

MARTHA DALILA SEDANO-PARTIDA

Chemical and biological potential of *Hyptis* Jacq. (Lamiaceae)

Potencial químico e biológico de *Hyptis* Jacq. (Lamiaceae)



São Paulo

2018

© Copyright by Martha Dalila Sedano-Partida 2018. All Rights Reserved



MARTHA DALILA SEDANO-PARTIDA

Chemical and biological potential of
Hyptis Jacq. (Lamiaceae)

Potencial químico e biológico de *Hyptis*
Jacq. (Lamiaceae)

A thesis submitted to the Institute of Biosciences of the University of São Paulo in partial fulfillment of the requirements for the degree of Doctor of Philosophy in Sciences focus on Economic Botanic.

Advisor: Dra. Cláudia Maria Furlan

São Paulo

2018

Sedano-Partida, Martha Dalila. Chemical and biological potential of *Hyptis* Jacq. (Lamiaceae). N° of pages 196

Thesis – Institute of Biosciences of the University of São Paulo. Botany Department.

Keywords: *Hyptis*; flavonoids; nepetoidins; rosmarinic acid; lithospermic acid A; antioxidant; anti-HIV; cytotoxicity; antibacterial.

The partial or total reproduction of this text is permitted for educational and research purposes on the condition that the source is cited.

Examining board

Prof(a). Dr(a).

Prof(a). Dr(a).

Prof(a). Dr(a).

Prof(a). Dr(a).

Profa Dra Cláudia Maria Furlan

*Dedico con todo mi corazón este trabajo a
mis padres, mi hermana,
mi pajarito, mi cuñado y al amor de mi vida,
por su invaluable e incondicional apoyo.*

Acknowledgments

Primeramente quiero agradecer a la Prof. Dra. Claudia Furlan por su ayuda y apoyo que fue mas allá del lado profesional, porque sin tu ayuda Clau no hubiera sido posible yo estar en la Universidad de São Paulo, por tu hospitalidad cuando llegué la primera vez a Brasil nunca voy a olvidarlo y sobre todo por el gran ejemplo que me llevo de ti de disciplina, organización, perfeccionismo, responsabilidad y compromiso ¡Muchas Gracias Clau!

A las agencias financieradoras de mi beca de estudios y de proyecto: “Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) y Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP), respectivamente.

Agradezco a la Prof. Dra. Deborah Y. A. C. dos Santos, Prof. Dra. Maria Luiza Faria Salatino y a la Dra. Paula Novaes por todas las sugerencias y correcciones durante mi examen de “qualificação” y por toda la ayuda en el laboratorio así como por la resolución de dudas y enseñanzas en sus disciplinas. Al Prof Dr. Antonio Salatino por compartir su conocimiento con nosotros y ser un excelente ejemplo de amor a la ciencia. Al Prof. Dr. Marcelo José Pena Ferreira, por el gran apoyo para la identificación de las substancias aisladas y por enseñarme tanto en su disciplina. A la Dra. Adne Righi por sus sugerencias y apoyo durante la “qualificação”.

A Mourissa por su alegría y su inmenso apoyo en todo lo referente al laboratorio durante la realización de la parte práctica de este trabajo y a Aline por dedicar su tiempo a enseñarme a usar el HPLC y CG-MS y sobre todo, a ambas por la compañía y conversaciones durante el café en la “copa”. A Leandro y Willian por la disposición en cualquier situación que acontecía en los laboratorios.

Agradezco a Norberto Palacios porque siempre me ayudó con los documentos necesarios para la renovación de mi visa demostrando buena disposición y dando apoyo sincero ¡Gracias Norberto!

Agradezco a la Dra. Cintia Luiza, por su ayuda en la identificación de las especies y por su participación como co-autora de los artículos que fueron obtenidos de este trabajo y a

la Dra. Caroline Cristina Fernandes da Silva por su orientación durante el ensayo de citotoxicidad y por la provisión del material y equipo para su realización ¡Gracias Carol!.

A la amiga que me dió el laboratorio de fitoquímica, muchas gracias Kátinha por todo el apoyo dentro y fuera de la USP, por enseñarme muchas cosas que me llevo para siempre, nunca voy a olvidar tu gran corazón y tu nobleza.

A mis grandes compañeros de la sala 143: Wilton, obrigada Wil por toda la ayuda que me diste, por ser siempre un excelente colega dispuesto a dar la mano y lo mismo para Pâmela, gracias a los dos por estar presentes en cualquier momento. A Fê Anselmo, Fê Rezende, Jana y al más nuevo Richard, los llevaré siempre en el corazón! Gracias por la compañía y fuerza en estos años.

A mis grandes compañeros de las salas 135 y 144: Gislaine, Lucas, Tamara, Cintia e Paulo. A las que ya terminaron, los queridos Carmen, Priscila, Alice y Miguel. Y una gran persona que nunca olvido, la querida Sarah Soares, por su gran amistad durante mi llegada a Brasil ¡A todos muchas gracias!

A mis amigos Carcassons que hicieron mi vida en Brasil muy alegre y divertida ¡Gracias por la compañía y la amistad incondicional!

A Isis y Roberto, por todo su apoyo y ayuda en este último año, por su interés y cariño sincero y por ser parte de mi familia, los quiero mucho ¡Gracias!

A mis queridos amigos “Butantinos”: por acompañarme y llevarme a casa o a las reuniones en mis salidas tarde de la USP, por su amor y hospitalidad durante mi llegada a Brasil, por los almuerzos, por fortalecer mi ánimo en la predicación y las reuniones, por su amistad sincera, por su compañía en los momentos difíciles y sobre todo por ser mi familia durante estos años.

A mis tíos y mis primos por hacer mis idas a México, mas divertidas, llenas de amor y por recargar mi ánimo. A mi tío Luis especialmente por todo el apoyo incondicional que siempre me ha dado. A yona Martha por mantener unida a mi familia y por darnos siempre una sonrisa a pesar de su cansancio, ir a su casa cuando están todos juntos, siempre me llena el corazón de alegría ¡Muchas gracias familia!

A mis amados abuelo y Erendira, que han sido el mas grande ejemplo de fuerza, motivación, amor a la vida y lucha incanzable, que nunca desistieron y me enseñaron con su actitud a nunca rendirme y a ver el lado positivo de la vida, verlos reír a pesar de las circunstancias era una lección de vida para mí día a día ¡Gracias por todo!

A mi hermana por ser mi compañera incondicional en este camino, compañera de cuarto, de peleas, de laboratório, de “bandejão”, de “republica”, en fin, no hay palabras, mi compañera de vida y le agradezco infinitamente por haber traído a mi familia la alegría más grande que podemos tener, mi amado sobrino Sebastián, a quien le agradezco cada uno de los momentos que ha alegrado en mi vida desde que nació, gracias pajarito por todas tus sonrisas y gracias hermana porque aunque eres la menor me has enseñado muchas cosas, sobre todo a no rendirme y a no tener miedo y has estado a mi lado en cada momento de esta etapa. A mi cu, por apoyarme en todos los sentidos siempre que he necesitado durante mi estadia en Brasil y por ser un amigo de verdad. ¡Gracias a ustedes tres por la hospitalidad y compañía a lo largo de estos años! ¡Los amo!

A mis padres por que sin ellos nada de esto sería posible, no existen palabras de agradecimiento para expresar el profundo amor y respeto que siento por ustedes, son todo para mi. Gracias por el ánimo que me dan siempre que lo necesito. Gracias por enseñarme a no rendirme y por apoyar todos mis sueños, sin ustedes yo no sería nada, este logro es suyo, mas que mío. A la Susy, la Nany y el Mateo que tanto nos llenan de alegría ¡Los amo!

Y por último, quiero agradecer a la persona que ha cambiado mi vida con su presencia y llegó para ser mi mejor amigo, mi compañero, mi soporte, quien me ha ayudado con cariño y profundo amor. Gracias por tu incondicionalidad, por estar siempre para mi, por no soltar mi mano nunca, por ser mi fuente de ánimo, mi bendición de Jehová. Gracias a ti mi esposo, Gaudi ¡Te amo mi amor!

Dalila Sedano

List of tables

CHAPTER II

Table 2.1. Ethnopharmacological/pharmacological, economic uses, and chemical composition reported for <i>Hyptis sensu</i> Harley and Pastore (2012). SciFinder, Web of Science and SciELO databases were as a source of information (1906 to 2018).....	22
--	----

CHAPTER III

Table 3.1. Yield (%) of crude ethanol extract (EE) and its phases from <i>H. radicans</i> and <i>H. multibracteata</i> : hexane phase (HP), ethyl acetate phase (EAP), and hydromethanol phase (HMP).....	66
--	----

Table 3.2. Content of phenolic compounds (mg g^{-1}) of crude ethanol extract (EE) and phases of <i>H. radicans</i> and <i>H. multibracteata</i> analyzed by HPLC-DAD detected in 280 nm: hexane phase (HP), ethyl acetate phase (EAP), hydromethanol phase (HMP).....	69
--	----

Table 3.3. Major constituents of crude ethanol extract (EE) and phases (EAP, ethyl acetate; HP, hexane; and HMP, hydromethanol) of <i>H. radicans</i> and <i>H. multibracteata</i> analyzed by HPLC-DAD (280 nm) and expressed as mg g^{-1} . RT: retention time.....	
---	--

Table 3.4. ^1H , ^{13}C -NMR and HMBC data of compound Lithospermic acid A (16) in CD_3OD compared to data from the literature in CD_3OD	80
--	----

Table 3.5. ^1H NMR data for Ethyl caffeate (20) in CD_3OD compared to data from the literature in $\text{DMSO-d}6$	82
---	----

Table 3.6. ^1H NMR data for Nepetoidin B (24) in CD_3OD compared to data from the literature in $\text{DMSO-d}6$	83
---	----

Table 3.7. ^1H NMR data for Cirsimarinin (25) in CDCl_3 compared to data from the literature in CDCl_3	84
---	----

Table 3.8. Relative abundance (%) and mass spectra fragmentation of constituents from crude ethanol extract (EE) and phases (EAP, ethyl acetate; HP, hexane; and HMP, hydromethanol) of <i>H. radicans</i> and <i>H. multibracteata</i> analyzed by GC-MS. RT: retention time.....	90
---	----

CHAPTER IV

Table 4.1. Contents of antioxidant substances equivalents to trolox, quercetin, and galic acid per gram of dry extract for DPPH, ABTS, FRAP, ORAC, and Metal chelating assays. in Ethanol crude extract (EE), hexane phase (HP), ethyl acetate phase (EAP), hydromethanol phase (HMP) from H.rad - <i>Hyptis radicans</i> ; H. mul - <i>Hyptis multibracteata</i>	145
--	-----

Table 4.2. Effective concentration of EE, HP, EAP and HMP, from of *H. radicans* and *H. multibracteata* to achieve 50% antioxidant activity (EC_{50} in $\mu\text{g mL}^{-1}$) for antioxidant assays: sequestration of DPPH and ABTS, FRAP, ORAC, metal chelator assay (MCh), lipid peroxidation (TBARS), Site-Specific (S-Spe) and nonsite-Specific (NS-Spe) OH[•] mediated 2-Deoxy-d-ribose degradation..... 146

Table 4.3. Percent cell inhibition of *H. radicans* and *H. multibracteata* extracts and phases in a concentration of 100 $\mu\text{g mL}^{-1}$ on RAW 264.7 Cell Lines (macrophages)..... 155

Table 4.4. Effective concentration of isolated substances from *H. radicans* and *H. multibracteata* to achieve 50% of antioxidant (EC_{50} $\mu\text{g mL}^{-1}$) and anti-acetylcholinesterase (EC_{50} ng mL^{-1}) activities tested by the methods of sequestration of DPPH, ORAC, Site-Specific (S-Spe), nonsite-Specific (NS-Spe) OH[•] mediated 2-Deoxy-d-ribose degradation, and anti-acetylcholinesterase enzyme assay (AChE)..... 157

CHAPTER V

Table 5.1. Yield (%) of EE and phases (HP, EAP and HMP) of *H. radicans* and *H. multibracteata*..... 179

Table 5.2. Content of phenolic compounds (mg g^{-1}) of EE and phases of *H. radicans* and *H. multibracteata* analyzed by HPLC-DAD detected in 280 nm: HP, EAP and HMP..... 181

Table 5.3. Minimal inhibitory Concentration ($\mu\text{g mL}^{-1}$) of EE and phases of *H. radicans* and *H. multibracteata* to inhibit 50% of Reverse Transcriptase activity (HIV1). HP, EAP and HMP..... 181

Table 5.4. Bacteriostatic effect (MIC_{50} $\mu\text{g mL}^{-1}$) of EE and phases (HP, EAP and HMP) of *H. radicans* (*H. rad*) and *H. multibracteata* (*H. mul*) against *Bacillus subtilis*, *Pseudomonas aeruginosa* and *Escherichia coli*..... 184

List of figures

CHAPTER I

- Figure 1.1** A simple schematic representation of the major secondary metabolites in plants. (Ncube and Van Staden, 2015)..... 7

CHAPTER III

- Figure 3.1.** (A) Geographic distribution of *Hyptis radicans* (Pohl) Harley & J.F.B.Pastore. This species is accepted, and is native to Brazil South, Brazil Southeast and Brazil West-Central. (B)Geographical distribution of *Hyptis multibracteata* Benth. This species is accepted, and is native to Brazil South, Brazil Northeast and Brazil Southeast. (Images: powo.science.kew.org by Royal Botanic Gardens & Plants of the World Online)..... 62

- Figure 3.2.** *Hyptis* species used in this study. A) *H. radicans* and B) *H. multibracteata* (Photos: Furlan)..... 63

- Figure 3.3.** Flowcharts of crude extract and phases of *H. radicans* and *H. multibracteata*: EE: crude ethanol extract; HP: hexane phase; EAP: ethyl acetate phase; HMP: hydromethanol phase..... 64

- Figure 3.4.** Chromatograms obtained by HPLC-DAD (280 nm) of extract and phases of: *H. radicans* - A. Ethanol extract, B. Ethyl acetate phase, C. Hexane phase, D. Hydromethanol phase; and *H. multibracteata*, E. Ethanol extract, F. Ethyl acetate phase, G. Hexane phase, H. Hydromethanol phase. Numbers above each peak correspond to identifications shown in **table 3.3**. All chromatograms are on the same scale for comparison purposes..... 67

- Figure 3.5.** Flowchart of crude ethanol extract from *H. radicans*, its fractions and sub-fractions from Column Chromatography (Sephadex) and semi-preparative HPLC. Grey boxes represent sub-fractions with isolated substances according to data from HPLC-DAD..... 72

- Figure 3.6.** Flowchart of crude ethanol extract from *H. multibracteata*, its fractions and sub-fractions from Column Chromatography (Sephadex) and semi-preparative HPLC. Grey boxes represent sub-fractions with isolated substances according to data from HPLC-DAD..... 73

- Figure 3.7.** UV/Vis absorption spectra of the major constituents detected by HPLC-DAD (280 nm) in the crude ethanol extract and phases of *H. radicans* and *H. multibracteata*. Numbers correspond to the identification shown in Table 3.3..... 75

- Figure 3.8.** Structure of **Lithospermic acid A**..... 80

- Figure 3.9.** Structure of **ethyl caffeoate**..... 82

- Figure 3.10.** Structure of **Nepetoidin B**..... 83

Figure 3.11. Structure of Cirsimaritin	84
Figure 3.12. CG/MS Chromatograms of extract and phases of: <i>H. radicans</i> - A. Ethanol extract, B. ethyl acetate phase, C. hexane phase, D. Hydromethanol phase; and <i>H. multibracteata</i> , E. Ethanol extract, F. ethyl acetate phase, G. hexane phase, H. hydromethanol phase). Numbers above each peak correspond to the possible identifications shown in table 3.7.....	89
CHAPTER III Supplemental information	
Supplemental figure 3.8.1. ^1H NMR spectra (CD_3OD) of Lithospermic acid A (16)	107
Supplemental figure 3.8.2. Amplification of signals between regions at δ 5.1 and δ 7.9 from ^1H NMR spectra (CD_3OD) of Lithospermic acid A (16)	108
Supplemental figure 3.8.3. ^{13}C NMR spectra (CD_3OD) of Lithospermic acid A (16)	109
Supplemental figure 3.8.4. Amplification of signals between regions at δ 30 and δ 175 from ^{13}C NMR spectra (CD_3OD) of Lithospermic acid A (16)	110
Supplemental figure 3.8.5. HSQC spectra (CD_3OD) of Lithospermic acid A (16)	111
Supplemental figure 3.8.6. Amplification of signals between regions at δ 1.5 and δ 8.0 from HSQC spectra (CD_3OD) of Lithospermic acid A (16)	112
Supplemental figure 3.8.7. HMBC spectra (CD_3OD) of Lithospermic acid A (16)	113
Supplemental figure 3.8.8. Amplification of signals between regions at δ 2.6 and δ 8.0 from HMBC spectra (CD_3OD) of Lithospermic acid A (16)	114
Supplemental figure 3.9.1. ^1H spectra (CD_3OD) of Ethyl caffeoate (20)	115
Supplemental figure 3.9.2. Amplification of signals between regions at δ_{H} 1 and δ_{H} 7.5 from ^1H NMR spectra (CD_3OD) of Ethyl caffeoate (20)	116
Supplemental figure 3.10.1. A): ^1H NMR spectra (CD_3OD) of Nepetoidin B (24)	117
Supplemental figure 3.10.2. Amplification of signals between regions at δ 5.6 and δ 7.8 from ^1H NMR spectra (CD_3OD) of Nepetoidin B (24)	118
Supplemental figure 3.11.1. ^1H NMR spectra (CDCl_3) of Cirsimaritin (25)	119
Supplemental figure 3.11.2. Amplification of signals between regions at δ 3.8 and δ 7.9 from ^1H NMR spectra (CDCl_3) of Cirsimaritin (25)	120
Supplemental figure 3.12.1. 1-Pentadecanone, 6,10,14-trimethyl (Hexahydrofarnesyl acetone) N° 7. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder [®]).....	121
Supplemental figure 3.12.2. n-Hexadecanoic acid methyl ester (Palmitic acid, methyl ester) N° 9. A) Experimental mass spectrum; B) Reference mass spectrum	

(SciFinder®).....	121
Supplemental figure 3.12.3. n-Hexadecanoic acid (Palmitic acid) N° 10. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	122
Supplemental figure 3.12.4. n-Hexadecanoic acid, ethyl ester (Palmitic acid, ethyl ester) N° 11. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	122
Supplemental figure 3.12.5. 3,7,11,15-Tetramethylhexadec-2-en-1-ol (Phytol) -> Phytol derivative N° 15. A) Experimental mass spectrum; B) Reference mass spectrum NIST Mass Spectrometry Data Center.....	123
Supplemental figure 3.12.6. n-Octadecanoic acid, methyl ester (Methyl stearate) N° 16. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	123
Supplemental figure 3.12.7. 9,12-Octadecadienoic acid (Z,Z)- (Linoleic acid) N° 17. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	124
Supplemental figure 3.12.8. 9,12-Octadecadienoic acid (Z,Z)-, ethyl Ester (Linoleic acid ethyl ester) N° 19. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	124
Supplemental figure 3.12.9. Ethyl 9,12,15-octadecatrienoate (Linolenic acid, ethyl Ester) N° 20. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	125
Supplemental figure 3.12.10. Octadecanoic acid, ethyl Ester (Stearic acid, ethyl ester) N° 22. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	125
Supplemental figure 3.12.11. n-Eicosanoic acid (Arachidic acid) N° 23. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	126
Supplemental figure 3.12.12. Hexadecanoic acid, 4-nitrophenyl ester (4-nitrophenyl palmitate) N° 24. A) Experimental mass spectrum; B) Reference mass spectrum (NIST Mass Spectrometry Data Center).....	126
Supplemental figure 3.12.13. Eicosanoic acid, ethyl ester (Arachidic acid, ethyl Ester). N° 25. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	127
Supplemental figure 3.12.14. Phenol, 2,2'-methylenebis[6-(1,1-dimethylethyl)-4-ethyl- N° 28. A) Experimental mass spectrum; B) Reference mass spectrum (NIST Mass Spectrometry Data Center).....	127
Supplemental figure 3.12.15. 1,2-Benzenedicarboxylic acid, diisooctyl ester (Phthalic acid, diisooctyl ester) N° 29. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	128

Supplemental figure 3.12.16. 24α-Methyl-5-cholest-en-3β-ol (Campesterol) N° 32. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	128
Supplemental figure 3.12.17. Stigmasta-5,22-dien-3β-ol (Stigmasterol) N° 33. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	129
Supplemental figure 3.12.18. Stigmast-5-en-3β-ol (β-sitosterol) N° 34. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	129
Supplemental figure 3.12.19. Olean-12-en-3β-ol (β-amyrin) N° 35. A) Experimental mass spectrum; B) Reference mass spectrum (NIST Mass Spectrometry Data Center)...	130
Supplemental figure 3.12.20. Urs-12-en-3β-ol (α-amyrin) N° 36. A) Experimental mass spectrum; B) Reference mass spectrum (NIST Mass Spectrometry Data Center)....	130
Supplemental figure 3.12.21. Stigmasta-3,5-dien-7-ona (Tremulone) N° 37. A) Experimental mass spectrum; B) Reference mass spectrum (SciFinder®).....	131

CHAPTER IV

Figure 4.1. Principal component analyses (PCA) using 13 variables (Fla: flavonoids; NeAB: nepetoidins A and B; CinD: cinnamic acid derivatives; ChlD: chlorogenic acid derivatives; PhA: phenolics acids; RA: rosmarinic acid; S-Spe and NS-Spe: site-specific and nonsite-specific hydroxyl radical-mediated 2-Deoxy-D-ribose degradation; TBARS: thiobarbituric acid reactive substances; MCh: metal chelating assay; ABTS: 2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulfonic acid scavenging assay; DPPH: 2,2-Diphenyl-1-picrylhydrazyl scavenging assay; ORAC: oxygen radical absorbance capacity; FRAP: ferric-reducing antioxidant potential) evaluated in four samples of <i>H. radicans</i> (Hrad-EE: crude ethanol extract; Hrad-HP: hexane phase; Hrad-EAP: ethyl acetate phase; Hrad-HMP: hydromethanol phase), and four samples of <i>H. multibracteata</i> (Hmul-EE: crude ethanol extract; Hmul-HP: hexane phase; Hmul-EAP: ethyl acetate phase; Hmul-HMP: hydromethanol phase). a) Percentage of variance explained by PCA, and percentage of expected variance estimated by the broken-stick test; and b) correlation coefficients between variables and axes 1 and 2.....	153
---	-----

Figure 4.2. Binding sites for trace metal (Image: Pietta, 2000).....	159
---	-----

Figure 4.3. Antioxidant structure-activity relationships including: <i>ortho</i> -dihydroxy arrangement in the B ring (a); C ₂ -C ₃ unsaturated bond combined with C ₄ carbonyl group in the C skeleton (b) and hydroxyl groups (c); O-methylation (d); this last structure belongs to Cirsimarinin (25). Adapted from Banjarnahor and Artanti (2014)....	160
--	-----

CHAPTER V

Figure 5.1. Principal component analyses (PCA) using 12 variables (**PhA**: phenolics acids; **RA**: rosmarinic acid; **NeAB**: nepetoidins A and B; **CinD**: cinnamic acid derivatives; **ChID**: chlorogenic acid derivatives; **Fla**: flavonoids; **NI**: no identified compounds; **Total**: total phenolic content; **E. coli**: *Escherichia coli*; **B. sub**: *Bacillus subtilis*; **P. aerug**: *Pseudomonas aeruginosa*; **HIV**: inhibit of Reverse Transcriptase activity); evaluated in four samples of *H. radicans* (**Hrad-EE**: crude ethanol extract; **Hrad-HP**: hexane phase; **Hrad-EAP**: ethyl acetate phase; **Hrad-HMP**: hydromethanol phase), and four samples of *H. multibracteata* (**Hmul-EE**: crude ethanol extract; **Hmul-HP**: hexane phase; **Hmul-EAP**: ethyl acetate phase; **Hmul-HMP**: hydromethanol phase). **a)** Percentage of variance explained by PCA, and percentage of expected variance estimated by the broken-stick test; and **b)** correlation coefficients between variables and axes 1 and 2.....

