) is stratigraphically located between the vulcanite of the Cerro Bastión Formation (middle Eocene, Sepúlveda, 1980) and the vulcanite of the Ventana Formation (late Oligocene, Cazau *et al.*, 1989). It reaches a thickness of more than 1900 m. It appears as if El Foyel Group has a chronostratigraphic range extending from the middle Eocene to the early Oligocene. This entity presents a variable vertical development, as evidenced by the margins of the Foyel River, where the succession reaches a thickness of 2000 m (Asensio *et al.*, 2005).

Río Foyel Section

The outcropping section of the Foyel river margins (42° S) (Figure 1) is central to the stratigraphic analysis of the Ñirihuau Basin. Although it has been known since the 19th century (Roth, 1899; Wherli, 1899), only in 2005 it was substantially studied and valued as having important implications within the sedimentary register of the basin. From the bottom up, it comprises Troncoso, Salto del Macho and Río Foyel formations (Figure 2).

The Troncoso Formation (Asensio *et al.*, 2005) reaches a thickness of 750 m and is constituted mainly of volcaniclastic sandstones and tobaceous clay rocks with intercalation of thick ignimbritic banks. A particularity of this unity is that it presents a divergence between its constitutive layers, which

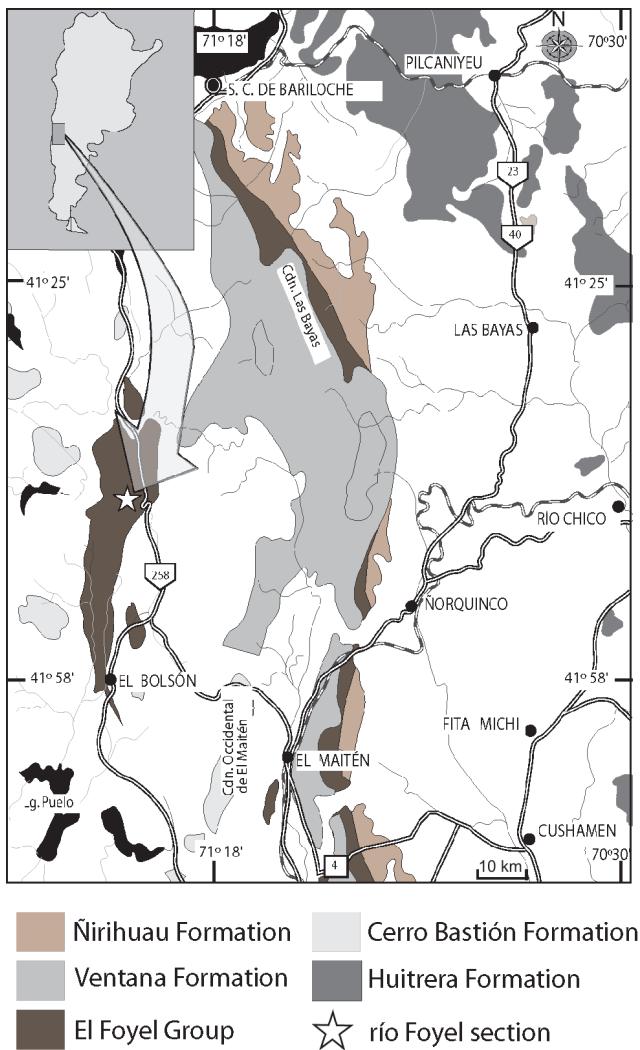


Figure 1. Location map of the study area with indication of the Río Foyel section.

can be inferred as resulting from synsedimentary tectonic activity. The contact with the Cerro Bastión Formation is sharp, although in most cases it is blurred by heavily forested areas. The age of this unit has not been established with precision but, as inferred from its stratigraphic relations, it is estimated to extend from the middle Eocene to late Eocene (Asensio *et al.*, 2005).

The Salto del Macho Formation reaches 560 m of thickness and is characterized by having tabular bodies-less than 10 m thick- with a predominance of the conglomeratic facies, among which there is intercalation of sandstone banks and, in lower proportion, carbonaceous shale. As evidenced by the comparison of facies and the stratigraphic relations within the section, it is possible to emphasize that the origin of this dense succession, mainly conglomeratic, is closely related to subaerial hyperconcentrated flows, associated to distal deltaic fans. The tectosedimentary development of the unit is linked with a strong extensional pulse occurred during middle Eocene-lower Oligocene and is related with the tectonics of the Farallón and the Sudamericana Plates (Asensio *et al.*, 2008). The age of this unit as inferred from

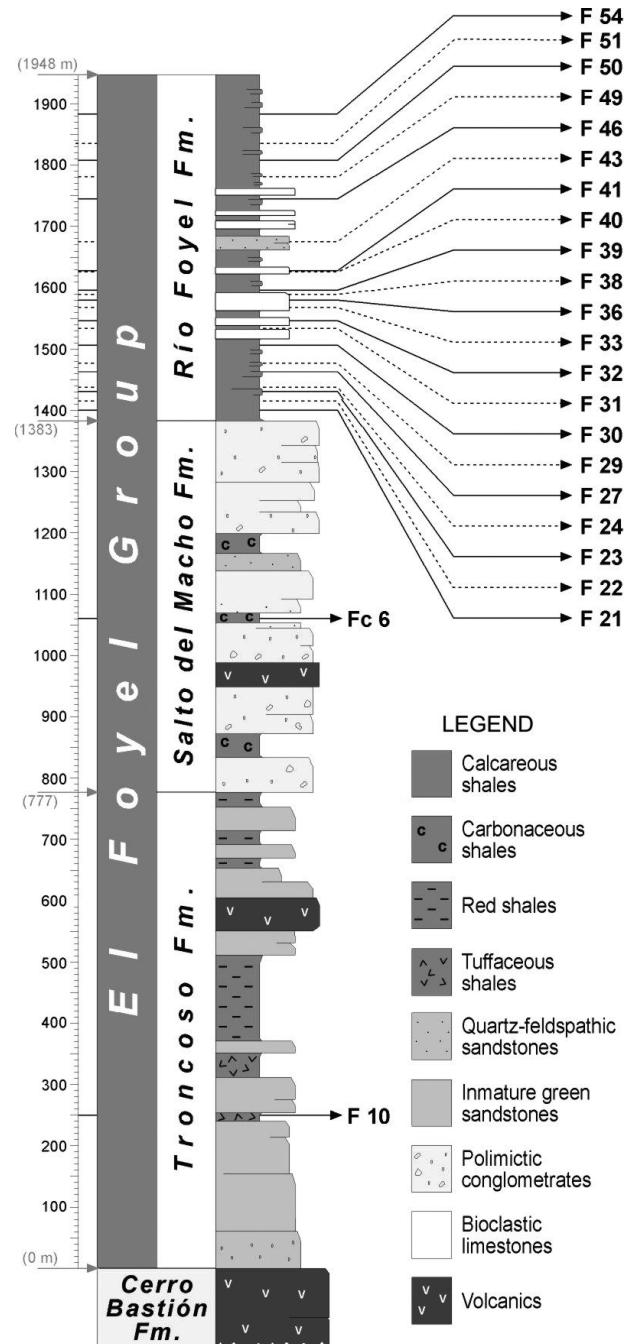


Figure 2. Study section and stratigraphic distribution of the palynological samples.

its stratigraphic relations extends from the middle Eocene to early Oligocene (Asensio *et al.*, 2005).

The Río Foyel Formation (Pöthe de Baldis, 1984) presents a minimum thickness higher than 600 m and is constituted mainly by black calcareous shales. The age is determined by a sampling $^{87}\text{Sr}/^{86}\text{Sr}$ of 30, 65 Ma (Griffin *in* Asensio *et al.*, 2010) on the valve of an *Ostrea* and on an andesite dyke that intrudes to the unit and which yielded the value of 31 Ma (K/Ar) (Giacosa *et al.*, 2001). It stands, with a maximum flooding surface, on the conglomerates of the Salto del Macho Formation and could be related to the sandstones of the Norquino Formation (Cazau, 1972).

MATERIAL AND METHODS

Physical and chemical extraction was performed using standard palynological processing techniques (Volkheimer & Melendi, 1976). Twenty-three samples were processed for palynological purpose, and sixteen yielded moderately to poorly preserved palynomorphs. Six from Troncoso Formation only one fertile, one from Salto del Macho Formation and the others from Río Foyel Formation. All figured specimens are lodged in the collections of the Palynological Laboratory (UNSP), Universidad Nacional del Sur, Bahía Blanca, Argentina. In the citation of specimens referred to or illustrated, the sample and slide number (e.g. 1980a) are given first and the position on the slide is indicated by an England Finder reference (e.g. G36/4). Additionally, the formations are given: Salto del Macho (SM), Troncoso (T), Río Foyel (RF).

The palynologic analysis for biostratigraphic (palynostratigraphic), paleoclimatical and paleoecological

approach is based on qualitative and quantitative data. The quantitative analysis comprises a minimum of 100 palynomorphs per sample, excepting those samples in which this was not possible due to the scarcity of palynomorphs. These are indicated in Table 1 as “presence”. The sporomorphs are arranged in Table 2 according to their botanical affinity.

Hammer's *et al.* (2001) program, *Palaeontological Statistics* (PAST), was used to determine similarities among El Foyel Group and other Danian-Miocene sporomorph assemblages from different units of southern South America, based on presence/absence data.

Each site in the cluster represents a sum of several samples with the exception of Troncoso, Salto del Macho (El Foyel Group) and San Julián formations in cabo Curioso (Santa Cruz Province). Using PAST, the Jaccard Coefficient was applied as a measure of similarity (between 0 to 1 - low to high similarity) between samples. The results of the cluster analyses are presented in dendograms (Figure 3).

