



Prevalence of parasitic isopods (Cymothoidae and Gnathiidae) on Actinopterygii fishes from the Alexandria coast off the Mediterranean Sea, Egypt

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ABSTRACT

During the year 2021, a total of 2152 Actinopterygii fish represented by eleven fish species were collected from the Egyptian waters of Alexandria. *Livoneca redmanii* Leach 1818, *Anilocra alloceraea* Koelbel 1878, *Ceratothoa oestroides* Rosso, 1827) (family: Cymothoidae), and *Gnathia* sp. (praniza larvae) (family: Gnathiidae) were discovered throughout the research period. The current study aims to provide insight on the distribution of parasitic isopods among infested fish on the Egyptian coast, through associations between parasitic species and their hosts. Two of the four species (*C. oestroides* and *A. alloceraea*) were reported for the first time on the Egyptian coast. Among the six host species, *Gnathia* sp. (praniza larva) has consistently been proven to be the most frequent species. Gills of four host species have been implicated in the discovery of *Livoneca redmanii*. The buccal cavity of *Boops boops* (Linnaeus, 1758) has been gathered to have *Ceratothoa oestroides*. whereas *Anilocra alloceraea* were reported on the outer body of *Sardina pilchardus* (Walbaum, 1792). *L. redmanii* and *Gnathia* sp. which parasitized of *Scomberomorus commerson* (Lacepède, 1800) and *Pomatomus saltatrix* (Linnaeus, 1766) had the greatest prevalence rates across their host species ($P=27.1\%$ & $P=17.7\%$), respectively. The bulk of the host species infection sites is found on the gill macroniche of the hosts, which account for 580 parasitic specimens out of a total of 978 for *Gnathia* sp. *L. redmanii* followed with 162 parasitic individuals out of a total of 174 individuals. Our investigation concluded that the parasites select a particular host to access it secretly, in a situation known as the host's specificity. New hosts that included *L. redmanii* on *P. saltatrix*, *S. commerson*, and *Umbrina cirrosa* (Linnaeus, 1758) have been reported.

INTRODUCTION

Recent years have seen an increase in interest in the parasitic isopod family Cymothoidae Leach, 1814 due to its ecological and commercial importance (Aneesh, et al., 2019, 2021; Hadfield & Smit, 2020). Members of cymothoid species are protandric hermaphrodites that feed on the blood, flesh, and mucous of several species of freshwater, marine, and estuary fishes, which are easily recognizable, although genera and species are frequently lost and incorrectly categorized (Aneesh et al., 2018; Hadfield & Smit, 2020). However, as it is challenging to validate or refute previous species reports, a large portion of

the present data on the biodiversity, distribution, and host records of this family need further investigation about its authenticity (Smit *et al.*, 2014). Despite recent studies (Hadfield & Smit, 2020), many cymothoid species still need to be corrected to provide accurate information for future studies on this key group of fish parasites from ecological and commercial perspectives.

The Egyptian parasitic isopod fauna near the Mediterranean is relatively poorly known, and thus far, investigations around this neglected group are still scarce on marine fishes from the Egyptian coasts. In total, 15 species from the family Cymothoidae have been reported from Morocco, 16 species from Algeria, 11 cymothoid species from Tunisia, and seven from Egypt (see Geba, *et al.*, 2019). Because cymothoid fauna are common in most marine environments, some cymothoid species, such as *Ceratothoa*, have been linked to significant outbreaks in endemic areas, e.g., the Adriatic Sea and the Aegean Sea. However, it appears that the cymothoids that reside on the fish's surface have fewer particular environmental needs and may thus be found in various positions. Manifestly, such a presumption certainly explains the paucity of information on the location of this group of parasites; most authors explain this simple estimate by positioning the parasites for major subdivisions of the body, e.g., head, trunk, or similar, as reported by Rameshkumar *et al.* (2016). On the subject of the nature of isopod parasite infestation in fish, few related papers are accessible in the following references (Williams & Williams 1994 and Cuyas *et al.*, 2004). However, marine fishes require more in-depth investigations on the distribution of parasitic isopods (Rameshkumar, *et al.*, 2013, 2016). From this vantage point, we were able to pinpoint the presence and distribution of a few parasitic isopod species among the commercially significant fish of the country. Hence, an attempt was made to provide new data on the distribution or prevalence of parasitic isopod species in Actinopterygii fish, which are of great economic importance, as well as to provide data on the host associations of these parasitic species in the Mediterranean of the Egyptian coast.

MATERIAL AND METHODS

The present study is conducted from January to December 2021, 2152, fish specimens belonging to eleven species of Actinopterygii fish were gathered and examined for parasitic isopods. Specimens were collected seasonally along the Alexandria coast of Egypt (Fig. 1). The living parasitic isopods were recovered from the host species in the field, fixed using the techniques described by Aneesh *et al.*, (2018), thereafter, preserved in 70% ethanol, and taxonomically identified to species-level using the proper taxonomic key. Fish specimens were delivered right away to the Invertebrate Lab (MiTA in Lab), Faculty of Science, Al-Azhar University for parasitological examination. In Lab, the total and standard length with weight were measured, the fish sex was determined for each fish specimen. Sources of fish taxonomy and host nomenclature were carried out according to Fish-Base (Froese & Pauly 2021) and the catalog of fish related to (Fricke *et al.*, 2021). The date, fish species, size, parasite micro-niche, and sample location were all documented. Parasitic isopod specimens were examined under a dissecting stereomicroscope (OPTIKA-SFX-33) and light microscopy (06AAGPV4541F1ZO) after being cleaned in lactic acid for two hours. The identification

and characterization of parasitic isopod species were performed by using modern classification keys based on morphological features and explained according to the following references (Hadfield *et al.*, 2013; Aneesh 2014). The host-parasite relationships were analyzed for the sex and size of the hosts. Infestation rates were evaluated using prevalence and mean intensity as defined by Margolis *et al.*, (1982) and its modification by Bush *et al.*, (1997).

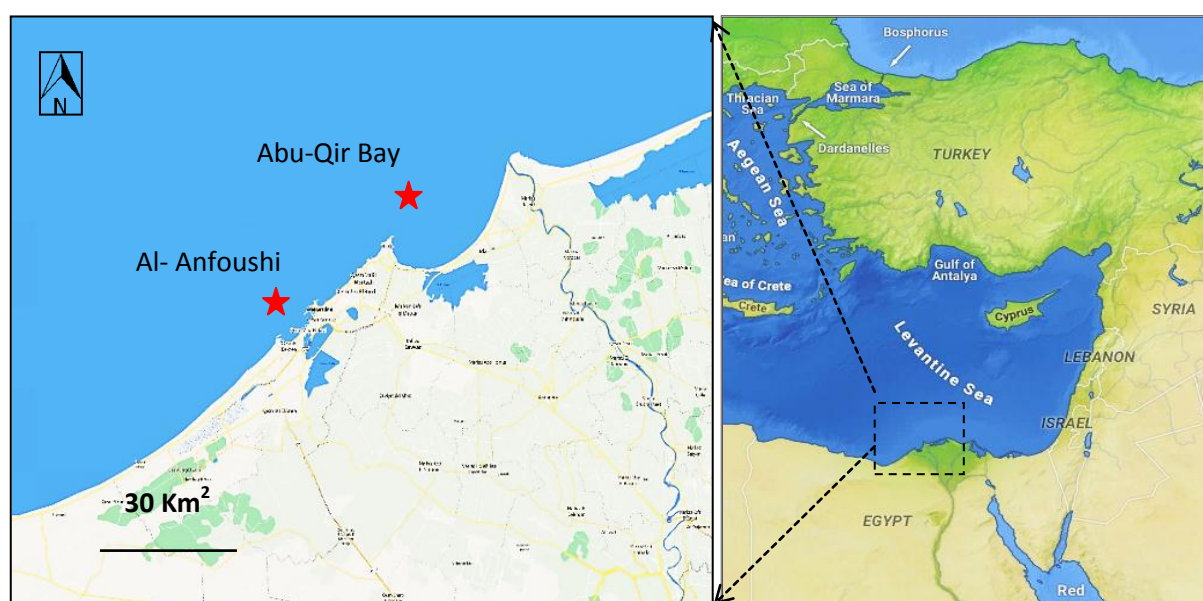


Fig. 1. Sampling sites, Abu-Qir Bay and Al-Anfoushi region indicated by red stars.

