

## Redisposition of some taxa of *Pseudolachnea*, with some other transfers

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**Abstract:** *Pseudolachnea juniperi* provides an older name for *Urceolella conspicua* and *U. magnifica*, and a new combination is proposed. The type of *Pseudolachnea pallida* turned out to provide a name for three problematic collections from Sweden, Scotland and Finland. As the hairs lose their structure in KOH, a new combination to *Urceolella* is proposed. *Pseudolachnea minutissima* and *Pseudolachnea echinata* are both shown to be synonyms of *Hyalopeziza pygmaea*. *Hyaloscypha amyloideopilosa* is shown to be a synonym of *Hyaloscypha priapi*. *Hamatocanthoscypha unci-pila* is divided into two taxa: *H. unci-pila* is a taxon on conifers showing MLZ+ asci and small spores; the excluded collections are from deciduous wood and they show larger spores and mostly MLZ- asci. *Helotium straminellum* offers a name for this taxon and a new combination to *Hamatocanthoscypha* is proposed.

**Zusammenfassung:** *Pseudolachnea juniperi* ist ein älterer Name für *Urceolella conspicua* und *U. magnifica*; eine Neukombination wird vorgeschlagen. Es stellte sich heraus, daß der Typus von *Pseudolachnea pallida* einen Namen für drei problematische Kollektionen von Schweden, Schottland und Finnland zur Verfügung stellt. Da die Haare ihre Struktur in KOH verlieren, wird die Neukombination zu *Urceolella* vorgeschlagen. *Pseudolachnea minutissima* und *Pseudolachnea echinata* sind beide Synonyme von *Hyalopeziza pygmaea*. *Hyaloscypha amyloideopilosa* ist ein Synonym von *Hyaloscypha priapi*. *Hamatocanthoscypha unci-pila* wird in zwei Taxa geteilt: *H. unci-pila* ist ein Taxon auf Koniferen mit MLZ+ Ascii und kleinen Sporen; die ausgegliederten Kollektionen sind von Laubholz und haben größere Sporen und meist MLZ- Ascii. *Helotium straminellum* bietet einen Namen für dieses Taxon und die Neukombination zu *Hamatocanthoscypha* wird vorgeschlagen.

In 1934 VELENOVSKY, in his *Monographia Discomycetum Bohemiae*, described a new genus *Pseudolachnea* for seven taxa, with minute apothecia beset with rigid, glassy hairs. In modern sense the genus is a mixtum of taxa, with widely different hair characteristics. Although the genus was considered close to *Lachnea* (FR.) GILLET, the included taxa mostly belong to the glassy-haired *Hyaloscyphaceae* NANNF. Some of the types were available for study during a visit to PRM in 1999. These provided some interesting results discussed below. Type studies on some KARSTEN's and SVRČEK's species also resulted in new synonymizations and combinations proposed below.

The abbreviations used are: CB Cotton blue, CR ammoniacal Congo red, MLZ Melzer's reagent, IKI 1 % Lugol's solution, KOH 10 % solution, H<sub>2</sub>O cells in living state in water.

***Urceolella juniperi* (VELEN.) HUHTINEN, comb. nova.** Figs. 1-2.

**Basionym:** *Pseudolachnea juniperi* VELEN., Monographia Discomycetum Bohemiae, p. 314, 1934.

**Lectotype** (designated here): Czech Republic, Bohemia, Mnichovice, *Juniperus communis* L. (needle), Sept. 1931, J. VELENOVSKÝ (PRM 152880, examined).

**Synonym:** *Urceolella conspicua* HUHTINEN, Mycotaxon 24: 277, 1987.

**Holotype:** Finland, Etelä-Häme, Tammela, Mustiala, on needles of *Juniperus* and leaves of *Populus*, 22. 10. 1866, P. KARSTEN (H, Herb. KARSTEN 2627).

**Synonym:** *Urceolella magnifica* RAITV. & GALÁN, Sydowia 45: 43, 1993.

**Holotype:** Spain, Guadalajara, Anquela del Ducado, on fallen leaves of *Quercus faginea* LAM., 24. 10. 1991, A. RAITVIIR & R. GALÁN (TAA, isotype in Herb. GALÁN, examined).