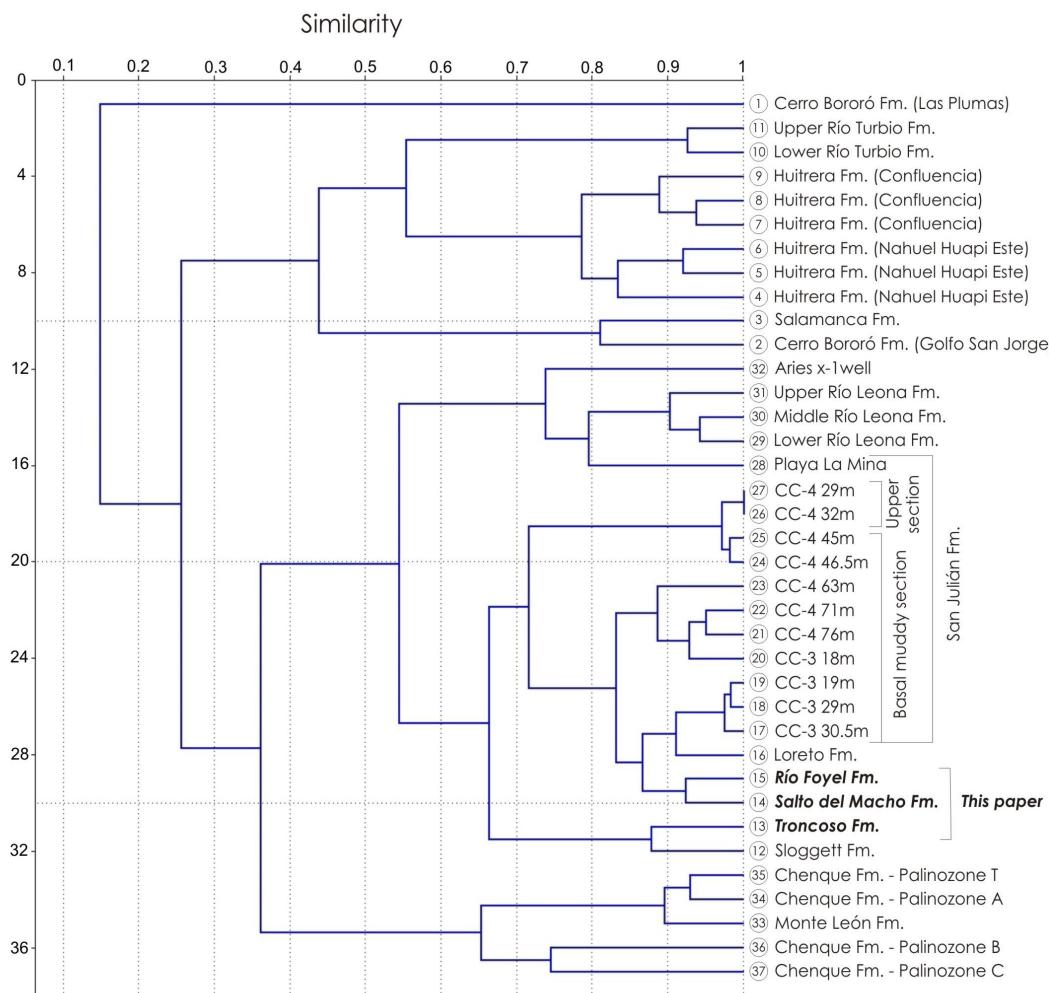


Figure 3. Dendrogram showing the clustering of palynological assemblages from different formations and/or localities, based on Jaccard coefficient and the PAST programme. References: 1, Volkheimer *et al.* (2007); 2-3, Archangelsky (1973, 1976); Archangelsky & Romero (1974); Archangelsky & Zamaloa (1986); 4-9, Melendi *et al.* (2003); 10-11, following in this paper the Lower and Upper Section (Río Turbio Formation) recognized by Hünicken (1955); Archangelsky (1972); Romero & Castro (1986); Romero & Zamaloa, 1985; Romero, 1977; 12, Olivero *et al.*, 1998; 13-15, Asensio *et al.*, 2005; this paper; 16, Fasola (1969); 17-27, Nañez *et al.* (2009); 28, Barreda (1997); 29-31, Barreda *et al.* (2009); 32, Palamarczuk & Barreda (2000); 33, Barreda & Palamarczuk (2000a); 34-37, Barreda (1996); Palamarczuk & Barreda (1998); Barreda & Palamarczuk (2000b) (modified from Nañez *et al.*, 2009).

Table 1. Distribution charts of palynomorphs from the río Foyel section.

Taxa	Formation / Samples	Río Foyel														
		2223 Tuncoso Marelo Sp. sp.	2297 Tuncoso Marelo Sp. sp.	2299 Tuncoso Marelo Sp. sp.	2226 Tuncoso Marelo Sp. sp.	2300 Tuncoso Marelo Sp. sp.	2227 Tuncoso Marelo Sp. sp.	2228 Tuncoso Marelo Sp. sp.	2229 Tuncoso Marelo Sp. sp.	2230 Tuncoso Marelo Sp. sp.	2231 Tuncoso Marelo Sp. sp.	2232 Tuncoso Marelo Sp. sp.	2233 Tuncoso Marelo Sp. sp.	2310 Tuncoso Marelo Sp. sp.	2234 Tuncoso Marelo Sp. sp.	2311 Tuncoso Marelo Sp. sp.
PTERIDOPHYTA & BRYOPHYTA	<i>Azolla</i> sp.	X														
	<i>Baculatisporites comaumensis</i>															
	<i>Baculatisporites turbioensis</i>	X														
	<i>Baculatisporites</i> sp.															
	<i>Biretisporites crassilabrus</i>															
	<i>Biretisporites</i> sp. 1	3.3														
	<i>Cingutriletes australis</i>	1.8														
	<i>Concavisporites</i> sp.															
	<i>Cyatheacidites annulatus</i>	4.41	1.7		X		10.6	8.5		2.3	2.3	X	3.3		1.3	1.2
	<i>Cyathidites subtilis</i>														11	1.4
	<i>Deltoidospora australis</i>	9.73	10	X	X		9.4	19.6	X	20.9	11.6	X	8.8	57.3	15.9	12.3
	<i>Deltoidospora minor</i>		5	X			4.7		X				6.6	13.3		1.4
	<i>Deltoidospora</i> spp.						X									2.7
	<i>Dictyophyllidites harrisii</i>	0.9														
	<i>Gleicheniidites argentinus</i>		1.7													2.7
	<i>Hymenophylleaceae</i>	0.9	1.7												4	1.2
	cf. <i>Hymenophyllum caudiculatum</i>															5.5
	<i>Ischyosporites areapunctatis</i>	3.3		X			7	4.9	X	4.6	2.3		6.6	8	9.8	6.8
	<i>Laevigatosporites ovatus</i>	3.3			X		4.9						2.2			
	<i>Leptolepidites macroverrucosus</i>															1.4
	<i>Matonisporites ornamentalis</i>															
GIMNOSPERMAE	<i>Muricungulispores chenquensis</i>															
	<i>Peromonolites vellosus</i>														X	
	<i>Polypodiisporites inangahuensis</i>	0.9	1.7		X		3.5	1.2							2.7	3.2
	<i>Reticuloidosporites tenellis</i>		1.7													
	Indeterminate trilete spore						2.4									3.2
	<i>Araucariacites australis</i>	33.6			X	X		6.1	X	2.3	2.3					1.6
	? <i>Dacrycarpites australis</i>		1.8					1.2								1.4
	<i>Inaperturopollenites</i> spp.															
	<i>Microcachryidites antarcticus</i>		1.7													1.2
	<i>Phyllocladidites mawsonii</i>	1.7			X		11.8	11					3.3	1.3	6.1	
Algae	<i>Podocarpidites marwickii</i>	10.6	6.7	X		X	5.9	4.9	X	4.6	4.6		2.2	11	13.7	1.6
	<i>Podocarpidites</i> spp.	0.9					3.7									13.7
Others	<i>Taxodiaceapollenites hiatus</i>					X	2.4			2.3	14	X			4.8	
	<i>Trisaccites microsaccatum</i>					X										
ANGIOSPERMAE	<i>Botryococcus</i> sp.	11.5						1.2								1.2
	? <i>Celyphus rillus</i>															
	Foraminiferal test linings							1.2						1.3		1.4
	? <i>Partitomorphitae</i>													4.9		
	<i>Baumannipollis</i> sp.												X			
	<i>Chenopodipollis chenopodiaceoides</i>	1.7														
	<i>Corsinipollenites atlantica</i>	1.7														
	<i>Graminidites</i> sp.	1.8		X			2.4	4.9		2.3			3.3			1.4
	<i>Haloragacidites</i> sp.								X							
	<i>Myrtaceidites verrucosus</i>	0.9	1.7		X	X	3.5	4.9	X	2.3	2.3				2.4	4.1
	Myrtaceidites sp. (cf. <i>Myrceugenia</i>)						1.2							1.3		1.4
	<i>Momipites</i> sp.							X								
	<i>Nothofagidites americanus</i>															
	<i>Nothofagidites amisoechinatus</i>															
	<i>Nothofagidites dorotensis</i>															1.3
	<i>Nothofagidites flemingii</i>															
	<i>Nothofagidites menziesii</i> type	1.7													2.2	1.4
	<i>Nothofagidites nanus</i>															
	<i>Nothofagidites rocaensis</i>	11.7		X	X	X	8.2	2.4		7	14	X	4.4	4	11	
	<i>Nothofagidites saraensis</i>	5		X		X	3.5	2.4					3.3		3.7	4.8
	<i>Nothofagidites</i> spp.	11.7					2.4		X	4.7					2.7	11.1
	<i>Propylipollis rythnthus</i>															
	<i>Proteacidites subscabratus</i>	0.9						X								
	<i>Proteacidites symphyonemoides</i>							X								
	<i>Proteacidites</i> spp.							1.2								
	<i>Pseudowinterapollis couperi</i>							X								
	<i>Psilamonocolpites grandis</i>														2.2	
	<i>Psilatricolporites salamanquensis</i>												X			
	<i>Psilatricolporites</i> spp.												X			1.2
	<i>Restioniidites</i> sp.	0.9							7							

Table 1. Cont.

