

How the Worm Got its Pharynx: Phylogeny, Classification and Bayesian Assessment of Character Evolution in Acoela

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Abstract.—Acoela are marine microscopic worms currently thought to be the sister taxon of all other bilaterians. Acoels have long been used as models in evolutionary scenarios, and generalized conclusions about acoel and bilaterian ancestral features are frequently drawn from studies of single acoel species. There is no extensive phylogenetic study of Acoela and the taxonomy of the 380 species is chaotic. Here we use two nuclear ribosomal genes and one mitochondrial gene in combination with 37 morphological characters in an analysis of 126 acoel terminals (about one-third of the described species) to estimate the phylogeny and character evolution of Acoela. We present an estimate of posterior probabilities for ancestral character states at 31 control nodes in the phylogeny. The overall reconstruction signal based on the shape of the posterior distribution of character states was computed for all morphological characters and control nodes to assess how well these were reconstructed. The body-wall musculature appears more clearly reconstructed than the reproductive organs. Posterior similarity to the root was calculated by averaging the divergence between the posterior distributions at the nodes and the root over all morphological characters. Diopisthoporidae is the sister group to all other acoels and has the highest posterior similarity to the root. Convolutidae, including several “model” acoels, is most divergent. Finally, we present a phylogenetic classification of Acoela down to the family level where six previous family level taxa are synonymized. [Acoela; ancestor; Bilateria; character evolution; divergence; posterior similarity; reconstruction signal.]

Acoela are small hermaphroditic marine worms (380 spp.) that occur commonly as part of the meiofauna of sandy or muddy sediments as well as among algae. Acoels have attracted considerable attention in recent years because analyses of ribosomal (Ruiz-Trillo et al. 1999; Telford et al. 2003; Wallberg et al. 2007) and protein coding nuclear genes (Ruiz-Trillo et al. 2002; Hejnol et al. 2009) have supported their position as one of the earliest branches within Bilateria and separate from the protostomian Platyhelminthes, in which they were previously classified. These molecular findings are consistent with their comparatively simple morphology: acoels lack excretory organs and other features of the Nephrozoa (Jondelius et al. 2002). Neurotransmitter patterns in acoels indicate a brain composed of transverse commissures and strengthened anterior parts of longitudinal cords different from the typical bilaterian pattern with a central neuropile surrounded by neurons (Raikova et al. 2001; Reuter et al. 2001). Reconstruction of acoel ancestral traits will likely provide important clues to the morphology of the last common bilaterian ancestor and the evolution of more complex bilaterians (Jondelius et al. 2002) but is restricted to inference from extant species since there is no acoel fossil record.

The study of acoels has its challenges, as the worms are small, ranging between 0.3 and 10 mm, and rather fragile. Identification of species often requires studying the reproductive organs of live specimens in a drop of sea water under a high-quality light microscope. The reproductive biology and general appearance of the worms often adds to the difficulty; interesting

specimens are often immature and their soft bodies easily disintegrate under a badly applied cover slip. Moreover, many cold water specimens die in less than half an hour at room temperature. Acoela have a limited number of organs but display almost unlimited variation in their configuration. Thus, there is relatively low morphological complexity but high disparity. Such disparity could be expected from a lineage as old as Nephrozoa and likely predating the Cambrian (Budd 2008; Peterson et al. 2008). The mouth can be located almost anywhere along the ventral midline: either in the anterior as in *Proporus*, ventrally as in most Acoela or posterotermally as in *Diopisthoporus*. A muscular pharynx exists in some taxa, but it has been considered nonhomologous among these groups due to anatomical differences (Todt 2009) implying independent appearance of a pharynx within Acoela. The acoel nervous system is highly variable in terms of position relative to the body wall and level of centralization (Kotikova and Raikova 2008). Acoela are hermaphrodites and there may be paired or single ovaries and testes or, as in *Diopisthoporus* and *Antigonaria* a single mixed gonad. Secondary female organs such as a copulatory bursa for storage of allosperm and a vagina, may be highly developed or completely absent. Male copulatory organs may take the shape of sclerotized stylets, muscular penes, or glandular structures of various shapes. One or two gonopores may be located anywhere along the ventral midline. Even the acoel spermatozoa are diverse. They have two incorporated flagella with either 9 + 2, 9 + 1 or 9 + 0 axoneme configurations and many

different arrangements of cytoplasmic microtubules and other inclusions. It is not surprising that 19th and 20th century systematists struggled with the classification of the group.

In spite of their pivotal position in animal phylogeny, there is no extensive phylogenetic study of the interrelationships within Acoela. The proliferation of family level taxa without reference to a phylogenetic hypothesis (e.g., Westblad 1948; Dörjes 1968; Kostenko 1989; Kostenko and Mamkaev 1990) has resulted in some confusion. In many cases, there are no credible synapomorphies, and the current classification has little predictive power: it is problematic to determine to which higher taxon a new specimen of an unknown species should be referred because classification into taxa such as families is more or less a matter of pigeonholing. Moreover, it is often difficult to identify a specimen belonging to a previously described species.

The largest study to date, which was based on partial 18S rDNA sequences from 32 species of 10 nominal acoel families, retrieved only 4 of these as monophyletic (Paratomellidae, Mecynostomidae, Sagittiferidae, and Anaperidae), whereas three families were reconstructed as nonmonophyletic (Actinoposthiidae, Convolutidae, and Haploposthiidae) and the status of the remaining three families represented was undetermined (Hooge et al. 2002). Hooge and coauthors observed that some features of the spermatozoa and body-wall musculature appeared to fit the topology particularly well, but without a more complete taxonomic sample it is difficult to hypothesize which states are ancestral and important for inferring the features of the last common bilaterian ancestor. There have been attempts to use some acoel species as “models” for Acoela and lower Bilateria, for example, when inferring a complex pattern with longitudinal, circular and U-shaped muscles in the acoel and bilaterian ancestors based on studies of *Iso-diametra pulchra* and *Symsagittifera roscoffensis* (Semmler et al. 2008) and when inferring separate origins of the bilaterian mouth and anus based on studies of the development of *Convolutriloba longifissura* (Hejnol and Martindale 2008). The choice of species for detailed study has been based on availability or ease of identification rather than on a hypothesis spanning as much as possible of the phylogenetic diversity. Reconstructions of ancestral states must be based on thorough knowledge of character distributions and a robust phylogenetic hypothesis for the Acoela, both of which are currently lacking. Consequently, explicit hypotheses of character evolution in this group are few, restricted mainly to the evolution of the pharynx (Todt 2009), spermatozoa (Hooge et al. 2002; Achatz et al. 2010), and body-wall musculature (Hooge et al. 2002), and phylogenetic hypothesis testing has generally not been feasible.

Here we set out to reconstruct the phylogeny of Acoela using a comprehensive data set consisting of ribosomal and mitochondrial DNA sequences and morphological characters. We aim to provide a revised and simplified classification of Acoela consistent with the phylogeny, study the evolution of morphological

characters, and reconstruct ancestral features of Acoela and of acoel subgroups. In particular, we compare some evolutionary properties of the body-wall musculature with those of the gonads because both organ systems have been highlighted as important for acoel phylogenetics and classification. The evolution of the acoel pharynx, as inferred by us, differs distinctly from the hypothesis put forward by Todt (2009) and we therefore evaluate alternative hypotheses on pharynx homology in more detail. We use a Bayesian framework to account for some of the uncertainty of the reconstruction (Ronquist 2004) and demonstrate how this approach can be extended to provide useful tools for systematic research such as estimating the reconstruction potential of characters at deep nodes, morphological divergence among clades and help delineate taxa.

MATERIALS AND METHODS

Specimens

The material represents a collecting effort spanning more than 10 years and many marine habitats and locations from all over the world (Table 1). Specimens were collected by a variety of methods ranging from beach sampling at low tide or snorkeling to scuba diving or dredging at larger depths. Live specimens were most often fully identified to species using light microscopy, but some unidentified specimens or specimens of unidentified species were only determined to genus. If available, photos of these unidentified worms can be found at the Acoela Scratchpad project (<http://acoela.myspecies.info/>).

Coding of Morphological Characters

General morphology such as size, shape, pigmentation, presence or absence of algal symbionts, eyes, pharynx, chordoid vacuoles, or frontal glands and most of the characters related to the gonads and genitals were coded for 104 species using original descriptions, live observations, and illustrations or photos of the actual specimen. Ultrastructural and histological information is still not available for many species, and was, with the exception of the musculature in *Diopisthoporus longitubus*, only included for those species for which these features have been described. Information about the presence or absence of interconnecting digestive cells located at the mouth or pharynx was coded for 6 species from Todt (2009); axoneme and microtubular arrangements of spermatozoa were coded for 36 species; (the 9 + 2 axoneme arrangement in *Hofstenia miamia* is our own unpublished observation); and properties of the body-wall musculature were fully coded for 46 species. The body-wall musculature of *Diopisthoporus lofolitis*, as described by Hooge and Smith (2004), was here attributed to the very similar *D. longitubus*, for which we had molecular data. See Table 2 for the full set of references used to code the morphological information. We concluded that information about the nervous system in acoels was still too fragmentary to

TABLE 1. Species and genes sampled for phylogenetic analyses

Group/species	Voucher ID	18S	28S	COI	Source/location
Acoela					
Actinoposthiidae Hooge, 2001					
<i>Actinoposthia beklemschevi</i> Mamkaev, 1965	—	AI012522	—	—	GenBank
<i>Actinoposthia haplocata</i> Dörjes, 1968	AWHel-30	FR837674	FR837757	FR837838	Offshore Kattegat, Sweden
<i>Atrioforontia polyacucula</i> Dörjes, 1968	—	AF102895	—	—	GenBank
<i>Pelophila lutheri</i> (Westblad, 1946)	—	AY078366	—	—	GenBank
<i>Philaetinoesthia coneyi</i> Hooge and Rocha, 2006	MHSc	FR837724	FR837809	FR837880	São Paulo, Brazil
<i>Philaetinoesthia</i> nsp. 2	MHBawiActinoTAN	FR837726	—	—	Zanzibar, Tanzania
<i>Philaetinoesthia</i> nsp. 3	AWBer-26	FR837725	FR837810	FR837881	Bermuda
<i>Philaetinoesthia saliens</i> (Graff, 1882)	UJ07-60	FR837727	FR837811	FR837881	Koster Area, Sweden
<i>Philaetinoesthia</i> sp.	UJ07-33	FR837731	FR837814	FR837884	Koster Area, Sweden
<i>Philaetinoesthia</i> sp. 2	UJ08-65	FR837728	FR837812	FR837882	West Coast, New Caledonia
<i>Philaetinoesthia</i> sp. 3	AWHel-35	FR837729	FR837813	FR837883	Offshore Kattegat, Sweden
<i>Philaetinoesthia</i> sp. 4	UJ04-25	FR837730	—	—	Sardinia, Italy
<i>Pseudactinoesthia sanguineum</i> (Beklemishev, 1915)	—	AY078369	—	—	GenBank
Anaperidae Dörjes, 1968					
<i>Anaperus biaculatus</i> Boguta, 1970	—	AI012527	AY157602	—	GenBank
<i>Anaperus gardineri</i> (Graff, 1911)	MHAg	AY078365	—	FR837840	Maine, USA
<i>Anaperus rubellus</i> Westblad, 1945	UJ07-47	FN552040	FN552040	FR837841	Koster Area, Sweden
<i>Anaperus singularis</i> Hooge and Smith, 2004	AWSp-24	—	FR837758	FR837842	Galiza, Spain
<i>Anaperus toaerinimensis</i> (Luther, 1912)	—	AF102898	—	—	GenBank
Chilidiidae Dörjes, 1968					
<i>Chilidia bracliyoposthium</i> (Westblad, 1942)	AWHel-27	FR837681	FR837764	FR837847	Offshore Kattegat, Sweden
<i>Chilidia crassum</i> (Westblad, 1942)	AWHel-08	FR837682	FR837765	—	Offshore Kattegat, Sweden
<i>Chilidia cycloposthium</i> (Westblad, 1942)	—	AF329178	AJ849494	—	GenBank
<i>Chilidia groenlandica</i> (Levinsen, 1879)	AWSp-13	AM701805	FR837766	—	Galiza, Spain
<i>Chilidia macroposthium</i> (Steinböck, 1931)	AWHel-28	FR837683	FR837767	FR837848	Offshore Kattegat, Sweden
<i>Chilidia</i> nsp. 1	UJ08-37	FR837684	FR837768	—	Noumea area, New Caledonia
<i>Chilidia</i> nsp. 2	AWSp-26	FR837685	FR837769	FR837849	Galiza, Spain
<i>Chilidia</i> sp.	AWHel-23	FR837686	FR837770	—	Offshore Kattegat, Sweden
<i>Chilidia submaculatum</i> (Westblad, 1942)	AWHel-02	FR837687	FR837771	FR837850	Offshore Kattegat, Sweden
<i>Chilidia trianguliferum</i> (Westblad, 1942)	AWHel-25	FR837688	FR837772	FR837851	Offshore Kattegat, Sweden
<i>Chilidia vivipara</i> Tekle et al., 2006	—	AY297954	AJ849498	—	GenBank
Convolutidae Graff, 1905					
<i>Amphiscolops bermudensis</i> Hyman, 1939	AWA14-Amp	FN552036	FN552037	FR837839	Lee Stocking Island, Bahamas
<i>Amphiscolops</i> sp.	—	AMB18SRNA	—	—	GenBank
<i>Amphiscolops</i> sp. "Hendelberg"	—	FN552041	—	—	Gullmar fiord, Sweden
<i>Convoluta cf. boehmigi</i> (Brauner, 1920)	UJ05-25	FR837689	FR837773	—	Lee Stocking Island, Bahamas
<i>Convoluta convoluta</i> (Abildgaard, 1806)	MHCc	FR837690	—	FR837852	Maine, USA
<i>Heterochaeris blumi</i> (Achatz et al., 2007)	MHAb	FN552038	FR837788	FR837864	Carrie Bow Cay, Belize
<i>Neochilidia fusca</i> Bush, 1975	MHnf	—	FR837801	FR837876	Maine, USA
<i>Prusaqitiffera naikaisensis</i> (Yamasu, 1982)	—	DEL18SRNAA	—	—	GenBank
<i>Stomatrichia hochbergi</i> Hooge, 2003	MHSh	FN552046	FN552047	FR837902	Queensland, Australia
<i>Waminioa brickneri</i> Ogunlana et al., 2005	MHWa	FR837756	FR837837	—	Eilat, Israel ^a
<i>Waminioa</i> sp.	—	AJ875221	—	—	GenBank
Dakuidae Hooge, 2003					
<i>Daku woorimensis</i> Hooge, 2003	MHDw	FR837691	FR837774	FR837854	Queensland, Australia
Diopisthoporidae Westblad, 1940					
<i>Diopisthoporus longitubus</i> Westblad, 1940	UJ06-06	FR837692	FR837775	FR837855	Offshore Skagerrak, Sweden
<i>Diopisthoporus</i> nsp.	AWSp-16	FR837693	FR837776	FR837856	Galiza, Spain
<i>Diopisthoporus psammophilus</i> Dörjes, 1968	UJ04-45	FR837694	FR837777	—	Sardinia, Italy
Hallangiidae Westblad, 1946					
<i>Hallangia proporioides</i> Westblad, 1946	UJ06-14	FR837698	FR837780	FR837858	Offshore Skagerrak, Sweden
Haploposthiidae Westblad, 1948					
<i>Haplogonaria "schillingi"</i>	MHRr	FR837700	FR837782	FR837859	Maine, USA
<i>Haplogonaria syltensis</i> Dörjes, 1968	—	AF102900	—	—	GenBank

