# Evaluation of antagonistic of *Trichoderma* spp. against *Fusarium* oxysporum f. sp. lycopersiciand *Fusarium* oxysporum f. sp. ciceri in in vitro

Preeti Sonkar

Research Scholar, Department ofBiological Sciences, Mahatma Gandhi Chitrakoot Gramodaya Vishwavidyalaya, Chitrakoot, Satna (M.P.) - 485334.India

Received: June 25, 2018

Accepted: August 06, 2018

**ABSTRACT** The objective of this paper was evaluated to antagonistic of 14 different strain of Trichoderma species against Fusarium oxysporum f. sp. lycopersiciand Fusarium oxysporum f. sp. ciceri in in vitro. However, Fusarium wilt is a disease and one of the major yield limited factorswhich are harmful to economic important crop (Tomato and Chickpea). The antagonist of Trichoderma had been determined base on dualand its interaction. Here, II-method of dual culture was measured by Percent Growth Inhibition (Korsten L.) and MIC (Sangoyomi T.). Consituently, Invitro result of all Trichoderma strain was excellent as biological control agents due to PGI value more than 80%. However, Trichoderma virens(96.27%) was reduced to mycelium of Fusarium oxysporum f. sp. lycopersici as compared to other strain. In another hand, Trichoderma harzianumand Trichoderma viride(93.75%) was reduced to Fusariumoxysporum f. sp. ciceri. Therefore, Scale of MIC values were effective for all strain of Trichoderma according to Sangoyomi T.Conclusively, finding of the study indicates that different strain of Trichoderma has varied degree of antagonistic for both pathogens.

Keywords: Trichoderma strain, Fusarium wilts, PGI, MIC, antagonist etc.

#### 1. Introduction

This crop, (Chickpea and Tomato) is also very important in diet against common disease and economic value for farmers, Soil-borne pathogens inflict a lot of disease and economic vield loss (Babalola and Glick,2012). Out of phytopathogens, specially, wilt disease caused by species of *Fusarium* remain to be a challenging task in terms of management (Barari,2016). Disease of wilt is a destructive mechanism in regard economically important crops caused by the soil borne fungus *Fusariumoxysporum*(Singha et al. 2016). Fusarium oxysporum Schlechtend Fr. f. sp. ciceri (Padwick) Matuo & K. Sato and Fusarium oxysporum f. sp. lycopersici (Sacc.) W.C. Snyder and H.N. Hans (FOL), is the most important soil-borne disease throughout the particularly world and in the Indian Subcontinent.It is a cosmopolitan, soil pathogen and saprophyte that feed on dead and decaying organic matter. Therefore, Fusarium oxysporum f. sp. ciceri and Fusarium oxysporum f. sp. lycopersici, is not easily to control and used of chemicals causes threat to environment and crop quality. However, biocontrol is the effective (Sonawane, Mahajan, and Renake 2015; Brandler et al. 2017) with best substitution at the place chemical. Recently, interest in biological control of soil borne fungus Fusariumoxysporum has increased and effort to find alternatives to use of

chemicals (Akila et al. 2011; Vinale et al. 2014). Recently.Soil-dwelling filamentous fungy. Trichoderma spp. are used in agriculture as biofungicides and induction of plant defense. Morever, Mycoparasitism of Trichoderma are considered to be the most important mechanisms, known as Trichoderma-mediated biological control (Mukherjee et al. 2012, Cucinotta 2014). Therefore, Trichoderma spp. is chosen to be the most promising biocontrol agents of plant diseases might be exploited for suitable disease management programs to save environmental risk.

The control of Fusarium oxysporum can be done by most promising *Trichoderma* spp., widely used as putative BCAs. In the present study, the biological potential of Trichoderma strain was evaluated with in in vitro experiments against two different Fusarium wilt pathogen (Fusarium f. *lycopersiciand* oxysporum sp. Fusarium oxvsporum f. sp. ciceri) (Altinok and Erdogan, 2015). Understanding the interaction of Trichoderma-Fusarium oxysporum in invitro, would be help in developing strains with superior biocontrol properties.

#### 2.Materials and methods

#### 2.1. Source of Trichoderma isolates

*Trichoderma* spp. were collected from rhizosphere zone of healthy plant tomato and chickpea plant according to Akhtar (1996) than

Purified on PDA plate according to watts et. al. and Rifai. After 5 days incubate. growing Microflora was observe under compound microscope, identified with available literature Rifai and Webster (1969), Barnett (1998) and identified Trichoderma spp. were purified by hypal tip culture technique and preserved in 5°C refrigerator at for further use (Mustafaet.al., 2009; Khanget.al. 2013).