Taxa		Formation / Samples		Río Foyel																
				Toroso Madero Salto del Salto del	2223	2297	2299	2226	2300	2227	2228	2229	2230	2231	2232	2233	2310	2234	2311	2312
ANGIOSPERMAE	<i>Rhoipites baculatus</i>	0.9									X									
	<i>Rhoipites minusculus</i>	2.65								X	1.2		2.3	4.6						
	<i>Rhoipites</i> sp. A	2.65													X					
	<i>Rhoipites</i> spp.		X																	
	cf. <i>Rousea patagonica</i>			1.7								X								
	<i>Spinizonocolpites</i> sp.																			
	<i>Striatricolporites</i> spp.										1.2									
	<i>Triatriopollenites bertelsii</i>																			
	<i>Triorites minor</i>																			
FUNGI	Indet. tricolporate pollen grain			1.7						X	1.2									
	<i>Desmidiospora willoughbyi</i>		X																	
	cf. <i>Diporicellaesporites navicularis</i>											X								
	<i>Diporisporites</i> cf. <i>psilatus</i>											X								
	<i>Fractisporonites</i> sp.																	X		
	<i>Hilidicellites</i> sp.		X																	
	<i>Hypoxylonites</i> sp.									X		X					X			
	<i>Inapertisporites cystoides</i>		X																	
	<i>Inapertisporites elencantense</i>		X							X										
DINOCYSTS	<i>Monoporites abruptus</i>								X											
	<i>Monoporites lagenarius</i>											X								
	<i>Multicellaesporites</i> sp.									X										
	<i>Multicellites</i> sp.														X					
	? <i>Apteodinium</i> sp.										1.2						1.2			
	? <i>Batiacasphaera baculata</i>															5.5				
	<i>Batiacasphaera micropapillata</i>	4.4		X												9.9				
	<i>Batiacasphaera</i> spp.															6.6				
	? <i>Cristadinium</i> sp.									X										
PALEOCOMMUNITIES AND PALEOCLIMATIC INFERENCES	<i>Escharisphaeridia psilata</i>	2.7			X															
	<i>Escharisphaeridia</i> spp.	2.7	3.3							X										
	<i>Hystrichostrogylion</i> sp.									X										
	<i>Lejeunecysta communis</i>	0.9		X						X										
	<i>Lejeunecysta convexa</i>									X										
	<i>Lejeunecysta fallax</i>										1.2									
	<i>Lejeunecysta globosa</i>									X										
	<i>Lejeunecysta</i> spp.											4.7								
	<i>Lingulodinium bergmannii</i>			X	X	X	X			2.4		X	4.7	7	X	12.1	1.2	12.3	47.6	
SPOROMORPHS	<i>Lingulodinium hemicystum</i>									X							1.4			
	cf. <i>Selenopemphix nephroides</i>																			
	<i>Spiniferites</i> sp.																2.3			
	<i>Tuberculodinium vancampoae</i>									X	1.2							1.4		
	Proximate indet. dinocysts								X	1.2		X							6.4	

The “non-metric multidimensional scaling” (N-MDS) (Kruskal & Wish, 1976) was used as well, which is based on rank-order of the elements of the similarity matrix, rather than on their absolute values. The procedure is based on a clear assumption: the greater the similarity between two objects, the closer to each other they will appear in the ordination (Kovach, 1989). The results presented in here correspond to the first two axes, which reflect the main variability of the data (Figure 4).

PALEOCOMMUNITIES AND PALEOCLIMATIC INFERENCES

Sporomorphs analysis (Figures 5-7) allows the characterization of the paleofloristic and paleoclimatic scenario of the studied section during the middle Eocene-Oligocene. It reflects a regional forest dominated by Nothofagaceae, Myrtaceae, Podocarpaceae and Palmae, developed under a temperate to warm-temperate and humid climate. The distribution

charts of palynomorphs and the list of identified species, and botanical affinities from the Río Foyel section are given in Tables 1 and 2, respectively.

In Río Foyel section, the species of *Nothofagidites* correspond to *N. saraensis* Menéndez & Caccavari and *N. rocaensis* Romero, which are both similar to the *Nothofagus* “Dombeyi type” and, among others, include the actual *N. dombeyi* (Mirb.) Oerst and *N. antarctica* (Forst.) Oerst. These two species are opposite ecologic indicators. The first corresponds to the evergreen forest and the second to the microtermic and deciduous forest. The paleoenvironmental inferences with these morpho-species then are limited.

Morphologically different pollen types are recognizable within the genus *Nothofagus* Blume, but a single pollen type can be found in a number of living species (Dettmann & Pocknall, 1990). This point is important, for the ecological tolerances apply to the species and if the identification is to more than one species, their ecological tolerances may not be the same (Martin, 1997).

Table 2. List of identified species, and botanical affinities.

Fossil taxon	Botanical affinity
Bryophyte and Pteridophyte spores	
<i>Azolla</i> sp.	Azollaceae (<i>Azolla filiculoides</i>)
<i>Baculatisporites comaumensis</i> (Cookson) Potonié, 1953	Osmundaceae
<i>Baculatisporites turbioensis</i> Archangelsky, 1972	?Osmundaceae
<i>Baculatisporites</i> sp. (in Barreda <i>et al.</i> , 2003)	
<i>Biretisporites crassilabratus</i> Archangelsky, 1972	?Hymenophylleaceae
<i>Biretisporites</i> sp. 1 (in Archangelsky, 1972)	
<i>Cingutriletes australis</i> (Cookson) Archangelsky, 1972	Sphagnales (<i>Sphagnum</i>)
<i>Concavisporites</i> sp.	?Filicopsida
<i>Cyatheacidites annulatus</i> Cookson, 1947	Lophosoriaceae (<i>Lophosoria</i>)
<i>Cyatheidites subtilis</i> Partridge, 1976	Cyatheaceae/Dicksoniaceae
<i>Deltoidospora australis</i> (Couper) Pocock, 1970	Polypodiaceae (<i>Acrostichum</i>)
<i>Deltoidospora minor</i> (Couper) Pocock, 1970	Polypodiaceae (<i>Acrostichum</i>)
<i>Dictyophyllidites harrisii</i> Couper, 1958	Matoniaceae (<i>Phlebopteris</i>)
<i>Gleicheniidites argentinus</i> Volkheimer, 1972	Gleicheniaceae
Hymenophylleaceae	Hymenophylleaceae
cf. <i>Hymenophyllum caudiculatum</i> Mart.	Hymenophylleaceae
<i>Ischyosporites aerapunctatis</i> (Stuchlik) Barreda, 1996	Dicksoniaceae
<i>Laevigatosporites ovatus</i> Wilson & Webster, 1946	Blechnaceae
<i>Leptolepidites macroverrucosus</i> Schulz, 1967	Dennstaedtiaceae
<i>Matonisporites ornamentalis</i> (Cookson) Partridge in Stover & Partridge, 1973	Dicksoniaceae
<i>Muricingulisporis chenquensis</i> Barreda, 1992	Pteridaceae (<i>Pteris semiadnata</i>)
<i>Peromonolites yellosus</i> Partridge, 1973	Blechnaceae
<i>Polypodiisporites inangahuensis</i> (Couper) Potonié, 1956	Polypodiaceae
emend. Pocknall & Mildenhall, 1984	
<i>Reticuloidosporites tenellis</i> Krutzsch, 1959	Polypodiaceae
Gimnosperm pollen	
<i>Araucariacites australis</i> Cookson, 1947	Araucariaceae (<i>Araucaria</i>)
? <i>Dacrycarpites australiensis</i> Cookson & Pike, 1953	Podocarpaceae
<i>Microcachryidites antarcticus</i> Cookson 1947 ex Couper, 1953	Podocarpaceae (<i>Microcachrys</i>)
<i>Phyllocladidites mawsonii</i> Cookson, 1947	Podocarpaceae (<i>Dacrydium franklinii</i>)
<i>Podocarpidites marwickii</i> Couper, 1953	Podocarpaceae
<i>Taxodiaceaepollenites hiatus</i> (Potonié) Kremp	Taxodiaceae
<i>Trisaccites microsaccatum</i> (Cookson) Couper, 1960	Podocarpaceae
Angiosperm pollen	
<i>Baumannipollis</i> sp.	Malvaceae
<i>Chenopodipollis chenopodiaceoides</i> (Martin) Truswell, 1985	Chenopodiaceae
<i>Corsinipollenites atlantica</i> Barreda, 1997	Onagraceae
<i>Graminidites</i> sp.	Poaceae
<i>Haloragacidites</i> sp.	Haloragaceae
<i>Myrtaceidites verrucosus</i> Partridge, 1973	Myrtaceae
<i>Myrtaceidites</i> sp.	Myrtaceae (cf. <i>Myrciogenia</i>)
<i>Momipites</i> sp.	Juglandaceae
<i>Nothofagidites americanus</i> Zamaloa, 1992	Nothofagaceae
<i>Nothofagidites anisoechinatus</i> Menéndez & Caccavari, 1975	Nothofagaceae
<i>Nothofagidites dorotensis</i> Romero, 1973	Nothofagaceae
<i>Nothofagidites flemingii</i> (Couper) Potonié, 1960	Nothofagaceae
<i>Nothofagidites menziesii</i> type	Nothofagaceae
<i>Nothofagidites nanus</i> Romero, 1977	Nothofagaceae
<i>Nothofagidites rocaensis</i> Romero, 1973	Nothofagaceae
<i>Nothofagidites saraensis</i> Menéndez & Caccavari, 1975	Nothofagaceae
<i>Propylipollis rynthius</i> (Stover y Partridge) Nañez <i>et al.</i> , 2009	Proteaceae
<i>Proteacidites subscaberratus</i> (Couper) Harris, 1965	Proteaceae
<i>Proteacidites symphyonemooides</i> Cookson, 1950	Proteaceae
<i>Pseudowinterapollis couperi</i> (Krutzsch) Mildenhall, 1979	Winteraceae
<i>Psilamonocolpites grandis</i> (Van der Hammen) Van der Hammen & García, 1966	Palmae
<i>Psilatricolporites salamanquensis</i> Archangelsky & Zamaloa, 1986	
<i>Restioniidites</i> sp.	Restionaceae
<i>Rhoipites baculatus</i> Archangelsky, 1973	Rutaceae-?Araliaceae
<i>Rhoipites minusculus</i> Archangelsky, 1973	Loganiaceae
<i>Rhoipites</i> sp. A (in Quattrocchio, 1980)	Rutaceae
cf. <i>Rousea patagonica</i> Archangelsky, 1973	Adoxaceae
<i>Spinizonocolpites</i> sp. (Archangelsky, 1973)	Palmae

Table 2. Cont.