Continued

TABLE 1. Continued

Group/species	Voucher ID	18S	28S	COI	Source/location
<i>Haplophragma viridis</i> Dörjes, 1968	UJ07-70	FR837701	FR837783	FR837860	Koster Area, Sweden
<i>Haplophragma lactomaculata</i> Tekle, 2004	UJ03-32	FR837702	FR837784	FR837861	Gullmar fiord, Sweden
<i>Haplophragma rubra</i> (An der Lan, 1936)	UJ03-16	FR837703	FR837785	FR837862	Koster Area, Sweden
<i>Haplophragma</i> sp.	UJ05-33	FR837704	FR837786	—	Lee Stocking Island, Bahamas
<i>Haplophragma vandula</i> Hooge et al., 2002	AWBer-12	FR837705	FR837787	FR837863	Bermuda
<i>Kuma albiventris</i> (Marcus, 1954)	AWBer-34	FR837712	FR837795	FR837872	Bermuda
<i>Kuma viridis</i> (An der Lan, 1936)	UJ03-34	FR837713	FR837796	FR837873	Gullmar fiord, Sweden
<i>Pseudohaplogonaria rodnani</i> Hooge et al., 2002	MHPH	FR837749	FR837830	—	Carrie Bow Cay, Belize
<i>Pseudohaplogonaria</i> sp.	UJ05-02	FR837750	FR837831	FR837898	Lee Stocking Island, Bahamas
<i>Simpliconormpha gigantiorhabditis</i> Dörjes, 1968	—	AF102894	—	—	—
Hofsteniidae Bock, 1923	—	—	—	—	—
<i>Hofstenia miamia</i> Correa, 1960	UJ05-34	AM701809	AM701808	FR837865	Lee Stocking Island, Bahamas
<i>Hofsteniella pardii</i> Papi, 1957	AWSp-08	AM701821	AM701822	FR837866	Galiza, Spain
Isodiametridae Hooze and Tyler, 2005	—	—	—	—	—
<i>Aphanostoma bruscai</i> Hooge and Tyler, 2003	MHMa5	AY078368	FR837759	—	Maine, USA
<i>Aphanostoma collinae</i> Hooge and Tyler, 2008	MHMa14	FR837675	—	FR837843	Bocas del Toro, Panama
<i>Aphanostoma</i> sp.	AWHel-19	FR837676	FR837760	FR837844	Offshore Kattegat, Sweden
<i>Aphanostoma virescens</i> Oersted, 1845	—	AJ012528	—	—	GenBank
<i>Archaphanostoma macrospiriferum</i> (Westblad, 1946)	UJ07-18	FR837677	FR837761	FR837845	Koster Area, Sweden
<i>Avogina marci</i> Dörjes and Karling, 1975	MHAM	FR837679	FR837763	—	São Paulo, Brazil
<i>Avogina</i> nsp.	UJ08-14	FR837680	—	FR837846	Noumea area, New Caledonia
<i>Faerlea glomerata</i> Westblad, 1945	MHHd	FR837697	—	—	Gullmar fiord, Sweden
<i>Haplocelis dichona</i> (Marcus, 1954)	MHHb	FR837699	FR837781	FR837867	São Paulo, Brazil
<i>Isodiametra bajaensis</i> Hooge and Eppinger, 2005	MHI1	FR837706	FR837789	—	Baja California, Mexico
<i>Isodiametra cernuus</i> Hooge and Tyler, 2008	MHMa1	FR837707	—	FR837868	São Paulo, Brazil
<i>Isodiametra divae</i> (Marcus, 1950)	—	AY078370	FR837790	—	GenBank
<i>Isodiametra hortulus</i> (Hooge and Tyler, 2003)	MHMa16	FR837708	—	FR837869	Bocas del Toro, Panama
<i>Isodiametra nicki</i> Hooge and Tyler, 2008	MHMa8	FR837709	FR837792	FR837870	Maine, USA
<i>Isodiametra norvegica</i> (Westblad, 1946)	MHCp	FR837710	FR837793	—	Maine, USA
<i>Isodiametra pulchra</i> (Smith and Bush, 1991)	UJCove	FR837711	FR837794	FR837871	Belgian coast, Belgium
<i>Isodiametra vexillaria</i> (Marcus, 1948)	MHCc	FR837718	FR837803	—	São Paulo, Brazil
<i>Otocelis erinae</i> Hooge and Rocha, 2006	MHOs	FR837719	FR837804	FR837878	Maine, USA
<i>Otocelis sandara</i> Hooge and Tyler, 2003	MHMa7	FR837739	FR837820	—	Bocas del Toro, Panama
<i>Praeconcolita bocensis</i> Hooge and Tyler, 2008	MHMa13	FR837738	FR837821	FR837890	Maine, USA
<i>Praeconcolita castinea</i> Hooge and Tyler, 2003	MHMa7	FR837739	FR837821	FR837891	Maine, USA
<i>Praeconcolita tigrina</i> Hooge and Tyler, 2003	MHMa6	FR837740	FR837822	—	GenBank
<i>Praeconcolita tornuwa</i> Tyler and Hooge, 1999	—	AY078374	—	—	Galiza, Spain
<i>Proaphanostoma tenuissima</i> (Westblad, 1946)	AWSp-22	FR837741	FR837823	FR837892	Galiza, Spain
<i>Pseudaphanostoma herringi</i> Hooge and Rocha, 2006	MHMa2	FR837747	FR837828	FR837895	São Paulo, Brazil
<i>Pseudaphanostoma smithii</i> Hooge and Tyler, 2003	MHPs	AY078375	—	FR837896	Maine, USA
<i>Raphidophallus actuosus</i> Kozloff, 1965	MHMa12	FR837751	—	FR837899	California, USA
Mecynostomidae Dörjes, 1968	—	—	—	—	—
<i>Eumecynostomum altitudi</i> Faubel and Regier, 1983	—	AY297955	AJ849492	—	GenBank
<i>Mecynostomum auritum</i> (Schultze, 1851)	—	AJ845244	AJ849493	—	GenBank
<i>Mecynostomum</i> sp.	AWSp-34	FR837715	FR837798	FR837874	Galiza, Spain
<i>Paedomecynostomum bruncum</i> Dörjes, 1968	AWK05-47	FR837720	FR837805	—	Gullmar fiord, Sweden
<i>Paramecynostomum diversicolor</i> (Oersted, 1845)	UJ07-36	FR837721	FR837806	FR837879	Koster Area, Sweden
<i>Paramecynostomum</i> sp.	UJ08-53	FR837722	FR837807	—	Noumea area, New Caledonia
<i>Philomecynostomum lapillum</i> Dörjes, 1968	—	AF102897	—	—	GenBank
<i>Philomecynostomum</i> sp.	AWSp-41	FR837734	FR837817	FR837887	Galiza, Spain
<i>Postmecynostomum pictum</i> Dörjes, 1968	AF102899	FR837734	—	—	GenBank
<i>Pseudomecynostomum bruceifilum</i> Faubel, 1974	AWSp-02	FR837748	FR837829	FR837897	Galiza, Spain
<i>Pseudomecynostomum phoca</i> Hooge and Tyler, 2003	—	AY078376	—	—	GenBank
Otocelididae Westblad, 1948	—	—	—	—	—
<i>Notocelis gullmarensis</i> (Westblad, 1946)	UJ07-37	—	FR837802	FR837877	Koster Area, Sweden
<i>Philocelis brueggemanni</i> Hooge and Tyler, 2003	MHPHb	—	FR837815	FR837885	Maine, USA

Continued

TABLE 1. Continued

Group/species	Voucher ID	18S	28S	COI	Source/location
<i>Philocelis karlingi</i> (Westblad, 1946)	—	AJ845243	—	—	GenBank
<i>Philocelis robrochii</i> Hooge and Rocha, 2006	MHSa	FR837733	FR837816	FR837886	São Paulo, Brazil
Paratomellidae Dörjes, 1966	—	—	—	—	—
<i>Paratomella rubra</i> Rieger & Ott, 1971	UJ04-55	FR837723	FR837808	AY228758	Sardinia, Italy
<i>Paratomella unichaeta</i> Dörjes, 1966	—	AY078379	—	—	GenBank
Polycanthiidae Hooge, 2003	—	—	—	—	—
<i>Polycanthus</i> sp.	UJ08-33	FR837736	—	FR837889	Noumea area, New Caledonia
<i>Polycanthus</i> sp. 2	UJ08-17	FR837735	FR837818	FR837888	Noumea area, New Caledonia
<i>Polycanthus torosus</i> Hooge, 2003	MHPt	FR837737	FR837819	—	Queensland, Australia
Proporidae	—	—	—	—	—
<i>Proporus bermudensis</i> Hooge et al., 2002	AWBer-02	FR837742	FR837824	FR837893	Bermuda
<i>Proporus brochii</i> Westblad, 1946	UJ06-18	FR837743	FR837825	—	Offshore Skagerrak, Sweden
<i>Proporus carolinensis</i> Hooge and Smith, 2004	MHPb	FR837744	FR837826	—	North Carolina, USA
<i>Proporus</i> sp. 2	UJ05-23	FR837746	FR837827	FR837894	Lee Stocking Island, Bahamas
Sagittiferidae Kostenko and Mamkaev, 1990	UJProporussp	FR837745	—	—	Offshore Skagerrak, Sweden
<i>Convolvuliribba hastifera</i> Winsor, 1990	—	EU710906	EU710919	EU710926	GenBank
<i>Convolvuliribba longifissura</i> Bartolomaeus and Balzer, 1997	AWColo	FN552044	FN552045	FR837853	Hawaii, USA ^b
<i>Convolvuliribba macropyga</i> Shannon and Achatz, 2007	—	EU710899	EU710916	EU710922	GenBank
<i>Convolvuliribba retrogenema</i> Hendelberg and Akesson, 1988	—	EU710909	EU710914	EU710924	GenBank
<i>Synsagittifera corsicae</i> Geschwenter et al., 2002	—	AJ319029	—	—	GenBank
<i>Synsagittifera psammophilula</i> (Beklemishev, 1957)	AWSp-04	FR837755	FR837835	FR837903	Galiza, Spain
<i>Synsagittifera roscoffensis</i> (Graff, 1891)	AWSp-37	AJ012530	FR837836	FR837904	Galiza, Spain
Solenofilomorphae Dörjes, 1968	—	—	—	—	—
<i>Endocincta</i> nsp.	AWSp-33	FR837695	FR837778	FR837857	Galiza, Spain
<i>Endocincta punctata</i> Crezée, 1975	UJ07-46	FR837696	FR837779	—	Koster area, Sweden
<i>Myopea</i> "callaeum"	MHMc	AY078380	FR837799	FR837875	Maine, USA
<i>Myopea</i> sp.	—	AM701823	—	—	GenBank
<i>Oligofilomorpha interstitiophilum</i> Faubel, 1974	—	FR837752	AM701824	—	GenBank
<i>Solenofilomorpha</i> "crezai"	UJ08-74	FR837754	FR837834	FR837901	Maine, USA
<i>Solenofilomorpha</i> nsp.	UJ08-05	FR837753	FR837832	FR837900	Amedee island, New Caledonia
<i>Solenofilomorpha</i> nsp. 2	—	—	—	—	Recif Iamanou, New Caledonia
Outgroups	—	—	—	—	—
Cnidaria	—	—	—	—	—
<i>Aurelia</i> sp.	—	AY920770	EU272547	EU366144	GenBank
<i>Montastraea franksi</i>	—	AY026382	AY026375	AY451358	GenBank
<i>Rosacea flaccida</i>	—	AY937328	EU305529	AY937376	GenBank
Nemertodermatida	—	—	—	—	—
<i>Ascoparia</i> sp.	AWHel-24	FR837678	FR837762	FR837905	Offshore Kattgat, Sweden
<i>Meara stichopi</i>	UJ03-52	FR837714	FR837797	FR837906	Bergen, Norway
<i>Nemertodermis</i> sp.	UJ06-16	FR837717	FR837800	FR837907	Offshore Skagerrak, Sweden
Nephrozoa	—	—	—	—	—
<i>Anialis entalis</i>	—	DQ279936	AY145388	AY260818	GenBank
<i>Apionsoma misakianum</i>	—	AY210440	AY210454	EU267000	GenBank
<i>Asterias forbesi</i>	—	DQ077937	AF212169	AF498646	GenBank
<i>Branchiostoma floridae</i>	—	M97571	AF061796	AF098298	GenBank
<i>Ciona intestinalis</i>	—	AB013017	AF212177	AJ517314	GenBank
<i>Nephrops norvegicus</i>	—	FJ174918	DQ079803	FJ174945	GenBank
<i>Priapulus caudatus</i>	—	X80234	AY210840	DQ087502	GenBank
<i>Ramazzottius oberhaueri</i>	—	AY582122	FJ435768	FJ435800	GenBank
<i>Terebellides stroemi</i>	—	AY577893	DQ790066	DQ209261	GenBank
<i>Terebratulina retusa</i>	—	U08324	AY839244	AJ245743	GenBank

Note: new sequences produced in this study are highlighted in bold.

^aCollected by Orit Barneah, Ben-Gurion University.

^bDonation from Andreas Hejnol, University of Hawaii.

TABLE 2. Primary literature sources used for coding acoel morphology

<i>General morphology</i>		
Abildgaard (1806)	Graff (1911)	Marcus (1950)
Achatz et al. (2007)	Gray and Rieger (1971)	Marcus (1954)
An der Lan (1936)	Gschwentner et al. (2002)	Oersted (1845)
Bartolomaeus and Balzer (1997)	Hendelberg and Åkesson (1988)	Ogunlana et al. (2005)
Beklemishev (1915)	Hooge (2003)	Papi (1957)
Beklemishev (1957)	Hooge and Eppinger (2005)	Schultze (1851)
Boguta (1970)	Hooge and Rocha (2006)	Shannon and Achatz (2007)
Brauner (1920)	Hooge and Smith (2004)	Smith and Bush (1991)
Bush (1975)	Hooge and Tyler (1999)	Steinböck (1931)
Correa (1960)	Hooge et al. (2002)	Tekle (2004)
Crezée (1975)	Hooge and Tyler (2003)	Tekle et al. (2006)
Dörjes (1966)	Hooge and Tyler (2005)	Tekle et al. (2007)
Dörjes (1968)	Hooge and Tyler (2008)	Uljanin (1870)
Dörjes and Karling (1975)	Hyman (1939)	Westblad (1940)
Faubel (1974)	Kozloff (1965)	Westblad (1942)
Faubel and Regier (1983)	Levinson (1879)	Westblad (1945)
Graff (1882)	Luther (1912)	Westblad (1946)
Graff (1891)	Mamkaev (1965)	Winsor (1990)
Graff (1905)	Marcus (1948)	Yamasu (1982)
<i>Body-wall musculature</i>		
Gschwentner et al. (2002)	Hooge and Rocha (2006)	Todt (2009)
Gschwentner et al. (2003)	Hooge and Smith (2004)	Tyler and Hooge (1999)
Hooge (2001)	Hooge and Tyler (2003)	Tyler and Rieger (1999)
Hooge (2003)	Shannon and Achatz (2007)	
Hooge and Eppinger (2005)	Tekle et al. (2005)	
<i>Sperm ultrastructure</i>		
Afzelius (1966)	Henley and Costello (1969)	Raikova et al. (1997)
Afzelius (1982)	Henley (1968)	Raikova et al. (1998)
Costello et al. (1969)	Henley (1974)	Raikova et al. (2001)
Crezée (1975)	Henley et al. (1968)	Raikova and Justine (1999)
Hendelberg (1969)	Hooge and Tyler (2005)	Silveira (1967)
Hendelberg (1974)	Mamkaev and Ivanov (1970)	Silveira (1969)
Hendelberg (1975)	Petrov et al. (2004)	Tekle et al. (2007)
Hendelberg (1977)	Raikova and Justine (1994)	

be included in the analysis. Coding and management of the data matrix was done with the web-based MX content management system (Yoder et al. 2006). Both binary and multistate characters were coded. The core data set consisted of 37 characters coded for 104 acoel species. Seven additional characters were coded as multistate versions of some binary characters to provide additional details at a few nodes. Three Nemertodermatida species were coded specifically to assess how inclusion of morphological information from this taxon would influence our inference. See Table 3 for the morphological characters and states and the Supplementary Data for the morphological matrix (available from <http://www.sysbio.oxfordjournals.org>).

Molecular Sequence Assembly and Alignment

We used the 18S rRNA, 28S rRNA, and COI genes as molecular markers for the phylogenetic analyses. Eighty-seven new sequences of about 1780 bp of the 18S rRNA gene and 82 new sequences of about 3200 bp of the 28S rRNA gene were produced for acoels according to the PCR protocols described in Wallberg et al. (2007). A few additional 28S primers were used: F2093 (alternative to U1846) 5'-CCAYAACCGCATCAGGT-3', R3182 (alternative

to L3449) 5'-AACACTTTGCGAATTCTGCTTCGC-3', and R3239 (alternative to L3449) 5'-CCTGTCTCACGACGGTCTAAACCC-3'. Sixty-seven new acoel sequences of about 660 bp of the COI gene were produced using the universal LCO/HCO primers and protocols from Folmer et al. (1994) and with minor variations of the annealing temperature around 50 °C. Contigs were produced with Staden v1.6.0 (Staden 1996) and the Phred/Phrap base calling and assembly modules (Ewing and Green 1998; Ewing et al. 1998). The three genes were also sequenced in the three nemertodermatids used as outgroups. Additional acoel and outgroup sequences were downloaded from GenBank (<http://www.ncbi.nlm.nih.gov/>). See Table 1 for the complete list of sequences included in the analyses. Both ribosomal genes and COI were aligned with Multiple Alignment using Fast Fourier Transform (MAFFT) (Kato and Toh 2008; Kato et al. 2009). The COI data were aligned using the nonstructural global Needleman-Wunsch algorithm. Because acoels are known to have unusually variable ribosomal sequences compared with most metazoans (Ruiz-Trillo et al. 1999), 18S and 28S sequences were aligned with alternative algorithms in MAFFT, using either the nonstructural method above or a structural option that aims to incorporate folding properties of the ribosomal genes

