

Ecological specificities and molecular diversity of truffles (genus *Tuber*) originating from mid-west of the Balkan Peninsula

Marjanović, Ž.^{1*}, Grebenc, T.², Marković, M.³, Glišić, A.⁴, and Milenković, M.⁵

1 Institute for Multidisciplinary Research, Kneza Višeslava 1a,
11 030 Beograd, Serbia,

2 Slovenian Forestry Institute, Vec̃na pot 2, SI-1000 Ljubljana, Slovenia
3 Makedonska 34, 26340 Bela Crkva, Serbia

4 Hajduk Veljkova 51/9, 14 000 Valjevo, Serbia

5 Institute for Biological Research "Siniša Stanković", Bulevar Despota
Stefana 142, 11000 Beograd, Serbia

Marjanović, Ž.¹, Grebenc, T.², Marković, M.³, Glišić, A.⁴, and Milenković, M.⁵ (2010) Ecological specificities and molecular diversity of truffles (genus *Tuber*) originating from mid-west of the Balkan Peninsula *Sydowia* 62 (1): 67–87

Little is known on the diversity and distribution of truffles at the Balkan Peninsula. A first detailed study of hypogeous fungi in Serbia has been started at 1992. To extend the knowledge on European truffles and their natural habitats, data on their diversity in Serbia, and to some extent in Montenegro and FYRO Macedonia are summarised within this paper. Twelve species of the genus *Tuber*, including five varieties of *T. rufum* Pico, are recorded and their habitats briefly described. Four species are reported for the Balkan Peninsula for the first time. In addition, parts of ribosomal DNA (ITS regions) of 46 specimens are sequenced and analysed to confirm the morphological determinations. ITS sequences of specimens morphologically identified as *Tuber fulgens* Qué! are reported for the first time. A phylogeny based on ITS sequences of the Balkan truffles and additional 29 records from the GenBank is calculated. The results are discussed towards expanding the information on distribution as well as ecological and molecular diversification of *Tuber spp.* in Europe.

Keywords: hypogeous Ascomycetes, phylogeny, ecology, Europe

Currently, 50 genera of hypogeous fungi are recorded in Europe which implies a lower diversity rate of hypogeous fungi than known in Australia or North America (Simpson 2000; Bougher & Lebel 2001). Recently, Montecchi & Sarasini (2000) described 178 European species, among them 22 species of *Tuber spp.* Based on morphological parameters, Rioussat *et al.* (2001) recognize 24 European species of this genus, and divide them in 6 taxonomical groups. Many misidentifications and synonyms based on the morphological descriptions of ascoma created unclear overview of species of genus *Tuber* in Europe (reviewed in Rioussat *et al.* 2001), and many authors tried to resolve this

by using different methods of comparing DNA sequences originating from European truffles (Henrion *et al.* 1994; Gandeboeuf *et al.* 1997; Roux *et al.* 1999, Mello *et al.* 2000, 2002; Halasz *et al.* 2005; Weden *et al.* 2005; Jeandroz *et al.* 2008). Still, the problem with connecting morphological and the molecular identification of *Tuber spp.* is continuing. Within the most recent comprehensive monographic publication on the *Tuber spp.* in Europe, Ceruti *et al.* (2003) list 32 species and refer to the Mediterranean and sub-Mediterranean zones of Europe (Iberian and Apennine Peninsulas and Southern France) as the regions of greatest diversity of *Tuberaceae*. The Balkan Peninsula, as the largest and the most continental peninsula of the South Europe until now has been almost unexplored for hypogeous fungi.

Due to its geographic position, its geological, topographical and climatic diversity, the biodiversity of Balkan Peninsula is one of the highest in Europe (Stevanović *et al.* 1995; Myers 1999). Moreover, the area served as a refugium site for many European plants in the times of glaciations, and has therefore been the natural centre of plant diversification during times of interglaciations (Bennet *et al.* 1991; Taberlet *et al.* 1998; Petit *et al.* 2002a and 2002b). The western part of the peninsula is dominated by continental climate, with Mediterranean influence that is declining from the Adriatic sea towards central peninsular regions. Calcareous and very high Dinaric Alps start steeply from the Adriatic coast and spread eastward till the Western Serbia and northern Former Yugoslav Republic of Macedonia (FYROM). They form natural barrier for the wet maritime air currents, which causes annual precipitation rates gradient, decreasing from more than 2000ml in Montenegro coast, over app. 1000 ml in Western Serbia, to as low as 500 ml in east Serbia and FYROM (Atlas klime SFRJ 1969). Average annual temperatures vary between 8.8–16.6°C in Montenegro, 9.5–11.7°C in Serbia and 8.8–14.6°C in FYROM (Atlas klime SFRJ 1969). Seasonal variations of precipitation and temperature are pronounced in the northern and central regions of the peninsula. Such climate conditions determined environment supportive for temperate ectomycorrhiza (ECM) forming trees that dominate forest communities in entire investigated area (Stevanović *et al.* 1995, Kojić *et al.* 1997). Only small areas on the predominantly steep Adriatic coast are covered by macchia vegetation on the hillsides, and pines (*Pinus pinea* L., *P. pinaster* Ait.) or Mediterranean oaks (*Quercus pubescens* Wild., *Q. ilex* L.) on the plane areas.

From already known truffle-producing areas, ecological requirements and eco-geographic limits of European truffles are well documented (Szemere 1965, Alvarez *et al.* 1993; Rioussset *et al.* 2001; Chevalier & Frochot 2002; Ceruti *et al.* 2003; Jeandroz *et al.* 2008). However, some data on truffle habitats in Serbia are contradictory to these findings (Milenković & Marjanović 2001). Records of *Tuber spp.* from the Balkan Peninsula have been sporadically published (Lindtner 1935;

Frančisković 1950; Hrka 1988; Pázmány 1991; Milenković *et al.* 1992; Glamočlija *et al.* 1997; Marjanović & Milenković 1998; Milenković & Marjanović 2001), but a number of data appeared to be unclear or out-dated.

The aim of this report was to provide a comprehensive list of species of *Tuber spp.* found in Serbia during 17 years of investigation, as well as their molecular determination and relation to specimens originating from other European territories. Results from Montenegro and FYROM are included as well. Furthermore, an overview of some differential to previously reported characteristics of the ecosystems that support truffle fructification in the mid-west of Balkan Peninsula are provided. Therefore the distribution, general ecological features and molecular diversity of some European truffles is re-evaluated.

Materials and Methods

Sampling, morphological identification, preservation

Calcareous areas of Serbia and to some extent of Montenegro and FYROM (Fig. 1) have been explored for truffles by few collectors since 1992 with a help of trained dogs. More than three hundred collections (362) of *Tuber spp.* have been supplemented with comprehensive photo documentation and data on microscopic characters. Specimens have been preserved either by drying, or immersion in a 2:1 ethanol-glycerol mixture. Collections are presently deposited at the Institute for Biological Research „Siniša Stanković“ and at Institute for Multidisciplinary Research in Belgrade, Serbia. Specimens (great majority of them dry) have been morphologically identified according to Szemere (1965), Pegler *et al.* (1993), Zambonelli *et al.* (2000), Montecchi & Sarasini (2000), Rioussset *et al.* (2001) and Ceruti *et al.* (2003). Only ecological data supplied together with specimens by collectors were analyzed: potential plant host as registered on the site, date of collection, approximate elevation and altitude of the site. Soil data were taken from Soil map of Serbia (Tanasijević *et al.* 1965, 1966) or registered by collector.

DNA isolation

About 50ng of dry fungal material of the inner part of ascomata was used for DNA extraction. Samples were taken using a binocular, to avoid contamination with soil particles or parasitic fungi. The DNA was extracted either by using 2% CTAB (Rogers & Bendlich 1985; Doyle & Doyle 1990) or by Plant DNeasy Mini Kit (Promega). After both procedures the DNA was suspended in pre-warmed, sterile Milli-Q water to the approximate final concentration 100 ng/μl.

