

EVALUATION OF LICHEN DIVERSITY ON LIANAS IN SELECTED SACRED GROVES OF WEST MIDNAPORE DISTRICT, WEST BENGAL

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ABSTRACT ■ The sacred groves are fairly well studied for the anthropological, ecological, botanical, zoological and conservation perspectives. However, there are a few publications on lichens of sacred groves. The lichens have long been considered sensitive indicators for monitoring environmental state. The present study documents 80 species of lichens representing 43 genera of 19 families in twenty one selected sacred groves of West Midnapore district, West Bengal. *Combretum decandrum* Jacq. exhibited the maximum diversity of lichens in form of 45 species. This work is likely to foster further studies on lichen-diversity in other regions of West Bengal for a better understanding of the associated biodiversity and environment.

Key words: Conservation, Diversity, Liana, Lichen, Sacred grove

INTRODUCTION

Lichens are interesting symbiotic association of green and non-green thallophytes, which thrive in a variety of habitats, being most abundant in subtropical and temperate climates. These require high levels of humidity for better growth and can grow easily on soil, bark and leaf of the plant, and on rocks. Growth-forms of lichen have many characteristic features. They are thalloid, attached to substratum in three forms: crustose, foliose and fruticose. Primarily lichens are symbiotic associations of fungi and algae. The major fungal component belongs to ascomycetes, for which most of the lichens are known as ascolichens (Ahmadjian, 1993). Lichens with cyanobacterial symbionts

contribute significantly to forest nitrogen fixation (Bergman et al., 1992). Besides many other uses including folk uses, lichens are also used as pollution monitors and in dating of rocks.

Lianas are woody climbers rooted in the soil which use other plants, especially trees, as support for growth away from the ground (Holbrook and Putz, 1996). It is mostly assumed that the particular advantage of this habit is to allow these plants to escape from deep shade and to reach the upper canopy of forests. This implies that their seeds would germinate in the shade and seedlings initially would grow upwards and develop in the shade. In the tropical south West Bengal, lush growth of lianas, however, is mostly found in

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undisturbed forests especially the sacred groves. Lianas in fact can slow down successions in the reinvasion of gaps, in that they suppress climax species, and thus, support the growth of pioneer species (Schnitzer et al., 2000). Growth of saplings of trees is not only inhibited by lianas via competition for light, but also below ground via competition in the rooting medium (Schnitzer et al., 2005). Perhaps, by clasping other plants they are just saving investment in thick stems which would provide independent support for their biomass.

Shoots of lianas wind around branches of a support or form coils, which are then modified by their secondary thickening, so that the wood with a thick bark develops in the form of bands forming rope or cable like structures which are resistant to torsion.

Sacred groves, the tribal community-based repositories of plant diversity, are fragments of landscape with distinctive ecological features to afford protection to biodiversity on the basis of sacredness or religious practice or faith (Bhakat et al., 2008). The groves are distributed uniformly in West Midnapore district, West Bengal, India, in the form of densely wooded natural patches, mainly of angiosperm flora with perennial water resources in vicinity. As a unique ecosystem, it helps in soil and water conservation, preserving the biological wealth. They are the treasure house of many cryptogamic and phanerogamic plants such as algae, fungi, lichen, bryophyta, pteridophyta and angiosperms (Nayaka and Upreti, 2004; Singh et al., 2012).

The general floristic composition and physiognomy of the vegetation of sacred groves are typically as the semi-evergreen forests (FSI, 2015). The vegetation in undisturbed groves is luxuriant and comprises several stories of trees mixed with shrubs,

lianas and herbs (Sen and Bhakat, 2012). The soil is rich in humus, and covered with thick litter. Such types of groves create microclimatic conditions, encouraging luxuriant growth of moisture-loving lichens including other growth forms, of immense ecological and economic values (Upreti et al., 2005; Kumar, 2009).

The distribution of lichens (as in other cryptogams) is largely influenced by microclimatic factors such as topography, land cover and water. In sacred groves, four major ecological factors that produce several microclimatic niches for the growth of lichens are substrate, vegetation, climate and altitude (Nayaka et al., 2013). The sacred grove provides a wide range of substrates for the growth of lichens, and hence, total 80 species of lichens belongs to corticolous category.

STUDY AREA

The West Midnapore district is located in the southern part of West Bengal, India. It is bounded by Bankura and Hooghly districts in the northern side, Howrah district in the northeastern side and East Midnapore district in the southeastern side. The western boundary is merged with Jhargram district of West Bengal (Fig. 1).