TABLE 3. Morphological characters

No.	Character	States
1	Size	0 = small, 1 = conventional, 2 = large
2	Body shape	0 = cylindrical, 1 = convoluted, 2 = flattened
3	Threadlike (slender)	0 = no, 1 = yes
4	Chordoid vesicles (large vacuoles)	0 = absent, 1 = present
5	Level of chordoid vesicles	0 = chordoid vesicles absent, 1 = weakly developed, 2 = well developed
6	Frontal glands	0 = absent, 1 = present
7	Level of frontal glands ^a	0 = frontal glands absent, 1 = weakly developed, 2 = well developed
8	Symbiotic algae	0 = absent, 1 = present
9	Sagittocysts	0 = absent, 1 = present
10	Pigmentation	0 = absent, 1 = present
11	Eyes	0 = absent, 1 = ocelli with platelets, 2 = pigmented eyespots
12	Mouth position	0 = anteroterminal or subterminal, 1 = ventral, 2 = posteroterminal or subterminal
13	Pharynx	0 = absent, 1 = present
14	Interconnecting cells (glandular cells at the pharynx epithelium)	0 = absent, 1 = present
15	Testis	0 = unpaired, 1 = paired
16	Seminal vesicle	0 = absent, 1 = present
17	Glandomuscular eversible copulatory organ	0 = absent, 1 = present
18	Copulatory organ partly inside seminal vesicle	0 = no, 1 = yes
19	Copulatory organ invaginated into seminal vesicle	0 = no, 1 = yes
20	Shape of glandomuscular copulatory organ ^a	0 = glandomuscular copulatory organ absent, 1 = isodiametric tube, 2 = conical tube, 3 = spherical, 4 = globular papilla, 5 = multilayered muscular seminal vesicle
21	Penis stylet (sclerotized copulatory organ)	0 = absent, 1 = present
22	Accessory penetrating hard structures	0 = absent, 1 = adenodactyls, 1 = needles
23	Antrum	0 = absent, 1 = present
24	Antrum ciliation ^a	0 = antrum absent, 1 = ciliated, 2 = not ciliated, 3 = shallow
25	Antrum shape ^a	0 = antrum absent, 1 = long and tubular, 2 = irregular or saclike
26	Male opening position	0 = anteroterminal or subterminal, 1 = midbody, 2 = posteroterminal or subterminal
27	Ovary	0 = unpaired, 1 = paired
28	Bursa seminalis (receptacle for allosperm)	0 = absent, 1 = present
29	Number of bursae ^a	0 = bursa absent, 1 = one, 2 = several
30	Bursal nozzle (sclerotized proximal opening of bursa)	0 = absent, 1 = present
31	Number of bursal nozzles ^a	0 = nozzle absent, 1 = one, 2 = several
32	Vagina	0 = absent, 1 = present
33	Separate female gonopore	0 = absent, 1 = present
34	Ovary positioned or reaches behind testes	0 = no, 1 = yes
35	Hermaphrodite gonad (testis and ovary mixed)	0 = no, 1 = yes
36	Asexual reproduction	0 = absent, 1 = present
37	Sperm axonemes	0 = 9 + 0, 1 = 9 + 1, 2 = 9 + 2
38	Sperm cytoplasmic microtubules	0 = cortical, 1 = axial, 2 = distal only, 3 = corticoaxial
39	Longitudinal muscles enwrap mouth	0 = absent, 1 = present
40	Ventral diagonal muscles	0 = absent, 1 = present
41	Ventral crossover muscles	0 = absent, 1 = present
42	Dorsal diagonal muscles	0 = absent, 1 = present
43	Longitudinal muscles outside of circular muscles	0 = absent, 1 = present
44	Longitudinal muscles between mouth and frontal pore	0 = absent, 1 = present

^aCharacter not included in posterior similarity and reconstruction signal computations for nodes.

to assess position homology by using the Vienna RNA package (Hofacker et al. 2002) and MXSCARNA (Tabei et al. 2008) from within MAFFT. To further assess the influence of ambiguously aligned regions in the ribosomal data on downstream phylogenetic analyses, these alignments were filtered using Aliscore (Misof K. and Misof A. 2009), treating gaps as ambiguous characters and using a sliding window size of 4. Thus, four different concatenated data sets were produced and analyzed, with respect to the ribosomal data: nonstructural unfiltered versus filtered alignments; and structural unfiltered versus filtered alignments, respectively. These data sets spanned 126 acoels and the

taxon sampling coverage of the 18S, 28S, and COI genes were 95%, 72% and 55%, respectively.

Substitution Models and Phylogenetic Analysis

We used Bayesian, maximum likelihood (ML), and cladistic methods of phylogenetic inference. For the model-based analyses of the genes, we used MrAIC (Nylander 2004) and PhyML v2.4.4 (Guindon and Gascuel 2003) to help select among commonly implemented substitution models. We used GTR + G for phylogenetic analysis of 18S, HKY + G for 28S, and GTR + G for COI. Phylogenies were inferred from the combined

data using either nemertodermatids or additional bilaterian and cnidarian taxa as the outgroup. Bayesian phylogenetic inference was performed using a CVS version of MrBayes v3.2 (Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003). We performed the analysis using the concatenated molecular data with and without morphological data and used a G model to allow variation in rates among the morphological characters. The default prior settings were used to estimate parameters such as nucleotide state frequencies, substitution rates, and the shape of the site-to-site rate distribution for each gene. We also analyzed the COI gene separately. For the concatenated data we ran two parallel runs of eight Markov chain Monte Carlo (MCMC) chains using a melting temperature of 0.1 because we found chain mixing to be very low and convergence among runs to be insufficient using the default MCMC settings of four chains and a temperature of 0.2. The combination of additional chains and lower temperature make the chains cover approximately the same range of temperatures as the default settings, but subdivided into more intervals, which in our case improved chain mixing and convergence. The COI analyses used the default settings. The chains ran for 10 million generations and trees and parameters were sampled every 1000 generation. We used AWTY (Nylander et al. 2008), LnL, the potential scale reduction factor, and the average standard deviation of split frequencies to monitor and infer convergence. At 10 million generations, the latter measure was always less than 1.5%. Rapid ML bootstrapping of the molecular data was done with the Pthreads version of RAxML v7.0.4 (Stamatakis 2006; Stamatakis et al. 2008). We ran 500 bootstrap replications using the default “fast and easy” settings and a separate GTR+G model for each data partition. Five-hundred parsimony jackknife replications were run in the software TNT (Goloboff et al. 2008), each of which used five random additions holding up to five trees, sectorial search, and a round of tree fusing (Goloboff 1999). We found that the implied weights algorithm generally produced majority-rule consensus trees that were more congruent with the model-based analysis. The ML bootstrap and parsimony jackknife sets of trees were compared with the Bayesian consensus tree.

Reconstruction of Ancestral Morphology

Reconstructions of ancestral morphological states were performed with BayesTraits (Pagel et al. 2004), and the inference of states at the root were compared with a cladistic reconstruction based on the majority-rule consensus tree in MacClade v4 (Maddison D.R. and Maddison W.P. 2001). We used the trees sampled from the stationary distribution in the Bayesian molecular phylogenetic analysis to represent the current best estimate of the interrelationships and evolutionary distances among taxa. To study the extent to which topological differences related to alignment ambiguities in the molecular data influenced our conclusions about

morphological evolution in acoels, we analyzed the posterior distribution of trees produced from all four alternative alignments in BayesTraits. Three aspects of morphological evolution were compared among the analyses: the inferred evolutionary rates and reconstruction signals (see below) of each morphological character, as well as the states inferred at ancestral nodes. For each analysis, the original sample of 10,000 trees was thinned to 2000 trees due to computational memory constraints, using an in-house Perl script. The outgroup terminals and uncoded acoel species were pruned and the trees were rooted on the first split inside Acoela using Phyutility (Smith and Dunn 2008). Ancestral states were specified to be estimated for nodes of particular interest using the AddMRCA option in BayesTraits and the Multistate version of the program. Two acoel specifiers were used for each node. We focused on the deep nodes in the tree, those nodes roughly corresponding to the traditional families and a few within-family nodes of particular interest. All in all, 31 such nodes were specified. We used the reversible-jump MCMC algorithm (Pagel and Meade 2006) and tried both the exponential and gamma hyperpriors to seed the prior distribution of rates. An MCMC analysis of 101 million generations was run separately for each character. The first million generations were considered burn-in and discarded. Posterior probabilities of states at all nodes were then sampled every 20,000 generation to avoid autocorrelation among samples, amounting to 5000 samples per character-state and node. The analyses were repeated several times but we noted very little difference in the posterior probabilities between runs using the exponential or gamma hyperprior. The final analyses used the default reversible jump setting (rjhp gamma 0 10 0 10). We repeated the analyses without *Hallangia proporooides* because this taxon was recovered at an unexpected part of the tree. To evaluate alternative hypotheses on the evolution and homology of the pharynx among acoel families, we constrained the states of various basal nodes and took the difference between the harmonic means of the marginal likelihoods among different MCMC runs (Bayes factors) as indications for how well different hypotheses fit our phylogenetic evidence. A difference of 2 indicates “positive” evidence to reject the worse hypothesis, whereas a difference of >5 can be taken as strong evidence (Kass and Raftery 1995). BTParser, a cross-platform Perl program, was written specifically to automate running BayesTraits and summarizing the output and can be downloaded from <http://code.google.com/p/btparser/>.

Morphological Divergence and Reconstruction Potential

We implemented a posterior similarity index that compares the posterior probabilities of states estimated at each control node in a set of trees with those of the other control nodes to study morphological divergence. The maximum difference between two posterior probability distributions is 2 and it is recovered when the nodes have full support for different states,

whereas minimal difference is 0, which is recovered when the nodes have identical posterior distributions among states. We inverted and normalized the scale between 0 (maximal divergence) and 1 (maximal similarity) to harmonize this index with other commonly used phylogenetic indexes. A benefit of this approach, compared with analyzing averaged branch lengths in a single morphological tree, is that divergence can be directly analyzed for individual characters or arbitrary groups of characters, even when a well-supported morphological tree is difficult to reconstruct with that data or incongruent with the genealogy. Overall posterior similarity between nodes representing aceol families and the root of the tree was computed by averaging the posterior similarities between the nodes and the root for the core set of morphological characters.

To assess how clearly characters were reconstructed over all control nodes and how well the ancestral worms were reconstructed at each control node, we used the Shannon index to capture the shape of the posterior distributions. Whereas posterior probabilities are mainly used to highlight those states that dominate the posterior, the Shannon index can be used to measure the full shape of the posterior and further help distinguish signal from noise. We here computed the reconstruction signal R to capture the ratio of signal to noise in the posterior distributions by taking 1 minus the evenness among the posterior probabilities (p_1, p_2, \dots, p_N) of all states ($1, 2, \dots, N$) for a given character at a given node:

$$R = 1 - \sum_{i=1}^N \left(-p_i \times \frac{\ln p_i}{\ln N} \right)$$

A flat posterior where states are recovered with nearly the same probabilities has no signal, whereas a highly unbalanced posterior distribution favoring a particular state has a strong signal (Fig. 1). The main benefit of this approach, compared with evaluating the posterior probabilities of only the dominating states at various nodes, is that the reconstruction signal can be computed for characters with an arbitrary set of states and therefore more naturally lends itself to summation over heterogeneous multistate characters to provide an approximation of how well a particular ancestor is reconstructed and understood. We calculated the overall signal at nodes by averaging over the core set of morphological characters and plotted the estimates on the Bayesian consensus tree. We used the Wilcoxon rank-sum test as implemented in R to test whether a group of nodes or characters appeared to be reconstructed with significantly higher signal than another group, given a common set of trees. Two comparisons were done. We first compared the reconstruction signals at 9 control nodes corresponding to families (Fig. 8a,b; gray nodes) with those of 11 deeper parent nodes (Fig. 8a,b; black nodes), most of which were strongly supported in the phylogenetic analyses. We then compared the reconstruction signals of the characters related to the body-wall musculature of the worms with those related to the reproductive system. BayesTraits draws

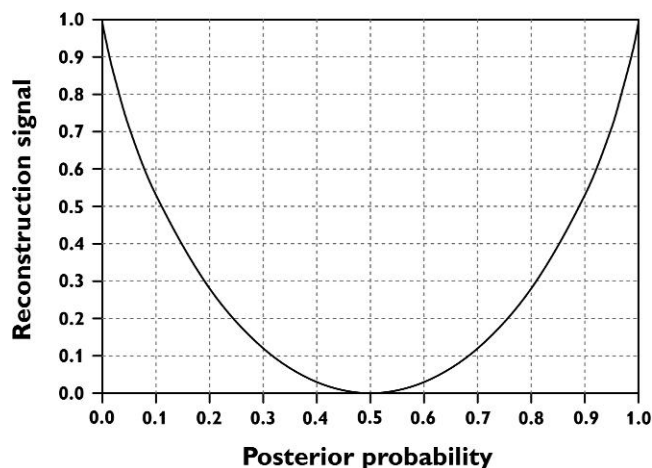


FIGURE 1. Relationship between reconstruction signal and posterior probabilities of states in a binary character. A flat posterior distribution among the two states with $P = 0.5$ has no signal, whereas a highly skewed posterior has a strong signal.

trees randomly from the pool of input trees, so the sequence of sampled trees differs among the MCMC chains of each character. When samples were structured to compare the overall signals of families with their parent nodes, the pattern of sequenced trees were identical among the two sets and estimates generated for all 2000 input trees were compared. To facilitate statistical evaluation between the musculature (six characters, fully sampled in 1109 trees) and reproductive signals (19 characters, fully sampled in 106 trees), we filtered the BayesTraits output to find trees that had been sampled by all 25 MCMC chains, resulting in 55 fully sampled trees. Statistical tests were conducted on a tree-by-tree basis or by pooling the reconstruction signals of each group of samples. In addition to using the full set of taxa in the comparison, we repeated the BayesTraits analyses keeping only those taxa for which the body-wall musculature was coded in the trees.

RESULTS

Phylogenetic Analyses of Molecular Data

The results of the phylogenetic analyses of the concatenated molecular data are summarized on a Bayesian majority-rule consensus tree inferred from the filtered structural alignment in Figure 2. The deep nodes are generally recovered with high support across Bayesian inference, ML bootstrapping, and parsimony jackknifing. Compared with this phylogeny, we detected only minor discrepancies in topologies and clade support among the sets of Bayesian and ML trees inferred from the alternative alignments (see Supplementary Data). Ribosomal sequence alignments are often rich in gaps due to insertions and deletions. For the structural 18S alignment, Aliscore filtered out about 15% of the original positions, yet purging only about 5% of the original sequence data. About 18% of the sequence data were removed from the more variable structural 28S

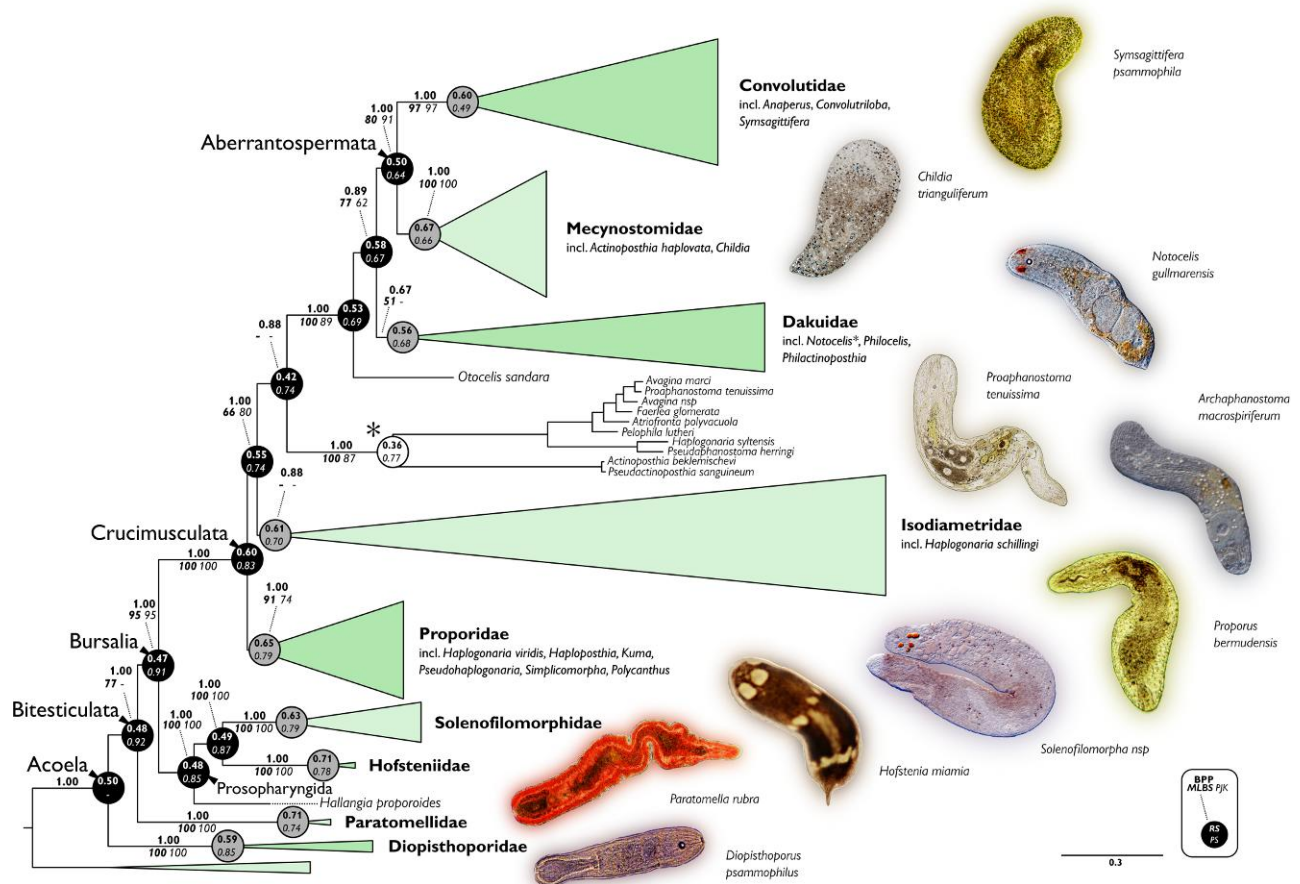


FIGURE 2. Overview of acoel phylogeny. Total-evidence Bayesian majority-rule consensus tree inferred from 18S, 28S, and COI sequences. Bayesian posterior probabilities are shown in bold above branches. ML bootstrap (bold italics) and parsimony jackknife (italics) support below branches. Deep nodes are marked with black circles, nodes corresponding to recognized families are gray. The white control node represents a poorly sampled clade with unstable position. The reconstruction signal (RS) (bold) and posterior similarity to the root (PS) (italics) is displayed on the nodes. A mixed clade with actinoposthiids and isodiametrids with unstable position among analyses of different alignments is marked with an asterisk. Diopisthoporidae is the sister taxon to all other acoels and has the highest similarity to the features reconstructed at the root. Convolutidae is the most derived taxon.

alignment. For the nonstructural alignments, 6% and 14% of the data were removed, respectively. For both the 18S and 28S genes, the structural alignment approach produced the longest filtered alignments (1796 positions and 2811 positions, respectively). However, the only discernible effect of the different data treatments is a slight escalation of topological uncertainties regarding two nodes in the tree: (i) a clade of actinoposthiids and isodiametrids grouping either inside (nonstructural alignments) or outside of Isodiametridae (structural alignments; Fig. 2; clade marked with an asterisk); and (ii) the unresolved placement of *Notocelis gullmarensis* either inside Dakuinae or basal to it; both of which are based on partially sequenced taxa lacking one of the two ribosomal genes (see further results and discussion below).