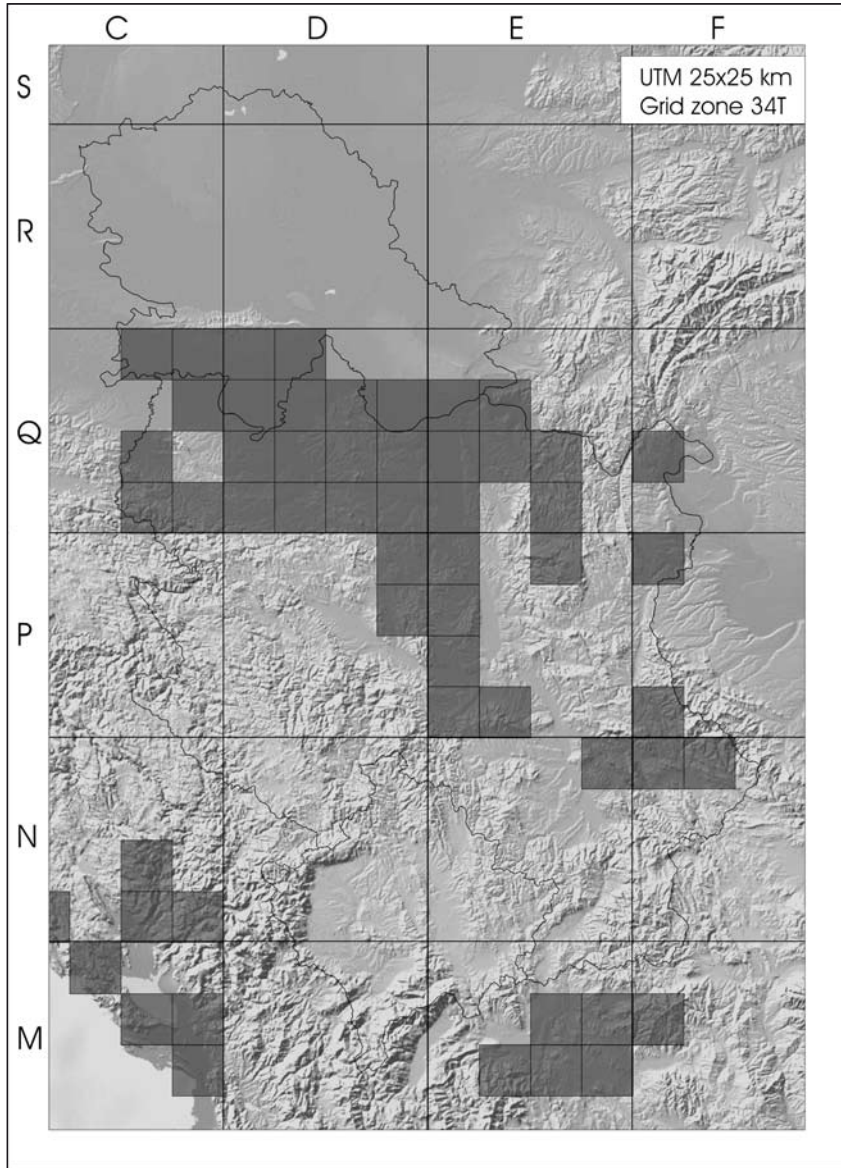


Figure 1. UTM map of the mid-west of Balkan Peninsula (Grid zone 34T). The codes of UTM quadrants are given. Shaded quadrants (size 25 x 25 km) represent area covered by investigation

PCR amplification

The fungal specific primers ITS1F (Gardes & Bruns 1993) and ITS4 (White *et al.* 1990) were used for PCR amplification of the ITS1-5.8S-ITS2 rDNA region. Amplifications were done using standard procedure described in White *et al.* (1990) in a total reaction volume of 40 µl with AmplyTaq Gold polymerase and modified according to Kraigher *et al.* (1995), using a PE 9700 DNA thermocycler and annealing temperature of 55°C. When no or little PCR product could be obtained the annealing temperature was lowered down to 50 °C to. Negative controls were run for each experiment to check for the contamination of reagents. Amplified DNA was purified and analysed as described in Grebenc *et al.* (2000).

Sequencing

Prior to sequencing the amplified DNA was separated on 2% agarose gel and purified using the Promega Wizard SV Gel and PCR Clean-Up System. For sequencing the primers mentioned above were used, sequencing was conducted using a ABI Prism 310 automatic sequencer (Applied Biosystems). Sequencer 4.8 software (Gene Codes) was used to read, identify and clean the consensus sequence from the two strands of each isolate. All consensus sequences were blasted against the GenBank nucleotide database using a nucleotide query (<http://www.ncbi.nlm.nih.gov/blast/Blast.cgi>). Sequences have been lodged in the EMBL database with accession numbers indicated in Tab. 2.

Phylogenetic analyses

Sequences were aligned using CLUSTAL W software package (Larkin *et al.* 2007). The NJ tree based on 393 informative characters was constructed applying *MEGA* version 4 (Tamura *et al.* 2007) using the K2P model presuming uniform substitution rates. Bootstrap values were calculated based on 10000 bootstrap replicates. Gaps were treated as missing data.

Results

Twelve species of the genus *Tuber* were recognised at the investigated area by morphological criteria according to Montecchi & Sarasini (2000). Four species (*T. maculatum*, *T. oligospermum*, *T. borchii* and *T. fulgens*) are new records for the Balkan Peninsula. The study also revealed first records of six species from Montenegro, and five species from FYROM (Tab. 1).

Table 1. – Diversity, distribution and ecological parameters of truffle species recorded in investigated part of Balkan Peninsula. Ascoma maturation periods expressed as Roman numbered months of the year.

Species (according to ascomata morphology)	Potential plant host	Maturation period distribution	Inclination altitude soil type
<i>Tuber aestivum</i> Vittad. (exemined 44 specimens)	<i>Quercus cerris</i> L.	I-XII	0°– 60°
	<i>Quercus pubescens</i> Wild.		
	<i>Quercus frainetto</i> Ten.	Serbia	0 – 1000 m a.s.l
	<i>Quercus robur</i> L.	Montenegro	
	<i>Fagus sylvatica</i> L.	FYROM	cambisol
	<i>Corylus avellana</i> L.		rendzina
	<i>Carpinus betulus</i> L.		alluvium
<i>Tuber mesentericum</i> Vittad. (exemined 13 specimens)	<i>Betula pendula</i> L.		lithosol
	<i>Tilia</i> sp.		vertisol
	<i>Quercus cerris</i> L.	I-III, VII-XII	20°– 60°
	<i>Quercus pubescens</i> Wild	Serbia	
	<i>Ostrya carpinifolia</i> Scop.	Montenegro	0 – 1000 m a.s.l
	<i>Carpinus betulus</i> L.	FYROM	
	<i>Populus alba</i> L.		rendzina
<i>Tuber brumale</i> Vittad. including <i>T. brumale</i> var. <i>moschatum</i> ; (exemined 70 specimens)	<i>Tilia</i> sp.		lithosol
	<i>Pinus pinea</i> L.		cambisol
	<i>Quercus robur</i> L.	I-V, IX-XII	0° – 60°
	<i>Quercus cerris</i> L.		
	<i>Quercus pubescens</i> Wild	Serbia	0 – 500 m a.s.l
	<i>Fagus sylvatica</i> L.	Montenegro	
	<i>Ostrya carpinifolia</i> Scop.	FYROM	semiglay
<i>Tuber macrosporum</i> Vittad. (exemined 45 specimens)	<i>Populus alba</i> L.		rendzina
	<i>Tilia</i> sp.		lithosol
	<i>Pinus pinea</i> L.		cambisol
	<i>Quercus cerris</i> L.	X – XII(I)	0° – 20°
	<i>Quercus robur</i> L.		
	<i>Carpinus betulus</i> L.	Serbia	0 – 500 m a.s.l
	<i>Corylus avellana</i> L.	Montenegro	
<i>Tuber magnatum</i> Vittad. (exemined 36 specimens)	<i>Populus alba</i> L.	FYROM	cambisol
	<i>Salix</i> sp.		aluvium
	<i>Quercus robur</i> L.	IX – XII (I)	0° – 5°
	<i>Carpinus betulus</i> L.		
	<i>Populus nigra</i> L.	Serbia	0 – 20 m a.s.l
	<i>Populus alba</i> L.		semiglay
	<i>Salix</i> sp. L.		humogley
<i>Tuber oligospermum</i> (Tul. & C. Tul.) Trappe (exemined 8 specimens)			alluvium
	<i>Quercus pubescens</i> Wild	XII – III	30° – 60°
	<i>Ostrya carpinifolia</i> Scop.		
	<i>Carpinus orientalis</i> Mill.	Serbia	200 – 500 m a.s.l;
	<i>Pinus halepensis</i> Mill.	Montenegro	rendzina