Location and Topography

The West Midnapore district exhibits regional diversity in terms of physiographic, agro-climatic characteristics and social composition etc. Geo-morphologically, the district is subdivided into three parts, viz. Chhotonagpur flanks with hills, mounds and rolling lands in the westernmost part, Rahr plain with lateritic uplands in the middle part and Alluvial plain of the east with recent deposits. It is hilly towards the north-west, represents low basins in the southeast and east. It has drought-affected dry areas in the west, but highly wet flood-affected in the east. Dense semi-

evergreen forest in the west is replaced by semi-aquatic vegetations of marshy lands in the east. It has barren lateritic, non-arable lands in the west and north-west, which gradually changes with highly productive alluvial soil areas in the central and eastern part of the district. It is the abode of tribes and primitive tribes (*Bhumij, Kheria, Lodha, Mahali, Munda* and *Santal*) in the western blocks, while most of the area is inhabited by almost all castes of the society, representing the diverse culture across blocks. Extremely rugged topography is seen in the western part of the district and rolling topography is experienced consisting of lateritic areas. These rolling plains gradually merge into flat alluvial and deltaic plains to the east and southeast of the district (Anon, 2011).

Climate and Soil

The climate of the region is tropical; the terrain is characterized by hard rock uplands, lateritic areas, and flat alluvial and deltaic plains with fairly fertile soil. The area experiences an annual rainfall of 1400 - 1500 mm, but is highly erratic for the last few years. The mean temperature of the area is between maximum of 44°C during peak summer and minimum 10°C during the coldest days of winter.

METHODOLOGY

Different species of lichens were opportunistically collected from ten different species of dicotyledonous lianas from twenty one selected sacred groves (area more than 1 hectare). Intensive sampling of lichens was carried out from January, 2015 to February, 2017. Lichens were collected along with substratum using a sharp knife. The specimens were procured very precisely without damaging the thallus. Various species of lichen were also encountered through collection of fallen branches and twigs on the ground. The specimens were cleaned carefully

by removing debris, sun-dried and deposited for preservation. Later, the species of lichens were studied using light compound binocular microscope and identified up to species level with the help of standard techniques such as spot tests, UV-light and Thin Layer Chromatography (TLC) (Elix and Ernst-Russel, 1993; Orange et al., 2001) and relevant literature including keys and technical monographs (Huneck and Yoshimura, 1996; Rout et al., 2010; Singh and Sinha, 2010; Vinayaka et al., 2010; Singh and Kumar, 2012).

RESULTS AND DISCUSSIONS

The lichen flora on lianas of the sacred groves studied was of tropical type (Table 1). Among the liana-species *Combretum decandrum* was found to provide suitable habitat for rich growth of lichens (45 species). The other lichen-accommodating lianas were *Acacia torta*, *Bauhinia vahlii* and *Derris scandens* (13 species each); *Hiptage benghalensis* (10 species); *Aganosma heynei* and *Vallaris solanacea* (8 species each); *Capparis zeylanica* (3 species); *Ventilago denticulata* and *Ziziphus oenopolia* (2 species each) (Table 3; Fig. 2).

The study revealed the occurrence of 19 families of lichens represented by 43 genera and 80 species. Graphidaceae (24 species) was the most dominant family, followed successively by Arthoniaceae and Trypetheliaceae (10 species each); Pyrenulaceae (9 species); Ramalinaceae (4 species); Lecanoraceae, Parmeliaceae and Physciaceae (3 species each); Letrouitiaceae, Pertusariaceae and Thelotremaaceae (2 species each) (Table 2; Fig. 3). Another set of eight families contains a single species, each namely Arthopyreniaceae, Chrysothricaceae, Collemataceae, Haematommataceae, Lecideaceae, Monoblastiaceae, Roccellaceae and Teloschistaceae (Table 2). Among the various growth forms crustose were the

Table 1. Occurrence of lichens in selected lianas of West Midnapore district, West Bengal, India.