Fossil taxon	Botanical affinity
Angiosperm pollen	
<i>Striaticolporites</i> spp. <i>Triatriopollenites bertelsii</i> Archangelsky, 1973 <i>Triorites minor</i> Couper, 1953	Myricaceae
Fungi	
<i>Desmidiospora willoughbyi</i> (Bradley, 1987) Ethridge Glass <i>et al.</i> , 1986 cf. <i>Diporicellaesporites navicularis</i> Katgutkar, 1993 <i>Diporisorites</i> cf. <i>psilatus</i> Kumar, 1990 <i>Fractisporonites</i> sp. <i>Hilidicellites</i> sp. <i>Hypoxylonites</i> sp. <i>Inapertisporites cystoides</i> (Ambwani) Kalgutkar & Jansonius, 2000 <i>Inapertisporites elencantense</i> Sepúlveda, 1980 <i>Monoporites abruptus</i> Sheffy & Dilcher, 1971 <i>Monoporisorites lagenarius</i> Song & Luo, 1989 <i>Multicellaesporites</i> sp. <i>Multicellites</i> sp.	
Organic walled paleomicroplankton	
? <i>Apteodinium</i> sp. ? <i>Batiacasphaera baculata</i> Drugg, 1970 <i>Batiacasphaera micropapillata</i> Stover, 1977 <i>Batiacasphaera</i> spp. ? <i>Cristadinium</i> sp. <i>Escharisphaeridia psilata</i> Kumar, 1986 <i>Hystrichostrogon</i> sp. <i>Lejeuneocysta communis</i> Biffi & Grignani, 1983 <i>Lejeuneocysta convexa</i> Matzuoka & Bujak, 1988 <i>Lejeuneocysta fallax</i> (Morgenroth) Biffi & Grignani, 1983 <i>Lejeuneocysta globosa</i> Biffi & Grignani, 1983 <i>Lingulodinium bergmannii</i> (Archangelsky) Quattrocchio & Sarjeant, 2003 <i>Lingulodinium hemicyustum</i> McMinn, 1991 cf. <i>Selenopemphix nephroides</i> (Benedek) Benedek & Sarjeant, 1981 <i>Spiniferites</i> sp. <i>Tuberculodinium vancampoae</i> (Rossignol) Wall, 1967	Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata Dinoflagellata
Algae	
<i>Botryococcus</i> sp. ? <i>Celyphus rillus</i>	Chlorococcales Rivulariaceae
Others	
Foraminiferal test linings	

The frequent presence of gymnosperm suggests that the parent plants were present in quite large numbers near to the site of deposition. Their anemophilous pollen grains (*Podocarpidites* spp. and *Microcachryidites*) characterize the regional input of palynomorphs in the depositional site.

The Araucariaceae community may have occupied ecotones associated with lowlands (García *et al.*, 2006). Some authors related them even with coastal (Abbink, 1998) and/or swampy (Whitaker *et al.*, 1992) communities. The morphologic and structural features of their pollen grains suggest that they are not suitable for transport over large distances or for eolian dispersion. The presence of araucariacean pollen grains in high proportions may be related to forests of altitude or to relatively lower areas where pollen was transported principally by fluvial currents (Martínez *et al.*, 1996). In this paper due to the increase of Araucariaceae associated with a high frequency of

pteridophytic spores in the regressive events, a lowlands scenario is suggested.

Phyllocladidites mawsonii Cookson is very similar to pollen of *Lagarostrobus franklinii* (Hook) Quinn. Today *L. franklinii* is restricted to high-rainfall areas in western Tasmania, where the annual precipitation is up to 2,500 mm. The habitat is banks of rivers and swampy flats to 750 m.

Most of the Myrtaceae pollen corresponds to *Myrtaceidites verrucosus*, a rainforest type (Martin, 1997).

The presence of Palmae, even though palms are found in tropical and subtropical climates, in Chile reaches the 35°30'S surviving -15°C during short periods in winter (Ottone, 2009).

Among the pteridophytas *Cyatheacidites annulatus* Cookson and *Deltoidospora minor* Couper and *D. australis* Couper are recognized. Dispersed spores of *Cyatheacidites* are found in rocks of Cretaceous and Cenozoic age, and are virtually identical to those of the living monotypic

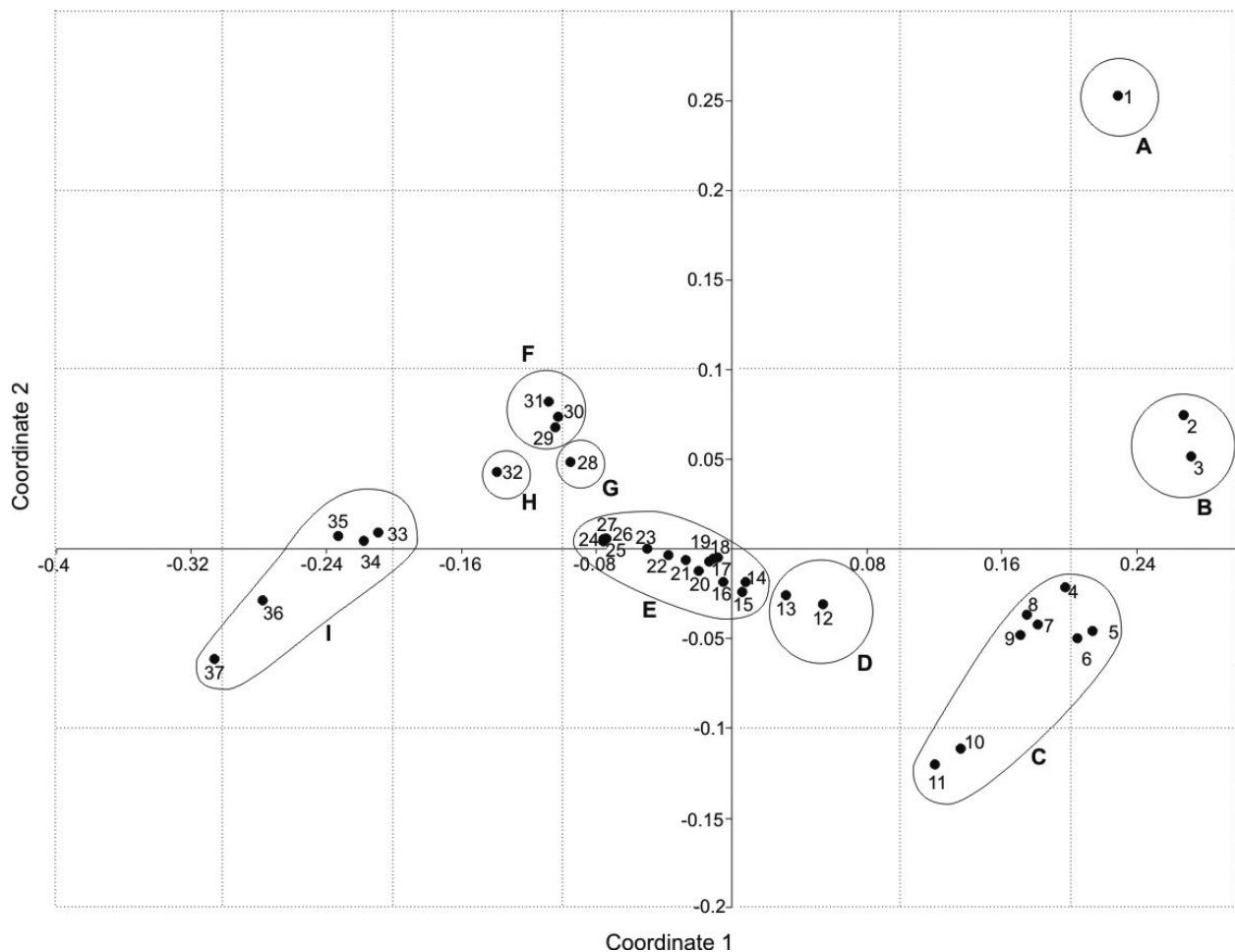


Figure 4. Groupings of palynological assemblages based on the non-parametric multidimensional scalin (N-MDS) technique. **A-B**, Danian, Cerro Bororó and Salamanca formations; **C**, Eocene, Huitrera and Río Turbio formations; **D**, late Eocene-?early Oligocene, Sloggett and Troncoso formations; **E**, Oligocene, Salto del Macho and Río Foyel formations, and “basal muddy section” of the San Julián Formation; **F-G**, late Oligocene, San Julián (Playa La Mina) and Río Leona formations; **H**, late Oligocene-early Miocene (Aries x-1 well); **I**, early-middle Miocene (Monte León and Chenque formations).

fern, *Lophosoria* according to Kurmann & Taylor (1987). Oligocene-early Miocene macrofossils of parts of a fertile frond are assigned to the extant South American species *Lophosoria quadripinnata* (Gmel.) C. Chr. These macrofossils bear the dispersed spore species *Cyatheacidites annulatus* Cookson ex Potonié (Hill *et al.*, 1999). Living *L. quadripinnata* is a large fern, now restricted to South America, where it ranges over more than 70° of latitude and occurs primarily in cloud forest in the tropics and in cool, wet regions further south. It grows in a variety of habitats and readily colonizes disturbed sites such as road cuttings and landslips, and may persist in logged forest, in pastures and in burned areas. *Deltoidospora* spores were found from Eocene-Oligocene polypodiaceous ferns close to *Acrostichum* by Collinson (1978). Anzótegui (2006) recognized a palustrine community with rooted herbaceous: *Acrostichum* cf. *A. aureum* L. in the middle to upper Miocene of the NW of Argentina. Presence of Pteridaceae, *Pteris semiadnata* Phil. (*Muricingulisperis chenquensis* Barreda) was registered. This plant inhabits humid areas, with almost constant rainfall between 5-1370 m.o.s.l. (metres over sea level), in Chile (Rodríguez, 1995).