RESULTS

Throughout the research period, 19 fish species from the Egyptian coast close to Alexandria were gathered and subjected to parasitological analysis. Four parasitic isopod species were discovered from eleven host species. The remaining eight fish species in Table (1) were not infested throughout the collection year. In total, 2152 specimens of Actinopterygii fish were recovered and classified into eleven different fish species across five orders and eight families (Table 1). Four parasitic species representing three genera (*Livoneca*, *Anilocra*, and *Ceratothoa*) of the family Cymothoidae and one genus (*Gnathia*) of the family Gnathiidae that attacked species of Actinopterygii fish were found (Table 2, and Fig. 2).

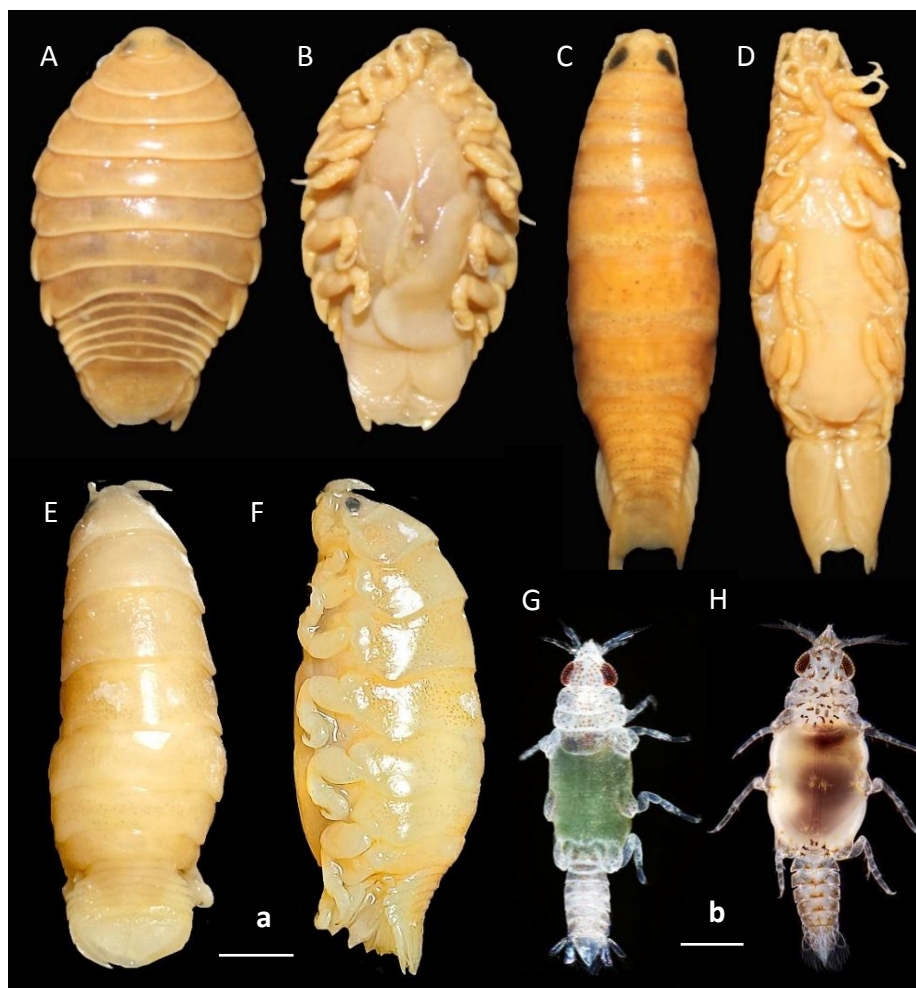


Fig. 2. Parasitic isopod species: (A & B) *Livoneca redmanii*; (C & D) *Anilocra alloceraea*; (E & F) *Ceratothoa oestroides*; and (G & H) *Gnathia* sp. (praniza larva). Scale bar, a= 3.0mm (A – F) and b= 0.5mm.

1. Annual prevalence of investigated isopod species

Throughout the research period, *Livoneca redmanii* Leach, 1818; *Anilocra alloceraea* (Koelbel, 1878); *Ceratothoa oestroides* (Risso, 1827), and *Gnathia* sp. (praniza larva) were discovered from 346 distinct host specimens (Tab. 3, and Fig. 2). *Sardina pilchardus*, the host species, had occasional penetrations of *A. alloceraea* on its body surface; the *Boops boops* buccal cavity yielded the discovery of *C. oestroides*, whereas various host species, including *Dicentrarchus labrax*, *Mugil cephalus*, *Pomatomus saltatrix*, *Scomberornorus commerson*, and *Umbrina cirrosa*, were discovered to accommodate *L. redmanii* parasitic species; additionally, *Dicentrarchus labrax*, *Mugil cephalus*, *Sparus aurata*, *Mullus surmuletus*, *Lithognathus mormyrus*, and *Pagellus erythrinus* were found to have parasitic *Gnathia* sp. (Figs., 4, 5, and 6). The annual distribution of parasitic isopods was determined according to the seasons. A total of 346 host specimens were investigated as a result of our research, which constituted 54.8% of host females and 45.2% of host males. These host species were found to carry 1295 isopod parasites, corresponding to an overall prevalence of 16.1%. Female and male host species alike had the same infestation rate ($P=16.1\%$)

($P=16.0\%$). Seasonal prevalence exhibited noticeable year-round changes. The summer season saw the highest infestation rate ($P=19.6\%$), while the autumn saw the lowest ($P=12.8\%$). The mean intensity varied from 3.4 at its lowest point in the summer to 4.4 at its highest point in the winter. Regarding the sex of the host, male predominance was highest in the summer ($P=20.2\%$) and lowest ($P=13.3\%$) in the spring. In female hosts, the frequency peaked in the winter at 20.5% and peaked in the fall at 12.3%. For male hosts during spring and for female hosts during winter, greater intensities (4.9 and 3.8) were noted (Table 2). Some clarifications on the distribution and attachment mode of the cymothoid and gnathiid species were addressed in relation to host species (Table 3) as follows.

2. Parasitological indices of parasitic isopod

A total of 1295 individuals of parasitic isopod species were collected throughout this survey. Of these, 174 *Livoneca redmanii* individuals were gathered from five distinct host species: *Dicentrarchus labrax*, *Mugil cephalus*, *Pomatomus saltatrix*, *Scomberomorus commerson*, and *Umbrina cirrosa*. This isopod exhibits a higher prevalence on host species, *S. commerson* ($P=27.1\%$), compared with its distribution on other host species, *M. cephalus* ($P=3.0\%$), and *P. saltatrix* ($P=8.8\%$) (Table 3). *Livoneca redmanii* are often discovered on host fish gills and cause negative effects on their hosts. A few individuals of *Livoneca redmanii* were observed attached to the host's trunk. They were discovered to be fixed to the host by hook-like projections on the first three pereopods, first maxillae, and mandibles. The body was always pointed toward the fish's front end when it was in the connected position. In other instances, numerous *L. redmanii* individuals were discovered from the gills on the same host, e.g., *U. cirrosa* (Fig. 4C), and these infections resulted in exceedingly serious side effects because of the known frequency or quantity of infections. However, *L. redmanii* has fewer microniches on their hosts, normally seen attached on/between the gill filaments (162 individuals), and the majority of them were obtained from the fish host of *S. commerson* (83 individuals) or the outer body surface (12 individuals) of their hosts.

Sardina pilchardus, the host species, revealed the presence of 139 individuals of *Anilocra alloceraea* on its outer body surface. According to parasitic distribution, the prevalence of this parasite was rather high ($P=23.0\%$), constituting 24.1% of the female host population and 21.6% of the male host population. Infections on the gill filaments themselves, on the outer or inner of the operculum, or anywhere else along the circumference of the gill zone were considered to fall under the definition of "gills", as shown in Fig. 5C. The presence of *A. alloceraea* within the gill filaments may have been entirely accidental, as no internal infection was observed on the gill filaments in the presence of the parasites, with the exception of one specimen that was too fragile to readily identify infections with it. However, the majority of *A. alloceraea* individuals were found on its ventral surfaces, particularly between the pectoral fins under the operculum (76 parasitic individuals) and were distributed close to the gill zonation (50 individuals). In certain instances, there were two or more individuals on the same host, as shown in Fig 5A.