Sl. No.	Lichen Species	Family	Habit	Habitat	Substratum/Host plant (Table 3)
1.	<i>Anisomeridium terminatum</i> (Nyl.) R. C. Harris	Monoblastiaceae	Cr.	C	Cz
2.	<i>Anthracotheccium thwaitesii</i> (Leight.) Müll. Arg.	Pyrenulaceae	Cr.	C	Hb
3.	<i>Arthonia translucens</i> Stirt.	Arthoniaceae	Cr.	C	Hb
4.	<i>Arthonia medusula</i> (Pers.) Nyl.	Arthoniaceae	Cr.	C	Vd/Zo
5.	<i>Arthothelium albescens</i> Patw. & Makhija	Arthoniaceae	Cr.	C	Bv/Cd
6.	<i>Arthothelium pycnocarpoides</i> Müll. Arg.	Arthoniaceae	Cr.	C	Hb
7.	<i>Bacidia alutacea</i> (Kremp.) Zahlbr.	Ramalinaceae	Cr.	C	Vd/Zo
8.	<i>Bacidia convexula</i> (Müll. Arg.) Zahlbr.	Ramalinaceae	Cr.	C	At/Bv/Ds
9.	<i>Bacidia phaeolomoides</i> (Müll. Arg.) Zahlbr.	Ramalinaceae	Cr.	C	Cd
10.	<i>Bacidiospora psorina</i> (Nyl. ex Hue) Kalb	Ramalinaceae	Cr.	C	At/Ds
11.	<i>Bathelium benguelense</i> Müll. Arg.	Trypetheliaceae	Cr.	C	Hb
12.	<i>Bathelium madreporiforme</i> (Eschw.) Trevis.	Trypetheliaceae	Cr.	C	Cd/Hb
13.	<i>Bulbothrix isidiza</i> (Nyl.) Hale	Parmeliaceae	Fo.	C	Cd
14.	<i>Caloplaca bassiae</i> (Ach.) Zahlbr.	Teloschistaceae	Cr.	C	Cd
15.	<i>Chapsa pseudophlyctis</i> (Nyl.) Frisch	Graphidaceae	Cr.	C	Cd
16.	<i>Chrysothrix candelaris</i> (L.) J. R. Laundon	Chrysothricaceae	Cr.	C	At/Bv/Ds
17.	<i>Cryptothecia bengalensis</i> Jagadeesh, G. P. Sinha & Kr. P. Singh	Arthoniaceae	Cr.	C	Cd
18.	<i>Cryptothecia effusa</i> (Müll. Arg.) R. Sant.	Arthoniaceae	Cr.	C	Ah/Vs
19.	<i>Cryptothecia involuta</i> Stirt.	Arthoniaceae	Cr.	C	Cd
20.	<i>Cryptothecia subtectata</i> Stirt.	Arthoniaceae	Cr.	C	Cd
21.	<i>Diorygma megasporum</i> Kalb, Staiger & Elix	Graphidaceae	Cr.	C	Cd
22.	<i>Dyplolabia afzelii</i> A. Massal.	Graphidaceae	Cr.	C	At/Bv/Ds
23.	<i>Glyphis cicatricosa</i> Ach.	Graphidaceae	Cr.	C	At/Bv/Ds
24.	<i>Glyphis scyphulifera</i> (Ach.) Staiger	Graphidaceae	Cr.	C	Cd
25.	<i>Graphina platycarpa</i> (Eschw.) Zahlbr.	Graphidaceae	Cr.	C	Cd
26.	<i>Graphis acharii</i> F'ee	Graphidaceae	Cr.	C	At/Bv/Ds
27.	<i>Graphis caesiella</i> Vainio	Graphidaceae	Cr.	C	Hb
28.	<i>Graphis distincta</i> Makhija & Adaw.	Graphidaceae	Cr.	C	Ah/Vs
29.	<i>Graphis filiformis</i> Adaw. & Makhiza	Graphidaceae	Cr.	C	Cd
30.	<i>Graphis glaucescens</i> Fee.	Graphidaceae	Cr.	C	Hb