Abstract

Flavonoids and other phenolics are groups of natural bioactive compounds widely distributed in edible plants and are well documented to possess biological potential. *Hyptis* (Lamiaceae) is used in Brazilian folk medicine to treat various diseases. The aim of this study was to evaluate the antioxidant, anti-acetylcholinesterase, cytotoxic, antiviral and antibacterial potential of *Hyptis radicans* and *Hyptis multibracteata* by isolating and characterizing major constituents and their biological activities. *H. radicans* and *H. multibracteata* were dried, powdered and macerated in 70% ethanol which resulted in a crude ethanol extract (EE) for each species. EE were dissolved in 50% methanol and then was fractionated by partition with hexane and ethyl acetate; were obtained three phases: hexane phase (HP), ethyl acetate phase (EAP) and hydromethanol phase (HMP). EAP from *H. radicans* was the sample that presented the highest levels of total phenolic content, especially flavonoids, and was the sample with the high antioxidant activity with promising values of EC₅₀: DPPH (32.12 µg mL⁻¹), ABTS (5.04 µg mL⁻¹), Metal chelator assay (42.36 µg mL⁻¹), TBARS (40.46 µg mL⁻¹) and nonsite-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation (NS-Spe) with EC₅₀ of 75.08 µg mL⁻¹. EE from *H. radicans* showed high antioxidant activity for FRAP and ORAC with EC₅₀ of 6.01 and 2.68 µg mL⁻¹, respectively and had the highest amount of rosmarinic acid (17.64 mg ρ-CE g⁻¹). HMP from *H. radicans* showed high antioxidant activity in Site-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation (S-Spe) assay with EC₅₀ of 0.32 µg mL⁻¹ and had the highest content of chlorogenic acid derivatives. Regarding the results of cytotoxicity, HP from *H. multibracteata* induced the death of more than 80% of RAW 264.7 Cell Lines at 100 µg mL⁻¹. Nepetoidin B, isolated from *H. multibracteata* had the best EC₅₀ (52.73 µg mL⁻¹) for anti-acetylcholinesterase activity. Antibacterial activity was evaluated *in vitro* against two Gram-negative bacteria, *Pseudomonas aeruginosa* and *Escherichia coli*, and a Gram-positive *Bacillus subtilis*. Phases from *H. multibracteata* were more effective on inhibiting *B. subtilis* with MIC₅₀ of 23.6 µg mL⁻¹ and 12.13 µg mL⁻¹ for HP and EAP, respectively. HP was also activity against *P. aeruginosa* with MIC₅₀ of 37.55 µg mL⁻¹. EE and HMP phase from *H. radicans* showed moderate anti-HIV-1 activity (MIC₅₀ 159 µg mL⁻¹; MIC₅₀ 180 µg mL⁻¹). Contents of total phenolic were not the main sample feature to define this activity, but there

was correlation between Rosmarinic acid contents and anti-HIV1 activity of *H. radicans*. Cirsimarinin and litospermic acid A were isolated for the first time, being the first time that they are described for the genus *Hyptis*. This study provides the first evidence of chemical and biological potential for these two Brazilian native species of *Hyptis*.

Keywords

Hyptis; flavonoids; nepetoidins; rosmarinic acid; lithospermic acid A; antioxidant; anti-HIV; cytotoxicity; antibacterial.

Resumo

Flavonoides e outros compostos fenólicos são grupos de compostos bioativos naturais amplamente distribuídos em plantas e estão bem documentados por possuírem potencial biológico. *Hyptis* (Lamiaceae) é usado na medicina popular brasileira para tratar várias doenças. O objetivo deste estudo foi avaliar o potencial antioxidante, anti-acetilcolinesterase, citotóxico, antiviral e antibacteriano de *Hyptis radicans* e *Hyptis multibracteata*, isolar e identificar substâncias e correlacionar as atividades biológicas com a quantidade de compostos fenólicos e substâncias isoladas. *H. radicans* e *H. multibracteata* foram secas, pulverizadas e maceradas em etanol 70%, resultando em extrato etanólico bruto (EE). EE foi dissolvido em metanol 50% e depois foi fracionado por partição com hexano e acetato de etila, o que resultou em três fases: fase hexânica (HP), fase acetato de etila (EAP) e fase hidrometanólica (HMP). EAP de *H. radicans* foi a amostra que apresentou os maiores teores de conteúdo fenólico, principalmente flavonoides, e foi a amostra com a maior atividade antioxidante, com valores promissores de EC₅₀: DPPH (32,12 µg mL⁻¹), ABTS (5,04 µg mL⁻¹), Quelante de metais (42,36 µg mL⁻¹), TBARS (40,46 µg mL⁻¹) e Degradação da 2-deoxy-D-ribose de sitio não específico mediada pelo radical hidroxil (NS-Spe) com EC₅₀ de 75,08 µg mL⁻¹. EE de *H. radicans* apresentou a maior atividade antioxidante para FRAP e ORAC com EC₅₀ de 6,01 e 2,68 µg mL⁻¹, respectivamente, e apresentou a maior quantidade de ácido rosmariníco (17,64 mg p-CE g⁻¹). HMP de *H. radicans* apresentou a mais alta atividade antioxidante no ensaio de Degradação da 2-deoxy-D-ribose de sitio específico mediada pelo radical hidroxil (S-Spe) com EC₅₀ de 0,32 µg mL⁻¹ e apresentou o maior teor de derivados de ácido clorogênico. Em relação aos resultados da citotoxicidade, HP de *H. multibracteata* induziu a morte de mais de 80% das células do tipo RAW 264.7 com uma concentração de 100 µg mL⁻¹. A Nepetoidina B isolada de *H. multibracteata* apresentou a melhor EC₅₀ (52,73 µg mL⁻¹) para atividade anti-acetilcolinesterase. A atividade antibacteriana foi avaliada *in vitro* contra duas bactérias Gram-negativas, *Pseudomonas aeruginosa* e *Escherichia coli*, e uma bactéria Gram-positiva, *Bacillus subtilis*. Fases de *H. multibracteata* foram mais eficazes na inibição de *B. subtilis* com MIC₅₀ 23,6 µg mL⁻¹ e 12,13 µg mL⁻¹ para HP e EAP, respectivamente. HP também apresentou atividade contra *P. aeruginosa* com MIC₅₀ de 37,55 µg mL⁻¹. EE e HMP de *H. radicans* mostraram moderada

atividade anti-HIV-1 (MIC_{50} 159 $\mu\text{g mL}^{-1}$; MIC_{50} 180 $\mu\text{g mL}^{-1}$). Não há correlação entre o conteúdo total de fenólicos e esta atividade biológica, mas sim entre a quantidade de ácido rosmarínico das fases e a atividade anti-HIV₁ de *H. radicans*. Foram isoladas pela primeira vez a Cirsimaritina e o ácido litospermico A, sendo esta a primeira vez que se descrevem para o gênero *Hyptis*. Este estudo fornece a primeira evidência do potencial químico e biológico para estas duas espécies nativas de *Hyptis*.

Palavras chave:

Hyptis; Flavonoides; nepetoidinas; ácido rosmarínico; ácido litospermico A; antioxidante, anti-HIV; citotoxicidade; antibacterial.

Summary

CHAPTER I

Plant-derived drug discovery and special metabolism.....	1
Historical perspectives.....	1
Natural products as drug candidates: significance and advantages against synthetic compounds.....	3
Where do these medicinal substances originate in plants?.....	5
What is a special (or secondary) metabolite and which is their role in plants?.....	5
Special metabolism and major groups of plant special metabolites.....	7
Terpenes.....	7
Nitrogenous compounds alkaloids.....	8
Phenolic compounds.....	9
Aims of the present study.....	10
References.....	12

CHAPTER II

Hyptis Jacq.: a general chemical profile review.....	17
Results and discussions.....	40
Conclusions.....	42
References.....	43
Supplementary information.....	56

CHAPTER III

Botanical aspects & chemical description of <i>Hyptis radicans</i> and <i>Hyptis multibracteata</i>	59
General taxonomic and morphological aspects of <i>Hyptis</i> Jaqc.....	59
Material and methods.....	63
Plant Material.....	63
Extraction and Fractionation (plant material and sample preparation).....	63
Chemical analyzes.....	64
Isolation of bioactive compounds and identification by NMR.....	65
Results and discussions.....	66
Extraction yield.....	66
Isolation and identification of non-volatiles constituents.....	67
First step to isolation: HPLC-DAD to see chemical profiles.....	67
Second step to isolation: exclusion column chromatography using Sephadex.....	70
After isolation...third step is identification.....	74
NMR results.....	79
Quantification and identification of nonpolar substances.....	87
Conclusions.....	97
References.....	98
Supplementary information.....	107

CHAPTER IV

Antioxidant, Anti-acetylcholinesterase and cytotoxic potential of <i>Hyptis</i> spp.....	133
Oxidation, reactive oxygen species (ROS), what does these mean and which are their consequences?.....	133

Antioxidant defense system.....	134
Why are antioxidant compounds important against inhibition of acetylcholinesterase?	135
Antioxidant potential in Lamiaceae.....	137
Free radical scavenging activity determination using DPPHo (2,2-Diphenyl-1-picrylhydrazyl).....	138
Free radical scavenging activity determination using ABTSo (2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulfonic acid).....	138
Ferric-reducing antioxidant potential (FRAP).....	139
Metal chelating assay.....	139
Determination of oxygen radical absorbance capacity (ORAC).....	139
Inhibition of lipid peroxidation assay (TBARS).....	140
Nonsite-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation.....	140
Site-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation.....	141
Acetylcholinesterase Inhibitory Activity.....	141
Cytotoxic activity.....	142
Results and discussions	143
Antioxidant activity results.....	143
PCA statistical analysis method.....	152
Cytotoxicity assay (MTT).....	154
Biological activity of isolated compounds.....	155
Conclusions.....	161
Reference.....	162

CHAPTER V

Anti-HIV-1 and antibacterial potential of <i>Hyptis radicans</i> (Pohl) Harley & J.F.B. Pastore and <i>Hyptis multibracteata</i> Benth. (Lamiaceae).....	173
Introduction.....	174
Material and methods.....	176
Plant Material.....	176
Extraction and Fractionation (plant material and sample preparation).....	176
Phenolic content.....	176
HIV-1 reverse transcriptase (RT) inhibitory bioassay.....	177
Antimicrobial assay.....	177
Statistical analysis.....	178
Results and discussions.....	178
Extraction yield.....	178
Phenolic content.....	179
Anti-HIV-1 capacity.....	181
Antibacterial potential.....	183
PCA analysis for antibacterial and anti-HIV activities.....	186
Conclusions.....	187
References.....	188

FINAL CONSIDERATIONS

195

CHAPTER I

Plant-derived drug discovery and special metabolism

Historical perspective

Throughout the history of the mankind, humans did depend on the nature in how to satisfy their basic needs. The first documented reports related to medicinal applications of plants dates back to 2,600 BCE and they report the existence of a sophisticated medical system in Mesopotamia, comprising about 1,000 plant-derived medicines. In these derivatives are included oils of *Cedrus* Trew species (cedar) and *Cupressus sempervirens* L. (cypress), *Glycyrrhiza glabra* L. (licorice), *Commiphora* Jacq. species (myrrh), and *Papaver somniferum* L. (poppy juice), all these which nowadays are still in use for the treatment of diseases who vary from coughs and colds to parasitic infections and inflammation. The Egyptian medicine dates from approximate 2,900 BCE, but the best-known record is the *Ebers Papyrus* dating from 1,500 BCE, which describes about 700 varieties of drugs, mostly originating of plants (Borchardt, 2002). The Chinese *Compendium of Materia Medica* has been extensively documented over the centuries (Huang, 1998), having the first report, known as the Wu Shi Er Bing Fang, dated from 1,100 BCE, and containing 52 prescriptions. Late, we have the, *Shennong Herbal* (~100 BCE), which contains 350 types of drugs, and the *Tang Herbal* written in 659 CE and containing 850 varieties of drugs. In the same way also exist documents where could be find the description of substances or plant-derived drugs in the Indian Ayurvedic system (dated from before 1,000 BCE). For example, the *Charaka Samhitas* and the *Sushruta Samhitas* report 341 and 516 drugs, respectively (Kapoor, 1990; Dev, 1999).

In Western world, the knowledge of the medicinal application of plants is mostly founded in two cultures: the Greek and Roman. In the first century the written documentation of the Greek physician Dioscorides was specifically important; for the Roman culture in the first century Pliny the Elder and Galen (2nd Century CE) were the two representatives in the area of medicinal plant descriptions (Sneader, 2005). Thanks to the Greek and Roman cultures, the Arabs, who already had medical experience, did preserve a great amount of knowledge during the Dark and Middle ages, in between the 5th and the 12th century and were improved with information received from traditional medicine from China and India.

Johannes Gutenberg assists in the revival of Greek and Roman knowledge in the 15th and 16th centuries with the invention of the printing press. As a result, several influential books on herbalism were compiled and widely distributed in Europe, for example *The Mainz Herbal* (1484) and *The German Herbal* (1485), both edited by Peter Schöffer, a Gutenberg's partner; the *Herbarium Vivaे Eicones* by Otto Brunfels (1530), the *Kreütter Buch* by Hieronymus Bock in 1546 (written in German) and *De Historia Stirpium* by Leonhart Fuchs that was published in Latin (1542) and also in German (1543) (Sneader, 2005).

During all this time, the use of medicinal plants was applied on empirical grounds, since there was no mechanistic knowledge of its pharmacological activities or active constituents.

It was in the 18th century that the foundations for rational clinical investigation of medicinal herbs were laid by researcher Anton von Störck, who studied poisonous herbs such as *Aconitum* and *Colchicum*, and William Whitering who studied foxglove (*Digitalis L.*) for the treatment of edema (Sneader, 2005).

Later, at the beginning of the 19th century, the German apothecary assistant Friedrich Sertürner succeeded in isolating morphine from opium, an analgesic and sleep inducing agent, which was named after the Greek God, Morpheus. This was the first isolated constituent of a plant and Sertürner published the method of isolation and crystallization, as well as the crystal structure and pharmacological properties of morphine (which he studied in stray dogs and self-experiments) (Sertürner, 1817).

As expected, thanks to this research, other similar studies on medicinal plants were originated in the following decades of the 19th century, and many bioactive natural products were isolated and identified, mainly alkaloids (quinine, caffeine, nicotine, codeine, atropine, colchicine, cocaine, capsaicin) (Felter and Lloyd, 1898; Hosztafi, 1997; Sneader, 2005; Kruse, 2007; Zenk and Juenger, 2007; Corson and Crews, 2007; Kaiser, 2008). Between 1981 and 2010, 1,073 new chemical entities were approved, of which only 36% were purely synthetic and more than 50% were derived or inspired by nature (Newman and Cragg, 2012). A substantial number of these compounds have been discovered in higher plants (Kinghorn et al., 2011).

At the moment we can find very remarkable examples related to the importance that have acquired the natural products and their importance for modern pharmacotherapy, specifically in the area of anticancer agents. An example of this is paclitaxel and its derivatives from yew (*Taxus L.*) species, vincristine and vinblastine from Madagascar

periwinkle (*Catharanthus roseus* (L.) G. Don), and camptothecin and their analogs originally discovered in the Chinese tree *Camptotheca acuminate* Decne (Cragg and Newman, 2013; Kinghorn et al., 2011).

Another notable example is galanthamine, which is a cholinesterase inhibitor and has been approved for the treatment of Alzheimer's disease, discovered in *Galanthus nivalis* L. (Mashkovsky and Kruglikova-Lvova, 1951 *apud* Atanasov et al., 2015), and the important antimalarial and potential anti-cancer agent artemisinin originally derived from the traditional Chinese herb *Artemisia annua* L. (Klayman et al., 1984).

Caventou and Pelletier were the first to report, in 1820, the isolation of the anti-malaria drug quinine from the bark of *Cinchona* L. species (e. g., *C. officinalis* L.) (Buss and Waigh, 1995). Quinine occurs naturally in the bark of *Cinchona* trees in South America and had long been used by indigenous groups in the Amazon for the treatment of fever. It was first introduced into Europe in the early 1600s for the treatment of malaria. Quinine formed the basis for the synthesis of the commonly used antimalarial drugs being one of the oldest malaria remedies known. Chloroquine and mefloquine replaced quinine in the mid-20th century, but with the emergence of resistance to both these drugs in many tropical regions, another plant long used in the treatment of fevers in Traditional Chinese Medicine, *A. annua*, gained prominence (Wongsrichanalai et al., 2002). As described, for millennia, medicinal plants have been an invaluable resource for therapeutic agents. Nowadays many therapeutic agents are botanical drugs or directly derived therefrom (Kinghorn et al., 2011).

Natural products as drug candidates: significance and advantages against synthetic compounds

There is a wealth of available and well-documented ethnopharmacological information on the traditional uses of natural drugs, which is a great advantage because it provides evidences for therapeutically effective compounds in humans (Heinrich and Gibbons, 2001; Corson and Crews, 2007; Heinrich, 2010; Kinghorn et al., 2011). According to the information above, 122 compounds derived from plants used worldwide as therapeutic agents were analyzed and it was revealed that 80% have an identical or related use to indications for which these pure compounds were prescribed in ethnomedicine (Farnsworth et al., 1985; Fabricant and Farnsworth, 2001).

In addition, it has been shown that natural products used for the development of medicines are highly likely to be used traditionally. An example is the discovery of the anti-

cancer agent taxol, from *Taxus brevifolia* Nutt., which discovery was done with a random screening approach. Later on, it came to light that the plant has been used by western Indian cultures as a medicine (Heinrich, 2010).

Because natural drugs are made by or in living organisms, these products possess properties that are evolutionarily optimized to serve in different biological functions, as they are part of the body's metabolism, for example binding to specific target proteins or other biomolecules (Hunter, 2008; Appendino et al., 2010).

Natural compounds are highly diverse and often provide highly specific biological activities. This stems from the proposition that essentially all natural products have some ability to bind to the receptor. The natural molecules, however, differ substantially from the synthetic ones. The main structural differences between natural and combinatorial compounds originate mainly from properties introduced to make combinatorial synthesis more efficient. These include the number of chiral centers, the prevalence of aromatic rings, the introduction of complex ring systems, and the degree of saturation of the molecule, as well as the number and proportions of different heteroatoms.

The chiral separation method is challenging and expensive. Therefore, the creation of molecules with a low number of chiral centers is favorable. Synthetic compounds tend to have a much smaller number of chiral centers, and in addition a lower molecular weight, a higher number of freely rotatable bonds, higher chain lengths, a lower number of rings, less oxygen but more nitrogen, sulfur and halogen atoms, a lower number of acceptors and donors of Lipinski-type H-bonds and higher calculated octanol-water partition coefficients (cLogP values). Other differences are the complexity of ring systems and the degree of saturation (Stahura et al., 2000; Feher and Schmidt, 2003; Atasanov et al., 2015).

For example, because of the stereospecificity of most biological targets, it is likely that many non-stereospecific synthetic analogues, created, for example, by the introduction of aromatic rings, represent non-optimal compromises, especially in terms of selectivity and this occurs more frequently in the case of combinatorial synthesis compounds. The greater flexibility of combinatorial products is likely to have entropic consequences detrimental to the binding of these compounds. It may also affect negatively their ability to induce conformational changes in the receptor required for biological function. Also, the production process of synthetic analogs radically alters the number and ratios of different types of atoms, such as nitrogen, oxygen, sulfides and halogens. These distributions in turn

have a direct impact on the donor and acceptor patterns available to complement the receptor surface properties (Feher and Schmidt, 2003).

These factors, which are structural differences, specifically the significant number of chiral centers, low size and high flexibility, make the synthetic products weaker and less specific than natural products (Feher and Schmidt, 2003). Natural products have selective and specific biological actions due to the binding affinities to relevant proteins in their biological functions, and during biosynthesis a greater diversity and chemical complexity are developed than for their synthetic analogues (Clardy and Walsh, 2004; Koehn and Carter, 2005). They often have less advantageous absorption, distribution, metabolism, excretion and toxicity properties. In view of these facts, it is interesting to consider that the search for the replacement of natural compounds with synthetic ones is usually based on exactly these kinds of ‘unfavorable’ modifications.