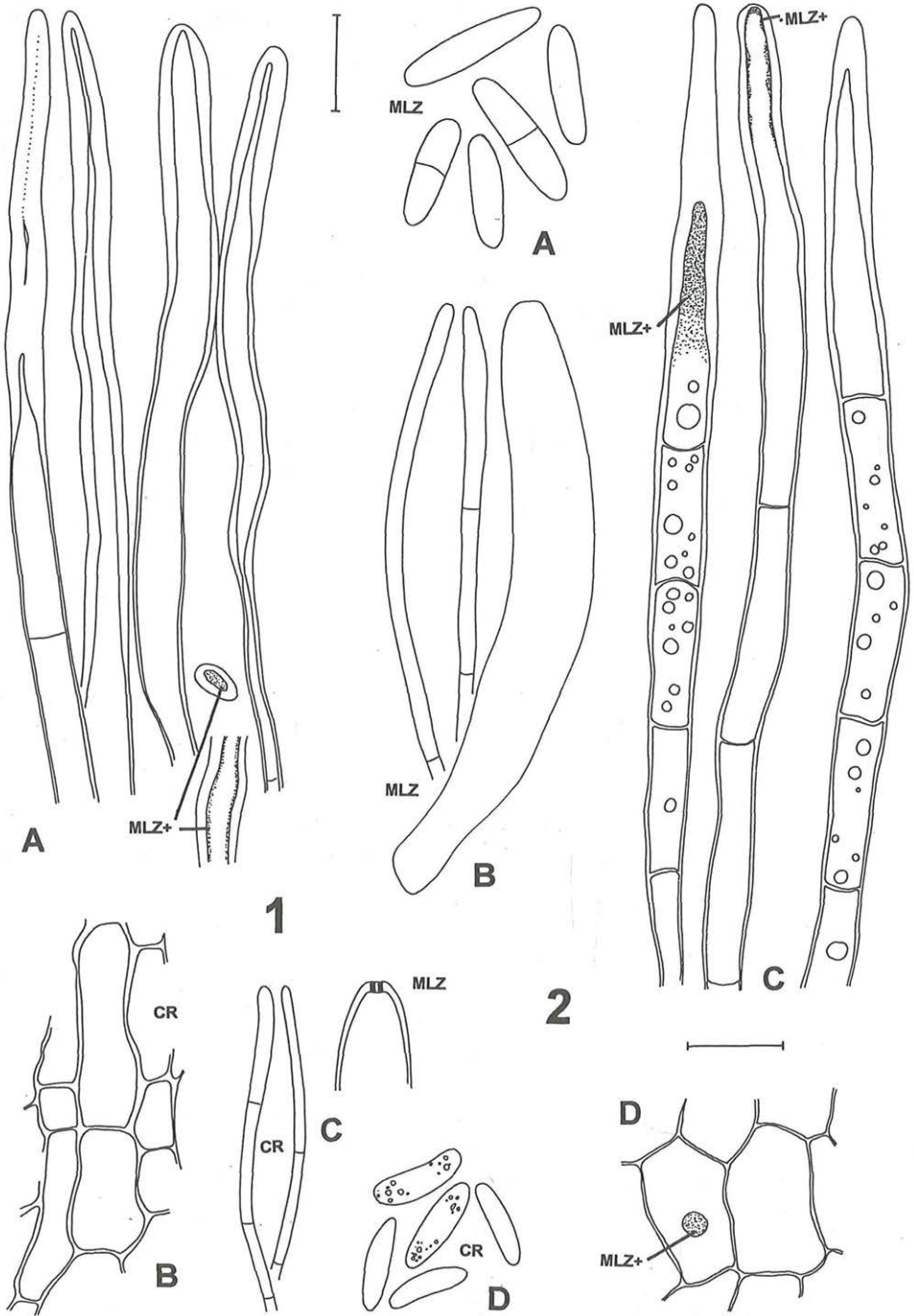
VELENOVSKÝ (1934) diagnosed *Pseudolachnea juniperi* on the basis of two collections. The first one was on *Juniperus* needles while the other was found on a small twig of *Juniperus*. The illustration was based on the former collection and is a good match to the latter specimen, which is here selected as a lectotype of the species. *Urceolella juniperi* is not, however, restricted to grow on juniper. KARSTEN's specimen showed normal apothecia on *Populus* leaves and H.-O. BARAL (pers. comm.) has collected material where the apothecia occur both on juniper needles and on leaves of the surrounding moss (*Hylocomium*). It seems that the mycelium can overtake, to some extent, also the surrounding litter.

Little can be added to previous treatments. The abundance of similarly shaped but totally thin-walled hairs at the margin was not noted by me in KARSTEN's material (HUHTINEN 1987). Nor was the very faint amyloid reaction inside the lumen, which was sporadically seen in the single mount made from the Czech specimen. When on top of each other, the hairs strengthened this reaction but it can also be observed in some of the solitary hairs (Fig. 1A). This reaction is magnificently clear in the type of *Urceolella magnifica* (RAITVIIR & GALAN 1993), where it can be clearly seen that the staining substance is not localized in the wall but on the inner walls (Fig. 2C). The only marked difference to other collections are the predominantly four-spored asci in *U. magnifica*. Developing asci are often 8-spored but when asci with totally mature spores only are observed, nearly all of them are four-spored.

***Urceolella pallida* (VELEN.) HUHTINEN, comb. nova.** Figs. 10-13.

**Basionym:** *Pseudolachnea pallida* VELEN., Monographia Discomycetum Bohemiae, p. 314, 1934.

**Holotype** (the only specimen mentioned): Czech Republic, Bohemia, Radotin, twig of *Cotoneaster*, 7. 9. 1924, J. VELENOVSKÝ (PRM 148675, examined).



Figs. 1-2. *Urceolella juniperi*. 1. Lectotype of *Pseudolachnea juniperi*. A Marginal hairs, B Ectal excipulum, C Ascus apices and paraphyses, D Spores. – 2. Isotype of *Urceolella magnifica*. A Spores, B Ascus and paraphyses, C Marginal hairs, D Ectal excipulum. – Bar: 10 µm.

**Synonym:** *Barbella pallida* VELEN., Monographia Discomycetum Bohemiae, p. 314, 1934.

**Apothecia:** gregarious, superficial, up to 500 µm in diam. when dry, cupulate, broadly sessile, disc Apricot (CAILLEUX 1981: L65) to Straw (K79), in one population dried apothecia pure white, margin somewhat raised, whitish due to a dense hair cover.

**Excipulum:** of textura prismatica – textura angularis close to margin, cells measuring 4-10 x 3-4 µm, typically of textura angularis on middle flanks, cells often measuring 4-6 x 3-5 µm, walls slightly thickened, hyaline, MLZ-, CB-.

**Hairs:** varying from straight-undulating to clearly curved, in one population also rarely bifurcate, up to 35 µm long, mostly shorter, typically aseptate in their exceeding part, basally slightly to clearly widened, up to 5 µm wide, apical part always narrow, tapering to a 0.8-1.0 µm wide apex, wall smooth, basally thin to somewhat thickened, typically thickened in the whole apical part, more rarely the apical part totally solidified or solidified with an inconspicuous lumen, occasional hairs with unevenly distributed wall thickenings; wall MLZ-, CB-, CR-, IKI+ (faintly blue), thickened parts clearly refractive in CB, CR, less so in MLZ, immediately losing their structure and swelling inwards in 10 % KOH.

**Asci:** eight-spored, clavate, 55-74 x 8.5-10.2 µm,  $x_{\text{mean}} = 66 \times 9.6 \mu\text{m}$  (n = 3) when fresh in water, 40-56 x 7.2-9.2 µm,  $x_{\text{mean}} = 46 \times 8.3 \mu\text{m}$  (n = 6) in CB, pore MLZ+ and IKI+ (blue), arising from croziers.

