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The Alysiinae (Hym. Braconidae) parasites of the Agromyzidae (Diptera)

IV. The parasites of Hexomyza ENDERLEIN, Melanagromyza HENDEL, Ophiomyia Braschnikov and Napomyza Westwood²

With textfigures 148-170

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Introduction

Three parts of this paper have already appeared in this journal (GRIFFITHS, 1964, 1966a and 1966b). The present fourth part deals with the three remaining European genera of Agromyzinae, that is *Hexomyza* ENDERLEIN, *Melanagromyza* HENDEL and *Ophiomyia* BRASCHNIKOV, and with one further genus of Phytomyzinae, *Napomyza* WESTWOOD. These genera contain very few species whose larvae form parenchymal leaf-mines (as in the majority of Agromyzidae). Most species of *Melanagromyza*, *Ophiomyia* and *Napomyza* feed in the stems of their host plant, either boring within the stem (in most *Melanagromyza* and *Napomyza*) or forming shallow mines beneath the epidermis (in many *Ophiomyia* species). However a few species of these genera have different biologies (for instance a few *Ophiomyia* and *Napomyza*, are all gall-causers.

The three genera of Agromyzinae are clearly monophyletic (being synapomorph in respect of their black halteres, reduction in the number of dorsocentral bristles and the form of the larval mandibles). Napomyza is without doubt phylogenetically disjunct from this group of genera (since the Agromyzinae are a monophyletic group), but it is convenient to consider its parasites at the same time because species of the Chorebus senilis group sensu lato and the C. cybele group are associated with both Napomyza and these genera of Agromyzinae.

² Part I in Beitr. Ent., 14, 823-914; 1964. – Part II in Beitr. Ent., 16, 551-605; 1966. – Part III in Beitr. Ent., 16 775-951; 1966.

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This similarity in parasite fauna appears to have arisen through the similarity in larval biology which has facilitated the transference of parasites from one group to the other.

All four host genera treated in this paper have recently been revised by SPENCER (1964, 1966a and 1966b). I have followed his nomenclature throughout. This involves some transference of species formerly included in *Melanagromyza* to *Ophiomyia*, as well as a new generic concept for which the name *Hexomyza* is available. As thus revised these three genera appear to represent monophyletic groups. My concept of *Napomyza* in this paper has been restricted (following NOWAKOWSKI and SPENCER) to those species monophyletic with the genotype *N. lateralis* FALLÉN. The parasites of the several leaf-mining species which were previously ascribed to this genus have already been treated in Part III (GRIFFITHS, 1966b).

Acknowledgements to most of those who have helped me with material for this paper have been given in previous parts. In addition to those already mentioned I would like to thank Drs. L. E. VAN'T SANT and Mr. J. G. C. BETHE of the Instituut voor Plantenziektenkundig Onderzoek at Wageningen, Holland, and Mr. J. LOUNSKY of the Station d'Entomologie at Gembloux, Belgium, whose help in obtaining parasite material from hosts of agricultural importance has been most valuable. My thanks are also due to Mr. J. C. DEEMING, who obtained the bred specimens of *Chorebus orbiculatae* sp. nov.

The abbreviations used in this paper have already been explained in the introduction to Part II (GRIFFITHS, 1966a).

Previous Records

As in previous parts of this paper I have prepared a table below explaining the discrepancies between my list of host records and the list given in FULMEK (1962). The comments exclude changes in the generic nomenclature, which affect some of the host names and nearly all the parasite names. Comments on some of the rejected records are also included under the descriptions of the species concerned. Leaf-mining species formerly included in *Napomyza* are not mentioned in this list, but have been dealt with in Part III (GRIFFITHS, 1966b). FULMEK's list excludes previous records for *Hexomyza*, whose larvae are gallcausers, and *Melanagromyza*, whose larvae are stem-borers (the two species listed by FULMEK as *Melanagromyza*, as shown below, are now placed in *Ophiomyia*). Previous records for those genera are referred to under the descriptions of *Chorebus senilis* (NEES), *C. brevicornis* (THOMSON) and *C. gedanensis* (RATZEBURG), and also in the footnote to table 17.

Dacnusa Haliday

Dacnusa pubescens (CURTIS)

In addition to the records of this species from *Phytomyza* hosts given in Part III, I have received the following material bred from *Napomyza*.

Host 5 — Napomyza carotae SPENCER

1 ${\rm J}$ from puparium in carrot (Daucus carota sativus), Nes, Friesland, Holland, em. 11. ix. 65, leg. Bethe (GCDG).

Table 16

Earlier Records of Alysiinae parasites of host species included in this paper in *Ophiomyia* and *Napomyza* (after FULMER, 1962) with comments thereon.

Host	Parasite	Comments
Melanagromyza beckeri HENDEL	Dacnusa leptogaster HALIDAY Rhizarcha areolarís NEES	host was <i>Ophiomyia cunctata</i> HENDEL not accepted (see Part III)
Melanagromyza simplex H. Löw	Dacnusa bathyzona MARSHALL Dacnusa rondanii GIARD	refers to Chorebus rondanii (GIARD) accepted
Napomyza lateralis Fallín	Dacnusa didas NIXON Dacnusa flavipes GOUREAU	host was N. scrophulariae SPENCER. • not accepted (described from Cerodontha (Dizygomyza) iraeos ROBINEAU DESVOIDY see Appendix VIII)
	Dacnusa leptogaster HALIDAY	refers to Chorebus glaber (NIXON) bred from N. cichorii SPENCER
	Dacnusa rufipes NEES	SIMM'S (1925) "Phytomyza lateralis FAL- LÉN" was the species now accepted as <i>P. nigra</i> MEIGEN. The parasite he called <i>rufipes</i> was probably the species described in Part III as <i>Chorebus aphantus</i> (MAR SHALL). SIMM's figure is of a teneral spe- cimen whose wing venation is not develop ed.
	Rhizarcha areolaris NEES	The host was <i>Phytomyza nigra</i> MEIGEN (called <i>lateralis</i> by SIMM, 1925).
Ophiomyia proboscidea STROBL	Dacnusa tarsalis Thomson	not accepted (parasite of <i>Phytomyza autum-</i> nalis GRIFFITHS and <i>P. farfarae</i> HENDEL)
Tylomyza pinguis FALLÉN	Dacnusa leptogaster HALIDAY	accepted

Host 6 — Napomyza cichorii Spencer

1 \bigcirc from puparium in roots of chicory (*Cichorium intybus*), Isle of Walcheren, Holland, em. iii. 66, leg. BETHE (GCDG). 9 ex. from roots and leaves of chicory (*Cichorium intybus*), 1963/64 and 1964/65, from Zaventem, Belgium, leg. LOUNSKY (GCDG and Station d'Entomologie, Gembloux).

Host 7 — Napomyza lateralis FALLÉN

2 ex. from puparia in *Matricaria chamomilla*, Zaventem, Belgium, 1964/65, leg. LOUNSKY (GCDG).

The Chorebus senilis group sensu lato

The concept of the *Chorebus senilis* group sensu lato here proposed includes all species included by NIXON (1944) in his *Dacnusa leptogaster* group, *Dacnusa* senilis group s. str. and *Dacnusa gracilis* group, as well as *C. petiolatus* (NEES), which NIXON classified as a "species sola" of *Dacnusa*. I do not include in this group either *C. fallax* (NIXON) (referred to the *lateralis/ovalis* complex, see Part III) or the species included in NIXON's "cytherea subgroup" of his senilis group. The senilis group s.l., as here redefined, is perhaps part of a wider monophyletic group which also includes the merella and cytherea groups, and species formerly included in "Gyrocampa", "Paragyrocampa" and "Chorebus" in the restricted sense. My reasons for suggesting this concept have

been given in Part I (GRIFFITHS, 1964, p. 849-850). The senilis group is distinguished from the other members of this suggested group by one distinctive apomorph character - the sides of the pronotum, at least along and below the oblique suture, are covered with dense, opaque, usually matted pubescence. (In most other Chorebus spp. the pronotal pubescence is sparser, fine and inconspicuous: however a few species of the ovalis/lateralis complex associated with Liriomyza hosts, e.g. C. misellus (MARSHALL), have also, through convergence, evolved similar dense pronotal pubescence.) In addition the senilis group is apooec, i.e. characterised by an innovation in host association. That the association of this group with hosts of the Melanagromyza-Ophiomyia-Hexomyza group (these three genera being monophyletic) was the result of a transference from non-Agromyzinae hosts is apparent from the fact that the Chorebus parasites of leaf-mining species of Agromyzinae, included in the paraphyletic "genus" Agromyza, belong to the lateralis/ovalis complex, and are thus more closely related to the parasites of leaf-mining Phytomyzinae than to the senilis group. I have already discussed this interesting question of priorities in host association in Part I (GRIFFITHS, 1964, p. 869-874), where I concluded that the senilis group was probably derived from ancestors associated with Cerodontha subgenus Dizygomyza.

It seems to me that my concept of the *senilis* group s.l. is established beyond reasonable doubt as a monophyletic group within *Chorebus* by the synapomorphy and synapooecy just mentioned. Other noteworthy morphological features for recognition of this group are as follows.

(i) Metapleural pubescence very dense, forming a well-defined rosette around the rugose swelling (compare figs. 22-23 in Part I): prododeal pubescence also very dense (synapomorph with the majority of species included in *Chorebus*).

(ii) Hind coxa with a well-defined tuft of matted pubescence near its base (? synapomorph with the *cytherea* group and those species formerly included in "Gyrocampa" and "Chorebus" in the restricted sense).

(iii) Petiole at least 1.6 times as long as wide, often very elongate, parallel-sided or only slightly widened towards its apex (? synapomorph with the *merella* and *cytherea* groups). (iv) Pubescence immediately above the base of the mandibles somewhat dense, in some species forming distinct tufts: in some species the entire back of the head is densely pubescent. (The development of rather dense pubescence above the base of the mandibles probably represents an apomorph character, associated with the development of the adjacent pronotal pubescence, in the groundplan of the *senilis* group s.l.; but the difference in the pubescence of the head between species of the *senilis* group which are relatively plesiomorph in this respect, for instance C. leptogaster (HALIDAY) and C. euryale (NIXON), and species of the *merella* and *cytherea* groups is not very great.)

(v) Wing with vein Cu_{1b} retained (fig. 154), though sometimes very short, forming a distinct angle with the transverse section of Cu_1 (plesiomorph). (Only in C. larides (NIXON) is Cu_{1b} weak, as in the lateralis/ovalis complex.)

(vi) Thorax elongate, at least 13 times as long as high.

NIXON (1943 and 1944) distinguished his "leptogaster-group" from his "senilisgroup s. str." primarily on the length and pubescence of the petiole. He states in his key to "Dacnusa" (NIXON, 1943, p. 164) that in the leptogaster-group the petiole is "very narrow, at least two and a half times as long as apically wide,

usually virtually bare . . .", while in the *senilis*-group the petiole is "less narrow, at most twice as long as apically wide, thickly hairy except along the middle line . . ." Some exceptions to this distinction were however made. C. larides (NIXON) was included in the "leptogaster-group" in spite of its relatively short, pubescent petiole: and C. glaber (NIXON) was included in the "senilis-group s. str." in spite of its bare petiole. Furthermore C. brevicornis (THOMSON), a species whose petiole does not agree well with either of the alternatives given in the key, was described under different synonyms in both groups. It seems to me therefore that NIXON's group classification requires revision.

I have also included the *posticus* group (= the *gracilis* group of NIXON, 1944) in my *senilis* group s. l. The *posticus* group is established beyond doubt as a monophyletic group by the strongly apomorph female gaster. But it also possesses all the characters appropriate to the *senilis* group s. l. (see above). I am therefore fully satisfied that the *posticus* group is a subordinate group to be included within the *senilis* group s. l. The evolution of the apomorph female gaster of the *posticus* group was probably consequent upon the change in host association from an Agromyzid host to *Psila* (Psilidae). I have already discussed this question in Part I (GRIFFITHS, 1964, pp. 850 and 870).

I am also fully satisfied that C. petiolatus (NEES) belongs to the senilis group s. l. (see also GRIFFITHS, 1964, p. 850). NIXON (1943) appears to have overlooked the presence of hind coxal tufts in this species when constructing his key to "Dacnusa". The life-history of petiolatus in unknown³, but it seems possible that its extremely large size is the result of transference from Ophiomyia to some non-Agromyzid host. The species appears synapomorph in respect of its extremely elongate petiole with species associated with the Ophiomyia pulicaria group.

Within my concept of the *senilis* group s. l. I think that four main subordinate groups may be recognised: (i) the *senilis* group sensu stricto, (ii) the *bathyzonus* group, (iii) the *posticus* group, and (iv) the *petiolatus* group. In addition there are seven species which I have not included in any subordinate group. The species included in these groups are shown in the key below.

The senilis group sensu stricto is apomorph in respect of its densely pubescent petiole: most species also have a stout, projecting ovipositor. The concept includes all species included in the senilis group s. str. by NIXON (1944) except C. glaber (NIXON), and also C. larides (NIXON), included by NIXON in his "lepto-gaster-group". The inclusion of C. brevicornis (THOMSON) in this group is tentative, as its petiole is less pubescent than in the other species.

The *bathyzonus* group is characterised by the reduction of the precoxal suture to an almost smooth well-defined linear groove. The species of this group are as far as known restricted to stem-mining species of *Ophiomyia*. There is one other species associated with a stem-mining species of *Ophiomyia* which is

³ GIARD (1904) concluded that the record of *petiolatus* as a parasite of *Platyparea poeciloptera* (SCHRANK) (Trypetidae) in stems of *Asparagus* was probably erroneous. See below under the description of *rondanii*.

probably monophyletic with the *bathyzonus* group as its precoxal suture is narrow, only weakly rugose-costate, approaching the condition found in that group: this is described below as C. *heringianus* sp. nov.

The posticus group, characterised by the strongly apomorph female gaster, has already been discussed above. There are three other species, C. tamsi (NIXON), C. rondanii (GIARD) and C. orbiculatae sp. nov. (not included in any group at present), which have mandibles similar to those of the posticus group (with tooth 1 strongly expanded, but teeth 3 and 4 relatively small). It seems probable that this feature represents synapomorphy, but this opinion is only tentative because similarity in the form of the mandibles is often the result of convergence. However if this suggested relationship is correct, it may be concluded that the change in host association which resulted in the formation of the posticus group was from Ophiomyia to Psila (not from Melanagromyza as I suggested in Part I).

In the *petiolatus* group I have included four species characterised by their extremely elongate petiole. Three of these species (*C. leptogaster* (HALIDAY), *C. femoratus* (TOBIAS) and *C. xiphidius* sp. nov.) are also synapomorph in respect of their wing venation, having a relatively short pterostigma and cell $2R_1$.

Of the three species not yet mentioned two, C. gedanensis (RATZEBURG) and C. caelebs (NIXON), are possibly synapomorph in respect of their strongly expanded cheeks. When more information on the parasites of *Hexomyza* is available their affinities may be clarified. The remaining species, C. glaber (NIXON), is unusual in being associated with a Napomyza host. Morphologically it is relatively plesiomorph, and I am unable to make any firm suggestions regarding its relationships at present (although it is very similar to leptogaster, there is no clear evidence of synapomorphy).

The key below will I hope help to identify European species of the *senilis* group s. l. I have not been able to see the types of TOBIAS' species, but I have included them in the key as best I can on the basis of their original descriptions (TOBIAS, 1962) and personal correspondance with their author. A few of the couplets in the key are based on NIXON'S (1944) keys to his "*leptogaster-group*" and "*senilis-group* s. str.", but I have made extensive revision. Some references to relevent figures in the works of NIXON (1944) and TOBIAS (1962) have been included in the key.