Table. 1. A list of taxonomy for Actinopterygii fish species with reference to their numbers that were gathered throughout the research period.

Order	Family	Species	No	Inf	Un-Inf
Perciformes	Moronidae	<i>Dicentrarchus labrax</i> (Linnaeus, 1758)	234	+	
		<i>Sparus aurata</i> (Linnaeus, 1758)	211	+	
	Sparidae	<i>Boops boops</i> (Linnaeus, 1758)	119	+	
		<i>Lithognathus mormyrus</i> (Linnaeus, 1758)	84	+	
		<i>Pagrus pagrus</i> (Linnaeus, 1758)	96		+
		<i>Diplodus sargus</i> Linnaeus, 1758	126		+
	Sphyraenidae	<i>Sphyraena viridensis</i> Cuvier, 1829	36		+
	Serranidae	<i>Epinephelus alexandrines</i> (Valenciennes, 1828)	16		+
Siganidae	<i>Siganus rivulatus</i> Forsskål & Niebuhr, 1775	62		+	
Clupeiformes	Clupeidae	<i>Sardina pilchardus</i> (Walbaum, 1792)	357	+	
		<i>Mullus surmuletus</i> (Linnaeus, 1758)	168	+	
	Mullidae	<i>Pagellus erythrinus</i> (Linnaeus, 1758)	164	+	
		<i>Pomatomus saltatrix</i> (Linnaeus, 1766)	160	+	
Acanthuriformes	Sciaenidae	<i>Umbrina cirrose</i> (Linnaeus, 1758)	145	+	
		<i>Argyrosomus regius</i> Asso, 1801	48		+
Mugiliformes	Mugilidae	<i>Mugil cephalus</i> Linnaeus, 1758	231	+	
Scombriformes	Scombridae	<i>Scomberomorus commerson</i> (Lacepède, 1800)	210	+	
Aulopiformes	Synodontidae	<i>Saurida undosquamis</i> (J. Richardson, 1848)	98		+
Beloniformes	Hemiramphidae	<i>Hemiramphus far</i> (Forsskål, 1775)	123		+

Furthermore, only three of the 119 fish specimens of *Boops boops* that were investigated had four *Ceratothoa oestroides* individuals, which had a significantly low distribution ($P=1.3\%$). Since *C. oestroides* only lived in the host fish's buccal cavity, their location is quite particular. During the study period, only four individuals of *C. oestroides* were recorded throughout the year (Tables 3, and 4). Three of the individuals were captured in a posture facing the mouth opening, and a fourth specimen protruded from the mouth.

Multiple host species, including *D. labrax*, *Sparus aurata*, *Mullus surmuletus*, *Lithognathus mormyrus*, *Pagellus erythrinus*, *M. cephalus*, and *Gnathia* sp. (praniza larva), were also discovered. *P. erythrinus* had the highest prevalence of this isopod parasite ($P=17.7\%$), whereas *D. labrax* had the lowest prevalence ($P=8.1\%$) (Table 3). The top-ranking and most prevalent parasite was *Gnathia* sp. (praniza larva), which took up residence in a number of microniches on its hosts (580 individuals on gills, 304 individuals in the buccal cavity and mouth); the remainder were gathered from various areas of the body's surface, particularly the

frontal region, which includes the head, the upper and lower pectoral fins, the region between the pectoral and pelvic fins, and the area under the dorsal fin. The majority of these parasite individuals were discovered on the gills of the fish hosts *S. aurata* (132 individuals), *P. erythrinus* (128 individuals), and *M. cephalus* (133 individuals) (Table 4).

Table 2. Seasonal variation in the prevalence of parasitic isopods according to the fish sex.

Season	Total			Female fish			Male fish		
	NEF	NIF	NCP	NEF	NIF	NCP	NEF	NIF	NCP
		P (%)	MI		P (%)	MI		P (%)	MI
Winter	410	77 (18.8)	337 (4.4)	219	45 (20.5)	220 (4.9)	191	32 (16.8)	117 (3.7)
Spring	589	77 (13.1)	297 (3.9)	325	42 (12.9)	165 (3.9)	264	35 (13.3)	132 (3.8)
Summer	654	128 (19.6)	438 (3.4)	367	70 (19.1)	232 (3.3)	287	58 (20.2)	206 (3.6)
Autumn	499	64 (12.8)	223 (3.5)	268	33 (12.3)	112 (3.4)	231	31 (13.4)	111 (3.6)
Total	2152	346 (16.1)	1295 (3.7)	1179	190 (16.1)	729 (3.8)	973	156 (16.0)	566 (3.6)

NEF = No. of Examined fish; NIF = No. of Infested fish; NCP = No. of collected parasites; MI = Mean Intensity and P (%) = Prevalence

The prevalence of isopod parasites in relation to the sex of hosts was calculated according to the different fish hosts. In male fishes, the highest prevalence (23.5%) was observed for *L. redmanii* on *S. commerson* and the lowest (2.9%) for the same parasite species on the host *Mugil cephalus*. In female fishes, the prevalence was maximum (30.4%) for *L. redmanii* on *S. commerson* and the minimum (1.4) for *L. redmanii* on all its fish hosts (Table 3). The mean intensity of isopod species was low and did not exceed 1.7 for *L. redmanii*, *Anilocra alloceraea* and *Ceratothoa oestroides* on most fish hosts. However, for *Gnathia* sp., the mean intensity was markedly high, ranging between 4.2 and 9.0 on the fish hosts *D. labrax* and *M. cephalus*, respectively (Tab. 3).

According to field observations of the researched hosts (Figs. 4, 5, and 6), the principal clinical symptoms of infested fish species were sluggish movement, swimming near the water's surface, debilitation with abundant mucus and becoming food intolerant, as well as rubbing against hard objects. Additionally, fish that were infected with parasitic isopods displayed increased opercular movement and gathered around air sources. Some fish gathered on the water's surface, while others gathered along the beach, particularly in the intertidal zone. Below is a list of some of these indicators that were seen throughout the inquiry period, including the lack of escape reflex, emaciation, degenerative and hyperplastic alterations in the skin, and the scale of the body surface.

Table 3. Parasitological indices of parasitic isopod on their hosts in relation to the sex of hosts.

Fish hosts	Parasitic isopods	Total			Female host fish			Male host fish		
		NE F	NIF	NC P	NE F	NIF	NC P	NE F	NIF	NC P
			P (%)	MI		P (%)	MI		P (%)	MI
<i>Dicentrarchus labrax</i>	<i>Livoneca redmanii</i>	234	23 (9.8)	30 (1.3)	134	14 (10.4)	20 (1.4)	100	9 (9.0)	10 (1.1)
	<i>Gnathia sp</i>		19 (8.1)	80 (4.2)		11 (8.2)	45 (4.1)		8 (8.0)	35 (4.4)
<i>Mugil cephalus</i>	<i>Livoneca redmanii</i>	231	7 (3.0)	8 (1.1)	128	4 (3.1)	5 (1.3)	103	3 (2.9)	3 (1.0)
	<i>Gnathia sp</i>		29 (12.6)	262 (9.0)		15 (11.5)	176 (11.7)		14 (13.6)	86 (6.1)
<i>Pomatomus saltatrix</i>	<i>Livoneca redmanii</i>	160	14 (8.8)	18 (1.3)	93	10 (10.8)	14 (1.4)	67	4 (6.0)	4 (1.0)
<i>Scomberornorus commerson</i>		210	57 (27.1)	95 (1.7)	112	34 (30.4)	57 (1.7)	98	23 (23.5)	38 (1.7)
<i>Umbrina cirrosa</i>		145	17 (11.7)	23 (1.4)	78	7 (9.0)	10 (1.4)	67	10 (14.9)	13 (1.3)
<i>Sparus aurata</i>	<i>Gnathia sp</i>	211	29 (13.7)	222 (7.7)	107	15 (14.0)	124 (8.3)	104	14 (13.5)	98 (7.0)
<i>Mullus surmuletus</i>		168	15 (8.9)	73 (4.9)	92	9 (9.8)	44 (4.9)	76	6 (7.9)	29 (4.8)
<i>Lithognthus mormyrus</i>		153	22 (14.4)	134 (6.1)	84	11 (13.1)	65 (5.9)	69	11 (15.9)	69 (6.3)
<i>Pagellus erythrinus</i>		164	29 (17.7)	207 (7.1)	85	12 (14.1)	90 (7.5)	79	17 (21.5)	117 (6.9)
<i>Sardina pilchardus</i>	<i>Anilocra alloceraea</i>	357	82 (23.0)	139 (1.7)	195	47 (24.1)	78 (1.7)	162	35 (21.6)	61 (1.7)
<i>Boops boops</i>	<i>Ceratothoa oestriodes</i>	119	3 (2.5)	4 (1.3)	71	1 (1.4)	1 (1.4)	48	2 (4.2)	3 (1.5)
Total		215 2	346 (16.1)	1295 (3.7)	1179	190 (16.1)	729 (3.8)	973	156 (16.0)	566 (3.6)