SEDIMENTARY CYCLES

The palynological data are discussed in order to reconstruct the paleo-oceanographic and paleoclimatic conditions of the studied section.

The parameter used here for the reconstruction of relative sea-level changes is the relation of terrigenous sporomorphs to marine palynomorphs (t/m). Within a section, the values of the t/m relation may fluctuate depending on relative sea level changes, phytoplankton productivity or stronger fluvial/eolic input of terrigenous sporomorphs into the basin. They also depend on episodic regional climatic changes, which produce changes in quantitative (and qualitative) sporomorphs input.

When, for example, a progradational package in a highstand systems tract is considered in detail, it is found to consist of a series of smaller packages of beds, each of which shows an upward shallowing, progradational trend. In the terminology of sequence stratigraphy, this group of beds is called a “parasequence”.

In the Río Foyel Formation, homogeneous pelite deposits were identified; the parasequences are identified on the basis of the biofacies characteristics, which indicate upward swallowing.

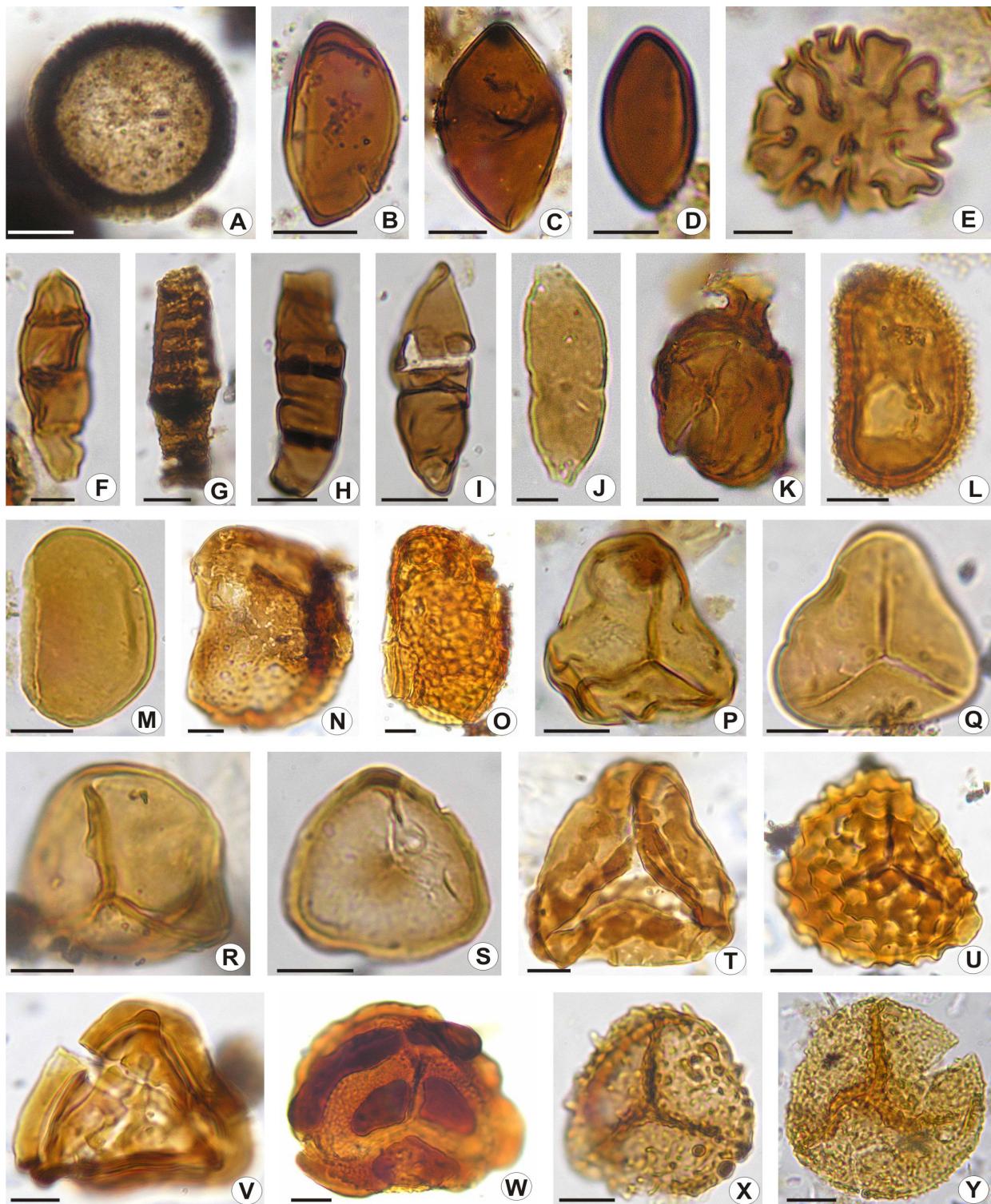


Figure 5. A, *Inapertisporites cystoides* (Ambwani) Katgutkar & Jansonius, UNSP RF 2312b: X6. B, *Hypoxylonites* sp., UNSP RF 2230c: P32. C, *Monoporites abruptus* Sheffy & Dilcher, UNSP T 2223b: P66/1. D, *Inapertisporites elencantense* Sepúlveda, UNSP T 2223b: Z56/4. E, *Desmidiospora willoughbyi* (Bradley) Ethridge Glass, Brown & Elsik, UNSP T 2223b: L19/4. F, *Multicellaesporites* sp., UNSP RF 2300c: T9. G, *Fractisporonites* sp., UNSP RF 2312b: W5/4. H, *Multicellaesporites* sp., UNSP SM 2297b: X51/2. I, *Hilidicellites* sp. UNSP T 2223c: V55. J, *Diporisporites* cf. *D. psilatus* Kumar, UNSP RF 2229c: V57. K, *Monoporisporites lagenarius* Song & Luo, UNSP RF 2229c: X64/4. L, *Peromonolites vellosus* Partridge in Stover & Partridge, UNSP RF 2311: V6. M, *Laevigatosporites ovatus* Wilson & Webster, UNSP RF 2228c: X28/3. N, *Reticuloidosporites tenellis* Krutzsch UNSP SM 2297b: R49. O, *Polypodiisporites inangahuensis* (Couper) Potonié emend. Pocknall & Mildenhall, UNSP RF 2311c: F45. P, *Cyathidites subtilis* Partridge, UNSP RF 2231c: S65/3. Q, *Deltoidospora minor* (Couper) Pocock, UNSP SM 2297b: K48/4. R, *Biretisporites crassilabratus* Archangelsky, UNSP SM 2297b: B30/4. S, *Cingulitrites australis* (Cookson) Archangelsky, UNSP SM 2297b: J54. T, *Matonisporites ornamentals* (Cookson) Partridge, in Stover & Partridge, UNSP SM 2297b: R52/1. U, *Ischyosporites areapunctatis* (Stuchlik) Barreda, UNSP SM 2297b: V9. V, *Muricingulisporis chenquensis* Barreda, UNSP SM 2297b: W53/1. W, *Cyatheacidites annulatus* Cookson, UNSP SM 2297b: V56. X, *Baculatisporites turbioensis* Archangelsky, UNSP SM 2297b: R58. Y, *Baculatisporites* sp., UNSP RF 2228c: S44. Scale bars = 10 µm, except in D, E, F, J = 5 µm.