The main focus of the pharmaceutical industry was for a time led to synthetic compound libraries and high throughput screening, with the aim of discovering new drug derivatives (Beutler, 2009; David et al., 2015). But, the results obtained did not meet expectations, and this is evident when the decreasing number of drugs that reach the market is observed (David et al., 2015). Because of this, the interest in products based on natural products has been revitalized for the discovery of new drugs, where broad interdisciplinary research approaches are required due to their high complexity, but at the same time high specificity as mentioned in previous paragraphs (Heinrich, 2010a).

Plants have been the basis for medical treatments through much of the human history. Nowadays, researchers are increasingly interested in medicinal plants as alternative medicine, due to their good pharmacological properties, fewer side effects, and low cost (Sayah et al., 2017).

Where do these medicinal substances originate in plants?

What is a special (or secondary) metabolite and which is their role in plants?

Land plants have colonized the vast majority of the Earth's surface due to rich levels of specialization and intricate relationships with other organisms. During this process land plants had (and still have) to face a number of challenges imposed by the terrestrial environment. These organisms are autotrophic stationary, dealing with biotic and abiotic stress factors such as the coexistence of herbivores and pathogens in their immediate environment, pollination and seed dispersal (specially angiosperms), and climate variations.

Therefore, and because of these challenges, land plants have developed special biochemical pathways that allow them to synthesize a series of chemicals, also called secondary metabolites or special metabolites, that are produced regularly in response to specific environmental stimuli, such as herbivore induced-damage, pathogens attack, enhanced concentration of air pollutants etc. (Reymond et al., 2000; Hermsmeier et al., 2001).

The function of these special metabolites is to increase the general plant ability plant to survive and overcome local challenges, allowing them to interact with their environment. They play no role in primary metabolic needs and may be unique to specific species or genera (Harborne, 2014). The energy invested in the synthesis of these special metabolites, which is usually much higher than that required to synthesize primary metabolites, is an indicator of the importance of these substances for the survival of plants (Gershenson, 2017; Hong et al., 2016). Among the functions of the special metabolites are: protective roles as antioxidant, free radical-scavenging, UV light-absorbing, and defend the plant against microorganisms such as bacteria, fungi, and virus. They also manage inter-plant relationships, acting as allelopathic defenders of the plant's growing space against competitor plants. More complex roles include dictating or modifying the plant's relationship with more complex organisms (Harborne, 2014; Wink, 2003; Tahara, 2007).

One of the main roles of special metabolism is feeding deterrence. For that reason, many of these substances are bitter and/or toxic to potential herbivores, affecting the central and peripheral nervous system of the herbivore. In this regard, special metabolites often act as agonists or antagonists of neurotransmitter systems or form structural analogs of endogenous hormones (Wink, 2000; Miller and Heyland, 2010; Rattan, 2010). In addition to that defense mechanism, plants also have to foster a number of symbiotic relationships. One of the obvious roles in this series of mechanisms is the attraction of pollinators and other symbionts, using colors and scents or indirect defenses, by attracting natural enemies of its herbivorous attackers. In this way, it provides an attractive chemical environment for the predator or alternatively, it may be a direct response to tissue damage by the herbivore, resulting in the synthesis and release of a set of substances that are attractive to natural herbivore predators (Harborne, 2014; Wink, 2003; Tahara, 2007). There are more than 100,000 special metabolites already described in plants, ranging from simple alkaloids (structurally) to phytosterols and more complex polyphenolic molecules (Dillard and German, 2000).

Special metabolism and major groups of plant special metabolites

All living cells possess similar pathways for the synthesis of components such as sugars, amino acids, nitrogenous bases, carbohydrates, proteins, and nucleotides, being these are molecules essential for energy production and cell constitution and plant development. Plant special metabolites are derived from the products of primary metabolism but have a much more limited taxonomic distribution. They can be broadly classified according to their structure and biosynthetic pathways; however, it should be appreciated that many special metabolites are also derived by combining elements of all these biosynthetic pathways (Fig. 1.1).

These diversified compounds can be divided into three main categories: terpenes, nitrogenous compounds and phenolic compounds, based on their chemical structure. Amines, cyanogenic glycosides, glucosinolates, acetylenes and psoralens, are other minor groups that cannot be included in these three large groups (Fang et al., 2011 *apud* Russell and Duthie, 2011).

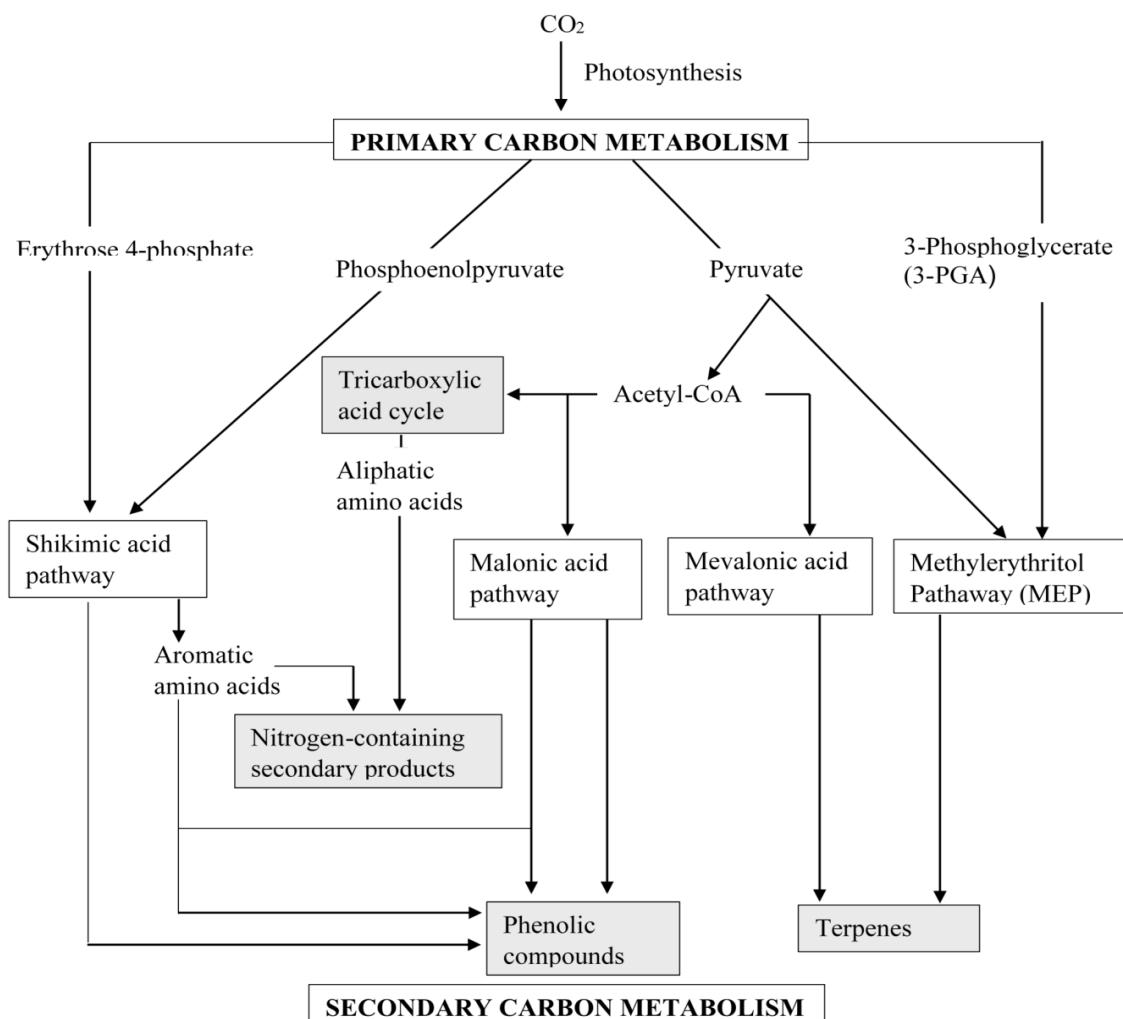


Figure 1.1. A simple schematic representation of the major secondary metabolites in plants (Ncube and Van Staden, 2015)

Terpenes

Terpenes represent the most abundant and structurally diverse group of plant special metabolites, in which more than 36,000 structures have been identified. They are a structurally diverse group of hydrocarbons derived from the five-carbon precursors: isopentyl diphosphate (IPP) or dimethylallyl diphosphate (DMPP), synthesized, in photosynthetic organisms, by mevalonate and methylerythritol phosphate pathways.

Terpenes are classified according to the degree of isoprene incorporation as follow: hemiterpenes (C_5), monoterpenes (C_5)₂, sesquiterpenes (C_5)₃, diterpenes (C_5)₄, sesterpenes (C_5)₅, triterpenes (C_5)₆, tetraterpenes (or carotenoids) (C_5)₈, and through to higher polymers such as rubber (C_5)_{>100}. Isoprene units are often joined in a head-to-tail and head-to-head linkage and a few terpene structures are formed by irregular head-to-middle linkage. After the basic terpene skeletons are formed, subsequent modifications occur which give rise to different structures such as steroids like cholesterol, ergosterol, sitosterol and stigmasterol, which are synthesized from a triterpene precursor. Among the modifications that the basic terpene skeleton receives are: reduction, isomerization, oxidation, conjugation and degradation (Grayson, 2000; Croteau et al., 2000; Maimone and Baran, 2007).

Pharmacological active molecules derived from terpenes include, for example, the herbal tranquilizer valtrate, the major component of valerian (*Valeriana officinalis* L.) and the anti-cancer drug taxol, extracted originally from the Pacific Yew (*Taxus brevifolia*) (Hayes et al., 2008).

Nitrogenous compounds: alkaloids

Alkaloids are a group of alkaline, low molecular weight and nitrogen containing compounds. They are the most widely distributed nitrogenous special metabolites and are found not only in plants, but also in microorganism, playing an important role in plant defense systems. Alkaloid containing plants were, for mankind, the original "materia medica" and many are still in use today as prescription drugs, such as vinblastine, quinine, atropine, and camptothecin. There are more than 12,000 alkaloids reported for 100 families of plants, being especially abundant in Fabaceae, Solanaceae, Menispermaceae, Papaveraceae, Ranunculaceae, Apocynaceae and Berberidaceae. Can be classified on the basis of the plants from which they were isolated, their chemical structures, and the biosynthetic origins. This last feature has an obvious advantage of reflecting the relationship between biosynthetic pathways and the chemical structures. Alkaloids could thus be further

classified into three groups according to their biosynthesis origin: true alkaloid, protoalkaloid, and pseudoalkaloid (Buchanan et al., 2000; Dewick, 2009).

The alkaloids, have contributed mainly providing neurotoxins, poisons and traditional psychedelics, among which are some that come from *Atropa belladonna* L. like, atropine, scopolamine, and hyoscyamine, to this chemical group also belong the most consumed social drugs, nicotine, caffeine, methamphetamine (ephedrine), cocaine and opiates (Zenk and Juenger, 2007). This group also provides the cholinesterase inhibiting treatments routinely prescribed for the cholinergic deregulation of Alzheimer's disease, such as galantamine, huperzine, physostigmine, and rivastigmine (Mukherjee et al., 2007).

Phenolic compounds

Phenolic compounds are ubiquitously found across plant, with ~10,000 structures identified. Structurally, they share at least 1 aromatic hydrocarbon ring with 1 or more hydroxyl groups attached and are synthesized via the shikimate pathway alone or in combination with the acetate-malonate pathway. The simplest compound with this structural motif is the phenol molecule, which itself does not occur in plants. Phenolic compounds range from simple low-molecular mass, such as the simple phenylpropanoids, coumarins, and benzoic acid derivatives, to more complex structures such as flavonoids, stilbenes, and tannins. Flavonoids represent the largest, most diverse group, encompassing some 6,000 compounds, all of which share a common underlying structure of two 6-carbon rings, with a 3-carbon bridge, which usually forms a 3rd ring. Flavonoids can then be subdivided according to modifications of this basic skeleton into: chalcones, flavones, flavonols, flavanones, isoflavones, flavan-3-ols, and anthocyanins (Bowsher and Tobin, 2008; Yang et al., 2012).

Phenolic compounds and flavonoids in particular, are ubiquitous in plants and therefore represent an important component of a normal human diet. Epidemiological studies have suggested associations between consumption of phenolic-rich foods or beverages and various diseases, such as stroke, cardiovascular disease, and cancer (Steffen, 2006) and neurologic disorders such as dementia/Alzheimer's disease (Commenges et al., 2000; Vingtdeux et al., 2008).

Aims of the present study

In the world there are a great number of plant species, which produce a diversity of bioactive compounds with different chemical scaffolds. According to previous estimates, only 6% of existing plant species have been systematically pharmacologically investigated, and only about 15% were studied phytochemically (Fabricant and Farnsworth, 2001; Verpoorte, 1998 and 2000). Although today the percentage of species is better characterized by increased interest in this phytochemical area, it is still conceivable that there are a large number of plant compounds that are not well pharmacologically researched, especially if we consider the approximately 310,000 plant species described (IUCN, 2015). Unfortunately, a significant decline in global plant species is expected in the coming years as a result of climate change and anthropogenic factors that jeopardize these potential sources of new natural drugs, and therefore urgent measures are needed to access different species (Maclean and Wilson, 2011; Thomas et al., 2004).

Another point to highlight is the ethnobotanical knowledge about the traditional pharmacological use that is disappearing. With the increase of globalization, this information is in danger of being lost forever and it is being lost faster than the loss of the biodiversity (Appendino et al., 2010).

In the context of the discovery of drugs of plant origin, it is highly advantageous when the species under study come from regions of high biodiversity and endemism, as the chemical diversity of natural products can reflect the biodiversity of their organisms of origin and an example of megadiverse country is Brazil (Barbosa et al., 2012; Henrich and Beutler, 2013). These estimates gain importance when considering the broad potential of the active principles contained in nature and which have not yet been identified and evaluated in a medical context. This fact attracts the attention of the pharmaceutical industry that sees in plant diversity a feasible source for new medicines.

The present study aimed to access phytochemically to Brazilian native species: *Hyptis radicans* Jacq. and *Hyptis multibracteata* Poit. Both are species of occurrence in the Atlantic Rainforest of State of São Paulo and are easily found in the Paranapiacaba region. Furthermore, both are species that do not count on studies on their chemical composition, have not been evaluated for their biological activities and, therefore, are promising models for prospecting studies of natural bioactive substances.

The main objectives of the present study were:

- To isolate and identify substances present in both species; and
- To evaluate the antioxidant, antibacterial and anti-HIV potential of *H. multibracteata* and *H. radicans*;

This study is divided in 5 chapters and a final consideration as follow:

Chapter 1: Plant-derived drug discovery and special metabolism.

Chapter 2: *Hyptis* Jacq.: a general chemical profile review - *manuscript submitted to Chemistry & Biodiversity*.

Chapter 3: Botanical aspects & chemical description of *H. radicans* and *H. multibracteata*.

Chapter 4: Antioxidant, anti-acetyl cholinesterase and cytotoxic potential of *Hyptis* spp – *part of the results published at Industrial Crops & Products 112 (2018) 705–715*.

Chapter 5: Anti-HIV-1 and antibacterial potential of *Hyptis radicans* (Pohl) Harley & J.F.B. Pastore and *Hyptis multibracteata* Benth. (Lamiaceae) – *manuscript submitted to Journal of Herbal Medicine*.

References

- Abedini, A., Roumy, V., Mahieux, S., Biabiany, M., Standaert-Vitse, A., Rivière, C., Sahpaz, S., Bailleul, F., Neut, C. & Hennebelle, T. (2013). Rosmarinic acid and its methyl ester as antimicrobial components of the hydromethanolic extract of *Hyptis atrorubens* Poit. (Lamiaceae). *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Abedini, A., Roumy, V., Neut, C., Biabiany, M., Joseph, H., Sahpaz, S., Hennebelle, T. & Hennebelle, T. (2012). Antibacterial activity of the methanolic extract of *Hyptis atrorubens* (Lamiaceae). *Planta Medica*, 78(11), PD135.
- Adjanohoun, E., Ahyi, M. R. A., Aké, A. L., Akpagana, K., Chibon, P., El- Hadj, A., Eymen, I., Goutote, E., Ginko, S., Hodouto, KK., Hougnon, P., Keita, A., Kéoula, Y., Klouga-Ocloo, W. P., Lo, I., Siamevi, K., Taffame, K. K., Garba, M., Gassita, J. N. & Gbeassor, M. (1986). Médecine traditionnelle et Pharmacopée: Contribution aux études ethnobotaniques et floristiques du Togo. ACCT, Paris, p. 671.
- Adjimani, J. P., & Asare, P. (2015). Antioxidant and free radical scavenging activity of iron chelators. *Toxicology reports*, 2, 721-728.
- Adomako-Bonsu, A. G., Chan, S. L., Pratten, M., & Fry, J. R. (2017). Antioxidant activity of rosmarinic acid and its principal metabolites in chemical and cellular systems: Importance of physico-chemical characteristics. *Toxicology In Vitro*, 40, 248-255.
- Agra, M. D. F., Silva, K. N., Basílio, I. J. L. D., Freitas, P. F. D., & Barbosa-Filho, J. M. (2008). Survey of medicinal plants used in the region Northeast of Brazil. *Revista brasileira de farmacognosia*, 18(3), 472-508.
- Ahmad, N., Feyes, D. K., Agarwal, R., Mukhtar, H., & Nieminen, A. L. (1997). Green tea constituent epigallocatechin-3-gallate and induction of apoptosis and cell cycle arrest in human carcinoma cells. *Journal of the National Cancer Institute*, 89(24), 1881-1886.
- Ait-Ouazzou, A., Lorán, S., Arakrak, A., Laglaoui, A., Rota, C., Herrera, A., Pagán, R., Conchello, P. (2012). Evaluation of the chemical composition and antimicrobial activity of *Mentha pulegium*, *Juniperus phoenicea*, and *Cyperus longus* essential oils from Morocco. *Food Research International*, 45(1), 313-319.
- Ali, S. S., Kasoju, N., Luthra, A., Singh, A., Sharanabasava, H., Sahu, A., & Bora, U. (2008). Indian medicinal herbs as sources of antioxidants. *Food Research International*, 41(1), 1-15.
- Alimi, H., Hfaiedh, N., Bouoni, Z., Sakly, M., & Rhouma, K. B. (2011). Evaluation of antioxidant and antiulcerogenic activities of *Opuntia ficus indica* f. *inermis* flowers extract in rats. *Environmental Toxicology and Pharmacology*, 32(3), 406-416.
- Almtorp, G. T., Hazell, A. C., & Torssell, K. B. (1991). A lignan and pyrone and other constituents from *Hyptis capitata*. *Phytochemistry*, 30(8), 2753-2756.
- Al-Sereiti, M. R., Abu-Amer, K. M., & Sena, P. (1999). Pharmacology of rosemary (*Rosmarinus officinalis* Linn.) and its therapeutic potentials.

- Alvarez, F., Tello, E., Bauer, K., E Diaz, L., Rodriguez, J., & Jimenez, C. (2015). Cytotoxic and antimicrobial diterpenes isolated from *Hyptis dilatata*. *Current Bioactive Compounds*, 11(3), 189-197.
- Andrade-Wartha, E. R. S. (2007). Capacidade antioxidante in vitro do pedúnculo de caju (*Anacardium Occidentale* L.) e efeito sobre as enzimas participantes do sistema antioxidante de defesa do organismo animal. *São Paulo*.
- Andrae-Marobela, K., Ghislain, F. W., Okatch, H., & Majinda, R. R. (2013). Polyphenols: a diverse class of multi-target anti-HIV-1 agents. *Current drug metabolism*, 14(4), 392-413.
- Appendino, G., Fontana, G., & Pollastro, F. (2010). 3.08—Natural products drug discovery. *Comprehensive Natural Products II*. Elsevier, Oxford, 205.
- Araújo, E. C., Lima, M. A. S., Nunes, E. P., & Silveira, E. R. (2005). Abietane diterpenes from *Hyptis platanifolia*. *Journal of the Brazilian Chemical Society*, 16(6B), 1336-1341.
- Arullappan, S., Rajamanickam, P., Thevar, N. & Kodimani, C. C. (2014). In vitro screening of cytotoxic, antimicrobial and antioxidant activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. *Tropical Journal of Pharmaceutical Research*, 13(9), 1455-1461.
- Aruoma, O. I., Grootveld, M., & Halliwell, B. (1987). The role of iron in ascorbate-dependent deoxyribose degradation. Evidence consistent with a site-specific hydroxyl radical generation caused by iron ions bound to the deoxyribose molecule. *Journal of Inorganic Biochemistry*, 29(4), 289-299.
- Atanasov, A. G., Waltenberger, B., Pferschy-Wenzig, E. M., Linder, T., Wawrosch, C., Uhrin, P., Temml, V., Wang, L., Schwaiger, S., Heiss, E. H., Rollinger, J. M., Schuster, D., Breuss, J. M., Bochkov, V., Mihovilovic, M. D., Kopp, B., Bauer, R., Dirsch, V. M. & Stuppner, H. (2015). Discovery and resupply of pharmacologically active plant-derived natural products: a review. *Biotechnology advances*, 33(8), 1582-1614.
- Atoui, A. K., Mansouri, A., Boskou, G. & Kefalas, P. (2005). Tea and herbal infusions: their antioxidant activity and phenolic profile. *Food chemistry*, 89(1), 27-36.
- Awaad, A. S., Al-Jaber, N. A., Moses, J. E., El-Meligy, R. M., & Zain, M. E. (2013). Antiulcerogenic activities of the extracts and isolated flavonoids of *Euphorbia cuneata* Vahl. *Phytotherapy Research*, 27(1), 126-130.
- Bae, G. U., Seo, D. W., Kwon, H. K., Lee, H. Y., Hong, S., Lee, Z. W., Ha, K. S., Lee, H. W. & Han, J. W. (1999). Hydrogen peroxide activates p70S6k signaling pathway. *Journal of Biological Chemistry*, 274(46), 32596-32602.
- Baghel, S. S., Shrivastava, N., Baghel, R. S., Agrawal, P., & Rajput, S. (2012). A review of quercetin: antioxidant and anticancer properties. *World J Pharm Pharmaceutical Sci*, 1(1), 146-60.
- Bakkali, F., Averbeck, S., Averbeck, D., & Idaomar, M. (2008). Biological effects of essential oils—a review. *Food and chemical toxicology*, 46(2), 446-475.