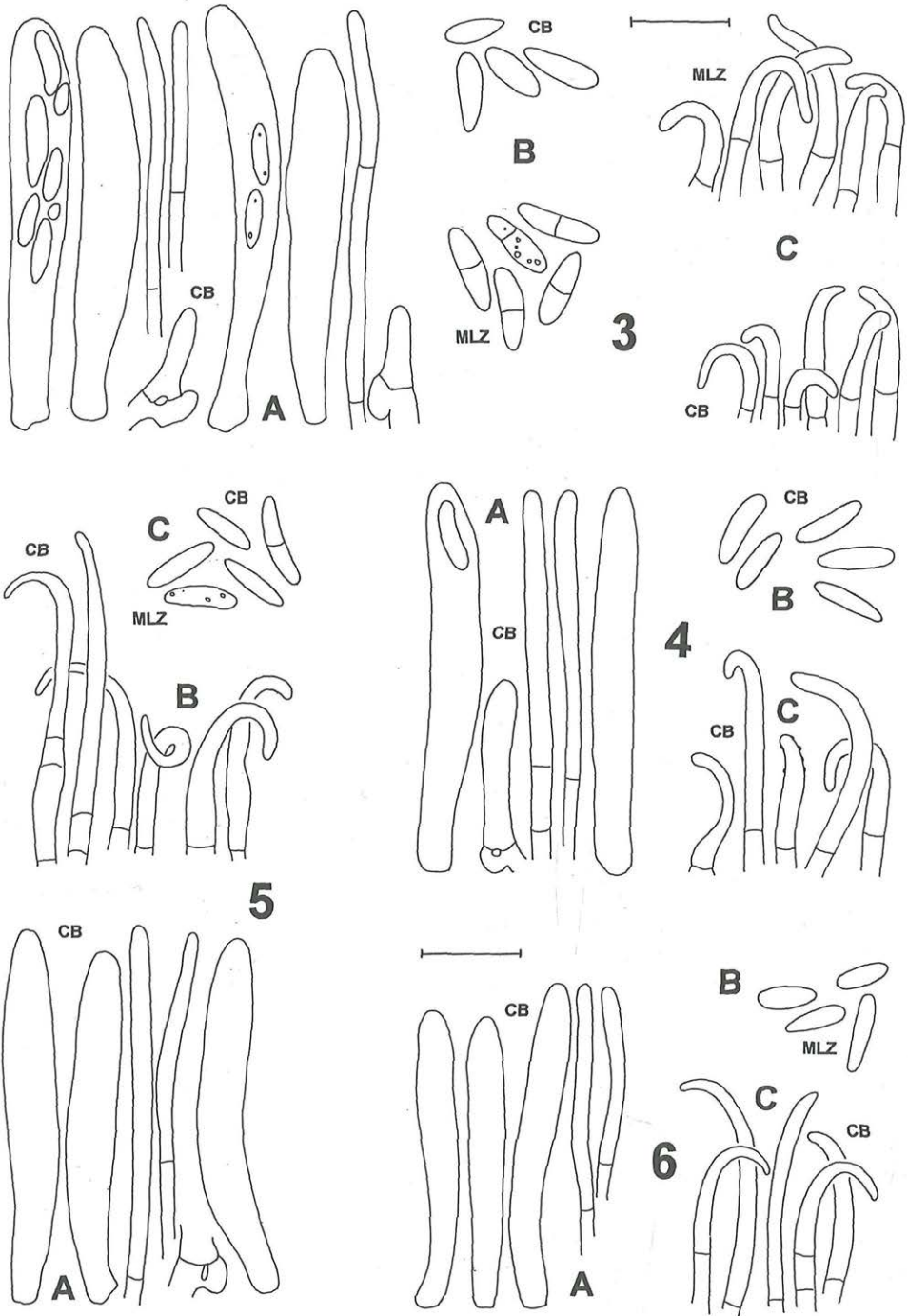
**Ascospores:** ellipsoid-elongate, clearly guttulate in CB, MLZ, frequently one-septate, septum MLZ-; 11.0-15.8 x 4.0-4.6 µm,  $x_{\text{mean}} = 13.2 \times 4.3$  (n = 10), Q = 2.7-4.0,  $Q_{\text{mean}} = 3.1$  when fresh in water; 10.0-14.4(-15.0) x 2.8-4.4 µm,  $x_{\text{mean}} = 12.0 \times 3.6 \mu\text{m}$  (n = 70), Q = 2.5-4.4,  $Q_{\text{mean}} = 3.4$  in CB, MLZ.

**Paraphyses:** narrowly cylindrical, simple, more rarely terminal cells irregularly undulating, 1.0-1.8 µm wide, appearing as empty in CB, MLZ, no superficial encrustations.

**Cultural characters** on 2 % malt agar (1 population, 5 single-spore strains): Radial growth 15.8 mm/month (n = 40, from 5 strains). Mats white, at early stages with creamy tint (K77), later pure white. Aerial mycelium lacking, margin slightly fimbriate, hyphal strands lacking. Anamorph and teleomorph not produced within four months.

A study of the scanty type specimen revealed a name for this long-problematic, lignicolous species. The French specimen on *Urtica* is placed here somewhat tentatively. In this collection the asci appeared first MLZ- but were MLZ+ after KOH-pretreatment. But since all the other characters were identical a wider ecological amplitude is best accepted for *U. pallida*.

The nomenclatural problems concerning this species are intriguing. The protologue indicates the taxon as *Pseudolachnea pallida*. The species is, however, grouped with *P. nivea* VELEN. under the additional generic name *Barbella* VELEN., the diagnosis of which is "Sporae ellipticae, biguttulatae, par. capillares, ramosae. – (*Barbella* VEL.)". In VELENOVSKÝ's plate 16 both taxa are cited as species of *Barbella*. In the index they



Figs. 3-6. *Hamatocanthoscypha straminella*. 3. Lectotype of *Helotium straminellum*. A Asci and paraphyses, B Spores, C Marginal hairs. – 4. REID 5. 10. 1973. A Asci and paraphyses, B Spores, C Marginal hairs. – 5. RAITVIIR 25. 9. 1979. A Asci and paraphyses, B Marginal hairs, C Spores. – 6. KARSTEN 16. 9. 1870. A Asci and paraphyses, B Spores, C Marginal hairs. – Bar: 10 µm.

are listed as *Pseudolachnea*, whereas *Barbella* is listed as a generic name without any included species. According to Art. 34.2. two alternative names are validly published because both species are grouped under *Pseudolachnea* (text and index) and *Barbella* (table and legends). *Barbella* is validly published as a genus, because for a diagnosis before 1. Jan. 1935 no latin diagnosis is required (Art. 36.1.).

