

Article

First Study on Marine Heterobranchia (Gastropoda, Mollusca) in Bangka Archipelago, North Sulawesi, Indonesia

Adelfia Papu ^{1,2,*}, Nani Undap ¹, Nancy Armas Martinez ¹, Marco R. Segre ³,
Ivan Galton Datang ³, Rendy Robert Kuada ³, Marco Perin ³, Nathalie Yonow ⁴
and Heike Wägele ¹

¹ Centre of Molecular Biodiversity, Zoological Research Museum Alexander Koenig, 53113 Bonn, Germany

² Faculty of Mathematics and Natural Sciences, Sam Ratulangi University, Manado 95115, Indonesia

³ Coral Eye, Bangka Island, North Sulawesi, Indonesia

⁴ Department of Biosciences, Swansea University, Singleton Park, Swansea SA2 8PP, Wales, UK

* Correspondence: A.Papu@leibniz-zfmk.de

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Abstract: As ephemeral, benthic, secondary consumers usually associated with sessile coral reef organisms, marine heterobranchs are good indicators of the health of marine tropical habitats. Thus, marine Heterobranchia have recently become a major target for monitoring programs. For this work, an extensive survey was made in Bangka Archipelago, the first of its kind in this area. Bangka Archipelago is composed of small islands and the adjacent coastline of North Sulawesi. The substrate is dominated by biogenic reefs and volcanic rocks, thus forming highly diverse habitats. In total, 149 species were collected and/or photo-documented in September 2017 and September 2018, which represents the dry season. They can be assigned to the Cephalaspidea, Aplysiida, Pleurobranchida, Nudibranchia, and Eupulmonata. Thirty-three species are new to science, highlighting the lack of knowledge about the group and especially about this region. Our data provide a baseline for future monitoring surveys, as the anthropogenic pressures on Bangka Archipelago are increasing due to enhanced touristic activities, but also due to large scale environmental changes caused by previous mining operation activities.

Keywords: biodiversity; marine Heterobranchia; Bangka Island; North Sulawesi; monitoring

1. Introduction

Indonesia is one of the mega-diverse countries; however, surveys covering marine invertebrates in particular are very rare, these documents are often not open-access, and they are frequently in the Indonesian language [1]. The largest open-access study on Indonesian marine life is the Rumphius Biohistorical Expedition to Ambon in 1990, which has been published as a series of reports on numerous marine taxa in *Zoologische Mededelingen*, e.g., Naturalis Biodiversity Center [2]. This series also comprised studies on marine Heterobranchia, increasing our knowledge on this particular group considerably in an Indonesian region [3–6].

Marine heterobranchs are interesting in many ways. Many of them have reduced or lost their shells and developed alternative defense or antifouling systems, by taking up natural compounds from their food, or by *de novo* synthesis. These compounds are of interest as potential drug leads for medical application [7–10]. The loss of the shell allows sea slugs to develop stunning body shapes and coloration, and they are therefore well-known to snorkelers, divers, and underwater photographers [11]. The diversity of heterobranchs serves as an indication of the diversity of other metazoan life forms such as sponges, cnidarians, ascidians and bryozoans, as they provide nutrition with high food specificity on the species level. These filter feeders or consumers on lower levels are

highly affected by changes in the environment, thus also affecting sea slug communities (e.g., [12–14]).

Burghardt et al. [15] recorded molluscan diversity for the first time in North Sulawesi (Bunaken National Park, BNP), including heterobranch taxa. More recently, several publications on heterobranch diversity have been published from North Sulawesi, including Bunaken National Park, Lembah Strait and Sangihe Island [1,16–18], which renders this area as one of the best investigated places in Indonesia with regards to this taxon. Our study in Bangka Archipelago increases our knowledge on the biodiversity of heterobranchs in North Sulawesi in an area that has never been documented before.

Bangka Archipelago comprises small islands located in the northeast of North Sulawesi, thus lying in between the recently well-studied area of Bunaken National Park, and the famous Lembah Strait, whereas Sangihe Island lies approximately 200 km to the north. All studies have shown that these areas differ in heterobranch composition due to the differing habitat compositions [16–18]. Bangka Island exhibits biogenic reefs, volcanic rocks, and a few areas with mangroves and sea grass meadows. The mainland facing Bangka Island is of volcanic origin with smaller fringing reefs. The central area of the archipelago is shallow, while the outer slopes drop down to more than 1000 m depth. The islands are covered by lush vegetation and are framed by fringing reefs, alternating with few mangroves and volcanic cliffs. The islands are home to some villages and a few resorts, of which the oldest one was built in 1987. Bangka, the largest island of this archipelago (approximately 48 km²), has a resident population of nearly 2500 inhabitants and only six resorts [19]. Therefore, this island is less affected by tourists than the more famous Bunaken National Park (BNP), Lembah Strait, or other Indonesian islands, such as Bali. However, implementation of mining operations began in 2012 in coastal areas, leading to deforestation of mangroves and broad land erosion within only 5 years. Unfortunately, there are no detailed data available for the diversity of marine organisms and coral reef health in Bangka Island before mining started; therefore, the impacts of the mining in areas close by cannot be assessed. This study therefore aims to serve as a baseline for future monitoring surveys concerning further environmental changes within the Bangka Archipelago.

2. Materials and Methods

Material was collected during the dry season, in September 2017 and September 2018. In total, 28 dives were performed during the day and 10 during the early night, with 3–5 divers and 60–120 minutes per dive in 19 different localities around Bangka Island and the adjacent mainland (Figure 1A,B). A depth range from the eulittoral down to ca. 20 m was covered. Table 1 summarizes the specific conditions of the various diving localities. Various substrates, such as macro algae and coral rubble, were collected for further analyses in the laboratory. Additionally, several hours were spent collecting and photo-documenting while snorkeling. In a few sites, only underwater photographs were taken of sea slugs for species recording. These species are included in our results on species numbers, but not in our overall specimen numbers.

Each collected specimen was documented with an Olympus TG5 under water (when possible) and additionally in the laboratory, provided with a unique identifier (abbreviation of name, year, location, and number of specimen), and preliminarily identified with the help of available literature (e.g., [3–5,11,15,16,20–27]). Gosliner et al. [24] depict many undescribed species. We used their species number for our internal identifier when we assigned one of our specimens to these undescribed species. However, we applied only a letter (e.g., sp. a, or sp. b) when a specific assignment to any undescribed species in this identification book was not possible and/or when our barcodes checked against the reference database in NCBI (National Center for Biotechnology Information) provided no hit. Validity of names was checked in the World Register of Marine Species [26]. Collected specimens were preserved in 96% EtOH and/or in formaldehyde/seawater.

Those families with documented cryptic speciation (e.g., Chromodorididae, Phyllidiidae) and a few other rare species of interest (e.g., *Moridilla*) were analyzed by using barcoding methods to verify

identification. Additionally, small specimens that were difficult to identify (if at all) or where no clear assignment was possible were also barcoded.

All specimens are registered in the collection of Sam Ratulangi University according to the year under the numbers SRU2017/01 and SRU2018/01. Metadata of each individual are documented in the database Diversity Collection (Diversity Workbench) using the data brokerage service of the German Federation for Biological Data (<https://www.gfbio.org/>) [28]. Data are publicly available at www.gfbio.org for browsing and the archived data can be downloaded at <https://doi.org/10.20363/heterobranchia-bangka-prj-1.1>

3. Results

In total, 484 specimens comprising 149 species of marine heterobranchs were collected in the two sampling events in 2017 and 2018. Five species can be assigned to Cephalaspidea, two to Aplysiida, 15 to Sacoglossa, two to Pleurobranchida, and 124 to Nudibranchia. Additionally, one species is a member of the eupulmonate taxon Onchidiidae. Thirteen species of the 149 were only recorded by photo-documentation in 2018 without collection. Thirty-three species cannot be assigned to any described species and are considered new species. Table 2 summarizes available information on all collected and photo-documented specimens, and also includes information about taxa authorities (these are therefore not mentioned in the text). Families and species are listed in the same order as in the text below. The table also provides information about the presence of the particular species in BNP, Sangihe, and Lembeh Strait according to the recently published surveys [16–18] to highlight those species that are only recorded from our study area. All species listed in Table 2 are also depicted by at least one specimen, together with the specimen identifier, if the animal was collected, and are available for further investigations. In the following sections, species and specimens are discussed and compared to other studies from North Sulawesi [1,16–18].

3.1. Animal Collections

3.1.1. Cephalaspidea (Five Species in Five Genera Belonging to Four Families)

The number of cephalaspidean species is low in comparison to the number from BNP (5 vs. 16).

Haminoeidae: *Haminoea* sp. (Figure 2A), collected at a depth of 7 m, has a translucent orange shell with orange and yellow marks, resembling *Haminoea* sp. 3 from Gosliner et al. [24].

Colpodaspidae: *Colpodaspis thompsoni* (Figure 2B; length 2–5 mm) was most often found crawling in the coral rubble at 1.5–9.2 m depth. With only five specimens, it was not as common as in BNP.

Aglajidae: Only two specimens of *Chelidonura amoena* (Figure 2C), a common species in sandy reefal habitats, were found in one site only. One specimen of *Odontoglaja* (Odsp.a18Ba-1, Figure 2D, E) was found in the coral rubble. It looks similar to *Odontoglaja guamensis* but lacks the distinct color pattern on the dorsum. Gosliner et al. [24] depict several *Odontoglaja* spp. with similar background patterns; however, the dorsal color patterns and especially the intensity of the color differ in all of these. We cannot assign our specimen to any of these taxa.

Gastropteridae: Two specimens of *Sagaminopteron psychedelicum* (Figure 2F) were found in coral rubble.

3.1.2. Aplysiida (= Anaspidea) (Two Species in Two Genera Belonging to One Family)

Aplysiidae: Members of this family mainly forage on algae in shallow water. Four specimens of probably *Aplysia* cf. *nigrocincta* (Figure 2G) (Appa18Ba-3, with a black line along the parapodia) were found under coral rubble. Recently, Golestani et al. [29] revised *Aplysia parvula* and identified 10 different lineages within this species. They resurrected the name *A. nigrocincta* Martens, 1880 for specimens from the Philippines and Papua New Guinea. However, molecular analyses still need to be performed on our material for correct assignment. The same holds true for the specimens

described in Eisenbarth et al. [16] as *A. parvula*. Two specimens of *Stylocheilus striatus* (Figure 2H) were found under coral rubble.

3.1.3. Sacoglossa (15 Species in Four Genera Belonging to Three Families)

Oxynoideae: *Lobiger* sp. 1 (Figure 3A), mimicking the green alga *Caulerpa*, has elongate leaf-like cerata and a green bubble-like shell. Our two specimens lack the blue lines typical for *L. viridis* (as depicted in Gosliner et al. [24]) and seem to be undescribed.

Hermaeidae: According to Bouchet et al. [30], the family Caliphyllidae is united with this family, and we follow their systematics. Two *Cyerce* species were collected: one specimen of *C. bourbonica* (Figure 3B) shows the characteristic opaque white body, dark brown marks along the cerata and rhinophores, and violet to pink color at the tip of cerata. The other species can be assigned to *C. nigra* (Figure 3C), based on the typical color and color pattern of the cerata.

Plakobranchidae: Only the genera *Elysia* and *Thuridilla* are present in our collections. *Elysia asbecki* (Figure 3D), *E. marginata* (Figure 3E), *E. cf. nigropunctata* (Figure 3F), *E. pusilla* (Figure 3G), and four unidentified *Elysia* species were sampled from various algae in the coral rubble or highly structured micro-habitats in the coral reef. *E. cf. nigropunctata* (Elni18Ba-1) was collected from a sponge, probably feeding on its epiphytic green algae. Four undescribed *Elysia* spp. (*Elysia* sp. 23, *Elysia* sp. 27, *Elysia* sp. a, and *Elysia* sp. b) in our collection indicate the need for further investigation with regard to this group: *Elysia* sp. 24 (Elsp24-18Ba-1; Figure 3H) and *Elysia* sp. 27 (Elsp27-18Ba-1; Figure 4A) are depicted in Gosliner et al. [24], whereas *Elysia* sp. a (Elsp.a18Ba-1; Figure 4B) and *Elysia* sp. b (Elsp.b18Ba-1; Figure 4C) are featured here for the first time. These species were not found previously in North Sulawesi. *Elysia* sp. 24 has numerous white dots on its body, the rhinophores have three white bands, and the light green parapodia are without a line at the edge. *Elysia* sp. 27 (Elsp27-18Ba-1), collected from *Halimeda*, is pale green with brown dots and numerous opaque white spots on the dorsum and rhinophores; an orange line runs along the edge of the parapodia. *Elysia* sp. a (Figure 4B) looks like *Elysia* sp. 24 [24] but has a red line along the mantle margin. *Elysia* sp. b (Figure 4C) has an opaque white body color with small green dots covering the dorsal notum and parapodia. The rhinophores are relatively short, with papillae, and a brown line running along the margin of the parapodia crosses the foot on the ventral side. Four *Thuridilla* species were collected, including *T. carlsoni* (Figure 4D), *T. flavomaculata* (Figure 4E), *T. vataae* (Figure 4F), and *T. cf. gracilis* (Figure 4G, H), the latter being the most common *Thuridilla* species in the study area. According to Martín-Hervás et al. [31], this species consists of a complex of 14 species. We cannot assign our specimens to any of these cryptic species, but we can provide more details on the coloration especially of the rhinophores that might help in future assignment of our material. Our specimens from Bangka have a black to dark brown background with narrow white to light green longitudinal lines, which are broader in certain areas, implicating transversal white interrupted patches. The parapodial edge exhibits a narrow orange line, but no blue spots on the outside of the parapodia. Some specimens have white rhinophores with a black tip apical to an orange band, and others have rhinophores only with orange tips (Figure 5A–E).

3.1.4. Pleurobranchida (Two Species in One Genus Belonging to One Family)

Pleurobranchida (Pleurobranchomorpha) are carnivorous, nocturnal animals usually large in size. We mainly observed the specimens during night dives.

Pleurobranchidae: *Pleurobranchus forskalii* (>200 mm) was only found during the night. The tubercles of this species (Figure 6A) are compound and brownish in color. The specimens of *P. peronii* (Figure 6B), also only found during night dives, have only simple tubercles.

3.1.5. Nudibranchia (124 Species)

a. Nudibranchia, Doridina (83 species in 29 genera belonging to eight families)

Doridina are represented in our study by nine families of the 17 listed in WoRMS (World Register of Marine Species) [26].

Hexabanchidae: One specimen of *Hexabanchus sanguineus* (Figure 7A) was found in coral rubble. It was a juvenile mimicking a *Hypselodoris maculosa* in coloration. Molecular analyses confirmed its assignment to *H. sanguineus*.

Polyceridae: The family is represented by 13 species in three genera. Compared to BNP and Sangihe, the genus *Nembrotha* is relatively common in Bangka Archipelago, with seven species records. Five specimens of *N. chamberlaini* (Figure 7B) and five specimens of *N. cristata* (Figure 7C) were collected at different sites and depths. *N. kubaryana* (Figure 7D), encountered quite often, is represented in our samples by 13 specimens, collected at 10 different sites. It exhibits some variation with regard to the presence of orange patterns or lines along the body. Two mating *N. lineolata* (Figure 7E) were only recorded by photographs. *N. milleri* (Figure 7F) was crawling on brown sponges near green tunicates. Two specimens of *N. mullineri* (Figure 7G) were collected only during two night dives, one in 2017 and one in 2018. In both cases, the animals were crawling on sand. One undescribed *Nembrotha* (Nesp1_17Ba-1; Figure 7H), similar to *Nembrotha* sp. 1 [24], was collected on a greenish tunicate; however, our specimen has bluish rhinophores and gills. *Tambja gabriellae* (Figure 8A) and *T. morosa* (Figure 8B) were collected at 8–17 m depth. The monotypic genus *Gymnodoris* within the Gymnodorididae comprises many similar species with at least 60 undescribed species [24]. The single specimen of *G. aurita* (Figure 8C) was identified only by photo-documentation. *Gymnodoris tuberculosa* (Figure 8D; 3 mm long) was crawling in coral rubble at 1 m depth. Two specimens of two *Gymnodoris* spp. (Figure 8E, F) are undescribed and depicted in Gosliner et al. [24] as *Gymnodoris* sp. 20 and *Gymnodoris* sp. 25, respectively.

Goniodorididae: Members of this family are quite rare and not represented by many species in North Sulawesi [16]. Some undescribed *Goniodoris* were recorded by Gosliner et al. [24]. We collected only one specimen (Figure 8G), which is very similar to *Goniodoris* sp. 7 in Gosliner et al. *Trapania armilla* (Figure 8H) was collected on a gray sponge in shallow water, whereas *T. safracornia* (Figure 9A) was collected in deeper areas.

Aegiridae: Both acknowledged aegirid genera were present in our collections: *Aegires* sp. (Figure 9B), depicted in Gosliner et al. [24] as *Aegires* sp. 7, and the “banana sea slug” *Notodoris minor* (Figure 9C) (the latter only by photo-documentation).

Discodorididae: The large and easily identifiable *Asteronotus cespitosus* (Figure 9D) was only recorded by photo-documentation. *Asteronotus mimeticus* (Figure 9E) is always associated with a phyllospongian sponge. It usually sits underneath and was more often observed during the night. The coloration of the sea slug matches perfectly with the color and structure of the sponge surface (Figure 9F). Two specimens of *Atagema intacta* (Figure 9G) were collected during night dives, one from sand substrate, the other on a sponge. Two specimens of *Discodoris cebuensis* (Figure 9H) were also collected during a night dive. *Halgerda batangas* (Figure 10A) is a common species in this location, while in other areas (e.g., BNP) the species is quite rare. The same is true for *Halgerda c.* (Figure 10B), represented by two specimens. *Jorunna funebris* (Figure 10C) was common in mangroves, and not all of the about 20 specimens seen were collected. *Paradoris liturata* (Figure 10D) is a member of the Discodorididae that mimics *Phyllidiella pustulosa*, illustrated in Figure 10E. *Paradoris liturata* has tubercles on the notum with black lines in between, as well as black rhinophores, similar to *P. pustulosa*. When the gills in *P. liturata* are retracted into the gill pockets, the opening looks like the anus of a *Phyllidiella* species. *Platydoris sanguinea* (Figure 10F) was found on coral rubble at 5.5 m depth.

Chromodorididae: This family is represented in our study by 263 specimens, which could be assigned to 37 species. *Ceratosoma tenue* (Figure 10G) was only identified by photo-documentation. Eight species of the genus *Chromodoris* are present in our collection. *Chromodoris diana* (Figure 10H), a very common species in BNP, was only represented by two specimens, collected at depths in the range of 13–15 m. Of all chromodorids, *C. annae* (Figure 11A, B) is the species with the highest specimen numbers and distribution (Table 2); rhinophores and gills vary in color from light yellow to a rather unusual orange. *Chromodoris elisabethina* (Figure 11C) is recorded only by photographs from Sempini. *Chromodoris lochi* (Figure 11D, E), the second most common *Chromodoris* species, is represented by two color morphs: rhinophores and gills are either pink or pale yellow, and the mantle

is always translucent white. Molecular barcoding has confirmed both their identities as *C. lochi*. The same two color morphs are also found in BNP [16]. One specimen of *C. magnifica* (Figure 11F) with the typical whitish background was found on sand underneath coral rubble. Another specimen was collected that looked in general very similar to *C. magnifica* with a discontinuous dark line in the middle of the notum, but with a distinct bluish body. According to molecular barcoding, both genes (*CO1* and *16S*) show a 99% similarity with sequences from NCBI assigned to *C. quadricolor* (Figure 11G). This would then indicate another record in the Indo-Pacific Ocean of a species that is primarily distributed in the Red Sea. Two specimens of *C. strigata* (Figure 11H) were collected at depths of 7–14 m, one during a night dive crawling on the green alga *Halimeda*. Only two specimens of *C. willani* (Figure 12A) were found close to each other at 8 m depth, another species much more common in BNP. Four specimens of *Doriprismatica atromarginata* (Figure 12B, C) with sizes of 17–74 mm were collected from different dive sites. In one of these dive sites, we observed a yellow flatworm *Pseudoceros* sp. (Figure 12D), which clearly mimics *D. atromarginata*. Only one specimen of *D. sibogae* (Figure 12E) was found, differing from *D. atromarginata* in gill color. *Glossodoris cincta* is a common species and widespread in the Indo-Pacific, and is also recorded from BNP, Sangihe, and Lembeh Strait. However, there are several species with very similar coloration [32]. We assign two specimens to this species (Figure 12F). One was collected at 6 m depth during a night dive underneath coral rubble, and one at 1 m depth. Both animals had a yellowish to green band followed by a black and then a white margin on the mantle (typical feature of *G. cincta*). *Glossodoris rufomarginata* (Figure 12G) was identified only by photo-documentation. It is described in the literature as having an orange-brown spackled pattern on the notum, brown marginal band, and white submarginal band [24]. The genus *Goniobranchus* was represented by six different species. *Goniobranchus coi* (Figure 12H) was only collected in 2017 during a night dive. It was not recorded from BNP [16] or Sangihe [18]. *Goniobranchus fidelis* (Figure 13A) is a smaller member of the genus, and our single animal was only 13 mm in length, collected at 17 m depth. *Goniobranchus geometricus* (Figure 13B) is more common in North Sulawesi, and we collected six specimens at 5–15 m depth. *G. kuniei* (Figure 13C) and *G. reticulatus* (Figure 13D) were collected at deeper than 20 m depth. Yonow [3] outlines the problems of correct identification of this species, and misidentification of *G. inopinata* and even *G. tinctorius* cannot be excluded: *G. tinctorius* usually has a yellow band along the edge of the notum [33], which is not present in our specimens.

In contrast to BNP, where only five species have been recently depicted [16], 12 species of *Hypselodoris* are recorded in Bangka Archipelago: *H. apolegma* (Figure 13F), *H. bullockii* (Figure 13G), *H. cerisae* (Figure 13H), *H. decorata* (Figure 14A), *H. emma* (Figure 14B), *H. iacula* (Figure 14C), *H. lacuna* (Figure 14D), *H. maculosa* (Figure 14E), *H. maridadilus* (Figure 14F), *H. zephyra* (Figure 14G), *H. tryoni* (Figure 14H) and *Hypselodoris* sp. a (Figure 15A). Many of the species are easily recognized by their distinct coloration and are not discussed in detail below. A recently described new species, *H. iba*, is very similar to *H. bullockii*; however, our animal, which was only photo-documented (Figure 13G), shows the features typical of *H. bullockii* also found in Ambon [5] and BNP [16]. However, confirmation of the identification of this individual would require specimens and possibly molecular analyses. One member of the newly described species *H. cerisae* (Figure 13H) [27] was collected in front of Coral Eye, and molecular analyses verified the identification based on 99% similarity of our specimen with the sequence of *H. cerisae* CASIZ 178350 deposited in NCBI. Our specimen is pale brown with some purple pigment concentrated at the anterior mantle and foot margin; white dots are aligned on three longitudinal brown lines on the notum; brownish diagonal lines are present on the posterior foot; there are nine gill branches. Externally, *H. decorata* (Figure 14A) is very similar to *H. maculosa* (Figure 14E). According to recent studies [27], they are closely related and form a clade with *H. juniperiae*. *H. decorata* can be distinguished from *H. maculosa* by the three reddish rhinophoral bands, versus only two reddish rings. Our specimen of *H. decorata* has these three rhinophoral rings, as well as the other typical color features, e.g., the white dots on the broad brown margin of the mantle and longitudinal lines on the middle part of the mantle with brown dots. Our single specimen of *H. iacula* (Figure 14C) has a light honey-colored body, with a brown band around the mantle margin, violet at the foot margin, and a white net-like pattern on the notum and top of the foot. Eight

gill branches arise from a relatively high gill pocket. Two specimens are assigned to *H. lacuna* (Figure 14D), whose name is based on the translucent areas on the notum that resemble holes in the body wall [27]. Both specimens have a white mantle, with many translucent patches and two large translucent circles posterior to the rhinophores with two others in front of the gills; further characteristics are blue dots on the lateral margin of the mantle, and white gills and rhinophores with reddish apices. The body of *Hypselodoris* sp. a (Figure 15A) is cream, as well as the rhinophores, which are more translucent at the base and with one orange ring in the cream area. Red dots spread along the margin of the notum and a few along the midline of the notum. Externally, the species is very similar to *Thorunna australis* and *Hypselodoris maculosa*. However, *Hypselodoris* sp. a lacks contrasting pigment in the middle of the notum, whereas *H. maculosa* shows dots all over the notum. Two specimens of *Mexichromis aurora* (Figure 15B) were sitting next to each other on corals at 8.6 m depth, but only one was collected. *M. trilineata* (Figure 15C) was only identified by photo-documentation.