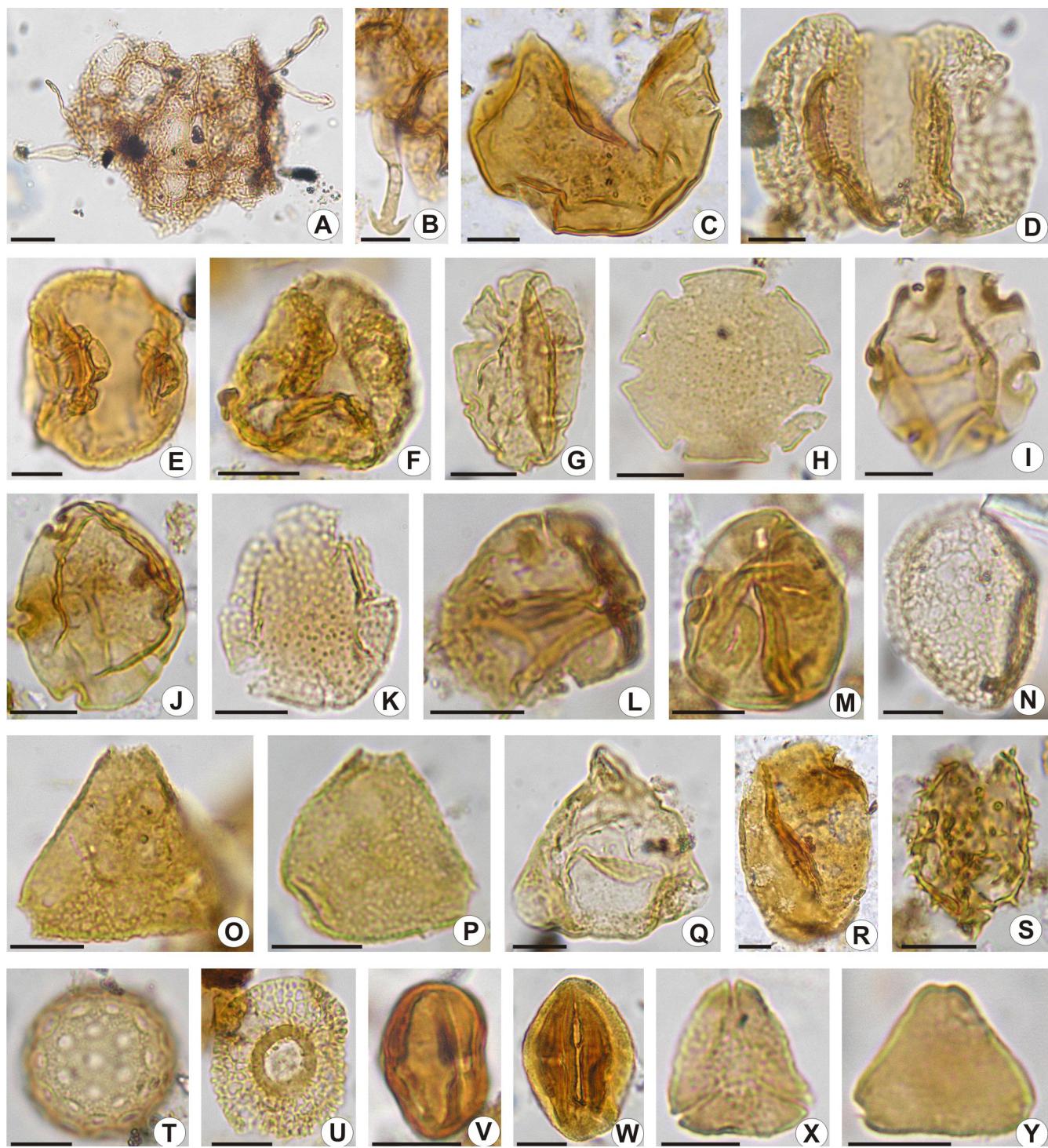


Figure 6. **A-B**, *Azolla* sp.: **A**, UNSP SM 2297: N36/2. **B**, Detail of *Glochidium*, UNSP SM 2297b: V20/2. **C**, *Taxodiaceae pollenites hiatus* (Potonié) Kremp, UNSP RF 2231c: D17. **D**, *Podocarpidites marwickii*, UNSP SM 2297: S11/3. **E**, *Phyllocladidites mawsonii*, UNSP SM 2297b: T30/1. **F**, *Trisaccites micosaccatum* (Cookson) Couper, UNSP RF 2300c: U9/3. **G**, *Nothofagidites* cf. *N. dorotensis* Romero, UNSP RF 2300c: Y50/1. **H**, *Nothofagidites fuegiensis* Menéndez & Caccavari, UNSP RF 2300d: X52/4. **I**, *Nothofagidites rocaensis* Romero, UNSP SM 2297b: Y58/2. **J**, *Nothofagidites saraensis* Menéndez & Caccavari, UNSP RF 2228c: O9/4. **K**, *Nothofagidites* "menziesii type" UNSP SM 2297b: N25/2. **L**, *Nothofagidites* sp., UNSP RF 2300c: W37/1. **M**, *Haloragacidites* sp., UNSP RF 2229c: U13. **N**, cf. *Rousea patagonica* Archangelsky, UNSP SM 2297: B29/4. **O**, *Proteacidites symphyonemoides* Cookson, UNSP RF 2300c: R12/1. **P**, *Proteacidites* sp., UNSP RF 2300d: A32. **Q**, *Corsinipollentites atlantica* Barreda, UNSP SM 2297b: R49. **R**, *Psilamonocolpites grandis* (Van der Hammen) Van der Hammen & García, UNSP RF 2233c: O46. **S**, *Spinizonocolpites* sp., UNSP RF 2229c: N30. **T**, *Chenopodipollis chenopodiaceoides* (Martin) Truswell, UNSP SM 2297b: X23/3. **U**, *Pseudowinterapolis couperi* (Krutzsch) Mildenhall, UNSP RF 2300c: B12. **V**, *Rhoipites minusculus* Archangelsky, UNSP RF 2234c: V21/1. **W**, *Rhoipites baculatus* Archangelsky, UNSP RF 2229c: U42. **X**, *Myrtaceidites verrucosus* Partridge, UNSP SM 2297b: S20/2. **Y**, *Triorites minor* Couper, UNSP RF 2300d: W44/3. Scale bars = 10 µm, except in A = 20 µm.

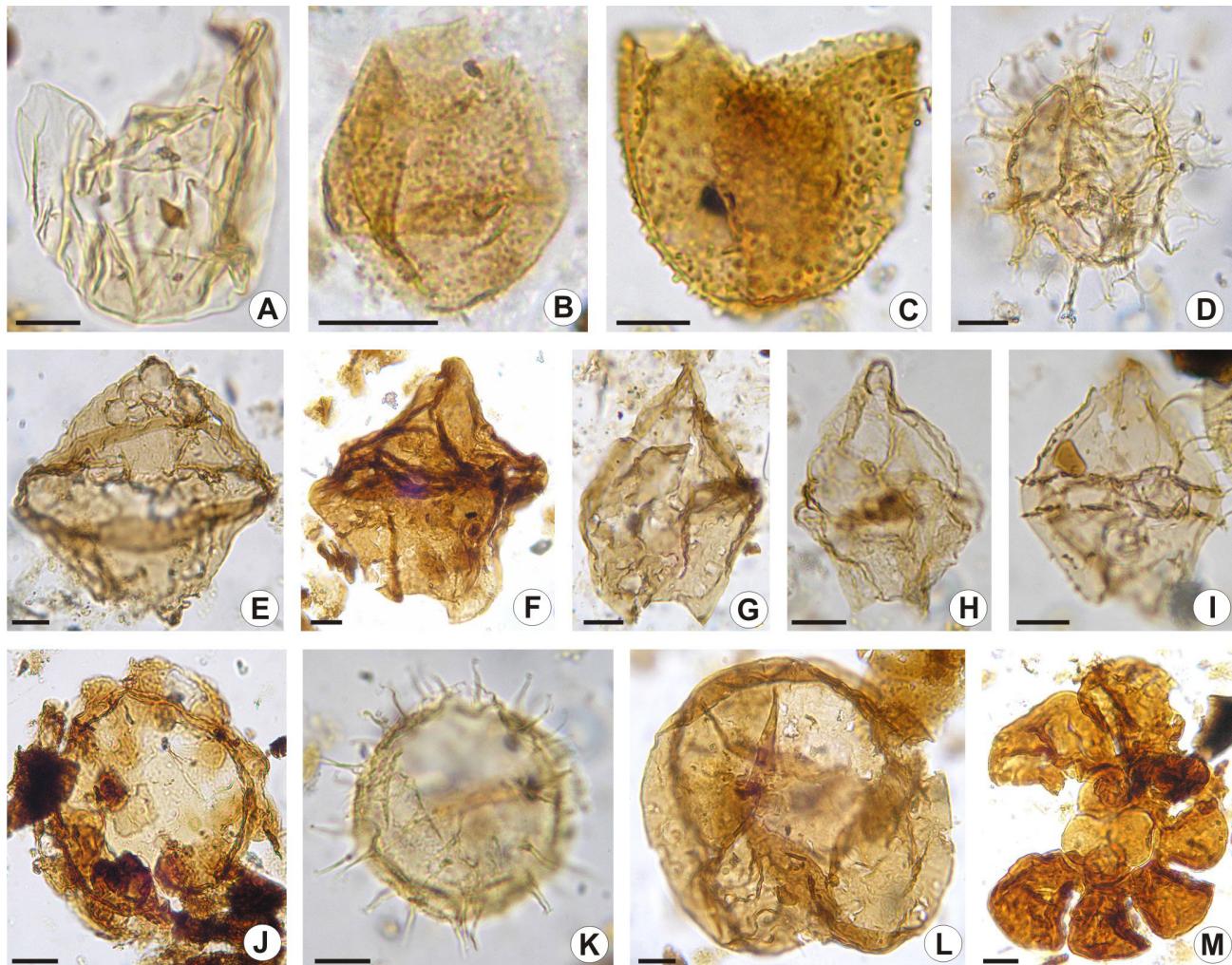


Figure 7. A, *Escharisphaeridia psilata* Kumar, UNSP T 2223c: T15. B, *Batiacasphaera micropapillata* Stover, UNSP RF 2233c: Z46. C, *?Batiacasphaera baculata* Drugg, UNSP RF 2228c: H24/3. D, *Hystrichostrogylon* sp., UNSP RF 2300d: O35/2. E, *Lejeuneucysta globosa* Biffi & Grignani, UNSP RF 2299b: K16/3. F, *Lejeuneucysta fallax* (Morgenroth) Biffi & Grignani, UNSP RF 2227c: U61/2. G, *Lejeuneucysta communis* Biffi & Grignani, UNSP RF 2299b: G37/4. H, *Lejeuneucysta convexa* Matsuoka & Bujak, UNSP RF 2300c: B53. I, *?Cristadinium* sp., UNSP RF 2300c: Y30/2. J, *Tuberculodinium vancampoae* (Rossignol) Wall, UNSP RF 2300c: Z52. K, *Lingulodinium bergmannii* (Archangelsky) Quattrocchio & Sarjeant, UNSP RF 2300c: Z26. L, cf. *Selenopemphix nephroides* (Benedek) Benedek & Sarjeant, UNSP RF 2227c: K33/3. M, Foraminiferal test linings UNSP RF 2228c: V43/4. Scale bars = 10 µm

These results are only an approach to this concept, due to limited fertile sampling. The bounding surfaces of parasequences are defined as surfaces of flooding which represent a relative sea level rise (marine flooding surfaces) (Figure 8).