NEF= No. of Examined fish; NIF= No. of Infested fish; NCP= No. of collected parasites; MI= Mean Intensity and P (%) = Prevalence.

Table 4. Attachment sites (micro-niche) of parasitic isopods on host species.

Parasitic sp.	Host species	Micro-niche					
		Gills	BCM	D. side	V. side	Tail	Head
<i>Anilocra alloceraea</i>	<i>S. pilchardus</i>	50	-	5	76	6	2
<i>Ceratothoa oestriodes</i>	<i>B. boops</i>	-	4	-	-	-	-
<i>Livoneca redmanii</i>	<i>D. labrax</i>	30	-	-	-	-	-
	<i>M. cephalus</i>	8	-	-	-	-	-
	<i>P. saltatrix</i>	18	-	-	-	-	-
	<i>S. commerson</i>	83	-	7	5	-	-
	<i>U. cirrosa</i>	23	-	-	-	-	-
	Total	162	-	7	5	-	-
<i>Gnathia</i> sp. (praniza larva)	<i>D. labrax</i>	58	15	-	3	-	4
	<i>S. aurata</i>	132	75	3	1	-	11
	<i>M. surmuletus</i>	49	20	3	1	-	-
	<i>L. mormyrus</i>	80	46	2	-	-	6
	<i>P. erythrinus</i>	128	64	-	2	-	13
	<i>M. cephalus</i>	133	84	10	16	5	14
	Total	580	304	18	23	5	48
Total		792	308	20	104	11	50

BCM=Buccal cavity and Mouth. D=Dorsal side. V=Ventral side.

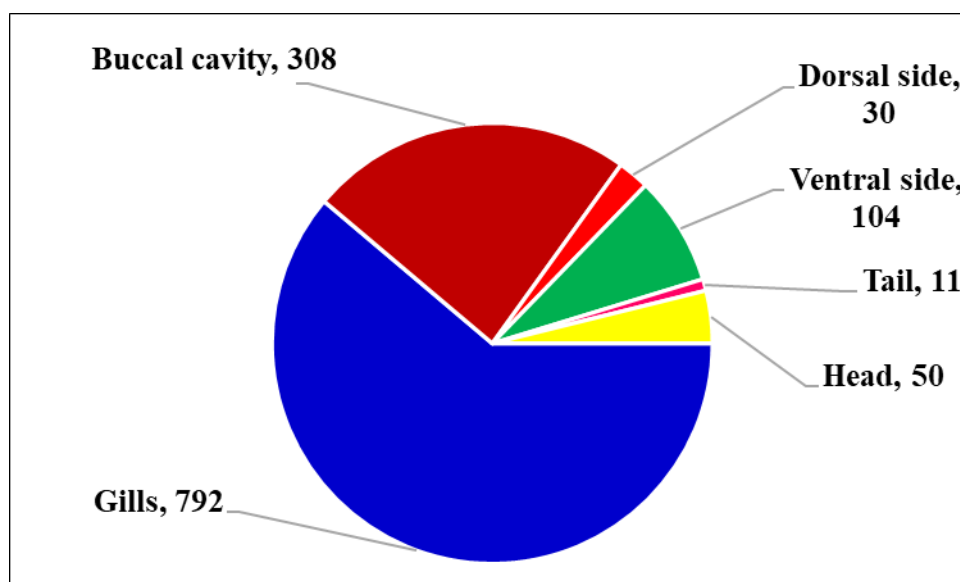


Fig. 3. Attachment sites (micro-niches) of parasitic isopods on Actinopterygii fish.

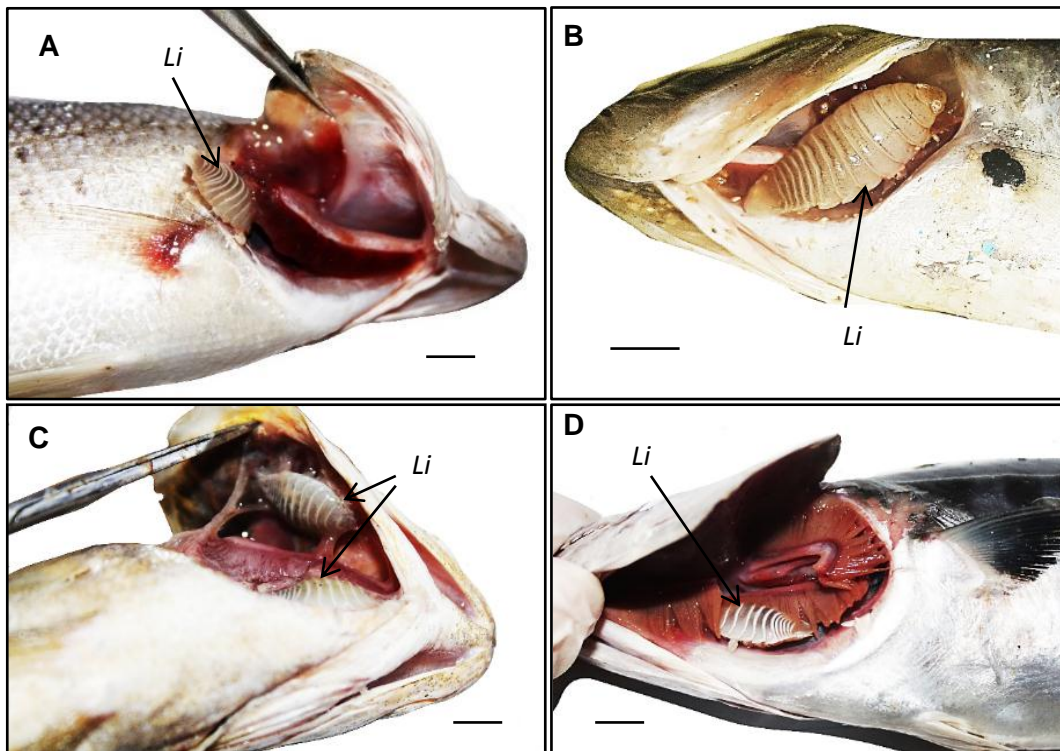


Fig. 4. *Livoneca redmanii* on their host gills. A) *D. labrax*, B) *M. cephalus*, C) *U. cirrosa*, and D) *S. commerson*. Scale bar for host species= 1cm.