Key to the Chorebus senilis group sensu lato

- 1 Petiole (compare fig. 170) with dense adpressed more or less matted pubescence covering all its surface except sometimes the central line (but somewhat sparser in *brevicornis*, fig. 169). Ovipositor (φ) stout and projecting beyond the apical tergite in the retracted position in most species (except *ares*, *brevicornis* and *larides*). Back of head pubescent, often very densely so, except in *euryale*. Precoxal suture rugose-costate. Gaster not laterally compressed *senilis* group sensu stricto
- Petiole largely bare (compare fig. 168) with pubescence mainly near its base (the species with the most extensively pubescent petioles included here are *fuscipennis* and *brevifemur*, see fig. 167: these will readily be distinguished from species of the

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	sensitis group s. str. by their virtually smooth precoxal suture). Ovipositor (\mathcal{Q}) not or hardly projecting beyond the apical tergite in the retracted position, except in <i>brevi-</i> <i>femur</i> , glaber, xiphidius and the posticus group (in the latter the gaster is strongly laterally compressed)
2	Gaster beyond petiole conspicuously yellow. Legs largely pale yellow with the base of the hind coxae and sometimes the apex of the hind femora infuscated. $31-32$ antennal segments (\mathcal{Q}). Ovipositor (\mathcal{Q}) stout, but only shortly projecting beyond the apical tergite in the retracted position (its sheaths not longer than segment 1 of the hind tarsus) C. stenocerus (THOMSON), comb. nov. ⁴ (= Dacnusa praeclara NIXON, 1944, syn. nov.)
	Gaster beyond petiole varying from yellow-brown or red-brown to black 3
3	Legs with femora and tibiae entirely yellow, at most the coxae and tarsal segments 5 infuscated. Ovipositor (2) strongly projecting
4	27–28 antennal segments (\mathcal{Q}). (\mathcal{J} unknown) C. pratensis (TOBIAS), comb. nov.
	Antennal segments: $\mathcal{J}, 37; \mathcal{Q}, 31$
-	Host: an Agromyzid in Hypochoeris radicata
5	Ovipositor (\mathcal{Q}) extremely long, projecting beyond the apiacl tergite by nearly the length of the petiole (NIXON, 1944, fig. 110). Back of head largely bare and shining (as in <i>leptogaster</i>). 29 antennal segments (\mathcal{Q}). Hind coxae and greater part of the hind femora blackish
-	Ovipositor (Q) shorter, in the retracted position not projecting beyond the apical tergite by more than two-thirds of the length of the petiole. Back of head more pub- escent
6	24-26 antennal segments (Q). Ovipositor (Q) not projecting beyond the apical tergite in the retracted position. Back of head pubescent, but not densely so. Mandibles not expanded, with teeth 3 and 4 relatively small. Basal flagellar segments very short. Hind legs largely infuscated
	30 or more antennal segments. Ovipositor (P) projecting except in <i>brevicornis</i> and ares
7	Mandibles small, with tooth 4 much reduced (fig. 161). Petiole with a band of pub- escence on either side, but broadly bare along its centre line and near its apex (fig. 169). Basal flagellar segments unusually short (see the table of biometric data). Ovipositor (Q) not or only slightly projecting beyond the apical tergite in the retract- ed position
	Host: Melanagromyza aeneoventris FALLÉN
	Mandibles not so. Petiole more densely pubsecent
8	Mandibles narrow, not at all widened towards their apex, with all teeth sharply pointed (fig. 159 and NIXON, 1944, fig. 79). Pubescence of back of head not so dense as in <i>senilis</i> and <i>ares</i> , but tending to form tufts above the base of the mandibles 9
	Mandibles (figs. 158 and 160) at least slightly widened towards their apex. Pubescence of back of head dense (extremely so in <i>senilis</i> and <i>ares</i>), but not forming distinct tufts near the base of the mandibles

⁴ As NIXON (1944) suspected his *praeclara* is a synonym of *stenocera* THOMSON. As well as the male referred to in Appendix V (in Part I), there is a female of this species in the THOMSON collection without a locality label. Possibly this is from the type locality Ringsjön (since there is no specimen labelled as from that locality). At any rate it fits the description well. I am therefore designating this specimen as the lectotype of *stenocera*. G. C. D. GRIFFITHS, The Alysiinae parasites of the Agromyzidae IV

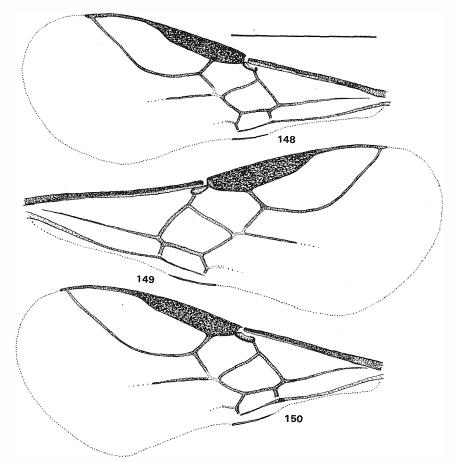
9	$38-42$ antennal segments (\mathcal{Q})
10	Antennal segments: $3, (36) - 37 - 41; 9, 35 - 36$. Legs paler, with the hind femora infuscated only apically and the hind tibiae entirely yellow or yellow-brown. Mand- ibles large (fig. 160), distinctly widened towards their apex
	Antennal segments: $3(31)-33-36$; Q , $(29)-31-34$. Legs darker, with the hind femora entirely infuscated and often the apex of the hind tibiae infuscated. Mandibles (fig. 158) only slightly widened towards their apex
11	Ovipositor (Q) projecting beyond the apical tergite in the retracted position by about two-thirds of the length of the petiole C. senilis (NEES)
	HOSTS: Melanagromyza aeneoveniris FALLÉN, Napomyza lateralis FALLÉN, N. scrophulariae SPENCER and N. cichorii SPENCER
-	Ovipositor (\mathcal{Q}) much shorter, not or only slightly projecting beyond the apical tergite in the retracted position
12	Precoxal suture visible as an almost smooth linear groove
-	Precoxal suture distinctly rugose-costate, at least on its anterior half 22
13	Mandibles narrow, with tooth 2 exceptionally long and pointed: tooth 3 weak or absent. Back of head bare or public entry is a set of the set of
	Certain species not referred to the <i>senilis</i> group s. 1. (e. g. C. <i>uliginosus</i> ((HALIDAY) and C. <i>fordi</i> (NIXON)) resemble the <i>bathyzonus</i> group (through convergence) in having a long smooth precoxal suture and densely public pronotum. These will be discussed in Part VI.
—	Mandibles not as above: tooth 2 not exceptionally long and pointed: tooth 3 always distinkt. Back of head densely public ent in all species <i>bathyzonos</i> group 14
14	Tooth 1 of mandibles enormously expanded, completely hiding the clypeus in lateral view of the head (NIXON, 1944, fig. 75). Legs bright yellow throughout. $36-37$ antennal segments (3). (Q unknown) C. sera (NIXON), comb. nov.
<u>.</u>	Tooth 1 of mandibles not or hardly expanded (compare fig. 162). Not more than 31 antennal segments 15
15	Cheeks in lateral view angularly produced (NIXON, 1944, fig. 78), bearing a conspicu- ous tuft of white pubescence above the base of the mandibles. In the male the seg- ments of at least the basal half of the flagellum are very shining and, at least on their dorsal surface, virtually bare. Petiole $3-3\frac{1}{2}$ times as long as wide. Gaster beyond petiole reddish yellow. Legs largely reddish yellow C. bathyzonus (MARSHALL) Host: Ophiomyia herakleivora SPENCER

⁵ It is difficult to judge to what extent the other differences between this species and *nomia* suggested by NIXON (1944) are reliable diagnostic criteria, since he only had a single specimen of *maculatus* before him. I have therefore thought it best to omit them from this key until further evidence is available.

^e I have seen five specimens of marsyas in the HALIDAY collection. They appear to be HALIDAY's (1839) Alysia (*Dacnusa*) senilis (NEES) Var. β , which he considered to represent *Dacnusa pulverosa* CURTIS. STELFOX was of this same opinion and labelled one of these specimens as *pulverosa*. I am considering *pulverosa* a nomen nudum, since the species was not described by CURTIS (the name was merely given in a check list). It hardly seems justifiable to consider the name available on account of HALIDAY's brief reference in synonymy.

⁷ The holotype of *Dacnusa (Dacnusa) stenocentra* THOMSON, 1895, from Arrie, Skåne, Sweden, is a very similar insect to *ares*. Its short ovipositor is extruded, but clearly would not project beyond the apical tergite if retracted. Probably this insect represents a different species from *ares* since its coloration is much paler (the basal antennal segments and legs are largely yellowish, and the gaster beyond petiole yellow-brown). Unfortunately the antennae are broken and most of the legs missing. I am not including the species in the key until further information is available.

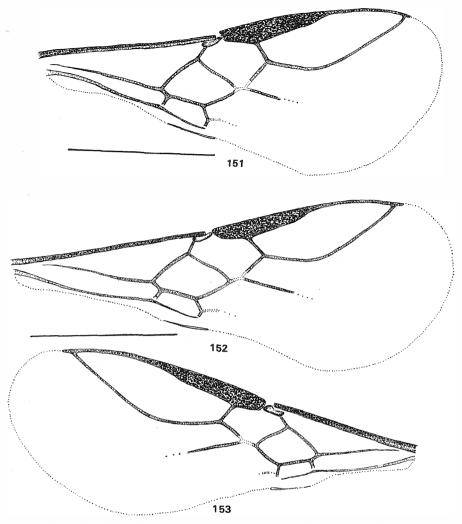
- Cheeks not or only weakly angulate; tuft of pubescence not so conspicuous. Basal
 flagellar segments pubescent in both sexes. Gaster beyond petiole usually darker 16
- a few scattered hairs on its dorsal surface (fig. 168 and Nrxon, 1944, fig. 100) . . . 18
- 17 Mandibles (fig. 162) hollowed and usually somewhat dilated posteriorly near their base below the tuft of pubescence (compare cyparissa). Ovipositor sheaths much shorter than the petiole C. fuscipennis (NIXON) Hosts: Ophiomyia heringi STARÝ, O. labiatarum HERING and O. sp. on Stachys palustris
- less dark brown with the coxae almost black. Mesoscutum entirely covered with



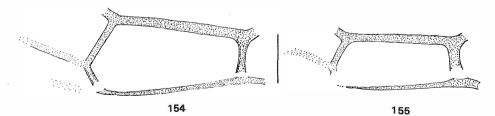
Figs. 148-150. Wings of Chorebus spp. 99: 148, C. leptogaster (HALIDAY); 149, C. glaber (NIXON); 150, C. fuscipennis (NIXON), (Scale 1 mm.)

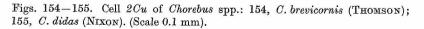
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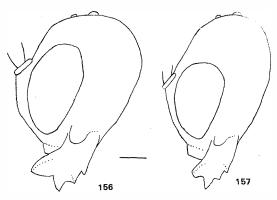




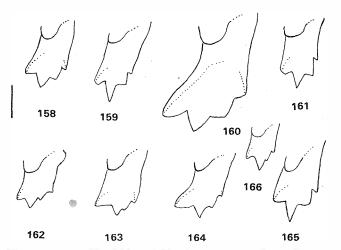
Figs. 151-153. Wings of Chorebus spp. QQ: 151, C. brevicornis (THOMSON); 152, C. senilis (NEES); 153, C. didas (NIXON). (Scale 1 mm.)







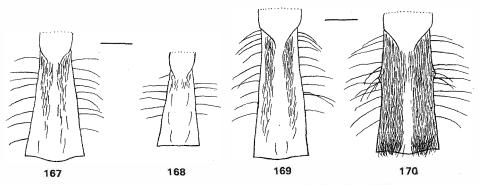
Figs. 156-157. Head and mandibles in lateral view of *Chorebus glaber* (NIXON) (to illustrate range of variation). (Scale 0.1 mm.)



Figs. 158-166. Mandibles of Chorebus spp.: 158, C. senilis (NEES); 159, C. nomia (NIXON); 160, C. marsyas (NIXON); 161, C. brevicornis (THOMSON); 162, C. fuscipennis (NIXON); 163, C. leptogaster (GALIDAY); 164, C. orbiculatae sp. nov.; 165, C. cybele (NIXON); 166, C. ibericus sp. nov. (Scale 0.1 mm.)

	Antennal segments: $29-31$; $9, 27-30$	19
19	Mandibles hollowed and dilated posteriorly near their base below the tuft of pub-	
	escence (NIXON, 1944, fig. 74). Gaster beyond petiole deep yellow. Legs 1 and 2 yel-	
	low, the hind legs light brown C. cyparissa (NIXON), comb. no	v.
	Mandibles hollowed posteriorly near their base, but not or hardly dilated	20
20	Lateral lobes of mesoscutum sculptured and densely pubescent. Legs light brown	
	with the hind coxae infuscated C. herbigradus (TOBIAS), comb. no	v.
	Lateral lobes of mesoscutum at least partly bare	21
43*		

G. C. D. GRIFFITHS, The Alysiinae parasites of the Agromyzidae IV



Figs. 167-170. Petioles of Chorebus spp.: 167, C. fuscipennis (NIXON); 168, C. caesariatus sp. nov.; 169, C. brevicornis (THOMSON); 170, C. senilis (NEES). (Scale 0.1 mm.)

- 22 Petiole extraordinarily elongate, $3-3\frac{1}{2}$ times as long as wide. Back of head more or less bare centrally, pubescent only at its sides (near the mandibles) 23 - Petiole not so elongate, less than three times as long as wide 26 23 Very large species, about 5 mm. long. Antennal segments: 3, 45-51; 9, 43-45. Tooth 1 of mandible much expanded. Ovipositor (9) not projecting beyond the apical tergite in the retracted position C. petiolatus (NEES), comb. nov. Smaller species (up to 2.8 mm. long). Not more than 33 antennal segments. Tooth 1 of mandibles hardly expanded (fig. 163). Wing with pterostigma and cell $2R_{1}$ 24 24 Coxae yellow. Gaster beyond petiole conspicuously yellow or yellow-brown. Ovipositor (Q) projecting beyond the apical tergite in the retracted position Host: Ophiomyia sp. on Picris
- 26 Gaster laterally compressed towards its apex in the female; ovipositor slender and upcurved, projecting beyond the apex of the gaster (NIXON, 1943, figs. 63 and 64; and TOBIAS, 1962, figs. 54 and 55). Tooth 1 of mandible very much expanded

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	(NIXON, 1944, fig. 145). Back of head more or less bare centrally, pubescent only at its sides (near the mandibles)
27	glaber
-	figs. 81 and 82); but the mandibles are relatively small (tooth 1 not expanded) 28 Head more transverse, or, if with somewhat expanded cheeks (<i>rondanii</i>), the mand-
28	ibles are also large, with tooth 1 distinctly expanded
_	Gaster beyond petiole red-brown or brown. Legs largely reddish yellow or yellow- brown, the hind pair being only slightly darker than legs 1 and 2. Back of head fairly densely public cent
29	Mesoscutum largely bare (with some fine pubescence on its anterior face and a few hairs along the former course of the notaulices: central lobe either bare or with a little fine scattered pubescence). Mandibles small, with tooth I only slightly expanded. Precoxal suture narrow, only weakly rugose-costate. Legs ochreous yellow or yellow-brown with the hind coxae slightly darker, brown or dark brown
_	Mesoscutum with at least its anterior face and central lobe fairly densely pubescent. Mandibles usually with tooth 1 strongly expanded (compare fig. 164) (but variable in glaber, figs. 156 and 157)
30	Legs entirely reddish yellow, except that the hind femora are slightly darkened at their apex. Gaster beyond petiole yellow. Basal flagellar segments markedly yellowish beneath. (Q unknown)
31	Gaster beyond petiole bright yellow. Legs 1 and 2 deep yellow, but the hind legs con- trastingly darker, with the femora, tibiae and tarsi uniformly dark brown. Petiole slightly widened towards its apex C. rondanii (GIARD)
	Host: Ophiomyia simplex LOEW
-	Gaster beyond petiole darker. Legs I and 2 yellow-brown or brown. Petiole par- allel-sided
32	Back of head virtually bare centrally (although densely pubescent at its sides, near the mandibles). Hind tibiae largely yellow-brown or red-brown, usually becoming

⁸ Note on the posticus group. The available names for species of this group are Alysia (Dacnusa) postica HALDAY, 1839, Dacnusa egregia MARSHALL, 1891, Dacnusa (Dacnusa) dentifera THOMSON, 1895, Dacnusa compressiventris TE-LENGA, 1935, Dacnusa selene NIXON, 1937, Dacnusa dentata TOBIAS, 1962, and Dacnusa cultrata TOBIAS, 1962. The earliest and best-known name, Alysia gracilis NEES, 1834, is a primary homonym of Alysia gracilis CURTIS, 1826, and must presumably be discarded unless reference is made to the International Commission on Zoological Nomenclature. The application of these names and their synonymy requires reappraisal. I suspect that the concept of gracilis in NIXON (1944) is composite, since the range of variation recorded in colour and the number of antennal segments is very wide. One species (usually called gracilis in the literature) is well known as a parasite of Psila rosae FABRICUS (Psilidae). It is possible that C. tamsi (NIXON), whose female is unknown, also belongs to the posticus group; but I have provisionally keyed it in couplet 30 below.