- Balouiri, M., Sadiki, M., & Ibsouda, S. K. (2016). Methods for in vitro evaluating antimicrobial activity: A review. *Journal of Pharmaceutical Analysis*, 6(2), 71-79.
- Banjarnahor, S. D., & Artanti, N. (2014). Antioxidant properties of flavonoids. *Medical Journal of Indonesia*, 23(4), 239.
- Baptistella, L. H. B., Imamura, P. M., Melo, L. V. D., & Castello, C. (2009). Preparação do (+)- \pm -terpineol a partir do (+)-limoneno: monoterpenos de odor agradável em um projeto para química orgânica experimental. *Química Nova*.
- Barbosa, W. L. R., do Nascimento, M. S., do Nascimento Pinto, L., Maia, F. L. C., Sousa, A. J. A., Júnior, J. O. C. S., Monteiro, M. M. & de Oliveira, D. R. (2012). Selecting medicinal plants for development of phytomedicine and use in primary health care. In *Bioactive Compounds in Phytomedicine*. InTech.
- Barros, L., Carvalho, A. M., & Ferreira, I. C. (2011). From famine plants to tasty and fragrant spices: Three Lamiaceae of general dietary relevance in traditional cuisine of Trás-os-Montes (Portugal). *LWT-Food Science and Technology*, 44(2), 543-548.
- Barth, S. A., Iselmann, G., Engemann, R., & Heidemann, H. T. (1991). Influences of *Ginkgo biloba* on cyclosporin A induced lipid peroxidation in human liver microsomes in comparison to vitamin E, glutathione and N-acetylcysteine. *Biochemical Pharmacology*, 41(10), 1521-1526.
- Başer, K. H. C., Demirci, B., & Dadandi, M. Y. (2008). Comparative essential oil composition of the natural hybrid *Phlomis* x *vuralii* Dadandi (Lamiaceae) and its parents. *Journal of Essential Oil Research*, 20(1), 57-62.
- Basílio, I. J. L. D., Agra, M. F., Rocha, E. A., Leal, C. K. A., & Abrantes, H. F. (2007). Estudo farmacobotânico comparativo das folhas de *Hyptis pectinata* (L.) Poit. e *Hyptis suaveolens* (L.) Poit (Lamiaceae). *acta farmacéutica bonaerense*, 25(4), 518.
- Batista, F. L., de Paula, J. R., Silva, J. G., Santos, S. C., Ferri, P. H., & Ferreira, H. D. (2003). Essential Oils of *Hyptidendron canum* (Pohl ex Benth.) R. Harley and *Hyptis velutina* Pohl ex Benth. from Brazilian Cerrado. *Journal of Essential Oil Research*, 15(2), 88-89.
- Baxter, J. W. (1961). North American Species of *Puccinia* on *Hyptis*. *Mycologia*, 53(1), 17-24.
- Beutler, J. A. (2009). Natural products as a foundation for drug discovery. *Current protocols in pharmacology*, 9-11.
- Bezerra, J. W. A., Costa, A. R., da Silva, M. A. P., Rocha, M. I., Boligon, A. A., da Rocha, J. B. T., Barros, L. M. & Kamdem, J. P. (2017). Chemical composition and toxicological evaluation of *Hyptis suaveolens* (L.) Poiteau (Lamiaceae) in *Drosophila melanogaster* and *Artemia salina*. *South African Journal of Botany*, 113, 437-442.
- BFG. Growing knowledge: an overview of Seed Plant diversity in Brazil. 2015. Rodriguésia 66(4), 1085-1113.
- Bhuiyan, M. N. I., & Begum, J. (2010). Chemical component studies on the leaf and inflorescence essential oil of *Hyptis brevipes* (Poit.). *Journal of Medicinal Plants Research*, 4(20), 2128-2131.

- Biggs, D. A., Porter, R. B., Reynolds, W. F., & Williams, L. A. (2008). A New Hyptadienic Acid Derivative from *Hyptis verticillata* (Jacq.) with Insecticidal Activity. *Natural Product Communications*, 3(11), 1759-1762.
- Biradi, M., & Hullatti, K. (2017). Bioactivity guided isolation of cytotoxic terpenoids and steroids from *Premna serratifolia*. *Pharmaceutical biology*, 55(1), 1375-1379.
- Birben, E., Sahiner, U. M., Sackesen, C., Erzurum, S., & Kalayci, O. (2012). Oxidative stress and antioxidant defense. *World Allergy Organization Journal*, 5(1), 9.
- Birkett, M. A., Bruce, T. J., & Pickett, J. A. (2010). Repellent activity of *Nepeta grandiflora* and *Nepeta clarkei* (Lamiaceae) against the cereal aphid, *Sitobion avenae* (Homoptera: Aphididae). *Phytochemistry Letters*, 3(3), 139-142.
- Boechat, C. L., Pistóia, V. C., Gianelo, C., & de Oliveira Camargo, F. A. (2016). Accumulation and translocation of heavy metal by spontaneous plants growing on multi-metal-contaminated site in the Southeast of Rio Grande do Sul state, Brazil. *Environmental Science and Pollution Research*, 23(3), 2371-2380.
- Bogninou-Agbidinoukoun, G.S.; Yedomonhan, H.; Avlessi, F.; Sohouunhloué, D.; Chalard, P.; Chalchat, J-C.; Delort, L.; Billard, H.; Caldefie-Chézet, F. & Figueredo, G. (2012). Volatile oil composition and antiproliferative activity of *Hyptis spicigera* Lam against human breast adenocarcinoma cells MCF-7. *Research Journal of Chemical Sciences*, Vol. 3(1), 27-31.
- Borchardt, J. K. (2002). The beginnings of drug therapy: Ancient mesopotamian medicine. *Drug News Perspect*, 15(3), 187-192.
- Borges Bubols, G., da Rocha Vianna, D., Medina-Remon, A., von Poser, G., Maria Lamuela-Raventos, R., Lucia Eifler-Lima, V., & Cristina Garcia, S. (2013). The antioxidant activity of coumarins and flavonoids. *Mini reviews in medicinal chemistry*, 13(3), 318-334.
- Borg-Karlson, A. K., Norin, T., & Talvitie, A. (1981). Configurations and conformations of torreyol (δ -cadinol), α -cadinol, T-muurolol and T-cadinol. *Tetrahedron*, 37(2), 425-430.
- Botrel, P. P., Pinto, J. E. B. P., Araújo, A. C. C. D., Bertolucci, S. K. V., Figueiredo, F. C., Ferri, P. H., & Costa, D. P. D. (2010). Variation in the content and volatile composition of *Hyptis marruboides* EPL: cultivated in field and greenhouse. *Química Nova*, 33(1), 33-37. (B)
- Botrel, P. P., Pinto, J. E. B. P., Ferraz, V., Bertolucci, S. K. V., & Figueiredo, F. C. (2010). Content and chemical composition of *Hyptis marruboides* essential oil in function of seasons. *Acta Scientiarum. Agronomy*, 32(3), 533-538. (A)
- Botrel, P. P., Pinto, J. E. B. P., Figueiredo, F. C., Bertolucci, S. K. V., & Ferri, P. H. (2009). Essential oil content and chemical composition in *Hyptis marruboides* Epl. of different genotypes. *Revista Brasileira de Plantas Medicinais*, 11(2), 164-169.
- Botrel, P. P., Rodrigues, M. A., Bertolucci, S. V., Lima, A. F., Alvarenga, I. C. A., & Pinto, J. P. (2013). Factors affecting in vitro propagation and chromatographic analysis of compounds in *Hyptis marruboides* epl., a threatened medicinal plant. In VIII

- International Symposium on In Vitro Culture and Horticultural Breeding 1083* (pp. 319-325).
- Boulila, A., Sanaa, A., Salem, I. B., Rokbeni, N., M'rabet, Y., Hosni, K., & Fernandez, X. (2015). Antioxidant properties and phenolic variation in wild populations of *Marrubium vulgare* L. (Lamiaceae). *Industrial Crops and Products*, 76, 616-622.
- Bowsher, C., Steer, M., & Tobin, A. (2008). *Plant biochemistry*. Garland Science.
- Bozin, B., Mimica-Dukic, N., Simin, N., & Anackov, G. (2006). Characterization of the volatile composition of essential oils of some Lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *Journal of Agricultural and Food Chemistry*, 54(5), 1822-1828.
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. L. W. T. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT-Food Science and Technology*, 28(1), 25-30.
- Buchanan, B. B., Gruissem, W., & Jones, R. L. (2000). *Biochemistry & molecular biology of plants* (Vol. 40). Rockville, MD: American Society of Plant Physiologists.
- Buettner, G. R. (1988). In the absence of catalytic metals ascorbate does not autoxidize at pH 7: ascorbate as a test for catalytic metals. *Journal of biochemical and biophysical methods*, 16(1), 27-40.
- Burkill, H. M. (1994). *The useful plants of west tropical Africa. Volume 2: Families EI* (No. Edn 2). Royal Botanic Gardens.
- Buss, A. D., Cox, B., & Waigh, R. D. (1995). Natural products as leads for new pharmaceuticals. *Burger's medicinal chemistry and drug discovery*.
- Butler, M. S., & Cooper, M. A. (2011). Antibiotics in the clinical pipeline in 2011. *The Journal of antibiotics*, 64(6), 413.
- Caldas, G. F. R., do Amaral Costa, I. M., da Silva, J. B. R., da Nóbrega, R. F., Rodrigues, F. F. G., da Costa, J. G. M., & Wanderley, A. G. (2011). Antiulcerogenic activity of the essential oil of *Hyptis martiusii* Benth. (Lamiaceae). *Journal of ethnopharmacology*, 137(1), 886-892.
- Carneiro, V. A., Santos, H. S. D., Arruda, F. V. S., Bandeira, P. N., Albuquerque, M. R. J. R., Pereira, M. O., Henriques, M., Cavada, B. S. & Teixeira, E. H. (2010). Casbane diterpene as a promising natural antimicrobial agent against biofilm-associated infections. *Molecules*, 16(1), 190-201.
- Castelluccio, C., Paganga, G., Melikian, N., Paul Bolwell, G., Pridham, J., Sampson, J., & Rice-Evans, C. (1995). Antioxidant potential of intermediates in phenylpropanoid metabolism in higher plants. *Febs Letters*, 368(1), 188-192.
- Castrillo, M., Vizcaíno, D., Moreno, E., & Latorraca, Z. (2001). Chlorophyll content in some cultivated and wild species of the family Lamiaceae. *Biologia plantarum*, 44(3), 423-425.

- Castrillo, M., Vizcaíno, D., Moreno, E., & Latorraca, Z. (2005). Specific leaf mass, fresh: dry weight ratio, sugar and protein contents in species of Lamiaceae from different light environments. *Revista de biología tropical*, 53(1-2), 23-28.
- Cayman chemical. (2016). Linolenic Acid methyl Ester. Available at: <https://www.caymanchem.com/product/9000290> accesed: 19th May 2018.
- Chan, K. W. K., & Ho, W. S. (2015). Anti-oxidative and hepatoprotective effects of lithospermic acid against carbon tetrachloride-induced liver oxidative damage in vitro and in vivo. *Oncology reports*, 34(2), 673-680.
- Chang, Y. T., Chang, W. N., Tsai, N. W., Huang, C. C., Kung, C. T., Su, Y. J., Lin, W. C., Cheng, B. C., Su, C. M., Chiang, Y. F. & Lu, C. H. (2014). The roles of biomarkers of oxidative stress and antioxidant in Alzheimer's disease: a systematic review. *BioMed Research International*, 2014:182303.
- Charles, D. J. (2012). *Antioxidant properties of spices, herbs and other sources*. Springer Science & Business Media.
- Chen, J. H., & Ho, C. T. (1997). Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds. *Journal of agricultural and food chemistry*, 45(7), 2374-2378.
- Cheng, J. C., Dai, F., Zhou, B., Yang, L., & Liu, Z. L. (2007). Antioxidant activity of hydroxycinnamic acid derivatives in human low density lipoprotein: mechanism and structure-activity relationship. *Food Chemistry*, 104(1), 132-139.
- Chobot, V. (2010). Simultaneous detection of pro-and antioxidative effects in the variants of the deoxyribose degradation assay. *Journal of agricultural and food chemistry*, 58(4), 2088-2094.
- Clardy, J., & Walsh, C. (2004). Lessons from natural molecules. *Nature*, 432(7019), 829.
- Clarkson, P. M., & Thompson, H. S. (2000). Antioxidants: what role do they play in physical activity and health?. *The American Journal of Clinical Nutrition*, 72(2), 637s-646s.
- Clifford, M. N., Johnston, K. L., Knight, S., & Kuhnert, N. (2003). Hierarchical scheme for LC-MS n identification of chlorogenic acids. *Journal of agricultural and food chemistry*, 51(10), 2900-2911.
- Commenges, D., Scotet, V., Renaud, S., Jacqmin-Gadda, H., Barberger-Gateau, P., & Dartigues, J. F. (2000). Intake of flavonoids and risk of dementia. *European journal of epidemiology*, 16(4), 357-363.
- Correa, M. P. (1931). *Dicionário das plantas úteis do Brasil e das exóticas cultivadas* (No. 581.981 C6).
- Corson, T. W., & Crews, C. M. (2007). Molecular understanding and modern application of traditional medicines: triumphs and trials. *Cell*, 130(5), 769-774.
- Costa, A. C., Rosa, M., Meguer, C. A., Silva, F. G., Pereira, F. D., & Otoni, W. C. (2014). A reliable methodology for assessing the *in vitro* photosynthetic competence of two Brazilian savanna species: *Hyptis marruboides* and *Hancornia speciosa*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 117(3), 443-454.

- Costa, H. N. R. D., Santos, M. C. D., Alcântara, A. F. D. C., Silva, M. C., França, R. C., & Piló-Veloso, D. (2008). Chemical constituents and antiedematogenic activity of *Peltodon radicans* (Lamiaceae). *Química Nova*, 31(4), 744-750.
- Coutinho, H. D., Costa, J. G., Siqueira-Júnior, J. P., & Lima, E. O. (2008). *In vitro* anti-staphylococcal activity of *Hyptis martiusii* Benth against methicillin-resistant *Staphylococcus aureus*: MRSA strains. *Revista Brasileira de Farmacognosia*, 18, 670-675.
- Cragg, G. M., & Newman, D. J. (2013). Natural products: a continuing source of novel drug leads. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1830(6), 3670-3695.
- Croteau, R., Kutchan, T. M., & Lewis, N. G. (2000). Natural products (secondary metabolites). *Biochemistry and molecular biology of plants*, 24, 1250-1319.
- Cudd, T., T. A. Estrada Mendoza, P. Sconcert-Hall, A. D. Richard, C. Richmond, J. P. Dupre, S. M. McMicken W. H. Dees, and O. E. Christian. McSU. J. Hightower. CPMC. (2014). Antibacterial and mosquitocidal activity of native Louisiana plant species. *The Louisiana Academy of Sciences* (Abstract), 4(1), 37.
- Cui, H. X., Tang, L., Cheng, F. R., & Yuan, K. (2017). Antitumor effects of ethanol extracts from *Hyptis rhomboidea* in H22 tumor-bearing mice. *Pharmacognosy magazine*, 13(52), 571.
- Cunha, A. P. D., Roque, O. R., & da Silva, A. P. (2006). *Plantas e produtos vegetais em fitoterapia*.
- Da Silva, A. C., de Souza, P. E., Pinto, J. E. B. P., da Silva, B. M., Amaral, D. C., & de Arruda Carvalho, E. (2012). Essential oils for preventative treatment and control of Asian soybean rust. *European journal of plant pathology*, 134(4), 865-871.
- David, B., Wolfender, J. L., & Dias, D. A. (2015). The pharmaceutical industry and natural products: historical status and new trends. *Phytochemistry reviews*, 14(2), 299-315.
- Davies, J., & Davies, D. (2010). Origins and evolution of antibiotic resistance. *Microbiology and molecular biology reviews*, 74(3), 417-433.
- de Lira Mota, K. S., Dias, G. E. N., Pinto, M. E. F., Luiz-Ferreira, Â., Monteiro Souza-Brito, A. R., Hiruma-Lima, C. A., Barbosa-Filho, J. M. & Batista, L. M. (2009). Flavonoids with gastroprotective activity. *Molecules*, 14(3), 979-1012.
- de Rodríguez, D. J., Salas-Méndez, E. D. J., Rodríguez-García, R., Hernández-Castillo, F. D., Díaz-Jiménez, M. L. V., Sáenz-Galindo, A., González-Morales, S., Flores-López, M.L., Villarreal-Quintanilla, J. A., Peña-Ramos, F. M. & Carrillo-Lomelí, D. A. (2017). Antifungal activity in vitro of ethanol and aqueous extracts of leaves and branches of *Flourensia* spp. against postharvest fungi. *Industrial Crops and Products*, 107, 499-508.
- de Siqueira, D. S., dos SPereira, A., de Aquino Neto, F. R., Cabral, J. A., Ferreira, C. A. C., Simoneit, B. R., & Elias, V. O. (2003). Determinação de compostos de massa molecular alta em folhas de plantas da Amazônia. *Química Nova*, 26(5), 633-640.
- Dees, W. H., J. P. Dupre, A. D. Richard, S. M. McMicken, O. E. Christian, C. W. Richmond and J. R. Woolman. McSU. J. Hightower. CPMC. (2014). The effects of botanical

- extracts on adult mosquitoes. *The Louisiana Academy of Sciences (Abstract)*, 4(1), 28. (B)
- Dees, W.H., J. P. Dupre, A. D. Richard, S. M. Mc Micken, O. E. Christian, C. W. Richmond and J. R. Woolman. McSU. J. Hightower. CPCM. (2014). The effects of botanical extracts on mosquito oviposition. *The Louisiana Academy of Sciences (Abstract)*, 4(1), 28-29. (A)
- Deng, Y. E., Balunas, M. J., Kim, J. A., Lantvit, D. D., Chin, Y. W., Chai, H., Sugiarto, S., Kardono, L. B. S., Fong, H. H. S., Pezzuto, J. M., Swanson, S. M., Carcache de Blanco, E. J. & Kinghorn, D. (2009). Bioactive 5, 6-dihydro- α -pyrone derivatives from *Hyptis brevipes*. *Journal of natural products*, 72(6), 1165-1169.
- Dev, S. (1999). Ancient-modern concordance in Ayurvedic plants: Some examples. *Environmental Health Perspectives*, 107, 783-789.
- Dewick, P. M. (2009). Medicinal natural products: a biosynthetic approach. 3ed. John Wiley & Sons Ltd. United Kingdom.
- Di Domenico, F., Pupo, G., Giraldo, E., Badia, M. C., Monllor, P., Lloret, A., Schinina, M. E., Giorgi, A., Cini, C., Tramutola, A., Butterfield, D. A., Viña, J. & Perlutti, M. (2016). Oxidative signature of cerebrospinal fluid from mild cognitive impairment and Alzheimer disease patients. *Free Radical Biology and Medicine*, 91, 1-9.
- Dillard, C. J., & German, J. B. (2000). Phytochemicals: nutraceuticals and human health. *Journal of the Science of Food and Agriculture*, 80(12), 1744-1756.
- Diniz, L. R. L., Vieira, C. F. X., dos Santos, E. C., Lima, G. C., Aragão, K. K. V., Vasconcelos, R. P., Oliveira, H. D., Portella, V. G. & Coelho-de-Souza, A. N. (2013). Gastroprotective effects of the essential oil of *Hyptis crenata* Pohl ex Benth. on gastric ulcer models. *Journal of ethnopharmacology*, 149(3), 694-700.
- Dongre, S. H., Badami, S., & Godavarthi, A. (2008). Antitumor activity of *Hypericum hookerianum* against DLA induced tumor in mice and its possible mechanism of action. *Phytotherapy Research*, 22(1), 23-29.
- Dorman, H. D., Bachmayer, O., Kosar, M., & Hiltunen, R. (2004). Antioxidant properties of aqueous extracts from selected Lamiaceae species grown in Turkey. *Journal of Agricultural and Food Chemistry*, 52(4), 762-770.
- Dorman, H. J. D., Deans, S. G., Noble, R. C., & Surai, P. (1995). Evaluation *in vitro* of plant essential oils as natural antioxidants. *Journal of Essential Oil Research*, 7(6), 645-651.
- DrugBank. (2018). Palmitic acid. Available at: <https://www.drugbank.ca/drugs/DB03796> accesed: 19th May 2018.
- Duarte, D. P. (2009). Process for manufacturing a naturally hydroplastic mask for removal or progressive reduction of hair growth. Assignee: Natureza Brasil Pesquisa e Desenvolvimento de Cosmeticos LTDA-ME, Brazil. Patent No. BR 2008000859.
- Dubey, N. K. (Ed.). (2014). *Plants as a source of natural antioxidants*. CABI.
- Dubois, M., Bailly, F., Mbemba, G., Mouscadet, J. F., Debysier, Z., Witvrouw, M., & Cotelle, P. (2008). Reaction of rosmarinic acid with nitrite ions in acidic conditions: discovery of

- nitro-and dinitrorosmarinic acids as new anti-HIV-1 agents. *Journal of medicinal chemistry*, 51(8), 2575-2579.
- Działo, M., Mierziak, J., Korzun, U., Preisner, M., Szopa, J., & Kulma, A. (2016). The potential of plant phenolics in prevention and therapy of skin disorders. *International journal of molecular sciences*, 17(2), 160.
- Džinić, T., & Dencher, N. A. (2018). Oxygen Concentration and Oxidative Stress Modulate the Influence of Alzheimer's Disease A β 1-42 Peptide on Human Cells. *Oxidative Medicine and Cellular Longevity*, 2018.
- Ellman, G. L., Courtney, K. D., Andres, V., & Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 7(2), 88IN191-9095.
- Erasmo, E. A. L., Pinheiro, L. L. A., & Costa, N. V. (2004). Phyto-sociological survey of weed communities in flooded rice areas cultivated under different management systems. *Planta Daninha*, 22(2), 195-201.
- Erkan, N., Ayrancı, G., & Ayrancı, E. (2008). Antioxidant activities of rosemary (*Rosmarinus Officinalis* L.) extract, blackseed (*Nigella sativa* L.) essential oil, carnosic acid, rosmarinic acid and sesamol. *Food Chemistry*, 110(1), 76-82.
- Fabricant, D. S., & Farnsworth, N. R. (2001). The value of plants used in traditional medicine for drug discovery. *Environmental health perspectives*, 109(Suppl 1), 69.
- Falcão, D. Q., & Menezes, F. S. (2003). Revisão etnofarmacológica, farmacológica e química do gênero *Hyptis*. *Revista Brasileira de Farmacognosia*, 84(3), 69-74.
- Falcão, D. Q., Fernandes, S. B. O., & Menezes, F. S. (2003). Triterpenos de *Hyptis fasciculata* Benth. *Revista Brasileira de Farmacognosia*, 13, 81-83.
- Falcão, D. Q., Menezes, F. S. (2003). The *Hyptis* genus: an ethnopharmacological and chemical review. *Revista Brasileira de Ciências Farmacêuticas*, 84, 69-74.
- Falcao, R. A., do Nascimento, P. L., de Souza, S. A., da Silva, T. M., de Queiroz, A. C., da Matta, C. B., Moreira, M. S. A., Camara, C. A. & Silva, T. M. S. (2013). Antileishmanial phenylpropanoids from the leaves of *Hyptis pectinata* (L.) Poit. *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Fang, X., Yang, C. Q., Wei, Y. K., Ma, Q. X., Yang, L., & Chen, X. Y. (2011). Genomics grand for diversified plant secondary metabolites. *Plant Div Res*, 33(1), 53-64.
- Farhoosh, R., Johnny, S., Asnaashari, M., Molaahmadibahraseman, N., & Sharif, A. (2016). Structure-antioxidant activity relationships of o-hydroxyl, o-methoxy, and alkyl ester derivatives of p-hydroxybenzoic acid. *Food chemistry*, 194, 128-134.
- Farnsworth, N. R., Akerele, O., Bingel, A. S., Soejarto, D. D., & Guo, Z. (1985). Medicinal plants in therapy. *Bulletin of the world health organization*, 63(6), 965.
- Feher, M., & Schmidt, J. M. (2003). Property distributions: differences between drugs, natural products, and molecules from combinatorial chemistry. *Journal of Chemical Information and Computer Sciences*, 43(1), 218-227.