Sl. No.	Lichen Species	Family	Habit	Habitat	Substratum/Host plant (Table 3)
31.	<i>Graphis japonica</i> (Müll. Arg.) A.W. Archer & Lücking	Graphidaceae	Cr.	C	At/Bv/Ds
32.	<i>Graphis librata</i> C. Knight	Graphidaceae	Cr.	C	Cd
33.	<i>Graphis perticosa</i> (Kremp.) A.W. Archer	Graphidaceae	Cr.	C	Cd
34.	<i>Graphis pinicola</i> Zahlbr.	Graphidaceae	Cr.	C	At/Bv/Ds
35.	<i>Graphis scripta</i> (L.) Ach.	Graphidaceae	Cr.	C	Cd
36.	<i>Graphis streblocarpa</i> (Bel) Nyl.	Graphidaceae	Cr.	C	Ah/Vs
37.	<i>Graphis subasahinae</i> Nagarkar & Patw.	Graphidaceae	Cr.	C	At/Bv/Ds
38.	<i>Graphis chlorotica</i> A. Massal.	Graphidaceae	Cr.	C	Hb
39.	<i>Haematomma wattii</i> (Stirt.) Zahlbr.	Haematommataceae	Cr.	C	Cd
40.	<i>Herpothallon isidiatum</i> Jagadeesh & G.P. Sinha	Arthoniaceae	Cr.	C	Ah/Vs
41.	<i>Heterodermia obscurata</i> (Nyl.) Trevis.	Physciaceae	Fo.	C	Ah/Vs
42.	<i>Heterodermia pseudospeciosa</i> (Kurok.) W. L. Culb.	Physciaceae	Fo.	C	Cd
43.	<i>Laurera cumingii</i> (Mont.) Zahlbr.	Trypetheliaceae	Cr.	C	At/Bv/Ds
44.	<i>Laurera keralensis</i> Upreti & Ajay Singh	Trypetheliaceae	Cr.	C	Cz
45.	<i>Lecanora cinereofusca</i> H. Magn.	Lecanoraceae	Cr.	C	Cd
46.	<i>Lecanora iseana</i> Rasanen	Lecanoraceae	Cr.	C	Ah/Vs
47.	<i>Lecidea granifera</i> (Nyl.) Zahlbr.	Lecideaceae	Cr.	C	Cd
48.	<i>Leiorreuma exaltatum</i> (Mont. & v.d. Bosch) Staiger	Graphidaceae	Cr.	C	Cd
49.	<i>Leptogium austroamericanum</i> (Malme) C.W. Dodge	Collemataceae	Fo.	C	Cd
50.	<i>Letrouitia leprolyta</i> (Nyl.) Hafellner	Letrouitiaceae	Cr.	C	Ah/Vs
51.	<i>Letrouitia transgressa</i> (Malme) Hafellner & Bellem.	Letrouitiaceae	Cr.	C	Cd
52.	<i>Mycomicrothelia conothele</i> (Nyl.) Hawksw.	Arthopyreniaceae	Cr.	C	Cd
53.	<i>Myelochroa xantholepis</i> (Mont. & Bosch) Elix & Hale	Parmeliaceae	Fo.	C	Cd
54.	<i>Myriotrema norstictideum</i> (Patw. & Nagarkar) D.D. Awasthi	Thelotremataceae	Cr.	C	Cd
55.	<i>Opegrapha rufescens</i> Pers.	Roccellaceae	Cr.	C	Cd
56.	<i>Pallidogramme chrysenteron</i> (Mont.) Staiger, Kalb & Lücking	Graphidaceae	Cr.	C	Cd
57.	<i>Parmotrema andinum</i> (Mull. Arg.) Hale	Parmeliaceae	Fo.	C	Cd
58.	<i>Pertusaria melastomella</i> Nyl.	Pertusariaceae	Cr.	C	Cd
59.	<i>Pertusaria quassiae</i> (Fée) Nyl.	Pertusariaceae	Cr.	C	Cd

Sl. No.	Lichen Species	Family	Habit	Habitat	Substratum/Host plant (Table 3)
60.	<i>Phaeographis brasiliensis</i> (A. Massal.) Kalb & Matthes-Leicht	Graphidaceae	Cr.	C	Hb
61.	<i>Platythecium grammitis</i> (Fée) Staiger	Graphidaceae	Cr.	C	Cd
62.	<i>Polymeridium proponens</i> (Nyl.) R.C. Harris	Trypetheliaceae	Cr.	C	Cd
63.	<i>Pyrenula acutalis</i> R.C. Harris	Pyrenulaceae	Cr.	C	Cd
64.	<i>Pyrenula anomala</i> (Ach.) Vain.	Pyrenulaceae	Cr.	C	Cd
65.	<i>Pyrenula introducta</i> (Stirt.) Zahlbr.	Pyrenulaceae	Cr.	C	At/Bv/Ds
66.	<i>Pyrenula leucotrypa</i> (Nyl.) Upreti	Pyrenulaceae	Cr.	C	Cd
67.	<i>Pyrenula nitens</i> (Fée) Fée	Pyrenulaceae	Cr.	C	Cd
68.	<i>Pyrenula sublaevigata</i> (Patw. & Makhija) Upreti	Pyrenulaceae	Cr.	C	Cd
69.	<i>Pyrenula subnitida</i> Müll. Arg.	Pyrenulaceae	Cr.	C	At/Bv/Ds
70.	<i>Pyrenula thelomorpha</i> Tuck.	Pyrenulaceae	Cr.	C	Ah/Vs
71.	<i>Pyxine coccifera</i> (Fée) Nyl.	Physciaceae	Fo.	C	Cd
72.	<i>Ramboldia russula</i> (Ach.) Kalb, Lumbsch & Elix	Lecanoraceae	Cr.	C	Cd
73.	<i>Reimnitzia sentensis</i> (Tuck.) Kalb.	Thelotremataceae	Cr.	C	Cd
74.	<i>Sarcographa tricola</i> (Ach.) Müll. Arg.	Graphidaceae	Cr.	C	At/Bv/Ds
75.	<i>Trypethelium eluteriae</i> Spreng.	Trypetheliaceae	Cr.	C	Cz
76.	<i>Trypethelium endosulphureum</i> Makhija & Patw.	Trypetheliaceae	Cr.	C	Cd
77.	<i>Trypethelium platystomum</i> Mont.	Trypetheliaceae	Cr.	C	Hb
78.	<i>Trypethelium pupula</i> (Ach.) R.C. Harris	Trypetheliaceae	Cr.	C	Cd
79.	<i>Trypethelium tropicum</i> (Ach.) Müll. Arg.	Trypetheliaceae	Cr.	C	Cd
80.	<i>Tylophoron protrudens</i> Nyl.	Arthoniaceae	Cr.	C	Cd