One small animal (Dorid18Ba-1; Figure 15D) was found in Yellow Coco at a depth of 20 m. It is characterized by a pale blue body color with an orange mantle rim, orange gills and rhinophores, as well as orange patches on the dorsal notum and tail. The body shape resembles some goniodorids. However, it could be identified as a juvenile of the species *Miamira magnifica*. One further small member of probably the same genus, *Miamira* sp. a (Figure 15E) (10 mm), was found at a depth of 4 m. It is reddish to brown in color, covered with darker spots and many tiny white dots on the mantle; the edge of the mantle is cream with brown dots. The rhinophores are translucent cream with white lines and an orange hue at the tip, whereas the translucent gills exhibit brown and white patches. *Verconia simplex* (Figure 15F), 6 mm in size, was sitting on black sponges at 5.2 m depth. Both specimens of *Thorunna furtiva* (Figure 15G) were found close to each other on a stone, covered by encrusting red sponge, and were observed to have vibrating gills, a typical character of the genus. *Dendrodoris nigra* (Figure 15H) is the only member of the family Dendrodorididae that was collected. It is a juvenile of only 6 mm length and was found crawling out of the coral rubble sorted in the laboratory.

Phyllidiidae: Members of this family are probably the most common sea slugs in tropical waters. More than 100 specimens were collected in the Bangka Archipelago, assigned to 16 species and three genera, *Phyllidia*, *Phyllidiella*, and *Phyllidiopsis*. *Phyllidia* cf. *babai* (Figure 16A) was found in Batu Belah and was only identified by photo-documentation. Eight specimens of *Phyllidia coelestis* (Figure 16B) were collected at 1–20 m depth. The other *Phyllidia* species represented in our collection are *P. elegans* (Figure 16C), *P. exquisita* (Figure 16D), *P. ocellata* (Figure 16E), and *P. picta* (Figure 16F). *Phyllidia picta* often co-occurred at the same locations as *P. ocellata*. *Phyllidia varicosa* (Figure 16G, H) was probably the most common *Phyllidia* species with 16 specimens collected at nine sites, ranging between 23 and 85 mm in size and between 2 and 20 m in depth. One specimen originally assigned to *Phyllidia picta* (Phsp18Ba-2; Figure 17A), based on external coloration, clusters with several other specimens from various regions of North Sulawesi in a separate clade, distinct from the *P. picta* clade. It therefore represents a new species with very similar appearance to *P. picta* (Papu et al., in preparation). Four species of *Phyllidiella* collected in Bangka Archipelago comprise *Phyllidiella* cf. *annulata* (Figure 17B), *P. lizae* (Figure 17C), *P. nigra* (Figure 17D) and *P. pustulosa* (Figure 17E, F). Only one specimen of *Phyllidiella* cf. *annulata* (Figure 17B) was found in Talisei Island, a collection site with strong tidal currents and ocean waves, as well as small compact coral formations. *Phyllidiella nigra* (Figure 17D) was collected from mangrove roots. It is characterized by the dark gray hyponotum and foot, and black notum with single pale pink round tubercles. *Phyllidiella pustulosa* (Figure 17E, F) is the most common species of the genus *Phyllidiella* and also within Phyllidiidae. However, this species was shown to represent a complex of several species [25], which are very difficult to distinguish by external characteristics alone. Figure 17G and H depict a specimen which we removed from the substrate to show feeding marks on the sponge. *Phyllidiopsis annae* (Figure 18A, B) is a small species, and our four specimens ranged in length from 3 to 12 mm. Further collected *Phyllidiopsis* species are *P. cf. burni* (Figure 18C), *P. krempfi* (Figure 18D, E), and *P. xishaensis* (Figure 18F).

b. Nudibranchia, Cladobranchia (42 species in 22 genera belonging to 12 families)

Arminidae: Six different species of *Dermatobranchus* were collected during the two studies: *D. caeruleomaculatus* (Figure 19A), *D. rudmani* (Figure 19B), *D. pustulosus* (Figure 19C), and three undescribed *Dermatobranchus* species (Figure 19D–F). Most of them were collected in 2018 close to Coral Eye under coral rubble.

Proctonotidae: One *Janolus* species (Figure 19G) exists only as a photo-documentation in our collection. It is already depicted in Gosliner et al. as *Janolus* sp. 1.

Bornellidae: Several individuals of *Bornella anguilla* (Figure 19H) (of which only two were collected) were found crawling on encrusting corals and tunicates, and some were mating at approximately 8 m depth. One *Bornella* specimen was found in coral rubble at 1 m depth, which resembles most *B. dotoides* when comparing the shape of the cerata (Figure 20A) [3].

Tethydidae: *Melibe bucephala* (Figure 20B) was found together with *B. stellifera* in the collected coral rubble. *Melibe engeli* (Figure 20C; length of 34 mm) is a well-camouflaged species due to its transparent body. The species is recorded to be associated with algae of the genus *Padina* or *Achantophora* [24]; however, our animal was collected from an encrusting sponge.

Dotidae: The brownish-cream to olive-colored species *Doto ussi* (Figure 20D) was always found at the base of the hydrozoan *Aglaophenia cupressina*.

Tritoniidae: Two undescribed species were collected, both at the same site, with many different soft coral species close by. The dorsal appendages of the undescribed *Marionia* species (Figure 20E, F) look like green algae, and the animal is very similar to *M.* sp. 2, depicted in Gosliner et al. [24]. The dorsal appendages of the undescribed *Tritonia* sp. (Figure 20G) resemble polyps of soft corals. The animal is similar to *T.* sp. 3 depicted by Gosliner et al. [24].

Flabellinidae: *Coryphellina exoptata* (Figure 20H), *C. rubrolineata* (Figure 21A), and two undescribed species (*Flabellina* sp. 2, *Flabellina* sp. 3) are members of the family Flabellinidae. The color of *Flabellina* sp. 2 (Figure 21B) (also depicted in Gosliner et al. [24]) is similar to *C. rubrolineata*, but our animal has only one ring around each cerata, and no lines in the middle and margin of the body. The specimen of *Flabellina* sp. 3 (Figure 21C, similar to *F.* sp. 3 in Gosliner et al. [24]) has a rather translucent body with a single red line in the middle of the body. The white cerata carry a few red dots, and the rhinophores are translucent with a tinge of white in the middle part.

Samliidae: *Samla riwo* (Figure 21D) was collected from a non-specific substrate.

Eubranchidae: *Eubranchus* sp. 22 (Figure 21E) was only found once, with one specimen on a hydrozoan of the family Plumulariidae [24].

Trinchesiidae: The conspicuous *Trinchesia yamasui* (Hamatani, 1993) (Figure 21F) was only recorded by photo-documentation. It is very similar to the original and subsequent descriptions [21,34,35]. The animal is grayish, with a black head followed by a violet band in front of the rhinophores. The cerata are black to gray, with a subapical orange ring, followed by a black apex. Two specimens of an undescribed *Cuthona* species (Figure 21G) depict a uniformly orange body color with the oral tentacles and rhinophores showing a darker orange on the distal parts, followed by a translucent apical part. The specimens resemble *Cuthona* sp. 57 in Gosliner et al. [24]. One unidentified aeolid specimen (Aeol18Ba-1; Figure 21H) was collected from coral rubble and is characterized by a pale white body and cerata with the digestive gland shining through in a darker shade.

Facelinidae: This cladobranch family is represented by the highest number of specimens (99) and species (20) in this study. Several new species were also recorded. One specimen probably represents an undescribed *Antonietta* species (Figure 22A). It measured only 2 mm alive and has an opaque white body, violet cerata with a dark violet ring on each cerata, and yellow rhinophores. One of the most common facelinid species is *Caloria indica* (Figure 22B), which we collected at depths of 5–14 m. An unidentified specimen probably belonging to the genus *Cratena* (Figure 22C) was collected during a night dive at 4 m depth. It is characterized by the orange color of the jaws, a typical character of the genus. Many members of *Favorinus* collected during this study were observed feeding on other nudibranch egg masses. *F. japonicus* (Figure 22D) was collected from the macroalga *Udotea* while laying eggs. *Favorinus* sp. (Figure 22E) was also found on *Udotea*, feeding on a large nudibranch egg mass. Our animal is similar to *F.* sp. 1 in Gosliner et al. [24]. *F. tsuruganus* (Figure 22F) was

collected from coral rubble. Two specimens of an undescribed *Moridilla* species (Figure 22G, H) were collected in 2017 and 2018. Both of these specimens are white, with white and yellow on the oral tentacles. The cerata are shorter than the rhinophores, which carry papillae along the posterior side. Molecular analyses clearly distinguish these specimens as a separate species and group them as sister taxa to a new *Moridilla* species recently described from Bunaken National Park, *M. jobeli* [36]. The animals exhibit an interesting behavior of thrusting and wriggling the cerata when disturbed, and swimming with lateral movements. Three undescribed *Noumeaella* species were collected: one looks very similar to *Noumeaella* sp. 2 depicted in Gosliner et al. [24] (Figure 23A), the second resembles *Noumeaella* sp. 3 (Figure 23B) [24], and the third is similar to *Noumeaella* sp. 13 (Figure 23C) [24]. The genus *Phyllodesmium* was represented in our collection by eight species. *P. briareum* (Figure 23D) is usually found on the soft coral *Briareum*, where the cerata are very well camouflaged by the tentacles of the octocoral. *P. cf. crypticum* (Figure 23E) was found while laying an egg mass at the base of an unidentified xeniid soft coral species. Three specimens of *P. lizardense* (Figure 23F) were collected at various depths, down to 10 m. One large *P. longicirrum* (Figure 23G; 125 mm) was collected on the sand flat at Efrata site at 25 m depth. It usually feeds on the soft coral *Sarcophyton*, which was also quite common on that sandy flat. *P. magnum* (Figure 22H) and *P. parangatum* (Figure 24A) were collected from members of the family Xeniidae, while *P. pecten* (Figure 24B) was extracted from coral rubble. Two specimens of *P. poindimiei* (Figure 24C, D) were collected from an unspecific substrate. *Pteraeolidia semperi* (Figure 24E) is one of the largest facelinid species and was the most common cladobranch species observed, due to an aggregation of more than 100 specimens in shallow water (Sempini), probably for mating.

3.1.6. Eupulmonata (One Species in One Genus Belonging to One Family)

Onchidiidae: One member of this family, an undescribed *Peronia* species (confirmed by barcoding) (Figure 24F), was collected in the mangrove. It lives in tidal areas and was nearly 40 mm in size. The notum is a cream background color, with pale green and orange patches. Small cream-colored tubercles are spread over the notum.

Figure 25 provides an overview of the species numbers in the various sampling sites, taking into consideration the number of sampling events per site. The high sampling effort at Coral Eye contributed considerably to overall species numbers in this area.



Figure 1. Location of the study area: (A) depiction of the 19 diving sites at Bangka Archipelago and mainland. Insert showing the region of North Sulawesi with Bangka Archipelago. (B) Indonesia and Sulawesi, with the blue circle indicating the close-up area in A.

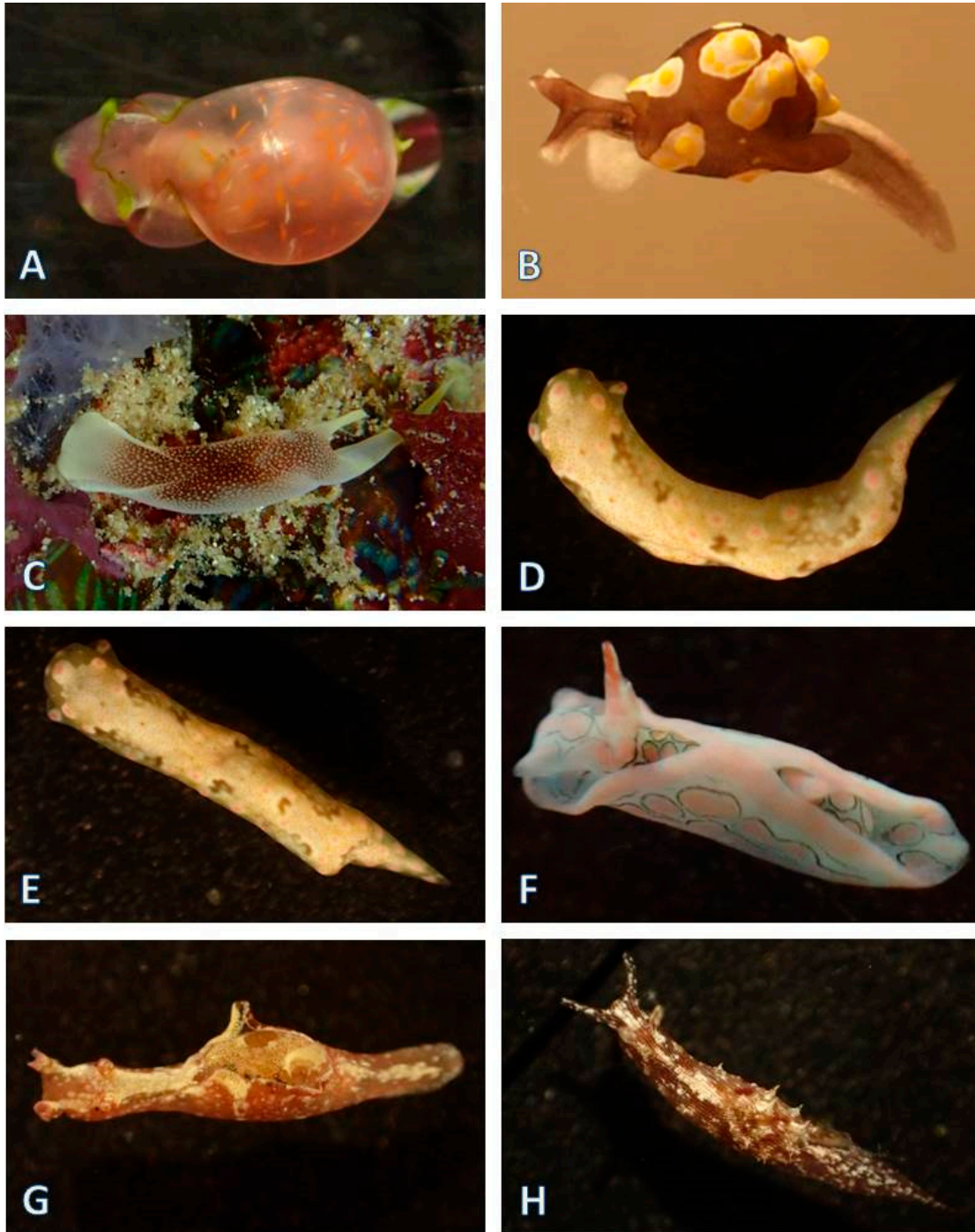


Figure 2. Cephalaspidea and Aplysiida: (A) *Haminoea* sp. (*Haminoea* sp. 3 in Gosliner et al. [24]: 34), Hasp18Ba-1; (B) *Colpodaspis thompsoni*, Coth18Ba-2; (C) *Chelidonura amoena*, Cham17Ba-1; (D, E) *Odontoglaja* sp. (*Odontoglaja* sp. a), Odsp.a18Ba-1; (F) *Sagaminopteron psychedelicum*, Saps18Ba-2; (G) *Aplysia* cf. *nigrocincta*, Appa18Ba-1; (H) *Stylocheilus striatus*, Stst18Ba-2.

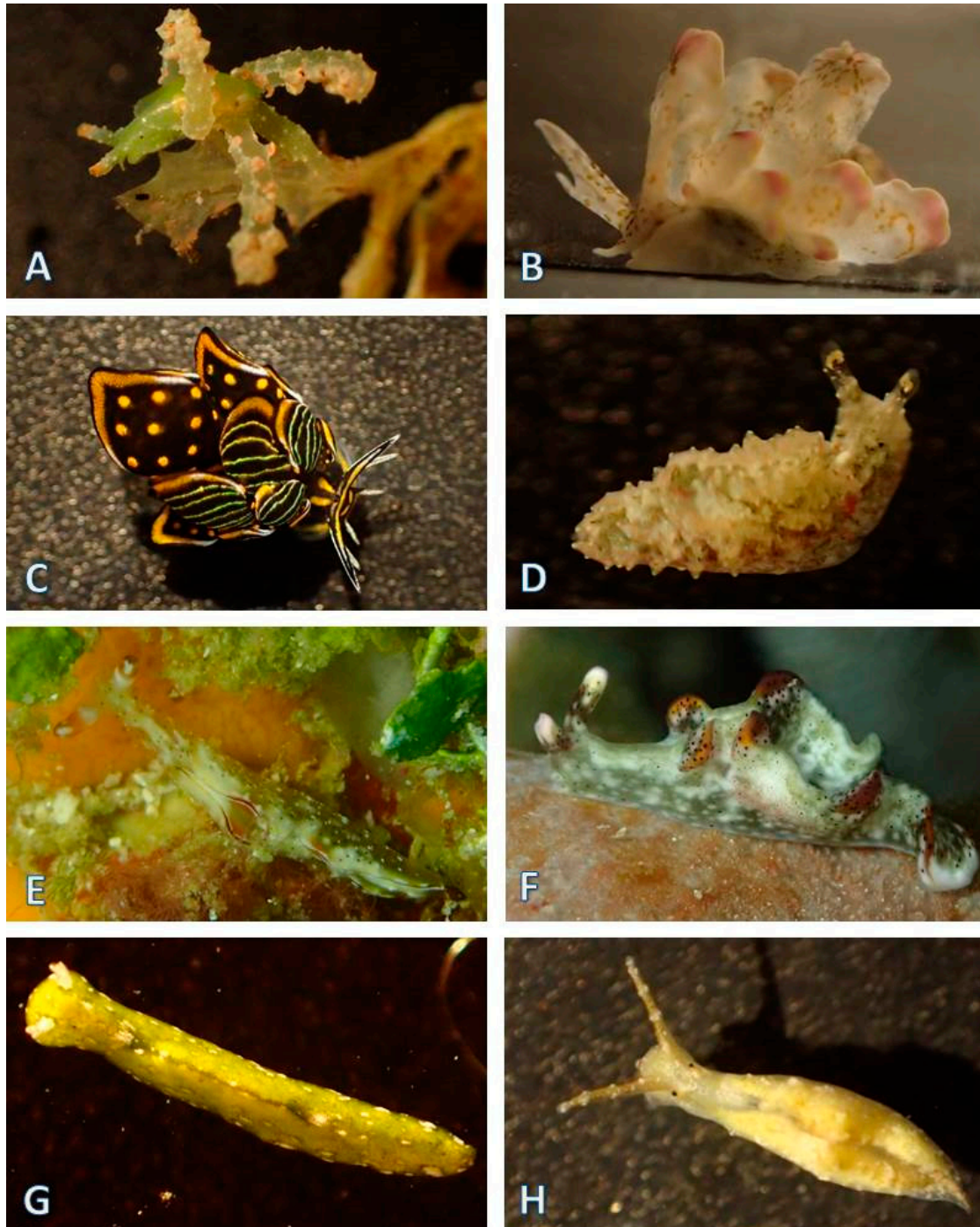


Figure 3. Sacoglossa: (A) *Lobiger* sp. (*Lobiger* sp. 1 in Gosliner et al. [24]: 70), Losp1-18Ba1; (B) *Cyerce bourbonica*, Cybo18Ba-1; (C) *Cyerce nigra*, Cyni18Ba-1; (D) *Elysia asbecki*, Elas18Ba-1; (E) *Elysia marginata*, Elma17Ba-1; (F) *Elysia* cf. *nigropunctata*, Elni18Ba-1; (G) *Elysia pusilla*, Elpu18Ba-2; (H) *Elysia* sp. (*Elysia* sp. 24 in Gosliner et al. [24]: 89), Elsp24-18Ba-1.

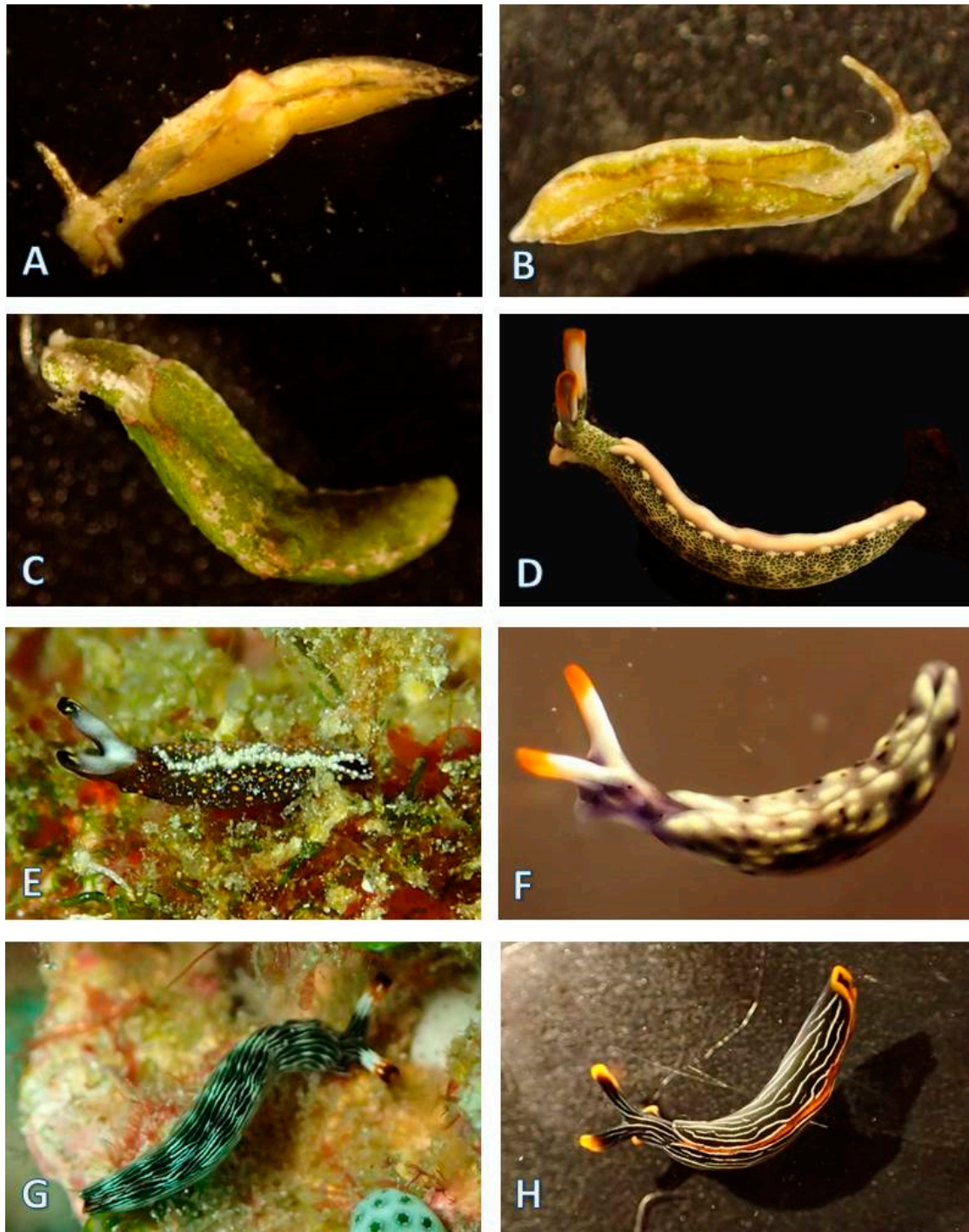


Figure 4. Sacoglossa: (A) *Elysia* sp. (*Elysia* sp. 27 in Gosliner et al. [24]: 89), Elsp27-18Ba-1; (B) *Elysia* sp. (*Elysia* sp. a), Elsp.a18Ba-1; (C) *Elysia* sp. (*Elysia* sp. b), Elsp.b18Ba-1; (D) *Thuridilla carlsoni*, Thca18Ba-1; (E) *Thuridilla flavomaculata*, Thfl17Ba-1; (F) *Thuridilla vataae*, Thva18Ba-3; (G, H) *Thuridilla gracilis*, Thgr18Ba-1, Thgr18Ba-4.

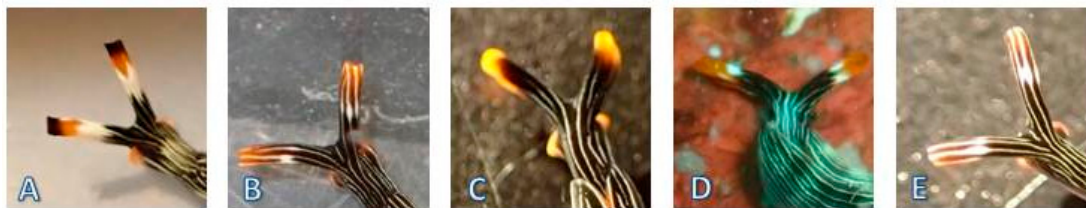


Figure 5. Variability of rhinophores in the *Thuridilla gracilis* species complex from Bangka Archipelago: (A) Thgr18Ba1; (B) Thgr17Ba-3; (C) Thgr18Ba4; (D) Thgr18Ba6; (E) Thgr18Ba-5.