In view of the nearly identical topologies produced from the analyses of the alternative alignments, we here chose to focus on the results of the phylogenetic analyses of the longer and potentially least ambigu-

ous filtered versions of the structural data sets. In the phylogeny, *Childia* is nested inside Mecynostomidae. *Anaperus* and those sagittiferid genera included in the analysis are nested inside Convolutidae. Most of the haploposthiid species form a group with *Proporus* and *Polycanthus* but some group within Isodiametridae. Members of the Actinoposthiidae are distributed across Mecynostomidae, Dakuinae, and Isodiametridae. Hofsteniidae and Solenofilomorphidae are recovered as sister taxa, as are Mecynostomidae and Convolutidae. Diopisthoporidae is found to be the sister taxon to all other acoels and further shortens the branch between acoels and other bilaterians. When cnidarians and other bilaterians are included as outgroups, acoels and the included nemertodermatids do not form a monophylum (see Supplementary Material), as was also reported by Wallberg et al. (2007). The molecular tree is presented in more detail in Figure 8a,b.

The Bayesian analysis of the mitochondrial COI gene alone results in a less well-supported consensus

TABLE 4. Reconstructions of ancestors at deep nodes (BPP cut-off = 0.6)

No.	Character	Acoela	BPP	Bitesticulata	BPP	Bursalia	BPP	Crucimusculata	BPP	Prosopharyngida	BPP	Aberrantopermata	BPP
1	Size	?	—	?	—	?	0.75	Small	0.75	?	—	?	—
2	Body shape	Cylindrical	0.93	Cylindrical	0.95	Cylindrical	0.95	Cylindrical	1	Cylindrical	0.96	?	—
3	Threadlike	No	0.76	No	0.71	No	0.76	No	0.8	No	0.76	No	0.78
4	Chordoid vacuoles	Absent	0.98	Absent	0.98	Absent	0.98	Absent	0.89	Absent	0.99	Absent	0.96
5	Level of chordoid vacuoles	Xhordoid vesicles absent	0.86	Chordoid vesicles absent	0.85	Chordoid vesicles absent	0.83	Chordoid vesicles absent	0.71	Chordoid vesicles absent	0.95	Chordoid vesicles absent	0.83
6	Frontal glands	Present	0.93	Present	0.85	Present	0.95	Present	0.96	Present	0.95	Present	0.95
7	Level of frontal glands	?	—	?	—	Well developed	0.76	Well developed	0.97	?	—	?	—
8	Symbiotic algae	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	0.68
9	Sagittocysts	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1
10	Pigmentation	Absent	0.81	Absent	0.73	Absent	0.8	Absent	0.7	Absent	0.82	Absent	0.62
11	Eyes	Absent	0.99	Absent	0.99	Absent	0.99	Absent	1	Absent	0.99	Absent	0.7
12	Mouth position	postero-terminal or sub-terminal	0.69	Antero-terminal or sub-terminal	0.75	Antero-terminal or sub-terminal	0.75	Antero-terminal or sub-terminal	0.82	Antero-terminal or sub-terminal	0.78	Ventral	0.9
13	Pharynx	Present	0.98	Present	0.93	Present	0.97	Present	0.92	Present	1	Absent	0.97
14	Interconnecting cells	?	—	?	—	?	—	?	—	?	—	?	—
15	Testis	?	—	Paired	0.92	Paired	0.83	Paired	0.98	?	—	Paired	0.99
16	Seminal vesicle	?	—	?	—	?	—	?	—	?	—	?	—
17	Glandomuscular eversible copulatory organ	absent	0.75	?	—	?	—	Absent	0.76	Present	0.72	Present	0.8
18	Copulatory organ partly inside seminal vesicle	No	0.91	No	0.88	No	0.84	No	0.95	No	0.76	No	0.95
19	Copulatory organ invaginated into seminal vesicle	No	0.99	No	0.97	No	0.95	No	0.98	No	0.91	No	0.99
20	Shape of glandomuscular copulatory organ	?	—	?	—	?	—	?	—	Spherical	0.72	?	—
21	Penis stylet	Present	0.95	Present	0.96	Present	0.95	Present	0.95	Present	0.95	Present	0.97
22	Accessory penetrating hard structures	Absent	0.7	Absent	0.72	Absent	0.74	Absent	0.87	Absent	0.74	Absent	0.63
23	Antrum	present	0.72	Present	0.72	Present	0.69	Present	0.97	?	—	Pbsent	0.92
24	Antrum ciliation	?	—	?	—	?	—	Ciliated	0.76	?	—	Antrum absent	0.85
25	Antrum shape	Long and tubular	0.95	Long and tubular	0.96	Long and tubular	0.94	Long and tubular	0.96	Long and tubular	0.86	Antrum absent	0.93

Continued

TABLE 4. Continued

No.	Character	Acoela	BPP	Bitesticulata	BPP	Bursalia	BPP	Crucimusculata	BPP	Prosopha- ryngida	BPP	Aberranto- spermata	BPP
26	Male opening position	Posteroterminal or sub-terminal	0.7	Midbody	0.64	?	-	Posteroterminal 1	-	midbody	0.83	Posteroterminal or sub-terminal	0.88
27	Ovary	Unpaired	0.74	Unpaired	0.63	?	-	Unpaired	0.67	?	-	Paired	0.89
28	Bursa	Absent	0.64	?	-	Present	0.65	?	-	Present	0.74	Present	0.87
29	seminalis	?	-	?	-	?	-	?	-	?	-	?	-
30	Number of bursae	Absent	0.98	Absent	0.99	Absent	0.99	Absent	0.99	Absent	0.99	Absent	0.65
31	Bursal nozzle	Absent	0.97	Absent	0.98	Absent	0.98	Absent	1	Absent	0.98	?	-
32	Number of bursal nozzles	Absent	0.78	?	-	?	-	?	-	?	-	?	-
33	Vagina Separate female	Absent	0.78	Absent	0.75	Absent	0.71	Absent	0.98	?	-	Present	0.88
34	gonopore positioned or reaches behind testes	Yes	0.97	Yes	0.98	Yes	0.98	No	1	Yes	0.98	No	0.97
35	Hermaphrodite	?	-	No	0.89	No	0.9	No	0.91	No	0.83	No	0.99
36	gonad Asexual reproduction	Absent	1	Absent	0.99	Absent	1	Absent	1	Absent	1	Absent	1
37	Sperm	9 + 2	1	9 + 2	1	9 + 2	1	9 + 2	1	9 + 2	0.98	?	-
38	Sperm axonemes	Cortical	0.96	Cortical	0.95	Cortical	0.86	Cortical	0.97	?	-	?	-
39	cytoplasmic microtubules	Absent	0.88	Absent	0.68	Absent	0.81	?	-	Absent	0.98	?	-
40	Longitudinal muscles	Present	1	Present	1	Present	1	Present	1	Absent	0.69	Present	1
41	mouth	Absent	0.96	Absent	0.89	?	-	Present	1	Absent	0.97	Present	0.99
42	Ventral diagonal muscles	Absent	0.83	Absent	0.69	Absent	0.79	Absent	0.64	Absent	0.99	Present	0.95
43	Dorsal diagonal muscles	Absent	1	Absent	1	Absent	1	Absent	1	Absent	0.99	Absent	0.99
44	Longitudinal muscles outside of circular muscles	Absent	0.6	Absent	0.6	Absent	0.6	?	-	Present	0.68	Absent	0.75

TABLE 5. Reconstruction of family ancestors (BPP cut-off = 0.6)

Nr	Character	Diopisthoporidae	BPP	Paratomellidae	BPP	Hofstiemiidae	BPP	Solenoflo-morphidae	BPP	Proporidae	BPP	Dakuidae	BPP	Convolutidae	BPP	Mecynostomidae	BPP
1	Size	?	0.72	Conventional	0.7	Large	0.7	?	0.74	Small	0	?	0	?	Conventional	0.64	
2	Body shape	Cylindrical	0.99	Cylindrical	0.99	Cylindrical	0.99	Cylindrical	0.99	Cylindrical	0.99	Cylindrical	0.99	Convolutated	0.79	Cylindrical	0.99
3	Threadlike	No	0.91	Yes	0.84	No	0.84	No	0.8	No	0.8	No	No	No	No	No	0.79
4	Chordoid vesicles	Absent	0.98	Absent	1	Absent	1	Absent	0.98	Absent	0.78	Present	0.79	Absent	1	Absent	0.81
5	Level of chordoid vesicles	Chordoid absent	0.93	Chordoid vesicles absent	0.99	Chordoid vesicles absent	0.99	Chordoid vesicles absent	0.98	Chordoid vesicles absent	0	?	0	Chordoid vesicles absent	0.99	Chordoid vesicles absent	0.66
6	Frontal glands	Present	0.97	Absent	1	Present	0.99	Present	0.81	Present	0.88	Present	0.97	Present	0.95	Present	0.99
7	Level of frontal glands	?	0	Frontal glands absent	0.99	Weakly developed	0.98	?	0	Well developed	0	Well developed	0.9	Weakly developed	0.95	Well developed	0.85
8	Symbiotic algae	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Present	0.96	Absent	1.00
9	Sagittocysts	absent	1	Present	1	Absent	1	Absent	1	Absent	1	Absent	1	1	Absent	1.00	0.95
10	Pigmentation	Absent	0.88	Present	0.61	Present	0.99	Absent	0.92	Present	0.89	Absent	0.85	Present	0.97	Present	0.99
11	Eyes	Absent	0.99	Absent	1	Absent	1	Absent	0.99	Absent	0.99	Absent	0.94	Ocelli with platelets	0.75	Absent	0.99
12	Mouth position	Posteroterminal or subterminal	0.88	Ventral	0.96	Anteroterminal or subterminal	0.97	Ventral	0.93	Anteroterminal or subterminal	0.85	Ventral	0.86	Ventral	0.95	Ventral	0.95
13	Pharynx	Present	1	Absent	0.99	Present	1	Present	1	Present	0.99	Absent	0.95	Absent	0.99	Absent	0.99
14	Interconnecting cells	?	0	Absent	0.62	Present	0.68	Present	0.61	?	0	?	0	?	0	?	0.00
15	Testis	Unpaired	0.98	Paired	1	Unpaired	0.68	Unpaired	1	Paired	0.75	Paired	0.98	Paired	0.99	Paired	0.99
16	Seminal vesicle	?	0	Present	0.67	Present	0.7	?	0	?	0	?	0	Absent	0.62	?	0.00
17	Glandomuscular eversible organ	Absent	0.89	Absent	0.98	Present	0.99	Absent	0.77	Absent	0.85	Absent	0.77	Present	0.86	Present	0.67
18	Copulatory organ partly inside seminal vesicle	No	0.95	No	0.99	No	0.88	No	0.91	No	0.65	No	0.95	No	0.85	No	0.98
19	Copulatory organ invaginated into seminal vesicle	No	0.99	No	1	No	1	No	0.96	No	0.84	No	0.99	No	0.94	No	1.00
20	Shape of glandomuscular organ	glandomuscular copulatory organ absent	0.65	Glandomuscular copulatory organ absent	0.92	Spherical	0.98	?	0	Glandomuscular copulatory organ absent	0.79	?	0	Conical	0.98	Globular papilla	0.87
21	Penis stylet	Absent	0.87	Present	1	Present	1	Absent	0.96	Absent	0.97	Present	0.99	Absent	0.97	Present	0.99
22	Accessory penetrating hard structures	Absent	0.74	Absent	0.88	Absent	0.9	Absent	0.82	Absent	0.85	Absent	0.74	?	0	Absent	0.85

Continued

TABLE 5. Continued

Nr	Character	Dipisthoporidae	Paratomelliidae	BPP	Hofsteniidae	Solenofilomorphidae	Preporidae	BPP	Isodiametriidae	BPP	Dakuiidae	BPP	Convolutidae	BPP	Mecynostomidae	BPP
23	Antrum	Present	Present	0.74	Present	0.98	Present	0.94	Present	0.84	Absent	0.88	Absent	0.93	Absent	0.96
24	Antrum	?	Ciliated	0	Ciliated	0.83	Ciliated	0.72	Ciliated	0.65	Antrum	0.8	Antrum	0.83	Antrum	0.91
25	Antrum	Long and tubular	Long and tubular	0.89	Long and tubular	1	Long and tubular	0.99	Long and tubular	0.69	Antrum	0.91	Antrum	0.89	Antrum	0.98
26	Male opening position	Postero-terminal or Sub-terminal	Midbody	0.94	Anterotermininal or subterminal	1	Postero-terminal or subterminal	0.93	Postero-terminal or subterminal	0.98	Postero-terminal or subterminal	0.91	?	0	Postero-terminal or subterminal	0.95
27	Ovary	Unpaired	Unpaired	0.85	Unpaired	0.71	Unpaired	0.85	Paired	0.83	Paired	0.82	Paired	0.94	Paired	0.86
28	<i>Bursa seminalis</i>	Absent	Absent	0.81	Absent	0.94	Absent	0.84	Present	0.74	Present	0.84	Present	0.9	Present	0.91
29	Number of bursae	?	Absent	0	Absent	0.71	?	0	?	0	?	0	?	0	One	0.61
30	Bursal nozzle	Absent	Absent	0.98	Absent	1	Absent	1	Absent	0.88	Present	0.97	Present	1	Absent	1.00
31	Number of bursal nozzles	Absent	Absent	0.97	Absent	1	Absent	0.99	Absent	0.95	One	0.81	Several	0.71	Absent	1.00
32	Vagina	?	?	0	Absent	0.62	?	0	?	0	?	0	?	0	?	0.00
33	Separate female gonopore	Absent	Absent	0.85	Absent	0.98	Absent	0.89	Absent	0.96	Present	0.83	Present	0.87	Present	0.87
34	Ovary positioned or reaches behind testes	No	Yes	0.93	Yes	1	No	1	No	0.98	No	0.95	No	0.99	No	0.98
35	Hermaphrodite gonad	Yes	No	0.99	No	1	No	1	No	1	No	0.99	No	1	No	1.00
36	Asexual reproduction	absent	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1.00
37	Sperm axonemes	9 + 2	9 + 2	0.97	9 + 2	0.99	9 + 2	0.96	9 + 2	0.99	9 + 2	0.97	9 + 0	1	9 + 1	0.98
38	Sperm cytoplasmic microtubules	Cortical	Cortical	0.9	?	0	Cortical	0	Cortical	0.85	Axial	0.64	Axial	0.93	Distal only	0.88
39	Longitudinal muscles	absent	Present	0.97	Absent	0.99	Absent	0.98	Present	0.92	Present	0.77	Present	0.91	?	0.00
40	Ventral mouth	Absent	Present	0.71	Absent	0.9	present	0.86	Present	0.99	Present	1	Absent	0.86	Present	1.00
41	Diagonal muscles	Absent	Absent	0.96	Absent	0.99	Present	1	Present	0.99	Present	0.99	Present	0.99	Present	0.96
42	Ventral crossover muscles	Absent	Present	0.98	Absent	0.99	Absent	1	Present	0.93	Present	0.91	Present	0.93	Present	0.97
43	Diagonal muscles	Absent	Absent	0.99	Absent	1	Absent	1	Absent	1	Absent	1	Absent	1	Absent	0.98
44	Longitudinal muscles outside of circular muscles	Present	Present	0.73	?	0	Present	0.82	Present	0.83	Present	0.63	Absent	0.6	Absent	0.88
	Longitudinal muscles between mouth and frontal pore															

Phylogenetic Analysis of Morphological Data

The Bayesian phylogenetic analysis of the morphological data is presented as a majority-rule consensus tree in Figure 4. A few traditional clades are recovered,

but families such as Isodiometridae, Mecynostomidae, and Proporidae are unresolved at the 0.5 posterior probability cutoff level. In a consensus tree where all compatible clades are resolved, Mecynostomidae groups with *Childia* and *Anaperus* groups with Convolutidae

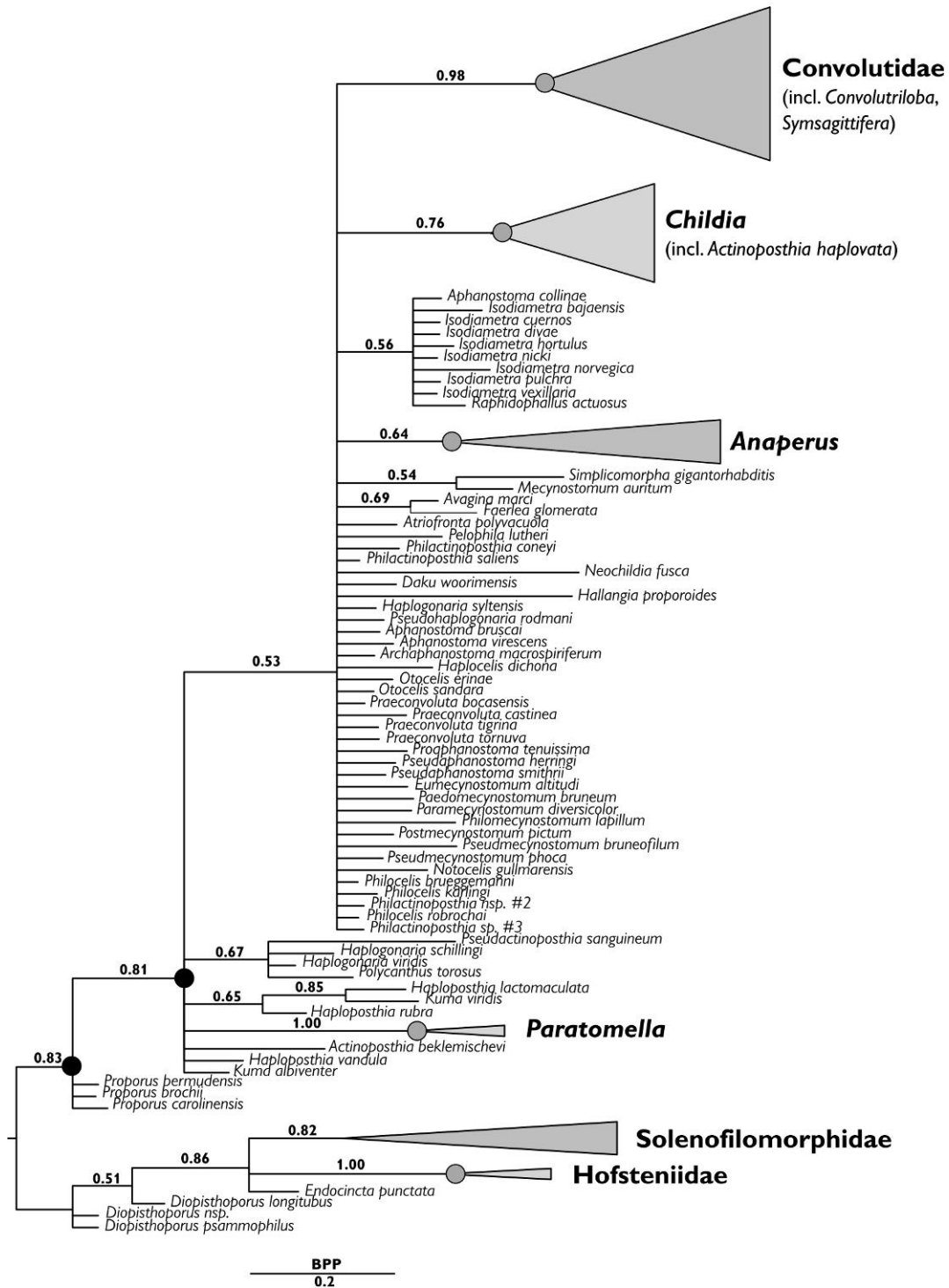


FIGURE 4. Bayesian majority-rule consensus tree inferred from morphological characters. Bayesian posterior probabilities are shown in bold above branches. Some families and genera are monophyletic, whereas the relationships among isodiometriids, dakuiids, and mecynostomiids are completely unresolved. *Paratomella* and *Hallangia* appear more deeply nested in the tree than in the molecular analyses.