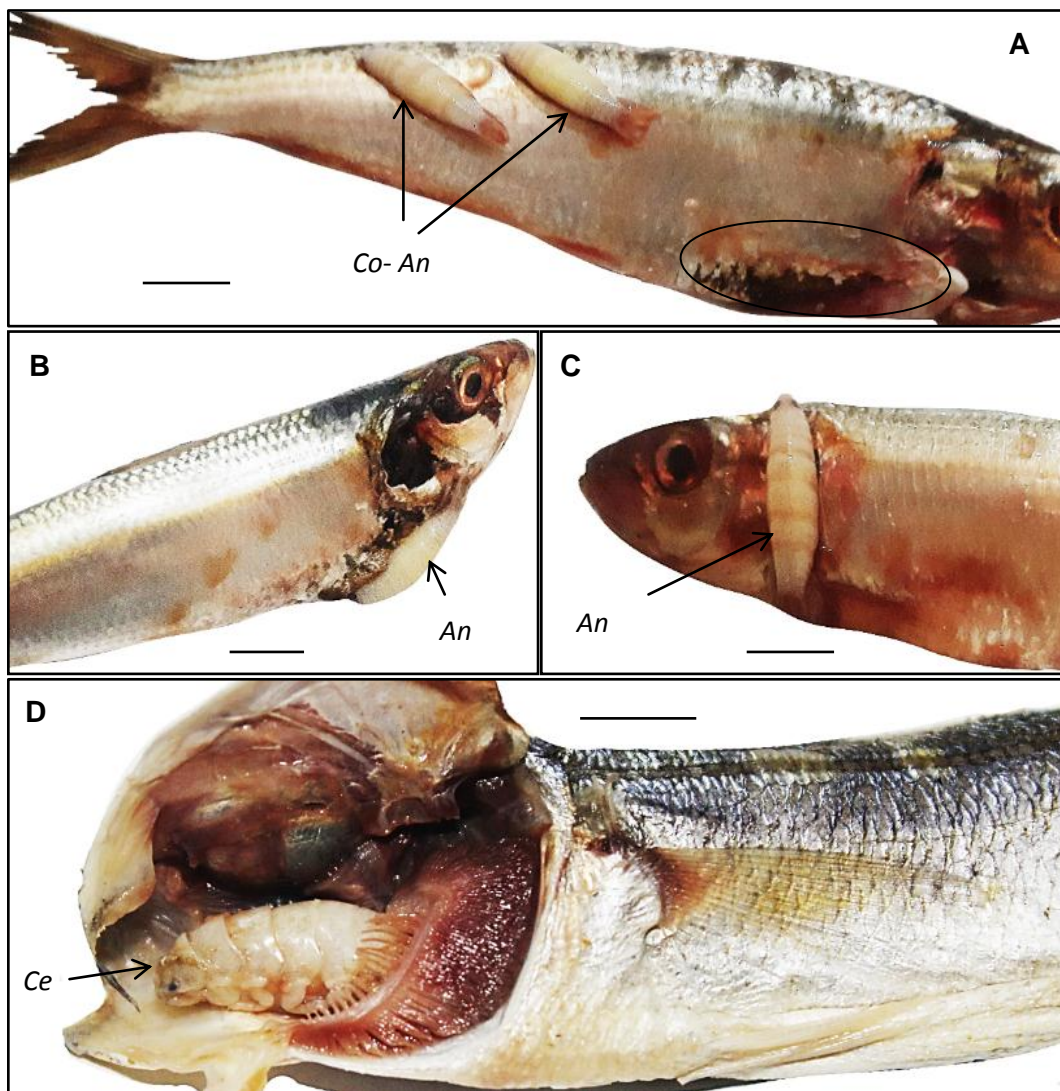


Fig. 5. *Anilocra alloceraea* in several macro-niche of *S. pilchardus* (A – C); and *Ceratothoa oestroides* on the host buccal cavity of *B. boops*. Scale bar for host species= 1cm.

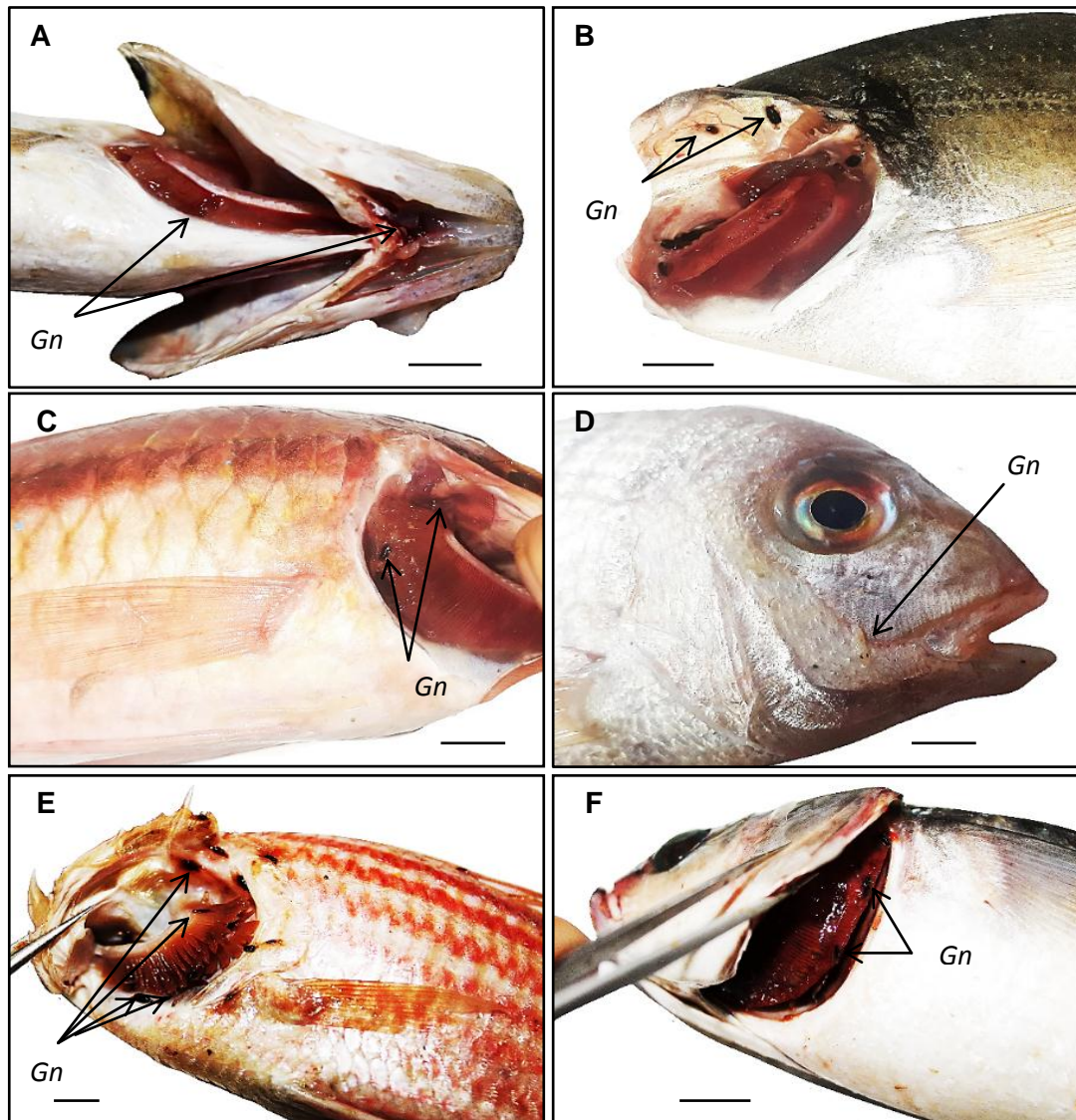


Fig. 6. *Gnathia* sp. (praniza larva) on their host species. A) *D. labrax*, B) *S. aurata*, C) *M. surmuletus*, D) *L. mormyrus*, E) *P. erythrinus* and F) *M. cephalus*. Scale bar for host species=2cm

DISCUSSION

Numerous fish species are parasitized by cymothoid isopods in a variety of habitats, which causes significant economic losses (Ravichandran *et al.*, 2011; Khalaf-Allah & Yousef, 2019). Eight out of the 19 fish species we examined in our survey were not parasitized by any isopods, while four different isopods parasitized eleven of the host species. The current study documents the presence of four species of parasitic isopods that feed on commercially significant marine fishes off the coast of Alexandria, Egypt; three of these species are in the family Cymothoidae, and one is in Gnathiidae. *Anilocra alloceraea* and *Ceratothoa oestroides* were recorded for the first time, and each parasitized just one host species; *Livoneca redmanii* and *Gnathia* sp. (praniza larva) were discovered on several distinct host species.