**Material studied:** **Finland:** Varsinais-Suomi, Parainen, *Alnus* or *Salix*, 19. 10. 1985, S. HUHTINEN 85/158 (TUR). **France:** Eure, Bosrobert, *Urtica*, 16. 1. 1999, J.-C. MALAVAL 98002 (TUR). **Great Britain:** Scotland, Angus, near Forfar, cortex of *Ulmus glabra* HUDS., 30. 10. 1979, B. COPPINS (E). **Sweden:** Uppland, Dalby par., Dalkarlskräret, *Salix cinerea* L., 22. 11. 1985, K. & L. HOLM 3835m (TUR).

***Hamatocanthoscypha straminella* (P. KARST.) HUHTINEN, comb. nova.** Figs. 3-6.

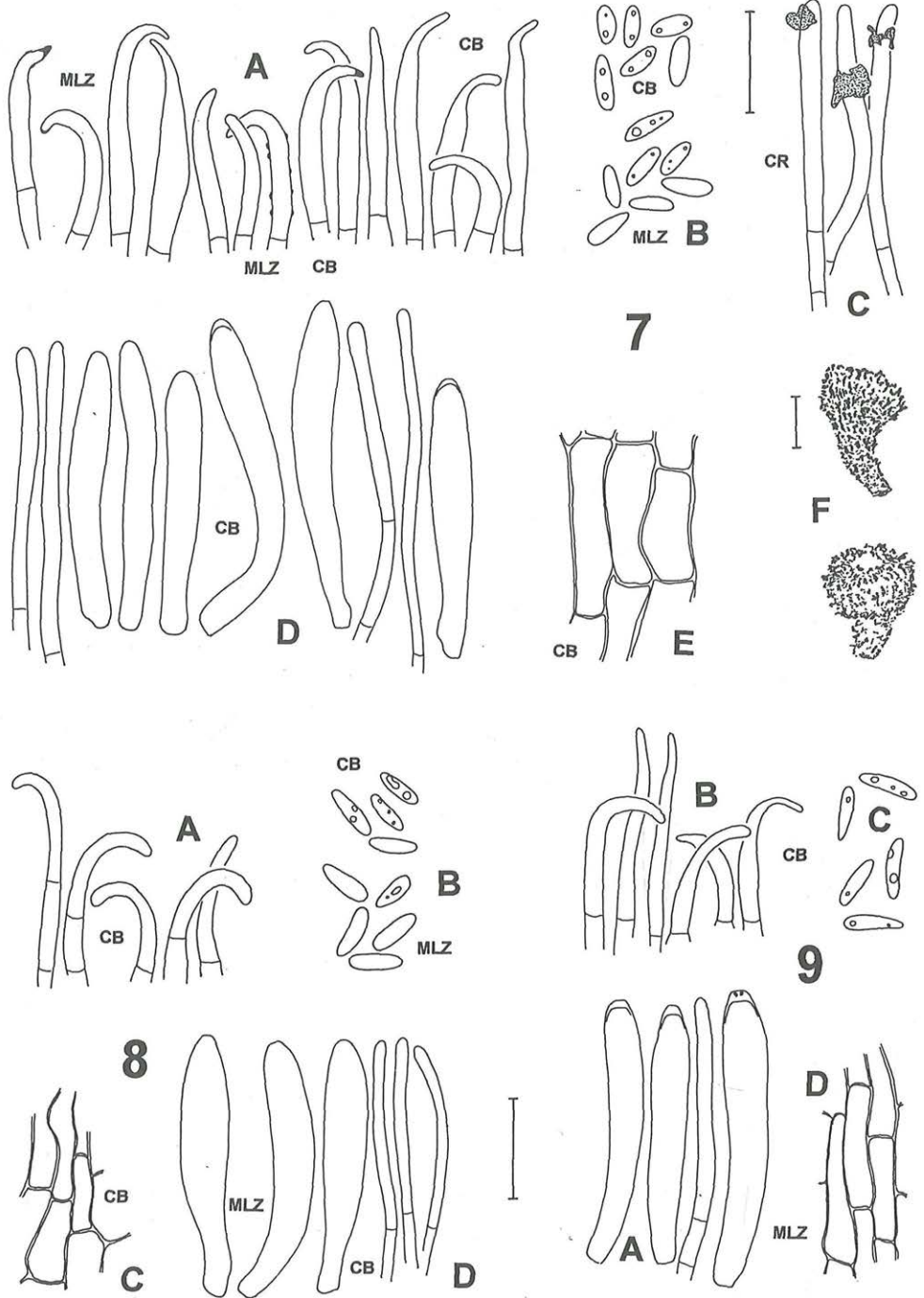
**Basionym:** *Helotium straminellum* P. KARST., Medd. Soc. Fauna Flora Fenn. **16:** 27, 1888. – *Phialea straminella* (P. KARST.) SACC., Syll. Fung. **8:** 273, 1889. – *Hymenoscyphus straminellus* (P. KARST.) O. KUNTZE, Revis. Gen. Pl. **3:** 486, 1898. – *Micropodia straminella* (P. KARST.) BOUD., Hist. Classific. Discomyc. Europe, p. 128, 1907.

**Lectotype** (designated by citation as holotype by CARPENTER 1981: 256; Art. 7.11.): Finland, Etelä-Häme, Tammela, Mustiala, ad lign. betulae, 2. 10. 1869, P. KARSTEN (H, Herb. KARSTEN 2065, examined).

**Synonyms:** *Chrysothallus spiralis* VELEN. sensu DENNIS, Kew Bull. **30:** 352, Fig. 3c, 1975. – *Pezizella viridiflavescens* REHM sensu RAITV., Fol. Crypt. Est. **9:** 1, 1977. – *Hamatocanthoscypha sulphureocitrina* SVRČEK, Česká Mykol. **37:** 65, 1983. – *Hamatocanthoscypha unci-pila* (LE GAL) HUHTINEN p.p., Karstenia **29:** 201-204, 1990.

The concept of *Hamatocanthoscypha unci-pila*, presented by HUHTINEN (1990), embraced two elements. This is seen in the variability of ascal MLZ-reaction, in the two-fold ecological amplitude and in the two maxima seen in the sporograph of *H. unci-pila* (Fig. 215). A restudy of the material revealed that this wide concept should be divided into two clear-cut taxa: one on deciduous wood (*H. straminella*), one on cone scales and coniferous wood (*H. unci-pila*). The former shows, with one exception, MLZ- asci and larger spores. The latter has smaller, MLZ+ asci and smaller spores.