## 2.2. Isolation and identification of *Fusarium* oxysporum

*F. oxysporum* was isolated from both infected tomato and chickpea plants according to Rahman *et.al.*2009; Ignjatov *et al.* 2012.and growing Microflora was observe under compound microscope,identified with available literature. All culture was preserved at 4 ° C until further study.

#### 2.3. Dual test

*Trichoderma* strains were tested against to both isolate *Fusarium oxysporum* from tomato and chickpea, known as *Fusarium oxysporum* f. sp. *lycopersici*(FOL) and *Fusarium oxysporum* f. sp. *ciceri*(FOC) based on host. The screening of antagonist was based on dual according to method of Dickinson and Skidmore (1976) and Rahman *et.al.*2009. Each antagonist and pathogen was set up in II-method according to Rahman *et.al.*2009 with triplicated. The inoculated plates were incubated at  $24\pm2^{\circ}$ C with a photoperiod.

Percent Growth Inhibition (PGI) of pathogen was also calculated as describe by korsten L.

## $PGI = \frac{R_1 - R_2}{R_1} \times 100$

 $R_1$  = Mycelium growth of both pathogen (*Fusarium oxysporum* f. sp. *lycopersici*(FOL) and *Fusarium oxysporum* f. sp. *ciceri*(FOC) without *Trichoderma* sp.

 $R_2$  = Mycelium growth of both pathogen (*Fusarium oxysporum* f. sp. *lycopersici*(FOL) and *Fusarium oxysporum* f. sp. *ciceri*(FOC) with *Trichoderma* sp.

After PGI was calculated, used to value in MIC (Minimum Inhibition Concentration) as measure effective in inhibition of Mycelium growth of both pathogen (*Fusarium oxysporum* f. sp. *lycopersici*(FOL) and *Fusarium oxysporum* f. sp. *ciceri*(FOC). Scale of measure was given by Sangoyomi T. as below (Nwankiti and Gwa; 2018)

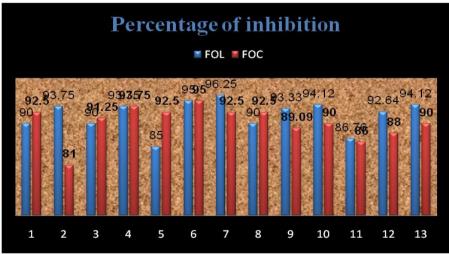
 $\leq 0\%$  Inhibition (Not effective)

> 0-20% Inhibition (Slightly effective)
>20-50% Inhibition (Moderately effective)
>50-<100% Inhibition (effective)</li>
100% Inhibition (Highly effective)

#### **Statistical Analysis**

All the data were statistically analyzed using by SPSS. Differences among treatments were determined by XLSTAT. Data are presented as mean values.

#### 3. Result and discussion



Pic.1. Percentage of inhibition represented between two pathogen Fol and Foc.

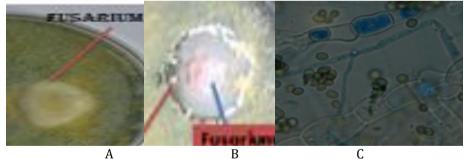
Here, we had been used to two type pathogen and monitored of percentage of inhibition based on dual technique. All Result of percentage inhibition was more than 80%. Thus all Trichoderma strain was excellent effective by measuring scale of Sangoyomi T. However, *T. virens*(2) was maximum (96.25) percentage of inhibition in case Fol, in compared to other species while *T. harzianum* (3) and *T. piluliferum* (3) was second highest (94.12) in % of inhibition. In case Foc, *T. harzianum* (2) was maximum 95% percentage of inhibition where *T. harzianum* (1) and *T. viride* (2) was

93.75. *T. harzianum* (1) and (2) was same for both pathogen Fol and Foc.

Data are availabled according to list of Table.1.with Pic.1.