In the studied section, the relation between terrestrial/marine palynomorphs in Troncoso Formation indicates the general dominance of terrigenous over marine elements. The frequent presences of gymnosperm pollen (46%) with Araucariaceae (33.6%) dominate the spectrum, indicating temperate-humid conditions. The abundance of bryophytic/pteridophytic spores in general denotes humid local condition, and the pteridophytic polypodiaceous ferns close to *Acrostichum* (*Deltoidospora*) reflect palustrine communities.

The dinocysts assemblage corresponds to only proximate cysts (*Escharisphaeridia* spp.) indicative of near shore conditions. The presence of Algae corresponds to *Botryococcus* sp. Colonies of *Botryococcus* are common in littoral areas where they accumulate in quiet and wind-protected sections (Tyson, 1995) with freshwater influence.

In Salto del Macho Formation, there is a retraction of the gymnosperm forest, associated with the dominance of *Nothofagus* Forest (30%), indicating temperate-humid condition. The pteridophytic spores (45%) correspond in particular to *Muricinguliporis chenquensis* (Pteridaceae). These plants inhabit humid areas, with almost constant rainfall.

The assemblage of dinocysts corresponds to *Escharisphaeridia* spp., indicative of a littoral marine environment. Similar conditions are inferred from 13 fertile samples of Salto del Macho Formation, in the same section, by Cornou *et al.* (2012).

In Río Foyel Formation, in the lower part of the section (from sample F21 to F23), only the “presence” (see Material and Methods) of palynomorph is registered (Pteridophytic, Gymnosperm, Nothofagaceae, Loganiaceae, Myrtaceae and dinocysts). In F23 the presence of *Tuberculodinium vancampoae* (R. Rossignol) Wall, suggest an estuarine and inner neritic environments, with warm-temperate to warm water temperatures (Wall *et al.*, 1977; Harland, 1983).

Samples F24 to F27, show an upward shallowing sequence,

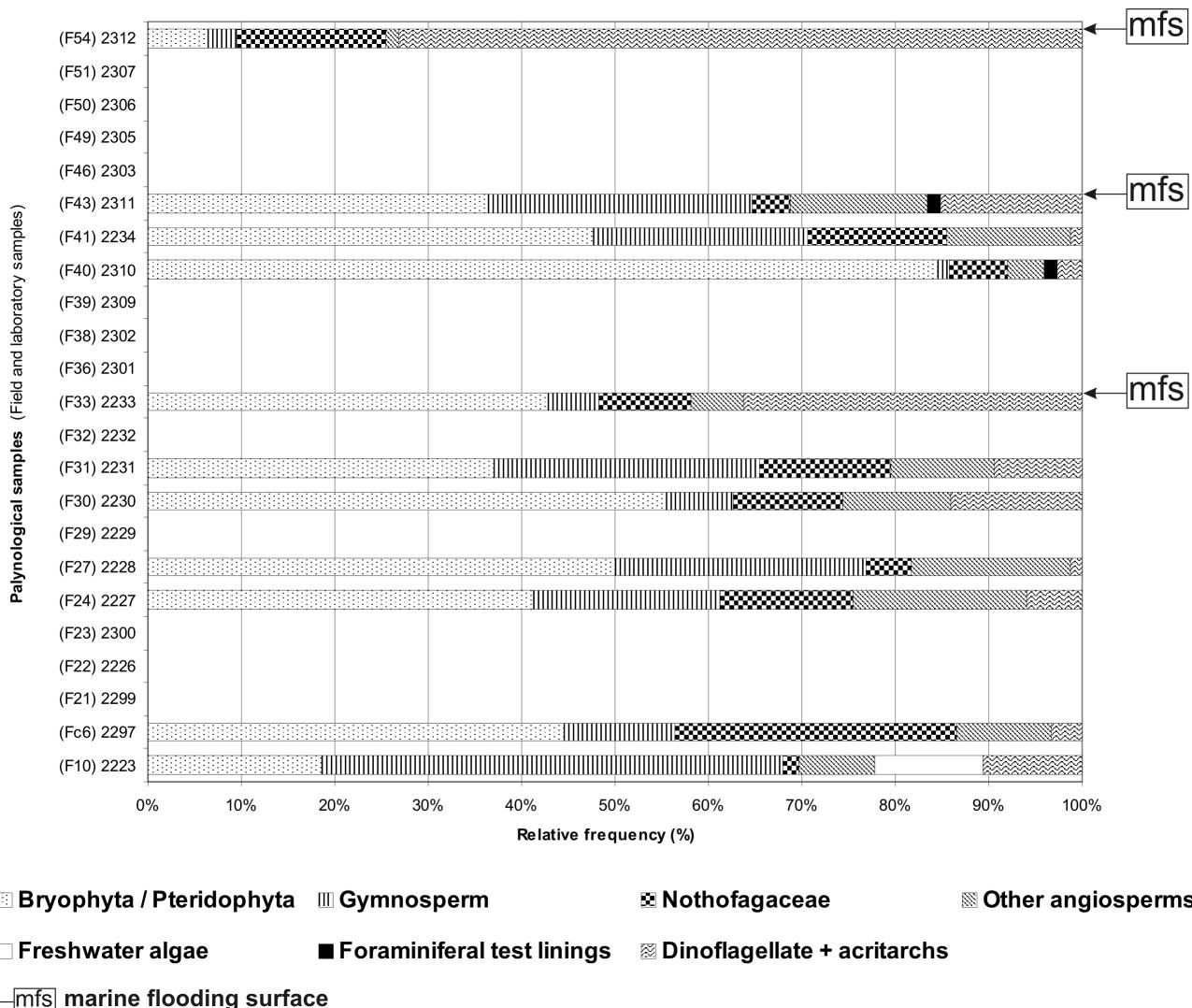


Figure 8. Relation between terrestrial and marine palynomorphs (suprageneric groups). Only stratigraphic levels with quantitative analysis are documented.

indicated by the decrease of dinocysts (6% in F24 to 1.2% in F27). The pteridophytic dominate the spectrum (41.2% in F24 to 50% in F27). In the regression (F27), the Araucariaceae (6.1%) income is associated with a decrease of the Nothofagaceae (14.1% in F24 to 4.8% in F27) and an increase of the Myrtaceae (3.5% in F24 to 6.1% in F27). *Phyllocladidites mawsonii* Cookson ex Couper have the same representativity (approximately 11% in both samples). This pulse could be associated with temperate to warm-humid climatic condition.

In F30, again, a marine influence is registered (dinocysts 14.1% with *Batiacasphaera* spp., *Lejeuneacysta globosa* Biffi & Grignani and *Lingulodinium bergmannii* Archangelsky), associated with the income of Palmae (4.6%), Nothofagaceae (11%), Myrtaceae (4.6%) and Poaceae (4.6%). The pteridophytic spores still dominate the spectrum (55.7%). Probably, this sample reflects warm conditions.

In sample F31, there is a relative decrease of marine influence (9.3%, only *Lejeuneacysta* spp. and *Lingulodinium bergmannii* are registered). An increase of gymnosperm

pollen (28%) of the montane forest indicates either less humid conditions or the generation of an altitude gradient. Due to a register of humid indicators as *Phyllocladidites mawsonii* (11.6%), Nothofagaceae (14%), and pteridophytic spores (37%), the latter could be the most valid explanation.

Probably, marine flooding surface could be detected in F33 where dinocyst reach 36.3%, with *?Batiacasphaera baculata* Drugg, *B. micropapillata* Stover, *Lejeuneacysta globosa* and *Lingulodinium bergmannii*. There is a register of Pteridophyta (42.9%), Nothofagaceae (9.9%) associated with gymnosperm pollen (5%), Palmae (2.2%) and Poaceae (3.3%). The presence of Palmae suggests relatively warm conditions.

A regressive event (dinocysts: 2.7% only *Lejeuneacysta globosa* is registered) is present in F40, where the pteridophytic spores (85.2%) correspond to pteridophytic polypodiaceous ferns close to *Acrostichum* (*Deltoidospora*) and reflect the presence of palustrine environments. The Nothofagaceae (6.3%), *Phyllocladidites mawsonii* (1.3%) and Proteaceae (6.3%) indicate humidity.

A regressive event is also registered at F41 (dinocysts: 2.4% only *Apteodinium* sp. and *Lingulodinium bergmannii* are registered), where the pteridophytic spores reach 47.6%, associated with an increase of the gymnosperm forest (23%). Nothofagaceae (14.7%) and Loganiaceae (3.7%) are registered.

At F43, a marine flooding surface (dinocysts 15% with *Lingulodinium bergmannii*, *L. hemicystum* McMinn and *Spiniferites* sp.) could be detected, associated with gymnospermous pollen from the montane forest (28%).

In F54, the maximum marine flooding surfaces of the studied section are registered (dinocysts: 73.1%, with *Lejeuneacysta globosa* and *Lingulodinium bergmannii* in high percentages). The Nothofagaceae (15.9%) have their highest representation associated to the minimum of the pteridophytic spores (6.4%) of the profile.