Livoneca redmanii has been misidentified several times by researchers in Egypt after being found in Lake Qarun, the Suez Canal, and the Mediterranean Sea (see **Geba, et al., 2019**). Five distinct fish species were parasitized by *L. redmanii* in the current investigation, and the prevalence rates for the examined hosts varied from 3.0% for the fish host *Mugil cephalus* to 27.10% for the fish host *Scomberomorus commerson*. The analysis of diversity indices revealed that the researched fish species had a high diversity and a moderate prevalence rate. Additionally, these cymothoid species favor gill location above other microniches. These results are commensurate with the findings given by **Samn et al., (2014)** for some Sparidae fish species infected by *Nerocila bivittata* (Risso, 1816) in the Mediterranean, **Khalaf-Allah & Yousef (2019)** for *L. redmanii* on *Solea solea* (Linnaeus, 1758) from Lake Qarun and **Hellal & Yousef (2018)** for *L. redmanii* on *Mugil cephalus* Linnaeus, 1758 from the same lake. Additionally, **Mahmoud et al., (2019)** examined certain Mugilidae fish from the Mediterranean that have *L. redmanii* infection. Different parasite species exhibit host and environment selection (microniches) to varying degrees. The morphological and physiological factors that affect an isopod's choice of location are still unknown for the vast majority of species (**Kabata, 1981; Youssef et al., 2018**). The results of the current investigation indicated that the majority of parasites were attached to the fish's gill, seldom to the fish's body surface.

Sardina pilchardus was infected with *Anilocra alloceraea* at a high prevalence rate ($P=23.0\%$), with the ventral body surface location of the host being the primary microniches for infection (54.7% of all individuals). **Bariche & Trilles (2006)**, *S. pilchardus*, which was obtained from an offshore area of Lebanon in the Mediterranean Sea, was heavily parasitized by *A. pilchardi*. According to **Bariche & Trilles (2006)**, **Welicky et al., (2017)**, and **Fujita et al., (2021)**, the genus *Anilocra* is more frequently found on the body surface than on the fins or in the mouth, which was also noted in the current study. The most prevalent species of tropical fish on the coast of Alexandria is *S. pilchardus*, which is also a strong swimmer and fast-growing epipelagic. *A. alloceraea* abundance among its constituents was probably due to the species' extensive occurrence in varied environments. Therefore, host specificity and the geographic location of the host may both have a role in the high frequency or abundance of *A. alloceraea* on *S. pilchardus*. As a result, our study supports the conclusions of **Welicky et al., (2017)**, **Hure & Mustas (2020)**, and **Fujita et al., (2021)**. In the buccal cavity of the host fish *Boops boops*, four individuals of *Ceratothoa oestroides* were discovered ($P=2.5\%$), suggesting that it may merely be a less frequent species in this host at that site. Considering that there are only 25 recognized *Ceratothoa* species and that they are only present in eight of the world's twelve nautical biogeographical zones, as reported by **Hadfield & Smit (2020)**, this behavior and rarity of *Ceratothoa* species may be reflected in the declining prevalence rates among their examined host species.

Gnathia sp. (praniza larvae) are a common ectoparasite of elasmobranchs and teleost fishes (**Genc et al., 2005**, **Gonzalez & Moreno 2005**; **Nagel & Grutter 2007**). They are blood-feeding isopod larvae found on various sites of their hosts, causing severe lesions (**Davies 1981**). In addition, these parasites have low host specificity and worldwide

distribution in different habitats, including the Mediterranean Sea (**Papoutsoglou 1975; Ayari 2004; Koukouras 2010**). Previously, they have been reported in many areas around the world (see **Tanaka 2004; Shimomura & Tanaka 2008; Hadfield & Smit 2008**). In the present study, *Gnathia* sp. (praniza larva) was reported from six fish species and showed the highest parasite abundance among the investigated hosts. This parasite prefers members of Sparidae as hosts and has a higher prevalence on *Pagellus erythrinus* ($P=17.7\%$) and *Lithognathus mormyrus* ($P=14.4\%$). Our investigation using a few indices revealed that the analyzed hosts had a high frequency and quantity of *Gnathia* sp. In fact, this analysis concurs with that made by **Grutter (1994); Chambers & Sikkell (2002)**. Although some articles have proven the opposite, **Honma *et al.*, (1991) and Martens & Moens (1995)** revealed that *Gnathia* sp. seldom injures the hosts they feed on (**Paperna *et al.*, 1984; Honma & Chiba 1991**). It is probable that what the first stated is compatible with our findings. *Gnathia* sp. is widely distributed over many hosts, and the severity of the varied sicknesses in the various regions of the host's body produces significant harm among the afflicted fish.

The host species of the fish that are most frequently sought after are abundant and widespread around the coast of Alexandria, although there are also fewer common species. The most common species in our study were *Dicentrarchus labrax* and *Mugil cephalus*, allowing us to analyze 234 and 231 specimens of each in only one year. These two species have the highest richness among the species assessed along the Alexandrian coast, with two parasitoids appearing. These findings agree with those made public by **Youssef *et al.*, (2018) and Raibaut *et al.*, (1998)**. Therefore, parasite prevalence rates and main intensity among the individuals of these host species may be influenced by population richness. The current study sought to provide light on the distribution of parasitic isopods among infested fish in Egyptian habitats and concluded that some parasitic isopod species may be predicted to be present on specific hosts but not others. This impression can be explained by the fact that parasites favor and are drawn to a particular host, and this coexistence or dependency between parasite animals and their hosts might result from environmental changes. In light of an awareness of the link between the parasite and its host, this view also clarifies a few ideas relating to biodiversity. As certain environmental variables overlap with other determinants, such as population density or geographic distribution, they may also have an impact on the frequency and severity of infections.

CONCLUSION

Aspects of several cymothoid species' distribution and prevalence in the Egyptian waters were discussed in the study. These aspects are crucial for identifying their host species, which increases their variety and geographic distribution. The gills of Actinopterygii fish captured off the Egyptian Mediterranean coast suffered serious damage as a result of the isolation of cymothoid species as obligatory parasites on their hosts. Understanding this behaviour is essential to comprehending the problem of biodiversity, particularly in areas where parasitic crustaceans are few. By performing more study to comprehend other concerns, such as those linked to the taxonomic aspects and the egregious issues experienced

by these parasitic species, this report assures that researchers are going forward in their efforts to improve the use of these parasites.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could influence the work presented in this paper.

Conflict of interest

I, as the corresponding author, declare, on behalf of all authors of the paper, that no financial conflict of interest exists concerning the work described.

Ethical approval

The specimen is not under the listed category of experimental animals, which requires ethical approval.

Sampling and field studies

All necessary permits for sampling and observational field studies were obtained by the authors from the competent authorities.

REFERENCE

- Aneesh, P.T.; Hadfield, K.A.; Smit, N.J. and Kumar, A.B.** (2021). Morphological description and molecular characterization of a new species of *Anilocra* Leach, 1818 (Crustacea: Isopoda: Cymothoidae) from India. *International Journal for Parasitology: Parasites and Wildlife* 14: 321–328.
- Aneesh, P.T.; Hadfield, K.A.; Smith, N.J. and Biju Kumar, A.** (2021). Morphological description and molecular characterization of a new species of *Anilocra* Leach, 1818 (Crustacea: Isopoda: Cymothoidae) from India. *International Journal for Parasitology: Parasites and Wildlife*, 14: 321–328.
- Aneesh, P.T.; Helna, A.K.; Kumar, A.B. and Trilles, J.P.** (2020). A taxonomic review of the branchial fish parasitic genus *Elthusa* Schioedte and Meinert, 1884 (Crustacea: isopoda: Cymothoidae) from Indian waters with the description of three new species. *Mar. Biodivers.* 50: e65.
- Aneesh, P.T.; Helna, A.K.; Sudha K. and Anilkumar, G.** (2018). *Agarna malayi* Tiwari 1952 (Crustacea: Isopoda: Cymothoidae) parasitizing the marine fish, *Tenualosa toli*