I able.1. PGI % and MIC of among Tricnoderma strain       S     News					
S.	Name	PGI %	MIC (FOL)	PGI%(FOC)	MIC(FOC)
No.	<i></i>	(FOL)			70 1000/
1.	Trichoderma	90	>50-<100% Inhibition	92.5	>50-<100%
	virense(1)		(effective)		Inhibition (effective)
2.	Trichoderma	93.75	>50-<100% Inhibition	81	>50-<100%
	viride(1)		(effective)		Inhibition (effective)
3.	Trichoderma	90	>50-<100% Inhibition	91.25	>50-<100%
	piluliferum(1)		(effective)		Inhibition (effective)
4.	Trichoderma	93.75	>50-<100% Inhibition	93.75	>50-<100%
	harzianum(1)		(effective)		Inhibition (effective)
5.	Trichoderma	85	>50-<100% Inhibition	92.5	>50-<100%
	piluliferum(2)		(effective)		Inhibition (effective)
6.	Trichoderma	95	>50-<100% Inhibition	95	>50-<100%
	harzianum		(effective)		Inhibition (effective)
	(2)				
7.	Trichoderma	96.25	>50-<100% Inhibition	92.5	>50-<100%
	virens(1)		(effective)		Inhibition (effective)
8.	Trichoderma	87.5	>50-<100% Inhibition	93.75	>50-<100%
	viride(2)		(effective)		Inhibition (effective)
9.	Trichoderma	90	>50-<100% Inhibition	92.5	>50-<100%
	harzianum(3)		(effective)		Inhibition (effective)
10.	Trichoderma	93.33	>50-<100% Inhibition	89.09	>50-<100%
	erinaccum		(effective)		Inhibition (effective)
11.	Trichoderma	94.12	>50-<100% Inhibition	90	>50-<100%
	harzianum(4)		(effective)		Inhibition (effective)
12.	Trichoderma	86.76	>50-<100% Inhibition	86	>50-<100%
	harzianum(5)		(effective)		Inhibition (effective)
13.	Trichoderma	92.64	>50-<100% Inhibition	88	>50-<100%
	harzianum(6)		(effective)		Inhibition (effective
14.	Trichoderma	94.12	>50-<100% Inhibition	90	>50-<100%
	piluliferum(3)		(effective)		Inhibition (effective)

### Table.1. PGI %and MIC of among Trichoderma strain



**Pic.2. A and B**: Type of interaction of *Trichodermaln vitro* (Dual) and **C**: Mycelium of pathogen and *Trichoderma* with conidia

## 3.1. Isolation and Identification both *Trichoderma* and pathogen

Number of isolate *Trichoderma* was 14.Method of Rifai was confirmed Species and also included ITCC, New Delhi. Both pathogens were *Fusarium oxysporum* f. sp. *lycopersici*(FOL) and *Fusarium oxysporum* f. sp. *ciceri*(FOC) which was isolated from infected plant. Identify to Pathogen and *Trichoderma* was based microscopic studies which are presented in Pic.2.C. Mycelium of *Fusarium oxysporum*was thick comparison to mycelium of *Trichoderma* strain, this work is supported by Poornimasharma, 2011, Cordova-Albores *et.al.* 2016.

#### 3.2. Screening of Dual test

*Trichoderma* species was first screened and evaluated in term for antagonistic ability. AccordingPic.1and Table.1, In this case of

Fusarium oxysporum f. sp. lycopersici, highest PGI values were recorded as 96.25% for *T. virens* in comparison other *Trichoderma* strain and lowest PGI value was 85% for Trichoderma harzianum. However, Fusarium oxysporum f. sp. ciceri, highest PGI was 93.75% for Trichoderma virideand Trichoderma piluliferumand Lowest PGI value was 81% T. virens. result of MIC was showed effective for all strain. Degree of antagonist was varied for each strain (Rahman et.al., 2009). Each strain of *Trichoderma* was recorded excellent efficacy against *Fusarium oxysporum* f. sp. *lycopersici*(FOL) and Fusarium oxysporum f. sp. ciceri(FOC) In-vitro (Altinok and Erdogan, 2015). These results have implications for use of such in-vitro test as a part of general screen for biological efficacy of different biocontrol agents.

Mycelium interaction is a basic method to assess antagonist according Pic.2. During experiment was observed two type interaction of *Trichoderma* strain with FOL and FOC In vitro. One was zone of inhibition and second was without zone of inhibition. These results proofed that all strains of Trichoderma antagonized FOL and FOC growth to various degrees and that different degrees of inhibition (Jinantara, 1995). This experiment was showed that PGI values varied but the ranking of MIC remained same for all Trichoderma strain. This task agreed with the work done by Mokhtar H et.al.arereported the inhibition of pathogen. *Trichoderma*, as one of the promising bio-control agent, has been described (Morsvet.al., 2009; Sabalparaet.al., 2009). Therefore, Many researcher has been proofed that *Trichoderma* are not equally effective in control of the pathogen in vitro by Biswas and Das. 1999: Ramezani.2008. The presence Zone of inhibition has been used as evidence of the production of either antibiotics or cell free metabolites by the Trichoderma strains (Jackson et. al. 1991; Crawford et.al., 1993). Barari (2016) has proofed that siderophore and antibiotics was secreted from Trichoderma strains which have played a crucial role in plant growth and sustainable disease management programs to save environmental risk.