- Felter, H. W., & Lloyd, J. U. (1898). *King's American dispensatory*. Eclectic Medical Publications.
- Fernández-Alonso, J. L., & Rivera-Díaz, O. (2006) Las labiadas (familia Labiatae). Libro rojo.
- Fernández-Alonso, J. L., Vega, N., Filgueira, J. J., & Pérez, G. (2003). Lectin prospecting in Colombian Labiatae. A systematic-ecological approach. *Biochemical Systematics and Ecology*, 31(6), 617-633.
- Ferrali, M., Signorini, C., Caciotti, B., Sugherini, L., Ciccoli, L., Giachetti, D., & Comporti, M. (1997). Protection against oxidative damage of erythrocyte membrane by the flavonoid quercetin and its relation to iron chelating activity. *FEBS letters*, 416(2), 123-129.
- Ferreira, E. C., Faria, L. C., Santos, S. C., Ferri, P. H., Silva, J. G., Paula, J. R., & Ferreira, H. D. (2005). Essential Oils of *Hyptis conferta* Pohl ex Benth. var. conferta and *Hyptis conferta* Pohl ex Benth. var. angustata (Briq.) Pohl ex Harley from Brazilian Cerrado. *Journal of Essential Oil Research*, 17(2), 145-146.
- Ferreira, H. D. Morfologia, taxonomia, filogenia, anatomia foliar e fitoquímica de espécies do gênero *Hyptis* Jacq. (Labiatae) ocorrentes em Goiás e Tocantins. Tese (Doutorado Ciências Biológicas) Universidade Federal de Goiás. 2009.
- Fidyt, K., Fiedorowicz, A., Strzadala, L., & Szumny, A. (2016). β -caryophyllene and β -caryophyllene oxide—natural compounds of anticancer and analgesic properties. *Cancer medicine*, 5(10), 3007-3017.
- Fierascu, R. C., Padure, I. M., Avramescu, S. M., Ungureanu, C., Bunghez, R. I., Ortan, A., Dinupirvu, C., Fierascu, I. & Soare, L. C. (2016). Preliminary assessment of the antioxidant, antifungal and germination inhibitory potential of *Heracleum sphondylium* L.(Apiaceae). *Farmacia*, 64(3), 403-408.
- Flora do Brasil. Lamiaceae in Flora do Brasil 2020 under construction. Botanical Garden of Rio de Janeiro. Available in <<http://reflora.jbrj.gov.br/reflora/floradobrasil/FB133004>>. Accessed on: Mar 30, 2018 (*Hyptis radicans*).
- Flora do Brasil. Lamiaceae in Flora do Brasil 2020 under construction. Botanical Garden of Rio de Janeiro. Available in: <<http://reflora.jbrj.gov.br/reflora/floradobrasil/FB8232>>. Accessed on: Mar 30, 2018 (*Hyptis multibracteata*).
- França, B. K., Alves, M. R. M., Souto, F. M. S., Tiziane, L., Boaventura, R. F., Guimarães, A., & Alves Jr, A. (2013). Peroxidação lipídica e obesidade: Métodos para aferição do estresse oxidativo em obesos. *GE jornal português de gastroenterologia*, 20(5), 199-206.
- Fronza, M. (2011). Phytochemical investigation of the roots of *Peltodon longipes* and *in vitro* cytotoxic studies of abietane diterpenes (Doctoral dissertation).
- Fronza, M., Lamy, E., Günther, S., Heinzmann, B., Laufer, S., & Merfort, I. (2012). Abietane diterpenes induce cytotoxic effects in human pancreatic cancer cell line MIA PaCa-2 through different modes of action. *Phytochemistry*, 78, 107-119.

- Fronza, M., Murillo, R., Ślusarczyk, S., Adams, M., Hamburger, M., Heinzmann, B., Laufer, S. & Merfort, I. (2011). *In vitro* cytotoxic activity of abietane diterpenes from *Peltodon longipes* as well as *Salvia miltiorrhiza* and *Salvia sahendica*. *Bioorganic & medicinal chemistry*, 19(16), 4876-4881.
- Fu, J., Cheng, K., Zhang, Z. M., Fang, R. Q., & Zhu, H. L. (2010). Synthesis, structure and structure-activity relationship analysis of caffeic acid amides as potential antimicrobials. *European journal of medicinal chemistry*, 45(6), 2638-2643.
- Fujiki, H., Suganuma, M., Okabe, S., Sueoka, E., Suga, K., Imai, K., & Kimura, S. (1999). Mechanistic findings of green tea as cancer preventive for humans. *Proceedings of the Society for Experimental Biology and Medicine*, 220(4), 225-228.
- Furlan, C. M., Santos, K. P., Sedano-Partida, M. D., da Motta, L. B., Santos, D. Y. A., Salatino, M. L. F., Negri, G., Berry, P. E., van Ee, B. W. & Salatino, A. (2015). Flavonoids and antioxidant potential of nine Argentinian species of *Croton* (Euphorbiaceae). *Brazilian Journal of Botany*, 38(4), 693-702.
- Gallegos-Monterrosa, R., Mhatre, E., & Kovács, Á. T. (2016). Specific *Bacillus subtilis* 168 variants form biofilms on nutrient-rich medium. *Microbiology*, 162(11), 1922-1932.
- Garambone, E., & Rosa, G. (2008). Possíveis benefícios do ácido clorogênico à saúde. *Alimentos e Nutrição Araraquara*, 18(2), 229-235.
- García-Granados, A., López, P. E., Melguizo, E., Parra, A., & Simeó, Y. (2007). Remote hydroxylation of methyl groups by regioselective cyclopalladation. Partial synthesis of hyptatic acid-A. *The Journal of organic chemistry*, 72(9), 3500-3509.
- Gaspar-Marques, C., Fátima Simões, M., Luísa Valdeira, M., & Rodriguez, B. (2008). Terpenoids and phenolics from *Plectranthus strigosus*, bioactivity screening. *Natural product research*, 22(2), 167-177.
- Gayathiri, K., Sangeetha, M., Sharanya, V. K., Prakash, G. S., Vimalavathini, R., Sudheer, J. G., & Kumar, S. K. (2016). A Review: Potential Pharmacological Uses of Natural Products from Lamiaceae. *Int J Pharma Res Rev [Internet]*, 21-34.
- Gershenzon, J. (2017). The cost of plant chemical defense against herbivory: a biochemical perspective. In *Insect-Plant Interactions* (1993) (pp. 121-190). CRC Press.
- Geuenich, S., Goffinet, C., Venzke, S., Nolkemper, S., Baumann, I., Plinkert, P., Reichling, J. & Keppler, O. T. (2008). Aqueous extracts from peppermint, sage and lemon balm leaves display potent anti-HIV-1 activity by increasing the virion density. *Retrovirology*, 5(1), 27.
- Ghalib, R. M., Hashim, R., Sulaiman, O., Mehdi, S. H., Anis, Z., Rahman, S. Z., ... & Abdul Majid, A. M. S. (2012). Phytochemical analysis, cytotoxic activity and constituents-activity relationships of the leaves of *Cinnamomum iners* (Reinw. ex Blume-Lauraceae). *Natural product research*, 26(22), 2155-2158.
- Gibson, G. E., & Huang, H. M. (2005). Oxidative stress in Alzheimer's disease. *Neurobiology of Aging*, 26(5), 575-578.

- Göçer, H., & Gülçin, İ. (2011). Caffeic acid phenethyl ester (CAPE): correlation of structure and antioxidant properties. *International Journal of Food Sciences and Nutrition*, 62(8), 821-825.
- Golembiovská, O., Tsurkan, A., & Vynogradov, B. (2014). Components of *Prunella vulgaris* L. Grown in Ukraine. *Journal of Pharmacognosy and Phytochemistry*, 2(6).
- Gonçalves, S. D. J., Rêgo, M., & Araújo, A. D. (1996). Abelhas sociais (Hymenoptera: Apidae) e seus recursos florais em uma região de mata secundária, Alcântara, MA, Brasil. *Acta Amazônica*, 26(1/2).
- González, A. G., Bazzocchi, I. L., Moujir, L., Ravelo, A. G., Correa, M. D., & Gupta, M. P. (1995). Xanthine oxidase inhibitory activity of some Panamanian plants from Celastraceae and Lamiaceae. *Journal of ethnopharmacology*, 46(1), 25-29.
- Goun, E., Cunningham, G., Chu, D., Nguyen, C., & Miles, D. (2003). Antibacterial and antifungal activity of Indonesian ethnomedical plants. *Fitoterapia*, 74(6), 592-596.
- Grayer, R. J., Eckert, M. R., Veitch, N. C., Kite, G. C., Marin, P. D., Kokubun, T., Simmonds, M. S. J. & Paton, A. J. (2003). The chemotaxonomic significance of two bioactive caffeic acid esters, nepetoidins A and B, in the Lamiaceae. *Phytochemistry*, 64(2), 519-528.
- Grayson, D. H. (2000). Monoterpeneoids. *Natural product reports*, 17(4), 385-419.
- Greff, D. 1998. Cosmetic compositions comprising at least one *Hyptis* extract. Assignee: Sederma S.A., Fr. Patent No. WO 9843608
- Grenand, P. Moretti, C., Jacquemin H., Prévost, M. F. (2004). Pharmacopées Traditionnelles en Guyane, *Institut de Recherche pour le Développement*. 2nd edition. Paris, France.
- Grundhöfer, P., Niemetz, R., Schilling, G., & Gross, G. G. (2001). Biosynthesis and subcellular distribution of hydrolyzable tannins. *Phytochemistry*, 57(6), 915-927.
- Gupta, M. P., Monge, A., Karikas, G. A., Lopez de Cerain, A., Solis, P. N., De Leon, E., Trujillo, M., Suarez, O., Wilson, F., Montenegro, G., Noriega, Y., Santana, A. L., Correa M. & Sanchez, C. (1996). Screening of Panamanian medicinal plants for brine shrimp toxicity, crown gall tumor inhibition, cytotoxicity and DNA intercalation. *International Journal of Pharmacognosy*, 34(1), 19-27.
- Gupta, M., Sharma, C., Meena, P., & Khatri, M. (2017). investigating the free radical scavenging and acetylcholinesterase inhibition activities of *Ellettaria cardamomum*, *Piper nigrum* and *Syzygium aromaticum*. *International Journal of Pharmaceutical Sciences and Research*, 8(7), 3180-3186.
- Gutteridge, J. M. (1984). Reactivity of hydroxyl and hydroxyl-like radicals discriminated by release of thiobarbituric acid-reactive material from deoxy sugars, nucleosides and benzoate. *Biochemical Journal*, 224(3), 761.
- Guzman, J. D. (2014). Natural cinnamic acids, synthetic derivatives and hybrids with antimicrobial activity. *Molecules*, 19(12), 19292-19349.
- Halliwell, B., & Gutteridge, J. M. (2015). *Free radicals in biology and medicine*. Oxford University Press, USA.

- Halliwell, B., & Whiteman, M. (2004). Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean?. *British Journal of Pharmacology*, 142(2), 231-255.
- Halliwell, B., Gutteridge, J. M., & Aruoma, O. I. (1987). The deoxyribose method: a simple "test-tube" assay for determination of rate constants for reactions of hydroxyl radicals. *Analytical Biochemistry*, 165(1), 215-219.
- Harborne, J. B. (2014). *Introduction to ecological biochemistry*. Academic press.
- Harborne, J. B., & Williams, C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*, 55(6), 481-504.
- Harley, R. M. (2012). Checklist and key of genera and species of the Lamiaceae of the Brazilian Amazon. *Rodriguésia*, 63(1), 129-144.
- Harley, R. M., & Pastore, J. F. B. (2012). A generic revision and new combinations in the Hyptidinae (Lamiaceae), based on molecular and morphological evidence. *Phytotaxa*, 58(1), 1-55.
- Harley, R. M., Atkins, S., Budantsev, A., Cantino, P. D., Conn, B. J., Grayer, R., Harley, M. M., De Kok, R., Kretovskaja, T., Morales, R., Paton, P. J., Ryding, O. & Upson, T. 2004. Labiateae. Pp. 167-275 in: Kadereit, J.W. (ed.), *The families and genera of vascular plants*, vol. 7. Berlin: Springer
- Harley, R. M., Reynolds, T., & Atkins, S. (1992). *Advances in Labiate science*. Royal Botanical Gardens, Kew.
- Harley, R., França, F., Santos, E. P., Santos, J. S., Pastore, J. F. 2015. Lamiaceae in Lista de Espécies da Flora do Brasil. Jardim Botânico do Rio de Janeiro.
- Hayes, J. D., Kelleher, M. O., & Eggleston, I. M. (2008). The cancer chemopreventive actions of phytochemicals derived from glucosinolates. *European journal of nutrition*, 47(2), 73-88.
- Heinrich, M. (2010). Ethnopharmacology in the 21st century-grand challenges. *Frontiers in pharmacology*, 1, 8.
- Heinrich, M., & Gibbons, S. (2001). Ethnopharmacology in drug discovery: an analysis of its role and potential contribution. *Journal of Pharmacy and Pharmacology*, 53(4), 425-432.
- Heldt, H. W., & Piechulla, B. (2004). *Plant biochemistry*. Academic Press.
- Henrich, C. J., & Beutler, J. A. (2013). Matching the power of high throughput screening to the chemical diversity of natural products. *Natural product reports*, 30(10), 1284-1298.
- Hermsmeier, D., Schittko, U., & Baldwin, I. T. (2001). Molecular interactions between the specialist herbivore *Manduca sexta* (Lepidoptera, Sphingidae) and its natural host *Nicotiana attenuata*. I. Large-scale changes in the accumulation of growth-and defense-related plant mRNAs. *Plant physiology*, 125(2), 683-700.
- Hinneburg, I., Dorman, H. D., & Hiltunen, R. (2006). Antioxidant activities of extracts from selected culinary herbs and spices. *Food chemistry*, 97(1), 122-129. Hossain, M.,

- Brunton, N., Barry-Ryan, C., Martin-Diana, A. B., & Wilkinson, M. (2008). Antioxidant activity of spice extracts and phenolics in comparison to synthetic antioxidants.
- Hintz, T., Matthews, K. K., & Di, R. (2015). The use of plant antimicrobial compounds for food preservation. *BioMed research international*, 2015.
- HMDB. (2018). Human metabolome Database. Arachidic acid. Available at: <http://www.hmdb.ca/metabolites/HMDB0002212> accsesed: 19th May, 2018. (A)
- HMDB. (2018). Human metabolome Database. Linoleic acid. Available at: <http://www.hmdb.ca/metabolites/HMDB0000673> accsesed: 19th May, 2018. (C)
- HMDB. (2018). Human metabolome Database. Methyl stearate. Available at: <http://www.hmdb.ca/metabolites/HMDB0034154> accsesed: 19th May, 2018. (B)
- Hong, J., Yang, L., Zhang, D., & Shi, J. (2016). Plant metabolomics: an indispensable system biology tool for plant science. *International journal of molecular sciences*, 17(6), 767.
- Hooker, C. W., Lott, W. B., & Harrich, D. (2001). Inhibitors of human immunodeficiency virus type 1 reverse transcriptase target distinct phases of early reverse transcription. *Journal of virology*, 75(7), 3095-3104.
- Hosztafi, S. (1997). The discovery of alkaloids. *Die Pharmazie*, 52(7), 546-550.
- Huang, K. C. (1998). *The pharmacology of Chinese herbs*. Routledge.
- Hunter, P. (2008). Harnessing Nature's wisdom: Turning to Nature for inspiration and avoiding her follies. *EMBO reports*, 9(9), 838-840.
- Hussain, S. P., Hofseth, L. J., & Harris, C. C. (2003). Radical causes of cancer. *Nature Reviews Cancer*, 3(4), 276-285.
- Igwe, O. U. (2014). Chromatographic and spectrometric characterization of bioactive compounds from the leaves of *Hyptis lanceolata* Poir. *International Journal of Chemistry and Pharmaceutical Sciences*, Vol.2(1): 547-553
- Iloki-Assanga, S. B., Lewis-Luján, L. M., Lara-Espinoza, C. L., Gil-Salido, A. A., Fernandez-Angulo, D., Rubio-Pino, J. L., & Haines, D. D. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L. and *Phoradendron californicum*. *BMC Research Notes*, 8(1), 396.
- Ito, H., Miyazaki, T., Ono, M., & Sakurai, H. (1998). Antiallergic activities of rhabdosin and its related compounds: chemical and biochemical evaluations. *Bioorganic & medicinal chemistry*, 6(7), 1051-1056.
- IUCN. Numbers of threatened species by major groups of organisms (1996–2015). (2015). Available at:http://cmsdocs.s3.amazonaws.com/summarystats/2015_2_Summary_Stats_Page_Documents/2015_2_RL_Stats_Table_1.pdf
- Izzo, A. A., & Capasso, F. (2007). Herbal medicines to treat Alzheimer's disease. *Trends in Pharmacological Sciences*, 28(2), 47-48.

- Jeong, G. H., Cho, J. H., Jo, C., Lee, S., Lee, S. S., Bai, H. W., Chung, B. Y. & Kim, T. H. (2018). Gamma irradiation-assisted degradation of rosmarinic acid and evaluation of structures and anti-adipogenic properties. *Food chemistry*, 258, 181-188.
- Jesus, N. Z. T. D., Lima, J. C. D. S., Silva, R. M. D., Espinosa, M. M., & Martins, D. T. D. O. (2009). Ethnobotanical survey of plants popularly used as anti-ulcer and anti-inflammatory in Pirizal, Nossa Senhora do Livramento, MT, Brazil. *Revista Brasileira de Farmacognosia*, 19(1A), 130-139.
- Judd, W., Campbell, C., Kellogg, E., Stevens, P., Donoghue, M., 2002. Plant Systematics: a Phylogenetic Approach, 2nd ed. *Sinauer Associates Sunderland*.
- Jupudi, S., Rao, P., & Prakasa, N. S. (2002). Database of Natural Products-Lead Identification, Optimization by Molecular Modeling Studies of Olguines Reported from Genus of Family.
- Kaiser, H. (2008). Von der Pflanze zur Chemie—Die Frühgeschichte der Rheumamittel “From the plant to chemistry—the early history of “rheumatic medication”. *Zeitschrift für Rheumatologie*, 67(3), 252-262.
- Kang, B. C., Yeam, I., & Jahn, M. M. (2005). Genetics of plant virus resistance. *Annu. Rev. Phytopathol.*, 43, 581-621.
- Kapoor, L. D. (1990). CRC handbook of Ayurvedic plants. *CSIR, USA*, 183.
- Kashiwada, Y., Wang, H. K., Nagao, T., Kitanaka, S., Yasuda, I., Fujioka, T., Yamagishi, T., Consentino, L. M., Kozuka, M., Okabe, H., Ikeshiro, Y., Hu, C. Q., Yeh, E. & Lee, K. H. (1998). Anti-AIDS agents. 30. Anti-HIV activity of oleanolic acid, pomolic acid, and structurally related triterpenoids. *Journal of Natural Products*, 61(9), 1090-1095.
- Kelley, C. J., Mahajan, J. R., Brooks, L. C., Neubert, L. A., Breneman, W. R., & Carmack, M. (1975). Polyphenolic acids of *Lithospermum ruderale* (Boraginaceae). I. Isolation and structure determination of lithospermic acid. *The Journal of Organic Chemistry*, 40(12), 1804-1815.
- Kerdudo, A., Njoh Ellong, E., Gonnnot, V., Boyer, L., Michel, T., Adenet, S., Rochefort, K. & Fernandez, X. (2016). Essential oil composition and antimicrobial activity of *Hyptis atrorubens* Poit. from Martinique (FWI). *Journal of Essential Oil Research*, 28(5), 436-444.
- Kim, G. D., Park, Y. S., Jin, Y. H., & Park, C. S. (2015). Production and applications of rosmarinic acid and structurally related compounds. *Applied microbiology and biotechnology*, 99(5), 2083-2092.
- Kinghorn, A. D., Pan, L., Fletcher, J. N., & Chai, H. (2011). The relevance of higher plants in lead compound discovery programs. *Journal of natural products*, 74(6), 1539-1555.
- Kini, F., Kam, B., Aycard, J. P., Gaydou, E. M., & Bombarda, I. (1993). Chemical composition of the essential oil of *Hyptis spicigera* Lam. from Burkina Faso. *Journal of Essential Oil Research*, 5(2), 219-221.