I have not been able on the limited information available to give a satisfactory definition of males of the *posticus* group in this couplet.

black only near their apex. Ovipositor (Q) stout, distinctly projecting beyond the the apical tergite in the retracted position C. glaber (NIXON) Host: Napomyza cichorii SPENCER

In the descriptions which follow the following common characters may be assumed, unless otherwise stated.

Clypeus dark, like the face. Face almost smooth, covered with dense, fine pubescence which is directed upwards over its centre but downwards along the eye-margins. Mandibles 4-toothed. Thorax elongate (at least 1.3 times as long as high). Precoxal suture long, reaching the hind margin of the mesepisternum. Sides of pronotum, at least along and below the oblique suture, and subalar callus covered with dense, opaque pubescence. Metapleural pubescence very dense, forming a well-defined rosette around the rugose swelling (compare figs. 22-23 in Part I): propodeum covered with similar dense pubescence. Tergite 3 without basal pubescence.

Wing (figs. 148-152 and 154) with vein Cu_{1b} retained (though sometimes very short), forming a distinct angle with the transverse section of Cu_1 .

Chorebus senilis (NEES), comb. nov.

Bassus senilis NEES, 1814

Alysia senilis (NEES), NEES, 1834

Alysia (Dacnusa) senilis (NEES), HALIDAY, 1839 (in part)

Dacnusa senilis (NEES), MARSHALL, 1891, 1895 and 1897 (in part), NIXON, 1937 and 1944 Dacnusa (Dacnusa) tomentosa THOMSON, 1895

Dacnusa nemesis MORLEY, 1924

Colour. Palpi varying from yellow to dark brown. Labrum deep yellow. Antennae entirely dark or becoming obscurely brown towards their base. Legs largely yellow-brown or brown, with the coxae, especially the hind coxae, darker, usually virtually black: hind femora dark brown or black, at least near their apex: hind tibiae often distinctly infuscated towards their apex. Gaster beyond petiole dark, sometimes with tergite 3 reddish.

Morphology. Back of head clothed with very dense adpressed pubescence, but no distinct tufts are formed above the base of the mandibles. Mandible (fig. 158) slightly expanded towards its apex, with all four teeth strongly developed. Antennal segments: 3, 33-36; 9, 31-34 (bred material only). Palpi long.

Mesoscutum conspicuously punctate on about its anterior half, with extremely dense fine pubescence covering most of its surface (although sometimes this tends to become sparse on the posterior half of the lateral lobes): notaulices not extending longitudinally (only their lateral extensions distinct). Sides of pronotum also entirely covered with extremely dense fine pubescence, beneath which its surface can be seen to be strongly punctate. Subalar callus and the

anterior edge of the mesepisternum with similar dense pubescence: precoxal suture rugose-costate anteriorly. Pubescence of metapleuron, propodeum and the base of the hind coxa extremely dense. Petiole 1.9-2.2 times as long as wide, more or less parallel sided, with its entire surface covered by extremely dense fine pubescence (fig. 170): this shows some tendency to become denser towards the apical corners (although no distinct tufts are formed). Ovipositor (\mathcal{Q}) usually very stout, projecting beyond the apical tergite in the retracted position by about two-thirds of the length of the petiole (but only by about one-third of the length of the petiole in the small female bred from Napomyza cichorii SPENCER).

Wing (fig. 152) with vein R_s weakly sinuate: 1m-cu rejected from cell R_s .

Breeding records

Host 1 – Melanagromyza aeneoventris FALLÉN (= cirsii RONDANI)

1 3 from puparium 15. ix. 61 in stem of *Cirsium palustre*, Llanrhidian, Gower, Wales, em. 6. iv. 62 (GCDG). 19 from puparium 23. ix. 61 in stem of *Cirsium vulgare*, Betchworth, Surrey, England, em. 15. x. 61 (GCDG). 19 from stem of *Cirsium palustre*, Oxford, England, em. 30. iv. 23, leg WATERS (BM). 5 ex. from puparia in stems of *Cirsium vulgare* (= *Carduus lanceolatus*), Farnham Royal and Cippenham, Bucks., England, em 20. i and 4-13. v. 38 (BM). 13 from puparium in stem of *Cirsium vulgare*, Reading, Berks., England, em. 4. v. 38 (BM).

Host 2 — Napomyza lateralis FALLÉN

19 from puparium in *Matricaria chamomilla*, Zaventem, Belgium, 1964/65, leg. LOUNSKY (Station d'Entomologie, Gembloux).

Host 3 — Napomyza cichorii Spencer

 $1 \, \varphi$ from puparium in leaves (chicons) of *Cichorium intybus*, Zaventem, Belgium, em. 13. v. 66, leg LOUNSKY (GCDG).

Host 4 — Napomyza scrophulariae Spencer

2 QQ from puparia 19. ix. 64 in stems of *Scrophularia nodosa* on dunes at Newcastle, Co. Down, Ireland, em. 7-8. v. 65 (GCDG): a dead male was found on the same occasion inside a hollow stem of this plant, clearly emerged from this same host.

ALLEN (1956) also recorded this species (det. M. W. R. DE V. GRAHAM) from puparia of a Napomyza species found in stems of Anthriscus sylvestris at Sunninghill, Berks., England.

There are a number of species similar to *senilis* whose life-history has not yet been established (see the key above). Important characters for distinguishing *senilis* from these are the number of antennal segments (contrast *marsyas*), the form of the mandibles (contrast *nomia*, *maculatus* and *marsyas*) and the projecting ovipositor (contrast *ares*).

I have reexamined the female (not male as stated in Appendix V in Part I) of THOMSON'S (1895) tomentosa from the type locality (Pålsjö, Sweden), and am satisfied that it represents this species, although the infuscation of its hind femora and tibiae is not very marked. (It is clear from the shape of the mandibles that it does not represent C. marsyas (NIXON)).

Chorebus pulchellus sp. nov.

Colour. Palpi yellow. Labrum and centre of mandibles orange-yellow. Clypeus red-brown or black. Basal antennal segments yellow-brown (clearly so at least as far as the fourth flagellar segment), this colour merging gradually into the darker colour of the more apical flagellar segments. Legs deep yellow, with only tarsal segments 5 and sometimes the base of the hind coxa infuscated. Gaster beyond petiole brown or red-brown.

Morphology. Back of head evenly clothed with fairly dense adpressed public pub

Mesoscutum strongly punctate on about its anterior half, with dense whitish adpressed pubescence covering all its surface except part of the posterior half of the lateral lobes: longitudinal extensions of notaulices absent or weak. Sides of pronotum punctate, entirely covered by dense pubescence similar to that of the mesoscutum. Precoxal suture narrow, but distinctly rugose-costate anteriorly. Pubescence of metapleuron, propodeum and base of the hind coxa whitish and extremely dense. Petiole 1.8-2.0 times as long as wide, slightly widened towards its apex, entirely covered (except for the narrow central keel) with short, very dense pubescence which shows a tendency to become denser towards the apical corners (as in *senilis*, fig. 170). Ovipositor (\mathfrak{Q}) stout, projecting beyond the apical tergite in the retracted position by about half the length of the petiole.

Wing with vein R_s sinuate: *1m-cu* rejected from cell R_s .

Holotype Q, 2 33 paratypes from puparia 10. vi. 36 in *Hypochoeris radicata*, Ribnitz/Körkwitz, Mecklenburg, Germany, em. 23. vi. 36, leg. BUHR (GCDG).

Dr. BUHR thought that the host was probably *Phytomyza cecidonomia* HE-RING, but the puparia were not retained. Since no species of the *senilis* group has been confirmed from any *Phytomyza* host, I suspect that BUHR's suggestion may not be correct. The question cannot be settled until additional bred material is obtained.

This species is a typical member of the *senilis* group sensu stricto, but is unusual in having entirely yellow legs. TOBIAS (1962) has described a yellowlegged species of this group as *Dacnusa pratensis*. But he states that his two females have 27 and 28 antennal segments. The description appears appropriate to the species before me in most respects, but the range of antennal segments (27-37) assumed if it is considered that only one species is involved, is wider than has been established for any species of the *senilis* group. I have therefore concluded that the material before me probably represents a distinct species.

Chorebus brevicornis (THOMSON)

Dacnusa (Dacnusa) brevicornis Thomson, 1895 Dacnusa chrysippe Nixon, 1944 Dacnusa ea Nixon, 1944 Chorebus brevicornis (Thomson), Griffiths, 1964

Colour. Palpi and labrum brown or yellow-brown. Antennae usually entirely dark (at most reddish at their base). Centre of mandibles red-brown. Legs 1 and 2 yellow-brown or reddish; hind legs contrastingly dark, varying from redbrown to virtually black. Gaster entirely dark.

Morphology. Back of head evenly clothed with dense adpressed pubescence, but no distinct tufts are formed above the base of the mandibles. Mandibles (fig. 161) small, not expanded, with tooth 4 much reduced. Antennal segments: \Im , (32)-33-36; \Im , 30-33: basal flagellar segments unusually short (see the table of biometric data).

Mesoscutum punctate anteriorly, with its entire surface covered with short dense pubescence which on the lateral lobes is partly directed laterally: notaulices with only their lateral extensions distinct, at most extending longitudinally as feeble smooth impressions. Precoxal suture narrow, weakly rugose-costate anteriorly. Petiole (fig. 169) 2.4—2.6 times as long as wide, more or less parallelsided, with a band of pubescence on either side but broadly bare and strongly shining along its centre-line and near its apex. Ovipositor (\mathfrak{Q}) stout but short, not or only slightly projecting beyond the apical tergite in the retracted position.

Wing with cell $2R_1$ rather elongate: vein R_s hardly sinuate: position of 1m-cu variable, sometimes distinctly rejected from cell R_s (as fig. 151), but often only narrowly rejected or almost interstitial.

Breeding records

Host – Melanagromyza aeneoventris FALLÉN (= cirsii RONDANI)

1 $\[mu]$ from puparium in stem of *Cirsium palustre*, Woodwalton Fen, Hunts., England, em. 3. iv. 61 (GCDG). 1 $\[mu]$ from puparium in stem of *Cirsium* sp., Scratch Wood, London, em. 11. v. 55, leg. SPENCER (GCDG). 2 $\[mu]$ (including the holotype of *Dacnusa ea* NIXON) from stems of *Cirsium vulgare* (= *Carduus lanceolatus*), Reading, Berks., England, em. 15. vi. 38 (BM). 3 $\[mu]$ from puparia 18. ix. 65 in stems of *Cirsium arvense*, Wöllnitz, Jena, Thuringia, Germany, em. 2. xi. 65, 9 and 11. iii, 66, leg. BUHR no. 2661 (GCDG). 1 $\[mu]$ from puparium in stem of *Cirsium arvense*, Mühlhausen, Thuringia, Germany, em. spring '65, leg BUHR no. 2322 (GCDG). 1 $\[mu]$, Neckarrems, Württemberg, Germany, em. 30. v. 55, leg GROSCHKE (STGT). 1 $\[mu]$, Stuttgart-Hofen, Germany, em. 30. v. 55, leg. GROSCHKE (STGT). 3 $\[mu]$ Feuerbach Tal, Stuttgart, em. 1. vi. 55, leg GROSCHKE (STGT).

This species will be readily distinguished from other species of the *senilis* group sensu stricto (including *C. senilis* (NEES) which is associated with the same host) by the public of its petiole (fig. 169), the form of its mandibles (fig. 161), its short basal flagellar segments and short ovipositor. The synonymy of NIXON'S (1944) two names was proposed in Part I of this paper (GRIFFITHS, 1964).

Chorebus rondanii (GIARD), comb. nov.

Dacnusa Rondanii GIARD, 1904

Dacnusa galba NIXON, 1944, syn. nov.

Colour. Palpi and labrum yellow. Basal antennal segments yellowish or reddish as far as about the third flagellar segment. Centre of mandibles yellowbrown. Legs 1 and 2 deep yellow: hind legs contrastingly darker, with the femora, tibiae and tarsi uniformly dark brown, but the coxae, trochanter and trochantellus paler, more or less yellow-brown. Gaster beyond petiole bright yellow, in strong contrast with the shining black petiole.

Morphology. Pubescence of head as described for *leptogaster*. Mandibles (compare fig. 164) with tooth 1 strongly expanded, but teeth 3 and 4 relatively small. Antennal segments: 3, 30 (holotype of *Dacnusa galba* NIXON); 9, 25, 26 (bred specimens). Palpi fairly long.

Mesoscutum with shallow sculpture on its anterior face extending onto the anterior part of the central lobe, with fairly dense pubescence covering its anterior face and central lobe, but the lateral lobes largely bare: notaulices distinct, reaching the posterior fovea as shallow V-shaped impressions. Precoxal suture narrow but conspicuously rugose-costate anteriorly. Dorsal face of propodeum partly with only sparse pubescence, so that its shining rugose surface is clearly visible: but its posterior face is very densely pubescent (as normally in the *senilis* group). Petiole 2.0-2.2 times as long as wide, slightly widened towards its apex, with a little pubescence on each side near its base but otherwise almost bare. Ovipositor (\mathfrak{Q}) directed upwards in the retracted position, slightly projecting beyond the apical tergite. Hind femur unusually short and thick.

Wing with cell $2R_1$ fairly elongate: vein R_s weakly sinuate: 1m-cu well rejected from cell R_s .

Breeding records

Host – Ophiomyia simplex LOEW

BARNES and WALTON (1934) recorded this species (as *Dacnusa bathyzona* MARSHALL) from the above host in stems of cultivated *Asparagus officinalis* at Harpenden, Herts., and Evesham, Worcs., England. Further breeding data for Harpenden material with some observations on the habits of the insect are given by BARNES (1937). The above description is based on my examination of 11 of BARNES' specimens from Harpenden (retained in my personal collection). These appear to represent the same species as NIXON'S *Dacnusa galba*.

GIARD's (1904) description of rondanii (from Argenteuil, France) is as follows.

"Ce parasite ressemble beaucoup, comme forme, comme couleur et comme ornamentation, à Dacnusa petiolata NEES, dont il est, en quelque sorte, une reduction. Sa taille est en effet à peu près moitié moindre. D. petiolata mesure 5 mill. Notre espèce est longue de 3 mill. environ; les antennes ont aussi 3 mill.; les ailes antérieures 2.5 mill. En outre, les nervures cubitale et postérieure, au lieu d'être interrompues, se prolongent jusqu'au bord de l'aile.