Species (according to ascomata morphology)	Potential plant host	Maturation period distribution	Inclination altitude soil type
<i>Tuber maculatum</i> Vittad. (exemined 15 specimens)	<i>Carpinus betulus</i> L. <i>Populus alba</i> L. <i>Populus sp.hybrid</i>	IV, VI, X – XII Serbia	0°– 5° 0 – 20 m a.s.l cambisol alluvium antropogenic soils
<i>Tuber borchii</i> Vittad. (exemined 2 specimens)	<i>Quercus cerris</i> L. <i>Quercus frainetto</i> Ten. <i>Carpinus betulus</i> L. <i>Populus tremula</i> L.	II – IV, XI Serbia	app 30° app 200 m a.s.l cambisol
<i>Tuber foetidum</i> Vittad. (exemined 3 specimens)	<i>Salix fragilis</i> L. <i>Salix alba</i> L. <i>Populus alba</i> L.	V, VI, XII Serbia	0° – 5° 0-20 m a.s.l humogley
<i>Tuber excavatum</i> Vittad. (exemined 54 specimens)	<i>Quercus robur</i> L. <i>Quercus cerris</i> L. <i>Quercus pubescens</i> Wild <i>Fagus sylvatica</i> L. <i>Corylus avellana</i> L. <i>Populus alba</i> L. <i>Salix</i> sp. <i>Pinus</i> sp.	VI – XII (I) Serbia Montenegro FYROM	0° – 30° 0 – 1200m a.s.l cambisol podzol alluvium rendzina
<i>Tuber fulgens</i> Quéf. (exemined 16 specimens)	<i>Fagus sylvatica</i> L. <i>Tilia cordata</i> Mill.	VII, IX – X Serbia	0° – 30° 0 – 1200 m a.s.l rendzina cambisol
<i>Tuber rufum</i> Pico var. <i>rufum</i> according to Montecchi & Sarrasini (2000) (exemined 35 specimens)	<i>Quercus robur</i> L. <i>Quercus cerris</i> L. <i>Quercus frainetto</i> Ten. <i>Quercus rubra</i> L. <i>Fagus sylvatica</i> L. <i>Carpinus betulus</i> L. <i>Corylus avellana</i> L. <i>Betula pendula</i> Roth. <i>Populus nigra</i> L. <i>Populus alba</i> L. <i>Abies alba</i> L.	I – III, VII – XII; Serbia Montenegro FYROM	0° – 60° 0 – 1000 m a.s.l cambisol alluvium rendzina lithosol atropogenic soils
<i>Tuber rufum</i> var. <i>apiculatum</i> E. Fisch. according to Montecchi & Sarrasini (2000) (exemined 5 specimens)	<i>Fagus sylvatica</i> L. <i>Picea abies</i> L.	IX – XI Serbia	20° – 60° 500 – 1000 ma.s.l. Cambisol

Species (according to ascomata morphology)	Potential plant host	Maturation period distribution	Inclination altitude soil type
<i>Tuber rufum</i> var. <i>ferrugineum</i> (Vittad.) Montecchi & Lazzari according to Montecchi & Sarrasini (2000) (examined 7 specimens)	<i>Quercus pubescens</i> Wild <i>Fagus sylvatica</i> L. <i>Tilia sp.</i> L.	IX, XI; Serbia	20° – 60° 500 – 1000 m a.s.l. cambisol Rendzina
<i>Tuber rufum</i> var. <i>lucidum</i> (Bonnet) Montecchi & Lazzari according to Montecchi & Sarrasini (2000) (examined 3 specimens)	<i>Quercus cerris</i> L. <i>Carpinus betulus</i> L.	VII – X Serbia	20° – 30° 200 – 500 m a.s.l. cambisol
<i>Tuber rufum</i> var. <i>nitidum</i> (Vittad.) Montecchi & Lazzari according to Montecchi & Sarrasini (2000) (examined 6 specimens)	<i>Quercus cerris</i> L. <i>Quercus robur</i> L. <i>Carpinus betulus</i> L. <i>Populus alba</i> L. <i>Populus nigra</i> L.	I, VI, XI – XII; Serbia	0° – 20° 0 – 500 m a.s.l. alluvium cambisol

Distribution and ecological preferences of *Tuber* species in mid-west Balkan Peninsula

Neutral to weakly basic soils and presence of appropriate ECM forming tree host encourage the occurrence of *Tuber spp.* As the distribution of the host trees in Balkan Peninsula is strongly determined by altitude, exposition and soil water status, truffles are consequently following same distribution pattern as their host plants. While some species could be found in most forests dominated by ECM trees (*T. rufum*, *T. excavatum*, *T. aestivum*, and *T. brumale*), other species are restricted to areas defined by their exposition. Lowlands, especially the wide river valleys, and hilly regions in the mid-west of Balkan Peninsula generally host qualitatively and quantitatively different truffle communities. *Tuber macrosporum*, *T. magnatum*, *T. foetidum*, and *T. maculatum* were found to have their ecological optimum in Serbian lowland forests, while *T. borchii*, *T. oligospermum*, *T. fulgens*, and *T. mesentericum* show preferences for hills and mountains of mid-west Balkan Peninsula. Distribution and some general ecological features of truffle species that were registered at Balkan Peninsula are given in Tab. 1. Specific details on every species are described below.

Tuber rufum Pico (species concept according to Montecchi & Sarasini (2000))

Though never abundant, *T. rufum* is probably the most widely distributed species in all studied forests, but also in polluted city parks,

or near high-traffic roads (Tab. 1). In Serbia, all varieties described in Montecchi & Sarasini (2000) were recorded, but the most common variety by far was var. *rufum* Pico, whereas var. *lucidum* (Bonnet) Montecchi & Lazzari, was recorded only twice. In Montenegro and FYROM only var. *rufum* Pico was recorded.

***Tuber excavatum* Vittad.**

In the Balkan Peninsula, *T. excavatum* is very common and plant host and soil non-specific truffle (Tab.1). Ascoma can be abundant, ripening in early summer to autumn and occurring in both broadleaved and conifer dominated forests. Particularly preferring moist seasons, this species forms large ascoma (radius up to 6cm) at the upper elevation ranges. On the soil surface of the lowland forests in late spring, small (radius less than 1cm) and very abundant ascoma were registered, but were disappearing in early summer from the spot.

***Tuber aestivum* Vittad.**

T. aestivum is widely distributed throughout ECM supporting forests in the entire investigated area. *T. uncinatum* Chatin (Chevalier & Frochot 2002), lately signed as morphological form of *T. aestivum* (Weden *et al.* 2005; Mello *et al.* 2002), is the most abundant in the lime or oak dominated forests on slopes exposed to north or west. In damp places with Mediterranean oaks in Serbia, or in the European beech dominating mountain forests of Montenegro, mature ascoma of *T. aestivum* were collected even in late winter.