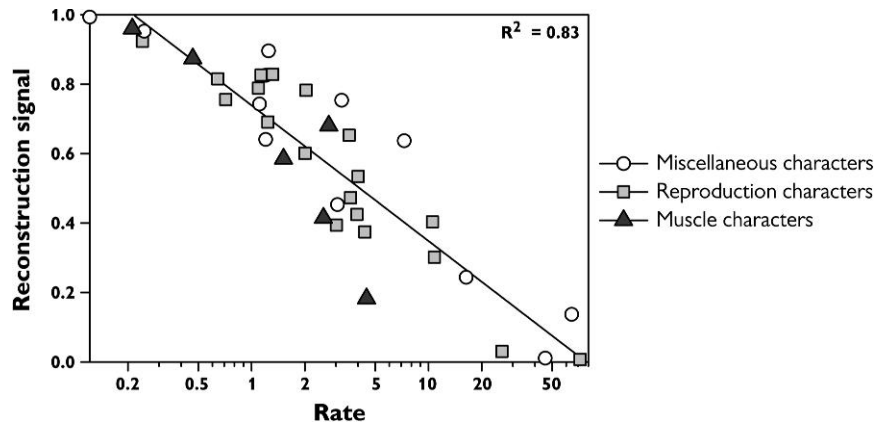


FIGURE 5. The correlation between reconstruction signal and evolutionary rates of morphological characters. There is a relationship between the rate of a character and how clearly it is reconstructed at the control nodes. Characters related to the reproductive organs cover a wider range of rates and signals than those of the body-wall musculature.

(see Supplementary Material) similar to the molecular data. Apart from the nested positions of *Paratomella* and *Hallangia*, there is no strongly supported conflict with the molecular data. It is evident that those morphological characters that we were able to score across all of Acoela are not sufficiently informative to recover a fully resolved tree alone.

Reconstruction of Ancestral Morphology

The insignificant topological disagreements among the alternative sets of posterior distributions of trees produced from the different alignments naturally result in striking similarities among the features of morpho-

logical evolution selected for comparison: (i) the rates and reconstruction signals of all characters are highly concordant among the sets (see Supplementary Data); and (ii) only 2 characters out of 44 are reconstructed with alternative states with posterior probabilities of 0.70 or higher, which occurs at only a single control node out of 30 (hence conflicts are restricted to two cells in a 1320-cell matrix). This ambiguity relates to details of the copulatory organ inferred at a node inside Isodiametridae depending on whether or not the clade containing many actinoposthiids groups inside Isodiametridae with high support (Figs. 2 and 8a,b and the Supplementary Data). As a consequence of these uniform results, we again chose to focus on the results inferred from the Bayesian trees generated from the filtered structural

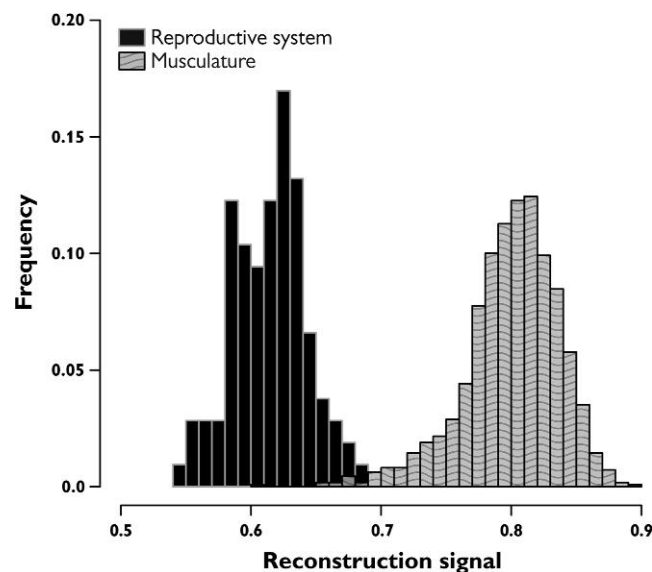


FIGURE 6. The reconstruction signals of the reproductive system and musculature. The characters of the body-wall musculature are reconstructed with significantly stronger signals than those of the reproductive system (Wilcoxon rank-sum test; pooled data: $p(H_0) < 2.2e^{-16}$). The histograms represent the distribution of mean reconstruction signals estimated for the reproductive system (19 characters) on 106 fully sampled trees and the musculature (six characters) on 1109 fully sampled trees, respectively, out of 2000 Bayesian input trees.

TABLE 6. Proportion of successfully reconstructed states at different nodes and confidence levels

Node	Confidence level				
	0.5	0.6	0.7	0.8	0.9
Higher taxon					
Acoela	0.93	0.77	0.70	0.55	0.45
Bitesticulata	0.91	0.77	0.66	0.52	0.41
Bursalia	0.91	0.75	0.68	0.52	0.41
Prosopharyngida	0.91	0.73	0.68	0.52	0.43
Hofsteniidae + Solenofilomorphidae	0.91	0.75	0.68	0.52	0.45
Crucimusculata	1.00	0.81	0.70	0.65	0.57
((Abberantospemata + Dakuidae) + Otocelis) + Isodiametridae + "Avaginaclade"	0.89	0.73	0.52	0.41	0.32
((Abberantospemata + Dakuidae) + Otocelis) + Isodiametridae	0.91	0.77	0.66	0.59	0.45
(Abberantospemata + Dakuidae) + Otocelis	0.91	0.80	0.68	0.59	0.48
Abberantospemata + Dakuidae	0.93	0.77	0.70	0.61	0.52
Abberantospemata	0.93	0.73	0.61	0.57	0.41
Average	0.92	0.76	0.66	0.55	0.45
Family					
Diopisthoporidae	0.98	0.84	0.82	0.70	0.52
Paratomellidae	1.00	0.98	0.86	0.82	0.70
Hofsteniidae	0.95	0.95	0.86	0.80	0.68
Solenofilomorphidae	0.95	0.84	0.80	0.75	0.59
Proporidae	1.00	0.91	0.86	0.73	0.57
Isodiametridae	0.93	0.86	0.77	0.70	0.48
Dakuidae	0.98	0.84	0.80	0.68	0.48
Convolutidae	0.95	0.86	0.82	0.73	0.59
Mecynostomidae	0.98	0.91	0.82	0.80	0.61
Average	0.97	0.93	0.86	0.78	0.66

data set. The overall reconstruction signals of all characters are plotted as a function of their evolutionary rates in Figure 5 (see Supplementary Material for the corresponding data table and detailed transition rates among states). The characters span a wide range of rates and, as expected, more rapidly evolving characters have less

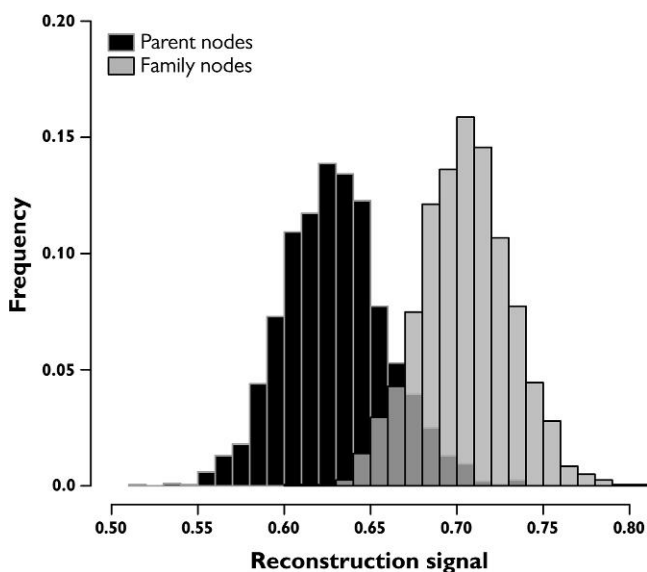


FIGURE 7. The reconstruction signals of all characters at different sets of nodes. Nodes corresponding to families are reconstructed with significantly stronger signals than their deeper parent nodes (Wilcoxon rank-sum test; pooled data: $p(H_0) < 2.2e^{-16}$). The histograms represent the distribution of mean reconstruction signals estimated for families (9 nodes) and parent nodes (11 nodes) on the same set of 2000 Bayesian input trees.

reconstruction signal at the control nodes. Although the characters of the body-wall musculature generally have lower rates, the rates of characters related to the reproductive system range across the whole spectrum. Some features of the reproductive organs, such as the absence or presence of a vagina or seminal vesicle are reconstructed with extremely high rates to fit their patchy distribution in extant taxa and provide virtually no signal at all as to what state would be most probable in the deeper parts of the tree. The characters of the muscle system are significantly more clearly reconstructed than those of the reproductive system at the control nodes (pooled data: $p(H_0) < 2.2e^{-16}$; tree-by-tree: 55 out of 55 trees $p(H_0) < 0.05$; see also Fig. 6), but the difference appears to be smaller at the family nodes than the deeper backbone nodes (see Supplementary Material for a more detailed breakdown of all data samples). Some of these family nodes, such as the Mecynostomidae and the Proporidae, show the opposite pattern in that the genitals are reconstructed with a slightly stronger signal than the muscles (Fig. 8a,b). The reconstructions of the acoel root and other deep nodes are presented in Table 4 and the reconstructions of the family ancestors are presented in Table 5 (see Supplementary Material for the reconstructions at all control nodes). The hypothetical common ancestor of a recognized family, such as that of Mecynostomidae or Convolutidae, is typically more clearly reconstructed than that of a deeper node (pooled data: $p(H_0) < 2.2e^{-16}$; tree-by-tree: 1247 out of 2000 trees $p(H_0) < 0.05$; no tree with significant support for the opposite pattern; see also Table 6 and Fig. 7), such as Abberantospemata, the parent node of both Mecynostomidae or Convolutidae (Fig. 8a,b). There is a clear relationship between cutoff levels of credibility

in the reconstructions and the proportion of characters that are successfully reconstructed at that level (Table 6). In the Bayesian analysis, the pharynx was inferred to be homologous among all families with high posterior probabilities (Tables 4 and 5) and alternative hypotheses were associated with likelihood values orders of magnitudes lower (Table 7).

DISCUSSION

Phylogeny

We were able to reconstruct a comprehensive and well-resolved phylogeny of Acoela using new and previously published ribosomal and mitochondrial sequence data from over 120 acoel species, about one-third of the described species. The tree recovered from

analysis of the mitochondrial COI gene is less well resolved than the ribosomal gene trees but is generally compatible with the tree of all combined data. In addition to what was previously understood about acoel interrelationships, there is now strong evidence for: (i) Diopisthoporidae being the sister taxon to all other acoels; (ii) a clade uniting *Hallangia*, Hofsteniidae, and Solenofilomorphae; (iii) the close relationship between *Proporus*, most haploposthiids and *Polycanthus*; (iv) the polyphyly of Actinoposthiidae with taxa distributed among Dakuidae and Mecynostomidae and possibly inside Isodiametriidae; (v) a clade uniting *Daku*, *Notocelis*, *Philactinoposthia*, and *Philocelis*; (vi) *Anaperus*, *Symsagittifera*, and *Convolutriloba* being part of Convolutidae; (vii) *Childia* being nested inside Mecynostomidae; and (viii) Mecynostomidae and Convolutidae being

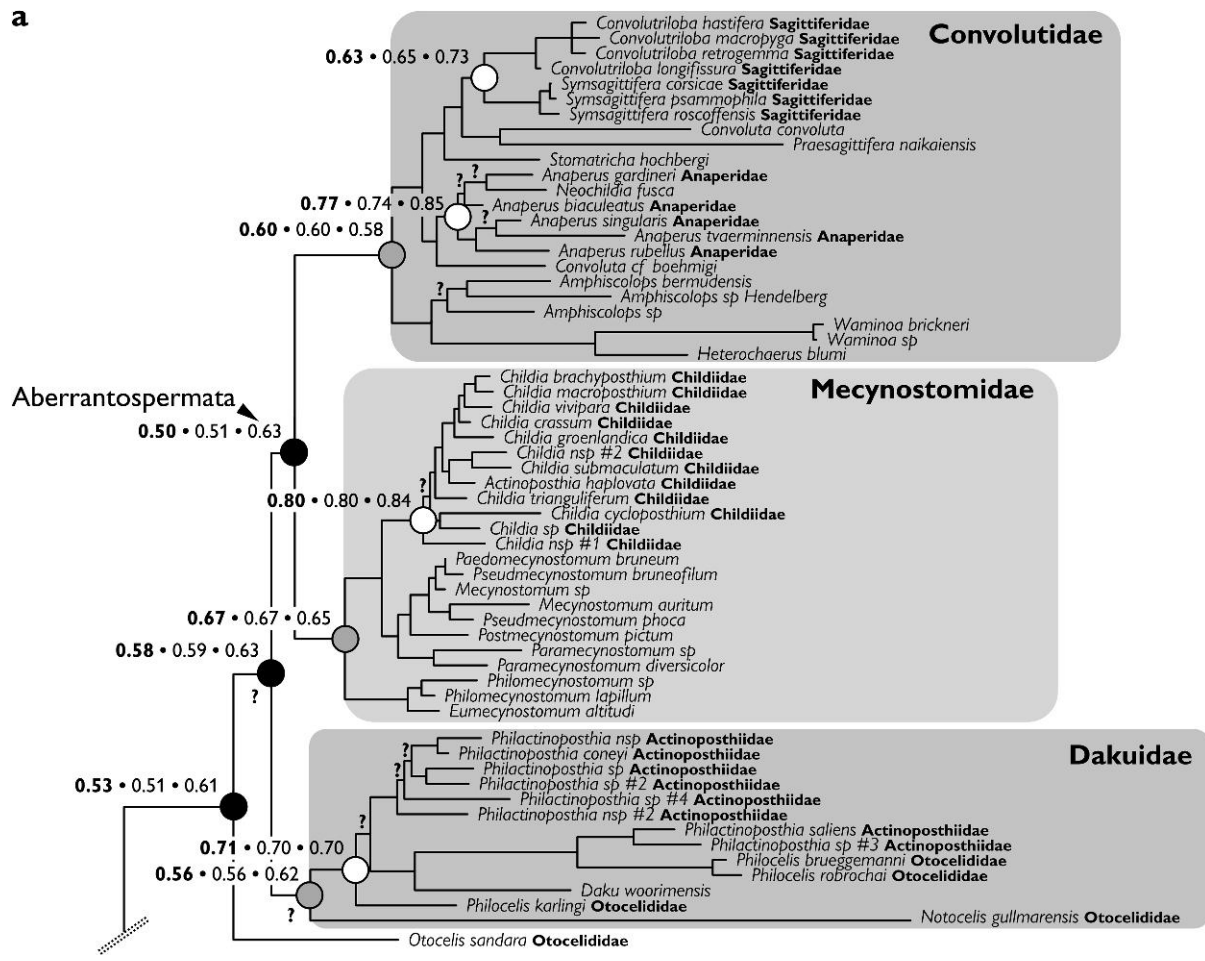


FIGURE 8. Detailed presentation of the acoel phylogeny. Total-evidence Bayesian majority-rule consensus tree inferred from 18S, 28S, and COI. All control nodes are displayed. Nodes corresponding to families are highlighted as gray, their deep parent nodes are black, and other control nodes are white. Nodes for which Bayesian posterior probabilities are below 0.90 are marked with a question mark. The overall reconstruction signals and the averaged reconstruction signals of the reproductive organs and body-wall musculature are displayed at all control nodes. Families in dotted boxes. Species transferred from other families or recovered outside their expected family clade are highlighted with their old family names in bold. Family nodes generally have a higher overall reconstruction signal than their parent nodes and body-wall musculature is generally reconstructed with stronger signals than the reproduction system. A mixed clade with actinoposthiids and isodiametriids with unstable position among analyses of different alignments is marked with an asterisk. The positions of *Hallangia proporoides* and *Actinoposthia beklemischevi* + *Pseudactinoposthia sanguineum* are associated with drops in the reconstruction signals of the reproductive organs.