- (Clupeidae) from India: redescription/description, life cycle and pattern of parasitic occurrence. *Zoological Studies*, 57: 25.
- Aneesh, P.T.; Helna, A.K.; Trilles, J.P. and Chandra, K.** (2019). Occurrence and redescription of *Anilocra leptosoma* bleeker, 1857 (Crustacea: isopoda: Cymothoidae) parasitizing the clupeid fish *Tenualosa toli* (valenciennes) from the Arabian Sea, India. *Mar. Biodivers.* 49: 443–450.
- Aneesh, P.T.; Kottarathil, H.A. and Kuma, A.B.** (2022). Simultaneous double parasitism by the parasitic cymothoids (Crustacea: Isopoda) of two genera on a single host fish *Tenualosa toli* from India. *The journal of the Brazilian Crustacean Society*. DOI 10.1590/2358-2936e2022013.
- Aneesh, P.T.; Sudha, K.; Arshad, K.; Anilkumar, G. and Trilles, J.P.** (2013). Seasonal fluctuation of the prevalence of cymothoids representing the genus *Nerocila* (Crustacea, Isopoda), parasitizing commercially exploited marine fishes from the Malabar Coast, India. *Acta Parasitol.*, 58: 80–90.
- Aneesh, P.T.; Sudha, K.; Helna, A.K.; Anilkumar, G. and Trilles, J.P.** (2014). Multiple parasitic crustacean infestation on belonid fish *Strongylura strongylura*. In: I.S. Wehrtmann, R.T. Bauer (eds), *Proceedings of the Summer Meeting of the Crustacean Society and the Latin American Association of Carcinology*, Costa Rica, July 2013. *ZooKeys*, 457: 339–353.
- Ayari R.** (2004). *Les peuplements macrozoobenthiques du petit golfe de Tunis: structure, Organization et Etat sanitaire* [MSc Thesis]. University of Carthage, Faculte des Sciences de Bizerte.
- Bariche, M. and Trilles, J.P.** (2005). Preliminary checklist of Cymothoids (Crustacea: Isopoda), parasites on marine fishes from Lebanon. *Zoology in the Middle East*, 34: 5–12.
- Bariche, M. and Trilles, J.P.** (2006). *Anilocra pilchardi* n. sp., a new parasitic cymothoid isopod from off Lebanon (Eastern Mediterranean). *Systematic Parasitology*. 64: 203–214.
- Bowman, T.E. and Tareen, I.U.** (1983). Cymothoidae from fishes of Kuwait (Arabian Gulf) (Crustacea: Isopoda). *Smithsonian Contributions to Zoology*, 382, 1–30.
- Boxshall G.A. and Halsey S.H.** (2004). *An Introduction to Copepod Diversity*. London: The Ray Society.
- Boyko, C. B.; Bruce, N. L.; Hadfield, K.A.; Merrin, K. L.; Ota, Y.; Poore, G.C.B.; Taiti, S.; Schotte, M. and Wilson, G.D.F.** (2008). Onward. World Marine, Fresh-water, and Terrestrial Isopod Crustaceans Database. *Gnathia* Leach, 1814. Accessed through: World Register of Marine Species at. <http://www.marinespecies.org/aphia.php?p=taxdetails&id=257166> on 2020-12-10.
- Bruce, N.L.** (1990). The genera *Catoessa*, *Elthusa*, *Ichthyoxenus*, *Idusa*, *Livoneca*, and *Norileca* n. gen. (Isopoda, Cymothoidae), crustacean parasites of marine fishes, with descriptions of eastern Australian species. *Records of the Australian Museum.*, 42(3): 247-300.

- Brusca, R.C.** (1981). A monograph on the Isopoda Cymothoidae (Crustacea) of the eastern Pacific. *Zoological Journal of the Linnean Society.*, 73(2): 117 – 199.
- Bush A.O.; Lafferty K.D.; Lotz J.M. and Shostak A.W.** (1997). Parasitology meets ecology on its own terms: Margolis *et al.*, revisited. *Journal of Parasitology* 83: 575–583.
- Bush, A.O.; Fernandez, J.C.; Esch, G.W. and Seed, J.R.** (2001). Parasitism: The diversity and ecology of animal parasites. Cambridge University Press, United Kingdom. Pp: 580.
- Chambers S.D. and Sikkel P.C.** (2002). Diel emergence patterns of ecologically important, fish-parasitic, gnathiid isopod larvae on Caribbean coral reefs. *Carib J Sci.* 38: 37–43.
- Cuyas C.; Castro J.J.; Ortega, A.T. and Carbonne, E.** (2004). Insular stock identification of *Serranus atricauda* (Pisces: Serranidae) through the presence of *Ceratothoa steindachneri* (Isopoda: Cymothoidae) and *Pentacapsula cutancea* (Myoxoa: Pentacapsulidae) in the Canary Islands. *Sci Mar Bare* 68: 159–163.
- Davies A.J.** (1981). A scanning electron microscope study of the praniza larva of *Gnathia maxillaries Montagu* (Crustacea, Isopoda, Gnathiidae), with special reference to the mouthparts. *J Nat His.* 15: 545–554.
- Fischer W.; Bauchot M. L. and Schneider M.** (1987). Fiches FAO d'identification des espèces pour les besoins de la pêche. (Révision 1). Méditerranée et Mer Noire. Zone de Pêche 37. Rome: FAO, 761–1529.
- Fricke, R.; Eschmeyer, W.N. and van der Laan, R.** (2021). Catalog of fishes: genera, species <http://research.calacademy.org/research/ichthyology/catalog/fishca>, tmain.asp accessed January 2021.
- Froese, R. and Pauly, D.** (2018). Fish-Base. <http://www.fishbase.org>.
- Froese, R. and Pauly, D.** (2021). FishBase. Version (02/2015). Worldwide Web electronic publication. Available from. <http://www.fishbase.org>. accessed January 2021.
- Geba, K. M.; Sheir, S. K.; Aguilar, R.; Ogburn, M. B.; Hines, A. H.; Khalafallah, H. J.; El-Kattan, A.; Hassab El-Nabi, S.E. and Galal-Khallaf. A.** (2019). Molecular and morphological confirmation of an invasive American isopod; *Livoneca redmanii* Leach, 1818, from the Mediterranean region to Lake Qaroun, Egypt. *Egyptian Journal of Aquatic Biology & Fisheries*, Vol. 23(4): 251 – 273.
- Genc E.; Genc M.A.; Can M.F.; Genc E., and Cengizler I.** (2005). A first documented record of gnathiid infestation on white grouper (*Epinephelus aeneus*) in Iskenderun Bay (northeastern Mediterranean), Turkey. *J App Ichthyol.* 21: 448–450.
- Gonzalez M.T. and Moreno C.A.** (2005). The distribution of the ectoparasite fauna of *Sebastes capensis* from the southern hemisphere does not correspond with zoogeographical provinces of free-living marine animals. *J Biogeogr.* 32: 1539–1547.
- Grutter A.S.** (1994). Spatial and temporal variations of the ectoparasites of seven reef fish species from Lizard Island and Heron Island, Australia. *Mar Ecol Prog Ser.* 115: 21–30.