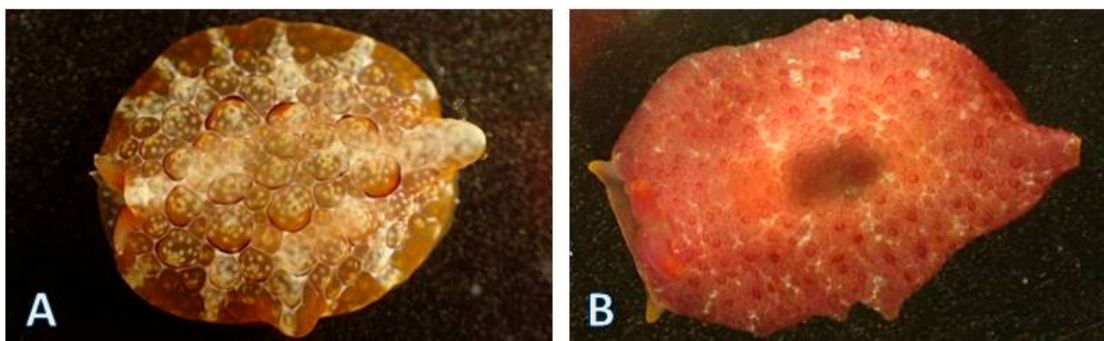


Figure 6. Pleurobranchida: (A) *Pleurobranchus forskalii*, Plfo18Ba-1; (B) *Pleurobranchus peronii*, Plpe18Ba-1.



Figure 7. Nudibranchia Doridina: (A) *Hexabranchnus sanguineus*, Glsp1_17Ba-1; (B) *Nembrotha chamberlaini*, Nech18Ba-1; (C) *Nembrotha cristata* Nocr17Ba-2; (D) *Nembrotha kubaryana*, Neku17Ba-4; (E) *Nembrotha lineolata**; (F) *Nembrotha milleri*, Nemi17Ba-1; (G) *Nembrotha mullineri*, Nemu17Ba-1; (H) *Nembrotha* sp. (*Nembrotha* sp. 1 in Gosliner et al. [24]: 122), Nesp1_17Ba-1. *Specimen not collected.

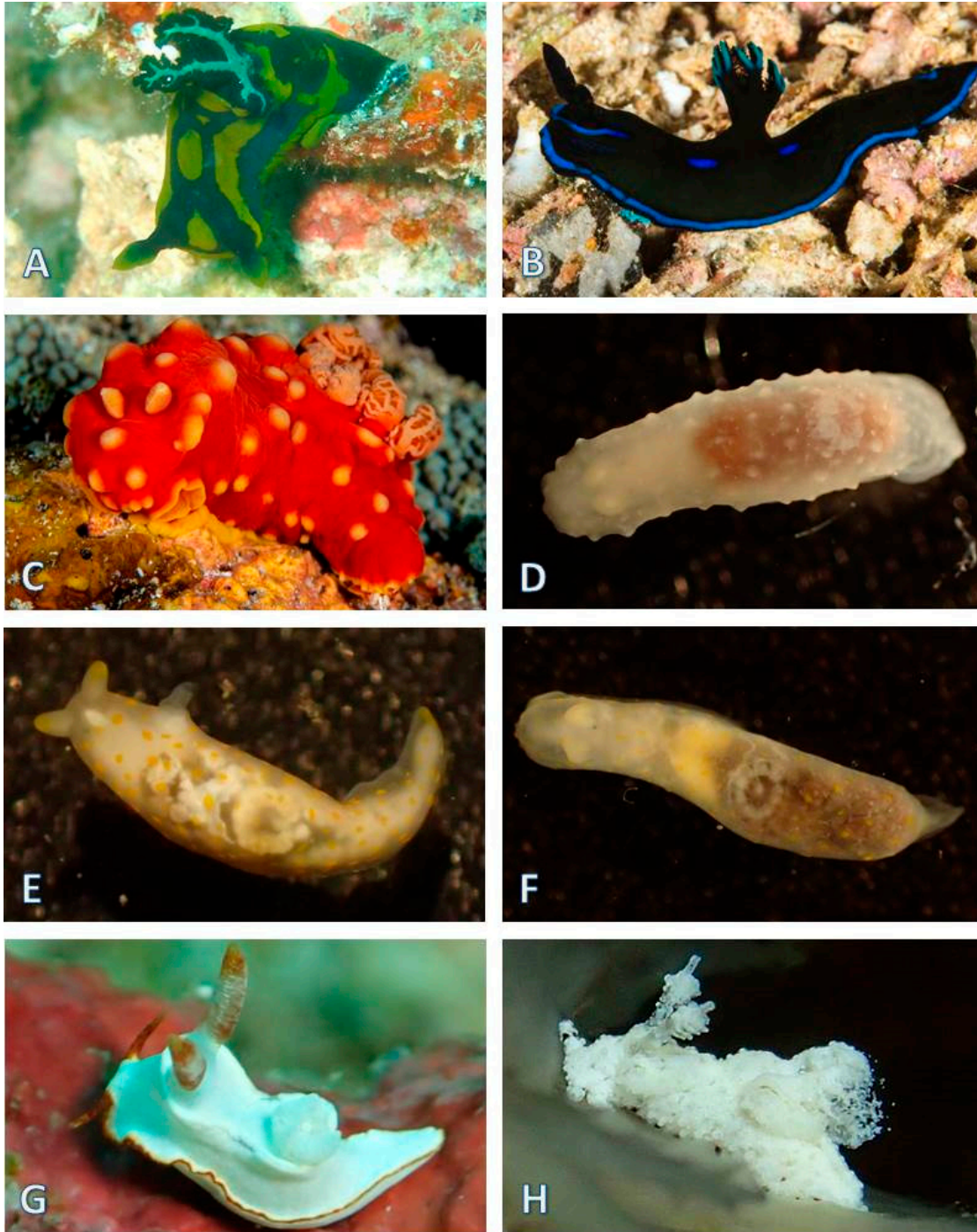


Figure 8. Nudibranchia Doridina: (A) *Tambja gabrielae*, Taga18Ba-1; (B) *Tambja morosa**; (C) *Gymnodoris aurita**; (D) *Gymnodoris tuberculosa*, Gytu18Ba-1; (E) *Gymnodoris* sp. (*Gymnodoris* sp. 20 in Gosliner et al. [24]: 156), Gysp20-18Ba1; (F) *Gymnodoris* sp. (*Gymnodoris* sp. 25 in Gosliner et al. [24]: 157), Gysp25-18Ba1; (G) *Goniodoris* sp. (*Goniodoris* sp. 7 in Gosliner et al. [24]: 153), Gosp7-18Ba1; (H) *Trapania armilla*, Trar17Ba-1. *Specimen not collected.

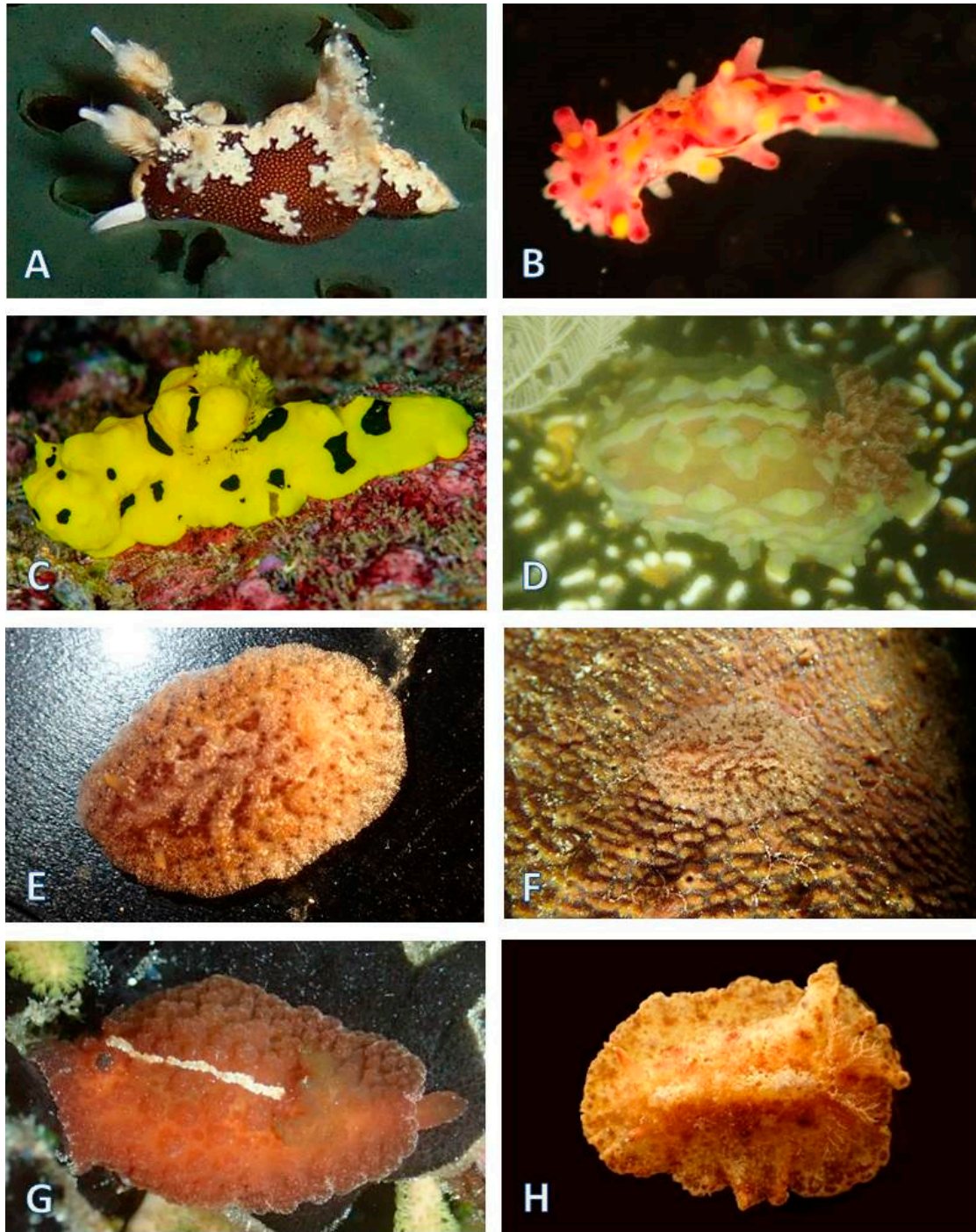


Figure 9. Nudibranchia Doridina: (A) *Trapania safracornia**; (B) *Aegires* sp. (*Aegires* sp. 7 in Gosliner et al. [24]: 149), Aesp7-18Ba-1; (C) *Notodoris minor**; (D) *Asteronotus cespitosus**; (E) *Asteronotus mimeticus*, Asmi18Ba-2; (F) *Asteronotus mimeticus* substrate, Asmi18Ba3S; (G) *Atagemia intecta*, Atin18Ba-2; (H) *Discodoris cebuensis*, Dice18Ba-1. *Specimen not collected.

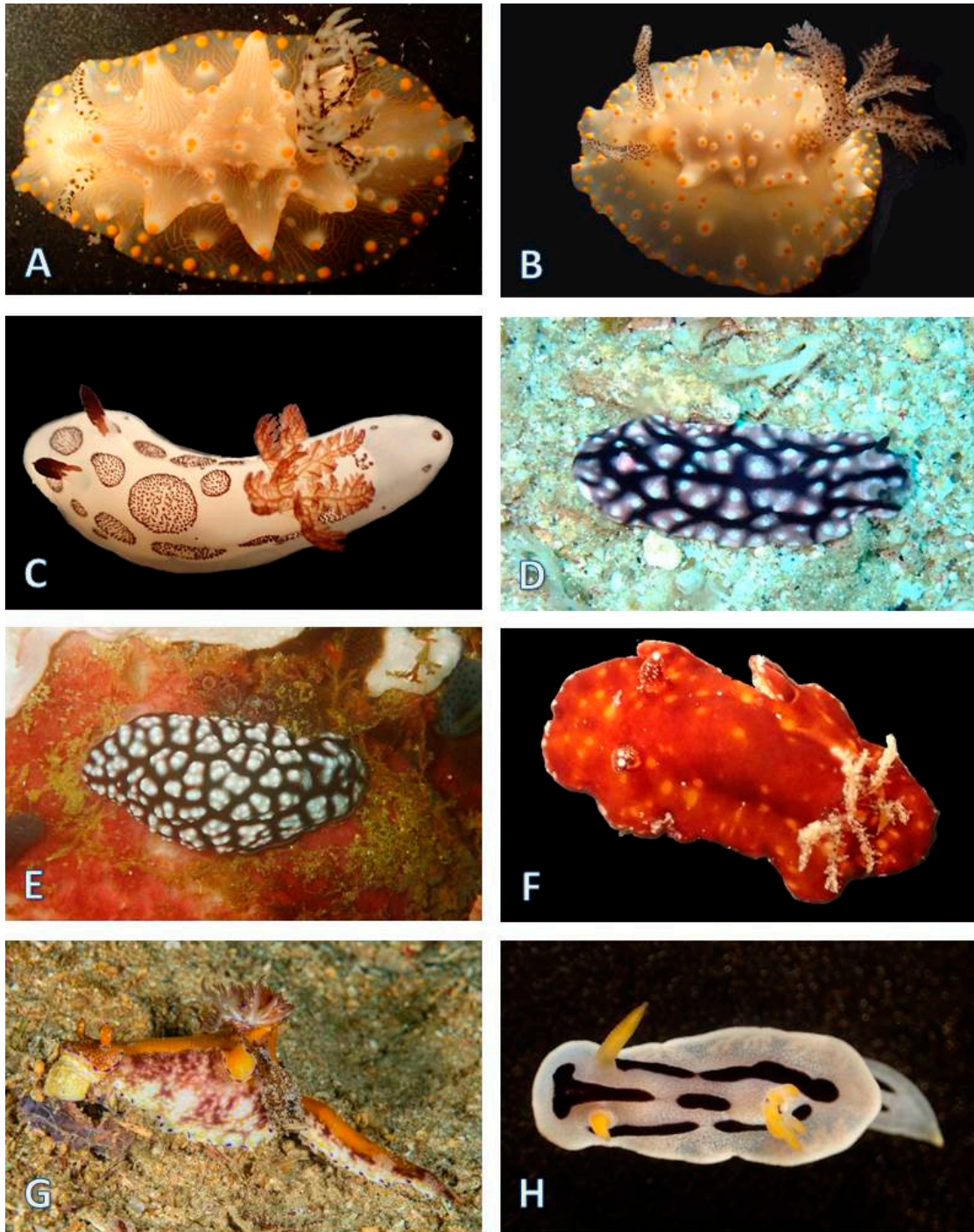


Figure 10. Nudibranchia Doridina: (A) *Halgerda batangas*, Haba18Ba-8; (B) *Halgerda carlsoni*, Haca17Ba-2; (C) *Jorunna funebris*, Jofu17Ba-1; (D) *Paradoris liturata* in situ, Phpu17Ba-6; (E) *Phyllidiella pustulosa* mimicked by *Paradoris liturata*; (F) *Platydoris sanguinea*, Plsa17Ba-1; (G) *Ceratosoma tenue**; (H) *Chromodoris diana*, Chdi18Ba-1. *Specimen not collected.

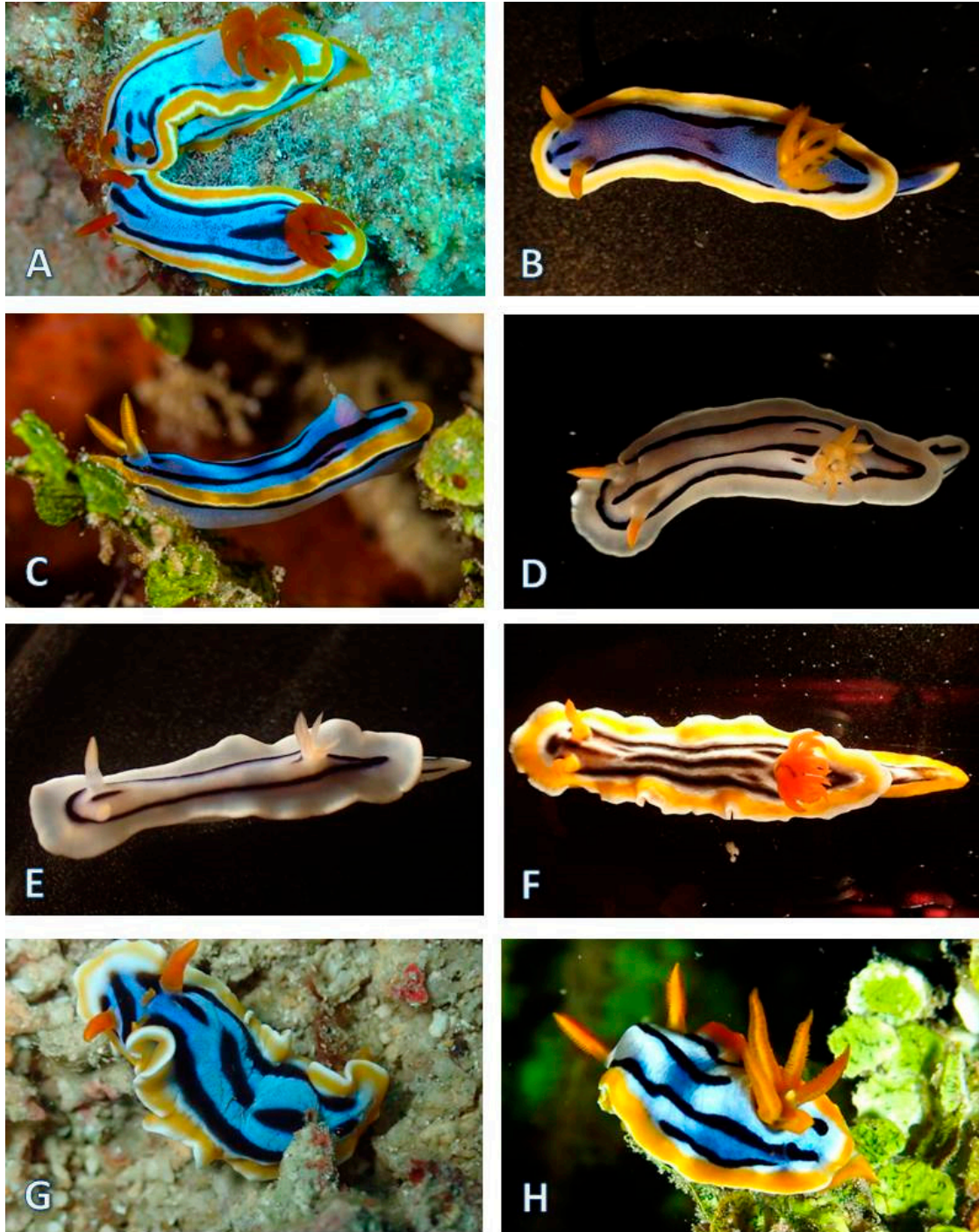


Figure 11. Nudibranchia Doridina: (A, B) *Chromodoris annae*, Chan18Ba-11-12, Chan18Ba-7; (C) *Chromodoris elisabethina**; (D, E) *Chromodoris lochi*, Chlo18Ba-6, Chlo18Ba-1; (F) *Chromodoris magnifica*, Chma18Ba-1; (G) *Chromodoris quadricolor*, Chma18a-2; (H) *Chromodoris strigata*, Chst17Ba-1. *Specimen not collected.

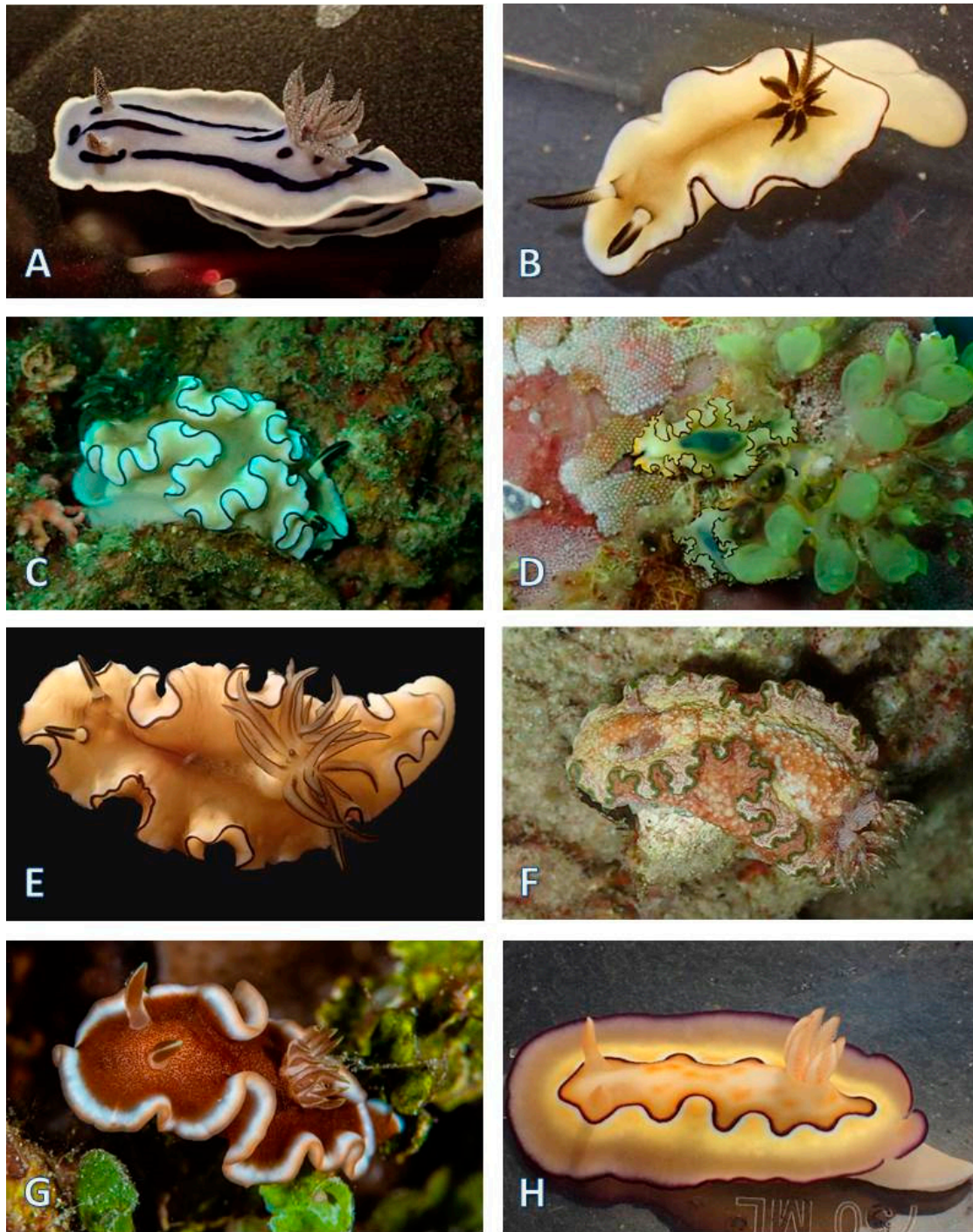


Figure 12. Nudibranchia Doridina: (A) *Chromodoris willani*, Chwi18Ba-1; (B) *Doriprismatica atromarginata*, Doat17Ba-1; (C) *Doriprismatica atromarginata* in situ; (D) Flatworm mimicking *D. atromarginata*; (E) *Doriprismatica sibogae*, Dosi17Ba-1; (F) *Glossodoris cincta*, Glic18Ba-1; (G) *Glossodoris rufomarginata**; (H) *Goniobranchus coi*, Gocho17Ba-1. *Specimen not collected.