Il me parait trés possible que ce *Dacnusa*, que j'apellerai *Dacnusa Rondanii*, soit celui qui a été indiqué par RONDANI sous le nom de *D. petiolata*, comme parasite de *Platyparea poeciloptera*.

Mais, comme les *Dacnusa* sont généralement parasites des *Agromyza*, je ne puis affirmer que notre espèce ne soit pas plutôt parasite d'*A. simplex* dont le présence dans les Asperges aurait échappé à RONDANI. De nouvelles recherches sont nécessaires pour lever toute espèce de dout à cet égard."

As a description this is entirely inadequate, but appears nevertheless to make the name *rondanii* available under the current rules of nomenclature. I have not been able to obtain any information on whether GIARD's material still exists. In the circumstances I have assumed on the evidence of host association that GIARD's species was the same as the species obtained by BARNES. Should it later be discovered that more than one species of the *senilis* group is associated with *O. simplex* LOEW, then this assumption could be challenged. But on present information it seems to me reasonable that this assumption should be made, since BARNES' extensive breedings suggest that, at least in England, there is only one species of *Chorebus* associated with this host.

This species should be readily recognised by its extensive yellow coloration (but with contrastingly dark hind legs), and the form of the mandibles.

Chorebus orbiculatae sp. nov.

Colour. Palpi and labrum deep yellow or yellow-brown. Antennae entirely dark. Centre of mandibles red-brown. Legs 1 and 2 more or less uniformly yellow-brown or brown: hind legs with the coxae, femora, tibiae and tarsi uniformly dark brown or almost black, but the trochanter and trochantellus contrastingly yellow-brown. Gaster beyond petiole red-brown, becoming darker towards its apex.

Morphology. Back of head distinctly publicate centrally, as well as at its sides: a distinct opaque whitish tuft of publications is formed on the posterior edge of the head near the base of the mandibles. Mandibles (fig. 164) with tooth 1 strongly expanded, but teeth 3 and 4 relatively small. Antennal segments: 3, 32; 9, 27 (2 ex.). Palpi fairly long.

Mesoscutum roughened anteriorly, with dense pubescence covering all or most of its surface (at most the posterior half of the lateral lobes partly bare): notaulices weak, hardly extending longitudinally on the dorsal surface of the mesoscutum. Precoxal suture narrow, weakly rugose-costate anteriorly. Petiole about 2.6 times as long as wide, parallel-sided or very slightly widened towards its apex, largely bare and shining, with some fine pubescence near its base but otherwise only about 4-5 pairs of scattered hairs on its dorsal surface. Ovipositor (\mathfrak{Q}) similar to that of *leptogaster*, directed upwards in the retracted position, only slightly projecting beyond the apical tergite.

Wing with vein R_s weakly sinuate: *1m-cu* rejected from cell R_s .

Breeding records

Host – Ophiomyia orbiculata HENDEL

Holotype Q and paratype Q from puparia 13. vii. 62 in roots and stems of cultivated *Pisum* sativum, Potters Bar, Herts., England, leg. DEEMING (GCDG). 1 Q paratype from puparium ix. 63, same plant and locality, em. 14. v. 64 (GCDG).

In addition I have received a male, also designated a paratype, caught on 20. vii. 50 at Portrane, Co. Dublin, Ireland (in Mr. A. W. STELFOX's collection).

This species may be recognised by its very dark coloration, expanded mandibles (fig. 164) and the presence of pubescence on the central part of the back of the head. The male may be confused with that of C. glaber (NIXON) if the last character is not properly appreciated (the females are readily separable by the length of the ovipositor).

Chorebus heringianus sp. nov.

Colour. Palpi ochreous yellow. Labrum orange. Antennae dark, with the scape and annellus and sometimes the first flagellar segment yellow-brown or red-brown. Centre of mandibles red or red-brown. Legs ochreous yellow or yellow-brown with the hind coxae and sometimes the tarsi slightly darker, brown or dark brown. Gaster beyond petiole dark brown.

Morphology. Back of head bare except at its sides (near the mandibles). Mandibles small, with tooth 1 only slightly expanded. Antennal segments: 3, 28 (2 ex.), 30; 9, 27 (2 ex.). Palpi fairly long.

Mesoscutum largely smooth, bare and shining, often flattened centrally, with some fine pubescence on its anterior face (mainly towards its sides) and a few hairs along the former course of the notaulices (the dense tufts on the hind margin of the mesoscutum are however retained as in other members of the *senilis* group): in two specimens there is also a little pubescence on the central lobe, but in the other three specimens these are virtually bare: notaulices distinct anteriorly only, although in two of the specimens some indication of V-shaped impressions can be seen as far as the posterior fovea. Sides of pronotum with densely matted pubescence along and below the oblique suture (as in other members of the *senilis* group), but largely bare and shining above the suture. Precoxal suture narrow, weakly rugose-costate at least anteriorly. Petiole 2.1-2.6 times as long as wide, more or less parallel-sided, strongly shining and almost bare, with at most a few fine hairs at its base and along its sides. Ovipositor (\mathfrak{Q}) not projecting beyond the apical tergite in the retracted position.

Wing with cell $2R_1$ elongate; vein R_s almost evenly curved, not at all sinuate; *1m-cu* rejected from cell R_s .

Host – Ophiomyia thalictricaulis HERING

Holotype Q, 2 33 paratypes from puparia in stems of *Thalictrum minus*, Ochsenburg, Kyffhäuser, Thuringia, Germany, em. 8–16. ii. 63, HERING no. 1914, leg. BUHR (GCDG). 1 3 paratype from puparium in stem of *Thalictrum minus*, Jenzig, Jena, Thuringia, em. 15. vii. 63, HERING no. 1967, leg BUHR (GCDG). 1 \bigcirc paratype from puparium 17. ix. 65 in stem of *Thalictrum minus*, Lobeda, Jena, Thuringia, em. 24. iii. 66, leg. BUHR no. 2664 (GCDG).

I have much pleasure in naming this species in honour of Professor Dr. E. M. HERING, who has contributed so much to our knowledge of leaf-mining insects.

This species has a very narrow precoxal suture, approaching the condition found in the *bathyzonus* group, but this is weakly rugose-costate at least anteriorly. It differs from all other species of the *senilis* group in having a largely bare mesoscutum (in strong contrast with the sides of the pronotum and the propodeum, which are densely publicent as normally in this group).

Chorebus caesariatus sp. nov.

Colour. Palpi and labrum yellow-brown. Antennae entirely dark. Centre of mandibles red or red-brown. Legs 1 and 2 largely brown, the hind legs somewhat darker, more or less dark brown, with the coxae almost black (sometimes the

hind femora and tibiae are also virtually black). Gaster beyond petiole yellowbrown, either unicolorous or with the apical tergites becoming darker.

Morphology. Back of head covered with dense whitish pubescence, which is fairly evenly distributed and does not form well-defined tufts near the base of the mandibles: the vertex and temples however bear only the normal two or three rows of pubescence. Mandibles not expanded, with all four teeth clearly defined. Antennal segments: \bigcirc , 23 (3 ex.), 24 (5 ex.).

Mesoscutum slightly flattened centrally, roughened anteriorly, entirely covered with extremely dense whitish pubescence which on the lateral lobes is largely directed laterally: notaulices reaching the posterior fovea (although much obscured by the pubescence). Precoxal suture visible as a smooth welldefined linear groove. Pubescence of metapleuron, propodeum and hind coxa extremely dense and whitish. Petiole (fig. 168) 2.3-2.5 times as long as wide, very narrow at its base and distinctly widened towards its apex, strongly shining and largely bare, with some fine pubescence near its base but otherwise only 3-5 pairs of long hairs on its dorsal surface. Ovipositor(\mathcal{Q}) not projecting beyond the apical tergite in the retracted position.

Wing with vein R_s weakly sinuate: *1m-cu* clearly rejected from cell R_s .

Host – Ophiomyia sp. ? curvipalpis ZETTERSTEDT

Holotype Q, 7 Q Q paratypes from puparia 18. viii. 63 in stems of *Medicago sativa*, Lido di Venezia, Italy, em. 23. viii-15. ix. 63 (GCDG).

This small species will readily be distinguished from other members of the *bathyzonus* group by its lower number of antennal segments and extremely dense mesoscutal pubescence.

Chorebus fuscipennis (NIXON), comb. nov.

Dacnusa fuscipennis NIXON, 1937 and 1944

Colour. Palpi and labrum yellow or ochreous yellow. Antennae more or less entirely dark, rarely becoming brownish towards their base. Centre of mandibles red-brown. Legs 1 and 2 largely ochreous yellow or yellow-brown: hind legs more or less uniformly brown except that the trochanter and trochantellus are often somewhat paler. Gaster beyond petiole uniformly coloured, varying from deep yellow to dark brown.

Morphology. Back of head clothed with fairly dense pubescence which becomes denser towards the base of the mandibles, where it forms whitish or grey tufts which are distinct in lateral view. Mandibles (fig. 162) hardly expanded towards their apex, but with a characteristic hollowed dilation posteriorly near their base below the tuft of pubescence (as also in *C. cyparissa* (NIXON)). Antennal segments: 3, (26)-28-29; 9, (23)-25-28. Palpi fairly long.

Mesoscutum usually not conspicuously flattened centrally (though this is so in a few specimens), usually slightly punctate anteriorly, completely covered with dense pubescence, which sometimes tends to be directed laterally on the lateral lobes: notaulices almost absent. Precoxal suture visible as a smooth 674

well-defined linear groove. Petiole (fig. 167) 2.3-2.5 times as long as wide, usually slightly widened towards its apex, rather densely pubescent on its basal half but becoming largely bare towards its apex. Ovipositor (\mathcal{Q}) not or hardly projecting beyond the apical tergite in the retracted position.

Wing (fig. 150) with cell $2R_1$ rather elongate: R_s hardly sinuate: 1m-cu rejected from cell R_s .

Breeding records

Host 1 – Ophiomyia heringi STARÝ

2 ex. from puparia 15. viii. 35 in stems of Lapsana communis, Dargun, Warsow, Mecklenburg, Germany, em. 20. viii. 35, leg. BUHR (GCDG). 1 ex. from pupariam 25. viii. 35, same plant and locality, em. 23. ii. 36, leg. BUHR (GCDG). 2 ex. from puparia 31. viii. 35 in stems of Lapsana communis, Laage, Mecklenburg, em 30. ix. 35 and 26. ii. 36, leg. BUHR (GCDG). 5 ex. from puparia 25 and 31. viii. 35 in stems of Lapsana communis, Teterow, Mecklenburg, em. 27. ii -18. iii. 36, leg. BUHR (GCDG). 5 ex. from puparia 3. ix. 35 in stems of Lapsana communis, Penzlin, Mecklenburg, em. 12-20. iii. 36, leg. BUHR (GCDG). 2 ex. from puparia 15. ix. 35 in stems of Lapsana communis, Penzlin, Mecklenburg, em. 12-20. iii. 36, leg. BUHR (GCDG). 2 ex. from puparia 15. ix. 35 in stems of Lapsana communis, Serrahn, Mecklenburg, em. 19-20. ii. 36, leg. BUHR (GCDG). 11 ex. from puparia 15. ix. 35 in stems of Mycelis muralis, Serrahn, Mecklenburg, em. 19. ii-1. iii. 36, leg. BUHR (GCDG). 4 ex. from puparia 15. ix. 35 in stems of Mycelis muralis, Serrahn, Mecklenburg, em. 19-20. ii. 35 in stems of Lapsana communis, Mecklenburg, em. 15-22. ii. 36, leg. BUHR (GCDG). 1 $_{\odot}$ from stem of Lapsana communis, Mühlhausen, Thuringia, Germany, em. 25. i. 65, leg. BUHR, HERING no. 2233 (GCDG). 2 QQ from puparia in stems of Campanula persicifolia, Hedlandet, Södermanlad, Sweden, em. 15 and 28. vii. 43, leg. LUNDQVIST (LUND).

Host 2 — Ophiomyia labiatarum HERING

 1_{\circ} from puparium 16. viii. 35 in stem of *Stachys silvatica*, Dargun, Warsow, Mecklenburg, Germany, em. 20. viii. 35, leg, BUHR (GCDG).

Host 3 - Ophiomyia sp.

 1_{\odot} 299 from puparia 28. ix. 60 in stems of *Stachys palustris*, Woodwalton Fen, Hunts., England, em. 6-23. iv. 61 (GCDG).

Host 3 was recorded in GRIFFITHS (1963a) and SPENCER (1964) as Ophiomyia labiatarum HERING, but I think it probable that it represents a different species, since the hind spiracles of the puparia have 9-11 bulbs in contrast with about 7 in the true labiatarum. No bred flies have yet been obtained. The series of fuscipennis bred from this host are darker coloured than NIXON's holotype and the other material before me (on which the above description has been based), having the legs, mandibles, labrum and palpi entirely dark brown or black. It is possible that they represent a different species, but I do not wish to offer a firm opinion on the basis of a single series.

NIXON (1944) has emphasised that this species has infumated wings: however this character is subject to individual variation and it does not seem to me very reliable for purposes of identification. An important character for identifying this species is that its petiole (fig. 167) is more densely public enter than in other species of the *bathyzonus* group except *C. brevifemur* (TOBIAS). Also the mandibles have a hollowed dilation near their base (fig. 162), as in *C. cyparissa* (NIXON).

Chorebus lychnidis sp. nov.

Colour. Palpi dull yellow or yellow-brown. Labrum orange-yellow. Antennae dark except for the yellow-brown scape and annellus. Centre of mandibles redbrown. Legs 1 and 2 ochreous yellow: hind legs uniformly brown except for the yellowish trochanter and trochantellus. Gaster beyond petiole largely brown (but tergite 3 yellow-brown in the male).

Morphology. Back of head clothed with short, fairly dense pubescence (but not so densely pubescent as for instance in *fuscipennis* and *caesariatus*): in lateral view a small whitish tuft of pubescence can be seen on the posterior edge of the head near the base of the mandibles. Cheeks not conspicuously angled (contrast *bathyzonus*). Mandibles hardly expanded towards their apex, with tooth 2 relatively large and pointed, rather conspicuously hollowed posteriorly near their base below the tuft of pubescence. Antennal segments: 3, 30; 2, 28: male flagellum normally pubescent (contrast *bathyzonus*). Palpi long.

Mesoscutum not distinctly flattened centrally, with punctate sculpture on its anterior face and the anterior part of its central lobe, with its anterior face and central lobe densely public public but the lateral lobes largely bare: notaulices distinct, V-shaped, reaching the posterior fovea. Precoxal suture visible as a smooth well-defined linear groove. Petiole very narrow and elongate, over 3 times as wide as long, parallel-sided, with a little public energy to the but otherwise only 3-4 pairs of hairs on its dorsal surface. Ovipositor (\mathfrak{Q}) hardly projecting beyond the apical tergite in the retracted position.

Wing with vein R_s weakly sinuate: *Im-cu* clearly rejected from cell R_s .

Host – Ophiomyia sp. ? melandricaulis HERING

Holotype \mathcal{F} , paratype \mathcal{Q} from puparia 9. vii. 61 in stems of *Lychnis flos-cuculi*, Woodwalton Fen, Hunts., England, em. 2. viii. 61 (GCDG).

These specimens were referred to in GRIFFITHS (1963a) as "Dacnusa sp. (leptogaster group)". They are very similar to C. nerissa (NIXON), but are darker coloured. I think they probably represent a different species, but this conclusion should be reappraised when more material is available.

Chorebus bathyzonus (MARSHALL), comb.nov.