Abbreviations: C-Corticolous, Cr-Crustose, Fo-Foliose

dominant (73 species; 91%) than foliose (7 species; 9%) (Fig. 4). On the basis of substratum, the studied all lichens on the lianas of the sacred groves was found to prefer corticolous (80 species of bark) habitat (Table 1).

The 14 dominant plant genera with descending species number (=2) are *Graphis* (13 species), *Pyrenula* (8 species), *Trypethelium* (5 species), *Cryptothecia* (4 species), *Bacidia* (3 species), and 9 genera *Arthonia*, *Arthothelium*,

Glyphis, *Lecanora*, *Letrouitia*, *Pertusaria*, *Heterodermia*, *Bathelium*, *Laurera* contain 2 species each. Remaining 29 genera contain single species each (Table 2; Fig. 5).

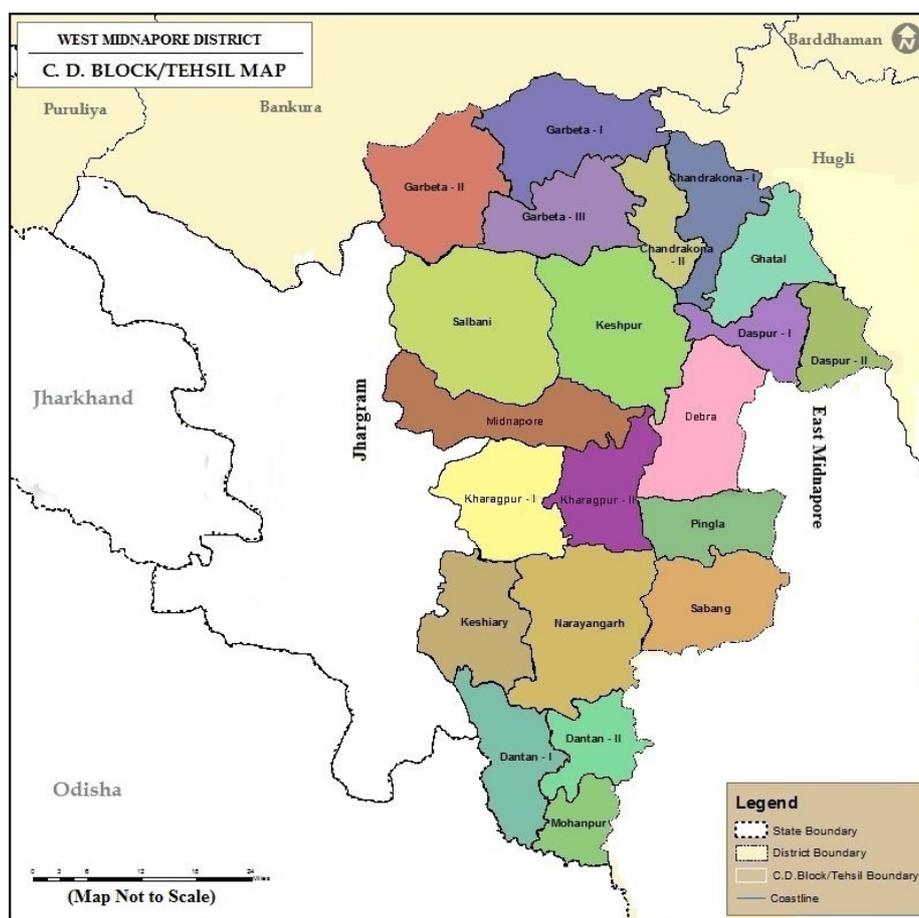
The rich lichen flora on a particular liana is dependent on a wide range of interrelated factors. The microclimate exhibited by various parts of encountering liana species, including the mature substratum, determines the lichen growth on a liana. The age, smoothness, roughness, and spongy nature of bark, along

Table 2. Enumeration of family, genus and species of lichen.