#### Conclusion

The study find out the antagonistic potentials of different strain of Trichoderma against toFusarium oxysporum f. sp. lycopersiciand Fusarium oxysporum f. sp. ciceri is varied and excellent. Morever, degree of PGI value is varied due toavailable different strain of *Trichoderma* antagonist with both pathogens. This investigation is anevidenced that could be easily Plantpathogen control to bv Trichoderma antagonist behavior, this is proved in vitro.

#### Acknowledgement

I wish to express my sincere gratitude and respect to **Dr. Arjun Singh, scientist (Agriculture microbiology), NBIAM, Mau (U.P.)** for facilities of Lab work.

#### References

- Akhtar, C.M. (1966). The isolation of soil fungi-1, A simple method of isolating fungi from soil. The needle Method. W. Pak. J. Agri. Res., 4: 122-131.
- 2. Rifai, M. A. and Webster (1996). A revision of the genus Trichoderma spp. Mycol. Paper series 16: 1-56, CMI, Kew, Surrey, England.
- Barnett, H. L., and B.B. Hunter (1998). Illustrated genera of imperfect fungi 4<sup>th</sup>Edition. St. Paul, MN, APS Press. pp:223.
- 4. Mokhtar H, Aid D (2013) Contribution in isolation and identification of some pathogenic fungi from wheat seeds, and evaluation ofantagonistic capability of Trichoderma harzianum against those isolatedfungi in vitro. Agric Boil J N Am 4(2): 145-154.
- Akila, R., L. Rajendran, S. Harish, K. Saveetha, T. Raguchander, and R. Samiyappan. (2011). "Combined Application of Botanical Formulations and Biocontrol Agents for the Management of Fusarium Oxysporum F. Sp. Cubense (Foc) Causing Fusarium Wilt in Banana." Biological Control 57 (3): 175–83. doi:10.1016/j.biocontrol.2011.02.010.
- Brandler, Daiani, Luan Junior Divensi, Thalita Pedrozo Pilla, Ines Rezendes, and Paola Mendes Milanesi. 2017. "Evaluation of Biological Control of Fusarium Wilt in Gerbera" (1): 234–39.
- 7. Cucinotta, Francis A. 2014. "Article @ Journals.Plos.Org." Plos One. doi:dx.doi.org/10.1371/journal.pntd.0002908.
- Ignjatov, Maja, Dragana Milosevic, Zorica Nikolic, Jelica Gvozdanovic-Varga, Dusica Jovicic, and Gordana Zdjelar. 2012. "Fusarium Oxysporum as Causal Agent of Tomato Wilt and Fruit Rot." Pesticidi I Fitomedicina 27 (1): 25–31. doi:10.2298/PIF12010251.
- Mukherjee, Mala, Prasun K. Mukherjee, Benjamin A. Horwitz, Christin Zachow, Gabriele Berg, and Susanne Zeilinger. (2012). "Trichoderma-Plant-Pathogen Interactions: Advances in Genetics of Biological Control." Indian Journal of Microbiology. doi:10.1007/s12088-012-0308-5.
- 10. Singha, I.M., Y. Kakoty, B.G. Unni, J. Das, and M.C. Kalita. (2016). "Identification and Characterization of Fusarium Sp. Using ITS and RAPD Causing Fusarium Wilt of Tomato Isolated from Assam, North East India." Iournal of Genetic Engineering and 99-105. Biotechnology 14 (1): doi:10.1016/j.jgeb.2016.07.001.
- 11. Sonawane, Anupama, Manali Mahajan, and Sonali Renake. (2015). "Original Research Article Antifungal Activity of a Fungal Isolates

against Pomegranate Wilt Pathogen Fusarium" 2 (2): 48–57.