Dacnusa bathyzona MARSHALL, 1891, NIXON, 1937 and 1944

Colour. Palpi and labrum yellow. Antennae almost entirely dark (with only the annellus and the ventral surface of the scape yellow-brown) in the male, but with their basal segments (as far as the first or second flagellar segment) yellowish in the female. Centre of mandibles red-brown. Legs largely deep yellow, but with the fifth tarsal segments of legs 1 and 2, the entire hind tarsi and the apex of the hind tibiae infuscated (and sometimes also the hind femora and the rest of the hind tibiae red-brown). Gaster beyond petiole largely bright reddish yellow.

Morphology. Back of head densely clothed with adpressed pubescence: in lateral view the cheeks appear angularly produced and bear a conspicuous tuft

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of matted white pubescence above the base of the mandibles. Mandibles hardly expanded, with all four teeth clearly defined, hollowed posteriorly near their base below the tuft of pubescence. Antennal segments: 3, 28-32; 9, 26-31: in the male the segments of at least the basal third of the flagellum are very shining and, at least on their dorsal surface, virtually bare. Palpi long.

Mesoscutum often somewhat flattened centrally, with its anterior face and central lobe punctate and very densely public entry, but the lateral lobes contrastingly smooth, shining and largely bare: notaulices distinct, V-shaped, reaching the posterior fovea. Precoxal suture visible as a smooth well-defined linear groove. Petiole very narrow and elongate, $3-3\frac{1}{2}$ times as long as wide, parallel-sided, virtually bare except for a little public entry base. Ovipositor (\mathfrak{P}) not or hardly projecting beyond the apical tergite in the retracted position.

Wing with vein R_s weakly sinuate: *1m-cu* clearly rejected from cell R_s .

Breeding records

Host – Ophiomyia heracleivora Spencer

1 3 from puparium 17. xi. 56 on *Heracleum sphondylium*, Bookham Common, Surrey, England, em. vi. 57, leg. SPENCER (BM). 6 ex. from puparia on *Heracleum sphondylium*, Sunninghill, Berks., England, leg. ALLEN (det. M.W.R. DE V. GRAHAM, recorded by ALLEN, 1956).

This species is easily distinguished from other members of the *bathyzonus* group by its brightly coloured gaster, the conspicuous tuft of matted white pubescence visible in lateral view on the rather angularly produced cheeks, and the sexual dimorphism of its antennal pubescence.

Records of "Dacnusa bathyzona" as a parasite of Ophiomyia simplex LOEW refer to C. rondanii (GIARD) (see under the description of that species above). I have no doubt that the record in TAVARES (1905) also refers to a different species (see the footnote to table 17).

Chorebus leptogaster (HALIDAY), comb. nov.

Alysia (Dacnusa) leptogaster HALIDAY, 1839

Dacnusa leptogaster (HALIDAY), MARSHALL, 1891, 1895 and 1897, NIXON, 1937 and 1944 Dacnusa naenia Morley, 1924

Dacnusa dinae BURGHELE, 1960, syn. nov.

Colour. Palpi yellow-brown, testaceous or brown. Labrum black. Antennae entirely black. Centre of mandibles red-black. Legs 1 and 2 varying from yellow-brown or testaceous to dark brown with the coxae and sometimes the tarsi virtually black: hind legs largely black or dark brown (the coxae always black), but usually with the trochantellus and the base of the tibiae red-brown. Gaster beyond petiole uniformly coloured, varying from reddish yellow to almost black.

Morphology. Back of head largely bare, with pubescence only at its sides (near the mandibles): in lateral view some fairly dense pubescence can be seen above the base of the mandibles (where the surface beneath is distinctly sculptur-

ed), but no distinct tufts are formed. Mandibles (fig. 163) with tooth 1 only slightly expanded. Antennal segments: 3, (29)-30-33; 9, (25)-26-30 (bred material only). Palpi long.

Sides of pronotum densely pubescent along and below the oblique suture, but with a shining, bare or only sparsely pubescent area above this. Mesoscutum with its anterior face and the anterior part of the central lobe roughened, with dense pubescence covering its anterior face and central lobe but the lateral lobes largely bare: notaulices usually indicated anteriorly only (but occasionally complete, reaching the posterior fovea). Precoxal suture distinctly rugosecostate anteriorly. Petiole extraordinarily long and narrow $(3-3\frac{1}{2})$ times as long as wide), parallel-sided, strongly shining and largely bare, with only a few fine hairs at its base and along its sides. Ovipositor (\mathfrak{P}) directed upwards in the retracted position, not or only slightly projecting beyond the apical tergite.

Wing (fig. 148) with the pterostigma rather short and almost parallel-sided: cell $2R_1$ rather short: vein R_s only weakly sinuate: *Im-cu* well rejected from cell R_s .

Breeding records

Host 1 – Ophiomyia pulicaria MEIGEN

4 ex from puparia 5. v. 63 on *Taraxacum* sp. in my garden at Barnet, London, em. 4 to 26. vi. 63 (GCDG). 1 \mathcal{J} from puparium 14. vii. 65 on *Taraxacum officinale*, Rieseninger, Mühlhausen, Thuringia, Germany, em 2. viii. 65, leg BUHR no. 2483 (GCDG). 3 $\mathcal{J}\mathcal{J}$ from puparia 14. v. 65 on *Taraxacum officinale*, Stadtwald, Mühlhausen, Thuringia, em. 2–7. vi. 65, leg BUHR no. 2363 (GCDG). 2 $\mathcal{Q}\mathcal{Q}$ from puparia 13. v. 66, same plant and locality, em. 4 and 9. vi. 66, leg. BUHR no. 2724 (GCDG). 1 \mathcal{J} from puparium 1. vi. 66, same plant and locality, em. 15. vi. 66, leg. BUHR no. 2809 (GCDG). 1 \mathcal{Q} from puparium 20. v. 66 on *Taraxacum officinale*, Ochsenburg, Süd-Kyffhäuser, Thuringia, em. 5. vi. 66, leg. BUHR no. 2751 (GCDG). 1 \mathcal{Q} , Stuttgart-O'türkheim, Germany, em. 8. viii. 55, leg. GROSCHKE (STGT).

Host 2 — Ophiomyia cunctata HENDEL

19 from puparium 14. vii. 65 on Sonchus oleraceus, Rieseninger, Mühlhausen, Thuringia, Germany, em. 28. vii. 65, leg BUHR no. 2465a (GCDG). 1 ex. from puparium 11. vii. 53 on Sonchus asper, Hampstead, London, em 2. viii. 53, leg SPENCER (BM) (host formerly recorded as O. beckeri HENDEL in GRIFFITHS, 1956).

Host 3 — Ophiomyia pinguis FALLÉN

About 30 ex. from puparia in cultivated chicory (*Cichorium intybus*), Zaventem, Belgium, 1964-1966, leg. LOUNSKY (GCDG and Station d'Entomologie, Gembloux).

Other breeding records from undetermined species of the *Ophiomyia pulicaria* group (probably *pulicaria* MEIGEN or *cunctata* HENDEL).

1 $\[mu]$ from puparium 14. ix. 61 on *Taraxacum* sp., Oxwich, Gower, Wales, em. 4. x. 61 (GCDG). 1 $\[mu]$ from *Urospermum capense*, Rostock Botanical Gardens, Mecklenburg, Germany, em. 25. vii. 36, leg. BUHR (GCDG). $\[mu]_{Q}$ from puparia 18. viii. 65 on *Taraxacum officinale*, Stadtwald, Mühlhausen, Thuringia, Germany, em. 28. viii and 17. ix. 65, leg. BUHR no. 2605 (GCDG). 1 $\[mu]$ from puparium 3. viii. 65 on *Lapsana communis*, same locality, em. 22. viii. 65, leg. BUHR no. 2566 (GCDG). 1 $\[mu]$ from puparium 18. v. 66 on *Picris hieracioides*, same locality, em. 7–8. vi. 66, leg. BUHR no. 2738 (GCDG). $\[mu]_{Q}$ from puparia 20. vii. 65 on *Sonchus oleraceus*, Katzentreppen, Mühlhausen, Thuringia, em. 2–5. viii. 65, leg. BUHR no. 2502 (GCDG).

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The synonymy of *Dacnusa naenia* MORLEY was stated in Appendix VII in Part II. Although I have not seen BURGHELE's (1960) material of "*Dacnusa dinae*", her description seems to me to agree fully with *leptogaster* and I have therefore proposed the synonymy of her name.

The record of a host of this species as "*Phytomyza continua* HENDEL" given in NIXON (1944) was doubtless the result of confusion with *Ophiomyia pinguis* FALLÉN. There are no puparia with the series in question (Geneva, Switzerland, 1934, leg. DESHUSSES), which is in the British Museum.

This species is similar to C. femoratus (TOBIAS) and C. xiphidius sp. nov. in respect of its extremely long petiole and the wing venation. The differences between these species are given in the key above.

Chorebus xiphidius sp. nov.

Similar to *C. leptogaster* (HALIDAY), with which it may be compared as follows. Colour. Palpi and labrum clear yellow. Centre of mandibles red-brown. Legs largely deep yellow or ochreous yellow, with the tarsi (especially the hind tarsi) and the apex of the hind tibiae infuscated, and the dorsal surface of the hind femora infuscated at least on their apical half (the coxae however are all yellow or ochreous yellow, not at all infuscated). Gaster beyond petiole conspicuously yellow or yellow-brown.

Morphology. Antennal segments: 3, 32-33; 9, 29-31. Palpi extremely long (see the table of biometric data).

Mesoscutal pubescence extending onto about the anterior half of the lateral lobes. Ovipositor (\mathfrak{P}) much longer, directed more or less horizontally and strongly projecting beyond the apical tergite in the retracted position, its sheaths about as long as the first segment of the hind tarsus.

Host — Ophiomyia sp. (pulicaria group)

1 \bigcirc paratype from puparium 2. v. 66 in leaf of *Picris hieracioides*, Stadtwald, Mühlhausen, Thuringia, Germany, em. 15. v. 66, leg. BUHR no. 2716 (GCDG). Holotype \bigcirc ; 2 \bigcirc 1 \bigcirc paratypes from puparia 13. v. 66, same plant and locality, em. 7–16. vi. 66, leg. BUHR no. 2726 (GCDG). 1 \bigcirc 4 \bigcirc paratypes from puparia 18. v. 66, same plant and locality, em. 7–23. vi. 66, leg. BUHR nos. 2735 and 2738 (GCDG).

This species is clearly monophyletic with C. leptogaster (HALIDAY) and C. femoratus (TOBIAS), being synapomorph in respect of the extremely elongate petiole and wing venation (fig. 148). The form of the petiole in conjunction with pale coloration and a projecting ovipositor in the female will enable the species to be easily recognised.

The host is an apparently undescribed species of the *Ophiomyia pulicaria* group whose puparia differ from those of the known species in having smaller spiracles (with only 6-7 bulbs on the hind spiracles).

Chorebus glaber (NIXON), comb. nov.

Dacnusa leptogaster (HALIDAY) sensu VAN DEN BRUEL, 1933 (nec Alysia (Dacnusa) leptogaster HALIDAY, 1839)

Dacnusa glabra NIXON, 1944

Colour. Palpi dark brown or black. Labrum black. Antennae entirely dark (except sometimes the annellus). Centre of mandibles red or red-brown. Legs with all coxae, trochanters and usually the trochantelli black: femora and tibiae of legs 1 and 2 largely yellow-brown or brown, at most with the dorsal surface of the femora infuscated; hind legs with black femora, but the tibiae are largely yellow-brown or red-brown, usually becoming black only near their apex: all tarsi infuscated. Gaster entirely black.

Morphology. Back of head bare centrally, but densely pubescent at its sides (near the mandibles): in lateral view the pubescence on the posterior edge of the head above the base of the mandibles appears to form tufts, but these are sometimes grey and rather inconspicuous. Mandibles very variable in size, sometimes large with tooth 1 strongly expanded (fig. 156), but often much smaller, not expanded (fig. 157). Antennal segments: 3, (29) - 30 - 32 - (33); 9, 25 - 29.

Sides of pronotum entirely covered with dense pubescence. Mesoscutum largely smooth, with only its anterior face punctate, with pubescence covering its anterior face and central lobe but the lateral lobes largely bare: notaulices weak, not extending longitudinally on the dorsal surface of the mesoscutum. Precoxal suture rugose-costate anteriorly. Petiole 2.0-2.6 times as long as wide, almost or completely parallel-sided, strongly shining, with its dorsal surface largely bare (except for some fine pubescence near its base and sometimes a few hairs along its sides). Ovipositor (\mathcal{Q}) stout, shortly projecting beyond the apical tergite in the retracted position (by up to a half of the length of the petiole).

Wing (fig. 149) with cell $2R_1$ relatively short; pterostigma broad, distinctly tapering towards its apex; vein R_s only weakly sinuate; Im-cu widely rejected from cell R_s .

Breeding records

Host — Napomyza cichorii Spencer

This species is the most numerous parasite of this host, which is a serious pest of cultivated chicory (Cichorium intybus) in Belgium and Holland. About 3,000 specimens have been bred by Mr. J. LOUNSKY of the Station d'Entomologie at Gembloux from material collected at Zaventem, Belgium (1963-1966). I have also received material from Holland (Isle of Walcheren and Isle of Overflakee, spring 1966) sent by Drs. VAN 'T SANT (leg. BETHE) of the Instituut voor Plantenziektenkundig Onderzoek at Wageningen. A sample of the above material is retained in my personal collection.

VAN DEN BRUEL'S (1933) figure of "Dacnusa leptogaster HAL." clearly refers to this species, but it is likely that he also had the true Chorebus leptogaster (HALIDAY) before him, since this is a parasite of Ophiomyia pinguis FALLÉN which was included in some of his breeding samples. The species has also been referred to as "Dacnusa gracilis NEES" in VAN 'T SANT, VIJZELMAN and BETHE (1961).

There are many other references to parasites of this host in the considerable literature concerning the control of chicory flies. It seems reasonable to assume that all such references to "Dacnusa" parasites of Napomyza on chicory refer wholly or for the most part to the present species. But it must be borne in mind when interpreting the results of experiments in which both Napomyza and Ophiomyia pinguis FALLÉN were present, that workers in this field have hitherto failed to distinguish glaber from the true leptogaster, which is a parasite of the Ophiomyia.

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There is an unusually wide range of variation in the size of the head and mandibles in this species (compare nos. 19-20 with nos. 21-22 in the table of biometric data and fig. 156 with fig. 157). Specimens with a large head and mandibles were bred by Mr. LOUNSKY mainly from puparia in the roots of chicory, while the majority of specimens bred from puparia in leaves had a relatively smaller head and mandibles. However this correlation is only partial and many intermediate specimens occur.

There is a specimen of *glaber* in the HALIDAY collection labelled "navicularis" (possibly doubtfully as the record was not published) by HALIDAY, but the species does not fit NEES' (1834) description of Alysia navicularis, which is stated to have a public public. NIXON'S (1944) holotype is a specimen with strongly expanded mandibles.

This species is well characterised in the female by its projecting ovipositor. The male might easily be confused with C. *leptogaster* (HALIDAY), but has a shorter petiole and the sides of the pronotum entirely covered with dense pubescence. Another dark-legged species which might be confused with *glaber* is described in this paper as C. *orbiculatae* sp. nov. (the differences are given in the key above).

Chorebus gedanensis (RATZEBURG), comb. nov.

Alysia Gedanensis RATZEBURG, 1852 Dacnusa Gedanensis (RATZEBURG), MARSHALL, 1891 Dacnusa anguligena NIXON, 1937 and 1944, syn. nov.

Colour. Palpi and labrum yellow-brown. Antennae entirely dark or becoming brownish towards their base. Centre of mandibles red-brown. Legs largely reddish yellow or yellow-brown, the hind pair being slightly darker than legs 1 and 2: coxae more or less brown. Gaster beyond petiole red-brown or brown, becoming darker towards its apex.