- Hadfield, K. A., Bruce, N. L., Smit, and N. J.** (2013). Review of the fish-parasitic genus *Cymothoa* Fabricius, 1793 (Isopoda, Cymothoidae, Crustacea) from the southwestern Indian Ocean, including a new species from South Africa. *Zootaxa* 3640: 152–176.
- Hadfield, K.A. and Smit, N.J.** (2008). Description of a new gnathiid, *Afrignathia multicavea* gen. et sp. n. (Crustacea: Isopoda: Gnathiidae), from South Africa. *African Zoology*, 43 (1): 81–89.
- Hellal, A. M. and Yousef, O. E.** (2018). Infestation Study of *Livoneca redmanii* (Isopoda, Cymothoidae) on *Mugil cephalus* in Lake Qarun, Egypt. *Egypt. Acad. J. Biol. Sci.*, 10(1): 1-17.
- Hiroki Fujita, H.; Umino, T. and Saito, N.** (2021). Molecular identification of the aegathoid stage of *Anilocra clupei* (Isopoda: Cymothoidae) parasitizing sweeper *Pempheris* sp. (Perciformes: Pempheridae). *Crustacean Research*; 50: 29–31.
- Honma Y.; Tsunaki S.; Chiba A., and Ho J.** (1991). Histological studies on the juvenile gnathiid (Isopoda, Crustacea) parasitic on the branchial chamber wall of the stingray *Dasyatis akajei*, in the Sea of Japan (East Sea). *Rep Sado Mar Biol Station Niigata Un.* 21: 37–47.
- Honma, Y. and Chiba, A.** (1991). Pathological changes in the branchial chamber wall of stingrays, *Dasyatis* spp., associated with the presence of juvenile gnathiids (Isopoda, Crustacea). *Fish Pathol.* 26: 9–16.
- Horton, T.; Diamant, A. and Galil, B.S.** (2005). *Ceratothoa steindachneri* (Isopoda, Cymothoidae): an unusual record from the Mediterranean. *Crustaceana.*, 77(9): 1145—1148.
- Hure, M. and Mustać, B.** (2020). Feeding ecology of *Sardina pilchardus* considering co-occurring small pelagic fish in the eastern Adriatic Sea. *Marine Biodiversity*, 50 (40): 1– 12.
- Kabata, Z.** (1981). Copepoda (Crustacea) parasitic on fishes: problems and perspectives. *Advances in Parasitology* 19: 1–71.
- Kabata, Z.** (1984). Diseases caused by metazoans: crustaceans. In: *Diseases of Marine Animals*. Kinne, O. (ed.). Hamburg, Germany: Biologische Anstalt Helgoland; P. 321-399.
- Kamiya, T.; Dwyer, K.; Nakagawa, S. and Poulin, R.** (2014). What determines species richness of parasitic organisms? A meta-analysis across animal, plant, and fungal hosts. *Biological Reviews* 89: 123–134.
- Khalaf-Allah, H.M.M. and Yousef, O.E.** (2019). Infestation Study of *Livoneca redmanii* (Isopoda, Cymothoidae) on *Solea solea* in Lake Qarun, Egypt. *Journal of the Egyptian Society of Parasitology*, 49(1): 105- 114.
- Koukouras, A.** (2010). Checklist of marine species from Greece. Aristotle University of Thessaloniki. Assembled in the framework of the EU FP7 PESI project. [Accessed 2015 Oct]. Available from: www.marinespecies.org.
- Mahmoud, N. E.; Fahmy, M. M.; Abuowarda, M. M.; Zaki, M. M.; Ismail, E. M. and Ismael, E. S.** (2019). Mediterranean Sea fry; a source of isopod infestation problem in Egypt with reference to the effect of salinity and temperature on the survival of

- Livoneca redmanii (Isopoda: Cymothoidae) juvenile stages. J. Egypt. Soc. Parasitol., 49(1): 235-242.
- Margolis L.; Esche G.W.; Holmes J. C.; Kuris, A.M. and Schrad, G.A.** (1982). The use of ecological terms in parasitology (report of an ad hoc committee of the American Society of Parasitologists). Journal of Parasitology 68: 131–133.
- Martens, E. and Moens J.** (1995). The metazoan ecto- and endoparasites of the rabbitfish, *Siganus sutor* (Cuvier and Valenciennes, 1835) of the Kenyan Coast I. Afr J Ecol. 33: 405–416.
- Nagel, L. and Grutter A.S.** (2007). Host preference and specialization in *Gnathia* sp., a common parasitic isopod of coral reef fishes. J Fish Biol. 70: 497–508.
- Paperna I.; Diamant A. and Overstreet R.M.** (1984). Monogenean infestations and mortality in wild and cultured Red Sea fishes. Helgol Meeresunters. 37:445–462.
- Papoutsoglou S. (1975).** Study of the metazoan parasites of the fish of Saronikos Gulf [Ph. D. Thesis]. University of Patras.
- Ramdane, Z.; Bensouilah, M.A. and Trilles, J.P.** (2007). The Cymothoidae (Crustacea, Isopoda), parasites on marine fishes, from Algerian fauna. Belg. J. Zool., 137 (1): 67-74.
- Rameshkumar, G.; Ravichandran S.; Sivasubramanian K. and Trilles J.P.** (2013). New occurrence of parasitic isopods from Indian fishes. J Parasit Dis 37(1): 42–46.
- Ravi, V. and Rajkumar, M.** (2007). Effect of isopod parasite, *Cymothoa indica* on gobiid fish, *Oxyurichthys microlepis* from Parangipettai coastal waters (South–east coast of India). Journal of Environmental Biology., 28(2): 251-256.
- Ravichandran, S.; Rameshkumar, G. and Kumaravel, K.** (2009). Variation in the morphological features of Isopod fish parasites. World Journal of Fish and Marine Sciences., 1(2): 137-140.
- Ravichandran, S.; Rameshkumar, G. and Trilles, J.P.** (2011). New records of two parasitic cymothoids from Indian fishes. J. Parasit. Dis., 35(2): 232-234.
- Ravichandran, S.; Rameshkumar, G. and Trilles, J.P.** (2011). New records of two parasitic cymothoids from Indian fishes. J. Parasit. Dis., 35(2): 232-234.
- Rhode, K.** (2005). Marine parasitology. CABI, Australia. Folia Parasitological., 53: 77–78.
- Samn, A.A.; Metwally, K.M.; Zeina, A.F. and Khalaf-Allah, H.M.M.** (2014). First occurrence of *Nerocila bivittata*: parasitic Isopods (skin shedders) on *Lithognathus mormyrus* (Osteichthyes, Sparidae) from Abu Qir Bay, Alexandria, Egypt. Journal of American Science., 10(7): 171-179.
- Shimomura, M. and Tanaka, K.** (2008). A new species of *Thaumastognathia* Monod, 1926 from Japan (Isopoda, Gnathiidae). Crustaceana 81(9): 1091–1097.
- Smit N.J.; Bruce N.L. and Hadfield K.A.** (2014). Global diversity of fish parasitic isopod crustaceans of the family Cymothoidae. Int J Parasitol Parasites Wildl 3: 188–197.
- Smit, N.J. and Davies, A.J.** (2004). The curious lifestyle of the parasitic stages of gnathiid isopods. Adv. Parasitol. 58, 289–391. [https://doi.org/10.1016/S0065-308X\(04\) 58005-3](https://doi.org/10.1016/S0065-308X(04) 58005-3).

- Smit, N.J.; Bruce, N.L. and Hadfield, K.A.** (2019). Introduction to parasitic Crustacea: State of knowledge and future trends. In: Smit, N.J., Bruce, N.L., Hadfield, K.A. (Eds.), Parasitic Crustacea: State of Knowledge and Future Trends. Springer Nature Switzerland AG, Cham, pp. 1–6.
- Tanaka, K.** (2004). A new species of Gnathia (Isopoda: Cymothoidea: Gnathiidae) from Ishigaki Island, the Ryukyus, southwestern Japan. Crustacean Research, 33: 51–60.
- Tanaka, K.** (2007). Life history of gnathiid isopods – current knowledge and future directions. Plankt. Benthos Res. 2: 1–11.
- Trilles, J.P.** (1986). Les Cymothoidea (Crustacea, Isopoda, Flabellifera) D’Afrique. Bull Mus National Hist Nat 8(3): 617–636.
- Trilles, J.P.** (1994). Les Cymothoidea (Crustacea, Isopoda) du Monde. Podrome pour une faune. Studia Marina, 21/22: 1 – 288.
- Trilles, J.P. and Bariche, M.** (2006). First record of the Indo-Pacific Cymothoa indica (Crustacea, Isopoda, Cymothoidea) a Lessepsian species in the Mediterranean Sea. Acta Parasitol., 51(3): 223-230.
- Welicky, R.L.; Hadfield, K.A.; Sikkel, P.C. and Smit, N.J.** (2017). Molecular assessment of three species of *Anilocra* (Isopoda, Cymothoidea) ectoparasites from Caribbean coral reef fishes, with the description of *Anilocra brillae* sp. n. ZooKeys; 663: 21–43.
- Williams, E.H. and Williams, L.B.** (1994). Four cases of unusual crustacean-fish associations and comments on parasitic processes. J Aquat Anim Health 6: 202–208.
- Wilson, G.D.F.** (2008). Global diversity of Isopod crustaceans (Crustacea; Isopoda) in freshwater. Hydrobiologia. 595: 231–240.
- Youssef, F.; Tlig Zouari, S. and Benmansour, B.** (2018). New host–parasite records of siphonostomatoid copepods infesting elasmobranch fishes in Tunisian waters. Journal of the Marine Biological Association of the United Kingdom 1–5. <https://doi.org/10.1017/S002531541800084X>.