Family	Genus	Species	
		Genus-wise	Total
Arthoniaceae Reichenb. ex Reichenb.	<i>Arthonia</i>	2	10
	<i>Arthothelium</i>	2	
	<i>Cryptothecia</i>	4	
	<i>Herpothallon</i>	1	
	<i>Tylophoron</i>	1	
Arthopyreniaceae W. Watson	<i>Mycomicrothelia</i>	1	1
Chrysothricaceae Zahlbr.	<i>Chrysothrix</i>	1	1
Collemataceae Zenker	<i>Leptogium</i>	1	1
Graphidaceae Dumort.	<i>Chapsa</i>	1	24
	<i>Diorygma</i>	1	
	<i>Dyplolabia</i>	1	
	<i>Glyphis</i>	2	
	<i>Graphina</i>	1	
	<i>Graphis</i>	13	
	<i>Leiorreuma</i>	1	
	<i>Pallidogramme</i>	1	
	<i>Phaeographis</i>	1	
	<i>Platythecium</i>	1	
	<i>Sarcographa</i>	1	
Haematommataceae Hafellner	<i>Haematomma</i>	1	1
Lecideaceae Chevall.	<i>Lecidea</i>	1	1
Lecanoraceae Korb.	<i>Lecanora</i>	2	3
	<i>Ramboldia</i>	1	
Letrouitiaceae Bellem. & Hafellner	<i>Letrouitia</i>	2	2
Monoblastiaceae W. Watson	<i>Anisomeridium</i>	1	1
Parmeliaceae Zenker	<i>Bulbothrix</i>	1	3
	<i>Parmotrema</i>	1	
	<i>Myelochroa</i>	1	
Pertusariaceae Korb. ex Korb.	<i>Pertusaria</i>	2	2
Physciaceae Zahlbr.	<i>Heterodermia</i>	2	3
	<i>Pyxine</i>	1	
Pyrenulaceae Rabenh.	<i>Anthracothecium</i>	1	9
	<i>Pyrenula</i>	8	
Ramalinaceae C. Agardh	<i>Bacidia</i>	3	4
	<i>Bacidiospora</i>	1	
Roccellaceae Chevall.	<i>Opegrapha</i>	1	1
Teloschistaceae Zahlbr.	<i>Caloplaca</i>	1	1
Thelotremataceae (Nyl.) Stizenb.	<i>Myriotrema</i>	1	2
	<i>Reimnitzia</i>	1	
Trypetheliaceae Zenker	<i>Bathelium</i>	2	10
	<i>Laurera</i>	2	
	<i>Polymeridium</i>	1	
	<i>Trypethelium</i>	5	
Total	19	43	80

Table 3. Total number of lichen species hosted by each liana.

Sl. No.	Name of the liana species	Abbreviation	Family	No. of species hosted
1.	<i>Acacia torta</i> (Roxb.) Craib	At	Fabaceae	13
2.	<i>Aganosma heynei</i> (Spreng.) I.M. Turner	Ah	Apocynaceae	08
3.	<i>Bauhinia vahlii</i> Wight & Arn.	Bv	Fabaceae	13
4.	<i>Capparis zeylanica</i> L.	Cz	Capparaceae	03
5.	<i>Combretum decandrum</i> Jacq.	Cd	Combretaceae	45
6.	<i>Derris scandens</i> (Roxb.) Benth.	Ds	Fabaceae	13
7.	<i>Hiptage benghalensis</i> (L.) Kurz	Hb	Malpighiaceae	10
8.	<i>Vallis solanacea</i> (Roth) Kuntze	Vs	Apocynaceae	08
9.	<i>Ventilago denticulata</i> Willd.	Vd	Rhamnaceae	02
10.	<i>Ziziphus oenopolia</i> (L.) Mill.	Zo	Rhamnaceae	02