Morphology. Back of head clothed with fairly dense pubescence: in lateral view the cheeks are produced backwards above the mandibles to form an angulate projection, which is rather densely pubescent. Mandibles not expanded, hollowed posteriorly near their base. Antennal segments: 3, 31-33; 9, 30-33.

Mesoscutum hardly sculptured, with fine pubescence covering much of its surface but absent from a large part of the lateral lobes: notaulices well-developed anteriorly, reaching the posterior fovea as smooth (sometimes very weak) impressions. Precoxal suture rugose-costate anteriorly. Sculpture of metapleuron and propodeum very coarse. Petiole about 2.2 times as long as wide, parallel-sided, with its dorsal surface almost bare and strongly shining. Ovipositor (\mathcal{Q}) not projecting beyond the apical tergite in the retracted position.

*Wing with cell $2R_1$ rather elongate: vein R_s weakly sinuate: pterostigma rather broad: 1m-cu well rejected from cell R_s .

Breeding records

Host — Hexomyza schineri GIRAUD

33329 from galls on *Populus tremula*, Hoddesdon, Herts., England, em. v-vi, leg. EAGLES and NIBLETT (BM) (the type series of *Dacnusa anguligena* NIXON). RATZEBURG (1852) refers to 2 ex. bred by BRISCHKE from galls on *Populus tremula*, em. 2 and 17. vi. 1849 (locality not stated but clearly Gdansk (Danzig) from the specific name), and further material bred by REISSIG (? locality), also from galls on *Populus tremula*.

RATZEBURG (1852) gives the host of this species as Saperda populnea (L.) (Coleoptera) or possibly "gleichzeitig mit ihm lebenden Dipteren". Galls of *Hexomyza* on Salicaceae can easily be confused on external appearance with coleopterous galls and clearly this is the explanation of the record. RATZEBURG's specimens have been destroyed, but I think there can be no reasonable doubt from his description that the species before him was the same as NIXON's *Dacnusa anguligena*. For instance his reference to the number of antennal segments, the extensive pubescence and the shape of the petiole all strongly support this interpretation. GIRAUD (1861), in his original description of the host fly bred from galls on *Populus alba* on the banks of the Danube (presumably near Vienna), states that he obtained a parasite "*Dachnusa agromyzae* m. n. sp.", but this is a nomen nudum since no description is given and does not preoccupy the use of the name *Dacnusa agromyzae* GAHAN for an American species.

Little difficulty will arise in identifying bred material of this species on account of the characteristic life-history. The shape of the head is perhaps its most distinctive morphological feature (see NIXON, 1944, fig. 81).

The Chorebus cybele group

The concept of the cybele group, first proposed by NIXON (1943 and 1944) for the two monophyletic species cybele and didas, is here extended to include three further species. I consider the species included in this group synapomorph in respect of the reduction of tooth 4 of the mandibles, their elongate form, and possibly also the upcurved ovipositor. They differ from the other group treated in this paper, the senilis group, in possessing at most fine inconspicuous pubescence on the sides of the pronotum (a plesiomorph character), in lacking a distinct rosette of metapleural pubescence (plesiomorph) (compare fig. 21), and in having vein Cu_{1b} weak or absent (fig. 155), so that cell 2Cu is more or less open at its lower distal corner (apomorph).

The inclusion in this group of C. cyclops (NIXON)⁹, a species of unknown lifehistory, is provisional. Its metapleural and propodeal pubescence are similar to those of the other species, and its mandibles may be derived from the type found in other species (through reduction of tooth 3 as well as tooth 4, as has also occurred in C. *ibericus* sp. nov.). However cyclops has an almost bare mesoscutum and a subcubical head. On the basis mainly of the last two characters NIXON (1943 and 1946) associated it with C. diremtus (NEES), a parasite of Cerodontha s. s., as the diremta group of "Dacnusa". That species however appears referable to

⁹ See also the table of biometric data.

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the *ovalis*/*lateralis* complex, as it has a well-defined metapleural rosette. I think therefore that the undoubted similarity of these two species in the shape of the head represents convergence.

The species which I include in the *cybele* group may be distinguished by the following key.

Key to the Chorebus cybele group

- 3 Mesoscutal pubescence almost confined to the former course of the notaulices. Head subcubical (about 1.3 times as wide as long). Antennal segments: ♂, 27-30; ♀, (22)-23-25. Mandibles with tooth 2 long and pointed but teeth 3 and 4 reduced. Petiole elongate, about twice as long as wide. . . C. cyclops (NIXON), comb. nov.
 Mesoscutum with at least its central lobe densely pubescent. Head more transverse

C. parvungulus (Thomson) (= acco Nixon)

Host: Napomyza lateralis FALLÉN and N. cichorii SPENCER

- Petiole more elongate, about 1.7 times as long as wide. 18 antennal segments $(2 \varphi \varphi)$. Mandibles with both teeth 3 and 4 small and indistinct (fig. 166). *C. ibericus* sp. nov. Host: *Ophiomyia beckeri* HENDEL

In the descriptions which follow the following common characters may be assumed.

Clypeus dark, like the face. Sides of pronotum shining, with only fine inconspicuous pubescence mainly below the oblique suture. Propodeal pubescence dense, usually rather short. Petiole more or less parallel-sided. Hind coxa not unusually densely pubescent, without any distinct tufts. Hind tarsus about as long as the hind tibia.

Wing (compare fig. 153) with cell $2R_1$ somewhat narrow and vein R_s not or only weakly sinuate: vein 2r branching rather remote from the base of the pterostigma: *Im-cu* clearly rejected from cell R_s : cell 2Cu (fig. 155) more or less open at its lower distal corner (Cu_{1b} weak or absent).

Chorebus cybele (NIXON), comb. nov.

Dacnusa cybele NIXON, 1937 and 1944

Colour. Palpi yellow or reddish yellow. Labrum orange or yellow-brown. Centre of mandibles red-brown. Antennae with the scape and pedicel redbrown, the flagellum usually entirely dark, but sometimes with its basal segments more or less red-brown. Legs largely golden yellow or reddish yellow with the hind tarsi and usually the base of the hind coxae and apex of the hind tibiae infuscated: occasionally the hind femora and tibiae are more or less brown.¹⁰ Gaster usually largely dark except for the yellow-brown or reddish base of tergite 3: occasionally this paler colour is more extensive.

Morphology. Antennal segments: 3, (34)-35-40-(42); 9, (32)-33-37-(38). Mandibles 4-toothed, not expanded, with tooth 4 reduced, obviously smaller than the large tooth 3 (fig. 165). Face shining, with only shallow punctate sculpture, covered with fairly dense fine pubescence which is directed downwards along the eye-margins but mostly inwards towards its centre, becoming sparse or absent along the centre-line. Palpi long.

Thorax extremely narrow and elongate (2.3-2.4 times as long as wide). Mesoscutum shining, with its anterior face and central lobe weakly punctate, with dense pubescence covering its anterior face, central lobe and about the anterior half of the lateral lobes, but the posterior half of the lateral lobes is often bare: posterior fovealong and deep, extending almost from the middle of the mesoscutum: notaulices with only their lateral extensions distinctly rugose, either extending to the posterior fovea as narrow V-shaped more or less smooth grooves or virtually absent from the dorsal surface of the mesoscutum. Mesepisternum with a long, fairly narrow, but distinctly rugose precoxal suture extending almost from the epicnemial suture to its posterior margin. Metapleural pubescence (fig. 21) not forming a distinct rosette. Petiole 1.7-1.9 times as long as wide, with fairly dense pubescence covering almost its entire surface (but occasionally tending to be absent from the centre-line posteriorly), often slightly denser at the apical corners, but no tufts are formed. Tergite 3 with few or no basal hairs. Ovipositor (\mathcal{Q}) long and upcurved, conspicuously projecting beyond the apical tergite (by a third to a half of the length of the petiole) in the retracted position.

Large species (wing length up to 4 mm.).

Breeding records

Host 1 — Melanagromyza lappae LOEW

2 ex. from puparia in stems of Arctium sp., Scratch Wood, London, em. 10. v. 56, leg. SPENCER (GCDG). 8 ex. from puparia in stems of Arctium lappa, Mühlhausen, Thuringia, Germany, em. 14. iv $(1\,\text{Q})$ and 7. v. 55, leg. BUHR, HERING nos. 867 and 870 (GCDG).

¹⁰ One of the two males bred from the *Melanagromyza* sp. in stems of *Chaerophyllum aureum* at Mühlhausen is remarkable in having dark palpi, the hind legs completely infuscated and the middle legs largely so. This insect must clearly be considered a variant, as the other male bred with it is normally coloured. (There is no question of confusion with *didas*, because that species has fewer antennal segments.)

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 $1 \notin 7$ QQ (including the holotype Q) from puparia in stems of Arctium sp., Bagley Wood, Berks., England, em. 24. v-3. vi. 32, leg. HAMM (HD and BM). 3 ex. from puparia in stems of Arctium vulgare, North Mimms, Herts., England, em. 12-19. v. 62 (GCDG).

Host 2 — Melanagromyza eupatorii Spencer

 $1 \, \bigcirc$ from puparium 22. i. 61 in stem of *Eupatorium cannabinum*, Woodwalton Fen, Hunts., England, em. 12. iii. 61 (GCDG). $1 \, \bigcirc$ from puparium 18. iii. 23 in stem of *Eupatorium cannabinum*, Cothill, Berks., England, em. 3. v. 23, leg. WATERS (HD). $1 \, \bigcirc$ from puparium ix. 58 in stem of *Eupatorium cannabinum*, Chippenham Fen, Cambs., England, em. spring 59, leg. SPENCER (GCDG). $1 \, \bigcirc$ from puparium in stem of *Eupatorium cannabinum*, Heidelberg, Germany, em. 5. v. 56, leg. SPENCER (GCDG).

Host 3 — Melanagromyza sp.

19 from puparium 18. ix. 64 in stem of *Chaerophyllum aureum*, Jena-Wöllnitz, Thuringia, Germany, em. 10. iii. 65, leg. BUHR (GCDG). 233 from puparia in stems of *Chaerophyllum aureum*, Mühlhausen, Thuringia, em. spring 66, leg. BUHR (GCDG).

Host 4 — Melanagromyza symphyti GRIFFITHS

48 ex. from larvae and puparia 17. ix. 61 in stems and leaf-stalks of Symphytum officinale, Woodwalton Fen, Hunts., England, em. 26. iv-25. vi. 62 (32 ex.) and 4-21. v. 63 (16 ex.) (GCDG).

The records for Woodwalton Fen were previously published in GRIFFITHS (1963a and 1936b). It is noteworthy that some specimens from the series bred from M. symphyti GRIFFITHS at that locality did not emerge until they had passed two winters in the host puparia. Host 3 is probably an undescribed species.

This species will be readily distinguished from the other species of this group treated in this paper by its large size, longer ovipositor and usually yellow legs.

Chorebus didas (NIXON), comb. nov.

Dacnusa didas NIXON, 1944

Similar to *cybele*, with which it may be compared as follows.

Colour darker. Palpi yellow-brown to almost black. Labrum brown. Antennae more or less entirely dark. Legs 1 and 2 largely brown or light brown: hind legs darker, varying from almost completely black to brown with only the coxae more or less black. Gaster entirely dark.

Morphology. Antennal segments: $\mathcal{J}, 33-35; \mathcal{Q}, 28-31$. Centre of mesepisternum bearing a band of fine hairs. Petiole more elongate, 2.1-2.3 times as long as wide. Ovipositor (\mathcal{Q}) shorter than in most specimens of *cybele*, not so strongly upcurved.

Smaller species (wing length not exceeding 3 mm.).

Breeding records

Host — Napomyza scrophulariae Spencer

4 dd from puparia in *Digitalis* sp., Hampstead, London, em. 10-23. iii. 66, leg. SPENCER (GCDG). Numerous specimens from seed capsules of *Digitalis* sp., Slough (Bucks.), Abinger (Surrey) and Wallington (Surrey), England 1951 (see WOODROFFE and SOUTHGATE, 1952). 1 \circ from puparium 27. ix. 64 in stem of *Mentha spicata* (garden mint), East Barnet, London, em. 7. v. 65 (GCDG).

In addition I have received one female labelled as bred from a puparium 25. iii. 56 of *Melanagromyza nibletti* SPENCER in stem of *Silaum silaus*, Bookham, Surrey, England, em. 20. iv. 56, leg. SPENCER (GCDG). I regard this record as requiring confirmation, since it is in conflict with the other information on the host association of this species and the host puparium was not preserved.

This species may be distinguished from the other dark-legged species which I have included in the *cybele* group by its extremely long and narrow thorax (as in *cybele*), distinctly projecting ovipositor and elongate densely public entipetiole.

Chorebus parcungulus (THOMSON), comb.nov.

Dacnusa (Dacnusa) parvungula THOMSON, 1895 Dacnusa acco Nixon, 1943 and 1946, syn. nov.

Colour. Palpi infuscated, more or less dark brown. Labrum brown or blackish. Centre of mandibles brown or red-brown. Antennae more or less entirely dark. Legs very dark, with at least the coxae and tarsi dark brown or virtually black; but usually the femora and tibiae of legs 1 and 2 and the hind tibiae are somewhat paler, more or less brown. Gaster with tergite 3 reddish or more or less entirely dark.

Morphology. Antennal segments: \mathcal{J} , (26) -27-29; \mathcal{Q} , 23-26. Face virtually smooth. Mandibles (compare fig. 165) with tooth 1 slightly expanded and tooth 3 relatively large (only tooth 4 markedly reduced).

Thorax not so narrow as in cybele and didas (about 2.0-2.1 times as long as wide). Mesoscutum largely smooth and shining with its anterior face densely pubescent and rather long pubescence covering its central lobe and extending onto the anterior part of the lateral lobes, although these are largely bare: notaulices hardly extending longitudinally on the dorsal surface of the mesoscutum. Precoxal suture well developed anteriorly but often not extending to the hind margin of the mesepisternum: centre of mesepisternum bearing a band of fine pubescence. Metapleural pubescence similar to that of cybele and didas (compare fig. 21). Petiole 1.3-1.6 times as long as wide, strongly shining with short inconspicuous pubescence at its sides but bare centrally: the base of tergite 3 also bears some fine pubescence. Ovipositor (\mathfrak{Q}) short and stout, slightly upcurved, hardly projecting beyond the apical tergite in the retracted position.

Breeding records

Host 1 — Napomyza lateralis FALLÉN

299 from puparia 19. viii. 35 in flowerheads of *Matricaria maritima inodora*, Schorrentin, Mecklenburg, Germany, em. 30. viii. 35, leg. BUHR no. 69 (GCDG). 1 3 from puparium in *Matricaria chamomilla*, Lüsewitz, Mecklenburg, em. 20. ix. 53, leg. BUHR no. 590 (BM). 299 from flowerheads of *Chrysanthemum leucanthemum*, 1953, Rothamsted Experimental Station, Herts., England (BM). 12 ex. from puparia in *Matricaria chamomilla*, Zaventem, Belgium, 1964/65, leg. LOUNSKY (GCDG and Station d'Entomologie, Gembloux).

Host 2 — Napomyza cichorii Spencer

 $1 \bigcirc$ from puparium 25. iii. 66 in *Cichorium intybus*, Isle of Overflakee, Holland, em. 19. iv. 66, leg. Bethe (GCDG).

This species may be readily distinguished from the other dark-legged species of the *cybele* group by the form and pubescence of its petiole.

NIXON (1946) considered that C. thusa (NIXON) (a parasite of Phytomyza rufipes MEIGEN, see Part III) was "very closely related" to this species (his acco). But thusa has a large mandible, very different from that of the species which I include in the cybele group. Its similarity to parvungulus in respect of its coloration, number of antennal segments and reduction of the notaulices does not seem to me firm evidence of synapomorphy, as agreement in these characters is clearly often the result of convergence in the Dacnusini. But although I do not consider that thusa can be the sister-species of parvungulus, it is possible that there is a close relationship between the cybele group and thusa (which also lacks a metapleural rosette). The question may be clarified when further information on the parasites of other stem-boring species of Phytomyza and Napo-myza is available.