The diagnosis and figs. 212-214, given by HUHTINEN (1990) apply for the present species with the modifications given below. Because the Canadian specimen has a deviating ascal plug reaction and somewhat smaller spores than the rest of the material, its characteristics are given separately below. The other collections show MLZ- asci measuring 35-47 x 4.8-6.0  $\mu\text{m}$ ,  $x_{\text{mean}} = 42.3 \times 5.3 \mu\text{m}$  ( $n = 24$ ),  $Q_{\text{mean}} = 7.7$  in MLZ; 31-48 x 4.0-6.0  $\mu\text{m}$ ,  $x_{\text{mean}} = 39.1 \times 5.0 \mu\text{m}$  ( $n = 22$ ),  $Q_{\text{mean}} = 8.0$  in CB. The living spores measure 8.0-11.5 x 2.3-3.0  $\mu\text{m}$ ,  $x_{\text{mean}} = 9.1 \times 2.8 \mu\text{m}$ ,  $Q_{\text{mean}} = 3.2$  in water; dead spores measure 4.8-9.0 x 1.8-2.8  $\mu\text{m}$ ,  $x_{\text{mean}} = 7.0 \times 2.1 \mu\text{m}$  ( $n = 100$ ),  $Q_{\text{mean}} = 3.4$  in CB and MLZ. The amount of one-septate spores varies between populations from almost none to ca. 90 %.



Figs. 7-9. *Hamatocanthoscypha uncipila*. 7. KORF & al. 28. 9. 1963. A Marginal hairs, B Spores, C Paraphyses, D Asci and paraphyses, E Ectal excipulum, F Dry apothecia. – 8. KORF & al. 30. 10. 1957. A Marginal hairs, B Spores, C Ectal excipulum, D Asci and paraphyses. – 9. LAESSØE 17. 8. 1996. A Asci and paraphysis, B Marginal hairs, C Spores, D Ectal excipulum. – Bar: 10 µm, for apothecia 100 µm.

In the Canadian collection living asci measure  $34-40 \times 5.3-6.4 \mu\text{m}$  ( $n = 5$ ) in water, which is clearly less than in specimen 86/134 where asci measure  $44-59 \times 6.2-7.3 \mu\text{m}$  ( $n = 6$ ). Spores measure  $5.0-7.0 \times 1.8-2.0 \mu\text{m}$  in MLZ which is somewhat overlapping to the rest of the material. As the cultural characters show no marked differences, the Canadian collection is treated as a marginal population of *H. straminella*. The cultural characters and anamorph production indicated in HUHTINEN (1990) thus refer to *H. straminella*, not to *H. uncipila*.

KARSTEN's original diagnosis cites the material seen by him as "prope Mustiala, m. Oct. 1869". Although he definitely used specimen #2065 in the making of the protologue, he did not cite just one element. Specimen #2066 includes similar chips of *Betula*, but lacks the exact collection day. It may be a fragment of the bigger collection, but may also be another collection from the same trunk. Hence, in the absence of definite proof that only #2065 was studied and cited, there cannot be a holotype (Art. 9.1.).

CARPENTER (1981) erroneously linked KARSTEN's species to *Unguicularia* HÖHN. This was due to the abundance of *U. millepunctata* (LIB.) DENNIS on the chips. The characteristics of these apothecia are, however, in contrast to the protologue and KARSTEN's notes on the convolute. Especially the presence of a clear stipe, equalling the disc diameter in length, is a convincing proof that *U. millepunctata* was not part of the protologue of *H. straminellum*. CARPENTER's citation of #2065 as holotype is to be taken as a selection of lectotype (Art. 7.11.).

**Material studied:** **Canada:** Alberta, Edmonton, Devonian Botanical Garden, *Salix*, 28. 9. 1987, S. HUHTINEN 87/179 (TUR). **Finland:** Varsinais-Suomi, Turku, Jäkärälä, cortex of *Populus tremula* L., 28. 9. 1986, S. HUHTINEN 86/134 (TUR); - Etelä-Häme, Tammela, Mustiala, *Betula*, 16. 9. 1870, P. KARSTEN (H, Herb. KARSTEN 2686); - - *Betula*, Oct. 1869, P. KARSTEN (H, Herb. KARSTEN 2066; possible isoelectotype of *H. straminella*); - Nokia, *Salix*, 8. 10. 1991, U. SÖDERHOLM 1887 (TUR). **Great Britain:** Bedfordshire, Heath and Reach, *Populus*, 5. 10. 1973, D. REID (K). **Estonia:** Distr. Rakvere, Vinni, *Quercus*, 17. 10. 1970, A. RAITVIIR (TAA 60857); - Heinassaar, *Populus*, 25. 9. 1979, A. RAITVIIR (TAA 60822).