DISCUSSION

The microfloristic assemblages of El Foyel Group (Troncoso, Salto del Macho and Río Foyel formations), were compared to the following stratigraphic units and localities (Figure 9): Salamanca-Cerro Bororó (Danian,

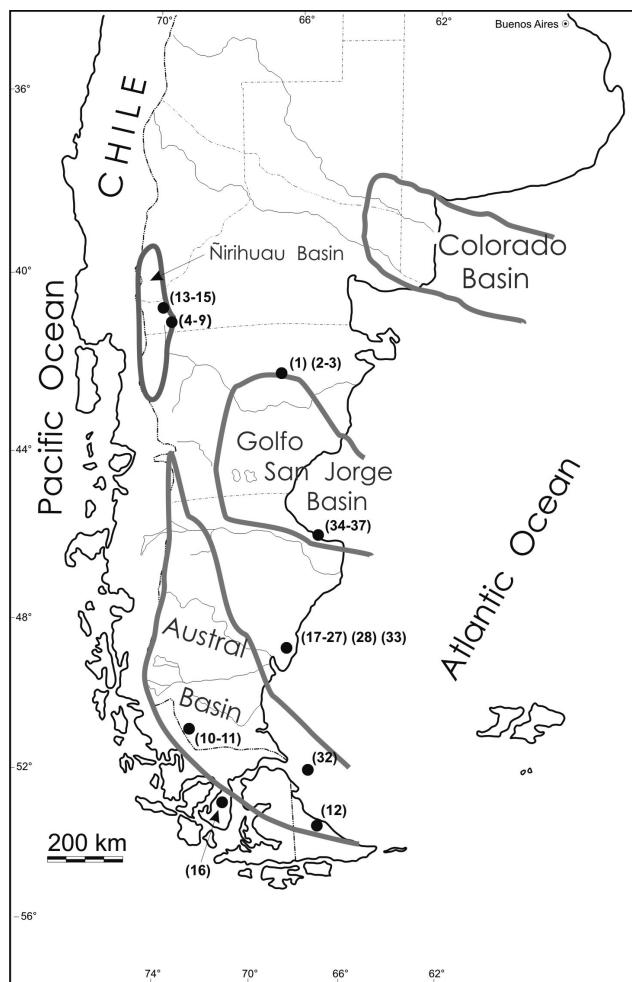


Figure 9. Map of Argentina indicating the sedimentary basins and localities compared in the text. For references of plotted numbers, see Figure 3.

Chubut Province), Huitrera (Eocene, Río Negro and Neuquén provinces), Río Turbio (Eocene, Santa Cruz Province), Sloggett (late Eocene-?early Oligocene, Tierra del Fuego) and Loreto (late Eocene-Oligocene; Magallanes, Chile) formations; El Foyel Group (Troncoso, Salto del Macho and Río Foyel formations, this paper), San Julián (Oligocene, “basal muddy section” and Playa La Mina, Santa Cruz Province); borehole Aries x-1, Tierra del Fuego (late Oligocene-early Miocene); Río Leona (early late Oligocene) and Monte León and Chenque (early-middle Miocene, Chubut Province) formations.

From the analysis of the similarity matrix and of the dendrogram (Figure 3), achieved by application of “cluster analysis”, it is evident that the higher values of similarity (close to 1) are between the samples of the Salto del Macho and Río Foyel formations and between Loreto (Fasola, 1969) and San Julián formations in CC3 (Nañez *et al.*, 2009). A second level of similarity is represented between the samples 21, 22 y 23 (FSJ in CC4), associated with the sample 20 (San Julián in CC3).

There is a close similarity between Troncoso and Sloggett formations assigned to the late Eocene-?early Oligocene. The Sloggett Formation is characterized by temperate to temperate-cold climate, very humid and without modern families, such as Poaceae, Onagraceae and Asteraceae (Olivero *et al.*, 1998).

Just as (similarity lower than 0.6) of the Salto del Macho, Troncoso and Río Foyel formations, with the San Julian Formation in Playa La Mina from the late Oligocene (Sample 28), Río Leona Formation (samples 29-31) from the late early Eocene and Aires x-1 (sample 32) from the late Oligocene-early Miocene.

Chenque and Monte León (Miocene) formations present values lower than 0.4 and, with the formations corresponding to the Eocene (Huitrera and Río Turbio formations) and to the Danian (Salamanca and Cerro Bororó formations), the values are lower than 0.3.

By means of the N-MDS (Figure 4), it was possible to determine that the samples are mainly grouped according to the analyzed formations. There are six clearly distinct groups, corresponding to the interval Paleocene-Miocene. (A-B) Danian, Cerro Bororó and Salamanca formations; (C), Eocene, Huitrera and Río Turbio formations; (D), late Eocene-?early Oligocene, Sloggett and Troncoso formations; (E), Oligocene, Salto del Macho and Río Foyel formations, and “basal muddy section” of the San Julián Formation; (F-G), late Oligocene, San Julián Formation (Playa La Mina) and Río Leona Formation; (H), late Oligocene - early Miocene (Aires x-1 well); (I), early-middle Miocene (Monte León and Chenque formations).

Then, both the Cluster Analysis and the N-MDS techniques (Figures 3, 4) yielded the same levels of similarity between the samples, which evidences a real difference between the different palynofloras here analyzed.

As in many other paleogene associations of the Patagonia

(Romero, 1986a, b; Baez *et al.*, 1990; Markgraf *et al.*, 1996), the analyzed formations have podocarpaceae linked to forms which are currently restricted to Tasmania and New Zealand (*Phyllocladidites*, *Dacrycarpites* and *Microcachryidites*), associated to the Nothofagaceae, which are currently part of the Subantarctic region. In general, this flora is characterized by Nothofagaceae, Podocarpaceae and, in lower proportion, by Araucariaceae, Myrtaceae and Proteaceae, and they do not present the diversity of angiosperms that appears in the late Oligocene.

In the late Oligocene, a community dominated by Myrtaceae, Palmae and Araucariaceae trees with Podocarpaceae and Nothofagaceae is recognized in San Julián Formation, at Playa La Mina, Santa Cruz Province (Barreda, 1997). However, the presence of small amounts of Anacardiaceae, Malvaceae, Symplocaceae, Ephedraceae, Poaceae, Asteraceae and Chenopodiaceae suggests the development of local open vegetation. The spore-pollen assemblage suggests warm and humid conditions. All of these families are present in the río Foyel section, with the exception of the local open vegetation.

As Barreda *et al.* (2003) pointed, the almost total absence of herbaceous and shrubby angiosperms in the río Foyel section (upper levels assigned to late Oligocene-early Miocene) is not consistent with the families observed in contemporary formations of the Atlantic coast. Most probably, the reason is that the paleoclimatic conditions allowed a nearly exclusive development of forest on the western side of Patagonia, while near the Atlantic coast some areas with open vegetation began to spread. Pollen assemblages from the Río Foyel Formation (northwestern area) are probably coeval or slightly older (Pöthe de Baldis, 1984; Barreda *et al.*, 2003).

Barreda *et al.* (2003) provides a detailed discussion on the marine transgression in the Ñirihuau Basin. A marine connection with the Pacific Ocean is proposed for the río Foyel section (Malumíán *et al.*, 2008; Asensio *et al.*, 2010), due to the presence of crystalline gypsum internal moulds, of both specimens and chambers of *Transversigerina* cf. *T. transversa*. Palaeoenvironmentally, *Transversigerina* is a foraminifera genus type of deep water environment, with a tendency towards upper and middle bathyal environments, which is compatible with that of other species present in large numbers in the upper levels of the Río Foyel Formation, such as *Sphaeroidina bulloides* d'Orbigny. A closed species is *T. tenua* (Cushman & Kleinpell), which characterizes the “Patagonian” in the Atlantic coast. In Chile, as opposed to Ñirihuau basin, *T. transversa* is abundant and frequent in Cholchol Formation, Temuco Basin SE of Temuco and Hueyusca Formation, Catamutún Basin, NE of Osorno, (cf. Marchant, 1990). These formations were assigned to the late middle Miocene.

Dinoflagellate assemblages exhibit similarities with those identified in the “Patagonian” deposits outcropping at the southern side of San Jorge Gulf (Barreda & Palamarczuk, 2000b), but there are no published registers of dinoflagellate coming from the Pacific Ocean at Ñirihuau Basin latitude.

CONCLUSIONS

The Age of El Foyel Group has not been established with precision and generates a lot of controversy. The present statistical results refine the stratigraphic model of Asensio *et al.* (2005), inferred from stratigraphic relations and absolute data.

In the Río Foyel Formation, homogeneous pelite deposits were registered; this palynological study allows to recognizing parasequences, identifiable on the basis of the biofacies characteristics, which indicate upward shallowing. The bounding surfaces of parasequences are defined as surfaces of flooding which represent a relative sea level rise (marine flooding surfaces) through the profile.

In general, in the Río Foyel Formation the regression events are indicated by the decrease on abundance and diversity of dinocysts and the dominance of pteridophytic spores and the gymnospermous pollen. In general, the increase of the Araucariaceae pollen corresponds to the decrease of the Nothofagaceae pollen.

The marine influence is registered by an increase on the abundance and diversity of dinocysts. In this event the Nothofagaceae, Myrtaceae, Palmae and Poaceae increase in abundance. Marine flooding surface could be detected through the Río Foyel Formation, associated with the dominance of the Nothofagaceae over the gymnospermous pollen; with the exceptions of F43, where the gymnosperm pollen dominate over the Nothofagaceae, probably due to the generation of an altitude gradient.

A comparison of these spore-pollen assemblages with others from Patagonia, using multivariate statistical techniques, yields strong similarities between the Troncoso Formation and the Sloggett Formation (late Eocene-?early Oligocene); and between Salto del Macho and Río Foyel formations with Loreto Formation (late Eocene-Oligocene) and San Julián Formation in cabo Curioso area “basal muddy section” (Oligocene).

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