***Tuber brumale* Vittad.**

T. brumale was found to be abundant in all habitats in Serbia and Montenegro that could be expected to host *T. melanosporum* (Mediterranean forests of *Quercus pubescens* Wild., *Q. ilex* L., *Q. coccifera* L., *Ostrya carpinifolia* Scop. and *Carpinus orientalis* Mill., on the shallow or stony calcareous soils). It is also very common in the Mediterranean coast forests dominated by *Pinus pinea* L. and *P. halepensis* Ait. or in continental lowland forests dominated by *Quercus robur* L. and *Populus spp.* In early winter, when it reaches fructification maximum, *T. brumale* is the most ubiquitous species in Serbia and Montenegro. A distinct difference in fragrance could be observed between specimens found in aerated, limestone-rich soils in hills, and those found in thick clay-rich soils of lowlands.

***Tuber macrosporum* Vittad. and *Tuber magnatum* Pico**

In autumn, *T. macrosporum* and *T. magnatum* are dominant and abundant in the Serbian lowland, mixed forests (Tab. 1). Sporadically,

T. macrosporum was also collected in forests typical for the hilly regions of the continental part of Mesian Balkans (Tab.1, Kojić *et al.* 1998) where *T. magnatum* has never been recorded. *T. macrosporum* specimens collected in Serbia seem to have significantly larger dimensions than those described in literature (Szemere 1965; Pegler *et al.* 1993; Montecchi & Sarasini 2000; Riuosset *et al.* 2001), obtaining often the weight up to 50g. In conditions of the continental part of Balkan Peninsula, *T. magnatum* is rather associated with the native poplars (predominantly *Populus alba* L.) than oaks (except *Q. robur* L.) or hornbeams, and prefers wet, often occasionally flooded, thick, clay-rich soils in the lowlands. *T. brumale*, *T. macrosporum* and *T. magnatum*, that are common in natural lowland poplar forests, have never been found in introduced tree plantations or parks.

***Tuber foetidum* Vittad**

In lowland forests with even higher soil moisture (Tab.1), additional localities of *T. foetidum* were found, heretofore recorded only once in Serbia (Milenković & Marjanović 2001).

***Tuber maculatum* Vittad.**

In the course of investigation of the plantations of different introduced hybrid poplars in the Serbian lowlands in late autumn and early winter of the very moist 2004/2005 season, *T. maculatum* could be recorded, for the first time at Balkan Peninsula. This was the only truffle species that has ever been recorded in the hybrid poplar plantations in investigated area. In the same season it was also found near the planted trees in parks and quite rarely in natural lowland forests (Tab.1).

***Tuber borchii* Vittad.**

First time recorded for the Balkan Peninsula, *T. borchii* could be found only once in Serbia, on the Avala mountain (Belgrade county) in the mixed forest of *Quercus cerris* L., *Q. frainetto* Ten. with some introduced conifers. Continuous examination of *T. magnatum* habitats for the *T. borchii* revealed no results.

***Tuber oligospermum* (Tul. & C. Tul.) Trappe**

T. oligospermum could be found for the first time at Balkan Peninsula, but with only few collections. Ascoma were collected in late winter, in natural mixed deciduous forests (Tab.1) and in planted forest of *Pinus halepensis* Mill. (not native tree species for the area), on the limestone hills of Western Serbia. The species was also recorded in early spring in *Quercus ilex* L. dominating communities on coastal sandy soils of Montenegro. Serbian samples were previously misiden-

tified as *T. puberulum* (Tab. 2) due to untypical habitat, but when a typical example was found in Montenegro, it became clear that all collections were *T. oligospermum*, which could be confirmed by molecular analysis.

***Tuber mesentericum* Vittad.**

In mid-west Balkan Peninsula, *T. mesentericum* occurs in environments similar to those of the sister-species *T. aestivum*, but always in soils with potentially high CaCO₃ content (Tanasijević *et al.* 1965; 1966). This truffle is not common but can be abundant and dominant in suitable habitats, for example in *Tilia sp.* dominated forests on wet calcareous sandy soils.

***Tuber fulgens* Quél.**

In the similar ecosystems as *T. mesentericum*, *T. fulgens* was collected in Western Serbia, which was the first record for the Balkan Peninsula. It is ubiquitous in mixed forests of *Fagus sylvatica* L. at altitudes more than 1000 m (mountain Tara), but also occurs in *Tilia sp.*-dominated forests in North-East Serbia, always sharing its habitat with *T. mesentericum*. Even though of a very different origin, the soils in both localities were very calcareous (Marjanovic *et al.* unpublished).

T. borchii, *T. fulgens*, *T. macrosporum*, *T. magnatum*, *T. foetidum* and *T. maculatum* have by now been recorded only in Serbia. *T. oligospermum* was not recorded in FYROM. The rest of the species were present in all investigated countries. *T. melanosporum* Vitt., previously reported from Serbia (Milenković *et al.* 1992; Glamočlija *et al.* 1997) was not encountered in the present study. Revision of morphological and molecular characters of previously published and herbarised collections revealed that all can be assigned to *T. brumale*. Coastal and central areas of Montenegro were frequently searched for truffles but no additional species to the listed ones were discovered.

Molecular characterisation of truffles originating from the mid-west of Balkan Peninsula.

A total of 46 new ITS1-5.8S-ITS2 rDNA sequences of *Tuber* spp. were generated (for accession numbers see Tab. 2). Sequences were aligned with additional 29 truffle ITS sequences retrieved from GenBank for comparison and identification. Amplification of ITS-DNA of *T. mesentericum* was repeatedly unsuccessful, and therefore this species was not included in the phylogenetic analysis. A neighbour joining tree is given in Fig. 2. As the tree was primarily constructed to support morphological identifications, distances and relations between taxa do not represent in-depth phylogenetic relations and therefore truffle phylogeny or taxonomy will not be discussed.