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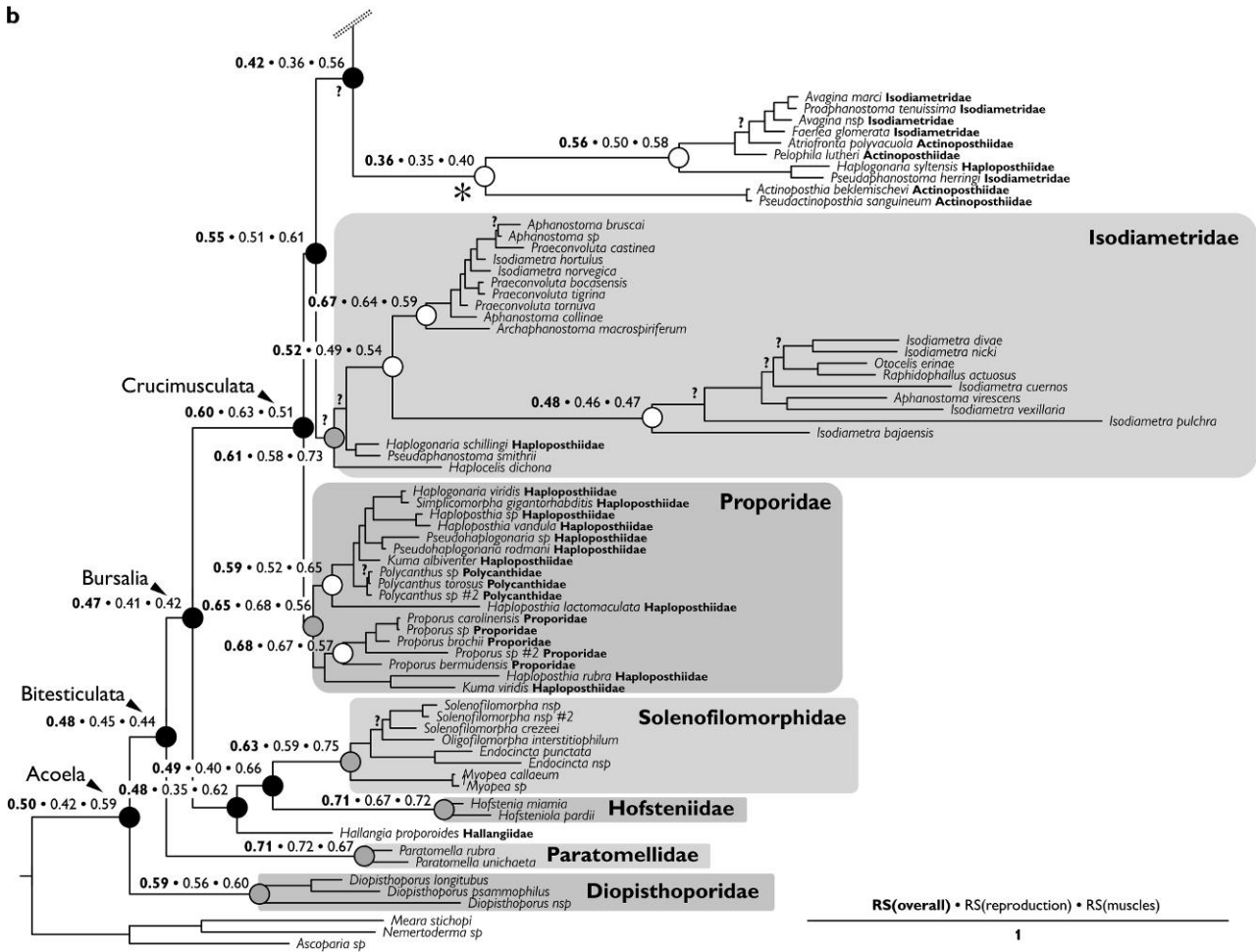


FIGURE 8. Continued

sister taxa. Although a few monotypic families are missing from our analysis, we believe that it is now possible to trace the evolution of morphological traits in Acoela with some accuracy.

Reconstruction of Ancestral Morphology

By using a Bayesian approach to character evolution, we aim to accommodate uncertainties in topological relationships and evolutionary distances among nodes and species when inferring ancestral states. Such uncertainties appear to be low in our phylogenetic inferences, given the currently available molecular data, alignment techniques, and the substitution models that we applied. The Bayesian approach to inference does not assign fixed synapomorphies to all clades. It instead assumes that all states are possible at all nodes and that their posterior probabilities are proportional to how well they fit the data and the model. Although this aims for more realistic inferences of evolutionary properties of the data when the true genealogy and evolutionary processes are not known, it leaves the job of assigning character states and changes to the observer. In the end, only one of several mutually exclusive states was actually present in an ancestral organism. To decide

if variations in posterior probabilities of states among adjacent nodes represent different states at these nodes, we have to find a level of confidence that balances credibility and information content. As we show, there is a correlation between the level of confidence and the number of characters available for discussion. We here focus our discussion of ancestral morphological features on characters with states reconstructed with average posterior probabilities of close to 0.7 or higher. This corresponds to a reconstruction signal of 0.12 or higher in a binary character.

The ancestral acoel was likely a cylindrical worm with frontal glands and a pharynx located in the posterior end of the body (Fig. 9 and Table 4). Chordoid vacuoles were absent. The body-wall musculature consisted of an orthogonal muscle grid with additional ventral diagonal fibers. It had an unpaired ovary, which may have formed a diffuse mixed gonad with the testis. There was likely a tubular antrum through which the sperm was transported to a posteroterminal or subterminal male opening. The spermatozoa had a 9 + 2 arrangement of axonemes and cortical microtubules. Copulation may have been assisted with a stylet but other accessory penetrating muscular or hard structures

TABLE 7. Evaluation of alternative hypotheses on the evolution of the acoel pharynx

Node constraint(s)	Mean of log Lh	Harmonic mean (H) of log Lh	Test statistic
None	-13.51	-14.59	-
Proporidae = absent	-14.80	-16.12	3.07
Acoela = absent	-16.32	-17.10	5.03
Bursalia = absent	-16.00	-17.28	5.39
H + S = absent, Prosopharyngida = absent	-21.25	-22.09	15
H + S = absent	-21.22	-22.09	15
Acoela, Bitesticulata, Bursalia, Sphinctostomata, Prosopharyngida, H + S = absent	-21.58	-22.85	16.53

were absent. Female accessory organs such as a separate female gonopore, bursa, and bursal nozzle were absent. Traits mainly associated with Convolutidae such as symbiotic algae, sagittocysts, adenodactyls, pigmentation, and eyes were absent. Extant species of *Diopisthoporus* possess many of these ancestral traits and of all the families included in the analysis, the node corresponding to Diopisthoporidae has the posterior distribution of states that is the most similar to the posterior distributions inferred at the root of the tree. Our parsimony-based reconstruction of unambiguous changes in the ancestral acoel (morphological characters optimized on Bayesian molecular tree) implies a cylindrical body with frontal glands, without chordoid vacuoles, orthogonal body-wall musculature without ventral diagonal fibres, an unpaired ovary, a ciliated male antrum, and 9 + 2 arrangement of axonemes and cortical microtubules in the spermatozoa. The parsimony reconstruction further entails absence of a copulatory stylet, absence of vagina, bursa and bursal nozzles, and absence of pigmentation, eyes, or symbionts. In contrast to the Bayesian reconstruction, the presence or absence of a pharynx, position of the mouth, and existence of a mixed gonad could not be determined under the parsimony framework. Except for the presence of a muscular pharynx, our Bayesian estimate of character states at the acoel root is largely consistent with Westblad's list of ancestral acoel features: a mixed gonad, lack of accessory female structures, and posteroterminal gonopore were all regarded as "primitive" by Westblad (1948). Many of the ancestral features, including the cylindrical body with frontal

glands, pharynx, the indistinct division between testes and ovary (a hermaphrodite gonad), tubular antrum opening into a posterior gonopore, penis stylet, and 9 + 2 sperm axoneme structure with cortical microtubules occur in deep nodes and "basal" families. In this context, the basal position of *Hallangia proporoidea* is unexpected. This species stands out as it has paired ovaries that do not mix with the testes and its copulatory organ and chordoid vesicles are similar to those of Isodiametridae. Consequently, some of the ancestral features shared among many deep nodes cannot be assumed to be homologous without recognizing some level of ambiguity in this region of the tree (see Supplementary Data for reconstructions with stronger signals without this species). The reconstruction of the ancestral acoel is nearly identical in the presence of Nemertodermatida (see Supplementary Data), with the exception that the position of the mouth opening is unresolved and that the testis may have been paired structures (BPP = 0.65). However, the sister group relationship between Acoela and Nemertodermatida (Ehlers 1985) was recently challenged (Wallberg et al. 2007) and we here prefer to infer ancestral features of Acoela from extant acoel species.

In our Bayesian reconstruction, the pharynx of Diopisthoporidae, *Hallangia*, Solenofilomorphae, Hofsteniidae, and *Proporus* is homologous. This provides an alternative to the hypothesis on the origin the acoel pharynx put forward by Todt (2009). Todt argued, based on anatomical differences in extant species, that the pharynges of different families had evolved independently, and she found it unlikely that a pharynx would have

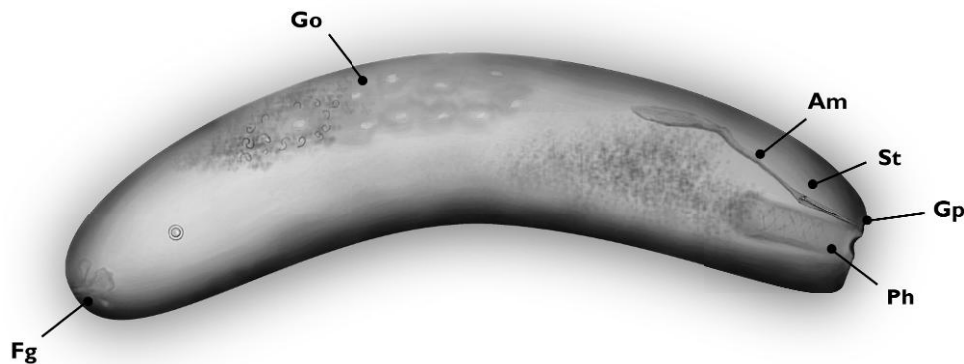


FIGURE 9. A representation of the ancestral acoel worm as inferred from molecular and morphological analyses. Fg = frontal glands; Go = mixed hermaphroditic gonads; Am = tubular antrum; St = copulatory stylet; Gp = posteroterminal gonopore; Ph = posteroterminal muscular pharynx. Illustration: Karolina Larsson. This figure is available in black and white in print and in color at *Systematic Biology* online.

been present in the acoel ancestor. Our phylogenetic analyses suggest the opposite: the pharynx is a feature that likely evolved only once ($q_0 \rightarrow 1 = 0.015$) but was lost multiple times ($q_1 \rightarrow 0 = 1.19$): once in *Paratomella*, twice in the clade comprising *Proporus* and various haploposthiids and once in the ancestor of Isodiametridae and its sister taxon (Tables 4 and 5; Fig. 8a,b). Multiple independent origins of the pharynx is a far less likely hypothesis (Table 7). To accommodate fully independent origins of pharynges among families, the relative transition rates between the absence and presence of a pharynx need to be reversed ($q_0 \rightarrow 1 = 0.218$ and $q_1 \rightarrow 0 = 0.083$, respectively). Under our reconstruction, the variation in pharynx tube muscle layers and associated tissues among the groups are better interpreted as autapomorphies. Most extant acoels lack a pharynx, but this condition is often associated with the presence of additional ventral muscles oriented in such a way that they could pull the posterior part of the body and mouth forward, forcing food items into the mouth. Thus, ventral longitudinal muscles crossing over from one side to the other just behind the mouth or U-shaped fibers enveloping the mouth may substitute for a specialized pharynx (Tyler and Rieger 1999). Is the gain of such muscles correlated with the loss of the pharynx in our reconstruction? U-shaped fibers have been found in the basal but pharynx-less *Paratomella* and most Crucimusculata species studied to date, but not in any acoel with a pharynx (Todt 2009). Loss of the pharynx represents independent events in these lineages and the appearance of U-shaped fibers are accordingly inferred to be convergent in our reconstructions. Todt (2009) noted that the U-shaped muscles of *Paratomella unichaeta* form a distinct layer of fibers not connected to the ventral longitudinal fibers, which is otherwise the case in Crucimusculata; a difference that could be explained by independent origins of these fibers among these lineages. Ventral crossover muscles are not present in the basal and pharyngeate Diopisthoporidae, Solenofilomorphidae, and Hofstenidae and is here inferred to have evolved only once along the lineage leading to Crucimusculata. The Bayesian inference reconstructs the pharynx of *Proporus* as homologous to that of the other acoels, whereas the parsimony analysis infers a separate origin. Although the latter hypothesis is not the preferred explanation in the Bayesian analysis, it is the best-fitting alternative solution among those explored here (Table 7). It should be noted that the pharynx in *Proporus* appears different from other acoel pharynges when observed in live specimens and some unique features such as the presence of pharynx retractors and the absence of oral sphincter muscles have been reported from *Proporus bermudensis* (Todt 2009), the single species of the genus that has been studied in detail. Although *P. bermudensis* does have the ventral crossover musculature associated with Crucimusculata, other ventral fibers of the orthogonal muscle grid shared among most acoels appear to be secondarily reduced (Hooge 2001). The pharynx and elaborate ventral muscle fibres seem to be coevolving and mutually exclusive. The pharynx is

likely an ancestral feature of the acoel feeding apparatus, but a separate origin of the divergent pharynx in *Proporus* cannot be completely ruled out, especially as it is consistent with the parsimony-based reconstruction. Comparative studies of the developmental program as well as the feeding mechanics and ecology in *Diopisthoporus*, *Hofstenia*, *Proporus*, and other acoels are needed to further elucidate the evolution of the acoel pharynx.

From our attempts to evaluate the reconstruction performance of characters related to the body-wall musculature relative to those of the reproductive system, we conclude that the musculature is generally easier to reconstruct at deep nodes than many features of the reproductive system, that is, they make up a more conservative set of characters. The difference was significant also when only species completely coded for both organ systems were included in the analyses (Supplementary Material). The higher evolutionary rates of reproductive system characters are consistent with the results of Achatz et al. (2010), who found indications of a high rate of coevolution between sperm morphology, secondary female reproductive organs and speciation in Convolutidae. When averaged over all characters, deeper nodes tend to have a weaker reconstruction signal than nodes at the family level. The family level nodes can be regarded as “outpost” nodes for which many aspects of the ancestor can still be reconstructed with high confidence. These nodes typically contain all sampled representatives of the family according to the traditional classification but in some cases also additional species, or in the case of Dakuidae constitute a completely new configuration of species. Although recovered in a nested position, Proporidae retains many plesiomorphic states and therefore has a high posterior similarity to the ancestral acoel (PS(non-structural) = 0.82; PS(structural) = 0.79), second only to Diopisthoporidae. In the analysis of only the morphological data sets, *Proporus* was basal even to *Paratomella*, even though the latter taxon holds a basal position in the molecular tree. Convolutidae (including *Anaperus* and *Sagittiferidae*) on the other hand are the acoels that diverged the most from the ancestral acoel with a posterior similarity score of only 0.50.