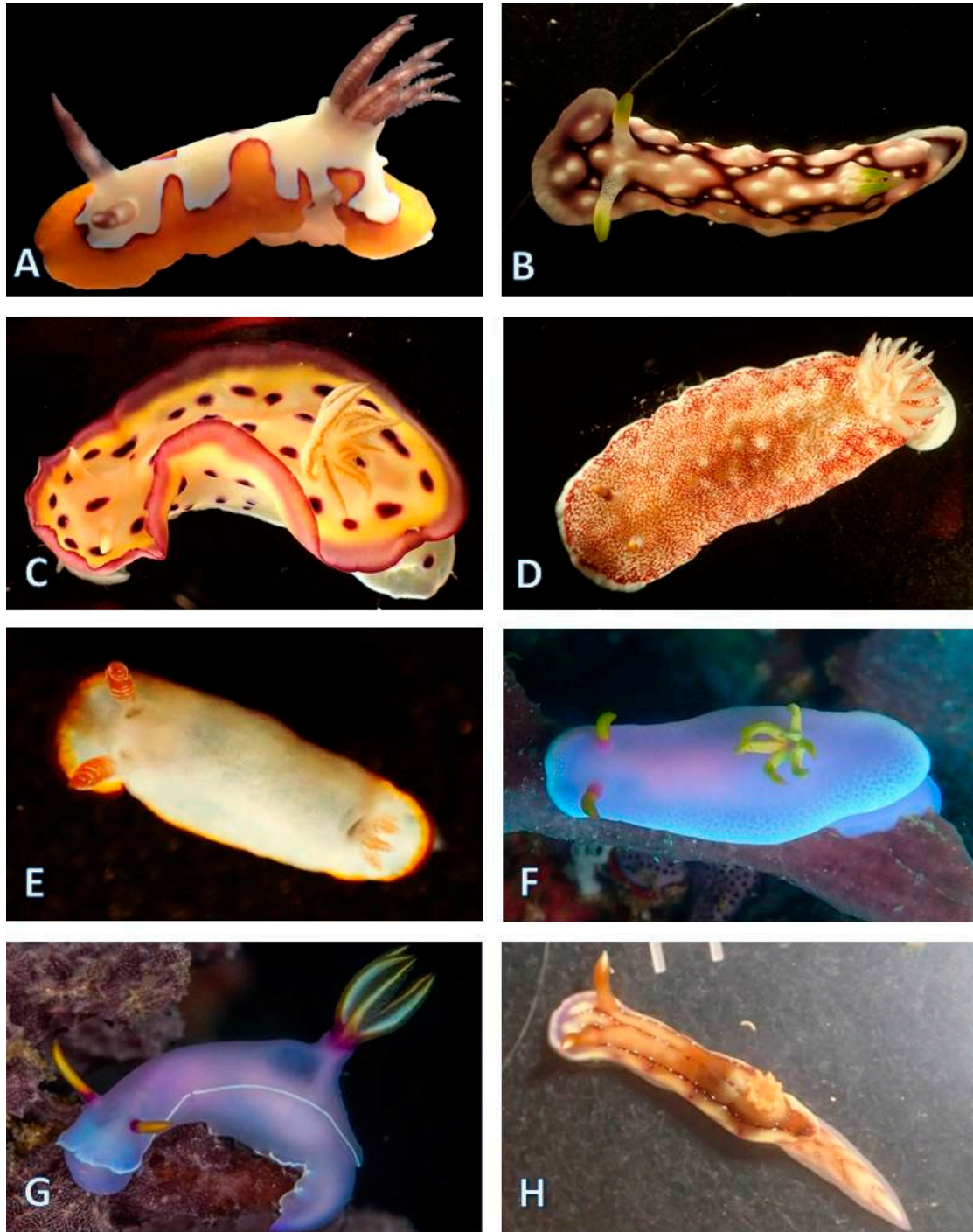


Figure 13. Nudibranchia Doridina: (A) *Goniobranchus fidelis*, Gofi17Ba-1; (B) *Goniobranchus geometricus*, Goge18Ba-5; (C) *Goniobranchus kuniei*, Goku18Ba-1; (D) *Goniobranchus reticulatus*, Gore18Ba-1; (E) *Goniobranchus verrieri*, Thho18Ba-1; (F) *Hypselodoris apolegma*, Hyap17Ba-1; (G) *Hypselodoris bullockii**; (H) *Hypselodoris cerisae*, Hysp1.17Ba-1. *Specimen not collected.

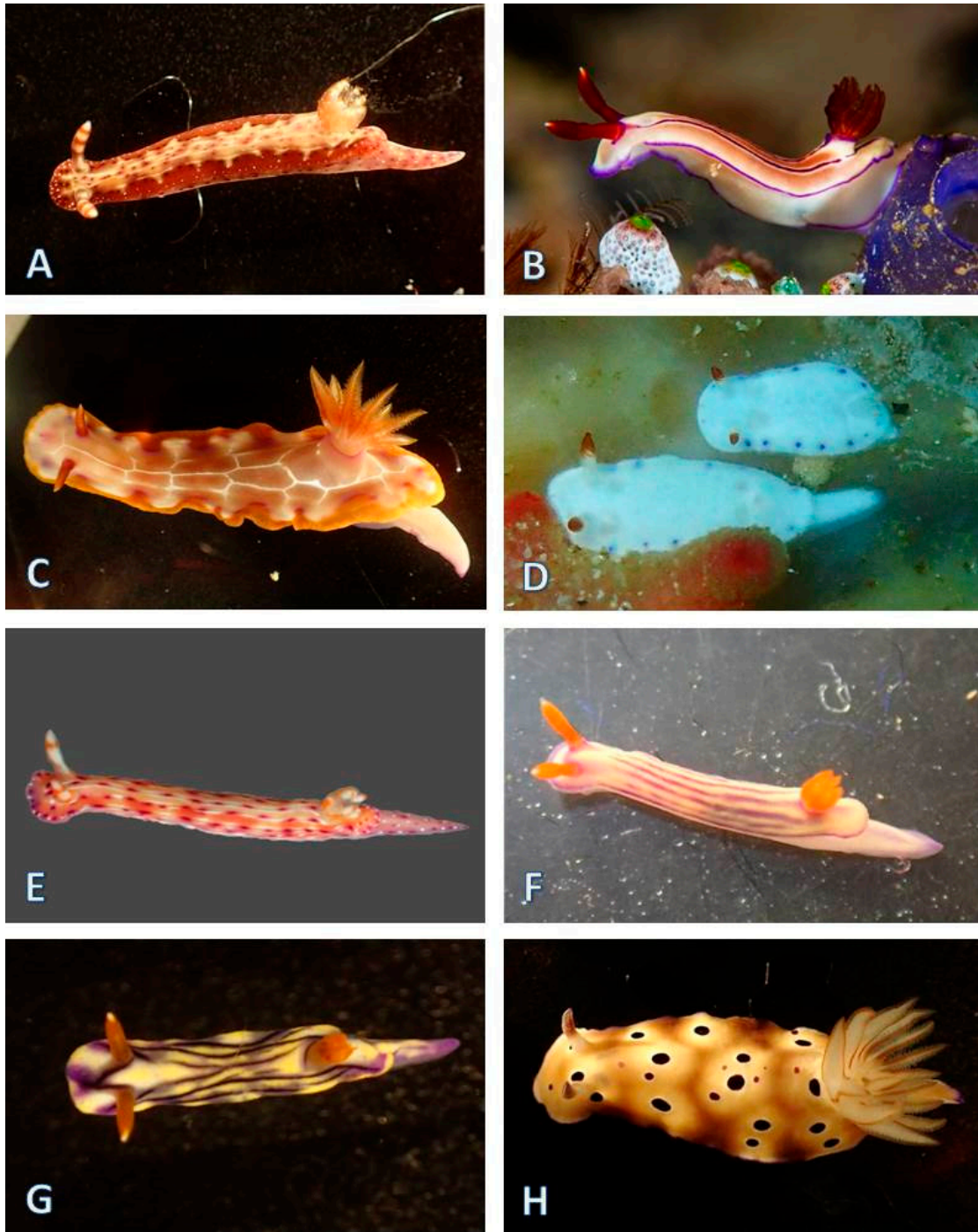


Figure 14. Nudibranchia Doridina: (A) *Hypselodoris decorata*, Hyma18Ba-1; (B) *Hypselodoris emma**; (C) *Hypselodoris iacula*, Chromos18Ba-1; (D) *Hypselodoris lacuna*, Hysp19_17Ba-1-2; (E) *Hypselodoris maculosa*, Hyma17Ba-1; (F) *Hypselodoris maridadilus*, Hymari17Ba-1; (G) *Hypselodoris zephyra*, Hyni18Ba-1; (H) *Hypselodoris tryoni*, Hytr18Ba-5. *Specimen not collected.

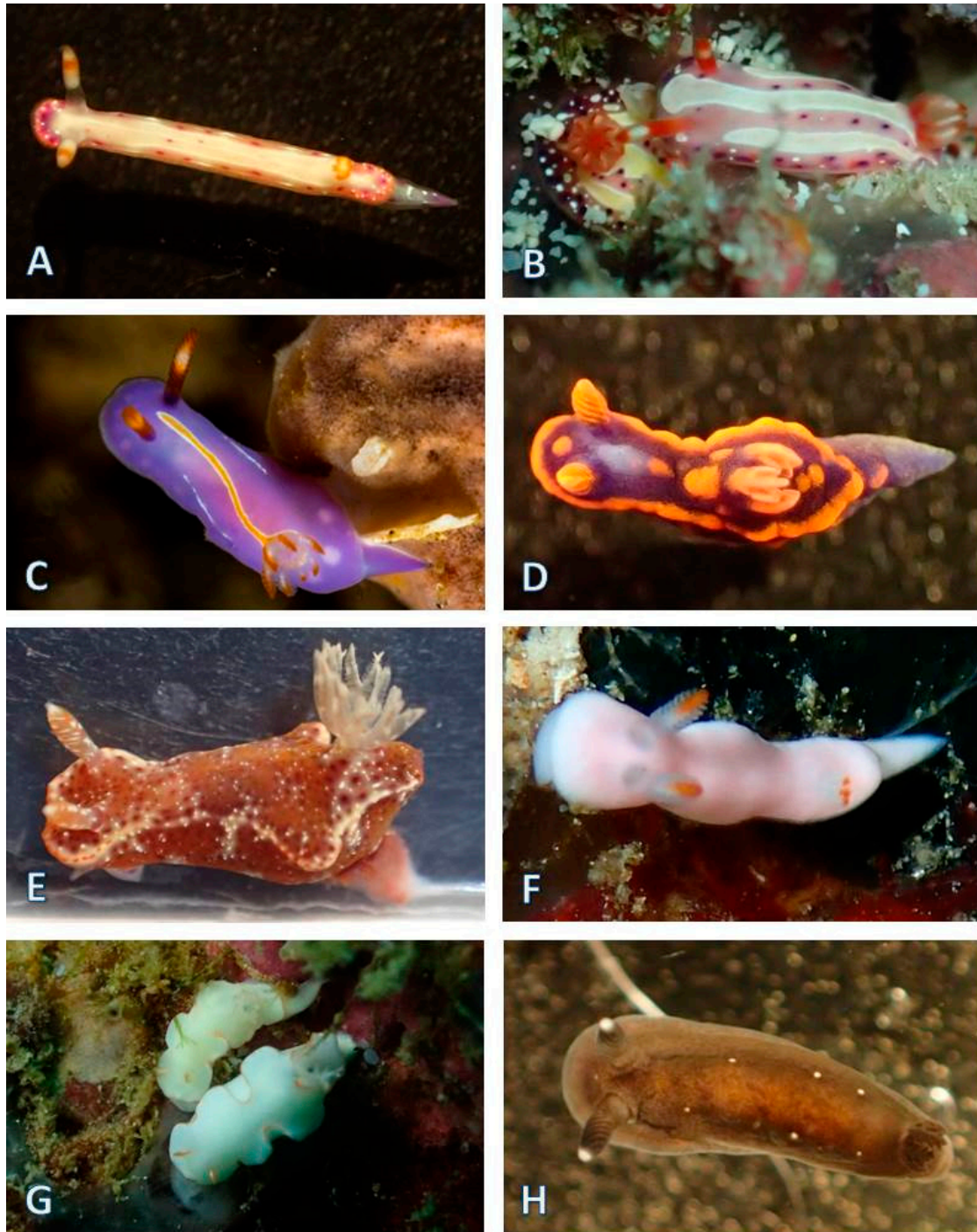


Figure 15. Nudibranchia Doridina: (A) *Hypselodoris* sp. a, Thsp1-18Ba-1; (B) *Mexichromis aurora*, Meau18Ba-1; (C) *Mexichromis trilineata**; (D) *Miamira magnifica*, Dorid18Ba-1; (E) *Miamira* sp. a, Misp17Ba-1; (F) *Verconia simplex*, Nosi17Ba-1; (G) *Thorunna furtiva*, Thfu18Ba-1-2; (H) *Dendrodoris nigra*, Deni18Ba-1. *Specimen not collected.

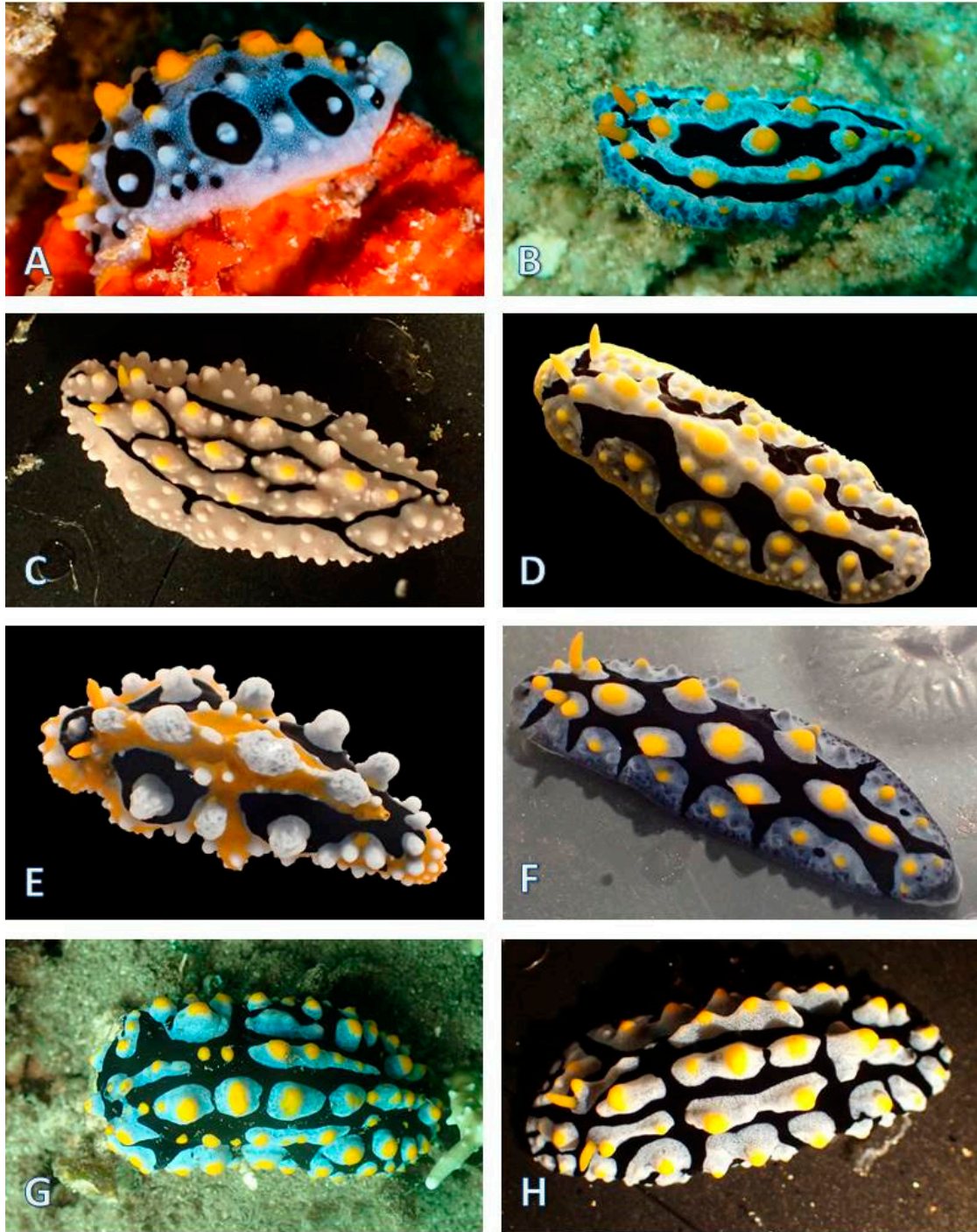


Figure 16. Nudibranchia Doridina: (A) *Phyllidia* cf. *babai**; (B) *Phyllidia coelestis*, Phco18Ba-1; (C) *Phyllidia elegans*, Phel18Ba-1; (D) *Phyllidia exquisita*, Phex17Ba-1; (E) *Phyllidia ocellata*, Phoc17Ba-1; (F) *Phyllidia picta*, Phpi17Ba-1; (G, H) *Phyllidia varicosa*, Phva18Ba-11, Phva18Ba-6. *Specimen not collected.

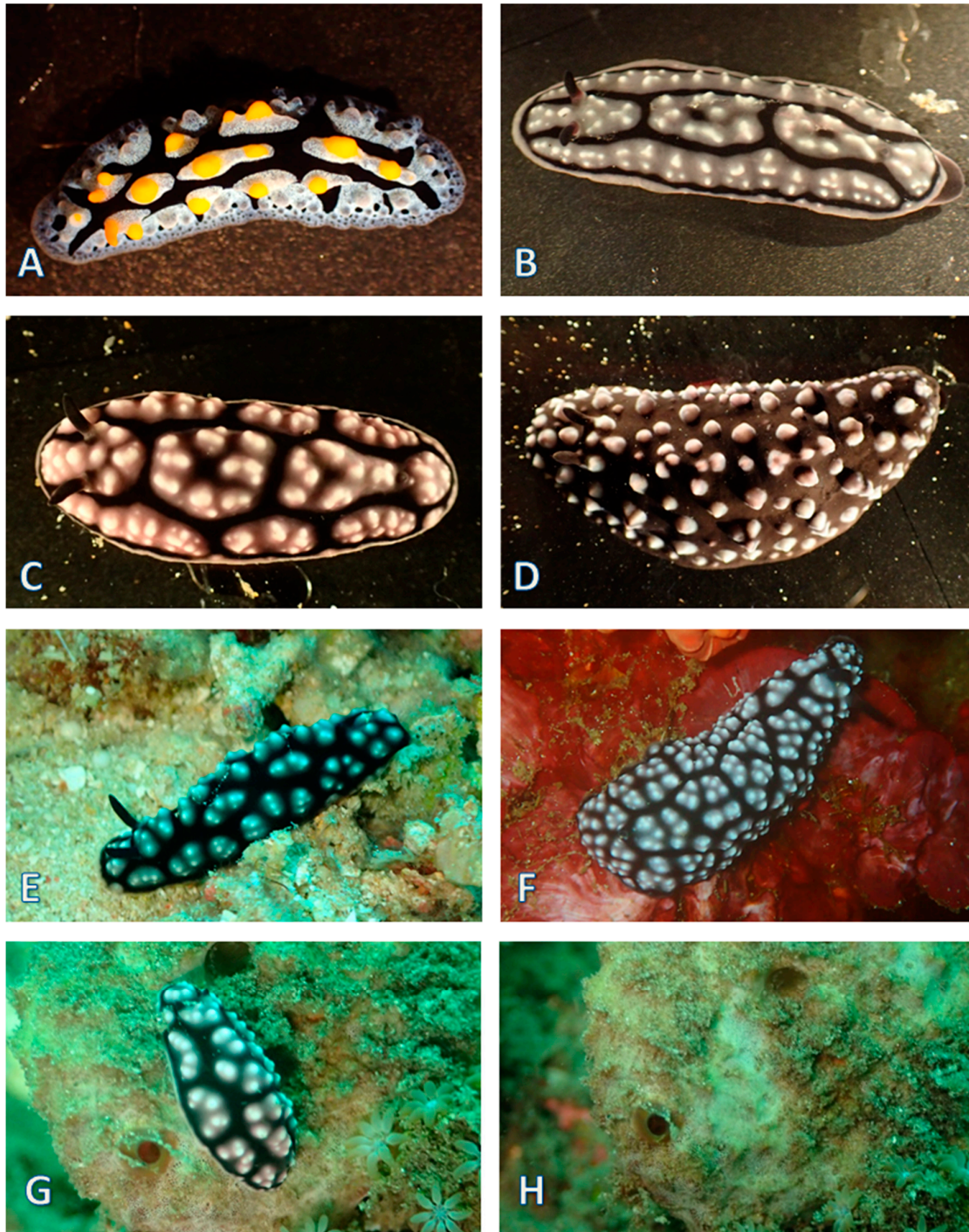


Figure 17. Nudibranchia Doridina: (A) *Phyllidia* sp. a, Phsp18Ba2; (B) *Phyllidiella annulata*, Phpu18Ba-12; (C) *Phyllidiella lizae*, Phli18Ba-1; (D) *Phyllidiella nigra*, Phni18Ba-1; (E, F) *Phyllidiella pustulosa*, Phpu18Ba-2, Phpu18Ba-4. (G) Specimen of *Phyllidiella pustulosa* in situ on a sponge; (H) Sponge viewed immediately after *P. pustulosa* was removed.

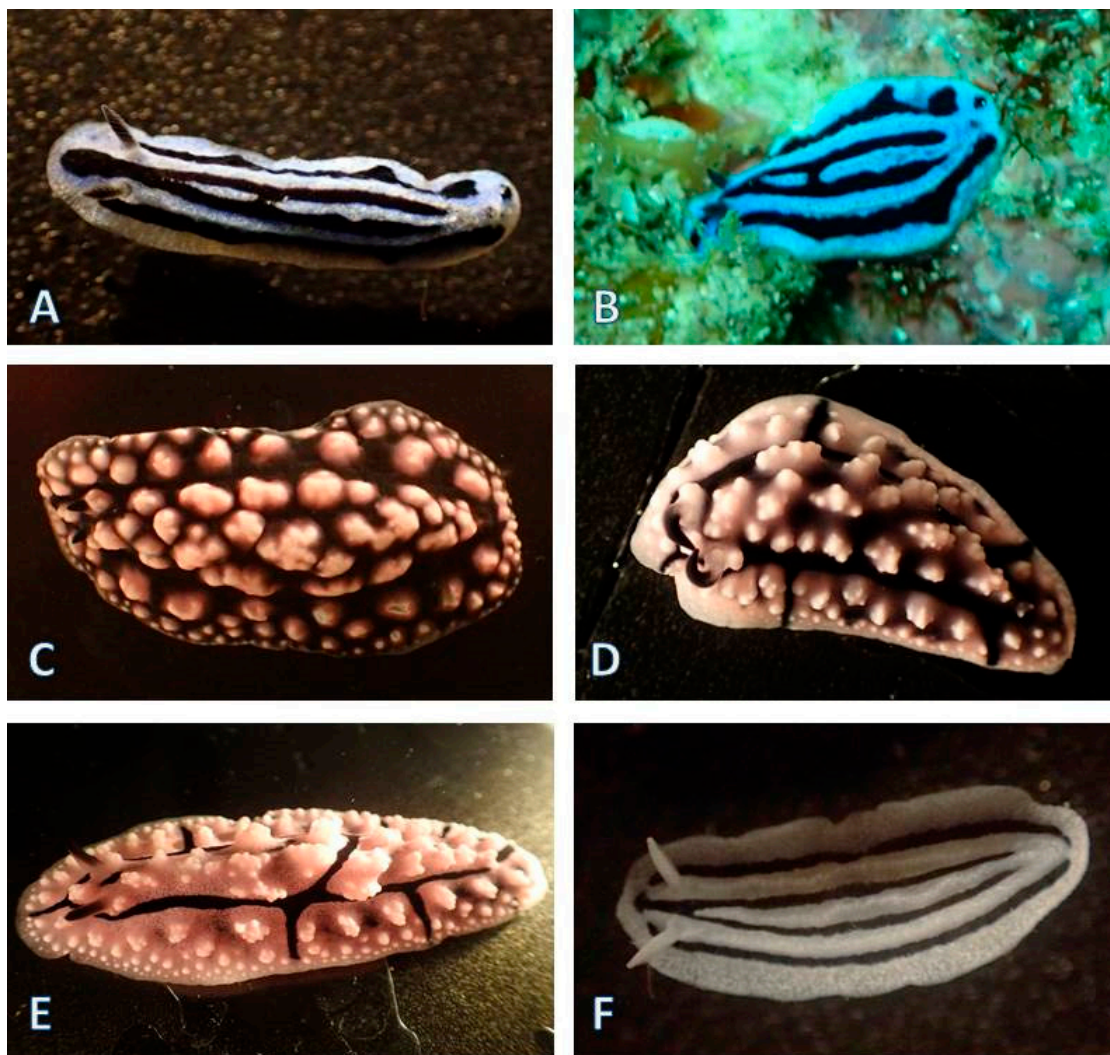


Figure 18. Nudibranchia Doridina: (A, B) *Phyllidiopsis annae*, Phsan18Ba-1 and in situ; (C, D) *Phyllidiopsis* cf. *burni*, Phssp18Ba-1; (E) *Phyllidiopsis krempfi*, Phskre18Ba-1, Phspi18Ba-1; (F) *Phyllidiopsis xishaensis*, Phsxi18Ba-1.