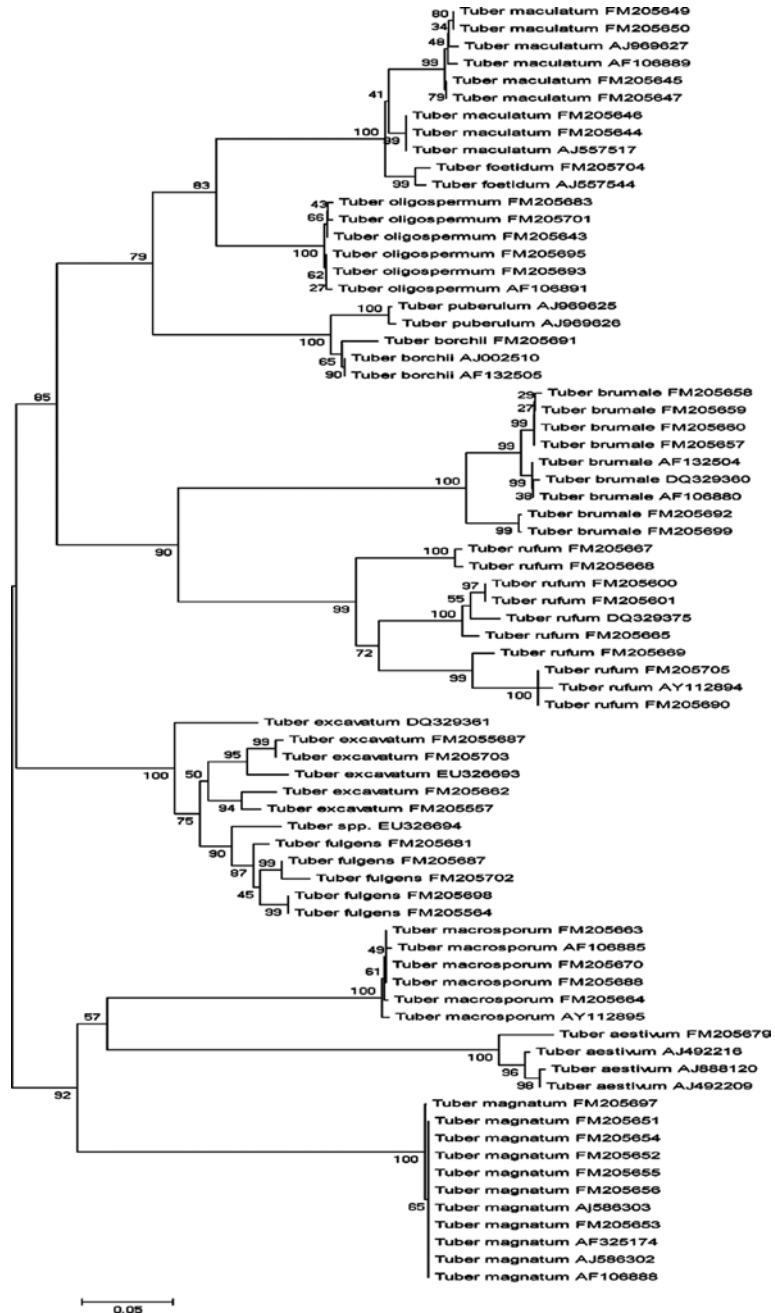


Figure 2. Unrooted NJ phylogenetic tree based on the alignment of ITS regions of truffles originating from Balkan Peninsula and an additional representative samples from other areas of Europe. Taxonomic names, GenBank accession numbers, and origin of the samples are listed in Tab. 2

Table 2. Morphological and molecular identifications of truffles recorded from Balkan Peninsula. GenBank accession numbers for rDNA ITS sequences included in the phylogenetic analysis are given.

Morphological determination	Identities according to BLAST search	GenBank	Geographic origin
Tuber puberulum Berk. & Broome	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205642	Dinaric Alps, West Serbia
Tuber puberulum Berk. & Broome	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205643	Dinaric Alps, West Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205644	West Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205645	West Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205646	North Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205647	Centa, Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205649	Obrenovac, Serbia
Tuber maculatum Vittad.	Tuber maculatum Vittad.	FM205650	Maljen, Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205651	West Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205652	West Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205653	Mid Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205654	Sumadija, Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205655	Sumadija, Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205656	Sumadija, Serbia
Tuber magnatum Pico	Tuber magnatum Pico	FM205697	West Serbia
Tuber brumale Vittad	Tuber brumale Vittad	FM205657	Montenegro
Tuber brumale Vittad	Tuber brumale Vittad	FM205658	Montenegro
Tuber brumale Vittad	Tuber brumale Vittad	FM205659	Montenegro
Tuber brumale Vittad	Tuber brumale Vittad	FM205660	Montenegro
Tuber brumale Vittad	Tuber brumale Vittad	FM205692	West Serbia
Tuber brumale Vittad	Tuber brumale Vittad	FM205699	West Serbia
Tuber macrosporum Vittad.	Tuber macrosporum Vittad.	FM205663	Belgrade, Serbia
Tuber macrosporum Vittad.	Tuber macrosporum Vittad.	FM205664	Central Serbia
Tuber macrosporum Vittad.	Tuber macrosporum Vittad.	FM205670	North Serbia
Tuber macrosporum Vittad.	Tuber macrosporum Vittad.	FM205688	West Serbia
Tuber rufum f. lucidum (Bonnet) Montecchi & Lazzari	Tuber rufum Pico	FM205665	Montenegro coast
Tuber rufum Pico	Tuber rufum Pico	FM205667	Tara, Serbia
Tuber rufum Pico	Tuber rufum Pico	FM205668	Tara, Serbia
Tuber rufum var. apiculatum E. Fisch	Tuber rufum Pico	FM205669	Tara, Serbia
Tuber rufum var. nitidum (Vittad.) Montecchi & Lazzari	Tuber rufum Pico	FM205677	Tara, Serbia
Tuber rufum var. rufum Pico	Tuber rufum Pico	FM205690	City park, Belgrade, Serbia
Tuber rufum var. rufum Pico	Tuber rufum Pico	FM205705	North Serbia
Tuber aestivum Vittad.	Tuber aestivum Vittad.	FM205679	West Serbia
Tuber fulgens Quél	no significant match	FM205681	Tara, Serbia
Tuber fulgens Quél	no significant match	FM205702	Tara, Serbia
Tuber fulgens Quél	no significant match	FM205698	West Serbia
Tuber fulgens Quél	no significant match	FM205685	West Serbia
Tuber excavatum Vittad.	Tuber excavatum Vittad.	FM205662	Tara, Serbia

Morphological determination	Identities according to BLAST search	GenBank	Geographic origin
Tuber excavatum Vittad.	Tuber excavatum Vittad.	FM205687	Ub, Serbia
Tuber excavatum	Tuber excavatum	FM205703	Tara, Serbia
Tuber oligospermum (Tul. & C. Tul.) Trappe	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205683	Montenegro
Tuber foetidum Vittad.	Tuber foetidum Vittad.	FM205704	West Serbia
Tuber borchii Vittad.	Tuber borchii Vittad.	FM205691	Avala, Serbia
Tuber puberulum Berk. & Broome	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205693	Dinaric Alps, West Serbia
Tuber puberulum Berk. & Broome	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205695	Dinaric Alps, West Serbia
Tuber sp.	Tuber oligospermum (Tul. & C. Tul.) Trappe	FM205701	West Serbia
Tuber fulgens Quél	no significant match	FM205564	Slovenia
Tuber sp.	no significant match	EU326694	Poland
Tuber excavatum Vittad.	Tuber excavatum Vittad.	FM205557	Slovenia
Tuber excavatum Vittad.	Tuber excavatum Vittad.	EU326693	Poland
Tuber excavatum Vittad.	Tuber excavatum Vittad.	DQ329361	Vaucluse, France
Tuber rufum Pico	Tuber rufum Pico	FM205600	Slovenia
Tuber rufum Pico	Tuber rufum Pico	FM205601	Slovenia
Tuber rufum Pico	Tuber rufum Pico	AY112894	Italy
Tuber maculatum Vittad	Tuber maculatum Vittad	AJ969627	Denmark
Tuber maculatum Vittad	Tuber maculatum Vittad	AF106889	Umbria, Italy
Tuber maculatum Vittad	Tuber maculatum Vittad	AJ557517	Hungary
Tuber foetidum Vittad	Tuber foetidum Vittad	AJ557544	Hungary
Tuber puberulum Berk. & Broome	Tuber puberulum Berk. & Broome	AJ969625	Suserup, South Zealand, Denmark:
Tuber puberulum Berk. & Broome	Tuber puberulum Berk. & Broome	AJ969626	Bromme Plantage, South Zealand, Denmark:
Tuber borchii Vittad.	Tuber borchii Vittad.	AJ002510	Italy
Tuber borchii Vittad.	Tuber borchii Vittad.	AF132505	France
Tuber oligospermum (Tul. & C. Tul.) Trappe	Tuber oligospermum (Tul. & C. Tul.) Trappe	AF106891	Italy
Tuber brumale Vittad.	Tuber brumale Vittad.	AF106880	Marche, Italy
Tuber brumale Vittad.	Tuber brumale Vittad.	DQ329360	Vaucluse, France
Tuber brumale Vittad.	Tuber brumale Vittad.	AF132504	France
Tuber macrosporium Vittad.	Tuber macrosporium	AF106885	Umbria, Italy
Tuber macrosporium Vittad.	Tuber macrosporium	AY112895	Italy
Tuber magnatum Pico	Tuber magnatum Pico	AJ586303	Piedmont, Italy
Tuber magnatum Pico	Tuber magnatum Pico	AF325174	Italy
Tuber magnatum Pico	Tuber magnatum Pico	AJ586302	Piedmont, Italy
Tuber magnatum Pico	Tuber magnatum Pico	AF106888	Umbria, Italy
Tuber aestivum Vittad.	Tuber aestivum Vittad.	AJ888120	United Kingdom
Tuber aestivum Vittad.	Tuber aestivum Vittad.	AJ492209	Campobasso, Italy
Tuber aestivum Vittad.	Tuber aestivum Vittad.	AJ492216	Teruel, Spain