**Fig. 1:** Location map of West Midnapore district.

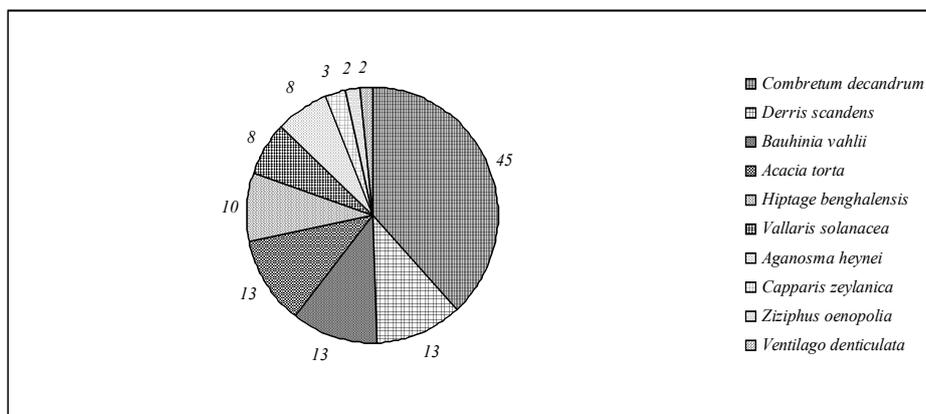


Fig. 2: Total number of lichen species hosted by each liana.

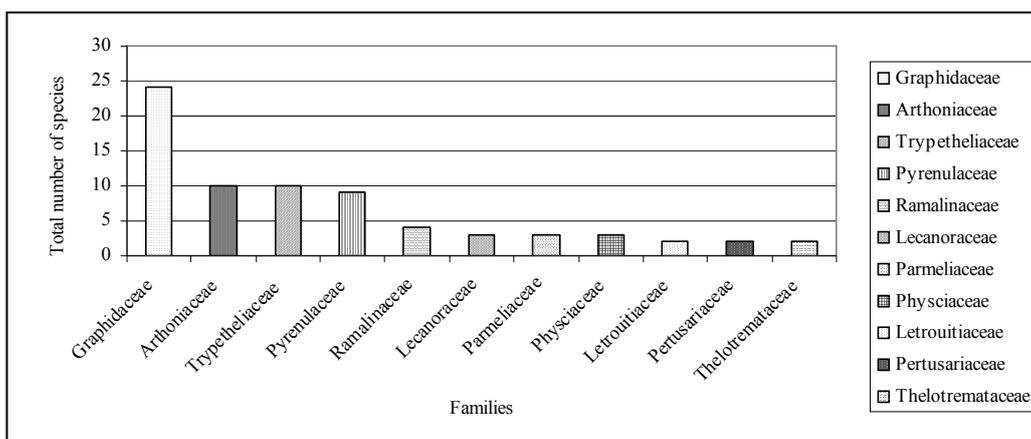


Fig. 3: Dominant lichen families with descending number of species (= 2 species).

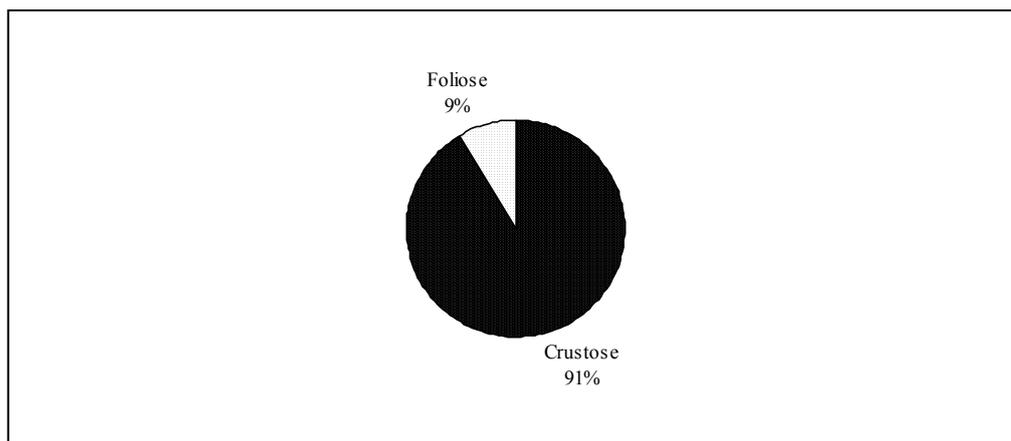


Fig. 4: Different growth forms of lianas.