- Klayman, D. L., Lin, A. J., Acton, N., Scovill, J. P., Hoch, J. M., Milhous, W. K. & Theoharides, A. D. (1984). Isolation of artemisinin (qinghaosu) from *Artemisia annua* growing in the United States. *Journal of natural products*, 47(4), 715-717.
- Koba, K., Raynaud, C., Millet, J., Chaumont, J. P., & Sanda, K. (2007). Chemical Composition of *Hyptis pectinata* L., *H. lanceolata* Poit, *H. suaveolens* (L) Poit and *H. spicigera* Lam. Essential Oils from Togo. *Journal of Essential Oil Bearing Plants*, 10(5), 357-364.
- Kobayashi, K., Umishio, K., Ota, M., Inomata, S., Satake, G. & Sekita, S. 2001. Antiaging cosmetics comprising gelatinase inhibitors. Assignee: *Shiseido Co., Ltd., Japan*. Patent No JP 2001172157. (B)
- Kobayashi, K., Umishio, K., Ota, M., Yoshida, Y., Satake, M. & Sekita, S. (2001). Serine protease inhibitors and skin preparations containing the inhibitors for treatment of rough skin. Assignee: *Shiseido Co., Ltd., Japan; Human Science Shinko Zaidan*. Patent No. JP 2001240551. (A)
- Koehn, F. E., & Carter, G. T. (2005). The evolving role of natural products in drug discovery. *Nature reviews Drug discovery*, 4(3), 206.
- Koury, J. C., & Donangelo, C. M. (2003). Zinco, estresse oxidativo e atividade física. *Revista de Nutrição*, 16(4), 433-441.
- Krinsky, N. I. (1992). Mechanism of action of biological antioxidants. *Proceedings of the Society for Experimental Biology and Medicine*, 200(2), 248-254.
- Kruse, P. R. (2007). Geschichte der Pharmazie. Vol. II: Von der Frühen Neuzeit bis zur Gegenwart-by Rudolf Schmitz. *Centaurus*, 49(2), 182-183.
- Kumar, S., & Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 2013.
- Kvist, L. P., & Holm-Nielsen, L. B. (1987). Ethnobotanical aspects of lowland Ecuador. *Opera Botanica*, 92(83), 107.
- Ladan, Z., Okonkwo, E. M., Amupitan, J. O., Ladan, E. O., & Aina, B. (2010). Physicochemical properties and fatty acid profile of *Hyptis spicigera* seed oil. *Research Journal of Applied Sciences*, 5(2), 123-125.
- Law, M. (2000). Plant sterol and stanol margarines and health. *BMJ: British Medical Journal*, 320(7238), 861
- Leclercq, P. A., Delgado, H. S., Garcia, J., Hidalgo, J. E., Cerrutti, T., Mestanza, M., Ríos, F., Nina, E., Nonato, L., Alvarado, R. & Menéndez, R. (2000). Aromatic Plant Oils of the Peruvian Amazon. Part 2. *Cymbopogon citratus* (DC) Stapf., *Renealmia* sp., *Hyptis recurvata* Poit. and *Tyanthus panurensis* (Bur.) Sandw. *Journal of Essential Oil Research*, 12(1), 14-18.
- Lee, K. H., Lin, Y. M., Wu, T. S., Zhang, D. C., Yamagishi, T., Hayashi, T., Hall, I. H., Chang, J. J., Wu, R. Y. & Yang, T. H. (1988). The cytotoxic principles of *Prunella vulgaris*, *Psychotria serpens*, and *Hyptis capitata*: ursolic acid and related derivatives. *Planta medica*, 54(04), 308-311.

- Li, B., & Olmstead, R. G. (2017). Two new subfamilies in Lamiaceae. *Phytotaxa*, 313(2), 222-226.
- Li, B., Cantino, P. D., Olmstead, R. G., Bramley, G. L., Xiang, C. L., Ma, Z. H., Tan, Y. H. & Zhang, D. X. (2016). A large-scale chloroplast phylogeny of the Lamiaceae sheds new light on its subfamilial classification. *Scientific reports*, 6, 34343.
- Li, X. F., Tang, L., Zheng, N. N., An, S., Wang, Y. X., Zhang, W. X., & Hu, Y. N. (2014). Determination of Antioxidant Activity *In Vitro* from *Hyptis rhomboidea* Mart. et Gal. Ethanol Extracts. In *Applied Mechanics and Materials* (Vol. 472, pp. 824-828). Trans Tech Publications.
- Lim, G. H., Singhal, R., Kachroo, A., & Kachroo, P. (2017). Fatty Acid-and Lipid-Mediated Signaling in Plant Defense. *Annual review of phytopathology*, 55, 505-536.
- Lima, G. C, Vasconcelos, Y. D. A. G., de Santana, S. M. T., Oliveira, A. S., Bomfim, R. R., de Albuquerque, J. R. L. C., Camargo, E. A., Portella, V. G., Coelho-de-Souza, A. N. & Diniz, L. R. L. (2018). Hepatoprotective Effect of Essential Oils from *Hyptis crenata* in Sepsis-Induced Liver Dysfunction. *Journal of Medicinal Food*.
- Lin, H. (2017). Efficient Chinese medicinal pesticide for garden plant and flower and preparation method thereof. By: Lin, Han. Assignee: Peop. Rep. China. Patent No.: CN 106857715
- Lin, J. K., Liang, Y. C., & Lin-Shiau, S. Y. (1999). Cancer chemoprevention by tea polyphenols through mitotic signal transduction blockade. *Biochemical Pharmacology*, 58(6), 911-915.
- Lin, Y. L., Chang, Y. Y., Kuo, Y. H., & Shiao, M. S. (2002). Anti-Lipid-Peroxidative Principles from *Tournefortia s armentosa*. *Journal of natural products*, 65(5), 745-747.
- Lin, Y. L., Lee, H. P., Huang, R. L., Ou, J. C., Kuo & Y. H. (1993). Studies on the Constituents of *Hyptis Rhomboidea*. *The Chinese Pharmaceutical Journal*, 45(1), 61-8.
- Lin, Y. L., Wang, C. N., Shiao, Y. J., Liu, T. Y., & Wang, W. Y. (2003). Benzolignanoid and polyphenols from *Origanum vulgare*. *Journal of the Chinese Chemical Society*, 50(5), 1079-1083.
- Lin, Y., Shi, R., Wang, X., & Shen, H. M. (2008). Luteolin, a flavonoid with potential for cancer prevention and therapy. *Current cancer drug targets*, 8(7), 634-646.
- Lipid home. (2018). A Lipid Primer - the Diversity of Natural Lipids. Available at:<http://www.lipidhome.co.uk/lipids/basics/whatlip/index.htm> accesed: 19th May 2018.
- Liu, A. H., Li, L., Xu, M., Lin, Y. H., Guo, H. Z., & Guo, D. A. (2006). Simultaneous quantification of six major phenolic acids in the roots of *Salvia miltiorrhiza* and four related traditional Chinese medicinal preparations by HPLC-DAD method. *Journal of pharmaceutical and biomedical analysis*, 41(1), 48-56.
- Liu, D. & Yang, C. (2015). Preparation method of hyptatic acid A and anti-tumor application thereof. Assignee: Nanjing Zelang Medical Technology Co., Ltd., Peop. Rep. China. Patent No. CN 104945461

- Liu, X., Chen, R., Shang, Y., Jiao, B., & Huang, C. (2008). Lithospermic acid as a novel xanthine oxidase inhibitor has anti-inflammatory and hypouricemic effects in rats. *Chemico-biological interactions*, 176(2-3), 137-142.
- Liu, Z. (2008). Preparation of botanical samples for biomedical research. *Endocrine, Metabolic & Immune Disorders-Drug Targets (Formerly Current Drug Targets-Immune, Endocrine & Metabolic Disorders)*, 8(2), 112-121.
- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy reviews*, 4(8), 118.
- Loizzo, M. R., Bonesi, M., Passalacqua, N. G., Saab, A., Menichini, F., & Tundis, R. (2013). Antiproliferative activities on renal, prostate and melanoma cancer cell lines of *Sarcopoterium spinosum* aerial parts and its major constituent tormentic acid. *Anti-Cancer Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Anti-Cancer Agents)*, 13(5), 768-776.
- Loizzo, M. R., Menichini, F., Conforti, F., Tundis, R., Bonesi, M., Saab, A. M., Statti, G. A., de Cindio, B., Houghton, P. J., Menichini, F. & Frega, N. G. (2009). Chemical analysis, antioxidant, antiinflammatory and anticholinesterase activities of *Origanum ehrenbergii* Boiss and *Origanum syriacum* L. essential oils. *Food Chemistry*, 117(1), 174-180.
- Luz, A. I. R., Zoghbi, M. G. B., Ramos, L. S., Maia, J. G. S., & Da Silva, M. L. (1984). Essential Oils of Some Amazonian Labiateae, I: Genus *Hyptis*. *Journal of Natural Products*, 47(4), 745-747.
- Mabry, T. J., Markham, K. R., & Thomas, M. B. (1970). The ultraviolet spectra of isoflavones, flavanones and dihydroflavonols. In *The systematic identification of flavonoids* (pp. 165-226). Springer, Berlin, Heidelberg.
- Maclean, I. M., & Wilson, R. J. (2011). Recent ecological responses to climate change support predictions of high extinction risk. *Proceedings of the National Academy of Sciences*, 108(30), 12337-12342.
- Maes, M., Loyter, A., & Friedler, A. (2012). Peptides that inhibit HIV-1 integrase by blocking its protein-protein interactions. *The FEBS journal*, 279(16), 2795-2809.
- Maimone, T. J., & Baran, P. S. (2007). Modern synthetic efforts toward biologically active terpenes. *Nature chemical biology*, 3(7), 396-407.
- Maisuthisakul, P., Suttajit, M., & Pongsawatmanit, R. (2007). Assessment of phenolic content and free radical-scavenging capacity of some Thai indigenous plants. *Food Chemistry*, 100(4), 1409-1418.
- Maiyoa, F., Moodley, R., & Singh, M. (2016). Phytochemistry, cytotoxicity and apoptosis studies of β -sitosterol-3-oglucoside and β -amyrin from *Prunus africana*. *African Journal of Traditional, Complementary and Alternative Medicines*, 13(4), 105-112.
- Manian, R., Anusuya, N., Siddhuraju, P., & Manian, S. (2008). The antioxidant activity and free radical scavenging potential of two different solvent extracts of *Camellia sinensis* (L.) O. Kuntz, *Ficus bengalensis* L. and *Ficus racemosa* L. *Food Chemistry*, 107(3), 1000-1007.

- Markham, K. R. (1982). Techniques of flavonoid identification. London, Academic press.
- Marrelli, M., Loizzo, M. R., Nicoletti, M., Menichini, F., & Conforti, F. (2014). In vitro investigation of the potential health benefits of wild Mediterranean dietary plants as anti-obesity agents with α -amylase and pancreatic lipase inhibitory activities. *Journal of the Science of Food and Agriculture*, 94(11), 2217-2224.
- Mashkovsky, M. D., & Kruglikova-Lvova, R. P. (1951). On the pharmacology of the new alkaloid galantamine. *Farmakologija Toxicologija (Moscow)*, 14, 27-30.
- Mathew, M., & Subramanian, S. (2014). In vitro screening for anti-cholinesterase and antioxidant activity of methanolic extracts of ayurvedic medicinal plants used for cognitive disorders. *PLoS One*, 9(1), e86804.
- Matkowski, A., & Piotrowska, M. (2006). Antioxidant and free radical scavenging activities of some medicinal plants from the Lamiaceae. *Fitoterapia*, 77(5), 346-353.
- Matsuse, I. T., Lim, Y. A., Hattori, M., Correa, M., & Gupta, M. P. (1998). A search for anti-viral properties in Panamanian medicinal plants: The effects on HIV and its essential enzymes. *Journal of ethnopharmacology*, 64(1), 15-22.
- McCue, P. P., & Shetty, K. (2004). Inhibitory effects of rosmarinic acid extracts on porcine pancreatic amylase in vitro. *Asia Pacific Journal of Clinical Nutrition*, 13(1).
- McGaw, L. J., Elgorashi, E. E., & Eloff, J. N. (2014). Cytotoxicity of African medicinal plants against normal animal and human cells. In *Toxicological Survey of African Medicinal Plants* (pp. 181-233).
- McNeil, M., Facey, P., & Porter, R. (2011). Essential oils from the *Hyptis* genus—a review (1909–2009). *Nat Prod Commun*, 6, 1775-1796.
- Medina, E., & Pieper, D. H. (2016). Tackling threats and future problems of multidrug-resistant bacteria. In *How to Overcome the Antibiotic Crisis* (pp. 3-33). Springer, Cham.
- Melo, A. M. (2003). Estudo fitoquímico e biológico da *Hyptis mutabilis* Salz. (Lamiaceae).
- Messana, I., Ferrari, F., e Souza, M. A. D. M., & Gács-Baitz, E. (1990). (-)-Salzol, an isopimarane diterpene, and a chalcone from *Hyptis salzmanii*. *Phytochemistry*, 29(1), 329-332.
- Miléo, L. J., Silva, J. F., Bentes, J. L. S., & Christoffoleti, P. J. (2007). Plantas daninhas hospedeiras alternativas de *Colletotrichum guanicola* em cultivos de guaraná no Estado do Amazonas. *Planta daninha*, 25, 771-782.
- Miller, A. E., & Heyland, A. (2010). Endocrine interactions between plants and animals: Implications of exogenous hormone sources for the evolution of hormone signaling. *General and comparative endocrinology*, 166(3), 455-461.
- Misra, T. N., Singh, R. S., & Upadhyay, J. (1983). Triterpenoids from *Hyptis suaveolens* roots. *Phytochemistry*, 22(2), 603-605.

- Misra, T. N., Singh, R. S., Oiha, T. N., & Upadhyay, J. (1981). Chemical constituents of *Hyptis suaveolens*. Part I. Spectral and biological studies on a triterpene acid. *Journal of Natural Products*, 44(6), 735-738.
- Mitani, Hiroaki; Soda, Makoto; Oshima, Koichi. 2000. Hyaluronidase inhibitors for cosmetic and other manufacturing. Assignee: Sansho Seiyaku Co., Ltd., Japan; Sumitomo Forestry Co., Ltd. JP 1106311.
- Moalin, M., Van Strijdonck, G. P., Beckers, M., Hagemen, G. J., Borm, P. J., Bast, A., & Haenen, G. R. (2011). A planar conformation and the hydroxyl groups in the B and C rings play a pivotal role in the antioxidant capacity of quercetin and quercetin derivatives. *Molecules*, 16(11), 9636-9650.
- Mohapatra, D. K., Kanikarapu, S., Naidu, P. R., & Yadav, J. S. (2015). Toward the synthesis of brevipolide H. *Tetrahedron Letters*, 56(8), 1041-1044.
- Moire, L., Rezzonico, E., Goepfert, S., & Poirier, Y. (2004). Impact of unusual fatty acid synthesis on futile cycling through β -oxidation and on gene expression in transgenic plants. *Plant physiology*, 134(1), 432-442.
- Moon, J. K., & Shibamoto, T. (2009). Antioxidant assays for plant and food components. *Journal of Agricultural and Food Chemistry*, 57(5), 1655-1666.
- Moosavi, F., Hosseini, R., Rajaian, H., Silva, T., e Silva, D. M., Saso, L., Edraki, N., Miri, R., Borges, F. & Firuzi, O. (2017). Derivatives of caffeic acid, a natural antioxidant, as the basis for the discovery of novel nonpeptidic neurotrophic agents. *Bioorganic & medicinal chemistry*, 25(12), 3235-3246.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1-2), 55-63.
- Mühlen, V. C. (2009). Índices de retenção em cromatografia gasosa bidimensional abragente. *Scientia chromatographica*, 1(3), 21-8. Available at: <http://scientiachromatographica.com/files/v1n3/v1n3a3.pdf>.
- Mukherjee, P. K., Kumar, V., Mal, M., & Houghton, P. J. (2007). Acetylcholinesterase inhibitors from plants. *Phytomedicine*, 14(4), 289-300.
- Murillo, E., Britton, G. B., & Durant, A. A. (2012). Antioxidant activity and polyphenol content in cultivated and wild edible fruits grown in Panama. *Journal of pharmacy & bioallied sciences*, 4(4), 313.
- Mussi de Mira, N. V., Cerdeira Barros, R. M., Schiocchet, M. A., Noldin, J. A., & Lanfer-Marquez, U. M. (2008). Extração, análise e distribuição dos ácidos fenólicos em genótipos pigmentados e não pigmentados de arroz (*Oryza sativa* L.). *Ciência e Tecnologia de Alimentos*, 28(4).
- Nakanishi, T., Nishi, M., Inada, A., Obata, H., Tanabe, N., Abe, S., & Wakashiro, M. (1990). Two new potent inhibitors of xanthine oxidase from leaves of *Perilla frutescens* Britton var. acuta Kudo. *Chemical and Pharmaceutical Bulletin*, 38(6), 1772-1774.

- Nayak, U. G., & Guha, P. C. (1952). Essential oil from *Hyptis Suaveolens* Poit.. *Journal of the Indian Chemical Society*, 29(3), 183-186.
- Nayeem, N., Asdaq, S. M. B., Salem, H. & AHEl-Alfqy, S. (2016) Gallic Acid: A Promising Lead Molecule for Drug Development. *Journal of Applied Pharmacy*, 8:213.
- Ncube, B., & Van Staden, J. (2015). Tilting plant metabolism for improved metabolite biosynthesis and enhanced human benefit. *Molecules*, 20(7), 12698-12731.
- Nedelkoska, T. V., & Doran, P. M. (2000). Characteristics of heavy metal uptake by plant species with potential for phytoremediation and phytomining. *Minerals engineering*, 13(5), 549-561.
- Newman, D. J., & Cragg, G. M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of natural products*, 75(3), 311-335.
- Nijveldt, R. J., Van Nood, E. L. S., Van Hoorn, D. E., Boelens, P. G., Van Norren, K., & Van Leeuwen, P. A. (2001). Flavonoids: a review of probable mechanisms of action and potential applications-. *The American journal of clinical nutrition*, 74(4), 418-425.
- NIST (2017). Available at: <http://nistmassspeclibrary.com/2017/05/identify-any-spectrum-with-our-nist-library-upgrade/> accessed: 19th may, 2018.
- Nobel prize. (2015). Press Release for the 2015 Nobel Prize in Medicine Laureates. Available:http://www.nobelprize.org/nobel_prizes/medicine/laureates/2015/press.html (Accessed: 8th July 2016).
- Notka, F., Meier, G. R., & Wagner, R. (2003). Inhibition of wild-type human immunodeficiency virus and reverse transcriptase inhibitor-resistant variants by *Phyllanthus amarus*. *Antiviral research*, 58(2), 175-186.
- Odonbayar, B., Murata, T., Matsumoto, N., Batkhuu, J., & Sasaki, K. (2017). Chemical constituents of aerial parts of *Thymus gobicus* and their cholinesterase inhibitory activities. *Mongolian Journal of Chemistry*, 17(43), 14-17.
- Ohkawa, H., Ohishi, N., Yagi, K. 1979. Assay for lipid peroxidation in animal tissues by thiobarbituric acid reaction. *Annals of Biochemistry*, 95, 351- 358.
- Okhale, S. E., Ode, S. S., Oladosu, P., & Ugbabe, G. E. (2018). Evaluation of the antioxidant and anti-proliferative chemical constituents of *Hyptis pectinata* (Linn.) aerial infusion. *International journal of pharmacognosy*.
- Oliveira, A. D. L. de. (2011). Estudo químico e avaliação biológica do óleo essencial de *Hyptis martiusii* Benth. (Lamiaceae). Dissertação (Mestrado em Bioprospecção Molecular) *Universidade Regional do Cariri – URCA*.
- Olivero-Verbel, J., & Pacheco-Londoño, L. (2002). Structure– activity relationships for the anti-HIV activity of flavonoids. *Journal of chemical information and computer sciences*, 42(5), 1241-1246.
- Onayade, O. A., Looman, A., Scheffer, J. J. C., & Svendsen, A. B. (1990). Composition of the herb essential oil of *Hyptis spicigera* Lam. *Flavour and fragrance journal*, 5(2), 101-105.