Discussion

Distribution and ecological preferences of *Tuber* species in mid-west Balkan Peninsula

The number of native species of *Tuber* reported for Europe ranges from 22 to 32, depending on the species interpretations of various authors (Montecchi & Sarasini 2000; Riuosett *et al.* 2001; Ceruti *et al.* 2003). The list of twelve *Tuber* species recorded for the investigated part of Balkan Peninsula is notable, when the relatively small area of investigation is taken into consideration. Among these, few species could be regarded as widespread throughout Europe: *T. rufum*, *T. aestivum*, *T. excavatum*, *T. borchii* and *T. maculatum* (Szemere 1965; Chevalier *et al.* 1986; Wojewoda & Lawrinowicz 1986; Pázmány 1991; Pegler *et al.* 1993; Montecchi & Sarasini 2000; Weden *et al.* 2000; Riuosett *et al.* 2001; Ceruti *et al.* 2003; Kers 2003; Jeandroz *et al.* 2008). The ecological features of these wide spread species in the investigated area, resembled those reported from other European areas (Szemere 1965; Wojewoda & Lawrinowicz 1986; Pázmány 1991; Pegler *et al.* 1993; Montecchi & Sarasini 2000; Riuosett *et al.* 2001; Chevalier & Frochot 2002; Ceruti *et al.* 2003). *Tuber mesentericum* and *Tuber fulgens* are restricted to South and Central Europe (Pegler *et al.* 1983; Ceruti *et al.* 2003; Jeandroz *et al.* 2008). Our results suggest that sister-species of *T. aestivum* and *T. excavatum*, represented by *T. mesentericum* and *T. fulgens* respectively, are adapted to the similar environments, but grow in soils of high CaCO₃ content.

In comparison to the rest of the Europe, the peculiarity of the mid-west Balkans in respect to the occurrence of *Tuber* species, is rarity of *T. borchii*, *T. maculatum* and *T. foetidum*, and the absence of *T. puberulum* and *T. dryophyllum*, which have been reported as common all over continental Europe and British Isles (Szemere, 1965; Pegler *et al.* 1993; Ceruti *et al.* 2003; Jeandroz *et al.* 2008). The Balkan Peninsula seems not to be a suitable environment for small white truffles. In the ecological constraints of the Balkan Peninsula, *T. maculatum* is often a pioneer species that may help ECM forming trees to establish in adverse environments (plantations, parks). As this species is assumed to be very tolerant to unfavourable environmental conditions (Montecchi & Sarasini 2000; Ceruti *et al.* 2003), it is not clear why is it found so rarely at Balkan Peninsula. Indeed, *T. oligospermum*, that has been assumed to be connected to Mediterranean ecosystems and climate (Montecchi & Sarasini; 2000; Riuosett *et al.* 2001; Ceruti *et al.* 2003; Jeandroz *et al.* 2008), is common in the continental conditions of Serbia.. Even if its habitats in Serbia resemble those from Mediterranean zones in matters of vegetation, soils and climate are quite different, particularly the winter season. It can be postulated that this truffle is more dependent on specific host-plant species than on abiotic factors,

as thought previously (Montecchi & Sarasini 2000, Ceruti *et al.* 2003). *T. foetidum*, reported to be distributed in Western Europe (Jeandroz *et al.* 2008) was collected few times at Balkan Peninsula.

All European commercially important truffles, except *T. melanosporum* and *T. borchii*, are widespread in Serbia. While in Southern Europe and bordering regions *T. brumale* and *T. macrosporum* are rather common, there are only few records in other parts of Europe (Pegler *et al.* 1993, Rioussset *et al.*, 2001, Ceruti *et al.* 2003). As unusually big *T. macrosporum* ascoma were recorded often in Serbian lowland forests, this species probably has its distribution optimum in the warmer climates, on the sites where soil water is not strongly limited. Even if the climate, soils and vegetation in Montenegro and Western Serbia resembles conditions optimal for *T. melanosporum*, this species has never been recorded in investigated area. Instead, *T. brumale* ascoma of a very good quality and unusual smell was recorded on hills with calcareous stony soils and Mediterranean vegetation. The historical reasons could be postulated as explanation – when following the routes of oaks recolonization after the last ice age, no oak haplotype that was carrying *T. melanosporum* was registered to originate from the Balkan Peninsula (Murat *et al.* 2004).

T. magnatum was assumed to be primarily distributed in Central and Northern Italy, and in Istria, Croatia (Hall *et al.* 1998, Ceruti *et al.* 2003; Mello *et al.* 2005), with some additional records from south-east of France and Ticino Canton in Switzerland (Lawrynowicz 1993; Zambonelli & Di Munno, 1991), but there are also records published from Serbia (Marjanović & Milenković 1998), Hungary (Bratek *et al.* 2003) and Slovenia (Piltaver & Ratoša 2006). Lowland ecosystems where *T. magnatum* is usually found in Serbia (clay rich, often flooded soils, Tab.1) are clearly different from hilly sites reported from Italy (silty, well aerated soils, Lulli & Primavera 2000). It appears that, the so called “Piemont” white truffle has much wider area of distribution and less restricted ecological requirements than previously claimed (Hall *et al.* 1998, Ceruti *et al.* 2003). However, much more detailed study is required for final conclusions on the ecological limits of this species.

In comparison to other European regions, with high truffle diversity (Ceruti *et al.* 2003), the western Balkan Peninsula is specific for truffle chorology and ecology in a few points: *T. magnatum* is widespread all over river valleys in clay-rich soils, but has never been recorded on hills and well aerated soils; *T. borchii*, *T. maculatum* and *T. foetidum* are very rare, while *T. puberulum* and *T. dryophylum* by now have not been recorded; *T. oligospermum* is not specific for Mediterranean coast-lines only – it is common at the interior continental parts of the peninsula as well; some true Mediterranean species (*T. asa*, *T. belonae*, *T. panniferum*, *T. malenconii*), as well as *T. melanosporum* are by now missing.

Molecular diversity of truffles in the mid-west of Balkan Peninsula

Phylogenetic analysis of our sequence data together with data from GenBank revealed a neighbour joining tree with an over all similar topology of trees already published (Jeandroz *et al.* 2008). Morphological determination of some small white truffles did not appear to be in accordance with molecular data. Two samples initially determined as *T. puberulum* Berk. & Broome due to their continental habitat, appeared to be morphologically very similar *T. oligospermum* (Tul. & C. Tul.) Trappe, the species that had been previously collected only in Mediterranean regions (Cerruti *et al.* 2003). Two very small herbarium samples determined as *T. borchii* (according to Montecchi & Sarasini 2000) belonged to the *T. maculatum* clade in the NJ-tree. Halász *et al.* (2005) reported almost clear correlation between morphological and molecular determination of small white truffles in Hungary. However, other authors that compared sequences from the other European regions, could not observe this correlation (Wang *et al.* 2007, Jeandroz *et al.* 2008). As all these analyses (including our) were performed on quite a small number of sequences, we suggest that more detailed analysis of small white truffles on the global level is necessary for clarifying species delimitation, and realistic phylogenetic relationships.