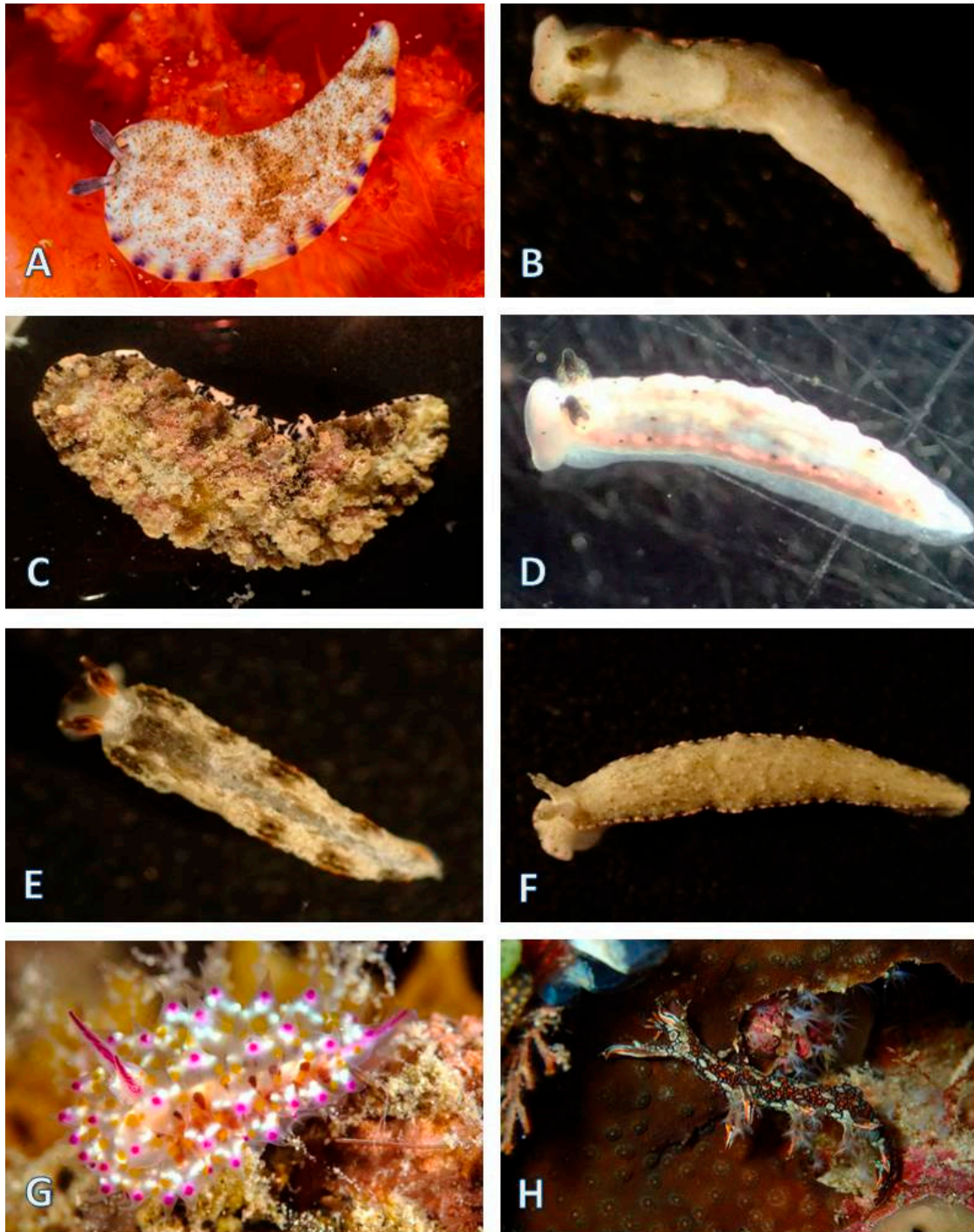


Figure 19. Nudibranchia Cladobranchia: (A) *Dermatobranchus caeruleomaculatus**; (B) *Dermatobranchus rodmani*, Dero18Ba-1; (C) *Dermatobranchus pustulosus*, Depu18Ba-1; (D) *Dermatobranchus* sp. a, Desp.a18Ba-1; (E) *Dermatobranchus* sp. b, Desp18Ba-1; (F) *Dermatobranchus* sp. c, Desp17Ba-1; (G) *Janolus* sp. (*Janolus* sp. 1 in Gosliner et al. [24]: 305)*; (H) *Bornella anguilla*, Boan17Ba-1. *Specimen not collected.

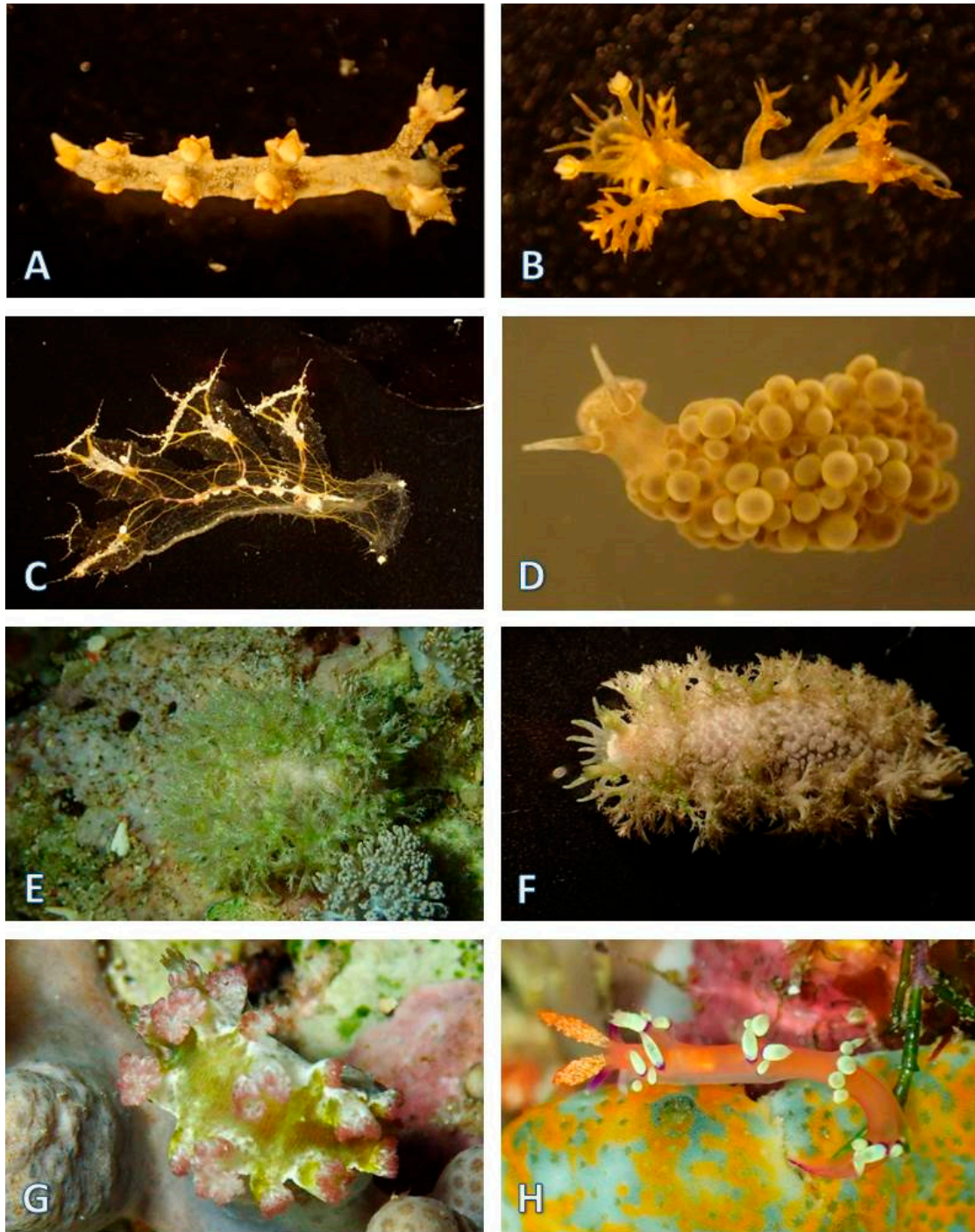


Figure 20. Nudibranchia Cladobranchia: (A) *Bornella dotoides*, Bohe18Ba-1; (B) *Melibe bucephala*, Mesp.a18Ba-1; (C) *Melibe engeli*, Mebu18Ba-1; (D) *Doto ussi*, Dous18Ba-2; (E, F) *Marionia* sp. (*Marionia* sp. 2 in Gosliner et al. [24]: 324), Masp2.18Ba-1; (G) *Tritonia* sp. (*Tritonia* sp. 3 in Gosliner et al. [24]: 320), Trsp3_17Ba-1; (H) *Coryphellina exoptata*, Flex17Ba-1.

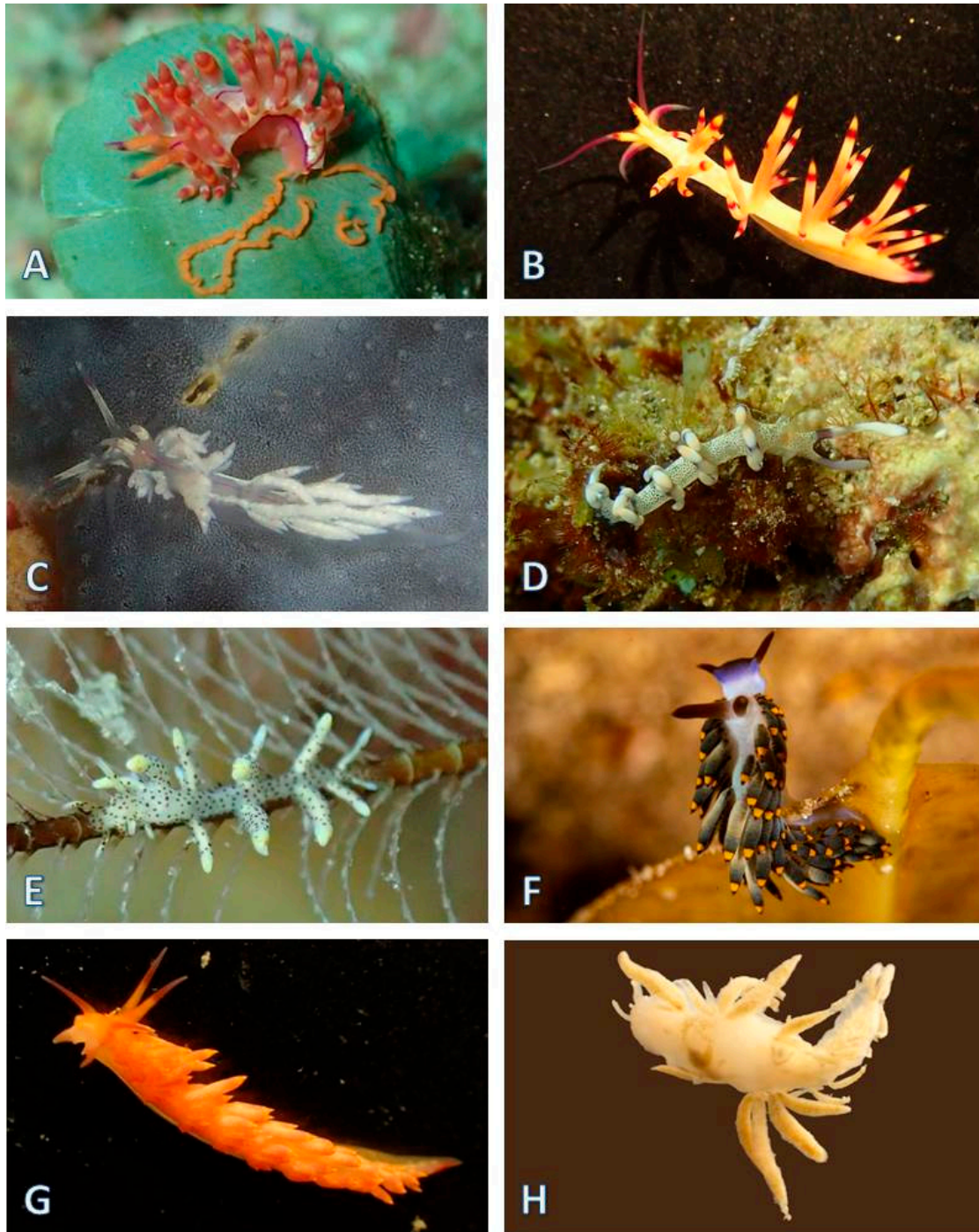


Figure 21. Nudibranchia Cladobranchia: (A) *Coryphellina rubrolineata*, Flru17Ba-1; (B) *Flabellina* sp. (*Flabellina* sp. 2 in Gosliner et al. [24]: 333), Flsp2-18Ba-1; (C) *Flabellina* sp. (*Flabellina* sp. 3 in Gosliner et al. [24]: 333), Flsp3-17Ba-1; (D) *Samla riwo*, Flri17Ba-3; (E) *Eubranchus* sp. (*Eubranchus* sp. 22 in Gosliner et al. [24]: 341), Eusp22-18Ba-1; (F) *Trinchesia yamasui**; (G) *Cuthona* sp. (*Cuthona* sp. 57 in Gosliner et al. [24]: 353), Cusp57-18Ba-1; (H) *Aeolidia* sp. A, Aeol18Ba-1. *Specimen not collected.

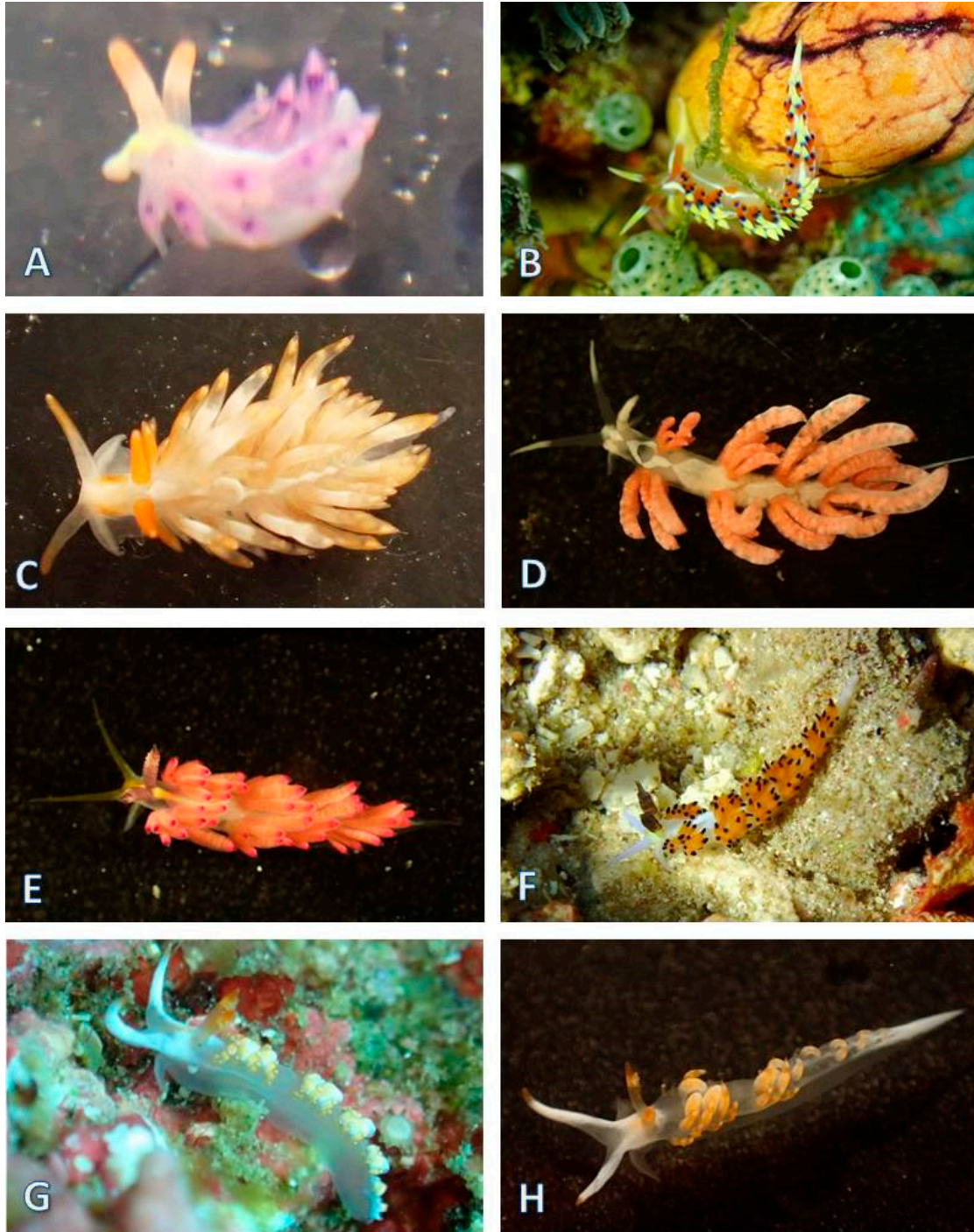


Figure 22. Nudibranchia Cladobranchia: (A) *Antonietta* sp. a, Ansp17Ba-1; (B) *Caloria indica*, Cain18Ba-3; (C) *Cratena* sp. a, Crsp17Ba-1; (D) *Favorinus japonicas*, Faja18Ba-1; (E) *Favorinus* sp. (*Favorinus* sp. 1 in Gosliner et al. [24]: 363), Fasp1.18Ba-1; (F) *Favorinus tsuruganus*, Fats17Ba-1; (G, H) *Moridilla* sp. a, Nosp1.17Ba-1, Mojo18Ba-1.

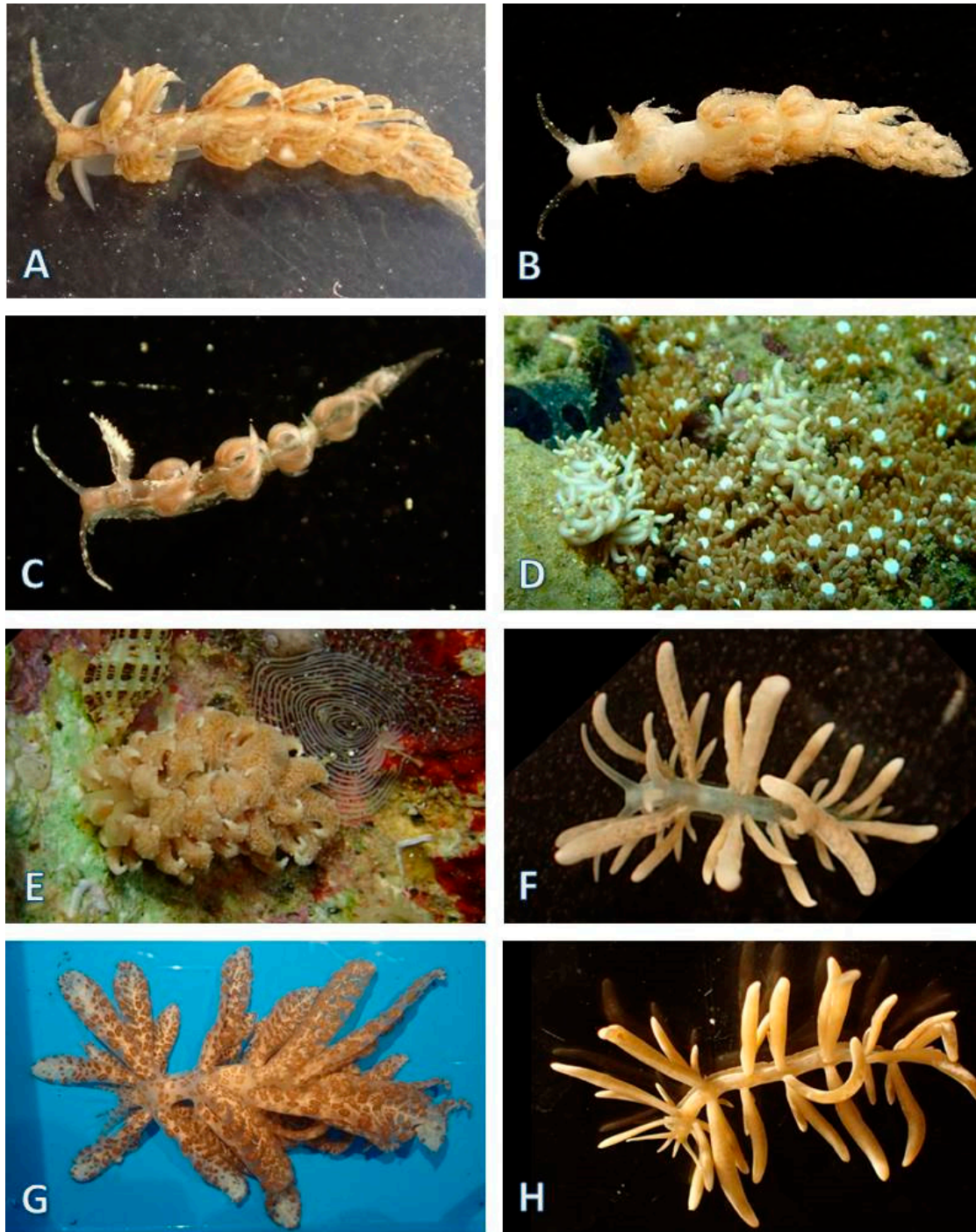


Figure 23. Nudibranchia Cladobranchia: (A) *Noumeaella* sp. (*Noumeaella* sp. 2 in Gosliner et al. [24]: 367), Nosp2_17Ba-1; (B) *Noumeaella* sp. (*Noumeaella* sp. 3 in Gosliner et al. [24]: 367), Nosp3Ba-3; (C) *Noumeaella* sp. (*Noumeaella* sp. 13 in Gosliner et al. [24]: 369), Nosp13Ba-4; (D) *Phyllodesmium briareum*, Phbr18Ba-2-16; (E) *Phyllodesmium* cf. *crypticum*, Phcr17Ba-4; (F) *Phyllodesmium lizardense*, Phliz18Ba-3; (G) *Phyllodesmium longicirrum*, Phlo18Ba-1; (H) *Phyllodesmium magnum*, Phma18Ba-1.

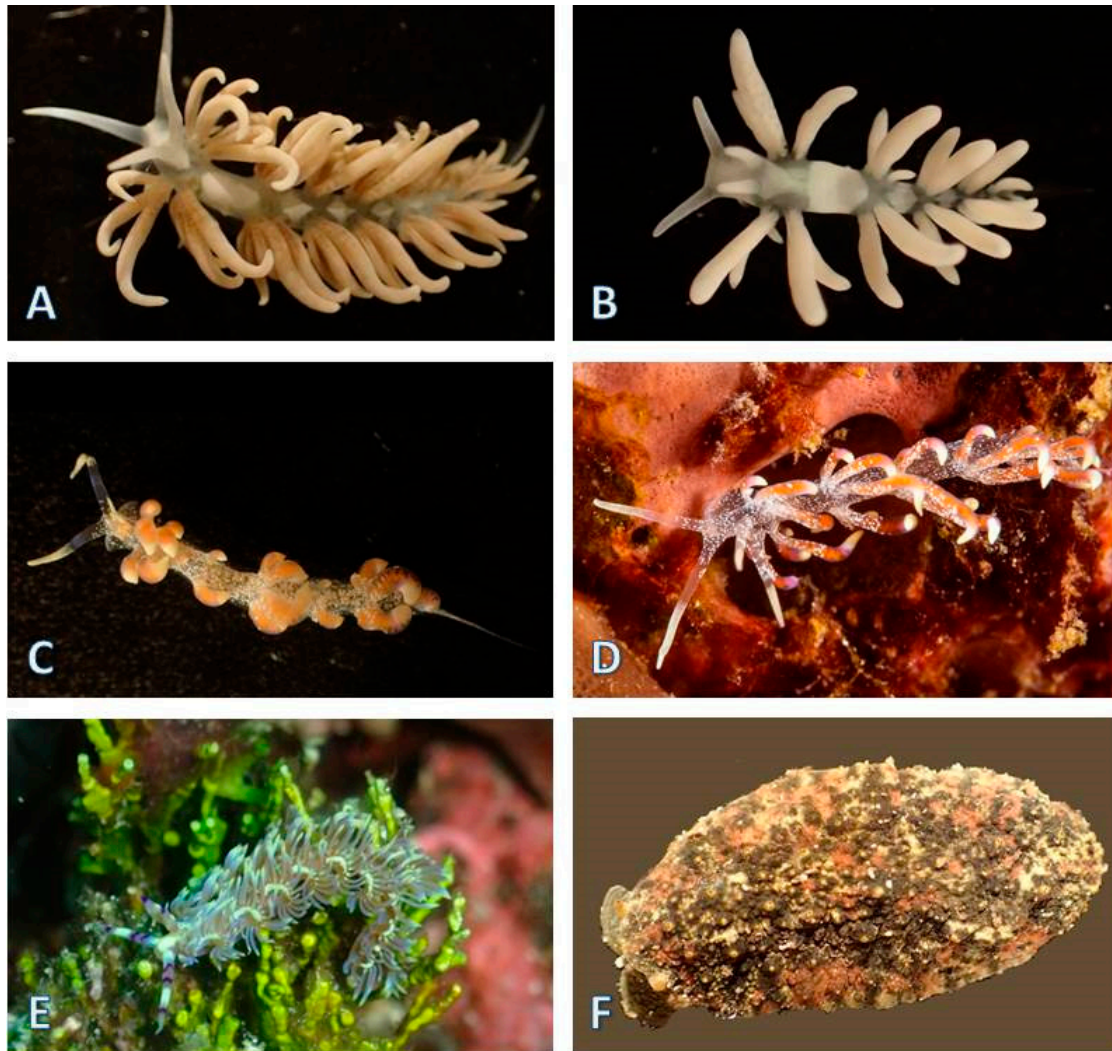


Figure 24. Nudibranchia Cladobranchia and Eupulmonata: (A) *Phyllodesmium parangatum*, Phpa18Ba-4; (B) *Phyllodesmium pecten*, Phpe18Ba-1; (C, D*) *Phyllodesmium poindimiei*, Phpo18Ba-1; (E) *Pteraeolidia semperi*, Ptse17Ba-2; (F) *Peronia* sp. a, Onsp18Ba-1.