THOMSON'S (1895) original description refers to both sexes from Örtofta, near Lund, Sweden. Two specimens from this locality, one of each sex, were sent to me for examination. The description is clearly based on the female, which is hereby designated lectotype. The male is a faded example of a species of the *senilis* group sensu stricto.

Chorebus ibericus sp. nov.

Similar to parvungulus, with which it may be compared as follows.

Morphology. 18 antennal segments $(2 \Im \Im)$. Facial pubescence rather coarse. Mandibles small, with tooth 2 relatively large and pointed but teeth 3 and 4 small and indistinct (fig. 166). Mesoscutal pubescence more extensive, only part of the posterior half of the lateral lobes bare. Petiole more elongate, about 1.7 times as long as wide. Ovipositor (\Im) distinctly upcurved, shortly projecting beyond the apical tergite in the retracted position.

Size very small (wing length about 1.9 mm.).

Host — Ophiomyia beckeri HENDEL

Holotype Q, paratype Q from puparia 24. iv. 55 on an unidentified species of Compositae, Algeciras, Spain, em. v. 55, leg. SPENCER (GCDG).

This species should be readily distinguished from *parvungulus* by its fewer antennal segments and more elongate petiole. Its somewhat modified mandibles (fig. 166) might cause doubt about its belonging to the *cybele* group, and it is important for purposes of identification that the pubescence of its metapleuron and petiole should be carefully checked. Its mandibles are similar to those of *C. cyclops* (NIXON), but that species has much less mesoscutal pubescence, a subcubical head and more numerous antennal segments.

Keys to the Dacnusini Parasites of particular Host Genera

Keys are here given to the Dacnusini parasites of *Melanagromyza*, *Ophiomyia* and *Napomyza*. No key to the parasites of *Hexomyza* can be given, since only a single species, *Chorebus gedanensis* (RATZEBURG), was available to me for study.

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1. Melanagromyza spp.

2. Ophiomyia spp.

	Metapleural pubescence very dense, forming a well-defined rosette around the rugose	
	swelling (compare figs. $22-23$). Sides of pronotum densely pubescent, at least along	
	and below the oblique suture. Mandibles not so	2
2	Tooth 1 of mandibles strongly expanded but teeth 3 and 4 relatively small (fig. 164).	
	Precoxal suture rugose, at least anteriorly	3
	Tooth 1 of mandibles not strongly expanded (figs. 162 and 163)	4
3	Gaster beyond petiole bright yellow. Legs 1 and 2 deep yellow, but the hind legs	
	contrastingly darker, with the femora, tibiae and tarsi uniformly dark brown.	
	Petiole $2.0-2.2$ times as long as wide, slightly widened towards its apex. Back of	
	head almost bare centrally Chorebus rondanii (GIAR	≀D)
	Host: O. simplex LOBW	

- 4 Pterostigma and cell $2R_1$ relatively short (fig. 148). Petiole extraordinarily long and narrow, 3-31/2 times as long as wide. Back of head virtually bare centrally, pubescent only at its sides (near the mandibles). Precoxal suture rugose-costate anteriorly. Lateral lobes of mesoscutum bare $\mathbf{5}$ 6 5 Coxae yellow. Gaster beyond petiole conspicuously yellow or yellow-brown. Ovipositor (Q) strongly projecting beyond the apical tergite in the retracted position Chorebus xiphidius sp. nov. Host: O. sp. on Picris - Coxae black. Gaster beyond petiole varying from reddish yellow to black. Ovipositor (9) not or only slightly projecting beyond the apical tergite in the retracted Hosts: O. cunctata HENDEL, O. pulicaria MEIGEN and O. pinguis FALLÉN 6 Mesoscutum largely bare (with some fine pubescence on its anterior face and a few hairs along the former course of the notaulices: central lobe either bare or with a little fine scattered pubescence). Back of head bare centrally, pubescent only at its sides (near the mandibles). Precoxal suture narrow, but weakly rugose-costate ant-. Chorebus heringianus sp. nov. eriorly Host: O. thalictricaulis HERING - Mesoscutum with its anterior face and central lobe densely publication. Back of head densely pubescent. Precoxal suture visible as an almost smooth well-defined linear 7 7 23-24 antennal segments (Q). Mesoscutum entirely covered with extremely dense whitish pubescence which on the lateral lobes is largely directed laterally. Petiole largely bare (fig. 168) Chorebus caesariatus sp. nov. Host: O. sp. on Medicago - Antennal segments more numerous 8 8 Cheeks in lateral view angularly produced (NIXON, 1944, fig. 78), bearing a conspicuous tuft of white pubescence above the base of the mandibles. In the male the segments of at least the basal third of the flagellum are very shining and, at least on their dorsal surface, virtually bare. Gaster beyond petiole reddish yellow. Legs largely reddish yellow. Chorebus bathyzonus (MARSHALL) Host: O. heracleivera SPENCER - Cheeks not thus produced; tuft of pubescence not so conspicuous. Flagellum 9 Mandibles (fig. 162) hollowed and somewhat dilated posteriorly near their base. 9 Lateral lobes of mesoscutum pubescent. Petiole somewhat densely pubescent on about its basal half (fig. 167) Chorebus fuscipennis (NIXON) Hosts: O. heringi STARY, O. labiatarum HERING and O. sp. on Stachys palustris - Mandibles hollowed posteriorly near their base, but not or hardly dilated. Lateral lobes of mesoscutum largely bare. Petiole largely bare. (Legs 1 and 2 ochreous yellow: hind legs uniformly brown except for the yellowish trochanter and trochantellus – contrast C. nerissa (NIXON)) Chorebus lychnidis sp. nov. Host: O. sp. on Lychnis 3. Napomyza spp.
 - 1 Metapleural pubescence long and dense, directed mainly downwards towards the hind coxa. Petiole subtriangular with dense pubescence similar to that of the meta-

- Petiole more or less parallel-sided. Rugose precoxal suture present. Pterostigma shorter, not blackened in the male. Mandibles 4-toothed, but differently shaped. Legs darker, with at least the hind coxae and femora infuscated.

- Thorax roughly 2.0-2.1 times as long as wide. Petiole broad, 1.3-1.6 times as long as wide, with short inconspicuous pubescence at its sides but bare centrally. Antennal segments: \mathcal{J} , (26)-27-29; \mathcal{Q} , 23-26. Ovipositor (\mathcal{Q}) hardly projecting beyond the apical tergite in the retracted position. . . Chorebus parvungulus (THOMSON) Hosts: N. lateralis FALLÉN and N. cichorii SPENCER
- 4 Petiole with its entire surface very densely pubescent (fig. 170). Back of head entirely covered with very dense pubescence Chorebus senilis (NEES) Hosts: N. lateralis FALLÉN, N. cichorii SPENCER and N. scrophulariae SPENCER (and also Melanagromyza aeneoventris FALLÉN)

Key to the Dacnusini Parasites of Chicory Flies

Since experimental work on the Agromyzid pests of chicory (Cichorium intybus) is often undertaken using mixed infestations of flies of different genera, it appears useful to include in this paper a key to all parasites which have been obtained from chicory flies. The material I have seen has been obtained from two species, Napomyza cichorii SPENCER and Ophiomyja pinguis FALLÉN.

- 1 Pterostigma very elongate (fig. 128 in Part III), blackened in the male. Precoxal suture absent. Metapleuron evenly covered with long, dense pubescence (which does not form a rosette). Petiole subtriangular with dense pubescence similar to that of the propodeum and metapleuron. Legs largely yellow (but with the base of the hind coxae infuscated) Dacnusa pubescens (CURTIS) Occasionally on Napomyza cichorii SPENCEE (also on other Napomyza spp. and certain Phytomyza spp.)
- Pterostigma shorter, not blackened in the male. Rugose precoxal suture present. Petiole more or less parallel-sided. Legs darker, with at least the hind coxae and femora infuscated

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- Metapleural pubescence forming a well-defined rosette (compare figs. 22-23). Sides of pronotum with conspicuous dense matted pubescence at least along and below the oblique suture. Petiole over twice as long as wide
- 4 Petiole 2.0–2.6 times as long as wide. Sides of pronotum entirely covered with dense pubescence. Ovipositor (\mathcal{Q}) long, distinctly projecting beyond the apical tergite in the retracted position (by up to a half of the length of the petiole). Pubescence on back of head near the base of the mandibles very dense. . . . Chorebus glaber (NIXON) The most common parasite of Napomyza cichorii SPENCER (known only from this host)

Host Association

Table 17 below lists the known associations of Dacnusini with the host genera treated in this paper.

The majority of the *Chorebus* spp. treated in this paper exhibit, as far as known, monophagy of the first or second degree. This conforms with the pattern of host association shown by the *Chorebus* parasites of the other genera of Agromyzidae treated in parts II and III of this paper. One species, *C. senilis* (NEES), exhibits disjunctive monophagy. The host association of other members of the *senilis* group suggests that the original association of this species was with *Melanagromyza* and that its association also with *Napomyza* spp. is secondary.

The host range of *Dacnusa pubescens* (CURTIS), associated with a number of *Phytomyza* species as well as *Napomyza*, is disjunctive and appears determined partly by the location of the host larvae — the known hosts are all stem-borers or feed in the midrib of the leaf with the exception of *Phytomyza atricornis* MEIGEN, which however does not seem to be a normal host.

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Table 17

Hosts	Dacnusa	Chorebus senilis group s. l.	Chorebus cybele group
I. Hexomyza	_* *	C. 47	-
schineri		gedanensis	
kiefferi		sp.11	
II. Melanagromyza			
aeneoventris		senilis, brevicornis	1
lappae			cybere
eupatorii			cybele
symphyti		2	cybele
sp. (Chaerophyllum aureum)			cybele
III. Ophiomyia			
orbiculata		orbiculatae	
simplex		rondanii	
beckeri			ibericus
sp. (Picris)		xiphidius	
cunctata		leptogaster	
pulicaria		leptogaster	1
vinguis		leptogaster	
thalictricaulis		heringianus	
heringi		fuscipennis	
abiatarum		fuscipennis	
sp. (Stachys palustris)		fuscipennis	5. S.
sp. (Medicago)		caesariatus	
sp. (Lychnis)		lychnidis	
heracleivora		bathyzonus	
IV. Napomyza			1
lateralis	pubescens	senilis	parvungulus
scrophulariae		senilis	didas
carotae	pubescens		
cichorii	pubescens	glaber, senilis	parrungulus

List of Records of Dacnusini Parasites of *Hexomyza*, *Melanagromyza*, *Ophiomyia* and *Napomyza*

¹¹ TAVARES (1905) records "Dacnusa bathyzona MARSHALL" as a parasite of his "Agromyza kiefferi", bred from galls on Cylisus. The host is doubtless a species of Hexomyza, probably a prior name for H. sarothamni HENDEL. Unfortunately I have not been able to obtain any material bred from this host. The identification of the parasite as bathyzona carries no authority, as the Dacnusini were very poorly known in 1905.

Summary

1. This paper, the fourth of a series, deals with the Dacnusini (Alysiinae) parasites of *Hexomyza* ENDERLEIN, *Melanagromyza* HENDEL, *Ophiomyia* BRASCHNIKOV (Agromyzinae) and *Napomyza* WESTWOOD (Phytomyzinae). Except for a single species of *Dacnusa* the parasites all belong to two disjunct groups of the genus *Chorebus*, the *C. senilis* group s.l. and the *C. cybele* group, which are both redefined. —

2. Revised keys are given to the European species of the two *Chorebus* groups treated. Keys are also given to the parasites of three of the host genera, and to the parasites of those flies associated with chicory (*Cichorium intybus*). -

3. Except for the single *Dacnusa* species (which also attacks certain *Phytomyza* spp.), all parasites treated in this paper exhibit a high degree of host specificity. A complete host/parasite list for Europe has been prepared, including revision of previous records. -

4. Seven new species are described, six in the *Chorebus senilis* group s.l. and one in the *Chorebus cybele* group.

Zusammenfassung

1. Dieser Artikel, der vierte einer Reihe, behandelt die Dacnusini-(Alysiinae)-Parasiten von *Hexomyza* ENDERLEIN, *Melanagromyza* HENDEL, *Ophiomyia* BRASCHNIKOV (Agromyzinae) und *Napomyza* WESTWOOD (Phytomyzinae). Außer einer einzigen Art von *Dacnusa* gehören diese Parasiten alle zu zwei getrennten Gruppen der Gattung *Chorebus*, der *C. senilis* Gruppe s.l. und der *C. cybele* Gruppe, die beide neu bestimmt werden. —

2. Es werden revidierte Bestimmungstabellen der europäischen Arten der beiden behandelten Chorebus-Gruppen mitgeteilt. Ferner werden Bestimmungstabellen angegeben für die Parasiten von drei der Wirtsgattungen und für die Parasiten der mit der Zichorie (Cichorium intybus) verbundenen Fliegen. -

3. Außer der einzigen Dacnusa-Art (die auch gewisse Phytomyza-Arten befällt) weisen alle in dieser Arbeit behandelten Parasiten einen hohen Grad von Wirtsspezifik auf. Es wurde eine vollständige Liste von Wirten und Parasiten für Europa aufgestellt, die die Revision früherer Berichte einschließt. —

4. Sieben neue Arten werden beschrieben, sechs in der *Chorebus senilis* Gruppe s.l. und eine in der *Chorebus cybele* Gruppe.

Резюме

Эта статья, четвёртая по очерели, занимается с паразитами Dacnusini (Alysiinae) родов *Hexomyza* Enderlein, *Melanagromyza* Hendel, *Ophiomyia* Braschnikov (Agromyzinae) и *Napomyza* Westwood (Phytomyzinae). Кроме одного вида *Dacnusa* все паразиты принадлежат двум отдельным гуппам рода *Chorebus*, группе *C. senilis* s.l. и группе *C. cybele*, которые оба определяются по новому. —

2. Даются ревидированные определительные таблицы европейских видов обоих групп *Chorebus*. Далее приводятся определительные таблицы для паразитов трёх родов хозяинов и для паразитов тех мух, которые связанны с *Cichorium intybus*. —

3. Кроме одного вида *Dacnusa* (которая паразитирует и некоторые виды *Phytomyza*) выявляют все эдесь обработанные паразиты высокую специфику к хозяинам. Составляется полный список всех хозяинов и празитов для Европы, который включает ревизию раньших данныхч. —

4. Семь новых видов описываются, шесть из группы Chorebus senilis s.l. и один из группы Chorebus cybele.