- Ortega, H. E., Shen, Y. Y., TenDyke, K., Rios, N., & Cubilla-Ríos, L. (2014). Polyhydroxylated macrolide isolated from the endophytic fungus *Pestalotiopsis mangiferae*. *Tetrahedron Letters*, 55(16), 2642-2645.
- Oshima, Ryo; Soda, Makoto. (2000). Antibacterial agent/Highly safe antibacterial agent obtained from plants. Assignee: Sumitomo Forestry Co., Ltd., Japan. Patent No. JP 2000136141.
- Otero, R., Núñez, V., Barona, J., Fonnegra, R., Jiménez, S. L., Osorio, R. G., Saldarriaga, M. & Diaz, A. (2000). Snakebites and ethnobotany in the northwest region of Colombia: Part III: Neutralization of the haemorrhagic effect of *Bothrops atrox* venom. *Journal of Ethnopharmacology*, 73(1-2), 233-241.
- Ożarowski, M., Piasecka, A., Gryszczyńska, A., Sawikowska, A., Pietrowiak, A., Opala, B., Mikołajczak, P. L., Kujawski, R., Kachlicki, P., Buchwald, W., & Seremak-Mrozikiewicz, A. (2017). Determination of phenolic compounds and diterpenes in roots of *Salvia miltiorrhiza* and *Salvia przewalskii* by two LC-MS tools: Multi-stage and high resolution tandem mass spectrometry with assessment of antioxidant capacity. *Phytochemistry Letters*, 20, 331-338.
- Özbek, H., Güvenalp, Z., Özak, T., Sevindik, H. G., Yuca, H., Yerdelen, K. Ö., & Demirezer, L. Ö. (2017). Chemical composition, antioxidant and anticholinesterase activities of the essential oil of *Origanum rotundifolium* Boiss. from Turkey. *Records of Natural Products*, 11(5), 9.
- Park, I. W., Han, C., Song, X., Green, L. A., Wang, T., Liu, Y., Cen, C., Song, X., Yang, B., Chen, G. & He, J. J. (2009). Inhibition of HIV-1 entry by extracts derived from traditional Chinese medicinal herbal plants. *BMC complementary and alternative medicine*, 9(1), 29.
- Park, Y. K., Alencar, S. M., & Aguiar, C. L. (2002). Botanical origin and chemical composition of Brazilian propolis. *Journal of Agricultural and Food Chemistry*, 50(9), 2502-2506.
- Parker, C., Caton, B. P., & Fowler, L. (2007). Ranking nonindigenous weed species by their potential to invade the United States. *Weed Science*, 55(4), 386-397.
- Parker, C., Caton, B. P., & Fowler, L. (2007). Ranking nonindigenous weed species by their potential to invade the United States. *Weed Science*, 55(4), 386-397.
- Pastore, J. F. B. (2010). Filogenia Molecular da Subtribo Hyptidinae (Labiatae) e suas Implicações Taxonômicas. Tese (Doutorado em Botânica) Universidade Estadual de Feira de Santana.
- Pedersen, J. A. (2000). Distribution and taxonomic implications of some phenolics in the family Lamiaceae determined by ESR spectroscopy. *Biochemical systematics and Ecology*, 28(3), 229-253.
- Pedroso, R. C. N., Branquinho, N. A. A., Hara, A. C., Costa, A. C., Silva, F. G., Pimenta, L. P., Silva, M. L. A., Cunha, W. R., Pauletti, P. M. & Januario, A. H. (2017). Impact of light quality on flavonoid production and growth of *Hyptis marruboides* seedlings cultivated in vitro. *Revista Brasileira de Farmacognosia*, 27(4), 466-470.

- Pereda-Miranda, R., & Delgado, G. (1990). Triterpenoids and flavonoids from *Hyptis albida*. *Journal of Natural Products*, 53(1), 182-185.
- Pereda-Miranda, R., Hernández, L., Villavicencio, M. J., Novelo, M., Ibarra, P., Chai, H., & Pezzuto, J. M. (1993). Structure and stereochemistry of pectinolides AC, novel antimicrobial and cytotoxic 5, 6-dihydro- α -pyrones from *Hyptis pectinata*. *Journal of natural products*, 56(4), 583-593.
- Perera, W. H., Bizzo, H. R., Gama, P. E., Alviano, C. S., Salimena, F. R. G., Alviano, D. S., & Leitão, S. G. (2017). Essential oil constituents from high altitude Brazilian species with antimicrobial activity: *Baccharis parvidentata* Malag., *Hyptis monticola* Mart. ex Benth. and *Lippia origanoides* Kunth. *Journal of Essential Oil Research*, 29(2), 109-116.
- Perini, J. Â. D. L., Stevanato, F. B., Sargi, S. C., Visentainer, J. E. L., Dalalio, M. M. D. O., Matshushita, M., Souza, N. E. D. & Visentainer, J. V. (2010). Ácidos graxos poli-insaturados n-3 e n-6: metabolismo em mamíferos e resposta imune. Omega-3 and omega-6 polyunsaturated fatty acids: metabolism in mammals and immune response. *Revista de Nutrição*, 23(6), 1075-1086.
- Petersen, M. (2013). Rosmarinic acid: new aspects. *Phytochemistry Reviews*, 12(1), 207- 227.
- Petersen, M., & Simmonds, M. S. (2003). Rosmarinic acid. *Phytochemistry*, 62(2), 121-125.
- Phaniendra, A., Jestadi, D. B., & Periyasamy, L. (2015). Free radicals: properties, sources, targets, and their implication in various diseases. *Indian Journal of Clinical Biochemistry*, 30(1), 11-26.
- Pharmacopoeia Commission of People's Republic of China. (2010). Pharmacopoeia of the People's Republic of China.; *Chemical Industry Press*: Beijing, China, 2010; p. 71.
- Pietta, P. G. (2000). Flavonoids as antioxidants. *Journal of Natural Products*, 63(7), 1035-1042.
- Pourmorad, F., Hosseini-mehr, S. J., & Shahabimajd, N. (2006). Antioxidant activity, phenol and flavonoid contents of some selected Iranian medicinal plants. *African Journal of Biotechnology*, 5(11).
- Poyton, R. O., Ball, K. A., & Castello, P. R. (2009). Mitochondrial generation of free radicals and hypoxic signaling. *Trends in Endocrinology & Metabolism*, 20(7), 332-340.
- Prasad, N. R., Karthikeyan, A., Karthikeyan, S., & Reddy, B. V. (2011). Inhibitory effect of caffeic acid on cancer cell proliferation by oxidative mechanism in human HT-1080 fibrosarcoma cell line. *Molecular and Cellular Biochemistry*, 349(1-2), 11-19.
- Procházková, D., Boušová, I., & Wilhelmová, N. (2011). Antioxidant and prooxidant properties of flavonoids. *Fitoterapia*, 82(4), 513-523.
- Quideau, S., Deffieux, D., Douat-Casassus, C., & Pouysegur, L. (2011). Plant polyphenols: chemical properties, biological activities, and synthesis. *Angewandte Chemie International Edition*, 50(3), 586-621.
- Radulović, N. S., & Blagojević, P. D. (2012). Volatile Secondary Metabolites of *Micromeria dalmatica* Benth.(Lamiaceae): Biosynthetical and Chemotaxonomical Aspects. *Chemistry & biodiversity*, 9(7), 1303-1319.

- Raghuveer, C., & Tandon, R. V. (2009). Consumption of functional food and our health concerns. *Pakistan Journal of Physiology*, 5(1), 76-83.
- Rao, B. G. V. N., & Nigam, S. S. (1972). Chemical examination of the fixed oil from the seeds of *Hyptis suaveolens*. *Indian Oil Soap J*, 37(12), 295-300.
- Rattan, R. S. (2010). Mechanism of action of insecticidal secondary metabolites of plant origin. *Crop protection*, 29(9), 913-920.
- Razzaghi-Asl, N., Garrido, J., Khazraei, H., Borges, F., & Firuzi, O. (2013). Antioxidant properties of hydroxycinnamic acids: a review of structure-activity relationships. *Current medicinal chemistry*, 20(36), 4436-4450.
- Rebelo, M. M., Silva, J. K. R. D., Andrade, E. H. A., & Maia, J. G. S. (2009). Antioxidant capacity and biological activity of essential oil and methanol extract of *Hyptis crenata* Pohl ex Benth. *Revista Brasileira de Farmacognosia*, 19(1B), 230-235.
- Reymond, P., Weber, H., Damond, M., & Farmer, E. E. (2000). Differential gene expression in response to mechanical wounding and insect feeding in *Arabidopsis*. *The Plant Cell*, 12(5), 707-719.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, 20(7), 933-956.
- Richardson, P. M. (1992). The chemistry of the Labiateae: an introduction and overview. *Advances in Labiateae Science*. Kew: Royal Botanic Gardens, 291-297.
- Rocha, G. S., Roughan, J., Leach, M., Flecknell, P., Ingram, C. D., & Brandt, K. (2009, November). Antinociceptive activities of *Hyptis crenata* pohl. In *II International Symposium on Medicinal and Nutraceutical Plants* 972 (pp. 105-110). (A)
- Rocha, G., Roughan, J. V., Leach, M. C., Flecknell, P. A., Ingram, C. D., & Brandt, K. (2009). Traditional use, chemical analysis and antinociceptive effects of *Hyptis crenata* Pohl. *Planta Medica*, 75(09), PA40. (B)
- Rohn, S., Rawel, H. M., & Kroll, J. (2002). Inhibitory effects of plant phenols on the activity of selected enzymes. *Journal of Agricultural and Food Chemistry*, 50(12), 3566-3571.
- Roman Junior W.A., Picolli, A., Morais, B., Loeblein, M., & Schönell, A. (2015). Atividade antiulcerogênica do extrato aquoso de *Salvia officinalis* L.(Lamiaceae). *Revista Brasileira de Plantas Medicinais*, 17(4 supl I), 774-781.
- Royal Botanic Gardens & Plants of the World Online. (2018). *Hyptis radicans* and *Hyptis multibracteata* (world distribution). Available at: <http://powo.science.kew.org> accesed: 19th May 2018.
- Ruhfel, B. R., Gitzendanner, M. A., Soltis, P. S., Soltis, D. E., & Burleigh, J. G. (2014). From algae to angiosperms—inferring the phylogeny of green plants (Viridiplantae) from 360 plastid genomes. *BMC Evolutionary Biology*, 14(1), 23.
- Russell, W., & Duthie, G. (2011). Plant secondary metabolites and gut health: the case for phenolic acids. *Proceedings of the Nutrition Society*, 70(3), 389-396.

- Saavedra, O. M., Vázquez, E. N. J., Vargas, M. R. B. G., Reyes, G. M. C., & Bolaina, E. M. (2010). Radicales libres y su papel en las enfermedades crónico-degenerativas. *Rev. Médica Univ. Veracruzana*, 10(2), 32-39.
- Sakr, H. H. & Roshdy, S. H. (2015). Effect of *Hyptis brevipes* (Lamiaceae) Methanol Extract on *Spodoptera littoralis* (Lepidoptera: Noctuidae) Larvae. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 6(6), 651. ISSN: 0975-8585
- Sakr, H. H., Roshdy, S. H. & El-Seedi H. R. (2013). *Hyptis brevipes* (Lamiaceae) Extracts Strongly Inhibit the Growth and Development of *Spodoptera littoralis* (Boisd.) Larvae (Lepidoptera: Noctuidae). *Journal of Applied Pharmaceutical Science*, 3 (10), pp. 083-088. DOI: 10.7324/JAPS.2013.31014
- Sala-Carvalho, W. R. (2017). Estudo *in vitro* dos potenciais antioxidante, antimicrobiano e anti-HIV de extratos de *Hyptis lacustris* A. St.-Hil. Ex Benth. (Lamiaceae). *Universidade de São Paulo*, Brasil.
- Sales, J. D. F., Pinto, J. E. B. P., Ferri, P. H., Silva, F. G., de Oliveira, C. B. A., & Botrel, P. P. (2009). Growth, production and chemical composition of the essential oil in hortelã-do-campo (*Hyptis marrubiooides* Epl.) in function of the irradiation level. *Semina: Ciencias Agrárias (Londrina)*, 30(2), 389-395. (A)
- Sales, J. D. F., Pinto, J. E. B. P., Oliveira, J. A. D., Botrel, P. P., Silva, F. G., & Corrêa, R. M. (2011). The germination of bush mint (*Hyptis marrubiooides* EPL.) seeds as a function of harvest stage, light, temperature and duration of storage. *Acta Scientiarum. Agronomy*, 33(4), 709-713.
- Sales, J. F., Pinto, J. E. B. P., Botrel, P. P., Silva, F. G., Correa, R. M., & de Carvalho, J. G. (2009). Biomass accumulation, foliar content of nutrients and yield of essential oil of hortelã-do-campo (*Hyptis marrubiooides* EPL.) cultivated under organic fertilization. *Bioscience Journal*, 25(1), 60-68. (B)
- Sales, J. F., Pinto, J. E. B., Botrel, P. P., Oliveira, C. B., Ferri, P. H., Paula, J. R., & Seraphin, J. C. (2007). Composition and chemical variability in the essential oil of *Hyptis marrubiooides* Epl. *Journal of Essential Oil Research*, 19(6), 552-556.
- Santos, K. P. D., Sedano-Partida, M. D., Sala-Carvalho, W. R., Loureiro, B. O. S. J., da Silva-Luz, C. L., & Furlan, C. M. (2018). Biological activity of *Hyptis* Jacq. (Lamiaceae) is determined by the environment. *Industrial Crops and Products*, 112, 705-715.
- Santos, K. P., Sedano-Partida, M. D., Motta, L. B., Cordeiro, I., & Furlan, C. M. (2016). Antioxidant activity of flavonoids from *Croton sphaerogynus* Baill. *Brazilian Journal of Botany*, 39(4), 1021-1030.
- Santos, M. R., & Mira, L. (2004). Protection by flavonoids against the peroxynitrite-mediated oxidation of dihydrorhodamine. *Free radical research*, 38(9), 1011-1018.
- Sayah, K., Marmouzi, I., Naceiri Mrabti, H., Cherrah, Y., & Faouzi, M. E. A. (2017). Antioxidant activity and inhibitory potential of *Cistus salvifolius* (L.) and *Cistus monspeliensis* (L.) Aerial parts extracts against key enzymes linked to Hyperglycemia. *BioMed research international*, 2017.

- Scio, E., Mendes, R. F., Motta, E. V., Bellozi, P. M., Aragão, D. M., Mello, J., Fabri, R. L., Moreira, J. R., de Assis, I. V. L. & Bouzada, M. L. M. (2012). Antimicrobial and antioxidant activities of some plant extracts. In *Phytochemicals as Nutraceuticals- Global Approaches to Their Role in Nutrition and Health*. InTech.
- Scramin, S., Saito, M. L., Pott, A., & Marques, M. O. M. (2000). Volatile constituents of *Hyptis crenata* Pohl (Labiatae) native in Brazilian pantanal. *Journal of Essential Oil Research*, 12(1), 99-101.
- Sedano-Partida, M. D., Santos, K. P., Loureiro, B. O. S. J., & Furlan, C. M. (2015). Anti-HIV activity of *Hyptis* Jacq. (Lamiaceae). *Planta Medica*, 81(16), PM_88.
- Serturner, F. W. (1817). Ueber das Morphium, eine neue salifahige Grundlage, und die Mekonsaure als Hauptbestandtheile des Opiums. *Annalen der Physik*, 25, 56-89.
- Seyoum, A., Pålsson, K., Kung'a, S., Kabiru, E. W., Lwande, W., Killeen, G. F., Hassanali, A. & Knots, B. G. J. (2002). Traditional use of mosquito-repellent plants in western Kenya and their evaluation in semi-field experimental huts against *Anopheles gambiae*: ethnobotanical studies and application by thermal expulsion and direct burning. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 96(3), 225-231.
- Shahrbandy, K., & Hosseinzadeh, R. (2007). *In vitro* Antioxidant Activity of *Polygonum hyrcanicum*, *Centaurea depressa*, *Sambucus ebulus*, *Mentha spicata* and *Phytolacca americana*. *Pakistan Journal of Biological Sciences*, 10(4), 637-640.
- Shami, N. J. I. E., & Moreira, E. A. M. (2004). Licopeno como agente antioxidante. *Rev Nutr*, 17(2), 227-36.
- Shan, B., Cai, Y. Z., Sun, M., & Corke, H. (2005). Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. *Journal of agricultural and food chemistry*, 53(20), 7749-7759.
- Sharififar, F., Dehghn-Nudeh, G., & Mirtajaldini, M. (2009). Major flavonoids with antioxidant activity from *Teucrium polium* L. *Food Chemistry*, 112(4), 885-888.
- Shekarchi, M., Hajimehdipoor, H., Saeidnia, S., Gohari, A. R., & Hamedani, M. P. (2012). Comparative study of rosmarinic acid content in some plants of Labiatae family. *Pharmacognosy magazine*, 8(29), 37.
- Shen, W. (2015). Germicidal composition containing thiophanate-methyl and *Hyptis rhomboidea* extract. By: Shen, Wei. Assignee: Huzhou Xulong Biochemistry Co., Ltd., Peop. Rep. China. Patent No. CN 105165912.
- Shepherd, G. J., 2011. FITOPAC 2 - Software package for multivariate analysis. V. 2.1.2 Campinas: UNICAMP.
- Shi, S., Zhao, Y., Zhou, H., Zhang, Y., Jiang, X., & Huang, K. (2008). Identification of antioxidants from *Taraxacum mongolicum* by high-performance liquid chromatography-diode array detection-radical-scavenging detection-electrospray ionization mass spectrometry and nuclear magnetic resonance experiments. *Journal of Chromatography A*, 1209(1-2), 145-152.

- Silva, A. C. D., de Souza, P. E., de Resende, M. L. V., Júnior, M. B. D. S., Vitorino, L. R. R., & Baroni, G. D. R. (2013). Decocts isolados e em mistura com fungicida no controle do ódio em minicepas de eucalipto. *Revista Caatinga*, 26(3), 73-79. (B)
- Silva, A. C. D., de Souza, P. E., Pinto, J. E. B. P., da Silva, B. M., Amaral, D. C., & de Arruda Carvalho, E. (2012). Essential oils for preventative treatment and control of Asian soybean rust. *European journal of plant pathology*, 134(4), 865-871. (B)
- Silva, A. C. D., Souza, P. E. D., Amaral, D. C., Zeviani, W. M., & Pinto, J. E. B. P. (2014). Essential oils from *Hyptis marruboides*, *Aloysia gratissima* and *Cordia verbenacea* reduce the progress of Asian soybean rust. *Acta Scientiarum. Agronomy*, 36(2), 159-166. (B)
- Silva, A. C. D., Souza, P. E. D., Machado, J. D. C., Silva, B. M. D., & Pinto, J. E. B. P. (2012). Effectiveness of essential oils in the treatment of *Colletotrichum truncatum*-infected soybean seeds. *Tropical Plant Pathology*, 37(5), 305-313. (A)
- Silva, A. C. R. D., Lopes, P. M., Azevedo, M. M. B. D., Costa, D. C. M., Alviano, C. S., & Alviano, D. S. (2012). Biological activities of α-pinene and β-pinene enantiomers. *Molecules*, 17(6), 6305-6316. (C)
- Silva, A. C., Souza, P. E., Resende, M. L. V., Silva, M. B., Ribeiro, P. M., & Zeviani, W. M. (2014). Local and systemic control of powdery mildew in *eucalyptus* using essential oils and decoctions from traditional Brazilian medicinal plants. *Forest pathology*, 44(2), 145-153. (A)
- Silva, C. G., Raulino, R. J., Cerqueira, D. M., Mannarino, S. C., Pereira, M. D., Panek, A. D., Silva, J. F. M., Menezes, F. S. & Eleutherio, E. C. A. (2009). In vitro and in vivo determination of antioxidant activity and mode of action of isoquercitrin and *Hyptis fasciculata*. *Phytomedicine*, 16(8), 761-767.
- Silva, R. F., Rezende, C. M., Santana, H. C., Vieira, R. F., & Bizzo, H. R. (2013). Scents from Brazilian Cerrado: chemical composition of the essential oil from the leaves of *Hyptis villosa* Pohl ex Benth (Lamiaceae). *Journal of Essential Oil Research*, 25(5), 415-418. (A)
- Silva-Luz, C. L. D., Gomes, C. G., Pirani, J. R., & Harley, R. M. (2012). Flora da Serra do Cipó, Minas Gerais: Lamiaceae. *Boletim de Botânica*, 30(2), 109-155.
- Simões, C. M. O. (2001). *Farmacognosia: da planta ao medicamento*. UFRGS; Florianópolis: UFSC.
- Siquet, C., Paiva-Martins, F., Lima, J. L., Reis, S., & Borges, F. (2006). Antioxidant profile of dihydroxy-and trihydroxyphenolic acids-A structure-activity relationship study. *Free radical research*, 40(4), 433-442.
- Skaltsa, H. D., Mavrommati, A., & Constantinidis, T. (2001). A chemotaxonomic investigation of volatile constituents in *Stachys* subsect. *Swainsonianeae* (Labiatae). *Phytochemistry*, 57(2), 235-244.
- Šliumpaitė, I., Venskutonis, P. R., Murkovic, M., & Ragažinskienė, O. (2013). Antioxidant properties and phenolic composition of wood betony (*Betonica officinalis* L., syn. *Stachys officinalis* L.). *Industrial Crops and Products*, 50, 715-722.

- Sneader, W. (2005). *Drug discovery: a history*. John Wiley & Sons.
- Soares, S. E. (2002). Ácidos fenólicos como antioxidantes. *Revista de Nutrição*.
- Son, S., & Lewis, B. A. (2002). Free radical scavenging and antioxidative activity of caffeic acid amide and ester analogues: Structure–activity relationship. *Journal of agricultural and food chemistry*, 50(3), 468-472.
- Souza, J. N., Silva, E. M., Loir, A., Rees, J. F., Rogez, H., & Larondelle, Y. (2008). Antioxidant capacity of four polyphenol-rich Amazonian plant extracts: A correlation study using chemical and biological in vitro assays. *Food Chemistry*, 106(1), 331-339.
- Souza, L. K. H., Oliveira, C. M. A. D., Ferri, P. H., Santos, S. C., Oliveira Júnior, J. G. D., Miranda, A. T. B., Lião, L. M. & Silva, M. D. R. R. (2002). Antifungal properties of Brazilian cerrado plants. *Brazilian Journal of Microbiology*, 33(3), 247-249.
- Souza, L. K., Oliveira, C., Ferri, P. H., Oliveira Júnior, J. G. D., Souza Júnior, A. H. D., Fernandes, O. D. F. L., & Silva, M. D. R. R. (2003). Antimicrobial activity of *Hyptis ovalifolia* towards dermatophytes. *Memorias do Instituto Oswaldo Cruz*, 98(7), 963-965.
- Stadnik, M. J., Bettoli, W., & Saito, M. L. (2003). Bioprospecting for plant and fungus extracts with systemic effect to control the cucumber powdery mildew/Bioprospektion von pflanzlichen und pilzlichen Extrakten mit systemischem Effekt zur Bekämpfung des Echten Mehltaupilzes an Gurken. *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz/Journal of Plant Diseases and Protection*, 383-393.
- Stahura, F. L., Godden, J. W., Xue, L., & Bajorath, J. (2000). Distinguishing between natural products and synthetic molecules by descriptor Shannon entropy analysis and binary QSAR calculations. *Journal of chemical information and computer sciences*, 40(5), 1245-1252.
- Steffen, L. M. (2006). Eat your fruit and vegetables. *The Lancet*, 367(9507), 278-279.
- Su, Liuhua. (2013). Method for extracting asprelllic acid a. Assignee: Nanjing Zelang Agriculture Development Co., Ltd., Peop. Rep. China. Patent No. CN 102887935
- Suárez-Ortiz, G. A., Cerda-García-Rojas, C. M., Fragoso-Serrano, M., & Pereda-Miranda, R. (2017). Complementarity of DFT Calculations, NMR Anisotropy, and ECD for the Configurational Analysis of Brevipolides K-O from *Hyptis brevipes*. *Journal of natural products*, 80(1), 181-189.
- Suárez-Ortiz, G. A., Cerda-García-Rojas, C. M., Hernández-Rojas, A., & Pereda-Miranda, R. (2013). Absolute configuration and conformational analysis of brevipolides, bioactive 5, 6-dihydro- α -pyrones from *Hyptis brevipes*. *Journal of natural products*, 76(1), 72-78.
- Tafurt-García, G., Jiménez-Vidal, L. F., & Calvo-Salamanca, A. M. (2015). Antioxidant capacity and total phenol content of *Hyptis* spp., *P. heptaphyllum*, *T. panamensis*, *T. rhoifolia* and *Ocotea* sp. *Revista Colombiana de Química*, 44(2), 28-33.
- Tafurt-García, G., Jiménez-Vidal, L. F., & Calvo-Salamanca, A. M. (2015). Antioxidant capacity and total phenol content of *Hyptis* spp., *P. heptaphyllum*, *T. panamensis*, *T. rhoifolia* and *Ocotea* sp. *Revista Colombiana de Química*, 44(2), 28-33.