Our results appear to be robust with respect to the small differences in the posterior distributions of trees inferred from molecular data sets aligned with different algorithms and optional filtering of ambiguous regions. Furthermore, they underscore the importance of broad taxonomic sampling and an informed choice of “models” for in-depth studies of for example, developmental biology in Acoela. Inferences about ancestral properties of Acoela should not be drawn from those species that share the fewest features with the common ancestor of Acoela. It is ironic to note that recent studies drawing conclusions about the developmental origin of the mouth and gonopores in acoels (Hejnol and Martindale 2008), the architecture of the ancestral acoel muscular system (Semmler et al. 2008), the phylogenetic significance of serotonergic nerve chords in Acoela (Gaerber et al. 2007), and the anatomy of the

TABLE 8. Family level classification of Acoela

Name and author	Nomenclatorial acts in this paper	Specifiers for recognized families	Emended diagnoses
Actinoposthiidae, Hooge 2001	<i>Philactinoposthia</i> is transferred to <i>Dakuidae</i> . <i>Actinoposthia haploata</i> is transferred to <i>Chilidia</i>	No specifiers can be provided as the taxon was insufficiently sampled	No changes
Anaperidae, Dörjes 1968	Anaperidae is synonymized with Convolutidae	No specifiers can be provided. This monotypic taxon was not available for study.	Synonymized No changes
Anthroposthiidae, Faubel 1976	None	No specifiers can be provided. This monotypic taxon was not available for study	No changes
Antigonariidae, Dörjes 1968	None	—	Synonymized
Chilidiidae, Dörjes 1968	Chilidiidae is synonymized with Mecynostomidae	The most inclusive clade that contains <i>Amphicolops bernudensis</i> and <i>Convolutriloba longifissura</i> but not <i>Chilidia macroposthium</i>	Spermatozoa with 9+0 axonemes. Pigmentation, algal symbionts and eye spots often present. Male antrum may have numerous stimulatory organs. Sagittocyts may be present. Includes large and active worms
Convolutidae, Graff 1905	Anaperidae and Sagittiferidae are synonymized with Convolutidae	—	Penis with sclerotized needles, muscular, or absent. Seminal vesicle may be highly muscular. Bursal nozzle present. Sperm with axial microtubules and 9+2 axonemes No changes
Dakuidae, Hooge 2003	<i>Philocelis</i> , <i>Philactinoposthia</i> , and <i>Notocelis</i> are transferred to this family.	The most inclusive clade that contains <i>Dakuaaorinensis</i> and <i>Philactinoposthia coneyi</i> but not <i>Convoluta convoluta</i>	No changes
Diopisthoporidae, Westblad 1940	None	The most inclusive clade that contains <i>Diopisthoporus longitubus</i> and <i>D. psammophilus</i> but not <i>Paratomella rubra</i>	No changes
Hallangriidae, Westblad 1946	None	No specifiers can be provided as this taxon was insufficiently sampled in this study	No changes
Haploposthiidae, Westblad 1948	Haploposthiidae is synonymized with Proporidae. <i>Haplogonaria schillingi</i> and <i>Haplogonaria syltensis</i> are transferred to Pseudophanostoma (Isodiametridae).	—	No changes
Hofstemiidae, Papi 1957	None	The most inclusive clade that contains <i>Hofstemia miamaia</i> and <i>H. paridi</i> but not <i>Myopea callacum</i> .	No changes
Isodiametridae, Hooge and Tyler 2005	<i>Haplogonaria schillingi</i> is transferred to <i>Pseudophanostoma</i> within this family.	The most inclusive clade that contains <i>Isodiametra norvegica</i> and <i>Haploctelis dictyona</i> , but not <i>Philactinoposthia saliensis</i>	Body-wall musculature with circular, longitudinal, and longitudinal crossover muscle fibers in both the dorsal and ventral body wall, and U-shaped fibers in the ventral body wall. Male copulatory organ invaginated into muscular seminal vesicle, usually a tubular muscular isodiametric penis, often glandular. Penis musculature with inner circular and outer nonanastomosing longitudinal fibers. Never with symbiotic algae. Ocelli, when present, do not contain platelets in the pigment cell
Mecynostomidae, Dörjes 1968	Chilidiidae is synonymized with Mecynostomidae	The most inclusive clade that contains <i>Paramecynostomum diversicolor</i> and <i>Chilidia macroposthium</i> , but not <i>Convolutriloba longifissura</i>	Spermatozoa with 9+1 axonemes and distal microtubules only. Four layers of dorsal body-wall muscles No changes
Nadiniidae, Dörjes 1968	None	No specifiers can be provided. This monotypic taxon was not available for study	No changes
Otocelididae	<i>Philocelis</i> and <i>Notocelis</i> are transferred to Dakuidae	No specifiers can be provided, as this taxon was insufficiently sampled in this study	No changes
Paratomellidae, Dörjes 1966	None	The most inclusive clade that contains <i>Paratomella rubra</i> and <i>P. unichuete</i> but not <i>Hofstemia miamaia</i>	No changes
Polycanthidae, Hooge 2003	Polycanthidae is synonymized with Proporidae. <i>Polycanthus</i> is transferred to Proporidae.	—	Synonymized
Proporidae, Graff 1882-kolla	Haploposthiidae and Polycanthidae are synonymized with Proporidae, <i>Haplogonaria viridis</i> is transferred to <i>Simplicomorpha</i>	The most inclusive clade that contains <i>Proporus brocchii</i> and <i>Polycanthus torosus</i> , but not <i>Haploctelis dichoma</i>	Male copulatory organ posteroterminal, seminal vesicle present. Pharynx, when present located anteriorly. Ovaries never reaching behind testes synonymized
Sagittiferidae, Kostenko and Mamkaev 1990	Sagittiferidae is synonymized with Convolutidae.	The most inclusive clade that contains <i>Oligofolomorpha interstitiophilum</i> and <i>Myopea callacum</i> but not <i>Hofstemia miamaia</i>	No changes
Solenofilomorphidae, Dörjes 1968	None	No specifiers can be provided. This monotypic taxon was not available for study	No changes
Taurididae, Kostenko 1989	None	—	No changes

brain (Bery et al. 2010) are all based on single species of the Sagittiferidae. Likewise, Ramachandra et al. (2002) made general inferences about acoel and early bilaterian morphogenesis based on observations restricted to *Neochildia fusca*. The stem cell system in Acoela and its phylogenetic implications were studied by Egger et al. (2009), who improved sampling by studying two species of Isodiametridae and one species of Sagittiferidae. Egger and coauthors suggested that the stem cell system in their acoel species and stem cells in Platyhelminthes constitute a synapomorphy conflicting with molecular phylogenies that place Acoela at the base of Bilateria (e.g., Telford et al. 2003; Wallberg et al. 2007; Dunn et al. 2008, Hejnol et al. 2009). Unfortunately, Isodiametridae does not appear to be a particularly plesiomorphic family. Some species of Sagittiferidae are among the few Acoela reported to reproduce asexually through fission (Åkesson et al. 2001), a process that requires considerable regenerative powers. This is a highly derived feature in our reconstruction. Thus, in addition to being part of the clade that diverged the most from the acoel ancestor, sagittiferids have special adaptations for regeneration. We propose that the similarity between stem cells in Platyhelminthes and stem cells in the Acoela studied by Egger and coworkers is best explained as convergence in view of the strong support for Acoela as a basal bilaterian clade in the DNA sequence-based studies, the specialized nature of sagittiferids, and the uncertainties regarding reconstruction of ancestral isodiametrid characters. We recommend that species from less divergent clades with high reconstruction scores

should be considered when choosing material for in-depth studies seeking to draw generalized conclusions about acoel biology and that such conclusions should be based on more than one species, spanning as much phylogenetic diversity as possible.

A formal reclassification of Acoela at the family level, consistent with the phylogenetic hypothesis presented here is given in Table 9. In order to achieve the goal of monophyletic family level groups, some reassignments of species and genera are necessary: all the formal nomenclatorial acts are reported in Table 8. We have kept the reclassifications to the minimum necessary for obtaining monophyletic family level taxa. Generally, the family level clades are strongly supported in the Bayesian, ML, and parsimony analyses. We have chosen not to reclassify the species of Actinoposthiidae that group within the Isodiametridae clade (Fig. 8a,b) as our sequence data for some of these species are incomplete. Their position is based solely on 18S rRNA sequences from GenBank and depending on underlying alignment algorithms of the molecular data, there are competing solutions indicating either inclusion of actinoposthiids in Isodiametridae or paraphyly of Isodiametridae with regard to a few isodiametrid species grouping with the actinoposthiids. Better sequence coverage and more thorough sampling preferably involving type species of the genera is required to determine the phylogenetic status of Actinoposthiidae, which we regard as unresolved at this point. A comprehensive phylogenetic system is essential for further studies of acoel biodiversity and evolution. Informal names for

TABLE 9. Phylogenetic classification of Acoela with diagnostic features and specifiers of suprafamilial clades

Clade
Acoela (digestive parenchyma, biflagellate spermatozoa)
Diopisthoporidae Westblad, 1940
Bitesticulata (Paired or follicular testes, ventral gonopore. The most inclusive clade that contains <i>Paratomella rubra</i> and <i>Symsagittifera roscoffensis</i> but not <i>Diopisthoporus longitubus</i>)
Paratomellidae Dörjes, 1966
Bursalia (Copulatory bursa often present. The most inclusive clade that contains <i>Oligofilomorpha interstitiophilum</i> and <i>Childia groenlandica</i> but not <i>Paratomella rubra</i>)
Prosopharyngida (Muscular pharynx in anterior part of body. The most inclusive clade that contains <i>Hofstena miamia</i> and <i>Oligofilomorpha interstitiophilum</i> but not <i>Haploposthia rubra</i>)
Hallangidae Westblad, 1946
Hofsteniidae Papi, 1957
Solenofilomorphidae Dörjes, 1968
Crucimusculata (Ventral crossover muscle fibres). The most inclusive clade that contains <i>Actinoposthia beklemishevi</i> and <i>Childia groenlandica</i> but not <i>Haploposthia rubra</i>)
Dakuidae Hooge 2003
Isodiametridae Hooge and Tyler 2005
Otocelididae Westblad, 1948
Proporidae Graff, 1882
Aberrantospemata (Spermatozoa with 9 + 0 or 9 + 1 axonemes. The most inclusive clade that contains <i>Neochildia fusca</i> and <i>Childia groenlandica</i> but not <i>Actinoposthia beklemishevi</i>)
Convolutidae Graff, 1905
Mecynostomidae Dörjes, 1968
Acoela Incertae cedis, not sampled in this study
Actinoposthiidae (see discussion)
Anthroposthiidae (single species)
Antigonariidae (single species)
Nadinidae (single species)
Tauridiidae (single species)

strongly supported higher level groups are introduced in Table 9, thereby completing a phylogenetic classification of Acoela down to the family level.

SUPPLEMENTARY MATERIAL

Supplementary material, including data files and/or online-only appendices, can be found at <http://www.sysbio.oxfordjournals.org/>.

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REFERENCES

- Abildgaard P.C. 1806. Zoologica danica seu animalium Danicae et Norwegiae rariorum ac mines notorum descriptorum et historia aus O. F. Mueller. Zool. Dan. 4:26.
- Achatz J.G., Hooge M.D., Tyler S. 2007. Convolutidae (Acoela) from Belize. Zootaxa. 1479:35–66.
- Achatz J.G., Hooge M.D., Wallberg A., Jondelius U., Tyler S. 2010. Systematic revision of acoels with 9+0 sperm ultrastructure (Convolutida) and the influence of sexual conflict on morphology. J. Zool. Syst. Evol. Res. 48:9–32.
- An der Lan H. 1936. Ergebnisse einer von E. Reisinger und O. Steinböck mit Hilfe des Rask-Orested Fonds durchgeführten Reise in Grönland 1926. 7. Acoela I. Vidensk. Medd. Dansk. Naturh. Foren. 99:289–329.
- Afzelius B.A. 1966. Anatomy of the cell. Chicago: University Chicago Press.
- Afzelius B.A. 1982. The flagellar apparatus of marine spermatozoa. Evolutionary and functional aspects. In: Amos W.B., Duckett J. G., editors. Prokaryotic and eukaryotic flagella. Cambridge: Cambridge University Press. p. 495–519.
- Åkesson B., Gschwentner R., Hendelberg J., Ladurner P., Mueller J., Rieger R. 2001. Fission in *Convolutriloba longifissura*: asexual reproduction in acelous turbellarians revisited. Acta Zool. (Stockh.) 82: 231–239.
- Bartolomaeus T., Balzer I. 1997. *Convolutriloba longifissura* nov. spec. (Acoela)—the first case of longitudinal fission in Plathelminthes. Microfauna Mar. 11: 7–18.
- Beklemishev V.N. 1915. Sur les turbellariés parasites de la cote Mouri-manne. I. Acoela. Trav. Soc. Imp. Natur. Petrograd. 43:103–172.
- Beklemishev V.N. 1957. *Convoluta psammophila* sp. nov. and the tendency toward juvenile oligomerisation of cellular elements in Turbellaria Acoela. Tr. Leningr. O-va Estestvoispyt. 73: 5–13.
- Bery A., Cardona A., Martinez P., & Hartenstein V. (2010). Structure of the central nervous system of a juvenile acoel, *Symsagittifera roscoffensis*. Dev. Genes Evol. 1–16.
- Boguta K.K. 1970. A new representative of turbellarians (Acoela) *Anaperus biaculeatus* sp. n. from the sublittoral of the White Sea. Zool. Zh. 49: 198–209.
- Brauner K. 1920. *Monchoerues chuni*, *Monchoerues boehmigi* und *Convoluta dubia*. Drei neue Turbellaria Acoela aus den Fangergebnissen der deutschen Tiefsee-Expedition des Jahres 1898. Zool. Anz. 52: 31–37.
- Budd G.E. 2008. The earliest fossil record of the animals and its significance. Philos. Trans. R. Soc. Lond. B. 363:1425–1434.
- Bush L.F. 1975. Biology of *Neochildia fusca* n. gen., n. sp. from the north-eastern coast of the United States (Plathelminthes: Turbellaria). Biol. Bull. 148:35–48.
- Correa D.D. 1960. Two new marine Turbellaria from Florida. Bull. Mar. Sci. Gulf Caribb. 10:208–216.
- Costello D.P., Henley C., Ault C.R. 1969. Microtubules in spermatozoa of *Childia* (Turbellaria, Acoela) revealed by negative staining. Science. 163:678–679.
- Crezée M. 1975. Monograph of the Solenofilomorphidae (Turbellaria, Acoela). Int. Rev. Gesamten. Hydrobiol. 60:769–845.
- Dörjes J. 1966. *Paratomella unichaeta* nov. gen. nov. spec., Vertreter einer neuen Familie der Turbellaria Acoela mit asexueller Fortpflanzung durch Paratomie. Veröff. Inst. Meeresforsch. Bremerh. Sonderb. 2:187–200.
- Dörjes J. 1968. Die Acoela (Turbellaria) der Deutschen Nordseeküste und ein neues system der Ordnung. Z. Zool. Syst. Evolutionsforsch. 6:56–452.
- Dörjes J., Karling T.G. 1975. Species of Turbellaria Acoela in the Swedish Museum of Natural History, with remarks on their anatomy, taxonomy and distribution. Zool. Scr. 4:175–189.
- Dunn C.W., Hejnol A., Matus D.Q., Pang K., Browne W.E., Smith S.A., Seaver E., Rouse G.W., Obst M., Edgecombe G.D., Sorensen M.V., Haddock S.H.D., Schmidt-Rhaesa A., Okusu A., Kristensen R.M., Wheeler W.C., Martindale M.Q., Giribet G. 2008. Broad phylogenomic sampling improves resolution of the animal tree of life. Nature. 452:745–749.
- Egger B., Steinke D., Tarui H., De Mulder K., Arendt D., Borgonie G., Funayama N., Gschwentner R., Hartenstein V., Hobmayer B., Hooge M., Hrouda M., Ishida S., Kobayashi C., Kuales G., Nishimura O., Pfister D., Rieger R., Salvenmoser W., Smith J., Technau U., Tyler S., Agata K., Salzburger W., Ladurner P. 2009. To be or not to be a flatworm: the acoel controversy. PLoS One 4:e5502.
- Ehlers U. 1985. Das Phylogenetische System der Plathelminthes. Stuttgart (Germany): G. Fischer.
- Ewing B., Green P. 1998. Base-calling of automated sequencer traces using phred. II. Error probabilities. Genome Res. 8:186–194.
- Ewing B., Hillier L., Wendl M.C., Green P. 1998. Base-calling of automated sequencer traces using phred. I. Accuracy assessment. Genome Res. 8:175–185.
- Faubel A. 1974. Die Acoela (Turbellaria) eines Sandstrandes der Nordseeinsel Sylt. Mikrofauna Meeresb. 32:1–58.
- Faubel A. 1976. Interstitielle Acoela (Turbellaria) aus dem Litoral der nordfriesischen Inseln Sylt und Amrum (Nordsee). Mitt. Hamb. Zool. Mus. Inst. 73:17–56.
- Faubel A., Regier S. 1983. Anatomy and taxonomy of new Acoela (Turbellaria) from the sublittoral of the northern North Sea. Senckenbergiana Marit. 15:1–17.
- Folmer O., Black M., Hoeh W., Lutz R., Vrijenhoek R. 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I form diverse metazoan invertebrates. Mol. Mar. Biol. Biotechnol. 3:294–299.
- Gaerber C.W., Salvenmoser W., Rieger R.M., Gschwentner R. 2007. The nervous system of *Convolutriloba* (Acoela) and its patterning during regeneration after asexual reproduction. Zoomorphology. 126: 73–87.
- Goloboff P. 1999. Analyzing large data sets in reasonable times: solutions for composite optima. Cladistics. 15:415–428.