- 12. Vinale. Francesco, Krishnapillai Sivasithamparam, Emilio L. Ghisalberti, Sheridan L. Woo, Marco Nigro, Roberta Marra, Lombardi,(2014). "Trichoderma Nadia Secondary Metabolites Active on Plants and Fungal Pathogens." The Open Mycology Iournal 127-39. 8 (1): doi:10.2174/1874437001408010127.
- Skidmore A.M., Dickinson C.H. (1976).Colony interactions and hyphal interference between Septoria nodorum and phylloplane fungi.Transaction of the British Mycological Society, 66: 57–64.
- 14. Mustafa A, MA Khan, M Inam-ul-Haq, MA Pervez and UD Umar.(2009). Usefulness Of Different Culture Media For In-Vitro Evaluation Of Trichoderma Spp. Against Seed-Borne Fungi Of Economic Importance. Pakistan Journal of Phytopathololy. 21(1): 83-88.
- 15. Khang T.V., Anh MY T.N., Pham Minh Tu M.P., Tham H. T. N., (2013).Isolation and selection of Trichoderma spp. exhibiting high antifungal activities against major pathogens in Mekong delt.Omonrice 19: 159-171.
- 16. Rahman MA, Begum MF, Alam MF (2009) Screening of Trichoderma isolates as a biological control agent against Ceratocystis paradoxa causing pineapple disease of sugarcane. Mycoscience 37(4):277-285.
- 17. Korsten L, De Jager ES (1995) Mode of action of Bacillus subtilis for control of avocado post harvest pathogens. S Afr Avocado Growers Assoc. 18: 124-130.
- Sangoyomi T. (2004) Post-harvest Fungal Deterioration of yam (Dioscorearotundata. Poir) and its Control. Ph.D. Thesis. University of Ibadan, Nigeria, pp. 179.
- 19. Jinantara, J. (1995). Evaluation of Malaysian isolates of Trichoderma harzianum Rifai and Glicocladium virens Miller, Giddensand Foster for the biological control of Sclerotium foot rot of chilli. Ph.D. Thesis. Universiti Putra Malaysia, Selangor, Malaysia.
- Cordova-Albores, L.C.; Zapotitla E. S.; Ríos M.Y.; Barrera-Nechaa, L.L.; Hernández-López M.; Bautista-Banos S. (2016) Microscopic study of the morphology and metabolic activity of Fusarium oxysporum f. sp. gladiolitreated with Jatrophacurcas oil and derivatives Journal of Microscopy and Ultrastructure 4 (2016) 28–35.

- 21. Sharma P.,(2011). Complexity of Trichoderma-Fusarium interaction and manifestation of biological control.AJCS 5(8): 1027-1038.
- 22. Altinok, Hacer Handan; Erdogan, Oktay (2015). Determination of the In vitro Effectof Trichoderma harzianum on Phytopathogenic Strains of Fusarium oxysporum. NotulaeBotanicaeHortiAgrobotani ci Cluj-Napoca, [S.l.], v. 43, n. 2, p. 494-500, dec. ISSN 1842-4309.
- 23. Nwankiti A, Gwa V. (2018) Evaluation of Antagonistic Effect of Trichoderma Harzianum against Fusarium oxysporum causal Agent ofWhite Yam (Dioscorearotundatapoir) Tuber Rot. Trends Tech Sci Res.; 1(1): 555554.
- 24. Barari, H. (2016). Biocontrol of tomato Fusarium wilt by Trichoderma species under in vitro and in vivo conditions. Cercetariagronomice in noldova vol. XLIX, No.1(165):91-88.
- 25. Babalola O.O., Glick B.R. (2012). Indigenous African Agriculture and plant associated microbes: current practice and future transgenic prospects. Science Res Essays, 7: 2431-2439.
- Biswas K.K., Das N.D., 1999 Biological control of pigeon pea wilt caused by Fusarium udum with Trichoderma spp. Ann. Plant Protection Science,7(1): 46-50.
- 27. Jackson A.M., Whips J.M., Lynch J.M., (1991).In vitro screening for identification of potential biocontrol agent of Allium white rot. Mycol Res, 95: 430-434.
- Crawford D.L., Lynch J.M., Whipps J.M., Osley M.A., (1993). Isolation and characterization of actinomycete antagonists of a fungal root pathogen. Appl Environ Microbiol, 59: 3899-3909.
- Morsy E.M., Abdel-Kawi K.A., Khalil M.N.A., 2009 - Efficacy of Trichoderma viride and Bacillus subtilis as biocontrol agents against Fusarium solani on tomato plants. Egypt. Journal of Plant Pathology, 37(1): 47-57.
- Sabalpara A.N., Priya J., Waghunde R.R., Pandya J.P., 2009 - Antagonism of Trichoderma against sugarcane wilt pathogen (Fusarium moniliformae), American-Eurasian J. Agric. Environ. Sci., 3(4): 637-638.
- Ramezani H., 2008 Biological control of rootrot of eggplant caused by Macrophominaphaseolina. American-Eurasian J. Agric. Environ. Sci, 4(2): 218-220.