4. Discussion

This study is the first of its kind in the Bangka Archipelago (BA). In total, 484 specimens representing 149 species are recorded for the first time from this area, with a continuous increase in species with every collection area (Figure 25). With 33 undescribed species, more than 20% of the total species record is not known to science. In comparison to the recent study of Bunaken National Park (BNP) [16], which recorded 69 undescribed species out of a total of 215 species, the percentage of new species in BA is lower. Summarizing our data from Bangka Archipelago and comparing them with previous studies from North Sulawesi (Figure 26), BA represents the second most diverse area with regard to marine heterobranchs. The species accumulation curve also indicates the increase in species around Bangka Archipelago not recorded before. In total, 333 recorded species are now recorded from North Sulawesi.

When comparing higher order levels in the studied regions of North Sulawesi (Figure 27), the distribution of species within the higher taxa is quite similar, with Doridina and Cladobranchia being the most common taxa, followed by Sacoglossa and Cephalaspidea. However, BA shows the highest number of Doridina, whereas Cladobranchia are more common in BNP.

Distribution of specific taxa is highly variable when comparing the different localities in BA (Table 2), with *Chromodoris annae* (46 individuals) and *Phyllidiella pustulosa* (39 individuals) dominating the overall collection and being encountered in almost all sites. Unpublished results confirm earlier results from Stoffel et al. [25] and clearly show cryptic speciation in *Phyllidiella pustulosa*, with sympatric occurrences of the various clades in Bangka Archipelago, BNP, and the island of Sangihe. If these clades are considered separate species, the diversity would increase by five to seven species and thus render *P. pustulosa* a much less common species. All available *C. annae* specimens from this study and the prior studies seem to cluster in one clade as one species (unpublished data). Other species were also found in high numbers, but not at many sites. A large population of *Pteraeolidia semperi* was found in one place (Sempini) with more than 100 specimens recorded, probably coming together for mating. More than 50 specimens of *Phyllodesmium briareum* were found on a single large soft coral colony of *Briareum*, which extended nearly 2 m². However, these larger aggregations are unusual. Many species (62) were found with only one individual.

With regard to animals encountered during diving, the family Phyllidiidae seems to be the most dominant one, with members found at almost all dive sites. The animals feed on sponges, on which they sometimes leave a scar (Figure 17G,H). This was already noted before [37,38] where, e.g., Yasman [37] interpreted the scars as an effect of extra-oral digestion.

A few genera are represented by a higher number of species than in other areas of North Sulawesi. Eight species of *Phyllodesmium* are now recorded from BA; this number is thus much higher than that recorded in BNP, with only four species. Members of *Phyllodesmium* feed on octocorals, which are very common in our study area.

Recording diversity can be biased by collection efforts (Figure 25). The highest number of species was collected in front of Coral Eye, due to the proximity of this site to the marine laboratory station; this enabled several night dives and also a more thorough search in the coral rubble, which was collected and examined in the laboratory. Coral rubble is an important habitat for smaller heterobranchs, providing a wide array of food items. Specific searches in this habitat yielded many species not found while diving, e.g., the rather cryptic *Melibe bucephala*, several *Gymnodoris* spp., *Dermatobranchus rodmani*, and *Dermatobranchus* species.

The lowest number of species and specimens occurred in mangrove areas (close to Sipi and Kinabuhutan). This is certainly because of lower food availability. However, a species typical of this habitat, *Jorunna funebris*, was recorded from the locality in high numbers (but not all were collected). The record of only one species from Batu Tiga is certainly due to the difficult diving situation on that particular day with strong currents. The locality is dominated by a coral sand flat at a depth of ca. 20 m with a few coral pinnacles. Snorkeling in front of the destroyed area at the mining site close to Sipi did not result in any records of nudibranchs, due to the still rather fresh stone blocks dumped in front of the site, where only a few corals and sponges were observed to have begun to colonize them. We did not include the mining site in our list of localities, because we did not perform any scuba diving at this particular locality.

Table 3 and Figure 28 list all recorded marine Heterobranchia species from North Sulawesi including our data from Bangka Archipelago (BA) and those published from the three other study areas: Bunaken National Park (BNP) [1,15,16], Sangihe Island (SA) [18], and Lembeh Strait (LS) [17]. Lembeh Strait is particularly famous for its richness in marine heterobranchs, although detailed studies are still lacking and most of the data are only available on the internet. For better comparison, we extracted additional information from Tono-zuka [39] and especially from seaslugforum.net [21]. Photographic records provided by many scientists and citizen scientists on this website were evaluated by Bill Rudman and the scientific community in the past. Using information from citizen scientists has become more important lately in documenting changes in species composition [13,40–42]. This is a state-of-the-art species list and enables future monitoring to assess earlier data from this locality. In the following section, we highlight some results from this comparison.

Only 10 species (*Thuridilla gracilis*, *Chromodoris annae*, *C. strigata*, *Glossodoris cincta*, *Goniobranchus geometricus*, *G. reticulatus*, *Hypselodoris tryoni*, *Phyllidia ocellata*, *P. varicosa*, and *Phyllidiella pustulosa*) are recorded from all areas. Of these, *C. annae*, *P. varicosa*, and *P. pustulosa* show the broadest

distribution with records from nearly all dive sites of the four study areas, and can probably be considered as the most dominant species in North Sulawesi. However, there are also distinct differences in species composition at each of the various study areas. A few of them we would like to address here, with an emphasis on Bunaken National Park (BNP) and Lembeh Strait (LS). A much higher number of cephalaspideans and sacoglossans are now recorded from BNP than from BA or LS. However, the sacoglossan *Thuridilla gracilis* is very common in BA and LS, but very rare in BNP. Future studies will show which of the recently identified 14 *T. gracilis* lineages [31] are present. In contrast, its congener *T. lineolata* is a common species in BNP, especially in the lagoons behind the fringing reefs. This habitat structure is less common in BA and LS and is probably the reason why *T. lineolata* was not found in BA (but recorded in LS).

Six species of *Nembrotha* were collected in BA, but only two of these species, the highly conspicuous *Nembrotha kubaryana* and *N. cristata*, are reported from BNP, which are also very common in BA. Members of the genus feed on ascidians, which usually need higher nutrition in the water column [43]. No bryozoan-feeding members of the genus *Polycera* are recorded from BA, although several species are recorded from BNP [16]. However, the bryozoan-feeding genus *Tambja* is represented by two species in BA, but only one in BNP.

Halgerda batangas was represented in BA with 10 specimens from five different localities, a much higher number than the records in BNP [16]. Localities in the two different study areas are similar in as much as all sites with *Halgerda* present were noted to have a highly diverse sponge community. Why diversity and also number of specimens of *Halgerda* species seems to be higher in BA and LS is therefore not clear, but can probably be explained by the distribution and environmental needs of the food sponges [24,44].

Within Chromodorididae, the genus *Hypselodoris* is quite common in BA and LS, with 11 and 10 species, respectively. In BNP, five species are recorded: the three species *H. apolegma*, *H. maculosa*, and *H. tryoni* are also distributed in BA and LS, but two undescribed species are only recorded from BNP.

Some taxa are small and very inconspicuous, and therefore seldom reported by citizen scientists. The genus *Dermatobranchus* has many small forms with cryptic coloration and is therefore easily overlooked. Our study revealed six different species in BA; Eisenbarth et al. [16] also recorded six species from BNP; however, the overlap of species includes only one undescribed species of *Dermatobranchus*. None of these *Dermatobranchus* species are recorded from LS [21,39].

Dotidae are represented by *Doto* and *Kabeiro* in North Sulawesi. Interestingly, the genus *Kabeiro* was quite common in BNP, but was not found in BA. Members of this genus exclusively feed on hydroids of the family Plumulariidae, which are less common in BA.

Many biotic and abiotic factors influence both the occurrence and distribution of sea slugs. The main island of Bunaken National Park, Bunaken Island, is formed by an old reef and is characterized by fringing reefs with wall-like structures, exposed to hydrodynamics varying in strength (e.g., high waves and strong currents). Lagoons behind the fringing reefs are formed by white sand partly covered by seagrass or mangroves and are influenced by the tides. Lembeh Strait, located in the south-east of the Bangka Archipelago, is of volcanic origin and dominated by slopes with coral patches, volcanic sand slopes, and beds. Fringing reefs, typical for Bunaken National Park, are uncommon in LS and BA. Bangka Island lies between these two areas and supports both habitats: it is partly fringed by reefs, and areas are partly dominated by volcanic sands. However, habitat structure and substrate are not the only factors that influence species composition; temperature and water currents also have a strong effect on the connectivity of populations within these three localities, affecting the distribution of both heterobranch predator and prey larvae [45–47]. Understanding how these factors contribute to the differences would require much more information about the lifestyle of those particular species that are more common or rare in the respective geographic areas.

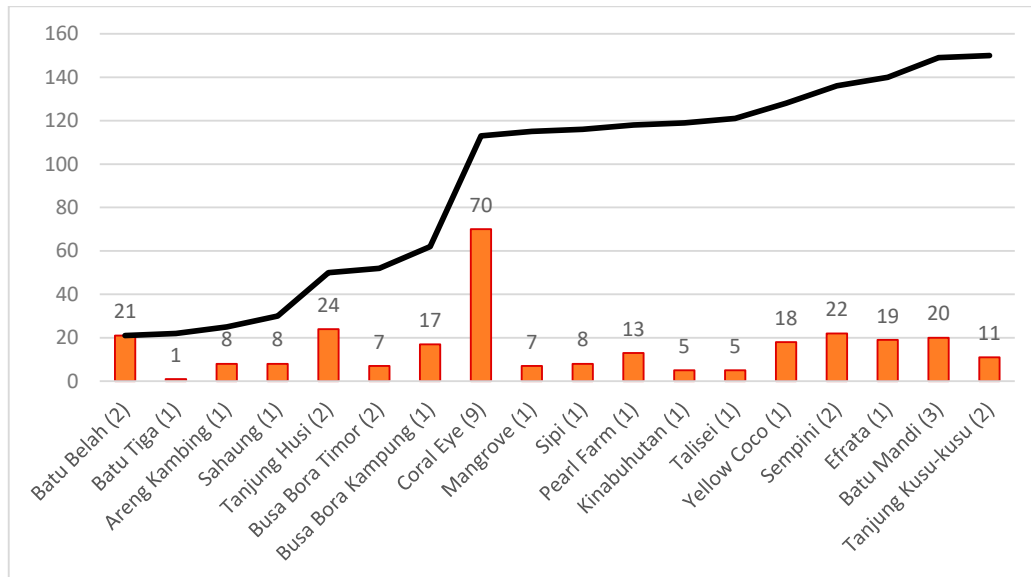


Figure 25. Species accumulation curve (black line) and number of species per locality (red bars). Numbers in brackets after the locality name indicate the number of sampling events at that particular site. Note the high number of sampling events at the locality Coral Eye.

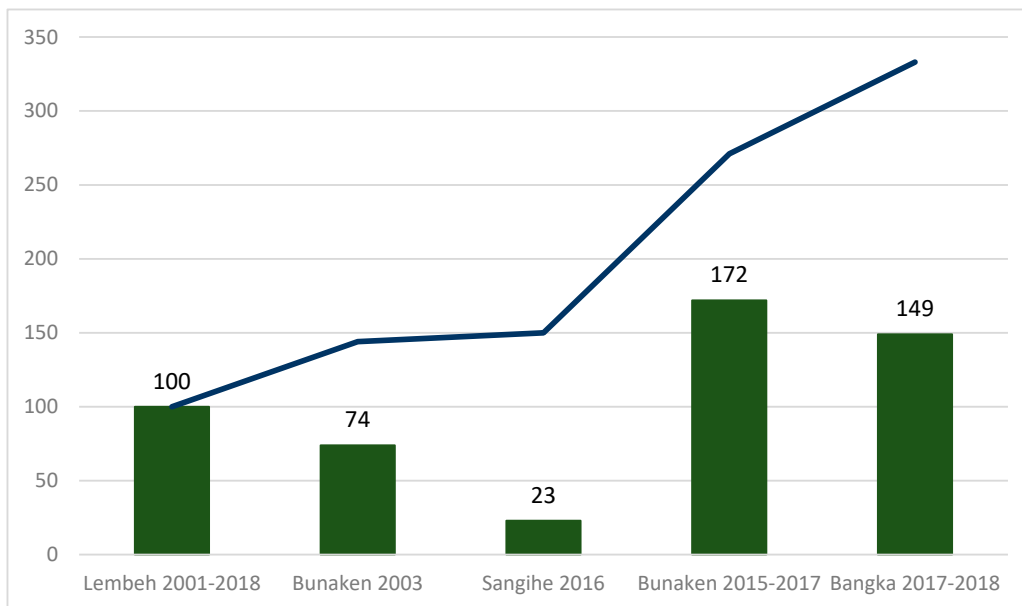


Figure 26. Species accumulation curve and number of species per locality based on the available studies from Lembeh Strait [17,21,39] with the earliest information on diversity from North Sulawesi, Bunaken National Park (BNP) [1,15,16], and Sangihe Island [18].

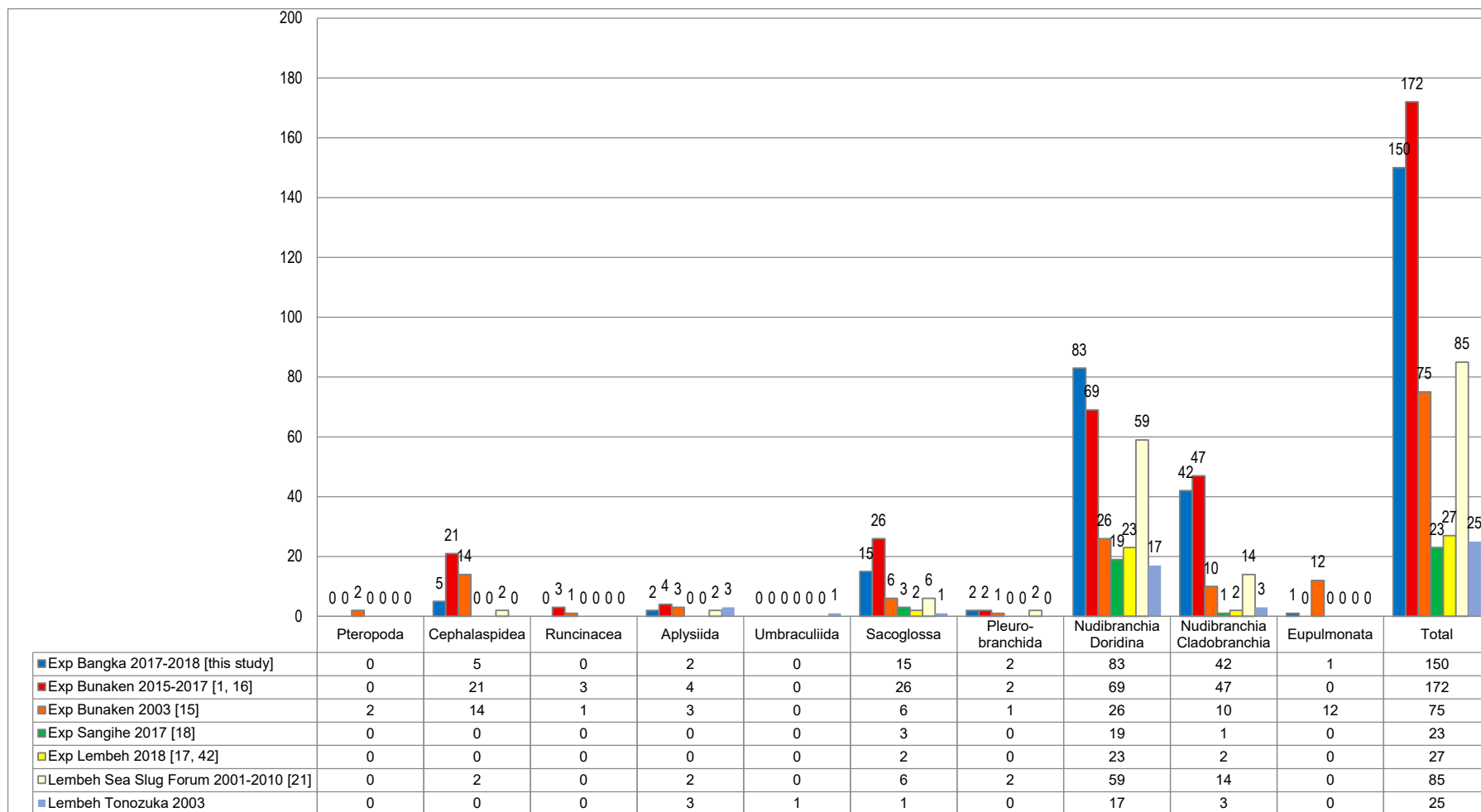


Figure 27. Higher taxa diversity of marine Heterobranchia in Bangka Archipelago compared with other areas in North Sulawesi. Considered are the expeditions to Bunaken National Park [1,15,16], to Sangihe Island [18], and to Lembeh Strait [17,39]. Additional information is accumulated for Lembeh Strait from Sea Slug Forum [21]. Numbers above columns indicate species numbers.

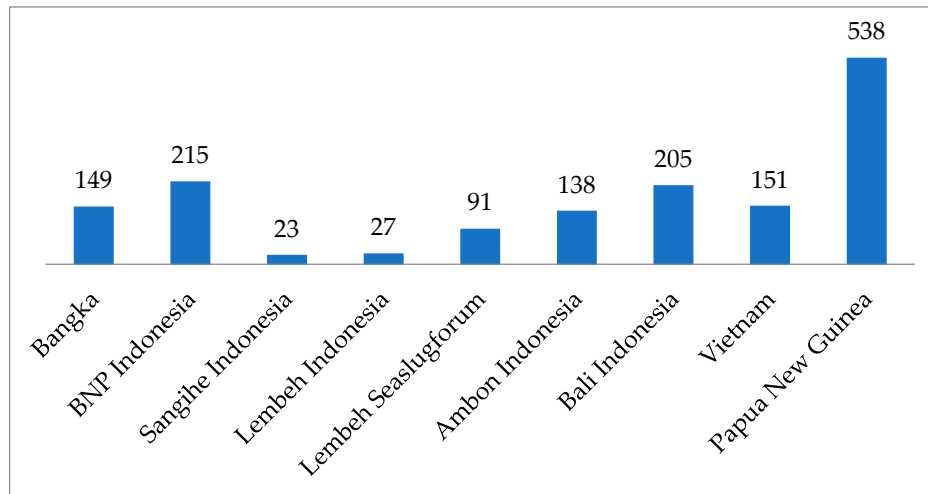


Figure 28. Species diversity from Bangka Archipelago compared with other studies in Indonesia and areas nearby. Data taken from the following literature: BNP, Indonesia [15,16]; Sangihe, Indonesia [18]; Lembeh Island, Indonesia [17,21]; Ambon, Indonesia [3–6]; Bali, Indonesia [39]; Vietnam [22]; and Papua New Guinea [48].

Table 1. Information about collection sites and dates in 2017–2018. *Batu Tiga is the only locality where no samples were collected, and specimens were recorded only by photo-documentation provided by Gianni Valenti.

Area of Collection Site	Location	Characteristics of the Habitat	Year of Collection
Bangka Archipelago			
Batu Belah	1°46'22.20"N 125°10'57.38"E	Sandy bottom with a deposit of large formations of stones richly covered with corals	2017–2018
Batu Tiga *	1°46'07.6"N 125°10'35.7"E	Coral pinnacles on a deep coral sand bottom	2018
Areng Kambing	1°46'07.8"N 125°10'34.6"E	Coral sand and coral rabble slope	2018
Sahaung	1°44'35.11"N 125°09'43.5"E	A shallow area made up of rocks and pinnacles of volcanic origin, richly covered with coral formations	2017
Tanjung Husi	1°44' 07.74"N 125°09'05.59"E	Volcanic rockslide, characterized by the presence of strong tidal currents and ocean swell	2017–2018
Busa Bora Timur	1°44'27.4"N 125°08'35.1"E	Fringing reef on a coral sand slope	2018
Busa Bora Kampung	1°44'39.8"N 125°08'24.4"E	Fringing reef on a coral sand slope in front of a little village	2018
Coral Eye	1°45'03.61"N 125°07'58.85"E	Fringing reef in front of a resort; 2012 was the last storm [1]. Dominant branching coral and little rubble. Coral grown over the tide surface.	2017–2018

Mangrove	1°45'46.9"N 125°07'50.0"E	Fringing reef on a coral sand slope that starts from the lagoon in front of the mangrove	2017–2018
Sipi	1°47'02.8"N 125°07'42.4"E	River mouth: mud/fringing reef, site impacted by mining operations	2018
Pearl Farm	1°48'41.3"N 125°06'45.1"E	Former abandoned pearl farm, fringing reef on a coral sand slope	2018
Kinabuhutan	1°50'45.44"N 125° 5'55.26"E	Fringing reef on a shallow coral sand slope	2018
Talisei	1°53'38.32"N 125°05'53.82"E	A vertical wall of volcanic rock that ends on a sandy bottom. The spot is subject to strong tidal currents and ocean swell, so the coral formations are small and compact.	2018
Mainland			
Yellow Coco	1°40'21.67"N 125° 8'6.60"E	Fringing reef on a mix of volcanic and coral sand slopes	2018
Sempini	1°41'4.83"N 125° 8'32.65"E	Fringing reef on a coral sand slope	2018
Sempini 2	1°40'51.06"N 125° 8'22.45"E	Fringing reef on a coral sand slope	2018
Efrata	1°40'52.34"N 125° 9'10.55"E	Volcanic rock beach, followed by volcanic sand	2018
Batu Mandi	1°41' 16.71"N 125° 09' 42.79"E	Volcanic rock covered by coral reef, sponge, and other invertebrates. Various topographies.	2017–2018
Tanjung Kusu-Kusu	1°41'23.01"N 125°09'55.26"E	Volcanic rock wall on a backdrop of rock and coral, characterized by the presence of strong tidal currents and ocean swell	2017

Table 2. List of species, specimen identifiers, and further metadata of the material; (*) indicate species that were identified only by photo-documentation. The last three columns indicate the number of specimens or occurrence of the respective species in other study areas nearby (information is taken from Eisenbarth et al. (2018) [16], Undap et al. (2019) [18], and Ompi et al. (2019) [17]: BNP, Lembah Strait, and Sangihe Island).