- Tafurt-Garcia, G., Munoz-Acevedo, A., Calvo, A. M., Jimenez, L. F., & Delgado, W. A. (2014). Componentes volátiles de *Eriope crassipes*, *Hyptis conferta*, *H. dilatata*, *H. brachiata*, *H. suaveolens* y *H. mutabilis* (Lamiaceae). *Boletín Latinoamericano y del Caribe de Plantas Medicinales y aromáticas*, 13(3).
- Taguchi, R., Hatayama, K., Takahashi, T., Hayashi, T., Sato, Y., Sato, D., Ohta, K., Nakano, H., Seki, C., Endo, Y., Tokuraku, K. & Uwai, K. (2017). Structure–activity relations of rosmarinic acid derivatives for the amyloid β aggregation inhibition and antioxidant properties. *European journal of medicinal chemistry*, 138, 1066-1075.
- Tahara, S. (2007). A journey of twenty-five years through the ecological biochemistry of flavonoids. *Bioscience, biotechnology, and biochemistry*, 71(6), 1387-1404.
- Tajkarimi, M. M., Ibrahim, S. A., & Cliver, D. O. (2010). Antimicrobial herb and spice compounds in food. *Food control*, 21(9), 1199-1218.
- Takeda, H., Tsuji, M., Matsumiya, T., & Kubo, M. (2002). Identification of rosmarinic acid as a novel antidepressive substance in the leaves of *Perilla frutescens* Britton var. acuta Kudo (Perillae Herba). *Nihon shinkei seishin yakurigaku zasshi= Japanese journal of psychopharmacology*, 22(1), 15-22.
- Tang, L., Li, X. F., Yang, S. X., Qiu, Y., & Yuan, K. (2014). Chemical constituents of *Hyptis rhomboidea* and their antifungal activity. *Zhongguo Zhong yao za zhi= Zhongguo zhongyao zazhi= China journal of Chinese materia medica*, 39(12), 2284-2288.
- Tarnawski, M., Depta, K., Grejciun, D., & Szelepin, B. (2006). HPLC determination of phenolic acids and antioxidant activity in concentrated peat extract—a natural immunomodulator. *Journal of Pharmaceutical and Biomedical Analysis*, 41(1), 182-188.
- Taylor, P., Arsenak, M., Abad, M. J., Fernández, Á., Milano, B., Gonto, R., Ruiz, M. C., Fraile, S., Taylor, S., Estrada, O. & Michelangeli, F. (2013). Screening of Venezuelan medicinal plant extracts for cytostatic and cytotoxic activity against tumor cell lines. *Phytotherapy Research*, 27(4), 530-539.
- Tchoumbougnang, F., Zollo, P. H., Boyom, F. F., Nyegue, M. A., Bessière, J. M., & Menut, C. (2005). Aromatic plants of tropical Central Africa. XLVIII. Comparative study of the essential oils of four *Hyptis* species from Cameroon: *H. lanceolata* Poit., *H. pectinata* (L.) Poit., *H. spicigera* Lam. and *H. suaveolens* Poit. *Flavour and fragrance journal*, 20(3), 340-343.
- Tepe, B. (2008). Antioxidant potentials and rosmarinic acid levels of the methanolic extracts of *Salvia virgata* (Jacq), *Salvia staminea* (Montbret & Aucher ex Benth) and *Salvia verbenaca* (L.) from Turkey. *Bioresource Technology*, 99(6), 1584-1588.
- Thomas, C. D., Cameron, A., Green, R. E., Bakkenes, M., Beaumont, L. J., Collingham, Y. C., Erasmus, B. F. N., de Siqueira, M. F., Grainger, A., Hannah, L., Hughes, L., Huntley, B., van Jaarsveld, A. S., Midgley, G. F., Miles, L., Ortega-Huerta, M. A., Townsend Peterson, A., Phillips, O. L. & Williams, S. E. (2004). Extinction risk from climate change. *Nature*, 427(6970), 145.
- Thomford, N. E., Dzobo, K., Adu, F., Chirikure, S., Wonkam, A., & Dandara, C. (2018). Bush mint (*Hyptis suaveolens*) and spreading hogweed (*Boerhavia diffusa*) medicinal plant

- extracts differentially affect activities of CYP1A2, CYP2D6 and CYP3A4 enzymes. *Journal of ethnopharmacology*, 211, 58-69.
- Thuong, P. T., Kang, K. W., Kim, J. K., Seo, D. B., Lee, S. J., Kim, S. H., & Oh, W. K. (2009). Lithospermic acid derivatives from *Lithospermum erythrorhizon* increased expression of serine palmitoyltransferase in human HaCaT cells. *Bioorganic & medicinal chemistry letters*, 19(6), 1815-1817.
- Tomás-Barberán, F. A., & Wollenweber, E. (1990). Flavonoid aglycones from the leaf surfaces of some Labiatae species. *Plant Systematics and Evolution*, 173(3-4), 109-118.
- Tsai, S. F., & Lee, S. S. (2014). Neolignans as xanthine oxidase inhibitors from *Hyptis rhombooides*. *Phytochemistry*, 101, 121-127.
- Tsao, R. (2010). Chemistry and biochemistry of dietary polyphenols. *Nutrients*, 2(12), 1231-1246.
- Upadhyay, R., Chaurasia, J. K., Tiwari, K. N., & Singh, K. (2014). Antioxidant property of aerial parts and root of *Phyllanthus fraternus* Webster, an important medicinal plant. *The Scientific World Journal*, 2014.
- Urones, J. G., Marcos, I. S., Diez, D., & Cubilla, L. (1998). Tricyclic diterpenes from *Hyptis dilatata*. *Phytochemistry*, 48(6), 1035-1038.
- Vaca, C. E., Wilhelm, J., Harms-Ringdahl, M. 1988. Interaction of lipid peroxidation products with DNA. A Review. *Mutation Research*, 195, 137-149.
- Valadeau, C., V., Pabon, A., Deharo, E., Albán-Castillo, J., Estevez, Y., Lores Fransis, A., Rojas, R., Gamboa, D., Sauvain, M. & Bourdy, G. (2009). Medicinal plants from the Yanesha (Peru): evaluation of the leishmanicidal and antimalarial activity of selected extracts. *Journal of ethnopharmacology*, 123(3), 413-422.
- Valant-Vetschera, K. M., Roitman, J. N., & Wollenweber, E. (2003). Chemodiversity of exudate flavonoids in some members of the Lamiaceae. *Biochemical systematics and ecology*, 31(11), 1279-1289.
- Van Overbeek, J.; Velez, I. (1946). Weed control in Puerto Rico with 2,4-D. *Inst. Agr. Tropical, Univ. Puerto Rico, Bol.* Volume 1 27 pp.
- Van Wyk, B. E. (2011). The potential of South African plants in the development of new medicinal products. *South African Journal of Botany*, 77(4), 812-829.
- Velasco, V., & Williams, P. (2011). Improving meat quality through natural antioxidants. *Chilean journal of agricultural research*, 71(2), 313.
- Vergallo, A., Giampietri, L., Baldacci, F., Volpi, L., Chico, L., Pagni, C., Giorgi, F. S., Ceravolo, R., Tognoni, G., Siciliano, G. & Bonuccelli, U. (2018). Oxidative Stress Assessment in Alzheimer's Disease: A Clinic Setting Study. *American Journal of Alzheimer's Disease & Other Dementias®*, 33(1), 35-41.
- Verpoorte, R. (1998). Exploration of nature's chemodiversity: the role of secondary metabolites as leads in drug development. *Drug Discovery Today*, 3(5), 232-238.

- Verpoorte, R. (2000). Pharmacognosy in the new millennium: leadfinding and biotechnology. *Journal of pharmacy and pharmacology*, 52(3), 253-262.
- Vingtdeux, V., Dreses-Werringloer, U., Zhao, H., Davies, P., & Marambaud, P. (2008). Therapeutic potential of resveratrol in Alzheimer's disease. *BMC neuroscience*, 9(2), S6.
- Violante, I. M. P., Hamerski, L., Garcez, W. S., Batista, A. L., Chang, M. R., Pott, V. J., & Garcez, F. R. (2012). Antimicrobial activity of some medicinal plants from the cerrado of the central-western region of Brazil. *Brazilian Journal of Microbiology*, 43(4), 1302-1308. (B)
- Violante, I. M., Garcez, W. S., Barbosa, S. C., & Garcez, F. R. (2012). Chemical composition and biological activities of essential oil from *Hyptis crenata* growing in the Brazilian cerrado. *Natural product communications*, 7(10), 1387-1389. (A)
- Vitorino, L. C., Silva, F. G., Lima, W. C., Soares, M. A., Pedroso, R. C. N., Silva, M. R., Dias, H. J., Crotti, A. E. M., Silva, M. L. A., Cunha, W. R., Pauletti, P. M. & Januário, A. H. (2013). Metabolic response induced by endophytic fungi and bacteria in *H. marrubiooides* Epling *in vitro* microplants. *Química Nova*, 36(7), 1014-1020.
- Walley, J. W., Kliebenstein, D. J., Bostock, R. M., & Dehesh, K. (2013). Fatty acids and early detection of pathogens. *Current opinion in plant biology*, 16(4), 520-526.
- Wang, H., Cao, G., & Prior, R. L. (1996). Total antioxidant capacity of fruits. *Journal of agricultural and food chemistry*, 44(3), 701-705.
- Wang, L., Deng, D. Y., Tang, L., Yuan, K., & Li, Y. X. (2015). Seed-Germination and Seedling-Growth Inhibition by Aqueous Extracts from *Hyptis Rhomboidea*. In *2015 International Conference on Industrial Technology and Management Science*. Atlantis Press.
- Watzke, A., O'Malley, S. J., Bergman, R. G., & Ellman, J. A. (2006). Reassignment of the configuration of salvianolic acid B and establishment of its identity with lithospermic acid B. *Journal of natural products*, 69(8), 1231-1233.
- WHO. 2016. HIV/AIDS. World Health Organization. Available at: <http://www.who.int/hiv/data/en/> (Accessed: 15th May, 2018)
- WHO. 2016. Antibiotic resistance. World Health Organization. Available at: <http://www.who.int/mediacentre/factsheets/antibiotic-resistance/en/> (Accessed: 14th February 2017).
- Wichner, D., Idris, H., Houssen, W. E., McEwan, A. R., Bull, A. T., Asenjo, J. A., Goodfellow, M., Jaspars, M., Ebel, R. & Rateb, M. E. (2017). Isolation and anti-HIV-1 integrase activity of lentzeosides A-F from extremotolerant lentzea sp. H45, a strain isolated from a high-altitude Atacama Desert soil. *The Journal of antibiotics*, 70(4), 448.
- Wink, M. (2000). Interference of alkaloids with neuroreceptors and ion channels. In *Studies in natural products chemistry* (Vol. 21, pp. 3-122). Elsevier.
- Wink, M. (2003). Evolution of secondary metabolites from an ecological and molecular phylogenetic perspective. *Phytochemistry*, 64(1), 3-19.

- Wong, F. C., Yong, A. L., Ting, E. P. S., Khoo, S. C., Ong, H. C., & Chai, T. T. (2014). Antioxidant, metal chelating, anti-glucosidase activities and phytochemical analysis of selected tropical medicinal plants. *Iranian journal of pharmaceutical research: IJPR*, 13(4), 1409.
- Wongsrichanalai, C., Pickard, A. L., Wernsdorfer, W. H., & Meshnick, S. R. (2002). Epidemiology of drug-resistant malaria. *The Lancet infectious diseases*, 2(4), 209-218.
- Wu, T., He, M., Zang, X., Zhou, Y., Qiu, T., Pan, S., & Xu, X. (2013). A structure-activity relationship study of flavonoids as inhibitors of *E. coli* by membrane interaction effect. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1828(11), 2751-2756 (B).
- Wu, T., Zang, X., He, M., Pan, S., & Xu, X. (2013). Structure-activity relationship of flavonoids on their anti-*Escherichia coli* activity and inhibition of DNA gyrase. *Journal of agricultural and food chemistry*, 61(34), 8185-8190 (A).
- Xu, D. H., Huang, Y. S., Jiang, D. Q., & Yuan, K. (2013). The essential oils chemical compositions and antimicrobial, antioxidant activities and toxicity of three *Hyptis* species. *Pharmaceutical biology*, 51(9), 1125-1130.
- Xu, Y. Y., Wan, R. Z., Lin, Y. P., Yang, L., Chen, Y., & Liu, C. X. (2007). Recent advance on research and application of *Salvia miltiorrhiza*. *Asian J Pharmacodyn Pharmacokinet*, 7(2), 99-130.
- Yamagishi, T., Zhang, D. C., Chang, J. J., McPhail, D. R., McPhail, A. T., & Lee, K. H. (1988). The cytotoxic principles of *Hyptis capitata* and the structures of the new triterpenes hyptatic acid-A and-B. *Phytochemistry*, 27(10), 3213-3216.
- Yang, C. Q., Fang, X., Wu, X. M., Mao, Y. B., Wang, L. J., & Chen, X. Y. (2012). Transcriptional regulation of plant secondary metabolism. *Journal of integrative plant biology*, 54(10), 703-712.
- Yasukawa, K., Yu, S. Y., Tsutsumi, S., Kurokawa, M., & Park, Y. K. (2012). Inhibitory effects of Brazilian propolis on tumor promotion in two-stage mouse skin carcinogenesis. *Journal of Pharmacy and Nutrition Sciences*, 2(1).
- Yesilada, E., Gürbüz, İ., & Toker, G. (2014). Anti-ulcerogenic activity and isolation of the active principles from *Sambucus ebulus* L. leaves. *Journal of Ethnopharmacology*, 153(2), 478-483.
- Yuasa, Y., & Yuasa, Y. (2006). A Practical Synthesis of d- α -terpineol via markovnikov addition of d-limonene using trifluoroacetic acid. *Organic process research & development*, 10(6), 1231-1232.
- Yun, Y. S., Fukaya, H., Nakane, T., Takano, A., Takahashi, S., Takahashi, Y., & Inoue, H. (2014). A New Bis-seco-abietane Diterpenoid from *Hyptis crenata* Pohl ex Benth. *Organic letters*, 16(23), 6188-6191.
- Zaidan, L. B., Dietrich, S., & Schwabe, W. W. (1991). Effects of temperature and photoperiod on flowering in *Hyptis brevipes*. *Physiologia Plantarum*, 81(2), 221-226.
- Zellner, B. D., Amorim, A. C. L., Miranda, A. L. P. D., Alves, R. J., Barbosa, J. P., Costa, G. L. D., & Rezende, C. M. (2009). Screening of the odour-activity and bioactivity of the

- essential oils of leaves and flowers of *Hyptis Passerina* Mart. from the Brazilian Cerrado. *Journal of the Brazilian Chemical Society*, 20(2), 322-332.
- Zenk, M. H., & Juenger, M. (2007). Evolution and current status of the phytochemistry of nitrogenous compounds. *Phytochemistry*, 68(22-24), 2757-2772.
- Zhang, H., & Tsao, R. (2016). Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. *Current Opinion in Food Science*, 8, 33-42.
- Zhang, X. (2016). A composite herbicide. By: Zhang, Xudong. Assignee: Qingdao Haiyicheng Management Technology Co., Ltd., Peop. Rep. China. Patent No. CN 105494457.
- Zhang, X., Yu, Y., Cen, Y., Yang, D., Qi, Z., Hou, Z., Han, S., Cai, Z. & Liu, K. (2018). Bivariate Correlation Analysis of the Chemometric Profiles of Chinese Wild *Salvia miltiorrhiza* Based on UPLC-Qqq-MS and Antioxidant Activities. *Molecules*, 23(3), 538.
- Zheng, W., & Wang, S. Y. (2001). Antioxidant activity and phenolic compounds in selected herbs. *Journal of Agricultural and Food chemistry*, 49(11), 5165-5170.
- Zhong, G. X., Li, P., Zeng, L. J., Guan, J. I. A., Li, D. Q., & Li, S. P. (2009). Chemical characteristics of *Salvia miltiorrhiza* (Danshen) collected from different locations in China. *Journal of agricultural and food chemistry*, 57(15), 6879-6887.
- Zhong, J. S., Li, J., Li, L., Conran, J. G., & Li, H. W. (2010). Phylogeny of *Isodon* (Schrad. ex Benth.) Spach (Lamiaceae) and related genera inferred from nuclear ribosomal ITS, *trnL-trnF* region, and *rps16* intron sequences and morphology. *Systematic Botany*, 35(1), 207-219.
- Zoghbi, M. D. G. B., Andrade, E. H. A., da Silva, M. H. L., Maia, J. G. S., Luz, A. I. R., & da Silva, J. D. (2002). Chemical variation in the essential oils of *Hyptis crenata* Pohl ex Benth. *Flavour and fragrance journal*, 17(1), 5-8.
- Zollo, P. H., Biyiti, L., Tchoumbougnang, F., Menut, C., Lamaty, G., & Bouchet, P. H. (1998). Aromatic plants of tropical Central Africa. Part XXXII. Chemical composition and antifungal activity of thirteen essential oils from aromatic plants of Cameroon. *Flavour and Fragrance Journal*, 13(2), 107-114.
- Zuccolotto, T. (2017). Identificação de constituintes químicos de *Baccharis organensis* baker e avaliação das atividades biológicas dos extratos e frações das partes aéreas de *Baccharis aracatubaensis* Malag. e *Baccharis organensis* baker (Asteraceae).

Final considerations

Natural products remain a source of novel compounds for drug discovery due to their lower toxicity (Park et al., 2009). Most drugs on the market are plant-derived (Newman and Cragg, 2016) and the area of infectious diseases is largely dependent on natural products and their structures for sources of better treatment.

Related to *Hyptis* species, although a considerable number of papers founded in databases: 879 in SciFinder, 528 in Web of Science and 96 in SciELO, only 20% of *Hyptis sensu* Harley and Pastore (2012) have been studied. Most species were studied regarding their volatile oil composition; remaining poorly explored the polar constituents.

Furthermore, based on the articles published, it was possible to notice that these species are characterized by the presence of substances with promising pharmacological potential, mainly antimicrobial, antifungal, cytotoxic, anti-inflammatory, and anti-HIV, pointing to a great relevance of *Hyptis* to bioprospecting studies.

This study corroborated rosmarinic acid, chlorogenic acids, and nepetoidins as common constituents of Nepetoideae. Furthermore, the results corroborate the presence of these constituents also in *Hyptis* species. Lithospermic acid A and cirsimarinin were described for the first time in this study for *Hyptis*, both found in *H. radicans*. Fatty acids and triterpenes are the most abundant kind of apolar substances in *H. radicans* and *H. multibracteata*. This differs from what is most reported in the literature; first because the majority of reports focused on volatile oils and in this study, we analyzed also the apolar constituents of aerial parts extracts.

The present research also provides, for the first time, a comprehensive report on the antioxidant and cytotoxic activities of *Hyptis* species. EAP from *H. radicans* was the sample that presented the highest levels of total phenolic content, especially flavonoids, being also the sample with the high antioxidant activity with promising EC₅₀: DPPH (32.12 µg mL⁻¹), ABTS (5.04 µg mL⁻¹), Metal chelator assay (42.36 µg mL⁻¹), TBARS (40.46 µg mL⁻¹) and nonsite-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation (NS-Spe) with a EC₅₀ of 75.08 µg mL⁻¹. EE from *H. radicans* showed the high antioxidant activity for FRAP and ORAC with EC₅₀ of 6.01 and 2.68 µg mL⁻¹, respectively and has the highest amount of rosmarinic acid (17.64 mg g⁻¹). HMP from *H. radicans* showed the high antioxidant activity

in Site-Specific Hydroxyl Radical-Mediated 2-Deoxy-D-ribose Degradation (S-Spe) assay with EC₅₀ of 0.32 µg mL⁻¹ and has the highest content of chlorogenic acid derivatives.

Lithospermic acid A isolated from *H. radicans* and rosmarinic acid and nepetoidin B from *H. multibracteata*, were substances with better antioxidant activity. Nepetoidin B isolated from *H. multibracteata* had the best EC₅₀ (52.73 µg mL⁻¹) for anti-acetylcholinesterase activity. Regarding the results of cytotoxicity, **HP** from *H. multibracteata* induced the death of more than 80% of RAW 264.7 Cell Lines turning **HP** as an interesting phase as promising cytotoxic agent.

The anti-HIV-1 and antibacterial results from the present study lend support for further investigation of the bioactive constituents of *H. radicans* and *H. multibracteata* to validate the use of these plants in traditional medicine as antiviral and/or as antibacterial. **EE** and **HMP** of *H. radicans* showed anti-HIV-1 activity but contents of total phenolic compound are not the main sample feature to define anti-HIV activity but there is a correlation between the presence of rosmarinic acid and their anti-HIV activity. **HP** of *H. multibracteata* was the sample with better antibacterial activity but there was no correlation between phenolic contents and its activity. With the great results on biological activity it can be concluded that *H. radicans* is a promising phytotherapeutic.