- Goloboff P.A., Farris J.S., Nixon K.C. 2008. TNT, a free program for phylogenetic analysis. *Cladistics*. 24:774–786.
- Graff L.v. 1882. Monographie der Turbellarien I. Rhabdoceolida. Leipzig (Germany): Verlag Wilhelm Engelmann. p. 1–442.
- Graff L.v. 1891. Die Organisation der Turbellaria Acoela. Leipzig (Germany): Verlag Von Wilhelm Engelmann. p. 1–90.
- Graff L.v. 1905. Turbellaria I. Acoela. In: Schulze F.E., editor. Das Tierreich. No. 23. Berlin (Germany): Kniglich preussischen Akademie der Wissenschaften zu Berlin.
- Graff L.v. 1911. Acoela, Rhabdoceola und Alloeoceola des Ostens der Vereinigten Staaten von Amerika. *Z. Wiss. Zool.* 99:321–428.
- Gray J.S., Rieger R.M. 1971. A quantitative study of the meiofauna of an exposed sandy beach at Robin Hood's Bay, Yorkshire. *J. Mar. Biol. Assoc. UK.* 51:1–19.
- Gschwentner R., Baric S., Rieger R. 2002. New model for the formation and function of sagittocysts: *Symsagittifera corsicae* n. sp. (Acoela). *Invertebr. Biol.* 121:95–103.
- Gschwentner R., Mueller J., Ladurner R., Rieger R., Tyler S. 2003. Unique patterns of longitudinal body-wall musculature in the Acoela (Platyhelminthes): the ventral musculature of *Convolutiloba longifissura*. *Zoomorphology*. 122:87–94.
- Guindon S., Gascuel O. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Syst. Biol.* 52:696–704.
- Hejnal A., Martindale M.Q. 2008. Acoel development indicates the independent evolution of the bilaterian mouth and anus. *Nature*. 456:382–386.
- Hejnal A., Obst M., Stamatakis A., Ott M., Rouse G.W., Edgecombe G.D., Martinez P., Baganà J., Bailly X., Jondelius U., Wiens M., Müller W.E., Seaver E., Wheeler W.C., Martindale M.Q., Giribet G., Dunn C.W. 2009. Assessing the root of bilaterian animals with scalable phylogenomic methods. *Proc. Biol. Sci.* 276:4261–70.
- Hendelberg J. 1969. On the development of different types of spermatozoa from spermatids with two flagella in the Turbellaria with remarks on the ultrastructure of the flagella. *Zool. Bidr. Upps.* 38:1–50.
- Hendelberg J. 1974. Spermiogenesis, sperm morphology, and biology of fertilization in the Turbellaria. In: Riser N.W., Morse M.P., editors. *Biology of the Turbellaria*. New York: McGraw Hill. p. 148–164.
- Hendelberg J. 1975. Functional aspects of flatworm sperm morphology. In: Afzelius B.A., editor. *The functional anatomy of the spermatozoon*. Oxford: Pergamon. p. 299–309.
- Hendelberg J. 1977. Comparative morphology of turbellarian spermatozoa studied by electron microscopy. *Acta Zool. Fenn.* 154:149–162.
- Hendelberg J., Åkesson B. 1988. *Convolutiloba retrogenemma* gen. et sp. n., a turbellarian (Acoela, Platyhelminthes) with reversed polarity of reproductive buds. *Forts. Zool.* 36:321–327.
- Henley C. 1968. Refractile bodies in the developing and mature spermatozoa of *Childia groenlandica* (Turbellaria, Acoela) and their possible significance. *Biol. Bull.* 134:382–397.
- Henley C. 1974. Platyhelminthes (Turbellaria). In: Giese A.C., Pearse J.S., editors. *Reproduction of marine invertebrates*. Volume 1. Acoelomate and pseudocoelomate metazoans. New York: Academic Press. p. 267–343.
- Henley C., Costello D.P. 1969. Microtubules in spermatozoa of some turbellarian flatworms. *Biol. Bull.* 137:405.
- Henley C., Costello D.P., Ault C.R. 1968. Microtubules in the axial filament complexes of Acoel turbellarian spermatozoa as revealed by negative staining. *Biol. Bull.* 135:422–423.
- Hofacker I.L., Fekete M., Stadler P.F. 2002. Secondary structure prediction for aligned RNA sequences. *J. Mol. Biol.* 319:3724–3732.
- Hooge M.D. 2001. Evolution of body-wall musculature in the Platyhelminthes (Acoelomorpha, Catenulida, Rhabditophora). *J. Morphol.* 249:171–194.
- Hooge M.D. 2003. Two new families, three new genera, and four new species of acoel flatworms (Acoela, Platyhelminthes) from Queensland, Australia. *Cah. Biol. Mar.* 44:275–298.
- Hooge M.D., Eppinger N. 2005. New species of Acoela (Acoelomorpha) from the Gulf of California. *Zootaxa*. 1009:1–14.
- Hooge M.D., Haye P., Tyler S., Litvaitis M.K., Kornfield I. 2002. Molecular systematics of the Acoela (Acoelomorpha, Platyhelminthes) and its concordance with morphology. *Mol. Phylogenet. Evol.* 24:333–342.
- Hooge M.D., Rocha C.E.F. 2006. Acoela (Acoelomorpha) from the northern beaches of the state of São Paulo, Brazil, and a systematic revision of the family Otocelididae. *Zootaxa*. 1335:1–50.
- Hooge M.D., Smith J.P.S., III. 2004. New acoels (Acoela, Acoelomorpha) from North Carolina. *Zootaxa*. 442:1–24.
- Hooge M.D., Tyler S. 1999. Body-wall musculature in *Praeconvoluta tornuva*, n. sp. and the use of muscle patterns in taxonomy of acoel turbellarians. *Invertebr. Biol.* 118:8–17.
- Hooge M.D., Tyler S. 2003. Acoels (Platyhelminthes, Acoela) from the Atlantic coast of North America. *Meiofauna Mar.* 12:7–36.
- Hooge M.D., Tyler S. 2005. New tools for resolving phylogenies: a systematic revision of the Convolutidae (Acoelomorpha, Acoela). *J. Zool. Syst. Evol. Res.* 43:100–113.
- Hooge M.D., Tyler S. 2008. Acoela (Acoelomorpha) from Bocas del Toro, Panama. *Zootaxa*. 1719:1–40.
- Huelsenbeck J.P., Ronquist F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics*. 17:754–755.
- Hyman L.H. 1939. Acoel and polyclad Turbellaria from Bermuda and the Sargassum. *Bull. Bingham. Oceanogr. Coll.* 7:1–26.
- Jondelius U., Ruiz-Trillo I., Baguna J., Riutort M. 2002. The Nemeritodermatida are basal bilaterians and not members of the Platyhelminthes. *Zool. Scr.* 31:201–215.
- Kass R.E., Raftery A.E. 1995. Bayes factors. *J. Am. Stat. Assoc.* 90, 773–795.
- Katoh K., Asimenos G., Toh H. 2009. Multiple alignment of DNA sequences with MAFFT. *Methods Mol. Biol.* 537:39–64.
- Katoh K., Toh H. 2008. Improved accuracy of multiple ncRNA alignment by incorporating structural information into a MAFFT-based framework. *BMC Bioinformatics*. 9:212.
- Kostenko A.G. 1989. The position of *Taurida fulvomaculata* (Ax, 1959) in the system of Acoela. *Taurididae* fam. nov. *Trudy Zool. Inst. Leningrad.* 195:14–25.
- Kostenko A.G., Mamkaev Yu.V. 1990. The position of green convolutids in the system of acoel turbellarians (Turbellaria, Acoela). 1. *Simsagittifera* gen. n. 2. *Sagittiferidae* fam. n. *Zool. Zh.* 69(6): 11–21; 69(7):5–16.
- Kotikova E.A., Raikova O.I. 2008. Architectonics of the central nervous system of Acoela, Platyhelminthes, and Rotifera. *J. Evol. Biochem. Phys.* 44:95–108.
- Kozloff E.N. 1965. New species of the acoel turbellarians from the Pacific coast. *Biol. Bull.* 129:151–166.
- Levens G.M.R. 1879. Bidrag til Kundskab om Gronlands Turbellariefauna. *Vidensk. Meddr. Dansk. Naturhist. Foren.* 1879–1880 31:165–204.
- Luther A. 1912. Studien über acoele Turbellarien aus dem Finnischen Meerbusen. *Acta. Soc. Fauna Flora Fenn.* 36:2–62.
- Maddison D.R., Maddison W.P. 2001. MacClade 4 analysis of phylogeny and character evolution: version 4.03. Sinauer.
- Mamkaev Y.V. 1965. Etude morphologique d'*Actinoposthia beklemishevi* n. sp. (Turbellaria Acoela). *Cah. Biol. Mar.* 6:23–50.
- Mamkaev Y.V., Ivanov V.P. 1970. Electron microscopy investigation of spermatozoa of *Convoluta convoluta* (Turbellaria, Acoela). *Annual report. Zool. Inst. Leningrad*:12–13.
- Marcus E. 1948. Turbellaria do Brasil. *Bol. Fac. Fil. Ci Letr. U. Sao Paulo Zool.* 13:111–243.
- Marcus E. 1950. Turbellaria Brasileiros (8). *Bol. Fac. Fil. Ci Letr. U. Sao Paulo Zool.* 15:5–192.
- Marcus E. 1954. Turbellaria Brasileiros (11). *Papeis Aruhos Dep. Zool. Secr. Agric. Sao Paulo.* 11:419–489.
- Misof K., Misof A. 2009. Monte Carlo approach successfully identifies randomness of multiple sequence alignments. *Syst. Biol.* 58:21–34.
- Nylander J.A.A. 2004. MRAIC pl. Program distributed by the author. *Evolutionary Biology Centre, Uppsala University.*
- Nylander J.A., Wilgenbusch J.C., Warren D.L., Swofford D.L. 2008. AWTY (are we there yet?): a system for graphical exploration of MCMC convergence in Bayesian phylogenetics. *Bioinformatics*. 24:581–583.
- Oersted A.S. 1845. Fortegnelse over Dyr, samlede i Christianafjord ved Dobrak fra 21–24 Julie 1844. *Naturh. Tidessk. udgivet af H Kroyer.* 2:400–427.
- Ogunlana M.V., Hooge M.D., Tekle Y.I., Benayahu Y., Barneah O., Tyler S. 2005. *Waminia brickneri* n. sp. (Acoela: Acoelomorpha) associated with corals in the Red Sea. *Zootaxa*. 1008:1–11.

- Pagel M., Meade A. 2006. Bayesian analysis of correlated evolution of discrete characters by reversible-jump Markov chain Monte Carlo. *Am. Nat.* 167:808–825.
- Pagel M., Meade A., Barker D. 2004. Bayesian estimation of ancestral character states on phylogenies. *Syst. Biol.* 53:673–684.
- Papi F. 1957. Sopra un nuovo Turbellario arcooforo di particolare significato filetico e sulla posizione della fam. Hofsteniidae nel sistema dei Turbellari. *Pubbl. Staz. Zool. Napoli* 30:132–148.
- Peterson K.J., Cotton J.A., Gehling J.G., Pisani D. 2008. The Ediacaran emergence of bilaterians: congruence between the genetic and the geological fossil records. *Philos. Trans. R. Soc. Lond. B.* 363:1435–1443.
- Petrov A., Hooge M.D., Tyler S. 2004. Ultrastructure of sperms in Acoela (Acoelomorpha) and its concordance with molecular systematics. *Invertebr. Biol.* 123:183–197.
- Raikova O.I., Falleni A., Justine J.-L. 1997. Spermiogenesis in *Paratomella rubra* (Platyhelminthes, Acoela): ultrastructural, immunocytochemical, cytochemical studies and phylogenetic implications. *Acta Zool.* 78:295–307.
- Raikova O.I., Flyachinskaya L.P., Justine J.-L. 1998. Acoel spermatozoa: ultrastructure and immunocytochemistry of tubulin. *Hydrobiologia.* 383:207–214.
- Raikova O.I., Justine J.-L. 1994. Ultrastructure of spermiogenesis and spermatozoa in three Acoels (Platyhelminthes). *Ann. Sci. Nat. Zool.* 15:63–75.
- Raikova O.I., Justine J.-L. 1999. Microtubular system during spermiogenesis and in the spermatozoon of *Convoluta saliens* (Platyhelminthes, Acoela): tubulin immunocytochemistry and electron microscopy. *Mol. Reprod. Dev.* 52:74–85.
- Raikova O.I., Reuter M., Justine J.-L. 2001. Contributions to the phylogeny and systematics of the Acoelomorpha. In: Littlewood D.T.J., Bray R.A., editors. *Interrelationships of the Platyhelminthes*. The Systematics Association Special Volume 60. London: Taylor & Francis. p. 13–23.
- Ramachandra N.B., Gates R.D., Ladurner P., Jacobs D.K., Hartenstein V. 2002. Embryonic development in the primitive bilaterian *Neochildia fusca*: normal morphogenesis and isolation of POU genes Brn-1 and Brn-3. *Dev. Genes Evol.* 212:55–69.
- Reuter M., Raikova O.I., Jondelius U., Gustafsson M.K.S., Maule A.G., Halton D.W. 2001. Organisation of the nervous system in the Acoela: an immunocytochemical study. *Tissue Cell.* 33:119–128.
- Ronquist F. 2004. Bayesian inference of character evolution. *Trends Ecol. Evol.* 19:475–481.
- Ronquist F., Huelsenbeck J.P. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics.* 19:1572–1574.
- Ruiz-Trillo I., Paps J., Loukota M., Ribera C., Jondelius U., Baguna J., Riutort M. 2002. A phylogenetic analysis of myosin heavy chain type II sequences corroborates that Acoela and Nemertodermatida are basal bilaterians. *Proc. Natl Acad. Sci. U.S.A.* 99:11246–11251.
- Ruiz-Trillo I., Riutort M., Littlewood D.T.J., Herniou E., Baguna J. 1999. Acoel flatworms: earliest extant bilaterian metazoans, not members of Platyhelminthes. *Science.* 283:1919–1923.
- Schultze M. 1851. *Beitraege zur Naturgeschichte der Turbellarien*. Greifswald (Germany): C.A. Koch's verlagshandlung.
- Semmler H., Bailly X., Wanninger A. 2008. Myogenesis in the basal bilaterian *Symsagittifera roscoffensis* (Acoela). *Front. Zool.* 5:14.
- Shannon T., Achatz J.G. 2007. *Convolutriloba macropyga* sp. nov., an uncommonly fecund acoel (Acoelomorpha) discovered in tropical aquaria. *Zootaxa.* 1525:1–17.
- Silveira M. 1967. Formation of structured secretory granules within the Golgi complex in an acoel Turbellarian. *J. Microscopie.* 6:95–100.
- Silveira M. 1969. Ultrastructural studies on a 'nine plus one' flagellum. *J. Ultrastr. Res.* 26:274–288.
- Smith J.P.S., Bush L. 1991. *Convoluta pulchra*, n. sp., (Turbellaria: Acoela) from the east coast of North America. *Trans. Am. Microsc. Soc.* 110:12–26.
- Smith S.A., Dunn C.W. 2008. Phyutility: a phyloinformatics tool for trees, alignments and molecular data. *Bioinformatics.* 24: 715–716.
- Staden R. 1996. The Staden sequence analysis package. *Mol. Biotechnol.* 5:233–241.
- Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics.* 22:2688–2690.
- Stamatakis A., Hoover P., Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML web servers. *Syst. Biol.* 57:758–771.
- Steinböck O. 1931. Marine Turbellaria. In: Jensen S., Lundbeck W., Mortensen Th., editors. *Zoology of the Faroes*. Volume 8. Copenhagen (Denmark): A.F. Høst & Søn p. 1–26.
- Tabei Y., Kiryu H., Kin T., Asai K. 2008. A fast structural multiple alignment method for long RNA sequences. *BMC Bioinformatics.* 9:33.
- Tekle Y.I. 2004. A new *Haploposthia* species (Acoela) from the Swedish west coast. *Sarsia.* 89:85–90.
- Tekle Y.I., Raikova O.I., Ahmadzadeh A., Jondelius U. 2005. Revision of the Childiidae (Acoela), a total evidence approach in reconstructing the phylogeny of acoels with reversed muscle layers. *J. Zool. Syst. Evol. Res.* 43:72–90.
- Tekle Y.I., Raikova O.I., Jondelius U. 2006. A new viviparous acoel *Childia vivipara* sp. nov. with observations on the developing embryos, sperm ultrastructure, body wall and stylet musculatures. *Acta Zool.* 87:121–130.
- Tekle Y.I., Raikova O.I., Justine J.-L., Hendelberg J., Jondelius U. 2007. Ultrastructural and immunocytochemical investigation of acoel sperms with 9 + 1 axoneme structure: new sperm characters for unraveling phylogeny in Acoela. *Zoomorphology.* 126:1–16.
- Tekle Y.I., Raikova O.I., Justine J.-L., Jondelius U. 2007. Ultrastructure and tubulin immunocytochemistry of the copulatory stylet-like structure in *Childia* species (Acoela). *J. Morphol.* 268: 166–180.
- Telford M.J., Lockyer N.E., Cartwright-Finch C., Littlewood D.T.J. 2003. Combined large and small subunit ribosomal RNA phylogenies support a basal position of the acoelomorph flatworms. *Proc. R. Soc. Lond. B.* 270:1077–1083.
- Todt C. 2009. Structure and Evolution of the Pharynx Simplex in Acoel Flatworms (Acoela). *J. Morphol.* 270:271–290.
- Tyler S., Hooge M.D. 1999. Taxonomic account: *Praeconvoluta tornuva* n. sp. *Invertebr. Biol.* 118:15–17.
- Tyler S., Rieger R.M. 1999. Functional morphology of musculature in the acoelomate worm, *Convoluta pulchra* (Plathelminthes). *Zoomorphology.* 119:127–141.
- Uljanin W. 1870. The Turbellaria from the Bay of Sebastopol. *Commun. Soc. Sci. Moscow* 4:1–96.
- Wallbor A., Curini-Galletti M., Ahmadzadeh A., Jondelius U. 2007. Dismissal of Acoelomorpha: Acoela and Nemertodermatida are separate early bilaterian clades. *Zool. Scr.* 36:509–523.
- Westblad E. 1940. Studien über skandinavische Turbellaria Acoela. I. *Ark. Zool.* 32A:1–28.
- Westblad E. 1942. Studien über skandinavische Turbellaria Acoela. II. *Ark. Zool.* 33A:1–48.
- Westblad E. 1945. Studien über skandinavische Turbellaria Acoela. III. *Ark. Zool.* 36A:1–56.
- Westblad E. 1946. Studien über skandinavische Turbellaria Acoela. IV. *Ark. Zool.* 38A:1–56.
- Westblad E. 1948. Studien über skandinavische Turbellaria Acoela. V. *Ark. Zool.* 41:191–273.
- Winsor L. 1990. Marine Turbellaria (Acoela) from North Queensland. *Mem. Queensland Mus.* 28:785–800.
- Yamasu T. 1982. Five new species of acoel flat worms from Japan. *Galaxea.* 1:29–43.
- Yoder M.J., Dole K., Seltmann K., Deans A. 2006-Present. Mx, a collaborative web based content management for biological systematists.