Higher Taxon Affiliation	Identifier	Species Name	Number of Specimens	Expedition in Bangka Archipelago and Mainland Nearby												Other Expeditions									
				Size (mm)	Depth (m)	Bangka Archipelago								Mainland				Exp Bunaken	Exp Sangihe	Exp Lembah					
						Batu Belah	Batu Tiga	Areng Kambang	Sahaung	Tanjung Husi	Busa Bora Timur	Busa Bora Kampung	Coral Eye	Mangrove	Sipi	Pearl Farm	Kinabuhutan				Talisei	Yellow Coco	Sempini	Efrata	Batu Maandi
Cephalaspidea (5 species)																									
Haminoeidae Pilsbry, 1895	Hasp18Ba-1	<i>Haminoea</i> sp. (<i>Haminoea</i> sp. 3 in Gosliner et al. [24]: 30)	1	2	7														1	x	-	-			
Colpodaspididae Oskars, Bouchet and Malaquias, 2015		<i>Colpodaspis thompsoni</i> Brown, 1979	5	2–5	1.5–9.2					2	1	1									x	-	-		
Aglajidae Pilsbry, 1895 (1847)	Odsp.a18Ba-1	<i>Chelidonura amoena</i> Bergh, 1905 <i>Odontoglaia</i> sp. a	2 1	11–25 15	4–9.1 1.5							2									x	-	-		
Gastropteridae Swainson, 1840		<i>Sagaminopteron psychedelicum</i> Carlson and Hoff, 1974	2	3	1														1		x	-	-		
Aplysiida (2 species)																									
Aplysiidae Lamarck, 1809		<i>Aplysia</i> cf. <i>nigrocincta</i> von Martens, 1880 <i>Stylocheilus striatus</i> (Quoy and Gaimard, 1832)	4 2	4–10 5–15	1 7–7.5														4 1			x	-	-	
Sacoglossa (15 species)																									
Oxynoidae Stoliczka, 1868 (1847)	Losp1-18Ba1; Losp1-18Ba2	<i>Lobiger</i> sp. (<i>Lobiger</i> sp. 1 in Gosliner et al. [24]: 70)	2	2–11	8.4					2											x	-	-		
Hermaeidae H. Adams and A. Adams, 1854		<i>Cyerce bourbonica</i> Yonow, 2012 <i>Cyerce nigra</i> Bergh, 1871	2 1	3–4 4	1–1.5 8.6														2		x	-	-		
Plakobranchidae Gray, 1840		<i>Elysia asbecki</i> Wägele, Stemmer, Burghardt and Händeler, 2010	1	3	8.5					1												x	-	-	
		<i>Elysia marginata</i> (Pease, 1871)	1	3	11.6																	1	x	-	-
		<i>Elysia</i> cf. <i>nigropunctata</i> (Pease, 1871)	1	21	2.6														1			-	-	-	
		<i>Elysia pusilla</i> (Bergh, 1871)	2	7–11	1–1.5							1	1									x	x	-	
		Els24-18Ba-1	<i>Elysia</i> sp. 24 (in Gosliner et al. [24]: 89)	1	7	7													1			-	-	-	
	Els27-18Ba-1	<i>Elysia</i> sp. 27 (in Gosliner et al. [24]: 89)	1	5	7													1			-	-	-		
	Els2.a18Ba-1	<i>Elysia</i> sp. a	1	8	1													1			x	-	-		
	Els2.b18Ba-1	<i>Elysia</i> sp. b	1	9	20																x	-	-		

	<i>Hypselodoris apolegma</i> (Yonow, 2001)	1	33	16.4						1	x	-	-
	<i>Hypselodoris bullockii</i> (Collingwood, 1881) *	X								X	-	-	-
Hysp1_17Ba-1	<i>Hypselodoris cerisae</i> Gosliner and Johnson, 2018	1	16	6.6			1					-	-
	<i>Hypselodoris decorata</i> Gosliner and Johnson, 2018	1	20	8.1	1	1			1		1	x	-
	<i>Hypselodoris emma</i> Rudman, 1977 *	X								X		-	-
Chromos18Ba-1	<i>Hypselodoris iacula</i> Gosliner and Johnson, 1999	1	50	15						1		-	-
Hysp19_17Ba-1-2	<i>Hypselodoris lacuna</i> Gosliner and Johnson, 2018	2	6–8	6.8						2		-	-
	<i>Hypselodoris maculosa</i> (Pease, 1871)	4	18–23	3–10.3	1							-	-
	<i>Hypselodoris maridadilus</i> Rudman, 1977	1	10	6.5					1			-	-
	<i>Hypselodoris zephyra</i> (Eliot, 1904)	1	9	14.2	1							-	-
	<i>Hypselodoris tryoni</i> (Garret 1873)	7	26–54	4–14.1					2	2	2	1	x
Thsp1-18Ba-1	<i>Hypselodoris</i> sp. a	1	8	11.5						1		-	-
	<i>Mexichromis aurora</i> (Johnson and Gosliner, 1998)	1	11	8.6					1			-	-
	<i>Mexichromis trilineata</i> (A. Adams and Reeve, 1850) *	X							X			-	-
Dorid18Ba-1	<i>Miamira magnifica</i>	1	2.5	20						1		x	-
Misp17Ba_1	<i>Miamira</i> sp. a	1	10	4					1			-	-
	<i>Verconia simplex</i> Pease, 1871	1	6	5.2					1			-	-
Thfu18Ba-1-2	<i>Thorunna furtiva</i> Bergh, 1878	2	14–16	13.8						2		x	-
Dendrodorididae O'Donoghue, 1924 (1864)	<i>Dendrodoris nigra</i> (Stimpson, 1855)	1	6	1					1			-	-
	<i>Phyllidia</i> cf. <i>babai</i> Brunckhorst, 1993 *	X				X						-	-
	<i>Phyllidia coelestis</i> Bergh, 1905	8	12–43	1.5–20	X		1	1	1	1	3	1	x
	<i>Phyllidia elegans</i> Bergh, 1869	3	34–38	2					1	2			x
	<i>Phyllidia exquisita</i> Brunckhorst, 1993	2	30–32	4.5–12.4			1		1				-
Phyllidiidae Rafinesque, 1814	<i>Phyllidia ocellata</i> Cuvier, 1804	8	14–40	2–13.4	1		1	2		1	2	1	x
	<i>Phyllidia picta</i> Pruvot-Fol, 1957	5	18–35	4–12	2			1	1		1		-
	<i>Phyllidia varicosa</i> Lamarck, 1801	18	23–85	2–20		1	5	1	2	1	5	1	1
Phsp18Ba-2-3	<i>Phyllidia</i> sp. a	2	33–34	6–13.5							1	1	-
	<i>Phyllidiella annulata</i> (Gray, 1853)	1	30	10						1			x
	<i>Phyllidiella lizae</i> Brunckhorst, 1993	4	15.26	5–20						1	1	2	-

	<i>Phyllidiella nigra</i> (van Hasselt 1824)	1	56	1					1										-	x	-	
	<i>Phyllidiella pustulosa</i> (Cuvier, 1804)	39	12–75	1.5–20	5	2	2	1	2	7		1	3	3	2	2	6	1	2	x	x	x
	<i>Phyllidiopsis annae</i> Brunckhorst, 1993	4	3–12	9.2–11.9					2						2					-	-	-
	<i>Phyllidiopsis cf. burni</i> Brunckhorst, 1993	1	39	20											1					-	-	-
	<i>Phyllidiopsis kremptfi</i> Pruvot-Fol, 1957	3	17–56	9.2–16.2		1									1		1			-	x	-
	<i>Phyllidiopsis xishaensis</i> (Lin, 1983)	4	9–17	3.9–16.8				1				2					1			x	-	-
Nudibranchia, Cladobranchia (42 species)																						
	<i>Dermatobranchus caeruleomaculatus</i> Gosliner and Fahey, 2011 *	X													X					-	-	-
Arminidae Iredale and O'Donoghue, 1923	<i>Dermatobranchus rodmanni</i> Gosliner and Fahey, 2011	2	3–4	1					2											-	-	-
	<i>Dermatobranchus pustulosus</i> van Hasselt, 1824	1	70	9.1	1															-	-	-
	Desp.a18Ba-1-2 <i>Dermatobranchus</i> sp. a	2	8–11	1					2											-	-	-
	Desp18Ba-1 <i>Dermatobranchus</i> sp. b	1	6	20.4											1					-	-	-
	Desp17Ba-1 <i>Dermatobranchus</i> sp. c	1	3	3					1											-	-	-
Proctonotidae Gray, 1853	<i>Janolus</i> sp. 1 (in Gosliner et al. [24]: 305) *	X													X					-	-	-
Bornellidae Bergh, 1874	<i>Bornella anguilla</i> Johnson, 1984	2	25–40	7.8–8.5	2															-	-	-
	<i>Bornella stellifera</i> Adams and Reeve in A. Adams, 1848)	1	8	1					1											-	-	-
Tethydidae Rafinesque, 1815	Mesp.a18Ba-1 <i>Melibe bucephala</i> Bergh, 1902	1	9	1					1											-	-	-
	<i>Melibe engeli</i> Riscecc, 1937	1	34	7					1											-	-	-
Dotidae Gray, 1853	<i>Doto ussi</i> Ortea, 1982	3	4–12	4–6.5					2			1								x	-	-
Tritoniidae Lamarck, 1809	Masp2.18Ba-1 <i>Marionia</i> sp. 2 (in Gosliner et al. [24]: 324)	1	40	2					1											-	-	-
	Trsp3_17Ba-1 <i>Tritonia</i> sp. 3 (in Gosliner et al. [24]: 320)	1	13	4					1											-	-	-
Flabellinidae Bergh, 1889	<i>Coryphellina exoptata</i> (Gosliner and Willan, 1991)	3	16–32	2–12.4	1				1			1								x	-	-
	<i>Coryphellina rubrolineata</i> O'Donoghue, 1929	8	20–40	6.3–20.4	1	2						2			1	1		1		x	-	x
	Flsp2-18Ba-1 <i>Flabellina</i> sp. 2 (in Gosliner et al. [24]: 333)	1	16	14.5					1											-	-	-
	Flsp3_17Ba-1 <i>Flabellina</i> sp. 3 (in Gosliner et al. [24]: 333)	1	11	13.6											1					-	-	-
Samlidae Korshunova, Martynov, Bakken, Evertsen, Fletcher,	<i>Samla riwo</i> (Gosliner and Willan, 1991)	3	9–13	4–8.6					2						1					x	-	-

Mudianta, Saito, Lundin, Schrödl and Picton, 2017																											
Eubrancheidae Odhner, 1934	<i>Eubrancheus</i> sp. 22 (in Gosliner et al. [24]: 341)	1	6	5.1		1											x	-	-								
Trinchesiidae F. Nordsieck, 1972	<i>Trinchesia yamasui</i> (Hamatani, 1993)	X															X	-	-	-							
Cuthonidae Odhner, 1934	<i>Cuthona</i> sp. 57 (in Gosliner et al. [24]: 353)	2	17–25	12.7															-	-	-						
	Aeol18Ba-1 <i>Aeolidia</i> sp. a	1	8	1.5																-	-	-					
	Ansp17Ba-1 <i>Antonieta</i> sp. a	1	2	6.4																	1	-	-	-			
	<i>Caloria indica</i> (Bergh, 1896)	15	2–38	5–13.4	5		2		3		1				1	2	1			x	-	-					
	Crsp17Ba-1 <i>Cratena</i> sp. a	1	8	4					1												-	-	-				
	<i>Favorinus japonicus</i> Baba, 1949	2	10–16	8.9–20							1				1					x	-	-					
	Fasp1-18Ba-1-2 <i>Favorinus</i> sp. 1 (in Gosliner et al. [24]: 363)	2	13–15	20													2				-	-	-				
	<i>Favorinus tsuruganus</i> Baba and Abe, 1964	1	11	2																	-	-	-				
	Mojo18Ba-1 <i>Moridilla</i> sp. a	2	12	6.8–11.2	1		1														-	-	-				
	<i>Noumeaella</i> sp. 2 (in Gosliner et al. [24]: 367)	1	11	4																	-	-	-				
	<i>Noumeaella</i> sp. 3 (in Gosliner et al. [24]: 367)	6	8–12	4–9																	x	-	-				
	<i>Noumeaella</i> sp. 13 (in Gosliner et al. [24]: 369)	7	7–16	3–9																	-	-	-				
Facelinidae Bergh, 1889	<i>Phylloidesmium briareum</i> (Bergh, 1896)	17	15–60	3–20											1						x	-	-				
	<i>Phylloidesmium</i> cf. <i>crypticum</i> Rudman, 1981	5	8–22	4																	-	-	-				
	<i>Phylloidesmium lizardense</i> Burghardt, Schrödl and Wägele, 2008	3	10–13	1–10						1	2										-	-	-				
	<i>Phylloidesmium longicirrum</i> (Bergh, 1905)	1	125	25																		1	-	-	-		
	<i>Phylloidesmium magnum</i> Rudman, 1991	2	50	5.5–11.5			1		1												-	-	-				
	<i>Phylloidesmium parangatum</i> Ortiz and Gosliner, 2003	5	8–15	1–7																	-	-	-				
	<i>Phylloidesmium pecten</i> Rudman, 1981	1	15	1																	-	-	-				
	<i>Phylloidesmium poindimiei</i> (Risbec, 1928)	2	14–21	4–8.9																		1	x	-	-		
	<i>Pteracolidia semperi</i> (Bergh, 1870)	25	13–54	5–8.9	2		2		1													15	2	3	x	-	-
Eupulmonata (1 species)																											
Onchidiidae	<i>Peronia</i> sp. a	1	39	1																		1	-	-	-		
Total		149			34	0	11	14	40	14	23	162	8	13	16	7	12	23	41	30	23	14	-	-	-		

Table 3. Compilation of the species recorded in this study and those recently published from Bunaken National Park [16], a previous study from Bunaken National Park [15], from Sangihe Island [18], and three sources on Lembeh Strait: Ompi et al. (2019) [17], species records from Lembeh Strait available from the seaslugforum.net [21] in the time period of 2001–2010, and data extracted from Tonozuka [39].

Expedition in Bangka Archipelago and Adjacent Islands									
Higher Taxon Affiliation	Identifier	Species Name	Exp Bangka 2017–2018	Exp Bunaken 2015–2017	Exp Bunaken 2003	Exp Sangihe 2016	Exp Lembeh 2018	Lembeh Sea Slug Forum 2001–2010	Tonozuka 2003
Pteropoda									
Limacinidae Gray, 1840		<i>Limacina</i> cf. <i>helicina</i> (Phipps, 1774)			X				
Cavoliniidae Gray, 1850 (1815)		<i>Cavolinia globulosa</i> Gray, 1850			X				
Cephalaspidea									
Bullidae Lamarck, 1801		<i>Bulla</i> cf. <i>ampulla</i> Linnaeus, 1758			X				
		<i>Alys</i> cf. <i>semistriata</i> Pease, 1860			X				
		<i>Haminoea curta</i> (Adams, 1850)			X				
	Hasp15Bu-1, Hasp16Bu-1	<i>Haminoea</i> sp. (<i>Haminoea</i> sp. 2 in Gosliner et al. [24]: 30)		X	X				
Haminoeidae Pilsbry, 1895		<i>Haminoeid</i> sp. (<i>Haminoeid</i> sp. 2 in Gosliner et al. [24]: 34)		X					
	Hasp18Ba-1	<i>Haminoea</i> sp. (<i>Haminoea</i> sp. 3 in Gosliner et al. [24]: 30)	X						
	Hasp2_15Bu-1	<i>Haminoea</i> sp.		X					
	Hasp2_16Bu-1	<i>Haminoea</i> sp.		X					
		<i>Limulatys</i> cf. <i>ooformis</i> Habe, 1952			X				
		<i>Phanerophthalmus</i> cf. <i>albocollaris</i>		X	X				

		Heller and Thompson, 1983				
		Phanerophthalmus olivaceus (Ehrenberg, 1828) as <i>P.</i> <i>smaragdinus</i>			X	
		<i>Phanerophthalmus</i> sp. (<i>Phanerophthalmu</i> s sp. 3 in Gosliner et al. [24]: 33)		X		
Philinidae Gray, 1850 (1815)	Ilsp17Bu-1	<i>Philine</i> sp.		X	X	
		<i>Philinoglossa</i> <i>marcusi</i> Challis, 1969			X	
Philinoglossidae Hertling, 1932		<i>Philinoglossa</i> sp. (in Gosliner et al. [24]: 103)			X	
Colpodaspididae Oskars, Bouchet and Malaquias, 2015		<i>Colpodaspis</i> <i>thompsoni</i> Brown, 1979	X	X	X	
	Agsp15Bu-1	<i>Aglajidae</i> sp.		X		
		<i>Chelidonura amoena</i> Bergh, 1905	X	X	X	X
		<i>Chelidonura</i> <i>hirundinina</i> (Quoy and Gaimard, 1833)		X	X	
Aglajidae Pilsbry, 1895 (1847)	Odsp.a18Ba-1	<i>Odontoglaja</i> cf. <i>guamensis</i> Rudman, 1978	X	X		
		<i>Philinopsis</i> <i>speciosa</i> Pease, 1860				X
	Phisp16Bu-1	<i>Tubulophilinopsis</i> sp.		X		
		<i>Gastropteron</i> sp. (<i>Gastropteron</i> sp. 5 in Gosliner et al. [24]: 56)		X		
Gastropteridae Swainson, 1840	Gasp5_16Bu-1	<i>Sagaminopteron</i> <i>psychedelicum</i>	X	X		

		Carlson and Hoff, 1974			
	Sasp17Bu-1	<i>Sagaminopteron</i> sp.	X		
		<i>Siphopteron brunneomarginatum</i> (Carlson and Hoff, 1974)	X		
		<i>Siphopteron ladrones</i> (Carlson and Hoff, 1974)	X		
		<i>Siphopteron nigromarginatum</i> Gosliner, 1989	X		
		<i>Siphopteron tigrinum</i> Gosliner, 1989	X	X	
	Sini15Bu-19+20	<i>Siphopteron</i> sp.	X		
			Runcinacea		
	Rusp15Bu-1	<i>Runcina</i> sp.	X	X	
	Rusp16Bu-1	<i>Runcina</i> sp.	X		
	Rusp2_16Bu-1	<i>Runcina</i> sp.	X		
	Rusp3_16Bu-1				
			Aplysiida		
		<i>Aplysia oculifera</i> A. Adams and Reeve, 1850			X
		<i>Bursatella leachii</i> Blainville, 1817		X	
		<i>Dolabella auricularia</i> (Lightfoot, 1786)			X
		<i>Petalifera ramosa</i> Baba, 1959			X
Aplysiidae Lamarck, 1809		<i>Aplysia</i> cf. <i>nigrocincta</i> von Martens, 1880	X	X	
		<i>Stylocheilus striatus</i> (Quoy and Gaimard, 1832)	X	X	X

		<i>Dolabella auricularia</i> (Lightfoot, 1786)	X	X	
		<i>Dolabrifera dolabrifera</i> (Rang, 1828)	X		
		<i>Phyllaplysia</i> sp.		X	
		<i>Syphonota geographica</i> (A. Adams and Reeve, 1850)			X
Umbraculida					
Umbraculiidae Dall, 1889 (1827)		<i>Umbraculum umbraculum</i> (Lightfoot, 1786)			X
Sacoglossa					
Cylindrobullidae Thiele, 1931	Assp1_17Bu-1-4	<i>Cylindrobulla</i> sp.	X		
		<i>Lobiger nevilli</i> Pilsbry, 1896	X		
Oxynoidae Stoliczka, 1868 (1847)	Losp1-18Ba1; Losp1-18Ba2	<i>Lobiger</i> sp. (<i>Lobiger</i> sp. 1 in Gosliner et al. [24]: 70)	X	X	
		<i>Cyerce bourbonica</i> Yonow, 2012	X	X	X as <i>C. sp. 1</i>
		<i>Cyerce elegans</i> Bergh, 1870			X
		<i>Cyerce nigra</i> Bergh, 1871	X		
Hermaeidae H. Adams and A. Adams, 1854	Cysp2_15Bu-5	<i>Cyerce</i> sp.	X		
		<i>Hermaea</i> sp. (<i>Hermaea</i> sp. 2 in Gosliner et al. [24]: 81); <i>Aplysiopsis</i> sp. 1 in Sea Slug Forum)			X
		<i>Sohgenia palauensis</i> Hamatani, 1991	X		
Costasiellidae Clarke, 1984	Cosp17Bu-1	<i>Costasiella kuroshimae</i> Ichikawa, 1993	X		X as <i>Chromodoris</i> sp. 3

	Cosp1_17Bu-1-2	<i>Costasiella</i> sp. (<i>Costasiella</i> sp. 1 in Gosliner et al. [24]: 79)		X				
	Cosp8_17Bu-1	<i>Costasiella</i> sp. (<i>Costasiella</i> sp. 8 in Gosliner et al. [24]: 81)		X				
	Cosp3_16Bu-1-5	<i>Costasiella</i> sp.		X				
Plakobranchidae Gray, 1840		<i>Elysia asbecki</i> Wägele, Stemmer, Burghardt and Händeler, 2010	X	X				
		<i>Elysia grandifolia</i> Kelaart, 1858					X	
		<i>Elysia marginata</i> (Pease, 1871)	X	X				
		<i>Elysia mercieri</i> (Pruvot-Fol, 1930)		X				
		<i>Elysia</i> cf <i>nigropunctata</i> (Pease, 1871)	X					
		<i>Elysia ornata</i> (Swainson, 1840)					X	
		<i>Elysia pusilla</i> (Bergh, 1871)	X	X	X	X		
		Elspl.a18Ba-1	<i>Elysia</i> sp. (<i>Elysia</i> sp. a)	X	X			
		Elspl.b18Ba-1	<i>Elysia</i> sp. (<i>Elysia</i> sp. b)	X	X			
		Elspl_16Bu-1	<i>Elysia</i> sp.		X			
		Elspl6Bu-1	<i>Elysia</i> sp.		X			
		Elspl4_16Bu-1	<i>Elysia</i> sp.		X			
		Elspl24-18Ba-1	<i>Elysia</i> sp. 24 (in Gosliner et al. [24]: 89)	X				
	Elspl27-18Ba-1	<i>Elysia</i> sp. 27 (in Gosliner et al. [24]: 89)	X					

	<i>Plakobranchnus ocellatus</i> van Hasselt, 1824		X				
	<i>Plakobranchnus</i> cf. <i>papua</i> Meyers-Muños and van der Velde, 2016					X	
	<i>Thuridilla albopustulosa</i> Gosliner, 1995		X				X
	<i>Thuridilla carlsoni</i> Gosliner, 1995	X					
	<i>Thuridilla flavomaculata</i> Gosliner, 1995	X	X				
	<i>Thuridilla vataae</i> (Risbec, 1928)	X	X				
	<i>Thuridilla gracilis</i> (Risbec, 1928)	X	X	X as <i>T.bayeri</i>	X		X
	<i>Thuridilla</i> cf. <i>hoffae</i> Gosliner, 1995			X			
	<i>Thuridilla lineolate</i> (Bergh, 1905)		X	X			X
	<i>Thuridilla livida</i> (Baba, 1955)		X				
	<i>Thuridilla undula</i> Gosliner, 1995		X				
	Pleurobranchida						
	<i>Berthellina citrina</i> (Pease, 1861)		X				
	<i>Berthella martensi</i> (Pilsbry, 1896)						X
Pleurobranchidae Gray, 1827	<i>Pleurobranchus forskalii</i> Rüppell and Leuckart, 1828	X	X				
	<i>Pleurobranchus peronii</i> Cuvier, 1804	X		X			X

Nudibranchia, Doridina						
Hexabranchidae Bergh, 1891	Glsp1_17Ba-1	<i>Hexabranchus sanguineus</i> (Rüppell and Leuckart, 1830)	X	X	X	
		<i>Kaloplocamus dokte</i> Vallès and Gosliner, 2006		X		
	Kalsp8_16Bu-1	<i>Kaloplocamus</i> sp. (<i>Kaloplocamus</i> sp. 8 in Gosliner et al. [24]: 116)		X		
		Kalsp9_16Bu-1	<i>Kaloplocamus</i> sp. (<i>Kaloplocamus</i> sp. 9 in Gosliner et al. [24]: 116)		X	
Polyceridae Alder and Hancock, 1845			<i>Nembrotha chamberlaini</i> Gosliner and Behrens, 1997	X		
		<i>Nembrotha cristata</i> Bergh, 1877	X	X		
		<i>Nembrotha kubaryana</i> Bergh, 1877	X	X	X	X
		<i>Nembrotha lineolata</i> Bergh, 1905	X			
		<i>Nembrotha megalocera</i> Yonow, 1990				X misidentified?
		<i>Nembrotha milleri</i> Gosliner and Behrens, 1997	X			
		<i>Nembrotha mullineri</i> Gosliner and Behrens, 1997	X			
		<i>Nembrotha purpureolineata</i> O'Donoghue, 1924				X as <i>N. rutilans</i>
	Nesp1_17Ba-1	<i>Nembrotha</i> sp. 1	X			