***Hamatocanthoscypha uncipila* (LE GAL) HUHTINEN.** Figs. 7-9.

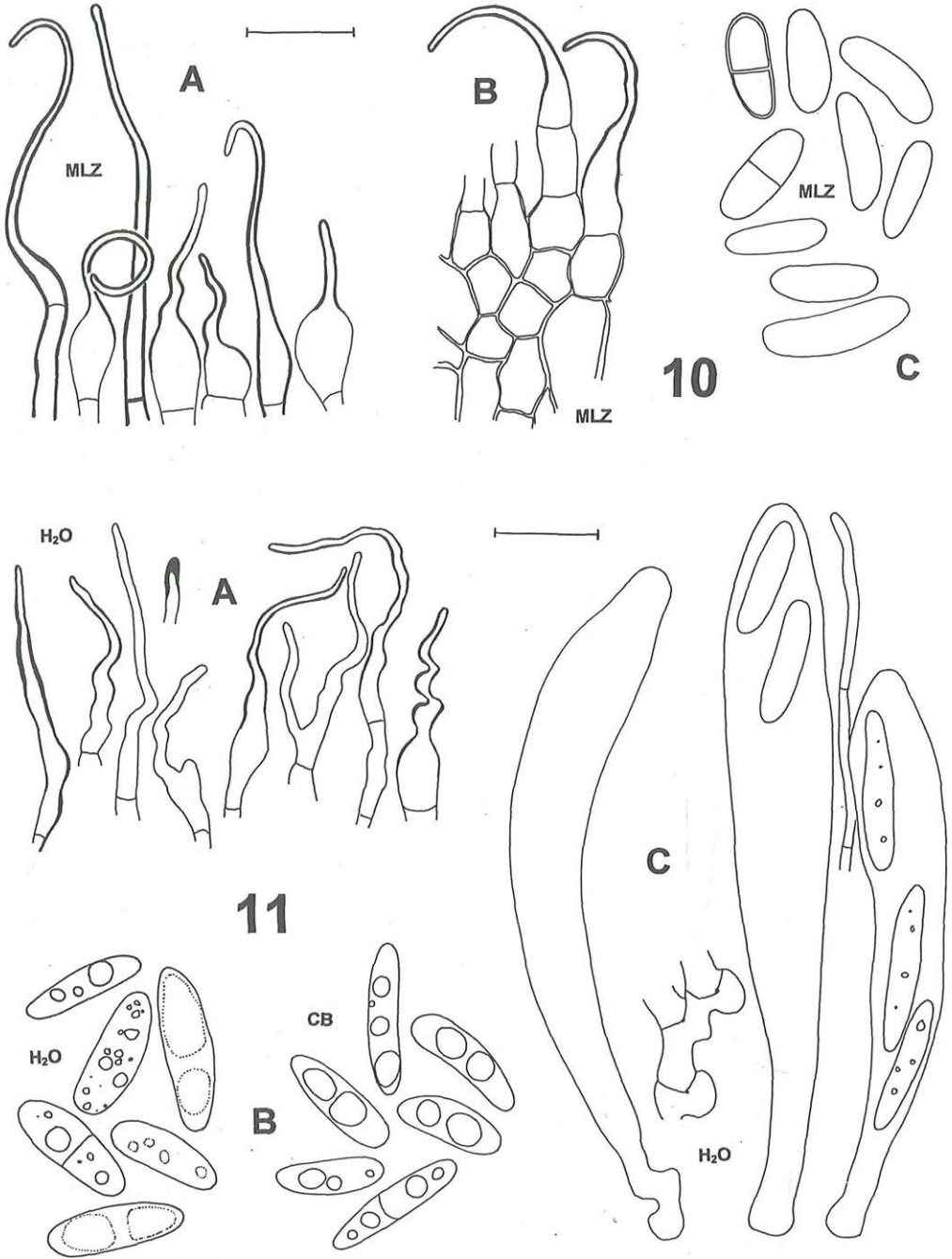
**Basionym:** *Hyaloscypha uncipila* LE GAL, Bull. Soc. Mycol. France **70**: 214, 1954. - *Uncinia uncipila* (LE GAL) RAITV., Scripta Mycol. **1**: 74, 1970.

**Lectotype:** Fig. 12, Bull. Soc. Mycol. France **70**: 215, 1954 (selected by HUHTINEN 1990: 201).

**Synonyms:** *Hyaloscypha sulphureopilosa* SVRČEK, Česká Mykol. **40**: 211, 1986. - *Hamatocanthoscypha uncipila* (LE GAL) HUHTINEN p.p., Karstenia **29**: 201-204, 1990.

From the wide concept presented by HUHTINEN (1990) this name embraces those specimens which occur on cone scales or on coniferous wood and show small MLZ+ asci and clearly smaller spores than those of *H. straminella*. The overall diagnosis given by HUHTINEN (1990) is a good match to the material, with the exceptions discussed below.





Figs. 10-11. *Urceolella pallida*. 10. Holotype of *Pseudolachnea pallida*. *A* Marginal hairs, *B* Hairs on lower flanks, *C* Spores. – 11. HUHTINEN 19. 10. 1985. *A* Marginal hairs, *B* Spores, *C* Asci and paraphysis. – Bar: 10  $\mu$ m.

The asci measure 28-38 x 4.0-5.0  $\mu\text{m}$ ,  $x_{\text{mean}} = 30.8 \times 4.8 \mu\text{m}$  ( $n = 11$ ),  $Q_{\text{mean}} = 6.5$  in MLZ; 24-38 x 3.4-4.6  $\mu\text{m}$ ,  $x_{\text{mean}} = 32 \times 4.1 \mu\text{m}$  ( $n = 39$ ),  $Q_{\text{mean}} = 7.9$  in CB. The spores measure 4.0-7.2(-8.0) x 1.4-2.2  $\mu\text{m}$ ,  $x_{\text{mean}} = 5.2 \times 1.8 \mu\text{m}$  ( $n = 60$ ),  $Q_{\text{mean}} = 2.9$  in MLZ and CB. In CUP-JA-2527 some apothecia show clearly brown-pigmented hairs, like those characterizing *Hamatocanthoscypha uncinata* (PHILL.) HUHTINEN var. *phaeotricha* (K. HOLM & L. HOLM) HUHTINEN. They occur, however, only at the base of the apothecium and show also marked variability between apothecia; some of which have only hyaline hairs. The spores of this specimen are also clearly different from those of var. *phaeotricha*.

Unfortunately *H. uncipila* can at present, awaiting the original collection to be located in P, be typified only by LE GAL's drawing. The likeness in morphological characters and in ecology is, however, good enough. The type of *Hyaloscypha sulphureopilosa* has not yet been restudied. The proposed synonymy is based on the pigmentation, ecology and spore size. SVRČEK illustrated mostly undulating marginal hairs, which are only slightly curved. Also the asci were reported larger, measuring 40-50 x 6-8  $\mu\text{m}$ . Until the type can be studied, the synonymy is somewhat vague.

**Material studied:** **Finland:** Etelä-Häme, Lempäälä, on *Picea* cone, 17. 8. 1986, T. LAESSØE (C). **Japan:** Nagasaki Pref., Unzen National Park, on a board, 30. 10. 1957, R. KORF & al. (CUP-JA-305); - Kushiro Prov., Akan National Park, on a branch of *Picea jezoensis* (SIEBOLD & ZUCC.) CARRIÈRE, 28. 9. 1963, R. KORF & al. (CUP-JA-2527). **Norway:** Svalbard, Longyearbyen, on a board, 17. 8. 1988, S. HUHTINEN 88/65 (TUR).