As reported before (Iotti *et al.* 2007; Wang *et al.* 2007), *T. rufum* appeared to be genetically very diverse species, but morphological forms (varieties) described by Montecchi & Sarasini (2000) were not supported by the molecular analysis (Fig. 2). *T. excavatum* and *T. fulgens* were not clearly separated in the NJ-tree. More specimens of both taxa must be investigated by morphological and molecular methods for a clear decision if *T. fulgens* is a distinct species, or just a variety of *T. excavatum*. From our analysis it seems likely that *Tuber sp.* published from Poland by Hilszczanska *et al.* 2008 (Acc. No. EU326694 Tab.1, Fig. 2) is *T. fulgens*.

We have observed minor, two base pairs variation within the ITS regions of *T. magnatum* (Acc. No. FM205697, Tab. 2) originating from Serbia (data not shown). This could be important as no ITS diversity was detected in samples from Italy and Istria by Mello *et al.* (2005) or Rubini *et al.* (2005). Still, much more sequences should be investigated in order to confirm possible diversity in ITS region of *T. magnatum*.

Comparing truffles originating from mid-west Balkan Peninsula with those from other parts of Europe, it seems likely that some species differ in their favoured ecological conditions, but only slight differences in the ITS sequences could be detected. An adaptation to the specific climatic, edaphic and vegetational conditions of the Balkan Peninsula can be speculated, due to climatic changes throughout European bio-geographic history. During periods of glaciation, there might have been refugial sites of truffles and their host plant hosts at the Balkan Peninsula (Petit *et al.* 2002a,b; Murat *et al.* 2004). The list

of truffles from the mid-west of Balkan Peninsula will be expanded with additional collectings in an extended area. More species can be expected from regions with strong Mediterranean influences, together with mountains and canyons, which could have been potential glacial refugia.

Acknowledgements

The present work was mostly financed by contributing authors and partially from EU supported Market oriented R&D project EU-REKA E!3835. Authors are grateful to all truffle hunters that contributed to the collection of specimens, especially to Mr. Momčilo Vićentijević and Mr. Ljubiša Milojević. We are especially thankful to Dr James Trappe for literature supply, support and critical reading of the manuscript, and to Dr Gerard Chevalier and Dr Zoltan Bratek for organizing several discussions on the subject. Dr Branko Karadžić provided map presented in this work, and we are deeply grateful to him for a lasting and successful collaboration. For logistic support we are thankful to Prof Dr. Željko Vučinić and Prof. Dr Hojka Kraigher.

References

- Alvarez I.F., Parlade J., Trappe J.M., Castellano M.A. (1993). Hypogeous mycorrhizal fungi of Spain. *Mycotaxon* **67**: 201–217.
- Atlas klime SFRJ (1969). Savezni hidrometeorološki zavod, Beograd,
- Bennet K.D., Tzedakis P.C., Willis K.J. (1991). Quaternary refugia of north European trees. *Journal of Biogeography* **18**: 103–115.
- Bougher N.L., Lebel T. (2001). Sequestrate (truffle-like) fungi of Australia and New Zealand. *Australian Systematic Botany* **14**, 439–484.
- Bratek Z., Gogan A., Halász K., Bagi I., Erdei V., Bujaku, G. (2004). The Northeast habitats of *Tuber magnatum* known from Hungary. Le premier symposium sur les champignons hypogés du basin méditerranéen, 6–8 Avril, Rabat.
- Ceruti A., Fontana A., Nosenzo, C. (2003). *Le Specie Europee del genere Tuber—Una Revisione Storica*. Museo Regionale de Scienze Naturali, Torino.
- Chevalier G., Rioussel L., Dupre, C. (1986). Taxonomie des truffes Européennes. In V. Gianinazzi-Pearson and S. Gianinazzi (eds.), *Proceedings of the 1st European Symposium on Mycorrhizae, Dijon, 1–5 July 1985* pp. 631–635. Paris; INRA.
- Chevalier G., Frochot, H. (2002). *La Truffe de Bourgogne* (*Tuber uncinatum* Chatin). Petrarque, Levallois – Perret Cedex.
- Doyle J.J., Doyle J.L. (1990). Isolation of plant DNA from fresh tissue. *Focus* **12**: 13–15.
- Frančičković S. (1950). Naši tartufi. *Šumarski List* **74**: 23–38.
- Gandeboeuf D., Dupre C., Roeckel-Drevet P., Nicolas P., Chevalier G. (1997). Grouping and identification of *Tuber* species using RAPD markers. *Canadian Journal of Botany* **75**: 36–45.
- Gardes M., Bruns T.D. (1993). ITS primers with enhanced specificity for basidiomycetes – application to the identification of mycorrhizae and rusts. *Molecular Ecology* **2**: 113–118.
- Glamočlija J., Vujičić R., Vukojević J. (1997). Evidence of truffles in Serbia. *Mycotaxon* **LXV**: 211–22.

- Grebenc T., Piltaver A., Kraigher, H. (2000). Establishment of the PCR-RFLP library for *Basidiomycetes*, *Ascomycetes* and their ectomycorrhizae on *Picea abies* (L.) Karst. *Phyton, Annales rei Botanicae* **40** (4): 79–82.
- Hall I.R., Zambonelli A., Primavera F. (1998). Ectomycorrhizal Fungi with Edible fruiting Bodies 3. *Tuber magnatum*, *Tuberaceae*. *Economic Botany* **52**, No. 2: 192–200.
- Halász K., Bratek Z., Szegoe D., Rudnóy S., Rác I., Lásztity D., Trappe J.M. (2005). Tests of species concepts of the small, white, European group of *Tuber* spp. based on morphology and rDNA ITS sequences with special reference to *Tuber rapaeodorum*. *Mycological Progress* **4** (4): 281–290.
- Henrion B., Chevalier G., Martin F. (1994). Typing truffle species by PCR amplification of the ribosomal DNA spacers. *Mycological Research* **98**: 37–43.
- Hilszczanska D., Sierota Z., Palenzona M. (2008). New *Tuber* species found in Poland. *Mycorrhiza* **18**: 223–226.
- Hrka J. (1988). *Tartufi – poznavanje, branje, uzgoj, pripremanje*. Mladost, Zagreb.
- Iotti M., Amicucci A., Bonito G., Bonuso E., Stocchi V., Zambonelli A. (2007). Selection of a set of primers for the identification of *Tuber rufum*: a truffle species with high genetic variability. *FEMS Microbiology Letters* **277**: 223–231.
- Jeandroz S., Murat C., Wang Y., Bonfante P., Le Tacon F. (2008). Molecular phylogeny and historical biogeography of the genus *Tuber*, the ‘true truffles’. *Journal of Biogeography* **35** (5): 815–829.
- Kers L.E. (2003). Tryfflarna *Tuber aestivum* och *T. mesentericum* i Sverige. [*Tuber aestivum* Vittad. and *T. mesentericum* Vittad. (Ascomycetes) in Sweden.]. *Svensk Botanisk Tidskr* **97**: 157–175.
- Kojić M., Popović R., Karadžić B. (1997). *Vaskularne biljke Srbije kao indikatori staništa*. Institut za istraživanja u poljoprivredi „Srbija“ i Institut za biološka istraživanja „Siniša Stanković“, Beograd, Serbia.
- Kraigher H., Javornik B., Agerer, R. (1995). Ectomycorrhizae of *Lactarius lignyotus* on Norway spruce, characterized by anatomical and molecular tools. *Mycorrhiza* **5** (3): 175–180.
- Larkin M.A., Blackshields G., Brown N.P., Chenna R., McGettigan P.A., McWilliam H., Valentin F., Wallace I.M., Wilm A., Lopez R., Thompson J.D., Gibson T.J., Higgins D.G. (2007). ClustalW and ClustalX version 2. *Bioinformatics* **23** (21): 2947–2948.
- Lulli L., Primavera F. (2000). *Tuber magnatum* Pico: environment of growth. *The 5th International Congress Science and Cultivation of Tuber and other edible hypogeous mushrooms Proceedings, Section Ecology*: 5.269–5.272.
- Lindtner V. (1935). Podzemne gljive u Srbiji. *Šumarski List* **59**: 15–18.
- Marjanović Ž., Milenković M. (1998). *Tuber magnatum* Pico and some similar species in Yugoslavia, *Abstracts of Second International Conference of Mycorrhiza*, Uppsala, Sweden: 114.
- Mello A., Cantisani A., Vizzini A., Bonfante, P. (2002). Genetic variability of *Tuber uncinatum* and its relatedness to other black truffles. *Environmental Microbiology* **4** (10): 584–594.
- Mello A., Murat C., Vizzini A., Gavazza V., Bonfante, P. (2005). *Tuber magnatum* Pico, a species of limited geographical distribution: its diversity inside and outside a truffle ground. *Environmental Microbiology* **7** (1): 55–65.
- Milenković M., Glamočlija J., Veljković V., Vukojević, J. (1992). Record of two *Tuber* (*T. aestivum* and *T. melanosporum*) species in Serbia. *Archives of Biological Sciences* **44**: 223–28.
20. Milenković, M. & Marjanović, Ž. (2000). Current results on *Tuber* spp. research in Yugoslavia. *The 5th International Congress Science and Cultivation of Tuber and other edible hypogeous mushrooms Proceedings, Section Ecology*: 4.218–4.225.