	(in Gosliner et al. [24]: 122)			
	<i>Polycera japonica</i> Baba, 1949		X	
	<i>Polycera risbeci</i> Odhner, 1941		X	
Posp516Bu-1	<i>Polycera</i> sp. (<i>Polycera</i> sp. 5 in Gosliner et al. [24]: 113)		X	
	<i>Roboastra gracilis</i> (Bergh, 1877)		X	X
	<i>Tambja gabriellae</i> Pola, Cervera and Gosliner, 2005	X		X
	<i>Tambja morosa</i> (Bergh, 1877)	X		X
	<i>Thecacera picta</i> Baba, 1972			X
	<i>Gymnodoris aurita</i> (Gould, 1852) *	X		
	<i>Gymnodoris citrina</i> (Bergh, 1877)			X
Gysp2_16Bu-1	<i>Gymnodoris</i> sp. (<i>Gymnodoris</i> sp. 2 in Gosliner et al. [24]: 152)		X	
Gysp16Bu-1	<i>Gymnodoris</i> sp. (<i>Gymnodoris</i> cf. sp. 35 in Gosliner et al. [24]: 159)		X	
Gysp1_17Bu-1 Gysp22_17Bu-1	<i>Gymnodoris</i> sp. (<i>Gymnodoris</i> cf. sp. 46 in Gosliner et al. [24]: 161)		X	
Gysp1_15Bu-2	<i>Gymnodoris</i> sp.		X	
	<i>Gymnodoris tuberculosa</i> Knutson and Gosliner, 2014	X	X	

		<i>Thecacera</i> sp.				X
	Gysp20-18Ba-1	<i>Gymmodoris</i> sp. 20 (in Gosliner et al. [24]: 156)	X			
	Gysp25-18Ba-1-2	<i>Gymmodoris</i> sp. 25 (in Gosliner et al. [24]: 157)	X			
	Gosp7-18Ba-1	<i>Goniodoris</i> sp. 7 (in Gosliner et al. [24]: 153)	X			
		<i>Okenia</i> <i>kendi</i> Gosliner, 2004				X
		<i>Okenia</i> <i>pellucida</i> Burn, 1967				X
Goniodorididae H. Adams and A. Adams, 1854		<i>Trapania armilla</i> Gosliner and Fahey, 2008	X			X
		<i>Trapania euryeia</i> Gosliner and Fahey, 2008			X	
		<i>Trapania safracornia</i> Fahey, 2004	X			
		<i>Aegires citrinus</i> Pruvot-Fol, 1930			X	
	Aesp7-18Ba-1	<i>Aegires</i> sp. 7 (in Gosliner et al. [24]: 149)	X			
Aegiridae P. Fischer, 1883		<i>Notodoris minor</i> Eliot, 1904 *	X			X
		<i>Notodoris serенаe</i> Gosliner and Behrens, 1997		X	X	
Doridoidea Rafinesque, 1815	Dosp17Bu-3	<i>Doridoidea</i> sp.		X		
	Scsp1_17Bu-1	<i>Doridoidea</i> sp.		X		
Cadlinidae Bergh, 1891		<i>Aldisa albatrossae</i> Elwood, Valdés and Gosliner, 2000				X
Discodorididae Bergh, 1891		<i>Asteronotus</i> <i>cespitosus</i> (van Hasselt, 1824) *	X	X		X

	<i>Asteronotus mimeticus</i> Gosliner and Valdés, 2002	X	X					
	<i>Atagema intecta</i> (Kelaart, 1859)	X						X
Disp1_Bu-1	<i>Diaulula</i> sp. (<i>Diaulula</i> sp. 1 in Gosliner et al. [24]: 197)		X					
	<i>Carminodoris estrelyado</i> (Gosliner and Behrens, 1998)					X		X
	<i>Discodoris cebuensis</i> Bergh, 1877	X						
	<i>Halgerda batangas</i> Carlson and Hoff, 2000	X	X	X		X	X	
	<i>Halgerda carlsoni</i> Rudman, 1978	X	X					
	<i>Halgerda okinawa</i> Carlson and Hoff, 2000						X	
	<i>Halgerda tessellate</i> (Bergh, 1880)		X					
	<i>Jorunna funebris</i> Kelaart, 1859	X				X	X	
	<i>Paradoris liturata</i> (Bergh, 1905)	X					X	
	<i>Platydoris ellioti</i> (Alder and Hancock, 1864)						X	
	<i>Platydoris formosa</i> (Alder and Hancock, 1864)							X
	<i>Platydoris inframaculata</i> (Abraham, 1877)					X		

		<i>Platydorid sanguinea</i> Bergh, 1905	X	X					
		<i>Sclerodoris</i> <i>tuberculata</i> Eliot, 1904						X	
	Scsp2_16Bu-1	<i>Sclerodoris</i> sp. (<i>Sclerodoris</i> sp. 2 in Gosliner et al. [24]: 195)		X					
		<i>Taringa halgerda</i> Gosliner and Behrens, 1998		X					
	Dosp17Bu-1	<i>Discodorididae</i> sp.		X					
	Dosp17Bu-2	<i>Discodorididae</i> sp.		X					
		<i>Ardeadoris averni</i> (Rudman, 1985)							X
		<i>Ardeadoris cruenta</i> (Rudman, 1986)						X	
		<i>Ceratosoma bicolor</i> Baba, 1949						X	
		<i>Ceratosoma tenue</i> Abraham, 1876 *	X					X	X
		<i>Ceratosoma</i> <i>trilobatum</i> (J.E. Gray, 1827)						X	X
Chromodorididae Bergh, 1891	Cesp2_15Bu-3 Cesp1_17Bu-1	<i>Ceratosoma</i> sp. (<i>Ceratosoma</i> sp. 1 in Gosliner et al. [24]: 266)		X					
		<i>Chromodoris annae</i> Bergh, 1877	X	X	X	X	X	X	
		<i>Chromodoris</i> cf. <i>boucheti</i> Rudman, 1982		X					
		<i>Chromodoris</i> <i>colemanni</i> Rudman, 1982						X	
		<i>Chromodoris diana</i> Gosliner and Behrens, 1998	X	X	X	X			

<i>Chromodoris elisabethina</i> Bergh, 1877 *	X						
<i>Chromodoris lochi</i> Rudman, 1982	X	X	X		X	X	X
<i>Chromodoris magnifica</i> (Quoy and Gaimard, 1832)	X					X	
<i>Chromodoris cf. michaeli</i> Gosliner and Behrens, 1998			X				
<i>Chromodoris quadricolor</i> (Rüppell and Leuckart, 1830)	X						
<i>Chromodoris strigata</i> Rudman, 1982	X	X	X	X			X
<i>Chromodoris willani</i> Rudman, 1982	X	X	X				X
<i>Doriprismatica atromarginata</i> (Cuvier, 1804)	X						
<i>Doriprismatica sibogae</i> Berg, 1905	X						
<i>Doriprismatica stellata</i> (Rudman, 1986)			X				
<i>Glossodoris cincta</i> (Bergh, 1888)	X	X		X	X		X
<i>Glossodoris hikuensis</i> (Pruvot-Fol, 1954)			X				
<i>Glossodoris rufomarginata</i> (Bergh, 1890) *	X						X
<i>Goniobranchus aureopurpureus</i>							X

	(Collingwood, 1881)						
	<i>Goniobranchus coi</i> (Risbec, 1956)	X					
	<i>Goniobranchus decorus</i> (Pease, 1860)			X			
	<i>Goniobranchus fidelis</i> (Kelaart, 1858)	X	X			X	X
	<i>Goniobranchus geometricus</i> (Risbec, 1928)	X	X	X	X	X	X
	<i>Goniobranchus hintuanensis</i> (Gosliner and Behrens, 1998)						X
	<i>Goniobranchus kuniei</i> (Pruvot-Fol, 1930)	X					X
	<i>Goniobranchus preciosus</i> (Kelaart, 1858)						X
	<i>Goniobranchus reticulatus</i> (Quoy and Gaimard, 1832)	X	X		X	X	X
	<i>Goniobranchus roboi</i> (Gosliner and Behrens, 1998)						X
	<i>Goniobranchus tinctorius</i> (Rüppell and Leuckart, 1830)			X			
	<i>Goniobranchus verrieri</i> (Crosse, 1875)	X					X
Gosp40_16Bu-1	<i>Goniobranchus</i> sp. (<i>Goniobranchus</i> sp. 40 in Gosliner et al. [24]: 230)		X				

	<i>Hypselodoris apolegma</i> (Yonow, 2001)	X	X	X		X
	<i>Hypselodoris bullockii</i> (Collingwood, 1881) *	X				
Hysp1_17Ba-1	<i>Hypselodoris cerisae</i> Gosliner and Johnson, 2018	X				X
	<i>Hypselodoris decorata</i> Gosliner and Johnson, 2018	X				
	<i>Hypselodoris emma</i> Rudman, 1977 *	X				
Chromos18Ba-1	<i>Hypselodoris iacula</i> Gosliner and Johnson, 1999	X				
	<i>Hypselodoris iba</i> Gosliner and Johnson, 2018					X
	<i>Hypselodoris infucata</i> (Rüppell and Leuckart, 1830)					X
	<i>Hypselodoris kanga</i> Rudman, 1977					X
	<i>Hypselodoris cf. krakatoa</i> Gosliner and Johnson, 1999					X
Hysp19_17Ba-1-2	<i>Hypselodoris lacuna</i> Gosliner and Johnson, 2018	X				
	<i>Hypselodoris maculosa</i> (Pease, 1871)	X	X			X
	<i>Hypselodoris zephyra</i> (Eliot, 1904)	X				

	<i>Hypselodoris nigrostriata</i> (Eliot, 1904)	X						X	
	<i>Hypselodoris purpureomaculosa</i> Hamatani, 1995							X	X
	<i>Hypselodoris tryoni</i> (Garret, 1873)	X	X	X	X	X		X	
Thsp1-18Ba-1	<i>Hypselodoris</i> sp. a	X	X					X	
Hysp16Bu-1	<i>Hypselodoris</i> sp.		X						
Hysp2_16Bu-1	<i>Hypselodoris</i> sp.		X						
	<i>Mexichromis aurora</i> (Johnson and Gosliner, 1998)	X							
	<i>Mexichromis multituberculata</i> (Baba, 1953)							X	
	<i>Mexichromis trilineata</i> (A. Adams and Reeve, 1850)	X						X	
	<i>Miamira sinuata</i> (van Hasselt, 1824)			X	X				
Dorid18Ba-1	<i>Miamira magnifica</i>	X							
Misp17Ba_1	<i>Miamira</i> sp. a	X							
	<i>Verconia simplex</i> Pease, 1871	X							
	<i>Thorunna florens</i> (Baba, 1949)							X	
	<i>Thorunna furtiva</i> Bergh, 1878	X	X						
	<i>Verconia varians</i> (Pease, 1871)			X	X				
Dosp17Bu-4	<i>Verconia</i> sp.		X						
Dendrodorididae O'Donoghue, 1924 (1864)	<i>Dendrodoris albobrunnea</i> Allan, 1933		X						

	<i>Dendrodoris carbunculosa</i> (Kelaart, 1858)						X	
	<i>Dendrodoris elongata</i> Baba, 1936			X				
	<i>Dendrodoris guttata</i> (Odhner, 1917)						X	
	<i>Dendrodoris nigra</i> (Stimpson, 1855)	X	X					
Rosp17Bu-1	<i>Dendrodoris</i> sp.		X					
	<i>Phyllidia</i> cf. <i>babai</i> Brunckhorst, 1993 *	X					X	X
	<i>Phyllidia coelestis</i> Bergh, 1905	X	X	X	X			
	<i>Phyllidia elegans</i> Bergh, 1869	X	X	X		X	X	
	<i>Phyllidia exquisita</i> Brunckhorst, 1993	X					X	
	<i>Phyllidia madangensis</i> Brunckhorst, 1993					X		
	<i>Phyllidia ocellata</i> Cuvier, 1804	X	X	X	X		X	
Phyllidiidae Rafinesque, 1814	<i>Phyllidia picta</i> Pruvot-Fol, 1957	X			X			
	<i>Phyllidia polkadotsa</i> Brunckhorst, 1993						X	
Phsp18Ba-2-3	<i>Phyllidia</i> sp. a	X	X		X			
	<i>Phyllidia varicosa</i> Lamarck, 1801	X	X	X	X	X	X	
	<i>Phyllidiella annulata</i> (Gray, 1853)	X	X					
	<i>Phyllidiella lizae</i> Brunckhorst, 1993	X		X	X			
	<i>Phyllidiella nigra</i> (van Hasselt 1824)	X		X	X			
	<i>Phyllidiella pustulosa</i> (Cuvier, 1804)	X	X	X	X	X	X	

	<i>Phyllidiopsis annae</i> Brunckhorst, 1993	X			X
Phssp.aBa-1	<i>Phyllidiopsis</i> cf. <i>burni</i> Brunckhorst, 1983	X			
	<i>Phyllidiopsis krempfi</i> Pruvot-Fol, 1957	X		X	
	<i>Phyllidiopsis pipeki</i> Brunckhorst, 1993		X		
	<i>Phyllidiopsis shireenae</i> Brunckhorst, 1990			X	X
	<i>Phyllidiopsis sphingis</i> Brunckhorst, 1993		X		
	<i>Phyllidiopsis xishaensis</i> (Lin, 1983)	X	X	X	
Nudibranchia, Cladobranchia					
	<i>Dermatobranchus caeruleomaculatus</i> Gosliner and Fahey, 2011 *	X			
	<i>Dermatobranchus diagonalis</i> Gosliner and Fahey, 2011		X		
Arminidae Iredale and O'Donoghue, 1923	<i>Dermatobranchus fasciatus</i> Gosliner and Fahey, 2011		X		
	<i>Dermatobranchus</i> cf. <i>kokonas</i> Gosliner and Fahey, 2011		X		
	<i>Dermatobranchus</i> cf. <i>piperoides</i> Gosliner and Fahey, 2011		X		

		<i>Dermatobranchus rodmani</i> Gosliner and Fahey, 2011	X		
		<i>Dermatobranchus pustulosus</i> van Hasselt, 1824	X		
	Desp8_17Bu-1	<i>Dermatobranchus</i> sp. (<i>Dermatobranchus</i> sp. 8 in Gosliner et al. [24]: 302)		X	
	Desp17Ba-1	<i>Dermatobranchus</i> sp.	X		
	Desp.a18Ba-1-2	<i>Dermatobranchus</i> sp. a	X		
	Desp1_16Bu-1 Desp18Ba-1	<i>Dermatobranchus</i> sp.	X	X	
		<i>Janolus</i> cf. <i>mirabilis</i> Baba and Abe, 1970		X	X
Proctonotidae Gray, 1853		<i>Janolus savinkini</i> Martynov and Korshunova, 2012			X as <i>Janolus</i> sp. 4
		<i>Janolus</i> sp. 1 (in Gosliner et al. [24]: 305)*	X	X	
Scyllaeidae Alder and Hancock, 1855	Cross16Bu-1-8	<i>Crosslandia daedali</i> Poorman and Mulliner, 1981		X	
		<i>Bornella anguilla</i> Johnson, 1984	X		
Bornellidae Bergh, 1874		<i>Bornella stellifera</i> (A. Adams and Reeve [in A. Adams], 1848)	X		X
		<i>Pseudovermis</i> cf. <i>mortoni</i> Challis, 1969			X
Pseudovermidae Thiele, 1931	Mesp.a18Ba-1	<i>Melibe bucephala</i> Bergh, 1902	X		
Tethydidae Rafinesque, 1815		<i>Melibe engeli</i> Risbec, 1937	X		

		<i>Melibe viridis</i> (Kelaart, 1858)					X
		<i>Doto ussi</i> Ortea, 1982	X	X			
		<i>Kabeiro</i> <i>rubroreticulata</i> Shipman and Gosliner, 2015				X	
Dotidae Gray, 1853	Dotosp15Bu-1	<i>Kabeiro</i> sp.				X	
	Kasp16Bu-1-15+17 Kasp17Bu-1	<i>Kabeiro</i> sp.				X	
	Kasp16Bu-16	<i>Kabeiro</i> sp.				X	
		<i>Marianina rosea</i> (Pruvot-Fol, 1930)				X	
	Masp2.18Ba-1	<i>Marionia</i> sp. 2 (in Gosliner et al. [24]: 324)	X				
	Trsp3_17Ba-1	<i>Tritonia</i> sp. 3 (in Gosliner et al. [24]: 320)	X				
Tritoniidae Lamarck, 1809	Trsp8_16Bu-1	<i>Tritonia</i> sp. (<i>Tritonia</i> sp. 9 in Gosliner et al. [24]: 321)				X	
	Trisp10_16Bu-1	<i>Tritonia</i> sp. (<i>Tritonia</i> sp. 10 in Gosliner et al. [24]: 321)				X	
	Trsp16Bu-1	<i>Tritonia</i> sp.				X	
		<i>Embletonia gracilis</i> Risbec, 1928				X	
Embletoniidae		<i>Coryphellina delicate</i> (Gosliner and Willan, 1991)				X	
Flabellinidae Bergh, 1889		<i>Coryphellina</i> <i>exoptata</i> (Gosliner and Willan, 1991)	X	X	X		X
		<i>Coryphellina</i> <i>rubrolineata</i>	X	X		X	X

		O'Donoghue, 1929		
	Flsp2-18Ba-1	<i>Flabellina</i> sp. 2 (in Gosliner et al. [24]: 333)	X	
	Flsp3_17Ba-1	<i>Flabellina</i> sp. 3 (in Gosliner et al. [24]: 333)	X	
		<i>Flabellina</i> sp. 8 (Sea Slug Forum)		X
		<i>Samla bicolor</i> (Kelaart, 1858)	X	
		<i>Samla bicolor</i> (Kelaart, 1858)		X
Samliidae Korshunova, Martynov, Bakken, Evertsen, Fletcher, Mudianta, Saito, Lundin, Schrödl and Picton, 2017		<i>Samla rivoo</i> (Gosliner and Willan, 1991)	X	X
		<i>Phestilla lugubris</i> (Bergh, 1870)		X
Trinchesiidae F. Nord sieck, 1972		<i>Phestilla minor</i> Rudman, 1981		X
		<i>Trinchesia yamasui</i> (Hamatani, 1993)	X	
		<i>Eubbranchus</i> sp. (<i>Eubbranchus</i> sp. 22 in Gosliner et al. [24]: 341)	X	X
Eubbranchidae Odhner, 1934	Cusp4_16Bu-1	<i>Cuthona</i> sp. (<i>Cuthona</i> sp. 4 in Gosliner et al. [24]: 343)		X
	Cusp54_16Bu-1	<i>Cuthona</i> sp. (<i>Cuthona</i> sp. 54 in Gosliner et al. [24]: 353)		X
Cuthonidae Odhner, 1934		<i>Cuthona</i> sp. 57 (in Gosliner et al. [24]: 353)	X	
	Cusp65_16Bu-1	<i>Cuthona</i> sp. (<i>Cuthona</i> cf. sp. 65 in		X

		Gosliner et al. [24]: 343)			
	Aeol18Ba-1	<i>Aeolidia</i> sp.	X		X X
Aeolidiidae Gray, 1827		<i>Bulbaeolidia alba</i> (Risbec, 1928)		X	X
		<i>Baeolidia australis</i> (Rudman, 1982)			X
		<i>Cerberilla</i> <i>ambonensis</i> Bergh, 1905			X
		Ansp17Ba-1	<i>Antonietta</i> sp.	X	
			<i>Caloria indica</i> (Bergh, 1896)	X	X
Facelinidae Bergh, 1889		<i>Caloria</i> sp. (<i>Caloria</i> sp. 1 in Gosliner et al. [24]: 362)		X	
		<i>Cratena</i> sp. (<i>Cratena</i> sp. 5 in Gosliner et al. [24]: 383)		X	
		Crsp17Ba-1	<i>Cratena</i> sp.	X	
			<i>Facelina rhodopos</i> Yonow, 2000		X
		Fasp3_16Bu-1+3-5	<i>Facelina</i> sp. (<i>Facelina</i> sp. 3 in Gosliner et al. [24]: 359)		X
	Fasp3_16Bu-2	<i>Facelina</i> sp. (<i>Facelina</i> sp. 4 in Gosliner et al. [24]: 359)		X	
	Fasp8_16Bu-1	<i>Facelina</i> sp. (<i>Facelina</i> sp. 8 in Gosliner et al. [24]: 360)		X	
		<i>Favorinus japonicus</i> Baba, 1949	X	X	
		<i>Favorinus mirabilis</i> Baba, 1955		X	

	<i>Favorinus tsuruganus</i> Baba and Abe, 1964	X				
Fasp1-18Ba-1-2	<i>Favorinus</i> sp. 1 (in Gosliner et al. [24]: 363)	X				
Mojo18Ba-1	<i>Moridilla</i> sp. a	X				
	<i>Noumeaella</i> sp. 2 (in Gosliner et al. [24]: 367)	X				
	<i>Noumeaella</i> sp. 3 (in Gosliner et al. [24]: 367)	X	X			
Nosp6_17Bu-1-2	<i>Noumeaella</i> sp. (<i>Noumeaella</i> cf. sp. 6 in Gosliner et al. [24]: 368)			X		
Nosp12_16Bu-1	<i>Noumeaella</i> sp. (<i>Noumeaella</i> sp. 12 in Gosliner et al. [24]: 367)			X		
	<i>Noumeaella</i> sp. 13 (in Gosliner et al. [24]: 369)	X				
Nosp2_15Bu 1 Nosp2_16Bu-1-6	<i>Noumeaella</i> sp.			X		
	<i>Phyllodesmium briareum</i> (Bergh, 1896)	X	X	X		X
	<i>Phyllodesmium</i> cf. <i>crypticum</i> Rudman, 1981	X				
	<i>Phyllodesmium jakobsenae</i> Burghardt and Wägele, 2004				X	
	<i>Phyllodesmium lizardense</i>	X				

	Burghardt, Schrödl and Wägele, 2008							
	<i>Phyllodesmium longicirrum</i> (Bergh, 1905)	X						
	<i>Phyllodesmium magnum</i> Rudman, 1991	X				X		
	<i>Phyllodesmium parangatum</i> Ortiz and Gosliner, 2003	X						
	<i>Phyllodesmium pecten</i> Rudman, 1981	X						
	<i>Phyllodesmium poindimiei</i> (Risbec, 1928)	X	X					
	<i>Phyllodesmium rudmani</i> Burghardt and Gosliner, 2006			X as sp 1			X as sp. 11	
	<i>Phyllodesmium undulatum</i> Moore and Gosliner, 2014							X
	<i>Phyllodesmium</i> sp. 9 (Sea Slug Forum)					X		
	<i>Pteraeolidia semperi</i> (Bergh, 1870) as <i>P. ianthina</i> (Burghardt and Sea Slug Forum)	X	X	X			X	X
Fasp17Bu-1	<i>Facelinidae</i> sp.		X					
Fasp17Bu-1	<i>Facelinidae</i> sp.		X					
Eupulmonata								
Onchidiidae	<i>Peronia</i> sp.	X						
Total number		149	172	75	23	27	85	25

5. Conclusions

When comparing our study in BA with other available studies from Indonesia, e.g., Ambon, Bali, and Papua New Guinea [2,16,38,48], we can consider the number of 149 species as reasonable for this region after only a few collection events. Only the study in Papua New Guinea exhibits a much higher number of species (Figure 28), probably due to the exceptionally high collection effort in this area [48]. We also cannot exclude the influence of seasons on the success of collection. Larkin et al. [49] showed a considerable difference in the abundance of marine Heterobranchia seen during the day and night, a factor that we took into consideration to a certain extent. However, we collected only during the late dry season, another factor that may influence overall diversity numbers [49,50].

The comparison between the various regions in North Sulawesi (Figure 27) and other Indonesian regions (Figure 28) allows the conclusion that the true number of species is actually certainly much higher than those recorded in these studies, not only in Bangka Archipelago but also in the other Indonesian areas sampled to date. A continuous increase in species records in the Indo-Pacific can already be seen when comparing the identification book of Gosliner et al. published in 2015 [24] with his follow-up study published in 2018 [51], with an increase of more than 100 species, from less than 2000 documented ones in the Indo-Pacific to more than 2100. Although we have mainly used the version from 2015, we subsequently checked our findings with the latest version, which did not reveal new information or species relevant for our study. Thus, our results contribute to the overall species numbers for the Indo-Pacific, and they form a baseline for future monitoring in the region, which still appears to be unaffected by environmental stress factors. According to the planktonic community index based on recent planktonic studies, the marine environment of BA is considered to be in a good condition [52]. However, Ponti et al. [19] consider the health of the corals in the area as critical, despite the low impact of disease, and refer to the general problems in BA of increased human activities, including mining, which irredeemably compromises reef health [19]. Snorkeling in front of the destroyed area at the mining site did not result in any records of nudibranchs, due to the recent stone blocks dumped in front of the site, where only a few corals and sponges were observed to colonize them. Probably due to land erosion and the lack of mangroves to filter nutrients and sediments, the water was extremely murky, and the high concentration of tiny jellyfish might be an additional indicator of eutrophication [50,53]. However, the nearby investigated areas, such as Sipi, revealed a pristine environment, and 13 heterobranch species were recorded from this locality (Table 2) when diving for approximately one hour with five divers involved. Since the mining project stopped recently, the highly disturbed habitat in front of the mines will provide a good study area for future studies of recolonization and recovery, and the results of this study with detailed information from each locality are a good baseline for comparison.

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