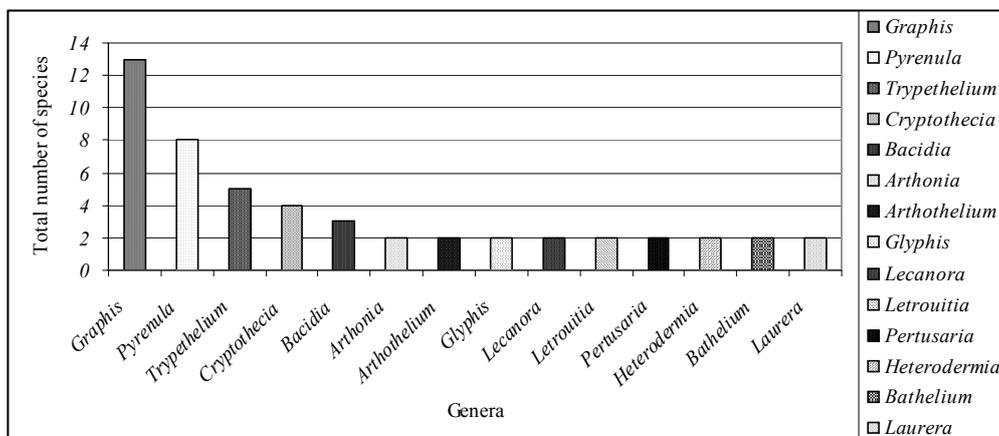


Fig. 5: Dominant lichen genera with descending number of species (=2 species).

with pH, nutrient status, buffer capacity and water holding capacity of soil are other important factors affecting the growth of lichens on the lianas (Satya et al., 2005). The reason for rich lichen flora harbored by *Combretum decandrum* could be due to the variation of the bark-quality in different parts of the liana.

Over the past three decades, nearly all arguments about the conservation of nature on this planet have involved the issue of biological diversity and the ways to preserve it (Margules and Pressey, 2000). Lichens have not remained immune from such inquiries, and several studies testify to the contemporary importance of conservation of lichens and their habitats (Wolseley, 1995). Ganderton et al. (2005) state “in terms of biogeography, conservation can be seen as one more element in the dynamic interactions between species and their natural environment”. Such an interaction tends to maintain a desired range of taxa and habitats often against prevailing ecological and environmental changes. Many aspects of our environment are changing locally or globally as a result of human activities. These include temperature, carbon dioxide, rainfall, UV radiation, ozone, acidification and

nitrification and changes in them are likely to have direct effects on lichen communities (Galloway, 1993). Conservation strategies, such as selection of protected areas, now need to take into account changes in environmental factors and human-induced events, such as rapid climate change that may shift the environmental conditions of a protected area in a way that may no longer support the taxa or ecosystems the protected area attempts to conserve (Mackey et al., 2008). Lichens over their long evolutionary history have seen many catastrophic changes in the planet’s terrestrial environments and are probably much better placed with their unique symbiotic systems to survive and see some future anthropogenic mass extinction episodes.

Threats and Conservation

Lichens being very sensitive to the environmental changes may in response show changes in their diversity, abundance, morphology, physiology, accumulation of pollutants etc. The main threats that apply to biodiversity in general are also true for lichens, e.g. habitat fragmentation, degradation and loss, overexploitation, air pollution and climate change (Scheidegger and Werth, 2009). The rapid urbanization,

agriculture, tourism and small scale industrialization in remote areas have been leading to the fast extinction of the forest vegetation as well as the lichens (Negi, 2003). Overgrazing by animals also leads to extinction of lichens. There has been a sharp increase in the demand of non-timber forest products from the forests and hence, the irrational harvesting of lichens has become a serious hazard to biodiversity. Lichens are slow growing organisms, and thus, if once removed from their habitat, they will take several years to reestablish or ultimately disappear. Air pollution is yet another major threat to the lichens as they are sensitive to different levels of pollution (Upreti et al., 2005; Sen, 2014). Conservation of their habitat is very important to prevent the extinction of lichens. It can be done by developing strategies for *in-situ* and *ex-situ* conservation. The overexploitation of natural resources should be reduced and conservation areas must be prioritized. It is important to create awareness among people about the importance and conservation of lichens.

CONCLUSION

Sacred groves are religiously protected areas that provide a comprehensive and rich ecological niche as repositories of genetic diversity. Moreover, it is felt that there are tremendous direct and indirect pressures at work on the groves threatening their existence. These threats can be related to increasing prospects of tourism, higher demands for non timber forest products, fuel wood collection, decrease in the religious faiths along with a fall in the commitment of the present generation towards such natural sacred areas, and lastly, the heavy burden of developmental interventions that are prepared to undertake. Microclimatic conditions of sacred groves play an important

role in the ecology of lichens. Availability of water, sunlight, moderate climate, unpolluted atmosphere, wind condition and type of substratum are the major factors responsible for the optimal growth of lichens. From the present study, it is evident that sacred grove abodes a good number of lichens which are getting impoverished due to various factors. The little attention of administrators towards the deteriorating condition of holy places and the groves add another dimension. Such gene pool reserves can definitely serve as icons of *in-situ* conservation under the prevailing times through a good mixture of scientific measures and awareness building efforts with the active involvement of the local community and the government.

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