- Montecchi A., Sarasini, M. (2000). *Funghi ipogei d'Europa*. Associazione Micologica Bresadola – Fondazione centro Studi Micologici, Trento-Vicenza.
- Murat C., Díez J., Luis P., Delaruelle C., Dupré C., Chevalier G., Bonfante P., Martin F. (2004). Polymorphism at the ribosomal DNA ITS and its relation to post-glacial re-colonization routes of the Perigord truffle *Tuber melanosporum*. *New Phytologist* **164** (2): 401–411.
- Myers N., Cowling R.M. (1999). *Mediterranean Basin*. In: *Hotspots: Earth's Biologically Richest and Most Endangered Terrestrial Ecoregions* (R.A. Mittermeier, N. Meyers, P. Robles Gil, and C.G. Mittermeier eds). CEMEX, Mexico 254–267.
- Pázmány D. (1990/1991). Conspectus fungorum hypogaeorum Transsilvaniae. *Notes on Botany, Horticulture and Agrobotany Cluj XX–XXI*.
- Pegler D.N., Spooner B.M., Young T.W.K. (1993) *British Truffles*. Royal Botanic Garden, Kew, London
- Petit R.J., Csaikl U.M., Bordács S., Burg K., Coart E., Cottrell J., van Dam B., Deans J. D., Dumolin-Lapègue S., Fineschi S. *et al.* (2002a). Chloroplast DNA variation in European white oaks: Phylogeography and patterns of diversity based on data from over 2600 populations. *Forest ecology and Management* **156**: 5–26.
- Petit R.J., Brewer S., Bordács S., Burg Cheddadi R.K., Coart E., Cottrell J., Csaikl U.M., van Dam B., Deans J.D., *et al.* (2002b). Identification of refugia and post-glacial colonisation routes of European white oaks based on chloroplast DNA and fossil pollen evidence. *Forest ecology and Management* **156**: 49–74.
- Piltaver A., Ratoša I. 2006. A contribution to better knowledge of hypogeous fungi in Slovenia. *Gozdarski vestnik*, **64** (7/8): 303–312, 329–330.
- RiOUSset L., RiOUSset G., Chevalier G., Bardet M.E. (2001). *Truffles d'Europe et de Chine*. INRA, Paris.
- Rogers S.O., Bendich A.J. (1985). Extraction of DNA from milligram amounts of fresh, herbarium and mummied plant tissues. *Plant Molecular Biology* **5**: 69–76
- Roux C., Sejalon-Delmas N., Martins M., Parguey-Leduc A., Dargent R., Becard G. (1999). Phylogenetic relationships between European and Chinese truffles based on parsimony and distance analysis of ITS sequences. *FEMS Microbiology Letters* **180**: 147–155.
- Rubini A., Paolocci F., Riccioni C., Vendramin G.G., Arcioni S. (2005) Genetic and Phylogeographic Structures of the Symbiotic Fungus *Tuber magnatum*, *Applied and Environmental Microbiology* **71** (11): 6584–6589.
- Simpson J.A. (2000). More on mycophagous birds. *Australasian Mycologist* **19**: 49–51.
- Stevanović V., Jovanović S., Lakušić D., Niketić, M. (1995). *Diverzitet vaskularne flore Jugoslavije sa pregledom vrsta od međunarodnog značaja*. In *Biodiverzitet Jugoslavije* (Stevanović, V I V. Vasić eds). Biološki Fakultet Univerziteta u Beogradu i Ecolibri, Beograd.
- Szemere L. (1965). *Die untererdishen Pilze des Karpatenbeckens*. Akademiai Kiado. Budapest.
- Taberlet P., Fumagalli L., Wust-Saucy A.G., Cossons J-F. (1998). Comparative phylogeography and postglacial colonization routes in Europe. *Molecular Ecology* **7**: 453–464.
- Tamura K, Dudley J, Nei M and Kumar S (2007). *MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0*. *Molecular Biology and Evolution* **24**: 1596–1599.
- Tanasijević Đ., Antonović G., Kovačević R., Aleksić Ž., Popović Ž. (1965). *Zemljišta basena Velike Morave i Mlave* Institut za proučavanje zemljišta. Beograd.

- Tanasijević Đ., Antonović G., Aleksić Ž., Pavičević N., Filipović Đ, Spasojević M. (1966). *Pedološki pokrivač Zapadne i Severozapadne Srbije*. Institut za proučavanje zemljišta. Beograd.
- Wang Y., Tan Z.M., Murat C., Jeandroz S., Le Tacon F. (2007). Molecular taxonomy of Chinese truffles belonging to the *Tuber rufum* and *Tuber puberulum* groups. *Fungal Diversity* **24**: 301–328.
- Weden C., Danell E., Camacho F.J., Backlund A. (2004). The population of the hypogeous fungus *Tuber aestivum* syn. *T. uncinatum* on the island of Gotland. *Mycorrhiza* **14** (1): 19–23.
- Wedén C., Danell E., Tibell, L. (2005). Species recognition in the truffle genus *Tuber* – the synonyms *Tuber aestivum* and *Tuber uncinatum*. *Environmental Microbiology* **7** (10): 1535–1546.
- White T.J., Bruns T., Lee S., Taylor J. (1990). *Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics*. In: *PCR Protocols. A guide to methods and applications*. Innis, M.A., Gelfand, D.H., Sninsky, J.J. & White, T.J. (Eds.). San Diego, Academic Press, p. 315–322.
- Wojewoda W., Lawrinowicz, M. (1986). *Red list of the threatened macrofungi in Poland*. In: *List of threatened plants in Poland* (Zarzyckiego K and W. Wojewoda eds) Polaska Akademia Nauk, Komitet Ochrony Przyrody & Institut Botaniki, PWN Warszawa.
- Zambonelli A., Riveti C., Percudani R., Otonello, S. (2000). TUBERKEY, A computer program for the description and recognition of truffles. *Mycotaxon* **74**: 57–76 (<http://www.truffle.org/tuberkey/html>).

(Manuscript accepted 2 Nov 2009; Corresponding Editor: M. Kirchmair)

ZOBODAT - www.zobodat.at

Zoologisch-Botanische Datenbank/Zoological-Botanical Database

Digitale Literatur/Digital Literature

Zeitschrift/Journal: [Sydowia](#)

Jahr/Year: 2010

Band/Volume: [062](#)

Autor(en)/Author(s): Marjanovic Z., Grebenc Tine, Markovic M. M., Glisic A., Milenkovic M.

Artikel/Article: [Ecological specificities and molecular diversity of truffles \(genus Tuber\) originating from mid-west of the Balkan Peninsula. 67-87](#)