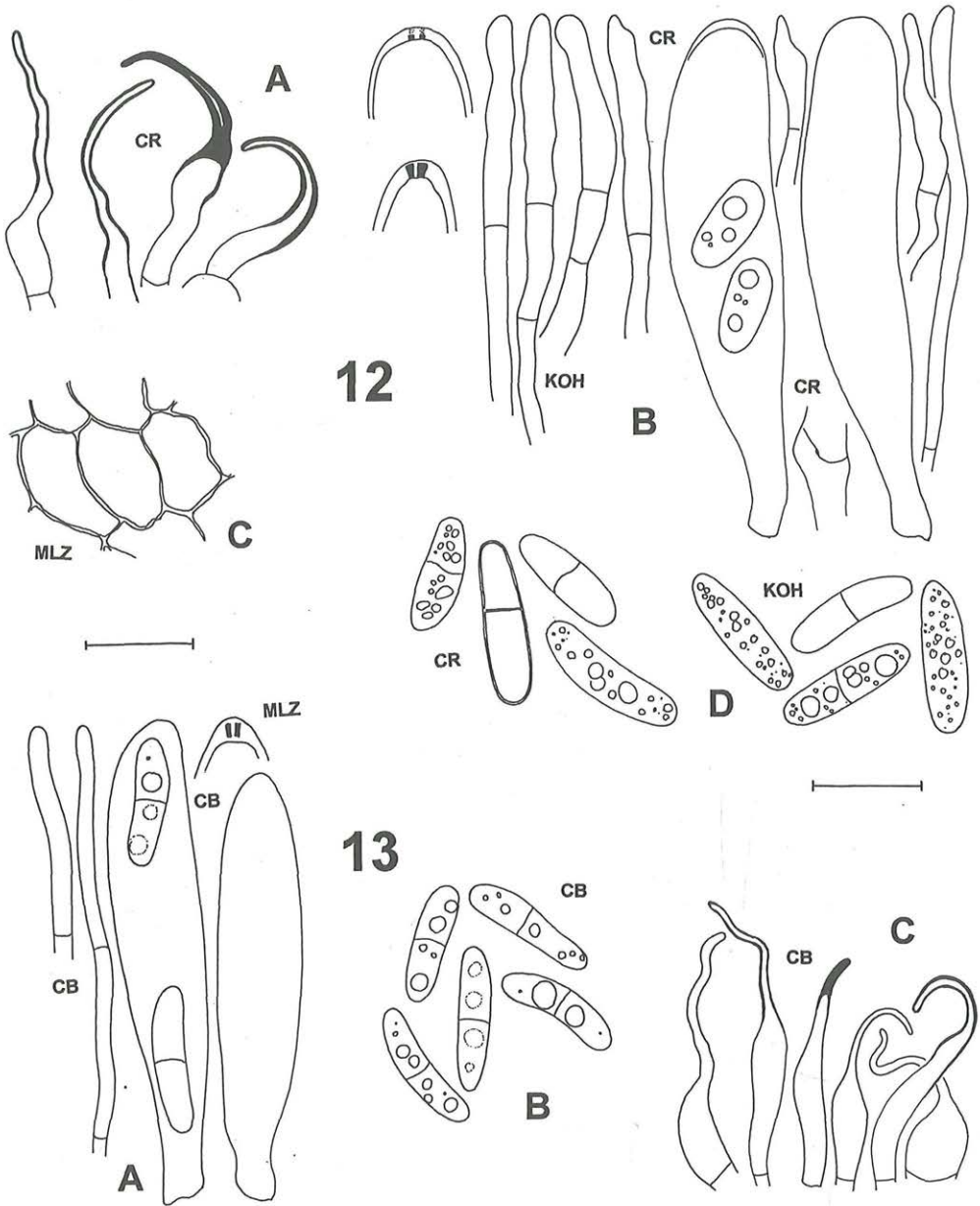
*Hyaloscypha priapi* VELEN. Figs. 14, 16.

**Synonym:** *Hyaloscypha amyloideopilosa* SVRČEK, Česká Mykol. 37: 65, 1983.

**Holotype:** Czech Republic, Bohemia, Poříčko apud Čerňovice, in valle rivi Křešický potok, ad lignum durum rami deiecti *Carpini betuli*, 18. 10. 1968, M. SVRČEK (PRM 829226, examined).

HUHTINEN (1990) suggested that *Hyaloscypha amyloideopilosa* SVRČEK would be conspecific with *Hyaloscypha aureliella* (NYL.) HUHTINEN. This was based on the original diagnosis, because the type of SVRČEK's species was not available for study at that time. The diagnosis was not accompanied with any illustrations (SVRČEK 1983). Most of the diagnostic characters matched those of *H. aureliella*. Especially interesting were SVRČEK's observations on the dark amyloid hair apices in *H. amyloideopilosa*. This matched the dark amyloid reactions seen in the excipulum of 29 % of the populations of *H. aureliella* and inside the hairs of 6 % of the populations (HUHTINEN 1990). The reported substrate (*Carpinus*) was considered as an error, as *H. aureliella* had not been observed to grow on deciduous wood.

A study of the holotype showed that SVRČEK's species is conspecific with *Hyaloscypha priapi* VEL. The hairs and excipulum in this taxon show occasional, clearly dextrinoid solidifications. Just as in *H. aureliella*, the variability between the different MLZ-reactions of the solidifications characterizes also *H. priapi*. Of the few populations studied so far, most show dextrinoid areas. The type of *H. amyloideopilosa* is the



Figs. 12-13. *Urceolella pallida*. 12. HOLM & HOLM 22. 11. 1985. *A* Marginal hairs, *B* Asci and paraphysis, *C* Ectal excipulum, *D* Spores. – 13. COPPINS 30. 10. 1979. *A* Asci and paraphyses, *B* Spores, *C* Marginal hairs. – Bar: 10 µm.

first population with deep amyloid areas. They occur in the excipulum and at hair apices. Especially the shorter hairs on lower flanks have solidified apices; marginal hairs more rarely so. Only one apothecium was mounted but the variability can be concluded from SVRČEK's original text where another apothecium was stated to have mostly inamyloid hairs. In contrast to what was stated in the original diagnosis, the hair wall proper is always inamyloid, while the lumen is apically filled with amyloid matter.

***Hyalopeziza pygmaea* (MOUTON) HUHTINEN.** Figs. 15, 17.

**Basionym:** *Trichopeziza pygmaea* MOUTON, Bull. Soc. Roy. Bot. Belg. **36**: 19, 1897. – *Hyaloscypha pygmaea* (MOUTON) BOUD., Hist. Classific. Discomyc. Europe, p. 127, 1907. – *Unguicularia pygmaea* (MOUTON) SVRČEK, Česká Mykol. **39**: 217, 1985.