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Tables of Biometric Data

Table 16 Biometric Data

		Absolute Measurements $(1 = 0.01 \text{ mm.})$																								
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
	3		Head	L	Eye-width (lateral)	Eyes	Width	es Width		nten: gme		Ma		ry Pa meni	lpus ts	T	'hora	x	H	ind I	Jeg			id Ti gme		
		Width	Length	Height	Eye-wid	Distance between Eyes	Clypeus Width	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5
1 2	% ♀	72 67	43 11	56 56	22 18	37 34	23 25	16 13	19 20	$15 \\ 15$	13 14	13 12	16 17	11 12	$12 \\ 13$	111 111	51 52	69 71	67 74	106 109	108 115	44 48	22 24	16 17	10 11	13 15
3 4	¢ ∿	65 60	37 37	50 48	21 19	34 34	22 22	13 11	20 21	17 16	14 14	11 11	17 16	11 9	11 9	$\begin{array}{c} 102 \\ 95 \end{array}$	46 44	61 67	65 58	100 89	102 —	41 41	22 21	15 14	10 —	14
5 6	¢ ℃	76 65	41 41	61 61	21 17	37 35	23 —	12 11	13 15	13 12	12 12	11 9	15 13	11 9	11 9	$\frac{122}{113}$	54 —	80 71	69 67	102 93	96 89	40 37	21 17	14 13	10 9	$\frac{14}{12}$
7 8	¢ 6	65 71	41 46	51 52	19 19	34 34	21 22	15 14	11 13	11 12	10 12	7 8	10 12	7 9	7 9	98 104	52 54	58 58	56 58	83 87	74	32 35	15 17	9 11	7	11
9 10	♀ ♀	66 65	35 39	50 49	15 21	34 33	22	17 16	12 13	12 12	$\frac{11}{12}$	- 8	 12	- 9	- 9	87 89	48 48	61 63	56 57	83 87	76 80	30 34	15 17	11 11	8 8	10 11
11 12	¢ ℃	62 58	33 31	51 43	19 15	31 27	21	13 11	12 13	12 11	11 11	8	11 13	9 8	9 7	100 91	60 49	63 52	56 48	84 74	76 73	30 29	17 15	11 10	7 7	11 10
13 14	₽ ₽	50 44	31 26	43 39	19 15	25 21	21 17	10 9	12 11	12 10	12 11	7 6	10 9	777	777	80 67	46 38	43 41	44 41	69 61	68 62	$\frac{26}{25}$	15 13	10 9	7 6	10 9
15	ð	50	26	41	13	25	18	10	12	12	11	7	12	9	9	74	40	47	44	69	67	27	13	9	7	9
16	ð	59	40	48	17	32	23	12	16	13	12	10	13	10	11	89	47	58	50	78	84	35	17	12	8	10
17	ę	44	27	41	13	22	17	9	12	11	11	-	11	7	8	78	41	44	41	68	65	26	12	9	7	10
18 19	రే రే	49 72	30 35	41 58	17 23	24	17 22	9 11	13 13	12 12	12 11	6 9	12 15	7 9	8 9	81 118	43 63	44 72	44 69	69 98	67 80	28 28	13 17	9 11	7 9	9 12
20	o ç	67	37	54	16	34	22	19	14	12	11	8	10	7	9	89	44	58	56	83	91	34	21	14	9	12
21	ð	79	43	60	15	38	26	21	19	16	13	11	13	11	11	113	55	78	67	104	112	43	25	19	11	16
$\frac{22}{23}$	₽ 8	69 71	36 38	56 61	19 21	38 39	22 26	13 14	16 17	13 15	12 14	10 9	$\frac{13}{14}$	9 9	10 9	108 111	53 59	73 76	64 65	96 98	102 103	39 39	23 22	16 17	11 11	14 14
24 25	0° +0	63 67	35 35	51 52	17 14	32 35	21 22	16 14	12 17	12 13	11 12	7 9	$12 \\ 13$	8 9	9 11	85 96	48 56	65 74	59 58	85 81	83 81	32 31	17 17	12 11	9 9	$\frac{12}{12}$
26 27	ର୍ଚ୍ଚ ଦୁ	54 58	30 32	44 48	13 17	29 30	19 21	12 12	$15 \\ 15$	12 12	12 12	12 12	14 13	9 9	11 12	78 83	43 46	56 56	50 56	73 81	80 85	32 34	17 19	12 12	8 9	11 11

Nos. 1-2. Chorebus senilis (NEES): 1, ex Melanagromyza aeneoventris FALLÉN, Gower, Wales; 2, ex Napomyza scrophulariae SPENCER, Ireland.

Nos. 3-4. Chorebus pulchellus sp. nov. (3 the holotype).

Nos. 5-6. Chorebus brevicornis (THOMSON) ex Melanagromyza aeneoventris FALLÉN: 5, Woodwalton, Hunts., England; 6, Jena, Germany.

Nos. 7-8. Chorebus rondanii (GIARD) ex Ophiomyia simplex LOEW, England.

Nos. 9-10. Chorebus orbiculatae sp. nov. ex Ophiomyia orbiculata HENDEL, Potters Bar, England (10 the holotype).

Nos. 11-12. Chorebus fuscipennis (NIXON) ex Ophiomyia heringi STARÝ, Germany: 11, Dargun; 12, Mühlhausen.

Nos. 13-14. Chorebus caesariatus sp. nov. (14 the holotype).

									······································			Ra	tios						
26	27	28	29	30	31	32	Α	в	C	D	Е	F	G	н	I	J		K	L
Hip Co:		ength	Length	Pet	iole	Total Body Length	of Head	Length/ Height of Head	Width of Head/ Distance between Eyes/ Width of Clypeus	of /Length oles/of Head	Antennal Segments	Height/Length of Thorax	/Head /Width /Body Length		Hind Tibia/Tarsus	Hind Tarsa	of Petiole/	Width/Length of Petiole	
Width	Length	Wing Length	Gaster Length	Width	Length	Total B	Length/ Width of	Length/ Height	Width o Distanc Eyes/ Width o	Width of Mandibles,	3 4 5	- Height/ Thorax	Thorax Width	Wing Length/	Hind Ti	1 2 3	4 5	Lengths Gaster	Width/J Petiole
19 19	32 32	300 300	152 145	22 26	49 48	305 300	1.7 1.6	1.3 1.4	2.0:1:0.6 2.0:1:0.7	340000 3	1.3:1:0.9 1.3:1:0.9	1.6 1.6	1.4 1.3	1.0 1.0	1.0 1.1	2.0:1:0. 2.0:1:0.		3.1 3.0	2.2 1.9
17 17	28 28	276 248	$\begin{array}{c} 135\\ 130 \end{array}$	22 24	44 44	276 276	1.8 1.8	$1.3 \\ 1.3$	1.9:1:0.7 1.8:1:0.7		1.2:1:0.8 1.4:1:0.9	1.7 1.4	1.4 1.3	1.0 1.1	1.0 _	1.9:1:0. 2.0:1:0.		3.0 2.9	$2.0 \\ 1.8$
 18	- 28	305 286	$\begin{array}{c} 155\\ 145\end{array}$	21 21	54 48	329 286	1.9 1.6	$1.5 \\ 1.5$	2.0:1:0.6 1.8:1:-	3.3 3.7	1.0:1:0.9 1.2:1:1.0	$1.5 \\ 1.6$	1.4	1.1 1.0	0.9 1.0	1.9:1:0. 2.1:1:0.		2.9 3.0	$2.6 \\ 2.4$
17 19	24 24	248 257	124 124	21 18	41 40	257 252	1.6 1.5	$1.2 \\ 1.1$	1.9:1:0.6 2.1:1:0.7	1	1.0:1:0.9 1.1:1:1:1.0	1.7 1.8	$1.3 \\ 1.3$	1.0 1.0	0.9 —	2.1:1:0. 2.1:1:0.		3.0 3.1	2.0 2.2
17 16	26 22	228 233	95 93	17 17	43 43	224 233	1.9 1.7	1.4 1.3	2.0:1:0.7 2.0:1:-	2.1	1.1:1:0.9 1.1:1:0.9		1.4 1.3	1.0 1.0	0.9 0.9	2.0:1:0. 2.0:1:0.		2.2 2.1	2.6 2.6
1714	25 21	248 228	95 115	19 17	46 39	233 238	1.9 1.9	1.5 1.4	$\begin{vmatrix} 2.1: 1 : 0.7 \\ 2.2: 1 : - \end{vmatrix}$	2.5	1.1:1:0.9 1.2:1:1.0	1.6 1.7	1.0 1.2	0.9 1.0	0.9 1.0	1.8:1:0. 1.9:1:0.		2.0 2.9	$2.5 \\ 2.3$
14 13	21 18	200 176	85 76	14 13	35 30	195 173	1.6 1.7	1.4 1.5	2.0:1:0.8 2.1:1:0.8		1.0:1:1.0 1.1:1:1.1	1	$1.1 \\ 1.2$	1.0 1.0	1.0 1.0	1.8:1:0. 1.9:1:0.		$2.5 \\ 2.6$	$2.5 \\ 2.3$
14	20	219	108	11	35	205	1.9	1.6	2.0:1:0.5	2.6	1.0:1:0.9	1.6	1.3	0.9	1.0	2.1:1:0	7:0.6:0.7	3.1	3.1
16	23	238	120	13	41	248	1.5	1.2	1.9:1:0.7	3.4	1.2:1:0.9	1.5	1.3	1.0	1.1	2.1:1:0	7:0.5:0.8	3.0	3.1
13 15	19 21	214 209	100 93	13 15	34 32	224 205	1.6 1.7	1.5 1.4	2.1:1:0.8 2.1:1:0.3	1	1.1:1:1:1.0 1.1:1:1:1.0		1.1 1.2	1.0 1.0	1.0 1.0	2.2:1:0. 2.1:1:0.		3.0 2.9	1.000
19	28	286	132	22	48	286	2.1	1.6	2.0:1:0.6	3.2	1.1:1:0.9	1.6	1.1	1.0	0.8	1.7:1:0	7:0.6:0.7	2.7	2.2
17	25	233	126	19	41	262	1	1.5	2.0:1:0.6		1.1:1:0.9		1.5	1.1	1.1	1.7:1:0		3.1	2.2
19	27	286	167	25	50	338	11	1.4	2.1:1:0.3	1	1.2:1:0.8			1.2	1.1	1.7:1:0 1.7:1:0			
17 19	27 27	276 271	135 157	19 21	48 48	286 295	11	1.5 1.6	1.8:1:0.6 1.8:1:0.7	1	$1.2: 1:0.9 \\ 1.1: 1:0.9$	5	1	1.0 1.1	1.1	1.7:1:0 1.7:1:0		3.3	1
$\frac{15}{15}$	22 24	238 248	103 124	14 16	44 48	228 267		1.5 1.5	2.0:1:0. 1.9:1:0.		1.1:1:0.9 1.3:1:0.9	1	1	1.0 1.1	1.0 1.0	1.9:1:0 1.8:1:0		2.3 2.6	a subscript
12 13	21 21	205 233	104 132	12 12	37 41	214 248	1.8 1.8	1.5 1.5	1.9:1:0.0 2.0:1:0.1		$1.2: 1:1.0 \\ 1.2: 1:1.0$	1		1.0 1.1	1.1 1.1	1.9:1:0 1.8:1:0		1	

No. 15. Chorebus lychnidis sp. nov. holotype.

No. 16. Chorebus bathyzonus (MARSHALL) ex Ophiomyia heracleivora SPENCER, England.

Nos. 17-18. Chorebus heringianus sp. nov., Kyffhäuser, Germany (17 the holotype).

No. 19. Chorebus gedanensis (RATZEBURG), Höör district, Sweden (BM).

Nos. 20-23. Chorebus glaber (NIXON) ex Napomyza cichorii SPENCER: 20, Walcheren, Holland; 21, Zaventem, Belgium; 22, Holland (? locality); 23, Isle of Overflakee, Holland.

Nos. 24-25. Chorebus leptogaster (HALIDAY) ex Ophiomyia pulicaria MEIGEN: 24, Barnet, England; 25, Mühlhunson Germany.

Nos. 26-27. Chorebus xiphidius sp. nov. (27 the holotype).

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Table 17 Biometric data

										Abs	olute	Mea	surei	nent	s (1	= 0.	01 n	.m.)									
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	
			Head	1	th (lateral)	Eyes	Width	es Width		nten egme		Ma	xilla Seg	ry Pa ment	alpus ts	r	Thora	ıx	H	ind I	eg		Hind Tarsal Segments				
		Width	Length	Height	Eye-width	Distance between]	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5	
1 2	40 OF	78 72	47	65 61	22 22	39 35	28 23	15 13	22 18	17 16	15 15	12 12	19 16	13 11	13 11	132 117	58 50	87 74	87 76	132 109	1	52 46	27 22	21 17	13 11	16 13	
3 4	ç ç	58 59	35 39	47 47	15 19	30 29	21 19	12 9	16 15	13 13	13 12	10 10	14 13	10 9	12 9	95 93	40 40	58 56	58 58	93 95	91 93	37 39	19 20	15 14	9 9	12 11	
5 6	ç ç	64 52	36 32	54 46	16 17	34 26	21 19	13 10	15 14	13 11	12 11	7 7	10 10	6 6	6 7	96 80	46 39	63 50	59 51	85 75	89 78	37 29	20 18	14 13	8 9	11 10	
7	Ŷ	46	27	38	14	25	17	8	11	12	11	5	7	4	5	63	34	44	39	62	63	23	13	Ģ	7	10	
8	ę	44	34	41	14	21	17	11	13	12	11	7	9	6	7	71	34	52	48	72	74	31	16	11	7	10	

Right part of table 17

														Rat	tios					3				
26	27	28	29	30	31	32	A	В	C	D		E			G	H	I	J					K	L
Hi Co		Length	Length	Pe	tiole	ody Length	/ of Head	of Head	Width of Head/ Distance between Eyes/ Width of Clypeus	of /Length les/of Head	rien			/Head Width	(Body Length	Tibia/Tarsus	Hind Tarsa			Segi	nents	of Pe	length of	
Width	Length	Wing Le	Gaster I	Width	Length	Total B	Length/ Width o	Length/ Height c	Width o Distance Eyes/ Width o	Width of Mandibles	3	4	5		Thorax Width	Wing Length/	Hind Til		2	3	4	5	Lengths Gaster	Width/Length Petiolo
21	39	376	170	30	52	362	1.7	1.4	2.0:1:0.	3.2	1.3	: 1	: 0.9	1.5	1.4	1.0	1.0	1.9	: 1	:0.8	:0.5	: 0.6	3.3	1.8
21	34	329	152	26	48	300	1.7	1.4	2.1:1:0.	3.3	1.2	: 1	· 0.9	1.6	1.4	0.9	1.0	2.1	: 1	:0.8	:0.5	: 0.6	3,2	1.9
14	29	286	126	16	36	262	1.7	1.4	1.9:1:0.	2.9	1.2	: 1	: 1.0	1.6	1.5	0.9	1.0	2.0	: 1	:0.8	:0.5	: 0.7	3.5	2.3
15	28	276	130	17	35	257	1.5	1.2	2.0:1:0.	4.2	1.1	: 1	: 0.9	1.7	1.5	0.9	1.0	2.0	: 1	:0.7	:0.4	: 0.6	3.7	2.1
16	27	-	132	25	36	262	1.8	1.5	1.9:1:0.0	3 2.8	1.1	: 1	: 0.9	1.5	1.4	-	1.0	1.9	: 1	:0.7	:0.4	: 0.6	3.6	1.4
14	22	224	104	22	32	209	1.6	1.4	2.0:1:0.	3.2	1.2	: 1	:1.0	1.6	1.3	0.9	1.0	1.6	: 1	:0.7	:0.5	: 0.6	3.3	1.4
9	17	186	78	15	25	162	1.7	1.4	1.8:1:0.5	3.3	0.9 :	: 1	: 0.9	1.4	1.4	0.9	1.0	1.8	: 1	:0.7	:0.6	: 0.8	3.1	1.7
15	22	209	98	16	32	213	1.3	1.2	2.1:1:0.8	3.1	1.1	: 1	: 0.9	1.4	1.3	1.0	1.0	1.9	: 1	:0.7	:0.5	: 0.6	3.1	2.0

Nos. 1-2. Chorebus cybele (NIXON): 1, ex Melanagromyza lappae LOEW, Herts., England; 2, ex M. eupatorii SPENCER, Woodwalton, England.

Nos. 3-4. Chorebus didas (NIXON): 3, ex Napomyza scrophulariae Spencer, Barnet, London; 4, Bookham, Surrey.

Nos. 5-6. Chorebus parvungulus (THOMSON): 5, ex Napomyza lateralis FALLÉN, Germany; 6, Sweden (holotype).

No. 7. Chorebus ibericus sp. nov. holotype.

No. 8. Chorebus cyclops (NIXON), Ashtead, Surrey, England (BM).

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