**Lectotype** (designated here): Belgium, Liege, Beaufays, feuille de charme (*Carpinus*), V. MOUTON 473 (BR, examined).

**Synonym:** *Hyaloscypha subtilis* VELEN. var. *drupacea* VELEN., Monogr. Discom. Bohemiae, p. 273, 1934.

**Synonym:** *Pseudolachnea minutissima* VELEN., Monogr. Discom. Bohemiae, p. 315, 1934.

**Holotype** (only one element cited in protologue): Czech Republic, Bohemia, Mnichovice, Hubačov, ad conum pinaceum, 13. 8. 1931, J. VELENOVSKÝ (PRM 152878, examined)

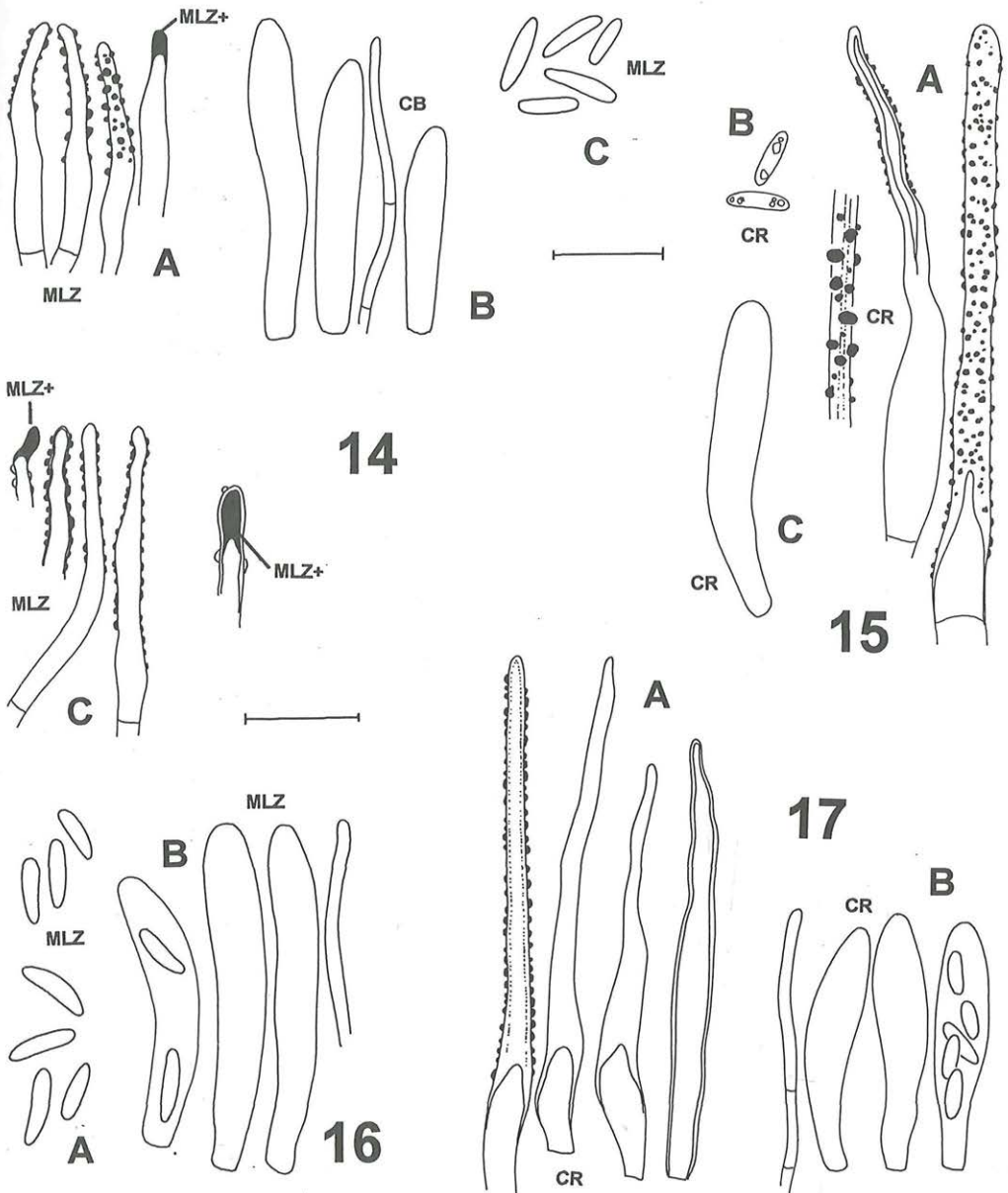
**Synonym:** *Pseudolachnea echinata* VELEN., Monogr. Discom. Bohemiae, p. 314, 1934.

**Lectotype** (selected here): Czech Republic, Bohemia, Mnichovice, "Bila Shala" infra Božkov, *Rosa* sp., 8. 12. 1928, J. VELENOVSKÝ (PRM 152881, examined).

Both types of *Pseudolachnea* show identical characteristics to the type of *T. pygmaea*. Lignicolous habit was not linked to any clear-cut difference to foliicolous collections. Both Czech types were studied in CR and they showed the same variability in hair vesture as discussed by HUHTINEN (1987). The specimen on cone scales had smooth and rough hairs in equal abundance. When mounted in MLZ, all hairs were clearly rough. The specimen on *Rosa* showed only rough hairs in CR. Hair walls appeared as CR+, as already noticed by RAITVIIR & GALÁN (1993).

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Figs 14-17. *Hyaloscypha priapi* and *Hyalopeziza pygmaea*. 14. Lectotype of *Hyaloscypha priapi*. A Marginal hairs, B Asci and paraphysis, C Spores. – 15. Lectotype of *Pseudolachnea echinata*. A Marginal hairs, B Spores, C Ascus. – 16. Holotype of *Hyaloscypha amyloideopilosa*. A Spores, B Asci and paraphysis, C Marginal hairs, with one detail showing the location of amyloid solidification. – 17. Holotype of *Pseudolachnea minutissima*. A Marginal hairs, B Asci and paraphysis. – Bar: 10 µm.

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